ANTI-NME ANTIBODY

BACKGROUND OF THE INVENTION

[0001] 1. Field of the Invention:

[0002] The present application relates to NME proteins, peptides derived from NME proteins, and antibodies generated from the peptides thereof or antibody or antibody fragments selected by virtue of their ability to bind to said peptides. The present application also relates to treating or preventing diseases associated with the expression of NME in a patient.

[0003] 2. General Background and State of the Art:

[0004] NDPK (nucleoside diphosphate protein kinase) proteins are a family of proteins grouped together because they all contain an NDPK domain. The first NME proteins discovered, previously called NM23 proteins, were NM23-H1 and NM23-H2. For decades it was unclear whether they induced differentiation or prevented differentiation of hematopoietic cells. The inventors previously discovered that NM23-H1 prevents differentiation when it is a dimer, which binds to the MUC1* growth factor receptor, but at higher concentrations NM23-H1 becomes a hexamer, which does not bind to MUC1*, and it induces differentiation. NM23 used to be called a metastasis suppressor when it was found that it was under-expressed in some very aggressive cancers. The present inventors previously disclosed that NM23-H1 dimers bind to and dimerize the extracellular domain of the MUC1* growth factor receptor that is over expressed on the vast majority of cancers and such binding promotes the growth of cancer cells. Conversely, at higher concentrations, NM23 forms tetramers and hexamers that do not bind to MUC1* and do not promote tumorigenesis. Very recently more NME family proteins (NME 1-10) have been discovered although until now, their functions have not been elucidated. NME7 is a newly discovered NME family protein, but its NDPK domain has no enzymatic activity, unlike other NME family members. NME7 is either not expressed at all in adult tissues or is expressed at extremely low levels.

SUMMARY OF THE INVENTION

[0005] The present application is directed to a method of treating or preventing cancer in a subject, comprising administering to the subject an antibody made against a member of the NME family. The NME family may be NME7 family. The antibody may bind to NME7. The

antibody may bind to NME7-AB or NME-AB-like protein. The antibody may bind to NME7-X1. The antibody may inhibit binding between NME7 and its cognate binding partner. The cognate binding partner may be MUC1*. The cognate binding partner may be PSMGFR portion of the MUC1* extracellular domain. In one aspect, the antibody may be generated or selected for its ability to bind to a peptide selected from those listed in Figures 16-19 (SEQ ID NOS:88 to 145). Preferably, the peptide may be selected from those listed in Figure 19 (SEQ ID NOS:141 to 145).

[0006] The peptide may be highly homologous to, or to which is added or subtracted up to 7, up to 6, up to 5, up to 4, up to 3, up to 2, or up to 1 amino acid residues at the N-terminus or C-terminus, of the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145). In one aspect, the antibody may be selected for its ability to bind to NME7-AB or NME7-X1 but not to NME1. The antibody may be polyclonal, monoclonal, bivalent, monovalent, bispecific, an antibody fragment containing the variable region, or an antibody mimic. The antibody may be human or humanized. The antibody may be a single chain scFv.

[0007] In another aspect, the invention is directed to a method of treating or preventing cancer in a subject, comprising administering to the subject a peptide that is highly homologous or identical to regions of NME7-AB. The peptide may be at least 80% homologous to one or more of the peptides listed in Figure 16. The peptide may be at least 80% homologous to one or more of the peptides listed in Figure 17. The peptide may be at least 80% homologous to one or more of the peptides listed in Figure 18. The peptide may be at least 80% homologous to one or more of the peptides listed in Figure 19. The peptide may be selected from peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145). The peptide may be selected from those listed in Figure 19 (SEQ ID NOS:141 to 145). Or, the peptide may be highly homologous to, or to which is added or subtracted up to 7 , up to 6, up to 5, up to 4, up to 3, up to 2, or up to 1 amino acid residues at the N-terminus or C-terminus, of the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145). The peptide may be connected to another peptide via a spacer or linker.

[0008] In another aspect, the invention is directed to a chimeric antigen receptor (CAR), for the treatment or prevention of cancer wherein the targeting extracellular portion of the CAR comprises at least a peptide fragment of a member of the NME family. NME family may be NME7 family. The member of the NME7 family may be NME7. Or, the member of the NME7 family may be NME7-AB or NME-AB-like protein. The member of the NME7 family may be also NME7-X1. The targeting extracellular portion of the CAR may include a peptide of the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145). The peptide may be

selected from those listed in Figure 19 (SEQ ID NOS:141 to 145). The peptide may include a peptide, which is highly homologous to, or to which is added or subtracted up to 7, up to 6, up to 5, up to 4, up to 3, up to 2, or up to 1 amino acid residues at the N-terminus or C-terminus, of the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145). The peptide may be connected to another peptide via a spacer or linker.

[0009] In yet another aspect, the invention is directed to a method of treating or preventing cancer or cancer metastasis, comprising engineering the chimeric antigen receptor according to claim 3, into an immune system cell and administering the cell to a subject in need thereof.

[0010] In another aspect, the invention is directed to a chimeric antigen receptor (CAR), for the treatment or prevention of cancer, wherein the targeting extracellular portion of the chimeric antigen receptor comprises a portion of an antibody that binds to NME7-AB, NME-AB-like protein or NME7-X1. The portion of the antibody may be a single chain scFv or may be human or humanized.

[0011] In yet another aspect, the invention is directed to a method of vaccinating a person against cancer or metastatic cancer comprising immunizing the person with a peptide fragment of a member of the NME family. The NME family may be NME7 family. The member of the NME7 family may be NME7 or NME7b. The member of the NME7 family may be NME7-AB or NME7-AB-like protein. The NME7 family may be NME7-X1. The immunizing peptide may be a peptide from the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145). Preferably, the peptide may be selected from those listed in Figure 19 (SEQ ID NOS:141 to 145). The immunizing peptide may include a peptide, which is highly homologous to, or to which is added or subtracted up to 7, up to 6, up to 5, up to 4, up to 3, up to 2, or up to 1 amino acid residues at the N-terminus or C-terminus, of the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145). The immunizing peptide may be connected to another peptide via a spacer or linker.

[0012] In yet another aspect, the invention is directed to a method of treating or preventing cancer in a subject, comprising administering to the subject a nucleic acid that inhibits the expression of NME7, NME7b, NME7-AB-like protein or NME7-X1. The nucleic acid may be an anti-sense nucleic acid that suppresses expression of NME7, NME7-AB-like protein or NME7-X1. The nucleic acid may be an inhibitory RNA, siRNA, RNAi, or shRNA that inhibits expression of NME7, NME7-AB-like protein or NME7-X1.

[0013] In another aspect, the invention is directed to a method of treating or preventing cancer in a subject, comprising administering to the subject genetically edited nucleic acids

that inhibit expression of NME7, NME7b, NME7-AB-like protein or NME7-X1. The genetically edited nucleic acids that inhibit expression of NME7, NME7b, NME7-AB-like protein or NME7-X1 may be inserted into cells that may be then administered to the patient. The genetically edited nucleic acids that inhibit expression of NME7, NME7b, NME7-AB-like protein or NME7-X1 may be inserted into cells using a viral vector. The viral vector may be a lentiviral system.

[0014] In another aspect, the invention is directed to a method of growing cancer cells comprising contacting the cells with NME7-AB, NME7b, NME7-AB-like protein or NME7-X1, 2i or 5i. The method may include culturing the cells in a medium that contains NME7-AB, NME7b, NME7-AB-like protein or NME7-X1, 2i or 5i, or growing cells in an animal that expresses human NME7-AB, NME7b, NME7-AB-like protein or NME7-X1, or to which NME7-AB, NME7b, NME7-AB-like protein or NME7-X1 is administered. The cancer cells may be breast, prostate, ovarian, colorectal, pancreatic, liver, melanoma or brain cancer cells. Drug candidates may be tested on the cells. Th efficacy of the drugs may be assessed by comparing cancer growth to a no drug control or comparing expression levels of metastatic markers or stem cell markers to a no drug control or comparing the ability of the resultant cells to form tumors in animals from low cell copy number compared to a no drug control and determining the efficacy of a candidate drug for the treatment of cancer or metastasis. The cells may be obtained from a patient being assessed for treatment for cancer and drugs that would be effective for that patient are selected based on results using methods described above. The cells may not be obtained from a patient being assessed for treatment for cancer but drugs that would be effective for that patient are selected based on results using the methods described above.

[0015] In another aspect, the invention is directed to a method of generating antibodies or antibody-like molecules from peptides or peptide mimics having a sequence derived from the sequence of NME. The NME may be NME7. The peptide may be used as an immunogen to generate antibodies or antibody-like molecules. The peptide may be administered to an animal to generate anti-NME7 antibodies. The peptide may be administered to a human to generate anti-NME7 antibodies. The peptide may have a sequence listed in Figures 16 to 19 (SEQ ID NOS:88 to 145). Preferably, the peptide may be selected from those listed in Figure 19 (SEQ ID NOS:141 to 145). The peptide may include a peptide, which is highly homologous to, or to which is added or subtracted up to 7, up to 6, up to 5, up to 4, up to 3, up to 2, or up to 1 amino acid residues at the N-terminus or C-terminus, of the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145).

[0016] In another aspect, the invention is directed to a method of detecting presence of cancer or the progression of cancer, comprising the steps of:

[0017] 1) obtaining a sample from a patient having cancer or at risk of developing a cancer;

[0018] 2) subjecting that sample to an assay capable of detecting or measuring levels of a member of the NME7 family, or levels of nucleic acids encoding a member of the NME7 family;

[0019] 3) comparing levels of the measured member of the NME7 family or the member of the NME7 family-encoding nucleic acids in the test sample to levels in control patients or control cells;

[0020] 4) determining that the levels of the member of the NME7 family or nucleic acids encoding the member of the NME7 family are elevated compared to the controls; and

[0021] 5) concluding that the donor of the test sample has cancer or has had a progression of cancer if the control to which the test was compared came from a donor previously diagnosed with a cancer. In this method, the detection of the member of the NME7 family in circulation or in a tissue may be an indicator of cancer in a patient. The member of the NME7 family may be NME7, NME7b, NME7-X1, or NME7-AB-like protein.

[0022] In yet another aspect, the invention is directed to a method comprising:

[0023] detecting presence of a member of the NME7 family or MUC1* in a patient; and

[0024] administering anti-NME7 or anti-MUC1* antibody or antibodies to the patient exhibiting the member of the NME7 family or MUC1* expression. The member of the NME7 family may be NME7, NME7b, NME7-X1, or NME7-AB-like protein.

[0025] In yet another aspect, the invention is directed to a method for treating or preventing cancer comprising:

[0026] 1) obtaining a sample from a patient suspected of having a cancer or at risk of developing a cancer or at risk of developing a metastatic cancer;

[0027] 2) measuring an amount of the member of an NME7 family or a member of the NME7 family encoding nucleic acid, wherein the measured levels are significantly above those measured in a control sample;

[0028] 3) determining that the patient has a cancer or has developed a more aggressive or a metastatic cancer;

[0029] 4) administering to the patient an effective amount of a therapeutic agent that suppresses expression of the member of the NME7 family, inhibits cleavage of NME7 or inhibits NME7 binding to its targets. The target of the member of the NME7 family may be

MUC1*. The target of the member of the NME7 family may be PSMGFR portion of the MUC1* extracellular domain. The member of the NME7 family may be NME7, NME7b, NME7-X1, or NME7-AB-like protein.

[0030] In any of the methods above regarding cancer, cancer may include breast, prostate, ovarian, colorectal, pancreatic, liver, melanoma or brain cancer.

BRIEF DESCRIPTION OF THE DRAWINGS

[0031] The present invention will become more fully understood from the detailed description given herein below, and the accompanying drawings which are given by way of illustration only, and thus are not limitative of the present invention, and wherein;

[0032] **Figures 1A-1D.** Photos of Western blot gels showing the expression of NME1 or NME7 in the cell lysate of: 1) BGO1V human embryonic stem cells cultured in NM23-H1 dimers over a surface coated with a MUC1* antibody surface (MN-C3 mab); 2) BGO1V human embryonic stem cells cultured according to standard protocol in bFGF over a layer of mouse feeder cells (MEFs); 3) T47D breast cancer cells cultured by standard method in RPMI media; and 4) recombinant human NM23-H1 wild type, "wt" (A, B). Bottom row (C, D) shows the results of a "pull-down" or an immuno-precipitation assay in which the cell lysates were separately incubated with beads to which was added an antibody to the MUC1 cytoplasmic tail, "Ab-5". Species captured by binding to the MUC1* peptide were separated by SDS-PAGE and blotted with antibodies against each respective NM23 protein. Same experiments were conducted with NME6 but data is not shown.

[0033] **Figures 2A-2E** show photos of Western blots in which cell lysates from T47D breast cancer cells, BGO1V and HES-3 human ES cells and human SC101-A1 iPS cells were probed for the presence of NME1, NME6 or NME7. NME1 in all cell lines ran with an apparent molecular weight of ~17kDa (A). In all cell lines, NME7 ~33kDa species and the 42kDa species (C, E) could be detected in all but the HES-3 cell line (cultured in FGF). Species that reacted with an NME6-specific antibody were detected in all cell lines except the HES-3 cell line, when visualization was enhanced using Super Signal.

[0034] **Figures 3A-3C** show panels of photos of Western blots of human embryonic stem (ES) cells (A) and induced pluripotent stem (iPS) cells (B, C) probed for the presence of NME7. Western blots show the presence of three forms of NME7 in the cell lysates. One with an apparent molecular weight of ~42kDa (full length), ~33kDa (NME7-AB domains

devoid of the N-terminal DH domain) and a small ~25kDa species. However, only the lower molecular weight species are in the conditioned media (B).

[0035] **Figures 4A-4C.** (A) is an elution profile of size exclusion chromatography purification of NME7-AB; (B) is non-reducing SDS-PAGE gel from NME7-AB peak fractions; (C) is the elution profile of size exclusion chromatography of the purified NME7-AB.

[0036] **Figure 5** shows graph of HRP signal from ELISA sandwich assay showing NME7-AB dimerizes MUC1* extra cellular domain peptide.

[0037] **Figures 6A-6G** show photos of MUC1*-positive cancer cells treated with nothing (Row A), Taxol (Row B) or an anti-NME7 antibody (Rows C-E); a graph showing cell count in response to treatment at 48 hours (F), and a dot-blot used to estimate antibody concentration used in the cancer cell inhibition experiment (G).

[0038] **Figures 7A-7K** show the 48 hour results of an experiment using an anti-NME7 antibody to inhibit cancer cell growth. Photos of the cells cultured in media alone (A), taxol (B), or anti-NME7 at the concentrations indicated (C-J); a graph of cell number obtained using a calcein AM assay is shown (K).

[0039] **Figures 8A-8K** show the 96 hour results of an experiment using an anti-NME7 antibody to inhibit cancer cell growth. Photos of the cells cultured in media alone (A), taxol (B), or anti-NME7 at the concentrations indicated (C-J); a graph of cell number obtained using a calcein AM assay is shown (K). The graph and the photos show anti-NME7 antibodies inhibit cancer cell growth at concentrations as low as in the nanomolar range.

[0040] **Figure 9** is a photo of a Western blot wherein stem cell lysates (odd numbered lanes) or cell-conditioned media (even numbered lanes) were probed for the presence of NME7. iPS (induced pluripotent stem) cells were cultured in FGF over MEFs (lanes 1,2), NM23-H1 dimers over an anti-MUC1* antibody (C3) surface (lanes 3,4) or NME7 over an anti-MUC1* antibody (C3) surface (lanes 5-8). HES-3 (human embryonic stem) cells were cultured in FGF over MEFs (lanes 9,10), NM23-H1 dimers over an anti-MUC1* antibody (C3) surface (lanes 11,12) or NME7 over an anti-MUC1* antibody (C3) surface (lanes 13,14). Mouse embryonic fibroblast (MEFs) cells were also probed (lanes 15,16). The Western blot shows that the cell lysates contain an NME7 species with molecular weight of ~42kDa, which corresponds to the full-length protein. However, the secreted species runs with an apparent MW of ~33kDa, which corresponds to an NME7 species that is devoid of the N-terminal leader sequence.

[0041] **Figures 10A-10B** show photos of Western blots of various cell lysates and corresponding conditioned media probed for the presence of NME7 using a mouse monoclonal antibody (A) or another monoclonal antibody that only recognizes the N-terminal DM10 sequence (B). The lack of binding of the DM10 specific antibody to the ~33kDa NME7 species in the samples from the conditioned media of the cells indicates that the secreted form of NME7 is devoid of most if not all of the N-terminal DM10 leader sequence.

[0042] **Figure 11** is a graph of RT-PCR measurements of gene expression for stem cell markers and cancer stem cell markers for T47D cancer cells after being cultured in traditional media or a media containing NME7, wherein cells that became non-adherent (floaters) were analyzed separate from those that remained adherent.

[0043] **Figure 12** is a graph of RT-PCR measurements of gene expression for stem cell marker SOX2 and cancer stem cell marker CXCR4 for T47D cancer cells. Cells were cultured either in traditional media or a media containing NME1 dimers or NME7 (NME7-AB). Cell types that were separately analyzed were floating cells, cells plus Rho kinase inhibitor (+Ri), which made all cells adhere, or cells that remained adherent after floaters were removed which was in the absence of rho kinase inhibitor (- Ri).

[0044] **Figure 13** is a graph of RT-PCR measurements of gene expression for a variety of stem and putative cancer stem cell markers for DU145 prostate cancer cells. Cells were cultured either in traditional media or a media containing NME1 dimers ("NM23") or NME7 (NME7-AB). Rho kinase inhibitor was not used because by passage 2, cells remained adherent.

[0045] **Figures 14A-14B** are a graphs of RT-PCR measurement of the metastatic markers and pluripotent stem cell markers showing that the 2i inhibitors (GSK3-beta and MEK inhibitors) (A) that were previously shown to revert stem cells to a more naïve state or bacterial NMEs (B) that have high sequence homology to human NME1 or human NME7, also transform cancer cells to a more metastatic state.

[0046] **Figure 15** is a sequence alignment between human NME1 and human NME7-A or –B domain.

[0047] **Figure 16** lists immunogenic peptides from human NME7 with low sequence identity to NME1 and selected for their ability to generate therapeutic anti-NME7 antibodies for the treatment or prevention of cancers.

[0048] **Figure 17** lists immunogenic peptides from human NME7 that may be important for structural integrity or for binding to MUC1* selected for their ability to generate therapeutic anti-NME7 antibodies for the treatment or prevention of cancers.

[0049] **Figure 18** lists immunogenic peptides from human NME1 that may be important for structural integrity or for binding to MUC1* and selected for their ability to generate therapeutic anti-NME7 antibodies for the treatment or prevention of cancers.

[0050] **Figure 19** lists immunogenic peptides from human NME7 selected for their low sequence identity to NME1 and for their homology to bacterial NME1 proteins that have been implicated in cancers. These peptides are preferred for their ability to generate therapeutic anti-NME7 antibodies for the treatment or prevention of cancers. The peptides shown in this Figure include and added Cysteine covalently bound at the C-terminal end.

[0051] **Figure 20** shows photographs of two female athymice nu/nu mice out of 24 that were xenografted with only 50 human breast cancer cells that had first been grown for 7 days in NME7-AB and showed greatly increased expression of CXCR4, CHD1 and stem cell markers. In addition, half the mice were also injected daily with human recombinant NME7-AB. 82% of the mice that were also injected daily with NME7-AB developed remote metastases as well as tumors at the site of injection.

[0052] **Figure 21** shows a table of the results of the experiment in which mice were xenografted with cancer cells that were transformed to a more metastatic state by pre-culture in a medium containing human NME7-AB.

[0053] **Figure 22A** shows a graph of tumor volume measurements for four (4) groups of immune-compromised nu/nu female mice implanted with either 50, 100, 1,000 or 10,000 cells subcutaneously in the flank wherein the cells that were implanted were human MUC1-positive breast cancer cells that were cultured for seven (7) days in recombinant human NME7-AB wherein the 'floaters' were collected and verified to overexpress metastasis receptor CXCR4 by more than 100-fold. Half the mice in each group were injected daily with human recombinant NME7-AB. Numbers within the graph refer to the mouse tracking number. 'M' denotes a mouse with multiple tumors.

[0054] **Figure 22B** shows a graph of tumor volume measurements for four (4) groups of immune-compromised nu/nu female mice implanted with either 50, 100, 1,000 or 10,000 cells subcutaneously in the flank wherein the cells that were implanted were human MUC1-positive breast cancer cells that were cultured for seven (7) days in recombinant human NME7-AB wherein the 'floaters' were collected and verified to overexpress metastasis receptor CXCR4 by more than 100-fold. Half the mice in each group were injected daily with human recombinant NME7-AB. Of the mice that received daily injections of NME7-AB, 80% developed multiple tumors. This graph shows the combined volumes of multiple

tumors in the same mouse. Numbers within the graph refer to the mouse tracking number. 'M' denotes a mouse with multiple tumors.

[0055] **Figure 23** shows Western blots of primary tumors as well as the remote bumps on mice xenografted with human breast cancer cells that were transformed to a more metastatic state by pre-culture in a medium containing human NME7-AB. Westerns show that the remote bumps were human breast tumors as VU4H5 antibody only stains human MUC1, not murine.

[0056] **Figure 24** shows Western blots of primary tumors on mice xenografted with human breast cancer cells that were transformed to a more metastatic state by pre-culture in a medium containing human NME7-AB. Westerns show that the visible bumps are human breast tumors as VU4H5 antibody only stains human MUC1, not murine.

[0057] **Figure 25** shows Western blots of organs harvested from mice xenografted with human breast cancer cells that were transformed to a more metastatic state by pre-culture in a medium containing human NME7-AB. Westerns show that some mice that did not appear to have remote tumors, have human MUC1-positive cancer in some of their organs.

[0058] **Figures 26A-26B** show graphs of ELISA assays in which either NME7-AB (A) or NME1 (B) is adsorbed to the plate and anti-NME7 antibodies generated by NME7 peptides A1, A2, B1, B2 and B3 are tested for their ability to bind to NME7 but not to NME1. C20 is an anti-NME1 antibody.

[0059] **Figure 27** shows graphs of ELISA assays in which anti-NME7 antibodies generated are tested for their ability to inhibit binding of NME7-AB to a surface immobilized MUC1* peptide but not inhibit binding of NME1.

[0060] **Figure 28** shows a graph of a cancer cell growth experiment in which breast cancer cells were grown in the presence or absence of NME7 antibodies or short peptides derived from NME7, which were used to generate or select the antibodies. In addition, an antibody generated by immunization with nearly the entire NME7-AB peptide, amino acids 100-376, was shown to inhibit cancer cell growth.

[0061] **Figure 29** shows a graph of a cancer cell growth experiment in which breast cancer cells were grown in the presence or absence of combinations of NME7 antibodies or combinations of the short peptides derived from NME7, which were used to generate or select the antibodies. Both antibodies as well as their immunizing NME7-AB peptides inhibited growth of cancer cells.

[0062] **Figure 30** shows a table of scientist observations when cancer cells were grown in either NME7-AB or 2i inhibitors, which both are able to transform cancer cells to a more

metastatic state, and in the presence or absence of NME7 derived peptides A1, A2, B1, B2 and B3. The NME7-AB peptides inhibited the transition of adherent cancer cells to the floater cells, which RT-PCR measurements show have increased expression of metastatic markers, especially CXCR4.

[0063] **Figures 31A-31C** show a graph of RT-PCR measurements of CXCR4 expression in T47D breast cancer cells that were grown in either NME7-AB or 2i inhibitors, each of which transform cancer cells to a more metastatic state, and the inhibitory effect of anti-NME7 antibodies on the metastatic transformation (A). A graph of RT-PCR measurements of CXCR4, CHD1 and SOX2 expression in T47D breast cancer cells that were grown in 2i inhibitors for 72 hours or 144 hours, shows that the NME7-AB immunizing peptides are themselves inhibitory to the metastatic transformation. Peptides A1, A2 and B1 which were used in the inhibitory Combo 2 and 3 in part (A) are also inhibitory as peptides. Peptide B3 is the most inhibitory and is the immunizing peptide for antibody 61 which was the most inhibitory antibody tested in part (A). In part (C), the scale of the Y-axis of the graph of part (B) is reduced.

[0064] **Figure 32** shows a table of recorded RNA levels in samples that were used for RT-PCR measurement of CXCR4 in Figure 31 as well as the threshold cycle number for CXCR4 expression as well as for the control housekeeping gene.

[0065] **Figure 33** shows a graph of RT-PCR measurement of the expression of NME7-X1 in a panel of human stem cells and cancer cells.

[0066] **Figure 34** shows a graph of RT-PCR measurement of the expression of NME7, NME7a, NME7b and NME7-X1 in a panel of human stem cells and cancer cells. NME7a is full-length NME7, NME7b is missing a small portion of the DM10 domain, NME7-X1 is missing all of the DM10 domain and a small portion of the N-terminus of the first NDPK A domain. The bar labeled NME7 means that primers were used that detected both NME7a and NME7b.

[0067] **Figures 35A-35C** show photographs of Western blots in which various cancer cell lines are probed for expression of NME7 species using antibodies generated by immunization with NME7 derived short peptides.

[0068] **Figures 36A-36B** show photographs of Western blots in which various cancer cell lines are probed for expression of NME7 species using commercially available antibodies.

[0069] **Figures 37A-37C** show graphs of RT-PCR measurements of metastatic markers in cancer cells after being cultured in a serum-free media containing NME7-AB compared to the standard media. A) SK-OV3, a MUC1-positive ovarian cancer cell line increased

expression of metastatic markers CXCR4, CDH1 aka E-cadherin, SOX2 and NME7-X1; B) OV-90 a MUC1-negative ovarian cancer cell line increased expression of metastatic markers CXCR4 and NME7-X1; C) MDA-MB a breast cancer cell line that expresses minimal levels of MUC1 increased expression of metastatic markers CDH1 aka E-cadherin and SOX2.

[0070] **Figures 38A-38F** show photographs of Western blots and cancer growth graphs. A) various cancer cell lines are probed for the expression of full-length MUC1 using an anti-tandem repeat antibody VU4H5. B) various cancer cell lines are probed for the expression of cleaved form MUC1* using anti-PSMGFR antibody. C) various cancer cell lines are probed for the expression of NME7 species using a commercially available anti-NME7 antibody B9, showing full-length NME7 as well as a 33kDa and 30kDa species, consistent with a naturally occurring NME7-AB-like species as well as NME7-X1. D) HER2 positive BT-474 breast cancer cells express little to no MUC1 or MUC1* until they acquire resistance to chemotherapy drugs and metastasize. Parent cells were made resistant to Herceptin, Taxol, Doxorubicin and cyclophosphamide by culturing the cells in sub-lethal levels of the drug. Part (D) shows that the expression level of HER2 has not changed but expression of MUC1* has dramatically increased. E) shows a graph of the growth of the parent BT-474 cells compared to the drug resistant metastatic cells in response to treatment with Herceptin in the presence or absence of an anti-MUC1* Fab. F) shows a graph of the growth of the parent BT-474 cells compared to the drug resistant metastatic cells in response to treatment with Taxol in the presence or absence of an anti-MUC1* Fab.

[0071] **Figures 39A-39E** show photographs of Western blots of a co-immunoprecipitation experiment. T47D breast cancer cell extracts were incubated with an antibody against the MUC1 cytoplasmic tail, Ab-5, or a control antibody, IgG, and co-immunoprecipitated. The gels were blotted with two different commercially available anti-NME7 antibodies B9 (A) and CF7 (B). Both gels show unique NME7 bands at ~33kDa and ~30kDa. The gels were stripped and re-probed with an antibody against the extracellular domain of MUC1*, anti-PSMGFR (C) and (D), which shows that the NME7 species and MUC1* interact. A recombinant NME7-AB and a recombinant NME7-X1 were mixed together and run on a gel, then probed with an anti-NME7 antibody, showing that the two unique NME7 species that are naturally occurring in breast cancer cells and that interact with MUC1* are an NME7-AB-like species and NME7-X1 (E).

[0072] **Figures 40A-40C** show photographs of Western blots of a co-immunoprecipitation experiment. Human induced pluripotent stem, iPS7, or embryonic stem, HES3, cell extracts were incubated with an antibody against the MUC1 cytoplasmic tail, Ab-

5, or a control antibody, IgG, and co-immunoprecipitated. The gel was blotted with a commercially available anti-NME7 antibody B9 (A). Both cell types show unique NME7 bands at ~33kDa and ~30kDa. The gel was stripped and re-probed with an antibody against the extracellular domain of MUC1*, anti-PSMGFR (B), which shows that the NME7 species and MUC1* interact. A recombinant NME7-AB and a recombinant NME7-X1 were mixed together and run on a gel, then probed with an anti-NME7 antibody, showing that the two unique NME7 species that are naturally occurring in breast cancer cells and that interact with MUC1* are an NME7-AB-like species and NME7-X1 (C).

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

[0073] Definitions

[0074] In the present application, “a” and “an” are used to refer to both single and a plurality of objects.

[0075] As used herein, “about” or “substantially” generally provides a leeway from being limited to an exact number. For example, as used in the context of the length of a polypeptide sequence, “about” or “substantially” indicates that the polypeptide is not to be limited to the recited number of amino acids. A few amino acids add to or subtracted from the N-terminus or C-terminus may be included so long as the functional activity such as its binding activity is present.

[0076] As used herein, administration "in combination with" one or more further therapeutic agents includes simultaneous (concurrent) and consecutive administration in any order.

[0077] As used herein, "amino acid" and "amino acids" refer to all naturally occurring L-α-amino acids. This definition is meant to include norleucine, ornithine, and homocysteine.

[0078] As used herein, in general, the term "amino acid sequence variant" refers to molecules with some differences in their amino acid sequences as compared to a reference (e.g. native sequence) polypeptide. The amino acid alterations may be substitutions, insertions, deletions or any desired combinations of such changes in a native amino acid sequence.

[0079] Substitutional variants are those that have at least one amino acid residue in a native sequence removed and a different amino acid inserted in its place at the same position. The substitutions may be single, where only one amino acid in the molecule has been

substituted, or they may be multiple, where two or more amino acids have been substituted in the same molecule.

[0080] Substitutes for an amino acid within the sequence may be selected from other members of the class to which the amino acid belongs. For example, the nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan and methionine. The polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine and glutamine. The positively charged (basic) amino acids include arginine, lysine and histidine. The negatively charged (acidic) amino acids include aspartic acid and glutamic acid. Also included within the scope of the invention are proteins or fragments or derivatives thereof which exhibit the same or similar biological activity and derivatives which are differentially modified during or after translation, e.g., by glycosylation, proteolytic cleavage, linkage to an antibody molecule or other cellular ligand, and so on.

[0081] Insertional variants are those with one or more amino acids inserted immediately adjacent to an amino acid at a particular position in a native amino acid sequence. Immediately adjacent to an amino acid means connected to either the α-carboxy or α-amino functional group of the amino acid.

[0082] Deletional variants are those with one or more amino acids in the native amino acid sequence removed. Ordinarily, deletional variants will have one or two amino acids deleted in a particular region of the molecule.

[0083] As used herein, “fragments” or “functional derivatives” refers to biologically active amino acid sequence variants and fragments of the polypeptide of the present invention, as well as covalent modifications, including derivatives obtained by reaction with organic derivatizing agents, post-translational modifications, derivatives with nonproteinaceous polymers, and immunoadhesins.

[0084] As used herein, "carriers" include pharmaceutically acceptable carriers, excipients, or stabilizers which are nontoxic to the cell or mammal being exposed thereto at the dosages and concentrations employed. Often the pharmaceutically acceptable carrier is an aqueous pH buffered solution. Examples of pharmaceutically acceptable carriers include without limitation buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid; low molecular weight (less than about 10 residues) polypeptide; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, arginine or lysine;

monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrins; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; salt-forming counterions such as sodium; and/or nonionic surfactants such as TWEEN®, polyethylene glycol (PEG), and PLURONICS®.

[0085] As used herein "pharmaceutically acceptable carrier and/or diluent" includes any and all solvents, dispersion media, coatings antibacterial and antifungal agents, isotonic and absorption delaying agents and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, use thereof in the therapeutic compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions.

[0086] It is especially advantageous to formulate parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the mammalian subjects to be treated; each unit containing a predetermined quantity of active material calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on (a) the unique characteristics of the active material and the particular therapeutic effect to be achieved, and (b) the limitations inherent in the art of compounding such an active material for the treatment of disease in living subjects having a diseased condition in which bodily health is impaired.

[0087] The principal active ingredient is compounded for convenient and effective administration in effective amounts with a suitable pharmaceutically acceptable carrier in dosage unit form. A unit dosage form can, for example, contain the principal active compound in amounts ranging from 0.5 μg to about 2000 mg. Expressed in proportions, the active compound is generally present in from about 0.5 μg/ml of carrier. In the case of compositions containing supplementary active ingredients, the dosages are determined by reference to the usual dose and manner of administration of the said ingredients.

[0088] As used herein, "vector", "polynucleotide vector", "construct" and "polynucleotide construct" are used interchangeably herein. A polynucleotide vector of this invention may be in any of several forms, including, but not limited to, RNA, DNA, RNA encapsulated in a retroviral coat, DNA encapsulated in an adenovirus coat, DNA packaged in another viral or viral-like form (such as herpes simplex, and adeno- structures, such as polyamides.

[0089] As used herein, "host cell" includes an individual cell or cell culture which can be or has been a recipient of a vector of this invention. Host cells include progeny of a single host cell, and the progeny may not necessarily be completely identical (in morphology or in total DNA complement) to the original parent cell due to natural, accidental, or deliberate mutation and/or change.

[0090] As used herein, "subject" is a vertebrate, preferably a mammal, more preferably a human.

[0091] As used herein, "mammal" for purposes of treatment refers to any animal classified as a mammal, including humans, domestic and farm animals, and zoo, sports, or pet animals, such as dogs, cats, cattle, horses, sheep, pigs, and so on. Preferably, the mammal is human.

[0092] As used herein, "treatment" is an approach for obtaining beneficial or desired clinical results. For purposes of this invention, beneficial or desired clinical results include, but are not limited to, alleviation of symptoms, diminishment of extent of disease, stabilized (i.e., not worsening) state of disease, delay or slowing of disease progression, amelioration or palliation of the disease state, and remission (whether partial or total), whether detectable or undetectable. "Treatment" can also mean prolonging survival as compared to expected survival if not receiving treatment. "Treatment" refers to both therapeutic treatment and prophylactic or preventative measures. Those in need of treatment include those already with the disorder as well as those in which the disorder is to be prevented. "Palliating" a disease means that the extent and/or undesirable clinical manifestations of a disease state are lessened and/or the time course of the progression is slowed or lengthened, as compared to a situation without treatment.

[0093] As used herein, “A1” peptide, “A2” peptide, “B1” peptide, “B2” peptide and “B3” peptide refer to peptides that bind to human NME7-AB, but not (or significantly less) to human NME1. The peptides used to generate these antibodies are common to both NME7-AB and NME7-X1, and are set forth as below.

[0094] A1 is NME7A peptide 1 (A domain): MLSRKEALDFHVDHQS (SEQ ID NO:141)

[0095] A2 is NME7A peptide 2 (A domain): SGVARTDASES (SEQ ID NO:142)

[0096] B1 is NME7B peptide 1 (B domain): DAGFEISAMQMFNMDRVNVE (SEQ ID NO:143)

[0097] B2 is NME7B peptide 2 (B domain): EVYKGVVTEYHDMVTE (SEQ ID NO:144)

[0098] B3 is NME7B peptide 3 (B domain):
AIFGKTKIQNAVHCTDLPEDGLLEVQYFF (SEQ ID NO:145)

[0099] Further, for the sake of clarity, NME7A (with capital letter "A") refers to the subunit A portion of NME7. NME7a (with small letter "a") refers to the full-length NME7 that is described elsewhere in this application. And, NME7B (with capital letter "B") refers to the subunit B portion of NME7. NME7b (with small letter "b") refers to a species of NME7 that is partially devoid of the DM10 region, which is described elsewhere in this application.

[00100] As used herein, the term "antibody-like" means a molecule that may be engineered such that it contains portions of antibodies but is not an antibody that would naturally occur in nature. Examples include but are not limited to CAR (chimeric antigen receptor) T cell technology and the Ylanthia® technology. The CAR technology uses an antibody epitope fused to a portion of a T cell so that the body's immune system is directed to attack a specific target protein or cell. The Ylanthia® technology consists of an "antibody-like" library that is a collection of synthetic human fabs that are then screened for binding to peptide epitopes from target proteins. The selected Fab regions can then be engineered into a scaffold or framework so that they resemble antibodies.

[00101] As used herein, an "effective amount of an agent to inhibit an NME family member protein" refers to the effective amount of the agent in hindering the activating interaction between the NME family member protein and its cognate receptor such as

[00102] As used herein, "NME derived fragment" refers to a peptide sequence that is either a fragment of the NME or is highly homologous to the peptide sequence that is a fragment of the NME.

[00103] As used herein, the "MUC1*" extra cellular domain is defined primarily by the PSMGFR sequence (GTINVHDVETQFNQYKTEAASRYNLTISDVSVSDVPFPFSAQSGA (SEQ ID NO:6)). Because the exact site of MUC1 cleavage depends on the enzyme that clips it, and that the cleavage enzyme varies depending on cell type, tissue type or the time in the evolution of the cell, the exact sequence of the MUC1* extra cellular domain may vary at the N-terminus.

[00104] As used herein, the term "PSMGFR" is an acronym for Primary Sequence of MUC1 Growth Factor Receptor as set forth as GTINVHDVETQFNQYKTEAASRYNLTISDVSVSDVPFPFSAQSGA (SEQ ID NO:6). In this regard, the "N-number" as in "N-10 PSMGFR", "N-15 PSMGFR", or "N-20 PSMGFR" refers to the number of amino acid residues that have been deleted at the N-terminal end of PSMGFR. Likewise "C-number" as in "C-10 PSMGFR", "C-15 PSMGFR", or "C-20

PSMGFR" refers to the number of amino acid residues that have been deleted at the C-terminal end of PSMGFR.

[00105] As used herein, the "extracellular domain of MUC1*" refers to the extracellular portion of a MUC1 protein that is devoid of the tandem repeat domain. In most cases, MUC1* is a cleavage product wherein the MUC1* portion consists of a short extracellular domain devoid of tandem repeats, a transmembrane domain and a cytoplasmic tail. The precise location of cleavage of MUC1 is not known perhaps because it appears that it can be cleaved by more than one enzyme. The extracellular domain of MUC1* will include most of the PSMGFR sequence but may have an additional 10-20 N-terminal amino acids.

[00106] As used herein, "high homology" is considered to be at least 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, or 97% identity in a designated overlapping region between any two polypeptides.

[00107] As used herein, "NME family proteins" or "NME family member proteins", numbered 1-10, are proteins grouped together because they all have at least one NDPK (nucleotide diphosphate kinase) domain. In some cases, the NDPK domain is not functional in terms of being able to catalyze the conversion of ATP to ADP. NME proteins were formerly known as NM23 proteins, numbered H1 and H2. Recently, as many as ten (10) NME family members have been identified. Herein, the terms NM23 and NME are interchangeable. Herein, terms NME1, NME2, NME5, NME6, NME7, NME8 and NME9 are used to refer to the native protein as well as NME variants. In some cases these variants are more soluble, express better in *E. coli* or are more soluble than the native sequence protein. For example, NME7 as used in the specification can mean the native protein or a variant, such as NME7-AB that has superior commercial applicability because variations allow high yield expression of the soluble, properly folded protein in *E. coli*. NME7-AB consists primarily of the NME7 A and B domains but is devoid of most of the DM10 domain (SEQ ID NO:39), which is at the N-terminus of the native protein. "NME1" as referred to herein is interchangeable with "NM23-H1". It is also intended that the invention not be limited by the exact sequence of the NME proteins. The mutant NME1-S120G, also called NM23-S120G, are used interchangeably throughout the application. The S120G mutants and the P96S mutant are preferred because of their preference for dimer formation, but may be referred to herein as NM23 dimers, NME1 dimers, or dimeric NME1, or dimeric NM23.

[00108] NME7 as referred to herein is intended to mean native NME7 having a molecular weight of about 42kDa.

[00109] A "family of NME7" refers to full length NME7 as well as naturally occurring or artificially created cleaved form having a molecular weight about 30kDa, 33kDa, or a cleaved form having a molecular weight of about 25kDa, a variant devoid or partially devoid of the DM10 leader sequence (SEQ ID NO:162), which is NME7 amino acids 1-91 of NME7 represented by SEQ ID NO:82 or 147, such as NME7b, NME7-X1, NME7-AB or a recombinant NME7 protein, or variants thereof whose sequence may be altered to allow for efficient expression or that increase yield, solubility or other characteristics that make the NME7 more effective or commercially more viable. The "family of NME7" may also include "NME7-AB-like" protein, which is a protein in the range of 30 to 33kDa that is expressed in cancer cells.

[00110] As used herein, an "an agent that maintains stem cells in the naïve state or reverts primed stem cells to the naïve state" refers to a protein, small molecule or nucleic acid that alone or in combination maintains stem cells in the naïve state, resembling cells of the inner cell mass of an embryo. Examples include but are not limited to human NME1 dimers, bacterial, fungal, yeast, viral or parasitic NME proteins that have high sequence identity to human NME proteins, especially NME1, NME7, NME7-X1, NME7-AB, NME6, 2i (Silva J et al, 2008; Hanna et al, 2010), 5i (Theunissen TW et al, 2014), nucleic acids such as siRNA that suppress expression of MBD3, CHD4 (Rais Y1 et al, 2013), BRD4, or JMJD6 (Liu W et al 2013).

[00111] As used herein, an "an agent that promotes pluripotency" or "reverts somatic cells to a stem-like or cancer-like state" refers to a protein, small molecule or nucleic acid that alone or in combination induces expression of or suppresses expression of certain genes such that the genetic signature shifts to one that more closely resembles stem cells or cancer cells. Examples include but are not limited to NME1 dimers, NME7, NME7-X1, NME7-AB, 2i, 5i, nucleic acids such as siRNA that suppress expression of MBD3, CHD4, BRD4, or JMJD6, microbial NME proteins that have high sequence homology to human NME1, NME2, NME5, NME6, NME7, NME8, or NME9, preferably with the regions that house NDPK domains.

[00112] As used herein, in reference to an agent being referred to as a "small molecule", it may be a synthetic chemical or chemically based molecule having a molecular weight between 50Da and 2000Da, more preferably between 150 Da and 1000 Da, still more preferably between 200Da and 750Da.

[00113] As used herein, in reference to an agent being referred to as a "natural product", it may be chemical molecule or a biological molecule, so long as the molecule exists in nature.

[00114] As used herein, FGF, FGF-2 or bFGF refer to fibroblast growth factor (Xu RH et al, 2005; Xu C et al, 2005).

[00115] As used herein, "Rho associated kinase inhibitors" may be small molecules, peptides or proteins (Rath N, et al, 2012). Rho kinase inhibitors are abbreviated here and elsewhere as ROCi or ROCKi, or Ri. The use of specific rho kinase inhibitors are meant to be exemplary and can be substituted for any other rho kinase inhibitor.

[00116] As used herein, the term "cancer stem cells" or "tumor initiating cells" refers to cancer cells that express levels of genes that have been linked to a more metastatic state or more aggressive cancers. The terms "cancer stem cells" or "tumor initiating cells" can also refer to cancer cells for which far fewer cells are required to give rise to a tumor when transplanted into an animal. Cancer stem cells and tumor initiating cells are often resistant to chemotherapy drugs.

[00117] As used herein, the terms "stem/cancer", "cancer-like", "stem-like" refers to a state in which cells acquire characteristics of stem cells or cancer cells, share important elements of the gene expression profile of stem cells, cancer cells or cancer stem cells. Stem-like cells may be somatic cells undergoing induction to a less mature state, such as increasing expression of pluripotency genes. Stem-like cells also refers to cells that have undergone some de-differentiation or are in a meta-stable state from which they can alter their terminal differentiation. Cancer like cells may be cancer cells that have not yet been fully characterized but display morphology and characteristics of cancer cells, such as being able to grow anchorage-independently or being able to give rise to a tumor in an animal.

[00118] As used herein, "spacers" or "linkers" of different lengths can be incorporated anywhere in the peptide. Spacer attachment is usually through an amide linkage but other functionalities are possible.

[00119] NME, NME7 and protein family of NME7

[00120] The present inventors discovered that NME7 is highly expressed in early human stem cells and also in most cancer cells (Figures 1, 2, 3, 35, 36, 38, 39 and 40 and Examples 2, 3, and 4). Further, we demonstrated that like NM23-H1, NME7 binds to and dimerizes the MUC1* growth factor receptor on both stem cells and cancer cells. Figure 15 shows a sequence alignment of NME1 and NME7 A and B domains.

[00121] The inventors recently discovered that NME7 is a primitive form of NME1 (NM23-H1) that is expressed in very early embryonic stem cells. NME7 is either not expressed at all, or is expressed at extremely low levels, in adult tissues. However, the inventors discovered that NME7 is expressed at high levels in cancerous cells and tissues and

at even higher levels in metastatic cancer cells and tissues. A cleaved form of NME7 may be a secreted form allowing it to bind to and activate extracellular receptors. We detect full-length NME7, MW 42kDa, as well as NME7 species that are approximately 33kDa and 30kDa. The 33kDa and 30kDa species are secreted from cancer cells. Western blots detect full-length NME7 in cell lysates, but smaller 30-33kDa NME7 species in their condition media (Figures 9 and 10). Western blots probed with either an antibody that recognizes NME7 or an antibody that only recognizes the DM10 domain show that the lower molecular weight NME7 species that are secreted into the conditioned media are devoid of the DM10 domain (Figure 10). These data are consistent with the idea that naturally occurring NME7 species are comparable to the recombinant NME7-AB we generated as they have nearly the same molecular weight, both are secreted and are both devoid of the 91 amino acids of the DM10 domain which may keep the protein retained within the cell.

[00122] We discovered a new NME7 isoform, NME7-X1, and also discovered that it is over-expressed in cancers and is particularly over-expressed in prostate cancers (Figure 33, 34). NME7-X1, molecular weight ~30kDa, comprises NME7 amino acids 125-376, whereas the recombinant NME7-AB, molecular weight ~33kDa, that we generated spans amino acids 92-376, so includes 33 more N-terminal amino acids. NME7b spans amino acids 37-376 and is devoid of only 37 amino acids of the DM10 domain is also overexpressed in prostate cancers (Figure 34). We generated a human recombinant NME7-X1 and show that it is the secreted 30kDa NME7 species in cancer cells that runs just lower than a naturally occurring ~33kDa NME7 species that appears to be a naturally occurring "NME7-AB-like" protein that is a cleavage product or alternative isoform.

[00123] We tested a panel of cancer cell lines and found that they express high levels of NME7 and lower molecular weight species that may be truncations similar to NME7-AB, such as NME7-AB-like protein, or alternate isoforms such as NME7-X1.

[00124] Whereas NM23-H1 (aka NME1) has to be a dimer, NME7 is a monomer with two binding sites for MUC1* extracellular domain. We generated a recombinant human NME7 that is devoid of the DM10 domain, which we call NME7-AB. Figures 4A-4C show the elution profile of size exclusion chromatography purification of NME7-AB, a non-reducing SDS-PAGE gel from NME7-AB peak fractions and the elution profile of size exclusion chromatography of the purified NME7-AB. A sandwich ELISA binding assay that shows that a recombinant NME7, NME7-AB simultaneously binds to two PSMGFR peptides wherein the extracellular domain of MUC1* is comprised of most or all of the PSMGFR sequence

(Figure 5, Example 6). In a nanoparticle binding assay, NME7 was also shown to be able to bind to and dimerize the PSMGFR portion of the MUC1* extracellular domain.

[00125] Agents that disable NME7, block its interaction with its binding partners or suppress its expression are potent anti-cancer therapeutics. Such agents may be antibodies, small molecules or nucleic acids. They may act on NME7 directly, on molecules that regulate NME7 expression, or on enzymes that cleave NME7 to cancer-promoting forms.

[00126] We discovered that like NM23-H1 dimers, a recombinant NME7-AB monomer was fully able to support pluripotent human stem cell growth in the absence of any other growth factor, cytokine or serum. Competitively inhibiting the interaction between NME7 and MUC1* extracellular domain, comprised essentially of the PSMGFR sequence, induced differentiation of stem cells, showing that it is the interaction of NME7 and MUC1* that promotes stem cell growth and inhibits differentiation.

[00127] Next, we showed that NME7-AB alone is also able to fully support human cancer cell growth. NME7-AB, when added to regular cancer cell growth media, stimulated cancer cell growth and in particular the growth of MUC1-positive and MUC1*-positive cancer cells. Inhibiting the interaction of NME7 with MUC1* inhibited cancer cell growth. Blocking the MUC1* growth factor receptor with an anti-MUC1* Fab potently inhibited cancer cell growth. Similarly, antibodies that bind to NME7 inhibit cancer cell growth. One example of inhibition of cancer growth by anti-NME7 antibody is shown in Figures 6-8 and Example 10. In this case, the polyclonal antibody was generated from immunizing an animal with the portion of NME7 that spans amino acids 100-376. However, we found that antibodies generated from immunizing with shorter peptides from NME7-AB or from NME7-X1 also inhibit cancer growth. In particular, they inhibit the growth of MUC1 and MUC1*-positive cancers.

[00128] NME7 Causes Cancer Metastasis

[00129] The inventors further discovered that culturing cancer cells in a minimal media containing NME7-AB induced a wide variety of cancer cells to become transformed to a more metastatic state. Evidence of this induced metastatic state include a change from adherent cell growth to no-adherent cell growth, aka, “floater” cells and accompanying up-regulation of specific metastatic markers that were especially upregulated in the floating cells. These metastatic markers that are upregulated after culture in NME7-AB include but are not limited to CXCR4, CHD1 aka E-cadherin, CD44, and pluripotent stem cell markers such as OCT4, SOX2, NANOG and KLF2/4. Cancer cells cultured in NME7-AB had dramatically higher engraftment rates when xenografted into test animals, which were over

90%. In addition, very low numbers of implanted cancer cells formed tumors in the test animals, which is evidence that NME7-AB had transformed them into cancer stem cells also known as metastatic cancer cells. Because cancer cells make either an NME7 cleavage product or alternative isoform that is essentially equivalent to NME7-AB, the methods described here are not limited to using NME7-AB; other NME7 species could work as well. For example, we discovered another NME7 isoform, NME7-X1, is expressed by cancer cells. It is identical to our recombinant NME7-AB with the exception that the X1 isoform is missing 33 amino acids from the N-terminus. NME7-X1 is expected to function like NME7-AB. "NME7-AB-like" protein has also been detected in cancer cells as being about 33Da species.

[00130] We note that the inventors' previous work showed that NME7-AB alone is able to revert human stem cells to an earlier naïve state. We discovered that culturing cancer cells in the presence of other reagents that make stem cells revert to a more naïve state, makes the cancer cells transform to a more metastatic state. We demonstrated that NME7-AB (Figure 11 and 12), "2i" inhibitors (Figure 14A), human NME1 dimers or bacterial NME1 dimers with high sequence homology to human NME1 or human NME7 (Figure 14B) are each able to transform regular cancer cells into metastatic cancer cells, which are also called cancer stem cells "CSCs" or tumor initiating cells "TICs" (Figures 11 - 14).

[00131] 2i is the name given to two biochemical inhibitors that researchers found made human stem cells revert to a more naïve state. 2i are MEK and GSK3-beta inhibitors PD0325901 and CHIR99021, which are added to culture medium to final concentrations of about 1 mM and 3 mM, respectively. NME7-AB and NME7-X1 are at a final concentration of about 4nM when added to separate batches of minimal medium to make cancer cells transform to metastatic cells, although lower and higher concentrations also work well in the range of about 1nM to 16nM. Human or bacterial NME1 dimers are used at a final concentration of 4nM to 32nM, with 16nM typically used in these experiments, wherein the human NME bears the S120G mutation. Lower concentrations may be required if using wild type. It is not intended that these exact concentrations are important. It is important that the NME1 proteins are dimers and the range of concentrations over which this happens is in the low nanomolar range although certain mutations allow higher concentrations to remain as dimers. Similarly, the concentrations of NME7 proteins can vary. NME7-AB and NME7-X1 are monomers and concentrations used to transform cancer cells to metastatic cells should allow the proteins to remain as monomers.

[00132] In addition to NME7, NME7-AB, NME7-X1, and the 2i inhibitors MEKi and GSK3i, other reagents and inhibitors have been shown by others to cause stem cells to revert to a more naïve state. These inhibitors, "i's" include JNKi, p38i, PKCi, ROCKi, BMPi, BRAFi, SRCi as well as growth factors activing and LIF (Gafni et al 2013, Chan et al 2013, Valamehr et al 2014, Ware et al 2014, Theunissen et al 2014). These reagents can also be used to make cancer cells progress to a more metastatic state. Cells that have been induced to transform to a more metastatic state using any single factor or combination of the inhibitors or growth factors, that make stem cells revert to a more naïve state, can then be used as discovery tools to identify or test drugs to treat or prevent cancer metastasis.

[00133] Various molecular markers have been proposed as being indicators of metastatic cancer cells. Different cancer types may have different molecules that are up-regulated. For example, the receptor CXCR4 is up-regulated in metastatic breast cancers while E-cadherin, also known as CHD1, is up-regulated more in metastatic prostate cancers. In addition to these specific metastasis markers, typical markers of pluripotency such as OCT4, SOX2, NANOG, and KLF4 are up-regulated as cancers become metastatic. The starting cancer cells and the later metastatic cancer cells are assayed by PCR to measure expression levels of these genes. We demonstrated that these cancer cells, cultured in agents such as NME7-AB that cause them to be transformed to a more metastatic state, as evidenced by increased expression of metastatic markers and pluripotent stem cell markers, function as metastatic cancer cells.

[00134] A functional test of whether or not a population of cancer cells is metastatic is to implant very low numbers, e.g. 200, of the cells in immuno-compromised mice and see if they develop into a tumor. Typically 5-6 million cancer cells are required to form a tumor in an immuno-compromised mouse. We showed that as few as 50 of the NME-induced metastatic cancer cells formed tumors in mice. In addition, mice that were injected throughout the test period with human NME7-AB, NME1, or NME7-X1 developed remote metastases.

[00135] In one particular experiment, T47D human breast cancer cells were cultured in standard RPMI media for 14 days with media changes every 48 hours and passed by trypsinization when approximately 75% confluent. The cells were then plated into 6-well plates and cultured in minimal stem cell media (see Example 1) that was supplemented with 4nM NME7-AB. Media was changed every 48 hours. By about Day 4, some cells become detached from the surface and float. Media is carefully changed so as to retain the "floaters" as these are the cells that have the highest metastatic potential as evidence by RT-PCR

measurement of metastatic markers. On Day 7 or 8, the floaters are harvested and counted. Samples are retained for RT-PCR measurement. The key marker measured is CXCR4, which is up-regulated by 40-200-times after being briefly cultured in NME7-AB.

[00136] The freshly harvested floater metastatic cells were xenografted into the flank of female nu/nu athymic mice that have been implanted with 90-day slow release estrogen pellets. Floater cells were xenografted with 10,000, 1,000, 100 or 50 cells each. Half of the mice in each group of 6 were also injected daily with 32nM NME7-AB near the original implantation site. The parent T47D cells that were cultured in RPMI media without NME7-AB were also implanted into mice at 6 million, 10,000 or 100 as controls. Mice implanted with the NME7-induced floater cells developed tumors even when as few as 50 cells were implanted. Mice that were implanted with the floater cells and that received daily injections of NME7-AB also developed remote tumors or remote metastases in various organs (Figure 20-25). 11 out of the 12 mice, or 92%, that were injected with human NME7-AB after implantation of the NME7-AB cultured cancer cells developed tumors at the injection site. Only 7 out of the 12 mice, or 58%, that were not injected with human NME7-AB after implantation developed tumors. 9 out of the 11 mice, or 82%, that exhibited tumors and were injected with human NME7-AB developed multiple tumors remote from the injection site. None of the mice that were not injected with NME7-AB developed multiple, visible tumors.

[00137] After sacrifice, RT-PCR and Western blots showed that the remote bumps on the mice injected with NME7-AB were indeed human breast tumors. Similar analysis of their organs showed that in addition to remote bumps, mice had randomly metastasized to the liver and lung with human breast cancer characteristic of the human breast cancer cells that were implanted. As expected, only the mice implanted with 6 million cells grew tumors.

[00138] We have demonstrated that human recombinant NME7-AB is comparable in size and sequence to NME7-X1 and to a 30-33kDa NME7 cleavage product. We have shown that NME7-AB promotes cancerous growth and causes cancer cells to accelerate to the highly metastatic cancer stem cell (CSC) state also called tumor initiating cells (TIC). Therefore, we conclude that NME7-X1 and an NME7 cleavage product that removes the DM10 domain also promote cancerous growth and causes cancer cells to accelerate to the highly metastatic cancer stem cell (CSC) state also called tumor initiating cells (TIC). In one example, NME7-AB was added to cancer cells in a serum-free media and in the absence of any other growth factors or cytokines. Within 7-10 days, the cancer cells had reverted to the highly metastatic CSCs/TICs as evidenced by more than 100-fold increase in the expression of molecular markers such as CXCR4, which are indicators of metastatic cancer cells. In one example,

T47D breast cancer cells were cultured in either standard RPMI media or in a Minimal Stem Cell Media (Example 1) to which was added recombinant NME7-AB to a final concentration of 16nM. After 10 days cells were collected and analyzed by RT-PCR for expression of molecular markers of CSCs which were elevated by 10-200-times (Figures 11, 12). This is a specific, detailed example of how we transformed one cancer cell type to a more metastatic state. It is not intended that the invention be limited by these details as there are a range of cancer cells that are transformed in this way, a range of reagents that revert stem cells to a more naïve state that also progress cancer cells to a more metastatic state and a range of concentrations over which the added reagents transform the cancer cells. Other types of cancer cells have required longer periods of culture in NME7-AB for dramatic upregulation of metastatic markers and ability to form tumors from very low numbers of cancer cells implanted. For example, prostate cancer cells cultured in NME7-AB, 2i, human NME1 or bacterial NME1 that has high homology to human NME1 or human NME7 showed dramatic increase in metastatic markers after 2-3 passages.

[00139] Metastasis marker CXCR4 is particularly elevated in metastatic breast cancer cells, while CHD1 is particularly elevated in metastatic prostate cancer. Here we show that pluripotent stem cell markers such as OCT4, SOX2, NANOG, KLF2/4 and TBX3 are also up-regulated when cancer cells transform to more metastatic cells.

[00140] DU145 prostate cancer cells were cultured similarly and those cells cultured in NME7-AB also showed dramatic increases in expression of CSC markers (Figure 13). In prostate cancer cells, CHD1 (aka E-cadherin) and CXCR4 were up-regulated compared to the control cancer cells, which were not grown in NME7-AB, along with other pluripotent stem cell markers. Ovarian cancer cells, pancreatic cancer cells and melanoma cells were also cultured in NME7-AB and were transformed to a more metastatic state after as few as 3 days in culture. Figures 37A-C shows that ovarian cancer cell lines SK-OV3, OV-90 and breast cancer cell line MDA-MB all transitioned from adherent to non-adherent floater cells and increased expression of metastatic markers after 72 or 144 hours in culture with NME7-AB.

[00141] Here we have shown that NME7-AB transforms a wide range of cancer cells to a more metastatic state. We have also shown that cancer cells express a naturally occurring species that is approximately the same molecular weight as recombinant NME7-AB 33kDa (Figures 33-36 and Figure 38) and is also devoid of the DM10 domain (Figure 10) like NME7-AB and also express an alternative isoform NME7-X1 30kDa which is the same sequence as NME7-AB except is missing 33 amino acids from the N-terminus. A co-immunoprecipitation experiment was performed on T47D breast cancer cells, wherein the

cell extracts were incubated with an antibody against the MUC1 cytoplasmic tail, Ab-5, or a control antibody, IgG, and co-immunoprecipitated. The immunoprecipitated species were separated by gel electrophoresis. The gels were blotted with two different commercially available anti-NME7 antibodies. Both gels show unique NME7 bands at ~33kDa and ~30kDa (Figure 39 A,B). The gels were stripped and re-probed with an antibody against the extracellular domain of MUC1*, anti-PSMGFR (Figure 39 C,D), which shows that the NME7 species and MUC1* interact. A recombinant NME7-AB and a recombinant NME7-X1 that we made were mixed together and run on a gel, then probed with an anti-NME7 antibody, showing that the two unique NME7 species that are naturally occurring in breast cancer cells and that interact with MUC1* are an NME7-AB-like species and NME7-X1 (Figure 39E). A similar experiment was carried out in human stem cells. Figures 40A-C show photographs of Western blots of a co-immunoprecipitation experiment. Human induced pluripotent stem, iPS7, or embryonic stem, HES3, cell extracts were incubated with an antibody against the MUC1 cytoplasmic tail, Ab-5, or a control antibody, IgG, and co-immunoprecipitated. The gel was blotted with a commercially available anti-NME7 antibody B9 (A). Both cell types show unique NME7 bands at ~33kDa and ~30kDa. The gel was stripped and re-probed with an antibody against the extracellular domain of MUC1*, anti-PSMGFR (B), which shows that the NME7 species and MUC1* interact. A recombinant NME7-AB and a recombinant NME7-X1 that we made were mixed together and run on a gel, then probed with an anti-NME7 antibody, showing that the two unique NME7 species that are naturally occurring in breast cancer cells and that interact with MUC1* are an NME7-AB-like species and NME7-X1 (C). Because NME7-AB is a recombinant protein, we do not know if the naturally occurring species may contain an extra 1-15 additional amino acids or devoid of 1-15 additional amino acids than the recombinant NME7-AB, yet run with the same apparent molecular weight. By "NME7-AB-like", we mean an NME7 species that runs with an apparent molecular weight of approximately 33kDa that is able to function the way the recombinant NME7-AB does, in that it is able to stimulate cancer cell growth, induce transition of cancer cells to a more metastatic state and is able to fully support pluripotent growth of human stem cells.

[00142] We conclude that cancer cell lines and cancer cell populations that express NME7 and lower molecular weight NME7 species contain some cancer cells that are CSCs or metastatic cancer cells. These cancers can be made more metastatic or increase the population of cells that are metastatic by culturing the cells in NME7-AB, NME7-X1 or lower molecular weight NME7 species. Figure 35 shows a Western blot of a panel of cancer cells all

expressing NME7 as well as lower molecular weight species NME7-AB-like at 33kDa and NME7-X1 at 30kDa. Figure 38 shows that cancer cell lines T47D breast cancer, PC3 and DU145 prostate cancer, BT-474 breast cancer, CHL-1 and A2058 both melanoma cell lines and CAPAN-2 and PANC-1 both pancreatic cell lines all express MUC1, MUC1* and NME7-AB-like species and NME7-X1. In Figure 38A, BT0474 cells appear not to express MUC1 or MUC1* however, we previously showed (Fessler et al 2009) that when these HER2 positive breast cancer cells become resistant to chemotherapy drugs, i.e. metastatic, they do so by increasing expression of MUC1* (Figure 38 D). Blocking the MUC1* receptor with an anti-MUC1* Fab reversed their resistance to Herceptin (Figure 38E), Taxol (Figure 38F) as well as other chemo agents. These cancer types and other cancer types that express NME7 and lower molecular weight NME7 species such as 33kDa, 30kDa can be made more metastatic or increase the population of cells that are metastatic by culturing the cells in NME7-AB, NME7-X1 or lower molecular weight NME7 species.

[00143] Conversely, the metastatic potential of these and other cancer types that express NME7 and lower molecular weight NME7 species such as 33kDa or 30kDa can be reversed by treating the cells with anti-NME7 antibodies. Anti-NME7 antibodies or antibodies that bind to NME7-AB or NME7-X1 are administered to a patient for the treatment or prevention of cancers including breast, prostate, ovarian, pancreatic and liver cancers. Because we have shown that NME7-AB exerts its tumorigenic effects by binding to and activating the MUC1* growth factor receptor, anti-NME7 antibodies will be effective against any MUC1*-positive cancers, which include but are not limited to breast, lung, liver, pancreatic, gastric colorectal, prostate, brain, melanoma, kidney and others. Anti-NME7, anti-NME7-AB or anti-NME7-X1 antibodies are administered to patients for the treatment or prevention of cancers that are NME7-AB, NME7-AB-like, or NME7-X1 positive or a MUC1* positive.

[00144] **Testing Patient Cancer Cells for Effective Therapies**

[00145] NME7-AB, NME7-X1 as well as 2i and other reagents that revert stem cells to a more naïve state also induce cancer cells to transform to a more metastatic state. After treatment with any one or combination of these reagents, cancer cells have a higher engraftment rate and require up to 100,000-times less cells to cause a tumor to form in a test animal. Therefore, methods described in this disclosure can be used to enable xenografting of a patient's primary tumor cells into a test animal.

[00146] Candidate therapeutic agents can then be tested on the recipient animal. Effective therapeutic agents identified in this way can be used to treat the donor patient or other patients with similar cancers. In one embodiment, a method of identifying effective

therapeutics for a particular patient or a particular type of cancer comprises the steps of: 1) cancer cells are obtained from a cell line, a patient or a patient to whom the therapeutic being tested will be administered; 2) cancer cells are cultured in NME7-AB, NME7-X1, human NME1, bacterial NME1 that has high homology to human NME1 or NME7, 2i, or other reagents shown to revert stem cells to a more naïve state; 3) resultant cancer cells are implanted into a test animal to which human NME7-AB, NME7-X, human NME1, bacterial NME1 that has high homology to human NME1 or NME7, 2i, or other reagents shown to revert stem cells to a more naïve state may also be administered or animal is transgenic for human NME7-AB or NME7-X1; 4) candidate anti-cancer therapeutic agents are administered to the animal; 5) efficacy of the therapeutic agents are assessed; and 6) effective therapeutic agent is administered to the donor patient or to another patient with similar cancer.

[00147] Anti-NME7 Antibodies

[00148] Anti-NME7 antibodies are potent anti-cancer agents. NME7 is a growth factor that promotes the growth of cancer cells and also promotes their progression to a more metastatic state or a more aggressive state. NME7 and a truncated form of NME7 that is ~ 33 kDa or 30 kDa have been shown to fully support cancer growth even in serum-free media devoid of any other growth factors or cytokines. In pull-down assays, ELISAs and nanoparticle binding experiments, we have shown that the growth factor receptor MUC1* is a binding partner of NME7 and NME7-AB. Promotion of this interaction by eliminating all other growth factors or cytokines increased expression of cancer stem cell markers. Blocking this interaction even in the presence of serum, using a polyclonal antibody that specifically binds to NME7 actively killed the cancer cells. Thus, anti-NME7 or anti-NME7-AB antibodies are potent anti-cancer agents that can be administered to a patient for the treatment or prevention of cancers. More than 75% of all cancers are MUC1* positive. MUC1* is the transmembrane cleavage product of MUC1 wherein most of the extracellular domain has been shed, leaving a portion of the extracellular domain that contains most of the PSMGFR sequence and may contain 9-20 additional amino acids N-terminal to the boundary of the of the PSMGFR sequence.

[00149] One aspect of the invention is a method of treating or preventing cancer in a subject, comprising administering to the subject an effective amount of an anti-NME7 antibody. In one instance, the anti-NME7 antibody is able to bind to NME7-AB. In another instance, the anti-NME7 antibody is able to bind to NME7-X1. In yet another instance, the anti-NME7 antibody that is administered to a patient inhibits or prevents its binding to its target in the promotion of cancers. In one case, the target is the extracellular domain of a

cleaved MUC1. More specifically, the NME7 target that promotes cancer is the PSMGFR region of the MUC1* extracellular domain. In one aspect, an effective therapeutic agent is one that disrupts or prevents the interaction between an NME7 species and MUC1* extracellular domain, consisting primarily of the PSMGFR portion of MUC1* or the PSMGFR peptide. Agents for the treatment or prevention of cancers are those agents that directly or indirectly inhibit the expression or function of NME7, an NME7-AB-like cleavage product or alternative isoform, including NME7-X1. In one case an effective anti-cancer therapeutic agent is one that binds to the NME7 species or disables its tumorigenic activity. An effective therapeutic agent for the treatment or prevention of cancers is an agent that binds to or disables NME7, an NME7-AB-like cleavage product or alternative isoform, or NME7-X1. In one aspect, the therapeutic agents that binds to the NME7 species is an antibody. The antibody may be polyclonal, monoclonal, bispecific, bivalent, monovalent, single chain, scFv, or an antibody mimic that may be animal in origin, human-animal chimera, humanized or human. The antibody can be generated by inoculation or immunization with an NME7 species or fragment thereof or selected, for example from a library or a pool of antibodies, for their ability to bind to an NME7 species, including NME7, an NME7-AB-like cleavage product or alternative isoform, including NME7-X1.

[00150] Generation of Anti-NME7 Antibodies

[00151] Anti-NME7 antibodies can be generated outside of the patient such as in a host animal or in a patient. Antibodies can be generated by immunization of NME7 or NME7 fragments or selected from a library or pool of antibodies that may be natural, synthetic, whole or antibody fragments based on their ability to bind to desired NME7 species such as NME7-AB or NME7-X1. In one aspect, the antibody is generated from immunization with, or selected for its ability to bind to, a peptide selected from those listed in Figures 16-19. In another aspect, the antibody is generated from peptides whose sequences are not identical to those of human NME1 or the antibodies are selected for their ability to bind to NME7 species and their inability to bind to human NME1.

[00152] One method used to identify NME7 or NME7-X1 derived peptides that give rise to antibodies that inhibit cancer growth and inhibit transition to metastasis or peptides that are themselves inhibitory is as follows: 1) protein sequences of human NME1, human NME7, human NME7-X1 and several bacterial or fungal NME proteins that have high sequence homology to either human NME1 or human NME7 are aligned; 2) regions of high sequence homology among all the NMEs are identified; 3) peptide sequences that are unique to NME7 or NME7-X1 but are flanking the regions of high sequence homology are identified. The

peptides are then synthesized and used to generate antibodies in a human or host animal. The resultant antibodies are tested for their ability to inhibit cancer growth or inhibit the transition to metastatic cancer cells.

[00153] Use of Anti-NME7 Antibody for Treatment of Cancer

[00154] Those antibodies that inhibit cancer growth or transition to a more metastatic state are selected for use as anti-cancer therapeutics and may be administered to a patient for the treatment or prevention of cancers. Selected antibodies may be further optimized for example by engineering or making human chimera antibodies or fully human antibodies. To demonstrate the efficacy of this approach, we selected NME7 peptides from regions of NME7 suspected to be critical to its cancerous function. We then generated antibodies using these peptides and then tested both the resultant antibodies as well as the immunizing peptides for their ability to: a) inhibit cancerous growth; and b) inhibit the induced transition from cancer cells to metastatic cancer cells. NME7 peptides were selected as immunizing agents for antibody production and as inhibitory agents themselves (Figure 19 and Example 11). Peptides A1 (SEQ ID NO:141), A2 (SEQ ID NO:142), B1 (SEQ ID NO:143), B2 (SEQ ID NO:144) and B3 (SEQ ID NO:145), wherein A refers to the domain from which the peptide is derived, i.e. the NDPK A domain and the B denotes that the peptide is derived from the NDPK B domain (Figure 15). Each peptide was used as an immunogen and injected into 2 rabbits each for production of polyclonal antibodies. The antibodies that were harvested from the blood of the immunized rabbits were purified over a column derivatized with the immunizing peptide. The purified antibodies were then tested for their ability to bind to human NME7. All of the resultant antibodies bound to human NME7 but not human NME1 as desired (Figure 26 A-B, Example 12). These results show that by choosing peptides whose sequence is found in NME7 but not exactly identical in NME1, antibodies are generated that specifically bind to NME7 but not NME1. Because NME1 has healthy function, it is in most cases desirable to generate antibodies that do not interfere with NME1. The antibodies were also tested for their ability to inhibit the binding of NME7 to a MUC1* extracellular domain peptide. The ELISA experiment shown in Figure 27 shows that the antibodies inhibited the binding of NME7-AB to a MUC1* extracellular domain peptide much more than they inhibited binding of NME1.

[00155] This is but one example of selecting peptides that generate antibodies that inhibit the cancerous function of NME7 and NME7 species. Sequence alignment among human NME1, human NME7, human NME7-X1 and bacterial NME proteins that had high sequence homology to human NME1 or NME7 identified five regions of homology. The fact that

peptides A1, A2, B1, B2 and B3 all generated antibodies that inhibited cancer growth or their transition to a metastatic state means that the five regions from which these peptides were derived are regions of NME7 that are important for its function in the promotion of cancer. Other peptides from these regions will also give rise to anti-NME7 antibodies that will inhibit cancer growth and metastasis and are therefore potent anti-cancer therapeutics. Antibodies generated from peptides A1, A2, B1, B2 and B3 were shown to inhibit cancer growth and inhibited the transition to a more metastatic state. Monoclonal antibodies generated by immunization with the same or similar peptides and subsequent testing of the monoclonals will identify antibodies that, after humanizing or other engineering known to those skilled in the art, would be administered to a patient for the treatment or prevention of cancers.

[00156] In a particular experiment, the antibodies generated by immunization with peptides A1, A2, B1, B2 and B3, as well as the immunizing peptides themselves, were added to cancer cells in culture to see if the addition of the antibodies or the immunizing peptides would inhibit cancer cell growth. At low concentrations and added separately, the antibodies as well as the immunizing peptides inhibited cancer cells growth (see Figure 28 for one example). However, when added at higher concentrations or combined, the antibodies as well as the immunizing peptides robustly inhibited cancer cell growth (Figure 29). The corresponding human NME7 amino acid numbers of immunizing peptides A1, A2, B1, B2 and B3 are 127-142, 181-191, 263-282, 287-301, 343-371, respectively, from human full-length NME7 having SEQ ID NO:82 or 147.

[00157] To clarify, when residue numbers of NME7 are discussed, they refer to the residue numbers of NME7 as set forth in SEQ ID NO:82 or 147.

[00158] The antibody used in the cancer growth inhibition experiments shown in Figure 6-8 and one of the antibodies shown in Figure 28 was generated by immunizing with NME7 peptide corresponding to amino acids 100-376 of NME7 (SEQ ID NO:82 or 147). To generate higher affinity and specific anti-NME7 antibodies, the following steps are followed: immunize animal with a peptide containing human NME7 amino acids 100-376, then: 1) de-select those antibodies that bind to human NME1; 2) select those antibodies that inhibit NME7-AB, 2i, or other NME induced transition of cancer cells to a more metastatic state; 3) select those antibodies that inhibit the growth of cancer cells; 4) select those antibodies that inhibit the growth of MUC1* positive cancer cells; 5) select those antibodies that inhibit binding of NME7-AB or NME7-X1 to MUC1* extracellular domain, essentially inhibit binding to the PSMGFR peptide; and/or 6) select those antibodies that bind to one or more of the peptides listed in Figure 19 - A1, A2, B1, B2 or B3 peptides.

[00159] Higher affinity monoclonal antibodies or monoclonal antibodies generated from longer peptides may be more effective antibody therapeutics. Alternatively, combinations of ani-NME7, anti-NME7-AB or anti-NME7-X1 antibodies are administered to a patient to increase efficacy.

[00160] Anti-NME7 antibodies inhibit the transition of cancer cells to metastatic cancer cells.

[00161] Anti-NME7 antibodies inhibit transition of cancer cells to metastatic cancer cells also called cancer stem cells (CSCs) or tumor initiating cells (TICs). Recall that we have demonstrated that culturing a wide variety of cancer cells in the presence of NME7-AB causes them to transition from regular cancer cells to the metastatic CSCs or TICs. Thus, antibodies that bind to NME7, NME7-AB or NME7-X1 will inhibit the progression of cancer cells to a more metastatic state.

[00162] Cancer cells transform to a more metastatic state when cultured in the presence of agents that revert stem cells to a more naïve state. We have demonstrated that culturing cancer cells in NME7-AB, human NME1 dimers, bacterial NME1 dimers or MEK and GSK3-beta inhibitors, called "2i", causes the cells to become more metastatic. As the cells transition to a more metastatic state, they become non-adherent or less adherent and float off of the culture plate. These floating cells, "floaters" were collected separately from those that were adherent and were shown to: a) express much higher levels of metastatic genes; and b) generated tumors when xenografted into mice at very low copy number. RT-PCR measurement of specific metastatic markers such as CXCR4 for breast cancers, CHD1 for prostate cancer, and other pluripotent stem cell markers such as OCT4, SOX2, NANOG, KLF4 and others were dramatically over-expressed in cancer cells that were cultured in NME7-AB and most over-expressed in the cells that became non-adherent, called "floaters" here and in figures.

[00163] In one example, NME7-AB specific antibodies, generated by immunization with NME7-derived peptides A1, A2, B1, B2 and B3, as well as the immunizing peptides themselves, were added into the media along with either NME7-AB or 2i to determine if they inhibited the transformation of regular cancer cells to metastatic cancer stem cells. Antibodies and peptides were separately added along with the agent that causes metastatic transformation; in this case NME7-AB or the 2i inhibitors PD0325901 and CHIR99021. NME7-AB and 2i were separately used to induce the cancer cells to be transformed to a more aggressive metastatic state. 2i was used so that it could not be argued that the antibodies that

were added to the media simply sopped up all of the NME7-AB so that the causative agent effectively was not there (Example 14).

[00164] Visual observation was independently recorded by two scientists as the experiment progressed (Figure 30). The most striking observation was that the antibodies and the peptides dramatically reduced the number of floater cells, which was the first indication that the antibodies and peptides inhibit the transformation to metastatic cancer cells. In particular, cells to which the antibody generated from immunization with the B3 peptide barely generated any floater cells. mRNA was extracted from both the floater cells, the adherent cells and the control cancer cells. The amount of mRNA, which indicates cell viability and growth, was measured. Cells that were treated with antibody had much less mRNA, indicating less live dividing cells (Figure 32), which confirms that anti-NME7-AB antibodies inhibit cancer cell growth as well as their transition to a more metastatic state. RT-PCR was used to measure expression levels of metastatic markers, including CXCR4. Treatment with the anti-NME7 antibodies greatly reduced the amount of metastatic markers, such as CXCR4, indicating that the anti-NME7 antibodies or peptides inhibit the transition to metastatic cancer (Figure 31 A-C). These results show that antibodies that bind to NME7-AB can be administered to a patient for the treatment or prevention of metastatic cancers.

[00165] Peptides derived from NME7-AB or NME7-X1 competitively inhibit the binding of intact NME7-AB and NME7-X1 and are anti-cancer agents.

[00166] In another aspect of the invention, therapeutic agents for the treatment or prevention of cancers are peptides derived from the NME7 sequence, which are administered to a patient for the treatment or prevention of cancers. In one aspect, the NME7-derived peptides are administered to a patient so that the peptides, which should be shorter than the entire NME7 and unable to confer the oncogenic activity of NME7, bind to the targets of NME7 and competitively inhibit the interaction of intact NME7 with its targets, wherein such interactions promote cancer. Since NME7-AB is fully able to confer oncogenic activity, the sequence of NME7-AB is preferred as the source for the shorter peptide(s), wherein it must be confirmed that the peptides themselves are not able to promote cancerous growth or other tumorigenic or oncogenic activity. In a preferred embodiment, one or more peptides having the sequence of a portion of NME7-AB and being preferably about 12-56 amino acids in length are administered to a patient. To increase half-life, the peptides may be peptide mimics, such as peptides with unnatural backbone or D-form amino acids for L. In yet another case, the anti-cancer therapeutic agent is a peptide or peptide mimic wherein the peptide has a sequence highly homologous to at least a portion of NME7, NME7-AB, or

NME7-X1 or its target the MUC1* extracellular domain, comprising the PSMGFR peptide, also called "FLR" in some cases herein.

[00167] Figures 16-19 provide a listing of preferred amino acid sequences that are predicted to inhibit NME7 binding to its cognate target. In a still more preferred embodiment, the peptides that are chosen for administration to a patient suffering from cancer or at risk of developing cancer are chosen because they bind to an NME7 binding partner and they do not themselves confer tumorigenic activity. In a yet more preferred embodiment, the NME7 binding partner is the extracellular domain of MUC1*. In a still more preferred embodiment, the NME7 binding partner is the PSMGFR peptide.

[00168] By the term "conferring tumorigenic activity or oncogenic activity", it is meant that the peptides themselves cannot support or promote the growth of cancers. Another way of testing whether or not a peptide or peptides derived from NME7 can promote tumorigenesis is to test whether or not the peptides can support pluripotent growth of human stem cells. NME proteins and peptides that support pluripotent human stem cell growth also support cancer growth. In yet another method, peptides are de-selected if they can cause somatic cells to revert to a less mature state.

[00169] Fragments of NME7-AB inhibit cancer cell growth and the transition of cancer cells to a more metastatic state. As a demonstration, NME7 peptides A1, A2, B1, B2 and B3 added separately (Figure 28) or in combinations (Figure 29) inhibit the growth of cancer cells. In addition, NME7 peptides A1, A2, B1, B2 and B3 inhibited the transition of cancer cell to a more metastatic state (Figure 31 B-C).

[00170] Thus, antibodies generated by immunizing with peptides specific to NME7, and specific to NME7-AB or NME7-X1 will block the cancerous action of NME7 species and will be potent anti-cancer agents. Similarly, these results show that the peptides specific to NME7, and specific to NME7-AB or NME7-X1 will block the cancerous action of NME7 species. In one aspect of the invention, the peptides are chosen from the list shown in Figure 16. In one aspect of the invention the peptides are chosen from the list shown in Figure 17. In one aspect of the invention the peptides are chosen from the list shown in Figure 18. In yet another aspect of the invention the peptides are chosen from the list shown in Figure 19.

[00171] Anti-NME7 antibodies for use in the treatment or prevention of cancers can be generated by standard methods known to those skilled in the art wherein those methods are used to generate antibodies or antibody-like molecules that recognize NME7, NME7-AB or a shorter form of NME7-AB wherein an additional 10-25 amino acids form the N-terminus are not present.

[00172] In another aspect of the invention, small molecules are anti-cancer agents that are selected for their ability to inhibit the tumorigenic effects of NME7, NME7-AB or NME7-X1. For example, a high throughput screen identifies small molecules that will treat cancer. In a multi-well plate, small molecules are separately added to wells in which cancer cells are cultured in a medium containing NME7-AB. If the small molecule diminishes the amount of cells that become floaters and/or reduces the expression of metastatic markers such as CXCR4, CHD1 or pluripotent stem cell markers, then that small molecule is an anti-cancer drug candidate. Another method of identifying small molecules that are anti-cancer agents is to select those small molecules that bind to NME7, NME7-AB or NME7-X1 or suppresses expression of the NME7 species. Yet another high throughput screen is to select for small molecules that inhibit the binding of NME7-AB to the PSMGFR peptide of the MUC1* extracellular domain and those small molecules will be anti-cancer agents.

[00173] The sequences of NME7-AB and NME7-X1 differ only in that NME7-X1 is missing some of the N-terminal sequence that NME7-AB has. Experiments show that there is a naturally occurring NME7 species that is nearly identical to NME7-AB, which we call NME-AB-like species. Antibodies that bind to NME7-X1 may also bind to the naturally occurring species that mimics NME7-AB, unless there are conformational differences that an antibody can differentiate. Therefore, if it is desired to inhibit NME7-X1 but not NME7-AB-like species, or vice versa, siRNA, anti-sense nucleic acids, or genetic editing techniques can be used to inhibit expression of one but not the other.

[00174] In one case, the anti-cancer therapeutic agent is a nucleic acid that directly or indirectly suppresses specific expression of NME7, NME7-X1 or NME7-AB-like species. Such nucleic acids can be siRNA, RNAi, anti-sense nucleic acids and the like that directly suppress the NME7 species. In another aspect of the invention, the nucleic acid can indirectly suppress the NME7 species for example by altering the expression of a molecule that regulates it. For example, the super enhancer BRD4 suppresses expression of NME7. Therefore, an effective therapeutic for the treatment or prevention of cancer is an agent that increases expression of BRD4. An effective therapeutic may be an agent that increases expression of BRD4's co-factor, JMJD6.

[00175] Peptides derived from NME7-AB or NME7-X1, or the entire protein, are used to generate anti-NME7 or anti-NME7-X1 antibodies in animals that we have demonstrated inhibit cancer growth and inhibit transition of cancer cells to metastatic cancer cells. Similarly, NME7 derived peptides can be administered to a human such that they generate antibodies that treat or prevent cancer or inhibit transition of cancer cells to metastatic cancer

cells. NME7 peptides or proteins are administered to a person as a type of vaccine to stimulate the production of anti-NME7, anti-NME7-AB or anti-NME7-X1 antibodies in the recipient. The results shown in Figures 28 and 29 indicate that immunizing a person with a collection of peptides derived from NME7, especially in the NME7-X1 or NME7-AB sequences may be a more effective vaccine than immunizing with a single peptide. Said peptides or proteins may further be conjugated to a carrier protein or other adjuvant, known to those skilled in the art to aid in the stimulation of an immune response.

[00176] NME7 peptides that lie outside of the DM10 domain are preferred to generate antibodies for the treatment or prevention of cancer. Peptides that can be administered to a patient for the prevention of cancer or metastasis contain sequences of the peptides listed in Figures 16-19. A1, A2, B1, B2 and B3 are examples of peptides that generate antibodies that bind to NME7-AB and NME7-X1 and are administered to a patient for the treatment or prevention of cancer. The invention is not limited to peptides of the exact sequence as is naturally occurring in NME7 or NME7-X1. As is known to those skilled in the art, substitution of several amino acids of a peptide sequence can still give rise to antibodies that specifically recognize the natural protein sequence. It is not intended that the invention be limited to the peptides demonstrated herein to inhibit cancer growth or inhibit the transition of regular cancer cells to metastatic cancer cells. The methods used here to identify peptides A1, A2, B1, B2 and B3 can also be used to identify other peptide sequences that could be equally or more effective than the peptides demonstrated here.

[00177] Chimeric antigen receptor molecules comprising portions of human NME7-AB or NME7-X1 or comprising an antibody fragment that binds to NME7-AB or NME7-X1 are anti-cancer therapeutics and are administered to a patient for the treatment or prevention of cancers.

[00178] In one instance, the recognition units or variable regions of anti-NME7 antibodies are fused to molecules of T cells using the technology known as CAR (chimeric antigen receptor) technology or CAR T technology. The salient feature of antibodies or fragments thereof that can be used therapeutically to treat or prevent cancers is the identification of antibody-like variable regions that recognize NME7 and prevent its interaction with targets that promote cancers. In one case, the target is the PSMGFR region of MUC1*.

[00179] Antibodies, antibody fragments or single chain antibodies can be engineered into chimeric molecules, including chimeric antigen receptors, also known as CARs, which molecules are then transfected or transduced into an immune system cell, such as a T cell, and administered to a patient. The humanized antibodies or antibody fragments, typically an

scFv comprises much of the extracellular domain of a CAR. The antibody fragment is biochemically fused to immune system signaling molecules, such as CD8 as the transmembrane domain and cytoplasmic signaling motifs such as T cell receptor signaling molecules also called activation domains, or co-stimulatory domains including but not limited to CD3-zeta, CD28, 41bb, OX40. CARs can be transfected into T cells or other cells, preferably immune system cells and administered to a patient. Here we describe CARs in which the extracellular portion contains an anti-NME7, anti-NME7-AB or anti-NME7-X1 antibody, antibody fragment or single chain, scFv antibody fragment. In a preferred embodiment, the antibody or antibody fragment is human or humanized.

[00180] Effective anti-NME7 or anti-NME7-X1 antibodies or fragments will have the ability to bind to native NME7, NME7-AB or NME7-X1. In practice, the parent antibody, from which the extracellular domain of the CAR is engineered, is generated by immunizing an animal with an NME7, NME7-AB or NME7-X1 derived peptide. In one aspect of the invention, the immunizing peptide is comprised of NME7 amino acids 1-376. In one aspect of the invention, the immunizing peptide is comprised of NME7 amino acids 92-376. In another aspect of the invention, the immunizing peptide is comprised of NME7 amino acids 125-376. In yet another aspect of the invention, the immunizing peptide is made up of sequences listed in Figures 16-18. In another aspect of the invention, the immunizing peptide is made up of sequences listed in Figures 19. Alternatively, the parent antibody or the antibody fragment is selected from a library or pool of antibodies, which may be natural, synthetic or fragments of either, wherein they are selected for their ability to bind to NME7, NME7-AB or NME7-X1, peptides listed in Figures 16-18, or peptides listed in Figure 19.

[00181] The targeting portion of a CAR need not be an antibody or antibody fragment. Here we describe a CAR wherein the extracellular domain contains an NME7 fragment. NME7-derived peptide(s) are engineered into a different sort of CAR wherein the targeting portion of the extracellular domain is a protein fragment or peptide rather than an antibody or antibody fragment. The peptide CARs are transfected or transduced into an immune system cell, typically a T cell. The NME7 fragments or NME7 derived peptides are selected for their ability to bind to their cognate binding partners but should not be able to function as intact NME7, NME7-AB or NME7-X1 and confer tumorigenic activity. NME7 fragments or NME7 derived peptides are biochemically fused to immune system signaling molecules, such as CD8 as the transmembrane domain and cytoplasmic signaling motifs such as T cell receptor signaling molecules also called activation domains, or co-stimulatory domains including but not limited to CD3-zeta, CD28, 41bb, OX40.

[00182] In one aspect of the invention, the NME7 fragment is most or all of the NME7 NDPK B domain. In another aspect of the invention, the NME7 fragment is an NME7 peptide that contains one or more of the peptide sequences listed in Figures 16-19. Experiments indicate that, for strategies that use NME7 or fragments of NME7, NME7-AB, or NME7-X1 as the targeting portion of a chimeric antigen receptor (CAR) for engineered immune cell therapeutics, fairly large fragments of NME7-AB or NME7-X1 would be more effective than shorter peptides, for example peptides less than 15 amino acids in length. Alternatively, a collection of CARs, each bearing a different NME7-AB derived peptide can collectively be transfected or transduced into an immune system cell and administered to a patient for the treatment or prevention of cancers. Experiments shown in Figures 28 and 29 support the validity of this approach.

[00183] CARs that contain an NME7 fragment in its extracellular domain are transfected or transduced into an immune system cell, typically a T cell, and administered to a patient for the treatment or prevention of cancers. In one aspect, the cancer is a MUC1*-positive cancer. In another aspect, the cancer is a metastatic cancer.

[00184] Agents that inhibit an enzyme that cleaves NME7 can be used to treat or prevent cancers. Some forms of NME7 are sequestered within the cell and therefore are not secreted from the cell whereupon they can act as growth factors to promote cancers. Full-length NME7 is 42kDa. However, we found that a ~33kDa NME7 species that is devoid of the DM10 domain and appears to be essentially identical to the recombinant NME7-AB that we generated, is secreted from cancer cells and stem cells. This ~33 kDa NME7 species and another ~25kDa NME7 species may be cleavage products that would be eliminated by an agent that inhibited cleavage of NME7.

[00185] The detection of elevated levels of NME7, or an ~33kDa NME7 species, which we call NME7-AB-like species, or NME7-X1 in a patient sample is diagnostic of the presence of cancer or its progression to a more aggressive or metastatic state. The inventors have discovered that both early stage, naïve stem cells and cancer cells, especially MUC1*-positive cancer cells, express high levels of a ~33kDa NME7 that is devoid of the DM10 domain and NME7-X1.

[00186] NME7-X1 was recently listed in a protein database as being a theoretical alternative isoform of NME7, however, it had never been detected in tissues or cells. We designed primers that differentiate NME7-X1 from NME7 by PCR. The expression levels of human NME7, NME7a, NME7b and NME7-X1 were measured by PCR in a panel of cells that included fibroblast cells, human embryonic stem cells, human iPS cells, T47D human

breast cancer cells, DU145 human prostate cancer cells, PC3 human prostate cancer cells, HEK295 human fetal liver cells, and other human stem cell lines. NME7 is expressed at higher levels in cancer cells than in stem cells. Particularly, NME7-X1 is expressed 10-fold higher in prostate cancer cells and 3-fold higher in breast cancer cells, than it is in fibroblast cells or stem cells. NME7-X1 is expressed ~5-fold higher in HEK293 fetal liver cells than it is in fibroblast cells or stem cells and therefore predicts that NME7-X1 is elevated in liver cancers. NME7b is expressed 17-25-times higher in prostate cancer cells than in stem cells.

[00187] Detection of elevated levels of NME7 species in a patient sample will be indicators that the patient has a cancer or is at risk of developing a cancer. Levels of NME7 species levels can be measured or assessed by PCR, hybridization schemes, cycling probe technologies, FISH, immunocytochemistry, IHC, Western blot, immunoprecipitation, sandwich assays, ELISA assays and the like. The patient sample may be a fluid sample, a blood sample, milk, urine, cells, liquid biopsy, biopsy and the like. In a patient diagnosed with cancer, elevated levels of NME7 species are indicators of increased metastatic potential. Elevated levels of NME7-X1 are indicators of prostate cancer. Antibodies of the invention are used to detect and distinguish NME7 species and are used as a diagnostic tool.

[00188] Because adult cells and tissues do not express significant levels of NME7 or secrete NME7, an effective way to diagnose cancer or to diagnose a more aggressive or metastatic form, or a shift to a more aggressive form, is to measure levels of NME7 in a sample from a patient, from a collection of cells or tissues or from cultured cells, compared to NME7 levels in a healthy sample or compared to levels of NME7 known to exist in healthy adult cells or tissues. Increased levels of NME7 indicate the presence of cancer, the presence of a metastatic cancer or the onset of metastasis. Increased levels of NME7 is also indicative of a MUC1*-positive cancer. The sample assayed for the presence of NME7 may be a collection of cells that may be cultured cell lines or cells from a patient, a bodily fluid, a blood sample, a tissue specimen, or a biopsy specimen. Therefore, a diagnostic assay that will detect the presence of cancer or the progression of cancer, comprises the steps of: 1) obtaining a sample from a patient having cancer or at risk of developing a cancer; 2) subjecting that sample to an assay capable of detecting or measuring levels of NME7, or levels of nucleic acids encoding NME7; 3) comparing levels of the measured NME7 protein or NME7-encoding nucleic acids in the test sample to levels in control patients or control cells; 4) determining that the levels of NME7 or nucleic acids encoding NME7 are elevated compared to the controls; and 5) concluding that the donor of the test sample has cancer or

has had a progression of cancer if the control to which the test was compared came from a donor previously diagnosed with a cancer.

[00189] In this assay, the control sample to which the test sample is compared can be non-cancerous cells, cultured cells, a sample from a healthy donor, a non-cancerous sample from the donor, or a sample from the donor of the test sample wherein the control sample was taken from the donor at a previous point in time. The source of such samples may be any specimen taken from the patient being tested for the presence or progression of cancer, including bodily fluids, cerebrospinal fluid, bone marrow samples, blood, tissues, cells, biopsy tissues or cells, cultured cells derived from a patient's cells and the like. The source of the sample to which the test sample is compared can be bodily fluids, cerebrospinal fluid, bone marrow samples, blood, tissues, cells, biopsy tissues or cells, or cultured cells that may be derived from a healthy donor or the test patient wherein the samples were taken at a previous point in time. The measured levels to which the test sample is compared may be from previously recorded data and compiled into lists for comparison to test samples.

[00190] Theranostics

[00191] Patients diagnosed with elevated levels of NME7 protein or nucleic acids encoding NME7 are then treated with therapeutic agents that suppress expression of NME7, inhibit cleavage of NME7 or inhibit NME7 binding to its targets, wherein such interaction promotes cancers. An important target of NME7 or a cleavage product of NME7, is MUC1*. NME7 binds to and dimerizes the extracellular domain of MUC1*. Therefore, patients diagnosed with elevated levels of NME7 will benefit from treatment with therapeutic agents that inhibit NME7 and/or therapeutic agents that inhibit the dimerization of a cleaved form of MUC1, whose extracellular domain is comprised of some or all of the PSMGFR sequence. Thus assessing suitability of cancer treatments and administration of an effective amount of a therapeutic for the treatment or prevention of cancers would consists of the steps of: 1) obtaining a sample from a patient suspected of having a cancer or at risk of developing a cancer or at risk of developing a metastatic cancer; 2) measuring an amount of NME7 or a cleavage product thereof or an NME7 encoding nucleic acid wherein the measured levels are significantly above those measured in a control sample; 3) determining that the patient has a cancer or has developed a more aggressive or a metastatic cancer; 4) administering to the patient an effective amount of a therapeutic agent that suppresses expression of NME7, inhibits cleavage of NME7 or inhibits NME7 binding to its targets and/or administering to the patient an effective amount of a therapeutic agent that suppresses expression of MUC1, inhibits cleavage of MUC1 to MUC1* or inhibits MUC1* binding to its targets. In a

preferred embodiment, the therapeutic agent that inhibits NME7 binding to its targets, inhibits its interaction with MUC1*. In a more preferred embodiments, it inhibits its interaction with the extracellular domain of MUC1* comprised essentially of the PSMGFR sequence. In a preferred embodiment, the therapeutic agent that inhibits MUC1* binding to its targets, inhibits the interaction between MUC1* and NME7. In a more preferred embodiment, the therapeutic agent that inhibits the interaction between MUC1* and NME7 inhibits the binding of MUC1* to the portion of NME7 that is comprised essentially of the sequence of NME7-AB.

[00192] Chemically modified peptides

[00193] Polypeptide or antibody therapeutics may suffer from short circulating half-life, and proteolytic degradation and low solubility. To improve the pharmacokinetics and pharmacodynamics properties of the inventive biopharmaceuticals, methods such as manipulation of the amino acid sequence may be made to decrease or increase immunogenicity and decrease proteolytic cleavage; fusion or conjugation of the peptides to immunoglobulins and serum proteins, such as albumin may be made; incorporation into drug delivery vehicles for the biopharmaceuticals such as the inventive peptides and antibodies for protection and slow release may also be made; and conjugating to natural or synthetic polymers are also contemplated. In particular, for synthetic polymer conjugation, pegylation or acylation, such as N-acylation, S-acylation and so forth are also contemplated.

[00194] Nucleic Acid Constructs

[00195] Also provided is an expression vector comprising a nucleic acid molecule of the invention as described herein, wherein the nucleic acid molecule is operatively linked to an expression control sequence. Also provided is a host-vector system for the production of a polypeptide which comprises the expression vector of the invention which has been introduced into a host cell suitable for expression of the polypeptide. The suitable host cell may be a bacterial cell such as *E. coli*, a yeast cell, such as *Pichia pastoris*, an insect cell, such as *Spodoptera frugiperda*, or a mammalian cell, such as a COS, HEK or CHO cell.

[00196] The present invention also provides for methods of producing the polypeptides of the invention by growing cells of the host-vector system described herein, under conditions permitting production of the polypeptide and recovering the polypeptide so produced. The polypeptides useful for practicing the present invention may be prepared by expression in a prokaryotic or eukaryotic expression system.

[00197] The recombinant gene may be expressed and the polypeptide purified utilizing any number of methods. The gene may be subcloned into a bacterial expression vector, such as for example, but not by way of limitation, pZErO.

[00198] The polypeptides may be purified by any technique which allows for the subsequent formation of a stable, biologically active protein. For example, and not by way of limitation, the factors may be recovered from cells either as soluble proteins or as inclusion bodies, from which they may be extracted quantitatively by 8M guanidinium hydrochloride and dialysis. In order to further purify the factors, any number of purification methods may be used, including but not limited to conventional ion exchange chromatography, affinity chromatography, different sugar chromatography, hydrophobic interaction chromatography, reverse phase chromatography or gel filtration.

[00199] When used herein, polypeptide includes functionally equivalent molecules in which amino acid residues are substituted for residues within the sequence resulting in a silent or conservative change. For example, one or more amino acid residues within the sequence can be substituted by another amino acid of a similar polarity, which acts as a functional equivalent, resulting in a silent or conservative alteration. Substitutes for an amino acid within the sequence may be selected from other members of the class to which the amino acid belongs. For example, the nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan and methionine. The polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine and glutamine. The positively charged (basic) amino acids include arginine, lysine and histidine. The negatively charged (acidic) amino acids include aspartic acid and glutamic acid. The potential glycosylation amino acids include serine, threonine, and asparagine. Also included within the scope of the invention are proteins or fragments or derivatives thereof which exhibit the same or similar biological activity and derivatives which are differentially modified during or after translation, e.g., by glycosylation, proteolytic cleavage, linkage to an antibody molecule or other cellular ligand, etc.

[00200] Any of the methods known to one skilled in the art for the insertion of DNA fragments into a vector may be used to construct expression vectors encoding the polypeptides of the invention using appropriate transcriptional/translational control signals and protein coding sequences. These methods may include *in vitro* recombinant DNA and synthetic techniques and *in vivo* recombinations (genetic recombination). Expression of nucleic acid sequence encoding the polypeptides of the invention may be regulated by a second nucleic acid sequence so that the polypeptide is expressed in a host transformed with

the recombinant DNA molecule. For example, expression of the polypeptides described herein may be controlled by any promoter/enhancer element known in the art. Promoters which may be used to control expression of the polypeptide include, but are not limited to the long terminal repeat as described in Squinto et al., (1991, Cell 65:1-20); the SV40 early promoter region (Bernoist and Chambon, 1981, Nature 290:304-310), the CMV promoter, the M-MuLV 5' terminal repeat the promoter contained in the 3'long terminal repeat of Rous sarcoma virus (Yamamoto, et al., 1980, Cell 22:787-797), the herpes thymidine kinase promoter (Wagner et al., 1981, Proc. Natl. Acad. Sci. U.S.A. 78:144-1445), the regulatory sequences of the metallothionein gene (Brinster et al., 1982, Nature 296:39-42); prokaryotic expression vectors such as the β-lactamase promoter (Villa-Kamaroff, et al., 1978, Proc. Natl. Acad. Sci. U.S.A. 75:3727-3731), or the tac promoter (DeBoer, et al., 1983, Proc. Natl. Acad. Sci. U.S.A. 80:21-25), see also "Useful proteins from recombinant bacteria" in Scientific American, 1980, 242:74-94; promoter elements from yeast or other fungi such as the Gal 4 promoter, the ADH (alcohol dehydrogenase) promoter, PGK (phosphoglycerol kinase) promoter, alkaline phosphatase promoter, and the following animal transcriptional control regions, which exhibit tissue specificity and have been utilized in transgenic animals: elastase I gene control region which is active in pancreatic acinar cells (Swift et al., 1984, Cell 38:639-646; Ornitz et al., 1986, Cold Spring Harbor Symp. Quant. Biol. 50:399-409; MacDonald, 1987, Hepatology 7:425-515); insulin gene control region which is active in pancreatic beta cells (Hanahan, 1985, Nature 315:115-122), immunoglobulin gene control region which is active in lymphoid cells (Grosschedl et al., 1984, Cell 38:647-658; Adames et al., 1985, Nature 318:533-538; Alexander et al., 1987, Mol. Cell. Biol. 7:1436-1444), mouse mammary tumor virus control region which is active in testicular, breast, lymphoid and mast cells (Leder et al., 1986, Cell 45:485-495), Sendai virus, lenti virus, albumin gene control region which is active in liver (Pinkert et al., 1987, Genes and Devel. 1:268-276), alpha-fetoprotein gene control region which is active in liver (Krumlauf et al., 1985, Mol. Cell. Biol. 5:1639-1648; Hammer et al., 1987, Science 235:53-58); alpha 1-antitrypsin gene control region which is active in the liver (Kelsey et al., 1987, Genes and Devel. 1:161-171), beta-globin gene control region which is active in myeloid cells (Mogram et al., 1985, Nature 315:338-340; Kollias et al., 1986, Cell 46:89-94); myelin basic protein gene control region which is active in oligodendrocyte cells in the brain (Readhead et al., 1987, Cell 48:703-712); myosin light chain-2 gene control region which is active in skeletal muscle (Shani, 1985, Nature 314:283-286), and gonadotropic releasing hormone gene control region which is active in the hypothalamus (Mason et al., 1986, Science 234:1372-1378).

[00201] Thus, according to the invention, expression vectors capable of being replicated in a bacterial or eukaryotic host comprising nucleic acids encoding a polypeptide as described herein, are used to transfect the host and thereby direct expression of such nucleic acid to produce polypeptides which may then be recovered in biologically active form. As used herein, a biologically active form includes a form capable of binding to the relevant receptor and causing a differentiated function and/or influencing the phenotype of the cell expressing the receptor.

[00202] Expression vectors containing the nucleic acid inserts can be identified by without limitation, at least three general approaches: (a) DNA-DNA hybridization, (b) presence or absence of "marker" gene functions, and (c) expression of inserted sequences. In the first approach, the presence of foreign nucleic acids inserted in an expression vector can be detected by DNA-DNA hybridization using probes comprising sequences that are homologous to an inserted nucleic acid sequences. In the second approach, the recombinant vector/host system can be identified and selected based upon the presence or absence of certain "marker" gene functions (e.g., thymidine kinase activity, resistance to antibiotics, transformation phenotype, occlusion body formation in baculovirus, etc.) caused by the insertion of foreign nucleic acid sequences in the vector. For example, if an *efl* nucleic acid sequence is inserted within the marker gene sequence of the vector, recombinants containing the insert can be identified by the absence of the marker gene function. In the third approach, recombinant expression vectors can be identified by assaying the foreign nucleic acid product expressed by the recombinant constructs. Such assays can be based, for example, on the physical or functional properties of the nucleic acid product of interest, for example, by binding of a ligand to a receptor or portion thereof which may be tagged with, for example, a detectable antibody or portion thereof or binding to antibodies produced against the protein of interest or a portion thereof.

[00203] The polypeptide, in particular modified of the present invention, may be expressed in the host cells transiently, constitutively or permanently.

[00204] Effective doses useful for treating the diseases or disorders indicated in the present application may be determined using methods known to one skilled in the art (see, for example, Fingl, et al., The Pharmacological Basis of Therapeutics, Goodman and Gilman, eds. Macmillan Publishing Co, New York, pp. 1-46 (1975). Pharmaceutical compositions for use according to the invention include the polypeptides described above in a pharmacologically acceptable liquid, solid or semi-solid carrier, linked to a carrier or targeting molecule (e.g., antibody, hormone, growth factor, etc.) and/or incorporated into

liposomes, microcapsules, and controlled release preparation prior to administration *in vivo*. For example, the pharmaceutical composition may comprise a polypeptide in an aqueous solution, such as sterile water, saline, phosphate buffer or dextrose solution. Alternatively, the active agents may be comprised in a solid (e.g. wax) or semi-solid (e.g. gelatinous) formulation that may be implanted into a patient in need of such treatment. The administration route may be any mode of administration known in the art, including but not limited to intravenously, intrathecally, subcutaneously, intrauterinely, by injection into involved tissue, intraarterially, intranasally, orally, or via an implanted device.

[00205] Administration may result in the distribution of the active agent of the invention throughout the body or in a localized area. For example, in some conditions, which involve distant regions of the nervous system, intravenous or intrathecal administration of agent may be desirable. In some situations, an implant containing active agent may be placed in or near the lesioned area. Suitable implants include, but are not limited to, gelfoam, wax, spray, or microparticle-based implants.

[00206] The present invention also provides for pharmaceutical compositions comprising the polypeptides described herein, in a pharmacologically acceptable vehicle. The compositions may be administered systemically or locally. Any appropriate mode of administration known in the art may be used, incuding, but not limited to, intravenous, intrathecal, intraarterial, intranasal, oral, subcutaneous, intraperitoneal, or by local injection or surgical implant. Sustained release formulations are also provided for.

[00207] **Gene Therapy**

[00208] Gene therapy refers to therapy performed by the administration to a subject of an expressed or expressible nucleic acid. In this embodiment of the invention, the nucleic acids produce their encoded protein that mediates a therapeutic effect.

[00209] Any of the methods for gene therapy available in the art can be used according to the present invention. Exemplary methods are described below.

[00210] For general reviews of the methods of gene therapy, see Goldspiel et al., Clinical Pharmacy 12:488-505 (1993); Wu and Wu, Biotherapy 3:87-95 (1991); Tolstoshev, Ann. Rev. Pharmacol. Toxicol. 32:573-596 (1993); Mulligan, Science 260:926-932 (1993); and Morgan and Anderson, Ann. Rev. Biochem. 62:191-217 (1993); May, TIBTECH 11(5):155-215 (1993). Methods commonly known in the art of recombinant DNA technology which can be used are described in Ausubel et al. (eds.), Current Protocols in Molecular Biology, John Wiley & Sons, NY (1993); and Kriegler, Gene Transfer and Expression, A Laboratory Manual, Stockton Press, NY (1990).

[00211] Delivery of the nucleic acids into a patient may be either direct, in which case the patient is directly exposed to the nucleic acid or nucleic acid- carrying vectors, or indirect, in which case, cells are first transformed with the nucleic acids *in vitro*, then transplanted into the patient. These two approaches are known, respectively, as *in vivo* or *ex vivo* gene therapy.

[00212] In a specific embodiment, the nucleic acid sequences are directly administered *in vivo*, where it is expressed to produce the encoded product. This can be accomplished by any of numerous methods known in the art, e.g., by constructing them as part of an appropriate nucleic acid expression vector and administering it so that they become intracellular, e.g., by infection using defective or attenuated retrovirals or other viral vectors, or by direct injection of naked DNA, or coating with lipids or cell-surface receptors or transfecting agents, encapsulation in liposomes, microparticles, or microcapsules, or by administering them in linkage to a peptide which is known to enter the nucleus, by administering it in linkage to a ligand subject to receptor-mediated endocytosis (see, e.g., Wu and Wu, J. Biol. Chem. 262:4429-4432 (1987)) (which can be used to target cell types specifically expressing the receptors) and so on. In another embodiment, nucleic acid-ligand complexes can be formed in which the ligand comprises a fusogenic viral peptide to disrupt endosomes, allowing the nucleic acid to avoid lysosomal degradation. In yet another embodiment, the nucleic acid can be targeted *in vivo* for cell specific uptake and expression, by targeting a specific receptor. Alternatively, the nucleic acid can be introduced intracellularly and incorporated within host cell DNA for expression, by homologous recombination (Koller and Smithies, Proc. Natl. Acad. Sci. USA 86:8932-8935 (1989); Zijlstra et al., Nature 342:435-438 (1989)).

[00213] In a specific embodiment, viral vectors that contain nucleic acid sequences encoding the polypeptide are used. The nucleic acid sequences encoding the polypeptide to be used in gene therapy are cloned into one or more vectors, which facilitates delivery of the gene into a patient. Lentiviral vectors, such as retroviral vectors, and other vectors such as adenoviral vectors and adeno-associated viruses are examples of viral vectors that may be used. Retroviral vectors contain the components necessary for the correct packaging of the viral genome and integration into the host cell DNA.

[00214] Adenoviruses are especially attractive vehicles for delivering genes to respiratory epithelia because they naturally infect respiratory epithelia where they cause a mild disease. Other targets for adenovirus-based delivery systems are liver, the central nervous system, endothelial cells, and muscle. Adenoviruses have the advantage of being capable of infecting non-dividing cells. In addition, adeno-associated virus (AAV) has also been proposed for use in gene therapy.

[00215] Another approach to gene therapy involves transferring a gene to cells in tissue culture by such methods as electroporation, lipofection, calcium phosphate mediated transfection, or viral infection. Usually, the method of transfer includes the transfer of a selectable marker to the cells. The cells are then placed under selection to isolate those cells that have taken up and are expressing the transferred gene. Those cells are then delivered to a patient.

[00216] In this embodiment, the nucleic acid is introduced into a cell prior to administration *in vivo* of the resulting recombinant cell. Such introduction can be carried out by any method known in the art, including but not limited to transfection, electroporation, microinjection, infection with a viral or bacteriophage vector containing the nucleic acid sequences, cell fusion, chromosome-mediated gene transfer, microcell-mediated gene transfer, spheroplast fusion and so on. Numerous techniques are known in the art for the introduction of foreign genes into cells and may be used in accordance with the present invention, provided that the necessary developmental and physiological functions of the recipient cells are not disrupted. The technique should provide for the stable transfer of the nucleic acid to the cell, so that the nucleic acid is expressible by the cell and preferably heritable and expressible by its cell progeny.

[00217] Cells into which a nucleic acid can be introduced for purposes of gene therapy encompass any desired, available cell type, and include but are not limited to epithelial cells, endothelial cells, keratinocytes, fibroblasts, muscle cells, hepatocytes; blood cells such as T-lymphocytes, B-lymphocytes, monocytes, macrophages, neutrophils, eosinophils, megakaryocytes, granulocytes; various stem or progenitor cells, in particular hematopoietic stem or progenitor cells, e.g., as obtained from bone marrow, umbilical cord blood, peripheral blood, fetal liver, and so on.

[00218] In a preferred embodiment, the cell used for gene therapy is autologous to the patient.

[00219] In an embodiment in which recombinant cells are used in gene therapy, nucleic acid sequences encoding the polypeptide are introduced into the cells such that they are expressible by the cells or their progeny, and the recombinant cells are then administered *in vivo* for therapeutic effect. In a specific embodiment, stem or progenitor cells are used. Any stem and/or progenitor cells which can be isolated and maintained *in vitro* can potentially be used in accordance with this embodiment of the present invention.

[00220] In a specific embodiment, the nucleic acid to be introduced for purposes of gene therapy comprises an inducible promoter operably linked to the coding region, such that

expression of the nucleic acid is controllable by controlling the presence or absence of the appropriate inducer of transcription.

[00221] Therapeutic Composition

[00222] The formulation of therapeutic compounds is generally known in the art and reference can conveniently be made to Remington's Pharmaceutical Sciences, 17th ed., Mack Publishing Co., Easton, Pa., USA. For example, from about 0.05 ng to about 20 mg per kilogram of body weight per day may be administered. Dosage regime may be adjusted to provide the optimum therapeutic response. For example, several divided doses may be administered daily or the dose may be proportionally reduced as indicated by the exigencies of the therapeutic situation. The active compound may be administered in a convenient manner such as by the oral, intravenous (where water soluble), intramuscular, subcutaneous, intra nasal, intra ocular, intradermal or suppository routes or implanting (eg using slow release molecules by the intraperitoneal route or by using cells e.g. monocytes or dendrite cells sensitized *in vitro* and adoptively transferred to the recipient). Depending on the route of administration, the peptide may be required to be coated in a material to protect it from the action of enzymes, acids and other natural conditions which may inactivate said ingredients.

[00223] For example, the low lipophilicity of the peptides will allow them to be destroyed in the gastrointestinal tract by enzymes capable of cleaving peptide bonds and in the stomach by acid hydrolysis. In order to administer peptides by other than parenteral administration, they will be coated by, or administered with, a material to prevent its inactivation. For example, peptides may be administered in an adjuvant, co-administered with enzyme inhibitors or in liposomes. Adjuvants contemplated herein include resorcinols, non-ionic surfactants such as polyoxyethylene oleyl ether and n-hexadecyl polyethylene ether. Enzyme inhibitors include pancreatic trypsin inhibitor, diisopropylfluorophosphate (DEP) and trasylol. Liposomes include water-in-oil-in-water CGF emulsions as well as conventional liposomes.

[00224] The active compounds may also be administered parenterally or intraperitoneally. Dispersions can also be prepared in glycerol liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations contain a preservative to prevent the growth of microorganisms.

[00225] The pharmaceutical forms suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. In all cases the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the

conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol and liquid polyethylene glycol, and the like), suitable mixtures thereof, and vegetable oils. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of superfactants. The prevention of the action of microorganisms can be brought about by various antibacterial and antifungal agents, for example, chlorobutanol, phenol, sorbic acid, theomersal and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the composition of agents delaying absorption, for example, aluminium monostearate and gelatin.

[00226] Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with various other ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the various sterile active ingredient into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and the freeze-drying technique which yield a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

[00227] When the peptides are suitably protected as described above, the active compound may be orally administered, for example, with an inert diluent or with an assimilable edible carrier, or it may be enclosed in hard or soft shell gelatin capsule, or it may be compressed into tablets, or it may be incorporated directly with the food of the diet. For oral therapeutic administration, the active compound may be incorporated with excipients and used in the form of ingestible tablets, buccal tablets, troches, capsules, elixirs, suspensions, syrups, wafers, and the like. Such compositions and preparations should contain at least 1% by weight of active compound. The percentage of the compositions and preparations may, of course, be varied and may conveniently be between about 5 to about 80% of the weight of the unit. The amount of active compound in such therapeutically useful compositions is such that a suitable dosage will be obtained. Preferred compositions or preparations according to the present invention are prepared so that an oral dosage unit form contains between about 0.1 μg and 2000 mg of active compound.

[00228] The tablets, pills, capsules and the like may also contain the following: A binder such as gum tragacanth, acacia, corn starch or gelatin; excipients such as dicalcium phosphate; a disintegrating agent such as corn starch, potato starch, alginic acid and the like; a lubricant such as magnesium stearate; and a sweetening agent such as sucrose, lactose or saccharin may be added or a flavoring agent such as peppermint, oil of wintergreen, or cherry flavoring. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar or both. A syrup or elixir may contain the active compound, sucrose as a sweetening agent, methyl and propylparabens as preservatives, a dye and flavoring such as cherry or orange flavor. Of course, any material used in preparing any dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In addition, the active compound may be incorporated into sustained-release preparations and formulations.

[00229] **Delivery Systems**

[00230] Various delivery systems are known and can be used to administer a compound of the invention, e.g., encapsulation in liposomes, microparticles, microcapsules, recombinant cells capable of expressing the compound, receptor-mediated endocytosis, construction of a nucleic acid as part of a retroviral or other vector, etc. Methods of introduction include but are not limited to intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, intranasal, intra ocular, epidural, and oral routes. The compounds or compositions may be administered by any convenient route, for example by infusion or bolus injection, by absorption through epithelial or mucocutaneous linings (e.g., oral mucosa, rectal and intestinal mucosa, etc.) and may be administered together with other biologically active agents. Administration can be systemic or local. In addition, it may be desirable to introduce the pharmaceutical compounds or compositions of the invention into the central nervous system by any suitable route, including intraventricular and intrathecal injection; intraventricular injection may be facilitated by an intraventricular catheter, for example, attached to a reservoir, such as an Ommaya reservoir. Pulmonary administration can also be employed, e.g., by use of an inhaler or nebulizer, and formulation with an aerosolizing agent.

[00231] In a specific embodiment, it may be desirable to administer the pharmaceutical compounds or compositions of the invention locally to the area in need of treatment; this may be achieved by, for example, and not by way of limitation, local infusion during surgery, topical application, e.g., in conjunction with a wound dressing after surgery, by injection, by

means of a catheter, by means of a suppository, or by means of an implant, said implant being of a porous, non-porous, or gelatinous material, including membranes, such as sialastic membranes, or fibers. Preferably, when administering a protein, including an antibody or a peptide of the invention, care must be taken to use materials to which the protein does not absorb. In another embodiment, the compound or composition can be delivered in a vesicle, in particular a liposome. In yet another embodiment, the compound or composition can be delivered in a controlled release system. In one embodiment, a pump may be used. In another embodiment, polymeric materials can be used. In yet another embodiment, a controlled release system can be placed in proximity of the therapeutic target, thus requiring only a fraction of the systemic dose.

[00232] Sequence Listing Free Text

[00233] As regards the use of nucleotide symbols other than a, g, c, t, they follow the convention set forth in WIPO Standard ST.25, Appendix 2, Table 1, wherein k represents t or g; n represents a, c, t or g; m represents a or c; r represents a or g; s represents c or g; w represents a or t and y represents c or t.

[00234] MTPGTQSPFF LLLLLTVLTV VTGSGHASST PGGEKETSAT QRSSVPSSTE KNAVSMTSSV LSSHSPGSGS STTQGQDVTL APATEPASGS AATWGQDVTS VPVTRPALGS TTPPAHDVTS APDNKPAPGS TAPPAHGVTS APDTRPAPGS TAPPAHGVTS APDNRPALGS

TAPPVHNVTS ASGSASGSAS TLVHNGTSAR ATTTPASKST PFSIPSHHSD TPTTLASHST KTDASSTHHS SVPPLTSSNH STSPQLSTGV SFFFLSFHIS NLQFNSSLED PSTDYYQELQ RDISEMFLQI YKQGGFLGLS NIKFRPGSVV VQLTLAFREG TINVHDVETQ FNQYKTEAAS RYNLTISDVS VSDVPFPFSA QSGAGVPGWG IALLVLVCVL VALAIVYLIA LAVCQCRRKN YGQLDIFPAR DTYHPMSEYP TYHTHGRYVP PSSTDRSPYE KVSAGNGGSS LSYTNPAVAA ASANL (SEQ ID NO:1) describes full-length MUC1 Receptor (Mucin 1 precursor, Genbank Accession number: P15941).

[00235] MTPGTQSPFFLLLLLTVLT (SEQ ID NO:2)

[00236] MTPGTQSPFFLLLLLTVLT VVTA (SEQ ID NO:3)

[00237] MTPGTQSPFFLLLLLTVLT VVTG (SEQ ID NO:4)

[00238] SEQ ID NOS:2, 3 and 4 describe N-terminal MUC-1 signaling sequence for directing MUC1 receptor and truncated isoforms to cell membrane surface. Up to 3 amino acid residues may be absent at C-terminal end as indicated by variants in SEQ ID NOS:2, 3 and 4.

[00239] GTINVHDVETQFNQYKTEAASRYNLTISDVSVSDVPFPFSAQSGAGVPGW GIALLVLVCVLVALAIVYLIALAVCQCRRKNYGQLDIFPARDTYHPMSEYPTYHTHG RYVPPSSTDRSPYEKVSAGNGGSSLSYTNPAVAAASANL (SEQ ID NO:5) describes a truncated MUC1 receptor isoform having nat-PSMGFR at its N-terminus and including the transmembrane and cytoplasmic sequences of a full-length MUC1 receptor.

[00240] GTINVHDVETQFNQYKTEAASRYNLTISDVSVSDVPFPFSAQSGA (SEQ ID NO:6) describes the extracellular domain of Native Primary Sequence of the MUC1 Growth Factor Receptor (nat-PSMGFR – an example of "PSMGFR"):

[00241] TINVHDVETQFNQYKTEAASRYNLTISDVSVSDVPFPFSAQSGA (SEQ ID NO:7) describes the extracellular domain of Native Primary Sequence of the MUC1 Growth Factor Receptor (nat-PSMGFR – An example of "PSMGFR"), having a single amino acid deletion at the N-terminus of SEQ ID NO:6).

[00242] GTINVHDVETQFNQYKTEAASPYNLTISDVSVSDVPFPFSAQSGA (SEQ ID NO:8) describes the extracellular domain of "SPY" functional variant of the native Primary Sequence of the MUC1 Growth Factor Receptor having enhanced stability (var-PSMGFR – An example of "PSMGFR").

[00243] TINVHDVETQFNQYKTEAASPYNLTISDVSVSDVPFPFSAQSGA (SEQ ID NO:9) describes the extracellular domain of "SPY" functional variant of the native Primary Sequence of the MUC1 Growth Factor Receptor having enhanced stability (var-PSMGFR –

An example of "PSMGFR"), having a single amino acid deletion at the C-terminus of SEQ ID NO:8).

[00244] tgtcagtgccgccgaaagaactacgggcagctggacatctttccagcccgggatacctaccatcctatgagcgagta ccccacctaccacacccatgggcgctatgtgccccctagcagtaccgatcgtagcccctatgagaaggtttctgcaggtaacggtggc agcagcctctcttacacaaacccagcagtggcagccgcttctgccaacttg (SEQ ID NO:10) describes MUC1 cytoplasmic domain nucleotide sequence.

[00245] CQCRRKNYGQLDIFPARDTYHPMSEYPTYHTHGRYVPPSSTDRSPYEKVS AGNGGSSLSYTNPAVAAASANL (SEQ ID NO:11) describes MUC1 cytoplasmic domain amino acid sequence.

[00246] gagatcctgagacaatgaatcatagtgaaagattcgttttcattgcagagtggtatgatccaaatgcttcacttcttcgac gttatgagcttttatttacccaggggatggatctgttgaaatgcatgatgtaaagaatcatcgcacctttttaaagcggaccaaatatgata acctgcacttggaagatttatttataggcaacaaagtgaatgtcttttctcgacaactggtattaattgactatggggatcaatatacagctc gccagctgggcagtaggaaagaaaaaacgctagccctaattaaaccagatgcaatatcaaaggctggagaaataattgaaataataa acaaagctggatttactataaccaaactcaaaatgatgatgctttcaaggaaagaagcattggattttcatgtagatcaccagtcaagacc ctttttcaatgagctgatccagtttattacaactggtcctattattgccatggagattttaagagatgatgctatatgtgaatggaaaagactg ctgggacctgcaaactctggagtggcacgcacagatgcttctgaaagcattagagccctctttggaacagatggcataagaaatgcag cgcatggccctgattcttttgcttctgcggccagagaaatggagttgtttttttccttcaagtggaggttgtgggccggcaaacactgctaa atttactaattgtacctgttgcattgttaaaccccatgctgtcagtgaaggtatgttgaatacactatattcagtacattttgttaataggagag caatgtttattttcttgatgtactttatgtatagaaaataa (SEQ ID NO:12) describes NME7 nucleotide sequence (NME7: GENBANK ACCESSION AB209049).

[00247] DPETMNHSERFVFIAEWYDPNASLLRRYELLFYPGDGSVEMHDVKNHRT FLKRTKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTARQLGSRKEKTLALIKPDAI SKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQSRPFFNELIQFITTGPIIAMEIL RDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIRNAAHGPDSFASAAREMELF FPSSGGCGPANTAKFTNCTCCIVKPHAVSEGMLNTLYSVHFVNRRAMFIFLMYFMY RK (SEQ ID NO:13) describes NME7 amino acid sequence (NME7: GENBANK ACCESSION AB209049).

[00248] atggtgctactgtctactttagggatcgtctttcaaggcgaggggcctcctatctcaagctgtgatacaggaaccatggc caactgtgagcgtaccttcattgcgatcaaaccagatggggtccagcggggtcttgtgggagagattatcaagcgttttgagcagaaag gattccgccttgttggtctgaaattcatgcaagcttccgaagatcttctcaaggaacactacgttgacctgaaggaccgtccattctttgcc ggcctggtgaaatacatgcactcagggccggtagttgccatggtctgggaggggctgaatgtggtgaagacgggccgagtcatgctc ggggagaccaaccctgcagactccaagcctgggaccatccgtggagacttctgcatacaagttggcaggaacattatacatggcagt gattctgtggagagtgcagagaaggagatcggcttgtggtttcaccctgaggaactggtagattacacgagctgtgctcagaactggat

ctatgaatga (SEQ ID NO:14) describes NM23-H1 nucleotide sequence (NM23-H1: GENBANK ACCESSION AF487339).

[00249] MVLLSTLGIVFQGEGPPISSCDTGTMANCERTFIAIKPDGVQRGLVGEIIKR FEQKGFRLVGLKFMQASEDLLKEHYVDLKDRPFFAGLVKYMHSGPVVAMVWEGL NVVKTGRVMLGETNPADSKPGTIRGDFCIQVGRNIIHGSDSVESAEKEIGLWFHPEEL VDYTSCAQNWIYE (SEQ ID NO:15) NM23-H1 describes amino acid sequence (NM23-H1: GENBANK ACCESSION AF487339).

[00250] atggtgctactgtctactttagggatcgtctttcaaggcgaggggcctcctatctcaagctgtgatacaggaaccatggc caactgtgagcgtaccttcattgcgatcaaaccagatggggtccagcggggtcttgtgggagagattatcaagcgttttgagcagaaag gattccgccttgttggtctgaaattcatgcaagcttccgaagatcttctcaaggaacactacgttgacctgaaggaccgtccattctttgcc ggcctggtgaaatacatgcactcagggccggtagttgccatggtctgggaggggctgaatgtggtgaagacgggccgagtcatgctc ggggagaccaaccctgcagactccaagcctgggaccatccgtggagacttctgcatacaagttggcaggaacattatacatggcggt gattctgtggagagtgcagagaaggagatcggcttgtggtttcaccctgaggaactggtagattacacgagctgtgctcagaactggat ctatgaatga (SEQ ID NO:16) describes NM23-H1 S120G mutant nucleotide sequence (NM23-H1: GENBANK ACCESSION AF487339).

[00251] MVLLSTLGIVFQGEGPPISSCDTGTMANCERTFIAIKPDGVQRGLVGEIIKR FEQKGFRLVGLKFMQASEDLLKEHYVDLKDRPFFAGLVKYMHSGPVVAMVWEGL NVVKTGRVMLGETNPADSKPGTIRGDFCIQVGRNIIHGGDSVESAEKEIGLWFHPEEL VDYTSCAQNWIYE (SEQ ID NO:17) describes NM23-H1 S120G mutant amino acid sequence (NM23-H1: GENBANK ACCESSION AF487339).

[00252] atggccaacctggagcgcaccttcatcgccatcaagccggacggcgtgcagcgcggcctggtgggcgagatcatc aagcgcttcgagcagaagggattccgcctcgtggccatgaagttcctccgggcctctgaagaacacctgaagcagcactacattgac ctgaaagaccgaccattcttccctgggctggtgaagtacatgaactcagggccggttgtggccatggtctgggaggggctgaacgtg gtgaagacaggccgagtgatgcttggggagaccaatccagcagattcaaagccaggcaccattcgtggggacttctgcattcaggtt ggcaggaacatcattcatggcagtgattcagtaaaaagtgctgaaaaagaaatcagcctatggtttaagcctgaagaactggttgacta caagtcttgtgctcatgactgggtctatgaataa (SEQ ID NO:18) describes NM23-H2 nucleotide sequence (NM23-H2: GENBANK ACCESSION AK313448).

[00253] MANLERTFIAIKPDGVQRGLVGEIIKRFEQKGFRLVAMKFLRASEEHLKQH YIDLKDRPFFPGLVKYMNSGPVVAMVWEGLNVVKTGRVMLGETNPADSKPGTIRG DFCIQVGRNIIHGSDSVKSAEKEISLWFKPEELVDYKSCAHDWVYE (SEQ ID NO:19) describes NM23-H2 amino acid sequence (NM23-H2: GENBANK ACCESSION AK313448).

[00254] Human NM23-H7-2 sequence optimized for *E. coli* expression:

[00255] (DNA)

[00256] atgcatgacgttaaaaatcaccgtacctttctgaaacgcacgaaatatgataatctgcatctggaagacctgtttattggc aacaaagtcaatgtgttctctcgtcagctggtgctgatcgattatggcgaccagtacaccgcgcgtcaactgggtagtcgcaaagaaaa aacgctggccctgattaaaccggatgcaatctccaaagctggcgaaattatcgaaattatcaacaaagcgggtttcaccatcacgaaac tgaaaatgatgatgctgagccgtaaagaagccctggattttcatgtcgaccaccagtctcgcccgtttttcaatgaactgattcaattcatc accacgggtccgattatcgcaatggaaattctgcgtgatgacgctatctgcgaatggaaacgcctgctgggcccggcaaactcaggtg ttgcgcgtaccgatgccagtgaatccattcgcgctctgtttggcaccgatggtatccgtaatgcagcacatggtccggactcattcgcat cggcagctcgtgaaatggaactgtttttcccgagctctggcggttgcggtccggcaaacaccgccaaatttaccaattgtacgtgctgta ttgtcaaaccgcacgcagtgtcagaaggcctgctgggtaaaattctgatggcaatccgtgatgctggctttgaaatctcggccatgcag atgttcaacatggaccgcgttaacgtcgaagaattctacgaagtttacaaaggcgtggttaccgaatatcacgatatggttacggaaatg tactccggtccgtgcgtcgcgatggaaattcagcaaaacaatgccaccaaaacgtttcgtgaattctgtggtccggcagatccggaaat cgcacgtcatctgcgtccgggtaccctgcgcgcaatttttggtaaaacgaaaatccagaacgctgtgcactgtaccgatctgccggaa gacggtctgctggaagttcaatacttttttcaaaattctggataattga (SEQ ID NO:20)

[00257] (amino acids)

[00258] MHDVKNHRTFLKRTKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTAR QLGSRKEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQSRP FFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIRNA AHGPDSFASAAREMELFFPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILMAIR DAGFEISAMQMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQNN ATKTFREFCGPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFKILD N- (SEQ ID NO:21)

[00259] Human NME7-A:

[00260] (DNA)

[00261] atggaaaaaacgctagccctaattaaaccagatgcaatatcaaaggctggagaaataattgaaataataaacaaagct ggatttactataaccaaactcaaaatgatgatgctttcaaggaaagaagcattggattttcatgtagatcaccagtcaagacccttttttcaat gagctgatccagtttattacaactggtcctattattgccatggagattttaagagatgatgctatatgtgaatggaaaagactgctgggacc tgcaaactctggagtggcacgcacagatgcttctgaaagcattagagccctctttggaacagatggcataagaaatgcagcgcatggc cctgattcttttgcttctgcggccagagaaatggagttgtttttttga (SEQ ID NO:22)

[00262] (amino acids)

[00263] MEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQS RPFFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIR NAAHGPDSFASAAREMELFF- (SEQ ID NO:23)

[00264] Human NME7-A1:

[00265] (DNA)

[00266] atggaaaaaacgctagccctaattaaaccagatgcaatatcaaaggctggagaaataattgaaataataaacaaagct ggatttactataaccaaactcaaaatgatgatgctttcaaggaaagaagcattggattttcatgtagatcaccagtcaagacccttttttcaat gagctgatccagtttattacaactggtcctattattgccatggagattttaagagatgatgctatatgtgaatggaaaagactgctgggacc tgcaaactctggagtggcacgcacagatgcttctgaaagcattagagccctctttggaacagatggcataagaaatgcagcgcatggc cctgattcttttgcttctgcggccagagaaatggagttgtttttttccttcaagtggaggttgtgggccggcaaacactgctaaatttacttga (SEQ ID NO:24)

[00267] (amino acids)

[00268] MEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQS RPFFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIR NAAHGPDSFASAAREMELFFPSSGGCGPANTAKFT- (SEQ ID NO:25)

[00269] Human NME7-A2:

[00270] (DNA)

[00271] atgaatcatagtgaaagattcgttttcattgcagagtggtatgatccaaatgcttcacttcttcgacgttatgagcttttatttt acccaggggatggatctgttgaaatgcatgatgtaaagaatcatcgcacctttttaaagcggaccaaatatgataacctgcacttggaag atttatttataggcaacaaagtgaatgtcttttctcgacaactggtattaattgactatggggatcaatatacagctcgccagctgggcagta ggaaagaaaaaacgctagccctaattaaaccagatgcaatatcaaaggctggagaaataattgaaataataaacaaagctggatttact ataaccaaactcaaaatgatgatgctttcaaggaaagaagcattggattttcatgtagatcaccagtcaagacccttttttcaatgagctgat ccagtttattacaactggtcctattattgccatggagattttaagagatgatgctatatgtgaatggaaaagactgctgggacctgcaaact ctggagtggcacgcacagatgcttctgaaagcattagagccctctttggaacagatggcataagaaatgcagcgcatggccctgattct ttgcttctgcggccagagaaatggagttgtttttttga (SEQ ID NO:26)

[00272] (amino acids)

[00273] MNHSERFVFIAEWYDPNASLLRRYELLFYPGDGSVEMHDVKNHRTFLKR TKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTARQLGSRKEKTLALIKPDAISKA GEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQSRPFFNELIQFITTGPIIAMEILRDD AICEWKRLLGPANSGVARTDASESIRALFGTDGIRNAAHGPDSFASAAREMELFF- (SEQ ID NO:27)

[00274] Human NME7-A3:

[00275] (DNA)

[00276] atgaatcatagtgaaagattcgttttcattgcagagtggtatgatccaaatgcttcacttcttcgacgttatgagcttttatttt acccaggggatggatctgttgaaatgcatgatgtaaagaatcatcgcacctttttaaagcggaccaaatatgataacctgcacttggaag atttatttataggcaacaaagtgaatgtcttttctcgacaactggtattaattgactatggggatcaatatacagctcgccagctgggcagta ggaaagaaaaaacgctagccctaattaaaccagatgcaatatcaaaggctggagaaataattgaaataataaacaaagctggatttact ataaccaaactcaaaatgatgatgctttcaaggaaagaagcattggattttcatgtagatcaccagtcaagacccttttttcaatgagctgat ccagtttattacaactggtcctattattgccatggagattttaagagatgatgctatatgtgaatggaaaagactgctgggacctgcaaact

ctggagtggcacgcacagatgcttctgaaagcattagagccctctttggaacagatggcataagaaatgcagcgcatggccctgattct tttgcttctgcggccagagaaatggagttgtttttttccttcaagtggaggttgtgggccggcaaacactgctaaatttacttga (SEQ ID NO:28)

[00277] (amino acids)

[00278] MNHSERFVFIAEWYDPNASLLRRYELLFYPGDGSVEMHDVKNHRTFLKR TKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTARQLGSRKEKTLALIKPDAISKA GEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQSRPFFNELIQFITTGPIIAMEILRDD AICEWKRLLGPANSGVARTDASESIRALFGTDGIRNAAHGPDSFASAAREMELFFPSS GGCGPANTAKFT- (SEQ ID NO:29)

[00279] Human NME7-B:

[00280] (DNA)

[00281] atgaattgtacctgttgcattgttaaaccccatgctgtcagtgaaggactgttgggaaagatcctgatggctatccgaga tgcaggttttgaaatctcagctatgcagatgttcaatatggatcgggttaatgttgaggaattctatgaagtttataaaggagtagtgaccg aatatcatgacatggtgacagaaatgtattctggcccttgtgtagcaatggagattcaacagaataatgctacaaagacatttcgagaattt tgtggacctgctgatcctgaaattgcccggcatttacgccctggaactctcagagcaatctttggtaaaactaagatccagaatgctgttc actgtactgatctgccagaggatggcctattagaggttcaatacttcttctga (SEQ ID NO:30)

[00282] (amino acids)

[00283] MNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAMQMFNMDRVNVEEFY EVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFCGPADPEIARHLRPGTL RAIFGKTKIQNAVHCTDLPEDGLLEVQYFF- (SEQ ID NO:31)

[00284] Human NME7-B1:

[00285] (DNA)

[00286] atgaattgtacctgttgcattgttaaaccccatgctgtcagtgaaggactgttgggaaagatcctgatggctatccgaga tgcaggttttgaaatctcagctatgcagatgttcaatatggatcgggttaatgttgaggaattctatgaagtttataaaggagtagtgaccg aatatcatgacatggtgacagaaatgtattctggcccttgtgtagcaatggagattcaacagaataatgctacaaagacatttcgagaattt tgtggacctgctgatcctgaaattgcccggcatttacgccctggaactctcagagcaatctttggtaaaactaagatccagaatgctgttc actgtactgatctgccagaggatggcctattagaggttcaatacttcttcaagatcttggataattagtga (SEQ ID NO:32)

[00287] (amino acids)

[00288] MNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAMQMFNMDRVNVEEFY EVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFCGPADPEIARHLRPGTL RAIFGKTKIQNAVHCTDLPEDGLLEVQYFFKILDN— (SEQ ID NO:33)

[00289] Human NME7-B2:

[00290] (DNA)

[00291] atgccttcaagtggaggttgtgggccggcaaacactgctaaatttactaattgtacctgttgcattgttaaaccccatgct gtcagtgaaggactgttgggaaagatcctgatggctatccgagatgcaggttttgaaatctcagctatgcagatgttcaatatggatcgg gttaatgttgaggaattctatgaagtttataaaggagtagtgaccgaatatcatgacatggtgacagaaatgtattctggcccttgtgtagc aatggagattcaacagaataatgctacaaagacatttcgagaattttgtggacctgctgatcctgaaattgcccggcatttacgccctgga actctcagagcaatctttggtaaaactaagatccagaatgctgttcactgtactgatctgccagaggatggcctattagaggttcaatactt cttctga (SEQ ID NO:34)

[00292] (amino acids)

[00293] MPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAM QMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFC GPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFF- (SEQ ID NO:35)

[00294] Human NME7-B3:

[00295] (DNA)

[00296] atgccttcaagtggaggttgtgggccggcaaacactgctaaatttactaattgtacctgttgcattgttaaaccccatgct gtcagtgaaggactgttgggaaagatcctgatggctatccgagatgcaggttttgaaatctcagctatgcagatgttcaatatggatcgg gttaatgttgaggaattctatgaagtttataaaggagtagtgaccgaatatcatgacatggtgacagaaatgtattctggcccttgtgtagc aatggagattcaacagaataatgctacaaagacatttcgagaattttgtggacctgctgatcctgaaattgcccggcatttacgccctgga actctcagagcaatctttggtaaaactaagatccagaatgctgttcactgtactgatctgccagaggatggcctattagaggttcaatactt cttcaagatcttggataattagtga (SEQ ID NO:36)

[00297] (amino acids)

[00298] MPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAM QMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFC GPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFKILDN-- (SEQ ID NO:37)

[00299] Human NME7-AB:

[00300] (DNA)

[00301] atggaaaaaacgctagccctaattaaaccagatgcaatatcaaaggctggagaaataattgaaataataaacaaagct ggatttactataaccaaactcaaaatgatgatgctttcaaggaaagaagcattggattttcatgtagatcaccagtcaagaccctttttcaat gagctgatccagtttattacaactggtcctattattgccatggagattttaagagatgatgctatatgtgaatggaaaagactgctgggacc tgcaaactctggagtggcacgcacagatgcttctgaaagcattagagccctctttggaacagatggcataagaaatgcagcgcatggc cctgattcttttgcttctgcggccagagaaatggagttgtttttttccttcaagtggaggttgtgggccggcaaacactgctaaatttactaatt gtacctgttgcattgttaaaccccatgctgtcagtgaaggactgttgggaaagatcctgatggctatccgagatgcaggttttgaaatctc agctatgcagatgttcaatatggatcgggttaatgttgaggaattctatgaagtttataaaggagtagtgaccgaatatcatgacatggtga cagaaatgtattctggcccttgtgtagcaatggagattcaacagaataatgctacaaagacatttcgagaattttgtggacctgctgatcct

gaaattgcccggcatttacgccctggaactctcagagcaatctttggtaaaactaagatccagaatgctgttcactgtactgatctgccag aggatggcctattagaggttcaatacttcttcaagatcttggataattagtga (SEQ ID NO:38)

[00302] (amino acids)

[00303] MEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQS RPFFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIR NAAHGPDSFASAAREMELFFPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILM AIRDAGFEISAMQMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQ NNATKTFREFCGPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFK ILDN-- (SEQ ID NO:39)

[00304] Human NME7-AB1:

[00305] (DNA)

[00306] atggaaaaaacgctagccctaattaaaccagatgcaatatcaaaggctggagaaataattgaaataataaacaaagct ggatttactataaccaaactcaaaatgatgatgctttcaaggaaagaagcattggattttcatgtagatcaccagtcaagacccttttttcaat gagctgatccagtttattacaactggtcctattattgccatggagattttaagagatgatgctatatgtgaatggaaaagactgctgggacc tgcaaactctggagtggcacgcacagatgcttctgaaagcattagagccctctttggaacagatggcataagaaatgcagcgcatggc cctgattcttttgcttctgcggccagagaaatggagttgtttttttccttcaagtggaggttgtgggccggcaaacactgctaaatttactaatt gtacctgttgcattgttaaaccccatgctgtcagtgaaggactgttgggaaagatcctgatggctatccgagatgcaggttttgaaatctc agctatgcagatgttcaatatggatcgggttaatgttgaggaattctatgaagtttataaaggagtagtgaccgaatatcatgacatggtga cagaaatgtattctggcccttgtgtagcaatggagattcaacagaataatgctacaaagacatttcgagaattttgtggacctgctgatcct gaaattgcccggcatttacgccctggaactctcagagcaatctttggtaaaactaagatccagaatgctgttcactgtactgatctgccag aggatggcctattagaggttcaatacttcttctga (SEQ ID NO:40)

[00307] (amino acids)

[00308] MEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQS RPFFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIR NAAHGPDSFASAAREMELFFPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILM AIRDAGFEISAMQMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQ NNATKTFREFCGPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFF- (SEQ ID NO:41)

[00309] Human NME7-A sequence optimized for *E. coli* expression:

[00310] (DNA)

[00311] atggaaaaaacgctggccctgattaaaccggatgcaatctccaaagctggcgaaattatcgaaattatcaacaaagcg ggtttcaccatcacgaaactgaaaatgatgatgctgagccgtaaagaagccctggattttcatgtcgaccaccagtctcgcccgtttttca atgaactgattcaattcatcaccacgggtccgattatcgcaatggaaattctgcgtgatgacgctatctgcgaatggaaacgcctgctgg

gcccggcaaactcaggtgttgcgcgtaccgatgccagtgaatccattcgcgctctgtttggcaccgatggtatccgtaatgcagcacat ggtccggactcattcgcatcggcagctcgtgaaatggaactgtttttctga (SEQ ID NO:42)

[00312] (amino acids)

[00313] MEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQS RPFFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIR NAAHGPDSFASAAREMELFF- (SEQ ID NO:43)

[00314] Human NME7-A1 sequence optimized for *E. coli* expression:

[00315] (DNA)

[00316] atggaaaaaacgctggccctgattaaaccggatgcaatctccaaagctggcgaaattatcgaaattatcaacaaagcg ggtttcaccatcacgaaactgaaaatgatgatgctgagccgtaaagaagccctggattttcatgtcgaccaccagtctcgcccgtttttca atgaactgattcaattcatcaccacgggtccgattatcgcaatggaaattctgcgtgatgacgctatctgcgaatggaaacgcctgctgg gcccggcaaactcaggtgttgcgcgtaccgatgccagtgaatccattcgcgctctgtttggcaccgatggtatccgtaatgcagcacat ggtccggactcattcgcatcggcagctcgtgaaatggaactgtttttcccgagctctggcggttgcggtccggcaaacaccgccaaatt tacctga (SEQ ID NO:44)

[00317] (amino acids)

[00318] MEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQS RPFFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIR NAAHGPDSFASAAREMELFFPSSGGCGPANTAKFT- (SEQ ID NO:45)

[00319] Human NME7-A2 sequence optimized for *E. coli* expression:

[00320] (DNA)

[00321] atgaatcactccgaacgctttgtttttatcgccgaatggtatgacccgaatgcttccctgctgcgccgctacgaactgct gttttatccgggcgatggtagcgtggaaatgcatgacgttaaaaatcaccgtacctttctgaaacgcacgaaatatgataatctgcatctg gaagacctgtttattggcaacaaagtcaatgtgttctctcgtcagctggtgctgatcgattatggcgaccagtacaccgcgcgtcaactg ggtagtcgcaaagaaaaaacgctggccctgattaaaccggatgcaatctccaaagctggcgaaattatcgaaattatcaacaaagcgg gtttcaccatcacgaaactgaaaatgatgatgctgagccgtaaagaagccctggattttcatgtcgaccaccagtctcgcccgtttttcaa tgaactgattcaattcatcaccacgggtccgattatcgcaatggaaattctgcgtgatgacgctatctgcgaatggaaacgcctgctggg cccggcaaactcaggtgttgcgcgtaccgatgccagtgaatccattcgcgctctgtttggcaccgatggtatccgtaatgcagcacatg gtccggactcattcgcatcggcagctcgtgaaatggaactgtttttctga (SEQ ID NO:46)

[00322] (amino acids)

[00323] MNHSERFVFIAEWYDPNASLLRRYELLFYPGDGSVEMHDVKNHRTFLKR TKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTARQLGSRKEKTLALIKPDAISKA GEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQSRPFFNELIQFITTGPIIAMEILRDD AICEWKRLLGPANSGVARTDASESIRALFGTDGIRNAAHGPDSFASAAREMELFF- (SEQ ID NO:47)

[00324] Human NME7-A3 sequence optimized for *E. coli* expression:

[00325] (DNA)

[00326] atgaatcactccgaacgctttgtttttatcgccgaatggtatgacccgaatgcttccctgctgcgccgctacgaactgct gttttatccgggcgatggtagcgtggaaatgcatgacgttaaaaatcaccgtacctttctgaaacgcacgaaatatgataatctgcatctg gaagacctgtttattggcaacaaagtcaatgtgttctctcgtcagctggtgctgatcgattatggcgaccagtacaccgcgcgtcaactg ggtagtcgcaaagaaaaaacgctggccctgattaaaccggatgcaatctccaaagctggcgaaattatcgaaattatcaacaaagcgg gtttcaccatcacgaaactgaaaatgatgatgctgagccgtaaagaagccctggattttcatgtcgaccaccagtctcgcccgtttttcaa tgaactgattcaattcatcaccacgggtccgattatcgcaatggaaattctgcgtgatgacgctatctgcgaatggaaacgcctgctggg cccggcaaactcaggtgttgcgcgtaccgatgccagtgaatccattcgcgctctgtttggcaccgatggtatccgtaatgcagcacatg gtccggactcattcgcatcggcagctcgtgaaatggaactgtttttcccgagctctggcggttgcggtccggcaaacaccgccaaatttt acctga (SEQ ID NO:48)

[00327] (amino acids)

[00328] MNHSERFVFIAEWYDPNASLLRRYELLFYPGDGSVEMHDVKNHRTFLKR TKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTARQLGSRKEKTLALIKPDAISKA GEIIEIINKAGFTITKLKMMMMLSRKEALDFHVDHQSRPFFNELIQFITTGPIIAMEILRDD AICEWKRLLGPANSGVARTDASESIRALFGTDGIRNAAHGPDSFASAAREMELFFPSS GGCGPANTAKFT- (SEQ ID NO:49)

[00329] Human NME7-B sequence optimized for *E. coli* expression:

[00330] (DNA)

[00331] atgaattgtacgtgctgtattgtcaaaccgcacgcagtgtcagaaggcctgctgggtaaaattctgatggcaatccgtg atgctggctttgaaatctcggccatgcagatgttcaacatggaccgcgttaacgtcgaagaattctacgaagtttacaaaggcgtggtta ccgaatatcacgatatggttacggaaatgtactccggtccgtgcgtcgcgatggaaattcagcaaaacaatgccaccaaaacgtttcgt gaattctgtggtccggcagatccggaaatcgcacgtcatctgcgtccgggtaccctgcgcgcaatttttggtaaaacgaaaatccagaa cgctgtgcactgtaccgatctgccggaagacggtctgctggaagttcaatacttttttctga (SEQ ID NO:50)

[00332] (amino acids)

[00333] MNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAMQMFNMDRVNVEEFY EVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFCGPADPEIARHLRPGTL RAIFGKTKIQNAVHCTDLPEDGLLEVQYFF- (SEQ ID NO:51)

[00334] Human NME7-B1 sequence optimized for *E. coli* expression:

[00335] (DNA)

[00336] atgaattgtacgtgctgtattgtcaaaccgcacgcagtgtcagaaggcctgctgggtaaaattctgatggcaatccgtg atgctggctttgaaatctcggccatgcagatgttcaacatggaccgcgttaacgtcgaagaattctacgaagtttacaaaggcgtggtta ccgaatatcacgatatggttacggaaatgtactccggtccgtgcgtcgcgatggaaattcagcaaaacaatgccaccaaaacgtttcgt gaattctgtggtccggcagatccggaaatcgcacgtcatctgcgtccgggtaccctgcgcgcaatttttggtaaaacgaaaatccagaa

cgctgtgcactgtaccgatctgccggaagacggtctgctggaagttcaatacttttcaaaattctggataattga (SEQ ID NO:52)

[00337] (amino acids)

[00338] MNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAMQMFNMDRVNVEEFY EVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFCGPADPEIARHLRPGTL RAIFGKTKIQNAVHCTDLPEDGLLEVQYFFKILDN- (SEQ ID NO:53)

[00339] Human NME7-B2 sequence optimized for *E. coli* expression:

[00340] (DNA)

[00341] atgccgagctctggcggttgcggtccggcaaacaccgccaaatttaccaattgtacgtgctgtattgtcaaaccgcac gcagtgtcagaaggcctgctgggtaaaattctgatggcaatccgtgatgctggctttgaaatctcggccatgcagatgttcaacatggac cgcgttaacgtcgaagaattctacgaagtttacaaaggcgtggttaccgaatatcacgatatggttacggaaatgtactccggtccgtgc gtcgcgatggaaattcagcaaaacaatgccaccaaaacgtttcgtgaattctgtggtccggcagatccggaaatcgcacgtcatctgcg tccgggtaccctgcgcgcaatttttggtaaaacgaaaatccagaacgctgtgcactgtaccgatctgccggaagacggtctgctggaa gttcaatacttttctga (SEQ ID NO:54)

[00342] (amino acids)

[00343] MPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAM QMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFC GPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFF- (SEQ ID NO:55)

[00344] Human NME7-B3 sequence optimized for *E. coli* expression:

[00345] (DNA)

[00346] atgccgagctctggcggttgcggtccggcaaacaccgccaaatttaccaattgtacgtgctgtattgtcaaaccgcac gcagtgtcagaaggcctgctgggtaaaattctgatggcaatccgtgatgctggctttgaaatctcggccatgcagatgttcaacatggac cgcgttaacgtcgaagaattctacgaagtttacaaaggcgtggttaccgaatatcacgatatggttacggaaatgtactccggtccgtgc gtcgcgatggaaattcagcaaaacaatgccaccaaaacgtttcgtgaattctgtggtccggcagatccggaaatcgcacgtcatctgcg tccgggtaccctgcgcgcaatttttggtaaaacgaaaatccagaacgctgtgcactgtaccgatctgccggaagacggtctgctggaa gttcaatacttttcaaaattctggataattga (SEQ ID NO:56)

[00347] (amino acids)

[00348] MPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAM QMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFC GPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFKILDN- (SEQ ID NO:57)

[00349] Human NME7-AB sequence optimized for *E. coli* expression:

[00350] (DNA)

[00351] atggaaaaaacgctggccctgattaaaccggatgcaatctccaaagctggcgaaattatcgaaattatcaacaaagcg ggtttcaccatcacgaaactgaaaatgatgatgctgagccgtaaagaagccctggattttcatgtcgaccaccagtctcgcccgtttttca atgaactgattcaattcatcaccacgggtccgattatcgcaatggaaattctgcgtgatgacgctatctgcgaatggaaacgcctgctgg gcccggcaaactcaggtgttgcgcgtaccgatgccagtgaatccattcgcgctctgtttggcaccgatggtatccgtaatgcagcacat ggtccggactcattcgcatcggcagctcgtgaaatggaactgtttttcccgagctctggcggttgcggtccggcaaacaccgccaaatt taccaattgtacgtgctgtattgtcaaaccgcacgcagtgtcagaaggcctgctgggtaaaattctgatggcaatccgtgatgctggcttt gaaatctcggccatgcagatgttcaacatggaccgcgttaacgtcgaagaattctacgaagtttacaaaggcgtggttaccgaatatca cgatatggttacggaaatgtactccggtccgtgcgtcgcgatggaaattcagcaaaacaatgccaccaaaacgtttcgtgaattctgtgg tccggcagatccggaaatcgcacgtcatctgcgtccgggtaccctgcgcgcaatttttggtaaaacgaaaatccagaacgctgtgcact gtaccgatctgccggaagacggtctgctggaagttcaatacttttttcaaaattctggataattga (SEQ ID NO:58)

[00352] (amino acids)

[00353] MEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQS RPFFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIR NAAHGPDSFASAAREMELFFPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILM AIRDAGFEISAMQMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQ NNATKTFREFCGPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFK ILDN- (SEQ ID NO:59)

[00354] Human NME7-AB1 sequence optimized for *E. coli* expression:

[00355] (DNA)

[00356] Atggaaaaaacgctggccctgattaaaccggatgcaatctccaaagctggcgaaattatcgaaattatcaacaaagc gggtttcaccatcacgaaactgaaaatgatgatgctgagccgtaaagaagccctggattttcatgtcgaccaccagtctcgcccgttttc aatgaactgattcaattcatcaccacgggtccgattatcgcaatggaaattctgcgtgatgacgctatctgcgaatggaaacgcctgctg ggcccggcaaactcaggtgttgcgcgtaccgatgccagtgaatccattcgcgctctgtttggcaccgatggtatccgtaatgcagcaca tggtccggactcattcgcatcggcagctcgtgaaatggaactgtttttcccgagctctggcggttgcggtccggcaaacaccgccaaat ttaccaattgtacgtgctgtattgtcaaaccgcacgcagtgtcagaaggcctgctgggtaaaattctgatggcaatccgtgatgctggctt tgaaatctcggccatgcagatgttcaacatggaccgcgttaacgtcgaagaattctacgaagtttacaaaggcgtggttaccgaatatca cgatatggttacggaaatgtactccggtccgtgcgtcgcgatggaaattcagcaaaacaatgccaccaaaacgtttcgtgaattctgtgg tccggcagatccggaaatcgcacgtcatctgcgtccgggtaccctgcgcgcaatttttggtaaaacgaaaatccagaacgctgtgcact gtaccgatctgccggaagacggtctgctggaagttcaatacttttttctga (SEQ ID NO:60)

[00357] (amino acids)

[00358] MEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQS RPFFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIR NAAHGPDSFASAAREMELFFPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILM AIRDAGFEISAMQMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQ

NNATKTFREFCGPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFF- (SEQ ID NO:61)

[00359] Mouse NME6

[00360] (DNA)

[00361] Atgacctccatcttgcgaagtccccaagctcttcagctcacactagccctgatcaagcctgatgcagttgcccaccca ctgatcctggaggctgttcatcagcagattctgagcaacaagttcctcattgtacgaacgagggaactgcagtggaagctggaggact gccggaggttttaccgagagcatgaagggcgttttttctatcagcggctggtggagttcatgacaagtgggccaatccgagcctatatc cttgcccacaaagatgccatccaactttggaggacactgatgggacccaccagagtatttcgagcacgctatatagccccagattcaat tcgtggaagtttgggcctcactgacacccgaaatactacccatggctcagactccgtggtttccgccagcagagagattgcagccttctt ccctgacttcagtgaacagcgctggtatgaggaggaggaaccccagctgcggtgtggtcctgtgcactacagtccagaggaaggtat ccactgtgcagctgaaacaggaggccacaaacaacctaacaaaacctag (SEQ ID NO:62)

[00362] (amino acids)

[00363] MTSILRSPQALQLTLALIKPDAVAHPLILEAVHQQILSNKFLIVRTRELQWK LEDCRRFYREHEGRFFYQRLVEFMTSGPIRAYILAHKDAIQLWRTLMGPTRVFRARY IAPDSIRGSLGLTDTRNTTHGSDSVVSASREIAAFFPDFSEQRWYEEEEPQLRCGPVHY SPEEGIHCAAETGGHKQPNKT- (SEQ ID NO:63)

[00364] Human NME6:

[00365] (DNA)

[00366] Atgacccagaatctggggagtgagatggcctcaatcttgcgaagccctcaggctctccagctcactctagccctgat caagcctgacgcagtcgcccatccactgattctggaggctgttcatcagcagattctaagcaacaagttcctgattgtacgaatgagag aactactgtggagaaaggaagattgccagaggttttaccgagagcatgaagggcgttttttctatcagaggctggtggagttcatggcc agcgggccaatccgagcctacatccttgcccacaaggatgccatccagctctggaggacgctcatgggacccaccagagtgttccga gcacgccatgtggccccagattctatccgtgggagtttcggcctcactgacacccgcaacaccacccatggttcggactctgtggtttc agccagcagagagattgcagccttcttccctgacttcagtgaacagcgctggtatgaggaggaagagccccagttgcgctgtggccct gtgtgctatagcccagagggaggtgtccactatgtagctggaacaggaggcctaggaccagcctga (SEQ ID NO:64)

[00367] (amino acids)

[00368] MTQNLGSEMASILRSPQALQLTLALIKPDAVAHPLILEAVHQQILSNKFLIV RMRELLWRKEDCQRFYREHEGRFFYQRLVEFMASGPIRAYILAHKDAIQLWRTLMG PTRVFRARHVAPDSIRGSFGLTDTRNTTHGSDSVVSASREIAAFFPDFSEQRWYEEEE PQLRCGPVCYSPEGGVHYVAGTGGLGPA- (SEQ ID NO:65)

[00369] Human NME6 1:

[00370] (DNA)

[00371] Atgacccagaatctggggagtgagatggcctcaatcttgcgaagccctcaggctctccagctcactctagccctgat caagcctgacgcagtcgcccatccactgattctggaggctgttcatcagcagattctaagcaacaagttcctgattgtacgaatgagag

aactactgtggagaaaggaagattgccagaggttttaccgagagcatgaagggcgttttttctatcagaggctggtggagttcatggcc agcgggccaatccgagcctacatccttgcccacaaggatgccatccagctctggaggacgctcatgggacccaccagagtgttccga gcacgccatgtggccccagattctatccgtgggagtttcggcctcactgacacccgcaacaccacccatggttcggactctgtggtttc agccagcagagagattgcagccttcttccctgacttcagtgaacagcgctggtatgaggaggaagagccccagttgcgctgtggccct gtgtga (SEQ ID NO:66)

[00372] (amino acids)

[00373] MTQNLGSEMASILRSPQALQLTLALIKPDAVAHPLILEAVHQQILSNKFLIV RMRELLWRKEDCQRFYREHEGRFFYQRLVEFMASGPIRAYILAHKDAIQLWRTLMG PTRVFRARHVAPDSIRGSFGLTDTRNTTHGSDSVVSASREIAAFFPDFSEQRWYEEEE PQLRCGPV- (SEQ ID NO:67)

[00374] Human NME6 2:

[00375] (DNA)

[00376] Atgctcactctagccctgatcaagcctgacgcagtcgcccatccactgattctggaggctgttcatcagcagattctaa gcaacaagttcctgattgtacgaatgagagaactactgtggagaaaggaagattgccagaggttttaccgagagcatgaagggcgtttt ttctatcagaggctggtggagttcatggccagcgggccaatccgagcctacatccttgcccacaaggatgccatccagctctggagga cgctcatgggacccaccagagtgttccgagcacgccatgtggccccagattctatccgtgggagtttcggcctcactgacacccgcaa caccacccatggttcggactctgtggtttcagccagcagagagattgcagccttcttccctgacttcagtgaacagcgctggtatgagg aggaagagccccagttgcgctgtggccctgtgtga (SEQ ID NO:68)

[00377] (amino acids)

[00378] MLTLALIKPDAVAHPLILEAVHQQILSNKFLIVRMRELLWRKEDCQRFYRE HEGRFFYQRLVEFMASGPIRAYILAHKDAIQLWRTLMGPTRVFRARHVAPDSIRGSF GLTDTRNTTHGSDSVVSASREIAAFFPDFSEQRWYEEEEPQLRCGPV- (SEQ ID NO:69)

[00379] Human NME6 3:

[00380] (DNA)

[00381] Atgctcactctagccctgatcaagcctgacgcagtcgcccatccactgattctggaggctgttcatcagcagattctaa gcaacaagttcctgattgtacgaatgagagaactactgtggagaaaggaagattgccagaggttttaccgagagcatgaagggcgtttt ttctatcagaggctggtggagttcatggccagcgggccaatccgagcctacatccttgcccacaaggatgccatccagctctggagga cgctcatgggacccaccagagtgttccgagcacgccatgtggccccagattctatccgtgggagtttcggcctcactgacacccgcaa caccacccatggttcggactctgtggtttcagccagcagagagattgcagccttcttccctgacttcagtgaacagcgctggtatgagg aggaagagccccagttgcgctgtggccctgtgtgctatagcccagagggaggtgtccactatgtagctggaacaggaggcctagga ccagcctga (SEQ ID NO:70)

[00382] (amino acids)

[00383] MLTLALIKPDAVAHPLILEAVHQQILSNKFLIVRMRELLWRKEDCQRFYRE HEGRFFYQRLVEFMASGPIRAYILAHKDAIQLWRTLMGPTRVFRARHVAPDSIRGSF GLTDTRNTTHGSDSVVSASREIAAFFPDFSEQRWYEEEEPQLRCGPVCYSPEGGVHY VAGTGGLGPA- (SEQ ID NO:71)

[00384] Human NME6 sequence optimized for *E. coli* expression:

[00385] (DNA)

[00386] Atgacgcaaaatctgggctcggaaatggcaagtatcctgcgctccccgcaagcactgcaactgaccctggctctgat caaaccggacgctgttgctcatccgctgattctggaagcggtccaccagcaaattctgagcaacaaatttctgatcgtgcgtatgcgcg aactgctgtggcgtaaagaagattgccagcgtttttatcgcgaacatgaaggccgtttcttttatcaacgcctggttgaattcatggcctct ggtccgattcgcgcatatatcctggctcacaaagatgcgattcagctgtggcgtaccctgatgggtccgacgcgcgtctttcgtgcacgt catgtggcaccggactcaatccgtggctcgttcggtctgaccgatacgcgcaataccacgcacggtagcgactctgttgttagtgcgtc ccgtgaaatcgcggccttttcccggacttctccgaacagcgttggtacgaagaagaagaaccgcaactgcgctgtggcccggtctgtt attctccggaaggtggtgtccattatgtggcgggcacgggtggtctgggtccggcatga (SEQ ID NO:72)

[00387] (amino acids)

[00388] MTQNLGSEMASILRSPQALQLTLALIKPDAVAHPLILEAVHQQILSNKFLIV RMRELLWRKEDCQRFYREHEGRFFYQRLVEFMASGPIRAYILAHKDAIQLWRTLMG PTRVFRARHVAPDSIRGSFGLTDTRNTTHGSDSVVSASREIAAFFPDFSEQRWYEEEE PQLRCGPVCYSPEGGVHYVAGTGGLGPA- (SEQ ID NO:73)

[00389] Human NME6 1 sequence optimized for *E. coli* expression:

[00390] (DNA)

[00391] Atgacgcaaaatctgggctcggaaatggcaagtatcctgcgctccccgcaagcactgcaactgaccctggctctgat caaaccggacgctgttgctcatccgctgattctggaagcggtccaccagcaaattctgagcaacaaatttctgatcgtgcgtatgcgcg aactgctgtggcgtaaagaagattgccagcgtttttatcgcgaacatgaaggccgtttcttttatcaacgcctggttgaattcatggcctct ggtccgattcgcgcatatatcctggctcacaaagatgcgattcagctgtggcgtaccctgatgggtccgacgcgcgtctttcgtgcacgt catgtggcaccggactcaatccgtggctcgttcggtctgaccgatacgcgcaataccacgcacggtagcgactctgttgttagtgcgtc ccgtgaaatcgcggccttttcccggacttctccgaacagcgttggtacgaagaagaagaaccgcaactgcgctgtggcccggtctga (SEQ ID NO:74)

[00392] (amino acids)

[00393] MTQNLGSEMASILRSPQALQLTLALIKPDAVAHPLILEAVHQQILSNKFLIV RMRELLWRKEDCQRFYREHEGRFFYQRLVEFMASGPIRAYILAHKDAIQLWRTLMG PTRVFRARHVAPDSIRGSFGLTDTRNTTHGSDSVVSASREIAAFFPDFSEQRWYEEEE PQLRCGPV- (SEQ ID NO:75)

[00394] Human NME6 2 sequence optimized for *E. coli* expression:

[00395] (DNA)

[00396] Atgctgaccctggctctgatcaaaccggacgctgttgctcatccgctgattctggaagcggtccaccagcaaattctg agcaacaaatttctgatcgtgcgtatgcgcgaactgctgtggcgtaaagaagattgccagcgtttttatcgcgaacatgaaggccgtttct ttatcaacgcctggttgaattcatggcctctggtccgattcgcgcatatatcctggctcacaaagatgcgattcagctgtggcgtaccctg atgggtccgacgcgcgtctttcgtgcacgtcatgtggcaccggactcaatccgtggctcgttcggtctgaccgatacgcgcaataccac gcacggtagcgactctgttgttagtgcgtcccgtgaaatcgcggccttttcccggacttctccgaacagcgttggtacgaagaagaag aaccgcaactgcgctgtggcccggtctga (SEQ ID NO:76)

[00397] (amino acids)

[00398] MLTLALIKPDAVAHPLILEAVHQQILSNKFLIVRMRELLWRKEDCQRFYRE HEGRFFYQRLVEFMASGPIRAYILAHKDAIQLWRTLMGPTRVFRARHVAPDSIRGSF GLTDTRNTTHGSDSVVSASREIAAFFPDFSEQRWYEEEEPQLRCGPV- (SEQ ID NO:77)

[00399] Human NME6 3 sequence optimized for *E. coli* expression:

[00400] (DNA)

[00401] Atgctgaccctggctctgatcaaaccggacgctgttgctcatccgctgattctggaagcggtccaccagcaaattctg agcaacaaatttctgatcgtgcgtatgcgcgaactgctgtggcgtaaagaagattgccagcgtttttatcgcgaacatgaaggccgtttct ttatcaacgcctggttgaattcatggcctctggtccgattcgcgcatatatcctggctcacaaagatgcgattcagctgtggcgtaccctg atgggtccgacgcgcgtctttcgtgcacgtcatgtggcaccggactcaatccgtggctcgttcggtctgaccgatacgcgcaataccac gcacggtagcgactctgttgttagtgcgtcccgtgaaatcgcggccttttcccggacttctccgaacagcgttggtacgaagaagaag aaccgcaactgcgctgtggcccggtctgttattctccggaaggtggtgtccattatgtggcgggcacgggtggtctgggtccggcatg a (SEQ ID NO:78)

[00402] (amino acids)

[00403] MLTLALIKPDAVAHPLILEAVHQQILSNKFLIVRMRELLWRKEDCQRFYRE HEGRFFYQRLVEFMASGPIRAYILAHKDAIQLWRTLMGPTRVFRARHVAPDSIRGSF GLTDTRNTTHGSDSVVSASREIAAFFPDFSEQRWYEEEEPQLRCGPVCYSPEGGVHY VAGTGGLGPA- (SEQ ID NO:79)

[00404] OriGene-NME7-1 full length

[00405] (DNA)

[00406] gacgttgtatacgactcctatagggcggccgggaattcgtcgactggatccggtaccgaggagatctgccgccgcg atcgccatgaatcatagtgaaagattcgttttcattgcagagtggtatgatccaaatgcttcacttcttcgacgttatgagcttttatttttaccc aggggatggatctgttgaaatgcatgatgtaaagaatcatcgcacctttttaaagcggaccaaatatgataacctgcacttggaagattta tttataggcaacaaagtgaatgtcttctctcgacaactggtattaattgactatggggatcaatatacagctcgccagctgggcagtagga aagaaaaaacgctagccctaattaaaccagatgcaatatcaaaggctggagaaataattgaaataataaacaaagctggatttactataa ccaaactcaaaatgatgatgctttcaaggaaagaagcattggattttcatgtagatcaccagtcaagacccttttcaatgagctgatcca gtttattacaactggtcctattattgccatggagattttaagagatgatgctatatgtgaatggaaaagactgctgggacctgcaaactctg

gagtggcacgcacagatgcttctgaaagcattagagccctctttggaacagatggcataagaaatgcagcgcatggccctgattctttt gcttctgcggccagagaaatggagttgtttttttccttcaagtggaggttgtgggccggcaaacactgctaaatttactaattgtacctgttg cattgttaaaccccatgctgtcagtgaaggactgttgggaaagatcctgatggctatccgagatgcaggttttgaaatctcagctatgcag atgttcaatatggatcgggttaatgttgaggaattctatgaagtttataaaggagtagtgaccgaatatcatgacatggtgacagaaatgta ttctggcccttgtgtagcaatggagattcaacagaataatgctacaaagacatttcgagaattttgtggacctgctgatcctgaaattgccc ggcatttacgccctggaactctcagagcaatctttggtaaaactaagatccagaatgctgttcactgtactgatctgccagaggatggcct attagaggttcaatacttcttcaagatcttggataatacgcgtacgcggccgctcgagcagaaactcatctcagaagaggatctggcag caaatgatatcctggattacaaggatgacgacgataaggtttaa (SEQ ID NO:80)

[00407] (amino acids)

[00408] MNHSERFVFIAEWYDPNASLLRRYELLFYPGDGSVEMHDVKNHRTFLKR TKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTARQLGSRKEKTLALIKPDAISKA GEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQSRPFFNELIQFITTGPIIAMEILRDD AICEWKRLLGPANSGVARTDASESIRALFGTDGIRNAAHGPDSFASAAREMELFFPSS GGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAMQMFNMDRVNV EEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFCGPADPEIARHLR PGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFKILDNTRTRRLEQKLISEEDLAAN DILDYKDDDDKV (SEQ ID NO:81)

[00409] Abnova NME7-1 Full length (amino acids)

[00410] MNHSERFVFIAEWYDPNASLLRRYELLFYPGDGSVEMHDVKNHRTFLKR TKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTARQLGSRKEKTLALIKPDAISKA GEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQSRPFFNELIQFITTGPIIAMEILRDD AICEWKRLLGPANSGVARTDASESIRALFGTDGIRNAAHGPDSFASAAREMELFFPSS GGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAMQMFNMDRVNV EEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFCGPADPEIARHLR PGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFKILDN (SEQ ID NO:82)

[00411] Abnova Partial NME7-B

[00412] (amino acids)

[00413] DRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREF CGPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFKIL (SEQ ID NO:83)

[00414] Histidine Tag

[00415] (ctcgag)caccaccaccaccaccactga (SEQ ID NO:84)

[00416] Strept II Tag

[00417] (accggt)tggagccatcctcagttcgaaaagtaatga (SEQ ID NO:85)

[00418] N-10 peptide:

[00419] QFNQYKTEAASRYNLTISDVSVSDVPFPFSAQSGA (SEQ ID NO:86)

[00420] C-10 peptide

[00421] GTINVHDVETQFNQYKTEAASRYNLTISDVSVSDV (SEQ ID NO:87)

[00422] LALIKPDA (SEQ ID NO:88)

[00423] MMMLSRKEALDFHVDHQS (SEQ ID NO:89)

[00424] ALDFHVDHQS (SEQ ID NO:90)

[00425] EILRDDAICEWKRL (SEQ ID NO:91)

[00426] FNELIQFITTGP (SEQ ID NO:92)

[00427] RDDAICEW (SEQ ID NO:93)

[00428] SGVARTDASESIRALFGTDGIRNAA (SEQ ID NO:94)

[00429] ELFFPSSGG (SEQ ID NO:95)

[00430] KFTNCTCCIVKPHAVSEGLLGKILMA (SEQ ID NO:96)

[00431] LMAIRDAGFEISAMQMFNMDRVNVEEFYEVYKGVVT (SEQ ID NO:97)

[00432] EFYEVYKGVVTEYHD (SEQ ID NO:98)

[00433] EIQQNNATKTFREFCGPADPEIARHLRPGTLRAIFGKTKIQNA (SEQ ID NO:99)

[00434] YSGPCVAM (SEQ ID NO:100)

[00435] FREFCGP (SEQ ID NO:101)

[00436] VHCTDLPEDGLLEVQYFFKILDN (SEQ ID NO:102)

[00437] IQNAVHCTD (SEQ ID NO:103)

[00438] TDLPEDGLLEVQYFFKILDN (SEQ ID NO:104)

[00439] PEDGLLEVQYFFK (SEQ ID NO:105)

[00440] EIINKAGFTITK (SEQ ID NO:106)

[00441] MLSRKEALDFHVDHQS (SEQ ID NO:107)

[00442] NELIQFITT (SEQ ID NO:108)

[00443] EILRDDAICEWKRL (SEQ ID NO:109)

[00444] SGVARTDASESIRALFGTDGI (SEQ ID NO:110)

[00445] SGVARTDASES (SEQ ID NO:111)

[00446] ALFGTDGI (SEQ ID NO:112)

[00447] NCTCCIVKPHAVSE (SEQ ID NO:113)

[00448] LGKILMAIRDA (SEQ ID NO:114)

[00449] EISAMQMFNMDRVNVE (SEQ ID NO:115)

[00450] EVYKGVVT (SEQ ID NO:116)

[00451] EYHDMVTE (SEQ ID NO:117)

[00452] EFCGPADPEIARHLR (SEQ ID NO:118)

[00453] AIFGKTKIQNAV (SEQ ID NO:119)

[00454] LPEDGLLEVQYFFKILDN (SEQ ID NO:120)

[00455] GPDSFASAAREMELFFP (SEQ ID NO:121)

[00456] Immunizing peptides derived from human NME7

[00457] ICEWKRL (SEQ ID NO:122)

[00458] LGKILMAIRDA (SEQ ID NO:123)

[00459] HAVSEGLLGK (SEQ ID NO:124)

[00460] VTEMYSGP (SEQ ID NO:125)

[00461] NATKTFREF (SEQ ID NO:126)

[00462] AIRDAGFEI (SEQ ID NO:127)

[00463] AICEWKRLLGPAN (SEQ ID NO:128)

[00464] DHQSRPFF (SEQ ID NO:129)

[00465] AICEWKRLLGPAN (SEQ ID NO:130)

[00466] VDHQSRPF (SEQ ID NO:131)

[00467] PDSFAS (SEQ ID NO:132)

[00468] KAGEIIEIINKAGFTITK (SEQ ID NO:133)

[00469] Immunizing peptides derived from human NME1

[00470] MANCERTFIAIKPDGVQRGLVGEIIKRFE (SEQ ID NO:134)

[00471] VDLKDRPF (SEQ ID NO:135)

[00472] HGSDSVESAEKEIGLWF (SEQ ID NO:136)

[00473] ERTFIAIKPDGVQRGLVGEIIKRFE (SEQ ID NO:137)

[00474] VDLKDRPFFAGLVKYMHSGPVVAMVWEGLN (SEQ ID NO:138)

[00475] NIIHGSDSVESAEKEIGLWFHPEELV (SEQ ID NO:139)

[00476] KPDGVQRGLVGEII (SEQ ID NO:140)

[00477] Immunizing peptide derived from human NME7, but which does not bind NME1

[00478] MLSRKEALDFHVDHQS (SEQ ID NO:141) peptide A1

[00479] SGVARTDASES (SEQ ID NO:142) peptide A2

[00480] DAGFEISAMQMFNMDRVNVE (SEQ ID NO:143) peptide B1

[00481] EVYKGVVTEYHDMVTE (SEQ ID NO:144) peptide B2

[00482] AIFGKTKIQNAVHCTDLPEDGLLEVQYFF (SEQ ID NO:145) peptide B3

[00483] Human NME7 a

[00484] (DNA)

[00485] atgaatcatagtgaaagattcgttttcattgcagagtggtatgatccaaatgcttcacttcttcgacgttatgagcttttatttt acccaggggatggatctgttgaaatgcatgatgtaaagaatcatcgcacctttttaaagcggaccaaatatgataacctgcacttggaag atttatttataggcaacaaagtgaatgtcttttctcgacaactggtattaattgactatggggatcaatatacagctcgccagctgggcagta ggaaagaaaaaacgctagccctaattaaaccagatgcaatatcaaaggctggagaaataattgaaataataaacaaagctggatttact ataaccaaactcaaaatgatgatgctttcaaggaaagaagcattggattttcatgtagatcaccagtcaagacccttttcaatgagctgat ccagtttattacaactggtcctattattgccatggagattttaagagatgatgctatatgtgaatggaaaagactgctgggacctgcaaact ctggagtggcacgcacagatgcttctgaaagcattagagccctctttggaacagatggcataagaaatgcagcgcatggccctgattct ttgcttctgcggccagagaaatggagttgtttttttccttcaagtggaggttgtgggccggcaaacactgctaaatttactaattgtacctgtt gcattgttaaaccccatgctgtcagtgaaggactgttgggaaagatcctgatggctatccgagatgcaggttttgaaatctcagctatgca gatgttcaatatggatcgggttaatgttgaggaattctatgaagtttataaaggagtagtgaccgaatatcatgacatggtgacagaaatgt attctggcccttgtgtagcaatggagattcaacagaataatgctacaaagacatttcgagaattttgtggacctgctgatcctgaaattgcc cggcatttacgccctggaactctcagagcaatctttggtaaaactaagatccagaatgctgttcactgtactgatctgccagaggatggc ctattagaggttcaatacttcttcaagatcttggataattag (SEQ ID NO:146)

[00486] (amino acids)

[00487] MNHSERFVFIAEWYDPNASLLRRYELLFYPGDGSVEMHDVKNHRTFLKR TKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTARQLGSRKEKTLALIKPDAISKA GEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQSRPFFNELIQFITTGPIIAMEILRDD AICEWKRLLGPANSGVARTDASESIRALFGTDGIRNAAHGPDSFASAAREMELFFPSS GGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAMQMFNMDRVNV EEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFCGPADPEIARHLR PGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFKILDN (SEQ ID NO:147)

[00488] Human NME7 b

[00489] (DNA)

[00490] atgcatgatgtaaagaatcatcgcacctttttaaagcggaccaaatatgataacctgcacttggaagatttatttataggc aacaaagtgaatgtcttttctcgacaactggtattaattgactatggggatcaatatacagctcgccagctgggcagtaggaaagaaaaa acgctagccctaattaaaccagatgcaatatcaaaggctggagaaataattgaaataataaacaaagctggatttactataaccaaactc aaaatgatgatgctttcaaggaaagaagcattggattttcatgtagatcaccagtcaagacccttttcaatgagctgatccagtttattaca actggtcctattattgccatggagattttaagagatgatgctatatgtgaatggaaaagactgctgggacctgcaaactctggagtggca cgcacagatgcttctgaaagcattagagccctctttggaacagatggcataagaaatgcagcgcatggccctgattcttttgcttctgcg gccagagaaatggagttgtttttttccttcaagtggaggttgtgggccggcaaacactgctaaatttactaattgtacctgttgcattgttaaa ccccatgctgtcagtgaaggactgttgggaaagatcctgatggctatccgagatgcaggttttgaaatctcagctatgcagatgttcaata tggatcgggttaatgttgaggaattctatgaagtttataaaggagtagtgaccgaatatcatgacatggtgacagaaatgtattctggccct tgtgtagcaatggagattcaacagaataatgctacaaagacatttcgagaattttgtggacctgctgatcctgaaattgcccggcatttac

gccctggaactctcagagcaatctttggtaaaactaagatccagaatgctgttcactgtactgatctgccagaggatggcctattagagg ttcaatacttcttcaagatcttggataattag (SEQ ID NO:148)

[00491] (amino acids)

[00492] MHDVKNHRTFLKRTKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTAR QLGSRKEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQSRP FFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIRNA AHGPDSFASAAREMELFFPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILMAIR DAGFEISAMQMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQNN ATKTFREFCGPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFKILD N (SEQ ID NO:149)

[00493] Human NME7-AB

[00494] (DNA)

[00495] atggaaaaaacgctagccctaattaaaccagatgcaatatcaaaggctggagaaataattgaaataataaacaaagct ggatttactataaccaaactcaaaatgatgatgctttcaaggaaagaagcattggattttcatgtagatcaccagtcaagacccttttcaat gagctgatccagtttattacaactggtcctattattgccatggagattttaagagatgatgctatatgtgaatggaaaagactgctgggacc tgcaaactctggagtggcacgcacagatgcttctgaaagcattagagccctctttggaacagatggcataagaaatgcagcgcatggc cctgattcttttgcttctgcggccagagaaatggagttgtttttttccttcaagtggaggttgtgggccggcaaacactgctaaatttactaatt gtacctgttgcattgttaaaccccatgctgtcagtgaaggactgttgggaaagatcctgatggctatccgagatgcaggttttgaaatctc agctatgcagatgttcaatatggatcgggttaatgttgaggaattctatgaagtttataaaggagtagtgaccgaatatcatgacatggtga cagaaatgtattctggcccttgtgtagcaatggagattcaacagaataatgctacaaagacatttcgagaattttgtggacctgctgatcct gaaattgcccggcatttacgccctggaactctcagagcaatctttggtaaaactaagatccagaatgctgttcactgtactgatctgccag aggatggcctattagaggttcaatacttcttcaagatcttggataattag (SEQ ID NO:150)

[00496] (amino acids)

[00497] MEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQS RPFFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIR NAAHGPDSFASAAREMELFFPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILM AIRDAGFEISAMQMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQ NNATKTFREFCGPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFK ILDN (SEQ ID NO:151)

[00498] Human NME7-X1

[00499] (DNA)

[00500] atgatgatgctttcaaggaaagaagcattggattttcatgtagatcaccagtcaagacccttttcaatgagctgatccag tttattacaactggtcctattattgccatggagattttaagagatgatgctatatgtgaatggaaaagactgctgggacctgcaaactctgg agtggcacgcacagatgcttctgaaagcattagagccctctttggaacagatggcataagaaatgcagcgcatggccctgattcttttgc

ttctgcggccagagaaatggagttgtttttccttcaagtggaggttgtgggccggcaaacactgctaaatttactaattgtacctgttgcat tgttaaaccccatgctgtcagtgaaggactgttgggaaagatcctgatggctatccgagatgcaggttttgaaatctcagctatgcagat gttcaatatggatcgggttaatgttgaggaattctatgaagtttataaaggagtagtgaccgaatatcatgacatggtgacagaaatgtatt ctggcccttgtgtagcaatggagattcaacagaataatgctacaaagacatttcgagaattttgtggacctgctgatcctgaaattgcccg gcatttacgccctggaactctcagagcaatctttggtaaaactaagatccagaatgctgttcactgtactgatctgccagaggatggccta ttagaggttcaatacttcttcaagatcttggataattag (SEQ ID NO:152)

[00501] (amino acids)

[00502] MMMLSRKEALDFHVDHQSRPFFNELIQFITTGPIIAMEILRDDAICEWKRL LGPANSGVARTDASESIRALFGTDGIRNAAHGPDSFASAAREMELFFPSSGGCGPANT AKFTNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAMQMFNMDRVNVEEFYEVYK GVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFCGPADPEIARHLRPGTLRAIFG KTKIQNAVHCTDLPEDGLLEVQYFFKILDN* (SEQ ID NO:153)

[00503] Human NME7 a (optimized for E coli expression)

[00504] (DNA)

[00505] atgaatcactccgaacgctttgtttttatcgccgaatggtatgacccgaatgcttccctgctgcgccgctacgaactgct gttttatccgggcgatggtagcgtggaaatgcatgacgttaaaaatcaccgtacctttctgaaacgcacgaaatatgataatctgcatctg gaagacctgtttattggcaacaaagtcaatgtgttctctcgtcagctggtgctgatcgattatggcgaccagtacaccgcgcgtcaactg ggtagtcgcaaagaaaaaacgctggccctgattaaaccggatgcaatctccaaagctggcgaaattatcgaaattatcaacaaagcgg gtttcaccatcacgaaactgaaaatgatgatgctgagccgtaaagaagccctggattttcatgtcgaccaccagtctcgcccgtttttcaa tgaactgattcaattcatcaccacgggtccgattatcgcaatggaaattctgcgtgatgacgctatctgcgaatggaaacgcctgctggg cccggcaaactcaggtgttgcgcgtaccgatgccagtgaatccattcgcgctctgtttggcaccgatggtatccgtaatgcagcacatg gtccggactcattcgcatcggcagctcgtgaaatggaactgtttttcccgagctctggcggttgcggtccggcaaacaccgccaaatttt accaattgtacgtgctgtattgtcaaaccgcacgcagtgtcagaaggcctgctgggtaaaattctgatggcaatccgtgatgctggcttt gaaatctcggccatgcagatgttcaacatggaccgcgttaacgtcgaagaattctacgaagtttacaaaggcgtggttaccgaatatca cgatatggttacggaaatgtactccggtccgtgcgtcgcgatggaaattcagcaaaacaatgccaccaaaacgtttcgtgaattctgtgg tccggcagatccggaaatcgcacgtcatctgcgtccgggtaccctgcgcgcaatttttggtaaaacgaaaatccagaacgctgtgcact gtaccgatctgccggaagacggtctgctggaagttcaatacttttttcaaaattctggataat (SEQ ID NO:154)

[00506] (amino acids)

[00507] MNHSERFVFIAEWYDPNASLLRRYELLFYPGDGSVEMHDVKNHRTFLKR TKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTARQLGSRKEKTLALIKPDAISKA GEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQSRPFFNELIQFITTGPIIAMEILRDD AICEWKRLLGPANSGVARTDASESIRALFGTDGIRNAAHGPDSFASAAREMELFFPSS GGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAMQMFNMDRVNV

EEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFCGPADPEIARHLR PGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFKILDNTG (SEQ ID NO:155)

[00508] Human NME7 b (optimized for E coli expression)

[00509] (DNA)

[00510] atgcatgacgttaaaaatcaccgtacctttctgaaacgcacgaaatatgataatctgcatctggaagacctgtttattggc aacaaagtcaatgtgttctctcgtcagctggtgctgatcgattatggcgaccagtacaccgcgcgtcaactgggtagtcgcaaagaaaa aacgctggccctgattaaaccggatgcaatctccaaagctggcgaaattatcgaaattatcaacaaagcgggtttcaccatcacgaaac tgaaaatgatgatgctgagccgtaaagaagccctggattttcatgtcgaccaccagtctcgcccgtttttcaatgaactgattcaattcatc accacgggtccgattatcgcaatggaaattctgcgtgatgacgctatctgcgaatggaaacgcctgctgggcccggcaaactcaggtg ttgcgcgtaccgatgccagtgaatccattcgcgctctgtttggcaccgatggtatccgtaatgcagcacatggtccggactcattcgcat cggcagctcgtgaaatggaactgtttttcccgagctctggcggttgcggtccggcaaacaccgccaaatttaccaattgtacgtgctgta ttgtcaaaccgcacgcagtgtcagaaggcctgctgggtaaaattctgatggcaatccgtgatgctggctttgaaatctcggccatgcag atgttcaacatggaccgcgttaacgtcgaagaattctacgaagtttacaaaggcgtggttaccgaatatcacgatatggttacggaaatg tactccggtccgtgcgtcgcgatggaaattcagcaaaacaatgccaccaaaacgtttcgtgaattctgtggtccggcagatccggaaat cgcacgtcatctgcgtccgggtaccctgcgcgcaatttttggtaaaacgaaaatccagaacgctgtgcactgtaccgatctgccggaa gacggtctgctggaagttcaatacttttttcaaaattctggataat (SEQ ID NO:156)

[00511] (amino acids)

[00512] MHDVKNHRTFLKRTKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTAR QLGSRKEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQSRP FFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIRNA AHGPDSFASAAREMELFFPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILMAIR DAGFEISAMQMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQNN ATKTFREFCGPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFKILD NTG (SEQ ID NO:157)

[00513] Human NME7-AB (optimized for E coli expression)

[00514] (DNA)

[00515] atggaaaaaacgctggccctgattaaaccggatgcaatctccaaagctggcgaaattatcgaaattatcaacaaagcg ggtttcaccatcacgaaactgaaaatgatgatgctgagccgtaaagaagccctggattttcatgtcgaccaccagtctcgcccgtttttca atgaactgattcaattcatcaccacgggtccgattatcgcaatggaaattctgcgtgatgacgctatctgcgaatggaaacgcctgctgg gcccggcaaactcaggtgttgcgcgtaccgatgccagtgaatccattcgcgctctgtttggcaccgatggtatccgtaatgcagcacat ggtccggactcattcgcatcggcagctcgtgaaatggaactgtttttcccgagctctggcggttgcggtccggcaaacaccgccaaatt taccaattgtacgtgctgtattgtcaaaccgcacgcagtgtcagaaggcctgctgggtaaaattctgatggcaatccgtgatgctggcttt gaaatctcggccatgcagatgttcaacatggaccgcgttaacgtcgaagaattctacgaagtttacaaaggcgtggttaccgaatatca cgatatggttacggaaatgtactccggtccgtgcgtcgcgatggaaattcagcaaaacaatgccaccaaaacgtttcgtgaattctgtgg

tccggcagatccggaaatcgcacgtcatctgcgtccgggtaccctgcgcgcaatttttggtaaaacgaaaatccagaacgctgtgcact gtaccgatctgccggaagacggtctgctggaagttcaatacttttttcaaaattctggataat (SEQ ID NO:158)

[00516] (amino acids)

[00517] MEKTLALIKPDAISKAGEIIEIINKAGFTITKLKMMMLSRKEALDFHVDHQS RPFFNELIQFITTGPIIAMEILRDDAICEWKRLLGPANSGVARTDASESIRALFGTDGIR NAAHGPDSFASAAREMELFFPSSGGCGPANTAKFTNCTCCIVKPHAVSEGLLGKILM AIRDAGFEISAMQMFNMDRVNVEEFYEVYKGVVTEYHDMVTEMYSGPCVAMEIQQ NNATKTFREFCGPADPEIARHLRPGTLRAIFGKTKIQNAVHCTDLPEDGLLEVQYFFK ILDNTG (SEQ ID NO:159)

[00518] Human NME7-X1 (optimized for E coli expression)

[00519] (DNA)

[00520] atgatgatgctgagccgtaaagaagccctggattttcatgtcgaccaccagtctcgcccgtttttcaatgaactgattcaa ttcatcaccacgggtccgattatcgcaatggaaattctgcgtgatgacgctatctgcgaatggaaacgcctgctgggcccggcaaactc aggtgttgcgcgtaccgatgccagtgaatccattcgcgctctgtttggcaccgatggtatccgtaatgcagcacatggtccggactcatt cgcatcggcagctcgtgaaatggaactgtttttcccgagctctggcggttgcggtccggcaaacaccgccaaatttaccaattgtacgt gctgtattgtcaaaccgcacgcagtgtcagaaggcctgctgggtaaaattctgatggcaatccgtgatgctggctttgaaatctcggcca tgcagatgttcaacatggaccgcgttaacgtcgaagaattctacgaagtttacaaaggcgtggttaccgaatatcacgatatggttacgg aaatgtactccggtccgtgcgtcgcgatggaaattcagcaaaacaatgccaccaaaacgtttcgtgaattctgtggtccggcagatccg gaaatcgcacgtcatctgcgtccgggtaccctgcgcgcaatttttggtaaaacgaaaatccagaacgctgtgcactgtaccgatctgcc ggaagacggtctgctggaagttcaatacttttttcaaaattctggataat (SEQ ID NO:160)

[00521] (amino acids)

[00522] MMMLSRKEALDFHVDHQSRPFFNELIQFITTGPIIAMEILRDDAICEWKRL LGPANSGVARTDASESIRALFGTDGIRNAAHGPDSFASAAREMELFFPSSGGCGPANT AKFTNCTCCIVKPHAVSEGLLGKILMAIRDAGFEISAMQMFNMDRVNVEEFYEVYK GVVTEYHDMVTEMYSGPCVAMEIQQNNATKTFREFCGPADPEIARHLRPGTLRAIFG KTKIQNAVHCTDLPEDGLLEVQYFFKILDNTG (SEQ ID NO:161)

[00523] DM10 domain of NME7

[00524] (amino acids)

[00525] MNHSERFVFIAEWYDPNASLLRRYELLFYPGDGSVEMHDVKNHRTFLKR TKYDNLHLEDLFIGNKVNVFSRQLVLIDYGDQYTARQLGSRK (SEQ ID NO:162)

EXAMPLES

[00526] **Example 1 - Components of minimal serum-free base ("MM") (500mls)**

[00527] 400 ml DME/F12/GlutaMAX I (Invitrogen# 10565-018)

[00528] 100 ml Knockout Serum Replacement (KO-SR, Invitrogen# 10828-028)

[00529] 5 ml 100x MEM Non-essential Amino Acid Solution (Invitrogen# 11140-050)

[00530] 0.9 ml (0.1mM) β-mercaptoethanol (55mM stock, Invitrogen# 21985-023)

[00531] Example 2 - Probing cancer and stem cells for the presence of NME1, NME6 and NME7

[00532] In this series of experiments, we probed the expression of NME6 and NME7 in stem cells and cancer cells. In addition, we identified MUC1* as the target of NME7. We first performed Western blot assays on cell lysates to determine the presence or absence of NME1, NME6 and NME7. In Figure 1A, lysates from BGO1v human embryonic stem cells that had been cultured in NME1 dimers over a surface coated with anti-MUC1* antibodies (Lane 1), or cultured in bFGF over MEFs (Lane 2) or T47D human breast cancer cell lysates (Lane 3) or NME1-wt as a positive control, were separated by SDS-PAGE then probed with an anti-NME1 specific antibody. The results show that NME1 is strongly expressed in human ES cells whether cultured in NME1 dimers or bFGF, and in T47D cancer cells. The same cell lysates are separated by SDS-PAGE and then probed with an anti-NME6 specific antibody (anti-NME6 from Abnova). No NME6 was detected (data not shown), however it was detected later in a more concentrated sample (see Figure 2).

[00533] In Figure 1B, the same cell lysates are separated by SDS-PAGE and then probed with an anti-NME7 specific antibody (nm23-H7 B9 from Santa Cruz Biotechnology, Inc). The results show that NME7 is strongly expressed in human ES cells cultured in NME1 dimers over an anti-MUC1* antibody surface (Lane 1), weakly expressed in the same ES cells that were cultured in bFGF over MEFs (Lane 2), and strongly expressed in breast cancer cells (Lane 3). Lane 4 in which NME1 was added is blank indicating that the NME7 antibody does not cross react with NME1. The fact that NME7 is expressed to a greater degree in stem cells cultured in NME1 dimers, which we have shown express markers indicating that they are in a more naïve state than cells cultured in bFGF, means that NME7 is expressed at a higher level in naïve cells, compared to its expression in primed cells.

[00534] To determine whether NME7 also functions as a growth factor with MUC1* as its target receptor, we performed pull-down assays. In these experiments, a synthetic MUC1* extra cellular domain peptide (His-tagged PSMGFR sequence) was immobilized on NTA-Ni magnetic beads. These beads were incubated with the cell lysates of BGO1v human embryonic stem cells that had been cultured in NME1 dimers over a surface coated with anti-MUC1* antibodies (Lane 1), or cultured in bFGF over MEFs (Lane 2) or T47D human breast

cancer cell lysates (Lane 3). Beads were rinsed and captured proteins were released by addition of imidazole. Proteins were separated by SDS-PAGE and then probed with either an anti-NME1 antibody (Figure 1C) or an NME7 antibody (Figure 1D). The results show that NME7 binds to the MUC1* extra cellular domain peptide. This means that in stem cells and cancer cells, NME7 via its portions of its two NDPK domains, activates pluripotency pathways by dimerizing the MUC1* extra cellular domain.

[00535] Example 3 - A MUC1 pull down assay shows that NME1, NME6 and NME7 bind to a MUC1 species protein.

[00536] A pull down assay using an antibody to the MUC1* cytoplasmic tail (Ab-5) was performed on a panel of cells. Results are shown in Figures 2A-2F. The proteins pulled down by the MUC1 antibody were separated by SDS-PAGE then probed with antibodies specific for NME1, NME6 and NME7, using Western blot technique. MUC1*-positive breast cancer cell line T47D cells (ATCC), human embryonic stem cell line BGO1v (LifeTechnologies), human ES cells (HES-3, BioTime Inc.), human iPS cells (SC101A-1, System Biosciences Inc.) and T47D cancer cells were grown according to ATCC protocol in RPMI-1640 (ATCC) plus 10% FBS (VWR). All stem cells were cultured in minimal stem cell media "MM" with 8nM NM23-RS (recombinant NME1 S120G dimers). Stem cells were grown on plasticware coated with 12.5 ug/mL anti-MUC1* C3 mab. Cells were lysed with 200uL RIPA buffer for 10 min on ice. After removal of cell debris by centrifugation, the supernatant was used in a co-immunoprecipitation assay. MUC1* was pulled down using the Ab-5 antibody (anti-MUC-1 Ab-5, Thermo Scientific), which recognizes the MUC1 cytoplasmic tail, coupled to Dynabeads protein G (Life Technologies). The beads were washed twice with RIPA buffer and resuspended in reducing buffer. A sample of the supernatant was subjected to a reducing SDS-PAGE followed by transfer of the protein to a PVDF membrane. In Figure 2, the membrane was then probed with: A) an anti-NM23-H1 (NME1) Antibody (C-20, Santa Cruz Biotechnology); B) anti-NME6 (Abnova); or C) anti-NM23-H7 Antibody (B-9, Santa Cruz Biotechnology); D) the staining of NME6 was enhanced using Supersignal (Pierce); and E) the staining of NME7 was enhanced using Supersignal.

[00537] After incubation with their respective secondary antibody coupled to HRP, the proteins were detected by chemiluminescence. The photos show that native NME1, NME6 and NME7 are present in MUC1*-positive breast cancer cells, in human ES cells and in human iPS cells and that they bind to MUC1*. Note that the number of cells present in the HES-3 pellet was less than the number present in the other samples.

[00538] Example 4 - Detection of NME7 in embryonic stem cells and iPS cells

[00539] Results are shown in Figure 3. Human ES cells (BGO1v and HES-3) were cultured in NME-based media wherein cells were plated over a layer of anti-MUC1* antibody. To identify NME7 species, cells were harvested and lysed with RIPA buffer (Pierce), supplemented with protease inhibitor (Pierce). Cell lysates (20 uL) were separated by electrophoresis on a 12% SDS-PAGE reducing gel and transferred to a PVDF membrane (GE Healthcare). The blot was blocked with PBS-T containing 3% milk and then incubated with primary antibody (anti NM23-H7 clone B-9, Santa Cruz Biotechnology) at 4°C overnight. After washing with PBS-T, the membrane was incubated with horseradish peroxidase (HRP)-conjugated secondary antibody (goat anti mouse, Pierce) for 1 hr at room temperature. Signals were detected with Immun-Star Chemiluminescence kit (Bio-Rad). Figure 3A shows that the lysates from human stem cells contain three NME7 species: full-length at 42kDa and two lower molecular weight NME7 species at ~33kDa and~30kDa. Figures 3B and C shows the difference between NME7 species that are secreted (B), and those that are retained within the cell (C). Figure 3B shows that only the 30-33kDa NME7 species are secreted from the cells. Figure 3C shows that the lysates of those same cells have both the full-length form and a lower molecular weight species that may be a cleavage product or alternate isoform of both. For part (B), iPS Conditioned media (20 uL) was separated by electrophoresis on either a 12% SDS-PAGE reducing gel and transferred to a PVDF membrane (GE Healthcare). The blot was blocked with PBS-T containing 3% milk and then incubated with primary antibody (anti NM23-H7 clone B-9, Santa Cruz Biotechnology) at 4°C overnight. After washing with PBS-T, the membrane was incubated with horseradish peroxidase (HRP)-conjugated secondary antibody (goat anti mouse, Pierce) for 1 hr at room temperature. Signals were detected with Immun-Star Chemiluminescence kit (Bio-Rad). For part (C) experiment was similarly performed except that the cell lysate was used instead of the conditioned media.

[00540] Example 5 - Generation of Protein Constructs

[00541] For generating recombinant NME7, first, constructs were made to make a recombinant NME7 that could be expressed efficiently and in soluble form. The first approach was to make a construct that would encode the native NME7 (a) or an alternative splice variant NME7 (b), which has an N-terminal deletion. In some cases, the constructs carried a histidine tag or a strep tag to aid in purification. NME7-a, full-length NME7 expressed poorly in *E. coli* and NME7-b did not express at all in *E. coli*. However, a novel

construct was made in which the DM10 sequence was deleted and the NME7 comprised essentially the NDPK A and B domains having a calculated molecular weight of 33kDa.

[00542] This novel NME7-AB expressed very well in *E. coli* and existed as the soluble protein. NME7-AB was first purified over an NTA-Ni column and then further purified by size exclusion chromatography (FPLC) over a Sephadex 200 column (Figure 4A). Fractions were collected and tested by SDS-PAGE to identify fractions with the highest and purest expression of NME7-AB (Figure 4B). Figure 4C shows the FPLC trace for the combined fractions that were the most pure. The purified NME7-AB protein was then tested and shown to fully support the growth of human stem cells and further reverts them to the most naïve, pre-X-inactivation state. The purified NME7-AB was also shown to accelerate the growth of cancer cells.

[00543] Example 6 - ELISA assay showing NME7-AB simultaneously binds to two MUC1* extra cellular domain peptides

[00544] Results are shown in Figure 5. The PSMGFR peptide bearing a C-terminal Cysteine (PSMGFR-Cys) was covalently coupled to BSA using Imject Maleimide activated BSA kit (Thermo Fisher). PSMGFR-Cys coupled BSA was diluted to 10ug/mL in 0.1M carbonate/bicarbonate buffer pH 9.6 and 50uL was added to each well of a 96 well plate. After overnight incubation at 4°C, the plate was washed twice with PBS-T and a 3% BSA solution was added to block remaining binding site on the well. After 1h at RT the plate was washed twice with PBS-T and NME7, diluted in PBS-T + 1% BSA, was added at different concentrations. After 1h at RT the plate was washed 3x with PBS-T and anti-NM23-H7 (B-9, Santa Cruz Biotechnology), diluted in PBS-T + 1% BSA, was added at 1/500 dilution. After 1h at RT the plate was washed 3x with PBS-T and goat anti mouse-HRP, diluted in PBS-T + 1% BSA, was added at 1/3333 dilution. After 1h at RT the plate was washed 3x with PBS-T and binding of NME7 was measured at 415nm using ABTS solution (Pierce).

[00545] ELISA MUC1* dimerization: The protocol for NME7 binding was used, and NME7 was used at 11.6ug/mL.

[00546] After 1h at RT the plate was washed 3x with PBS-T and HisTagged PSMGFR peptide (PSMGFR-His) or biotinylated PSMGFR peptide (PSMGFR-biotin), diluted in PBS-T + 1% BSA, was added at different concentration. After 1h at RT the plate was washed 3x with PBS-T and anti-Histag-HRP (Abcam) or streptavidin-HRP (Pierce), diluted in PBS-T + 1% BSA, was added at a concentration of 1/5000. After 1h at RT the plate was washed 3x with PBS-T and binding of PSMGFR peptide to NME7 already bound to another PSMGFR

peptide (which could not signal by anti-His antibody or by streptavidin) coupled BSA was measured at 415nm using a ABTS solution (Pierce).

[00547] Example 7 - Functional testing of human recombinant NME7-AB

[00548] For testing recombinant NME7-AB for ability to maintain pluripotency and inhibit differentiation, a soluble variant of NME7, NME7-AB, was generated and purified. Human stem cells (iPS cat# SC101a-1, System Biosciences) were grown per the manufacturer's directions in 4ng/ml bFGF over a layer of mouse fibroblast feeder cells for four passages. These source stem cells were then plated into 6-well cell culture plates (Vita™, Thermo Fisher) that had been coated with 12.5 ug/well of a monoclonal anti-MUC1* antibody, MN-C3. Cells were plated at a density of 300,000 cells per well. The base media was Minimal Stem Cell Media consisting of: 400 ml DME/F12/GlutaMAX I (Invitrogen# 10565-018), 100 ml Knockout Serum Replacement (KO-SR, Invitrogen# 10828-028), 5 ml 100x MEM Non-essential Amino Acid Solution (Invitrogen# 11140-050) and 0.9 ml (0.1mM) β-mercaptoethanol (55mM stock, Invitrogen# 21985-023). The base media can be any media. In a preferred embodiment, the base media is free of other growth factors and cytokines. To the base media was added either 8nM of NME7-AB or 8nM NM23-H1 refolded and purified as stable dimers. Media was changed every 48 hours and due to accelerated growth, had to be harvested and passaged at Day 3 post-plating. Comparable pluripotent stem cell growth was achieved when stem cells were grown in NM23-H1 dimers or in NME7 monomers.

[00549] NME7 and NM23-H1 (NME1) dimers both grew pluripotently and had no differentiation even when 100% confluent. As can be seen in the photos, NME7 cells grew faster than the cells grown in NM23-H1 dimers. Cell counts at the first harvest verified that culture in NME7 produced 1.4-times more cells than culture in NM23-H1 dimers. ICC staining for the typical pluripotent markers confirmed that NME7-AB fully supported human stem cell growth, pluripotency, and resisted differentiation.

[00550] The NME7 species of ~30-33kDa may be an alternative splice isoform or a post translational modification such as cleavage, which may enable secretion from the cell.

[00551] Example 8 - Inducing transition of cancer cells to metastatic cancer cells by culturing cells under conditions that revert stem cells to a more naïve state

[00552] Cancer cells are normally cultured in a serum-containing media such as RPMI. We discovered that culturing cancer cells in the presence of reagents that make stem cells revert to a more naïve state, makes the cancer cells transform to a more metastatic state.

[00553] We demonstrated that NME7-AB, human NME1 dimers, bacterial NME1 dimers, NME7-X1 and "2i" inhibitors were each able to transform regular cancer cells into metastatic

cancer cells, which are also called cancer stem cells "CSCs" or tumor initiating cells "TICs". 2i is the name given to two biochemical inhibitors that researchers found made human stem cells revert to a more naïve state. 2i are MEK and GSK3-beta inhibitors PD0325901 and CHIR99021, which are added to culture medium to final concentrations of about 1mM and 3mM, respectively.

[00554] NME7-AB and NME7-X1 are at a final concentration of about 4nM when added to separate batches of minimal medium to make cancer cells transform to metastatic cells, although lower and higher concentrations also work well in the range of about 1nM to 16nM. Human or bacterial NME1 dimers are used at a final concentration of 4nM to 32nM, with 16nM typically used in these experiments, wherein the human NME bears the S120G mutation. Lower concentrations may be required if using wild type. It is not intended that these exact concentrations are important. It is important that the NME1 proteins are dimers and the range of concentrations over which this happens is in the low nanomolar range although certain mutations allow higher concentrations to remain as dimers.

[00555] Similarly, the concentrations of NME7 proteins can vary. NME7-AB and NME7-X1 are monomers and concentrations used to transform cancer cells to metastatic cells should allow the proteins to remain as monomers. Various molecular markers have been proposed as being indicators of metastatic cancer cells. Different cancer types may have different molecules that are up-regulated. For example, the receptor CXCR4 is up-regulated in metastatic breast cancers while E-cadherin, also known as CHD1, is up-regulated more in metastatic prostate cancers.

[00556] In addition to these specific metastasis markers, typical markers of pluripotency such as OCT4, SOX2, NANOG, and KLF4 are up-regulated as cancers become metastatic. The starting cancer cells and the later metastatic cancer cells can be assayed by PCR to measure expression levels of these genes.

[00557] Figure 11 shows a graph of RT-PCR measurements of T47D breast cancer cells that were cultured in a media that contained NME7-AB. A rho I kinase inhibitor, ROCi, ROCKi or Ri, was added to prevent the transformed cells from floating off the plate. Expression levels of various metastatic markers as well as pluripotent stem cell markers were measured for the parent cells and for the NME7-AB cultured cells. The results show that the floater cells express higher amounts of metastatic and pluripotency markers compared to the cells that received ROCi. We reasoned it was because those measurements were the average of cells that did not transform and those that did but the ROCi made them remain adherent.

This can clearly be seen in Figure 12 wherein "-Ri" means adherent cells that did not receive ROCi and so were not mixed with the highly metastatic cells that float.

[00558] Prostate cancer cells also transitioned to a more metastatic state when cultured in media containing NM23, aka NME1, or NME7-AB. Here we show that for every cell line tested so far, culture in NME7-AB, human NME1 dimers, or bacterial NMEs that have high sequence homology to human, induces transition to a more metastatic state.

[00559] Figure 14A shows a graph of RT-PCR measurements of expression levels of metastatic and pluripotency markers for breast cancer cells that are cultured in media containing either 2i inhibitors, NME7-AB or both. As can be seen, 2i inhibitors are also able to induce the transition of cancer cells to a more metastatic state. Figure 14B shows a graph of RT-PCR measurements of expression levels of metastatic and pluripotency markers for breast cancer cells that were cultured in media containing an NME1 from bacteria HSP593, whose sequence is highly homologous to human NME1 and NME7-AB, showing that bacterial NMEs with high sequence homology can mimic the effect of human NME1 and NME7-AB in that they induce transition to a more metastatic state. Ovarian cancer cell lines SK-OV3, OV-90, pancreatic cancer cell lines CAPAN-2 and PANC-1, breast cancer cell line MDA-MB all displayed the morphological transition of going from adherent to non-adherent when cultured in NME7-AB and or 2i inhibitors.

[00560] Figure 37 shows graphs of RT-PCR measurement of metastatic or pluripotency markers for various cancer cell lines cultured for 72 or 144 hours in NME7-AB. Figure 37A shows that SK-OV3 cells increase expression of metastatic markers CHD1, SOX2 and NME7-X1 when cultured in NME7-AB. Figure 37B shows that OV-90 cells increase expression of metastatic markers CXCR4 and NME7-X1 after culture in NME7-AB.

[00561] Example 9 - Demonstration that cancer cells cultured in NME7 become metastatic

[00562] A functional test of whether or not a population of cancer cells is metastatic is to implant very low numbers, e.g. 200, of the cells in immuno-compromised mice and see if they develop into a tumor. Typically 5-6 million cancer cells are required to form a tumor in an immuno-compromised mouse. We showed that as few as 50 of the NME-induced metastatic cancer cells formed tumors in mice. In addition, mice that were injected throughout the test period with human NME7-AB, NME1, or NME7-X1 developed remote metastases.

[00563] T47D human breast cancer cells were cultured in standard RPMI media for 14 days with media changes every 48 hours and passed by trypsinization when approximately

75% confluent. The cells were then plated into 6-well plates and cultured in minimal stem cell media (see Example 1) that was supplemented with 4nM NME7-AB. Media was changed every 48 hours. By about Day 4, some cells become detached from the surface and float. Media is carefully changed so as to retain the "floaters" as these are the cells that have the highest metastatic potential as evidenced by RT-PCR measurement of metastatic markers. On Day 7 or 8, the floaters are harvested and counted. Samples are retained for RT-PCR measurement. The key marker measured is CXCR4 which is up-regulated by 40-200 times after being briefly cultured in NME7-AB.

[00564] The freshly harvested floater metastatic cells are xenografted into the flank of female nu/nu athymic mice that have been implanted with 90-day slow release estrogen pellets. Floater cells were xenografted as 10,000, 1,000, 100 or 50 cells each. Half of the mice in each group of 6 were also injected daily with 32nM NME7-AB near the original implantation site. The parent T47D cells that were cultured in RPMI media without NME7-AB were also implanted into mice as 6 million, 10,000 or 100 as controls. Mice implanted with the NME7-induced floater cells developed tumors even when as few as 50 cells were implanted. Mice that were implanted with the floater cells and that received daily injections of NME7-AB also developed remote tumors or remote metastases in various organs (Figure 20-25). 11 out of the 12 mice, or 92%, that were injected with human NME7-AB after implantation of the NME7-AB cultured cancer cells, developed tumors at the injection site. Only 7 out of the 12 mice, or 58%, that were not injected with human NME7-AB after implantation developed tumors. 9 out of the 11 mice, or 82%, that got tumors and were injected with human NME7-AB developed multiple tumors remote from the injection site. None of the mice that were not injected with NME7-AB developed multiple, visible tumors.

[00565] After sacrifice, RT-PCR and Western blots showed that the remote bumps on the mice injected with NME7-AB were indeed human breast tumors. Similar analysis of their organs showed that in addition to remote bumps, mice had randomly metastasized to the liver and lung with human breast cancer characteristic of the human breast cancer cells that were implanted. As expected, only the mice implanted with 6 million cells grew tumors.

[00566] Several experiments like the one described above were performed with essentially the same results. In each experiment, there were either 24 or 52 mice, including all proper controls.

[00567] Example 10 - Anti-NME7 antibodies inhibit cancer cell growth.

[00568] T47D breast cancer cells and DU145 prostate cancer cells were cultured according to recommended protocols by ATCC. Cells were grown to ~30% confluency. An anti-

NME7 polyclonal rabbit antibody was raised against a fragment of NME7 that encompasses nearly the entire protein: amino acids 100 to 376. This polyclonal antibody was added to the cancer cells at concentrations between 2.7 to 375ng/mL. Taxol was used as the positive control. Cells were photographed and counted at 48 hours (Figures 6 and 7) and after 96 hours (Figure 8). The photos and cell counts show that the antibody potently inhibited the growth of breast and prostate cancer cells. However, because there was no attempt to select immunizing peptides that were unique to NME7, this antibody could be exerting cytotoxic effects by binding to and inhibiting both NME7-AB-like species and NME1.

[00569] Example 11 - Peptides selected because their sequence is unique to NME7, A1, A2, B1, B2 and B3, inhibit the binding of NME7 species to MUC1* extracellular domain peptide.

[00570] NME7 peptides were selected as immunizing agents for antibody production. NME7 peptides A1, A2, B1, B2 and B3 (Figure 19) were chosen using a process of sequence alignment among human NME1, human NME7 and several bacterial NMEs that were homologous to human NME1 or human NME7. Five regions that had high sequence homology among all were identified. However, to prevent selecting peptides that would give rise to antibodies that would inhibit human NME1 as well as human NME7, we chose NME7 sequences that were adjacent to the homologous regions wherein those peptides had sequences that were different from human NME1. We did ELISA assays to see if the peptides on their own could bind to a synthetic MUC1* peptide on the surface and inhibit the binding of human NME7 or human NME1 to the immobilized peptide (Figure 27). Figure 27 shows that the peptides inhibited the binding of NME7 and NME1 to the immobilized peptide. This showed that those regions from which the peptides were derived were the regions that interacted with MUC1* and would give rise to antibodies that would bind to those regions of NME7 and inhibit its binding to MUC1* receptor.

[00571] In another experiment, the free peptides A1, A2, B1, B2 and B3 were added to cancer cells in culture that were undergoing transition to a more metastatic state by culturing in either NME7-AB or 2i. Figure 30 shows a table of scientist observations when cancer cells are grown in either NME7-AB or 2i inhibitors, and shows that the free peptides inhibited the morphological change from adherent cells to floaters, which for breast cancer cells is directly correlated to increased expression of metastatic markers, especially CXCR4. RT-PCR measurements confirm that the NME7-AB peptides inhibited the increase in expression of metastasis marker CXCR4.

[00572] Figure 31 shows a graph of RT-PCR measurements of CXCR4 expression in T47D breast cancer cells that were grown in either NME7-AB or 2i inhibitors, each of which transform cancer cells to a more metastatic state, and the inhibitory effect of NME7-derived peptides, A1, A2, B1, B2 and B3, on the metastatic transformation. Figure 32 shows a table of recorded RNA levels in samples that were used for RT-PCR measurement of CXCR4 in Figure 31 as well as the threshold cycle number for CXCR4 expression as well as for the control housekeeping gene.

[00573] Example 12 - Anti-NME7 antibodies specifically bind to human NME7 but not to human NME1

[00574] A standard ELISA assay was performed to determine whether or not the NME7 antibodies we generated by immunization with NME7-AB peptides A1, A2, B1, B2, and B3 would bind specifically to NME7-AB, but not to human NME1 as it has healthy functions and it may be detrimental to a human to block it with an antibody. The ELISA of Figure 26 shows that all of the NME7 antibodies we generated from peptides A1, A2, B1, B2, and B3 bind to human NME7-AB (A) but not to human NME1 (B). The peptides used to generate these antibodies are common to both NME7-AB and NME7-X1. This assays show that the antibodies generated from peptides A1, A2, B1, B2, and B3 specifically bind to NME7-AB and by extension will bind to NME7-X1.

[00575] **NME7A peptide 1** (A domain): MLSRKEALDFHVDHQS (SEQ ID NO:141)

[00576] **NME7A peptide 2** (A domain)**:** SGVARTDASES (SEQ ID NO:142)

[00577] **NME7B peptide 1** (B domain)**:** DAGFEISAMQMFNMDRVNVE (SEQ ID NO:143)

[00578] **NME7B peptide 2** (B domain)**:** EVYKGVVTEYHDMVTE (SEQ ID NO:144)

[00579] **NME7B peptide 3** (B domain)**:** AIFGKTKIQNAVHCTDLPEDGLLEVQYFF (SEQ ID NO:145)

[00580] Example 13 - Anti-NME7 specific antibodies and the peptides that generated them inhibit cancer cell growth

[00581] Rabbits were immunized with NME7 peptides A1, A2, B1, B2, and B3 and antibodies were generated, collected and purified over a column to which the immunizing peptide had been conjugated. T47D breast cancer cells were plated and cultured according to ATCC protocols in RPMI media supplemented with serum. Antibodies generated from immunization with peptides A1, A2, B1, B2, and B3 were added at the concentrations indicated in Figure 28. Immunizing peptides A1, A2, B1, B2, and B3, and the PSMGFR extracellular domain peptide of MUC1*, "FLR" here, were also added separately to growing

T47D breast cancer cells. Taxol and the E6 anti-MUC1* Fab were added as controls. The graph of Figure 28 shows that the antibodies generated, as well as the free peptides, potently inhibited the growth of the cancer cells. Note the comparison to inhibition using Taxol, which is a chemotherapy agent that kills healthy and cancer cells alike. Also, for comparison, a polyclonal antibody generated using a large stretch of NME7 from amino acid 100 to 376 is shown. Although this antibody is a potent inhibitor of cancer growth it could have non-specific effects since it can bind to NME1 as well as to NME7.

[00582] In a similar experiment, combinations of the antibodies generated from immunization with peptides A1, A2, B1, B2, and B3 as well as the peptides themselves were added to growing cancer cells at the concentrations indicated. The graphs of cell growth shown in Figure 29 show that the combinations of antibodies and peptides potently inhibited the growth of cancer cells. In these two experiments, the cells were MUC1* positive breast cancer cells.

[00583] Example 14 - Anti-NME7 antibodies inhibit the transition of cancer cells to metastatic cancer cells

[00584] Cancer cells transform to a more metastatic state when cultured in the presence of agents that revert stem cells to a more naïve state. We have demonstrated that culturing cancer cells in NME7-AB, human NME1 dimers, bacterial NME1 dimers or MEK and GSK3-beta inhibitors, called "2i", causes the cells to become more metastatic. As the cells transition to a more metastatic state, they become non-adherent and float off of the culture plate. These floating cells, "floaters" were collected separately from those that were adherent and were shown to: a) express much higher levels of metastatic genes; and b) when xenografted into mice, the floater cells were able to generate tumors when implanted at very low numbers. RT-PCR measurement of specific metastatic markers such as CXCR4 in breast cancers, CHD1 in prostate cancer, and other pluripotent stem cell markers such as OCT4, SOX2, NANOG, KLF4, c-Myc and others were dramatically over-expressed in cancer cells that were cultured in NME7-AB and most over-expressed in the cells that became non-adherent, called "floaters" here and in figures.

[00585] Here we show that the NME7-specific antibodies, generated by immunization with NME7-derived peptides A1, A2, B1, B2 and B3, as well as the peptides themselves, inhibit the transition from cancer cell to metastatic cancer cells. In the first of these experiments, the antibodies generated by immunization with A1, A2, B1, B2 and B3 were tested for their ability to inhibit the metastatic transition induced by culture of T47D breast cancer cells in NME7-AB or in 2i inhibitors. The most striking observation was that the

antibodies and the peptides dramatically reduced the number of floater cells, which was the first indication that the antibodies and peptides had inhibited the transformation to metastatic cancer cells. In particular, cells to which the antibody generated from immunization with the B3 peptide barely generated any floater cells.

[00586] Figure 30 shows the recorded observations of the percentage of floater cells visible for each antibody relative to the control wells that did not receive any antibody treatment. mRNA was extracted from both the floater cells and the adherent cells. RT-PCR was used to measure expression levels of metastatic markers, including CXCR4. Treatment with the anti-NME7 antibodies greatly reduced the amount of metastatic markers, such as CXCR4, indicating the antibodies inhibited the transition to metastatic cancer. (See Figure 31). Notably, the antibody generated by immunization with peptide B3, aka antibody #61, essentially completely inhibited the transition to a more metastatic state. Figure 31B shows that breast cancer cells that were treated with the NME7-AB peptides, A1, A2, B1, B2 and B3, alone were able to potently inhibit the transition to a more metastatic state induced by culturing the cells in a media containing the 2i inhibitors. Peptide B3 was especially effective as was antibody #61 that it generated. Figure 31C shows the same graph but with the Y-axis expanded to show the peptide inhibition of metastatic markers. The amount of mRNA, which indicates cell viability and growth, was measured. Cells that were treated with antibody had much less mRNA, indicating that in addition to inhibiting the transition to a more metastatic state, the anti-NME7-AB antibodies inhibited the growth of the cancer cells. Figure 32 shows a table of the amounts of RNA recovered for the inhibition experiment shown in Figure 31A.

[00587] Example 15 - Anti-NME7 antibodies generated with NME7-derived peptides A1, A2, B1, B2 and B3 identify novel NME7 species not detectable using any commercially available antibodies.

[00588] As is known to those skilled in the art, some antibodies recognize a linear portion of the target protein and can be used in Western blot assays while other antibodies recognize a non-linear conformational motif and can be used in pull-down or immunoprecipitation assays. Previous to this application, cleaved NME7 or isoform NME7-X1 was not known to exist. Using antibodies that were commercially available at the time of filing shows that existing antibodies could not specifically detect these important NME7 species. B9 (Santa Cruz Biotechnology) is a monoclonal antibody raised against NME7 amino acids 100-376. Figure 36A shows that it only detects full-length 42kDa NME7. Another commercially available antibody, H278, is a rabbit polyclonal raised against NME7 amino acids 100-376, which includes amino acid sequences that are not unique to NME7. Figure 36B shows that

this antibody also stains NME1, which is 17kDa as well as full-length NME7 and other bands that do not appear to be specific to NME7-AB.

[00589] NME7 antibodies generated by immunization with NME7-AB peptides A1, A2, B1, B2 or B3 identify new NME7 species including the full-length 42kDa protein, a ~33kDa NME7 species that may be a cleavage product or alternative isoform, a ~30kDa NME7 species that may be a cleavage product or alternative isoform, wherein the ~30kDa species appears to be NME7-X1. Figure 35A-C shows that antibodies generated by peptides A1, B1 and B3 identify the secreted forms of NME7, NME7-AB and NME7-X1 in a wide range of cancer cell lines, including T47D breast cancer cells, PC3 and DU145 prostate cancer cells, HEK293 fetal liver cells, and leukemia cells IM-9, K562, and MV411.

[00590] All of the references cited herein are incorporated by reference in their entirety.

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[00591] Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention specifically described herein. Such equivalents are intended to be encompassed in the scope of the claims.

What is claimed is:

1. A method of treating or preventing cancer in a subject, comprising administering to the subject an antibody made against a member of the NME family.
2. The method according to claim 1, wherein the NME family is NME7 family.
3. The method according to claim 2, wherein the antibody binds to NME7.
4. The method according to claim 2, wherein the antibody binds to NME7-AB or NME-AB-like protein.
5. The method according to claim 2, wherein the antibody binds to NME7-X1.
6. The method according to claim 1, wherein the antibody inhibits binding between NME7 and its cognate binding partner.
7. The method according to claim 6, wherein the cognate binding partner is MUC1*.
8. The method according to claim 6, wherein the cognate binding partner is PSMGFR portion of the MUC1* extracellular domain.
9. The method according to claim 1, wherein the antibody is generated or selected for its ability to bind to a peptide selected from those listed in Figures 16-19 (SEQ ID NOS:88 to 145).
10. The method according to claim 9, wherein the peptide is selected from those listed in Figure 19 (SEQ ID NOS:141 to 145).
11. The method according to claim 9, wherein the peptide comprises a peptide, which is highly homologous to, or to which is added or subtracted up to 7 amino acid residues at the N-terminus or C-terminus, of the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145).
12. The method according to claim 1, wherein the antibody is selected for its ability to bind to NME7-AB or NME7-X1 but not to NME1.
13. The method according to claim 1, wherein the antibody is polyclonal, monoclonal, bivalent, monovalent, bispecific, an antibody fragment containing the variable region, or an antibody mimic.
14. The method according to claim 1, wherein the antibody is human or humanized.
15. The method according to claim 1, wherein the antibody is a single chain scFv.
16. A method of treating or preventing cancer in a subject, comprising administering to the subject a peptide that is highly homologous or identical to regions of NME7-AB.
17. The method according to claim 16, wherein the peptide is at least 80% homologous to one or more of the peptides listed in Figure 16.

18. The method according to claim 16, wherein the peptide is at least 80% homologous to one or more of the peptides listed in Figure 17.

19. The method according to claim 16, wherein the peptide is at least 80% homologous to one or more of the peptides listed in Figure 18.

20. The method according to claim 16, wherein the peptide is at least 80% homologous to one or more of the peptides listed in Figure 19.

21. The method according to claim 16, wherein the peptide is selected from peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145).

22. The method according to claim 21, wherein the peptide is selected from those listed in Figure 19 (SEQ ID NOS:141 to 145).

23. The method according to claim 16, wherein the peptide comprises a peptide, which is highly homologous to, or to which is added or subtracted up to 7 amino acid residues at the N-terminus or C-terminus, of the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145).

24. The method according to claim 16, wherein the peptide is connected to another peptide via a spacer or linker.

25. A chimeric antigen receptor (CAR), for the treatment or prevention of cancer wherein the targeting extracellular portion of the CAR comprises at least a peptide fragment of a member of the NME family.

26. The chimeric antigen receptor according to claim 25, wherein the NME family is NME7 family.

27. The chimeric antigen receptor according to claim 26, wherein the member of the NME7 family is NME7.

28. The chimeric antigen receptor according to claim 26, wherein the member of the NME7 family is NME7-AB or NME-AB-like protein.

29. The chimeric antigen receptor according to claim 26, wherein the member of the NME7 family is NME7-X1.

30. The chimeric antigen receptor according to claim 25, wherein the targeting extracellular portion of the CAR comprises a peptide of the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145).

31. The chimeric antigen receptor according to claim 30, wherein the peptide is selected from those listed in Figure 19 (SEQ ID NOS:141 to 145).

32. The chimeric antigen receptor according to claim 25, wherein the peptide comprises a peptide, which is highly homologous to, or to which is added or subtracted up to 7 amino acid

residues at the N-terminus or C-terminus, of the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145).

33. The chimeric antigen receptor according to claim 25, wherein the peptide is connected to another peptide via a spacer or linker.

34. A method of treating or preventing cancer or cancer metastasis, comprising engineering the chimeric antigen receptor according to claim 25, into an immune system cell and administering the cell to a subject in need thereof.

35. A chimeric antigen receptor (CAR), for the treatment or prevention of cancer, wherein the targeting extracellular portion of the chimeric antigen receptor comprises a portion of an antibody that binds to NME7-AB, NME-AB-like protein or NME7-X1.

36. The chimeric antigen receptor according to claim 35, wherein the portion of the antibody is a single chain scFv.

37. The chimeric antigen receptor according to claim 35, wherein the antibody is human or humanized.

38. A method of vaccinating a person against cancer or metastatic cancer comprising immunizing the person with a peptide fragment of a member of the NME family.

39. The method according to claim 38, wherein the NME family is NME7 family.

40. The method according to claim 39, wherein the member of the NME7 family is NME7 or NME7b.

41. The method according to claim 39, wherein the wherein the member of the NME7 family is NME7-AB or NME7-AB-like protein.

42. The method according to claim 39, wherein the member of the NME7 family is NME7-X1.

43. The method according to claim 38, wherein the immunizing peptide is a peptide from the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145).

44. The method according to claim 43, wherein the peptide is selected from those listed in Figure 19 (SEQ ID NOS:141 to 145).

45. The method according to claim 38, wherein the immunizing peptide comprises a peptide, which is highly homologous to, or to which is added or subtracted up to 7 amino acid residues at the N-terminus or C-terminus, of the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145).

46. The method according to claim 38, wherein the immunizing peptide is connected to another peptide via a spacer or linker.

47. A method of treating or preventing cancer in a subject, comprising administering to the subject a nucleic acid that inhibits the expression of NME7, NME7b, NME7-AB-like protein or NME7-X1.

48. The method according to claim 47, wherein the nucleic acid is an anti-sense nucleic acid that suppresses expression of NME7, NME7-AB-like protein or NME7-X1.

49. The method according to claim 47, wherein the nucleic acid is an inhibitory RNA, siRNA, RNAi, or shRNA that inhibits expression of NME7, NME7-AB-like protein or NME7-X1.

50. A method of treating or preventing cancer in a subject, comprising administering to the subject genetically edited nucleic acids that inhibit expression of NME7, NME7b, NME7-AB-like protein or NME7-X1.

51. The method according to claim 50, wherein the genetically edited nucleic acids that inhibit expression of NME7, NME7b, NME7-AB-like protein or NME7-X1 are inserted into cells that are then administered to the patient.

52. The method according to claim 50, wherein the genetically edited nucleic acids that inhibit expression of NME7, NME7b, NME7-AB-like protein or NME7-X1 are inserted into cells using a viral vector.

53 The method according to claim 50, wherein the viral vector is a lentiviral system.

54. A method of growing cancer cells comprising contacting the cells with NME7-AB, NME7b, NME7-AB-like protein or NME7-X1, 2i or 5i.

55. The method according to claim 54, comprising culturing the cells in a medium that contains NME7-AB, NME7b, NME7-AB-like protein or NME7-X1, 2i or 5i.

56. The method according to claim 54, comprising growing cells in an animal that expresses human NME7-AB, NME7b, NME7-AB-like protein or NME7-X1, or to which NME7-AB, NME7b, NME7-AB-like protein or NME7-X1 is administered.

57. The method according to claim 54, wherein the cancer cells are breast, prostate, ovarian, colorectal, pancreatic, liver, melanoma or brain cancer cells.

58. The method according to claim 54, wherein drug candidates are tested on the cells.

59. The method according to claim 58, wherein the efficacy of the drugs is assessed by comparing cancer growth to a no drug control or comparing expression levels of metastatic markers or stem cell markers to a no drug control or comparing the ability of the resultant cells to form tumors in animals from low cell copy number compared to a no drug control and determining the efficacy of a candidate drug for the treatment of cancer or metastasis.

60. The method according to claim 54, wherein the cells are obtained from a patient being assessed for treatment for cancer and drugs that would be effective for that patient are selected based on results of claim above.

61. The method according to claim 54, wherein the cells are not obtained from a patient being assessed for treatment for cancer but drugs that would be effective for that patient are selected based on results of claim above.

62. A method of generating antibodies or antibody-like molecules from peptides or peptide mimics having a sequence derived from the sequence of NME.

63. The method according to claim 62, wherein the NME is NME7.

64. The method according to claim 62, wherein the peptide is used as an immunogen to generate antibodies or antibody-like molecules.

65. The method according to claim 63, wherein the peptide is administered to an animal to generate anti-NME7 antibodies.

66. The method according to claim 65, wherein the peptide is administered to a human to generate anti-NME7 antibodies.

67. The method according to claim 62, wherein the peptide has a sequence listed in Figures 16 to 19 (SEQ ID NOS:88 to 145).

68. The method according to claim 67, wherein the peptide is selected from those listed in Figure 19 (SEQ ID NOS:141 to 145).

69. The method according to claim 67, wherein the peptide comprises a peptide, which is highly homologous to, or to which is added or subtracted up to 7 amino acid residues at the N-terminus or C-terminus, of the peptides listed in Figures 16-19 (SEQ ID NOS:88 to 145).

70. A method of detecting presence of cancer or the progression of cancer, comprising the steps of:

1) obtaining a sample from a patient having cancer or at risk of developing a cancer;

2) subjecting that sample to an assay capable of detecting or measuring levels of a member of the NME7 family, or levels of nucleic acids encoding a member of the NME7 family;

3) comparing levels of the measured member of the NME7 family or the member of the NME7 family-encoding nucleic acids in the test sample to levels in control patients or control cells;

4) determining that the levels of the member of the NME7 family or nucleic acids encoding the member of the NME7 family are elevated compared to the controls; and

5) concluding that the donor of the test sample has cancer or has had a progression of cancer if the control to which the test was compared came from a donor previously diagnosed with a cancer.

71. The method according to claim 70, wherein detection of the member of the NME7 family in circulation or in a tissue is an indicator of cancer in a patient.

72. The method according to claim 70, wherein the member of the NME7 family is NME7, NME7b, NME7-X1, or NME7-AB-like protein.

73. A method comprising:

detecting presence of a member of the NME7 family or MUC1* in a patient; and

administering anti-NME7 or anti-MUC1* antibody or antibodies to the patient exhibiting the member of the NME7 family or MUC1* expression.

74. The method according to claim 73, wherein the member of the NME7 family is NME7, NME7b, NME7-X1, or NME7-AB-like protein.

75. A method for treating or preventing cancer comprising:

1) obtaining a sample from a patient suspected of having a cancer or at risk of developing a cancer or at risk of developing a metastatic cancer;

2) measuring an amount of the member of an NME7 family or a member of the NME7 family encoding nucleic acid, wherein the measured levels are significantly above those measured in a control sample;

3) determining that the patient has a cancer or has developed a more aggressive or a metastatic cancer;

4) administering to the patient an effective amount of a therapeutic agent that suppresses expression of the member of the NME7 family, inhibits cleavage of NME7 or inhibits NME7 binding to its targets.

76. The method according to claim 75, wherein the target of the member of the NME7 family is MUC1*.

77. The method according to claim 76, wherein the target of the member of the NME7 family is PSMGFR portion of the MUC1* extracellular domain.

78. The method according to claim 75, wherein the member of the NME7 family is NME7, NME7b, NME7-X1, or NME7-AB-like protein.