

**PROCESSES FOR PREPARING (1
-(3-FLUORO-2-(TRIFLUOROMETHYL)ISONICOTINYL)PIPERIDINE-4-ONE)**

TECHNICAL FIELD

This invention relates to processes and intermediates for making {1-{1-[3-
5 fluoro-2-(trifluoromethyl)isonicotinoyl]piperidin-4-yl}-3-[4-(7H-pyrrolo[2,3-
d]pyrimidin-4-yl)-1H-pyrazol-1-yl]azetidin-3-yl} acetonitrile, useful in the treatment
of diseases related to the activity of Janus kinases (JAK) including inflammatory
disorders, autoimmune disorders, cancer, and other diseases.

BACKGROUND

10 Protein kinases (PKs) regulate diverse biological processes including cell
growth, survival, differentiation, organ formation, morphogenesis, neovascularization,
tissue repair, and regeneration, among others. Protein kinases also play specialized
roles in a host of human diseases including cancer. Cytokines, low-molecular weight
polypeptides or glycoproteins, regulate many pathways involved in the host
15 inflammatory response to sepsis. Cytokines influence cell differentiation,
proliferation and activation, and can modulate both pro-inflammatory and anti-
inflammatory responses to allow the host to react appropriately to pathogens.
Signaling of a wide range of cytokines involves the Janus kinase family (JAKs) of
protein tyrosine kinases and Signal Transducers and Activators of Transcription
20 (STATs). There are four known mammalian JAKs: JAK1 (Janus kinase-1), JAK2,
JAK3 (also known as Janus kinase, leukocyte; JAKL; and L-JAK), and TYK2
(protein-tyrosine kinase 2).

Cytokine-stimulated immune and inflammatory responses contribute to
pathogenesis of diseases: pathologies such as severe combined immunodeficiency
25 (SCID) arise from suppression of the immune system, while a hyperactive or
inappropriate immune/inflammatory response contributes to the pathology of
autoimmune diseases (*e.g.*, asthma, systemic lupus erythematosus, thyroiditis,
myocarditis) and illnesses such as scleroderma and osteoarthritis (Ortmann, R. A., T.
Cheng, *et al.* (2000) *Arthritis Res* 2(1): 16-32).

Deficiencies in expression of JAKs are associated with many disease states. For example, Jak1^{-/-} mice are runted at birth, fail to nurse, and die perinatally (Rodig, S. J., M. A. Meraz, *et al.* (1998) *Cell* 93(3): 373-83). Jak2^{-/-} mouse embryos are anemic and die around day 12.5 postcoitum due to the absence of definitive erythropoiesis.

The JAK/STAT pathway, and in particular all four JAKs, are believed to play a role in the pathogenesis of asthmatic response, chronic obstructive pulmonary disease, bronchitis, and other related inflammatory diseases of the lower respiratory tract. Multiple cytokines that signal through JAKs have been linked to inflammatory diseases/conditions of the upper respiratory tract, such as those affecting the nose and sinuses (*e.g.*, rhinitis and sinusitis) whether classically allergic reactions or not. The JAK/STAT pathway has also been implicated in inflammatory diseases/conditions of the eye and chronic allergic responses.

Activation of JAK/STAT in cancers may occur by cytokine stimulation (*e.g.* IL-6 or GM-CSF) or by a reduction in the endogenous suppressors of JAK signaling such as SOCS (suppressor of cytokine signaling) or PIAS (protein inhibitor of activated STAT) (Boudny, V., and Kovarik, J., *Neoplasia*. 49:349-355, 2002).

Activation of STAT signaling, as well as other pathways downstream of JAKs (*e.g.*, Akt), has been correlated with poor prognosis in many cancer types (Bowman, T., *et al.* *Oncogene* 19:2474-2488, 2000). Elevated levels of circulating cytokines that signal through JAK/STAT play a causal role in cachexia and/or chronic fatigue. As such, JAK inhibition may be beneficial to cancer patients for reasons that extend beyond potential anti-tumor activity.

JAK2 tyrosine kinase can be beneficial for patients with myeloproliferative disorders, *e.g.*, polycythemia vera (PV), essential thrombocythemia (ET), myeloid metaplasia with myelofibrosis (MMM) (Levin, *et al.*, *Cancer Cell*, vol. 7, 2005: 387-397).

Inhibition of the JAK2V617F kinase decreases proliferation of hematopoietic cells, suggesting JAK2 as a potential target for pharmacologic inhibition in patients with PV, ET, and MMM.

Inhibition of the JAKs may benefit patients suffering from skin immune disorders such as psoriasis, and skin sensitization. The maintenance of psoriasis is believed to depend on a number of inflammatory cytokines in addition to various

chemokines and growth factors (JCI, 113:1664-1675), many of which signal through JAKs (*Adv Pharmacol.* 2000;47:113-74).

JAK1 plays a central role in a number of cytokine and growth factor signaling pathways that, when dysregulated, can result in or contribute to disease states. For example, IL-6 levels are elevated in rheumatoid arthritis, a disease in which it has been suggested to have detrimental effects (Fonesca, J.E. et al., *Autoimmunity Reviews*, 8:538-42, 2009). Because IL-6 signals, at least in part, through JAK1, antagonizing IL-6 directly or indirectly through JAK1 inhibition is expected to provide clinical benefit (Guschin, D., N., et al *Embo J* 14:1421, 1995; Smolen, J. S., et al. *Lancet* 371:987, 2008). Moreover, in some cancers JAK1 is mutated resulting in constitutive undesirable tumor cell growth and survival (Mullighan CG, *Proc Natl Acad Sci U S A*.106:9414-8, 2009; Flex E., et al. *J Exp Med*. 205:751-8, 2008). In other autoimmune diseases and cancers, elevated systemic levels of inflammatory cytokines that activate JAK1 may also contribute to the disease and/or associated symptoms. Therefore, patients with such diseases may benefit from JAK1 inhibition. Selective inhibitors of JAK1 may be efficacious while avoiding unnecessary and potentially undesirable effects of inhibiting other JAK kinases.

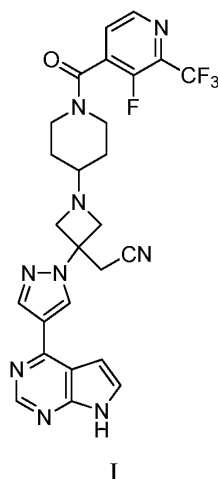
Selective inhibitors of JAK1, relative to other JAK kinases, may have multiple therapeutic advantages over less selective inhibitors. With respect to selectivity against JAK2, a number of important cytokines and growth factors signal through JAK2 including, for example, erythropoietin (Epo) and thrombopoietin (Tpo) (Parganas E, et al. *Cell*. 93:385-95, 1998). Epo is a key growth factor for red blood cells production; hence a paucity of Epo-dependent signaling can result in reduced numbers of red blood cells and anemia (Kaushansky K, *NEJM* 354:2034-45, 2006). Tpo, another example of a JAK2-dependent growth factor, plays a central role in controlling the proliferation and maturation of megakaryocytes – the cells from which platelets are produced (Kaushansky K, *NEJM* 354:2034-45, 2006). As such, reduced Tpo signaling would decrease megakaryocyte numbers (megakaryocytopenia) and lower circulating platelet counts (thrombocytopenia). This can result in undesirable and/or uncontrollable bleeding. Reduced inhibition of other JAKs, such as JAK3 and Tyk2, may also be desirable as humans lacking functional version of these kinases have been shown to suffer from numerous maladies such as severe-combined

immunodeficiency or hyperimmunoglobulin E syndrome (Minegishi, Y, et al. *Immunity* 25:745-55, 2006; Macchi P, et al. *Nature*. 377:65-8, 1995). Therefore a JAK1 inhibitor with reduced affinity for other JAKs would have significant advantages over a less-selective inhibitor with respect to reduced side effects involving immune suppression, anemia and thrombocytopenia.

Due to the usefulness of JAK inhibitors, there is a need for development of new processes for making JAK inhibitors. This invention is directed towards this need and others.

SUMMARY

JAK inhibitors are described in US 2011/0224190, which is incorporated herein by reference in its entirety, including {1-{1-[3-fluoro-2-(trifluoromethyl)isonicotinoyl]piperidin-4-yl}-3-[4-(7H-pyrrolo[2,3-d]pyrimidin-4-yl)-1H-pyrazol-1-yl]azetidin-3-yl}acetonitrile, which is depicted below as Formula I.



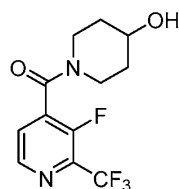
The present invention provides, *inter alia*, processes and intermediates for making the compound of Formula I.

In particular, the present invention provides a process comprising reacting a compound of Formula III:



III

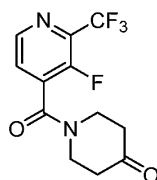
or a salt thereof, with 4-hydroxypiperidine to form a compound of Formula IV:



IV

5 or a salt thereof.

The processes described herein can further comprise reacting the compound of Formula IV, or a salt thereof, under oxidation conditions to form a compound of Formula V:



V

10

or a salt thereof.

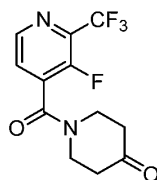
The present invention also provides a process comprising reacting a compound of Formula III:



III

15

or a salt thereof, with 4-piperidone, or a salt thereof, to form a compound of Formula V:

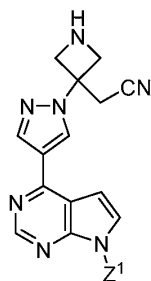


V

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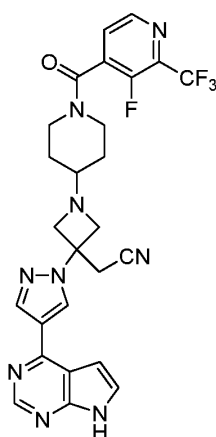
or a salt thereof.

In some embodiments, the processes described herein further comprise reacting the compound of Formula V with a compound of Formula VI:



VI

5 or a salt thereof, in the presence of a reducing agent, to form a compound of Formula I;

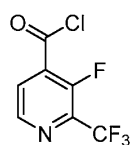


I

or a salt thereof;

wherein Z^1 is H or a protecting group.

The present invention also provides a compound of Formula III:



III

or a salt thereof.

DETAILED DESCRIPTION

It is appreciated that certain features of the invention, which are, for clarity, described in the context of separate embodiments, can also be provided in combination in a single embodiment. Conversely, various features of the invention which are, for brevity, described in the context of a single embodiment, can also be provided separately or in any suitable subcombination.

The processes described herein can be monitored according to any suitable method known in the art. For example, product formation can be monitored by spectroscopic means, such as nuclear magnetic resonance spectroscopy (e.g., ^1H or ^{13}C), infrared spectroscopy, or spectrophotometry (e.g., UV-visible); or by chromatography such as high performance liquid chromatography (HPLC) or thin layer chromatography (TLC) or other related techniques.

As used herein, the term "reacting" is used as known in the art and generally refers to the bringing together of chemical reagents in such a manner so as to allow their interaction at the molecular level to achieve a chemical or physical transformation. In some embodiments, the reacting involves two reagents, wherein one or more equivalents of second reagent are used with respect to the first reagent. The reacting steps of the processes described herein can be conducted for a time and under conditions suitable for preparing the identified product.

Preparation of compounds can involve the protection and deprotection of various chemical groups. The need for protection and deprotection, and the selection of appropriate protecting groups can be readily determined by one skilled in the art. The chemistry of protecting groups can be found, for example, in Greene, et al., *Protective Groups in Organic Synthesis*, 4d. Ed., Wiley & Sons, 2007, which is incorporated herein by reference in its entirety. Adjustments to the protecting groups and formation and cleavage methods described herein may be adjusted as necessary in light of the various substituents.

The reactions of the processes described herein can be carried out in suitable solvents which can be readily selected by one of skill in the art of organic synthesis. Suitable solvents can be substantially nonreactive with the starting materials (reactants), the intermediates, or products at the temperatures at which the reactions are carried out, e.g., temperatures which can range from the solvent's freezing

temperature to the solvent's boiling temperature. A given reaction can be carried out in one solvent or a mixture of more than one solvent. Depending on the particular reaction step, suitable solvents for a particular reaction step can be selected. In some embodiments, reactions can be carried out in the absence of solvent, such as when at least one of the reagents is a liquid or gas.

Suitable solvents can include halogenated solvents such as carbon tetrachloride, bromodichloromethane, dibromochloromethane, bromoform, chloroform, bromochloromethane, dibromomethane, butyl chloride, dichloromethane, tetrachloroethylene, trichloroethylene, 1,1,1-trichloroethane, 1,1,2-trichloroethane, 1,1-dichloroethane, 2-chloropropane, α,α,α -trifluorotoluene, 1,2-dichloroethane, 1,2-dibromoethane, hexafluorobenzene, 1,2,4-trichlorobenzene, 1,2-dichlorobenzene, chlorobenzene, fluorobenzene, mixtures thereof and the like.

Suitable ether solvents include: dimethoxymethane, tetrahydrofuran, 1,3-dioxane, 1,4-dioxane, furan, diethyl ether, ethylene glycol dimethyl ether, ethylene glycol diethyl ether, diethylene glycol dimethyl ether, diethylene glycol diethyl ether, triethylene glycol dimethyl ether, anisole, t-butyl methyl ether, mixtures thereof and the like.

Suitable protic solvents can include, by way of example and without limitation, water, methanol, ethanol, 2-nitroethanol, 2-fluoroethanol, 2,2,2-trifluoroethanol, ethylene glycol, 1-propanol, 2-propanol, 2-methoxyethanol, 1-butanol, 2-butanol, i-butyl alcohol, t-butyl alcohol, 2-ethoxyethanol, diethylene glycol, 1-, 2-, or 3- pentanol, neo-pentyl alcohol, t-pentyl alcohol, diethylene glycol monomethyl ether, diethylene glycol monoethyl ether, cyclohexanol, benzyl alcohol, phenol, or glycerol.

Suitable aprotic solvents can include, by way of example and without limitation, tetrahydrofuran (THF), N,N-dimethylformamide (DMF), N,N-dimethylacetamide (DMA), 1,3-dimethyl-3,4,5,6-tetrahydro-2(1H)-pyrimidinone (DMPU), 1,3-dimethyl-2-imidazolidinone (DMI), N-methylpyrrolidinone (NMP), formamide, N-methylacetamide, N-methylformamide, acetonitrile, dimethyl sulfoxide, propionitrile, ethyl formate, methyl acetate, hexachloroacetone, acetone, ethyl methyl ketone, ethyl acetate, sulfolane, N,N-dimethylpropionamide, tetramethylurea, nitromethane, nitrobenzene, or hexamethylphosphoramide.

Suitable hydrocarbon solvents include benzene, cyclohexane, pentane, hexane, toluene, cycloheptane, methylcyclohexane, heptane, ethylbenzene, m-, o-, or p-xylene, octane, indane, nonane, or naphthalene.

The reactions of the processes described herein can be carried out at
5 appropriate temperatures which can be readily determined by the skilled artisan. Reaction temperatures will depend on, for example, the melting and boiling points of the reagents and solvent, if present; the thermodynamics of the reaction (e.g., vigorously exothermic reactions may need to be carried out at reduced temperatures); and the kinetics of the reaction (e.g., a high activation energy barrier may need
10 elevated temperatures). “Elevated temperature” refers to temperatures above room temperature (about 22 °C).

The reactions of the processes described herein can be carried out in air or under an inert atmosphere. Typically, reactions containing reagents or products that are substantially reactive with air can be carried out using air-sensitive synthetic
15 techniques that are well known to the skilled artisan.

In some embodiments, preparation of compounds can involve the addition of acids or bases to affect, for example, catalysis of a desired reaction or formation of salt forms such as acid addition salts.

Example acids can be inorganic or organic acids. Inorganic acids include
20 hydrochloric acid, hydrobromic acid, sulfuric acid, phosphoric acid, and nitric acid. Organic acids include formic acid, acetic acid, propionic acid, butanoic acid, benzoic acid, 4-nitrobenzoic acid, methanesulfonic acid, p-toluenesulfonic acid, benzenesulfonic acid, tartaric acid, trifluoroacetic acid, propiolic acid, butyric acid, 2-butynoic acid, vinyl acetic acid, pentanoic acid, hexanoic acid, heptanoic acid,
25 octanoic acid, nonanoic acid and decanoic acid.

Example bases include lithium hydroxide, sodium hydroxide, potassium hydroxide, lithium carbonate, sodium carbonate, potassium carbonate, and sodium bicarbonate. Some example strong bases include, but are not limited to, hydroxide, alkoxides, metal amides, metal hydrides, metal dialkylamides and arylamines,
30 wherein; alkoxides include lithium, sodium and potassium salts of methyl, ethyl and t-butyl oxides; metal amides include sodium amide, potassium amide and lithium amide; metal hydrides include sodium hydride, potassium hydride and lithium

hydride; and metal dialkylamides include sodium and potassium salts of methyl, ethyl, n-propyl, i-propyl, n-butyl, t-butyl, trimethylsilyl and cyclohexyl substituted amides.

5 The intermediates and products may also include salts of the compounds disclosed herein. As used herein, the term "salt" refers to a salt formed by the addition of an acceptable acid or base to a compound disclosed herein. In some embodiments, the salts are pharmaceutically acceptable salts. As used herein, the phrase "pharmaceutically acceptable" refers to a substance that is acceptable for use in pharmaceutical applications from a toxicological perspective and does not
10 adversely interact with the active ingredient. Pharmaceutically acceptable salts, including mono- and bi- salts, include, but are not limited to, those derived from organic and inorganic acids such as, but not limited to, acetic, lactic, citric, cinnamic, tartaric, succinic, fumaric, maleic, malonic, mandelic, malic, oxalic, propionic, hydrochloric, hydrobromic, phosphoric, nitric, sulfuric, glycolic, pyruvic,
15 methanesulfonic, ethanesulfonic, toluenesulfonic, salicylic, benzoic, and similarly known acceptable acids. Lists of suitable salts are found in Remington's Pharmaceutical Sciences, 17th ed., Mack Publishing Company, Easton, Pa., 1985, p. 1418 and Journal of Pharmaceutical Science, 66, 2 (1977), each of which is incorporated herein by reference in their entireties.

20 Upon carrying out preparation of compounds according to the processes described herein, the usual isolation and purification operations such as concentration, filtration, extraction, solid-phase extraction, recrystallization, chromatography, and the like may be used, to isolate the desired products.

In some embodiments, the compounds described herein and salts thereof, are
25 substantially isolated. By "substantially isolated" is meant that the compound is at least partially or substantially separated from the environment in which it was formed or detected. Partial separation can include, for example, a composition enriched in the compound of the invention. Substantial separation can include compositions containing at least about 50%, at least about 60%, at least about 70%, at least about
30 80%, at least about 90%, at least about 95%, at least about 97%, or at least about 99% by weight of the compound of the invention, or a salt thereof. Methods for isolating compounds and their salts are routine in the art.

Processes for preparing some of the intermediates can be found in U.S. Patent No. 8,987,443, issued March 24, 2015; U.S. Patent No. 9,487,521, issued November 8, 2016; U.S. Patent No. 8,410,265, issued April 2, 2013, and U.S. Patent No. 8,765,734, issued July 1, 2014, each of which is incorporated herein by reference in its entirety.

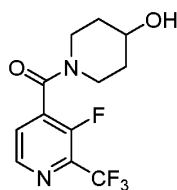
Processes and Intermediates

The present invention provides, *inter alia*, processes and intermediates for making the compound of Formula I. Accordingly, in one aspect, the provided herein is a process, comprising reacting a compound of Formula III:



III

or a salt thereof, with 4-hydroxypiperidine to form a compound of Formula IV:



IV

or a salt thereof.

In some embodiments, the reacting with 4-hydroxypiperidine is conducted in the presence of a base.

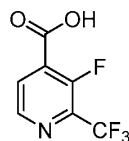
In some embodiments, the base is a tertiary amine.

In some embodiments, the tertiary amine is *N,N*-diisopropylethylamine.

In some embodiments, the reacting with 4-hydroxypiperidine is conducted in a solvent component which comprises dichloromethane.

In some embodiments, the reacting with 4-hydroxypiperidine is conducted at a temperature of from about 25 °C to about 35 °C.

In some embodiments, the compound of Formula III is formed by a process comprising reacting a compound of Formula II:



II

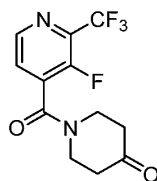
or a salt thereof, with oxalyl chloride to form the compound of Formula III, or a salt thereof.

5 In some embodiments, the reacting of the compound of Formula II with oxalyl chloride is conducted in the presence of a catalytic amount of dimethyl formamide (DMF). In some embodiments, the DMF is present at between about 0.01 and about 0.03 molar equivalents with respect to the compound of Formula II.

10 In some embodiments, the reacting of the compound of Formula II with oxalyl chloride is conducted in a solvent component comprising dichloromethane.

In some embodiments, the reacting of the compound of Formula II with oxalyl chloride is conducted at a temperature of from about 15 °C to about 25 °C.

The processes described herein can further comprise reacting the compound of Formula IV, or a salt thereof, under oxidation conditions to form a compound of
15 Formula V:



V

or a salt thereof.

20 In some embodiments, the oxidation conditions comprise at least one oxidizing agent (i.e., a first oxidizing agent). In some embodiments, the oxidation conditions comprise a second oxidizing agent.

In some embodiments, the oxidizing agent is trichloroisocyanuric acid (TCIC). In some embodiments, the TCIC is present at between about 0.5 and about 0.7 molar equivalents with respect to the compound of Formula IV.

25 . In some embodiments, the oxidizing agent is 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO). In some embodiments, the TEMPO is present at between about 0.005 and about 0.05 molar equivalents with respect to the compound of

Formula IV. In some embodiments, the TEMPO is present at between about 0.015 and about 0.025 molar equivalents with respect to the compound of Formula IV. In some embodiments, the TEMPO is present at about 0.020 molar equivalents with respect to the compound of Formula IV.

5 In some embodiments, the oxidation conditions comprise at least TCIC (a first oxidizing agent) and TEMPO (a second oxidizing agent). In some embodiments, the oxidation conditions comprise a metal bromide, e.g., sodium bromide. In some embodiments, the sodium bromide is present at between about 0.005 and about 0.015 molar equivalents with respect to the compound of Formula IV. In some
10 embodiments, the sodium bromide is present at about 0.01 equivalents with respect to the compound of Formula IV.

In some embodiments, the oxidation conditions further comprise a base. In some embodiments, the base is sodium bicarbonate and/or sodium carbonate. In some embodiments, the sodium bicarbonate is present at between about 1 and about 10
15 molar equivalents with respect to the compound of Formula IV. In some embodiments, the sodium bicarbonate is present at about 5 molar equivalents with respect to the compound of Formula IV. In some embodiments, the sodium carbonate is present at between about 0.1 and about 1 molar equivalents with respect to the compound of Formula IV. In some embodiments, the sodium carbonate is present at
20 about 0.5 molar equivalents with respect to the compound of Formula IV.

In some embodiments, the reacting of the compound of Formula IV under oxidation conditions further comprises one or more of sodium bicarbonate, sodium carbonate, and sodium bromide. In some embodiments, the reacting of the compound of Formula IV under oxidation conditions further comprises sodium bicarbonate,
25 sodium carbonate, and sodium bromide.

In some embodiments, the reacting of the compound of Formula IV under oxidation conditions further comprises a solvent component comprising dichloromethane. In some embodiments, the solvent component further comprises water.

30 In some embodiments, the oxidation conditions comprise addition of trichloroisocyanuric acid (TCIC) to a solution comprising the compound of Formula IV and TEMPO at a temperature of from about 0 °C to about 5 °C. In some

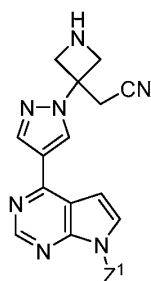
embodiments, the addition of trichloroisocyanuric acid comprises adding the trichloroisocyanuric acid in at least two portions. In some embodiments, the solution is stirred after said addition at a temperature of from about 0 °C to about 5 °C for about 30 min. In some embodiments, the process further comprises, after said

5 stirring, warming said solution to a temperature of from about 20 °C to about 25 °C for a time of about one hour to about two hours.

In some embodiments, the oxidation conditions result in the compound of Formula V having a purity of greater than about 99%.

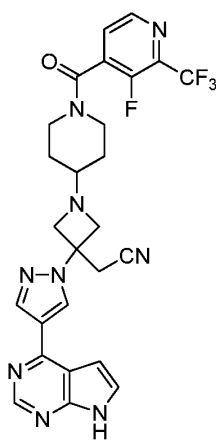
In some embodiments, the processes described herein further comprise

10 reacting a compound of Formula V with a compound of Formula VI:



VI

or a salt thereof, in the presence of a reducing agent, to form a compound of Formula I:



I

or a salt thereof;

wherein Z¹ is H or a protecting group.

In some embodiments, Z^1 is H.

In some embodiments, the reducing agent is sodium cyanoborohydride or sodium triacetoxyborohydride. In some embodiments, the reducing agent is sodium triacetoxyborohydride. In some embodiments, greater than 1 equivalent (e.g., 2
5 equivalents) of sodium triacetoxyborohydride is used based on the compound of Formula VI.

The reducing agent can be any reducing agent suitable for use in reductive amination, including various borohydride and borane reducing agents, such as those in Ellen W. Baxter and Allen B. Reitz, Reductive Aminations of Carbonyl Compounds
10 with Borohydride and Borane Reducing Agents, Organic Reactions, Chapter 1, pages 1-57 (Wiley, 2002), which is incorporated herein by reference in its entirety. Non-limiting classes of appropriate reducing agents include borohydride, cyanoborohydride, tri(C_{1-4} acyl)oxyborohydride (e.g., triacetoxyborohydride derivatives), 9-borobicyclo[3.3.1]nonane hydride, tri(C_{1-4} alkyl)borohydride, and
15 disopinocampteylcyanoborohydride derivatives, amino boranes, borane-pyridine complex, and alkylamine boranes. Non-limiting examples of appropriate reducing agents include sodium cyanoborohydride, sodium triacetoxyborohydride, sodium cyano-9-borobicyclo[3.3.1]nonane hydride, tetrabutylammonium cyanoborohydride, cyanoborohydride on a solid support, tetramethylammonium triacetoxyborohydride,
20 sodium triacetoxyborohydride, lithium triethylborohydride, lithium tri(sec-butyl)borohydride, sodium disopinocampteylcyanoborohydride, catechol borane, borane tetrahydrofuran, sodium borohydride, potassium borohydride, lithium borohydride, palladium in the presence of hydrogen gas, 5-ethyl-2-methylpyridine borane (PEMB), 2-picoline borane or polymer-supported triacetoxyborohydride. In
25 some embodiments, any of the aforementioned, and preferably sodium cyanoborohydride, is used in combination with a titanium (IV) additive, dehydrating agent, or a zinc halide additive. In some embodiments, the reducing agent is a tetra(C_{1-4} alkyl)ammonium cyanoborohydride or triacetoxyborohydride, an alkali metal cyanoborohydride or triacetoxyborohydride, or an alkaline earth
30 cyanoborohydride or triacetoxyborohydride. In some embodiments, the reducing agent is an alkali metal cyanoborohydride. In some embodiments, the reducing agent is selected from sodium cyanoborohydride and sodium triacetoxyborohydride. In

some embodiments, the reducing agent is sodium triacetoxyborohydride. As used herein, a titanium (IV) additive is a Lewis acid containing a titanium (IV) metal (e.g., titanium tetrachloride, titanium isopropoxide, titanium ethoxide, and the like).

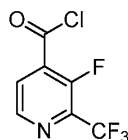
In some embodiments, the compound of Formula VI, or salt thereof, is 2-(3-
5 (4-(7H-pyrrolo[2,3-d]pyrimidin-4-yl)-1H-pyrazol-1-yl)azetidin-3-yl)acetonitrile dihydrochloride salt. In some embodiments, the reacting is carried out in the presence of at least two equivalents of a second base. In some embodiments, the second base is a tertiary amine (e.g., triethylamine). As used herein, “second” in the phrase “second base” is used to differentiate the base from other bases used in earlier or later steps of
10 the process and does not indicate that two bases must be present.

In some embodiments, greater than 1 equivalent of the compound of Formula V is used based on the compound of Formula VI, or salt thereof.

In some embodiments, reaction of a compound of Formula VI, or salt thereof, with a compound of Formula V is performed in dichloromethane solvent.

In some embodiments, the process further comprises reacting the compound of
15 Formula I with adipic acid to form the adipate salt of the compound of Formula I.

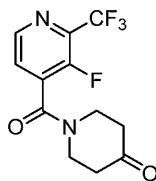
In another aspect, provided herein is a process comprising reacting a compound of Formula III:



III

20

or a salt thereof, with 4-piperidone, or a salt thereof, to form a compound of Formula V:



V

25 or a salt thereof.

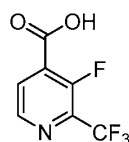
In some embodiments, the 4-piperidone, or a salt thereof, is 4-piperidone hydrochloride. In some embodiments, the 4-piperidone, or a salt thereof, is 4-piperidone hydrochloride monohydrate.

In some embodiments, the reacting of the compound of Formula III with the 4-piperidone further comprises a base. In some embodiments, the base is sodium carbonate.

In some embodiments, the reacting of the compound of Formula III with the 4-piperidone is conducted in a solvent component comprising dichloromethane.

In some embodiments, the reacting of the compound of Formula III with the 4-piperidone is conducted at a temperature of from about 0 °C to about 5 °C.

In some embodiments, the compound of Formula III is formed by a process comprising reacting a compound of Formula II:



II

5 or a salt thereof, with oxalyl chloride to form the compound of Formula III, or a salt thereof.

In some embodiments, the reacting of the compound of Formula II with oxalyl chloride is conducted in the presence of a catalytic amount of dimethyl formamide (DMF). In some embodiments, the DMF is present at between about 0.01 and about
10 0.03 molar equivalents with respect to the compound of Formula II.

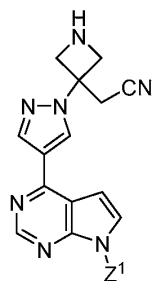
In some embodiments, the reacting of the compound of Formula II with oxalyl chloride is conducted in a solvent component comprising dichloromethane.

In some embodiments, the reacting of the compound of Formula II with oxalyl chloride is conducted at a temperature of from about 15 °C to about 25 °C.

15 In some embodiments, the compound of Formula III is not isolated before reacting the compound of Formula III with 4-piperidone.

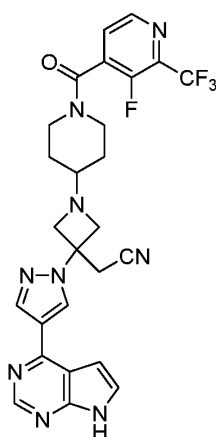
In some embodiments, the reacting of the compound of Formula II with oxalyl chloride and the reacting of the compound of Formula III with 4-piperidone are conducted in a single reactor.

In some embodiments, the process described herein further comprise reacting the compound of Formula V with a compound of Formula VI:



VI

- 5 or a salt thereof, in the presence of a reducing agent, to form a compound of Formula I:



I

or a salt thereof;

wherein Z^1 is H or a protecting group.

In some embodiments, Z^1 is H.

- In some embodiments, the reducing agent is sodium cyanoborohydride or sodium triacetoxyborohydride. In some embodiments, the reducing agent is sodium triacetoxyborohydride. In some embodiments, greater than 1 equivalent (e.g., 2 equivalents) of sodium triacetoxyborohydride is used based on the compound of Formula VI.

In some embodiments, the compound of Formula VI, or salt thereof, is 2-(3-(4-(7H-pyrrolo[2,3-d]pyrimidin-4-yl)-1H-pyrazol-1-yl)azetidin-3-yl)acetonitrile

dihydrochloride salt. In some embodiments, the reacting is carried out in the presence of at least two equivalents of a second base. In some embodiments, the second base is a tertiary amine (e.g., triethylamine).

In some embodiments, greater than 1 equivalent of the compound of Formula V is used based on the compound of Formula VI, or salt thereof.

In some embodiments, reaction of a compound of Formula VI, or salt thereof, with a compound of Formula V is performed in dichloromethane solvent.

In some embodiments, the process further comprises reacting the compound of Formula I with adipic acid to form the adipate salt of the compound of Formula I.

In an aspect, provided herein is a compound of Formula III:



or a salt thereof.

Uses

The compound of Formula I, {1-{1-[3-fluoro-2-(trifluoromethyl)isonicotinoyl]piperidin-4-yl}-3-[4-(7H-pyrrolo[2,3-d]pyrimidin-4-yl)-1H-pyrazol-1-yl]azetidin-3-yl}acetonitrile, is an inhibitor of JAK (e.g., JAK1, JAK2). JAK inhibitors are useful in treating various JAK-associated diseases or disorders. Examples of JAK-associated diseases include diseases involving the immune system including, for example, organ transplant rejection (e.g., allograft rejection and graft versus host disease). Further examples of JAK-associated diseases include autoimmune diseases such as multiple sclerosis, rheumatoid arthritis, juvenile arthritis, psoriatic arthritis, type I diabetes, lupus, psoriasis, inflammatory bowel disease, ulcerative colitis, Crohn's disease, myasthenia gravis, immunoglobulin nephropathies, myocarditis, autoimmune thyroid disorders, chronic obstructive pulmonary disease (COPD), and the like. In some embodiments, the autoimmune disease is an autoimmune bullous skin disorder such as pemphigus vulgaris (PV) or bullous pemphigoid (BP).

Further examples of JAK-associated diseases include allergic conditions such as asthma, food allergies, eszematous dermatitis, contact dermatitis, atopic dermatitis (atropic eczema), and rhinitis. Further examples of JAK-associated diseases include viral diseases such as Epstein Barr Virus (EBV), Hepatitis B, Hepatitis C, HIV, HTLV 1, Varicella-Zoster Virus (VZV) and Human Papilloma Virus (HPV). Further examples of JAK-associated disease include diseases associated with cartilage turnover, for example, gouty arthritis, septic or infectious arthritis, reactive arthritis, reflex sympathetic dystrophy, algodystrophy, Tietze syndrome, costal athroopathy, osteoarthritis deformans endemica, Mseleni disease, Handigodu disease, degeneration resulting from fibromyalgia, systemic lupus erythematosus, scleroderma, or ankylosing spondylitis.

Further examples of JAK-associated disease include congenital cartilage malformations, including hereditary chrondrolysis, chrondrodysplasias, and pseudochrondrodysplasias (e.g., microtia, enotia, and metaphyseal chrondrodysplasia).

Further examples of JAK-associated diseases or conditions include skin disorders such as psoriasis (for example, psoriasis vulgaris), atopic dermatitis, skin rash, skin irritation, skin sensitization (e.g., contact dermatitis or allergic contact dermatitis). For example, certain substances including some pharmaceuticals when topically applied can cause skin sensitization. In some embodiments, co-administration or sequential administration of at least one JAK inhibitor of the invention together with the agent causing unwanted sensitization can be helpful in treating such unwanted sensitization or dermatitis. In some embodiments, the skin disorder is treated by topical administration of at least one JAK inhibitor of the invention.

Further examples of JAK-associated diseases or conditions include those characterized by solid tumors (e.g., prostate cancer, renal cancer, hepatic cancer, pancreatic cancer, gastric cancer, breast cancer, lung cancer, cancers of the head and neck, thyroid cancer, glioblastoma, Kaposi's sarcoma, Castleman's disease, uterine leiomyosarcoma, melanoma etc.), hematological cancers (e.g., lymphoma, leukemia such as acute lymphoblastic leukemia (ALL), acute myelogenous leukemia (AML) or multiple myeloma), and skin cancer such as cutaneous T-cell lymphoma (CTCL) and cutaneous B-cell lymphoma. Example CTCLs include Sezary syndrome and mycosis

fungoides. Other examples of JAK-associated diseases or conditions include pulmonary arterial hypertension.

Other examples of JAK-associated diseases or conditions include inflammation-associated cancers. In some embodiments, the cancer is associated with inflammatory bowel disease. In some embodiments, the inflammatory bowel disease is ulcerative colitis. In some embodiments, the inflammatory bowel disease is Crohn's disease. In some embodiments, the inflammation-associated cancer is colitis-associated cancer. In some embodiments, the inflammation-associated cancer is colon cancer or colorectal cancer. In some embodiments, the cancer is gastric cancer, gastrointestinal carcinoid tumor, gastrointestinal stromal tumor (GIST), adenocarcinoma, small intestine cancer, or rectal cancer.

JAK-associated diseases can further include those characterized by expression of: JAK2 mutants such as those having at least one mutation in the pseudo-kinase domain (*e.g.*, JAK2V617F); JAK2 mutants having at least one mutation outside of the pseudo-kinase domain; JAK1 mutants; JAK3 mutants; erythropoietin receptor (EPOR) mutants; or deregulated expression of CRLF2.

JAK-associated diseases can further include myeloproliferative disorders (MPDs) such as polycythemia vera (PV), essential thrombocythemia (ET), myelofibrosis with myeloid metaplasia (MMM), primary myelofibrosis (PMF), chronic myelogenous leukemia (CML), chronic myelomonocytic leukemia (CMML), hypereosinophilic syndrome (HES), systemic mast cell disease (SMCD), and the like. In some embodiments, the myeloproliferative disorder is myelofibrosis (*e.g.*, primary myelofibrosis (PMF) or post polycythemia vera/essential thrombocythemia myelofibrosis (Post-PV/Post-ET MF)). In some embodiments, the myeloproliferative disorder is post-essential thrombocythemia myelofibrosis (Post-ET MF). In some embodiments, the myeloproliferative disorder is post polycythemia vera myelofibrosis (Post-PV MF).

Other examples of JAK-associated diseases or conditions include ameliorating the dermatological side effects of other pharmaceuticals by administration of the compound of the invention. For example, numerous pharmaceutical agents result in unwanted allergic reactions which can manifest as acneiform rash or related dermatitis. Example pharmaceutical agents that have such undesirable side effects

include anti-cancer drugs such as gefitinib, cetuximab, erlotinib, and the like. The compounds of the invention can be administered systemically or topically (e.g., localized to the vicinity of the dermatitis) in combination with (e.g., simultaneously or sequentially) the pharmaceutical agent having the undesirable dermatological side effect. In some embodiments, the compound of the invention can be administered topically together with one or more other pharmaceuticals, where the other pharmaceuticals when topically applied in the absence of a compound of the invention cause contact dermatitis, allergic contact sensitization, or similar skin disorder. Accordingly, compositions of the invention include topical formulations containing the compound of the invention and a further pharmaceutical agent which can cause dermatitis, skin disorders, or related side effects.

Further JAK-associated diseases include inflammation and inflammatory diseases. Example inflammatory diseases include sarcoidosis, inflammatory diseases of the eye (e.g., iritis, uveitis, scleritis, conjunctivitis, or related disease), inflammatory diseases of the respiratory tract (e.g., the upper respiratory tract including the nose and sinuses such as rhinitis or sinusitis or the lower respiratory tract including bronchitis, chronic obstructive pulmonary disease, and the like), inflammatory myopathy such as myocarditis, and other inflammatory diseases. In some embodiments, the inflammation disease of the eye is blepharitis.

Further JAK-associated diseases include ischemia reperfusion injuries or a disease or condition related to an inflammatory ischemic event such as stroke or cardiac arrest, endotoxin-driven disease state (e.g., complications after bypass surgery or chronic endotoxin states contributing to chronic cardiac failure), anorexia, cachexia, fatigue such as that resulting from or associated with cancer, restenosis, sclerodermitis, fibrosis, conditions associated with hypoxia or astrogliosis such as, for example, diabetic retinopathy, cancer, or neurodegeneration, and other inflammatory diseases such as systemic inflammatory response syndrome (SIRS) and septic shock. Other JAK-associated diseases include gout and increased prostate size due to, e.g., benign prostatic hypertrophy or benign prostatic hyperplasia, as well as bone resorption diseases such as osteoporosis or osteoarthritis, bone resorption diseases associated with: hormonal imbalance and/or hormonal therapy, autoimmune disease (e.g. osseous sarcoidosis), or cancer (e.g. myeloma).

Further JAK-associated diseases include a dry eye disorder. As used herein, “dry eye disorder” is intended to encompass the disease states summarized in a recent official report of the Dry Eye Workshop (DEWS), which defined dry eye as “a multifactorial disease of the tears and ocular surface that results in symptoms of discomfort, visual disturbance, and tear film instability with potential damage to the ocular surface. It is accompanied by increased osmolarity of the tear film and inflammation of the ocular surface.” Lemp, “The Definition and Classification of Dry Eye Disease: Report of the Definition and Classification Subcommittee of the International Dry Eye Workshop”, *The Ocular Surface*, 5(2), 75-92 April 2007, which is incorporated herein by reference in its entirety. In some embodiments, the dry eye disorder is selected from aqueous tear-deficient dry eye (ADDE) or evaporative dry eye disorder, or appropriate combinations thereof. In some embodiments, the dry eye disorder is Sjogren syndrome dry eye (SSDE). In some embodiments, the dry eye disorder is non-Sjogren syndrome dry eye (NSSDE).

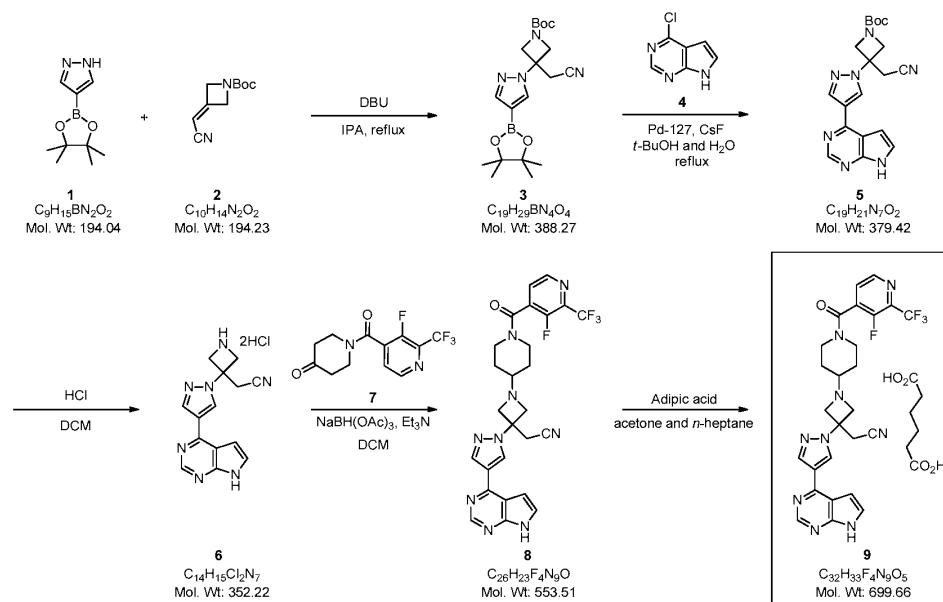
Further JAK-associated diseases include conjunctivitis, uveitis (including chronic uveitis), chorioiditis, retinitis, cyclitis, scleritis, episcleritis, or iritis. Other JAK-associated diseases include respiratory dysfunction or failure associated with viral infection, such as influenza and SARS.

EXAMPLES

The invention will be described in greater detail by way of specific examples. The following examples are offered for illustrative purposes, and are not intended to limit the invention in any manner. Those of skill in the art will readily recognize a variety of noncritical parameters which can be changed or modified to yield essentially the same results.

Example 1. Synthesis of 2-(3-(4-(7*H*-Pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)-1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)azetidin-3-yl)acetonitrile Adipate (9)

Scheme I



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***tert*-Butyl 3-(cyanomethyl)-3-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1*H*-pyrazol-1-yl)azetidine-1-carboxylate (3).** To a 1-L flask equipped with a nitrogen inlet, a thermocouple, and a mechanical stirrer were sequentially added isopropanol (IPA, 200 mL), 1,8-diazabicyclo[5,4,0]undec-ene (DBU, 9.8 g, 64.4 mmol, 0.125 equiv), 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1*H*-pyrazole (**1**, 101 g, 520.51 mmol, 1.01 equiv) and *tert*-butyl 3-(cyanomethylene)azetidine-1-carboxylate (**2**, 100 g, 514.85 mmol) at ambient temperature to generate a reaction mixture as a suspension. The resulting reaction mixture was heated to reflux in 30 minutes to provide a homogenous solution and the mixture was maintained at reflux for an additional 2 – 3 hours. After the reaction was complete as monitored by HPLC, *n*-heptane (400 mL) was gradually added to the reaction mixture in 45 minutes while maintaining the mixture at reflux. Solids were precipitated out during the *n*-heptane addition. Once *n*-heptane addition was complete, the mixture was gradually cooled to ambient temperature and stirred at ambient temperature for an additional 1 hour. The

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solids were collected by filtration, washed with *n*-heptane (200 mL), and dried under vacuum at 50 °C with nitrogen sweeping to constant weight to afford *tert*-butyl 3-(cyanomethyl)-3-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1*H*-pyrazol-1-yl)azetidine-1-carboxylate (**3**, 181 g, 199.9 g theoretical, 90.5%) as a white to pale yellow solid. For **3**: ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.31 (s, 1H), 7.74 (s, 1H), 4.45 – 4.23 (m, 2H), 4.23 – 4.03 (m, 2H), 3.56 (s, 2H), 1.38 (s, 9H), 1.25 (s, 12H) ppm; ¹³C NMR (101 MHz, DMSO-*d*₆) δ 155.34, 145.50, 135.88, 116.88, 107.08 (br), 83.15, 79.36, 58.74 (br), 56.28, 27.96, 26.59, 24.63 ppm; C₁₉H₂₉BN₄O₄ (MW 388.27), LCMS (EI) *m/e* 389 (M⁺ + H).

***tert*-Butyl 3-(4-(7*H*-pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)-3-(cyanomethyl)-azetidine-1-carboxylate (**5**).**

To a 1-L flask equipped with a nitrogen inlet, a thermocouple, and a mechanical stirrer were added 4-chloro-7*H*-pyrrolo[2,3-*d*]pyrimidine (**4**, 39.6 g, 257.6 mmol), *tert*-butyl 3-(cyanomethyl)-3-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1*H*-pyrazol-1-yl)azetidine-1-carboxylate (**3**, 100 g, 257.6 mmol, 1.0 equiv), cesium fluoride (136.9 g, 901.4 mmol, 3.5 equiv), *tert*-butanol (250 mL), water (250 mL), and [1,1'-bis(di-cyclohexylphosphino)ferrocene]dichloropalladium(II) (Pd-127, 351.4 mg, 0.46 mmol, 0.0018 equiv) at ambient temperature. The resulting reaction mixture was de-gassed and refilled with nitrogen for 3 times before being heated to reflux and maintained at reflux under nitrogen for 20 – 24 hours. When HPLC showed the reaction was complete, the reaction mixture was cooled to 45 – 55 °C in 30 minutes, the two phases were separated, and the aqueous phase was discarded. To the organic phase was added *n*-heptane (125 mL) in 30 minutes at 45 – 55 °C. The resulting mixture was slowly cooled to ambient temperature in one hour and stirred at ambient temperature for an additional 2 hours. The solids were collected by filtration, washed with *n*-heptane (100 mL), and dried under vacuum at 50 °C with nitrogen sweeping to constant weight to afford *tert*-butyl 3-(4-(7*H*-pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)-3-(cyanomethyl)-azetidine-1-carboxylate (**5**, 96.8 g, 97.7 g theoretical, 99%) as a pale yellow solid. For **5**: ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.89 (s, 1H), 8.68 (s, 1H), 8.44 (s, 1H), 7.60 (d, *J* = 3.5 Hz, 1H), 7.06 (d, *J* = 3.6 Hz, 1H), 4.62 – 4.41 (m, 2H), 4.31 – 4.12 (m, 2H), 3.67 (s, 2H), 1.39 (s, 9H) ppm; ¹³C NMR (101

MHz, DMSO-*d*₆) δ 155.40, 152.60, 150.63, 149.15, 139.76, 129.53, 127.65, 122.25, 116.92, 113.21, 99.71, 79.45, 58.34 (br), 56.80, 27.99, 26.83 ppm; C₁₉H₂₁N₇O₂ (MW 379.4), LCMS (EI) *m/e* 380 (M⁺ + H).

5 **2-(3-(4-(7*H*-Pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)azetidin-3-yl)acetonitrile dihydrochloride salt (6).** To a 0.5-L flask equipped with a nitrogen inlet, a thermocouple, an additional funnel, and a mechanical stirrer were added *tert*-butyl 3-(4-(7*H*-pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)-3-(cyanomethyl)azetidine-1-carboxylate (**5**, 15 g, 39.5 mmol), water (7.5 mL, 416
10 mmol) and dichloromethane (75 mL) at room temperature. The mixture was stirred at room temperature to generate a suspension. To the suspension was added a solution of 5 M hydrogen chloride (HCl) in isopropanol (55 mL, 275 mmol, 7.0 equiv) in 5 minutes. The resulting reaction mixture was then heated to gentle reflux and maintained at reflux for 3-4 hours. After the reaction was completed as monitored by
15 HPLC, *tert*-butyl methyl ether (TBME, 45 mL) was added to the reaction suspension. The mixture was gradually cooled to room temperature, and stirred for an additional one hour. The solids were collected by filtration, washed with *tert*-butyl methyl ether (TBME, 45 mL) and dried under vacuum at 50 °C with nitrogen sweeping to constant weight to afford 2-(3-(4-(7*H*-pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)azetidin-
20 3-yl)acetonitrile dihydrochloride salt (**6**, 13.6 g, 13.9 g theoretical, 98%) as an off-white to light yellow solid. For **6**: ¹H NMR (400 MHz, D₂O) δ 8.96 (s, 1H), 8.81 (s, 1H), 8.49 (s, 1H), 7.78 (d, *J* = 3.8 Hz, 1H), 7.09 (d, *J* = 3.7 Hz, 1H), 4.93 (d, *J* = 12.8 Hz, 2H), 4.74 (d, *J* = 12.5 Hz, 2H), 3.74 (s, 2H) ppm; ¹³C NMR (101 MHz, D₂O) δ
25 151.35, 143.75, 143.33, 141.33, 132.03, 131.97, 115.90, 114.54, 113.85, 103.18, 59.72, 54.45 (2C), 27.02 ppm; C₁₄H₁₅Cl₂N₇ (C₁₄H₁₃N₇ for free base, MW 279.30), LCMS (EI) *m/e* 280 (M⁺ + H).

30 **2-(3-(4-(7*H*-Pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)-1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)azetidin-3-yl)acetonitrile (8, Free Base).** To a 0.5-L flask equipped with a nitrogen inlet, a thermocouple, an additional funnel, and a mechanical stirrer were added 2-(3-(4-(7*H*-pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)azetidin-3-yl)acetonitrile dihydrochloride salt (**6**, 20 g, 56.78

mmol), dichloromethane (200 mL) and triethylamine (TEA, 16.62 mL, 119.2 mmol, 2.1 equiv) at ambient temperature. The mixture was stirred at ambient temperature for 30 minutes before 1-(3-fluoro-2-(trifluoromethyl)-isonicotinoyl)piperidin-4-one (**7**, 17.15 g, 57.91 mmol, 1.02 equiv) was added to the mixture. The mixture was then

5 treated with sodium triacetoxyborohydride (25.34 g, 113.6 mmol, 2.0 equiv) in 5 minutes at ambient temperature (below 26 °C). The resulting reaction mixture was stirred at ambient temperature for 2 hours. After the reaction was complete as monitored by HPLC, the reaction mixture was quenched with saturated NaHCO₃ aqueous solution (200 mL). The two phases were separated and the aqueous phase

10 was extracted with methylene chloride (200 mL). The combined organic phase was washed with 4% brine (100 mL) followed by solvent switch of methylene chloride to acetone by distillation. The resulting solution of the desired crude product (**8**) in acetone was directly used for the subsequent adipate salt formation. A small portion of solution was purified by column chromatography (SiO₂, 0 – 10% of MeOH in

15 EtOAc gradient elution) to afford the analytically pure 2-(3-(4-(7*H*-pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)-1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)azetidin-3-yl)acetonitrile (**8** free base) as an off-white solid. For **8**: ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.17 (d, *J* = 2.8 Hz, 1H), 8.85 (s, 1H), 8.70 (m, 2H), 8.45 (s, 1H), 7.93 (t, *J* = 4.7 Hz, 1H), 7.63 (dd, *J* = 3.6, 2.3

20 Hz, 1H), 7.09 (dd, *J* = 3.6, 1.7 Hz, 1H), 4.10 (m, 1H), 3.78 (d, *J* = 7.9 Hz, 2H), 3.61 (t, *J* = 7.9 Hz, 1H), 3.58 (s, 2H), 3.46 (m, 1H), 3.28 (t, *J* = 10.5 Hz, 1H), 3.09 (ddd, *J* = 13.2, 9.5, 3.1 Hz, 1H), 2.58 (m, 1H), 1.83 – 1.75 (m, 1H), 1.70 – 1.63 (m, 1H), 1.35 – 1.21 (m, 2H) ppm; ¹³C NMR (101 MHz, DMSO-*d*₆) δ 160.28, (153.51, 150.86), 152.20, 150.94, 149.62, (146.30, 146.25), 139.48, (134.78, 134.61), (135.04, 134.92,

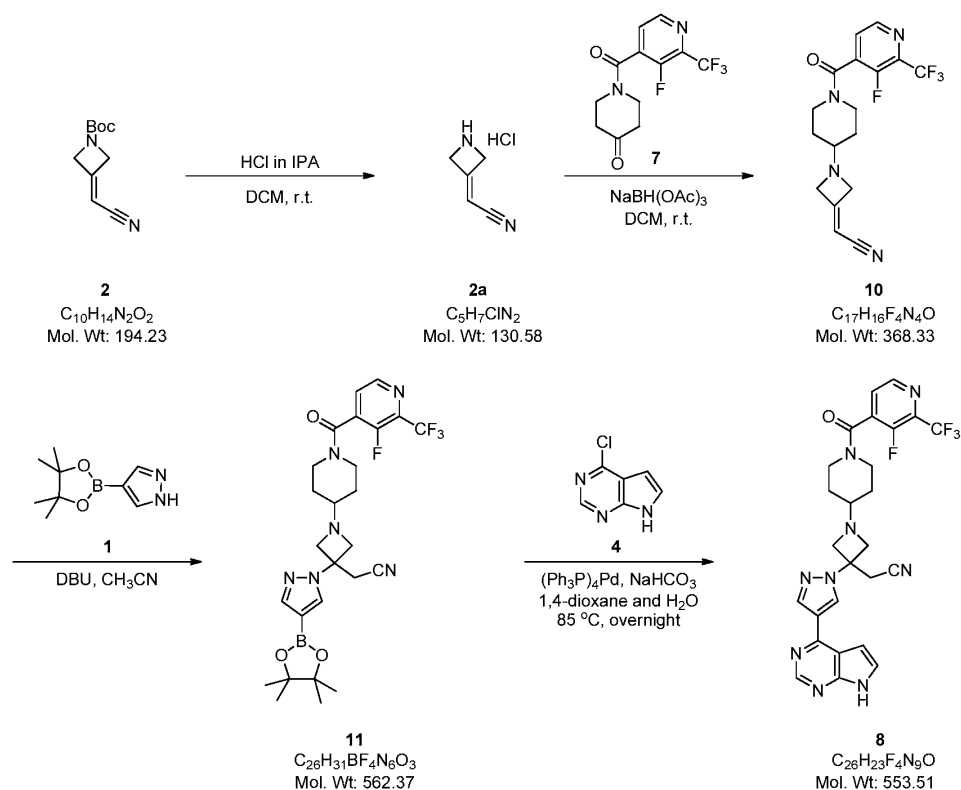
25 134.72, 134.60, 134.38, 134.26, 134.03, 133.92), 129.22, 127.62, 126.84, 121.99, 122.04, (124.77, 122.02, 119.19, 116.52), 117.39, 113.00, 99.99, 61.47, 60.49, 57.05, 44.23, 28.62, 27.88, 27.19 ppm; C₂₆H₂₃F₄N₉O (MW, 553.51), LCMS (EI) *m/e* 554.1 (M⁺ + H).

30 **2-(3-(4-(7*H*-Pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)-1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)azetidin-3-yl)acetonitrile Adipate (**9**).** To a 0.5-L flask equipped with a mechanical stirrer, a thermocouple, an addition

funnel, and a nitrogen inlet was added a solution of crude 2-(3-(4-(7*H*-pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)-1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)azetidin-3-yl)acetonitrile (**8** free base, 31.38 g, 56.7 mmol) in acetone (220 mL) and adipic acid (8.7 g, 59.53 mmol, 1.05 equiv) at ambient temperature. The reaction mixture was then heated to reflux to give a solution. *n*-Heptane (220 mL) was gradually added to the reaction mixture at 40 – 50 °C in one hour. The resulting mixture was gradually cooled to ambient temperature in one hour and stirred at ambient temperature for an additional 16 hours. The solids were collected by filtration, washed with *n*-heptane (2 X 60 mL), and dried under vacuum at 50 °C with nitrogen sweeping to constant weight to afford 2-(3-(4-(7*H*-Pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)-1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)azetidin-3-yl)acetonitrile adipate (**9**, 34.0 g, 39.7 g theoretical, 85.6% for two steps) as a white to off-white solid. **9**: ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.16 (s, 1H), 12.05 (brs, 2H), 8.85 (s, 1H), 8.72 (s, 1H), 8.69 (d, *J* = 4.7 Hz, 1H), 8.45 (s, 1H), 7.93 (t, *J* = 4.7 Hz, 1H), 7.63 (dd, *J* = 3.6, 2.3 Hz, 1H), 7.09 (dd, *J* = 3.6, 1.7 Hz, 1H), δ 4.11 (dt, *J* = 11.0, 4.4 Hz, 1H), 3.77 (d, *J* = 7.8 Hz, 2H), 3.60 (t, *J* = 7.8 Hz, 2H), 3.58 (s, 2H), 3.44 (dt, *J* = 14.4, 4.6 Hz, 1H), 3.28 (t, *J* = 10.4 Hz, 1H), 3.09 (ddd, *J* = 13.2, 9.6, 3.2 Hz, 1H), 2.58 (tt, *J* = 8.6, 3.5 Hz, 1H), 2.28 – 2.17 (m, 4H), 1.83 – 1.74 (m, 1H), 1.67 (d, *J* = 11.0 Hz, 1H), 1.59 – 1.46 (m, 4H), 1.37 – 1.21 (m, 2H) ppm; ¹³C NMR (101 MHz, DMSO-*d*₆) δ 174.38, 160.29, (153.52, 150.87), 152.20, 150.94, 149.63, (146.30, 146.25), 139.48, (134.79, 134.62), (135.08, 134.97, 134.74, 134.62, 134.38, 134.28, 134.04, 133.93), 129.21, 127.62, 126.84, 122.05, (124.75, 122.02, 119.29, 116.54), 117.39, 113.01, 99.99, 61.47, 60.50, 57.06, 44.24, 33.42, 30.70, 28.63, 27.89, 27.20, 24.07 ppm; C₃₂H₃₃F₄N₉O₅ (MW 699.66; C₂₆H₂₃F₄N₉O for free base, MW, 553.51), LCMS (EI) *m/e* 554.0 (M⁺ + H).

Example 2: Alternative Synthesis of 2-(3-(4-(7H-Pyrrolo[2,3-d]pyrimidin-4-yl)-1H-pyrazol-1-yl)-1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)azetidin-3-yl)acetonitrile

Scheme II



2-(Azetidin-3-ylidene)acetonitrile hydrochloride (2a). To a 0.5-L flask equipped with a nitrogen inlet, a thermocouple, and a mechanical stirrer were added *tert*-butyl 3-(cyanomethylene)azetidine-1-carboxylate (**2**, 30 g, 154.46 mmol) and methylenechloride (300 mL) at ambient temperature. The solution was then treated with a solution of 5 M hydrogen chloride (HCl) in isopropanol solution (294.2 mL, 1.54 mol, 10 equiv) at ambient temperature and the resulting reaction mixture was stirred at ambient temperature for 18 hours. After the reaction was complete as monitored by HPLC, the suspension was added *tert*-butyl methyl ether (TBME, 150 mL), and the mixture was stirred at ambient temperature for 2 hours. The solids was collected by filtration, washed with *n*-heptane (2 X 100 mL), and dried on the

filtration funnel at ambient temperature for 3 hours to afford 2-(azetidin-3-ylidene)acetonitrile hydrochloride (**2a**, 13.7 g, 20.2 g theoretical, 67.8 %) as a white solid. For **2a**: ^1H NMR (500 MHz, DMSO- d_6) δ 9.99 (s, 2H), 5.94 (p, $J = 2.5$ Hz, 1H), 4.85 – 4.80 (m, 2H), 4.77 – 4.71 (m, 2H) ppm; ^{13}C NMR (126 MHz, DMSO- d_6) δ 155.65, 114.54, 94.78, 55.26, 54.63 ppm; $\text{C}_5\text{H}_7\text{ClN}_2$ (MW 130.58; $\text{C}_5\text{H}_6\text{N}_2$ for free base, MW 94.11), LCMS (EI) m/e 95 ($\text{M}^+ + \text{H}$).

2-(1-(1-(3-Fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)azetidin-3-ylidene)acetonitrile (10). To a 0.25-L flask equipped with a nitrogen inlet, a thermocouple, and a magnetic stirrer were added 2-(azetidin-3-ylidene)acetonitrile hydrochloride (**2a**, 4.5 g, 34.46 mmol), 1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-one (**7**, 10 g, 34.46 mmol, 1.0 equiv), and methylenechloride (100 mL) at ambient temperature and the resulting mixture was then treated with sodium triacetoxyborohydride (14.6 g, 68.93 mmol, 2.0 equiv) at ambient temperature. The reaction mixture was stirred at ambient temperature for 2 hours before being quenched with saturated sodium bicarbonate (NaHCO_3) aqueous solution (50 mL). The two phases were separated and the aqueous phase was extracted with dichloromethane (200 mL). The combined organic phase was washed with water (50 mL) and brine (50 mL) and concentrated under reduced pressure to afford the crude desired product (**10**), which was purified by column chromatography (SiO_2 , 0 – 10 % of ethyl acetate in hexane gradient elution) to afford 2-(1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)azetidin-3-ylidene)acetonitrile (**10**, 9.5 g, 12.7 g theoretical, 74.8 %) as a white solid. For **10**: ^1H NMR (400 MHz, CDCl_3) δ 8.57 (d, $J = 4.7$ Hz, 1H), 7.54 (t, $J = 4.6$ Hz, 1H), 5.29 (p, $J = 2.4$ Hz, 1H), 4.18 – 4.08 (m, 1H), 4.08 – 4.03 (m, 2H), 3.98 – 3.94 (m, 2H), 3.57 – 3.39 (m, 2H), 3.17 – 3.04 (m, 1H), 2.56 (tt, $J = 7.4, 3.5$ Hz, 1H), 1.86 – 1.77 (m, 1H), 1.75 – 1.64 (m, 1H), 1.54 – 1.43 (m, 1H), 1.43 – 1.31 (m, 1H) ppm; ^{13}C NMR (101 MHz, CDCl_3) δ 161.34, 160.73, 152.62 (d, $J = 269.1$ Hz), 145.75 (d, $J = 6.1$ Hz), 136.73 (qd, $J = 36.1, 12.0$ Hz), 134.56 (d, $J = 16.9$ Hz), 126.89, 120.58 (qd, $J = 275.0, 4.9$ Hz), 115.11, 92.04, 62.05, 60.57 (2C), 44.47, 39.42, 29.38, 28.47 ppm; $\text{C}_{17}\text{H}_{16}\text{F}_4\text{N}_4\text{O}$ (MW 368.33), LCMS (EI) m/e 369 ($\text{M}^+ + \text{H}$).

2-(1-(1-(3-Fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)-3-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1H-pyrazol-1-yl)azetidin-3-yl)acetonitrile (11). To a 25 mL flask equipped with a nitrogen inlet, a thermocouple, and a magnetic stirrer were added 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1H-pyrazole (**1**, 210 mg, 1.08 mmol, 1.08 equiv), 2-(1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)azetidin-3-ylidene)acetonitrile (**10**, 370 mg, 1.0 mmol) and acetonitrile (3 mL) at ambient temperature. The solution was then treated with 1,8-diazabicyclo[5,4,0]undec-ene (DBU, 173 mg, 0.17 mL, 1.12 mmol, 1.12 equiv) at ambient temperature and the resulting reaction mixture was warmed to 50 °C and stirred at 50 °C for overnight. When the reaction was complete as monitored by HPLC, the reaction mixture was directly load on a silica gel (SiO₂) column for chromatographic purification (0 – 2.5 % MeOH in ethyl acetate gradient elution) to afford 2-(1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)-3-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1H-pyrazol-1-yl)azetidin-3-yl)acetonitrile (**11**, 263 mg, 562.4 mg theoretical, 46.7 %) as a white solid. For **11**: ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.64 (d, *J* = 4.7 Hz, 1H), 8.22 (d, *J* = 0.6 Hz, 1H), 7.88 (dd, *J* = 4.7 Hz, 1H), 7.69 (s, 1H), 4.10 – 3.99 (m, 1H), 3.58 (d, *J* = 7.8 Hz, 2H), 3.52 – 3.42 (m, 2H), 3.44 (s, 2H), 3.41 – 3.33 (m, 1H), 3.28 – 3.15 (m, 1H), 3.03 (ddd, *J* = 12.9, 9.2, 3.2 Hz, 1H), 2.51 – 2.44 (m, 1H), 1.77 – 1.66 (m, 1H), 1.64 – 1.54 (m, 1H), 1.28 – 1.17 (m, 2H), 1.24 (s, 12H) ppm; ¹³C NMR (101 MHz, DMSO-*d*₆) δ 160.22, 152.13 (d, *J* = 265.8 Hz), 146.23 (d, *J* = 5.7 Hz), 145.12, 135.41, 134.66 (d, *J* = 16.9 Hz), 134.43 (qd, *J* = 35.0, 11.7 Hz), 127.58, 120.61 (qd, *J* = 274.4, 4.6 Hz), 117.35, 106.59 (br), 83.10, 61.40, 60.53 (2C), 56.49, 44.17, 38.99, 28.55, 27.82, 27.02, 24.63 ppm; C₂₆H₃₁BF₄N₆O₃ (MW 562.37), LCMS (EI) *m/e* 563 (M⁺ + H).

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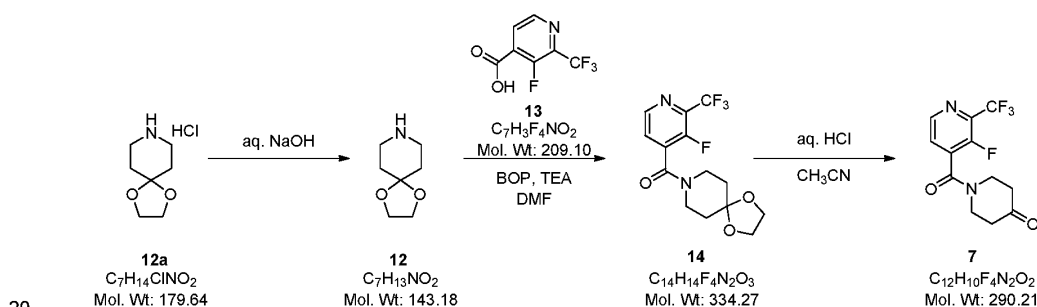
2-(3-(4-(7H-Pyrrolo[2,3-*d*]pyrimidin-4-yl)-1H-pyrazol-1-yl)-1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)azetidin-3-yl)acetonitrile (8). To a 25-mL flask equipped with a nitrogen inlet, a thermocouple, an additional funnel, and a magnetic stirrer were added 2-(1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)-3-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1H-pyrazol-1-yl)azetidin-3-yl)acetonitrile (**11**, 307 mg, 0.546 mmol), 4-chloro-7H-pyrrolo[2,3-*d*]pyrimidine (**4**, 84.8 mg, 0.548 mmol, 1.0 equiv), sodium bicarbonate

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(NaHCO₃, 229 mg, 2.72 mmol, 5.0 equiv), water (1.6 mL), and 1,4-dioxane (1.6 mL) at ambient temperature. The mixture was then treated with tetrakis(triphenylphosphine)palladium(0) (12.8 mg, 0.011 mmol, 0.02 equiv) at ambient temperature and the resulting reaction mixture was de-gassed and refilled with nitrogen for 3 times before being heated to 85 °C. The reaction mixture was stirred at 85 °C under nitrogen for overnight. When the reaction was complete as monitored by HPLC, the reaction mixture was concentrated to dryness under reduced pressure and the desired product, 2-(3-(4-(7*H*-pyrrolo[2,3-*d*]pyrimidin-4-yl)-1*H*-pyrazol-1-yl)-1-(1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-yl)azetidin-3-yl)acetonitrile (**8** free base, 135 mg, 302.2 mg theoretical, 44.6 %), was obtained as off-white solids by direct silica gel (SiO₂) column chromatography (0 – 10% of ethyl acetate in hexane gradient elution) purification of the dried reaction mixture. The compound obtained by this synthetic approach is identical in every comparable aspect to the compound 8 manufactured by the synthetic method as described above in Example 1.

Example 3. Synthesis of 1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-one (**7**)

Scheme III



(3-Fluoro-2-(trifluoromethyl)pyridin-4-yl)(1,4-dioxo-8-azaspiro[4,5]decan-8-yl)methanone (**14**). To a 30 L reactor equipped with a mechanic stirrer, an addition funnel and a septum was charged sodium hydroxide (NaOH, 1.4 kg, 35 mol, 2.0 equiv) and water (7 L) and the resulting solution was treated with 1,4-dioxo-8-azaspiro[4,5]decane hydrochloride (3.13 kg, 17.43 mol) at ambient temperature. The

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resulting mixture was then stirred at ambient temperature for 30 minutes before being saturated with solid sodium chloride (1.3 kg) and extracted with 2-methyl-tetrahydrofuran (3 x 7 L). The combined organic phase was dried with anhydrous sodium sulfate (Na_2SO_4 , 1.3 kg) and concentrated under reduced pressure (70 mmHg) at 50 °C after removal of the drying reagent, sodium sulfate (Na_2SO_4), by filtration. The yellow oil thus obtained was distilled under reduced pressure (80 mmHg, bp 115 to 120 °C) to afford 1,4-dioxo-8-azaspiro[4.5]decane (2.34 kg, 2.496 kg theoretical, 93.8%) as a clear oil, which was used directly in the subsequent coupling reaction. To a dried 100 L reactor equipped with a mechanic stirrer, an addition funnel, a thermometer and a vacuum outlet was charged 3-fluoro-2-(trifluoromethyl)isonicotinic acid (**13**, 3.0 kg, 14.35 mol), benzotriazol-1-yloxytris(dimethylamino)phosphonium hexafluorophosphate (BOP reagent, 7.6 kg, 17.2 mol, 1.2 equiv), 1,4-dioxo-8-azaspiro[4.5]decane (2.34 kg, 16.36 mol, 1.14 equiv) and *N,N*-dimethylformamide (DMF, 18 L) at ambient temperature. The resulting solution was then stirred at ambient temperature for 20 minutes before being cooled to 5 to 10 °C. Triethylamine (Et_3N , 4 L, 28.67 mol, 2.0 equiv) was then added to the reaction mixture over 1 hour and the internal temperature was kept between 5 °C and 10 °C during the addition of triethylamine. The dark brown solution thus obtained was stirred for 12 h at ambient temperature (approximately 20 °C) and then chilled to around 10 °C. With vigorous stirring, 18 L of the saturated sodium bicarbonate (NaHCO_3) aqueous solution and 36 L of water were sequentially added to the chilled reaction mixture and the internal temperature was kept under 15 °C. The precipitation (filter cake) thus obtained was collected by filtration. The aqueous phase was then saturated with 12 kg of solid sodium chloride (NaCl) and extracted with EtOAc (2 x 18 L). The combined organic layer was washed with saturated sodium bicarbonate (NaHCO_3) aqueous solution (18 L), and water (2 x 18 L) in sequence. The filter cake collected was then dissolved back in the organic phase and the resulting dark brown solution was washed with water (2 x 18 L) before being concentrated under reduced pressure (40 – 50 °C, 30 mm Hg) to afford approximately 5.0 kg of the crude desired product (**14**) as a viscous brown oil. The crude product obtained above was then dissolved in EtOH (8.15 L) at 50 °C and the resulting solution was treated with water (16.3 L) over 30 minutes at around 50 °C. The brown

solution was seeded before being gradually cooled to ambient temperature (approximately 20 °C) over 3 hours with stirring and stirred at ambient temperature for 12 h. The solids were collected by filtration, washed with a mixture of EtOH and water (EtOH : H₂O = 1 : 20, 2 L) and dried under reduced pressure (50 mmHg) at approximately 60 °C for 24 h to afford (3-fluoro-2-(trifluoromethyl)pyridin-4-yl)(1,4-dioxo-8-azaspiro[4,5]decan-8-yl)methanone (**14**, 3.98 kg, 4.797 kg theoretical, 83.0%) as a white solid. For **14**: ¹H NMR (300 MHz, DMSO-*d*₆) δ 8.64 (d, ³J_{HH} = 4.68 Hz, 1H, NCH in pyridine), 7.92 (dd, ³J_{HH} = 4.68 Hz, ⁴J_{HF} = 4.68 Hz, 1H, NCCH in pyridine), 3.87 - 3.91 (m, 4H, OCH₂CH₂O), 3.70 (br s, 2H, one of NCH₂ in piperidine ring, one of another NCH₂ in piperidine ring, both in axial position), 3.26 (t, ³J_{HH} = 5.86 Hz, 2H, one of NCH₂ in piperidine ring, one of another NCH₂ in piperidine ring, both in equatorial position), 1.67 (d, ³J_{HH} = 5.86 Hz, 2H, one of NCCH₂ in piperidine ring, one of another NCCH₂ in piperidine ring, both in equatorial position), 1.58 (br s, 2H, one of NCCH₂ in piperidine ring, one of another NCCH₂ in piperidine ring, both in axial position) ppm; ¹³C NMR (75 MHz, DMSO-*d*₆) δ 161.03 (N-C=O), 151.16 (d, ¹J_{CF} = 266.03 Hz, C-F), 146.85 (d, ⁴J_{CF} = 4.32 Hz, NCH in pyridine), 135.24 (d, ²J_{CF} = 11.51 Hz, C-C=O), 135.02 (quartet, ²J_{CF} = 34.57 Hz, NCCF₃), 128.24 (d, ⁴J_{CF} = 7.48 Hz, NCCH in pyridine), 119.43 (d × quartet, ¹J_{CF} = 274.38 Hz, ³J_{CF} = 4.89 Hz, CF₃), 106.74 (OCO), 64.60 (OCCO), 45.34 (NC in piperidine ring), 39.62 (NC in piperidine ring), 34.79 (NCC in piperidine ring), 34.10 (NCC in piperidine ring) ppm; ¹⁹F NMR (282 MHz, DMSO-*d*₆) δ -64.69 (d, ⁴J_{FF} = 15.85 Hz, F₃C), -129.26 (d × quartet, ⁴J_{FF} = 15.85 Hz, ⁴J_{FH} = 3.96 Hz, FC) ppm; C₁₄H₁₄F₄N₂O₃ (MW, 334.27), LCMS (EI) *m/e* 335.1 (M⁺ + H).

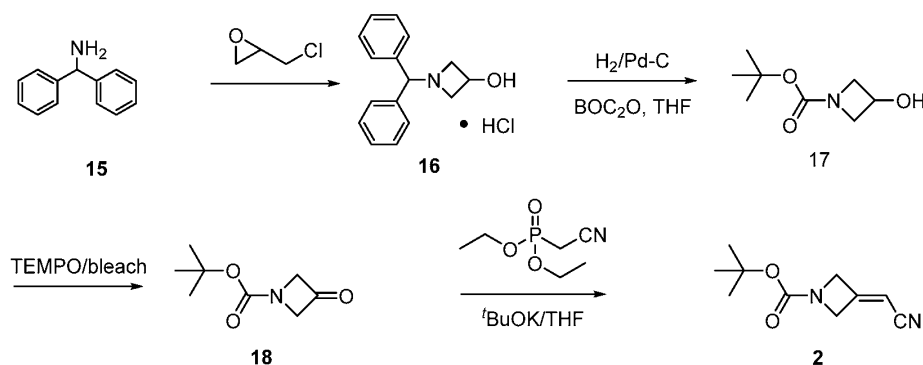
1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-one (7). In a 5 L 4-necked round bottom flask equipped with a mechanical stirrer, a thermocouple, an addition funnel and a nitrogen inlet was charged (3-fluoro-2-(trifluoromethyl)pyridin-4-yl)(1,4-dioxo-8-azaspiro[4,5]decan-8-yl)methanone (**14**, 100 g, 0.299 mol) in acetonitrile (ACN, 400 mL) at ambient temperature. The resultant solution was cooled to below 10 °C before being treated with 6.0 N aqueous hydrochloric acid (HCl) solution (450 mL, 2.70 mol, 9.0 equiv) while the internal temperature was kept at below 10 °C. The resulting reaction mixture was then gradually warmed to room

temperature and an additional amount of 6.0 N aqueous hydrochloric acid (HCl) solution (1050 mL, 6.30 mol, 21.0 equiv) was slowly introduced to the reaction mixture at ambient temperature over 8 hours *via* the addition funnel. When the reaction was complete as monitored by HPLC, the reaction mixture was then cooled to 0 °C before being treated with 30% aqueous sodium hydroxide (NaOH, 860 mL, 8.57 mmol, 28.6 equiv) while the internal temperature was kept at below 10 °C. The resulting reaction mixture was subsequently warmed to ambient temperature prior to addition of solid sodium bicarbonate (NaHCO₃, 85.0 g, 1.01 mol, 3.37 equiv) over 1 hour. The mixture was then extracted with EtOAc (2 x 1.2 L), and the combined organic phase was washed with 16% aqueous sodium chloride solution (2 x 800 mL) and concentrated to approximately 1.0 L by vacuum distillation. *n*-Heptane (2.1 L) was added to the residue, and the resulting mixture was concentrated to 1.0 L by vacuum distillation. To the concentrated mixture was added *n*-heptane (2.1 L). The resulting white slurry was then concentrated to 1.0 L by vacuum distillation. To the white slurry was then added methyl *tert*-butyl ether (MTBE, 1.94 L). The white turbid was heated to 40 °C to obtain a clear solution. The resulting solution was concentrated to about 1.0 L by vacuum distillation. The mixture was stirred at room temperature for 1 hour. The white precipitate was collected by filtration, washed with *n*-heptane (400 mL) and dried on the filter under nitrogen with pulling vacuum to afford 1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-one (7, 78.3 g, 86.8 g theoretical resulting in 90.2 % yield, and 98 % purity as measured by HPLC) as an off-white solid. For 7: ¹H NMR (300 MHz, DMSO-*d*₆) δ 8.68 (d, ³*J*_{HH} = 4.69 Hz, 1H, NCH in pyridine), 7.97 (dd, ³*J*_{HH} = 4.69 Hz, ⁴*J*_{HF} = 4.69 Hz, 1H, NCCH in pyridine), 3.92 (br s, 2H, one of NCH₂ in piperidine ring, one of another NCH₂ in piperidine ring, both in axial position), 3.54 (t, ³*J*_{HH} = 6.15 Hz, 2H, one of NCH₂ in piperidine ring, one of another NCH₂ in piperidine ring, both in equatorial position), 2.48 (t, ³*J*_{HH} = 6.44 Hz, 2H, NCCH₂), 2.34 (t, ³*J*_{HH} = 6.15 Hz, 2H, NCCH₂) ppm; ¹³C NMR (75 MHz, DMSO-*d*₆) δ 207.17 (C=O), 161.66 (N-C=O), 151.26 (d, ¹*J*_{CF} = 266.89 Hz, C-F), 146.90 (d, ⁴*J*_{CF} = 6.05 Hz, NCH in pyridine), 135.56 (C-C=O), 134.78 -135.56 (m, NCCF₃), 128.27 (d, ³*J*_{CF} = 7.19 Hz, NCCH in pyridine), 119.52 (d× quartet, ¹*J*_{CF} = 274.38 Hz, ³*J*_{CF} = 4.89 Hz, CF₃), 45.10 (NC in piperidine ring) ppm, one carbon (NCC in piperidine ring) missing due to overlap with (CD₃)₂SO; ¹⁹F NMR (282 MHz, DMSO-

d_6) δ -64.58 (d, $^4J_{FF} = 15.85$ Hz, F_3C), -128.90 (d $\times$ quartet, $^4J_{FF} = 15.85$ Hz, $^4J_{FH} = 4.05$ Hz, FC) ppm; $C_{12}H_{10}F_4N_2O_2$ (MW, 290.21), LCMS (EI) m/e 291.1 ($M^+ + H$).

Example 4. Synthesis of *tert*-Butyl 3-(cyanomethylene)azetidine-1-carboxylate

5 Scheme IV



1-Benzhydrylazetidin-3-ol hydrochloride (16). A solution of diphenylmethanamine (2737 g, 15.0 mol, 1.04 equiv) in methanol (MeOH, 6 L) was treated with 2-(chloromethyl)oxirane (1330 g, 14.5 mol) from an addition funnel at ambient temperature. During the initial addition a slight endotherm was noticed. The resulting reaction mixture was stirred at room temperature for 3 days before being warmed to reflux for an additional 3 days. When TLC showed that the reaction was deemed complete, the reaction mixture was first cooled down to room temperature and then to 0 – 5 °C in an ice bath. The solids were collected by filtration and washed with acetone (4 L) to give the first crop of the crude desired product (1516 g). The filtrate was concentrated under reduced pressure and the resulting semisolid was diluted with acetone (1 L). This solid was then collected by filtration to give the second crop of the crude desired product (221 g). The crude product, 1-benzhydrylazetidin-3-ol hydrochloride (1737 g, 3998.7 g theoretical, 43.4 % yield), was found to be sufficiently pure to be used in the subsequent reaction without further purification.

1H NMR (300 MHz, $DMSO-d_6$) δ 12.28 (br. d, 1H), 7.7 (m, 5H), 7.49 (m, 5H), 6.38 (d, 1H), 4.72 (br. s, 1H), 4.46 (m, 1H), 4.12 (m, 2H), 3.85 (m, 2H) ppm; $C_{16}H_{18}ClNO$ (MW 275.77; $C_{16}H_{17}NO$ for free base, MW, 239.31), LCMS (EI) m/e 240 ($M^+ + H$).

***tert*-Butyl 3-hydroxyazetidine-1-carboxylate (17).** A suspension of 1-benzhydrylazetidin-3-ol hydrochloride (625 g, 2.27 mol) in a 10 % solution of aqueous sodium carbonate (Na_2CO_3 , 5 L) and dichloromethane (CH_2Cl_2 , 5 L) was stirred at room temperature until all solids were dissolved. The two layers were separated, and the aqueous layer was extracted with dichloromethane (CH_2Cl_2 , 2 L). The combined organics extracts were dried over sodium sulfate (Na_2SO_4) and concentrated under reduced pressure. The resulting crude 1-benzhydrylazetidin-3-ol free base was then dissolved in THF (6 L) and the solution was placed into a large Parr bomb. Di-*tert*-butyl dicarbonate (BOC_2O , 545 g, 2.5 mol, 1.1 equiv) and 20 % palladium (Pd) on carbon (125 g, 50 % wet) were added to the Parr bomb. The vessel was charged to 30 psi with hydrogen gas (H_2) and stirred under steady hydrogen atmosphere (vessel was recharged three times to maintain the pressure at 30 psi) at room temperature for 18 h. When HPLC showed that the reaction was complete (no more hydrogen was taken up), the reaction mixture was filtered through a Celite pad and the Celite pad was washed with THF (4 L). The filtrates were concentrated under reduced pressure to remove the solvent and the residue was loaded onto a Biotage 150 column with a minimum amount of dichloromethane (CH_2Cl_2). The column was eluted with 20 – 50 % ethyl acetate in *n*-heptane and the fractions containing the pure desired product, *tert*-butyl 3-hydroxyazetidine-1-carboxylate, were collected and combined. The solvents were removed under reduced pressure to afford *tert*-butyl 3-hydroxyazetidine-1-carboxylate (357 g, 393.2 g theoretical, 90.8 % yield) as a colorless oil, which solidified upon standing at ambient temperature in vacuum. ^1H NMR (300 MHz, CDCl_3), δ 4.56 (m, 1H), 4.13 (m, 2H), 3.81 (m, 2H), 1.43 (s, 9H) ppm.

***tert*-Butyl 3-oxoazetidine-1-carboxylate (18).** A solution of *tert*-butyl 3-hydroxyazetidine-1-carboxylate (50 g, 289 mmol) in ethyl acetate (400 mL) was cooled to 0 °C. The resulting solution was then treated with solid TEMPO (0.5 g, 3.2 mmol, 0.011 equiv) and a solution of potassium bromide (KBr, 3.9 g, 33.2 mmol, 0.115 equiv) in water (60 mL) at 0 – 5 °C. While keeping the reaction temperature between 0 - 5 °C, a solution of saturated aqueous sodium bicarbonate (NaHCO_3 , 450 mL) and an aqueous sodium hypochlorite solution (NaClO , 10 - 13 % available

chlorine, 450 mL) were added. Once the solution of sodium hypochlorite was added, the color of the reaction mixture was changed immediately. When additional amount of sodium hypochlorite solution was added, the color of the reaction mixture was gradually faded. When TLC showed that all of the starting material was consumed, the color of the reaction mixture was no longer changed. The reaction mixture was then diluted with ethyl acetate (EtOAc, 500 mL) and two layers were separated. The organic layer was washed with water (500 mL) and the saturated aqueous sodium chloride solution (500 mL) and dried over sodium sulfate (Na₂SO₄). The solvent was then removed under reduced pressure to give the crude product, *tert*-butyl 3-oxoazetidine-1-carboxylate (48 g, 49.47 g theoretical, 97% yield), which was found to be sufficiently pure and was used directly in the subsequent reaction without further purification. ¹HNMR (CDCl₃, 300 MHz) δ 4.65 (s, 4H), 1.42 (s, 9H) ppm.

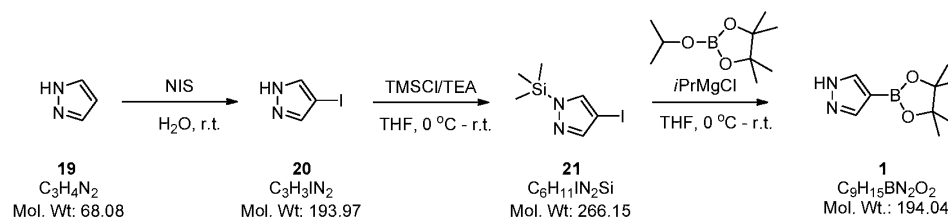
***tert*-Butyl 3-(cyanomethylene)azetidine-1-carboxylate (2).** Diethyl cyanomethyl phosphate (745 g, 4.20 mol, 1.20 equiv) and anhydrous tetrahydrofuran (THF, 9 L) were added to a four-neck flask equipped with a thermowell, an addition funnel and the nitrogen protection tube at room temperature. The solution was cooled with an ice-methanol bath to - 14 °C and a 1.0 M solution of potassium *tert*-butoxide (*t*-BuOK) in anhydrous tetrahydrofuran (THF, 3.85 L, 3.85 mol, 1.1 equiv) was added over 20 min keeping the reaction temperature below - 5 °C. The resulting reaction mixture was stirred for 3 h at - 10 °C and a solution of 1-*tert*-butoxycarbonyl-3-azetidinone (600 g, 3.50 mol) in anhydrous tetrahydrofuran (THF, 2 L) was added over 2 h keeping the internal temperature below - 5 °C. The reaction mixture was stirred at - 5 to - 10 °C over 1 h and then slowly warmed up to room temperature and stirred at room temperature for overnight. The reaction mixture was then diluted with water (4.5 L) and saturated aqueous sodium chloride solution (NaCl, 4.5 L) and extracted with ethyl acetate (EtOAc, 2 x 9 L). The combined organic layers were washed with brine (6 L) and dried over anhydrous sodium sulfate (Na₂SO₄). The solvent was removed under reduced pressure and the residue was diluted with dichloromethane (CH₂Cl₂, 4 L) before being absorbed onto silica gel (SiO₂, 1.5 Kg). The crude product, which was absorbed on silica gel, was purified by flash column chromatography (SiO₂, 3.5 Kg, 0 – 25% EtOAc/hexanes gradient elution) to afford

tert-butyl 3-(cyanomethylene)azetidine-1-carboxylate (**2**, 414.7 g, 679.8 g theoretical, 61% yield) as a white solid. For **2**: ^1H NMR (300MHz, CDCl_3) δ 5.40 (m, 1H), 4.70 (m, 2H), 4.61 (m, 2H), 1.46 (s, 9H) ppm; $\text{C}_{10}\text{H}_{14}\text{N}_2\text{O}_2$ (MW, 194.23), LCMS (EI) m/e 217 ($\text{M}^+ + \text{Na}$).

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Example 5. Synthesis of 4-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)-1H-pyrazole

Scheme V



10

4-Iodopyrazole (20). A flask equipped with a nitrogen inlet, an addition funnel, a thermowell, and a mechanical stirrer was charged with pyrazole (**1**, 450 g, 6.62 mol) and tetrahydrofuran (THF, 5 L) at ambient temperature. The mixture was then cooled to 10 °C and *N*-iodosuccinimide (NIS, 1490 g, 6.62 mol, 1.0 equiv) was added to the mixture in portions as a solid at approximately 10 °C. The resulting reaction mixture was then stirred at ambient temperature for 1 hour (longer reaction times may be necessary depending on ambient temperature). The mixture was then filtered and the THF was removed under reduced pressure. The residue was suspended in ethyl acetate (6 L) and insoluble materials were filtered. The dark filtrate was sequentially washed with saturated aqueous sodium thiosulfate solution (2 x 3 L) (organic layer lightens to a pale yellow), water (2 x 3 L), and brine (2 L). The resulting organic layer was then dried over sodium sulfate, filtered, and concentrated under reduced pressure to afford 4-iodopyrazole (1138 g, 1284.1 g theoretical, 88.6%) as a white to pale yellow solid after being dried in a vacuum oven at approximately 30 °C overnight. ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 13.17 (bs, 1H), 7.93 (bs, 1H), 7.55 (bs, 1H) ppm; $\text{C}_3\text{H}_3\text{IN}_2$ (MW, 193.97), LCMS (EI) m/e 195 ($\text{M}^+ + \text{H}$).

25

1-Trimethylsilyl-4-iodopyrazole (21). To a flask equipped with a reflux condenser, a nitrogen inlet, mechanical stirrer, and a thermowell was charged 4-iodopyrazole (200 g, 1.03 mol) and THF (2 L) at ambient temperature. To this solution was added triethylamine (TEA, 158 mL, 1.13 mol, 1.1 equiv) and the resulting solution was
5 cooled to 0 °C in an ice-brine bath. To this solution was added chlorotrimethylsilane (TMS-Cl, 137 mL, 1.08 mol, 1.05 equiv) with vigorous stirring allowing the temperature to reach 18 °C. (The reaction becomes very thick and difficult to stir, but becomes manageable after over time). When the exothermic process had subsided, the cold bath was removed and the reaction was warmed to room temperature. The
10 reaction was followed by GC and was found to be deemed complete after about 1 hour (sampling of reaction must be done out of air and diluted with dry solvent to prevent TMS hydrolysis). The reaction mixture was then diluted with *n*-heptane (2 L) before being filtered under nitrogen. The solvent was removed from the filtrate under reduced pressure venting the rotovap with nitrogen. The residual oil was diluted with
15 *n*-heptane (1 L) and re-concentrated. If the solids formed upon adding the *n*-heptane, a second filtration was necessary. The residue was then distilled under the reduced pressure (70 - 90 °C at about 0.5 Torr) using a Kugelrohr to afford 1-trimethylsilyl-4-iodopyrazole (263 g, 274.1 g theoretical, 96%) as a colorless oil. This material must be kept under nitrogen at all times since the TMS group rapidly hydrolyzes.
20 Subsequently, it was found that 1-trimethylsilyl-4-iodopyrazole can be prepared by heating the iodopyrazole with 2 equivalents of hexamethyldisilazane for 1 hr.

4-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)-1H-pyrazole (1). A flask equipped with a mechanical stirrer, a nitrogen inlet, an addition funnel and a
25 thermowell was charged with 1-trimethylsilyl-4-iodopyrazole (225.1 g, 0.85 mol) and THF (2200 mL) at ambient temperature. This mixture was cooled to approximately -6 °C in an ice/salt/brine bath before a solution of isopropyl magnesium chloride in THF (2 M solution in THF, 510 mL, 1.02 mol, 1.2 equiv) was added at a rate such that the internal temperature did not exceed 0 °C. The extent of metal/halogen
30 exchange was monitored by GC and was found complete after about 10 min. To the orange brown solution was then added 2-isopropoxy-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (isopropylpinacolborate, 347 mL, 1.7 mol, 2.0 equiv) slowly at first

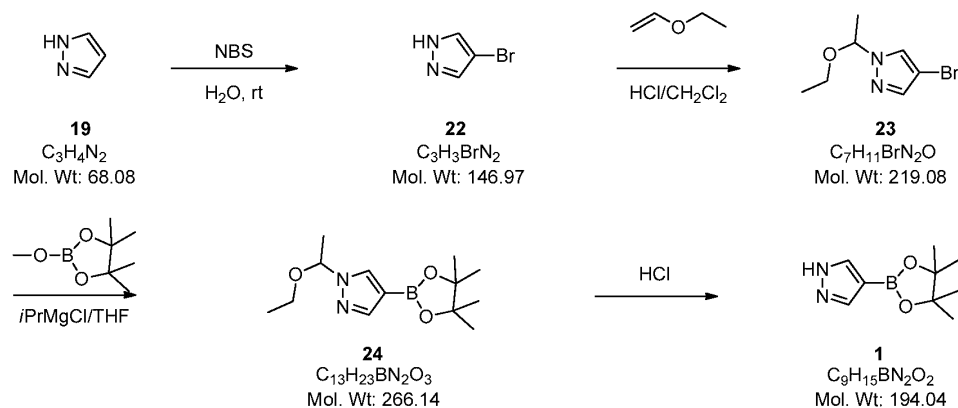
keeping the temperature below 0 °C and then fairly rapidly after about half of the compound was added allowing the temperature to reach 5 °C (the reaction becomes quite thick and then thins out slowly). The reaction is then stirred at 0 °C for 10 min before being warmed to ambient temperature over 1 h and stirred at ambient

5 temperature for an additional 1 h. The reaction mixture was cooled to approximately 6 °C and the saturated aqueous ammonium chloride solution (NH₄Cl, 2.2 L) was added with a temperature increase to 25 °C. The mixture was stirred for 5 minutes before being diluted with toluene (10 L). The layers were separated (a large amount of solid is present in the aqueous layer) and the organic layer was sequentially washed

10 with water (6 x 2.2 L) and brine (2 x 2.2 L) before being dried over sodium sulfate (Na₂SO₄). The drying reagent, sodium sulfate (Na₂SO₄), was removed by filtration and the solution was concentrated under reduced pressure. Residual toluene was co-evaporated with *n*-heptane to afford 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1*H*-pyrazole (**1**, 90.3 g, 164.9 g theoretical, 54.8%) as a white solid. For **1**: ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.08 (bs, 1H), 7.94 (s, 1H), 7.62 (s, 1H), 1.23 (s, 12H) ppm; C₉H₁₅BN₂O₂ (MW, 194.04), LCMS (EI) *m/e* 195 (M⁺ + H).

Example 6. Alternative Synthesis of 4-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)-1*H*-pyrazole

Scheme VI



4-Bromopyrazole (22). Pyrazole (**19**, 34.0 g, 0.5 mol) and NBS (89.0 g, 0.5 mol, 1.0 equiv) were suspended in water (625 ml) at ambient temperature. The resulting suspension was stirred at ambient temperature for overnight. The reaction mixture

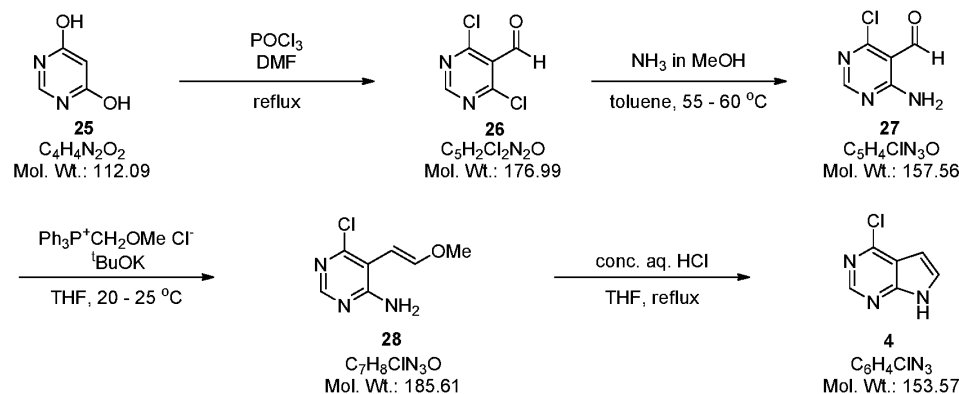
was then extracted with EtOAc (2 x 100 mL). The combined EtOAc extracts were washed with aqueous Na₂S₂O₃ and brine, dried over Na₂SO₄, and concentrated under reduced pressure to afford crude 4-bromopyrazole (72.0 g, 73.5 g theoretical, 98% yield) as white solids (GC purity: >98%), which was directly used in the subsequent reaction without further purification.

4-Bromo-1-(ethoxyethyl)-1H-pyrazole (23). To a solution of 4-bromopyrazole (70.0 g, 0.476 mol) in CH₂Cl₂ (600 mL) was added a solution of 3.1 M HCl in dioxane (4 mL) and ethyl vinyl ether (41 g, 0.569 mol, 1.2 equiv) at ambient temperature. The resulting reaction mixture was stirred at ambient temperature for 3 h. The reaction was quenched with aqueous NaHCO₃ and the two layers were separated. The organic layer was washed with water, dried over Na₂SO₄, and concentrated under reduced pressure to dryness to afford 4-bromo-1-(ethoxyethyl)-1H-pyrazole (113 g, 104.3 g theoretical, 97% yield) as an oil (GC purity: 89%), which was directly used in the subsequent reaction without further purification.

1-(Ethoxyethyl)-4-(4,4,5,5-tetramethyl[1,3,2]dioxaborolan-2-yl)-1H-pyrazole (24). To a 100 mL solution of *i*PrMgCl.LiCl (50 mmol, 1.8 equiv) in THF was added 4-bromo-1-(ethoxyethyl)-1H-pyrazole (6.15 g, 28 mmol) at ambient temperature. The resulting reaction mixture was stirred at ambient temperature for 12 h and then cooled to -20 °C. Methoxy pinacolborate (10.6 g, 67 mmol, 2.4 equiv) was then added to the reaction mixture at -20 °C. The resulting mixture was stirred at 0 - 10 °C for 1 h. Aqueous NH₄Cl was added to quench the reaction. The mixture was then extracted with petroleum ether (PE). The combined PE extracts were washed with saturated NaHCO₃, dried over Na₂SO₄ and concentrated under reduced pressure. The crude product was crystallized in PE to afford 1-(ethoxyethyl)-4-(4,4,5,5-tetramethyl[1,3,2]dioxaborolan-2-yl)-1H-pyrazole (**24**, 4.2 g, 7.45 g theoretical, 56.4% yield) as a white to off-white solid (GC purity: 99%). For **24**: ¹H NMR (DMSO-*d*₆, 400 MHz) δ 8.09 (s, 1H), 8.58 (s, 1H), 7.62 (s, 1H), 5.55 (q, 1H, *J* = 6.1 Hz), 3.37 (dq, 1H, *J* = 7.1, 9.6 Hz), 3.12 (dq, 1H, *J* = 7.0, 9.7 Hz), 1.56 (d, 3H, *J* = 6.0 Hz), 1.24 (s, 12H), 1.00 (t, 3H, *J* = 7.0 Hz) ppm; C₁₃H₂₃BN₂O₃ (MW, 266.14), LCMS (EI) *m/e* 267 (M⁺ + H).

4-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)-1H-pyrazole (1). To a mixture of 2,3-dimethylbutane-2,3-diol (25.0 kg, 211.6 mol) and 1-(1-ethoxyethyl)-4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1H-pyrazole (**24**, 55.0 kg, 206.7 mol) in 1,2-dichloroethane (750 kg) was slowly added a solution of HCl in MTBE (25.0 kg, 20 – 30% of HCl) at 0 – 5 °C. The resulting reaction mixture was then stirred at 10 – 20 °C for 3 – 5 hours. After the selective deprotection reaction was complete as monitored by HPLC (**1**: below 1%), the reaction mixture was degassed and refilled with nitrogen before being cooled to – 15 °C. The cooled reaction mixture was then added triethylamine (TEA, 30.0 kg, 296.5 mol) to adjust pH to 7 – 8. The mixture was then gradually warmed to ambient temperature before being treated with water (150 kg). The two phases were separated and the organic layer was washed with brine (60 kg) and dried over sodium sulfate (Na₂SO₄). The drying reagent, sodium sulfate (Na₂SO₄), was removed by filtration and the resulting solution was concentrated under reduced pressure at 40 – 50 °C to a thick oil. The residue was warmed to 60 – 70 °C and diluted with petroleum ether (100 kg) at the same temperature. The resulting mixture was then gradually cooled to ambient temperature and subsequently to – 5 °C and stirred at the same temperature for 3 hours. The solids were collected by centrifugation and dried at 50 – 60 °C under vacuum to afford the crude desired product (**1**, 33.75 kg, 40.11 kg theoretical, 84.1%). The crude desired product was then suspended in 1,2-dichloroethane (30 kg) and the resulting mixture was heated to reflux until a clear solution was formed. To the hot solution was then added petroleum ether (150 kg) at the same temperature. The resulting mixture was then gradually cooled to ambient temperature and subsequently to – 5 °C and stirred and the same temperature for 3 hours. The solids were collected by centrifugation and dried under vacuum at 50 – 60 °C to afford 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1H-pyrazole (**1**, 31.0 kg, 40.11 kg theoretical, 77.3%) as an off-white solid, which is identical in every comparable aspect to the material synthesized by the synthetic method as described above in Example 5.

30

Example 7. Synthesis of 4-Chloro-7H-[pyrrolo[2,3-d]pyrimidine**Scheme VII**

- 5 **4,6-Dichloropyrimidine-5-carbaldehyde (26).** In a 5 L 4-neck flask equipped with a mechanical stirrer, an addition funnel, a condenser, a thermocouple, and a N₂ sweep into an aqueous NaOH scrubbing solution, phosphorous oxychloride (POCl₃, 1 L, 10.572 mol, 4.82 equiv) was charged and cooled in an ice/salt bath. *N,N*-
- 10 Dimethylformamide (DMF, 320 mL, 4.138 mol, 1.85 equiv) was then added dropwise to the flask at 0 ± 2 °C. After addition of approximately 100 mL of DMF over approximately 0.5 h, crystallization occurred and the reaction temperature was increased from 0 to 10 °C. Addition was stopped and the mixture was allowed to re-cool to approximately 2 °C. The remaining DMF was added over 2.5 h at below 8 °C. The suspension became very thick making stirring difficult. When addition of DMF
- 15 was complete, the mixture was stirred at 3 – 5 °C for 0.5 h. 4,6-Dihydroxypyrimidine (250 g, 2.232 mol) was added portion wise as a solid. After about one third of 4,6-dihydroxypyrimidine was added, the reaction mixture became more mobile, and a slow exothermic phenomena occurred with the reaction temperature increasing to approximately 12 °C over 0.5 h. The remaining 4,6-dihydroxypyrimidine was added
- 20 portion wise over 0.25 h with the reaction temperature increasing from 12 to 27 °C. The reaction temperature was maintained at 25 – 27 °C with intermittent cooling during which time the yellow suspension became thinner, then thicker once again. After the exothermic phenomenon subsided in about 1 h, the reaction mixture was heated slowly. At about 55 °C the reaction mixture became extremely thick and the

second mild exothermic phenomenon was occurred. The heating mantle was removed while the reaction temperature continued to increase to about 63 °C and remained at this temperature for several minutes before dropping. Heating of the mixture was resumed until gentle reflux (about 100°C) was attained. At about 95 °C a steady,
5 fairly rapid evolution of HCl gas began and the reaction mixture gradually thinned and darkened. After about 0.5 h, a clear brown solution developed with the reflux temperature slowly increasing to 115 °C over 1.25 h. After a total of 2.5 h at reflux, the reaction mixture was cooled to ambient temperature and stirred overnight at ambient temperature. Excess amount of POCl₃ (as much as possible) was removed
10 under reduced pressure (bath temperature 45 – 50 °C). The thick residual brown oil was poured very slowly into cold H₂O (5 L) in a 20 L separation funnel, adding ice as needed to maintain the aqueous mixture near room temperature. The aqueous mixture was extracted with EtOAc (2 x 3 L followed by 1 x 2 L). The combined EtOAc extracts were washed with H₂O (2 x 2.5 L), saturated NaHCO₃ aqueous solution (1
15 L), brine (1 L), dried over Na₂SO₄, filtered, and concentrated under reduced pressure (bath temperature at 35°C) to afford the crude 4,6-dichloropyrimidine-5-carbaldehyde (270 g, 395 g theoretical, 68.4%) as yellow-orange solids. A 20 g portion of this crude material was purified by Kugelrohr distillation (oven temperature at 90 – 100 °C, 225 mTorr) to give 15.3 g of pure 4,6-dichloropyrimidine-5-carbaldehyde as a white solid
20 that turned yellow on standing at room temperature. ¹H NMR (300 MHz, CDCl₃) δ 10.46 (s, 1H), 8.89 (s, 1H) ppm.

4-Amino-6-chloropyrimidine-5-carbaldehyde (27). A solution of 7 M NH₃ in MeOH (265 mL, 1.855 mol, 2.0 equiv) was added over 1.25 h to a solution of 4,6-
25 dichloropyrimidine-5-carbaldehyde (163.7 g, 0.9301 mol) in toluene (3 L) at ambient temperature. The reaction temperature slowly increased from 20 to 26 °C and a yellow suspension formed. Mild cooling was applied to maintain the reaction temperature at below 26 °C. The suspension was stirred at ambient temperature for 3.5 h before the solids were collected by filtration. The solids were washed with
30 EtOAc (1 L). The filtrate was concentrated under reduced pressure, and the solids were triturated with toluene and *n*-heptane (2:1 v/v, 600 mL), filtered and dried to give 71.1 g of 4-amino-6-chloropyrimidine-5-carbaldehyde as a yellow solid. The

original solid filtered from the reaction mixture contained additional amount of 4-amino-6-chloropyrimidine-5-carbaldehyde. The product was extracted from the filtered solid by stirring in EtOAc (1.25 L) for 1.5 h, filtering, then stirring in THF (750 mL) for 1 h and again filtering. Both EtOAc and THF filtrates were concentrated under reduced pressure, and the resulting solids were triturated with toluene and *n*-heptane (2:1 v/v, 450 mL), filtered and dried to give an additional 44.1 g of 4-amino-6-chloropyrimidine-5-carbaldehyde as a yellow solid. The combined yield of 4-amino-6-chloropyrimidine-5-carbaldehyde (115.2 g, 146.5 g theoretical) was 78.6%. ¹HNMR (300 MHz, DMSO-*d*₆) δ 10.23 (s, 1H), 8.71 (bs, 1H), 8.55 (bs, 1H), 8.39 (s, 1H) ppm; C₅H₄ClN₃O (MW, 157.56), LCMS (EI) *m/e* 158 (M⁺ + H).

6-Chloro-5-(2-methoxyvinyl)pyrimidin-4-ylamine (28). A suspension of (methoxymethyl)triphenylphosphonium chloride (276.0 g, 0.807 mol, 1.1 equiv) in THF (1.5 L) was cooled in an ice/salt bath to -2 °C and 1 M potassium *tert*-butoxide (KO^tBu) in THF (807 mL, 0.807 mol, 1.1 equiv) was added over 1.5 h at -2 to -3 °C. The deep red-orange mixture was stirred at -2 to -3 °C for 1 h. 4-Amino-6-chloropyrimidine-5-carbaldehyde (115.2 g, 0.7338 mol, 1.0 equiv) was then added portion wise to the reaction mixture as a solid form using THF (200 mL) to rinse the container and funnel. During the addition the reaction temperature increased from -3 to 13 °C and a brown color developed. When the reaction temperature dropped to 10 °C, the cooling bath was removed and the reaction mixture was allowed to warm to ambient temperature and stirred at ambient temperature for 42 h. The reaction mixture was cooled to -2 °C before being quenched by the slow addition of saturated NH₄Cl aqueous solution (750 mL). The mixture was concentrated under reduced pressure to remove most of the THF. The residue was partitioned between EtOAc (3 L) and H₂O (1 L). The organic phase was filtered to remove insoluble material at the interface, then extracted with 2 N HCl (4 x 250 mL) followed by 3 N HCl (2 x 250 mL). The combined HCl extracts were back-extracted with EtOAc (500 mL) then filtered through Celite to remove insoluble material. The filtrate was cooled in an ice/brine bath, adjusted to pH 8 with a 6 N aqueous NaOH solution and extracted with EtOAc (3 x 1 L). The combined EtOAc extracts were washed with brine (1 L), dried over Na₂SO₄, stirred with charcoal (10 g) and silica gel (10 g) for 1 h. The mixture was

filtered through Celite, washing the Celite pad with EtOAc (1 L). The filtrate was concentrated, co-evaporating residual EtOAc with *n*-heptane (500 mL). The resulting tan solid was pumped under high vacuum for 2 h to afford crude 6-chloro-5-(2-methoxyvinyl)pyrimidin-4-ylamine (72.3 g, 136.2 g theoretical, 53.1%). The crude
5 desired product was used in the following reaction without further purification. A sample of crude product (2.3 g) was purified by silica gel column chromatography on, eluting with 0 – 35% EtOAc/*n*-heptane to give 1.7 g of pure 6-chloro-5-(2-methoxyvinyl)pyrimidin-4-ylamine as a white solid, which was found to be a 1 to 2 mixture of *E/Z* isomers. ¹H NMR (300 MHz, DMSO-*d*₆) for *E*-isomer: δ 8.02 (s, 1H),
10 7.08 (bs, 2H), 6.92 (d, 1H, *J* = 13.1), 5.35 (d, 1H, *J* = 13.0 Hz), 3.68 (s, 3H) ppm and for *Z*-isomer: δ 8.06 (s, 1H), 7.08 (bs, 2H), 6.37 (d, 1H, *J* = 6.8 Hz), 5.02 (d, 1H, *J* = 6.7 Hz), 3.69 (s, 3H) ppm; C₇H₈ClN₃O (MW, 185.61), LCMS (EI) *m/e* 186/188 (M⁺ + H).

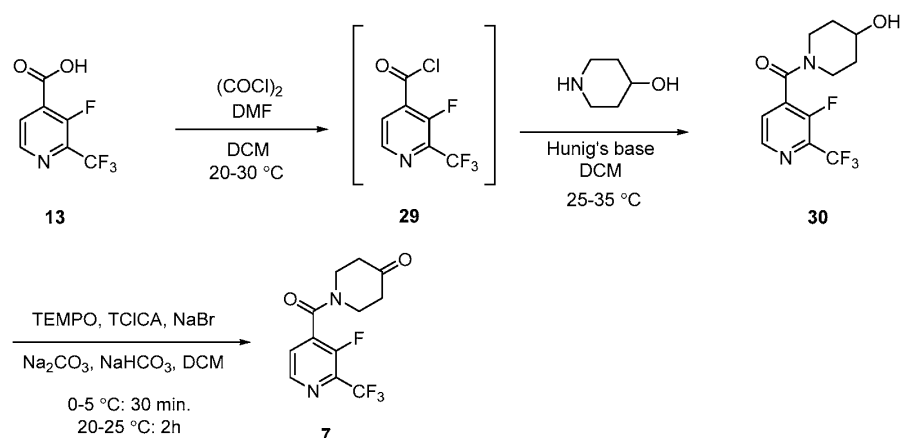
15 **4-Chloro-7H-[pyrrolo[2,3-*d*]pyrimidine (4).** Concentrated HCl (5 mL) was added to a solution of crude 6-chloro-5-(2-methoxyvinyl)pyrimidin-4-ylamine (70.0 g, 0.3784 mol) in THF (700 mL) and the resulting reaction mixture was heated to reflux for 7.5 h. On warming a light suspension was formed that gradually re-dissolved. When the
20 reaction was deemed complete as monitored by HPLC, the reaction mixture was cooled to ambient temperature and stirred at ambient temperature for overnight. Solid NaHCO₃ (15 g) was added to the reaction mixture and the resulting mixture was stirred at ambient temperature for 1 h. Charcoal (7 g), silica gel (7 g) and Na₂SO₄ (20 g) were added and the mixture was heated to 40 °C for 1 h. The mixture was then cooled to ambient temperature and filtered through Celite, washing the Celite pad
25 with THF (1 L). The filtrate was concentrated under reduced pressure and the resulting solid was dried under reduced pressure to afford crude 4-chloro-7H-[pyrrolo[2,3-*d*]pyrimidine (**4**, 58.1 g, 58.1 g theoretical, 100%) as a yellow-brown solid. This crude desired product was dissolved in EtOAc (1 L) at 50 – 55 °C and treated with activated charcoal (3 g). The mixture was filtered while warm through
30 Celite and the Celite pad was washed with warm EtOAc (250 mL). The filtrate was concentrated to about 500 mL and the suspension was allowed to stand at ambient temperature for overnight. The suspension was subsequently cooled to 0 – 5 °C for 2

h before the solids were collected by filtration. The solids were dried to afford pure 4-chloro-7*H*-[pyrrolo[2,3-*d*]pyrimidine (**4**, 54.5 g, 58.1 g theoretical, 94%) as yellow-brown crystals. ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.58 (bs, 1H), 8.58 (s, 1H), 7.69 (d, 1H, *J* = 3.5 Hz), 6.59 (d, 1H, *J* = 3.5 Hz) ppm; LCMS (EI) *m/e* 154/156 (*M*⁺ + H).

5

Example 8. Alternative Synthesis of 1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-one (7**)**

Scheme VIII



10

Step 1: (3-fluoro-2-(trifluoromethyl)pyridin-4-yl)(4-hydroxypiperidin-1-yl)methanone (30**).** To a solution of 3-fluoro-2-(trifluoromethyl)isonicotinic acid (**13**) (54.01 g, 258.3 mmol) in dichloromethane (270 mL) was added *N,N*-dimethylformamide (0.34g, 4.65 mmol) at room temperature. To this solution was added oxalyl chloride (34.41 g, 271.2 mmol) in dichloromethane (81 mL) over 30 minutes via addition funnel while the internal temperature was maintained at 15 to 25 °C. The addition funnel was rinsed with dichloromethane (27 mL). The mixture was stirred at room temperature for two hours to give a brown solution. The dichloromethane was distilled at a temperature of 30 °C under reduced pressure. The residue was dissolved in dichloromethane (270 mL) and the solvent was distilled off under reduced pressure at 30 °C. The resulting residue was dissolved in dichloromethane (270 mL) to give a dichloromethane solution of the 3-fluoro-2-(trifluoromethyl)isonicotinoyl chloride (**29**). To another flask was added 4-hydroxypiperidine (33.18 g, 328 mmol), dichloromethane (270 mL), and *N,N*-

20

diisopropylethylamine (108 mL, 619.9 mmol). The mixture was heated to 33 °C to form a solution. The mixture was cooled to room temperature. To the solution was added the the 3-fluoro-2-(trifluoromethyl)isonicotinoyl chloride (29) in dichloromethane at a temperature of 25 to 35 °C. After the addition, the mixture was stirred at 25 to 35 °C for another hour. To a container was added water (430 mL) and 37% hydrochloric acid (62.4 g (633 mmol). The diluted hydrochloric acid was added to the reaction mixture at internal temperature below 25 °C. After stirred the mixture for 30 minutes, the organic phase was collected by separation. The organic phase was washed with 9.5% brine (210 g). The aqueous phases were combined and extracted with dichloromethane (430 mL). The organic phases were combined and washed with 4.5% brine (210 ml) and water (215 mL). Dichloromethane was removed by solvent swap into 2-methoxy-2-methylpropane (TBME). The residue in TBME (135 mL) was heated to 60 °C for 1 hour. The mixture was gradually cooled to 0 °C to crystallize (3-fluoro-2-(trifluoromethyl)pyridin-4-yl)(4-hydroxypiperidin-1-yl)methanone. The mixture was filtered and the wet cake was washed with TBME (27 mL). The solids were dried at 50 °C to give (3-fluoro-2-(trifluoromethyl)pyridin-4-yl)(4-hydroxypiperidin-1-yl)methanone (**30**). (69.95 g, 92%) as a light brown solids. HPLC-MS: 293.0 (M+H). ¹H NMR (400 MHz, DMSO-*d*₆) δ 3.09 – 3.99 (m, 4H); 1.37 – 1.80 (m, 4H); 3.75 (m, 1H); 4.84 (d, 1H (b)); 8.65 (d, 1H, J = 4.7Hz); 7.89 (dd, 1H, J = 4.7, J = 4.7 Hz). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 33.5, 34.3, 38.9, 44.0, 65.0, 120.6, 127.6, 134.5, 134.7, 146.2, 152.2; 160.2. C₁₂H₁₂F₄N₂O₂ (MW 292.23), LCMS (EI) *m/e* 293.0 (M⁺ + H). The purity of compound 30 via this process was determined to be greater than about 98% as measured by HPLC.

Step 2: 1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-one (7). To a flask was added (3-fluoro-2-(trifluoromethyl)pyridin-4-yl)(4-hydroxypiperidin-1-yl)methanone (**30**) (50 g, 171.1 mmol), dichloromethane (733 mL), water (766 mL), sodium bicarbonate (71.4 g, 849.7), sodium carbonate 99.1 g, 85.2 mmol), sodium bromide (1.76 g, 17.1 mmol), and 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO) (0.535 g, 3.42 mmol) at 15 to 25 °C. The mixture was cooled to 0 to 5 °C. To the mixture was added 1,3,5-trichloro-1,3,5-triazinane-4,4,6-trione (23.8 g, 102 mmol) in four portions over 10 minutes. The mixture was stirred at 0 to 5 °C for 30 minutes and

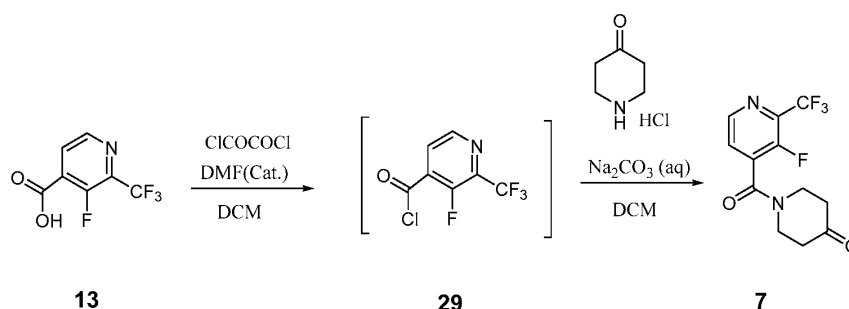
then the mixture was warmed to 20 to 25 °C in 30 minutes. After stirring the mixture at 20 to 25 °C for another hour, the reaction was quenched by addition of methanol (26.23 mL, 647.5 mmol) at 20 to 25 °C. The mixture was stirred for 20 minutes and filtered over Celite bed. The Celite (20 g) bed was washed with dichloromethane (50 mL). The organic phase was separated. The organic phase was washed sequentially with 6% brine (266 g) and water (250 mL). Activated charcoal (3.5 g) was added to the organic phase. After stirring the mixture at room temperature for 30 minutes, it was filtered over Celite (20 g) bed. The filter bed was washed with dichloromethane (50 mL). The dichloromethane was removed by solvent swap into 2-methoxy-2-methylpropane (TBME) and the residue in TBME (180 mL) was heated to 50 to 60 °C. Heptane (500 mL) was gradually added to the warm mixture (500 mL) while the internal temperature was maintained above 50 °C to crystallize the product. The mixture was gradually cooled to 10 °C and filtered. The wet cake was washed with heptane (2 x 75 mL). The solids were dried under reduced pressure to give 1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-one (7) (49.7 g, 87.9% yield) as off-white to brown solids. : ¹H NMR (300 MHz, DMSO-*d*₆) δ 8.68 (d, ³*J*_{HH} = 4.69 Hz, 1H, NCH in pyridine), 7.97 (dd, ³*J*_{HH} = 4.69 Hz, ⁴*J*_{HF} = 4.69 Hz, 1H, NCCH in pyridine), 3.92 (br s, 2H, one of NCH₂ in piperidine ring, one of another NCH₂ in piperidine ring, both in axial position), 3.54 (t, ³*J*_{HH} = 6.15 Hz, 2H, one of NCH₂ in piperidine ring, one of another NCH₂ in piperidine ring, both in equatorial position), 2.48 (t, ³*J*_{HH} = 6.44 Hz, 2H, NCCH₂), 2.34 (t, ³*J*_{HH} = 6.15 Hz, 2H, NCCH₂) ppm; ¹³C NMR (75 MHz, DMSO-*d*₆) δ 207.17 (C=O), 161.66 (N-C=O), 151.26 (d, ¹*J*_{CF} = 266.89 Hz, C-F), 146.90 (d, ⁴*J*_{CF} = 6.05 Hz, NCH in pyridine), 135.56 (C-C=O), 134.78 -135.56 (m, NCCF₃), 128.27 (d, ³*J*_{CF} = 7.19 Hz, NCCH in pyridine), 119.52 (d × quartet, ¹*J*_{CF} = 274.38 Hz, ³*J*_{CF} = 4.89 Hz, CF₃), 45.10 (NC in piperidine ring) ppm, one carbon (NCC in piperidine ring) missing due to overlap with (CD₃)₂SO; ¹⁹F NMR (282 MHz, DMSO-*d*₆) δ -64.58 (d, ⁴*J*_{FF} = 15.85 Hz, F₃C), -128.90 (d × quartet, ⁴*J*_{FF} = 15.85 Hz, ⁴*J*_{FH} = 4.05 Hz, FC) ppm; C₁₂H₁₀F₄N₂O₂ (MW, 290.21), LCMS (EI) *m/e* 291.1 (M⁺ + H). The purity of compound 7 via this process was determined to be

It is noted that the increased use of oxidizing agent (e.g., TEMPO) can result in increased impurity formation and decreased isolation yield. Longer oxidation times

can also lead to increased impurity formation and decreased isolation yield.

Example 9. Alternative Synthesis of 1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-one (7)

Scheme IX



1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-one (7). To a solution of 3-fluoro-2-(trifluoromethyl)isonicotinic acid (**13**) (20 g, 96.65 mmol) in dichloromethane (150 mL) was added *N,N*-dimethylformamide (0.13 g, 1.72 mmol) at room temperature. To this solution was added oxalyl chloride (12.75 g, 100.4 mmol) in dichloromethane (40 mL) over 30 minutes via addition funnel while the internal temperature was maintained at 15 to 25 °C. The addition funnel was rinsed with dichloromethane (10 mL). The mixture was stirred at room temperature for 2 hours to give a brown solution of intermediate 3-fluoro-2-(trifluoromethyl)isonicotinoyl chloride. To the solution was charged 4-piperidone monohydrate hydrochloride (19.1 g, 124.3 mmol). The mixture was cooled to 0 to 5 °C and an aqueous solution of sodium carbonate (20.28 g, 191.3 mmol) in water (200 mL) was added. After the addition, the mixture was warmed to room temperature and stirred for 2 hours. The organic phase was separated and sequentially washed with 6% brine (110 g) and water (100 mL). To the organic phase was added activated charcoal (1.4 g). After stirring the mixture at room temperature for 60 minutes, the mixture was filtered over Celite (5 g) bed. The filter bed was washed with dichloromethane (40 mL). The dichloromethane was removed by solvent swap into 2-methoxy-2-methylpropane (TBME) and the residue in TBME (100 mL) was heated to 50 to 60 °C. Heptane (250 mL) was gradually added to the warm mixture while maintaining the internal temperature above 50 °C to crystallize the product. The mixture was gradually cooled

to 10 °C and filtered. The wet cake was washed with heptane (2 x 40 mL). The solids were dried under reduced pressure to give 1-(3-fluoro-2-(trifluoromethyl)isonicotinoyl)piperidin-4-one (7) (25.8 g, 92.7% yield) as off-white to brown solids. ¹H NMR (300 MHz, DMSO-*d*₆) δ 8.68 (d, ³*J*_{HH} = 4.69 Hz, 1H, NCH in pyridine), 7.97 (dd, ³*J*_{HH} = 4.69 Hz, ⁴*J*_{HF} = 4.69 Hz, 1H, NCCH in pyridine), 3.92 (br s, 2H, one of NCH₂ in piperidine ring, one of another NCH₂ in piperidine ring, both in axial position), 3.54 (t, ³*J*_{HH} = 6.15 Hz, 2H, one of NCH₂ in piperidine ring, one of another NCH₂ in piperidine ring, both in equatorial position), 2.48 (t, ³*J*_{HH} = 6.44 Hz, 2H, NCCH₂), 2.34 (t, ³*J*_{HH} = 6.15 Hz, 2H, NCCH₂) ppm; ¹³C NMR (75 MHz, DMSO-*d*₆) δ 207.17 (C=O), 161.66 (N-C=O), 151.26 (d, ¹*J*_{CF} = 266.89 Hz, C-F), 146.90 (d, ⁴*J*_{CF} = 6.05 Hz, NCH in pyridine), 135.56 (C-C=O), 134.78-135.56 (m, NCCF₃), 128.27 (d, ³*J*_{CF} = 7.19 Hz, NCCH in pyridine), 119.52 (d× quartet, ¹*J*_{CF} = 274.38 Hz, ³*J*_{CF} = 4.89 Hz, CF₃), 45.10 (NC in piperidine ring) ppm, one carbon (NCC in piperidine ring) missing due to overlap with (CD₃)₂SO; ¹⁹F NMR (282 MHz, DMSO-*d*₆) δ -64.58 (d, ⁴*J*_{FF} = 15.85 Hz, F₃C), -128.90 (d × quartet, ⁴*J*_{FF} = 15.85 Hz, ⁴*J*_{FH} = 4.05 Hz, FC) ppm; C₁₂H₁₀F₄N₂O₂ (MW, 290.21), LCMS (EI) *m/e* 291.1 (M⁺ + H). The purity of compound 7 via this process was determined to be greater than about 99% as measured by HPLC.

Example A: *In vitro* JAK Kinase Assay

The compound of Formula I was tested for inhibitory activity of JAK targets according to the following *in vitro* assay described in Park *et al.*, *Analytical Biochemistry* **1999**, 269, 94-104. The catalytic domains of human JAK1 (a.a. 837-1142) and JAK2 (a.a. 828-1132) with an N-terminal His tag were expressed using baculovirus in insect cells and purified. The catalytic activity of JAK1 and JAK2 was assayed by measuring the phosphorylation of a biotinylated peptide. The phosphorylated peptide was detected by homogenous time resolved fluorescence (HTRF). IC₅₀s of compounds were measured for each kinase in the 40 microL reactions that contain the enzyme, ATP and 500 nM peptide in 50 mM Tris (pH 7.8) buffer with 100 mM NaCl, 5 mM DTT, and 0.1 mg/mL (0.01%) BSA. For the 1 mM IC₅₀ measurements, ATP concentration in the reactions was 1 mM. Reactions were carried out at room temperature for 1 hr and then stopped with 20 μL 45 mM EDTA,

300 nM SA-APC, 6 nM Eu-Py20 in assay buffer (Perkin Elmer, Boston, MA).

Binding to the Europium labeled antibody took place for 40 minutes and HTRF signal was measured on a Fusion plate reader (Perkin Elmer, Boston, MA). The compound of Formula I and the adipic acid salt had an IC₅₀ at JAK1 of ≤ 5 nM (measured at 1 mM ATP) with a JAK2/JAK1 ratio of > 10 (measured at 1 mM ATP).

Example B: Cellular Assays

Cancer cell lines dependent on cytokines and hence JAK/STAT signal transduction, for growth, can be plated at 6000 cells per well (96 well plate format) in RPMI 1640, 10% FBS, and 1 nG/mL of appropriate cytokine. Compounds can be added to the cells in DMSO/media (final concentration 0.2% DMSO) and incubated for 72 hours at 37 °C, 5% CO₂. The effect of compound on cell viability is assessed using the CellTiter-Glo Luminescent Cell Viability Assay (Promega) followed by TopCount (Perkin Elmer, Boston, MA) quantitation. Potential off-target effects of compounds are measured in parallel using a non-JAK driven cell line with the same assay readout. All experiments are typically performed in duplicate.

The above cell lines can also be used to examine the effects of compounds on phosphorylation of JAK kinases or potential downstream substrates such as STAT proteins, Akt, Shp2, or Erk. These experiments can be performed following an overnight cytokine starvation, followed by a brief preincubation with compound (2 hours or less) and cytokine stimulation of approximately 1 hour or less. Proteins are then extracted from cells and analyzed by techniques familiar to those schooled in the art including Western blotting or ELISAs using antibodies that can differentiate between phosphorylated and total protein. These experiments can utilize normal or cancer cells to investigate the activity of compounds on tumor cell survival biology or on mediators of inflammatory disease. For example, with regards to the latter, cytokines such as IL-6, IL-12, IL-23, or IFN can be used to stimulate JAK activation resulting in phosphorylation of STAT protein(s) and potentially in transcriptional profiles (assessed by array or qPCR technology) or production and/or secretion of proteins, such as IL-17. The ability of compounds to inhibit these cytokine mediated effects can be measured using techniques common to those schooled in the art.

Compounds herein can also be tested in cellular models designed to evaluate their potency and activity against mutant JAKs, for example, the JAK2V617F mutation found in myeloid proliferative disorders. These experiments often utilize cytokine dependent cells of hematological lineage (*e.g.* BaF/3) into which the wild-
5 type or mutant JAK kinases are ectopically expressed (James, C., *et al. Nature* 434:1144-1148; Staerk, J., *et al. JBC* 280:41893-41899). Endpoints include the effects of compounds on cell survival, proliferation, and phosphorylated JAK, STAT, Akt, or Erk proteins.

Certain compounds herein can be evaluated for their activity inhibiting T-cell
10 proliferation. Such as assay can be considered a second cytokine (*i.e.* JAK) driven proliferation assay and also a simplistic assay of immune suppression or inhibition of immune activation. The following is a brief outline of how such experiments can be performed. Peripheral blood mononuclear cells (PBMCs) are prepared from human whole blood samples using Ficoll Hypaque separation method and T-cells (fraction
15 2000) can be obtained from PBMCs by elutriation. Freshly isolated human T-cells can be maintained in culture medium (RPMI 1640 supplemented with 10% fetal bovine serum, 100 U/ml penicillin, 100 µg/ml streptomycin) at a density of 2×10^6 cells/ml at 37 °C for up to 2 days. For IL-2 stimulated cell proliferation analysis, T-cells are first treated with Phytohemagglutinin (PHA) at a final concentration of 10 µg/mL for 72h.
20 After washing once with PBS, 6000 cells/well are plated in 96-well plates and treated with compounds at different concentrations in the culture medium in the presence of 100 U/mL human IL-2 (ProSpec-Tany TechnoGene; Rehovot, Israel). The plates are incubated at 37 °C for 72h and the proliferation index is assessed using CellTiter-Glo Luminescent reagents following the manufactory suggested protocol (Promega;
25 Madison, WI).

Example C: *In vivo* anti-tumor efficacy

Compounds herein can be evaluated in human tumor xenograft models in immune compromised mice. For example, a tumorigenic variant of the INA-6
30 plasmacytoma cell line can be used to inoculate SCID mice subcutaneously (Burger, R., *et al. Hematol J.* 2:42-53, 2001). Tumor bearing animals can then be randomized into drug or vehicle treatment groups and different doses of compounds can be

administered by any number of the usual routes including oral, i.p., or continuous infusion using implantable pumps. Tumor growth is followed over time using calipers. Further, tumor samples can be harvested at any time after the initiation of treatment for analysis as described above (Example B) to evaluate compound effects
5 on JAK activity and downstream signaling pathways. In addition, selectivity of the compound(s) can be assessed using xenograft tumor models that are driven by other know kinases (*e.g.* Bcr-Abl) such as the K562 tumor model.

Example D: Murine Skin Contact Delayed Hypersensitivity Response Test

10 Compounds herein can also be tested for their efficacies (of inhibiting JAK targets) in the T-cell driven murine delayed hypersensitivity test model. The murine skin contact delayed-type hypersensitivity (DTH) response is considered to be a valid model of clinical contact dermatitis, and other T-lymphocyte mediated immune disorders of the skin, such as psoriasis (*Immunol Today*. 1998 Jan;19(1):37-44).
15 Murine DTH shares multiple characteristics with psoriasis, including the immune infiltrate, the accompanying increase in inflammatory cytokines, and keratinocyte hyperproliferation. Furthermore, many classes of agents that are efficacious in treating psoriasis in the clinic are also effective inhibitors of the DTH response in mice (*Agents Actions*. 1993 Jan;38(1-2):116-21).
20 On Day 0 and 1, Balb/c mice are sensitized with a topical application, to their shaved abdomen with the antigen 2,4,dinitro-fluorobenzene (DNFB). On day 5, ears are measured for thickness using an engineer's micrometer. This measurement is recorded and used as a baseline. Both of the animals' ears are then challenged by a topical application of DNFB in a total of 20 μ L (10 μ L on the internal pinna and 10
25 μ L on the external pinna) at a concentration of 0.2%. Twenty-four to seventy-two hours after the challenge, ears are measured again. Treatment with the test compounds is given throughout the sensitization and challenge phases (day -1 to day 7) or prior to and throughout the challenge phase (usually afternoon of day 4 to day 7). Treatment of the test compounds (in different concentration) is administered
30 either systemically or topically (topical application of the treatment to the ears). Efficacies of the test compounds are indicated by a reduction in ear swelling comparing to the situation without the treatment. Compounds causing a reduction of

20% or more were considered efficacious. In some experiments, the mice are challenged but not sensitized (negative control).

The inhibitive effect (inhibiting activation of the JAK-STAT pathways) of the test compounds can be confirmed by immunohistochemical analysis. Activation of the JAK-STAT pathway(s) results in the formation and translocation of functional transcription factors. Further, the influx of immune cells and the increased proliferation of keratinocytes should also provide unique expression profile changes in the ear that can be investigated and quantified. Formalin fixed and paraffin embedded ear sections (harvested after the challenge phase in the DTH model) are subjected to immunohistochemical analysis using an antibody that specifically interacts with phosphorylated STAT3 (clone 58E12, Cell Signaling Technologies). The mouse ears are treated with test compounds, vehicle, or dexamethasone (a clinically efficacious treatment for psoriasis), or without any treatment, in the DTH model for comparisons. Test compounds and the dexamethasone can produce similar transcriptional changes both qualitatively and quantitatively, and both the test compounds and dexamethasone can reduce the number of infiltrating cells. Both systemically and topical administration of the test compounds can produce inhibitive effects, *i.e.*, reduction in the number of infiltrating cells and inhibition of the transcriptional changes.

Example E: *In vivo* anti-inflammatory activity

Compounds herein can be evaluated in rodent or non-rodent models designed to replicate a single or complex inflammation response. For instance, rodent models of arthritis can be used to evaluate the therapeutic potential of compounds dosed preventatively or therapeutically. These models include but are not limited to mouse or rat collagen-induced arthritis, rat adjuvant-induced arthritis, and collagen antibody-induced arthritis. Autoimmune diseases including, but not limited to, multiple sclerosis, type I-diabetes mellitus, uveoretinitis, thyroiditis, myasthenia gravis, immunoglobulin nephropathies, myocarditis, airway sensitization (asthma), lupus, or colitis can also be used to evaluate the therapeutic potential of compounds herein. These models are well established in the research community and are familiar to those schooled in the art (Current Protocols in Immunology, Vol 3., Coligan, J.E. *et al*,

Wiley Press.; *Methods in Molecular Biology*: Vol. 225, Inflammation Protocols., Winyard, P.G. and Willoughby, D.A., Humana Press, 2003.).

**Example F: Animal Models for the Treatment of Dry Eye, Uveitis, and
Conjunctivitis**

Agents can be evaluated in one or more preclinical models of dry eye known to those schooled in the art including, but not limited to, the rabbit concanavalin A (ConA) lacrimal gland model, the scopolamine mouse model (subcutaneous or transdermal), the Botulinum mouse lacrimal gland model, or any of a number of spontaneous rodent auto-immune models that result in ocular gland dysfunction (e.g. NOD-SCID, MRL/lpr, or NZB/NZW) (Barabino et al., *Experimental Eye Research* 2004, 79, 613-621 and Schrader et al., *Developmental Ophthalmology*, Karger 2008, 41, 298-312, each of which is incorporated herein by reference in its entirety).

Endpoints in these models can include histopathology of the ocular glands and eye (cornea, etc.) and possibly the classic Schirmer test or modified versions thereof (Barabino et al.) which measure tear production. Activity can be assessed by dosing via multiple routes of administration (e.g. systemic or topical) which can begin prior to or after measurable disease exists.

Agents can be evaluated in one or more preclinical models of uveitis known to those schooled in the art. These include, but are not limited to, models of experimental autoimmune uveitis (EAU) and endotoxin induced uveitis (EIU). EAU experiments can be performed in the rabbit, rat, or mouse and can involve passive or activate immunization. For instance, any of a number of retinal antigens can be used to sensitize animals to a relevant immunogen after which animals can be challenged ocularly with the same antigen. The EIU model is more acute and involves local or systemic administration of lipopolysaccharide at sublethal doses. Endpoints for both the EIU and EAU models can include fundoscopic exam, histopathology amongst others. These models are reviewed by Smith et al. (*Immunology and Cell Biology* 1998, 76, 497-512, which is incorporated herein by reference in its entirety). Activity is assessed by dosing via multiple routes of administration (e.g. systemic or topical) which can begin prior to or after measurable disease exists. Some models listed above can also develop scleritis/episcleritis, choroiditis, cyclitis, or iritis and are

therefore useful in investigating the potential activity of compounds for the therapeutic treatment of these diseases.

Agents can also be evaluated in one or more preclinical models of conjunctivitis known those schooled in the art. These include, but are not limited to, rodent models utilizing guinea-pig, rat, or mouse. The guinea-pig models include those utilizing active or passive immunization and/or immune challenge protocols with antigens such as ovalbumin or ragweed (reviewed in Groneberg, D.A., et al., Allergy 2003, 58, 1101-1113, which is incorporated herein by reference in its entirety). Rat and mouse models are similar in general design to those in the guinea-pig (also reviewed by Groneberg). Activity can be assessed by dosing via multiple routes of administration (e.g. systemic or topical) which can begin prior to or after measurable disease exists. Endpoints for such studies can include, for example, histological, immunological, biochemical, or molecular analysis of ocular tissues such as the conjunctiva.

Example G: *In vivo* protection of bone

Compounds can be evaluated in various preclinical models of osteopenia, osteoporosis, or bone resorption known to those schooled in the art. For example, ovariectomized rodents can be used to evaluate the ability of compounds to affect signs and markers of bone remodeling and/or density (W.S.S. Jee and W. Yao, J Musculoskel. Nueron. Interact., 2001, 1(3), 193-207, which is incorporated herein by reference in its entirety). Alternatively, bone density and architecture can be evaluated in control or compound treated rodents in models of therapy (e.g. glucocorticoid) induced osteopenia (Yao, et al. Arthritis and Rheumatism, 2008, 58(6), 3485-3497; and *id.* 58(11), 1674-1686, both of which are incorporated herein by reference in its entirety). In addition, the effects of compounds on bone resorption and density can be evaluable in the rodent models of arthritis discussed above (Example E). Endpoints for all these models can vary but often include histological and radiological assessments as well as immunohisotology and appropriate biochemical markers of bone remodeling.

A number of embodiments of the invention have been described. Nevertheless, it will be understood that various modifications can be made without departing from the spirit and scope of the invention. Accordingly, other embodiments are within the scope of the following claims.