compositions comprising one or more antibodies purified by a method described herein.

One embodiment of the present invention is directed to a process for purifying adalimumab from a fermentation harvest of cultured mammalian cells grown in culture expressing said adalimumab, said process comprising:

a) binding adalimumab from said fermentation harvest to a Protein A resin,

b) eluting the bound adalimumab at an elution pH of 3.2-4, and

c) incubating the eluted adalimumab at a pH of 3-4 for 0.5 to 2 hours.

One embodiment of the present invention is directed to adalimumab when purified according to a process of the invention.

One embodiment of the present invention is directed toward a method of purifying an antibody or antigen-binding portion thereof from a sample matrix such that the resulting antibody composition is substantially free of host cell proteins ("HCPs"). In one aspect, the sample matrix (or simply "sample") comprises a cell line harvest wherein the cell line is employed to produce specific antibodies of the present invention. In a particular aspect, the sample matrix is prepared from a cell line used to produce anti-IL-12 antibodies; in another aspect, the sample matrix is prepared from a cell line used to produce anti-TNFα antibodies; and in another aspect the sample matrix is prepared from a cell line used to produce anti-IL-18 antibodies.

One method of the present invention involves subjecting a sample matrix comprising the putative antibody of interest or antigen-binding portion thereof to a pH adjustment. In one aspect, the pH is adjusted to an acidic pH. An example of a suitable pH is between about a pH of 3 and 5, preferably a pH of about 3.5. This primary recovery is performed, in part, to reduce or inactivate pH-sensitive viruses. In addition to reducing and/or inactivating viruses, the acidic conditions facilitate the removal of cells and cellular debris thus forming a primary recovery sample. After a suitable period of time, the pH can be adjusted toward a more neutral or basic pH and the primary recovery sample is subjected to affinity chromatography, preferably Protein A chromatography. In one aspect, the affinity chromatography sample is collected and further subjected to subsequent chromatographic steps such as ion exchange and hydrophobic interactive chromatography.

In one embodiment, the affinity chromatography step comprises subjecting the primary recovery sample to a column comprising a suitable affinity chromatographic support. Non-limiting examples of such chromatographic supports include, but are not limited to Protein A resin, Protein G resin, affinity supports comprising the antigen against which the antibody of interest was raised, and affinity supports comprising an Fc binding protein.

Protein A resin is useful for affinity purification and isolation of antibodies (IgG). In one aspect, a Protein A column is equilibrated with a suitable buffer prior to sample loading. An example of a suitable buffer is a Tris/NaCl buffer, pH around 7.2. Following this equilibration, the sample can be loaded onto the column. Following the loading of the column, the column can be washed one or multiple times using, e.g., the equilibrating buffer. Other washes