A DOSING REGIME AND FORMULATIONS FOR TYPE B ADENOVIRUSES

The present disclosure relates to a method of treating a patient, for example with a replication capable oncolytic adenovirus employing a dosing regimen designed to allow the virus to have a suitable therapeutic effect and/or minimise adverse events *in vivo*. The disclosure also extends to formulations described herein, methods of preparing said formulations and use of the same in treatment, in particular the treatment of cancer.

BACKGROUND

Cancer is a leading cause of death and serious illness worldwide. There are over 200 different types of cancer and the type of treatment is dependent on the type of cancer. Typically, treatment will involve surgery, chemotherapy and/or radiotherapy. These treatments are often unsuccessful or are only partially successful and have significant side effects. Five year survival rates for cancer can range from less than 5% to over 95% depending on the type of cancer (CRUK statistics, 2000-2001). For example, between 2005-2009, patients with colorectal cancer, which accounts for 13% of all cancers in men and women in the UK, had a five year survival rate of approximately 55% in the UK. This drops to just 12% for patients with metastatic colorectal cancer.

The management of metastatic cancer is mainly palliative and involves a combination of palliative surgery, chemotherapy, radiation and supportive care. Clinical outcomes such as overall survival, response and toxicity are important, but alternative outcomes such as progression-free survival, quality of life, convenience, acceptability and patient choice are also important. New therapies are clearly needed to improve these outcomes.

During transformation, cancer cells acquire certain mutations which render them more permissive to virus infection. Cancer cells also induce the suppression of host anti-tumour activity. Changes within the tumour cells and the local micro-environment create a potential vulnerability and expose the tumour to infection by viruses (Liu et al 2007; Liu et al 2008; Roberts, 2006).

There is a long history of using viruses to treat cancer beginning with anecdotal reports of temporary cancer remission after natural viral infections or viral vaccinations. The earliest report seems to be a 1912 account of the regression of cervical cancer in a patient vaccinated for rabies. Similar results were seen in cancer patients receiving smallpox vaccinations, or following natural virus infections such as mumps or measles. Based on these reports as well as animal data, inoculations of live viruses into patients for cancer treatment were initiated in the late 1940s and early 1950s.

The usual experience, however, was that after occasional temporary tumour regression, the tumour regrew and the patient died. These inoculations seldom resulted in long-lasting complete remissions. In 1957, Albert B. Sabin, M.D., who developed the live oral polio vaccine commented, “The most disappointing aspect is the fact that even when a virus is oncolytic and it punches a hole in a tumour, the immune response of the individual to the virus occurs so fast that the effects are quickly wiped out and the tumour continues to grow.”

At the present time, a number of oncolytic viruses have been identified but the only virus that has been approved for clinical use anywhere in the world to date is Oncorine (H101) a subgroup C adenovirus modified by E1B-55KD deletion enabling conditional replication in P53-deficient cancer cells (H101 is a close analogue of ONYX015 as described by Bischoff et al 1996). Oncorine is administered by intratumoural injection for head and neck cancer.

Talimogene laherparepvec (Tvec), is an oncolytic virus based on Herpes simplex virus type-1 carrying ICP34.5 & ICP47 deletions, expressing US11 as an immediate early gene and encoding GM-CSF. The OPTiM trial is a multi-national, open label, randomized study designed to assess the efficacy and safety of treatment with talimogene laherparepvec as an intratumoural treatment compared to subcutaneously administered GM-CSF in patients with unresectable stage III (b-c) and Stage IV (M1a-c) disease. On interim analysis, talimogene laherparepvec elicited a durable response rate in 16% of patients compared to 2% in those receiving GM-CSF.

Other oncolytic viruses for intra-tumoural administration currently in development include (Sheridan 2013):

- **Reolysin**, an oncolytic reovirus serotype 3 (Dearing strain)
- **PV701**, an oncolytic Newcastle disease virus
- **CG0070**, a conditionally replicating adenovirus encoding GM-CSF
- **Pexastimogene devacirepvec** (Pexa-Vec, JX-594), a thymidine kinase-deleted vaccinia virus encoding GM-CSF
- **Cavatak**, an unmodified Coxsackievirus A21
- **Seprehvir** (HSV1716), a conditionally replicating Herpes simplex type 1 carrying an ICP34.5 deletion
- **DNX-2401**, a conditionally replicating adenovirus encoding an integrin-binding peptide
- **CGTG-102,** a conditionally replicating adenovirus encoding GM-CSF

ColoAd1 is a chimeric (Ad11/Ad3) serogroup B adenovirus, which was developed using the process of directed evolution and it is thought to be suitable for the treatment of cancers of epithelial origin and metastatic forms thereof, including colorectal cancer (Kuhn, I et al. 2008).

To date, clinical studies of oncolytic viruses have primarily investigated intra-tumoural injection of the virus. In a review of clinical studies by Aghi & Martuza (2005) 25 of 36 studies used intra-tumoural injection to administer the virus. However, this method is only practical for treating easily accessible tumours and in patients where the structure of the tumour, such as tissue stroma and necrotic areas therein, do not limit spread of the virus within a tumour (Ries & Korn 2002).

Death from cancer is often the result of inaccessible tumours or metastases. Oncolytic viruses administered intra-tumourally rely on systemic dissemination from the tumour to reach these secondary tumours. However, dissemination has proved transient and often ineffective (Ferguson et al 2012).

Thus, intra-tumoural injection is only suitable for a limited number of cancers and is not suitable for treatment of, for example of many metastatic cancers.

Where intravenous administration of oncolytic viruses has been employed generally it has been associated with acute toxicity and rapid clearance. For example, in the case of the group C adenovirus Ad5, for which uptake is mediated by the ubiquitous coxsackie adenovirus

receptor (CAR), side effects including acute liver toxicity, influenza like illness and haematological changes have been regularly reported, whilst rapid hepatic clearance and immunological neutralisation are also well described.

The currently established view is that repeated doses are required in order to produce and maintain efficacy. In all the oncolytic cancer treatments under investigation it is generally envisaged that the treatment will be chronic, with repeat administrations over many weeks, months or years. For example, in the case of PV701, treatment in at least one patient continued in cycles for about 10 month with 6 days between finishing the previous treatment cycle and initiating the next treatment cycle.

Neumanatis et al (2001) report administration of up to 24 weekly cycles of an intravenous infusion of ONYX-015 to cancer patients. In an on-going phase III clinical study (Clinicaltrials.gov identifier NCT01708993), Reolysin® (an oncloytic reovirus) was infused over a 1 hour period on days 1 to 3 and then every 3 weeks until progression. In a recently reported phase I clinical trial (Clinicaltrials.gov identifier NCT01380600) JX-594 was administered intravenously every 2 weeks on four occasions, and in a second on-going phase I/II clinical study (Clinicaltrials.gov identifier NCT01394939) JX-594 is administered intravenously weekly for 5 weeks followed by up to 3 intra-tumoural boosts to liver metastases of patients with metastatic colorectal cancer. In the on-going OPTIM clinical trial talimogene laherparepvec was administered intra-tumourally every two weeks for up to 18 months (Clinicaltrials.gov identifier NCT00769704).

SUMMARY OF THE INVENTION

In a first aspect of the disclosure there is provided a method of treating a human patient, said method comprising the steps of:

systemically administering multiple doses of a parenteral formulation of a replication capable oncolytic adenovirus of subgroup B in a single treatment cycle,

wherein the total dose given in each administration is in the range of $1x10^{10}$ to $1x10^{14}$ viral particles per dose, and

wherein each dose of virus is administered such that the rate of viral particle delivery is in the range of $2x10^{10}$ particles per minute to $2x10^{12}$ particles per minute.

In an independent aspect the present disclosure relates to ColoAd1 for use in treating ovarian cancer, for example administering a therapeutically effective amount of ColoAd1 to a patient with ovarian cancer, for example employing a dosing regimen described herein.

In a further independent aspect the present disclosure relates to a combination therapy comprising oncolytic type B adenovirus, such as ColoAd1, and a chemotherapeutic agent which does not interfere with the adenovirus activity, such as viral replication *in vivo*.

In one embodiment the combination therapy is employed for treatment of cancer, in particular a cancer described herein, in particular colorectal cancer or ovarian cancer, including metastatic forms thereof.

In one embodiment ColoAd1 in a combination therapy is dosed according to a regimen described herein.

Also provided is a parenteral formulation of a replication capable oncolytic adenovirus of subgroup B, for use in treatment as described herein.

The present disclosure also extends to use of a parenteral formulation of a replication capable oncolytic adenovirus of subgroup B, for the manufacture of a medicament, as described herein and for use in treatments described herein.

In one aspect there is also provided a unit dose in the range $1x10^{10}$ to $1x10^{14}$, such as $6x10^{12}$ viral particles of a replication capable oncolytic adenovirus of subgroup B.

Also provided is an infusion or injection rate for dosing the viral particles of $2x10^{9}$ to $2x10^{12}$ virus particles (VP) per minute, for example $1.5x10^{11}$ VP per minute.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 Shows the cytotoxicity profile on A549 cells of ColoAd1 in the presence of fresh whole human blood.

Figure 2 Biodistribution of 1e11 ($1x10^{11}$) particles of ColoAd1 in normal BalbC mice 24 hours post injection.

Figure 3 Biodistribution of ColoAd1 and ColoAd1CJ132 in CD46 transgenic mice at 1 hr and 72 hrs post-injection.

Figure 4 Clearance kinetics of ColoAd1 from primary organs liver, spleen and lungs in CD46-expressing mice followed for 65 days.

Figure 5 Kinetics of ColoAd1 in mice either with or without co-administration of neutralising serum.

Figure 6 Cytokine levels after the first and subsequent equal therapeutic doses in a pre-clinical toxicology study in CD-1 mice:

Figure 7A-C Cytokine levels (ng/L) (TNF (A), gamma interferon (B) and IL6 (C)) over time in human cancer patients with metastatic solid epithelial tumours after intravenous doses of ColoAd1.

Figure 8A Systemic pharmacokinetics of ColoAd1 (Genome copies per mL of blood) in human cancer patients with metastatic solid epithelial tumours.

Figure 8B Systemic pharmacokinetics of ColoAd1 (Genome copies per mL of blood) in human cancer patients with metastatic solid epithelial tumours.

Figure 9A Systemic pharmacokinetics of ColoAd1 (Genome copies per mL of blood) in human cancer patients administered with 1e10 ($1x10^{10}$) ColoAd1 viral particles over 5 minutes.

Figure 9B Systemic pharmacokinetics of ColoAd1 (Genome copies per mL of blood) in human cancer patients administered with 1e11 ($1x10^{11}$) ColoAd1 viral particles over 5 minutes.

Figure 9C Systemic pharmacokinetics of ColoAd1 (Genome copies per mL of blood) in human cancer patients administered with 1e12 ($1x10^{12}$) ColoAd1 viral particles over 5 minutes.

Figure 9D Systemic pharmacokinetics of ColoAd1 (Genome copies per mL of blood) in human cancer patients administered with 1e13 ($1x10^{13}$) ColoAd1 viral particles over 5 minutes.

Figure 9E Systemic pharmacokinetics of ColoAd1 (Genome copies per mL of blood) in human cancer patients administered with 3e12 ($3x10^{12}$) ColoAd1 viral particles over 5 minutes.

Figure 9F Systemic pharmacokinetics of ColoAd1 (Genome copies per mL of blood) in human cancer patients administered with 3e12 ($3x10^{12}$) ColoAd1 viral particles over 20 minutes.

Figure 9G Systemic pharmacokinetics of ColoAd1 (Genome copies per mL of blood) in human cancer patients administered with 6e12 ($6x10^{12}$) ColoAd1 viral particles over 40 minutes.

Figure 10 Slower infusion of the same dose lowers the cMax level at the end of infusion (cohort 5 vs 6)

Figure 11 MCP1 levels (ng/L) over time in human cancer patients with metastatic solid epithelial tumours after intravenous doses of ColoAd1.

Figure 12 Schematic diagram showing ColoAd1 replication cycle in cell.

Figure 13 ColoAd1 infection of cancer cells shown as nuclear staining in a colorectal cell line following in vitro infection with the virus.

Figure 14A Nuclear staining (hexon staining) of ColoAd1 in colorectal tissue from a patient with colorectal cancer after administration of ColoAd1 by IT.

Figure 14B Isotype control staining for Figure 14A.

Figure 14C Colorectal tissue (hexon staining) showing no nuclear staining in stromal cells (following IV administration of ColoAd1 to a colorectal cancer patient.

Figure 14D Isotype control for Figure 14C.

Figure 15 320 compounds (clinically approved or compounds in development) that were analysed for their impact on viral replication.

Figure 16A-D An *in vivo* murine model showing the effects of Paclitaxel and ColoAd1 combination therapy (and controls).

Figure 17-18 *In vivo* data for ColoAd1 and chemotherapy in a murine model.

DETAILED DESCRIPTION

In one embodiment the dose administered is in the range of 1 $x10^{10}$ to 1 x 10^{13}, such as 1 $x10^{10}$ to 1 x 10^{12} viral particles.

In one embodiment the total dose administered in one treatment cycle is $1x10^{12}$, $2x10^{12}$, $3x10^{12}$, $4x10^{12}$, $5x10^{12}$, $6x10^{12}$, $7x10^{12}$, $8x10^{12}$ or $9x10^{12}$ viral particles.

In one embodiment the total dose administered in one treatment cycle is $6x10^{12}$ viral particles.

It is hypothesised by the present inventors that what may be most critical for efficacy is to establish a productive infection within the tumour at an early stage, for example before an anti-viral immune response has developed.

The dosing regimen thus has to balance delivering sufficient virus to generate, for example adequate plasma levels of virus for a period long enough to seed viral infection in the cancer cells, whilst not eliciting toxicity and severe adverse events in the patient (or minimising the same).

The present inventors have shown for the first time that infection of the tumour by type B adenovirus can be established by doses of viral particles administered intravenously. Support for this conclusion is provided herein where patients with colorectal cancer that received treatment with ColoAd1 intravenously were shown to have virus infection in the nucleus of cancer cells, when the cells were stained for hexon and also when analysed independently by PCR. Virus in the nucleus indicates the virus life cycle of the virus is in progress and an increase in the viral load in the patients indicates the virus is able to replicate.

When administering an oncolytic adenovirus systemically to a patient, a number of dosing variables need to be considered. These dosing variables include, but are not limited to: the route of virus administration; the dose of virus administered; the rate of viral administration for each dose; the interval between individual viral administrations in a given cycle; the number of viral administrations per treatment cycle; the interval between treatment cycles; the number of treatment cycles; and finally the use of any co-medicaments or other supportive care used to enhance efficacy or minimise adverse effects. Each of the dosing parameters is in turn dependent upon the specific characteristics of the type of oncolytic virus under investigation.

The key parameters will include, but are not limited to: the relative degree and avidity of any binding of the virus to tumour cells versus non-tumour cells; the relative selectivity and potency of the virus in tumour cells versus non-tumour cells; the rate of active uptake and clearance of the virus by reticuloendothelial cells (for example liver Kupffer cells) and any specific or non-specific binding of blood elements to the virus.

These key parameters are in turn driven by important physical and phenotypic characteristics of the specific virus type, which include but are not limited to: the receptor specificity of the virus; the charge carried on the viral coat; the presence or absence of an envelope; the size of the viral particle; the immunogenicity of the viral particle; the inflammatory potential of the viral particle; the tumour specificity of the virus; the replicative speed of the virus; and the killing potency of the virus.

Therefore, the suitability of any given dosing regime will vary with different types of virus and the most appropriate regime may be specific to the type of virus being administered. For example, Zhang *et al* (2012) describe an Ad5-Ad48 chimeric virus created to reduce hexon binding with blood coagulation factor X in order to eliminate liver sequestration, enhance circulation and decrease toxicity, whilst maintaining anti-tumour activity. Likewise, Shashkova et al (2009) describe significant differences between wild-type human adenoviral serotypes 5, 6, 11, and 35 when investigated as potential anticancer agents. It is thus anticipated that different viral types will behave with significant differences when administered systemically to humans and thus the optimum dosing strategy cannot be predicted *a priori* without *in vivo* data and preferably supporting clinical data.

The dose regimen described herein may be particularly suitable to achieve this for group B adenoviruses when compared to, for example the current practice of more regularly spaced and long term repeated dosing.

The objective of an optimised dose regimen for any given oncolytic adenovirus is thus to maximise delivery of the virus to the tumour cells whilst minimising both the induction of side

effects (adverse events) and antiviral immunity, in order to produce a suitable risk benefit treatment profile whilst still allowing for repeat viral administrations, as appropriate to the therapy. The optimised dose regimen will thus differ between virus types and specifically between adenoviral subtypes due to the differences in the viral coat.

Much work has been performed in the prior art based on Ad5, which is a subgroup C adenovirus, the infectivity for which is mediated by the Coxsackie Adenovirus Receptor. When delivered systemically, over 90% of the delivered dose is taken up by the liver. The rapid and extensive loss to the liver reduces the virus uptake by tumours and diminishes therapeutic efficacy. The vast majority of this dose is taken up by cytokine producing innate immune cells such as Kupffer cells, which are specialised macrophages resident in the liver. Ad5 also exhibits liver toxicity and causes necrosis and subsequent depletion of Kupffer cells.

Shoshkova et al 2009 showed that depletion of Kupffer cells by Ad5 increased levels of hepatocyte transduction with subsequent delivery of Ad5 vectors and also suggests that the mechanisms elucidated for Ad5 are not necessarily relevant to adenoviruses from subgroup B, for example Ad11 and Ad35. The data therein suggests that subgroup C adenoviruses interact with Kupffer cells in a similar manner, whereas subgroup B adenoviruses either are not well recognized by Kupffer cells or do not cause death of these cells. In particular, Shoshkova suggests that pre-dosing with Ad11 based viruses does not have the same beneficial effect upon Kupffer cells as Ad5. This paper concludes that whilst there may be some binding for subgroup B adenoviruses (including Ad11) the impact of this is in fact minimal.

Whilst not wishing to be bound by theory the present inventors believe that, contrary to the prior art suggestions, cytokine producing innate immune cells such as Kupffer cells may play a role in the clearance of subgroup B adenoviruses.

Furthermore, binding of blood coagulation factor X to Ad5 hexon is a mechanism of infection of hepatocytes and this mechanism may also be relevant to other adenoviruses *in vivo* (see for example Molecular Therapy vol. 17 no. 10, 1683-1691 October 2009) but generally is not a mechanism of hepatic uptake for adenoviruses from subgroup B.

High global seroprevalence of Ad5 (high Ad5 neutralizing antibody titres in human populations) and certain other adenoviral serotypes represent a significant concern for the systemic application of high seroprevelance adeno-based therapies, because such blood-borne viruses can be neutralised by pre-existing antibodies, Vogels et al Journal of Virology, Aug 2003 Vol 77, No. 15 page 8263-8271.

Subgroup B adenoviruses have certain inherent advantages in this respect, in that they are associated with lower seroprevalence (Stone et al Journal of Virology 2005 Vol 79 No. 8 page 5090-5104) and have lower inflammatory potential. Initial dosing may thus be far more efficient than with Ad5, for example. However, the ability to avoid the immune system after systemic delivery may still become an issue with repeat dosing. Thus, even with the local suppression of the immune system by the cancer, avoidance of the immune system is still probably the biggest obstacle to the long term success of oncolytic virus therapy based on subgroup B adenoviruses.

The data generated by the present inventors supports the position that the therapeutic effect of the oncolytic adenoviruses of subgroup B can thus be improved and/or the elimination of

neutralisation of the adenovirus by the immune system can be minimised by employing an appropriate dosing regimen.

In one embodiment the dosing regimens herein may also minimise side-effects, for example flu like symptoms and inflammatory responses.

In one embodiment, the replication competent adenovirus is administered repeatedly during an early "dosing window" before a specific anti-viral immune response has been developed, and that later dosing windows may again be exploited when the specific anti-viral immune response has waned once more. That is to say several treatments in a short period of time followed by a period of time before initiating subsequent treatment cycles.

Advantageously, by administering the replication competent adenovirus in such a way, the viral blood levels are sufficient to establish a self-amplifying infection within the tumour (which is known to be an immunosuppressed environment) thereby potentially avoiding the need for chronic repeat administrations with the oncolytic virus. In order to establish a self-amplifying infection within the tumour, it is beneficial to maintain the level of virus within the patients' blood stream at a level above an effective infective concentration for as long as possible but without producing adverse events. This concept is akin to identifying a therapeutic window for the virus, i.e. a range of doses or dosing regimens where the therapeutic effect is optimised and the side effects are minimised.

This can be achieved by optimising both the dose administered and the infusion rate of the virus. In one embodiment, the rate of infusion of the virus is equal to or greater than the rate of clearance of the virus by the body.

Once an infection is established inside the tumour the virus is relatively protected from neutralising antibodies and is afforded a potentially permissive environment to replicate and to produce a therapeutic effect without dose limiting toxicities.

In addition, it is hypothesised by the inventors that peaks in virus concentration (C_{max}) contribute to side effects and that a flatter pharmacological profile may be desirable.

In one embodiment the C_{max} is kept below a specific value, for example 3 $x10^8$ DNA copies per ml. It appears that a C_{max} level above the relevant threshold is more likely to induce serious adverse events or toxicity in some patients.

In one embodiment the rate of infusion has more influence than the absolute amount of virus administered.

Based on the data generated in the clinic, the present inventors also believe that virus can be delivered at a rate above the rate of clearance and up to 1.5 to $2x10^{11}$ viral particles per minute over a prolonged period such as up to 72 hours or more (wherein the total dose of virus delivered is above $6x10^{12}$ virus particles) without eliciting serious adverse events in the patient.

In one embodiment, the C_{max} of the viral genome in the blood is maintained at a level of less than $3x10^8$ genomes per ml of blood.

The present inventors have evaluated the initial rate of clearance of virus in a number of scenarios and believe that the estimated α-half-life is in the region of 18 minutes.

The use of prophylactic anti-inflammatories during oncolytic viral therapy is controversial. On the one hand it has been proposed that their use may minimise adverse events and thus

enhance tolerability of oncolytic Newcastle Disease Virus (Lorence et al 2007). On the other hand, there have been reports that the occurrence of fever may be associated with enhanced oncolytic efficacy for adenoviruses (Yu *et al* 2007).

The present inventors have found that the use of prophylactic or therapeutic agents (including anti-inflammatories, steroids, antiemetics, antidiarrheals or analgesics) administered during this treatment cycle may enhance the tolerability of this regime, particularly allowing for higher or more frequent doses.

In one embodiment, steroids are administered during the treatment cycle.

The present inventors thus hypothesise that six parameters, used either individually or in concert, are important in achieving the goal of suitable delivery of an oncolytic subgroup B adenovirus:

a) the number of virus particles administered with each dose,
b) the rate that each virus dose is administered (the number of viral particles delivered per minute),
c) the number of individual doses of virus in the treatment cycle,
d) the interval between each individual dose within the treatment cycles,
e) the use of prophylactic anti-inflammatory medicines during the treatment cycle, and
f) the time period between treatment cycles.

These parameters can be balanced against each other i.e. an increased dose can be given at a slower infusion rate to off-set the negative effect of the increase.

If the dose is too low then the level of viral particles is not sufficient to establish an effective infection of the cancer cells. If the rate of administration is too slow then then the viral particles can readily be cleared by natural viral sinks (for example cytokine producing innate immune cells such as hepatic Kupffer cells or blood components) and an effective infection of the cancer/tumour cells is not achieved. If the viral dose is too high and/or if the rate of administration is too fast then the number of adverse events is likely to increase because of the high concentration of viral particles. The latter then induce an inflammatory cytokine response, which may increase the side-effects experienced by the patient. A moderate infusion rate can thus optimise the dose delivered.

On average the rate of clearance of type B adenoviruses such as ColoAd1 have an α-half-life of about 18 mins.

A single dose of virus may fail to establish an infection, but may adequately occupy or remove viral sinks (for example cytokine producing innate immune cells such as hepatic Kupffer cells or blood components). If viral sinks have been adequately occupied or removed, and if subsequent doses are administered soon thereafter, the viral kinetics may be altered for the later doses, with a longer circulating half-life and/or higher peak plasma levels. In this case, one or more doses administered shortly after the first dose may more effectively establish an effective infection of the cancer cells.

However, if the subsequent doses are administered too far apart (for example greater than 14 days apart) then the viral sinks may have time to replenish and the benefit of the prior dose may be lost and/or a specific anti-viral immune response may have developed. Depletion of, for example cytokine producing innate immune cells such as hepatic Kupffer cells with this

form of dosing regimen may have an important secondary benefit in that Kupffer mediated cytokine release can be greatly reduced on subsequent viral doses such that these doses are better tolerated even in the face of higher viral plasma levels.

Thus the present inventors are advocating the administration of a given treatment cycle over a relatively short period of time, for example as described below.

From the work completed by the present inventors it appears that for a group B adenovirus multiple doses in a treatment cycle, with each dose administered over a relatively short period of time, with each dose administered as a moderately fast infusion, optionally in combination with prophylactic agents and with each dose separated by a relatively short period of time, is suitable for infecting cancer cells with oncolytic type B adenovirus with minimal toxicity.

Treatment cycles may be repeated as required.

The present inventors have monitored the inflammatory cytokines TNF, gamma interferon, IL-6 and MCP-1 as markers of acute toxicity and believe that by the second or subsequent doses, there is reduced toxicity and increased potential for the virus to infect the cancer cells in each case because the non-cancerous viral sinks are either removed or occupied by both the first and the second doses, provided these doses are administered at an appropriate dose level, rate and frequency.

In one embodiment three doses are employed in the treatment cycle, and in a further embodiment more than three doses are employed in the treatment cycle.

In one embodiment, a dose is administered on any or all of days 1, 3, 5, 14, and 21.

In another embodiment, a follow-up dose is administered as a maintenance or booster dose, for example, biweekly, weekly, once every two weeks, or every 3 weeks, such as every week or every 3 weeks, for a suitable period, in particular whilst the treatment is beneficial to the patient as maintenance therapy, for example whilst a patient remains in remission.

The skilled addressee will appreciate that various modifications to the treatment cycle can be made depending on the needs of the individual patient.

The present disclosure also extends to a replication capable oncolytic adenovirus of subgroup B for use in treatment of a human patient by systemically administering at least one dose, such as multiple doses of a parenteral formulation comprising the adenovirus in a single treatment cycle, wherein the total dose given in each administration is in the range $1x10^{10}$ to $7x10^{12}$, for example $1x10^{10}$ to $5x10^{12}$ viral particles, and is administered over a period of 1 minute to 90 minutes.

In a further aspect the disclosure extends to use of a replication capable oncolytic adenovirus of subgroup B for the manufacture of a medicament for use in the treatment of a human patient by systemically administering at least one dose, such as multiple doses of a parenteral formulation comprising the adenovirus in a single treatment cycle, wherein the total dose given in each dose is in the range $1x10^{10}$ to $1x10^{13}$ for example $1x10^{10}$ to $7 x 10^{12}$, such as $1x10^{10}$ to $5x10^{12}$, or $6x10^{12}$ viral particles, and is administered over a period of 1 minute to 90 minutes.

In one embodiment the first dose in the treatment of a given cycle is a lower dose than the dose administered in subsequent treatments in the cycle.

It appears that contrary to Shoshkova's suggestion, based on work in mice, that pre-dosing with Ad11 based viruses does not have a beneficial priming effect upon cytokine producing innate immune cells such as Kupffer cells. In fact, optimisation of the dose and timing between administration of group B oncolytic adenovirus doses may be employed to minimize side effects and hence be beneficial.

In one embodiment the dose administered is $6x10^{12}$, for example over a period of 20 to 60 minutes, such as 40 minutes.

In one embodiment a high first and second dose (i.e. which may correspond to a normal therapeutic dose) may be desirable to fully occupy cytokine producing innate immune cells, such as Kupffer cells (and/or other viral sinks) and thus optimise delivery for subsequent doses. To put it another way, the first and second dose may be equal.

In one embodiment all the doses administered contain an equal number of viral particles. This may be particularly advantageous in that it simplifies manufacture of the viral formulation, reduces the risk of dosing errors, and may in fact provide a highly effective treatment regime.

In one embodiment a follow-up cycle of treatment is provided 1 month to 6 months after completion of the previous treatment cycle, for example 2, 3, 4, 5 months thereafter in order to allow the immune response to wane.

In one embodiment a follow-up cycles may be a single dose administered weekly or bi-weekly, for a period of 1 month to 5 years, such 6, 7, 8, 9, 10, 11, 12, 18, 24, 30 or 36 months.

In one embodiment the following-up treatment cycle is initiated within about 14 days of administering the last dose in the first treatment cycle.

The follow-up cycles may also act as maintenance doses, thereby helping to maintain viral load at a level sufficient to provide a therapeutic effect.

In one embodiment there are 1, 2, 3, 4, 5 or more subsequent treatment cycles, for example 1 or 2.

In one embodiment there is only one treatment cycle with no subsequent treatment cycles.

In one embodiment there is provided a liquid parenteral formulation for infusion or injection of a replication capable oncolytic subgroup B adenovirus (such as ColoAd1) wherein the formulation provides a dose in the range of $1x10^{10}$ to $1x10^{14}$ viral particles per volume of dose, such as $6x10^{12}$ viral particles per dose.

Also disclosed is a method for treating a patient by administering a parenteral formulation according to the present disclosure comprising a replication capable oncolytic subgroup B adenovirus, for example containing a dose described herein, such as $6x10^{12}$ viral particles per dose.

Also disclosed is a method for treating a patient by administered parenteral formulation according to the present disclosure comprising a replication capable oncolytic subgroup B adenovirus said method comprising the co-administration to the patient of one or more substances or medicaments selected from the group comprising anti-inflammatory, steroid, anti-histamines, anti-pyretic medicaments and fluids for hydration.

Also disclosed is a method of determining when it is suitable to administer to a subject subsequent cycles of the parenteral formulation according to the disclosure comprising a replication capable oncolytic subgroup B adenovirus, said method comprising the steps of:

determining the pre-existing titre of the patient's specific antiviral immunity prior to a first treatment cycle,

serially determining the patient's specific antiviral immunity subsequent to the first treatment cycle, and

delaying any subsequent treatment cycles until the patient's specific antiviral immunity has reduced to a pre-specified percentage of baseline.

The term "serially determining" as used herein refers to determining a patient's antiviral immunity at multiple time points, which may be regularly or irregularly spaced apart. The multiple readings obtained may be used to generate an average titre over a particular period of time for example.

The term "pre-specified percentage of baseline" as used herein refers to a viral titre which is defined as a threshold or limit for a particular patient, taking into account factors such as a baseline measured before treatment is initiated, the patient's prognosis, ongoing cancer therapy, any adverse side effects, etc.

In one embodiment, the "pre-specified percentage of baseline" is 90% or less of the patient's baseline viral titre, such as 80% or less, 70% or less, 60% or less, 50% or less, 40% or less, 30% or less, 20% or less, or 10% or less.

In an alternative embodiment, no testing is performed prior to administering the subsequent treatment cycles.

In one embodiment there is provided use of a glass or plastic syringe with an internal volume in the range of 3 to 50ml, said syringe containing a parenteral formulation comprising 1 $x10^{10}$ to $1x10^{14}$, for example $1x10^{10}$ to $7x10^{12}$ (such as $1x10^{10}$ to $6x10^{12}$ or $1x10^{10}$ to $5x10^{12}$, or $1x10^{10}$ to 4 $x10^{12}$, or $1x10^{10}$ to $3x10^{12}$, or $1x10^{10}$ to $2x10^{12}$, or $1x10^{10}$ to $1x10^{12}$) viral particles, of a replication capable oncolytic adenovirus of subgroup B, wherein the formulation is sterile and was filled into the syringe under aseptic conditions, for use in treatment, in particular for use in the manufacture of a medicament which is capable of injection or intravenous infusion to a human subject.

The skilled person will appreciate that the formulations may include an overage of the viral particles, for example to compensate for viral particles that may adhere to the surface of the syringe and which are not subsequently administered.

Advantageously, such a prefilled syringe would significantly enhance the usability and cost effectiveness for a manufactured oncolytic adenovirus of subgroup B, by removing the need for dose preparation in specialised pharmacies using expensive resources such as specialised equipment (including extraction hoods) and trained personnel.

The disclosure also extends to pre-filled vials of the said formulation, in particular vials each containing a single dose, in the range defined herein.

In one embodiment the virus formulation is provided in a concentrated form, for example concentrated liquid, suitable for diluting with a sterile isotonic diluent, such as saline, glucose or similar locally before administration to a patient.

Advantageously, the dosing regimen herein is suitable for delivering a therapeutically effective amount of subgroup B oncolytic virus to the cancer target. In particular the dosing regimen herein may minimise neutralisation and/or clearance of the oncolytic virus by, for example

blood born agents, sinks, cytokine producing innate immune cells such as Kupffer cells and the immune system. The latter may lead to a better availability of the therapeutic dose of the oncolytic virus and, overall, an improved prognosis for the patient and/or improved survival. Advantageously the present regimen may also provide an improved quality of life for patients by minimising adverse events and/or side effects during treatment.

In one embodiment a patient who receives treatment according to the present disclosure shows an increased survival rate in comparison to a patient receiving the current standard treatment at the time of filing, for example a statistically significant increase in survival.

In one embodiment a patient who receives treatment according to the present disclosure shows a decreased tumour burden, in comparison to the standard treatment at the time of filing, for example a statistically significant decrease.

In one embodiment the a patient who receives treatment according to the present disclosure shows an increased likelihood of going into remission, in comparison to the standard treatment at the time of filing, for example a statistically significant increase.

In one embodiment the amount or extent of metastasis is reduced, for example is statistically significantly reduced in a patient who receives treatment according to the present disclosure in comparison to the standard treatment at the time of filing.

Whilst not wishing to be bound by theory, it is thought that cells of the mononuclear phagocyte system, and in particular cytokine producing innate immune cells such as Kupffer cells, may be responsible for the clearance of type B oncolytic viruses from the circulation, even though the prior art suggest otherwise.

Furthermore, the mouse studies conducted, by the present inventors, leads them to believe that after the first or second dose in a treatment regimen, cytokine producing innate immune cells such as the Kupffer cells are depleted or occupied such that they are unable to efficiently clear, for example the third dose and subsequent doses if those doses are administered in a short time frame after the second dose, or alternatively a lower toxicity may be observed or both. It is hypothesised that the cytokine markers indicate the latter, in that the levels of the cytokines are not significantly elevated after administration of the second or third dose when compared to the first dose, provided that the doses are administered within a relatively short time period. The present inventors take this to be an indication that the mechanisms for clearing the virus may be subdued after the first and second dose.

Whilst studies in mice do not always parallel what is seen in the human system, particularly with viruses, in this instance the human observations seem to correlate well with those in the murine model performed by the present inventors. The impact of the dosing regimen on cytokine responses and pharmacokinetics of ColoAd1 has also been exemplified in human subjects, by the present inventors.

As employed herein, “method of treating a patient by systemically administering” is intended to refer to a method of administering a therapeutic agent to a human to effect entry of the entity into the patient’s circulatory system, in particular wherein the treatment is intended to prevent or slow the progression of, ameliorate or cure a malignancy, such as cancer or complications or symptoms associated therewith, for example direct administration to the circulatory system by intravenous administration.

In one embodiment systemic delivery affords the opportunity to treat a primary tumour, any overt, inaccessible or undiagnosed tumours and/or metastases. This is particularly advantageous because it may lead to a better overall prognosis for the patient and/or improved survival.

Thus systemic delivery as employed herein does not refer to treatment which is localised in the tumour or within a body cavity, such as the peritoneal cavity. Examples of systemic delivery include intravenous infusion and intramuscular and subcutaneous injection.

Parenteral formulation means a formulation designed not to be delivered through the GI tract nor through topical administration. Typical parenteral delivery routes include injection, implantation or infusion. In one embodiment the formulation is provided in a form for bolus delivery.

In one embodiment the parenteral formulation is in the form of an injection. Injection includes intravenous, subcutaneous, intra-tumoural or intramuscular injection. Injection as employed herein means the insertion of liquid into the body via a syringe. In one embodiment the method of the present disclosure does not involve intra-tumoural injection. An injection will generally involve the administration of 150mL of fluid or less over a short period of time, for example 1.5 minutes or less.

In one embodiment the formulation is delivered into the peritoneal cavity.

For head and neck cancer, or brain metastases of epithelial cancers, intracranial injection may be necessary.

In one embodiment the parenteral formulation is in the form of an infusion.

Infusion as employed herein means the administration of fluids at a slower rate by drip, infusion pump, syringe driver or equivalent device. In one embodiment the infusion is administered over a period in the range of 1.5 minutes to 90 minutes, such as 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55 or 60 minutes.

In one embodiment the volume of formulation administered is 100mLs or less, in particular 50mLs or less, for example 30mls, 10ml, 5ml or less, such as 3ml, such as administered by a syringe driver. The latter may be referred to as a slow injection.

In one embodiment the infusion is delivered at a rate in the range of 0.5 to 6ml per minute, for example 0.75ml per minute

In one embodiment, the infusion is delivered at a rate in the range of $2x10^9$ to $2x10^{12}$ virus particles (VP) per minute, for example $1.5x10^{11}$ VP per minute.

In one embodiment the injection is administered as a slow injection, for example over a period of 1.5 to 30 or 1.5 to 40 minutes.

In one embodiment the formulation is for intravenous administration. This route is particularly effective for delivery of oncolytic virus because it allows rapid access to the majority of the organs and tissue and is particular useful for the treatment of metastases, for example established metastases especially those located in highly vascularised regions such as the liver and lungs.

In one embodiment a combination of administration methods are employed, for example IV and intra-tumourally or intraperitoneally and intra-tumourally, or IV and intra-peritoneally.

Thus in one embodiment systemic administration of the present disclosure may be employed in combination with other routes of administration, such as intra-tumoural administration either concomitantly or sequentially, for example a first pre-treatment cycle may be intra-tumoural and the second treatment cycle may be systemic according to the present disclosure. Alternatively, the first treatment cycle may be according to the present disclosure and subsequent cycles or boosts may be intra-tumoural, as appropriate.Therapeutic formulations typically will be sterile and stable under the conditions of manufacture and storage. The composition can be formulated as a solution, microemulsion, liposome, or other parenteral formulation suitable for administration to a human and may be formulated as a pre-filled device such as a syringe or vial, particular as a single dose.

In one embodiment 2 or more doses are employed in the treatment cycle, for example 2, 3, 4, 5 or 6 doses are employed in each treatment cycle and for example may be provided as a kit. Each dose administered in a given treatment cycle may be referred to herein as a treatment.

In one embodiment a lower first dose is employed in comparison to the subsequent doses administered in the cycle, for example a lower dose may be in the range of 30-95% of the subsequent dose or doses, for example 50, 60, 70 or 80%.

In one embodiment a higher first is employed in comparison to the subsequent doses administered in the cycle, which may be desirable to full occupy cytokine producing innate immune cells such as the Kupffer cells and thus optimise delivery for subsequent doses.

A higher dose means more than 100% of the subsequent dose, for example 105 to 150% of the subsequent dose, such as 110%, 115%, 120%, 125%, 130%, 135%, 140% or 145% of the subsequent dose.

In one embodiment 1, 2, 3 or all the doses administered contains an equal number of viral particles. This may be particularly advantageous in that it simplifies manufacture of the viral formulation and may in fact provide a highly effective treatment regime.

In one embodiment the “same dose” i.e. the same number of viral particles are administered in one or more doses, such as all the doses in a treatment cycle, however, the doses may be administered at different rates, for example as described herein.

Treatment cycle as employed herein is the period of treatment between a period of rest in a course of treatment repeated in accordance with a schedule with periods of rest there-between. A treatment cycle generally refers to multiple (i.e. at least two) treatments administered as part of a program or schedule of treatment, administered over a relatively short period of time, for example about 1 to 4 weeks, such as 3 weeks, 2 weeks, or 1 week. Generally, a given treatment cycle will be a part of a larger treatment regime.

In one embodiment the treatment cycle is a period of 14 days or less, for example 10, 9, 8, 7 or 5 days, such as 7 or 5 days.

In one embodiment each further dose or doses is/are administered at approximately 48 hour intervals, such as every 40 to 56 hours. This is advantageous since it allows dosing to occur within a normal working week or within an outpatient setting.

In one embodiment the first dose is administered on day 1 and the further therapeutic doses are administered every second day thereafter, such as on days 1, 3, 5, 7, 9, 11 and 13, or once every approximately 48 hours thereafter, such as every 40 to 56 hours.

In one embodiment the plasma levels of virus in the patient after administration of the dose (such as the second or subsequent dose) is at least 2 x10^6 viral particles per ml, for example for a period of 15 minutes or longer, for example 20, 30, 40, 50, 60 minutes or more.

In vitro studies performed by the inventors (see Figure 1) suggest that for virus particles in whole human blood at 37°C, killing drops below 50% at < 2x10^6 particles ml. Furthermore, the inventors have been able to show the presence of live viral particles in patient blood using plaque assays when viral genome levels are above for example 1.6e6 to 1e8, and can be consistently detected.

In one embodiment there is at least 14 days between treatment cycles.

The formulation will generally comprise a pharmaceutically acceptable diluent or carrier, for example a non-toxic, isotonic carrier that is compatible with the virus, and in which the virus is stable for the requisite period of time.

The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a dispersant or surfactant such as lecithin or a non-ionic surfactant such as polysorbate 80 or 40. In dispersions the maintenance of the required particle size may be assisted by the presence of a surfactant. Examples isotonic agents include sugars, polyalcohols such as mannitol, sorbitol, or sodium chloride in the composition.

In one embodiment a sterile isotonic diluent such as saline or glucose (for example 5% glucose is employed).

In one embodiment parenteral formulations employed in the method may comprise one or more of the following a buffer, for example 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid, a phosphate buffer, and/or a Tris buffer, a sugar for example dextrose, mannose, sucrose or similar, a salt such as sodium chloride, magnesium chloride or potassium chloride, a detergent such as a non-ionic surfactant such as briji®, PS-80, PS-40 or similar. The formulation may also comprise a preservative such as EDTA or ethanol or a combination of EDTA and ethanol, which are thought to prevent one or more pathways of possible degradation.

In one embodiment the formulation will comprise purified oncolytic virus, for example 1x10^{10} to 1x10^{14} viral particles per dose, such as 1x10^{10} to 7x10^{12} viral particles per dose, in particular 1x10^{10} to 1x10^{12} viral particles per dose, including overage as necessary.

In one embodiment the formulation according to the present disclosure comprises 6x10^{12} viral particles.

In one embodiment the concentration of virus in the formulation is in the range 2 x 10^8 to 2 x 10^{14} vp/mL, such as 2 x 10^{12} vp/ml.

In one embodiment the parenteral formulation comprises glycerol.

In one embodiment the formulation comprises oncolytic adenovirus from subgroup B, HEPES (N-2-hydroxyethylpiperazine-N´-2-ethanesulfonic acid), glycerol and buffer.

In one embodiment the parenteral formulation consists of virus, HEPES for example 5mM, glycerol for example 5-20% (v/v), hydrochloric acid, for example to adjust the pH into the range 7-8 and water for injection.

In one embodiment 0.7 mL of ColoAd1 at a concentration of 2 x 10^{12} vp/mL is formulated in 5 mM HEPES, 20% glycerol with a final pH of 7.8.

Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, monostearate salts and gelatin.

Thus the oncolytic adenoviruses employed herein may be administered in a time release formulation, for example in a composition which includes a slow release polymer. The oncolytic adenovirus can be prepared with carriers that will protect it against neutralisation and/or prevent rapid release, such as a controlled release formulation, such as implants and microencapsulated delivery systems.

Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, polylactic acid and polylactic, polyglycolic copolymers (PLG). Biocompatible non-degradable polymers such as Polyethylene glycol and poly(N-(2-hydroxypropyl)methacrylamide) can also be used. Many methods for the preparation of such formulations are known to those skilled in the art.

Sterile injectable solutions can be prepared by incorporating the oncolytic adenovirus in the required amount in an appropriate solvent, for example with one or a combination of ingredients described herein, as relevant, followed by filtered sterilisation. Generally, dispersions are prepared by incorporating the oncolytic adenovirus into a sterile vehicle which contains a basic dispersion medium and the required other ingredients.

Generally the parenteral formulation according to the disclosure is a sterile liquid formulation, such as an aqueous formulation, substantially free of particulates, for example prepared aseptically and sterilised by passing through a 0.2 micron filter.

In one embodiment the therapeutic parenteral formulation is administered to minimise the contact of the formulation with the epidermis of the patient, for example employing a sheathed needle or via a cannula. This precaution is thought to minimise the immune response of the patient to the oncolytic virus, for example by minimising contact with Langerhan cells in the skin.

Replication capable as employed herein is a virus that can replicate in a host cell. In one embodiment replication capable encompasses replication competent and replication selective viruses.

Replication competent as employed herein is intended to mean an oncolytic adenovirus that is capable of replicating in a human cell, such as a cancer cell, without any additional complementation to that required by wild-type viruses, for example without relying on defective cellular machinery. That is, they are tumour selective by infecting tumour cells in preference to non-tumour cells. ColoAd1 is an example of a replication competent virus.

Replication selective or selective replication as employed herein is intended to mean an oncolytic adenovirus that is able to replicate in cancer cells employing an element which is specific to said cancer cells or upregulated therein, for example defective cellular machinery, such as a p53 mutation, thereby allowing a degree of selectivity over healthy/normal cells.

Oncolytic subgroup B adenovirus as employed herein refers to an adenovirus comprising at least the hexon and fiber from subgroup B (see Shenk et al and Table 1) that preferentially infects and/or lyses tumour cells compared with normal cells. Thus an oncolytic subgroup B

adenovirus as employed herein includes a chimeric, a mutant or a variant, with the fiber and hexon of a group B adenovirus and which retains oncolytic properties.

Adenovirus or adenoviral serotype as used herein refers to any of the human adenoviral serotypes currently known (51) or isolated in the future. See for example, Strauss (1984) and Shenk (2001). Adenovirus serotypes are classified into subgroups as shown in Table 1.

Table 1 shows the division of adenovirus serotypes:

Subgroup	Adenoviral Serotype
A	12, 18, 31
B	**3, 7, 11, 14, 16, 21, 34, 35, 51**
C	1, 2, 5, 6
D	8-10, 13, 15, 17, 19, 20, 22-30, 32, 33, 36-39, 42-50
E	4
F	40, 41

Examples of subgroup B viruses include Ad11 (wild-type) such as Ad11a and Ad11p (Genbank Accession No: AF532578) and the chimeric adenovirus ColoAd1. The latter is disclosed in WO 2005/118825 and the full sequence for the virus is provided in SEQ ID NO: 1 therein.

Thus in one embodiment the virus employed in the method according to the present disclosure is a chimeric virus.

Chimeric adenovirus as employed herein refers to adenoviruses which have DNA from two or more different adenovirus serotypes such as those generated using the method of WO2005/118825 which is incorporated herein by reference.

In one embodiment the chimeric adenovirus is ColoAd1. ColoAd1 is thought to kill tumour cells by a mechanism which more closely resembles necrosis than apoptosis (unpublished data produced at the University of Oxford). This has a number of potential beneficial effects (Kirn et al 2001; Small et al 2006; Reid et al 2002; Liu et al 2007; Ferguson et al 2012):

- ColoAd1 has been shown to be potent in multi-drug resistant cancer cell lines and in cancer stem-cell like cells, which are known to have a resistance to apoptosis;
- An inflammatory necrotic cell death may be more suitable for the generation of a specific anti-tumoural immune response;
- ColoAd1 exits tumour cells very rapidly, even before target cell death, and may thus have enhanced ability to spread.

ColoAd1 is a chimera of Ad11 and Ad3 but has an outer capsule which is entirely homologous with that of Ad11. The viral kinetics, inflammatory potential and immunological characteristics of ColoAd1 thus most closely resemble and predict those of Ad11 and other subgroup B adenoviruses.

In one embodiment the oncolytic virus employed in the method of the present disclosure is deleted in the E3 and/or E4 region or part thereof. This may be beneficial because it may allow more rapid replication of the virus *in vivo*.

In addition the E3 deletion may contribute to the rapid clearance of the virus from non-cancer cells as the E3 region encoded proteins which may be relevant to avoiding the immunity of the host.

In one embodiment the virus employed in the method of the present disclosure is based on Ad11 or derived therefrom such that the hexon and fibre are substantially similar to Ad11, such as Ad11p. Furthermore since the serotype designation of adenovirus is based on the exterior properties of the virus i.e. hexon and fibre properties, the present disclosure is useful in type B adenovirus which have similar surface properties.

In one embodiment the type B adenovirus is OvAd1 or OvAd2 which are disclosed in SEQ ID NO: 1 and SEQ ID NO: 2 respectively in WO2008/080003, incorporated herein by reference.

Substantially similar as employed herein refers to an amino acid sequence for a relevant protein or proteins which is/are at least 95% identical (e.g. 96, 97, 98, 99 or 100% identical) over the "whole" of the particular protein. The protein(s) being compared may be part of a larger entity but the comparison will be the whole length of the relevant fragment or component.

Adenovirus type 5 (Ad5) generally enter the cell via the coxsackie-adenovirus receptor (CAR). However, Adenovirus serotype 11 (Ad11) is a subgroup B adenovirus that targets a different receptor (CD46) which is expressed at low levels in all nucleated cells. In normal cells CD46 is often hidden on the basolateral surfaces of cells and is thus not available for virus binding (Varela JC, et al Int J Cancer 2008 Sep 15;123(6):1357-63; Maisner et al., 1997). However, in tumour cells it typically has enhanced surface expression, particularly in more advanced and aggressive tumours (Kinugasa et al., 1999). Therefore, Ad11 efficiently infects carcinoma cell lines, for example from lung epithelial carcinoma (A549 cells), hepatoma (HepG2), prostatic cancer (DU 145 and LNCaP), laryngeal cancer (Hep2) and breast cancer (CAMA and MG7) and also to glioblastoma, medulloblastoma and neuroblastoma cells (Mei et al 2003). Thus Ad11 preferentially infects tumour cells and viruses derived therefrom are thought to be useful in the treatment of at least one or more of the above cancers. As a chimera of Ad11 and Ad3, ColoAd1 shares these characteristics with Ad11.

In one embodiment a virus employed in the method of the present disclosure comprises a transgene (in particular one or more transgenes), for example a therapeutic transgene, for expression *in vivo*. A transgene gene as employed herein is intended to refer to a gene not found in the parent or wild type virus. Such genes may perform a function as a marker or reporter for tracking efficacy of viral infection. Alternatively the gene may perform a role in improving the efficacy of the virus. Alternatively the gene may deliver a cytotoxic agent to the cell.

The therapeutic transgene may express a therapeutic agent in the cell, for example siRNA; shRNA; a polypeptide; tumour associated antigen (TAA), cytokine; antibody; or an anti-angiogenesis factor.

Examples of therapeutic antibodies include anti-VGEF antibodies such as bevacizumab, anti-EGFR antibodies such as cetuximab, an anti-CD20 antibody such as rituximab, or an immune system activator modulator such as anti-CTLA4 (e.g. ipilimumab), anti-PD-1 and anti-PD-L1 amongst others. Single chain antibodies, antibody subunits, antibody fragments and TRAPs

may also be encoded as well as full length antibodies. Importantly for the current disclosure, the inclusion of these proteins does not change the surface properties of the virus and therefore can readily be incorporated into the genome without deleterious effects upon the dosing as described herein whilst providing additional therapeutic mechanisms for attacking the cancer cells.

Examples of cytokines include interferon-alpha, interferon-gamma and IL-2 amongst others.

As the RNA, antibody, polypeptide, TAA or cytokine will be expressed in the tumour it is thought that this presents an opportunity to change the microenvironment of the tumour but avoid systemic side effects of the delivered agent. For example, it may be possible to stimulate the local immune system to attack the cancer. It is possible to modulate this local effect by altering whether or not the RNA, antibody, polypeptide, TAA or cytokine is secreted from the cell and when during the viral life cycle it is expressed.

In one embodiment the transgene encodes thymidine kinase, for example from a non-human origin or cytosine deaminase, for example from bacterial origin or from a yeast.

In one embodiment the antibody, polypeptide or cytokine or similar is non-human in origin and is not humanised. The latter is not likely to detrimentally effect the activity of the entity in the cancer cell and has the advantage that material that may escape the cancer cell will attract the attention of the immune system locally and will be rapidly cleared.

In one embodiment the virus encodes and expresses *in vivo* a visible or visualisable protein, for example a fluorescent protein, such as GFP or similar. Given the virus selectively infects cancerous cells, when it expresses a visible or visualisable protein then it can be used to highlight the area of cancerous tissue for resection or radiation.

In one embodiment, the viruses may be armed with therapeutic genes capable of eliciting anti-tumour immune function, inhibition of tumour neovascularization, or prodrug activation.

Therapeutic dose as employed herein refers to the amount of oncolytic adenovirus that is suitable for achieving the intended therapeutic effect when employed in a suitable treatment regimen, for example ameliorates symptoms or conditions of a disease. A dose may be considered a therapeutic dose in the treatment of cancer or metastases when the number of viral particles may be sufficient to result in the following: tumour or metastatic growth is slowed or stopped, or the tumour or metastasis is found to shrink in size, and/or the life span of the patient is extended. Suitable therapeutic doses are generally a balance between therapeutic effect and tolerable toxicity, for example where the side-effect and toxicity are tolerable given the benefit achieved by the therapy.

In one embodiment the therapeutic dose range does not have a dose limiting toxicity.

Dose limiting toxicity as employed herein means the appearance of side effects during treatment that are severe enough to prevent any one of the following: further increase in dosage, frequency or strength or to prevent continuation of treatment at any dosage level. Toxicity effects which are intolerable, for example associated with a high dose mean the latter is not suitable for use as a therapeutic dose in the context of the present disclosure.

In one embodiment pre-existing immunity to the Ad11 capsid is weak permitting effective administration of further therapeutic doses on or after day 7.

In one embodiment the poor immune stimulatory properties of the Ad11 capsid permits effective administration of further therapeutic doses on or after day 7.

In one embodiment intravenous delivery of the virus is less immunogenic in terms of antiviral immunogenicity than sub-cutaneous or intramuscular delivery of virus.

It is generally believed that the toxicity of Ad11 may be lower than certain other adenoviruses, such as Ad5. This together with the lower seroprevalence is beneficial but this may not be sufficient to allow Ad11 to evade immune responses. Even though the literature suggests that the subgroup B adenoviruses are not toxic to liver cells it may be that macrophages in the lungs, liver (Kupffer cells) and spleen clear oncolytic viruses after systemic delivery.

It is thought that the rapid delivery of at least two doses of the oncolytic virus may be beneficial in generating sufficient levels of virus that are sustained for a period which allows adequate infection of the target cells, namely cancer cells.

Providing at least two doses in quick succession may allow one or more the following beneficial events to occur a) the immune mechanisms are occupied by the first dose, which may then allow the second dose to escape the full onslaught of the immune system to reach the target and/or b) at least two doses in quick succession allow the biodistribution of the virus to reach sufficient levels for a sufficient period to reach the target cells *in vivo*, either way once the virus reaches and infects the target cells it is able to replicate.

Biodistribution as employed herein means the distribution *in vivo.*

Whilst not wishing to be bound by theory the inventors believe that the first dose of virus may down regulate clearance, for example mechanisms such as those employing cytokine producing innate immune cells such as Kupffer cells thereby improving the bioavailablity for the further therapeutic dose(s). The first dose of virus may thus “deplete” the phagocytic ‘sinks’ for circulating virus thereby achieving better delivery and/or increased efficacy. Depleting the phagocytic sinks also reduces the tendency to release cytokines on subsequent doses and thus allows higher viral blood levels to be achieved without excessive toxicity.

Bioavailability as employed herein means the amount of virus available to perform its intended therapeutic function *in vivo*.

In one embodiment the method herein wherein at least three doses are administered minimises side-effects and/or toxicity in the patient.

In one embodiment the adenovirus is stealthed by coating said virus with a polymer, for example to at least partially avoid the patient’s immune system.

Stealthed as employed herein means that the adenovirus’s exterior surface has been modified to avoid the patient’s immune response, for example employing a polymer. Examples of suitable polymers are disclosed in WO98/19710, WO00/74722, WO2010/067041, WO2010/067081, and WO2006/008513 incorporated herein by reference.

In one embodiment the oncolytic virus is conjugated to a cytotoxic or immunomodulatory agent.

In one embodiment the oncolytic adenovirus is provided which is pegylated, for example to reduce immunogenenicity and/or increase half-life.

In one embodiment the method of treatment is for use in the treatment of a tumour.

Tumour as employed herein is intended to refer to an abnormal mass of tissue that results from excessive cell division that is uncontrolled and progressive, also called a neoplasm. They may be either benign (not cancerous) or malignant. Tumour encompasses all forms of cancer and metastases.

In one embodiment the tumour is a solid tumour. The solid tumour may be localised or metastasised.

In one embodiment the tumour is of epithelial origin.

In one embodiment the tumour is a solid tumour.

In one embodiment the tumour is a malignancy, such as colorectal cancer, hepatoma (liver cancer), prostate cancer, pancreatic cancer, breast cancer, ovarian cancer, thyroid cancer, renal cancer, bladder cancer, head and neck cancer or lung cancer.

In one embodiment the tumour is a colorectal malignancy.

Malignancy as employed herein means cancerous cells.

In one embodiment the cancer is colorectal cancer and/or metastatic forms thereof such as liver metastasis.

In one embodiment the cancer is liver cancer and/or metastatic forms thereof.

In one embodiment the cancer is lung cancer and/or metastatic forms thereof.

In one embodiment the cancer is ovarian cancer and/or metastatic forms thereof, such as lung metastasis.

In one embodiment the cancer is renal cancer and/or metastatic forms thereof.

In one embodiment the cancer is bladder cancer and/or metastatic forms thereof.

In one embodiment the cancer is throat cancer.

In one embodiment the cancer is skin cancer, such as melanoma. In one embodiment the cancer is Leukemia. In one embodiment the cancer is glioblastoma, medulloblastoma or neuroblastoma. In one embodiment the cancer is a neuroendocrine cancer. In one embodiment the cancer is Hodgkin's or non-Hodgkins lymphoma.

In one embodiment the oncolytic adenovirus is employed in the treatment or prevention of metastasis.

In one embodiment the oncolytic adenoviruses described herein are suitable for the treatment of cancerous cells that have migrated to the lymph node. The present inventors have shown that oncolytic virus administered to colorectal cancer patients can infect cancerous cells that have migrated to the lymph nodes.

In one embodiment the virus, formulations and regimens according to the present disclosure are suitable for treating abnormal pre-cancerous cells.

In one embodiment the method or formulation herein is employed in the treatment of drug resistant cancers.

In one embodiment the method or formulation is employed in to sensitise drug resistant to cancers to said drugs.

CANCER TYPES IN MORE DETAIL

Lung cancer

Lung cancers are classified according to histological type and are categorized by the size and appearance of the malignant cells seen by a histopathologist under a microscope. For therapeutic purpose, two broad classes are distinguished: non-small cell lung carcinoma and small cell lung carcinoma.

In one embodiment the epithelial cancer is lung cancer, for example small-cell lung cancer (SCLC) and non-small-cell lung cancer (NSCLC).

Non-small-cell lung carcinoma-The three main subtypes of NSCLC are adenocarcinoma, squamous-cell carcinoma and large-cell carcinoma.

Nearly 40% of lung cancers are adenocarcinoma, which usually originates in peripheral lung tissue. A subtype of adenocarcinoma, the bronchioloalveolar carcinoma, is more common in female never-smokers, and may have a better long term survival.

Squamous-cell carcinoma accounts for about 30% of lung cancers. They typically occur close to large airways. A hollow cavity and associated cell death are commonly found at the center of the tumour. About 9% of lung cancers are large-cell carcinoma. These are so named because the cancer cells are large, with excess cytoplasm, large nuclei and conspicuous nucleoli.

Small-cell lung carcinoma- In small-cell lung carcinoma (SCLC), the cells contain dense neurosecretory granules (vesicles containing neuroendocrine hormones), which give this tumour an endocrine/paraneoplastic syndrome association. Most cases arise in the larger airways (primary and secondary bronchi). These cancers grow quickly and spread early in the course of the disease. Sixty to seventy percent have metastatic disease at presentation.

In one embodiment the cancer is non-small lung carcinoma.

Liver cancer

In one embodiment the cancer is liver cancer, for example a liver metastasis from a primary cancer, for example colon cancer, which has spread to the liver. In one embodiment the liver cancer is hepatocellular carcinoma (HCC).

Renal cancer

In one embodiment there is provided treatment of renal cancer, for example renal cell carcinoma and/or urothelial cell carcinoma using an oncolytic adenovirus as disclosed herein. Other examples of renal cancer include squamous cell carcinoma, juxtaglomerular cell tumour (reninoma), angiomyolipoma, renal oncocytoma, Bellini duct carcinoma, clear-cell sarcoma of the kidney, mesoblastic nephroma, Wilms' tumour, mixed epithelial stromal tumour, clear cell adenocarcinoma, transitional cell carcinoma, inverted papilloma, renal lymphoma, teratoma, carcinosarcoma, and carcinoid tumour of the renal pelvis.

Bladder cancer

In one embodiment the cancer is bladder cancer, for example is any of several types of malignancy arising from the epithelial lining (i.e., the urothelium) of the urinary bladder. About 90% of bladder cancers are transitional cell carcinoma. The other 10% are squamous cell carcinoma, adenocarcinoma, sarcoma, small cell carcinoma, and secondary deposits from cancers elsewhere in the body. The staging of is given below.

T (Primary tumour)

- TX Primary tumour cannot be assessed
- T0 No evidence of primary tumour

- Ta Non-invasive papillary carcinoma
- Tis Carcinoma in situ ('flat tumour')
- T1 Tumour invades subepithelial connective tissue
- T2a Tumour invades superficial muscle (inner half)
- T2b Tumour invades deep muscle (outer half)
- T3 Tumour invades perivesical tissue:
- T3a Microscopically
- T3b Macroscopically (extravesical mass)
- T4a Tumour invades prostate, uterus or vagina
- T4b Tumour invades pelvic wall or abdominal wall

N (Lymph nodes)

- NX Regional lymph nodes cannot be assessed
- N0 No regional lymph node metastasis
- N1 Metastasis in a single lymph node 2 cm or less in greatest dimension
- N2 Metastasis in a single lymph node more than 2 cm but not more than 5 cm in greatest dimension, or multiple lymph nodes, none more than 5 cm in greatest dimension
- N3 Metastasis in a lymph node more than 5 cm in greatest dimension

M (Distant metastasis)

- MX Distant metastasis cannot be assessed
- M0 No distant metastasis
- M1 Distant metastasis.

The current disclosure extends to any stage of bladder cancer.

Ovarian cancer

In an independent aspect the present disclosure relates to ColoAd1, a formulation of the same or a combination therapy comprising ColoAd1, for use in treating ovarian cancer, for example administering a therapeutically effective amount of ColoAd1 to a patient with ovarian cancer, for example employing a dosing regimen described herein.

There are more than 30 different types of ovarian cancer which are classified according to the type of cell from which they start. Cancerous ovarian tumours can start from three common cell types:

- Surface Epithelium - cells covering the lining of the ovaries
- Germ Cells - cells that are destined to form eggs
- Stromal Cells - Cells that release hormones and connect the different structures of the ovaries

The present disclosure relates to treatment of ovarian cancer from any source, for example as described herein, in particular epithelium cells. Epithelial ovarian carcinomas (EOCs) account for 85 to 90 percent of all cancers of the ovaries.

Common Epithelial Tumours - Epithelial ovarian tumours develop from the cells that cover the outer surface of the ovary. Most epithelial ovarian tumours are benign (noncancerous). There are several types of benign epithelial tumours, including serous adenomas, mucinous adenomas, and Brenner tumours. Cancerous epithelial tumours are carcinomas - meaning

they begin in the tissue that lines the ovaries. These are the most common and most dangerous of all types of ovarian cancers. Unfortunately, almost 70 percent of women with the common epithelial ovarian cancer are not diagnosed until the disease is advanced in stage.

There are some ovarian epithelial tumours whose appearance under the microscope does not clearly identify them as cancerous. These are called borderline tumours or tumours of low malignant potential (LMP tumours). The method of the present disclosure includes treatment of the latter.

Germ Cell Tumours - Ovarian germ cell tumours develop from the cells that produce the ova or eggs. Most germ cell tumours are benign (non-cancerous), although some are cancerous and may be life threatening. The most common germ cell malignancies are maturing teratomas, dysgerminomas, and endodermal sinus tumours. Germ cell malignancies occur most often in teenagers and women in their twenties. Today, 90 percent of patients with ovarian germ cell malignancies can be cured and their fertility preserved.

Stromal Tumours - Ovarian stromal tumours are a rare class of tumours that develop from connective tissue cells that hold the ovary together and those that produce the female hormones, estrogen and progesterone. The most common types are granulosa-theca tumours and Sertoli-Leydig cell tumours. These tumours are quite rare and are usually considered low-grade cancers, with approximately 70 percent presenting as Stage I disease (cancer is limited to one or both ovaries).

Primary Peritoneal Carcinoma-The removal of one's ovaries eliminates the risk for ovarian cancer, but not the risk for a less common cancer called Primary Peritoneal Carcinoma. Primary Peritoneal Carcinoma is closely rated to epithelial ovarian cancer (most common type). It develops in cells from the peritoneum (abdominal lining) and looks the same under a microscope. It is similar in symptoms, spread and treatment.

Stages of Ovarian Cancer

Once diagnosed with ovarian cancer, the stage of a tumour can be determined during surgery, when the doctor can tell if the cancer has spread outside the ovaries. There are four stages of ovarian cancer - Stage I (early disease) to Stage IV (advanced disease). The treatment plan and prognosis (the probable course and outcome of your disease) will be determined by the stage of cancer you have.

Following is a description of the various stages of ovarian cancer:

Stage I - Growth of the cancer is limited to the ovary or ovaries.

Stage IA - Growth is limited to one ovary and the tumour is confined to the inside of the ovary. There is no cancer on the outer surface of the ovary. There are no ascites present containing malignant cells. The capsule is intact.

Stage IB - Growth is limited to both ovaries without any tumour on their outer surfaces. There are no ascites present containing malignant cells. The capsule is intact.

Stage IC - The tumour is classified as either Stage IA or IB and one or more of the following are present: (1) tumour is present on the outer surface of one or both ovaries; (2) the capsule has ruptured; and (3) there are ascites containing malignant cells or with positive peritoneal washings.

Stage II - Growth of the cancer involves one or both ovaries with pelvic extension.

Stage IIA - The cancer has extended to and/or involves the uterus or the fallopian tubes, or both.

Stage IIB - The cancer has extended to other pelvic organs.

Stage IIC - The tumour is classified as either Stage IIA or IIB and one or more of the following are present: (1) tumour is present on the outer surface of one or both ovaries; (2) the capsule has ruptured; and (3) there are ascites containing malignant cells or with positive peritoneal washings.

Stage III - Growth of the cancer involves one or both ovaries, and one or both of the following are present: (1) the cancer has spread beyond the pelvis to the lining of the abdomen; and (2) the cancer has spread to lymph nodes. The tumour is limited to the true pelvis but with histologically proven malignant extension to the small bowel or omentum.

Stage IIIA - During the staging operation, the practitioner can see cancer involving one or both of the ovaries, but no cancer is grossly visible in the abdomen and it has not spread to lymph nodes. However, when biopsies are checked under a microscope, very small deposits of cancer are found in the abdominal peritoneal surfaces.

Stage IIIB - The tumour is in one or both ovaries, and deposits of cancer are present in the abdomen that are large enough for the surgeon to see but not exceeding 2 cm in diameter. The cancer has not spread to the lymph nodes.

Stage IIIC - The tumour is in one or both ovaries, and one or both of the following is present: (1) the cancer has spread to lymph nodes; and/or (2) the deposits of cancer exceed 2 cm in diameter and are found in the abdomen.

Stage IV - This is the most advanced stage of ovarian cancer. Growth of the cancer involves one or both ovaries and distant metastases (spread of the cancer to organs located outside of the peritoneal cavity) have occurred. Finding ovarian cancer cells in pleural fluid (from the cavity which surrounds the lungs) is also evidence of stage IV disease.

In one embodiment the ovarian cancer is: type I, for example IA, IB or IC; type II, for example IIA, IIB or IIC; type III, for example IIIA, IIIB or IIIC; or type IV.

The present disclosure relates to treatment of any stage of ovarian cancer, in particular as described herein.

COMBINATION THERAPY

In one embodiment the virus is administered in combination with the administration of a further cancer treatment or therapy.

"In combination" as employed herein is intended to encompass where the oncolytic virus is administered before, concurrently and/or post cancer treatment or therapy.

In one embodiment the oncolytic adenovirus is employed in combination with high intensity focused ultrasound (HIFU) treatment.

Cancer therapy includes surgery, radiation therapy, targeted therapy and/or chemotherapy.

Cancer treatment as employed herein refers to treatment with a therapeutic compound or biological agent, for example an antibody intended to treat the cancer and/or maintenance therapy thereof.

In one embodiment the cancer treatment is selected from any other anti-cancer therapy including a chemotherapeutic agent, a targeted anticancer agent, radiotherapy, radio-isotope therapy or any combination thereof.

In a further independent aspect the present disclosure relates to a combination therapy comprising oncolytic type B adenovirus, such as ColoAd1, and a chemotherapeutic agent which does not interfere with the adenovirus activity. Type B adenovirus, such as ColoAd1 as employed herein includes formulations thereof, for example pharmaceutical formulations thereof.

Activity as employed herein refers to any beneficial property or characteristic of the virus, for example the oncolytic activity and or the ability of the virus to replicate in cancer cells, such as viral replication *in vivo*.

In one embodiment the ColoAd1 in the combination therapy is dosed according to a regimen described herein.

Generally, the combination therapy will be provided as a formulation of the adenovirus and a formulation of the chemotherapeutic agent. Thus the administration of the adenovirus and the chemotherapeutic will suitably be separate events. These administrations may be on the same or different days.

In one embodiment the adenovirus is administered in a suitable regime one week and the chemotherapeutic is administer a following week, for example the next.

In one or more embodiments the chemotherapeutic agent and the adenovirus may have a synergistic therapeutic effect.

The oncolytic adenovirus may be used as a pre-treatment to the therapy, such as a surgery (neoadjuvant therapy), to shrink the tumour, to treat metastasis and/or prevent metastasis or further metastasis. The oncolytic adenovirus may be used after the therapy, such as a surgery (adjuvant therapy), to treat metastasis and/or prevent metastasis or further metastasis.

Concurrently as employed herein is the administration of the additional cancer treatment at the same time or approximately the same time as the oncolytic adenovirus formulation. The treatment may be contained within the same formulation or administered as a separate formulation.

In one embodiment the virus is administered in combination with the administration of a chemotherapeutic agent, for example as described herein, such as paclitaxel, abraxane or similar.

Chemotherapeutic agent as employed herein is intended to refer to specific antineoplastic chemical agents or drugs that are selectively destructive to malignant cells and tissues. For example alkylating agents, antimetabolites, anthracyclines, plant alkaloids, topoisomerase inhibitors, and other antitumour agents. Other examples of chemotherapy include doxorubicin, 5-fluorouracil (5-FU), paclitaxel, capecitabine, irinotecan, and platins such as cisplatin and oxaliplatin. The preferred dose may be chosen by the practitioner based on the nature of the cancer being treated.

Surprisingly the present inventors have established that certain classes of therapeutic agents can inhibit viral replication, for example topoisomerase or parp inhibitors, may inhibit the replication of the virus *in vivo*. Given it is thought to be desirable to establish a viral infection in a cancer cell such that the virus can replicate, then co-administration of compounds that inhibit viral replication is likely to be undesirable.

In one embodiment the chemotherapeutic agent is not an enzyme inhibitor. Thus in one embodiment the combination therapy does not employ a topoisomerase inhibitor.

In one embodiment he chemotherapeutic agent is not a parp inhibitor.

In one embodiment the combination therapy employs a platinum containing chemotherapeutic agent, for example cisplatin, carboplatin or oxaliplatin.

In one embodiment the combination employs a microtubule inhibitor, for example vincristine sulphate, epothilone A, N-[2-[(4-Hydroxyphenyl)amino]-3-pyridinyl]-4-methoxybenzenesulfonamide (ABT-751), ataxol derived chemotherapeutic agent, for example paclitaxel, abraxane, or docetaxel or a combination thereof.

In one embodiment the combination employs an mTor inhibitor. Examples of mTor inhibitors include: everolimus (RAD001), WYE-354, KU-0063794, papamycin (Sirolimus), Temsirolimus, Deforolimus(MK-8669), AZD8055 and BEZ235(NVP-BEZ235).

In one embodiment the combination employs a Pi3 Kinase inhibitor. Examples of Pi3 kinases inhibitors include: GDC-0941, ZSTK474, PIK-90, LY294002, TG100-115, XL147, GDC-0941, ZSTK474, PIK-90, LY294002, TG100-115, XL147, AS-605240, PIK-293, AZD6482, PIK-93, TGX-221, IC-87114, AS-605240, PIK-293, AZD6482, PIK-93, TGX-221, IC-87114 and compounds disclosed in WO2011/048111 incorporated herein by reference including 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-ethynylquinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(3-(2-(2-methoxyethoxy)ethoxy)prop-1-yn-1-yl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(6-morpholino-6-oxohex-1-yn-1-yl)quinazolin-4(3*H*)-one; 6-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)hex-5-ynoic acid; 2-((4-Amino-3-(4-hydroxyphenyl) -1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(6-morpholino-6-oxohex-1-yn-1-yl)quinazolin-4(3*H*)-one; 3-((2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-(3-(2-(2-hydroxyethoxy)ethoxy)prop-1-yn-1-yl)-4-oxoquinazolin-3(4*H*)-yl)methyl) benzonitrile; 2-((4-amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(3-(2-morpholinoethoxy)prop-1-ynyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chloro benzyl)-5-ethynylquinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(3-chlorobenzyl) -5-ethynylquinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(3-chlorobenzyl) -5-ethynyl quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(2-fluorobenzyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxy phenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(2-fluorobenzyl) quinazolin-4 (3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-

ethynyl-3-(3-methoxybenzyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(3-methoxybenzyl) quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(3-(trifluoromethyl)benzyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(3-(trifluoromethyl) benzyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(4-chlorobenzyl) -5-ethynyl quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(4-(methylsulfonyl) benzyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(4-(methylsulfonyl)benzyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(4-(trifluoromethyl) benzyl)quinazolin-4(3*H*)-one; 3-((2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-4-oxo-quinazolin-3(4*H*)-yl)methyl) benzonitrile; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(3-(methyl-sulfonyl)benzyl)quinazolin-4(3*H*)-one; 3-((2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-4-oxo-quinazolin-3(4*H*)-yl)methyl)benzonitrile; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(4-chlorobenzyl) -5-ethynylquinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(4-chlorobenzyl) -5-(3-methoxy-prop-1-ynyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(3-methoxybenzyl)-5-(3-methoxyprop-1-ynyl) quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl) -5-(3-methoxyprop-1-ynyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(4-(trifluoro methyl)benzyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(3-(2-methoxyethoxy)prop-1-ynyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-((5-methylisoxazol-3-yl)methyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-((5-methylisoxazol-3-yl)methyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(3-chloro-2-fluoro-benzyl)-5-ethynylquinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2,6-difluorobenzyl)-5-ethynylquinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(4-chloro-2-fluorobenzyl)-5-ethynylquinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-fluoro-4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-ethynylquinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-(3-methoxyprop-1-ynyl)-3-(3-(trifluoromethyl)benzyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(4-fluorobenzyl) quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(3-cyclopentylprop-1-ynyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-(3-(benzyloxy)prop-1-ynyl)-3-(2-chloro-benzyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl) -5-(5-hydroxypent-1-ynyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-

(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(2-fluoro-5-methoxybenzyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(3,4-dichlorobenzyl)-5-ethynylquinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-benzyl-5-ethynylquinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(2-trifluoromethylbenzyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(4-methoxybenzyl)quinazolin-4(3*H*)-one; 4-((2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-4-oxoquinazolin-3(4*H*)-yl)methyl) benzonitrile; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-ethynyl-3-(2-fluoro-4-methoxybenzyl)quinazolin-4(3*H*)-one; 1-(3-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)prop-2-ynyl)urea; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-fluorobenzyl)-5-(3-(2-(2-methoxyethoxy)ethoxy)prop-1-ynyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-fluoro-3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-ethynyl-quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(3-phenoxyprop-1-ynyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-fluorobenzyl)-5-(6-morpholino-6-oxohex-1-yn-1-yl)quinazolin-4(3*H*)-one; 6-(2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*-(2-methoxyethyl)hex-5-ynamide; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(7-morpholino-7-oxohept-1-yn-1-yl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(5-morpholino-5-oxopent-1-yn-1-yl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-((5-methylpyrazin-2-yl)methyl)-5-(6-morpholino-6-oxohex-1-yn-1-yl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(6-oxo-6-(piperidin-1-yl)hex-1-yn-1-yl)quinazolin-4(3*H*)-one; 6-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N,N*-diethylhex-5-ynamide; 7-(2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)hept-6-ynoic acid; 2-Acetamido-N-(3-(2-((4-amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)prop-2-yn-1-yl)acetamide; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(3-methoxy-5-(trifluoromethyl) benzyl)-5-(6-morpholino-6-oxohex-1-yn-1-yl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-methoxy phenethyl)-5-(6-morpholino-6-oxohex-1-yn-1-yl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(benzo[*b*]thiophen-2-ylmethyl)-5-(6-morpholino-6-oxohex-1-yn-1-yl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-fluoro-3-methoxybenzyl)-5-(6-morpholino-6-oxohex-1-yn-1-yl)quinazolin-4(3*H*)-one; Methyl 3-((2-((4-amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-(6-morpholino-6-oxohex-1-yn-1-yl)-4-oxoquinazolin-3(4*H*)-yl)methyl)benzoate; 2-((4-Amino-3-(3-

hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-((1-methyl-1*H*-pyrazol-4-yl)methyl)-5-(6-morpholino-6-oxohex-1-yn-1-yl) quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(benzofuran-5-ylmethyl)-5-(6-morpholino-6-oxohex-1-yn-1-yl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-((2-methylthiazol-4-yl)methyl)-5-(6-morpholino-6-oxohex-1-yn-1-yl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(6-(4-methylpiperazin-1-yl)-6-oxohex-1-yn-1-yl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(6-(4-morpholinopiperidin-1-yl)-6-oxohex-1-yn-1-yl)quinazolin-4(3*H*)-one; 5-(6-(4-Acetylpiperazin-1-yl)-6-oxohex-1-yn-1-yl)-2-((4-amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)quinazolin-4(3*H*)-one; *N*-(4-(2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)but-3-yn-1-yl)morpholine-4-carboxamide; 5-(6-(4-Acetyl-piperazin-1-yl)-6-oxohex-1-yn-1-yl)-2-((4-amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)quinazolin-4(3*H*)-one; *N*-(4-(2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)but-3-yn-1-yl)morpholine-4-carboxamide; 2-((4-Amino-3-(4-hydroxy-phenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-5-(5-(bis(2-methoxyethyl) amino)pent-1-ynyl)-3-(2-chlorobenzyl)quinazolin-4(3*H*)-one; 6-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*-cyclopentylhex-5-ynamide; 6-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*-(tetrahydro-2*H*-pyran-4-yl)hex-5-ynamide; 6-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d]*pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*-(2-morpholinoethyl)hex-5-ynamide; 2-((4-Amino-3-(4-hydroxy phenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(6-(4-(2-methoxyethyl)piperazin-1-yl)-6-oxohex-1-ynyl)quinazolin-4(3*H*)-one; 6-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*-(2-(dimethylamino)ethyl)hex-5-ynamide; 6-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*-(pyridin-4-yl)hex-5-ynamide; 6-(2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*-(pyridin-4-yl)hex-5-ynamide; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(6-(4-(dimethylamino)piperidin-1-yl)-6-oxohex-1-ynyl)quinazolin-4(3*H*)-one; 6-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*,*N*-bis(2-methoxyethyl)hex-5-ynamide; 6-(2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*,*N*-bis(2-methoxyethyl)hex-5-ynamide; 6-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*-(2-(4-methylpiperazin-1-yl)ethyl)hex-5-ynamide; 6-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*-methyl-*N*-(2-(4-methylpiperazin-1-yl)ethyl)hex-5-ynamide; 6-(2-((4-

Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl) -4-oxo-3,4-dihydroquinazolin-5-yl)-*N*-isopropylhex-5-ynamide; 6-(2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*-isopropylhex-5-ynamide; 6-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*,*N*-dimethylhex-5-ynamide; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(6-oxo-6-(pyrrolidin-1-yl)hex-1-yn-1-yl)quinazolin-4(3*H*)-one; 6-(2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-4-oxo-3,4-dihydroquinazolin-5-yl)-*N*-(pyrrolidin-3-yl)hex-5-ynamide; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(6-(3-(dimethylamino)pyrrolidin-1-yl)-6-oxohex-1-ynyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(6-(3-(dimethylamino)pyrrolidin-1-yl)-6-oxohex-1-ynyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(4-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl) -5-(6-(4-methyl-1,4-diazepan-1-yl)-6-oxohex-1-ynyl)quinazolin-4(3*H*)-one; 2-((4-Amino-3-(3-hydroxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl) -5-(6-(4-methyl-1,4-diazepan-1-yl)-6-oxohex-1-ynyl)quinazolin-4(3*H*)-one, 2-((4-Amino-3-(4-hydroxy-3-methoxyphenyl)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)methyl)-3-(2-chlorobenzyl)-5-(6-morpholino-6-oxohex-1-ynyl)quinazolin-4(3*H*)-one

or a pharmaceutically acceptable salt thereof, including all stereoisomers, tautomers and isotopic derivatives thereof

In one embodiment the combination employs a MEK inhibitor. Examples of MEK inhibitors include: AS703026, CI-1040 (PD184352), AZD6244 (Selumetinib), PD318088, PD0325901, AZD8330, PD98059, U0126-EtOH, BIX 02189 or BIX 02188.

In one embodiment the combination employs an AKT inhibitor. Examples of AKT inhibitors include: MK-2206 and AT7867.

In one embodiment the combination employs an aurora kinase inhibitor. Examples of aurora kinase inhibitors include: Aurora A Inhibitor I, VX-680, AZD1152-HQPA(Barasertib), SNS-314 Mesylate, PHA-680632, ZM-447439, CCT129202 and Hesperadin.

In one embodiment the combination employs a p38 inhibitor, for example as disclosed in WO2010/038086, such as *N*-[4-({4-[3-(3-*tert*-Butyl-1-*p*-tolyl-1*H*-pyrazol-5-yl)ureido]naphthalen-1-yloxy}methyl)pyridin-2-yl]-2-methoxyacetamide.

In one embodiment the combination employs a Bcl-2 inhibitor. Examples of Bcl-2 inhibitors include: obatoclax mesylate, ABT-737, ABT-263(navitoclax) and TW-37.

In one embodiment the combination employs an antimetabolite. Examples of an antimetabolite include: capecitabine (xeloda), fludarabine phosphate, fludarabine(fludara), decitabine, raltitrexed(tomudex), gemcitabine hydrochloride and cladribine.

In one embodiment the therapeutic agent is ganciclovir, which may assist in controlling immune responses and/or tumour vasculation.

In one embodiment one or more therapies employed in the method herein are metronomic, that is a continuous or frequent treatment with low doses of anticancer drugs, often given concomitant with other methods of therapy.

Subgroup B oncolytic adenoviruses, in particular Ad11 and those derived therefrom such as ColoAd1 may be particularly synergistic with chemotherapeutics because they seem to have a mechanism of action that is largely independent of apoptosis, killing cancer cells by a predominantly necrolytic mechanism. Moreover, the immunosuppression that occurs during chemotherapy may allow the oncolytic virus to function with greater efficiency.

In one embodiment the chemotherapeutic agent is administered parenterally.

In one embodiment the chemotherapeutic agent is administered separately to the virus, either temporally or by an alternate method of administration or both. Treatment can be concurrent or sequential.

In one embodiment the cancer treatment is a targeted agent, for example a monoclonal antibody such as bevacizumab, cetuximab or panitumumab or antibody conjugate, such as an antibody drug conjugate, in particular of the type where the antibody or binding fragment is linked to a toxin.

In one embodiment the cancer treatment is an immunotherapeutic agent, for example ipilimumab or other anti-CTLA4, anti-PD-1, anti-PD-L1, or other checkpoint inhibitors, or a cytokine or a cytokine analogue.

Checkpoint inhibitor as employed herein is intended to refer to agents that inhibit signalling from T-cell membrane proteins that act to inhibit or downregulate T-cell activation and function.

In one embodiment the virus is administered in combination with the administration of radiotherapy.

Radiotherapy as employed herein is intended to refer to the medical use of ionising radiation.

Cancer cells are generally undifferentiated and stem cell-like; they reproduce more than most healthy differentiated cells, and have a diminished ability to repair sub-lethal damage. DNA damage is then passed on through cell division; damage to the cancer cells' DNA accumulates, causing them to die or reproduce more slowly.

In one embodiment the radiotherapy is administered concurrently.

In one embodiment the radiotherapy is administered sequentially.

In one embodiment the virus is administered in combination with therapy complimentary to the cancer therapy, for example a treatment for cachexia, such as cancer cachexia, for example S-pindolol, S-mepindolol or S-bopindolol. Suitable doses may be in the range of 2.5mg to 100mg, such as 2.5mg to 50mg per day provided a single dose or multiple doses given as multiple doses administered during the day.

In one embodiment the virus is administered in combination with the administration of one or more prophylactic agents, for example selected from an antipyretic, an antihistamine, an antiemetic, an antidiarrheal, steroid and an analgesic.

Antipyretics include aspirin and non-steroidal anti-inflammatories, for example ibuprofen, naproxen and ketoprofen.

Antihistamines include acrivastine, azalastine, brompheniramine, buclizine, bromodiphenhydramine, carbinoxamine, cetirizine, chlorpromazine, cyclizine, chlorpheniramine, chlorodiphenhydramine, clemastine, cyproheptadine, desloratadine, dexbrompheniramine, deschlorpheniramine, dexchlorpheniramine, dimenhydrinate,

dimetindene, diphenhydramine, doxylamine, ebstine, embramine, fexofenadine, levocetirizine, loratadine, meclizine, mirtazapinem olopatadrine, pheninidamine, pheniramine, phenyltoloxamine, promethazine, pyrilamine, quetiapine, rupatadine, tripelennamine and triprolidine.

Antiemetics include dolasetron, granietron, ondansetron, tropisetron, palonoestron, mirtazapine, domperidone, olanzapine, droperidol, metoclopramide, alizapride, prochloperazine. In some instances antihistamines may be employed as antiemetics.

Antidiarrheals include methylcellulose, attapulgite, bismuth subsalicylate, atropine/diphenoxylate, loperamide and other opioids such as codeine and morphine.

Analgesics include non-steriodal anti-inflammatories, paracetamol, cox-2 inhibitors, opiates and morphinomimetics, such as morphine, codeine, oxycodone, hydrocodone, dihydromorphine, pethidine, buprenorphine, tramadol and the like.

In one embodiment viral treatment is employed in combination with a course of steroids.

Steroids include hydrocortisone, cortisone, prednisone, prednisolone, methylprednisolone, dexamethasone and the like.

Prophylactic as employed herein is intended to refer to preventive medicine or care, for example consisting of measures taken to prevent or ameliorate side effects during or following administration of the virus.

In one embodiment the prophylaxis is administered separately to the virus, either temporally or by an alternate method of administration or both. Treatment can be concurrent or sequential.

In one embodiment additional hydration is provided in combination with the administration of the virus, either concurrently or sequentially.

Additional hydration as employed herein means the patient is supplied with fluids beyond those included in the formulation. This may be any form of suitable liquid, for example, a saline or glucose infusion.

In one embodiment the virus therapy herein is administered in combination with an anti-inflammatory, for example a steroid or non-steroidal anti-inflammatory.

In one embodiment the virus therapy according to the present disclosure is administered in combination with an anti-pyretic.

In one embodiment the viral treatment is administered in combination with hydration therapy, for example intravenous administration of fluids, in particular isotonic saline or glucose.

In one embodiment the method is suitable for treating the patient as an outpatient.

Outpatient as employed herein is a patient who is not hospitalised during the treatment phase, but instead comes to a physician's office, clinic or day surgery for treatment.

In one embodiment there is provided a method of treating a patient with a pharmaceutical formulation described herein comprising ColoAd1 said method comprising the steps of intravenously administering to said patient: a dose on day 1 followed by, a dose on day 3, and a third dose on day 5.

In one embodiment there is provided a parenteral formulation of a replication capable oncolytic subgroup B adenovirus described herein, for use in treatment, such as a tumour

and/or malignancy and/or cancer treatment by administering: a first dose of said formulation described herein, followed by one or more further therapeutic doses thereof

wherein the first dose and further doses are administered within a period of 14 days, in particular as described *supra*.

In one embodiment, there is provided the use of multiple cycles of treatment with a replication capable oncolytic subgroup B adenovirus. A treatment cycle is to be interpreted herein as a series of viral doses administered to a patient over a relatively short period of time after which the patient's response will be assessed. Treatment cycles can be repeated multiple times provided the risk benefit is determined to be in the patient's best interests.

In one embodiment, there is provided a method to assess the suitability of repeated cycles of treatment with a replication capable oncolytic subgroup B adenovirus by determining the level of the specific antiviral titre and comparing it to the pre-treatment titre, such that a titre below a certain percentage of the pre-treatment titre will indicate a positive risk benefit profile for retreatment.

In one embodiment there is provided use of parenteral formulation of a replication capable oncolytic subgroup B adenovirus in the manufacture of a medicament for treatment of a tumour and/or malignancy and/or cancer treatment by employing a treatment regimen herein.

In one embodiment the formulation is employed in the treatment or prophylaxis of metastasis.

In one embodiment, there is provided a formulation presented as a sterile prefilled and packaged syringe of appropriate dose and volume in order to circumvent the need for complex and expensive dose preparation under sterile conditions and using appropriate air handling such as extraction hoods prior to administration to the patient.

In the context of this specification "comprising" is to be interpreted as "including".

Aspects of the invention comprising certain elements are also intended to extend to alternative embodiments "consisting" or "consisting essentially" of the relevant elements.

Any positive embodiment or combination thereof described herein may be the basis of a negative exclusion i.e. a disclaimer.

EXAMPLES

Preclinical potency and selectivity

Table 2 shows the IC_{50} of ColoAd1 on a variety of epithelial cell lines.

Cell Name	Cell Type	IC50
HT-29[1]	Colorectal cancer	0.06
HT-29[2]	Colorectal cancer	0.04
DLD-1[1]	Colorectal cancer	0.35
LS1034[1]	Colorectal cancer	0.21
HCT116[1]	Colorectal cancer	0.02
LS174T[1]	Colorectal cancer	0.57
SW48[1]	Colorectal cancer	0.06
SW403[1]	Colorectal cancer	1
HepG2[2]	Hepatoma	0.05
PC-3[1]	Prostate cancer	0.23
DU145[1]	Prostate cancer	5
Panc-1[1]	Pancreatic cancer	12
MDA231[1]	Breast cancer	0.84
OVCAR-3[1]	Ovarian cancer	3
A549[2]	Lung cancer	2
HMEC[1]	Capillary endothelial	575
HUVE[1]	Umbilical endothelial	50
HUVE[2]	Umbilical endothelial	60
Hepatocytes[2]	Normal liver cells	1050
W138[2]	Fibroblast	350

The IC_{50} of ColoAd1 on a range of epithelial derived cancer and normal cells. The number of ColoAd1 particles required to kill 50% of cell (IC_{50}) was determined in vitro using a standard 6 day MTS assay.

1 Results performed by Schering AG and published in Kuhn et al., 2008.

2 Repeat and additional studies performed by the University of Oxford (unpublished).

Table 3 shows the IC_{50} of ColoAd1 on a variety of non-epithelial cell lines.

Cell name	Cell type	IC50
Colo320DM[1]	neuroendocrine	105
501[2]	melanoma	430
IG37[2]	melanoma	>500
IG39[2]	melanoma	470
U87MG[2]	Glioblastoma	>1000
BBA[2]	Glioblastoma	>1000
BBB[2]	Glioblastoma	>1000
K562[2]	Leukaemia	>1000

The IC_{50} of ColoAd1 on a range of non-epithelial derived cancers. The number of particles required to kill 50% of cell (IC_{50}) was determined in vitro using a standard 5 day MTS assay.

1 Results performed by Schering AG and published in Kuhn et al., 2008.

2 Repeat and additional studies performed by the University of Oxford (unpublished).

Table 4 shows ColoAd1 replication in a variety of normal non-cancer human cell lines.

Human Cell Types	Genomes (cells)	% of control	Genomes (supernatant)	% of control	Successful Re-infection of HT29 Cells
HT29 (+ve control)	3.18E+08	100	4.29E+06	100	YES
hepatocytes	6.83E+03	0.0021	2.47E+02	0.01	No
glomerular endothelial cells	4.78E+02	0.0002	1.22E+03	0.03	No
dermal microvascular cells	1.03E+03	0.0003	1.45E+03	0.03	No
cardiac microvascular cells	5.36E+02	0.0002	1.29E+03	0.03	No
corneal epithelial	2.51E+06	0.7889	7.06E+04	1.65	No
bronchial epithelial	5.48E+05	0.1722	2.88E+04	0.67	No
renal cortical epithelial	3.68E+04	0.0116	3.67E+03	0.09	No
mesangial cells	1.18E+03	0.0004	9.71E+02	0.02	No
intestinal myofibroblasts	9.50E+02	0.0003	1.49E+03	0.03	No
ovarian epithelial	2.38E+06	0.7479	9.99E+04	2.33	No
astrocytes	7.00E+02	0.0002	9.26E+02	0.02	No
aortic smooth muscle Cells	8.65E+02	0.0003	9.42E+02	0.02	No
cardiac myocytes	1.28E+03	0.0004	7.99E+02	0.02	No
renal proximal	8.48E+05	0.2666	4.46E+03	0.10	No
CD34+	1.59E+03	0.0005	2.84E+03	0.07	No
PBMC	1.25E+03	0.0004	1.02E+03	0.02	No

Human cells growing in monolayers *in vitro* were exposed to ColoAd1 for 72 hours. The total number of ColoAd1 genome copies was then determined by qPCR. The data are presented as total genome copies and as a % relative to a carcinoma cell positive control (HT29). The ColoAd1 materials derived from these normal human cells were then tested for viability on HT29 carcinoma cells. In all cases ColoAd1 material so recovered could not be shown to replicate in HT29 cells.

Pre-clinical circulation kinetics

ColoAd1 circulation kinetics were obtained in CD-1 mice. Mice (3 per group) were administered virus particles via the tail vein and circulating genomes in whole blood samples were determined by quantitative PCR (qPCR). ColoAd1 half-life in this model is dose-dependent. At the lower input doses ($1 \times 10^9 – 2 \times 10^{10}$ on multiple dosing days), the mean alpha half-life is 1.8 +/- 0.5 minutes, consistent with values previously reported for other adenoviruses (Green 2004). At higher doses (over 2×10^{11}), saturation of clearance appears to occur, giving rise to longer circulation levels (mean alpha half-life 7.8 +/- 2 minutes). Saturation in the ColoAd1 study described here is reflected via multiple pharmacokinetic parameters

Table 5 demonstrates significant increases in Area Under the Curve (AUC), half-life and percentage of particles retained at the 30 min time-point when ColoAd1 doses were

administered over 2 x 10^{11}. It was noted in particular that the optimal kinetics (AUC, percentage retained particles at 30 min and mean alpha t1/2) were all achieved when three equal high doses were administered as opposed to a low priming dose followed by higher doses. From this data, given that blood circulation times are significantly longer in humans than in mice, it was anticipated that the half-life in humans would be considerably longer and that low priming doses were unlikely to be of value for a sub-group B adenovirus. In addition, in tumour bearing human patients, it was anticipated that replication of this virus in cancer cells, with subsequent release, would also result in further amplification of the virus at later time points. The clinical studies were planned accordingly.

Table 5: ***Circulation kinetics of ColoAd1 following multipleintra-venous injections in CD-1 mice (3 mice per group)***

Study	Dose	Mean Alpha t1/2 (minutes)	Mean AUC ml^{-1} min^{-1}	Mean % of input virus at 30 min
multi-dose study in CD1 mice $1x10^9$ on day 1 then $1x10^{10}$ on days 3 and 5	$1x10^9$ (d1)	2.2	$1.04x10^9$	0.77
	$1x10^{10}$ (d3)	2.6	$9.17x10^9$	0.52
	$1x10^{10}$ (d5)	2.6	$1.20x10^{10}$	0.42
multi-dose study in CD1 mice $1x10^{10}$ on day 1 then $1x10^{11}$ on days 3 and 5	$1x10^{10}$ (d1)	1.2	$1.06x10^{10}$	0.10
	$1x10^{11}$ (d3)	1.3	$9.20x10^{10}$	0.75
	$1x10^{11}$ (d5)	1.2	$1.06x10^{11}$	0.97
multi-dose study in CD1 mice $2x10^{10}$ on day 1 then $2x10^{11}$ on days 3 and 5	$2x10^{10}$ (d1)	1.7	$1.71x10^{10}$	0.29
	$2x10^{11}$ (d3)	3.7	$3.00x10^{11}$	4.52
	$2x10^{11}$ (d5)	4.0	$6.31x10^{11}$	13.62
multi-dose in CD1 mice all three doses at $2x10^{11}$ on days 1, 3 and 5	$2x10^{11}$ (d1)	6.5	$1.09x10^{12}$	27.43
	$2x10^{11}$ (d3)	10.1	$1.21x10^{12}$	28.67
	$2x10^{11}$ (d5)	6.8	$7.875x10^{11}$	10.48

Pre-clinical interaction studies

Virus particles can interact with components of human blood including antibodies, complement and blood cells leading to rapid neutralisation (Lyons 2005, Carlisle 2009). These events are species-specific and cannot be modelled effectively in animals.

To evaluate neutralisation in human blood, ColoAd1 may be incubated in freshly isolated whole human blood from individuals before being applied to permissive cells (HT29 colorectal tumour cells). A range of virus concentrations can be chosen to cover the target clinical dose range (2 x 10^6 to 2 x 10^9 particles per ml of human blood, and assuming a range of human blood volumes). Residual virus potency can be determined by cytotoxicity and compared to virus infection in the absence of incubation in human blood (media alone) and potency levels within a concentration in the range 2 x 10^6 to 2 x 10^9 viral particles per ml is desirable.

The data in **Figure 1** further demonstrate that ColoAd1 was only marginally affected by human blood. Fresh human blood was collected from 9 subjects (A – I) using lithium heparin tubes. ColoAd1 virus particles were added to the blood samples at 10 fold dilutions from $2x10^9$ VP/mL, which reflects a potential equivalent human dose of $1x10^{13}$ assuming that the dose is fully diluted in the total blood volume (assumed to be 5L of blood). After 20 minutes incubation at 37°C, the virus / blood mixture was added to A549 tumour cells growing in a 96-well plate. The proportion of viable A549 cells remaining was then determined after 5 days and plotted as a percentage. The IC_{50} occurs at a level of approximately equivalent to a viral blood concentration of $2x10^6$ VP/mL. This level of virus was thus determined as a minimum target level to achieve in the human clinical studies

Several *in vitro* studies were also conducted of the interaction of ColoAd1 with human blood cells. Fresh blood was obtained from 4 individuals, and erythrocytes, platelets and leukocytes were washed and re-suspended in PBS at physiological cell concentrations ($5x10^9$, $2x10^8$ and $6x10^6$ per mL, respectively) for use in individual experiments. qPCR analysis revealed that over 80% (82% ± 8%) of the ColoAd1 was associated with human blood cells, primarily to erythrocytes and leukocytes. There was no significant difference in the fraction of ColoAd1 bound to blood cells after a 5 or 30 minutes of incubation. Ad5 showed comparatively higher levels of binding to human blood cells (95.5 ± 1.2%) than ColoAd1. Based on relative fluorescence, pre-incubation of ColoAd1-gfp with human blood cells for 30 minutes significantly inhibited (>90%) the infection of SW480 tumour cells, which express only low levels of CD46, the cellular receptor for ColoAd1. Infection of HT29 cells, which express higher levels of CD46, was inhibited to a much lesser extent (~41%), probably because of the higher level of expression of the ColoAd1 receptor on these tumour cells. Finally, the infection of leukocytes, which may express high levels of CD46 and thus serve as a "sink" for ColoAd1, was assessed using ColoAd1-gfp to determine the extent of transgene expression. After 24 hours no evidence of transgene expression was observed in leukocytes. In contrast, previous studies have shown that Ad5 is able to efficiently infect monocytes *in vitro* under the same conditions. In summary, these studies suggest that the interaction of ColoAd1 with cellular blood components is limited and significantly different to that of Ad5. Again, the clinical studies were designed taking this into account.

Pre-clinical Biodistribution of ColoAd1

Biodistribution and clearance of ColoAd1 has been determined in normal mice and transgenic mice expressing the primary virus receptor CD46 (a receptor for group B adenoviruses that is not expressed in normal mice). Following tail vein administration of 1×10^{11} virus particles in normal mice, virus particles were predominantly found in the liver, spleen and lungs after 24 hrs. (Figure 2) indicates viral copies per mg and so these larger organs represent the predominant site of total viral distribution on a percentage basis). Similar distribution to the same target organs was observed in CD46 transgenic mice (Figure 3), showing that the CD46 receptor is not a significant determinant of distribution. The distribution of a non-replicating mutant (ColoAd1CJ132) was identical to that of ColoAd1 indicating replication was not responsible for any of this distribution effect. However, in tumour bearing human patients, it was anticipated that replication of this virus in cancer cells, with subsequent release, would

also result in further amplification of the virus at later time points and so the clinical studies were planned accordingly.

Pre-clinical Viral clearance

To identify the time to complete virus clearance, a long-term particle clearance study was carried out in normal Balbc mice. The dominant organs for virus distribution: liver, spleen and lungs, were chosen for analysis. Here, the total virus particles per organ are recorded as a percentage of the input dose at each time point such that the results are not normalised to organ weight. At 1 hour the majority of the input virus has already been sequestered in the Liver, with less than 5% in the spleen and less than 0.1% in the lungs. At 24 hours post injection virus particles have rapidly been cleared from these organs with less than 1% of the input virus genomes remaining. Beyond day 65 post-injection, no significant levels of virus were detectable in any tissues and levels were not significantly above background. No virus particles could be recovered for any tissues at day 65 post-administration. The kinetics of viral clearance (data presented as % of input dose per organ) are summarised in Figure 4.

CD46 transgenic mice were administered ColoAd1 on a single occasion at a dose of $1x10^{10}$ vp/mouse by tail vein. n = 3 animals per time-point. Genome copies (measured by qPCR) are presented as a percentage of the input dose of genome copies.

Pre-clinical immunogenicity

It is possible that the development of a specific anti-viral immune response may significantly impact the circulation kinetics. To examine this possibility, a group of mice were administered ColoAd1 repeatedly over several months in order to produce a pool of hyper-immune serum. A second group of mice were then passively immunised against ColoAd1 using the hyper-immune serum administered by i.v injection of 10 or 20ul. These mice were then rested for 10 minutes before being administered $5x10^{10}$ ColoAd1 i.v. Blood was collected from each mouse at 2, 10 and 30 minutes post-injection of ColoAd1 then analysed by qPCR. The results are shown in Figure 5 and show that an immune response to ColoAd1 will have a significant impact upon the kinetics and delivery of ColoAd1, thus demonstrating the importance of administering doses before such a response occurs.

Pre-clinical safety and toxicity

Several safety and toxicity studies have been conducted with ColoAd1, including pilot studies in CD-1 and Balb/c mice, CD46 transgenic mice. In a final toxicity study in male and female CD-1 mice, ColoAd1 was administered as three doses given over a 5-day period (on Days 1, 3 and 5) to model the intended clinical dosing regimen. Male and female CD-1 mice received intravenous bolus injections (dose volume = 100µL) of ColoAd1 or the formulation buffer as shown in **Table 6**, which shows the final study design after the unscheduled deaths of two Group 4 males on Day 1 led to the lowering of the dose in that particular group.

Table 6: ***Toxicity Study Design***

Group No.	Main Study No. of Animals		Recovery Group No. of Animals		Satellite Group No. of Animals		Test Item	Total Dosage (vp/animal)
	Males	Females	Males	Females	Males	Females		
1	8	8	8	8	9	9	Vehicle	-
2	8	8	8	8	9	9	ColoAd1	$6.6x10^9$
3	8	8	8	8	9	9	ColoAd1	$6.6x10^{10}$
4	2[a]	-	4[b]	-	2[b]	-	ColoAd1	2.2×10^{11}
	6[c]	-	4[c]	-	7[c]	-	ColoAd1	$3.59x10^{11}$
	-	-	-	-	3[d]	-	ColoAd1	$2.09x10^{11}$
	-	8[e]	-	8[e]	9[e]	9[e]	ColoAd1	$2.09x10^{11}$

a: found dead on Day 1 after treatment; received a single dose of ColoAd1 on Day 1 at 2.2 x 10^{11}vp

b: deemed unsuitable for further dosing and euthanised on Day 2; received a single ColoAd1 dose on Day 1 at 2.2 x 10^{11}vp

c: received a single dose of $2.2x10^{11}$vp ColoAd1 on Day 1, then subsequent doses of $6.96x10^{10}$vp on Days 3 and 5

d: additional males added for cytokine assessment received $6.96x10^{10}$ vp on Days 1, 3 and 5

e:at the top dose all females and all satellite group males received $6.96x10^{10}$ vp on Days 1, 3 and 5

A standard set of safety endpoints, including clinical signs, body weight, plasma cytokine levels, clinical pathology and gross and microscopic examinations were done periodically. A standard list of tissues and organs was collected at necropsy on Days 6 and 17.

No significant clinical signs were observed in males and females in Groups 1, 2 or 3 on any treatment day. Clinical signs of adverse effect were seen in Group 4 after the first dose on Day 1 at doses of both $2.2x10^{11}$ and $6.96x10^{10}$ vp/animal, but - with the exception of one Group 4 male on Day 3 - further adverse clinical signs were not seen. Dose-related body weight loss on Day 2 was seen in all ColoAd1-treated groups except for Group 2 males, though body weight was subsequently unaffected in any treatment group. Haematological and liver function changes, when recorded, occurred over a longer time course but had returned to normal range by the end of the recovery period. In summary, the most significant clinical signs were seen following the first dose, with subsequent doses being better tolerated.

Cytokine responses over time in this study are shown in Figure 6. Elevations of the cytokine MCP-1 were most marked and seen in Groups 3 and 4 on Day 1 at 6 and 24 hours post-first treatment and 6 hours after treatment on Day 5. No consistent elevation in any other cytokine was seen in Group 2 animals. Smaller, but dose-related, increases in IL-6, IFNγ and TNFα were seen in Group 3 and 4 animals only, most commonly at low concentrations compared to MCP-1 and often in only some animals in each dose group, particularly in Group 3.

The cytokine pattern seen in this study is thus consistent with the clinical signs observed, showing that after the first dose, each subsequent dose is better tolerated, even though the doses are equal.

Clinical Studies

At the time of filing, two clinical studies are being conducted to examine the safety and efficacy of ColoAd1 when delivered intravenously to human subjects with metastatic cancer.

The Evolve study (ColoAd1-1001) is a phase I/II clinical study with the phase I dose escalation component conducted in patients with an epithelially derived metastatic tumour (of any origin) and who have no further treatment options. Patients in this phase I dose escalation part of the study have been dosed with three equal doses of intravenous ColoAd1 on days 1, 3 and 5 (48 hours apart). A slow intravenous infusion has been used, and in the early cohorts, each patient was infused with 30ml of viral suspension over a 5 minute period (6ml per minute). Initially each cohort of three patients was dosed at one log increments starting at 1 $x10^{10}$ viral particles per dose until adverse events suggested dose limiting toxicity. Each patient also received a regimen of symptomatic prophylaxis, including supplemental fluids and a set regimen of anti-inflammatories (acetaminophen/paracetamol and ibuprofen). The safety and tolerability of this dosing regimen at each dose level was assessed using physical examinations (including blood pressure, pulse and temperature) and by eliciting all adverse events, as well as by assessing haematology, biochemistry and cytokine profile changes. Viral kinetics and excretion were assessed using regular blood, urine, stool and sputum samples. Efficacy was assessed by serial CT imaging according to objective criteria. Later stages of this study will go on to examine the safety and efficacy of the intravenous Maximum Tolerated Dose (MTD) of ColoAd1 in patients with metastatic colorectal cancer.

A second clinical study (ColoAd1-1002) is a phase 0 “window of opportunity” study to compare intravenous delivery with direct intra-tumoural delivery of ColoAd1 in patients with a newly diagnosed primary (non-metastatic) colorectal tumour. Patients in this study will be dosed pre-surgically with ColoAd1 and the resected tumours will then be examined post-surgically to examine the extent of viral delivery, replication and spread following the two different delivery and dosage regimens. The measures of safety and viral kinetics in this study are broadly similar to those of ColoAd1-1001.

The phase I dose escalation patients in study ColoAd1-1001 were dosed intravenously with ColoAd1 at dose levels up to and including $1x10^{13}$ viral particles, in 7 patient cohorts (i.e. Cohorts 1 to 7) as shown in table 7 below.

Table 7: Dosage regimes for Cohorts 1 to 7 in the phase I dose escalation component of the ColoAd1-1001 clinical study.

Dose cohort number	Dose regime (as repeated on days 1, 3 and 5)					Dose tolerability
	Total number of viral particles (VP) administered per dose	Infusion Volume (ml)	Infusion duration (min)	Infusion rate	Rate of viral particle delivery (VP/minute)	
1	1e10 ($1x10^{10}$)	30 ml	5 min	6 ml/min	2e9 VP/min	Well tolerated
2	1e11 ($1x10^{11}$)	30 ml	5 min	6 ml/min	2e10 VP/min	Well tolerated
3	1e12 ($1x10^{12}$)	30 ml	5 min	6 ml/min	2e11 VP/min	Well tolerated
4	1e13 ($1x10^{13}$)	30 ml	5 min	6 ml/min	2e12 VP/min	Not tolerated (dose limiting toxicity)
5	3e12 ($3x10^{12}$)	30 ml	5 min	6 ml/min	6e11 VP/min	Well tolerated
6	3e12 ($3x10^{12}$)	30 ml	20 min	1.5ml/min	1.5e11 VP/min	Well tolerated
7	6e12 ($6x10^{12}$)	30 ml	40 min	0.75ml/min	1.5e11 VP/min	Well tolerated

The side effect profile of ColoAd1 in this study has included fever, flu like illness, transaminitis, thrombocytopenia, neutropenia, diarrhoea and vomiting. However, at a dose level of $1x10^{13}$ viral particles infused over 5 minutes, the dose was not well tolerated. In particular, two patients suffered dose limiting toxicities (DLT) including a cytokine mediated acute lung injury at this dose and could not tolerate more than a single dose. One patient required steroids to treat this condition. Patients at this dose level also suffered chills, hypertension, pain, transaminitis, PPT prolongation, and D-dimer increases, although all resolved with time. As a result of these toxic effects at this poorly tolerated dose, the dose of ColoAd1 was reduced and then re-escalated using slower infusion rates. Using this strategy, doses of $3x10^{12}$ VP over either 5 (cohort 5) or 20 minutes (cohort 6) and $6x10^{12}$ VP infused over 40 minutes (cohort 7) were all shown to be well tolerated.

This safety data is preliminary at the time of writing, but is supportive of a very similar profile to that seen in mice. However, some patients continued to have fever and asthenia into the second week despite no ongoing dosing, a phenomenon that is consistent with on-going viral replication in the human tumours (a phenomenon that would not be seen in non-tumour bearing mice). The final maximum tolerated dose for humans is thus anticipated to be between 1×10^{12} and 1×10^{13} viral particles administered on days 1, 3 and 5 at an infusion rate of up to $6x10^{11}$ VP/min and with each patient also receiving prophylactic anti-inflammatory medication

and intravenous fluids as per the ColoAd1-1001 protocol. The final optimal dose regimen is now the subject of further confirmatory studies.

Table 8 summarises the key viral pharmacokinetic parameters as measured with qPCR for each patient in the phase I dose escalation component of the ColoAd1-1001 clinical study. These results were largely consistent with the pre-clinical data. In summary there was a dose dependent cMAX and AUC, and the average alpha half-life was approximately 18 minutes although there was an indication of possible saturation kinetics at the higher doses studied.

Table 8: ColoAd1 Pharmacokinetics for Cohorts 1 to 7

Cohort	Patient number *	Dose (viral particles)	Infusion time (min)	α-half life (min)	EOI whole blood viral load (DNA copies/ml)	C_{max} (DNA copies/ml)	AUC (DNA copies L/min)
1	1	$1x10^{10}$	5	28.22	$1.75x10^{6}$	$1.75x10^{6}$	$8.08x10^{10}$
	2	$1x10^{10}$	5	nd	$1.20x10^{6}$	nd	nd
	3	$1x10^{10}$	5	20.2	$9.84x10^{5}$	$9.86x10^{5}$	$4.08x10^{10}$
2	4	$1x10^{11}$	5	4.913	$1.60x10^{7}$	$1.60x10^{7}$	$3.70x10^{11}$
	5	$1x10^{11}$	5	7.218	$7.42x10^{6}$	$7.41x10^{6}$	$2.22x10^{11}$
	6	$1x10^{11}$	5	6.006	$1.73x10^{6}$	$2.67x10^{6}$	$5.54x10^{10}$
3	7	$1x10^{12}$	5	26.66	$1.08x10^{8}$	$1.20x10^{8}$	$4.84x10^{12}$
	8	$1x10^{12}$	5	8.046	$1.26x10^{8}$	$1.26x10^{8}$	$3.42x10^{12}$
	9	$1x10^{12}$	5	6.031	$2.18x10^{8}$	$2.18x10^{8}$	$2.98x10^{12}$
4	10	$1x10^{13}$	5	11.7	$7.30x10^{8}$	$7.26x10^{8}$	$1.95x10^{13}$
	11	$1x10^{13}$	5	7.085	$3.27x10^{8}$	$4.96x10^{8}$	$4.52x10^{13}$
	12	$1x10^{13}$	5	67.42	$3.57x10^{8}$	$3.47x10^{8}$	$2.99x10^{13}$
	13	$1x10^{13}$	5	19.86	$1.27x10^{9}$	$1.23x10^{9}$	$4.82x10^{13}$
5	14	$3x10^{12}$	5	3.746	$4.79x10^{8}$	$4.79x10^{8}$	$7.61x10^{12}$
	15	$3x10^{12}$	5	7.754	$1.31x10^{8}$	$1.31x10^{8}$	$2.54x10^{12}$
	16	$3x10^{12}$	5	11.96	$2.06x10^{8}$	$2.04x10^{8}$	$1.12x10^{13}$
6	17	$3x10^{12}$	20	6.779	$1.10x10^{8}$	$1.10x10^{8}$	$6.84x10^{12}$
	18	$3x10^{12}$	20	9.062	$2.35x10^{7}$	$3.37x10^{7}$	$2.23x10^{12}$
	19	$3x10^{12}$	20	6.151	$2.34x10^{8}$	$2.34x10^{8}$	$1.12x10^{13}$
7	20	$6x10^{12}$	40	46.54	$1.80x10^{8}$	$1.80x10^{8}$	$1.21x10^{13}$
	21	$6x10^{12}$	40	22.72	$5.68x10^{7}$	$5.68x10^{7}$	$4.82x10^{12}$
	22	$6x10^{12}$	40	51.34	$6.16x10^{7}$	$6.37x10^{7}$	$5.56x10^{12}$

* an indicative number for each patient

nd: not determined.

Figure 7 shows the cytokine pattern observed in the cancer patients dosed in the ColoAd1-1001 clinical study in the initial dose escalation phase (up to and including identification of the

dose limiting toxicity). As in the mouse studies, the inflammatory cytokine response seen in humans peaks after the first administration of ColoAd1 and then reduces for subsequent administrations. Interestingly, this initial priming effect of ColoAd1 is not reliably seen at the lower doses but is clearly seen at the higher dose, supporting the assertion that a repeated high dose regimen with equal dose levels may be optimal for the intravenous administration of subgroup B adenoviruses to human cancer patients.

In particular **Figure 7** shows cytokine levels (μg/L) over time in human cancer patients with metastatic solid epithelial tumours after intravenous doses of ColoAd1 administered as a 5 minute infusion of 30ml of viral suspension on Days 1, 3 and 5 (dose points indicated by arrows) at four different dose levels (1e10, 1e11, 1e12 and 1e13 viral particles respectively). Each patient also received prophylactic anti-inflammatory medication and intravenous fluids. Patients at doses up to and including 1e12 tolerated these doses well, but two out of four patients who received the 1e13 dose experienced cytokine mediated dose limiting toxicity and were unable to receive more than a single dose. For the individual patients, raised TNF and gamma interferon levels correlated well with tolerability, but raised IL6 did not (data not shown). It was thus determined that rates up to 2e11 viral particles per minute could be regarded as a well-tolerated infusion rate. Panel A: TNF; Panel B: gamma interferon; Panel C: IL-6.

Figure 8 shows systemic pharmacokinetics of ColoAd1 (Genome copies per mL of blood) in human cancer patients with metastatic solid epithelial tumours. Genome copies measured by qPCR.

In particular, **Figure 8A** shows the mean plasma level of the three patients from the ColoAd1-1001 dosed with a well-tolerated dose (1e12 VP per dose) administered at 2e11 VP/min as equal intravenous doses of ColoAd1 administered as a 5 minute infusion of 30ml of viral suspension on Days 1, 3 and 5 (dose time points indicated by arrows). The trend towards increasing viral concentration peaks with each subsequent dose is typical.

This clearly shows the beneficial effect of the claimed dosing regimen on viral pharmacokinetics, with the peak levels of virus after the second and third dose being increasingly higher than the peak levels of virus after the first dose. This demonstrates the benefit of occupying or removing the non-cancerous viral sinks with the earlier doses. This dose was well tolerated in these three patients.

Figure 8B shows the mean initial pharmacokinetics (viral DNA copies/ml) following the first dose of virus at four different dose levels for patient cohorts 1 to 4 ($1x10^{10}$, $1x10^{11}$, $1x10^{12}$ and $1x10^{13}$ viral particles respectively) of the ColoAd1-1001 clinical study. In each case the dose is administered over 5 minutes and so the viral infusion rate increases from $2x10^{9}$ viral particles per minute at the lowest dose to $2x10^{12}$ viral particles per minute at the top dose. At the top two doses, the blood viral concentration remains above the $2x10^{6}$ viral particles per mL level for a prolonged period. This is the minimum blood level (as predicted from preclinical studies and as shown in Figure 1) to be effective at establishing an infection within the tumour. For the $1x10^{12}$ dose this target level is achieved for between 1 to 2 hours, whilst for the $1x10^{13}$ dose this target level is maintained for over 6 hours. However, the $1x10^{13}$ viral particle dose administered as $2x10^{12}$ viral particles per minute was poorly tolerated, with two patients

suffering acute cytokine mediated dose limiting toxicity and so this dose regimen is not optimal. A dose regimen using a well-tolerated infusion rate (such as $2x10^{11}$ viral particles per minute or slower) may allow the administration of doses up to and possibly higher than $1x10^{13}$ viral particles. Using this data, pharmacokinetic modelling can then be used to show that a dose of $1x10^{13}$ viral particles infused over one hour ($1.67x10^{11}$ viral particles per minute) will maintain viral blood levels above the $2x10^{6}$ target level for three hours or more in most patients.

Figures 9A to 9H shows the pharmacokinetics from patients in the ColoAd1-1001 clinical trial. Patients were administered the first dose of ColoAd1 and the viral load was then assessed with serial blood draws using qPCR. The following treatment regimens were tested:

Figure 9A: 1e10 ($1x10^{10}$) viral particles administered over 5 minutes (Cohort 1); **Figure 9B:** 1e11 ($1x10^{11}$) viral particles administered over 5 minutes (Cohort 2); **Figure 9C:** 1e12 ($1x10^{12}$) viral particles administered over 5 minutes (Cohort 3); **Figure 9D:** 1e13 ($1x10^{13}$) viral particles administered over 5 minutes (Cohort 4); **Figure 9E:** 3e12 ($3x10^{12}$) viral particles administered over 5 minutes (Cohort 5); **Figure 9F:** 3e12 ($3x10^{12}$) viral particles administered over 20 minutes (Cohort 6); and **Figure 9G:** 6e12 ($6x10^{12}$) viral particles administered over 40 minutes (Cohort 7).

Each curve represents the viral blood levels measured for an individual test subject per unit time prior to receiving ColoAd1 and up to about 6 hours following treatment.

Adverse side effects were first observed in patients when viral blood levels exceeded a threshold of about 3e8 viral genomes per mL.

Hence, a range of about 3e7 to 3e8 viral genomes per ML was determined to be an ideal therapeutic range and that a regimen which maintains the viral blood levels within this range for as long as possible would maximise viral blood levels, whilst minimising toxic side effects.

As can be seen from the pharmacokinetic curves, Figure 9G (6e12 particles administered over 40 minutes) shows a particularly suitable profile with the viral blood levels maintained within the therapeutic range for the longest.

Figure 10 shows a comparison between C_{Max} levels when the same dose is administered to patients in the ColoAd1-1001 study as either a slow infusion or a fast infusion. Cohorts 5 and 6 were both administered a total dose of $3x10^{12}$ viral particles but for cohort 5 the dose was infused over 5 mins (fast infusion) and cohort 6 the dose was infused over 20 mins (slow infusion). It can be seen that slowing the infusion rate can effectively result in less variation of the C_{Max} and a lower mean C_{Max} level, and so limiting the infusion rate of a Group B adenovirus is thus relevant when higher C_{Max} levels are associated with toxicity.

Figure 11 shows MCP1 levels (μg/L) over time in human cancer patients after intravenous doses of ColoAd1 administered on Days 1, 3 and 5 (dose points indicated by arrows) in ther ColoAd1-1001 clinical study. The graph shows the comparison between the different patient cohorts (1 to 7) that were each administered with a different dose regimen shown in Table 7.

Measurements of MCP1 levels (μg/L) were taken at the following time points: at 0 hours, 6 hours, 12 hours, 24 hours, 48 hours, 54 hours, 60 hours, 72 hours, 96 hours, 102 hours, 108 hours, 120 hours, 168 hours and 336 hours. This human data reflects a similar pattern to that

seen in mice with reducing levels of MCP1 after each dose for every dose tested, thus supporting the specific benefits of the claimed dose regimen.

The cytokine pattern seen in Figure 12 is consistent with the cytokine patterns previously observed in the mice studies (see Figure 6), showing that after the first dose, each subsequent dose is better tolerated even when each of the 3 doses is equal.

Studies showing that infection of tumours by type B adenovirus can be established by doses of viral particles administered by IV

Figure 12 shows the replication cycle typical for an adenovirus. Adenovirus structural proteins e.g. Hexon, which makes 90% of the virus capsid, are only expressed late during infection after replication has occurred. The proteins are then transported back to the nucleus for assembly. The nucleus thus has the highest concentration of hexon and other structural proteins during replication. Therefore, nuclear hexon staining can be used for the quantification of adenovirus and as a marker for cells that have been successfully infected with ColoAd1.

In the ColoAd1-1002 clinical study, patients with primary (non-metastatic) colorectal tumours received ColoAd1 by either intratumoural (IT) delivery or intravenous (IV) delivery. In the IT group, the virus was administered via a colonoscope at a dose of up to 1e8 VP as multiple injections (actual dose was dependent upon the tumour size). In the IV group the dose was 1e12 VP administered as an infusion over 5 min on days 1, 3 and 5. Then, 7 to 14 days after the first dose of ColoAd1, the primary tumour was resected and was sent for pathological examination including immunohistochemical (IHC) staining for ColoAd1 hexon.

Sections of formalin-fixed, paraffin-embedded human tumour samples were analyzed for the presence of virus using an anti-hexon antibody (ab8251).

Staining was carried out under using a validated assay with a Ventana Benchmark Ultra. Strong nuclear staining indicates the presence of hexon undergoing capsid assembly.

Isotypes controls were processed at the same time and under the same conditions.

Figure 13 shows a transmission EM image of a colorectal cancer cell line infected in vitro with ColoAd1.

Figure 14A shows cell staining images of a tumour sample which has been infected with ColoAd1 after intratumoural injection (IT), and then stained for Hexon. As can be seen, there is substantial nuclear staining in carcinoma cells whereas there is no nuclear staining in stroma cells. **Figure 14B** shows the corresponding isotype control. Together these slides show that ColoAd1 infects tumour cells selectively, without infecting normal cells following direct intratumoural delivery.

Figure 14C shows a cell staining image of a tumour sample which has been infected with ColoAd1 after intravenous (IV) administration, and then stained for Hexon. As can be seen, there is substantial nuclear staining in carcinoma cells whereas there is no nuclear staining in stroma cells. **Figure 14D** is the corresponding isotype control.

Therefore, these images provide clear evidence that ColoAd1 can be selectively delivered to tumour cells in a manner that is equivalent to intratumoural delivery when using the claimed intravenous dosing regimen.

Example 2 Drug Combination

ColoAd1 virus replication in the presence of 320 clinically approved compounds or compounds in development was assessed in the colon carcinoma cell line, HT-29. HT-29 cell were seeded at a density of 3.0e4 cells per well in 96 well plates and incubated at 37°C, 5% CO_2. After 4-6hrs incubation virus and drug compound mixtures prepared in cell media were diluted onto the cells to give final doses of 10 ColoAd1 virus particles per cell (ppc) and 0.1μM of drug compound. The cells were incubated for 18hrs and then the total virus genomes in the cells were assessed by qPCR. The relative fold change in ColoAd1 replication, compared to ColoAd1 virus alone, is plotted for all compounds in Figure 15. The inset shows an increase in virus replication after 18 hrs in the presence of microtubule inhibitors and a decrease in virus replication in the presence of topoisomerase inhibitors.

The effect of paclitaxel or cisplatin treatment on ColoAd1 efficacy in a tumour model was assessed in an IP model of ovarian cancer. SCID mice were implanted with 2.5e6 luciferase-expressing SKOV-3 human ovarian carcinoma cells. Tumour burden was assessed by luciferase expression. Mice were imaged on day 5, on the day before each set of treatments and at least every 5-7 days for the duration of the study. All ColoAd1 treatments were carried out using 5e9 virus particles delivered by intra-peritoneal injection and in the combined treatment groups, paclitaxel (0.4mg) or cisplatin (0.04mg) was delivered the day after virus treatment. Disease progression was assessed by luciferase imaging using an IVIS imaging system. Images of the relative luminescence in mice dosed via IP injection with either PBS (A), paclitaxel (B), ColoAd1 (C) or paclitaxel and ColoAd1 (D) is shown in Figure 16 and the relative luminescence tracked over time for each dosing group is shown in Figure 17. The relative luminescence in mice dosed via IP injection with either PBS (Group 1), ColoAd1 then cisplatin (Group 2), cisplatin then ColoAd1 (Group 3) or Paclitaxel and ColoAd1 (Group 4) are shown in Figure 18. Dosing schedules are detailed in the Figures 17 and 18.

CLAIMS

1. A method of treating a human patient, said method comprising the steps of:
systemically administering multiple doses of a parenteral formulation of a replication capable oncolytic adenovirus of subgroup B in a single treatment cycle,
wherein the total dose given in each administration is in the range of $1x10^{10}$ to $1x10^{14}$ viral particles per dose, and
wherein each dose of virus is administered such that the rate of viral particle delivery is in the range of $2x10^{10}$ particles per minute to $2x10^{12}$ particles per minute.

2. A method according to claim 1, wherein the total dose given in each administration is in the range of $1x10^{12}$ to $1x10^{13}$ viral particles per dose, for example the total dose given in each administration is in the range of $3x10^{12}$ to $9x10^{12}$ viral particles per dose, in particular wherein the total dose given in each administration is $6x10^{12}$ viral particles per dose.

3. A method according to claims 1 or 2, wherein the period between each dose administration is in the range of 6 hours to 72 hours, such as 48 hours.

4. A method according to any one of claims 1 to 3, wherein the multiple doses are 2, 3, 4, 5, 6 or 7 doses in a single treatment cycle.

5. A method according to any one of claims 1 to 4, wherein the treatment cycle is a period of 14 days or less, such as 7 days or 5 days.

6. A method according to any one of claims 1 to 5, wherein each dose of virus is administered such that the rate of viral particle delivery is in the range of $1x10^{11}$ viral particles per minute to $3x10^{11}$ viral particles per minute, such as $2x10^{11}$ viral particles per minute or $1.67 x10^{11}$ viral particles per minute, per dose, in particular each dose of virus is administered such that the rate of viral particle delivery is $1.5x10^{11}$ viral particles per minute, per dose.

7. A method according to any one of claims 1 to 6, wherein $1 x 10^{13}$ viral particles are administered over a 60 minute period per dose or wherein $6x10^{12}$ viral particles are administered over a 40 minute period per dose.

8. A method according to any one of claims 1 to 7, wherein the plasma level of virus after administration of the second and optionally subsequent doses reaches a level of at least $2x10^{6}$ viral particles per mL.

9. A method according to claim 8, wherein the plasma level of viral particles is maintained for 15 minutes or greater, for example 20, 30, 40, 50 or 60 minutes.

10. A method according to any one of claims 1 to 9, wherein the volume of the formulation administered is 100mL or less, for example 30mLs or less, such as about 3mLs or 5mLs.

11. A method according to any one of claims 1 to 10, wherein a first dose is administered on day 1 and the further therapeutic doses are administered every second day thereafter, for example wherein the first dose is administered on day 1 and the further therapeutic doses are administered on days 3 and 5.

12. A method according to any one of claims 1 to 11, wherein the adenovirus is a chimeric adenovirus, for example wherein the chimeric adenovirus is ColoAd1.

13. A method according to any one of claims 1 to 12, wherein the virus is administered in combination with the administration of any other anti-cancer therapy including a chemotherapeutic agent, an immunotherapeutic agent such as an antibody or fragment thereof, a small molecule inhibitor such as a kinase inhibitor or an mTOR inhibitor, radiotherapy, radio-isotope therapy or any combination thereof.

14. A method according to any one of claims 1 to 13, wherein the virus is administered in combination with the administration of one or more prophylactic agents, for example selected from an antipyretic, an antiemetic, a steroid and an analgesic.

15. A method according to any one of claims 1 to 14 for the treatment of a tumour, for example a solid tumour, such as a colorectal malignancy.

16. A method according to any one of claims 1 to 23 wherein the first and therapeutic dose is in the range 10^9 to 10^{14} viral particles.

17. A method according to any one of claims 1 to 16, wherein the adenovirus is formulated to at least partially avoid the patient's immune system.

18. A method according to any one of claims 1 to 17, wherein the adenovirus contains a transgene.

19. A method according to any one of claims 1 to 17, wherein the treatment results in the necrosis of the cancer cells accompanied by an increase in plasma cytokine levels.

20. A method of treating a patient with a pharmaceutical formulation comprising ColoAd1 comprising the steps of intravenously administering to said patient
a first therapeutic dose on day 1 followed by
a second therapeutic dose on day 3 and
a third therapeutic dose on day 5.

21. A method of treating a human patient, said method comprising systemically administering multiple doses of a parenteral formulation comprising a replication

capable oncolytic adenovirus of subgroup B in a single treatment cycle, wherein the total dose given in each administration is in the range $1x10^{10}$ to $1x10^{13}$ viral particles, and is administered over a period of 1 minute to 90 minutes, for example wherein the total dose given in each administration is $6x10^{12}$ viral particles, and is administered over a period of 40 minutes.

22. A replication capable oncolytic adenovirus of subgroup B for use in treatment of a human patient by systemically administering multiple doses of a parenteral formulation comprising the adenovirus in a single treatment cycle, wherein the total dose given in each administration is in the range $1x10^{10}$ to $1x10^{14}$ or $1x10^{10}$ to $1x10^{13}$ viral particles, and is administered over a period of 1 minute to 90 minutes.

23. A parenteral formulation of a replication capable oncolytic adenovirus of subgroup B, for use according to claim 22, wherein the total dose administered is in the range 1 $x10^{10}$ to 1 $x10^{14}$ viral particles per dose and each dose of virus is administered such that the rate of particle delivery is in the range of $2x10^{10}$ particles per minute to $2x10^{12}$ per minute.

24. Use of a replication capable oncolytic adenovirus of subgroup B for the manufacture of a medicament use in treatment of a human patient by systemically administering multiple doses of a parenteral formulation comprising the adenovirus in a single treatment cycle, wherein the total dose given in each administration is in the range $1x10^{10}$ to $6x10^{12}$ viral particles, and is administered over a period of 1 minute to 90 minutes.

25. Use of a glass or plastic syringe with an internal volume in the range of 3 to 50ml, said syringe containing a parenteral formulation comprising $1x10^{10}$ to $6x10^{12}$ viral particles of a replication capable oncolytic adenovirus of subgroup B, wherein the formulation is sterile and was filled into the syringe under aseptic conditions, for use in treatment, in particular for use in the manufacture of a medicament which is capable of injection or intravenous infusion to a human subject, for example according to any one of claims 1 to 21.

26. ColoAd1 for use in treating ovarian cancer, for example administering a therapeutically effective amount of ColoAd1 to a patient with ovarian cancer, for example employing a dosing regimen described herein.

27. A combination therapy comprising oncolytic type B adenovirus, such as ColoAd1, and a chemotherapeutic agent which does not interfere with the adenovirus activity, such as viral replication *in vivo*.