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USE OF STREPTOMYCES WYEC 108 TO CONTROL PLANT PATHOGENS

FIELD OF THE INVENTION

The present invention relates to a new strain of *Streptomyces* bacteria that is capable of inhibiting the growth of soil borne plant pathogens and enhancing plant growth.

BACKGROUND OF THE INVENTION

Fungal phytopathogens are a cause of severe economic losses in the agricultural and horticultural industries. Many different types of fungal phytopathogens have been described: these pathogens cause plant diseases such as damping-off, white-rot, brown-rot and root-rot. Such diseases can kill emerging seedlings, reduce plant vigor and adversely affect crop yields.

To minimize fungal infections, bedding-plant nurseries may grow seedlings in steam sterilized or chemically treated soils. However, such treatments also remove beneficial microorganisms from the soil, including microorganisms that would normally compete with soil fungi. In such cases, if a fungal pathogen is accidentally introduced, it may spread rapidly and produce widespread disease.

In agricultural settings, soils infested with phytopathogenic fungi may be unsuitable for growing certain crops. For example, soybean production in Michigan and in other soybean growing states is often severely limited by *Phytophthora* root rot caused by the fungus *Phytophthora megasperma* (Filinow and Lockwood, 1985). Species of *Pythium* fungi are widespread in soils in parts of California, Washington State and Idaho. *Pythium ultimum* is the most common pathogenic species encountered and is associated with pre- and post-emergence damping-off of seedlings. This species is a serious pathogen of wheat, peas and chickpeas and other crop plants grown in these soils and in soils in other states and other countries (Trapero-Casas et al., 1990; Stanghellini and Hancock, 1970; Kraft and Burke, 1971; Westerdal et al., 1988). The use of chemical agents to control fungal phytopathogens is often not practical due to high costs, lack of efficacy and the emergence of resistant strains of the fungi. Additionally, the use of chemical fungicides is not desirable from an environmental viewpoint.

It is an object of the present invention to provide a biological control means of reducing fungal pathogen infection of plants.

SUMMARY OF THE INVENTION

The foregoing object has been achieved by the isolation of a number of actinomycete bacteria that are shown to be effective in inhibiting the growth of fungal phytopathogens. In particular, one of the isolated actinomycete bacteria, herein named *Streptomyces* WYEC 108 (also referred to herein as WYEC 108), is shown to exhibit strong antagonism towards a wide range of fungal plant pathogens, including pathogens that cause the plant diseases commonly known as damping-off, root rot, white rot and brown rot. Thus, one aspect of the present invention is a biologically pure culture of *Streptomyces* WYEC 108.



5 The present invention also sets forth various compositions suitable for treating plant seeds or plant roots with *Streptomyces* WYEC 108. Such compositions are useful to reduce the susceptibility of plants to fungal infection and to enhance the growth of treated plants. In a preferred embodiment, such compositions comprise a biologically pure culture of *Streptomyces* WYEC 108 and a delivery medium. In particular embodiments, the delivery medium may comprise alginate gel, peat moss, sand or corinnal. In one embodiment, the present invention encompasses a delivery medium which comprises peat moss, sand and corinnal together with *Streptomyces* WYEC 108. In another embodiment, the delivery medium comprises at least 10⁷ colony forming units per gram of delivery medium.

10 In another embodiment, the present invention encompasses alginate gel pellets containing *Streptomyces* WYEC 108. Such pellets can be added directly to the roots of growing plants or to horticultural or agricultural soils to reduce damage to plants caused by phytopathogenic fungi.

15 The present invention also encompasses methods for reducing the susceptibility of a plant to fungal infection. In one embodiment, this method comprises delivering *Streptomyces* WYEC 108 to the roots of a plant. In another embodiment, the method comprises immersing seeds in a composition that contains *Streptomyces* WYEC 108 and thereafter planting the coated seeds in a suitable growth medium. In this method, a suitable composition comprises alginate gel containing *Streptomyces* WYEC 108.

20 BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 comprises scanning electron photomicrographs showing the chain spores (top) and the surface of spores (bottom) of *Streptomyces* WYEC 108.

25 FIG. 2 shows chickpea plants grown in soil infected with *Pythium* species. Plants on the right germinated from seeds treated with *Streptomyces* WYEC 108. Plants shown on the left germinated from untreated seeds.

DETAILED DESCRIPTION OF THE INVENTION

30 The present invention describes the isolation of a number of actinomycete strains from soils. A number of these strains are shown to be effective in reducing the effects of fungal pathogens on plants, including lettuce, chickpea and pepper. In particular, the present invention sets forth the isolation of a strain herein referred to as *Streptomyces* WYEC 108. Strain WYEC 108 is shown to exhibit strong antagonism towards a wide range of fungal phytopathogens, including pathogens that cause pre- and post-emergence damping off of seedlings, root rot, brown rot and white rot. As such, strain WYEC 108 is particularly suitable as a biocontrol agent that can be used to protect plants against infection by these phytopathogens. Thus, *Streptomyces* WYEC 108 is useful in methods for reducing the susceptibility of plants to fungal infection. As such, plants treated with *Streptomyces* WYEC 108 will show reduced effects of fungal infection. Fungal infection of susceptible untreated plants affects certain growth characteristics of such plants. For instance, untreated plants



exposed to fungal pathogens may show significant reductions in plant height, plant biomass and crop yield compared to plants not exposed to the fungal pathogen. In preferred embodiments of the present of the invention, plants treated with *Streptomyces* WYEC 108 and subsequently exposed to the fungal pathogen will show less severe reductions in plant height, plant biomass and crop yield than untreated plants exposed to the fungal pathogen. In more preferred
 5 embodiments, plants treated with *Streptomyces* WYEC and exposed to the fungal pathogen will show growth characteristics similar to the untreated, unexposed plants. In most preferred embodiments, plants treated with *Streptomyces* WYEC 108 and exposed to the fungal pathogen will show growth characteristics superior to the untreated, unexposed plants.

10 Strain WYEC 108 colonizes plant roots in the presence of competition from rhizosphere microflora. Strain WYEC 108 is shown to enhance the growth of lettuce plants growing in steam sterilized soil and pepper plants growing in an agricultural field.

Also encompassed by this invention are means of producing vegetative cells or spores of strain WYEC 108 suitable for incorporation into a delivery medium. The
 15 composition comprising the vegetative cells and spores of WYEC 108 and the delivery medium is shown to have a long shelf life and to be suitable for delivering strain WYEC 108 to plants.

Materials and Methods

Bacterial Growth Media

All bacterial growth media were prepared using distilled water and sterilized by
 20 autoclaving prior to use. All bacterial samples were handled using standard aseptic laboratory techniques to maintain purity.

YGM (yeast extract/glucose/mineral salts) medium comprised 0.6% (wt/vol) yeast extract (Difco Laboratories, Detroit, Michigan), 1.0% (wt/vol) glucose, and phosphate mineral salt solution (5.3 g of Na_2HPO_4 , 1.98 g of KH_2PO_4 , 0.2 g of $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 0.2 g of
 25 NaCl , 0.05 g $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$, plus 1.0 ml of trace elements (Pridham and Gottlieb, 1948) per liter of deionized H_2O ; pH 7.1 to 7.2). The solution of trace elements consisted of 0.64 g of $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$, 0.11 g of $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$, 0.79 g of $\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$, 0.15 g of $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ in 100 ml of distilled water.

WYE (water/yeast extract/agar) medium, modified from Reddi and Rao (1971)
 30 contained yeast extract (Oxoid, 0.25 g/l) as the sole carbon and nitrogen source, and agar (Oxoid, 18.0 g/l). The medium was buffered to pH 7.2-7.4 with K_2HPO_4 (0.5 g/l).

WYEC (water/yeast extract/cellulose/agar) was WYE agar to which a thin overlay agar was added. The overlay agar contained 0.25 g/l of cellulose (Solka Floe, Sigma Chemical Co.) and 18.0 g/l agar in distilled water.

35 CYD (casamino acids/yeast extract/dextrose agar) medium contained casamino acids (Difco: 0.5 g/l), yeast extract (Oxoid or Difco: 0.8 g/l), D-glucose (0.4 g/l), K_2HPO_4 (2.0 g/l; pH 7.2-7.4), and 18.0 g/l agar in distilled water.



YCRD (casamino acids/yeast extract/dextrose/agar; modified from Reddi and Rao (1971) contained yeast extract (Oxoid, 0.3 g/l), casamino acids (Difco, 0.3 g/l), D-glucose (0.3 g/l), and agar (Oxoid, 18.0 g/l). The medium was buffered with K_2HPO_4 (2.0 g/l).

5 CYPC (cellulose/yeast extract/peptone/compost extract/agar) contained cellulose (Solka Flock, Sigma Chemical Co; 5.0 g/l), yeast extract (1.0 g/l), peptone (Oxoid, 1.0 g/l), phosphate buffer (K_2HPO_4 , 0.75 g/l), agar (18.0 g/l), and compost extract (100 ml/l) replacing 100 ml of distilled water in the medium. It was poured directly and not used as an overlay agar.

10 MSSC (mineral salts/starch/casein/agar; Turhan, 1981) contained a mineral salts solution consisting of NaCl (2.0 g/l), $MgSO_4 \cdot 7H_2O$ (0.05 g/l), $CaCO_3$ (0.02 g/l), $FeSO_4 \cdot 18H_2O$ (0.01 g/l), and KNO_3 (2.0 g/l), plus organic constituents including soluble starch (10.0 g/l) and casein (0.3 g/l), plus agar (18.0 g/l). The medium was buffered with K_2HPO_4 (2.0 g/l).

15 Sporulation medium (ATCC Medium #5) contained yeast extract (1.0 g/l), beef extract (1.0 g/l), tryptose (2.0 g/l), $FeSO_4$ (0.01 g/l), glucose (10.0 g/l), and agar (15.0 g/l). The medium was adjusted to pH 7.2 prior to autoclaving. (17th Edition ATCC Catalogue of Bacteria and Bacteriophages).

CYG medium contained Casamino acids (acid hydrolysate) (5.0 g/l), yeast extract (5.0 g/l) and glucose (10.0 g/l) in distilled water, adjusted to pH 7.1-7.2.

20 Delivery medium (comprising sand/commeneal/water or peat moss/sand/commeneal in ratios as set forth below) was sterilized by steam sterilization prior to use. Sterilization was typically performed by autoclaving 3 times, each time by 90 minutes.

Harvesting of bacterial growth

25 For mycelial growth of *Streptomyces* WYEC 108, one liter Erlenmeyer flasks containing 500 ml YGM medium (pH 7.1-7.2), were inoculated with 20 ml of stock culture (prepared as described in Example II) and incubated with shaking at 250 rpm at 30°C for three days. Mycelia were then harvested by centrifugation at 5,000 rpm for 10 minutes. Alternatively, mycelia were harvested by permitting the culture to stand until mycelia and spores settled to the bottom of the Erlenmeyer flask. Supernatant media was then decanted off and the concentrated suspension of mycelia and spores was used directly to inoculate 30 delivery medium.

Cells and spores were also produced by growth on solid medium (for example sporulation agar). Mycelia and spores were harvested from sporulation agar by scraping the surface of the agar into distilled water. This suspension of spores and mycelia was then mixed directly into the delivery medium.

35 For the production of spores of *Streptomyces* WYEC 108 two liter Erlenmeyer flasks containing 1,200 ml YGM medium were each inoculated with 50 ml of stock culture and incubated with shaking at 250 rpm at 30°C for 12-18 days. Spores were harvested by centrifugation at 9,000 rpm for 10 minutes.



Fungal Pathogens

- Pythium ultimum* Pulk. was obtained from the culture collection of the Department of Microbiology and Crop Protection at Horticulture Research International, Worthing Road, Litchamption, West Sussex BN17 6LP, United Kingdom. White-rot fungi *Phanerochaete chrysosporium* and *Corticium verticillium*, brown-rot fungi *Postia placenta*, *Calderonius jamae*, and *Gloeophyllum trabeum*; soil born fungal pathogens *Rhizoctonia solani*, *Fusarium sambucinum*, *Geotrichum candidum*, and *Verticillium dahliae* came from the culture collection of professor Don L. Crawford, Department of Bacteriology, University of Idaho, Moscow, ID. *Pythium irregulare*, *Phytophthora capsici*, *Phytophthora cinnamomi*, *Phytophthora parasitica*, *Sclerotinia cepivorum*, and *Sclerotinia sclerotiorum* came from the culture collection of Dr. Wesley Chum, Department of Plant Soil Pathology Science, University of Idaho, Moscow, ID. *Fusarium oxysporum* came from the culture collection of Dr. Arthur D. Partridge, Department of Forest Resources, University of Idaho, Moscow, ID. All cultures were maintained on potato dextrose agar or corn meal agar and grown at 25 °C. These strains were identified as "pathogens" when obtained, but were not retested for their pathogenicity.

Bioassay Soil

- For use in bioassays, soil naturally infested with *Pythium ultimum* was collected from several sites in the Palouse region near Moscow, Idaho. This soil was collected from the top 15cm from fields that had been cropped with wheat and pea in the previous two seasons. The soil population of *Pythium* species was determined as follows: A soil dilution of 1.0 g air-dried soil in 50 ml sterilized distilled-water was thoroughly mixed with a Vortex tube mixer. A 0.1 ml sample of the well mixed dilution was placed in small droplets on 3-day-old 2% water agar plates (Shanghalini and Hancock, 1970). Plates were incubated at 25 °C and read periodically using a low power (x10) dissecting microscope with fluorescent illumination to determine the identity and numbers of *Pythium* species present. Colonies on each plate were checked after 12, 48 and 72 hours of incubation, before the final population was estimated. Identification was based upon the morphological characteristics of fungal mycelium of *Pythium* species under microscope and the growth pattern on 2% (w/v) water agar plates. Fungal colonies of a pure culture growing on 2% (w/v) water agar served as a control for visual identification purposes (Shanghalini and Hancock, 1970; Stenz et al., 1980).

- Examination of this soil indicated that the population densities of *P. ultimum* and *P. irregulare* were 354 ± 15 and 194 ± 11 cfu/g of air-dried soil at the time of seeding (Spring, 1992), respectively. Population density of other *Pythium* species was 57 ± 9 cfu/g of air-dried soil. *P. ultimum* and *P. irregulare* were the most prevalent species isolated from the collected soil.



EXAMPLE I

Isolation of Actinomycete Strains Exhibiting Antagonism Towards Fungal Phytopathogens

Actinomycete strains were isolated from four rhizosphere-associated and four non-rhizosphere-associated soil samples. These strains were then tested for utility as inhibitors of fungal phytopathogens.

Isolation of Actinomycetes

Actinomycete isolates were isolated from 8 different soils by serial-dilution/spread-plate technique. Dilutions (10^{-5} to 10^{-7}) were plated onto various agar isolation media. The composition of these media is set forth in "Materials and Methods" above. Actinomycete isolates were designated according to the isolation medium on which they were isolated. For example, WYEC 108 was isolated on WYEC medium and YCED 9 was isolated on YCED medium. In general, such media are poor in organic carbon, which effectively controls eubacterial and fungal growth and aids in isolating the slower growing actinomycetes. Since WYE and YCED agars were particularly effective isolation media, they were used predominantly. Dilution plates were incubated at 25°C for 4 to 10 days to allow the actinomycetes to sporulate, and then colonies were picked and streaked onto WYE or YCED agar plates for purification. Pure colonies were transferred from these plates to YCED agar slants or CYD agar slants, incubated at 25°C or 37°C until sporulated, and stored at 5°C until used. Stock cultures were transferred every 3 to 4 weeks.

Soils

(i) Non-rhizosphere-associated soils.

Samples (100 to 200 g) of soil were taken from the top 7.5 to 10 cm of the soil profile at 4 sites in the United Kingdom, including a cultivated rose garden (Soil S1) in Rustington, West Sussex; between rows in a wheat field (Soil S2) at the Horticulture Research International (H.R.I.) farm in Littlehampton, West Sussex; a forest soil (Soil S6) from the Wynd Cliff hardwood forest reserve, South Wales; and a grassland (Soil S7) sometimes grazed by sheep at Hastings Hill, South Downs, West Sussex. These soils were considered non-rhizospheric, although they did contain plant roots in varying amounts.

(ii) Rhizosphere-associated soils.

Rhizosphere-associated soil samples from 4 places were prepared essentially by the method of Miller et al. (1990). Soil 3 (S3) was associated with the roots of a dandelion plant (*Taraxacum officinale*) in a rose garden at H.R.I. in West Sussex, England. Soil (S5) was associated with wheat roots and was taken from the same field as S2, a wheat field at the H.R.I.'s farm in Littlehampton, West Sussex. Soil 4 (S4) was also associated with wheat roots, but the soil was from a field on Bignor Hill along the South Downs Way, West Sussex. Soil 8 (S8) was associated with the roots of linseed plants and was taken from a field adjacent to the



S7 sampling site, a grassland sometimes grazed by sheep at Hastings Hill, South Downs, West Sussex.

Actinomycetes isolated from the soils could be divided into those isolated from non-rhizosphere- (S1, S2, S6, and S7) or rhizosphere-associated (S3, S4, S5, and S8) soils. All soils were characterized for their moisture content by drying 3 g (wet weight) samples (three replicates) at 100°C for 58 h and then reweighing them. Soil pH was determined by thoroughly mixing a 1:1 soil:water slurry, allowing the solids to settle for 2 h, and taking the pH of the supernatant solution. After collection, soils were stored at 4°C until used (24 to 48 h). The isolates were confirmed to be actinomycete strains by visual examination which showed that the colonies formed by these strains were typical actinomycete colonies (hard and leathery with an aerial mycelium containing spores). Further confirmation of the identity of these strains as actinomycetes was obtained by microscopy.

Determination of pH range for growth.

Each actinomycete isolate was tested for its ability to grow at pH 5.5 to 8.0. Cultures were spot-inoculated onto plates of CYD agar, buffered to pH 5.5, 6.0, 6.5, 7.0, and 8.0 with combinations of K_2HPO_4 and KH_2PO_4 buffers at 100 mM concentration. The final pH of each medium was adjusted to its final value just prior to autoclaving. Cultures were checked for growth after 5 to 7 days incubation at 25 or 37°C. Plates were evaluated visually for little or no observable growth ($\pm$), some growth (+ or ++), or excellent growth (+++).

As shown in Table I, the rhizosphere-associated soils gave almost twice as many isolates as the non-rhizosphere-associated soils. Each isolate was tested for growth on CYD agar media ranging from pH 5.5 to pH 8.0. All isolates grew from pH 6.5 to 8.0. Only 9 failed to grow at pH 6.0, while S7 (21%) failed to grow at pH 5.5. Of those that grew at pH 5.5, growth varied from poor to excellent depending upon the isolate. The ability of the isolates to sporulate strongly on CYD agar was also determined by visual and microscopic observation of colonies after 5-10 days of incubation.



TABLE I

		Selective medium			Totals
	Soil	Soil pH	YCED	WYE	
Non-rhizosphere-associated soils					77
5	1	7.5	8	16	24
	2	5.4	5	6	11
	6	7.2	20	8	28
	7	7.4	13	1	14
Rhizosphere-associated soils					140
10	3	7.0	17	22	39
	4	7.6	32	33	65
	5	6.5	11	15	26
	8	7.3	8	2	10
Total isolates			114	103	217

15

In Vitro Antagonism Assay

Eighty-two isolates were chosen on the basis of their ability to grow well and sporulate strongly on CYD agar.

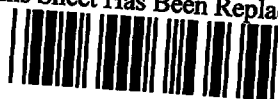
To test the ability of these isolates to inhibit the growth of *P. ultimum*, an in vitro plate assay was used. Each actinomycete was streak-inoculated on corn meal agar (CMA) plates, to one side of center. The culture was incubated at 25 °C for about 8 days or until the culture had sporulated. A CMA agar block (0.5 cm²) containing actively growing *P. ultimum* mycelium was then aseptically placed in the center of the plate. Incubation was continued for 96 h. After 48 and 96 h the plate was examined for inhibition in the growth of *P. ultimum*. Inhibition was indicated when *P. ultimum* mycelial growth in the direction of the actinomycete colony was retarded or prevented. The results of this test are shown in Table II.

25

TABLE II

	Culture	Source (soil)	Growth at pH 5.5 (+ or -)	Antagonism observed ^a	
				48 hr	96 hr
	Antagonistic (at 96 hr)				
30	WYEC 108	8	+	+++	+++
	YCED 1	1	+	++	++
	YCED 9	2	+	+++	+++
	YCED 35	4	+	+	+
35	YCED 48	4	+	+	+
	YCED 95	7	+	+	+
	YCED 106	7	+	++	+++
40	WYE 21	4	+	+	+
	WYE 22	4	+	+	+

This Sheet Has Been Replaced



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	Culture	Source (soil)	Growth at pH 5.5 (+ or -)	Antagonism observed ^a	
				48 hr	96 hr
5	WYE 30	4	-	+	+
	WYE 31	4	+	+	+
	WYE 78	1	+	++	++
	WYE 88	1	+	+	+
	WYE 90	6	+	+++	+++
	WYE 91	6	+	+++	+++
	WYE 97	1	+	++	++
10	MSSC 1	2	+	+	+
	MSSC 2	2	+	+	+
Nonantagonistic (at 96 hr)					
15	WYE 6	1	+	+	-
	WYE 9	3	+	-	-
	WYE 11	3	+	+	-
	WYE 12	4	+	+	-
	WYE 13	4	+	-	-
	WYE 20	4	+	-	-
	WYE 23	3	+	-	-
20	WYE 27	4	-	-	-
	WYE 28	3	+	-	-
	WYE 29	4	+	+	±
	WYE 34	3	+	-	-
	WYE 35	3	+	-	-
	WYE 38	4	+	-	-
	WYE 42	3	+	-	-
25	WYE 43	4	+	-	-
	WYE 45	3	+	-	-
	WYE 47	3	+	-	-
	WYE 53	4	+	-	-
	WYE 54	3	+	-	-
	WYE 56	3	+	-	-
	WYE 68	5	+	-	-
35	WYE 69	6	+	-	-
	WYE 73	6	+	-	-
	WYE 75	2	+	-	-
	WYE 77	2	+	±	±
	WYE 84	2	+	-	-
	WYE 85	2	+	-	-
	WYE 93	1	+	-	-
40	WYE 94	1	+	-	-
	WYE 120	8	+	-	-
	WYE 121	7	+	-	-
45	YCED 11	1	+	-	-
	YCED 15	4	+	-	-
	YCED 16	4	+	-	-
	YCED 17	3	+	-	-
	YCED 25	4	+	-	-
	YCED 28	4	+	-	-
	YCED 29	3	+	-	-
50	YCED 30	3	+	-	-
	YCED 31	3	+	-	-
	YCED 32	4	+	-	-



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Culture	Source (act)	Growth at pH 5.5 (+ or -)	Antagonism observed ^a 48 hr 96 hr
YCSD 41	4	+	-
YCSD 44	4	+	±
YCSD 54	4	+	-
YCSD 56	4	+	-
YCSD 62	5	+	-
YCSD 64	5	+	-
YCSD 71	5	+	-
YCSD 73	5	+	-
YCSD 85	6	-	-
YCSD 88	5	+	-
YCSD 93	7	+	-
YCSD 96	7	+	-
YCSD 98	8	+	-
YCSD 105	7	+	-
WYEC 101	4	+	-
WYEC 104	8	+	-
WYEC 107	7	-	±
WYEC 111	8	+	-
WYEC 113	8	+	±
WYEC 116	8	+	-
WYEC 118	7	+	-
CYFC 2	6	+	-
CYFC 5	6	+	-

^a Inhibition of *P. ultimum* defined as typical growth less abundant and growth retarded slightly on area of plate to the side where the actinomycete had been grown.

++ very strong inhibition with zone of inhibition ≥ 2.0 cm
 ++ strong inhibition with zone of inhibition ≥ 1.0 cm
 + growth definitely retarded, with obvious zone of inhibition near colony
 ± minor inhibition of *P. ultimum* (typical growth less abundant and growth retarded
 - no inhibition

35 After 96 hr, five isolates (WYEC108, YCSD9, WYEC91, WYEC90, and YCSD106) showed very strong antagonism towards *P. ultimum*, four (YCSD1, YCSD106, WYEC97, and WYEC95) showed strong antagonism, and ten others showed weak antagonism. The remaining isolates were either not antagonistic, or only very weakly so. The cultures that clearly inhibited growth of *P. ultimum* were about equally divided between those isolated from rhizosphere-associated soils and non-rhizosphere-associated soils.

40 Seventy of the isolates that grew at pH 5.5 were also tested for their *in vitro* antagonism against the white-rot fungus *Phanerochaete chrysosporium* on cornmeal agar (CMA). Thirteen of the isolates showed some degree of antagonism of the white-rot fungus as shown in Table III. The degree of antagonism varied from very strong (++) to relatively weak (+) as defined by the size of the inhibition zone. Five of the cultures that showed antagonism against *P. chrysosporium* (WYEC108, WYEC78, WYEC90, YCSD9, and MSSC2) were further tested on CMA against an additional white-rot fungus (*Coriolus versicolor*) and two

45



types of brown-rot fungi (*Postia placenta* and *Gloeophyllum trabeum*). Four isolates (MSSC2, YCED9, WYE90, WYEC108) showed very strong antagonism towards the above-mentioned white- and brown-rot fungi. One isolate, WYEC78, showed strong antagonism only against the two white-rot fungi.

5

TABLE III

	Culture	Source (soil)	Growth at pH 5.5 (+ or -)	Antagonism observed ^a	
				48 hr	96 hr
	Antagonistic WYEC 108 ^b	8	+	+++	+++
10	WYE 22	4	+	++	++
	WYE 78 ^c	1	+	+	+
	WYE 90 ^b	6	+	+++	+++
	WYE 97	1	+	++	++
15	YCED 9 ^b	2	+	+++	+++
	YCED 29	3	+	+	+
	YCED 41	4	+	+	+
	YCED 48	4	+	+	+
	YCED 95	7	+	++	+
20	CYPC 2	6	+	++	++
	CYPC 5	6	+	+	+
25	MSSC 2 ^b	2	+	+++	++

^aInhibition of *P. chrysosporium* defined as hyphal growth less abundant and growth retarded slightly on area of plate to the side where the actinomycete had been grown.

+++ very strong inhibition with zone of inhibition ≥ 2.0 cm

++ strong inhibition with zone of inhibition ≥ 1.0 cm

30 + growth definitely retarded, with obvious zone of inhibition near colony

± minor inhibition of *P. chrysosporium* (hyphal growth less abundant and growth retarded)

- no inhibition

^bInhibition of *Coriolus versicolor*, *Postia placenta* and *Gloeophyllum trabeum* in addition to *P. chrysosporium*.

35 ^cInhibition of *Postia placenta* and *Gloeophyllum trabeum* in addition to *P. chrysosporium*.

In Vivo Bioassay to Determine Activity of Actinomycete Isolates on Lettuce Seedlings

The biocontrol assay procedure of Lynch et al. (1991, 1992) was used to assay 12
40 of the isolates for their effect on the germination and outgrowth of lettuce (*Lactuca sativa*)
seeds.

For control plants, 9-cm-diameter plastic pots were filled with lettuce potting mix
and tamped down with a petri dish. Ten lettuce seeds were placed on top, lightly pressed into
the potting mix, and lightly covered with loose mix. The pots were then placed on trays in a
45 bed of water, and incubated in the dark at 20 to 22°C until germination was evident



(approximately 3 days). The trays were then transferred to a capillary matting in a greenhouse maintained at 15 to 25°C and watered as required. The numbers of germinated seedlings in each pot were counted periodically for up to 18 days.

5 For plants treated with the actinomycete isolates, pots were filled with potting mix inoculated with the spores of a specific actinomycete. Potting mix was inoculated with spores from CYD stock plants, to an average level of 10^8 to 10^9 cfu/g (dry weight) of potting mix. CfU/g of potting mix was determined by viable counts on CYD agar plates at the time of inoculation. Lettuce seeds were then planted as above, covered with a small amount of inoculated potting mix, and thereafter treated similarly to controls.

10 For plants treated with a specific actinomycete and the damping-off fungus *Pythium ultimum* (strain PaMXL, Lynch et al., 1991), the mix was also inoculated with the fungal pathogen at ca. 200 sporangia/g (dry weight) of mix. The procedure of Lynch et al. (1991, 1992) was used to produce the pathogen sporangia and inoculate the potting mix. Sporangial counts were made with a haemocytometer, and the pathogenicity of the *P. ultimum* strain was confirmed before use by passage through lettuce.

15 In all treatments, pots were prepared in replicates of five. In the glasshouse, pots were set in a random block arrangement surrounded by "guard plants"; guard plants served to provide uniformity of conditions and to act as a buffer. Eighteen days after planting, the plants were harvested, the emergence of the final stand was measured, and wet and dry weights (aboveground leaves and stem) were determined. Plant wet and dry weights were recorded as
20 total mg biomass per pot, and as average mg biomass per plant. Values were reported as averages of the five replicates $\pm$ standard deviation. Thus, each value was based upon 50 seeds planted per treatment (five replicate pots planted with 10 seeds each). Percent seed germination and final plant stand values were calculated similarly.



TABLE IV

Actino-mycete strain	Concentration $\times 10^5$ cfu g ⁻¹ dry compost	Number of Healthy plants per pot (n)	Shoot weight				
			Fresh		Dry		
			-P1	+P1*	-P1	+P1	
Unknown-Inoculated Control		9.8	3.6	1.34	0.75	0.074	0.034
YCED 85	5.19	10.0	6.6 ^a	1.44	0.98	0.087	0.054 ^a
WYE 27	3.02	10.0	5.0	1.54	0.72	0.090	0.039
YCED 64	3.21	9.8	5.6 ^a	1.44	0.91	0.084	0.057 ^a
WYEC 107	2.88	9.8	5.6 ^a	1.68 ^a	0.81	0.096 ^a	0.041
WYE 41	1.18	10.0	4.2	1.45	0.73	0.087	0.047
YCED 106	1.30	10.0	5.4 ^a	1.49	0.69	0.088	0.040
YCED 71	5.25	10.0	6.2 ^a	1.42	0.85	0.082	0.050
WYE 88	1.44	10.0	6.2 ^a	1.42	0.73	0.082	0.042
MSSC 1	2.13	9.8	4.4	1.38	1.16 ^a	0.079	0.060 ^a
WYE 21	9.56	10.0	6.6 ^a	1.49	1.09 ^a	0.091	0.059 ^a
WYE 30	18.90	10.0	5.0	1.53	0.86	0.088	0.046
WYE 28	0.18	10.0	3.6	1.39	0.90	0.079	0.049

P1* *P. ultimum*: 200 sporangia g⁻¹ dry potting mix.

Where n = the average number of healthy plants per pot out of a total of 5 replicate pots of 10 plants.

* Significantly different from the corresponding control value at the $P = 0.05$ level.

Degrees of freedom = 100

In the absence of pathogens (denoted as "-P1" in Table IV) there were no significant differences between actinomycete-inoculated and control pots (without actinomycetes) in the percentage of seeds germinating and producing healthy plants. Seed germination and outgrowth averaged $\geq 98\%$ in all cases. However, the presence of the actinomycetes sometimes delayed seed outgrowth by 1 to 3 days (data not shown). Similarly, in the absence of pathogens there were generally no significant differences between actinomycete-inoculated and control pots in plant shoot weight, measured either as fresh or dry weight. The exception was for pots inoculated with WYEC 107, where the presence of the actinomycetes significantly enhanced the plant biomass yield.

In the presence of pathogen (+P1), the number of healthy plants per pot 20 days after planting averaged 3.6 of 10.0 in the control pots, as compared to 9.8 of 10.0 for control pots without pathogen. In the presence of specific actinomycetes, however, some significant improvements in the number of healthy plants were seen in pathogen-inoculated pots. Seven of the 12 actinomycetes (YCED 85, YCED 64, WYEC 107, YCED 106, YCED 71, WYE 88, AND WYE 21) significantly improved the yield of healthy plants. Of these, YCED 85, YCED 64, and WYE 21 also significantly improved plant dry weight yields relative to pathogen-only controls. WYE 21 also significantly improved fresh weight yields relative to pathogen-only controls. One strain that did not significantly improve the yield of healthy plants (MSSC 1) did improve both fresh and dry weight plant shoot yields (Table IV, columns 5 and 7). Thus,



MSSC 1 may prove useful in protecting plants against lower doses of *P. ultimum* than used here.

EXAMPLE II

Isolation of Streptomyces WYEC 108

5 Strain WYEC 108 was identified as *Streptomyces* species on the basis of the morphological characteristics of the genus *Streptomyces*, as defined by Bergey's *Manual of Systematic Bacteriology* (1986). WYEC 108 is a filamentous bacterium that produces chains of spores in an aerial mycelium. As described above, *Streptomyces* WYEC 108 was isolated as one of a number of actinomycete strains isolated from soil taken from eight different sites in Great Britain. Along with other actinomycetes, *Streptomyces* WYEC 108 was isolated by the serial-dilution/spread-plate technique from rhizosphere soil associated with the roots of linseed plants in a field on Hastings Hill, South Downs, West Sussex, England. Dilutions (10^5 to 10^7) of this soil were plated onto isolation agar medium WYB. Dilution plates were incubated at 25 °C for 4 to 10 days to allow actinomycete colonies to grow and sporulate. Then the colonies were picked and streaked onto WYEC agar plates for purification. Pure colonies of WYEC 108 were transferred from these plates to CYD agar slants, incubated at 25 °C until sporulated, and stored at 4 °C until used. Stock cultures were transferred every 3 to 4 weeks.

Identity of Streptomyces WYEC 108

As described above, the actinomycete strains isolated were examined for their ability to grow well and sporulate strongly on CYD agar. Subsequently, a number of isolates were tested for ability to inhibit the in vivo growth of the phytopathogen *Pythium ultimum*. The isolates were also tested for in vitro antagonism against the white rot fungi *Phanerochaete chrysosporium* and *Coriolus versicolor* as well as the brown rot fungi *Postia placens* and *Gloeophyllum trabeum*. As a result of these tests, one of these strains, herein referred to as *Streptomyces* WYEC 108 was selected based on its favorable characteristics.

Colonies of Streptomyces WYEC 108 grown on Casamino Acid/Yeast

Extract/Dextrose (CYD) plates were examined by scanning electron microscopy. The sample preparation was as follows:

30 (1) colonies of *Streptomyces* WYEC 108 on CYD plates were covered with a 1.5% solution of glutaraldehyde in 0.2 M sodium cacodylate buffer and fixed for at least two hours; (2) the colonies were completely washed (2x) with 0.2 M sodium cacodylate buffer by aspirating the previous liquid with a pipette and replacing it with the buffer solution. Care was taken to insure that the sample did not dry out; (3) the colonies were then removed by taking "plugs" of agar, containing the colonies; (4) the "plugs" were placed into individual mesh containers and carried through dehydration in 100% ethanol (2x) (G.T. Baker Inc., Phillipsburg, NJ);

35 (5) the samples were then dried in a critical point drying "bomb" and mounted on individual specimen stubs using colloidal silver conducting paint. They were then coated with 60/40 gold-palladium and observed with a scanning electron microscope. FIG. 1 is scanning electron



photomicrographs showing the chains of spores (top) and the surface of spores (bottom) of *Streptomyces* WYEC 108. The surface of spores was observed to be relatively smooth.

- Various physiological characteristics of strain WYEC 108 were also determined: strain WYEC 108 did not produce melanin or H_2S on Peptone-Yeast-Iron Agar and Peptone-Iron Agar (Difco Lab., Detroit, Michigan), respectively. The color of the spore mass produced by *Streptomyces* WYEC 108 on CYD plates was gray. This strain did not grow at 45 °C. *Streptomyces* WYEC 108 may belong to the species *Streptomyces hyalans* as defined by Bergey's Manual of Determinative Bacteriology (1986). Accordingly, this organism may be referred to as *Streptomyces hyalans* WYEC 108. For brevity, the organism is referred to herein as *Streptomyces* WYEC 108, or simply WYEC 108. (21)

ATCC Accession Number.

- A deposit of *Streptomyces* WYEC 108 was made under the terms of the Budapest Treaty with the American Type Culture Collection (ATCC), Rockville, MD on June 29, 1993. This strain has been designated ATCC Accession No. 55445.

- 15 Preparation of Stock Cultures of *Streptomyces* WYEC 108

For short-term use, *Streptomyces* WYEC 108 was inoculated on CYD agar or sporulation agar slant at 25 °C until sporulated and stored at 4 °C until used. For long-term storage of cultures, 10 ml spore suspensions were prepared by suspending spores from a single agar slant or plate in 10ml sterile YGM medium. This spore suspension was then used to inoculate 250 ml Erlenmeyer flasks containing 100 ml YGM (yeast extract/glucose/mineral salts) medium. The flasks were then incubated with shaking at 250 rpm for 32-36 hours at 30 °C to provide a standard inoculum.

Samples from the YGM grown standard inoculum were also used for making glycerol cultures suitable for long-term storage at -70 °C and for lyophilization.

- 25

EXAMPLE III

In Vitro Antagonism of Fungal Phytopathogens by *Streptomyces* WYEC 108

- The ability of *Streptomyces* WYEC 108 to inhibit the growth of a number of selected fungal phytopathogens was measured in terms of colony growth inhibition. *Streptomyces* WYEC 108 was streak-inoculated to one side of the center of corn meal agar (CMA) (Difco Lab., Detroit, Michigan) plates. Inoculated plates were incubated at 25 °C for about 8-12 days until the cultures had sporulated. Sporulation was detectable as a mass of grey aerial mycelium and spores by observation with the naked eye. Sporulation was observed by phase contrast microscopy ($\times 1,000$). A 5-mm-diameter CMA agar disc containing actively growing mycelium of a specific fungal phytopathogen was taken from the leading edge of a fungal culture and aseptically placed in the center of the agar plate. The plates were incubated at 25 °C until the test fungus reached the edge of a control plate not containing *Streptomyces* WYEC 108. Inhibition of fungal growth was quantified by determining the ratio of radial



growth of fungal pathogen under the influence of *Streptomyces* WYEC 108 versus growth alone on the control plates. Percent inhibition of fungal growth was recorded after 48, 96, and 192 hr incubation depending on selected pathogenic fungus. The bioassay was replicated on five plates, inhibition was measured separately, and recorded as an average $\pm$ standard deviation.

5 The results of these in vitro bioassays are shown in Table V. This data shows that *Streptomyces* WYEC 108 exhibits very strong antagonism towards a wide range of fungal plant pathogens, including damping off (*Pythium ultimum*), root rot (*Pythium ultimum*, *Rhizoctonia solani*, *Fusarium solani*, and *Phytophthora cinnamomi*), white rot (*Phanerochaete chrysosporium* and *Coriolus versicolor*), brown rot (*Postia placenta* and *Gloeophyllum trabeum*) and leaf and stem rot (*Sclerotinia* sp.) fungi.

TABLE V

Fungal pathogens	Percent inhibition $\pm$ standard deviation ^a Antagonism observed ^b	
	48 hr	96 hr
15 <i>Pythium irregulare</i>	100 $\pm$ 0.0	100 $\pm$ 0.0
<i>Pythium ultimum</i>	100 $\pm$ 0.0	100 $\pm$ 0.0
20 <i>Rhizoctonia solani</i>	100 $\pm$ 0.0	84 $\pm$ 0.0
<i>Fusarium oxysporum</i>	26 $\pm$ 2.5	26 $\pm$ 3.6
<i>Fusarium sambucinctum</i>	44 $\pm$ 2.4	35 $\pm$ 2.4
<i>Fusarium solani</i>	36 $\pm$ 2.5	19 $\pm$ 2.5
25 <i>Phytophthora capsici</i>	100 $\pm$ 0.0	100 $\pm$ 0.0
<i>Phytophthora cinnamomi</i>	100 $\pm$ 0.0	100 $\pm$ 0.0
<i>Phytophthora parasitica</i>	100 $\pm$ 0.0	100 $\pm$ 0.0
30 <i>Sclerotinia cepivorum</i>	100 $\pm$ 0.0	95 $\pm$ 1.5
<i>Sclerotinia sclerotiorum</i>	100 $\pm$ 0.0	100 $\pm$ 0.0
<i>Phanerochaete chrysosporium</i>	100 $\pm$ 0.0	100 $\pm$ 0.0
<i>Coriolus versicolor</i>	100 $\pm$ 0.0	100 $\pm$ 0.0
35 <i>Postia placenta</i>	100 $\pm$ 0.0 ^c	100 $\pm$ 0.0 ^d
<i>Caldariomyces fumago</i>	100 $\pm$ 0.0 ^c	100 $\pm$ 0.0 ^d
<i>Gloeophyllum trabeum</i>	100 $\pm$ 0.0 ^c	100 $\pm$ 0.0 ^d
40 <i>Geotrichum candidum</i>	47 $\pm$ 2.1	45 $\pm$ 2.1
<i>Verticillium dahliae</i>	73 $\pm$ 2.0 ^c	59 $\pm$ 2.0 ^d

^a Values based upon averages of individual values of five replicated plates. Individual values were determined by separately measuring the mycelial growth from each plate.

45 ^b Inhibition of fungal pathogens defined as hyphal growth of pathogens under the influence of *Streptomyces* WYEC 108 versus growth alone on the control CMA plates.

^c and ^d % inhibition at 96 hr and 192 hr incubation, respectively.



EXAMPLE IV

Use of *Streptomyces* WYEC 108 as a Seed Treatment

The efficacy of *Streptomyces* WYEC 108 cells to protect plants against phytopathogens was determined by applying strain WYEC 108 to ungerminated chickpea seeds and then planting these seeds in soil infested with the fungal phytopathogens *P. ultimum* and *P. irregulare*.

Extracellular metabolites produced by strain WYEC 108 were extracted from cultures by ether extraction. The effect of these metabolites on fungal infection of emerging chickpea seedlings was also determined.

10 Growth of *Streptomyces* WYEC108

For growth of strain WYEC 108 cells, one liter Erlenmeyer flasks containing 500 ml YGM (pH 7.1-7.2) were inoculated with 20 ml of stock culture and incubated with shaking at 250 rpm at 30°C for 3 days for the production of cell mass. For production of antifungal metabolites, one liter Erlenmeyer flasks containing 500 ml CYD (pH 7.1-7.2) were inoculated with 20 ml of stock culture and incubated with shaking at 250 rpm at 30°C for 7 days.

15 Treatment of Seeds with *Streptomyces* WYEC108 and Antifungal Metabolites

A mycelial suspension of *Streptomyces* WYEC 108 was harvested by centrifugation at 5,000 rpm for 10 minutes from a 500 ml 3-day-old YGM liquid culture. The harvested mycelia were resuspended in 200-300 ml of sterilized 3% (w/v) sodium alginate solution to a culture density of 1.0-1.2 x 10⁴ cfu/ml. Chickpea seeds were then added to the well mixed cell-alginate suspension and the seeds were transferred one by one into sterilized 0.25 M CaCl₂ in distilled water. These seeds were used in the biocontrol assay described below.

Antifungal metabolites produced by *Streptomyces* WYEC 108 were obtained in purified form as follows. A 7-day-old 500 ml culture was filtered to remove cells and subsequently extracted with 150 ml ether using an extraction funnel. The ether was then removed by vacuum evaporation and the resulting extracts were redissolved in 1.5 ml distilled water. This solution was then filter sterilized through a sterile 0.45 µm filter and was added into 10 ml 3% (w/v) sodium alginate solution. The purified antifungal metabolites may be used to protect plants from fungal infection. Preferably, the antifungal metabolites will be purified so as to be substantially free from WYEC 108 cells. However, a preparation of WYEC 108 cells and/or spores together with antifungal metabolites is also expected to be effective against fungal phytopathogens. The antifungal metabolite-alginate suspension was applied as described above to chickpea seeds for use in the biocontrol assay.

In Vivo Biocontrol Assay

35 Soil naturally infested with *P. ultimum* and *P. irregulare* is described in "Materials and Methods", above. This agricultural soil was used in this vivo biocontrol assays. Soil pH was determined to be pH 5.6 by thoroughly mixing a soil:water slurry (1:1), allowing the solids to settle



for 2 h, and taking the pH of the supernatant solution. The soil was chopped, mixed thoroughly and then placed in seedling pots (10 cm deep x 10 cm diameter).

The in vivo biocontrol assay was carried out by planting ungerminated chickpea seeds treated with either *Streptomyces* WYEC 108 or the antifungal metabolites in the infested soil.

5 Untreated seeds planted in the same soil were used as a control. This procedure involved the following steps:

1) One cm of peat moss was placed in the bottom of each pot to prevent loss of soil while still providing for aeration and drainage.

2) Seedling pots were filled with the infested soil.

10 3) The soil was then watered to saturation from the bottom. After saturation of the soil surface, untreated and treated chickpea seeds were placed on the soil and covered 1.5-2.0cm deep with the same soil. The topping was allowed to become wet by capillary action from the column of wet soil beneath. Ten seeds were planted in each of three replicate seedling pots. No fertilizer was added to the soil. To minimize drying and prevent crusting the pots were covered
15 with clean plastic until seedling emergence. Additional water was sprayed on the top of the pots as needed, beginning after seedling emergence. Experiments were performed in a greenhouse at 15-30°C with a 12 hr light and 12 hr dark cycle photoperiod (16,000 lux).

Emergence counts of chickpea seedlings were made periodically, and final emergence counts were taken after 20 days. Emergence data were reported as the average for each treatment.

20 The ability of *Streptomyces* WYEC 108 to act as a biocontrol agent was based on total emergence, plant height, and plant fresh weight, as compared to the control plants grown from untreated seeds with the biocontrol agent. The results of this biocontrol assay are shown in Table VI.

TABLE VI

25	Treatment	Damping-off (%)		Emergence (%)	Height (cm)	Fresh weight (g/plant)
		Preemergence	Postemergence			
	Control	86.7	6.6	6.7	4.3 ^x	0.34 ^x
	<i>Streptomyces</i> WYEC 108 ^b	36.7	0.0	63.3	11.3	1.05
	3% Alginate ^d	83.3	10.0	6.7	4.1 ^x	0.32 ^x
30	Antifungal metabolites ^c	63.3	3.3	33.3	8.9	0.66

^aIn 3% alginate, coated on seeds.

^bNot containing WYEC 108.

35 ^xMeans so marked within column were not significantly different at the $P = 0.05$ level.

Both *Streptomyces* WYEC 108 cells and the antifungal metabolites produced by these cells reduced *Pythium* damping-off of the chickpeas.



Plants showed vigorous growth, when seeds were coated with *Streptomyces* WYEC 108 cells. There was a significant reduction in height and fresh weight of the plants that emerged from the control (untreated) chickpea seeds as compared to those of the plants germinated from seeds coated with *Streptomyces* WYEC 108 cells. Emergence of untreated chickpea seeds was extremely reduced (6.7% emergence) because of seed rot and preemergence damping-off disease caused by *P. ultimum* when seeds were planted in soil naturally infested with *P. ultimum* and *P. irregulare*. In contrast, emergence of seeds treated with *Streptomyces* WYEC 108 cells before seeding was 63.3%. Seeds treated with alginate alone did not show increased emergence. Symptoms typical of *Pythium* root rot, including root hair loss and root discoloration, were evident in harvested chickpea roots germinated from control seeds, but these symptoms were absent from plants grown from seeds treated with *Streptomyces* WYEC 108 cells. In the controls, damage to chickpea was mainly in the form of seed decay, and preemergence damping-off. Chickpea seedlings that did emerge and grow were stunted, and their roots were severely infected with *P. ultimum*. FIG. 2 shows a side by side comparison of chickpea plants taken from the biocontrol assay. The control plant, shown on the left, which was germinated from untreated seeds showed extensive root infection and lack of secondary roots and root hairs whereas the plants emerging from seeds coated with *Streptomyces* WYEC 108, shown on the right, showed good growth and normal formation of secondary roots and root hairs.

Emergence of chickpea seeds treated with antifungal metabolites in the form of ether soluble metabolite was higher (33.3%) than that of control seeds (6.7%), but lower than that of seeds coated with *Streptomyces* WYEC 108 cells (63.3%). Plants that emerged from seeds treated with antifungal metabolites showed vigorous growth, longer root, and a higher density of root hair development as compared to control plants.

25

EXAMPLE V

Effects of *Streptomyces* WYEC 108 on Root Infection and Seed Rot by *Pythium ultimum*

Chickpea seedlings grown as described in Example IV in soil naturally infested with *Pythium ultimum* and with and without pretreatment with WYEC 108 were examined to determine the effects of strain WYEC 108 on *P. ultimum* infection.

30

Root infection by *P. ultimum* was studied with control (untreated) chickpea seedlings harvested after 20-days of growth. *P. ultimum* causing root rot was also isolated from the rotted roots of these control plants. Isolation of *P. ultimum* was accomplished by first washing soil from the chickpea roots (rootlets and root hairs) with tap water and rinsing them twice with sterile distilled-water. Decolorized and rotted roots were cut out aseptically with a razor blade and placed on 3-day-old 2% water agar plates. The plates were incubated for 24-48 hrs at room temperature and observed under phase contrast microscope (x40). *P. ultimum*

35



growing from the infected chickpea roots was subcultured to fresh 2% water agar and then identified as described previously (Ingaram and Cook, 1990).

Seed rot by *P. ultimum* was studied with rotted control seeds harvested after 20-days. A part of a rotted seed was aseptically placed on 3-day-old 2% water agar plates with a sterilized toothpick, and incubated for 24-48 hrs at room temperature. The plates were examined as described above.

Pythium was observed to infect the roots of untreated chickpea plants growing in the naturally-infested soil. *P. ultimum* was the most prevalent species isolated from rotted seeds and roots. *P. irregularis* was less commonly observed.

Root colonization by *Streptomyces* WYEC 108 was examined in roots of 20-day-old chickpea plants that germinated from seeds treated with the *Streptomyces* WYEC 108 call alginate suspension described in Example II. These plants were grown in soil naturally infested with *P. ultimum* and *P. irregularis* as described in Example II. The plants were removed from the seedling pots and gently washed with tap water to remove adhering rhizosphere soil. Then they were rinsed with sterile distilled-water. Root samples were prepared for microscopy by placing a portion of roots on a glass slide, adding a drop of methylene blue, and then covering this with a coverslip. Prepared samples were then observed with a phase contrast microscope (x1000).

Streptomyces WYEC 108 present on seeds came into contact with the emerging roots. As the roots elongated, the *Streptomyces* was carried along with the elongating root hairs and tips. *Streptomyces* WYEC 108 was observed to have extensively colonized the main root, secondary roots, root hairs and tips. Plants that emerged from seeds coated with *Streptomyces* WYEC 108 were healthier, had longer roots and root hairs were present in greater density than in control plants emerged from seeds not coated with the WYEC 108. This difference was clearly associated with colonization of the roots by the biocontrol agent. Roots colonized by the biocontrol agent did not show any symptoms of root diseases.

Streptomyces WYEC 108 showed excellent root colonization in the presence of competition from the indigenous rhizosphere microflora.

Antifungal activity

In addition to the ability of *Streptomyces* WYEC 108 to colonize roots of plants and to produce antifungal metabolites, WYEC 108 has also been observed to lyse fungal cell walls and fungal spores. Using scanning electron microscopy, it has been shown that the hyphae of *Streptomyces* WYEC 108 colonize the surface of fungal hyphae or spores, including hyphae and spores of *Pythium ultimum*. The colonized hyphae and spores are degraded by *Streptomyces* WYEC 108, most likely as a result of the secretion of extracellular enzymes by WYEC 108, such as chitinases and cellulases. *Streptomyces* WYEC 108 has been shown to produce both of these enzymes. It is likely that *Streptomyces* WYEC 108 also produces a range of other extracellular degradative enzymes.



EXAMPLE VI

Incorporation of *Streptomyces* WYEC 108 into Delivery Medium

A composition suitable for the long term storage of viable *Streptomyces* WYEC 108 spores and use in agricultural practices was formulated as follows. One liter Erlenmeyer
5 flasks containing 500 ml YGM medium (pH 7.0-7.1) were inoculated with 20 ml of stock culture and incubated with shaking at 250 rpm at 30°C for three days. After incubation, the culture was harvested by centrifugation at 5,000 rpm for 10 minutes. The harvested material was resuspended in 1600 ml of 10% YGM and mixed with sterilized 8 g NH_4Cl dissolved in
10 400 ml distilled water. The two liters of cell and NH_4Cl mixture were then inoculated into a plastic container containing 4 kg sterilized delivery medium consisting of a sand-water-cornmeal mixture in a 9-2-1 (w/w) ratio. The delivery medium was sterilized twice (3 hr at 121°C) before incubation of the culture. This mixture was then incubated for 10-14 days at 25°C to maximize the number of spores present in the mixture. *Streptomyces* WYEC 108
15 produced spores during the 10-14 days incubation resulting in increased cfu/g of delivery medium (to an average level of 10^8 to 10^9 cfu/g of delivery medium (dry weight)). This mixture was then stored at 4°C until use.

Alternatively, instead of YGM medium, cells and spores can be produced in CYG medium. As an alternative to harvesting cells by centrifugation, culture flasks may also
20 be allowed to stand so that the bacterial mycelia and spores settle. Thereafter, the clear supernatant is decanted and the concentrated mycelia/spore suspension is inoculated directly into the delivery medium. When this harvesting procedure is utilized, it is not necessary to add NH_4Cl to the medium since the bacterial growth medium (YGM or CYG) is a suitable source of nitrogen.

25

EXAMPLE VII

Influence of *Streptomyces* WYEC 108 on Emergence and Fresh Weight of Lettuce Seedlings

To determine the effect of *Streptomyces* WYEC 108 on the growth of lettuce, lettuce seeds were grown either in the delivery medium containing *Streptomyces* WYEC 108 as
30 described in Example VI above, or in steam sterilized soil. A total of 30 seeds was planted for each growth medium (one seed per 4 x 13.5 cm pot) and emergence was recorded after 21 days of growth. Fresh weight was determined by harvesting above ground plant growth after 35 days of growth. Fresh weight was recorded as a mean value. The results are shown in Table VII below.



TABLE VII

	Treatment	Number of Emerged Plant			Emergence (%)	Fresh weight (g/plant)
		7-day	9-day	21-day		
	Sterile Soil + Seed	13	17	21	(21/30) 70	0.81
5	Delivery Medium + WYEC 108 + Seed	23	27	29	(29/30) 97	1.41

- 10 These results indicate that treatment of lettuce seed with *Streptomyces* WYEC 108 enhances both the emergence frequency and fresh weight of the seedlings.

EXAMPLE VIII

Use of *Streptomyces* WYEC 108 in Delivery Medium in the Field

- 15 In vivo biocontrol assays were carried out to determine the effectiveness of *Streptomyces* WYEC 108 as a biocontrol agent when incorporated into the delivery medium described above.

- 20 *Streptomyces* WYEC 108 was produced and incorporated into the delivery medium as described in Example VII above, using *Streptomyces* WYEC 108 grown for three days in YGM medium. The initial population of *Streptomyces* WYEC 108 in the delivery medium was determined to be approximately $1.0-1.2 \times 10^5$ cfu/g of soil by plate counts on CYD plates immediately prior to planting seedlings in the pots. This treated soil was then used to fill seedling pots (4 cm by 13.5 cm).

- 25 Control plants were grown in pots containing steam sterilized soil (sterilized at 100°C for 60 minutes).

- 30 Pepper seedlings (green, hot peppers) were then planted in the seedling pots containing either steam sterilized soil only or a mixture of steam sterilized soil inoculated with the delivery medium containing *Streptomyces* WYEC 108. After six weeks of growth in a greenhouse, these plants were transplanted to an agricultural field. In some instances, 100 g of the delivery medium containing 450 ± 17 cfu/g of *Phytophthora parasitica* was inoculated into the planting hole prior to transplantation.

- 35 Plant height was determined 55 days after transplantation and recorded as a mean value. Plant biomass was determined by harvesting and measuring fresh weight of the plants after 110 days of cultivation post transplantation. At that time, the number of peppers formed and the weight of peppers per plant were recorded as a mean value. The results of this field trial are shown in Table VIII.



TABLE VIII

	Treatment	Plant Height (cm)	Biomass fresh weight (g/plant)	# of pepper per plant	Yield of pepper fresh weight (g/plant)
5	^b Control: Untreated	22.4k ^y	274.1k ^y	19.1k ^y	158.8k ^y
	^c Treated with WYEC 108	29.1m	353.3m	36.5m	279.8m
10	^b Untreated + <i>P. parasitica</i> ^d	20.3k	238.5k	13.8k	151.2k
15	^c Treated with WYEC 108 + <i>P. parasitica</i> ^d	28.6m	266.6k	22.7k	221.6k

^bPepper plants grown in pots (4 cm x 13.5 cm) containing steam sterilized soil only.

^cPepper plants grown in pots containing a mixture of steam sterilized soil and delivery medium including *Streptomyces* WYEC 108.

^dOne hundred g of delivery medium containing *Phytophthora parasitica* was inoculated into individual hole before transplantation (450 ± 17 cfu/g of the delivery medium).

^yMeans in a column followed by the same letter are not significantly different at the $P = 0.05$ level.

As shown in Table VIII, treatment of pepper seedlings with WYEC 108 in the absence of *P. parasitica* produced statistically significant increases in plant height, biomass, number of peppers and yield of peppers compared to control plants that did not receive WYEC 108. A comparison of horizontal rows 3 and 4 of Table V shows that strain WYEC 108 protected peppers against the deleterious effects of *P. parasitica*. Additionally, there was a significant enhancement of the growth of plants treated with strain WYEC 108 in the absence of *P. parasitica* compared to untreated plants without *P. parasitica*.

In addition to the experiments described in this example, WYEC 108 has been shown to be effective in protecting a range of plants against fungal infection. Plants tested include those shown in Table IX below.



TABLE IX

Crops Protected from Fungal Infection by WYEC 108

Plants Tested with WYEC 108^a

5	Plant	Field Test (+/-)	Greenhouse/Lab Test (+/-)
	Lettuce	-	+
10	Chickpea	-	+
	Green Pea	-	+
	Pepper	+	-
	Cotton	+	+
	Turf Grass	+	+
15	Onion	+	+
	Potato	+	+

^a Tests were performed either in a greenhouse or in the field, or both, as indicated. The tests compared WYEC 108-treated plants with untreated control plants and/or with plants treated with WYEC 108 and a specific fungal pathogen (*Pythium*, *Aphanomyces*, *Rhizoctonia*, *Fusarium*, *Phytophthora*, or *Phytophthora*). In each of these cases WYEC 108 protected the plants against fungal diseases.

EXAMPLE IX

25 Production of Spores of *Streptomyces* WYEC 108
in Liquid Media

Biocontrol agents must survive for extended periods of time to meet shipping needs and the timing patterns of agricultural uses. The use of spores of strain WYEC 108 rather than vegetative cells in particular biocontrol formulations enhances the shelf-life of the biocontrol formulation since the spores retain viability under adverse conditions and over long periods of time.

Typically, spores of *Streptomyces* species are only produced on solid media. However, as set forth below, the following method was found suitable for producing spores in liquid culture.

35 Two liter Erlenmeyer flasks containing 1,200 ml YGM medium (pH 6.5) were each inoculated with 50 ml of stock culture (produced as described in Example II) and incubated with shaking at 250 rpm at 30°C for 12-18 days. Spore production in the culture was monitored by observing with phase-contrast microscope (x 1,000, and stained with methylene blue). Spores were harvested by centrifugation at 9,000 rpm for 10 minutes.

40 Thereafter, the spores were resuspended in 1,600 ml of sterilized 10% YGM liquid medium and 400 ml of a sterile solution comprising 8 g NH₄Cl in distilled water was added (to produce a final spore density of 1.0-1.2 x 10⁷ cfu/ml). This mixture was then directly inoculated into 4 kg of sterilized delivery medium consisting of sand, water and cornmeal in a 9:2:1 (w/w) ratio. The delivery medium was sterilized twice in an autoclave (3 hours at 121°C) prior to inoculation with the spores.



The production of spores directly in liquid culture in the described manner avoids the need for a further incubation of the mixture. The delivery medium containing spores was then stored at 4 °C until used.

5 Spores produced by the liquid culture method described were tested for viability after four months of storage at 4 °C. One ml of spore suspension was inoculated into flasks containing 100 ml of sterilized 10% YGM liquid medium (pH6.5) and incubated with shaking at 250 rpm at 30 °C. Germination of spores was observed by phase-contrast microscopy (x 1,000, stained with methylene blue). Spores were completely germinated in approximately 8 days. This simple observation test showed no loss in viability after this period of storage.

10 *Streptomyces* WYEC 108 incorporated into the delivery medium (sand, water, cornmeal; 9:2:1) was tested for viability as follows. A 1.0 g sample of the delivery medium containing *Streptomyces* WYEC 108 was serially diluted and plated on CYD agar plates. Plates were incubated at 25 °C until colonies were formed. An average level of 10^8 to 10^9 cfu/g of delivery medium (dry weight) were recorded with samples stored for 30 days.

15 Alternatively, the spores from an agar plate of sporulation agar were resuspended in 10-20 ml of sterile distilled water or YGM broth and mixed into 10-100 grams of delivery medium, to obtain a viable count of 10^{12} to 10^{14} cfu/g of medium. This mixture was then air-dried, mixed thoroughly and stored at 4 °C until used. This formulation is a concentrated product that can be diluted with additional delivery medium to any desired lower cfu/g final
20 viable count.

EXAMPLE X

Stability of Alginate Gel Formulation

Mycelia of *Streptomyces* WYEC 108 were harvested by centrifugation at 5,000 rpm for 10 minutes from a 500 ml 3-day-old YGM liquid culture. The harvested mycelia were
25 resuspended in 125 ml of 10% YGM and added 125 ml of sterilized 5% (w/v) sodium alginate solution to a culture density of 1.0 - 1.2×10^4 cfu/ml. Alginate pellets containing mycelia of *Streptomyces* WYEC 108 were formed by adding cell-alginate suspension drop by drop into sterilized 0.25 M CaCl_2 in distilled water.

To determine the viability of the alginate pellets formed by this method, alginate
30 pellets containing the culture were subsequently spread on a sterilized plastic petri dish (10 cm x 10 cm) and dried for one hour in a laminar flow sterile air hood. The pelletized *Streptomyces* WYEC 108 sporulated readily following storage at 25 °C for 6 to 8 months (to an average level of 10^8 to 10^9 cfu/g dried alginate beads). These spores were readily germinated when they were incubated in sterilized water at 25 °C. Germination of the spores were
35 observed by phase-contrast microscopy (x 1,000 magnification, stained with methylene blue).



EXAMPLE XI

Preferred Formulation of Delivery Medium Including *Streptomyces* WYEC 108

Having set forth above methods for isolating actinomycete strains, testing these strains for utility as biocontrol agents, methods for producing these biocontrol agents in mycelial form and as spores, suitable delivery media and, in a preferred embodiment, *Streptomyces* WYEC 108, it will be apparent to one skilled in the art that the present invention can be modified in a number of ways without departing from the spirit of the invention.

Set forth below are examples of alternative embodiments of the present inventions, together with descriptions of particularly preferred embodiments.

Optimum Culture Conditions

Optimal conditions for growth of strain WYEC 108 include temperatures between 20°C and 30°C, at pHs between 5.5 and 7.5, and at fermenter agitation speeds between 200 rpm and 300 rpm. *Streptomyces* WYEC 108 typically achieves maximal cell mass yields of about 5.3 dry weight grams of biomass/liter in YGM liquid medium with culture conditions of 30°C, pH 6.5, and shaking at 200 rpm for 72 hr (to the end of log phase). Doubling time during logarithmic growth phase is approximately 10 hours. The 72 hr incubation time may be significantly reduced by using higher inoculum levels of log phase cells.

Alternatively, spores may be produced on solid agar media such as sporulation agar. These spores may be directly harvested by scraping into a suitable liquid medium such as 10% YGM and then directly introduced into the delivery medium. This approach avoids the need for liquid growth of the culture and thereby shortens the production process.

Preferred and Alternative Delivery Media

Streptomyces WYEC 108 may be incorporated into a delivery medium for use in horticultural and agricultural settings. Example VI describes one formulation of the delivery medium which comprises sand-water-cornmeal in a 9-2-1 (w/w) ratio. It will be understood by one skilled in the art that the formulation of the delivery medium will be dictated by the particular application for which the biocontrol agent is intended. For example, various organic and inorganic fillers such as clay, vermiculite, wheat bran, corn cobs or chitin can be added to the delivery medium. The ratio of components of a delivery medium will be determined on the basis of texture and physical properties required. For example, properties such as moisture holding ability, light weight for easy handling and transportation, porosity to provide space for mycelial and plant root growth and spread may be important. Alternatively, vegetative mycelia or spores of *Streptomyces* WYEC 108 can be added to an alginate suspension to produce alginate-entrapped pellets on this strain. Methods of producing alginate pellets are known in the art and are described further in U.S. Patent No. 4,668,512 to Lewis et al. Other ingredients, such as fertilizers, may also be incorporated into these pellets.



In a preferred embodiment, the present inventors have determined that a delivery medium comprising peat moss-sand-cornmeal in a 1:3.5:1 weight/weight ratio is particularly suitable. This ratio provides an appropriate density and water holding capacity for the use of this product in agricultural and horticultural applications. However, as stated above, other ratios of these components and of other components are also acceptable as delivery media. For example, an effective alternative delivery medium comprises peat moss (620 g) - sand (3380 g) - cornmeal (270 g) - chitin (10 g).

In one embodiment, approximately 1.6 liters of harvested culture broth (log-phase cells: e.g., about 72 hr culture) containing *Streptomyces* WYEC 108 mycelium grown in YGM broth as described above is supplemented with 400 ml of a sterile solution of NH_4Cl (containing 8 g of NH_4Cl in 400 ml distilled water) and inoculated into plastic containers containing 4 kg sterilized Delivery Medium consisting of peat moss, sand, and cornmeal. The Delivery Medium is sterilized twice (3 hours at 121°C) before inoculation of *Streptomyces* WYEC 108. Inoculated containers are incubated at 30°C for 10 to 14 days to maximize spore formation. Containers are then stored at 4°C until used.

The use of NH_4Cl in the delivery medium provides a nitrogen source of germinating spores of *Streptomyces* WYEC 108. It will be apparent to one skilled in the art that other nitrogen sources besides NH_4Cl can be used for this purpose. For example, and as described herein, when spores are resuspended in bacterial growth medium (such as 10% YGM) prior to incorporation in the delivery medium, the addition of this nitrogen source is unnecessary. In preferred embodiments of the present invention, the delivery medium comprises a sufficient amount of a nitrogen source. It will be apparent to one skilled in the art that the determination of what comprises "a sufficient amount" of a nitrogen source can be made by determining the effects on germination frequency of increasing or decreasing the amount of a particular nitrogen source or the effects of changing the nitrogen source. A sufficient amount of a nitrogen source is that amount of a particular nitrogen source which facilitates germination of the spores or *Streptomyces* WYEC 108.

In an alternative embodiment, as described in Example XI, spores of *Streptomyces* WYEC 108 are produced in liquid medium and directly incorporated into the preferred delivery medium which is then stored at 4°C .

In a preferred embodiment of the present invention, *Streptomyces* WYEC 108 is added to the delivery medium to a final concentration of at least 1×10^5 cfu/g. In more preferred embodiments, the final concentration of *Streptomyces* WYEC 108 in the delivery medium is between 1×10^5 cfu/g and 1×10^8 cfu/g.



EXAMPLE XII

Detailed Description of Formulation of Delivery Medium Including Streptomyces WYEC 108

A preferred formulation of the delivery medium containing *Streptomyces* WYEC 108 is produced on a large scale by the procedure set forth below. All of the procedures described are performed using standard aseptic technique (e.g., in a UV light-sterilized laminar flow chamber) to assure asepsis until the packaged bags are opened by final users.

Production of Cells

1) Suspend the spores from a CYD slant of *Streptomyces* WYEC 108 in 10 ml of sterile YGM or CYG broth (pH 6.5). This inoculum suspension is used to inoculate the flask cultures.

2) Inoculate six 250 ml flasks containing 100 ml YGM (pH 6.5). Use 10 ml of spore suspension per flask as inoculum. After inoculation, flasks are incubated with shaking at 200 rpm and at 30 °C for about 36 hrs.

3) Inoculate six 2.0 liter flasks each containing 1.1 liter of YGM broth (pH 6.5) with the mycelial inoculum prepared above (100 ml of inoculum per flask). After inoculation, flasks are incubated with shaking at 200 rpm and at 30 °C for about 24 to 48 hrs or longer (up to 4 days). This becomes the inoculum for the fermenter.

Fermentation

Approximately 7.2 liter of the stock culture prepared above is inoculated into a fermenter containing 40 liter of sterile YGM broth (pH 6.5) (= a 15% inoculum by volume; the approach is to inoculate with as high a density of cell suspension as practical). The fermenter is operated with agitation (200 rpm) at 30 °C for about 72 hours (to near the end of log phase).

Fermenter Harvest

1) The fermentation culture broth containing the WYEC 108 cells (after about 72 hrs incubation) is aseptically harvested in sterile 20 liter plastic bottles.

2) Sterile NH_4Cl solution is added to the harvested culture broth, which still contains the WYEC 108 cells (Use 16 g NH_4Cl dissolved in 800 ml distilled water per 3.2 liter of harvested culture broth; pre-sterilized by autoclaving). The resulting 1.2 liter volume of NH_4Cl -containing cell suspension is then mixed well by shaking the bottle before it is inoculated into the previously prepared delivery medium.

Preparation of the Delivery Medium

1) Each component of the Delivery Medium is measured separately and added into a large size sterilizable pan or other suitable container. The combined mixture is defined as the delivery medium. It consists of peat moss, sand, and cornmeal (540g:2700g:540g; 1:3.5:1 w/w ratio).

2) The delivery medium is thoroughly mixed and covered with sturdy aluminum foil or cotton batting and then sterilized twice (90 minutes at a time at 121 °C with 12 hours between sterilization periods).

3) The delivery medium is cooled to room temperature after the second
5 sterilization and before inoculation of the harvested culture broth containing strain WYEC 108 and NH_4Cl solution (prepared above).

Incorporation of *Streptomyces* WYEC 108 into the delivery medium to Create a Formulation of Peat Moss, Sand, Water, Cornmeal, and NH_4Cl

1) About 0.5 liters of harvested culture broth containing strain WYEC
10 108- NH_4Cl solution (prepared above) is thoroughly incorporated into each of as many as needed presterilized plastic containers containing of 3.78 kg of delivery medium.

2) The inoculated containers are then incubated at 30 °C for 10-14 days (up to 20 days incubation may be optimal) after which they can be stored at 4 °C until used (the formulation is stable for months).

15 **Handling and Transportation**

1) The completed formulation containing *Streptomyces* WYEC 108 is aseptically transferred into sterile triple-layer plastic bags using a small sterilized shovel or equivalent tool, preferably in a UV-sterilized laminar flow hood.

2) The filled bags are next tied and put into 1.5 ft³ moving boxes for shipping.
20 Each box is then sealed with strong tape.

EXAMPLE XIII

Incorporation of the Formulation Containing *Streptomyces* WYEC 108 into Seedling Nursery Beds

The formulation containing the *Streptomyces* WYEC 108 biocontrol agent and
25 delivery medium as described in Example XII is mixed with a nursery bedding soil or potting mix to a final *Streptomyces* concentration of $\geq 1.0-1.2 \times 10^5$ or more cfu/g-soil). The seedling procedure is as follows.

1) About 1.0 cm of peat moss is placed in the bottom of each pot (or bed) to prevent loss of soil (or potting mix) while still providing for aeration and drainage.

2) Seedling pots are then filled with the agricultural (nursery or potting mix) soil
30 up to about 3.0 cm below from the top of the pots (or beds). The pots (beds) are then watered to saturation.

3) About 1.5 cm of the formulation containing *Streptomyces* WYEC 108 and delivery medium is then added to the top of each pot (bed). If desired, the formulation can
35 also pre-mixed with nursery bedding soil or potting soil to increase the volume and adjust the cfu/g count. However, for optimum efficacy, the cfu/g should be maintained at at least 10^5 cfu/g in the final mix.



4) Seeds are placed on the surface of the prepared seedling pots or beds and then covered with an additional 1.5 cm (approximate) of nursery bedding soil or potting soil/mix.

5) A small amount of water is then added to wet the soil and seeds.

5 6) To minimize drying and to prevent crusting, the pots are typically covered with clean black plastic until seedling emergence (This may not necessary if moisture is controlled).

7) Additional water is sprayed on the top of the pots (or beds) as needed after seedling emergence.

10 Having provided examples of embodiments of this invention and preferred embodiments, it will be apparent to those skilled in the art that changes and modifications may be made without departing from the present invention and its broader aspects. We therefore intend the appended claims to cover all such changes and modifications falling within the true spirit and scope of the present invention.

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CLAIMS

1. A biologically pure culture of a microorganism *Streptomyces* WYEC 108, having the identifying characteristics of ATCC 55445.
2. The microorganism of claim 1 wherein the microorganism is capable of conferring protection against fungal infection in a susceptible plant.
3. A composition comprising:
 - a biologically pure culture of *Streptomyces* WYEC 108; and
 - a delivery medium.
4. The composition of claim 3 wherein the delivery medium comprises at least one of the following components: alginate gel, peat moss, sand and cornmeal.
5. The composition of claim 3 wherein the delivery medium comprises a sufficient amount of a nitrogen source.
6. The composition of claim 4 wherein the delivery medium comprises peat moss, sand, cornmeal and a sufficient amount of a nitrogen source.
7. The composition of claim 6 wherein the nitrogen source is ammonium chloride.
8. The composition of claim 3 wherein the biologically pure culture of *Streptomyces* WYEC 108 comprises spores of *Streptomyces* WYEC 108.
9. A composition useful for protecting plants against fungal infection comprising a suitable *Streptomyces* species and a delivery means in a suitable container.
10. The composition of claim 9 wherein the *Streptomyces* species is *Streptomyces* WYEC 108.
11. A composition comprising:
 - a purified preparation of antifungal metabolites produced by *Streptomyces* WYEC 108; and
 - a delivery medium.
12. The composition of claim 11 wherein the composition further comprises cells and spores of *Streptomyces* WYEC 108.
13. A method for reducing the susceptibility of a plant to fungal infection wherein the method comprises delivering *Streptomyces* WYEC 108 to the roots of the plant.
14. A method for reducing the susceptibility of plants to fungal infection wherein the method comprises the steps of:
 - immersing seeds in a composition comprising *Streptomyces* WYEC 108; and
 - planting the seeds in a suitable growth medium.
15. The method of claim 14 wherein the composition further comprises alginate gel.





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16. A method of protecting plants against fungal infection comprising the steps of:

supplying a culture of *Streptomyces* WYEC 108;
preparing a delivery medium comprising said culture; and
delivering said delivery medium to plants.

5

17. A method of enhancing the growth of plants comprising providing to seeds or seedlings a composition comprising a delivery medium and at least one component selected from the group consisting of: *Streptomyces* WYEC 108 cells; *Streptomyces* WYEC 108 spores; and antifungal metabolites produced by *Streptomyces* WYEC 108.

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