

Antibodies against IL-13 receptor alpha1 and uses thereof

The present invention relates to human antibodies against IL-13 receptor alpha1 (IL-13Ra (alpha)1), methods for their production, and uses.

5 Background of the Invention

10 IL-13 is a secreted monomeric peptide produced mainly by Th2 cells but also by mast cells and NK cells. Biological functions of IL-13 include regulation of IgE production and modulation of Th2 development. IL-13 binds to a receptor complex consisting of IL-13 receptor alpha1 (IL-13Ra1) chain and IL-4 receptor
15 alpha (IL-4Ra) chain. IL-13 binding triggers signal transduction events mainly through STAT6. IL-13 binds with low affinity to the IL-13 Ra1 alone and does not bind to IL-4Ra1. Contrary to this, IL-4 binds to IL-4Ra alone and does not bind to IL-13Ra1 alone. Another receptor for IL-13 has been described, the IL-13Ra2. IL-13 binds with high affinity to this receptor. Likely this receptor acts as a decoy receptor.

20 Inducible overexpression of IL-13 in transgenic mice results in a phenotype that shares many characteristics with asthmatic patients. They show mucus metaplasia, macrophage, lymphocyte and eosinophil-rich inflammation, upregulation of proteases like MMP-9, -12, -13, -2 and -14, cathepsin B, H, K and S and they also
25 present subepithelial fibrosis. Knockout mice for IL-13 show a significant reduction in Th2 cytokine production due to impairment in Th2 development. These mice do not develop airway hyperreactivity (AHR) in spite of the presence of eosinophil inflammation. The AHR was restored by administration of IL-13, indicating that IL-13 is necessary and sufficient for the induction of AHR in mouse. Other
30 important biological functions of IL-13 in relationship with asthma include the induction of goblet cell metaplasia and mucus production. It acts directly on airway epithelial cells, fibroblasts and airway smooth muscle cells and induces different transcriptional programs in each of this cell types. Interestingly, IL-13 decreases the alpha-adrenergic response in smooth muscle cells, contributing to airway narrowing. IL-13 promoter polymorphism is associated with increased risk of allergic asthma. Polymorphisms in the IL-13 gene are associated with high serum

IgE levels. Single nucleotide polymorphism in the intergene sequence between the IL-4 and IL-13 genes is associated with atopic asthma.

IL-13 antagonists have been utilized in animal models. For example a soluble mouse IL-13R α 2-IgGFc fusion protein has been used to show efficacy in completely reversing ovalbumin-induced AHR and the number of mucus containing cells. The reversal was obtained even if the treatment is given after full development of the phenotype. In addition, treatment of mice with an IL-13 fusion cytotoxin molecule resulted in reduction of all features of airway disease in a chronic fungal-induced allergic inflammation. In conclusion, IL-13 is a critical mediator of the effector arm of the allergic response.

IL-13R α 1 is a member of the hemopoietin receptor superfamily (type 1 cytokine receptor family) and identified and described by Obiri N. I., et al., J. Biol. Chem., 270 (1995) 8797-8804) and WO 96/29417. It is a protein of 427 amino acids including the signal sequence. Its DNA and protein sequences are described in WO 97/15663 and SwissProt No. P78552. IL-13R α 1 is a glycosylated protein binding to IL-13 with low affinity, but, when linked with IL-4R α to a heterodimer, it binds IL-13 with high affinity. This complex is also a receptor for IL-4.

Antibodies against IL-13R α 1 are known from WO 96/29417, WO 97/15663, WO 03/080675, Graber P., et al., Eur. J. Immunol., 28 (1998) 4286-4298; Poudrier J., et al., J. Immunol., 163 (1999) 1153-1161; Poudrier J., et al., Eur. J. Immunol., 30 (2000) 3157-3164; Aikawa M., et al., Cytokine, 13 (2001) 75-84. Antibodies against IL-13R α 1 are commercially available from R&D Systems Inc. USA.

Summary

The disclosure comprises an antibody binding to IL-13R α 1 and inhibiting IL-13 bioactivity, characterized in that the variable heavy chain amino acid sequence CDR3 of said antibody is selected from the group consisting of the heavy chain CDR3 sequences of SEQ ID NO: 1, 3, 5, 7 or 9.

In one aspect, there is provided an antibody binding to IL-13R α 1 and inhibiting IL-13, characterized in that the

variable heavy chain amino acid sequence CDR3 of said antibody is selected from the group consisting of the heavy chain CDR3 sequences of SEQ ID NO: 1, 3 or 7, whereby said antibody is obtained from hybridoma cell line DSM ACC2709, DSM ACC2710 or DSM ACC2712.

In another aspect, there is provided an antibody binding to IL-13R α 1 and inhibiting IL-13, characterized in that the variable heavy chain amino acid sequence CDR3 of said antibody is selected from the group consisting of the heavy chain CDR3 sequences of SEQ ID NO: 1, 3, 7 or 9 and comprises as heavy chain variable region SEQ ID NO: 1 and as light chain variable region SEQ ID NO: 2, as heavy chain variable region SEQ ID NO: 3 and as light chain variable region of SEQ ID NO: 4, as heavy chain variable region SEQ ID NO: 7 and as light chain variable region SEQ ID NO: 8 or as heavy variable region SEQ ID NO: 9 and as light chain variable region SEQ ID NO: 10.

The antibody is preferably a human antibody.

The antibody is preferably characterized by an affinity of 10^{-9} M (K_D) or less, preferably of 10^{-9} to 10^{-13} M for binding to IL-13R α 1.

5 Preferably the antibody is characterized in that its heavy chain CDR1, CDR2 and CDR3 sequences are selected from the group consisting of the heavy chain CDR1, CDR2 and CDR3 sequences of SEQ ID NO: 1, 3, 5, 7 or 9.

The antibody is preferably characterized in that the variable light chain amino acid sequences CDR1, CDR2 and CDR3 of said antibody are selected from the group consisting of the light chain CDR sequences of SEQ ID NO: 2, 4, 6, 8 or 10.

10 The antibody is preferably characterized in that the variable heavy chain amino acid sequences CDR1, CDR2 and CDR3 of said antibody are selected from the group consisting of the heavy chain CDR sequences of SEQ ID NO: 1, 3, 5, 7 or 9 and the variable light chain amino acid sequences CDR1, CDR2 and CDR3, of said antibody are selected from the group consisting of the light chain CDR sequences of SEQ ID NO: 2, 4, 6, 8 or 10.

15 The CDR sequences are preferably selected independently of each other and are separated by FR (framework) regions.

20 The antibody is preferably characterized in comprising as heavy chain CDRs the CDRs of SEQ ID NO: 1 and as light chain CDRs the CDRs of SEQ ID NO: 2, as heavy chain CDRs the CDRs of SEQ ID NO: 3 and as light chain CDRs the CDRs of SEQ ID NO: 4, as heavy chain CDRs the CDRs of SEQ ID NO: 5 and as light chain CDRs the CDRs of SEQ ID NO: 6, as heavy chain CDRs the CDRs of SEQ ID NO: 7 and as light chain CDRs the CDRs of SEQ ID NO: 8 or as heavy chain CDRs the CDRs of SEQ ID NO: 9 and as light chain CDRs the CDRs of SEQ ID NO: 10.

25 The CDR sequences can be determined according to the standard definition of Kabat et al., Sequences of Proteins of Immunological Interest, 5th ed., Public Health Service, National Institutes of Health, Bethesda, MD (1991). On this basis, the complementarity determining regions (CDRs) of SEQ ID NO: 1-8 have the following sequences:

Heavy chain CDRs: CDR1 (aa 31-35) of SEQ ID NO: 1, 3, 5, 7, 9, CDR2 (aa 50-66) of SEQ ID NO: 1, 3, 5, 7, 9, CDR3 (aa 99-108) of SEQ ID NO: 1, 3, 9, CDR3 (aa 99-107) of SEQ ID NO: 5, CDR3 (aa 99-112) of SEQ ID NO: 7;
Light chain CDRs: CDR1 (aa 24-34) of SEQ ID NO: 2, 4, 6, 10, CDR1 (aa 24-35) of SEQ ID NO: 8, CDR2 (aa 50-56) of SEQ ID NO: 2, 4, 6, 10, CDR2 (aa 51-57) of SEQ ID NO: 8 and CDR3 (aa 89-97) of SEQ ID NO: 2, 4, 6, 10, CDR3 (aa 90-97) of SEQ ID NO: 8.

Preferably, the disclosure provides an antibody comprising as complementarity determining regions (CDRs) the following sequences:

- a) an antibody heavy chain comprising heavy chain CDRs of SEQ ID NO: 1, 3, 5, 7 or 9;
- b) an antibody light chain comprising light chain CDRs of SEQ ID NO: 2, 4, 6, 8 or 10,

wherein the CDRs are selected independently of each other.

The antibody is preferably characterized in comprising as heavy chain variable region SEQ ID NO: 1 and as light chain variable region SEQ ID NO: 2, as heavy chain variable region SEQ ID NO: 3 and as light chain variable region of SEQ ID NO: 4, as heavy chain variable region SEQ ID NO: 5 and as light chain variable region SEQ ID NO: 6, as heavy chain variable region SEQ ID NO: 7 and as light chain variable region SEQ ID NO: 8 or as heavy variable region SEQ ID NO: 9 and as light chain variable region SEQ ID NO: 10.

The antibody is preferably characterized in comprising

a) as heavy chain variable region SEQ ID NO: 1, as light chain variable region SEQ ID NO: 2, as κ light chain constant region SEQ ID NO: 11 and as $\gamma 1$ heavy chain constant region SEQ ID NO: 12 optionally with mutations L234A and L235A or D265A and N297A,

b) as heavy chain variable region SEQ ID NO: 3 and as light chain variable region of SEQ ID NO: 4, as κ light chain constant region SEQ ID NO: 11 and as $\gamma 1$ heavy chain constant region SEQ ID NO: 12 optionally with mutations L234A and L235A or D265A and N297A,

c) as heavy chain variable region SEQ ID NO: 5 and as light chain variable region SEQ ID NO: 6, as κ light chain constant region SEQ ID NO: 11 and as $\gamma 1$ heavy chain constant region SEQ ID NO: 12 optionally with mutations L234A and L235A or D265A and N297A,

5 d) as heavy chain variable region SEQ ID NO: 7 and as light chain variable region SEQ ID NO: 8, as κ light chain constant region SEQ ID NO: 11 and as $\gamma 1$ heavy chain constant region SEQ ID NO: 12 optionally with mutations L234A and L235A or D265A and N297A, or

10 e) as heavy variable region SEQ ID NO: 9 and as light chain variable region SEQ ID NO: 10, as κ light chain constant region SEQ ID NO: 11 and as $\gamma 1$ heavy chain constant region SEQ ID NO: 12 optionally with mutations L234A and L235A or D265A and N297A.

15 Preferably the antibody is characterized in binding to IL-13R $\alpha 1$ in competition to antibody LC5002-002, LC5002-003, LC5002-005, LC5002-007 and/or LC5002-018. Preferably the antibody is characterized in comprising as variable regions the variable regions of LC5002-002, LC5002-003, LC5002-005, LC5002-007 or LC5002-018. The variable regions of these antibodies are shown in SEQ ID NO: 1-10. Useful constant regions are well known in the state of the art. Examples are shown in SEQ
20 ID NO: 11-12.

The antibody is preferably a monoclonal or a recombinantly produced antibody.

In one embodiment of the disclosure the antibody is a class-altered human antibody.

25 In a preferred embodiment of the disclosure the antibody contains a human $\gamma 1$ heavy chain comprising

- a) amino acid sequence Pro₂₃₃Val₂₃₄Ala₂₃₅ with deletion of Gly₂₃₆ and/or amino acid sequence Gly₃₂₇Leu₃₂₈Pro₃₂₉Ser₃₃₀Ser₃₃₁,
- b) amino acid sequence Ala₂₃₄Ala₂₃₅ or
- c) amino acids Ala₂₆₅ and Ala₂₉₇.

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Preferably the antibody according to the disclosure inhibits IL-13 induced Stat-6 phosphorylation with an IC₅₀ value of 6 nM or lower, inhibits IL-13 induced eotaxin production with an IC₅₀ value of 20 nM or lower and/or inhibits IL-13 or

IL-4 induced cell proliferation, preferably of TF-1 cells (ATCC CRL 2003) with an IC_{50} value of 10 nM or lower (IL-13) and 60 nM or lower (IL-4). IL-13 induced Stat-6 phosphorylation, eotaxin production and induction of cell proliferation are determined according to examples 6 to 8.

The antibody according to the disclosure preferably does not bind to denatured IL-13 α 1 (K_D for binding affinity 10^{-6} M or higher). The antibody is preferably characterized by showing substantially no crossreactivity with IL-13 α 2 and IL-4 α (K_D for binding affinity 10^{-6} M or higher).

The disclosure further provides hybridoma cell lines which produce antagonistic monoclonal antibodies against IL-13 α 1.

The preferred hybridoma cell lines according to the disclosure (hu-MAB<h-IL-13R alpha>LC.5002-002 (DSM ACC2709), hu-MAB<h-IL-13Ralpha>LC.5002-003 (DSM ACC2710), hu-MAB<h-IL-13Ralpha>LC.5002-005 (DSM ACC2711), hu-MAB<h-IL-13R alpha>LC.5002-007 (DSM ACC2712)) were deposited 13.01.2005 with Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (DSMZ), Germany.

In a further aspect, there is provided a hybridoma cell line DSM ACC2709, DSM ACC2710 or DSM ACC2712.

The antibodies obtainable from said cell lines are embodiments of the disclosure.

The disclosure further provides nucleic acids encoding polypeptides of which antibodies according to the disclosure are comprised, expression vectors comprising said nucleic acids, and host cells for the recombinant production of such antibodies.

In a further aspect, there is provided a nucleic acid encoding an antibody binding to IL-13 α 1 and inhibiting IL-13, comprising as heavy chain variable region SEQ ID NO: 1 and as light chain variable region SEQ ID NO: 2, as heavy chain variable region SEQ ID NO: 3 and as light chain variable region of SEQ ID NO: 4, as heavy chain variable region SEQ ID NO: 7 and as light chain variable region SEQ ID NO: 8 or as heavy variable region SEQ ID NO: 9 and as light chain variable region SEQ ID NO: 10.

In a further aspect, there is provided an expression vector comprising a nucleic acid as defined above, capable of expressing said nucleic acid in a prokaryotic or eukaryotic host cell.

In a further aspect, there is provided a prokaryotic or eukaryotic host cell comprising the vector as defined above.

In a further aspect, there is provided a recombinant host cell capable of producing a recombinant antibody as defined above.

The disclosure further provides methods for the recombinant production of such antibodies.

Hence, in a further aspect, there is provided a method for the production of an antibody binding to IL-13R α 1 and inhibiting the binding of IL-13 to IL-13R α 1, characterized by expressing a nucleic acid encoding a heavy chain and a nucleic acid encoding a light chain as defined above in a prokaryotic or eukaryotic host cell and recovering said antibody from said cell.

The polypeptides encoded by the nucleic acids according to the disclosure are

- a) an antibody heavy chain comprising heavy chain CDRs of SEQ ID NO: NO:1, 3, 5, 7 or 9 and
- b) an antibody light chain comprising light chain CDRs of SEQ ID NO: 2, 4, 6, 8 or 10.

These polypeptides are capable of assembling together with the respective other antibody chain to generate an antibody.

Antibodies according to the disclosure show benefits for patients in need of corticosteroid therapy. The antibodies according to the disclosure have new and inventive properties causing a benefit for a patient suffering from asthma or an allergic disease.

The disclosure further provides methods for treating asthma and allergic diseases.

In a further aspect, there is provided an antibody as defined above for use in the treatment of a patient in need of an asthma or antiallergic therapy.

The disclosure further comprises the use of an antibody according to the disclosure for asthma treatment and, in a further aspect, for the manufacture of a pharmaceutical composition according to the disclosure. In addition, the disclosure, in a further aspect, comprises a method for the manufacture of a pharmaceutical composition according to the disclosure.

Still additionally, in a further aspect, there is provided a method for the preparation of a pharmaceutical composition characterized in producing an antibody as defined above by means of recombinant expression, recovering said antibody and combining said antibody with a pharmaceutical acceptable buffer and/or adjuvant.

The disclosure, in a further aspect, comprises a pharmaceutical composition comprising an antibody according to the disclosure. The pharmaceutical composition may comprise an antibody with a pharmaceutically effective amount, optionally together with a buffer and/or an adjuvant useful for the formulation of antibodies for pharmaceutical purposes.

The disclosure further provides pharmaceutical compositions comprising such antibodies in a pharmaceutically acceptable carrier. In one embodiment, the pharmaceutical composition may be included in an article of manufacture or kit.

The disclosure further comprises a vector comprising a nucleic acid according to the disclosure, capable of expressing said nucleic acid in a prokaryotic or eukaryotic host cell.

The disclosure further comprises a prokaryotic or eukaryotic host cell comprising a vector according to the disclosure.

The disclosure further comprises a method for the production of a recombinant human antibody according to the disclosure, characterized by expressing a nucleic acid according to the disclosure in a prokaryotic or eukaryotic host cell and recovering said antibody from said cell. The disclosure further comprises the antibody obtainable by such a recombinant method.

The disclosure further comprises a method for the preparation of a pharmaceutical composition characterized in selecting an antibody against IL-13R α 1 from a

plurality of antibodies against IL-13R α 1 when compared to such an assay without said antibody, producing said antibody by means of recombinant expression, recovering said antibody and combining said antibody with a pharmaceutical acceptable buffer and/or adjuvant. Preferably the antibody has one or more of the above mentioned additional properties.

Detailed Description of the Invention

The terms "IL-13R α 1, murine IL-13R α 1, IL-13, IL-13R α 2 and IL-4R α " and their domains are well known in the state of the art and e.g. defined by SwissProt P78552, O09030, P35225, Q14627 and P24394. If not otherwise stated, the terms "IL-13R α 1, IL-13, IL-13R α 2 and IL-4R α " therefore denotes the human polypeptides IL-13R α 1, IL-13, IL-13R α 2 and IL-4R α .

The term "human antibody", as used herein, includes antibodies having variable and constant regions (domains) which can be assigned to defined human germ line immunoglobulin sequences because of their high sequence similarity or identity with these germ line sequences. Human antibodies are well-known in the state of the art (van Dijk, M.A., and van de Winkel, J.G., Curr. Opin. Chem. Biol. 5 (2001) 368-374). Human antibodies can also be produced in transgenic animals (e.g. mice) that are capable, upon immunization, of producing a full repertoire or a selection of human antibodies in the absence of endogenous immunoglobulin production. Transfer of the human germ-line immunoglobulin gene array in such germ-line mutant mice results in the production of human antibodies upon antigen challenge (see, e.g., Jakobovits, A., et al., Proc. Natl. Acad. Sci. USA 90 (1993) 2551-2555; Jakobovits, A., et al., Nature 362 (1993) 255-258; Bruggemann, M., et al., Year Immunol. 7 (1993) 33-40). Human antibodies can also be produced in phage display libraries (Hoogenboom, H.R., and Winter, G., J. Mol. Biol. 227 (1992) 381-388; Marks, J.D., et al., J. Mol. Biol. 222 (1991) 581-597). The techniques of Cole et al. and Boerner et al. are also available for the preparation of human monoclonal antibodies (Cole et al., Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, p. 77 (1985); and Boerner, P., et al., J. Immunol. 147 (1991) 86-95). A human antibody encompasses the various forms of antibodies, preferably monoclonal antibodies including but not being limited to whole antibodies, antibody fragments, class-altered antibodies and genetically engineered antibodies (variant or mutant antibodies) as long as the characteristic properties according to the invention are

retained. Especially preferred are recombinant human antibodies. The term "monoclonal antibody" as used herein refers to a preparation of antibody molecules all having substantially the same amino acid sequence.

5 The term "recombinant human antibody", as used herein, is intended to include all human antibodies that are prepared, expressed, created or isolated by recombinant means, such as antibodies isolated from a host cell such as a NS0 or CHO cell or from an animal (e.g. a mouse) that is transgenic for human immunoglobulin genes or antibodies expressed using a recombinant expression vector transfected into such a host cell. Such recombinant human antibodies have variable and constant regions in a rearranged form. The recombinant human antibodies according to the invention have been subjected to in vivo somatic hypermutation. Thus, the amino acid sequences of the VH and VL regions of the recombinant antibodies are sequences that can be assigned to defined human germ line VH and VL sequences, but may not naturally exist within the human antibody germ line repertoire in vivo.

15 The term "class-altered antibody" refers to a monoclonal antibody, preferably a human antibody, comprising a variable region, i.e., binding region, from one source or germ line and at least a portion of a constant region that matches a constant region of an antibody from a different source or germ line, usually prepared by recombinant DNA techniques. Such class-altered antibodies are not naturally occurring and therefore not available directly from xenograft mice. Forms of "class-altered antibodies" encompassed by the present invention are those in which the constant region has differences from the wild-type constant region sequence that result in an antibody having different properties according to the invention, especially in regard to C1q binding and/or Fc receptor (FcR) binding, i.e. by change or mutation of Fc. Class-altered antibodies are the product of expressed immunoglobulin genes comprising DNA segments encoding immunoglobulin variable regions and DNA segments encoding immunoglobulin constant regions. Methods for producing class-altered antibodies involve conventional recombinant DNA and gene transfection techniques are well known in the art (see, e.g., Morrison, S.L., et al., Proc. Natl. Acad. Sci. USA 81 (1984) 6851-6855; US Patent Nos. 5,202,238 and 5,204,244).

The "variable region" (variable region of a light chain (VL), variable region of a heavy chain (VH)) as used herein denotes the part of each of the pair of light and

heavy chains which is involved directly in binding the antibody to the antigen. The domains of variable human light and heavy chains have the same general structure and each domain comprises four framework (FR) regions whose sequences are widely conserved, connected by three "hypervariable regions" (or complementarity determining regions, CDRs). The framework regions adopt a β (beta)-sheet conformation and the CDRs may form loops connecting the β -sheet structure. The CDRs in each chain are held in their three-dimensional structure by the framework regions and form together with the CDRs from the other chain the antigen binding site. The antibody heavy and light chain CDR3 regions, preferably the heavy chain CDR3, play a particularly important role in the binding specificity/affinity of the antibodies according to the invention and therefore provide a further object of the invention.

The terms "hypervariable region" or "antigen-binding portion of an antibody" when used herein refer to the amino acid residues of an antibody which are responsible for antigen-binding. The hypervariable region comprises amino acid residues from the "complementarity determining regions" or "CDRs". "Framework" or "FR" regions are those variable domain regions other than the hypervariable region residues as herein defined. Therefore, the light and heavy chains of an antibody comprise from N- to C-terminus the domains FR1, CDR1, FR2, CDR2, FR3, CDR3, and FR4. CDR and FR regions are determined according to the standard definition of Kabat et al., Sequences of Proteins of Immunological Interest, 5th ed., Public Health Service, National Institutes of Health, Bethesda, MD (1991).

The "constant domains" are not involved directly in binding of an antibody to an antigen, but exhibit various effector functions. Depending on the amino acid sequence of the constant region of their heavy chains, antibodies or immunoglobulins are divided into the classes: IgA, IgD, IgE, IgG and IgM, and several of these may be further divided into subclasses (isotypes), e.g. IgG1, IgG2, IgG3, and IgG4, IgA1 and IgA2. The heavy chain constant regions that correspond to the different classes of immunoglobulins are called μ , δ , γ , α , and ϵ , respectively. The antibodies according to the invention are preferably of IgG1 type.

The Fc part of an antibody is directly involved in complement activation, C1q binding, C3 activation and Fc receptor binding. Binding to C1q is caused by defined binding sites in the Fc part. Such binding sites are known in the state of the

art and described e.g. by Lukas, T.J., et al., *J. Immunol.* 127 (1981) 2555-2560; Brunhouse, R., and Cebra, J.J., *Mol. Immunol.* 16 (1979) 907-917; Burton, D.R., et al., *Nature* 288 (1980) 338-344; Thommesen, J.E., et al., *Mol. Immunol.* 37 (2000) 995-1004; Idusogie, E.E., et al., *J. Immunol.* 164 (2000) 4178-4184; Hezareh, M., et al., *J. Virol.* 75 (2001) 12161-12168; Morgan, A., et al., *Immunology* 86 (1995) 319-324; and EP 0 307 434. Such binding sites are e.g. L234, L235, D270, N297, E318, K320, K322, P331 and P329 (numbering according to EU index of Kabat, see below). Antibodies of subclass IgG1, IgG2 and IgG3 usually show complement activation, C1q binding and C3 activation, whereas IgG4 antibodies do not activate the complement system, do not bind C1q and do not activate C3. As used herein the term "Fc part derived from human origin" denotes a Fc part which preferably has an amino acid sequence of a Fc part of a human antibody of the subclass IgG1 modified in such a way that no C1q binding, C3 activation and/or FcR binding can be detected or binding is at least reduced for 50%, preferably 70%, compared to a human IgG1 antibody. An "Fc part of an antibody" is a term well known to the skilled artisan and defined on the basis of papain cleavage of antibodies. The antibodies according to the invention contain as Fc part, preferably with an amino acid sequence of a Fc part derived from human origin and preferably all other parts of the human constant regions. Preferably the Fc part is a mutated human Fc part from human IgG1 subclass. Mostly preferred are Fc parts comprising a γ 1-heavy chain constant region (an example is shown in SEQ ID NO: 11) with mutations L234A and L235A or D265A and N297A (WO99/51642).

Human constant chains, e.g. γ 1-heavy chains are described in detail by Kabat et al., *Sequences of Proteins of Immunological Interest*, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, MD. (1991), and by Brüggemann, M., et al., *J. Exp. Med.* 166 (1987) 1351-1361; Love, T.W., et al., *Methods Enzymol.* 178 (1989) 515-527. The constant domains preferred in the invention provide no complement binding. The "variable region" (variable region of a light chain (VL), variable region of a heavy chain (VH)) as used herein denotes each of the pair of light and heavy chains which is involved directly in binding the antibody to the antigen.

The term nucleic acid or nucleic acid molecule, as used herein, is intended to include DNA molecules and RNA molecules. A nucleic acid molecule may be single-stranded or double-stranded, but preferably is double-stranded DNA.

A nucleic acid is "operably linked" when it is placed into a functional relationship with another nucleic acid sequence. For example, DNA for a presequence or secretory leader is operably linked to DNA for a polypeptide if it is expressed as a preprotein that participates in the secretion of the polypeptide; a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the sequence; or a ribosome binding site is operably linked to a coding sequence if it is positioned so as to facilitate translation. Generally, "operably linked" means that the DNA sequences being linked are cis, and, in the case of a secretory leader, contiguous and in reading frame. However, enhancers do not have to be contiguous. Linking is accomplished by ligation at convenient restriction sites. If such sites do not exist, the synthetic oligonucleotide adaptors or linkers are used in accordance with conventional practice.

As used herein, the expressions "cell," "cell line," and "cell culture" are used interchangeably and all such designations include progeny. Thus, the words "transformants" and "transformed cells" include the primary subject cell and cultures derived therefrom without regard for the number of transfers. It is also understood that all progeny may not be precisely identical in DNA content, due to deliberate or inadvertent mutations. Variant progeny that have the same function or biological activity as screened for in the originally transformed cell are included. Where distinct designations are intended, it will be clear from the context.

The term "binding to IL-13R α 1" as used herein means the binding of the antibody to IL-13R α 1 in an in vitro assay, preferably in a binding assay in which the antibody is bound to a surface and binding of IL-13R α 1 is measured by Surface Plasmon Resonance (SPR). Binding means a binding affinity (K_D) of 10^{-8} M or less, preferably 10^{-13} to 10^{-9} M. "No binding" means a K_D of 10^{-6} M or more. The antibodies according to the invention bind to the extracellular domain of human IL-13R α 1 and preferably also of mouse IL-13R α 1.

Binding to IL-13R α 1 can be investigated by a BIAcore assay (Pharmacia Biosensor AB, Uppsala, Sweden). The affinity of the binding is defined by the terms k_a (rate constant for the association of the antibody from the antibody/antigen complex), k_d (dissociation rate), and K_D (k_d/k_a).

The binding of IL-13 to IL-13R α 1 is inhibited by the antibodies according to the invention. The inhibition is measured as IC₅₀ in an ELISA for binding of IL-13 to IL-13R α 1/IL-4R α heterodimer. For performing such an assay IL-13R α 1 is immobilized and IL-13 and IL-4R α are added. The IC₅₀ values of the antibodies according to the invention for the binding of IL-13 to IL-13R α 1 are no more than 6 nM. IC₅₀ values are measured as average or median values of at least three independent measurements. Single IC₅₀ values may be out of the scope.

The antibodies according to the invention show preferably a binding to the same epitopes of IL-13R α 1 as an antibody selected from the group consisting of antibodies LC5002-002, LC5002-003, LC5002-005, LC5002-007 or LC5002-018 or are inhibited in binding to IL-13R α 1 due to steric hindrance of binding by these antibodies. Binding inhibition can be detected by an SPR assay using an immobilized antibody selected from the group consisting of antibodies LC5002-002, LC5002-003, LC5002-005, LC5002-007 or LC5002-018 and IL-13R α 1 at a concentration of 20-50 nM and the antibody to be detected at a concentration of 100 nM. A signal reduction of 50% or more shows that the antibody competes with an antibody selected from the group consisting of antibodies LC5002-002, LC5002-003, LC5002-005, LC5002-007 or LC5002-018. The term "epitope" means a protein determinant capable of specific binding to an antibody. Epitopes usually consist of chemically active surface groupings of molecules such as amino acids or sugar side chains and usually have specific three dimensional structural characteristics, as well as specific charge characteristics. Conformational and nonconformational epitopes are distinguished in that the binding to the former but not the latter is lost in the presence of denaturing solvents. The invention comprises also a human antibody binding to IL-13R α 1 and inhibiting IL-13 bioactivity, characterized by an affinity of 10⁻⁹ M (K_D) or less, preferably of 10⁻⁹ to 10⁻¹³ M for binding to IL-13R α 1 and by an affinity of 10⁻⁷ M (K_D) or less, preferably of 10⁻⁸ to 10⁻⁹ M for binding to murine IL-13R α 1.

In a preferred embodiment of the invention, the antibodies according to the invention are further characterized by one or more of the characteristics selected from the group selected from the binding parameters k_a, k_d and K_D, binding to the same epitope to which an antibody selected from the group consisting of antibodies LC5002-002, LC5002-003, LC5002-005, LC5002-007 or LC5002-018 binds.

- The antibodies according to the invention are preferably produced by recombinant means. Such methods are widely known in the state of the art and comprise protein expression in prokaryotic and eukaryotic cells with subsequent isolation of the antibody polypeptide and usually purification to a pharmaceutically acceptable purity. For the protein expression, nucleic acids encoding light and heavy chains or fragments thereof are inserted into expression vectors by standard methods. Expression is performed in appropriate prokaryotic or eukaryotic host cells like CHO cells, NS0 cells, SP2/0 cells, HEK293 cells, COS cells, yeast, or E.coli cells, and the antibody is recovered from the cells (supernatant or cells after lysis).
- Recombinant production of antibodies is well-known in the state of the art and described, for example, in the review articles of Makrides, S.C., *Protein Expr. Purif.* 17 (1999) 183-202; Geisse, S., et al., *Protein Expr. Purif.* 8 (1996) 271-282; Kaufman, R.J., *Mol. Biotechnol.* 16 (2000) 151-161; Werner, R.G., *Drug Res.* 48 (1998) 870-880.
- The antibodies may be present in whole cells, in a cell lysate, or in a partially purified or substantially pure form. Purification is performed in order to eliminate other cellular components or other contaminants, e.g. other cellular nucleic acids or proteins, by standard techniques, including alkaline/SDS treatment, CsCl banding, column chromatography, agarose gel electrophoresis, and others well known in the art. See Ausubel, F., et al., ed. *Current Protocols in Molecular Biology*, Greene Publishing and Wiley Interscience, New York (1987).
- Expression in NS0 cells is described by, e.g., Barnes, L.M., et al., *Cytotechnology* 32 (2000) 109-123; and Barnes, L.M., et al., *Biotech. Bioeng.* 73 (2001) 261-270. Transient expression is described by, e.g., Durocher, Y., et al., *Nucl. Acids. Res.* 30 (2002) E9. Cloning of variable domains is described by Orlandi, R., et al., *Proc. Natl. Acad. Sci. USA* 86 (1989) 3833-3837; Carter, P., et al., *Proc. Natl. Acad. Sci. USA* 89 (1992) 4285-4289; and Norderhaug, L., et al., *J. Immunol. Methods* 204 (1997) 77-87. A preferred transient expression system (HEK 293) is described by Schlaeger, E.-J., and Christensen, K., in *Cytotechnology* 30 (1999) 71-83 and by Schlaeger, E.-J., in *J. Immunol. Methods* 194 (1996) 191-199.
- The control sequences that are suitable for prokaryotes, for example, include a promoter, optionally an operator sequence, and a ribosome binding site.

Eukaryotic cells are known to utilize promoters, enhancers and polyadenylation signals.

5 The monoclonal antibodies are suitably separated from the culture medium by conventional immunoglobulin purification procedures such as, for example, protein A-Sepharose, hydroxylapatite chromatography, gel electrophoresis, dialysis, or affinity chromatography. DNA and RNA encoding the monoclonal antibodies is readily isolated and sequenced using conventional procedures. The hybridoma cells can serve as a source of such DNA and RNA. Once isolated, the DNA may be inserted into expression vectors, which are then transfected into host cells such as 10 CHO cells, HEK 293 cells, or myeloma cells that do not otherwise produce immunoglobulin protein, to obtain the synthesis of recombinant monoclonal antibodies in the host cells.

15 The antibodies according to the invention include, in addition, such antibodies having "conservative sequence modifications", nucleotide and amino acid sequence modifications which do not affect or alter the above-mentioned characteristics of the antibody according to the invention. Modifications can be introduced by standard techniques known in the art, such as site-directed mutagenesis and PCR-mediated mutagenesis. Conservative amino acid substitutions include ones in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have 20 been defined in the art. These families include amino acids with basic side chains (e.g., lysine, arginine, histidine), acidic side chains (e.g., aspartic acid, glutamic acid), uncharged polar side chains (e.g. glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine, tryptophan), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine), beta-branched side chains (e.g., threonine, valine, isoleucine) and aromatic side chains (e.g., tyrosine, 25 phenylalanine, tryptophan, histidine). Thus, a predicted nonessential amino acid residue in a human anti-IL-13R α 1 antibody can be preferably replaced with another amino acid residue from the same side chain family.

30 Amino acid substitutions can be performed by mutagenesis based upon molecular modeling as described by Riechmann, L., et al., Nature 332 (1988) 323-327 and Queen, C., et al., Proc. Natl. Acad. Sci. USA 86 (1989)10029-10033.

Amino acid sequence variants of human IL-13R α 1 antibody are prepared by introducing appropriate nucleotide changes into the antibody DNA, or by peptide synthesis. Such modifications can be performed, however, only in a very limited range, e.g. as described above. For example, the modifications do not alter the abovementioned antibody characteristics such as the IgG isotype and epitope binding, but may improve the yield of the recombinant production, protein stability or facilitate the purification.

Any cysteine residue not involved in maintaining the proper conformation of the anti-IL-13R α 1 antibody also may be substituted, generally with serine, to improve the oxidative stability of the molecule and prevent aberrant crosslinking. Conversely, cysteine bond(s) may be added to the antibody to improve its stability (particularly where the antibody is an antibody fragment such as an Fv fragment).

Another type of amino acid variant of the antibody alters the original glycosylation pattern of the antibody. By altering is meant deleting one or more carbohydrate moieties found in the antibody, and/or adding one or more glycosylation sites that are not present in the antibody. Glycosylation of antibodies is typically N-linked. N-linked refers to the attachment of the carbohydrate moiety to the side chain of an asparagine residue. The tripeptide sequences asparagine-X-serine and asparagine-X-threonine, where X is any amino acid except proline, are the recognition sequences for enzymatic attachment of the carbohydrate moiety to the asparagine side chain. Thus, the presence of either of these tripeptide sequences in a polypeptide creates a potential glycosylation site. Addition of glycosylation sites to the antibody is conveniently accomplished by altering the amino acid sequence such that it contains one or more of the above-described tripeptide sequences (for N-linked glycosylation sites).

Nucleic acid molecules encoding amino acid sequence variants of anti-IL-13R α 1 antibody are prepared by a variety of methods known in the art. These methods include, but are not limited to, isolation from a natural source (in the case of naturally occurring amino acid sequence variants) or preparation by oligonucleotide-mediated (or site-directed) mutagenesis, PCR mutagenesis, and cassette mutagenesis of an earlier prepared variant or a non-variant version of anti-IL-13R α 1 antibody.

Another type of covalent modification involves chemically or enzymatically coupling glycosides to the antibody. These procedures are advantageous in that they do not require production of the antibody in a host cell that has glycosylation capabilities for N- or O-linked glycosylation. Depending on the coupling mode used, the sugar(s) may be attached to (a) arginine and histidine, (b) free carboxyl groups, (c) free sulfhydryl groups such as those of cysteine, (d) free hydroxyl groups such as those of serine, threonine, or hydroxyproline, (e) aromatic residues such as those of phenylalanine, tyrosine, or tryptophan, or (f) the amide group of glutamine. These methods are described in WO 87/05330, and in Aplin, J.D., and Wriston, J.C. Jr., *CRC Crit. Rev. Biochem.* (1981) 259-306.

Removal of any carbohydrate moieties present on the antibody may be accomplished chemically or enzymatically. Chemical deglycosylation can be accomplished by exposing the antibody to trifluoromethanesulfonic acid, or an equivalent compound. This treatment results in the cleavage of most or all sugars except the linking sugar (N-acetylglucosamine or N-acetylgalactosamine), while leaving the antibody intact. Chemical deglycosylation is described by Sojahn, H.T., and Bahl, O.P., *Arch. Biochem. Biophys.* 259 (1987) 52-57 and by Edge, A.S., et al. *Anal. Biochem.* 118 (1981) 131-137. Enzymatic cleavage of carbohydrate moieties on antibodies can be achieved by the use of a variety of endo- and exo- glycosidases as described by Thotakura, N.R., and Bahl, O.P., *Meth. Enzymol.* 138 (1987) 350-359.

Another type of covalent modification of the antibody comprises linking the antibody to one of a variety of nonproteinaceous polymers, e.g., polyethylene glycol, polypropylene glycol, or polyoxyalkylenes, in the manner set forth in US Patent Nos. 4,640,835; 4,496,689; 4,301, 144; 4,670,417; 4,791,192 or 4,179,337.

In yet another aspect, the invention provides isolated B-cells from a transgenic non-human animal, e.g. a transgenic mouse, which express the human anti-IL-13R α 1 antibodies according to the invention. Preferably, the isolated B cells are obtained from a transgenic non-human animal, e.g., a transgenic mouse, which has been immunized with a purified or enriched preparation of IL-13R α 1 antigen and/or cells expressing IL-13R α 1. Preferably, the transgenic non-human animal, e.g. a transgenic mouse, has a genome comprising a human heavy chain transgene and a human light chain transgene encoding all or a portion of an antibody of the

invention. The isolated B-cells are then immortalized to provide a source (e.g. a hybridoma) of human anti-IL-13R α 1 antibodies. Accordingly, the present invention also provides a hybridoma capable of producing human monoclonal antibodies according to the invention. In one embodiment, the hybridoma includes
5 a B cell obtained from a transgenic non-human animal, e.g., a transgenic mouse having a genome comprising a human heavy chain transgene and a human light chain transgene encoding all or a portion of an antibody of the invention, fused to an immortalized cell.

10 In a particular embodiment, the transgenic non-human animal is a transgenic mouse having a genome comprising a human heavy chain transgene and a human light chain transgene encoding all or a portion of an antibody of the invention. The transgenic non-human animal can be immunized with a purified or enriched preparation of IL-13R α 1 antigen and/or cells expressing IL-13R α 1. Preferably, the transgenic non-human animal, e.g. the transgenic mouse, is capable of producing
15 IgG1 isotypes of human monoclonal antibodies to IL-13R α 1.

The human monoclonal antibodies according to the invention can be produced by immunizing a transgenic non-human animal, e.g. a transgenic mouse, having a genome comprising a human heavy chain transgene and a human light chain transgene encoding all or a portion of an antibody of the invention, with a purified
20 or enriched preparation of IL-13R α 1 antigen and/or cells expressing IL-13R α 1. B cells (e.g. splenic B cells) of the animal are then obtained and fused with myeloma cells to form immortal, hybridoma cells that secrete human monoclonal antibodies against IL-13R α 1.

25 In a preferred embodiment, human monoclonal antibodies directed against IL-13R α 1 can be generated using transgenic mice carrying parts of the human immune system rather than the mouse system. These transgenic mice, referred to herein as "HuMab" mice, contain a human immunoglobulin gene miniloci that encodes unrearranged human immunoglobulin genes which include the heavy (μ and γ) and κ light chain (constant region genes), together with targeted mutations that
30 inactivate the endogenous μ and κ chain loci (Lonberg, N., et al., Nature 368 (1994) 856-859). Accordingly, the mice exhibit reduced expression of mouse IgM or K, and in response to immunization, the introduced human heavy and light chain transgenes undergo class switching and somatic mutation to generate high affinity

human IgG monoclonal antibodies (Lonberg, N., et al., *Nature* 368 (1994) 856-859; reviewed in Lonberg, N., *Handbook of Experimental Pharmacology* 113 (1994) 49-101; Lonberg, N., and Huszar, D., *Intern. Rev. Immunol.* 25 (1995) 65-93; and Harding, F., and Lonberg, N., *Ann. N. Acad. Sci.* 764 (1995) 536-546). The preparation of HuMab mice is described in Taylor, L., et al., *Nucleic Acids Research* 20 (1992) 6287-6295; Chen, J., et al., *International Immunology* 5 (1993) 647-656; Tuaillon, N., et al., *Proc. Natl. Acad. Sci USA* 90 (1993) 3720-3724; Choi, T.K., et al., *Nature Genetics* 4 (1993) 117-123; Chen, J., et al., *EMBO J.* 12 (1993) 821-830; Tuaillon, N., et al., *Immunol.* 152 (1994) 2912-2920; Lonberg, N., et al., *Nature* 368 (1994) 856-859; Lonberg, N., *Handbook of Experimental Pharmacology* 113 (1994) 49-101; Taylor, L., et al., *Int. Immunol.* 6 (1994) 579-591; Lonberg, N., and Huszar, D., *Intern. Rev. Immunol.* 25 (1995) 65-93; Harding, F., and Lonberg, N., *Ann. N. Acad. Sci.* 764 (1995) 536-546; Fishwild, D.M., et al., *Nat. Biotechnol.* 14 (1996) 845-851, the contents of all of which are hereby incorporated by reference in their entirety. See further, US Patent Nos. 5,545,806; 5,569,825; 5,625,126; 5,633,425; 5,789,650; 5,877,397; 5,661,016; 5,814,318; 5,874,299; 5,545,807; 5,770,429; WO 98/24884; WO 94/25585; WO 93/1227; WO 92/22645; and WO 92/03918.

To generate fully human monoclonal antibodies to IL-13R α 1, HuMab mice can be immunized with a purified or enriched preparation of IL-13R α 1 antigen and/or cells expressing IL-13R α 1 in accordance with the general method, as described by Lonberg, N., et al., *Nature* 368 (1994) 856-859; Fishwild, D.M., et al., *Nat. Biotechnol.* 14 (1996) 845-851 and WO 98/24884. Preferably, the mice will be 6-16 weeks of age upon the first immunization. For example, a purified or enriched preparation of soluble IL-13R α 1 antigen (e.g. purified from IL-13R α 1-expressing cells) can be used to immunize the HuMab mice intraperitoneally. In the event that immunizations using a purified or enriched preparation of IL-13R α 1 antigen do not result in antibodies, mice can also be immunized with cells expressing IL-13R α 1, e.g., a tumor cell line, to promote immune responses. Cumulative experience with various antigens has shown that the HuMab transgenic mice respond best when initially immunized intraperitoneally (i.p.) with antigen in complete Freund's adjuvant, followed by every other week alternatingly i.p. or s.c. immunizations (for example, up to a total of 6) with antigen in incomplete Freund's adjuvant. The immune response can be monitored over the course of the immunization protocol with plasma samples being obtained by retroorbital bleeds. The plasma can be screened by ELISA, and mice with sufficient titers of anti-IL-13R α 1 human

immunoglobulin can be used for immortalization of corresponding B cells. Mice can be boosted intravenously with antigen 3 to 4 days before sacrifice and removal of the spleen and lymph nodes. It is expected that 2-3 fusions for each antigen may need to be performed. Several mice will be immunized for each antigen. For example, a total of five to twelve HuMab mice of the HCo7 and HCo12 strains can be immunized.

The HCo7 mice have a JKD disruption in their endogenous light chain (kappa) genes (as described in Chen, J., et al., EMBO J. 12 (1993) 821-830), a CMD disruption in their endogenous heavy chain genes (as described in Example 1 of WO 01/14424), a KCo5 human kappa light chain transgene (as described in Fishwild, D.M., et al., Nat. Biotechnol. 14 (1996) 845-851), and a HCo7 human heavy chain transgene (as described in US Patent No. 5,770,429).

The HCo12 mice have a JKD disruption in their endogenous light chain (kappa) genes (as described in Chen, J., et al., EMBO J. 12 (1993) 821-830), a CMD disruption in their endogenous heavy chain genes (as described in Example 1 of WO 01/14424), a KCo5 human kappa light chain transgene (as described in Fishwild, D.M., et al., Nat. Biotechnol. 14 (1996) 845-851), and a HCo12 human heavy chain transgene (as described in Example 2 of WO 01/14424).

The mouse lymphocytes can be isolated and fused with a mouse myeloma cell line using PEG based on standard protocols to generate hybridomas. The resulting hybridomas are then screened for the production of antigen-specific antibodies. For example, single cell suspensions of splenic and lymph node-derived lymphocytes from immunized mice are fused to one-sixth the number of SP 2/0 nonsecreting mouse myeloma cells (ATCC, CRL 1581) with 50% PEG. Cells are plated at approximately 2×10^5 in flat bottom microtiter plate, followed by about two weeks incubation in selective medium.

Individual wells are then screened by ELISA for human anti-IL-13R α 1 monoclonal IgM and IgG antibodies. Once extensive hybridoma growth occurs, medium is analyzed, usually after 10-14 days. The antibody secreting hybridomas are replated, screened again, and if still positive for human IgG, anti-IL-13R α 1 monoclonal antibodies, can be subcloned at least twice by limiting dilution. The stable

subclones are then cultured in vitro to produce antibody in tissue culture medium for characterization.

5 Because CDR sequences are responsible for antibody-antigen interactions, it is possible to express recombinant antibodies according to the invention by constructing expression vectors that include the CDR sequences according to the invention onto framework sequences from a different human antibody (see, e.g.,
10 Riechmann, L., et al., Nature 332 (1998) 323-327; Jones, P., et al., Nature 321 (1986) 522-525; and Queen, C., et al., Proc. Natl. Acad. Sci. U.S.A. 86 (1989) 10029-10033). Such framework sequences can be obtained from public DNA databases that include germline human antibody gene sequences. These germline sequences will differ from mature antibody gene sequences because they will not include completely assembled variable genes, which are formed by V(D)J joining during B cell maturation. Germline gene sequences will also differ from the sequences of a high affinity secondary repertoire antibody at individual evenly across the variable
15 region.

The invention preferably comprises a nucleic acid fragment encoding a polypeptide binding to IL-13R α 1, whereby said polypeptide inhibits the binding of IL-13 to IL-13R α 1, selected from the group consisting of

- 20 a) an antibody heavy chain comprising heavy chain CDRs of SEQ ID NO: 1, 3, 5, 7 or 9;
b) an antibody light chain comprising light chain CDRs of SEQ ID NO: 2, 4, 6, 8 or 10.

25 The reconstructed heavy and light chain variable regions are combined with sequences of promoter, translation initiation, constant region, 3' untranslated, polyadenylation, and transcription termination to form expression vector constructs. The heavy and light chain expression constructs can be combined into a single vector, co-transfected, serially transfected, or separately transfected into host cells which are then fused to form a single host cell expressing both chains.

30 Accordingly, the invention provides a method for the production of a recombinant human antibody according to the invention, comprising expressing a nucleic acid encoding

- a) an antibody heavy chain comprising heavy chain CDRs of SEQ ID NO:1, 3, 5, 7 or 9;
- b) an antibody light chain comprising light chain CDRs of SEQ ID NO: 2, 4, 6, 8 or 10.

5

The invention further comprises the use of an antibody according to the invention for the detection of IL-13R α 1 in vitro, preferably by an immunological assay determining the binding between IL-13R α 1 of a sample and the antibody according to the invention.

10

In another aspect, the present invention provides a composition, e.g. a pharmaceutical composition, comprising one or a combination of human monoclonal antibodies, or the antigen-binding portion thereof, of the present invention, formulated together with a pharmaceutically acceptable carrier.

15

As used herein, "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like that are physiologically compatible. Preferably, the carrier is suitable for intravenous, intramuscular, subcutaneous, parenteral, spinal or epidermal administration (e.g. by injection or infusion).

20

A "pharmaceutically acceptable salt" refers to a salt that retains the desired biological activity of the antibody and does not impart any undesired toxicological effects (see e.g. Berge, S.M., et al., J. Pharm. Sci. 66 (1977) 1-19). Such salts are included in the invention. Examples of such salts include acid addition salts and base addition salts. Acid addition salts include those derived from nontoxic inorganic acids, such as hydrochloric salts.

25

A composition of the present invention can be administered by a variety of methods known in the art. As will be appreciated by the skilled artisan, the route and/or mode of administration will vary depending upon the desired results.

30

To administer a compound of the invention by certain routes of administration, it may be necessary to coat the compound with, or coadminister the compound with, a material to prevent its inactivation. For example, the compound may be administered to a subject in an appropriate carrier, for example, liposomes, or a

diluent. Pharmaceutically acceptable diluents include saline and aqueous buffer solutions.

5 Pharmaceutically acceptable carriers include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. The use of such media and agents for pharmaceutically active substances is known in the art.

10 The phrases "parenteral administration" and "administered parenterally" as used herein means modes of administration other than enteral and topical administration, usually by injection, and includes, without limitation, intravenous, intramuscular, intraarterial, intrathecal, intracapsular, intraorbital, intracardiac, intradermal, intraperitoneal, transtracheal, subcutaneous, subcuticular, intraarticular, subcapsular, subarachnoid, intraspinal, epidural and intrasternal injection and infusion.

15 These compositions may also contain adjuvants such as preservatives, wetting agents, emulsifying agents and dispersing agents. Prevention of presence of microorganisms may be ensured both by sterilization procedures, supra, and by the inclusion of various antibacterial and antifungal agents, for example, paraben, chlorobutanol, phenol, sorbic acid, and the like. It may also be desirable to include isotonic agents, such as sugars, sodium chloride, and the like into the compositions. 20 In addition, prolonged absorption of the injectable pharmaceutical form may be brought about by the inclusion of agents which delay absorption such as aluminum monostearate and gelatin.

25 Regardless of the route of administration selected, the compounds of the present invention, which may be used in a suitable hydrated form, and/or the pharmaceutical compositions of the present invention, are formulated into pharmaceutically acceptable dosage forms by conventional methods known to those of skill in the art.

30 Actual dosage levels of the active ingredients in the pharmaceutical compositions of the present invention may be varied so as to obtain an amount of the active ingredient which is effective to achieve the desired therapeutic response for a particular patient, composition, and mode of administration, without being toxic

5 to the patient. The selected dosage level will depend upon a variety of pharmacokinetic factors including the activity of the particular compositions of the present invention employed, or the ester, salt or amide thereof, the route of administration, the time of administration, the rate of excretion of the particular compound being employed, the duration of the treatment, other drugs, compounds and/or materials used in combination with the particular compositions employed, the age, sex, weight, condition, general health and prior medical history of the patient being treated, and like factors well known in the medical arts.

10 The composition must be sterile and fluid to the extent that the composition is deliverable by syringe. In addition to water, the carrier can be an isotonic buffered saline solution, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), and suitable mixtures thereof.

15 Proper fluidity can be maintained, for example, by use of coating such as lecithin, by maintenance of required particle size in the case of dispersion and by use of surfactants. In many cases, it is preferable to include isotonic agents, for example, sugars, polyalcohols such as mannitol or sorbitol, and sodium chloride in the composition. Long-term absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate or gelatin.

20 The following examples, references, sequence listing and figures are provided to aid the understanding of the present invention, the true scope of which is set forth in the appended claims. It is understood that modifications can be made in the procedures set forth without departing from the spirit of the invention.

Description of the Sequence Listing

25 SEQ ID NO:1 heavy chain variable domain of HuMab LC5002-002

SEQ ID NO:2 light chain variable domain of HuMab LC5002-002

SEQ ID NO:3 heavy chain variable domain of HuMab LC5002-003

SEQ ID NO:4 light chain variable domain of HuMab LC5002-003

- SEQ ID NO:5 heavy chain variable domain of HuMab LC5002-005
- SEQ ID NO:6 light chain variable domain of HuMab LC5002-005
- SEQ ID NO:7 heavy chain variable domain of HuMab LC5002-007
- SEQ ID NO:8 light chain variable domain of HuMab LC5002-007
- 5 SEQ ID NO:9 heavy chain variable domain of HuMab LC5002-018
- SEQ ID NO:10 light chain variable domain of HuMab LC5002-018
- SEQ ID NO:11 κ light chain constant region
- SEQ ID NO:12 γ 1 heavy chain constant region

Description of the Figures

- 10 Figure 1 shows binding of anti-IL-13R α 1 antibodies to immobilized recombinant human IL-13R α 1 polypeptide. Included are polyclonal rabbit-anti-human IL-13R α 1 antibody AF152 (R&D systems) and anti-KLH as a negative control HuMab.
- 15 Figure 2 shows inhibition of IL-13 binding to immobilized IL-13R α 1/IL-4R α receptor by anti-IL-13R α 1 antibodies.
- Figure 3 shows the blockade of IL-13 binding to CHO cells (expressing IL-13R α 1 and IL-4R α 2) by anti-IL-13R α 1 antibodies. As a positive control, a commercially available polyclonal anti-IL-13R α 1 antibody (AF152, R&D Systems, Minneapolis, MN) was included.
- 20 Figure 4 shows binding of anti-IL-13R α 1 antibodies to hIL-13R α 1 and binding properties to functionally related receptors hIL-13R α 2 and hIL-4R α .

Figure 5 shows the capacity of anti-IL-13R α 1 antibodies to bind to immobilized recombinant murine IL-13R α 1 polypeptide. Included are polyclonal goat-anti-human IL-13R α 1 antibody AF152 (R&D Systems) and anti-KLH as a negative control HuMab.

Examples

Example 1

Generation of hybridomas

The human monoclonal antibodies according to the invention can be produced by immunizing a transgenic non-human animal, e. g. a transgenic mouse having a genome comprising a human heavy chain transgene and a human light chain transgene encoding all or a portion of an antibody of the invention, with cells expressing human IL-13R α 1. B cells (e.g. splenic B cells) of the animal are then obtained and fused with myeloma cells to form immortal, hybridoma cells that secrete human monoclonal antibodies against IL-13R α 1. Human monoclonal antibodies directed against human IL-13R α 1 can be generated using transgenic mice carrying parts of the human immune system rather than the mouse system. These transgenic mice, referred to herein as "HuMab" mice, contain a human immunoglobulin gene minilocus that encodes unrearranged human immunoglobulin genes which include the heavy (μ and γ) and κ (kappa) light chain (constant region genes), together with targeted mutations that inactivate the endogenous μ and kappa chain loci (Lonberg N., et al., Nature 368 (1994) 856-859). Accordingly, the mice exhibit reduced expression of mouse IgM or κ , and in response to immunization, the introduced human heavy and light chain transgenes undergo class switching and somatic mutation to generate high affinity human IgG monoclonal antibodies. To generate fully human monoclonal antibodies to human IL-13R α 1, HuMab mice can be immunized with cells expressing human IL-13R α 1 in accordance with the general method, as described by Lonberg, N., et al., Nature 368 (1994) 856-859; Fishwild, D.M., et al., Nat. Biotechnol. 14 (1996) 845-851 and WO 98/24884. Preferably, the mice will be 6-16 weeks of age upon the first immunization. For example, IL-13R α 1 transfected cells can be used to immunize the HuMab mice intraperitoneally. The immune response can be monitored over the course of the immunization protocol with plasma samples being obtained by

retroorbital bleeds. The plasma can be screened by ELISA and/or FACS. Mice with sufficient titers of anti-human IL-13R α 1 human immunoglobulin can be used for immortalization of corresponding B cells. Mice can be boosted intravenously with antigen 3 to 4 days before sacrifice and removal of the spleen and lymph nodes. For example, HuMab mice of the HCo7 or HCo12 strain can be immunized. The HCo7 mice have a JKD disruption in their endogenous light chain (kappa) genes (as described in Chen et al. (1993) EMBO J. 12: 821-830), a CMD disruption in their endogenous heavy chain genes (as described in Example 1 of WO 01/14424), a KCo5 human kappa light chain transgene (as described in Fishwild, D.M., et al. (1996) Nature Biotechnology 14:845-851), and a HCo7 human heavy chain transgene (as described in US 5,770,429). The HCo12 mice have a JKD disruption in their endogenous light chain (kappa) genes (as described in Chen, J., et al., EMBO J. 12 (1993) 821-830), a CMD disruption in their endogenous heavy chain genes (as described in Example 1 of WO 01/14424)) a KCo5 human kappa light chain transgene (as described in Fishwild, D.M., et al., Nat. Biotechnol. 14 (1996) 845-851, and a HCo12 human heavy chain transgene (as described in Example 2 of WO 01/14424). The mouse lymphocytes can be isolated and fused with a mouse myeloma cell line using PEG based on standard protocols to generate hybridomas. The resulting hybridomas are then screened for the production of antigen-specific antibodies. For example, single cell suspensions of splenic and lymph node derived lymphocytes from immunized mice are fused to SP 2/0 nonsecreting mouse myeloma cells (ATCC, CRL 1581) with 50% PEG. Cells are plated at approximately 0.75×10^7 in flat bottom micro titer plate, followed by about two weeks incubation in selective medium. Individual wells are then screened by ELISA and/or FACS for human anti-IL-13R α 1 monoclonal IgM and IgG antibodies. Once extensive hybridoma growth occurs, the antibody secreting hybridomas are replated, screened again, and if still positive for human IgG, anti-IL-13R α 1 monoclonal antibodies, can be subcloned at least twice by limiting dilution. The stable subclones are then cultured in vitro to produce antibody in tissue culture medium for characterization.

Immunization procedure of transgenic mice

Three HCo7 mice (3 males), strain GG2201 (Medarex, San Jose, CA, USA) and 2 HCo12 mice (1 male and 1 female), strain GG2198 (Medarex, San Jose, CA, USA) were immunized with 1×10^6 HEK293 cells, transfected with an expression vector

for IL-13R α 1. In total eight immunizations were given alternating intraperitoneally (i.p.) and subcutaneous (s.c.) at the tail base. For the first immunization, 100 μ l of 1×10^6 HEK293: IL-13R α 1 cells, was mixed with 100 μ l complete Freund's adjuvant (CFA; Difco Laboratories, Detroit, USA). For all other immunizations, 100 μ l of cells in PBS was mixed with 100 μ l incomplete Freund's adjuvant (ICFA; Difco).

Boosting of mice

When serum titers of anti-IL-13R α 1 were found to be sufficient, mice were additionally boosted twice with 1×10^6 HEK293: IL-13R α 1 cells in 200 μ l PBS intravenously (i.v.) 4 and 3 days before fusion.

Example 2

Testing the binding of HuMab to immobilized IL-13R α 1 by ELISA

To determine the ability of the antibodies of the invention to bind to recombinant IL-13R α 1, the extracellular domain of IL-13R α 1 (R&D Systems, UK) was dissolved in PBS (1 μ g/ml) and allowed to adsorb to microtiter plates (NUNC Maxisorb) by incubation over night at 4°C. After washing the plates with washing buffer (WB = 0.9 % NaCl; 0.1% Tween® 20) unspecific binding sites were blocked by addition of 100 μ l incubation buffer (IB = PBS with 1% crotein C and 0.1% Tween® 20) and incubation for 30 min at room-temperature (RT). Then, serially diluted HuMab and control antibodies (100 μ l /well; dilutions in IB) were added and incubated for 1 hour at RT. The plates were again washed, and bound human antibodies were detected by incubation with peroxidase-conjugated rabbit anti-human kappa (DAKO, Denmark) in a final dilution of 1:500 in IB. Polyclonal goat anti-hIL-13R α 1 antibodies were detected with peroxidase-conjugated polyclonal donkey anti-goat IgG (Santa Cruz; dilution 1:1000 in IB). After incubation for 1h at RT and a subsequent washing step, the plates were developed with ready-to-use ABTS® solution (Roche Diagnostics GmbH) at RT in the dark. Absorbance was measured at 405 nm after absorbance of the highest concentration reached a sufficient OD.

All antibodies against IL-13R α 1 tested were able to bind to immobilized extracellular domains of human IL-13R α 1. The EC50 values determined were in the range of 0.5 - 2 nM for the various LC antibodies tested. The negative control HuMab anti-KLH did not bind to the immobilized extracellular domains of IL-

13R α 1. Polyclonal goat-anti human IL-13R α 1 antibody, included as positive control, also bound efficiently to the immobilized extracellular domains of IL-13R α 1 (Fig.1).

Example 3

5 Inhibition of IL-13 binding to IL-13R α 1/IL-4R α heterodimer (ELISA)

Microtiter plates were coated with 100 μ l hIL-13R α 1:hFc chimeric protein (R&D Systems, UK) in PBS at 3 μ g/ml at 4°C overnight on a shaker. After washing the plates with WB, serially diluted HuMab and control antibodies (100 μ l /well; dilutions in IB) were added and incubated for 30 min at RT. The plates were again
10 washed, and then a mixture of IL-13 (R&D Systems, UK; 0.5 μ g/ml; dilution with IB) and IL-4R α (R&D Systems, UK; 0.75 μ g/ml; dilution with IB) were added and incubated for 1 h at RT. After washing the plates 100 μ l biotinylated anti IL-13 antibody (BAF213; R&D Systems, UK) in a concentration of 0.4 μ g/ml was added and incubated for 1h at RT. After washing the plates, bound IL-13 was detected by
15 peroxidase-coupled streptavidin (Roche Diagnostics GmbH, DE) in a dilution of 1:5000 in IB (incubation period 1h at RT). Finally, plates were washed and developed with ready-to-use ABTS[®] solution (Roche Diagnostics GmbH, DE) at room temperature (RT) in the dark. Absorbance was measured at 405 nm after 45 to 60 minutes.

20 Antibodies LC5002-002, LC5002-003, LC5002-005, LC5002-007 and LC5002-018 were able to inhibit binding of IL-13 to the heterodimeric receptor with maximal inhibition values ranging from approximately 50% to 80-85%. Positive control was AF152 (polyclonal rabbit antibody). As expected, the negative control anti-KLH did not inhibit binding of IL-13 to the heterodimeric receptor. The IC₅₀ values obtained
25 were between 1.5 nM and 10.1 nM for LC5002-002, LC5002-003, LC5002-005, LC5002-007 and LC5002-018 (Fig.2).

Example 4

Radioligand Binding Assay

30 ¹²⁵I-IL-13 binding assay was performed using CHO cells expressing human IL-13R α 1 and human IL-4R α in binding buffer (25 mM HEPES, 150 mM NaCl, 1 mM CaCl₂, 5 mM MgCl₂, and 0.5% bovine serum albumin, adjusted to pH 7.2). 1x10⁵

cells per well were mixed with the antibodies and preincubated for 15 minutes to 1 hour. 0.1 nM ^{125}I -IL-13 was added, and the mix was incubated at 4°C for 4 hours. The concentration of ^{125}I -IL-13 used in the assay was determined from saturation binding analysis, competition analysis and determination of input ^{125}I -IL-13 to reach equilibrium binding with the cell line. Samples were harvested onto a GF/C filter plate pretreated with 1% PEI/0.5% BSA and counted on Packard TopCount Scintillation counter. Data analysis was performed in PRISM using nonlinear regression curve fit (GraphPad Software, San Diego, CA).

All antibodies against IL-13R α 1 tested block binding of labeled IL-13 to the IL-13R α 1/IL-4R α complex. The calculated IC₅₀ values for antibodies LC5002-002, LC5002-003, LC5002-005, LC5002-007 and LC5002-018 were between 0.09 nM and 0.32 nM and 84.8nM for AF152 (Fig.3).

Example 5

Inhibition of IL-13 induced upregulation of CD23 on human B cells and monocytes by HuMab

Peripheral blood mononuclear cells (PBMC) were isolated by a Ficoll Hypaque density gradient. After washing the cells with RPMI they were resuspended in RPMI/10% FCS and distributed at 3×10^5 PBMC/well (volume 50 μl) in 96 well flat bottom microtiter plates (Corning Incorporated Costar). Then, 25 μl of an anti-human CD40 antibody (Immunotech) in a final concentration 0.5 $\mu\text{g}/\text{ml}$ in RPMI/10% FCS and 25 μl of an anti-human IgA+IgG+IgM antibody (Immunoresearch) in a final concentration of 10 $\mu\text{g}/\text{ml}$ in RPMI/10%FCS were added. Then serially diluted HuMab and control antibodies (50 $\mu\text{l}/\text{well}$; dilutions in RPMI/10% FCS) were added and the cells incubated for 30 min in the incubator (37°C; 5% CO₂). Then recombinant human IL-13 (R&D Systems) in a final concentration of 0.67 ng/ml was added (50 $\mu\text{l}/\text{well}$) and the cells incubated for 72h at 37°C/5% CO₂. After this incubation the plates were centrifuged and the medium aspirated. For detachment of adherent cells 200 μl of Accutase (PAA) was added and the cells incubated for approximately 5 min at 37°C; 5% CO₂. The cells were detached by repeated flushing and transferred to a round bottom plate. After centrifugation and aspiration of the supernatants the cells were incubated with 200 μl of a mixture of anti-CD23-PE, anti-CD20-FITC and anti-CD14-APC (all from BD Biosciences Pharmingen, San Diego, CA). The cells were incubated for 30 min

at 4°C, then centrifuged and the supernatants aspirated. This washing step was repeated once, and finally the cells were resuspended in 200 µl of PBS/0.1% human serum albumin and analysed in a FACS Calibur flow cytometer (BD Biosciences Pharmingen, San Diego, CA) using the CellQuest software. In most cases, 10 000 events were acquired and gated on a light scatter gate to include only viable lymphocytes and monocytes. The cells were pregated on a CD19 positive cluster for B lymphocytes or a CD14 positive cluster for monocytes and analyzed further for CD23 expression.

The observed IC₅₀ values for inhibiting of CD 23 upregulation on B-lymphocytes were between 0.5 nM and >70 nM for antibodies LC5002-002, LC5002-003, LC5002-005, LC5002-007 and LC5002-018, and 13.6 nM for AF152. A similar profile was found for inhibition of IL-13 induced CD23 upregulation on human monocytes. On monocytes the IC₅₀ values were between 0.1 nM and 62.8 nM for antibodies LC5002-002, LC5002-003, LC5002-005, LC5002-007 and LC5002-018, and 62.9 nM for AF152.

Example 6

TF-1 proliferation assay in response to IL-13 or IL-4 as stimulus

TF-1 cells (ATCC # CRL 2003) were grown in media containing ATCC modified RPMI, 10% FBS, 1X Pencillin/Streptomycin, 2ng/ml GM-CSF. A day prior to the assay the cells were maintained in GM-CSF free media. 5 x 10³ cells per well were incubated with appropriate concentrations of anti-IL-13Rα1 antibodies at 37°C for 1 hour. Then the cells were stimulated with 2ng/ml of human IL-13 (R&D Systems, Minneapolis, MN) or 0.1 ng/ml of human IL-4 (R&D Systems, Minneapolis, MN) and incubated at 37°C for 48 hours. The cells were pulsed with 0.5 µCi ³H-Thymidine and incubated at 37°C for 16-18 hours. Samples were harvested onto GFC plates pretreated with 1% PEI/0.25% BSA using Perkin Elmer Filtermate 96 harvester. The GFC plates were counted on a Perkin Elmer Top count Scintillation counter. Data analysis was performed in PRISM using nonlinear regression curve fit (GraphPad Software, San Diego, CA).

Anti-KLH antibody did not show any inhibition in this assay. The same was true for LC5002-007. All other antibodies inhibited the response, even though LC5002-007 inhibited the response with higher IC value than the other antibodies. The observed

IC₅₀ values for the different antibodies were: 13.50 nM for AF152, 9.21 nM for LC5002-002, 3.07 nM for LC5002-003 and 0.39 nM for LC5002-005. A similar profile was found for IL-4-induced TF-1 cell proliferation, however the potency of the antibodies was decreased compared to IL-13-induced responses. IC₅₀ values for IL-4-induced proliferation were 0.02 nM for anti-IL4R antibody, 74.37 nM for AF152 and for antibodies LC5002-002, LC5002-003, LC5002-005, LC5002-007 and LC5002-018 between 4.68 nM and 60 nM.

Example 7

Inhibition of eotaxin production in response to IL-13 by human lung fibroblast

The assay was performed using HFL-1 cells (Human Lung Fibroblast, ATCC # CCL-153). Cells were plated at a density of 100,000 cells per well in a 12-well plate and incubated at 37°C for 72 hours to reach confluency. Cells were then starved in serum free medium for 24 h and treated with anti-IL-13R α 1 antibody at 37°C for 1 h. Following this treatment cells were stimulated with 10 ng/ml IL-13 (R&D Systems, Minneapolis, MN) at 37°C for 48 h. Supernatants were collected and eotaxin determinations were done using commercially available ELISA from R&D Systems (Cat. No. DTX00). Absorbance was read using Spectromax microplate reader and the data was analyzed using PRISM (GraphPad Software, San Diego, CA).

The antibodies tested showed different capability to inhibit eotaxin release. With the exception of LC5002-007 all other antibodies tested showed some inhibition. The calculated average IC₅₀ values from 3 to 4 different experiments were 11.5 nM for AF152 and between 2.45 nM and 19.8nM for antibodies LC5002-002, LC5002-003, LC5002-005, LC5002-007 and LC5002-018.

Example 8

Inhibition of IL-13-induced Stat-6 phosphorylation in human bronchial smooth muscle cells

Human Bronchial Smooth Muscle Cells (BSMC; Clonetics, Cat. No CC-2576) were grown following the manufacturer's instructions. Cells were grown in 12-well tissue culture plates until they reached confluency. Cells were starved for 24h in serum-free medium and variable amounts of antibody were added. Plates were incubated

for 1h and then stimulated with 2.5 ng/ml IL-13 (R&D System). After 15 min incubation, supernatant was removed, cells were washed with phosphate buffer and 100 µl of lysis buffer was added. The mix was briefly sonicated on ice and centrifuged. Lysate was used for Western Blot detection of phosphorylated Stat-6. Equal amounts of protein were loaded onto an SDS-gel, run and transferred to a membrane. Anti-Stat-6 antibody was from Santa Cruz Biotechnology (Cat. No. SC-11762R) and a secondary antibody coupled to peroxidase was used. Detection was done using the ECL Plus System from Amersham (Cat No RPN 2132). Quantitation was done in a Typhoon 9400 Imager.

- Both HuMabs tested in this assay (LC50002-003 and LC5002-005) inhibited IL-13-induced Stat-6 phosphorylation. The potency found in this assay was similar to that of other functional assays. The calculated IC_{50} values were 18.64 nM for AF152, 5.98 nM for LC5002-003 and 1.18 nM for LC5002-005.

Example 9

- Cloning and sequence analysis of anti-hIL-13R α 1 HuMab variable domains (κ -light and γ 1-heavy chains)

The nucleotide sequences coding for the light chain variable region V_L and the heavy chain variable region V_H of the anti hIL-13R α 1 HuMabs were isolated by a standard cDNA synthesis / PCR procedure. Total RNA was prepared from 1×10^6 – 1×10^7 hybridoma cells using the GeneRacer™ Kit (Invitrogen). Hybridoma derived RNA was used as a template for the 1st strand cDNA synthesis and ligation of the GeneRacer™ Oligo-dT Primer. 2nd-strand cDNA synthesis and further PCR amplification of V_L and V_H encoding cDNA fragments were performed with reverse light and heavy chain primers complementary to nucleotide sequences of the κ -light and γ 1-heavy chain constant region and 5'-specific GeneRacer™ primers, respectively. The PCR products were cloned using the TOPO™ TA cloning kit from Invitrogen™ Life Technologies and pCR4-TOPO™ as a cloning vector. Cloned PCR products were identified by restriction mapping of the appropriate plasmids using EcoRI for digestion and expected / calculated DNA fragment sizes of about 740 and 790 bp for V_L and V_H , respectively. The DNA sequence of cloned PCR fragments was determined by double strand sequencing. The GCG (Genetics Computer Group, Madison, Wisconsin) software package version 10.2 and Vector-NTI 8 (InforMax, Inc) was used for general data processing. DNA and protein

sequences were aligned using the GCG modul CLUSTALW. Sequence alignments were made using the program GENEDOC (version 2.1).

Example 10

Construction of expression plasmids for an anti-hIL-13R α 1 IgG1 HuMab

5 The anti-hIL-13R α 1 HuMab light and heavy chain encoding genes were separately assembled in mammalian cell expression vectors. Thereby the gene segments encoding the anti-hIL-13R α 1 HuMab light chain variable region (V_L) and the human κ -light chain constant region (C_L, SEQ ID NO: 11) were joined as were gene segments for the anti-hIL-13R α 1 HuMab heavy chain variable region (V_H) and the
10 human γ 1-heavy chain constant region (C_{H1}-Hinge-C_{H2}-C_{H3}, SEQ ID NO: 12). General information regarding the nucleotide sequences of human light and heavy chains from which the codon usage is given in: Kabat, E. A., Wu, T. T., Perry, H. M., Gottesman, K. S., and Foeller, C. ,(1991) Sequences of Proteins of Immunological Interest, Fifth Ed., NIH Publication No. 91-3242. The transcription unit of the anti-
15 hIL-13R α 1 HuMab κ -light chain is composed of the following elements:

- The immediate early enhancer and promoter from the human cytomegalovirus (HCMV),
- A synthetic 5'-UT including a Kozak sequence,
- A murine immunoglobulin heavy chain signal sequence including the signal
20 sequence intron,
- The cloned anti-hIL-13R α 1 HuMab variable light chain cDNA arranged with a unique BsmI restriction site at the 5' end and a splice donor site and a unique NotI restriction site at the 3' end,
- The genomic human κ -gene constant region, including the intron 2 mouse
25 Ig- κ enhancer [Picard, D., and Schaffner, W., Nature 307 (1984) 80-82] and
- The human immunoglobulin κ -polyadenylation ("poly A") signal sequence.

The transcription unit of the anti-hIL-13R α 1 HuMab γ 1-heavy chain is composed of the following elements:

- 30 • The immediate early enhancer and promoter from the human cytomegalovirus (HCMV),
- A synthetic 5'-UT including a Kozak sequence,

- A modified murine immunoglobulin heavy chain signal sequence including the signal sequence intron,
- The cloned anti-hIL-13R α 1 HuMab variable heavy chain cDNA arranged with a unique BsmI restriction site at the 5' and a splice donor site and a unique NotI restriction site at the 3' end,
- The genomic human γ 1-heavy gene constant region, including the mouse Ig μ -enhancer (Neuberger, M.S., EMBO J. 2 (1983) 1373-1378),
- The human γ 1-immunoglobulin polyadenylation ("poly A") signal sequence.

Functional elements of the anti-hIL-13R α 1 HuMab κ -light chain and γ 1-heavy chain expression plasmids:

Besides the anti-hIL-13R α 1 HuMab κ -light chain or γ 1-heavy chain expression cassette these plasmids contain

- A hygromycin resistance gene
- An origin of replication, oriP, of Epstein-Barr virus (EBV)
- An origin of replication from the vector pUC18 which allows replication of this plasmid in *E. coli*, and
- A β -lactamase gene which confers ampicillin resistance in *E. coli*.

Example 11

Construction of expression plasmids for mutant (variant) anti-hIL-13R α 1 IgG1

Expression plasmids encoding mutant anti-hIL-13R α 1 γ 1-heavy chains can be created by site-directed mutagenesis of the wild type expression plasmids using the QuickChangeTM Site-Directed mutagenesis Kit (Stratagene) and are described in table 1. Amino acids are numbered according to EU numbering [Edelman, G.M., et al., Proc. Natl. Acad. Sci. USA 63 (1969) 78-85; Kabat, E. A., Wu, T. T., Perry, H. M., Gottesman, K. S., and Foeller, C., (1991) Sequences of Proteins of Immunological Interest, Fifth Ed., NIH Publication No. 91-3242].

Table 1

Mutation	Description
PVA-236; GLPSS331 as specified by E233P; L234V; L235A; delta G236;	The amino acid sequence Glu ₂₃₃ Leu ₂₃₄ Leu ₂₃₅ Gly ₂₃₆ of the human γ 1-heavy chain is replaced by the amino acid sequence Pro ₂₃₃ Val ₂₃₄ Ala ₂₃₅ of the human γ 2-heavy chain.
A327G; A330S; P331S	The amino acid sequence Ala ₃₂₇ Leu ₃₂₈ Pro ₃₂₉ Ala ₃₃₀ Pro ₃₃₁ of the human γ 1-heavy chain is replaced by the amino acid sequence Gly ₃₂₇ Leu ₃₂₈ Pro ₃₂₉ Ser ₃₃₀ Ser ₃₃₁ of the human γ 4-heavy chain
L234A; L235A	The amino acid sequence Leu ₂₃₄ Leu ₂₃₅ of the human γ 1-heavy chain is replaced by the amino acid sequence Ala ₂₃₄ Ala ₂₃₅

Example 12

Production of recombinant anti-hIL-13R α 1 HuMabs

- 5 Recombinant HuMabs were generated by transient transfection of adherent HEK293-EBNA cells (ATTC CRL-10852) cultivated in DMEM (Gibco) supplemented with 10 % ultra-low IgG FCS (Gibco), 2 mM Glutamine (Gibco), 1% v/v nonessential aminoacids (Gibco) and 250 μ g/ml G418 (Roche Diagnostics GmbH, DE). For transfection Eugene[™] 6 (Roche Diagnostics GmbH, DE)
- 10 transfection reagent was used in a ratio of reagent (μ l) to DNA (μ g) ranging from 3:1 to 6:1. Immunoglobulin light and heavy chains were expressed from two different plasmids using a molar ratio of light chain to heavy chain encoding plasmid from 1:2 to 2:1. HuMab containing cell culture supernatants were harvested at day 4 to 11 after transfection.
- 15 General information regarding the recombinant expression of human antibody in e.g. HEK293 is given in: Meissner, P., et al., Biotechnol Bioeng 75 (2001) 197-203.

Example 13

a) Affinity analysis of HuMabs LC5002-003, -005, and -007 using chimeric hIL-13R α 1:hFc

For interaction analysis a Biacore 3000 instrument was used. As running and reaction buffer, HBS-P (10 mM HEPES, 150 mM NaCl, 0.005 % polysurfactant P, pH 7.4) at 25°C was used. Capturing molecules (goat anti-human-IgG, Fc γ specific) were amine-coupled at a concentration of 20 μ g/ml at a flow rate of 5 μ l/min for 20 minutes. HuMabs were injected at a concentration of 1 μ g/ml at a flow rate of 10 μ l/min for 1 minute. Blocking of the free goat anti human IgG, Fc γ was achieved by injecting human gamma globulin at 500 nM and 30 μ l/min for 3 minutes. Analyte (hIL-13R α 1:hFc chimeric protein) was injected for two minutes at five concentrations between 5.63 nM and 90 nM and washed with HBS-P for five minutes. Regeneration of the surface was accomplished by two injections of 100 mM HCl for 1 min each. The chip, assay format and sequence of injections and kinetic data correspond to the description in table 2. Negative control data (e.g. buffer curves) were subtracted from sample curves for correction of system intrinsic baseline drift and for noise signal reduction. BiaEvaluation version 4.01 was used for analysis of sensorgrams and for calculation of affinity data. Kinetic data were calculated by fitting kinetic data to a 1:1 Langmuir binding model (Table 2).

20

Table 2:

Affinity analysis of HuMabs using hIL-13R α 1:hFc. Data analysis based on 1:1 Langmuir binding model.

Chip	Capturing	Ligand	Analyte	k_a (1/Ms)	k_d (1/s)	K_D (M)
CM5	Anti-hFc γ	LC5002-003	hIL-13 R α 1:hFc	2.1×10^5	1.4×10^{-6}	$< 6.5 \times 10^{-12}$
CM5	Anti-hFc γ	LC5002-005	hIL-13 R α 1:hFc	1.73×10^5	3.12×10^{-6}	1.8×10^{-11}
CM5	Anti-hFc γ	LC5002-007	hIL-13 R α 1:hFc	1.19×10^5	1×10^{-6}	$< 8.4 \times 10^{-12}$

b) Affinity analysis of HuMabs LC5002-003, -005 and -007 using the extracellular domain of hIL-13R α 1 cleaved from of hIL-13R α 1:hFc chimeric protein

For interaction analysis, a Biacore 3000 instrument was used. Running and reaction buffer was HBS-P (10 mM HEPES, 150 mM NaCl, 0.005 % polysurfactant P, pH 7.4) at 25°C. Capturing antibody molecules (anti-hFc γ) were amine-coupled at a concentration of 100 μ g/ml at a flow rate of 5 μ l/min for 20 minutes. HuMabs were injected at concentrations of 10 μ g/ml at a flow rate of 10 μ l/min for 30 seconds. Cleaved hIL-13R α 1 molecules (M_w 40kDa) were injected for 200 seconds at seven concentrations between 1.56 nM and 100 nM and washed with HBS-P for five minutes. Regeneration of the surface was accomplished by two injections of 100 mM HCl for 1 min each at a flow rate of 10 μ g/ml. The chip, assay format and sequence of injections and kinetic data correspond to the description in the following Table 3. Kinetic data were calculated by fitting kinetic data to a 1:1 Langmuir binding model.

Table 3:

Chip	Capturing	Ligand	Analyte	k_a (1/Ms)	k_d (1/s)	K_D (M)
C1	Anti-hFc γ	LC5002-002	hIL-13R α 1	1.1×10^6	6.5×10^{-4}	6.2×10^{-10}
C1	Anti-hFc γ	LC5002-003	hIL-13R α 1	1.3×10^6	5.1×10^{-4}	3.9×10^{-10}
C1	Anti-hFc γ	LC5002-005	hIL-13R α 1	1.4×10^6	3.0×10^{-4}	2.2×10^{-10}
C1	Anti-hFc γ	LC5002-007	hIL-13R α 1	1.9×10^5	8.3×10^{-4}	4.4×10^{-9}
C1	Anti-hFc γ	LC5002-005, mutant L234A; L235A	hIL-13R α 1	1.4×10^6	2.9×10^{-4}	2.1×10^{-10}

- c) Comparative affinity analysis of recombinant variants of LC5002-005 using the extracellular domain of hIL-13R α 1 cleaved from hIL-13R α 1:hFc chimeric protein

In these experiments, the affinity of the original IgG1 derived from hybridoma was compared with the affinities of the recombinant variant IgG1-Ala-Ala. For interaction analysis, a Biacore 3000 instrument was used. Running and reaction buffer was HBS-P (10 mM HEPES, 150 mM NaCl, 0.005 % polysurfactant P, pH 7.4) at 25°C. Capturing antibody molecules (anti-hFc γ) were amine-coupled at a concentration of 20 μ g/ml at a flow rate of 5 μ l/min for 20 minutes. HuMabs were injected at concentrations of 10 μ g/ml at a flow rate of 10 μ l/min for 1 minute. Cleaved hIL-13 R α 1 molecules (analyte) were injected for five minutes at eight concentrations between 1,56 nM and 200 nM and washed with HBS-P for five minutes. Regeneration of the surface was accomplished by two injections of 100 mM HCl for 1 min each. The chip, assay format and sequence of injections and kinetic data correspond to the description in Table 4. Kinetic data were calculated by fitting kinetic data to a bivalent analyte binding model.

Table 4:

Chip	Capturing	Ligand	Analyte	k_{a1} (1/Ms)	k_{d1} (1/s)	k_{a2} (1/RUs)	k_{d2} (1/s)	K_D (M)
CM5	Anti-hFc γ	LC5002-005	hIL-13 R α 1	1.33×10^{-5}	3.6×10^{-4}	5.8×10^{-3}	0.06	2.7×10^{-9}
CM5	Anti-hFc γ	IgG1 ala-ala LC5002-005	hIL-13 R α 1	1.53×10^{-5}	4.2×10^{-4}	4.1×10^{-3}	0.04	2.8×10^{-9}

Example 14

Testing the crossreactivity of HuMab with hIL-13R α 2 and hIL-4R α by ELISA

Chimeric proteins hIL-13R α 2:hFc and hIL-4R α :hFc (R&D Systems, UK) were dissolved in PBS (1 μ g/ml) and allowed to adsorb to microtiter plates (NUNC Maxisorb) by incubation over night at 4°C. After washing the plates with washing

buffer (WB = 0.9 % NaCl; 0.1% Tween® 20) unspecific binding sites were blocked by addition of 100 µl incubation buffer (IB = PBS with 1% crotein C and 0.1% Tween® 20) and incubation for 30 min at room-temperature (RT). Then serially diluted HuMab and control antibodies (100 µl/well; dilutions in IB) were added and incubated for 1 hour at RT. The plates were again washed and bound antibody was detected by incubation with peroxidase conjugated rabbit anti-human kappa (DAKO, Denmark) in a final dilution of 1:500 in IB. After incubation for 1h at RT and a subsequent washing step, the plates were developed with ready-to-use ABTS® solution (Roche Diagnostics GmbH, DE) at RT in the dark. Absorbance was measured at 405 nm after absorbance of the highest concentration reached a sufficient OD.

All antibodies against IL-13Rα1 tested were able to bind to immobilized extracellular domains of human IL-13Rα1, but not neither to hIL-13Rα2 nor to hIL-4Rα (Fig. 4).

Example 15

Crossreactivity of HuMab with murine IL-13Rα1

Chimeric protein murine IL-13Rα1:hFc (R&D Systems, UK) was dissolved in PBS (1 µg/ml) and allowed to adsorb to microtiter plates (NUNC Maxisorb) by incubation over night at 4°C. After washing the plates with washing buffer (WB = 0.9 % NaCl; 0.1% Tween® 20), unspecific binding sites were blocked by addition of 100 µl incubation buffer (IB = PBS with 1% Crotein C and 0.1% Tween® 20) and incubation for 30 min at room-temperature (RT). Then, serially diluted HuMab and control antibodies (HuMab anti-KLH and polyclonal goat anti-hIL-13Rα1 (R&D Systems)) were added to the wells (100 µl /well; dilutions in IB) and incubated for 1 hour at RT. The plates were again washed and bound human antibodies were detected by incubation with peroxidase conjugated rabbit anti-human kappa (DAKO, Denmark) in a final dilution of 1:500 in IB. Goat anti-hIL-13Rα1 antibodies bound to the plates were detected by peroxidase-conjugated donkey anti-goat IgG (Santa Cruz; 1:1000 in IB). After incubation for 1h at RT and a subsequent washing step, the plates were developed with ready-to-use ABTS® solution (Roche Diagnostics GmbH, DE) at RT in the dark. Absorbance was measured at 405 nm after absorbance of the highest concentration reached a sufficient OD (Fig. 5).

Example 16

Crossreactivity of HuMab with Cynomolgus IL-13R α 1

5 The gene coding for IL-13R α 1 was isolated by RT-PCR from Cynomolgus tissue and transfected into in the murine cell line Ba/F3. In order to test whether the HuMabs crossreact with Cynomolgus IL-13R α 1, the stably transfected Ba/F3 cells as well as the parental Ba/F3 cells were incubated with 10 μ g/ml of HuMab and control antibodies. As positive control, a polyclonal goat-anti hIL-13R α 1(R&D Systems) was used. Negative controls included were: a human IgG1 myeloma protein (Nordic) and normal goat serum. Bound antibodies were detected by FACS
10 analysis using an antibody directed against human IgG conjugated with FITC to detect HuMabs and an antibody directed against goat IgG conjugated with FITC to detect goat antibodies. Mean fluorescence intensities (MFI) were compared for the individual antibodies tested on the transfected Ba/F3 line versus the parental line.

15 All HuMabs of the invention were able to bind to Cynomolgus IL-13R α 1 expressed in transfected Ba/F3 cells. As expected from the close homology between human and Cynomolgus IL-13R α 1, the polyclonal AF152 antibody also bound to the Cynomolgus IL-13R α 1. The negative control antibodies showed only a marginal increase in MFI when tested with the transfected Ba/F3 cell line (Table 5).

Table 5:

	Antibody	MFI Ba/F3	MFIBa/F3_Cyno_IL- 13Rα1	Fold Increase in MFI in the presence of Cyno_IL-13Rα1
HuMabs	LC5002-002	3.9	83.7	79.8
	LC5002-003	3.4	82.4	79.0
	LC5002-005	14.8	101.5	86.7
	LC5002-007	4.1	19	14.9
Controls	API52	3.2	21.2	18.0
	Normal goat IgG	3.3	5.7	2.4
	Normal human IgG1 (Nordic)	3.5	10.2	6.7
	Anti human IgG- FITC only	3.3	5.5	2.2

Example 17

Binding of IL-13Rα1 HuMabs to Fcγ receptors (binding to FcγRIIIa on NK cells)

- 5 To determine the ability of the antibodies of the invention to bind to FcγRIIIa (CD16) on Natural Killer (NK) cells, Peripheral Blood Mononuclear Cells (PBMCs) were isolated and incubated with 20 µg/ml of HuMab antibody and control antibodies in the presence or absence of 20 µg/ml of a blocking mouse antibody to FcγRIIIa (anti-CD16, clone 3G8, RDI, Flanders, NJ), to verify binding
- 10 via FcγRIIIa. As negative controls, human IgG2 and IgG4 (The Binding Site), that do not bind FcγRIIIa, were used. Human IgG1 and IgG3 (The Binding Site) were included as positive controls for FcγRIIIa binding. Bound antibodies on NK cells were detected by FACS analysis using a PE-labeled mouse anti-human CD56 (NK-cell surface marker) antibody (BD Biosciences Pharmingen, San Diego, CA) in
- 15 combination with a FITC-labeled goat F(ab)₂ anti-human IgG (Fc) antibody (Protos immunoresearch, Burlingame, CA). Maximum binding at 20 µg/ml (Bmax: MFI±st.dev) of the HuMab tested was determined.

LC5002-005 was able to bind to FcγRIIIa efficiently (comparable to the control IgG1 antibody) as indicated by a Bmax (MFI) value of 580.6 ± 245.8 . Addition of a blocking antibody against FcγRIIIa dramatically reduced binding of LC5002-005 to NK cells (Bmax (MFI) value of 260.4 ± 95.90) indicating specific binding to FcγRIIIa.

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Patent Claims

1. An antibody binding to IL-13R α 1 and inhibiting IL-13, characterized in that the variable heavy chain amino acid sequence CDR3 of said antibody is selected from the group consisting of the heavy chain CDR3 sequences of SEQ ID NO: 1, 3 or 7, whereby said antibody is obtained from hybridoma cell line DSM ACC2709, DSM ACC2710 or DSM ACC2712.
2. An antibody binding to IL-13R α 1 and inhibiting IL-13, characterized in that the variable heavy chain amino acid sequence CDR3 of said antibody is selected from the group consisting of the heavy chain CDR3 sequences of SEQ ID NO: 1, 3, 7 or 9 and comprises as heavy chain variable region SEQ ID NO: 1 and as light chain variable region SEQ ID NO: 2, as heavy chain variable region SEQ ID NO: 3 and as light chain variable region of SEQ ID NO: 4, as heavy chain variable region SEQ ID NO: 7 and as light chain variable region SEQ ID NO: 8 or as heavy variable region SEQ ID NO: 9 and as light chain variable region SEQ ID NO: 10.