

Antigen Binding Molecules comprising a TNF family ligand trimer

FIELD OF THE INVENTION

The invention relates to novel TNF family ligand trimer-containing antigen binding molecules comprising (a) at least one moiety capable of specific binding to a target cell antigen and (b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecules are characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or two fragments thereof that are connected to each other by a peptide linker and in that the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof. The invention further relates to methods of producing these molecules and to methods of using the same.

BACKGROUND

Ligands interacting with molecules of the TNF (tumor necrosis factor) receptor superfamily have pivotal roles in the organization and function of the immune system. While regulating normal functions such as immune responses, hematopoiesis and morphogenesis, the TNF family ligands (also called cytokines) play a role in tumorigenesis, transplant rejection, septic shock, viral replication, bone resorption, rheumatoid arthritis and diabetes (Aggarwal, 2003). The TNF ligand family comprises 18 genes encoding 19 type II (i.e. intracellular N terminus and extracellular C-terminus) transmembrane proteins, characterized by the presence of a conserved C-terminal domain coined the 'TNF homology domain' (THD). This domain is responsible for receptor binding and is thus critical for the biological activity of the TNF ligand family members. The sequence identity between family members is ~20–30% (Bodmer, 2002). Members of the TNF ligand family exert their biological function as self-assembling, noncovalent trimers (Banner et al, Cell 1993, 73, 431-445). Thus, the TNF family ligands form a trimer that is able to bind to and to activate the corresponding receptors of TNFR superfamily.

4-1BB (CD137), a member of the TNF receptor superfamily, has been first identified as a molecule whose expression is induced by T-cell activation (Kwon and Weissman, 1989). Subsequent studies demonstrated expression of 4-1BB in T- and B-lymphocytes (Snell et al., 2011; Zhang et al., 2010), NK-cells (Lin et al., 2008), NKT-cells (Kim et al., 2008), monocytes (Kienzle and von Kempis, 2000; Schwarz et al., 1995), neutrophils (Heinisch et al., 2000), mast (Nishimoto et al., 2005) and dendritic cells as well as cells of non-hematopoietic origin such as endothelial and smooth muscle cells (Broll et al., 2001; Olofsson et al., 2008). Expression of 4-

1BB in different cell types is mostly inducible and driven by various stimulatory signals, such as T-cell receptor (TCR) or B-cell receptor triggering, as well as signaling induced through co-stimulatory molecules or receptors of pro-inflammatory cytokines (Diehl et al., 2002; von Kempis et al., 1997; Zhang et al., 2010).

5 Expression of 4-1BB ligand (4-1BBL or CD137L) is more restricted and is observed on professional antigen presenting cells (APC) such as B-cells, dendritic cells (DCs) and macrophages. Inducible expression of 4-1BBL is characteristic for T-cells, including both $\alpha\beta$ and $\gamma\delta$ T-cell subsets, and endothelial cells (reviewed in Shao and Schwarz, 2011).

10 CD137 signaling is known to stimulate IFN γ secretion and proliferation of NK cells (Buechele et al., 2012; Lin et al., 2008; Melero et al., 1998) as well as to promote DC activation as indicated by their increased survival and capacity to secrete cytokines and upregulate co-stimulatory molecules (Choi et al., 2009; Futagawa et al., 2002; Wilcox et al., 2002). However, CD137 is best characterized as a co-stimulatory molecule which modulates TCR-induced activation in both the CD4+ and CD8+ subsets of T-cells. In combination with TCR triggering,
15 agonistic 4-1BB-specific antibodies enhance proliferation of T-cells, stimulate lymphokine secretion and decrease sensitivity of T-lymphocytes to activation-induced cells death (reviewed in (reviewed in Snell et al., 2011).

20 In line with these co-stimulatory effects of 4-1BB antibodies on T-cells in vitro, their administration to tumor bearing mice leads to potent anti-tumor effects in many experimental tumor models (Melero et al., 1997; Narazaki et al., 2010). However, 4-1BB usually exhibits its potency as an anti-tumor agent only when administered in combination with other immunomodulatory compounds (Curran et al., 2011; Guo et al., 2013; Morales-Kastresana et al., 2013; Teng et al., 2009; Wei et al., 2013), chemotherapeutic reagents (Ju et al., 2008; Kim et al., 2009), tumor-specific vaccination (Cuadros et al., 2005; Lee et al., 2011) or radiotherapy (Shi
25 and Siemann, 2006). In vivo depletion experiments demonstrated that CD8+ T-cells play the most critical role in anti-tumoral effect of 4-1BB-specific antibodies. However, depending on the tumor model or combination therapy, which includes anti-4-1BB, contributions of other types of cells such as DCs, NK-cells or CD4+ T-cells have been reported (Melero et al., 1997; Murillo et al., 2009; Narazaki et al., 2010; Stagg et al., 2011).

30 In addition to their direct effects on different lymphocyte subsets, 4-1BB agonists can also induce infiltration and retention of activated T-cells in the tumor through 4-1BB-mediated upregulation of intercellular adhesion molecule 1 (ICAM1) and vascular cell adhesion molecule 1 (VCAM1) on tumor vascular endothelium (Palazon et al., 2011).

4-1BB triggering may also reverse the state of T-cell anergy induced by exposure to soluble antigen that may contribute to disruption of immunological tolerance in the tumor micro-environment or during chronic infections (Wilcox et al., 2004).

It appears that the immunomodulatory properties of 4-1BB agonistic antibodies in vivo
5 require the presence of the wild type Fc-portion on the antibody molecule thereby implicating Fc-receptor binding as an important event required for the pharmacological activity of such reagents as has been described for agonistic antibodies specific to other apoptosis-inducing or immunomodulatory members of the TNFR-superfamily (Li and Ravetch, 2011; Teng et al., 2009). However, systemic administration of 4-1BB-specific agonistic antibodies with the
10 functionally active Fc domain also induces expansion of CD8+ T-cells associated with liver toxicity (Dubrot et al., 2010) that is diminished or significantly ameliorated in the absence of functional Fc-receptors in mice. In human clinical trials (ClinicalTrials.gov, NCT00309023), Fc-competent 4-1BB agonistic antibodies (BMS-663513) administered once every three weeks for 12 weeks induced stabilization of the disease in patients with melanoma, ovarian or renal cell
15 carcinoma. However, the same antibody given in another trial (NCT00612664) caused grade 4 hepatitis leading to termination of the trial (Simeone and Ascierto, 2012).

Collectively, the available pre-clinical and clinical data clearly demonstrate that there is a high clinical need for effective 4-1BB agonists. However, new generation drug candidates should not only effectively engage 4-1BB on the surface of hematopoietic and endothelial cells
20 but also be capable of achieving that through mechanisms other than binding to Fc-receptors in order to avoid uncontrollable side effects. The latter may be accomplished through preferential binding to and oligomerization on tumor-specific or tumor-associated moieties.

Fusion proteins composed of one extracellular domain of a 4-1BB ligand and a single chain antibody fragment (Mueller et al., 2008; Hornig et al., 2012) or a single 4-1BB ligand
25 fused to the C-terminus of a heavy chain (Zhang et al, 2007) have been made. WO 2010/010051 discloses the generation of fusion proteins that consist of three TNF ligand ectodomains linked to each other and fused to an antibody part.

However, there is still a need of new antigen binding molecules that combine a moiety capable of preferred binding to tumor-specific or tumor-associated targets with a moiety capable
30 of forming a costimulatory TNF ligand trimer and that have sufficient stability to be pharmaceutically useful. The antigen binding molecules of the present invention comprise both and surprisingly they provide a trimeric and thus biologically active TNF ligand, although one of the trimerizing TNF ligand ectodomains is located on another polypeptide than the other two TNF ligand ectodomains of the molecule.

SUMMARY OF THE INVENTION

In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising

(a) at least one moiety capable of specific binding to a target cell antigen and

- 5 (b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or two fragments thereof that are connected to each other by a peptide linker and in that the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof.

10 In a particular aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising

(a) at least one moiety capable of specific binding to a target cell antigen,

(b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises two
15 ectodomains of a TNF ligand family member or two fragments thereof that are connected to each other by a peptide linker and in that the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof, and

(c) an Fc domain composed of a first and a second subunit capable of stable association.

20 In a further aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, comprising

(a) at least one moiety capable of specific binding to a target cell antigen and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that

- 25 (i) the first polypeptide contains a CH1 or CL domain and the second polypeptide contains a CL or CH1 domain, respectively, wherein the second polypeptide is linked to the first polypeptide by a disulfide bond between the CH1 and CL domain, and wherein the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the CH1 or CL domain by a peptide linker and wherein the second polypeptide comprises one ectodomain of said TNF
30 ligand family member or a fragment thereof connected via a peptide linker to the CL or CH1 domain of said polypeptide, or

- (ii) the first polypeptide contains a CH3 domain and the second polypeptide contains a CH3 domain, respectively, and wherein the first polypeptide comprises two
35 ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the C-terminus of the CH3 domain by a peptide linker and wherein the second polypeptide comprises only one ectodomain of said TNF ligand family member

or a fragment thereof connected via a peptide linker to C-terminus of the CH3 domain of said polypeptide, or

(iii) the first polypeptide contains a VH-CL or a VL-CH1 domain and the second polypeptide contains a VL-CH1 domain or a VH-CL domain, respectively, wherein the second polypeptide is linked to the first polypeptide by a disulfide bond between the CH1 and CL domain, and wherein the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to to VH or VL by a peptide linker and wherein the second polypeptide comprises one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to VL or VH of said polypeptide.

In a particular aspect, the TNF ligand family member is one that costimulates human T-cell activation. Thus, the TNF family ligand trimer-containing antigen binding molecule comprises (a) at least one moiety capable of specific binding to a target cell antigen and (b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or two fragments thereof that are connected to each other by a peptide linker and in that the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof, wherein the TNF ligand family member costimulates human T-cell activation. More particularly, the TNF ligand family member is selected from 4-1BBL and OX40L.

In one aspect, the TNF ligand family member is 4-1BBL.

In a further aspect, the ectodomain of a TNF ligand family member comprises the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO: 2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:96, SEQ ID NO: 373, SEQ ID NO:374 and SEQ ID NO:375, particularly the amino acid sequence of SEQ ID NO:1 or SEQ ID NO:96.

In another aspect, the ectodomain of a TNF ligand family member or fragment thereof comprises the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3 , SEQ ID NO:4 and SEQ ID NO:96, particularly the amino acid sequence of SEQ ID NO:1 or SEQ ID NO:96. More particularly, the ectodomain of a TNF ligand family member comprises the amino acid sequence of SEQ ID NO:96.

In a further aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

- (a) at least one moiety capable of specific binding to a target cell antigen and
- (b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:97, SEQ ID NO:98 and SEQ ID NO:99 and in that the second polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:96, SEQ ID NO:3 and SEQ ID NO:4.

In one aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

- (a) at least one moiety capable of specific binding to a target cell antigen and
- (b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence of SEQ ID NO:5 and in that the second polypeptide comprises the amino acid sequence of SEQ ID NO:6.

In a further aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

- (a) at least one moiety capable of specific binding to a target cell antigen and
 - (b) a first and a second polypeptide that are linked to each other by a disulfide bond,
- wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence of SEQ ID NO:5 and in that the second polypeptide comprises the amino acid sequence of SEQ ID NO:183.

In yet a further aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

- (a) at least one moiety capable of specific binding to a target cell antigen and
 - (b) a first and a second polypeptide that are linked to each other by a disulfide bond,
- wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence of SEQ ID NO:97 and in that the second polypeptide comprises the amino acid sequence of SEQ ID NO:184 or SEQ ID NO:185.

In another aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

- (a) at least one moiety capable of specific binding to a target cell antigen,
 - (b) a first polypeptide containing a CH1 or CL domain and a second polypeptide containing a CL or CH1 domain, respectively, wherein the second polypeptide is linked to the first polypeptide by a disulfide bond between the CH1 and CL domain,
- and wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or two fragments thereof that are connected to each other and to the CH1 or CL domain by a peptide linker and in that the second polypeptide

comprises only one ectodomain of said TNF ligand family member or a fragment thereof connected by a peptide linker to the CL or CH1 domain of said polypeptide.

In one aspect, provided is a TNF family ligand trimer-containing antigen binding molecule comprising

- 5 (a) at least one moiety capable of specific binding to a target cell antigen,
(b) a first polypeptide containing a CH1 domain and a second polypeptide containing a CL domain, wherein the second polypeptide is linked to the first polypeptide by a disulfide bond between the CH1 and CL domain,
and wherein the antigen binding molecule is characterized in that the first polypeptide comprises
10 two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the CH1 domain by a peptide linker and in that the second polypeptide comprises one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to the CL domain of said polypeptide.

In another aspect, provided is a TNF family ligand trimer-containing antigen binding
15 molecule comprising

- (a) at least one moiety capable of specific binding to a target cell antigen,
(b) a first polypeptide containing a CL domain and a second polypeptide containing a CH1 domain, wherein the second polypeptide is linked to the first polypeptide by a disulfide bond between the CH1 and CL domain,
20 and wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the CL domain by a peptide linker and in that the second polypeptide comprises one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to the CH1 domain of said polypeptide.

25 In a further aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule as defined herein before, wherein the moiety capable of specific binding to a target cell antigen is selected from the group consisting of an antibody, an antibody fragment and a scaffold antigen binding protein.

In one aspect, the invention provides a TNF family ligand trimer-containing antigen
30 binding molecule as defined herein before, wherein the moiety capable of specific binding to a target cell antigen is an antibody fragment.

In particular, the moiety capable of specific binding to a target cell antigen is selected from the group consisting of an antibody fragment, a Fab molecule, a crossover Fab molecule, a single chain Fab molecule, a Fv molecule, a scFv molecule, a single domain antibody, an aVH and a
35 scaffold antigen binding protein.

In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule as defined herein before, wherein the moiety capable of specific binding to a target cell antigen is a scaffold antigen binding protein.

5 In a particular aspect, the invention is concerned with a TNF family ligand trimer-containing antigen binding molecule as defined above, wherein the moiety capable of specific binding to a target cell antigen is a Fab molecule capable of specific binding to a target cell antigen.

10 The invention provides a TNF family ligand trimer-containing antigen binding molecule that comprises at least one moiety capable of specific binding to a target cell antigen. In a particular aspect, the TNF family ligand trimer-containing antigen binding molecule comprises one moiety capable of specific binding to a target cell antigen. In another aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising two moieties capable of specific binding to a target cell antigen.

15 In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule of the invention, wherein the target cell antigen is selected from the group consisting of Fibroblast Activation Protein (FAP), Carcinoembryonic Antigen (CEA), Melanoma-associated Chondroitin Sulfate Proteoglycan (MCSP), Epidermal Growth Factor Receptor (EGFR), CD19, CD20 and CD33.

In a particular aspect, the target cell antigen is Fibroblast Activation Protein (FAP).

20 In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, wherein the moiety capable of specific binding to FAP comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:7 or SEQ ID NO:100, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:8 or SEQ ID NO:101, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:9 or SEQ ID NO:102, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:10 or SEQ ID NO:103, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:11 or SEQ ID NO:104, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:12 or SEQ ID NO:105.

30 In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, wherein the moiety capable of specific binding to FAP comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:7, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:8 and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:9, and a VL domain comprising (iv) CDR-L1 comprising

the amino acid sequence of SEQ ID NO:10, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:11 and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:12.

In a particular aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, wherein the moiety capable of specific binding to FAP comprises a
5 VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:100, (ii) CDR-H2 comprising the amino acid sequence SEQ ID NO:101, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:102, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:103, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:104, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID
10 NO:105.

In one aspect, provided is a TNF family ligand trimer-containing antigen binding molecule as defined herein before, wherein the moiety capable of specific binding to FAP comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:16 and a variable light chain comprising an amino acid sequence of SEQ ID NO:17 or wherein the moiety capable of
15 specific binding to FAP comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:106 and a variable light chain comprising an amino acid sequence of SEQ ID NO:107.

In a further aspect, provided is a TNF family ligand trimer-containing antigen binding molecule according to the invention, wherein a peptide comprising two ectodomains of a TNF
20 ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus to the CH1 or CL domain of a heavy chain by a second peptide linker and wherein one ectodomain of said TNF ligand family member or a fragment thereof is fused at the its C-terminus the CL or CH1 domain on a light chain by a third peptide linker.

In a particular aspect, the invention relates to a TNF family ligand trimer-containing
25 antigen binding molecule as defined above, wherein the peptide linker is (G4S)₂, i.e. a peptide linker of SEQ ID NO:13. In one aspect, the first peptide linker is (G4S)₂, the second peptide linker is GSPGSSSSGS (SEQ ID NO:57) and the third peptide linker is (G4S)₂. In another aspect, the first, the second and the third peptide linker is (G4S)₂.

The invention is further concerned with a TNF family ligand trimer-containing antigen
30 binding molecule as defined herein before, comprising an Fc domain composed of a first and a second subunit capable of stable association.

In particular, the TNF family ligand trimer-containing antigen binding molecule of the invention comprising (c) an Fc domain composed of a first and a second subunit capable of stable association further comprises (a) a Fab molecule capable of specific binding to a target

cell antigen, wherein the Fab heavy chain is fused at the C-terminus to the N-terminus of a CH2 domain in the Fc domain.

In a further aspect, the Fc domain is an IgG, particularly an IgG1 Fc domain or an IgG4 Fc domain. More particularly, the Fc domain is an IgG1 Fc domain. In a particular aspect, the Fc domain comprises a modification promoting the association of the first and second subunit of the Fc domain.

In another aspect, the invention is concerned with a TNF family ligand trimer-containing antigen binding molecule as defined herein before, comprising (c) an Fc domain composed of a first and a second subunit capable of stable association, wherein the Fc domain comprises one or more amino acid substitution that reduces binding to an Fc receptor, in particular towards Fc γ receptor.

In particular, the Fc domain comprises amino acid substitutions at positions 234 and 235 (EU numbering) and/or 329 (EU numbering) of the IgG heavy chains. More particularly, provided is a trimeric TNF family ligand-containing antigen binding molecule according to the invention which comprises an IgG1 Fc domain with the amino acid substitutions L234A, L235A and P329G (EU numbering).

In a further aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, wherein the antigen binding molecule comprises a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific binding to a target cell antigen, a first peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker fused at its C-terminus by a second peptide linker to a second heavy or light chain, and a second peptide comprising one ectodomain of said TNF ligand family member fused at its C-terminus by a third peptide linker to a second light or heavy chain, respectively.

In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the first peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by a second peptide linker to a CH1 domain that is part of a heavy chain, and the second peptide comprising one ectodomain of said TNF ligand family member or a fragment thereof is fused at its C-terminus by a third peptide linker to a CL domain that is part of a light chain.

In yet another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the first peptide comprising two ectodomains of a TNF ligand family member

or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by a second peptide linker to a CL domain that is part of a heavy chain, and the second peptide comprising one ectodomain of said TNF ligand family member or a fragment thereof is fused at its C-terminus by a third peptide linker to a CH1 domain that is part of a light chain.

In a further aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, wherein the first peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by a second peptide linker to a VH domain that is part of a heavy chain, and the second peptide comprising one ectodomain of said TNF ligand family member or a fragment thereof is fused at its C-terminus by a third peptide linker to a VL domain that is part of a light chain.

Provided is further a TNF family ligand trimer-containing antigen binding molecule, wherein in the CL domain adjacent to the TNF ligand family member the amino acid at position 123 (EU numbering) has been replaced by arginine (R) and the amino acid at position 124 (EU numbering) has been substituted by lysine (K), and wherein in the CH1 domain adjacent to the TNF ligand family member the amino acids at position 147 (EU numbering) and at position 213 (EU numbering) have been substituted by glutamic acid (E).

In a further aspect, provided is a TNF family ligand trimer-containing antigen binding molecule as described herein before, wherein the antigen binding molecule comprises (a) a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific binding to a target cell antigen, (b) a second heavy chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:97, SEQ ID NO:98 and SEQ ID NO:99, and a second light chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:96, SEQ ID NO:3 and SEQ ID NO:4.

In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, wherein the antigen binding molecule comprises (a) a Fab molecule capable of specific binding to FAP, and (b) a second heavy chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:97, SEQ ID NO:98 and SEQ ID NO:99, and a second light chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:96, SEQ ID NO:3 and SEQ ID NO:4.

In a particular aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising a moiety capable of specific binding to FAP. In one aspect,

provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the antigen binding molecule comprises

(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:16 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:17 or

a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:106 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:107,

(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:14, SEQ ID NO:108, SEQ ID NO:111, SEQ ID NO:113, SEQ ID NO:115, SEQ ID NO:139 and SEQ ID NO:148, and

(iii) a second light chain comprising the amino acid sequence of SEQ ID NO:15, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:112, SEQ ID NO:114 and SEQ ID NO:115.

In another aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, wherein the antigen binding molecule comprises

(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:16 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:17 or

a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:106 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:107,

(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:115, SEQ ID NO:117, SEQ ID NO:119 and SEQ ID NO:173, and

(iii) a second light chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:116, SEQ ID NO:118, SEQ ID NO:120 and SEQ ID NO:174.

In yet another aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, comprising

(a) at least one moiety capable of specific binding to a target cell antigen, and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide contains a CH3 domain and the second polypeptide contains a CH3 domain, respectively, and wherein the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the C-terminus of the CH3 domain by a peptide linker and wherein the second polypeptide comprises one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to C-terminus of the CH3 domain of said polypeptide.

In particular, such a TNF family ligand trimer-containing antigen binding molecule comprises two moieties capable of specific binding to a target cell antigen.

In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising two moieties capable of specific binding to FAP. In particular,
5 provided is a TNF family ligand trimer-containing antigen binding molecule as described herein before comprises

(i) a first heavy chain comprising the amino acid sequence of SEQ ID NO:121, a second heavy chain comprising the amino acid sequence of SEQ ID NO:122, and two light chains comprising the amino acid sequence of SEQ ID NO:19, or

10 (ii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:123, a second heavy chain comprising the amino acid sequence of SEQ ID NO:124, and two light chains comprising the amino acid sequence of SEQ ID NO:125, or

(iii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:126, a second heavy chain comprising the amino acid sequence of SEQ ID NO:127, and two light chains comprising
15 the amino acid sequence of SEQ ID NO:125.

In another particular aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule as described herein before, wherein the target cell antigen is CD19.

In one aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the moiety capable of specific binding to CD19 comprises a VH domain comprising (i)
20 CDR-H1 comprising the amino acid sequence of SEQ ID NO:195 or SEQ ID NO:252, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:196 or SEQ ID NO:253, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:197 or SEQ ID NO:254, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:198 or
SEQ ID NO:249, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:199 or SEQ
25 ID NO:250, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:200 or SEQ ID NO:251.

In a further aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the moiety capable of specific binding to CD19 comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:201 and a variable light chain
30 comprising an amino acid sequence of SEQ ID NO:202 or wherein the moiety capable of specific binding to FAP comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:357 and a variable light chain comprising an amino acid sequence of SEQ ID NO:358.

In a particular aspect, provided is a TNF family ligand trimer-containing antigen binding
35 molecule, wherein the antigen binding molecule comprises

(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:201 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:202 or

5 a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:357 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:358,

(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:14, SEQ ID NO:108, SEQ ID NO:111 and SEQ ID NO:113, and

10 (iii) a second light chain comprising the amino acid sequence of SEQ ID NO:15, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:112 and SEQ ID NO:114.

In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the antigen binding molecule comprises

15 (i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:201 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:202 or

a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:357 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:358,

20 (ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:115, SEQ ID NO:117, SEQ ID NO:119 and SEQ ID NO:173, and

(iii) a second light chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:116, SEQ ID NO:118, SEQ ID NO:120 and SEQ ID NO:174.

In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising two moieties capable of specific binding to CD19. In particular, 25 provided is a TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 29, 30 and 32 to 34, wherein the antigen binding molecule comprises

(i) a first heavy chain comprising the amino acid sequence of SEQ ID NO:209, a second heavy chain comprising the amino acid sequence of SEQ ID NO:210, and two light chains comprising the amino acid sequence of SEQ ID NO:206, or

30 (ii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:213, a second heavy chain comprising the amino acid sequence of SEQ ID NO:214, and two light chains comprising the amino acid sequence of SEQ ID NO:206, or

(iii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:309, a second heavy chain comprising the amino acid sequence of SEQ ID NO:310, and two light chains comprising 35 the amino acid sequence of SEQ ID NO:279, or

(iv) a first heavy chain comprising the amino acid sequence of SEQ ID NO:313, a second heavy

chain comprising the amino acid sequence of SEQ ID NO:314, and two light chains comprising the amino acid sequence of SEQ ID NO:279.

In another particular aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule as described herein before, wherein the target cell antigen is CEA.

5 In one aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the moiety capable of specific binding to CEA comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:321, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:322, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:323, and a VL domain comprising (iv) CDR-L1 comprising the amino acid
10 sequence of SEQ ID NO:324, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:325, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:326.

In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the moiety capable of specific binding to CEA comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:329 and a variable light chain
15 comprising an amino acid sequence of SEQ ID NO:330.

In one aspect, provided is a TNF family ligand trimer-containing antigen binding molecule as described herein before, wherein the antigen binding molecule comprises
(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:329 and a first light chain comprising the VL domain comprising the amino acid
20 sequence of SEQ ID NO:330,
(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:14, SEQ ID NO:108, SEQ ID NO:111 and SEQ ID NO:113, and
(iii) a second light chain comprising the amino acid sequence of SEQ ID NO:15, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:112 and SEQ ID NO:114.

25 In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the antigen binding molecule comprises
(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:329 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:330,
30 (ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:115, SEQ ID NO:117, SEQ ID NO:119 and SEQ ID NO:173, and
(iii) a second light chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:116, SEQ ID NO:118, SEQ ID NO:120 and SEQ ID NO:174.

In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising two moieties capable of specific binding to CEA. Particularly, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the antigen binding molecule comprises

- 5 (i) a first heavy chain comprising the amino acid sequence of SEQ ID NO:337, a second heavy chain comprising the amino acid sequence of SEQ ID NO:338, and two light chains comprising the amino acid sequence of SEQ ID NO:334, or
(ii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:341, a second heavy chain comprising the amino acid sequence of SEQ ID NO:342, and two light chains comprising
10 the amino acid sequence of SEQ ID NO:334.

In a further aspect, provided is a TNF family ligand trimer-containing antigen binding molecule as described herein before, wherein the TNF ligand family member is OX40L. In one aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the ectodomain of a TNF ligand family member comprises the amino acid sequence of SEQ ID
15 NO:53 or SEQ ID NO:54, particularly the amino acid sequence of SEQ ID NO:53.

In a further aspect, provided is a TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 5, 10 to 24, 29, 30, 32 to 34, 38 to 40, 44 and 45, comprising
(a) at least one moiety capable of specific binding to a target cell antigen and
(b) a first and a second polypeptide that are linked to each other by a disulfide bond,
20 wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence of SEQ ID NO:371 or SEQ ID:372 and in that the second polypeptide comprises the amino acid sequence of SEQ ID NO:53 or SEQ ID NO:54.

In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the target cell antigen is Fibroblast Activation Protein (FAP) and the moiety
25 capable of specific binding to FAP comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:7 or SEQ ID NO:100, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:8 or SEQ ID NO:101, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:9 or SEQ ID NO:102, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:10 or SEQ ID NO:103, (v) CDR-
30 L2 comprising the amino acid sequence of SEQ ID NO:11 or SEQ ID NO:104, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:12 or SEQ ID NO:105.

Particularly, provided is a TNF family ligand trimer-containing antigen binding molecule as described herein, wherein the antigen binding molecule comprises

- (i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ

ID NO:16 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:17 or

a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:106 and a first light chain comprising the VL domain comprising the amino acid sequence of
5 SEQ ID NO:107,

(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:355, and

(iii) a second light chain comprising the amino acid sequence of SEQ ID NO:356.

According to another aspect of the invention, there is provided an isolated polynucleotide
10 encoding a TNF family ligand trimer-containing antigen binding molecule as defined herein before. The invention further provides a vector, particularly an expression vector, comprising the isolated polynucleotide of the invention and a host cell comprising the isolated polynucleotide or the vector of the invention. In some embodiments the host cell is a eukaryotic cell, particularly a mammalian cell.

15 In another aspect, provided is a method for producing the TNF family ligand trimer-containing antigen binding molecule of the invention, comprising the steps of (i) culturing the host cell of the invention under conditions suitable for expression of the antigen binding molecule, and (ii) recovering the antigen binding molecule. The invention also encompasses a TNF family ligand trimer-containing antigen binding molecule produced by the method of the
20 invention.

The invention further provides a pharmaceutical composition comprising the TNF family ligand trimer-containing antigen binding molecule of the invention and at least one pharmaceutically acceptable excipient.

Also encompassed by the invention is the TNF family ligand trimer-containing antigen
25 binding molecule of the invention, or the pharmaceutical composition of the invention, for use as a medicament. In one aspect is provided the TNF family ligand trimer-containing antigen binding molecule of the invention, or the pharmaceutical composition of the invention, for use in the treatment of a disease in an individual in need thereof. In a specific embodiment, provided is the TNF family ligand trimer-containing antigen binding molecule of the invention, or the
30 pharmaceutical composition of the invention, for use in the treatment of cancer.

Also provided is the use of the TNF family ligand trimer-containing antigen binding molecule of the invention for the manufacture of a medicament for the treatment of a disease in an individual in need thereof, in particular for the manufacture of a medicament for the treatment of cancer, as well as a method of treating a disease in an individual, comprising administering to

said individual a therapeutically effective amount of a composition comprising the TNF family ligand trimer-containing antigen binding molecule of the invention in a pharmaceutically acceptable form. In a specific embodiment, the disease is cancer. In any of the above embodiments the individual is preferably a mammal, particularly a human.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the components for the assembly of split trimeric human 4-1BB ligands. Figure (1A) shows the dimeric ligand that is fused at the C-terminus to a human CH1 or CL domain or a VL or VH domain and Figure (1B) shows the monomeric ligand fused to human CL or CH1 domain or a VL or VH domain. Figure (1C) shows the dimeric ligand that is fused at the N-terminus to a human CH3 domain and Figure (1D) shows the monomeric ligand fused at the N-terminus to a human CH3 domain.

Figure 2 shows the 4-1BBL-trimer-containing antigen binding molecules Constructs 1.1 to 1.10 of the invention. The preparation and production of these constructs is described in Example 1. The VH and VL domains are those of anti-FAP antibody 28H1, the thick black point stands for the knob-into-hole modification. * symbolizes amino acid modifications in the CH1 and CL domain (so-called charged residues).

Figure 3 shows the components for the assembly of split trimeric murine 4-1BB ligands. Figure (3A) shows the dimeric ligand that is fused at the C-terminus to murine CL domain and Figure (3B) shows the monomeric ligand fused at the C-terminus to murine CH1 domain.

Components for the assembly of FAP targeted split trimeric murine 4-1BB ligand. Figure (3C) shows the assembled murine 4-1BBL-trimer-containing antigen binding molecules as described in more detail in Example 1.3.

Figure 4 shows the 4-1BBL-trimer-containing antigen binding molecules Constructs 2.1 to 2.6 of the invention. The preparation and production of these constructs is described in Example 2. The VH and VL domains are those of anti-FAP antibody 4B9, the thick black point stands for the knob-into-hole modification. * symbolizes amino acid modifications in the CH1 and CL domain (so-called charged residues).

Figure 5A and **Figure 5B** show the “untargeted” variants of Constructs 1.1 and 1.2 comprising a DP47 Fab molecule instead of the anti-FAP Fab molecule. The molecules are named Control A and Control B, respectively. The preparation is described in Example 1.4. **Figure 5C** is a drawing of the monomeric 4-1BB Fc(kih) construct as prepared in Example 3.

Figure 6 relates to the binding of FAP-targeted 4-1BB ligand trimer-containing Fc(kih) fusion antigen binding molecule (FAP split 4-1BBL trimer, filled circles) or DP-47 untargeted 4-

1BB ligand trimer-containing Fc(kih) fusion antigen binding molecule (DP47 split 4-1BBL trimer, open circles) to resting (naïve) or activated human PMBCs. Specifically, the binding to resting (naïve) or activated human CD8+ T cells is shown in Figure (6A), to resting (naïve) or activated human CD4+ T cells in Figure (6B) and to resting (naïve) or activated human NK cells in Figure (6C). Shown is the binding as Median of fluorescence intensity (MFI) of red macrophytic algae Phycoerythrin (R-PE)-labeled anti-human IgG Fcγ-specific goat IgG F(ab')₂ fragment which is used as secondary detection antibody. MFI was measured by flow cytometry and baseline corrected by subtracting the MFI of the blank control.

The binding of different FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs to human 4-1BB expressing T cells from PHA-L and Proleukin pre-activated and anti-human CD3/anti-human CD28 re-activated human PBMCs is shown in **Figure 7**. Binding was detected with R-Phycoerythrin-fluorochrome conjugated anti-human IgG Fcγ-specific goat IgG F(ab')₂ fragment. Shown is the median of fluorescence intensity (MFI) versus the concentration of tested Constructs 1.1 to 1.10 of Example 1. For a better display the binding curves are split in four different blots with Construct 1.1 (monovalent FAP-targeted split trimeric human 4-1BB ligand Fc (kih)) and Control B (monovalent untargeted split trimeric human 4-1BB ligand Fc (kih) with CH-CL cross and charged residues) as comparison curves. Binding was monitored on CD3+ CD8+ T cells (Figure 7A) and CD3+ CD4+ T cells (Figure 7B). The 4-1BB expression level on CD8 T cells is normally higher than on CD4 T cells. All versions bind with a quite similar affinity to human 4-1BB.

Figure 8 shows the binding of different FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs to CD4+ or CD8+ T cells from fresh PBMCs (Figure 8A) or to human 4-1BB expressing PHA-L and Proleukin pre-activated and anti-human CD3/anti-human CD28 re-activated human PBMCs (Figure 8B). Binding was detected with R-Phycoerythrin-fluorochrome conjugated anti-human IgG Fcγ-specific goat IgG F(ab')₂ fragment. Shown is the median of fluorescence intensity (MFI) versus the concentration of tested Constructs 2.1, 2.3, 2.4, 2.5 and 2.6 of Example 2 and control molecules Control B, Control C, Control E and Control F. For a better display the binding curves are split in two different blots with construct 2.1 (monovalent FAP-targeted split trimeric human 4-1BB ligand Fc (kih)) and control B (monovalent untargeted split trimeric human 4-1BB ligand Fc (kih) with CH-CL cross and charged residues) as comparison curves. Binding was monitored on CD45+ CD3+ CD8+ T cells (blots on the bottom) and CD45+ CD3+ CD4+ T cells (blots on the top). The 4-1BB expression level on CD8 T cells is normally higher than on CD4 T cells. All constructs bind with a quite similar affinity to human 4-1BB, whereas the bivalent construct 2.3 and its untargeted control C show a lower MFI. This can be due to sterical hindrance of 4-1BB-binding and/or less detection due to the 2nd detection antibody induced by the Fc-conjugated split 4-1BB ligand.

In **Figure 9** the binding of FAP-targeted 4-1BB ligand trimer-containing Fc(kih) fusion antigen binding molecules (FAP split 4-1BBL trimer, filled circles) or DP47 untargeted 4-1BB ligand trimer-containing Fc(kih) fusion antigen binding molecules (DP47 split 4-1BBL trimer; open circles) to activated mouse splenocytes is shown. In particular, the binding to activated mouse CD4+ T cells is shown in Figure (9A) and to activated mouse CD8+ T cells in Figure (9B). An anti-mouse CD137-specific human IgG1 P329G LALA antibody (clone Lob12.3) was used as positive control (Triangles). The binding is characterized by plotting the MFI of R-PE-labeled anti-human IgG Fc γ -specific goat IgG F(ab')₂ fragment that is used as secondary detection antibody versus the concentration in nM of the tested split 4-1BBL trimer constructs. MFI was measured by flow cytometry and baseline corrected by subtracting the MFI of the blank control.

Figure 10 shows the binding of 4-1BB ligand trimer-containing Fc(kih) fusion antigen binding molecules (filled circles: FAP-targeted 4-1BB ligand trimer-containing Fc(kih) fusion antigen binding molecule Construct 1.1, open circles: DP47 untargeted 4-1BB ligand trimer-containing Fc(kih) fusion antigen binding molecule Control A to fibroblast activation protein (FAP)-expressing human melanoma (A) MV-3 cell line and (B) WM-266-4 cell line. The binding is characterized by plotting the MFI of R-PE-labeled anti-human IgG Fc γ -specific goat IgG F(ab')₂ fragment that is used as secondary detection antibody versus the concentration in nM of tested split 4-1BBL trimer constructs. MFI was measured by flow cytometry and baseline corrected by subtracting the MFI of the blank control.

In **Figure 11** the binding of different FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs to human-FAP expressing human melanoma MV-3 cells (Figure 11A) and/or NIH/3T3-huFAP clone 39 transfected mouse embryonic fibroblast cells (Figure 11B) is shown. Binding was detected with R-Phycoerythrin-fluorochrome or fluorescein-fluorochrome conjugated anti-human IgG Fc γ -specific goat IgG F(ab')₂ fragments. Shown is the median of fluorescence intensity (MFI) versus the concentration of tested constructs. For a better display binding curves were distributed to four (Figure 11A) or two blots (Figure 11B), whereas construct 1.1 (monovalent FAP-targeted split trimeric human 4-1BB ligand Fc (kih)) is used a comparison curve. All constructs bind with a similar affinity to human FAP except the bivalent FAP-targeted constructs (constructs 1.5, 1.7 and 1.8). They showed a tendency to have lower EC50 values and lower median fluorescence intensity. This can be explained with their bivalent targeting (higher avidity, less molecules can bind at the same time due to occupancy of two epitopes resulting in a lower MFI). Structural differences may also explain the difference between Construct 1.8 (complete bivalent targeting) and Constructs 1.5 and 1.7 (only partial bivalent targeting).

In **Figure 12** the binding of different FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) Constructs 2.1, 2.3, 2.4, 2.5 and 2.6 to human-FAP expressing human melanoma MV-3 cells (Figure 12A) and WM-266-4 cells (Figure 12B) is shown. Binding was detected with R-Phycoerythrin-fluorochrome conjugated anti-human IgG Fc γ -specific goat IgG F(ab')₂ fragments. Shown is the median of fluorescence intensity (MFI) versus the concentration of tested constructs. For a better display binding curves were distributed to two blots, whereas construct 2.1 (monovalent FAP-targeted split trimeric human 4-1BB ligand Fc (kih)) is used as comparison curve. All constructs bind with a similar affinity to human FAP except the bivalent FAP-targeted construct 2.3. It has a tendency to show lower EC₅₀ values and lower median fluorescence intensity. This can be explained with its bivalent targeting, which results in higher avidity but less occupancy of FAP molecules on the cell surface resulting in a lower MFI.

Figure 13 shows the binding of different FAP-targeted or untargeted split trimeric mouse 4-1BB ligand Fc (kih) constructs to CD4⁺ or CD8⁺ T cells from fresh splenocytes (Figure 13A) or to mouse 4-1BB expressing anti-mouse CD3/anti-mouse CD28 monoclonal agonistic antibodies activated mouse splenocytes (Figure 13B). Binding was detected with FITC-fluorochrome conjugated anti-mouse IgG Fc γ -specific goat IgG F(ab')₂ fragment. Shown is the median of fluorescence intensity (MFI) versus the concentration of tested constructs. Binding was monitored on CD3⁺ CD8⁺ T cells (left blot) and CD3⁺ CD4⁺ T cells (right blot). The 4-1BB expression level on CD8 T cells is normally higher than on CD4 T cells. All constructs bind with a quite similar affinity to mouse 4-1BB.

The binding of different FAP-targeted or untargeted split trimeric mouse 4-1BB ligand Fc (kih) constructs to human FAP expressing tumor cells is demonstrated in **Figure 14**. Binding was detected with FITC-fluorochrome conjugated anti-mouse IgG Fc γ -specific goat IgG F(ab')₂ fragment. Shown is the median of fluorescence intensity (MFI) versus the concentration of tested constructs. Binding was monitored on MV-3 cells (Figure 14A) and WM-266-4 cells (Figure 14B). FAP-targeted split trimeric mouse 4-1BB ligand Fc (kih) constructs M.1 and M.2 bind with a quite similar affinity to FAP.

Figure 15 shows a scheme that illustrates the general principal of the NF κ B activity assay described in Example 6.1 using a reporter cell line. Shown is the activation assay set up with human 4-1BB expressing HeLa reporter cell line. A crosslinking of 4-1BB expressed on the reporter cells induces NF κ B activation and NF κ B-mediated Luciferase expression. After lysis of the cells Luciferase can catalyze the oxidation of Luciferin to Oxyluciferin. This chemical reaction correlates positively with the strength of NF κ B-mediated luciferase expression and can be measured by the strength of light emission (units of released light). The ratio of FAP-expressing tumor cells to the reporter cell line HeLa-huCD137-NF κ B-luc was 5 to 1.

In **Figure 16** it is shown that the activation of the NFkB signaling pathway by FAP-targeted 4-1BB ligand trimer-containing Fc(kih) fusion antigen binding molecule (Construct 1.1) is strictly dependent on its binding to FAP-expressing target cells. Human CD137 expressing NFkB reporter HeLa cells were co-cultured with the indicated tumor cells exhibiting different levels of cell surface FAP expression. Luciferase activity was assessed as described in Example 6.1 after culturing cells in the absence or presence of 4-1BBL-containing molecules at the indicated concentrations for 6 hours. Filled circles refer to Construct 1.1. Open circles refer to DP47 untargeted 4-1BB ligand trimer-containing Fc(kih) fusion antigen binding molecule (Control A). Cell line NIH/3T3-human FAP clone 39 was used as target cells in Graph (A), Graph (B) shows the activation with MV3 cell line as target cells and Graph (C) with WM-266-4 cell line as target cells. Activity is characterized by blotting the units of released light (URL) measured during 0.5 s versus the concentration in nM of tested split 4-1BBL trimer constructs. URLs are emitted due to luciferase-mediated oxidation of luciferin to oxyluciferin.

Figure 17 shows the NFkB-activation-induced Luciferase expression and activity as measured with the assay described in Example 6.1. Counts of released light per seconds (CPS) are measured for 0.5 s/well and plotted against the used concentration of FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs. Human 4-1BB-expressing HeLa-reporter cells were incubated for 6 h in the absence (**Figure 17A**) or presence of crosslinking human-FAP expressing human melanoma cell line MV-3 (**Figure 17B**) or WM-266-4 (**Figure 17C**). CPS were measured and blotted against the concentrations of different FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) Constructs. The cell ratio is one human 4-1BB-expressing HeLa reporter cell to five tumor cells. For better display, activation curves were split to four different display-blots with construct 1.1 (monovalent FAP-targeted split trimeric human 4-1BB ligand Fc (kih)) and control B (monovalent untargeted split trimeric human 4-1BB ligand Fc (kih) with CH-CL cross and charged residues) as comparison curves. **Figure 17A** shows the activation without crosslinking FAP-expressing tumor cells, **Figure 17B** shows the activation in the presence of crosslinking FAP-expressing MV-3 tumor cells and **Figure 17C** shows the activation in the presence of crosslinking FAP-expressing WM-266-4 tumor cells.

Figure 18 shows the NFkB-activation-induced Luciferase expression and activity as measured for the constructs of Example 2. Units of released light (URL) are measured for 0.5 s/well and plotted against the used concentration of FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs. Human 4-1BB-expressing HeLa-reporter cells were incubated for 6 h in the absence or presence of crosslinking human-FAP expressing human melanoma cell line MV-3 or WM-266-4. URLs were measured and blotted against the concentrations of different FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs 2.1, 2.3, 2.4, 2.5 and 2.6 and Controls B, C, E and F. The cell ratio is one 4-1BB-

expressing HeLa reporter cell to five tumor cells. For better display activation curves were split to two different display-blot with construct 2.1 (monovalent FAP-targeted split trimeric human 4-1BB ligand Fc (kih)).

In **Figure 19** the activation assay set up with cynomolgus monkey 4-1BB expressing T293-HEK reporter cell line is shown. A crosslinking of cynomolgus monkey 4-1BB expressed on the reporter cells induces NFκB activation and NFκB-mediated Luciferase expression. After lysis of the cells Luciferase can catalyze the oxidation of Luciferin to Oxyluciferin. This chemical reaction correlates positively with the strength of NFκB-mediated luciferase expression and can be measured by the strength of light emission (units of released light).

Figure 20 shows the NFκB-activation-induced Luciferase expression and activity. Units of released light (URL) are measured for 0.5 s/well and plotted against the used concentration of FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs. Cynomolgus monkey 4-1BB-expressing T293-HEK-reporter cells were incubated for 6 h in the absence or presence of crosslinking human-FAP expressing human melanoma cell line MV-3 or WM-266-4. URLs were measured and blotted against the concentrations of different FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs. The cell ratio is one 4-1BB-expressing T293-HEK reporter cell to five MV-3 or two WM-266-4 cells. For better display activation curves were split to two different blots with Construct 2.1 as comparison curve.

Figure 21: This scheme illustrates the principal of the T-cell activation assay described in Example 6.3. Shown is the schematic assay activation set up with HLA-A2-NLV-specific CD8 T cells and NLV-pulsed HLA-A2+ FAP+ human melanoma cell line MV-3 in the presence of different titrated concentration of FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs. Cells were incubated for 28 h, the last 4 h in the presence of monensin-containing Golgi-Stop. The ratio of NLV-specific CD8 T cells to MV-3 tumor cells is 1:8.

Figures 22A-E and 23A-E relate to the Activation assay with HLA-A2-NLV-specific CD8 T cells and NLV-pulsed HLA-A2+ FAP+ human melanoma cell line MV-3 in the presence of different titrated concentration of different FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs as prepared in Example 1. For better display expression curves were split to several different display-blot with Construct 1.1 (monovalent FAP-targeted split trimeric human 4-1BB ligand Fc (kih)) and Control B (monovalent untargeted split trimeric human 4-1BB ligand Fc (kih)) as comparison curves. Results were obtained in four independent similar experiments and show that prolonged IFNγ secretion and CD137 expression of NLV-specific CD8+ T cells is strictly dependent on simultaneous activation of T-cells via recognition of NLV-HLA-A2 complexes (signal 1) and 4-1BB-triggering by FAP-targeted human split 4-1BBL (signal 2). The effect of 4-1BB upregulation is shown in graphs of Figure 22, whereas the

effect of INF γ expression of CD8+ T cells is presented in graphs of Figure 23. Shown is always the frequency in percentage of positive cells in the total CD8+ T cell population. All FAP-targeted variants induced a similar activation improvement of NLV-peptide activated CD8 T cells shown in Figure 22 as 4-1BB-upregulation (positive feedback loop) and in Figure 23 as IFN γ expression after 24 h of stimulation. Differences of curves lie in the range of normal error deviation and are not significant.

Figures 24 and 25 refer to the Activation assay with HLA-A2-NLV-specific CD8 T cells and NLV-pulsed HLA-A2+ FAP+ human melanoma cell line MV-3 in the presence of titrated concentration of different FAP-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs of Example 2. For better display expression curves were split to two different display-blots with Construct 2.1 (monovalent FAP-targeted split trimeric human 4-1BB ligand Fc (kih)) and Control B as comparison curves. All FAP-targeted split trimeric human 4-1BB ligand Fc (kih) constructs show a similar activation improvement of HLA-A2-NLV-peptide specific CD8 T cells shown in Figure 24 as 4-1BB-upregulation (positive feedback loop) and in Figure 25 as IFN γ expression after 24 h of stimulation. Differences of curves lie in the range of normal error deviation and are not significant.

Figure 26: This scheme illustrates the experiment as described in Example 6.4.

Figure 27 shows the induction of CD8+ T cell proliferation. Shown is the frequency of proliferating CD8+ T cells versus the concentration of tested constructs.

Figure 28A relates to the single dose PK experiment of Construct 1.2 and Control B in healthy NOG mice. Shown is the decline in Construct concentration over the time. **Figure 28B** shows the results of the single dose PK experiment of Constructs 2.1, 2.3, Control B and Control C in tumor bearing NOG mice humanized with stem cells. Figure 28C relates to the single dose PK experiment comparing Construct 2.1 and 2.3 in healthy NOG mice.

Figure 29 shows components for the assembly of split trimeric human 4-1BB ligands. Figure (29A) shows the dimeric ligand that is fused at the C-terminus to a human CL domain with mutations E123R and Q124K (charged residues) and Figure (29B) shows the monomeric 4-1BB ligand fused to human CH1 domain with mutations K147E and K213E (charged residues). Components for the assembly of bivalent CD19-targeted split trimeric human 4-1BB ligand (71-254) antigen binding molecule (construct 3.3). Figure (29C) shows the dimeric ligand being fused to the C-terminus of human IgG1 Fc hole chain. Figure (29D) shows the monomeric ligand being fused to the C-terminus of human IgG1 Fc knob chain.

Figure 30 shows the CD19-targeted 4-1BBL-trimer-containing antigen binding molecules Constructs 3.1 to 3.6 of the invention. The preparation and production of these constructs is

described in Example 3. The VH and VL domains are those of anti-CD19 antibody 8B8-018, the thick black point stands for the knob-into-hole modification. * symbolizes amino acid modifications in the CH1 and CL domain (so-called charged residues).

In **Figure 31A** is illustrated the randomization strategy for the CDR regions of the parental clone 8B8. Shown are the variable domains of the parental clone 8B8 and the CDR regions (boxed) according to the numbering of Kabat. (X) represents the randomized positions. **Figure 31B** shows the schematic description of the library generation strategies. Shown is the PCR amplification and cloning strategy used for the generation of the 8B8-based library with A) randomized CDR1 and CDR2 regions in the light and heavy chain or B) randomized CDR1 and CDR3 regions in the light and CDR3 region in the heavy chain. Respective enzymes used for cloning into the phagemide are indicated.

Figure 32 shows the alignment of the parental anti-CD19 clone 8B8 with the selected affinity-matured binders. Shown are the sequences of clone 8B8 and all selected affinity-matured binders. CDRs of both heavy and light chains are framed.

Figure 33 relates to the SPR analysis of the parental 8B8 clone and its affinity-matured variants. Shown are the sensorgrams of clone 8B8 and its affinity-matured derivatives that are devoid of the LCDR1 N27d and N28 hotspots.

Figure 34 illustrates the setup of the assay measuring Simultaneous binding of CD19 targeted trimeric split 4-1BBL to hu4-1BB and huCD19 (Example 8.2).

The graphs in **Figure 35** show simultaneous binding of the CD19 targeted trimeric 4-1BBL FC fusion antigen binding molecules Constructs 3.1, 3.3, 3.4, 3.5, 3.6 and 4.4 (Analyte 1) to immobilized human 4-1BB and human CD19 (Analyte 2).

Figure 36 shows the binding of different CD19-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs to 4-1BB-expressing CD4 and CD8 T cells of PHA-L and Proleukin pre-activated and anti-human CD3/anti-human CD28 re-activated human PBMCs. Binding was detected with R-Phycoerythrin-fluorochrome conjugated anti-human IgG Fc γ -specific goat IgG F(ab')₂ fragment. Shown is the median of fluorescence intensity (MFI) versus the concentration of tested constructs. For a better display the binding curves are split in three different blots with construct 3.4 and control F (Isotype control huIgG1 P329G LALA) as comparison curves. Binding was monitored on CD45⁺ CD3⁺ CD8⁺ T cells (Figure 36A) and CD45⁺ CD3⁺ CD4⁺ T cells (Figure 36B). The 4-1BB expression level on CD8 T cells is normally higher than on CD4 T cells. All constructs bind with a quite similar affinity to human 4-1BB.

Figure 37 shows the binding of CD19-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) antigen binding molecules to human-CD19 expressing B cell lymphoma cell lines: diffuse large non-Hodgkin B cell lymphoma SU-DHL-8 (37A), acute B cell precursor lymphoid leukemia Nalm6 (37B), diffuse large cell lymphoblast lymphoma Toledo (37C) and diffuse large B cell lymphoma OCI-Ly18 (37D). Binding was detected with R-Phycoerythrin-fluorochrome conjugated anti-human IgG Fc γ -specific goat IgG F(ab')₂ fragments. Shown is the median of fluorescence intensity (MFI) versus the concentration of tested constructs. For a better display the binding curves are split in three different blots with construct 3.4 and control F (Isotype control huIgG1 P329G LALA) as comparison curves. All constructs bind with a quite similar affinity to human CD19.

Figure 38 relates to NF κ B-activation-induced Luciferase expression and activity of CD19-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) antigen binding molecules. Units of released light (URL) are measured for 0.5 s/well and plotted against the used concentration of CD19-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs 3.1 and 3.3 and control molecules B and C. Human 4-1BB-expressing HeLa-reporter cells were incubated for 7.5 h in the absence presence of crosslinking human-CD19 expressing SU-DHL-8 or Pfeiffer cells. URLs were measured and blotted against the concentrations of different CD19-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs. The cell ratio is one 4-1BB-expressing HeLa reporter cell to 2.5 or five tumor cells.

Figure 39 shows the binding of different humanized variants of T84.66 IgG on CEA-expressing human gastric adenocarcinoma cells. Based on the data humanized variant 1 was selected for including it into CEA-targeted trimeric human 4-1BB ligand Fc (kih) antigen binding molecules.

Figure 40 shows the CEA targeted 4-1BBL-trimer-containing antigen binding molecules Constructs 5.1 to 5.6 of the invention. The preparation and production of these constructs is described in Example 11. The VH and VL domains are those of anti-CEA antibody T84.66-LCHA, the thick black point stands for the knob-into-hole modification. * symbolizes amino acid modifications in the CH1 and CL domain (so-called charged residues).

Figure 41A shows a schematic description of human NA3B3A2-avi His, the antigen used to assess binding of CEA-targeted trimeric split 4-1BBL Fc (kih) antigen binding molecules. **Figure 41B** illustrates the setup of the assay measuring simultaneous binding of CEA-targeted trimeric split 4-1BBL to hu4-1BB and human NA3B3A2 (Example 12.1).

The graphs in **Figure 42** show simultaneous binding of the CEA targeted trimeric 4-1BBL Fc fusion antigen binding molecules Constructs 5.4, 5.6, 5.7 and 5.8 (Analyte 1) to immobilized human 4-1BB and human NA3B3A2 (Analyte 2).

Binding of different CEA-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs to 4-1BB-expressing CD4 and CD8 T cells of PHA-L and Proleukin pre-activated and anti-human CD3/anti-human CD28 re-activated human PBMCs is shown in **Figure 43**. Binding was detected with R-Phycoerythrin-fluorochrome conjugated anti-human IgG Fc γ -specific goat IgG F(ab')₂ fragment. Shown is the median of fluorescence intensity (MFI) versus the concentration of tested constructs. For a better display the binding curves are split in two different blots with construct 5.4 and control F (Isotype control huIgG1 P329G LALA) as comparison curves. Binding was monitored on CD45+ CD3+ CD8+ T cells (blots on the bottom) and CD45+ CD3+ CD4+ T cells (blots on the top). The 4-1BB expression level on CD8 T cells is normally higher than on CD4 T cells. All constructs bind with quite similar affinity to human 4-1BB.

Figure 44 shows the binding of CEA-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs to human-CEA expressing human gastric cell line MKN-45 (left) and human colorectal adenocarcinoma cells line LS180 (right). Binding was detected with R-Phycoerythrin-fluorochrome conjugated anti-human IgG Fc γ -specific goat IgG F(ab')₂ fragments. Shown is the median of fluorescence intensity (MFI) versus the concentration of tested constructs.

Figure 45 relates to NF κ B-activation-induced Luciferase expression and activity of CEA-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) antigen binding molecules. Units of released light (URL) are measured for 0.5 s/well and blotted against the used concentration of CEA-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs 5.4, 5.6, 5.7 and 5.8 and control molecules. Human 4-1BB-expressing HeLa-reporter cells were incubated for 6 h in the absence or presence of crosslinking human-CEA expressing human gastric cancer cell line MKN-45. The cell ratio is one 4-1BB-expressing HeLa reporter cell to three tumor cells.

In **Figures 46A and 46B** are shown the components for the assembly of monovalent FAP targeted split trimeric human OX40 ligand (construct 6.1). Figure 46A relates to dimeric ligand fused to human IgG1-CL domain, Figure 46B relates to monomeric ligand fused to human IgG1-CH1 domain. **Figure 46C** shows the FAP targeted OX40L-trimer-containing antigen binding molecule Construct 6.1. In **Figure 46D** is shown the DP47 "untargeted" human IgG1 PGLALA (control F).

Figure 47A shows the binding of FAP targeted split trimeric human Ox40L to FAP positive WM-266-4 cells. WM-266-4 cells express high levels of human fibroblast activation protein (huFAP). Only FAP targeted OX40 ligand Fc (kih) constructs (filled square) but not control 5 (filled diamond) bound to WM-266-4 cells. Shown is the binding as median of fluorescence intensity (MFI) of Fluorescein isothiocyanate (FITC)-labeled anti-human IgG Fc γ -specific goat

IgG F(ab')₂ fragment which is used as secondary detection antibody. MFI was measured by flow cytometry. The x-axis shows the concentration of antibody constructs. **Figure 47B** shows the binding of FAP targeted OX40 ligand Fc (kih) construct to human FAP human OX40 negative A549 NucLight Red cells. FAP targeted OX40 ligand Fc (kih) construct showed no binding to OX40 negative FAP negative A549 tumor cells. Shown is the binding as median of fluorescence intensity (MFI) of FITC labeled anti-human IgG Fcγ-specific goat IgG F(ab')₂ fragment which is used as secondary detection antibody. MFI was measured by flow cytometry and baseline corrected by subtracting the MFI of the blank control.

In **Figure 48A** the binding of FAP-OX40L to resting and activated human CD4 T cells is shown. OX40 is not expressed on resting human CD4 T cells (left side). In the absence of human OX40 expressing cells no binding was observed (left graphs). After activation of human PBMCs OX40 is up-regulated on CD4+ T cells (right side). FAP-OX40L bound to OX40+ activated CD4 T cells. Shown is the binding as median of fluorescence intensity (MFI) of FITC labeled anti-human IgG Fcγ-specific goat IgG F(ab')₂ fragment which is used as secondary detection antibody. MFI was measured by flow cytometry and baseline corrected by subtracting the MFI of the blank control. The x-axis shows the concentration of antibody constructs. **Figure 48B** shows that OX40 is not expressed on resting human CD8 T cells (left side). In the absence of human OX40 expressing cells no binding was observed (left graphs). After activation of human PBMCs OX40 is up-regulated on CD8+ T cells (right side). OX40 expression on human CD8+ T cells is lower than on CD4+ T cells and varies between donors and time points. Expression of OX40 was low on the depicted CD8 T cells. FAP-OX40L bound to OX40+ activated CD8 T cells. Shown is the binding as median of fluorescence intensity (MFI) of FITC labeled anti-human IgG Fcγ-specific goat IgG F(ab')₂ fragment which is used as secondary detection antibody. MFI was measured by flow cytometry and baseline corrected by subtracting the MFI of the blank control. The x-axis shows the concentration of antibody constructs

In **Figure 49** the activation of NFκB signaling pathway by the FAP targeted split trimeric human OX40L antigen binding molecule (FAP-OX40L) in HeLa_hOX40_NFκB_Luc1 reporter cells is demonstrated. Shown is the activation with (right graph) or without (left graph) crosslinking by secondary antibody. The reporter cells were cultured for 5 hours in the presence of FAP-OX40L at the indicated concentrations with or without crosslinking secondary polyclonal anti-huIgG1 Fcγ-specific goat IgG F(ab')₂ fragment in a 1:2 ratio. Luciferase activity was assessed as described in Example 6.1. Activity is characterized by blotting the units of released light (URL) measured during 0.5 s versus the concentration in nM of tested construct. URLs are emitted due to luciferase-mediated oxidation of luciferin to oxyluciferin.

The activation of NFκB by FAP-OX40L in HeLa_hOX40_NFκB_Luc1 reporter cells in the presence of FAP positive cells is shown in **Figure 50A**. Shown is the activation of NFκB

signaling pathway in the reporter cells by FAP-OX40L in the presence of low FAP expressing
expressing NIH-3T3 human FAP cells (ratio 3 FAP+ tumor cells to 1 reporter cell). The NFκB-
mediated luciferase activity was characterized by blotting the units of released light (URL),
measured during 0.5 s, versus the concentration in nM of tested compounds. URLs are emitted
5 due to luciferase-mediated oxidation of luciferin to oxyluciferin. Values are baseline corrected
by subtracting the URLs of the blank control. For a better comparison the area under the curve of
the respective blotted dose-response curves were quantified as a marker for the agonistic
capacity of each construct. The comparison is illustrated in **Figure 50B**. The area was calculated
using GraphPad Prism. Values are baseline corrected by subtracting the value of the blank
10 control.

Figure 51 shows the OX40 mediated costimulation of suboptimally TCR triggered resting
human PBMC (Example 15.5). Hyper-crosslinking of FAP-Ox40L by the present NIH/3T3-
huFAP clone 39 cells strongly promoted survival and proliferation in human CD4 and CD8 T
cells. Shown is the event count of vital CD4+ (left) and CD8+ (right) T cells. Baseline values of
15 samples containing only the anti-human CD3 (clone V9, huIgG1), resting human PBMC and
NIH/3T3-huFAP clone 39 were subtracted. Thus the enhancing effect of OX40 co-stimulation
but not the effect of suboptimal anti-CD3 stimulation per se is shown here. In the Figures on the
bottom the rescue of suboptimal TCR stimulation of resting human PBMC with cell surface
immobilized FAP-Ox40L – Proliferation is shown.

DETAILED DESCRIPTION OF THE INVENTION

Definitions

Unless defined otherwise, technical and scientific terms used herein have the same
meaning as generally used in the art to which this invention belongs. For purposes of interpreting
this specification, the following definitions will apply and whenever appropriate, terms used in
25 the singular will also include the plural and vice versa.

As used herein, the term "**antigen binding molecule**" refers in its broadest sense to a
molecule that specifically binds an antigenic determinant. Examples of antigen binding
molecules are antibodies, antibody fragments and scaffold antigen binding proteins.

As used herein, the term "**moiety capable of specific binding to a target cell antigen**"
30 refers to a polypeptide molecule that specifically binds to an antigenic determinant. In one
aspect, the antigen binding moiety is able to activate signaling through its target cell antigen. In a
particular aspect, the antigen binding moiety is able to direct the entity to which it is attached
(e.g. the TNF family ligand trimer) to a target site, for example to a specific type of tumor cell or

tumor stroma bearing the antigenic determinant. Moieties capable of specific binding to a target cell antigen include antibodies and fragments thereof as further defined herein. In addition, moieties capable of specific binding to a target cell antigen include scaffold antigen binding proteins as further defined herein, e.g. binding domains which are based on designed repeat proteins or designed repeat domains (see e.g. WO 2002/020565).

In relation to an antibody or fragment thereof, the term "moiety capable of specific binding to a target cell antigen" refers to the part of the molecule that comprises the area which specifically binds to and is complementary to part or all of an antigen. A moiety capable of specific antigen binding may be provided, for example, by one or more antibody variable domains (also called antibody variable regions). Particularly, a moiety capable of specific antigen binding comprises an antibody light chain variable region (VL) and an antibody heavy chain variable region (VH).

The term "**antibody**" herein is used in the broadest sense and encompasses various antibody structures, including but not limited to monoclonal antibodies, polyclonal antibodies, monospecific and multispecific antibodies (e.g., bispecific antibodies), and antibody fragments so long as they exhibit the desired antigen-binding activity.

The term "**monoclonal antibody**" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical and/or bind the same epitope, except for possible variant antibodies, e.g. containing naturally occurring mutations or arising during production of a monoclonal antibody preparation, such variants generally being present in minor amounts. In contrast to polyclonal antibody preparations, which typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody of a monoclonal antibody preparation is directed against a single determinant on an antigen.

The term "**monospecific**" antibody as used herein denotes an antibody that has one or more binding sites each of which bind to the same epitope of the same antigen. The term "**bispecific**" means that the antigen binding molecule is able to specifically bind to at least two distinct antigenic determinants. Typically, a bispecific antigen binding molecule comprises two antigen binding sites, each of which is specific for a different antigenic determinant. In certain embodiments the bispecific antigen binding molecule is capable of simultaneously binding two antigenic determinants, particularly two antigenic determinants expressed on two distinct cells.

The term "**valent**" as used within the current application denotes the presence of a specified number of binding sites in an antigen binding molecule. As such, the terms "bivalent", "tetravalent", and "hexavalent" denote the presence of two binding sites, four binding sites, and six binding sites, respectively, in an antigen binding molecule.

The terms “full length antibody”, “intact antibody”, and “whole antibody” are used herein interchangeably to refer to an antibody having a structure substantially similar to a native antibody structure. “**Native antibodies**” refer to naturally occurring immunoglobulin molecules with varying structures. For example, native IgG-class antibodies are heterotetrameric glycoproteins of about 150,000 daltons, composed of two light chains and two heavy chains that are disulfide-bonded. From N- to C-terminus, each heavy chain has a variable region (VH), also called a variable heavy domain or a heavy chain variable domain, followed by three constant domains (CH1, CH2, and CH3), also called a heavy chain constant region. Similarly, from N- to C-terminus, each light chain has a variable region (VL), also called a variable light domain or a light chain variable domain, followed by a light chain constant domain (CL), also called a light chain constant region. The heavy chain of an antibody may be assigned to one of five types, called α (IgA), δ (IgD), ϵ (IgE), γ (IgG), or μ (IgM), some of which may be further divided into subtypes, e.g. $\gamma 1$ (IgG1), $\gamma 2$ (IgG2), $\gamma 3$ (IgG3), $\gamma 4$ (IgG4), $\alpha 1$ (IgA1) and $\alpha 2$ (IgA2). The light chain of an antibody may be assigned to one of two types, called kappa (κ) and lambda (λ), based on the amino acid sequence of its constant domain.

An “**antibody fragment**” refers to a molecule other than an intact antibody that comprises a portion of an intact antibody that binds the antigen to which the intact antibody binds. Examples of antibody fragments include but are not limited to Fv, Fab, Fab', Fab'-SH, F(ab')₂; diabodies, triabodies, tetrabodies, cross-Fab fragments; linear antibodies; single-chain antibody molecules (e.g. scFv); and single domain antibodies. For a review of certain antibody fragments, see Hudson et al., Nat Med 9, 129-134 (2003). For a review of scFv fragments, see e.g. Plückthun, in The Pharmacology of Monoclonal Antibodies, vol. 113, Rosenberg and Moore eds., Springer-Verlag, New York, pp. 269-315 (1994); see also WO 93/16185; and U.S. Patent Nos. 5,571,894 and 5,587,458. For discussion of Fab and F(ab')₂ fragments comprising salvage receptor binding epitope residues and having increased in vivo half-life, see U.S. Patent No. 5,869,046. Diabodies are antibody fragments with two antigen-binding sites that may be bivalent or bispecific, see, for example, EP 404,097; WO 1993/01161; Hudson et al., Nat Med 9, 129-134 (2003); and Hollinger et al., Proc Natl Acad Sci USA 90, 6444-6448 (1993). Triabodies and tetrabodies are also described in Hudson et al., Nat Med 9, 129-134 (2003). Single-domain antibodies are antibody fragments comprising all or a portion of the heavy chain variable domain or all or a portion of the light chain variable domain of an antibody. In certain embodiments, a single-domain antibody is a human single-domain antibody (Domantis, Inc., Waltham, MA; see e.g. U.S. Patent No. 6,248,516 B1). Antibody fragments can be made by various techniques, including but not limited to proteolytic digestion of an intact antibody as well as production by recombinant host cells (e.g. E. coli or phage), as described herein.

Papain digestion of intact antibodies produces two identical antigen-binding fragments, called “Fab” fragments containing each the heavy- and light-chain variable domains and also the

constant domain of the light chain and the first constant domain (CH1) of the heavy chain. As used herein, Thus, the term “**Fab fragment**” refers to an antibody fragment comprising a light chain fragment comprising a VL domain and a constant domain of a light chain (CL), and a VH domain and a first constant domain (CH1) of a heavy chain. Fab’ fragments differ from Fab fragments by the addition of a few residues at the carboxy terminus of the heavy chain CH1 domain including one or more cysteins from the antibody hinge region. Fab’-SH are Fab’ fragments in which the cysteine residue(s) of the constant domains bear a free thiol group. Pepsin treatment yields an F(ab')₂ fragment that has two antigen-combining sites (two Fab fragments) and a part of the Fc region.

The term “**cross-Fab fragment**” or “xFab fragment” or “crossover Fab fragment” refers to a Fab fragment, wherein either the variable regions or the constant regions of the heavy and light chain are exchanged. Two different chain compositions of a crossover Fab molecule are possible and comprised in the bispecific antibodies of the invention: On the one hand, the variable regions of the Fab heavy and light chain are exchanged, i.e. the crossover Fab molecule comprises a peptide chain composed of the light chain variable region (VL) and the heavy chain constant region (CH1), and a peptide chain composed of the heavy chain variable region (VH) and the light chain constant region (CL). This crossover Fab molecule is also referred to as CrossFab_(VLVH). On the other hand, when the constant regions of the Fab heavy and light chain are exchanged, the crossover Fab molecule comprises a peptide chain composed of the heavy chain variable region (VH) and the light chain constant region (CL), and a peptide chain composed of the light chain variable region (VL) and the heavy chain constant region (CH1). This crossover Fab molecule is also referred to as CrossFab_(CLCH1).

A “single chain Fab fragment” or “**scFab**” is a polypeptide consisting of an antibody heavy chain variable domain (VH), an antibody constant domain 1 (CH1), an antibody light chain variable domain (VL), an antibody light chain constant domain (CL) and a linker, wherein said antibody domains and said linker have one of the following orders in N-terminal to C-terminal direction: a) VH-CH1-linker-VL-CL, b) VL-CL-linker-VH-CH1, c) VH-CL-linker-VL-CH1 or d) VL-CH1-linker-VH-CL; and wherein said linker is a polypeptide of at least 30 amino acids, preferably between 32 and 50 amino acids. Said single chain Fab fragments are stabilized via the natural disulfide bond between the CL domain and the CH1 domain. In addition, these single chain Fab molecules might be further stabilized by generation of interchain disulfide bonds via insertion of cysteine residues (e.g. position 44 in the variable heavy chain and position 100 in the variable light chain according to Kabat numbering).

A “crossover single chain Fab fragment” or “**x-scFab**” is a polypeptide consisting of an antibody heavy chain variable domain (VH), an antibody constant domain 1 (CH1), an antibody light chain variable domain (VL), an antibody light chain constant domain (CL) and a

linker, wherein said antibody domains and said linker have one of the following orders in N-terminal to C-terminal direction: a) VH-CL-linker-VL-CH1 and b) VL-CH1-linker-VH-CL; wherein VH and VL form together an antigen-binding site which binds specifically to an antigen and wherein said linker is a polypeptide of at least 30 amino acids. In addition, these x-scFab molecules might be further stabilized by generation of interchain disulfide bonds via insertion of cysteine residues (e.g. position 44 in the variable heavy chain and position 100 in the variable light chain according to Kabat numbering).

A “**single-chain variable fragment (scFv)**” is a fusion protein of the variable regions of the heavy (V_H) and light chains (V_L) of an antibody, connected with a short linker peptide of ten to about 25 amino acids. The linker is usually rich in glycine for flexibility, as well as serine or threonine for solubility, and can either connect the N-terminus of the V_H with the C-terminus of the V_L , or *vice versa*. This protein retains the specificity of the original antibody, despite removal of the constant regions and the introduction of the linker. scFv antibodies are, e.g. described in Houston, J.S., *Methods in Enzymol.* 203 (1991) 46-96). In addition, antibody fragments comprise single chain polypeptides having the characteristics of a V_H domain, namely being able to assemble together with a V_L domain, or of a V_L domain, namely being able to assemble together with a V_H domain to a functional antigen binding site and thereby providing the antigen binding property of full length antibodies.

“**Scaffold antigen binding proteins**” are known in the art, for example, fibronectin and designed ankyrin repeat proteins (DARPin) have been used as alternative scaffolds for antigen-binding domains, see, e.g., Gebauer and Skerra, *Engineered protein scaffolds as next-generation antibody therapeutics. Curr Opin Chem Biol* 13:245-255 (2009) and Stumpp et al., *Darpins: A new generation of protein therapeutics. Drug Discovery Today* 13: 695-701 (2008). In one aspect of the invention, a scaffold antigen binding protein is selected from the group consisting of CTLA-4 (Evibody), Lipocalins (Anticalin), a Protein A-derived molecule such as Z-domain of Protein A (Affibody), an A-domain (Avimer/Maxibody), a serum transferrin (*trans*-body); a designed ankyrin repeat protein (DARPin), a variable domain of antibody light chain or heavy chain (single-domain antibody, sdAb), a variable domain of antibody heavy chain (nanobody, aVH), V_{NAR} fragments, a fibronectin (AdNectin), a C-type lectin domain (Tetranectin); a variable domain of a new antigen receptor beta-lactamase (V_{NAR} fragments), a human gamma-crystallin or ubiquitin (Affilin molecules); a kunitz type domain of human protease inhibitors, microbodies such as the proteins from the knottin family, peptide aptamers and fibronectin (adnectin).

CTLA-4 (Cytotoxic T Lymphocyte-associated Antigen 4) is a CD28-family receptor expressed on mainly CD4+ T-cells. Its extracellular domain has a variable domain- like Ig fold. Loops corresponding to CDRs of antibodies can be substituted with heterologous sequence to

confer different binding properties. CTLA-4 molecules engineered to have different binding specificities are also known as Evibodies (e.g. US7166697B1). Evibodies are around the same size as the isolated variable region of an antibody (e.g. a domain antibody). For further details see Journal of Immunological Methods 248 (1-2), 31-45 (2001).

5 Lipocalins are a family of extracellular proteins which transport small hydrophobic molecules such as steroids, bilins, retinoids and lipids. They have a rigid beta-sheet secondary structure with a number of loops at the open end of the conical structure which can be engineered to bind to different target antigens. Anticalins are between 160-180 amino acids in size, and are derived from lipocalins. For further details see Biochim Biophys Acta 1482: 337-350 (2000),
10 US7250297B1 and US20070224633.

An affibody is a scaffold derived from Protein A of *Staphylococcus aureus* which can be engineered to bind to antigen. The domain consists of a three-helical bundle of approximately 58 amino acids. Libraries have been generated by randomization of surface residues. For further details see Protein Eng. Des. Sel. 17, 455-462 (2004) and EP 1641818A1.

15 Avimers are multidomain proteins derived from the A-domain scaffold family. The native domains of approximately 35 amino acids adopt a defined disulfide bonded structure. Diversity is generated by shuffling of the natural variation exhibited by the family of A-domains. For further details see Nature Biotechnology 23(12), 1556 - 1561 (2005) and Expert Opinion on Investigational Drugs 16(6), 909-917 (June 2007).

20 A transferrin is a monomeric serum transport glycoprotein. Transferrins can be engineered to bind different target antigens by insertion of peptide sequences in a permissive surface loop. Examples of engineered transferrin scaffolds include the Trans-body. For further details see J. Biol. Chem 274, 24066-24073 (1999).

25 Designed Ankyrin Repeat Proteins (DARPs) are derived from Ankyrin which is a family of proteins that mediate attachment of integral membrane proteins to the cytoskeleton. A single ankyrin repeat is a 33 residue motif consisting of two alpha-helices and a beta-turn. They can be engineered to bind different target antigens by randomizing residues in the first alpha-helix and a beta-turn of each repeat. Their binding interface can be increased by increasing the number of modules (a method of affinity maturation). For further details see J. Mol. Biol. 332, 489-503
30 (2003), PNAS 100(4), 1700-1705 (2003) and J. Mol. Biol. 369, 1015-1028 (2007) and US20040132028A1.

A single-domain antibody is an antibody fragment consisting of a single monomeric variable antibody domain. The first single domain were derived from the variable domain of the antibody heavy chain from camelids (nanobodies or V_HH fragments). Furthermore, the term

single-domain antibody includes an autonomous human heavy chain variable domain (aVH) or V_{NAR} fragments derived from sharks.

Fibronectin is a scaffold which can be engineered to bind to antigen. Adnectins consists of a backbone of the natural amino acid sequence of the 10th domain of the 15 repeating units of human fibronectin type III (FN3). Three loops at one end of the .beta.-sandwich can be engineered to enable an Adnectin to specifically recognize a therapeutic target of interest. For further details see Protein Eng. Des. Sel. 18, 435- 444 (2005), US20080139791, WO2005056764 and US6818418B1.

Peptide aptamers are combinatorial recognition molecules that consist of a constant scaffold protein, typically thioredoxin (TrxA) which contains a constrained variable peptide loop inserted at the active site. For further details see Expert Opin. Biol. Ther. 5, 783-797 (2005).

Microbodies are derived from naturally occurring microproteins of 25-50 amino acids in length which contain 3-4 cysteine bridges - examples of microproteins include KalataBI and conotoxin and knottins. The microproteins have a loop which can be engineered to include upto 25 amino acids without affecting the overall fold of the microprotein. For further details of engineered knottin domains, see WO2008098796.

An "**antigen binding molecule that binds to the same epitope**" as a reference molecule refers to an antigen binding molecule that blocks binding of the reference molecule to its antigen in a competition assay by 50% or more, and conversely, the reference molecule blocks binding of the antigen binding molecule to its antigen in a competition assay by 50% or more.

The term "**antigen binding domain**" refers to the part of an antigen binding molecule that comprises the area which specifically binds to and is complementary to part or all of an antigen. Where an antigen is large, an antigen binding molecule may only bind to a particular part of the antigen, which part is termed an epitope. An antigen binding domain may be provided by, for example, one or more variable domains (also called variable regions). Preferably, an antigen binding domain comprises an antibody light chain variable region (VL) and an antibody heavy chain variable region (VH).

As used herein, the term "**antigenic determinant**" is synonymous with "antigen" and "epitope," and refers to a site (e.g. a contiguous stretch of amino acids or a conformational configuration made up of different regions of non-contiguous amino acids) on a polypeptide macromolecule to which an antigen binding moiety binds, forming an antigen binding moiety-antigen complex. Useful antigenic determinants can be found, for example, on the surfaces of tumor cells, on the surfaces of virus-infected cells, on the surfaces of other diseased cells, on the surface of immune cells, free in blood serum, and/or in the extracellular matrix (ECM). The

proteins useful as antigens herein can be any native form the proteins from any vertebrate source, including mammals such as primates (e.g. humans) and rodents (e.g. mice and rats), unless otherwise indicated. In a particular embodiment the antigen is a human protein. Where reference is made to a specific protein herein, the term encompasses the “full-length”, unprocessed protein as well as any form of the protein that results from processing in the cell. The term also encompasses naturally occurring variants of the protein, e.g. splice variants or allelic variants.

By “**specific binding**” is meant that the binding is selective for the antigen and can be discriminated from unwanted or non-specific interactions. The ability of an antigen binding molecule to bind to a specific antigen can be measured either through an enzyme-linked immunosorbent assay (ELISA) or other techniques familiar to one of skill in the art, e.g. Surface Plasmon Resonance (SPR) technique (analyzed on a BIAcore instrument) (Liljeblad et al., Glyco J 17, 323-329 (2000)), and traditional binding assays (Heeley, Endocr Res 28, 217-229 (2002)). In one embodiment, the extent of binding of an antigen binding molecule to an unrelated protein is less than about 10% of the binding of the antigen binding molecule to the antigen as measured, e.g. by SPR. In certain embodiments, an molecule that binds to the antigen has a dissociation constant (K_d) of $\leq 1 \mu\text{M}$, $\leq 100 \text{ nM}$, $\leq 10 \text{ nM}$, $\leq 1 \text{ nM}$, $\leq 0.1 \text{ nM}$, $\leq 0.01 \text{ nM}$, or $\leq 0.001 \text{ nM}$ (e.g. 10^{-8} M or less, e.g. from 10^{-8} M to 10^{-13} M , e.g. from 10^{-9} M to 10^{-13} M).

“**Affinity**” or “binding affinity” refers to the strength of the sum total of non-covalent interactions between a single binding site of a molecule (e.g. an antibody) and its binding partner (e.g. an antigen). Unless indicated otherwise, as used herein, “binding affinity” refers to intrinsic binding affinity which reflects a 1:1 interaction between members of a binding pair (e.g. antibody and antigen). The affinity of a molecule X for its partner Y can generally be represented by the dissociation constant (K_d), which is the ratio of dissociation and association rate constants (k_{off} and k_{on} , respectively). Thus, equivalent affinities may comprise different rate constants, as long as the ratio of the rate constants remains the same. Affinity can be measured by common methods known in the art, including those described herein. A particular method for measuring affinity is Surface Plasmon Resonance (SPR).

A “**target cell antigen**” as used herein refers to an antigenic determinant presented on the surface of a target cell, for example a cell in a tumor such as a cancer cell or a cell of the tumor stroma. In certain embodiments, the target cell antigen is an antigen on the surface of a tumor cell. In one embodiment, target cell antigen is selected from the group consisting of Fibroblast Activation Protein (FAP), Carcinoembryonic Antigen (CEA), Melanoma-associated Chondroitin Sulfate Proteoglycan (MCSP), Epidermal Growth Factor Receptor (EGFR), CD19, CD20 and CD33. In particular, the target cell antigen is Fibroblast Activation Protein (FAP).

The term “**Fibroblast activation protein (FAP)**”, also known as Prolyl endopeptidase FAP or Seprase (EC 3.4.21), refers to any native FAP from any vertebrate source, including mammals such as primates (e.g. humans) non-human primates (e.g. cynomolgus monkeys) and rodents (e.g. mice and rats), unless otherwise indicated. The term encompasses “full-length,”
5 unprocessed FAP as well as any form of FAP which results from processing in the cell. The term also encompasses naturally occurring variants of FAP, e.g., splice variants or allelic variants. In one embodiment, the antigen binding molecule of the invention is capable of specific binding to human, mouse and/or cynomolgus FAP. The amino acid sequence of human FAP is shown in UniProt (www.uniprot.org) accession no. Q12884 (version 149, SEQ ID NO:20), or NCBI
10 (www.ncbi.nlm.nih.gov/) RefSeq NP_004451.2. The extracellular domain (ECD) of human FAP extends from amino acid position 26 to 760. The amino acid and nucleotide sequences of a His-tagged human FAP ECD is shown in SEQ ID NOs 15 and 16, respectively. The amino acid sequence of mouse FAP is shown in UniProt accession no. P97321 (version 126, SEQ ID NO:23), or NCBI RefSeq NP_032012.1. The extracellular domain (ECD) of mouse FAP extends
15 from amino acid position 26 to 761. SEQ ID NOs 24 and 25 show the amino acid and nucleotide sequences, respectively, of a His-tagged mouse FAP ECD. SEQ ID NOs 26 and 27 show the amino acid and nucleotide sequences, respectively, of a His-tagged cynomolgus FAP ECD. Preferably, an anti-FAP binding molecule of the invention binds to the extracellular domain of FAP. Exemplary anti-FAP binding molecules are described in International Patent Application
20 No. WO 2012/020006 A2.

The term “**Carcinoembryonic antigen (CEA)**”, also known as Carcinoembryonic antigen-related cell adhesion molecule 5 (CEACAM5), refers to any native CEA from any vertebrate source, including mammals such as primates (e.g. humans) non-human primates (e.g. cynomolgus monkeys) and rodents (e.g. mice and rats), unless otherwise indicated. The amino
25 acid sequence of human CEA is shown in UniProt accession no. P06731 (version 151, SEQ ID NO:28). CEA has long been identified as a tumor-associated antigen (Gold and Freedman, J Exp Med., 121:439-462, 1965; Berinstein N. L., J Clin Oncol., 20:2197-2207, 2002). Originally classified as a protein expressed only in fetal tissue, CEA has now been identified in several normal adult tissues. These tissues are primarily epithelial in origin, including cells of the
30 gastrointestinal, respiratory, and urogenital tracts, and cells of colon, cervix, sweat glands, and prostate (Nap et al., Tumour Biol., 9(2-3):145-53, 1988; Nap et al., Cancer Res., 52(8):2329-23339, 1992). Tumors of epithelial origin, as well as their metastases, contain CEA as a tumor associated antigen. While the presence of CEA itself does not indicate transformation to a cancerous cell, the distribution of CEA is indicative. In normal tissue, CEA is generally
35 expressed on the apical surface of the cell (Hammarström S., Semin Cancer Biol. 9(2):67-81 (1999)), making it inaccessible to antibody in the blood stream. In contrast to normal tissue, CEA tends to be expressed over the entire surface of cancerous cells (Hammarström S., Semin

Cancer Biol. 9(2):67-81 (1999)). This change of expression pattern makes CEA accessible to antibody binding in cancerous cells. In addition, CEA expression increases in cancerous cells. Furthermore, increased CEA expression promotes increased intercellular adhesions, which may lead to metastasis (Marshall J., Semin Oncol., 30(a Suppl. 8):30-6, 2003). The prevalence of

5 CEA expression in various tumor entities is generally very high. In concordance with published data, own analyses performed in tissue samples confirmed its high prevalence, with approximately 95% in colorectal carcinoma (CRC), 90% in pancreatic cancer, 80% in gastric cancer, 60% in non-small cell lung cancer (NSCLC, where it is co-expressed with HER3), and 40% in breast cancer; low expression was found in small cell lung cancer and glioblastoma.

10 CEA is readily cleaved from the cell surface and shed into the blood stream from tumors, either directly or via the lymphatics. Because of this property, the level of serum CEA has been used as a clinical marker for diagnosis of cancers and screening for recurrence of cancers, particularly colorectal cancer (Goldenberg D M., The International Journal of Biological Markers, 7:183-188, 1992; Chau I., et al., J Clin Oncol., 22:1420-1429, 2004; Flamini et al., Clin

15 Cancer Res; 12(23):6985-6988, 2006).

The term “**Melanoma-associated Chondroitin Sulfate Proteoglycan (MCSP)**”, also known as Chondroitin Sulfate Proteoglycan 4 (CSPG4) refers to any native MCSP from any vertebrate source, including mammals such as primates (e.g. humans) non-human primates (e.g. cynomolgus monkeys) and rodents (e.g. mice and rats), unless otherwise indicated. The amino

20 acid sequence of human MCSP is shown in UniProt accession no. Q6UVK1 (version 103, SEQ ID NO:29). The term “**Epidermal Growth Factor Receptor (EGFR)**”, also named Proto-oncogene c-ErbB-1 or Receptor tyrosine-protein kinase erbB-1, refers to any native EGFR from any vertebrate source, including mammals such as primates (e.g. humans) non-human primates (e.g. cynomolgus monkeys) and rodents (e.g. mice and rats), unless otherwise indicated. The

25 amino acid sequence of human EGFR is shown in UniProt accession no. P00533 (version 211, SEQ ID NO:30).

The term “**CD19**” refers to B-lymphocyte antigen CD19, also known as B-lymphocyte surface antigen B4 or T-cell surface antigen Leu-12 and includes any native CD19 from any vertebrate source, including mammals such as primates (e.g. humans) non-human primates (e.g.

30 cynomolgus monkeys) and rodents (e.g. mice and rats), unless otherwise indicated. The amino acid sequence of human CD19 is shown in Uniprot accession no. P15391 (version 160, SEQ ID NO:31). The term encompasses “full-length” unprocessed human CD19 as well as any form of human CD19 that results from processing in the cell as long as the antibody as reported herein binds thereto. CD19 is a structurally distinct cell surface receptor expressed on the surface of

35 human B cells, including, but not limited to, pre-B cells, B cells in early development {i.e., immature B cells), mature B cells through terminal differentiation into plasma cells, and

malignant B cells. CD19 is expressed by most pre-B acute lymphoblastic leukemias (ALL), non-Hodgkin's lymphomas, B cell chronic lymphocytic leukemias (CLL), pro-lymphocytic leukemias, hairy cell leukemias, common acute lymphocytic leukemias, and some Null-acute lymphoblastic leukemias. The expression of CD19 on plasma cells further suggests it may be expressed on differentiated B cell tumors such as multiple myeloma. Therefore, the CD19 antigen is a target for immunotherapy in the treatment of non-Hodgkin's lymphoma, chronic lymphocytic leukemia and/or acute lymphoblastic leukemia.

“**CD20**” refers to B-lymphocyte antigen CD20, also known as membrane-spanning 4-domains subfamily A member 1 (MS4A1), B-lymphocyte surface antigen B1 or Leukocyte surface antigen Leu-16, and includes any native CD20 from any vertebrate source, including mammals such as primates (e.g. humans) non-human primates (e.g. cynomolgus monkeys) and rodents (e.g. mice and rats), unless otherwise indicated. The amino acid sequence of human CD20 is shown in Uniprot accession no. P11836 (version 149, SEQ ID NO:32). “**CD33**” refers to Myeloid cell surface antigen CD33, also known as SIGLEC3 or gp67, and includes any native CD33 from any vertebrate source, including mammals such as primates (e.g. humans) non-human primates (e.g. cynomolgus monkeys) and rodents (e.g. mice and rats), unless otherwise indicated. The amino acid sequence of human CD33 is shown in Uniprot accession no. P20138 (version 157, SEQ ID NO:33).

The term “**variable region**” or “variable domain” refers to the domain of an antibody heavy or light chain that is involved in binding the antigen binding molecule to antigen. The variable domains of the heavy chain and light chain (VH and VL, respectively) of a native antibody generally have similar structures, with each domain comprising four conserved framework regions (FRs) and three hypervariable regions (HVRs). See, e.g., Kindt et al., Kuby Immunology, 6th ed., W.H. Freeman and Co., page 91 (2007). A single VH or VL domain may be sufficient to confer antigen-binding specificity.

The term “**hypervariable region**” or “HVR,” as used herein refers to each of the regions of an antibody variable domain which are hypervariable in sequence and/or form structurally defined loops (“hypervariable loops”). Generally, native four-chain antibodies comprise six HVRs; three in the VH (H1, H2, H3), and three in the VL (L1, L2, L3). HVRs generally comprise amino acid residues from the hypervariable loops and/or from the “complementarity determining regions” (CDRs), the latter being of highest sequence variability and/or involved in antigen recognition. Exemplary hypervariable loops occur at amino acid residues 26-32 (L1), 50-52 (L2), 91-96 (L3), 26-32 (H1), 53-55 (H2), and 96-101 (H3). (Chothia and Lesk, *J. Mol. Biol.* 196:901-917 (1987).) Exemplary CDRs (CDR-L1, CDR-L2, CDR-L3, CDR-H1, CDR-H2, and CDR-H3) occur at amino acid residues 24-34 of L1, 50-56 of L2, 89-97 of L3, 31-35B of H1, 50-65 of H2, and 95-102 of H3. (Kabat et al., *Sequences of Proteins of Immunological Interest*,

5th Ed. Public Health Service, National Institutes of Health, Bethesda, MD (1991).)

Hypervariable regions (HVRs) are also referred to as complementarity determining regions (CDRs), and these terms are used herein interchangeably in reference to portions of the variable region that form the antigen binding regions. This particular region has been described by Kabat et al., U.S. Dept. of Health and Human Services, "Sequences of Proteins of Immunological Interest" (1983) and by Chothia *et al.*, *J. Mol. Biol.* 196:901-917 (1987), where the definitions include overlapping or subsets of amino acid residues when compared against each other.

Nevertheless, application of either definition to refer to a CDR of an antibody or variants thereof is intended to be within the scope of the term as defined and used herein. The appropriate amino acid residues which encompass the CDRs as defined by each of the above cited references are set forth below in Table A as a comparison. The exact residue numbers which encompass a particular CDR will vary depending on the sequence and size of the CDR. Those skilled in the art can routinely determine which residues comprise a particular CDR given the variable region amino acid sequence of the antibody.

TABLE A. CDR Definitions¹

CDR	Kabat	Chothia	AbM ²
V _H CDR1	31-35	26-32	26-35
V _H CDR2	50-65	52-58	50-58
V _H CDR3	95-102	95-102	95-102
V _L CDR1	24-34	26-32	24-34
V _L CDR2	50-56	50-52	50-56
V _L CDR3	89-97	91-96	89-97

¹ Numbering of all CDR definitions in Table A is according to the numbering conventions set forth by Kabat et al. (see below).

² "AbM" with a lowercase "b" as used in Table A refers to the CDRs as defined by Oxford Molecular's "AbM" antibody modeling software.

Kabat *et al.* also defined a numbering system for variable region sequences that is applicable to any antibody. One of ordinary skill in the art can unambiguously assign this system of "Kabat numbering" to any variable region sequence, without reliance on any experimental data beyond the sequence itself. As used herein, "Kabat numbering" refers to the numbering system set forth by Kabat et al., U.S. Dept. of Health and Human Services, "Sequence of Proteins of Immunological Interest" (1983). Unless otherwise specified, references to the numbering of specific amino acid residue positions in an antibody variable region are according to the Kabat numbering system.

With the exception of CDR1 in VH, CDRs generally comprise the amino acid residues that form the hypervariable loops. CDRs also comprise “specificity determining residues,” or “SDRs,” which are residues that contact antigen. SDRs are contained within regions of the CDRs called abbreviated-CDRs, or a-CDRs. Exemplary a-CDRs (a-CDR-L1, a-CDR-L2, a-CDR-L3, a-CDR-H1, a-CDR-H2, and a-CDR-H3) occur at amino acid residues 31-34 of L1, 50-55 of L2, 89-96 of L3, 31-35B of H1, 50-58 of H2, and 95-102 of H3. (See Almagro and Fransson, *Front. Biosci.* 13:1619-1633 (2008).) Unless otherwise indicated, HVR residues and other residues in the variable domain (e.g., FR residues) are numbered herein according to Kabat et al., *supra*.

As used herein, the term “**affinity matured**” in the context of antigen binding molecules (e.g., antibodies) refers to an antigen binding molecule that is derived from a reference antigen binding molecule, e.g., by mutation, binds to the same antigen, preferably binds to the same epitope, as the reference antibody; and has a higher affinity for the antigen than that of the reference antigen binding molecule. Affinity maturation generally involves modification of one or more amino acid residues in one or more CDRs of the antigen binding molecule. Typically, the affinity matured antigen binding molecule binds to the same epitope as the initial reference antigen binding molecule.

“**Framework**” or “FR” refers to variable domain residues other than hypervariable region (HVR) residues. The FR of a variable domain generally consists of four FR domains: FR1, FR2, FR3, and FR4. Accordingly, the HVR and FR sequences generally appear in the following sequence in VH (or VL): FR1-H1(L1)-FR2-H2(L2)-FR3-H3(L3)-FR4.

An “**acceptor human framework**” for the purposes herein is a framework comprising the amino acid sequence of a light chain variable domain (VL) framework or a heavy chain variable domain (VH) framework derived from a human immunoglobulin framework or a human consensus framework, as defined below. An acceptor human framework “derived from” a human immunoglobulin framework or a human consensus framework may comprise the same amino acid sequence thereof, or it may contain amino acid sequence changes. In some embodiments, the number of amino acid changes are 10 or less, 9 or less, 8 or less, 7 or less, 6 or less, 5 or less, 4 or less, 3 or less, or 2 or less. In some embodiments, the VL acceptor human framework is identical in sequence to the VL human immunoglobulin framework sequence or human consensus framework sequence.

The term “**chimeric**” antibody refers to an antibody in which a portion of the heavy and/or light chain is derived from a particular source or species, while the remainder of the heavy and/or light chain is derived from a different source or species.

The “**class**” of an antibody refers to the type of constant domain or constant region possessed by its heavy chain. There are five major classes of antibodies: IgA, IgD, IgE, IgG, and

IgM, and several of these may be further divided into subclasses (isotypes), e.g. IgG₁, IgG₂, IgG₃, IgG₄, IgA₁, and IgA₂. The heavy chain constant domains that correspond to the different classes of immunoglobulins are called α , δ , ϵ , γ , and μ respectively..

A “**humanized**” antibody refers to a chimeric antibody comprising amino acid residues from non-human HVRs and amino acid residues from human FRs. In certain embodiments, a humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the HVRs (e.g., CDRs) correspond to those of a non-human antibody, and all or substantially all of the FRs correspond to those of a human antibody. A humanized antibody optionally may comprise at least a portion of an antibody constant region derived from a human antibody. A “**humanized form**” of an antibody, e.g., a non-human antibody, refers to an antibody that has undergone humanization. Other forms of “humanized antibodies” encompassed by the present invention are those in which the constant region has been additionally modified or changed from that of the original antibody to generate the properties according to the invention, especially in regard to C1q binding and/or Fc receptor (FcR) binding.

A “**human**” antibody is one which possesses an amino acid sequence which corresponds to that of an antibody produced by a human or a human cell or derived from a non-human source that utilizes human antibody repertoires or other human antibody-encoding sequences. This definition of a human antibody specifically excludes a humanized antibody comprising non-human antigen-binding residues.

The term “Fc domain” or “**Fc region**” herein is used to define a C-terminal region of an antibody heavy chain that contains at least a portion of the constant region. The term includes native sequence Fc regions and variant Fc regions. An IgG Fc region comprises an IgG CH2 and an IgG CH3 domain. The “CH2 domain” of a human IgG Fc region usually extends from an amino acid residue at about position 231 to an amino acid residue at about position 340. In one embodiment, a carbohydrate chain is attached to the CH2 domain. The CH2 domain herein may be a native sequence CH2 domain or variant CH2 domain. The “CH3 domain” comprises the stretch of residues C-terminal to a CH2 domain in an Fc region (i.e. from an amino acid residue at about position 341 to an amino acid residue at about position 447 of an IgG). The CH3 region herein may be a native sequence CH3 domain or a variant CH3 domain (e.g. a CH3 domain with an introduced “protuberance” (“knob”) in one chain thereof and a corresponding introduced “cavity” (“hole”) in the other chain thereof; see US Patent No. 5,821,333, expressly incorporated herein by reference). Such variant CH3 domains may be used to promote heterodimerization of two non-identical antibody heavy chains as herein described. In one embodiment, a human IgG heavy chain Fc region extends from Cys226, or from Pro230, to the carboxyl-terminus of the heavy chain. However, the C-terminal lysine (Lys447) of the Fc region may or may not be

present. Unless otherwise specified herein, numbering of amino acid residues in the Fc region or constant region is according to the EU numbering system, also called the EU index, as described in Kabat et al., Sequences of Proteins of Immunological Interest, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, MD, 1991.

5 The “**knob-into-hole**” technology is described e.g. in US 5,731,168; US 7,695,936; Ridgway et al., Prot Eng 9, 617-621 (1996) and Carter, J Immunol Meth 248, 7-15 (2001). Generally, the method involves introducing a protuberance (“knob”) at the interface of a first polypeptide and a corresponding cavity (“hole”) in the interface of a second polypeptide, such that the protuberance can be positioned in the cavity so as to promote heterodimer formation and
10 hinder homodimer formation. Protuberances are constructed by replacing small amino acid side chains from the interface of the first polypeptide with larger side chains (e.g. tyrosine or tryptophan). Compensatory cavities of identical or similar size to the protuberances are created in the interface of the second polypeptide by replacing large amino acid side chains with smaller ones (e.g. alanine or threonine). The protuberance and cavity can be made by altering the nucleic
15 acid encoding the polypeptides, e.g. by site-specific mutagenesis, or by peptide synthesis. In a specific embodiment a knob modification comprises the amino acid substitution T366W in one of the two subunits of the Fc domain, and the hole modification comprises the amino acid substitutions T366S, L368A and Y407V in the other one of the two subunits of the Fc domain. In a further specific embodiment, the subunit of the Fc domain comprising the knob modification
20 additionally comprises the amino acid substitution S354C, and the subunit of the Fc domain comprising the hole modification additionally comprises the amino acid substitution Y349C. Introduction of these two cysteine residues results in the formation of a disulfide bridge between the two subunits of the Fc region, thus further stabilizing the dimer (Carter, J Immunol Methods 248, 7-15 (2001)). The numbering is according to EU index of Kabat et al, Sequences of Proteins
25 of Immunological Interest, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, MD, 1991.

A "region equivalent to the Fc region of an immunoglobulin" is intended to include naturally occurring allelic variants of the Fc region of an immunoglobulin as well as variants having alterations which produce substitutions, additions, or deletions but which do not decrease
30 substantially the ability of the immunoglobulin to mediate effector functions (such as antibody-dependent cellular cytotoxicity). For example, one or more amino acids can be deleted from the N-terminus or C-terminus of the Fc region of an immunoglobulin without substantial loss of biological function. Such variants can be selected according to general rules known in the art so as to have minimal effect on activity (see, e.g., Bowie, J. U. et al., Science 247:1306-10 (1990)).

35 The term “**effector functions**” refers to those biological activities attributable to the Fc region of an antibody, which vary with the antibody isotype. Examples of antibody effector

functions include: C1q binding and complement dependent cytotoxicity (CDC), Fc receptor binding, antibody-dependent cell-mediated cytotoxicity (ADCC), antibody-dependent cellular phagocytosis (ADCP), cytokine secretion, immune complex-mediated antigen uptake by antigen presenting cells, down regulation of cell surface receptors (e.g. B cell receptor), and B cell activation.

An “**activating Fc receptor**” is an Fc receptor that following engagement by an Fc region of an antibody elicits signaling events that stimulate the receptor-bearing cell to perform effector functions. Activating Fc receptors include FcγRIIIa (CD16a), FcγRI (CD64), FcγRIIa (CD32), and FcαRI (CD89). A particular activating Fc receptor is human FcγRIIIa (see UniProt accession no. P08637, version 141).

The term “**TNF ligand family member**” or “TNF family ligand” refers to a proinflammatory cytokine. Cytokines in general, and in particular the members of the TNF ligand family, play a crucial role in the stimulation and coordination of the immune system. At present, nineteen cytokines have been identified as members of the TNF (tumour necrosis factor) ligand superfamily on the basis of sequence, functional, and structural similarities. All these ligands are type II transmembrane proteins with a C-terminal extracellular domain (ectodomain), N-terminal intracellular domain and a single transmembrane domain. The C-terminal extracellular domain, known as TNF homology domain (THD), has 20-30% amino acid identity between the superfamily members and is responsible for binding to the receptor. The TNF ectodomain is also responsible for the TNF ligands to form trimeric complexes that are recognized by their specific receptors.

Members of the TNF ligand family are selected from the group consisting of Lymphotoxin α (also known as LTA or TNFSF1), TNF (also known as TNFSF2), LTβ (also known as TNFSF3), OX40L (also known as TNFSF4), CD40L (also known as CD154 or TNFSF5), FasL (also known as CD95L, CD178 or TNFSF6), CD27L (also known as CD70 or TNFSF7), CD30L (also known as CD153 or TNFSF8), 4-1BBL (also known as TNFSF9), TRAIL (also known as APO2L, CD253 or TNFSF10), RANKL (also known as CD254 or TNFSF11), TWEAK (also known as TNFSF12), APRIL (also known as CD256 or TNFSF13), BAFF (also known as CD257 or TNFSF13B), LIGHT (also known as CD258 or TNFSF14), TL1A (also known as VEGI or TNFSF15), GITRL (also known as TNFSF18), EDA-A1 (also known as ectodysplasin A1) and EDA-A2 (also known as ectodysplasin A2). The term refers to any native TNF family ligand from any vertebrate source, including mammals such as primates (e.g. humans), non-human primates (e.g. cynomolgus monkeys) and rodents (e.g. mice and rats), unless otherwise indicated. In specific embodiments of the invention, the TNF ligand family member is selected from the group consisting of OX40L, FasL, CD27L, TRAIL, 4-1BBL, CD40L and GITRL. In a particular embodiment, the TNF ligand family member is selected from 4-1BBL and OX40L.

Further information, in particular sequences, of the TNF ligand family members may be obtained from publically accessible databases such as Uniprot (www.uniprot.org). For instance, the human TNF ligands have the following amino acid sequences: human Lymphotoxin α (UniProt accession no. P01374, SEQ ID NO:34), human TNF (UniProt accession no. P01375, SEQ ID NO:35), human Lymphotoxin β (UniProt accession no. Q06643, SEQ ID NO:36), human OX40L (UniProt accession no. P23510, SEQ ID NO:37), human CD40L (UniProt accession no. P29965, SEQ ID NO:38), human FasL (UniProt accession no. P48023, SEQ ID NO:39), human CD27L (UniProt accession no. P32970, SEQ ID NO:40), human CD30L (UniProt accession no. P32971, SEQ ID NO:41), 4-1BBL (UniProt accession no. P41273, SEQ ID NO:42), TRAIL (UniProt accession no. P50591, SEQ ID NO:43), RANKL (UniProt accession no. O14788, SEQ ID NO:44), TWEAK (UniProt accession no. O43508, SEQ ID NO:45), APRIL (UniProt accession no. O75888, SEQ ID NO:46), BAFF (UniProt accession no. Q9Y275, SEQ ID NO:47), LIGHT (UniProt accession no. O43557, SEQ ID NO:48), TL1A (UniProt accession no. O95150, SEQ ID NO:49), GITRL (UniProt accession no. Q9UNG2, SEQ ID NO:50) and ectodysplasin A (UniProt accession no. Q92838, SEQ ID NO:51).

An “**ectodomain**” is the domain of a membrane protein that extends into the extracellular space (i.e. the space outside the target cell). Ectodomains are usually the parts of proteins that initiate contact with surfaces, which leads to signal transduction. The ectodomain of TNF ligand family member as defined herein thus refers to the part of the TNF ligand protein that extends into the extracellular space (the extracellular domain), but also includes shorter parts or fragments thereof that are responsible for the trimerization and for the binding to the corresponding TNF receptor. The term “ectodomain of a TNF ligand family member or a fragment thereof” thus refers to the extracellular domain of the TNF ligand family member that forms the extracellular domain or to parts thereof that are still able to bind to the receptor (receptor binding domain).

The term “**costimulatory TNF ligand family member**” or “costimulatory TNF family ligand” refers to a subgroup of TNF ligand family members, which are able to costimulate proliferation and cytokine production of T-cells. These TNF family ligands can costimulate TCR signals upon interaction with their corresponding TNF receptors and the interaction with their receptors leads to recruitment of TNFR-associated factors (TRAF), which initiate signalling cascades that result in T-cell activation. Costimulatory TNF family ligands are selected from the group consisting of 4-1BBL, OX40L, GITRL, CD70, CD30L and LIGHT, more particularly the costimulatory TNF ligand family member is selected from 4-1BBL and OX40L.

As described herein before, 4-1BBL is a type II transmembrane protein and one member of the TNF ligand family. Complete or full length 4-1BBL having the amino acid sequence of SEQ ID NO:42 has been described to form trimers on the surface of cells. The formation of trimers is

enabled by specific motives of the ectodomain of 4-1BBL. Said motives are designated herein as “trimerization region”. The amino acids 50-254 of the human 4-1BBL sequence (SEQ ID NO:52) form the extracellular domain of 4-1BBL, but even fragments thereof are able to form the trimers. In specific embodiments of the invention, the term “ectodomain of 4-1BBL or a fragment thereof” refers to a polypeptide having an amino acid sequence selected from SEQ ID NO:4 (amino acids 52-254 of human 4-1BBL), SEQ ID NO:1 (amino acids 71-254 of human 4-1BBL), SEQ ID NO:3 (amino acids 80-254 of human 4-1BBL) and SEQ ID NO:2 (amino acids 85-254 of human 4-1BBL) or a polypeptide having an amino acid sequence selected from SEQ ID NO:96 (amino acids 71-248 of human 4-1BBL), SEQ ID NO:375 (amino acids 52-248 of human 4-1BBL), SEQ ID NO:374 (amino acids 80-248 of human 4-1BBL) and SEQ ID NO:373 (amino acids 85-248 of human 4-1BBL), but also other fragments of the ectodomain capable of trimerization are included herein.

As described herein before, OX40L is another type II transmembrane protein and a further member of the TNF ligand family. Complete or full length human OX40L has the amino acid sequence of SEQ ID NO:37. The amino acids 51-183 of the human OX40L sequence (SEQ ID NO:53) form the extracellular domain of OX40L, but even fragments thereof that are able to form the trimers. In specific embodiments of the invention, the term “ectodomain of OX40L or a fragment thereof” refers to a polypeptide having an amino acid sequence selected from SEQ ID NO:53 (amino acids 51-183 of human OX40L) or SEQ ID NO:54 (amino acids 52-183 of human OX40L), but also other fragments of the ectodomain capable of trimerization are included herein.

The term “**peptide linker**” refers to a peptide comprising one or more amino acids, typically about 2 to 20 amino acids. Peptide linkers are known in the art or are described herein. Suitable, non-immunogenic linker peptides are, for example, $(G_4S)_n$, $(SG_4)_n$ or $G_4(SG_4)_n$ peptide linkers, wherein “n” is generally a number between 1 and 10, typically between 1 and 4, in particular 2, i.e. the peptides selected from the group consisting of GGGGS (SEQ ID NO:128), GGGGSGGGGS (SEQ ID NO:13), SGGGSGGGG (SEQ ID NO:55) and GGGGSGGGGSGGGG (SEQ ID NO:56), but also include the sequences GSPGSSSSGS (SEQ ID NO:57), GSGSGSGS (SEQ ID NO:58), GSGSGNGS (SEQ ID NO:59), GGSGSGSG (SEQ ID NO:60), GGSGSG (SEQ ID NO:61), GGSG (SEQ ID NO:62), GGSGNGSG (SEQ ID NO:63), GGNGSGSG (SEQ ID NO:64) and GGNGSG (SEQ ID NO:65). Peptide linkers of particular interest are $(G_4S)_1$ or GGGGS (SEQ ID NO:128), $(G_4S)_2$ or GGGGSGGGGS (SEQ ID NO:13) and GSPGSSSSGS (SEQ ID NO:57), more particularly $(G_4S)_2$ or GGGGSGGGGS (SEQ ID NO:13) and GSPGSSSSGS (SEQ ID NO:57).

The term “**amino acid**” as used within this application denotes the group of naturally occurring carboxy α -amino acids comprising alanine (three letter code: ala, one letter code: A), arginine (arg, R), asparagine (asn, N), aspartic acid (asp, D), cysteine (cys, C), glutamine (gln,

Q), glutamic acid (glu, E), glycine (gly, G), histidine (his, H), isoleucine (ile, I), leucine (leu, L), lysine (lys, K), methionine (met, M), phenylalanine (phe, F), proline (pro, P), serine (ser, S), threonine (thr, T), tryptophan (trp, W), tyrosine (tyr, Y), and valine (val, V).

A **“single chain fusion protein”** as used herein refers to a single chain polypeptide
5 composed of one or two ectodomains of said TNF ligand family member fused to a part of antigen binding moiety or Fc part. The fusion may occur by directly linking the N or C-terminal amino acid of the antigen binding moiety via a peptide linker to the C- or N-terminal amino acid of the ectodomain of said TNF ligand family member.

By “fused” or “connected” is meant that the components (e.g. a polypeptide and an
10 ectodomain of said TNF ligand family member) are linked by peptide bonds, either directly or via one or more peptide linkers.

“Percent (%) amino acid sequence identity” with respect to a reference polypeptide (protein) sequence is defined as the percentage of amino acid residues in a candidate sequence that are identical with the amino acid residues in the reference polypeptide sequence, after
15 aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not considering any conservative substitutions as part of the sequence identity. Alignment for purposes of determining percent amino acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST, BLAST-2, ALIGN, SAWI or Megalign (DNASTAR)
20 software. Those skilled in the art can determine appropriate parameters for aligning sequences, including any algorithms needed to achieve maximal alignment over the full length of the sequences being compared. For purposes herein, however, % amino acid sequence identity values are generated using the sequence comparison computer program ALIGN-2. The ALIGN-2 sequence comparison computer program was authored by Genentech, Inc., and the source code
25 has been filed with user documentation in the U.S. Copyright Office, Washington D.C., 20559, where it is registered under U.S. Copyright Registration No. TXU510087. The ALIGN-2 program is publicly available from Genentech, Inc., South San Francisco, California, or may be compiled from the source code. The ALIGN-2 program should be compiled for use on a UNIX operating system, including digital UNIX V4.0D. All sequence comparison parameters are set by
30 the ALIGN-2 program and do not vary. In situations where ALIGN-2 is employed for amino acid sequence comparisons, the % amino acid sequence identity of a given amino acid sequence A to, with, or against a given amino acid sequence B (which can alternatively be phrased as a given amino acid sequence A that has or comprises a certain % amino acid sequence identity to, with, or against a given amino acid sequence B) is calculated as follows:

100 times the fraction X/Y

where X is the number of amino acid residues scored as identical matches by the sequence alignment program ALIGN-2 in that program's alignment of A and B, and where Y is the total number of amino acid residues in B. It will be appreciated that where the length of amino acid sequence A is not equal to the length of amino acid sequence B, the % amino acid sequence identity of A to B will not equal the % amino acid sequence identity of B to A. Unless specifically stated otherwise, all % amino acid sequence identity values used herein are obtained as described in the immediately preceding paragraph using the ALIGN-2 computer program.

In certain embodiments, **amino acid sequence variants** of the TNF ligand trimer-containing antigen binding molecules provided herein are contemplated. For example, it may be desirable to improve the binding affinity and/or other biological properties of the TNF ligand trimer-containing antigen binding molecules. Amino acid sequence variants of the TNF ligand trimer-containing antigen binding molecules may be prepared by introducing appropriate modifications into the nucleotide sequence encoding the molecules, or by peptide synthesis. Such modifications include, for example, deletions from, and/or insertions into and/or substitutions of residues within the amino acid sequences of the antibody. Any combination of deletion, insertion, and substitution can be made to arrive at the final construct, provided that the final construct possesses the desired characteristics, e.g., antigen-binding. Sites of interest for substitutional mutagenesis include the HVRs and Framework (FRs). Conservative substitutions are provided in Table B under the heading "Preferred Substitutions" and further described below in reference to amino acid side chain classes (1) to (6). Amino acid substitutions may be introduced into the molecule of interest and the products screened for a desired activity, e.g., retained/improved antigen binding, decreased immunogenicity, or improved ADCC or CDC.

TABLE B

Original Residue	Exemplary Substitutions	Preferred Substitutions
Ala (A)	Val; Leu; Ile	Val
Arg (R)	Lys; Gln; Asn	Lys
Asn (N)	Gln; His; Asp, Lys; Arg	Gln
Asp (D)	Glu; Asn	Glu
Cys (C)	Ser; Ala	Ser
Gln (Q)	Asn; Glu	Asn
Glu (E)	Asp; Gln	Asp
Gly (G)	Ala	Ala
His (H)	Asn; Gln; Lys; Arg	Arg
Ile (I)	Leu; Val; Met; Ala; Phe; Norleucine	Leu

Original Residue	Exemplary Substitutions	Preferred Substitutions
Leu (L)	Norleucine; Ile; Val; Met; Ala; Phe	Ile
Lys (K)	Arg; Gln; Asn	Arg
Met (M)	Leu; Phe; Ile	Leu
Phe (F)	Trp; Leu; Val; Ile; Ala; Tyr	Tyr
Pro (P)	Ala	Ala
Ser (S)	Thr	Thr
Thr (T)	Val; Ser	Ser
Trp (W)	Tyr; Phe	Tyr
Tyr (Y)	Trp; Phe; Thr; Ser	Phe
Val (V)	Ile; Leu; Met; Phe; Ala; Norleucine	Leu

Amino acids may be grouped according to common side-chain properties:

- (1) hydrophobic: Norleucine, Met, Ala, Val, Leu, Ile;
- (2) neutral hydrophilic: Cys, Ser, Thr, Asn, Gln;
- (3) acidic: Asp, Glu;
- (4) basic: His, Lys, Arg;
- (5) residues that influence chain orientation: Gly, Pro;
- (6) aromatic: Trp, Tyr, Phe.

Non-conservative substitutions will entail exchanging a member of one of these classes for another class.

The term “**amino acid sequence variants**” includes substantial variants wherein there are amino acid substitutions in one or more hypervariable region residues of a parent antigen binding molecule (*e.g.* a humanized or human antibody). Generally, the resulting variant(s) selected for further study will have modifications (*e.g.*, improvements) in certain biological properties (*e.g.*, increased affinity, reduced immunogenicity) relative to the parent antigen binding molecule and/or will have substantially retained certain biological properties of the parent antigen binding molecule. An exemplary substitutional variant is an affinity matured antibody, which may be conveniently generated, *e.g.*, using phage display-based affinity maturation techniques such as those described herein. Briefly, one or more HVR residues are mutated and the variant antigen binding molecules displayed on phage and screened for a particular biological activity (*e.g.* binding affinity). In certain embodiments, substitutions, insertions, or deletions may occur within one or more HVRs so long as such alterations do not substantially reduce the ability of the antigen binding molecule to bind antigen. For example, conservative alterations (*e.g.*,

conservative substitutions as provided herein) that do not substantially reduce binding affinity may be made in HVRs. A useful method for identification of residues or regions of an antibody that may be targeted for mutagenesis is called "alanine scanning mutagenesis" as described by Cunningham and Wells (1989) *Science*, 244:1081-1085. In this method, a residue or group of target residues (e.g., charged residues such as Arg, Asp, His, Lys, and Glu) are identified and replaced by a neutral or negatively charged amino acid (e.g., alanine or polyalanine) to determine whether the interaction of the antibody with antigen is affected. Further substitutions may be introduced at the amino acid locations demonstrating functional sensitivity to the initial substitutions. Alternatively, or additionally, a crystal structure of an antigen-antigen binding molecule complex to identify contact points between the antibody and antigen. Such contact residues and neighboring residues may be targeted or eliminated as candidates for substitution. Variants may be screened to determine whether they contain the desired properties.

Amino acid sequence insertions include amino- and/or carboxyl-terminal fusions ranging in length from one residue to polypeptides containing a hundred or more residues, as well as intrasequence insertions of single or multiple amino acid residues. Examples of terminal insertions include TNF family ligand trimer-containing antigen binding molecule with an N-terminal methionyl residue. Other insertional variants of the molecule include the fusion to the N- or C-terminus to a polypeptide which increases the serum half-life of the TNF ligand trimer-containing antigen binding molecules.

In certain embodiments, the TNF family ligand trimer-containing antigen binding molecules provided herein are altered to increase or decrease the extent to which the antibody is glycosylated. Glycosylation variants of the molecules may be conveniently obtained by altering the amino acid sequence such that one or more glycosylation sites is created or removed. Where the TNF ligand trimer-containing antigen binding molecule comprises an Fc region, the carbohydrate attached thereto may be altered. Native antibodies produced by mammalian cells typically comprise a branched, biantennary oligosaccharide that is generally attached by an N-linkage to Asn297 of the CH2 domain of the Fc region. See, e.g., Wright et al. *TIBTECH* 15:26-32 (1997). The oligosaccharide may include various carbohydrates, e.g., mannose, N-acetyl glucosamine (GlcNAc), galactose, and sialic acid, as well as a fucose attached to a GlcNAc in the "stem" of the biantennary oligosaccharide structure. In some embodiments, modifications of the oligosaccharide in TNF family ligand trimer-containing antigen binding molecule may be made in order to create variants with certain improved properties. In one aspect, variants of TNF family ligand trimer-containing antigen binding molecules are provided having a carbohydrate structure that lacks fucose attached (directly or indirectly) to an Fc region. Such fucosylation variants may have improved ADCC function, see e.g. US Patent Publication Nos. US 2003/0157108 (Presta, L.) or US 2004/0093621 (Kyowa Hakko Kogyo Co., Ltd). Further variants of the TNF family ligand trimer-containing antigen binding molecules of the invention

include those with bisected oligosaccharides, e.g., in which a biantennary oligosaccharide attached to the Fc region is bisected by GlcNAc. Such variants may have reduced fucosylation and/or improved ADCC function., see for example WO 2003/011878 (Jean-Mairet et al.); US Patent No. 6,602,684 (Umana et al.); and US 2005/0123546 (Umana *et al.*). Variants with at least one galactose residue in the oligosaccharide attached to the Fc region are also provided. Such antibody variants may have improved CDC function and are described, e.g., in WO 1997/30087 (Patel et al.); WO 1998/58964 (Raju, S.); and WO 1999/22764 (Raju, S.).

In certain embodiments, it may be desirable to create **cysteine engineered variants** of the TNF family ligand trimer-containing antigen binding molecule of the invention, e.g., “thioMAbs,” in which one or more residues of the molecule are substituted with cysteine residues. In particular embodiments, the substituted residues occur at accessible sites of the molecule. By substituting those residues with cysteine, reactive thiol groups are thereby positioned at accessible sites of the antibody and may be used to conjugate the antibody to other moieties, such as drug moieties or linker-drug moieties, to create an immunoconjugate. In certain embodiments, any one or more of the following residues may be substituted with cysteine: V205 (Kabat numbering) of the light chain; A118 (EU numbering) of the heavy chain; and S400 (EU numbering) of the heavy chain Fc region. Cysteine engineered antigen binding molecules may be generated as described, e.g., in U.S. Patent No. 7,521,541.

In certain aspects, the TNF family ligand trimer-containing antigen binding molecules provided herein may be further modified to contain additional non-proteinaceous moieties that are known in the art and readily available. The moieties suitable for derivatization of the antibody include but are not limited to water soluble polymers. Non-limiting examples of water soluble polymers include, but are not limited to, polyethylene glycol (PEG), copolymers of ethylene glycol/propylene glycol, carboxymethylcellulose, dextran, polyvinyl alcohol, polyvinyl pyrrolidone, poly-1, 3-dioxolane, poly-1,3,6-trioxane, ethylene/maleic anhydride copolymer, polyaminoacids (either homopolymers or random copolymers), and dextran or poly(n-vinyl pyrrolidone)polyethylene glycol, propylene glycol homopolymers, polypropylene oxide/ethylene oxide co-polymers, polyoxyethylated polyols (e.g., glycerol), polyvinyl alcohol, and mixtures thereof. Polyethylene glycol propionaldehyde may have advantages in manufacturing due to its stability in water. The polymer may be of any molecular weight, and may be branched or unbranched. The number of polymers attached to the antibody may vary, and if more than one polymer is attached, they can be the same or different molecules. In general, the number and/or type of polymers used for derivatization can be determined based on considerations including, but not limited to, the particular properties or functions of the antibody to be improved, whether the bispecific antibody derivative will be used in a therapy under defined conditions, etc. In another aspect, conjugates of an antibody and non-proteinaceous moiety that may be selectively heated by exposure to radiation are provided. In one embodiment,

the non-proteinaceous moiety is a carbon nanotube (Kam, N.W. et al., Proc. Natl. Acad. Sci. USA 102 (2005) 11600-11605). The radiation may be of any wavelength, and includes, but is not limited to, wavelengths that do not harm ordinary cells, but which heat the non-proteinaceous moiety to a temperature at which cells proximal to the antibody-non-proteinaceous moiety are
5 killed.

In another aspect, immunoconjugates of the TNF family ligand trimer-containing antigen binding molecules provided herein maybe obtained. An “**immunoconjugate**” is an antibody conjugated to one or more heterologous molecule(s), including but not limited to a cytotoxic agent.

10 The term “**polynucleotide**” refers to an isolated nucleic acid molecule or construct, e.g. messenger RNA (mRNA), virally-derived RNA, or plasmid DNA (pDNA). A polynucleotide may comprise a conventional phosphodiester bond or a non-conventional bond (e.g. an amide bond, such as found in peptide nucleic acids (PNA). The term “nucleic acid molecule” refers to any one or more nucleic acid segments, e.g. DNA or RNA fragments, present in a polynucleotide.

15 By “**isolated**” nucleic acid molecule or polynucleotide is intended a nucleic acid molecule, DNA or RNA, which has been removed from its native environment. For example, a recombinant polynucleotide encoding a polypeptide contained in a vector is considered isolated for the purposes of the present invention. Further examples of an isolated polynucleotide include recombinant polynucleotides maintained in heterologous host cells or purified (partially or
20 substantially) polynucleotides in solution. An isolated polynucleotide includes a polynucleotide molecule contained in cells that ordinarily contain the polynucleotide molecule, but the polynucleotide molecule is present extrachromosomally or at a chromosomal location that is different from its natural chromosomal location. Isolated RNA molecules include in vivo or in vitro RNA transcripts of the present invention, as well as positive and negative strand forms, and
25 double-stranded forms. Isolated polynucleotides or nucleic acids according to the present invention further include such molecules produced synthetically. In addition, a polynucleotide or a nucleic acid may be or may include a regulatory element such as a promoter, ribosome binding site, or a transcription terminator.

By a nucleic acid or polynucleotide having a nucleotide sequence at least, for example,
30 95% “identical” to a reference nucleotide sequence of the present invention, it is intended that the nucleotide sequence of the polynucleotide is identical to the reference sequence except that the polynucleotide sequence may include up to five point mutations per each 100 nucleotides of the reference nucleotide sequence. In other words, to obtain a polynucleotide having a nucleotide sequence at least 95% identical to a reference nucleotide sequence, up to 5% of the nucleotides
35 in the reference sequence may be deleted or substituted with another nucleotide, or a number of

nucleotides up to 5% of the total nucleotides in the reference sequence may be inserted into the reference sequence. These alterations of the reference sequence may occur at the 5' or 3' terminal positions of the reference nucleotide sequence or anywhere between those terminal positions, interspersed either individually among residues in the reference sequence or in one or more contiguous groups within the reference sequence. As a practical matter, whether any particular polynucleotide sequence is at least 80%, 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to a nucleotide sequence of the present invention can be determined conventionally using known computer programs, such as the ones discussed above for polypeptides (e.g. ALIGN-2).

The term "**expression cassette**" refers to a polynucleotide generated recombinantly or synthetically, with a series of specified nucleic acid elements that permit transcription of a particular nucleic acid in a target cell. The recombinant expression cassette can be incorporated into a plasmid, chromosome, mitochondrial DNA, plastid DNA, virus, or nucleic acid fragment. Typically, the recombinant expression cassette portion of an expression vector includes, among other sequences, a nucleic acid sequence to be transcribed and a promoter. In certain embodiments, the expression cassette of the invention comprises polynucleotide sequences that encode bispecific antigen binding molecules of the invention or fragments thereof.

The term "**vector**" or "expression vector" is synonymous with "expression construct" and refers to a DNA molecule that is used to introduce and direct the expression of a specific gene to which it is operably associated in a target cell. The term includes the vector as a self-replicating nucleic acid structure as well as the vector incorporated into the genome of a host cell into which it has been introduced. The expression vector of the present invention comprises an expression cassette. Expression vectors allow transcription of large amounts of stable mRNA. Once the expression vector is inside the target cell, the ribonucleic acid molecule or protein that is encoded by the gene is produced by the cellular transcription and/or translation machinery. In one embodiment, the expression vector of the invention comprises an expression cassette that comprises polynucleotide sequences that encode bispecific antigen binding molecules of the invention or fragments thereof.

The terms "**host cell**", "host cell line," and "host cell culture" are used interchangeably and refer to cells into which exogenous nucleic acid has been introduced, including the progeny of such cells. Host cells include "transformants" and "transformed cells," which include the primary transformed cell and progeny derived therefrom without regard to the number of passages. Progeny may not be completely identical in nucleic acid content to a parent cell, but may contain mutations. Mutant progeny that have the same function or biological activity as screened or selected for in the originally transformed cell are included herein. A host cell is any type of cellular system that can be used to generate the bispecific antigen binding molecules of the

present invention. Host cells include cultured cells, e.g. mammalian cultured cells, such as CHO cells, BHK cells, NS0 cells, SP2/0 cells, YO myeloma cells, P3X63 mouse myeloma cells, PER cells, PER.C6 cells or hybridoma cells, yeast cells, insect cells, and plant cells, to name only a few, but also cells comprised within a transgenic animal, transgenic plant or cultured plant or
5 animal tissue.

An "**effective amount**" of an agent refers to the amount that is necessary to result in a physiological change in the cell or tissue to which it is administered.

A "**therapeutically effective amount**" of an agent, e.g. a pharmaceutical composition, refers to an amount effective, at dosages and for periods of time necessary, to achieve the desired
10 therapeutic or prophylactic result. A therapeutically effective amount of an agent for example eliminates, decreases, delays, minimizes or prevents adverse effects of a disease.

An "**individual**" or "subject" is a mammal. Mammals include, but are not limited to, domesticated animals (e.g. cows, sheep, cats, dogs, and horses), primates (e.g. humans and non-human primates such as monkeys), rabbits, and rodents (e.g. mice and rats). Particularly, the
15 individual or subject is a human.

The term "**pharmaceutical composition**" refers to a preparation which is in such form as to permit the biological activity of an active ingredient contained therein to be effective, and which contains no additional components which are unacceptably toxic to a subject to which the formulation would be administered.

20 A "**pharmaceutically acceptable excipient**" refers to an ingredient in a pharmaceutical composition, other than an active ingredient, which is nontoxic to a subject. A pharmaceutically acceptable excipient includes, but is not limited to, a buffer, a stabilizer, or a preservative.

The term "**package insert**" is used to refer to instructions customarily included in commercial packages of therapeutic products, that contain information about the indications,
25 usage, dosage, administration, combination therapy, contraindications and/or warnings concerning the use of such therapeutic products.

As used herein, "**treatment**" (and grammatical variations thereof such as "treat" or "treating") refers to clinical intervention in an attempt to alter the natural course of the individual being treated, and can be performed either for prophylaxis or during the course of clinical
30 pathology. Desirable effects of treatment include, but are not limited to, preventing occurrence or recurrence of disease, alleviation of symptoms, diminishment of any direct or indirect pathological consequences of the disease, preventing metastasis, decreasing the rate of disease progression, amelioration or palliation of the disease state, and remission or improved prognosis.

In some embodiments, the molecules of the invention are used to delay development of a disease or to slow the progression of a disease.

The term “**cancer**” as used herein refers to proliferative diseases, such as lymphomas, carcinoma, lymphoma, blastoma, sarcoma, leukemia, lymphocytic leukemias, lung cancer, non-
5 small cell lung (NSCL) cancer, bronchioloalviolar cell lung cancer, bone cancer, pancreatic cancer, skin cancer, cancer of the head or neck, cutaneous or intraocular melanoma, uterine cancer, ovarian cancer, rectal cancer, cancer of the anal region, stomach cancer, gastric cancer, colorectal cancer (CRC), pancreatic cancer, breast cancer, triple-negative breast cancer, uterine
10 carcinoma of the fallopian tubes, carcinoma of the endometrium, carcinoma of the cervix, carcinoma of the vagina, carcinoma of the vulva, Hodgkin's Disease, cancer of the esophagus, cancer of the small intestine, cancer of the endocrine system, cancer of the thyroid gland, cancer of the parathyroid gland, cancer of the adrenal gland, sarcoma of soft tissue, cancer of the urethra, cancer of the penis, prostate cancer, cancer of the bladder, cancer of the kidney or ureter, renal cell carcinoma, carcinoma of the renal pelvis, mesothelioma, hepatocellular cancer, biliary
15 cancer, neoplasms of the central nervous system (CNS), spinal axis tumors, brain stem glioma, glioblastoma multiforme, astrocytomas, schwannomas, ependymomas, medulloblastomas, meningiomas, squamous cell carcinomas, pituitary adenoma and Ewings sarcoma, melanoma, multiple myeloma, B-cell cancer (lymphoma), chronic lymphocytic leukemia (CLL), acute lymphoblastic leukemia (ALL), hairy cell leukemia, chronic myeloblastic leukemia, including
20 refractory versions of any of the above cancers, or a combination of one or more of the above cancers.

TNF family ligand trimer-containing antigen binding molecules of the invention

The invention provides novel TNF family ligand trimer-containing antigen binding molecules with particularly advantageous properties such as producibility, stability, binding
25 affinity, biological activity, targeting efficiency and reduced toxicity.

In a first aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising

(a) at least one moiety capable of specific binding to a target cell antigen and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

30 wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or two fragments thereof that are connected to each other by a peptide linker and in that the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof.

In a particular aspect, the invention provides a TNF family ligand trimer-containing
35 antigen binding molecule comprising

- (a) at least one moiety capable of specific binding to a target cell antigen,
(b) a first and a second polypeptide that are linked to each other by a disulfide bond,
wherein the antigen binding molecule is characterized in that the first polypeptide comprises two
ectodomains of a TNF ligand family member or two fragments thereof that are connected to each
5 other by a peptide linker and in that the second polypeptide comprises only one ectodomain of
said TNF ligand family member or a fragment thereof, and
(c) an Fc domain composed of a first and a second subunit capable of stable association.

In a particular aspect, the TNF family ligand trimer-containing antigen binding molecule
comprises (a) at least one moiety capable of specific binding to a target cell antigen and
10 (b) a first and a second polypeptide that are linked to each other by a disulfide bond,
wherein the antigen binding molecule is characterized in that the first polypeptide comprises two
ectodomains of a TNF ligand family member or two fragments thereof that are connected to each
other by a peptide linker and in that the second polypeptide comprises only one ectodomain of
said TNF ligand family member or a fragment thereof, wherein the TNF ligand family member
15 is costimulates human T-cell activation.

In another particular aspect, the TNF family ligand trimer-containing antigen binding
molecule comprises (a) at least one moiety capable of specific binding to a target cell antigen
and
(b) a first and a second polypeptide that are linked to each other by a disulfide bond,
20 wherein the antigen binding molecule is characterized in that the first polypeptide comprises two
ectodomains of a TNF ligand family member or two fragments thereof that are connected to each
other by a peptide linker and in that the second polypeptide comprises only one ectodomain of
said TNF ligand family member or a fragment thereof, wherein the ectodomains of a TNF ligand
family member are identical in all instances.

25 In a further aspect, provided is a TNF family ligand trimer-containing antigen binding
molecule of claim 1, comprising

- (a) at least one moiety capable of specific binding to a target cell antigen and
(b) a first and a second polypeptide that are linked to each other by a disulfide bond,
wherein the antigen binding molecule is characterized in that

30 (i) the first polypeptide contains a CH1 or CL domain and the second polypeptide contains
a CL or CH1 domain, respectively, wherein the second polypeptide is linked to the first
polypeptide by a disulfide bond between the CH1 and CL domain, and wherein the first
polypeptide comprises two ectodomains of a TNF ligand family member or fragments
thereof that are connected to each other and to the CH1 or CL domain by a peptide
35 linker and wherein the second polypeptide comprises one ectodomain of said TNF

ligand family member or a fragment thereof connected via a peptide linker to the CL or CH1 domain of said polypeptide, or

- (ii) the first polypeptide contains a CH3 domain and the second polypeptide contains a CH3 domain, respectively, and wherein the first polypeptide comprises two
 5 ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the C-terminus of the CH3 domain by a peptide linker and wherein the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to C-terminus of the CH3 domain of said polypeptide, or
- 10 (iii) the first polypeptide contains a VH-CL or a VL-CH1 domain and the second polypeptide contains a VL-CH1 domain or a VH-CL domain, respectively, wherein the second polypeptide is linked to the first polypeptide by a disulfide bond between the CH1 and CL domain, and wherein the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to
 15 to VH or VL by a peptide linker and wherein the second polypeptide comprises one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to VL or VH of said polypeptide.

In a particular aspect, the TNF family ligand trimer-containing antigen binding molecule comprises a TNF ligand family member that costimulates human T-cell activation which is
 20 selected from 4-1BBL and OX40L. More particularly, the TNF ligand family member is 4-1BBL.

In another aspect, wherein the ectodomain of a TNF ligand family member comprises the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO: 2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:96, SEQ ID NO: 373, SEQ ID NO:374 and SEQ ID NO:375, particularly the amino acid sequence of SEQ ID NO:1 or SEQ ID NO:96. In one aspect,
 25 the ectodomain of a TNF ligand family member or fragment thereof comprises the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO: 2, SEQ ID NO:3, SEQ ID NO:4 and SEQ ID NO:96, particularly the amino acid sequence of SEQ ID NO:1 or SEQ ID NO:96. In a particular aspect, the the ectodomain of a TNF ligand family member or fragment thereof comprises the amino acid sequence of SEQ ID NO:96.

30 In a further aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

- (a) at least one moiety capable of specific binding to a target cell antigen and
- (b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises the
 35 amino acid sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:97, SEQ ID NO:98 and SEQ ID NO:99 and in that the second polypeptide comprises the amino acid

sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:96, SEQ ID NO:3 and SEQ ID NO:4. In a particular aspect, the first polypeptide comprises the amino acid sequence of SEQ ID NO:97 and the second polypeptide comprises the amino acid sequence of SEQ ID NO:96.

5 In one aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

(a) at least one moiety capable of specific binding to a target cell antigen and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises the
10 amino acid sequence of SEQ ID NO:5 and in that the second polypeptide comprises the amino acid sequence of SEQ ID NO:6.

In a further aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

(a) at least one moiety capable of specific binding to a target cell antigen and

15 (b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence of SEQ ID NO:5 and in that the second polypeptide comprises the amino acid sequence of SEQ ID NO:183.

In yet a further aspect, the TNF family ligand trimer-containing antigen binding molecule
20 of the invention comprises

(a) at least one moiety capable of specific binding to a target cell antigen and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence of SEQ ID NO:97 and in that the second polypeptide comprises the amino
25 acid sequence of SEQ ID NO:184 or SEQ ID NO:185.

In another aspect, the TNF ligand family member is OX40L. In a particular aspect, provided is TNF family ligand trimer-containing antigen binding molecule, wherein the ectodomain of a TNF ligand family member comprises the amino acid sequence of SEQ ID NO:53 or SEQ ID NO:54, particularly the amino acid sequence of SEQ ID NO:53.

30 In one aspect, the invention relates to a TNF family ligand trimer-containing antigen binding molecule comprising

(a) at least one moiety capable of specific binding to a target cell antigen and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence of SEQ ID NO:371 or SEQ ID:372 and in that the second polypeptide comprises the amino acid sequence of SEQ ID NO:53 or SEQ ID NO:54, respectively.

In one aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

(a) at least one moiety capable of specific binding to a target cell antigen,
(b) a first polypeptide containing a CH1 or CL domain and a second polypeptide containing a CL or CH1 domain, respectively, wherein the second polypeptide is linked to the first polypeptide by a disulfide bond between the CH1 and CL domain,

and wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the CH1 or CL domain by a peptide linker and in that the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to the CL or CH1 domain of said polypeptide.

In one aspect, provided is a TNF family ligand trimer-containing antigen binding molecule comprising

(a) at least one moiety capable of specific binding to a target cell antigen,
(b) a first polypeptide containing a CH1 domain and a second polypeptide containing a CL domain, wherein the second polypeptide is linked to the first polypeptide by a disulfide bond between the CH1 and CL domain,

and wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the CH1 domain by a peptide linker and in that the second polypeptide comprises one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to the CL domain of said polypeptide.

In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule comprising

(a) at least one moiety capable of specific binding to a target cell antigen,
(b) a first polypeptide containing a CL domain and a second polypeptide containing a CH1 domain, wherein the second polypeptide is linked to the first polypeptide by a disulfide bond between the CH1 and CL domain,

and wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the CL domain by a peptide linker and in that the second polypeptide comprises one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to the CH1 domain of said polypeptide.

In another aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising

(a) one moiety capable of specific binding to a target cell antigen and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

5 wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or two fragments thereof that are connected to each other by a peptide linker and in that the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof.

In yet another aspect, the invention provides a TNF family ligand trimer-containing
10 antigen binding molecule comprising

(a) more than one moiety capable of specific binding to a target cell antigen and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or two fragments thereof that are connected to each
15 other by a peptide linker and in that the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to said polypeptide.

In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising

20 (a) two moieties capable of specific binding to a target cell antigen and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or two fragments thereof that are connected to each other by a peptide linker and in that the second polypeptide comprises only one ectodomain of
25 said TNF ligand family member or a fragment thereof.

In a particular aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, comprising

(a) at least one moiety capable of specific binding to a target cell antigen, and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

30 wherein the antigen binding molecule is characterized in that the first polypeptide contains a CH3 domain and the second polypeptide contains a CH3 domain, respectively, and wherein the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the C-terminus of the CH3 domain by a peptide linker and wherein the second polypeptide comprises one ectodomain of said TNF ligand family
35 member or a fragment thereof connected via a peptide linker to C-terminus of the CH3 domain

of said polypeptide. Particularly, such TNF family ligand trimer-containing antigen binding molecule comprises two moieties capable of specific binding to a target cell antigen.

In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising

- 5 (a) two moieties capable of specific binding to a target cell antigen and
(b) a first and a second polypeptide that are linked to each other by a disulfide bond,
wherein the antigen binding molecule is characterized in that the first polypeptide comprises two
ectodomains of a TNF ligand family member or two fragments thereof that are connected to each
other by a peptide linker and in that the second polypeptide comprises one ectodomain of said
10 TNF ligand family member or a fragment thereof,
wherein the two moieties capable of specific binding to a target cell antigen bind to two different
target cell antigens.

- In a further aspect, the invention provides a TNF family ligand trimer-containing antigen
binding molecule as defined herein before, wherein the moiety capable of specific binding to a
15 target cell antigen is selected from the group consisting of an antibody, an antibody fragment and
a scaffold antigen binding protein.

- In one aspect, provided is a TNF family ligand trimer-containing antigen binding molecule
as described herein before, wherein the moiety capable of specific binding to a target cell antigen
is selected from the group consisting of an antibody fragment, a Fab molecule, a crossover Fab
20 molecule, a single chain Fab molecule, a Fv molecule, a scFv molecule, a single domain
antibody, an aVH and a scaffold antigen binding protein. In one aspect, the moiety capable of
specific binding to a target cell antigen is an aVH or a scaffold antigen binding protein. In one
aspect, the moiety capable of specific binding to a target cell antigen is a scaffold antigen
binding protein capable of specific binding to a target cell antigen.

- 25 In particular, the TNF family ligand trimer-containing antigen binding molecule comprises
one or two moieties capable of specific binding to a target cell antigen.

- In a particular aspect, provided is a TNF family ligand trimer-containing antigen binding
molecule, wherein the moiety capable of specific binding to a target cell antigen is a Fab
molecule or a crossover Fab molecule capable of specific binding to a target cell antigen. In
30 particular, the moiety capable of specific binding to a target cell antigen is a Fab capable of
specific binding to a target cell antigen.

Furthermore, provided is TNF family ligand trimer-containing antigen binding molecule as
described herein, wherein the target cell antigen is selected from the group consisting of
Fibroblast Activation Protein (FAP), Melanoma-associated Chondroitin Sulfate Proteoglycan

(MCSP), Epidermal Growth Factor Receptor (EGFR), Carcinoembryonic Antigen (CEA), CD19, CD20 and CD33.

In a further aspect, provided is a TNF family ligand trimer-containing antigen binding molecule according to the invention, wherein a peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus to the CH1 domain of a heavy chain by a second peptide linker and wherein one ectodomain of said TNF ligand family member or a fragment thereof is fused at the its C-terminus to the CL domain on a light chain by a third peptide linker.

In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule according to the invention, wherein a peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus to the CL domain of a heavy chain by a second peptide linker and wherein one ectodomain of said TNF ligand family member or a fragment thereof is fused at the its C-terminus to the CH1 domain on a light chain by a third peptide linker.

In a further aspect, the invention is concerned with a TNF family ligand trimer-containing antigen binding molecule according to the invention, wherein a peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus to the CL domain of a light chain by a second peptide linker and wherein one ectodomain of said TNF ligand family member or a fragment thereof is fused at the its C-terminus to the CH1 domain of the heavy chain by a third peptide linker.

In a particular aspect, the invention relates to a TNF family ligand trimer-containing antigen binding molecule as defined above, wherein the peptide linker is (G4S)₂. In one aspect, the first peptide linker is (G4S)₂ (SEQ ID NO:13), the second peptide linker is GSPGSSSSGS (SEQ ID NO:57) and the third peptide linker is (G4S)₂ (SEQ ID NO:13). In particular, the invention relates to a TNF ligand trimer-containing antigen binding molecule as defined above, wherein the first peptide linker is (G4S)₂ (SEQ ID NO:13), the second peptide linker is (G4S)₂ (SEQ ID NO:13), and the third peptide linker is (G4S)₂ (SEQ ID NO:13).

In another aspect, the TNF family ligand trimer-containing antigen binding molecule as defined herein before comprises an Fc domain composed of a first and a second subunit capable of stable association.

In particular, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises (a) a Fab molecule capable of specific binding to a target cell antigen, wherein the Fab heavy chain is fused at the C-terminus to the N-terminus of a CH2 domain in

the Fc domain and (c) an Fc domain composed of a first and a second subunit capable of stable association.

In a further aspect, the Fc domain is an IgG, particularly an IgG1 Fc domain or an IgG4 Fc domain. More particularly, the Fc domain is an IgG1 Fc domain. In a particular aspect, the Fc domain comprises a modification promoting the association of the first and second subunit of the Fc domain.

Fc domain modifications reducing Fc receptor binding and/or effector function

The Fc domain of the TNF family ligand trimer-containing antigen binding molecules of the invention consists of a pair of polypeptide chains comprising heavy chain domains of an immunoglobulin molecule. For example, the Fc domain of an immunoglobulin G (IgG) molecule is a dimer, each subunit of which comprises the CH2 and CH3 IgG heavy chain constant domains. The two subunits of the Fc domain are capable of stable association with each other.

The Fc domain confers favorable pharmacokinetic properties to the antigen binding molecules of the invention, including a long serum half-life which contributes to good accumulation in the target tissue and a favorable tissue-blood distribution ratio. At the same time it may, however, lead to undesirable targeting of the bispecific antibodies of the invention to cells expressing Fc receptors rather than to the preferred antigen-bearing cells. Accordingly, in particular aspects, the Fc domain of the TNF family ligand trimer-containing antigen binding molecule of the invention exhibits reduced binding affinity to an Fc receptor and/or reduced effector function, as compared to a native IgG1 Fc domain. In one aspect, the Fc does not substantially bind to an Fc receptor and/or does not induce effector function. In a particular aspect the Fc receptor is an Fc γ receptor. In one aspect, the Fc receptor is a human Fc receptor. In a specific aspect, the Fc receptor is an activating human Fc γ receptor, more specifically human Fc γ RIIIa, Fc γ RI or Fc γ RIIa, most specifically human Fc γ RIIIa. In one aspect, the Fc domain does not induce effector function. The reduced effector function can include, but is not limited to, one or more of the following: reduced complement dependent cytotoxicity (CDC), reduced antibody-dependent cell-mediated cytotoxicity (ADCC), reduced antibody-dependent cellular phagocytosis (ADCP), reduced cytokine secretion, reduced immune complex-mediated antigen uptake by antigen-presenting cells, reduced binding to NK cells, reduced binding to macrophages, reduced binding to monocytes, reduced binding to polymorphonuclear cells, reduced direct signaling inducing apoptosis, reduced dendritic cell maturation, or reduced T cell priming.

In certain aspects, one or more amino acid modifications may be introduced into the Fc region of a TNF family ligand trimer-containing antigen binding molecule provided herein, thereby generating an Fc region variant. The Fc region variant may comprise a human Fc region

sequence (e.g., a human IgG1, IgG2, IgG3 or IgG4 Fc region) comprising an amino acid modification (e.g. a substitution) at one or more amino acid positions.

In a particular aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising

- 5 (a) at least one moiety capable of specific binding to a target cell antigen,
(b) a first and a second polypeptide that are linked to each other by a disulfide bond,
wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or two fragments thereof that are connected to each other by a peptide linker and in that the second polypeptide comprises only one ectodomain of
10 said TNF ligand family member or a fragment thereof, and
(c) an Fc domain composed of a first and a second subunit capable of stable association, wherein the Fc domain comprises one or more amino acid substitution that reduces binding to an Fc receptor, in particular towards Fcγ receptor.

In one aspect, the Fc domain of the TNF family ligand trimer-containing antigen binding
15 molecule of the invention comprises one or more amino acid mutation that reduces the binding affinity of the Fc domain to an Fc receptor and/or effector function. Typically, the same one or more amino acid mutation is present in each of the two subunits of the Fc domain. In particular, the Fc domain comprises an amino acid substitution at a position of E233, L234, L235, N297, P331 and P329 (EU numbering). In particular, the Fc domain comprises amino acid substitutions
20 at positions 234 and 235 (EU numbering) and/or 329 (EU numbering) of the IgG heavy chains. More particularly, provided is a trimeric TNF family ligand-containing antigen binding molecule according to the invention which comprises an Fc domain with the amino acid substitutions L234A, L235A and P329G ("P329G LALA", EU numbering) in the IgG heavy chains. The amino acid substitutions L234A and L235A refer to the so-called LALA mutation. The "P329G
25 LALA" combination of amino acid substitutions almost completely abolishes Fcγ receptor binding of a human IgG1 Fc domain and is described in International Patent Appl. Publ. No. WO 2012/130831 A1 which also describes methods of preparing such mutant Fc domains and methods for determining its properties such as Fc receptor binding or effector functions. "EU numbering" refers to the numbering according to EU index of Kabat et al , Sequences of Proteins
30 of Immunological Interest, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, MD, 1991.

Fc domains with reduced Fc receptor binding and/or effector function also include those with substitution of one or more of Fc domain residues 238, 265, 269, 270, 297, 327 and 329 (U.S. Patent No. 6,737,056). Such Fc mutants include Fc mutants with substitutions at two or
35 more of amino acid positions 265, 269, 270, 297 and 327, including the so-called "DANA" Fc mutant with substitution of residues 265 and 297 to alanine (US Patent No. 7,332,581).

In another aspect, the Fc domain is an IgG4 Fc domain. IgG4 antibodies exhibit reduced binding affinity to Fc receptors and reduced effector functions as compared to IgG1 antibodies. In a more specific aspect, the Fc domain is an IgG4 Fc domain comprising an amino acid substitution at position S228 (Kabat numbering), particularly the amino acid substitution S228P.

5 In a more specific aspect, the Fc domain is an IgG4 Fc domain comprising amino acid substitutions L235E and S228P and P329G (EU numbering). Such IgG4 Fc domain mutants and their Fc γ receptor binding properties are also described in WO 2012/130831.

Mutant Fc domains can be prepared by amino acid deletion, substitution, insertion or modification using genetic or chemical methods well known in the art. Genetic methods may
10 include site-specific mutagenesis of the encoding DNA sequence, PCR, gene synthesis, and the like. The correct nucleotide changes can be verified for example by sequencing.

Binding to Fc receptors can be easily determined e.g. by ELISA, or by Surface Plasmon Resonance (SPR) using standard instrumentation such as a BIAcore instrument (GE Healthcare), and Fc receptors such as may be obtained by recombinant expression. A suitable such binding
15 assay is described herein. Alternatively, binding affinity of Fc domains or cell activating bispecific antigen binding molecules comprising an Fc domain for Fc receptors may be evaluated using cell lines known to express particular Fc receptors, such as human NK cells expressing Fc γ IIIa receptor.

Effector function of an Fc domain, or bispecific antibodies of the invention comprising an
20 Fc domain, can be measured by methods known in the art. A suitable assay for measuring ADCC is described herein. Other examples of in vitro assays to assess ADCC activity of a molecule of interest are described in U.S. Patent No. 5,500,362; Hellstrom et al. Proc Natl Acad Sci USA 83, 7059-7063 (1986) and Hellstrom et al., Proc Natl Acad Sci USA 82, 1499-1502 (1985); U.S. Patent No. 5,821,337; Bruggemann et al., J Exp Med 166, 1351-1361 (1987). Alternatively, non-
25 radioactive assays methods may be employed (see, for example, ACTITM non-radioactive cytotoxicity assay for flow cytometry (CellTechnology, Inc. Mountain View, CA); and CytoTox 96® non-radioactive cytotoxicity assay (Promega, Madison, WI)). Useful effector cells for such assays include peripheral blood mononuclear cells (PBMC) and Natural Killer (NK) cells. Alternatively, or additionally, ADCC activity of the molecule of interest may be assessed in vivo,
30 e.g. in a animal model such as that disclosed in Clynes et al., Proc Natl Acad Sci USA 95, 652-656 (1998).

In some embodiments, binding of the Fc domain to a complement component, specifically to C1q, is reduced. Accordingly, in some embodiments wherein the Fc domain is engineered to have reduced effector function, said reduced effector function includes reduced CDC. C1q
35 binding assays may be carried out to determine whether the bispecific antibodies of the invention

is able to bind C1q and hence has CDC activity. See e.g., C1q and C3c binding ELISA in WO 2006/029879 and WO 2005/100402. To assess complement activation, a CDC assay may be performed (see, for example, Gazzano-Santoro et al., J Immunol Methods 202, 163 (1996); Cragg et al., Blood 101, 1045-1052 (2003); and Cragg and Glennie, Blood 103, 2738-2743 (2004)).

In a particular aspect, the Fc domain comprises a modification promoting the association of the first and second subunit of the Fc domain.

Fc domain modifications promoting heterodimerization

In one aspect, the TNF family ligand trimer-containing antigen binding molecules of the invention comprise (a) at least one moiety capable of specific binding to a target cell antigen, (b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or two fragments thereof that are connected to each other by a peptide linker and in that the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof, and (c) an Fc domain composed of a first and a second subunit capable of stable association, wherein the Fc domain comprises one or more amino acid substitution that reduces binding to an Fc receptor, in particular towards Fcγ receptor. Thus, they comprise different moieties, fused to one or the other of the two subunits of the Fc domain that are typically comprised in two non-identical polypeptide chains (“heavy chains”). Recombinant co-expression of these polypeptides and subsequent dimerization leads to several possible combinations of the two polypeptides. To improve the yield and purity of the TNF family ligand trimer-containing antigen binding molecules in recombinant production, it will thus be advantageous to introduce in the Fc domain of the TNF family ligand trimer-containing antigen binding molecules of the invention a modification promoting the association of the desired polypeptides.

Accordingly, the Fc domain of the TNF family ligand trimer-containing antigen binding molecules of the invention comprises a modification promoting the association of the first and the second subunit of the Fc domain. The site of most extensive protein-protein interaction between the two subunits of a human IgG Fc domain is in the CH3 domain of the Fc domain. Thus, said modification is particularly in the CH3 domain of the Fc domain.

In a specific aspect, said modification is a so-called “knob-into-hole” modification, comprising a “knob” modification in one of the two subunits of the Fc domain and a “hole” modification in the other one of the two subunits of the Fc domain. Thus, in a particular aspect, the invention relates to a TNF family ligand trimer-containing antigen binding molecule as described herein before which comprises an IgG molecule, wherein the Fc part of the first heavy

chain comprises a first dimerization module and the Fc part of the second heavy chain comprises a second dimerization module allowing a heterodimerization of the two heavy chains of the IgG molecule and the first dimerization module comprises knobs and the second dimerization module comprises holes according to the knob into hole technology.

5 The knob-into-hole technology is described e.g. in US 5,731,168; US 7,695,936; Ridgway et al., Prot Eng 9, 617-621 (1996) and Carter, J Immunol Meth 248, 7-15 (2001). Generally, the method involves introducing a protuberance ("knob") at the interface of a first polypeptide and a corresponding cavity ("hole") in the interface of a second polypeptide, such that the protuberance can be positioned in the cavity so as to promote heterodimer formation and hinder
10 homodimer formation. Protuberances are constructed by replacing small amino acid side chains from the interface of the first polypeptide with larger side chains (e.g. tyrosine or tryptophan). Compensatory cavities of identical or similar size to the protuberances are created in the interface of the second polypeptide by replacing large amino acid side chains with smaller ones (e.g. alanine or threonine).

15 Accordingly, in a particular aspect, in the CH3 domain of the first subunit of the Fc domain of the TNF family ligand trimer-containing antigen binding molecules of the invention an amino acid residue is replaced with an amino acid residue having a larger side chain volume, thereby generating a protuberance within the CH3 domain of the first subunit which is positionable in a cavity within the CH3 domain of the second subunit, and in the CH3 domain of the second
20 subunit of the Fc domain an amino acid residue is replaced with an amino acid residue having a smaller side chain volume, thereby generating a cavity within the CH3 domain of the second subunit within which the protuberance within the CH3 domain of the first subunit is positionable.

The protuberance and cavity can be made by altering the nucleic acid encoding the polypeptides, e.g. by site-specific mutagenesis, or by peptide synthesis.

25 In a specific aspect, in the CH3 domain of the first subunit of the Fc domain the threonine residue at position 366 is replaced with a tryptophan residue (T366W), and in the CH3 domain of the second subunit of the Fc domain the tyrosine residue at position 407 is replaced with a valine residue (Y407V). More particularly, in the second subunit of the Fc domain additionally the threonine residue at position 366 is replaced with a serine residue (T366S) and the leucine
30 residue at position 368 is replaced with an alanine residue (L368A). More particularly, in the first subunit of the Fc domain additionally the serine residue at position 354 is replaced with a cysteine residue (S354C), and in the second subunit of the Fc domain additionally the tyrosine residue at position 349 is replaced by a cysteine residue (Y349C). The introduction of these two cysteine residues results in the formation of a disulfide bridge between the two subunits of the Fc

domain. The disulfide bridge further stabilizes the dimer (Carter, J Immunol Methods 248, 7-15 (2001)).

In an alternative aspect, a modification promoting association of the first and the second subunit of the Fc domain comprises a modification mediating electrostatic steering effects, e.g. as described in PCT publication WO 2009/089004. Generally, this method involves replacement of one or more amino acid residues at the interface of the two Fc domain subunits by charged amino acid residues so that homodimer formation becomes electrostatically unfavorable but heterodimerization electrostatically favorable.

Modifications in the CH1/CL domains

To further improve correct pairing, the TNF family ligand trimer-containing antigen binding molecules can contain different charged amino acid substitutions (so-called “charged residues”). These modifications are introduced in the crossed or non-crossed CH1 and CL domains. In a particular aspect, the invention relates to a TNF family ligand trimer-containing antigen binding molecule, wherein in one of CL domains the amino acid at position 123 (EU numbering) has been replaced by arginine (R) and the amino acid at position 124 (EU numbering) has been substituted by lysine (K) and wherein in one of the CH1 domains the amino acids at position 147 (EU numbering) and at position 213 (EU numbering) have been substituted by glutamic acid (E).

More particularly, the invention relates to a TNF family ligand trimer-containing antigen binding molecule, wherein in the CL domain adjacent to the TNF ligand family member the amino acid at position 123 (EU numbering) has been replaced by arginine (R) and the amino acid at position 124 (EU numbering) has been substituted by lysine (K), and wherein in the CH1 domain adjacent to the TNF ligand family member the amino acids at position 147 (EU numbering) and at position 213 (EU numbering) have been substituted by glutamic acid (E).

Particular TNF family ligand trimer-containing antigen binding molecules

In another aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, wherein the antigen binding molecule comprises a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific binding to a target cell antigen, a first peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker fused at its C-terminus by a second peptide linker to a second heavy or light chain, and a second peptide comprising one ectodomain of said TNF ligand family member fused at its C-terminus by a third peptide linker to a second light or heavy chain, respectively.

In a further aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the first peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by a second peptide linker to a CH1 domain that is part of a heavy chain, and the second peptide
5 comprising one ectodomain of said TNF ligand family member or a fragment thereof is fused at its C-terminus by a third peptide linker to a CL domain that is part of a light chain.

In yet another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the first peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by
10 a second peptide linker to a CL domain that is part of a heavy chain, and the second peptide comprising one ectodomain of said TNF ligand family member or a fragment thereof that is fused at its C-terminus by a third peptide linker to a CH1 domain that is part of a light chain.

In a further aspect, provided is a TNF family ligand trimer-containing antigen binding
15 molecule, wherein the first peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by a second peptide linker to a VH domain that is part of a heavy chain, and the second peptide comprising one ectodomain of said TNF ligand family member or a fragment thereof is fused at its C-terminus by a third peptide linker to a VL domain that is part of
20 a light chain.

In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule of claims 21 to 23, wherein in the CL domain adjacent to the TNF ligand family member the amino acid at position 123 (EU numbering) has been replaced by arginine (R) and the amino acid at position 124 (EU numbering) has been substituted by lysine (K), and
25 wherein in the CH1 domain adjacent to the TNF ligand family member the amino acids at position 147 (EU numbering) and at position 213 (EU numbering) have been substituted by glutamic acid (E). These modifications lead to so-called charged residues with advantageous properties that avoid undesired effects such as for example mispairing.

Furthermore, provided is TNF family ligand trimer-containing antigen binding molecule as
30 described herein, wherein the target cell antigen is selected from the group consisting of Fibroblast Activation Protein (FAP), Melanoma-associated Chondroitin Sulfate Proteoglycan (MCSP), Epidermal Growth Factor Receptor (EGFR), Carcinoembryonic Antigen (CEA), CD19, CD20 and CD33.

TNF family ligand trimer-containing antigen binding molecules, wherein the target cell antigen is FAP

In a particular aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the target cell antigen is Fibroblast Activation Protein (FAP).

5 In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, wherein the moiety capable of specific binding to FAP comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:7 or SEQ ID NO:100, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:8 or SEQ ID NO:101, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:9 or SEQ ID NO:102, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:10 or SEQ ID NO:103, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:11 or SEQ ID NO:104, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:12 or SEQ ID NO:105.

15 In a particular aspect, provided is a TNF family ligand trimer-containing antigen binding molecule of the invention, wherein the moiety capable of specific binding to a target cell antigen is a Fab molecule capable of specific binding to FAP and comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:7, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:8 and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:9, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:10, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:11 and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:12.

25 In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule of the invention, wherein the moiety capable of specific binding to a target cell antigen is a Fab molecule capable of specific binding to FAP and comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:100, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:101 and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:102, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:103, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:104 and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:105.

30 In a further aspect, the moiety capable of specific binding to FAP comprises a heavy chain variable region comprising an amino acid sequence that is at least about 95%, 96%, 97%, 98%, 99% or 100% identical to the amino acid sequence of SEQ ID NO:16 and a light chain variable region comprising an amino acid sequence that is at least about 95%, 96%, 97%, 98%, 99% or 100% identical to the amino acid sequence of SEQ ID NO:17.

In another aspect, the moiety capable of specific binding to FAP comprises a heavy chain variable region comprising an amino acid sequence that is at least about 95%, 96%, 97%, 98%, 99% or 100% identical to the amino acid sequence of SEQ ID NO:106 and a light chain variable region comprising an amino acid sequence that is at least about 95%, 96%, 97%, 98%, 99% or 100% identical to the amino acid sequence of SEQ ID NO:107.

In one aspect, the moiety capable of specific binding to FAP comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:16 and a variable light chain comprising an amino acid sequence of SEQ ID NO:17 or a variable heavy chain comprising an amino acid sequence of SEQ ID NO:106 and a variable light chain comprising an amino acid sequence of SEQ ID NO:107.

In a particular aspect, the moiety capable of specific binding to FAP comprises a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:16 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:17. In another particular aspect, the moiety capable of specific binding to FAP comprises a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:106 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:107. In a specific aspect, the moiety capable of specific binding to FAP comprises a VH domain consisting of amino acid sequence of SEQ ID NO:106 and a VL domain consisting of the amino acid sequence of SEQ ID NO:107.

In a further aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

(a) at least one moiety capable of specific binding to a target cell antigen comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:16 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:17, and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:97, SEQ ID NO:98 and SEQ ID NO:99 and in that the second polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:96, SEQ ID NO:3 and SEQ ID NO:4.

In a particular aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

(a) at least one moiety capable of specific binding to a target cell antigen comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:16 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:17, and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence of SEQ ID NO:97 and the second polypeptide comprises the amino acid sequence of SEQ ID NO:96.

In another aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

(a) at least one moiety capable of specific binding to a target cell antigen comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:106 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:107, and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:97, SEQ ID NO:98 and SEQ ID NO:99 and in that the second polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:96, SEQ ID NO:3 and SEQ ID NO:4.

In a particular aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

(a) at least one moiety capable of specific binding to a target cell antigen comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:106 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:107, and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence of SEQ ID NO:97 and the second polypeptide comprises the amino acid sequence of SEQ ID NO:96.

In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the antigen binding molecule comprises

a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific binding to a target cell antigen,

a second heavy chain comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker that is fused at its C-terminus by a

second peptide linker to a CH1 domain, and a second light chain comprising one ectodomain of said TNF ligand family member or a fragment thereof is fused at its C-terminus by a third peptide linker to a CL domain, and wherein the antigen binding molecule comprises

(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:16 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:17 or

a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID

NO:106 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:107,

(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:14, SEQ ID NO:108, SEQ ID NO:111 and SEQ ID NO:113, and

5 (iii) a second light chain comprising the amino acid sequence of SEQ ID NO:15, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:112 and SEQ ID NO:114.

In a further particular aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, and wherein the antigen binding molecule comprises

a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific

10 binding to a target cell antigen,

a second heavy chain comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by a second peptide linker to a CL domain, and a second light chain comprising one ectodomain of said TNF

15 linker to a CH1 domain, and wherein the molecule comprises

(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:16 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:17 or

20 a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:106 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:107,

(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:115, SEQ ID NO:117, SEQ ID NO:119 and SEQ ID NO:173, and

25 (iii) a second light chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:116, SEQ ID NO:118, SEQ ID NO:120 and SEQ ID NO:174.

More particularly, provided is a TNF family ligand trimer-containing antigen binding molecule comprising

(a) a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific binding to a target cell antigen, wherein the first heavy chain comprises the VH domain

30 comprising the amino acid sequence of SEQ ID NO:106 and the first light chain comprises the VL domain comprising the amino acid sequence of SEQ ID NO:107, and

(b) a second heavy chain comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by a second peptide linker to a CL domain, and a second light chain comprising one ectodomain of

35 said TNF ligand family member or a fragment thereof that is fused at its C-terminus by a third peptide linker to a CH1 domain, wherein the second heavy chain comprises the amino acid sequence of SEQ ID NO:119 or SEQ ID NO:173, and the second light chain comprises the

amino acid sequence of SEQ ID NO:120 or SEQ ID NO:174. In particular, the second heavy chain comprises the amino acid sequence of SEQ ID NO:119 and the second light chain comprises the amino acid sequence of SEQ ID NO:120.

Furthermore, the invention provides a TNF family ligand trimer-containing antigen binding molecule, comprising

(a) at least one moiety capable of specific binding to a target cell antigen, and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide contains a CH3 domain and the second polypeptide contains a CH3 domain, respectively, and wherein the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the C-terminus of the CH3 domain by a peptide linker and wherein the second polypeptide comprises one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to C-terminus of the CH3 domain of said polypeptide.

In a particular aspect, such a TNF family ligand trimer-containing antigen binding molecule comprises two moieties capable of specific binding to a target cell antigen.

More particular, such TNF family ligand trimer-containing antigen binding molecule comprises

- (i) a first heavy chain comprising the amino acid sequence of SEQ ID NO:121, a second heavy chain comprising the amino acid sequence of SEQ ID NO:122, and two light chains comprising the amino acid sequence of SEQ ID NO:19, or
- (ii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:123, a second heavy chain comprising the amino acid sequence of SEQ ID NO:124, and two light chains comprising the amino acid sequence of SEQ ID NO:125, or
- (iii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:126, a second heavy chain comprising the amino acid sequence of SEQ ID NO:127, and two light chains comprising the amino acid sequence of SEQ ID NO:125.

In a further aspect, the invention relates to a TNF family ligand trimer-containing antigen binding molecule, selected from the group consisting of:

a) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:18, a first light chain comprising the amino acid sequence of SEQ ID NO:19, a second heavy chain comprising the amino acid sequence of SEQ ID NO:14 and a second light chain comprising the amino acid sequence of SEQ ID NO:15;

b) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:18, a first light chain comprising the amino acid sequence of SEQ ID NO:19, a second

heavy chain comprising the amino acid sequence of SEQ ID NO:115 and a second light chain comprising the amino acid sequence of SEQ ID NO:116;

c) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:135, a first light chain comprising the amino acid sequence of SEQ ID NO:136, a second heavy chain comprising the amino acid sequence of SEQ ID NO:108 and a second light chain comprising the amino acid sequence of SEQ ID NO:109;

d) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:18, a first light chain comprising the amino acid sequence of SEQ ID NO:19, a second heavy chain comprising the amino acid sequence of SEQ ID NO:139 and a second light chain comprising the amino acid sequence of SEQ ID NO:140;

e) a molecule comprising two light chains comprising the amino acid sequence of SEQ ID NO:19, a first heavy chain comprising the amino acid sequence of SEQ ID NO:121 and a second heavy chain comprising the amino acid sequence of SEQ ID NO:122;

f) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:18, a first light chain comprising the amino acid sequence of SEQ ID NO:19, a second heavy chain comprising the amino acid sequence of SEQ ID NO:108 and a second light chain comprising the amino acid sequence of SEQ ID NO:110;

g) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:145, a first light chain comprising the amino acid sequence of SEQ ID NO:19, a second heavy chain comprising the amino acid sequence of SEQ ID NO:115 and a second light chain comprising the amino acid sequence of SEQ ID NO:116;

h) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:18, a first light chain comprising the amino acid sequence of SEQ ID NO:19, a second heavy chain comprising the amino acid sequence of SEQ ID NO:148 and a second light chain comprising the amino acid sequence of SEQ ID NO:149;

i) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:18, a first light chain comprising the amino acid sequence of SEQ ID NO:19, a second heavy chain comprising the amino acid sequence of SEQ ID NO:111 and a second light chain comprising the amino acid sequence of SEQ ID NO:112; and

j) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:18, a first light chain comprising the amino acid sequence of SEQ ID NO:19, a second heavy chain comprising the amino acid sequence of SEQ ID NO:113 and a second light chain comprising the amino acid sequence of SEQ ID NO:114.

In another aspect, the invention relates to a TNF family ligand trimer-containing antigen binding molecule, selected from the group consisting of:

a) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:164, a first light chain comprising the amino acid sequence of SEQ ID NO:125, a second

heavy chain comprising the amino acid sequence of SEQ ID NO:115 and a second light chain comprising the amino acid sequence of SEQ ID NO:116;

b) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:164, a first light chain comprising the amino acid sequence of SEQ ID NO:125, a second heavy chain comprising the amino acid sequence of SEQ ID NO:117 and a second light chain comprising the amino acid sequence of SEQ ID NO:118;

c) a molecule comprising two light chains comprising the amino acid sequence of SEQ ID NO:125, a first heavy chain comprising the amino acid sequence of SEQ ID NO:123 and a second heavy chain comprising the amino acid sequence of SEQ ID NO:124;

d) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:164, a first light chain comprising the amino acid sequence of SEQ ID NO:125, a second heavy chain comprising the amino acid sequence of SEQ ID NO:119 and a second light chain comprising the amino acid sequence of SEQ ID NO:120;

e) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:164, a first light chain comprising the amino acid sequence of SEQ ID NO:125, a second heavy chain comprising the amino acid sequence of SEQ ID NO:173 and a second light chain comprising the amino acid sequence of SEQ ID NO:174; and

f) a molecule comprising two light chains comprising the amino acid sequence of SEQ ID NO:125, a first heavy chain comprising the amino acid sequence of SEQ ID NO:126 and a second heavy chain comprising the amino acid sequence of SEQ ID NO:127.

In particular, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:164, a first light chain comprising the amino acid sequence of SEQ ID NO:125, a second heavy chain comprising the amino acid sequence of SEQ ID NO:119 and a second light chain comprising the amino acid sequence of SEQ ID NO:120.

In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the TNF ligand family member is OX40L and wherein the target cell antigen is Fibroblast Activation Protein (FAP) and the moiety capable of specific binding to FAP comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:7 or SEQ ID NO:100, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:8 or SEQ ID NO:101, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:9 or SEQ ID NO:102, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:10 or SEQ ID NO:103, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:11 or SEQ ID NO:104, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:12 or SEQ ID NO:105.

In a particular aspect, the TNF family ligand trimer-containing antigen binding molecule of comprises

(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:16 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:17 or

a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:106 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:107,

(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:355, and

(iii) a second light chain comprising the amino acid sequence of SEQ ID NO:356.

TNF family ligand trimer-containing antigen binding molecules, wherein the target cell antigen is CD19

In a particular aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the target cell antigen is CD19.

In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, wherein the moiety capable of specific binding to CD19 comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:195 or SEQ ID NO:252, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:196 or SEQ ID NO:253, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:197 or SEQ ID NO:254, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:198 or SEQ ID NO:249, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:199 or SEQ ID NO:250, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:200 or SEQ ID NO:251.

In a particular aspect, provided is a TNF family ligand trimer-containing antigen binding molecule of the invention, wherein the moiety capable of specific binding to a target cell antigen is a Fab molecule capable of specific binding to CD19 and comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:195, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:196 and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:197, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:198, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:199 and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:200.

In a further aspect, provided is a TNF family ligand trimer-containing antigen binding molecule of the invention, wherein the moiety capable of specific binding to a target cell antigen is a Fab molecule capable of specific binding to CD19 and comprises a VH domain comprising

(i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:252, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:253 and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:254, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:249, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:250 and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:251.

In another aspect, the moiety capable of specific binding to CD19 comprises a heavy chain variable region comprising an amino acid sequence that is at least about 95%, 96%, 97%, 98%, 99% or 100% identical to the amino acid sequence of SEQ ID NO:201 and a light chain variable region comprising an amino acid sequence that is at least about 95%, 96%, 97%, 98%, 99% or 100% identical to the amino acid sequence of SEQ ID NO:202.

In a further aspect, the moiety capable of specific binding to CD19 comprises a heavy chain variable region comprising an amino acid sequence that is at least about 95%, 96%, 97%, 98%, 99% or 100% identical to the amino acid sequence of SEQ ID NO:357 and a light chain variable region comprising an amino acid sequence that is at least about 95%, 96%, 97%, 98%, 99% or 100% identical to the amino acid sequence of SEQ ID NO:358.

In one aspect, the moiety capable of specific binding to CD19 comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:201 and a variable light chain comprising an amino acid sequence of SEQ ID NO:202 or wherein the moiety capable of specific binding to CD19 comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:357 and a variable light chain comprising an amino acid sequence of SEQ ID NO:358.

In a particular aspect, the moiety capable of specific binding to CD19 comprises a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:201 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:202. In another particular aspect, the moiety capable of specific binding to CD19 comprises a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:357 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:358.

In another aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

(a) at least one moiety capable of specific binding to a target cell antigen comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:201 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:202, and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:97, SEQ

ID NO:98 and SEQ ID NO:99 and in that the second polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:96, SEQ ID NO:3 and SEQ ID NO:4.

In a particular aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

(a) at least one moiety capable of specific binding to a target cell antigen comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:201 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:202, and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence of SEQ ID NO:97 and the second polypeptide comprises the amino acid sequence of SEQ ID NO:96.

In another aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

(a) at least one moiety capable of specific binding to a target cell antigen comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:357 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:358, and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:97, SEQ ID NO:98 and SEQ ID NO:99 and in that the second polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:96, SEQ ID NO:3 and SEQ ID NO:4.

In a particular aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

(a) at least one moiety capable of specific binding to a target cell antigen comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:357 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:358, and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond,

wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence of SEQ ID NO:97 and the second polypeptide comprises the amino acid sequence of SEQ ID NO:96.

In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the antigen binding molecule comprises

a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific

binding to a target cell antigen,

a second heavy chain comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker that is fused at its C-terminus by a second peptide linker to a CH1 domain, and a second light chain comprising one ectodomain of

5 said TNF ligand family member or a fragment thereof is fused at its C-terminus by a third peptide linker to a CL domain, and wherein the antigen binding molecule comprises

(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:201 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:202 or

10 a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:357 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:358,

(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:14, SEQ ID NO:108, SEQ ID NO:111 and SEQ ID NO:113, and

15 (iii) a second light chain comprising the amino acid sequence of SEQ ID NO:15, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:112 and SEQ ID NO:114.

In a further particular aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, and wherein the antigen binding molecule comprises

a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific

20 binding to a target cell antigen,

a second heavy chain comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by a second peptide linker to a CL domain, and a second light chain comprising one ectodomain of said TNF ligand family member or a fragment thereof that is fused at its C-terminus by a third peptide

25 linker to a CH1 domain, and wherein the molecule comprises

(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:201 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:202 or

30 a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:357 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:358,

(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:115, SEQ ID NO:117, SEQ ID NO:119 and SEQ ID NO:173, and

35 (iii) a second light chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:116, SEQ ID NO:118, SEQ ID NO:120 and SEQ ID NO:174.

Furthermore, the invention provides a TNF family ligand trimer-containing antigen binding molecule, comprising

(a) at least one moiety capable of specific binding to a target cell antigen, and
(b) a first and a second polypeptide that are linked to each other by a disulfide bond,
wherein the antigen binding molecule is characterized in that the first polypeptide contains a
CH3 domain and the second polypeptide contains a CH3 domain, respectively, and wherein the
5 first polypeptide comprises two ectodomains of a TNF ligand family member or fragments
thereof that are connected to each other and to the C-terminus of the CH3 domain by a peptide
linker and wherein the second polypeptide comprises one ectodomain of said TNF ligand family
member or a fragment thereof connected via a peptide linker to C-terminus of the CH3 domain
of said polypeptide.

10 In a particular aspect, such a TNF family ligand trimer-containing antigen binding
molecule comprises two moieties capable of specific binding to a target cell antigen.

More particular, such TNF family ligand trimer-containing antigen binding molecule
comprises

- 15 (i) a first heavy chain comprising the amino acid sequence of SEQ ID NO:209, a second heavy
chain comprising the amino acid sequence of SEQ ID NO:210, and two light chains comprising
the amino acid sequence of SEQ ID NO:206, or
(ii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:213, a second heavy
chain comprising the amino acid sequence of SEQ ID NO:214, and two light chains comprising
the amino acid sequence of SEQ ID NO:206, or
20 (iii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:309, a second heavy
chain comprising the amino acid sequence of SEQ ID NO:310, and two light chains comprising
the amino acid sequence of SEQ ID NO:279, or
(iv) a first heavy chain comprising the amino acid sequence of SEQ ID NO:313, a second heavy
chain comprising the amino acid sequence of SEQ ID NO:314, and two light chains comprising
25 the amino acid sequence of SEQ ID NO:279.

In a further aspect, the invention relates to a TNF family ligand trimer-containing antigen
binding molecule, selected from the group consisting of:

- a) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ
ID NO:205, a first light chain comprising the amino acid sequence of SEQ ID NO:206, a second
30 heavy chain comprising the amino acid sequence of SEQ ID NO:115 and a second light chain
comprising the amino acid sequence of SEQ ID NO:116;
b) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ
ID NO:205, a first light chain comprising the amino acid sequence of SEQ ID NO:206, a second
heavy chain comprising the amino acid sequence of SEQ ID NO:117 and a second light chain
35 comprising the amino acid sequence of SEQ ID NO:118;

c) a molecule comprising two light chains comprising the amino acid sequence of SEQ ID NO:206, a first heavy chain comprising the amino acid sequence of SEQ ID NO:209 and a second heavy chain comprising the amino acid sequence of SEQ ID NO:210;

5 d) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:205, a first light chain comprising the amino acid sequence of SEQ ID NO:206, a second heavy chain comprising the amino acid sequence of SEQ ID NO:119 and a second light chain comprising the amino acid sequence of SEQ ID NO:120;

10 e) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:205, a first light chain comprising the amino acid sequence of SEQ ID NO:206, a second heavy chain comprising the amino acid sequence of SEQ ID NO:173 and a second light chain comprising the amino acid sequence of SEQ ID NO:174; and

f) a molecule comprising two light chains comprising the amino acid sequence of SEQ ID NO:206, a first heavy chain comprising the amino acid sequence of SEQ ID NO:213 and a second heavy chain comprising the amino acid sequence of SEQ ID NO:214.

15 In particular, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:205, a first light chain comprising the amino acid sequence of SEQ ID NO:206, a second heavy chain comprising the amino acid sequence of SEQ ID NO:119 and a second light chain comprising the amino acid sequence of SEQ ID NO:120.

20 In another aspect, the invention relates to a TNF family ligand trimer-containing antigen binding molecule, selected from the group consisting of:

a) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:357, a first light chain comprising the amino acid sequence of SEQ ID NO:358, a second heavy chain comprising the amino acid sequence of SEQ ID NO:115 and a second light chain
25 comprising the amino acid sequence of SEQ ID NO:116;

b) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:357, a first light chain comprising the amino acid sequence of SEQ ID NO:358, a second heavy chain comprising the amino acid sequence of SEQ ID NO:117 and a second light chain comprising the amino acid sequence of SEQ ID NO:118;

30 c) a molecule comprising two light chains comprising the amino acid sequence of SEQ ID NO:358, a first heavy chain comprising the amino acid sequence of SEQ ID NO:209 and a second heavy chain comprising the amino acid sequence of SEQ ID NO:210;

d) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:357, a first light chain comprising the amino acid sequence of SEQ ID NO:358, a second
35 heavy chain comprising the amino acid sequence of SEQ ID NO:119 and a second light chain comprising the amino acid sequence of SEQ ID NO:120;

e) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:357, a first light chain comprising the amino acid sequence of SEQ ID NO:358, a second heavy chain comprising the amino acid sequence of SEQ ID NO:173 and a second light chain comprising the amino acid sequence of SEQ ID NO:174; and

- 5 f) a molecule comprising two light chains comprising the amino acid sequence of SEQ ID NO:358, a first heavy chain comprising the amino acid sequence of SEQ ID NO:213 and a second heavy chain comprising the amino acid sequence of SEQ ID NO:214.

In particular, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:357,
10 a first light chain comprising the amino acid sequence of SEQ ID NO:358, a second heavy chain comprising the amino acid sequence of SEQ ID NO:119 and a second light chain comprising the amino acid sequence of SEQ ID NO:120.

TNF family ligand trimer-containing antigen binding molecules, wherein the target cell antigen is CEA

- 15 In a particular aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the target cell antigen is CEA.

In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule, wherein the moiety capable of specific binding to CD19 comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:321, (ii)
20 CDR-H2 comprising the amino acid sequence of SEQ ID NO:322, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:323, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:324, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:325, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:326.

- 25 In a particular aspect, provided is a TNF family ligand trimer-containing antigen binding molecule of the invention, wherein the moiety capable of specific binding to a target cell antigen is a Fab molecule capable of specific binding to CEA and comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:321, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:322 and (iii) CDR-H3 comprising the amino acid sequence
30 of SEQ ID NO:323, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:324, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:325 and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:326.

In a further aspect, the moiety capable of specific binding to CEA comprises a heavy chain variable region comprising an amino acid sequence that is at least about 95%, 96%, 97%, 98%, 99% or 100% identical to the amino acid sequence of SEQ ID NO:327 and a light chain variable region comprising an amino acid sequence that is at least about 95%, 96%, 97%, 98%, 99% or 100% identical to the amino acid sequence of SEQ ID NO:328.

In one aspect, the moiety capable of specific binding to CEA comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:327 and a variable light chain comprising an amino acid sequence of SEQ ID NO:328.

In a further aspect, the moiety capable of specific binding to CEA comprises a heavy chain variable region comprising an amino acid sequence that is at least about 95%, 96%, 97%, 98%, 99% or 100% identical to the amino acid sequence of SEQ ID NO:329 and a light chain variable region comprising an amino acid sequence that is at least about 95%, 96%, 97%, 98%, 99% or 100% identical to the amino acid sequence of SEQ ID NO:330.

In one aspect, the moiety capable of specific binding to CEA comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:329 and a variable light chain comprising an amino acid sequence of SEQ ID NO:330.

In another aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

- (a) at least one moiety capable of specific binding to a target cell antigen comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:329 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:330, and
- (b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:97, SEQ ID NO:98 and SEQ ID NO:99 and in that the second polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:96, SEQ ID NO:3 and SEQ ID NO:4.

In a particular aspect, the TNF family ligand trimer-containing antigen binding molecule of the invention comprises

- (a) at least one moiety capable of specific binding to a target cell antigen comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:329 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:330, and
- (b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises the

the amino acid sequence of SEQ ID NO:97 and the second polypeptide comprises the amino acid sequence of SEQ ID NO:96.

In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule, wherein the antigen binding molecule comprises

- 5 a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific binding to a target cell antigen,
- a second heavy chain comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker that is fused at its C-terminus by a second peptide linker to a CH1 domain, and a second light chain comprising one ectodomain of
- 10 said TNF ligand family member or a fragment thereof is fused at its C-terminus by a third peptide linker to a CL domain, and wherein the antigen binding molecule comprises
- (i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:329 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:330,
- 15 (ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:14, SEQ ID NO:108, SEQ ID NO:111 and SEQ ID NO:113, and
- (iii) a second light chain comprising the amino acid sequence of SEQ ID NO:15, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:112 and SEQ ID NO:114.

In a further particular aspect, provided is a TNF family ligand trimer-containing antigen

- 20 binding molecule, and wherein the antigen binding molecule comprises
- a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific binding to a target cell antigen,
- a second heavy chain comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by a second
- 25 peptide linker to a CL domain, and a second light chain comprising one ectodomain of said TNF ligand family member or a fragment thereof that is fused at its C-terminus by a third peptide linker to a CH1 domain, and wherein the molecule comprises
- i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:329 and a first light chain comprising the VL domain comprising the amino acid sequence of
- 30 SEQ ID NO:330,
- (ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:115, SEQ ID NO:117, SEQ ID NO:119 and SEQ ID NO:173, and
- (iii) a second light chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:116, SEQ ID NO:118, SEQ ID NO:120 and SEQ ID NO:174.

- 35 Furthermore, the invention provides a TNF family ligand trimer-containing antigen binding molecule, comprising

(a) at least one moiety capable of specific binding to a target cell antigen, and
(b) a first and a second polypeptide that are linked to each other by a disulfide bond,
wherein the antigen binding molecule is characterized in that the first polypeptide contains a
CH3 domain and the second polypeptide contains a CH3 domain, respectively, and wherein the
5 first polypeptide comprises two ectodomains of a TNF ligand family member or fragments
thereof that are connected to each other and to the C-terminus of the CH3 domain by a peptide
linker and wherein the second polypeptide comprises one ectodomain of said TNF ligand family
member or a fragment thereof connected via a peptide linker to C-terminus of the CH3 domain
of said polypeptide.

10 In a particular aspect, such a TNF family ligand trimer-containing antigen binding
molecule comprises two moieties capable of specific binding to a target cell antigen.

More particular, such TNF family ligand trimer-containing antigen binding molecule
comprises

(i) a first heavy chain comprising the amino acid sequence of SEQ ID NO:337, a second heavy
15 chain comprising the amino acid sequence of SEQ ID NO:338, and two light chains comprising
the amino acid sequence of SEQ ID NO:334, or
(ii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:341, a second heavy
chain comprising the amino acid sequence of SEQ ID NO:342, and two light chains comprising
the amino acid sequence of SEQ ID NO:334.

20 In a further aspect, the invention relates to a TNF family ligand trimer-containing antigen
binding molecule, selected from the group consisting of:

a) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ
ID NO:333, a first light chain comprising the amino acid sequence of SEQ ID NO:334, a second
heavy chain comprising the amino acid sequence of SEQ ID NO:115 and a second light chain
25 comprising the amino acid sequence of SEQ ID NO:116;

b) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ
ID NO:333, a first light chain comprising the amino acid sequence of SEQ ID NO:334, a second
heavy chain comprising the amino acid sequence of SEQ ID NO:117 and a second light chain
comprising the amino acid sequence of SEQ ID NO:118;

30 c) a molecule comprising two light chains comprising the amino acid sequence of SEQ ID
NO:334, a first heavy chain comprising the amino acid sequence of SEQ ID NO:337 and a
second heavy chain comprising the amino acid sequence of SEQ ID NO:338;

d) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ
ID NO:333, a first light chain comprising the amino acid sequence of SEQ ID NO:334, a second
35 heavy chain comprising the amino acid sequence of SEQ ID NO:119 and a second light chain
comprising the amino acid sequence of SEQ ID NO:120;

e) a molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:333, a first light chain comprising the amino acid sequence of SEQ ID NO:334, a second heavy chain comprising the amino acid sequence of SEQ ID NO:173 and a second light chain comprising the amino acid sequence of SEQ ID NO:174; and

- 5 f) a molecule comprising two light chains comprising the amino acid sequence of SEQ ID NO:334, a first heavy chain comprising the amino acid sequence of SEQ ID NO:341 and a second heavy chain comprising the amino acid sequence of SEQ ID NO:342.

In particular, the invention provides a TNF family ligand trimer-containing antigen binding molecule comprising a first heavy chain comprising the amino acid sequence of SEQ ID NO:333,
10 a first light chain comprising the amino acid sequence of SEQ ID NO:334, a second heavy chain comprising the amino acid sequence of SEQ ID NO:119 and a second light chain comprising the amino acid sequence of SEQ ID NO:120.

Polynucleotides

The invention further provides isolated polynucleotides encoding a TNF family ligand
15 trimer-containing antigen binding molecule as described herein or a fragment thereof.

The isolated polynucleotides encoding TNF ligand trimer-containing antigen binding molecules of the invention may be expressed as a single polynucleotide that encodes the entire antigen binding molecule or as multiple (e.g., two or more) polynucleotides that are co-expressed. Polypeptides encoded by polynucleotides that are co-expressed may associate
20 through, e.g., disulfide bonds or other means to form a functional antigen binding molecule. For example, the light chain portion of an immunoglobulin may be encoded by a separate polynucleotide from the heavy chain portion of the immunoglobulin. When co-expressed, the heavy chain polypeptides will associate with the light chain polypeptides to form the immunoglobulin.

25 In some aspects, the isolated polynucleotide encodes the entire TNF family ligand trimer-containing antigen binding molecule according to the invention as described herein. In particular, the isolated polynucleotide encodes a polypeptide comprised in the TNF family ligand trimer-containing antigen binding molecule according to the invention as described herein.

30 In one aspect, the present invention is directed to an isolated polynucleotide encoding a TNF family ligand trimer-containing antigen binding molecule, wherein the polynucleotide comprises (a) a sequence that encodes a moiety capable of specific binding to a target cell antigen, (b) a sequence that encodes a polypeptide comprising two ectodomains of a TNF ligand

family member or two fragments thereof that are connected to each other by a peptide linker and (c) a sequence that encodes a polypeptide comprising one ectodomain of said TNF ligand family member or a fragment thereof.

In another aspect, provided is an isolated polynucleotide encoding a 4-1BB ligand trimer-
5 containing antigen binding molecule, wherein the polynucleotide comprises (a) a sequence that encodes a moiety capable of specific binding to a target cell antigen, (b) a sequence that encodes a polypeptide comprising two ectodomains of 4-1BBL or two fragments thereof that are connected to each other by a peptide linker and (c) a sequence that encodes a polypeptide comprising one ectodomain of 4-1BBL or a fragment thereof.

10 In a further aspect, the invention is directed to an isolated polynucleotide comprising a sequence that encodes a polypeptide comprising two 4-1BBL fragments comprising an amino acid sequence that is at least about 90%, 95%, 98% or 100% identical to an amino acid sequence shown in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4 or SEQ ID NO:96, and to a polynucleotide comprising a sequence that encodes a polypeptide comprising one 4-1BBL
15 fragment comprising an amino acid sequence that is is at least about 90%, 95%, 98% or 100% identical to an amino acid sequence shown in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4 or SEQ ID NO:96.

Furthermore, provided is an isolated polynucleotide encoding a OX40 ligand trimer-
20 containing antigen binding molecule, wherein the polynucleotide comprises (a) a sequence that encodes a moiety capable of specific binding to a target cell antigen, (b) a sequence that encodes a polypeptide comprising two ectodomains of OX40L or two fragments thereof that are connected to each other by a peptide linker and (c) a sequence that encodes a polypeptide comprising one ectodomain of OX40L or a fragment thereof.

In another aspect, the invention is directed to an isolated polynucleotide comprising a
25 sequence that encodes a polypeptide comprising two 4-1BBL fragments comprising an amino acid sequence that is at least about 90%, 95%, 98% or 100% identical to an amino acid sequence shown in SEQ ID NO:53 or SEQ ID NO:54, and to a polynucleotide comprising a sequence that encodes a polypeptide comprising one 4-1BBL fragment comprising an amino acid sequence that is is at least about 90%, 95%, 98% or 100% identical to an amino acid sequence shown in
30 SEQ ID NO:53 or SEQ ID NO:54.

In further aspects, the invention relates to the polynucleotides comprising a sequence that is at least about 90%, 95%, 98% or 100% identical to the specific cDNA sequences disclosed herein. In a particular aspect, the invention relates to a polynucleotide comprising a sequence that is identical to one of the specific cDNA sequences disclosed herein.

In other aspects, the nucleic acid molecule comprises or consists of a nucleotide sequence that encodes an amino acid sequence as set forth in any one of SEQ ID NOs: 5, 6, 97, 98, 99, 183, 184 or 185. In a further aspect, the nucleic acid molecule comprises or consists of a nucleotide sequence that encodes an amino acid sequence as set forth in any one of SEQ ID NOs:14, 15, 108, 109, 110, 111, 112, 113, 114, 115,116, 117, 118, 119, 120, 173 or 174.

In still other aspects, the nucleic acid molecule comprises or consists of a nucleotide sequence selected from the group consisting of SEQ ID NOs: 66, 67, 68, 69, 129, 130, 131, 132, 133, 134, 137, 138, 141, 142, 143, 144, 146, 147, 150, 151, 152, 153, 162, 163, 165, 166, 167, 168, 169, 170, 171, 172, 175, 176, 177, 178, 203, 204, 207, 208, 211, 212, 215, 216, 273, 274, 277, 278, 281, 282, 285, 286, 289, 290, 293, 294, 297, 298, 301, 302, 305, 307, 308, 311, 312, 315, 316, 331, 332, 335, 336, 339, 340, 343, 344, 347, 348, 353 or 354.

In certain aspects, the polynucleotide or nucleic acid is DNA. In other embodiments, a polynucleotide of the present invention is RNA, for example, in the form of messenger RNA (mRNA). RNA of the present invention may be single stranded or double stranded.

Recombinant Methods

TNF family ligand trimer-containing antigen binding molecules of the invention may be obtained, for example, by solid-state peptide synthesis (e.g. Merrifield solid phase synthesis) or recombinant production. For recombinant production one or more polynucleotide encoding the TNF family ligand trimer-containing antigen binding molecule or polypeptide fragments thereof, e.g., as described above, is isolated and inserted into one or more vectors for further cloning and/or expression in a host cell. Such polynucleotide may be readily isolated and sequenced using conventional procedures. In one aspect of the invention, a vector, preferably an expression vector, comprising one or more of the polynucleotides of the invention is provided. Methods which are well known to those skilled in the art can be used to construct expression vectors containing the coding sequence of the TNF family ligand trimer-containing antigen binding molecule (fragment) along with appropriate transcriptional/translational control signals. These methods include in vitro recombinant DNA techniques, synthetic techniques and in vivo recombination/genetic recombination. See, for example, the techniques described in Maniatis et al., MOLECULAR CLONING: A LABORATORY MANUAL, Cold Spring Harbor Laboratory, N.Y. (1989); and Ausubel et al., CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, Greene Publishing Associates and Wiley Interscience, N.Y. (1989). The expression vector can be part of a plasmid, virus, or may be a nucleic acid fragment. The expression vector includes an expression cassette into which the polynucleotide encoding the TNF family ligand trimer-containing antigen binding molecule or polypeptide fragments thereof (i.e. the coding region) is cloned in operable association with a promoter and/or other transcription or translation control

elements. As used herein, a "coding region" is a portion of nucleic acid which consists of codons translated into amino acids. Although a "stop codon" (TAG, TGA, or TAA) is not translated into an amino acid, it may be considered to be part of a coding region, if present, but any flanking sequences, for example promoters, ribosome binding sites, transcriptional terminators, introns, 5' and 3' untranslated regions, and the like, are not part of a coding region. Two or more coding regions can be present in a single polynucleotide construct, e.g. on a single vector, or in separate polynucleotide constructs, e.g. on separate (different) vectors. Furthermore, any vector may contain a single coding region, or may comprise two or more coding regions, e.g. a vector of the present invention may encode one or more polypeptides, which are post- or co-translationally separated into the final proteins via proteolytic cleavage. In addition, a vector, polynucleotide, or nucleic acid of the invention may encode heterologous coding regions, either fused or unfused to a polynucleotide encoding the TNF family ligand trimer-containing antigen binding molecule of the invention or polypeptide fragments thereof, or variants or derivatives thereof. Heterologous coding regions include without limitation specialized elements or motifs, such as a secretory signal peptide or a heterologous functional domain. An operable association is when a coding region for a gene product, e.g. a polypeptide, is associated with one or more regulatory sequences in such a way as to place expression of the gene product under the influence or control of the regulatory sequence(s). Two DNA fragments (such as a polypeptide coding region and a promoter associated therewith) are "operably associated" if induction of promoter function results in the transcription of mRNA encoding the desired gene product and if the nature of the linkage between the two DNA fragments does not interfere with the ability of the expression regulatory sequences to direct the expression of the gene product or interfere with the ability of the DNA template to be transcribed. Thus, a promoter region would be operably associated with a nucleic acid encoding a polypeptide if the promoter was capable of effecting transcription of that nucleic acid. The promoter may be a cell-specific promoter that directs substantial transcription of the DNA only in predetermined cells. Other transcription control elements, besides a promoter, for example enhancers, operators, repressors, and transcription termination signals, can be operably associated with the polynucleotide to direct cell-specific transcription.

Suitable promoters and other transcription control regions are disclosed herein. A variety of transcription control regions are known to those skilled in the art. These include, without limitation, transcription control regions, which function in vertebrate cells, such as, but not limited to, promoter and enhancer segments from cytomegaloviruses (e.g. the immediate early promoter, in conjunction with intron-A), simian virus 40 (e.g. the early promoter), and retroviruses (such as, e.g. Rous sarcoma virus). Other transcription control regions include those derived from vertebrate genes such as actin, heat shock protein, bovine growth hormone and rabbit α -globin, as well as other sequences capable of controlling gene expression in eukaryotic cells. Additional suitable transcription control regions include tissue-specific promoters and

enhancers as well as inducible promoters (e.g. promoters inducible tetracyclins). Similarly, a variety of translation control elements are known to those of ordinary skill in the art. These include, but are not limited to ribosome binding sites, translation initiation and termination codons, and elements derived from viral systems (particularly an internal ribosome entry site, or IRES, also referred to as a CITE sequence). The expression cassette may also include other features such as an origin of replication, and/or chromosome integration elements such as retroviral long terminal repeats (LTRs), or adeno-associated viral (AAV) inverted terminal repeats (ITRs).

Polynucleotide and nucleic acid coding regions of the present invention may be associated with additional coding regions which encode secretory or signal peptides, which direct the secretion of a polypeptide encoded by a polynucleotide of the present invention. For example, if secretion of the TNF family ligand trimer-containing antigen binding molecule or polypeptide fragments thereof is desired, DNA encoding a signal sequence may be placed upstream of the nucleic acid encoding a TNF family ligand trimer-containing antigen binding molecule of the invention or polypeptide fragments thereof. According to the signal hypothesis, proteins secreted by mammalian cells have a signal peptide or secretory leader sequence which is cleaved from the mature protein once export of the growing protein chain across the rough endoplasmic reticulum has been initiated. Those of ordinary skill in the art are aware that polypeptides secreted by vertebrate cells generally have a signal peptide fused to the N-terminus of the polypeptide, which is cleaved from the translated polypeptide to produce a secreted or "mature" form of the polypeptide. In certain embodiments, the native signal peptide, e.g. an immunoglobulin heavy chain or light chain signal peptide is used, or a functional derivative of that sequence that retains the ability to direct the secretion of the polypeptide that is operably associated with it. Alternatively, a heterologous mammalian signal peptide, or a functional derivative thereof, may be used. For example, the wild-type leader sequence may be substituted with the leader sequence of human tissue plasminogen activator (TPA) or mouse β -glucuronidase.

DNA encoding a short protein sequence that could be used to facilitate later purification (e.g. a histidine tag) or assist in labeling the fusion protein may be included within or at the ends of the polynucleotide encoding a TNF family ligand trimer-containing antigen binding molecule of the invention or polypeptide fragments thereof.

In a further aspect of the invention, a host cell comprising one or more polynucleotides of the invention is provided. In certain embodiments a host cell comprising one or more vectors of the invention is provided. The polynucleotides and vectors may incorporate any of the features, singly or in combination, described herein in relation to polynucleotides and vectors, respectively. In one aspect, a host cell comprises (e.g. has been transformed or transfected with) a vector comprising a polynucleotide that encodes (part of) a TNF family ligand trimer-

containing antigen binding molecule of the invention of the invention. As used herein, the term "host cell" refers to any kind of cellular system which can be engineered to generate the fusion proteins of the invention or fragments thereof. Host cells suitable for replicating and for supporting expression of antigen binding molecules are well known in the art. Such cells may be transfected or transduced as appropriate with the particular expression vector and large quantities of vector containing cells can be grown for seeding large scale fermenters to obtain sufficient quantities of the antigen binding molecule for clinical applications. Suitable host cells include prokaryotic microorganisms, such as *E. coli*, or various eukaryotic cells, such as Chinese hamster ovary cells (CHO), insect cells, or the like. For example, polypeptides may be produced in bacteria in particular when glycosylation is not needed. After expression, the polypeptide may be isolated from the bacterial cell paste in a soluble fraction and can be further purified. In addition to prokaryotes, eukaryotic microbes such as filamentous fungi or yeast are suitable cloning or expression hosts for polypeptide-encoding vectors, including fungi and yeast strains whose glycosylation pathways have been "humanized", resulting in the production of a polypeptide with a partially or fully human glycosylation pattern. See Gerngross, *Nat Biotech* 22, 1409-1414 (2004), and Li et al., *Nat Biotech* 24, 210-215 (2006).

Suitable host cells for the expression of (glycosylated) polypeptides are also derived from multicellular organisms (invertebrates and vertebrates). Examples of invertebrate cells include plant and insect cells. Numerous baculoviral strains have been identified which may be used in conjunction with insect cells, particularly for transfection of *Spodoptera frugiperda* cells. Plant cell cultures can also be utilized as hosts. See e.g. US Patent Nos. 5,959,177, 6,040,498, 6,420,548, 7,125,978, and 6,417,429 (describing PLANTIBODIESTM technology for producing antibodies in transgenic plants). Vertebrate cells may also be used as hosts. For example, mammalian cell lines that are adapted to grow in suspension may be useful. Other examples of useful mammalian host cell lines are monkey kidney CV1 line transformed by SV40 (COS-7); human embryonic kidney line (293 or 293T cells as described, e.g., in Graham et al., *J Gen Virol* 36, 59 (1977)), baby hamster kidney cells (BHK), mouse sertoli cells (TM4 cells as described, e.g., in Mather, *Biol Reprod* 23, 243-251 (1980)), monkey kidney cells (CV1), African green monkey kidney cells (VERO-76), human cervical carcinoma cells (HELA), canine kidney cells (MDCK), buffalo rat liver cells (BRL 3A), human lung cells (W138), human liver cells (Hep G2), mouse mammary tumor cells (MMT 060562), TRI cells (as described, e.g., in Mather et al., *Annals N.Y. Acad Sci* 383, 44-68 (1982)), MRC 5 cells, and FS4 cells. Other useful mammalian host cell lines include Chinese hamster ovary (CHO) cells, including dhfr- CHO cells (Urlaub et al., *Proc Natl Acad Sci USA* 77, 4216 (1980)); and myeloma cell lines such as YO, NS0, P3X63 and Sp2/0. For a review of certain mammalian host cell lines suitable for protein production, see, e.g., Yazaki and Wu, *Methods in Molecular Biology*, Vol. 248 (B.K.C. Lo, ed., Humana Press, Totowa, NJ), pp. 255-268 (2003). Host cells include cultured cells, e.g., mammalian cultured

cells, yeast cells, insect cells, bacterial cells and plant cells, to name only a few, but also cells comprised within a transgenic animal, transgenic plant or cultured plant or animal tissue. In one embodiment, the host cell is a eukaryotic cell, preferably a mammalian cell, such as a Chinese Hamster Ovary (CHO) cell, a human embryonic kidney (HEK) cell or a lymphoid cell (e.g., Y0,
5 NS0, Sp20 cell). Standard technologies are known in the art to express foreign genes in these systems. Cells expressing a polypeptide comprising either the heavy or the light chain of an immunoglobulin, may be engineered so as to also express the other of the immunoglobulin chains such that the expressed product is an immunoglobulin that has both a heavy and a light chain.

10 In one aspect, a method of producing a TNF family ligand trimer-containing antigen binding molecule of the invention or polypeptide fragments thereof is provided, wherein the method comprises culturing a host cell comprising polynucleotides encoding the TNF family ligand trimer-containing antigen binding molecule of the invention or polypeptide fragments thereof, as provided herein, under conditions suitable for expression of the TNF family ligand
15 trimer-containing antigen binding molecule of the invention or polypeptide fragments thereof, and recovering the TNF family ligand trimer-containing antigen binding molecule of the invention or polypeptide fragments thereof from the host cell (or host cell culture medium).

In the TNF family ligand trimer-containing antigen binding molecule of the invention, the components (at least one moiety capable of specific binding to a target cell antigen, one
20 polypeptide comprising two ectodomains of a TNF ligand family member or fragments thereof and a polypeptide comprising one ectodomain of said TNF family ligand family member or a fragment thereof) are not genetically fused to each other. The polypeptides are designed such that its components (two ectodomains of a TNF ligand family member or fragments thereof and other components such as CH or CL) are fused to each other directly or through a linker
25 sequence. The composition and length of the linker may be determined in accordance with methods well known in the art and may be tested for efficacy. Examples of linker sequences between different components of the antigen binding molecules of the invention are found in the sequences provided herein. Additional sequences may also be included to incorporate a cleavage site to separate the individual components of the fusion protein if desired, for example an
30 endopeptidase recognition sequence.

In certain embodiments the moieties capable of specific binding to a target cell antigen (e.g. Fab fragments) forming part of the antigen binding molecule comprise at least an immunoglobulin variable region capable of binding to an antigen. Variable regions can form part of and be derived from naturally or non-naturally occurring antibodies and fragments thereof.
35 Methods to produce polyclonal antibodies and monoclonal antibodies are well known in the art (see e.g. Harlow and Lane, "Antibodies, a laboratory manual", Cold Spring Harbor Laboratory,

1988). Non-naturally occurring antibodies can be constructed using solid phase-peptide synthesis, can be produced recombinantly (e.g. as described in U.S. patent No. 4,186,567) or can be obtained, for example, by screening combinatorial libraries comprising variable heavy chains and variable light chains (see e.g. U.S. Patent. No. 5,969,108 to McCafferty).

5 Any animal species of immunoglobulin can be used in the invention. Non-limiting immunoglobulins useful in the present invention can be of murine, primate, or human origin. If the fusion protein is intended for human use, a chimeric form of immunoglobulin may be used wherein the constant regions of the immunoglobulin are from a human. A humanized or fully human form of the immunoglobulin can also be prepared in accordance with methods well
10 known in the art (see e. g. U.S. Patent No. 5,565,332 to Winter). Humanization may be achieved by various methods including, but not limited to (a) grafting the non-human (e.g., donor antibody) CDRs onto human (e.g. recipient antibody) framework and constant regions with or without retention of critical framework residues (e.g. those that are important for retaining good antigen binding affinity or antibody functions), (b) grafting only the non-human specificity-determining
15 regions (SDRs or a-CDRs; the residues critical for the antibody-antigen interaction) onto human framework and constant regions, or (c) transplanting the entire non-human variable domains, but "cloaking" them with a human-like section by replacement of surface residues. Humanized antibodies and methods of making them are reviewed, e.g., in Almagro and Fransson, *Front Biosci* 13, 1619-1633 (2008), and are further described, e.g., in Riechmann et al., *Nature* 332, 323-329 (1988); Queen et al., *Proc Natl Acad Sci USA* 86, 10029-10033 (1989); US Patent Nos. 5,821,337, 7,527,791, 6,982,321, and 7,087,409; Jones et al., *Nature* 321, 522-525 (1986); Morrison et al., *Proc Natl Acad Sci* 81, 6851-6855 (1984); Morrison and Oi, *Adv Immunol* 44, 65-92 (1988); Verhoeven et al., *Science* 239, 1534-1536 (1988); Padlan, *Molec Immun* 31(3), 169-217 (1994); Kashmiri et al., *Methods* 36, 25-34 (2005) (describing SDR (a-CDR) grafting);
20 Padlan, *Mol Immunol* 28, 489-498 (1991) (describing "resurfacing"); Dall'Acqua et al., *Methods* 36, 43-60 (2005) (describing "FR shuffling"); and Osbourn et al., *Methods* 36, 61-68 (2005) and Klimka et al., *Br J Cancer* 83, 252-260 (2000) (describing the "guided selection" approach to FR shuffling). Particular immunoglobulins according to the invention are human immunoglobulins. Human antibodies and human variable regions can be produced using various techniques known
30 in the art. Human antibodies are described generally in van Dijk and van de Winkel, *Curr Opin Pharmacol* 5, 368-74 (2001) and Lonberg, *Curr Opin Immunol* 20, 450-459 (2008). Human variable regions can form part of and be derived from human monoclonal antibodies made by the hybridoma method (see e.g. *Monoclonal Antibody Production Techniques and Applications*, pp. 51-63 (Marcel Dekker, Inc., New York, 1987)). Human antibodies and human variable regions
35 may also be prepared by administering an immunogen to a transgenic animal that has been modified to produce intact human antibodies or intact antibodies with human variable regions in response to antigenic challenge (see e.g. Lonberg, *Nat Biotech* 23, 1117-1125 (2005)). Human

antibodies and human variable regions may also be generated by isolating Fv clone variable region sequences selected from human-derived phage display libraries (see e.g., Hoogenboom et al. in *Methods in Molecular Biology* 178, 1-37 (O'Brien et al., ed., Human Press, Totowa, NJ, 2001); and McCafferty et al., *Nature* 348, 552-554; Clackson et al., *Nature* 352, 624-628 (1991)).

5 Phage typically display antibody fragments, either as single-chain Fv (scFv) fragments or as Fab fragments.

In certain aspects, the moieties capable of specific binding to a target cell antigen (e.g. Fab fragments) comprised in the antigen binding molecules of the present invention are engineered to have enhanced binding affinity according to, for example, the methods disclosed in PCT
10 publication WO 2012/020006 (see Examples relating to affinity maturation) or U.S. Pat. Appl. Publ. No. 2004/0132066. The ability of the antigen binding molecules of the invention to bind to a specific antigenic determinant can be measured either through an enzyme-linked
15 immunosorbent assay (ELISA) or other techniques familiar to one of skill in the art, e.g. surface plasmon resonance technique (Liljeblad, et al., *Glyco J* 17, 323-329 (2000)), and traditional
20 binding assays (Heeley, *Endocr Res* 28, 217-229 (2002)). Competition assays may be used to identify an antigen binding molecule that competes with a reference antibody for binding to a particular antigen. In certain embodiments, such a competing antigen binding molecule binds to the same epitope (e.g. a linear or a conformational epitope) that is bound by the reference antigen
25 binding molecule. Detailed exemplary methods for mapping an epitope to which an antigen binding molecule binds are provided in Morris (1996) "Epitope Mapping Protocols", in *Methods in Molecular Biology* vol. 66 (Humana Press, Totowa, NJ). In an exemplary competition assay, immobilized antigen is incubated in a solution comprising a first labeled antigen binding
30 molecule that binds to the antigen and a second unlabeled antigen binding molecule that is being tested for its ability to compete with the first antigen binding molecule for binding to the antigen. The second antigen binding molecule may be present in a hybridoma supernatant. As a control, immobilized antigen is incubated in a solution comprising the first labeled antigen binding
35 molecule but not the second unlabeled antigen binding molecule. After incubation under conditions permissive for binding of the first antibody to the antigen, excess unbound antibody is removed, and the amount of label associated with immobilized antigen is measured. If the
40 amount of label associated with immobilized antigen is substantially reduced in the test sample relative to the control sample, then that indicates that the second antigen binding molecule is competing with the first antigen binding molecule for binding to the antigen. See Harlow and Lane (1988) *Antibodies: A Laboratory Manual* ch.14 (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY).

35 TNF ligand trimer-containing antigen binding molecules of the invention prepared as described herein may be purified by art-known techniques such as high performance liquid chromatography, ion exchange chromatography, gel electrophoresis, affinity chromatography,

size exclusion chromatography, and the like. The actual conditions used to purify a particular protein will depend, in part, on factors such as net charge, hydrophobicity, hydrophilicity etc., and will be apparent to those having skill in the art. For affinity chromatography purification an antibody, ligand, receptor or antigen can be used to which the TNF ligand trimer-containing antigen binding molecule binds. For example, for affinity chromatography purification of fusion proteins of the invention, a matrix with protein A or protein G may be used. Sequential Protein A or G affinity chromatography and size exclusion chromatography can be used to isolate an antigen binding molecule essentially as described in the Examples. The purity of the TNF ligand trimer-containing antigen binding molecule or fragments thereof can be determined by any of a variety of well-known analytical methods including gel electrophoresis, high pressure liquid chromatography, and the like. For example, the TNF ligand trimer-containing antigen binding molecules expressed as described in the Examples were shown to be intact and properly assembled as demonstrated by reducing and non-reducing SDS-PAGE.

Assays

The antigen binding molecules provided herein may be identified, screened for, or characterized for their physical/chemical properties and/or biological activities by various assays known in the art.

1. Affinity assays

The affinity of the TNF family ligand trimer-containing antigen binding molecule provided herein for the corresponding TNF receptor can be determined in accordance with the methods set forth in the Examples by surface plasmon resonance (SPR), using standard instrumentation such as a BIAcore instrument (GE Healthcare), and receptors or target proteins such as may be obtained by recombinant expression. The affinity of the TNF family ligand trimer-containing antigen binding molecule for the target cell antigen can also be determined by surface plasmon resonance (SPR), using standard instrumentation such as a BIAcore instrument (GE Healthcare), and receptors or target proteins such as may be obtained by recombinant expression. A specific illustrative and exemplary embodiment for measuring binding affinity is described in Example 4. According to one aspect, K_D is measured by surface plasmon resonance using a BIACORE® T100 machine (GE Healthcare) at 25 °C.

2. Binding assays and other assays

Binding of the TNF family ligand trimer-containing antigen binding molecule provided herein to the corresponding receptor expressing cells may be evaluated using cell lines expressing the particular receptor or target antigen, for example by flow cytometry (FACS). In one aspect, fresh peripheral blood mononuclear cells (PBMCs) expressing the TNF receptor are

used in the binding assay. These cells are used directly after isolation (naïve PMBCs) or after stimulation (activated PMBCs). In another aspect, activated mouse splenocytes (expressing the TNF receptor molecule) were used to demonstrate the binding of the TNF family ligand trimer-containing antigen binding molecule of the invention to the corresponding TNF receptor
5 expressing cells.

In a further aspect, cancer cell lines expressing the target cell antigen, for example FAP, were used to demonstrate the binding of the antigen binding molecules to the target cell antigen.

In another aspect, competition assays may be used to identify an antigen binding molecule that competes with a specific antibody or antigen binding molecule for binding to the target or
10 TNF receptor, respectively. In certain embodiments, such a competing antigen binding molecule binds to the same epitope (e.g., a linear or a conformational epitope) that is bound by a specific anti-target antibody or a specific anti-TNF receptor antibody. Detailed exemplary methods for mapping an epitope to which an antibody binds are provided in Morris (1996) "Epitope Mapping Protocols," in *Methods in Molecular Biology* vol. 66 (Humana Press, Totowa, NJ).

15 **3. Activity assays**

In one aspect, assays are provided for identifying TNF family ligand trimer-containing antigen binding molecules that bind to a specific target cell antigen and to a specific TNF receptor having biological activity. Biological activity may include, e.g., agonistic signalling through the TNF receptor on cells expressing the target cell antigen. TNF family ligand trimer-
20 containing antigen binding molecules identified by the assays as having such biological activity in vitro are also provided.

In certain aspects, a TNF family ligand trimer-containing antigen binding molecule of the invention is tested for such biological activity. Assays for detecting the biological activity of the molecules of the invention are those described in Example 6. Furthermore, assays for detecting
25 cell lysis (e.g. by measurement of LDH release), induced apoptosis kinetics (e.g. by measurement of Caspase 3/7 activity) or apoptosis (e.g. using the TUNEL assay) are well known in the art. In addition the biological activity of such complexes can be assessed by evaluating their effects on survival, proliferation and lymphokine secretion of various lymphocyte subsets such as NK cells, NKT-cells or $\gamma\delta$ T-cells or assessing their capacity to modulate phenotype and
30 function of antigen presenting cells such as dendritic cells, monocytes/macrophages or B-cells.

Pharmaceutical Compositions, Formulations and Routes of Administration

In a further aspect, the invention provides pharmaceutical compositions comprising any of the TNF family ligand trimer-containing antigen binding molecules provided herein, e.g., for use

in any of the below therapeutic methods. In one embodiment, a pharmaceutical composition comprises any of the TNF family ligand trimer-containing antigen binding molecules provided herein and at least one pharmaceutically acceptable excipient. In another embodiment, a pharmaceutical composition comprises any of the TNF family ligand trimer-containing antigen binding molecules provided herein and at least one additional therapeutic agent, e.g., as described below.

Pharmaceutical compositions of the present invention comprise a therapeutically effective amount of one or more TNF family ligand trimer-containing antigen *binding* molecules dissolved or dispersed in a pharmaceutically acceptable excipient. The phrases "pharmaceutical or pharmacologically acceptable" refers to molecular entities and compositions that are generally non-toxic to recipients at the dosages and concentrations employed, i.e. do not produce an adverse, allergic or other untoward reaction when administered to an animal, such as, for example, a human, as appropriate. The preparation of a pharmaceutical composition that contains at least one TNF family ligand trimer-containing antigen binding molecule and optionally an additional active ingredient will be known to those of skill in the art in light of the present disclosure, as exemplified by Remington's Pharmaceutical Sciences, 18th Ed. Mack Printing Company, 1990, incorporated herein by reference. In particular, the compositions are lyophilized formulations or aqueous solutions. As used herein, "pharmaceutically acceptable excipient" includes any and all solvents, buffers, dispersion media, coatings, surfactants, antioxidants, preservatives (e.g. antibacterial agents, antifungal agents), isotonic agents, salts, stabilizers and combinations thereof, as would be known to one of ordinary skill in the art.

Parenteral compositions include those designed for administration by injection, e.g. subcutaneous, intradermal, intralesional, intravenous, intraarterial intramuscular, intrathecal or intraperitoneal injection. For injection, the TNF family ligand trimer-containing antigen binding molecules of the invention may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hanks' solution, Ringer's solution, or physiological saline buffer. The solution may contain formulatory agents such as suspending, stabilizing and/or dispersing agents. Alternatively, the fusion proteins may be in powder form for constitution with a suitable vehicle, e.g., sterile pyrogen-free water, before use. Sterile injectable solutions are prepared by incorporating the fusion proteins of the invention in the required amount in the appropriate solvent with various of the other ingredients enumerated below, as required. Sterility may be readily accomplished, e.g., by filtration through sterile filtration membranes. Generally, dispersions are prepared by incorporating the various sterilized active ingredients into a sterile vehicle which contains the basic dispersion medium and/or the other ingredients. In the case of sterile powders for the preparation of sterile injectable solutions, suspensions or emulsion, the preferred methods of preparation are vacuum-drying or freeze-drying techniques which yield a powder of the active ingredient plus any additional desired ingredient from a previously sterile-

filtered liquid medium thereof. The liquid medium should be suitably buffered if necessary and the liquid diluent first rendered isotonic prior to injection with sufficient saline or glucose. The composition must be stable under the conditions of manufacture and storage, and preserved against the contaminating action of microorganisms, such as bacteria and fungi. It will be appreciated that endotoxin contamination should be kept minimally at a safe level, for example, less than 0.5 ng/mg protein. Suitable pharmaceutically acceptable excipients include, but are not limited to: buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid and methionine; preservatives (such as octadecyldimethylbenzyl ammonium chloride; hexamethonium chloride; benzalkonium chloride; benzethonium chloride; phenol, butyl or benzyl alcohol; alkyl parabens such as methyl or propyl paraben; catechol; resorcinol; cyclohexanol; 3-pentanol; and m-cresol); low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, histidine, arginine, or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrans; chelating agents such as EDTA; sugars such as sucrose, mannitol, trehalose or sorbitol; salt-forming counter-ions such as sodium; metal complexes (e.g. Zn-protein complexes); and/or non-ionic surfactants such as polyethylene glycol (PEG). Aqueous injection suspensions may contain compounds which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, dextran, or the like. Optionally, the suspension may also contain suitable stabilizers or agents which increase the solubility of the compounds to allow for the preparation of highly concentrated solutions. Additionally, suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes.

Active ingredients may be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization, for example, hydroxymethylcellulose or gelatin-microcapsules and poly-(methylmethacrylate) microcapsules, respectively, in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nanoparticles and nanocapsules) or in macroemulsions. Such techniques are disclosed in Remington's Pharmaceutical Sciences (18th Ed. Mack Printing Company, 1990). Sustained-release preparations may be prepared. Suitable examples of sustained-release preparations include semipermeable matrices of solid hydrophobic polymers containing the polypeptide, which matrices are in the form of shaped articles, e.g. films, or microcapsules. In particular embodiments, prolonged absorption of an injectable composition can be brought about by the use in the compositions of agents delaying absorption, such as, for example, aluminum monostearate, gelatin or combinations thereof.

Exemplary pharmaceutically acceptable excipients herein further include interstitial drug dispersion agents such as soluble neutral-active hyaluronidase glycoproteins (sHASEGP), for example, human soluble PH-20 hyaluronidase glycoproteins, such as rHuPH20 (HYLENEX®, Baxter International, Inc.). Certain exemplary sHASEGPs and methods of use, including
5 rHuPH20, are described in US Patent Publication Nos. 2005/0260186 and 2006/0104968. In one aspect, a sHASEGP is combined with one or more additional glycosaminoglycanases such as chondroitinases.

Exemplary lyophilized antibody formulations are described in US Patent No. 6,267,958. Aqueous antibody formulations include those described in US Patent No. 6,171,586 and
10 WO2006/044908, the latter formulations including a histidine-acetate buffer.

In addition to the compositions described previously, the fusion proteins may also be formulated as a depot preparation. Such long acting formulations may be administered by implantation (for example subcutaneously or intramuscularly) or by intramuscular injection. Thus, for example, the fusion proteins may be formulated with suitable polymeric or
15 hydrophobic materials (for example as an emulsion in an acceptable oil) or ion exchange resins, or as sparingly soluble derivatives, for example, as a sparingly soluble salt.

Pharmaceutical compositions comprising the fusion proteins of the invention may be manufactured by means of conventional mixing, dissolving, emulsifying, encapsulating, entrapping or lyophilizing processes. Pharmaceutical compositions may be formulated in
20 conventional manner using one or more physiologically acceptable carriers, diluents, excipients or auxiliaries which facilitate processing of the proteins into preparations that can be used pharmaceutically. Proper formulation is dependent upon the route of administration chosen.

The TNF family ligand trimer-containing antigen binding molecules may be formulated into a composition in a free acid or base, neutral or salt form. Pharmaceutically acceptable salts
25 are salts that substantially retain the biological activity of the free acid or base. These include the acid addition salts, e.g. those formed with the free amino groups of a proteinaceous composition, or which are formed with inorganic acids such as for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric or mandelic acid. Salts formed with the free carboxyl groups can also be derived from inorganic bases such as for example, sodium,
30 potassium, ammonium, calcium or ferric hydroxides; or such organic bases as isopropylamine, trimethylamine, histidine or procaine. Pharmaceutical salts tend to be more soluble in aqueous and other protic solvents than are the corresponding free base forms.

The composition herein may also contain more than one active ingredients as necessary for the particular indication being treated, preferably those with complementary activities that do not

adversely affect each other. Such active ingredients are suitably present in combination in amounts that are effective for the purpose intended.

The formulations to be used for in vivo administration are generally sterile. Sterility may be readily accomplished, e.g., by filtration through sterile filtration membranes.

5 **Therapeutic methods and compositions**

Any of the TNF family ligand trimer-containing antigen binding molecules provided herein may be used in therapeutic methods.

For use in therapeutic methods, TNF family ligand trimer-containing antigen binding molecules of the invention can be formulated, dosed, and administered in a fashion consistent with good medical practice. Factors for consideration in this context include the particular disorder being treated, the particular mammal being treated, the clinical condition of the individual patient, the cause of the disorder, the site of delivery of the agent, the method of administration, the scheduling of administration, and other factors known to medical practitioners.

In one aspect, TNF family ligand trimer-containing antigen binding molecules of the invention for use as a medicament are provided. In further aspects, TNF family ligand trimer-containing antigen binding molecules of the invention for use in treating a disease, in particular for use in the treatment of cancer, are provided. In certain aspects, TNF family ligand trimer-containing antigen binding molecules of the invention for use in a method of treatment are provided. In one aspect, the invention provides a TNF family ligand trimer-containing antigen binding molecule as described herein for use in the treatment of a disease in an individual in need thereof. In certain aspects, the invention provides a TNF family ligand trimer-containing antigen binding molecule for use in a method of treating an individual having a disease comprising administering to the individual a therapeutically effective amount of the fusion protein. In certain aspects, the disease to be treated is cancer. Examples of cancers include solid tumors, bladder cancer, renal cell carcinoma, brain cancer, head and neck cancer, pancreatic cancer, lung cancer, breast cancer, ovarian cancer, uterine cancer, cervical cancer, endometrial cancer, esophageal cancer, colon cancer, colorectal cancer, rectal cancer, gastric cancer, prostate cancer, blood cancer, skin cancer, squamous cell carcinoma, bone cancer, and kidney cancer, melanoma, B-cell lymphoma, B-cell leukemia, non-Hodgkin lymphoma and acute lymphoblastic leukemia. Thus, a TNF family ligand trimer-containing antigen binding molecule as described herein for use in the treatment of cancer is provided. The subject, patient, or "individual" in need of treatment is typically a mammal, more specifically a human.

In another aspect, provided is a TNF family ligand trimer-containing antigen binding molecule as described herein for use in the treatment of infectious diseases, in particular for the treatment of viral infections. In a further aspect, provided is a TNF family ligand trimer-containing antigen binding molecule as described herein for use in the treatment of autoimmune diseases such as for example Lupus disease.

In one aspect, provided is a TNF family ligand trimer-containing antigen binding molecule according to the invention for use in treating head and neck squamous cell carcinoma (HNSCC), breast cancer, colorectal cancer (CRC), pancreatic cancer (PAC), gastric cancer, non-small-cell lung carcinoma (NSCLC) and Mesothelioma, wherein the target cell antigen is FAP.

In a further aspect, the invention relates to the use of a TNF family ligand trimer-containing antigen binding molecule in the manufacture or preparation of a medicament for the treatment of a disease in an individual in need thereof. In one aspect, the medicament is for use in a method of treating a disease comprising administering to an individual having the disease a therapeutically effective amount of the medicament. In certain embodiments the disease to be treated is a proliferative disorder, particularly cancer. Thus, in one aspect, the invention relates to the use of a TNF family ligand trimer-containing antigen binding molecule of the invention in the manufacture or preparation of a medicament for the treatment of cancer. Examples of cancers include solid tumors, bladder cancer, renal cell carcinoma, brain cancer, head and neck cancer, pancreatic cancer, lung cancer, breast cancer, ovarian cancer, uterine cancer, cervical cancer, endometrial cancer, esophageal cancer, colon cancer, colorectal cancer, rectal cancer, gastric cancer, prostate cancer, blood cancer, skin cancer, squamous cell carcinoma, bone cancer, and kidney cancer, melanoma, B-cell lymphoma, B-cell leukemia, non-Hodgkin lymphoma and acute lymphoblastic leukemia.. Other cell proliferation disorders that can be treated using a TNF family ligand trimer-containing antigen binding molecule of the present invention include, but are not limited to neoplasms located in the: abdomen, bone, breast, digestive system, liver, pancreas, peritoneum, endocrine glands (adrenal, parathyroid, pituitary, testicles, ovary, thymus, thyroid), eye, head and neck, nervous system (central and peripheral), lymphatic system, pelvic, skin, soft tissue, spleen, thoracic region, and urogenital system. Also included are pre-cancerous conditions or lesions and cancer metastases. In certain embodiments the cancer is chosen from the group consisting of renal cell cancer, skin cancer, lung cancer, colorectal cancer, breast cancer, brain cancer, head and neck cancer. A skilled artisan may recognize that in some cases the TNF family ligand trimer-containing antigen binding molecule may not provide a cure but may only provide partial benefit. In some aspects, a physiological change having some benefit is also considered therapeutically beneficial. Thus, in some aspects, an amount of TNF family ligand trimer-containing antigen binding molecule that provides a physiological change is considered an "effective amount" or a "therapeutically effective amount".

In a further aspect, the invention relates to the use of a TNF family ligand trimer-containing antigen binding molecule as described herein in the manufacture or preparation of a medicament for the treatment of infectious diseases, in particular for the treatment of viral infections or for the treatment of autoimmune diseases, for example Lupus disease.

5 In a further aspect, the invention provides a method for treating a disease in an individual, comprising administering to said individual a therapeutically effective amount of a TNF family ligand trimer-containing antigen binding molecule of the invention. In one aspect a composition is administered to said individual, comprising a fusion protein of the invention in a pharmaceutically acceptable form. In certain aspects, the disease to be treated is a proliferative
10 disorder. In a particular aspect, the disease is cancer. In another aspect, the disease is an infectious disease or an autoimmune disease. In certain aspects, the method further comprises administering to the individual a therapeutically effective amount of at least one additional therapeutic agent, e.g. an anti-cancer agent if the disease to be treated is cancer. An “individual” according to any of the above embodiments may be a mammal, preferably a human.

15 For the prevention or treatment of disease, the appropriate dosage of a TNF family ligand trimer-containing antigen binding molecule of the invention (when used alone or in combination with one or more other additional therapeutic agents) will depend on the type of disease to be treated, the route of administration, the body weight of the patient, the type of fusion protein, the severity and course of the disease, whether the fusion protein is administered for preventive or
20 therapeutic purposes, previous or concurrent therapeutic interventions, the patient's clinical history and response to the fusion protein, and the discretion of the attending physician. The practitioner responsible for administration will, in any event, determine the concentration of active ingredient(s) in a composition and appropriate dose(s) for the individual subject. Various dosing schedules including but not limited to single or multiple administrations over various
25 time-points, bolus administration, and pulse infusion are contemplated herein.

The TNF family ligand trimer-containing antigen binding molecule is suitably administered to the patient at one time or over a series of treatments. Depending on the type and severity of the disease, about 1 µg/kg to 15 mg/kg (e.g. 0.1 mg/kg – 10 mg/kg) of TNF family ligand trimer-containing antigen binding molecule can be an initial candidate dosage for
30 administration to the patient, whether, for example, by one or more separate administrations, or by continuous infusion. One typical daily dosage might range from about 1 µg/kg to 100 mg/kg or more, depending on the factors mentioned above. For repeated administrations over several days or longer, depending on the condition, the treatment would generally be sustained until a desired suppression of disease symptoms occurs. One exemplary dosage of the fusion protein
35 would be in the range from about 0.005 mg/kg to about 10 mg/kg. In other examples, a dose may also comprise from about 1 µg/kg body weight, about 5 µg/kg body weight, about 10 µg/kg body

weight, about 50 µg/kg body weight, about 100 µg/kg body weight, about 200 µg/kg body weight, about 350 µg/kg body weight, about 500 µg/kg body weight, about 1 mg/kg body weight, about 5 mg/kg body weight, about 10 mg/kg body weight, about 50 mg/kg body weight, about 100 mg/kg body weight, about 200 mg/kg body weight, about 350 mg/kg body weight, about 500 mg/kg body weight, to about 1000 mg/kg body weight or more per administration, and any range derivable therein. In examples of a derivable range from the numbers listed herein, a range of about 5 mg/kg body weight to about 100 mg/kg body weight, about 5 µg/kg body weight to about 500 mg/kg body weight etc., can be administered, based on the numbers described above. Thus, one or more doses of about 0.5 mg/kg, 2.0 mg/kg, 5.0 mg/kg or 10 mg/kg (or any combination thereof) may be administered to the patient. Such doses may be administered intermittently, e.g. every week or every three weeks (e.g. such that the patient receives from about two to about twenty, or e.g. about six doses of the fusion protein). An initial higher loading dose, followed by one or more lower doses may be administered. However, other dosage regimens may be useful. The progress of this therapy is easily monitored by conventional techniques and assays.

The TNF family ligand trimer-containing antigen binding molecules of the invention will generally be used in an amount effective to achieve the intended purpose. For use to treat or prevent a disease condition, the TNF family ligand trimer-containing antigen binding molecules of the invention, or pharmaceutical compositions thereof, are administered or applied in a therapeutically effective amount. Determination of a therapeutically effective amount is well within the capabilities of those skilled in the art, especially in light of the detailed disclosure provided herein.

For systemic administration, a therapeutically effective dose can be estimated initially from in vitro assays, such as cell culture assays. A dose can then be formulated in animal models to achieve a circulating concentration range that includes the IC₅₀ as determined in cell culture. Such information can be used to more accurately determine useful doses in humans.

Initial dosages can also be estimated from in vivo data, e.g., animal models, using techniques that are well known in the art. One having ordinary skill in the art could readily optimize administration to humans based on animal data.

Dosage amount and interval may be adjusted individually to provide plasma levels of the TNF family ligand trimer-containing antigen binding molecules which are sufficient to maintain therapeutic effect. Usual patient dosages for administration by injection range from about 0.1 to 50 mg/kg/day, typically from about 0.5 to 1 mg/kg/day. Therapeutically effective plasma levels may be achieved by administering multiple doses each day. Levels in plasma may be measured, for example, by HPLC.

In cases of local administration or selective uptake, the effective local concentration of the TNF family ligand trimer-containing antigen binding molecule may not be related to plasma concentration. One skilled in the art will be able to optimize therapeutically effective local dosages without undue experimentation.

5 A therapeutically effective dose of the TNF family ligand trimer-containing antigen binding molecules described herein will generally provide therapeutic benefit without causing substantial toxicity. Toxicity and therapeutic efficacy of a fusion protein can be determined by standard pharmaceutical procedures in cell culture or experimental animals. Cell culture assays and animal studies can be used to determine the LD₅₀ (the dose lethal to 50% of a population)
10 and the ED₅₀ (the dose therapeutically effective in 50% of a population). The dose ratio between toxic and therapeutic effects is the therapeutic index, which can be expressed as the ratio LD₅₀/ED₅₀. TNF family ligand trimer-containing antigen binding molecules that exhibit large therapeutic indices are preferred. In one embodiment, the TNF family ligand trimer-containing antigen binding molecule according to the present invention exhibits a high therapeutic index.
15 The data obtained from cell culture assays and animal studies can be used in formulating a range of dosages suitable for use in humans. The dosage lies preferably within a range of circulating concentrations that include the ED₅₀ with little or no toxicity. The dosage may vary within this range depending upon a variety of factors, e.g., the dosage form employed, the route of administration utilized, the condition of the subject, and the like. The exact formulation, route of
20 administration and dosage can be chosen by the individual physician in view of the patient's condition (see, e.g., Fingl et al., 1975, in: The Pharmacological Basis of Therapeutics, Ch. 1, p. 1, incorporated herein by reference in its entirety).

The attending physician for patients treated with fusion proteins of the invention would know how and when to terminate, interrupt, or adjust administration due to toxicity, organ
25 dysfunction, and the like. Conversely, the attending physician would also know to adjust treatment to higher levels if the clinical response were not adequate (precluding toxicity). The magnitude of an administered dose in the management of the disorder of interest will vary with the severity of the condition to be treated, with the route of administration, and the like. The severity of the condition may, for example, be evaluated, in part, by standard prognostic
30 evaluation methods. Further, the dose and perhaps dose frequency will also vary according to the age, body weight, and response of the individual patient.

Other agents and treatments

The TNF family ligand trimer-containing antigen binding molecules of the invention may be administered in combination with one or more other agents in therapy. For instance, a fusion
35 protein of the invention may be co-administered with at least one additional therapeutic agent.

The term “therapeutic agent” encompasses any agent that can be administered for treating a symptom or disease in an individual in need of such treatment. Such additional therapeutic agent may comprise any active ingredients suitable for the particular indication being treated, preferably those with complementary activities that do not adversely affect each other. In certain
5 embodiments, an additional therapeutic agent is another anti-cancer agent.

Such other agents are suitably present in combination in amounts that are effective for the purpose intended. The effective amount of such other agents depends on the amount of fusion protein used, the type of disorder or treatment, and other factors discussed above. The TNF family ligand trimer-containing antigen binding molecules are generally used in the same
10 dosages and with administration routes as described herein, or about from 1 to 99% of the dosages described herein, or in any dosage and by any route that is empirically/clinically determined to be appropriate.

Such combination therapies noted above encompass combined administration (where two or more therapeutic agents are included in the same or separate compositions), and separate
15 administration, in which case, administration of the TNF family ligand trimer-containing antigen binding molecule of the invention can occur prior to, simultaneously, and/or following, administration of the additional therapeutic agent and/or adjuvant.

Articles of Manufacture

In another aspect of the invention, an article of manufacture containing materials useful for
20 the treatment, prevention and/or diagnosis of the disorders described above is provided. The article of manufacture comprises a container and a label or package insert on or associated with the container. Suitable containers include, for example, bottles, vials, syringes, IV solution bags, etc. The containers may be formed from a variety of materials such as glass or plastic. The container holds a composition which is by itself or combined with another composition effective
25 for treating, preventing and/or diagnosing the condition and may have a sterile access port (for example the container may be an intravenous solution bag or a vial having a stopper that is pierceable by a hypodermic injection needle). At least one active agent in the composition is a TNF ligand trimer-containing antigen binding molecule of the invention.

The label or package insert indicates that the composition is used for treating the condition
30 of choice. Moreover, the article of manufacture may comprise (a) a first container with a composition contained therein, wherein the composition comprises a TNF ligand trimer-containing antigen binding molecule of the invention; and (b) a second container with a composition contained therein, wherein the composition comprises a further cytotoxic or otherwise therapeutic agent. The article of manufacture in this embodiment of the invention may

further comprise a package insert indicating that the compositions can be used to treat a particular condition.

Alternatively, or additionally, the article of manufacture may further comprise a second (or third) container comprising a pharmaceutically-acceptable buffer, such as bacteriostatic water for injection (BWFI), phosphate-buffered saline, Ringer's solution and dextrose solution. It may further include other materials desirable from a commercial and user standpoint, including other buffers, diluents, filters, needles, and syringes.

Table C (Sequences):

SEQ ID NO:	Name	Sequence
1	Human (hu) 4-1BBL (71-254)	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVY YVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLG VHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLP SPRSE
2	hu 4-1BBL (85-254)	LDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVY YVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLP SPRSE
3	hu 4-1BBL (80-254)	DPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVY YVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLP SPRSE
4	hu 4-1BBL (52-254)	PWAVSGARASPGSAASPRLREGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVY YVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLP SPRSE
5	dimeric hu 4-1BBL (71-254) connected by (G4S) ₂ linker	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVY YVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLG VHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLP SPRSEGGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVY YVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLP SPRSE
6	monomeric hu 4-1BBL (71-254) plus (G4S) ₂ linker	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVY YVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLG

SEQ ID NO:	Name	Sequence
		VHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLP SPRSEGGGGSGGGGS
7	FAP(28H1) CDR-H1	SHAMS
8	FAP(28H1) CDR-H2	AIWASGEQYYADSVKG
9	FAP(28H1) CDR-H3	GWLGNFY
10	FAP(28H1) CDR-L1	RASQSVRSYLA
11	FAP(28H1) CDR-L2	GASTRAT
12	FAP(28H1) CDR-L3	QQGQVIPPT
13	(G4S) ₂	GGGGSGGGGS
14	dimeric hu 4-1BBL (71-254)-CH1 Fc knob chain	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLIDGP LSWYSDPGLAGVSLTGGLSYKEDTKELVVAAGVY YVFFQLELRRVAVAGEGSGSVSLALHLQPLRSAAGAA ALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLG VHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLP SPRSEGGGGSGGGGSREGPELSPDDPAGLLDLRQGM FAQLVAQNVLIDGPLSWYSDPGLAGVSLTGGLSYK EDTKELVVAAGVYVFFQLELRRVAVAGEGSGSVSL ALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGF QGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATV LGLFRVTPEIPAGLPSPRSEGGGGSGGGGSASTKGPS VFPLAPSSKSTSGGTAALGCLVKDYFPEPTVSWNSG ALTSQVHTFPAVLQSSGLYSLSVTVPSSSLGTQTYI CNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAA GGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPE VKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLT VLHQDWLNGKEYKCKVSNKALGAPIEKTISKAKGQP REPQVYTLPPCRDELTKNQVSLWCLVKGFYPSDIAV EWESNGQPENNYKTTTPVLDSDGSFLLYSLKLTVDKS RWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK
15	monomeric hu 4-1BBL (71-254)-CL	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLIDGP LSWYSDPGLAGVSLTGGLSYKEDTKELVVAAGVY YVFFQLELRRVAVAGEGSGSVSLALHLQPLRSAAGAA ALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLG VHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLP SPRSEGGGGSGGGGSRTVAAPSVFIFPPSDEQLKSGT ASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVT EQDSKDYSLSTLTLSKADYEKHKVYACEVTHQG LSSPVTKSFNREGEC
16	FAP(28H1) VH	EVQLLESQGGGLVQPGGSLRLSCAASGFTFSSHA MSWVRQAPGKGLEWVSAIWASGEQYYADSVK GRFTISRDNKNTLYLQMNSLRAEDTAVYYCAK GWLGNFYWGQGTLLTVSS
17	FAP(28H1) VL	EIVLTQSPGTLSPGERATLSCRASQSVRSYLA WYQQKPGQAPRLLIIGASTRATGIPDRFSGSGSG TDFTLTISRLEPEDFAVYYCQQGQVIPPTFGQGT KVEIK
18	anti-FAP(28H1) Fc hole chain	see Table 2
19	anti-FAP(28H1) light chain	see Table 2
20	Human (hu) FAP	UniProt no. Q12884

SEQ ID NO:	Name	Sequence
21	hu FAP ectodomain+poly-lys-tag+his ₆ -tag	RPSRVHNSEENTMRALTLKDILNGTFSYKTFFPNWIS GQEYLHQSDNNIVLYNIETGQSYTILSNRTMKSVNA SNYGLSPDRQFVYLESYKLRYSYATYYIYDLS NGEFVRGNELPRPIQYLCWSPVGSKLAYVYQNNIYL KQRPDPPFQITFNGRENKIFNGIPDWVYEEEMLATK YALWWSPNGKFLAYAEFNDTDIPVIAYSYYGDEQYP RTINIPYPKAGAKNPVVRIFIIDTTYPAYVGPQEVVVP AMIASDYYFSWLTWVTDERVCLQWLKRVQNVSVL SICDFREDWQTWDCPKTQEHIEESRTGWAGGFFVST PVFSYDAISYYKIFSDKDGKHIHYIKDTVENAIQITS GKWEAINIFRVTQDSLFISSNEFEEYPGRRNIYRISGS YPPSKKCVTCHLRKERCQYYTASFSDYAKYYALVCY GPGIPISTLHDGRDQEIKILEENKELENALKNIQLPKE EIKKLEVDEITLWYKMILPPQFDRSKKYPLLIQVYGG PCSQSVRSVFAVNWISYLASKEGMVIALVDGRGTAF QGDKLLYAVYRKLGVYEVEDQITAVRKFIEMGFIDE KRIAIWGSYGGYVSSLALASGTGLFKCGIAVAPVSS WEYYASVYTERFMGLPTKDDNLEHYKNSTVMARAE YFRNVDYLLIHGTADDNVHFQNSAQIAKALVNAQV DFQAMWYSDQNHGLSGLSTNHLTYTHMTHFLKQCFS LSDGKKKKKKKGHHHHHH
22	nucleotide sequence hu FAP ectodomain+poly-lys-tag+his ₆ -tag	CGCCCTTCAAGAGTTCATAACTCTGAAGAAAATAC AATGAGAGCACTCACACTGAAGGATATTTTAAATG GAACATTTTCTTATAAAACATTTTTTCCAAACTGGA TTTCAGGACAAGAATATCTTCATCAATCTGCAGAT AACAAATATAGTACTTTATAATATTGAAACAGGACA ATCATATACCATTTTGAGTAATAGAACCATGAAAA GTGTGAATGCTTCAAATTACGGCTTATCACCTGAT CGGCAATTTGTATATCTAGAAAGTGATTATTCAA GCTTTGGAGATACTCTTACACAGCAACATATTACA TCTATGACCTTAGCAATGGAGAATTTGTAAGAGGA AATGAGCTTCCTCGTCCAATTCAGTATTTATGCTGG TCGCTGTGTTGGGAGTAAATTAGCATATGTCTATCA AAACAATATCTATTTGAAACAAAGACCAGGAGAT CCACCTTTTCAAATAACATTTAATGGAAGAGAAAA TAAATATTTAATGGAATCCCAGACTGGGTTTATG AAGAGGAAATGCTTGCTACAAAATATGCTCTCTGG TGGTCTCCTAATGGAAAATTTTGGCATATGCGGA ATTTAATGATACGGATATACCAAGTTATTGCCTATTC CTATTATGGCGATGAACAATATCCTAGAACAAATAA ATATTCCATACCCAAAGGCTGGAGCTAAGAATCCC GTTGTTTCGGATATTTATTATCGATACCACTTACCCT GCGTATGTAGGTCCCCAGGAAGTGCCTGTTCCAGC AATGATAGCCTCAAGTGATTATTATTTCAGTTGGC TCACGTGGGTACTGATGAACGAGTATGTTTGCAG TGGCTAAAAAGAGTCCAGAATGTTTCGGTCTCTGTC TATATGTGACTTCAGGGAAGACTGGCAGACATGGG ATTGTCCAAAGACCCAGGAGCATATAGAAGAAAG CAGAACTGGATGGGCTGGTGGATTCTTTGTTTCAA CACCAGTTTTTCAGCTATGATGCCATTTTCGTAATA AAATATTTAGTGACAAGGATGGCTACAAACATATT CACTATATCAAAGACACTGTGGAAAATGCTATTCA

SEQ ID NO:	Name	Sequence
		AATTACAAGTGGCAAGTGGGAGGCCATAAATATA TTCAGAGTAACACAGGATTCAGTGTATTTATTCTAG CAATGAATTTGAAGAATACCTTGAAGAAGAAAC ATCTACAGAATTAGCATTGGAAGCTATCCTCCAAG CAAGAAGTGTGTTACTTGCCATCTAAGGAAAGAAA GGTGCCAATATTACACAGCAAGTTTCAGCGACTAC GCCAAGTACTATGCACTTGTCTGCTACGGCCCAGG CATCCCCATTTCACCCCTTCATGATGGACGCACTG ATCAAGAAATTAAAATCCTGGAAGAAAACAAGGA ATTGGAAAATGCTTTGAAAAATATCCAGCTGCCTA AAGAGGAAATTAAGAACTTGAAGTAGATGAAAT TACTTTATGGTACAAGATGATTCTTCCTCCTCAATT TGACAGATCAAAGAAGTATCCCTTGCTAATTCAAG TGTATGGTGGTCCCTGCAGTCAGAGTGTAAAGGTCT GTATTTGCTGTTAATTGGATATCTTATCTTGCAAGT AAGGAAGGGATGGTCATTGCCTTGGTGGATGGTCG AGGAACAGCTTTCCAAGGTGACAACTCCTCTATG CAGTGTATCGAAAGCTGGGTGTTTATGAAGTTGAA GACCAGATTACAGCTGTCAGAAAATTCATAGAAAT GGGTTTCATTGATGAAAAAAGAATAGCCATATGGG GCTGGTCCTATGGAGGATACGTTTCATCACTGGCC CTTGCACTCTGGAAGTGGTCTTTTCAAATGTGGTATA GCAGTGGCTCCAGTCTCCAGCTGGGAATATTACGC GTCTGTCTACACAGAGAGATTCATGGGTCTCCCAA CAAAGGATGATAATCTTGAGCACTATAAGAATTCA ACTGTGATGGCAAGAGCAGAATATTTTCAGAAATGT AGACTATCTTCTCATCCACGGAACAGCAGATGATA ATGTGCACTTTCAAACTCAGCACAGATTGCTAAA GCTCTGGTTAATGCACAAGTGGATTTCAGGCAAT GTGGTACTCTGACCAGAACCACGGCTTATCCGGCC TGTCACGAACCACTTATACACCCACATGACCCAC TTCCTAAAGCAGTGTCTTCTTTGTCAGACGGCAA AAAGAAAAAGAAAAAGGGCCACCACCATCACCAT CAC
23	mouse FAP	UniProt no. P97321
24	Murine FAP ectodomain+poly-lys- tag+his ₆ -tag	RPSRVYKPEGNTKRALTLKDILNGTFSYKTYFPNWIS EQEYLHQSEDDNIVFYNIETRESYHLSNSTMKSVNAT DYGLSPDRQFVYLESDYSLWRYSYTATYYIYDLQN GEFVRGYELPRPIQYLCWSPVGSKLAYVYQNNIYLK QRPGDPPFQITYTGRENRIENGIPDWVYEEEMLATKY ALWWSPDGKFLAYVEFNDSDIPIAYSYYGDGQYPR TINIPYPKAGAKNPVVRVFIVDTTYPHVGPMVEVPVP EMIASSDYFWSLTVWSSERVCLQWLKRVQNVSVL SICDFREDWHAWECPKNQEHVEESRTGWAGGFFVST PAFSQDATSYKIFSDKDGKHHYIKDTVENAIQITS GKWEAIYIFRVTQDSLFISSNEFEGYPGRNRIYRISIG NSPPSKKCVTCHLRKERCQYYTASFYSYKAKYYALVC YGPGLPISTLHDGRDQEIQVLEENKELENSLRNIQLP KVEIKKLKDGGLTFWYKMILPPQFDRSKKYPLLIQVY GGPCSQSVKSVFAVNWITYLASKEGIVIALVDGRGTA FQGDKFLHAVYRKLGVEVEDQLTAVRKFIEMGFID EERIAIWGWSYGGYVSSLALASGTGLFKCGIAVAPVS

SEQ ID NO:	Name	Sequence
		SWEYYASIYSERFMGLPTKDDNLEHYKNSTVMARA EYFRNVDYLLIHGTADDNVHFQNSAQIAKALVNAQV DFQAMWYSDQNHGILSGRSQNHLYTHMTHFLKQCF SLSDGKKKKKKGHHHHHH
25	nucleotide sequence Murine FAP ectodomain+poly-lys- tag+his ₆ -tag	CGTCCCTCAAGAGTTTACAAACCTGAAGGAAACAC AAAGAGAGCTCTTACCTTGAAGGATATTTTAAATG GAACATTCTCATATAAAACATATTTTCCCAACTGG ATTCAGAACAAAGAATATCTTCATCAATCTGAGGA TGATAACATAGTATTTTATAATATTGAAACAAGAG AATCATATATCATTTTGAGTAATAGCACCATGAAA AGTGTGAATGCTACAGATTATGGTTTGTACCTGA TCGGCAATTTGTGTATCTAGAAAGTGATTATTCAA AGCTCTGGCGATATTCATACACAGCGACATACTAC ATCTACGACCTTCAGAATGGGGAATTTGTAAGAGG ATACGAGCTCCCTCGTCCAATTCAGTATCTATGCT GGTCGCCTGTTGGGAGTAAATTAGCATATGTATAT CAAAACAATATTTATTTGAAACAAAGACCAGGAG ATCCACCTTTTCAAATAACTTATACTGGAAGAGAA AATAGAATATTTAATGGAATACCAGACTGGGTTTA TGAAGAGGAAATGCTTGCCACAAAATATGCTCTTT GGTGGTCTCCAGATGGAAAATTTTGGCATATGTA GAATTTAATGATTCAGATATACCAATTATTGCCTA TTCTTATTATGGTGATGGACAGTATCCTAGAACTA TAAATATTCCATATCCAAAGGCTGGGGCTAAGAAT CCGGTTGTTTCGTGTTTTTATTGTTGACACCACCTAC CCTCACCACGTGGGCCCAATGGAAGTGCCAGTTCC AGAAATGATAGCCTCAAGTGACTATTATTTAGCT GGCTCACATGGGTGTCCAGTGAACGAGTATGCTTG CAGTGGCTAAAAAGAGTGCAGAATGTCTCAGTCCT GTCTATATGTGATTTAGGGAAGACTGGCATGCAT GGGAATGTCCAAAGAACCAGGAGCATGTAGAAGA AAGCAGAACAGGATGGGCTGGTGGATTCTTTGTTT CGACACCAGCTTTTAGCCAGGATGCCACTTCTTAC TACAAAATATTTAGCGACAAGGATGGTTACAAACA TATTCACATACATCAAAGACACTGTGGAATGCTA TTCAAATTACAAGTGGCAAGTGGGAGGCCATATAT ATATTCCGCGTAACACAGGATTCAGTGTTTATTCT AGCAATGAATTTGAAGGTTACCTGGAAGAAGAA ACATCTACAGAATTAGCATTGGAAACTCTCCTCCG AGCAAGAAGTGTGTTACTTGCCATCTAAGGAAAGA AAGGTGCCAATATTACACAGCAAGTTTCAGCTACA AAGCCAAGTACTATGCACTCGTCTGCTATGGCCCT GGCCTCCCCATTTCCACCCCTCCATGATGGCCGCAC AGACCAAGAAATACAAGTATTAGAAGAAAACAAA GAACTGGAAAATTTCTCTGAGAAATATCCAGCTGCC TAAAGTGGAGATTAAGAAGCTCAAAGACGGGGGA CTGACTTTCTGGTACAAGATGATTCTGCCTCCTCAG TTTGACAGATCAAAGAAGTACCCTTTGCTAATTCA AGTGTATGGTGGTCTTGTAGCCAGAGTGTTAAGT CTGTGTTTGCTGTTAATTGGATAACTTATCTCGCAA GTAAGGAGGGGATAGTCATTGCCCTGGTAGATGGT CGGGGCACTGCTTTCCAAGGTGACAAATTCCTGCA

SEQ ID NO:	Name	Sequence
		TGCCGTGTATCGAAAACCTGGGTGTATATGAAGTTG AGGACCAGCTCACAGCTGTCAGAAAATTCATAGA AATGGGTTTCATTGATGAAGAAAGAATAGCCATAT GGGGCTGGTCCTACGGAGGTTATGTTTCATCCCTG GCCCTTGCATCTGGAACCTGGTCTTTTCAAATGTGG CATAGCAGTGGCTCCAGTCTCCAGCTGGGAATATT ACGCATCTATCTACTCAGAGAGATTTCATGGGCCTC CCAACAAAGGACGACAATCTCGAACACTATAAAA ATTCAACTGTGATGGCAAGAGCAGAATATTTTCAGA AATGTAGACTATCTTCTCATCCACGGAACAGCAGA TGATAATGTGCACCTTCAGAACTCAGCACAGATTG CTAAAGCTTTGGTTAATGCACAAGTGGATTTCAG GCGATGTGGTACTCTGACCAGAACCATGGTATATT ATCTGGGCGCTCCCAGAATCATTTATATACCCACA TGACGCACTTCCTCAAGCAATGCTTTTCTTTATCAG ACGGCAAAAAGAAAAAGAAAAAGGGCCACCACCA TCACCATCAC
26	Cynomolgus FAP ectodomain+poly-lys- tag+his ₆ -tag	RPPRVHNSEENTMRALTLKDILNGTFSYKTFFPNWIS GQEYLHQADNNIVLYNIETGQSYTILSNRTMKSVNA SNYGLSPDRQFVYLESDYSKLWRYSYTATYYIYDLS NGEFVRGNELPRPIQYLCWSPVGSKLAYVYQNNIYL KQRPDPPFQITFNGRENKIFNGIPDWVYEEEMLATK YALWWSPNGKFLAYAEFNDTDIPVIAYSYYGDEQYP RTINIPYPKAGAKNPFVRIFIIDTTYPAYVGPQEVVVP AMIASDYYFSWLTWVTDERVCLQWLKRVQNVSVL SICDFREDWQTWDCPKTQEHIIESRTGWAGGFFVST PVFSYDAISYYKIFSDKDGYKHHYIKDTVENAIQITS GKWEAINIFRVTQDSL FYSSNEFEDYPGRRNIYRISIG SYPPSKKCVTCHLRKERCQYYTASFSDYAKYYALVC YGP GIPISTLHDGRTDQEIKILEENKELENALKNIQLP KEEIKKLEVDEITLWYKMILPPQFDRSKKYPLLIQVY GGPCSQSVRSVFAVNWISYLASKEGMVIALVDGRGT AFQGDKLLYAVYRKLGVYEVEDQITAVRKFIEMGFI DEKRIAIWGSYGGYVSSLALASGTGLFKCGIAVAP VSSWEYYASVYTERFMGLPTKDDNLEHYKNSTVMA RAEYFRNVLYLLIHGTADDNVHVFQNSAQIAKALVNA QVDFQAMWYSDQNHGLSGLSTNHL YTHMTHFLKQ CFSLSDGK KKKKKKGHHHHHH
27	nucleotide sequence Cynomolgus FAP ectodomain+poly-lys- tag+his ₆ -tag	CGCCCTCCAAGAGTTCATAACTCTGAAGAAAATAC AATGAGAGCACTCACACTGAAGGATATTTTAAATG GGACATTTTCTTATAAAACATTTTTTCCAAACTGGA TTTCAGGACAAGAATATCTTCATCAATCTGCAGAT AACAAATATAGTACTTTATAATATTGAAACAGGACA ATCATATACCATTTTGAGTAACAGAACCATGAAAA GTGTGAATGCTTCAAATTATGGCTTATCACCTGAT CGGCAATTTGTATATCTAGAAAGTGATTATTCAAA GCTTTGGAGATACTCTTACACAGCAACATATTACA TCTATGACCTTAGCAATGGAGAATTTGTAAGAGGA AATGAGCTTCCTCGTCCAATTCAGTATTTATGCTGG TCGCCTGTTGGGAGTAAATTAGCATATGTCTATCA AAACAATATCTATTTGAAACAAAGACCAGGAGAT CCACCTTTTCAAATAACATTTAATGGAAGAGAAAA

SEQ ID NO:	Name	Sequence
		TAAATATTTAATGGAATCCCAGACTGGGTTTATG AAGAGGAAATGCTTGCTACAAAATATGCTCTCTGG TGGTCTCCTAATGGAAAATTTTGGCATATGCGGA ATTTAATGATACAGATATACCAGTTATTGCCTATTC CTATTATGGCGATGAACAATATCCCAGAACAATAA ATATTCCATACCCAAAGGCCGGAGCTAAGAATCCT TTTGTTTCGGATATTTATTATCGATACCACTTACCCT GCGTATGTAGGTCCCCAGGAAGTGCCTGTTCCAGC AATGATAGCCTCAAGTGATTATTATTTTCAGTTGGC TCACGTGGGTACTGATGAACGAGTATGTTTGCAG TGGCTAAAAAGAGTCCAGAATGTTTCGGTCTTGTC TATATGTGATTTTCAGGGAAGACTGGCAGACATGGG ATTGTCCAAAGACCCAGGAGCATATAGAAGAAAG CAGAACTGGATGGGCTGGTGGATTCTTTGTTTCAA CACCAGTTTTCAGCTATGATGCCATTTTCATACTACA AAATATTTAGTGACAAGGATGGCTACAAACATATT CACTATATCAAAGACACTGTGGAAAATGCTATTCA AATTACAAGTGGCAAGTGGGAGGCCATAAATATA TTCAGAGTAACACAGGATTCAGTGTTCCTCTAG CAATGAATTTGAAGATTACCCTGGAAGAAGAAAC ATCTACAGAATTAGCATTGGAAGCTATCCTCCAAG CAAGAAGTGTGTTACTTGCCATCTAAGGAAAGAAA GGTGCCAATATTACACAGCAAGTTTCAGCGACTAC GCCAAGTACTATGCACTTGTCTGCTATGGCCCAGG CATCCCCATTTCCACCCTTCATGACGGACGCACTG ATCAAGAAATTAAAATCCTGGAAGAAAACAAGGA ATTGGAATAATGCTTTGAAAAATATCCAGCTGCCTA AAGAGGAAATTAAGAACTTGAAGTAGATGAAAT TACTTTATGGTACAAGATGATTCTTCCTCCTCAATT TGACAGATCAAAGAAGTATCCCTTGCTAATTCAAG TGTATGGTGGTCCCTGCAGTCAGAGTGTAAGGTCT GTATTTGCTGTTAATTGGATATCTTATCTTGCAAGT AAGGAAGGGATGGTCATTGCCTTGGTGGATGGTCG GGGAACAGCTTTCCAAGGTGACAACTCCTGTATG CAGTGTATCGAAAGCTGGGTGTTTATGAAGTTGAA GACCAGATTACAGCTGTCAGAAAATTCATAGAAAT GGGTTTCATTGATGAAAAAAGAATAGCCATATGGG GCTGGTCTATGGAGGATATGTTTCATCACTGGCC CTTGCACTCTGGAAGTGGTCTTTTCAAATGTGGGAT AGCAGTGGCTCCAGTCTCCAGCTGGGAATATTACG CGTCTGTCTACACAGAGAGATTCATGGGTCTCCCA ACAAAGGATGATAATCTTGAGCACTATAAGAATTC AACTGTGATGGCAAGAGCAGAATATTTTCAGAAAT GTAGACTATCTTCTCATCCACGGAACAGCAGATGA TAATGTGCACTTTCAAAACTCAGCACAGATTGCTA AAGCTCTGGTTAATGCACAAGTGGATTTCAGGCA ATGTGGTACTCTGACCAGAACCACGGCTTATCCGG CCTGTCCACGAACCACTTATACACCCACATGACCC ACTTCCTAAAGCAGTGTTCCTCTTTGTTCAGACGGC AAAAAGAAAAAGAAAAAGGGCCACCACCATCACC ATCAC
28	human CEA	UniProt no. P06731

SEQ ID NO:	Name	Sequence
29	human MCSP	UniProt no. Q6UVK1
30	human EGFR	UniProt no. P00533
31	human CD19	UniProt no. P15391
32	human CD20	Uniprot no. P11836
33	human CD33	UniProt no. P20138
34	human Lymphotoxin α	UniProt no. P01374
35	human TNF	UniProt no. P01375
36	human Lymphotoxin β	UniProt no. Q06643
37	human OX40L	UniProt no. P23510
38	human CD40L	UniProt no. P29965
39	human FasL	UniProt no. P48023
40	human CD27L	UniProt no. P32970
41	human CD30L	UniProt no. P32971
42	human 4-1BBL	UniProt no. P41273
43	human TRAIL	UniProt no. P50591
44	human RANKL	UniProt no. O14788
45	human TWEAK	UniProt no. O43508
46	human APRIL	UniProt no. O75888
47	human BAFF	UniProt no. Q9Y275
48	human LIGHT	UniProt no. O43557
49	human TL1A	UniProt no. O95150
50	human GITRL	UniProt no. Q9UNG2
51	human ectodysplasin A	UniProt no. Q92838
52	hu 4-1BBL (50-254)	ACPWAVSGARASPGSAASPRLREGPELSPDDPAGLL DLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLT GGLSYKEDTKELVVAKAGVYYVFFQLELRRVVAGE GSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEAR NSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQL TQGATVLGLFRVTPEIPAGLPSPRSE
53	hu OX40L (51-183)	QVSHRYPRIQSIKVQFTEYKKEKGFILTSQKEDEIMK VQNNSVIINCDFYLISLKGYSQEVNLSLHYQKDEEP LFQLKKVRSVNSLMVASLTYKDKVYLNVTDDNTSL DDFHVNGGELILIHQNPGEFCVL
54	hu OX40L (52-183)	VSHRYPRIQSIKVQFTEYKKEKGFILTSQKEDEIMKV QNNSVIINCDFYLISLKGYSQEVNLSLHYQKDEEPL FQLKKVRSVNSLMVASLTYKDKVYLNVTDDNTSLD DFHVNGGELILIHQNPGEFCVL
55	Peptide linker (SG4) ₂	SGGGSGGGG
56	Peptide linker G4(SG4) ₂	GGGGSGGGSGGGG
57	Peptide linker	GSPGSSSSGS
58	Peptide linker (G4S) ₄	GGGGSGGGSGGGSGGGGS
59	Peptide linker	GSGSGNGS
60	Peptide linker	GGSGSGSG
61	Peptide linker	GGSGSG
62	Peptide linker	GGSG
63	Peptide linker	GGSGNGSG
64	Peptide linker	GGNGSGSG

SEQ ID NO:	Name	Sequence
65	Peptide linker	GGNGSG
66	nucleotide sequence dimeric hu 4-1BBL (71-254)-CH1 Fc knob chain	See Table 2
67	nucleotide sequence monomeric hu 4-1BBL (71-254)-CL1	See Table 2
68	nucleotide sequence anti-FAP(28H1) Fc hole chain	See Table 2
69	nucleotide sequence anti-FAP (28H1) light chain	See Table 2
70	Murine (mu) 4-1BBL	UniProt no. Q3U1Z9-1
71	nucleotide sequence dimeric mu 4-1BBL (104-309, C137,160,246S)-CH1 Fc knob chain	See Table 13
72	nucleotide sequence monomeric mu 4-1BBL (104-309, C137,160,246S)-CL	See Table 13
73	nucleotide sequence anti-FAP Fc KK chain	See Table 13
74	nucleotide sequence anti-FAP light chain	See Table 13
75	dimeric mu 4-1BBL (104-309, C137,160,246S) – CL Fc DD chain	See Table 13
76	monomeric mu 4-1BBL (104-309, C137,160,246S)-CL1	See Table 13
77	anti-FAP Fc KK chain	See Table 13
78	anti-FAP light chain	See Table 13
79	nucleotide sequence DP47 Fc-hole chain	See Table 18
80	nucleotide sequence DP47 light chain	See Table 18
81	DP47 Fc-hole chain	See Table 18
82	DP47 light chain	See Table 18
83	Human 4-1BB Fc(kih)	See Table 31
84	Cynomolgus 4-1BB Fc(kih)	See Table 31
85	Murine 4-1BB Fc(kih)	See Table 31
86	nucleotide sequence Fc hole chain	See Table 32
87	nucleotide sequence Human 4-1BB Fc(kih)	See Table 32
88	nucleotide sequence	See Table 32

SEQ ID NO:	Name	Sequence
	Cynomolgus 4-1BB Fc(kih)	
89	nucleotide sequence Murine 4-1BB Fc(kih)	See Table 32
90	Fc hole chain	See Table 32
91	Human 4-1BB Fc(kih)	See Table 32
92	Cynomolgus 4-1BB Fc(kih)	See Table 32
93	Murine 4-1BB Fc(kih)	See Table 32
94	nucleotide sequence Human 4-1BB His	See Table 33
95	Human 4-1BB His	See Table 33
96	Human (hu) 4-1BBL (71-248)	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVY YVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGL
97	dimeric hu 4-1BBL (71-248) connected by (G4S) ₂ linker	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVY YVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGL
98	dimeric hu 4-1BBL (80-254) connected by (G4S) ₂ linker	DPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSEGGGGSGGGSDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSE
99	dimeric hu 4-1BBL (52-254) connected by (G4S) ₂ linker	PWAVSGARASPGSAASPRLREGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSEGGGGSGGGGSPWAVSGARASPGSAASPRLREGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSE
100	FAP(4B9) CDR-H1	SYAMS
101	FAP(4B9) CDR-H2	AIIGSGASTYYADSVKG

SEQ ID NO:	Name	Sequence
102	FAP(4B9) CDR-H3	GWFGGFNY
103	FAP(4B9) CDR-L1	RASQSVTSSYLA
104	FAP(4B9) CDR-L2	VGSRRAT
105	FAP(4B9) CDR-L3	QQGIMLPPT
106	FAP(4B9) VH	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSYA MSWVRQAPGKGLEWVSAIIGSGASTYYADSVK GRFTISRDN SKNTLYLQMNSLRAEDTAVYYCAK GWFGGFNYWGQGT LVT VSS
107	FAP(4B9) VL	EIVLTQSPGTL SLSPGERATL SCRASQSVTSSYLA WYQQKPGQAPRLIN VGSRRATGIPDRFSGSGSG TDFTLTISRLEPEDFAVYYCQQGIMLPPTFGQGT KVEIK
108	dimeric hu 4-1BBL (71-254)-CH1* Fc knob chain	see Table 4
109	monomeric hu 4-1BBL (71-254)-CL*	see Table 4
110	monomeric hu 4-1BBL (71-254)-(G4S)1-CL*	see Table 7
111	dimeric hu 4-1BBL (52-254)-CH1* Fc knob chain	see Table 10
112	monomeric hu 4-1BBL (52-254)-CL*	see Table 10
113	dimeric hu 4-1BBL (80-254)-CH1* Fc knob chain	see Table 11
114	monomeric hu 4-1BBL (80-254)-CL*	see Table 11
115	dimeric hu 4-1BBL (71-254)-CL* Fc knob chain	see Table 3
116	monomeric hu 4-1BBL (71-254)-CH1*	see Table 3
117	dimeric hu 4-1BBL (71-254)-CL Fc knob chain	see Table 22
118	monomeric hu 4-1BBL (71-254)-CH1	see Table 22
119	dimeric hu 4-1BBL (71-248)-CL* Fc knob chain	see Table 24
120	monomeric hu 4-1BBL (71-248)-CH1*	see Table 24
121	anti-FAP (28H1) Fc hole chain fused to dimeric 4-1BBL (71-254)	see Table 6
122	anti-FAP (28H1) Fc knob chain fused to monomeric 4-1BBL (71-254)	see Table 6
123	anti-FAP (4B9) Fc hole chain fused to dimeric 4-1BBL (71-254)	see Table 23

SEQ ID NO:	Name	Sequence
124	anti-FAP (4B9) Fc knob chain fused to monomeric 4-1BBL (71-254)	see Table 23
125	anti-FAP (4B9) light chain	see Table 21
126	anti-FAP (4B9) Fc hole chain fused to dimeric 4-1BBL (71-248)	see Table 26
127	anti-FAP (4B9) Fc knob chain fused to monomeric 4-1BBL (71-248)	see Table 26
128	Peptide linker	GGGGS
129	nucleotide sequence dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 3
130	nucleotide sequence monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
131	nucleotide sequence dimeric hu 4-1BBL (71-254) – CH1* Fc knob chain	see Table 4
132	nucleotide sequence monomeric hu 4-1BBL (71-254) – CL*	see Table 4
133	nucleotide sequence anti-FAP (28H1) (VHCL) Fc hole chain	see Table 4
134	nucleotide sequence anti-FAP (28H1) (VLCH1) light chain	see Table 4
135	anti-FAP (VHCL) (28H1) Fc hole chain	see Table 4
136	anti-FAP (VLCH1) (28H1) light chain	see Table 4
137	nucleotide sequence monomeric hu 4-1BBL (71-254) – CH1* Fc knob chain	see Table 5
138	nucleotide sequence dimeric hu 4-1BBL (71-254) – CL*	see Table 5
139	monomeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 5
140	dimeric hu 4-1BBL (71-254) – CL*	see Table 5
141	nucleotide sequence anti-FAP (28H1) Fc hole chain fused to dimeric hu 4-1BBL (71-254)	see Table 6
142	nucleotide sequence anti-	see Table 6

SEQ ID NO:	Name	Sequence
	FAP (28H1) Fc knob chain fused to monomeric hu 4-1BBL (71-254)	
143	nucleotide sequence monomeric hu 4-1BBL (71-254) –(G4S) ₁ - CL*	see Table 7
144	nucleotide sequence [anti-FAP (28H1)] ₂ Fc hole chain	see Table 8
145	[anti-FAP (28H1)] ₂ Fc hole chain	see Table 8
146	nucleotide sequence dimeric hu 4-1BBL (71-254) – FAP (VHCL*) Fc knob chain	see Table 9
147	nucleotide sequence monomeric hu 4-1BBL (71-254) –FAP (VLCH1*)	see Table 9
148	dimeric hu 4-1BBL (71-254) – FAP (VHCL*) Fc knob chain	see Table 9
149	monomeric hu 4-1BBL (71-254) –FAP (VLCH1*)	see Table 9
150	nucleotide sequence dimeric hu 4-1BBL (52-254) – CH1* Fc knob chain	see Table 10
151	nucleotide sequence Monomeric hu 4-1BBL (52-254) –CL*	see Table 10
152	nucleotide sequence dimeric hu 4-1BBL (80-254) – CH1* Fc knob chain	see Table 11
153	nucleotide sequence Monomeric hu 4-1BBL (80-254) –CL*	see Table 11
154	nucleotide sequence DP47 FC KK chain	see Table 14
155	nucleotide sequence DP47 light chain	see Table 14
156	DP47 FC KK chain	see Table 14
157	DP47 light chain	see Table 14
158	nucleotide sequence dimeric mu 4-1BBL (104-309, C160S) – CL Fc DD chain	see Table 15
159	nucleotide sequence monomeric murine 4-1BBL (104-309, C160S) – CH1	see Table 15
160	dimeric mu 4-1BBL (104-309, C160S) – CL Fc DD	see Table 15

SEQ ID NO:	Name	Sequence
	chain	
161	monomeric murine 4-1BBL (104-309, C160S) – CH1	see Table 15
162	nucleotide sequence anti-FAP (4B9) Fc hole chain	see Table 21
163	nucleotide sequence anti-FAP (4B9) light chain	see Table 21
164	anti-FAP (4B9) Fc hole chain	see Table 21
165	nucleotide sequence dimeric hu 4-1BBL (71-254) – CL Fc knob chain	see Table 22
166	nucleotide sequence monomeric hu 4-1BBL (71-254) – CH1	see Table 22
167	nucleotide sequence anti-FAP (4B9) Fc hole chain fused to dimeric hu 4-1BBL (71-254)	see Table 23
168	nucleotide sequence anti-FAP (4B9) Fc knob chain fused to monomeric hu 4-1BBL (71-254)	see Table 23
169	nucleotide sequence dimeric hu 4-1BBL (71-248) – CL* Fc knob chain	see Table 24
170	nucleotide sequence monomeric hu 4-1BBL (71-248) – CH1*	see Table 24
171	nucleotide sequence dimeric hu 4-1BBL (71-248) – CL Fc knob chain	see Table 25
172	nucleotide sequence monomeric hu 4-1BBL (71-248) – CH1	see Table 25
173	Dimeric hu 4-1BBL (71-248) – CL Fc knob chain	see Table 25
174	Monomeric hu 4-1BBL (71-248) – CH1	see Table 25
175	nucleotide sequence anti-FAP (4B9) Fc hole chain fused to dimeric hu 4-1BBL (71-248)	see Table 26
176	nucleotide sequence anti-FAP (4B9) Fc knob chain	see Table 26

SEQ ID NO:	Name	Sequence
	fused to monomeric hu 4-1BBL (71-248)	
177	nucleotide sequence DP47 Fc hole chain fused to dimeric hu 4-1BBL (71-254)	see Table 27
178	nucleotide sequence DP47 Fc knob chain fused to monomeric hu 4-1BBL (71-254)	see Table 27
179	DP47 Fc hole chain fused to dimeric hu 4-1BBL (71-254)	see Table 27
180	DP47 Fc knob chain fused to monomeric hu 4-1BBL (71-254)	see Table 27
181	nucleotide sequence DP47 heavy chain (hu IgG1 PGLALA)	see Table 29
182	DP47 heavy chain (hu IgG1 PGLALA)	see Table 29
183	monomeric hu 4-1BBL (71-254) plus (G4S) ₁ linker	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGP LSWYSDPGLAGVSLTGGLSYKEDTKELVVAAGVY YVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAA ALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLG VHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLP SPRSEGGGGS
184	monomeric hu 4-1BBL (71-248) plus (G4S) ₂ linker	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGP LSWYSDPGLAGVSLTGGLSYKEDTKELVVAAGVY YVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAA ALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLG VHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLG GGGSGGGGS
185	monomeric hu 4-1BBL (71-248) plus (G4S) ₁ linker	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGP LSWYSDPGLAGVSLTGGLSYKEDTKELVVAAGVY YVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAA ALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLG VHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLG GGGS
186	Nucleotide sequence human CD19 antigen Fc knob chain avi tag	see Table 43
187	Polypeptide sequence human CD19 antigen Fc knob chain avi tag	see Table 43
188	Nucleotide sequence cynomolgus CD19 antigen Fc knob chain avi tag	see Table 43
189	Polypeptide sequence cynomolgus CD19 antigen Fc knob chain avi tag	see Table 43

SEQ ID NO:	Name	Sequence
190	humanized CD19 (8B8) HVR-L1	NSNGNT
191	humanized CD19 (8B8) HVR-H2	KFNG
192	humanized CD19 (8B8) var.1 to 9 HVR-H2	TEKFQGRVTM
193	humanized CD19 (8B8) var.5 HVR-L1	LENPNGNT
194	humanized CD19 (8B8) var.9 HVR-L1	LENPSGNT
195	CD19 (8B8-018) CDR-H1	DYIMH
196	CD19 (8B8-018) CDR-H2	YINPYNDGSKYTEKFQG
197	CD19 (8B8-018) CDR-H3	GTYYYGSALFDY
198	CD19 (8B8-018) CDR-L1	KSSQSLENPNGNTYLN
199	CD19 (8B8-018) CDR-L2	RVSKRFS
200	CD19 (8B8-018) CDR-L3	LQLTHVPYT
201	CD19 (8B8-018) VH	QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYIMH WVRQAPGQGLEWMGYINPYNDGSKYTEKFQGRVT MTSDTSISTAYMELSRLRSDDTAVYYCARGTYYYGS ALFDYWGQGTTVTVSS
202	CD19 (8B8-018) VL	DIVMTQTPLSLSVTPGQPASISCKSSQSLENPNGNTYL NWYLRKPGQSPQLLIYRVSKRFSGVPDRFSGSGSGTD FTLKISRVEAEDVGVYYCLQLTHVPYTFGQGTKLEIK
203	Nucleotide sequence anti-CD19(8B8-018) Fc hole chain	see Table 47
204	Nucleotide sequence anti-CD19(8B8-018) light chain	see Table 47
205	anti-CD19(8B8-018) Fc hole chain	see Table 47
206	anti-CD19(8B8-018) light chain	see Table 47
207	Nucleotide sequence anti-CD19(8B8-018) Fc hole dimeric ligand chain	see Table 49
208	Nucleotide sequence anti-CD19(8B8-018) Fc knob monomeric ligand	see Table 49
209	anti-CD19(8B8-018) Fc hole dimeric ligand chain	see Table 49
210	anti-CD19(8B8-018) Fc knob monomeric ligand	see Table 49
211	Nucleotide sequence anti-CD19(8B8-018) Fc hole dimeric ligand (71-248)	see Table 52

SEQ ID NO:	Name	Sequence
	chain	
212	Nucleotide sequence anti-CD19(8B8-018) Fc knob monomeric (71-248) ligand	see Table 52
213	anti-CD19(8B8-018) Fc hole dimeric ligand (71-248) chain	see Table 52
214	anti-CD19(8B8-018) Fc knob monomeric (71-248) ligand	see Table 52
215	Nucleotide sequence CD19 (8B8) VH Parental clone	GAGGTCCAGCTGCAGCAGTCTGGACCTGAGCTGGTAAAGCCTGGGGCTTCAGTGAAGATGGCCTGCAAGGCTTCTGGATACACATTCAGTACTATATTATGCACTGGGTGAAGCAGAAAGACTGGGCAGGGCCTTGAGTGGATTGGATATATTAATCCTTACAATGATGGTTCTAAGTACACTGAGAAGTTCAACGGCAAGGCCACACTGACTTCAGACAAATCTTCCATCACAGCCTACATGGAGCTCAGCAGCCTGACCTCTGAGGACTCTGCGGTCTATTACTGTGCAAGAGGGACCTATTATTATGGTAGCGCCCTCTTTGACTACTGGGGCCAAGGCACCACTCTCACAGTCTCCTCG
216	Nucleotide sequence CD19 (8B8) VL Parental clone	GATGCTGTGATGACCCAAACTCCACTCTCCCTGCCGTGTCAGTCTTGGAGATCAAGCCTCCATCTCTTGCAAGTCTAGTCAGAGCCTTGAAAACAGTAATGGAAACACCTATTTGAACTGGTACCTCCAGAAACCAGGCCAGTCTCCACAACCTCCTGATCTACAGGGTTTCCAAACGATTTTCTGGGGTCCTAGACAGGTTTCAGTGGTAGTGGATCAGGGACAGATTTTCACTGAAAATCAGCAGAGTGGAGGCTGAGGATTTGGGAGTTTATTTCTGCTACAACCTTACACATGTCCCGTACACGTTTCGGAGGGGGACCAAGCTGGAAATAAAA
217	CD19 L1 reverse random	see Table 53
218	CD19 L2 forward random	see Table 53
219	CD19 H1 reverse random	see Table 53
220	CD19 H2 forward random	see Table 53
221	CD19 H3 reverse constant	see Table 53
222	LMB3	see Table 53
223	D19 L1 forward constant	see Table 54
224	CD19 L3 reverse random	see Table 54
225	CD19 L3 forward constant	see Table 54
226	CD19 H3 reverse random	see Table 54
227	Nucleotide sequence SNAP tag human CD19 ECD-PDGFR	GGCCGCCGCTAGCGGCATCGACTACAAGGACGACGATGACAAGGCCGGCATCGATGCCATCATGGACAAGACTGCGAAATGAAGCGCACCAACCCTGGATAG

SEQ ID NO:	Name	Sequence
		CCCTCTGGGCAAGCTGGAAGTGTCTGGGTGCGAAC AGGGCCTGCACGAGATCAAGCTGCTGGGCAAAGG AACATCTGCCGCCGACGCCGTGGAAGTGCCTGCCC CAGCCGCCGTGCTGGGCGGACCAGAGCCACTGAT GCAGGCCACCGCCTGGCTCAACGCCTACTTTCACC AGCCTGAGGCCATCGAGGAGTTCCCTGTGCCAGCC CTGCACCACCCAGTGTTCCAGCAGGAGAGCTTTAC CCGCCAGGTGCTGTGGAACTGCTGAAAGTGGTGA AGTTCGAGAGGTCATCAGCTACCAGCAGCTGGCC GCCCTGGCCGGCAATCCCGCCGCCACCGCCGCCGT GAAAACCGCCCTGAGCGGAAATCCCGTGCCCATTC TGATCCCCTGCCACCGGGTGGTGTCTAGCTCTGGC GCCGTGGGGGGCTACGAGGGCGGGCTCGCCGTGA AAGAGTGGCTGCTGGCCACGAGGGCCACAGACT GGGCAAGCCTGGGCTGGGTGATATCCCCGAGGAA CCCCTGGTCGTGAAGGTGGAAGAGGGCGACAATG CCGTGCTGCAGTGCCTGAAGGGCACCTCCGATGGC CCTACCCAGCAGCTGACCTGGTCCAGAGAGAGCCC CCTGAAGCCCTTCCTGAAGCTGTCTCTGGGCCTGC CTGGCCTGGGCATCCATATGAGGCCTCTGGCCATC TGGCTGTTTCATCTTCAACGTGTCCAGCAGATGGG CGGCTTCTACCTGTGTGTCAGCCTGGCCCCCATCTG AGAAGGCTTGGCAGCCTGGCTGGACCGTGAACGT GGAAGGATCCGGCGAGCTGTTCCGGTGGAACGTGT CCGATCTGGGCGGCCTGGGATGCGGCCTGAAGAA CAGATCTAGCGAGGGCCCCAGCAGCCCCAGCGGC AACTGATGAGCCCCAAGCTGTACGTGTGGGCCAA GGACAGACCCGAGATCTGGGAGGGCGAGCCTCCT TGCCTGCCCCCTAGAGACAGCCTGAACCAGAGCCT GAGCCAGGACCTGACAATGGCCCCCTGGCAGCACA CTGTGGCTGAGCTGTGGCGTGCCACCCGACTCTGT GTCTAGAGGCCCTCTGAGCTGGACCCACGTGCACC CTAAGGGCCCTAAGAGCCTGCTGAGCCTGGAAGT AAGGACGACAGGCCCGCCAGAGATATGTGGGTCA TGAAAACCGGCCTGCTGCTGCCTAGAGCCACAGCC CAGGATGCCGGCAAGTACTACTGCCACAGAGGCA ACCTGACCATGAGCTTCCACCTGGAAATCACCGCC AGACCCGTGCTGTGGCACTGGCTGCTGAGAACAGG CGGCTGGAAGGTCGACGAACAAAACTCATCTCA GAAGAGGATCTGAATGCTGTGGGCCAGGACACGC AGGAGGTCATCGTGGTGCCACACTCCTTGCCCTTT AAGGTGGTGGTGATCTCAGCCATCCTGGCCCTGGT GGTGCTCACCATCATCTCCCTTATCATCCTCATCAT GCTTTGGCAGAAGAAGCCACGT
228	Nucleotide sequence SNAP tag cynomolgus CD19 ECD- PDGFR	CCGGCCGCCGCTAGCGGCATCGACTACAAGGACG ACGATGACAAGGCCGGCATCGATGCCATCATGGA CAAAGACTGCGAAATGAAGCGCACCACCCTGGAT AGCCCTCTGGGCAAGCTGGAAGTGTCTGGGTGCGA ACAGGGCCTGCACGAGATCAAGCTGCTGGGCAA GGAACATCTGCCGCCGACGCCGTGGAAGTGCCTGC CCCAGCCGCCGTGCTGGGCGGACCAGAGCCACTG ATGCAGGCCACCGCCTGGCTCAACGCCTACTTTCA

SEQ ID NO:	Name	Sequence
		CCAGCCTGAGGCCATCGAGGAGTTCCTGTGCCAG CCCTGCACCACCCAGTGTTCAGCAGGAGAGCTTT ACCCGCCAGGTGCTGTGGAAACTGCTGAAAGTGGT GAAGTTCGGAGAGGTCATCAGCTACCAGCAGCTG GCCGCCCTGGCCGGCAATCCCGCCGCCACCGCCGC CGTGAAAACCGCCCTGAGCGGAAATCCCGTGCCCA TTCTGATCCCCTGCCACCGGGTGGTGTCTAGCTCTG GCGCCGTGGGGGGCTACGAGGGCGGGGCTCGCCGT GAAAGAGTGGCTGCTGGCCCACGAGGGCCACAGA CTGGGCAAGCCTGGGCTGGGTGATATCCCCCAGGA ACCCCTGGTCGTGAAGGTGGAAGAGGGCGACAAT GCCGTGCTCCAGTGTCTCGAGGGCACCTCCGATGG CCCTACACAGCAGCTCGTGTGGTGCAGAGACAGCC CCTTCGAGCCCTTCCTGAACCTGTCTCTGGGCCTGC CTGGCATGGGCATCAGAATGGGCCCTCTGGGCATC TGGCTGCTGATCTTCAACGTGTCCAACCAGACCGG CGGCTTCTACCTGTGTTCAGCCTGGCCTGCCAAGCG AGAAGGCTTGGCAGCCTGGATGGACCGTGTCCGTG GAAGGATCTGGCGAGCTGTTCCGGTGGAACGTGTC CGATCTGGGCGGCCTGGGATGCGGCCTGAAGAAC AGAAGCAGCGAGGGCCCTAGCAGCCCCAGCGGCA AGCTGAATAGCAGCCAGCTGTACGTGTGGGCCAA GGACAGACCCGAGATGTGGGAGGGCGAGCCTGTG TGTGGCCCCCTAGAGATAGCCTGAACCAGAGCCT GAGCCAGGACCTGACAATGGCCCCCTGGCAGCACA CTGTGGCTGAGCTGTGGCGTGCCACCCGACTCTGT GTCCAGAGGCCCTCTGAGCTGGACACACGTGCGGC CTAAGGGCCCTAAGAGCAGCCTGCTGAGCCTGGA ACTGAAGGACGACCGGCCCCGACCGGGATATGTGG GTGGTGGATACAGGCCTGCTGCTGACCAGAGCCAC AGCCCAGGATGCCGGCAAGTACTACTGCCACAGA GGCAACTGGACCAAGAGCTTTTACCTGGAATCAC CGCCAGACCCGCCCTGTGGCACTGGCTGCTGAGAA TCGGAGGCTGGAAGGTCGACGAGCAGAAGCTGAT CTCCGAAGAGGACCTGAACGCCGTGGGCCAGGAT ACCCAGGAAGTGATCGTGGTGCCCCACAGCCTGCC CTCAAGGTGGTCTGATCAGCGCCATTCTGGCCC TGGTGGTGCTGACCATCATCAGCCTGATCATCCTG ATTATGCTGTGGCAGAAAAAGCCCCGC
229	Polypeptide sequence SNAP tag human CD19 ECD- PDGFR	PAAASGIDYKDDDDKAGIDAIMDKDCMKRTTLDSP LGKLELSGCEQGLHEIKLLGKGTSAADAVEVPAPAA VLGGPEPLMQATAWLNAYFHQPEAIEEFPVPALHHP VFQQESFTRQVLWKKLVVKFGEVISYQQLAALAGN PAATAAVKTAALSGNPVPILIPCHRVVSSSGAVGGYEG GLAVKEWLLAHEGHRLGKPLGDIPEEPLVVKVEEG DNAVLQCLKGTSDGPTQQLTWSRESPLKPFLKLSLG LPGLGIHMRPLAIWLFIFNVSQQMGGFYLCQPGPPSE KAWQPGWTVNVEGSGELFRWNVSDLGGLGCGLKN RSSEGPSSPSGKLMSPKLYVWAKDRPEIWEGEPPCLP PRDSL NQSLSQDLTMAPGSTLWLSCGVPPDSVSRGPL

SEQ ID NO:	Name	Sequence
		SWTHVHPKGPKSLLSLELKDDRPARDMWVMETGLL LPRATAQDAGKYYCHRGNTMSFHLEITARPVLWH WLLRTGGWKVDEQKLISEEDLNAVVGQDTQEIVVVP HSLPFKVVVISAILALVVLTIISLIILIMLWQKKPR
230	Polypeptide sequence SNAP tag cynomolgus CD19 ECD- PDGFR	PAAASGIDYKDDDDKAGIDAIMDKDCMKRTTLDSP LGKLELSGCEQGLHEIKLLGKGTSAADAVEVPAPAA VLGGPEPLMQATAWLNAYFHQPEAIEEFPVPALHHP VFQQESFTRQVLWKLKVVKFGEVISYQQLAALAGN PAATAAVKTALSGNPVPIIPCHRVVSSSGAVGGYEG GLAVKEWLLAHEGHRLGKPGLDIPQEPLVVKVEEG DNAVLQCLEGTSDGPTQQLVWCRDSPFEPFLNLSLG LPGMGIRMGPLGIWLLIFNVSNQTGGFYLCQPGLPSE KAWQPGWTVSVEGSGELFRWNVSDLGGLGCGLKN RSSEGPSSPSGKLNSSQLYVWAKDRPEMWEGEPVCG PPRDSL NQSLSQDLTMAPGSTLWLSCGVPPDSVSRGP LSWTHVRPKGPKSLLSLELKDDRPDRDMWVVDTG LLLTRATAQDAGKYYCHRGNTKSFYLEITARPAL WHWLLRIGGWKVDEQKLISEEDLNAVVGQDTQEIVV VPHSLPFKVVVISAILALVVLTIISLIILIMLWQKKPR
231	CD19 (8B8-5H09) CDR-L1	see Table 56
232	CD19 (8B8-5H09) CDR-L2	see Table 56
233	CD19 (8B8-5H09) CDR-L3	see Table 56
234	CD19 (8B8-5H09) CDR-H1	see Table 57
235	CD19 (8B8-5H09) CDR-H2	see Table 57
236	CD19 (8B8-5H09) CDR-H3	see Table 57
237	CD19 (8B8-7H07) CDR-L1	see Table 56
238	CD19 (8B8-7H07) CDR-L2	see Table 56
239	CD19 (8B8-7H07) CDR-L3	see Table 56
240	CD19 (8B8-7H07) CDR-H1	see Table 57
241	CD19 (8B8-7H07) CDR-H2	see Table 57
242	CD19 (8B8-7H07) CDR-H3	see Table 57
243	CD19 (8B8-2B03) CDR-L1	see Table 56
244	CD19 (8B8-2B03) CDR-L2	see Table 56
245	CD19 (8B8-2B03) CDR-L3	see Table 56
246	CD19 (8B8-2B03) CDR-H1	see Table 57
247	CD19 (8B8-2B03) CDR-H2	see Table 57
248	CD19 (8B8-2B03) CDR-H3	see Table 57
249	CD19 (8B8-2B11) CDR-L1	see Table 56
250	CD19 (8B8-2B11) CDR-L2	see Table 56
251	CD19 (8B8-2B11) CDR-L3	see Table 56
252	CD19 (8B8-2B11) CDR-H1	see Table 57
253	CD19 (8B8-2B11) CDR-H2	see Table 57
254	CD19 (8B8-2B11) CDR-H3	see Table 57
255	CD19 (8B8-5A07) CDR-L1	see Table 56

SEQ ID NO:	Name	Sequence
256	CD19 (8B8-5A07) CDR-L2	see Table 56
257	CD19 (8B8-5A07) CDR-L3	see Table 56
258	CD19 (8B8-5A07) CDR-H1	see Table 57
259	CD19 (8B8-5A07) CDR-H2	see Table 57
260	CD19 (8B8-5A07) CDR-H3	see Table 57
261	CD19 (8B8-5B08) CDR-L1	see Table 56
262	CD19 (8B8-5B08) CDR-L2	see Table 56
263	CD19 (8B8-5B08) CDR-L3	see Table 56
264	CD19 (8B8-5B08) CDR-H1	see Table 57
265	CD19 (8B8-5B08) CDR-H2	see Table 57
266	CD19 (8B8-5B08) CDR-H3	see Table 57
267	CD19 (8B8-5D08) CDR-L1	see Table 56
268	CD19 (8B8-5D08) CDR-L2	see Table 56
269	CD19 (8B8-5D08) CDR-L3	see Table 56
270	CD19 (8B8-5D08) CDR-H1	see Table 57
271	CD19 (8B8-5D08) CDR-H2	see Table 57
272	CD19 (8B8-5D08) CDR-H3	see Table 57
273	nucleotide sequence CD19 (8B8) parental light chain	see Table 58
274	nucleotide sequence CD19 (8B8) parental heavy chain	see Table 58
275	CD19 (8B8) parental light chain	see Table 58
276	CD19 (8B8) parental heavy chain	see Table 58
277	nucleotide sequence CD19 (8B8-2B11) light chain	see Table 59
278	nucleotide sequence CD19 (8B8-2B11) heavy chain	see Table 59
279	CD19 (8B8-2B11) light chain	see Table 59
280	CD19 (8B8-2B11) heavy chain	see Table 59
281	nucleotide sequence CD19 (8B8-7H07) light chain	see Table 59
282	nucleotide sequence CD19 (8B8-7H07) heavy chain	see Table 59
283	CD19 (8B8-7H07) light chain	see Table 59
284	CD19 (8B8-7H07) heavy chain	see Table 59
285	nucleotide sequence CD19 (8B8-2B03) light chain	see Table 59
286	nucleotide sequence CD19 (8B8-2B03) heavy chain	see Table 59
287	CD19 (8B8-2B03) light chain	see Table 59

SEQ ID NO:	Name	Sequence
288	CD19 (8B8-2B03) heavy chain	see Table 59
289	nucleotide sequence CD19 (8B8-5A07) light chain	see Table 59
290	nucleotide sequence CD19 (8B8-5A07) heavy chain	see Table 59
291	CD19 (8B8-5A07) light chain	see Table 59
292	CD19 (8B8-5A07) heavy chain	see Table 59
293	nucleotide sequence CD19 (8B8-5D08) light chain	see Table 59
294	nucleotide sequence CD19 (8B8-5D08) heavy chain	see Table 59
295	CD19 (8B8-5D08) light chain	see Table 59
296	CD19 (8B8-5D08) heavy chain	see Table 59
297	nucleotide sequence CD19 (8B8-5B08) light chain	see Table 59
298	nucleotide sequence CD19 (8B8-5B08) heavy chain	see Table 59
299	CD19 (8B8-5B08) light chain	see Table 59
300	CD19 (8B8-5B08) heavy chain	see Table 59
301	nucleotide sequence CD19 (8B8-5H09) light chain	see Table 59
302	nucleotide sequence CD19 (8B8-5H09) heavy chain	see Table 59
303	CD19 (8B8-5H09) light chain	see Table 59
304	CD19 (8B8-5H09) heavy chain	see Table 59
305	Nucleotide sequence anti-CD19(8B8-2B11) Fc hole chain	see Table 62
306	anti-CD19(8B8-2B11) Fc hole chain	see Table 62
307	Nucleotide sequence anti-CD19(8B8-2B11) Fc hole dimeric ligand chain	see Table 64
308	Nucleotide sequence anti-CD19(8B8-2B11) Fc knob monomeric ligand	see Table 64
309	anti-CD19(8B8-2B11) Fc	see Table 64

SEQ ID NO:	Name	Sequence
	hole dimeric ligand chain	
310	anti-CD19(8B8-2B11) Fc knob monomeric ligand	see Table 64
311	Nucleotide sequence anti-CD19(8B8-2B11) Fc hole dimeric ligand (71-248) chain	see Table 67
312	Nucleotide sequence anti-CD19(8B8-2B11) Fc knob monomeric (71-248) ligand	see Table 67
313	anti-CD19(8B8-2B11) Fc hole dimeric ligand (71-248) chain	see Table 67
314	anti-CD19(8B8-2B11) Fc knob monomeric (71-248) ligand	see Table 67
315	nucleotide sequence CD19(8B8-018) heavy chain (huIgG1 PGLALA)	see Table 72
316	CD19(8B8-018) heavy chain (huIgG1 PGLALA)	see Table 72
317	anti-mu CEA T84.66 VH	MKCSWVIFFL MAVVTGVNSE VQLQQSGAEL VEPGASVKLS CTASGFNIKD TYMHWVKQRP EQGLEWIGRI DPANGNSKYV PKFQGKATIT ADTSSNTAYL QLTSLTSED AVYYCAPFGY YVSDYAMAYW GQGTSVTVSS
318	anti-mu CEA T84.66 VL	METDTLLLVV LLLWVPGSTG DIVLTQSPAS LAVSLGQRAT MSCRAGESVD IFGVGFLHWY QQKPGQPPKL LIYRASNLES GIPVRFSGTG SRDFTLIID PVEADDVATY YCQQTNEPDY TFGGGTKLEI K
319	IGHV1-69*08 IMGT Acc No. Z14309	TAAGGGGCTT CCTAGTCCTA AGGCTGAGGA AGGGATCCTG GTTTAGTTAA AGAGGATTTT ATTCACCCCT GTGTCCTCTC CACAGGTGTC CAGTCCCAGG TCCAGCTGGT GCAATCTGGG GCTGAGGTGA AGAAGCCTGG GTCCTCGGTG AAGGTCTCCT GCAAGGCTTC TGGAGGCACC TTCAGCAGCT ATACTATCAG CTGGGTGCGA CAGGCCCTG GACAAGGGCT TGAGTGGATG GGAAGGATCA TCCCTATCCT TGGTACAGCA AACTACGCAC AGAAGTTCCA GGCAGAGTC ACGATTACCG CGGACAAATC CACGAGCACA GCCTACATGG AGCTGAGCAG CCTGAGATCT GAGGACACGG CCGTGTATTA CTGTGCGAGA GA
320	IGKV3-11*01 IMGT Acc No.	CTGCAGCTGG AAGCTCAGCT CCCACCCAGC TGCTTTGCAT GTCCCTCCCA GCTGCCCTAC CTCCAGAGC CCATATCAAT GCCTGTGTCA GAGCCCTGGG GAGGAACTGC TCAGTTAGGA

SEQ ID NO:	Name	Sequence
		CCCAGAGGGA ACCATGGAAG CCCCAGCTCA GCTTCTCTTC CTCCTGCTAC TCTGGCTCCC AGGTGAGGGG AACATGAGGT GGTTTTGCAC ATTAGTGAAA ACTCTTGCCA CCTCTGCTCA GCAAGAAATA TAATTAAAAT TCAAAGTATA TCAACAATTT TGGCTCTACT CAAAGACAGT TGGTTTGATC TTGATTACAT GAGTGCATTT CTGTTTTATT TCCAATTTCA GATACCACCG GAGAAATTGT GTTGACACAG TCTCCAGCCA CCCTGTCTTT GTCTCCAGGG GAAAGAGCCA CCCTCTCCTG CAGGGCCAGT CAGAGTGTTA GCAGCTACTT AGCCTGGTAC CAACAGAAAC CTGGCCAGGC TCCCAGGCTC CTCATCTATG ATGCATCCAA CAGGGCCACT GGCATCCCAG CCAGGTTTCA TGGCAGTGGG TCTGGGACAG ACTTCACTCT CACCATCAGC AGCCTAGAGC CTGAAGATTT TGCAGTTTAT TACTGTCAGC AGCGTAGCAA CTGGCCTCCC ACAGTGATTC CACATGAAAC AAAAACCCCA ACAAGACCAT CAGTGTTTAC TAGATTATTA TACCAGCTGC TTCCTTTACA GACAGCTAGT GGGGTGGCCA CTCAGTGTTA GCATCTCAGC TCTATTTGGC CATTTTGGAG TTCAAGT
321	CEA CDR-H1	see Table 81
322	CEA CDR-H2	see Table 81
323	CEA CDR-H3	see Table 81
324	CEA CDR-L1	see Table 81
325	CEA CDR-L2	see Table 81
326	CEA CDR-L3	see Table 81
327	Parental CEA binder VH	see Table 81
328	Parental CEA binder VL	see Table 81
329	Humanized CEA binder VH	see Table 81
330	Humanized CEA binder VL	see Table 81
331	Nucleotide sequence anti-CEA(T84.66-LCHA) Fc hole chain	see Table 82
332	Nucleotide sequence anti-CEA(T84.66-LCHA) light chain	see Table 82
333	anti- CEA (T84.66-LCHA) Fc hole chain	see Table 82
334	anti- CEA (T84.66-LCHA) light chain	see Table 82
335	Nucleotide sequence anti-CEA(T84.66-LCHA) Fc hole dimeric ligand chain	see Table 84

SEQ ID NO:	Name	Sequence
336	Nucleotide sequence anti-CEA(T84.66-LCHA) Fc knob monomeric ligand	see Table 84
337	anti- CEA(T84.66-LCHA) Fc hole dimeric ligand chain	see Table 84
338	anti- CEA(T84.66-LCHA) Fc knob monomeric ligand	see Table 84
339	Nucleotide sequence anti-CEA (T84.66-LCHA) Fc hole dimeric ligand (71-248) chain	see Table 87
340	Nucleotide sequence anti-CEA (T84.66-LCHA) Fc knob monomeric (71-248) ligand	see Table 87
341	anti- CEA (T84.66-LCHA) Fc hole dimeric ligand (71-248) chain	see Table 87
342	anti- CEA (T84.66-LCHA) Fc knob monomeric (71-248) ligand	see Table 87
343	Nucleotide sequence anti-CEA(T84.66) Fc hole chain	see Table 88
344	Nucleotide sequence anti-CEA(T84.66) light chain	see Table 88
345	anti- CEA (T84.66) Fc hole chain	see Table 88
346	anti- CEA (T84.66) light chain	see Table 88
347	Nucleotide sequence anti-CEA(T84.66) Fc hole dimeric ligand chain	see Table 89
348	Nucleotide sequence anti-CEA(T84.66) Fc knob monomeric ligand	see Table 89
349	anti- CEA(T84.66) Fc hole dimeric ligand chain	see Table 89
350	anti- CEA(T84.66) Fc knob monomeric ligand	see Table 89
351	nucleotide sequence hu NA3B3A2-avi His	see Table 92

SEQ ID NO:	Name	Sequence
352	human NA3B3A2-avi-His	see Table 92
353	Nucleotide sequence Dimeric hu OX40L (51-183) – CL* Fc knob chain	see Table 97
354	Nucleotide sequence Monomeric hu OX40L (51-183) – CH1*	see Table 97
355	Dimeric hu OX40L (51-183) – CL* Fc knob chain	see Table 97
356	Monomeric hu OX40L (51-183) – CH1*	see Table 97
357	CD19 (8B8-2B11) VH	QVQLVQSGAEVKKPGASVKVSCASGYTFTDYIMH WVRQAPGQGLEWMGYINPYNDGSKYTEKFQGRVT MTSDTSISTAYMELSRLRSDDTAVYYCARGTYYYGP QLFDYWGGQTTVTVSS
358	CD19 (8B8-2B11) VL	DIVMTQTPLSLSVTPGQPASISCKSSQSLETSTGTTYL NWY LQKPGQSPQLLIYRVSKRFSGVPDRFSGSGSGTD FTLKISRVEAEDVGVYYCLQLLEDPTYTFGQGTKLEIK
359	CD19 (8B8-7H07) VH	QVQLVQSGAEVKKPGASVKVSCASGYTFTDYIMH WVRQAPGQGLEWMGYINPYNDGSKYTEKFQGRVT MTSDTSISTAYMELSRLRSDDTAVYYCARGTYYYGS ELFDYWGGQTTVTVSS
360	CD19 (8B8-7H07) VL	DIVMTQTPLSLSVTPGQPASISCKSSQSLETSTGN TYL NWY LQKPGQSPQLLIYRVSKRFSGVPDRFSGSGSGTD FTLKISRVEAEDVGVYYCLQATHIPYTFGQGTKLEIK
361	CD19 (8B8-2B03) VH	QVQLVQSGAEVKKPGASVKVSCASGYTFTDYITH WVRQAPGQGLEWMGYINPYNDGSKYTEKFQGRVT MTSDTSISTAYMELSRLRSDDTAVYYCARGTYYYGP DLFDYWGGQTTVTVSS
362	CD19 (8B8-2B03) VL	DIVMTQTPLSLSVTPGQPASISCKSSQSLETSTGN TYL NWY LQKPGQSPQLLIYRVSKRFSGVPDRFSGSGSGTD FTLKISRVEAEDVGVYYCLQLTHVPYTFGQGXKLEIK
363	CD19 (8B8-5A07) VH	QVQLVQSGAEVKKPGASVKVSCASGYTFTDYIMH WVRQAPGQGLEWMGYINPYNDGSKYTEKFQGRVT MTSDTSISTAYMELSRLRSDDTAVYYCARGTYYYGS ALFDYWGGQTTVTVSS
364	CD19 (8B8-5A07) VL	DIVMTQTPLSLSVTPGQPASISCKSSQSLETSTGN TYL NWY LQKPGQSPQLLIYRVSKRFSGVPDRFSGSGSGTD FTLKISRVEAEDVGVYYCLQPGHYPGTFGQGTKLEIK
365	CD19 (8B8-5D08) VH	QVQLVQSGAEVKKPGASVKVSCASGYTFTDYIMH WVRQAPGQGLEWMGYINPYNDGSKYTEKFQGRVT MTSDTSISTAYMELSRLRSDDTAVYYCARGTYYYGS ELFDYWGGQTTVTVSS
366	CD19 (8B8-5D08) VL	DIVMTQTPLSLSVTPGQPASISCKSSQSLETSTGN TYL NWY LQKPGQSPQLLIYRVSKRFSGVPDRFSGSGSGTD FTLKISRVEAEDVGVYYCLQLTHEPYTFGQGTKLEIK
367	CD19 (8B8-5B08) VH	QVQLVQSGAEVKKPGASVKVSCASGYTFTDYIMH

SEQ ID NO:	Name	Sequence
		WVRQAPGQGLEWMGYINPYNDGSKYTEKFQGRVT MTSDTSISTAYMELSRLRSDDTAVYYCARGTYYYGP QLFDYWGGQTTVTVSS
368	CD19 (8B8-5B08) VL	DIVMTQTPLSLSVTPGQPASISCKSSQSLETSTGNTYL NWYLVQKPGQSPQLLIYRVSKRFSGVPDRFSGSGSGTD FTLKISRVEAEDVGVYYCLQLDSYPNTFGQGGTKLEIK
369	CD19 (8B8-5H09) VH	QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYIMH WVRQAPGQGLEWMGYINPYNDGSKYTEKFQGRVT MTSDTSISTAYMELSRLRSDDTAVYYCARGTYYYGS ALFDYWGGQTTVTVSS
370	CD19 (8B8-5H09) VL	DIVMTQTPLSLSVTPGQPASISCKSSQSLESSTGNTYL NWYLVQKPGQSPQLLIYRVSKRFSGVPDRFSGSGSGTD FTLKISRVEAEDVGVYYCLQLIDYPVTFGQGGTKLEIK
371	dimeric huOX40L (51-183) connected by (G4S) ₂ linker	QVSHRYPRIQSIKVQFTEYKKEKGFIILTSQKEDEIMK VQNNSVIINCDFYLISLKGYSQEVNLSLHYQKDEEPL LFQLKKVRSVNSLMVASLTYKDKVYLNVTDDNTSL DDFHVNGGELILIHQNPGEFCVLGGGGSGGGGSQVS HRYPRIQSIKVQFTEYKKEKGFIILTSQKEDEIMKVQN NSVIINCDFYLISLKGYSQEVNLSLHYQKDEEPLFQ LKKVRSVNSLMVASLTYKDKVYLNVTDDNTSLDDF HVNGGELILIHQNPGEFCVL
372	dimeric huOX40L (52-183) connected by (G4S) ₂ linker	VSHRYPRIQSIKVQFTEYKKEKGFIILTSQKEDEIMKV QNNSVIINCDFYLISLKGYSQEVNLSLHYQKDEEPL FQLKKVRSVNSLMVASLTYKDKVYLNVTDDNTSLD DFHVNGGELILIHQNPGEFCVLGGGGSGGGGSVSHR YPRIQSIKVQFTEYKKEKGFIILTSQKEDEIMKVQNN SVIINCDFYLISLKGYSQEVNLSLHYQKDEEPLFQLK KRSVNSLMVASLTYKDKVYLNVTDDNTSLDDFHV NGGELILIHQNPGEFCVL
373	hu 4-1BBL (85-248)	LDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSL TGGLSYKEDTKELVVAAGVYYYVFFQLELRRVVAG EGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSE ARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQ LTQGATVLGLFRVTPEIPAGL
374	hu 4-1BBL (80-248)	DPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGL AGVSLTGGLSYKEDTKELVVAAGVYYYVFFQLELRR VVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPP ASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARAR HAWQLTQGATVLGLFRVTPEIPAGL
375	hu 4-1BBL (52-248)	PWAVSGARASPGSAASPRREGPELSPDDPAGLLDLR QGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGL SYKEDTKELVVAAGVYYYVFFQLELRRVVAGEGSG SVSLALHLQPLRSAAGAAALALTVDLPPASSEARN SAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQ GATVLGLFRVTPEIPAGL

General information regarding the nucleotide sequences of human immunoglobulins light and heavy chains is given in: Kabat, E.A., et al., Sequences of Proteins of Immunological Interest, 5th ed., Public Health Service, National Institutes of Health, Bethesda, MD (1991).

Amino acids of antibody chains are numbered and referred to according to the EU numbering systems according to Kabat (Kabat, E.A., et al., Sequences of Proteins of Immunological Interest, 5th ed., Public Health Service, National Institutes of Health, Bethesda, MD (1991)) as defined above.

EXAMPLES

The following are examples of methods and compositions of the invention. It is understood that various other embodiments may be practiced, given the general description provided above.

Recombinant DNA techniques

5 Standard methods were used to manipulate DNA as described in Sambrook et al.,
Molecular cloning: A laboratory manual; Cold Spring Harbor Laboratory Press, Cold Spring
Harbor, New York, 1989. The molecular biological reagents were used according to the
manufacturer's instructions. General information regarding the nucleotide sequences of human
immunoglobulin light and heavy chains is given in: Kabat, E.A. et al., (1991) Sequences of
10 Proteins of Immunological Interest, Fifth Ed., NIH Publication No 91-3242.

DNA sequencing

DNA sequences were determined by double strand sequencing.

Gene synthesis

Desired gene segments were either generated by PCR using appropriate templates or were
15 synthesized by Geneart AG (Regensburg, Germany) from synthetic oligonucleotides and PCR
products by automated gene synthesis. In cases where no exact gene sequence was available,
oligonucleotide primers were designed based on sequences from closest homologues and the
genes were isolated by RT-PCR from RNA originating from the appropriate tissue. The gene
segments flanked by singular restriction endonuclease cleavage sites were cloned into standard
20 cloning / sequencing vectors. The plasmid DNA was purified from transformed bacteria and
concentration determined by UV spectroscopy. The DNA sequence of the subcloned gene
fragments was confirmed by DNA sequencing. Gene segments were designed with suitable
restriction sites to allow sub-cloning into the respective expression vectors. All constructs were
designed with a 5'-end DNA sequence coding for a leader peptide which targets proteins for
25 secretion in eukaryotic cells.

Cell culture techniques

Standard cell culture techniques were used as described in Current Protocols in Cell
Biology (2000), Bonifacino, J.S., Dasso, M., Harford, J.B., Lippincott-Schwartz, J. and Yamada,
K.M. (eds.), John Wiley & Sons, Inc.

Protein purification

Proteins were purified from filtered cell culture supernatants referring to standard protocols. In brief, antibodies were applied to a Protein A Sepharose column (GE healthcare) and washed with PBS. Elution of antibodies was achieved at pH 2.8 followed by immediate neutralization of the sample. Aggregated protein was separated from monomeric antibodies by size exclusion chromatography (Superdex 200, GE Healthcare) in PBS or in 20 mM Histidine, 150 mM NaCl pH 6.0. Monomeric antibody fractions were pooled, concentrated (if required) using e.g., a MILLIPORE Amicon Ultra (30 MWCO) centrifugal concentrator, frozen and stored at -20°C or -80°C. Part of the samples were provided for subsequent protein analytics and analytical characterization e.g. by SDS-PAGE, size exclusion chromatography (SEC) or mass spectrometry.

SDS-PAGE

The NuPAGE® Pre-Cast gel system (Invitrogen) was used according to the manufacturer's instruction. In particular, 10% or 4-12% NuPAGE® Novex® Bis-TRIS Pre-Cast gels (pH 6.4) and a NuPAGE® MES (reduced gels, with NuPAGE® Antioxidant running buffer additive) or MOPS (non-reduced gels) running buffer was used.

Analytical size exclusion chromatography

Size exclusion chromatography (SEC) for the determination of the aggregation and oligomeric state of antibodies was performed by HPLC chromatography. Briefly, Protein A purified antibodies were applied to a Tosoh TSKgel G3000SW column in 300 mM NaCl, 50 mM $\text{KH}_2\text{PO}_4/\text{K}_2\text{HPO}_4$, pH 7.5 on an Agilent HPLC 1100 system or to a Superdex 200 column (GE Healthcare) in 2 x PBS on a Dionex HPLC-System. The eluted protein was quantified by UV absorbance and integration of peak areas. BioRad Gel Filtration Standard 151–1901 served as a standard.

Mass spectrometry

This section describes the characterization of the multispecific antibodies with VH/VL exchange (VH/VL CrossMabs) with emphasis on their correct assembly. The expected primary structures were analyzed by electrospray ionization mass spectrometry (ESI-MS) of the deglycosylated intact CrossMabs and deglycosylated/plasmin digested or alternatively deglycosylated/limited LysC digested CrossMabs.

The VH/VL CrossMabs were deglycosylated with N-Glycosidase F in a phosphate or Tris buffer at 37°C for up to 17 h at a protein concentration of 1 mg/ml. The plasmin or limited LysC (Roche) digestions were performed with 100 µg deglycosylated VH/VL CrossMabs in a Tris buffer pH 8 at room temperature for 120 hours and at 37°C for 40 min, respectively. Prior to

mass spectrometry the samples were desalted via HPLC on a Sephadex G25 column (GE Healthcare). The total mass was determined via ESI-MS on a maXis 4G UHR-QTOF MS system (Bruker Daltonik) equipped with a TriVersa NanoMate source (Advion).

Determination of binding and binding affinity of multispecific antibodies to the respective antigens using surface plasmon resonance (SPR) (BIAcore)

Binding of the generated antibodies to the respective antigens is investigated by surface plasmon resonance using a BIAcore instrument (GE Healthcare Biosciences AB, Uppsala, Sweden). Briefly, for affinity measurements Goat-Anti-Human IgG, JIR 109-005-098 antibodies are immobilized on a CM5 chip via amine coupling for presentation of the antibodies against the respective antigen. Binding is measured in HBS buffer (HBS-P (10 mM HEPES, 150 mM NaCl, 0.005% Tween 20, pH 7.4), 25°C (or alternatively at 37°C). Antigen (R&D Systems or in house purified) was added in various concentrations in solution. Association was measured by an antigen injection of 80 seconds to 3 minutes; dissociation was measured by washing the chip surface with HBS buffer for 3 - 10 minutes and a KD value was estimated using a 1:1 Langmuir binding model. Negative control data (e.g. buffer curves) are subtracted from sample curves for correction of system intrinsic baseline drift and for noise signal reduction. The respective BIAcore Evaluation Software is used for analysis of sensorgrams and for calculation of affinity data.

Example 1

1.1 Preparation of targeted human 4-1BB ligand trimer-containing Fc fusion antigen binding molecules

Different fragments of the DNA sequence encoding part of the ectodomain (amino acids 71-254, 52-254 and 80-254) of human 4-1BB ligand were synthesized according to the P41273 sequence of Uniprot database (SEQ ID NO:42).

As components for the assembly of a TNF ligand trimer-containing antigen binding molecule a polypeptide comprising two ectodomains of 4-1BB ligand, separated by (G₄S)₂ linkers, and fused to the human IgG1-CH1 or CL domain, was cloned as depicted in Figure 1A- (human 4-1BB ligand, (G₄S)₂ connector, human 4-1BB ligand, (G₄S)₂ connector, human CH1 or CL) or as depicted in Figure 1C (human CH3, (G₄S)₂ connector, human 4-1BB ligand, (G₄S)₂ connector, human 4-1BB ligand).

A polypeptide comprising one ectodomain of 4-1BB ligand and fused to the human IgG1-CL or CH1 domain, was cloned as described in Figure 1B (human 4-1BB ligand, (G₄S)₂ connector, human CL or CH1) or as depicted in Figure 1D (human CH3, (G₄S)₂ connector, human 4-1BB ligand).

The polypeptides were subcloned in frame with the human IgG1 heavy chain CH2 and CH3 domains with optional peptide linkers, for example for construct 1 the polypeptide encoding the dimeric 4-1BB ligand fused to a human CH1 domain was subcloned in frame with the human IgG1 heavy chain CH2 and CH3 domains on the knob (Merchant, Zhu et al. 1998) using a linker (G₄S)₂ of SEQ ID NO:13 or (GSPGSSSSGS) of SEQ ID NO:57.

The variable region of heavy and light chain DNA sequences encoding a binder specific for fibroblast activation protein (FAP), i.e. 28H1, were subcloned in frame with either the constant heavy chain of the hole (Carter, J. Immunol. Methods (2001), 248, 7-15) or the constant light chain of human IgG1. The generation and preparation of the FAP binders is described in WO 2012/020006 A2, which is incorporated herein by reference.

Table 1 summarizes the characteristics of the constructs produced. The constructs 1 to 10 differ in their geometry, valency for FAP, 4-1BB ligand ectodomain, crossover of the CH1 and CL domain (CrossMab technology), mutations in the CH1 and CL domains and different peptide linkers in the polypeptide comprising one ectodomain of 4-1BB ligand (monomeric 4-1BBL chain).

Table 1: Characteristics of produced TNF ligand trimer-containing antigen binding molecules (FAP split 4-1BBL trimers)

Construct	Valency for FAP	FAP binder	4-1BBL ectodomain	Crossed CH1-CL domains	Charged residues	Linker to 4-1BBL in light chain
1.1	monovalent	28H1	71-254	no	no	(G4S) ₂
1.2	monovalent	28H1	71-254	yes (Ligand)	yes (Ligand)	(G4S) ₂
1.3	monovalent	28H1	71-254	yes (FAP Fab)	yes (Ligand)	(G4S) ₂
1.4	monovalent	28H1	71-254	no	yes (Ligand)	(G4S) ₂
1.5	bivalent	28H1	71-254	no	no	(G4S) ₂
1.6	monovalent	28H1	71-254	no	yes (Ligand)	(G4S) ₁
1.7	bivalent	28H1	71-254	yes (Ligand)	yes (Ligand)	(G4S) ₂
1.8	bivalent	28H1	71-254	yes (FAP Fab fused to Ligand)	yes (Ligand)	(G4S) ₂
1.9	monovalent	28H1	52-254	no	yes (Ligand)	(G4S) ₂
1.10	monovalent	28H1	80-254	no	yes (Ligand)	(G4S) ₂

In order to avoid mispairing, in most of the constructs one pair of CH1 and CL domains was replaced by each other (domain crossover) as described in WO 2009/080253 A1.

To further improve correct pairing, different charged amino acid substitutions were introduced in the crossed or non-crossed CH1 and CL domains as charged residues in constructs 2 to 4 and 6 to 10. In the human CL domain the mutations E123R and Q124K were introduced, whereas the mutations K147E and K213E were cloned into the human CH1 domain.

For all constructs the knobs into holes heterodimerization technology was used with the S354C/T366W mutations in the CH3 domain of the knob chain and the corresponding Y349C/T366S/L368A/Y407V mutations in the CH3 domain of the hole chain (Carter, J Immunol Methods 248, 7-15 (2001)).

The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in International Patent Appl. Publ. No. WO 2012/130831 A1.

For example, in construct 1 the combination of the ligand-Fc knob chain containing the S354C/T366W mutations in the first CH3 domain, with the targeted anti-FAP-Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations in the second CH3 domain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a FAP binding Fab (Figure 2, Graph 1.1).

Table 2 shows the cDNA and amino acid sequences of the monovalent FAP-targeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecule (Construct 1.1).

Table 2: Sequences of FAP-targeted human 4-1BB ligand trimer containing Fc (kih) fusion molecule Construct 1.1

SEQ ID NO:	Description	Sequence
66	Dimeric hu 4-1BBL (71-254) – CH1 Fc knob chain	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAACTGGTGG TGGCCAAGGCCGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGCCCGGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCTG

		<p> TTCAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCT CCAAGAAGCGAAGGCGGAGGCGGATCTGGCGGCGGAG GATCTAGAGAGGGACCCGAACGTGCCCCGACGATCCA GCCGGGCTGCTGGATCTGAGACAGGGAATGTTGCCCA GCTGGTGGCTCAGAATGTGCTGCTGATTGACGGACCTCT GAGCTGGTACTCCGACCCAGGGCTGGCAGGGGTGTCCC TGACTGGGGGACTGTCCTACAAAGAAGATACAAAAGAA CTGGTGGTGGCTAAAGCTGGGGTGTACTATGTGTTTTT CAGCTGGAAGTGAAGCGGGTGGTGGCTGGGGAGGGCTC AGGATCTGTGTCCCTGGCTCTGCATCTGCAGCCACTGCG CTCTGCTGCTGGCGCAGCTGCACTGGCTCTGACTGTGGA CCTGCCACCAGCCTCTAGCGAGGCCAGAAACAGCGCCT TCGGGTTCGAAGGACGCCTGCTGCATCTGAGCGCCGGAC AGCGCCTGGGAGTGCATCTGCATACTGAAGCCAGAGCC CGGCATGCTTGGCAGCTGACTCAGGGGGCAACTGTGCTG GGACTGTTTCGCGTGACACCTGAGATCCCTGCCGGACTG CCAAGCCCTAGATCAGAAGGGGGCGGAGGAAGCGGAG GGGGAGGAAGTGCTAGCACCAAGGGCCCTAGCGTGTTT CCTCTGGCCCCCTAGCAGCAAGAGCACAAGTGGAGGAAC AGCCGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGA GCCCCGTGACCGTGTCTGGAATTCTGGCGCCCTGACAAG CGGCGTGCACACATTTCCAGCCGTGCTGCAGAGCAGCG GCCTGTACTCTCTGAGCAGCGTCGTGACCGTGCCCTCTA GCTCTCTGGGCACCCAGACCTACATCTGCAACGTGAACC ACAAGCCCAGCAACACCAAAGTGGACAAGAAGGTGGA ACCCAAGAGCTGCGACAAGACCCACACCTGTCCCCCTTG CCCTGCCCCCTGAAGCTGCTGGTGGCCCTTCCGTGTTCT GTTCCCCCCTAAAGCCCAAGGACACCCTGATGATCAGCC GGACCCCCGAAGTGAAGTGAAGTTCAATTGGTACGTGGAC ACGAGGACCCTGAAGTGAAGTTCAATTGGTACGTGGAC GGCGTGGAAGTGCACAATGCCAAGACCAAGCCGCGGGA GGAGCAGTACAACAGCACGTACCGTGTGGTCAGCGTCC TCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGGAG TACAAGTGCAAGGTCTCCAACAAAGCCCTCGGCGCCCC CATCGAGAAAACCATCTCCAAGCCAAAGGGCAGCCCC GAGAACCACAGGTGTACACCCTGCCCCCATGCCGGGAT GAGCTGACCAAGAACCAGGTGAGCCTGTGGTGCCTGGT CAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGG AGAGCAATGGGCAGCCGGAGAACAACTACAAGACCACG CCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCTCTACA GCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGG AACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCAC AACCCTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGT AAA </p>
67	Monomeric hu 4-1BBL (71-254) - CL	<p> AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCTTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAAGTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGCCCGGAATAGCGCATTGGGTTT </p>

		CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGTGGGCCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCT CCAAGAAGCGAAGGCGGAGGCGGATCTGGCGGCGGAG GATCTCGTACGGTGGCTGCACCATCTGTCTTCATCTTCCC GCCATCTGATGAGCAGTTGAAATCTGGAAGTGCCTCTGT TGTGTGCCTGCTGAATAACTTCTATCCCAGAGAGGCCAA AGTACAGTGGAAGGTGGATAACGCCCTCCAATCGGGTA ACTCCCAGGAGAGTGTACAGAGCAGGACAGCAAGGAC AGCACCTACAGCCTCAGCAGCACCCCTGACGCTGAGCAA AGCAGACTACGAGAAACACAAAGTCTACGCCTGCGAAG TCACCCATCAGGGCCTGAGCTCGCCCGTCACAAAGAGCT TCAACAGGGGAGAGTGT
68	anti-FAP Fc hole chain	GAAGTGCAGCTGCTGGAATCCGGCGGAGGCCTGGTGCA GCCTGGCGGATCTCTGAGACTGTCTTGCGCCGCTCCGG CTTCACCTTCTCCTCCCACGCCATGTCTGGGTCCGACA GGCTCCTGGCAAAGGCCTGGAATGGGTGTCCGCCATCTG GGCCTCCGGCGAGCAGTACTACGCCGACTCTGTGAAGG GCCGGTTCACCATCTCCCGGGACAACCTCCAAGAACACCC TGTACCTGCAGATGAACTCCCTGCGGGCCGAGGACACC GCCGTGTACTACTGTGCCAAGGGCTGGCTGGGCAACTTC GACTACTGGGGACAGGGCACCCCTGGTCACCGTGTCCAG CGCTAGCACCAAGGGCCCCCTCCGTGTTCCCCCTGGCCCC CAGCAGCAAGAGCACCAGCGGCGGCACAGCCGCTCTGG GCTGCCTGGTCAAGGACTACTTCCCCGAGCCCGTGACCG TGTCTTGAACAGCGGAGCCCTGACCTCCGGCGTGCAC ACCTTCCCCGCCGTGCTGCAGAGTTCTGGCCTGTATAGC CTGAGCAGCGTGGTCACCGTGCCTTCTAGCAGCCTGGGC ACCCAGACCTACATCTGCAACGTGAACCACAAGCCCAG CAACACCAAGGTGGACAAGAAGGTGGAGCCCAAGAGCT GCGACAAAACCTCACACATGCCACCGTGCCAGCACCT GAAGCTGCAGGGGGACCGTCAGTCTTCCCTCTTCCCCCA AAACCAAGGACACCCTCATGATCTCCCGGACCCCTGA GGTCACATGCGTGGTGGTGGACGTGAGCCACGAAGACC CTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGAG GTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTA CAACAGCACGTACCGTGTGGTCAGCGTCCTACCGTCCCT GCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCA AGGTCTCCAACAAAGCCCCTCGGCGCCCCCATCGAGAAA ACCATCTCCAAGCCAAAGGGCAGCCCCGAGAACCACA GGTGTGCACCCTGCCCCCATCCCGGGATGAGCTGACCAA GAACCAGGTCAGCCTCTCGTGCGCAGTCAAAGGCTTCTA TCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGC AGCCGGAGAACAACTACAAGACCACGCCTCCCGTGCTG GACTCCGACGGCTCCTTCTTCTCGTGAGCAAGCTCACC GTGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTC ATGCTCCGTGATGCATGAGGCTCTGCACAACCACTACAC GCAGAAGAGCCTCTCCCTGTCTCCGGGTAAA
69	anti-FAP light chain	GAGATCGTGCTGACCCAGTCCCCCGGCACCCTGTCTCTG AGCCCTGGCGAGAGAGCCACCCTGTCTGAGAGCCTC CCAGTCCGTGTCCCGGTCCTACCTCGCCTGGTATCAGCA

		GAAGCCCGGCCAGGCCCTCGGCTGCTGATCATCGGCG CCTCTACCAGAGCCACCGGCATCCCTGACCGGTTCTCCG GCTCTGGCTCCGGCACC GACTTCACCCTGACCATCTCCC GGCTGGAACCCGAGGACTTCGCCGTGTACTACTGCCAGC AGGGCCAGGTCATCCCTCCCACCTTTGGCCAGGGCACCA AGGTGGAAATCAAGCGTACGGTGGCTGCACCATCTGTCT TCATCTTCCCGCCATCTGATGAGCAGTTGAAATCTGGAA CTGCCTCTGTTGTGTGCCTGCTGAATAACTTCTATCCCAG AGAGGCCAAAGTACAGTGAAGGTGGATAACGCCCTCC AATCGGGTAACTCCCAGGAGAGTGTACAGAGCAGGAC AGCAAGGACAGCACCTACAGCCTCAGCAGCACCCCTGAC GCTGAGCAAAGCAGACTACGAGAAACACAAAGTCTACG CCTGCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCA CAAAGAGCTTCAACAGGGGAGAGTGT
14	Dimeric hu 4- 1BBL (71-254) – CH1 Fc knob chain	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLPSRSEGGGSGGGGSREGPEL SPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLA GVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELRRVVAG EGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARN SAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATV LGLFRVTPEIPAGLPSRSEGGGSGGGGSASTKGPSVFPLA PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHT FPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTK VDKKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPPKPKDTL MISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKTK PREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALG APIEKTISKAKGQPREPQVYTLPPCRDELTKNQVSLWCLVK GFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLT VDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK
15	Monomeric hu 4- 1BBL (71-254) – CL1	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLPSRSEGGGSGGGGSRTVAAP SVFIFPPSDEQLKSGTASVCLLNNFYPPREAKVQWKVDNA LQSGNSQESVTEQDSKSTYSLSSTLTLSKADYEKHKVYA CEVTHQGLSSPVTKSFNRGEC
18	anti-FAP(28H1) Fc hole chain	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSHAMSWVRQA PGKGLEWVSAIWASGEQYYADSVKGRFTISRDN SKNTLYL QMNSLR AEDTAVYYCAKGWLG NFDYW GQGTLTVSSAS TKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNS GALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICN VNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVF LFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDG VEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYK CKVSNKALGAPIEKTISKAKGQPREPQVCTLPSPRDELTKN QVSLSCAVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD GSFFLVSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKS

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		LSLSPGK
19	anti-FAP (28H1) light chain	EIVLTQSPGTLSSLSPGERATLSCRASQSVSRSYLAWYQQKP GQAPRLLIIGASTRATGIPDRFSGSGSGTDFTLTISRLEPEDF AVYYCQQGQVIPPTFGQGTKVEIKRTVAAPSVFIFPPSDEQ LKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESV TEQDSKSTYSLSSITLTLKADYEKHKVYACEVTHQGLSS PVTKSFNRGEC

Table 3 shows the cDNA and amino acid sequences of acid sequences of the monovalent FAP-targeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecule (Construct 1.2) with CH1-CL crossover and charged residues.

5 **Table 3: Sequences of FAP-targeted human 4-1BB ligand trimer containing Fc (kih) fusion molecule Construct 1.2**

SEQ ID NO:	Description	Sequence
129	Dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCAGTACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAACTGGTGG TGGCCAAGGCCGCGGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGCCCGGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGTGGGCCCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCT CCAAGAAGCGAAGGCGGAGGCGGATCTGGCGGCGGAG GATCTAGAGAGGGACCCGAAGTGTCCCTGACGATCCA GCCGGGCTGCTGGATCTGAGACAGGGAATGTTCCGCCA GCTGGTGGCTCAGAAATGTGCTGCTGATTGACGGACCTCT GAGCTGGTACTCCGACCCAGGGCTGGCAGGGGTGTCCC TGACTGGGGGACTGTCCTACAAAGAAGATACAAAAGAA CTGGTGGTGGCTAAAGCTGGGGTGTACTATGTGTTTTT CAGCTGGAAGTGAAGGCGGGTGGTGGCTGGGGAGGGCTC AGGATCTGTGTCCCTGGCTCTGCATCTGCAGCCACTGCG CTCTGCTGCTGGCGCAGCTGCACTGGCTCTGACTGTGGA CCTGCCACCAGCCTCTAGCGAGGCCAGAAACAGCGCCT TCGGGTTCGAAGGACGCCTGCTGCATCTGAGCGCCGGAC AGCGCCTGGGAGTGCATCTGCATACTGAAGCCAGAGCC CGGCATGCTTGGCAGCTGACTCAGGGGGCAACTGTGCTG GGACTGTTTCGCGTGACACCTGAGATCCCTGCCGGACTG CCAAGCCCTAGATCAGAAGGGGGCGGAGGTTCCGGAGG GGGAGGATCTCGTACGGTGGCTGCACCATCTGTCTTTAT

		<p>CTTCCCACCCAGCGACCGGAAGCTGAAGTCTGGCACAG CCAGCGTCGTGTGCCTGCTGAATAACTTCTACCCCCGCG AGGCCAAGGTGCAGTGGAAGGTGGACAATGCCCTGCAG AGCGGCAACAGCCAGGAAAGCGTGACCGAGCAGGACA GCAAGGACTCCACCTACAGCCTGAGCAGCACCCCTGACC CTGAGCAAGGCCGACTACGAGAAGCACAAGGTGTACGC CTGCGAAGTGACCCACCAGGGCCTGTCTAGCCCCGTGAC CAAGAGCTTCAACCGGGGCGAGTGCGACAAGACCCACA CCTGTCTTCCATGCCCTGCCCTGAAGCTGCTGGCGGCC CTAGCGTGTTCCTGTTCCCCCAAAGCCCAAGGACACCC TGATGATCAGCCGGACCCCTGAAGTGACCTGCGTGGTGG TGGATGTGTCCACAGAGGACCCTGAAGTGAAGTTCAATT GGTACGTGGACGGCGTGGAAGTGCACAATGCCAAGACC AAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGT GGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAA TGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCC TCGGCGCCCCATCGAGAAAACCATCTCCAAGCCAAA GGGCAGCCCCGAGAACCACAGGTGTACACCCTGCCCCC ATGCCGGGATGAGCTGACCAAGAACCAGGTCAGCCTGT GGTGCCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCG TGGAGTGGGAGAGCAATGGGCAGCCGGAGAACAACTAC AAGACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTC TTCCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTG GCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGA GGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCT GTCTCCGGGTAAA</p>
130	Monomeric hu 4- 1BBL (71-254) – CH1*	<p>AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCTTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAACTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCTCTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGCCCGGAATAGCGCATTTGGGTTT CAAGGCCGGCTGCTGCACCTGTCTGCCGGCCAGAGACT GGGAGTGCATCTGCACACAGAGGCCAGAGCCAGGCACG CCTGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCTG TTCAGAGTGACCCCCGAGATTCTTGCCGGCCTGCCTAGC CCTAGATCTGAAGGCGGCGGAGGTTCCGGAGGCGGAGG ATCTGCTAGCACAAAGGGCCCCAGCGTGTTCCCTCTGGC CCCTAGCAGCAAGAGCACATCTGGCGGAACAGCCGCC TGGGCTGCCTGGTGGGAAGATTACTTCCCCGAGCCCGTGA CCGTGTCCTGGAATTCTGGCGCCCTGACAAGCGGCGTGC ACACCTTTCCAGCCGTGCTGCAGAGCAGCGGCCTGTACT CTCTGAGCAGCGTCGTGACAGTGCCCAGCAGCTCTCTGG GCACCCAGACCTACATCTGCAACGTGAACCACAAGCCC AGCAACACCAAGGTGGACGAGAAGGTGGAACCCAAGTC CTGC</p>
68	anti-FAP Fc hole chain	See Table 2

69	anti-FAP light chain	See Table 2
115	Dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSEGGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSEGGGGSGGGGSRTVAAPSVFIFP PSDRKLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGN SQESVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTH QGLSSPVTKSFNRGECDKTHTCPPCPAPEAAGGPSVFLFPP KPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEV HNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKV SNKALGAPIEKTISKAKGQPREPQVYTLPPCRDELTKNQVSLWCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKLTVDKSRWQQGNVVFSCSVMEALHNHYTQKSLSLSPGK
116	Monomeric hu 4-1BBL (71-254) – CH1*	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSEGGGGSGGGGSASTKGPSVFPLAPSSKSTSGGTAALGCLVEDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDEKVEPKSC
18	anti-FAP(28H1) Fc hole chain	See Table 2
19	anti-FAP (28H1) light chain	See Table 2

Table 4 shows the cDNA and amino acid sequences of the monovalent FAP-targeted 4-1BB ligand trimer-containing Fc (kih) fusion molecule Construct 1.3 (FAP split trimer with CH1-CL crossover in anti-FAP Fab and charged residues on the 4-1BBL containing chains).

Table 4: Sequences of FAP-targeted human 4-1BB ligand trimer containing Fc (kih) fusion molecule Construct 1.3

SEQ ID NO:	Description	Sequence
131	Dimeric hu 4-1BBL (71-254) – CH1* Fc knob chain	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAAGTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGCCCCGGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGTGGGCCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCT CCAAGAAGCGAAGGCGGAGGCGGATCTGGCGGCGGAG GATCTAGAGAGGGACCCGAAGTGTCCCCTGACGATCCA GCCGGGCTGCTGGATCTGAGACAGGGAATGTTGCCCA GCTGGTGGCTCAGAATGTGCTGCTGATTGACGGACCTCT GAGCTGGTACTCCGACCCAGGGCTGGCAGGGGTGTCCC TGACTGGGGGACTGTCCTACAAAGAAGATACAAAAGAA CTGGTGGTGGCTAAAGCTGGGGTGTACTATGTGTTTTTT CAGCTGGAACTGAGGCGGGTGGTGGCTGGGGAGGGCTC AGGATCTGTGTCCCTGGCTCTGCATCTGCAGCCACTGCG CTCTGCTGCTGGCGCAGCTGCACTGGCTCTGACTGTGGA CCTGCCACCAGCCTCTAGCGAGGCCAGAAACAGCGCCT TCGGGTTCGAAGGACGCCTGCTGCATCTGAGCGCCGGAC AGCGCCTGGGAGTGCATCTGCATACTGAAGCCAGAGCC CGGCATGCTTGGCAGCTGACTCAGGGGGCAACTGTGCTG GGACTGTTTCGCGTGACACCTGAGATCCCTGCCGGACTG CCAAGCCCTAGATCAGAAGGGGGCGGAGGAAGCGGAG GGGGAGGAAGTGCTAGCACCAAGGGCCCCCTCCGTGTTC CCCCTGGCCCCCAGCAGCAAGAGCACCAGCGGCGGCAC AGCCGCTCTGGGCTGCCTGGTTCGAGGACTACTTCCCCGA GCCCCGTGACCGTGTCTGGAACAGCGGAGCCCTGACCTC CGGCGTGCACACCTTCCCCGCCGTGCTGCAGAGTTCTGG CCTGTATAGCCTGAGCAGCGTGGTCACCGTGCCTTCTAG CAGCCTGGGCACCCAGACCTACATCTGCAACGTGAACC ACAAGCCCAGCAACACCAAGGTGGACGAGAAGGTGGA GCCCAAGAGCTGCGACAAAACCTCACACATGCCACCGT GCCCAGCACCTGAAGCTGCAGGGGGGACCGTCAGTCTTC CTCTTCCCCCCTAAAACCCAAGGACACCCTCATGATCTCC CGGACCCCTGAGGTACATGCGTGGTGGTGGACGTGAG CCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGTGG ACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGG GAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGCGT CCTCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGG AGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGGCGCC

		CCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCAGCC CCGAGAACCACAGGTGTACACCCTGCCCCCATGCCGGG ATGAGCTGACCAAGAACCAGGTCAGCCTGTGGTGCCTG GTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGG GAGAGCAATGGGCAGCCGAGAAACAACACTACAAGACCAC GCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCTCTAC AGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGGG GAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCA CAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGG TAAA
132	Monomeric hu 4-1BBL (71-254) – CL*	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAAGACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAAGTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGCCCGGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGTGGGCGCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCT CCAAGAAGCGAAGGCGGAGGCGGATCTGGCGGCGGAG GATCTCGTACGGTGGCTGCACCATCTGTCTTCATCTTCCC GCCATCTGATCGGAAGTTGAAATCTGGAAGTGCCTCTGT TGTGTGCCTGCTGAATAACTTCTATCCCAGAGAGGCCAA AGTACAGTGGAAAGGTGGATAACGCCCTCCAATCGGGTA ACTCCCAGGAGAGTGTACAGAGCAGGACAGCAAGGAC AGCACCTACAGCCTCAGCAGCACCCCTGACGCTGAGCAA AGCAGACTACGAGAAACACAAAGTCTACGCCTGCGAAG TCACCCATCAGGGCCTGAGCTCGCCCCGTCACAAAGAGCT TCAACAGGGGAGAGTGT
133	anti-FAP (VHCL) (28H1) Fc hole chain	GAAGTGCAGCTGCTGGAATCCGGCGGAGGCCTGGTGCA GCCTGGCGGATCTCTGAGACTGTCTTGCGCCGCTCCGG CTTCACCTTCTCCTCCCACGCCATGTCTGGGTCCGACA GGCTCCTGGCAAAGGCCTGGAATGGGTGTCCGCCATCTG GGCTCCGGCGAGCAGTACTACGCCGACTCTGTGAAGG GCCGGTTACCATCTCCCGGGACAACCTCCAAGAACACCC TGTACCTGCAGATGAACTCCCTGCGGGCCGAGGACACC GCCGTGTACTACTGTGCCAAGGGCTGGCTGGGCAACTTC GACTACTGGGGACAGGGCACCCCTGGTCACCGTGTCCAG CGCTAGCGTGGCCGCTCCCAGCGTGTTCATCTTCCCACC CAGCGACGAGCAGCTGAAGTCCGGCACAGCCAGCGTGG TGTGCCTGCTGAACAACCTTCTACCCCCGCGAGGCCAAGG TGCAGTGGAAAGGTGGACAACGCCCTGCAGAGCGGCAAC AGCCAGGAATCCGTGACCGAGCAGGACAGCAAGGACTC CACCTACAGCCTGAGCAGCACCCCTGACCCCTGAGCAAGG CCGACTACGAGAAGCACAAGGTGTACGCCTGCGAAGTG ACCCACCAGGGCCTGTCCAGCCCCGTGACCAAGAGCTTC AACCGGGGCGAGTGCACAAAGACCCACACCTGTCCCCC TTGCCCTGCCCCCTGAAGCTGCTGGTGGCCCTTCCGTGTT

		CCTGTTCCCCCAAAGCCCAAGGACACCCTGATGATCAG CCGGACCCCCGAAGTGACCTGCGTGGTGGTCGATGTGTC CCACGAGGACCCTGAAGTGAAGTTCAATTGGTACGTGG ACGGCGTGGAAGTGCACAATGCCAAGACCAAGCCGCGG GAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGCGT CCTCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGG AGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGGCGCC CCCATCGAGAAAACCATCTCCAAGCCAAAGGGCAGCC CCGAGAACCACAGGTGTGCACCCTGCCCCCATCCCGGG ATGAGCTGACCAAGAACCAGGTCAGCCTCTCGTGCACA GTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGG GAGAGCAATGGGCAGCCGGAGAACAACACTACAAGACCAC GCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCTCGT GAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGG GGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGC ACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGG GTAAG
134	anti-FAP (VLCH1) (28H1) light chain	GAGATCGTGCTGACCCAGTCTCCCGGCACCCTGAGCCTG AGCCCTGGCGAGAGAGCCACCCTGAGCTGCAGAGCCAG CCAGAGCGTGAGCCGGAGCTACCTGGCCTGGTATCAGC AGAAGCCCCGGCCAGGCCCCCAGACTGCTGATCATCGGC GCCAGCACCCGGGCCACCGGCATCCCCGATAGATTGAG CGGCAGCGGCTCCGGCACCGACTTCACCCTGACCATCAG CCGGCTGGAACCCGAGGACTTCGCCGTGTACTACTGCCA GCAGGGCCAGGTGATCCCCCCCCACCTTCGGCCAGGGCA CCAAGGTGGAATCAAGAGCAGCGCTTCCACCAAAGGC CCTTCCGTGTTTCTCTGGCTCCTAGCTCCAAGTCCACCT CTGGAGGCACCGCTGCTCTCGGATGCCTCGTGAAGGATT ATTTTCTGAGCCTGTGACAGTGTCTGGAATAGCGGAG CACTGACCTCTGGAGTGCATACTTTCCCCGCTGTGCTGC AGTCTCTGGACTGTACAGCCTGAGCAGCGTGGTGACAG TGCCCAGCAGCAGCCTGGGCACCCAGACCTACATCTGC AACGTGAACCACAAGCCCAGCAACACCAAGGTGGACAA GAAGGTGGAACCCAAGTCTTGT
108	Dimeric hu 4- 1BBL (71-254) – CH1* Fc knob chain	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAAGVYYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLPSRSEGGGGSGGGGSREGPEL SPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLA GVSLTGGLSYKEDTKELVVAAGVYYYVFFQLELRRVVAG EGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARN AFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATV LGLFRVTPEIPAGLPSRSEGGGGSGGGGSASTKGPSVFPLA PSSKSTSGGTAALGCLVEDYFPEPVTVSWNSGALTSGVHTF PAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKV DEKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPPKPKDTLM ISRTPETCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKP REEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGA PIEKTISKAKGQPREPQVYTLPPCRDELTKNQVSLWCLVKG FYPSTDAVEWESNGQPENNYKTPPVLDSDGSFFLYSKLTV DKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK

109	Monomeric hu 4-1BBL (71-254) – CL*	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLPSPRSEGGGGSGGGGSRTVAAP SVFIFPPSDRKLKSGTASVVCLLNNFYPREAKVQWKVDNA LQSGNSQESVTEQDSKDYSLSTLTLSKADYEKHKVYA CEVTHQGLSSPVTKSFNRGEC
135	anti-FAP (VHCL) (28H1) Fc hole chain	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSHAMSWVRQA PGKGLEWVSAIWASGEQYYADSVKGRFTISRDNKNTLYL QMNSLRAEDTAVYYCAKGWLGNFYWGQGTILVTVSSAS VAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWK VDNALQSGNSQESVTEQDSKDYSLSTLTLSKADYEKHK KVYACEVTHQGLSSPVTKSFNRGECDKTHTCPPCPAPEAA GGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFN WYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWL NGKEYCKCKVSNKALGAPIEKTISKAKGQPREPQVCTLPSSR DELTKNQVSLSCAVKGFYPSDIAVEWESNGQPENNYKTT PVLDSDGSFFLVSKLTVDKSRWQQGNVFCFSVMHEALHN HYTQKSLSLSPGK
136	anti-FAP (VLCH1) (28H1) light chain	EIVLTQSPGTLISLSPGERATLSCRASQSVSRSYLAWYQQK PQAPRLLIIGASTRATGIPDRFSGSGSGTDFLTISRLEPEDF AVYYCQQGQVIPPTFGQGTKVEIKSSASTKGPSVFPLAPSS KSTSGGTAAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPA VLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDK KVEPKSC

Table 5 shows the cDNA and amino acid sequences of the monovalent FAP-targeted 4-1BB ligand trimer-containing Fc (kih) fusion molecule Construct 1.4 (FAP split trimer with anti-FAP Fab, monomeric 4-1BB ligand fused to CH1-knob chain and charged residues on the 4-1BBL containing chains).

Table 5: Sequences of FAP-targeted human 4-1BB ligand trimer containing Fc (kih) fusion molecule Construct 1.4

SEQ ID NO:	Description	Sequence
137	Monomeric hu 4-1BBL (71-254) – CH1* Fc knob chain	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAAGTGGTGG TGGCCAAGGCCGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCTCTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGCCCGGAATAGCGCATTTGGGTTT CAAGGCCGGCTGCTGCACCTGTCTGCCGGCCAGAGACT

		GGGAGTGCATCTGCACACAGAGGCCAGAGCCAGGCACG CCTGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCTG TTCAGAGTGACCCCCGAGATTCTGCGCGCCTGCCTAGC CCTAGATCTGAAGGCGGCGGAGGTTCCGGAGGCGGAGG ATCTGCTAGCACCAAGGGCCCCCTCCGTGTTCCCCCTGGC CCCCAGCAGCAAGAGCACCAAGCGGCGGCACAGCCGCTC TGGGCTGCCTGGTCGAGGACTACTTCCCCGAGCCCGTGA CCGTGTCCTGGAACAGCGGAGCCCTGACCTCCGGCGTGC ACACCTTCCCCGCCGTGCTGCAGAGTTCTGGCCTGTATA GCCTGAGCAGCGTGGTCACCGTGCCTTCTAGCAGCCTGG GCACCCAGACCTACATCTGCAACGTGAACCACAAGCCC AGCAACACCAAGGTGGACGAGAAGGTGGAGCCCAAGA GCTGCGACAAAACCTCACACATGCCACCGTGCCAGCA CCTGAAGCTGCAGGGGGACCGTCAGTCTTCTCTTCCCC CCAAAACCCAAGGACACCCCTCATGATCTCCCGGACCCCT GAGGTCACATGCGTGGTGGTGGACGTGAGCCACGAAGA CCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGG AGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCAG TACAACAGCACGTACCGTGTGGTCAGCGTCCTACCGTC CTGCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTG CAAGGTCTCCAACAAAGCCCTCGGCGCCCCCATCGAGA AAACCATCTCCAAAGGCCAAAGGGCAGCCCCGAGAACCA CAGGTGTACACCTGCCCCCATGCCGGGATGAGCTGACC AAGAACCAGGTCAGCCTGTGGTGCCTGGTCAAAGGCTTC TATCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGG GCAGCCGGAGAACAACCTACAAGACCACGCCTCCCGTGC TGGACTCCGACGGCTCCTTCTTCTCTACAGCAAGCTCA CCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTC TCATGCTCCGTGATGCATGAGGCTCTGCACAACCACTAC ACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAA
138	Dimeric hu 4- 1BBL (71-254) – CL*	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAAGTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGCCCCGGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCT CCAAGAAGCGAAGGCGGAGGCGGATCTGGCGGCGGAG GATCTAGAGAGGGACCCGAAGTGTCCCCTGACGATCCA GCCGGGCTGCTGGATCTGAGACAGGGAATGTTCCGCCA GCTGGTGGCTCAGAATGTGCTGCTGATTGACGGACCTCT GAGCTGGTACTCCGACCCAGGGCTGGCAGGGGTGTCCC TGACTGGGGGACTGTCCTACAAAGAAGATACAAAAGAA CTGGTGGTGGCTAAAGCTGGGGTGTACTATGTGTTTTT CAGCTGGAAGTGAAGCGGGTGGTGGCTGGGGAGGGCTC AGGATCTGTGTCCCTGGCTCTGCATCTGCAGCCACTGCG CTCTGCTGCTGGCGCAGCTGCACTGGCTCTGACTGTGGA

		CCTGCCACCAGCCTCTAGCGAGGCCAGAAACAGCGCCT TCGGGTTCCAAGGACGCCTGCTGCATCTGAGCGCCGGAC AGCGCCTGGGAGTGCATCTGCATACTGAAGCCAGAGCC CGGCATGCTTGGCAGCTGACTCAGGGGGCAACTGTGCTG GGACTGTTTCGCGTGACACCTGAGATCCCTGCCGGACTG CCAAGCCCTAGATCAGAAGGGGGCGGAGGTTCCGGAGG GGGAGGATCTCGTACGGTGGCTGCACCATCTGTCTTCAT CTTCCCGCCATCTGATCGGAAGTTGAAATCTGGAAGTGC CTCTGTTGTGTGCCTGCTGAATAACTTCTATCCCAGAGA GGCCAAAGTACAGTGGAAGGTGGATAACGCCCTCCAAT CGGGTAACTCCCAGGAGAGTGTACAGAGCAGGACAGC AAGGACAGCACCTACAGCCTCAGCAGCACCCCTGACGCT GAGCAAAGCAGACTACGAGAAACACAAAGTCTACGCCT GCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCACA AAGAGCTTCAACAGGGGAGAGTGT
68	anti-FAP (28H1) Fc hole chain	see Table 2
69	anti-FAP (28H1) light chain	See Table 2
139	Monomeric hu 4- 1BBL (71-254) – CL* Fc knob chain	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLPSRSEGGGGSGGGGSREGPEL SPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLA GVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELRRVVAG EGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARN AFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATV LGLFRVTPEIPAGLPSRSEGGGGSGGGGSASTKGPSVFPLA PSSKSTSGGTAALGCLVEDYFPEPVTVSWNSGALTSGVHTF PAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKV DEKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPPKPKDTLM ISRTPETCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKP REEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGA PIEKTISKAKGQPREPQVYTLPPCRDELTKNQVSLWCLVKG FYPDIQAVESWESNGQPENNYKTTTPVLDSDGSFFLYSKLTV DKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK
140	Dimeric hu 4- 1BBL (71-254) – CL*	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLPSRSEGGGGSGGGGSREGPEL SPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLA GVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELRRVVAG EGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARN AFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATV LGLFRVTPEIPAGLPSRSEGGGGSGGGGSRTVAAPSVFIFP PSDRKLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGN SQESVTEQDSKSTYSLSSSTLTLSKADYEKHKVYACEVTH

		QGLSSPVTKSFNRGEC
18	anti-FAP (28H1) Fc hole chain	see Table 2
19	anti-FAP (28H1) light chain	see Table 2

Table 6 shows the cDNA and amino acid sequences of the bivalent FAP-targeted 4-1BB ligand trimer-containing Fc (kih) fusion molecule Construct 1.5 (FAP split trimer with 2 anti-FAP Fabs, dimeric and monomeric 4-1BB ligand fused at the C-terminus of each heavy chain, respectively).

Table 6: Sequences of FAP-targeted human 4-1BB ligand trimer containing Fc (kih) fusion molecule Construct 1.5

SEQ ID NO:	Description	Sequence
141	anti-FAP (28H1) Fc hole chain fused to dimeric hu 4-1BBL (71-254)	GAAGTGCAGCTGCTGGAATCCGGCGGAGGCCTGGTGCA GCCTGGCGGATCTCTGAGACTGTCCTGCGCCGCCTCCGG CTTCACCTTCTCCTCCCACGCCATGTCCTGGGTCCGACA GGCTCCTGGCAAAGGCCTGGAATGGGTGTCCGCCATCTG GGCCTCCGGCGAGCAGTACTACGCCGACTCTGTGAAGG GCCGGTTCACCATCTCCCGGGACAACCTCCAAGAACACCC TGTACCTGCAGATGAACTCCCTGCGGGCCGAGGACACC GCCGTGTACTACTGTGCCAAGGGCTGGCTGGGCAACTTC GACTACTGGGGACAGGGCACCCCTGGTCACCGTGTCCAG CGCTAGCACCAAGGGCCCCCTCCGTGTTCCCCCTGGCCCC CAGCAGCAAGAGCACCAGCGGCGGCACAGCCGCTCTGG GCTGCCTGGTCAAGGACTACTTCCCCGAGCCCGTGACCG TGTCCTGGAACAGCGGAGCCCTGACCTCCGGCGTGAC ACCTTCCCCGCCGTGCTGCAGAGTTCTGGCCTGTATAGC CTGAGCAGCGTGGTCACCGTGCCTTCTAGCAGCCTGGGC ACCCAGACCTACATCTGCAACGTGAACCACAAGCCCAG CAACACCAAGGTGGACAAGAAGGTGGAGCCCAAGAGCT GCGACAAAACCTCACACATGCCCACCGTGCCCAGCACCT GAAGCTGCAGGGGGACCGTCAGTCTTCCCTCTTCCCCCA AAACCCAAGGACACCCTCATGATCTCCCGGACCCCTGA GGTCACATGCGTGGTGGTGGACGTGAGCCACGAAGACC CTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGAG GTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTA CAACAGCACGTACCGTGTGGTCAGCGTCCTACCGTCCT GCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCA AGGTCTCCAACAAAGCCCTCGGCGCCCCCATCGAGAAA ACCATCTCCAAGCCAAAGGGCAGCCCCGAGAACCACA GGTGTGCACCCTGCCCCCATCCCGGGATGAGCTGACCAA

		GAACCAGGTCAGCCTCTCGTGCGCAGTCAAAGGCTTCTA TCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGC AGCCGGAGAACAAC TACAAGACCACGCCTCCCGTGCTG GACTCCGACGGCTCCTTCTTCTCGTGAGCAAGCTCACC GTGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTC ATGCTCCGTGATGCATGAGGCTCTGCACAACCACTACAC GCAGAAGAGCCTCTCCCTGTCTCCGGGTGGAGGCGGCG GAAGCGGAGGAGGAGGATCCAGAGAGGGCCCTGAGCTG AGCCCCGATGATCCTGCTGGACTGCTGGACCTGCGGCAG GGCATGTTTGCTCAGCTGGTGGCCAGAACGTGCTGCTG ATCGATGGCCCCCTGTCTGGTACAGCGATCCTGGACTG GCTGGCGTGTCACTGACAGGCGGCCTGAGCTACAAAGA GGACACCAAAGAACTGGTGGTGGCCAAGGCCGGCGTGT ACTACGTGTTCTTTCAGCTGGAAGTGCAGAGAGTGGTGG CCGGCGAAGGATCTGGCTCTGTGTCTCTGGCCCTGCATC TGCAGCCTCTGAGAAGCGCTGCTGGCGCTGCAGCTCTGG CACTGACAGTGGATCTGCCTCCTGCCAGCTCCGAGGCCC GGAATAGCGCATTTGGGTTTCAAGGCAGGCTGCTGCACC TGTCTGCCGGCCAGAGGCTGGGAGTGCATCTGCACACA GAGGCCAGGGCTAGACACGCCTGGCAGCTGACACAGGG CGCTACAGTGCTGGGCCTGTTCAAGAGTGACCCCCGAGAT TCCAGCCGGCCTGCCTTCTCCAAGAAGCGAAGGCGGAG GCGGATCTGGCGGGCGGAGGATCTAGAGAGGGACCCGAA CTGTCCCCTGACGATCCAGCCGGGCTGCTGGATCTGAGA CAGGGAATGTTTCGCCAGCTGGTGGCTCAGAATGTGCTG CTGATTGACGGACCTCTGAGCTGGTACTCCGACCCAGGG CTGGCAGGGGTGTCCCTGACTGGGGGACTGTCCTACAAA GAAGATACAAAAGAACTGGTGGTGGCTAAAGCTGGGGT GTACTATGTGTTTTTTCAGCTGGAAGTGAAGCGGGTGGT GGCTGGGGAGGGCTCAGGATCTGTGTCCCTGGCTCTGCA TCTGCAGCCACTGCGCTCTGCTGCTGGCGCAGCTGCACT GGCTCTGACTGTGGACCTGCCACCAGCCTCTAGCGAGGC CAGAAACAGCGCCTTCGGGTTCGAAGGACGCCTGCTGC ATCTGAGCGCCGACAGCGCCTGGGAGTGCATCTGCAT ACTGAAGCCAGAGCCCGGCATGCTTGGCAGCTGACTCA GGGGGCAACTGTGCTGGGACTGTTTCGCGTGACACCTGA GATCCCTGCCGACTGCCAAGCCCTAGATCAGAA
142	anti-FAP (28H1) Fc knob chain fused to monomeric hu 4- 1BBL (71-254)	GAAGTGCACTGCTGGAATCCGGCGGAGGCCTGGTGCA GCCTGGCGGATCTCTGAGACTGTCTTGCGCCGCCTCCGG CTTCACCTTCTCCTCCCACGCCATGTCTGGGTCCGACA GGCTCCTGGCAAAGGCCTGGAATGGGTGTCCGCCATCTG GGCCTCCGGCGAGCAGTACTACGCCGACTCTGTGAAGG GCCGGTTCACCATCTCCCGGGACAAC TCAAGAACACCC TGTACCTGCAGATGAACTCCCTGCGGGCCGAGGACACC GCCGTGTACTACTGTGCCAAGGGCTGGCTGGGCAACTTC GACTACTGGGGACAGGGCACCCCTGGTCACCGTGTCCAG CGCTAGCACCAAGGGCCCATCGGTCTTCCCCCTGGCACC CTCCTCCAAGAGCACCTCTGGGGGCACAGCGGCCCTGG GCTGCCTGGTCAAGGACTACTTCCCCGAACCGGTGACGG TGTCGTGGAAGTCAAGGCGCCCTGACCAGCGGCGTGAC ACCTTCCCCGGCTGTCTTACAGTCCTCAGGACTCTACTCC CTCAGCAGCGTGGTGACCGTGCCCTCCAGCAGCTTGGGC ACCCAGACCTACATCTGCAACGTGAATCACAAGCCCAG CAACACCAAGGTGGACAAGAAAGTTGAGCCCAAATCTT

		<p>GTGACAAAACCTCACACATGCCCACCGTGCCCAGCACCT GAAGCTGCAGGGGGACCGTCAGTCTTCCTCTTCCCCCA AAACCCAAGGACACCCTCATGATCTCCCGGACCCCTGA GGTCACATGCGTGGTGGTGGACGTGAGCCACGAAGACC CTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAG GTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTA CAACAGCACGTACCGTGTGGTCAGCGTCCTACCGTCCT GCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCA AGGTCTCCAACAAAGCCCTCGGCGCCCCCATCGAGAAA ACCATCTCCAAGCCAAAGGGCAGCCCCGAGAACCACA GGTGTACACCCTGCCCCCTGCAGAGATGAGCTGACCA AGAACCAGGTGTCCCTGTGGTGTCTGGTCAAGGGCTTCT ACCCAGCGATATCGCCGTGGAGTGGGAGAGCAACGGC CAGCCTGAGAACAACACTACAAGACCACCCCCCTGTGCT GGACAGCGACGGCAGCTTCTTCCTGTACTCCAACTGAC CGTGGACAAGAGCCGGTGGCAGCAGGGCAACGTGTTCA GCTGCAGCGTGATGCACGAGGCCCTGCACAACCACTAC ACCCAGAAGTCCCTGAGCCTGAGCCCCGGCGGAGGCGG CGGAAGCGGAGGAGGAGGATCCAGAGAGGGCCCTGAG CTGAGCCCCGATGATCCTGCTGGACTGCTGGACCTGCGG CAGGGCATGTTTGCTCAGCTGGTGGCCAGAACGTGCTG CTGATCGATGGCCCCCTGTCTGGTACAGCGATCCTGGA CTGGCTGGCGTGTCACTGACAGGCGGCCTGAGCTACAA AGAGGACACCAAAGAACTGGTGGTGGCCAAGGCCGGCG TGTA CTACGTGTTCTTTCAGCTGGA ACTGCGGAGAGTGG TGGCCGGCGAAGGATCTGGCTCTGTGTCTCTGGCCCTGC ATCTGCAGCCTCTGAGAAGCGCTGCTGGCGCTGCAGCTC TGGCACTGACAGTGGATCTGCCTCCTGCCAGCTCCGAGG CCCGGAATAGCGCATTTGGGTTTCAAGGCAGGCTGCTGC ACCTGTCTGCCGCCAGAGGCTGGGAGTGCATCTGCAC ACAGAGGCCAGGGCTAGACACGCCTGGCAGCTGACACA GGGCGCTACAGTGCTGGGCCTGTTCAAGAGTGACCCCCG AGATTCCAGCCGGCCTGCCTTCTCCAAGAAGCGAA</p>
69	anti-FAP (28H1) light chain	see Table 2
121	anti-FAP (28H1) Fc hole chain fused to dimeric hu 4-1BBL (71- 254)	<p>EVQLLESGGGLVQPGGSLRLSCAASGFTFSSHAMSWVRQA PGKGLEWVSAIWASGEQYYADSVKGRFTISRDN SKNTLYL QMNSLR AEDTAVYYCAKGWLG NFDYW GQGLVTVSSAS TKGPSVFPLAPSSKSTSGGTAA LGCLVKDYFPEPVT VSWNS GALTSGVHTFPAVLQSSGLYSLSSV VTPSSSLGTQTYICN VNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVF LFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDG VEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYK CKVSNKALGAPIEKTISKAKGQPREPQVCTLP SRDELTKN QVSLSCAVKGFYPSDIAVEWESNGQPENNYKTPPVLDSD GSFFLVSKLTVDKSRWQQGNV FSCVMHEALHNHYTQKS LSLSPGGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQ LVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELV VAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAA GAAALALTVDLPASSEARNSAFGFQGRLLHLSAGQRLGV HLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSEG GGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQLVAQNV</p>

		LLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGV YYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALA LTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEA RARHAWQLTQGATVLGLFRVTPEIPAGLPSRSE
122	anti-FAP (28H1) Fc knob chain fused to monomeric hu 4- 1BBL (71-254)	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSHAMSWVRQA PGKGLEWVSAIWASGEQYYADSVKGRFTISRDN SKNTLYL QMNSLRAEDTAVYYCAKGWLG NFDYWGQGLTVTVSSAS TKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNS GALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICN VNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVF LFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDG VEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYK CKVSNKALGAPIEKTISKAKGQPREPQVYTLPPCRDELTKN QVSLWCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDS GSFFLYSKLTVDKSRWQQGNV FSCVMHEALHNHYTQKS LSLSPGGGGGSGGGGSGREGPELSPDDPAGLLDLRQGMFAQ LVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELV VAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAA GAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGV HLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSE
19	anti-FAP (28H1) light chain	see Table 2

Table 7 shows the cDNA and amino acid sequences of the monovalent FAP-targeted 4-1BB ligand trimer-containing Fc (kih) fusion molecule Construct 1.6 (FAP split trimer with anti-FAP Fab, monomeric 4-1BB ligand fused to CL* via a (G4S)-linker).

5 **Table 7: Sequences of FAP-targeted human 4-1BB ligand trimer containing Fc (kih) fusion molecule Construct 1.6**

SEQ ID NO:	Description	Sequence
131	Dimeric hu 4-1BBL (71-254) – CH1* Fc knob chain	see Table 4
143	Monomeric hu 4-1BBL (71-254) – (G4S) ₁ - CL*	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAACTGGTGG TGGCCAAGGCCGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT

		GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGCCCGGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGTGGGCCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCT CCAAGAAGCGAAGGCGGAGGCGGATCTCGTACGGTGGC TGCACCATCTGTCTTCATCTTCCCGCCATCTGATCGGAA GTTGAAATCTGGAAGTGCCTCTGTTGTGTGCCTGCTGAA TAACTTCTATCCCAGAGAGGCCAAAGTACAGTGGAAGG TGGATAACGCCCTCCAATCGGGTAACTCCCAGGAGAGT GTCACAGAGCAGGACAGCAAGGACAGCACCTACAGCCT CAGCAGCACCTGACGCTGAGCAAAGCAGACTACGAGA AACACAAAGTCTACGCCTGCGAAGTCACCCATCAGGGC CTGAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGAGA GTGT
68	anti-FAP (28H1) Fc hole chain	see Table 2
69	anti-FAP (28H1) light chain	see Table 2
108	Dimeric hu 4- 1BBL (71-254) – CH1* Fc knob chain	see Table 4
110	Monomeric hu 4- 1BBL (71-254) – (G4S) ₁ - CL*	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLPSPRSEGGGGSRTVAAPSVFIFP PSDRKLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGN SQESVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTH QGLSPVTKSFNRGEC
18	anti-FAP (28H1) Fc hole chain	see Table 2
19	anti-FAP (28H1) light chain	see Table 2

Table 8 shows the cDNA and amino acid sequences of the bivalent FAP-targeted 4-1BB ligand trimer-containing Fc (kih) fusion molecule Construct 7 (FAP split trimer with double anti-

FAP on the N-terminus of Fc hole chain and charged residues on crossed CH1 and CL fused to 4-1BB ligands).

Table 8: Sequences of FAP-targeted human 4-1BB ligand trimer containing Fc (kih) fusion molecule Construct 1.7

SEQ ID NO:	Description	Sequence
129	Dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 3
130	Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
144	[anti-FAP (28H1)] ₂ Fc hole chain	GAAGTGCAGCTGCTGGAATCCGGCGGAGGCCTGGTGCA GCCTGGCGGATCTCTGAGACTGTCCTGCGCCGCCTCCGG CTTCACCTTCTCCTCCCACGCCATGTCCTGGGTCCGACA GGCTCCTGGCAAAGGCCTGGAATGGGTGTCCGCCATCTG GGCCTCCGGCGAGCAGTACTACGCCGACTCTGTGAAGG GCCGGTTCACCATCTCCCGGGACAACCTCCAAGAACACCC TGTACCTGCAGATGAACTCCCTGCGGGCCGAGGACACC GCCGTGTACTACTGTGCCAAGGGCTGGCTGGGCAACTTC GACTACTGGGGACAGGGCACCCCTGGTCACCGTGTCCAG CGCTAGCACAAAGGGACCTAGCGTGTTCCTCCCTGGCCCC CAGCAGCAAGTCTACATCTGGCGGAACAGCCGCCCTGG GCTGCCTCGTGAAGGACTACTTTCCCGAGCCCGTGACCG TGTCCTGGAACCTCTGGCGCTCTGACAAGCGGCGTGCACA CCTTTCCAGCCGTGCTGCAGAGCAGCGGCCTGTACTCTC TGAGCAGCGTCGTGACAGTGCCAGCAGCTCTCTGGGC ACCCAGACCTACATCTGCAACGTGAACCACAAGCCAG CAACACCAAGGTGGACAAGAAGGTGGAACCCAAGAGCT GCGACGGCGGAGGGGGATCTGGCGGCGGAGGATCCGAA GTGCAGCTGCTGGAATCCGGCGGAGGCCTGGTGCAGCC TGGCGGATCTCTGAGACTGTCCTGCGCCGCCTCCGGCTT CACCTTCTCCTCCCACGCCATGTCCTGGGTCCGACAGGC TCCTGGCAAAGGCCTGGAATGGGTGTCCGCCATCTGGGC CTCCGGCGAGCAGTACTACGCCGACTCTGTGAAGGGCC GGTTCACCATCTCCCGGGACAACCTCCAAGAACACCCTGT ACCTGCAGATGAACTCCCTGCGGGCCGAGGACACCGCC GTGTACTACTGTGCCAAGGGCTGGCTGGGCAACTTCGAC TACTGGGGACAGGGCACCCCTGGTCACCGTGTCCAGCGCT AGCACCAAGGGCCCCCTCCGTGTTCCCCCTGGCCCCCAGC AGCAAGAGCACCAGCGGCGGCACAGCCGCTCTGGGCTG CCTGGTCAAGGACTACTTCCCCGAGCCCGTGACCGTGTCT CTGGAACAGCGGAGCCCTGACCTCCGGCGTGCACACCTT

		CCCC GCCGTGCTGCAGAGTTCTGGCCTGTATAGCCTGAG CAGCGTGGTCACCGTGCCTTCTAGCAGCCTGGGCACCCA GACCTACATCTGCAACGTGAACCACAAGCCCAGCAACA CCAAGGTGGACAAGAAGGTGGAGCCCAAGAGCTGCGAC AAAACTCACACATGCCACCGTGCCAGCACCTGAAGC TGCAGGGGGACCGTCAGTCTTCCTCTTCCCCCAAACC CAAGGACACCCTCATGATCTCCCGGACCCCTGAGGTCAC ATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGG TCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCAT AATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAACA GCACGTACCGTGTGGTCAGCGTCCTCACCGTCCTGCACC AGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTC TCCAACAAAGCCCTCGGCGCCCCCATCGAGAAAACCAT CTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTGT GCACCCTGCCCCCATCCCGGGATGAGCTGACCAAGAAC CAGGTCAGCCTCTCGTGCGCAGTCAAAGGCTTCTATCCC AGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCC GGAGAACAACTACAAGACCACGCCTCCCGTGTGGACT CCGACGGCTCCTTCTTCTCGTGAGCAAGCTCACCGTGG ACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGC TCCGTGATGCATGAGGCTCTGCACAACCACTACACGCAG AAGAGCCTCTCCCTGTCTCCGGGTAAA
69	anti-FAP (28H1) light chain	see Table 2
115	Dimeric hu 4- 1BBL (71-254) – CL* Fc knob chain	see Table 3
116	Monomeric hu 4- 1BBL (71-254) – CH1*	see Table 3
145	[anti-FAP (28H1)] ₂ Fc hole chain	EVQLLES GGGVLVQPGGSLRLS CAASGFTFSSHAMSWVRQA PGKGLEWVSAIWASGEQYYADSVKGRFTISRDN SKNTLYL QMNSLRAEDTAVYYCAKGWLG NFDYW GQGTLVTVSSAS TKGPSVFPLAPSSKSTSGGTAA LGCLVKDYFPEPVTVSWNS GALTSGVHTFPAVLQSSGLYSLSSV VTPSSSLGTQTYICN VNHKPSNTKVDKKVEPKSCDGGGGSGGGGSEVQLLESGG GLVQPGGSLRLS CAASGFTFSSHAMSWVRQAPGKGLEWV SAIWASGEQYYADSVKGRFTISRDN SKNTLYLQMNSLRAE DTAVYYCAKGWLG NFDYW GQGTLVTVSSASTKGPSVFPL APSSKSTSGGTAA LGCLVKDYFPEPVTVSWNSGALTSGVH TFP AVLQSSGLYSLSSV VTPSSSLGTQTYICNVNHKPSNT KVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPPKPKDT LMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKT KPREEQYNSTYRVVSVLTVLHQD WLN GKEYKCKVSNKAL GAPIEKTISKAKGQPREPQVCTLP SRDELTKNQVSLSCAV KGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLVSKL TVDKSRWQQGNV FSCSV MHEALHNHYTQKSLSLSPGK

19	anti-FAP (28H1) light chain	see Table 2
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Table 9 shows the cDNA and amino acid sequences of the bivalent FAP-targeted 4-1BB ligand trimer-containing Fc (kih) fusion molecule Construct 1.8 (FAP split trimer with 4-1BB ligands fused to anti-FAP CrossFab, with charged residues, on knob chain).

5 **Table 9: Sequences of FAP-targeted human 4-1BB ligand trimer containing Fc (kih) fusion molecule Construct 1.8**

SEQ ID NO:	Description	Sequence
146	Dimeric hu 4-1BBL (71-254) – FAP (VHCL*) Fc knob chain	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAAGTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGGCCCGGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGTGGGCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCT CCAAGAAGCGAAGGCGGAGGCGGATCTGGCGGCGGAG GATCTAGAGAGGGACCCGAAGTGTCCCCTGACGATCCA GCCGGGCTGCTGGATCTGAGACAGGGAATGTTCCGCCA GCTGGTGGCTCAGAATGTGCTGCTGATTGACGGACCTCT GAGCTGGTACTCCGACCCAGGGCTGGCAGGGGTGTCCC TGACTGGGGGACTGTCCTACAAAGAAGATACAAAAGAA CTGGTGGTGGCTAAAGCTGGGGTGTACTATGTGTTTTT CAGCTGGAAGTGAAGCGGGTGGTGGCTGGGGAGGGCTC AGGATCTGTGTCCCTGGCTCTGCATCTGCAGCCACTGCG CTCTGCTGCTGGCGCAGCTGCACTGGCTCTGACTGTGGA CCTGCCACCAGCCTCTAGCGAGGCCAGAAACAGCGCCT TCGGGTTCGAAGGACGCCTGCTGCATCTGAGCGCCGGAC AGCGCCTGGGAGTGCATCTGCATACTGAAGCCAGAGCC CGGCATGCTTGGCAGCTGACTCAGGGGGCAACTGTGCTG GGACTGTTTCGCGTGACACCTGAGATCCCTGCCGGACTG CCAAGCCCTAGATCAGAAGGGGGCGGAGGTTCCGGAGG CGGAGGATCTGAGGTGCAGCTGCTGGAATCCGGCGGAG GCCTGGTGCAGCCTGGCGGATCTCTGAGACTGTCCTGCG CCGCTCCGGCTTCACCTTCTCCTCCACGCCATGTCCTG GGTCCGACAGGCTCCTGGCAAAGGCCTGGAATGGGTGT CCGCCATCTGGGCCTCCGGCGAGCAGTACTACGCCGACT CTGTGAAGGGCCGGTTCACCATCTCCCGGGACAACCTCCA

		AGAACACCCTGTACCTGCAGATGAACTCCCTGCGGGCC GAGGACACCGCCGTGTACTACTGTGCCAAGGGCTGGCT GGGCAACTTCGACTACTGGGGCCAGGGCACCCTGGTCA CCGTGTCCAGCGCTAGCGTGGCTGCACCATCTGTCTTTA TCTTCCCACCCAGCGACCGGAAGCTGAAGTCTGGCACA GCCAGCGTCGTGTGCCTGCTGAATAACTTCTACCCCCGC GAGGCCAAGGTGCAGTGGAAAGGTGGACAATGCCCTGCA GAGCGGCAACAGCCAGGAAAGCGTGACCGAGCAGGAC AGCAAGGACTCCACCTACAGCCTGAGCAGCACCCTGAC CCTGAGCAAGGCCGACTACGAGAAGCACAAGGTGTACG CCTGCGAAGTGACCCACCAGGGCCTGTCTAGCCCCGTGA CCAAGAGCTTCAACCGGGGCGAGTGGCACAAGACCCAC ACCTGTCTCCATGCCCTGCCCCCTGAAGCTGCTGGCGGC CCTAGCGTGTTCCTGTTCCCCCCAAAGCCCAAGGACACC CTGATGATCAGCCGGACCCCTGAAGTGACCTGCGTGGTG GTGGATGTGTCCACAGAGGACCCTGAAGTGAAGTTCAAT TGGTACGTGGACGGCGTGGAAGTGCACAATGCCAAGAC CAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTG TGGTCAGCGTCCTACCGTCCTGCACCAGGACTGGCTGA ATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCC CTCGGCGCCCCCATCGAGAAAACCATCTCCAAAGCCAA AGGGCAGCCCCGAGAACCACAGGTGTACACCCTGCCCC CATGCCGGGATGAGCTGACCAAGAACCAGGTGACGCTG TGGTGCCTGGTCAAAGGCTTCTATCCCAGCGACATCGCC GTGGAGTGGGAGAGCAATGGGCAGCCGGAGAACAATA CAAGACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTT CTTCCTCTACAGCAAGCTACCGTGGACAAGAGCAGGT GGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATG AGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCC CTGTCTCCGGGTAAA
147	Monomeric hu 4-1BBL (71-254) – FAP (VLCH1*)	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAAGTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCTCTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGGCCCGGAATAGCGCATTTGGGTTT CAAGGCCGGCTGCTGCACCTGTCTGCCGGCCAGAGACT GGGAGTGCATCTGCACACAGAGGCCAGAGCCAGGCACG CCTGGCAGCTGACACAGGGCGCTACAGTGTGGGCCCTG TTCAGAGTGACCCCCGAGATTCTGCCGGCCTGCCTAGC CCTAGATCTGAAGGCGGCGGAGGTTCCGGAGGCGGAGG ATCTGAGATCGTGCTGACCCAGTCTCCCGGCACCCTGAG CCTGAGCCCTGGCGAGAGAGCCACCCTGAGCTGCAGAG CCAGCCAGAGCGTGAGCCGGAGCTACCTGGCCTGGTAT CAGCAGAAGCCCGGCCAGGCCCCCAGACTGCTGATCAT CGGCGCCAGCACCCGGGCCACCGGCATCCCCGATAGAT TCAGCGGCAGCGGCTCCGGCACCGACTTCACCCTGACCA TCAGCCGGCTGGAACCCGAGGACTTCGCCGTGTACTACT GCCAGCAGGGCCAGGTGATCCCCCCCACCTTCGGCCAG GGCACCAAGGTGGAAATCAAGTCCTCTGCTAGCACAAA

		GGGCCCCAGCGTGTTCCCTCTGGCCCCTAGCAGCAAGAG CACATCTGGCGGAACAGCCGCCCTGGGCTGCCTGGTGG AAGATTACTTCCCCGAGCCCGTGACCGTGTCTTGGAATT CTGGCGCCCTGACAAGCGGCGTGCACACCTTTCAGCCG TGCTGCAGAGCAGCGGCCTGTACTCTCTGAGCAGCGTCC TGACAGTGCCCAGCAGCTCTCTGGGCACCCAGACCTACA TCTGCAACGTGAACCACAAGCCCAGCAACACCAAGGTG GACGAGAAGGTGGAACCCAAGTCCTGC
68	anti-FAP (28H1) Fc hole chain	see Table 2
69	anti-FAP (28H1) light chain	see Table 2
148	Dimeric hu 4- 1BBL (71-254) – FAP (VHCL*) Fc knob chain	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLPSRSEGGGGSGGGGSREGPEL SPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLA GVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELRRVVAG EGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARN AFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATV LGLFRVTPEIPAGLPSRSEGGGGSGGGGSEVQLLESGGGL VQPGGSLRLSCAASGFTFSSHAMSWVRQAPGKGLEWVSAI WASGEQYYADSVKGRFTISRDN SKNTLYLQMNSLRAEDT AVYYCAKGWLG NFDYWGQGTLVTVSSASVAAPSVFIFPPS DRKLKSGTASVVCLLN NFYPREAKVQWKVDNALQSGNSQ ESVTEQDSKDS TYSLSS TLTL SKADYEKHKVYACEVTHQG LSSPVT KSFNRGEC DKHTHCPPCPAPEAAGGPSVFLFPPKPK DTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNA KTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNK ALGAPIEKTISKAKGQPREPQVYTLPPCRDELTKNQVSLWC LVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYS KLTVDKSRWQQGNV FSCSVMHEALHNHYTQKSLSLSPGK
149	Monomeric hu 4- 1BBL (71-254) – FAP (VLCH1*)	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLPSRSEGGGGSGGGGSEIVLTQS PGTSLSPGERATLSCRASQSVRSYLA WYQQKPGQAPRL LIIGASTRATGIPDRFSGSGSGTDFTLTISRLEPEDFAVYYCQ QGQVIPPTFGQG TKVEIKSSASTKGPSVFPLAPSSKSTSGGT AALGCLVEDYFPEPVT VSWNSGALTSGVHTFPAVLQSSGL YSLSSVTV PSSLGTQTYICNVNHKPSNTKVDEKVEPKSC
18	anti-FAP (28H1) Fc hole chain	see Table 2
19	anti-FAP (28H1) light chain	see Table 2

Table 10 shows the cDNA and amino acid sequences of the monovalent FAP-targeted 4-1BB ligand (52-254) trimer-containing Fc (kih) fusion molecule Construct 1.9 (FAP split trimer with 4-1BBL ectodomain amino acids 52-254 and charged residues on ligand chains).

5 **Table 10: Sequences of FAP-targeted human 4-1BB ligand trimer containing Fc (kih) fusion molecule Construct 1.9**

SEQ ID NO:	Description	Sequence
150	Dimeric hu 4-1BBL (52-254) – CH1* Fc knob chain	CCTTGGGCTGTGTCTGGCGCTAGAGCCTCTCCTGGATCT GCCGCCAGCCCCAGACTGAGAGAGGGACCTGAGCTGAG CCCCGATGATCCTGCCGGAAGTGTGGATCTGAGACAGG GCATGTTCCGCCAGCTGGTGGCCAGAACGTGCTGCTGA TCGATGGCCCCCTGTCTGGTACAGCGATCCTGGACTGG CTGGCGTGTCACTGACAGGCGGCCTGAGCTACAAAGAG GACACCAAAGAACTGGTGGTGGCCAAGGCCGGCGTGTA CTACGTGTTCTTTCAGCTGGAAGTGCAGAGAGTGGTGGC CGGCGAGGGATCTGGATCTGTGTCTCTGGCCCTGCATCT GCAGCCCCCTGAGAAGCGCTGCTGGCGCTGCAGCTCTGG CACTGACAGTGGATCTGCCTCCTGCCAGCTCCGAGGCCC GGAATAGCGCATTTGGGTTTCAAGGCAGACTGCTGCACC TGTCTGCCGGCCAGAGGCTGGGAGTGCATCTGCACACA GAGGCCAGGGCTAGACACGCCTGGCAGCTGACACAGGG CGCTACAGTGTCTGGGCTGTTTCAAGAGTGACCCCGAGAT TCCAGCCGGACTGCCAGCCCTAGATCTGAAGGCGGCG GAGGAAGCGGAGGCGGAGGATCCCCCTGGGCTGTGTCT GGCGCTAGAGCCTCTCCTGGATCTGCCGCCAGCCCCAGA CTGAGAGAGGGACCTGAGCTGAGCCCCGATGATCCTGC CGGACTGCTGGACCTGCGGCAGGGAATGTTGCTCAGCT GGTGGCTCAGAATGTGCTGCTGATTGACGGACCTCTGTC CTGGTACTCCGACCCTGGCCTGGCAGGGGTGTCCCTGAC TGGGGGACTGTCTACAAAGAAGATACAAAAGAACTGG TGGTGGCTAAAGCTGGGGTGTACTATGTGTTTTTTCAGC TGGAAGTGAAGCGGGTGGTGGCTGGGGAGGGCTCAGGA TCTGTGTCCCTGGCTCTGCATCTGCAGCCTCTGCGCTCTG CTGCTGGCGCAGCTGCACTGGCTCTGACTGTGGACCTGC CACCAGCCTCTAGCGAGGCCAGAAACAGCGCCTTCGGG TTCCAAGGACGGCTGCTGCATCTGAGCGCCGGACAGCG CCTGGGAGTGCATCTGCATACTGAAGCCAGAGCCCGGC ATGCTTGGCAGCTGACCCAGGGGGCAACTGTGCTGGGA CTGTTTCGCGTGACACCTGAGATCCCCGCTGGCCTGCCT AGCCCAAGAAGTGAAGGGGGAGGCGGATCTGGCGGAG GGGGATCTGCTAGACCAAGGGCCCCCTCCGTGTTCCCCC TGGCCCCCAGCAGCAAGAGCACCAGCGGCGGCACAGCC GCTCTGGGCTGCCTGGTCGAGGACTACTTCCCCGAGCCC GTGACCGTGTCTTGAACAGCGGAGCCCTGACCTCCGG CGTGACACACCTTCCCCGCCGTGCTGCAGAGTTCTGGCCT GTATAGCCTGAGCAGCGTGGTCACCGTGCCTTCTAGCAG CCTGGGCACCCAGACCTACATCTGCAACGTGAACCACA

		AGCCCAGCAACACCAAGGTGGACGAGAAGGTGGAGCCC AAGAGCTGCGACAAAACCTCACACATGCCCACCGTGCCC AGCACCTGAAGCTGCAGGGGGACCGTCAGTCTTCTCTT CCCCCAAAAACCAAGGACACCCTCATGATCTCCCGGAC CCCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACG AAGACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGC GTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGA GCAGTACAACAGCACGTACCGTGTGGTCAGCGTCTTCAC CGTCTTGCACCAGGACTGGCTGAATGGCAAGGAGTACA AGTGCAAGGTCTCCAACAAAGCCCTCGGCGCCCCCATC GAGAAAACCATCTCCAAGCCAAAGGGCAGCCCCGAGA ACCACAGGTGTACACCCTGCCCCCATGCCGGGATGAGCT GACCAAGAACCAGGTCAGCCTGTGGTGCCTGGTCAAAG GCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGC AATGGGCAGCCGGAGAACAACACTACAAGACCACGCCTCC CGTGCTGGACTCCGACGGCTCCTTCTTCTCTACAGCAA GCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACG TCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACC ACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAA
151	Monomeric hu 4-1BBL (52-254) – CL*	CCTTGGGCTGTGTCTGGCGCTAGAGCCTCTCCTGGATCT GCCGCCAGCCCCAGACTGAGAGAGGGACCTGAGCTGAG CCCCGATGATCCTGCCGGAAGTGTGGATCTGAGACAGG GCATGTTCCGCCAGCTGGTGGCCCAGAACGTGCTGCTGA TCGATGGCCCCCTGTCTGGTACAGCGATCCTGGACTGG CTGGCGTGTCACTGACAGGCGGCCTGAGCTACAAAGAG GACACCAAAGAAGTGGTGGTGGCCAAGGCCGGCGTGTA CTACGTGTTCTTTCAGCTGGAAGTGCAGGAGAGTGGTGGC CGGCGAGGGATCTGGATCTGTGTCTCTGGCCCTGCATCT GCAGCCCCCTGAGAAGCGCTGCTGGCGCTGCAGCTCTGG CACTGACAGTGGATCTGCCTCCTGCCAGCTCCGAGGCCC GGAATAGCGCATTTGGGTTTCAAGGCAGGCTGCTGCACC TGTCTGCCGGCCAGAGGCTGGGAGTGCATCTGCACACA GAGGCCAGGGCTAGACACGCCTGGCAGCTGACACAGGG CGCTACAGTGTGGGCTGTTCAAGAGTGACCCCCGAGAT TCCAGCCGGCCTGCCTTCTCCAAGAAGCGAAGGCGGAG GCGGATCTGGCGGCGGAGGATCTCGTACGGTGGCTGCA CCATCTGTCTTCATCTTCCCGCCATCTGATCGGAAGTTGA AATCTGGAAGTGCCTCTGTTGTGTGCCTGCTGAATAACT TCTATCCCAGAGAGGCCAAAGTACAGTGGAAAGGTGGAT AACGCCCTCCAATCGGGTAACTCCCAGGAGAGTGTAC AGAGCAGGACAGCAAGGACAGCACCTACAGCCTCAGCA GCACCCTGACGCTGAGCAAAGCAGACTACGAGAAACAC AAAGTCTACGCCTGCGAAGTACCCATCAGGGCCTGAG CTCGCCCGTCACAAAGAGCTTCAACAGGGGAGAGTGT
68	anti-FAP (28H1) Fc hole chain	see Table 2
69	anti-FAP (28H1) light chain	see Table 2
111	Dimeric hu 4-1BBL (52-254) –	PWAVSGARASPGSAASPRLREGPELSPDDPAGLLDLRQGM FAQLVAQNVLIDGPLSWYSDPGLAGVSLTGGLSYKEDTK ELVVAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLR

	CH1* Fc knob chain	SAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSPRSEGGGGSGGGGSPWAVSGARASPGSAASPRLREGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSPRSEGGGGSGGGGSASTKGPSVFPLAPSSKSTSGGTAALGCLVEDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVD EKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPPKPKDTLMI SRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGAP IEKTISKAKGQPREPQVYTLPPCRDELTKNQVSLWCLVKGF YPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKLTVD KSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK
112	Monomeric hu 4-1BBL (52-254) – CL*	PWAVSGARASPGSAASPRLREGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSPRSEGGGGSGGGGSRTVAAPSVFIFPPSDRKLKSGTASVVCL LNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYS LSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC
18	anti-FAP (28H1) Fc hole chain	see Table 2
19	anti-FAP (28H1) light chain	see Table 2

Table 11 shows the cDNA and amino acid sequences of the monovalent FAP-targeted 4-1BB ligand (80-254) trimer-containing Fc (kih) fusion molecule Construct 1.10 (FAP split trimer with 4-1BBL ectodomain amino acids 80-254 and charged residues on ligand chains).

5 **Table 11: Sequences of FAP-targeted human 4-1BB ligand trimer containing Fc (kih) fusion molecule Construct 10**

SEQ ID NO:	Description	Sequence
152	Dimeric hu 4-1BBL (80-254) – CH1* Fc knob chain	GATCCTGCCGGCCTGCTGGATCTGCGGCAGGGAATGTTT GCCCAGCTGGTGGCCCAGAACGTGCTGCTGATCGATGG CCCCTGAGCTGGTACAGCGATCCTGGACTGGCTGGCGT GTCAC TGACAGGCGGCCTGAGCTACAAAGAGGACACCA AAGAACTGGTGGTGGCCAAGGCCGGCGTGTACTACGTG TTCTTTCAGCTGGAAGTGC GGAGAGTGGTGGCCGGCGAA GGATCTGGCTCTGTGTCTCTGGCCCTGCATCTGCAGCCC CTGAGAAGCGCTGCTGGCGCTGCAGCTCTGGCACTGAC

		AGTGGATCTGCCTCCTGCCAGCTCCGAGGCCCCGGAATAG CGCATTTGGGTTTCAAGGCAGACTGCTGCACCTGTCTGC CGGCCAGAGGCTGGGAGTGCATCTGCACACAGAGGCCA GGGCTAGACACGCCTGGCAGCTGACACAGGGCGCTACA GTGCTGGGCCTGTTCAGAGTGACCCCCGAGATTCCAGCC GGACTGCCCAGCCCTAGATCTGAAGGCGGCGGAGGAAG CGGAGGCGGAGGATCCGACCCAGCTGGACTGCTGGACC TGCGGCAGGGAATGTTCGCTCAGCTGGTGGCTCAGAATG TGCTGCTGATTGACGGACCTCTGTCTGGTACTCCGACC CTGGCCTGGCAGGGGTGTCCCTGACTGGGGGACTGTCTT ACAAAGAAGATACAAAAGAACTGGTGGTGGCTAAAGCT GGGGTGTACTATGTGTTTTTTCAGCTGGAAGTGAAGCGG GTGGTGGCTGGGGAGGGCTCAGGATCTGTGTCCCTGGCT CTGCATCTGCAGCCTCTGCGCTCTGCTGCTGGCGCAGCT GCACTGGCTCTGACTGTGGACCTGCCACCAGCCTCTAGC GAGGCCAGAAACAGCGCCTTCGGGTTCGAAGGACGGCT GCTGCATCTGAGCGCCGACAGCGCCTGGGAGTGCATC TGCATACTGAAGCCAGAGCCCCGGCATGCTTGGCAGCTG ACCCAGGGGGCAACTGTGCTGGGACTGTTTCGCGTGACA CCTGAGATCCCCGCTGGCCTGCCTAGCCCAAGAAGTGA AGGGGGAGGCGGATCTGGCGGAGGGGGATCTGCTAGCA CCAAGGGCCCCCTCCGTGTTCCCCCTGGCCCCCAGCAGCA AGAGCACCAGCGGCGGCACAGCCGCTCTGGGCTGCCTG GTTCGAGGACTACTTCCCCGAGCCCGTGACCGTGTCTTGG AACAGCGGAGCCCTGACCTCCGGCGTGCACACCTTCCCC GCCGTGCTGCAGAGTTCTGGCCTGTATAGCCTGAGCAGC GTGGTCACCGTGCCTTCTAGCAGCCTGGGCACCCAGACC TACATCTGCAACGTGAACCACAAGCCCAGCAACACCAA GGTGGACGAGAAGGTGGAGCCCAAGAGCTGCGACAAAA CTCACACATGCCACCGTGCCAGCACCTGAAGCTGCAG GGGGACCGTCAGTCTTCCTCTTCCCCCAAAACCCAAGG ACACCCTCATGATCTCCCGGACCCCTGAGGTACATGCG TGGTGGTGGACGTGAGCCACGAAGACCCTGAGGTCAAG TTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCC AAGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGT ACCGTGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACT GGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAAC AAAGCCCTCGGCGCCCCCATCGAGAAAACCATCTCCAA AGCCAAAGGGCAGCCCCGAGAACCACAGGTGTACACCC TGCCCCCATGCCGGGATGAGCTGACCAAGAACCAGGTC AGCCTGTGGTGCCTGGTCAAAGGCTTCTATCCCAGCGAC ATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAA CAACTACAAGACCACGCCTCCCGTGCTGGACTCCGACG GCTCCTTCTTCTCTACAGCAAGCTCACCGTGGACAAGA GCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGA TGCATGAGGCTCTGCACAACCACTACACGCAGAAGAGC CTCTCCCTGTCTCCGGGTAAA
153	Monomeric hu 4-1BBL (80-254) – CL*	GATCCTGCCGGCCTGCTGGATCTGCGGCAGGGAATGTTT GCCCAGCTGGTGGCCAGAACGTGCTGCTGATCGATGG CCCCCTGAGCTGGTACAGCGATCCTGGACTGGCTGGCGT GTCAGTACAGGCGGCCTGAGCTACAAAGAGGACACCA AAGAACTGGTGGTGGCCAAGGCCGCGCTGTACTACGTG TTCTTTCAGCTGGAAGTGCAGGAGAGTGGTGGCCGCGCAA GGATCTGGCTCTGTGTCTCTGGCCCTGCATCTGCAGCCC

		CTGAGAAGCGCTGCTGGCGCTGCAGCTCTGGCACTGAC AGTGGATCTGCCTCCTGCCAGCTCCGAGGCCCCGGAATAG CGCATTTGGGTTTCAAGGCAGGCTGCTGCACCTGTCTGC CGGCCAGAGGCTGGGAGTGCATCTGCACACAGAGGCCA GGGCTAGACACGCCTGGCAGCTGACACAGGGCGCTACA GTGCTGGGCCTGTTCAGAGTGACCCCCGAGATTCCAGCC GGCCTGCCTTCTCCAAGAAGCGAAGGCGGAGGCGGATC TGGCGGCGGAGGATCTCGTACGGTGGCTGCACCATCTGT CTTCATCTTCCCGCCATCTGATCGGAAGTTGAAATCTGG AACTGCCTCTGTTGTGTGCCTGCTGAATAACTTCTATCCC AGAGAGGCCAAAGTACAGTGGAAGGTGGATAACGCCCT CCAATCGGGTAACCTCCAGGAGAGTGTACAGAGCAGG ACAGCAAGGACAGCACCTACAGCCTCAGCAGCACCTTG ACGCTGAGCAAAGCAGACTACGAGAAACACAAAGTCTA CGCCTGCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGT CACAAAGAGCTTCAACAGGGGAGAGTGT
68	anti-FAP (28H1) Fc hole chain	see Table 2
69	anti-FAP (28H1) light chain	see Table 2
113	Dimeric hu 4- 1BBL (80-254) – CH1* Fc knob chain	DPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGV SLTGGLSYKEDTKELVVAKAGVYYVFFQLELRRVVAGEGS GSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGF QGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLF RVTPEIPAGLPSPRSEGGGGSGGGGSDPAGLLDLRQGMFA QLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKEL VVAKAGVYYVFFQLELRRVVAGEGSVSLALHLQPLRSA AGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLG VHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSPRSE GGGGSGGGGSASTKGPSVFPLAPSSKSTSGGTAALGCLVE DYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTV PSSSLGTQTYICNVNHKPSNTKVDEKVEPKSCDKTHTCPPC PAPEAAGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHED PEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVL HQDWLNGKEYKCKVSNKALGAPIEKTISKAKGQPREPQVY TLPPCRDELTKNQVSLWCLVKGFYPSDIAVEWESNGQPEN NYKTTTPVLDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMH EALHNHYTQKSLSLSPGK
114	Monomeric hu 4- 1BBL (80-254) – CL*	DPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGV SLTGGLSYKEDTKELVVAKAGVYYVFFQLELRRVVAGEGS GSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGF QGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLF RVTPEIPAGLPSPRSEGGGGSGGGGSRTVAAPSVFIAPPDR KLKSGTASVVCLLNNFYPPREAKVQWKVDNALQSGNSQES VTEQDSKDSSTYSLSSTLTLSKADYEKHKVYACEVTHQGLS SPVTKSFNRGEC
18	anti-FAP (28H1) Fc hole chain	see Table 2

19	anti-FAP (28H1) light chain	see Table 2
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1.2 Production of FAP (28H1) targeted split trimeric 4-1BB ligand Fc fusion constructs

The targeted TNF ligand trimer-containing Fc (kih) fusion antigen binding molecule encoding sequences were cloned into a plasmid vector, which drives expression of the insert from an MPSV promoter and contains a synthetic polyA sequence located at the 3' end of the CDS. In addition, the vector contains an EBV OriP sequence for episomal maintenance of the plasmid.

The targeted TNF ligand trimer-containing Fc (kih) fusion antigen binding molecule was produced by co-transfecting HEK293-EBNA cells with the mammalian expression vectors using polyethylenimine. The cells were transfected with the corresponding expression vectors at a 1:1:1:1 ratio (e.g. "vector dimeric ligand-(CH1 or CL)- knob chain": "vector monomeric ligand fusion-(CL or CH1)": "vector anti-FAP Fab-hole heavy chain": "vector anti-FAP light chain") for the Constructs 1, 2, 3, 4, 6, 7, 8, 9, 10. For the bivalent Construct 5, a 1:1:1 ratio ("vector hole heavy chain": "vector knob heavy chain": "vector anti-FAP light chain") was used.

For production in 500 mL shake flasks, 300 million HEK293 EBNA cells were seeded 24 hours before transfection. For transfection cells were centrifuged for 10 minutes at 210 x g, and the supernatant was replaced by 20 mL pre-warmed CD CHO medium. Expression vectors (200 µg of total DNA) were mixed in 20 mL CD CHO medium. After addition of 540 µL PEI, the solution was vortexed for 15 seconds and incubated for 10 minutes at room temperature. Afterwards, cells were mixed with the DNA/PEI solution, transferred to a 500 mL shake flask and incubated for 3 hours at 37 °C in an incubator with a 5% CO₂ atmosphere. After the incubation, 160 mL of Excell medium supplemented with 6 mM L-Glutamine, 5 g/L PEPSOY and 1.2 mM valproic acid was added and cells were cultured for 24 hours. One day after transfection 12% Feed 7 and Glucose (final concentration 3 g/L) were added. After culturing for 7 days, the supernatant was collected by centrifugation for 30-40 minutes at 400 x g. The solution was sterile filtered (0.22 µm filter), supplemented with sodium azide to a final concentration of 0.01 % (w/v), and kept at 4 °C.

The targeted TNF ligand trimer-containing Fc (kih) fusion antigen binding molecule was purified from cell culture supernatants by affinity chromatography using Protein A, followed by size exclusion chromatography. For affinity chromatography, the supernatant was loaded on a MabSelect Sure column (CV = 5-15 mL, resin from GE Healthcare) equilibrated with 20 mM

sodium phosphate, 20 mM sodium citrate buffer (pH 7.5). Unbound protein was removed by washing with at least 6 column volumes of the same buffer. The bound protein was eluted using either a linear gradient (20 CV) or a step elution (8 CV) with 20 mM sodium citrate, 100 mM Sodium chloride, 100 mM Glycine buffer (pH 3.0). For the linear gradient an additional 4
5 column volumes step elution was applied.

The pH of collected fractions was adjusted by adding 1/10 (v/v) of 0.5M sodium phosphate, pH 8.0. The protein was concentrated prior to loading on a HiLoad Superdex 200 column (GE Healthcare) equilibrated with 20 mM Histidine, 140 mM sodium chloride, 0.01% (v/v) Tween20 solution of pH 6.0.

The protein concentration was determined by measuring the optical density (OD) at 280 nm, using a molar extinction coefficient calculated on the basis of the amino acid sequence. Purity and molecular weight of the targeted TNF ligand trimer-containing Fc (kih) fusion antigen binding molecule was analyzed by SDS-PAGE in the presence and absence of a reducing agent (5 mM 1,4-dithiotreitol) and staining with Coomassie SimpleBlue™ SafeStain (Invitrogen USA)
15 or or CE-SDS using Caliper LabChip GXII (Perkin Elmer).. The aggregate content of samples was analyzed using a TSKgel G3000 SW XL analytical size-exclusion column (Tosoh) equilibrated in 25 mM K₂HPO₄, 125 mM NaCl, 200 mM L-Arginine Monohydrochloride, 0.02 % (w/v) NaN₃, pH 6.7 running buffer at 25°C.

Table 12 summarizes the yield and final monomer content of the FAP-targeted 4-1BBL
20 trimer-containing Fc (kih) fusion antigen binding molecules.

Table 12: Biochemical Analysis of the FAP (28H1)-targeted 4-1BBL trimer-containing Fc (kih) fusion antigen binding molecules

Construct	Yield [mg/l]	Monomer [%] (SEC)
Construct 1.1	12.7	95
Construct 1.2	25.2	97
Construct 1.3	22	92
Construct 1.4	14.2	99
Construct 1.5	14	99
Construct 1.6	12	98
Construct 1.7	3.4	99
Construct 1.8	5.4	98
Construct 1.9	11.2	98
Construct 1.10	19.8	99

1.3 Preparation of targeted murine 4-1BB ligand trimer-containing Fc fusion antigen binding molecules

Similarly to targeted human 4-1BB ligand trimer-containing Fc fusion antigen binding molecules, murine FAP-targeted 4-1BBL trimer-containing Fc fusion antigen binding molecules were prepared.

The DNA sequence encoding part of the ectodomain (amino acids 104-309) of murine 4-1BB ligand was synthesized according to the Q3U1Z9-1 sequence of Uniprot database (SEQ ID NO:70). For Construct M.1 the cysteines at positions 137, 160 and 246 were mutated to Serine by standard PCR methods, whereas for Construct M.2 the cysteine at position 160 was mutated to Serine (C160S).

The murine ligand was assembled as described for the human 4-1BBL and as depicted in Figure 3A and 3B. The dimeric 4-1BBL, separated by (G4S)₂ linkers, was fused to the murine IgG1-CL domain (Figure 3A) and the monomeric 4-1BBL was fused to murine IgG1-CH domain (Figure 3B). The polypeptide encoding the dimeric 4-1BB ligand fused to murine CL domain was subcloned in frame with the murine IgG1 heavy chain CH2 and CH3 domains to build the Constructs as depicted in Figure 3C.

For the murine constructs, mutations Lys392Asp and Lys409Asp (DD) were introduced in the heavy chain containing the murine 4-1BBL and mutations Glu356Lys and Asp399Lys (KK) were introduced in the heavy chain containing the anti-FAP Fab to obtain asymmetric molecules (Gunasekaran K. et al, J Biol. Chem., 2010, Jun 18;285(25):19637-46).

Mutations Asp265Ala and Pro329Gly (DAPG) were introduced in the constant region of the heavy chains to abrogate binding to Fc gamma receptors.

Table 13 shows, respectively, the cDNA and amino acid sequences of the FAP-targeted murine 4-1BB ligand trimer-containing Fc fusion antigen binding molecule Construct M.1.

Table 13: Sequences of FAP-targeted murine Construct M.1

SEQ ID NO:	Description	Sequence
71	Dimeric murine 4-1BBL (104-309, C137,160,246S) – CL Fc DD chain	AGAACCGAGCCCAGACCCGCCCTGACCATCACCACCAG CCCTAACCTGGGCACCAGAGAGAACAACGCCGACCAAG TGACCCCCGTGTCCACATCGGCAGCCCCAATACCACAC AGCAGGGCAGCCCTGTGTTTCGCCAAGCTGCTGGCCAAG AACCAGGCCAGCCTGAGCAACACCACCCTGAACTGGCA CAGCCAGGATGGCGCCGGAAGCAGCTATCTGAGCCAGG GCCTGAGATACGAAGAGGACAAGAAAGAACTGGTGGTG GACAGCCCTGGCCTGTACTACGTGTTCTGGAAGTGAAG

		CTGAGCCCCACCTTCACCAACACCGGCCACAAGGTGCA GGGCTGGGTGTCAGTGGTGCTGCAGGCCAAACCCCAGG TGGACGACTTCGACAACCTGGCCCTGACCGTGGAAGTGT TCCCCAGCAGCATGGAAAAACAAGCTGGTGGATCGGAGC TGGTCCCAGCTTCTGCTGCTGAAGGCCGGACACAGACTG AGCGTGGGCCTGAGGGCTTATCTGCACGGCGCCAGGA CGCCTACAGAGACTGGGAGCTGAGCTACCCCAACACAA CCAGCTTCGGCCTGTTCCCTCGTGAAGCCCGACAACCCCT GGGAAGGCGGCGGAGGATCTGGCGGAGGCGGATCTAGA ACAGAGCCTCGGCCTGCCCTGACAATTACCACATCCCCC AATCTGGGCACCCGGGAAAAACAATGCAGATCAAGTGAC ACCTGTGTCTCATATTGGCTCCCCAAACACTACCCAGCA GGGCTCCCCCGTGTGTTGCTAAACTGCTGGCTAAAAATCA GGCCTCCCTGTCTAACACAACACTGAACTGGCACTCCCA GGACGGCGCTGGCAGCTCTTACCTGAGTCAGGGACTGC GCTATGAGGAAGATAAGAAAGAAGTGGTGGTGGATTCC CCCGGACTGTACTATGTGTTTCTGGAAGTGAAGTGTCC CCTACCTTTACAAATACCGGGCACAAAGTGCAGGGATG GGTGTCCCTGGTGTGTCAGGCTAAGCCTCAGGTGGACGA TTTGTATAATCTGGCTCTGACAGTGGAACTGTTTCTAG CAGCATGGAAAAACAAGCTGGTGGACAGAAGCTGGTCCC AGCTCCTGCTGCTGAAGGCCGGACACAGACTGAGCGTG GGCCTGAGAGCCTATCTGCACGGCGCCAGGACGCCTA CAGAGACTGGGAGCTGAGCTACCCCAACACAACCAGCT TCGGCCTGTTCCCTCGTGAAGCCCGACAACCCCTTGGGAAG GCGGCGGAGGATCTGGCGGAGGCGGATCCAGAGCTGAT GCTGCCCTACCGTGTCCATCTTCCCACCCAGCAGCGAG CAGCTGACATCTGGGGGAGCTAGCGTCGTGTGCTTCCTG AACAACTTCTACCCCAAGGACATCAACGTGAAGTGGAA GATCGACGGCAGCGAGCGGCAGAACGGCGTGCTGAATA GCTGGACCGACCAGGACAGCAAGGACTCCACCTACAGC ATGAGCAGCACCTGACCCTGACCAAGGACGAGTACGA GCGGCACAACAGCTACACATGCGAGGCCACCCACAAGA CCAGCACCAAGCCCCATCGTGAAGTCCTTCAACCGGAAC GAGTGCGTGCCAGAGACTGCGGCTGCAAGCCTTGCAT CTGCACCGTGCCCTGAGGTGTCCAGCGTGTTTCATCTTCCC ACCCAAGCCCAAGGACGTGCTGACCATCACCTGACAC CCAAAGTGACCTGCGTGGTGGTGGCCATCAGCAAGGAT GACCCCGAGGTGCAGTTTCAGTTGGTTTCGTGGACGACGTG GAAGTGCACACCGCTCAGACCAAGCCCAGAGAGGAACA GATCAACAGCACCTTCAGAAGCGTGTCCGAGCTGCCCCAT CATGCACCAGGACTGGCTGAACGGCAAAGAATTCAAGT GCAGAGTGAACAGCGCCGCCTTTGGCGCCCCCTATCGAG AAAACCATCTCCAAGACCAAGGGCAGACCCAAGGCCCC CCAGGTGTACACAATCCCCCCCACCCAAGAACAGATGG CCAAGGACAAGGTGTCCCTGACCTGCATGATCACCATT TCTTCCCAGAGGATATCACCGTGGAATGGCAGTGGAAAC GGCCAGCCCGCCGAGAACTACGACAACACCCAGCCTAT CATGGACACCGACGGCTCCTACTTCGTGTACAGCGACCT GAACGTGCAGAAGTCCAACCTGGGAGGCCGGCAACACCT TCACCTGTAGCGTGCTGCACGAGGGCCTGCACAACCACC ACACCGAGAAGTCCCTGTCCCACAGCCCTGGCAAG
72	Monomeric murine 4-1BBL	AGAACCGAGCCCAGACCCGCCCTGACCATCACCACCAG CCCTAACCTGGGCACCAGAGAGACAACGCCGACCAAG

	(104-309, C137,160,246S) – CL	TGACCCCCGTGTCCACATCGGCAGCCCCAATACCACAC AGCAGGGCAGCCCTGTGTTCGCCAAGCTGCTGGCCAAG AACCAGGCCAGCCTGAGCAACACCACCCTGAACTGGCA CAGCCAGGATGGCGCCGGAAGCAGCTATCTGAGCCAGG GCCTGAGATACGAAGAGGACAAGAAAGAACTGGTGGTG GACAGCCCTGGCCTGTACTACGTGTTCTGGAAGTGAAG CTGAGCCCCACCTTCACCAACACCGGCCACAAGGTGCA GGGCTGGGTGTCACTGGTGCTGCAGGCCAAACCCCAGG TGGACGACTTCGACAACCTGGCCCTGACCGTGGAAGTGT TCCCCAGCAGCATGGAAAACAAGCTGGTGGATCGGAGC TGGTCCCAGCTTCTGCTGCTGAAGGCCGGACACAGACTG AGCGTGGGCCTGAGGGCCTATCTGCATGGCGCCAGGA CGCCTACAGAGACTGGGAGCTGAGCTACCCCAACACAA CCAGCTTCGGCCTGTTCCTCGTGAAGCCCGACAACCCCT GGGAAGGCGGCGGAGGCTCCGGAGGAGGCGGAAGCGC TAAGACCACCCCCCCCCAGCGTGTACCCTCTGGCCCCCTGG ATCTGCCGCCAGACCAACAGCATGGTGACCCTGGGCT GCCTGGTGAAGGGCTACTTCCCCGAGCCTGTGACCGTGA CCTGGAACAGCGGCAGCCTGAGCAGCGGCGTGCACACC TTCCAGCCGTGCTGCAGAGCGACCTGTACACCCTGAGC AGCTCCGTGACCGTGCCTAGCAGCACCTGGCCCAGCCA GACAGTGACCTGCAACGTGGCCACCCCTGCCAGCAGCA CCAAGGTGGACAAGAAAATCGTGCCCCGGGACTGC
73	anti-FAP (28H1) Fc KK heavy chain	GAAGTGCAGCTGCTGGAATCCGGCGGAGGCCTGGTGCA GCCTGGCGGATCTCTGAGACTGTCTGCGCCGCCTCCGG CTTCACCTTCTCCTCCCACGCCATGTCTGGGTCCGACA GGCTCCTGGCAAAGGCCTGGAATGGGTGTCCGCCATCTG GGCCTCCGGCGAGCAGTACTACGCCGACTCTGTGAAGG GCCGGTTCACCATCTCCCGGGACAACCTCCAAGAACACCC TGTACCTGCAGATGAACTCCCTGCGGGCCGAGGACACC GCCGTGTACTACTGTGCCAAGGGCTGGCTGGGCAACTTC GACTACTGGGGACAGGGCACCCCTGGTCACCGTGTCCAG CGCTAAGACCACCCCCCTAGCGTGTACCCTCTGGCCCC TGGATCTGCCGCCAGACCAACAGCATGGTGACCCTGG GCTGCCTGGTGAAGGGCTACTTCCCCGAGCCTGTGACCG TGACCTGGAACAGCGGCAGCCTGAGCAGCGGCGTGCAC ACCTTTCCAGCCGTGCTGCAGAGCGACCTGTACACCCTG AGCAGCTCCGTGACCGTGCCTAGCAGCACCTGGCCCAG CCAGACAGTGACCTGCAACGTGGCCACCCCTGCCAGCA GCACCAAGGTGGACAAGAAAATCGTGCCCCGGGACTGC GGCTGCAAGCCCTGCATCTGCACCGTGGCCGAGGTGTCC AGCGTGTTTCATCTTCCACCCAAGCCCAAGGACGTGCTG ACCATCACCTGACCCCCAAAGTGACCTGCGTGGTGGTG GCCATCAGCAAGGACGACCCCGAGGTGCAGTTCTCTTG GTTTGTGGACGACGTGGAGGTGCACACAGCCCAGACAA AGCCCCGGGAGGAACAGATCAACAGCACCTTCAGAAGC GTGTCCGAGCTGCCCATCATGCACCAGGACTGGCTGAAC GGCAAAGAATTCAAGTGCAGAGTGAACAGCGCCGCCCTT CGGCGCCCCCATCGAGAAAACCATCAGCAAGACCAAGG GCAGACCCAAGGCCCCCAGGTGTACACCATCCCCCA CCCAAAAAACAGATGGCCAAGGACAAGGTGTCCCTGAC CTGCATGATCACCAACTTTTTCCCCGAGGACATCACCGT GGAGTGGCAGTGAATGGCCAGCCCGCCGAGAAGTACA AGAACACCCAGCCCATCATGAAGACCGACGGCAGCTAC

		TTCGTGTACAGCAAGCTGAACGTGCAGAAGTCCAACTG GGAGGCCGGCAACACCTTCACCTGTAGCGTGTCTGCACG AGGGCCTGCACAACCACACACCGAGAAGTCCCTGAGC CACTCCCCCGGCAAG
74	anti-FAP (28H1) light chain	GAGATCGTGTGCTGACCCAGTCCCCCGGCACCCTGTCTCTG AGCCCTGGCGAGAGAGCCACCCTGTCCTGCAGAGCCTC CCAGTCCGTGTCCCGGTCCTACCTCGCCTGGTATCAGCA GAAGCCCCGGCCAGGCCCTCGGCTGCTGATCATCGGCG CCTCTACCAGAGCCACCGGCATCCCTGACCGGTTCTCCG GCTCTGGCTCCGGCACCGACTTCACCCTGACCATCTCCC GGCTGGAACCCGAGGACTTCGCCGTGTACTACTGCCAGC AGGGCCAGGTCATCCCTCCCACCTTTGGCCAGGGCACCA AGGTGGAAATCAAGCGTGCCGATGCTGCACCAACTGTA TCGATTTTCCCACCATCCAGTGAGCAGTTAACATCTGGA GGTGCCTCAGTCGTGTGCTTCTTGAACAACCTTCTACCCC AAAGACATCAATGTCAAGTGGAAGATTGATGGCAGTGA ACGACAAAATGGCGTCCTGAACAGTTGGACTGATCAGG ACAGCAAAGACAGCACCTACAGCATGAGCAGCACCTC ACGTTGACCAAGGACGAGTATGAACGACATAACAGCTA TACCTGTGAGGCCACTCACAAGACATCAACTTCACCCAT TGTCAGAGCTTCAACAGGAATGAGTGT
75	Dimeric murine 4-1BBL (104- 309, C137,160,246S) – CL Fc DD chain	RTEPRPALTITTSPTNLGTRENNADQVTPVSHIGSPNTTQQGS PVFAKLLAKNQASLSNTTLNWHSQDGAGSSYLSQGLRYEE DKKELVVDSPGLYYVFLELKLSPFTNTGCHKVQGWVSLVL QAKPQVDDFDNLALTVELFPSSMENKLVDRSWSQLLLLKA GHRLSVGLRAYLHGAQDAYRDWELSYPNNTTSFGLFLVKP DNPWEGGGGSGGGGSRTEPRPALTITTSPTNLGTRENNADQ VTPVSHIGSPNTTQQGSPVFAKLLAKNQASLSNTTLNWHS QDGAGSSYLSQGLRYEEDKKELVVDSPGLYYVFLELKLSP FTNTGCHKVQGWVSLVLQAKPQVDDFDNLALTVELFPSS MENKLVDRSWSQLLLLKAGHRLSVGLRAYLHGAQDAYR DWELSYPNNTTSFGLFLVKPDNPWEGGGGSGGGGSRADAA PTVSIFPPSSEQLTSGGASVVCFLNNFYPKDINVKWKIDGSE RQNGVLNSWTDQDSKDSTYSMSSTLTLTKDEYERHNSYT CEATHKTSTSPIVKSFNRNECVPRDCGCKPCICTVPEVSSVF IFPPKPKDVLITLTPKVTCVVVAISKDDPEVQFSWFVDDV EVHTAQTKPREEQINSTRSVSELPIMHQDWLNGKEFKCR VNSAAFGAPIEKTISKTKGRPKAPQVYTIPPPKEQMAKDKV SLTCMITNFFPEDITVEWQWNGQPAENYDNTQPIMDTDGS YFVYSDLNVQKSNWEAGNTFTCSVLHEGLHNHHTKSL HSPGK
76	Monomeric murine 4-1BBL (104-309, C137,160,246S) – CL	RTEPRPALTITTSPTNLGTRENNADQVTPVSHIGSPNTTQQGS PVFAKLLAKNQASLSNTTLNWHSQDGAGSSYLSQGLRYEE DKKELVVDSPGLYYVFLELKLSPFTNTGCHKVQGWVSLVL QAKPQVDDFDNLALTVELFPSSMENKLVDRSWSQLLLLKA GHRLSVGLRAYLHGAQDAYRDWELSYPNNTTSFGLFLVKP DNPWEGGGGSGGGGSAKTTPPSVYPLAPGSAAQTNSMVT LGCLVKGYFPEPVTVTWNSGSLSSGVHTFPAVLQSDLYTL SSSVTVPSSTWPSQTVTCNVAPASSTKVDDKIVPRDC
77	anti-FAP (28H1) Fc KK chain	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSHAMSWSVRQA PGKGLEWVSAIWASGEQYYADSVKGRFTISRDNKNTLYL QMNSLR AEDTAVYYCAKGWLGNFYWGQGTLVTVSSAK

		TTPPSVYPLAPGSAAQTNSMVTLGCLVKGYFPEPVTVTWN SGLSSGVHTFPAVLQSDLYTLSSSVTVPSSTWPSQTVTCN VAHPASSTKVDDKKIVPRDCGCKPCICTVPEVSSVFIFFPKPK DVLITLTPKVTCVVVAISKDDPEVQFSWFVDDVEVHTAQ TKPREEQINSTFRSVSELPIMHQDWLNGKEFKCRVNAAAFG APIEKTISKTKGRPKAPQVYTIPPPKKQMAKDKVSLTCMIT NFFPEDITVEWQWNGQPAENYKNTQPIMKTDGSYFVYSKL NVQKSNWEAGNTFTCSVLHEGLHNHHTKSLSHSPGK
78	anti-FAP (28H1) light chain	EIVLTQSPGTLSPGERATLSCRASQSVRSYLAWEYQQKP GQAPRLLIIGASTRATGIPDRFSGSGSGTDFTLTISRLEPEDF AVYYCQQGQVIPPTFGQGTKVEIKRADAAPTVSIFPPSSEQ LTSGGASVVCFLNNFYPKDINVKWKIDGSERQNGVLNSWT DQDSKDSTYSMSSTLTLTKEDEYERHNSYTCEATHKTSTSPI VKSFNRNEC

Table 14 shows, respectively, the cDNA and amino acid sequences of the untargeted (DP47) murine 4-1BB ligand trimer-containing Fc fusion antigen binding molecule Control M.1.

Table 14: Sequences of untargeted murine Control M.1

SEQ ID NO:	Description	Sequence
71	Dimeric murine 4-1BBL (104-309, C137,160,246S) – CL Fc DD chain	See Table 13
72	Monomeric murine 4-1BBL (104-309, C137,160,246S) – CH1	See Table 13
154	DP47 Fc KK chain	GAGGTGCAATTGTTGGAGTCTGGGGGAGGCTTGGTACA GCCTGGGGGGTCCCTGAGACTCTCCTGTGCAGCCTCCGG ATTCACCTTTAGCAGTTATGCCATGAGCTGGGTCCGCCA GGCTCCAGGGAAGGGGCTGGAGTGGGTCTCAGCTATTA GTGGTAGTGGTGGTAGCACATACTACGCAGACTCCGTGA AGGGCCGGTTCACCATCTCCAGAGACAATTCCAAGAAC ACGCTGTATCTGCAGATGAACAGCCTGAGAGCCGAGGA CACGGCCGTATATTACTGTGCGAAAGGCAGCGGATTGTA CTAAGACCAACCCCCCTAGCGTGTACCTCTGGCCCCCTG GATCTGCCGCCCAGACCAACAGCATGGTGACCCTGGGC TGCCTGGTGAAGGGCTACTTCCCCGAGCCTGTGACCCTG ACCTGGAACAGCGGCAGCCTGAGCAGCGGCGTGACAC CTTTCCAGCCGTGCTGCAGAGCGACCTGTACACCCTGAG CAGCTCCGTGACCGTGCCTAGCAGCACCTGGCCCCAGCCA GACAGTGACCTGCAACGTGGCCACCCCTGCCAGCAGCA

		CCAAGGTGGACAAGAAAATCGTGCCCCGGGACTGCGGC TGCAAGCCCTGCATCTGCACCGTGCCCCGAGGTGTCCAGC GTGTTTCATCTTCCCACCCAAGCCCAAGGACGTGCTGACC ATCACCCCTGACCCCCAAAGTGACCTGCGTGTTGGTGGCC ATCAGCAAGGACGACCCCCGAGGTGCAGTTCTCTTGGTTT GTGGACGACGTGGAGGTGCACACAGCCCAGACAAAGCC CCGGGAGGAACAGATCAACAGCACCTTCAGAAGCGTGT CCGAGCTGCCCATCATGCACCAGGACTGGCTGAACGGC AAAGAATTCAAGTGCAGAGTGAACAGCGCCGCCTTCGG CGCCCCCATCGAGAAAACCATCAGCAAGACCAAGGGCA GACCCAAGGCCCCCCAGGTGTACACCATCCCCCACCCA AAAAACAGATGGCCAAGGACAAGGTGTCCCTGACCTGC ATGATCACCAACTTTTTCCCCGAGGACATCACCGTGGAG TGGCAGTGGAATGGCCAGCCCCGCCGAGAACTACAAGAA CACCCAGCCCATCATGAAGACCGACGGCAGCTACTTCGT GTACAGCAAGCTGAACGTGCAGAAGTCCAAGTGGGAGG CCGGCAACACCTTCACCTGTAGCGTGCTGCACGAGGGCC TGCACAACCACACACCGAGAAGTCCCTGAGCCACTCC CCCGGCAAG
155	DP47 light chain	GAAATCGTGTTAACGCAGTCTCCAGGCACCCTGTCTTTG TCTCCAGGGGAAAGAGCCACCCTCTCTTGCAGGGCCAGT CAGAGTGTTAGCAGCAGCTACTTAGCCTGGTACCAGCAG AAACCTGGCCAGGCTCCCAGGCTCCTCATCTATGGAGCA TCCAGCAGGGCCACTGGCATCCCAGACAGGTTTCAGTGG CAGTGGATCCGGGACAGACTTCACTCTCACCATCAGCAG ACTGGAGCCTGAAGATTTTGCAGTGTATTACTGTCAGCA GTATGGTAGCTCACCGCTGACGTTCCGGCCAGGGGACCA AAGTGGAATCAAACGTGCCGATGCTGCACCAACTGTA TCGATTTTCCCACCATCCAGTGAGCAGTTAACATCTGGA GGTGCCTCAGTCGTGTGCTTCTTGAACAACCTTCTACCCC AAAGACATCAATGTCAAGTGGAAGATTGATGGCAGTGA ACGACAAAATGGCGTCCTGAACAGTTGGACTGATCAGG ACAGCAAAGACAGCACCTACAGCATGAGCAGCACCCCTC ACGTTGACCAAGGACGAGTATGAACGACATAACAGCTA TACCTGTGAGGCCACTCACAAGACATCAACTTCACCCAT TGTCAGAGCTTCAACAGGAATGAGTGT
75	Dimeric murine 4-1BBL (104- 309, C137,160,246S) – CL Fc DD chain	see Table 13
76	Monomeric murine 4-1BBL (104-309, C137,160,246S) – CH1	See Table 13
156	DP47 Fc KK chain	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSYAMSWVRQA PGKGLEWVSAISGSGGSTYYADSVKGRFTISRDN SKNTLYL QMNSLRAEDTAVYYCAKGS GFDYWGQGTLVTVSSAKTTP PSVYPLAPGSAAQTNSMVTLGCLVKGYFPEPVTVTVNSGS LSSGVHTFPAVLQSDLYTLSSSVTVPSSTWPSQTVTCNVAH

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		PASSTKVDKKIVPRDCGCKPCICTVPEVSSVFIFPPKPKDVL TITLTPKVTCVVVAISKDDPEVQFSWFVDDVEVHTAQTKP REEQINSTFRSVSELPIMHQDWLNGKEFKCRVNSAAFGAPI EKTISKTKGRPKAPQVYTIPPPKKQMAKDKVSLTCMITNFF PEDITVEWQWNGQPAENYKNTQPIMKTDGSYFVYSKLN QKSNWEAGNTFTCSVLHEGLHNHHTKSLSHSPGK
157	DP47 light chain	EIVLTQSPGTLSPGERATLSCRASQSVSSSYLAWYQQKP GQAPRLLIYGASSRATGIPDRFSGSGSGTDFTLTISRLEPEDF AVYYCQQYGSSPLTFGQGTKVEIKRADAAPTVSIFPPSSEQ LTSGGASVVCFLNNFYPKDINVKWKIDGSERQNGVLNSWT DQDSKDSTYSMSSTLTTLTKDEYERHNSYTCEATHKTSTSPI VKSFNRNEC

Table 15 shows the cDNA and amino acid sequences of the FAP-targeted murine 4-1BB ligand trimer-containing Fc fusion antigen binding molecule Construct M.2.

Table 15: Sequences of FAP-targeted murine Construct M.2

SEQ ID NO:	Description	Sequence
158	Dimeric murine 4-1BBL (104-309, C160S) – CL Fc DD chain	AGAACCGAGCCCAGACCCGCCCTGACCATCACCACCAG CCCTAACCTGGGCACCAGAGAGAACAACGCCGACCAAG TGACCCCCGTGTCCACATCGGCTGCCCCAATACCACAC AGCAGGGCAGCCCTGTGTTGCGCAAGCTGCTGGCCAAG AACCAGGCCAGCCTGAGCAACACCACCCTGAACTGGCA CAGCCAGGATGGCGCCGGAAGCAGCTATCTGAGCCAGG GCCTGAGATACGAAGAGGACAAGAAAGAACTGGTGGTG GACAGCCCTGGCCTGTACTACGTGTTCTGGAAGTGAAG CTGAGCCCCACCTTCACCAACACCGGCCACAAGGTGCA GGGCTGGGTGTCAGTGGTGCTGCAGGCCAAACCCAGG TGGACGACTTCGACAACCTGGCCCTGACCGTGGAAGTGT TCCCCTGCAGCATGGAAAACAAGCTGGTGGATCGGAGC TGGTCCCAGCTTCTGCTGCTGAAGGCCGGACACAGACTG AGCGTGGGCCTGAGGGCTTATCTGCACGGCGCCAGGA CGCCTACAGAGACTGGGAGCTGAGCTACCCCAACACAA CCAGCTTCGGCCTGTTCTCGTGAAGCCCGACAACCCCT GGGAAGGCGGCGGAGGCTCCGGAGGAGGCGGATCTAGA ACAGAGCCTCGGCCTGCCCTGACAATTACCACATCCCCC AATCTGGGCACCCGGGAAAACAATGCAGATCAAGTGAC ACCTGTGTCTCATATTGGGTGCCCAACACTACCCAGCA GGGGTCCCCAGTGTTTGCTAAACTGCTGGCTAAAAATCA GGCCTCCCTGTCTAACACAACACTGAATTGGCAGTAGTCA GGACGGGGCTGGCAGCAGCTACCTGTCTCAGGGACTGC GCTATGAGGAAGATAAGAAAGAACTGGTGGTGGATTCC CCCGGACTGTACTATGTGTTTCTGGAAGTGAAGTGTCC CCTACCTTTACAAATACCGGGCACAAAGTGCAGGGATG GGTGTCCCTGGTGCTGCAGGCTAAGCCTCAGGTGGACGA TTTGTGATAATCTGGCTCTGACAGTGAAGTGTTCCTTGC TCTATGGAAAACAAGTGGTGGACCGCTCTTGGAGCCA GTTGCTGCTGCTGAAAGCTGGCCACCGGCTGTCTGTGGG

		<p> ACTGAGAGCATACCTGCATGGGGCACAGGATGCCTACC GGGATTGGGAACTGTCCTACCCTAACACTACTTCCTTCG GACTGTTCCCTCGTGAAACCTGATAATCCCTGGGAGGGCG GAGGCGGAAGTGGCGGAGGGGGATCCAGAGCTGATGCT GCCCCTACCGTGTCCATCTTCCCACCCAGCAGCGAGCAG CTGACATCTGGGGGAGCTAGCGTCGTGTGCTTCCTGAAC AACTTCTACCCCAAGGACATCAACGTGAAGTGAAGAT CGACGGCAGCGAGCGGCAGAACGGCGTGCTGAATAGCT GGACCGACCAGGACAGCAAGGACTCCACCTACAGCATG AGCAGCACCTGACCCTGACCAAGGACGAGTACGAGCG GCACAACAGCTACACATGCGAGGCCACCCACAAGACCA GCACCAGCCCCATCGTGAAGTCCTTCAACCGGAACGAG TCGGTGCCCAGAGACTGCGGCTGCAAGCCTTGCATCTGC ACCGTGCCTGAGGTGTCCAGCGTGTTCATCTTCCCACCC AAGCCCAAGGACGTGCTGACCATCACCTGACACCCAA AGTGACCTGCGTGGTGGTGGCCATCAGCAAGGATGACC CCGAGGTGCAGTTCAGTTGGTTCGTGGACGACGTGGAA GTGCACACCGCTCAGACCAAGCCCAGAGAGGAACAGAT CAACAGCACCTTCAGAAGCGTGTCCGAGCTGCCCATCAT GCACCAGGACTGGCTGAACGGCAAAGAATTCAAGTGCA GAGTGAACAGCGCCGCCTTTGGCGCCCCATCGAGAAA ACCATCTCCAAGACCAAGGGCAGACCCAAAGGCCCCCCA GGTGTACACAATCCCCCCCACCCAAAGAACAGATGGCCA AGGACAAGGTGTCCCTGACCTGCATGATCACCAATTTCT TCCCAGAGGATATACCGTGGAATGGCAGTGAACGGC CAGCCCGCCGAGAACTACGACAACACCCAGCCTATCAT GGACACCGACGGCTCCTACTTCGTGTACAGCGACCTGAA CGTGCAGAAGTCCAACCTGGGAGGCCGGCAACACCTTCA CCTGTAGCGTGCTGCACGAGGGCCTGCACAACCACCAC ACCGAGAAGTCCCTGTCCCACAGCCCTGGCAAG </p>
159	<p> Monomeric murine 4-1BBL (104-309, C160S) – CH1 </p>	<p> AGAACCGAGCCCAGACCCGCCCTGACCATCACCACCAG CCCTAACCTGGGCACCAGAGAGAACAACGCCGACCAAG TGACCCCCGTGTCCACATCGGCTGCCCCAATACCACAC AGCAGGGCAGCCCTGTGTTGCGCAAGCTGTGGCCAAG AACCAGGCCAGCCTGAGCAACACCACCCTGAACTGGCA CAGCCAGGATGGCGCCGGAAGCAGCTATCTGAGCCAGG GCCTGAGATACGAAGAGGACAAGAAAGAACTGGTGGTG GACAGCCCTGGCCTGTACTACGTGTTCTTGGAACTGAAG CTGAGCCCCACCTTCACCAACACCGGCCACAAGGTGCA GGGCTGGGTGTCACTGGTGCTGCAGGCCAAACCCCAGG TGGACGACTTCGACAACCTGGCCCTGACCGTGGAAGTGT TCCCCTGCAGCATGGAAAACAAGCTGGTGGATCGGAGC TGGTCCCAGCTTCTGCTGCTGAAGGCCGGACACAGACTG AGCGTGGGCCTGAGGGCTTATCTGCACGGCGCCAGGA CGCCTACAGAGACTGGGAGCTGAGCTACCCCAACACAA CCAGCTTCGGCCTGTTCTCGTGAAGCCCGACAACCTT GGGAAGGCGGCGGAGGCTCCGGAGGAGGCGGAAGCGC TAAGACCACCCCCCCCAGCGTGTACCCTCTGGCCCCCTGG ATCTGCCGCCAGACCAACAGCATGGTGACCTGGGCT GCCTGGTGAAGGGCTACTTCCCCGAGCCTGTGACCGTGA CCTGGAACAGCGGCAGCCTGAGCAGCGGCGTGCACACC TTTCCAGCCGTGCTGCAGAGCGACCTGTACACCCTGAGC AGCTCCGTGACCGTGCCTAGCAGCACCTGGCCCAGCCA GACAGTGACCTGCAACGTGGCCACCCCTGCCAGCAGCA </p>

		CCAAGGTGGACAAGAAAATCGTGCCCCGGGACTGC
73	anti-FAP (28H1) Fc KK chain	see Table 13
74	anti-FAP (28H1) light chain	see Table 13
160	Dimeric murine 4-1BBL (104- 309, C160S) – CL Fc DD chain	RTEPRPALTITTTSPNLGTRENNADQVTPVSHIGCPNTTQQGS PVFAKLLAKNQASLSNTTLNWHSQDGAGSSYLSQGLRYEE DKKELVVDSPGLYYVFLELKLSPFTNTGCHKVQGWVSLVL QAKPQVDDFDNLALTVELFPCSMENKLVDRSWSQLLLLK AGHRLSVGLRAYLHGAQDAYRDWELSYPNNTTSFGLFLVK PDNPWEGGGGSGGGGSRTEPRPALTITTTSPNLGTRENNAD QVTPVSHIGCPNTTQQGSPVFAKLLAKNQASLSNTTLNWH SQDGAGSSYLSQGLRYEEDKKELVVDSPGLYYVFLELKL PTFTNTGCHKVQGWVSLVLQAKPQVDDFDNLALTVELFPCS MENKLVDRSWSQLLLLKAGHRLSVGLRAYLHGAQDAYR DWELSYPNNTTSFGLFLVKPDNPWEGGGGSGGGGSRADAA PTVSIFPPSSEQLTSGGASVVCFLNNFYPKDINVKWKIDGSE RQNGVLNSWTDQDSKDSTYSMSSTLTCLKDEYERHNSYT CEATHKTSTSPIVKSFNRECVPRDCGCKPCICTVPEVSSVF IFPPKPKDVLITLTPKVTCVVVAISKDDPEVQFSWFVDDV EVHTAQTKPREEQINSTFRSVSELPIMHQDWLNGKEFKCR VNSAAFGAIEKTISKTKGRPKAPQVYTIPPPKEQMAKDKV SLTCMITNFFPEDITVEWQWNGQPAENYDNTQPIMDTDGS YFVYSDLNVQKSNWEAGNTFTCSVLHEGLHNHHTKSL HSPGK
161	Monomeric murine 4-1BBL (104-309, C160S) – CH1	RTEPRPALTITTTSPNLGTRENNADQVTPVSHIGCPNTTQQGS PVFAKLLAKNQASLSNTTLNWHSQDGAGSSYLSQGLRYEE DKKELVVDSPGLYYVFLELKLSPFTNTGCHKVQGWVSLVL QAKPQVDDFDNLALTVELFPCSMENKLVDRSWSQLLLLK AGHRLSVGLRAYLHGAQDAYRDWELSYPNNTTSFGLFLVK PDNPWEGGGGSGGGGSAKTTPPSVYPLAPGSAAQTNSMV TLGCLVKGYFPEPVTVTWNSGSLSSGVHTFPAVLQSDLYT LSSSVTVPSSTWPSQTVTCNVVHPASSTKVDKKIVPRDC
77	anti-FAP (28H1) Fc KK chain	see Table 13
78	anti-FAP (28H1) light chain	see Table 13

Table 16 shows the cDNA and amino acid sequences of the DP47-untargeted murine 4-1BB ligand trimer-containing Fc fusion antigen binding molecule Construct Control M.2.

Table 16: Sequences of FAP-targeted murine Control M.2

SEQ ID NO:	Description	Sequence
158	Dimeric mu 4-1BBL (104-309, C160S) – CL Fc DD chain	see Table 15
159	Monomeric mu 4-1BBL (104-309, C160S) – CH1	see Table 15
154	DP47 Fc KK chain	see Table 14
155	DP47 light chain	see Table 14
160	Dimeric mu 4-1BBL (104-309, C160S) – CL Fc DD chain	see Table 15
161	Monomeric mu 4-1BBL (104-309, C160S) – CH1	see Table 15
156	DP47 Fc KK chain	see Table 14
157	DP47 light chain	see Table 14

The murine 4-1BB ligand trimer-containing Fc fusion antigen binding molecules were produced and purified as described herein before for the human 4-1BBL constructs.

- 5 Table 17 summarizes the yield and final monomer content of the FAP-targeted and untargeted murine 4-1BBL trimer-containing Fc fusion antigen binding molecule.

Table 17: Summary of the production of the FAP-targeted and untargeted murine 4-1BBL trimer-containing Fc fusion antigen binding molecules

Construct	Yield [mg/l]	Monomer [%] (SEC)
Construct M.1	2.6	95
Control M.2	2.3	96
Construct M.2	8.5	98
Control M.2	8.1	97

1.4 Preparation and purification of untargeted human 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (Control molecules)

The control molecules were prepared as described above for the FAP-targeted Constructs 1 and 2, with the only difference that the anti-FAP binder (VH-VL) was replaced by a germline control, termed DP47, not binding to the antigen. The control is an untargeted monovalent split trimeric human 4-1BB ligand Fc (kih) (Control A, Figure 5A) and for Control B, the construct also contains a CH-CL crossover with charged residues (Figure 5B). The variable region of heavy and light chain DNA sequences of the FAP binder were replaced with those of the germline control (DP47) and subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1.

The untargeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules were produced as described above for the FAP-targeted constructs. The cells were transfected with the corresponding expression vectors at a 1:1:1:1 ratio (“vector dimeric ligand-CH1 or CL*-knob chain”: “vector monomeric ligand fusion-CL or CH1*”: “vector DP47 Fab-hole chain”: “vector DP47 light chain”).

Table 18 shows, respectively, the cDNA and amino acid sequences of the DP47-untargeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecule Control A.

Table 18: Sequences of DP47 untargeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecule (DP47 split 4-1BBL trimer) Control A

SEQ ID NO:	Description	Sequence
66	Dimeric hu 4-1BBL (71-254) – CH1 Fc knob chain	See Table 2
67	Monomeric hu 4-1BBL (71-254) - CL	see Table 2
79	DP47 Fc hole chain	GAGGTGCAATTGTTGGAGTCTGGGGGAGGCTTGGTACA GCCTGGGGGGTCCCTGAGACTCTCCTGTGCAGCCTCCGG ATTCACCTTTAGCAGTTATGCCATGAGCTGGGTCCGCCA GGCTCCAGGGAAGGGGCTGGAGTGGGTCTCAGCTATTA GTGGTAGTGGTGGTAGCACATACTACGCAGACTCCGTGA AGGGCCGGTTACCATCTCCAGAGACAATTCCAAGAAC ACGCTGTATCTGCAGATGAACAGCCTGAGAGCCGAGGA CACGGCCGTATATTACTGTGCGAAAGGCAGCGGATTGTA CTACTGGGGCCAAGGAACCCTGGTCACCGTCTCGAGTGC TAGCACCAAGGGCCCATCGGTCTTCCCCCTGGCACCCCTC CTCCAAGAGCACCTCTGGGGGCACAGCGGCCCTGGGCT GCCTGGTCAAGGACTACTTCCCCGAACCGGTGACGGTGT

		CGTGGAACCTCAGGCGCCCTGACCAGCGGCGTGACACACC TTCCCGGCTGTCTACAGTCCTCAGGACTCTACTCCCTC AGCAGCGTGGTGACCGTGCCCTCCAGCAGCTTGGGAC CCAGACCTACATCTGCAACGTGAATCACAAGCCAGCA ACACCAAGGTGGACAAGAAAGTTGAGCCCAAATCTTGT GACAAAACCTCACACATGCCACCGTGCCAGCACCTGA AGCTGCAGGGGGACCGTCAGTCTTCTCTTCCCCCAA ACCCAAGGACACCCTCATGATCTCCCGGACCCCTGAGGT CACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTG AGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTG CATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAA CAGCACGTACCGTGTGGTCAGCGTCTCACCCTGCTGCA CCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGG TCTCCAACAAAGCCCTCGGCGCCCCCATCGAGAAAACC ATCTCCAAGCCAAAGGGCAGCCCCGAGAACCACAGGT GTGCACCCTGCCCCCATCCCGGGATGAGCTGACCAAGA ACCAGGTCAGCCTCTCGTGCGCAGTCAAAGGCTTCTATC CCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAG CCGGAGAACAACTACAAGACCACGCCTCCCGTGCTGGA CTCCGACGGCTCCTTCTTCTCGTGAGCAAGCTCACCGT GGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCAT GCTCCGTGATGCATGAGGCTCTGCACAACCACTACACGC AGAAGAGCCTCTCCCTGTCTCCGGGTAAA
80	DP47 light chain	GAAATCGTGTTAACGCAGTCTCCAGGCACCCTGTCTTTG TCTCCAGGGGAAAGAGCCACCCTCTCTTGACGGGCCAGT CAGAGTGTTAGCAGCAGCTACTTAGCCTGGTACCAGCAG AAACCTGGCCAGGCTCCCAGGCTCCTCATCTATGGAGCA TCCAGCAGGGCCACTGGCATCCAGACAGGTTCAAGTGG CAGTGGATCCGGGACAGACTTCACTCTCACCATCAGCAG ACTGGAGCCTGAAGATTTTGCAGTGTATTACTGTCAGCA GTATGGTAGCTCACCGCTGACGTTCCGGCCAGGGGACCA AAGTGGAAATCAAACGTACGGTGGCTGCACCATCTGTCT TCATCTTCCCGCCATCTGATGAGCAGTTGAAATCTGGAA CTGCCTCTGTTGTGTGCCTGCTGAATAACTTCTATCCCAG AGAGGCCAAAGTACAGTGGAAAGGTGGATAACGCCCTCC AATCGGGTAACTCCCAGGAGAGTGTACAGAGCAGGAC AGCAAGGACAGCACCTACAGCCTCAGCAGCACCCCTGAC GCTGAGCAAAGCAGACTACGAGAAACACAAAGTCTACG CCTGCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCA CAAAGAGCTTCAACAGGGGAGAGTGT
14	Dimeric hu 4-1BBL (71-254) – CH1 Fc knob chain	See Table 2
15	Monomeric hu 4-1BBL (71-254) - CL	See Table 2
81	DP47 Fc hole chain	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSYAMSWVRQA PGKGLEWVSAISGSGGSTYYADSVKGRFTISRDN SKNTLYL QMNSLR AEDTAVYYCAKGS GFDYWGQGLTVTVSSASTKG PSVFPLAPSSKSTSGGTAAALGCLVKDYFPEPVTVSWNSGAL TSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNH

		KPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPP KPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEV HNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKV SNKALGAPIEKTISKAKGQPREPQVCTLPPSRDELTKNQVS LSCAVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFF LVSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLS PGK
82	DP47 light chain	EIVLTQSPGTLSSLSPGERATLSCRASQSVSSSYLAWYQQKP GQAPRLLIYGASSRATGIPDRFSGSGSGTDFLTISRLEPEDF AVYYCQQYGSSPLTFGQGTKVEIKRTVAAPSVFIFPPSDEQ LKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESV TEQDSKSTYSLSSLTLSKADYEKHKVYACEVTHQGLSS PVTKSFNRGEC

Table 19 shows the cDNA and amino acid sequences of the DP47-untargeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecule with CH1-CL crossover and charged residues in the 4-1BB ligand containing arms (Control B).

5 Table 19: Sequences of DP47 untargeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecule (DP47 split 4-1BBL trimer) Control B

SEQ ID NO:	Description	Sequence
96	Dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 3
97	Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
79	DP47 Fc hole chain	see Table 18
80	DP47 light chain	see Table 18
98	Dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 3
99	Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
81	DP47 Fc hole chain	see Table 18
82	DP47 light chain	see Table 18

Table 20 summarizes the yield and final monomer content of the DP47 untargeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules.

Table 20: Production Characteristics of DP47 untargeted 4-1BBL trimer-containing Fc (kih) fusion antigen binding molecules (Control molecules)

Construct	Monomer [%] (SEC)	Yield [mg/l]	LC/MS (non red)
Control A	97	3.7	<i>Theoretical*: 179069.7 Da Experimental: 179116.2 Da * without terminal lysines</i>
Control B	99	15.4	

5

Example 2

2.1 Preparation of FAP (4B9) targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules

Different fragments of the DNA sequence encoding part of the ectodomain (amino acid 71-254 and 71-248) of human 4-1BB ligand were synthesized according to the P41273 sequence of Uniprot database (SEQ ID NO:42).

2.1.1 Preparation of monovalent FAP (4B9) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains with charged residues (Construct 2.1)

A polypeptide containing two ectodomains of 4-1BB ligand (71-254), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned as depicted in Figure 1A: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL.

A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the human IgG1-CH1 domain, was cloned as described in Figure 1B: human 4-1BB ligand, (G4S)₂ connector, human CH.

The polypeptide encoding the dimeric 4-1BB ligand fused to human CL domain was subcloned in frame with the human IgG1 heavy chain CH2 and CH3 domains on the knob (Merchant, Zhu et al. 1998) using a linker (G4S)₂, or alternatively, (GSPGSSSSGS).

To improve correct pairing the following mutations have been introduced in the crossed CH-CL. In the dimeric 4-1BB ligand fused to human CL the mutations E123R and Q124K were introduced. In the monomeric 4-1BB ligand fused to human CH1, the mutations K147E and K213E were cloned into the human CH1 domain.

- 5 The variable region of heavy and light chain DNA sequences encoding a binder specific for fibroblast activation protein (FAP), clone 4B9, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1.

The generation and preparation of the FAP binders is described in WO 2012/020006 A2, which is incorporated herein by reference.

- 10 The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in International Patent Appl. Publ. No. WO 2012/130831.

- 15 For all constructs the knobs into hole heterodimerization technology was used with the the S354C/T366W mutations in the knob chain and the corresponding Y349C/T366S/L368A/Y407V mutations in the hole chain.

- 20 Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-FAP-Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations and the anti-FAP light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a FAP binding Fab (Figure 4, Construct 2.1).

Table 21 shows the cDNA and amino acid sequences of the monovalent FAP (4B9)-human 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule containing CH1-CL crossover and charged residues (Construct 2.1).

- 25 **Table 21: Sequences of monovalent FAP(4B9)-targeted human 4-1BB ligand (71-254) containing Fc (kih) fusion molecule Construct 2.1**

SEQ ID NO:	Description	Sequence
129	Dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 3
130	Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3

162	anti-FAP (4B9) Fc hole chain	GAGGTGCAGCTGCTCGAAAGCGGCGGAGGACTGGTGCA GCCTGGCGGCAGCCTGAGACTGTCTTGCGCCGCCAGCG GCTTCACCTTCAGCAGCTACGCCATGAGCTGGGTCCGCC AGGCCCCTGGCAAGGGACTGGAATGGGTGTCCGCCATC ATCGGCTCTGGCGCCAGCACCTACTACGCCGACAGCGTG AAGGGCCGGTTCACCATCAGCCGGGACAACAGCAAGAA CACCTGTACCTGCAGATGAACAGCCTGCGGGCCGAGG ACACCGCCGTGTACTACTGCGCCAAGGGATGGTTCGGC GGCTTCAACTACTGGGGACAGGGCACCCCTGGTCACAGT GTCCAGCGCTAGCACCAAGGGCCCCTCCGTGTTCCCCCT GGCCCCCAGCAGCAAGAGCACCCAGCGGCGGCACAGCCG CTCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAGCCCG TGACCGTGTCTTGGAAACAGCGGAGCCCTGACCTCCGGC GTGCACACCTTCCCCGCCGTGCTGCAGAGTTCTGGCCTG TATAGCCTGAGCAGCGTGGTCACCGTGCCTTCTAGCAGC CTGGGCACCCAGACCTACATCTGCAACGTGAACCACAA GCCCAGCAACACCAAGGTGGACAAGAAGGTGGAGCCCA AGAGCTGCGACAAAACCTCACACATGCCACCGTGCCCA GCACCTGAAGCTGCAGGGGGACCGTCAGTCTTCCTCTTC CCCCCAAACCCAAGGACACCCCTCATGATCTCCCGGACC CCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGA AGACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCG TGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAG CAGTACAACAGCACGTACCGTGTGGTCAGCGTCCTCACC GTCCTGCACCAGGACTGGCTGAATGGCAAGGAGTACAA GTGCAAGGTCTCCAACAAAGCCCTCGGCGCCCCCATCG AGAAAACCATCTCCAAAGCCAAAGGGCAGCCCCGAGAA CCACAGGTGTGCACCCTGCCCCCATCCCGGGATGAGCTG ACCAAGAACCAGGTCAGCCTCTCGTGCAGTCAAAGG CTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGCA ATGGGCAGCCGGAGAACAACACTACAAGACCACGCCTCCC GTGCTGGACTCCGACGGCTCCTTCTTCCTCGTGAGCAAG CTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGT CTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCA CTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAA
163	anti-FAP (4B9) light chain	GAGATCGTGCTGACCCAGTCCCCCGGCACCCTGTCTCTG AGCCCTGGCGAGAGAGCCACCCTGTCTTGCAGAGCCTC CCAGTCCGTGACCTCCTCCTACCTCGCCTGGTATCAGCA GAAGCCCGGCCAGGCCCCCTCGGCTGCTGATCAACGTGG GCAGTCGGAGAGCCACCGGCATCCCTGACCGGTTCCTCCG GCTCTGGCTCCGGCACCGACTTCACCCTGACCATCTCCC GGCTGGAACCCGAGGACTTCGCCGTGTACTACTGCCAGC AGGGCATCATGCTGCCCCCACCTTTGGCCAGGGCACCA AGGTGGAAATCAAGCGTACGGTGGCTGCACCATCTGTCT TCATCTTCCCGCCATCTGATGAGCAGTTGAAATCTGGAA CTGCCTCTGTTGTGTGCCTGCTGAATAACTTCTATCCCAG AGAGGCCAAAGTACAGTGGAAAGGTGGATAACGCCCTCC AATCGGGTAACTCCCAGGAGAGTGTACAGAGCAGGAC AGCAAGGACAGCACCTACAGCCTCAGCAGCACCCCTGAC GCTGAGCAAAGCAGACTACGAGAAACACAAAGTCTACG CCTGCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCA CAAAGAGCTTCAACAGGGGAGAGTGT
115	Dimeric hu 4-	see Table 3

	1BBL (71-254) – CL* Fc knob chain	
116	Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
164	anti-FAP (4B9) Fc hole chain	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSYAMSWVRQA PGKGLEWVSAIIGSGASTYYADSVKGRFTISRDN SKNTLYL QMNSLRAEDTAVYYCAKGWFGGFNYWGQGT LVTVSSAS TKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVT VSWNS GALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICN VNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVF LFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDG VEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYK CKVSNKALGAPIEKTISKAKGQPREPQVCTLPSSRDELTKN QVSLSCAVKGFYPSDIAVEWESNGQPENNYKTPPVLDSD GSFFLVSKLTVDKSRWQQGNV FSCSV MHEALHNHYTQKS LSLSPGK
125	anti-FAP (4B9) light chain	EIVLTQSPGTL SLSPGERATL SCRASQSVTSSYLAWYQQKP GQAPRLLINVGSRRTGIPDRFSGSGSGTDFLTISRLEPEDF AVYYCQQGIMLPPTFGQGTKVEIKRTVAAPSVFIFPPSDEQ LKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESV TEQDSKSTYSLSTLTLSKADYEEKHKVYACEVTHQGLSS PVTKSFNRGEC

2.1.2 Preparation of monovalent FAP (4B9) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains without charged residues (Construct 2.2)

5 A polypeptide containing two ectodomains of 4-1BB ligand (71-254), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned as depicted in Figure 1A: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL.

A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the human IgG1-CH1 domain, was cloned as described in Figure 1B: human 4-1BB ligand, (G4S)₂ connector, human CH1.

The polypeptide encoding the dimeric 4-1BB ligand fused to human CL domain was subcloned in frame with the human IgG1 heavy chain CH2 and CH3 domains on the knob (Merchant, Zhu et al. 1998) using a linker (G4S)₂ or, alternatively (GSPGSSSSGS).

The variable region of heavy and light chain DNA sequences encoding a binder specific for fibroblast activation protein (FAP), clone 4B9, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1.

The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors (WO 2012/130831).

Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations,
 5 the monomeric CH1 fusion, the targeted anti-FAP-Fc hole chain containing the
 Y349C/T366S/L368A/Y407V mutations and the anti-FAP light chain allows generation of a
 heterodimer, which includes an assembled trimeric 4-1BB ligand and a FAP binding Fab (Figure
 4, Construct 2.2).

Table 22 shows the cDNA and amino acid sequences of the monovalent FAP (4B9)-human
 10 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule containing CH1-CL crossover
 without charged residues (Construct 2.2).

Table 22: Sequences of monovalent FAP(4B9)-targeted human 4-1BB ligand (71-254) containing Fc (kih) fusion molecule Construct 2.2

SEQ ID NO:	Description	Sequence
165	Dimeric hu 4- 1BBL (71-254) – CL Fc knob chain	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAACTGGTGG TGGCCAAGGCCGCGGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGCCCCGGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCT CCAAGAAGCGAAGGCGGAGGCGGATCTGGCGGCGGAG GATCTAGAGAGGGACCCGAAGTGTCCCCTGACGATCCA GCCGGGCTGCTGGATCTGAGACAGGGAATGTTCCGCCA GCTGGTGGCTCAGAATGTGCTGCTGATTGACGGACCTCT GAGCTGGTACTCCGACCCAGGGCTGGCAGGGGTGTCCC TGACTGGGGGACTGTCCTACAAAGAAGATACAAAAGAA CTGGTGGTGGCTAAAGCTGGGGTGTACTATGTGTTTTT CAGCTGGAAGTGAAGGCGGGTGGTGGCTGGGGAGGGCTC AGGATCTGTGTCCCTGGCTCTGCATCTGCAGCCACTGCG CTCTGCTGCTGGCGCAGCTGCACTGGCTCTGACTGTGGA CCTGCCACCAGCCTCTAGCGAGGCCAGAAACAGCGCCT TCGGGTTCCAAGGACGCCTGCTGCATCTGAGCGCCGGAC AGCGCCTGGGAGTGCATCTGCATACTGAAGCCAGAGCC CGGCATGCTTGGCAGCTGACTCAGGGGGCAACTGTGCTG GGACTGTTTCGCGTGACACCTGAGATCCCTGCCGGACTG

		<p>CCAAGCCCTAGATCAGAAGGGGGCGGAGGTTCCGGAGG GGGAGGATCTCGTACGGTGGCCGCTCCCTCCGTGTTTAT CTTTCCCCCATCCGATGAACAGCTGAAAAGCGGCACCGC CTCCGTCTGTGTCTGCTGAACAATTTTACCCTAGGGA AGCTAAAGTGCAGTGGAAAGTGGATAACGCACTGCAGT CCGGCAACTCCCAGGAATCTGTGACAGAACAGGACTCC AAGGACAGCACCTACTCCCTGTCCTCCACCCTGACACTG TCTAAGGCTGATTATGAGAAACACAAAGTCTACGCCTGC GAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCACAAA GAGCTTCAACAGGGGAGAGTGTGACAAGACCCACACCT GTCCCCCTTGTCTGCCCCCTGAAGCTGCTGGCGGCCCTT CTGTGTTCTGTCTCCCCCAAGGCCAAGGACACCCTGA TGATCAGCCGGACCCCCGAAGTGACCTGCGTGGTGGTG GATGTGTCCACGAGGACCCTGAAGTGAAGTTCAATTGG TACGTGGACGGCGTGGAAAGTGCACAATGCCAAGACCAA GCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGG TCAGCGTCTCACCGTCTGCACCAGGACTGGCTGAATG GCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTC GGCGCCCCATCGAGAAAACCATCTCCAAAGCCAAAGG GCAGCCCCGAGAACCACAGGTGTACACCCTGCCCCCAT GCCGGGATGAGCTGACCAAGAACCAGGTCAGCCTGTGG TGCCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGTG GAGTGGGAGAGCAATGGGCAGCCGGAGAACAACACTACAA GACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTT CCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGC AGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGG CTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGT CTCCGGGTAAA</p>
166	Monomeric hu 4- 1BBL (71-254) – CH1	<p>AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAAGTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCTCTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGCCCGGAATAGCGCATTTGGGTTT CAAGGCCGGCTGCTGCACCTGTCTGCCGGCCAGAGACT GGGAGTGCATCTGCACACAGAGGCCAGAGCCAGGCACG CCTGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCTG TTCAGAGTGACCCCCGAGATTCTGCGCGCCTGCCTAGC CCTAGATCTGAAGGCGGCGGAGGTTCCGGAGGCGGAGG ATCTGCTAGCACCAAAGGCCCTTCCGTGTTTCTCTGGC TCCTAGCTCCAAGTCCACCTCTGGAGGCACCGCTGCTCT CGGATGCCTCGTGAAGGATTATTTTCTGAGCCTGTGAC AGTGTCTTGAATAGCGGAGCACTGACCTCTGGAGTGC ATACTTTCCCCGCTGTGCTGCAGTCTCTGGACTGTACA GCCTGAGCAGCGTGGTGACAGTGCCAGCAGCAGCCTG GGCACCCAGACCTACATCTGCAACGTGAACCACAAGCC CAGCAACACCAAGGTGGACAAGAAGGTGGAACCCAAGT CTTGT</p>

162	anti-FAP (4B9) Fc hole chain	see Table 21
163	anti-FAP (4B9) light chain	see Table 21
117	Dimeric hu 4- 1BBL (71-254) – CL Fc knob chain	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLPSRSEGGGGSGGGGSREGPEL SPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLA GVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELRRVVAG EGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARN AFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATV LGLFRVTPEIPAGLPSRSEGGGGSGGGGSRTVAAPSVFIFP PSDEQLKSGTASVCLLN NFYPREAKVQWKVDNALQSGN SQESVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTH QGLSSPVTKSFNREGCDKTHTCPPCPAPEAAGGPSVFLFP KPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEV HNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKV SNKALGAPIEKTISKAKGQPREPQVYTLPPCRDELTKNQVS LWCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSF FLYSKLTVDKSRWQQGNV FSCSV MHEALHNHYTQKSLSL SPGK
118	Monomeric hu 4- 1BBL (71-254) – CH1	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLPSRSEGGGGSGGGGSASTKGP SVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALT SGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHK PSNTKVDKCKVEPKSC
164	anti-FAP (4B9) Fc hole chain	see Table 21
125	anti-FAP (4B9) light chain	see Table 21

2.1.3 Preparation of bivalent FAP (4B9) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding molecule with the dimeric and monomeric 4-1BB ligands fused at the C-terminus of each heavy chain (Construct 2.3)

A polypeptide containing two ectodomains of 4-1BB ligand (71-254), separated by (G4S)₂ linkers was fused to the C-terminus of human IgG1 Fc hole chain, as depicted in Figure 1C:
 5 human IgG1 Fc hole, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human 4-1BB

ligand. A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the C-terminus of human IgG1 Fc knob chain as described in Figure 1D: human IgG1 Fc knob, (G4S)₂ connector, human 4-1BB ligand.

The polypeptide encoding the dimeric 4-1BB ligand was subcloned in frame at the C-terminus of human IgG1 heavy chain CH2 and CH3 domains on the hole (Merchant, Zhu et al. 1998) using a (G4S)₂ connector. The polypeptide encoding the monomeric 4-1BB ligand was subcloned in frame at the C-terminus of human IgG1 heavy chain CH2 and CH3 domains on the knob (Merchant, Zhu et al. 1998) using a (G4S)₂ connector.

The variable region of heavy and light chain DNA sequences encoding a binder specific for fibroblast activation protein (FAP), clone 4B9, were subcloned in frame with either the constant heavy chain of the hole, the knob or the constant light chain of human IgG1.

The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831.

Combination of the anti-FAP huIgG1 hole dimeric ligand chain containing the Y349C/T366S/L368A/Y407V mutations, the anti-FAP huIgG1 knob monomeric ligand chain containing the S354C/T366W mutations and the anti-FAP light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and two FAP binding Fabs (Figure 4, Construct 2.3)

Table 23 shows the cDNA and amino acid sequences of the bivalent FAP (4B9)-targeted 4-1BB ligand trimer-containing Fc (kih) fusion molecule Construct 2.3 (FAP split trimer with 2 anti-FAP Fabs, dimeric and monomeric 4-1BB ligand fused at the C-terminus of each heavy chain, respectively).

Table 23: Sequences of bivalent FAP(4B9)-targeted human 4-1BB ligand (71-254) containing Fc (kih) fusion molecule Construct 2.3

SEQ ID NO:	Description	Sequence
167	anti-FAP (4B9) Fc hole chain fused to dimeric hu 4-1BBL (71-254)	GAGGTGCAGCTGCTCGAAAGCGGCGGAGGACTGGTGCA GCCTGGCGGCAGCCTGAGACTGTCTTGCGCCGCCAGCG GCTTCACCTTCAGCAGCTACGCCATGAGCTGGGTCCGCC AGGCCCTGGCAAGGGACTGGAATGGGTGTCCGCCATC ATCGGCTCTGGCGCCAGCACCTACTACGCCGACAGCGTG AAGGGCCGGTTCACCATCAGCCGGGACAACAGCAAGAA CACCCTGTACCTGCAGATGAACAGCCTGCGGGCCGAGG ACACCGCCGTGTACTACTGCGCCAAGGGATGGTTCCGGC GGCTTCAACTACTGGGGACAGGGCACCCCTGGTCACAGT

		<p>GTCCAGCGCTAGCACCAAGGGCCCCTCCGTGTTCCCCCT GGCCCCCAGCAGCAAGAGCACCAGCGGCGGCACAGCCG CTCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAGCCCG TGACCGTGTCTTGGAACAGCGGAGCCCTGACCTCCGGC GTGCACACCTTCCCCGCCGTGCTGCAGAGTTCTGGCCTG TATAGCCTGAGCAGCGTGGTCACCGTGCCTTCTAGCAGC CTGGGCACCCAGACCTACATCTGCAACGTGAACCACAA GCCCAGCAACACCAAGGTGGACAAGAAGGTGGAGCCCA AGAGCTGCGACAAAACCTCACACATGCCACCGTGCCCA GCACCTGAAGCTGCAGGGGGACCGTCAGTCTTCCTCTTC CCCCAAAACCCAAGGACACCCTCATGATCTCCCGGACC CCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGA AGACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCG TGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAG CAGTACAACAGCACGTACCGTGTGGTCAGCGTCCTCACC GTCCTGCACCAGGACTGGCTGAATGGCAAGGAGTACAA GTGCAAGGTCTCCAACAAAGCCCTCGGCGCCCCCATCG AGAAAACCATCTCCAAAGCCAAAGGGCAGCCCCGAGAA CCACAGGTGTGCACCCTGCCCCCATCCCGGGATGAGCTG ACCAAGAACCAGGTCAGCCTCTCGTGCAGTCAAAGG CTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGCA ATGGGCAGCCGGAGAACAACCTACAAGACCACGCCTCCC GTGCTGGACTCCGACGGCTCCTTCTTCCTCGTGAGCAAG CTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGT CTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCA CTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTGGAGG CGGCGGAAGCGGAGGAGGAGGATCCAGAGAGGGCCCT GAGCTGAGCCCCGATGATCCTGCTGGACTGCTGGACCTG CGGCAGGGCATGTTTGCTCAGCTGGTGGCCAGAACGTG CTGCTGATCGATGGCCCCCTGTCCTGGTACAGCGATCCT GGACTGGCTGGCGTGTCACTGACAGGCGGCCTGAGCTA CAAAGAGGACACCAAGAAGAACTGGTGGTGGCCAAGGCCG GCGTGTA CTACGTGTTCTTTCAGCTGGAAGTGCAGGAGAG TGGTGGCCGGCGAAGGATCTGGCTCTGTGTCTCTGGCCC TGCATCTGCAGCCTCTGAGAAGCGCTGCTGGCGCTGCAG CTCTGGCACTGACAGTGGATCTGCCTCCTGCCAGCTCCG AGGCCCGGAATAGCGCATTTGGGTTTCAAGGCAGGCTG CTGCACCTGTCTGCCGGCCAGAGGCTGGGAGTGCATCTG CACACAGAGGCCAGGGCTAGACACGCCTGGCAGCTGAC ACAGGGCGCTACAGTGCTGGGCCTGTTCAAGAGTGACCC CCGAGATTCCAGCCGGCCTGCCTTCTCCAAGAAGCGAA GGCGGAGGCGGATCTGGCGGCGGAGGATCTAGAGAGGG ACCCGAAGTGTCCCCTGACGATCCAGCCGGGCTGCTGGA TCTGAGACAGGGAATGTTCCGCCAGCTGGTGGCTCAGA ATGTGCTGCTGATTGACGGACCTCTGAGCTGGTACTCCG ACCCAGGGCTGGCAGGGGTGTCCCTGACTGGGGGACTG TCCTACAAAGAAGATACAAAAGAAGTGGTGGTGGCTAA AGCTGGGGTGTACTATGTGTTTTTTCAGCTGGAAGTGA GCGGGTGGTGGCTGGGGAGGGCTCAGGATCTGTGTCCCT GGCTCTGCATCTGCAGCCACTGCGCTCTGCTGCTGGCGC AGCTGCACTGGCTCTGACTGTGGACCTGCCACCAGCCTC TAGCGAGGCCAGAAACAGCGCCTTCGGGTTCGAAGGAC GCCTGCTGCATCTGAGCGCCGGACAGCGCCTGGGAGTG CATCTGCATACTGAAGCCAGAGCCCGGCATGCTTGGCA</p>
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		GCTGACTCAGGGGGCAACTGTGCTGGGACTGTTTCGCGT GACACCTGAGATCCCTGCCGGACTGCCAAGCCCTAGATC AGAA
168	anti-FAP (4B9) Fc knob chain fused to monomeric hu 4- 1BBL (71-254)	GAGGTGCAGCTGCTCGAAAGCGGCGGAGGACTGGTGCA GCCTGGCGGCAGCCTGAGACTGTCTTGCGCCGCCAGCG GCTTCACCTTCAGCAGCTACGCCATGAGCTGGGTCCGCC AGGCCCCTGGCAAGGGACTGGAATGGGTGTCCGCCATC ATCGGCTCTGGCGCCAGCACCTACTACGCCGACAGCGTG AAGGGCCGGTTCACCATCAGCCGGGACAACAGCAAGAA CACCTGTACCTGCAGATGAACAGCCTGCGGGCCGAGG ACACCGCCGTGTACTACTGCGCCAAGGGATGGTTCGGC GGCTTCAACTACTGGGGACAGGGCACCCCTGGTCACAGT GTCCAGCGCTAGCACCAAGGGCCCATCGGTCTTCCCCCT GGCACCCCTCCTCCAAGAGCACCTCTGGGGGCACAGCGG CCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGG TGACGGTGTGCTGGAATCAGGCGCCCTGACCAGCGGC GTGCACACCTTCCCGGCTGTCCTACAGTCCTCAGGACTC TACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGCAGC TTGGGCACCCAGACCTACATCTGCAACGTGAATCACAA GCCCAGCAACACCAAGGTGGACAAGAAAGTTGAGCCCA AATCTTGTGACAAAACCTCACACATGCCACCGTGCCAG CACCTGAAGCTGCAGGGGGACCGTCAGTCTTCCTCTTCC CCCCAAAACCCAAGGACACCCTCATGATCTCCCGGACCC CTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGAA GACCCTGAGGTCAAGTCAACTGGTACGTGGACGGCGT GGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGC AGTACAACAGCACGTACCGTGTGGTCAGCGTCCTCACCG TCCTGCACCAGGACTGGCTGAATGGCAAGGAGTACAAG TGCAAGGTCTCCAACAAAGCCCTCGGCGCCCCCATCGA GAAAACCATCTCCAAGCCAAAGGGCAGCCCCGAGAAC CACAGGTGTACACCCTGCCCCCTGCAGAGATGAGCTG ACCAAGAACCAGGTGTCCCTGTGGTGTCTGGTCAAGGGC TTCTACCCAGCGATATCGCCGTGGAGTGGGAGAGCAA CGGCCAGCCTGAGAACAACCTACAAGACCACCCCCCTG TGCTGGACAGCGACGGCAGCTTCTTCTGTACTCCAAAC TGACCGTGGACAAGAGCCGGTGGCAGCAGGGCAACGTG TTCAGCTGCAGCGTGATGCACGAGGCCCTGCACAACCA CTACACCCAGAAGTCCCTGAGCCTGAGCCCCGGCGGAG GCGGCGGAAGCGGAGGAGGAGGATCCAGAGAGGGCCC TGAGCTGAGCCCCGATGATCCTGCTGGACTGCTGGACCT GCGGCAGGGCATGTTTGCTCAGCTGGTGGCCAGAACGT GCTGCTGATCGATGGCCCCCTGTCTTGGTACAGCGATCC TGGACTGGCTGGCGTGTCACTGACAGGCGGCCTGAGCTA CAAAGAGGACACCAAAGAACTGGTGGTGGCCAAGGCCG GCGTGTACTACGTGTTCTTTCAGCTGGAAGTGCAGGAGAG TGGTGGCCGGCGAAGGATCTGGCTCTGTGTCTCTGGCCC TGCATCTGCAGCCTCTGAGAAGCGCTGCTGGCGCTGCAG CTCTGGCACTGACAGTGGATCTGCCTCCTGCCAGCTCCG AGGCCCGGAATAGCGCATTTGGGTTTCAAGGCAGGCTG CTGCACCTGTCTGCCGGCCAGAGGCTGGGAGTGCATCTG CACACAGAGGCCAGGGCTAGACACGCCTGGCAGCTGAC ACAGGGCGCTACAGTGCTGGGCCTGTTCAAGAGTGACCC

		CCGAGATTCCAGCCGGCCTGCCTTCTCCAAGAAGCGAA
163	anti-FAP (4B9) light chain	see Table 21
123	anti-FAP (4B9) Fc hole chain fused to dimeric hu 4-1BBL (71- 254)	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSYAMSWVRQA PGKGLEWVSAIIGSGASTYYADSVKGRFTISRDN SKNTLYL QMNSLRAEDTAVYYCAKGWFGGFNYWGQGLTVTVSSAS TKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNS GALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICN VNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVF LFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDG VEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYK CKVSNKALGAPIEKTISKAKGQPREPQVCTLPSPRDELTKN QVSLSCAVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSD GSFFLVSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKS LSLSPGGGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQ LVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELV VAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAA GAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGV HLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSEG GGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQLVAQNV LLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELV VAKAGV YYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALA LTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEA RARHAWQLTQGATVLGLFRVTPEIPAGLPSRSE
124	anti-FAP (4B9) Fc knob chain fused to monomeric hu 4- 1BBL (71-254)	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSYAMSWVRQA PGKGLEWVSAIIGSGASTYYADSVKGRFTISRDN SKNTLYL QMNSLRAEDTAVYYCAKGWFGGFNYWGQGLTVTVSSAS TKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNS GALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICN VNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVF LFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDG VEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYK CKVSNKALGAPIEKTISKAKGQPREPVYTLPPCRDELTKN QVSLWCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSD GSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKS LSLSPGGGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQ LVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELV VAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAA GAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGV HLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSE
125	anti-FAP (4B9) light chain	see Table 21

2.1.4 Preparation of monovalent FAP (4B9) targeted 4-1BB ligand (71-248) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains with charged residues (Construct 2.4)

A polypeptide containing two ectodomains of 4-1BB ligand (71-248), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned as depicted in Figure 1A: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL. A polypeptide containing one ectodomain of 4-1BB ligand (71-248) and fused to the human IgG1-CH domain, was cloned as described in Figure 1B: human 4-1BB ligand, (G4S)₂ connector, human CH.

The polypeptide encoding the dimeric 4-1BB ligand fused to human CL domain was subcloned in frame with the human IgG1 heavy chain CH2 and CH3 domains on the knob (Merchant, Zhu et al. 1998) using a linker (G4S)₂ or, alternatively (GSPGSSSSGS). To improve correct pairing the following mutations have been introduced in the crossed CH-CL. In the dimeric 4-1BB ligand fused to human CL, E123R and Q124K. In the monomeric 4-1BB ligand fused to human CH1, K147E and K213E.

The variable region of heavy and light chain DNA sequences encoding a binder specific for fibroblast activation protein (FAP), clone 4B9, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1.

The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831.

Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-FAP-Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations and the anti-FAP light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a FAP binding Fab (Figure 4, Construct 2.4).

Table 24 shows the cDNA and amino acid sequences of the monovalent FAP (4B9)-human 4-1BB ligand (71-248) Fc (kih) fusion antigen binding molecule containing CH1-CL crossover with charged residues (Construct 2.4).

Table 24: Sequences of monovalent FAP(4B9)-targeted human 4-1BB ligand (71-248) containing Fc (kih) fusion molecule Construct 2.4

SEQ ID NO:	Description	Sequence
169	Dimeric hu 4-1BBL (71-248) – CL* Fc knob chain	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAACTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGATCTGCT GCTGGCGCCGCTGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCAGCGAGGCCCCGGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGTGGGCCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGACTGGGAGG CGGCGGATCTGGCGGCGGAGGATCTAGAGAAGGACCCG AGCTGTCCCCTGACGATCCAGCCGGGCTGCTGGATCTGA GACAGGGAATGTTCCGCCAGCTGGTGGCTCAGAATGTG CTGCTGATTGACGGACCTCTGAGCTGGTACTCCGACCCA GGGCTGGCAGGGGTGTCCCTGACTGGGGGACTGTCCTAC AAAGAAGATACAAAAGAACTGGTGGTGGCTAAAGCTGG GGTGTACTATGTGTTTTTTCAGCTGGAAGTGAAGCGGGT GGTGGCTGGGGAGGGCTCAGGATCTGTGTCCCTGGCTCT GCATCTGCAGCCACTGCGCTCTGCAGCAGGGGCTGCAG CACTGGCCCTGACTGTGGACCTGCCCCCAGCTTCTTCCG AGGCCAGAAACAGCGCCTTCGGGTTCCAAGGACGCCTG CTGCATCTGAGCGCCGGACAGCGCCTGGGAGTGCATCT GCATACTGAAGCCAGAGCCCGGCATGCTTGGCAGCTGA CTCAGGGGGCAACTGTGCTGGGACTGTTTCGCGTGACAC CTGAGATCCCCGCTGGACTGGGCGGAGGCGGTTCCGGA GGGGGAGGATCTCGTACGGTGGCTGCACCATCTGTCTTT ATCTTCCCACCCAGCGACCGGAAGCTGAAGTCTGGCAC AGCCAGCGTCGTGTGCCTGCTGAATAACTTCTACCCCCG CGAGGCCAAGGTGCAGTGGAAGGTGGACAATGCCCTGC AGAGCGGCAACAGCCAGGAAAGCGTGACCGAGCAGGA CAGCAAGGACTCCACCTACAGCCTGAGCAGCACCTGA CCCTGAGCAAGGCCGACTACGAGAAGCACAAGGTGTAC GCCTGCGAAGTGACCCACCAGGGCCTGTCTAGCCCCGTG ACCAAGAGCTTCAACCGGGGCGAGTGCGACAAGACCCA CACCTGTCTCCATGCCCTGCCCCCTGAAGCTGCTGGCGG CCCTAGCGTGTTCTGTTCCTTCCCCCAAGCCCAAGGACAC CCTGATGATCAGCCGGACCCCTGAAGTGACCTGCGTGGT GGTGGATGTGTCCACGAGGACCCCTGAAGTGAAGTTCA ATTGGTACGTGGACGGCGTGGAAGTGCACAATGCCAAG ACCAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCG TGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCT GAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAG CCCTCGGCGCCCCCATCGAGAAAACCATCTCCAAGGCC

		AAAGGGCAGCCCCGAGAACCACAGGTGTACACCCTGCC CCCATGCCGGGATGAGCTGACCAAGAACCAGGTCAGCC TGTGGTGCCTGGTCAAAGGCTTCTATCCCAGCGACATCG CCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAACAAC TACAAGACCACGCCTCCCGTGCTGGACTCCGACGGCTCC TTCTTCCTCTACAGCAAGCTCACCGTGGACAAGAGCAGG TGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCAT GAGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTC CCTGTCTCCGGGTAAA
170	Monomeric hu 4- 1BBL (71-248) – CH1*	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAAGTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGATCTGCT GCTGGCGCCGCTGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCAGCGAGGCCCCGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGTGGGCCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGACTGGGAGG CGGAGGTTCCGGAGGCGGAGGATCTGCTAGCACAAAGG GCCCCAGCGTGTTCCCTCTGGCCCCCTAGCAGCAAGAGCA CATCTGGCGGAACAGCCGCCCTGGGCTGCCTGGTGGAA GATTACTTCCCCGAGCCCGTGACCGTGTCTTGGAAATTCT GGCGCCCTGACAAGCGGCGTGCACACCTTTCCAGCCGTG CTGCAGAGCAGCGGCCTGTACTCTCTGAGCAGCGTCGTG ACAGTGCCCAGCAGCTCTCTGGGCACCCAGACCTACATC TGCAACGTGAACCACAAGCCCAGCAACACCAAGGTGGA CGAGAAGGTGGAACCCAAGTCCTGC
162	anti-FAP (4B9) Fc hole chain	see Table 21
163	anti-FAP (4B9) light chain	see Table 21
119	Dimeric hu 4- 1BBL (71-248) – CL* Fc knob chain	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAAGVYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLGGGGSGGGGSREGPELSPDDPA GLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTG GLSYKEDTKELVVAAGVYVFFQLELRRVVAGEGSGSV SLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQ RLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRV TPEIPAGLGGGGSGGGGSRTVAAPSVFIFPPSDRKLKSGTAS VVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKD STYLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNR GECDKTHTCPPCPAPEAAGGPSVFLFPPKPKDTLMISRTPE

		VTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQY NSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGAPIEKTI SKAKGQPREPQVYTLPPCRDELTKNQVSLWCLVKGFYPSD IAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKLTVDKSR WQQGNVFSCSVMHEALHNHYTQKSLSLSPGK
120	Monomeric hu 4- 1BB (71-248) – CH1*	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWY SDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELR RVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASS EARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLT QGATVLGLFRVTPEIPAGLGGGGSGGGGSASTKGPSVFPLA PSSKSTSGGTAALGCLVEDYFPEPVTVSWNSGALTSGVHTF PAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKV DEKVEPKSC
164	anti-FAP (4B9) Fc hole chain	see Table 21
125	anti-FAP (4B9) light chain	see Table 21

2.1.5 Preparation of monovalent FAP (4B9) targeted 4-1BB ligand (71-248) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains without charged residues (Construct 2.5)

5 A polypeptide containing two ectodomains of 4-1BB ligand (71-248), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned as depicted in Figure 1A: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL. A polypeptide containing one ectodomain of 4-1BB ligand (71-248) and fused to the human IgG1-CH domain, was cloned as described in Figure 1B: human 4-1BB ligand, (G4S)₂ connector,
10 human CH.

The polypeptide encoding the dimeric 4-1BB ligand fused to human CL domain was subcloned in frame with the human IgG1 heavy chain CH2 and CH3 domains on the knob (Merchant, Zhu et al. 1998) using a linker (G4S)₂ or, alternatively (GSPGSSSSGS).

15 The variable region of heavy and light chain DNA sequences encoding a binder specific for fibroblast activation protein (FAP), clone 4B9, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1.

The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831.

Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-FAP-Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations and the anti-FAP light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a FAP binding Fab (Figure 4, Construct 2.5)

Table 25 shows the cDNA and amino acid sequences of the monovalent FAP (4B9)-human 4-1BB ligand (71-248) Fc (kih) fusion antigen binding molecule containing CH1-CL crossover without charged residues (Construct 2.5).

Table 25: Sequences of monovalent FAP(4B9)-targeted human 4-1BB ligand (71-248) containing Fc (kih) fusion molecule Construct 2.5

SEQ ID NO:	Description	Sequence
171	nucleotide sequence dimeric hu 4-1BBL (71-248) – CL Fc knob chain	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAACTGGTGG TGGCCAAGGCCGCGGTGTACTACGTGTTCTTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGATCTGCT GCTGGCGCCGCTGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCAGCGAGGCCCCGGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGTGGGCCCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGACTGGGAGG CGGCGGATCTGGCGGCGGAGGATCTAGAGAAGGACCCG AGCTGTCCCCTGACGATCCAGCCGGGCTGCTGGATCTGA GACAGGGAATGTTCCGCCAGCTGGTGGCTCAGAATGTG CTGCTGATTGACGGACCTCTGAGCTGGTACTCCGACCCA GGGCTGGCAGGGGTGTCCCTGACTGGGGGACTGTCCTAC AAAGAAGATACAAAAGAACTGGTGGTGGCTAAAGCTGG GGTGTACTATGTGTTTTTTCAGCTGGAAGTGAAGCGGGT GGTGGCTGGGGAGGGCTCAGGATCTGTGTCCCTGGCTCT GCATCTGCAGCCACTGCGCTCTGCAGCAGGGGCTGCAG CACTGGCCCTGACTGTGGACCTGCCCCCAGCTTCTTCCG AGGCCAGAAACAGCGCCTTCGGGTTCCAAGGACGCCTG CTGCATCTGAGCGCCGGACAGCGCCTGGGAGTGCATCT GCATACTGAAGCCAGAGCCCGGCATGCTTGGCAGCTGA CTCAGGGGGCAACTGTGCTGGGACTGTTTCGCGTGACAC CTGAGATCCCCGCTGGACTGGGCGGAGGCGGTTCCGGA GGGGGAGGATCTCGTACGGTGGCCGCTCCCTCCGTGTTT ATCTTTCCCCCATCCGATGAACAGCTGAAAAGCGGCACC GCCTCCGTCTGTGTCTGCTGAACAATTTTACCCTAGG GAAGCTAAAGTGCAGTGGAAAGTGGATAACGCACTGCA GTCCGGCAACTCCCAGGAATCTGTGACAGAACAGGACT

		CCAAGGACAGCACCTACTCCCTGTCCTCCACCCTGACAC TGTCTAAGGCTGATTATGAGAAACACAAAGTCTACGCCT GCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCACA AAGAGCTTCAACAGGGGAGAGTGTGACAAGACCCACAC CTGTCCCCCTTGTCTGCCCCCTGAAGCTGCTGGCGGCCC TTCTGTGTTCTGTTCCTGTTCCCCCAAGCCCAAGGACACCCT GATGATCAGCCGACCCCCGAAGTGACCTGCGTGTTGG TGGATGTGTCCACAGAGGACCCTGAAGTGAAGTTCAATT GGTACGTGGACGGCGTGGAAGTGCACAATGCCAAGACC AAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGT GGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAA TGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCC TCGGCGCCCCCATCGAGAAAACCATCTCCAAGCCAAA GGGCAGCCCCGAGAACCACAGGTGTACACCCTGCCCCC ATGCCGGGATGAGCTGACCAAGAACCAGGTACGCCTGT GGTGCCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCG TGGAGTGGGAGAGCAATGGGCAGCCGGAGAACAACACTAC AAGACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTC TTCCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTG GCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGA GGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCT GTCTCCGGGTAAA
172	Monomeric hu 4-1BBL (71-248) – CH1	AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAAGTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGATCTGCT GCTGGCGCCGCTGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCAGCGAGGCCCCGGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCTG TTCAGAGTGACCCCCGAGATTCCAGCCGACTGGGAGG CGGAGGTTCCGGAGGCGGAGGATCTGCTAGCACCAAAG GCCCTTCCGTGTTTCTCTGGCTCCTAGCTCCAAGTCCAC CTCTGGAGGCACCGCTGCTCTCGGATGCCTCGTGAAGGA TTATTTTCCTGAGCCTGTGACAGTGTCTTGAATAGCGG AGCACTGACCTCTGGAGTGCATACTTTCCCCGCTGTGCT GCAGTCCTCTGGACTGTACAGCCTGAGCAGCGTGGTGAC AGTGCCAGCAGCAGCCTGGGCACCCAGACCTACATCT GCAACGTGAACCACAAGCCCAGCAACACCAAGGTGGAC AAGAAGGTGGAACCCAAGTCTTGT
162	anti-FAP (4B9) Fc hole chain	see Table 21
163	anti-FAP (4B9) light chain	see Table 21

173	Dimeric hu 4-1BBL (71-248) – CL Fc knob chain	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLGGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLGGGGSGGGGSRTVAAPSVFIFPPSDEQLKSGTASVVCLLNFPYPREAKVQWKVDNALQSGNSQESVTEQDSKDESTYLSSTLTLSKADYEKHKVYACEVTHQGLSPPVTKSFNRGECDKTHTCPPCPAPEAAGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGAPIEKTI SKAKGQPREPQVYTLPPCRDELTKNQVSLWCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKLTVDKSRWQQGNVVFSCSVMHEALHNHYTQKSLSLSPGK
174	Monomeric hu 4-1BBL (71-248) – CH1	REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFFQLELRVVAGEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLGGGGSGGGGSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKKVEPKSC
164	anti-FAP (4B9) Fc hole chain	see Table 21
125	anti-FAP (4B9) light chain	see Table 21

2.1.6 Preparation of bivalent FAP (4B9) targeted 4-1BB ligand (71-248) trimer-containing Fc (kih) fusion antigen binding molecule with the dimeric and monomeric 4-1BB ligands fused at the C-terminus of each heavy chain (Construct 2.6)

- 5 A polypeptide containing two ectodomains of 4-1BB ligand (71-248), separated by (G4S)₂ linkers was fused to the C-terminus of human IgG1 Fc hole chain, as depicted in Figure 1C: human IgG1 Fc hole, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand. A polypeptide containing one ectodomain of 4-1BB ligand (71-248) and fused to the C-terminus of human IgG1 Fc knob chain as described in Figure 1D: human IgG1 Fc knob, (G4S)₂ connector, human 4-1BB ligand.
- 10

The polypeptide encoding the dimeric 4-1BB ligand was subcloned in frame at the C-terminus of human IgG1 heavy chain CH2 and CH3 domains on the hole (Merchant, Zhu et al. 1998) using a (G4S)2 connector. The polypeptide encoding the monomeric 4-1BB ligand was subcloned in frame at the C-terminus of human IgG1 heavy chain CH2 and CH3 domains on the knob (Merchant, Zhu et al. 1998) using a (G4S)2 connector.

The variable region of heavy and light chain DNA sequences encoding a binder specific for fibroblast activation protein (FAP), clone 4B9, were subcloned in frame with either the constant heavy chain of the hole, the knob or the constant light chain of human IgG1.

The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831.

Combination of the anti-FAP huIgG1 hole dimeric ligand chain containing the Y349C/T366S/L368A/Y407V mutations, the anti-FAP huIgG1 knob monomeric ligand chain containing the S354C/T366W mutations and the anti-FAP light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and two FAP binding Fabs (Figure 4, Construct 2.6).

Table 26 shows the cDNA and amino acid sequences of the bivalent FAP (4B9)-targeted 4-1BB ligand trimer-containing Fc (kih) fusion molecule Construct 2.6 (FAP split trimer with 2 anti-FAP Fabs, dimeric and monomeric 4-1BB ligand fused at the C-terminus of each heavy chain, respectively).

Table 26: Sequences of bivalent FAP(4B9)-targeted human 4-1BB ligand (71-248) containing Fc (kih) fusion molecule Construct 2.6

SEQ ID NO:	Description	Sequence
175	nucleotide sequence of anti-FAP (4B9) Fc hole chain fused to dimeric hu 4-1BBL (71-248)	GAGGTGCAGCTGCTCGAAAGCGGCGGAGGACTGGTGCA GCCTGGCGGCAGCCTGAGACTGTCTTGCGCCGCCAGCG GCTTCACCTTCAGCAGCTACGCCATGAGCTGGGTCCGCC AGGCCCTGGCAAGGGACTGGAATGGGTGTCCGCCATC ATCGGCTCTGGCGCCAGCACCTACTACGCCGACAGCGTG AAGGGCCGGTTCACCATCAGCCGGGACAACAGCAAGAA CACCTGTACCTGCAGATGAACAGCCTGCGGGCCGAGG ACACCGCCGTGTACTACTGCGCCAAGGGATGGTTCGGC GGCTTCACTACTGGGGACAGGGCACCCCTGGTCACAGT GTCCAGCGCTAGCACCAAGGGCCCCTCCGTGTTCCCCCT GGCCCCAGCAGCAAGAGCACCAGCGGCGGCACAGCCG CTCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAGCCCG TGACCGTGTCTGGAACAGCGGAGCCCTGACCTCCGGC GTGCACACCTTCCCCGCCGTGCTGCAGAGTTCTGGCCTG

		<p> TATAGCCTGAGCAGCGTGGTCACCGTGCCTTCTAGCAGC CTGGGCACCCAGACCTACATCTGCAACGTGAACCACAA GCCCAGCAACACCAAGGTGGACAAGAAGGTGGAGCCCA AGAGCTGCGACAAAACCTCACACATGCCACCGTGCCCA GCACCTGAAGCTGCAGGGGGACCGTCAGTCTTCCTCTTC CCCCAAAACCCAAGGACACCCTCATGATCTCCCGGACC CCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGA AGACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCG TGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAG CAGTACAACAGCACGTACCGTGTGGTCAGCGTCCTCACC GTCCTGCACCAGGACTGGCTGAATGGCAAGGAGTACAA GTGCAAGGTCTCCAACAAAGCCCTCGGCGCCCCCATCG AGAAAACCATCTCCAAAGCCAAAGGGCAGCCCCGAGAA CCACAGGTGTGCACCCTGCCCCCATCCCGGGATGAGCTG ACCAAGAACCAGGTACGCCTCTCGTGCAGTCAAAGG CTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGCA ATGGGCAGCCGGAGAACAACATAAGACCACGCCTCCC GTGCTGGACTCCGACGGCTCCTTCTTCCTCGTGAGCAAG CTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGT CTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCA CTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTGGAGG CGGCGGAAGCGGAGGAGGAGGATCCAGAGAGGGCCCT GAGCTGAGCCCCGATGATCCTGCTGGACTGCTGGACCTG CGGCAGGGCATGTTTGCTCAGCTGGTGGCCAGAACGTG CTGCTGATCGATGGCCCCCTGTCCTGGTACAGCGATCCT GGACTGGCTGGCGTGTCACTGACAGGCGGCCTGAGCTA CAAAGAGGACACCAAAGAAGTGGTGGTGGCCAAGGCCG GCGTGTACTACGTGTTCTTTCAGCTGGAAGTGCAGGAGAG TGGTGGCCGGCGAAGGATCTGGCTCTGTGTCTCTGGCCC TGCATCTGCAGCCTCTGAGAAGCGCTGCTGGCGCTGCAG CTCTGGCACTGACAGTGGATCTGCCTCCTGCCAGCTCCG AGGCCCGGAATAGCGCATTTGGGTTTCAAGGCAGGCTG CTGCACCTGTCTGCCGGCCAGAGGCTGGGAGTGCATCTG CACACAGAGGCCAGGGCTAGACACGCCTGGCAGCTGAC ACAGGGCGCTACAGTGCTGGGCCTGTTCAAGAGTGACCC CCGAGATTCCAGCCGGCCTGGGCGGAGGCGGATCTGGC GGCGGAGGATCTAGAGAGGGACCCGAAGTGTCCCTGA CGATCCAGCCGGGCTGCTGGATCTGAGACAGGGAATGT TCGCCCAGCTGGTGGCTCAGAATGTGCTGCTGATTGACG GACCTCTGAGCTGGTACTCCGACCCAGGGCTGGCAGGG GTGTCCCTGACTGGGGGACTGTCTACAAAGAAGATAC AAAAGAAGTGGTGGTGGCTAAAGCTGGGGTGTACTATG TGTTTTTTCAGCTGGAAGTGAAGCGGGTGGTGGCTGGGG AGGGCTCAGGATCTGTGTCCCTGGCTCTGCATCTGCAGC CACTGCGCTCTGCTGCTGGCGCAGCTGCACTGGCTCTGA CTGTGGACCTGCCACCAGCCTCTAGCGAGGCCAGAAAC AGCGCCTTCGGGTCCAAGGACGCCTGCTGCATCTGAGC GCCGGACAGCGCCTGGGAGTGCATCTGCATACTGAAGC CAGAGCCCGGCATGCTTGGCAGCTGACTCAGGGGGCAA CTGTGCTGGGACTGTTTCGCGTGACACCTGAGATCCCTG CCGGACTG </p>
176	nucleotide sequence anti-	<p> GAGGTGCAGCTGCTCGAAAGCGGCGGAGGACTGGTGCA GCCTGGCGGCAGCCTGAGACTGTCTTGCGCCGCCAGCG </p>

	FAP (4B9) Fc knob chain fused to monomeric hu 4-1BBL (71-248)	GCTTCACCTTCAGCAGCTACGCCATGAGCTGGGTCCGCC AGGCCCCTGGCAAGGGACTGGAATGGGTGTCCGCCATC ATCGGCTCTGGCGCCAGCACCTACTACGCCGACAGCGTG AAGGGCCGGTTCACCATCAGCCGGGACAACAGCAAGAA CACCTGTACCTGCAGATGAACAGCCTGCGGGCCGAGG ACACCGCCGTGTACTACTGCGCCAAGGGATGGTTTCGGC GGCTTCAACTACTGGGGACAGGGCACCCCTGGTCACAGT GTCCAGCGCTAGCACCAAGGGCCCATCGGTCTTCCCCCT GGCACCTCCTCCAAGAGCACCTCTGGGGGCACAGCGG CCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGG TGACGGTGTCTGGAACCTCAGGCGCCCTGACCAGCGGC GTGCACACCTTCCCGGCTGTCTTACAGTCCTCAGGACTC TACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGCAGC TTGGGCACCCAGACCTACATCTGCAACGTGAATCACAA GCCCAGCAACACCAAGGTGGACAAGAAAGTTGAGCCCA AATCTTGTGACAAAACCTCACACATGCCACCGTGCCAG CACCTGAAGCTGCAGGGGGACCGTCAGTCTTCCCTCTCC CCCCAAAACCAAGGACACCTCATGATCTCCCGGACCC CTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGAA GACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGT GGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGC AGTACAACAGCACGTACCGTGTGGTCAGCGTCTCACCG TCCTGCACCAGGACTGGCTGAATGGCAAGGAGTACAAG TGCAAGGTCTCCAACAAAGCCCTCGGCGCCCCCATCGA GAAAACCATCTCCAAGCCAAAGGGCAGCCCCGAGAAC CACAGGTGTACACCCTGCCCCCTGCAGAGATGAGCTG ACCAAGAACCAGGTGTCCCTGTGGTGTCTGGTCAAGGGC TTCTACCCCAGCGATATCGCCGTGGAGTGGGAGAGCAA CGGCCAGCCTGAGAACAACCTACAAGACCACCCCCCTG TGCTGGACAGCGACGGCAGCTTCTTCTGTACTCCAAAC TGACCGTGGACAAGAGCCGGTGGCAGCAGGGCAACGTG TTCAGCTGCAGCGTGATGCACGAGGCCCTGCACAACCA CTACACCCAGAAGTCCCTGAGCCTGAGCCCCGGCGGAG GCGGCGGAAGCGGAGGAGGAGGATCCAGAGAGGGCCC TGAGCTGAGCCCCGATGATCCTGCTGGACTGCTGGACCT GCGGCAGGGCATGTTTGCTCAGCTGGTGGCCCAGAACGT GCTGCTGATCGATGGCCCCCTGTCTGGTACAGCGATCC TGGACTGGCTGGCGTGTCACTGACAGGCGGCCTGAGCTA CAAAGAGGACACCAAAGAAGTGGTGGTGGCCAAGGCCG GCGTGTACTACGTGTTCTTTCAGCTGGAAGTGCAGGAGAG TGGTGGCCGGCGAAGGATCTGGCTCTGTGTCTTGGCCC TGATCTGCAGCCTCTGAGAAGCGCTGCTGGCGCTGCAG CTCTGGCACTGACAGTGATCTGCCTCCTGCCAGCTCCG AGGCCCCGAATAGCGCATTTGGGTTTCAAGGCAGGCTG CTGCACCTGTCTGCCGGCCAGAGGCTGGGAGTGCATCTG CACACAGAGGCCAGGGCTAGACACGCCTGGCAGCTGAC ACAGGGCGCTACAGTGCTGGGCCTGTTTCAAGAGTGACCC CCGAGATTCCAGCCGGCCTG
163	anti-FAP (4B9) light chain	see Table 21

126	anti-FAP (4B9) Fc hole chain fused to dimeric hu 4-1BBL (71- 248)	EVQLLESGGGLVQPGGSLRLSCAASGFTSSYAMSWVRQA PGKGLEWVSAIIGSGASTYYADSVKGRFTISRDNKNTLYL QMNSLRAEDTAVYYCAKGWFGGFNYWGQGLTVTVSSAS TKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNS GALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICN VNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVF LFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDG VEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYK CKVSNKALGAPIEKTISKAKGQPREPQVCTLPSSRDELTKN QVSLSCAVKGFYPSDIAVEWESNGQPENNYKTPPVLDSD GSFFLVSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKS LSLSPGGGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQ LVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELV VAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAA GAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGV HLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLGGGGSG GGGSREGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGP LSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGVYYVFF QLELRRVVAGEGSGSVSLALHLQPLRSAAAGAAALALTVDL PPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHA WQLTQGATVLGLFRVTPEIPAGL
127	anti-FAP (4B9) Fc knob chain fused to monomeric hu 4- 1BBL (71-248)	EVQLLESGGGLVQPGGSLRLSCAASGFTSSYAMSWVRQA PGKGLEWVSAIIGSGASTYYADSVKGRFTISRDNKNTLYL QMNSLRAEDTAVYYCAKGWFGGFNYWGQGLTVTVSSAS TKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNS GALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICN VNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVF LFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDG VEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYK CKVSNKALGAPIEKTISKAKGQPREPQVYTLPPCRDELTKN QVSLWCLVKGFYPSDIAVEWESNGQPENNYKTPPVLDSD GSFFLYSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKS LSLSPGGGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQ LVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELV VAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAA GAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGV HLHTEARARHAWQLTQGATVLGLFRVTPEIPAGL
125	anti-FAP (4B9) light chain	see Table 21

2.2 Preparation of untargeted human 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (Control molecules)

Further control molecules were prepared as described in Example 1.4 above for Control A
5 and B. A bivalent variant Control C was prepared in analogy to the bivalent Construct 2.3 and
2.6 and a monovalent variant Control E was prepared in analogy to Construct 2.5 (containing a

4-1BB ligand (71-248) trimer), with the only difference that the anti-FAP binder (VH-VL) was replaced by a germline control, termed DP47, not binding to the antigen.

Table 27 shows the cDNA and amino acid sequences of the bivalent DP47-untargeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion molecule Control C. Table 28 shows the cDNA and amino acid sequences of the monovalent DP47-untargeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion molecule Control E.

Table 27: Sequences of bivalent DP47-untargeted human 4-1BB ligand (71-254) containing Fc (kih) fusion molecule Control C

SEQ ID NO:	Description	Sequence
177	nucleotide sequence DP47 Fc hole chain fused to dimeric hu 4-1BBL (71-254)	GAGGTGCAATTGTTGGAGTCTGGGGGAGGCTTGGTACA GCCTGGGGGGTCCCTGAGACTCTCCTGTGCAGCCTCCGG ATTCACCTTTAGCAGTTATGCCATGAGCTGGGTCCGCCA GGCTCCAGGGAAGGGGCTGGAGTGGGTCTCAGCTATTA GTGGTAGTGGTGGTAGCACATACTACGCAGACTCCGTGA AGGGCCGGTTACCATCTCCAGAGACAATTCCAAGAAC ACGCTGTATCTGCAGATGAACAGCCTGAGAGCCGAGGA CACGGCCGTATATTACTGTGCGAAAGGCAGCGGATTTGA CTACTGGGGCCAAGGAACCCTGGTCACCGTCTCGAGTGC TAGACCAAGGGCCCCCTCCGTGTTCCCCCTGGCCCCCAG CAGCAAGAGCACCAGCGGCGGCACAGCCGCTCTGGGCT GCCTGGTCAAGGACTACTTCCCCGAGCCCGTGACCGTGT CCTGGAACAGCGGAGCCCTGACCTCCGGCGTGCACACC TTCCCCGCCGTGCTGCAGAGTTCTGGCCTGTATAGCCTG AGCAGCGTGGTCACCGTGCCTTCTAGCAGCCTGGGCACC CAGACCTACATCTGCAACGTGAACCACAAGCCCAGCAA CACCAAGGTGGACAAGAAGGTGGAGCCCAAGAGCTGCG ACAAAACCTCACACATGCCCACCGTGCCCAGCACCTGAA GCTGCAGGGGGACCGTCAGTCTTCCTCTTCCCCCAAAA CCCAAGGACACCCTCATGATCTCCCGGACCCCTGAGGTG ACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGA GGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGC ATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAAC AGCACGTACCGTGTGGTCAGCGTCCTACCGTCCTGCAC CAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGT CTCCAACAAAGCCCTCGGCGCCCCATCGAGAAAACCA TCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTG TGCACCCTGCCCCCATCCCGGGATGAGCTGACCAAGAA CCAGGTCAGCCTCTCGTGCGCAGTCAAAGGCTTCTATCC CAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGC CGGAGAACAACTACAAGACCACGCCTCCCGTGCTGGAC TCCGACGGCTCCTTCTTCTCGTGAGCAAGCTCACCGTG GACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATG CTCCGTGATGCATGAGGCTCTGCACAACCACTACACGCA GAAGAGCCTCTCCCTGTCTCCGGGTGGAGGCGGCGGAA GCGGAGGAGGAGGATCCAGAGAGGGCCCTGAGCTGAGC CCCGATGATCCTGCTGGACTGCTGGACCTGCGGCAGGGC ATGTTTGCTCAGCTGGTGGCCAGAACGTGCTGCTGATC

		GATGGCCCCCTGTCCTGGTACAGCGATCCTGGACTGGCT GGCGTGTCACTGACAGGCGGCCTGAGCTACAAAGAGGA CACCAAAGAACTGGTGGTGGCCAAAGGCCGGCGTGTACT ACGTGTTCTTTCAGCTGGAAGTGCAGAGAGTGGTGGCCG GCGAAGGATCTGGCTCTGTGTCTCTGGCCCTGCATCTGC AGCCTCTGAGAAGCGCTGCTGGCGCTGCAGCTCTGGCAC TGACAGTGGATCTGCCTCCTGCCAGCTCCGAGGCCCGGA ATAGCGCATTTGGGTTTCAAGGCAGGCTGCTGCACCTGT CTGCCGGCCAGAGGCTGGGAGTGCATCTGCACACAGAG GCCAGGGCTAGACACGCCTGGCAGCTGACACAGGGCGC TACAGTGCTGGGCCTGTTCAAGAGTGACCCCGAGATTCC AGCCGGCCTGCCTTCTCCAAGAAGCGAAGGCGGAGGCG GATCTGGCGGCGGAGGATCTAGAGAGGGACCCGAAGTCTG TCCCCTGACGATCCAGCCGGGCTGCTGGATCTGAGACAG GGAATGTTTCGCCAGCTGGTGGCTCAGAATGTGCTGCTG ATTGACGGACCTCTGAGCTGGTACTCCGACCCAGGGCTG GCAGGGGTGTCCCTGACTGGGGGACTGTCCTACAAAGA AGATACAAAAGAACTGGTGGTGGCTAAAGCTGGGGTGT ACTATGTGTTTTTTCAGCTGGAAGTGGGCGGGTGGTGG CTGGGGAGGGCTCAGGATCTGTGTCCCTGGCTCTGCATC TGCAGCCACTGCGCTCTGCTGCTGGCGCAGCTGCACTGG CTCTGACTGTGGACCTGCCACCAGCCTCTAGCGAGGCCA GAAACAGCGCCTTTCGGGTTCCAAGGACGCCTGCTGCATC TGAGCGCCGGACAGCGCCTGGGAGTGCATCTGCATACT GAAGCCAGAGCCCGGCATGCTTGGCAGCTGACTCAGGG GGCAACTGTGCTGGGACTGTTTCGCGTGACACCTGAGAT CCCTGCCGGACTGCCAAGCCCTAGATCAGAA
178	nucleotide sequence DP47 Fc knob chain fused to monomeric hu 4- 1BBL (71-254)	GAGGTGCAATTGTTGGAGTCTGGGGGAGGCTTGGTACA GCCTGGGGGGTCCCTGAGACTCTCCTGTGCAGCCTCCGG ATTCACCTTTAGCAGTTATGCCATGAGCTGGGTCCGCCA GGCTCCAGGGAAGGGGCTGGAGTGGGTCTCAGCTATTA GTGGTAGTGGTGGTAGCACATACTACGCAGACTCCGTGA AGGGCCGGTTCACCATCTCCAGAGACAATTCCAAGAAC ACGCTGTATCTGCAGATGAACAGCCTGAGAGCCGAGGA CACGGCCGTATATTACTGTGCGAAAGGCAGCGGATTTGA CTACTGGGGCCAAGGAACCCTGGTCACCGTCTCGAGTGC TAGCACCAAGGGCCCATCGGTCTTCCCCCTGGCACCCCTC CTCCAAGAGCACCTCTGGGGGACAGCGGCCCTGGGCT GCCTGGTCAAGGACTACTTCCCCGAACCGGTGACGGTGT CGTGGAACCTCAGGCGCCCTGACCAGCGGCGTGACACACC TTCCCGGCTGTCTACAGTCCTCAGGACTCTACTCCCTC AGCAGCGTGGTGACCGTGCCCTCCAGCAGCTTGGGCAC CCAGACCTACATCTGCAACGTGAATCACAAGCCAGCA ACACCAAGGTGGACAAGAAAGTTGAGCCCAAATCTTGT GACAAAACCTCACACATGCCCACCGTGCCAGCACCTGA AGCTGCAGGGGGACCGTCAGTCTTCTCTTCCCCCAAA ACCCAAGGACACCCTCATGATCTCCCGGACCCCTGAGGT CACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTG AGGTCAAGTTCAACTGGTACGTGGACGGCGTGAGGTG CATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAA CAGCACGTACCGTGTGGTCAGCGTCCTCACCCTCCTGCA CCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGG TCTCCAACAAAGCCCTCGGCGCCCCCATCGAGAAAACC ATCTCCAAGCCAAAGGGCAGCCCCGAGAACCACAGGT

		GTACACCCTGCCCCCTGCAGAGATGAGCTGACCAAGA ACCAGGTGTCCCTGTGGTGTCTGGTCAAGGGCTTCTACC CCAGCGATATCGCCGTGGAGTGGGAGAGCAACGGCCAG CCTGAGAACAACACTACAAGACCACCCCCCTGTGCTGGA CAGCGACGGCAGCTTCTTCTGTACTCCAAACTGACCGT GGACAAGAGCCGGTGGCAGCAGGGCAACGTGTTTCAGCT GCAGCGTGATGCACGAGGCCCTGCACAACCACTACACC CAGAAGTCCCTGAGCCTGAGCCCCGGCGGAGGCGGCGG AAGCGGAGGAGGAGGATCCAGAGAGGGCCCTGAGCTGA GCCCCGATGATCCTGCTGGACTGCTGGACCTGCGGCAGG GCATGTTTGCTCAGCTGGTGGCCCAGAACGTGCTGCTGA TCGATGGCCCCCTGTCTGGTACAGCGATCCTGGACTGG CTGGCGTGTCACTGACAGGCGGCCTGAGCTACAAAGAG GACACCAAAGAACTGGTGGTGGCCAAGGCCGGCGTGTA CTACGTGTTCTTTCAGCTGGAAGTGCAGGAGAGTGGTGGC CGGCGAAGGATCTGGCTCTGTGTCTCTGGCCCTGCATCT GCAGCCTCTGAGAAGCGCTGCTGGCGCTGCAGCTCTGGC ACTGACAGTGGATCTGCCTCCTGCCAGCTCCGAGGCCCG GAATAGCGCATTTGGGTTTCAAGGCAGGCTGCTGCACCT GTCTGCCGGCCAGAGGCTGGGAGTGCATCTGCACACAG AGGCCAGGGCTAGACACGCCTGGCAGCTGACACAGGGC GCTACAGTGCTGGGCCTGTTTCAGAGTGACCCCCGAGATT CCAGCCGGCCTGCCTTCTCCAAGAAGCGAA
80	nucleotide sequence DP47 light chain	see Table 18
179	DP47 Fc hole chain fused to dimeric hu 4- 1BBL (71-254)	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSYAMSWVRQA PGKGLEWVSAIIGSGASTYYADSVKGRFTISRDN SKNTLYL QMNSLRAEDTAVYYCAKGWFGGFNYWGQGLVTVSSAS TKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNS GALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICN VNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVF LFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDG VEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYK CKVSNKALGAPIEKTISKAKGQPREPQVCTLPSPRDELTKN QVSLSCAVKGFYPSDIAVEWESNGQPENNYKTPPVLDSD GSFFLVSKLTVDKSRWQQGNV FSCVMHEALHNHYTQKS LSLSPGGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQ LVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELV VAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAA GAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGV HLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSEG GGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQLVAQNV LLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELV VAKAGV YYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALA LTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEA RARHAWQLTQGATVLGLFRVTPEIPAGLPSRSE
180	DP47 Fc knob chain fused to monomeric hu 4- 1BBL (71-254)	EVQLLESGGGLVQPGGSLRLSCAASGFTFSSYAMSWVRQA PGKGLEWVSAIIGSGASTYYADSVKGRFTISRDN SKNTLYL QMNSLRAEDTAVYYCAKGWFGGFNYWGQGLVTVSSAS TKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNS GALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICN VNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVF

		LFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDG VEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYK CKVSNKALGAPIEKTISKAKGQPREPQVYTLPPCRDELTKN QVSLWCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD GSFFLYSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKS LSLSPGGGGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQ LVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELV VAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAA GAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGV HLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSE
82	DP47 light chain	see Table 18

Table 28: Sequences of monovalent untargeted human 4-1BB ligand (71-248) containing Fc (kih) fusion molecule Control E

SEQ ID NO:	Description	Sequence
171	Dimeric hu 4-1BBL (71-248) – CL Fc knob chain	see Table 25
172	Monomeric hu 4-1BBL (71-248) – CH1	see Table 25
79	DP47 Fc hole chain	see Table 18
80	DP47 light chain	see Table 18
173	Dimeric hu 4-1BBL (71-248) – CL Fc knob chain	see Table 25
174	Monomeric hu 4-1BBL (71-248) – CH1	see Table 25
81	DP47 Fc hole chain	see Table 18
82	DP47 light chain	see Table 18

5 2.3 Preparation of untargeted human IgG1 as Control F

An additional control molecule used in the assays was an untargeted DP47, germline control, human IgG1, containing the Pro329Gly, Leu234Ala and Leu235Ala mutations, to abrogate binding to Fc gamma receptors according to the method described in International Patent Appl. Publ. No. WO 2012/130831).

Table 29 shows the cDNA and amino acid sequences of the cDNA and amino acid sequences of the untargeted DP47 huIgG1 PGLALA (Control F).

Table 29: Sequences of untargeted DP47 huIgG1 (Control F)

SEQ ID NO:	Description	Sequence
181	nucleotide sequence DP47 heavy chain (hu IgG1 PGLALA)	GAGGTGCAATTGTTGGAGTCTGGGGGAGGCTTGGTACA GCCTGGGGGGTCCCTGAGACTCTCCTGTGCAGCCTCCGG ATTCACCTTTAGCAGTTATGCCATGAGCTGGGTCCGCCA GGCTCCAGGGAAGGGGCTGGAGTGGGTCTCAGCTATTA GTGGTAGTGGTGGTAGCACATACTACGCAGACTCCGTGA AGGGCCGGTTCACCATCTCCAGAGACAATTCCAAGAAC ACGCTGTATCTGCAGATGAACAGCCTGAGAGCCGAGGA CACGGCCGTATATTACTGTGCGAAAGGCAGCGGATTTGA CTACTGGGGCCAAGGAACCCTGGTCACCGTCTCGAGTGC TAGACCAAGGGCCCATCGGTCTTCCCCCTGGCACCCCTC CTCCAAGAGCACCTCTGGGGGCACAGCGGCCCTGGGCT GCCTGGTCAAGGACTACTTCCCCGAACCGGTGACGGTGT CGTGGAACCTCAGGCGCCCTGACCAGCGGCGTGCACACC TCCCCGGCTGTCCTACAGTCCTCAGGACTCTACTCCCTC AGCAGCGTGGTGACCGTGCCCTCCAGCAGCTTGGGCAC CCAGACCTACATCTGCAACGTGAATCACAAGCCCAGCA ACACCAAGGTGGACAAGAAAGTTGAGCCCAAATCTTGT GACAAAACCTCACACATGCCCACCGTGCCCGAGCACCTGA AGCTGCAGGGGGACCGTCAGTCTTCCTCTTCCCCCAAAA ACCCAAGGACACCCTCATGATCTCCCGGACCCCTGAGGT CACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTG AGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTG CATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAA CAGCACGTACCGTGTGGTCAGCGTCCTACCGTCCTGCA CCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGG TCTCCAACAAAGCCCTCGGCGCCCCCATCGAGAAAACC ATCTCCAAGCCAAAGGGCAGCCCCGAGAACCACAGGT GTACACCCTGCCCCCATCCCGGGATGAGCTGACCAAGA ACCAGGTCAGCCTGACCTGCCTGGTCAAAGGCTTCTATC CCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAG CCGGAGAACAACCTACAAGACCACGCCTCCCGTGCTGGA CTCCGACGGCTCCTTCTTCTCTACAGCAAGCTCACCGT GGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCAT GCTCCGTGATGCATGAGGCTCTGCACAACCACTACACGC AGAAGAGCCTCTCCCTGTCTCCGGGTAAA
80	DP47 light chain	see Table 18
182	DP47 heavy chain (hu IgG1 PGLALA)	EVQLLESGGGLVQPGGSLRLSCAASGFTSSYAMSWVRQA PGKGLEWVSAISGSGGSTYYADSVKGRFTISRDN SKNTLYL QMNSLR AEDTAVYYCAKGS GFDYWGQGT LVTVSSASTKG PSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVT VSWNSGAL TSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNH KPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPP

		KPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEV HNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKV SNKALGAPIEKTISKAKGQPREPQVYTLPPSRDELTKNQVS LTCLVKGFYPSDIAVEWESNGQPENNYKTTPVLDSGDSFF LYSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLS PGK
82	DP47 light chain	see Table 18

2.4 Production of monovalent and bivalent FAP (4B9) targeted split trimeric 4-1BB ligand Fc fusion constructs and control molecules

The targeted and untargeted split trimeric 4-1BB ligand Fc (kih) fusion encoding
 5 sequences were cloned into a plasmid vector, which drives expression of the insert from an MPSV promoter and contains a synthetic polyA sequence located at the 3' end of the CDS. In addition, the vector contains an EBV OriP sequence for episomal maintenance of the plasmid.

The split trimeric 4-1BB ligand Fc (kih) fusion was produced by co-transfecting HEK293-
 EBNA cells with the mammalian expression vectors using polyethylenimine. The cells were
 10 transfected with the corresponding expression vectors. For Constructs 2.1, 2.2., 2.4 and 2.5 and corresponding control molecules, a 1:1:1:1 ratio (e.g. "vector dimeric ligand-CL- knob chain": "vector monomeric ligand fusion-CH1": "vector anti-FAP Fab-hole chain": "vector anti-FAP light chain") was used. For Constructs 2.3 and 2.6 and its control molecule, a 1:1:1 ratio ("vector huIgG1 Fc hole dimeric ligand chain": "vector huIgG1 Fc knob monomeric ligand chain":
 15 "vector anti-FAP light chain") was taken. Human IgGs, used as control in the assay, were produced as for the bispecific constructs (for transfection only a vector for light and a vector for heavy chain were used).

For production in 500 mL shake flasks, 300 million HEK293 EBNA cells were seeded 24
 hours before transfection. For transfection cells were centrifuged for 10 minutes at 210 x g, and
 20 the supernatant was replaced by 20mL pre-warmed CD CHO medium. Expression vectors (200 µg of total DNA) were mixed in 20 mL CD CHO medium. After addition of 540 µL PEI, the solution was vortexed for 15 seconds and incubated for 10 minutes at room temperature. Afterwards, cells were mixed with the DNA/PEI solution, transferred to a 500 mL shake flask and incubated for 3 hours at 37 °C in an incubator with a 5% CO₂ atmosphere. After the
 25 incubation, 160 mL of Excell medium supplemented with 6mM L-Glutamine, 5g/L PEPSOY and 1.2mM valproic acid was added and cells were cultured for 24 hours. One day after transfection 12% Feed 7 and Glucose (final conc. 3g/L) were added. After culturing for 7 days, the supernatant was collected by centrifugation for 30-40 minutes at least 400 x g. The solution

was sterile filtered (0.22 μ m filter), supplemented with sodium azide to a final concentration of 0.01 % (w/v), and kept at 4 °C.

The targeted and untargeted TNF ligand trimer-containing Fc (kih) fusion antigen binding molecules and the human IgG1 were purified from cell culture supernatants by affinity chromatography using Protein A, followed by size exclusion chromatography. For affinity chromatography, the supernatant was loaded on a MabSelect Sure column (CV = 5-15 mL, resin from GE Healthcare) equilibrated with Sodium Phosphate (20 mM), Sodium Citrate (20 mM) buffer (pH 7.5). Unbound protein was removed by washing with at least 6 column volumes of the same buffer. The bound protein was eluted using either a linear gradient (20 CV) or a step elution (8 CV) with 20 mM sodium citrate, 100 mM Sodium chloride, 100 mM Glycine buffer (pH 3.0). For the linear gradient an additional 4 column volumes step elution was applied.

The pH of collected fractions was adjusted by adding 1/10 (v/v) of 0.5M sodium phosphate, pH8.0. The protein was concentrated prior to loading on a HiLoad Superdex 200 column (GE Healthcare) equilibrated with 20 mM Histidine, 140 mM sodium chloride, 0.01% (v/v) Tween20 solution of pH 6.0.

The protein concentration was determined by measuring the optical density (OD) at 280 nm, using a molar extinction coefficient calculated on the basis of the amino acid sequence. Purity and molecular weight of the targeted trimeric 4-1BB ligand Fc (kih) fusion was analyzed by SDS-PAGE in the presence and absence of a reducing agent (5 mM 1,4-dithiotreitol) and staining with Coomassie SimpleBlue™ SafeStain (Invitrogen USA) or CE-SDS using Caliper LabChip GXII (Perkin Elmer). The aggregate content of samples was analyzed using a TSKgel G3000 SW XL analytical size-exclusion column (Tosoh) equilibrated in 25 mM K₂HPO₄, 125 mM NaCl, 200 mM L-Arginine Monohydrochloride, 0.02 % (w/v) NaN₃, pH 6.7 running buffer at 25 °C.

Table 30 summarizes the yield and final monomer content of the FAP (4B9) targeted and untargeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules and control molecules.

Table 30 Biochemical analysis of FAP (4B9) targeted and untargeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules and control molecules

Construct	Monomer [%] (SEC)	Yield [mg/l]
Construct 2.1	95	15.8

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Construct 2.3	97	11.5
Construct 2.4	97	14.1
Construct 2.5	100	16.5
Control C (bivalent)	98	12.6
Control E (monovalent)	93	4.1
Control F (germline DP47 human IgG1 PGLALA)	100	50

Example 3

Preparation, purification and characterization of 4-1BB

DNA sequences encoding the ectodomains of human, mouse or cynomolgus 4-1BB (Table 31) were subcloned in frame with the human IgG1 heavy chain CH2 and CH3 domains on the knob (Merchant et al., 1998). An AcTEV protease cleavage site was introduced between an antigen ectodomain and the Fc of human IgG1. An Avi tag for directed biotinylation was introduced at the C-terminus of the antigen-Fc knob. Combination of the antigen-Fc knob chain containing the S354C/T366W mutations, with a Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations allows generation of a heterodimer which includes a single copy of 4-1BB ectodomain containing chain, thus creating a monomeric form of Fc-linked antigen (Figure 5C). Table 32 shows the cDNA and amino acid sequences of the antigen Fc-fusion constructs.

Table 31: Amino acid numbering of antigen ectodomains (ECD) and their origin

SEQ ID NO:	Construct	Origin	ECD
83	human 4-1BB ECD	Synthetized according to Q07011	aa 24-186
84	cynomolgus 4-1BB ECD	isolated from cynomolgus blood	aa 24-186
85	murine 4-1BB ECD	Synthetized according to P20334	aa 24-187

Table 32: cDNA and Amino acid sequences of monomeric antigen Fc(kih) fusion molecules

SEQ ID NO:	Antigen	Sequence
86	Nucleotide sequence	GACAAAACTCACACATGCCCACCGTGCCCAGCACCTGA ACTCCTGGGGGGACCGTCAGTCTTCCTCTTCCCCCAAA ACCCAAGGACACCCTCATGATCTCCCGGACCCCTGAGGT CACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTG

	Fc hole chain	AGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTG CATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAA CAGCACGTACCGTGTGGTCAGCGTCCTCACCGTCCTGCA CCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGG TCTCCAACAAAGCCCCTCCCAGCCCCCATCGAGAAAACC ATCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGT GTGCACCCTGCCCCCATCCCGGGATGAGCTGACCAAGA ACCAGGTCAGCCTCTCGTGCGCAGTCAAAGGCTTCTATC CCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAG CCGGAGAACAACACTACAAGACCACGCCTCCCGTGCTGGA CTCCGACGGCTCCTTCTTCTCGTGAGCAAGCTCACCGT GGACAAGAGCAGGTGGCAGCAGGGGAACGCTTCTCAT GCTCCGTGATGCATGAGGCTCTGCACAACCACTACACGC AGAAGAGCCTCTCCCTGTCTCCGGGTAAA
87	Nucleotide sequence human 4-1BB antigen Fc knob chain	CTGCAGGACCCCTGCAGCAACTGCCCTGCCGGCACCTTC TGCGACAACAACCGGAACCAGATCTGCAGCCCCTGCCC CCCCAACAGCTTCAGCTCTGCCGGCGGACAGCGGACCT GCGACATCTGCAGACAGTGCAAGGGCGTGTTCAGAACC CGGAAAGAGTGCAGCAGCACCAGCAACGCCGAGTGCGA CTGCACCCCCGGCTTCCATTGTCTGGGAGCCGGCTGCAG CATGTGCGAGCAGGACTGCAAGCAGGGCCAGGAACTGA CCAAGAAGGGCTGCAAGGACTGCTGCTTCGGGCACCTTC AACGACCAGAAGCGGGGCATCTGCCGGCCCTGGACCAA CTGTAGCCTGGACGGCAAGAGCGTGCTGGTCAACGGCA CCAAAGAACGGGACGTCTGTGTGCGGCCCCAGCCCTGCT GATCTGTCTCCTGGGGCCAGCAGCGTGACCCCTCCTGCC CCTGCCAGAGAGCCTGGCCACTCTCCTCAGGTCGACGAA CAGTTATATTTTCAGGGCGGCTCACCCAAATCTGCAGAC AAAACCTCACACATGCCACCGTGCCAGCACCTGAACTC CTGGGGGGGACCGTCAGTCTTCTCTTCCCCCAAAACCC AAGGACACCCTCATGATCTCCCGGACCCCTGAGGTCACA TGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGGT CAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATA ATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGC ACGTACCGTGTGGTCAGCGTCCTCACCGTCCTGCACCAG GACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTC CAACAAAGCCCTCCCAGCCCCCATCGAGAAAACCATCT CCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTGTAC ACCCTGCCCCCATGCCGGGATGAGCTGACCAAGAACCA GGTCAGCCTGTGGTGCCTGGTCAAAGGCTTCTATCCCAG CGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGG AGAACAACACTACAAGACCACGCCTCCCGTGCTGGACTCC GACGGCTCCTTCTTCTCTACAGCAAGCTCACCGTGGAC AAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTC CGTGATGCATGAGGCTCTGCACAACCACTACACGCAGA AGAGCCTCTCCCTGTCTCCGGGTAAATCCGGAGGCCTGA ACGACATCTTCGAGGCCCAGAAGATTGAATGGCACGAG
88	Nucleotide sequence cynomolgus 4- 1BB antigen	TTGCAGGATCTGTGTAGTAACTGCCAGCTGGTACATTC TGTGATAATAACAGGAGTCAGATTTGCAGTCCCTGTCTT CCAAATAGTTTCTCCAGCGCAGGTGGACAAAGGACCTGT GACATATGCAGGCAGTGTAAGGTGTTTTCAAGACCAG GAAGGAGTGTTCTTCCACCAGCAATGCAGAGTGTGACT GCATTTCAAGGTATCACTGCCTGGGGGCAGAGTGCAGC ATGTGTGAACAGGATTGTAAACAAGGTCAAGAATTGAC

	Fc knob chain	AAAAAAAGGTTGTAAAGACTGTTGCTTTGGGACATTTAA TGACCAGAAACGTGGCATCTGTCGCCCCCTGGACAAACT GTTCTTTGGATGGAAAGTCTGTGCTTGTGAATGGGACGA AGGAGAGGGACGTGGTCTGCGGACCATCTCCAGCCGAC CTCTCTCCAGGAGCATCCTCTGCGACCCCGCCTGCCCT GCGAGAGAGCCAGGACACTCTCCGCAGGTCGACGAACA GTTATATTTTCAGGGCGGCTCACCCAAATCTGCAGACAA AACTCACACATGCCCACCGTGCCCAGCACCTGAACTCCT GGGGGGACCGTCAGTCTTCCTCTTCCCCCAAAACCCAA GGACACCTCATGATCTCCCGGACCCCTGAGGTCACATG CGTGGTGGTGGACGTGAGCCACGAAGACCTGAGGTCA AGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAAT GCCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGCA CGTACCGTGTGGTCAGCGTCTCACCCTGCTGCACCAGG ACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCC AACAAAGCCCTCCCAGCCCCATCGAGAAAACCATCTC CAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTGTACA CCCTGCCCCCATGCCGGGATGAGCTGACCAAGAACCAG GTCAGCCTGTGGTGCCTGGTCAAAGGCTTCTATCCCAGC GACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGA GAACAATAACAAGACCACGCCTCCCGTGCTGGACTCCG ACGGCTCCTTCTTCTCTACAGCAAGCTCACCGTGGACA AGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCC GTGATGCATGAGGCTCTGCACAACCACTACACGCAGAA GAGCCTCTCCCTGTCTCCGGGTAAATCCGGAGGCCTGAA CGACATCTTCGAGGCCCAAGAAGATTGAATGGCACGAG
89	murine 4-1BB antigen Fc knob chain	GTGCAGAACAGCTGCGACAACCTGCCAGCCCGGCACCTT CTGCCGGAAGTACAACCCCGTGTGCAAGAGCTGCCCCC CCAGCACCTTCAGCAGCATCGGCGGCCAGCCCAACTGC AACATCTGCAGAGTGTGCGCCGGCTACTTCCGGTTCAAG AAGTTCTGCAGCAGCACCCACAACGCCGAGTGCAGAGTG CATCGAGGGCTTCCACTGCCTGGGCCCCCAGTGCACCAG ATGCGAGAAGGACTGCAGACCCGGCCAGGAAGTACCA AGCAGGGCTGTAAGACCTGCAGCCTGGGCACCTTCAAC GACCAGAACGGGACCGGCGTGTGCCGGCCTTGGACCAA TTGCAGCCTGGACGGGAGAAGCGTGCTGAAAACCGGCA CCACCGAGAAGGACGTGCTGTGCGGCCCTCCCGTGGTGT CCTTCAGCCCTAGCACCAACCATCAGCGTGACCCCTGAAG GCGGCCCTGGCGGACACTCTCTGCAGGTCTTGGTCGACG AACAGTTATATTTTCAGGGCGGCTCACCCAAATCTGCAG AAAAACTCACACATGCCCACCGTGCCCAGCACCTGAA CTCCTGGGGGGACCGTCAGTCTTCCTCTTCCCCCAAAA CCCAAGGACACCCTCATGATCTCCCGGACCCCTGAGGTC ACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGA GGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGC ATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAAC AGCACGTACCGTGTGGTCAGCGTCCTCACCGTCCTGCAC CAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGT CTCCAACAAAGCCCTCCCAGCCCCATCGAGAAAACCA TCTCCAAGGCCAAAGGGCAGCCCCGAGAACCACAGGTG TACACCCTGCCCCCATGCCGGGATGAGCTGACCAAGAA CCAGGTCAGCCTGTGGTGCCTGGTCAAAGGCTTCTATCC CAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGC CGGAGAACAATAACAAGACCACGCCTCCCGTGCTGGAC

		TCCGACGGCTCCTTCTTCTCTACAGCAAGCTCACCGTG GACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATG CTCCGTGATGCATGAGGCTCTGCACAACCACTACACGCA GAAGAGCCTCTCCCTGTCTCCGGGTAATCCGGAGGCCT GAACGACATCTTCGAGGCCAGAAAGATTGAATGGCACG AG
90	Fc hole chain	DKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCV VVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYR VVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKG QPREPQVCTLPPSRDELTKNQVSLSCAVKGFYPSDIAVEWE SNGQPENNYKTTTPVLDSDGSFFLVSKLTVDKSRWQQGNV FSCSVMEALHNHYTQKSLSLSPGK
91	human 4-1BB antigen Fc knob chain	LQDPCSNCPAGTFCDDNRRNQICSPCPPNSFSSAGGQRTCDI CRQCKGVFRTRKECSSTSNAECDCTPGFHCLGAGCSMCEQ DCKQGQELTKKGCKDCCFGTFNDQKRGICRPWTNCSLDG KSVLVNGTKERDVVCGPSPADLSPGASSVTPPAPAREPGHS PQVDEQLYFQGGSPKSADKTHTCPPCPAPELLGGPSVFLFP PKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVE VHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCK VSNKALPAPIEKTISKAKGQPREPQVYTLPPCRDELTKNQV SLWCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGS FFLYSKLTVDKSRWQQGNVFSCSVMEALHNHYTQKSLS LSPGKSGGLNDIFEAQKIEWHE
92	cynomolgus 4- 1BB antigen Fc knob chain	LQDLCSNCPAGTFCDDNRRSQICSPCPPNSFSSAGGQRTCDIC RQCKGVFKTRKECSSTSNAECDICISGYHCLGAECMSMCEQD CKQGQELTKKGCKDCCFGTFNDQKRGICRPWTNCSLDGK SVLVNGTKERDVVCGPSPADLSPGASSATPPAPAREPGHSP QVDEQLYFQGGSPKSADKTHTCPPCPAPELLGGPSVFLFPP KPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEV HNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKV SNKALPAPIEKTISKAKGQPREPQVYTLPPCRDELTKNQVS LWCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSF FLYSKLTVDKSRWQQGNVFSCSVMEALHNHYTQKSLSL SPGKSGGLNDIFEAQKIEWHE
93	murine 4-1BB antigen Fc knob chain	VQNSCDNCQPGTFCRKYNPVCKSCPPSTFSSIGGQPNCNIC RVCAGYFRFKFCSSSTHNAECECIEGFHCLGPQCTRCEKDC RPGQELTKQGCKTCSLGTENDQNGTGVCRPWTNCSLDGR SVLKTGTTEKDVVCGPPVVSFSPSTTISVTPEGGPGGHS VLVDEQLYFQGGSPKSADKTHTCPPCPAPELLGGPSVFLFP PKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVE VHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCK VSNKALPAPIEKTISKAKGQPREPQVYTLPPCRDELTKNQV SLWCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGS FFLYSKLTVDKSRWQQGNVFSCSVMEALHNHYTQKSLS LSPGKSGGLNDIFEAQKIEWHE

All 4-1BB-Fc-fusion molecule encoding sequences were cloned into a plasmid vector, which drives expression of the insert from an MPSV promoter and contains a synthetic polyA signal sequence located at the 3' end of the CDS. In addition, the vector contains an EBV OriP
5 sequence for episomal maintenance of the plasmid.

For preparation of the biotinylated monomeric antigen/Fc fusion molecules, exponentially growing suspension HEK293 EBNA cells were co-transfected with three vectors encoding the two components of fusion protein (knob and hole chains) as well as BirA, an enzyme necessary for the biotinylation reaction. The corresponding vectors were used at a 2 : 1 : 0.05 ratio

5 (“antigen ECD-AcTEV- Fc knob” : “Fc hole” : “BirA”).

For protein production in 500 ml shake flasks, 400 million HEK293 EBNA cells were seeded 24 hours before transfection. For transfection cells were centrifuged for 5 minutes at 210 g, and the supernatant was replaced by pre-warmed CD CHO medium. Expression vectors were resuspended in 20 mL of CD CHO medium containing 200 µg of vector DNA. After addition of
10 540 µL of polyethylenimine (PEI), the solution was vortexed for 15 seconds and incubated for 10 minutes at room temperature. Afterwards, cells were mixed with the DNA/PEI solution, transferred to a 500 mL shake flask and incubated for 3 hours at 37°C in an incubator with a 5 % CO₂ atmosphere. After the incubation, 160 mL of F17 medium was added and cells were cultured for 24 hours. The production medium was supplemented with 5µM kifunensine. One
15 day after transfection, 1 mM valproic acid and 7 % Feed 1 with supplements were added to the culture. After 7 days of culturing, the cell supernatant was collected by spinning down cells for 15 min at 210 g. The solution was sterile filtered (0.22 µm filter), supplemented with sodium azide to a final concentration of 0.01 % (w/v), and kept at 4°C.

Secreted proteins were purified from cell culture supernatants by affinity chromatography
20 using Protein A, followed by size exclusion chromatography. For affinity chromatography, the supernatant was loaded on a HiTrap ProteinA HP column (CV = 5 mL, GE Healthcare) equilibrated with 40 mL 20 mM sodium phosphate, 20 mM sodium citrate pH 7.5. Unbound protein was removed by washing with at least 10 column volumes of 20 mM sodium phosphate, 20 mM sodium citrate, 0.5 M sodium chloride containing buffer (pH 7.5). The bound protein was
25 eluted using a linear pH-gradient of sodium chloride (from 0 to 500 mM) created over 20 column volumes of 20 mM sodium citrate, 0.01 % (v/v) Tween-20, pH 3.0. The column was then washed with 10 column volumes of 20 mM sodium citrate, 500 mM sodium chloride, 0.01 % (v/v) Tween-20, pH 3.0.

The pH of collected fractions was adjusted by adding 1/40 (v/v) of 2M Tris, pH8.0. The
30 protein was concentrated and filtered prior to loading on a HiLoad Superdex 200 column (GE Healthcare) equilibrated with 2mM MOPS, 150 mM sodium chloride, 0.02 % (w/v) sodium azide solution of pH 7.4.

For affinity determination to the human receptor, the ectodomain of human 4-1BB was also subcloned in frame with an avi (GLNDIFEAQKIEWHE) and a hexahistidine tag.

Protein production was performed as described above for the Fc-fusion protein. Secreted proteins were purified from cell culture supernatants by chelating chromatography, followed by size exclusion chromatography. The first chromatographic step was performed on a NiNTA Superflow Cartridge (5ml, Qiagen) equilibrated in 20 mM sodium phosphate, 500 nM sodium chloride, pH7.4. Elution was performed by applying a gradient over 12 column volume from 5 % to 45 % of elution buffer (20 mM sodium phosphate, 500 nM sodium chloride, 500 mM Imidazole, pH7.4). The protein was concentrated and filtered prior to loading on a HiLoad Superdex 75 column (GE Healthcare) equilibrated with 2 mM MOPS, 150 mM sodium chloride, 0.02 % (w/v) sodium azide solution of pH 7.4 (Table 33).

Table 33: Sequences of monomeric human 4-1BB His molecule

SEQ ID NO:	antigen	Sequence
94	nucleotide sequence human 4-1BB His	CTGCAGGACCCCTGCAGCAACTGCCCTGCCGGCACCTTCTG CGACAACAACCGGAACCAGATCTGCAGCCCCCTGCCCCCCC AACAGCTTCAGCTCTGCCGGCGGACAGCGGACCTGCGACA TCTGCAGACAGTGCAAGGGCGTGTTTACAACCCGGAAAGA GTGCAGCAGCACCAGCAACGCCGAGTGCGACTGCACCCCC GGCTTCCATTGTCTGGGAGCCGGCTGCAGCATGTGCGAGC AGGACTGCAAGCAGGGCCAGGAACTGACCAAGAAGGGCT GCAAGGACTGCTGCTTCGGCACCTTCAACGACCAGAAGCG GGGCATCTGCCGGCCCTGGACCAACTGTAGCCTGGACGGC AAGAGCGTGCTGGTCAACGGCACCAAAGAACGGGACGTCG TGTGCGGCCCCAGCCCTGCTGATCTGTCTCCTGGGGCCAGC AGCGTGACCCCTCCTGCCCTGCCAGAGAGCCTGGCCACTC TCCTCAGGTCGACGAACAGTTATATTTTCAGGGCGGCTCAG GCCTGAACGACATCTTCGAGGCCCAGAAGATCGAGTGGCA CGAGGCTCGAGCTCACCACCATCACCATCAC
95	human 4-1BB His	LQDPCSNCPAGTFCDNNRNQICSPCPPNSFSSAGGQRTCDICR QCKGVFRTRKECSSTSNAECDCTPGFHCLGAGCSMCEQDCK QGQELTKKGCKDCCFGTFNDQKRGICRPWTNCSLDGKSVLV NGTKERDVVCGPSPADLSPGASSVTPPAPAREPGHSPQVDEQL YFQGGSGLNDIFEAQKIEWHEARAHHHHHH

Example 4

Biochemical characterization of FAP-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecule by surface plasmon resonance

The binding of FAP-targeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules to recombinant 4-1BB was assessed by surface plasmon resonance (SPR). All SPR experiments were performed on a Biacore T100 at 25 °C with HBS-EP as a running buffer (0.01 M HEPES pH 7.4, 0.15 M NaCl, 3 mM EDTA, 0.005 % Surfactant P20, Biacore, Freiburg/Germany).

The avidity of the interaction between the FAP-targeted or untargeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules and recombinant 4-1BB (human, cyno and murine) was determined as described below. The data demonstrated that both targeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules as well as untargeted DP47 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules bind with comparable avidities to human and cynomolgus 4-1BB but negligibly to the mouse homolog.

Recombinant biotinylated human, cynomolgus and murine 4-1BB Fc(kih) fusion molecules were directly coupled on a SA chip using the standard coupling instruction (Biacore, Freiburg/Germany). The immobilization level was about 30 RU. FAP-targeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules, or the DP47 untargeted controls, were passed at a concentration range from 0.39 to 200 nM with a flow of 30 μ L/minutes through the flow cells over 120 seconds. The dissociation was monitored for 180 seconds. Bulk refractive index differences were corrected for by subtracting the response obtained on a reference empty flow cell.

For affinity measurement, direct coupling of around 7200 resonance units (RU) of an anti-human Fc specific antibody was performed on a CM5 chip at pH 5.0 using the standard amine coupling kit (GE Healthcare). FAP-targeted or untargeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules, at 50 nM were captured with a flow rate of 30 μ L/min for 60 sec on flow cell 2. A dilution series (1.95 to 1000 nM) of human 4-1BB-avi-His was passed on both flow cells at 30 μ L/min for 180 sec to record the association phase. The dissociation phase was monitored for 180 s and triggered by switching from the sample solution to HBS-EP. The chip surface was regenerated after every cycle using a double injection of 60 sec 10 mM Glycine-HCl pH 2.1. Bulk refractive index differences were corrected for by subtracting the response obtained on reference flow cell 1. For the interaction between the 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules and hu4-1BB avi His, the affinity constants were derived from the rate constants by fitting to a 1:1 Langmuir binding curve using the Biaeval software (GE Healthcare).

Table 34: Fittings to 1:1 Langmuir binding and Affinity constants

Ligand	Analyte	ka (1/Ms)	kd (1/s)	KD (M)
FAP split 4-1BBL trimer (Construct 1.1)	hu4-1BB	4.8E+04	2.6E-02	5.5E-07
DP47 split 4-1BBL trimer (Control A)	hu4-1BB	6.2E+04	3.3E-02	5.2E-07

Example 5**Functional characterization of the targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules****5.1. Binding on naïve versus activated human PMBCs of the FAP-targeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules**

Buffy coats were obtained from the Zürich blood donation center. To isolate fresh peripheral blood mononuclear cells (PBMCs) the buffy coat was diluted with the same volume of DPBS (Gibco by Life Technologies, Cat. No. 14190 326). 50 mL polypropylene centrifuge tubes (TPP, Cat.-No. 91050) were supplied with 15 mL Histopaque 1077 (SIGMA Life Science, Cat.-No. 10771, polysucrose and sodium diatrizoate, adjusted to a density of 1.077 g/mL) and the diluted buffy coat solution was layered above the Histopaque 1077. The tubes were centrifuged for 30 min at 400 x g. PBMCs were then collected from the interface, washed three times with DPBS and resuspended in T cell medium consisting of RPMI 1640 medium (Gibco by Life Technology, Cat. No. 42401-042) supplied with 10 % Fetal Bovine Serum (FBS, Gibco by Life Technology, Cat. No. 16000-044, Lot 941273, gamma-irradiated, mycoplasma-free and heat inactivated at 56 °C for 35 min), 1 % (v/v) GlutaMAX-I (GIBCO by Life Technologies, Cat. No. 35050 038), 1 mM Sodium Pyruvate (SIGMA, Cat. No. S8636), 1 % (v/v) MEM non-essential amino acids (SIGMA, Cat.-No. M7145) and 50 µM β-Mercaptoethanol (SIGMA, M3148).

PBMCs were used directly after isolation or stimulated to induce 4-1BB expression at the cell surface of T and NK cells by culturing for 4 days in T cell medium supplemented with 200 U/mL Proleukin (Novartis Pharma Schweiz AG, CHCLB-P-476-700-10340) and 2 µg/mL PHA-L (SIGMA Cat.-No. L2769) in a 6-well tissue culture plate and then 1 day in a 6-well tissue culture plate coated with 10 µg/mL anti-human CD3 (clone OKT3, BioLegend, Cat.-No. 317315) and 2 µg/mL anti-human CD28 (clone CD28.2, BioLegend, Cat.-No.: 302928) in T cell medium at 37 °C and 5% CO₂.

To determine binding of 4-1BBL trimer-containing Fc fusion antigen binding molecules to human PBMCs, 0.1 x 10⁶ naïve or activated PBMCs were added to each well of a round-bottom suspension cell 96-well plates (Greiner bio-one, cellstar, Cat. No. 650185). Plates were centrifuged 4 minutes with 400 x g and at 4 °C. Supernatant was discarded. Afterwards cells were stained in 100 µL/well DPBS containing 1:1000 diluted LIVE/DEAD Fixable Blue Dead Cell Stain Kit, for UV excitation (Life Technologies, Molecular Probes, L-23105) or Fixable Viability Dye eF660 (eBioscience 65-0864-18) or LIVE/DEAD Fixable Green Dead Cell Stain Kit (Life Technologies, Molecular Probes, L-23101) for 30 minutes at 4 °C in the dark. If DAPI was used as Live/Death stain, this staining step was skipped. Cells were washed once with 200

μL cold FACS buffer (DPBS supplied with 2 % (v/v) FBS, 5 mM EDTA pH8 (Amresco, Cat. No. E177) and 7.5 mM sodium azide (Sigma-Aldrich S2002).

Next, 50 μL/well of 4 °C cold FACS buffer containing different titrated concentrations of 4-1BBL trimer-containing Fc fusion antigen binding molecules were added and cells were
 5 incubated for 120 minutes at 4 °C, washed four times with 200 μL/well 4 °C FACS buffer and resuspended. Cells were further stained with 50 μL/well of 4 °C cold FACS buffer containing 0.67 μg/mL anti-human CD3-PerCP-Cy5.5 (clone UCHT1, mouse IgG1κ, BioLegend, Cat.-No. 300430) or 0.16 μL anti-human CD3-PE/Cy7 (clone SP34-2, mouse IgG1 κ, BD Pharmingen, Cat.-No. 557749, Lot 33324597), 0.67 μg/mL anti-human CD45-AF488 (clone HI30, mouse
 10 IgG1κ, BioLegend, Cat.-No. 304017) or 0.12 μg/mL anti-human CD56-FITC (clone NCAM16.2, mouse IgG2bκ, BD Pharmingen, Cat.-No. 345811) or 1 μL anti-human CD56-APC (clone B159, mouse IgG1 κ, BD Pharmingen, Cat.-No. 555518, Lot 3098894), 0.25 μg/mL anti-human CD4-BV421 (clone RPA-T4, mouse IgG1κ, BioLegend, Cat.-No. 300532) or 0.23 μg/mL anti-human CD4-BV421 (clone OKT4, mouse IgG2bκ, BioLegend, Cat.-No. 317434), 0.25 μL anti-human
 15 CD8a-APC (clone RPA-T8, mouse IgG1κ, BD Pharmingen, Cat.-No. 555369) or 0.67 μL anti-human CD8a-APC/Cy7 (clone RPA-T8, mouse IgG1κ, BioLegend, Cat.-No. 301016) or 0.83 ng/mL anti-human CD8a-BV711 (clone RPA-T8, mouse IgG1κ, BD Pharmingen, Cat.-No. 301044) and 5 μg/mL PE-conjugated AffiniPure anti-human IgG Fcγ-fragment-specific goat IgG F(ab')₂ fragment (Jackson ImmunoResearch, Cat. No. 109 116 098 or 109 116 170). Cells were
 20 washed twice with FACS-buffer. If cells were stained with fixable viability dyes, they were fixed with 50 μL/well DPBS containing 1 % formaldehyde (Sigma, HT501320-9.5L). Cells were resuspended in FACS buffer and acquired the next or the same day using a 5-laser LSR-Fortessa (BD Bioscience with DIVA software) or 3-laser Miltenyi Quant Analyzer 10 (Miltenyi Biotec) and Flow Jo (FlowJo X 10.0.7). If DAPI staining was used to detect dead cells, they were
 25 resuspended in 80 μL/well FACS buffer containing 0.2 μg/mL DAPI (Santa Cruz Biotech, Cat. No. Sc-3598) and acquired the same day using a 5-laser LSR-Fortessa (BD Bioscience with DIVA software).

As shown in Figure 6, both FAP-targeted or untargeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules did not bind to resting human CD4+ T cells and showed no
 30 detectable binding to resting CD8+ T cells and NK cells. In contrast, both constructs bound strongly to activated NK, CD8+ or CD4+ T cells, although the latter showed approximately 10 fold lower intensity of specific fluorescence as compared to the NK cells and 20 fold decreased intensity of specific fluorescence as compared to CD8+ T cells.

Figures 7A and 7B show the binding of Constructs 1.1 to 1.10 as prepared in Example 1 on
 35 4-1BB-expressing activated human CD3+ CD8+ T cells and 4-1BB-expressing activated human

CD3+ CD4+ T cells, respectively. Table 35 shows the EC₅₀ values as measured for Constructs 1.1 to 1.10.

Table 35: Binding on activated human CD3+ CD8+ T cells and CD3+ CD4+ T cells

Construct	EC ₅₀ [nM] 4-1BB ⁺ CD8 ⁺	EC ₅₀ [nM] 4-1BB ⁺ CD4 ⁺
Control B	0.11	16.21
1.1	0.43	4.99
1.2	0.18	20.79
1.3	0.07	2.82
1.4	0.19	0.34
1.5	0.17	2.67
1.6	0.19	0.95
1.7	0.26	16.47
1.8	0.14	2.77
1.9	0.18	12.92
1.10	0.12	0.3

5 Figures 8A and 8B show the binding of Constructs 2.1, 2.3, 2.4, 2.5 and 2.6 as prepared in Example 2 on CD4+ and CD8+ from fresh human blood and on activated 4-1BB-expressing CD4+ T cells and CD8 + T cells, respectively. Gates were set on living CD45+ CD3+ CD4+ or CD45+ CD3+ CD8+ T cells and MFI of PE-conjugated AffiniPure anti-human IgG IgG Fcγ-fragment-specific goat F(ab')₂ fragment were blotted against the titrated concentration of
10 targeted split trimeric 4-1BB ligand Fc fusion variants. Table 36 shows the EC₅₀ values as measured for Constructs 2.1, 2.3, 2.4, 2.5 and 2.6.

Table 36: Binding on activated 4-1BB-expressing CD4+ T cells and CD8 + T cells

Construct	EC ₅₀ [nM] 4-1BB ⁺ CD8 ⁺	EC ₅₀ [nM] 4-1BB ⁺ CD4 ⁺
Control B	0.36	0.42
Control C	0.39	0.41
Control E	0.57	0.76
2.1	0.21	0.24
2.3	0.44	0.3
2.4	0.3	0.38

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2.5	0.35	0.68
2.6	0.33	0.24

5.2 Binding of FAP-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecule to activated mouse splenocytes

Mouse spleens were collected in 3 mL PBS and a single cell suspension was generated using gentle MACS tubes (Miltenyi Biotec Cat.-No. 130-096-334) and gentleMACS Octo Dissociator (Miltenyi Biotec). Afterwards splenocytes were filtered through a 30 µm Pre-Separation Filters (Miltenyi Biotec Cat.-No. 130-041-407) and centrifuged for 7 min at 350 x g and 4°C. Supernatant was aspirated and cells were resuspended in RPMI 1640 medium supplied with 10 % (v/v) FBS, 1 % (v/v) GlutaMAX-I, 1 mM Sodium-Pyruvate, 1 % (v/v) MEM non-essential amino acids, 50 µM β-Mercaptoethanol and 10 % Penicillin-Streptomycin (SIGMA, Cat.-No. P4333). 10⁶ cells/mL were cultured for 2 days in a 6-well tissue culture plate coated with 10 µg/mL anti-mouse CD3ε Armenian Hamster IgG (clone 145-2C11, BioLegend, Cat.-No. 100331) and 2 µg/mL anti-mouse CD28 Syrian Hamster IgG (clone 37.51, BioLegend, Cat.-No. 102102).

Activated mouse splenocytes were harvested, washed in DPBS, counted and 0.1 x 10⁶ cells were transferred to each well of a 96 U-bottom non-tissue culture treated well plate. Supernatant was removed and cells were stained in 100 µL/well DPBS containing 1:5000 diluted Fixable Viability Dye eF660 (Bioscience, Cat.-No. 65-0864-18) for 30 min at 4°C. Cells were washed with PBS and stained in 50 µL FACS buffer containing different concentration of FAP-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (FAP split 4-1BBL trimer), untargeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (DP47 split 4-1BBL trimer) or anti-mouse CD137 human IgG1 P329G LALA mAb (clone Lob.12.3, BioXcell Catalog#: BE0169). Cells were incubated for 120 min at 4°C. Cells were washed four times with FACS buffer and stained in 50 µL/well FACS buffer containing 10 µg/mL purified anti-mouse CD16/CD32 rat IgG-Fc-Block (BD Pharmingen, Cat.-No. 553142 clone 2.4G2), 5 µg/mL anti-mouse CD8b rat IgG2bκ-FITC (BioLegend, Cat.-No. 126606, clone YTS156.7.7), 0.67 µg/mL anti-mouse CD3 rat IgG2bκ-APC-Cy7 (BioLegend, Cat.-No. 100222, clone 17A2), 0.67 µg/mL anti-mouse CD4 rat IgG2bκ-PE-Cy7 (BioLegend, Cat.-No. 100422, clone GK1.5), 2 µg/mL anti-mouse NK1.1 Mouse (C3H x BALB/c) IgG2aκ-PerCp-Cy5.5 (BioLegend, Cat.-No. 108728, clone PK136) and 10 µg/mL PE-conjugated AffiniPure polyclonal F(ab')₂ Fragment goat anti-human IgG, Fcγ fragment specific, minimal cross-reactive to bovine mouse and rabbit serum proteins (Jackson ImmunoResearch, Cat.-No. 109-116-170) for 30 min at 4°C. Cells were washed twice with 200 µL/well cold FACS buffer. Cells were fixed with 50 µL/well DPBS

containing 1 % formaldehyde. Cells were resuspended in FACS-buffer and acquired the next day using a 5-laser LSR-Fortessa (BD Bioscience with DIVA software).

As shown in Figure 9, FAP-targeted hu4-1BB ligand trimer-containing Fc fusion antigen binding molecules (FAP split hu4-1BBL trimer) and untargeted hu4-1BB ligand trimer-
5 containing Fc fusion antigen binding molecules (DP47 split hu4-1BBL trimer) do not bind to mouse 4-1BB. Therefore activity cannot be tested in immune competent mice. For in vivo mode of action studies either humanized mouse models in immune incompetent mice or surrogates containing mouse 4-1BBL trimers as shown in Figure 3 have to be used.

5.3 Binding to FAP-expressing tumor cells

10 For binding assays on FAP expressing cells, the human melanoma cell line MV-3 (see Ruiter et al., Int. J. Cancer 1991, 48(1), 85-91), WM-266-4 (ATTC CRL-1676) or NIH/3T3-huFAP clone 39 cell line were used. To generate the latter cell line, NIH/3T3 cells were transfected with human FAP (NIH/3T3-huFAP clone 39). The cells were generated by transfection of mouse embryonic fibroblast NIH/3T3 cells (ATCC CRL-1658) with the
15 expression pETR4921 plasmid encoding human FAP under a CMV promoter. Cells were maintained in the presence of 1.5 µg/mL puromycin (InvivoGen, Cat.-No.: ant-pr-5). 0.1×10^6 of FAP expressing tumor cells were added to each well of a round-bottom suspension cell 96-well plates (Greiner bio-one, cellstar, Cat.-No. 650185). Cells were washed once with 200 µL DPBS and pellets were resuspended. 100 µL/well of 4 °C cold DPBS buffer containing 1:5000 diluted
20 Fixable Viability Dye eFluor 450 (eBioscience, Cat.-No. 65-0863-18) or Fixable Viability Dye eFluor 660 (eBioscience, Cat.-No. 65-0864-18) were added and plates were incubated for 30 minutes at 4 °C. Cells were washed once with 200 µL 4 °C cold DPBS buffer and resuspended in 50 µL/well of 4 °C cold FACS buffer (DPBS supplied with 2 % (v/v) FBS, 5 mM EDTA pH 8 (Amresco, Cat.-No. E177) and 7.5 mM Sodium azide (Sigma-Aldrich S2002) containing
25 different concentrations of titrated 4-1BBL trimer-containing Fc fusion antigen binding molecules, followed by incubation for 1 hour at 4 °C. After washing four times with 200 µL/well, cells were stained with 50 µL/well of 4 °C cold FACS buffer containing 30 µg/mL FITC-conjugated AffiniPure anti-human IgG Fcγ-fragment-specific goat F(ab')₂ fragment (Jackson ImmunoResearch, Cat.-No. 109-096-098) or 5 µg/mL PE-conjugated AffiniPure anti-
30 human IgG Fcγ-fragment-specific goat F(ab')₂ fragment (Jackson ImmunoResearch, Cat. No. 109-116-098 or 109-116-170) for 30 minutes at 4 °C. Cells were washed twice with 200 µL 4 °C FACS buffer and then resuspended in 50 µL/well DPBS containing 1 % formaldehyde. The same or the next day cells were resuspended in 100 µL FACS-buffer and acquired using 5-laser LSR-Fortessa (BD Bioscience with DIVA software) or 3-laser Miltenyi Quant Analyzer 10
35 (Miltenyi Biotec) and Flow Jo (FlowJo X 10.0.7).

As shown in Figure 10, the FAP-targeted 4-1BB ligand trimer-containing Fc(kih) fusion antigen binding molecule (FAP split 4-1BBL trimer) Construct 1.1, but not the untargeted, DP47-Fab-containing construct (DP47 split 4-1BBL trimer) Control A, efficiently bound to human fibroblast activation protein (FAP)-expressing melanoma (10A) MV-3 cells or (10B) WM-266-4 cells.

Figure 11A shows the binding of Constructs 1.1 to 1.10 as prepared in Example 1 to human-FAP expressing human melanoma MV-3 cells and in Figure 11B the binding of Construct 1.1., 1.2, 1.3 and 1.5 to human FAP expressing NIH/3T3-huFAP clone 39 transfected mouse embryonic fibroblast cells is presented. Table 37 shows the EC₅₀ values as measured for Constructs 1.1 to 1.10.

Table 37: Binding to human FAP-expressing tumor cells

Construct	EC ₅₀ [nM] FAP ⁺ MV-3	EC ₅₀ [nM] NIH/3T3-hu FAP
1.1	4.14	12.2
1.2	5.36	9.35
1.3	-	14.97
1.4	5.13	-
1.5	0.53	10.06
1.6	8.16	-
1.7	4.09	-
1.8	2.79	-
1.9	4.22	-
1.10	4.31	-

Figure 12 shows the binding of different FAP (4B9)-targeted or untargeted split trimeric human 4-1BB ligand Fc (kih) constructs to human-FAP expressing human melanoma MV-3 cells (Figure 12A) and WM-266-4 cells (Figure 12B). The constructs 2.1, 2.3, 2.4, 2.5 and 2.6 were prepared as described in Example 2 and Controls were prepared as described herein before. Gates were set on living tumor cells and MFI of PE-conjugated AffiniPure anti-human IgG Fcγ-fragment-specific goat F(ab')₂ fragment were blotted against the titrated concentration of targeted split trimeric 4-1BB ligand Fc fusion constructs. Table 38 shows the EC₅₀ values as measured.

Table 38: Binding to human FAP-expressing tumor cells

Construct	EC ₅₀ [nM] FAP ⁺ MV-3	EC ₅₀ [nM] FAP ⁺ WM-266-4
2.1	1.66	0.99
2.3	0.53	0.42
2.4	0.83	0.59
2.5	1.66	1.2

5.4 Functional characterization of the murine targeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules

5.4.1 Binding to activated mouse splenocytes

Mouse spleens were collected in 3 mL PBS and a single cell suspension was generated using gentle MACS tubes (Miltenyi Biotec Cat.-No. 130-096-334) and gentleMACS Octo Dissociator (Miltenyi Biotec). Afterwards splenocytes were filtered through 30 µm Pre-Separation Filters (Miltenyi Biotec Cat.-No. 130-041-407) and centrifuged for 7 min at 350 x g and 4°C. Supernatant was aspirated and cells were resuspended in RPMI 1640 medium supplied with 10 % (v/v) FBS, 1 % (v/v) GlutaMAX-I, 1 mM Sodium-Pyruvate, 1 % (v/v) MEM non-essential amino acids, 50 µM β-Mercaptoethanol .

For binding on fresh mouse splenocytes cells were used directly. To induce mouse 4-1BB expression on T cells, mouse splenocytes were activated as following: 10⁶ cells/mL were cultured for 2 days in a 6-well tissue culture plate coated with 10 µg/mL anti-mouse CD3ε Armenian Hamster IgG (clone 145-2C11, BioLegend, Cat.-No. 100331) and 2 µg/mL anti-mouse CD28 Syrian Hamster IgG (clone 37.51, BioLegend, Cat.-No. 102102).

Fresh mouse splenocytes or activated mouse splenocytes were collected, washed in DPBS (Gibco life technologies, Cat.-No. 14190-136), counted and 0.1 x 10⁶ cells were transferred to each well of a 96 U-bottom non-tissue culture treated well plate (Greiner bio-one, cell star, Cat.-No. 650185). Supernatant was removed and cells were stained in 100 µL/well 4 °C cold DPBS containing 1:1000 diluted LIVE/DEAD Fixable Aqua Dead Cell Stain Kit (Life Technologies, L34957) for 30 min at 4°C. Cells were washed with cold DPBS and stained in 50 µL/well cold FACS buffer (DPBS supplied with 2 % (v/v) FBS, 5 mM EDTA pH8 (Amresco, Cat. No. E177) and 7.5 mM Sodium azide (Sigma-Aldrich S2002)) containing different concentration of mouse 4-1BB ligand trimer-containing Fc(kih) fusion molecules or mouse IgG1 κ Isotype control (BioLegend, Cat.-No. 400153, clone MOPC-21). Cells were incubated for 120 min at 4°C, washed four times with cold DPBS and stained in 50 µL/well cold FACS buffer

containing 30 µg/mL FITC-conjugated anti-mouse IgG Fc-gamma-specific goat IgG F(ab')₂ (Jackson ImmunoResearch, Cat.-No. 115-096-071) for 30 min at 4°C. Afterwards cells were washed twice with cold DPBS and stained with 50 µL/well FACS buffer supplied with 10 µg/mL purified anti-mouse CD16/CD32 rat IgG-Fc-Block (BD Pharmingen, Cat.-No. 553142 clone 2.4G2), 0.67 µg/mL anti-mouse CD8a-APC-Cy7 (BioLegend, Cat.-No. 100714, clone 53-6.7), 0.67 µg/mL anti-mouse CD3ε-PerCP-Cy5.5 (BioLegend, Cat.-No. 100328, clone 145-2C11), 0.67 µg/mL anti-mouse CD4 rat IgG2bκ-PE-Cy7 (BioLegend, Cat.-No. 100422, clone GK1.5) for 30 min at 4°C. Cells were washed twice with 200 µL/well cold DPBS, fixed with 50 µL/well DPBS containing 1 % Formaldehyde and resuspended in FACS-buffer. Cells were acquired using 3-laser MACSQuant Analyzer 10 (Miltenyi Biotech) and Flow Jo v10.0.7 (FlowJo LLC). Gates were set on CD3⁺ CD8⁺ or CD3⁺ CD4⁺ T cells and the median fluorescence intensity (MFI) of FITC-conjugated anti-mouse IgG Fc-gamma-specific goat IgG F(ab')₂ was analyzed and normalized by the subtraction of the MFI of the blank control (no addition of mouse 4-1BB ligand trimer-containing Fc(kih) fusion molecule). The MFI was plotted against the concentration of used mouse 4-1BB ligand trimer-containing Fc(kih) fusion molecules to display the binding to mouse 4-1BB cell-bound molecule.

As can be seen in Figure 13, the murine 4-1BBL Constructs M.1 and M.2 as well as corresponding control molecules Control M.1 and Control M.2 bind with a quite similar affinity to mouse 4-1BB. Table 39 shows the EC₅₀ values as measured for Constructs M.1 and M.2 and the control molecules.

Table 39: Binding on activated 4-1BB-expressing CD4⁺ T cells and CD8⁺ T cells

Construct	EC ₅₀ [nM] 4-1BB ⁺ CD8 ⁺	EC ₅₀ [nM] 4-1BB ⁺ CD4 ⁺
Control M.1	0.95	0.74
M.1	0.87	0.52
Control M.2	0.78	0.6
M.2	0.54	0.42

5.4.2 Binding on FAP-expressing tumor cells

For binding assays on FAP expressing cells, the human melanoma cell line MV-3 (see Ruiter et al., Int. J. Cancer 1991, 48(1), 85-91) and WM-266-4 (ATTC CRL-1676) were used (anti-FAP specific clone 28H1 is mouse/human-crossreactive). 0.1 x 10⁶ of FAP expressing tumor cells were added to each well of a round-bottom suspension cell 96-well plates (Greiner bio-one, cellstar, Cat.-No. 650185). Cells were washed once with 200 µL cold DPBS and pellets

were resuspended in 100 μ L/well of 4 °C cold DPBS buffer containing 1:1000 diluted LIVE/DEAD Fixable Aqua Dead Cell Stain Kit (Life Technologies, L34957) and incubated for 30 min at 4°C. Cells were washed once with 200 μ L cold DPBS buffer and resuspended in 50 μ L/well of cold FACS buffer (DPBS supplied with 2 % (v/v) FBS, 5 mM EDTA pH8 (Amresco, Cat. No. E177) and 7.5 mM Sodium azide (Sigma-Aldrich S2002)) containing murine 4-1BB ligand trimer-containing Fc(kih) fusion molecules at a series of concentrations followed by incubation for 1 hour at 4 °C. After washing four times with 200 μ L DPBS/well, cells were stained with 50 μ L/well of 4 °C cold FACS buffer containing 30 μ g/mL FITC-conjugated anti-mouse IgG Fc-gamma-specific goat IgG F(ab')₂ (Jackson ImmunoResearch, Cat.-No. 115-096-071) for 30 min at 4 °C. Cells were washed twice with 200 μ L/well cold DPBS buffer, fixed with 50 μ L/well DPBS containing 1 % Formaldehyde and resuspended in FACS-buffer. Cells were acquired using 3-laser MACSQuant Analyzer 10 (Miltenyi Biotech) and Flow Jo v10.0.7 (FlowJo LLC). Gates were set on living cells and the median fluorescence intensity (MFI) of FITC-conjugated anti-mouse IgG Fc-gamma-specific goat IgG F(ab')₂ was analyzed and normalized by the subtraction of the MFI of the blank control (no addition of mouse 4-1BB ligand trimer-containing Fc(kih) fusion molecule). The MFI was plotted against the concentration of used murine 4-1BB ligand trimer-containing Fc(kih) fusion molecules to display the binding to murine 4-1BB cell-bound molecule. As expected, the murine 4-1BBL constructs M.1 and M.2 bind with a quite similar affinity to FAP whereas the control molecules do not bind.

Figure 14 shows the binding of the FAP-targeted or untargeted split trimeric murine 4-1BB ligand Fc (kih) Constructs M.1 and M.2 to human-FAP expressing human melanoma MV-3 cells (Figure 14A) and WM-266-4 cells (Figure 14B). Table 40 shows the EC₅₀ values as measured.

Table 40: Binding to human FAP-expressing tumor cells

Construct	EC ₅₀ [nM]	EC ₅₀ [nM]
	FAP ⁺ MV-3	FAP ⁺ WM-266-4
M.1	7.26	5.14
M.2	6.9	5.63

Example 6

Biological activity of the targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules

6.1. NF- κ B activation in HeLa cells expressing human 4-1BB

5 Generation of HeLa cells expressing human 4-1BB and NF- κ B-luciferase

The cervix carcinoma cell line HeLa (ATCC CCL-2) was transduced with a plasmid based on the expression vector pETR10829, which contains the sequence of human 4-1BB (Uniprot accession Q07011) under control of a CMV-promoter and a puromycin resistance gene. Cells were cultured in DMEM medium supplemented with 10 % (v/v) FBS, 1 % (v/v) GlutaMAX-I and 3 μ g/mL Puromycin.

4-1BB-transduced HeLa cells were tested for 4-1BB expression by flow cytometry: 0.2×10^6 living cells were resuspended in 100 μ L FACS buffer containing 0.1 μ g PerCP/Cy5.5 conjugated anti-human 4-1BB mouse IgG1 κ clone 4B4-1 (BioLegend Cat.-No.309814) or its isotype control (PerCP/Cy5.5 conjugated mouse IgG1 κ isotype control antibody clone MOPC-21, BioLegend Cat.-No.400150) and incubated for 30 minutes at 4 °C. Cells were washed twice with FACS buffer, resuspended in 300 μ L FACS buffer containing 0.06 μ g DAPI (Santa Cruz Biotec, Cat. No. Sc-3598) and acquired using a 5-laser LSR-Fortessa (BD Bioscience, DIVA software). Limited dilutions were performed to generate single clones as described: human-4-1BB-transduced HeLa cells were resuspended in medium to a density of 10, 5 and 2.5 cells/ml and 200 μ l of cell suspensions were transferred to round bottom tissue-culture treated 96-well plates (6 plates/cell concentration, TPP Cat.-No. 92697). Single clones were harvested, expanded and tested for 4-1BB expression as described above. The clone with the highest expression of 4-1BB (clone 5) was chosen for subsequent transfection with the NF- κ B-luciferase expression-vector 5495p Tranlucent HygB. The vector confers transfected cells both with resistance to Hygromycin B and capacity to express luciferase under control of NF- κ B-response element (back bone vector Panomics, Cat.-No. LR0051 with introduced HyB resistance). Human-4-1BB HeLa clone 5 cells were cultured to 70 % confluence. 50 μ g (40 μ L) linearized (restriction enzymes AseI and SalI) 5495p Tranlucent HygB expression vector were added to a sterile 0.4 cm Gene Pulser/MicroPulser Cuvette (Biorad, Cat.-No, 165-2081). 2.5×10^6 human-4-1BB HeLa clone 5 cells in 400 μ l supplement-free DMEM medium were added and mixed carefully with the plasmid solution. Transfection of cells was performed using a Gene Pulser Xcell total system (Biorad, Cat-No. 165-2660) under the following settings: exponential pulse, capacitance 500 μ F, voltage 160 V, resistance ∞ . Immediately after the pulse transfected cells were transferred to a 75 cm² tissue culture flask (TPP, Cat.-No. 90075) with 15 mL 37°C warm DMEM medium supplied with 10 % (v/v) FBS and 1 % (v/v) GlutaMAX-I. Next day, culture medium containing

3 µg/mL Puromycin and 200 µg/mL Hygromycin B (Roche, Cat.-No. 10843555001) was added. Surviving cells were expanded and limited dilution was performed as described above to generate single clones.

Clones were tested for 4-1BB expression as described above and for NF-κB-Luciferase activity as following: Clones were harvested in selection medium and counted using a Cell Counter Vi-cell xr 2.03 (Beckman Coulter, Cat.-No. 731050). Cells were set to a cell density of 0.33×10^6 cells/mL and 150 µL of this cell suspension were transferred to each well of a sterile white 96-well flat bottom tissue culture plate with lid (greiner bio-one, Cat.-No. 655083) and - as a control - to normal 96-well flat bottom tissue culture plate (TPP Cat.-No. 92096) to test survival and cell density the next day. Cells were incubated at 37 °C and 5 % CO₂ overnight. The next day 50 µL of medium containing different concentrations of recombinant human tumor necrosis factor alpha (rhTNF-α, PeproTech, Cat.-No. 300-01A) were added to each well of a 96-well plate resulting in final concentration of rhTNF-α of 100, 50, 25, 12.5, 6.25 and 0 ng/well. Cells were incubated for 6 hours at 37 °C and 5 % CO₂ and then washed three times with 200 µL/well DPBS. Reporter Lysis Buffer (Promega, Cat-No: E3971) was added to each well (40 µl) and the plates were stored over night at -20 °C. The next day frozen cell plates and Detection Buffer (Luciferase 1000 Assay System, Promega, Cat.-No. E4550) were thawed to room temperature. 100 uL of detection buffer were added to each well and the plate was measured as fast as possible using a SpectraMax M5/M5e microplate reader and the SoftMax Pro Software (Molecular Devices). Measured units of released light for 500 ms/well (URLs) above control (no rhTNF-α added) were taken as luciferase activity. The NF-κB-luc-4-1BB-HeLa clone 26 exhibiting the highest luciferase activity and a considerable level of 4-1BB-expression and was chosen for further use.

25 NF-κB activation in Hela cells expressing human 4-1BB co-cultured with FAP-expressing tumor cells

NF-κB-luciferase human-4-1BB HeLa cells were harvested and resuspended in DMEM medium supplied with 10 % (v/v) FBS and 1 % (v/v) GlutaMAX-I to a concentration of 0.2×10^6 cells/ml. 100 µl (2×10^4 cells) of this cell suspension were transferred to each well of a sterile white 96-well flat bottom tissue culture plate with lid (greiner bio-one, Cat. No. 655083) and the plate were incubated at 37 °C and 5 % CO₂ overnight. The next day 50 µL of medium containing titrated concentrations of FAP-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (FAP split 4-1BBL trimer) or DP47-untargeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (DP47 split 4-1BBL trimer) were added. FAP-expressing tumor cells (MV3, WM-266-4 or NIH/3T3-huFAP clone 39) were resuspended in DMEM medium supplied with 10 % (v/v) FBS and 1 % (v/v) GlutaMAX-I to a concentration of 2×10^6 cells/ml.

Suspension of FAP-expressing tumor cell (50 μ l, final ratio 1:5) or only medium were added to each well and plates were incubated for 6 hours at 37 °C and 5 % CO₂. Cells were washed two times with 200 μ L/well DPBS. 40 μ l freshly prepared Reporter Lysis Buffer (Promega, Cat-No: E3971) were added to each well and the plate were stored over night at -20 °C. The next day frozen cell plate and Detection Buffer (Luciferase 1000 Assay System, Promega, Cat. No. E4550) were thawed at room temperature. 100 μ L of detection buffer were added to each well and luciferase activity was measured as fast as possible using a SpectraMax M5/M5e microplate reader and a SoftMax Pro Software (Molecular Devices) counting light emission in URL (units of released light for 0.5s/well) or Victor3 1420 multilabel counter plate reader (Perkin Elmer) and the Perkin Elmer 2030 Manager Software counting light emission as counts per seconds (CPS) and blotted against the concentration of tested constructs.

FAP-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecule (FAP split 4-1BBL trimer) triggered activation of the NF κ B signaling pathway in the reporter cell line in the presence of FAP-expressing tumor cells. In contrast, the untargeted variant of the same molecule failed to trigger such an effect at any of the tested concentrations (Figure 16). This activity of targeted 4-1BBL was strictly dependent on the expression of FAP at the cell surface of tumor cells as no NF- κ B activation could be detected upon culturing of the NF- κ B reporter cell line with FAP-negative tumor cells even in the presence of FAP-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecule. The activities as measured for Constructs 1.1 to 1.10 are shown in Figure 17 and the data as measured for Constructs 2.1, 2.4 and 2.5 are presented in Figure 18.

6.2. NF κ B activation in HEK T293 cells expressing cynomolgus monkey 4-1BB

Generation of HEK T293 cells expressing cynomolgus monkey 4-1BB and NF κ B-luciferase

For the production of viral-like particles (VLP) the Human Embryonic Kidney (HEK) T293/17 (ATCC CRL-11268) was transfected using Lipofectamine® LTX Reagent with PLUS™ Reagent (Life Technologies, Cat.-No. 15338100) with the vector pETR14372 encoding a NF κ B-luciferase-IRIS-GFP reporter gene cassette (NF κ B-luc-GFP) accordingly to the manufacture's protocol. 6 hours later DMEM supplied with 10% FBS medium replacement was performed and VLP were harvested 4 days later. Fresh HEK 293T cells were transduced at a confluency of 70-80 % with the produced pETR14372-VLP and 4 μ g/mL polybrene. Cells were cultured for 24 h and a medium exchange was performed. The transduced HEK T293/17 cells were harvested and a limited dilution of 1 cell/well was performed to screen for stable single clones. The single clones were stimulated with 25 ng/mL TNF- α (PeproTech Inc. Cat.-No. 300-01A) in the medium and were screened for a positive GFP signal over time using the Incucyte

Zoom Fluorescence Microscope System (Essen Bioscience). After GFP signal recording cells were tested for luciferase activity using the Nano Glo Luciferase Kit (Promega, N1120) accordingly to the manufacture's protocol. Luciferase activity was measured using Victor3 1420 multilabel counter plate reader (Perkin Elmer) and the Perkin Elmer 2030 Manager Software.

- 5 Light emission was counted in counts per seconds (CPS) for 0.5 sec/well. The clone 61 showed the highest expression of GFP and Luciferase after TNF- α activation and was further used for the reporter cell line generation.

- As described above, new VLP were produced using the vector pETR14879 encoding cynomolgus monkey 4-1BB and a puromycine resistance and the HEK 293T NF κ B-fluc-GFP
- 10 clone 61 cell line was transduced at a confluency of 70-80% with the produced pETR14879-VLP and 4 μ g/mL polybrene. Cells were cultured for 24 h and a medium exchange was performed. Four days after transduction the cells were stained with PE-conjugated anti-human cynomolgus-crossreactive 4-1BB antibody (mouse IgG1 κ , clone MOPC-21, BioLegend, Cat.-No. 309804) in DPBS containing 1% FBS, were sorted by FACS (ARIA, BD) and seeded with 5 cells/well in
- 15 DMEM supplied with 10% FBS medium containing 1 μ g/mL Puromycine (InvivoGen, Cat.-No. ant-pr). Growing clones were tested as described for GFP and Luciferase activity after TNF- α stimulation and for high cynomolgus monkey 4-1BB expression by flow cytometry. Double positive clones were chosen and tested for Luciferase activity in the presence of monovalent FAP-targeted Construct 2.1 or Control B and FAP-expressing MV-3 or WM-266-4 cells. HEK
- 20 T293/17-NF- κ B-luc-GFP-cy4-1BB expressing Clone 61-13 was chosen to be used for all further experiments.

NF κ B activation of HEK T293/17 reporter cells expressing cynomolgus monkey 4-1BB co-cultured with FAP-expressing tumor cells

- HEK T293/17-NF κ B-luc-GFP-cy4-1BB expressing Clone 61-13 cells were harvested and
- 25 resuspended in DMEM medium supplied with 10 % (v/v) FBS and 1 % (v/v) GlutaMAX-I to a concentration of 0.2×10^6 cells/mL. 100 μ l (2×10^4 cells) of this cell suspension were transferred to each well of a sterile white 96-well flat bottom tissue culture plate with lid (greiner bio-one, Cat. No. 655083) and the plate were incubated at 37 °C and 5 % CO₂ overnight. The next day 50 μ L of medium containing different titrated concentrations of FAP-targeted or untargeted 4-1BB
- 30 ligand trimer-containing Fc fusion antigen binding molecules were added. FAP-expressing tumor cells (MV3 and WM-266-4) were resuspended in medium to a concentration of 2×10^6 cells/mL. Suspension of FAP-expressing tumor cell (50 μ l) was added to each well and plates were incubated for 6 hours at 37 °C and 5 % CO₂. The principle of the assay is shown in Figure 19. After incubation cells were washed three times with 200 μ L/well DPBS. 40 μ l freshly prepared
- 35 Reporter Lysis Buffer (Promega, Cat-No: E3971) were added to each well and plates were stored over night at -20 °C. The next day frozen cell plates and detection buffer (Luciferase 1000 Assay

System, Promega, Cat. No. E4550) were thawed to room temperature. 100 μ L of detection buffer were added to each well and luciferase activity was measured as fast as possible using SpectraMax M5/M5e (Molecular Devices) microplate reader (500 ms integration time, no filter collecting all wavelength). Light emission was counted in units of released light (URL) for 0.5 sec/well and blotted against the concentration of tested FAP-targeted or untargeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules. The results for Constructs of Example 2 are shown in **Figure 20**.

6.3 Antigen-specific CD8⁺ T cell-based assay

Isolation and culture of antigen-specific CD8 T cells

Fresh blood was obtained from a HLA-A2⁺ CMV-infected volunteer. PBMCs were isolated as described above. CD8 T cells were purified from PBMCs using a negative selection human CD8 T cell isolation Kit according to manufacturer's recommendations (Miltenyi Biotec, Cat. No. 130-094-156). Ten million of isolated CD8 T cells were resuspended in 1 mL sterile DPBS supplemented with 1 % (v/v) FBS along with 50 μ L of PE-labeled HLA-A2-pentamer containing the CMV-derived NLVPMVATV peptide (ProImmune, Cat. No. F008-2B) and incubated for 10 min at room temperature. Cells were washed twice with 3 mL sterile DPBS supplied with 1 % (v/v) FBS. Cells were resuspended in 1 mL cells DPBS supplied with 1 % (v/v) FBS containing 1 μ g/mL anti-human CD8-FITC (clone LT8, Abcam, Cat. No. Ab28010) and incubated for 30 minutes at 4°C. Cells were washed twice, resuspended to a concentration of 5×10^6 cells/mL in DPBS supplied with 1 % (v/v) FBS, and filtrated through a 30 μ m pre-separation nylon-net cell strainer (Miltenyi Biotec, Cat. No. 130-041-407). NLV-peptide-specific CD8⁺ T cells were isolated by FACS sorting using an ARIA cell sorter (BD Bioscience with DIVA software) with the following settings: 100 μ m nozzle and purity sort mask. Sorted cells were collected in a 15 ml polypropylene centrifuge tube (TPP, Cat. No. 91015) containing 5 ml RPMI 1640 medium supplied with 10 % (v/v) FBS, 1 % (v/v) GlutaMAX-I and 400 U/mL Proleukin. Sorted cells were centrifuged for 7 minutes at 350 x g at room temperature and resuspended in same medium to a concentration of 0.53×10^6 cells/mL. 100 μ L/well of this cell suspension were added to each well of a previously prepared feeder plate.

PHA-L-activated irradiated allogeneic feeder cells were prepared from PBMCs as previously described (Levitsky et al., 1998) and distributed to 96 well culture plates at 2×10^5 feeder cells per well.

After one day of culturing 100 μ L medium/well were removed from well containing sorted CD8⁺ T-cells and replaced by new RPMI 1640 medium supplemented with 10 % (v/v) FBS and 1 % (v/v) GlutaMAX-I and 400 U/mL Proleukin, this was repeated during culture on a regular basis (every 2-4 days). As soon as cells start to proliferate, they were transferred to 24-well flat-

bottom tissue culture plate (TPP, 92024). Cells were expanded/split and reactivated with new feeder cell preparation on a regular basis.

Activation assay of antigen-specific CD8+ T cells

MV3 cells were harvested and washed with DPBS and 2×10^7 cells were resuspended in
 5 250 μ L C diluent of the PKH-26 Red Fluorescence Cell linker Kit (Sigma, Cat.-No. PKH26GL).
 1 μ L PKH26-Red-stain solution was diluted with 250 μ L C diluent and added to the suspension
 of MV3 cells which were then incubated for 5 min at room temperature in the dark. This was
 followed by addition of 0.5 mL FBS and cells were incubated for 1 minute and washed once
 with T cell medium consisting of RPMI 1640 medium supplemented with 10 % (v/v) FBS, 1 %
 10 (v/v) GlutaMAX-I, 1 mM Sodium-Pyruvate, 1 % (v/v) MEM non-essential amino acids and 50
 μ M β -Mercaptoethanol. 1×10^6 MV3 cells/mL were resuspended in T cell medium and
 separated into three tubes. Synthetic NLVPMVATV peptide (obtained from thinkpeptides) was
 added to a final concentration of 1×10^{-9} M or 1×10^{-8} M and cells were incubated for 90 min.
 MV3 cells were washed once with T cell medium and resuspended to a density of 0.5×10^6
 15 cells/mL, distributed (100 μ L/well) to a 96-well round bottom cell-suspension plate (Greiner bio-
 one, cellstar, Cat.-No. 650185) and incubated over night at 37 °C and 5 % CO₂. The principle of
 the assay is shown in Figure 21.

The next day, 50 μ L/well T cell medium containing different titrated concentrations of
 targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules were added. NLV-
 20 specific CD8 T cells were harvested, CFDA-SE (5(6)-Carboxyfluoresceindiacetate-N-
 succinimidylester, SIGMA-Aldrich, Cat.-No. 21888-25MG-F) was added to a final concentration
 of 40 nM and cells were incubated under rotation for 15 min at 37 °C. Labeling was stopped by
 adding FBS, cells were washed and resuspended in T cell medium to a final concentration of
 0.125×10^6 cells/mL. 50 μ L of this CFSE-labeled CD8 T cell suspension were added to each
 25 well (final E:T ratio = 1:8). Cell plates were incubated for 24 h, 50 μ L/well were removed and 50
 μ L T cell medium containing 2.64 μ L/mL Golgi stop (Protein Transport Inhibitor containing
 Monesin, BD Bioscience, Cat.-No. 554724) were added to each well (final concentration 0.66
 μ L/mL). Cells were incubated for 4 h and then plates were washed with 200 μ L/well DPBS and
 stained with 100 μ L/well 4 °C DPBS containing 1:5000 diluted Fixable Viability Dye-eF450
 30 (eBioscience, Cat.-No. 65-0864) for 30 minutes at 4°C. Cell plates were washed with 200
 μ L/well DPBS followed by staining with fluorescent dye-conjugated antibodies: anti-human
 CD137-PerCP/Cy5.5 (clone 4B4-1, mouse IgG1 κ , BioLegend, Cat.-No. 309814), anti-human
 CD8-BV605 (clone RPA-T8, mouse IgG1 κ , BioLegend, Cat.-No. 301012) or 0.67 μ g/mL anti-
 human CD8a-APC/Cy7 (clone RPA-T8, mouse IgG1 κ , BioLegend, Cat.-No. 301016) and anti-
 35 human CD25 PE/Cy7 (clone BC96, mouse IgG1 κ , BioLegend, Cat.-No. 302612). After
 incubation for 30 min at 4°C, cells were washed twice with 200 μ L/well FACS buffer,

resuspended in 50 μ L/well freshly prepared FoxP3 Fix/Perm buffer (eBioscience Cat.-No. 00-5123 and 00-5223) and incubated for 30 min at 4 °C. Plates were washed twice with 200 μ L/well Perm-Buffer (DPBS supplied with 2 % (v/v) FBS, 1 % (w/v) saponin (Sigma Life Science, S7900) and 1 % (w/v) sodium azide (Sigma-Aldrich, S2002) and stained with 50 μ L/well Perm-Buffer (eBioscience, Cat.-No. 00-8333-56) containing 0.25 μ g/mL anti-human IFN γ -APC (clone B27, mouse IgG1 κ , BioLegend, Cat.-No. 506510) or 0.33 μ g/mL anti-human IFN γ -BV510 (clone 4S.B3, mouse IgG1 κ , BioLegend, Cat.-No.502543). Plates were incubated for 1 h at 4 °C and washed twice with 200 μ L/well Perm-Buffer. For fixation, 50 μ L/well DPBS containing 1 % formaldehyde were added. The same or the next day, cells were resuspended in 100 μ L/well FACS buffer and acquired using a 5-laser Fortessa flow cytometer (BD Bioscience with DIVA software) or 3-laser Miltenyi Quant Analyzer 10 (Miltenyi Biotec) and Flow Jo (FlowJo X 10.0.7).

As shown in Figures 22 and 23 for Constructs 1.1 to 1.10 and in Figures 24 and 25 for Constructs 2.1, 2.3 and 2.4, antigen-specific CD8⁺ T cells, but not unstimulated controls, exhibited increased levels of surface 4-1BB expression in the presence of FAP-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecule (FAP split 4-1BBL trimer). This effect of 4-1BBL was dose dependent and required FAP-targeting as addition of the untargeted control molecule did not affect the level of 4-1BB expression. Furthermore, T-cells activated at the higher peptide concentration (1×10^{-8} M) showed sustained secretion of INF γ in the presence of FAP-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecule (FAP split 4-1BBL trimer). Collectively, these data demonstrate that the antigen-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecule modulates the surface phenotype and responsiveness of antigen specific T-cells in a targeting dependent manner.

6.4 Comparison of cell-targeted and untargeted mouse 4-1BBL Fc fusion antigen binding molecules

Targeted and untargeted mouse 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (FAP split mouse 4-1BBL trimer and DP47 split mouse 4-1BBL trimer) were prepared as described in Example 1.3.

To compare the bioactivity of cell-targeted and untargeted mouse 4-1BB ligand trimer-containing Fc fusion antigen binding molecules, Proliferation Dye eFlour670- labeled (eBioscience, Cat.-No.65-0840-90) or CellTrace Violet Cell Proliferation dye-labeled (Cell tracer, Cat.-No. C34557) fresh mouse splenocytes were cocultured for 3-4 days in 96 well tissue culture U-bottom plates (TTP, Cat.-No. 92097) with adherent 50 Gy irradiated NIH/3T3-huFAP clone 39 cells (generation see 5.3) in RPMI 1640 medium (Gibco, Cat.-No. 42401-042) supplied with 10 % (v/v) FBS, 1 % (v/v) GlutaMAX-I, 1 mM Sodium-Pyruvate, 1 % (v/v) MEM non-

essential amino acids and 50 μ M β -Mercaptoethanol in the presence of 0.5 μ g/mL anti-mouse CD3 Syrian hamster IgG (clone 145-2C11, BD, Cat.-No. 553057) and the indicated drug candidate molecule added at a range of concentrations (Figure 26). After three or four days, cells were washed with FACS buffer and stained for 30 min at 4°C in 25 μ L FACS buffer/well containing anti-mouse CD8 ratIgG2a-BV711 (BioLegend, Cat.-No. 100747, clone 53-6.7,) and anti-mouse CD4 ratIgG2a-BV421 (BioLegend, Cat.-No. 100544, clone RM4-5) and 0.67 μ g/mL anti-mouse CD137 (4-1BB) Syrian hamster IgG-PE (BioLegend, Cat.-No. 106106, clone 17B5) and anti-mouse CD25-PErCP-Cy5.5 ratIgG2b (BioLegend, Cat.-No. 1019112). Cells were washed and incubated for 1 h at room temperature in prepared Fix/Perm Buffer (Foxp3 / Transcription Factor Staining Buffer Set, eBioscience, Cat.-Ni. 00-5523-00). Cells were washed twice with freshly prepared Perm buffer and co-stained with 25 μ L/well Perm-buffer containing fluorescently-labeled antibodies against the cytotoxic lineage transcription factor Eomes, i.e. anti-mouse Eomes ratIgG2a-AlexaFluor488 (eBioscience, Cat.-No. 534875, clone Dan11mag) and – if CD137 was not stained - against the cytotoxic effector molecule granzyme B, i.e. anti-mouse ratIgG2a granzyme B-PE (eBioscience, Cat.-No. 128822, clone 16G6) for 1 h at room temperature. Cells were then washed twice, resuspended in FACS buffer and acquired using laser Fortessa flow cytometer (BD Bioscience with DIVA software) or the 3-laser MACSQuant Analyzer 10 (Miltenyi Biotech) and Flow Jo v10.0.7 (FlowJo LLC). Gates were set on living CD8+ T cells and CD4+ T cells and the frequency of proliferating cells was determined as well as the expression levels of CD25, Eomes and granzyme B or CD137. The proliferation frequency and frequencies and MFIs of activation markers were blotted against the concentration of used mouse 4-1BB ligand trimer-containing Fc(kih) fusion molecules to display the functional activity. As can be seen in Figure 27, an increase in proliferating CD8+ T cells could be observed for Constructs M.1 and M.2.

6.5 Liver changes in mice treated with anti-murine 4-1BB antibody Lob 12.3 (muIgG1 Wt) or with Construct M.2

C57BL/6 mice bearing MC38-muFAP (murine colorectal cancer model) s.c. were treated once per week for 3 weeks with agonistic anti-murine 4-1BB antibodies targeted to FAP (Efficacy Study 020-GA1401: “Experiment to show efficacy of 4-1BB targeted therapy in combination with a-PD-L1 in MC38-muFAP s.c. model in C57B6 mice.”). Antibodies used were Lob 12.3 muIgG1 Wt (with “wildtype” Fc, clone Lob 12.3 from BioXcell Catalog#: BE0169) or Construct M.2 with DAPG mutation (inactive Fc). The two antibodies were administered once weekly for three consecutive weeks. Four animals/group were sacrificed 7 days after last treatment and livers examined microscopically.

Liver changes were observed only in animals receiving Lob 12.3 muIgG1 Wt, consisting in foci of hepatocellular degeneration with accumulation of F4/80 positive macrophages and a

lower amount of mixed population of inflammatory cells (mainly lymphocytes) frequently showing a vasocentric distribution. Occasionally single cell necrosis of hepatocytes, and perivascular mononuclear cell infiltrates in portal spaces were noted. No treatment related findings were observed in the liver of animals receiving Construct M.2 (Table 41).

5 **Table 41: Incidence of Histopathological Findings (n=4/group)**

Treatment	Vehicle	Lob 12.3 muIgG1 Wt	Construct M.2
Foci of hepatocellular degeneration with macrophages and inflammatory cells	-	4	-
Perivascular inflammatory cells infiltrates	-	4	-
Single cell necrosis	-	4	-

Hepatitis, attributed to crosslinking by Fc γ R_s in the liver, has been observed in patients treated with Urelumab BMS-663513 (Ascierto P.A. et al. 2010) and in mouse using the mouse surrogate. The absence of liver findings in animals treated with an antibody with inactive Fc support this hypothesis.

6.6 Determination of pharmacokinetic parameters of human 4-1BB ligand trimer-containing Fc fusion antigen binding molecules

In order to test if the human 4-1BB ligand trimer-containing Fc fusion antigen binding molecules of the invention are suitable for pharmaceutical use, the pharmacokinetic parameters (PK data) such as clearance, volume of distribution or elimination half-time ($t_{1/2}$) in mice were determined. Thus, the following experiments were carried out:

Experiment A: Single dose PK of Construct 1.2 and Control B in healthy NOG mice

NOG female mice at an average age of 8 to 10 weeks at start of experiment (purchased from Taconic, SOPF facility) were maintained under specific-pathogen-free condition with daily cycles of 12 h light /12 h darkness according to committed guidelines (GV-Solas; Felasa; TierschG). Experimental study protocol was reviewed and approved by local government (P 2011128). After arrival animals were maintained for one week to get accustomed to new environment and for observation. Continuous health monitoring was carried out on regular basis.

A single dose pharmacokinetic study (SDPK) was performed to evaluate exposure of Construct 1.2 and Control B. An i.v. bolus administration of 2.5 mg/kg was administered to NOG mice and blood samples were taken at selected time points for pharmacokinetic evaluation. Mouse serum samples were analyzed by ELISA. Biotinylated human 4-1BB, test samples, Digoxigenin labelled anti-huCH1 antibody and anti-Digoxigenin detection antibody (POD) were added stepwise to a 96-well streptavidin-coated microtiter plate and incubated after every

step for 1h at roomtemperature. The plate was washed three times after each step to remove unbound substances. Finally, the peroxidase-bound complex was visualized by adding ABTS substrate solution to form a colored reaction product. The reaction product intensity which was photometrically determined at 405 nm (with reference wavelength at 490 nm) is proportional to the analyte concentration in the serum sample. The calibration range of the standard curve for the constructs was 0.156 to 10 ng/ml, where 3 ng/ml is the lower limit of quantification (LLOQ). Figure 28A shows the decrease in concentration over the time as observed in this experiment.

Experiment B: Single dose PK of Constructs 2.1, 2.3, Control B and Control C in tumor bearing NOG mice humaniced with stem cells

A single dose pharmacokinetic study (SDPK) was performed to evaluate exposure of Construct 2.1, 2.3, Control B and Control C. NSG female mice transferred with human stem cells were delivered by Jackson Laboratories. Mice were maintained under specific-pathogen-free condition with daily cycles of 12 h light /12 h darkness according to committed guidelines (GV-Solas; Felasa; TierschG). Experimental study protocol was reviewed and approved by local government (ZH193-2014). After arrival animals were maintained for one week to get accustomed to new environment and for observation. Continuous health monitoring was carried out on regular basis.

Human MKN45 cells (human gastric carcinoma) were originally obtained from ATCC and after expansion deposited in the Glycart internal cell bank. Cells were cultured in DMEM containing 10% FCS. Cells were cultured at 37 °C in a water-saturated atmosphere at 5 % CO₂. In vitro passage 9 was used for subcutaneous injection, at a viability of 97%. Human fibroblasts NIH-3T3 were engineered at Roche Nutley to express human FAP. Clone 39 was used at an in vitro passage number 12 and at a viability of 98%. 50 microliters cell suspension (1x10⁶ MKN45 cells + 1x10⁶ 3T3-huFAP) mixed with 50 microliters Matrigel were injected subcutaneously in the flank of anaesthetized mice. An i.v. bolus administration of 10 mg/kg was administered to humaniced mice when tumor reached an average size of 190 mm³. Blood samples were taken at selected time points for pharmacokinetic evaluation. Mouse serum samples were analyzed by ELISA. Biotinylated human 4-1BB, test samples, Digoxigenin labelled anti-huCH1 antibody and anti-Digoxigenin detection antibody (POD) were added stepwise to a 96-well streptavidin-coated microtiter plate and incubated after every step for 1h at room temperature. The plate was washed three times after each step to remove unbound substances. Finally, the peroxidase-bound complex is visualized by adding ABTS substrate solution to form a colored reaction product. The reaction product intensity which was photometrically determined at 405 nm (with reference wavelength at 490 nm) is proportional to the analyte concentration in the serum sample. The calibration range of the standard curve for the constructs was 0.156 to 10 ng/ml, where 3 ng/ml is the lower limit of quantification (LLOQ).

Figure 28B shows the decrease in concentration of the constructs over the time as observed in this experiment.

Experiment C: Single dose PK of Construct 2.1 and 2.3 in healthy NOG mice

NOG female mice at an average age of 8-10 weeks at start of experiment (purchased from Taconic, SOPF facility) were maintained under specific-pathogen-free condition with daily cycles of 12 h light /12 h darkness according to committed guidelines (GV-Solas; Felasa; TierschG). Experimental study protocol was reviewed and approved by local government (P 2011128). After arrival animals were maintained for one week to get accustomed to new environment and for observation. Continuous health monitoring was carried out on regular basis.

A single dose pharmacokinetic study (SDPK) was performed to evaluate exposure of Construct 2.1 and 2.3. An i.v. bolus administration of 2.5 mg/kg was administered to NOG mice and blood samples were taken at selected time points for pharmacokinetic evaluation. Mouse serum samples were analyzed by ELISA. Biotinylated human 4-1BB, test samples, Digoxigenin labelled anti-huCH1 antibody and anti-Digoxigenin detection antibody (POD) were added stepwise to a 96-well streptavidin-coated microtiter plate and incubated after every step for 1h at roomtemperature. The plate is washed three times after each step to remove unbound substances. Finally, the peroxidase-bound complex is visualized by adding ABTS substrate solution to form a colored reaction product. The reaction product intensity, which is photometrically determined at 405 nm (with reference wavelength at 490 nm), is proportional to the analyte concentration in the serum sample. The calibration range of the standard curve for the constructs was 0.156 to 10 ng/ml, where 3 ng/ml is the lower limit of quantification (LLOQ). Figure 28C shows the observed decrease in concentration over the time.

The tested constructs 2.1 and 2.3 are stable enough in the body and possess PK parameters in a suitable range for pharmaceutical development. It can also be concluded from the results that construct 2.1 is slightly more stable.

6.7 FAP prevalence in human tumors

The prevalence of FAP in human tumors was evaluated as described in WO 2014/161845 to get an understanding on possible clinical use of FAP-targeted constructs.

Rat anti-human Seprase antibody (IgG2a, clone D8) from Vitatex (MABS1001) was used to immunostain 2,5 µm FFPET sections from various tumour indications on the Ventana Benchmark XT. Sections were subjected to standard CC1 treatment followed by antibody incubation for 60' at 37°C at a concentration of 5 µg/mL in Dako antibody diluent (S3022) and positive staining was detected using the Ultraview DAB detection system (Ventana #760-4456).

Matched isotype antibody from Abcam (ab18450) was used as the negative control. FAP+ stromal infiltrate was present in human tumors of different indications including head and neck squamous cell carcinoma (HNSCC), breast cancer, colorectal cancer (CRC), pancreatic cancer (PAC), gastric cancer, non-small-cell lung carcinoma (NSCLC) and Mesothelioma marking

5 potentially interesting clinical indications for a FAP-targeted constructs (Table 42).

Table 42: FAP prevalence in human tumors

Tumor Type	% cases with moderate to high grade of FAP ⁺ infiltrate	No. of samples investigated
HNSCC	90	10
Breast Cancer	77	105
triple negative BC	80	7
CRC	77	90
PAC	74	19
Gastric Cancer	68	28
NSCLC	66	90
Mesothelioma	60	10

Example 7**7.1 Preparation of CD19 (8B8-018) targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules****7.1.1 Preparation, purification and characterization of CD19 antigen Fc fusion for phage display campaign**

In order to express and purify the human and cynomolgus CD19 ectodomain in a monomeric state (human CD19 see SEQ ID NO:31), the respective DNA fragment was fused to a human IgG1 Fc gene segment containing the “knob” mutations (human: SEQ ID NO: 186; cynomolgus: SEQ ID NO: 188) and was transfected with an “Fc-hole” (SEQ ID NO: 86) counterpart (Merchant et al., 1998). An IgA cleavage site (PTPPTP) was introduced between an antigen ectodomain and the Fc knob chain. An Avi tag for directed biotinylation was introduced at the C-terminus of the antigen-Fc knob chain and mutations H435R and Y436F were introduced in the Fc hole for purification purposes (Jendeberg L. et al, J. Immunological methods, 1997). Combination of the antigen-Fc knob chain containing the S354C/T366W mutations (human: SEQ ID NO: 187; cynomolgus: SEQ ID NO: 189), with a Fc hole chain containing the Y349C/T366S/L368A/ Y407V mutations (SEQ ID NO: 90) allows generation of a heterodimeric Fc fusion fragment which includes a single copy of the CD19 ectodomain (in analogy to the 4-1BB construct in Figure 5C). Table 43 lists the cDNA and amino acid sequences of the antigen Fc-fusion construct.

Table 43: cDNA and Amino acid sequences of monomeric human and cynomolgus CD19 antigen Fc(kih) fusion molecule

SEQ ID NO:	Antigen	Sequence
86	Nucleotide sequence Fc hole chain	see Table 32
186	Nucleotide sequence human CD19 antigen Fc knob chain avi tag	CCGAGGAACCCCTGGTCGTGAAGGTGGAAGAGGGCGACAAT GCCGTGCTGCAGTGCCTGAAGGGCACCTCCGATGGCCCTACC CAGCAGCTGACCTGGTCCAGAGAGAGCCCCCTGAAGCCCTTC CTGAAGCTGTCTCTGGGCCTGCCTGGCCTGGGCATCCATATG AGGCCTCTGGCCATCTGGCTGTTTCATCTTCAACGTGTCCCAG CAGATGGGCGGCTTCTACCTGTGTGTCAGCCTGGCCCCCATCT GAGAAGGCTTGGCAGCCTGGCTGGACCGTGAACGTGGAAGGA TCCGGCGAGCTGTTCCGGTGGAACGTGTCCGATCTGGGCGGC CTGGGATGCGGCCTGAAGAACAGATCTAGCGAGGGCCCCAGC AGCCCCAGCGGCAAACTGATGAGCCCCAAGCTGTACGTGTGG GCCAAGGACAGACCCGAGATCTGGGAGGGCGAGCCTCCTTGC CTGCCCCCTAGAGACAGCCTGAACCAGAGCCTGAGCCAGGAC

		<p>CTGACAATGGCCCCTGGCAGCACACTGTGGCTGAGCTGTGGC GTGCCACCCGACTCTGTGTCTAGAGGCCCTCTGAGCTGGACC CACGTGCACCCTAAGGGCCCTAAGAGCCTGCTGAGCCTGGAA CTGAAGGACGACAGGCCCGCCAGAGATATGTGGGTGATGGAA ACCGGCCTGCTGCTGCCTAGAGCCACAGCCCAGGATGCCGGC AAGTACTACTGCCACAGAGGCAACCTGACCATGAGCTTCCAC CTGGAAATCACCGCCAGACCCGTGCTGTGGCACTGGCTGCTG AGAACAGGCGGCTGGAAGGTCGACGCTAGCGGTGGTAGTCCG ACACCTCCGACACCCGGGGGTGGTTCTGCAGACAAAACCTCAC ACATGCCCACCGTGCCAGCACCTGAAGCCGCAGGGGGACCG TCAGTCTTCTCTTCCCCCAAAACCCAAGGACACCCTCATG ATCTCCCGGACCCCTGAGGTACATGCGTGGTGGTGGACGTG AGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGTGGAC GGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAG CAGTACAACAGCACGTACCGTGTGGTCAGCGTCCTCACCGTC CTGCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAG GTCTCCAACAAAGCCCTCGGAGCCCCATCGAGAAAACCATC TCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTGTACACC CTGCCCCCATGCCGGGATGAGCTGACCAAGAACCAGGTGAGC CTGTGGTGCCTGGTCAAAGGCTTCTATCCAGCGACATCGCC GTGGAGTGGGAGAGCAATGGGCAGCCGAGAACAACTACAAG ACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCTC TACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGG AACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAAC CACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAATCC GGAGGCCTGAACGACATCTTCGAGGCCCAGAAGATTGAATGG CACGAG</p>
90	Polypeptide sequence Fc hole chain	see Table 32
187	Polypeptide sequence human CD19 antigen Fc knob chain avi tag	<p>PEEPLVVKVEEGDNAVLQCLKGTS DGPTQQLTWSRESPLKPF LKLSLGLPLGLIHMRPLAIWLFIFNVSQQMGGFYLCQPGPPS EKAWQPGWTVNVEGSGELFRWNVSDLGGLGCLKNRSSEGPS SPSGKLMSPKLYVWAKDRPEIWEGEPPCLPPRDSL NQSLSD LTMAPGSTLWLSCGVPPDSVSRGPLSWTHVHPKGP KSLLSLE LKDDRPARDMWVMTG LLLPRATAQDAGKY YCHRG NLTMSFH LEITARPVLWHWLLRTGGWKVDASGGSPTPPTPGGGSADKTH TCPPCPAPEAAGGPSVFLFPPKPKDTLMISRTPEVTCVVVDV SHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTV LHQDWLNGKEYKCKVSNKALGAP IEKTI SKAKGQPREPQVYT LPPCRDELTKNQVSLWCLVKGFYPSDIAVEWESNGQPENNYK TTPPVLDSDGSFFLYSKLTVDKSRWQQGNV FSCSVMHEALHN HYTQKSLSLSPGKSGGLNDIFEAQKIEWHE</p>
188	Nucleotide sequence cynomolgus CD19 antigen	<p>CCCCAGGAACCCCTGGTCGTGAAGGTGGAAGAGGGCGACAAT GCCGTGCTCCAGTGCTGGAAGGCACCTCCGATGGCCCTACA CAGCAGCTCGTGTGGTGCAGAGACAGCCCCCTTCGAGCCCTTC CTGAACCTGTCTCTGGGCCTGCCTGGCATGGGCATCAGAATG GGCCCTCTGGGCATCTGGCTGCTGATCTTCAACGTGTCCAAC CAGACCGGCGGCTTCTACCTGTGTCAGCCTGGCCTGCCAAGC</p>

	Fc knob chain avi tag	GAGAAGGCTTGGCAGCCTGGATGGACCGTGTCCGTGGAAGGA TCTGGCGAGCTGTTCCGGTGGAACTGTCCGATCTGGGCGGC CTGGGATGCGGCCTGAAGAACAGAAGCAGCGAGGGCCCTAGC AGCCCCAGCGGCAAGCTGAATAGCAGCCAGCTGTACGTGTGG GCCAAGGACAGACCCGAGATGTGGGAGGGCGAGCCTGTGTGT GGCCCCCTAGAGATAGCCTGAACCAGAGCCTGAGCCAGGAC CTGACAATGGCCCCCTGGCAGCACACTGTGGCTGAGCTGTGGC GTGCCACCCGACTCTGTGTCCAGAGGCCCTCTGAGCTGGACA CACGTGCGGCCAAAGGGGCCCTAAGAGCAGCCTGCTGAGCCTG GAACTGAAGGACGACCGGCCCGACCGGGATATGTGGGTGGTG GATACAGGCCTGCTGCTGACCAGAGCCACAGCCCAGGATGCC GGCAAGTACTACTGCCACAGAGGCAACTGGACCAAGAGCTTT TACCTGGAAATCACCGCCAGACCCGCCCTGTGGCACTGGCTG CTGAGAATCGGAGGCTGGAAGGTCGACGCTAGCGGTGGTAGT CCGACACCTCCGACACCCGGGGTGGTTCTGCAGACAAACT CACACATGCCCACCGTGCCAGCACCTGAAGCCGCAGGGGA CCGTCAGTCTTCCTCTTCCCCCAAACCCAAAGGACACCCTC ATGATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGAC GTGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGTG GACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAG GAGCAGTACAACAGCACGTACCGTGTGGTCAGCGTCCTCACC GTCCTGCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGC AAGGTCTCCAACAAAGCCCTCGGAGCCCCATCGAGAAAACC ATCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTGTAC ACCCTGCCCCCATGCCGGGATGAGCTGACCAAGAACCAGGTC AGCCTGTGGTGCCTGGTCAAAGGCTTCTATCCAGCGACATC GCCGTGGAGTGGGAGAGCAATGGGCAGCCGAGAACAACTAC AAGACCACGCCTCCCGTGTGGACTCCGACGGCTCCTTCTTC CTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAG GGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCAC AACCCTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAA TCCGGAGGCCTGAACGACATCTTCGAGGCCCAGAAGATTGAA TGGCACGAG
189	Polypeptide sequence cynomolgus CD19 antigen Fc knob chain avi tag	PQEPLVVKVEEGDNAVLQCLEGTS DGPTQQLVWCRDSPFEPF LNL SLGLPGM GIRMGLGIWLLIFNVS NQTGGFYLCQPGLPS EKAWQPGWTVSVEGSGELFRWNVSDLGGLGCGLKNRSSEGPS SPSGKLNSSQLYVWAKDRPEMWE GEPVCGPPRDSL NQSLSQD LTMAPGSTLWLS CGVPPDSVSRGPLSWTHVRPKGPKSSL LSL ELKDDRPRDRM WVVD TGLLLTRATAQDAGKY YCHRGNWTKSF YLEITARPALWHWLLRIGGWKVDASGGSP TPPTPGGGSADKT HTCPPCPAPEAAGGPSVFLFP PKP KDTLMISRTPEVTCVVVD VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLT VLHQDWLNGKEYKCKVSNKALGAPI EKTISKAKGQPREPQVY TLPPCRDELTKNQVSLWCLVKGFYPSDIAVEWESNGQPENNY KTPPVLDSDGSFFLYSKLTVDKSRWQQGNV FSCSVMHEALH NHYTQKSLSLSPGKSGGLNDIFE AQKIEWHE

For the production of the monomeric antigen/Fc fusion molecules, exponentially growing suspension CHO cells were co-transfected with two plasmids encoding the two components of fusion protein (knob and hole chains) using standard methods.

5 Secreted protein was purified from cell culture supernatant by affinity chromatography using Protein A, followed by size exclusion chromatography. For affinity chromatography, the supernatant was loaded on a MabSelect Sure column volume (CV) = 5-15 mL, resin from GE Healthcare) equilibrated with Sodium Phosphate (20 mM), Sodium Citrate (20 mM), 0.5M sodium chloride buffer (pH 7.5). Unbound protein was removed by washing with at least 6 column volumes of the same buffer. The bound protein was eluted using a linear gradient; step 1,
10 10 CV from 0 to 60% elution buffer (20 mM sodium citrate, 500 mM Sodium chloride buffer (pH 2.5)); step 2, 2 CV from 60 to 100% elution buffer. For the linear gradient an additional 2 column volumes step elution with 100% elution buffer was applied.

The pH of collected fractions was adjusted by adding 1/40 (v/v) of 2M Tris, pH8.0. The protein was concentrated and filtered prior to loading on a HiLoad Superdex 200 column (GE
15 Healthcare) equilibrated with 2mM MOPS, 150 mM sodium chloride, 0.02 % (w/v) sodium azide solution of pH 7.4.

Table 44 summarizes the yield and final monomer content of monomeric human and cynomolgus CD19 antigen Fc(kih) fusion protein.

20 **Table 44 - Biochemical analysis of monomeric human and cynomolgus CD19 antigen Fc(kih) fusion protein**

Construct	Monomer [%] (SEC)	Yield [mg/l]
monomeric human CD19 Fc(kih) fusion protein	91	0.2
monomeric cynomolgus CD19 Fc(kih) fusion protein	95	3.56

Part of the purified antigen was in vitro biotinylated using the BirA biotin-protein ligase standard reaction kit (Avidity, Cat. # BirA500) according to the manufacturer's instructions. The biotinylation degree for the human CD19-containing fusion was 94 %, for the respective
25 cynomolgus CD19 construct 100%. The biotinylated protein was then used for selection, screening and characterization of affinity-matured 8B8-derived clones devoid of the deamidation hotspots N27d and N28.

7.1.2 Generation of anti-CD19 clone 8B8-018

7.1.2.1 Immunization and generation of mouse anti-human CD19 antibodies (hybridomas)

Balb/c mice were immunized six times and boosted with CD19-transfected HEK293 cells (mean receptor density 35,000 per cell). The immune response was monitored by testing serum samples with a CD19-cell-ELISA on human CD19-transfected NIH-3T3 cells. Spleen cells from mice with sufficient titers of anti-human CD19 antibody were used for immortalization by fusion with mouse myeloma cell line P3X63 Ag8.653. Three fusions were carried out and hybridoma supernatants screened by cell-ELISA on human CD19-transfected NIH-3T3 cells and FACS binding assay using Daudi (CD19+) and CD19- cells for anti-human CD19 specific antibodies (see Example 1 of WO 2011/147834).

7.1.2.2 Hybridoma screening and cell biological functional evaluation of anti-CD19 antibody

Cell-ELISA for screening antibodies against human CD19

- A cell ELISA was applied for screening of hybridomas, and to identify those hybridomas that secrete antibodies against human-CD19. NIH3T3 cells transfected with human-CD19 were used as positive cells; non-transfected NIH3T3 cells were used as negative control cells. For the assessment of the positive hybridomas the OD ratio between transfected and non-transfected NIH3T3 cells was quantified.
- Culture Medium: DMEM high glucose (4.5 mg/ml), 10 % FCS, Na-Pyruvate, NEAA, Glutamine
 - Antibodies positive control: anti CD19 monoclonal antibody (IgG1) Pharmingen Cat# 555409 c = 1 mg/ml
 - Detection antibody: Goat anti-Mouse IgG (H+L) HRP Conjugate Bio-Rad Cat# 170-06516
 - Dilution 1: 2000 in 1x ELISA Blocking Reagent
 - Other reagents: Fibronectin Roche Cat# 838039 c = 1 mg/ml
 - Glutardialdehyde: 25 % stock solution // Grade Agar Scientific #R102 final concentration: 0.05 % in PBS
 - ELISA Blocking Reagent: 10x stock solution // Roche Cat# 1112589
 - TMB substrate: Roche Cat# 11432559
 - Stop Solution: 1 M H2SO4
 - BioRad Cat# 170-6516 Dilution 1: 2000 in 1x ELISA Blocking Reagent

Day 1:

- Fibronectin coating: 5 $\mu\text{g}/\text{cm}^2$ in PBS; 96well plate = 32 cm^2 ; 160 $\mu\text{g}/\text{plate}$ in 6 ml
- PBS, 50 $\mu\text{l}/\text{well}$
- incubate 45 min at RT, aspirate coating solution
- 5 - Seed 1.25×10^4 cells/well in 50 μl culture medium in a 96well plate
- incubate 40 hours at 37 °C
- add to upper half of the plate: NIH3T3 cells expressing CD19
- add to lower half of the plate: non-transfected NIH3T3 cells

Day 3:

- 10 - Addition of positive control antibody or samples (supernatant or mouse serum) in 50 μl culture medium
- incubate for 2 h at 4 °C
- Remove medium, fix cells with 100 μl Glutardialdehyde (0.05 % in PBS)
- Wash two times with 200 μl PBS
- 15 - Addition of detection antibody 1:2000, 50 $\mu\text{l}/\text{well}$
- incubate 2 h at RT
- wash three times with 200 μl PBS
- add 50 μl TMB, incubate for 30 min. at RT,
- stop by addition of 25 μl 1 M H_2SO_4 ; read extinction at 450nm/620nm
- 20 - Calculation of results: ratio OD NIH3T3 CD19 : OD NIH3T3 non-transfected

The selected antibody demonstrated specific binding to CD19 transfected NIH3T3 cells as compared to untransfected NIH3T3 cells (see Example 2 of WO 2011/147834).

7.1.2.3 Humanization of anti-CD19 antibody

- 25 The CD19 binding specificity of the murine antibody was transferred onto a human acceptor framework to eliminate potential immunogenicity issues arising from sequence stretches that the human body will recognize as foreign. This was done by engrafting the entire complementary determining regions (CDR) of the murine (donor) antibody onto a human (acceptor) antibody framework, and is called CDR-grafting or antibody humanization.

- 30 The murine amino acid sequence was aligned with a collection of human germ-line antibody V genes, and sorted according to sequence identity and homology. Before selecting one particular acceptor sequence, the so-called canonical loop structures of the donor antibody have to be determined (Morea, V., et al., Methods, Vol 20, Issue 3 (2000) 267-279). These canonical

loop structures are determined by the type of residues present at the so-called canonical positions. These positions lie (partially) outside of the CDR regions, and have to be kept functionally equivalent in the final construct in order to retain the CDR conformation of the parental (donor) antibody. The human germ-line sequence VBASE_VH1_1 was chosen as the acceptor for the heavy chain and sequence VBASE_VK2_5 was chosen for the light chain.

7.1.2.4 Removal of deamidation hotspots

It has been found that the wild-type humanized anti-human CD19 antibody has three deamidation hotspots in the HVR-L1: **NSNGNT** (SEQ ID NO: 190). Additionally it has been found that in the HVR-H2 a further deamidation hotspot is present: **KFNG** (SEQ ID NO: 191).

To address the deamidation hotspot in the HVR-H2 an N (Asn) to Q (Gln) point mutation at position 64 (numbering according to Kabat) has been introduced. Thus, the antibody as reported herein has a HVR-H2 comprising the amino acid sequence **TEKFQGRVTM** (SEQ ID NO: 192).

To address the deamidation hotspots in the light chain and to obtain a humanized anti-human CD19 antibody with improved deamidation stability individual mutations at Kabat position 27d, 27e, 28 and 29 and a double mutation at positions 27e and 28 (numbering according to Kabat) were introduced. In total 9 variants (var.1 to var.9) of the wild-type humanized antibody (var.0) have been generated (see Table 45).

Table 45: Variants of humanized wild-type CD19 antibody

Kabat position	2222	Kabat position	6
LC:	7789	HC:	4
de			
var.0:wt	QSLNSNGNTYLNW		TEKFNGKATM
var.1:N27dH	QSLE H SNGNTYLNW		TEKF Q GRVTM
var.2:N27dQ	QSLE Q SNGNTYLNW		TEKF Q GRVTM
var.3:S27eA	QSLN A NGNTYLNW		TEKF Q GRVTM
var.4:S27eV	QSLN V NGNTYLNW		TEKF Q GRVTM
var.5:S27eP	QSLN P NGNTYLNW		TEKF Q GRVTM
var.6:N28Q	QSLNS Q GNTYLNW		TEKF Q GRVTM
var.7:G29A	QSLNSN A NTYLNW		TEKF Q GRVTM
var.8:G29V	QSLNSN V NTYLNW		TEKF Q GRVTM
var.9:S27eP/N28S	QSLN P SNGNTYLNW		TEKF Q GRVTM

variant →	0	1	2	3	4	5	6	7	8	9
↓parameter										
K _D (BIAcore) [nM]	5	250	136	2	1	6	54	4	16	45
t _{1/2} [min]	-	0.1	1.1	105.2	191.5	43.6	4.4	51.5	17.6	4

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variant →	0	1	2	3	4	5	6	7	8	9
↓parameter										
human CD19 binding after pH 7.4 incubation [%]	46	0	75	84	85	95	91	72	83	83
human CD19 binding after pH 6.0 incubation [%]	90	0	95	95	97	99	97	86	91	87
SEC main peak after incubation [%]	> 95	> 95	> 95	> 95	> 95	> 95	> 95	> 95	> 95	-

It has been found that with a single mutation at position 27e according to Kabat from S (serine) to P (proline) all deamidation hotspots in the HVR-L1 can be addressed. This is a mutation not of the deamidation prone N (asparagine) residue but of a neighboring residue.

- 5 Thus, the antibody as reported herein has a HVR-L1 comprising the amino acid sequence LENPNGNT (SEQ ID NO: 193). In one embodiment the humanized anti-human CD19 antibody comprises a HVR-L1 that has the amino acid sequence LENPSGNT (SEQ ID NO: 194).

Additionally these antibodies maintain the cross-reactivity to cynomolgus CD19 as shown in the following **Table 46**.

EC50 [μg/ml]	var.0	var.5	var.9
huCD19 ECD	0.087	0.084	0.089
cyCD19 ECD	0.313	0.255	0.435

10

- The wild-type humanized anti-human CD19 antibody (var.0) shows after purification approx. 7.5 % deamidation. After storage for two weeks at pH 7.4 the amount of deamidated antibody is increased to approx. 18.5 %. The variant antibody with an S27eP mutation (var.5) shows approx. 2% deamidation and 2% succinimide formation after purification. During storage at pH 7.4 for two weeks only approx. 7.5 % deamidated antibody is present. Var. 5 is named clone 8B8-018 and was elected for the preparation of CD19-targeted TNF family ligand trimer-containing antigen binding molecules.
- 15

7.1.3 Preparation of monovalent CD19(8B8-018) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains with charged residues (Construct 3.1)

A polypeptide containing two ectodomains of 4-1BB ligand (71-254), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned as depicted in Figure 29A: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL. A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the human IgG1-CH domain, was cloned as described in Figure 29B: human 4-1BB ligand, (G4S)₂ connector, human CH.

The polypeptide encoding the dimeric 4-1BB ligand fused to human CL domain was subcloned in frame with the human IgG1 heavy chain CH2 and CH3 domains on the knob (Merchant, Zhu et al. 1998). To improve correct pairing the following mutations have been introduced in the crossed CH-CL. In the dimeric 4-1BB ligand fused to human CL, E123R and Q124K. In the monomeric 4-1BB ligand fused to human CH1, K147E and K213E.

The variable region of heavy and light chain DNA sequences encoding a binder specific for CD19, **clone 8B8-018**, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831.

Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-CD19-Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations and the anti-CD19 light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a CD19 binding Fab (**Figure 30**, Construct 3.1).

Table 47 shows the cDNA and amino acid sequences of the monovalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule with crossed CH-CL and charged residues (construct 3.1).

Table 47: cDNA and amino acid sequences of monovalent CD19 (8B8-018) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion containing CH-CL cross with charged residues (construct 3.1). * for charged residues

SEQ ID NO:	Description	Sequence
129	Nucleotide sequence Dimeric hu 4-1BBL (71-	see Table 3

	254) – CL* Fc knob chain	
130	Nucleotide sequence Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
203	Nucleotide sequence anti-CD19(8B8-018) Fc hole chain	<p>CAGGTCCAGCTGGTGCAGTCCGGCGCCGAGGTCAAGAA ACCCGGGGCTTCTGTGAAGGTTTCATGCAAGGCAAGCG GATACACCTTCACCGACTATATCATGCATTGGGTCAGGC AGGCCCTGGCCAAGGTCTCGAATGGATGGGCTACATTA ACCCATATAATGATGGCTCCAAATACACCGAGAAGTTTC AGGGAAGAGTCACTATGACATCTGACACCAGTATCAGC ACTGCTTACATGGAGCTGTCCCGCCTTCGGTCTGATGAC ACCGCAGTGTATTACTGTGCCAGGGGCACATATTACTAC GGCTCAGCTCTGTTCGACTATTGGGGGCAGGGAACCACA GTAACCGTGAGCTCCGCTAGCACCAAGGGCCCCTCCGTG TTCCCCCTGGCCCCCAGCAGCAAGAGCACCAGCGGCGG CACAGCCGCTCTGGGCTGCCTGGTCAAGGACTACTTCCC CGAGCCCGTGACCGTGTCTTGGAACAGCGGAGCCCTGA CCTCCGGCGTGACACCTTCCCCGCCGTGCTGCAGAGTT CTGGCCTGTATAGCCTGAGCAGCGTGGTCACCGTGCCTT CTAGCAGCCTGGGACCCAGACCTACATCTGCAACGTG AACCACAAGCCCAGCAACACCAAGGTGGACAAGAAGGT GGAGCCCAAGAGCTGCGACAAAACCTCACACATGCCAC CGTGCCAGCACCTGAAGCTGCAGGGGGACCGTCAGTC TTCCTCTTCCCCCAAAACCCAAGGACACCCTCATGATC TCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTG AGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGT GGACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGC GGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGC GTCCTCACCGTCTCTGCACCAGGACTGGCTGAATGGCAAG GAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGGCGC CCCCATCGAGAAAACCATCTCCAAGGCCAAAGGGCAGC CCCGAGAACCACAGGTGTGCACCCTGCCCCATCCCGG GATGAGCTGACCAAGAACCAGGTCAGCCTCTCGTGCGC AGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTG GGAGAGCAATGGGCAGCCGGAGAACAACCTACAAGACC ACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCCTC GTGAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCA GGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCT GCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCC GGGTAAA</p>
204	Nucleotide sequence anti-CD19(8B8-018) light chain	<p>GACATCGTCATGACCCAGACACCCCTGTCCCTCTCTGTG ACCCCTGGCCAGCCAGCCTCAATTAGCTGCAAGTCCTCT CAAAGTCTGGAGAACCCCAATGGGAACACTTACCTTAAT TGGTATCTGCAGAAACCCGGACAATCCCCCTCAACTCCTG ATCTACAGGGTCTCTAAGAGATTCTCAGGCGTGCCAGAT CGCTTTAGCGGTTCCGGGTCTGGCACAGACTTCACCTTG AAGATTAGTCGGGTTGAAGCTGAGGATGTGGGAGTCTA TACTGTCTGCAGCTCACTCATGTGCCCTACACCTTTGGT CAGGGCACAAAACCTGGAGATCAAGCGGACCGTGGCCGC</p>

		TCCCTCCGTGTTTCATCTTCCCACCCTCCGACGAGCAGCT GAAGTCCGGCACCGCCAGCGTGGTGTGCCTGCTGAACA ACTTCTACCCCCGCGAGGCCAAGGTGCAGTGGAAGGTG GACAACGCCCTGCAGTCCGGCAACTCCCAGGAATCCGT GACCGAGCAGGACTCCAAGGACAGCACCTACTCCCTGT CCTCCACCCTGACCCTGTCCAAGGCCGACTACGAGAAGC ACAAGGTGTACGCCTGCGAAGTGACCCACCAGGGCCTG TCCAGCCCCGTGACCAAGTCCTTCAACCGGGGCGAGTGC
115	Dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 3
116	Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
205	anti-CD19(8B8-018) Fc hole chain	QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYIMHWVRQ APGQGLEWMGYINPYNDGSKYTEKFQGRVTMTSDTSISTA YMELSRLRSDDTAVYYCARGTYYYGSALFDYWQGQTTVT VSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVT VSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVTPSSSLGTQ TYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAG GPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNW YVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLN GKEYKCKVSNKALGAPIEKTISKAKGQPREPQVCTLPSPRD ELTKNQVSLSCAVKGFYPSDIAVEWESNGQPENNYKTPP VLDSGDSFFLVSKLTVDKSRWQQGNVFSVSMHEALHNH YTQKSLSLSPGK
206	anti-CD19(8B8-018) light chain	DIVMTQTPLSLSVTPGQPASISCKSSQSLENPNNGNTYLNWY LQKPGQSPQLLIYRVSKRFSGVDPDRFSGSGSGTDFTLKISR EAEDVGVYYCLQLTHVPYTFGQGTKLEIKRTVAAPSVFIFP PSDEQLKSGTASVCLLNFPYPREAKVQWKVDNALQSGN SQESVTEQDSKSTYLSSTLTLSKADYEKHKVYACEVTH QGLSSPVTKSFNRGEC

7.1.4 Preparation of monovalent CD19(8B8-018) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains without charged residues (Construct 3.2)

- 5 A polypeptide containing two ectodomains of 4-1BB ligand (71-254), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned in analogy as depicted in Figure (29A), but without amino acid mutations in the CL domain: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL. A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the human IgG1-CH1 domain, was cloned in analogy as depicted in Figure (29B), but without amino acid mutations in the CH1 domain:
- 10 human 4-1BB ligand, (G4S)₂ connector, human CH1.

The variable region of heavy and light chain DNA sequences encoding a binder specific for CD19, clone 8B8-018, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1.

The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the
 5 constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831. Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-CD19-Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations and the anti-
 10 CD19 light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a CD19-binding Fab (Figure 30, Construct 3.2).

Table 48 shows the cDNA and amino acid sequences of the monovalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule containing crossed CH-CL cross without charged residues (construct 3.2).

**Table 48: cDNA and amino acid sequences of monovalent CD19(8B8-018) targeted split
 15 trimeric 4-1BB ligand (71-254) Fc (kih) fusion containing CH-CL cross without charged residues (construct 3.2).**

SEQ ID NO:	Description	Sequence
165	Nucleotide sequence dimeric ligand (71-254)–CL Fc knob chain	see Table 22
166	Nucleotide sequence monomeric hu 4-1BBL (71-254) – CH1	see Table 22
203	Nucleotide sequence anti-CD19(8B8-018) Fc hole chain	see Table 47
204	Nucleotide sequence anti-CD19(8B8-018) light chain	see Table 47
117	Dimeric ligand (71-254) - CL Fc knob chain	see Table 22
118	Monomeric ligand (71-254) -	see Table 22

	CH1	
205	anti-CD19(8B8-018) Fc hole chain	see Table 47
206	anti-CD19(8B8-018) light chain	see Table 47

7.1.5 Preparation of bivalent CD19(8B8-018) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding (Construct 3.3)

A polypeptide containing two ectodomains of 4-1BB ligand (71-254), separated by (G4S)₂ linkers was fused to the C-terminus of human IgG1 Fc hole chain, as depicted in Figure 29C: human IgG1 Fc hole, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand. A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the C-terminus of human IgG1 Fc knob chain as described in Figure 29D: human IgG1 Fc knob, (G4S)₂ connector, human 4-1BB ligand.

The variable region of heavy and light chain DNA sequences encoding a binder specific for CD19, clone 8B8-018, were subcloned in frame with either the constant heavy chain of the hole, the knob or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831. Combination of the anti-CD19 huIgG1 hole dimeric ligand chain containing the Y349C/T366S/L368A/Y407V mutations, the anti-CD19 huIgG1 knob monomeric ligand chain containing the S354C/T366W mutations and the anti-CD19 light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and two CD19 binding Fabs (Figure 30, construct 3.3).

Table 49 shows the cDNA and amino acid sequences of the bivalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule (construct 3.3).

Table 49: Base pair sequences of bivalent CD19(8B8-018) targeted split trimeric 4-1BB ligand Fc (kih) PGLALA fusion (construct 3.3)

SEQ ID NO:	Description	Sequence
207	Nucleotide sequence anti-CD19(8B8-018) Fc hole dimeric	CAGGTCCAGCTGGTGCAGTCCGGCGCCGAGGTCAAGAA ACCCGGGGCTTCTGTGAAGGTTTCATGCAAGGCAAGCG GATACACCTTCACCGACTATATCATGCATTGGGTCAGGC AGGCCCTGGCCAAGGTCTCGAATGGATGGGCTACATTA ACCCATATAATGATGGCTCCAAATACACCGAGAAGTTTC

	ligand chain	AGGGAAGAGTCACTATGACATCTGACACCAGTATCAGC ACTGCTTACATGGAGCTGTCCCGCCTTCGGTCTGATGAC ACCGCAGTGTATTACTGTGCCAGGGGCACATATTACTAC GGCTCAGCTCTGTTCGACTATTGGGGGCAGGGAACCACA GTAACCGTGAGCTCCGCTAGCACCAAGGGCCCCCTCCGTG TTCCCCCTGGCCCCCAGCAGCAAGAGCACCAGCGGCGG CACAGCCGCTCTGGGCTGCCTGGTCAAGGACTACTTCCC CGAGCCCCGTGACCGTGTCTTGAACAGCGGAGCCCTGA CCTCCGGCGTGCACACCTTCCCCGCCGTGCTGCAGAGTT CTGGCCTGTATAGCCTGAGCAGCGTGGTCACCGTGCCTT CTAGCAGCCTGGGACCCAGACCTACATCTGCAACGTG AACCACAAGCCCAGCAACACCAAGGTGGACAAGAAGGT GGAGCCCAAGAGCTGCGACAAAACCTCACACATGCCAC CGTGCCAGCACCTGAAGCTGCAGGGGGACCGTCAGTC TTCCTCTTCCCCCAAAACCCAAGGACACCCTCATGATC TCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTG AGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGT GGACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGC GGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGC GTCCTCACCGTCTGCACCAGGACTGGCTGAATGGCAAG GAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGGCGC CCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCAGC CCCGAGAACCACAGGTGTGCACCCTGCCCCATCCCGG GATGAGCTGACCAAGAACCAGGTGAGCCTCTCGTGCGC AGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTG GGAGAGCAATGGGCAGCCGGAGAACAACCTACAAGACC ACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCTC GTGAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCA GGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCT GCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCC GGGTGGAGGCGGCGGAAGCGGAGGAGGAGGATCCAGA GAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGGACT GCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGTGGC CCAGAACGTGCTGCTGATCGATGGCCCCCTGTCTTGTA CAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGGCG GCCTGAGCTACAAAGAGGACACCAAAAGAACTGGTGGTG GCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGGAA CTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCTGT GTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCTGC TGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCTCC TGCCAGCTCCGAGGCCCGGAATAGCGCATTTGGGTTTCA AGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCTGG GAGTGCATCTGCACACAGAGGCCAGGGCTAGACACGCC TGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCTGTTC AGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCTCCA AGAAGCGAAGGCGGAGGCGGATCTGGCGGCGGAGGATC TAGAGAGGGACCCGAACCTGTCCCCTGACGATCCAGCCG GGCTGCTGGATCTGAGACAGGGAATGTTCCGCCAGCTG GTGGCTCAGAATGTGCTGCTGATTGACGGACCTCTGAGC TGGTACTCCGACCCAGGGCTGGCAGGGGTGTCCCTGACT GGGGGACTGTCTACAAAGAAGATACAAAAGAACTGGT GGTGGCTAAAGCTGGGGTGTACTATGTGTTTTTTCAGCT GGAACCTGAGGCGGGTGGTGGCTGGGGAGGGCTCAGGAT CTGTGTCCCTGGCTCTGCATCTGCAGCCACTGCGCTCTG
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		CTGCTGGCGCAGCTGCACTGGCTCTGACTGTGGACCTGC CACCAGCCTCTAGCGAGGCCAGAAACAGCGCCTTCGGG TTCCAAGGACGCCTGCTGCATCTGAGCGCCGGACAGCG CCTGGGAGTGCATCTGCATACTGAAGCCAGAGCCCCGGC ATGCTTGGCAGCTGACTCAGGGGGCAACTGTGCTGGGA CTGTTTCGCGTGACACCTGAGATCCCTGCCGGACTGCCA AGCCCTAGATCAGAA
208	Nucleotide sequence anti- CD19(8B8-018) Fc knob monomeric ligand	CAGGTCCAGCTGGTGCAGTCCGGCGCCGAGGTCAAGAA ACCCGGGGCTTCTGTGAAGGTTTCATGCAAGGCAAGCG GATACACCTTCACCGACTATATCATGCATTGGGTACAGGC AGGCCCTGGCCAAGGTCTCGAATGGATGGGCTACATTA ACCCATATAATGATGGCTCCAAATACACCGAGAAGTTTC AGGGAAGAGTCACTATGACATCTGACACCAGTATCAGC ACTGCTTACATGGAGCTGTCCCGCCTTCGGTCTGATGAC ACCGCAGTGTATTACTGTGCCAGGGGGCACATATTACTAC GGCTCAGCTCTGTTCGACTATTGGGGGCAGGGAACCACA GTAACCGTGAGCTCCGCTAGCACCAAGGGCCCATCGGT CTTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGGGGG CACAGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCC CGAACCGGTGACGGTGTCTGTGGAACCTCAGGCGCCCTGA CCAGCGGCGTGCACACCTTCCCGGCTGTCTACAGTCCT CAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCCT CCAGCAGCTTGGGACCCAGACCTACATCTGCAACGTG AATCACAAGCCAGCAACACCAAGGTGGACAAGAAAGT TGAGCCCAAATCTTGTGACAAAACCTCACACATGCCACC GTGCCCAGCACCTGAAGCTGCAGGGGGACCGTCAGTCT TCCTCTTCCCCCAAACCCAAAGGACACCCTCATGATCT CCCGGACCCCTGAGGTACATGCGTGGTGGTGGACGTG AGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGT GGACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGC GGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGC GTCTCACCCTCCTGCACCAGGACTGGCTGAATGGCAAG GAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGGCGC CCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCAGC CCCGAGAACCACAGGTGTACACCCTGCCCCCTGCAGA GATGAGCTGACCAAGAACCAGGTGTCCCTGTGGTGTCTG GTCAAGGGCTTCTACCCCAGCGATATCGCCGTGGAGTGG GAGAGCAACGGCCAGCCTGAGAACAACCTACAAGACCAC CCCCCTGTGCTGGACAGCGACGGCAGCTTCTTCCTGTA CTCCAAACTGACCGTGGACAAGAGCCGGTGGCAGCAGG GCAACGTGTTTACGCTGCAGCGTGATGCACGAGGCCCTG CACAACCACTACACCCAGAAGTCCCTGAGCCTGAGCCC CGGCGGAGGCGGCGGAAGCGGAGGAGGAGGATCCAGA GAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGGACT GCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGTGGC CCAGAACGTGCTGCTGATCGATGGCCCCCTGTCTGGTA CAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGGCG GCCTGAGCTACAAAGAGGACACCAAAGAACTGGTGGTG GCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGGAA CTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCTGT GTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCTGC TGGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCTCC TGCCAGCTCCGAGGCCCGGAATAGCGCATTTGGGTTTCA AGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCTGG

		GAGTGCATCTGCACACAGAGGCCAGGGCTAGACACGCC TGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCTGTTC AGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCTCCA AGAAGCGAA
204	Nucleotide sequence anti- CD19(8B8-018) light chain	see Table 47
209	anti-CD19(8B8- 018) Fc hole dimeric ligand chain	QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYIMHWVRQ APGQGLEWMGYINPYNDGSKYTEKFQGRVTMTSDTSISTA YMELSRLRSDDTAVYYCARGTYYYGSALFDYWQGTTVT VSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVT VSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQ TYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAG GPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNW YVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLN GKEYKCKVSNKALGAPIEKTISKAKGQPREPQVCTLPSPRD ELTKNQVSLSCAVKGFYPSDIAVEWESNGQPENNYKTPP VLDSGDSFFLVSKLTVDKSRWQQGNVFSCSVMHEALHNH YTQKSLSLSPGGGGGSGGGGSREGPELSPDDPAGLLDLRQ GMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKED TKELVVAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQP LRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAG QRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLP SPRSEGGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQL VAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVV AKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAG AAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVH LHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSPRSE
210	anti-CD19(8B8- 018) Fc knob monomeric ligand	QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYIMHWVRQ APGQGLEWMGYINPYNDGSKYTEKFQGRVTMTSDTSISTA YMELSRLRSDDTAVYYCARGTYYYGSALFDYWQGTTVT VSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVT VSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQ TYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAG GPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNW YVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLN GKEYKCKVSNKALGAPIEKTISKAKGQPREPQVYTLPPCRD ELTKNQVSLWCLVKGFYPSDIAVEWESNGQPENNYKTPP VLDSGDSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNH YTQKSLSLSPGGGGGSGGGGSREGPELSPDDPAGLLDLRQ GMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKED TKELVVAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQP LRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAG QRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLP SPRSE
206	anti-CD19(8B8- 018) light chain	see Table 47

7.1.6 Preparation of monovalent CD19(8B8-018) targeted 4-1BB ligand (71-248) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains with charged residues (Construct 3.4)

A polypeptide containing two ectodomains of 4-1BB ligand (71-248), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned in analogy to the one depicted in Figure 29A: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL. A polypeptide containing one ectodomain of 4-1BB ligand (71-248) and fused to the human IgG1-CH domain, was cloned in analogy to the one described in Figure 29B: human 4-1BB ligand, (G4S)₂ connector, human CH.

The polypeptide encoding the dimeric 4-1BB ligand fused to human CL domain was subcloned in frame with the human IgG1 heavy chain CH2 and CH3 domains on the knob (Merchant, Zhu et al. 1998). To improve correct pairing the following mutations have been introduced in the crossed CH-CL. In the dimeric 4-1BB ligand fused to human CL, E123R and Q124K. In the monomeric 4-1BB ligand fused to human CH1, K147E and K213E.

The variable region of heavy and light chain DNA sequences encoding a binder specific for CD19, clone 8B8-018, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831. Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-CD19-Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations and the anti-CD19 light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a CD19 binding Fab (Figure 30, construct 3.4).

Table 50 shows the cDNA and amino acid sequences of the monovalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion antigen binding molecule with crossed CH-CL and charged residues (construct 3.4).

Table 50: cDNA and amino acid sequences of monovalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion containing CH-CL cross with charged residues (construct 3.4). * charged residues

SEQ ID NO:	Description	Sequence
169	Nucleotide sequence dimeric ligand (71-248) – CL* Fc knob	see Table 24

	chain	
170	Nucleotide sequence monomeric hu 4-1BBL (71-248) – CH1*	see Table 24
203	Nucleotide sequence anti-CD19(8B8-018) Fc hole chain	see Table 47
204	Nucleotide sequence anti-CD19(8B8-018) light chain	see Table 47
119	Dimeric ligand (71-248) – CL* Fc knob chain	see Table 24
120	Monomeric ligand (71-248)-CH1*	see Table 24
205	anti-CD19(8B8-018) Fc hole chain	see Table 47
206	anti-CD19(8B8-018) light chain	see Table 47

7.1.7 Preparation of monovalent CD19(8B8-018) targeted 4-1BB ligand (71-248) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains without charged residues (Construct 3.5)

- 5 A polypeptide containing two ectodomains of 4-1BB ligand (71-248), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned in analogy as depicted in Figure (29A), but without amino acid mutations in the CL domain: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL. A polypeptide containing one ectodomain of 4-1BB ligand (71-248) and fused to the human IgG1-CH1 domain, was cloned in
10 analogy as depicted in Figure (29B), but without amino acid mutations in the CH1 domain: human 4-1BB ligand, (G4S)₂ connector, human CH1.

The variable region of heavy and light chain DNA sequences encoding a binder specific for CD19, clone 8B8-018, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala
15 mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831.

Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-CD19-Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations and the anti-CD19 light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a CD19-binding Fab

5 (Figure 30, Construct 3.5).

Table 51 shows the cDNA and amino acid sequences of the monovalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion antigen binding molecule containing crossed CH-CL cross without charged residues (construct 3.5).

10 **Table 51: cDNA and amino acid sequences of monovalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion containing CH-CL cross without charged residues (construct 3.5).**

SEQ ID NO:	Description	Sequence
171	Nucleotide sequence dimeric ligand (71-248) – CL Fc knob chain	see Table 25
172	Nucleotide sequence monomeric ligand (71-248)-CH1	see Table 25
203	Nucleotide sequence anti-CD19(8B8-018) Fc hole chain	see Table 47
204	Nucleotide sequence anti-CD19(8B8-018) light chain	see Table 47
173	Dimeric ligand (71-248) – CL Fc knob chain	see Table 25
174	Monomeric ligand (71-248)-CH1	see Table 25
205	anti-CD19(8B8-018) Fc hole chain	see Table 47
206	anti-CD19(8B8-018) light chain	see Table 47

7.1.8 Preparation of bivalent CD19(8B8-018) targeted 4-1BB ligand (71-248) trimer-containing Fc (kih) fusion antigen binding (Construct 3.6)

A polypeptide containing two ectodomains of 4-1BB ligand (71-248), separated by (G4S)₂ linkers was fused to the C-terminus of human IgG1 Fc hole chain, as depicted in **Figure 29C**:

- 5 human IgG1 Fc hole, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand. A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the C-terminus of human IgG1 Fc knob chain as described in **Figure 29D**: human IgG1 Fc knob, (G4S)₂ connector, human 4-1BB ligand.

- The variable region of heavy and light chain DNA sequences encoding a binder specific for CD19, clone 8B8-018, were subcloned in frame with either the constant heavy chain of the hole, the knob or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831. Combination of the anti-CD19 huIgG1 hole dimeric ligand chain containing the Y349C/T366S/L368A/Y407V mutations, the anti-CD19 huIgG1 knob monomeric ligand chain containing the S354C/T366W mutations and the anti-CD19 light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and two CD19 binding Fabs (Figure 30, construct 3.6).

- Table 52 shows the cDNA and amino acid sequences of the bivalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion antigen binding molecule (construct 3.6).

Table 52: cDNA and amino acid sequences of bivalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion (construct 3.6)

SEQ ID NO:	Description	Sequence
211	Nucleotide sequence anti-CD19(8B8-018) Fc hole dimeric ligand (71-248) chain	CAGGTCCAGCTGGTGCAGTCCGGCGCCGAGGTCAAGAA ACCCGGGGCTTCTGTGAAGGTTTCATGCAAGGCAAGCG GATACACCTTCACCGACTATATCATGCATTGGGTCAGGC AGGCCCTGGCCAAGGTCTCGAATGGATGGGCTACATTA ACCCATATAATGATGGCTCCAAATACACCGAGAAGTTTC AGGGAAGAGTCACTATGACATCTGACACCAGTATCAGC ACTGCTTACATGGAGCTGTCCCGCCTTCGGTCTGATGAC ACCGCAGTGTATTACTGTGCCAGGGGACATATTACTAC GGCTCAGCTCTGTTCGACTATTGGGGGCAGGGAACCACA GTAACCGTGAGCTCCGCTAGCACCAAGGGCCCCCTCCGTG TCCCCCTGGCCCCCAGCAGCAAGAGCACCAGCGGCGG CACAGCCGCTCTGGGCTGCCTGGTCAAGGACTACTTCCC CGAGCCCGTGACCGTGTCTGGAACAGCGGAGCCCTGA CCTCCGGCGTGACACCTTCCCCGCCGTGCTGCAGAGTT CTGGCCTGTATAGCCTGAGCAGCGTGGTCACCGTGCCTT

		CTAGCAGCCTGGGACCCAGACCTACATCTGCAACGTG AACCACAAGCCCAGCAACACCAAGGTGGACAAGAAGGT GGAGCCCAAGAGCTGCGACAAAACCTCACACATGCCAC CGTGCCCAAGCACCTGAAGCTGCAGGGGGACCGTCAGTC TTCCTCTTCCCCCAAAACCCAAGGACACCCTCATGATC TCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTG AGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGT GGACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGC GGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGC GTCCTCACCGTCTTGCACCAGGACTGGCTGAATGGCAAG GAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGGCGC CCCCATCGAGAAAACCATCTCCAAGGCCAAAGGGCAGC CCCGAGAACCACAGGTGTGCACCCTGCCCCATCCCGG GATGAGCTGACCAAGAACCAGGTGACGCTCTCGTGCGC AGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTG GGAGAGCAATGGGCAGCCGGAGAACAACCTACAAGACC ACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCCTC GTGAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCA GGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCT GCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCC GGGTGGAGGCGGCGGAAGCGGAGGAGGAGGATCCAGA GAGGGCCCTGAGCTGAGCCCTGATGATCCTGCCGACT GCTGGACCTGCGGCAGGGAATGTTTGCCAGCTGGTGGC CCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTGGTA CAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGGCG GCCTGAGCTACAAAGAGGACACCAAAAGAACTGGTGGTG GCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGGAA CTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCTGT GTCTCTGGCCCTGCATCTGCAGCCTCTGAGATCTGCTGC TGGCGCCGCTGCTCTGGCACTGACAGTGGATCTGCCTCC TGCCAGCAGCGAGGCCCGGAATAGCGCATTGGGTTTCA AGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCTGG GAGTGCATCTGCACACAGAGGCCAGGGCTAGACACGCC TGGCAGCTGACACAGGGCGCTACAGTGTGGGCTGTTC AGAGTGACCCCCGAGATTCCAGCAGGCCTGGGAGGCGG CGGATCTGGCGGCGGAGGATCTAGAGAAGGACCCGAGC TGTCCCCCGACGATCCCGCTGGGCTGCTGGATCTGAGAC AGGGCATGTTTCGCTCAGCTGGTGGCTCAGAATGTGCTGC TGATTGACGGACCTCTGAGCTGGTACTCCGACCCAGGGC TGGCAGGGGTGTCCCTGACTGGGGGACTGTCTACAAA GAAGATACAAAAGAACTGGTGGTGGCTAAAGCTGGGGT GTACTATGTGTTTTTTCAGCTGGAAGTGGGCGGGTGGT GGCTGGGGAGGGCTCAGGATCTGTGTCCCTGGCTCTGCA TCTGCAGCCACTGCGCTCTGCAGCAGGGGCTGCAGCACT GGCCCTGACTGTGGACCTGCCCCAGCTTCTTCCGAGGC CAGAAACAGCGCCTTCGGGTTCCAAGGACGCCTGCTGC ATCTGAGCGCCGGACAGCGCCTGGGAGTGCATCTGCAT ACTGAAGCCAGAGCCCGGCATGCTTGGCAGCTGACTCA GGGGGCAACTGTGCTGGGACTGTTTCGCGTGACACCTGA GATCCCAGCCGGGCTC
212	Nucleotide sequence anti- CD19(8B8-018)	CAGGTCCAGCTGGTGCAGTCCGGCGCCGAGGTCAAGAA ACCCGGGGCTTCTGTGAAGGTTTCATGCAAGGCAAGCG GATACACCTTCACCGACTATATCATGCATTGGGTGAGGC AGGCCCTGGCCAAGGTCTCGAATGGATGGGCTACATTA

	Fc knob monomeric (71-248) ligand	ACCCATATAATGATGGCTCCAAATACACCGAGAAGTTTC AGGGAAGAGTCACTATGACATCTGACACCAGTATCAGC ACTGCTTACATGGAGCTGTCCCGCCTTCGGTCTGATGAC ACCGCAGTGTATTACTGTGCCAGGGGCACATATTACTAC GGCTCAGCTCTGTTCGACTATTGGGGGCAGGGAACCACA GTAACCGTGAGCTCCGCTAGCACCAAGGGCCCATCGGT CTTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGGGGG CACAGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCC CGAACCGGTGACGGTGTCGTGGAACCTCAGGCGCCCTGA CCAGCGGCGTGACACACCTTCCCGGCTGTCCTACAGTCCT CAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCCT CCAGCAGCTTGGGACCCAGACCTACATCTGCAACGTG AATCACAAGCCCAGCAACACCAAGGTGGACAAGAAAGT TGAGCCCCAAATCTTGTGACAAAACCTCACACATGCCACC GTGCCACGACCTGAAGCTGCAGGGGGACCGTCAGTCT TCCTCTTCCCCCAAAACCCAAGGACACCCTCATGATCT CCCGGACCCCTGAGGTACATGCGTGGTGGTGGACGTG AGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGT GGACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGC GGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGC GTCCTCACCGTCTGTCACCAGGACTGGCTGAATGGCAAG GAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGGCGC CCCCATCGAGAAAACCATCTCCAAGGCCAAAGGGCAGC CCCGAGAACCACAGGTGTACACCCTGCCCCCTGCAGA GATGAGCTGACCAAGAACCAGGTGTCCCTGTGGTGTCTG GTCAAGGGCTTCTACCCCAGCGATATCGCCGTGGAGTGG GAGAGCAACGGCCAGCCTGAGAACAACCTACAAGACCAC CCCCCTGTGCTGGACAGCGACGGCAGCTTCTTCTCTGTA CTCCAAACTGACCGTGGACAAGAGCCGGTGGCAGCAGG GCAACGTGTTTCACTGTCAGCGTGATGCACGAGGCCCTG CACAACCACTACACCCAGAAGTCCCTGAGCCTGAGCCC CGGCGGAGGCGGCGGAAGCGGAGGAGGAGGATCCAGA GAGGGCCCTGAGCTGAGCCCTGATGATCCTGCCGGACT GCTGGACCTGCGGCAGGGAATGTTTGCCAGCTGGTGGC CCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTGGTA CAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGGCG GCCTGAGCTACAAAGAGGACACCAAGAAGTGGTGGTG GCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGGAA CTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCTGT GTCTCTGGCCCTGCATCTGCAGCCTCTGAGATCTGCTGC TGGCGCCGCTGCTCTGGCACTGACAGTGGATCTGCCTCC TGCCAGCAGCGAGGCCCGGAATAGCGCATTTGGGTTTCA AGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCTGG GAGTGCATCTGCACACAGAGGCCAGGGCTAGACACGCC TGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCTGTTC AGAGTGACCCCCGAGATTCTGCCGGGCTC
204	Nucleotide sequence anti-CD19(8B8-018) light chain	see Table 47
213	anti-CD19(8B8-018) Fc hole dimeric ligand	QVQLVQSGAEVVKKPGASVKVSKASGYTFTDYIMHWVRQ APQGQLEWMGYINPYNDGSKYTEKFQGRVTMTSDTSISTA YMELSRRLRSDDTAVYYCARGTYYYGSALFDYWQGQTTVT

-261-

	(71-248) chain	VSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVT VSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQ TYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAG GPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNW YVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLN GKEYKCKVSNKALGAPIEKTISKAKGQPREPQVCTLPSPRD ELTKNQVSLSCAVKGFYPSDIAVEWESNGQPENNYKTPP VLDSGDSFFLVSKLTVDKSRWQQGNVFSVSMHEALHNH YTQKSLSLSPGGGGGSGGGGSREGPELSPDDPAGLLDLRQ GMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKED TKELVVAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQP LRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAG QRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGL GGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQLVAQNV LLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKAGV YYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALA LTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEA RARHAWQLTQGATVLGLFRVTPEIPAGL
214	anti-CD19(8B8-018) Fc knob monomeric (71-248) ligand	QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYIMHWVRQ APGQGLEWMGYINPYNDGSKYTEKFQGRVTMTSDTSISTA YMELSRLRSDDTAVYYCARGTYYYGSALFDYWGQGTTVT VSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVT VSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQ TYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAG GPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNW YVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLN GKEYKCKVSNKALGAPIEKTISKAKGQPREPQVYTLPPCRD ELTKNQVSLWCLVKGFYPSDIAVEWESNGQPENNYKTPP VLDSGDSFFLYSKLTVDKSRWQQGNVFSVSMHEALHNH YTQKSLSLSPGGGGGSGGGGSREGPELSPDDPAGLLDLRQ GMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKED TKELVVAKAGVYYVFFQLELRRVVAGEGSGSVSLALHLQP LRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAG QRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGL
206	anti-CD19(8B8-018) light chain	see Table 47

7.2 Preparation of CD19 (8B8-derived affinity matured) targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules and corresponding control molecules

5 7.2.1 Generation of 8B8-derived affinity-matured anti-CD19 binders devoid of hotspots

7.2.1.1 Selection of affinity matured CD19-specific antibodies

De-amidation of the asparagine residues at positions 27d and 28, located in CDR1 of the light chain of the humanized clone 8B8, leads to a significant reduction in the biological activity.

Therefore, 2 phage display libraries were generated in which a) both asparagine residues at positions 27d and 28 were eliminated and b) additional CDRs of heavy and light chain were randomized in order to select for 8B8 variants with an improved affinity.

7.2.1.2 Generation of 8B8 affinity maturation libraries devoid of LCDR1 hotspots

5 Generation of affinity-matured 8B8-derived antibodies without the de-amidation sites N27d and N28, located in LCDR1, was carried out by phage display using standard protocols (Silacci et al, 2005). In a first step, the VL and VH DNA sequences of the humanized parental clone 8B8 (SEQ ID NO: 215 and SEQ ID NO: 216) were cloned into our phagemid which was then used as a template for randomization. In a next step, two libraries were generated for the
10 selection of favourable clones by phage display. In order to eliminate the above-mentioned hotspot positions, a LCDR1 randomization primer (SEQ ID NO: 217) that only allowed amino acids S T Q E at positions 27d and 28 was used for both libraries. Maturation library 1 was randomized in CDR1 and 2 of both the light and the heavy chain, while maturation library 2 was randomized in CDR1 and 3 of the light chain and in CDR3 of the heavy chain. The randomized
15 positions in the respective CDR regions are shown in **Figure 31A**. For the generation of the maturation library 1, randomized in CDR1 and 2 of both the light and the heavy chain, three fragments were assembled by “splicing by overlapping extension” (SOE) PCR and cloned into the phage vector (**Figure 31B**). The following primer combinations were used to generate the library fragments: fragment 1 (LMB3 (SEQ ID NO: 222) and CD19 L1 reverse random (SEQ ID
20 NO: 217), fragment 2 (CD19 L2 forward random (SEQ ID NO: 218) and CD19 H1 reverse random (SEQ ID NO: 219), and fragment 3 (CD19 H2 forward random (SEQ ID NO: 220) and CD19 H3 reverse constant (SEQ ID NO: 221) (**Table 53**). After assembly of sufficient amounts of full length randomized fragment, it was digested with *NcoI/NheI* alongside with identically treated acceptor phagemid vector. A 3-fold molar excess of library insert was ligated with 10 µg
25 of phagemid vector. Purified ligations were used for 20 transformations resulting in 2×10^9 transformants. Phagemid particles displaying the 8B8 affinity maturation library were rescued and purified by PEG/NaCl purification to be used for selections.

The generation of the second library, randomized in CDR1 and 3 of the light chain and in CDR3 of the heavy chain, was done similarly. The following primer combinations were used to
30 generate the library fragments: fragment 1 (LMB3 (SEQ ID NO: 222) and CD19 L1 reverse random (SEQ ID NO: 217), fragment 2 (CD19 L1 forward constant (SEQ ID NO 223) and CD19 L3 reverse random (SEQ ID NO 224), and fragment 3 (CD19 L3 forward constant (SEQ ID NO: 225) and CD19 H3 reverse random (SEQ ID NO: 226) (**Table 54**). After assembly of sufficient amounts of full length randomized fragment, it was digested with *NcoI/KpnI* alongside with
35 identically treated acceptor phagemid vector. A 3-fold molar excess of library insert was ligated with 20ug of phagemid vector. Purified ligations were used for 40 transformations resulting in 2

× 10 exp9 transformants. Phagemid particles displaying the 8B8 affinity maturation library were rescued and purified by PEG/NaCl purification to be used for selections.

Table 53: Primers for 8B8 affinity maturation and hotspot removal library L1_L2 / H1_H2

SEQ ID	Name	Sequence
217	CD19 L1 reverse random	CAG CTG CGG GCT CTG ACC CGG TTT CTG GAG ATA CCA GTT CAG 1 CGT 2 GCC 3 GGA 4 TTC CAG AGA TTG GCT GGA TTT GCA AGA AAT G 1: 40% Y, 6% A/S/T/G/P/D/N/E/Q/V, 2: 40% N, 6% A/S/T/Y/G/P/D/E/Q/V, 3: 25% S/T/Q/E, 4: 25% S/T/Q/E
218	CD19 L2 forward random	CTC CAG AAA CCG GGT CAG AGC CCG CAG CTG CTG ATC TAC 5 GTA TCT 6 CGC 7 8 GGC GTT 9 GAT CGT TTC AGC GGT TCT GGA TCC GGC ACC 5: 30% R, 20% E, 5% A/S/T/Y/G/P/D/N/Q/V, 6: 30% K, 20% S, 5% A/N/T/Y/G/P/D/E/Q/V, 7: 40% F, 5% A/S/T/Y/G/P/D/E/Q/V/I/L, 8: 40% S, 6.6% A/T/Y/G/P/D/E/Q/V, 9: 50% P, 50% L
219	CD19 H1 reverse random	CAT CCA CTC CAG ACC CTG GCC CGG GGC CTG ACG AAC CCA 10 CAT 11 12 13 14 GAA 15 GTA ACC AGA TGC TTT GCA GCT CAC TTT AAC GGA AGC 10: 52% H, 4% G/A/S/P/T/N/Y/D/E/Q/V/I, 11: 30% I, 15% Y, 5% G/A/S/T/P/N/H/D/E/Q/V, 12: 52% Y, 4% G/A/S/P/T/N/H/D/E/Q/V/I, 13: 30% D, 15% G, 5% A/S/P/Y/N/H/D/E/Q/V/I, 14: 52% T, 4% G/A/S/P/Y/N/H/D/E/Q/V/I, 15: 52% T, 4% G/A/S/P/Y/N/H/D/E/Q/V/I
220	CD19 H2 forward random	CAG GCC CCG GGC CAG GGT CTG GAG TGG ATG GGC 16 ATT 17 CCA 18 19 20 21 TCC 22 TAT ACC 23 AAA TTC CAG GGC CGC GTC ACG ATG ACC 16: 45% Y, 5% A/S/P/T/N/H/D/E/Q/V/I, 17: 52% N, 4% G/A/S/P/Y/T/H/D/E/Q/V/I, 18: 40% Y, 5% G/A/S/P/T/N/H/D/E/Q/V/I, 19: 30% N, 15% S, 5% G/A/T/P/Y/H/D/E/Q/V/I, 20: 30% D, 15% G, 5% A/S/T/P/Y/N/H/E/Q/V/I, 21: 52% G, 4% N/A/S/P/Y/T/H/D/E/Q/V/I, 22: 30% K, 15% N, 4% G/A/S/P/Y/T/H/D/E/Q/V/I, 23: 30% E, 15% Q, 5% G/A/S/T/P/Y/N/H/D/V/I
221	CD19 H3 reverse constant	CGTCACCGGTTCCGGGAAGTAGTCCTTGACCAG
222	LMB3	CAGGAAACAGCTATGACCATGATTAC

Table 54: Primers for 8B8 affinity maturation and hotspot removal library L1_L3 / H3

SEQ ID	Name	Sequence
223	CD19 L1 forward constant	TGGTATCTCCAGAAACCGGGTCAGAGCCCGCAG
217	CD19 L1 reverse random	See Table 53
224	CD19 L3 reverse random	TTT AAT TTC CAG TTT AGT TCC TTG ACC GAA GGT 24 25 26 27 28 29 CTG CAG ACA ATA GTA GAC GCC AAC GTC TTC AGC 24: 52% Y, 4% G/A/S/T/N/P/D/E/Q/V/L/I, 25: 52% P, 4% G/A/S/T/Y/N/H/D/E/Q/V/I, 26: 42% V, 10% L, 4% G/A/S/T/Y/N/P/D/E/Q/V/I, 27: 52% H, 4% G/A/S/T/Y/N/P/D/E/Q/V/I, 28: 42% T, 10% I, 4% G/A/S/T/Y/N/P/D/E/Q/V/L, 29: 45% L, 11% G, 4% A/S/T/Y/N/P/D/E/Q/V/I
225	CD19 L3 forward constant	ACCTTCGGTCAAGGAACTAACTGGAAATTAAACG
226	CD19 H3 reverse random	TT GGT GCT AGC AGA GCT TAC GGT CAC CGT GGT ACC TTG GCC CCA GTA ATC AAA 30 31 32 33 34 35 36 37 38 GCG TGC ACA ATA GTA AAC AGC GGT GTC 30: 50% L, 3.8% G/A/S/T/P/H/Y/N/D/E/Q/V/I, 31: 50% A, 4.2% G/S/T/P/H/Y/N/D/E/Q/V/I, 32: 50% S, 4.2% G/A/T/P/H/Y/N/D/E/Q/V/I, 33: 50% G, 4.2% S/A/T/P/H/Y/N/D/E/Q/V/I, 34: 50% Y, 4.2% G/A/T/P/H/S/N/D/E/Q/V/I, 35: 50% Y, 4.2% G/A/T/P/H/S/N/D/E/Q/V/I, 36: 50% Y, 4.2% G/A/T/P/H/S/N/D/E/Q/V/I, 37: 50% T, 4.2% G/A/Y/P/H/S/N/D/E/Q/V/I, 38: 50% G, 4.2% Y/A/T/P/H/S/N/D/E/Q/V/I
222	LMB3	See Table 53

7.2.1.3 Selection of affinity matured 8B8-derived clones devoid of LCDR1 hotspots N27d and N28

5

For the selection of affinity-matured clones devoid of the LCDR1 hotspots N27d and N28, two selection approaches by phage display were performed:

In the first approach, the selection was executed on human CD19-Fc fusion protein using both phage display libraries. Panning rounds were performed in solution according to the following pattern: 1. binding of $\sim 10^{12}$ phagemid particles to 30nM biotinylated CD19-Fc

10

protein for 0.5 h in a total volume of 1ml, 2. capture of biotinylated CD19-Fc protein and specifically bound phage particles by addition of 5.4×10^7 streptavidin-coated magnetic beads for 10 min, 3. washing of beads using 5x 1ml PBS/Tween20 and 5x 1ml PBS, 4. elution of phage particles by addition of 1ml 100mM TEA for 10 min and neutralization by adding 500ul 1M Tris/HCl pH 7.4, 5. re-infection of exponentially growing *E. coli* TG1 bacteria, and 6. infection with helperphage VCSM13 and subsequent PEG/NaCl precipitation of phagemid particles to be used in subsequent selection rounds. Selections were carried out over 3 rounds using decreasing antigen concentrations (30×10^{-9} M, 10×10^{-9} M, and 3×10^{-9} M). In round 2 and 3, capture of antigen:phage complexes was performed using neutravidin plates instead of streptavidin beads. Neutravidin plates were washed with 5x PBS/Tween20 and 5x PBS. In round 3, the neutravidin plate was incubated overnight in 2 liters PBS for an “off-rate” selection before phage was eluted from the plate. Furthermore, cynomolgus CD19-Fc protein was used in round 2 in order to enrich cross-reactive binders.

In the second selection approach, the phage panning was executed on cells transiently expressing either the human or cynomolgus CD19 ECD on the cell surface. For the transient transfection of HEK cells, expression plasmids were generated that harbor the DNA sequences (from 5' to 3') for the following protein segments: A Flag tag, a SNAP tag, the CD19 ECD of either human or cynomolgus origin, and the transmembrane region of the Platelet-derived growth factor receptor (PDGFR) (SEQ ID NOs: 227 and 228). The expression of the respective proteins (SEQ ID NOs: 229 and 230) on the cell surface was confirmed by flow cytometry using an anti-Flag antibody for detection. Both libraries were exposed in the first selection round to cells either expressing the human or cynomolgus CD19 ECD-containing protein fusion. For the subsequent panning rounds, the species of the CD19 ECD was alternated accordingly. Cells transiently transfected with an irrelevant membrane protein were used for pre-clearing.

Panning rounds were performed according to the following pattern:

1. Transfection of HEK cells with constructs expressing either CD19 ECD or an irrelevant transmembrane protein according to the standard procedure described before,
2. Incubation of the cells for total 48h at 37°C in an incubator with a 5% CO₂ atmosphere, 3. Isolation of cells by centrifugation (3 min at 250×g) and re-suspension of 1×10^7 CD19 ECD-positive cells and 1×10^7 negative cells in PBS/5% BSA, respectively,
3. Pre-clearing of unspecific phage by incubating the phage library with 1×10^7 CD19-negative cells for 60 min at 4°C using a gently rotating tube rotator,
4. Centrifugation of cells at 250×g for 3min and transfer of supernatant into a fresh tube and addition of 1×10^7 CD19-positive cells and incubation for 60 min at 4 °C by gentle rotation on a tube rotator,
5. Washing of cells by centrifugation for 1 min at 250×g, aspiration of the supernatant, and re-suspension in 1 ml PBS (8 times),

6. Phage elution with 1 ml 100mM TEA, incubation for 5 min at RT, and neutralization of the eluate with 500 ul 1M Tris-HCl, pH7.6,
7. re-infection of exponentially growing *E. coli* TG1 bacteria, and
8. infection with helperphage VCSM13 and subsequent PEG/NaCl precipitation of phagemid particles to be used in subsequent selection rounds. Selections were carried out over 3 rounds.

For both selection approaches, specific binders were identified by ELISA as follows: 100 ul of 30 nM biotinylated CD19-Fc protein per well were coated on neutravidin plates. Fab-containing bacterial supernatants were added and binding Fabs were detected via their Flag-tags using an anti-Flag/HRP secondary antibody.

- 10 Clones that were ELISA-positive on recombinant human CD19 were further tested in a cell-based ELISA using cells that were transiently transfected with the human CD19 ECD-containing expression plasmid (SEQ ID NO: 227). This analysis was performed as follows: 48 h after transfection, HEK cells were harvested and centrifuged at 250xg for 5 min. Cells were then re suspended in ice-cold PBS BSA 2% to 4×10^6 cells/ml and incubated for 20 min on ice to
- 15 block unspecific binding sites. 4×10^5 cells in 100ul were distributed to each well of a 96 well plate and centrifuged at 250xg and 4°C for 3 min. Supernatant was aspirated off and 50ul bacterial supernatant containing soluble Fab fragments was diluted with 50ul ice-cold PBS/BSA 2%, added to the plate, mixed with the cells and incubated for 1 h at 4°C. Afterwards, cells were
- 20 washed 3 times with ice cold PBS before 100ul PBS BSA 2% per well containing a 1:2000 dilution of anti-Fab-HRP antibody were added. After an incubation time of 1 h, cells were washed again 3 times with ice-cold PBS. For the development, 100ul “1-step ultra TMB-ELISA” substrate was added per well. After an incubation time of 10 minutes, supernatant was
- 25 transferred to a new 96-well plate containing 40ul H₂SO₄ 1M per well and absorbance was measured 450 nM. Clones exhibiting significant signals over background were subjected to a kinetic screening experiment by SPR-analysis using ProteOn XPR36.

7.2.1.4 Identification of affinity-matured 8B8-derived variants by SPR

In order to further characterize the ELISA-positive clones, the off-rate was measured by surface plasmon resonance and compared with the parental humanized clone 8B8.

- 30 For this experiment, 7000 RU of polyclonal anti-human Fab antibody were immobilized on all 6 channels of a GLM chip by Amine coupling (NaAcetate pH4.5, 25 µl/min, 240s) (vertical orientation). Each antibody-containing bacterial supernatant was filtered and 2-fold diluted with PBS, and then injected for 360s at 25 µl/minute to achieve immobilization levels of between 100 and 400 response units (RU) in vertical orientation. Injection of monomeric CD19-Fc: For one-shot kinetics measurements, injection direction was changed to horizontal orientation, three-fold
- 35 dilution series of purified monomeric CD19-Fc (varying concentration ranges between 150 and 6

nM) were injected simultaneously at 50 μ l/min along separate channels 1-4, with association times of 180 s, and dissociation times of 300 s. A human IgG Fc fragment (150nM) was injected in channel 5 as a negative control for specific binding to monomeric CD19-Fc Buffer (PBST) was injected along the sixth channel to provide an “in-line” blank for referencing. Regeneration was performed by two pulses of 10mM glycine pH 1.5 and 50mM NaOH for 30s at 90ul/min (horizontal orientation). Dissociation rate constants (k_{off}) were calculated using a simple one-to-one Langmuir binding model in ProteOn Manager v3.1 software by simultaneously fitting the sensorgrams. Clones expressing Fabs with the slowest dissociation rate constants were identified (**Table 55**). Of note, the dissociation rate constants of clones 5A07 and 5B08 could not be determined due to inadequate fitting. Nevertheless, both clones were selected because results obtained suggested a very slow dissociation. The variable domains of the corresponding phagemids were sequenced. Importantly, both asparagine residue in LCDR1 (position 27d and 28) were replaced by a serine or a threonine, demonstrating that both de-amidation sites were removed. An alignment is shown in **Figure 32**. The CDRs of the best clones are listed in **Table 56** (variable regions of the light chain) and **Table 57** (variable regions of the heavy chain) (clone 5H09: (SEQ ID NO:231-236); clone 7H07: (SEQ ID NO:237-242); clone 2B03: (SEQ ID NO: 243-248); clone 2B11: (SEQ ID NO:249-254); clone 5A07: (SEQ ID NO:255-260); clone 5B08: (SEQ ID NO:261-266); clone 5D08: (SEQ ID NO:267-272).

Table 55: Dissociation constants of selected clones obtained in screening analysis with bacterial supernatant

clone	Dissociation constant k_d (1/s)
Parental 8B8	3.01E-4
5H09	2.58E-4
7H07	5.75E-5
2B03	3.24E-5
2B11	4.37E-6
5A07	n.d.
5B08	n.d.
5D08	1.95E-4

Table 56: CDR sequences of the selected 8B8 light chains

clone	SEQ ID NO	CDR-L1	SEQ ID NO	CDR-L2	SEQ ID NO	CDR-L3
5H09	231	KSSQSLESSTGNTYLN	232	RVSKRFS	233	LQLIDYPVT
7H07	237	KSSQSLETSTGNTYLN	238	RVSKRFS	239	LQATHIPYT
2B03	243	KSSQSLETSTGNTYLN	244	RVSKRFS	245	LQLTHVPYT
2B11	249	KSSQSLETSTGTTYLN	250	RVSKRFS	251	LQLLEDPYT
5A07	255	KSSQSLETSTGNTYLN	256	RVSKRFS	257	LQPGHYPGT
5B08	261	KSSQSLETSTGNTYLN	262	RVSKRFS	263	LQLDSYPNT
5D08	267	KSSQSLETSTGNTYLN	268	RVSKRFS	269	LQLTHEPYT

Table 57: CDR sequences of the selected 8B8 heavy chains

5

clone	SEQ ID NO	CDR-H1	SEQ ID NO	CDR-H2	SEQ ID NO	CDR-H3
5H09	234	DYIMH	235	YINPYNDGSKYTEKFQG	236	GTYYYYGSALFDY
7H07	240	DYIMH	241	YINPYNDGSKYTEKFQG	242	GTYYYYGSELDY
2B03	246	DYITH	247	YINPYNDGSKYTEKFQG	248	GTYYYYGPDLDY
2B11	252	DYIMH	253	YINPYNDGSKYTEKFQG	254	GTYYYYGPQLFDY
5A07	258	DYIMH	259	YINPYNDGSKYTEKFQG	260	GTYYYYGSALFDY
5B08	264	DYIMH	265	YINPYNDGSKYTEKFQG	266	GTYYYYGPQLFDY
5D08	270	DYIMH	271	YINPYNDGSKYTEKFQG	272	GTYYYYGSELDY

7.2.2 Characterization of affinity-matured 8B8-derived antibodies

7.2.2.1 Cloning of variable antibody domains into expression vectors

10 The variable regions of heavy and light chain DNA sequences of the selected anti-CD19 binders were subcloned in frame with either the constant heavy chain or the constant light chain of human IgG1. In the heavy chain, Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in order to abrogate binding to Fc gamma receptors according to the method described in International Patent Appl. Publ. No. WO 2012/130831 A1.

The cDNA and amino acid sequences of the anti-CD19 IgGs are shown in **Table 58** and **Table 59**, respectively. All antibody-encoding sequences were cloned into an expression vector, which drives transcription of the insert with a chimeric MPSV promoter and contains a synthetic polyA signal sequence located at the 3' end of the CDS. In addition, the vector contains an EBV OriP sequence for episomal maintenance of the plasmid.

Table 58: cDNA and amino acid sequences of anti-CD19 clone 8B8 in P329GLALA human IgG1 format

SEQ ID NO:	Clone and Chain	Sequence
273	8B8 Parental light chain	GATGCTGTGATGACCCAAACTCCACTCTCCCTGCCTGTCAGTCTTGGA GATCAAGCCTCCATCTCTTGCAGGTCTAGTCAGAGCCTTGAAAACAGT AATGGAAACACCTATTTGAACTGGTACCTCCAGAAACCAGGCCAGTC TCCACAACCTCCTGATCTACAGGGTTTCCAAACGATTTTCTGGGGTCTCT AGACAGGTTCAAGTGGTAGTGGATCAGGGACAGATTTACACTGAAAA TCAGCAGAGTGGAGGCTGAGGATTTGGGAGTTTATTTCTGCCTACAA CTTACACATGTCCCGTACACGTTTCGGAGGGGGGACCAAGCTGGAAAT AAAACGTACGGTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGATG AGCAGTTGAAATCTGGAAGTGCCTCTGTTGTGTGCCTGCTGAATAACTTCT ATCCCAGAGAGGCCAAAGTACAGTGGAAGGTGGATAACGCCCTCCAATCG GGTAACTCCCAGGAGAGTGTACAGAGCAGGACAGCAAGGACAGCACCT ACAGCCTCAGCAGCACCTGACGCTGAGCAAAGCAGACTACGAGAAACAC AAAGTCTACGCCTGCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCAC AAAGAGCTTCAACAGGGGAGAGTGT
274	8B8 parental heavy chain	GAGGTCCAGCTGCAGCAGTCTGGACCTGAGCTGGTAAAGCCTGGGGC TTCAGTGAAGATGGCCTGCAAGGCTTCTGGATACACATTCAGTGACTA TATTATGCACTGGGTGAAGCAGAAGACTGGGCAGGGCCTTGAGTGGA TTGGATATATTAATCCTTACAATGATGGTTCTAAGTACACTGAGAAGT TCAACGGCAAGGCCACACTGACTTCAGACAAATCTTCCATCACAGCC TACATGGAGCTCAGCAGCCTGACCTCTGAGGACTCTGCGGTCTATTAC TGTGCAAGAGGGACCTATTATTATGGTAGCGCCCTCTTTGACTACTGG GGCCAAGGCACCACTCTCACAGTCTCCTCGGCTAGCACCAAGGGCCCA TCGGTCTTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGGGGGCACAGC GGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGGTGACGGTG TCGTGGAACTCAGGCGCCCTGACCAGCGGCGTGCACACCTTCCCGGCTG TCCTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCC TCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACGTGAATCACAAAGCC CAGCAACACCAAGGTGGACAAGAAAGTTGAGCCCAAATCTTGTGACAAAA CTCACACATGCCCACCGTGCCAGCACCTGAAGCTGCAGGGGGACCGTC AGTCTTCTCTTCCCCCCTCAAGGACACCCCTCATGATCTCCCGGAC CCCTGAGGTACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCCTGAG GTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCAAGAC AAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGCGTC CTACCGTCTCTGCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAA GGTCTCCAACAAAGCCCTCGGCGCCCCCATCGAGAAAACCATCTCCAAAG CCAAAGGGCAGCCCCGAGAACCACAGGTGTACACCTGCCCCCATCCCG GGATGAGCTGACCAAGAACCAGGTGACCTGACCTGCCTGGTCAAAGGCT TCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGA GAACAACTACAAGACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCT TCCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAA

		<i>CGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACTACACGCA GAAGAGCCTCTCCCTGTCTCCGGGTA</i>
275	8B8 Parental light chain	<i>DAVMTQTPLSLPVSLGDQASISCRSSQSLENSNGNTYLNWYLQKPGQSP QLLIYRVSKRFSGLDRFSGSGSGTDFTLKISRVEAEDLGVYFCLQLTHVP YTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQW KVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQGL SSPVTKSFNRGEC</i>
276	8B8 parental heavy chain	<i>EVQLQQSGPELVKPGASVKMACKASGYTFTDYIMHWVKQKTGQGLEWI GYINPYNDGSKYTEKFNGKATLTSDKSSITAYMELSSLTSEDSAVYYCAR GTYYYGSALFDYWQGTTLTVSSASTKGPSVFPLAPSSKSTSGGTAALGCL VKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYIC NVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPPKPKDTL MISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVV SVLTVLHQDWLNGKEYKCKVSNKALGAPIEKTISKAKGQPREPQVYTLPPSRD ELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTPPVLDSDGSFFLYSK LTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPGK</i>

Table 59: cDNA and amino acid sequences of affinity matured anti-CD19 clones in P329GLALA human IgG1 format

SEQ ID NO:	Clone and Chain	Sequence
277	2B11 light chain	<i>GATATTGTCATGACTCAAACCTCCACTGTCTCTGTCCGTGACCCCGGGT CAGCCAGCGAGCATTTCTTGCAAATCCAGCCAATCTCTGGAAACCTCC ACCGGCACCACGTACCTGAACCTGGTATCTCCAGAAACCGGGTCAGAG CCCGCAGCTGCTGATCTACCGTGTATCTAAGCGCTTCTCCGGCGTTCC TGATCGTTTCAGCGGTTCTGGATCCGGCACCGACTTTACTCTGAAAAT CAGCCGTGTGGAAGCTGAAGACGTTGGCGTCTACTATTGTCTGCAGCT GCTGGAAGATCCATACACCTTCGGTCAAGGAACGAACTGGAAATTA AACGTACGGTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGATG AGCAGTTGAAATCTGGAACCTGCCTCTGTTGTGTGCCTGCTGAATAACT TCTATCCCAGAGAGGCCAAAGTACAGTGGAAGGTGGATAACGCCCTC CAATCGGGTAACTCCCAGGAGAGTGTACAGAGCAGGACAGCAAGG ACAGCACCTACAGCCTCAGCAGCACCTGACGCTGAGCAAAGCAGAC TACGAGAAACACAAAGTCTACGCCTGCGAAGTCACCCATCAGGGCCT GAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGAGAGTGT</i>
278	2B11 heavy chain	<i>CAGGTGCAATTGGTTCAATCTGGTGCTGAAGTAAAAAAACCGGGCGC TTCCGTTAAAGTGAGCTGCAAAGCATCTGGTTACACCTTCACTGACTA TATCATGCACTGGGTTTCGTCAGGCCCCGGGCCAGGGTCTGGAGTGGA TGGGCTACATTAACCCATACAACGACGGTTCCAAATATACCGAGAAA TTCCAGGGCCGCGTCACGATGACCAGCGACACTTCTATCTCCACCGCG TACATGGAACCTGTCTAGACTGCGTTCTGACGACACCGCTGTTTACTAT TGTGCACGCGGTACCTACTACTACGGTCCACAGCTGTTTGATTACTGG GGCCAAGGTACCACGGTGACCGTAAGCTCTGCTAGCACCAAGGGCCC ATCGGTCTTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGGGGGCAC AGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGGTGA CGGTGTCGTGGAACCTCAGGCGCCCTGACCAGCGGCGTGCACACCTTC CCGGCTGTCCTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTG ACCGTGCCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACGT GAATCACAAGCCCAGCAACACCAAGGTGGACAAGAAAGTTGAGCCC AAATCTTGTGACAAAACCTCACACATGCCACCGTGCCAGCACCTGA</i>

		AGCTGCAGGGGGACCGTCAGTCTTCCTCTTCCCCCAAAACCCAAGG ACACCTCATGATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTG GACGTGAGCCACGAAGACCCCTGAGGTCAAGTTCAACTGGTACGTGGA CGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCAG TACAACAGCACGTACCGTGTGGTCAGCGTCCTCACCGTCCTGCACCA GGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAA GCCCTCGGCGCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCA GCCCCGAGAACCACAGGTGTACACCCTGCCCCCATCCCGGGATGAGC TGACCAAGAACCAGGTCAGCCTGACCTGCCTGGTCAAAGGCTTCTAT CCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGA ACAACCTACAAGACCACGCCTCCCGTGTGCTGGACTCCGACGGCTCCTTCT TCCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGG AACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACTAC ACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAA
279	2B11 light chain	DIVMTQTPLSLSVTPGQPASISCKSSQSLETSTGTTYLNWYLQKPGQSPQL LIYRVSKRFSGVDPDRFSGSGSDFTLKISRVEAEDVGVYYCLQLLEDPY TFGQGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKV QWKVDNALQSGNSQESVTEQDSKDYSLSTLTLSKADYEKHKVYAC EVTHQGLSSPVTKSFNRGEC
280	2B11 heavy chain	QVQLVQSGAEVKKPGASVKVSCKASGYTFDYMHWVRQAPGQGLEW MGYINPYNDGSKYTEKFQGRVTMTSDTSISTAYMELSRLSDDTAVYYC ARGTYYYGPQLFDYWGQGTITVTVSSASTKGPSVFPLAPSSKSTSGGTAA LGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPS SLGTQTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVF LFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKT KPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGAPIEKTISK AKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPE NNYKTTTPVLDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHY TQKSLSLSPGK
281	7H07 light chain	GATATTGTTATGACTCAAACCTCCACTGTCTCTGTCCGTGACCCCGGGT CAGCCAGCGAGCATTTCTTGCAAATCCAGCCAATCTCTGGAAACCTCC ACCGGCAACACGTACCTGAACTGGTATCTCCAGAAACCGGGTCAGAG CCCGCAGCTGCTGATCTACCGTGTATCTAAGCGCTTCTCCGGCGTTCC TGATCGTTTCAGCGGTTCTGGATCCGGCACCGACTTTACTCTGAAAAT CAGCCGTGTGGAAGCTGAAGACGTTGGCGTCTACTATTGTCTGCAGG CAACCCATATCCCATACACCTTCGGTCAAGGAATAAAGTGGAAATT AAACGTACGGTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGAT GAGCAGTTGAAATCTGGAAGTGCCTCTGTTGTGTGCCTGCTGAATAAC TTCTATCCCAGAGAGGCCAAAGTACAGTGGAAAGGTGGATAACGCCCT CCAATCGGGTAACTCCCAGGAGAGTGTACAGAGCAGGACAGCAAG GACAGCACCTACAGCCTCAGCAGCACCTGACGCTGAGCAAAGCAGA CTACGAGAAACACAAAGTCTACGCCTGCGAAGTCACCCATCAGGGCC TGAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGAGAGTGT
282	7H07 heavy chain	CAGGTGCAATTGGTTCAATCTGGTGCTGAAGTAAAAAAACCGGGCGC TTCCGTTAAAGTGAGCTGCAAAGCATCTGGTTACACCTTCACTGACTA TATCATGCACTGGGTTTCGTACAGGCCCGGGCCAGGGTCTGGAGTGGA TGGGCTACATTAACCCATACAAACGACGGTTCCAAATATACCGAGAAA TTCCAGGGCCGCGTCACGATGACCAGCGACACTTCTATCTCCACCGCG TACATGGAAGTGTCTAGACTGCGTTCTGACGACACCGCTGTTTACTAT TGTGCACGCGGTACCTACTACTACGGTTCTGAAGTGTGTTGATTACTGG GGCCAAGGTACCACGGTGACCGTAAGCTCTGCTAGCACCAAGGGCCC ATCGGTCTTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGGGGGCAC AGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGGTGA CGGTGTCGTGGAAGTCAAGGCGCCCTGACCAGCGGCGTGCACACCTTC

		CCGGCTGTCCTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTG ACCGTGCCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACGT GAATCACAAGCCCAGCAACACCAAGGTGGACAAGAAAGTTGAGCCC AAATCTTGTGACAAAACCTCACACATGCCCACCGTGCCCAGCACCTGA AGCTGCAGGGGGACCGTCAGTCTTCCCTCTTCCCCCAAAACCCAAGG ACACCCTCATGATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTG GACGTGAGCCACGAAGACCCCTGAGGTCAAGTTCAACTGGTACGTGGA CGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCAG TACAACAGCACGTACCGTGTGGTCAGCGTCCTCACCCTCCTGCACCA GGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAA GCCCTCGGCGCCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCA GCCCCGAGAACCACAGGTGTACACCCTGCCCCCATCCCGGGATGAGC TGACCAAGAACCAGGTGAGCCTGACCTGCCTGGTCAAAGGCTTCTAT CCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGA ACAACCTACAAGACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCT TCCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGG AACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACTAC ACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAA
283	7H07 light chain	DIVMTQTPLSLSVTPGQPASISCKSSQSLETSTGNTYLNWYLQKPGQSPQL LIYRVSKRFSGVDPDRFSGSGSDFTLKISRVEAEDVGVYYCLQATHIPYT FGQGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQ WKVDNALQSGNSQESVTEQDSKDSYSTLSSTLTLSKADYEEKHKVYACE VTHQGLSSPVTKSFNRGEC
284	7H07 heavy chain	QVQLVQSGAEVKKPGASVKVSCKASGYTFDYMHWVRQAPGQGLEW MGYINPYNDGSKYTEKFQGRVTMTSDTSISTAYMELSRLRSDDTAVYYC ARGTYYYGSELFDYWGQGTITVTVSSASTKGPSVFPLAPSSKSTSGGTAA LGCLVKDYFPEPVTWSNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSS SLGTQTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVF LFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKT KPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGAPIEKTISK AKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPE NNYKTTTPVLDSDGSFFLYSKLTVDKSRWQQGNVVFSCSVMHLEAHNHY TQKSLSLSPGK
285	2B03 light chain	GATATTGTTATGACTCAAACCTCCACTGTCTCTGTCCGTGACCCCGGGT CAGCCAGCGAGCATTTCTTGCAAATCCAGCCAATCTCTGGAAACCTC CACCGGCAACACGTACCTGAACTGGTATCTCCAGAAACCGGGTCAGA GCCCCGAGCTGCTGATCTACCGTGTATCTAAGCGCTTCTCCGGCGTTC CTGATCGTTTCAGCGGTTCTGGATCCGGCACCGACTTTACTCTGAAAA TCAGCCGTGTGGAAGCTGAAGACGTTGGCGTCTACTATTGTCTGCAG TTGACCCACGTTCCGTACACCTTCGGTCAAGGAANNAACTGGAAAT TAAACGTACGGTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGA TGAGCAGTTGAAATCTGGAAGTGCCTCTGTTGTGTGCCTGCTGAATAA CTTCTATCCCAGAGAGGGCCAAAGTACAGTGGAAGGTGGATAACGCCC TCCAATCGGGTAACTCCAGGAGAGTGTACAGAGCAGGACAGCAA GGACAGCACCTACAGCCTCAGCAGCACCCCTGACGCTGAGCAAAGCA GACTACGAGAAACACAAAGTCTACGCCTGCGAAGTCACCCATCAGGG CCTGAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGAGAGTGT
286	2B03 heavy chain	CAGGTGCAATTGGTTC AATCTGGTGCTGAAGTAAAAAAACCGGGCGC TTCCGTTAAAGTGAGCTGCAAAGCATCTGGTTACACCTTCACTGACTA TATCACGCACTGGGTTCGTCAGGCCCGGGCCAGGGTCTGGAGTGGA TGGGCTACATTAACCCATACAAACGACGGTTCCAAATATACCGAGAAA TTCCAGGGCCGCGTCACGATGACCAGCGACACTTCTATCTCCACCGC GTACATGGAAGTGTCTAGACTGCGTTCTGACGACACCGCTGTTTACTA TTGTGCACGCGGTACCTACTACTACGGTCCAGATCTGTTTGATTACTG

		GGGCCAAGGTACCACGGTGACCGTAAGCTCTGCTAGCACCAAGGGCC CATCGGTCCTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGGGGGCA CAGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGGTG ACGGTGTCGTGGAACCTCAGGCGCCCTGACCAGCGGCGTGCACACCTT CCCGGCTGTCTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGT GACCGTGCCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACG TGAATCACAAGCCCAGCAACACCAAGGTGGACAAGAAAGTTGAGCC CAAATCTTGTGACAAAACCTCACACATGCCACCGTGCCAGCACCTG AAGCTGCAGGGGGACCGTCAGTCTTCTCTTCCCCCAAACCCAAAG GACACCCTCATGATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGT GGACGTGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGTGG ACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCA GTACAACAGCACGTACCGTGTGGTCAGCGTCCTACCGTCTGCACC AGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAA AGCCCTCGGCGCCCCCATCGAGAAAACCATCTCCAAGGCCAAAGGGC AGCCCCGAGAACCACAGGTGTACACCCTGCCCCCATCCCGGGATGAG CTGACCAAGAACCAGGTGAGCCTGACCTGCCTGGTCAAAGGCTTCTA TCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAG AACTACAAGACCACGCTCCCGTGTGGTCCGACGGCTCCTT CTTCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGG GGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACT ACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTA
287	2B03 light chain	DIVMTQTPLSLSVTPGQPASISCKSSQSLETSTGNTYLNWYLQKPGQSPQ LLIYRVSKRFSGVDPDRFSGSGSGTDFTLKISRVEAEDVGVYYCLQLTHVP YTFGQGXKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAK VQWKVDNALQSGNSQESVTEQDSKSTYLSSTLTLSKADYEKHKVYA CEVTHQGLSSPVTKSFNRGEC
288	2B03 heavy chain	QVQLVQSGAEVKKPGASVKVSCKASGYTFDTHWVRQAPGQGLEW MGYINPYNDGSKYTEKFQGRVTMTSDTSISTAYMELSRLSDDTAVYYC ARGTYYYGPDLDYWGQGTFTVTVSSASTKGPSVFPLAPSSKSTSGGTAA LGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSS SLGTQTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSV FLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKT KPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGAPIEKTISK AKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQP ENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFCFSVMHEALHNH YTQKSLSLSPGK
289	5A07 light chain	GATATTGTTATGACTCAAACCTCCACTGTCTCTGTCCGTGACCCCGGGT CAGCCAGCGAGCATTTCTTGCAAATCCAGCCAATCTCTGGAAACCTC CACCGGCAACACGTACCTGAACTGGTATCTCCAGAAACCGGGTCAGA GCCCCGAGCTGCTGATCTACCGTGTATCTAAGCGCTTCTCCGGCGTTC CTGATCGTTTCAGCGGTTCTGGATCCGGCACCGACTTTACTCTGAAAA TCAGCCGTGTGGAAGCTGAAGACGTTGGCGTCTACTATTGTCTGCAG CCAGGTCATTACCCAGGTACCTTCGGTCAAGGAATAAAGTGGAAAT TAAACGTACGGTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGA TGAGCAGTTGAAATCTGGAAGTGCCTCTGTTGTGTGCCTGCTGAATAA CTTCTATCCCAGAGAGGCCAAAGTACAGTGGAAGGTGGATAACGCC TCCAATCGGGTAACTCCAGGAGAGTGTACAGAGCAGGACAGCAA GGACAGCACCTACAGCCTCAGCAGCACCCCTGACGCTGAGCAAAGCA GACTACGAGAAACACAAAGTCTACGCCTGCGAAGTCACCCATCAGGG CCTGAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGAGAGTGT
290	5A07 heavy chain	CAGGTGCAATTGGTTCAATCTGGTGCTGAAGTAAAAAACCAGGGCGC TTCCGTTAAAGTGAGCTGCAAAGCATCTGGTTACACCTTCACTGACTA TATCATGCACTGGGTTTCGTACAGCCCCGGGCCAGGGTCTGGAGTGGA

		<p>TGGGCTACATTAACCCATACAACGACGGTTCCAAATATACCGAGAAA TTCCAGGGCCGCGTCACGATGACCAGCGACACTTCTATCTCCACCGC GTACATGGAAGTGTCTAGACTGCGTTCTGACGACACCGCTGTTTACTA TTGTGCACGCGGTACTTACTACTACGGTTCCGCCCTCTTTGATTACTG GGGCCAAGGTACCACGGTGACCGTAAGCTCTGCTAGCACCAAGGGCC CATCGGTCTTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGGGGGCA CAGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGGTG ACGGTGTCGTGGAAGTCAGGCGCCCTGACCAGCGGCGTGACACACCTT CCCGGTGTCTTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGT GACCGTGCCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACG TGAATCACAAGCCCAGCAACACCAAGGTGGACAAGAAAAGTTGAGCC CAAATCTTGTGACAAAACCTCACACATGCCACCCGTGCCCAGCACCTG AAGCTGCAGGGGGACCGTCAGTCTTCCTCTTCCCCCAAAACCCAAG GACACCCTCATGATCTCCCGGACCCCTGAGGTACATGCGTGGTGGT GGACGTGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGTGG ACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCA GTACAACAGCACGTACCGTGTGGTCAGCGTCCTACCGTCCTGCACC AGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAA AGCCCTCGGCGCCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGC AGCCCCGAGAACCACAGGTGTACACCCTGCCCCCATCCCGGGATGAG CTGACCAAGAACCAGGTACGCTGACCTGCCTGGTCAAAGGCTTCTA TCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAG AACAACTACAAGACCACGCTCCCGTGCTGGACTCCGACGGCTCCTT CTTCCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGG GGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACT ACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAA</p>
291	5A07 light chain	<p>DIVMTQTPLSLSVTPGQPASISCKSSQSLETSTGNTYLNWYLQKPGQSPQ LLIYRVSKRFSGVPDRFSGSGSGTDFTLKISRVEAEDVGVYYCLQPGHYP GTFGQGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAK VQWKVDNALQSGNSQESVTEQDSKDSSTYLSSTLTLSKADYEKHKVYA CEVTHQGLSSPVTKSFNRGEC</p>
292	5A07 heavy chain	<p>QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYIMHWVRQAPGQGLEW MGYINPYNDGSKYTEKFQGRVTMTSDTSISTAYMELSRLRSDDTAVYYC ARGTYYYGSALFDYWQGTTVTVSSASTKGPSVFPLAPSSKSTSGGTAA LGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSS SLGTQTYICNVNHKPSNTKVDKKVEPKSCDKHTHTCPPCPAPEAAAGPSV FLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKT KPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGAPIEKTISK AKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQP ENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFCSCVMHEALHNH YTQKSLSLSPGK</p>
293	5D08 light chain	<p>GATATTGTTATGACTCAAACCTCCACTGTCTCTGTCCGTGACCCCGGGT CAGCCAGCGAGCATTTCTTGCAAATCCAGCCAATCTCTGGAAACCTC CACCGGCAACACGTACCTGAAGTGGTATCTCCAGAAACCGGGTCAGA GCCCCGAGCTGCTGATCTACCGTGTATCTAAGCGCTTCTCCGGCGTTC CTGATCGTTTCAGCGGTTCTGGATCCGGCACCGACTTTACTCTGAAAA TCAGCCGTGTGGAAGCTGAAGACGTTGGCGTCTACTATTGTCTGCAG CTGACCCATGAACCATAACCTTCGGTCAAGGAACTAACTGGAAAT TAAACGTACGGTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGA TGAGCAGTTGAAATCTGGAAGTGCCTCTGTTGTGTGCTGCTGAATAA CTTCTATCCCAGAGAGGCCAAAGTACAGTGGAAGGTGGATAACGCC TCCAATCGGGTAACTCCCAGGAGAGTGTACAGAGCAGGACAGCAA GGACAGCACCTACAGCCTCAGCAGCACCCCTGACGCTGAGCAAAGCA GACTACGAGAAACACAAAGTCTACGCCTGCGAAGTCACCCATCAGGG</p>

		CCTGAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGAGAGTGT
294	5D08 heavy chain	CAGGTGCAATTGGTTC AATCTGGT GCTGAAGTAAAAAAACCGGGCGC TTCCGTTAAAGTGAGCTGCAAAGCATCTGGTTACACCTTCACTGACTA TATCATGCACTGGGTTCGTCAGGCCCGGGCCAGGGTCTGGAGTGGA TGGGCTACATTAACCCATAACAACGACGGTCCAAATATACCGAGAAA TTCCAGGGCCGCGTCACGATGACCAGCGACACTTCTATCTCCACCGC GTACATGGAAGTGTCTAGACTGCGTTCTGACGACACCGCTGTTTACTA TTGTGCACGCGGTACCTACTACTACGGTTCTGAACTGTTTGATTACTG GGGCCAAGGTACCACGGTGACCGTAAGCTCTGCTAGCACC AAGGGCC CATCGGTCTTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGGGGGCA CAGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGGTG ACGGTGTCGTGGAAGTACAGGCGCCCTGACCAGCGGCGTGACACCTT CCCGGTGTCTTACAGTCTCAGGACTCTACTCCCTCAGCAGCGTGGT GACCGTGCCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACG TGAATCACAAGCCCAGCAACACCAAGGTGGACAAGAAAGTTGAGCC CAAATCTTGTGACAAAACCTCACACATGCCCACCGTGCC CAGCACCTG AAGCTGCAGGGGGACCGTCAGTCTTCTCTTCCCCCAA AACC AAG GACACCCTCATGATCTCCCGGACCCCTGAGGTCAAGTTCAACTGGTACGTGG GGACGTGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGTGG ACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCA GTACAACAGCACGTACCGTGTGGTCAGCGTCCTCACCGTCTGCACC AGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAA AGCCCTCGGCGCCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGC AGCCCCGAGAACCACAGGTGTACACCCTGCCCCCATCCCGGGATGAG CTGACCAAGAACCAGGTACGCCTGACCTGCCTGGTCAAAGGCTTCTA TCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAG AACAACTACAAGACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTT CTTCCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGG GGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACT ACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAA
295	5D08 light chain	DIVMTQTPLSLSVTPGQPASISCKSSQSLETSTGNTYLNWYLQKPGQSPQ LLIYRVSKRFSGV PDRFSGSGSGTDFTLKISRVEAEDVGVYYCLQLTHEP YTFGQGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAK VQWKVDNALQSGNSQESVTEQDSKSTYLSSTLTLSKADYEKHKVYA CEVTHQGLSSPVTKSFNRGEC
296	5D08 heavy chain	QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYIMHWVRQAPGQGLEW MGYINPYNDGSKYTEKFQGRVTMTSDTSISTAYMELSR LRSDDTAVYYC ARGTYYYGSELFDYWGQGT VTVSSASTKGPSVFPLAPSSKSTSGGTAA LGCLVKDYFPEPVT VSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSS SLGTQTYICNVNHKPSNTKVDDKKVEPKSCDKTHTCPPCPAPEAAGGPSV FLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKT KPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGAPIEKTISK AKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQP ENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNV FSCSV MHEALHNH YTQKSLSLSPGK
297	5B08 light chain	GATATTGTTATGACTCAAAC TCCACTGTCTCTGTCCGTGACCCCGGGT CAGCCAGCGAGCATTTCTTGCAAATCCAGCCAATCTCTGGAAACCTC CACCGGCAACACGTACCTGAACTGGTATCTCCAGAAACCGGGTCAGA GCCCCGAGCTGCTGATCTACCGTGTATCTAAGCGCTTCTCCGGCGTTC CTGATCGTTTCAGCGGTTCTGGATCCGGCACCGACTTTACTCTGAAAA TCAGCCGTGTGGAAGCTGAAGACGTTGGCGTCTACTATTGTCTGCAG CTGGATTCTTACCCAAACACCTTCGGTCAAGGAAC TAACTGGAAAT TAAACGTACGGTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGA TGAGCAGTTGAAATCTGGAAGTGCCTCTGTTGTGTGCCTGCTGAATAA

		CTTCTATCCCAGAGAGGCCAAAGTACAGTGGAAGGTGGATAACGCCC TCCAATCGGGTAACTCCCAGGAGAGTGTACAGAGCAGGACAGCAA GGACAGCACCTACAGCCTCAGCAGCACCCCTGACGCTGAGCAAAGCA GACTACGAGAAACACAAAGTCTACGCCTGCGAAGTCACCCATCAGGG CCTGAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGAGAGTGT
298	5B08 heavy chain	CAGGTGCAATTGGTTCAATCTGGTGCTGAAGTAAAAAAACCGGGCGC TTCCGTTAAAGTGAGCTGCAAAGCATCTGGTTACACCTTCACTGACTA TATCATGCACTGGGTTTCGTCAGGCCCGGGCCAGGGTCTGGAGTGGA TGGGCTACATTAACCCATACAACGACGGTTCCAAATATACCGAGAAA TTCCAGGGCCGCGTCACGATGACCAGCGACACTTCTATCTCCACCGC GTACATGGAAGTGTCTAGACTGCGTTCTGACGACACCGCTGTTTACTA TTGTGCACGCGGTACCTACTACTACGGTCCACAGCTGTTTGATTACTG GGGCCAAGGTACCACGGTGACCGTAAGCTCTGCTAGCACCAAGGGCC CATCGGTCTTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGGGGGCA CAGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGGTG ACGGTGTCGTGGAAGTACAGGCGCCCTGACCAGCGGCGTGACACCTT CCCGGTGTCTTACAGTCTCAGGACTCTACTCCCTCAGCAGCGTGGT GACCGTGCCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACG TGAATCACAAGCCCAGCAACACCAAGGTGGACAAGAAAAGTTGAGCC CAAATCTTGTGACAAAAGTACACATGCCCACCGTGCCCAGCACCTG AAGCTGCAGGGGGACCGTCAGTCTTCCTCTTCCCCCAAACCCAAG GACACCCTCATGATCTCCCGGACCCCTGAGGTACATGCGTGGTGGT GGACGTGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGTGG ACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCA GTACAACAGCACGTACCGTGTGGTCAGCGTCCTACCGTCTCTGCACC AGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAA AGCCCTCGGCGCCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGC AGCCCCGAGAACCACAGGTGTACACCCTGCCCCCATCCCGGGATGAG CTGACCAAGAACCAGGTACGCTGACCTGCCTGGTCAAAGGCTTCTA TCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAG AACAACTACAAGACCACGCTCCCGTGTGGACTCCGACGGCTCCTT CTTCCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGG GGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACT ACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAA
299	5B08 light chain	DIVMTQTPLSLSVTPGQPASISCKSSQSLETSTGNTYLNWYLQKPGQSPQ LLIYRVSKRFSGVDPDRFSGSGSGTDFTLKISRVEAEDVGVYYCLQDSYP NTFGQGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAK VQWKVDNALQSGNSQESVTEQDSKSTYLSSTLTLSKADYEKHKVYA CEVTHQGLSSPVTKSFNRGEC
300	5B08 heavy chain	QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYIMHWVRQAPGQGLEW MGYINPYNDGSKYTEKFQGRVTMTSDTSISTAYMELSRLRSDDTAVYYC ARGTYYYGPQLFDYWGQGTITVTVSSASTKGPSVFPLAPSSKSTSGGTAA LGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSS SLGTQTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSV FLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKT KPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGAPIEKTISK AKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQP ENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVVFSCSVMHLEALHNH YTQKSLSLSPGK
301	5H09 light chain	GATATTGTTATGACTCAAACCTCCACTGTCTCTGTCCGTGACCCCGGGT CAGCCAGCGAGCATTTCTTGCAAATCCAGCCAATCTCTGGAATCTTCC ACCGGCAACACGTACCTGAACTGGTATCTCCAGAAACCGGGTCAGAG CCCGCAGCTGCTGATCTACCGTGTATCTAAGCGCTTCTCCGGCGTTCC TGATCGTTTCAGCGGTTCTGGATCCGGCACCGACTTTACTCTGAAAAT

		CAGCCGTGTGGAAGCTGAAGACGTTGGCGTCTACTATTGTCTGCAGC TGATCGATTACCCAGTTACCTTCGGTCAAGGAACTAAACTGGAAATT AAACGTACGGTGGCTGCACCATCTGTCTTCATCTTCCC GCCATCTGAT GAGCAGTTGAAATCTGGAAGTGCCTCTGTTGTGTGCCTGCTGAATAA CTTCTATCCCAGAGAGGGCCAAAGTACAGTGGAAGGTGGATAACGCCC TCCAATCGGGTAACTCCCAGGAGAGTGTACAGAGCAGGACAGCAA GGACAGCACCTACAGCCTCAGCAGCACCCCTGACGCTGAGCAAAGCA GACTACGAGAAACACAAAGTCTACGCCTGCGAAGTCACCCATCAGGG CCTGAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGAGAGTGT
302	5H09 heavy chain	CAGGTGCAATTGGTTCAATCTGGTGCTGAAGTAAAAAAACCGGGCGC TTCCGTTAAAGTGAGCTGCAAAGCATCTGGTTACACCTTCACTGACTA TATCATGCACTGGGTTTCGTCAGGCCCGGGCCAGGGTCTGGAGTGGA TGGGCTACATTAACCCATACAAACGACGGTTCCAAATATACCGAGAAA TTCCAGGGCCGCGTCACGATGACCAGCGACACTTCTATCTCCACCGC GTACATGGAAGTGTCTAGACTGCGTTCTGACGACACCGCTGTTTACTA TTGTGCACGCGGTACCTACTACTACGGTTCTGCACTGTTTGATTACTG GGGCCAAGGTACCACGGTGACCGTAAGCTCTGCTAGCACCAAGGGCC CATCGGTCTTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGGGGGCA CAGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGGTG ACGGTGTCGTGGAAGTCAAGGCGCCCTGACCAGCGGCGTGCACACCTT CCCGGCTGTCTACAGTCTCAGGACTCTACTCCCTCAGCAGCGTGGT GACCGTGCCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACG TGAATCACAAGCCCAGCAACACCAAGGTGGACAAGAAAGTTGAGCC CAAATCTTGTGACAAAACCTCACACATGCCACCGTGCCCAGCACCTG AAGCTGCAGGGGGACCGTCAGTCTTCTCTTCCCCCAAACCCAAAG GACACCCTCATGATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGT GGACGTGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGTGG ACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCA GTACAACAGCACGTACCGTGTGGTCAGCGTCCTACCGTCTCTGCACC AGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAA AGCCCTCGGCGCCCCCATCGAGAAAACCATCTCCAAGCCAAAGGGC AGCCCCGAGAACCACAGGTGTACACCCTGCCCCCATCCCGGGATGAG CTGACCAAGAACCAGGTACGCCTGACCTGCCTGGTCAAAGGCTTCTA TCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAG AACAACACAAGACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTT CTTCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGG GGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACT ACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAG
303	5H09 light chain	DIVMTQTPLSLSVTPGQPASISCKSSQSLESSTGNTYLNWYLQKPGQSPQ LLIYRVSKRFSGVPRDFSGSGSDFTLTKISRVEAEDVG VYYCLQLIDYP VTFGQGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAK VQWKVDNALQSGNSQESVTEQDSKSTYSLSSLTSLSKADYEKHKVYA CEVTHQGLSSPVTKSFNRGEC
304	5H09 heavy chain	QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYIMHWVRQAPGQGLEW MGYINPYNDGSKYTEKFQGRVTMTSDTSISTAYMELSRLRSDDTAVYYC ARGTYYYGSALFDYWQGTTTVTVSSASTKGPSVFPLAPSSKSTSGGTAA LGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSS SLGTQTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSV FLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNAKT KPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGAPIEKTISK AKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQP ENNYKTTTPVLDSDGSFFLYSKLTVDKSRWQQGNVFCFSVMHEALHNH YTQKSLSLSPGK

7.2.2.2 Affinity determination of selected antibodies by SPR

For the exact determination of the affinities by SPR, the selected anti-CD19 antibodies were produced by co-transfecting HEK293-EBNA cells with the mammalian expression vectors using polyethylenimine. The cells were transfected with the corresponding expression vectors in a 1:1 ratio (“vector heavy chain”: “vector light chain”) according to the standard procedure. 7 days after transfection, the antibody titer in the supernatant was measured and all titers were equilibrated to 10 µg/ml.

The Affinity (K_D) of the parental antibody 8B8 as well as its derivatives was measured by SPR using a ProteOn XPR36 instrument (Biorad) at 25°C. 7000 RU of polyclonal anti-human Fab antibody were immobilized on all 6 channels of a GLM chip by Amine coupling (NaAcetate pH4.5, 25µl/min, 240s) (vertical orientation). Each antibody-containing HEK supernatant was filtered, diluted with PBST (10 mM phosphate, 150 mM sodium chloride pH 7.4, 0.005% Tween 20) to a concentration of 10µg/ml, and then injected at a flow rate of 25 µl/minute to achieve immobilization levels between 500 and 800 response units (RU) in vertical orientation. Injection of monomeric CD19-Fc: For one-shot kinetics measurements, injection direction was changed to horizontal orientation, three-fold dilution series of purified monomeric CD19-Fc (varying concentration ranges between 150 and 6 nM) were injected simultaneously at 50µl/min along separate channels 1-4, with association times of 180s, and dissociation times of 300s. A human IgG Fc fragment (150nM) was injected in channel 5 as a negative control for specific binding to monomeric CD19-Fc. Buffer (PBST) was injected along the sixth channel to provide an “in-line” blank for referencing. An overview of the respective sensorgrams is shown in **Figure 33**. Regeneration was performed by two pulses of 10mM glycine pH 1.5 and 50mM NaOH for 30s at 90µl/min (vertical orientation). Association rate constants (k_{on}) and dissociation rate constants (k_{off}) were calculated using a simple one-to-one Langmuir binding model in ProteOn Manager v3.1 software by simultaneously fitting the association and dissociation sensorgrams. The equilibrium dissociation constant (K_D) was calculated as the ratio k_{off}/k_{on} . A summary of the kinetic and thermodynamic data is shown in **Table 60**. The dissociation constant of all affinity-matured clones was improved compared to their parental clone 8B8.

Table 60: Summary of the kinetic and thermodynamic data for the interaction between anti-CD19 huIgG1 and human CD19

clone	k_a (1/Ms)	k_d (1/s)	K_D (M)
Parental 8B8	5.66E+4	1.34E-4	2.36E-9
5H09	7.91E+4	1.50E-5	1.89E-10
7H07	7.45E+4	5.57E-5	7.47E-10

clone	ka (1/Ms)	kd (1/s)	KD (M)
2B03	6.02E+4	5.00E-5	8.31E-10
2B11	6.34E+4	3.14E-5	4.95E-10
5A07	6.98E+4	3.07E-5	4.40E-10
5B08	6.81E+4	5.26E-5	7.72E-10
5D08	8.88E+4	8.44E-5	9.51E-10

7.2.2.3 Preparation and purification of anti-CD19 IgG1 P329G LALA

The selected anti-CD19 antibodies were produced by co-transfecting HEK293-EBNA cells with the mammalian expression vectors using polyethylenimine. The cells were transfected with the corresponding expression vectors in a 1:1 ratio (“vector heavy chain”: “vector light chain”).

For the production in 500 mL shake flasks, 400 million HEK293 EBNA cells were seeded 24 hours before transfection. Before the transfection, cells were centrifuged for 5 minutes at 210 x g, and the supernatant was replaced by pre-warmed CD CHO medium. Expression vectors (200 µg of total DNA) were mixed in 20 mL CD CHO medium. After addition of 540 µL PEI, the solution was vortexed for 15 seconds and incubated for 10 minutes at room temperature. Afterwards, cells were mixed with the DNA/PEI solution, transferred to a 500 mL shake flask and incubated for 3 hours at 37°C in an incubator with a 5% CO₂ atmosphere. After the incubation, 160 mL of F17 medium was added and cells were cultured for 24 hours. One day after transfection 1 mM valproic acid and 7% Feed with supplements were added. After culturing for 7 days, the supernatant was collected by centrifugation for 15 minutes at 210 x g. The solution was sterile filtered (0.22 µm filter), supplemented with sodium azide to a final concentration of 0.01 % (w/v), and kept at 4 °C.

Purification of antibody molecules from cell culture supernatants was carried out by affinity chromatography using Protein A as described above for purification of antigen Fc fusions. The protein was concentrated and filtered prior to loading on a HiLoad Superdex 200 column (GE Healthcare) equilibrated with 20mM Histidine, 140mM NaCl solution of pH 6.0.

The protein concentration of purified antibodies was determined by measuring the OD at 280 nm, using the molar extinction coefficient calculated on the basis of the amino acid sequence. Purity and molecular weight of the antibodies were analyzed by CE-SDS in the presence and absence of a reducing agent (Invitrogen, USA) using a LabChipGXII (Caliper). The aggregate content of antibody samples was analyzed using a TSKgel G3000 SW XL analytical size-exclusion column (Tosoh) equilibrated in a 25 mM K₂HPO₄, 125 mM NaCl, 200mM L-Arginine Monohydrochloride, 0.02 % (w/v) NaN₃, pH 6.7 running buffer at 25°C (**Table 61**).

Table 61: Biochemical analysis of anti-CD19 P329G LALA IgG1 clones

Clone	Yield [mg/l]	Monomer [%]	CE-SDS (non red)
Parental 8B8	25.3	100	99.1
2B11	35.4	100	98.4
7H07	89.8	100	99.4
2B03	182	100	100
5A07	90.2	100	99.4
5D08	90.2	100	99.3
5B08	24.1	99.6	100
5H09	29.9	100	98.1

For the preparation of bispecific constructs clone 2B11 was chosen because it lacks the three deamidation hotspots.

- 5 The DNA sequence encoding part of the ectodomain (amino acid 71-254 and 71-248) of human 4-1BB ligand was synthesized according to the P41273 sequence of Uniprot database.

7.2.3 Preparation of monovalent CD19 (8B8-2B11) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains with charged residues (Construct 4.1)

- 10 The construct 4.1 was prepared as described for construct 3.1 (**Figure 30**), but using the variable region of heavy and light chain DNA sequences encoding a binder specific for CD19, clone 8B8-2B11.

- Table 62** shows the cDNA and amino acid sequences of the monovalent CD19(8B8-2B11) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule with
15 crossed CH-CL and charged residues (construct 4.1).

Table 62: cDNA and amino acid sequences of monovalent CD19(8B8-2B11) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion containing CH-CL cross with charged residues (construct 4.1). * for charged residues

SEQ ID NO:	Description	Sequence
129	Nucleotide sequence Dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 3
130	Nucleotide sequence Monomeric hu	see Table 3

	4-1BBL (71-254) – CH1*	
305	Nucleotide sequence anti- CD19(8B8-2B11) Fc hole chain	CAGGTGCAATTGGTTCAATCTGGTGCTGAAGTAAAAAA ACCGGGCGCTTCCGTTAAAGTGAGCTGCAAAGCATCTGG TTACACCTTCACTGACTATATCATGCACTGGGTTCGTCA GGCCCCGGGCCAGGGTCTGGAGTGGATGGGCTACATTA ACCCATACAACGACGGTTCCAAATATACCGAGAAATTC CAGGGCCGCGTCACGATGACCAGCGACACTTCTATCTCC ACCGCGTACATGGAAGTGTCTAGACTGCGTTCTGACGAC ACCGCTGTTTACTATTGTGCACGCGGTACCTACTACTAC GGTCCACAGCTGTTTGATTACTGGGGCCAAGGTACCACG GTGACCGTAAGCTCTGCTAGCACCAAGGGCCCCCTCCGTG TTCCCCCTGGCCCCCAGCAGCAAGAGCACCAGCGGCGG CACAGCCGCTCTGGGCTGCCTGGTCAAGGACTACTTCCC CGAGCCCCGTGACCGTGTCTGGAACAGCGGAGCCCTGA CCTCCGGCGTGCACACCTTCCCCGCCGTGCTGCAGAGTT CTGGCCTGTATAGCCTGAGCAGCGTGGTCACCGTGCCTT CTAGCAGCCTGGGACCCAGACCTACATCTGCAACGTG AACCACAAGCCCAGCAACACCAAGGTGGACAAGAAGGT GGAGCCCAAGAGCTGCGACAAAACCTACACATGCCAC CGTGCCCAGCACCTGAAGCTGCAGGGGGACCGTCAGTC TTCCTCTTCCCCCAAAACCCAAGGACACCCTCATGATC TCCCGGACCCCTGAGGTACATGCGTGGTGGTGGACGTG AGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGT GGACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGC GGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGC GTCCTCACCGTCTCTGCACCAGGACTGGCTGAATGGCAAG GAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGGCGC CCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCAGC CCCGAGAACCACAGGTGTGCACCCTGCCCCCATCCCGG GATGAGCTGACCAAGAACCAGGTGACGCTCTCGTGCGC AGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTG GGAGAGCAATGGGCAGCCGGAGAACAACCTACAAGACC ACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCCTC GTGAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCA GGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCT GCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCC GGGTAAA
277	Nucleotide sequence anti- CD19(8B8-2B11) light chain	see Table 59
115	Dimeric hu 4- 1BBL (71-254) – CL* Fc knob chain	see Table 3
116	Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
306	anti-CD19(8B8-	QVQLVQSGAEVKKPGASVKVSKASGYTFTDYIMHWVRQ APGQGLEWMGYINPYNDGSKYTEKFQGRVTMTSDTSISTA

	2B11) Fc hole chain	YMELSRRLRSDDTAVYYCARGTYYYGPQLFDYWGGGTTVT VSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVT VSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQ TYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAG GPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNW YVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLN GKEYCKVSNKALGAPIEKTISKAKGQPREPQVCTLPSPRD ELTKNQVSLSCAVKGFYPSDIAVEWESNGQPENNYKTTPP VLDSGDGSFFLVSKLTVDKSRWQQGNVFCFSVMHEALHNH YTQKSLSLSPGK
206	anti-CD19(8B8-2b11) light chain	see Table 59

7.2.4 Preparation of monovalent CD19(8B8-2B11) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains without charged residues (Construct 4.2)

- 5 The construct 4.2 was prepared as described for construct 3.2 (**Figure 30**), but using the variable region of heavy and light chain DNA sequences encoding a binder specific for CD19, clone 8B8-2B11.

- 10 **Table 63** shows the cDNA and amino acid sequences of the monovalent CD19(8B8-2B11) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule containing crossed CH-CL cross without charged residues (construct 4.2).

Table 63: cDNA and amino acid sequences of monovalent CD19(8B8-2B11) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion containing CH-CL cross without charged residues (construct 4.2).

SEQ ID NO:	Description	Sequence
165	Nucleotide sequence dimeric ligand (71-254)–CL Fc knob chain	see Table 22
166	Nucleotide sequence monomeric hu 4-1BBL (71-254) – CH1	see Table 22
305	Nucleotide sequence anti-CD19(8B8-2B11) Fc hole chain	see Table 62

277	Nucleotide sequence anti-CD19(8B8-2B11) light chain	see Table 59
117	Dimeric ligand (71-254) - CL Fc knob chain	see Table 22
118	Monomeric ligand (71-254) - CH1	see Table 22
306	anti-CD19(8B8-2B11) Fc hole chain	see Table 62
279	anti-CD19(8B8-018) light chain	see Table 59

7.2.5 Preparation of bivalent CD19(8B8-2B11) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding (Construct 4.3)

The construct 4.3 was prepared as described for construct 3.3 (Figure 30), but using the variable region of heavy and light chain DNA sequences encoding a binder specific for CD19, clone 8B8-2B11.

Table 64 shows the cDNA and amino acid sequences of the bivalent CD19 (8B8-2B11) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule (construct 4.3).

Table 64: cDNA and amino acid sequences of bivalent CD19(8B8-2B11) targeted split trimeric 4-1BB ligand Fc (kih) PGLALA fusion (construct 4.3)

SEQ ID NO:	Description	Sequence
307	Nucleotide sequence anti-CD19(8B8-2B11) Fc hole dimeric ligand chain	CAGGTGCAATTGGTTCAATCTGGTGCTGAAGTAAAAAA ACCGGGCGCTTCCGTTAAAGTGAGCTGCAAAGCATCTGG TTACACCTTCACTGACTATATCATGCACTGGGTTCGTCA GGCCCCGGGCCAGGGTCTGGAGTGGATGGGCTACATTA ACCCATACAACGACGGTTCCAAATATACCGAGAAATTC CAGGGCCGCGTCACGATGACCAGCGACACTTCTATCTCC ACCGCGTACATGGAAGTGTCTAGACTGCGTTCTGACGAC ACCGCTGTTTACTATTGTGCACGCGGTACCTACTACTAC GGTCCACAGCTGTTTGATTACTGGGGCCAAGGTACCACG GTGACCGTAAGCTCTGCTAGCACCAAGGGCCCCCTCCGTGT TCCCCCTGGCCCCCAGCAGCAAGAGCACCAGCGGCGGCAC AGCCGCTCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAG CCCGTGACCGTGTCTGGAACAGCGGAGCCCTGACCTCCG GCGTGACACCTTCCCCGCCGTGCTGCAGAGTTCTGGCCT

		<p> GTATAGCCTGAGCAGCGTGGTCACCGTGCCCTTAGCAGCC TGGGCACCCAGACCTACATCTGCAACGTGAACCACAAGCCC AGCAACACCAAGGTGGACAAGAAGGTGGAGCCCAAGAGCT GCGACAAAACCTCACACATGCCACCGTGCCAGCACCTGAA GCTGCAGGGGGACCGTCAGTCTTCTCTTCCCCCAAAC CAAGGACACCCTCATGATCTCCCGGACCCCTGAGGTCACAT GCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGGTCAA GTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCA AGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCG TGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGA ATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTC GGCGCCCCCATCGAGAAAACCATCTCCAAGCCAAAGGGC AGCCCCGAGAACCACAGGTGTGCACCCTGCCCCCATCCCG GGATGAGCTGACCAAGAACCAGGTGAGCCTCTCGTGCACA GTCAAAGGCTTCTATCCAGCGACATCGCCGTGGAGTGGGA GAGCAATGGGCAGCCGGAGAACAATAAGACCACGCCT CCCGTGCTGGACTCCGACGGCTCCTTCTTCTCGTGAGCAA GCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGTC TTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCTA CACGCAGAAGAGCCTCTCCCTGTCTCCGGGTGGAGGCGGC GGAAGCGGAGGAGGAGGATCCAGAGAGGGCCCTGAGCTG AGCCCCGATGATCCTGCTGGACTGCTGGACCTGCGGCAGG GCATGTTTGCTCAGCTGGTGGCCCAGAACGTGCTGCTGATC GATGGCCCCCTGTCTGGTACAGCGATCCTGGACTGGCTG GCGTGTCAGTACAGGCGGCCTGAGCTACAAAGAGGACAC CAAAGAACTGGTGGTGGCCAAGGCCGGCGTGTACTACGTG TTCTTTCAGCTGGAAGTGCAGGAGTGGTGGCCGGCGAAG GATCTGGCTCTGTGTCTCTGGCCCTGCATCTGCAGCCTCTG AGAAGCGCTGCTGGCGCTGCAGCTCTGGCACTGACAGTGG ATCTGCCTCCTGCCAGCTCCGAGGCCCGGAATAGCGCATTT GGGTTTCAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGA GGCTGGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACA CGCCTGGCAGCTGACACAGGGCGCTACAGTGTGGGCCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCTCC AAGAAGCGAAGGCGGAGGCGGATCTGGCGGCGGAGGATC TAGAGAGGGACCCGAAGTGTCCCTGACGATCCAGCCGGG CTGCTGGATCTGAGACAGGGAATGTTGCCCCAGCTGGTGG CTCAGAATGTGCTGCTGATTGACGGACCTCTGAGCTGGTAC TCCGACCCAGGGCTGGCAGGGGTGTCCCTGACTGGGGGAC TGTCTACAAAGAAAGATACAAAAGAACTGGTGGTGGCTAAA GCTGGGGTGTACTATGTGTTTTTTCAGCTGGAAGTGGGCG GGTGGTGGCTGGGGAGGGCTCAGGATCTGTGTCCCTGGCT CTGCATCTGCAGCCACTGCGCTCTGCTGCTGGCGCAGCTG CACTGGCTCTGACTGTGGACCTGCCACCAGCCTCTAGCGAG GCCAGAAACAGCGCCTTCGGGTCCAAGGACGCCTGCTGC ATCTGAGCGCCGGACAGCGCCTGGGAGTGCATCTGCATAC TGAAGCCAGAGCCCGGCATGCTTGGCAGCTGACTCAGGGG GCAACTGTGCTGGGACTGTTTCGCGTGACACCTGAGATCCC TGCCGGACTGCCAAGCCCTAGATCAGAA </p>
308	Nucleotide sequence anti- CD19(8B8-2B11) Fc knob	<p> CAGGTGCAATTGGTTCAATCTGGTGTGCTGAAGTAAAAAA ACCGGGCGCTTCCGTTAAAGTGAGCTGCAAAGCATCTGG TTACACCTTCACTGACTATATCATGCACTGGGTTCGTCA GGCCCCGGGCCAGGGTCTGGAGTGGATGGGCTACATTA ACCCATACAACGACGGTTCCAAATATACCGAGAAATTC </p>

	monomeric ligand	CAGGGCCGCGTCACGATGACCAGCGACACTTCTATCTCC ACCGCGTACATGGAAGTGTCTAGACTGCGTTCTGACGAC ACCGCTGTTTACTATTGTGCACGCGGTACCTACTACTAC GGTCCACAGCTGTTTGATTACTGGGGCCAAGGTACCACG GTGACCGTAAGCTCTGCTAGCACCAAGGGCCCATCGGTCT TCCCCCTGGCACCCTCCTCCAAGAGCACCTCTGGGGGAC AGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAA CCGGTGACGGTGTCGTGGAAGTCAAGGCGCCCTGACCAGCG GCGTGACACCTTCCCGGCTGTCTACAGTCTCAGGACTC TACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGCAGCTT GGGCACCCAGACCTACATCTGCAACGTGAATCACAAGCCCA GCAACACCAAGGTGGACAAGAAAGTTGAGCCCAATCTTGT GACAAAAGTACACATGCCACCGTGCCAGCACCTGAAGC TGCAGGGGGACCGTCAGTCTTCTCTTCCCCCAAAACCCA AGGACACCCTCATGATCTCCCGGACCCCTGAGGTACATGC GTGGTGGTGGACGTGAGCCACGAAGACCCTGAGGTCAAGT TCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCAA GACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGT GTGGTCAGCGTCCTCACCCTCTGCACCAGGACTGGCTGA ATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTC GGCGCCCCCATCGAGAAAACCATCTCCAAGGCCAAAGGGC AGCCCCGAGAACCACAGGTGTACACCCTGCCCCCTGCAG AGATGAGCTGACCAAGAACCAGGTGTCCCTGTGGTGTCTGG TCAAGGGCTTCTACCCCAGCGATATCGCCGTGGAGTGGGA GAGCAACGGCCAGCCTGAGAACAACCTACAAGACCACCCCC CCTGTGCTGGACAGCGACGGCAGCTTCTTCTGTACTCCAA ACTGACCGTGGACAAGAGCCGGTGGCAGCAGGGCAACGTG TTCAGCTGCAGCGTGATGCACGAGGCCCTGCACAACCACTA CACCCAGAAGTCCCTGAGCCTGAGCCCCGGCGGAGGCGG CGGAAGCGGAGGAGGAGGATCCAGAGAGGGGCCCTGAGCT GAGCCCCGATGATCCTGCTGGACTGCTGGACCTGCGGCAG GGCATGTTTGCTCAGCTGGTGGCCAGAACGTGCTGCTGAT CGATGGCCCCCTGTCTGGTACAGCGATCCTGGACTGGCT GGCGTGTCACTGACAGGCGGCCTGAGCTACAAAGAGGACA CCAAAGAACTGGTGGTGGCCAAGGCCGGCGTGTACTACGT GTTCTTTCAGCTGGAAGTGCAGGAGAGTGGTGGCCGGCGAA GGATCTGGCTCTGTGTCTCTGGCCCTGCATCTGCAGCCTCT GAGAAGCGCTGCTGGCGCTGCAGCTCTGGCACTGACAGTG GATCTGCCTCCTGCCAGCTCCGAGGCCCGGAATAGCGCAT TGGGTTTCAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAG AGGCTGGGAGTGATCTGCACACAGAGGCCAGGGCTAGAC ACGCCTGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCT GTTCAAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCTC CAAGAAGCGAA
277	Nucleotide sequence anti- CD19(8B8-018) light chain	see Table 59
309	anti-CD19(8B8- 2B11) Fc hole dimeric ligand chain	QVQLVQSGAEVKKPGASVKVSKASGYTFTDYIMHWVRQ APGQGLEWMGYINPYNDGSKYTEKFQGRVTMTSDTSISTA YMELSRRLRSDDTAVYYCARGTYYYGPQLFDYWQGTTVT VSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSW NSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVN

		<i>HKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPP KPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNA KTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALG APIEKTISKAKGQPREPQVCTLPSPRDELTKNQVSLSCAVKGFY PSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLVSKLTVDKSR WQQGNVFSCSVMHEALHNHYTQKSLSLSPGGGGGSGGGGSR EGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDP GLAGVSLTGGLSYKEDTKELVVAAGVYYYVFFQLELRRVAGE GSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQ GRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTP EIPAGLPSRSEGGGGSGGGGSREGPELSPDDPAGLLDLRQG MFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKEL VVAAGVYYYVFFQLELRRVAGEGSGSVSLALHLQPLRSAAGA AALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTE ARARHAWQLTQGATVLGLFRVTP EIPAGLPSRSE</i>
310	anti-CD19(8B8-2B11) Fc knob monomeric ligand	<i>QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYIMHWVRQ APGQGLEWMGYINPYNDGSKYTEKFQGRVTMTSDTSISTA YMELSRLRSDDTAVYYCARGTYYYGPQLFDYWGGQTTVT VSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPTVSW NSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVN HKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPP KPKDTLMISRTPEVTCVVDVSHEDPEVKFNWYVDGVEVHNA KTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALG APIEKTISKAKGQPREPQVYTLPPCRDELTKNQVSLWCLVKGF YPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKS RWQQGNVFSCSVMHEALHNHYTQKSLSLSPGGGGGSGGGGS REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSD PGLAGVSLTGGLSYKEDTKELVVAAGVYYYVFFQLELRRVAG EGS SVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGF QGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVT PEIPAGLPSRSE</i>
279	anti-CD19(8B8-018) light chain	see Table 59

7.2.6 Preparation of monovalent CD19(8B8-2B11) targeted 4-1BB ligand (71-248) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains with charged residues (Construct 4.4)

- 5 The construct 4.4 was prepared as described for construct 3.4 (Figure 30), but using the variable region of heavy and light chain DNA sequences encoding a binder specific for CD19, clone 8B8-2B11.

Table 65 shows the cDNA and amino acid sequences of the monovalent CD19(8B8-2B11) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion antigen binding molecule with
 10 crossed CH-CL and charged residues (construct 4.4).

Table 65: cDNA and amino acid sequences of monovalent CD19(8B8-2B11) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion containing CH-CL cross with charged residues (construct 4.4). * charged residues

SEQ ID NO:	Description	Sequence
169	Nucleotide sequence dimeric ligand (71-248) – CL* Fc knob chain	see Table 24
170	Nucleotide sequence monomeric hu 4-1BBL (71-248) – CH1*	see Table 24
305	Nucleotide sequence anti-CD19(8B8-2B11) Fc hole chain	see Table 62
277	Nucleotide sequence anti-CD19(8B8-2B11) light chain	see Table 59
119	Dimeric ligand (71-248) – CL* Fc knob chain	see Table 24
120	Monomeric ligand (71-248)-CH1*	see Table 24
306	anti-CD19(8B8-2B11) Fc hole chain	see Table 62
279	anti-CD19(8B8-2B11) light chain	see Table 59

5 7.2.7 Preparation of monovalent CD19(8B8-2B11) targeted 4-1BB ligand (71-248) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains without charged residues (Construct 4.5)

The construct 4.5 was prepared as described for construct 3.5 (Figure 30), but using the variable region of heavy and light chain DNA sequences encoding a binder specific for CD19, clone 8B8-2B11.

Table 66 shows the cDNA and amino acid sequences of the monovalent CD19(8B8-2B11) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion antigen binding molecule containing crossed CH-CL cross without charged residues (construct 4.5).

5 **Table 66: cDNA and amino acid sequences of monovalent CD19(8B8-2B11) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion containing CH-CL cross without charged residues (construct 4.5).**

SEQ ID NO:	Description	Sequence
171	Nucleotide sequence dimeric ligand (71-248) – CL Fc knob chain	see Table 25
172	Nucleotide sequence monomeric ligand (71-248)-CH1	see Table 25
305	Nucleotide sequence anti-CD19(8B8-2B11) Fc hole chain	see Table 62
277	Nucleotide sequence anti-CD19(8B8-2B11) light chain	see Table 59
173	Dimeric ligand (71-248) – CL Fc knob chain	see Table 25
174	Monomeric ligand (71-248)-CH1	see Table 25
306	anti-CD19(8B8-2B11) Fc hole chain	see Table 62
279	anti-CD19(8B8-2B11) light chain	see Table 59

7.2.8 Preparation of bivalent CD19(8B8-2B11) targeted 4-1BB ligand (71-248) trimer-containing Fc (kih) fusion antigen binding (Construct 4.6)

10 The construct 4.6 was prepared as described for construct 3.6 (Figure 30), but using the variable region of heavy and light chain DNA sequences encoding a binder specific for CD19, clone 8B8-2B11.

Table 67 shows the cDNA and amino acid sequences of the bivalent CD19(8B8-2B11) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion antigen binding molecule (construct 3.6).

Table 67: cDNA and amino acid sequences of bivalent CD19(8B8-2B11) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion (construct 4.6)

SEQ ID NO:	Description	Sequence
311	Nucleotide sequence anti-CD19(8B8-2B11) Fc hole dimeric ligand (71-248) chain	CAGGTGCAATTGGTTCAATCTGGTGCTGAAGTAAAAAA ACCGGGCGCTTCCGTTAAAGTGAGCTGCAAAGCATCTGG TTACACCTTCACTGACTATATCATGCACTGGGTTCGTCA GGCCCCGGGCCAGGGTCTGGAGTGGATGGGCTACATTA ACCCATACAACGACGGTTCCAAATATACCGAGAAATTC CAGGGCCGCGTCACGATGACCAGCGACACTTCTATCTCC ACCGCGTACATGGAAGTGTCTAGACTGCGTTCTGACGAC ACCGCTGTTTACTATTGTGCACGCGGTACCTACTACTAC GGTCCACAGCTGTTTGATTACTGGGGCCAAGGTACCACG GTGACCGTAAGCTCTGCTAGCACCAAGGGCCCCCTCCGTGT TCCCCCTGGCCCCCAGCAGCAAGAGCACCAGCGGCGGCAC AGCCGCTCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAG CCCGTGACCGTGTCTTGAACAGCGGAGCCCTGACCTCCG GCGTGACACCTTCCCCGCCGTGCTGCAGAGTTCTGGCCT GTATAGCCTGAGCAGCGTGGTCACCGTGCCCTCTAGCAGCC TGGGCACCCAGACCTACATCTGCAACGTGAACCACAAGCCC AGCAACACCAAGGTGGACAAGAAGGTGGAGCCCAAGAGCT GCGACAAAACCTCACACATGCCACCGTGCCAGCACCTGAA GCTGCAGGGGGACCGTCAGTCTTCTCTTCCCCCAAAACC CAAGGACACCCTCATGATCTCCCGGACCCCTGAGGTACAT GCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGGTCAA GTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCA AGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCG TGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGA ATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTC GGCGCCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGC AGCCCCGAGAACCACAGGTGTGACCCTGCCCCCATCCCG GGATGAGCTGACCAAGAACCAGGTGAGCCTCTCGTGCGCA GTCAAAGGCTTCTATCCAGCGACATCGCCGTGGAGTGGGA GAGCAATGGGCAGCCGGAGAACAATAAAGACCACGCCT CCCGTGCTGGACTCCGACGGCTCCTTCTTCTCTCGTGAGCAA GCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGTC TTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACTA CACGCAGAAGAGCCTCTCCCTGTCTCCGGGTGGAGGCGGC GGAAGCGGAGGAGGAGGATCCAGAGAGGGGCCCTGAGCTG AGCCCTGATGATCCTGCCGGACTGCTGGACCTGCGGCAGG GAATGTTTGCCCAGCTGGTGGCCCAGAACGTGCTGCTGATC GATGGCCCCCTGTCTGGTACAGCGATCCTGGACTGGCTG GCGTGTCATGACAGGCGGCCTGAGCTACAAAGAGGACAC CAAAGAACTGGTGGTGGCCAAGGCCGGCGTGTACTACGTG TTCTTTCAGCTGGAAGTGCAGGAGAGTGGTGGCCGGCGAAG GATCTGGCTCTGTGTCTCTGGCCCTGCATCTGCAGCCTCTG AGATCTGCTGCTGGCGCCGCTGCTCTGGCACTGACAGTGG

		<p> ATCTGCCTCCTGCCAGCAGCGAGGCCCGGAATAGCGCATTT GGGTTTCAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGA GGCTGGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACA CGCCTGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCTG TTCAGAGTGACCCCCGAGATTCCAGCAGGCCTGGGAGGCG GCGGATCTGGCGGCGGAGGATCTAGAGAAGGACCCGAGCT GTCCCCCGACGATCCCGCTGGGCTGCTGGATCTGAGACAG GGCATGTTCGCTCAGCTGGTGGCTCAGAATGTGCTGCTGAT TGACGGACCTCTGAGCTGGTACTCCGACCCAGGGCTGGCA GGGGTGTCCCTGACTGGGGGACTGTCCTACAAAGAAGATAC AAAAGAACTGGTGGTGGCTAAAGCTGGGGTGTACTATGTGT TTTTTCAGCTGGAACCTGAGGCGGGTGGTGGCTGGGGAGGG CTCAGGATCTGTGTCCCTGGCTCTGCATCTGCAGCCACTGC GCTCTGCAGCAGGGGCTGCAGCACTGGCCCTGACTGTGGA CCTGCCCCCAGCTTCTTCCGAGGCCAGAAACAGCGCCTTCG GGTTCCAAGGACGCCTGCTGCATCTGAGCGCCGGACAGCG CCTGGGAGTGATCTGCATACTGAAGCCAGAGCCCGGCAT GCTTGGCAGCTGACTCAGGGGGCAACTGTGCTGGGACTGT TTCGCGTGACACCTGAGATCCCAGCCGGGCTC </p>
312	Nucleotide sequence anti- CD19(8B8-2B11) Fc knob monomeric (71- 248) ligand	<p> CAGGTGCAATTGGTTCAATCTGGTGCTGAAGTAAAAAA ACCGGGCGCTTCCGTTAAAGTGAGCTGCAAAGCATCTGG TTACACCTTCACTGACTATATCATGCACTGGGTTCGTCA GGCCCCGGGCCAGGGTCTGGAGTGGATGGGCTACATTA ACCCATACAACGACGGTTCCAAATATACCGAGAAATTC CAGGGCCGCGTCACGATGACCAGCGACACTTCTATCTCC ACCGCGTACATGGAACCTGTCTAGACTGCGTTCTGACGAC ACCGCTGTTTACTATTGTGCACGCGGTACCTACTACTAC GGTCCACAGCTGTTTGATTACTGGGGCCAAGGTACCACG GTGACCGTAAGCTCTGCTAGCACCAAGGGCCCATCGGTCT TCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGGGGGAC AGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAA CCGGTGACGGTGTCGTGGAACCTCAGGCGCCCTGACCAGCG GCGTGACACCTTCCCGGCTGTCTACAGTCTCAGGACTC TACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGCAGCTT GGGCACCCAGACCTACATCTGCAACGTGAATCACAAGCCCA GCAACACCAAGGTGGACAAGAAAGTTGAGCCCAAATCTTGT GACAAAACCTCACACATGCCCAACCGTGCCCAGCACCTGAAGC TGCAGGGGGACCGTCAGTCTTCTTCCCCCCCCAAAACCCA AGGACACCCCTCATGATCTCCCGGACCCCTGAGGTACATGC GTGGTGGTGGACGTGAGCCACGAAGACCCTGAGGTCAAGT TCAACTGGTACGTGGACGGCGTGAGGTGCATAATGCCAA GACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGT GTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGA ATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTC GGCGCCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGC AGCCCCGAGAACCACAGGTGTACACCCTGCCCCCTGCAG AGATGAGCTGACCAAGAACCAGGTGTCCCTGTGGTGTCTGG TCAAGGGCTTCTACCCCAGCGATATCGCCGTGGAGTGGGA GAGCAACGGCCAGCCTGAGAACTACAAGACCACCCCC CCTGTGCTGGACAGCGACGGCAGCTTCTTCTGTACTCCAA ACTGACCGTGGACAAGAGCCGGTGGCAGCAGGGCAACGTG TTCAGCTGCAGCGTGATGCACGAGGCCCTGCACAACCACTA CACCCAGAAAGTCCCTGAGCCTGAGCCCCGGCGGAGGCGG CGGAAGCGGAGGAGGAGGATCCAGAGAGGGCCCTGAGCT </p>

		<p>GAGCCCTGATGATCCTGCCGGACTGCTGGACCTGCGGCAG GGAATGTTTGCCAGCTGGTGGCCCAGAACGTGCTGCTGAT CGATGGCCCCCTGTCTGGTACAGCGATCCTGGACTGGCT GGCGTGTCACTGACAGGCGGCCTGAGCTACAAAGAGGACA CCAAAGAACTGGTGGTGGCCAAGGCCGGCGTGTACTACGT GTTCTTTCAGCTGGAAGTGCAGGAGAGTGGTGGCCGGCGAA GGATCTGGCTCTGTGTCTCTGGCCCTGCATCTGCAGCCTCT GAGATCTGCTGCTGGCGCCGCTGCTCTGGCACTGACAGTG GATCTGCCTCCTGCCAGCAGCGAGGCCCGGAATAGCGCAT TTGGGTTTCAAGGCAGGCTGCTGCACCTGTCTGCCGGCCA GAGGCTGGGAGTGCATCTGCACACAGAGGCCAGGGCTAGA CACGCCTGGCAGCTGACACAGGGCGCTACAGTGCTGGGCC TGTTCAAGAGTGACCCCCGAGATTCCTGCCGGGCTC</p>
277	Nucleotide sequence anti- CD19(8B8-2B11) light chain	see Table 59
313	anti-CD19(8B8- 2B11) Fc hole dimeric ligand (71-248) chain	<p>QVQLVQSGAEVKKPGASVKVSKASGYTFTDYIMHWVRQ APGQGLEWMGYINPYNDGSKYTEKFQGRVTMTSDTSISTA YMELSRRLRSDDTAVYYCARGTYYYGPQLFDYWGQGTTVT VSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSW NSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVN HKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPP KPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNA KTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALG APIEKTISKAKGQPREPQVCTLPSSRDELTKNQVSLSCAVKGFY PSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLVSKLTVDKSR WQQGNVFSCSVMHEALHNHYTQKSLSLSPGGGGGSGGGGSR EGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSDP GLAGVSLTGGLSYKEDTKELVVAKAGVYVFFQLELRRVVAGE GSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGFQ GRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTP EIPAGLGGGGSGGGGSGREGPELSPDDPAGLLDLRQGMFAQL VAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAKA GVYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAAALALT VDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLHTEARARHA WQLTQGATVLGLFRVTP EIPAGL</p>
314	anti-CD19(8B8- 2B11) Fc knob monomeric (71- 248) ligand	<p>QVQLVQSGAEVKKPGASVKVSKASGYTFTDYIMHWVRQ APGQGLEWMGYINPYNDGSKYTEKFQGRVTMTSDTSISTA YMELSRRLRSDDTAVYYCARGTYYYGPQLFDYWGQGTTVT VSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSW NSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVN HKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPP KPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNA KTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALG APIEKTISKAKGQPREPQVYTLPPCRDELTKNQVSLWCLVKGF YPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLVSKLTVDKS RWQQGNVFSCSVMHEALHNHYTQKSLSLSPGGGGGSGGGGS REGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYSD PGLAGVSLTGGLSYKEDTKELVVAKAGVYVFFQLELRRVVAG EGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFGF QGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRVT</p>

		<i>PEIPAGL</i>
279	anti-CD19(8B8-018) light chain	see Table 59

7.3 Preparation of untargeted split trimeric 4-1BB ligand Fc fusion and human IgG as control molecules

7.3.1 Preparation of untargeted human 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (Control molecules)

These control molecules were prepared as described above for the CD19 targeted construct 3.1 (termed control B), 3.3 (termed control C), 3.4 (termed control D) and 3.5 (termed control E) with the only difference that the anti-CD19 binder (VH-VL) was replaced by a germline control, termed DP47, not binding to the antigen (see Figure 30).

Table 68 shows, respectively, the cDNA and amino acid sequences of the monovalent DP47-untargeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion containing crossed CH-CL with charged residues, control B.

Table 69 shows, respectively, the cDNA and amino acid sequences of the bivalent DP47-untargeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion, control C.

Table 70 shows, respectively, the cDNA and amino acid sequences of the monovalent DP47-untargeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion containing CH-CL cross with charged residues, control D.

Table 71 shows, respectively, the cDNA and amino acid sequences of the monovalent DP47-untargeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion without charged residues in the CH-CL cross, control E.

Table 68: cDNA and amino acid sequences of monovalent DP47 untargeted split trimeric human 4-1BB ligand (71-254) Fc (kih) fusion with CH-CL cross and with charged residues (control B). * charges residues

SEQ ID NO:	Description	Sequence
96	nucleotide sequence dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 3
97	nucleotide sequence monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
79	nucleotide sequence DP47 Fc hole chain	see Table 18
80	nucleotide sequence DP47 light chain	see Table 18
98	Dimeric hu 4-1BBL (71-254) – CL* Fc	see Table 3

	knob chain	
99	Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
81	DP47 Fc hole chain	see Table 18
82	DP47 light chain	see Table 18

Table 69: cDNA and amino acid sequences of bivalent DP47 untargeted split trimeric human 4-1BB ligand (71-254) Fc (kih) fusion (control C).

SEQ ID NO:	Description	Sequence
177	nucleotide sequence DP47 Fc hole chain fused to dimeric hu 4-1BBL (71-254)	see Table 27
178	nucleotide sequence DP47 Fc knob chain fused to monomeric hu 4-1BBL (71-254)	see Table 27
80	nucleotide sequence DP47 light chain	see Table 18
179	DP47 Fc hole chain fused to dimeric hu 4-1BBL (71-254)	see Table 27
180	DP47 Fc knob chain fused to monomeric hu 4-1BBL (71-254)	see Table 27
82	DP47 light chain	see Table 18

Table 70: cDNA and amino acid sequences of monovalent DP47 untargeted split trimeric human 4-1BB ligand (71-248) Fc (kih) fusion with CH-CL cross and with charged residues (control D). * charged residues

SEQ ID NO:	Description	Sequence
169	nucleotide sequence dimeric hu 4-1BBL (71-248) – CL* Fc knob chain	see Table 24
170	nucleotide sequence monomeric hu 4-1BBL (71-248) – CH1*	see Table 24
79	nucleotide sequence DP47 Fc hole chain	see Table 18
80	nucleotide sequence DP47 light chain	see Table 18
119	Dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 24
120	Monomeric hu 4-1BBL (71-254) – CH1*	see Table 24
81	DP47 Fc hole chain	see Table 18
82	DP47 light chain	see Table 18

Table 71: cDNA and amino acid sequences of monovalent DP47 untargeted split trimeric human 4-1BB ligand (71-248) Fc (kih) fusion with CH-CL cross and without charged residues (control E).

SEQ ID NO:	Description	Sequence
171	nucleotide sequence dimeric hu 4-1BBL (71-248) – CL Fc knob chain	see Table 25
172	nucleotide sequence monomeric hu 4-1BBL (71-248) – CH1	see Table 25
79	nucleotide sequence DP47 Fc hole chain	see Table 18
80	nucleotide sequence DP47 light chain	see Table 18
173	Dimeric hu 4-1BBL (71-248) – CL Fc knob chain	see Table 25
174	Monomeric hu 4-1BBL (71-248) – CH1	see Table 25
81	DP47 Fc hole chain	see Table 18
82	DP47 light chain	see Table 18

5 7.3.2 Antibodies as Control Molecules

Two control human IgG1 containing PGLALA were prepared.

Table 72 shows the cDNA and amino acid sequences of the anti-CD19 huIgG1 PGLALA (clone 8B8-018), i.e. control G.

Table 73 shows the cDNA and amino acid sequences of germline control DP47 huIgG1 PGLALA (control F).

Table 72: cDNA and amino acid sequences of anti-CD19(8B8-018) huIgG1 PGLALA (control G)

SEQ ID NO:	Description	Sequence
315	nucleotide sequence CD19(8B8-018) heavy chain (huIgG1 PGLALA)	CAGGTCCAGCTGGTGCAGTCCGGCGCCGAGGT CAAGAAACCCGGGGCTTCTGTGAAGGTTTCAT GCAAGGCAAGCGGATACACCTTCACCGACTAT ATCATGCATTGGGTCAGGCAGGCCCTGGCCA AGGTCTCGAATGGATGGGCTACATTAACCCAT ATAATGATGGCTCCAAATACACCGAGAAGTTT CAGGGAAGAGTCACTATGACATCTGACACCAG TATCAGCACTGCTTACATGGAGCTGTCCCGCC TTCGGTCTGATGACACCGCAGTGTATTACTGT

		<p>GCCAGGGGCACATATTACTACGGCTCAGCTCT GTTCTGACTATTGGGGGCAGGGAACCCACAGTAA CCGTGAGCTCCGCAAGTACTAAGGGCCCATCG GTCTTCCCCCTGGCACCCCTCCTCCAAGAGCAC CTCTGGGGGGCACAGCGGCCCTGGGCTGCCTGG TCAAGGACTACTTCCCCGAACCGGTGACGGTG TCGTGGAACCTCAGGCGCCCTGACCAGCGGCGT GCACACCTTCCCGGCTGTCCTACAGTCCTCAG GACTCTACTCCCTCAGCAGCGTGGTGACCGTG CCCTCCAGCAGCTTGGGCACCCAGACCTACAT CTGCAACGTGAATCACAAGCCCAGCAACACCA AGGTGGACAAGAAAGTTGAGCCCAAATCTTGT GACAAAACCTCACACATGCCACCCGTGCCCAGC ACCTGAAGCAGCTGGGGGACCGTCAGTCTTCC TCTTCCCCCAAAACCCAAGGACACCCCTCATG ATCTCCCGGACCCCTGAGGTCACATGCGTGGT GGTGGACGTGAGCCACGAAGACCCCTGAGGTC AAGTTCAACTGGTACGTGGACGGCGTGAGGT GCATAATGCCAAGACAAAGCCGCGGGAGGAG CAGTACAACAGCACGTACCGTGTGGTCAGCGT CCTCACCGTCTTGCACCAGGACTGGCTGAATG GCAAGGAGTACAAGTGCAAGGTCTCCAACAA AGCCCTCGGAGCCCCCATCGAGAAAACCATCT CCAAAGCCAAAGGGCAGCCCCGAGAACCACA GGTGTACACCCTGCCCCCATCCCGGGATGAGC TGACCAAGAACCAGGTCAGCCTGACCTGCCTG GTCAAAGGCTTCTATCCCAGCGACATCGCCGT GGAGTGGGAGAGCAATGGGCAGCCGGAGAAC AACTACAAGACCACGCCTCCCGTGCTGGACTC CGACGGCTCCTTCTTCTCTACAGCAAGCTCAC CGTGGACAAGAGCAGGTGGCAGCAGGGGAAC GTCTTCTCATGCTCCGTGATGCATGAGGCTCTG CACAACCACTACACGCAGAAGAGCCTCTCCCT GTCCCCGGGCAAA</p>
204	nucleotide sequence CD19(8B8-018) light chain	see Table 47
316	CD19(8B8-018) heavy chain (huIgG1 PGLALA)	<p>QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYI MHWVRQAPGQGLEWMGYINPYNDGSKYTEKF QGRVTMTSDTSISTAYMELSRLSDDTAVYYCA RGTYYYGSALFDYWQGTTVTVSSASTKGPSVF PLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNS GALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLG TQTYICNVNHKPSNTKVDKKVEPKSCDKHTCP PCPAPEAAGGPSVFLFPPKPKDTLMISRTPEVTCV VVDVSHEDPEVKFNWYVDGVEVHNAKTKPREE QYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNK ALGAPIEKTISKAKGQPREPQVYTLPPSRDELTK NQVSLTCLVKGFYPSDIAVEWESNGQPENNYKT TPPVLDSDGSFFLYSKLTVDKSRWQQGNVFS VMHEALHNHYTQKSLSLSPGK</p>
206	CD19(8B8-018) light	see Table 47

	chain	
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Table 73: cDNA and amino acid sequences of germline control DP47 huIgG1 PGLALA (control F)

SEQ ID NO:	Description	Sequence
181	nucleotide sequence DP47 heavy chain (hu IgG1 PGLALA)	see Table 29
80	DP47 light chain	see Table 18
182	DP47 heavy chain (hu IgG1 PGLALA)	see Table 29
82	DP47 light chain	see Table 18

5 **7.4 Production of CD19-targeted split trimeric 4-1BB ligand Fc fusion antigen binding molecules and their control molecules**

The targeted and untargeted split trimeric 4-1BB ligand Fc (kih) fusion antigen binding molecule encoding sequences were cloned into a plasmid vector, which drives expression of the insert from an MPSV promoter and contains a synthetic polyA sequence located at the 3' end of the CDS. In addition, the vector contains an EBV OriP sequence for episomal maintenance of the plasmid.

The split trimeric 4-1BB ligand Fc (kih) fusion antigen binding molecule was produced by co-transfecting HEK293-EBNA cells with the mammalian expression vectors using polyethylenimine. The cells were transfected with the corresponding expression vectors. For variants 1,2,4,5 and it's control B, D and E, at a 1:1:1:1 ratio ("vector dimeric ligand-CL- knob chain": "vector monomeric ligand fusion-CH1": "vector anti-CD19 Fab-hole chain": "vector anti-CD19 light chain"). For variant 3, 6 and it's control C, at a 1:1:1 ratio ("vector huIgG1 Fc hole dimeric ligand chain": "vector huIgG1 Fc knob monomeric ligand chain": "vector anti-CD19 light chain"). Human IgGs, used as control in the assay, were produced as for the bispecific construct (for transfection only a vector for light and a vector for heavy chain were used at a 1:1 ratio).

For production in 500 mL shake flasks, 300 million HEK293 EBNA cells were seeded 24 hours before transfection. For transfection cells were centrifuged for 10 minutes at 210 x g, and the supernatant was replaced by 20mL pre-warmed CD CHO medium. Expression vectors (200 µg of total DNA) were mixed in 20 mL CD CHO medium. After addition of 540 µL PEI, the solution was vortexed for 15 seconds and incubated for 10 minutes at room temperature.

Afterwards, cells were mixed with the DNA/PEI solution, transferred to a 500 mL shake flask and incubated for 3 hours at 37 °C in an incubator with a 5% CO₂ atmosphere. After the incubation, 160 mL of Excell medium supplemented with 6 mM L-Glutamine, 5g/L PEPSOY and 1.2mM valproic acid was added and cells were cultured for 24 hours. One day after

5 transfection 12% Feed (amino acid and glucose) were added. After culturing for 7 days, the supernatant was collected by centrifugation for 30-40 minutes at least 400 x g. The solution was sterile filtered (0.22 µm filter), supplemented with sodium azide to a final concentration of 0.01 % (w/v), and kept at 4 °C.

The split trimeric 4-1BB ligand Fc (kih) fusion antigen binding molecule, as well as the

10 IgG, was purified from cell culture supernatants by affinity chromatography using Protein A, followed by size exclusion chromatography. For affinity chromatography, the supernatant was loaded on a MabSelect Sure column (CV = 5-15 mL, resin from GE Healthcare) equilibrated with sodium phosphate (20 mM), sodium citrate (20 mM) buffer (pH 7.5). Unbound protein was removed by washing with at least 6 column volumes of the same buffer. The bound protein was

15 eluted using either a linear gradient (20 CV) or a step elution (8 CV) with 20 mM sodium citrate, 100 mM sodium chloride, 100 mM glycine buffer (pH 3.0). For the linear gradient an additional 4 column volumes step elution was applied.

The pH of collected fractions was adjusted by adding 1/10 (v/v) of 0.5M sodium phosphate, pH8.0. The protein was concentrated prior to loading on a HiLoad Superdex 200 column (GE

20 Healthcare) equilibrated with 20 mM histidine, 140 mM sodium chloride, 0.01% (v/v) Tween20 solution of pH 6.0.

The protein concentration was determined by measuring the optical density (OD) at 280 nm, using a molar extinction coefficient calculated on the basis of the amino acid sequence. Purity and molecular weight of the targeted trimeric 4-1BB ligand Fc (kih) fusion was analyzed

25 by SDS-PAGE in the presence and absence of a reducing agent (5 mM 1,4-dithiotreitol) and staining with Coomassie SimpleBlue™ SafeStain (Invitrogen USA). The aggregate content of samples was analyzed using a TSKgel G3000 SW XL analytical size-exclusion column (Tosoh) equilibrated in 25 mM K₂HPO₄, 125 mM NaCl, 200 mM L-arginine monohydrochloride, 0.02 % (w/v) NaN₃, pH 6.7 running buffer at 25°C.

30 **Table 74** summarizes the yield and final monomer content of the CD19 targeted split trimeric 4-1BB ligand Fc (kih) fusion antigen molecules.

Table 74: Biochemical analysis of CD19 targeted split trimeric 4-1BB ligand Fc (kih) fusion antigen binding molecules

Construct	Monomer [%] (SEC)	Yield [mg/l]
monovalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-254) Fc fusion anitgcontaining CH-CL cross with charged residues (construct 3.1)	98	8.6
bivalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-254) Fc fusion (construct 3.3)	100	11.3
monovalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-248) Fc fusion containing CH-CL cross with charged residues (construct 3.4)	99	11.5
monovalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-248) Fc fusion containing CH-CL cross without charged residues (construct 3.5)	97	13.3
bivalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-248) Fc fusion (construct 3.6)	96	19.9
monovalent CD19(8B8-2B11) targeted split trimeric 4-1BB ligand (71-248) Fc fusion containing CH-CL cross with charged residues (construct 4.4)	99.2	21.2

Table 75 summarizes the yield and final monomer content of the DP47 untargeted split trimeric 4-1BB ligand Fc (kih) fusion, both monovalent (control B, D and E) and bivalent (control C).

Table 75: Biochemical analysis of DP47 untargeted split trimeric 4-1BB ligand Fc (kih) fusion

Construct	Monomer [%] (SEC)	Yield [mg/l]
monovalent DP47-untargeted split trimeric human 4-1BB ligand (71-254) Fc (kih) fusion (control B)	99	15.4
bivalent DP47 untargeted split trimeric human 4-1BB ligand (71-254) Fc (kih) fusion (control C)	98	12.6
monovalent DP47-untargeted split trimeric human 4-1BB ligand (71-254) Fc (kih) fusion (control D)	99.5	25.9

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monovalent DP47-untargeted split trimeric human 4-1BB ligand (71-254) Fc (kih) fusion (control E)	93.3	4.1
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Table 76 summarizes the yield and final monomer content of anti-CD19 (8B8-018) and germline DP47 human IgG1 PGLALA (control F).

Table 76: Biochemical analysis of control human IgG1 PGLALA

Construct	Monomer [%] (SEC)	Yield [mg/l]
anti-CD19(8B8-018) huIgG1 PGLALA	100	36.6
germline DP47 human IgG1 PGLALA	100	50

5

Example 8

Functional characterization of the CD19 targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules

8.1. Surface plasmon resonance (affinity)

10 Binding of CD19 targeted split trimeric 4-1BB ligand Fc fusion antigen binding molecules (constructs 3.4 and 3.6) to the recombinant 4-1BB Fc(kih) and CD19 was assessed by surface plasmon resonance (SPR). All SPR experiments were performed on a Biacore T200 at 25 °C with HBS-EP as running buffer (0.01 M HEPES pH 7.4, 0.15 M NaCl, 3 mM EDTA, 0.005% Surfactant P20, Biacore, Freiburg/Germany).

15 Interaction with human and cynomolgus 4-1BB

Anti-human Fab antibody (Biacore, Freiburg/Germany) was directly coupled on a CM5 chip at pH 5.0 using the standard amine coupling kit (Biacore, Freiburg/Germany). The immobilization level was approximately 8000 RU. The CD19 targeted split trimeric 4-1BB ligand Fc fusions were captured for 60 seconds at 2 and 5 nM (control D was also injected).
 20 Recombinant human or cynomolgus 4-1BB avi His was passed at a concentration range from 2.7 to 2000 nM (3-fold dilution) with a flow of 30 µL/minutes through the flow cells over 120 seconds. The dissociation was monitored for 180 seconds. Bulk refractive index differences were corrected for by subtracting the response obtained on reference flow cell. Here, the antigens were flown over a surface with immobilized anti-human Fab antibody but on which HBS-EP has been
 25 injected rather than the antibodies.

Interaction with human CD19

Anti-human Fab antibody (Biacore, Freiburg/Germany) was directly coupled on a CM5 chip at pH 5.0 using the standard amine coupling kit (Biacore, Freiburg/Germany). The immobilization level was approximately 8000 RU. The CD19 targeted split trimeric 4-1BB ligand Fc fusions, or the control antibody (anti-CD19(8B8-018) huIgG1 PGLALA) were captured for 60 seconds at 20 nM. Recombinant human CD19-Fc(kih) was passed at a concentration range from 7.8 to 500 nM (2-fold dilution) with a flow of 30 μ L/minutes through the flow cells over 120 seconds. The dissociation was monitored for 120/1800 seconds. Bulk refractive index differences were corrected for by subtracting the response obtained on reference flow cell. Here, the antigens were flown over a surface with immobilized anti-human Fab antibody but on which HBS-EP has been injected rather than the antibodies.

Kinetic constants were derived using the Biacore T200 Evaluation Software (vAA, Biacore AB, Uppsala/Sweden), to fit rate equations for 1:1 Langmuir binding by numerical integration.

The bispecific constructs 3.4, 3.6 and control D bind similarly to 4-1BB. **Table 77** shows the average with standard deviation (in parenthesis) from the two experiments (using the construct capture solution either at 2 nM or 5 nM). The bispecific constructs 3.4 and 3.6 bind human CD19 with a similar affinity as the IgG. Affinity constants for the interaction were determined by fitting to a 1:1 Langmuir binding. For measurements with hu4-1BB and cy4-1BB, average and standard deviation (in parenthesis) are shown (two experiments with 2 or 5 nM capture solution).

Table 77: Binding of CD19 targeted split trimeric 4-1BB ligand Fc fusion to recombinant human (hu) 4-1BB, cynomolgus (cy) 4-1BB and human (hu) CD19.

	Antigen	ka (1/Ms)	kd (1/s)	KD (M)
monovalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion containing CH-CL cross with charged residues (construct 3.4)	hu 4-1BB	7.2E+04 (5.9E+03)	2.5E-02 (1.0E-05)	3.4E-07 (2.8E-08)
	cy 4-1BB	1.2E+05 (8.6E+03)	1.3E-02 (1.8E-04)	1.1E-07 (9.9E-09)
	hu CD19	2.77E+04	2.67E-04	9.64E-09
bivalent CD19(8B8-018) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion (construct 3.6)	hu 4-1BB	6.9E+04 (1.7E+03)	2.4E-02 (1.5E-04)	3.5E-07 (1.1E-08)
	cy 4-1BB	1.1E+05 (7.7E+03)	1.4E-02 (3.1E-04)	1.3E-07 (1.3E-08)
	hu CD19	2.55E+04	2.69E-04	1.06E-08

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monovalent DP47 untargeted split trimeric human 4-1BB ligand (71- 248) Fc (kih) fusion with CH-CL cross and with charged residues (control D)	hu 4- 1BB	7.3E+04 (3.9E+03)	2.6E-02 (6.3E-04)	3.5E-07 (1.0E-08)
	cy 4- 1BB	1.2E+05 (1.9E+03)	1.4E-02 (1.0E-04)	1.2E-07 (2.9E-09)
anti-CD19(8B8-018) huIgG1 PGLALA	hu CD19	2.12E+04	2.61E-04	1.23E-08

8.2. Surface plasmon resonance (simultaneous binding)

The capacity of binding simultaneously human 4-1BB Fc(kih) and human CD19 was assessed by surface plasmon resonance (SPR). All SPR experiments were performed on a

5 Biacore T200 at 25 °C with HBS-EP as running buffer (0.01 M HEPES pH 7.4, 0.15 M NaCl, 3 mM EDTA, 0.005% Surfactant P20, Biacore, Freiburg/Germany). Biotinylated human 4-1BB Fc(kih) was directly coupled to a flow cell of a streptavidin (SA) sensor chip. Immobilization levels up to 250 resonance units (RU) were used.

10 The CD19 targeted trimeric split 4-1BBL constructs (constructs 3.1, 3.3, 3.4, 3.5, 3.6, 4.4) were passed at a concentration range of 200 nM with a flow of 30 µL/minute through the flow cells over 90 seconds and dissociation was set to zero sec. Human CD19 was injected as second analyte with a flow of 30 µL/minute through the flow cells over 90 seconds at a concentration of 500 nM (**Figure 34A**). The dissociation was monitored for 120 sec. Bulk refractive index differences were corrected for by subtracting the response obtained in a reference flow cell,

15 where no protein was immobilized.

As can be seen in the graphs of **Figure 35**, all bispecific constructs could bind simultaneously human 4-1BB and human CD19.

Example 9

Functional characterization of the CD-19 targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules

20

9.1. Binding on activated human PMBCs of the CD19-targeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules

To determine binding of 4-1BBL trimer-containing Fc fusion antigen binding molecules to human PMBCs, different titrated concentrations of the CD19-targeted 4-1BBL trimer-containing

25 Fc fusion antigen binding molecules were used in the assay as described in Example 5.2.

Figures 36A and 36B show the binding of Constructs 3.1, 3.3, 3.4, 3.5 and 3.6 as prepared in Example 7 on activated 4-1BB-expressing CD4⁺ T cells and CD8⁺ T cells, respectively. Gates were set on living CD45⁺ CD3⁺ CD4⁺ or CD45⁺ CD3⁺ CD8⁺ T cells and MFI of PE-conjugated AffiniPure anti-human IgG IgG Fc γ -fragment-specific goat F(ab')₂ fragment were blotted against the titrated concentration of targeted split trimeric 4-1BB ligand Fc fusion variants. **Table 78** shows the EC₅₀ values as measured for Constructs 3.1, 3.3, 3.4, 3.5 and 3.6 and control molecules.

Table 78: Binding on activated 4-1BB-expressing CD4⁺ T cells and CD8⁺ T cells

Construct	EC ₅₀ [nM] 4-1BB ⁺ CD8 ⁺	EC ₅₀ [nM] 4-1BB ⁺ CD4 ⁺
Control B	0.05	0.26
Control C	0.02	0.30
Control D	0.04	0.28
Control E	0.13	1.22
3.1	0.03	0.28
3.3	0.01	0.29
3.4	0.15	2.04
3.5	0.04	1.03
3.6	0.05	0.21

9.2 Binding to CD19-expressing tumor cells

For binding assays on CD19-expressing tumor cells, the following human CD19-expressing lymphoma cell lines were used: diffuse large non-Hodgkin B cell lymphoma (B-NHL) cell line SU-DHL-8 (DSMZ ACC573), acute B cell precursor lymphoid leukemia cell line Nalm6 (DSMZ ACC-128), diffuse large cell lymphoblast lymphoma cell line Toledo (ATCC CRL-2631) and diffuse large B cell lymphoma cell line OCI-Ly18 (DSMZ ACC-699). The assays were performed as described for the FAP-expressing MV-3 and WM-266-4 tumor cell lines in Example 5.3.

Gates were set on living tumor cells and MFI of PE-conjugated AffiniPure anti-human IgG IgG Fc γ -fragment-specific goat F(ab')₂ fragment were blotted against the titrated concentration of targeted split trimeric 4-1BB ligand Fc fusion constructs.

Figure 37A shows the binding of Constructs 3.1, 3.3, 3.4, 3.5 and 3.6 as prepared in Example 7.1 to diffuse large non-Hodgkin B cell lymphoma (B-NHL) cell line SU-DHL-8 and in

Figure 37B the binding of Constructs 3.1, 3.3, 3.4, 3.5 and 3.6 to acute B cell precursor lymphoid leukemia cell line Nalm6 is presented. **Figure 37C** shows the binding of Constructs 3.1, 3.3, 3.4, 3.5 and 3.6 to diffuse large cell lymphoblast lymphoma cell line Toledo and **Figure 37D** shows the binding of Constructs 3.1, 3.3, 3.4, 3.5 and 3.6 to diffuse large B cell lymphoma cell line OCI-Ly18. **Table 79** shows the EC₅₀ values as measured for Constructs 3.1, 3.3, 3.4, 3.5 and 3.6 and control molecules.

Table 79: Binding to CD19-expressing tumor cells

Construct	EC ₅₀ [nM] SU-DHL-8	EC ₅₀ [nM] Nalm6	EC ₅₀ [nM] Toledo	EC ₅₀ [nM] OCI-Ly18
3.1	0.64	0.43	0.29	0.29
3.3	0.15	0.14	0.10	0.09
3.4	0.31	0.39	0.29	0.26
3.5	0.54	0.43	0.27	0.31
3.6	0.14	0.12	0.09	0.10
control G	0.09	0.10	0.06	0.07

Example 10

10 Biological activity of the CD19-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules

10.1. NF- κ B activation in HeLa cells expressing human 4-1BB

HeLa cells expressing human 4-1BB and NF- κ B-luciferase were generated as described in Example 6.1.

15 NF- κ B activation in Hela cells expressing human 4-1BB co-cultured with CD19-expressing tumor cells

NF- κ B-luciferase human-4-1BB HeLa cells were harvested and resuspended in DMEM medium supplied with 10 % (v/v) FBS and 1 % (v/v) GlutaMAX-I to a concentration of 0.2×10^6 cells/ml. 100 μ l (2×10^4 cells) of this cell suspension were transferred to each well of a sterile white 96-well flat bottom tissue culture plate with lid (greiner bio-one, Cat. No. 655083) and the plate were incubated at 37 °C and 5 % CO₂ overnight. The next day 50 μ L of medium containing titrated concentrations of CD19-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (CD19 split 4-1BBL trimer) or DP47-untargeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (DP47 split 4-1BBL trimer) were added. CD19-

expressing B cell lymphoma cell lines (diffuse large non-Hodgkin B cell lymphoma (B-NHL) cell line SU-DHL-8 (DSMZ ACC573) and human non-Hodgkin's B cell lymphoma cell line Pfeiffer (ATCC CRL-2632)) were resuspended in DMEM medium supplied with 10 % (v/v) FBS and 1 % (v/v) GlutaMAX-I to a concentration of 2×10^6 cells/ml.

- 5 Suspension of CD19-expressing B cell lymphoma cell (50 μ l, final ratio 1:5) or only medium were added to each well and plates were incubated for 6 hours at 37 °C and 5 % CO₂. Cells were washed two times with 200 μ L/well DPBS. 40 μ l freshly prepared Reporter Lysis Buffer (Promega, Cat-No: E3971) were added to each well and the plate were stored over night at -20 °C. The next day frozen cell plate and Detection Buffer (Luciferase 1000 Assay System, 10 Promega, Cat. No. E4550) were thawed at room temperature. 100 μ L of detection buffer were added to each well and luciferase activity was measured as fast as possible using a SpectraMax M5/M5e microplate reader and a SoftMax Pro Software (Molecular Devices) counting light emission in URL (units of released light for 0.5s/well) or Victor3 1420 multilabel counter plate reader (Perkin Elmer) and the Perkin Elmer 2030 Manager Software counting light emission as 15 counts per seconds (CPS) and blotted against the concentration of tested constructs.

CD19-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules Constructs 3.1 and 3.3 triggered activation of the NF- κ B signaling pathway in the reporter cell line in the presence of CD19-expressing B cell lymphoma cells. In contrast, the untargeted control molecules failed to trigger such an effect at any of the tested concentrations (Figure 38).

Example 11

11.1 Preparation of CEA (T84.66-LCHA) targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules

11.1.1 Humanization of anti-CEA clone T84.66

5 Novel humanized variants of the murine antibody T84.66 (Wagener et al., J Immunol 130, 2308 (1983), Neumaier et al., J Immunol 135, 3604 (1985)) were developed by grafting of the CDRs onto human germline framework acceptor sequences.

Humanization of an antibody from non-human origin consists essentially of transplanting the CDR residues from the non-human antibody (donor) onto the framework of a human (acceptor) antibody. Normally the acceptor framework is selected by aligning the sequence of the donor to a collection of potential acceptor sequences and choosing one that has either reasonable homology to the donor, or shows similar amino acids at some positions critical for structure and activity. In the present case, the search for the antibody acceptor framework was performed by aligning the mouse T84.66 protein (NCBI Acc No: CAA36980 for the heavy chain (SEQ ID NO:317), and CAA36979 (SEQ ID NO:318) for the light chain) sequence to a collection of human germ-line sequences and picking that human sequence that showed high sequence identity. Here, the sequence IGHV1-69*08 from the IMGT database was chosen as the heavy chain framework acceptor sequence (IMGT Acc No. Z14309, SEQ ID NO:319), and the IGKV3-11*01 sequence (IMGT Acc No. X01668, SEQ ID NO:320) was chosen to be the framework acceptor for the light chain. Onto these two acceptor frameworks, the three complementary determining regions (CDRs) of the mouse heavy and light variable domains were grafted. Since the framework 4 (FR4) region is not part of the variable region of the germ line V gene, the alignment for that position was done individually. The JH4 sequence was chosen for the heavy chain, and the JK2 sequence was chosen for the light chain.

25 11.1.2 Binding of different humanized variants of T84.66 IgG to cells

The binding of different humanized variants of T84.66 IgG was tested on CEA-expressing human gastric adenocarcinoma cells (MKN45, DSMZ ACC 409).

30 Cells were harvested, counted, checked for viability and re-suspended at 2×10^6 cells/ml in FACS buffer (100 μ l PBS 0.1% BSA). 100 μ l of cell suspension (containing 0.2×10^6 cells) were incubated in round-bottom 96-well plate for 30 min at 4°C with increasing concentrations of the CEA IgG (4 ng/ml– 60 μ g/ml), washed twice with cold PBS 0.1% BSA, re-incubated for further 30 min at 4°C with the PE-conjugated AffiniPure F(ab')₂ Fragment goat anti-human IgG Fc γ Fragment Specific secondary antibody (Jackson Immuno Research Lab PE #109-116-170), washed twice with cold PBS 0.1% BSA and immediately analyzed by FACS using a FACS

CantoII (Software FACS Diva). Binding curves and EC50 values were obtained and calculated using GraphPadPrism5.

Figure 39 shows the different binding pattern of selected humanized variants of the T84.66 IgG to human CEA, expressed on MKN45 cells. Based on the calculated EC50 binding values (Table 80), the humanized variant 1 was selected for further evaluation.

Table 80: Binding of different humanized variants of T84.66 IgGs to cells (EC50 values, based on binding curves shown in Figure 39, calculated by Graph Pad Prism).

	EC50 (µg/ml)
Parental chimeric T84.66	0.99
Humanized variant 1	1.5
Humanized variant 2	8.6
Humanized variant 3	1.4
Humanized variant 4	3.1
Humanized variant 5	-
Humanized variant 6	-

Humanized variant 1 is termed in the following T84.66-LCHA. The amino acid sequences of its CDRs and of the VH and VL as well as the amino acid sequences of the VH and VL domain of the parental chimeric T84.66 clone are shown in Table 81.

Table 81: Amino acid sequences of the variable domains of CEA clone T84.66-LCHA and its parental antibody T84.66

Description	Sequence	SEQ ID NO:
CEA CDR-H1	DTYMH	321
CEA CDR-H2	RIDPANGNSKYVPKFQG	322
CEA CDR-H3	FGYYVSDYAMAY	323
CEA CDR-L1	RAGESVDIFGVGFLH	324
CEA CDR-L2	RASN RAT	325
CEA CDR-L3	QQTNEDPYT	326
Parental CEA binder VH	EVQLQQSGAELVEPGASVKLSCTASGFNIKDTYMHVWKQRPEQ GLEWIGRIDPANGNSKYVPKFQKGKATITADTSSNTAYLQLTSLTS EDTAVYYCAPFGYYVSDYAMAYWGQGTSVTVSS	327
Parental CEA binder VL	DIVLTQSPASLAVSLGQRATMSCRAGESVDIFGVGFLHWYQQKP GQPPKLLIYRASNLSESGIPVRFSGTGSRTDFTLIIDPVEADDVATY YCQQTNEDPYTFGGGTKLEIK	328
Humanized	QVQLVQSGAEVKKPGSSVKVSCASGFNIKDTYMHVWRQAPGQ GLEWMGRIDPANGNSKYVPKFQGRVTITADTSTSTAYMELSSLR	329

CEA binder CEA (T84.66- LCHA) VH	SEDTAVYYCAPFGYYVSDYAMAYWGQGLTVSS	
Humanized CEA binder CEA (T84.66- LCHA) VL	EIVLTQSPATLSLSPGERATLSCRAGESVDIFGVGFLHWYQQKPG QAPRLLIYRASNRATGIPARFSGSGSGTDFTLTSSLEPEDFAVYY CQQTNEDPYTFGQGTKLEIK	330

11.2 Preparation of CEA (T84.66-LCHA) targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules

Different fragments of the DNA sequence encoding part of the ectodomain (amino acid 71-254 and 71-248) of human 4-1BB ligand were synthesized according to the P41273 sequence of Uniprot database (SEQ ID NO:42).

11.2.1 Preparation of monovalent CEA (T84.66-LCHA) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains with charged residues (Construct 5.1)

A polypeptide containing two ectodomains of 4-1BB ligand (71-254), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned as depicted in **Figure 29A**: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL. A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the human IgG1-CH domain, was cloned as described in Figure 29B: human 4-1BB ligand, (G4S)₂ connector, human CH.

To improve correct pairing the following mutations have been introduced in the crossed CH-CL. In the dimeric 4-1BB ligand fused to human CL, E123R and Q124K. In the monomeric 4-1BB ligand fused to human CH1, K147E and K213E.

The variable region of heavy and light chain DNA sequences encoding a binder specific for CEA, clone T84.66-LCHA, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831.

Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-CEA-Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations and the anti-CEA light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a CEA binding Fab (**Figure 40**, Construct 5.1).

Table 82 shows the cDNA and amino acid sequences of the monovalent CEA (T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule with crossed CH-CL and charged residues (construct 5.1).

Table 82: cDNA and amino acid sequences of monovalent CEA(T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion containing CH-CL cross with charged residues (construct 5.1). * for charged residues

SEQ ID NO:	Description	Sequence
129	Nucleotide sequence Dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 3
130	Nucleotide sequence Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
331	Nucleotide sequence anti-CEA(T84.66-LCHA) Fc hole chain	CAGGTGCAGCTGGTGCAGTCTGGCGCCGAAGTGAAGAA ACCCGGCAGCAGCGTGAAGGTGTCCTGCAAGGCCAGCG GCTTCAACATCAAGGACACCTACATGCACTGGGTGCGCC AGGCCCTGGACAGGGACTGGAATGGATGGGCAGAATC GACCCCGCCAACGGCAACAGCAAATACGTGCCCAAGTT CCAGGGCAGAGTGACCATCACCGCCGACACCAGCACCT CCACCGCCTACATGGAAGTGAAGCAGCCTGCGGAGCGAG GACACCGCCGTGTACTACTGTGCCCCCTTCGGCTACTAC GTGTCCGACTACGCCATGGCCTATTGGGGCCAGGGCAC ACTCGTGACCGTGTCTCTGCTAGCACCAAGGGCCCCCTC CGTGTTCCTCCCTGGCCCCCAGCAGCAAGAGCACCAGCG GCGGCACAGCCGCTCTGGGCTGCCTGGTCAAGGACTACT TCCCCGAGCCCGTGACCGTGTCTTGAACAGCGGAGCC CTGACCTCCGGCGTGCACACCTTCCCCGCCGTGCTGCAG AGTTCTGGCCTGTATAGCCTGAGCAGCGTGGTCACCGTG CCTTCTAGCAGCCTGGGCACCCAGACCTACATCTGCAAC GTGAACCACAAGCCCAGCAACACCAAGGTGGACAAGAA GGTGGAGCCCAAGAGCTGCGACAAAACCTCACACATGCC CACCGTGCCAGCACCTGAAGCTGCAGGGGGACCGTCA GTCTTCTCTTCCCCCCTTCCCAAGGACACCCCTCATG ATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGAC GTGAGCCACGAAGACCCCTGAGGTCAAGTTCAACTGGTA CGTGGACGGCGTGGAGGTGCATAATGCCAAGACAAAGC CGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTC AGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGC AAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGG CGCCCCATCGAGAAAACCATCTCCAAGGCCAAAGGGC AGCCCCGAGAACCACAGGTGTGCACCTGCCCCCATCCC GGGATGAGCTGACCAAGAACCAGGTGAGCCTCTCGTGC GCAGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGA

		GTGGGAGAGCAATGGGCAGCCGGAGAACAACACTACAAGACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCCTCGTGAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAA
332	Nucleotide sequence anti-CEA(T84.66-LCHA) light chain	GAGATCGTGCTGACCCAGAGCCCTGCCACCCTGTCACTGTCTCCAGGCGAGAGAGCCACCCTGAGCTGTAGAGCCGGCGAGAGCGTGGACATCTTCGGCGTGGGATTCTGCACTGTATCAGCAGAAGCCCCGGCCAGGCCCCCAGACTGCTGATCTACAGAGCCAGCAACCGGGCCACAGGCATCCCCGCCAGATTTTCTGGCTCTGGCAGCGGCACCGACTTCACCCTGACAATCAGCAGCCTGGAACCCGAGGACTTCGCCGTGTACTACTGCCAGCAGACCAACGAGGACCCCTACACCTTTGGCCAGGGCACCAAGCTGGAAATCAAGCGTACGGTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGATGAGCAGTTGAAATCTGGAAGTGCCTCTGTTGTGTGCCTGCTGAATAACTTCTATCCCAGAGAGGCCAAAGTACAGTGGAAGGTGGATAACGCCCTCCAATCGGGTAACTCCCAGGAGAGTGTACAGAGCAGGACAGCAAGGACAGCACCTACAGCCTCAGCAGCACCTGACGCTGAGCAAAGCAGACTACGAGAAACACAAAGTCTACGCCTGCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGAGAGTG T
115	Dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 3
116	Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
333	anti- CEA (T84.66-LCHA) Fc hole chain	QVQLVQSGAEVKKPGSSVKVSKASGFNIKDTYMHWRQAPGQGLEWMGRIDPANGNSKYVPKFQGRVTITADTSTSTAYMELSSLRSEDTAVYYCAPFGYYVSDYAMAYWGQGLTVTVSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALGAPIEKTISKAKGQPREPQVCTLPSPSRDELTKNQVSLSCAVKGFYPSDIAVEWESNGQPENNYKTPPVLDSDGSFFLVSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK
334	anti- CEA (T84.66-LCHA) light chain	EIVLTQSPATLSLSPGERATLSCRAGESVDIFGVGFLHWYQKPGQAPRLLIYRASNRATGIPARFSGSGGTDFLTISLLEPEDFAVYYCQQTNEDPYTFGQGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSYSTLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC

11.2.2 Preparation of monovalent CEA (T84.66-LCHA) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains without charged residues (Construct 5.2)

5 A polypeptide containing two ectodomains of 4-1BB ligand (71-254), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned in analogy as depicted in Figure (29A), but without amino acid mutations in the CL domain: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL. A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the human IgG1-CH1 domain, was cloned in
10 analogy as depicted in Figure (29B), but without amino acid mutations in the CH1 domain: human 4-1BB ligand, (G4S)₂ connector, human CH1.

The variable region of heavy and light chain DNA sequences encoding a binder specific for CEA, clone T84.66-LCHA, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1.

15 The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831. Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-CEA-Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations and the anti-
20 CEA light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a CEA-binding Fab (**Figure 40**, Construct 5.2).

Table 83 shows the cDNA and amino acid sequences of the monovalent CEA (T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule containing crossed CH-CL cross without charged residues (construct 5.2).

25 **Table 83: cDNA and amino acid sequences of monovalent CEA (T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion containing CH-CL cross without charged residues (construct 5.2)**

SEQ ID NO:	Description	Sequence
165	Nucleotide sequence dimeric ligand (71-254)– CL Fc knob chain	see Table 22
166	Nucleotide sequence monomeric hu 4-1BBL (71-254) – CH1	see Table 22
331	Nucleotide sequence anti- CEA	see Table 82

	(T84.66-LCHA) Fc hole chain	
332	Nucleotide sequence anti- CEA (T84.66-LCHA) light chain	see Table 82
117	Dimeric ligand (71-254) - CL Fc knob chain	see Table 22
118	Monomeric ligand (71-254) -CH1	see Table 22
333	anti- CEA (T84.66-LCHA) Fc hole chain	see Table 82
334	anti- CEA (T84.66-LCHA) light chain	see Table 82

11.2.3 Preparation of bivalent CEA(T84.66-LCHA) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding (Construct 5.3)

A polypeptide containing two ectodomains of 4-1BB ligand (71-254), separated by (G4S)₂ linkers was fused to the C-terminus of human IgG1 Fc hole chain, as depicted in Figure 29C: human IgG1 Fc hole, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand. A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the C-terminus of human IgG1 Fc knob chain as described in Figure 29D: human IgG1 Fc knob, (G4S)₂ connector, human 4-1BB ligand.

The variable region of heavy and light chain DNA sequences encoding a binder specific for CEA, clone T84.66-LCHA, were subcloned in frame with either the constant heavy chain of the hole, the knob or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831. Combination of the anti-CEA huIgG1 hole dimeric ligand chain containing the Y349C/T366S/L368A/Y407V mutations, the anti-CEA huIgG1 knob monomeric ligand chain containing the S354C/T366W mutations and the anti-CEA light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and two CEA binding Fabs (Figure 40, construct 5.3).

Table 84 shows the cDNA and amino acid sequences of the bivalent CEA(T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule (construct 5.3).

Table 84: cDNA and amino acid sequences of bivalent CEA(T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) PGLALA fusion (construct 5.3)

SEQ ID NO:	Description	Sequence
335	Nucleotide sequence anti-CEA(T84.66-LCHA) Fc hole dimeric 4-1BBL (71-254) chain	CAGGTGCAGCTGGTGCAGTCTGGCGCCGAAGTGAAGAA ACCCGGCAGCAGCGTGAAGGTGTCCTGCAAGGCCAGCG GCTTCAACATCAAGGACACCTACATGCACTGGGTGCGCC AGGCCCCTGGACAGGGACTGGAATGGATGGGCAGAATC GACCCCGCCAACGGCAACAGCAAATACGTGCCCAAGTT CCAGGGCAGAGTGACCATCACCGCCGACACCAGCACCT CCACCGCCTACATGGAAGTGAAGCAGCTGCGGAGCGAG GACACCGCCGTGTACTACTGTGCCCCCTTCGGCTACTAC GTGTCCGACTACGCCATGGCCTATTGGGGCCAGGGCAC ACTCGTGACCGTGTCTCTGCTAGCACCAAGGGCCCCCTCC GTGTTCCCCCTGGCCCCCAGCAGCAAGAGCACCAAGCGGCG GCACAGCCGCTCTGGGCTGCCTGGTCAAGGACTACTTCCC CGAGCCCGTGACCGTGTCTCTGGAACAGCGGAGCCCTGACC TCCGGCGTGCACACCTTCCCCGCCGTGCTGCAGAGTTCTG GCCTGTATAGCTGAGCAGCGTGGTCACCGTGCCCTTCTAGC AGCCTGGGCACCCAGACCTACATCTGCAACGTGAACCACAA GCCCAGCAACACCAAGGTGGACAAGAAGGTGGAGCCCAAG AGCTGCGACAAAACCTCACACATGCCACCGTGCCACGACCC TGAAGCTGCAGGGGGACCGTCAGTCTTCTCTTCCCCCAA AACCAAGGACACCCCTCATGATCTCCCGGACCCCTGAGGTC ACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGG TCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAAT GCCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGT ACCGTGTGGTCAGCGTCCTCACCGTCTGCAACAGGACTG GCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAG CCCTCGGCGCCCCCATCGAGAAAACCATCTCCAAGGCCAAA GGGCAGCCCCGAGAACCACAGGTGTGCACCCTGCCCCCAT CCCGGGATGAGCTGACCAAGAACCAGGTGAGCCTCTCGTG CGCAGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGT GGGAGAGCAATGGGCAGCCGGAGAACAACTACAAGACCAC GCCTCCCGTGCTGGACTCCGACGGCTCCTTCTCTCTCGTGA GCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAA CGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACC ACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTGGAGG CGGCGGAAGCGGAGGAGGAGGATCCAGAGAGGGCCCTGA GCTGAGCCCCGATGATCCTGCTGGACTGCTGGACCTGCGG CAGGGCATGTTTGCTCAGCTGGTGGCCAGAACGTGCTGCT GATCGATGGCCCCCTGTCTGCTGACAGCGATCCTGGACTG GCTGGCGTGTCACTGACAGGCGGCCTGAGCTACAAAGAGG ACACCAAAGAAGTGGTGGTGGCCAAGGCCGGCGTGTACTA CGTGTCTTTTCACTGGAAGTGCAGGAGAGTGGTGGCCGGC GAAGGATCTGGCTCTGTGTCTCTGGCCCTGCATCTGCAGCC TCTGAGAAGCGCTGCTGGCGCTGCAGCTCTGGCACTGACA GTGGATCTGCCTCCTGCCAGCTCCGAGGCCCGGAATAGCG CATTTGGGTTTCAAGGCAGGCTGCTGCACCTGTCTGCCGGC CAGAGGCTGGGAGTGCATCTGCACACAGAGGCCAGGGCTA GACACGCCTGGCAGCTGACACAGGGCGCTACAGTGCTGGG CCTGTTTCAAGTGAACCCCGAGATTCCAGCCGGCCTGCCTT

		<p>CTCCAAGAAGCGAAGGCGGAGGCGGATCTGGCGGCGGAG GATCTAGAGAGGGACCCGAACGTGCCCTGACGATCCAGC CGGGCTGCTGGATCTGAGACAGGGAATGTTCCGCCAGCTG GTGGCTCAGAATGTGCTGCTGATTGACGGACCTCTGAGCTG GTACTCCGACCCAGGGCTGGCAGGGGTGTCCCTGACTGGG GGACTGTCCTACAAAGAAGATACAAAAGAACTGGTGGTGGC TAAAGCTGGGGTGTACTATGTGTTTTTCAGCTGGAACCTGAG GCGGGTGGTGGCTGGGGAGGGCTCAGGATCTGTGTCCCTG GCTCTGCATCTGCAGCCACTGCGCTCTGCTGCTGGCGCAG CTGCACTGGCTCTGACTGTGGACCTGCCACCAGCCTCTAGC GAGGCCAGAAACAGCGCCTTCGGGTTCAGGACGCCTGC TGCATCTGAGCGCCGGACAGCGCCTGGGAGTGCATCTGCA TACTGAAGCCAGAGCCCGGCATGCTTGGCAGCTGACTCAG GGGGCAACTGTGCTGGGACTGTTTCGCGTGACACCTGAGAT CCCTGCCGGACTGCCAAGCCCTAGATCAGAA</p>
336	Nucleotide sequence anti- CEA(T84.66- LCHA) Fc knob monomeric 41- BBL (71-254)	<p>CAGGTGCAGCTGGTGCAGTCTGGCGCCGAAGTGAAGAA ACCCGGCAGCAGCGTGAAAGGTGTCCTGCAAGGCCAGCG GCTTCAACATCAAGGACACCTACATGCACTGGGTGCGCC AGGCCCTGGACAGGGACTGGAATGGATGGGCAGAATC GACCCCGCCAACGGCAACAGCAAATACGTGCCCAAGTT CCAGGGCAGAGTGACCATCACCGCCGACACCAGCACCT CCACCGCCTACATGGAAGTGAAGCAGCTGCGGAGCGAG GACACCGCCGTGTACTACTGTGCCCCCTTCGGCTACTAC GTGTCCGACTACGCCATGGCCTATTGGGGCCAGGGCAC ACTCGTGACCGTGTCTCTGCTAGCACCAAGGGCCCATCG GTCTTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGGGG GCACAGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCC CGAACCGGTGACGGTGTGCTGGAAGTCAAGGCGCCCTGACC AGCGGCGTGACACCTTCCCAGGTGTCTACAGTCTCTCAG GACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGC AGCTTGGGCACCCAGACCTACATCTGCAACGTGAATCACAA GCCCAGCAACACCAAGGTGGACAAGAAAGTTGAGCCCAAA CTTGTGACAAAACCTCACACATGCCACCGTGCCAGCACCT GAAGCTGCAGGGGGACCGTCAGTCTTCTCTTCCCCCAAA ACCCAAGGACACCCTCATGATCTCCCGGACCCCTGAGGTCA CATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGGT CAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATG CCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTA CCGTGTGGTCAGCGTCTCACCGTCTGCAACCAGGACTGG CTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGC CCTCGGCGCCCCCATCGAGAAAACCATCTCCAAGCCAAAG GGCAGCCCCGAGAACCACAGGTGTACACCCTGCCCCCTG CAGAGATGAGCTGACCAAGAACCAGGTGTCCCTGTGGTGTG TGGTCAAGGGCTTCTACCCAGCGATATCGCCGTGGAGTG GGAGAGCAACGGCCAGCCTGAGAACAATAAGACCACC CCCCCTGTGCTGGACAGCGACGGCAGCTTCTTCTGTACTC CAAAGTACCGTGGACAAGAGCCGGTGGCAGCAGGGCAAC GTGTTCACTGCAGCGTGATGCACGAGGCCCTGCACAACC ACTACACCAGAAAGTCCCTGAGCCTGAGCCCCGGCGGAGG CGGCGGAAGCGGAGGAGGAGGATCCAGAGAGGGCCCTGA GCTGAGCCCCGATGATCCTGCTGGACTGCTGGACCTGCGG CAGGGCATGTTTGCTCAGCTGGTGGCCAGAACGTGCTGCT GATCGATGGCCCCCTGTCTTGGTACAGCGATCCTGGACTG GCTGGCGTGTCACTGACAGGCGGCCTGAGCTACAAAGAGG</p>

		ACACCAAAGAACTGGTGGTGGCCAAGGCCGGCGTGTACTA CGTGTTCTTTCAGCTGGAAGCTGCGGAGAGTGGTGGCCGGC GAAGGATCTGGCTCTGTGTCTCTGGCCCTGCATCTGCAGCC TCTGAGAAGCGCTGCTGGCGCTGCAGCTCTGGCACTGACA GTGGATCTGCCTCCTGCCAGCTCCGAGGCCCGGAATAGCG CATTTGGGTTTCAAGGCAGGCTGCTGCACCTGTCTGCCGGC CAGAGGCTGGGAGTGCATCTGCACACAGAGGCCAGGGCTA GACACGCCTGGCAGCTGACACAGGGCGCTACAGTGCTGGG CCTGTTTCAGAGTGACCCCGAGATTCCAGCCGGCCTGCCTT CTCCAAGAAGCGAA
332	Nucleotide sequence anti- CEA(T84.66- LCHA) light chain	see Table 82
337	anti- CEA(T84.66- LCHA) Fc hole dimeric 41-BBL (71-254) chain	QVQLVQSGAEVKKPGSSVKVSCASGFNIKDTYMHWVRQ APGQGLEWMGRIDPANGNSKYVPKFQGRVTITADTSTSTA YMESSLRSEDYAVYYCAPFGYYVSDYAMAYWGQGLV TVSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVS WNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNV NHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFP PKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHN AKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKAL GAPIEKTISKAKGQPREPQVCTLPISRDELTKNQVSLSCAVKG FYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLVSKLTVDK SRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGGGGGSGGGG SREGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYS DPGLAGVSLTGGLSYKEDTKELVVAAGVYVFFQLELRRVVA GEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFG FQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRV TPEIPAGLPSRSEGGGGSGGGGSREGPELSPDDPAGLLDLR QGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDT KELVVAAGVYVFFQLELRRVVAEGEGSGSVSLALHLQPLRSA AGAAALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHL HTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSE
338	anti- CEA(T84.66- LCHA) Fc knob monomeric 4- 1BBL (71-254) chain	QVQLVQSGAEVKKPGSSVKVSCASGFNIKDTYMHWVRQ APGQGLEWMGRIDPANGNSKYVPKFQGRVTITADTSTSTA YMESSLRSEDYAVYYCAPFGYYVSDYAMAYWGQGLV TVSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVS WNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNV NHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGGPSVFLFP PKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHN AKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKAL GAPIEKTISKAKGQPREPQVYTLPCRDELTKNQVSLWCLVKG FYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKLTVDK SRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGGGGGSGGGG SREGPELSPDDPAGLLDLRQGMFAQLVAQNVLLIDGPLSWYS DPGLAGVSLTGGLSYKEDTKELVVAAGVYVFFQLELRRVVA GEGSGSVSLALHLQPLRSAAGAAALALTVDLPPASSEARNSAFG FQGRLLHLSAGQRLGVHLHTEARARHAWQLTQGATVLGLFRV TPEIPAGLPSRSE

334	anti-CEA(T84.66-LCHA) light chain	see Table 82
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11.2.4 Preparation of monovalent CEA(T84.66-LCHA) targeted 4-1BB ligand (71-248) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains with charged residues (Construct 5.4)

5 A polypeptide containing two ectodomains of 4-1BB ligand (71-248), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned in analogy to the one depicted in Figure 29A: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL. A polypeptide containing one ectodomain of 4-1BB ligand (71-248) and fused to the human IgG1-CH domain, was cloned in analogy to the one described in Figure 29B: human 4-
10 1BB ligand, (G4S)₂ connector, human CH.

The polypeptide encoding the dimeric 4-1BB ligand fused to human CL domain was subcloned in frame with the human IgG1 heavy chain CH₂ and CH₃ domains on the knob (Merchant, Zhu et al. 1998). To improve correct pairing the following mutations have been introduced in the crossed CH-CL. In the dimeric 4-1BB ligand fused to human CL, E123R and
15 Q124K. In the monomeric 4-1BB ligand fused to human CH1, K147E and K213E.

The variable region of heavy and light chain DNA sequences encoding a binder specific for CEA, clone T84.66-LCHA, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to
20 abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831. Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-CD19-Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations and the anti-CD19 light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a CEA binding Fab
25 (**Figure 40**, construct 5.4).

Table 85 shows the cDNA and amino acid sequences of the monovalent CEA(T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion antigen binding molecule with crossed CH-CL and charged residues (construct 5.4).

Table 85: cDNA and amino acid sequences of monovalent CEA(T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion containing CH-CL cross with charged residues (construct 5.4). * charged residues

SEQ ID NO:	Description	Sequence
169	Nucleotide sequence dimeric ligand (71-248) – CL* Fc knob chain	see Table 24
170	Nucleotide sequence monomeric hu 4-1BBL (71-248) – CH1*	see Table 24
331	Nucleotide sequence anti-CEA(T84.66-LCHA) Fc hole chain	see Table 82
332	Nucleotide sequence anti-CEA(T84.66-LCHA) light chain	see Table 82
119	Dimeric ligand (71-248) – CL* Fc knob chain	see Table 24
120	Monomeric ligand (71-248)-CH1*	see Table 24
333	anti- CEA(T84.66-LCHA) Fc hole chain	see Table 62
334	anti- CEA(T84.66-LCHA) light chain	see Table 59

5 11.2.5 Preparation of monovalent CEA(T84.66-LCHA) targeted 4-1BB ligand (71-248) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains without charged residues (Construct 5.5)

A polypeptide containing two ectodomains of 4-1BB ligand (71-248), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned in analogy as depicted in Figure (29A), but without amino acid mutations in the CL domain: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL. A polypeptide containing one ectodomain of 4-1BB ligand (71-248) and fused to the human IgG1-CH1 domain, was cloned in analogy as depicted in Figure (29B), but without amino acid mutations in the CH1 domain: human 4-1BB ligand, (G4S)₂ connector, human CH1.

15 The variable region of heavy and light chain DNA sequences encoding a binder specific for CEA, clone T84.66-LCHA, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831.

20 Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-CEA-Fc hole chain containing the

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Y349C/T366S/L368A/Y407V mutations and the anti-CEA light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a CD19-binding Fab (Figure 40, Construct 5.5).

Table 86 shows the cDNA and amino acid sequences of the monovalent CEA(T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion antigen binding molecule containing crossed CH-CL cross without charged residues (construct 5.5).

Table 86: cDNA and amino acid sequences of monovalent CEA(T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion containing CH-CL cross without charged residues (construct 5.5).

SEQ ID NO:	Description	Sequence
171	Nucleotide sequence dimeric ligand (71-248) – CL Fc knob chain	see Table 25
172	Nucleotide sequence monomeric ligand (71-248)-CH1	see Table 25
331	Nucleotide sequence anti-CEA(T84.66-LCHA) Fc hole chain	see Table 82
332	Nucleotide sequence anti-CEA(T84.66-LCHA) light chain	see Table 82
173	Dimeric ligand (71-248) – CL Fc knob chain	see Table 25
174	Monomeric ligand (71-248)-CH1	see Table 25
333	anti-CEA(T84.66-LCHA) Fc hole chain	see Table 82
334	anti-CEA(T84.66-LCHA) light chain	see Table 82

10

11.2.6 Preparation of bivalent CEA(T84.66-LCHA) targeted 4-1BB ligand (71-248) trimer-containing Fc (kih) fusion antigen binding (Construct 5.6)

A polypeptide containing two ectodomains of 4-1BB ligand (71-248), separated by (G4S)₂ linkers was fused to the C-terminus of human IgG1 Fc hole chain, as depicted in **Figure 29C**: human IgG1 Fc hole, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand. A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the C-terminus of human IgG1 Fc knob chain as described in **Figure 29D**: human IgG1 Fc knob,

(G4S)2 connector, human 4-1BB ligand.

The variable region of heavy and light chain DNA sequences encoding a binder specific for CEA, clone T84.66-LCHA, were subcloned in frame with either the constant heavy chain of the hole, the knob or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831. Combination of the anti-CEA huIgG1 hole dimeric ligand chain containing the Y349C/T366S/L368A/Y407V mutations, the anti-CEA huIgG1 knob monomeric ligand chain containing the S354C/T366W mutations and the anti-CEA light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and two CEA binding Fabs (Figure 40, construct 5.6).

Table 87 shows the cDNA and amino acid sequences of the bivalent CEA(T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion antigen binding molecule (construct 5.6).

Table 87: cDNA and amino acid sequences of bivalent CEA (T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion (construct 5.6)

SEQ ID NO:	Description	Sequence
339	Nucleotide sequence anti-CEA (T84.66-LCHA) Fc hole dimeric 4-1BBL (71-248) chain	CAGGTGCAGCTGGTGCAGTCTGGCGCCGAAGTGAAGAA ACCCGGCAGCAGCGTGAAGGTGTCCTGCAAGGCCAGCG GCTTCAACATCAAGGACACCTACATGCACTGGGTGCGCC AGGCCCTGGACAGGGACTGGAATGGATGGGCAGAATC GACCCCGCCAACGGCAACAGCAAATACGTGCCCAAGTT CCAGGGCAGAGTGACCATCACCGCCGACACCAGCACCT CCACCGCCTACATGGAAGTGAAGCAGCCTGCGGAGCGAG GACACCGCCGTGTACTACTGTGCCCCCTTCGGCTACTAC GTGTCCGACTACGCCATGGCCTATTGGGGCCAGGGCAC ACTCGTGACCGTGTCTCTGCTAGCACCAAGGGCCCCCTC CGTGTTCCCCCTGGCCCCCAGCAGCAAGAGCACCAGCG GCGGCACAGCCGCTCTGGGCTGCCTGGTCAAGGACTACT TCCCCGAGCCCGTGACCGTGTCTTGGAACAGCGGAGCC CTGACCTCCGGCGTGCACACCTTCCCCGCCGTGCTGCAG AGTTCTGGCCTGTATAGCCTGAGCAGCGTGGTCACCGTG CCTTCTAGCAGCCTGGGCACCCAGACCTACATCTGCAAC GTGAACCACAAGCCCAGCAACACCAAGGTGGACAAGAA GGTGGAGCCCAAGAGCTGCGACAAAACCTCACACATGCC CACCGTGCCAGCACCTGAAGCTGCAGGGGGACCGTCA GTCTTCTCTTCCCCCAAAACCAAGGACACCTCATG ATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGAC GTGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTA CGTGGACGGCGTGGAGGTGCATAATGCCAAGACAAAGC CGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTC AGCGTCCTCACCGTCTGCACCAGGACTGGCTGAATGGC AAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGG

		CGCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGC AGCCCCGAGAACCACAGGTGTGCACCCCTGCCCCATCCC GGGATGAGCTGACCAAGAACCAGGTCAGCCTCTCGTGC GCAGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGA GTGGGAGAGCAATGGGCAGCCGGAGAACAACACTACAAGA CCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCC TCGTGAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAG CAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCT CTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCT CCGGGTGGAGGCGGCGGAAGCGGAGGAGGAGGATCCA GAGAGGGCCCTGAGCTGAGCCCTGATGATCCTGCCGGA CTGCTGGACCTGCGGCAGGGAATGTTTGCCAGCTGGTG GCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCTGG TACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGGC GGCCTGAGCTACAAAGAGGACACCAAAGAACTGGTGGT GGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGGA ACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCTG TGTCTCTGGCCCTGCATCTGCAGCCTCTGAGATCTGCTG CTGGCGCCGCTGCTCTGGCACTGACAGTGGATCTGCCTC CTGCCAGCAGCGAGGCCCGGAATAGCGCATTTGGGTTTC AAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCTG GGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACGC CTGGCAGCTGACACAGGGCGCTACAGTGTGGGCCTGTT CAGAGTGACCCCCGAGATTCCAGCAGGCCTGGGAGGCG GCGGATCTGGCGGGCGGAGGATCTAGAGAAGGACCCGAG CTGTCCCCCGACGATCCCGCTGGGCTGCTGGATCTGAGA CAGGGCATGTTTCGCTCAGCTGGTGGCTCAGAAATGTGCTG CTGATTGACGGACCTCTGAGCTGGTACTCCGACCCAGGG CTGGCAGGGGTGTCCCTGACTGGGGGACTGTCCTACAAA GAAGATACAAAAGAAGTGGTGGTGGCTAAAGCTGGGGT GTACTATGTGTTTTTTCAGCTGGAAGTGAAGCGGGTGGT GGCTGGGGAGGGCTCAGGATCTGTGTCCCTGGCTCTGCA TCTGCAGCCACTGCGCTCTGCAGCAGGGGCTGCAGCACT GGCCCTGACTGTGGACCTGCCCCCAGCTTCTTCCGAGGC CAGAAACAGCGCCTTCGGGTTCGAAGGACGCCTGCTGC ATCTGAGCGCCGGACAGCGCCTGGGAGTGCATCTGCAT ACTGAAGCCAGAGCCCAGCATGCTTGGCAGCTGACTCA GGGGGCAACTGTGCTGGGACTGTTTCGCGTGACACCTGA GATCCCAGCCGGGCTC
340	Nucleotide sequence anti- CEA (T84.66- LCHA) Fc knob monomeric (71- 248) 4-1BBL chain	CAGGTGCAGCTGGTGCAGTCTGGCGCCGAAGTGAAGAA ACCCGGCAGCAGCGTGAAGGTGTCCTGCAAGGCCAGCG GCTTCAACATCAAGGACACCTACATGCACTGGGTGCGCC AGGCCCTGGACAGGGACTGGAATGGATGGGCAGAATC GACCCCGCCAACGGCAACAGCAAATACGTGCCCAAGTT CCAGGGCAGAGTGACCATCACCGCCGACACCAGCACCT CCACCGCCTACATGGAAGTGAAGCAGCCTGCGGAGCGAG GACACCGCCGTGTACTACTGTGCCCCCTTCGGCTACTAC GTGTCCGACTACGCCATGGCCTATTGGGGCCAGGGCAC ACTCGTGACCGTGTCTCTGCTAGCACCAAGGGCCCATC GGTCTTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGG GGGCACAGCGGCCCTGGGCTGCCTGGTCAAGGACTACT TCCCCGAACCGGTGACGGTGTCTGTGGAAGTCAAGGCGCC CTGACCAGCGGCGTGCACACCTTCCCGGCTGTCTTACAG TCCTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTG

		CCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAAC GTGAATCACAAGCCCAGCAACACCAAGGTGGACAAGAA AGTTGAGCCCAAATCTTGTGACAAAACCTCACACATGCCC ACCGTGCCCAGCACCTGAAGCTGCAGGGGGACCGTCAG TCTTCCTCTTCCCCCAAAACCCAAGGACACCCTCATGA TCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACG TGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTAC GTGGACGGCGTGGAGGTGCATAATGCCAAGACAAAGCC GCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCA GCGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGCA AGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGGC GCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCA GCCCCGAGAACCACAGGTGTACACCCTGCCCCCTGCA GAGATGAGCTGACCAAGAACCAGGTGTCCCTGTGGTGT CTGGTCAAGGGCTTCTACCCCAGCGATATCGCCGTGGAG TGGGAGAGCAACGGCCAGCCTGAGAACAACCTACAAGAC CACCCCCCTGTGCTGGACAGCGACGGCAGCTTCTTCCT GTACTCCAACTGACCGTGGACAAGAGCCGGTGGCAGC AGGGCAACGTGTTTCACTGCAGCGTGATGCACGAGGCC CTGCACAACCACTACACCCAGAAGTCCCTGAGCCTGAG CCCCGGCGGAGGCGGCGGAAGCGGAGGAGGAGGATCC AGAGAGGGCCCTGAGCTGAGCCCTGATGATCCTGCCGG ACTGCTGGACCTGCGGCAGGGAATGTTTGCCCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAAGTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGATCTGCT GCTGGCGCCGCTGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCAGCGAGGCCCCGAATAGCGCATTTGGGTTT CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGCTGGGCCTG TTCAGAGTGACCCCCGAGATTCCTGCCGGGCTC
332	Nucleotide sequence anti- CEA (T84.66- LCHA) light chain	see Table 82
341	anti- CEA (T84.66-LCHA) Fc hole dimeric 4- 1BBL (71-248) chain	QVQLVQSGAEVKKPGSSVKVSCKASGFNIKDTYMHWVRQ APGQGLEWMGRIDPANGNSKYVPKFQGRVTITADTSTSTA YMELSSLRSEDYAVYYCAPFGYYVSDYAMAYWGQGLV TVSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPV TVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGT QTYICNVNHKPSNTKVDKKVEPKSCDKHTCTPPCPAPEAA GGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFN WYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWL NGKEYKCKVSNKALGAPIEKTISKAKGQPREPQVCTLPPSR DELTKNQVSLSCAVKGFYPSDIAVEWESNGQPENNYKTP PVLDSGDSFFLVSKLTVDKSRWQQGNVFSQSVMEALHN HYTQKSLSLSPGGGGSGGGGSREGPELSPDDPAGLLDLR QGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKE

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		DTKELVVAKAGVYYVFFQLELRRVVAGEGSGSVSLALHL QPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLS AGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPA GLGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQLVA QNVLLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVAK AGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGAA ALALTVDLPPASSEARNSAFGFQGRLLHLSAGQRLGVHLH TEARARHAWQLTQGATVLGLFRVTPEIPAGL
342	anti- CEA (T84.66-LCHA) Fc knob monomeric (71- 248) 4-1BBL chain	QVQLVQSGAEVKKPGSSVKVSCKASGFNIKDTYMHVVRQ APGQGLEWMGRIDPANGNSKYVPKFQGRVTITADTSTSTA YMELSSLRSEDYAVYYCAPFGYYVSDYAMAYWGQGTIV TVSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPV TVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGT QTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAA GGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFN WYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWL NGKEYKCKVSNKALGAPIEKTISKAKGQPREPQVYTLPPCR DELTKNQVSLWCLVKGFYPSDIAVEWESNGQPENNYKTP PVLDSDGSEFLYSKLTVDKSRWQQGNVFCFSVMHEALHN HYTQKSLSLSPGGGGSGGGGSREGPELSPDDPAGLLDLR QGMFAQLVAQNVLLIDGPLSWYSDPGLAGVSLTGGLSYKE DTKELVVAKAGVYYVFFQLELRRVVAGEGSGSVSLALHL QPLRSAAGAAALALTVDLPPASSEARNSAFGFQGRLLHLS AGQRLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPA GL
334	anti-CD19(8B8- 018) light chain	see Table 82

11.2.7 Preparation of monovalent CEA(T84.66) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding molecule with crossed CH1-CL domains with charged residues (Construct 5.7)

- 5 A polypeptide containing two ectodomains of 4-1BB ligand (71-254), separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned as depicted in Figure 29A: human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human CL. A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the human IgG1-CH domain, was cloned as described in Figure 29B: human 4-1BB ligand, (G4S)₂ connector,
- 10 human CH.

To improve correct pairing the following mutations have been introduced in the crossed CH-CL. In the dimeric 4-1BB ligand fused to human CL, E123R and Q124K. In the monomeric 4-1BB ligand fused to human CH1, K147E and K213E.

- 15 The variable region of heavy and light chain DNA sequences encoding a binder specific for CEA, clone T84.66, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala

mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831.

Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-CD19-Fc hole chain containing the

- 5 Y349C/T366S/L368A/Y407V mutations and the anti-CD19 light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and a CEA binding Fab. Construct 5.7 corresponds to Construct 5.1 as shown in **Figure 40**.

Table 88 shows the cDNA and amino acid sequences of the monovalent CEA(T84.66) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule with
10 crossed CH-CL and charged residues (construct 5.7).

Table 88: cDNA and amino acid sequences of monovalent CEA(T84.66) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion containing crossed CH-CL with charged residues (construct 5.7). * for charged residues

SEQ ID NO:	Description	Sequence
129	Nucleotide sequence Dimeric hu 4-1BBL (71-254) – CL* Fc knob chain	see Table 3
130	Nucleotide sequence Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
343	Nucleotide sequence anti-CEA(T84.66) Fc hole chain	GAGGTGCAGCTGCAGCAGTCTGGCGCCGAAGTGGTGGA ACCTGGCGCCTCTGTGAAGCTGAGCTGTACCGCCAGCGG CTTCAACATCAAGGACACCTACATGCACTGGGTCAAGC AGCGGCCTGAGCAGGGCCTGGAATGGATCGGCAGAATC GACCCCGCCAACGGCAACAGCAAATACGTGCCCAAGTT CCAGGGCAAGGCCACCATCACCGCCGACACCAGCAGCA ACACAGCCTACCTGCAGCTGACCAGCCTGACCTCCGAG GACACCGCCGTGTACTACTGCGCCCCCTTCGGCTACTAC GTGTCCGACTACGCCATGGCCTATTGGGGCCAGGGCAC AAGCGTGACCGTGTCTCTGCTAGCACCAAGGGCCCCCTC CGTGTTCCCCCCTGGCCCCCAGCAGCAAGAGCACCAGCG GCGGCACAGCCGCTCTGGGCTGCCTGGTCAAGGACTACT TCCCCGAGCCCGTGACCGTGTCTTGAACAGCGGAGCC CTGACCTCCGGCGTGCACACCTTCCCCGCCGTGCTGCAG AGTTCTGGCCTGTATAGCCTGAGCAGCGTGGTCACCGTG CCTTCTAGCAGCCTGGGCACCCAGACCTACATCTGCAAC GTGAACCACAAGCCCAGCAACACCAAGGTGGACAAGAA GGTGGAGCCCAAGAGCTGCGACAAAACCTCACACATGCC

		CACCGTGCCACGACCTGAAGCTGCAGGGGGACCGTCA GTCTTCCTCTTCCCCCAAAACCCAAGGACACCCTCATG ATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGAC GTGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTA CGTGGACGGCGTGGAGGTGCATAATGCCAAGACAAAGC CGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTC AGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGC AAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGG CGCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGC AGCCCCGAGAACCACAGGTGTGCACCCTGCCCCATCCC GGGATGAGCTGACCAAGAACCAGGTACGCCTCTCGTGC GCAGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGA GTGGGAGAGCAATGGGCAGCCGGAGAACAACACTACAAGA CCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCC TCGTGAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAG CAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCT CTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCT CCGGGTAAA
344	Nucleotide sequence anti- CEA(T84.66) light chain	GACATCGTGCTGACCCAGAGCCCTGCCTCTCTGGCCGTG TCTCTGGGACAGAGGGGCCACCATGTCTTGCAGAGCCGG CGAGAGCGTGGACATCTTCGGCGTGGGATTCTGCACTG GTATCAGCAGAAGCCCGGCCAGCCCCCAAGCTGCTGA TCTACAGAGCCAGCAACCTGGAAAGCGGCATCCCCGTG CGGTTTAGCGGCACCGGCAGCAGAACCGACTTCACCCT GATCATCGACCCCGTGGAAGCCGACGACGTGGCCACCT ACTACTGCCAGCAGACCAACGAGGACCCCTACACCTTTG GCGGAGGCACCAAGCTGGAAATCAAGCGTACGGTGGCT GCACCATCTGTCTTCATCTTCCCGCCATCTGATGAGCAG TTGAAATCTGGAAGTGCCTCTGTTGTGTGCCTGCTGAAT AACTTCTATCCCAGAGAGGCCAAAGTACAGTGGAAGGT GGATAACGCCCTCCAATCGGGTAACTCCCAGGAGAGTG TCACAGAGCAGGACAGCAAGGACAGCACCTACAGCCTC AGCAGCACCTGACGCTGAGCAAAGCAGACTACGAGAA ACACAAAGTCTACGCCTGCGAAGTCACCCATCAGGGCC TGAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGAGAG TGT
115	Dimeric hu 4- 1BBL (71-254) – CL* Fc knob chain	see Table 3
116	Monomeric hu 4-1BBL (71-254) – CH1*	see Table 3
345	anti- CEA (T84.66) Fc hole chain	EVQLQQSGAELVEPGASVKLSCTASGFNIKDTYMHVVKQ RPEQGLEWIGRIDPANGNSKYVPKFQGKATITADTSSNTAY LQLTSLTSEDYAVYYCAPFGYYVSDYAMAYWGQGTSTVTV SSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTV SWNSGALTSGVHTFPAVLQSSGLYSLSSVTVTPSSSLGTQT YICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGG PSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWY VDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNG KEYKCKVSNKALGAPIEKTISKAKGQPREPQVCTLPISRDE

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		LTKNQVSLSCAVKGFYPSDIAVEWESNGQPENNYKTTPPV LDSDGSFFLVSKLTVDKSRWQQGNVFSCSVMHEALHNHY TQKSLSLSPGK
346	anti- CEA (T84.66) light chain	DIVLTQSPASLA VSLGQRATMSCRAGESVDIFGVGFLHWY QQKPGQPPKLLIYRASNLESGIPVRFSGTGSRTDFTLIIDPVE ADDVATYYCQQTNE DPYTFGGGTKLEIKRTVAAPS VFIFPP SDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNS QESVTEQDSKDSYSLSSSTLTLSKADYEKHKVYACEVTHQ GLSSPVTKSFNRGEC

11.2.8 Preparation of bivalent CEA(T84.66) targeted 4-1BB ligand (71-254) trimer-containing Fc (kih) fusion antigen binding molecule (Construct 5.8)

A polypeptide containing two ectodomains of 4-1BB ligand (71-254), separated by (G4S)₂ linkers was fused to the C-terminus of human IgG1 Fc hole chain, as depicted in Figure 29C: human IgG1 Fc hole, (G4S)₂ connector, human 4-1BB ligand, (G4S)₂ connector, human 4-1BB ligand. A polypeptide containing one ectodomain of 4-1BB ligand (71-254) and fused to the C-terminus of human IgG1 Fc knob chain as described in Figure 29D: human IgG1 Fc knob, (G4S)₂ connector, human 4-1BB ligand.

The variable region of heavy and light chain DNA sequences encoding a binder specific for CEA, clone T84.66, were subcloned in frame with either the constant heavy chain of the hole, the knob or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831. Combination of the anti-CEA huIgG1 hole dimeric ligand chain containing the Y349C/T366S/L368A/Y407V mutations, the anti-CEA huIgG1 knob monomeric ligand chain containing the S354C/T366W mutations and the anti-CEA light chain allows generation of a heterodimer, which includes an assembled trimeric 4-1BB ligand and two CEA binding Fabs. Construct 5.8 corresponds to Construct 5.3 as shown in **Figure 40**.

Table 89 shows the cDNA and amino acid sequences of the bivalent CEA(T84.66) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion antigen binding molecule (construct 5.8).

Table 89: cDNA and amino acid sequences of bivalent CEA(T84.66) targeted split trimeric 4-1BB ligand (71-254) Fc (kih) PGLALA fusion (construct 5.8)

SEQ ID NO:	Description	Sequence
347	Nucleotide sequence anti-CEA(T84.66) Fc	GAGGTGCAGCTGCAGCAGTCTGGCGCCGAAGTGGTGGA ACCTGGCGCCTCTGTGAAGCTGAGCTGTACCGCCAGCGG CTTCAACATCAAGGACACCTACATGCACTGGGTCAAGC AGCGGCCTGAGCAGGGCCTGGAATGGATCGGCAGAATC

	hole dimeric 4-1BBL (71-254) chain	GACCCCGCCAACGGCAACAGCAAATACGTGCCCAAGTT CCAGGGCAAGGCCACCATCACCGCCGACACCAGCAGCA ACACAGCCTACCTGCAGCTGACCAGCCTGACCTCCGAG GACACCGCCGTGTACTACTGCGCCCCCTTCGGCTACTAC GTGTCCGACTACGCCATGGCCTATTGGGGCCAGGGCAC AAGCGTGACCGTGTCTCTGCTAGCACCAAGGGCCCCCTC CGTGTTCCCCCCTGGCCCCCAGCAGCAAGAGCACCAGCG GCGGCACAGCCGCTCTGGGCTGCCTGGTCAAGGACTACT TCCCCGAGCCCGTGACCGTGTCTTGAACAGCGGAGCC CTGACCTCCGGCGTGCACACCTTCCCCGCCGTGCTGCAG AGTTCTGGCCTGTATAGCCTGAGCAGCGTGGTCACCGTG CCTTCTAGCAGCCTGGGCACCCAGACCTACATCTGCAAC GTGAACCACAAGCCCAGCAACACCAAGGTGGACAAGAA GGTGGAGCCCAAGAGCTGCGACAAAACCTCACACATGCC CACCGTGCCAGCACCTGAAGCTGCAGGGGGACCGTCA GTCTTCCTCTTCCCCCAAAACCCAAGGACACCCTCATG ATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGAC GTGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTA CGTGGACGGCGTGGAGGTGCATAATGCCAAGACAAAGC CGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTC AGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGC AAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGG CGCCCCATCGAGAAAACCATCTCCAAGGCCAAAGGGC AGCCCCGAGAACCACAGGTGTGCACCCTGCCCCATCCC GGGATGAGCTGACCAAGAACCAGGTACGCCTCTCGTGC GCAGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGA GTGGGAGAGCAATGGGCAGCCGGAGAACAACCTACAAGA CCACGCCTCCCGTGTCTGGACTCCGACGGCTCCTTCTTCC TCGTGAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAG CAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCT CTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCT CCGGGTGGAGGCGGCGGAAGCGGAGGAGGAGGATCCA GAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGGA CTGCTGGACCTGCGGCAGGGCATGTTTGCTCAGCTGGTG GCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCCTGG TACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGGC GGCCTGAGCTACAAAGAGGACACCAAGAAGTGGTGGT GGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGGA ACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCTG TGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCTG CTGGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCTC CTGCCAGCTCCGAGGCCCGGAATAGCGCATTTGGGTTC AAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCTG GGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACGC CTGGCAGCTGACACAGGGCGCTACAGTGTCTGGGCCTGTT CAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCTCC AAGAAGCGAAGGCGGAGGCGGATCTGGCGGCGGAGGA TCTAGAGAGGGACCCGAACGTGCCCTGACGATCCAGC CGGGCTGCTGGATCTGAGACAGGGAATGTTCCGCCAGCT GGTGGCTCAGAATGTGCTGCTGATTGACGGACCTCTGAG CTGGTACTCCGACCCAGGGCTGGCAGGGGTGTCCCTGAC TGGGGGACTGTCTACAAAGAAGATACAAAAGAAGTGG TGGTGGCTAAAGCTGGGGTGTACTATGTGTTTTTTCAGC TGGAAGTGAAGCGGGTGGTGGCTGGGGAGGGCTCAGGA
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		TCTGTGTCCCTGGCTCTGCATCTGCAGCCACTGCGCTCT GCTGCTGGCGCAGCTGCACTGGCTCTGACTGTGGACCTG CCACCAGCCTCTAGCGAGGCCAGAAACAGCGCCTTCGG GTTCCAAGGACGCCTGCTGCATCTGAGCGCCGGACAGC GCCTGGGAGTGCATCTGCATACTGAAGCCAGAGCCCGG CATGCTTGGCAGCTGACTCAGGGGGCAACTGTGCTGGG ACTGTTTCGCGTGACACCTGAGATCCCTGCCGGACTGCC AAGCCCTAGATCAGAA
348	Nucleotide sequence anti- CEA(T84.66) Fc knob monomeric 4-1BBL (72-254) chain	GAGGTGCAGCTGCAGCAGTCTGGCGCCGAAGTGGTGGA ACCTGGCGCCTCTGTGAAGCTGAGCTGTACCGCCAGCGG CTTCAACATCAAGGACACCTACATGCACTGGGTCAAGC AGCGGCCTGAGCAGGGCCTGGAATGGATCGGCAGAATC GACCCCGCCAACGGCAACAGCAAATACGTGCCCAAGTT CCAGGGCAAGGCCACCATCACCGCCGACACCAGCAGCA ACACAGCCTACCTGCAGCTGACCAGCCTGACCTCCGAG GACACCGCCGTGTACTACTGCGCCCCCTTCGGCTACTAC GTGTCCGACTACGCCATGGCCTATTGGGGCCAGGGCAC AAGCGTGACCGTGTCTCTGCTAGACCAAGGGCCCATC GGTCTTCCCCCTGGCACCCCTCCTCCAAGAGCACCTCTGG GGGCACAGCGGCCCTGGGCTGCCTGGTCAAGGACTACT TCCCCGAACCGGTGACGGTGTCTGTGGAAGTCAAGGCGCC CTGACCAGCGGCGTGCACACCTTCCCCGGCTGTCTTACAG TCCTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTG CCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAAC GTGAATCACAAGCCCAGCAACACCAAGGTGGACAAGAA AGTTGAGCCCAAATCTTGTGACAAAACCTCACACATGCCC ACCGTGCCAGCACCTGAAGCTGCAGGGGGGACCGTCAG TCTTCCCTCTTCCCCCAAAACCAAGGACACCCTCATGA TCTCCCGGACCCCTGAGGTACATGCGTGGTGGTGGACG TGAGCCACGAAGACCCCTGAGGTCAAGTTCAACTGGTAC GTGGACGGCGTGGAGGTGCATAATGCCAAGACAAAGCC GCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCA GCGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGCA AGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCGGC GCCCCATCGAGAAAACCATCTCCAAGCCAAAGGGCA GCCCCGAGAACCACAGGTGTACACCCTGCCCCCTGCA GAGATGAGCTGACCAAGAACCAGGTGTCCCTGTGGTGT CTGGTCAAGGGCTTCTACCCCAGCGATATCGCCGTGGAG TGGGAGAGCAACGGCCAGCCTGAGAACAACCTACAAGAC CACCCCCCTGTGCTGGACAGCGACGGCAGCTTCTTCCCT GTACTCCAAACTGACCGTGGACAAGAGCCGGTGGCAGC AGGGCAACGTGTTTACGCTGCAGCGTGATGCACGAGGCC CTGCACAACCACTACACCCAGAAGTCCCTGAGCCTGAG CCCCGGCGGAGGCGGCGGAAGCGGAGGAGGAGGATCC AGAGAGGGCCCTGAGCTGAGCCCCGATGATCCTGCTGG ACTGCTGGACCTGCGGCAGGGCATGTTTGTCTCAGCTGGT GGCCCAGAACGTGCTGCTGATCGATGGCCCCCTGTCTTG GTACAGCGATCCTGGACTGGCTGGCGTGTCACTGACAGG CGGCCTGAGCTACAAAGAGGACACCAAAGAAGTGGTGG TGGCCAAGGCCGGCGTGTACTACGTGTTCTTTCAGCTGG AACTGCGGAGAGTGGTGGCCGGCGAAGGATCTGGCTCT GTGTCTCTGGCCCTGCATCTGCAGCCTCTGAGAAGCGCT GCTGGCGCTGCAGCTCTGGCACTGACAGTGGATCTGCCT CCTGCCAGCTCCGAGGCCCGGAATAGCGCATTTGGGTTT

		CAAGGCAGGCTGCTGCACCTGTCTGCCGGCCAGAGGCT GGGAGTGCATCTGCACACAGAGGCCAGGGCTAGACACG CCTGGCAGCTGACACAGGGCGCTACAGTGTCTGGGCCTG TTCAGAGTGACCCCCGAGATTCCAGCCGGCCTGCCTTCT CCAAGAAGCGAA
344	Nucleotide sequence anti- CEA(T84.66) light chain	see Table 88
349	anti- CEA(T84.66) Fc hole dimeric 4- 1BBL(71-254) chain	EVQLQQSGAELVEPGASVKLSCTASGFNIKDTYMHVVKQ RPEQGLEWIGRIDPANGNSKYVPKFQ GKATITADTSSNTAY LQLTSLTSED TAVYYCAPFGYYVSDYAMAYWGQGTSTVTV SSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVT SWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQT YICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGG PSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWY VDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNG KEYKCKVSNKALGAPIEKTISKAKGQPREPQVCTLPSSRDE LTKNQVSLSCAVKGFYPSDIAVEWESNGQPENNYKTTPPV LDS DGSFFLVSKLTVDKSRWQQGNVFSCSVMHEALHNHY TQKSLSLSPGGGGGSGGGGSREGPELSPDDPAGLLDLRQG MFAQLVAQNVLIDGPLSWYSDPGLAGVSLTGGLSYKEDT KELVVA KAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPL RSAAGAAALALTVDLPASSEARNSAFGFQGRLLHLSAGQ RLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPS PRSEGGGGSGGGGSREGPELSPDDPAGLLDLRQGMFAQLV AQNVLIDGPLSWYSDPGLAGVSLTGGLSYKEDTKELVVA KAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPLRSAAGA AALALTVDLPASSEARNSAFGFQGRLLHLSAGQRLGVHL HTEARARHAWQLTQGATVLGLFRVTPEIPAGLPSRSE
350	anti- CEA(T84.66) Fc knob monomeric 4-1BBL (71-254) chain	EVQLQQSGAELVEPGASVKLSCTASGFNIKDTYMHVVKQ RPEQGLEWIGRIDPANGNSKYVPKFQ GKATITADTSSNTAY LQLTSLTSED TAVYYCAPFGYYVSDYAMAYWGQGTSTVTV SSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVT SWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQT YICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEAAGG PSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFNWY VDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNG KEYKCKVSNKALGAPIEKTISKAKGQPREPQVYTLPPCRDE LTKNQVSLWCLVKGFYPSDIAVEWESNGQPENNYKTTPPV LDS DGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHY TQKSLSLSPGGGGGSGGGGSREGPELSPDDPAGLLDLRQG MFAQLVAQNVLIDGPLSWYSDPGLAGVSLTGGLSYKEDT KELVVA KAGVYYVFFQLELRRVVAGEGSGSVSLALHLQPL RSAAGAAALALTVDLPASSEARNSAFGFQGRLLHLSAGQ RLGVHLHTEARARHAWQLTQGATVLGLFRVTPEIPAGLPS PRSE
346	anti- CEA(T84.66) light chain	see Table 88

11.3 Preparation of untargeted split trimeric 4-1BB ligand Fc fusion molecules and human IgG as control molecules

11.3.1 Preparation of untargeted human 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (Control molecules)

5 These control molecules were prepared as described above for the CEA targeted construct 3.1 (termed control B), 3.3 (termed control C), 3.4 (termed control D) and 3.5 (termed control E) with the only difference that the anti-CD19 binder (VH-VL) was replaced by a germline control, termed DP47, not binding to the antigen (see Figure 40). The cDNA and amino acid sequences of control B, the monovalent DP47-untargeted split trimeric 4-1BB ligand (71-254) Fc (kih)
 10 fusion containing crossed CH-CL with charged residues, are shown in **Table 68** above (see Example 7.3.1). **Table 69** shows the cDNA and amino acid sequences of the bivalent DP47-untargeted split trimeric 4-1BB ligand (71-254) Fc (kih) fusion, control C. **Table 70** shows the cDNA and amino acid sequences of the monovalent DP47-untargeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion containing CH-CL cross with charged residues, control D. **Table 71**
 15 shows the cDNA and amino acid sequences of the monovalent DP47-untargeted split trimeric 4-1BB ligand (71-248) Fc (kih) fusion without charged residues in the CH-CL cross, control E.

11.3.2 Antibodies as control molecules

An additional control used in the assays, termed control F, was an untargeted DP47, germline control, human IgG1, containing the Pro329Gly, Leu234Ala and Leu235Ala mutations,
 20 to abrogate binding to Fc gamma receptors. The cDNA and amino acid sequences of control F can be found in **Table 73** above.

11.4 Production of CEA- targeted split trimeric 4-1BB ligand Fc fusion antigen binding molecules and their control molecules

25 The targeted and untargeted split trimeric 4-1BB ligand Fc (kih) fusion antigen binding molecule encoding sequences were cloned into a plasmid vector, which drives expression of the insert from an MPSV promoter and contains a synthetic polyA sequence located at the 3' end of the CDS. In addition, the vector contains an EBV OriP sequence for episomal maintenance of the plasmid.

30 The split trimeric 4-1BB ligand Fc (kih) fusion was produced by co-transfecting HEK293-EBNA cells with the mammalian expression vectors using polyethylenimine. The cells were transfected with the corresponding expression vectors. For variants 1,2,4,5 and its control B, D and E, at a 1:1:1:1 ratio ("vector dimeric ligand-CL- knob chain": "vector monomeric ligand fusion-CH1": "vector anti-CEA Fab-hole chain": "vector anti-CEA light chain"). For variant 3,

6 and it's control C, at a 1:1:1 ratio ("vector huIgG1 Fc hole dimeric ligand chain": "vector huIgG1 Fc knob monomeric ligand chain": "vector anti-CEA light chain"). Human IgGs, used as control in the assay, were produced as for the bispecific construct (for transfection only a vector for light and a vector for heavy chain were used at a 1:1 ratio).

- 5 For production in 500 mL shake flasks, 300 million HEK293 EBNA cells were seeded 24 hours before transfection. For transfection cells were centrifuged for 10 minutes at 210 x g, and the supernatant was replaced by 20mL pre-warmed CD CHO medium. Expression vectors (200 µg of total DNA) were mixed in 20 mL CD CHO medium. After addition of 540 µL PEI, the solution was vortexed for 15 seconds and incubated for 10 minutes at room temperature.
- 10 Afterwards, cells were mixed with the DNA/PEI solution, transferred to a 500 mL shake flask and incubated for 3 hours at 37°C in an incubator with a 5% CO₂ atmosphere. After the incubation, 160 mL of Excell medium supplemented with 6mM L-Glutamine, 5g/L PEPSOY and 1.2 mM valproic acid was added and cells were cultured for 24 hours. One day after transfection 12% Feed (amino acid and glucose) were added. After culturing for 7 days, the
- 15 supernatant was collected by centrifugation for 30-40 minutes at least 400 x g. The solution was sterile filtered (0.22 µm filter), supplemented with sodium azide to a final concentration of 0.01 % (w/v), and kept at 4 °C.

- The split trimeric 4-1BB ligand Fc (kih) fusion, as well as the IgG, was purified from cell culture supernatants by affinity chromatography using Protein A, followed by size exclusion
- 20 chromatography. For affinity chromatography, the supernatant was loaded on a MabSelect Sure column (CV = 5-15 mL, resin from GE Healthcare) equilibrated with Sodium Phosphate (20 mM), Sodium Citrate (20 mM) buffer (pH 7.5). Unbound protein was removed by washing with at least 6 column volumes of the same buffer. The bound protein was eluted using either a linear gradient (20 CV) or a step elution (8 CV) with 20 mM sodium citrate, 100 mM Sodium chloride,
- 25 100 mM Glycine buffer (pH 3.0). For the linear gradient an additional 4 column volumes step elution was applied.

- The pH of collected fractions was adjusted by adding 1/10 (v/v) of 0.5M sodium phosphate, pH8.0. The protein was concentrated prior to loading on a HiLoad Superdex 200 column (GE Healthcare) equilibrated with 20 mM Histidine, 140 mM sodium chloride, 0.01% (v/v) Tween20
- 30 solution of pH 6.0.

- The protein concentration was determined by measuring the optical density (OD) at 280 nm, using a molar extinction coefficient calculated on the basis of the amino acid sequence. Purity and molecular weight of the targeted trimeric 4-1BB ligand Fc (kih) fusion antigen binding molecule was analyzed by SDS-PAGE in the presence and absence of a reducing agent
- 35 (5 mM 1,4-dithiotreitol) and staining with Coomassie SimpleBlue™ SafeStain (Invitrogen USA)

or CE-SDS using Caliper LabChip GXII (Perkin Elmer). The aggregate content of samples was analyzed using a TSKgel G3000 SW XL analytical size-exclusion column (Tosoh) equilibrated in 25 mM K₂HPO₄, 125 mM NaCl, 200 mM L-Arginine Monohydrochloride, 0.02 % (w/v) NaN₃, pH 6.7 running buffer at 25°C.

- 5 **Table 90** summarizes the yield and final monomer content of the CEA targeted split trimeric 4-1BB ligand Fc (kih) fusion antigen binding molecules.

Table 90: Biochemical analysis of CEA targeted split trimeric 4-1BB ligand Fc (kih) fusion.

Construct	Monomer [%] (SEC)	Yield [mg/l]
monovalent CEA(T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-248) Fc fusion containing CH-CL cross with charged residues (construct 5.4)	98	1.4
bivalent CEA(T84.66-LCHA) targeted split trimeric 4-1BB ligand (71-248) Fc fusion (construct 5.6)	98	0.4
monovalent CEA(T84.66) targeted split trimeric 4-1BB ligand (71-254) Fc fusion containing CH-CL cross with charged residues (construct 5.7)	97	15
bivalent CEA(T84.66) targeted split trimeric 4-1BB ligand (71-254) Fc fusion (construct 5.8)	96	2

- 10 **Table 91** summarizes the yield and final monomer content of the DP47 untargeted split trimeric 4-1BB ligand Fc (kih) fusion molecules, both monovalent (control B, D and E) and bivalent (control C), and of the germline DP47 human IgG1 PGLALA (control F).

Table 91: Biochemical analysis of DP47 untargeted split trimeric 4-1BB ligand Fc (kih) fusion

Construct	Monomer [%] (SEC)	Yield [mg/l]
monovalent DP47-untargeted split trimeric human 4-1BB ligand (71-254) Fc (kih) fusion (control B)	99	15.4
bivalent DP47 untargeted split trimeric human 4-1BB ligand (71-254) Fc (kih) fusion (control C)	98	12.6
monovalent DP47-untargeted split trimeric human 4-1BB ligand (71-254) Fc (kih) fusion (control D)	99.5	25.9
monovalent DP47-untargeted split trimeric human 4-1BB ligand (71-254) Fc (kih) fusion (control E)	93.3	4.1
germline DP47 human IgG1 PGLALA	100	50

Example 12

Functional characterization of the CEA targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules

12.1 Surface plasmon resonance (simultaneous binding)

5 Production of hu NA3B3A2 as antigen for CEA targeted trimeric split 4-1BBL constructs

The antigen used to assess binding by SPR to CEA was a hybrid molecule composed of A3 and B3 domains from human CEACAM5 (CEA) and N and A2 domains from human CEACAM1 (BGP1) similarly to what has been described for NABA (Durbin H. et al, Proc Natl Acad Sci U S A. 1994 May 10;91(10):4313-7). The antigen is termed here NA3B3A2 and a
10 schematic description can be found in **Figure 41A**.

Table 92 shows the nucleotide and amino acid sequences of hu NA3B3A2-avi-His.

Table 92: Nucleotide and amino acid sequences of hu NA3B3A2-avi-His

SEQ ID NO:	Antigen	Sequence
351	nucleotide sequence hu NA3B3A2-avi His	CAGCTGACCACCGAGTCCATGCCCTTCAACGTGGCCG AGGGCAAAGAGGTGCTGCTGCTGGTCCACAACCTGCC CCAGCAGCTGTTTCGGCTACAGCTGGTACAAGGGCGAG CGGGTGGACGGCAACCGGCAGATCGTGGGCTACGCCA TCGGCACCCAGCAGGCCACACCCGGCCCTGCCAATAG CGGCAGAGAGACAATCTACCCCAACGCCAGCCTGCTG ATCCAGAACGTGACCCAGAACGACACCGGCTTCTACA CACTCCAAGTCATCAAGAGCGACCTGGTCAACGAGGA AGCCACCGGCCAGTTCCACGTGTACCCCGAGCTGCCC AAGCCAGCATCAGCAGCAACAACAGCAAGCCCGTGG AAGATAAGGACGCCGTGGCCTTTACCTGCGAGCCCGA GGCCAGAACACCACCTACCTGTGGTGGGTCAACGGC CAGAGCCTGCCCGTGTCCCCCAGACTCCAGCTGAGCA ACGGCAACAGAACCCCTGACCCTGTTCAACGTGACCCG GAATGACGCCAGAGCCTACGTGTGCGGCATCCAGAAC AGCGTGTCCGCCAACCGCAGCGACCCCGTGACCCTGG ATGTGCTGTACGGCCCCGACACCCCCATCATCAGCCCC CCTGACAGCAGCTACCTGAGCGGCGCCAACCTGAACC TGAGCTGCCACAGCGCCAGCAACCCCGAGCCCTCAGTA CAGCTGGCGGATCAACGGCATCCCCCAGCAGCACACC CAGGTGCTGTTTATCGCCAAGATCACCCCCAACAACA ACGGCACCTACGCCTGCTTCGTGTCCAACCTGGCCACC GGCCGGAACAACAGCATCGTGAAGTCCATCACCGTGT CCGCCTCCCTGAGCCCCGTGGTGGCCAAGCCTCAGAT CAAGGCCAGCAAGACCACCGTGACCGGCGACAAGGA CAGCGTGAACCTGACCTGCTCCACCAACGATACCGGC ATCAGCATCCGGTGGTTCTTCAAGAATCAGTCCCTGCC

-332-

		CAGCAGCGAGCGGATGAAGCTGAGCCAGGGCAACAT CACCCTGTCCATCAACCCCGTGAAAAGAGAGGACGCC GGCACCTATTGGTGCGAGGTGTTCAACCCCATCAGCA AGAACCAGAGCGACCCCATCATGCTGAACGTGAACTA CAACGCCCTGCCCAAGAAAACCTGATCAATGTTGAT CTGGAAGTGCTGTTCCAGGGCCCAGGCAGCGGCCTGA ACGACATCTTCGAAGCCCAGAAAATCGAGTGGCACGA GGCCAGAGCCCACCACCACCATCACCAC
352	human NA3B3A2-avi- His	QLTTESMPFNVAEGKEVLLL VHNLPQQLFGYSWKGER VDGNRQIVGYAIGTQQATPGPANSGRETIYPNASLLIQNV TQNDTGfYTLQVIKSDLVNEEATGQFHVYPELPKPSISSN NSKPVEDKDAVAFTCEPEAQNTTYLWWVNGQSLPVSPR LQLSNGNRTLTLFNVTRNDARAYVCGIQNSVSANRSDPV TLDVLYGPDTPHSPDSSYLSGANLNLCHSASNPSPQYS WRINGIPQQHTQVLFIKITPNNNGTYACFVSNLATGRN NSIVKSITVSASLSPVVAKPQIKASKTTVTGDKDSVNLTC STNDTGISIRWFFKNQSLPSSERMKLSQGNITLSINPVKRE DAGTYWCEVFNPISKNQSDPIMLNVNYNALPQENLINVD LEVLFQGPGLNDIFEAQKIEWHEARAHHHHHH

Protein production was performed as described above for the Fc-fusion protein (Example 7.1.1). Secreted proteins were purified from cell culture supernatants by chelating chromatography, followed by size exclusion chromatography. The first chromatographic step was performed on a NiNTA Superflow Cartridge (5ml, Qiagen) equilibrated in 20 mM sodium phosphate, 500 nM sodium chloride, pH7.4. Elution was performed by applying a gradient over 12 column volume from 5% to 45% of elution buffer (20 mM sodium phosphate, 500 nM sodium chloride, 500 mM Imidazole, pH7.4).

The protein was concentrated and filtered prior to loading on a HiLoad Superdex 75 column (GE Healthcare) equilibrated with 20 mM Histidine, 140 mM NaCl, 0.01% Tween-20 pH 6.0. **Table 93** summarizes the yield and final monomer content of human NA3B3A2-avi-His.

Table 93: Biochemical analysis of human NA3B3A2-avi-His

Construct	Monomer [%] (SEC)	Yield [mg/l]
human NA3B3A2-avi-His	88	14.1

The capacity of binding simultaneously human 4-1BB Fc (kih) and human NA3B3A2 was assessed by surface plasmon resonance (SPR). All SPR experiments were performed on a Biacore T200 at 25 °C with HBS-EP as running buffer (0.01 M HEPES pH 7.4, 0.15 M NaCl, 3 mM EDTA, 0.005% Surfactant P20, Biacore, Freiburg/Germany). Biotinylated human 4-1BB Fc (kih) was directly coupled to a flow cell of a streptavidin (SA) sensor chip. Immobilization levels up to 250 resonance units (RU) were used.

The CEA targeted trimeric split 4-1BBL constructs (constructs 5.4, 5.6, 5.7 and 5.8) were passed at a concentration range of 200 nM with a flow of 30 μ L/minute through the flow cells over 90 seconds and dissociation was set to zero seconds. Human NA3B3A2 was injected as second analyte with a flow of 30 μ L/minute through the flow cells over 90 seconds at a concentration of 500 nM (**Figure 41B**). The dissociation was monitored for 120 seconds. Bulk refractive index differences were corrected for by subtracting the response obtained in a reference flow cell, where no protein was immobilized.

As can be seen in the graphs of **Figure 42**, all bispecific constructs could bind simultaneously human 4-1BB and human NA3B3A2.

12.2. Binding on activated human PMBCs of the CEA-targeted 4-1BB ligand trimer-containing Fc (kih) fusion antigen binding molecules

To determine binding of 4-1BBL trimer-containing Fc fusion antigen binding molecules to human PBMCs, different titrated concentrations of the CEA-targeted 4-1BBL trimer-containing Fc fusion antigen binding molecules were used in the assay as described in Example 5.2.

Figure 43 shows the binding of Constructs 5.4, 5.6, 5.7 and 5.8 as prepared in Example 11 on activated 4-1BB-expressing CD4⁺ T cells and CD8⁺ T cells, respectively. Gates were set on living CD45⁺ CD3⁺ CD4⁺ or CD45⁺ CD3⁺ CD8⁺ T cells and MFI of PE-conjugated AffiniPure anti-human IgG IgG Fc γ -fragment-specific goat F(ab')₂ fragment were blotted against the titrated concentration of targeted split trimeric 4-1BB ligand Fc fusion variants.

Table 94 shows the EC₅₀ values as measured for Constructs 5.4, 5.6, 5.7 and 5.8 and control molecules.

Table 94: Binding on activated 4-1BB-expressing CD4⁺ T cells and CD8⁺ T cells

Construct	EC ₅₀ [nM] 4-1BB ⁺ CD8 ⁺	EC ₅₀ [nM] 4-1BB ⁺ CD4 ⁺
Control B	0.05	0.26
Control C	0.02	0.30
Control D	0.04	0.28
Control E	0.13	1.22
5.4	0.13	0.35
5.6	0.06	0.34
5.7	0.0004	0.36
5.8	0.17	0.38

12.2 Binding to CEA-expressing tumor cells

For binding assays on CEA-expressing tumor cells, the following human CEA-expressing lymphoma cell lines were used: CEA-expressing tumor cell lines human gastric cancer cell line MKN-45 (ATCC TCP-1008) and human colorectal adenocarcinoma cell line LS180 (ATCC CL-187). The assays were performed as described for the FAP-expressing MV-3 and WM-266-4 tumor cell lines in Example 5.3.

Gates were set on living tumor cells and MFI of PE-conjugated AffiniPure anti-human IgG IgG Fc γ -fragment-specific goat F(ab')₂ fragment were blotted against the titrated concentration of targeted split trimeric 4-1BB ligand Fc fusion constructs.

Figure 44 shows the binding of Constructs 5.7 as prepared in Example 11.2.7 to human-CEA expressing human gastric cell line MKN-45 (left) and human colorectal adenocarcinoma cells line LS180 (right). **Table 95** shows the EC₅₀ values as measured for human-CEA expressing human gastric cell line MKN-45.

Table 95: Binding to CEA-expressing tumor cells

Construct	EC ₅₀ [nM]	EC ₅₀ [nM]
	MKN45-8	LS180
5.7	11.6	14.4

Example 13

Biological activity of the CEA-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules

13.1. NF- κ B activation in HeLa cells expressing human 4-1BB

HeLa cells expressing human 4-1BB and NF- κ B-luciferase were generated as described in Example 6.1.

NF- κ B activation in Hela cells expressing human 4-1BB co-cultured with human CEA-expressing tumor cells

NF- κ B-luciferase human-4-1BB HeLa cells were harvested and resuspended in DMEM medium supplied with 10 % (v/v) FBS and 1 % (v/v) GlutaMAX-I to a concentration of 0.2 x 10⁶ cells/ml. 100 μ l (2 x 10⁴ cells) of this cell suspension were transferred to each well of a sterile white 96-well flat bottom tissue culture plate with lid (greiner bio-one, Cat. No. 655083) and the plate were incubated at 37 °C and 5 % CO₂ overnight. The next day 50 μ L of medium

containing titrated concentrations of CEA-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (CEA split 4-1BBL trimer) or DP47-untargeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules (DP47 split 4-1BBL trimer) were added. CEA-expressing tumor cell lines human gastric cancer cell line MKN-45 (ATCC TCP-1008) was
 5 resuspended in DMEM medium supplied with 10 % (v/v) FBS and 1 % (v/v) GlutaMAX-I to a concentration of 2×10^6 cells/ml.

Suspension of CEA-expressing B cell lymphoma cell (50 μ l, final ratio 1:5) or only medium were added to each well and plates were incubated for 6 hours at 37 °C and 5 % CO₂. Cells were washed two times with 200 μ L/well DPBS. 40 μ l freshly prepared Reporter Lysis
 10 Buffer (Promega, Cat-No: E3971) were added to each well and the plate were stored over night at -20 °C. The next day frozen cell plate and Detection Buffer (Luciferase 1000 Assay System, Promega, Cat. No. E4550) were thawed at room temperature. 100 μ L of detection buffer were added to each well and luciferase activity was measured as fast as possible using a SpectraMax
 15 M5/M5e microplate reader and a SoftMax Pro Software (Molecular Devices) counting light emission in URL (units of released light for 0.5s/well) or Victor3 1420 multilabel counter plate reader (Perkin Elmer) and the Perkin Elmer 2030 Manager Software counting light emission as counts per seconds (CPS) and blotted against the concentration of tested constructs.

CEA-targeted 4-1BB ligand trimer-containing Fc fusion antigen binding molecules
 Constructs 5.7 and 5.8 triggered activation of the NF-kB signaling pathway in the reporter cell
 20 line in the presence of human gastric cancer cell line MKN-45 cells. In contrast, the untargeted control molecules failed to trigger such an effect at any of the tested concentrations (Figure 45). Table 96 shows the corresponding EC₅₀ values.

Table 96: Binding to CEA-expressing tumor cells

Construct	EC ₅₀ [nM] MKN-45 no tumor cells	EC ₅₀ [nM] MKN45
5.4	3.1	0.34
5.6	2.05	0.21
5.7	0.85	0.05
5.8	1.52	0.45

Example 14

14.1 Preparation of FAP targeted OX40 ligand trimer-containing Fc fusion antigen binding molecules

The DNA sequence encoding part of the ectodomain (amino acids 51-183) of human OX40 ligand was synthesized according to the P23510 sequence of Uniprot database. To decrease heterogeneity of human Ox40 ligand due to glycosylation asparagine residues at position 90 and 114 were mutated to aspartic acid by site-directed mutagenesis (according to Compaa D.M., Hymowitz S.G., Structure (2006) 14(8), 1321-30).

A polypeptide containing two ectodomains of OX40 ligand, separated by (G4S)₂ linkers, and fused to the human IgG1-CL domain, was cloned as depicted in **Figure 46A**: human OX40 ligand, (G4S)₂ connector, human OX40 ligand, (G4S)₂ connector, human CL.

A polypeptide containing one ectodomain of OX40 ligand and fused to the human IgG1-CH domain, was cloned as described in **Figure 46B**: human OX40 ligand, (G4S)₂ connector, human CH.

To improve correct pairing the following mutations have been introduced in the crossed CH-CL. In the dimeric 4-1BB ligand fused to human CL, E123R and Q124K. In the monomeric 4-1BB ligand fused to human CH1, K147E and K213E.

The generation and preparation of the FAP binders is described in WO 2012/020006 A2, which is incorporated herein by reference.

The variable region of heavy and light chain DNA sequences encoding a binder specific for FAP, clone 28H11, were subcloned in frame with either the constant heavy chain of the hole or the constant light chain of human IgG1. The Pro329Gly, Leu234Ala and Leu235Ala mutations have been introduced in the constant region of the knob and hole heavy chains to abrogate binding to Fc gamma receptors according to the method described in WO 2012/130831.

Combination of the dimeric ligand-Fc knob chain containing the S354C/T366W mutations, the monomeric CH1 fusion, the targeted anti-FAP-Fc hole chain containing the Y349C/T366S/L368A/Y407V mutations and the anti-FAP light chain allows generation of a heterodimer, which includes an assembled trimeric OX40 ligand and a FAP binding Fab (**Figure 46C**, Construct 6.1).

Table 97 shows the cDNA and amino acid sequences of the monovalent CEA (T84.66-LCHA) targeted split trimeric OX40 ligand (51-183) Fc (kih) fusion antigen binding molecule with crossed CH-CL and charged residues (construct 6.1).

Table 97: cDNA and amino acid sequences of monovalent FAP(28H1) targeted split trimeric OX40 ligand Fc (kih) fusion containing CH-CL cross with charged residues (construct 6.1). * for charged residues

SEQ ID NO:	Description	Sequence
353	Nucleotide sequence Dimeric hu OX40L (51-183) – CL* Fc knob chain	CAGGTGTCCACAGATACCCCAGAATCCAGAGCATCAA GGTGCAGTTCACCGAGTACAAGAAAGAGAAGGGCTTCA TCCTGACCAGCCAGAAAGAGGACGAGATCATGAAGGTG CAGGACAACAGCGTGATCATCAACTGCGACGGCTTCTA CCTGATCAGCCTGAAGGGCTACTTCAGCCAGGAAGTGG ACATCAGCCTGCACTACCAGAAGGACGAGGAACCCCTG TTCCAGCTGAAGAAAGTGCGGAGCGTGAACAGCCTGAT GGTGGCCAGCCTGACCTACAAGGACAAGGTGTACCTGA ACGTGACCACCGACAACACCAGCCTGGACGACTTCCAC GTGAACGGCGCGGAGCTGATCCTGATTCACCAGAACCC CGGCGAGTTCTGCGTGCTGGGAGGCGGAGGATCTGGCG GAGGCGGATCTCAGGTGTACACCCGCTACCCCCGGATTCT AGTCCATTAAAGGTGCAGTTTACAGAGTATAAGAAAGAA AAAGGCTTTATTCTGACTTCCCAGAAAGAAGATGAGATT ATGAAGGTGCAGGATAATTCTGTGATCATCAATTGTGAC GGCTTCTACCTGATCAGCCTGAAGGGCTACTTCAGCCAG GAAGTGGACATCAGCCTGCACTACCAGAAGGACGAGGA ACCCCTGTTCCAGCTGAAGAAAGTGCGGAGCGTGAACA GCCTGATGGTGGCCAGCCTGACCTACAAGGACAAGGTG TACCTGAACGTGACCACCGACAACACCAGCCTGGACGA CTTCCACGTGAACGGCGGCGAGCTGATCCTGATCCACCA GAACCCTGGCGAGTTCTGCGTGCTGGGAGGCGGAGGCT CCGGAGGGGGAGGATCTCGTACGGTGGCTGCACCATCT GTCTTTATCTTCCCACCCAGCGACCGGAAGCTGAAGTCT GGCACAGCCAGCGTCGTGTGCCTGCTGAATAACTTCTAC CCCCGCGAGGCCAAGGTGCAGTGGAAGGTGGACAATGC CCTGCAGAGCGGCAACAGCCAGGAAAGCGTGACCGAGC AGGACAGCAAGGACTCCACCTACAGCCTGAGCAGCACC CTGACCCTGAGCAAGGCCGACTACGAGAAGCACAAGGT GTACGCCTGCGAAGTGACCCACCAGGGCCTGTCTAGCCC CGTGACCAAGAGCTTCAACCGGGGCGAGTGCGACAAGA CCCACACCTGTCTCCATGCCCTGCCCTGAAGCTGCTG GCGGCCCTAGCGTGTTCTGTTCCTCCCAAAGCCCAAGG ACACCCTGATGATCAGCCGGACCCCTGAAGTGACCTGC GTGGTGGTGGATGTGTCCACGAGGACCCTGAAGTGAA GTTCAATTGGTACGTGGACGGCGTGGAAGTGACAATG CCAAGACCAAGCCGCGGGAGGAGCAGTACAACAGCACG TACCGTGTGGTCAGCGTCCTCACCGTCCTGCACCAGGAC TGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAA CAAAGCCCTCGGCGCCCCCATCGAGAAAACCATCTCCA AAGCCAAAGGGCAGCCCCGAGAACCACAGGTGTACACC CTGCCCCCATGCCGGGATGAGCTGACCAAGAACCAGGT CAGCCTGTGGTGCCTGGTCAAAGGCTTCTATCCCAGCGA CATCGCCGTGGAGTGAGGAGCAATGGGCAGCCGGAGA ACAACTACAAGACCACGCCTCCCGTGCTGGACTCCGAC

		GGCTCCTTCTTCCTCTACAGCAAGCTCACCGTGGACAAG AGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTG ATGCATGAGGCTCTGCACAACCACTACACGCAGAAGAG CCTCTCCCTGTCTCCGGGTAAG
354	Nucleotide sequence Monomeric hu OX40L (51-183) – CH1*	CAGGTGTCCACAGATACCCAGAATCCAGAGCATCAA GGTGCAGTTCACCGAGTACAAGAAAGAGAAGGGCTTCA TCCTGACCAGCCAGAAAGAGGACGAGATCATGAAGGTG CAGGACAACAGCGTGATCATCAACTGCGACGGCTTCTA CCTGATCAGCCTGAAGGGCTACTTCAGCCAGGAAGTGG ACATCAGCCTGCACTACCAGAAGGACGAGGAACCCCTG TTCCAGCTGAAGAAAGTGCAGGAGCGTGAACAGCCTGAT GGTGGCCAGCCTGACCTACAAGGACAAGGTGTACCTGA ACGTGACCACCGACAACACCAGCCTGGACGACTTCCAC GTGAACGGCGGCGAGCTGATCCTGATTACCAAGAACCC CGGCGAGTTCTGCGTGCTGGGAGGCGGAGGTTCGGAG GCGGAGGATCTGCTAGCACAAAGGGCCCCAGCGTGTTT CCTCTGGCCCCTAGCAGCAAGAGCACATCTGGCGGAAC AGCCGCCCTGGGCTGCCTGGTGGAAGATTACTTCCCCGA GCCCCGTGACCGTGTCTGGAATTCTGGCGCCCTGACAAG CGGCGTGCACACCTTTCCAGCCGTGCTGCAGAGCAGCG GCCTGTACTCTCTGAGCAGCGTCGTGACAGTGCCCAGCA GCTCTCTGGGCACCCAGACCTACATCTGCAACGTGAACC ACAAGCCCAGCAACACCAAGGTGGACGAGAAGGTGGA ACCCAAGTCCTGC
68	Nucleotide sequence anti- FAP(28H1) Fc hole chain	see Table 2
69	Nucleotide sequence anti- FAP(28H1) light chain	see Table 2
355	Dimeric hu OX40L (51-183) – CL* Fc knob chain	QVSHRYPRIQSIKVQFTEYKKEKGFIILTSQKEDEIMKVQDN SVIINCDGFYLISLKGYSQEVDISLHYQKDEEPLFQLKKVR SVNSLMVASLTYKDKVYLNVTDDNTSLDDFHVNGGELILI HQNPGEFCVLGGGGSGGGGSQVSHRYPRIQSIKVQFTEYK KEKGFIILTSQKEDEIMKVQDNSVIINCDGFYLISLKGYSQE VDISLHYQKDEEPLFQLKKVRSVNSLMVASLTYKDKVYLN VTDDNTSLDDFHVNGGELILIHQNPGEFCVLGGGGSGGGG SRTVAAPSVFIFPPSDRKLKSGTASVCLLNPFYFPAKVK WKVDNALQSGNSQESVTEQDSKDSSTLSLTLSKADYE KHKVYACEVTHQGLSPVTKSFNRGECDKTHCPPCPAPE AAGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVK FNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQD WLNGKEYKCKVSNKALGAPIEKTISKAKGQPREPQVYTL PCRDELTKNQVSLWCLVKGFYPSDIAVEWESNGQPENNYK TTPPVLDSDGSFFLYSKLTVDKSRWQQGNVVFSCSVMHEAL HNHYTQKSLSLSPGK
356	Monomeric hu OX40L (51-183)	QVSHRYPRIQSIKVQFTEYKKEKGFIILTSQKEDEIMKVQDN SVIINCDGFYLISLKGYSQEVDISLHYQKDEEPLFQLKKVR SVNSLMVASLTYKDKVYLNVTDDNTSLDDFHVNGGELILI

	– CH1*	HQNPGEFCVLGGGGSGGGGSASTKGPSVFPLAPSSKSTSGG TAALGCLVEDYFPEPVTVSWNSGALTSGVHTFPAVLQSSG LYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDEKVEPKS C
18	anti- FAP(28H1) Fc hole chain	see Table 2
19	anti- FAP(28H1) light chain	see Table 2

14.2 Preparation of untargeted human IgG1 as Control F

A control molecule used in the assays, termed control F (**Figure 46D**), was an untargeted DP47, germline control, human IgG1, containing the Pro329Gly, Leu234Ala and Leu235Ala mutations, to abrogate binding to Fc gamma receptors according to the method described in International Patent Appl. Publ. No. WO 2012/130831). Its preparation is described in Example 2.3, **Table 29** shows the cDNA and amino acid sequences of the cDNA and amino acid sequences of the untargeted DP47 huIgG1 PGLALA (Control F).

14.3 Production of FAP- targeted split trimeric OX40 ligand Fc fusion antigen binding molecules and their control molecules

The targeted and untargeted split trimeric OX40 ligand Fc (kih) fusion encoding sequences were cloned into a plasmid vector, which drives expression of the insert from an MPSV promoter and contains a synthetic polyA sequence located at the 3' end of the CDS. In addition, the vector contains an EBV OriP sequence for episomal maintenance of the plasmid.

The split trimeric Ox40 ligand Fc (kih) fusion was produced by co-transfecting HEK293-EBNA cells with the mammalian expression vectors using polyethylenimine. The cells were transfected with the corresponding expression vectors. For variants 1,2,4,5 and it's control B, D and E, at a 1:1:1:1 ratio ("vector dimeric ligand-CL- knob chain": "vector monomeric ligand fusion-CH1": "vector anti- FAP Fab-hole chain": "vector anti-FAP light chain"). For variant 3, 6 and it's control C, at a 1:1:1 ratio ("vector huIgG1 Fc hole dimeric ligand chain": "vector huIgG1 Fc knob monomeric ligand chain": "vector anti-FAP light chain"). Human IgGs, used as control in the assay, were produced as for the bispecific construct (for transfection only a vector for light and a vector for heavy chain were used at a 1:1 ratio).

For production in 500 mL shake flasks, 300 million HEK293 EBNA cells were seeded 24 hours before transfection. For transfection cells were centrifuged for 10 minutes at 210 x g, and the supernatant was replaced by 20mL pre-warmed CD CHO medium. Expression vectors (200 µg of total DNA) were mixed in 20 mL CD CHO medium. After addition of 540 µL PEI, the

solution was vortexed for 15 seconds and incubated for 10 minutes at room temperature.

Afterwards, cells were mixed with the DNA/PEI solution, transferred to a 500 mL shake flask and incubated for 3 hours at 37°C in an incubator with a 5% CO₂ atmosphere. After the

incubation, 160 mL of Excell medium supplemented with 6mM L-Glutamine, 5g/L PEPSOY

5 and 1.2mM valproic acid was added and cells were cultured for 24 hours. One day after transfection 12% Feed (amino acid and glucose) were added. After culturing for 7 days, the supernatant was collected by centrifugation for 30-40 minutes at least 400 x g. The solution was sterile filtered (0.22 µm filter), supplemented with sodium azide to a final concentration of 0.01 % (w/v), and kept at 4°C.

10 The split trimeric OX40 ligand Fc (kih) fusion antigen binding molecule, as well as the IgG, was purified from cell culture supernatants by affinity chromatography using Protein A, followed by size exclusion chromatography. For affinity chromatography, the supernatant was loaded on a MabSelect Sure column (CV = 5-15 mL, resin from GE Healthcare) equilibrated with Sodium Phosphate (20 mM), Sodium Citrate (20 mM) buffer (pH 7.5). Unbound protein
15 was removed by washing with at least 6 column volumes of the same buffer. The bound protein was eluted using either a linear gradient (20 CV) or a step elution (8 CV) with 20 mM sodium citrate, 100 mM Sodium chloride, 100 mM Glycine buffer (pH 3.0). For the linear gradient an additional 4 column volumes step elution was applied.

The pH of collected fractions was adjusted by adding 1/10 (v/v) of 0.5M sodium phosphate,
20 pH8.0. The protein was concentrated prior to loading on a HiLoad Superdex 200 column (GE Healthcare) equilibrated with 20 mM Histidine, 140 mM sodium chloride, 0.01% (v/v) Tween20 solution of pH 6.0.

The protein concentration was determined by measuring the optical density (OD) at 280 nm, using a molar extinction coefficient calculated on the basis of the amino acid sequence.

25 Purity and molecular weight of the targeted trimeric 4-1BB ligand Fc (kih) fusion was analyzed by SDS-PAGE in the presence and absence of a reducing agent (5 mM 1,4-dithiotreitol) and staining with Coomassie SimpleBlue™ SafeStain (Invitrogen USA) or CE-SDS using Caliper LabChip GXII (Perkin Elmer). The aggregate content of samples was analyzed using a TSKgel G3000 SW XL analytical size-exclusion column (Tosoh) equilibrated in 25 mM K₂HPO₄, 125
30 mM NaCl, 200 mM L-Arginine Monohydrochloride, 0.02 % (w/v) NaN₃, pH 6.7 running buffer at 25°C.

Table 98 summarizes the yield and final monomer content of the FAP targeted split trimeric Ox40 ligand Fc (kih) fusion antigen binding molecule, and of the germline DP47 human IgG1 PGLALA (control F).

Table 98: Biochemical analysis of CEA targeted split trimeric 4-1BB ligand Fc (kih) fusion.

Construct	Monomer [%] (SEC)	Yield [mg/l]
monovalent FAP(28H1) targeted split trimeric Ox40 ligand Fc fusion containing CH-CL cross with charged residues (construct 6.1)	93.8	19.7
germline DP47 human IgG1 PGLALA	100	50

Example 15**Functional characterization of the targeted OX40 ligand trimer-containing Fc fusion antigen binding molecule****15.1 Binding to human FAP-expressing tumor cells**

The binding to cell surface FAP was tested using WM-266-4 cells (ATCC CRL-1676). 0.5 x 10⁵ WM-266-4 cells were added to each well of a round-bottom suspension cell 96-well plates (greiner bio-one, cellstar, Cat. No. 650185). Cells were stained for 120 minutes at 4°C in the dark in 50 µL/well 4 °C cold FACS buffer (DPBS (Gibco by Life Technologies, Cat. No. 14190 326) w/ BSA (0.1 % v/w, Sigma-Aldrich, Cat. No. A9418) containing titrated anti –Ox40 antibody construct. After three times washing with excess FACS buffer, cells were stained for 45 minutes at 4°C in the dark in 25 µL/well 4 °C cold FACS buffer containing Fluorescein isothiocyanate (FITC)-conjugated AffiniPure anti-human IgG Fcγ-fragment-specific goat IgG F(ab')₂ fragment (Jackson ImmunoResearch, Cat. No. 109 096 098).

Plates were finally resuspended in 90 µL/well FACS-buffer containing 0.2 µg/mL DAPI (Santa Cruz Biotech, Cat. No. Sc-3598) and acquired the same day using 5-laser LSR-Fortessa (BD Bioscience with DIVA software).

As shown in **Figure 47A**, the monovalent FAP(28H1) targeted split trimeric Ox40 ligand Fc (kih) fusion antigen binding molecule (FAP-OX40L) but not the negative control F efficiently bound to human FAP-expressing target cells. EC₅₀ values of binding to FAP positive WM-266-4 was [6.9 nM].

15.2 Binding to OX40 and FAP negative tumor cells

The lack of binding to OX40 negative FAP negative tumor cells was tested using A549 NucLight™ Red Cells (Essenbioscience, Cat. No. 4491) expressing the NucLight Red fluorescent protein restricted to the nucleus to allow separation from unlabeled human FAP positive WM266-4 cells. Parental A549 (ATCC CCL-185) were transduced with the Essen

CellPlayer NucLight Red Lentivirus (Essenbioscience, Cat. No. 4476; EF1 α , puromycin) at an MOI of 3 (TU/cell) in the presence of 8 μ g/ml polybrene following the standard Essen protocol.

A mixture of 5 x 10⁴ unlabeled WM266-4 cells and unlabeled A549 NucLight™ Red Cells in FACS buffer were added to each well of a round-bottom suspension cell 96-well plates and
5 binding assay was performed as described in section 15.1.

As shown in **Figure 47B**, FAP-OX40L did not bind to OX40 negative FAP negative human tumor cells.

15.3 Binding to human OX40 expressing cells: naïve and activated human peripheral mononuclear blood leukocytes (PBMCs)

10 Buffy coats were obtained from the Zürich blood donation center. To isolate fresh peripheral blood mononuclear cells (PBMCs) the buffy coat was diluted with the same volume of DPBS (Gibco by Life Technologies, Cat. No. 14190 326). 50 mL polypropylene centrifuge tubes (TPP, Cat.-No. 91050) were supplied with 15 mL Histopaque 1077 (SIGMA Life Science, Cat.-No. 10771, polysucrose and sodium diatrizoate, adjusted to a density of 1.077 g/mL) and
15 the buffy coat solution was layered above the Histopaque 1077. The tubes were centrifuged for 30 min at 400 x g, room temperature and with low acceleration and no break. Afterwards the PBMCs were collected from the interface, washed three times with DPBS and resuspended in T cell medium consisting of RPMI 1640 medium (Gibco by Life Technology, Cat. No. 42401-042) supplied with 10 % Fetal Bovine Serum (FBS, Gibco by Life Technology, Cat. No. 16000-044, Lot 941273, gamma-irradiated, mycoplasma-free and heat inactivated at 56 °C for 35 min), 1 %
20 (v/v) GlutaMAX I (GIBCO by Life Technologies, Cat. No. 35050 038), 1 mM Sodium-Pyruvate (SIGMA, Cat. No. S8636), 1 % (v/v) MEM non-essential amino acids (SIGMA, Cat.-No. M7145) and 50 μ M β -Mercaptoethanol (SIGMA, M3148).

PBMCs were used directly after isolation (binding on resting human PBMCs) or they were
25 stimulated to receive a strong human Ox40 expression on the cell surface of T cells (binding on activated human PBMCs). Therefore naïve PBMCs were cultured for four days in T cell medium supplied with 200 U/mL Proleukin (Novartis) and 2 μ g/mL PHA-L (Sigma-Aldrich, L2769-10) in 6-well tissue culture plate and then over night on pre-coated 6-well tissue culture plates [4 μ g/mL] anti-human CD3 (clone OKT3, eBioscience, Ca.No. 16-0037-85) and [2 μ g/mL] anti-
30 human CD28 (clone CD28.2, eBioscience, Cat No.16-0289-85] in T cell medium supplied with 200 U/mL Proleukin at 37 °C and 5% CO₂.

For detection of Ox40 naïve human PBMC and activated human PBMC were mixed. To enable distinction of naïve from activated human PBMC naïve cells were labeled prior to the binding assay using the eFluor670 cell proliferation dye (eBioscience, Cat.-No.65-0840-85).

For labeling cells were harvested, washed with pre-warmed (37°C) DPBS and adjusted to a cell density of 1×10^7 cells/mL in DPBS. eFluor670 cell proliferation dye (eBioscience, Cat.-No.65-0840-85) was added to the suspension of naïve human PBMC at a final concentration of 2.5 mM and a final cell density of 0.5×10^7 cells/mL in DPBS. Cells were then incubated for 10 min at room temperature in the dark. To stop labeling reaction 4 mL heat inactivated FBS were added and cells were washed three times with T cell medium. A two to one mixture of 1×10^5 resting eFluor670 labeled human PBMC and 0.5×10^5 unlabeled activated human PBMC were then added to each well of a round-bottom suspension cell 96-well plates (greiner bio-one, cellstar, Cat. No. 650185).

Cells were stained for 120 minutes at 4°C in the dark in 50 µL/well 4 °C cold FACS buffer containing titrated anti –Ox40 antibody constructs. After three times washing with excess FACS buffer, cells were stained for 45 minutes at 4°C in the dark in 25 µL/well 4 °C cold FACS buffer containing a mixture of fluorescently labeled anti-human CD4 (clone RPA-T4, mouse IgG1 k, BioLegend, Cat.-No. 300532), anti-human CD8 (clone RPa-T8, mouse IgG1k, BioLegend, Cat.-No. 3010441) and Fluorescein isothiocyanate (FITC)-conjugated AffiniPure anti-human IgG Fcγ-fragment-specific goat IgG F(ab')₂ fragment (Jackson ImmunoResearch, Cat.-No. 109-096-098).

Plates were finally resuspended in 90 µL/well FACS-buffer containing 0.2 µg/mL DAPI (Santa Cruz Biotech, Cat. No. Sc-3598) and acquired the same day using 5-laser LSR-Fortessa (BD Bioscience with DIVA software).

As shown in **Figures 48A and 48 B**, FAP-OX40L did not bind to resting human CD4+ T-cells or CD8+ T-cells, which are negative for OX40. In contrast, FAP-OX40L bound to activated CD8+ or CD4+ T-cells, which do express OX40. Binding to CD4+ T-cells was much stronger than that to CD8+ T cells. Activated human CD8+ T cells do express only a fraction of the OX40 levels detected on activated CD4+ T cells. Expression levels for OX40 are depending on kinetic and strength of stimulation and conditions were here optimized for OX40 expression on CD4+ T cells but not for CD8+ T cells. Thus, only little OX40 expression was induced on CD8 T cells. The EC 50 value of binding to OX40 positive CD4+ or CD8+ T cells was [0.15 nM].

15.4 NFκB activation in HeLa cells expressing human OX40 and reporter gene NFκB-luciferase

Agonistic binding of Ox40 to its ligand induces downstream signaling via activation of nuclear factor kappa B (NFκB) (A. D. Weinberg et al., J. Leukoc. Biol. 2004, 75(6), 962-972). The recombinant reporter cell line HeLa_hOx40_NFκB_Luc1 was generated to express human Ox40 on its surface. Additionally, it harbors a reporter plasmid containing the luciferase gene under the control of an NFκB-sensitive enhancer segment. Ox40 triggering induces dose-

dependent activation of NFκB, which translocates in the nucleus, where it binds on the NFκB sensitive enhancer of the reporter plasmid to increase expression of the luciferase protein. Luciferase catalyzes luciferin-oxidation resulting in oxyluciferin which emits light. This can be quantified by a luminometer. Thus, the capacity of the various anti-Ox40 molecules to induce NFκB activation in HeLa_hOx40_NFκB_Luc1 reporter cells was analyzed as a measure for bioactivity.

Adherent HeLa_hOx40_NFκB_Luc1 cells were harvested using cell dissociation buffer (Invitrogen, Cat.-No. 13151-014) for 10 minutes at 37 °C. Cells were washed once with DPBS and were adjusted to a cell density of 1.33×10^5 in assay media comprising of MEM (Invitrogen, Cat.-No. 22561-021), 10 % (v/v) heat-inactivated FBS, 1 mM Sodium-Pyruvate and 1% (v/v) non-essential amino acids. Cells were seeded in a density of 0.2×10^5 cells per well in a sterile white 96-well flat bottom tissue culture plate with lid (greiner bio-one, Cat. No. 655083) and kept over night at 37 °C and 5% CO₂ in an incubator (Hera Cell 150).

The next day, HeLa_hOx40_NFκB_Luc1 were stimulated for 5 hours by adding assay medium containing titrated FAP-Ox40L or negative control F. For testing the effect of hyper-crosslinking on anti-Ox40 antibodies, 25 μL/well of medium containing secondary antibody anti-human IgG Fcγ-fragment-specific goat IgG F(ab')₂ fragment (Jackson ImmunoResearch, 109-006-098) were added in a 1:2 ratio (2 times more secondary antibody than the primary antibody). After incubation, supernatant was aspirated and plates washed two times with DPBS. Quantification of light emission was done using the luciferase 100 assay system and the reporter lysis buffer (both Promega, Cat.-No. E4550 and Cat-No: E3971) according to manufacturer instructions. Briefly, cells were lysed for 10 minutes at -20 °C by addition of 30 uL per well 1x lysis buffer. Cells were thawed for 20 minutes at 37 °C before 90 uL per well provided luciferase assay reagent was added. Light emission was quantified immediately with a SpectraMax M5/M5e microplate reader (Molecular Devices, USA) using 500ms integration time, without any filter to collect all wavelengths. Emitted relative light units (URL) were corrected by basal luminescence of HeLa_hOx40_NFκB_Luc1 cells and were blotted against the logarithmic primary antibody concentration using Prism4 (GraphPad Software, USA). Curves were fitted using the inbuilt sigmoidal dose response.

As shown in **Figure 49**, a limited, dose dependent NFκB activation was induced already by addition of FAP-Ox40L (left side) to the reporter cell line. Hyper-crosslinking of FAP-Ox40L by anti-human IgG specific secondary antibodies increased the induction of NFκB-mediated luciferase-activation in a concentration-dependent manner (right side).

Consequently, we tested the NFκB activating capacity of FAP-Ox40L with hyper-crosslinking of the constructs by FAP+ tumor cell lines.

Tested tumor cell line was NIH/3T3-huFAP clone 39. NIH/3T3-huFAP clone 39 was generated by the transfection of the mouse embryonic fibroblast NIH/3T3 cell line (ATCC CRL-1658) with the expression vector pETR4921 to express huFAP under 1.5 µg/mL Puromycin selection. The surface expression of FAP was quantified using the Quifikit (Dako Cat. No. K0078) according to manufactures instructions. The primary antibody used to detect cell surface FAP expression was the human/ mouse crossreactive clone F11-24 (mouse IgG1, Calbiochem, Ca. No. OP188). The surface expression on NIH/3T3-huFAP clone 39 was app. 90000 huFAP per cell.

As described herein before, adherent HeLa_hOx40_NFkB_Luc1 cells were cultured over night at a cell density of 0.2×10^5 cells per well and were stimulated for 5 hours with assay medium containing titrated FAP-Ox40L. To test the effect of hyper-crosslinking by cell surface FAP binding 25 µL/well of medium containing FAP+ tumor cells NIH/3T3-huFAP clone 39 were co-cultured in a 3 to 1 ratio (three times as much FAP+ tumor cells than reporter cells per well). Activated NFkB was quantified by measuring light emission using luciferase 100 assay system and the reporter lysis buffer (both Promega, Cat.-No. E4550 and Cat-No: E3971).

As shown in **Figure 50A**, the presence of FAP-expressing tumor cells strongly increased induction of NFkB-mediated luciferase-activation when FAP-Ox40L was added. Area under the curve of the respective blotted dose-response curves was quantified as a marker for the agonistic capacity of each construct. As shown in **Figure 50A**, the presence of cell surface presented FAP ensured higher cross-linking and thus a better agonistic effect of FAP-Ox40L then addition of an Fc specific secondary antibody.

15.5 OX40 mediated costimulation of suboptimally TCR triggered resting human PBMC and hypercrosslinking by cell surface FAP

It was shown in Example 15.4 that addition of FAP+ tumor cells can strongly increase the NFkB activity induced by FAP targeted OX40L in a human Ox40 positive reporter cell lines by providing strong oligomerization of OX40 receptors. Likewise, we tested FAP-OX40L constructs in the presence of NIH/3T3-huFAP clone 39 cells for their ability to rescue suboptimal TCR stimulation of resting human PBMC cells.

Human PBMC preparations contain (1) resting Ox40 negative CD4+ and CD8+ T cells and (2) antigen presenting cells with various Fc-γ receptor molecules on their cell surface e.g. B cells and monocytes. Anti-human CD3 antibody of human IgG1 isotype can bind with its Fc part to the present Fc-γ receptor molecules and mediate a prolonged TCR activation on resting Ox40 negative CD4+ and CD8+ T cells. These cells then start to express Ox40 within several hours. Functional agonistic compounds against Ox40 can signal via the Ox40 receptor present on activated CD8+ and CD4+ T cells and support TCR-mediated stimulation.

Resting CFSE-labeled human PBMC were stimulated for five days with a suboptimal concentration of anti-CD3 antibody in the presence of irradiated FAP+ NIH/3T3-huFAP clone 39 cells and titrated FAP-Ox40L. Effects on T-cell survival and proliferation were analyzed through monitoring of total cell counts and CFSE dilution in living cells by flow cytometry.

Mouse embryonic fibroblast NIH/3T3-huFAP clone 39 cells (see Example 15.4) were harvested using cell dissociation buffer (Invitrogen, Cat.-No. 13151-014) for 10 minutes at 37 °C. Cells were washed once with DPBS. NIH/3T3-huFAP clone 39 cells were cultured at a density of 0.2×10^5 cells per well in T cell media in a sterile 96-well round bottom adhesion tissue culture plate (TPP, Cat. No. 92097) over night at 37 °C and 5% CO₂ in an incubator (Hera Cell 150). The next day they were irradiated in an xRay irradiator using a dose of 4500 RAD to prevent later overgrowth of human PBMC by the tumor cell line.

Human PBMCs were isolated by ficoll density centrifugation as described in Example 15.3. Cells were then labeled with CFSE at a cell density of 1×10^6 cells/ mL with CFDA-SE (Sigma-Aldrich, Cat.-No. 2188) at a final concentration of [50 nM] for 10 minutes at 37 °C. Thereafter, cells were washed twice with excess DPBS containing FBS (10% v/v). Labeled cells were rested in T-cell media at 37 °C for 30 minutes. Thereafter, non-converted CFDA-SE was removed by two additional washing steps with DPBS. CFSE labeled resting human PBMC were added to each well at a density of 0.5×10^5 cells per well. Anti-human CD3 antibody (clone V9, human IgG1, described in Rodrigues et al., Int J Cancer Suppl 7, 45-50 (1992) and US patent No. 6,054,297) at a final concentration of [20 nM] and FAP-OX40L were added at the indicated concentrations. Cells were activated for five days at 37 °C and 5% CO₂ in an incubator (Hera Cell 150). Then, Cells were surface-stained with fluorescent dye-conjugated antibodies anti-human CD4 (clone RPA-T4, BioLegend, Cat.-No. 300532) and CD8 (clone RPa-T8, BioLegend, Cat.-No. 3010441) for 20 min at 4°C. After a washing step with FACS buffer, cells were resuspended in 85 µL/well FACS buffer and acquired using a 5-laser Fortessa flow cytometer (BD Bioscience with DIVA software).

As shown in **Figure 51**, hyper-crosslinking of FAP-OX40L constructs by the present NIH/3T3-huFAP clone 39 cells strongly promoted proliferation (see “Events” graphs on the top) and survival (see “proliferation” graphs on the bottom) in TCR stimulated human CD4 and CD8 T cells. In line with a lower expression of OX40 on human CD8+ T cells the agonistic effect of FAP-OX40L was less strong on CD8+ T cells than on CD4+ T cells.

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Claims

1. A TNF family ligand trimer-containing antigen binding molecule comprising
 - (a) at least one moiety capable of specific binding to a target cell antigen and
 - 5 (b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or a fragment thereof that are connected to each other by a peptide linker and in that the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof.
- 10 2. The TNF family ligand trimer-containing antigen binding molecule of claim 1, further comprising
 - (c) an Fc domain composed of a first and a second subunit capable of stable association.
3. The TNF family ligand trimer-containing antigen binding molecule of claims 1 or 2, comprising
 - 15 (a) at least one moiety capable of specific binding to a target cell antigen and
 - (b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that
 - (i) the first polypeptide contains a CH1 or CL domain and the second polypeptide contains a CL or CH1 domain, respectively, wherein the second polypeptide is linked to the first polypeptide by a disulfide bond between the CH1 and CL domain, and wherein the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the CH1 or CL domain by a peptide linker and wherein the second polypeptide comprises one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to the CL or
 - 20 CH1 domain of said polypeptide, or
 - (ii) the first polypeptide contains a CH3 domain and the second polypeptide contains a CH3 domain, respectively, and wherein the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the C-terminus of the CH3 domain by a peptide linker and wherein the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to C-terminus of the CH3 domain of said polypeptide, or
 - 25 (iii) the first polypeptide contains a VH-CL or a VL-CH1 domain and the second polypeptide contains a VL-CH1 domain or a VH-CL domain, respectively, wherein the second polypeptide is linked to the first polypeptide by a disulfide bond between the
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CH1 and CL domain, and wherein the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to to VH or VL by a peptide linker and wherein the second polypeptide comprises one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to VL or VH of said polypeptide.

4. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 3, wherein the TNF ligand family member costimulates human T-cell activation.
5. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 4, wherein the TNF ligand family member is selected from 4-1BBL and OX40L.
6. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 5, wherein the TNF ligand family member is 4-1BBL.
7. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 6, wherein the ectodomain of a TNF ligand family member comprises the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO: 2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:96, SEQ ID NO: 373, SEQ ID NO:374 and SEQ ID NO:375, particularly the amino acid sequence of SEQ ID NO:1 or SEQ ID NO:96.
8. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 7, wherein the ectodomain of a TNF ligand family member comprises the amino acid sequence of SEQ ID NO:96.
9. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 8, comprising
 - (a) at least one moiety capable of specific binding to a target cell antigen and
 - (b) a first and a second polypeptide that are linked to each other by a disulfide bond,wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:97, SEQ ID NO:98 and SEQ ID NO:99 and in that the second polypeptide comprises the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:96, SEQ ID NO:3 and SEQ ID NO:4.
10. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 9, comprising
 - (a) at least one moiety capable of specific binding to a target cell antigen, and

- (b) a first polypeptide containing a CH1 or CL domain and a second polypeptide containing a CL or CH1 domain, respectively, wherein the second polypeptide is linked to the first polypeptide by a disulfide bond between the CH1 and CL domain, and wherein the antigen binding molecule is characterized in that the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the CH1 or CL domain by a peptide linker and in that the second polypeptide comprises only one ectodomain of said TNF ligand family member or a fragment thereof connected via a peptide linker to the CL or CH1 domain of said polypeptide.
- 10 11. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 10, wherein the moiety capable of specific binding to a target cell antigen is selected from the group consisting of an antibody fragment, a Fab molecule, a crossover Fab molecule, a single chain Fab molecule, a Fv molecule, a scFv molecule, a single domain antibody, an aVH and a scaffold antigen binding protein.
- 15 12. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 11, wherein the molecule comprises one moiety capable of specific binding to a target cell antigen.
- 20 13. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 12, wherein the moiety capable of specific binding to a target cell antigen is a Fab molecule capable of specific binding to a target cell antigen.
- 25 14. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 13, wherein the target cell antigen is selected from the group consisting of Fibroblast Activation Protein (FAP), Melanoma-associated Chondroitin Sulfate Proteoglycan (MCSP), Epidermal Growth Factor Receptor (EGFR), Carcinoembryonic Antigen (CEA), CD19, CD20 and CD33.
15. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, wherein the target cell antigen is Fibroblast Activation Protein (FAP).
- 30 16. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 15, wherein the moiety capable of specific binding to FAP comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:7 or SEQ ID NO:100, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:8 or SEQ ID NO:101, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:9 or SEQ ID

NO:102, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:10 or SEQ ID NO:103, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:11 or SEQ ID NO:104, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:12 or SEQ ID NO:105.

- 5 17. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 16, wherein the moiety capable of specific binding to FAP comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:16 and a variable light chain comprising an amino acid sequence of SEQ ID NO:17 or wherein the moiety capable of specific binding to FAP comprises a variable heavy chain comprising an amino acid sequence of SEQ ID
10 NO:106 and a variable light chain comprising an amino acid sequence of SEQ ID NO:107.
18. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 2 to 17, wherein the Fc domain is an IgG, particularly an IgG1 Fc domain or an IgG4 Fc domain.
19. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 2 to 18, wherein the Fc domain is an IgG1 Fc domain comprising the amino acid substitutions at
15 positions 234 and 235 (EU numbering) and/or 329 (EU numbering).
20. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 19, wherein the antigen binding molecule comprises
a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific binding to a target cell antigen,
20 a first peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker fused at its C-terminus by a second peptide linker to a second heavy or light chain,
and a second peptide comprising one ectodomain of said TNF ligand family member fused at its C-terminus by a third peptide linker to a second light or heavy chain, respectively.
- 25 21. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 20, wherein the first peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by a second peptide linker to a CH1 domain that is part of a heavy chain,
and the second peptide comprising one ectodomain of said TNF ligand family member or a
30 fragment thereof is fused at its C-terminus by a third peptide linker to a CL domain that is part of a light chain.

22. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 20, wherein the first peptide comprising two ectodomains of a TNF ligand family member or fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by a second peptide linker to a CL domain that is part of a heavy chain,
5 and the second peptide comprising one ectodomain of said TNF ligand family member or a fragment thereof is fused at its C-terminus by a third peptide linker to a CH1 domain that is part of a light chain.
23. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 20, wherein the first peptide comprising two ectodomains of a TNF ligand family member or
10 fragments thereof connected to each other by a first peptide linker is fused at its C-terminus by a second peptide linker to a VH domain that is part of a heavy chain,
and the second peptide comprising one ectodomain of said TNF ligand family member or a fragment thereof is fused at its C-terminus by a third peptide linker to a VL domain that is part of a light chain.
- 15 24. The TNF family ligand trimer-containing antigen binding molecule of claims 21 to 23, wherein in the CL domain adjacent to the TNF ligand family member the amino acid at position 123 (EU numbering) has been replaced by arginine (R) and the amino acid at position 124 (EU numbering) has been substituted by lysine (K), and wherein in the CH1 domain adjacent to the TNF ligand family member the amino acids at position 147 (EU
20 numbering) and at position 213 (EU numbering) have been substituted by glutamic acid (E).
25. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 24, wherein the antigen binding molecule comprises
(a) a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific binding to a target cell antigen,
25 (b) a second heavy chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:97, SEQ ID NO:98 and SEQ ID NO:99, and a second light chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:96, SEQ ID NO:3 and SEQ ID NO:4.
26. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 24, wherein the antigen binding molecule comprises
30 (a) a first heavy chain and a first light chain, both comprising a Fab molecule capable of specific binding to a target cell antigen,
(b) a second heavy chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:97, SEQ ID NO:98 and SEQ ID NO:99, and

a second light chain comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:96, SEQ ID NO:3 and SEQ ID NO:4.

27. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 21, wherein the antigen binding molecule comprises

(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:16 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:17 or

a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:106 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:107,

(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:14, SEQ ID NO:108, SEQ ID NO:111 and SEQ ID NO:113, and

(iii) a second light chain comprising the amino acid sequence of SEQ ID NO:15, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:112 and SEQ ID NO:114.

28. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 20 and 22, wherein the antigen binding molecule comprises

(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:16 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:17 or

a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:106 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:107,

(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:115, SEQ ID NO:117, SEQ ID NO:119 and SEQ ID NO:173, and

(iii) a second light chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:116, SEQ ID NO:118, SEQ ID NO:120 and SEQ ID NO:174.

29. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 19, comprising

(a) at least one moiety capable of specific binding to a target cell antigen, and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide contains a CH3 domain and the second polypeptide contains a CH3 domain, respectively, and wherein the first polypeptide comprises two ectodomains of a TNF ligand family member or fragments thereof that are connected to each other and to the C-terminus of the CH3 domain by a peptide linker and wherein the second polypeptide comprises one ectodomain of said

TNF ligand family member or a fragment thereof connected via a peptide linker to C-terminus of the CH3 domain of said polypeptide.

30. The TNF family ligand trimer-containing antigen binding molecule of claim 29, comprising two moieties capable of specific binding to a target cell antigen.
- 5 31. The TNF family ligand trimer-containing antigen binding molecule of claim 29, wherein the antigen binding molecule comprises
- (i) a first heavy chain comprising the amino acid sequence of SEQ ID NO:121, a second heavy chain comprising the amino acid sequence of SEQ ID NO:122, and two light chains comprising the amino acid sequence of SEQ ID NO:19, or
- 10 (ii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:123, a second heavy chain comprising the amino acid sequence of SEQ ID NO:124, and two light chains comprising the amino acid sequence of SEQ ID NO:125, or
- (iii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:126, a second heavy chain comprising the amino acid sequence of SEQ ID NO:127, and two light chains
- 15 comprising the amino acid sequence of SEQ ID NO:125.
32. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 18 to 26 and 29 to 30, wherein the target cell antigen is CD19.
33. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 18 to 26, 29 to 30 and 32, wherein the moiety capable of specific binding to CD19
- 20 comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:195 or SEQ ID NO:252, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:196 or SEQ ID NO:253, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:197 or SEQ ID NO:254, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:198 or SEQ ID NO:249, (v) CDR-L2
- 25 comprising the amino acid sequence of SEQ ID NO:199 or SEQ ID NO:250, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:200 or SEQ ID NO:251.
34. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 18 to 26, 29 to 30, 32 and 33, wherein the moiety capable of specific binding to CD19 comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:201
- 30 and a variable light chain comprising an amino acid sequence of SEQ ID NO:202 or wherein the moiety capable of specific binding to FAP comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:357 and a variable light chain comprising an amino acid sequence of SEQ ID NO:358.

35. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 18 to 26 and 32 to 34, wherein the antigen binding molecule comprises
- (i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:201 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:202 or
- 5 a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:357 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:358,
- (ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:14, SEQ ID NO:108, SEQ ID NO:111 and SEQ ID NO:113, and
- 10 (iii) a second light chain comprising the amino acid sequence of SEQ ID NO:15, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:112 and SEQ ID NO:114.
36. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 18 to 26 and 32 to 35, wherein the antigen binding molecule comprises
- 15 (i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:201 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:202 or
- a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:357 and a first light chain comprising the VL domain comprising the amino acid
- 20 sequence of SEQ ID NO:358,
- (ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:115, SEQ ID NO:117, SEQ ID NO:119 and SEQ ID NO:173, and
- (iii) a second light chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:116, SEQ ID NO:118, SEQ ID NO:120 and SEQ ID NO:174.
- 25 37. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 29, 30 and 32 to 34, wherein the antigen binding molecule comprises
- (i) a first heavy chain comprising the amino acid sequence of SEQ ID NO:209, a second heavy chain comprising the amino acid sequence of SEQ ID NO:210, and two light chains comprising the amino acid sequence of SEQ ID NO:206, or
- 30 (ii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:213, a second heavy chain comprising the amino acid sequence of SEQ ID NO:214, and two light chains comprising the amino acid sequence of SEQ ID NO:206, or
- (iii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:309, a second heavy chain comprising the amino acid sequence of SEQ ID NO:310, and two light chains
- 35 comprising the amino acid sequence of SEQ ID NO:279, or
- (iv) a first heavy chain comprising the amino acid sequence of SEQ ID NO:313, a second

heavy chain comprising the amino acid sequence of SEQ ID NO:314, and two light chains comprising the amino acid sequence of SEQ ID NO:279.

38. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 18 to 26 and 29 to 30, wherein the target cell antigen is CEA.
- 5 39. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 18 to 26, 29 to 30 and 38, wherein the moiety capable of specific binding to CEA comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:321, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:322, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:323, and a VL domain
10 comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:324, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:325, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:326.
40. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 18 to 26, 29 to 30, 38 and 39, wherein the moiety capable of specific binding to CEA
15 comprises a variable heavy chain comprising an amino acid sequence of SEQ ID NO:329 and a variable light chain comprising an amino acid sequence of SEQ ID NO:330.
41. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 18 to 26 and 38 to 40, wherein the antigen binding molecule comprises
20 (i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:329 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:330,
(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:14, SEQ ID NO:108, SEQ ID NO:111 and SEQ ID NO:113, and
(iii) a second light chain comprising the amino acid sequence of SEQ ID NO:15, SEQ ID
25 NO:109, SEQ ID NO:110, SEQ ID NO:112 and SEQ ID NO:114.
42. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 18 to 26 and 38 to 41, wherein the antigen binding molecule comprises
30 (i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:329 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:330,
(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:115, SEQ ID NO:117, SEQ ID NO:119 and SEQ ID NO:173, and

(iii) a second light chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:116, SEQ ID NO:118, SEQ ID NO:120 and SEQ ID NO:174.

43. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 14, 29, 30 and 38 to 40, wherein the antigen binding molecule comprises

(i) a first heavy chain comprising the amino acid sequence of SEQ ID NO:337, a second heavy chain comprising the amino acid sequence of SEQ ID NO:338, and two light chains comprising the amino acid sequence of SEQ ID NO:334, or

(ii) a first heavy chain comprising the amino acid sequence of SEQ ID NO:341, a second heavy chain comprising the amino acid sequence of SEQ ID NO:342, and two light chains comprising the amino acid sequence of SEQ ID NO:334.

44. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 5, 10 to 24, 29, 30, 32 to 34 and 38 to 40, wherein the TNF ligand family member is OX40L.

45. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 5, 10 to 24, 29, 30, 32 to 34, 38 to 40 and 44, wherein the ectodomain of a TNF ligand family member comprises the amino acid sequence of SEQ ID NO:53 or SEQ ID NO:54, particularly the amino acid sequence of SEQ ID NO:53.

46. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 5, 10 to 24, 29, 30, 32 to 34, 38 to 40, 44 and 45, comprising

(a) at least one moiety capable of specific binding to a target cell antigen and

(b) a first and a second polypeptide that are linked to each other by a disulfide bond, wherein the antigen binding molecule is characterized in that the first polypeptide comprises the amino acid sequence of SEQ ID NO:371 or SEQ ID:372 and in that the second polypeptide comprises the amino acid sequence of SEQ ID NO:53 or SEQ ID NO:54.

47. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 5, 10 to 24, 29, 30 and 44 to 46, wherein the target cell antigen is Fibroblast Activation Protein (FAP) and the moiety capable of specific binding to FAP comprises a VH domain comprising (i) CDR-H1 comprising the amino acid sequence of SEQ ID NO:7 or SEQ ID NO:100, (ii) CDR-H2 comprising the amino acid sequence of SEQ ID NO:8 or SEQ ID NO:101, and (iii) CDR-H3 comprising the amino acid sequence of SEQ ID NO:9 or SEQ ID NO:102, and a VL domain comprising (iv) CDR-L1 comprising the amino acid sequence of SEQ ID NO:10 or SEQ ID NO:103, (v) CDR-L2 comprising the amino acid sequence of SEQ ID NO:11 or SEQ ID NO:104, and (vi) CDR-L3 comprising the amino acid sequence of SEQ ID NO:12 or SEQ ID NO:105.

48. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 5, 10 to 24, 29, 30 and 44 to 47, wherein the antigen binding molecule comprises
(i) a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:16 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:17 or
5 a first heavy chain comprising the VH domain comprising the amino acid sequence of SEQ ID NO:106 and a first light chain comprising the VL domain comprising the amino acid sequence of SEQ ID NO:107,
(ii) a second heavy chain comprising the amino acid sequence selected from the group consisting of SEQ ID NO:355, and
10 (iii) a second light chain comprising the amino acid sequence of SEQ ID NO:356.
49. An isolated polynucleotide encoding the TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 48.
50. A vector, particularly an expression vector, comprising the isolated polynucleotide of claim 15 49.
51. A host cell comprising the isolated polynucleotide of claim 49 or the vector of claim 50.
52. A method for producing the TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 48, comprising the steps of
(i) culturing the host cell of claim 51 under conditions suitable for expression of the antigen binding molecule, and
20 (ii) recovering the antigen binding molecule.
53. A pharmaceutical composition comprising the TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 48 and at least one pharmaceutically acceptable excipient.
- 25 54. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 48, or the pharmaceutical composition of claim 53, for use as a medicament.
55. The TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 48, or the pharmaceutical composition of claim 53, for use in the treatment of cancer.
- 30 56. Use of the TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 48 for the manufacture of a medicament for the treatment of cancer.

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57. A method of treating a disease in an individual, comprising administering to said individual a therapeutically effective amount of a composition comprising a TNF family ligand trimer-containing antigen binding molecule of any one of claims 1 to 48 in a pharmaceutically acceptable form.

5 58. The method of claim 57, wherein said disease is cancer.
