ENHANCED PRODUCTION OF RECOMBINANT PROTEINS BY TRANSIENT TRANSFECTION OF SUSPENSION-GROWING MAMMALIAN CELLS

FIELD OF THE INVENTION

The invention relates to processes for producing recombinant proteins, in particular to a new process for an enhanced transient expression of a recombinant protein in host mammalian cells, and to new expression vectors, cell lines and culture media adapted to carrying out the process.

BACKGROUND OF THE INVENTION

Mammalian cells are an established expression system in the biotechnology industry for the production of recombinant proteins (r-proteins). In contrast to lower eukaryotes or prokaryotes, mammalian cells provide active r-proteins that possess relevant post-translational modifications. However, in order to obtain sufficient amount of protein for structure/activity analyses or high-throughput screenings, one needs to go through the long and tedious process of stable transfectoma isolation and characterization. As an alternative, the small-scale transient transfection of mammalian cells grown in monolayers can generate significant amount of r-proteins (Cullen B. R., *Methods Enzymol.*, **152**, 684 – 704 (1987); Blasey H. D. et al., *Cytotechnology*, **18**, 183 – 192 (1996); Cachianes G. et al., *Biotechniques*, **15**, 255 – 259 (1993)), but scalability of this process is limited by culture surface availability. The use of the well-established calcium phosphate precipitation technique or the recently described cationic polymer polyethylenimine (PEI) (Boussif O. et al., *Proc, Natl. Acad. Sci. USA*, **92**, 7297 – 7301 (1995)) provides cost-effective ways of introducing plasmid DNA into mammalian cells. A major breakthrough has recently emerged for the fast production of milligram amounts of recombinant proteins when these gene transfer vehicles were shown to be effective for large-scale transient transfection of mammalian cells grown in suspension culture (Jordan M. et al., *Cytotechnology*, **26**, 39 – 47 (1998); Schlaeger E-J. et al, *Cytotechnology*, **30**, 71 –

83 (1999); Wurm F. et al., *Curr. Opin. Biotechnol.*, **10**, 156 – 159 (1999)).

For an optimal large-scale transient transfection and r-protein expression in mammalian cells, four key aspects are to be taken into account, namely 1) the cell line, 2) the expression vector, 3) the transfection vehicle and 4) the culture medium. The human 293 cell line (a human embryonic kidney cell line containing the E1 region of human Ad5 adenovirus DNA) is widely used for r-protein production as it offers many advantages, such as high transfection yields with most common gene transfer vehicles, is easily grown in suspension culture, and can be adapted to serum-free media. Moreover, two genetic variants of the 293 cell line, the 293E and 293T cell lines, expressing the Epstein-Barr virus (EBV) Nuclear Antigen 1 (EBNA1) and the SV40 large-T antigen, respectively, allow episomal (extrachromosomal) amplification of plasmids containing the viral EBV (293E) or SV40 (293T) origins of replication. These cell lines are therefore expected to increase r-protein expression levels, by permitting more plasmid copies to persist in the transfected cells throughout the production phase (Van Craenenbroeck H. et al., *Eur. J. Biochem.*, **267**, 5665 – 5678 (2000)).

The second important issue for high level r-protein expression is the use of vectors having promoters that are highly active in the host cell line, such as the human cytomegalovirus (CMV) promoter (Foecking M. K. et al, *Gene*, **45**, 101 – 105 (1985)). This promoter is particularly powerful in 293 cells, where it has been shown to be strongly transactivated by the constitutively expressed adenovirus E1a protein (Gorman C. M. et al., *Virology*, **171**, 377 – 385 (1989)). Moreover, a highly efficient expression cassette using this promoter has been recently described that provides adenovirus-mediated transgene expression levels reaching up to 20% of total cell proteins (TCP) (Massie B. et al., *J. Virol.*, **72**, 2289 – 2296 (1998); Massie B. et al., *Cytotechnology*, **28**, 53 – 64 (1998)).

The third aspect is related to gene transfer reagent efficacy. Even though many highly effective gene transfer reagents are commercially available, only few are cost-effective when considering operations at the multi-liters scale. For large-scale

transient transfection applications, these reagents should also be simple to use, effective with suspension growing cells and have minimal cytotoxic effects. PEI satisfies most of these criteria, as it has high gene transfer activity in many cell lines while displaying low cytotoxicity (Boussif O., supra), is cost-effective, and efficiently transfects suspension growing 293 cells (Schlaeger E-J., supra). This polymer is available as both linear and branched isomers with a wide range of molecular weights and polydispersities, which physicochemical parameters are critical for efficient gene transfer activity (Godbey W. T. et al., *J. Control Release*, **60**, 149 – 160 (1999).

The last key aspect for efficient r-protein expression by transient transfection relates to the culture medium. Some gene transfer reagents work only in serum-free media whereas others are less sensitive to the presence of serum. Also, as the presence of cellular by-products in conditioned medium is associated with poor transfection yield, it is often necessary to perform a complete medium change prior to transfection. However, this step does not satisfy the need for a robust large-scale transient transfection process.

Transient protein expression system are known in the prior art, for example the transient expression system disclosed in US 5,024,939. However, these systems generally suffer from the above-discussed and other drawbacks and limitations and are not well suited to large-scale, high-throughput production of r-proteins. The present invention provides a transient expression system and process which is free of many such prior art limitations.

SUMMARY OF THE INVENTION

The invention provides a new process for the production of recombinant proteins, by transfection of suspension-growing eukaryotic cells with an expression vector comprising a first DNA sequence coding for the desired protein, said first DNA sequence being under control of a suitable promoter, and a second DNA sequence enhancing transcriptional activity of the promoter and increasing nuclear import of the expression vector. In a preferred embodiment, the second DNA sequence

additionally supports an episomal replication of the vector in the transfected cells. The eukaryotic cells are preferably mammalian cells, more preferably the human embryonic kidney 293 cell line and its genetic variants, more preferably genetic variants stably expressing the EBNA1 protein or a fragment thereof. The expression vector is preferably a plasmid, comprising the first DNA sequence as a part of an expression cassette, the cassette further comprising the promoter, preferably a cytomegalovirus (CMV) promoter, most preferably the CMV5 promoter. The second DNA sequence is preferably of a viral origin, more preferably the oriP sequence of Epstein-Barr virus (EBV) or a fragment thereof. The transfection is preferably carried out using polyethylenimine (PEI) as a transfection reagent, more preferably using the 25 kDa linear isoform of PEI. The process combines in a single step the cell growth, transfection and protein expression, is carried out using suspension-growing cells without changing the culture medium, and allows to achieve high expression levels in a short period of time. The process may be carried out in a serum-free culture medium, is easily scalable, compatible with continuous production processes, and fully adapted to high-throughput production of milligram quantities of recombinant proteins.

Thus, according to one aspect, the invention provides a process for the preparation of a recombinant protein, said process comprising the steps of: providing eukaryotic host cells suspension-growing in a culture medium; transfecting the host cells in the presence of a transfection reagent with an expression vector, said vector comprising a first DNA sequence encoding the recombinant protein, said first DNA sequence being under control of a promoter; culturing the transfected cells under conditions favoring expression of the recombinant protein, and harvesting the expressed protein.

According to another aspect, the invention provides an expression vector for an enhanced expression of a recombinant protein in a mammalian cell, said vector comprising a first DNA sequence encoding the recombinant protein, said first DNA sequence being under control of a promoter, said expression vector further comprising a second DNA sequence enhancing the transcriptional activity of the promoter and increasing the nuclear import of the expression vector.

According to still another aspect, the invention provides a human embryonic kidney cell line derived from the 293SF-3F6 cell line (ATCC Accession No. CRL-12585), said line constitutively expressing the EBNA1 protein or a fragment thereof.

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BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 is a graph showing effects of DNA to PEI ratio on transfection efficiency. 293E cells were transfected with linear (A) or branched (B) 25 kDa PEI at various DNA (pGFP plasmid) concentrations, as described in Material and Methods. DNA 10 concentration (μg ml^{-1}) used were: 0.25 (circles), 0.50 (squares), 1.0 (closed diamonds), 1.5 (triangles), and 2.0 (open diamonds). Transfection efficiencies were determined by flow cytometry analysis 72 hpt.

Fig. 2 is a graph showing effects of cell line and vector on transient SEAP 15 expression. (A) shows genetic maps of pCEP5 (left) and pTT (right) vectors drawn to scale. The pCEP5 vector backbone is identical to pCEP4 vector except for the transgene expression cassette. The pTT vector was obtained following removal of the hygromycin and EBNA1 expression cassettes and replacement of the ColE1 origin for the pMB1 ori, as described in Material and Methods. TPL: tripartite leader; 20 enh MLP: major late promoter enhancer; SD: splice donor; SA: splice acceptor; DS: dyad symmetry; FR: family of repeats. (B) Cells were transfected with 1 μg of DNA and 2 μg of linear PEI and SEAP activity measured 72 hpt. The pGFP plasmid (0.1 μg) was also added in each condition to monitor for transfection efficiency and SEAP activities were normalized accordingly. Empty boxes: pcDNA3.1/SEAP; hatched 25 boxes: pCEP4/SEAP; gray boxes: pCEP5/SEAP; black boxes: pTT/SEAP vector.

Fig. 3 is a graph showing effects of serum on transgene expression. 293E cells were transfected with pTT/GFPq (A) or pTT/SEAP (B) vectors using 1.0 μg of DNA and 2.0 μg of linear PEI (hatched boxes) or 1.5 μg and 2.0 μg of branched PEI (gray 30 boxes) in fresh serum-free or serum-supplemented media. In one experiment (0→1%), cells were transfected in serum-free media and serum was added 3 hours later to a final concentration of 1%. GFP-positive cells and SEAP activity were measured 72 hpt.

Fig. 4 is a graph illustrating the progress of transfection of suspension growing cells. Cells were resuspended in 10 ml of fresh HSFM containing 1% BCS to a density of 1 x 10^6 ml^{-1} in 125 ml Erlenmeyer flask. Three hours later, 1 ml of the DNA-PEI complexes were added and the culture incubated for an additional 3 h. The volume was then completed to 20 ml with fresh culture medium. The DNA-PEI complexes were as follows: 40 µg of linear or branched PEI was added to 1 ml of HEPES-supplemented HSFM containing 18 µg of pTT/SEAP and 2 µg of pEGFP or 27 µg of pTT/SEAP and 3 µg of pEGFP, respectively. Open symbols: linear PEI; closed symbols: branched PEI. Circles: SEAP activity; squares: % GFP-positive cells.

Fig. 5 is a graph showing effects of cell density and of conditioned medium. (A) Transfection efficiency and relative total GFP expression (in percent) obtained following transfection using standard conditions (hatched bars: 10 ml of cells at 1 x 10^6 ml^{-1} followed by addition of 10 ml of fresh medium 3 h after transfection) or using cells at 5 x 10^5 ml^{-1} in 20 ml of culture medium (gray bars). GFP was monitored 72 hpt. Relative total GFP was obtained following multiplication of percent GFP-positive cells by the mean fluorescence intensity. (B) Cells were seeded in 20 ml of 1% BCS-supplemented HSFM at a density 2.5 x 10^5 ml^{-1} 24 h before transfection. The medium was then left unchanged (conditioned: open circles) or replaced with 20 ml of fresh medium (closed circles). Three hours later, cells were transfected by the addition of 2 ml of DNA-PEI complexes (20 µg of pTT/SEAP and 40 µg of linear PEI).

Fig. 6 is a graph showing the contribution of FR and DS domains on transient gene expression. The pTT(delta DS) vector was obtained by *ApaI* digestion and re-circularisation. The pTT(delta FR) was obtained by *MluI* – *EcoR*I digestion, fill-in and re-circularisation. The pTT(delta oriP) vector was obtained by *ApaI* – *EcoR*I digestion, fill-in and re-circularisation. The cDNAs encoding GFP and SEAP were cloned into these vectors, followed by transfection of 293E cells. SEAP and GFP were measured 3 days post-transfection.

Fig. 7 is a graph showing the effect of various oriP truncations on transient gene expression. The pTTm/GFP vector was obtained by digesting pTT/GFP vector with

EcoRV – *MluI*, followed by fill-in and re-circularisation. In this construct, the oriP still contains the complete FR and DS domains. The pTTn/GFP vector was obtained by digesting pTTm/GFP vector with *BstX*I, followed by re-circularisation. This construct has an FR fragment containing only 9 EBNA1 binding sites (see Fig. 8). The pTTo/GFP vector was obtained by digesting pTT/GFP vector with *BspM*I – *EcoRV*. This construct contains intact FR and DS domains. 293E cells were transfected with these constructs and GFP levels measured 3 days later. Results are expressed relative to pTT/GFP vector.

Fig. 8 shows the sequence of the oriP's Family of Repeats (FR). The FR contains 20 EBNA1 binding sites (EBS) (boxed). Spacers between EBS are shadowed. Doubly underlined regions indicate *BstX*I restriction sites. Nucleotide in bold font indicate mutations in the EBS. Shadowed box indicates *EcoR*I site.

Fig. 9 is a graph showing the effect of the presence of various FR fragments on transient gene expression. The FR vector constructs used are shown in panel A. The pTT(delta oriP)/GFP vector was derived from pTT/GFP vector following *EcoR*I – *Apa*I digestion, T4 DNA polymerase treatment and re-circularisation. The pTT4a/GFP vector contains the *BstX*I FR fragment (containing 10 EBS) cloned in the *EcoR*I site of pTT(delta oriP)/GFP vector. The pTT4b/GFP vector contains the *BstX*I FR fragment cloned in the *Sal*I site of pTT(delta oriP)/GFP vector. The pTT4c/GFP vector contains an FR fragment containing 9 EBS and was derived from pTT(delta oriP)/GFP vector by *BstX*I - *Apa*I digestion, T4 DNA polymerase treatment and re-circularisation. 293E cells were transfected with vectors shown in panel A and with pTT/GFP vector. GFP was analyzed by flow cytometry 72 hours later and values expressed relative to the value obtained with pTT/GFP vector (containing the complete oriP) are shown in panel C.

Fig. 10 is a graph showing the effect of oriP on nuclear import of plasmids and gene expression. 293E cells were transfected with pcDNA3.1 plasmid encoding SEAP with or without various ratio of pTTΔ vector (containing the oriP) or pTT-ΔΔ vector (no oriP). SEAP activity was measured 72 hours later.

Fig. 11 is a graph showing the effect of peptones on 293E-GFP cells growth. Peptones were tested at 1% (w/v) in HSFM/1% serum. Cells (293E cells stably expressing GFP) were seeded at 1000 cells/well in 96-well plates and fluorescence was monitored daily using a fluorescence microplate reader. Increases in fluorescence indicate cell growth. Control is without peptones. MP: meat peptones; CP: casein peptones; TN: tryptone; GP: gelatin peptones; SP: soy peptones; WP: wheat peptones; ME: malt peptones; PP: plant peptones; YE: yeast extract.

Fig. 12 is a graph showing the effect of peptones and their concentration on transient transfection of SFE cells. Peptones were first tested at 1% (w/v) in HSFM. The effect of concentration of selected peptones was then tested at concentrations 0.5%, 1.0% and 1.5%. GFP was monitored 72 hours later by flow cytometry. Cell agglomeration was significant when using meat and casein peptones (data not shown). Control is without peptone or serum addition.

Fig. 13 is a graph illustrating a transient transfection in a 3.5-liters bioreactor. (A) 293E cells were seeded at a density of 2.5×10^5 ml^{-1} in 2.85 l of fresh HSFM supplemented with 1% BCS. Twenty-four hours later, the transfection mixture (6 mg of linear PEI added to 150 ml HSFM containing 2.85 mg pTT/SEAP and 150 µg pEGFP plasmids) was added to the bioreactor (solid lines). One hour later, 25 ml of culture was withdrawn from the bioreactor and transferred in a shake flask as a control (dashed lines). SEAP activity (circles) and GFP-positive cells (squares) were determined as described in Materials and Methods. (B) Growth curves (diamonds), viability (triangles) and yO_2 (gray line) in the 3.5-l bioreactor (solid lines) and shaker flask (dashed lines).

Fig. 14 is a photograph showing results of SEAP purification and production of other secreted and intracellular r-proteins. (A) SEAP purification by IMAC. One liter of culture medium from the 3.5-l bioreactor harvest (Fig. 13) was loaded onto a TALON™ IMAC column (10 ml bed volume). Following extensive washing, bound material was eluted with 150 mM imidazole (20 ml). Ten microliters of culture medium (lane 1), flow-through (lane 2) and eluted material (lane 3) were resolved in

duplicate on a 3 – 8% NuPAGE Tris-acetate gradient gel. One half of the gel was directly stained with Coomassie blue R-250 (left panel) whereas the other half was transferred onto a nitrocellulose membrane and probed with anti-Myc antibody (right panel). (B) Expression of secreted C-terminal Myc-$(His)_6$-tagged r-protein in a 14-l bioreactor. Lane 1, human Neurophilin (1 – 824; upper band) and VEGF (1 – 165; lower band) co-transfection in a 1:1 ratio; lane 2, human Tie2 (1 – 723); lane 3, human Cripto (1 – 173); lane 4, human c-Met (1 – 931). Transfections were performed as described in Materials and Methods and culture medium harvested 120 hpt. Fifteen microliters of culture medium were loaded per lane and tagged proteins detected using anti-Myc antibody. (C) Expression of intracellular r-proteins. Lane 1, pTT/sgGFP; lane 2, pTT/RR1; lane 3, pTT empty vector; lane 4, pcDNA3.1/$G_{\alpha q}$; lane 5, pTT/$G_{\alpha q}$; lane 6, pTT/$p27^{Kip1}$; lane 7, pTT/PYC; lane 8, pTT/$E1B^{19K}$; lane 9, pTT/hexokinase; lane 10, pTT/glucokinase. Cells were harvested 72 hpt, rinsed with PBS and solubilized in NuPAGE sample buffer followed by sonication (lanes 1 – 5) or extracted in lysis buffer (lanes 6 – 10) as indicated in Materials and Methods. Proteins were resolved on a 4 – 12% Bis-Tris NuPAGE gradient gel and stained with Coomassie blue R-250.

Fig. 15 is a graph showing the effect of an antibiotic resistance cassette added to a vector. When an antibiotic resistance cassette is added to the vector (in this example the pTTz vector and a zeocin expression cassette) and the antibiotic is added to the cell culture after transfection, a stable population of cells expressing the transgene (in this example GFP) can be obtained is less than a month.

Fig. 16 is a graph showing transient gene expression levels reached in various SFE clones compared to the parental 293SF-3F6 cell line (ATCC Accession No. CRL-12585). Cells were transfected with the pTT/SEAP plasmid. SFE clones were obtained following transfection of the 293SF-3F6 cell line with the pIRES-neo (Clontech) vector encoding the full-length EBNA1 protein and selection using 50 µg/ml of geneticin for two weeks. Resistant cells were seeded at 1cell/well in 96 well plates and emerging clones amplified and tested for transient gene expression. Of the 20 clones so isolated, only four are shown in the graph, the clone 41 (deposited

under IDAC Accession No. 020502) being the one showing the highest transgene expression.

Fig. 17 is a graph showing the effect of the addition of serum sub-fractions on transient gene expression. Panel A shows the bovine calf serum (BCS, Hyclone) protein profile following gel filtration chromatography using a Superdex 200 HR 16/60 column. Fractions were tested at a final concentration of 40% (v/v). Panel B, 293E cells were transfected with the pTT/SEAP vector and SEAP measured 72 hours later. Active fractions were pooled for further analysis (11-14: "fraction A" and 24-25: "fraction B").

Fig. 18 is a graph showing the effect of the combination of serum "fraction A" and "fraction B" on transient gene expression. 293E cells were transfected with pTT/SEAP in the absence or presence of various ratio of "fraction A" and "fraction B" (see Fig.17B). SEAP activity was measured 96 hours later. Positive control was medium with 1% BCS and negative control was plain medium.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention provides a new process for an enhanced transient expression of recombinant proteins (r-proteins) in eukaryotic cells, preferably in mammalian cells, most preferably in suspension-growing 293 cell lines.

The process was developed and optimized by investigating the effects of various parameters of the process on r-protein expression, by transient transfection of suspension-growing cells using the polycationic polymer polyethylenimine (PEI) as transfection reagent. In a preferred embodiment, by combining the optimized oriP-containing pTT expression plasmid with the 293E cell line, expression levels of intracellular r-protein representing up to 20% of total cellular proteins (TCP) have been achieved. To the inventors' knowledge, such high expression levels have never been achieved in 293 cells using transient transfection and these levels rival those obtained using virus-mediated transgene expression (Massie B. et al.,

Cytotechnology, **28**, 53 – 64 (1998)). Expression of the human placental secreted alkaline phosphatase (SEAP) protein, one of several proteins expressed using the process of the present invention was found to be at levels exceeding 20 mg/l.

It would be obvious to persons skilled in the art that many different eukaryotic cell lines, in particular mammalian and human cell lines, could be transfected using PEI as transfecting agent and that such cell lines could be used for the process of the invention. However, the efficacy of transfection, and protein expression levels as a result, vary considerably for various cell lines (see, for example Boussif et al., *Gene Therapy*, **3**, 1074 – 1080 (1996)) and is one of the highest for human embryonic kidney (HEK) 293 cell line. Also the activity of the CMV promoter appears to be one of the highest for HEK 293 cell line, as compared with other mammalian cell lines, which considerably improves expression levels of the recombinant protein when this promoter is used in combination with a human embryonic kidney (HEK) cell line. An additional improvement may be achieved by using the HEK 293E cell line (a genetic variant of 293 cell line, constitutively expressing the Epstein-Barr virus (EBV) EBNA1 protein), in combination with an expression vector comprising the EBV oriP-sequence or a fragment thereof containing EBNA1 binding sites (see Fig. 2B).

Particularly advantageous for carrying out the process of the invention proved to be the SFE cell line, a derivative of the cell line HEK293SF-3F6 (ATCC Accession No. CRL-12585) stably expressing the EBNA1 protein. The SFE cell line was developed by transfecting the 293SF-3F6 cell line with the pIRES-neo vector encoding the full-length EBNA1 protein and isolating and amplifying geneticin-resistant clones so obtained, following procedures well known to those skilled in the art. The isolated clones were then tested for transient expression of the SEAP gene. A clone (clone 41) showing the highest expression levels (see Fig. 16) was deposited under IDAC Accession No. 020502. The SFE cell line offers the advantage over the commercially available HEK 293E cell line of being capable of growing in a serum-free medium. Production of secreted r-proteins in a serum-free medium considerably facilitates their subsequent purification.

The use of amplifiable expression cassettes in mammalian cells, such as the dihydrofolate reductase or glutamine synthetase systems, have been shown to result in the isolation of stable call lines showing very high levels of r-protein expression. As an alternative to these stable amplified systems, vectors with viral-derived elements that allow for episomal replication and amplification, such as the large-T antigen/SV40 ori, or the EBNA1/oriP, are well suited when using transient expression systems (Van Craenenbroeck K. et al., *Eur. J. Biochem.*, **267**, 5665 – 5678 (2000)). Although plasmid DNA containing the SV40 ori was shown to replicate in the large–T antigen expressing 293T cell line (Heinzel S. S. et al., *J. Virol.*, **62**, 3738 – 3746 (1988)), it was now shown that it did not provide higher transgene expression in 293T cells when compared with the 293 parental cell line. In contrast, the use of oriP-containing plasmids in 293E cells significantly increased transgene expression compared with the non-permissive 293 cells. This suggests that the increased transgene expression obtained using EBV replicon-containing plasmids might be mediated by a phenomenon distinct from its ability to support episomal replication. This is further supported by the fact that removal of DS domain of oriP, which is responsible for initiation of DNA replication in EBNA1 positive cells (Wysokensky D. A. et al., *J. Virol.*, **63**, 2657 – 2666 (1989)), did not significantly reduce transgene expression (see Fig. 6). One likely mechanism for this oriP-mediated increased expression could arise from the described EBNA1-dependent enhancer activity of oriP (Reisman D. et al., *Moll. Cell. Biol.*, **6**, 3838 – 3846 (1986); Sugden B. et al., *J. Virol.*, **63**, 2644 – 2649 (1989); Gahn T. A. et al., *J. Virol.*, **69**, 2633 – 2636 (1995)). The EBV oriP contains 24 EBNA1 binding sites (Mackey D. et al., *Methods Enzymol.*, **306**, 308 – 328 (1999)). As EBNA1 has an efficient nuclear localization signal (Ambinder R. F. et al., *J. Virol.*, **65**, 1466 – 1478 (1991); Längle-Rouault F. et al., *J. Virol.*, **72**, 6181 – 6185 (1998)), its binding to plasmids bearing oriP may also increase their nuclear import, thus enhancing transgene expression. This effect is illustrated in Fig. 10, where co-transfection of the pcDNA3.1/SEAP plasmid (no oriP) with an oriP empty vector in a ratio of only 1:9 maintained specific SEAP production. In contrast, a co-transfection using the same ratio with an empty vector without oriP lead to a five-fold decrease in specific SEAP production. This suggests that the presence of an oriP vector in PEI-DNA complexes is sufficient to increase nuclear

import of non-oriP vectors that are present in the same complexes, thus increasing protein expression. Indeed, the most important barrier to transfection seems to be the limited migration of plasmid DNA from the cytoplasm to the nucleus (Zabner J. et al., *J. Biol. Chem.*, **270**, 18997 – 19007 (1995)). Contribution of this mechanism to the enhanced transgene expression could be partially hindered when using PEI as the transfection reagent, as this polymer was also shown to actively undergo nuclear localization (Pollard H. et al., *J. Biol. Chem.*, **273**, 7507 – 7511 (1998); Godbey W. T. et al., *Proc. Natl. Acad. Sci. USA*, **96**, 5177 – 5181 (1999)). However, data presented in Fig. 9 clearly show a significant contribution of oriP to an enhanced nuclear transport of plasmid DNA.

Whereas linear 25 kDa PEI was reported to efficiently mediate gene transfer in the presence of serum (Boussif O. et al., *Gene Ther.*, **3**, 1074 – 1080 (1996)), transgene expression mediated by the branched isoform was shown to be reduced 3-fold in its presence (Schlaeger E-J. et al., *Cytotchnology*, **30**, 71 – 83 (1999)). This contrasts with findings of the present invention showing that gene transfer was also significantly increased using the branched 25 kDa PEI.

A positive effect of serum as a component of the culture medium on transfection efficiency and protein expression was also observed (see Fig. 3). The mechanism by which serum increases gene delivery and/or transgene expression is not yet clear. Serum might contribute to augment transcriptional activity of the promoter as the CMV immediate early enhancer contains multiple binding sites for serum-activated transcription factors (Boshart M. et al., *Cell*, **41**, 521 – 530 (1985); Brightwell G. et al., *Gene*, **194**, 115 – 123 (1997)). However, only a partial recovery of transgene expression was obtained when serum was added to the cells 3 hrs after their transfection in serum-free medium. This suggests that, in addition to the potential serum-mediated CMV promoter transcription activation, some serum component(s) might increase transfection efficacy of DNA-PEI complexes. The results shown in Fig. 17 & 18 demonstrate that following serum fractionation, inhibitory components (such as BSA) can be removed, and fractions enhancing transfection and/or transgene expression can be isolated. Further purification of the active components

will allow to obtain an additive with minimal protein content (compared to whole serum), that will greatly increase production yields.

As attempts to adapt the commercially available 293E cell line to serum-free medium were unsuccessful, it was decided to create an EBNA1-expressing cell line growing in a serum-free medium by stably transfecting the serum-free adapted 293SF-3F6 clone with an EBNA1 expression plasmid (pIRESneo/EBNA1). Among multiple clones tested, the clone 41 showed the highest transgene expression following transient transfection of the pTT/SEAP plasmid (a ten-fold increase in SEAP expression compared to the 293SF-3F6 parental clone; see Fig. 16). Using this clone, the serum-free medium formulation was further improved in order to reach higher transient gene expression. Of various peptones tested as additives to the serum-free medium, the gelatin peptones GPN3 proved to be the most suitable for this purpose. Other peptones were similarly effective (see Fig. 12), but induced significant cell agglomeration, an undesirable phenomenon in suspension cultures. BSA was removed form the culture medium, as this protein proved to significantly inhibit the transfection and gene expression (data not shown).

A major drawback of using polycations or cationic lipids is the inhibitory effect of conditioned medium on gene delivery. In the case of cationic lipids, this inhibition was shown to be mediated by the presence of secreted glycosaminoglycans (Rupoen M. et al., *Biochim. Biophys. Acta*, **1415**, 331 – 341 (1999); Belting M. et al., *J. Biol. Chem.*, **274**, 19375 0 19382 (1999)), which are expected to efficiently displace DNA from lipid complexes. Whereas it was shown that conditional medium adversely reduced PEI-mediated transfection of 293E cells (Schlaeger E-J. et al., supra), no significant effect was observed by the inventors. The reason for this discrepancy is not clear, but might result from the type of culture medium used, the age of the culture, or from the cells themselves. The fact that, according to the invention, transfection of cells in their 24hr-conditioned medium does not reduce gene transfer and expression, greatly simplifies process scale up.

In conclusion, a significant improvement in transgene expression following transient transfection of suspension-growing cells using PEI was obtained by combining optimized parameters, such as the pTT expression vector, the 293E or 293SFE cell

lines, the culture medium, and the transfection process. Under these conditions, ~60 mg of purified SEAP could be obtained from a 3-l culture following a single IMAC purification step. Volumetric expressions of the intracellular proteins GFP and RR1 were, respectively, 20 and 50 mg/l at 72 hpt, representing up to 20% of TCP. As this technology is robust, inexpensive and easy to perform, it is fully adapted for high-throughput production of milligram quantities of r-proteins needed for biochemical or structural studies and high-throughput screenings.

EXPERIMENTAL

MATERIALS AND METHODS

Chemicals

A 25 kDa branched PEI was obtained from Aldrich (Milwaukee, WI) and 25 kDa linear PEI from Polysciences (Warrington PA). Stock solutions (1 mg ml^{-1}) were prepared in water, neutralized with HCl, sterilized by filtration (0.22 µm), aliquoted and stored at -80°C.

Cell culture

Human embryonic kidney 293S (293) cells (Côté J. et al., *Biotechnol. Bioeng.*, **59**, 567 – 5765 (1998)) and genetic variants stably expressing EBNA1 (293E) (Invitrogen, Carlsbad, CA) or the large-T antigen (293T) (DuBridge R. B. et al., *Mol. Cell. Biol.*, **7**, 379 – 387 (1987)) were adapted to suspension culture in low-calcium-hybridoma serum-free medium (HSFM) (Côté J. et al., supra) supplemented with 1% bovine calf serum (BCS), 50 µg ml^{-1} Geneticin (for 293E and 293T cells), 0.1% Pluronic F-68 (Sigma, Oakville, Ontario, Canada) and 10 mM HEPES. For culture in bioreactors, HEPES was omitted from the medium. Cells were cultured in Erlenmeyer flasks (50 or 125 ml) using 15 - 25% of the nominal volume at 110 – 130 r.p.m. (Thermolyne's BigBill orbital shaker, TekniScience Inc., Terrebonne, Québec, Canada) under standard humidified conditions (37°C and 5% CO_2).

Vectors

The pIRESpuro/EGFP (pEGFP) and pSEAP basic vectors were obtained from Clontech (Palo Alto, CA), and pcDNA3.1, pcDNA3.1/Myc-$(His)_6$ and pCEP4 vectors were from Invitrogen. The SuperGlo GFP variant (sgGFP) was from Q•Biogene (Carlsbad, CA). Construction of pCEP5 vector was as follows: the CMV promoter and polyadenylation signal of pCEP4 were removed by sequential digestion and self-ligation using *SalI* and *XbaI* enzymes, resulting in plasmid pCEP4Δ. A *BglII* fragment from pAdCMV5 (Massie B. et al., *J. Virol.*, **72**, 2289 – 2296 (1998) 11) encoding the CMV5-poly(A) expression cassette was ligated in *BglII*-linearized pCEP4Δ, resulting in pCEP5 vector. The pTT vector was generated following deletion of the hygromycin (*BsmI* and *SalI* excision followed by fill-in and ligation) and EBNA1 (*ClaI* and *NsiI* excision followed by fill-in and ligation) expression cassettes. The ColE1 origin (*FspI* - *SalI* fragment, including the 3' end of ß-lactamase ORF) was replaced with a *FspI* - *SalI* fragment from pcDNA3.1 containing the pMB1 origin (and the same 3' end of ß-lactamase ORF). A Myc-$(His)_6$ C-terminal fusion tag was added to SEAP (*HindIII* - *HpaI* fragment from pSEAP-basic), following in-frame ligation in pcDNA3.1/Myc-His (Invitrogen) digested with *HindIII* and *EcoRV*. To insert a SV40 promoter – zeocin - SV40 polyA expression cassette into the pTT vector (resulting in pTTz vector), the cassette was first amplified from pZeo(SV2+) vector (Invitrogen) using primers with BspHI sites at their extremities. The amplified cassette was then ligated between the BspHI sites of pTT vector. All plasmids were amplified in *Escherichia coli* (DH5α) grown in LB medium and purified using MAXI prep columns (Qiagen, Mississauga, Ontario, Canada). For quantification, plasmids were diluted in 50mM Tris-HCl pH 7.4 and the absorbances at 260 and 280 nm measured. Only plasmid preparations with A_{260}/A_{280} ratios between 1.75 and 2.00 were used.

Small-scale transient transfections

Three hours before transfection, cells were centrifuged and resuspended in fresh HSFM medium supplemented with 1% BCS at a density of 1.0×10^6 cells ml^{-1}. Five hundred microliters, or 10 ml, of cell suspension was distributed per well of a 12 well plate, or in a 125 ml shaker flask, respectively. DNA was diluted in fresh serum-free HSFM (in a volume equivalent to one-tenth of the culture to be transfected), PEI was

added, and the mixture immediately vortexed and incubated for 10 min at room temperature prior to its addition to the cells. Following a 3 h incubation with DNA-PEI complexes, culture medium was completed to 1 ml (12-well plate) or 20 ml (shaker flask) by the addition of HSFM supplemented with 1% BCS.

Transfection in bioreactors

A 3.5-l bioreactor containing 2.85 l of HSFM supplemented with 1% BCS was seeded with 293E cells to obtain a final cell density of 2.5 x 10^5 ml^{-1}. Twenty-four hours later, cells were transfected with 150 ml of a mixture of pTT/SEAP:pEGFP plasmids (19:1, 3 mg total) and PEI (6 mg). Agitation was at 70 r.p.m. using a helical ribbon impeller (Kamen A. A. et al., *Chem. Eng. Sci.*, **27**, 2375 – 2380 (1992)). Dissolved oxygen was maintained at 40% by surface aeration using a nitrogen/oxygen mixture (300 ml/min) and pH was maintained at 7.2 by addition of CO_2 in the head space and sodium bicarbonate (10% w/v in water) injection in the culture medium. The same conditions were used for transfection in 14-l bioreactors.

Flow cytometry

GFP was analyzed by flow cytometry using an EPICS Profile II (Coulter, Hialeah, FL, USA) equipped with a 15-mW argon-ion laser. Only viable cells were analyzed for the expression of GFP. Data are representative of at least two independent experiments. Error bars represent ±SEM of one experiment done in duplicate

SEAP analysis

Determination of SEAP activity was performed essentially as previously described (Durocher et al., *Anal. Biochem.*, **284**, 316 – 326 (2000)). Briefly, culture medium was diluted in water as required (typically 1/50 to 1/1000) and 50 µl were transferred to a 96-well plate. Fifty microliters of SEAP assay solution containing 20 mM paranitrophenylphosphate (pNPP), 1 mM $MgCl_2$, 10 mM l-homoarginine and 1 M diethanolamine pH 9.8 were then added and absorbance read at 410 nm at 1-2 min intervals at room temperature to determine pNPP hydrolysis rates. Data are representative of at least two independent experiments. Error bar represent ±SEM of one experiment done in duplicate. For the bioreactor run, error bars represent ±SEM of two SEAP measurements.

Electrophoresis, western analyses and quantification

Immunodetection of C-terminal Myc-$(His)_6$-tagged SEAP was done using the anti-Myc 9E10 antibody (Santa Cruz). For analysis of intracellular proteins, cells were directly lysed in NuPAGE sample buffer (Novex) or extracted with lysis buffer (50 mM HEPES pH 7.4, 150 mM NaCl, 1% Thesit and 0.5% sodium deoxycholate). Insoluble material was removed from lysates by centrifuagtion at 12 000 g at 4°C for 5 min. Concentrated NuPAGE buffer (4x) was added to clear lysates. All samples were heated for 3 min at 95°C. Proteins were resolved on 4 – 12% Bis-Tris or 3 – 8% Tris-acetate NuPAGE gradient gels as recommended by the manufacturer. GFP and other non-tagged proteins were quantified relative to purified bovine serum albumin (BSA) following electrophoresis and Coomassie blue R250 staining using the Kodak Digital Science Image Station 440cf equipped with the Kodak Digital Science 1D image analysis software version 3.0 (Eastman Kodak, NY, USA). RR1 was quantified by slot-blot relatively to a homogeneity-purified RR1 standard detected by using a monoclonal anti-RR1 antibody. Other Myc-$(His)_6$-tagged proteins were quantified relative to purified SEAP-Myc-$(His)_6$.

EXAMPLES

Transfection with linear and branched 25 kDa PEI

Preliminary results showed that linear and branched 25 kDa PEI were the most effective among various polymers tested (including branched 70 kDa, branched 50 – 100 kDa and branched 10 kDa; data not shown). In view of the above, transfection of 293E cells was optimized with both linear or branched 25 kDa PEI polymers using a plasmid encoding the enhanced GFP (pEGFP). Transfections were performed using cells grown as monolayers in 12-well plates and GFP expression was measured 72 hours later by flow cytometry. The effect of DNA to PEI ratios on transfection efficiency is shown in Figure 1 using linear (A) or branched (B) PEI. The indicated amounts of DNA and polymers are for one well containing $5x10^5$ cells. Only 0.25 µg of DNA per well was sufficient to reach a 50% transfection efficiency when using linear PEI, whereas a minimum of 1.0 µg was necessary using the branched isoform.

Transfection efficiencies of ~70% were reached with both linear and branched polymers at DNA:PEI (µg:µg) ratios of 1.0:1.5 and 1.5:2.0, respectively. Increasing the amounts of both DNA and PEI did not lead to higher transfection yield.

Cell line and expression vectors

Two commercially available expression vectors containing viral sequences allowing for episomal DNA replication in permissive cell lines were tested. The first vector, pcDNA3.1, contains the SV40 origin of replication that allows cellular polymerases to replicate the DNA up to 1000-copies in cells expressing the large T antigen (Chittenden T. et al., *J. Virol.*, **65**, 5944 – 5951 (19991)). The second vector, pCEP4, contains the EBV origin of replication oriP that replicates plasmid DNA up to 90-copies in cells expressing the EBNA1 protein (Yates J. L. et al., *Nature*, **313**, 812 – 815 (1985)). Also generated was the pCEP5 vector (Fig. 2A, left) by using an improved CMV expression cassette, as described in the adenoviral transfer vector pAdCMV5 (Massie B. et al., *Biotechnology*, **13**, 602 – 608 (1995)). This expression cassette has been shown to confer very high levels of r-protein expression in 293 cells (Massie B. et al., *Cytotechnology*, **28**, 53 – 64 (1998) 12). The pCEP5 vector was further modified (see Materials and Mathods) to yield the pTT vector (Fig. 2A, right) that is 4.6 kb smaller, hence providing more space for large cDNA cloning. The cDNA encoding for the reporter protein SEAP was then cloned in each of these four vectors and its expression level monitored following transient transfection in 293, 293T or 293E cells. As shown in Fig. 2B, transfection of 293T cell line with the SV40 ori-containing plasmid pcDNA3.1 did not translate into an increased transgene expression when compared with transfection of the parental 293 cells. However, transfection of 293E cells with pCEP4 vector resulted in a 2-3-fold increase in SEAP expression compared with transfection of 293 or 293T cells with the same vector. In addition, the use of pCEP5 vector further increased SEAP expression by a factor of 2-6-fold, depending on the cell line. Finally, the use of the pTT vector in 293E cells resulted in a 33% increase in transgene expression compared with the pCEP5 vector. The overall SEAP expression level in 293E cells was 10-fold higher with the pTT vector compared with pcDNA3.1 vector.

Effect of serum

The effect of serum on transfection efficiency (GFP) and r-protein production (SEAP) mediated by both linear and branched PEI was evaluated. Fig. 3 shows that when transfection mixture was added to cells in fresh 1% serum-containing medium, a 4-5-fold increase in SEAP activity 72 hpt is obtained compared with its addition to cells in serum-free medium. Increasing serum concentration to 5% further improved PEI-mediated transfection efficiency and production. When transfection mixture was added to cells in serum-free media followed 3 hours later by serum addition to a concentration of 1% (0→1%), a 2-fold increase in transgene expression was obtained; however, this level was only 50% of that obtained in 1% serum.

Process optimization for transfection in suspension

Next evaluated was gene transfer efficiency of both linear and branched PEI on suspension-growing 293E cells grown in 1% BCS-supplemented HSFM. Shaker flask cultures were co-transfected with a mixture of pTT/SEAP:pEGFP (9:1) plasmids (pEGFP was added to monitor for transfection efficiency). With both linear and branched PEI, SEAP accumulated in the culture medium for up to 96 hours post-transfection (hpt) (Fig. 4), but gene transfer and expression level were 50% higher using the linear isoform. These results clearly demonstrate that linear, and to a lesser extent branched PEI are effective for gene transfer in suspension-growing cells. In addition, SEAP expression levels obtained with suspension-growing cells using linear PEI were comparable with those obtained with adherent-growing cells. For all experiments discussed below, only linear PEI was used.

In order to design a robust, simple and scalable transfection process, two steps had to be simplified: the 3 hrs incubation of DNA-PEI complexes with cells in a reduced culture volume, and the medium change 3 hrs prior to transfection. The first step was performed with the assumption that it would promote interaction of the DNA-PEI complexes with the cells and thus increase transfection efficiency. The second was done according to reports showing deleterious effect of conditioned medium on transfection efficiency (Schlaeger E-J. et al., *Cytotechnology*, **30**, 71 – 83 (1999); Ruponen M. et al., *Biochim. Biophys. Acta*, **1415**, 331 – 341 (1999)). Whereas

medium exchange is simple to perform on a small scale, this step represents a significant hurdle at scales greater than a few liters.

The effect of cell density at the time of transfection was first evaluated (Fig. 5A) by transfecting high density (hatched bars; 10 ml at 1 x 10^6 cells ml^{-1}) or low density cultures (gray bars; 20 ml at 2.5 x 10^5 cells ml^{-1}) in shaker flasks. Three hours later, the high cell density flask was diluted to 5 x 10^5 cells ml^{-1} with fresh medium, and GFP expression monitored 72 hrs later. This experiment showed that cell concentrations prior to transfection could be omitted, as only a slight decrease (<10%) in transfection efficiency and a 15% decrease in GFP expression level was observed when cells were transfected in a larger culture volume.

The next evaluated was the effect of conditioned medium on SEAP expression using suspension growing cells. For this study, cells were seeded in shaker flasks at a density of 2.5 x 10^5 cells ml^{-1}. Twenty-four hours later, transfection was performed with or without a complete medium exchange. As shown in Fig. 5B, no significant difference in SEAP expression was observed when the transfection was carried out in medium conditioned for 24 hrs, indicating that medium exchange is not necessary.

Transfection in bioreactors

To demonstrate the scalability of the process, a 3.5-l bioreactor culture was transfected with a mixture of pTT/SEAP:pEGFP plasmids (19:1).One hour later, a sample (25 ml) was withdrawn and transferred into a shaker flask as a control. In the bioreactor (Fig. 13A, solid lines), SEAP (circles) accumulated up to 144 hpt and then reached a plateau, whereas accumulation continued up to 216 hpt in the control shaker flask (dashed lines). The percentage of GFP-positive cells (squares) at 96 hpt reached 54 and 50% for the bioreactor and the shaker flask, respectively. At the end of the culture , cell density was 4.1 and 4.7 x 10^6 cells ml^{-1} with a viability of 62 and 72% for the bioreactor and the shaker flask, respectively (Fig. 13B). Although viable cell density was 25% lower in the bioreactor compared with the shaker flask, volumetric SEAP productivity was almost 2-fold higher. Similar results were systematically observed in five independent experiments (results not shown),

indicating that the productivity of secreted proteins might be increased when using a controlled environment.

Purification of SEAP and production of other r-proteins

Purification of Myc-$(His)_6$-tagged SEAP harvested from the bioreactor run (Fig. 13) by immobilized metal affinity chromatography (IMAC) is shown in Fig. 14A. The left panel shows Coomassie blue-stained protein pattern from the culture medium before loading on the column (lane 1), flow-through (lane 2) and eluted material using 150 mM imidazole (lane 3). The right panel shows immunodetection of SEAP in the same fractions using anti-Myc antibody. This figure shows that all of the His-tagged SEAP was retained on the column, whereas very few, if any, serum protein bound to it (SEAP migrates with an apparent molecular weight slightly higher than BSA). SEAP quantification in the eluted fraction using the Lowry protein assay showed that ~60 mg of His-tagged SEAP could be recovered by IMAC from the 3-l bioreactor culture. As shown in Fig. 14B, high expression levels in bioreactor were also obtained with other secreted r-proteins. Fourteen- (lanes 1, 3 and 4) or 3.5-liter (lane 2) bioreactors were transfected with pTT plasmids encoding for Neurophilin-1 and VEGF (1:1 ratio, lane 1), Tie2 (lane 2), Cripto (lane 3) and c-Met (lane 4). All cultures were harvested 5 days post-transfection. With the exception of Cripto, which has been reported highly glycosylated on serine, threonine and asparagine (Schiffer S. G. et al., J. Biol. Chem., 276, 37769 – 37778 (2001) 22), glycosylation of the expressed proteins appeared to be relatively homogenous, as suggested by their migration behaviour following SDS-PAGE. High expression levels of intracellular r-proteins were also obtained as shown in Fig. 14C. In this experiment, 293E cells were transfected with pTT plasmids encoding for sgGFP (lane 1), herpes simplex virus ribonucleotide reductase (RR1, lane 2), mouse $G_{\alpha q}$ (lane 5), human $p27^{Kip1}$ (lane 6), yeast pyruvate carboxylase (PYC, lane 7), adenovirus $E1B^{19K}$ (lane 8), human hexokinase 1 (HK, lane 9) and human glucokinase (GK, lane 10). Three days after transfection, cells were rinsed with PBS, solubilized in sample buffer (GFP, RR1 and $G_{\alpha q}$) or extracted with lysis buffer ($p27^{Kip1}$, PYC, $E1B^{19K}$, HK and GK), and proteins analyzed by SDS-PAGE. Quantification of r-proteins shown in Fig. 14 is summarized in Table 1.

Table 1. Summary of r-protein expression level

r-Protein	Tag	Localization	Culture mode	Concentration (mg/l)
Human SEAP	Myc-$(His)_6$	Secreted	3-l bioreactor	20[a]
Human Neuropilin-1	Myc-$(His)_6$	Secreted	14-l bioreactor	8[b]
Human VEGF	Myc-$(His)_6$	Secreted	14-l bioreactor	10[b]
Human Tie2	Myc-$(His)_6$	Secreted	3-l bioreactor	9
Human Cripto	Myc-$(His)_6$	Secreted	14-l bioreactor	9
Human c-Met	Myc-$(His)_6$	Secreted	14-l bioreactor	1
sgGFP	None	Intracellular	Shaker flask	20
Herpes virus RR1	None	Intracellular	Shaker flask	50
Mouse $G\alpha_q$	None	Membrane	T-flask	16
Human $p27^{Kip1}$	None	Intracellular	T-flask	14
Human hexokinase	None	Intracellular	Shaker flask	40
Human glucokinase	None	Intracellular	Shaker flask	30
Yeast PYC	None	Intracellular	1-l bioreactor	4
Adenovirus $E1B^{19K}$	None	Intracellular	T-flask	3

[a] After purification by IMAC

[b] Neurophilin-1 and VEGF were co-transfected

In the case of RR1, volumetric production was 50 mg/l, representing 20% of total cell protein (TCP). The mouse $G\alpha_q$ was expressed at 16 mg/l, compared with a barely detectable level (by Coomassie staining) when expressed from pcDNA3.1 vector (lane 4).

Although various particular embodiments of the present invention have been described hereinbefore for purposes of illustration, it would be apparent to those skilled in the art that numerous variations may be made thereto without departing from the spirit and scope of the invention, as defined in the appended claims.

WHAT IS CLAIMED IS:

1. An expression vector for an enhanced expression of a recombinant protein in human cells which stably express EBNA1 protein or a fragment thereof, said vector comprising a first DNA sequence encoding the recombinant protein, said first DNA sequence being under control of a CMV5 promoter, said expression vector further comprising a second DNA sequence of the oriP sequence of Epstein-Barr virus (EBV) or a fragment thereof enhancing transcriptional activity of the promoter.

2. An expression vector according to claim 1, wherein the vector is a plasmid.

3. An expression vector according to claim 1, wherein the fragment of the EBV oriP sequence comprises at least one EBNA1 binding site (EBS).

4. An expression vector according to claim 1, wherein the fragment of the oriP sequence is selected from the group consisting of BstX I FR fragment, a BstX I EcoR I FR fragment, the FR domain, and fragments thereof.

5. A human kidney cell line designated 293SFE and deposited under IDAC Accession No. 020502, cells of said cell line transfected with an expression vector as defined in claim 1.

6. A process for preparing transfected human kidney cells of the 293 cell line, which process comprises transfecting said human kidney cells with an expression vector according to claim 1.

7. A process according to claim 6, when carried out in the presence of polyethylenimine (PEI) as a transfection reagent.

8. A process according to claim 7, wherein the polyethylenimine is a linear polyethylenimine or a branched polyethylenimine.

9. A process according to claim 8, wherein the average molecular weight of the polyethylenimine is from about 10 to about 100 kDa.

10. A process according to claim 9, wherein the average molecular weight of the polyethylenimine is about 25 kDa.

11. A process for preparing a recombinant protein, which process comprises culturing transfected human kidney cells prepared by the process of claim 6 to yield said protein.

12. A process according to claim 11, wherein the cells are cultured in a serum-free culture medium.

13. A process according to claim 11, wherein the cells are cultured in a culture medium further comprising a serum or a subfraction thereof.

14. A process according to claim 11 wherein the cells are cultured in a culture medium further comprising a peptone.

15. A process according to claim 14, wherein the peptone is the gelatin peptone N3.

16. A process according to claim 11, wherein the process is carried out in a culture medium without changing the culture medium.

17. A process for preparing a recombinant protein, which process comprises transfecting human kidney cells of the 293 cell line with an expression vector according to claim 1 and then culturing said transfected human kidney cells to yield said protein.

18. A process for preparing a protein, which process comprises culturing the human kidney cell line as defined in claim 5 to yield said protein.