WHAT IS CLAIMED IS:

1. A method comprising the steps of:

(a) subjecting harvested Dupilumab to affinity chromatography;

(b) subjecting said Dupilumab pooled from eluate of step (a) to viral inactivation at a pH from about 3 to about 4 and then adjusting the pH to from about 5 to about 8;

(c) subjecting said Dupilumab pooled from step (b) to anion exchange chromatography in flowthrough mode;

(d) subjecting said Dupilumab pooled from flowthrough fractions of step (c) to cation exchange chromatography in bind and elute mode;

(e) subjecting said Dupilumab pooled from eluate of step (d) to hydrophobic interaction chromatography in flowthrough mode; and

(f) subjecting said Dupilumab pooled from flowthrough fractions of step (e) to virus retentive filtration to produce Dupilumab.

2. The method of claim 1, further comprising a harvest pre-treatment step prior to step (a).

3. The method of claim 2, wherein said harvest pre-treatment step includes adjusting said Dupilumab to a transient pH level from about 4 to 5.5.

4. The method of claim 1, wherein the concentration of said Dupilumab pooled from flowthrough fractions in step (f) is between about 4 g/L to 12 g/L.

5. The method of claim 1, further comprising subjecting said Dupilumab to ultrafiltration and diafiltration (UF/DF) after step (f).

6. The method of claim 5, wherein said UF/DF includes a diafiltration buffer having a pH between 4.0 and 4.5.

7. The method of claim 5, wherein a concentrated Dupilumab pool following UF/DF has a pH of about 5.3.

8. The method of claim 5, wherein said UF/DF includes a diafiltration buffer comprising between about 4 mM acetate and about 6 mM acetate.

9. The method of claim 1, wherein said affinity chromatography is Protein A chromatography.

10. The method of claim 9, wherein a Protein A resin is selected from the group consisting of MabSelect PrismA, MabSelect SuRe, MabSelect SuRe LX, MabSelect, MabSelect SuRe pcc, MabSelect Xtra, rProtein A Sepharose, ProSep HC, ProSep Ultra, ProSep Ultra Plus, MabCapture, and Amsphere A3.

11. The method of claim 9, wherein a Protein A resin is selected that is capable of receiving a load at a concentration above 55 g/L.

12. The method of claim 1, wherein about 180 g to 200 g of Dupilumab is loaded per liter of HIC resin.

13. The method of claim 1, wherein the amount of PLBD2 in the HIC eluate is reduced compared to the amount of PLBD2 in the HIC load, optionally wherein the amount of PLBD2 in the HIC eluate is reduced to below 100 ppm, reduced to below 30 ppm, reduced to below 4 ppm, reduced to below 1 ppm, or reduced by about 40x-310x compared to the amount of PLBD2 in the HIC load.

14. The method of claim 9, wherein a Protein A column load pH is between 6 and 8.

15. The method of claim 1, wherein said Dupilumab comprises three heavy chain complementarity determining region (HCDR) sequences comprising SEQ ID NOs: 3, 4, and 5, and three light chain complementarity determining region (LCDR) sequences comprising SEQ ID NOs: 6, 7, and 8.

16. The method of claim 1, wherein said Dupilumab comprises a heavy chain variable region (HCVR) comprising the amino acid sequence of SEQ ID NO: 1 and a light chain variable region (LCVR) comprising the amino acid sequence of SEQ ID NO: 2.

17. A method comprising the steps of:

(a) culturing cells expressing Dupilumab;

(b) subjecting said cells to transient pH levels from about 4.0 to 5.5, then raising pH levels to from about 5.5 to 6.5;

(c) harvesting said cells by centrifugation to separate cell debris from clarified media comprising said Dupilumab;

(d) subjecting said clarified media to affinity chromatography;

(e) subjecting said Dupilumab pooled from eluate of step (d) to viral inactivation at a pH from about 3 to about 4 and then adjusting the pH to from about 5 to about 8;

(f) subjecting said Dupilumab pooled from step (e) to anion exchange chromatography in flowthrough mode;

(g) subjecting said Dupilumab pooled from flowthrough fractions of step (f) to cation exchange chromatography in bind and elute mode;

(h) subjecting said Dupilumab pooled from eluate of step (g) to hydrophobic interaction chromatography in flowthrough mode; and

(i) subjecting said Dupilumab pooled from flowthrough fractions of step (h) to virus retentive filtration to produce Dupilumab.

18. The method of claim 17, further comprising subjecting said Dupilumab to ultrafiltration and diafiltration (UF/DF) after step (i).

19. The method of claim 17, wherein said affinity chromatography is Protein A chromatography.

20. The method of claim 19, wherein a Protein A resin is selected from the group consisting of MabSelect PrismA, MabSelect SuRe, MabSelect SuRe LX, MabSelect, MabSelect SuRe pcc, MabSelect Xtra, rProtein A Sepharose, ProSep HC, ProSep Ultra, ProSep Ultra Plus, MabCapture, and AmsphereA3.

21. The method of claim 17, wherein an anion exchange resin is selected from the group consisting of Q Sepharose Fast Flow, Poros 50PI, Poros 50HQ, Capto Q Impres, Capto DEAE, Toyopearl QAE-550, Toyopearl DEAE-650, Toyopearl GigaCap Q-650, Fractogel EMD TMAE Hicap, Sartobind STIC PA nano, Sartobind Q nano, CUNO BioCap, and XOHC.

22. The method of claim 17, wherein a cation exchange resin is selected from the group consisting of Fractogel Hicap, Capto SP ImpRes, Capto S ImpAc, CM Hyper D grade F, Eshmuno S, Nuvia C Prime, Nuvia S, Poros HS, and Poros XS.

23. The method of claim 17, further comprising passing said Dupilumab through a LifeAssure filter after the viral inactivation of step (e) and prior to the anion exchange chromatography of step (f).

24. The method of claim 18, wherein said UF/DF step comprises a membrane filter device selected from the group consisting of Pellicon 2, Pellicon 3 cassettes with 10 kD, 30 kD or 50 kD membranes, Kvick 10 kD, 30 kD or 50 kD membrane cassettes, and Centramate and Centrasette 10 kD, 30 kD or 50 kD cassettes.

25. The method of claim 18, wherein said UF/DF step does not include addition of arginine.

26. The method of claim 17, wherein said HIC step comprises HIC media selected from the group consisting of Capto Phenyl, Capto Phenyl High Sub, Phenyl Sepharose™ 6 Fast Flow,

Phenyl Sepharose™ High Performance, Octyl Sepharose High Performance, Fractogel EMD Propyl, Fractogel EMD Phenyl, Macro-Prep Methyl, Macro-Prep t-Butyl columns, WP HI-Propyl (C3), Toyopearl ether, phenyl or butyl, Toyo PPG; Toyo Phenyl; Toyo Butyl, and Toyo Hexyl.

27. The method of claim 17, wherein said Dupilumab comprises three heavy chain complementarity determining region (HCDR) sequences comprising SEQ ID NOs: 3, 4, and 5, and three light chain complementarity determining region (LCDR) sequences comprising SEQ ID NOs: 6, 7, and 8.

28. The method of claim 17, wherein said Dupilumab comprises a heavy chain variable region (HCVR) comprising the amino acid sequence of SEQ ID NO: 1 and a light chain variable region (LCVR) comprising the amino acid sequence of SEQ ID NO: 2.

29. A method for producing an anti-IL4Rα antibody or antigen-binding fragment thereof, comprising:

(a) subjecting a harvested antibody or antigen-binding fragment thereof to affinity chromatography;

(b) subjecting said antibody or antigen-binding fragment thereof pooled from eluate of step (a) to viral inactivation;

(c) subjecting said antibody or antigen-binding fragment thereof pooled from step (b) to anion exchange chromatography in flowthrough mode;

(d) subjecting said antibody or antigen-binding fragment thereof pooled from flowthrough fractions of step (c) to cation exchange chromatography in bind and elute mode;

(e) subjecting said antibody or antigen-binding fragment thereof pooled from eluate of step (d) to hydrophobic interaction chromatography in flowthrough mode;

(f) subjecting said antibody or antigen-binding fragment thereof pooled from flowthrough fractions of step (e) to virus retentive filtration; and

(g) subjecting said antibody or antigen-binding fragment thereof pooled from step (f) to ultrafiltration and diafiltration (UF/DF) to produce an anti-IL4Rα antibody or antigen-binding fragment thereof, wherein said UF/DF step does not include addition of arginine.

30. The method of claim 29, wherein a diafiltration buffer of step (g) has a pH between 4.0 and 4.5.

31. The method of claim 30, wherein said diafiltration buffer comprises between about 4 mM acetate and about 6 mM acetate.

32. The method of claim 29, wherein a concentrated pool following UF/DF has a pH of about 5.3.

33. The method of claim 29, wherein said UF/DF step comprises a membrane filter device selected from the group consisting of Pellicon 2, Pellicon 3 cassettes with 10 kD, 30 kD or 50 kD membranes, Kvick 10 kD, 30 kD or 50 kD membrane cassettes, and Centramate and Centrasette 10 kD, 30 kD or 50 kD cassettes.

34. A method comprising the steps of:

(a) subjecting a harvested antibody to affinity chromatography;

(b) subjecting said antibody pooled from eluate of step (a) to viral inactivation at a pH from about 3 to about 4 and then adjusting the pH to from about 5 to about 8;

(c) subjecting said antibody pooled from step (b) to anion exchange chromatography in flowthrough mode;

(d) subjecting said antibody pooled from flowthrough fractions of step (c) to cation exchange chromatography in bind and elute mode;

(e) subjecting said antibody pooled from eluate of step (d) to hydrophobic interaction chromatography in flowthrough mode; and

(f) subjecting said antibody pooled from flowthrough fractions of step (e) to virus retentive filtration to produce an anti-IL4Rα antibody.

35. The method of claim 34, further comprising a harvest pre-treatment step prior to step (a).

36. The method of claim 35, wherein said harvest pre-treatment step includes adjusting said antibody to a transient pH level from about 4 to 5.5.

37. The method of claim 34, wherein the concentration of said antibody pooled from flowthrough fractions in step (f) is between about 4 g/L to 12 g/L.

38. The method of claim 34, further comprising subjecting said antibody to ultrafiltration and diafiltration (UF/DF) after step (f).

39. The method of claim 38, wherein said UF/DF includes a diafiltration buffer having a pH between 4.0 and 4.5.

40. The method of claim 38, wherein a concentrated antibody pool following UF/DF has a pH of about 5.3.

41. The method of claim 38, wherein said UF/DF includes a diafiltration buffer comprising between about 4 mM acetate and about 6 mM acetate.

42. The method of claim 34, wherein said affinity chromatography is Protein A chromatography.

43. The method of claim 42, wherein a Protein A resin is selected from the group consisting of MabSelect PrismA, MabSelect SuRe, MabSelect SuRe LX, MabSelect, MabSelect SuRe pcc, MabSelect Xtra, rProtein A Sepharose, ProSep HC, ProSep Ultra, ProSep Ultra Plus, MabCapture, and Amsphere A3.

44. The method of claim 42, wherein a Protein A resin is selected that is capable of receiving a load at a concentration above 55 g/L.

45. The method of claim 34, wherein about 180 g to 200 g of antibody is loaded per liter of HIC resin.

46. The method of claim 34, wherein the amount of PLBD2 in the HIC eluate is reduced compared to the amount of PLBD2 in the HIC load, optionally wherein the amount of PLBD2 in the HIC eluate is reduced to below 100 ppm, reduced to below 30 ppm, reduced to below 4 ppm, reduced to below 1 ppm, or reduced by about 40x-310x compared to the amount of PLBD2 in the HIC load.

47. The method of claim 42, wherein a Protein A column load pH is between 6 and 8.

48. The method of claim 34, wherein said anti-IL-4Rα antibody comprises three heavy chain complementarity determining region (HCDR) sequences comprising SEQ ID NOs: 3, 4, and 5, and three light chain complementarity determining region (LCDR) sequences comprising SEQ ID NOs: 6, 7, and 8.

49. The method of claim 34, wherein said anti-IL-4Rα antibody comprises a heavy chain variable region (HCVR) comprising the amino acid sequence of SEQ ID NO: 1 and a light chain variable region (LCVR) comprising the amino acid sequence of SEQ ID NO: 2.

50. The method of claim 34, wherein said anti-IL-4Rα antibody is Dupilumab.