

## PEPTIDE SYNTHESIS OF ALPHA-HELIXES ON PEG RESIN

The present invention relates to a method for synthesizing particular types of peptide on a solid phase, and to the respective peptide solid-phase conjugates.

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A successful peptide drug, acting as an inhibitor of such a widespread human viral disease such as HIV infection, is T-20 (also named DP-178), which is: Ac-Tyr-Thr-Ser-Leu-Ile-His-Ser-Leu-Ile-Glu-Glu-Ser-Gln-Asn-Gln-Gln-Glu-Lys-Asn-Glu-Gln-Glu-Leu-Leu-Glu-Leu-Asp-Lys-Trp-Ala-Ser-Leu-Trp-Asn-Trp-Phe-NH<sub>2</sub>. The peptide's sequence is taken from an  $\alpha$ -helical, fusogenic sequence segment, a so-called 'heptad repeat', corresponding to amino acids 638-673 of the transmembrane receptor protein gp41 from HIV-1. gp41 is crucial for mediating viral infection of cells.

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Accordingly, efficient large scale production is needed but has so far only been achieved by a combination of partial synthesis of segments of T-20 on solid phase followed by segment condensation typically in the liquid phase (see WO 99/48513). Efficient full-length synthesis on solid-phase has not been demonstrated so far, such approach typically failing to produce more proper product than useless sideproducts, purification of impurities posing further problems and again diminishing final yields.

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Tam, J. et al.(Dep. Microbiology Vanderbilt University; Organic Letters ,2002, Vol.4: 4167-4170) describes inter alia, the preparation of DP-178 and related 30-mer peptides with the purpose of preparing three-helix bundles.

Authors just mentioned that a general Fmoc synthesis method is applied, giving the coupling reagents and resin cleavage conditions used. Nothing is said on the type of resin and/or resin-handle employed, as well as the excess of reagents used. No chromatogram is showed and no purification details nor final purity is given. No indication of yields obtained are given and are entirely left subject to speculation. Since the work pertained only to lab-scale experimentation, no apparent need for devising an efficient synthetic process was given though.

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It is an object of the present invention to devise an improved, efficient method for synthesis of such  $\alpha$ -helical proteins and in particular T-20. This object is solved by the method of the present invention.

**CONFIRMATION COPY**

According to the method of the present invention for preparing a peptide, it comprises the step in that said peptide which may have protected and/or unprotected amino acid side chains is prepared bound to a solid phase which solid phase is a amphiphilic PEG resin and wherein the peptide comprises a prolin-free, helix-forming segment consisting of a run of six contiguous amino acid residues comprising at least four protected or unprotected helix-forming amino acids residues.

Fig. 1 shows the HPLC chromatogram and low level of impurities obtained from the product synthesized in this way, allowed of high-yield synthesis.

PEG resins are offered by different companies, e.g. Matrix Innovations Inc. from Quebec, Canada ('ChemMatrix' brand) or Versamatrix A.S. from Denmark. Such resins have been described e.g. in US2003078372 A1 or, as regards the preferred types of PEG resins, in WO 02/40559.- For linkage of peptide, such PEG resin may comprise terminal 'native' hydroxymethyl radicals or derived thereof 'quasi-native' amino-methyl, carboxyl or bromomethyl or iodomethyl radicals for instance, or may be derivatized by known integral or grafted linkers or handles for solid phase linkage such as e.g. Wang resins, 4-hydroxymethylbenzoic acid (the latter requiring attachment of the first amino acid by means of p-dimethylaminopyridine catalysed esterification, see Atherton et al., 1981, J. Chem. cos. Chem. Commun., p.336ff), 2-chloro-trityl-chloride (CTC) and related, preferably alkoxylated, trityl halogenid resins, Bayer's 4-carboxytrityl linker, 4-methylbenzylhydramine resin, or e.g. the amide generating PAL or Rink or Sieber amide linkers as are used in Fmoc chemistry, or where Boc chemistry is to be used, e.g. Pam handle or the amide-generating BHA handle.

Preferably, the PEG resin used in the present invention comprises a linker comprising a reactive NH- radical which generates upon cleavage of peptide from resin an acid amide, be it after initial coupling of the C $\alpha$ -carboxylic acid function of the C-terminal amino acid residue of the peptide to such linker or be it from the  $\omega$ -carboxylic acid side chain function of aspartic or glutamic acid, yielding asparagine or glutamine upon cleavage after initial side-chain anchoring and protection of the peptide's C-terminus. In the present context, such linkers are named amide-generating linkers. More preferably, and particularly for synthesis of T20, such amide-generating linker is PAL (Albericio et al., 1987, Int. J. Pept. Protein Research 30, 206-216),

Sieber (Tetrahedron Lett. 1987, 28, 2107-2110) or similar 9-amino-xanthenyl-type resin linker, or Rink amide (4-(2',4'-dimethoxyphenyl-Fmoc-aminomethyl)phenoxy; Rink et al, 1987, Tetrahedron Lett. 28,3787 ff) linker.

- 5 It is also possible that such PEG resin may also be a mixed polyether-polyester or polyether-polyamide resin such as described e.g. in Kempe et al., J. Am. Chem. Soc. 1996, 118, 7083 and in US5,910,554.

Preferably, the PEG resin is a substantially pure PEG resin that is devoid of eventually further substituted polystyrene moieties as a defining structural element and, more preferably, further does also not comprise any internal ester or amide functional groups but only polyether functional groups. Notably, in the latter definition terminal ester or amide bonds bridging the resin directly or via a handle to the peptide are not accounted for, since they are not internal structural, crosslinking features and accordingly do not affect the definition of 'pure PEG' in this regard.

PEG resins offer optimal chemical stability along with good compatibility and ease of handling with standard solvents during normal Fmoc synthesis. According to the present inventions, they allow of synthesizing e.g. T-20 in exceptional yield and purity.

20 A helix-forming segment comprises helix-forming amino acid residues as defined above; helix-forming residues in their meaning in the present context are those amounting to strong helix formers, namely being annotated as H $\alpha$  or h $\alpha$ -type amino acids in the helix propensity annotation and prediction scheme according to Chou and Fasman, Ann. Rev. Biochem. 47, 258 (1978). Such strong helix formers simply have an empirically determined helical propensity of >1 indicating that such strong helix forming amino acid residue occurs with greater than average frequency in an  $\alpha$  Helix in X-ray protein structures. For each of the natural 20 amino acids, an empirical numerical propensity value can be found tabulated in Chou, supra. Strong helix formers accordingly are Ala, Gln, Glu, Ile, Leu, Lys, Met, Phe, Trp and Val. In contrast, His, Arg and Asp occurred with statistical frequency, meaning a propensity of about 1, in  $\alpha$ -helices whilst Asn was found to have a truly negative impact on helix formation, namely a propensity of 0.67 only. For prediction of helical or helix-forming peptide segments, a

comparatively simple but reliable prediction algorithm was defined by Chou et al, 1978.: A cluster of 4 strongly helix forming amino acids residues, with the exceptional rule that two His and/or Asp make up for one strong helix forming residue, within six contiguous amino acid residues will nucleate a helix. Calculating the average helix propensity for every overlapping tetrapeptide segment of such helical segment, such helix may then be predicted to extend to both directions until the averaged propensity for the terminal tetrapeptide segment falls below 1.00 . Prolines are helix-breakers and may only occur at the ends of a helix. Preferably, an accordingly predicted helix-forming segment has an average helix propensity that is larger than a similarly predictable averaged beta-sheet propensity value according to Chou, supra.

Whilst it has often been contemplated in the field of peptide synthesis that beta-sheet structures, leading to aggregation of adjacent peptide chains by inter-chain bonding, may negatively affect coupling reactions during and deprotection reactions after synthesis, intra-chain helix formation has not been perceived as a likely obstacle for linear solid-phase synthesis.

Coupling reactions, coupling reagents employed and cleavage from resin/side chain deprotection are to be conducted with standard solid phase synthesis protocols, employing e.g. Boc or , preferably, Fmoc chemistry. Notably, coupling efficiency after each coupling step should be controlled during synthesis by means of e.g. Ninhydrin test and individual couplings showing unexpected low coupling efficiency should be repeated prior to continuing with further cycle of deprotection and coupling; in particular after critical coupling steps, preemptive capping of still unreacted N $\alpha$ -functions could be favorably carried out prior to the next synthetic cycle.

After on-resin synthesis, in particular in case of T-20, N-terminal acetylation commonly using acetic anhydride or similar modification may be carried out. After usually one-step global cleavage from PEG resin and deprotection where not using special, orthogonal protection groups for side chains, the peptide is harvested and isolated or further processed by customary methods.

Preferably, the peptide to be synthesized is T20. However, other peptides comprising helix-forming segments are also accessible by the present method. Apart from T-20, further examples of similar peptides having such helix-forming segments are overlapping or related, but not identical peptides from said gp41 heptad repeat domains having 15- 40 amino acid

residues and overlapping with T-20 by at least 10 amino acid residues.

Preferably, in addition to the requirement of comprising a helical peptide segment, the total length of the peptide to be synthesized and which peptides comprises afore said helical peptide segment as specified is at least 15 amino acids, more preferably it is at least 24 amino acids, most preferably it is at least 30 amino acids long.

Preferably, the peptide to be synthesized is Thymosin  $\alpha_1$  having the sequence

1-Ser-Asp-Ala-Ala-Val-Asp-Thr-Ser-Ser-Glu-Ile-Thr-Thr-Lys-Asp-Leu-Lys-Glu-Lys-Lys-Glu-Val-Val-Glu-Glu-Ala-Glu-Asn-28

wherein the individual amino acid chains are unprotected or are suitably protected, as need may be for a given individual amino acid residue as is common knowledge in the art (see Bodansky, *infra*, for instance). More preferably, such peptide is N-terminally acetylated in a step subsequent to linear synthesis but prior to cleavage from resin. Natural thymalfasin as is also used as a pharmaceutical is N-terminally acetylated.

Notably, the present inventors have found that it is mainly the C-terminal half of the thymalfasin peptide, it is about residues 19-28, that incur most of the impurities by chain deletions and hence losses in final product yield generated during linear phase synthesis. This part of the peptide has been found to be extremely difficult to synthesize.

Vice versa, during linear synthesis, it is also feasible to use such oxazolidine dipeptides according to Wöhr et al., (Wöhr et al., J. Am. Chem. Soc. 118, 9218).

## EXAMPLES

### Solid-phase synthesis of T-20

General Procedures. ChemMatrix resin from Matrix Innovation (Quebec, Canada), HCTU from Luxembourg Industries Ltd. (Tel Aviv, Israel), Protected Fmoc-amino acid derivatives and Rink handle from IRIS Biotech (Marktredwitz, Germany).

Manual solid-phase synthesis were carried out in a polypropylene syringes (10 mL) fitted with a

polyethylene porous disc. Solvents and soluble reagents were removed by suction. Removal of the Fmoc group was carried out with piperidine-DMF (2:8, v/v) (1 x 2 min, 2 x 10 min). Washings between deprotection, coupling, and, again, deprotection steps were carried out with DMF (5 x 0.5 min) and CH<sub>2</sub>Cl<sub>2</sub> (5 x 0.5 min) using each time 10 mL solvent/g resin. Peptide synthesis transformations and washes were performed at 25 °C.

Automatic solid-phase syntheses were carried out in an ABI 433A peptide synthesizer using a FastMoc program using 0.10 mmol of resin.

*Removal* of the Fmoc was carried out with 22% piperidien in DMF for 2 + 7.6 min. The resin is washed with DCM (6 times for 0.5 min) and DMF (4 times).

*Coupling.* In a cartridge, Fmoc-amino acid (0.9 mmol, 10 equiv) were dissolved in DMF and 0.9 mmol of 0.45 M HCTU in DMF were added to the cartridge. Then, 1 mL of 2 M DIEA in NMP is added and the solution is transferred to the reaction vessel. The coupling takes place for 30 min, where the resin is washed (6 times) with DMF.

HPLC reversed phase columns Symmetry<sup>TM</sup> C<sub>18</sub> 4,6 x 150 mm, 5 µm (column A) and were from Waters (Ireland). Analytical HPLC was carried out on a Waters instrument comprising two solvent delivery pumps (Waters 1525), automatic injector (Waters 717 autosampler), dual wavelength detector (Waters 2487), and system controller (Breeze V3.20). UV detection was at 220 nm, and linear gradients of CH<sub>3</sub>CN (+0.036% TFA) into H<sub>2</sub>O (+0.045% TFA), from 30% to 70% in 15 min.

MALDI-TOF and ES-MS analysis of peptide samples were performed in a PerSeptive Biosystems Voyager DE RP, using ACH matrix. Fig. 1 shows purity and yield of product in a) HPLC chromatogram and b,c) MS spectra. Yield of pure product (RT 7,078 was 50% relative to impurities in HPLC analysis.

#### Example 1

**Ac-Tyr-Thr-Ser-Leu-Ile-His-Ser-Leu-Ile-Glu-Glu-Ser-Gln-Asn-Gln-Gln-Glu-Lys-Asn-Glu-Gln-Glu-Leu-Leu-Glu-Leu-Asp-Lys-Trp-Ala-Ser-Leu-Trp-Asn-Trp-Phe-NH<sub>2</sub> (T-20)**

#### Step 1

***Fmoc-Rink-ChemMatrix-resin***

ChemMatrix-resin (0.1 mmol, 0.45 mmol/g) was placed in the polypropylene syringe. The resin was subjected to the following washings/treatments with CH<sub>2</sub>Cl<sub>2</sub> (3 x 0.5 min), DMF (3 x 0.5 min), and DMF (5 x 0.5 min). Then, Fmoc-Rink handle (3 equiv) and DIEA (6 equiv) in DMF (1 mL) were added, followed by PyBOP (3 equiv) and HOAt (3 equiv) in DMF (5 mL). The mixture was left to stir mechanically for 2 h and the resin was washed with DMF (3 x 0.5 min) and the ninhydrin test was negative.

**Step 2**

***H-Tyr(tBu)-Thr(tBu)-Ser(tBu)-Leu-Ile-His(Trt)-Ser(tBu)-Leu-Ile-Glu(OtBu)-Glu(OtBu)-Ser(tBu)-Gln(Trt)-Asn(Trt)-Gln(Trt)-Gln(Trt)-Glu(OtBu)-Lys(Boc)-Asn(Trt)-Glu(OtBu)-Gln(Trt)-Glu(OtBu)-Leu-Leu-Glu(OtBu)-Leu-Asp(OtBu)-Lys(Boc)-Trp(Boc)-Ala-Ser(tBu)-Leu-Trp(Boc)-Asn(Trt)-Trp(Boc)-Phe-NH-Rink-ChemMatrix-resin***

The Fmoc group was removed and Fmoc-aa-OH's were sequentially added to the above peptidyl-resin (step 1) in the ABI automatic synthesizer 433A as described above.

**Step 3**

***Ac-Tyr(tBu)-Thr(tBu)-Ser(tBu)-Leu-Ile-His(Trt)-Ser(tBu)-Leu-Ile-Glu(OtBu)-Glu(OtBu)-Ser(tBu)-Gln(Trt)-Asn(Trt)-Gln(Trt)-Gln(Trt)-Glu(OtBu)-Lys(Boc)-Asn(Trt)-Glu(OtBu)-Gln(Trt)-Glu(OtBu)-Leu-Leu-Glu(OtBu)-Leu-Asp(OtBu)-Lys(Boc)-Trp(Boc)-Ala-Ser(tBu)-Leu-Trp(Boc)-Asn(Trt)-Trp(Boc)-Phe-NH-Rink-ChemMatrix-resin***

The final acetylation was carried out with a solution of Ac<sub>2</sub>O-DIEA-DMF (10:5:85) in DMF (5 mL) for 15 min with sporadic manual stirring, where ninhydrin was negative.

**Step 4**

*Ac-Tyr-Thr-Ser-Leu-Ile-His-Ser-Leu-Ile-Glu-Glu-Ser-Gln-Asn-Gln-Gln-Glu-Lys-Asn-Glu-Gln-Glu-Leu-Leu-Glu-Leu-Asp-Lys-Trp-Ala-Ser-Leu-Trp-Asn-Trp-Phe-NH<sub>2</sub> (T-20)*

5 The peptide-resin was suspended in TFA-iPr<sub>3</sub>SiH-H<sub>2</sub>O (95:2.5:2.5) and the mixture was allowed to stir for 2 h. Then., ether/hexane was added, the peptide precipitated, the solvent was removed after centrifugation, and extra ether/hexane was added and removed after centrifugation. This operation was repeated several times. Finally, H<sub>2</sub>O (5 mL) was added and lyophilized.

EI-MS, calcd. 4489. Found:  $m/z$  [M+2H]<sup>2+</sup>/2 2248

**Claims**

1. Method of preparing a peptide, characterized in that said peptide which peptide may have individually protected and/or unprotected amino acid side chains is prepared  
5 bound to a solid phase which solid phase is an amphiphilic PEG resin and wherein the peptide comprises a prolin-free, helix-forming segment consisting of a run of six contiguous amino acid residues comprising at least four protected or unprotected helix-forming amino acids residues.
- 10 2. Method according to claim 1, characterized in that the loading capacity of the PEG resin is at least 0.5 mmol/g.
3. Method according to claim1, characterized in that it is an Fmoc synthesis.
- 15 4. Method according to claim 1, characterized in that the helix-forming segment is at least 10, more preferably is at least 12 residues long.
5. Method according to one of the preceding claims, characterized in that the peptide is linked to the solid phase via a functional linker or handle.
- 20 6. Method according to claim 5, characterized in that the linker is an amide-generating linker, preferably is Sieber or Rink amide linker.
7. Method according to claim 1, characterized in that the PEG resin is an amino-alkyl-  
25 functionalized PEG resin to which amino function the peptide is bound forming a C $\alpha$ -terminal peptidyl-carboxamide.
8. Method according to claim 2 or 5, characterized in that the resin is a PEG resin wherein the is resin is not a polystyrene-PEG mixed or co-polymer or block copolymer,  
30 preferably that the resin is a substantially pure PEG resin.
9. Peptide-PEG resin conjugate according to claim 1.

10. Peptide-PEG resin conjugate according to claim 9, characterized in that the peptide moiety has the amino acid sequence is R1- Tyr-Thr-Ser-Leu-Ile-His-Ser-Leu-Ile-Glu-Glu-Ser-Gln-Asn-Gln-Gln-Glu-Lys-Asn-Glu-Gln-Glu-Leu-Leu-Glu-Leu-Asp-Lys-Trp-Ala-Ser-Leu-Trp-Asn-Trp-Phe and wherein R1 is H, acetyl- or a removable protection group, most preferably R1 is acetyl, and wherein the individual amino acid residues may further comprise a side-chain protection group.
11. Method according to claim 1 or 6, characterized in that the peptide has the amino acid sequence R1- Tyr-Thr-Ser-Leu-Ile-His-Ser-Leu-Ile-Glu-Glu-Ser-Gln-Asn-Gln-Gln-Glu-Lys-Asn-Glu-Gln-Glu-Leu-Leu-Glu-Leu-Asp-Lys-Trp-Ala-Ser-Leu-Trp-Asn-Trp-Phe and wherein R1 is H, acetyl- or a removable protection group, and wherein further the peptide is cleaved from the resin and deprotected.
12. Method according to step claim 11, characterized in that the cleavage is a one-step cleavage and deprotection.