ANTIBODIES TO RECEPTOR OF ADVANCED GLYCATION END PRODUCTS (RAGE) AND USES THEREOF

TECHNICAL FIELD

The present application relates to antibodies, particularly monoclonal antibodies, and in particular CDR grafted, humanized versions thereof, that may be used in the treatment and diagnosis of Alzheimer's Disease (AD), central nervous system cell degeneration, impaired learning and memory, abnormal transport of amyloid ß and other neuroinflammatrory conditions associated with the Receptor of Advanced Glycation End Products (RAGE). In particular, the present invention relates to antibodies and fragments thereof that bind to RAGE.

BACKGROUND INFORMATION

Alzheimer's Disease (AD) is the most frequent cause for dementia among the aged, with an incidence of about 10% of the population in those above 65 years of age. With increasing age, the probability of disease also rises. Globally, there are about 15 million people affected with the disease and further increases in life expectancy are expected to increase the number of people affected with the disease to about three-fold over the next decades. In view of the foregoing, there is a tremendous and immediate need for a treatment for AD. With such treatment, affected patients may be able to maintain a functional and active lifestyle for many years beyond that which is not possible without such treatment. Thus, not only are there financial implications for such a treatment but "quality of life" implications as well, for the patients as well as for their caregivers.

From a molecular point of view, AD is characterized by a deposit of abnormally aggregated proteins. In the case of extra-cellular amyloid plaques, these deposits consist mostly of amyloid-β-peptide filaments (Aß), and in the case of the intracellular neurofibrillary tangles (NFTs), mostly of the tau protein. AD is also characterized by an increased neuronal expression of RAGE. RAGE is a multi-ligand receptor of the immunoglobulin family which functions as a signal-transducing cell surface acceptor for Aß.

*A*ß40 infusion in mice has been shown by several groups to lead to vasoconstriction of cerebral vessels and a decrease of cerebral blood flow (CBF). Patients suffering from AD also have a decreased cerebral blood flow. In mouse models of AD where the transgenic animals overexpress the protein Amyloid Precursor Protein (APP) that leads to disease causing plaque formation, RAGE has been implicated as a pathogenic factor in the disease progression (Deane et al. Nature Medicine 9(7) pp 907-913, 2003; Arancio et al. EMBOJ, 1-10, 2004).

RAGE has been shown to bind to Aß-peptides. Inhibition of this interaction suppresses accumulation of Aß in the transgenic animal model; therefore RAGE is believed to be involved in AD. Treatment with sRAGE (soluble RAGE) as well as anti-RAGE antibodies has been shown to lower plaque numbers (Deane et al, 2003). Blocking the interaction of RAGE with amyloid by antibodies could become a treatment for AD patients; however, existing polyclonal antibodies generated from animal serum are not suited for the chronic treatment of humans.

Interaction of RAGE with Aß is disclosed in WO 2006/077101 A1, which describes competition of RAGE lacking the v-domain for the binding of Aß to RAGE, as well as the competition of peptides representing parts of the C-terminal domain of RAGE, mostly the C1-domain. Interaction of anti-RAGE antibodies with the v-domain of RAGE is disclosed in WO2007109749(A2); which also describes that binding of different ligands (S100b, HMGB1 (High Mobility Group Box 1 protein), amyloid aβ) would bind to RAGE *via* binding to this domain.

WO 2008/137552 A2 discloses certain monoclonal anti-RAGE antibodies binding to different domains of RAGE. Most of said antibodies inhibit the interaction of human RAGE and a complex of HMGB1and CpG DNA.

WO 2006/077101 relates to the identification, functionality and use of peptides designated AGER-RME and AGER-CDP of RAGE. Said peptides are inter alia applicable for identifying and preparing RAGE binding ligands like anti-RAGE antibodies.The present invention describes novel monoclonal antibodies that bind to the C-domains of RAGE and the specific interaction and competition with the binding of Aβ with monoclonal antibodies for the C1 and C2-domain in RAGE.

SUMMARY OF THE INVENTION

The present invention provides binding molecules, in particular antibodies, that bind specifically to RAGE; representative anti-RAGE antibodies of the invention

may comprise at least one of the antibody variable region amino acid sequences shown in SEQ ID NOs: 1, 5, 9, 13, 17, and 21, or individual CDRs thereof or related CDR sequences, as specified in more detail below.

Specifically the present invention provides monoclonal antibodies that bind to RAGE, more specifically monoclonal antibodies that bind to the C-domain of RAGE.

Included in the present invention are anti-RAGE antibodies that bind specifically to RAGE and comprise a light chain variable region having an amino acid sequence that is at least 90% identical to any of SEQ ID NOs.: 5, 13, and 21, or is a RAGE-binding fragment of an antibody comprised in said sequences.

Also included are anti-RAGE antibodies that bind specifically to RAGE and comprise a heavy chain variable region having an amino acid sequence that is at least 90% identical to any of SEQ ID NOs.: 1, 9, and 17, or is a RAGE-binding fragment of an antibody comprised in said sequences.

A particular embodiment of the present invention is represented by several monoclonal antibodies that are able to bind to the C-domain of RAGE, and to block the binding of Aß -globulomers. More specifically the present invention describes monoclonal antibody 11E6, which binds to the C-2 domain of RAGE, does not bind to peptides with amino acid sequences used to generate polyclonal antibodies, and is able to neutralize *in vivo* the effect of Aß1-40 on cerebral vasculature in mice.

The anti-RAGE antibodies of the invention include antibodies that bind specifically to the C-domain of RAGE.

The anti-RAGE antibodies of the invention include an anti-RAGE antibody or a RAGE-binding fragment as described above, which is selected from the group consisting of chimeric antibody, a CDR-grafted or humanized antibody, a single chain antibody, a fusion protein, and a human antibody.

In various embodiments, the antibodies of the invention are recombinant antibodies or monoclonal antibodies. Particular neutralizing antibodies of the present application are referred to herein as mAb7F9, mAb11E6, and mAb4E5 and functional antibody fragments thereof, and other antibodies and functional antibody fragments with equivalent properties to mAb7F9, mAb11E6, and mAb4E5, such as high affinity binding to RAGE with low dissociation kinetics and high neutralizing capacity, are intended as part of the present invention. The human antibodies of the present application, however, may include amino acid residues not encoded by human germline immunoglobulin immunogenic RAGE polypeptide or fragment thereof, that

may be determined by any method known in the art. For example, the binding affinity can be measured by competitive ELISAs, c RIAs, BIAcore or KinExA technology. The dissociation rate also can be measured by BIAcore or KinExA technology. The binding affinity and dissociation rate are measured by surface plasmon resonance using, e.g., BIAcore.

One of the monoclonal antibodies of the present application, the mAb7F9 antibody, has at least 90% amino acid sequence identity with a sequence comprising a heavy chain variable region (VH region) comprising the sequence of SEQ ID NO: 1; and SEQ ID NOs. 2, 3, and 4 which are residues 31-35, 50-68, and 101-108 of SEQ ID NO: 1, respectively. The mAb7F9 antibody of the present invention has at least 90% amino acid sequence identity with a sequence comprising a light chain variable region (VL region) comprising the sequence of SEQ ID NO: 5, and SEQ ID NOs. 6, 7, and 8 which are residues 24-34, 50-56, 89-97 of SEQ ID NO: 5, respectively.

Another of the monoclonal antibodies of the present application, the mAb11E6 antibody, has at least 90% amino acid sequence identity with a sequence comprising a heavy chain variable region (VH region) comprising the sequence of SEQ ID NO: 9; and SEQ ID NOs. 10, 11, and 12 which are residues 31-35, 50-66, and 99-109 of SEQ ID NO: 9, respectively. The mAb11E6 antibody of the present invention has at least 90% amino acid sequence identity with a sequence comprising a light chain variable region (VL region) comprising the sequence of SEQ ID NO: 13, and SEQ ID NOs. 14, 15, and 16 which are residues 24-34, 50-56, 89-97 of SEQ ID NO: 13, respectively. The mAb11E6 binds to the C-2 domain of RAGE, does not bind to peptides with amino acid sequences used to generate polyclonal antibodies, and is able to neutralize *in vivo* the effect of Aß1-40 on cerebral vasculature in mice.

Another of the monoclonal antibodies of the present application, the mAb4E5 antibody, has at least 90% amino acid sequence identity with a sequence comprising a heavy chain variable region (VH region) comprising the sequence of SEQ ID NO: 17; and SEQ ID NOs. 18, 19, and 20 which are residues 31-35, 50-66, and 99-109 of SEQ ID NO: 17, respectively. The mAb4E5 antibody of the present invention has at least 90% amino acid sequence identity with a sequence comprising a light chain variable region (VL region) comprising the sequence of SEQ ID NO: 21, and SEQ ID NOs. 22, 23, and 24 which are residues 24-34, 50-56, 89-97 of SEQ ID NO: 21, respectively.

It is also intended that the isolated monoclonal antibodies that interact with RAGE of the present application may be a glycosylated binding protein wherein the antibody or antigen-binding portion thereof comprises one or more carbohydrate residues. Nascent *in vivo* protein production may undergo further processing, known as post-translational modification. In particular, sugar (glycosyl) residues may be added enzymatically, a process known as glycosylation. The resulting proteins bearing covalently linked oligosaccharide side chains are known as glycosylated proteins or glycoproteins. Protein glycosylation depends on the amino acid sequence of the protein of interest, as well as the host cell in which the protein is expressed. Different organisms may produce different glycosylation enzymes (eg., glycosyltransferases and glycosidases), and have different substrates (nucleotide sugars) available. Due to such factors, protein glycosylation pattern, and composition of glycosyl residues, may differ depending on the host system in which the particular protein is expressed. Glycosyl residues useful in the invention may include, but are not limited to, glucose, galactose, mannose, fucose, n-acetylglucosamine and sialic acid.

The antibodies of the present application comprise a heavy chain constant region, such as an IgG1, IgG2, IgG3, IgG4, IgA, IgE, IgM or IgD constant region. Furthermore, the antibody can comprise a light chain constant region, either a kappa light chain constant region or a lambda light chain constant region. Particularly, the antibody comprises a kappa light chain constant region. Alternatively, the antibody portion can be, for example, a Fab fragment or a single chain Fv fragment. Replacements of amino acid residues in the Fc portion to alter antibody effector function are known in the art (Winter, et al. U.S. Pat. Nos. 5,648,260; 5,624,821). The Fc portion of an antibody mediates several important effector functions. e.g. cytokine induction, ADCC, phagocytosis, complement dependent cytotoxicity (CDC) and half-life/clearance rate of antibody and antigen-antibody complexes. In some cases these effector functions are desirable for therapeutic antibodies but in other cases might be unnecessary or even deleterious, depending on the therapeutic objectives. Certain human IgG isotypes, particularly IgG1 and IgG3, mediate ADCC and CDC via binding to Fcγ Rs and complement C1q, respectively. Neonatal Fc receptors (FcRn) are the critical components determining the circulating half-life of antibodies. In still another embodiment at least one amino acid residue is replaced in the constant region

of the antibody, for example the Fc region of the antibody, such that effector functions of the antibody are altered.

BRIEF DESCRIPTION OF THE FIGURES

Figures 1A -1C show ELISA binding of recombinant and hybridoma-derived anti-human RAGE monoclonal antibodies 7F9, 11E6, and 4E5 to recombinant human RAGE.

Figure 2 shows characterization of monoclonal antibodies 7F9, 11E6, and 4E5 by dot blot binding.

Figures 3A - 3C show HTRF assay results showing sRAGE - Aβ-globulomers – monoclonal antibodies competition.

Figure 4 shows HTRF sRAGE - Aβ-globulomer binding.

Figure 5 shows binding of Aβ-globulomer to sRAGE-Fc and to RAGE v-domain.

Figures 6A - 6C show ELISA binding experiments of monoclonal antibodies of the invention to different RAGE fragments.

Figures 7A – 7C show ELISA binding experiments of monoclonal antibodies of the invention to different RAGE fragments.

Figures 8a – 8f show changes of Cerebral Blood Flow induced by different doses of Aß1-40.

Figures 9a – 9c show effects of 11E6 in Aß1-40-induced changes in Cerebral Blood Flow.

Figures 10 show changes of Cerebral Blood Flow observed in Aged Tg2576 mice treated with different doses of 11E6 or control antibody.

Figure 11 shows that antibody 11E6 protects hippocampal neurons against Aß induced dynamin cleavage. Upper panel: Samples representing repeated experimental conditions from a single experiment are shown and treatment concentrations (in μM) are indicated above. Cells without addition of Aß show mostly intact (~100 kDa) dynamin I signals (1st bar), which is decreased and reverted to a ~90kDa cleavage product in cells treated with 5μM Aß (2nd bar). Antibody 11E6 treatment (3rd bar) prevents the cleavage, whereas an Ig1 control antibody (4th bar) is without a pronounced protective effect. Lower panel: Quantification of the dynamin signal of three independent experiments (expressed as % dynamin +/- SEM after normalization

to 0μM Aß treatment) revealed a statistically significant protective effect of 11E6 (One-way ANOVA, Kruskal Wallis test followed by Dunns test; * indicates p<0,05)

Figure 12 shows the influence of 11E6 (A) or Control antibody (B) on the Globulomer-induced strong suppression of synaptic transmission in rat hippocampal slice culture. 0.1 μM 11E6 completely reversed the globulomer-induced deficits (see (A)).

Figure 13 shows the effect of 12 week treatment with antibody 11E6 on amyloid plaque deposits in Tg2576 mice. Area covered with plaques (A,B) and number of plaques (C,D) as detected by labelling with the anti Aβ antibody 6G1 after 11E6 or IgG1 control antibody treatment (n=19/group) is shown. Treatment with 11E6 reduced area covered by and number of deposits in the neocortex by 24.5% (A) and 26.8% (C), respectively. Statistical analysis revealed a strong trend (asterisks in brackets, p<0.06; Mann-Whitney U-test). The reduction was statistically significant in the frontal cortex (asterisks, p<0.05; Mann-Whitney U-test) where the area of deposits was reduced by 23.5% (B) and their number by 26.8% (D) after 11E6 treatment.

LIST OF SEQUENCES

SEQ ID NO: 1: amino acid sequence of mAb VH 7F9
SEQ ID NO: 2: amino acid sequence of mAb VH 7F9 CDR-H1
SEQ ID NO: 3: amino acid sequence of mAb VH 7F9 CDR-H2
SEQ ID NO: 4: amino acid sequence of mAb VH 7F9 CDR-H3
SEQ ID NO: 5: amino acid sequence of mAb VL 7F9
SEQ ID NO: 6: amino acid sequence of mAb VL 7F9 CDR-L1
SEQ ID NO: 7: amino acid sequence of mAb VL 7F9 CDR-L2
SEQ ID NO: 8: amino acid sequence of mAb VL 7F9 CDR-L3
SEQ ID NO: 9: amino acid sequence of mAb VH 11E6
SEQ ID NO: 10: amino acid sequence of mAb VH 11E6 CDR-H1
SEQ ID NO: 11: amino acid sequence of mAb VH 11E6 CDR-H2
SEQ ID NO: 12: amino acid sequence of mAb VH 11E6 CDR-H3
SEQ ID NO: 13: amino acid sequence of mAb VL 11E6
SEQ ID NO: 14: amino acid sequence of mAb VL 11E6 CDR-L1
SEQ ID NO: 15: amino acid sequence of mAb VL 11E6 CDR-L2
SEQ ID NO: 16: amino acid sequence of mAb VL 11E6 CDR-L3
SEQ ID NO: 17: amino acid sequence of mAb VH 4E5

SEQ ID NO: 18: amino acid sequence of mAb VH 4E5 CDR-H1

SEQ ID NO: 19: amino acid sequence of mAb VH 4E5 CDR-H2

SEQ ID NO: 20: amino acid sequence of mAb VH 4E5 CDR-H3

SEQ ID NO: 21: amino acid sequence of mAb VL 4E5

SEQ ID NO: 22: amino acid sequence of mAb VL 4E5 CDR-L1

SEQ ID NO: 23: amino acid sequence of mAb VL 4E5 CDR-L2

SEQ ID NO: 24: amino acid sequence of mAb VL 4E5 CDR-L3

SEQ ID NO: 25: amino acid sequence of human Ig gamma1 heavy chain constant region

SEQ ID NO: 26: amino acid sequence of human Ig kappa light chain constant region

SEQ ID NO: 27: Full plasmid nucleotide sequence of Construct #1 (bold) encoding OmpA-[RAGE (23-340)]-6His

SEQ ID NO: 28: Full plasmid nucleotide sequence of Construct # 2 (bold) encoding 6His-(Thr)-[RAGE (24-129)]

SEQ ID NO: 29: Full plasmid nucleotide sequence of Construct # 3 (bold) encoding 6His-(Thr)-[RAGE (24-234)]

SEQ ID NO: 30: Full plasmid nucleotide sequence of Construct # 4 (bold) encoding 6His-(Thr)-[RAGE (24-336)]

SEQ ID NO: 31: Full plasmid nucleotide sequence of Construct # 5 (bold) encoding 6His-(Thr)-[RAGE (130-234)]

SEQ ID NO: 32: Full plasmid nucleotide sequence of Construct # 6 (bold) encoding 6His-(Thr)-[RAGE (130-336)]

SEQ ID NO: 33: Full plasmid nucleotide sequence of Construct # 7 (bold) encoding 6His-(Thr)-[RAGE (235-336)]

SEQ ID NO: 34: encoded amino acid sequence RAGE protein # 1

SEQ ID NO: 35: encoded amino acid sequence RAGE protein # 2

SEQ ID NO: 36: encoded amino acid sequence RAGE protein # 3

SEQ ID NO: 37: encoded amino acid sequence RAGE protein # 4

SEQ ID NO: 38: encoded amino acid sequence RAGE protein # 5

SEQ ID NO: 39: encoded amino acid sequence RAGE protein # 6

SEQ ID NO: 40: encoded amino acid sequence RAGE protein # 7

SEQ ID NO:41: Ig gamma-1 constant region mutant amino acid sequence

SEQ ID NO:42: Ig gamma-2 constant region amino acid sequence

SEQ ID NO:43: framework amino acid sequence VH7-4.1/JH6 FR1

SEQ ID NO:44: framework amino acid sequence VH7-4.1/JH6 FR2 and VH1-2/JH6 FR2

SEQ ID NO:45: framework amino acid sequence VH7-4.1/JH6 FR3

SEQ ID NO:46: framework amino acid sequence VH7-4.1/JH6 FR4 and VH1-2/JH6 FR4

SEQ ID NO:47: framework amino acid sequence VH1-2/JH6 FR1

SEQ ID NO:48: framework amino acid sequence VH1-2/JH6 FR3

SEQ ID NO:49: framework amino acid sequence 1-12/L5/JK2 FR1

SEQ ID NO:50: framework amino acid sequence 1-12/L5/JK2 FR2

SEQ ID NO:51: framework amino acid sequence 1-12/L5/JK2 FR3

SEQ ID NO:52: framework amino acid sequence 1-12/L5/JK2 FR4 and 3-15/L2/JK2 FR4

SEQ ID NO:53: framework amino acid sequence 3-15/L2/JK2 FR1

SEQ ID NO:54: framework amino acid sequence 3-15/L2/JK2 FR2

SEQ ID NO:55: framework amino acid sequence 3-15/L2/JK2 FR3

SEQ ID NO:56: CDR-grafted amino acid sequence VH 11E6.1-GL

SEQ ID NO:57: CDR-grafted amino acid sequence VH 11E6.2-GL

SEQ ID NO:58: CDR-grafted amino acid sequence VL 11E6.1-GL

SEQ ID NO:59: CDR-grafted amino acid sequence VL 11E6.2-GL

SEQ ID NO: 60: amino acid sequence of hRAGE

SEQ ID NO: 61: amino acid sequence of a husRAGE fragment

SEQ ID NO: 62: humanized antibody sequence VH h11E6.1

SEQ ID NO: 63: humanized antibody sequence VL h11E6.1

SEQ ID NO: 64: humanized antibody sequence VL h11E6.2

SEQ ID NO: 65: humanized antibody sequence VL h11E6.3

SEQ ID NO: 66: humanized antibody sequence VL h11E6.4

SEQ ID NO: 67: humanized antibody sequence VH h11E6.5

SEQ ID NO: 68: humanized antibody sequence VH h11E6.9

SEQ ID NO: 69: humanized antibody sequence VH h11E6.16

SEQ ID NO:70: amino acid sequence of RAGE-derived peptide NtermR31

SEQ ID NO:71: amino acid sequence of RAGE-derived peptide 1

SEQ ID NO:72: amino acid sequence of RAGE-derived peptide 2

SEQ ID NO:73: amino acid sequence of RAGE-derived peptide 3

SEQ ID NO:74: amino acid sequence of RAGE-derived peptide 4

SEQ ID NO:75: amino acid sequence of RAGE-derived peptide 5
SEQ ID NO:76: amino acid sequence of RAGE-derived peptide 6
SEQ ID NO:77: amino acid sequence of RAGE-derived peptide 7
SEQ ID NO:78: amino acid sequence of RAGE-derived peptide 8
SEQ ID NO:79: amino acid sequence of RAGE-derived peptide 9
SEQ ID NO:80: amino acid sequence of RAGE-derived peptide 10
SEQ ID NO:81: nucleotide sequence of oligonucleotide primers
SEQ ID NO:82: nucleotide sequence of oligonucleotide primers
SEQ ID NO:83: nucleotide sequence of oligonucleotide primers
SEQ ID NO:84: nucleotide sequence of oligonucleotide primers
SEQ ID NO:85: nucleotide sequence of oligonucleotide primers
SEQ ID NO:86: nucleotide sequence of oligonucleotide primers
SEQ ID NO:87: nucleotide sequence of oligonucleotide primers
SEQ ID NO:88: nucleotide sequence of oligonucleotide primers
SEQ ID NO:89: nucleotide sequence of oligonucleotide primers
SEQ ID NO:90: nucleotide sequence of oligonucleotide primers
SEQ ID NO:91: nucleotide sequence of oligonucleotide primers
SEQ ID NO:92: nucleotide sequence of oligonucleotide primers
SEQ ID NO:93: nucleotide sequence of oligonucleotide primers
SEQ ID NO:94: nucleotide sequence of oligonucleotide primers

DETAILED DESCRIPTION

1. General definitions

Unless otherwise defined herein, scientific and technical terms used in connection with the present invention shall have the meanings that are commonly understood by those of ordinary skill in the art. The meaning and scope of the terms should be clear, however, in the event of any latent ambiguity, definitions provided herein take precedent over any dictionary or extrinsic definition. Further, unless otherwise required by context, singular terms shall include pluralities and plural terms shall include the singular. In this application, the use of "or" means "and/or" unless stated otherwise. Furthermore, the use of the term "including", as well as other forms, such as "includes" and "included", is not limiting. Also, terms such as "element" or "component" encompass both elements and components comprising one

unit and elements and components that comprise more than one subunit unless specifically stated otherwise.

Generally, nomenclatures used in connection with, and techniques of, cell and tissue culture, molecular biology, immunology, microbiology, genetics and protein and nucleic acid chemistry and hybridization described herein are those well known and commonly used in the art. The methods and techniques of the present invention are generally performed according to conventional methods well known in the art and as described in various general and more specific references that are cited and discussed throughout the present specification unless otherwise indicated. Enzymatic reactions and purification techniques are performed according to manufacturer's specifications, as commonly accomplished in the art or as described herein. The nomenclatures used in connection with, and the laboratory procedures and techniques of, analytical chemistry, synthetic organic chemistry, and medicinal and pharmaceutical chemistry described herein are those well known and commonly used in the art. Standard techniques are used for chemical syntheses, chemical analyses, pharmaceutical preparation, formulation, and delivery, and treatment of patients.

That the present invention may be more readily understood, selected terms are defined below.

The term "polypeptide" as used herein, refers to any polymeric chain of amino acids. The terms "peptide" and "protein" are used interchangeably with the term polypeptide and also refer to a polymeric chain of amino acids. The term "polypeptide" encompasses native or artificial proteins, protein fragments and polypeptide analogs of a protein sequence. A polypeptide may be monomeric or polymeric.

The term "isolated protein" or "isolated polypeptide" is a protein or polypeptide that by virtue of its origin or source of derivation is not associated with naturally associated components that accompany it in its native state; is substantially free of other proteins from the same species; is expressed by a cell from a different species; or does not occur in nature. Thus, a polypeptide that is chemically synthesized or synthesized in a cellular system different from the cell from which it naturally originates will be "isolated" from its naturally associated components. A protein may also be rendered substantially free of naturally associated components by isolation, using protein purification techniques well known in the art.

The term "recovering" as used herein, refers to the process of rendering a chemical species such as a polypeptide substantially free of naturally associated components by isolation, e.g., using protein purification techniques well known in the art.

The term "Receptor for Advanced Glycation Endproducts (RAGE)" designates a multiligand receptor in the immunoglobulin family, which binds soluble Aß peptide, S100b, and HMGB1 (also known as amphoterin) among others. RAGE mediates patho-physiologically relevant cellular changes in response to binding to these ligands. Transgenic animals overexpressing RAGE and human APP display early abnormalities in spatial learning and memory, indicating that RAGE is a cofactor for Aß-induced neuronal perturbation in Alzheimer-type pathologies, and suggesting that RAGE is a potential therapeutic target to ameliorate cellular dysfunction.

The structure of RAGE has not been solved. Homology to other proteins leads to a model where RAGE has several domains. These domains are named in analogy to immunoglobulins: (i) V-like domain at the N-terminus: this equivalent domain in immunoglobulins binds antigen and represents the only binding region within these proteins. In RAGE, this domain binds to some ligands like S100 (Ostendorp et al. EMBOJ. 26,3875,2007; Leclerc et al. JBC 282,31317,2007). A monoclonal antibody binding to the v-like domain in RAGE competes with binding of different ligands S100b, HMGB1, and amyloid β (WO2007109749(A2)) implying that these ligands would also bind to RAGE via this same domain. (ii) First C2-like domain: Two domains within RAGE have homology to the C2 domains of immunoglobulins; one of these domains has been called C1 (nomenclature also used by Ostendorp et al. BBRC 2006). Several ligands binding to this domain have been described, for instance, S100A12 (also called ENRAGE or Calgranulin C), which binds with an affinity of Kd = 90 nM (Hofmann et al. Cell 97, 889, 1999; Xie et al. JBC, 282, 4218, 2007). Aβ competes with S100A12 for this domain, suggesting that Aβ also binds to the C1-domain. (iii) Second C2-like domain: this domain is called C2 (also used by Ostendorp et al. BBRC 2006). RAGE ligand S100A6 binds to the C2 domain and an antibody generated against a peptide from the C2 domain was shown to compete againt S100A6 for this binding; the antibody lead to reduced signal transduction in SH-SY5Y (Leclerc et al. JBC 282,31317,2007).

A "RAGE domain" may be defined in line with different definitions provided in the state of the art:

According to Xie et al. 2007, J. Biol. Chem., Vol. 282:4218-4231 the following definition of h RAGE domains applies:

- the V domain (amino acids 24-129 of SEQ ID NO: 60),
- the C1 domain (amino acids 130-234 of SEQ ID NO:60),
- the C2 domain (amino acids 235-336 of SEQ ID NO:60),

According to a more recent definition by Hudson et al, The FASEB Journal. 2008;22:1572-1580, h RAGE (404 amino acid residues according to SEQ ID NO:60) has an extracellular region (amino acids 1–342 of SEQ ID NO:60) composed of

- a signal peptide (amino acids 1–22 of SEQ ID NO:60), followed by three immunoglobulin-like domains, including
- an Ig-like V-type domain (amino acids 23–116 of SEQ ID NO:60) and
- two Ig-like C2-type 1/2 domains (amino acids 124–221 of SEQ ID NO:60; also designated C1 domain; and amino acids 227–317 of SEQ ID NO:60; also designated C2 domain);
- a single transmembrane domain (amino acids 343–363 of SEQ ID NO:60), and
- a short cytoplasmic tail (amino acids 364–404 of SEQ ID NO:60).

In view of the high degree of identity, in particular with respect to the definition of domains V, C1 and C2, and unless otherwise stated, each of the definitions may be applied in order to define the binding characteristics of the binding proteins of the present invention.

As indicated above, RAGE is capable of binding different ligands via different domains. Results from competition experiments with other ligands seem to indicate that Aβ binds to the C1-domain (Hofmann et al. Cell 97, 889, 1999 or Xie et al. JBC, 282, 4218, 2007). As non-limiting examples of RAGE ligands there may be mentioned:

- Advanced glycation end products (AGEs), (Baynes J.W., 1991, Diabetes. 1991, 40:405-412; Ahmed K.A., 2007, J Clin Biochem Nutr. 41 (2):97-105);
- Members of the S100/calgranulin family (e.g., calgranulin C (also known as ENRAGE and S100A12), S100A1, S100A4, S100A11, S100A13, S100B, and S100P);
- Amyloid-ß-peptide (Aß), as for example Aß 1-40 peptide

- Amyloid-ß globulomers; as for example Aß1-42, Aß12-42, Aß20-42 globulomers (see Barghorn et al., Globular amyloid ß peptide1-42 oligomer - a homogenous and stable neuropathological protein in Alzheimer's disease) Journal of Neurochemistry. 95(3):834-847, November 2005; WO2007/062852; WO2008/150949; all incorporated by reference);
- eukocyte integrins (e.g., Mac-1)

The term "RAGE" as used herein, particularly refers to human RAGE, also designated "hRAGE" or "huRAGE". Unless otherwise stated the term "RAGE" also encompasses RAGE molecules isolated or obtained from other, different from human, species, as for example, rodents, like mice or rats; or bovine RAGE molecules.

The term "sRAGE" refers to a soluble form of RAGE, derived from the extra cellular domain of RAGE. For example, a sRAGE molecule derived from human RAGE, also designated as husRAGE comprises amino acid residues 1 to 331 (see SEQ ID NO: 61) of human RAGE (see SEQ ID NO: 60).

"Biological activity " as used herein, refers to all inherent biological properties of RAGE as defined herein.

The terms "specific binding" or "specifically binding", as used herein, in reference to the interaction of an antibody, a protein, or a peptide with a second chemical species, mean that the interaction is dependent upon the presence of a particular structure (e.g., an "antigenic determinant" or "epitope" as defined below) on the chemical species; for example, an antibody recognizes and binds to a specific protein structure rather than to proteins generally. If an antibody is specific for epitope "A", the presence of a molecule containing epitope A (or free, unlabeled A), in a reaction containing labeled "A" and the antibody, will reduce the amount of labeled A bound to the antibody.

The term "antibody", as used herein, broadly refers to any immunoglobulin (Ig) molecule comprised of four polypeptide chains, two heavy (H) chains and two light (L) chains, or any functional fragment, mutant, variant, or derivation thereof, which retains the essential epitope binding features of an Ig molecule. Such mutant, variant, or derivative antibody formats are known in the art. Nonlimiting embodiments of which are discussed below. An antibody is said to be "capable of binding" a molecule if it is capable of specifically reacting with the molecule to thereby bind the molecule to the antibody.

A "monoclonal antibody" as used herein is intended to refer to a preparation of antibody molecules, which share a common heavy chain and common light chain amino acid sequence, in contrast with "polyclonal" antibody preparations that contain a mixture of different antibodies. Monoclonal antibodies can be generated by several novel technologies like phage, bacteria, yeast or ribosomal display, as well as classical methods exemplified by hybridoma-derived antibodies (e.g., an antibody secreted by a hybridoma prepared by hybridoma technology, such as the standard Kohler and Milstein hybridoma methodology ((1975) Nature 256:495-497).

In a full-length antibody, each heavy chain is comprised of a heavy chain variable region (abbreviated herein as HCVR or VH) and a heavy chain constant region. The heavy chain constant region is comprised of three domains, CH1, CH2 and CH3. Each light chain is comprised of a light chain variable region (abbreviated herein as LCVR or VL) and a light chain constant region. The light chain constant region is comprised of one domain, CL. The VH and VL regions can be further subdivided into regions of hypervariability, termed complementarity determining regions (CDR), interspersed with regions that are more conserved, termed framework regions (FR). Each VH and VL is composed of three CDRs and four FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1, CDR1, FR2, CDR2, FR3, CDR3, FR4. Immunoglobulin molecules can be of any type (e.g., IgG, IgE, IgM, IgD, IgA and IgY), class (e.g., IgG 1, IgG2, IgG 3, IgG 4, IgA1 and IgA2) or subclass.

The term "antigen-binding portion" or "antigen-binding fragment"of an antibody (or simply "antibody portion" or "antibody fragment"), as used herein, refers to one or more fragments of an antibody that retain the ability to specifically bind to an antigen (*e.g.*, RAGE). It has been shown that the antigen-binding function of an antibody can be performed by fragments of a full-length antibody. Such antibody embodiments may also be bispecific, dual specific, or multi-specific formats; specifically binding to two or more different antigens. Examples of binding fragments encompassed within the term "antigen-binding portion" of an antibody include (i) a Fab fragment, a monovalent fragment consisting of the VL, VH, CL and CH1 domains; (ii) a $F(ab')_2$ fragment, a bivalent fragment comprising two Fab fragments linked by a disulfide bridge at the hinge region; (iii) a Fd fragment consisting of the VH and CH1 domains; (iv) a Fv fragment consisting of the VL and

VH domains of a single arm of an antibody, (v) a dAb fragment (Ward *et al.*, (1989) *Nature* 341:544-546, Winter et al., PCT publication WO 90/05144 A1 herein incorporated by reference), which comprises a single variable domain; and (vi) an isolated complementarity determining region (CDR). Furthermore, although the two domains of the Fv fragment, VL and VH, are coded for by separate genes, they can be joined, using recombinant methods, by a synthetic linker that enables them to be made as a single protein chain in which the VL and VH regions pair to form monovalent molecules (known as single chain Fv (scFv); see *e.g.*, Bird *et al.* (1988) *Science* 242:423-426; and Huston *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:5879-5883). Such single chain antibodies are also intended to be encompassed within the term "antigen-binding portion" of an antibody. Other forms of single chain antibodies, such as diabodies are also encompassed. Diabodies are bivalent, bispecific antibodies in which VH and VL domains are expressed on a single polypeptide chain, but using a linker that is too short to allow for pairing between the two domains on the same chain, thereby forcing the domains to pair with complementary domains of another chain and creating two antigen binding sites (see *e.g.*, Holliger, P., *et al.* (1993) *Proc. Natl. Acad. Sci. USA* 90:6444-6448; Poljak, R.J., *et al.* (1994) *Structure* 2:1121-1123). Such antibody binding portions are known in the art (Kontermann and Dubel eds., Antibody Engineering (2001) Springer-Verlag. New York. 790 pp. (ISBN 3-540-41354-5).

The term "antibody construct" as used herein refers to a polypeptide comprising one or more the antigen binding portions of the invention linked to a linker polypeptide or an immunoglobulin constant domain. Linker polypeptides comprise two or more amino acid residues joined by peptide bonds and are used to link one or more antigen binding portions. Such linker polypeptides are well known in the art (see *e.g.*, Holliger, P., *et al.* (1993) *Proc. Natl. Acad. Sci. USA* 90:6444-6448; Poljak, R.J., *et al.* (1994) *Structure* 2:1121-1123). An immunoglobulin constant domain refers to a heavy or light chain constant domain. Human IgG heavy chain and light chain constant domain amino acid sequences are known in the art and represented in Table 1.

Table 1: Sequence of human IgG heavy chain constant domain and light chain constant domain

Protein	Sequence Identifier	Sequence
		123456789012345678901234567890
Ig gamma-1 constant region	SEQ ID NO.:25	ASTKGPSVFPLAPSSKSTSGGTAALGCLVK DYFPEPVTVSWNSGALTSGVHTFPAVLQSS GLYSLSSVVTVPSSSLGTQTYICNVNHKPS NTKVDKKVEPKSCDKTHTCPPCPAPELLGG PSVFLFPPKPKDTLMISRTPEVTCVVVDVS HEDPEVKFNWYVDGVEVHNAKTKPREEQYN STYRVVSVLTVLHQDWLNGKEYKCKVSNKA LPAPIEKTISKAKGQPREPQVYTLPPSREE MTKNQVSLTCLVKGFYPSDIAVEWESNGQP ENNYKTTPPVLDSDGSFFLYSKLTVDKSRW QQGNVFSCSVMHEALHNHYTQKSLSLSPGK
Ig gamma-1 constant region mutant	SEQ ID NO.:41	ASTKGPSVFPLAPSSKSTSGGTAALGCLVK DYFPEPVTVSWNSGALTSGVHTFPAVLQSS GLYSLSSVVTVPSSSLGTQTYICNVNHKPS NTKVDKKVEPKSCDKTHTCPPCPAPEAAGG PSVFLFPPKPKDTLMISRTPEVTCVVVDVS HEDPEVKFNWYVDGVEVHNAKTKPREEQYN STYRVVSVLTVLHQDWLNGKEYKCKVSNKA LPAPIEKTISKAKGQPREPQVYTLPPSREE MTKNQVSLTCLVKGFYPSDIAVEWESNGQP ENNYKTTPPVLDSDGSFFLYSKLTVDKSRW QQGNVFSCSVMHEALHNHYTQKSLSLSPGK
Ig gamma-2 constant region	SEQ ID NO.:42	ASTKGPSVFPLAPCSRSTSESTAALGCLVK DYFPEPVTVSWNSGALTSGVHTFPAVLQSS GLYSLSSVVTVPSSNFGTQTYTCNVDHKPS NTKVDKTVERKCCVECPPCPAPPVAGPSVF LFPPKPKDTLMISRTPEVTCVVVDVSHEDP EVQFNWYVDGVEVHNAKTKPREEQFNSTFR VVSVLTVVHQDWLNGKEYKCKVSNKGLPAP IEKTISKTKGQPREPQVYTLPPSREEMTKN QVSLTCLVKGFYPSDIAVEWESNGQPENNY KTTPPMLDSDGSFFLYSKLTVDKSRWQQGN VFSCSVMHEALHNHYTQKSLSLSPGK
Ig Kappa constant region	SEQ ID NO.:26	TVAAPSVFIFPPSDEQLKSGTASVVCLLNN FYPREAKVQWKVDNALQSGNSQESVTEQDS KDSTYSLSSTLTLSKADYEKHKVYACEVTH QGLSSPVTKSFNRGEC

Still further, an antibody or antigen-binding portion thereof may be part of a larger immunoadhesion molecules, formed by covalent or noncovalent association of the antibody or antibody portion with one or more other proteins or peptides. Examples of such immunoadhesion molecules include use of the streptavidin core region to make a tetrameric scFv molecule (Kipriyanov, S.M., *et al.* (1995) *Human Antibodies and Hybridomas* 6:93-101) and use of a cysteine residue, a marker peptide

and a C-terminal polyhistidine tag to make bivalent and biotinylated scFv molecules (Kipriyanov, S.M., *et al.* (1994) *Mol. Immunol.* 31:1047-1058). Antibody portions, such as Fab and $F(ab')_2$ fragments, can be prepared from whole antibodies using conventional techniques, such as papain or pepsin digestion, respectively, of whole antibodies. Moreover, antibodies, antibody portions and immunoadhesion molecules can be obtained using standard recombinant DNA techniques, as described herein.

An "isolated antibody", as used herein, is intended to refer to an antibody that is substantially free of other antibodies having different antigenic specificities (*e.g.*, an isolated antibody that specifically binds human RAGE is substantially free of antibodies that specifically bind antigens other than human RAGE). An isolated antibody that specifically binds human RAGE may, however, have cross-reactivity to other antigens, such as RAGE molecules from other species. Moreover, an isolated antibody may be substantially free of other cellular material and/or chemicals.

The term "human antibody", as used herein, is intended to include antibodies having variable and constant regions derived from human germline immunoglobulin sequences. The human antibodies of the invention may include amino acid residues not encoded by human germline immunoglobulin sequences (e.g., mutations introduced by random or site-specific mutagenesis in vitro or by somatic mutation in vivo), for example in the CDRs and in particular CDR3. However, the term "human antibody", as used herein, is not intended to include antibodies in which CDR sequences derived from the germline of another mammalian species, such as a mouse, have been grafted onto human framework sequences.

The term "recombinant human antibody", as used herein, is intended to include all human antibodies that are prepared, expressed, created or isolated by recombinant means, such as antibodies expressed using a recombinant expression vector transfected into a host cell (described further below), antibodies isolated from a recombinant, combinatorial human antibody library (Hoogenboom H.R., (1997) *TIB Tech.* 15:62-70; Azzazy H., and Highsmith W.E., (2002) *Clin. Biochem.* 35:425-445; Gavilondo J.V., and Larrick J.W. (2002) *BioTechniques* 29:128-145; Hoogenboom H., and Chames P. (2000) *Immunology Today* 21:371-378), antibodies isolated from an animal (e.g., a mouse) that is transgenic for human immunoglobulin genes (see e.g., Taylor, L. D., et al. (1992) Nucl. Acids Res. 20:6287-6295; Kellermann S-A., and Green L.L. (2002) *Current Opinion in Biotechnology* 13:593-597; Little M. et al

(2000) *Immunology Today* 21:364-370) or antibodies prepared, expressed, created or isolated by any other means that involves splicing of human immunoglobulin gene sequences to other DNA sequences. Such recombinant human antibodies have variable and constant regions derived from human germline immunoglobulin sequences. In certain embodiments, however, such recombinant human antibodies are subjected to in vitro mutagenesis (or, when an animal transgenic for human Ig sequences is used, in vivo somatic mutagenesis) and thus the amino acid sequences of the VH and VL regions of the recombinant antibodies are sequences that, while derived from and related to human germline VH and VL sequences, may not naturally exist within the human antibody germline repertoire in vivo.

The term "chimeric antibody" refers to antibodies which comprise heavy and light chain variable region sequences from one species and constant region sequences from another species, such as antibodies having murine heavy and light chain variable regions linked to human constant regions. The chimeric antibody can be produced through recombinant molecular biological techniques, or may be physically conjugated together.

The term "CDR-grafted antibody" refers to antibodies which comprise heavy and light chain variable region sequences from one species but in which the sequences of one or more of the CDR regions of VH and/or VL are replaced with CDR sequences of another species, such as antibodies having murine heavy and light chain variable regions in which one or more of the murine CDRs (*e.g.*, CDR3) has been replaced with human CDR sequences.

The terms "Kabat numbering", "Kabat definitions and "Kabat labeling" are used interchangeably herein. These terms, which are recognized in the art, refer to a system of numbering amino acid residues which are more variable (*i.e.* hypervariable) than other amino acid residues in the heavy and light chain variable regions of an antibody, or an antigen binding portion thereof (Kabat *et al.* (1971) *Ann. NY Acad, Sci.* 190:382-391 and , Kabat, E.A., *et al.* (1991) *Sequences of Proteins of Immunological Interest, Fifth Edition*, U.S. Department of Health and Human Services, NIH Publication No. 91-3242). For the heavy chain variable region, the hypervariable region ranges from amino acid positions 31 to 35 for CDR1, amino acid positions 50 to 65 for CDR2, and amino acid positions 95 to 102 for CDR3. For the light chain variable region, the hypervariable region ranges from amino acid positions

24 to 34 for CDR1, amino acid positions 50 to 56 for CDR2, and amino acid positions 89 to 97 for CDR3.

As used herein, the terms "acceptor" and "acceptor antibody" refer to the antibody or nucleic acid sequence providing or encoding at least 50, 55, 60, 65, 70, 75 or 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% of the amino acid sequences of one or more of the framework regions. In some embodiments, the term "acceptor" refers to the antibody amino acid or nucleic acid sequence providing or encoding the constant region(s). In yet another embodiment, the term "acceptor" refers to the antibody amino acid or nucleic acid sequence providing or encoding one or more of the framework regions and the constant region(s). In a specific embodiment, the term "acceptor" refers to a human antibody amino acid or nucleic acid sequence that provides or encodes at least 50, 55, 60, 65, 70, 75 or 80%, particularly, at least 85%, at least 90%, at least 95%, at least 98%, or 100% of the amino acid sequences of one or more of the framework regions. In accordance with this embodiment, an acceptor may contain at least 1, at least 2, at least 3, least 4, at least 5, or at least 10 amino acid residues that does (do) not occur at one or more specific positions of a human antibody. An acceptor framework region and/or acceptor constant region(s) may be, e.g., derived or obtained from a germline antibody gene, a mature antibody gene, a functional antibody (e.g., antibodies well-known in the art, antibodies in development, or antibodies commercially available).

As used herein, the term "CDR" refers to the complementarity determining region within antibody variable sequences. There are three CDRs in each of the variable regions of the heavy chain and the light chain, which are designated CDR1, CDR2 and CDR3, for each of the variable regions. The term “CDR set” as used herein refers to a group of three CDRs that occur in a single variable region capable of binding the antigen. The exact boundaries of these CDRs have been defined differently according to different systems. The system described by Kabat (Kabat et al., Sequences of Proteins of Immunological Interest (National Institutes of Health, Bethesda, Md. (1987) and (1991)) not only provides an unambiguous residue numbering system applicable to any variable region of an antibody, but also provides precise residue boundaries defining the three CDRs. These CDRs may be referred to as Kabat CDRs. Chothia and coworkers (Chothia &Lesk, J. Mol. Biol. 196:901-917 (1987) and Chothia et al., Nature 342:877-883 (1989)) found that certain sub- portions within Kabat CDRs adopt nearly identical peptide backbone conformations, despite

having great diversity at the level of amino acid sequence. These sub-portions were designated as L1, L2 and L3 or H1, H2 and H3 where the "L" and the "H" designates the light chain and the heavy chains regions, respectively. These regions may be referred to as Chothia CDRs, which have boundaries that overlap with Kabat CDRs. Other boundaries defining CDRs overlapping with the Kabat CDRs have been described by Padlan (FASEB J. 9:133-139 (1995)) and MacCallum (J Mol Biol 262(5):732-45 (1996)). Still other CDR boundary definitions may not strictly follow one of the above systems, but will nonetheless overlap with the Kabat CDRs, although they may be shortened or lengthened in light of prediction or experimental findings that particular residues or groups of residues or even entire CDRs do not significantly impact antigen binding. The methods used herein may utilize CDRs defined according to any of these systems, although particular embodiments use Kabat or Chothia defined CDRs.

As used herein, the term "canonical" residue refers to a residue in a CDR or framework that defines a particular canonical CDR structure as defined by Chothia et al. (J. Mol. Biol. 196:901-907 (1987); Chothia et al., J. Mol. Biol. 227:799 (1992), both are incorporated herein by reference). According to Chothia et al., critical portions of the CDRs of many antibodies have nearly identical peptide backbone confirmations despite great diversity at the level of amino acid sequence. Each canonical structure specifies primarily a set of peptide backbone torsion angles for a contiguous segment of amino acid residues forming a loop.

As used herein, the terms "donor" and "donor antibody" refer to an antibody providing one or more CDRs. In a particular embodiment, the donor antibody is an antibody from a species different from the antibody from which the framework regions are obtained or derived. In the context of a humanized antibody, the term "donor antibody" refers to a non-human antibody providing one or more CDRs.

As used herein, the term "framework" or "framework sequence" refers to the remaining sequences of a variable region minus the CDRs. Because the exact definition of a CDR sequence can be determined by different systems, the meaning of a framework sequence is subject to correspondingly different interpretations. The six CDRs (CDR-L1, -L2, and -L3 of light chain and CDR-H1, -H2, and -H3 of heavy chain) also divide the framework regions on the light chain and the heavy chain into four sub-regions (FR1, FR2, FR3 and FR4) on each chain, in which CDR1 is positioned between FR1 and FR2, CDR2 between FR2 and FR3, and CDR3 between

FR3 and FR4. Without specifying the particular sub-regions as FR1, FR2, FR3 or FR4, a framework region, as referred by others, represents the combined FR's within the variable region of a single, naturally occurring immunoglobulin chain. As used herein, a FR represents one of the four sub- regions, and FRs represents two or more of the four sub- regions constituting a framework region.

Human heavy chain and light chain acceptor sequences are known in the art. In one embodiment of the invention the human heavy chain and light chain acceptor sequences are selected from the sequences described in Table 2 and Table 3. Different combinations for human framework sequences FR1 to FR4 are stated in said tables.

Table 2: Human heavy chain acceptor sequences

SEQ ID No.	Protein region	Sequence
		123456789**0**123456789**0**123456789**0**12
43	VH7-4.1/JH6 FR1	QVQLVQSGSELKKPGASVKVSCKASGYTFT
44	VH7-4.1/JH6 FR2	WVRQAPGQGLEWMG
45	VH7-4.1/JH6 FR3	RFVFSLDTSVSTAYLQICSLKAEDTAVYYCAR
46	VH7-4.1/JH6 FR4	WGQGTTVTVSS
47	VH1-2/JH6 FR1	QVQLVQSGAEVKKPGASVKVSCKASGYTFT
44	VH1-2/JH6 FR2	WVRQAPGQGLEWMG
48	VH1-2/JH6 FR3	RVTMTRDTSISTAYMELSRLRSDDTAVYYCAR
46	VH1-2/JH6 FR4	WGQGTTVTVSS

Table 3: Human light chain acceptor sequences

SEQ ID No.	Protein region	Sequence
		123456789012345678901234567890 12
49	1-12/L5/JK2 FR1	DIQMTQSPSSVSASVGDRVTITC
50	1-12/L5/JK2 FR2	WYQQKPGKAPKLLIY
51	1-12/L5/JK2 FR3	GVPSRFSGSGSGTDFTLTISSLQPEDFATYYC
52	1-12/L5/JK2 FR4	FGQGTKLEIKR
53	3-15/L2/JK2 FR1	EIVMTQSPATLSLSPGERATLSC
54	3-15/L2/JK2 FR2	WYQQKPGQAPRLLIY
55	3-15/L2/JK2 FR3	GIPARFSGSGSGTDFTLTISSLQSEDFAVYYC
52	3-15/L2/JK2 FR4	FGQGTKLEIKR

As used herein, the term "germline antibody gene" or "gene fragment" refers to an immunoglobulin sequence encoded by non-lymphoid cells that have not

undergone the maturation process that leads to genetic rearrangement and mutation for expression of a particular immunoglobulin. (See, e.g., Shapiro et al., Crit. Rev. Immunol. 22(3): 183-200 (2002); Marchalonis et al., Adv Exp Med Biol. 484:13-30 (2001)). One of the advantages provided by various embodiments of the present invention stems from the recognition that germline antibody genes are more likely than mature antibody genes to conserve essential amino acid sequence structures characteristic of individuals in the species, hence less likely to be recognized as from a foreign source when used therapeutically in that species.

As used herein, the term "key" residues refer to certain residues within the variable region that have more impact on the binding specificity and/or affinity of an antibody, in particular a humanized antibody. A key residue includes, but is not limited to, one or more of the following: a residue that is adjacent to a CDR, a potential glycosylation site (can be either N- or O-glycosylation site), a rare residue, a residue capable of interacting with the antigen, a residue capable of interacting with a CDR, a canonical residue, a contact residue between heavy chain variable region and light chain variable region, a residue within the Vernier zone, and a residue in the region that overlaps between the Chothia definition of a variable heavy chain CDR1 and the Kabat definition of the first heavy chain framework.

The term “humanized antibody” generally refers to antibodies which comprise heavy and light chain variable region sequences from a non-human species (*e.g.*, a mouse) but in which at least a portion of the VH and/or VL sequence has been altered to be more “human-like”, *i.e.*, more similar to human germline variable sequences. One type of humanized antibody is a CDR-grafted antibody, in which human CDR sequences are introduced into non-human VH and VL sequences to replace the corresponding nonhuman CDR sequences.

In particular, the term "humanized antibody" as used herein, is an antibody or a variant, derivative, analog or fragment thereof which immunospecifically binds to an antigen of interest and which comprises a framework (FR) region having substantially the amino acid sequence of a human antibody and a complementary determining region (CDR) having substantially the amino acid sequence of a non-human antibody. As used herein, the term "substantially" in the context of a CDR refers to a CDR having an amino acid sequence at least 50, 55, 60, 65, 70, 75 or 80%, particularly at least 85%, at least 90%, at least 95%, at least 98% or at least 99% identical to the amino acid sequence of a non-human antibody CDR. A humanized

antibody comprises substantially all of at least one, and typically two, variable domains (Fab, Fab', F(ab') 2, FabC, Fv) in which all or substantially all of the CDR regions correspond to those of a non-human immunoglobulin (i.e., donor antibody) and all or substantially all of the framework regions are those of a human immunoglobulin consensus sequence. Particularly, a humanized antibody also comprises at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin. In some embodiments, a humanized antibody contains both the light chain as well as at least the variable domain of a heavy chain. The antibody also may include the CH1, hinge, CH2, CH3, and CH4 regions of the heavy chain. In some embodiments, a humanized antibody only contains a humanized light chain. In some embodiments, a humanized antibody only contains a humanized heavy chain. In specific embodiments, a humanized antibody only contains a humanized variable domain of a light chain and/or humanized heavy chain.

The humanized antibody can be selected from any class of immunoglobulins, including IgY, IgM, IgG, IgD, IgA and IgE, and any isotype, including without limitation IgA1, IgA2, IgG1, IgG2, IgG3 and IgG4. The humanized antibody may comprise sequences from more than one class or isotype, and particular constant domains may be selected to optimize desired effector functions using techniques well-known in the art.

The framework and CDR regions of a humanized antibody need not correspond precisely to the parental sequences, e.g., the donor antibody CDR or the consensus framework may be mutagenized by substitution, insertion and/or deletion of at least one amino acid residue so that the CDR or framework residue at that site does not correspond to either the donor antibody or the consensus framework. In a particular embodiment, such mutations, however, will not be extensive. Usually, at least 50, 55, 60, 65, 70, 75 or 80%, particularly at least 85%, more particularly at least 90%, and in particular at least 95% of the humanized antibody residues will correspond to those of the parental FR and CDR sequences. As used herein, the term "consensus framework" refers to the framework region in the consensus immunoglobulin sequence. As used herein, the term "consensus immunoglobulin sequence" refers to the sequence formed from the most frequently occurring amino acids (or nucleotides) in a family of related immunoglobulin sequences (See e.g., Winnaker, From Genes to Clones (Verlagsgesellschaft, Weinheim, Germany 1987). In a family of immunoglobulins, each position in the consensus sequence is occupied by

the amino acid occurring most frequently at that position in the family. If two amino acids occur equally frequently, either can be included in the consensus sequence.

As used herein, "Vernier" zone refers to a subset of framework residues that may adjust CDR structure and fine-tune the fit to antigen as described by Foote and Winter (1992, J. Mol. Biol. 224:487-499, which is incorporated herein by reference). Vernier zone residues form a layer underlying the CDRs and may impact on the structure of CDRs and the affinity of the antibody.

The term "inhibition of binding" of RAGE to one of his ligands as used herein encompasses partial (as for example by about 1 to10% or more, in particular about 20%, 30%, 40%, 50%, 60%, 70%, 80%, 85%, 90%, or 95% or more) or complete reduction of said ligand binding activity. Said "inhibition of binding" may be determined by any suitable method available in the art, preferably by any method as exemplified herein, as for example HTRF assays described herein.

As used herein, the term "neutralizing" refers to neutralization of biological activity of a target protein when a binding protein specifically binds the target protein. Neutralizing may be the result of different ways of binding of said binding protein to the target. For example, neutralizing may be caused by binding of the binding protein in a region of the target, which does not affect receptor binding to the target molecule. Alternatively binding of a binding protein may result in a blockade of the receptor binding to the target, which blockade finally neutralizes the target protein activity. Each of said different mechanism may occur according to the invention.

Particularly a neutralizing binding protein is a neutralizing antibody whose binding to RAGE results in neutralization of a biological activity of RAGE Particularly the neutralizing binding protein binds RAGE and reduces a biologically activity of RAGE by at least about 1 to10%, at least about 20%, 30%, 40%, 50%, 60%, 70%, 80%, 85% or more. Neutralization of a biological activity of RAGE by a neutralizing binding protein can be assessed by measuring one or more indicators of RAGE biological activity well known in the art, and/or exemplified in the experimental part, as in particular, in Examples 5, 6, 10, and 16 to 19. For example neutralization of RAGE neutralizes the binding of RAGE to Aß-globulomers as measured in Examples 5 and 6 below.

An "inhibition of soluble Aß1-40 peptide-induced reduction of cerebral blood volume" determined in vivo in an animal model relates to a partial or complete inhibition, as for example by at least about 1 to10%, at least about 15%, 20%, 30%,

40%, 50%, 60%, 70%, 80%, 85% or more, if compared to a non-treated control, or in particular, if compared to a negative, monoclonal or polyclonal, immunoglobulin-isotype control.

An "improvement of the cerebral blood volume" as in vivo in an animal model over-expressing human APP, relates to a statistically significant improvement of CBV, if compared to a non-treated control, or in particular, if compared to a negative, monoclonal or polyclonal, immunoglobulin-isotype control.

A "reduction of amyloid plaque number and /or amyloid plaque area" as measured in vivo in an animal model over-expressing human APP relates to a partial or complete reduction, as for example by at least about 1 to10%, at least about 15%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 85% or more, if compared to a non-treated control, or in particular, if compared to a negative, monoclonal or polyclonal, immunoglobulin-isotype control.

An "inhibition of aggregated Aß1-40 peptide-induced dynamin cleavage of hippocampal neurons in vitro; relates to a partial or complete inhibition, as for example by at least about 1 to10%, at least about 15%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 85% or more, if compared to a non-treated control, or in particular, if compared to a negative, monoclonal or polyclonal, immunoglobulin-isotype control.

A "reversal of Aß1-42 globulomer-induced reduction of synaptic transmission" in vitro, relates to a partial or complete reversial, as for example by at least about 1 to10%, at least about 15%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 85% or more, if compared to a non-treated control, or in particular, if compared to a negative, monoclonal or polyclonal, immunoglobulin-isotype control.

A "neutralizing monoclonal antibody" as used herein is intended to refer to a preparation of antibody molecules, which upon binding to the specific antigen are able to compete and inhibit the binding of the natural ligand for said antigen. In a particular embodiment of the present application, the neutralizing antibodies of the present invention are capable of competing with RAGE for binding to at least one of its ligands, in particular a ligand selected from Aß-peptides, Aß-globulomers, S100b and Amphoterin, and to prevent RAGE biological activity or function.

The term "activity" includes activities such as the binding specificity/affinity of an antibody for an antigen, for example, an anti-RAGE antibody that binds to an RAGE antigen and/or the neutralizing potency of an antibody, for example, an anti-RAGE antibody whose binding to RAGE neutralizes the biological activity of RAGE.

The "biological function" or "activity" of RAGE may be described as that of a signal transducing cell surface receptor for Aß or a receptor that mediates transport of proteins into or through the cell.

The term "epitope" or "antigenic determinant" includes any polypeptide determinant capable of specific binding to an immunoglobulin or T-cell receptor. In certain embodiments, epitope determinants include chemically active surface groupings of molecules such as amino acids, sugar side chains, phosphoryl, or sulfonyl, and, in certain embodiments, may have specific three dimensional structural characteristics, and/or specific charge characteristics. An epitope is a region of an antigen that is bound by an antibody. In certain embodiments, an antibody is said to specifically bind an antigen when it preferentially recognizes its target antigen in a complex mixture of proteins and/or macromolecules.

The term "surface plasmon resonance", as used herein, refers to an optical phenomenon that allows for the analysis of real-time biospecific interactions by detection of alterations in protein concentrations within a biosensor matrix, for example using the BIAcore system (Pharmacia Biosensor AB, Uppsala, Sweden and Piscataway, NJ). For further descriptions, see Jönsson, U., *et al.* (1993) *Ann. Biol. Clin.* 51:19-26; Jönsson, U., *et al.* (1991) *Biotechniques* 11:620-627; Johnsson, B., *et al.* (1995) *J. Mol. Recognit.* 8:125-131; and Johnnson, B., *et al.* (1991) *Anal. Biochem.* 198:268-277.

The term "k_{on}", as used herein, is intended to refer to the on rate constant for association of an antibody to the antigen to form the antibody/antigen complex as is known in the art.

The term "k_{off}", as used herein, is intended to refer to the off rate constant for dissociation of an antibody from the antibody/antigen complex as is known in the art.

The term "K_d", as used herein, is intended to refer to the dissociation constant of a particular antibody-antigen interaction as is known in the art.

The term "labelled binding protein" as used herein, refers to a protein with a label incorporated that provides for the identification of the binding protein. Particularly, the label is a detectable marker, e.g., incorporation of a radiolabelled amino acid or attachment to a polypeptide of biotinyl moieties that can be detected by marked avidin (e.g., streptavidin containing a fluorescent marker or enzymatic activity that can be detected by optical or colorimetric methods). Examples of labels

for polypeptides include, but are not limited to, the following: radioisotopes or radionuclides (e.g., ^{3}H, ^{14}C, ^{35}S, ^{90}Y, ^{99}Tc, ^{111}In, ^{125}I, ^{131}I, ^{177}Lu, ^{166}Ho, or ^{153}Sm); fluorescent labels (e.g., FITC, rhodamine, lanthanide phosphors), enzymatic labels (e.g., horseradish peroxidase, luciferase, alkaline phosphatase); chemiluminescent markers; biotinyl groups; predetermined polypeptide epitopes recognized by a secondary reporter (e.g., leucine zipper pair sequences, binding sites for secondary antibodies, metal binding domains, epitope tags); and magnetic agents, such as gadolinium chelates.

The term “antibody conjugate” refers to a binding protein, such as an antibody, chemically linked to a second chemical moiety, such as a therapeutic or cytotoxic agent. The term "agent" is used herein to denote a chemical compound, a mixture of chemical compounds, a biological macromolecule, or an extract made from biological materials. Particularly the therapeutic or cytotoxic agents include, but are not limited to, pertussis toxin, taxol, cytochalasin B, gramicidin D, ethidium bromide, emetine, mitomycin, etoposide, tenoposide, vincristine, vinblastine, colchicin, doxorubicin, daunorubicin, dihydroxy anthracin dione, mitoxantrone, mithramycin, actinomycin D, 1-dehydrotestosterone, glucocorticoids, procaine, tetracaine, lidocaine, propranolol, and puromycin and analogs or homologs thereof.

The terms “crystal”, and “crystallized” as used herein, refer to an antibody, or antigen binding portion thereof, that exists in the form of a crystal. Crystals are one form of the solid state of matter, which is distinct from other forms such as the amorphous solid state or the liquid crystalline state. Crystals are composed of regular, repeating, three-dimensional arrays of atoms, ions, molecules (e.g., proteins such as antibodies), or molecular assemblies (e.g., antigen/antibody complexes). These three-dimensional arrays are arranged according to specific mathematical relationships that are well-understood in the field. The fundamental unit, or building block, that is repeated in a crystal is called the asymmetric unit. Repetition of the asymmetric unit in an arrangement that conforms to a given, well-defined crystallographic symmetry provides the "unit cell" of the crystal. Repetition of the unit cell by regular translations in all three dimensions provides the crystal. See Giege, R. and Ducruix, A. Barrett, Crystallization of Nucleic Acids and Proteins, a Practical Approach, 2nd ea., pp. 20 1-16, Oxford University Press, New York, New York, (1999)."

The term "polynucleotide" as referred to herein means a polymeric form of two or more nucleotides, either ribonucleotides or deoxynucleotides or a modified form of either type of nucleotide. The term includes single and double stranded forms of DNA but particularly is double-stranded DNA.

The term "isolated polynucleotide" as used herein shall mean a polynucleotide (e.g., of genomic, cDNA, or synthetic origin, or some combination thereof) that, by virtue of its origin , the "isolated polynucleotide": is not associated with all or a portion of a polynucleotide with which the "isolated polynucleotide" is found in nature; is operably linked to a polynucleotide that it is not linked to in nature; or does not occur in nature as part of a larger sequence.

The term "vector", as used herein, is intended to refer to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments may be ligated. Another type of vector is a viral vector, wherein additional DNA segments may be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (e.g., non-episomal mammalian vectors) can be integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "recombinant expression vectors" (or simply, "expression vectors"). In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" may be used interchangeably as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors, such as viral vectors (e.g., replication defective retroviruses, adenoviruses and adeno-associated viruses), which serve equivalent functions.

The term "operably linked" refers to a juxtaposition wherein the components described are in a relationship permitting them to function in their intended manner. A control sequence "operably linked" to a coding sequence is ligated in such a way that expression of the coding sequence is achieved under conditions compatible with the control sequences. "Operably linked" sequences include both expression control sequences that are contiguous with the gene of interest and expression control

sequences that act in *trans* or at a distance to control the gene of interest. The term "expression control sequence" as used herein refers to polynucleotide sequences, which are necessary to effect the expression and processing of coding sequences to which they are ligated. Expression control sequences include appropriate transcription initiation, termination, promoter and enhancer sequences; efficient RNA processing signals such as splicing and polyadenylation signals; sequences that stabilize cytoplasmic mRNA; sequences that enhance translation efficiency (i.e., Kozak consensus sequence); sequences that enhance protein stability; and when desired, sequences that enhance protein secretion. The nature of such control sequences differs depending upon the host organism; in prokaryotes, such control sequences generally include promoter, ribosomal binding site, and transcription termination sequence; in eukaryotes, generally, such control sequences include promoters and transcription termination sequence. The term "control sequences" is intended to include components whose presence is essential for expression and processing, and can also include additional components whose presence is advantageous, for example, leader sequences and fusion partner sequences.

"Transformation", as defined herein, refers to any process by which exogenous DNA enters a host cell. Transformation may occur under natural or artificial conditions using various methods well known in the art. Transformation may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method is selected based on the host cell being transformed and may include, but is not limited to, viral infection, electroporation, lipofection, and particle bombardment. Such "transformed" cells include stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome. They also include cells which transiently express the inserted DNA or RNA for limited periods of time.

The term "recombinant host cell" (or simply "host cell"), as used herein, is intended to refer to a cell into which exogenous DNA has been introduced. It should be understood that such terms are intended to refer not only to the particular subject cell, but, to the progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term "host cell" as used herein. Particularly host cells include

prokaryotic and eukaryotic cells selected from any of the Kingdoms of life. Particular eukaryotic cells include protist, fungal, plant and animal cells. In particular host cells include but are not limited to the prokaryotic cell line E. coli; mammalian cell lines CHO, HEK 293 and COS; the insect cell line Sf9; and the fungal cell Saccharomyces cerevisiae.

Standard techniques may be used for recombinant DNA, oligonucleotide synthesis, and tissue culture and transformation (e.g., electroporation, lipofection). Enzymatic reactions and purification techniques may be performed according to manufacturer's specifications or as commonly accomplished in the art or as described herein. The foregoing techniques and procedures may be generally performed according to conventional methods well known in the art and as described in various general and more specific references that are cited and discussed throughout the present specification. See e.g., Sambrook et al. Molecular Cloning: A Laboratory Manual (2d ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989)), which is incorporated herein by reference for any purpose.

"Transgenic organism", as known in the art and as used herein, refers to an organism having cells that contain a transgene, wherein the transgene introduced into the organism (or an ancestor of the organism) expresses a polypeptide not naturally expressed in the organism. A "transgene" is a DNA construct, which is stably and operably integrated into the genome of a cell from which a transgenic organism develops, directing the expression of an encoded gene product in one or more cell types or tissues of the transgenic organism.

The term "regulate" and "modulate" are used interchangeably, and, as used herein, refers to a change or an alteration in the activity of a molecule of interest (e.g., the biological activity of RAGE). Modulation may be an increase or a decrease in the magnitude of a certain activity or function of the molecule of interest. Exemplary activities and functions of a molecule include, but are not limited to, binding characteristics, enzymatic activity, cell receptor activation, and signal transduction.

Correspondingly, the term "modulator," as used herein, is a compound capable of changing or altering an activity or function of a molecule of interest (e.g., the biological activity of RAGE). For example, a modulator may cause an increase or decrease in the magnitude of a certain activity or function of a molecule compared to the magnitude of the activity or function observed in the absence of the modulator.

The term "agonist", as used herein, refers to a modulator that, when contacted with a molecule of interest, causes an increase in the magnitude of a certain activity or function of the molecule compared to the magnitude of the activity or function observed in the absence of the agonist. Particular agonists of interest may include, but are not limited to, RAGE polypeptides or polypeptides, nucleic acids, carbohydrates, or any other molecules that bind to RAGE.

The term "antagonist", as used herein, refer to a modulator that, when contacted with a molecule of interest causes a decrease in the magnitude of a certain activity or function of the molecule compared to the magnitude of the activity or function observed in the absence of the antagonist. Exemplary antagonists include, but are not limited to, proteins, peptides, antibodies, peptibodies, carbohydrates or small organic molecules. Peptibodies are described, e.g., in WO01/83525.

Particular antagonists of interest include those that block or modulate the biological or immunological activity of RAGE. Antagonists of RAGE may include, but are not limited to, proteins, nucleic acids, carbohydrates, or any other molecules, which bind to RAGE, particularly monoclonal antibodies that interact with the RAGE molecule. It should be noted that the interaction with RAGE may result in binding and neutralization of other ligands/cell membrane components, and may be useful for additive or synergistic functioning against multiple diseases.

As used herein, the term "effective amount" refers to the amount of a therapy which is sufficient to reduce or ameliorate the severity and/or duration of a disorder or one or more symptoms thereof, prevent the advancement of a disorder, cause regression of a disorder, prevent the recurrence, development, onset or progression of one or more symptoms associated with a disorder, detect a disorder, or enhance or improve the prophylactic or therapeutic effect(s) of another therapy (e.g. , prophylactic or therapeutic agent).

The term "sample", as used herein, is used in its broadest sense. A “biological sample”, as used herein, includes, but is not limited to, any quantity of a substance from a living thing or formerly living thing. Such living things include, but are not limited to, humans, mice, rats, monkeys, dogs, rabbits and other animals. Such substances include, but are not limited to, blood, serum, urine, synovial fluid, cells, organs, tissues, bone marrow, lymph nodes and spleen.

2. Specific Embodiments

Specific embodiments of the invention are listed below:

1. An isolated binding protein that dissociates from human RAGE with a K_D of 1×10^{-7} M or less and a k_{off} rate constant of 1×10^{-2} s^{-1} or less, both determined by surface plasmon resonance.

2. i) The binding protein of embodiment 1, that binds to human RAGE and modulates, in particular inhibits, the ability of RAGE to bind to at least one of its ligands, as determined in a standard in vitro assay, as for example, a HTRF assay, as for example described in more detail in the experimental part, in particular examples 4 and 5, and references cited therein.
ii) The binding protein of embodiment1 being capable of inhibiting a RAGE-mediated biological activity.
iii) A binding protein, in particular, according to one of the preceding embodiments, having at least one of the following biological activities:
   a. inhibition of soluble Aß1-40 peptide-induced reduction of cerebral blood volume (CBV) in vivo in an animal model, like C57BL/6 female mice, as for example described in more detail in the experimental part, in particular example 10 and references cited therein;
   b. improvement of the cerebral blood volume in vivo in an animal model over-expressing human APP, like the transgenic Tg2576 mouse model, as for example described in more detail in the experimental part, in particular example 16 and references cited therein;
   c. reduction of amyloid plaque number and /or amyloid plaque area in vivo in an animal model over-expressing human APP, like the transgenic Tg2576 mouse model, as for example described in more detail in the experimental part, in particular example 19 and references cited therein;
   d. inhibition of aggregated Aß1-40 peptide-induced dynamin cleavage of hippocampal neurons in vitro, like hippocampal neurons as obtained from embryonic rats, as for example described in more detail in the experimental part, in particular example 17 and references cited therein;

e. reversal of Aß1-42 globulomer-induced reduction of synaptic transmission in vitro, determined in hippocampal slice cultures, as for example described in more detail in the experimental part, in particular example 18 and references cited therein.

3. The binding protein of embodiment 1 or 2, wherein the ligand is selected from Aß peptides, Aß-globulomers, S100b and Amphoterin.

4. The binding protein of one of the preceding embodiments, which is a neutralizing binding protein.

5. The binding protein of one of the preceding embodiments, which is capable of blocking, in particular inhibiting, the binding of Aß globulomer to human RAGE.

6. The binding protein of one of the preceding embodiments wherein said wherein said Aß globulomer is Aß1-42.

7. The binding protein of one of the preceding embodiments, wherein said binding protein interacts with at least one, as for example 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, or 12amino acid residue of the C1- and /or C2-domain of human RAGE.

8. The binding protein according to one of the preceding embodiments, which is a humanized antibody.

9. The binding protein according to one of the preceding embodiments, comprising an antigen binding domain, said binding protein capable of binding an epitope of a human RAGE molecule, said antigen binding domain comprising at least one CDR comprising an amino acid sequence selected from

the CDR-H3 group of amino acid sequences consisting of SEQ ID NO.: 4, 12 and 20; and modified CDR amino acid sequences having a sequence identity

of at least 50%, as for example at least 55, 60, 65, 70, 75, 80, 85, 90, 95 % identity, to one of said sequences;
the CDR-L3 group of amino acid sequences consisting of SEQ ID NO.: 8, 16 and 24; andmodified CDR amino acid sequences having a sequence identity of at least 50%, as for example at least 55, 60, 65, 70, 75, 80, 85, 90, 95 % identity, to one of said sequences.

10. A binding protein comprising an antigen binding domain, said binding protein capable of binding an epitope of a human RAGE molecule, said antigen binding domain comprising at least one CDR comprising an amino acid sequence selected from:
the CDR-H3 group of amino acid sequences consisting of SEQ ID NO.: 4, 12 and 20; and modified CDR amino acid sequences having a sequence identity of at least 50%, as for example at least 55, 60, 65, 70, 75, 80, 85, 90, 95 % identity, to one of said sequences;
the CDR-L3 group of amino acid sequences consisting of SEQ ID NO.: 8, 16 and 24; and modified CDR amino acid sequences having a sequence identity of at least 50%, as for example at least 55, 60, 65, 70, 75, 80, 85, 90, 95 % identity, to one of said sequences.

11. The binding protein according to one of the preceding embodiments, further comprising at least one CDR comprising an amino acid sequence selected from the CDR-H1 group consisting of SEQ ID NO: 2, 10, 18; or selected from the CDR-H2 group consisting of SEQ ID NO: 3, 11, 19; or selected from the CDR-L1 group consisting of SEQ ID NO: 6, 14, 22; or selected from the CDR-L2 group consisting of SEQ ID NO: 7, 15, 23;
and modified CDR amino acid sequences having a sequence identity of at least 50%, as for example at least 55, 60, 65, 70, 75, 80, 85, 90, 95 % identity, to one of said sequences.

12. The binding protein according to any one of the preceding embodiments, wherein said at least one CDR comprises an amino acid sequence selected from the group consisting of:

SEQ ID NO.: 2	Residues 31-35 of SEQ ID NO.:1
SEQ ID NO.: 3	Residues 50-68 of SEQ ID NO.:1
SEQ ID NO.: 4	Residues 101-108 of SEQ ID NO.:1
SEQ ID NO.: 6	Residues 24-34 of SEQ ID NO.:5
SEQ ID NO.: 7	Residues 50-56 of SEQ ID NO.:5
SEQ ID NO.: 8	Residues 89-97 of SEQ ID NO.:5
SEQ ID NO.: 10	Residues 31-35 of SEQ ID NO.:9
SEQ ID NO.: 11	Residues 50-66 of SEQ ID NO.:9
SEQ ID NO.: 12	Residues 97-109 of SEQ ID NO.:9
SEQ ID NO.: 14	Residues 24-34 of SEQ ID NO.:13
SEQ ID NO.: 15	Residues 50-56 of SEQ ID NO.:13
SEQ ID NO.: 16	Residues 89-97 of SEQ ID NO.:13
SEQ ID NO.: 18	Residues 31-35 of SEQ ID NO.:17
SEQ ID NO.: 19	Residues 50-66 of SEQ ID NO.:17
SEQ ID NO.: 20	Residues 99-109 of SEQ ID NO.:17
SEQ ID NO.: 22	Residues 24-34 of SEQ ID NO.:21
SEQ ID NO.: 23	Residues 50-56 of SEQ ID NO.:21
SEQ ID NO.: 24	Residues 89-97 of SEQ ID NO.:21

13. The binding protein according to embodiment 12, comprising at least 3 CDRs which are selected from a variable domain CDR set consisting of:

VH 7F9 set		
VH 7F9 CDR-H1	Residues 31-35 of SEQ ID NO.:1	SEQ ID NO: 2
VH 7F9 CDR-H2	Residues 50-68 of SEQ ID NO.:1	SEQ ID NO: 3
VH 7F9 CDR-H3	Residues 101-108 of SEQ ID NO.:1	SEQ ID NO: 4
VL 7F9 set		
VL 7F9 CDR-L1	Residues 24-34 of SEQ ID NO.:5	SEQ ID NO: 6
VL 7F9 CDR-L2	Residues 50-56 of SEQ ID NO.:5	SEQ ID NO: 7
VL 7F9 CDR-L3	Residues 89-97 of SEQ ID NO.:5	SEQ ID NO: 8
VH 11E6 set		
VH 11E6 CDR-H1	Residues 31-35 of SEQ ID NO.:9	SEQ ID NO: 10
VH 11E6 CDR-H2	Residues 50-66 of SEQ ID NO.:9	SEQ ID NO: 11
VH 11E6 CDR-H3	Residues 99-109 of SEQ ID NO.:9	SEQ ID NO: 12
VL 11E6 set		
VL 11E6 CDR-L1	Residues 24-34 of SEQ ID NO.:13	SEQ ID NO: 14
VL 11E6 CDR-L2	Residues 50-56 of SEQ ID NO.:13	SEQ ID NO: 15
VL 11E6 CDR-L3	Residues 89-97 of SEQ ID NO.:13	SEQ ID NO: 16
VH 4E5 set		
VH 4E5 CDR-H1	Residues 31-35 of SEQ ID NO.:17	SEQ ID NO: 18
VH 4E5 CDR-H2	Residues 50-66 of SEQ ID NO.:17	SEQ ID NO: 19
VH 4E5 CDR-H3	Residues 99-109 of SEQ ID NO.:17	SEQ ID NO: 20
VL 4E5 set		
VL 4E5 CDR-L1	Residues 24-34 of SEQ ID NO.:21	SEQ ID NO: 22
VL 4E5 CDR-L2	Residues 50-56 of SEQ ID NO.:21	SEQ ID NO: 23
VL 4E5 CDR-L3	Residues 89-97 of SEQ ID NO.:21	SEQ ID NO: 24

or a variable domain set wherein at least one of said 3 CDRs is a modified CDR amino acid sequence having a sequence identity of at least 50%, as for example at least 55, 60, 65, 70, 75, 80, 85, 90, 95 % identity, to the parent sequence.

14. The binding protein according to embodiment 13, comprising at least two variable domain CDR sets.

15. The binding protein according to embodiment 14, wherein said at least two variable domain CDR sets are selected from a group consisting of:

VH 7F9 set & VL 7F9 set;
VH 4E5 set & VL 4E5 and
VH 11E6 set & VL 11E6 set.

16. The binding protein according to one of the preceding embodiments, further comprising a human acceptor framework.

17. The binding protein according to embodiment 16, wherein said human acceptor framework comprises at least one amino acid sequence selected from the group consisting of SEQ ID NO: 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53 54 and 55.

18. The binding protein of any one of the preceding embodiments comprising at least one heavy chain variable domain selected from SEQ ID NO: 56 and 57; and/or at least one light chain variable domain selected from SEQ ID NO: 58 and 59.

19. The binding protein of embodiment 18, wherein said binding protein comprises two variable domains, wherein said two variable domains have amino acid sequences selected from :
SEQ ID NOs: 56 & 58 ; 56 & 59 ;
SEQ ID NOs: .57 & 58 ; 57 & 59.

20. The binding protein according to any one of the embodiment 16 to 19, wherein said human acceptor framework comprises at least one framework region amino acid substitution at a key residue, said key residue selected from the group consisting of:

a residue adjacent to a CDR;

a glycosylation site residue;

a rare residue;

a residue capable of interacting with a RAGE epitope;

a residue capable of interacting with a CDR;

a canonical residue;

a contact residue between heavy chain variable region and light chain variable region;

a residue within a Vernier zone;

an N-terminal residue capable of para-glutamate formation; and

a residue in a region that overlaps between a Chothia-defined variable heavy chain CDR1 and a Kabat-defined first heavy chain framework.

21. The binding protein according to embodiment 20, wherein said key residue are selected from the group consisting

(heavy chain sequence position): 1, 2, 68, 70, 72, 76, 85, 89, 95
(light chain sequence position): 11, 13, 43, 49, 58, 70, 87.

22. The binding protein of any one of the preceding embodiments, wherein the binding protein is a consensus human variable domain.

23. The binding protein of any one of the embodiments 16 to 22, wherein said human acceptor framework comprises at least one framework region amino acid substitution, as for example 1 to 20, 1 to15, 1 to 10, or 2, 3, 4, 5, 6, 7, 8 or 9 substitutions, wherein the amino acid sequence of the framework is at least 65% identical to the sequence of said human acceptor framework and comprises at least 70 amino acid residues identical to said human acceptor framework.

24. The binding protein of any one of the preceding embodiments, wherein said binding protein comprises at least one (framework mutated) variable domain having an amino acid sequence selected from the group consisting of:
(heavy chain sequences) SEQ ID NO: 62, 67, 68 and 69;
(light chain sequences) SEQ ID NO: 63, 64, 65 and 66.

25. The binding protein of embodiment 24, wherein said binding protein comprises two variable domains, wherein said two variable domains have amino acid sequences selected from the groups consisting of:
SEQ ID NOs: 62 & 63 ; 62 & 64 ; 62 & 65 ; 62 & 66 ;;
SEQ ID NOs: 67 & 63 ; 67 & 64 ; 67 & 65 ; 67 & 66 ;
SEQ ID NOs: 68 & 63 ; 68 & 64 ; 68 & 65 ; 68 & 66 ;

26. The binding protein of any one of the preceding embodiments, wherein said binding protein is capable of binding a target, selected from RAGE molecules.

27. The binding protein of any one of the preceding embodiments capable of binding to human RAGE.

28. The binding protein of embodiment 27, having at least one of the following additional functional characteristics:
binding to mouse and rat RAGE.

29. The binding protein of any one of the preceding embodiments, wherein the binding protein is capable of modulating, in particular neutralizing, a biological function of a target, selected from RAGE molecules.

30. The binding protein of embodiment 29, wherein said binding protein modulates, in particular inhibits, the ability of RAGE to bind to at least one of its ligands.

31. The binding protein of embodiment 30, wherein said binding protein modulates, in particular inhibits, at least one of the following interactions:

binding of human RAGE to Aß peptides, Aß-globulomers, S100b and amphoterin.

32. The binding protein of any one of the preceding embodiments, wherein said binding protein is capable of neutralizing a RAGE biological activity, as for example Aβ-induced cleavage of dynamin in primary neurons, Globulomer induced synaptic deficits in hippocampal slices, Aβ-induced decrease in CBV

33. The binding protein of embodiment 32, wherein the RAGE molecule is RAGE or a RAGE fragment, like sRAGE.

34. The binding protein of embodiment 33, wherein the RAGE is selected from human, rat and mouse.

35. The binding protein of any one of the preceding embodiments, wherein said binding protein has an on rate constant (k_{on}) to said target selected from the group consisting of: at least about $10^2 M^{-1}s^{-1}$; at least about $10^3 M^{-1}s^{-1}$; at least about $10^4 M^{-1}s^{-1}$; at least about $10^5 M^{-1}s^{-1}$; at least about $10^6 M^{-1}s^{-1}$, and at least about $10^7 M^{-1}s^{-1}$ as measured by surface plasmon resonance.

36. The binding protein of any one of the preceding embodiments, wherein said binding protein has an off rate constant (k_{off}) to said target selected from the group consisting of: at most about $10^{-2}s^{-1}$; at most about $10^{-3}s^{-1}$; at most about $10^{-4}s^{-1}$; at most about $10^{-5}s^{-1}$; and at most about $10^{-6}s^{-1}$, as measured by surface plasmon resonance.

37. The binding protein of any one of the preceding embodiments, wherein said binding protein has a dissociation constant (K_D) to said target selected from the group consisting of: at most about 10^{-7} M; at most about 10^{-8} M; at most about 10^{-9} M; at most about 10^{-10} M; at most about 10^{-11} M; at most about 10^{-12} M; and at most 10^{-13} M.

38. An antibody construct comprising a binding protein described in any one of the preceding embodiments, said antibody construct further comprising a linker polypeptide or an immunoglobulin constant domain.

39. The antibody construct according to embodiment 38, wherein said binding protein is selected from the group consisting of:
an immunoglobulin molecule,
a monoclonal antibody,
a chimeric antibody,
a CDR-grafted antibody,
a humanized antibody,
a Fab,
a Fab',
a F(ab')2,
a Fv,
a disulfide linked Fv,
a scFv,
a single domain antibody,
a diabody,
a multispecific antibody,
a dual specific antibody,
a dual variable domain immunoglobulin, and
a bispecific antibody.

40. The antibody construct according to any on of the embodiments 38 and 39, wherein said binding protein comprises a heavy chain immunoglobulin constant domain selected from the group consisting of;
a human IgM constant domain,
a human IgG1 constant domain,
a human IgG2 constant domain,
a human IgG3 constant domain,
a human IgG4 constant domain,
a human IgE constant domain,
a human IgD constant domain,

a human IgA1 constant domain
a human IgA2 constant domain
a human IgY constant domain and
and corresponding mutated domains.

41. The antibody construct according to any on of the embodiments 38 to 40, comprising an immunoglobulin constant domain having an amino acid sequence selected from the group consisting of: SEQ ID NO: 25, 41, 42, and 26.

42. An antibody conjugate comprising an antibody construct described in any one of embodiments 38 to 41, said antibody conjugate further comprising an agent selected from the group consisting of; an immunoadhension molecule, an imaging agent, a therapeutic agent, and a cytotoxic agent.

43. The antibody conjugate according to embodiment 42, wherein said agent is an imaging agent selected from the group consisting of a radiolabel, an enzyme, a fluorescent label, a luminescent label, a bioluminescent label, a magnetic label, and biotin.

44. The antibody conjugate according to embodiment 43, wherein said imaging agent is a radiolabel selected from the group consisting of: ^{3}H, ^{14}C, ^{35}S, ^{90}Y, ^{99}Tc, ^{111}In, ^{125}I, ^{131}I, ^{177}Lu, ^{166}Ho, and ^{153}Sm.

45. The antibody conjugate according to embodiment 42, wherein said agent is a therapeutic or cytotoxic agent selected from the group consisting of; an anti-metabolite, an alkylating agent, an antibiotic, a growth factor, a cytokine, an anti-angiogenic agent, an anti-mitotic agent, an anthracycline, toxin, and an apoptotic agent.

46. The antibody construct according to any on of the embodiments 38 to 41, wherein said binding protein possesses a human glycosylation pattern.

47. The antibody conjugate according to any on of the embodiments 42 to 45, wherein said binding protein possesses a human glycosylation pattern.

48. The binding protein according to any one of the embodiments 1 to 37, wherein said binding protein exists as a crystal.

49. The antibody construct according to any one of the embodiments 38 to 41, wherein said antibody construct exists as a crystal.

50. The antibody conjugate according to any one of the embodiments 42 to 45, wherein said antibody construct exists as a crystal.

51. The binding protein according to embodiment 48, wherein said crystal is a carrier-free pharmaceutical controlled release crystal.

52. The antibody construct according to embodiment 49, wherein said crystal is a carrier-free pharmaceutical controlled release crystal.

53. The antibody conjugate according to embodiment 50, wherein said crystal is a carrier-free pharmaceutical controlled release crystal.

54. The binding protein according to embodiment 48, wherein said binding protein has a greater half life in vivo than the soluble counterpart of said binding protein.

55. The antibody construct according to embodiment 49, wherein said antibody construct has a greater half life in vivo than the soluble counterpart of said antibody construct.

56. The antibody conjugate according to embodiment 50, wherein said antibody conjugate has a greater half life in vivo than the soluble counterpart of said antibody conjugate.

57. The binding protein according to embodiment 48, wherein said binding protein retains biological activity.

58. The antibody construct according to embodiment 49, wherein said antibody construct retains biological activity.

59. The antibody conjugate according to embodiment 50, wherein said antibody conjugate retains biological activity.

60. An isolated nucleic acid encoding a binding protein amino acid sequence of any one of embodiments 1-37.

61. An isolated nucleic acid encoding an antibody construct amino acid sequence of any one of embodiments 38-41.

62. An isolated nucleic acid encoding an antibody conjugate amino acid sequence of any one of embodiments 42-45.

63. A vector comprising an isolated nucleic acid according to any one of embodiments 60 to 62.

64. The vector of embodiment 63 wherein said vector is selected from the group consisting of pcDNA, pTT, pTT3, pEFBOS, pBV, pJV, pHybE, and pBJ.

65. A host cell comprising a vector according to any one of embodiments 63 and 64.

66. The host cell according to embodiment 65, wherein said host cell is a prokaryotic cell.

67. The host cell according to embodiment 66, wherein said host cell is E. coli.

68. The host cell according to embodiment 67, wherein said host cell is a eukaryotic cell.

69. The host cell according to embodiment 68, wherein said eukaryotic cell is selected from the group consisting of protist cell, animal cell, plant cell and fungal cell.

70. The host cell according to embodiment 69, wherein said eukaryotic cell is an animal cell selected from the group consisting of; a mammalian cell, an avian cell, and an insect cell.

71. The host cell according to embodiment 69, wherein said host cell is selected from HEK Cells, CHO cells COS cells and yeast cells.

72. The host cell according to embodiment 71, wherein said yeast cell is *Saccharomyces cerevisiae*.

73. The host cell according to embodiment 70, wherein said host cell is an insect Sf9 cell.

74. A method of producing a protein capable of binding RAGE, comprising culturing a host cell of any one of embodiments 65 to 73 in culture medium under conditions sufficient to produce a binding protein capable of binding RAGE.

75. A protein produced according to the method of embodiment 74.

76. A composition for the release of a binding protein said composition comprising
(a) a formulation, wherein said formulation comprises a crystallized product protein according to any one of embodiments 48 to 50, and an ingredient; and
(b) at least one polymeric carrier.

77. The composition according to embodiment 76, wherein said polymeric carrier is a polymer selected from one or more of the group consisting of: poly (acrylic acid), poly (cyanoacrylates), poly (amino acids), poly (anhydrides),

poly (depsipeptide), poly (esters), poly (lactic acid), poly (lactic-co-glycolic acid) or PLGA, poly (b-hydroxybutryate), poly (caprolactone), poly (dioxanone); poly (ethylene glycol), poly ((hydroxypropyl) methacrylamide, poly [(organo) phosphazene], poly (ortho esters), poly (vinyl alcohol), poly (vinylpyrrolidone), maleic anhydride- alkyl vinyl ether copolymers, pluronic polyols, albumin, alginate, cellulose and cellulose derivatives, collagen, fibrin, gelatin, hyaluronic acid, oligosaccharides, glycaminoglycans, sulfated polyeaccharides, blends and copolymers thereof.

78. The composition according to embodiment 76, wherein said ingredient is selected from the group consisting of albumin, sucrose, trehalose, lactitol, gelatin, hydroxypropyl-ß- cyclodextrin, methoxypolyethylene glycol and polyethylene glycol.

79. A method for treating a mammal comprising the step of administering to the mammal an effective amount of the composition according to any one of the embodiments 77 and 78.

80. A pharmaceutical composition comprising the product of any one of embodiments 1 to 59, and a pharmaceutically acceptable carrier.

81. The pharmaceutical composition of embodiment 80, wherein said pharmaceutically acceptable carrier functions as adjuvant useful to increase the absorption, or dispersion of said binding protein.

82. The pharmaceutical composition of embodiment 81, wherein said adjuvant is hyaluronidase.

83. The pharmaceutical composition of embodiment 82 further comprising at least one additional therapeutic agent for treating a disorder in which RAGE activity is detrimental.

84. The pharmaceutical composition of embodiment 83, wherein said additional agent is selected from the group consisting of: Therapeutic agent, imaging

agent, cytotoxic agent, angiogenesis inhibitors; kinase inhibitors; co-stimulation molecule blockers; adhesion molecule blockers; anti-cytokine antibody or functional fragment thereof; methotrexate; cyclosporin; rapamycin; FK506; detectable label or reporter; a TNF antagonist; an antirheumatic; a muscle relaxant, a narcotic, a non-steroid anti-inflammatory drug (NSAID), an analgesic, an anesthetic, a sedative, a local anesthetic, a neuromuscular blocker, an antimicrobial, an antipsoriatic, a corticosteriod, an anabolic steroid, an erythropoietin, an immunization, an immunoglobulin, an immunosuppressive, a growth hormone, a hormone replacement drug, a radiopharmaceutical, an antidepressant, an antipsychotic, a stimulant, an asthma medication, a beta agonist, an inhaled steroid, an epinephrine or analog, a cytokine, and a cytokine antagonist. Further examples are: Dimebon, anti-Aβ-antibodies, beta-secretase inhibitors, tau-modulators, cognition enhancers like e.g. 5-HT6 antagonists, cholesterinase inhibitor (e.g., tactrine, donepezil, rivastigmine or galantamine) , a partial NMDA receptor blocker (e.g., memantine) , a glycosaminoglycan mimetic (e.g., Alzhemed) , an inhibitor or allosteric modulator of gamma secretase (e.g., R- flurbiprofen) , a luteinizing hormone blockade gonadotropin releasing hormone agonist (e.g., leuprorelin) , a serotinin 5- HTlA receptor antagonist, a chelatin agent, a neuronal selective L-type calcium channel blocker, an immunomodulator, an amyloid fibrillogenesis inhibitor or amyloid protein deposition inhibitor (e.g., M266) , another antibody (e.g., bapineuzumab) , a 5-HTla receptor antagonist, a PDE4 inhibitor, a histamine agonist, a receptor protein for advanced glycation end products, a PARP stimulator, a serotonin 6 receptor antagonist, a 5-HT4 receptor agonist, a human steroid, a glucose uptake stimulant which enhanced neuronal metabolism, a selective CBl antagonist, a partial agonist at benzodiazepine receptors, an amyloid beta production antagonist or inhibitor, an amyloid beta deposition inhibitor, a NNR alpha-7 partial antagonist, a therapeutic targeting PDE4, a RNA translation inhibitor, a muscarinic agonist, a nerve growth factor receptor agonist, a NGF receptor agonist and a gene therapy modulator.

85. A method for reducing human RAGE activity comprising contacting human RAGE with the product of any one of embodiments 1 to 59 such that human RAGE activity is reduced.

86. A method for decreasing hRAGE binding to at least one ligand selected from Aß peptides, globulomers, S100b and Amphoterin in a subject in need thereof, comprising the step of administering to the subject a product of any one of embodiments 1 to 59.

87. A method of treating a subject for a disorder associated with RAGE activity comprising the step of administering alone or in combination with other therapeutic agents a product of any one of embodiments 1 to 59.

88. A method for reducing RAGE activity in a subject suffering from a disorder in which RAGE activity is detrimental, comprising administering to the subject a product of any one of embodiments 1 to 59, alone or in combination with other therapeutic agents.

89. The method of embodiment 88, wherein the disorder comprises neurological diseases selected from the group comprising Amytropic Lateral Sclerosis, Brachial Plexus Injury, Brain Injury, including traumatic brain injury, Cerebral Palsy, Friedrich's Ataxia, Guillain Barre, Leukodystrophies, Multiple Sclerosis, Post Polio, Spina Bifida, Spinal Cord Injury, Spinal Muscle Atrophy, Spinal Tumors, Stroke, Transverse Myelitits, dementia, senile dementia, mild cognitive impairment, Alzheimer-related dementia, Huntington's chorea, tardive dyskinesia, hyperkinesias, manias, Morbus Parkinson, steel-Richard syndrome, Down's syndrome, myasthenia gravis, nerve trauma, vascular amyloidosis, cerebral hemorrhage I with amyloidosis, brain inflammation, Friedrich's ataxia, acute confusion disorder, amyotrophic lateral sclerosis, glaucoma, Alzheimer's disease, diabetic nephropathy, sepsis, rheumatoid arthritis and related inflammatory diseases; Diabetes and resulting complications like diabetic retinopathy, nephropathy, vascular complications; atherosclerotic complications, pulmonary fibrosis, Cancer especially melanomas, other amyloidoses

90. An isolated CDR of a binding protein as defined in any one of the embodiments 1 to 51.

91. An isolated binding protein that specifically interacts to at least one epitope of a Receptor of Advanced Glycation Endproducts (RAGE) protein.

92. The isolated binding protein of embodiment 91, wherein the isolated protein is a monoclonal antibody or antigen binding fragment thereof.

93. The monoclonal antibody or antigen binding fragment according to embodiment 92, which comprises a VH and a VL domain.

94. The monoclonal antibody according to embodiment 92 wherein said monoclonal antibody diminishes the ability of RAGE to bind to its ligands.

95. The ligands according to embodiment 94, wherein the ligands comprise Aß peptides, globulomers, S100b and amphoterin.

96. The monoclonal antibody according to embodiment 92 wherein said antibody is capable of blocking the binding of Aß globulomer to RAGE.

97. The monoclonal antibody or antigen binding fragment thereof according to embodiment 92 wherein the antibody or antigen binding fragment comprises:
a heavy chain variable region having an amino acid sequence selected from the group consisting of SEQ ID NO. 1, SEQ ID NO. 9, and SEQ ID NO. 17;
a light chain variable region having an amino acid sequence selected from the group consisting of SEQ ID NO. 5, SEQ ID NO. 13, and SEQ ID NO. 21;
a human Immunoglobulin gamma 1 heavy chain constant region with amino acid sequence SEQ ID No. 25; and
a human Immunoglobulin kappa light chain constant region with amino acid sequence SEQ ID NO 26.

98. The monoclonal antibody of embodiment 93, wherein the antigen binding domain comprises at least one complementarity determining region (CDR) comprising an amino acid sequence with at least 90% homology with the sequence selected form the group consisting of SEQ ID NOs: 2, 3, 4, 6, 7, 8, 10, 11, 12, 14, 15, 16, 18, 19, and 20.

99. The monoclonal antibody according to embodiment 93 wherein said VH domain comprises a heavy chain variable region having an amino acid sequence that has at least 90% homology with the sequence selected from the group consisting of SEQ ID NO. 1, SEQ ID NO. 9, and SEQ ID NO. 17.

100. The monoclonal antibody according to embodiment 99 wherein said VH domain comprises at least one CDR region comprising an amino acid sequence with at least 90% homology with the sequence selected form the group consisting of SEQ ID NOs: 2, 3, 4, 10, 11, 12, 18, 19, and 20.

101. The monoclonal antibody according to embodiment 100 wherein said VH domain comprises at least three CDR regions selected from the set of SEQ ID NOs: 2, 3,4; SEQ ID NOs. 10, 11, 12; SEQ ID NOs. 18, 19, and 20.

102. The monoclonal antibody according to embodiment 93 wherein said VL domain comprises a light chain variable region having an amino acid sequence that has at least 90% homology with the sequence selected from the group consisting of SEQ ID NO. 5, SEQ ID NO. 13, and SEQ ID NO. 21.

103. The monoclonal antibody according to embodiment 102 wherein said VL domain comprises at least one CDR region comprising an amino acid sequence with at least 90% homology with the sequence selected form the group consisting of SEQ ID NOs: 6, 7, 8, 14, 15, 16, 22, 23, and 24.

104. The monoclonal antibody according to embodiment 103 wherein said VL domain comprises at least three CDR regions selected from the set of SEQ ID NOs: 6, 7, 8; SEQ ID NOs. 14, 15, 16; SEQ ID NOs. 22, 23, and 24.

105. The antibody or antigen-binding fragment of embodiment 92, wherein said antibody or antigen-binding fragment is a mouse antibody, a humanized antibody, a fully human, a chimeric antibody, an antigen-binding fragment of a humanized antibody, or an antigen-binding fragment of a chimeric antibody.

106. The antibody or antigen-binding fragment of embodiment 92, wherein said antibody or antigen-binding fragment is an antigen-binding fragment selected from the group consisting of a Fab fragment, a Fab' fragment, a $F(ab')_2$ fragment and a Fv fragment.

107. A hybridoma cell line that produces a monoclonal antibody or antigen-binding fragment thereof according to embodiment 96.

108. The hybridoma cell line of embodiment 107, wherein the hybridoma is selected from the group consisting of mouse, human, rat, sheep, pig, cattle, goat, and horse hybridoma.

109. A hybridoma cell line that produces a monoclonal antibody, which specifically binds to at least one epitope of a RAGE protein.

110. The hybridoma cell line of embodiment 107, wherein the hybridoma is selected from the group consisting of mouse and human hybridoma.

111. The hybridoma cell line of embodiment 107, wherein the hybridoma is selected from the group consisting of rat, sheep, pig, cattle, goat, and horse hybridoma.

112. A vector comprising the isolated nucleic acid comprising the isolated nucleic acid that encodes any of the amino acid sequences of embodiment 97, wherein said vector is selected form the group consisting of pcDNA; pTT; pTT3; pEFBOS; pBV; pJV; and pBJ.

113. A host cell transformed with the vector according to embodiment 112, wherein the host cell is selected form the group consisting of protist cell, animal cell, plant cell and fungal cell.

114. The host cell of embodiment 113 wherein the animal cell is a mammalian cell selected form the group comprising HEK293, CHO and COS.

115. A method of producing the isolated binding protein according to embodiment 91, comprising culturing a host cell in a culture medium under conditions sufficient to produce the binding protein, collecting the culture media, and purifying the produced isolated binding protein.

116. A pharmaceutical composition comprising the monoclonal antibody or antigen-binding portion according to any of embodiments 99 or 102 and a pharmaceutically acceptable carrier.

117. A method of treating a disease or disorder comprising administering the monoclonal antibodies of embodiments 99 or 102 that bind to the C2-domain in RAGE.

118. The method of embodiment 117 wherein the disorder comprises neurological diseases selected from the group comprising Amytropic Lateral Sclerosis, Brachial Plexus Injury, Brain Injury, including traumatic brain injury, Cerebral Palsy, Friedrich's Ataxia, Guillain Barre, Leukodystrophies, Multiple Sclerosis, Post Polio, Spina Bifida, Spinal Cord Injury, Spinal Muscle Atrophy, Spinal Tumors, Stroke, Transverse Myelitits, dementia, senile dementia, mild cognitive impairment, Alzheimer-related dementia, Huntington's chorea, tardive dyskinesia, hyperkinesias, manias, Morbus Parkinson, steel-Richard syndrome, Down's syndrome, myasthenia gravis, nerve trauma, vascular amyloidosis, cerebral hemorrhage I with amyloidosis, brain inflammation, Friedrich's ataxia, acute confusion disorder, amyotrophic lateral sclerosis, glaucoma, Alzheimer's disease, diabetic nephropathy, sepsis, rheumatoid arthritis and related inflammatory diseases.

119. The antibody of embodiment 105, comprising at least one VH region comprising an amino acid sequence selected from SEQ ID NO: 56 and 57.

120. The antibody of embodiment 105, comprising at least one VL region comprising an amino acid sequence selected from SEQ ID NO: 58 and 59.

121. The antibody of embodiment 119 or 120, additionally modified by 1 to 10 mutations in an VH or VL sequence.

122. The antibody of embodiment 121, wherein the mutations are selected from framework back mutations and mutations of Vernier and VH/VL interfacing residues.

123. An antibody or binding protein of one of the preceding embodiments which inibits the binding of RAGE to an HMGB1-CpG DNA complex; or an antibody or binding protein of one of the preceding embodiments which does not inhibit the binding of RAGE to an HMGB1-CpG DNA complex.

3. Generation of anti-RAGE antibodies

3.1. General

Antibodies of the application can be generated by immunization of a suitable host (e.g., vertebrates, including humans, mice, rats, sheep, goats, pigs, cattle, horses, reptiles, fishes, amphibians, and in eggs of birds, reptiles and fish). To generate the antibodies of the present application, the host is immunized with an immunogenic RAGE polypeptide or fragment thereof of the invention. The term "immunization" refers herein to the process of presenting an antigen to an immune repertoire whether that repertoire exists in a natural genetically unaltered organism, or a transgenic organism, including those modified to display an artificial hum5an immune repertoire. Similarly, an "immunogenic preparation" is a formulation of antigen that contains adjuvants or other additives that would enhance the immunogenicity of the antigen.

Immunization of animals may be done by any method known in the art. See, e.g., Harlow and Lane, Antibodies: A Laboratory Manual, New York: Cold Spring Harbor Press, 1990. Methods for immunizing non- human animals such as mice, rats,

sheep, goats, pigs, cattle and horses are well known in the art. See, e.g., Harlow and Lane and U.S. Pat. No. 5, 994,619. In a particular embodiment, the RAGE antigen is administered with an adjuvant to stimulate the immune response. Such adjuvants include complete or incomplete Freund's adjuvant, RIBI (muramyl dipeptides) or ISCOM (immunostimulating complexes). Such adjuvants may protect the polypeptide from rapid dispersal by sequestering it in a local deposit, or they may contain substances that stimulate the host to secrete factors that are chemotactic for macrophages and other components of the immune system. Particularly, if a polypeptide is being administered, the immunization schedule will involve two or more administrations of the polypeptide, spread out over several weeks.

It is contemplated that the animal host is immunized with the antigen associated with the cell membrane of an intact or disrupted cell and antibodies of the present application are identified by binding to an immunogenic polypeptide of the invention. After immunization of the animal host with the antigen, antibodies may be obtained from the animal. The antibody-containing serum is obtained from the animal by bleeding or sacrificing the animal. The serum may be used as it is obtained from the animal, an immunoglobulin fraction may be obtained from the serum, or the antibodies may be purified from the serum. Serum or immunoglobulins obtained in this manner are polyclonal, thus having a heterogeneous array of properties.

3.2 Anti-RAGE monoclonal antibodies using Hybridoma technology

Monoclonal antibodies can be prepared using a wide variety of techniques known in the art including the use of hybridoma, recombinant, and phage display technologies, or a combination thereof. For example, monoclonal antibodies can be produced using hybridoma techniques including those known in the art and taught, for example, in Harlow et al. , Antibodies: A Laboratory Manual, (Cold Spring Harbor Laboratory Press, 2nd ed. 1988); Hammerling, et al., in: Monoclonal Antibodies and T-Cell Hybridomas 563-681 (Elsevier, N.Y., 1981) (said references incorporated by reference in their entireties). The term "monoclonal antibody" as used herein is not limited to antibodies produced through hybridoma technology. The term "monoclonal antibody" refers to an antibody that is derived from a single clone, including any eukaryotic, prokaryotic, or phage clone, and not the method by which it is produced.

Methods for producing and screening for specific antibodies using hybridoma technology are routine and well known in the art. In one embodiment,

the present invention provides methods of generating monoclonal antibodies as well as antibodies produced by the method comprising culturing a hybridoma cell secreting an antibody of the invention wherein, particularly, the hybridoma is generated by fusing splenocytes isolated from a mouse immunized with an antigen of the invention with myeloma cells and then screening the hybridomas resulting from the fusion for hybridoma clones that secrete an antibody able to bind a polypeptide of the invention. Briefly, mice can be immunized with an RAGE antigen. In a particular embodiment, the RAGE antigen is administered with a adjuvant to stimulate the immune response. Such adjuvants include complete or incomplete Freund's adjuvant, RIBI (muramyl dipeptides) or ISCOM (immunostimulating complexes). Such adjuvants may protect the polypeptide from rapid dispersal by sequestering it in a local deposit, or they may contain substances that stimulate the host to secrete factors that are chemotactic for macrophages and other components of the immune system. Particularly, if a polypeptide is being administered, the immunization schedule will involve two or more administrations of the polypeptide, spread out over several weeks.

Once an immune response is detected, e.g., antibodies specific for the antigen RAGE are detected in the mouse serum, the mouse spleen is harvested and splenocytes isolated. The splenocytes are then fused by well-known techniques to any suitable myeloma cells, for example cells from cell line SP20 available from the ATCC. Hybridomas are selected and cloned by limited dilution. The hybridoma clones are then assayed by methods known in the art for cells that secrete antibodies capable of binding RAGE. Ascites fluid, which generally contains high levels of antibodies, can be generated by immunizing mice with positive hybridoma clones.

In another embodiment, antibody-producing immortalized hybridomas may be prepared from the immunized animal. After immunization, the animal is sacrificed and the splenic B cells are fused to immortalized myeloma cells as is well known in the art. See, e.g., Harlow and Lane, *supra.* In a particular embodiment, the myeloma cells do not secrete immunoglobulin polypeptides (a non-secretory cell line). After fusion and antibiotic selection, the hybridomas are screened using RAGE or a portion thereof, or a cell expressing RAGE. In a particular embodiment, the initial screening is performed using an enzyme-linked immunoassay (ELISA) or a radioimmunoassay (RIA), particularlyan ELISA. An example of ELISA screening is provided in WO 00/37504, herein incorporated by reference.

Anti- RAGE antibody-producing hybridomas are selected, cloned and further screened for desirable characteristics, including robust hybridoma growth, high antibody production and desirable antibody characteristics, as discussed further below. Hybridomas may be cultured and expanded *in vivo* in syngeneic animals, in animals that lack an immune system, e.g., nude mice, or in cell culture *in vitro*. Methods of selecting, cloning and expanding hybridomas are well known to those of ordinary skill in the art.

In a particular embodiment, the hybridomas are mouse hybridomas, as described above. In another particular embodiment, the hybridomas are produced in a non-human, non-mouse species such as rats, sheep, pigs, goats, cattle or horses. In another embodiment, the hybridomas are human hybridomas, in which a human non-secretory myeloma is fused with a human cell expressing an anti- RAGE antibody.

Antibody fragments that recognize specific epitopes may be generated by known techniques. For example, Fab and F(ab')2 fragments of the invention may be produced by proteolytic cleavage of immunoglobulin molecules, using enzymes such as papain (to produce Fab fragments) or pepsin (to produce F(ab')2 fragments). F(ab')2 fragments contain the variable region, the light chain constant region and the CHI domain of the heavy chain.

3.3 Anti- RAGE monoclonal antibodies using SLAM

In another aspect of the invention, recombinant antibodies are generated from single, isolated lymphocytes using a procedure referred to in the art as the selected lymphocyte antibody method (SLAM), as described in U.S. Patent No. 5,627,052, PCT Publication WO 92/02551 and Babcock, J.S. *et al.* (1996) *Proc. Natl. Acad. Sci. USA* 93:7843-7848. In this method, single cells secreting antibodies of interest, *e.g.*, lymphocytes derived from any one of the immunized animals described above, are screened using an antigen-specific hemolytic plaque assay, wherein the antigen RAGE, a subunit of RAGE, or a fragment thereof, is coupled to sheep red blood cells using a linker, such as biotin, and used to identify single cells that secrete antibodies with specificity for RAGE. Following identification of antibody-secreting cells of interest, heavy- and light-chain variable region cDNAs are rescued from the cells by reverse transcriptase-PCR and these variable regions can then be expressed, in the context of appropriate immunoglobulin constant regions (*e.g.*, human constant regions), in mammalian host cells, such as COS or CHO cells. The host cells

transfected with the amplified immunoglobulin sequences, derived from *in vivo* selected lymphocytes, can then undergo further analysis and selection *in vitro*, for example by panning the transfected cells to isolate cells expressing antibodies to RAGE. The amplified immunoglobulin sequences further can be manipulated *in vitro*, such as by *in vitro* affinity maturation methods such as those described in PCT Publication WO 97/29131 and PCT Publication WO 00/56772.

3.4 Anti- RAGE monoclonal antibodies using transgenic animals

In another embodiment of the instant invention, antibodies are produced by immunizing a non-human animal comprising some, or all, of the human immunoglobulin locus with an RAGE antigen. In a particular embodiment, the non-human animal is a XENOMOUSE transgenic mouse, an engineered mouse strain that comprises large fragments of the human immunoglobulin loci and is deficient in mouse antibody production. *See, e.g.,* Green et al. *Nature Genetics* 7:13-21 (1994) and United States Patents 5,916,771, 5,939,598, 5,985,615, 5,998,209, 6,075,181, 6,091,001, 6,114,598 and 6,130,364. See also WO 91/10741, published July 25,1991, WO 94/02602, published February 3, 1994, WO 96/34096 and WO 96/33735, both published October 31, 1996, WO 98/16654, published April 23, 1998, WO 98/24893, published June 11, 1998, WO 98/50433, published November 12, 1998, WO 99/45031, published September 10, 1999, WO 99/53049, published October 21, 1999, WO 00 09560, published February 24, 2000 and WO 00/037504, published June 29, 2000. The XENOMOUSE transgenic mouse produces an adult-like human repertoire of fully human antibodies, and generates antigen-specific human Mabs. The XENOMOUSE transgenic mouse contains approximately 80% of the human antibody repertoire through introduction of megabase sized, germline configuration YAC fragments of the human heavy chain loci and x light chain loci. See Mendez et al., *Nature Genetics* 15:146-156 (1997), Green and Jakobovits *J. Exp. Med.* 188:483-495 (1998), the disclosures of which are hereby incorporated by reference.

3.5 Anti- RAGE monoclonal antibodies using recombinant antibody libraries

In vitro methods also can be used to make the antibodies of the invention, wherein an antibody library is screened to identify an antibody having the desired binding specificity. Methods for such screening of recombinant antibody libraries are

well known in the art and include methods described in, for example, Ladner *et al.* U.S. Patent No. 5,223,409; Kang *et al.* PCT Publication No. WO 92/18619; Dower *et al.* PCT Publication No. WO 91/17271; Winter *et al.* PCT Publication No. WO 92/20791; Markland *et al.* PCT Publication No. WO 92/15679; Breitling *et al.* PCT Publication No. WO 93/01288; McCafferty *et al.* PCT Publication No. WO 92/01047; Garrard *et al.* PCT Publication No. WO 92/09690; Fuchs *et al.* (1991) *Bio/Technology* 9:1370-1372; Hay *et al.* (1992) *Hum Antibod Hybridomas* 3:81-85; Huse *et al.* (1989) *Science* 246:1275-1281; McCafferty *et al.*, *Nature* (1990) 348:552-554; Griffiths *et al.* (1993) *EMBO J* 12:725-734; Hawkins *et al.* (1992) *J Mol Biol* 226:889-896; Clackson *et al.* (1991) *Nature* 352:624-628; Gram *et al.* (1992) *PNAS* 89:3576-3580; Garrad *et al.* (1991) *Bio/Technology* 9:1373-1377; Hoogenboom *et al.* (1991) *Nuc Acid Res* 19:4133-4137; and Barbas *et al.* (1991) *PNAS* 88:7978-7982, US patent application publication 20030186374, and PCT Publication No. WO 97/29131, the contents of each of which are incorporated herein by reference.

The recombinant antibody library may be from a subject immunized with RAGE, or a portion of RAGE. Alternatively, the recombinant antibody library may be from a naïve subject, i.e., one who has not been immunized with RAGE, such as a human antibody library from a human subject who has not been immunized with human RAGE. Antibodies of the invention are selected by screening the recombinant antibody library with the peptide comprising human RAGE to thereby select those antibodies that recognize RAGE. Methods for conducting such screening and selection are well known in the art, such as described in the references in the preceding paragraph. To select antibodies of the invention having particular binding affinities for hRAGE, such as those that dissociate from human RAGE with a particular k_{off} rate constant, the art-known method of surface plasmon resonance can be used to select antibodies having the desired k_{off} rate constant. To select antibodies of the invention having a particular neutralizing activity for hRAGE , such as those with a particular an IC_{50}, standard methods known in the art for assessing the inhibition of hRAGE activity may be used.

In one aspect, the invention pertains to an isolated antibody, or an antigen-binding portion thereof, that binds human RAGE. Particularly, the antibody is a neutralizing antibody. In various embodiments, the antibody is a recombinant antibody or a monoclonal antibody.

For example, the antibodies of the present invention can also be generated using various phage display methods known in the art. In phage display methods, functional antibody domains are displayed on the surface of phage particles, which carry the polynucleotide sequences encoding them. In a particular, such phage can be utilized to display antigen-binding domains expressed from a repertoire or combinatorial antibody library (e. g., human or murine). Phage expressing an antigen binding domain that binds the antigen of interest can be selected or identified with antigen, e.g., using labeled antigen or antigen bound or captured to a solid surface or bead. Phage used in these methods are typically filamentous phage including fd and M13 binding domains expressed from phage with Fab, Fv or disulfide stabilized Fv antibody domains recombinantly fused to either the phage gene III or gene VIII protein. Examples of phage display methods that can be used to make the antibodies of the present invention include those disclosed in Brinkman et al., J. Immunol. Methods 182:41-50 (1995); Ames et al., J. Immunol. Methods 184:177-186 (1995); Kettleborough et al., Eur. J. Immunol. 24:952-958 (1994); Persic et al., Gene 187 9-18 (1997); Burton et al., Advances in Immunology 57:191-280 (1994); PCT application No. PCT/GB91/01134; PCT publications WO 90/02809; WO 91/10737; WO 92/01047; WO 92/18619; WO 93/11236; WO 95/15982; WO 95/20401; and U.S. Pat. Nos. 5,698,426; 5,223,409; 5,403,484; 5,580,717; 5,427,908; 5,750,753; 5,821,047; 5,571,698; 5,427,908; 5,516,637; 5,780, 225; 5,658,727; 5,733,743 and 5,969,108; each of which is incorporated herein by reference in its entirety.

As described in the above references, after phage selection, the antibody coding regions from the phage can be isolated and used to generate whole antibodies including human antibodies or any other desired antigen binding fragment, and expressed in any desired host, including mammalian cells, insect cells, plant cells, yeast, and bacteria, e.g., as described in detail below. For example, techniques to recombinantly produce Fab, Fab' and $F(ab')_2$ fragments can also be employed using methods known in the art such as those disclosed in PCT publication WO 92/22324; Mullinax et al., BioTechniques 12(6):864-869 (1992); and Sawai et al., AJRI 34:26-34 (1995); and Better et al., Science 240:1041-1043 (1988) (said references incorporated by reference in their entireties). Examples of techniques, which can be used to produce single-chain Fvs and antibodies include those described in U.S. Pat. 4,946,778 and 5,258, 498; Huston et al., Methods in Enzymology 203:46-88 (1991);

Shu et al., PNAS 90:7995-7999 (1993); and Skerra et al., Science 240:1038-1040 (1988).

Alternative to screening of recombinant antibody libraries by phage display, other methodologies known in the art for screening large combinatorial libraries can be applied to the identification of dual specificity antibodies of the invention. One type of alternative expression system is one in which the recombinant antibody library is expressed as RNA-protein fusions, as described in PCT Publication No. WO 98/31700 by Szostak and Roberts, and in Roberts, R.W. and Szostak, J.W. (1997) *Proc. Natl. Acad. Sci. USA* 94:12297-12302. In this system, a covalent fusion is created between an mRNA and the peptide or protein that it encodes by *in vitro* translation of synthetic mRNAs that carry puromycin, a peptidyl acceptor antibiotic, at their 3' end. Thus, a specific mRNA can be enriched from a complex mixture of mRNAs (*e.g.*, a combinatorial library) based on the properties of the encoded peptide or protein, *e.g.*, antibody, or portion thereof, such as binding of the antibody, or portion thereof, to the dual specificity antigen. Nucleic acid sequences encoding antibodies, or portions thereof, recovered from screening of such libraries can be expressed by recombinant means as described above (*e.g.*, in mammalian host cells) and, moreover, can be subjected to further affinity maturation by either additional rounds of screening of mRNA-peptide fusions in which mutations have been introduced into the originally selected sequence(s), or by other methods for affinity maturation *in vitro* of recombinant antibodies, as described above.

In another approach the antibodies of the present invention can also be generated using yeast display methods known in the art. In yeast display methods, genetic methods are used to tether antibody domains to the yeast cell wall and display them on the surface of yeast. In particular, such yeast can be utilized to display antigen-binding domains expressed from a repertoire or combinatorial antibody library (e. g., human or murine). Examples of yeast display methods that can be used to make the antibodies of the present invention include those disclosed Wittrup, et al. U.S. Patent No. 6,699,658 incorporated herein by reference.

4. Production of particular recombinant RAGE antibodies of the invention

Antibodies of the present invention may be produced by any of a number of techniques known in the art. For example, expression from host cells, wherein expression vector(s) encoding the heavy and light chains is (are) transfected into a

host cell by standard techniques. The various forms of the term "transfection" are intended to encompass a wide variety of techniques commonly used for the introduction of exogenous DNA into a prokaryotic or eukaryotic host cell, *e.g.*, electroporation, calcium-phosphate precipitation, DEAE-dextran transfection and the like. Although it is possible to express the antibodies of the invention in either prokaryotic or eukaryotic host cells, expression of antibodies in eukaryotic cells is preferable, and most preferable in mammalian host cells, because such eukaryotic cells (and in particular mammalian cells) are more likely than prokaryotic cells to assemble and secrete a properly folded and immunologically active antibody.

Particular mammalian host cells for expressing the recombinant antibodies of the invention include Chinese Hamster Ovary (CHO cells) (including dhfr- CHO cells, described in Urlaub and Chasin, (1980) *Proc. Natl. Acad. Sci. USA* 77:4216-4220, used with a DHFR selectable marker, *e.g.*, as described in R.J. Kaufman and P.A. Sharp (1982) *Mol. Biol.* 159:601-621), NS0 myeloma cells, COS cells and SP2 cells. When recombinant expression vectors encoding antibody genes are introduced into mammalian host cells, the antibodies are produced by culturing the host cells for a period of time sufficient to allow for expression of the antibody in the host cells or, in particular, secretion of the antibody into the culture medium in which the host cells are grown. Antibodies can be recovered from the culture medium using standard protein purification methods.

Host cells can also be used to produce functional antibody fragments, such as Fab fragments or scFv molecules. It will be understood that variations on the above procedure are within the scope of the present invention. For example, it may be desirable to transfect a host cell with DNA encoding functional fragments of either the light chain and/or the heavy chain of an antibody of this invention. Recombinant DNA technology may also be used to remove some, or all, of the DNA encoding either or both of the light and heavy chains that is not necessary for binding to the antigens of interest. The molecules expressed from such truncated DNA molecules are also encompassed by the antibodies of the invention. In addition, bifunctional antibodies may be produced in which one heavy and one light chain are an antibody of the invention and the other heavy and light chain are specific for an antigen other than the antigens of interest by crosslinking an antibody of the invention to a second antibody by standard chemical crosslinking methods.

In a particular system for recombinant expression of an antibody, or antigen-binding portion thereof, of the invention, a recombinant expression vector encoding both the antibody heavy chain and the antibody light chain is introduced into dhfr-CHO cells by calcium phosphate-mediated transfection. Within the recombinant expression vector, the antibody heavy and light chain genes are each operatively linked to CMV enhancer/AdMLP promoter regulatory elements to drive high levels of transcription of the genes. The recombinant expression vector also carries a DHFR gene, which allows for selection of CHO cells that have been transfected with the vector using methotrexate selection/amplification. The selected transformant host cells are cultured to allow for expression of the antibody heavy and light chains and intact antibody is recovered from the culture medium. Standard molecular biology techniques are used to prepare the recombinant expression vector, transfect the host cells, select for transformants, culture the host cells and recover the antibody from the culture medium. Still further the invention provides a method of synthesizing a recombinant antibody of the invention by culturing a host cell of the invention in a suitable culture medium until a recombinant antibody of the invention is synthesized. The method can further comprise isolating the recombinant antibody from the culture medium.

4.1 Anti RAGE antibodies

Table 4 is a list of amino acid sequences of VH and VL regions of particular anti-hRAGE antibodies of the invention.

Table 4: List of Amino Acid Sequences of VH and VL regions of anti-huRAGE antibodies

Seq. ID No.	Protein Region		Sequence
			12345678901234567890123456789 01
1	**VH 7F9**		EEKLEESGGGLVQLGGSMKISCVASGFTLS**N YWMD**WVRQSPEKGLEWIA**EIRLKSNYYSTHY AESVKG**RFSISRDDSKGSVSLQMDNLTAEDT GIYFCAR**NAYWYFDV**WGTGTTVTVSS
	VH 7F9 CDR-H1	Residues 31-35 of SEQ ID NO.:1	**NYWMD**

Seq. ID No.	Protein Region		Sequence
			123456789012345678901234567890 1
	VH 7F9 CDR-H2	Residues 50-68 of SEQ ID NO.:1	**EIRLKSNYYSTHYAESVKG**
	VH 7F9 CDR-H3	Residues 101-108 of SEQ ID NO.:1	**NAYWYFDV**
5	**VL 7F9**		DIVMTQSHKFMSTSVGDRVSATC**KASQDVGT SVA**WYQQKLGQSPKLLIY**WTSTRHT**GVPDRF TGSGSGTDFTLTISNVQSEDLADYFC**QQYNN YPLT**FGDGTKLELKR
	VL 7F9 CDR-L1	Residues 24-34 of SEQ ID NO.:5	**KASQDVGTSVA**
	VL 7F9 CDR-L2	Residues50-56 of SEQ ID NO.:5	**WTSTRHT**
	VL 7F9 CDR-L3	Residues89-97 of SEQ ID NO.:5	**QQYNNYPLT**
9	**VH 11E6**		QIQLVQSGPELKKPGETVKISCKASGYTFT**N FGMN**WVKQAPGKGLKWMG**YINTNTGESIYSE EFKG**RFAFSLETSASTAYLQINNLKNEDTAT YFCAR**SRMVTAYGMDY**WGQGTSVTVSS
	VH 11E6 CDR-H1	Residues 31-35 of SEQ ID NO.:9	**NFGMN**
	VH 11E6 CDR-H2	Residues 50-66 of SEQ ID NO.:9	**YINTNTGESIYSEEFKG**
	VH 11E6 CDR-H3	Residues 99-109 of SEQ ID NO.:9	**SRMVTAYGMDY**
13	**VL 11E6**		DIVMTQSQKFMSTSVGDRVSITC**KASQNVGT AVA**WYQQRPGQSPKLLIF**SASNRYT**GVPDRF TGSGSGTDFTLTLSNMQPEDLADYFC**QQYSS YPLT**FGVGTKLELKR
	VL 11E6 CDR-L1	Residues24-34 of SEQ ID NO.:13	**KASQNVGTAVA**
	VL 11E6 CDR-L2	Residues50-56 of SEQ ID NO.:13	**SASNRYT**
	VL 11E6 CDR-L3	Residues89-97 of SEQ ID NO.:13	**QQYSSYPLT**

Seq. ID No.	Protein Region		Sequence
			1234567890**123456789**0**123456789**0**1**
17	**VH 4E5**		QVQLQQSGAELVRPGTSVKVSCKASGYAFN**N YLIE**WIKQRPGQGLEWIG**VINPGSGGTNHNE KFKV**KATLTADKSSSTAYIQLSSLTSDDSAV YFCAR**SAGTARARFAY**WGQGTLVTVSA
	VH 4E5 CDR-H1	Residues 31-35 of SEQ ID NO.:17	**NYLIE**
	VH 4E5 CDR-H2	Residues 50-66 of SEQ ID NO.:17	**VINPGSGGTNHNEKFKV**
	VH 4E5 CDR-H3	Residues 99-109 of SEQ ID NO.:17	**SAGTARARFAY**
21	**VL 4E5**		DIQMTQSPSSLSASLGERVSLTC**RASQDIGS SLN**WLQQEPDGTIKRLIY**ATSSLDS**GVPKRF SGSRSGSDYSLTISSLESEDFVDYYC**LQYAS FPFT**FGSGTKLEIKR
	VL 4E5 CDR-L1	Residues24-34 of SEQ ID NO.:21	**RASQDIGSSLN**
	VL 4E5 CDR-L2	Residues50-56 of SEQ ID NO.:21	**ATSSLDS**
	VL 4E5 CDR-L3	Residues89-97 of SEQ ID NO.:21	**LQYASFPFT**

The foregoing isolated anti-RAGE antibody CDR sequences establish a novel family of RAGE binding proteins, isolated in accordance with this invention. To generate and to select CDR's of the invention having particular RAGE binding and/or neutralizing activity with respect to hRAGE, standard methods known in the art for generating binding proteins of the present invention and assessing the RAGE binding and/or neutralizing characteristics of those binding protein may be used, including but not limited to those specifically described herein.

4.2 Anti RAGE Chimeric antibodies

A chimeric antibody is a molecule in which different portions of the antibody are derived from different animal species, such as antibodies having a variable region derived from a murine monoclonal antibody and a human immunoglobulin constant region. Methods for producing chimeric antibodies are known in the art. See e.g.,

Morrison, Science 229:1202 (1985); Oi et al., BioTechniques 4:214 (1986); Gillies et al., (1989) J. Immunol. Methods 125:191-202; U.S. Pat. Nos. 5,807,715; 4,816,567; and 4,816,397, which are incorporated herein by reference in their entireties. In addition, techniques developed for the production of "chimeric antibodies" (Morrison et al., 1984, Proc. Natl. Acad. Sci. 81:851-855; Neuberger et al., 1984, Nature 312:604-608; Takeda et al., 1985, Nature 314:452-454 which are incorporated herein by reference in their entireties) by splicing genes from a mouse antibody molecule of appropriate antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used.

In one embodiment, the chimeric antibodies of the invention are produced by replacing the heavy chain constant region of the murine monoclonal anti human RAGE antibodies described herein with a human IgG1 constant region.

4.3 Anti RAGE CDR grafted antibodies

CDR-grafted antibodies of the invention comprise heavy and light chain variable region sequences from a human antibody wherein one or more of the CDR regions of V_H and/or V_L are replaced with CDR sequences of non- human, as for example, murine antibodies of the invention. A framework sequence from any human antibody may serve as the template for CDR grafting. However, straight chain replacement onto such a framework often leads to some loss of binding affinity to the antigen. The more homologous a human antibody is to the original murine antibody, the less likely the possibility that combining the murine CDRs with the human framework will introduce distortions in the CDRs that could reduce affinity. Therefore, it is preferable that the human variable framework that is chosen to replace the murine variable framework apart from the CDRs have at least a 65% sequence identity with the murine antibody variable region framework. It is more preferable that the human and murine variable regions apart from the CDRs have at least 70% sequence identify. It is even more preferable that the human and murine variable regions apart from the CDRs have at least 75% sequence identity. It is most preferable that the human and murine variable regions apart from the CDRs have at least 80% sequence identity. Methods for producing CDR-grafted antibodies are known in the art (Jones et al., Nature 321:522-525 (1986); U.S. Pat. Nos. 5,225,539).

In a specific embodiment the invention provides CDR grafted antibodies with V_H and/or V_L chains as described in Table 5.

Table 5: CDR Grafted antibodies

SEQ ID No.	Protein region	Sequence
		12345678901234567890123456789 0
56 (43) (44) (45) (46)	**VH 11E6.1-GL** (VH7-4.1/JH6 FR1) (VH7-4.1/JH6 FR2) (VH7-4.1/JH6 FR3) (VH7-4.1/JH6 FR4)	QVQLVQSGSELKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RFVFSLDTSVSTAYLQICSLKAED TAVYYCAR**SRMVTAYGMDY**WGQGTTVTVSS
57 (47) (44) (48) (46)	**VH 11E6.2-GL** (VH1-2/JH6 FR1) (VH1-2/JH6 FR2) (VH1-2/JH6 FR3) (VH1-2/JH6 FR4)	QVQLVQSGAEVKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RVTMTRDTSISTAYMELSRLRSDD TAVYYCAR**SRMVTAYGMDY**WGQGTTVTVSS
58 (49) (50) (51) (52)	**VL 11E6.1-GL** (1-12/L5/JK2 FR1) (1-12/L5/JK2 FR2) (1-12/L5/JK2 FR3) (1-12/L5/JK2 FR4)	DIQMTQSPSSVSASVGDRVTITC**KASQNVG** **TAVA**WYQQKPGKAPKLLIY**SASNRYT**GVPS RFSGSGSGTDFTLTISSLQPEDFATYYC**QQ** **YSSYPLT**FGQGTKLEIKR
59 (53) (54) (55) (52)	**VL 11E6.2-GL** (3-15/L2/JK2 FR1) (3-15/L2/JK2 FR2) (3-15/L2/JK2 FR3) (3-15/L2/JK2 FR4)	EIVMTQSPATLSVSPGERATLSC**KASQNVG** **TAVA**WYQQKPGQAPRLLIY**SASNRYT**GIPA RFSGSGSGTEFTLTISSLQSEDFAVYYC**QQ** **YSSYPLT**FGQGTKLEIKR

CDR sequences derived from mAb 11E6 are stated in bold letters. Reference is also made to the specific framework sequences (FR1 to FR4) by stating the corresponding SEQ ID NOs (see also Tables 2 and 3)

4.4 Anti RAGE Humanized antibodies

Humanized antibodies are antibody molecules from non-human species antibody that binds the desired antigen having one or more complementarity determining regions (CDRs) from the non-human species and framework regions from a human immunoglobulin molecule. Known human Ig sequences are disclosed, e.g., www.ncbi.nlm.nih.gov/entrez- /query.fcgi; www.atcc.org/phage/hdb.html; www.sciquest.com/; www.abcam.com/; www.antibodyresource.com/onlinecomp.html; www.public.iastate.edu/.about.pedro/research_tools.html; www.mgen.uni-heidelberg.de/SD/IT/IT.html; www.whfreeman.com/immunology/CH-05/kuby05.htm; www.library.thinkquest.org/12429/Immune/Antibody.html;

www.hhmi.org/grants/lectures/1996/vlab/; www.path.cam.ac.uk/.about.mrc7/m-ikeimages.html; www.antibodyresource.com/; mcb.harvard.edu/BioLinks/Immunology.html.www.immunologylink.com/; pathbox.wustl.edu/.about.hcenter/index.-html; www.biotech.ufl.edu/.about.hcl/; www.pebio.com/pa/340913/340913.html- ; www.nal.usda.gov/awic/pubs/antibody/; www.m.ehime-u.acjp/.about.yasuhito-/Elisa.html; www.biodesign.com/table.asp; www.icnet.uk/axp/facs/davies/links.html; www.biotech.ufl.edu/.about.fccl/protocol.html; www.isac-net.org/sites_geo.html; aximtl.imt.uni-marburg.de/.about.rek/AEP- Start.html; baserv.uci.kun.nl/.about.jraats/linksl.html; www.recab.uni-hd.de/immuno.bme.nwu.edu/; www.mrc-cpe.cam.ac.uk/imt-doc/public/INTRO.html; www.ibt.unam.mx/vir/V_mice.html; imgt.cnusc.fr:8104/; www.biochem.ucl.ac.uk/.about.martin/abs/index.html; antibody.bath.ac.uk/; abgen.cvm.tamu.edu/lab/wwwabgen.html; www.unizh.ch/.about.honegger/AHOseminar/Slide01.html; www.cryst.bbk.ac.uk/.about.ubcg07s/; www.nimr.mrc.ac.uk/CC/ccaewg/ccaewg.htm; www.path.cam.ac.uk/.about.mrc7/humanisation/TAHHP.html; www.ibt.unam.mx/vir/structure/stat_aim.html; www.biosci.missouri.edu/smithgp/index.html; www.cryst.bioc.cam.ac.uk/.about.fmolina/Web-pages/Pept/spottech.html; www.jerini.de/fr roducts.htm; www.patents.ibm.com/ibm.html.Kabat et al., Sequences of Proteins of Immunological Interest, U.S. Dept. Health (1983), each entirely incorporated herein by reference. Such imported sequences can be used to reduce immunogenicity or reduce, enhance or modify binding, affinity, on-rate, off-rate, avidity, specificity, half-life, or any other suitable characteristic, as known in the art.

Framework residues in the human framework regions may be substituted with the corresponding residue from the CDR donor antibody to alter, particularly improve, antigen binding. These framework substitutions are identified by methods well known in the art, e.g., by modeling of the interactions of the CDR and framework residues to identify framework residues important for antigen binding and sequence comparison to identify unusual framework residues at particular positions. (See, e.g., Queen et al., U.S. Pat. No. 5,585,089; Riechmann et al., Nature 332:323 (1988), which are incorporated herein by reference in their entireties.) Three-dimensional immunoglobulin models are commonly available and are familiar to those skilled in the art. Computer programs are available which illustrate and display probable three-dimensional conformational structures of selected candidate immunoglobulin

sequences. Inspection of these displays permits analysis of the likely role of the residues in the functioning of the candidate immunoglobulin sequence, i.e., the analysis of residues that influence the ability of the candidate immunoglobulin to bind its antigen. In this way, FR residues can be selected and combined from the consensus and import sequences so that the desired antibody characteristic, such as increased affinity for the target antigen(s), is achieved. In general, the CDR residues are directly and most substantially involved in influencing antigen binding. Antibodies can be humanized using a variety of techniques known in the art, such as but not limited to those described in Jones et al., Nature 321:522 (1986); Verhoeyen et al., Science 239:1534 (1988)), Sims et al., J. Immunol. 151: 2296 (1993); Chothia and Lesk, J. Mol. Biol. 196:901 (1987), Carter et al., Proc. Natl. Acad. Sci. U.S.A. 89:4285 (1992); Presta et al., J. Immunol. 151:2623 (1993), Padlan, Molecular Immunology 28(4/5):489-498 (1991); Studnicka et al., Protein Engineering 7(6):805-814 (1994); Roguska. et al. , PNAS 91:969-973 (1994); PCT publication WO 91/09967, PCT/: US98/16280, US96/18978, US91/09630, US91/05939, US94/01234, GB89/01334, GB91/01134, GB92/01755; WO90/14443, WO90/14424, WO90/14430, EP 229246, EP 592,106; EP 519,596, EP 239,400, U.S. Pat. Nos. 5,565,332, 5,723,323, 5,976,862, 5,824,514, 5,817,483, 5814476, 5763192, 5723323, 5,766886, 5,714,352, 6,204,023, 6,180,370, 5,693,762, 5,530,101, 5,585,089, 5,225,539; 4,816,567, each entirely incorporated herein by reference, included references cited therein.

5. Further Embodiments of Antibodies of the Invention

5.1 Fusion Antibodies and Immunoadhesins

The present application also describes a fusion antibody or immunoadhesin that may be made which comprises all or a portion of a RAGE antibody of the present application linked to another polypeptide. In some embodiments, only the variable region of the RAGE antibody is linked to the polypeptide. In other embodiments, the VH domain of a RAGE antibody of this application is linked to a first polypeptide, while the VL domain of the antibody is linked to a second polypeptide that associates with the first polypeptide in a manner that permits the VH and VL domains to interact with one another to form an antibody binding site. In other embodiments, the VH domain is separated from the VL domain by a linker that permits the VH and VL domains to interact with one another (see below under Single Chain Antibodies). The VH -linker- VL antibody is then linked to a polypeptide of interest. The fusion

antibody is useful to directing a polypeptide to a cell or tissue that expresses a RAGE. The polypeptide of interest may be a therapeutic agent, such as a toxin, or may be a diagnostic agent, such as an enzyme; that may be easily visualized, such as horseradish peroxidase. In addition, fusion antibodies can be created in which two (or more) single-chain antibodies are linked to one another. This is useful if one wants to create a divalent or polyvalent antibody on a single polypeptide chain, or if one wants to create a bispecific antibody.

One embodiment provides a labelled binding protein wherein an antibody or antibody portion of the present application is derivatized or linked to another functional molecule (e.g., another peptide or protein). For example, a labelled binding protein of the present application can be derived by functionally linking an antibody or antibody portion of the present application (by chemical coupling, genetic fusion, noncovalent association or otherwise) to one or more other molecular entities, such as a nucleic acid, another antibody (e.g., a bispecific antibody or a diabody), a detectable agent, a cytotoxic agent, a pharmaceutical agent, and/or a protein or peptide that can mediate association of the antibody or antibody portion with another molecule (such as a streptavidin core region or a polyhistidine tag).

Useful detectable agents with which an antibody or antibody portion of the present application may be derivatized include fluorescent compounds. Exemplary fluorescent detectable agents include fluorescein, fluorescein isothiocyanate, rhodamine, 5-dimethylamine-1-napthalenesulfonyl chloride, phycoerythrin and the like. An antibody may also be derivatized with detectable enzymes, such as alkaline phosphatase, horseradish peroxidase, glucose oxidase and the like. When an antibody is derivatized with a detectable enzyme, it is detected by adding additional reagents that the enzyme uses to produce a detectable reaction product. For example, when the detectable agent horseradish peroxidase is present, the addition of hydrogen peroxide and diaminobenzidine leads to a colored reaction product, which is detectable. An antibody may also be derivatized with a nucleic acid, biotin, and detected through indirect measurement of avidin or streptavidin binding.

5.2 Single Chain Antibodies

The present application includes a single chain antibody (scFv) that binds an immunogenic RAGE of the invention. To produce the scFv, VH- and V-encoding DNA is operatively linked to DNA encoding a flexible linker, e.g., encoding the

amino acid sequence (Gly4-Ser), such that the VH and VL sequences can be expressed as a contiguous single-chain protein, with the VL and VH regions joined by the flexible linker (see e.g., Bird et al. (1988) Science 242:423-42 6; Huston et al. (1988) Proc. Natl. Acad. Sci. USA 85: 5879-5883; McCafferty et al., 30 Nature (1990) 34 8: 552- 554). The single chain antibody may be monovalent, if only a single VH and VL are used, bivalent, if two VH and VL are used, or polyvalent, if more than two VH and VL are used. Two of said scFv fragments coupled via a linker are called "diabody" which form is also encompassed by the invention.

5.3 Bispecific Antibodies

The present application further includes a bispecific antibody or antigen-binding fragment thereof in which one specificity is for an immunogenic RAGE polypeptide of the present application. For example, a bispecific antibody can be generated that specifically binds to an immunogenic RAGE polypeptide of the invention through one binding domain and to a second molecule through a second binding domainIn addition, a single chain antibody containing more than one VH and VL may be generated that binds specifically to an immunogenic polypeptide of the invention and to another molecule that is associated with attenuating myelin mediated growth cone collapse and inhibition of neurite outgrowth and sprouting. Such bispecific antibodies can be generated using techniques that are well known for example, Fanger et al. Immunol Methods 4: 72-81 (1994) and Wright and Harris, 20 (supra).

In some embodiments, the bispecific antibodies are prepared using one or more of the variable regions from an antibody of the invention. In another embodiment, the bispecific antibody is prepared using one or more CDR regions from said antibody.

5.4 Derivatized and Labeled Antibodies

An antibody or an antigen- binding fragment of the present application can be derivatized or linked to another molecule (e.g., another peptide or protein). In general, the antibody or antigen-binding fragment is derivatized such that binding to an immunogenic polypeptide of the invention is not affected adversely by the derivatization or labeling.

For example, an antibody or antibody portion of the present application can be functionally linked (by chemical coupling, genetic fusion, noncovalent association or otherwise) to one or more other molecular entities, such as another antibody (e.g., a bispecific antibody or a diabody), a detection reagent, a cytotoxic agent, a pharmaceutical agent, and/or a protein or peptide that can mediate association of the antibody or antigen-binding fragment with another molecule (such as a streptavidin core region or a polyhistidine tag). Still further, an antibody or antigen-binding portion thereof may be part of a larger immunoadhesion molecule, formed by covalent or non-covalent association of the antibody or antibody portion with one or more other or different proteins or peptides. Examples of such immunoadhesion molecules include use of the streptavidin core region to make a tetrameric scFv molecule (Kipriyanov et al. (1995) Human Antibodies and Hybridomas 6:93-101) and use of a cysteine residue, a marker peptide and a C-terminal polyhistidine tag to make bivalent and biotinylated scFv molecules (Kipriyanov et al. (1994) Molecular Immunology 31:1047-1058). Antibody portions, such as Fab and $F(ab')_2$ fragments, can be prepared from whole antibodies using conventional techniques, such as papain or pepsin digestion, respectively, of whole antibodies. Moreover, antibodies, antibody portions and immunoadhesion molecules can be obtained using standard recombinant DNA techniques.

A derivatized antibody may be produced by crosslinking two or more antibodies (of the same type or of different types, e. g., to create bispecific antibodies). Suitable crosslinkers include those that are heterobifunctional, having two distinctly reactive groups separated by an appropriate spacer (e.g. m-maleimidobenzoyl-N-hydroxysuccinimide ester) or homobifunctional (e.g., disuccinimidyl suberate). Such linkers are available from Pierce Chemical Company, Rockford, Ill.

A derivatized antibody may also be a labeled antibody. For instance, detection agents with which an antibody or antibody portion of the invention may be derivatized are fluorescent compounds, including fluorescein, fluorescein isothiocyanate, rhodamine, 5-dimethylamine-l-napthalenesulfonyl chloride, phycoerythrin, lanthanide phosphors and the like. An antibody also may be labeled with enzymes that are useful for detection, such as horseradish peroxidase, galactosidase, luciferase, alkaline phosphatase, glucoseoxidase and the like. In embodiments that are labeled with a

detectable enzyme, the antibody is detected by adding additional reagents that the enzyme uses to produce a detectable reaction product. For example, horseradish peroxidase with hydrogen peroxide and diaminobenzidine. An antibody also may be labeled with biotin, and detected through indirect measurement of avidin or streptavidin binding. An antibody may also be labeled with a predetermined polypeptide epitope recognized by a secondary reporter (e. g., leucine zipper pair sequences, binding sites for secondary antibodies, metal binding domains, epitope: tags). An RAGE antibody or an antigen fragment thereof also may be labeled with a radio-labeled amino acid. The radiolabel may be used for both diagnostic and therapeutic purposes. The radio-labeled RAGE antibody may be used diagnostically, for example, for determining RAGE receptor levels in a subject. Further, the radio-labeled RAGE antibody may be used therapeutically for treating spinal cord injury.

Examples of labels for polypeptides include, but are not limited to, the following radioisotopes or radionucleotides ^{15}N, ^{35}S, ^{90}Y, ^{99}Tc, ^{111}In, ^{125}I, ^{131}I, ^{177}Lu, ^{166}Ho, ^{153}Sm. A RAGE antibody or an antigen fragment thereof may also be derivatized with a chemical group such as polyethylene glycol (PEG), a methyl or ethyl group, or a carbohydrate group. These groups may be useful to improve the biological characteristics of the antibody, e.g., to increase serum half-life or to increase tissue binding. Also, a label for polypeptides can include a nucleic acid, for example DNA for detection by PCR, or enhancing gene expression, or siRNA to suppress gene expression in RAGE-bearing cells or tissues.

The class and subclass of RAGE antibodies may be determined by any method known in the art. In general, the class and subclass of an antibody may be determined using antibodies that are specific for a particular class and subclass of antibody. Such antibodies are available commercially. The class and subclass can be determined by ELISA, Western Blot as well as other techniques. Alternatively, the class and subclass may be determined by sequencing all or a portion of the constant domains of the heavy and/or light chains of the antibodies, comparing their amino acid sequences to the known amino acid sequences of various classes and subclasses of immunoglobulins, and determining the class and subclass of the antibodies.

5.5 Dual Variable Domain Immunoglobulins

Dual variable domain (DVD) binding proteins or immunoglobulins as used herein, are binding proteins that comprise two or more antigen binding sites and are

tetravalent or multivalent binding proteins, as for example divalent and tetravalent. The term "multivalent binding protein" is used in this specification to denote a binding protein comprising two or more antigen binding sites. The multivalent binding protein is particularly engineered to have the two or more antigen binding sites, and is generally not a naturally occurring antibody. The term "multispecific binding protein" refers to a binding protein capable of binding two or more related or unrelated targets. Such DVDs may be monospecific, i.e capable of binding one antigen or multispecific, i.e. capable of binding two or more antigens. DVD binding proteins comprising two heavy chain DVD polypeptides and two light chain DVD polypeptides are referred to a DVD Ig. Each half of a DVD Ig comprises a heavy chain DVD polypeptide, and a light chain DVD polypeptide, and two antigen binding sites. Each binding site comprises a heavy chain variable domain and a light chain variable domain with a total of 6 CDRs involved in antigen binding per antigen binding site. DVD binding proteins and methods of making DVD binding proteins are disclosed in US. Patent Application No. 11/507,050 and incorporated herein by reference. It is intended that the present invention comprises a DVD binding protein comprising binding proteins capable of binding RAGE. Particularly the DVD binding protein is capable of binding RAGE and a second target. The second target is selected from the group consisting of anti inflammatory MAB activities (IL-1, IL-6, IL-8, IL-11, IL-12,IL-17, IL-18, IL-23,TNF alpha/beta, IFN-beta, gamma, LIF, OSM, CNTF, PF-4, Platelet basic protein (PBP), NAP-2, beta-TG, MIP-1, MCP2/3, RANTES, lymphotactin), of transport-mediating proteins (insulin receptor, transferrin receptor, thrombin receptor, leptin receptor, LDL receptor), of other neuroregenerative MABs (NgR, Lingo, p75, CSPG (e.g. NG-2, neurocan, brevican, versican, aggrecan) hyaluronic acid, mAG, tenascin, NI-35, NI-250, IMP, perlecan, neurocan, phosphacan, nogo-A, OMGP, Sema4D, Sema 3A, ephrin B3, ephrin A2, ephrin A5, MAG, EphA4,plexin B1, TROY, wnts, ryk rec., BMP-2, BMP-4, BMP-7), of neuroprotective MAB activities(EGF, EGFR, Sema 3), of anti-amyloid beta MABs (e.g. m266, 3D6 (bapineuzumab), anti-globulomer MABs 7C6), of CNS located receptors and transporters (serotonin receptors, dopamine receptors, DAT, Asc-1, GlyT1).

5.6 Dual-specific antibodies

The present application also describes "dual-specific antibody" technology. Dual-specific antibodies may serve as agonists, antagonists, or both in different combinations. Dual-specific antibodies are antibodies in which the VH chain binds to a first antigen and the VL chain binds to another antigen as exemplified in WO2008082651.

5.7 Crystallized Antibodies

Another embodiment of the present application provides a crystallized binding protein. The term "crystallized" as used herein, refer to an antibody, or antigen binding portion thereof, that exists in the form of a crystal. Crystals are one form of the solid state of matter, which is distinct from other forms such as the amorphous solid state or the liquid crystalline state. Crystals are composed of regular, repeating, three-dimensional arrays of atoms, ions, molecules (e.g., proteins such as antibodies), or molecular assemblies (e.g., antigen/antibody complexes). These three-dimensional arrays are arranged according to specific mathematical relationships that are well understood in the field. The fundamental unit, or building block, that is repeated in a crystal is called the asymmetric unit. Repetition of the asymmetric unit in an arrangement that conforms to a given, well-defined crystallographic symmetry provides the "unit cell" of the crystal. Repetition of the unit cell by regular translations in all three dimensions provides the crystal. See Giege, R. and Ducruix, A. Barrett, Crystallization of Nucleic Acids and Proteins, a Practical Approach, 2nd ed., pp. 20 1-16, Oxford University Press, New York, New York, (1999).

Particularly the present application describes crystals of whole RAGE antibodies and fragments thereof as disclosed herein, and formulations and compositions comprising such crystals. In one embodiment the crystallized binding protein has a greater half-life in vivo than the soluble counterpart of the binding protein. In another embodiment the binding protein retains biological activity after crystallization.

Crystallized binding protein of the invention may be produced according methods known in the art and as disclosed in WO 02072636, incorporated herein by reference.

5.8 Glycosylated Antibodies

Another embodiment of the invention provides a glycosylated binding protein wherein the antibody or antigen-binding portion thereof comprises one or more carbohydrate residues. Nascent *in vivo* protein production may undergo further processing, known as post-translational modification. In particular, sugar (glycosyl) residues may be added enzymatically, a process known as glycosylation. The resulting proteins bearing covalently linked oligosaccharide side chains are known as glycosylated proteins or glycoproteins. Antibodies are glycoproteins with one or more carbohydrate residues in the Fc domain, as well as the variable domain. Carbohydrate residues in the Fc domain have important effect on the effector function of the Fc domain, with minimal effect on antigen binding or half-life of the antibody (R. Jefferis, *Biotechnol. Prog.* **21** (2005), pp. 11–16). In contrast, glycosylation of the variable domain may have an effect on the antigen binding activity of the antibody. Glycosylation in the variable domain may have a negative effect on antibody binding affinity, likely due to steric hindrance (Co, M.S., et al., Mol. Immunol. (1993) 30:1361- 1367), or result in increased affinity for the antigen (Wallick, S.C., et al., Exp. Med. (1988) 168:1099-1109; Wright, A., et al., EMBO J. (1991) 10:2717 2723).

One aspect of the present invention is directed to generating glycosylation site mutants in which the O- or N-linked glycosylation site of the binding protein has been mutated. One skilled in the art can generate such mutants using standard well-known technologies. Glycosylation site mutants that retain the biological activity but have increased or decreased binding activity are another object of the present invention.

In still another embodiment, the glycosylation of the antibody or antigen-binding portion of the invention is modified. For example, an aglycoslated antibody can be made (i.e., the antibody lacks glycosylation). Glycosylation can be altered to, for example, increase the affinity of the antibody for antigen. Such carbohydrate modifications can be accomplished by, for example, altering one or more sites of glycosylation within the antibody sequence. For example, one or more amino acid substitutions can be made that result in elimination of one or more variable region glycosylation sites to thereby eliminate glycosylation at that site. Such aglycosylation may increase the affinity of the antibody for antigen. Such an approach is described in further detail in PCT Publication WO2003016466A2, and U.S. Pat. Nos. 5,714,350 and 6,350,861, each of which is incorporated herein by reference in its entirety.

Additionally or alternatively, a modified antibody of the invention can be made that has an altered type of glycosylation, such as a hypofucosylated antibody

having reduced amounts of fucosyl residues or an antibody having increased bisecting GlcNAc structures. Such altered glycosylation patterns have been demonstrated to increase the ADCC ability of antibodies. Such carbohydrate modifications can be accomplished by, for example, expressing the antibody in a host cell with altered glycosylation machinery. Cells with altered glycosylation machinery have been described in the art and can be used as host cells in which to express recombinant antibodies of the invention to thereby produce an antibody with altered glycosylation. See, for example, Shields, R. L. et al. (2002) J. Biol. Chem. 277:26733-26740; Umana et al. (1999) Nat. Biotech. 17:176-1, as well as, European Patent No: EP 1,176,195; PCT Publications WO 03/035835; WO 99/54342 80, each of which is incorporated herein by reference in its entirety.

Protein glycosylation depends on the amino acid sequence of the protein of interest, as well as the host cell in which the protein is expressed. Different organisms may produce different glycosylation enzymes (eg., glycosyltransferases and glycosidases), and have different substrates (nucleotide sugars) available. Due to such factors, protein glycosylation pattern, and composition of glycosyl residues, may differ depending on the host system in which the particular protein is expressed. Glycosyl residues useful in the invention may include, but are not limited to, glucose, galactose, mannose, fucose, n-acetylglucosamine and sialic acid. Particularly the glycosylated binding protein comprises glycosyl residues such that the glycosylation pattern is human.

It is known to those skilled in the art that differing protein glycosylation may result in differing protein characteristics. For instance, the efficacy of a therapeutic protein produced in a microorganism host, such as yeast, and glycosylated utilizing the yeast endogenous pathway may be reduced compared to that of the same protein expressed in a mammalian cell, such as a CHO cell line. Such glycoproteins may also be immunogenic in humans and show reduced half-life *in vivo* after administration. Specific receptors in humans and other animals may recognize specific glycosyl residues and promote the rapid clearance of the protein from the bloodstream. Other adverse effects may include changes in protein folding, solubility, susceptibility to proteases, trafficking, transport, compartmentalization, secretion, recognition by other proteins or factors, antigenicity, or allergenicity. Accordingly, a practitioner may prefer a therapeutic protein with a specific composition and pattern of glycosylation,

for example glycosylation composition and pattern identical, or at least similar, to that produced in human cells or in the species-specific cells of the intended subject animal.

Expressing glycosylated proteins different from that of a host cell may be achieved by genetically modifying the host cell to express heterologous glycosylation enzymes. Using techniques known in the art a practitioner may generate antibodies or antigen-binding portions thereof exhibiting human protein glycosylation. For example, yeast strains have been genetically modified to express non-naturally occurring glycosylation enzymes such that glycosylated proteins (glycoproteins) produced in these yeast strains exhibit protein glycosylation identical to that of animal cells, especially human cells (U.S patent applications 20040018590 and 20020137134 and PCT publication WO2005100584 A2).

Further, it will be appreciated by one skilled in the art that a protein of interest may be expressed using a library of host cells genetically engineered to express various glycosylation enzymes, such that member host cells of the library produce the protein of interest with variant glycosylation patterns. A practitioner may then select and isolate the protein of interest with particular novel glycosylation patterns. Particularly, the protein having a particularly selected novel glycosylation pattern exhibits improved or altered biological properties.

5.9 Anti-idiotypic Antibodies

In addition to the binding proteins, the present invention is also directed to an anti-idiotypic (anti-Id) antibody specific for such binding proteins of the invention. An anti-Id antibody is an antibody, which recognizes unique determinants generally associated with the antigen-binding region of another antibody. The anti-Id can be prepared by immunizing an animal with the binding protein or a CDR containing region thereof. The immunized animal will recognize, and respond to the idiotypic determinants of the immunizing antibody and produce an anti-Id antibody. The anti-Id antibody may also be used as an "immunogen" to induce an immune response in yet another animal, producing a so-called anti-anti-Id antibody.

6. Uses of the Antibodies

Given their ability to bind to human RAGE, the neutralizing antibodies of the present application, or portions thereof, can be used to detect human RAGE (e.g., in a biological sample, such as serum or plasma), using a conventional immunoassay, such

as an enzyme linked immunosorbent assays (ELISA), a radioimmunoassay (RIA) or tissue immunohistochemistry. The present application provides a method for detecting human RAGE in a biological sample comprising contacting a biological sample with an antibody, or antibody portion, of the invention and detecting either the antibody (or antibody portion) bound to human RAGE or unbound antibody (or antibody portion), to thereby detect human RAGE in the biological sample. The antibody is directly or indirectly labeled with a detectable substance to facilitate detection of the bound or unbound antibody. Suitable detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, beta-galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; and examples of suitable radioactive material include ^{3}H, ^{14}C, ^{35}S, ^{90}Y, ^{99}Tc, ^{111}In, ^{125}I, ^{131}I, ^{177}Lu, ^{166}Ho, ^{153}Sm.

The antibodies and antibody portions of the present application particularly are capable of neutralizing human RAGE activity both in vitro and in vivo. Accordingly, such antibodies and antibody portions of the invention can be used to inhibit RAGE binding to its ligands and therefore neutralize the resulting activity.

In another embodiment, the present application provides a method for reducing RAGE activity in a subject, advantageously from a subject suffering from a disease or disorder in which RAGE resulting activity is detrimental. The present application provides methods for reducing RAGE activity in a subject suffering from such a disease or disorder, by preventing RAGE binding to at least one of its ligands, like Aß-globulomers, through the use of the monoclonal antibodies of the present application. The antibodies of the present invention, in particular, the humanized antibodies disclosed herein, can be administered to a human subject for therapeutic purposes. Moreover, the antibodies of the present application can be administered to a non-human mammal expressing an RAGE with which the antibody is capable of binding for veterinary purposes or as an animal model of human disease. Regarding the latter, such animal models may be useful for evaluating the therapeutic efficacy of antibodies of the invention (e.g., testing of dosages and time courses of administration).

As used herein, the term "a disorder in which RAGE activity is detrimental" is intended to include diseases and other disorders in which the presence of RAGE or its resulting activity in a subject suffering from the disorder has been shown to be or is suspected of being either responsible for the pathophysiology of the disorder or a factor that contributes to a worsening of the disorder. Accordingly, a disorder in which RAGE activity is detrimental is a disorder in which reduction of RAGE activity is expected to alleviate the symptoms and/or progression of the disorder. Non-limiting examples of disorders that can be treated with the antibodies of the invention include those disorders discussed in the section below pertaining to pharmaceutical compositions of the antibodies of the invention.

It is recognized that RAGE plays an important role in the pathology associated with a variety of diseases involving neurological diseases selected from the group comprising Amytropic Lateral Sclerosis, Brachial Plexus Injury, Brain Injury, including traumatic brain injury, Cerebral Palsy, Friedrich's Ataxia, Guillain Barre, Leukodystrophies, Multiple Sclerosis, Post Polio, Spina Bifida, Spinal Cord Injury, Spinal Muscle Atrophy, Spinal Tumors, Stroke, Transverse Myelitits, dementia, senile dementia, mild cognitive impairment, Alzheimer-related dementia, Huntington's chorea, tardive dyskinesia, hyperkinesias, manias, Morbus Parkinson, steel-Richard syndrome, Down's syndrome, myasthenia gravis, nerve trauma, vascular amyloidosis, cerebral hemorrhage I with amyloidosis, brain inflammation, Friedrich's ataxia, acute confusion disorder, amyotrophic lateral sclerosis, glaucoma , Alzheimer's disease, diabetic nephropathy, sepsis, rheumatoid arthritis and related inflammatory diseases. Diabetes and resulting complications like diabetic retinopathy, nephropathy, vascular complications; atherosclerotic complications, pulmonary fibrosis, Cancer especially melanomas, other amyloidoses. (See for example the following references: Amyloidosis, cancer, arthritis, Crohn's disease, chronic and acute inflammatory diseases: Schmidt AM et al: J Clin Invest. 2001 Oct;108(7):949-55.; cardiovascular diseases, diabetes, diabetic complications: Yan SD et al: Eur J Clin Invest. 1997 Mar;27(3):179-81; Prion-associated diseases: Sasaki N et al:Neurosci Lett. 2002 Jun 28;326(2):117-20; vascularitis, nephropathies, retinopathies and neuropathies: Thornalley PJ.:Int Rev Neurobiol. 2002;50:37-57; alzheimer disease: Weldon DT et al: Geriatrics. 1997 Sep;52 Suppl 2:S13-6; Yan SD et al: Biochim Biophys Acta. 2000 Jul 26;1502(1):145-57; rheumatoid arthritis, osteoarthritis: Drinda S et al:.Rheumatol Int. 2004 Mar 26; bowel disease: Foell D et al:Gut. 2003 Jun;52(6):847-53; multiple sclerosis: Yan SS et al:Nat Med. 2003 Mar;9(3):287-93; psoriasis: Foell D et al: Rheumatology (Oxford). 2003 Nov;42(11):1383-9 : lupus : Tanji N et al:J Am Soc

Nephrol. 2000 Sep;11(9):1656-66; general autoimmune diseases, sepsis: Liliensiek B et al:J Clin Invest. 2004 Jun;113(11):1641-50; arteriosclerosis and restenosis : Schmidt AM et al: Circ Res. 1999 Mar 19;84(5):489-97).

Also, as previously discussed, DVD immunoglobulins, or dual-specific antibodies between any one of the partners described above may be of use. Such antibody preparations as described above may be useful for the treatment of such diseases.

The antibodies of the present application may also be combined with peptides allowing the trans-membrane transfer to include targeting of intracellular target proteins. Such peptide sequences may include, but are not limited to, tat, antennapedia, poly-args, some anti-microbial peptides. Such peptides may allow transfer through membranes, including cellular plasma membranes, but also epithelia and endothelial membranes, including the blood-brain-barrier, gut mucosa, meninges, and others.

An antibody, or antibody portion, of the present application also can be administered with one or more additional small molecule therapeutic agents useful in the treatment of disorders in which RAGE activity is involved as discussed in the foregoing paragraphs. It should be understood that the antibodies of the present application or antigen binding portion thereof can be used alone or in combination with an additional agent, e.g., a therapeutic agent, said additional agent being selected by the skilled artisan for its intended purpose. For example, the additional agent can be a therapeutic agent art-recognized as being useful to treat the disease or condition being treated by the antibody of the present invention. The additional agent also can be an agent that imparts a beneficial attribute to the therapeutic composition e.g., an agent that affects the viscosity of the composition.

7. Pharmaceutical Compositions

The invention also provides pharmaceutical compositions comprising an antibody, or antigen-binding portion thereof, of the invention and a pharmaceutically acceptable carrier. The pharmaceutical compositions comprising antibodies of the invention are for use in, but not limited to, diagnosing, detecting, or monitoring a disorder, in preventing, treating, managing, or ameliorating of a disorder or one or more symptoms thereof, and/or in research. In a specific embodiment, a composition comprises one or more antibodies of the invention. In another embodiment, the

pharmaceutical composition comprises one or more antibodies of the invention and one or more prophylactic or therapeutic agents other than antibodies of the invention for treating a disorder in which RAGE activity is detrimental. Particularly, the prophylactic or therapeutic agents known to be useful for or having been or currently being used in the prevention, treatment, management, or amelioration of a disorder or one or more symptoms thereof. In accordance with these embodiments, the composition may further comprise of a carrier, diluent or excipient.

The antibodies and antibody-portions of the invention can be incorporated into pharmaceutical compositions suitable for administration to a subject. Typically, the pharmaceutical composition comprises an antibody or antibody portion of the invention and a pharmaceutically acceptable carrier. As used herein, "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like that are physiologically compatible. Examples of pharmaceutically acceptable carriers include one or more of water, saline, phosphate buffered saline, dextrose, glycerol, ethanol and the like, as well as combinations thereof. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as mannitol, sorbitol, or sodium chloride in the composition. Pharmaceutically acceptable carriers may further comprise minor amounts of auxiliary substances such as wetting or emulsifying agents, preservatives or buffers, which enhance the shelf life or effectiveness of the antibody or antibody portion.

Various delivery systems are known and can be used to administer one or more antibodies of the invention or the combination of one or more antibodies of the invention and a prophylactic agent or therapeutic agent useful for preventing, managing, treating, or ameliorating a disorder or one or more symptoms thereof, e.g., encapsulation in liposomes, microparticles, microcapsules, recombinant cells capable of expressing the antibody or antibody fragment, receptor- mediated endocytosis (see, e. g., Wu and Wu, J. Biol. Chem. 262:4429-4432 (1987)), construction of a nucleic acid as part of a retroviral or other vector, etc. Methods of administering a prophylactic or therapeutic agent of the invention include, but are not limited to, parenteral administration (e.g., intradermal, intramuscular, intraperitoneal, intravenous and subcutaneous) , epidurala administration, intratumoral administration, and mucosal adminsitration (e.g., intranasal and oral routes). In addition, pulmonary administration can be employed, e.g., by use of an inhaler or nebulizer, and

formulation with an aerosolizing agent. See, e.g., U.S. Pat. Nos. 6, 019,968, 5,985, 320, 5,985,309, 5,934, 272, 5,874,064, 5,855,913, 5,290, 540, and 4,880,078; and PCT Publication Nos. WO 92/19244, WO 97/32572, WO 97/44013, WO 98/31346, and WO 99/66903, each of which is incorporated herein by reference their entireties. In one embodiment, an antibody of the invention, combination therapy, or a composition of the invention is administered using Alkermes AIR® pulmonary drug delivery technology (Alkermes, Inc., Cambridge, Mass.). In a specific embodiment, prophylactic or therapeutic agents of the invention are administered intramuscularly, intravenously, intratumorally, orally, intranasally, pulmonary, or subcutaneously. The prophylactic or therapeutic agents may be administered by any convenient route, for example by infusion or bolus injection, by absorption through epithelial or mucocutaneous linings (e.g. , oral mucosa, rectal and intestinal mucosa, etc.) and may be administered together with other biologically active agents. Administration can be systemic or local.

In a specific embodiment, it may be desirable to administer the prophylactic or therapeutic agents of the invention locally to the area in need of treatment; this may be achieved by, for example, and not by way of limitation, local infusion, by injection, or by means of an implant, said implant being of a porous or non-porous material, including membranes and matrices, such as sialastic membranes, polymers, fibrous matrices (e.g., Tissel®), or collagen matrices. In one embodiment, an effective amount of one or more antibodies of the invention antagonists is administered locally to the affected area to a subject to prevent, treat, manage, and/or ameliorate a disorder or a symptom thereof. In another embodiment, an effective amount of one or more antibodies of the invention is administered locally to the affected area in combination with an effective amount of one or more therapies (e. g., one or more prophylactic or therapeutic agents) other than an antibody of the invention of a subject to prevent, treat, manage, and/or ameliorate a disorder or one or more symptoms thereof.

In another embodiment, the prophylactic or therapeutic agent can be delivered in a controlled release or sustained release system. In one embodiment, a pump may be used to achieve controlled or sustained release (see Langer, supra; Sefton, 1987, CRC Crit. Ref. Biomed. Eng. 14:20; Buchwald et al., 1980, Surgery 88:507; Saudek et al., 1989, N. Engl. J. Med. 321:574). In another embodiment, polymeric materials can be used to achieve controlled or sustained release of the therapies of the invention (see e.g., Medical Applications of Controlled Release, Langer and Wise (eds.), CRC

Pres., Boca Raton, Fla. (1974); Controlled Drug Bioavailability, Drug Product Design and Performance, Smolen and Ball (eds.), Wiley, New York (1984); Ranger and Peppas, 1983, J., Macromol. Sci. Rev. Macromol. Chem. 23:61; see also Levy et al., 1985, Science 228:190; During et al., 1989, Ann. Neurol. 25:351; Howard et al., 1989, J. Neurosurg. 7 1:105); U.S. Pat. No. 5,679,377; U.S. Pat. No. 5, 916,597; U. S. Pat. No. 5,912,015; U.S. Pat. No. 5,989,463; U.S. Pat. No. 5,128,326; PCT Publication No. WO 99/15154; and PCT Publication No. WO 99/20253. Examples of polymers used in sustained release formulations include, but are not limited to, poly(2-hydroxy ethyl methacrylate), poly(methyl methacrylate), poly(acrylic acid), poly(ethylene-co-vinyl acetate), poly(methacrylic acid), polyglycolides (PLG), polyanhydrides, poly(N- vinyl pyrrolidone), poly(vinyl alcohol), polyacrylamide, poly(ethylene glycol), polylactides (PLA), poly(lactide-co-glycolides) (PLGA), and polyorthoesters. In a preferred embodiment, the polymer used in a sustained release formulation is inert, free of leachable impurities, stable on storage, sterile, and biodegradable. In yet another embodiment, a controlled or sustained release system can be placed in proximity of the prophylactic or therapeutic target, thus requiring only a fraction of the systemic dose (see, e.g., Goodson, in Medical Applications of Controlled Release, supra, vol. 2, pp. 115-138 (1984)).

Controlled release systems are discussed in the review by Langer (1990, Science 249:1527-1533). Any technique known to one of skill in the art can be used to produce sustained release formulations comprising one or more therapeutic agents of the invention. See, e.g., U. S. Pat. No. 4,526, 938, PCT publication WO 91/05548, PCT publication WO 96/20698, Ning et al. , 1996, "Intratumoral Radioimmunotheraphy of a Human Colon Cancer Xenograft Using a Sustained-Release Gel," Radiotherapy &Oncology 39:179-189, Song et al., 1995, "Antibody Mediated Lung Targeting of Long- Circulating Emulsions," PDA Journal of Pharmaceutical Science &Technology 50:372-397, Cleek et al., 1997, "Biodegradable Polymeric Carriers for a bFGF Antibody for Cardiovascular Application," Pro. Int'l. Symp. Control. Rel. Bioact. Mater. 24:853-854, and Lam et al., 1997, "Microencapsulation of Recombinant Humanized Monoclonal Antibody for Local Delivery," Proc. Int'l. Symp. Control Rel. Bioact. Mater. 24:759- 760, each of which is incorporated herein by reference in their entireties.

In a specific embodiment, where the composition of the invention is a nucleic acid encoding a prophylactic or therapeutic agent, the nucleic acid can be

administered in vivo to promote expression of its encoded prophylactic or therapeutic agent, by constructing it as part of an appropriate nucleic acid expression vector and administering it so that it becomes intracellular, e.g., by use of a retroviral vector (see U. S. Pat. No. 4,980,286), or by direct injection, or by use of microparticle bombardment (e.g., a gene gun; Biolistic, Dupont), or coating with lipids or cell-surface receptors or transfecting agents, or by administering it in linkage to a homeobox-like peptide which is known to enter the nucleus (see, e.g., Joliot et al., 1991, Proc. Natl. Acad. Sci. USA 88:1864-1868). Alternatively, a nucleic acid can be introduced intracellularly and incorporated within host cell DNA for expression by homologous recombination.

A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include, but are not limited to, parenteral, e.g., intravenous, intradermal, subcutaneous, oral, intranasal (e.g., inhalation), transdermal (e.g., topical), transmucosal, and rectal administration. In a specific embodiment, the composition is formulated in accordance with routine procedures as a pharmaceutical composition adapted for intravenous, subcutaneous, intramuscular, oral, intranasal, or topical administration to human beings. Typically, compositions for intravenous administration are solutions in sterile isotonic aqueous buffer. Where necessary, the composition may also include a solubilizing agent and a local anesthetic such as lignocamne to ease pain at the site of the injection.

If the compositions of the invention are to be administered topically, the compositions can be formulated in the form of an ointment, cream, transdermal patch, lotion, gel, shampoo, spray, aerosol, solution, emulsion, or other form well known to one of skill in the art. See, e.g., Remington's Pharmaceutical Sciences and Introduction to Pharmaceutical Dosage Forms, 19th ed., Mack Pub. Co., Easton, Pa. (1995). For non- sprayable topical dosage forms, viscous to semi-solid or solid forms comprising a carrier or one or more excipients compatible with topical application and having a dynamic viscosity particularly greater than water are typically employed. Suitable formulations include, without limitation, solutions, suspensions, emulsions, creams, ointments, powders, liniments, salves, and the like, which are, if desired, sterilized or mixed with auxiliary agents (e.g., preservatives, stabilizers, wetting agents, buffers, or salts) for influencing various properties, such as, for example, osmotic pressure. Other suitable topical dosage forms include sprayable aerosol

preparations wherein the active ingredient, particularly in combination with a solid or liquid inert carrier, is packaged in a mixture with a pressurized volatile (e.g., a gaseous propellant, such as freon) or in a squeeze bottle. Moisturizers or humectants can also be added to pharmaceutical compositions and dosage forms if desired. Examples of such additional ingredients are well known in the art.

If the method of the invention comprises intranasal administration of a composition, the composition can be formulated in an aerosol form, spray, mist or in the form of drops. In particular, prophylactic or therapeutic agents for use according to the present invention can be conveniently delivered in the form of an aerosol spray presentation from pressurized packs or a nebuliser, with the use of a suitable propellant (e.g., dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide or other suitable gas). In the case of a pressurized aerosol the dosage unit may be determined by providing a valve to deliver a metered amount. Capsules and cartridges (composed of, e.g., gelatin) for use in an inhaler or insufflator may be formulated containing a powder mix of the compound and a suitable powder base such as lactose or starch.

If the method of the invention comprises oral administration, compositions can be formulated orally in the form of tablets, capsules, cachets, gelcaps, solutions, suspensions, and the like. Tablets or capsules can be prepared by conventional means with pharmaceutically acceptable excipients such as binding agents (e.g., pregelatinised maize starch, polyvinylpyrrolidone, or hydroxypropyl methylcellulose); fillers (e.g., lactose, microcrystalline cellulose, or calcium hydrogen phosphate) ; lubricants (e.g., magnesium stearate, talc, or silica); disintegrants (e.g., potato starch or sodium starch glycolate) ; or wetting agents (e.g., sodium lauryl sulphate). The tablets may be coated by methods well-known in the art. Liquid preparations for oral administration may take the form of, but not limited to, solutions, syrups or suspensions, or they may be presented as a dry product for constitution with water or other suitable vehicle before use. Such liquid preparations may be prepared by conventional means with pharmaceutically acceptable additives such as suspending agents (e.g., sorbitol syrup, cellulose derivatives, or hydrogenated edible fats); emulsifying agents (e.g., lecithin or acacia); non-aqueous vehicles (e.g., almond oil, oily esters, ethyl alcohol, or fractionated vegetable oils); and preservatives (e.g., methyl or propyl-p- hydroxybenzoates or sorbic acid). The preparations may also contain buffer salts, flavoring, coloring, and sweetening agents as appropriate.

Preparations for oral administration may be suitably formulated for slow release, controlled release, or sustained release of a prophylactic or therapeutic agent(s).

The method of the invention may comprise pulmonary administration, e.g., by use of an inhaler or nebulizer, of a composition formulated with an aerosolizing agent. See, e.g., U.S. Pat. Nos. 6,019, 968, 5,985, 320, 5, 985,309, 5,934,272, 5,874,064, 5,855,913, 5,290,540, and 4,880,078; and PCT Publication Nos. WO 92/19244, WO 97/32572, WO 97/44013, WO 98/31346, and WO 99/66903, each of which is incorporated herein by reference their entireties. In a specific embodiment, an antibody of the invention, combination therapy, and/or composition of the invention is administered using Alkermes AIR® pulmonary drug delivery technology (Alkermes, Inc., Cambridge, Mass.).

The method of the invention may comprise administration of a composition formulated for parenteral administration by injection (e. g., by bolus injection or continuous infusion). Formulations for injection may be presented in unit dosage form (e.g., in ampoules or in multi-dose containers) with an added preservative. The compositions may take such forms as suspensions, solutions or emulsions in oily or aqueous vehicles, and may contain formulatory agents such as suspending, stabilizing and/or dispersing agents. Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle (e.g., sterile pyrogen-free water) before use. The methods of the invention may additionally comprise of administration of compositions formulated as depot preparations. Such long acting formulations may be administered by implantation (e.g., subcutaneously or intramuscularly) or by intramuscular injection. Thus, for example, the compositions may be formulated with suitable polymeric or hydrophobic materials (e.g., as an emulsion in an acceptable oil) or ion exchange resins, or as sparingly soluble derivatives (e.g., as a sparingly soluble salt).

The methods of the invention encompass administration of compositions formulated as neutral or salt forms. Pharmaceutically acceptable salts include those formed with anions such as those derived from hydrochloric, phosphoric, acetic, oxalic, tartaric acids, etc., and those formed with cations such as those derived from sodium, potassium, ammonium, calcium, ferric hydroxides, isopropylamine, triethylamine, 2- ethylamino ethanol, histidine, procaine, etc.

Generally, the ingredients of compositions are supplied either separately or mixed together in unit dosage form, for example, as a dry lyophilized powder or water

free concentrate in a hermetically sealed container such as an ampoule or sachette indicating the quantity of active agent. Where the mode of administration is infusion, composition can be dispensed with an infusion bottle containing sterile pharmaceutical grade water or saline. Where the mode of administration is by injection, an ampoule of sterile water for injection or saline can be provided so that the ingredients may be mixed prior to administration.

In particular, the invention also provides that one or more of the prophylactic or therapeutic agents, or pharmaceutical compositions of the invention is packaged in a hermetically sealed container such as an ampoule or sachette indicating the quantity of the agent. In one embodiment, one or more of the prophylactic or therapeutic agents, or pharmaceutical compositions of the invention is supplied as a dry sterilized lyophilized powder or water free concentrate in a hermetically sealed container and can be reconstituted (e.g., with water or saline) to the appropriate concentration for administration to a subject. Particularly, one or more of the prophylactic or therapeutic agents or pharmaceutical compositions of the invention is supplied as a dry sterile lyophilized powder in a hermetically sealed container at a unit dosage of at least 5 mg, more particularly at least 10 mg, at least 15 mg, at least 25 mg, at least 35 mg, at least 45 mg, at least 50 mg, at least 75 mg, or at least 100 mg. The lyophilized prophylactic or therapeutic agents or pharmaceutical compositions of the invention should be stored at between 2° C. and 8° C. in its original container and the prophylactic or therapeutic agents, or pharmaceutical compositions of the invention should be administered within 1 week, particularly within 5 days, within 72 hours, within 48 hours, within 24 hours, within 12 hours, within 6 hours, within 5 hours, within 3 hours, or within 1 hour after being reconstituted. In an alternative embodiment, one or more of the prophylactic or therapeutic agents or pharmaceutical compositions of the invention is supplied in liquid form in a hermetically sealed container indicating the quantity and concentration of the agent. Particularly, the liquid form of the administered composition is supplied in a hermetically sealed container at least 0.25 mg/ml, more particularly at least 0.5 mg/ml, at least 1 mg/ml, at least 2.5 mg/ml, at least 5 mg/ml, at least 8 mg/ml, at least 10 mg/ml, at least 15 mg/kg, at least 25 mg/ml, at least 50 mg/ml, at least 75 mg/ml or at least 100 mg/ml. The liquid form should be stored at between 2° C. and 8° C. in its original container.

The antibodies and antibody-portions of the invention can be incorporated into a pharmaceutical composition suitable for parenteral administration. Particularly, the

antibody or antibody-portions will be prepared as an injectable solution containing 0.1-250 mg/ml antibody. The injectable solution can be composed of either a liquid or lyophilized dosage form in a flint or amber vial, ampule or pre-filled syringe. The buffer can be L-histidine (1-50 mM), optimally 5-10mM, at pH 5.0 to 7.0 (optimally pH 6.0). Other suitable buffers include but are not limited to, sodium succinate, sodium citrate, sodium phosphate or potassium phosphate. Sodium chloride can be used to modify the toxicity of the solution at a concentration of 0-300 mM (optimally 150 mM for a liquid dosage form). Cryoprotectants can be included for a lyophilized dosage form, principally 0-10% sucrose (optimally 0.5-1.0%). Other suitable cryoprotectants include trehalose and lactose. Bulking agents can be included for a lyophilized dosage form, principally 1-10% mannitol (optimally 2-4%). Stabilizers can be used in both liquid and lyophilized dosage forms, principally 1-50 mM L-Methionine (optimally 5-10 mM). Other suitable bulking agents include glycine, arginine, can be included as 0-0.05% polysorbate-80 (optimally 0.005-0.01%). Additional surfactants include but are not limited to polysorbate 20 and BRIJ surfactants. The pharmaceutical composition comprising the antibodies and antibody-portions of the invention prepared as an injectable solution for parenteral administration, can further comprise an agent useful as an adjuvant, such as those used to increase the absorption, or dispersion of a therapeutic protein (e.g., antibody). A particularly useful adjuvant is hyaluronidase, such as Hylenex® (recombinant human hyaluronidase).Addition of hyaluronidase in the injectable solution improves human bioavailability following parenteral administration, particularly subcutaneous administration. It also allows for greater injection site volumes (i.e. greater than 1 ml) with less pain and discomfort, and minimum incidence of injection site reactions. (see WO2004078140, US2006104968 incorporated herein by reference).

The compositions of this invention may be in a variety of forms. These include, for example, liquid, semi-solid and solid dosage forms, such as liquid solutions (*e.g.*, injectable and infusible solutions), dispersions or suspensions, tablets, pills, powders, liposomes and suppositories. The particular form depends on the intended mode of administration and therapeutic application. Typical particular compositions are in the form of injectable or infusible solutions, such as compositions similar to those used for passive immunization of humans with other antibodies. A particular mode of administration is parenteral (*e.g.*, intravenous, subcutaneous, intraperitoneal, intramuscular). In a particular embodiment, the antibody is

administered by intravenous infusion or injection. In another particular embodiment, the antibody is administered by intramuscular or subcutaneous injection.

Therapeutic compositions typically must be sterile and stable under the conditions of manufacture and storage. The composition can be formulated as a solution, microemulsion, dispersion, liposome, or other ordered structure suitable to high drug concentration. Sterile injectable solutions can be prepared by incorporating the active compound (*i.e.*, antibody or antibody portion) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle that contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile, lyophilized powders for the preparation of sterile injectable solutions, particular methods of preparation are vacuum drying and spray-drying that yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof. The proper fluidity of a solution can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prolonged absorption of injectable compositions can be brought about by including, in the composition, an agent that delays absorption, for example, monostearate salts and gelatin.

The antibodies and antibody-portions of the present invention can be administered by a variety of methods known in the art, although for many therapeutic applications, a particular route/mode of administration is subcutaneous injection, intravenous injection or infusion. As will be appreciated by the skilled artisan, the route and/or mode of administration will vary depending upon the desired results. In certain embodiments, the active compound may be prepared with a carrier that will protect the compound against rapid release, such as a controlled release formulation, including implants, transdermal patches, and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Many methods for the preparation of such formulations are patented or generally known to those skilled in the art. See, *e.g.*, *Sustained and Controlled Release Drug Delivery Systems*, J.R. Robinson, ed., Marcel Dekker, Inc., New York, 1978.

In certain embodiments, an antibody or antibody portion of the invention may be orally administered, for example, with an inert diluent or an assimilable edible

carrier. The compound (and other ingredients, if desired) may also be enclosed in a hard or soft shell gelatin capsule, compressed into tablets, or incorporated directly into the subject's diet. For oral therapeutic administration, the compounds may be incorporated with excipients and used in the form of ingestible tablets, buccal tablets, troches, capsules, elixirs, suspensions, syrups, wafers, and the like. To administer a compound of the invention by other than parenteral administration, it may be necessary to coat the compound with, or co-administer the compound with, a material to prevent its inactivation.

Supplementary active compounds can also be incorporated into the compositions. In certain embodiments, an antibody or antibody portion of the invention is coformulated with and/or coadministered with one or more additional therapeutic agents that are useful for treating disorders in which RAGE activity is detrimental. For example, an anti-RAGE antibody or antibody portion of the invention may be coformulated and/or coadministered with one or more additional antibodies that bind other targets (*e.g.*, antibodies that bind cytokines or that bind cell surface molecules). Furthermore, one or more antibodies of the invention may be used in combination with two or more of the foregoing therapeutic agents. Such combination therapies may advantageously utilize lower dosages of the administered therapeutic agents, thus avoiding possible toxicities or complications associated with the various monotherapies.

In certain embodiments, an antibody to RAGE or fragment thereof is linked to a half-life extending vehicle known in the art. Such vehicles include, but are not limited to, the Fc domain, polyethylene glycol, and dextran. Such vehicles are described, e.g., in U.S. Application Serial No. 09/428,082 and published PCT Application No. WO 99/25044, which are hereby incorporated by reference for any purpose.

In a specific embodiment, nucleic acid sequences comprising nucleotide sequences encoding an antibody of the invention or another prophylactic or therapeutic agent of the invention are administered to treat, prevent, manage, or ameliorate a disorder or one or more symptoms thereof by way of gene therapy. Gene therapy refers to therapy performed by the administration to a subject of an expressed or expressible nucleic acid. In this embodiment of the invention, the nucleic acids produce their encoded antibody or prophylactic or therapeutic agent of the invention that mediates a prophylactic or therapeutic effect.

Any of the methods for gene therapy available in the art can be used according to the present invention. For general reviews of the methods of gene therapy, see Goldspiel et al., 1993, Clinical Pharmacy 12:488-505; Wu and Wu, 1991, Biotherapy 3:87-95; Tolstoshev, 1993, Ann. Rev. Pharmacol. Toxicol. 32:573-596; Mulligan, Science 260:926- 932 (1993); and Morgan and Anderson, 1993, Ann. Rev. Biochem. 62:191-217; May, 1993, TIBTECH 11(5):155-215. Methods commonly known in the art of recombinant DNA technology which can be used are described in Ausubel et al. (eds.), Current Protocols in Molecular Biology, John Wiley &Sons, NY (1993); and Kriegler, Gene Transfer and Expression, A Laboratory Manual, Stockton Press, NY (1990). Detailed description of various methods of gene therapy are disclosed in US20050042664 A1 which is incorporated herein by reference.

RAGE plays a critical role in the pathology associated with a variety of diseases as defined herein above. Infusion of amyloid Aβ-peptides into animals leads to responses like inflammatory responses in arteriolose, decrease in cerebral blood flow. These effects could be prevented by antibodies against RAGE (Rhodin, J. et al. World Congress for Microcirculation, submitted Papers, 7th, Sydney, Australia, Aug. 19-22, 2001 ,543-547; Deane et al. Nature med. 2003). RAGE is upregulated in the microvasculature of AD patients and in transgenice mice where the human APP gene has been overexpressed (Deane et al. Nature med. 2003). Using double-transgenic mice where the human APP gene is expressed and RAGE is overexpressed it was shown that overexpression of the normal RAGE gene leads to impairment in learning, increase in plaques whereas overexpression of a dominant-negative signalling defective RAGE variant leads to improvement in learning and lower plaque levels (Arancio et al. 2004 EMBO J. 2004). Experimentation in animal models of both Type 1 and 2 diabetes reveals that antagonism of the ligand-RAGE axis suppresses the development and progression of vascular and inflammatory cell perturbation in the diabetic milieu, e.g. RAGE knock-out mice and Anti-RAGE antibodies have been used to show an improvement in animal models for e.g. diabetic nephropathy (Ravichandran R. et al CANADIAN JOURNAL OF DIABETES. 2006;30(4):422, Myint Khin et al. Diabetes (2006), 55(9), 2510; De-Vriese et al. Journal of the American Society of Nephrology 2003, 14/8, 2109, Jensen et al. Renal effects of a neutralising RAGE-antibody in long-term streptozotocin-diabetic mice. The Journal of endocrinology, 2006, 188, 493). Positive long-term renal effects of a neutralizing

RAGE antibody in obese type 2 diabetic mice were shown by Flyvbjerg et al (Diabetes, 2004, 53, 1, p. 166-72). RAGE knock-out mice were used to show an involvement of RAGE in sepsis (Birgit Liliensiek et al. J Clin Invest. 2004 June 1; 113(11): 1641–1650; Receptor for advanced glycation end products (RAGE) regulates sepsis but not the adaptive immune response). Blocking $F(ab)_2$ fragments derived from anti-RAGE IgG reduces the inflammatory response in MOG or MBP-induced EAE (Yan, S.S., et al. 2003. Nat. Med. 9:287–293.) Involvement of RAGE in Cancer was shown (Abe-R et al. Journal of Investigative Dermatology, 2004, 122/2 (461-467). In tumor-bearing mice, survival rates were prolonged, and spontaneous pulmonary metastases were inhibited by treatment using anti-RAGE neutralizing antibodies.

The antibodies, and antibody portions of the invention can be used to treat humans suffering from such a diseases.

It should be understood that the antibodies of the invention or antigen binding portion thereof can be used alone or in combination with an additional agent, e.g., a therapeutic agent, said additional agent being selected by the skilled artisan for its intended purpose. For example, the additional agent can be a therapeutic agent art-recognized as being useful to treat the disease or condition being treated by the antibody of the present invention. The additional agent also can be an agent that imparts a beneficial attribute to the therapeutic composition e.g., an agent, which effects the viscosity of the composition.

It should further be understood that the combinations which are to be included within this invention are those combinations useful for their intended purpose. The agents set forth below are illustrative for purposes and not intended to be limited. The combinations, which are part of this invention, can be the antibodies of the present invention and at least one additional agent selected from the lists below. The combination can also include more than one additional agent, e.g., two or three additional agents if the combination is such that the formed composition can perform its intended function.

Non-limiting examples of therapeutic agents for multiple sclerosis with which an antibody, or antibody portion, of the invention can be combined include the following: corticosteroids; prednisolone; methylprednisolone; azathioprine; cyclophosphamide; cyclosporine; methotrexate; 4-aminopyridine; tizanidine; interferon-ß1a (AVONEX; Biogen); interferon-ß1b (BETASERON; Chiron/Berlex);

interferon α-n3) (Interferon Sciences/Fujimoto), interferon-α (Alfa Wassermann/J&J), interferon ß1A-IF (Serono/Inhale Therapeutics), Peginterferon α 2b (Enzon/Schering-Plough), Copolymer 1 (Cop-1; COPAXONE; Teva Pharmaceutical Industries, Inc.); hyperbaric oxygen; intravenous immunoglobulin; clabribine; antibodies to or antagonists of other human cytokines or growth factors and their receptors, for example, TNF, LT, IL-1, IL-2, IL-6, IL-7, IL-8, IL-23, IL-15, IL-16, IL-18, EMAP-II, GM-CSF, FGF, and PDGF. Antibodies of the invention, or antigen binding portions thereof, can be combined with antibodies to cell surface molecules such as CD2, CD3, CD4, CD8, CD19, CD20, CD25, CD28, CD30, CD40, CD45, CD69, CD80, CD86, CD90 or their ligands. The antibodies of the invention, or antigen binding portions thereof, may also be combined with agents, such as methotrexate, cyclosporine, FK506, rapamycin, mycophenolate mofetil, leflunomide, NSAIDs, for example, ibuprofen, corticosteroids such as prednisolone, phosphodiesterase inhibitors, adensosine agonists, antithrombotic agents, complement inhibitors, adrenergic agents, agents which interfere with signalling by proinflammatory cytokines such as TNFalpha or IL-1 (e.g. IRAK, NIK, IKK, p38 or MAP kinase inhibitors), IL-1ß converting enzyme inhibitors, TACE inhibitors, T-cell signaling inhibitors such as kinase inhibitors, metalloproteinase inhibitors, sulfasalazine, azathioprine, 6-mercaptopurines, angiotensin converting enzyme inhibitors, soluble cytokine receptors and derivatives thereof (e.g. soluble p55 or p75 TNF receptors, sIL-1RI, sIL-1RII, sIL-6R) and antiinflammatory cytokines (e.g. IL-4, IL-10, IL-13 and TGFbeta).

Particular examples of therapeutic agents for multiple sclerosis in which the antibody or antigen binding portion thereof can be combined to include interferon-beta, for example, IFNß1a and IFNß1b; copaxone, corticosteroids, caspase inhibitors, for example inhibitors of caspase-1, IL-1 inhibitors, TNF inhibitors, and antibodies to CD40 ligand and CD80.

Particularly, the binding proteins and antibodies of the present invention may be utilized to treat an amyloidosis, for example, Alzheimer's disease and Down's syndrome. It should be understood that the binding proteins and antibodies of the invention can be used alone or in combination with at least one additional agents suitable for treating one of the above diseases. Said at least one additional agent may be selected by the skilled artisan for its intended purpose. For example, the additional

agent can be a therapeutic agent such as a cholesterinase inhibitor (e.g., tactrine, donepezil, rivastigmine or galantamine) , a partial NMDA receptor blocker (e.g., memantine) , a glycosaminoglycan mimetic (e.g., Alzhemed) , an inhibitor or allosteric modulator of gamma secretase (e.g., R- flurbiprofen) , a luteinizing hormone blockade gonadotropin releasing hormone agonist (e.g., leuprorelin) , a serotinin 5-HTlA receptor antagonist, a chelatin agent, a neuronal selective L-type calcium channel blocker, an immunomodulator, an amyloid fibrillogenesis inhibitor or amyloid protein deposition inhibitor (e.g., M266) , another antibody (e.g., bapineuzumab) , a 5-HTla receptor antagonist, a PDE4 inhibitor, a histamine agonist, a receptor protein for advanced glycation end products, a PARP stimulator, a serotonin 6 receptor antagonist, a 5-HT4 receptor agonist, a human steroid, a glucose uptake stimulant which enhanced neuronal metabolism, a selective CBl antagonist, a partial agonist at benzodiazepine receptors, an amyloid beta production antagonist or inhibitor, an amyloid beta deposition inhibitor, a NNR alpha-7 partial antagonist, a therapeutic targeting PDE4, a RNA translation inhibitor, a muscarinic agonist, a nerve growth factor receptor agonist, a NGF receptor agonist and a gene therapy modulator (i.e., those agents currently recognized, or in the future being recognized, as useful to treat the disease or condition being treated by the antibody or binding protein of the present invention) . The additional agent also can be an agent that imparts a beneficial attribute to the therapeutic composition e.g., an agent that affects the viscosity of the composition.

The antibodies of the invention, or antigen binding portions thereof, may also be combined with agents, such as alemtuzumab, dronabinol, Unimed, daclizumab, mitoxantrone, xaliproden hydrochloride, fampridine, glatiramer acetate, natalizumab, sinnabidol, a-immunokine NNSO3, ABR-215062, AnergiX.MS, chemokine receptor antagonists, BBR-2778, calagualine, CPI-1189, LEM (liposome encapsulated mitoxantrone), THC.CBD (cannabinoid agonist) MBP-8298, mesopram (PDE4 inhibitor), MNA-715, anti-IL-6 receptor antibody, neurovax, pirfenidone allotrap 1258 (RDP-1258), sTNF-R1, talampanel, teriflunomide,TGF-beta2, tiplimotide, VLA-4 antagonists (for example, TR-14035, VLA4 Ultrahaler, Antegran-ELAN/Biogen), interferon gamma antagonists, IL-4 agonists.

The pharmaceutical compositions of the invention may include a "therapeutically effective amount" or a "prophylactically effective amount" of an

antibody or antibody portion of the invention. A "therapeutically effective amount" refers to an amount effective, at dosages and for periods of time necessary, to achieve the desired therapeutic result. A therapeutically effective amount of the antibody or antibody portion may be determined by a person skilled in the art and may vary according to factors such as the disease state, age, sex, and weight of the individual, and the ability of the antibody or antibody portion to elicit a desired response in the individual. A therapeutically effective amount is also one in which any toxic or detrimental effects of the antibody, or antibody portion, are outweighed by the therapeutically beneficial effects. A "prophylactically effective amount" refers to an amount effective, at dosages and for periods of time necessary, to achieve the desired prophylactic result. Typically, since a prophylactic dose is used in subjects prior to or at an earlier stage of disease, the prophylactically effective amount will be less than the therapeutically effective amount.

Dosage regimens may be adjusted to provide the optimum desired response (*e.g.*, a therapeutic or prophylactic response). For example, a single bolus may be administered, several divided doses may be administered over time or the dose may be proportionally reduced or increased as indicated by the exigencies of the therapeutic situation. It is especially advantageous to formulate parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the mammalian subjects to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on (a) the unique characteristics of the active compound and the particular therapeutic or prophylactic effect to be achieved, and (b) the limitations inherent in the art of compounding such an active compound for the treatment of sensitivity in individuals.

An exemplary, non-limiting range for a therapeutically or prophylactically effective amount of an antibody or antibody portion of the invention is 0.1-20 mg/kg, more particularly 1-10 mg/kg. It is to be noted that dosage values may vary with the type and severity of the condition to be alleviated. It is to be further understood that for any particular subject, specific dosage regimens should be adjusted over time according to the individual need and the professional judgment of the person

administering or supervising the administration of the compositions, and that dosage ranges set forth herein are exemplary only and are not intended to limit the scope or practice of the claimed composition.

It will be readily apparent to those skilled in the art that other suitable modifications and adaptations of the methods of the invention described herein are obvious and may be made using suitable equivalents without departing from the scope of the invention or the embodiments disclosed herein. Having now described the present invention in detail, the same will be more clearly understood by reference to the following examples, which are included for purposes of illustration only and are not intended to be limiting of the invention.

EXPERIMANTAL PART

Example 1: Preferred anti-huRAGE antibodies

1.1 Production of hybridomas and antibodies.

Balb/c and A/J mice, 4-6 weeks of age, were immunized and boosted subcutaneously with human RAGE. Animals were injected every three weeks, beginning with a primary injection of 30 µg in complete Freund's adjuvant and injection boosts of 30 µg in Incomplete Freund's Adjuvant. Mice selected for fusion were injected intravenously with 10 µg hRAGE in saline, four days prior to fusion. Spleens from immunized animals were removed and single cell suspensions were prepared. SP2/0 myeloma cells were harvested from culture and washed. Spleen cells and tumor cells were mixed at a ratio of 5:1 and fused using 50% PEG 3000 using standard techniques (Kohler and Milstein, 1975). Fused cells were seeded in 96 well plates in selective media, at a density of $2.5x10^5$ spleen cells per well. Fusions were incubated at 37°C for 7-10 days. When macroscopic colonies were observed, supernatants were removed and tested in the hRAGE ELISA.

Hybridomas that were producing mAbs with desired characteristics were subcloned by the limiting dilution method. Supernatant containing subclones were assayed for binding to hRAGE by ELISA. Heavy and light chain subclasses of the mAbs were determined using the Zymed EIA Isotyping kit.

1.2. Determination of the amino acid sequence of the variable region for each murine anti-human RAGE mAb.

For each amino acid sequence determination, approximately $10x10^6$ hybridoma cells were isolated by centrifugation and processed to isolate total RNA with Trizol (Gibco BRL/Invitrogen, CA.) following manufacturer's instructions. Total RNA was subjected to first strand DNA synthesis using the SuperScript First-Strand Synthesis System (Invitrogen, CA) per the manufacturers instructions. Oligo(dT) was used to prime first-strand synthesis to select for poly(A)$^+$ RNA. The first-strand cDNA product was then amplified by PCR with primers designed for amplification of murine immunoglobulin variable regions (Ig-Primer Sets, Novagen, WI). PCR products were resolved on an agarose gel, excised, purified, and then subcloned with the TOPO Cloning kit into pCR2.1-TOPO vector (Invitrogen, CA) and transformed into TOP10 chemically competent *E. coli* (Invitrogen, CA). Colony PCR was performed on the transformants to identify clones containing insert. Plasmid DNA was isolated from clones containing insert using a QIAprep Miniprep kit (Qiagen, Valencia, CA). Inserts in the plasmids were sequenced on both strands to determine the variable heavy or variable light chain DNA sequences using M13 forward and M13 reverse primers (Fermentas Life Sciences, Hanover MD). Variable heavy and variable light chain sequences of the 3 monoclonal antibodies 7F9, 11E6 and 4E5 and their three variable heavy chain CDRs and three variable light chain CDRs are listed in Table 4, above.

1.3. Construction and expression of recombinant anti human RAGE antibodies

The DNA encoding the heavy chain constant region of murine anti-human RAGE monoclonal antibodies 7F9, 11E6 and 4E5 was replaced by a cDNA fragment encoding the human IgG1 constant region by homologous recombination in bacteria. The light chain constant region of each of these antibodies was replaced by a human kappa constant region (Table 1, above). Full-length chimeric antibodies were transiently expressed in COS cells or 293 cells by co-transfection of chimeric heavy and light chain cDNAs ligated into the pBOS or pTT3 expression plasmid (Mizushima and Nagata, Nucleic Acids Research 1990, Vol 18, pg 5322). Cell supernatants containing recombinant chimeric antibody were purified by Protein A Sepharose chromatography and bound antibody was eluted by addition of acid buffer. Antibodies were neutralized and dialyzed into PBS.

1.4. ELISA binding of recombinant anti-human RAGE mAbs and of hybridoma-derived anti-human RAGE mAbs.

The purified chimeric anti-human RAGE monoclonal antibodies were tested for their ability to bind human RAGE in a competition ELISA. Recombinant chimeric anti-human RAGE monoclonal antibodies or hybridoma-derived anti-human RAGE monoclonal antibodies were diluted in PBST + 10% Superblock (Pierce Biotech, Rockford, IL) and made up as a 2x stock at various concentrations ranging from 320 µg/mL to 0.0156 µg /mL (7F9 and 11E6) and from 160 µg /mL to 0.0078 µg /mL (4E5). Biotinylated hybridoma-derived anti-human RAGE monoclonal antibodies (7F9-biotin, 11E6-biotin and 4E5-biotin) were prepared at 8 µg /mL in PBST+10% Superblock. Equal volumes (50 µL) of each recombinant chimeric anti-human RAGE monoclonal antibodies or hybridoma-derived anti-human RAGE monoclonal antibodies and each corresponding biotinylated hybridoma-derived anti-RAGE mAbs were mixed. 50 µL of this mixture was then added to ELISA plates pre-coated with recombinant human RAGE at 2 µg /mL and incubated for 1.5 hours at room temperature. Wells were washed three times with PBS+ 0.05% Tween-20. Streptavidin HRP (1 mg/mL) was diluted 1:16000 in PBST + 10% Superblock; 50 L/well was added and the plates incubated for 1 hour at room temperature. Plates were washed 3 times with PBS+ 0.05% Tween-20. 50 µL of TMB solution (Sigma, St Louis, MO) was added to each well and incubated for 10 minutes at room temperature. The reaction was stopped by addition of 1N sulphuric acid. Plates were read spectrophotometrically at a wavelength of 450 nm. Results are shown in Figures 1A, 1B, and 1C.

Example 2: Generation of recombinant human sRAGE (husRAGE)

Recombinant husRAGE protein 293/6.1 sRAGE His 6 was expressed and purified in HEK293 cell (ATCC CRL-1573). The expression vector used for the generation of the stable expression was "pcDNA3 (-) 6.1 C HIS A".

Molecular biological standard techniques were used according to Sambrook and Russel (Molecular Cloning: A Laboratory Manual, 3rd edition, Cold Spring Harbor Laboratory Press. 2001). Total RNA from human lymphocytes (PBL) was reverse transcribed into cDNA using Superscript RT-PCR system (Invitrogen, Carlsbad USA). Using the oligonucleodtide primers RAGE-SE : CCG AAT TCC GGA AGC AGG ATG GCA GCC G (SEQ ID NO: 81) and RAGE-AS: CCC TCG AGC CCC TCA AGG CCC TCA GTA CTA CT (SEQ ID NO: 82), the RAGE cDNA was amplified from the cDNA (obtained above) yielding the RAGE cDNA as described in reference sequence NM-001136. The PCR-fragment was run on an

agarose gel, purified and extracted with QIAquick Gelextraction Kit (Qiagen GmbH, Germany). Afterwards the cDNA was cut with restriction endonucleases EcoR1 and XhoI. The resulting fragment was gel purified and ligated into vector pcDNA 3 (Invitrogen, USA) which had been precut with XhoI/EcoR1. After transformation into E.coli XL-1 blue cells (Invitrogen, USA) a positive recombinant clone was identified. The sequence of this clone was verified and the pcDNA3/RAGE 2.6 plasmid DNA isolated using plasmid mini-Kit (Qiagen, Germany). The coding region for the extracellular part of RAGE, husRAGE, was amplified from pcDNA3/RAGE 2.6 using PCR and primers N-SE A: AGT AAC GGC CGC CAG TGT GCT GGA ATT CGG A (SEQ ID NO: 83) and C-SE B: CCG GTA CCA CCT GCA GTT GGC CCC TCC TCG CC (SEQ ID NO: 84). The resulting PCR product was cut with restriction endonucleases EcoR1 und Kpn1, gel purified as described above and ligated into "pcDNA3.1(-) Myc HIS“ (Invitrogen, USA) which had been precut with EcoR1/Kpn1. The resulting plasmid "pcDNA 3 (-) 6.1 C HIS A“ was transfected into HEK293 cells using Superfect (Qiagen, Germany) according to manufacturers instructions. Selection of resistant cells was done using 800 µg/ml G418 in MEM Medium (#M4528, Sigma, Germany) + 10% FCS, 2 mM L-Glutamin, 100 U/ml Pennicillin/Streptavidin (Invitrogen, USA). Cloning of single cells through serial dilutions of cell suspensions lead to the identification of a clone "293/6.1 sRAGE His 6“ that secreted husRAGE into the cell culture medium as confirmed by Western Blot using a RAGE specific antibody (Santa Cruz; # sc5563). For expression and purification of sufficient amounts of husRAGE protein this clone was grown in serum containing cell culture medium (see above) in cell factories (Nunc, Germany). Cells were then switched to serum-free medium Pro293a-CDM (#12-764Q, BioWhittaker, Belgiun) and incubated for 3 days at 37 °C. 80 liters of cell free medium was harvested and concentrated using Hemoflow F-Series High-Flux columns (Fresenisus Medical Care AG, Germany) to a volume of 1400 ml.

Protein purification was done using immobilized metal ion affinity chromatography (IMAC) by Diarect AG (Freiburg, Germany) and sepharose FF for chelation (Amersham-Bioscience, Sweden). Equilibration of the column and binding of the hexa-His containing protein from cell supernatants to the matrix were done according to instructions by the manufacturer. Elution of the protein was done using step gradients with increasing concentrations of imidazole. Eluted fractions were analyzed for protein containing hexa-His using Western Blots and anti HIS

antibodies. Purified husRAGE eluted specifically at 250 –500 mM imidazole. Positive fractions were combined, concentrated and dialyzed 3 times against PBS (2 x 4h, 1 x 16 h).

A N-terminal shortened version of husRAGE (102-331-sRAGE-HIS) missing the first 101 amino acids of human RAGE was generated by standard techniques in molecular biology as described above for the husRAGE protein. This protein was generated by the same basic procedure used for husRAGE (1-331). Using the plasmid described above (pcDNA3/RAGE 2.6) and two primers (CGA AGC TTG ATG AAC AGG AAT GGA AAG GAG ACC AAG (SEQ ID NO: 85) and TCC TCG AGC ACC TGC AGT TGG CCC CTC CTC GCC T (SEQ ID NO: 86)) the shorter version of the DNA for husRAGE was amplified by PCR. After agarose gel and elution of the fragment the resulting pure fragment was cleaved with restriction endonucleases HindIII and XhoI and purified again using agarose gel and elution. The fragment was ligated into psecTAG 2A (Invitrogen, USA) that was precut with restriction endonucleases HindIII and XhoI. After transformation into E.coli "TOP10 One Shot" cells (Invitrogen, USA) a positive clone was picked and the plasmid DNA isolated. The DNA in the expression vector was transfected into HEK293 F cells using the Freestyle expression system (Invitrogen, USA). After 96 hours of expression the cell free supernatant was used for the purification using Ni-NTA Superflow beads (Quiagen, Germany). Equilibration and binding were done according to instruction by the manufacturer. Bound protein was eluted in buffer (PBS, 160 mM NaCl, 150 mM Imidazole, pH8.0). Fractions containing protein were combined and dialyzed over night at 4°C against TBS (Tris-buffered saline; pH 7,4). Purified husRAGE (102-331) concentrations were determined spectrophotometrically.

A C-terminal shortened version of husRAGE-fusion protein (1-130-sRAGE-Fc) missing the amino acids following amino acid 130 of human RAGE was generated by standard techniques in molecular biology as described above for the husRAGE protein. Using the plasmid described above (pcDNA3/RAGE 2.6) and two primers (GCACCATGGCAGCCGGAACAGCAGTTG (SEQ ID NO: 87) and GAGTCTCGAGGCAGAATCTACAATTTCTG (SEQ ID NO: 88)) the shorter version of the DNA for husRAGE was amplified by PCR. After agarose gel and elution of the fragment the resulting pure fragment was cleaved with restriction endonucleases NcoI and XhoI and purified again using agarose gel and elution. The fragment was then ligated into the plasmid pENTR4 that was precut with restriction

endonucleases NcoI and XhoI. The ligation mixtures were transformed into E.coli "TOP10 One Shot" cells (Invitrogen, USA) to generate pENTR4-RAGE 1-130. A positive clone was picked and the plasmid DNA isolated. Using site specific recombination and the gateway cloning system (Invitrogen, Carlsbad, USA; attL x attR) with DNA of the clone pENTR4 hRAGE 1-130 and DNA of the vector pcDNA3.1(+)Zeo hIgG lambda hc 257-Stop a plasmid was constructed (see below) that after transformation into "TOP10 One Shot" cells (Invitrogen, USA) and purification encoded for husRAGE-1-130-Fc (plasmid called: pEXP hRAGE 1-130/hIgG lambda hc 257-Stop). Expression of this plasmid using the Freestyle Expression system and 293F cells (Example 2.1) and purification of the resulting protein from the cell supernatant using Protein G-beads (Example 2.2) resulted in a protein with > 95 % purity.

Example 3: Construction of pcDNA3.1(+)Zeo hIgG lambda hc 257-Stop

2 oligonucleotide primers (gtacgatatcgagggacgaatggatccaccgtgcccagcacc (SEQ ID NO: 91); ctagtctagatcatttacccggagacagggag (SEQ ID NO: 92)) were used to amplify the DNA sequence for hIgG lambda heavy chain from a human placenta cDNA Library (Clontech # HL5014a) using EasyA Polymerase in a PCR, polymerase chain reaction. The resulting DNA was gel purified (as described above), cloned into pcDNA3.1 V5-His TOPO Vektor (pcDNA3.1/V5/His TOPO TA Expression Kit Invitrogen #K4800-01) using instruction from the manufacturer and transformed into E.coli TOP10 cells as described above. Positive clones were identified and the resulting plasmid DNA purified (named: pcDNA3.1(V5His) FC/hIgG lambda hc Nr.2/7) using PCR and olgionucleotide primers (gtacgatatcgagggacgaatggatccaccgtgcccagcacc (SEQ ID NO: 93); ctagtctagatcatttacccggagacagggag (SEQ ID NO: 94)). The hIgG lambda hc part of the DNA was amplified, cut with EcoRV/XbaI, ligated to EcoRV/XbaI precut pcDNA3.1(+)Zeo vector DNA and transformed into E.coli TOP10 cells. The resulting plasmid was named: pcDNA3.1(+)Zeo hIgG lambda hc 257-Stop and used for further work to express proteins N-terminally fused in frame to a C-terminal part of immunoglobulin IgG heavy chains.

3.1. Transfection and Expression of proteins in HEK293F cells

HEK 293F cells that had been grown in culture for 2 – 3 days in Free Style 293 Expression Medium were centrifuged at 400 g and the supernatant discarded. The cell pellet was resuspended in medium and adjusted to 3x107 cells in 28 ml fresh

medium, transferred to a 125 ml Erlenmeyer and incubated in an incubator at 37°C, 8% CO2 on an orbital shaker at 150 rpm until the transfection mixture was set up.

Transfection mixtures with 293fectin-DNA complex were set up as follows:

(i) 30 µg of DNA were diluted with Opti-MEM I to a total volume of 1000 µl (control 1000 µl Opti-MEM I) and mixed.

(ii) 35 µl of 293fectin (Invitrogen #12347-019; 1ml) were diluted with Opti-MEM I to a total volume of 1000 µl, mixed and incubated for 5 min at room temperature.

DNA mixture and 239fectin-solution from (i) and (ii) were transferred to a new tube, mixed slightly and after incubation for 25 minutes at room temperature were added to the cells in the Erlenmeyer.

Cells were incubated with this transfection mixture for the indicated time in an incubator at 37°C, 8% CO2 on an orbital shaker at 150 rpm. Cell supernatants were harvested by centrifugation at 400 g for 10 minutes.

3.2. Purification of RAGE-Fc fusion proteins using Protein G-Sepharose

To couple the protein from cell supernatants to beads, beads (protein G-sepharose 4 Fast Flow (Amersham Bioscience) were washed 3 times in PBS by suspending the beads in PBS and centrifugation at 13,500 rpm, discarding the supernatant. Beads were incubated with the respective cell supernatants (300 ml cell supernatants per ml beads) to be coupled for 1 – 2 hours on a rotator at room temperature. The beads were washed 3 times with PBS and incubated with the cell supernatants for 12 hours or overnight at 4°C. After incubation the beads were washed 3 times with PBS as above. Bound protein was eluted by adding 200 µl 140mM NaCl + 0.1M glycine to the bead pellet and incubating for 30 minutes on a rotator. After centrifugation the supernatant was immediately neutralized by adding 2 M Tris to adjust the pH to 7.1 – pH 7.4. The bead pellet was discarded. Obtained Probes were dialyzed against PBS and stored frozen in aliquots at –20°C. RAGE-Fc fusion protein containing the full extracellular ectodomain of RAGE was obtained from R&D systems (no. 1145-RG; Recombinant Human RAGE/Fc Chimera).

3.3. Dot blot binding of antibodies to peptides or fragments of RAGE in a non denatured form.

Dot blots were used to evaluate the binding of antibodies to peptides or fragments of RAGE in a non-denatured form. Proteins used were either sRAGE-Protein (1-331 sRAGE-HIS) or a N-terminal shortened version (102-331-sRAGE-

HIS). Peptides were ordered and synthesized by Biotrend according to standard methods (solid phases peptide synthesis on AMS 222 synthesizer using Fmoc/tBu-chemistry) containing a free carboxyl terminus. Peptides were HPLC-purified and analysis of every peptide for purity was done using RP-HPLC. All peptides had a purity of >80%. The identities of peptides were verified by mass spectrometry.

Peptides used were 30mers spanning the extracellular region of the human RAGE protein. Net charge of most peptides was similar.

NtermR31: QNITARIGEPLVLKCKGAPKKPPQRLEWKLN
Net charge:: +7 (SEQ ID NO: 70)

Peptide 1: KLNTGRTEAWKVLSPQGGGPWDSVARVLPN
Net charge: +2 (SEQ ID NO: 71)

Peptide 2: LPNGSLFLPAVGIQDEGIFRCQAMNRNGKE
Net charge: 0 (SEQ ID NO: 72)

Peptide 3: GKETKSNYRVRVYQIPGKPEIVDSASELTA
Net charge: +1 (SEQ ID NO: 73)

Peptide 4: LTAGVPNKVGTCVSEGSYPAGTLSWKLDGK
Net charge: +1 (SEQ ID NO: 74)

Peptide 5: DGKPLVPNEKGVSVKEQTRRHPETGLFTLQ
Net charge: +2 (SEQ ID NO: 75)

Peptide 6: TLQSELMVTPARGGDPRPTFSCSFSPGLPR
Net charge: +1 (SEQ ID NO: 76)

Peptide 7: LPRHRALRTAPIQPRVWEPVPLEEVQLVVE
Net charge: +2 (SEQ ID NO: 77)

Peptide 8: VVEPEGGAVAPGGTVTLTCEVPAQPSPQIH
Net charge: -2 (SEQ ID NO: 78)

Peptide 9: QIHWMKDGVPLPLPPSPVLILPEIGPQDQG
Net charge: +0 (SEQ ID NO: 79)

Peptide 10: DQGTYSCVATHSSHGPQESRAVSISIIEPG
Net charge: -1 (SEQ ID NO: 80)

Dots consisting of different amount of protein / peptide (30 ng, 10 ng, 3 ng, 1 ng, 0.3 ng, 0.1 ng, 0.03 ng, and 0.01 ng) in a volume of 1 µl in 1xPBS were spotted onto a Hybond-ECL Nitrocellulose Membrane (Amersham, RPN68D) in duplicates. Membranes were dried and unspecific binding was blocked by shaking the

membranes for 1 hour at room temperature with Western Blocking reagent (Roche, no. 1921673). Blocking reagent was discarded and the membranes were incubated with antibodies in a concentration of 7.14 nM (shaking, 1 hour at room temperature). Monoclonal antibodies were ML37-7F9, ML37-11E6, ML37-4E5, commercially available antibodies from R&D systems (e.g. AF1145). Blots were washed 4 times (each time 5 minute incubation with shaking at room temperature) with 1xPBS. Blots were then incubated with goat anti mouse IgG AP secondary antibody (Sigma no. A-7434) diluted 1:2000 in Western Blocking Reagent (Roche, no. 192173). Incubation for 1 hour was as before (shaking, room temperature). The filters were washed 4 times (5 minutes each) in 1X PBS. Development of signals was according to manufacturers instructions with NBT/BCIP substrate solution (Roche, no. 1697471). Color development was stopped after 10 minutes with bi-distilled water. See Figure 2.

Although husRAGE was detected in dot blots by all three monoclonal antibodies of the present invention and by polyclonal antibody AF1145 (from a commercial source, R&D) none of the peptides were detected by the monoclonal antibodies of the present invention. However the polyclonal antibody detected several peptides. Peptide 9, which did include the amino acid sequence used to generate polyclonal antibodies as described by Ostendorp et al. (EMBOJ. 26,3875,2007), was clearly detected by the commercially available polyclonal antibody. These results indicate that the monoclonal antibodies of the present invention clearly recognize a different epitope than currently available antibodies.

Further characterization of the binding was done by analysis of RAGE mutants of human sRAGE expressed in *E. coli*. Monoclonal antibodies 11E6 and 4E5 bind to a region around the C2 domain, since binding is lost in deletion mutants lacking amino acids 235-336 and binding, is apparent in mutant RAGE protein consisting of the amino acids 235-336.

Example 4: Interaction between Aβ1-42-globulomer and protein derived from husRAGE using HTRF technology.

The assay is based on the HTRF (Homogeneous time resolved fluorescence) technology available from CIS Bio International (Bagnols, France),

HTRF Donor- and Acceptor-components, Anti-6HIS-Europiumcryptate (CIS Bio catalogue no.: 61HISKLA; 500 wells/13 µg) and Streptavidin XL – 665 (CIS Bio catalogue no.: 611SAXLA, 500 wells/250µg), were each dissolved in 250 µl bi-

distilled water. These stock solutions were diluted 100-fold in PBS, 0.1 % BSA, pH 7.4 to obtain working solutions with final concentrations of 3.7 nM Anti6His-Cryptate and 60.6 nM Streptavidin XL-665. Solutions of 10 µM, 5 µM, 2.5 µM, 1.25 µM, 0.625 µM, 312.5 nM, 156.25 nM of the biotinylated Aß-globulomer (1/5 of Aβ1-42 peptide used to prepare the Aß-globulomers were Biotinyl-Amyloid ß-Protein (1-42) (Bachem no. H-5642) were prepared according to Barghorn et al. (J. Neurochemistry, vol. 95, no 3, pp. 834-847, 2005 and WO/2007/062852; International Application No. PCT/EP2006/011530); the used globulomer concentration was calculated based on the concentration of Aß1-42 monomers, which were used for the generation of the globulomers. Solutions of 10 µM, 5 µM, 2.5 µM, 1.25 µM, 0.625 µM, 0.312 µM, 0.156 µM, of the biotinylated Aß-globulomer was prepared in the same buffer (PBS, 0.1% BSA, pH 7.4). 4 µl of these solutions or 4 µl of buffer were mixed with 4 µl of 1 µM recombinant husRAGE protein and the solution incubated at room temperature for 1h, followed by the addition of 4 µl of each of the solutions (3.7 nM Anti6His-Cryptate and 60.6 nM Streptavidin XL-665).

The assay was incubated for 2 hours at 4°C. After addition of 4 µl of a 2M KF stock solution the HTRF signal was measured in HTRF mode in a BMG Pherastar fluorescence instrument (BMG Labtech GmbH, Germany). Maximum signal curves without antibody and background results using only Anti6HIS-Cryptate – or Streptavidin XL – solution were used. %DeltaF values were calculated according to instructions by the manufacturer CisBio using GraphPad Prism 4 (GraphPad Software, San Diego, USA).

Example 5: Inhibition of Aβ1-42-globulomer binding to husRAGE by antibodies using HTRF technology

The basic protocol as described above was used with few modifications. HTRF Donor- and Acceptor-components were diluted 40-fold to 10.25 nM for Anti-6HIS-Europiumcryptate and 151.5 nM for Streptavidin XL-665 in PBS, pH 7.4, 0.1% BSA.

Purified monoclonal antibodies (MABs) were used against husRAGE or control immunoglobulins (mouse IgG1 and mouse IgG2a; no. M-5284 rsp. No. M-5409; Sigma, Germany) as control antibodies.

The assay was performed in a total volume of 20 µl in 384 well plates. For each assay point: 4 µl of 1 µM husRAGE was incubated with 4 µl of test antibody or

IgG control-antibodies in concentrations of 2 µM, 1 µM, 0.5 µM, 0.25 µM, 0.125 µM, 62.5 nM, 31.25 nM, 15.62 nM, 7.81 nM, 3.9 nM for 1 hour at room temperature. Control for background was done without husRAGE and without antibodies. Maximum signal was obtained without antibodies. Subsequently, 4 µl of 800 nM biotinylated Aß-Globulomer was added, as well as 2 µl of 10.25 nM for Anti-6HIS-Europiumcryptate and 151.5 nM for Streptavidin XL-665. Differences in volume were adjusted by adding binding buffer (1xPBS pH 7.4; 0.1% BSA). The assay was incubated for another hour. After addition of 4 µl of a 2M KF stock solution the HTRF signal was measured in HTRF mode in a BMG Pherastar fluorescence instrument (BMG Labtech GmbH, Germany). Maximum signal curves without antibody and background results using only Anti6HIS-Cryptate – or Streptavidin XL – solution were used. %Delta F values were calculated according to instructions by the manufacturer CisBio using GraphPad Prism 4 (GraphPad Software, San Diego, USA). Results are shown in Figures 3A, 3B, and 3C. Concentrations indicated in the figures are the final concentrations of the proteins in 20 µl assay volume.

As shown in figure 4, husRAGE expressing all three domains of RAGE did bind to amyloid Aβ-globulomers. A RAGE mutant protein consisting of human sRAGE lacking most of the v-domain (RAGE102-331) did bind with higher affinity to amyloid Aβ-globulomers indicating that the domain within human RAGE for binding to Aβ-globulomers is within the C-terminus.

Example 6: Binding of Aβ-globulomers to RAGE proteins using the ALPHA screen assay technology

This assay was performed in assay buffer (25 mM HEPES, 100 mM NaCl pH 7.4 and 0.1% BSA) in a volume of 20 µl. Donor beads used were Streptavidin coated (Perkin Elmer; 6760002S) and acceptor beads used were Protein A ALPHALISA (Perkin Elmer; CUSM64133000EA), 4 µl of each of the beads was pre-diluted with 196 µl of assay buffer.

Using a 384-well Proxi-Plate (Perkin Elmer, no. 6006280) donor beads were loaded with biotin-aβ-globulomers (see above) using 4 µl of the prediluted donor beads and 6µl of a 200 nM solution of biotinylated-aβ-globulomers.

Acceptor beads were loaded with different amounts of RAGE-Fc fusion proteins using 4 µl of the pre-diluted acceptor beads and 6µl of different dilutions of RAGE-Fc fusion proteins (starting with e.g. 100 µg/ml).

Loading (binding of the proteins to the beads) was done in the dark at room temperature for 30 minutes.

Binding of Aβ-globulomers to RAGE started by combining pre-loaded donor and acceptor bead preparations for additional 180 minutes in the dark. Signals were measured in an ALPHA-Quest instrument (Perkin Elmer) with a time delay of 1 second. Further analyses were done using GraphPadPrism software. In a different experiment but using the same technology, the binding of Aβ-globulomers to RAGE-Fc consisting of all three domains, was compared to the binding of Aβ-globulomers to RAGE-Fc mutant protein consisting of the v-domain only (amino acids 1-130 of huRAGE). As shown in figure 5, binding of amyloid Aβ-globulomers to the three domains of soluble RAGE was strong and binding of amyloid Aβ-globulomers to the v-domain of RAGE was negligible. Since the binding of Aβ-globulomers to RAGE takes place in the C-terminal region, antibodies preferably binding to these domains would be predicted to compete with this binding.

Example 7: Construction, expression and purification of the E. coli RAGE fragments

7.1. Preparation of constructs

The E. coli RAGE constructs listed in Table 6 were generated as follows. Construct 1 was created by PCR amplification from the template plasmid pcDNA 3 (-) 6.1 C HIS A using the forward primer (atgctacatatgaaaaagacagctatcgcgatt gcagtggcactggctggtttcgctaccgtagcgcaggccgctcaaaacatcacagcc (SEQ ID NO: 89)) and reverse primer (atgctactcgagtcagtggtggtgg tggtggtgagttcccagccctgatcctcccacagagcctgcagttggcccctcc (SEQ ID NO: 90)) which introduced Nde I and Xho I restriction sites which were utilized for subcloning into the analogous sites of pET29. The remaining constructs (#2 - #7, Table 6) were generated using Construct 1 as a template. Sequences encoding RAGE amino acid residues 24-129, 24-234, 24-336, 130-234, 130-336, 235-336 were PCR amplified from Construct 1. The resulting DNA fragments were run on a 1.0% agarose gel, and the DNA purified using the QIAquick Gel Extraction Kit from Qiagen. The DNA fragments were digested with NdeI and XhoI, and ligated into similarly digested pET28a. The ligation mix was transformed into Max Efficiency DH5α competent cells and plated onto LB agar plates containing 50 mg/L kanamycin. After overnight incubation at 37°C, three colonies for each clone were inoculated into 3 ml of LB broth containing 50 mg/L kanamycin and shaken overnight at 37°C. The DNA was

isolated using the QIAprep Spin Miniprep Kit from Qiagen, and the insert sequenced using T7 promoter and T7 terminator specific primers. The DNA sequence of the plasmids encoding constructs #1 - #7 are listed as SEQ ID Nos.: 27-33, and the corresponding translated regions are listed as SEQ ID Nos 34-40.

Table 6: RAGE constructs

Construct #	Protein Form Name	Protein SEQ ID	Plasmid SEQ ID
1	OmpA-[RAGE (23-340)]-6His	34	27
2	6His-(Thr)-[RAGE (24-129)]	35	28
3	6His-(Thr)-[RAGE (24-234)]	36	29
4	6His-(Thr)-[RAGE (24-336)]	37	30
5	6His-(Thr)-[RAGE (130-234)]	38	31
6	6His-(Thr)-[RAGE (130-336)]	39	32
7	6His-(Thr)-[RAGE (235-336)]	40	33

E. coli strain BL21(DE3) was transformed with Construct #1 plasmid DNA, plated on LB plates containing kanamycin (50 mg/L), and incubated at 37°C overnight. The next day 14 fernbach flasks, each containing 1L of Terrific Broth with kanamycin (50 mg/L), were inoculated with a CFU and placed shaking (180 rpm) in an incubator at 37°C. When the cultures reached an OD_{600nm} of 0.47, the flasks were transferred to a 30°C incubator (still shaking at 180 rpm) and expression was induced by addition of 0.4 mM IPTG. Cells were harvested 4 hours after induction by centrifugation (15,900 g, 8 minutes, 4°C), and the cell paste then frozen at -80°C until purification.

Purification of RAGE Construct 1 proceeded by first thawing and resuspending an ~20g cell pellet in 180 ml of lysis buffer [50 mM Tris pH 7.6, 300 mM NaCl, 10% glycerol, 0.1% triton X-100, 0.5 mM $MgCl_2$, 20 mM imidazole, 1X Roche EDTA-free protease inhibitors, 20 U/ml DNase I]. Cells were lysed by passing the suspension three consecutive times through an Avestin Emulsiflex microfluidizer at 3°C. Clarified lysate was then loaded onto a 5 ml HiTrap IMAC-column (GE Healthcare, 17-5255-01) at 2 ml/min. The column was then washed with 10 CV of wash buffer [50 mM Tris pH 7.6, 300 mM NaCl, 10% glycerol, 20 mM imidazole]. Following the wash step, RAGE was gradient eluted using elution buffer [50 mM Tris pH 7.6, 300 mM NaCl, 10% glycerol, 500 mM imidazole]. Fractions containing RAGE were pooled and then dialyzed against 50mM Tris pH 7.6, 20mM NaCl, 10%

glycerol. Mass-spec analysis of the purified material confirmed the OmpA-leader had been processed off and that the purified material began with residue 23 of RAGE (i.e. the sequence A-Q-N-...) as expected.

Plasmids encoding Constructs #2- #7 were separately transformed into *E. coli* strain BL21(DE3), plated onto LB plates containing kanamycin (50mg/L), and incubated at 37°C overnight. The next day 1L of Overnight Express Instant TB Medium (Novagen) was inoculated with a colony and shaken for 19 hours at 30°C. The cells were pelleted by centrifugation (15,900 xg, 10 minutes, 4°C) and then frozen at -80°C. The pellets (5-6 grams each) were thawed and resuspended in 50 ml of lysis buffer [50 mM Tris, pH 8, 300 mM NaCl, 0.1% Triton X-100, 10% glycerol, 0.2 mg/ml lysozyme, 1 ml of protease inhibitor cocktail set III (Calbiochem), 20U/ml benzonase, 5 mM B-mercaptoethanol]. The lysates were sonicated on a Vibra Cell Sonicator for 2 minutes, followed by centrifugation at 20K x g for 30 minutes. Econo-Pac 10 columns from Bio-Rad were filled with a 2 ml bed volume of ProBond Nickel Resin and equilibrated with lysis buffer. The clarified lysates were passed through the columns 3 consecutive times, followed by washing with 3 x 10 column volumes (60 ml total) of wash buffer [2X PBS, 20 mM imidazole, 10% glycerol, 5 mM B-mercaptoethanol]. The proteins were eluted off the columns with 5 x 1 column volume (10 ml total) of elution buffer [2X PBS, 500 mM imidazole, 10% glycerol, 5 mM B-mercaptoethanol]. The eluted material was transferred into PBS, 10% glycerol, and 1 mM DTT using Bio-Rad Econo-Pac 10DG Columns.

7.2. Expression of anti-RAGE monoclonal antibodies 11E6, 4E5 and 7F9

The media used for hybridoma cell expansion consisted of BD Cell MAb Quantum Yield Medium (Becton Dickenson – catalog # 220511) containing 10% ultra low IgG fetal bovine serum (Invitrogen - catalog #16250-078). Briefly, multiple 300 ml seed cultures of the murine hybridoma cell line expressing RAGE monoclonal antibody 11E6 were expanded in a 2 L roller bottle shaking in an incubator (65 rpm, 8% CO_2, 37°C) until reaching a density of $1.0x10^6$ cell/ml. Cells were then seeded into 20L of media at a density 0.06 x 10^6 cells/mL in a 25L Wave BioReactor with operational settings of 14 rocks/minute, a rock angle of 6°, temperature of 37°C, and an 8% CO_2 sparge-rate of 0.15 Lpm. After two days, the culture was further expanded by addition of media to a final volume of 24 L, resulting in a new cell density of 0.43 x 10^6 cells/mL. The culture was harvested 12 days after being expanded to full volume. Cells were removed by continuous centrifugation (Carr

ViaFuge, 6000 rpm, 1.7 Lpm). After addition of 5 mM NaN_3 (from a 1 M NaN_3 stock – Hampton Research) to the clarified media, the material immediately was utilized in the purification process.

The media used for hybridoma cell expansion consisted of BD Cell MAb Quantum Yield Medium (Becton Dickenson – catalog # 220511) containing 10% ultra low IgG fetal bovine serum (Invitrogen - catalog #16250-078). Briefly, multiple 300ml seed cultures of the murine hybridoma cell line expressing RAGE monoclonal antibody 4E5 were expanded in a 2 L roller bottle shaking in an incubator (65 rpm, 8% CO_2, 37°C) until reaching a density of $1.0x10^6$ cell/ml. Cells were then seeded into 5L of media at a density 0.12 x 10^6 cells/mL in a 25L Wave BioReactor with operational settings of 12 rocks/minute, a rock angle of 6°, temperature of 37°C, and an 8% CO_2 sparge-rate of 0.15 Lpm. After four days, the culture was further expanded by addition of media to a final volume of 24 L, resulting in a new cell density of 0.24 x 10^6 cells/mL. The rock-rate was increased to 14 rocks/minute. The culture was harvested 12 days after being expanded to full volume. Cells were removed by continuous centrifugation (Carr ViaFuge, 6000 rpm, 1.7 Lpm). After addition of 5mM NaN_3 (from a 1 M NaN_3 stock – Hampton Research) to the clarified media, the material immediately was utilized in the purification process.

The media used for hybridoma cell expansion consisted of BD Cell MAb Quantum Yield Medium (Becton Dickenson – catalog # 220511) containing 10% ultra low IgG fetal bovine serum (Invitrogen - catalog #16250-078). Briefly, multiple 300ml seed cultures of the murine hybridoma cell line expressing RAGE monoclonal antibody 7F9 were expanded in a 2 L roller bottle shaking in an incubator (65 rpm, 8% CO_2, 37°C) until reaching a density of $1.0x10^6$ cell/ml. Cells were then seeded into 10L of media at a density 0.05 x 10^6 cells/mL in a 25L Wave BioReactor with operational settings of 12 rocks/minute, a rock angle of 6°, temperature of 37°C, and an 8% CO_2 sparge-rate of 0.15 Lpm. After four days, the culture was further expanded by addition of media to a final volume of 25 L, resulting in a new cell density of 0.25 x 10^6 cells/mL. The rock-rate was increased to 14 rocks/minute. The culture was harvested 10 days after being expanded to full volume. Cells were removed by continuous centrifugation (Carr ViaFuge, 6000 rpm, 1.7 Lpm). After addition of 5mM NaN_3 (from a 1 M NaN_3 stock – Hampton Research) to the clarified media, the material immediately was utilized in the purification process.

7.3. Purification of anti RAGE monoclonal antibodies 11E6, 4E5, 7F9

To the clarified hybridoma culture media, glycine and NaCl were added to final concentrations of 3M and 1.5M, respectively. The pH was adjusted to 8.0 with NaOH. Material is filtered using a 5 μm Pall Capsule #120 membrane filter and loaded onto a 200 mL BioSepra Protein A column. Protein solution was washed with 11 CV of wash buffer (20 mM sodium phosphate pH 8.0, 1mM sodium azide) and eluted using a step gradient of 50 mM glycine (pH 3.0). Fractions of 100 mL size were collected and protein concentration determined by measuring the A280nm. All column-processing steps were run at 4° C. Pooled material was dialyzed against 20 L of 10 mM Tris pH 8.0 overnight at 4° C. Further chromatographic polishing was achieved using a SartoBind Q strong basic anion exchanger Singlesep Mini cartridge (Sartorius) in flow through mode at 10 mL/min. The antibody does not bind the column and is collected as one pool. Following this step, the material was concentrated to 5 mg/ml using an Amicon stirred pressure cell (5000 MWCO membrane, 75 psi, 4°C). Finally, the antibody was dialyzed two times against 20L of PBS buffer (10 mM phophate, 0.138 M NaCl, 0.0027 M KCl, pH 7.4), each cycle for 24h at 4°C.

7.4. ELISA Binding Experiments:

Antigens (purified construct #1-7 proteins) were diluted into Coating Buffer [100 mM $NaHCO_3$, pH 8.2] to 1μg/ml, and 100 μl of the resulting solution was then aliquoted into a 96-well Nunc Immuno Plate (Maxi-Sorb Surface, flat bottom, catalog # 439454). The plate was sealed with sealing film and incubated at 4°C overnight. The next day, the plate wells were each washed 3 times with 150 μL PBST buffer [Sigma PBS + 0.05% Tween 20]. Then 300ul of blocking solution (3% NFDM in PBST) was added to each well. The plate was incubated for 2 hours at room temperature and shaking at 100 rpm. After the incubation step, each well was again washed three times with 150 uL PBST. 100 ul of the corresponding antibody to be tested (i.e. 7F9, 11E6, and 4E5) was added at various dilutions made in PBST/0.5% BSA. The plate was then sealed with sealing film and incubated for 2 hours at room temperature and shaking at 100 rpm. Next the antibody solution was drained out of the wells, and each well then washed three times with 200ul PBST. To each well then 200ul of a 1:5000 dilution (in PBST/1% NFDM) of conjugated secondary antibody [Donkey anti-mouse HRPO conjugate, Jackson Immuno Research, Catalog #715-035-150] was added. The plate was covered and allowed to incubate for 1 hour at room

temperature while shaking at 100 rpm. Following this incubation, the solution was removed from the wells and each well was washed three times with 200ul PBST. To each well 100µL of HRPO substrate [3,3',5',5'-Tetramethylbenzidine Liquid Substrate (TMB), Supersensitive for ELISA, Sigma Catalog #T4444] solution was added and the plate incubated at room temperature for 10 minutes. Finally, 50 µL of 2M H_2SO_4 was added to each well to stop the reaction and the A_{540nm} of each well was measured using a microtiter plate reader.

Results of these binding experiments are shown in Figures 6 A, 6B and 6C, and Figure 7A, 7B, and 7C. Figure 6A, 6B, and 6C shows that RAGE residues 24-234 were not involved in binding of RAGE monoclonal antibodies 11E6, 4E5, or 7F9. Conversely, as shown in Figure 7A, 7B, and 7C, RAGE residues 235-336 were sufficient for binding of RAGE monoclonal antibodies 11E6 and 4E5. RAGE mAb 7F9 did not show any binding to any of these (E. coli expressed) RAGE fragments.

Example 8: Epitope mapping

8.1. Immobilization of the 11E6, 4E5 and 7F9 Antibodies

Approximately 20 mg of CNBr-activated Sepharose fast flow resin was weighed into three compact reaction columns fitted with 35 µm frits. The resins were permitted to swell in 200 µL of 1 mM HCl before washing 3 times with 200 µL of 1 mM HCl. The resins were subsequently rinsed 3 times with 200 µL of 100 mM sodium bicarbonate buffer (pH 8.3) containing 500 mM sodium chloride (buffer A). Once completed, the solutions were flushed from the columns so that only a thin layer of buffer remained on the surface of each resin. Approximately 5.5 nmoles of the antibodies were immobilized to the resins, which required the addition of 235 µL of 7F9 (3.4 mg/mL), 500 µL of 11E6 (1.6 mg/mL), and 200 µL of 4E5 (3.75 mg/mL). For the 7F9 and 4E5 antibody immobilizations, 200 µL of buffer A was also included. The compact reaction columns were sealed and permitted to mix through inversion at room temperature for 4 hours. Once completed, the compact reaction columns were unsealed and flushed with three additions of 200 µL of buffer A to remove unbound antibody. After the flush, 200 µL of a buffer containing 100 mM Tris-HCl (pH 8) and 500 mM sodium chloride (buffer B) was added to each column. The columns were resealed and allowed to mix through inversion at room temperature to block unbound but activated sites on the resins. After 2 hours, the columns were unsealed and flushed first with 200 µL of 100 mM sodium acetate buffer (pH 4) containing 500 mM

sodium chloride (buffer C) followed by 200 μL of buffer B. This process was repeated an additional two times to ensure complete removal of unbound antibody and to fully block the remaining sites of attachment on the resins. The resins were then washed four times with 200 μL of 100 mM sodium bicarbonate buffer (pH 8.3) containing 100 mM sodium chloride (buffer D) prior to coupling the antigen.

8.2. Proteolytic Excision of the E.coli and BacMam-Expressed sRAGE Antigens

The sRAGE antigens were permitted to bind to the antibody columns by adding 75 μL of the *E.coli*-expressed antigen to the 11E6 and 4E5 resins and adding 125 μL of the *BacMam*-expressed antigen to the 11E6, 4E5 and 7E9 resins. The columns were sealed and the samples allowed to mix by inversion at room temperature for 2 hours. After this time, the columns were unsealed and flushed with 4 additions of 200 μL of buffer D. After flushing through the rinses, the resins were resuspended in 200 μL of buffer D as well as with the proteases, generated as 0.1 mg/mL solutions of either Trypsin, endoproteinase Glu-C or Chymotrypsin. The amounts of the proteases varied between the experiments to attenuate the digestions, but ranged from 200-400 fold excess of the antigen over the protease by weight. Proteolysis was permitted to occur at room temperature with mixing by inversion for 12 hours. After this time, the columns were unsealed and the proteolytic solution was flushed through and saved for further analysis. For samples subjected to dual digestion, the resins were resuspended in 200 μL of buffer D and a second protease before the next steps. For those samples not treated in a second protease, the columns were subjected to two individual 200 μL washes in buffer D followed by a 200 μL wash in buffer A then a final 200 μL wash in buffer D. Each wash was retained separately for later analysis. The epitope containing peptides were eluted from the column with three individual 200 μL washes in 2% formic acid. Each elution sample was retained separately for later analysis.

8.3. Mass Spectrometry Analysis of the Epitope-Containing Peptides

The samples were analyzed using both a Bruker Apex QE 7T Fourier-Transform Ion Cyclotron Resonance (FT-ICR) mass spectrometer as well as an Applied Biosystems Q-STAR Pulsar I mass spectrometer. For the FT-ICR mass spectrometric analysis, 8 μL of the epitope excision samples were injected onto a Jupiter C4 reversed phase column (0.5 x 150 mm, 5μ particle size, 300 Å pore size)

by an Agilent series 1100 capillary HPLC. The samples were washed for 5 minutes in 90% water with 0.1% formic acid (solvent A) / 10% acetonitrile with 0.1% formic acid (solvent B) at a 5 μL/min flow rate to desalt. The peptides were eluted into the mass spectrometer by changing the mobile phase composition to 5% solvent A / 95% solvent B. Samples requiring direct infusion for tandem mass spectrometry were injected onto a protein Microtrap (Michrom) equilibrated in 98% water, 1% acetonitrile and 1% formic acid. The samples were washed with 1 mL of the equilibration solvent before being eluted in 300 μL of 60% acetonitrile, 40% water and 0.1% formic acid. The eluent was directly infused into the FT-ICR mass spectrometer at 2 μL/min. For the Q-STAR Pulsar mass spectrometer, between 5-30 μL of sample was injected onto a protein Microtrap (Michrom) by an Agilent series 1100 HPLC. The samples were washed in 95% solvent A / 5% solvent B for 1 minute prior to elution of the bound peptides into the mass spectrometer in 5% solvent A / 95% solvent B.

Proteolytic excision of the E.coli-expressed sRAGE bound to the 11E6 antibody and elution of epitope-containing peptides revealed the presence of a peptide with a mass of 12,204.5 Da. Mass selection and collisionally-activated dissociation of the 10^+ charge state confirmed the identity of this peptide, which corresponded to the residues Val^{229}-His^{346} (this His residue is due to adding the Hexa-His-tag to the sRAGE protein).Further epitope mapping using Trypsin followed by Chymotrypsin proteolysis revealed cleavage of the C-terminal hexahistidine tag, thus refining the epitope to residues Val^{229}-His^{341} (this His residue is due to adding the Hexa-His-tag to the sRAGE protein). No further refinement of this epitope could be obtained using proteolysis. Similarly executed proteolytic excision of the E.coli-expressed sRAGE bound to the 4E5 antibody revealed the same peptide of 12,204.5 Da that was observed for the 11E6 antibody epitope. Correspondingly, excision of the BacMam-expressed sRAGE bound to either the 11E6 or 4E5 antibody revealed two major epitope containing peptides of masses 10,671.9 Da and 10,614.0 Da. These peptides match to the C-terminal of the BacMam-expressed construct, spanning residues Val^{229}-Gly^{331} and Val^{229}-Ala^{330}, respectively.

Proteolytic excision of the *BacMam*-expressed sRAGE bound to the 7F9 antibody and elution of epitope-containing peptides revealed multiple species representing several overlapping peptides. Deconvolution of the mass spectrum

revealed peptides of masses 12,079.6 Da, 12,372.9 Da, 13,477.3 Da and 24,132.3 Da. These masses match to residues Asn^{105}-Arg^{216}, Asn^{105}-Arg^{218}, Asn^{105}-Arg^{228} and Asn^{105}-Gly^{331}, respectively, suggesting a minimal epitope spanning residues Asn^{105}-Arg^{216}.

These results indicate that antibodies 11E6 and 4E5 possess epitopes on the C-termini of both E.coli and BacMam-expressed sRAGE, and that antibody 7F9 recognizes an epitope on the center domain of BacMam-expressed sRAGE.

Example 9: Biacore and surface plasmon resonance measurements

The affinity of the three monoclonal antibodies of the present invention, i.e. 7F9, 11E6 and 4E5, was measured using Biacore and surface plasmon resonance measurements.

9.1. Materials and Methods for antiRAGE binding kinetic determinations:

Biacore 2000 instrument was used to measure mouse anti-RAGE mAb binding kinetics. The assay format for mAb affinity analysis was Fc-based capture via immobilized anti-Fc antibodies. A standard amine coupling protocol was employed to immobilize Fc-specific IgG via primary amines to the carboxy-methyl (CM) dextran surface of CM5 sensorchips (Biacore). For the study of mouse anti-RAGE mAbs, anti-mouse Fc (Biacore, anti-mouse, BR-1005-14) was used as the immobilized capture reagent. An automated protocol, available on the Biacore 2000, was used to immobilize 8000-10000RU of capture reagent in all 4 flowcells of the sensorchip. Briefly, the CM–dextran surfaces were activated by freshly prepared 1:1 50mM N-hydroxysuccinimide (NHS):200mM 3-(N,N-dimethylamino) propyl-N-ethylcarbodiimide (EDC). Then the anti-Fc IgG capture reagent (20ug/ml in 10mM sodium acetate, pH4.5) was applied to the surface followed by deactivation of the surface and blocking of the residual reactive sites with 1M ethanolamine (pH 8.5).

The running buffer employed was HBS-EP+ [10mM HEPES, pH 7.4, 150mM NaCl, 3mM EDTA, 0.05% P20 surfactant (Biacore)] for the mouse anti-RAGE mAbs. Each experimental cycle consisted of the following steps: 1) anti-RAGE mAbs were captured in flowcells 2, 3 or 4 to a level of 50-200RU (depending on the antigen). All measurements were referenced against flowcell 1 which had no captured anti-RAGE mAb. 2) Antigen was injected through all 4 flowcells, 180ul at 60ul/min. After the antigen injection, dissociation was monitored for 600 seconds at 60ul/min. 3) the anti-Fc capture surface was regenerated with low pH glycine. For kinetic

determinations, RAGE injections were a 2 fold dilution series from 20nM – 0.31nM and buffer only in randomized duplicates.

9.2. Evaluation and results

Data were processed using Biacore evaluation software. Briefly, the data were double referenced by first, subtracting the signal from the reference cell and second, by subtracting the signal from buffer-only injections. The double referenced data from the RAGE injection series were then fit globally to a 1:1 (Langmuir) binding model, which included a mass transfer term, to determine the binding kinetic rate constants, ka and kd, and the affinity, K_D.

Table 7 shows that 4E5 did not bind to mouse RAGE (Mu-RAGE). 11E6 and 4E5 cross-competed with each other for binding to RAGE. 7F9 does not bind to RAGE produced in E.coli lacking glycosylation.

Table 7 also shows the specific epitopes to which the three antibodies of the present invention bound to human RAGE, via epitope mapping using protection of human sRAGE from proteolytic digestion and identification of the protected peptides with mass spectrometry. MAb 7F9 bound to C1 (Asn^{105} – Pro^{215}); mAb 4E5 bound to C2 (Val^{229} – Thr^{340} in E.coli RAGE; Val^{229} – Gly^{331} in husRAGE produced in mammalian cells), and 11E6 bound to C2 (Val^{238} – Arg^{314} in E.coli RAGE; Val^{229} – Gly^{331} in husRAGE produced in mammalian cells).

Table 7: epitopes to which the antibodies are binding

Mab	Affinity (nM)	Epitope	Mouse-RAGE
7F9	0.11	C1; glycosylation sensitive	+
11E6	0.29	C2; overlap with 4E5	+
4E5	2.2	C2; overlap with 11E6	-

Example 10: *In vivo* cerebral blood volume (CBV) studies in C57BL/6 female mice

10.1. Animals

C57BL/6 female mice (4-6 months old; Taconic, Germantown, New York, USA) were maintained on standard sterile wood chip bedding in a quiet room under conditions of 12 h lights on/12 h lights off (on at 06:00), with food and water available *ad libitum*. A total of 33 mice were used in fMRI-CBV studies. All studies

were approved and closely monitored by the Abbott institutional Animal Care and Use Committee, adhering to National Institutes of Health Guide for Care and Use of Laboratory Animals guidelines in facilities accredited by the Association for the Assessment and Accreditation of Laboratory Animal Care.

10.2. Soluble Aβ peptide preparation

Aβ1-40 (>99% pure) was synthesized at Abbott Laboratories. Briefly, E. coli. BL21(DE3) were induced with 1mM IPTG and expressed for 3 hours at 41°C for 3 hours in an 18L fermentor run. 185 gm of cell paste was harvested. The peptide was expressed as inclusion bodies. Cells were lysed in 0.1M tris buffer containing 0.1% triton X100. The pellet was then washed 3X with 50mM tris buffer and once with water. The washes were discarded and the final pellet was resuspended into water and lyophilized. The lyophilized pellet was extracted into 500mL of DMSO and diluted to 1L with 0.2% ammonia in water. This 1L sample was dialyzed against 19L of 10% ethanol containing 0.1% ammonia. Dialysis was continued with just 0.1% ammonia in water for a total time of 5 hours at room temperature. The 1.4L sample was diluted with 0.1% ammonia to 2L and applied to a 2.5 X 25cm PLRP-S HPLC column, (Polymer Labs, Amherst, MA), equilibrated with 0.1% ammonia in water. Elution was with acetonitrile containing 0.1% ammonia. Aβ1-40 eluted during a gradient from 10 to 30%B over 200 minutes. The pooled material was lyophilized. MALDI analysis was used to confirm the identity and purity of the material. The material was purified as Aβ Met-1-40, N15 labeled. A small amount of methionine sulfoxide was present at +16 mass units in the sample.138 mg were purified in this single run as the ammonium salt of the peptide. The reverse sequence, Aβ40-1 (>99% pure), was purchased from Sigma Chemical Co. (St. Louis, MO, USA). Aβ peptides (0.01 or 0.1 mg) were separately dissolved in fresh phosphate buffered saline (0.1 mL; PBS) immediately prior to every fMRI-CBV experiment. For initial studies, mice were randomly assigned into five treatment groups: PBS control, Aβ1-40 (0.01 or 0.1 mg/mouse) or Aβ40-1 (0.01 or 0.1 mg/mouse), with five animals in each group.

10.3. Antibody preparation

A negative IgG control antibody and the anti-RAGE antibody, 11E6 (both >99% pure), were synthesized at Abbott Laboratories.

10.4. CBV measurement using fMRI

All fMRI experiments were performed on a 7.0 T/21 cm horizontal magnet with a 20 G/cm magnetic field gradient insert (Biospec Bruker, Billerica, MA).

Animals were first anesthetized with meditomidine (1 mg/kg, i.p.; Pfizer Animal Health, Exton, Pennsylvania, USA)+ketamine (75 mg/kg, i.p.; Fort Dodge Animal Health, Iowa, USA) and then placed in a dual-coil small animal restrainer (Insight Neuroimaging Systems, LLC, Worcester, MA), which contains a volume coil for transmitting and a surface coil for receiving. Respiration rates and waveforms were continuously monitored via a force transducer. Rectal temperature was monitored and maintained at 37 ± 1 °C via a feedback-regulated, circulating water pad. All imaging was performed during the light phase. Coil-to-coil electromagnetic interaction was actively decoupled. Anatomical images were acquired using the fast spin-echo rapid acquisition relaxation enhanced (RARE) pulse sequence with TR = 3 s, effective TE = 100 ms, matrix = 256 x 256, FOV = 2.56 cm x 2.56 cm, nine 1.0-mm slices, and four averages. Gradient echo single-shot echo-planar imaging (EPI) was used for fMRI-CBV image acquisition with TR = 2 s, TE = 13 ms, matrix = 64 x 64, FOV = 2.56 cm x 2.56 cm, and giving an in-plane resolution = 400 μm x 400 μm. 10 mg Fe/kg ultrasmall superparamagnetic iron oxide (USPIO) contrast agent (SH U555C, Schering AG, Germany) was administered intravenously 2 min into an 18 min image acquisition. Soluble Aβ and PBS were administered via the tail vein 6 min after the contrast agent using a syringe pump (0.1 mL/min for 1 min) and changes in CBV were then detected over a subsequent 10 min period. Control IgG1 antibody or 11E6 was administered ip to mice in their home cages, 3 hours prior to the commencement of imaging studies.

10.5. fMRI Data analysis

Data analysis was performed using the Analysis of Functional NeuroImages (AFNI) software package (Cox R W, Comput Biomed Res 29:162-173, 1996). To identify time-dependent relative CBV change, rCBV(t), was calculated from time course raw data based on the relationship (Mandeville et al., 1999),

$$rCBV(t) = \ln[s(t)/s_0(t)]/\ln[s_0(t)/s_{pre}] \qquad \text{Eq.1}$$

where s(t) is the signal intensity after Aβ or PBS infusion, $S_0(t)$ is the baseline signal before the Aβ or PBS infusion, and S_{pre} is the mean sigal intensity before the administration of contrast agent. The time-course rCBV changes were detrended with a linear function to account for elimination of contrast agent from the blood (Cox, 1996).

Subsequently, the rCBV signal for each voxel in every mouse was fitted to a nonlinear differential exponential model (Eq.2) reflecting the drug's kinetics (Luo et al., 2004) where t_0 is the time delay of response, k is the multiplicative coefficient, α_1 is the elimination rate and α_2 the absorption rate.

$$y(t) = k(e^{-\alpha_1(t-t_0)} - e^{-\alpha_2(t-t_0)}) \qquad t \geq t_0 , \qquad \text{Eq. 2.}$$

The initial values fitted to parameters t_0, k, α_1 and α_2 were 0-45 seconds, -500-500, 0-0.15, and 0.15 -0.5, respectively, based on known Aβ kinetics (Shiiki et al., J Neurosci 24:9632-9637, 2004). Final values for t_0, k, α_1 and α_2 were automatically determined using AFNI based on maximal significance of model fitting. Activated rCBV voxels were then determined at $p < 0.05$ after Bonferroni correction.

Results shown in Figure 8 indicate that $A\beta_{1-40}$ decreased CBV in a dose-dependent and region-specific manner (with 0.01 mg Fig. 8a and with 0.1 mg Fig.8b). The extent of the decreased CBV was significantly greater when mice were treated with the high dose (Fig. 8b) compared with the lower dose (Fig. 8a) of $A\beta_{1-40}$ although similar brain regions (e.g. frontal cortex, caudate, thalamus, hippocampus) were affected. In contrast to the dose-dependent effects of $A\beta_{1-40}$, the reverse peptide, $A\beta_{40-1}$, did not significantly affect CBV when tested at the same doses (with 0.01 mg Fig. 8c and with 0.1 mg Fig. 8d) and the extent of any decrease in CBV observed did not differ significantly from the PBS-treated control group (PBS, Fig. 8e). During a 12-minute time course, the amplitude of the decreases in CBV induced by $A\beta_{1-40}$, which ranged from 10-20 % for affected voxels across several brain regions, consistently reached a maximum within 5 min following administration, remained depressed for the duration of the study (representative data shown for the hippocampus in Fig.8 f) and was similar for both doses of $A\beta_{1-40}$. These data are consistent with an additional study using the invasive laser Doppler flowmetry technique (not shown).

To test the effects of 11E6 on CBV using fMRI, the effects of pre-administration of a control IgG1 antibody was evaluated first. As anticipated, pre-administration of an IgG1 antibody 3 hours prior to commencement of imaging, did not block the effect of a challenge with $A\beta_{1-40}$ (with 0.01 mg Fig. 9a). In contrast, 11E6 at a dose of 0.1 mg/mouse given 3 hours prior to commencement of imaging, completely blocked the decrease in CBV normally elicited by challenge with $A\beta_{1-40}$ (with 0.1 mg Fig. 9b). Similarly, decreases in CBV amplitude were also abolished by pretreatment

with the anti-RAGE antibody, 11E6, but not by the control antibody (Fig. 9c). These data demonstrate *in vivo* functional activity of the antibody, 11E6, in an animal model relevant for Alzheimer's disease.

EXAMPLE 11: Construction of CDR-grafted antibodies

By applying standard methods well known in the art, the CDR sequences of VH and VL chains of monoclonal antibody 11E6 (see Table 4 above) are grafted into different different human heavy and light chain acceptor sequences. Based on sequence VH and VL alignments with the VH and VL sequences of monoclonal antibody 11E6 of the present invention the following known human sequences are selected:

a) VH7-4.1 and VH1-2 as well as the joining sequences hJH6 for constructing heavy chain acceptor sequences (according to Table 2 above);
b) 1-12/L5 and 3-15/L2 as well as hJK2 for constructing light chain acceptor sequences (according to Table 3 above).

By grafting the corresponding VH and VL CDRs of 11E6 into said acceptor sequences the following CDR grafted, humanized, modified VH and VL sequences were prepared (see also Table 5, above): VH 11E6.1-GL, VH 11E6.2-GL, VL 11E6.1-GL and VL 11E6.2-GL.

EXAMPLE 12: Construction of framework back mutations in CDR-grafted antibodies

To generate humanized antibody framework mutations, mutations are introduced into the CDR-grafted antibodies using *de novo* synthesis of variable domains and/or mutagenic primers and PCR, using methods well known in the art. Different combinations of back mutations and other mutations are constructed for each of the CDR-grafts as follows.

For heavy chain VH 11E6.1-GL one or more of the following Vernier and VH/VL interfacing residues are back mutated as follows: V2→I, and/or Y95→F.

For heavy chain VH 11E6.2-GL one or more of the following Vernier and VH/VL interfacing residues are back mutated as follows: V2→I, V68→F, M70→F, R72→L, Y95→F.

For light chain VL 11E6.1-GL one or more of the following Vernier and VH/VL interfacing residues are back mutated as follows: A43→S, Y49→F, Y87→F.

For light chain VL 11E6.2-GL one or more of the following Vernier and VH/VL interfacing residues are back mutated as follows: A43→S, Y49→F, I58→V, Y87→F.

Additional mutations include the following:

for heavy chain

VH 11E6.1-GL, Q1→E, and for

VH 11E6.2-GL, Q1→E, I76→T, R85→S, D89→E;

for light chain

VL 11E6.1-GL, V11→L , and

VL 11E6.2-GL, V13→L, E70→D.

EXAMPLE 13: Construction and expression of recombinant humanized anti RAGE antibodies

pHybE expression vectors harboring heavy and light chains containing framework back mutations were co-transfected into 293-6E cells to transiently produce full-length humanized antibodies. Mutations were introduced into the CDR-grafted antibody sequences as prepared according to Example 11, by *de novo* synthesis of the variable domain and/or using mutagenic primers and PCR, and methods well known in the art. The amino acid sequences of the VH and VL regions of the humanized antibodies are disclosed in Table 8.

Table 8: Expression of humanized antibodies

SEQ ID No.	Protein region	Sequence
		123456789012345678901234567890
62	**VH h11E6.1**	EVQLVQSGSELKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY SEEFKG**RFVFSLDTSVSTAYLQICSLKAED TAVYYCAR**SRMVTAYGMDY**WGQGTTVTVSS
63	**VL h11E6.1**	DIQMTQSPSSLSASVGDRVTITC**KASQNVG TAVA**WYQQKPGKAPKLLIY**SASNRYT**GVPS RFSGSGSGTDFTLTISSLQPEDFATYYC**QQ YSSYPLT**FGQGTKLEIKR
62	**VH h11E6.2**	EVQLVQSGSELKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY SEEFKG**RFVFSLDTSVSTAYLQICSLKAED TAVYYCAR**SRMVTAYGMDY**WGQGTTVTVSS

64	VL h11E6.2	DIQMTQSPSSLSASVGDRVTITC**KASQNVG** **TAVA**WYQQKPGKSPKLLIF**SASNRYT**GVPS RFSGSGSGTDFTLTISSLQPEDFATYFC**QQ** **YSSYPLT**FGQGTKLEIKR
62	VH h11E6.3	EVQLVQSGSELKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RFVFSLDTSVSTAYLQICSLKAED TAVYYCAR**SRMVTAYGMDY**WGQGTTVTVSS
65	VL h11E6.3	EIVMTQSPATLSLSPGERATLSC**KASQNVG** **TAVA**WYQQKPGQAPRLLIY**SASNRYT**GIPA RFSGSGSGTDFTLTISSLQSEDFAVYYC**QQ** **YSSYPLT**FGQGTKLEIKR
62	VH h11E6.4	EVQLVQSGSELKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RFVFSLDTSVSTAYLQICSLKAED TAVYYCAR**SRMVTAYGMDY**WGQGTTVTVSS
66	VL h11E6.4	EIVMTQSPATLSLSPGERATLSC**KASQNVG** **TAVA**WYQQKPGQSPRLLIF**SASNRYT**GVPA RFSGSGSGTDFTLTISSLQSEDFAVYFC**QQ** **YSSYPLT**FGQGTKLEIKR
67	VH h11E6.5	EIQLVQSGSELKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RFVFSLDTSVSTAYLQICSLKAED TAVYFCAR**SRMVTAYGMDY**WGQGTTVTVSS
63	VL h11E6.5	DIQMTQSPSSLSASVGDRVTITC**KASQNVG** **TAVA**WYQQKPGKAPKLLIY**SASNRYT**GVPS RFSGSGSGTDFTLTISSLQPEDFATYYC**QQ** **YSSYPLT**FGQGTKLEIKR
67	VH h11E6.6	EIQLVQSGSELKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RFVFSLDTSVSTAYLQICSLKAED TAVYFCAR**SRMVTAYGMDY**WGQGTTVTVSS
64	VL h11E6.6	DIQMTQSPSSLSASVGDRVTITC**KASQNVG** **TAVA**WYQQKPGKSPKLLIF**SASNRYT**GVPS RFSGSGSGTDFTLTISSLQPEDFATYFC**QQ** **YSSYPLT**FGQGTKLEIKR
67	VH h11E6.7	EIQLVQSGSELKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RFVFSLDTSVSTAYLQICSLKAED TAVY**F**CAR**SRMVTAYGMDY**WGQGTTVTVSS
65	VL h11E6.7	EIVMTQSPATLSLSPGERATLSC**KASQNVG** **TAVA**WYQQKPGQAPRLLIY**SASNRYT**GIPA RFSGSGSGTDFTLTISSLQSEDFAVYYC**QQ** **YSSYPLT**FGQGTKLEIKR
67	VH h11E6.8	EIQLVQSGSELKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RFVFSLDTSVSTAYLQICSLKAED TAVYFCAR**SRMVTAYGMDY**WGQGTTVTVSS
66	VL h11E6.8	EIVMTQSPATLSLSPGERATLSC**KASQNVG** **TAVA**WYQQKPGQSPRLLIF**SASNRYT**GVPA RFSGSGSGTDFTLTISSLQSEDFAVYFC**QQ** **YSSYPLT**FGQGTKLEIKR
68	VH h11E6.9	EVQLVQSGAEVKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RVTMTRDTSTSTAYMELSSLRSED TAVYYCAR**SRMVTAYGMDY**WGQGTSVTVSS

63	VL h11E6.9	DIQMTQSPSSLSASVGDRVTITC**KASQNVG** **TAVA**WYQQKPGKAPKLLIY**SASNRYT**GVPS RFSGSGSGTDFTLTISSLQPEDFATYYC**QQ** **YSSYPLT**FGQGTKLEIKR
68	VH h11E6.10	EVQLVQSGAEVKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RVTMTRDTSTSTAYMELSSLRSED TAVYYCAR**SRMVTAYGMDY**WGQGTSVTVSS
64	VL h11E6.10	DIQMTQSPSSLSASVGDRVTITC**KASQNVG** **TAVA**WYQQKPGKSPKLLIF**SASNRYT**GVPS RFSGSGSGTDFTLTISSLQPEDFATYFC**QQ** **YSSYPLT**FGQGTKLEIKR
68	VH h11E6.11	EVQLVQSGAEVKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RVTMTRDTSTSTAYMELSSLRSED TAVYYCAR**SRMVTAYGMDY**WGQGTSVTVSS
65	VL h11E6.11	EIVMTQSPATLSLSPGERATLSC**KASQNVG** **TAVA**WYQQKPGQAPRLLIY**SASNRYT**GIPA RFSGSGSGTDFTLTISSLQSEDFAVYYC**QQ** **YSSYPLT**FGQGTKLEIKR
68	VH h11E6.12	EVQLVQSGAEVKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RVTMTRDTSTSTAYMELSSLRSED TAVYYCAR**SRMVTAYGMDY**WGQGTSVTVSS
66	VL h11E6.12	EIVMTQSPATLSLSPGERATLSC**KASQNVG** **TAVA**WYQQKPGQSPRLLIF**SASNRYT**GVPA RFSGSGSGTDFTLTISSLQSEDFAVYFC**QQ** **YSSYPLT**FGQGTKLEIKR
69	VH h11E6.13	EIQLVQSGAEVKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RFTFTLDTSTSTAYMELSSLRSED TAVYFCAR**SRMVTAYGMDY**WGQGTSVTVSS
63	VL h11E6.13	DIQMTQSPSSLSASVGDRVTITC**KASQNVG** **TAVA**WYQQKPGKAPKLLIY**SASNRYT**GVPS RFSGSGSGTDFTLTISSLQPEDFATYYC**QQ** **YSSYPLT**FGQGTKLEIKR
69	VH h11E6.14	EIQLVQSGAEVKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RFTFTLDTSTSTAYMELSSLRSED TAVYFCAR**SRMVTAYGMDY**WGQGTSVTVSS
64	VL h11E6.14	DIQMTQSPSSLSASVGDRVTITC**KASQNVG** **TAVA**WYQQKPGKSPKLLIF**SASNRYT**GVPS RFSGSGSGTDFTLTISSLQPEDFATYFC**QQ** **YSSYPLT**FGQGTKLEIKR
69	VH h11E6.15	EIQLVQSGAEVKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RFTFTLDTSTSTAYMELSSLRSED TAVYFCAR**SRMVTAYGMDY**WGQGTSVTVSS
65	VL h11E6.15	EIVMTQSPATLSLSPGERATLSC**KASQNVG** **TAVA**WYQQKPGQAPRLLIY**SASNRYT**GIPA RFSGSGSGTDFTLTISSLQSEDFAVYYC**QQ** **YSSYPLT**FGQGTKLEIKR

69	VH h11E6.16	EIQLVQSGAEVKKPGASVKVSCKASGYTFT **NFGMN**WVRQAPGQGLEWMG**YINTNTGESIY** **SEEFKG**RFTFTLDTSTSTAYMELSSLRSED TAVYFCAR**SRMVTAYGMDY**WGQGTSVTVSS
66	VL h11E6.16	EIVMTQSPATLSLSPGERATLSC**KASQNVG** **TAVA**WYQQKPGQSPRLLIF**SASNRYT**GVPA RFSGSGSGTDFTLTISSLQSEDFAVYFC**QQ** **YSSYPLT**FGQGTKLEIKR

Specifically, for the heavy chains:

VH h11E6.1, VH h11E6.2, VH h11E6.3, and VH h11E6.4 contain VH 11E6.1-GL, with a Q1→E mutation.

VH h11E6.5, VH h11E6.6, VH h11E6.7 and VH h11E6.8 contain VH 11E6.1-GL, with a Q1→E mutation and the following Vernier and VH/VL interfacing residue back mutations: V2→I and Y95→F.

VH h11E6.9, VH h11E6.10, VH h11E6.11, and VH h11E6.12 contain VH 11E6.2-GL, with a Q1→E, I76→T, R85→S, and D89→E mutation.

VH h11E6.13, VH h11E6.14, VH h11E6.15, and VH h11E6.16 contain VH 11E6.2-GL, with a Q1→E, I76→T, R85→S, D89→E mutation and the following Vernier and VH/VL interfacing residue back mutations: V2→I, V68→F, M70→F, R72→L, and Y95→F.

For the light chains:

VL h11E6.1, VL h11E6.5, VL h11E6.9, and VL h11E6.13 contain VL 11E6.1-GL with a V11→L mutation.

VL h11E6.2, VL h11E6.6, VL h11E6.10, and VL h11E6.14 contain VL 11E6.1-GL with a V11→L mutation and the following Vernier and VH/VL interfacing residue back mutations: A43→S, Y49→F, and Y87→F.

VL h11E6.3, VL h11E6.7, VL h11E6.11, and VL h11E6.15 contain VL 11E6.2-GL with V13→L and E70→D mutations.

VL h11E6.4, VL h11E6.8, VL h11E6.12, and VL h11E6.16 contain VL 11E6.2-GL with V13→L and E70→D mutations and the following Vernier and VH/VL interfacing residue back mutations: A43→S, Y49→F, I58→V, and V87→F.

Example 14: Characterization of humanized 11E6 antibodies using competition ELISA

ELISA plates (Costar 3369) were coated overnight at 4°C with 50 µl/well of 2µg/ml hRAGE (1-331) in 0.2 M sodium carbonate-bicarbonate buffer, pH 9.4,

washed with Wash Buffer (PBS containing 0.1% Tween 20), and blocked for 1 hr at room temperature with 200 µl/well of 2% nonfat dry milk in PBS. After washing with Wash Buffer, a mixture of a biotinylated m11E6 (0.3 µg/ml final concentration) and unlabelled competitor test antibody starting at 81 µg/ml final concentration and serially diluted 3-fold) in 50 µl/well of ELISA buffer was added in duplicate. After incubating the plates for 1 hr at room temperature, and washing with Wash Buffer, bound antibodies were detected using 100 µl/well of 1:10,000 dilution of HRP-conjugated streptavidin (Fitzgerald) in ELISA buffer. After incubating for 1 hr at room temperature, and washing with Wash Buffer, color development was performed by adding 100 µl/well of TMB Buffer (Zymed). After incubating for 15 min at room temperature, color development was stopped by adding 50 µl/well of 1N hydrochloric acid. Absorbance was read at 490 nm. Table 9 shows the IC_{50} values of humanized 11E6 antibodies obtained using the computer software GraphPad Prism (GraphPad Software Inc., San Diego, CA).

Table 9: IC50 values of humanized 11E6 antibodies in a competitive ELISA

Antibody	IC50 (nM)	Antibody	IC50 (nM)
h11E6.1	19.7	h11E6.9	17.9
h11E6.2	N/A	h11E6.10	N/A
h11E6.3	18.8	h11E6.11	19.9
h11E6.4	14.1	h11E6.12	11.3
h11E6.5	16.1	h11E6.13	16.8
h11E6.6	N/A	h11E6.14	N/A
h11E6.7	15.5	h11E6.15	14.0
h11E6.8	10.8	h11E6.16	9.2

Example 15: Determination of binding constants for antiRAGE mAb interaction with RAGE

Biacore 2000 and Biacore T100 instruments were used to measure anti-RAGE mAb binding kinetics. The assay format for mAb affinity analysis was Fc-based capture via immobilized anti-Fc antibodies. A standard amine coupling protocol was

employed to immobilize Fc-specific IgG via primary amines to the carboxy-methyl (CM) dextran surface of CM5 sensorchips (Biacore). For the study of mouse anti-RAGE mAbs, anti-mouse Fc (Biacore, anti-mouse, BR-1005-14) was used as the immobilized capture reagent and for the study of humanized anti-RAGE mAbs, anti-human Fc (Pierce 31125) was used as the immobilized capture reagent. An automated protocol, available on the Biacore 2000 and Biacore T100, was used to immobilize 8000-10000 RU of capture reagent in all 4 flowcells of the sensorchip. Briefly, the CM–dextran surfaces were activated by freshly prepared 1:1 50mM N-hydroxysuccinimide (NHS):200mM 3-(N,N-dimethylamino) propyl-N-ethylcarbodiimide (EDC). Then the anti-Fc IgG capture reagent (20ug/ml in 10mM sodium acetate, pH4.5) was applied to the surface followed by deactivation of the surface and blocking of the residual reactive sites with 1M ethanolamine (pH 8.5).

The running buffer employed was PBS-P [1X PBS (Sigma P3813), pH 7.4, 0.005% P20 surfactant (Biacore)] for the humanized antibodies. Each experimental cycle consisted of the following steps: 1) anti-RAGE mAbs were captured in flowcells 2, 3 or 4 to a level of 50-200 RU (depending on the antigen). All measurements were referenced against flowcell 1 which had no captured anti-RAGE mAb. 2) Antigen was injected through all 4 flowcells, 240ul at 80ul/min. After the antigen injection, dissociation was monitored for 600 seconds at 80ul/min. 3) the anti-Fc capture surface was regenerated with low pH glycine. For kinetic determinations, antigen injections were a 3 fold dilution series from either 30nM – 0.12nM (for sRAGE [RAGE (1-331)] and buffer only in randomized duplicates.

Data were processed using either Biacore evaluation software or Scrubber 2.0 software (BioLogic Software). Briefly, the data were double referenced by first, subtracting the signal from the reference cell and second, by subtracting the signal from buffer-only injections. The double referenced data for the RAGE injection series were then fit globally to a 1:1 (Langmuir) binding model, which included a mass transfer term, to determine the binding kinetic rate constants, k_a and k_d, and the affinity, K_D ($k_a = k_{on}$; $k_d = k_{off}$)

Table 10: Biacore data

11E6 mAb	k_{on} (M^{-1} s^{-1})	k_{off} (s^{-1})	K_D (pM)	Resid Stdev
h11E6.8	2.5E+07	8.4E-04	33	1.6

h11E6.12	2.0E+07	1.7E-03	83	1.8
h11E6.16	2.8E+07	1.4E-03	50	1.9
mouse 11E6	1.8E+07	1.3E-03	68	1.4

The K_D values are double-digit pM for all three mAbs and there does not seem to be a significant distinction between the three mAbs regarding their binding kinetics.

In a concurrent experiment, the original mouse 11E6 mAb was evaluated with this antigen (human sRAGE 1-331, V#400) and was also double-digit pM K_D.

The original mouse 11E6 mAb was previously reported to be 290pM. However those early experiments used a different buffer system (Biacore buffer HBS-EP+: 10mM HEPES, pH 7.4, 150mM NaCl, 3mM EDTA, 0.05%P20). The buffer choice can, of course, affect the kinetics.

Example 16: *In vivo* cerebrovascular blood volume (CBV) studies in aged Tg2576 mice

16.1. Animals

The Tg2576 mouse model of Alzheimer's disease (Hsiao et al.,1996) expresses the Swedish mutation of APP (APPK670N,M671L) at high level under control of the hamster prion protein (PrP) promoter. It is well established that this mutation causes concomitant increases in secreted Ab42 and Ab40. As Tg2576 mice age, plaques appear that are similar to those seen in Alzheimer's disease. In addition, Tg2576 mice develop age dependent behavioural deficits as assessed by Y maze, T maze, and Morris water maze testing (Hsiao et al. (1996) Correlative memory deficits, Ab elevation, and amyloid plaques in transgenic mice. Science 274:99 –102.)

16.2 Antibody preparation

A negative IgG control antibody and the anti-RAGE antibody, 11E6 (both >99% pure), were synthesized at Abbott Laboratories.

16.3. CBV measurement using fMRI

fMRI-CBV experiments were performed on a 7.0 T/21 cm horizontal magnet with a 20 G/cm magnetic field gradient insert (Biospec Bruker, Billerica, MA). Aged Tg2576 mice (19-20 months old) were first anesthetized with meditomidine (1 mg/kg, i.p.; Pfizer Animal Health, Exton, Pennsylvania, USA)+ketamine (75 mg/kg, i.p.; Fort Dodge Animal Health, Iowa, USA) and then placed in a dual-coil small animal

restrainer (Insight Neuroimaging Systems, LLC, Worcester, MA), which contains a volume coil for transmitting and a surface coil for receiving. Respiration rates and waveforms were continuously monitored via a force transducer. Rectal temperature was monitored and maintained at 37 ± 1 °C via a feedback-regulated, circulating water pad. All imaging was performed during the light phase. Coil-to-coil electromagnetic interaction was actively decoupled. Anatomical images were acquired using the fast spin-echo rapid acquisition relaxation enhanced (RARE) pulse sequence with TR = 3 s, effective TE = 100 ms, matrix = 256 x 256, FOV = 2.56 cm x 2.56 cm, nine 1.0-mm slices, and four averages. Gradient echo single-shot echo-planar imaging (EPI) was used for fMRI-CBV image acquisition with TR = 2 s, TE = 13 ms, matrix = 64 x 64, FOV = 2.56 cm x 2.56 cm, and giving an in-plane resolution = 400 μm x 400 μm. 10 mg Fe/kg ultrasmall superparamagnetic iron oxide (USPIO) contrast agent (SH U555C, Schering AG, Berlin, Germany) was administered intravenously 2 min into an 18 min image acquisition. Mouse 11E6 or nonspecific mouse IgG1 (control antibody) were administered via the tail vein 6 min after the contrast agent using a syringe pump (0.1 mL/min for 1 min) and changes in CBV were then detected over a subsequent 10 min period.

16.4 fMRI Data analysis

Data analysis was performed using the Analysis of Functional NeuroImages (AFNI) software package (Cox, 1996). To identify time-dependent relative CBV change, rCBV(t), was calculated from time course raw data based on the relationship (Mandeville et al., 1999),

$$rCBV(t) = \ln[s(t)/s_0(t)]/\ln[s_0(t)/s_{pre}] \qquad \text{Eq.1}$$

where s(t) is the signal intensity after Aβ or PBS infusion, $S_0(t)$ is the baseline signal before the Aβ or PBS infusion, and S_{pre} is the mean sigal intensity before the administration of contrast agent. The time-course rCBV changes were detrended with a linear function to account for elimination of contrast agent from the blood (Cox, 1996).

Subsequently, the rCBV signal for each voxel in every mouse was fitted to a nonlinear differential exponential model (Eq.2) reflecting the drug's kinetics (Luo et al., 2004) where t_0 is the time delay of response, k is the multiplicative coefficient, α_1 is the elimination rate and α_2 the absorption rate.

$$y(t) = k(e^{-\alpha_1(t-t_0)} - e^{-\alpha_2(t-t_0)}) \qquad t \geq t_0, \qquad \text{Eq. 2.}$$

Final values for t_0, k, α_1 and α_2 were automatically determined using AFNI based on maximal significance of model fitting. Activated rCBV voxels were then determined at $p < 0.05$ after Bonferroni correction. Whole brain activated voxels with CBV increase were recorded. Since the untransformed data do not confirm with the ANOVA assumptions, Box-Cox transformation was employed to ensure adequate normality and variance homogeneity. The 11E6 effect is statistically significant versus IgG1 ($p<0.05$) in 19 month old TG2576 mice in the fMRI-CBV model.

The results are shown in the attached Figure 10. Our data extend results by Deane (Deane et al, 2003) showing that polyclonal anti-RAGE antibodies targeting a variety of epitopes within RAGE can enhance cerebral blood perfusion in APP-transgenic mice. Our data show for the first time that a monoclonal antibody 11E6 targeting the C2-domain within RAGE increases cerebral blood perfusion in APP transgenic mice. This effect is probably mediated by competing binding of high level of Aβ to RAGE. High level of Aβ exist in the brain and plasma of due to overexpression of human APP. Treatment of AD patients with 11E6 may thus increase cerebral perfusion in these patients potentially leading to improvement of neuronal functions. At the same time treatment of patients could be potentially monitored by following cerebral blood flow during treatment.

Example 17: Protection of hippocampal neurons against Aß induced dynamin cleavage by antibody 11E6

17.1. Culture of hippocampal neurons

Rat hippocampal neurons were prepared according to literature (Goslin and Banker. (1991) Rat Hippocampal Neurons in Low-Density Culture. In: Banker G and Goslin K (ed). *Culturing Nerve Cells*, MIT Press, Cambridge) with slight modification. Briefly, hippocampi of 19d old embryonic rats were dissected and freed from meninges. Hippocampal neurons were obtained by trypsination of tissue (0,1% trypsin / 17-20min / 37 °C) followed by trituration with fire-polished Pasteur pipettes. Hippocampal neurons were plated at a density of 0.2-1.0 x 10^5 cells into poly-D-lysine coated 6well or 24well plates (Biocoat™ plate; BD Biosciences, Heidelberg, Germany) using 0,5-3ml of serum-free culture medium (Neurobasal medium, B27 supplement , 2mM L-Glutamine;1% Penicillin-Streptomycin; Invitrogen, Karlsruhe,

Germany). Cells were cultivated at 37 °C, 5% CO_2 for at least 21d and one third of the medium was exchanged once a week.

17.2. Aß induced dynamin cleavage

Aß was aggregated according to the literature (Kelly, B. L., and Ferreira, A. (2006) *J Biol Chem* **281**(38), 28079-28089; Kelly, B. L., Vassar, R., and Ferreira, A. (2005) *J Biol Chem* **280**(36), 31746-31753) with slight modifications. Briefly, Aß1-40 (American Peptide, Sunnyvale, Ca) was dissolved in serum free culture medium at 0,1mg/ml and incubated for 4d at 37 °C. Anti-RAGE monoclonal antibody 11E6, an IgG1 isotype monoclonal control antibody directed against KLH (Keyhole Limpet Hemocyanin from Megathura crenulata, Abbott) or PBS were incubated with the aggregated Aß for 1h at 25°C under constant agitation in a final volume of 225µl-1ml. The mixtures were added to the culture medium resulting in a final concentration of 5µM Aß (calculated as monomer) and 2µM antibody, respectively. Every treatment was performed in triplicate and wells without addition of Aß or antibodies were included as further controls. Cells were cultivated for another 24h and briefly inspected by light microscopy before being processed for Western blot. Treatment with Aß did not induce overt neuronal death during the incubation period.

17.3. Western blot, quantification of dynamin cleavage, and statistical analysis

Culture medium was removed and cells were washed with PBS once. Cells were lyzed by the addition of cold buffer (50mM Tris-HCl pH 7.5 ; 150mM NaCl ; 1% NP-40 ; 1% Triton X-100 ; 2mM EDTA) containing protease and phosphatase inhibitor cocktails (Roche, Mannheim, Germany). Cells were scraped and the homogenate was centrifuged at 13000g at 4°C for 5min.. The supernatant was removed and total protein concentration was determined by the Bradford method using a commercial kit (Bio-Rad, München, Germany). Protein was diluted to 1µg/µl into loading buffer (Bio-Rad, München, Germany) and boiled for 5min. 25µg of each sample was run on a 4-20% SDS-PAGE (Invitrogen, Karlsruhe, Germany) and transferred onto nitrocellulose membranes using the iBlot system (Invitrogen, Karlsruhe, Germany). Alternatively, cells were directly lyzed on 96 well plates,diluted with loading buffer,and and 1/4 to 1/5 of the protein was loaded on SDS-PAGE. Membranes were blocked at room temperature for 1-2 h using 1x Blocking Reagent (Roche, Mannheim, Germany) and were then incubated with primary antibody against dynamin I (PA-1-660; 1:1000 dilution; Affinity BioReagents, Golden, Co) at 4°C overnight. Subsequently, a horseradish peroxidase-conjugated secondary antibody

(goat anti-rabbit IgG, Jackson ImmunoResearch, West Grove, PA) was applied to blots at room temperature for 1 h and detected using enhanced chemiluminescence (SuperSignal West Pico Chemiluminescent Substrate; Pierce, Rockford, Il). Immunoblot signals were visualized by a VersaDoc system (Bio-Rad, München, Germany) and the signal of intact dynamin I (~100 kDa) was quantified using Quantity One software (Bio-Rad, München, Germany) . For normalization, the blots were stripped (Restore Western Blot Stripping Buffer; Pierce, Rockford, Il) for 30 min at 37°C, washed in PBS, and re-probed with a primary antibody directed against ßIII tubulin (TuJ-I, 1:1000 dilution, Abcam, Cambridge, MA), and secondary antibody (donkey anti-mouse IgG, Jackson ImmunoResearch, West Grove, PA). The mean of the normalized dynamin I expression of cells not treated with Aß was set to 100%. Percent data of three separate experiments were analyzed for statistical significance by a One-Way ANOVA (Kruskal-Wallis test) followed by Dunn's test (GraphPad Prism™; GraphPad Software, San Diego, Ca).

17.4. Anti-RAGE antibody 11E6 protects hippocampal neurons against Aß induced dynamin cleavage

Aggregated Aß1-40 had recently been shown to induce cleavage of the synaptic marker protein dynamin I in hippocampal neurons (Kelly et al, 2005; Kelly & Ferreira, 2006). In full accordance with the published results, we observed a marked decrease in the amount of intact (~100kDa) dynamin I after incubation of hippocampal neurons with aggregated Aß for 24h and a concomitant increase of a ~90 kDa cleavage product (Fig. 11, upper panel). Pre-incubation of the Aß with the anti-RAGE antibody 11E6 prevented the cleavage to about 70%. In contrast, a RAGE-unrelated murine IgG1 isotype control antibody did not provide any protection (Fig. 11, upper panel). Densitometric scanning of the triplicate samples, normalization to the amount a control protein (ßIII tubulin), and data analysis of three independent experiments revealed the statistical significance of the observed protective effect (Fig. 11, lower panel; One-Way ANOVA; $p<0,05$).

Example 18: Effect of antibody 11E6 on globulomer-induced suppression of synaptic transmission

18.1. Test A

Organotypic hippocampal slice cultures were prepared in a modified protocol of Stoppini et al (Journal of Neuroscience Methods, 37, Issue 2, April 1991, Pages 173-182 "A simple method for organotypic cultures of nervous tissue" L. Stoppinia,

P.-A. Buchsa and D. Muller) and cultured on millicell-CM membranes (Millipore, Billerica, USA) in high potassium medium for 3 days and later in supplemented neurobasal A medium in the liquid/gas interface at 34°C / 5%CO2.

Rat hippocampal slice cultures were prepared from 9 day old Wistar rats and used at 15-16 days in vitro. Slice cultures were incubated over night with either

- 1 μM 1-42 globulomer,
- 0.1 μM 11E6 (RAGE mAb ML 39-11E6 purification #4194, sample #6116) + 1 μM 1-42 globulomer
- control (globulomer ultrafiltrate + SDS)

In the co-incubation group the antibody was applied to slice culture medium 2 hours before globulomer. Recordings were performed in an interface recording chamber under continuous perfusion with artificial cerebrospinal fluid. Field excitatiory postsynaptic potentials were recorded from the CA1 region after stimulation of the Schaffer collateral with biphasic pulses at different voltage intensities. The Schaffer collateral was stimulated with biphasic pulses (0.1 ms/phase) using a 0.5M bipolar Tungsten electrode (WPI; Saraosta USA), and fEPSP amplitudes were recorded with glass electrodes filled with aCSF (0.7-1.1 Megaohm, GC150F-15, Harvard Apparatus, Hugstetten, Germany). Signals were digitized using a power CED 1401 (Cambridge Electronic Design Ltd., Cambridge, UK) and analyzed using Signal 2.14 (Cambridge Electronic Design Ltd., Cambridge, UK).

The results are shown in Figure 12A. Globulomer-application strongly suppressed synaptic transmission. Co-application of 0.1 μM 11E6 completely reversed the globulomer-induced deficits. Thus, 11E6 can reverse globulomer-induced deficits in synaptic transmission.

18.2. Test B

Rat hippocampal slice cultures were prepared from 9 day old Wistar rats and used at 16-18 days in vitro. Slice cultures were incubated over night with either

- 1 μM 1-42 globulomer
- 0.1 μM IgG1 mAb H35C206 (KLH) + 1 μM 1-42 globulomer
- control (SDS)

Recordings were performed (in artificial cerebrospinal fluid) from CA1 after stimulation of the Schaffer collateral at different intensities.

The results are shown in Figure 12B. Globulomer-application strongly suppressed synaptic transmission. Co-application of IgG1 did not reverse the globulomer-induced deficits. Thus, the IgG control antibody does not reverse globulomer-induced deficits in synaptic transmission at 0.1 µM.

Example 19: In situ analysis of the effect of antibody 11E6 on amyloid plaques in the frontal cortex of Tg2576 mice.

For these experiments 14.5 month old Tg2576 mice (Hsiao et al., 1996, Science; 274(5284), 99-102) was used. The mice overexpress human APP with the so-called Swedish mutation (K670N/M671L) and formed β amyloid deposits in the brain parenchyma at about 11 months of age. Starting at 12 months of age the mice were injected with 500 µg 11E6 (n=19) i.p. (intraperitoneal) or an IgG1 control antibody (n=19) once weekly for 12 weeks. After the last injection, the animals were deeply anaesthetized and transcardially perfused with 0.1 M phosphate-buffered saline (PBS) to flush the blood. Then the brain was removed from the cranium and divided longitudinally. One hemisphere of the brain was shock-frozen, the other fixated by immersion into 4% paraformaldehyde. The immersion-fixated hemisphere was cryoprotected by soaking in 30% sucrose in PBS and mounted on a freezing microtome. The entire forebrain was cut into 40 µm section which were collected in PBS and mounted on Superfrost® Plus glass slides (Menzel Glaeser, Braunschweig, Germany), for the subsequent staining procedure. Staining of Aβ containing amyloid plaques was performed with the mouse monoclonal antibody 6G1 raised against monomeric Aβ (Barghorn et al., 2005, J. Neurochem. On an automatic staining device (Ventana Discovery®, Roche Diagnostics GmbH, Mannheim, Germany) in accordance with the following protocol:

- the sections on the glass slides were thoroughly air-dried and transferred to the Ventana machine
- an automatic protocol provided by Ventana for the chromogenic diamino benzidine (DAB) procedure was used that contained washing and blocking steps and staining with the DAB MapTM Kit was used; antigen retrieval and primary and secondary immunohistochemistry were included in the automatized protocol by the experimentator:
- antigen retrieval was obtained in the presence of conditioner #2 (citrat-based buffer, pH 6.0) at 95 °C for 45 minutes

- incubation with 6G1 (1:500) in antibody diluent (Roche Diagnostics GmbH, Mannheim, Germany) at 37°C for 3 hours
- incubation with biotinylated secondary antibody donkey anti-mouse IgG (1:500; Jackson ImmunoResearch, Newmarket, UK) at 37°C for 30 minutes
- after finalization of automated staining, slides were washed in normal water, dehydrated in graded ethanols, cleared in XTRA-Solve® (J.T. Baker, Griesheim, Germany), and coverslipped with UltraKitt® (J.T. Baker, Griesheim, Germany)

Plaque staining was quantified in 3 histological sections of the neocortex using the ImagePro 5.0 image analysis system. The experimentator was blinded to the treatment of the mouse under analysis and determined the following parameters: area of the neocortex, area covered with 6G1 positive staining and number of stained plaques. These parameters were variable and not normally distributed. Therefore, a reduction of plaque load was statistically evaluated by a on-sided Mann-Whitney U-test. Results of the Aβ deposit staining in Tg2576 mice are shown in Figure 13.

Evaluation of brown DAB deposits showed that the anti-RAGE antibody reduced the number and area of amyloid plaques in the neocortex by 24.5% and 26.8%, respectively ($p<0.1$). The reduction in plaque number and area was most evident in the frontal neocortex ($p<0.05$).

Documents as cited herein are incorporated by reference.

CLAIMS

1. An isolated monoclonal antibody comprising an antigen binding domain, said antibody capable of binding an epitope of a human RAGE molecule, said antigen binding domain comprising at least one CDR comprising an amino acid sequence selected from:

a) the CDR-H3 group of amino acid sequences consisting of SEQ ID NO.: 4, 12 and 20; modified CDR amino acid sequences having a sequence identity of at least 50% to one of said sequences; and/or

b) the CDR-L3 group of amino acid sequences consisting of SEQ ID NO.: 8, 16 and 24; and modified CDR amino acid sequences having a sequence identity of at least 50% to one of said sequences.

2. The antibody according to claim 1, further comprising at least one CDR comprising an amino acid sequence selected from the CDR-H1 group consisting of SEQ ID NO: 2, 10, 18; or selected from the CDR-H2 group consisting of SEQ ID NO: 3, 11, 19; or selected from the CDR-L1 group consisting of SEQ ID NO: 6, 14, 22; or selected from the CDR-L2 group consisting of SEQ ID NO: 7, 15, 23;

and modified CDR amino acid sequences having a sequence identity of at least 50% to one of said sequences.

3. The antibody according to claim 2, comprising at least 3 CDRs which are selected from a variable domain CDR set consisting of:

VH 7F9 set		
VH 7F9 CDR-H1	Residues 31-35 of SEQ ID NO.:1	SEQ ID NO: 2
VH 7F9 CDR-H2	Residues 50-68 of SEQ ID NO.:1	SEQ ID NO: 3
VH 7F9 CDR-H3	Residues 101-108 of SEQ ID NO.:1	SEQ ID NO: 4
VL 7F9 set		
VL 7F9 CDR-L1	Residues 24-34 of SEQ ID NO.:5	SEQ ID NO: 6
VL 7F9 CDR-L2	Residues 50-56 of SEQ ID NO.:5	SEQ ID NO: 7
VL 7F9 CDR-L3	Residues 89-97 of SEQ ID NO.:5	SEQ ID NO: 8
VH 11E6 set		
VH 11E6 CDR-H1	Residues 31-35 of SEQ ID NO.:9	SEQ ID NO: 10
VH 11E6 CDR-H2	Residues 50-66 of SEQ ID NO.:9	SEQ ID NO: 11
VH 11E6 CDR-H3	Residues 99-109 of SEQ ID NO.:9	SEQ ID NO: 12
VL 11E6 set		
VL 11E6 CDR-L1	Residues 24-34 of SEQ ID NO.:13	SEQ ID NO: 14
VL 11E6 CDR-L2	Residues 50-56 of SEQ ID NO.:13	SEQ ID NO: 15

VL 11E6 CDR-L3	Residues 89-97 of SEQ ID NO.:13	SEQ ID NO: 16
VH 4E5 set		
VH 4E5 CDR-H1	Residues 31-35 of SEQ ID NO.:17	SEQ ID NO: 18
VH 4E5 CDR-H2	Residues 50-66 of SEQ ID NO.:17	SEQ ID NO: 19
VH 4E5 CDR-H3	Residues 99-109 of SEQ ID NO.:17	SEQ ID NO: 20
VL 4E5 set		
VL 4E5 CDR-L1	Residues 24-34 of SEQ ID NO.:21	SEQ ID NO: 22
VL 4E5 CDR-L2	Residues 50-56 of SEQ ID NO.:21	SEQ ID NO: 23
VL 4E5 CDR-L3	Residues 89-97 of SEQ ID NO.:21	SEQ ID NO: 24

or a variable domain set wherein at least one of said 3 CDRs is a modified CDR amino acid sequence having a sequence identity of at least 50% to the parent sequence.

4. The antibody according to claim 3, wherein said at least two variable domain CDR sets are selected from a group consisting of:

VH 7F9 set & VL 7F9 set;

VH 4E5 set & VL 4E5 and

VH 11E6 set & VL 11E6 set.

5. The antibody according to one of the preceding claims, further comprising a human acceptor framework.

6. The antibody of any one of the preceding claims comprising at least one heavy chain variable domain selected from SEQ ID NO: 56 and 57; and/or at least one light chain variable domain selected from SEQ ID NO: 58 and 59.

7. The antibody according to claim 6, wherein said binding protein comprises at least one framework mutation selected from the group consisting of:

(heavy chain sequence position): 1, 2, 68, 70, 72, 76, 85, 89, 95
(light chain sequence position): 11, 13, 43, 49, 58, 70, 87.

8. The antibody of any one of the preceding claims, wherein said binding protein comprises at least one (framework mutated) variable domain having an amino acid sequence selected from the group consisting of:
(heavy chain sequences) SEQ ID NO: 62, 67, 68 and 69;

(light chain sequences) SEQ ID NO: 63, 64, 65 and 66.

9. The antibody of claim 1, which is selected from the group consisting of antibodies 11E6, 4E5 and 7F9.

10. An isolated nucleic acid encoding the amino acid sequence of the antibody of any one of claims 1-9.

11. A vector comprising an isolated nucleic acid according to claims 10.

12. A host cell comprising a vector according to claim 11.

13. A method of producing a protein capable of binding RAGE, comprising culturing a host cell according to claim 12 in culture medium under conditions sufficient to produce a binding protein capable of binding RAGE.

14. A protein produced according to the method of 13.

15. A composition for the release of an antibody said composition comprising
(a) a formulation, wherein said formulation comprises the antibody of claim 1 to 9 in crystallized form as an ingredient; and
(b) at least one polymeric carrier.

16. A pharmaceutical composition comprising the antibody of any one of claims 1 to 9, and a pharmaceutically acceptable carrier.

17. A method for treating a mammal comprising the step of administering to the mammal an effective amount of the composition according to claim 15 or 16.

18. The method of claim 17, wherein the method is for treating neurological diseases selected from the group comprising Amytropic Lateral Sclerosis, Brachial Plexus Injury, Brain Injury, including traumatic brain injury, Cerebral Palsy, Friedrich's Ataxia, Guillain Barre, Leukodystrophies, Multiple Sclerosis, Post Polio, Spina

Bifida, Spinal Cord Injury, Spinal Muscle Atrophy, Spinal Tumors, Stroke, Transverse Myelitits, dementia, senile dementia, mild cognitive impairment, Alzheimer-related dementia, Huntington's chorea, tardive dyskinesia, hyperkinesias, manias, Morbus Parkinson, steel-Richard syndrome, Down's syndrome, myasthenia gravis, nerve trauma, vascular amyloidosis, cerebral hemorrhage I with amyloidosis, brain inflammation, Friedrich's ataxia, acute confusion disorder, amyotrophic lateral sclerosis, glaucoma, Alzheimer's disease, diabetic nephropathy, sepsis, rheumatoid arthritis and related inflammatory diseases.

19. The method of claim 17, wherein the method is for treating Alzheimer's disease.

20. The method of claim 19, wherein the composition comprises 11E6.