1. A method for purifying transgenic human Factor VII (TgFVII) and/or transgenic activated human Factor VII (TgFVIIa) produced in the milk of transgenic non-human animal, comprising the steps of :

(a) a step of affinity chromatography comprising the steps of:

(i) contacting the milk containing human TgFVII and/or human TgFVIIa with a ligand which is specific to human TgFVII and/or human TgFVIIa, under conditions allowing the human TgFVII and/or human TgFVIIa to bind the ligand , and

(ii) recovering human TgFVII and/or human TgFVIIa by disrupting in a non destructive way the interaction with said ligand,

wherein said ligand is an antigen-binding protein directed to at least one human TgFVII or human TgFVIIa epitope, or a functional fragment or derivative thereof, and

wherein said ligand is derived from heavy immunoglobulin chains of Camelidae,

(b) chromatographic purification of the eluted human TgFVII and /or human TgFVIIa on an ion exchanger, under conditions wherein human TgFVII and/or human TgFVIIa are retained on the ion exchanger,

(c) chromatographic separation of the eluted human TgFVII and/or human TgFVIIa on pseudo-affinity resin, under conditions wherein human TgFVII and/or human TgFVIIa are retained on the pseudo-affinity resin, and

(d) chromatographic separation of the eluted human TgFVII and/or human TgFVIIa on a size exclusion support.

2. The method according to claim 1, wherein said ligand comprises two heavy polypeptide chains forming two complete antigen binding sites and is devoid of light polypeptide chains.

3. The method according to claims 1 to 2, wherein step (ii) comprises at least one washing step for removing unbound material before eluting human TgFVII or human TgFVIIa from the ligand, and further optionally comprises a second washing step for removing weak bound material.

4. The method according to claim 1, wherein the animal is a mammal female, and preferably a rabbit female.

5. The method according to claim 1, further comprising the preliminary steps, before step (a), of collecting and clarifying milk.

6. The method according to claim 5, wherein the milk clarification step is performed by citrate salt addition, followed by filtration.

7. The method according to claim 1, wherein the ion exchanger of step (b) is an anion exchanger.

8. The method according to claim 1, wherein the pseudo-affinity resin of step (c) is hydroxyapatite.

9. The method according to claims 1 to 8, further comprising at least one step of eliminating and/or inactivating viruses, preferably by nanofiltration and/or by Solvent/Detergent treatment.

10. The method according to claims 1 to 9, further comprising the final steps of formulating, sterilizing and lyophilizing the purified human TgFVII and/or human TgFVIIa.

11. The method according to claims 1 to 10, wherein the step of affinity chromatography comprises the steps of:
(a) loading a chromatographic resin on which a ligand as defined in any of claims 1 to 3 is immobilized, with milk from a transgenic female rabbit containing recombinantly produced human TgFVII and/or human TgFVIIa, under conditions allowing the ligand to bind human TgFVII and/or human TgFVIIa;
(b1) washing the unbound material;
(b2) washing the weak bound material; and
(b3) eluting human TgFVII and/or human TgFVIIa from the resin.

12. The method according to claim 11, wherein, during said affinity chromatography step, said second washing step (b2) for removing weak bound material is performed with a buffer comprising from 10% to 50% of a hydrophobic agent, and the ionic strength of which is ranging from 200 to 600 mM.

13. The method according to claim 11, wherein, during said affinity chromatography step, said elution step (b3) for eluting human TgFVII and/or human TgFVIIa from the resin is

performed with a buffer comprising from 20% to 70% of a hydrophobic agent, and the ionic strength of which is ranging from 500 to 2500 mM.

14. The method according to claims 1 to 13, wherein the resulting purified transgenic FVII is an almost fully activated, high purity human FVII, comprising a low percentage of degraded, proteolyzed or oxidized forms.

15. The method according to claims 1 to 14, characterized in that the affinity chromatography step and/or of ion exchange chromatography step and/or of size exclusion chromatography step allow to obtain a viral reduction factor superior to 3 $\log_{10}$, preferably superior to 4 $\log_{10}$, more preferably superior to 5 $\log_{10}$.