

Novel LHRH antagonists having improved solubility
properties

The invention relates to LHRH antagonists having
5 improved solubility properties, processes for the
preparation of these compounds, medicaments in which
these compounds are contained, and the use of the
medicaments for the treatment of hormone-dependent
tumours and hormone-influenced non-malignant disorders
10 such as benign prostate hyperplasia (BPH) and
endometriosis.

The nomenclature used for the definition of the
peptides agrees with that nomenclature explained by the
15 IUPAC-IUB Commission on Biochemical Nomenclature
(European J. Biochem. 1984, 138, 9-37), in which, in
agreement with the conventional representation, the
amino groups at the N terminus appear to the left and
the carboxyl group at the C terminus appears to the
20 right. The LH-RH antagonists such as the peptides
according to the invention include naturally occurring
and synthetic amino acids, the former including Ala,
Val, Leu, Ile, Ser, Thr, Lys, Arg, Asp, Asn, Glu, Gln,
Cys, Met, Phe, Tyr, Pro, Trp and His. The abbreviations
25 for the individual amino acid residues are based on the
trivial names of the amino acids and are Ala=alanine,
Arg=arginine, Gly=glycine, Leu=leucine, Lys=lysine,
Pal(3)=3-(3-pyridyl)alanine, Nal(2)=3-(2-naphthyl)-
alanine, Phe=phenylalanine, Cpa=4-chlorophenylalanine,
30 Pro=proline, Ser=serine, Thr=threonine, Trp=tryptophan,
Try=tyrosine and Sar=sarcosine. All amino acids
described here originate from the L series, if not
mentioned otherwise. For example, D-Nal(2) is the
abbreviation for 3-(2-naphthyl)-D-alanine and Ser is
35 the abbreviation for L-serine. Substitutions on the ϵ
amino group in the side chain of lysine are represented
by a term placed in brackets behind Lys, if appropriate
in the form of an abbreviation.



Other abbreviations used are:

	Ac	Acetyl
	Atz	3-Amino-1,2,4-triazole-5-carbonyl
	B	4-(4-Amidinophenyl)amino-1,4-dioxobutyl
5	Boc	tert-Butyloxycarbonyl
	Bop	Benzotriazol-1-oxy-tris(dimethylamino)- phosphonium hexafluorophosphate
	DCC	Dicyclohexylcarbodiimide
	DCM	Dichloromethane
10	Ddz	Dimethoxyphenyl-dimethylmethylenoxy-carbonyl (Dimethoxy-dimethyl-Z)
	DIC	Diisopropylcarbodiimide
	DIPEA	N,N-Diisopropylethylamine
	DMF	Dimethylformamide
15	Fmoc	Fluorenylmethyloxycarbonyl
	HF	Hydrofluoric acid
	HOBt	1-Hydroxybenzotriazole
	HPLC	High-pressure liquid chromatography
	Me	Methyl
20	TFA	Trifluoroacetic acid
	Z	Benzyloxycarbonyl

The peptides according to the invention are analogues
of the luteinizing-hormone-releasing hormone (LH-RH),
25 which has the following structure:
p-Glu-His-Trp-Ser-Tyr-Gly-Leu-Arg-Pro-Gly-NH₂, [LH-RH,
gonadorelin].

For more than 20 years, researchers have sought
30 selective potent antagonists of the LH-RH decapeptide
[M. Karten and J.E. Rivier, Endocrine Reviews 7, 44-66
(1986)]. The high interest in such antagonists is based
on their usefulness in the field of endocrinology,
gynaecology, contraception and cancer. A large number
35 of compounds have been prepared as potential LH-RH
antagonists. The most interesting compounds which have
been found to date are those compounds whose structures
are a modification of the LH-RH structure.

The first series of potent antagonists was obtained by the introduction of aromatic amino acid residues into the positions 1, 2, 3 and 6 or 2, 3 and 6. The customary way of writing the compounds is as follows:
5 the amino acids are first indicated which have taken the place of the amino acids originally present in the peptide chain of LH-RH, the positions in which the exchange took place being marked by superscripted figures. Furthermore, by the notation "LH-RH" placed
10 afterwards it is expressed that these are LH-RH analogues in which the exchange has taken place.

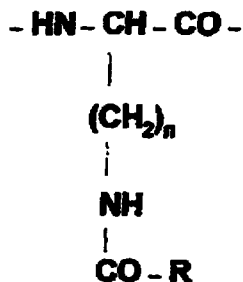
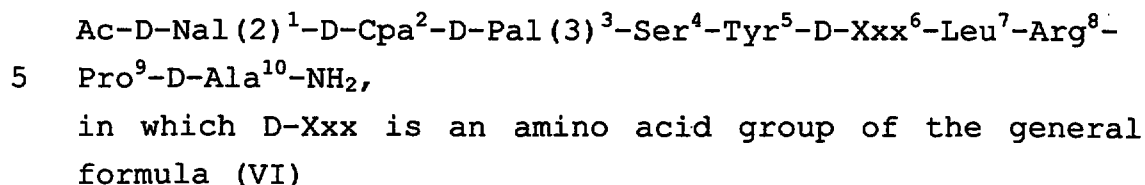
Known antagonists are:

[Ac-D-Cpa^{1,2}, D-Trp^{3,6}] LH-RH (D.H. Coy et al., In:
15 Gross, E. and Meienhofer, J. (Eds) Peptides; Proceedings of the 6th American Peptide Symposium, pp. 775-779, Pierce Chem. Co., Rockville III. (1979):
[Ac-Pro¹, D-Cpa², D-Nal(2)^{3,6}] LH-RH (US Patent No. 4,419,347) and [Ac-Pro¹, D-Cpa², D-Trp^{3,6}] LH-RH
20 (J.L. Pineda, et al., J. Clin. Endocrinol. Metab. 56, 420, 1983).

In order to improve the action of antagonists, basic amino acids, for example D-Arg, were later introduced
25 into the 6 position. For example [Ac-D-Cpa^{1,2}, D-Trp³, D-Arg⁶, D-Ala¹⁰] LH-RH (ORG-30276) (D.H. Coy, et al., Endocrinology 100, 1445, 1982); and
[Ac-D-Nal(2)¹, D-Phe(4-F)², D-Trp³, D-Arg⁶] LH-RH (ORF 18260) (J.E. Rivier et al., in: Vickery B.H. Nestor, Jr. J.J., Hafez, E.S.E (Eds). LHRH and its Analogs,
30 pp. 11-22 MTP Press, Lancaster, UK 1984).

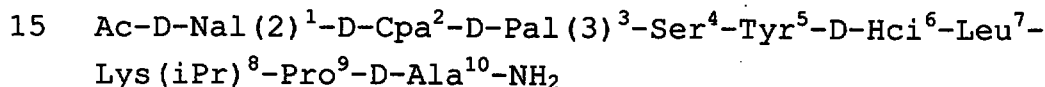
Further potent LH-RH antagonists are described in
WO 92/19651, WO 94/19370, WO 92/17025, WO 94/14841,
35 WO 94/13313, US-A 5,300,492, US-A 5,140,009,
EP 0 413 209 A1 and DE 195 44 212 A1.

The latter discloses compounds having a modified ornithine or lysine unit in position 6 and which correspond to the following formula:

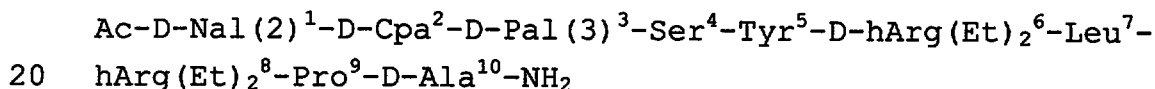


10 Further known LH-RH antagonists are antarelix, ganirelix and cetrorelix.

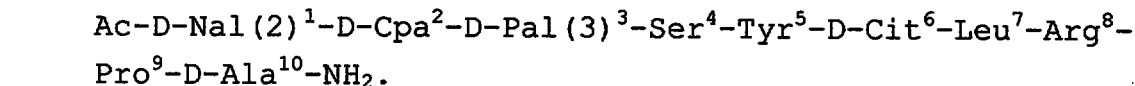
Antarelix:



Ganirelix:



Cetrorelix:

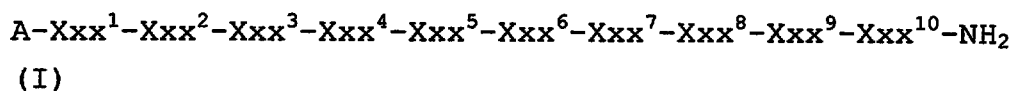


The aim of the invention is to create novel LH-RH antagonists which have an increased enzymatic stability and significantly improved water solubility.

30 This object is achieved by compounds of the following general formula (I)

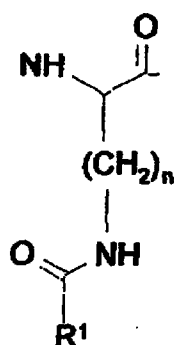
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in which

- 5 A is an acetyl or a 3-(4-fluorophenyl)propionyl group,
 X_{xx}^1 is D-Nal(1) or D-Nal(2),
 $X_{xx}^2-X_{xx}^3$ is D-Cpa-D-Pal(3) or a single bond,
 X_{xx}^4 is Ser,
 X_{xx}^5 is N-Me-Tyr,
 10 X_{xx}^6 is D-Cit, D-Hci or a D-amino acid group of the
 general formula (II)

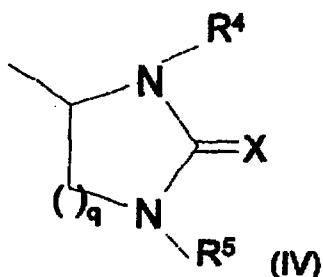


(II)

- in which n is the number 3 or 4, where R^1 is a group
 15 having the general formula III



- where p is an integer from 1 to 4, R^2 is hydrogen or an
 20 alkyl group and R^3 is an unsubstituted or substituted
 aryl group or heteroaryl group, or R^1 is a 3-amino-
 1,2,4-triazole-5-carbonyl group or R^1 is a ring of the
 general formula (IV)



in which q is the number 1 or 2, R⁴ is a hydrogen atom or an alkyl group, R⁵ is a hydrogen atom or an alkyl group and X is an oxygen or sulphur atom,

Xxx⁷ is Leu or Nle,

5 Xxx⁸ is Arg or Lys(iPr),

Xxx⁹ is Pro and

Xxx¹⁰ is Ala or Sar,

and their salts with pharmaceutically acceptable acids, in particular the acetates, embonates and

10 trifluoroacetates.

Among the compounds according to the invention, those are particularly preferred in which Xxx⁶ is D-[ε-N'-(imidazolidin-2-on-4-yl)formyl]-Lys, D-(3-amino-1,2,4-
15 triazole-3-carbonyl)-Lys, abbreviated D-Lys(Atz) or D-[ε-N'-4-(4-Amidinophenyl)amino-1,4-dioxobutyl]-Lys, abbreviated D-Lys(B).

Further particularly preferred compounds according to
20 the invention are:

Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Hci⁶-Nle⁷-Arg⁸-Pro⁹-D-Ala¹⁰-NH₂,

Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Lys(Atz)⁶-Leu⁷-Arg⁸-Pro⁹-D-Ala¹⁰-NH₂,

25 Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Lys(B)⁶-Leu⁷-Lys(iPr)⁸-Pro⁹-D-Ala¹⁰-NH₂,

Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Lys(B)⁶-Leu⁷-Arg⁸-Pro⁹-D-Ala¹⁰-NH₂,

30 Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Hci⁶-Nle⁷-Lys(iPr)⁸-Pro⁹-Ala¹⁰-NH₂,

Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Hci⁶-Nle⁷-Lys(iPr)⁸-Pro⁹-Sar¹⁰-NH₂,

Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Hci⁶-Nle⁷-Arg⁸-Pro⁹-Sar¹⁰-NH₂,

35 3-(4-Fluorophenyl)propionyl-D-Nal(1)¹-Ser⁴-N-Me-Tyr⁵-D-Lys(Atz)⁶-Leu⁷-Arg⁸-Pro⁹-D-Ala¹⁰-NH₂,

and also their salts with the abovementioned pharmaceutically acceptable acids.

The compounds according to the invention can be used for the treatment of hormone-dependent tumours, in particular prostate carcinoma or breast cancer, and
5 also for non-malignant indications whose treatment necessitates LH-RH hormone suppression. For this, they are mixed with the customary vehicles and excipients and formulated as medicaments.

10 The synthesis of compounds according to formula (I) can both be carried out either by classical fragment condensation or by solid-phase synthesis according to Merrifield with synthesis following one another using D-lysine already acylated in the side chain with the
15 carboxylic acid of the general formula $R^1\text{-COOH}$ or by reaction of a decapeptide unit with the appropriate carboxylic acids by amide linkage in the side chain of D-lysine⁶. Accordingly, the introduction of the $R^1\text{-CO-}$ group can be performed in three different positions in
20 the process: before the condensation of the individual units to give the peptide, after the incorporation of lysine or ornithine in the peptide chain, but before the condensation of the next unit or after condensation of all units.

25

The compounds of the formula (I) are synthesized according to the known methods, such as, for example, by pure solid-phase technique, partly solid-phase technique (so-called fragment condensation) or by the
30 classical solution couplings (see M. Bodanszky, "Principles of Peptide Synthesis", Springer Verlag 1984).

For example, the methods of solid-phase synthesis are described in the textbook "Solid Phase Peptide
35 Synthesis", J.M. Stewart and J.D. Young, Pierce Chem. Company, Rockford, III, 1984, and in G. Barany and R.B. Merrifield "The Peptides", Ch. 1, pp. 1-285, 1979, Academic Press Inc. Classical solution syntheses are

described in detail in the treatment "Methoden der Organischen Chemie [Methods of Organic Chemistry] (Houben-Weyl), Synthese von Peptiden" [Synthesis of Peptides] E. Wunsch (Editor) 1974, Georg Thieme Verlag, 5 Stuttgart, FRG.

The stepwise synthesis is carried out, for example, by first covalently bonding the carboxy-terminal amino acid whose α -amino group is protected to an insoluble support which is customary for this, removing the 10 α -amino protective group of this amino acid, bonding the free amino group thus obtained to the next protected amino acid via its carboxyl group, and in this manner linking the customary amino acids of the 15 peptide to be synthesized in the correct sequence step for step, and after linkage of all amino acids removing the finished peptide from the support and removing any further side function protective groups which may be present. The stepwise condensation is carried out in a 20 conventional manner by synthesis from the corresponding, customarily protected amino acids.

The linkage of the individual amino acids to one another is carried out according to the methods 25 customary for this; those particularly suitable are:

- Symmetrical anhydride method in the presence of dicyclohexylcarbodiimide or diisopropylcarbodiimide (DCC, DIC)
- Carbodiimide method generally
- 30 • Carbodiimide/hydroxybenzotriazole method
(see The Peptides, Volume 2, Ed. E. Gross and J. Meienhofer).

In the fragment coupling, the azide coupling, which proceeds without racemization, or the DCC-1- 35 hydroxybenzotriazole or DCC-3-hydroxy-4-oxo-3,4-dihydro-1,2,3-benzotriazine method is preferably used. Activated esters of fragments can also be employed.

- Esters of N-protected amino acids, such as, for example, N-hydroxysuccinimide esters or 2,4,5-trichlorophenyl esters, are particularly highly suitable for the stepwise condensation of amino acids.
- 5 The aminolysis can be very well catalysed by N-hydroxy compounds which have approximately the acidity of acetic acid, such as, for example, 1-hydroxybenzotriazole.
- 10 Intermediate amino protective groups which present themselves are groups which are removed by hydrogenation, such as, for example, the benzyloxycarbonyl radical (= Z radical) or groups which can be removed by weak acid. Suitable protective groups
- 15 for the α -amino groups are, for example: tertiary butyloxycarbonyl groups, fluorenylmethyloxycarbonyl groups, carbobenzoxy groups or carbobenzoethio groups (if appropriate in each case having a p-bromo [sic] or p-nitrobenzyl radical), the
- 20 trifluoroacetyl group, the phthalyl radical, the o-nitrophenoxyacetyl group, the trityl group, the p-toluenesulphonyl group, the benzyl group, benzyl radicals substituted in the benzene nucleus (p-bromo [sic] or p-nitrobenzyl radical) and the α -phenylethyl
- 25 radical. Reference is also made here to P. Greenstein and Milton Winitz, Chemistry of Amino Acids, New York 1961, John Wiley and Sons, Inc., Volume 2, for example page 883 et seq., "Principles of Ppetide Synthesis", Springer Verlag 1984, "Solid Phase Peptide Synthesis",
- 30 J.M. Stewart and J.D. Young, Pierce Chem. Company, Rockford, III, 1984, G. Barany and R.B. Merrifield "The Peptides", Ch. 1, pp. 1-285, 1979, Academic Press Inc., and also The Peptides, Volume 2, Ed. E. Gross and J. Maierhofer, Academic Press, New York. These
- 35 protective groups are fundamentally also suitable for the protection of further functional side groups (OH groups, NH_2 groups) of the corresponding amino acids.

Hydroxyl groups present (serine, threonine) are preferably protected by benzyl groups and similar groups. Further amino groups not in the α -position (for example amino groups in the ω -position, guanidino group of arginine) are preferably orthogonally protected.

The individual amino acid units, excluding lysine or ornithine modified by the R^1 -CO-group, are commercially obtainable. A possible course of the process for the preparation of the latter compounds is as follows:

1. The α -carboxylic acid group is amidated.
2. The ϵ -amino group is protected by the Z group.
3. The α -amino group is protected by the Boc group such that a selectivity with respect to the later removal of the amino protective groups results.
4. The Z group on the ϵ -amino group is removed.
5. The desired group R^4 -CO- is introduced on the ϵ -amino group.
6. The Boc group on the α -amino group is removed.
7. The α -amino group is provided with the Z group.

For the introduction of the R^1 -CO-group by reaction of the amino group of the lysine with appropriate carboxylic acid, suitable processes are fundamentally the same processes as described above for the linkage of the amino acids. However, condensation using carbodiimide, for example 1-ethyl-3-(3-dimethylamino-propyl)carbodiimide, and 1-hydroxybenzotriazole is particularly preferred.

The reaction for the linkage of the amino acids takes place in an inert solvent or suspending agent which is customary for this (for example dichloromethane), it being possible to add dimethylformamide, if necessary, to improve the solubility.

Suitable synthetic supports are insoluble polymers, for example polystyrene resin in bead form, which can be

swollen in organic solvents (for example a copolymer of polystyrene and 1% divinylbenzene). The synthesis of a protected decapeptide amide on a methylbenzhydrylamide [sic] resin (MBHA resin, i.e. polystyrene resin provided with methylbenzhydrylamide [sic] groups), which affords the desired C-terminal amide function of the peptide after HF cleavage from the support, can be carried out according to the following flow diagram:

10 Flow diagram
Peptide synthesis protocol

Stage	Function	Solvent/Reagent (v/v)	Time
1	Washing	Methanol	2 × 2 min
2	Washing	DCM	3 × 3 min
3	Removal	DCM/TFA (1:1)	1 × 30 min
4	Washing	Isopropanol	2 × 2 min
5	Washing	Methanol	2 × 2 min
6	Washing	DCM	2 × 3 min
7	Neutralization	DCM/DIPEA (9:1)	3 × 5 min
8	Washing	Methanol	2 × 2 min
9	Washing	DCM	3 × 3 min
10	STOP	Addition of the Boc-As in DCM + DIC + HOBt	
11	Coupling	DCM, optionally DCM/DCF	approx. 90 min
12	Washing	Methanol	3 × 2 min
13	Washing	DCM	2 × 3 min

The N α -Boc-protected amino acids are customarily coupled in a three fold molar excess in the presence of diisopropylcarbodiimide (DIC) and 1-hydroxybenzotriazole (HOBt) in CH₂Cl₂/DMF in the course of 90 min, and the Boc-protected group is removed by action of 50% trifluoroacetic acid (TFA) in CH₂Cl₂ for half an hour. To check for complete conversion, the chloranil test according to Christensen and the Kaiser's ninhydrin test can be used. Radicals of free amino function [sic]

- are blocked by acetylation in a five fold excess of acetylimidazole in CH_2Cl_2 . The sequence of the reaction steps of the peptide synthesis on the resin follows from the flow diagram. For the removal of the resin-bound peptides, the respective final product of the solid phase synthesis is dried in vacuo over P_2O_5 and treated at 0°C for 60 min in a 500-fold excess of HF/anisole 10:1/v:v.
- 10 After distilling of HF and anisole in vacuo, the peptide amides are obtained as white solids by washing with anhydrous ethyl ether with stirring, and the removal of polymeric support additionally obtained is carried out by washing with 50% strength aqueous acetic acid. By careful concentration of the acetic acid solutions in vacuo, the respective peptides can be obtained as highly viscous oils, which are converted into white solids after addition of abs. ether in the cold.
- 20 Further purification is carried out by routine methods of preparative high-pressure liquid chromatography (HPLC).
- 25 The conversion of the peptides into their acid addition salts can be effected in a manner known per se by reaction thereof with acids. Conversely, free peptides can be obtained by reaction of their acid addition salts with bases. Peptide embonates can be prepared by reaction of trifluoroacetic acid salts (TFA salts) of the peptide with free embonic acid (pamoic acid) or the corresponding disodium salt of embonic acid. For this, the peptide TFA salt is treated in aqueous solution with the solution of disodium embonate in polar aprotic medium, preferably dimethylacetamide, and the pale yellow precipitate formed is isolated.
- 35

The following examples serve to illustrate the invention without restricting it.

Example 1

5

Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Hci⁶-Nle⁷-
Arg⁸-Pro⁹-D-Ala¹⁰-NH₂

10 The synthesis was carried out according to a solid-
phase flow diagram (Peptide Synthesis Protocol, p. 11)
with DIC/HOBt coupling, starting from 3.3 g of MBHA
resin (loading density 1.08 mmol/g). After HF cleavage
from the polymeric support, 3.4 g of crude peptide were
15 obtained, which were purified by standard processes of
preparative HPCI [sic]. After subsequent freeze-drying,
1.43 g of HPLC-uniform product of the empirical formula
C₇₂, H₉₆, N₁₇, O₁₄, Cl [sic] having correct FAB-MS:
1458.7 (M+H⁺) (calc: 1457.7), and corresponding ¹H-NMR
spectrum were obtained.

20

¹H-NMR (500 MHz, D₂O/DMSO-d₆, δ in ppm):

8.7 to 7.2, several m, arom. H and incompletely
exchanged NH; 6.92 and 6.58, 2d, 2x2H, arom. H p-Cl-
Phe; 5.2 to 3.5, several m, Cα-H and aliph. H; 3.2 to
25 2.6, several m, arom. Cβ-H 2.1 to 0.7, several m,
residual aliph. H; 1.70, s, 3H, acetyl; 1.20, d, 3H,
Cβ-H Ala; 0.8, m, Cδ-H Leu

Example 2

30

Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Lys(B)⁶-
Leu⁷-Lys(iPr)⁸-Pro⁹-D-Ala¹⁰-NH₂

35 The synthesis was carried out according to a flow
diagram (Peptide Synthesis Protocol, p. 11) with
DIC/HOBt coupling, starting from 4.0 g of MBHA resin
(loading density 1.11 mmol/g). After HF cleavage from
the polymeric support, 4.87 g of crude peptide were

obtained, which were purified by standard processes of preparative HPLC [sic]. After subsequent freeze-drying, 0.93 g of HPLC-uniform product was obtained, which was reacted with 4-amidinophenylamino-4-oxobutyric acid in the presence of BOP as a coupling reagent to give the desired compound. After fresh HPLC purification, 148 mg of target compound of the empirical formula C₈₅, H₁₁₂, N₁₇, O₁₅, Cl [sic] having correct ESI-MS: 1647.6 (M+H⁺) (calc: 1645.8), and corresponding ¹H-NMR spectrum were obtained.

¹H-NMR (500 MHz, DMSO-d₆, δ in ppm):
10.4, s, 1H and 9.13, s, 2H, and 8.94, s, 2H, NHs of 4-amidinoaniline; 8.6 to 7.35, several m, arom. H and NH; 7.22 and 7.18, 2d, 4H, arom. H (pCl)Phe; 6.95 and 6.58, 2d, 4H, arom. H Tyr; 5.2 to 3.5, several m, Cα-H and aliphatic. H; 3.3 to 2.4, several m, Cβ-H and N-CH₃; 2.1 to 1.1, several m, residual aliphatic. H; 1.68, s, 3H, acetyl; 1.20, d, 3H, Cβ-H Ala; 0.83, dd, 6H, Cδ-H Leu

Example 3

Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Lys(B)⁶-
Leu⁷-Arg⁸-Pro⁹-D-Ala¹⁰-NH₂

The synthesis was carried out according to a solid-phase flow diagram (Peptide Synthesis Protocol, p. 11) with DIC/HOBt coupling, starting from 4.0 g of MBHA resin (loading density 0.97 mmol/g). After HF cleavage from the polymeric support, 4.0 g of crude peptide were obtained, which were purified by standard processes of preparative HPLC [sic]. After subsequent freeze-drying, 1.39 g of HPLC-uniform product were obtained, which was [sic] reacted with 4-amidinophenylamino-4-oxobutyric acid in the presence of BOP as a coupling reagent to give the desired compound. After fresh HPLC purification, 440 mg of target compound of the

empirical formula C82, H106, N19, O15, Cl [sic] having correct ESI-MS: 1632.7 (M+H⁺) (calc: 1631.7), and corresponding ¹H-NMR spectrum were obtained.

- 5 ¹H-NMR (500 MHz, DMSO-d₆, δ in ppm):
10.4, s, 1H and 9.15, s, 2H, and 9.0, s, 2H, NHs of 4-amidinoaniline; 8.60, m, 2H, arom. H; 8.3 to 7.2, several m, arom. H and NH; 7.27 and 7.20, 2d, 4H, arom. H (pCl)Phe; 6.96 and 6.60, 2d, 4H, arom. H Tyr; 5.2 to
10 3.5, several m, Cα-H and aliphatic. H; 3.2 to 2.4, several m, Cβ-H and N-CH₃; 2.1 to 1.1, several m, residual aliphatic. H; 1.70, s, 3H, acetyl; 1.20, d, 3H, Cβ-H Ala; 0.85, dd, 6H, Cδ-H Leu

15 Example 4

Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Hci⁶-Nle⁷-Lys(iPr)⁸-Pro⁹-D-Ala¹⁰-NH₂

- 20 The synthesis was carried out according to a solid-phase flow diagram (Peptide Synthesis Protocol, p. 11) with DIC/HOBt coupling, starting from 2.5 g of MBHA resin (loading density 1.08 mmol/g). After HF cleavage from the polymeric support, 2.78 g of crude peptide
25 were obtained, which were purified by standard processes of preparative HPLC [sic]. After subsequent freeze-drying, 400 mg of HPLC-uniform product of the empirical formula C75, H102, N15, O14, Cl [sic] having correct ESI-MS: 1472.6 (M+H⁺) (calc: 1471.7), and
30 corresponding ¹H-NMR spectrum were obtained.

- ¹H-NMR (500 MHz, D₂O/DMSO-d₆, δ in ppm):
8.62, m, 2H, 8.30, m, 2H, 7.80, m, 4H, 7.66, s, 1H, 7.47, m, 2H, 7.36, d, 1H, aromat. H; 7.25 and 7.20,
35 2 d, 4H, arom. H (pCl)Phe; 6.96 and 6.63, 2d, 4H, aromat. H Tyr; 5.10 to 4.0, several m, Cα-H and aliphatic. H; 3.75 to 2.65, several m, Cβ-H and N-CH₃; 2.1 to 1.05, several m, residual aliphatic. H; 1.74, s,

3H, acetyl; 1.23, d, 3H, C β -H Ala; 1.20, m, CH₃ isoprop.; 0.8, m, 3H, C δ -H Nle

Example 5

5

Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Hci⁶-Nle⁷-
Lys(iPr)⁸-Pro⁹-Sar¹⁰-NH₂

10 The synthesis was carried out according to a solid-
phase flow diagram (Peptide Synthesis Protocol, p. 11)
with DIC/HOBt coupling, starting from 2.5 g of MBHA
resin (loading density 1.08 mmol/g). After HF cleavage
from the polymeric support, 2.74 g of crude peptide
15 were obtained, which were purified by standard
processes of preparative HPCI [sic]. After subsequent
freeze-drying, 840 mg of HPLC-uniform product of the
empirical formula C₇₅, H₁₀₂, N₁₅, O₁₄ Cl [sic] having
correct ESI-MS: 1472.6 (M+H⁺) (calc: 1471.7), and
corresponding ¹H-NMR spectrum were obtained.

20

¹H-NMR (500 MHz, D₂O/DMSO-d₆, δ in ppm):

8.6, m, 2H, 8.3, m, 2H, 7.85, m, 2H, 7.8, m, 2H, 7.65,
s, 1H, 7.46, m, 2H, 7.35, d, 1H, aromat. H; 7.23 and
7.17, 2 d, 4H, arom. H (pCl)Phe; 7.0 and 6.6, 2d, 4H,
25 aromat. H Tyr; 5.10 to 3.8, several m, C α -H and
aliphat. H; 3.75 to 2.6, several m, C β -H and N-CH₃; 2.2
to 1.05, several m, residual aliphat. H; 1.70, s, 3H,
acetyl; 1.23, d, 3H, C β -H Ala; 1.20, m, CH₃ isoprop.;
0.8, m, 3H, C δ -H Nle

Example 6

3-(4-Fluorophenyl)propionyl-D-Nal(1)¹-Ser⁴-N-Me-Tyr⁵-D-Lys(Atz)⁶-Leu⁷-Arg⁸-Pro⁹-D-Ala¹⁰-NH₂

5

The synthesis was carried out according to a solid-phase flow diagram (Peptide Synthesis Protocol, p. 11) with DIC/HOBt coupling, starting from 9.2 g of MBHA resin (loading density 1.08 mmol/g). After HF cleavage
10 from the polymeric support, 5.8 g of crude peptide were obtained, which were purified by standard processes of preparative HPCI [sic]. After subsequent freeze-drying, 2.0 g of HPLC-uniform unsubstituted octapeptide were obtained, of which 0.4 mmol was reacted with 0.5 mmol
15 of 3-amino-1,2,4-triazole-5-carboxylic acid in the presence of PyBOP as a coupling reagent to give 790 mg of crude product of the desired compound. After fresh HPLC purification, 200 mg of target compound of the empirical formula C₆₄, H₈₆, N₁₇, O₁₂, F [sic] having
20 correct FAB-MS: 1304.6 (M+H⁺) (calc: 1303.6) were obtained.

¹H-NMR (500 MHz, D₂O/DMSO-d₆, δ in ppm):

8.14, m, 1H, 7.90, m, 1H, 7.80, m, 1H, 7.50, m, 2H,
25 7.35, m, 2H, 7.0, m, 6H, 7.63, m, 2H, aromat. H; 5.0, m, 1H, 4.83, m, 2H, 4.41, m, 1H, 4.30 - 4.05, several m, 4H, Cα-H; 3.66 to 2.25, several m, aliphatic and aromat. side-chain H; 2.95, s, and 2.75, s, N-Me; 2.05 to 1.1, several m, residual aliphatic H; 1.20, d, Cβ-H
30 Ala; 0.75, m, 6H, Cδ-H Leu

The compounds according to formula I according to the invention were investigated for their receptor binding. The process closely followed the process described in
35 Beckers et al., Eur. J. Biochem. 231, 535-543 (1995). Cetrorelix obtained according to the synthesis disclosed above was iodinated with [¹²⁵I] (Amersham; specific activity 80.5 Bq/fmol) using the IodoGen

reagent (Pierce). The reaction mixture was purified by reverse-phase high-performance liquid chromatography, monoiodinated cetorelix being obtained without unlabelled peptide. In each case, about 80% of the
5 $[^{125}\text{I}]$ -cetorelix and the unlabelled compound according to the invention were suitable for the specific receptor association.

The compounds according to the invention can be tested
10 for their in-vitro action using the following Methods 1 and 2, the binding affinities in the binding assay being determined with $[^{125}\text{I}]$ -Cetorelix (Method 1) and the functional activities being determined with triptorelin as an agonist stimulus (Method 2).

15

Method 1.

Receptor binding assay according to Beckers, T., Marheineke, K., Reiländer, H., Hilgard P. (1995)
20 "Selection and characterization of mammalian cell lines with stable overexpression of human pituitary receptors for gonadoliberin (GnRH)" Eur. J. Biochem. 231, 535-543.

25 For investigation of the receptor binding, cetorelix was iodinated using the IodoGen reagent (Pierce) with $[^{125}\text{I}]$ (Amersham; 80.5 Bq/fmol specific activity). The reaction mixture was purified by high-performance liquid chromatography with exchanged phases,
30 monoiodinated cetorelix being obtained without unlabelled peptide. About 80% of the $[^{125}\text{I}]$ cetorelix was capable of specific receptor association.

The receptor binding assay was carried out under
35 physiological conditions as described (Beckers et al., 1995) using intact cells. Subconfluent cultures of stably transfected LTK⁻ cells, which express the human LHRH receptor, were separated off by incubation in

NaCl/P_i (137 mM NaCl, 2.7 mM KCl, 8.1 mM Na₂HPO₄,
11.47 mM KH₂PO₄)/1 mM EDTA and collected by
centrifugation. The cell pellet was resuspended in
binding buffer (DMEM without H₂CO₃, with 4.5 g/l of
5 glucose, 10 mM Hepes pH 7.5, 0.5% (mass/volume) BSA,
1 g/l bacitracin, 0.1 g/l SBTI, 0.1% (mass/volume)
NaN₃). For displacement assays, 0.25×10^6 cells/100 μ l
were incubated with approximately 225 pM of the [¹²⁵I]-
cetrorelix (specific activity 5-10 $\times 10^5$ dpm/pmol) and
10 various concentrations of unlabelled compound according
to the invention as competitor. The cell suspension in
100 μ l of binding medium was layered in 400 μ l assay
tubes over 200 μ l of 84% by volume silicone oil (Merck
Type 550)/16% by volume paraffin oil. After incubation
15 for 1 h at 37°C with slow, continuous shaking, the
cells were separated from the incubation medium by
centrifugation for 2 min at 9000 rpm (rotor type
HTA13.8; Heraeus Sepatec, Osterode/Germany). The tips
of the tubes which contained the cell pellet were cut
20 off. Cell pellet and supernatants were then analysed by
counting the γ radiation. The amount of non-
specifically bound material was determined at a final
concentration of 1 μ M with inclusion of unlabelled
cetrorelix and was typically $\leq 10\%$ of the total bound
25 material. The analysis of the binding data was carried
out using the EBDA/ligand analysis programme (Biosoft
V3.0).

Method 2.

30

Functional assay for the determination of the
antagonistic activity

The assay was carried out, provided with some
35 modifications, as described in Beckers, T., Reiländer,
H., Hilgard, P. (1997) "Characterization of
gonadotropin-releasing hormone analogs based on a
sensitive cellular luciferase reporter gene assay",

Analyt. Biochem. 251, 17-23 (Beckers et al., 1997).
10,000 cells per well, which express the human LHRH
receptor and a luciferase reporter gene, were cultured
for 24 h in microtitre plates using DMEM with additives
5 and 1% (v:v) FCS_i. The cells were then stimulated with
1 nM [D-Trp⁶] LHRH for 6 h. Antagonistic compounds
according to the invention were added before the
stimulation and the cells were lysed at the end for the
quantification of the cellular Luc activity. The
10 calculation of the IC₅₀ values from dose-effect curves
was carried out by non-linear regression analysis using
the Hill model (Programme EDX 2.0 from C. Grunwald,
Arzneimittelwerk Dresden).

15 The quantification of the Luc activity was carried out
in duplicate essentially as described (Promega
Technical Bulletins #101/161) using the respective
luciferase assay system (Promega E4030). Owing to
addition of coenzyme A (CoA), an oxidation of
20 luciferyl-CoA takes place with advantageous kinetics.
After the removal of the culture medium from the
microtitre plate, the cells were lysed by addition of
100 µl of lysis buffer (25 mM tris-phosphate pH 7.8,
2 mM dithiothreitol, 2 mM 1,2-diaminocyclohexane-
25 N,N,N',N'-tetraacetic acid (CDTA), 10% (v:v) glycerol,
1% (v:v) Triton X-100). After incubation at room
temperature for 15 min, 10 µl of cell lysate were
transferred into a white microtitre plate suitable for
luminometric detection (Dynatech). The enzymatic
30 reaction was initiated by addition of 50 µl of assay
buffer (20 mM tricine pH 7.8, 1.07 mM (MgCO₃)₄Mg(OH)₂,
2.67 mM MgSO₄, 0.1 mM ethylenediaminetetraacetic acid
(EDTA), 33.3 mM dithiothreitol, 270 µM coenzyme A,
470 µM glow-worm (Photinus pyralis) luciferin, 530 µM
35 rATPNa₂). After one minute, the luminescence was
determined for a total time of one second with a signal
half-life of five minutes using the EG&G Berthold
MicroLumat LB 96 P.

In this way, the following in-vitro data were obtained, K_D being the binding affinities and IC_{50} being the functional activity and pM being picomoles per litre:

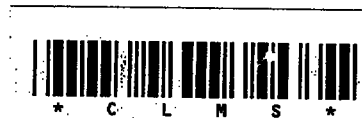
5

Compound	K_D [pM]	IC_{50} [pM]
cetrorelix	170 (21)	198 (5)
Example 1 (Acetate salt)	n.d.	242 (3)
Example 2	181 (1)	684 (2)
Example 3	154 (1)	492 (2)
Example 6	n.d.	221 (2)

n.d. = not determined

() = number of independent experiments

Patent Claims



1. Compounds of the general formula I

5 A-Xxx¹-Xxx²-Xxx³-Xxx⁴-Xxx⁵-Xxx⁶-Xxx⁷-Xxx⁸-Xxx⁹-Xxx¹⁰-
NH₂ (I)

in which

10 A is an acetyl or a 3-(4-fluorophenyl)propionyl
group,

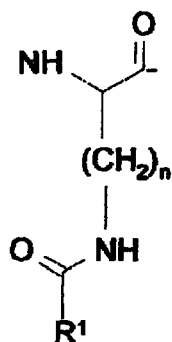
Xxx¹ is D-Nal(1) or D-Nal(2),

Xxx²-Xxx³ is D-Cpa-D-Pal(3) or a single bond,

Xxx⁴ is Ser,

Xxx⁵ is N-Me-Tyr,

15 Xxx⁶ is D-Cit, D-Hci or a D-amino acid group of
the general formula (II)



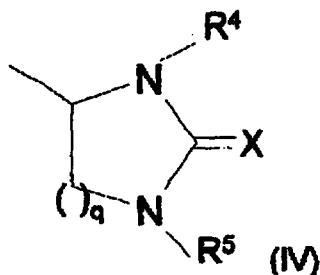
(II)

20 in which n is the number 3 or 4, where R¹ is a
group having the general formula III



25 where p is an integer from 1 to 4, R² is hydrogen
or an alkyl group and R³ is an unsubstituted or
substituted aryl group or heteroaryl group, or R¹
is a 3-amino-1,2,4-triazole-5-carbonyl group or R¹
is a ring of the general formula (IV)

30



in which q is the number 1 or 2, R⁴ is a hydrogen atom or an alkyl group, R⁵ is a hydrogen atom or an alkyl group and X is an oxygen or sulphur atom,
 5 Xxx⁷ is Leu or Nle,
 Xxx⁸ is Arg or Lys(iPr),
 Xxx⁹ is Pro and
 Xxx¹⁰ is Ala or Sar,
 10 and their salts with pharmaceutically acceptable acids.

2. Compounds according to Claim 1, in which the salt is an acetate, trifluoroacetate or embonate.
- 15 3. Compounds according to Claim 1 or 2, in which Xxx⁶ is D-[ε-N'-(imidazolidin-2-on-4-yl)formyl]-Lys, D-(3-amino-1,2,4-triazole-3-carbonyl)-Lys, abbreviated D-Lys(Atz) or D-[ε-N'-4-(4-amidino-phenyl)amino-1,4-dioxobutyl]-Lys, abbreviated D-Lys(B).
- 20 4. Compound according to Claim 1 having the formula:
 Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Hci⁶-
 25 Nle⁷-Arg⁸-Pro⁹-D-Ala¹⁰-NH₂.
5. Compound according to Claim 1 having the formula:
 Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-
 Lys(Atz)⁶-Leu⁷-Arg⁸-Pro⁹-D-Ala¹⁰-NH₂.
- 30 6. Compound according to Claim 1 having the formula:
 Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-
 Lys(B)⁶-Leu⁷-Lys(iPr)⁸-Pro⁹-D-Ala¹⁰-NH₂.



7. Compound according to Claim 1 having the formula:
Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-
Lys(B)⁶-Leu⁷-Arg⁸-Pro⁹-D-Ala¹⁰-NH₂.
8. Compound according to Claim 1 having the formula:
Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Hci⁶-
Nle⁷-Lys(iPr)⁸-Pro⁹-D-Ala¹⁰-NH₂.
9. Compound according to Claim 1 having the formula:
Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Hci⁶-
Nle⁷-Lys(iPr)⁸-Pro⁹-Sar¹⁰-NH₂.
10. Compound according to Claim 1 having the formula:
Ac-D-Nal(2)¹-D-Cpa²-D-Pal(3)³-Ser⁴-N-Me-Tyr⁵-D-Hci⁶-
Nle⁷-Arg⁸-Pro⁹-Sar¹⁰-NH₂.
11. Compound according to Claim 1 having the formula:
3-(4-fluorophenyl)propionyl-D-Nal(1)¹-Ser⁴-N-Me-
Tyr⁵-D-Lys(Atz)⁶-Leu⁷-Arg⁸-Pro⁹-D-Ala¹⁰-NH₂.
12. Pharmaceutical composition comprising a compound
according to one of Claims 1 to 11.
13. Process for the preparation of compounds of the
general formula I according to Claim 1, in which
fragments from units Xxx^m provided with suitable
protective groups, in which m is an integer from 1
to 10 and Xxx¹ is acetylated, are synthesized on a
solid phase or in solution according to customary
processes, then the fragments are linked to a
solid phase by segment coupling and after
conclusion of the coupling the compounds of the
general formula I are removed from the solid phase
using customary processes with amidation on the
unit Xxx¹⁰.



14. Use of the substances according to Claims 1 to 11
for producing medicaments for the treatment of
hormone-dependent tumours, in particular prostate
carcinoma or breast cancer, and also for non-
malignant indications whose treatment necessitates
LH-RH hormone suppression.
15. Process for producing medicaments, in which
compounds according to Claims 1 to 11 are mixed
with the customary vehicles and excipients and
formulated as medicaments.