

COMPOUND - DIAGNOSTIC MARKER FOR KIDNEY CANCER, METHOD FOR DETECTING ENZYMATICAIVITY, METHOD FOR DIAGNOSIS OF KIDNEY CANCER, KIT COMPRISING THE COMPOUND, USES OF THE COMPOUND AND METHOD FOR THE TREATMENT OF KIDNEY CANCER

5 The invention relates to a new chemical compound, a diagnostic marker, for use in medicine, more specifically in cancer diagnostics, in particular diagnosis of kidney cancer. The invention also relates to an *in vitro* method for detecting enzymatic activity present in a subject's body fluid, in particular deriving from kidney cancer cells, using the compound, an *in vitro* method for the diagnosis of kidney cancer using the compound, a kit comprising
10 the compound, use of the compound for the detection of enzymatic activity specific to kidney cancer, use of the compound for the diagnosis of kidney cancer, the compound for use as a diagnostic marker for kidney cancer. The invention further relates to a method for the treatment of kidney cancer comprising a step of carrying out the method for the diagnosis of kidney cancer as specified above.

15 **Background of the invention**

In 2020, more than 430 000 patients worldwide developed kidney cancer and it was the fourteenth most commonly diagnosed malignant neoplasms worldwide. Kidney cancer is a relatively rare cancer and accounts for 2.4 % of new diagnoses among solid tumours worldwide. Malignant kidney neoplasms are divided into primary and secondary ones, the
20 most common being renal cell carcinoma (RCC), which accounts for approximately 90% of all malignant kidney neoplasms. RCC represents a heterogeneous group of lesions that differ in both histological structure and clinical manifestations and molecular alterations. The most common histological subtype is clear cell renal cell carcinoma (ccRCC), which accounts for 70-80% of all renal cell carcinomas and is also the most aggressive of the subtypes, with the
25 highest rate of local invasion, metastasis and the highest mortality. In early-stage kidney cancer, the disease is not accompanied by pain or other characteristic symptoms. More than 30 % of patients with RCC have metastases at the time of diagnosis, and in 30 % of patients with RCC confined to the organ, metastases develop after local treatment. RCC remains a diagnostic and treatment challenge. In the majority of patients, this cancer is detected
30 incidentally as a focal lesion during imaging studies (X-ray, CT, MRI) performed intentionally or for other reasons.

The prognosis depends on the extent of the neoplasm. Early diagnosis is very important. 75% of patients diagnosed at an early stage survive five years or longer.

There is no laboratory test, and no “cancer marker”, or a set of tests which would enable and early and reliable diagnosis of kidney cancer. Imaging methods such as X-ray, CT scan, MRI or positron magnetic resonance imaging are used for effective diagnosis, followed by biopsy of the tumour. A small population of patients (5-10%) shows changes in the expression of specific genes which predisposes this group to genetic testing.

It is known that the process of initiation, growth and dissemination of cancer cells involves many factors, including many enzymes, in particular hydrolytic enzymes, especially proteolytic enzymes. Such enzymes catalyse enzymatic cleavage (hydrolytic or proteolytic) of proteins and peptides into smaller fragments thereof. This process enables cancer cells to expand by colonizing new tissues, enhancing the process of blood vessels formation (angiogenesis), which enables effective delivery of nutrients to a tumour. Moreover, these enzymes are present as a result of death of healthy cells due to a tumour growth process. All these processes form a characteristic and specific profile of the enzymatic (proteolytic) activity of cancer cells, characteristic to a tumour.

In this field, there are known chromogenic peptide molecules that undergo enzymatic breakdown into smaller fragments resulting in a change or increase in the colour of the solution being tested. This chromogenic effect is a consequence of the release of a chromophore (e.g. 4-nitroanilide or 2-aminobenzoic acid) from a chromogenic peptide molecule.

This type of chromogenic molecules and their uses are known, for example, from the publication by Erlanger BF, Kokowsky N, Cohen W., “The preparation and properties of two new chromogenic substrates of trypsin”, Arch Biochem Biophys., November 1961; 95:271-8 and Hojo K, Maeda M, Iguchi S, Smith T, Okamoto H, Kawasaki K. Amino acids and peptides. XXXV. “Facile preparation of p-nitroanilide analogs by the solid-phase method”, Chem Pharm Bull (Tokyo), November 2000; 48(11):1740-4.

However, the use of this class of compounds in the diagnosis of kidney cancer has not been described so far.

Methods for obtaining chromogenic peptides which consist in attaching individual components in appropriate time and stoichiometric conditions are also known in the prior art. The process of attaching consists of subsequent steps in which individual elements (amino acid derivatives) are attached, residues are washed off and protecting groups are sequentially removed and washed again. This cycle is repeated for each amino acid residue. The obtained peptide is separated from resin by a reaction in acidic conditions. Subsequently,

the solution is separated from resin in the filtration process and then the peptide is precipitated from the solution by means of a non-polar solvent.

Chromogenic peptide compounds appropriate for a specific and early diagnosis of kidney cancer or methods to obtain them are however not known in the prior art.

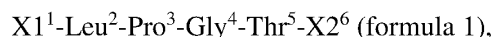
- 5 Therefore, in this field there is an urgent need for “a cancer marker” for kidney cancer, which would enable an early, sensitive and specific diagnosis of kidney cancer in a non-invasive and reliable manner, and for diagnostic methods and treatment methods using such a diagnostic marker.

The object of the present invention is to provide a novel, specific diagnostic marker for
10 kidney cancer and diagnostic methods using such a marker for a non-invasive, quick, sensitive and specific, early detection of kidney cancer, which would also be appropriate for screening tests, as well as treatment methods using such a marker.

These objects have been achieved by the inventions defined in the attached patent claims, whereas preferred variants thereof are defined in the dependent claims.

15 **Summary of the invention**

The invention provides a compound having formula 1:



wherein X1 comprises or consists of molecule C1, and X2 comprises or consists of molecule C2,

- 20 wherein the pair of molecules C1 and C2 is a pair of florescence donor and fluorescence acceptor,

and wherein the compound undergoes enzymatic cleavage into the fragments X1-Leu-Pro-Gly-Thr-OH (Fragment 1) and X2 (Fragment 2) with a generation of a measurable optical signal upon spatial separation of molecules C1 and C2.

- 25 The compound according to the invention preferably undergoes hydrolytic cleavage, more preferably proteolytic.

Preferably, in the compound according to the invention the pair of molecules C1 and C2 is selected from the group consisting of: 2-aminobenzoic acid (ABZ)/ 5-amino-2-nitrobenzoic (ANB), (ABZ)/pNA, ABZ/ANB-NH₂, ABZ/DNP, ABZ/EDDNP, EDANS/DABCYL,
30 TAM/DANSYL, ABZ/Tyr(3-NO₂), more preferably the pair of C1 and C2 is (ABZ)/pNA or ABZ/ANB-NH₂.

Preferably, the compound according to the invention is a compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ (formula 2) or a compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA (formula 3).

More preferably, the compound according to the invention undergoes hydrolytic cleavage with the generation of the following fragment 1: ABZ-Leu-Pro-Gly-Thr-OH and fragment 2: ANB-NH₂.

The invention further provides an *in vitro* method for detecting enzymatic activity present in a subject's body fluid, in particular deriving from kidney cancer cells, comprising:

a) contacting a sample of body fluid with the compound having formula 1:

10 $X1^1\text{-Leu}^2\text{-Pro}^3\text{-Gly}^4\text{-Thr}^5\text{-X2}^6$ (formula 1),

wherein X1 comprises or consists of molecule C1 and X2 comprises or consists of molecule C2,

wherein the pair of molecules C1 and C2 is a pair of a fluorescence donor and a fluorescence acceptor,

15 and wherein the said compound undergoes enzymatic cleavage into the fragments X1-Leu-Pro-Gly-Thr-OH (fragment 1) and X2 (fragment 2), and

b) detecting a measurable optical signal which is generated upon spatial separation of molecules C1 and C2.

In the method for detecting enzymatic activity according to the invention, enzymatic activity is preferably hydrolytic activity, more preferably proteolytic activity.

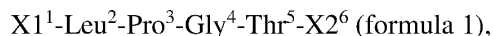
In the method for detecting enzymatic activity according to the invention as the said compound the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ (formula 2) or a compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA (formula 3) is preferably used.

25 In the method for detecting enzymatic activity according to the invention as the said body fluid preferably a urine, more preferably human urine, is used.

The invention also relates to an *in vitro* method for the diagnosis of kidney cancer, wherein the presence or absence of kidney cancer in a subject is detected by measuring enzymatic activity specific to kidney cancer in a body fluid sample from an examined subject, and
30 wherein the absence of the said enzymatic activity indicates the absence of kidney cancer, whereas the presence of the said enzymatic activity indicates the presence of kidney cancer.

In the method for detecting/diagnosis of kidney cancer according to the invention, the detection of enzymatic activity is carried out by the method for detecting enzymatic activity as defined above.

In the method for detecting/diagnosis of kidney cancer according to the invention, the measurement of the said enzymatic activity is performed using the compound having formula 1:



wherein X1 comprises or consists of molecule C1 and X2 comprises or consists of molecule C2,

wherein the pair of molecules C1 and C2 is a pair of a fluorescence donor and a fluorescence acceptor,

and wherein the said compound undergoes enzymatic cleavage into the fragments X1-Leu-Pro-Gly-Thr-OH (fragment 1) and X2 (fragment 2) with the generation of a measurable optical signal upon spatial separation of molecules C1 and C2.

In the method for detecting/diagnosis of kidney cancer according to the invention, the said body fluid sample is preferably incubated with the said compound in a measurement buffer having neutral or alkaline pH, preferably physiological, within the range of sample-to-measurement buffer ratio of from 1:2 to 1:10, preferably 1:5.

In the method for detecting/diagnosis of kidney cancer according to the invention, the said compound is preferably used at a concentration of 0.1-10 mg/mL, in particular 0.25-7.5 mg/mL.

In the method for detecting/diagnosis of kidney cancer according to the invention, as the said compound the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ (formula 2) or a compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA (formula 3) is preferably used.

In the method for detecting/diagnosis of kidney cancer according to the invention, as the said sample a urine sample, more preferably human urine, is preferably used.

In the method for detecting/diagnosis of kidney cancer according to the invention, the measurement of the said enzymatic activity preferably comprises the measurement of absorbance intensity within the range of 300-500 nm, more preferably 380-430 nm, in particular 405 nm, during 40-60 minutes, at a temperature within the range of 25-40° C, more preferably 36-38° C.

The invention further provides a kit comprising any compound according to the invention as defined above and a measurement buffer.

In the kit according to the invention, the said compound is preferably the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ or a compound having formula 3:
5 ABZ-Leu-Pro-Gly-Thr-pNA.

The invention also provides the use of any compound according to the invention as defined above for the detection of enzymatic activity specific to kidney cancer.

The invention also provides the use of any compound according to the invention as defined above for the diagnosis of kidney cancer.

10 Preferably, in such use the diagnosis of kidney cancer comprises the detection of primary kidney cancer, detection of Minimal Residual Disease after surgical resection of kidney cancer and/or detection of kidney cancer recurrence.

Preferably, the compound in the uses according to the invention is the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ or a compound having formula 3:
15 ABZ-Leu-Pro-Gly-Thr-pNA.

The invention further provides any of the compounds according to the invention as defined above for use as a diagnostic marker for the detection of kidney cancer.

Preferably, the compound for use as the diagnostic marker according to the invention is the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ or a compound having
20 formula 3: ABZ-Leu-Pro-Gly-Thr-pNA.

The invention further provides a method for the treatment of kidney cancer, wherein

a) the presence of enzymatic activity specific to kidney cancer is detected by any method as defined above in a body fluid sample from an examined subject, and
b) if the presence of the said enzymatic activity is found in the said sample, a treatment for
25 kidney cancer is applied in the subject.

Preferably, in the method for the treatment according to the invention, after the end of the treatment in accordance with point b), the said enzymatic activity specific to kidney cancer is monitored at predetermined time intervals.

Preferably, in the method for the treatment according to the invention, a urine sample,
30 preferably human urine, is used as the sample.

Preferably, in the method for the treatment according to the invention, the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ or a compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA is used as the said compound.

Detailed description of the invention

It is to be understood that the present invention is defined in the appended claims. The present description illustrates various, non-limiting embodiments and examples of the invention. The present invention is not limited to any particular methodology, protocol or reagents used to carry it out, unless indicated otherwise. The terms as well as scientific and technical expressions as used herein have meanings commonly known and used by persons skilled in the art of the present invention. For the sake of clarity however, the following expressions/terms and acronyms used in the patent shall be understood as follows:

5 A chromogenic compound or a chromogenic molecule means a compound having chromogenic properties. Chromogenic properties mean the ability of a compound to form a coloured product.

A fluorescent compound or a fluorescent molecule means a compound having fluorogenic properties. Fluorogenic properties mean the ability of a compound to form a product emitting fluorescence.

15 NMP stands for *N*-methylpyrrolidone; DMF stands for dimethylformamide; DCM stands for methylene chloride or dichloromethane; pNA stands for 4-nitroaniline or para-nitroaniline; ABZ stands for 2-aminobenzoic acid, ANB-NH₂ stands for amide of 5-amino-2-nitrobenzoic acid; Boc stands for tert-butyloxycarbonyl group; Fmoc stands for 9-fluorenylmethoxycarbonyl group; and TFA stands for trifluoroacetic acid.

20 In the context of this invention, the term kidney cancer shall be understood to mean primary kidney cancer (malignant neoplasm) that develops from tissues located in the kidney. Kidney cancer is most frequently renal cell carcinoma (about 90). The term: kidney cancer as used herein comprises thus all malignant kidney neoplasms which develop from the tissues located in a kidney.

25 In the context of the present invention, the term diagnosis of kidney cancer shall be understood to mean identification of this disease, in particular at its early stage at which other diagnostic methods are not sensitive and/or specific enough. As used herein, the diagnosis of kidney cancer also comprises the detection of Minimal Residual Disease (MRD) after surgical resection of kidney cancer and detection of kidney cancer recurrence after previously completed treatment of kidney cancer.

30 In the context of the present invention, the term: treatment of kidney cancer shall be understood to mean a treatment at an early stage of progression of the disease, which makes

it possible to significantly prolong survival time and improve the quality of life of the diseased subjects.

In the context of the present invention, the term: monitoring shall be understood to mean the diagnosing of Minimal Residual Disease (MRD)) – the presence of a small number of cancer
5 cells that have survived in the organism (during treatment or remission), in the amounts undetectable by means of standard diagnostic methods.

In the context of the present invention, the term: subject shall be understood to mean a human subject or a mammal that is suspected to have kidney cancer, or alternatively a human subject or a mammal belonging to a group with an increased risk of kidney cancer, or a human
10 subject or a mammal after resection of kidney cancer or after a finished treatment of kidney cancer. The subject is preferably a human subject.

The compounds according to the invention have chromogenic properties due to the presence of a chromophore, and fluorogenic properties, i.e. they contain molecules of a fluorescence donor and acceptor. Due to their structure, developed in such a way that an increase in colour
15 is observed in the wavelength range of 380-440 nm, specifically as a result of contact of a tested body fluid sample from an examined subject with kidney cancer, whereas such an effect is not observed in the reaction with a body fluid sample from a healthy subject or a subject with the diagnosis of another type of cancer, these compounds make it possible to detect enzymatic activity specific to kidney cancer, and in particular to diagnose kidney
20 cancer in a specific and sensitive manner, also at an early stage of progression of this cancer. The examined subject is preferably a human subject. The body fluid is preferably urine, more preferably human urine.

In the first aspect of the present invention, a novel chemical compound is provided which compound has formula 1:

25
$$X1^1\text{-Leu}^2\text{-Pro}^3\text{-Gly}^4\text{-Thr}^5\text{-X2}^6 \text{ (formula 1),}$$

wherein X1 is an amino acid derivative or a peptide fragment comprising molecule C1, or X1 consists of such a molecule C1, and X2 is an amino acid derivative or a peptide fragment comprising molecule C2, or X2 consists of such a molecule C2, wherein the pair of molecules C1 and C2 is a pair of a fluorescence donor and a fluorescence acceptor. The
30 superscripts indicate subsequent positions of residues in the compound according to the invention and the sequence of attaching of the residues during synthesis. In accordance with the present invention, in this context chemical formula 1 can be alternatively written without indicating the numbering of residues. The core of all compounds according to the invention

is tetrapeptide with the indicated sequence of 4 amino acids (Leu-Pro-Gly-Thr), which is also presented in the Sequence Listing as sequence no. 1.

The compound according to the invention undergoes enzymatic cleavage into the fragments: X1-Leu-Pro-Gly-Thr-OH (fragment 1) and X2 (fragment 2) with the generation of a measurable optical signal upon spatial separation of molecules C1 and C2. The measurable optical signal is measured by a method for measuring a change in absorbance/fluorescence after the enzymatic cleavage of the compound. Preferably, molecules C1 and C2 are separated from each other by not more than 10 amino acid residues, which ensure efficient quenching of the fluorescence donor by the fluorescence acceptor. It is obvious for the skilled person that the key factor is the distance between the fluorescence donor and acceptor. Therefore, where the amino acid sequence separating molecules C1 and C2 is folded into a twisted or condensed secondary structure, resulting in a proximity of molecules C1 and C2 relative to the primary structure, the distance between molecules C1 and C2 can be greater than 10 amino acid residues.

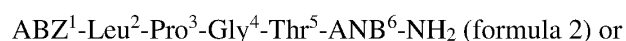
The compound, due to its chromogenic properties and the presence of a reactive site at the position 5 enabling enzymatic (preferably proteolytic) cleavage into smaller fragments, is particularly suitable for use as a diagnostic marker, in particular a specific diagnostic biomarker for kidney cancer, in particular for use in the early diagnosis of kidney cancer.

In preferred embodiments, the compound according to the invention undergoes hydrolytic cleavage, more preferably proteolytic cleavage.

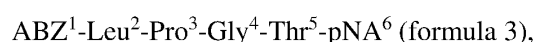
In preferred embodiments the pair of molecules C1 and C2 is selected from a group consisting of: 2-aminobenzoic acid (ABZ)/5-amino-2-nitrobenzoic acid (ANB), (ABZ)/pNA, ABZ/ANB-NH₂, ABZ/DNP, ABZ/EDDNP, EDANS/DABCYL, TAM/DANSYL, ABZ/Tyr(3-NO₂), more preferably the pair of molecules C1 and C2 is ABZ/pNA or ABZ/ANB-NH₂.

In preferred embodiments, the compound according to the invention is:

the compound having formula 2:



the compound having formula 3:



wherein ABZ stands for 2-aminobenzoic acid, ANB-NH₂ stands for amide of 5-amino-2-nitrobenzoic acid, and pNA stands for 4-nitroaniline.

The compound is subject to hydrolytic cleavage with the generation of the following fragment 1: ABZ-Leu-Pro-Gly-Thr-OH and fragment 2: ANB-NH₂ in the case of the compound having formula 2, whereas in the case of the compound having formula 3, with the generation of the following fragment 1: ABZ-Leu-Pro-Gly-Thr-OH and fragment 2: pNA. Thus, fragments 2 are free chromophores.

Spatial separation of molecules C1 and C2 being a result of the enzymatic cleavage of the compound according to the invention causes generation of a measurable optical signal because fluorescence emitted from the fluorescence donor is no longer quenched by the fluorescence acceptor. Such measurable optical signal can be detected preferably at a wavelength of 300-500 nm, more preferably 380-430 nm.

The compounds according to the invention can be obtained by known methods. For example, they can be obtained using a method for obtaining chromogenic peptides which consists in carrying out the process on a solid support in the form of a resin having an Fmoc group, which is removed in the course of the reaction. For example, it can be an amide resin, e.g. Teenage S RAM or RinkAmide, but any other commercially available resin can also be used. The resin used to carry out the process should be properly prepared. The preparation of the resin consists in increasing its volume by repeated washing with hydrophobic solvents. Preferably, a resin with a deposition of 0.23 mmol/g is used. The Fmoc protecting group must be removed from the resin by washing it with a 20% solvent solution.

Then, the known processes for obtaining chromogenic peptides comprise attaching individual components in appropriate time and stoichiometric conditions. The attaching process consists of subsequent steps in which individual elements (amino acid derivatives) are attached, residuals are washed off and protecting groups are successively removed and washed again. This cycle is repeated for each amino acid residue. The obtained peptide is separated from the resin by a reaction under acidic conditions. Then, the solution is separated from the resin in the filtration process and then the peptide is precipitated from the obtained solution by means of a non-polar solvent. The peptide precipitate obtained in this way is centrifuged.

Exemplary, detailed but non-limiting, method for the synthesis of the compounds according to the invention is described below and in Example 1 below.

The method for the synthesis of the compound according to the invention consists in that the process is carried out on a solid support in the form of a resin, preferably having an Fmoc group, wherein before the start of the process the solid support is prepared by increasing its

volume by repeated washing with hydrophobic solvents, preferably dimethylformamide, methylene chloride or N-methylpyrrolidone, and removing the Fmoc protecting group, preferably by washing with 10 – 30 % piperidine solution, in solvents such as dimethylformamide, methylene chloride or N-methylpyrrolidone.

5 Then, the method is carried out in subsequent steps:

a) Deposition of 5-amino-2-nitrobenzoic acid, ANB (or another chromophore suitable for use according to the invention as defined in the claims) on the resin is preceded by washing the solid support with a 3-6% solution of *N*-methyldmorpholine (NMM) in DMF, and then DMF, after which a solution of ANB in DMF is prepared to which TBTU, DMAP
10 and finally diisopropylethylamine (DIPEA) are successively added in the following excess relative to the polymer deposition: ANB/TBTU/DMAP/DIPEA, 3:3:2:6; the mixture prepared in this way is added to the resin and is mixed until homogenous, after which the resin is filtered off under reduced pressure and is washed with solvents such as DMF, DCM and isopropanol, and then the attaching of ANB to the resin is continued using
15 hexafluorophosphate-*O*-(7-azabenzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium (HATU), and then hexafluorophosphate-*O*-(benzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium (HBTU) in excess, and after finishing, the solid support is washed successively with DMF, DCM and isopropanol, and is gently dried;

b) Attachment of the amino acid residue to ANB is carried out by a reaction with the
20 amino acid derivative - Fmoc-Thr(OtBu)-OH, wherein at least fivefold molar excess of the amino acid derivative relative to the resin is dissolved in anhydrous pyridine and is contacted with the resin with deposited ANB, after which the whole is cooled to a temperature not lower than -20°C, and then POCl₃ is added in the ratio 1:1 relative to the amount of the amino acid derivative used and the whole is mixed, after which the mixing process is carried
25 out at room temperature and then at an elevated temperature and when the reaction is completed, the resin is filtered off under reduced pressure, washed with DMF and MeOH and gently dried, after which the obtained intermediate compound is subjected to the process of acylation, successively attaching a fragment of Leu-Pro-Gly;

c) Acylation of the obtained intermediate compound is carried out using an amino acid
30 derivative, preferably Fmoc-Gly-OH, and subsequently Fmoc-Pro-OH, then Fmoc-Leu-OH and, at the final step of the synthesis, Boc-Abz-OH, the acylation being carried out in steps from residue 6 to 1, using diisopropylcarbodiimide as a coupling agent, which is used in excess, and after finishing each step the resin is washed with DMF and preferably is

subjected to the chloranil test (a test for the presence of free amino groups) in which the attachment of the amino acid derivative is monitored;

- d) Removal of the Fmoc protecting group is carried out by washing with 10 - 30% piperidine solution in DMF and subsequent washings with each of the solvents: DMF, isopropanol and methylene chloride;
- e) Separation of the peptide from the resin is carried out using the mixture: TFA:phenol:water:TIPS while maintaining the ratios of 88:5:5:2 v/v/v/v, respectively, the mixture is stirred for the period of at least one hour, preferably three hours, and the obtained precipitate filtered off under reduced pressure, after which it is washed with diethyl ether and the obtained peptide is centrifuged;
- f) Preparation of the finished product is carried out by dissolving the peptide in water by means of ultrasound and then it is subjected to lyophilisation.

The second aspect of the present invention provides an *in vitro* method for detecting enzymatic, preferably proteolytic, activity, present in a subject's body fluid, in particular deriving from kidney cancer cells, the method comprising a) contacting the body fluid sample with the compound according to the invention and b) detecting a measurable optical signal which is generated upon spatial separation of molecules C1 and C2 present in the compound according to the invention. In a preferred embodiment of this aspect, the examined subject in this case is a human subject. In another preferred embodiment of this aspect, the body fluid is urine, in particular human urine.

In the preferred embodiment of this aspect, the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ or a compound having formula 3: ABZ-Leu-Pro-Gly-Thr -pNA is used.

The third aspect of the present invention provides an *in vitro* method for the diagnosis of kidney cancer in which the presence or absence of kidney cancer in a subject is detected by measuring enzymatic activity specific to kidney cancer in a body fluid sample from the examined subject, wherein the absence of the said enzymatic activity indicates the absence of kidney cancer whereas the presence of the said enzymatic activity indicates the presence of kidney cancer. Such detection of enzymatic activity is preferably carried out using methods for detecting enzymatic activity as discussed above. In the preferred embodiment of this aspect the subject is a human subject. In the preferred embodiment of this aspect the body fluid is urine, in particular human urine. In the preferred embodiment of this aspect the enzymatic activity specific to kidney cancer is proteolytic activity. In the preferred

embodiment of this aspect the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ or a compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA is used.

Moreover, in the preferred embodiment of this aspect, the measurement of the said enzymatic activity in the methods according to the invention comprises the measurement of absorbance intensity within the range of 300-500 nm, preferably 380-430 nm, in particular 405 nm, during 40-60 minutes, at a temperature within the range of 25-40° C, preferably 36-38° C. This enables obtaining a maximally intensive measurable optical signal resulting from an increase in absorbance or fluorescence.

Furthermore, in the preferred embodiments of the said methods according to the invention the measurement of the said enzymatic activity is performed using the compound according to the invention in the range of concentrations of 0.1 - 10 mg/mL, more preferably at the concentration of 1 mg/mL. In the preferred embodiments of the said methods according to the invention the tested sample is incubated with the compound according to the invention in a measurement buffer having a neutral or alkaline pH, preferably physiological, with a body fluid sample, preferably human urine, with the sample (e.g. of urine) to measurement buffer ratio ranging from 1:2 to 1:10, preferably 1:5. The sample is preferably taken from a subject with a referral for the diagnosis of kidney cancer. Preferably, absorbance intensity is measured within the range of 300-500 nm, preferably 380-430 nm, in particular 405 nm, during 40-60 minutes, at a temperature within the range of 25-40° C, preferably 36-38° C. In the aforementioned conditions, a maximally intensive measurable optical signal is obtained resulting from an increase in absorbance or fluorescence.

In the fourth aspect, the present invention provides a kit comprising any compound according to the invention and a measurement buffer. Measurement buffers are known in this art and a buffer suitable for use in the kit according to the invention is, for example, but without limitation, the Tris-HCl buffer. In the preferred embodiment, in the kit according to the invention the said compound is the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ or a compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA.

In the fifth aspect, the present invention provides use of the compound according to the invention for the detection of enzymatic activity specific to kidney cancer. In the sixth aspect, the present invention provides use of the compound according to the invention for the diagnosis of kidney cancer. Preferably, the diagnosis of kidney cancer comprises, according

to the invention, the detection of primary kidney cancer, detection of Minimal Residual Disease after surgical resection of kidney cancer and/or detection of kidney cancer recurrence after previously completed kidney cancer treatment.

5 In the seventh aspect, the present invention provides the compound according to the invention for use as a diagnostic marker for the detection of kidney cancer. In the preferred embodiments of this aspect the said compound is the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ or a compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA.

10 In the eighth aspect, the present invention provides a method for the treatment of kidney cancer wherein

a) the presence of enzymatic activity specific to kidney cancer is detected by any method according to the invention as defined above, in a body fluid sample from the examined subject, and

15 b) when the said enzymatic activity is found to be present in the said sample, a treatment of kidney cancer is applied in the subject.

In the preferred embodiment of the method for the treatment, after completing the treatment according to point b), the said enzymatic activity specific to kidney cancer is monitored at predetermined time intervals as known in the art, e.g. every week, every several weeks, every month, every several months, every year or at any other intervals considered to be
20 appropriate by the skilled person, in order to detect Minimal Residual Disease after surgical resection of kidney cancer or recurrence. Furthermore, in the preferred embodiment of the method, a urine sample, preferably human urine, is used as the test sample. In the preferred embodiment of the method for the treatment the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ (formula 2) or a compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA (formula 3) is used as the said compound.
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The advantages of the present invention consist in providing a novel chemical compound having properties that make it suitable for use for specific and sensitive detection of enzymatic activity specific to kidney cancer, for use as a diagnostic biomarker for the detection of kidney cancer, for use in a fast, non-invasive diagnosis of kidney cancer, while
30 enabling the detection of kidney cancer at an early stage of its progression. Another advantage is that the diagnostic methods according to the invention can be successfully used in screening tests. This enables full diagnosis at an early stage of cancer progression and consequently a more effective treatment. Early diagnosis enables surgical treatment, which

significantly prolongs patient's survival time. It is also important when monitoring the effectiveness of the applied surgical treatment and/or chemotherapy of kidney cancer since it is possible to detect Minimal Residual Disease or recurrence, if any.

The present invention will now be illustrated in the following figures and the following examples, which however are not intended to limit, in any manner, the scope of the invention as defined in the claims.

Brief description of the figures

Fig. 1 shows the results of chromatographic analysis of the substrate cleavage, i.e. ABZ-Leu-Pro-Gly-Thr-ANB-NH₂, in a sample of urine from a subject with kidney cancer.

Fig. 2 shows the rate of hydrolysis of the substrate - ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ – in the samples of urine from subjects with diagnosed kidney cancer (samples N1 - N20) and urine taken from healthy subjects (samples 21 – 40). Arabic numerals indicate the number of the selected urine sample.

Fig. 3 shows the selectivity of hydrolysis of the substrate - ABZ¹-Leu²-Pro³-Gly⁴-Thr⁵-ANB⁶-NH₂ (i.e. compound of formula 2) in urine samples from subjects with diagnosed kidney cancer (sample 1) and urine samples taken from subjects with the diagnosis of another neoplastic disease (samples 2 - 9). Arabic numerals indicate the numbers of given cancer types. The samples tested for each type of cancer were derived from 20 different patients for each of the tested cancers. The results are mean values for given cancer types.

The results show selectivity of substrate cleavage in the case of urine from patients with kidney cancer as compared to urine samples from patients suffering from other neoplasms.

Fig. 4 shows the dependence of the hydrolysis level of the substrate - ABZ¹-Leu²-Pro³-Gly⁴-Thr⁵-ANB⁶-NH₂, on pH conditions.

Examples

The invention is illustrated by the following non-limiting examples. Unless indicated otherwise, the examples below use known and/or commercially available devices, methods, reaction conditions, reactants and sets, which are commonly used in the field to which the present invention belongs and which are recommended by the producers of respective reactants and kits.

Example 1: Synthesis of the compound according to the invention

This example presents the synthesis of one representative compound according to the present invention, namely the compound: ABZ¹-Leu²-Pro³-Gly⁴-Thr⁵-ANB⁶-NH₂. The remaining peptides according to the invention can be synthesized in an analogous way. The superscripts

indicate subsequent positions of residues in the compound according to the invention and the sequence of attachment of the residues during synthesis. The compounds according to the invention can be alternatively represented by an analogous formula without the indication of residue positions, which does not change the sequence of residues in the compounds according to the invention, as it remains unchanged.

1. Obtaining a chromogenic peptide

a) The first step of the synthesis was to obtain the chromogenic peptide, which was obtained by solid phase synthesis, on a solid support, using Fmoc/tBu chemistry, i.e. with the use of protection.

10 A compound having the sequence $ABZ^1\text{-Leu}^2\text{-Pro}^3\text{-Gly}^4\text{-Thr}^5\text{-ANB}^6\text{-NH}_2$, wherein ABZ is 2-aminobenzoic acid and ANB-NH₂ is amide of 5-amino-2-benzoic acid and ANB is 5-amino-2-benzoic acid, was obtained in the process of solid phase chemical synthesis using the following amino acid derivatives:

Boc-ABZ, Fmoc-Leu, Fmoc-Pro, Fmoc-Gly, Fmoc-Thr(OtBu).

15 The synthesis of the compound according to the invention, i.e. a diagnostic marker for the detection of kidney cancer, was carried out on a solid support enabling the conversion of 5-amino-2-benzoic acid into ANB-NH₂ amide, namely amide resin TentaGel S RAM from RAPP Polymere (Germany), with a deposition of 0.23 mmol/g. However, it is possible to use any other amide resin, e.g. Rink Amide (Germany).

20 The synthesis of the compound was carried out manually using a laboratory shaker. In most of the steps a 25 mL sintered syringe for solid phase synthesis was used as a reactor.

All the obtained final compounds contained 2-aminobenzoic acid (ABZ) at the position 1 of their sequence, i.e. at the N-terminus, and a 5-amino-2-nitrobenzoic acid (ANB) molecule at the position 6, i.e. at the C-terminus. ABZ acts here as a fluorescence donor, while ANB acts as a fluorescence quencher and simultaneously a chromophore. The peptides contained at least and preferably one reactive site in their sequence, located between the amino acid residues Thr-ANB-NH₂, i.e. at the position 5 of the compound. The synthesis consisting in attaching amino acid derivatives is carried out from residue 6 to 1, i.e. from the C- to N-terminus.

30 b) Deposition of ANB on TentaGel S RAM resin:

The synthesis of peptides was performed on TentaGel S RAM resin from Rapp Polymere with a deposition of 0.23 mmol/g. In the first step, the resin was prepared, including loosening it by the wash cycle. Subsequently, the protection of the Fmoc amino group was

removed from the solid support with the 20% solution of piperidine in NMP. Then, the solvent washing cycle was carried out. In order to confirm the presence of free amino groups, a chloranil test was performed.

Solvent wash cycle:

- 5 DMF 1 x 10 minutes; IsOH 1 x 10 minutes; DCM 1 x 10 minutes.

Removal of Fmoc protection:

DMF 1 x 5 minutes; 20% piperidine in NMP 1 x 3 minutes; 20% piperidine in NMP 1 x 8 minutes.

Solvent wash cycle:

- 10 DMF 3 x 2 minutes; IsOH 3 x 2 minutes; DCM 3 x 2 minutes.

c) Chloranil test:

The chloranil test consisted in transferring, by means of a spatula, several grains of resin from the reactor – a syringe, into a glass ampule, to which subsequently 100 µL of saturated solution of p-chloranil in toluene and 50 µL of fresh acetaldehyde were added. After 10

- 15 minutes, the control of grains colour was carried out.

At that stage, after performing the test, a green colour of the grains was obtained, which evidenced the presence of free amino groups. After confirming the removal of 9-fluorenylmethoxycarbonyl protection from the resin, it was possible to proceed to the next step, the attachment of the ANB derivative (5-amino-2-nitrobenzoic acid).

- 20 d) Deposition of 5-amino-2-nitrobenzoic acid on solid support:

The first step in the synthesis of the peptide library – a mixture of peptides was deposition of ANB on 1 g of resin. Before attaching the chromophore, the resin used for the reaction was washed with the following solvents: DMF, DCM and again DMF, after which the Fmoc-protection was removed from the functional group of the solid support. One cycle of

- 25 removing the Fmoc- protection comprised the following steps:

Removal of Fmoc- protection:

20% piperidine in NMP 1 x 3 minutes; 20% piperidine in NMP 1 x 8 minutes.

e) Washing:

DMF 3 x 2 minutes; IsOH 3 x 2 minutes; DCM 3 x 2 minutes.

- 30 f) Chloranil test:

The resin with a free amino group was washed with 5% solution of *N*-methylmorpholine (NMM) in DMF, and then DMF. The procedure of removing the Fmoc- protection and the wash cycle were carried out in a Merrifield vessel. In a separate flask, ANB was dissolved

- in DMF, and TBTU, DMAP and finally diisopropylethylamine (DIPEA) were subsequently added in the following excess relative to polymer deposition: ANB/TBTU/DMAP/DIPEA, 3:3:2:6 v/v/v/v. The mixture prepared in this way was added to the resin and was stirred for 3 hours. The resin was filtered off under reduced pressure, washed with DMF, DCM and isopropanol, and the entire acylation procedure was repeated twice. To carry out subsequent reactions of attaching ANB to the resin, hexafluorophosphate-*O*-(7-azabenzotriazol-1-yl)-N,N,N',N'-tetramethyluronium (HATU) and then hexafluorophosphate-*O*-(benzotriazol-1-yl)-N,N,N',N'-tetramethyluronium (HATU) were used. In the last step, the resin was washed successively with DMF, DCM and isopropanol, and was air dried.
- g) Attachment of the C-terminal amino acid residue (Fmoc-Thr(OtBu)-OH) to ANB: The corresponding amino acid derivative (9-fold molar excess relative to resin deposition) was dissolved in pyridine and was transferred to the flask containing the resin with deposited ANB. The whole was cooled until the temperature of -15°C was reached (ice bath: 1 part by weight of NH₄Cl, 1 part by weight of NaNO₃, 1 part by weight of ice). After the desired temperature was reached, POCl₃ was added (in 1:1 ratio to the amount of amino acid derivative used) and the whole was stirred on a magnetic stirrer: for 20 minutes at -15°C, 30 minutes at room temperature and 6 hours at 40°C (oil bath). When the reaction was completed, the resin was filtered off under reduced pressure, washed with DMF and MeOH and left to dry. In the next stage, the residue was attached in P2 position (Fmoc-Gly).
- Every attachment of amino acid residue was preceded by washing the resin with DMF for 5 minutes. Diisopropylcarbodiimide was used as a coupling agent in subsequent attachments. The procedure was repeated twice.
- After each acylation, a resin wash cycle was started and then the chloranil test was performed in order to monitor the attachment of the amino acid derivative to free amino acid groups of the resin.
- Solvent wash cycle:
DMF 3 x 2 minutes; IsOH 3 x 2 minutes; DCM 3 x 2 minutes.
- Chloranil test:
As a result of the performed tests, after the first two coupling procedures, the colour of the grains was first green and then grey, so it was necessary to perform another acylation, as a result of which the resin grains tested by the chloranil test were colourless. This evidenced the attachment of ANB to the TentaGel S RAM resin, and thus it was possible to proceed to the next step of peptide synthesis.

h) Attachment of subsequent protected amino acid residues:

The resin together with the attached fragment ANB-Thr(OtBu), located in the reactor, was washed with DMF, which was followed by deprotection of the Fmoc from the amino group in order to attach the protected amino acid derivative Gly.

5 Removal of Fmoc protection:

DMF 1 x 5 minutes; 20% piperidine in NMP 1 x 3 minutes; 20% piperidine in NMP 1 x 8 minutes.

Solvent wash cycle:

DMF 3 x 2 minutes; IsOH 3 x 2 minutes; DCM 3 x 2 minutes.

10 Chloranil test:

The chloranil test produced a positive result, which was evidenced by the green colour of the resin grains. Thus, it was possible to proceed to the next step – attachment of the amino acid residue Fmoc-Pro-OH.

Attachment of the amino acid derivative

15 The process of coupling was preceded by washing the resin with DMF. The composition of the coupling mixture remained unchanged when attaching the protected serine residue.

After the end of each acylation, a solvent wash cycle was performed according to the specified procedure and then the chloranil test for the presence of free amino acid groups in the solution was performed.

20 Solvent wash cycle

DMF 3 x 2 minutes; IsOH 3 x 2 minutes; DCM 3 x 2 minutes.

Chloranil test:

During the test performed after the second acylation the resin grains were colourless, and thus it was possible to proceed to the next step of the synthesis i.e. the introduction of another

25 protected amino acid derivative – Fmoc-Leu and 2-aminobenzoic acid molecule. The coupling processes were performed according to the procedure discussed earlier.

Tests carried out after attaching the above-mentioned residues showed positive results - the resin grains were colourless.

2. Removal of the peptide from the solid support

30 After the synthesis, the amide of ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ peptide was removed from the solid support and simultaneously the side protection was removed using the mixture: TFA:phenol:water:TIPS (88:5:5:2, v/v/v/v) in a round-bottom flask on a magnetic stirrer.

After 3 hours, the content of the flask was filtered off under reduced pressure in sintered (Schott) funnels and washed with diethyl ether. The obtained sediment was centrifuged on a SIGMA 2K30 centrifuge (Laboratory Centrifuges) for 20 minutes. The precipitate obtained after centrifugation was dissolved in water by means of ultrasound and then it was subjected to lyophilisation. The remaining compounds according to the invention can be obtained in an analogous way.

The identity/characteristics of the novel compound according to the invention were confirmed using the HPLC analysis. The conditions of the HPLC analysis were as follows: RP Bio Wide Pore Supelco C8 column, 250 mm 4 mm, a phase system A 0.1% TFA in water, B: 80% acetonitrile in A), flow rate 1 mL/min, UV detection at 226 nm.

The analysis carried out confirmed that the compound according to the invention was obtained.

Example 2: Testing the properties of the peptide according to the invention as a cancer marker

The activity of the novel compounds according to the invention was studied in a group of 20 subjects diagnosed with kidney cancer using the representative compound according to the invention. The mechanism of action of the compounds according to the invention, including the representative compound having formula 2, consists in specific enzymatic cleavage, more specifically enzymatic hydrolysis, at the position that leads to the release of free molecules of respective chromophores: ANB-NH₂ (amide of 5-amino-2-nitrobenzoic acid) in the case of the compound having formula 2 or pNA (para-nitroanilide) in the case of the compound having formula 3, which exhibit absorbance at a wavelength of 320-480 nm, especially 380-430 nm, in particular 405 nm. The remaining compounds according to the invention are characterized by the analogous mechanism of action. For this purpose, the representative compound according to the invention, ABZ¹-Leu²-Pro³-Gly⁴-Thr⁵-ANB⁶-NH₂, was dissolved in dimethyl sulfoxide (at a concentration of 0.5 mg/mL) and then 50 µL of the solution was mixed with 120 µL of a buffer (200 mM Tris-HCl, pH 8.0) and 80 µL of urine from a subject suffering from kidney cancer. The measurement was performed on a 96-well plate designed for measuring absorbance and each sample was analysed in triplicate at the temperature of 37 °C. The duration of the measurement was 60 minutes. During the measurement, the wavelength characteristic for the chromophore (ANB-NH₂) being released was monitored at the wavelength 405 nm (range 380-430 nm).

A shown in Fig. 1, the RP HPLC analysis of a randomly selected system comprising urine taken from a person diagnosed with kidney cancer indicates that the compound according to the invention cleaves into the peptide fragment ABZ-Leu-Pro-Gly-Thr-OH and the chromophore group of the compound (ANB-NH₂).

- 5 The measurement showed that the colour intensity of the solution increased with time in all urine samples taken from persons diagnosed with kidney cancer. The observed magnitude of increase in absorbance in time is different for each of the examined samples. A different effect was obtained for 20 samples from healthy subjects (samples 21-40) since no increase in absorbance within the diagnostic range was observed in any of the 20 tested urine samples.
- 10 The tests carried out show that all samples N1-N20 from persons suffering from kidney cancer underwent cleavage, but in the case of samples N3 and N6, cleavage of the substrate, i.e. ABZ-Leu-Pro-Gly-Thr-ANB-NH₂, proceeded less efficiently than in the case of samples N5 or N20 (Fig. 2, Table 1). Such a result may be due to the difference in the activity as well as the amount of enzymes responsible for the enzymatic cleavage (proteolysis). Furthermore,
- 15 the results presented in Table 1 below indicate that incubating the substrate solution – the compound according to the invention – with urine samples taken from healthy persons (without a cancer diagnosis, marked with Arabic numerals sequentially from 21 to 40) does not lead to an increase in absorbance and thus hydrolysis of the tested compound does not take place. The result indicates the absence of proteolytic enzymes specific/characteristic of
- 20 kidney cancer.

Table 1. Results of absorbance analysis

N1	0,027000	0,019000	0,023000
N2	0,021000	0,014000	0,019000
N3	0,004400	0,002100	0,003600
N4	0,032000	0,029000	0,030000
N5	0,034000	0,032000	0,029000
N6	0,003300	0,002700	0,003000
N7	0,020000	0,021000	0,018000
N8	0,002000	0,004400	0,003500
N9	0,031000	0,030000	0,035000

N10	0,002600	0,003700	0,003100
N11	0,012000	0,010000	0,015000
N12	0,009800	0,011100	0,012000
N13	0,004000	0,005100	0,004500
N14	0,003600	0,003300	0,003200
N15	0,004300	0,003700	0,004000
N16	0,003900	0,003700	0,003500
N17	0,013000	0,013500	0,013600
N18	0,004000	0,003200	0,003800
N19	0,003800	0,004000	0,003700
N20	0,031000	0,031000	0,026000
21	0,000000	0,000000	0,000000
22	0,000000	0,000000	0,000000
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35	0,000000	0,000000	0,000000
36	0,000000	0,000000	0,000000
37	0,000000	0,000000	0,000000
38	0,000000	0,000000	0,000000
39	0,000000	0,000000	0,000000
40	0,000000	0,000000	0,000000

Furthermore, the dependence of the cleavage selectivity of the substrate, i.e. the compound according to the invention, on the cancer type being examined, was studied. The results of the performed tests are presented in Fig. 3 and they indicate that the tested substrate, i.e. ABZ¹-Leu²-Pro³-Gly⁴-Thr⁵-ANB⁶-NH₂, incubated with the samples taken from patients with diagnosed following cancers: testicular cancer, prostate cancer, pancreatic cancer, biliary duct cancer, lung cancer, ovary cancer, rectum cancer, is not subject to cleavage and does not cause an increase in absorbance within the specified range. The samples tested were, in each case, a mixture of 20 samples derived from each of the cancers studied. This indicates cleavage selectivity of the compounds according to the invention, which makes them suitable for the specific detection of enzymatic activity specific to kidney cancer and specific diagnosis of kidney cancer.

Table 2 below presents the obtained measurements results for each sample in triplicate.

Table 2. Results of analysis of selectivity of the cleavage

Kidney 1	0,01416	0,01476	0,01874
Stomach 2	0	0	0
Lung 3	0	0	0
Intestine 4	0	0	0
Prostate 5	0	0	0
Bladder 6	0	0	0
Pancreas 7	0	0	0
Ovary 8	0	0	0
Liver 9	0	0	0

15

Furthermore, measurements concerning the dependence of the proteolytic activity of the representative compound according to the invention on the reaction pH were performed. The experiment has shown that the studied material has at least one enzyme exhibiting maximum activity at alkaline pH (Fig. 4).

20 The analyses carried out confirmed suitability of the compounds according to the invention for the sensitive and specific detection of enzymatic activity specific to kidney cancer and, by the same, their suitability also for the specific diagnosis of kidney cancer, and as a diagnostic marker for kidney cancer. The mechanism of action of the compounds according to the invention consists in their specific enzymatic cleavage at the position that leads to the

release of free chromophore molecules, which generates a measurable optical signal that can be used for diagnostic purposes, in particular in the diagnosis of kidney cancer according to the present invention.

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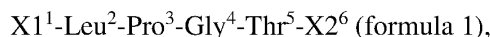
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Claims

1. A compound having formula 1:



wherein X1 comprises or consists of molecule C1 and X2 comprises or consists of molecule C2,

wherein the pair of molecules C1 and C2 is a pair of a fluorescence donor and a fluorescence acceptor,

and wherein the compound undergoes enzymatic cleavage into the fragments X1-Leu-Pro-Gly-Thr-OH (fragment 1) and X2 (fragment 2) with a generation of a measurable optical signal upon spatial separation of molecules C1 and C2.

2. The compound according to claim 1, which compound undergoes hydrolytic cleavage, preferably proteolytic.

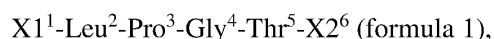
3. The compound according to claim 1 or 2, in which compound the pair of molecules C1 and C2 is selected from the group consisting of: 2-aminobeznoic acid (ABZ)/5-amino-2-nitrobenzoic acid (ANB), (ABZ)/pNA, ABZ/ANB-NH₂, ABZ/DNP, ABZ/EDDNP, EDANS/DABCYL, TAM/DANSYL, ABZ/Tyr(3-NO₂), preferably the pair of C1 and C2 is ABZ/pNA or ABZ/ANB-NH₂.

4. The compound according to any one of claims 1 to 3, which compound is the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ (formula 2) or a compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA (formula 3).

5. The compound according to claim 4, which compound undergoes hydrolytic cleavage with the generation of the following fragment 1: ABZ-Leu-Pro-Gly-Thr-OH and fragment 2: ANB-NH₂.

6. An *in vitro* method for detecting enzymatic activity present in a subject's body fluid, in particular deriving from kidney cancer cells, comprising:

- a) contacting the body fluid sample with the compound having formula 1:



wherein X1 comprises or consists of molecule C1 and X2 comprises or consists of molecule C2,

wherein the pair of molecules C1 and C2 is a pair of a fluorescence donor and a fluorescence acceptor,

and wherein the compound undergoes enzymatic cleavage into the fragments X1-Leu-Pro-Gly-Thr-OH (fragment 1) and X2 (fragment 2), and

b) detecting a measurable optical signal which is generated upon spatial separation of molecules C1 and C2.

7. The *in vitro* method according to claim 6, wherein the enzymatic activity is hydrolytic activity, preferably proteolytic activity.

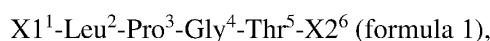
8. The *in vitro* method according to claim 6 or 7, wherein as the said compound the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ (formula 2) or the compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA (formula 3) is used.

9. The method according to any one of claims 6 to 8, wherein as the said body fluid urine, preferably human urine, is used.

10. An *in vitro* method for diagnosis of kidney cancer, wherein the presence or absence of kidney cancer in a subject is detected by measuring enzymatic activity specific to kidney cancer in a body fluid sample from the examined subject, and wherein the absence of the said enzymatic activity indicates the absence of kidney cancer whereas the presence of the said enzymatic activity indicates the presence of kidney cancer.

11. The method according to claim 10, wherein the detection of enzymatic activity is carried out by the method as defined in any one of claims 6 to 9.

12. The method according to claim 10 or 11, wherein the measurement of the said enzymatic activity is performed using the compound having formula 1:



wherein X1 comprises or consists of molecule C1 and X2 comprises or consists of molecule C2,

wherein the pair of molecules C1 and C2 is a pair of fluorescence donor and fluorescence acceptor,

and wherein the said compound undergoes enzymatic cleavage into the fragments X1-Leu-Pro-Gly-Thr-OH (fragment 1) and X2 (fragment 2) with the generation of a measurable optical signal upon spatial separation of molecules C1 and C2.

13. The method according to any one of claims 10 to 12, wherein the said body fluid sample is incubated with the said compound in a measurement buffer having neutral or alkaline pH, preferably physiological, within the range of sample-to-measurement buffer ratio of 1:2 to 1:10, preferably 1:5.

14. The method according to any one of claims 10 to 13, wherein the said compound is used at a concentration of 0.1-10 mg/mL, in particular 0.25-7.5 mg/mL.

15. The method according to claims 10 to 14, wherein as the said compound the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ (formula 2) or the compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA (formula 3) is used.

16. The method according to any one of claims 10 to 15, wherein as the said sample a urine sample, preferably human urine, is used.

17. The method according to any one of claims 10 to 16, wherein the measurement of the said enzymatic activity comprises the measurement of absorbance intensity in the range of 300-500 nm, preferably 380-430 nm, in particular 405 nm, during 40-60 minutes, at a temperature within the range of 25-40° C, preferably 36-38° C.

18. A kit comprising the compound as defined in any one of claims 1 to 5 and a measurement buffer.

19. The kit according to claim 18, wherein the compound is the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ (formula 2) or the compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA (formula 3).

20. Use of the compound as defined in any one of claims 1 to 5 for the detection of enzymatic activity specific to kidney cancer.

21. Use of the compound as defined in any one of claims 1 to 5 for the diagnosis of kidney cancer.

22. Use according to claim 21, wherein the diagnosis of kidney cancer comprises the detection of primary kidney cancer, detection of Minimal Residual Disease after surgical resection of kidney cancer and/or detection of kidney cancer recurrence.

23. Use according to any one of claims 21 to 22, wherein the compound is the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ (formula 2) or the compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA (formula 3).

24. The compound as defined in any one of claims 1 to 5 for use as a diagnostic marker for the detection of kidney cancer.

25. The compound for use according to claim 24, which compound is the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ (formula 2) or the compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA (formula 3).

26. A method for the treatment of kidney cancer, wherein

a) the presence of enzymatic activity specific to kidney cancer is detected by the method as defined in any one of claims 6 to 9 in a body fluid sample from the examined subject and,

b) if the presence of the said enzymatic activity is found in the said sample, a treatment of kidney cancer is applied in the subject.

27. The method according to claim 26, wherein after the end of the treatment in accordance with point b), the said enzymatic activity specific to kidney cancer is monitored at predetermined time intervals.

28. The method according to claim 26 or 27, characterized in that as the sample a urine sample, preferably human urine, is used.

29. The method according to any one of claims 26 to 28, wherein as the said compound the compound having formula 2: ABZ-Leu-Pro-Gly-Thr-ANB-NH₂ (formula 2) or the compound having formula 3: ABZ-Leu-Pro-Gly-Thr-pNA (formula 3) is used.