

HUMAN MONOCLONAL ANTIBODIES TO PROGRAMMED DEATH LIGAND 1 (PD-L1)

Cross-Reference to Related Applications

This application claims the benefit of U.S. Provisional Patent Application No.
5 60/696,426, filed July 1, 2005, which is incorporated herein by reference in its entirety.

Background

Programmed death 1 (PD-1) is a member of the CD28 family of receptors, which includes CD28, CTLA-4, ICOS, PD-1, and BTLA. The initial members of the family, CD28 and ICOS, were discovered by functional effect on augmenting T cell proliferation
10 following the addition of monoclonal antibodies (Hutloff *et al.* (1999) *Nature* 397:263-266; Hansen *et al.* (1980) *Immunogenetics* 10:247-260). Two cell surface glycoprotein ligands for PD-1 have been identified, PD-L1 and PD-L2, and have been shown to downregulate T cell activation and cytokine secretion upon binding to PD-1 (Freeman *et al.* (2000) *J Exp Med* 192:1027-34; Latchman *et al.* (2001) *Nat Immunol* 2:261-8; Carter
15 *et al.* (2002) *Eur J Immunol* 32:634-43; Ohigashi *et al.* (2005) *Clin Cancer Res* 11:2947-53). Both PD-L1 (B7-H1) and PD-L2 (B7-DC) are B7 homologs that bind to PD-1, but do not bind to other CD28 family members (Blank *et al.* (2004). Expression of PD-L1 on the cell surface has also been shown to be upregulated through IFN- γ stimulation.

PD-L1 expression has been found in several murine and human cancers, including
20 human lung, ovarian and colon carcinoma and various myelomas (Iwai *et al.* (2002) *PNAS* 99:12293-7; Ohigashi *et al.* (2005) *Clin Cancer Res* 11:2947-53). PD-L1 has been suggested to play a role in tumor immunity by increasing apoptosis of antigen-specific T-cell clones (Dong *et al.* (2002) *Nat Med* 8:793-800). It has also been suggested that PD-L1 might be involved in intestinal mucosal inflammation and inhibition of PD-L1
25 suppresses wasting disease associated with colitis (Kanai *et al.* (2003) *J Immunol* 171:4156-63).

Summary

The present invention provides isolated monoclonal antibodies, in particular human monoclonal antibodies that bind to PD-L1 and exhibit numerous desirable

properties. These properties include high affinity binding to human PD-L1. Still further, antibodies of the invention have been shown to increase T-cell proliferation, IFN- γ secretion, and IL-2 secretion in a mixed lymphocyte reaction.

5 In one aspect, the invention pertains to an isolated monoclonal antibody, or an antigen-binding portion thereof, wherein the antibody exhibits at least one of the following properties:

- (a) binds to human PD-L1 with a K_D of 1×10^{-7} M or less;
- (b) increases T-cell proliferation in a mixed lymphocyte reaction (MLR) assay;
- (c) increases interferon- γ production in an MLR assay;
- 10 (d) increases IL-2 secretion in an MLR assay;
- (e) stimulates antibody responses; or
- (f) reverses the effect of T regulatory cells on T cell effector cells and /or dendritic cells.

Preferably the antibody is a human antibody, although in alternative embodiments the
15 antibody can be, for example, a murine antibody, a chimeric antibody or humanized antibody.

In particular embodiments, the antibody binds to human PD-L1 with a K_D of 5×10^{-8} M or less, binds to human PD-L1 with a K_D of 1×10^{-8} M or less, binds to human PD-L1 with a K_D of 5×10^{-9} M or less, binds to human PD-L1 with a K_D of 5×10^{-9} M or
20 less, or binds to human PD-L1 with a K_D of between 1×10^{-8} M and 1×10^{-10} M.

In another embodiment, the invention provides an isolated monoclonal antibody, or antigen binding portion thereof, wherein the antibody cross-competes for binding to PD-L1 with a reference antibody comprising:

- (a) the human heavy chain variable region comprises an amino acid sequence
25 selected from the group consisting of SEQ ID NOs:1, 2, 3, 4, 5, 6, 7, 8, 9, and 10;
and
- (b) the human light chain variable region comprises an amino acid sequence
selected from the group consisting of SEQ ID NOs:11, 12, 13, 14, 15, 16, 17, 18,
19, and 20.

30 In various embodiments, the reference antibody comprises:

- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:1; and

- (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:11;
- or the reference antibody comprises:
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:2; and
- 5 (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:12;
- or the reference antibody comprises:
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:3; and
- 10 (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:13;
- or the reference antibody comprises:
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:4; and
- 15 (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:14;
- or the reference antibody comprises:
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:5; and
- 20 (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:15;
- or the reference antibody comprises:
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:6; and
- 25 (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:16;
- or the reference antibody comprises:
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:7; and
- 30 (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:17;
- or the reference antibody comprises:

- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:8; and
 - (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:18;
- 5 or the reference antibody comprises:
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:9; and
 - (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:19;
- 10 or the reference antibody comprises:
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:10; and
 - (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:20.
- 15 In another aspect, the invention pertains to an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising a heavy chain variable region that is the product of or derived from a human V_H 1-18 gene, wherein the antibody specifically binds PD-L1. The invention further provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising a heavy chain variable region that is the
- 20 product of or derived from a human V_H 1-69 gene, wherein the antibody specifically binds PD-L1. The invention further provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising a heavy chain variable region that is the product of or derived from a human V_H 1-3 gene, wherein the antibody specifically binds PD-L1. The invention further provides an isolated monoclonal antibody, or an antigen-
- 25 binding portion thereof, comprising a heavy chain variable region that is the product of or derived from a human V_H 3-9 gene, wherein the antibody specifically binds PD-L1. The invention further provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising a light chain variable region that is the product of or derived from a human V_K L6 gene, wherein the antibody specifically binds PD-L1. The invention
- 30 further provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising a light chain variable region that is the product of or derived from a human V_K L15 gene, wherein the antibody specifically binds PD-L1. The invention further provides an isolated monoclonal antibody, or an antigen-binding portion thereof,

comprising a light chain variable region that is the product of or derived from a human V_K A27 gene, wherein the antibody specifically binds PD-L1. The invention further provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising a light chain variable region that is the product of or derived from a human
5 V_K L18 gene, wherein the antibody specifically binds PD-L1.

In a particularly preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising:

- (a) a heavy chain variable region of a human V_H 1-18 gene; and
 - (b) a light chain variable region of a human V_K L6 gene;
- 10 wherein the antibody specifically binds to PD-L1.

In another preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising:

- (a) a heavy chain variable region of a human V_H 1-69 gene; and
 - (b) a light chain variable region of a human V_K L6 gene;
- 15 wherein the antibody specifically binds to PD-L1.

In another preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising:

- (a) a heavy chain variable region of a human V_H 1-3 gene; and
 - (b) a light chain variable region of a human V_K L15 gene;
- 20 wherein the antibody specifically binds to PD-L1.

In another preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising:

- (a) a heavy chain variable region of a human V_H 1-69 gene; and
 - (b) a light chain variable region of a human V_K A27 gene;
- 25 wherein the antibody specifically binds to PD-L1.

In another preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising:

- (a) a heavy chain variable region of a human V_H 3-9 gene; and
 - (b) a light chain variable region of a human V_K L15 gene;
- 30 wherein the antibody specifically binds to PD-L1.

In another preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising:

- (a) a heavy chain variable region of a human V_H 3-9 gene; and

(b) a light chain variable region of a human V_K L18 gene;
wherein the antibody specifically binds to PD-L1.

In another aspect, the invention provides an isolated monoclonal antibody, or antigen binding portion thereof, comprising:

- 5 a heavy chain variable region that comprises CDR1, CDR2, and CDR3 sequences;
and a light chain variable region that comprises CDR1, CDR2, and CDR3
sequences, wherein:
- (a) the heavy chain variable region CDR3 sequence comprises an amino acid
sequence selected from the group consisting of SEQ ID NOs:41, 42, 43, 44, 45,
10 46, 47, 48, 49, and 50, and conservative modifications thereof;
- (b) the light chain variable region CDR3 sequence comprises an amino acid
sequence selected from the group consisting of SEQ ID NOs:71, 72, 73, 74, 75,
76, 77, 78, 79, and 80, and conservative modifications thereof; and
- (c) the antibody specifically binds to human PD-L1.
- 15 Preferably, the heavy chain variable region CDR2 sequence comprises an amino acid
sequence selected from the group consisting of amino acid sequences of SEQ ID NOs:31,
32, 33, 34, 35, 36, 37, 38, 39, and 40, and conservative modifications thereof; and the
light chain variable region CDR2 sequence comprises an amino acid sequence selected
from the group consisting of amino acid sequences of SEQ ID NOs:61, 62, 63, 64, 65, 66,
20 67, 68, 69, and 70, and conservative modifications thereof. Preferably, the heavy chain
variable region CDR1 sequence comprises an amino acid sequence selected from the
group consisting of amino acid sequences of SEQ ID NOs:21, 22, 23, 24, 25, 26, 27, 28,
29, and 30, and conservative modifications thereof; and the light chain variable region
CDR1 sequence comprises an amino acid sequence selected from the group consisting of
25 amino acid sequences of SEQ ID NOs:51, 52, 53, 54, 55, 56, 57, 58, 59, and 60, and
conservative modifications thereof.

In yet another aspect, the invention provides an isolated monoclonal antibody, or antigen binding portion thereof, comprising a heavy chain variable region and a light chain variable region, wherein:

- 30 (a) the heavy chain variable region comprises an amino acid sequence that is at
least 80% homologous to an amino acid sequence selected from the group
consisting of SEQ ID NOs:1, 2, 3, 4, 5, 6, 7, 8, 9, and 10;

- (b) the light chain variable region comprises an amino acid sequence that is at least 80% homologous to an amino acid sequence selected from the group consisting of SEQ ID NOs:11, 12, 13, 14, 15, 16, 17, 18, 19, and 20; and
- (c) the antibody binds to human PD-L1 with a K_D of $1 \times 10^{-7} M$ or less.

5 In a preferred embodiment, the antibodies additionally comprise at least one of the following properties:

 (a) the antibody increases T-cell proliferation in a mixed lymphocyte reaction (MLR) assay;

 (b) the antibody increases interferon- γ production in an MLR assay; or

10 (c) the antibody increases IL-2 secretion in an MLR assay.

 In preferred embodiments, the invention provides an isolated monoclonal antibody, or antigen binding portion thereof, comprising:

 (a) a heavy chain variable region CDR1 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:21, 22, 23, 24, 25, 26, 27, 28, 15 29, and 30;

 (b) a heavy chain variable region CDR2 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:31, 32, 33, 34, 35, 36, 37, 38, 39, and 40;

 (c) a heavy chain variable region CDR3 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:41, 42, 43, 44, 45, 46, 47, 48, 20 49, and 50;

 (d) a light chain variable region CDR1 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:51, 52, 53, 54, 55, 56, 57, 58, 59, and 60;

25 (e) a light chain variable region CDR2 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:61, 62, 63, 64, 65, 66, 67, 68, 69, and 70; and

 (f) a light chain variable region CDR3 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:71, 72, 73, 74, 75, 76, 77, 78, 30 79, and 80;

 wherein the antibody specifically binds PD-L1.

 A preferred combination comprises:

 (a) a heavy chain variable region CDR1 comprising SEQ ID NO:21;

- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:31;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:41;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:51;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:61; and
- 5 (f) a light chain variable region CDR3 comprising SEQ ID NO:71.

Another preferred combination comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:22;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:32;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:42;
- 10 (d) a light chain variable region CDR1 comprising SEQ ID NO:52;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:62; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:72.

Another preferred combination comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:23;
- 15 (b) a heavy chain variable region CDR2 comprising SEQ ID NO:33;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:43;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:53;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:63; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:73.

20 Another preferred combination comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:24;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:34;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:44;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:54;
- 25 (e) a light chain variable region CDR2 comprising SEQ ID NO:64; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:74.

Another preferred combination comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:25;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:35;
- 30 (c) a heavy chain variable region CDR3 comprising SEQ ID NO:45;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:55;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:65; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:75.

Another preferred combination comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:26;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:36;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:46;
- 5 (d) a light chain variable region CDR1 comprising SEQ ID NO:56;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:66; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:76.

Another preferred combination comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:27;
- 10 (b) a heavy chain variable region CDR2 comprising SEQ ID NO:37;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:47;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:57;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:67; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:77.

15 Another preferred combination comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:28;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:38;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:48;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:58;
- 20 (e) a light chain variable region CDR2 comprising SEQ ID NO:68; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:78.

Another preferred combination comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:29;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:39;
- 25 (c) a heavy chain variable region CDR3 comprising SEQ ID NO:49;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:59;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:69; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:79.

Another preferred combination comprises:

- 30 (a) a heavy chain variable region CDR1 comprising SEQ ID NO:30;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:40;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:50;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:60;

(e) a light chain variable region CDR2 comprising SEQ ID NO:70; and

(f) a light chain variable region CDR3 comprising SEQ ID NO:80.

Other preferred antibodies of the invention, or antigen binding portions thereof comprise:

- 5 (a) a heavy chain variable region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:1, 2, 3, 4, 5, 6, 7, 8, 9, and 10; and
(b) a light chain variable region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:11, 12, 13, 14, 15, 16, 17, 18, 19, and 20; wherein the antibody specifically binds PD-L1.

A preferred combination comprises:

- 10 (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:1; and
(b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:11.

Another preferred combination comprises:

- 15 (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:2; and
(b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:12.

Another preferred combination comprises:

- 20 (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:3; and
(b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:13.

Another preferred combination comprises:

- 25 (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:4; and
(b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:14.

Another preferred combination comprises:

- 30 (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:5; and
(b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:15.

Another preferred combination comprises:

(a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:6; and

5 (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:16.

Another preferred combination comprises:

(a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:7; and

10 (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:17.

Another preferred combination comprises:

(a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:8; and

15 (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:18.

Another preferred combination comprises:

(a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:9; and

20 (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:19.

Another preferred combination comprises:

(a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:10; and

25 (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:20.

In another aspect of the instant disclosure, antibodies, or antigen-binding portions thereof, are provided that compete for binding to PD-L1 with any of the aforementioned antibodies.

30 The antibodies of the instant disclosure can be, for example, full-length antibodies, for example of an IgG1 or IgG4 isotype. Alternatively, the antibodies can be antibody fragments, such as Fab or Fab'2 fragments, or single chain antibodies.

The instant disclosure also provides an immunoconjugate comprising an antibody of the invention, or antigen-binding portion thereof, linked to a therapeutic agent, such as

a cytotoxin or a radioactive isotope. The invention also provides a bispecific molecule comprising an antibody, or antigen-binding portion thereof, of the invention, linked to a second functional moiety having a different binding specificity than said antibody, or antigen binding portion thereof.

5 Compositions comprising an antibody, or antigen-binding portion thereof, or immunoconjugate or bispecific molecule of the instant disclosure and a pharmaceutically acceptable carrier are also provided.

 Nucleic acid molecules encoding the antibodies, or antigen-binding portions thereof, of the invention are also encompassed by the invention, as well as expression
10 vectors comprising such nucleic acids and host cells comprising such expression vectors. Moreover, the invention provides a transgenic mouse comprising human immunoglobulin heavy and light chain transgenes, wherein the mouse expresses an antibody of the invention, as well as hybridomas prepared from such a mouse, wherein the hybridoma produces the antibody of the invention.

15 In yet another aspect, the invention provides a method of modulating an immune response in a subject comprising administering to the subject the antibody, or antigen-binding portion thereof, of the invention such that the immune response in the subject is modulated. Preferably, the antibody of the invention enhances, stimulates or increases the immune response in the subject.

20 In a further aspect, the invention provides a method of inhibiting growth of tumor cells in a subject, comprising administering to a subject a therapeutically effective amount of an anti-PD-L1 antibody, or antigen-binding portion thereof. The antibodies of the invention are preferred for use in the method although other anti-PD-L1 antibodies can be used instead (or in combination with an anti-PD-L1 antibody of the invention). For
25 example, a chimeric, humanized or fully human anti-PD-L1 antibody can be used in the method of inhibiting tumor growth.

 In a further aspect, the invention provides a method of treating an infectious disease in a subject, comprising administering to a subject a therapeutically effective amount of an anti-PD-L1 antibody, or antigen-binding portion thereof. The antibodies of
30 the invention are preferred for use in the method although other anti-PD-L1 antibodies can be used instead (or in combination with an anti-PD-L1 antibody of the invention). For example, a chimeric, humanized or fully human anti-PD-L1 antibody can be used in the method of treating an infectious disease.

Still further, the invention provides a method of enhancing an immune response to an antigen in a subject, comprising administering to the subject: (i) the antigen; and (ii) an anti-PD-L1 antibody, or antigen-binding portion thereof, such that an immune response to the antigen in the subject is enhanced. The antigen can be, for example, a tumor antigen,
5 a viral antigen, a bacterial antigen or an antigen from a pathogen. The antibodies of the invention are preferred for use in the method although other anti-PD-L1 antibodies can be used instead (or in combination with an anti-PD-L1 antibody of the invention). For example, a chimeric, humanized or fully human anti-PD-L1 antibody can be used in the method of enhancing an immune response to an antigen in a subject.

10 The invention also provides methods for making "second generation" anti-PD-L1 antibodies based on the sequences of the anti-PD-L1 antibodies provided herein. For example, the invention provides a method for preparing an anti-PD-L1 antibody comprising:

(a) providing: (i) a heavy chain variable region antibody sequence comprising a
15 CDR1 sequence that is selected from the group consisting of SEQ ID NOs:21, 22, 23, 24, 25, 26, 27, 28, 29, and 30, a CDR2 sequence that is selected from the group consisting of SEQ ID NOs:31, 32, 33, 34, 35, 36, 37, 38, 39, and 40; and a CDR3 sequence that is selected from the group consisting of SEQ ID NOs:41, 42, 43, 44, 45, 46, 47, 48, 49, and 50; or (ii) a light chain variable region antibody sequence comprising a CDR1 sequence
20 that is selected from the group consisting of SEQ ID NOs:51, 52, 53, 54, 55, 56, 57, 58, 59, and 60, a CDR2 sequence that is selected from the group consisting of SEQ ID NOs:61, 62, 63, 64, 65, 66, 67, 68, 69, and 70, and a CDR3 sequence that is selected from the group consisting of SEQ ID NOs:71, 72, 73, 74, 75, 76, 77, 78, 79, and 80;

(b) altering at least one amino acid residue within at least one variable region
25 antibody sequence, said sequence being selected from the heavy chain variable region antibody sequence and the light chain variable region antibody sequence, to create at least one altered antibody sequence; and

(c) expressing the altered antibody sequence as a protein.

Other features and advantages of the instant invention will be apparent from the
30 following detailed description and examples which should not be construed as limiting. The contents of all references, Genbank entries, patents and published patent applications cited throughout this application are expressly incorporated herein by reference.

Brief Description of the Drawings

Figure 1A shows the nucleotide sequence (SEQ ID NO:81) and amino acid sequence (SEQ ID NO:1) of the heavy chain variable region of the 3G10 human monoclonal antibody. The CDR1 (SEQ ID NO:21), CDR2 (SEQ ID NO:31) and CDR3 (SEQ ID NO:41) regions are delineated and the V, D and J germline derivations are indicated.

Figure 1B shows the nucleotide sequence (SEQ ID NO:91) and amino acid sequence (SEQ ID NO:11) of the light chain variable region of the 3G10 human monoclonal antibody. The CDR1 (SEQ ID NO:51), CDR2 (SEQ ID NO:61) and CDR3 (SEQ ID NO:71) regions are delineated and the V and J germline derivations are indicated.

Figure 2A shows the nucleotide sequence (SEQ ID NO:82) and amino acid sequence (SEQ ID NO:2) of the heavy chain variable region of the 12A4 human monoclonal antibody. The CDR1 (SEQ ID NO:22), CDR2 (SEQ ID NO:32) and CDR3 (SEQ ID NO:42) regions are delineated and the V and J germline derivations are indicated.

Figure 2B shows the nucleotide sequence (SEQ ID NO:92) and amino acid sequence (SEQ ID NO:12) of the light chain variable region of the 12A4 human monoclonal antibody. The CDR1 (SEQ ID NO:52), CDR2 (SEQ ID NO:62) and CDR3 (SEQ ID NO:72) regions are delineated and the V and J germline derivations are indicated.

Figure 3A shows the nucleotide sequence (SEQ ID NO:83) and amino acid sequence (SEQ ID NO:3) of the heavy chain variable region of the 10A5 human monoclonal antibody. The CDR1 (SEQ ID NO:23), CDR2 (SEQ ID NO:33) and CDR3 (SEQ ID NO:43) regions are delineated and the V and J germline derivations are indicated.

Figure 3B shows the nucleotide sequence (SEQ ID NO:93) and amino acid sequence (SEQ ID NO:13) of the light chain variable region of the 10A5 human monoclonal antibody. The CDR1 (SEQ ID NO:53), CDR2 (SEQ ID NO:63) and CDR3 (SEQ ID NO:73) regions are delineated and the V and J germline derivations are indicated.

Figure 4A shows the nucleotide sequence (SEQ ID NO:84) and amino acid sequence (SEQ ID NO:4) of the heavy chain variable region of the 5F8 human

monoclonal antibody. The CDR1 (SEQ ID NO:24), CDR2 (SEQ ID NO:34) and CDR3 (SEQ ID NO:44) regions are delineated and the V and J germline derivations are indicated.

5 Figure 4B shows the nucleotide sequence (SEQ ID NO:94) and amino acid sequence (SEQ ID NO:14) of the light chain variable region of the 5F8 human monoclonal antibody. The CDR1 (SEQ ID NO:54), CDR2 (SEQ ID NO:64) and CDR3 (SEQ ID NO:74) regions are delineated and the V and J germline derivations are indicated.

10 Figure 5A shows the nucleotide sequence (SEQ ID NO:85) and amino acid sequence (SEQ ID NO:5) of the heavy chain variable region of the 10H10 human monoclonal antibody. The CDR1 (SEQ ID NO:25), CDR2 (SEQ ID NO:35) and CDR3 (SEQ ID NO:45) regions are delineated and the V and J germline derivations are indicated.

15 Figure 5B shows the nucleotide sequence (SEQ ID NO:95) and amino acid sequence (SEQ ID NO:15) of the light chain variable region of the 10H10 human monoclonal antibody. The CDR1 (SEQ ID NO:55), CDR2 (SEQ ID NO:65) and CDR3 (SEQ ID NO:75) regions are delineated and the V and J germline derivations are indicated.

20 Figure 6A shows the nucleotide sequence (SEQ ID NO:86) and amino acid sequence (SEQ ID NO:6) of the heavy chain variable region of the 1B12 human monoclonal antibody. The CDR1 (SEQ ID NO:26), CDR2 (SEQ ID NO:36) and CDR3 (SEQ ID NO:46) regions are delineated and the V and J germline derivations are indicated.

25 Figure 6B shows the nucleotide sequence (SEQ ID NO:96) and amino acid sequence (SEQ ID NO:16) of the light chain variable region of the 1B12 human monoclonal antibody. The CDR1 (SEQ ID NO:56), CDR2 (SEQ ID NO:66) and CDR3 (SEQ ID NO:76) regions are delineated and the V and J germline derivations are indicated.

30 Figure 7A shows the nucleotide sequence (SEQ ID NO:87) and amino acid sequence (SEQ ID NO:7) of the heavy chain variable region of the 7H1 human monoclonal antibody. The CDR1 (SEQ ID NO:27), CDR2 (SEQ ID NO:37) and CDR3 (SEQ ID NO:47) regions are delineated and the V and J germline derivations are indicated.

Figure 7B shows the nucleotide sequence (SEQ ID NO:97) and amino acid sequence (SEQ ID NO:17) of the light chain variable region of the 7H1 human monoclonal antibody. The CDR1 (SEQ ID NO:57), CDR2 (SEQ ID NO:67) and CDR3 (SEQ ID NO:77) regions are delineated and the V and J germline derivations are indicated.

Figure 8A shows the nucleotide sequence (SEQ ID NO:88) and amino acid sequence (SEQ ID NO:8) of the heavy chain variable region of the 11E6 human monoclonal antibody. The CDR1 (SEQ ID NO:28), CDR2 (SEQ ID NO:38) and CDR3 (SEQ ID NO:48) regions are delineated and the V and J germline derivations are indicated.

Figure 8B shows the nucleotide sequence (SEQ ID NO:98) and amino acid sequence (SEQ ID NO:18) of the light chain variable region of the 11E6 human monoclonal antibody. The CDR1 (SEQ ID NO:58), CDR2 (SEQ ID NO:68) and CDR3 (SEQ ID NO:78) regions are delineated and the V and J germline derivations are indicated.

Figure 9A shows the nucleotide sequence (SEQ ID NO:89) and amino acid sequence (SEQ ID NO:9) of the heavy chain variable region of the 12B7 human monoclonal antibody. The CDR1 (SEQ ID NO:29), CDR2 (SEQ ID NO:39) and CDR3 (SEQ ID NO:49) regions are delineated and the V and J germline derivations are indicated.

Figure 9B shows the nucleotide sequence (SEQ ID NO:99) and amino acid sequence (SEQ ID NO:19) of the light chain variable region of the 12B7 human monoclonal antibody. The CDR1 (SEQ ID NO:59), CDR2 (SEQ ID NO:69) and CDR3 (SEQ ID NO:79) regions are delineated and the V and J germline derivations are indicated.

Figure 10A shows the nucleotide sequence (SEQ ID NO:90) and amino acid sequence (SEQ ID NO:10) of the heavy chain variable region of the 13G4 human monoclonal antibody. The CDR1 (SEQ ID NO:30), CDR2 (SEQ ID NO:40) and CDR3 (SEQ ID NO:50) regions are delineated and the V and J germline derivations are indicated.

Figure 10B shows the nucleotide sequence (SEQ ID NO:100) and amino acid sequence (SEQ ID NO:20) of the light chain variable region of the 13G4 human monoclonal antibody. The CDR1 (SEQ ID NO:60), CDR2 (SEQ ID NO:70) and CDR3

(SEQ ID NO:80) regions are delineated and the V and J germline derivations are indicated.

Figure 11 shows the alignment of the amino acid sequence of the heavy chain variable region of 3G10 with the human germline V_H 1-18 amino acid sequence (SEQ ID NO:101).

Figure 12 shows the alignment of the amino acid sequence of the heavy chain variable region of 12A4 with the human germline V_H 1-69 amino acid sequence (SEQ ID NO:102).

Figure 13 shows the alignment of the amino acid sequence of the heavy chain variable region of 10A5 with the human germline V_H 1-3 amino acid sequence (SEQ ID NO:103).

Figure 14 shows the alignment of the amino acid sequence of the heavy chain variable region of 5F8 with the human germline V_H 1-69 amino acid sequence (SEQ ID NO:102).

Figure 15 shows the alignment of the amino acid sequence of the heavy chain variable region of 10H10 with the human germline V_H 3-9 amino acid sequence (SEQ ID NO:104).

Figure 16 shows the alignment of the amino acid sequence of the heavy chain variable region of 1B12 with the human germline V_H 1-69 amino acid sequence (SEQ ID NO:102).

Figure 17 shows the alignment of the amino acid sequence of the heavy chain variable region of 7H1 with the human germline V_H 1-69 amino acid sequence (SEQ ID NO:102).

Figure 18 shows the alignment of the amino acid sequence of the heavy chain variable region of 11E6 with the human germline V_H 1-69 amino acid sequence (SEQ ID NO:102).

Figure 19 shows the alignment of the amino acid sequence of the heavy chain variable region of 12B7 with the human germline V_H 1-69 amino acid sequence (SEQ ID NO:102).

Figure 20 shows the alignment of the amino acid sequence of the heavy chain variable region of 13G4 with the human germline V_H 3-9 amino acid sequence (SEQ ID NO:104).

Figure 21 shows the alignment of the amino acid sequence of the light chain variable region of 3G10 with the human germline V_k L6 amino acid sequence (SEQ ID NO:105).

5 Figure 22 shows the alignment of the amino acid sequence of the light chain variable region of 12A4 with the human germline V_k L6 amino acid sequence (SEQ ID NO:105).

Figure 23 shows the alignment of the amino acid sequence of the light chain variable region of 10A5 with the human germline V_k L15 amino acid sequence (SEQ ID NO:106).

10 Figure 24 shows the alignment of the amino acid sequence of the light chain variable region of 5F8 with the human germline V_k A27 amino acid sequence (SEQ ID NO:107).

Figure 25 shows the alignment of the amino acid sequence of the light chain variable region of 10H10 with the human germline V_k L15 amino acid sequence (SEQ ID
15 NO:106).

Figure 26 shows the alignment of the amino acid sequence of the light chain variable region of 1B12 with the human germline V_k L6 amino acid sequence (SEQ ID NO:105).

Figure 27 shows the alignment of the amino acid sequence of the light chain
20 variable region of 7H1 with the human germline V_k L6 amino acid sequence (SEQ ID NO:105).

Figure 28 shows the alignment of the amino acid sequence of the light chain variable region of 11E6 with the human germline V_k A27 amino acid sequence (SEQ ID NO:107).

25 Figure 29 shows the alignment of the amino acid sequence of the light chain variable region of 11E6a (SEQ ID NO:109) with the human germline V_k A27 amino acid sequence (SEQ ID NO:107).

Figure 30 shows the alignment of the amino acid sequence of the light chain variable region of 12B7 with the human germline V_k L6 amino acid sequence (SEQ ID
30 NO:105).

Figure 31 shows the alignment of the amino acid sequence of the light chain variable region of 13G4 with the human germline V_k L18 amino acid sequence (SEQ ID NO:108).

Figures 32A-C show the results of flow cytometry experiments demonstrating that the human monoclonal antibodies 3G10, 10A5, and 12A4, directed against human PD-L1, binds the cell surface of CHO cells transfected with full-length human PD-L1. (A) Flow cytometry plot for 3G10 (B) Flow cytometry plot for 10A5 (C) Flow cytometry plot for 12A4.

Figure 33 shows the results of flow cytometry experiments demonstrating that the human monoclonal antibodies 3G10, 10A5, and 12A4, directed against human PD-L1, binds the cell surface of CHO cells transfected with full-length human PD-L1 in a concentration dependent manner.

Figure 34 shows the results of ELISA experiments demonstrating that the human monoclonal antibodies 3G10, 10A5, and 12A4, directed against human PD-L1, binds to PD-L1-Fc fusion protein.

Figure 35 shows the results of experiments demonstrating HuMab titration on stimulated human CD4+ T cells.

Figure 36 shows the results of experiments demonstrating HuMab titration on stimulated cynomolgus PBMC.

Figures 37A-C shows the results of flow cytometry experiments demonstrating that the human monoclonal antibodies 3G10, 10A5, and 12A4, directed against human PD-L1, binds to PD-L1 on the cell surface of activated T cells. (A) Flow cytometry plot for 3G10 (B) Flow cytometry plot for 10A5 (C) Flow cytometry plot for 12A4.

Figure 38 demonstrates binding of HuMabs to ES-2 cells.

Figures 39A-D shows the results of experiments demonstrating that human monoclonal antibodies against human PD-L1 promote T-cell proliferation, IFN- γ secretion and IL-2 secretion in a mixed lymphocyte reaction assay. Figure 39A is a bar graph showing concentration dependent T-cell proliferation using HuMab 10A5; Figure 39B is a bar graph showing concentration dependent IFN- γ secretion using HuMab 10A5; Figure 39C is a bar graph showing IFN- γ secretion using HuMabs 3G10 and 12A4; Figure 39D is a bar graph showing concentration dependent IL-2 secretion using HuMab 10A5.

Figure 40 demonstrates the effect of human anti-PD-L1 antibody on proliferation and IFN- γ secretion in the MLR using allogeneic dendritic cells and T cells (CD4+ effector T cells) Dendritic Cells.

Figures 41A-B shows the results of experiments demonstrating that human monoclonal antibodies against human PD-L1 promote T-cell proliferation and IFN- γ secretion in MLR containing T regulatory cells. Figure 41A is a bar graph showing concentration dependent T-cell proliferation using HuMAb 10A5; Figure 41B is a bar graph showing concentration dependent IFN- γ secretion using HuMAb 10A5.

Figure 42 demonstrates the results of anti-PD-L1 antibodies on cell proliferation in a Mixed Lymphocyte Reaction in the presence of regulatory T cells.

Figure 43 demonstrates the results of anti-PD-L1 antibodies on cytokine production in a Mixed Lymphocyte Reaction in the presence of regulatory T cells.

Figure 44 demonstrates the results of anti-PD-L1 antibodies on CMV lysate stimulated human PBMC IFN- γ secretion.

Figure 45 shows the results of flow cytometry experiments demonstrating that human monoclonal antibodies against human PD-L1 block the binding of PD-L1 to CHO transfected cells expressing PD-1.

Figure 46 shows that anti-PD-L1 antibodies block binding of PD-1 to IFN γ treated ES-2 cells.

Figure 47 shows the effect of anti-PD-L1 antibodies on tumor growth *in vivo*.

Detailed Description

In one aspect, the present disclosure relates to isolated monoclonal antibodies, particularly human monoclonal antibodies that bind specifically to PD-L1. In certain embodiments, the antibodies of the invention exhibit one or more desirable functional properties, such as high affinity binding to PD-L1, the ability to augment T cell proliferation, IFN- γ and/or IL-2 secretion in mixed lymphocyte reactions, the ability to inhibit binding of PD-L1 to the PD-1 receptor, the ability to stimulate antibody responses and/or the ability to reverse the suppressive function of T regulatory cells. Additionally or alternatively, the antibodies of the invention are derived from particular heavy and light chain germline sequences and/or comprise particular structural features such as CDR regions comprising particular amino acid sequences.

The instant disclosure provides, for example, isolated antibodies, methods of making such antibodies, immunoconjugates and bispecific molecules comprising such antibodies and pharmaceutical compositions containing the antibodies, immunoconjugates or bispecific molecules of the invention.

In another aspect, the disclosure pertains to methods of inhibiting growth of tumor cells in a subject using anti-PD-L1 antibodies. The invention also relates to methods of using the antibodies to modify an immune response, as well as to treat diseases such as cancer or infectious disease, or to stimulate a protective autoimmune response or to stimulate antigen-specific immune responses (e.g., by coadministration of anti-PD-L1 with an antigen of interest).

In order that the present disclosure may be more readily understood, certain terms are first defined. Additional definitions are set forth throughout the detailed description.

The term "immune response" refers to the action of, for example, lymphocytes, antigen presenting cells, phagocytic cells, granulocytes, and soluble macromolecules produced by the above cells or the liver (including antibodies, cytokines, and complement) that results in selective damage to, destruction of, or elimination from the human body of invading pathogens, cells or tissues infected with pathogens, cancerous cells, or, in cases of autoimmunity or pathological inflammation, normal human cells or tissues.

A "signal transduction pathway" refers to the biochemical relationship between a variety of signal transduction molecules that play a role in the transmission of a signal from one portion of a cell to another portion of a cell. As used herein, the phrase "cell surface receptor" includes, for example, molecules and complexes of molecules capable of receiving a signal and the transmission of such a signal across the plasma membrane of a cell. An example of a "cell surface receptor" of the present invention is the PD-L1 receptor.

The term "antibody" as referred to herein includes whole antibodies and any antigen binding fragment (*i.e.*, "antigen-binding portion") or single chains thereof. An "antibody" refers to a glycoprotein comprising at least two heavy (H) chains and two light (L) chains inter-connected by disulfide bonds, or an antigen binding portion thereof. Each heavy chain is comprised of a heavy chain variable region (abbreviated herein as V_H) and a heavy chain constant region. The heavy chain constant region is comprised of three domains, C_{H1} , C_{H2} and C_{H3} . Each light chain is comprised of a light chain variable region (abbreviated herein as V_L) and a light chain constant region. The light chain constant region is comprised of one domain, C_L . The V_H and V_L regions can be further subdivided into regions of hypervariability, termed complementarity determining regions (CDR), interspersed with regions that are more conserved, termed framework regions

(FR). Each V_H and V_L is composed of three CDRs and four FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1, CDR1, FR2, CDR2, FR3, CDR3, FR4. The variable regions of the heavy and light chains contain a binding domain that interacts with an antigen. The constant regions of the antibodies may mediate the binding of the immunoglobulin to host tissues or factors, including various cells of the immune system (e.g., effector cells) and the first component (C1q) of the classical complement system.

The term "antigen-binding portion" of an antibody (or simply "antibody portion"), as used herein, refers to one or more fragments of an antibody that retain the ability to specifically bind to an antigen (e.g., PD-L1). It has been shown that the antigen-binding function of an antibody can be performed by fragments of a full-length antibody. Examples of binding fragments encompassed within the term "antigen-binding portion" of an antibody include (i) a Fab fragment, a monovalent fragment consisting of the V_L , V_H , C_L and C_{H1} domains; (ii) a $F(ab')_2$ fragment, a bivalent fragment comprising two Fab fragments linked by a disulfide bridge at the hinge region; (iii) a Fd fragment consisting of the V_H and C_{H1} domains; (iv) a Fv fragment consisting of the V_L and V_H domains of a single arm of an antibody, (v) a dAb fragment (Ward *et al.*, (1989) *Nature* 341:544-546), which consists of a V_H domain; and (vi) an isolated complementarity determining region (CDR). Furthermore, although the two domains of the Fv fragment, V_L and V_H , are coded for by separate genes, they can be joined, using recombinant methods, by a synthetic linker that enables them to be made as a single protein chain in which the V_L and V_H regions pair to form monovalent molecules (known as single chain Fv (scFv); see e.g., Bird *et al.* (1988) *Science* 242:423-426; and Huston *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:5879-5883). Such single chain antibodies are also intended to be encompassed within the term "antigen-binding portion" of an antibody. These antibody fragments are obtained using conventional techniques known to those with skill in the art, and the fragments are screened for utility in the same manner as are intact antibodies.

An "isolated antibody," as used herein, is intended to refer to an antibody that is substantially free of other antibodies having different antigenic specificities (e.g., an isolated antibody that specifically binds PD-L1 is substantially free of antibodies that specifically bind antigens other than PD-L1). An isolated antibody that specifically binds PD-L1 may, however, have cross-reactivity to other antigens, such as PD-L1 molecules

from other species. Moreover, an isolated antibody may be substantially free of other cellular material and/or chemicals.

The terms "monoclonal antibody" or "monoclonal antibody composition" as used herein refer to a preparation of antibody molecules of single molecular composition. A
5 monoclonal antibody composition displays a single binding specificity and affinity for a particular epitope.

The term "human antibody," as used herein, is intended to include antibodies having variable regions in which both the framework and CDR regions are derived from human germline immunoglobulin sequences. Furthermore, if the antibody contains a
10 constant region, the constant region also is derived from human germline immunoglobulin sequences. The human antibodies of the invention may include amino acid residues not encoded by human germline immunoglobulin sequences (*e.g.*, mutations introduced by random or site-specific mutagenesis *in vitro* or by somatic mutation *in vivo*). However, the term "human antibody," as used herein, is not intended to include
15 antibodies in which CDR sequences derived from the germline of another mammalian species, such as a mouse, have been grafted onto human framework sequences.

The term "human monoclonal antibody" refers to antibodies displaying a single binding specificity which have variable regions in which both the framework and CDR regions are derived from human germline immunoglobulin sequences. In one
20 embodiment, the human monoclonal antibodies are produced by a hybridoma which includes a B cell obtained from a transgenic nonhuman animal, *e.g.*, a transgenic mouse, having a genome comprising a human heavy chain transgene and a light chain transgene fused to an immortalized cell.

The term "recombinant human antibody," as used herein, includes all human
25 antibodies that are prepared, expressed, created or isolated by recombinant means, such as (a) antibodies isolated from an animal (*e.g.*, a mouse) that is transgenic or transchromosomal for human immunoglobulin genes or a hybridoma prepared therefrom (described further below), (b) antibodies isolated from a host cell transformed to express the human antibody, *e.g.*, from a transfectoma, (c) antibodies isolated from a
30 recombinant, combinatorial human antibody library, and (d) antibodies prepared, expressed, created or isolated by any other means that involve splicing of human immunoglobulin gene sequences to other DNA sequences. Such recombinant human antibodies have variable regions in which the framework and CDR regions are derived

from human germline immunoglobulin sequences. In certain embodiments, however, such recombinant human antibodies can be subjected to *in vitro* mutagenesis (or, when an animal transgenic for human Ig sequences is used, *in vivo* somatic mutagenesis) and thus the amino acid sequences of the V_H and V_L regions of the recombinant antibodies are
5 sequences that, while derived from and related to human germline V_H and V_L sequences, may not naturally exist within the human antibody germline repertoire *in vivo*.

As used herein, "isotype" refers to the antibody class (e.g., IgM or IgG1) that is encoded by the heavy chain constant region genes.

The phrases "an antibody recognizing an antigen" and "an antibody specific for an
10 antigen" are used interchangeably herein with the term "an antibody which binds specifically to an antigen."

The term "human antibody derivatives" refers to any modified form of the human antibody, e.g., a conjugate of the antibody and another agent or antibody.

The term "humanized antibody" is intended to refer to antibodies in which CDR
15 sequences derived from the germline of another mammalian species, such as a mouse, have been grafted onto human framework sequences. Additional framework region modifications may be made within the human framework sequences.

The term "chimeric antibody" is intended to refer to antibodies in which the variable region sequences are derived from one species and the constant region sequences
20 are derived from another species, such as an antibody in which the variable region sequences are derived from a mouse antibody and the constant region sequences are derived from a human antibody.

As used herein, an antibody that "specifically binds to human PD-L1" is intended to refer to an antibody that binds to human PD-L1 with a K_D of 1×10^{-7} M or less, more
25 preferably 5×10^{-8} M or less, more preferably 1×10^{-8} M or less, more preferably 5×10^{-9} M or less, even more preferably between 1×10^{-8} M and 1×10^{-10} M or less.

The term "K_{assoc}" or "K_a," as used herein, is intended to refer to the association rate of a particular antibody-antigen interaction, whereas the term "K_{dis}" or "K_d," as used
herein, is intended to refer to the dissociation rate of a particular antibody-antigen
30 interaction. The term "K_D," as used herein, is intended to refer to the dissociation constant, which is obtained from the ratio of K_d to K_a (i.e., K_d/K_a) and is expressed as a molar concentration (M). K_D values for antibodies can be determined using methods well established in the art. A preferred method for determining the K_D of an antibody is by

using surface plasmon resonance, preferably using a biosensor system such as a Biacore® system.

As used herein, the term "high affinity" for an IgG antibody refers to an antibody having a K_D of 10^{-8} M or less, more preferably 10^{-9} M or less and even more preferably 10^{-10} M or less for a target antigen. However, "high affinity" binding can vary for other antibody isotypes. For example, "high affinity" binding for an IgM isotype refers to an antibody having a K_D of 10^{-7} M or less, more preferably 10^{-8} M or less, even more preferably 10^{-9} M or less.

As used herein, the term "subject" includes any human or nonhuman animal. The term "nonhuman animal" includes all vertebrates, *e.g.*, mammals and non-mammals, such as nonhuman primates, sheep, dogs, cats, horses, cows, chickens, amphibians, reptiles, *etc.*

Various aspects of the disclosure are described in further detail in the following subsections.

15 Anti-PD-L1 Antibodies

The antibodies of the invention are characterized by particular functional features or properties of the antibodies. For example, the antibodies bind specifically to human PD-L1. Preferably, an antibody of the invention binds to PD-L1 with high affinity, for example with a K_D of 1×10^{-7} M or less. The anti-PD-L1 antibodies of the invention preferably exhibit one or more of the following characteristics:

- (a) binds to human PD-L1 with a K_D of 1×10^{-7} M or less;
- (b) increases T-cell proliferation in a mixed lymphocyte reaction (MLR) assay;
- (c) increases interferon- γ production in an MLR assay;
- (d) increases IL-2 secretion in an MLR assay
- 25 (e) stimulates antibody responses; and/or
- (f) reverses the effect of T regulatory cells on T cell effector cells and /or dendritic cells..

Preferably, the antibody binds to human PD-L1 with a K_D of 5×10^{-8} M or less, binds to human PD-L1 with a K_D of 1×10^{-8} M or less, binds to human PD-L1 with a K_D of 5×10^{-9} M or less, binds to human PD-L1 with a K_D of 4×10^{-9} M or less, binds to human PD-L1 with a K_D of 2×10^{-9} M or less, or binds to human PD-L1 with a K_D of between 1×10^{-9} M and 1×10^{-10} M or less.

Standard assays to evaluate the binding ability of the antibodies toward PD-L1 are known in the art, including for example, ELISAs, Western blots and RIAs. Suitable assays are described in detail in the Examples. The binding kinetics (e.g., binding affinity) of the antibodies also can be assessed by standard assays known in the art, such as by Biacore[®] analysis.

Monoclonal Antibodies 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4.

Preferred antibodies of the invention are the human monoclonal antibodies 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4, isolated and structurally characterized as described in Examples 1 and 2. The V_H amino acid sequences of 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4 are shown in SEQ ID NOs:1, 2, 3, 4, 5, 6, 7, 8, 9, and 10, respectively. The V_L amino acid sequences of 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4 are shown in SEQ ID NOs:11, 12, 13, 14, 15, 16, 17, 18, 19, and 20, respectively.

Given that each of these antibodies can bind to PD-L1, the V_H and V_L sequences can be "mixed and matched" to create other anti-PD-L1 binding molecules of the invention. PD-L1 binding of such "mixed and matched" antibodies can be tested using the binding assays described above and in the Examples (e.g., ELISAs). Preferably, when V_H and V_L chains are mixed and matched, a V_H sequence from a particular V_H/V_L pairing is replaced with a structurally similar V_H sequence. Likewise, preferably a V_L sequence from a particular V_H/V_L pairing is replaced with a structurally similar V_L sequence.

Accordingly, in one aspect, the invention provides an isolated monoclonal antibody, or antigen binding portion thereof comprising:

- (a) a heavy chain variable region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:1, 2, 3, 4, 5, 6, 7, 8, 9, and 10; and
- (b) a light chain variable region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:11, 12, 13, 14, 15, 16, 17, 18, 19, and 20; wherein the antibody specifically binds PD-L1, preferably human PD-L1.

Preferred heavy and light chain combinations include:

- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:1; and (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:11; or

- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:2; and (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:12; or
- 5 (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:3; and (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:13; or
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:4; and (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:14; or
- 10 (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:5; and (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:15; or
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:6; and (b) a light chain variable region comprising the amino acid sequence of
- 15 SEQ ID NO:16; or
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:7; and (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:17; or
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID
- 20 NO:8; and (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:18; or
- (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:9; and (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:19; or
- 25 (a) a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:10; and (b) a light chain variable region comprising the amino acid sequence of SEQ ID NO:20.

In another aspect, the invention provides antibodies that comprise the heavy chain and light chain CDR1s, CDR2s and CDR3s of 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4, or combinations thereof. The amino acid sequences of the

30 V_H CDR1s of 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4 are shown in SEQ ID NOs:21, 22, 23, 24, 25, 26, 27, 28, 29, and 30, respectively. The amino acid sequences of the V_H CDR2s of 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6,

12B7, and 13G4 are shown in SEQ ID NOs:31, 32, 33, 34, 35, 36, 37, 38, 39, and 40, respectively. The amino acid sequences of the V_H CDR3s of 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4 are shown in SEQ ID NOs:41, 42, 43, 44, 45, 46, 47, 48, 49, and 50, respectively. The amino acid sequences of the V_k CDR1s of 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4 are shown in SEQ ID NOs:51, 52, 53, 54, 55, 56, 57, 58, 59, and 60, respectively. The amino acid sequences of the V_k CDR2s of 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4 are shown in SEQ ID NOs:61, 62, 63, 64, 65, 66, 67, 68, 69, and 70, respectively. The amino acid sequences of the V_k CDR3s of 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4 are shown in SEQ ID NOs:71, 72, 73, 74, 75, 76, 77, 78, 79, and 80, respectively. The CDR regions are delineated using the Kabat system (Kabat, E. A., *et al.* (1991) Sequences of Proteins of Immunological Interest, Fifth Edition, U.S. Department of Health and Human Services, NIH Publication No. 91-3242).

Given that each of these antibodies can bind to PD-L1 and that antigen-binding specificity is provided primarily by the CDR1, CDR2, and CDR3 regions, the V_H CDR1, CDR2, and CDR3 sequences and V_k CDR1, CDR2, and CDR3 sequences can be "mixed and matched" (*i.e.*, CDRs from different antibodies can be mixed and match, although each antibody must contain a V_H CDR1, CDR2, and CDR3 and a V_k CDR1, CDR2, and CDR3) to create other anti-PD-L1 binding molecules of the invention. PD-L1 binding of such "mixed and matched" antibodies can be tested using the binding assays described above and in the Examples (*e.g.*, ELISAs, Biacore analysis). Preferably, when V_H CDR sequences are mixed and matched, the CDR1, CDR2 and/or CDR3 sequence from a particular V_H sequence is replaced with a structurally similar CDR sequence(s). Likewise, when V_k CDR sequences are mixed and matched, the CDR1, CDR2 and/or CDR3 sequence from a particular V_k sequence preferably is replaced with a structurally similar CDR sequence(s). It will be readily apparent to the ordinarily skilled artisan that novel V_H and V_L sequences can be created by substituting one or more V_H and/or V_L CDR region sequences with structurally similar sequences from the CDR sequences disclosed herein for monoclonal antibodies 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4.

Accordingly, in another aspect, the invention provides an isolated monoclonal antibody, or antigen binding portion thereof comprising:

- (a) a heavy chain variable region CDR1 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:21, 22, 23, 24, 25, 26, 27, 28, 29, and 30;
- (b) a heavy chain variable region CDR2 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:31, 32, 33, 34, 35, 36, 37, 38, 39, and 40;
- (c) a heavy chain variable region CDR3 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:41, 42, 43, 44, 45, 46, 47, 48, 49, and 50;
- (d) a light chain variable region CDR1 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:51, 52, 53, 54, 55, 56, 57, 58, 59, and 60;
- (e) a light chain variable region CDR2 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:61, 62, 63, 64, 65, 66, 67, 68, 69, and 70; and
- (f) a light chain variable region CDR3 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:71, 72, 73, 74, 75, 76, 77, 78, 79, and 80;

wherein the antibody specifically binds PD-L1, preferably human PD-L1.

In a preferred embodiment, the antibody comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:21;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:31;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:41;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:51;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:61; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:71.

In another preferred embodiment the antibody comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:22;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:32;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:42;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:52;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:62; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:72.

In another preferred embodiment the antibody comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:23;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:33;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:43;
- 5 (d) a light chain variable region CDR1 comprising SEQ ID NO:53;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:63; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:73.

In another preferred embodiment the antibody comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:24;
- 10 (b) a heavy chain variable region CDR2 comprising SEQ ID NO:34;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:44;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:54;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:64; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:74.

15 In another preferred embodiment the antibody comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:25;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:35;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:45;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:55;
- 20 (e) a light chain variable region CDR2 comprising SEQ ID NO:65; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:75.

In another preferred embodiment the antibody comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:26;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:36;
- 25 (c) a heavy chain variable region CDR3 comprising SEQ ID NO:46;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:56;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:66; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:76.

In another preferred embodiment the antibody comprises:

- 30 (a) a heavy chain variable region CDR1 comprising SEQ ID NO:27;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:37;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:47;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:57;

- (e) a light chain variable region CDR2 comprising SEQ ID NO:67; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:77.

In another preferred embodiment the antibody comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:28;
- 5 (b) a heavy chain variable region CDR2 comprising SEQ ID NO:38;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:48;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:58;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:68; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:78.

10 In another preferred embodiment the antibody comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:29;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:39;
- (c) a heavy chain variable region CDR3 comprising SEQ ID NO:49;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:59;
- 15 (e) a light chain variable region CDR2 comprising SEQ ID NO:69; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:79.

In another preferred embodiment the antibody comprises:

- (a) a heavy chain variable region CDR1 comprising SEQ ID NO:30;
- (b) a heavy chain variable region CDR2 comprising SEQ ID NO:40;
- 20 (c) a heavy chain variable region CDR3 comprising SEQ ID NO:50;
- (d) a light chain variable region CDR1 comprising SEQ ID NO:60;
- (e) a light chain variable region CDR2 comprising SEQ ID NO:70; and
- (f) a light chain variable region CDR3 comprising SEQ ID NO:80.

It is well known in the art that the CDR3 domain, independently from the CDR1
25 and/or CDR2 domain(s), alone can determine the binding specificity of an antibody for a
cognate antigen and that multiple antibodies can predictably be generated having the
same binding specificity based on a common CDR3 sequence. *See, for example, Klimka
et al., British J. of Cancer* 83(2):252-260 (2000) (describing the production of a
humanized anti-CD30 antibody using only the heavy chain variable domain CDR3 of
30 murine anti-CD30 antibody Ki-4); Beiboer *et al., J. Mol. Biol.* 296:833-849 (2000)
(describing recombinant epithelial glycoprotein-2 (EGP-2) antibodies using only the
heavy chain CDR3 sequence of the parental murine MOC-31 anti-EGP-2 antibody);
Rader *et al., Proc. Natl. Acad. Sci. U.S.A.* 95:8910-8915 (1998) (describing a panel of

humanized anti-integrin $\alpha_v\beta_3$ antibodies using a heavy and light chain variable CDR3 domain of a murine anti-integrin $\alpha_v\beta_3$ antibody LM609 wherein each member antibody comprises a distinct sequence outside the CDR3 domain and capable of binding the same epitope as the parent murine antibody with affinities as high or higher than the parent murine antibody); Barbas *et al.*, *J. Am. Chem. Soc.* 116:2161-2162 (1994) (disclosing that the CDR3 domain provides the most significant contribution to antigen binding); Barbas *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 92:2529-2533 (1995) (describing the grafting of heavy chain CDR3 sequences of three Fabs (SI-1, SI-40, and SI-32) against human placental DNA onto the heavy chain of an anti-tetanus toxoid Fab thereby replacing the existing heavy chain CDR3 and demonstrating that the CDR3 domain alone conferred binding specificity); and Ditzel *et al.*, *J. Immunol.* 157:739-749 (1996) (describing grafting studies wherein transfer of only the heavy chain CDR3 of a parent polyspecific Fab LNA3 to a heavy chain of a monospecific IgG tetanus toxoid-binding Fab p313 antibody was sufficient to retain binding specificity of the parent Fab). Each of these references is hereby incorporated by reference in its entirety.

Accordingly, within certain aspects, the present invention provides monoclonal antibodies comprising one or more heavy and/or light chain CDR3 domain from a non-human antibody, such as a mouse or rat antibody, wherein the monoclonal antibody is capable of specifically binding to PD-L1. Within some embodiments, such inventive antibodies comprising one or more heavy and/or light chain CDR3 domain from a non-human antibody (a) are capable of competing for binding with; (b) retain the functional characteristics; (c) bind to the same epitope; and/or (d) have a similar binding affinity as the corresponding parental non-human antibody.

Within other aspects, the present invention provides monoclonal antibodies comprising one or more heavy and/or light chain CDR3 domain from a first human antibody, such as, for example, a human antibody obtained from a non-human animal, wherein the first human antibody is capable of specifically binding to PD-L1 and wherein the CDR3 domain from the first human antibody replaces a CDR3 domain in a human antibody that is lacking binding specificity for PD-L1 to generate a second human antibody that is capable of specifically binding to PD-L1. Within some embodiments, antibodies of the instant disclosure comprising one or more heavy and/or light chain CDR3 domain from the first human antibody (a) are capable of competing for

binding with; (b) retain the functional characteristics; (c) bind to the same epitope; and/or (d) have a similar binding affinity as the corresponding parental first human antibody.

Antibodies Having Particular Germline Sequences

5 In certain embodiments, an antibody of the invention comprises a heavy chain variable region from a particular germline heavy chain immunoglobulin gene and/or a light chain variable region from a particular germline light chain immunoglobulin gene.

For example, in a preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising a heavy chain variable region that is the product of or derived from a human V_H 1-18 gene, wherein the
10 antibody specifically binds PD-L1. In another preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising a heavy chain variable region that is the product of or derived from a human V_H 1-69 gene, wherein the antibody specifically binds PD-L1. In another preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-
15 binding portion thereof, comprising a heavy chain variable region that is the product of or derived from a human V_H 1-3 gene, wherein the antibody specifically binds PD-L1. In another preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising a heavy chain variable region that is the product of or derived from a human V_H 3-9 gene, wherein the antibody specifically binds
20 PD-L1. In yet another preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising a light chain variable region that is the product of or derived from a human V_K L6 gene, wherein the antibody specifically binds PD-L1. In yet another preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof,
25 comprising a light chain variable region that is the product of or derived from a human V_K L15 gene, wherein the antibody specifically binds PD-L1. In yet another preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising a light chain variable region that is the product of or derived from a human V_K A27 gene, wherein the antibody specifically binds PD-L1. In
30 yet another preferred embodiment, the invention provides an isolated monoclonal antibody, or an antigen-binding portion thereof, comprising a light chain variable region that is the product of or derived from a human V_K L18 gene, wherein the antibody

specifically binds PD-L1. In yet another preferred embodiment, the invention provides an isolated monoclonal antibody, or antigen-binding portion thereof, wherein the antibody:

(a) comprises a heavy chain variable region that is the product of or derived from a human V_H 1-18, 1-69, 1-3 or 3-9 gene (which encodes the amino acid sequences set forth in SEQ ID NOs:101, 102, 103 and 104, respectively);

(b) comprises a light chain variable region that is the product of or derived from a human V_K L6, L15, A27 or L18 gene (which encodes the amino acid sequences set forth in SEQ ID NOs:105, 106, 107 and 108, respectively); and

(c) specifically binds to PD-L1, preferably human PD-L1.

10 An example of an antibody having V_H and V_K of V_H 1-18 and V_K L6, respectively, is 3G10. Examples of antibodies having V_H and V_K of V_H 1-69 and V_K L6, respectively, 12A4, 1B12, 7H1, and 12B7. An example of an antibody having V_H and V_K of V_H 1-3 and V_K L15, respectively, is 10A5. Examples of antibodies having V_H and V_K of V_H 1-69 and V_K A27, respectively, are 5F8, 11E6 and 11E6a. An example of an antibody having V_H and V_K of V_H 3-9 and V_K L15, respectively, is 10H10. An example of an antibody having V_H and V_K of V_H 1-3 and V_K L15, respectively, is 10A5. An example of an antibody having V_H and V_K of V_H 3-9 and V_K L18, respectively, is 13G4.

As used herein, a human antibody comprises heavy or light chain variable regions that is "the product of" or "derived from" a particular germline sequence if the variable regions of the antibody are obtained from a system that uses human germline immunoglobulin genes. Such systems include immunizing a transgenic mouse carrying human immunoglobulin genes with the antigen of interest or screening a human immunoglobulin gene library displayed on phage with the antigen of interest. A human antibody that is "the product of" or "derived from" a human germline immunoglobulin sequence can be identified as such by comparing the amino acid sequence of the human antibody to the amino acid sequences of human germline immunoglobulins and selecting the human germline immunoglobulin sequence that is closest in sequence (*i.e.*, greatest % identity) to the sequence of the human antibody. A human antibody that is "the product of" or "derived from" a particular human germline immunoglobulin sequence may contain amino acid differences as compared to the germline sequence, due to, for example, naturally-occurring somatic mutations or intentional introduction of site-directed mutation. However, a selected human antibody is generally at least 90% identical in amino acids sequence to an amino acid sequence encoded by a human

germline immunoglobulin gene and contains amino acid residues that identify the human antibody as being human when compared to the germline immunoglobulin amino acid sequences of other species (e.g., murine germline sequences). In certain cases, a human antibody may be at least 95%, or even at least 96%, 97%, 98%, or 99% identical in amino acid sequence to the amino acid sequence encoded by the germline immunoglobulin gene. In certain embodiments, a human antibody derived from a particular human germline sequence will display no more than 10 amino acid differences from the amino acid sequence encoded by the human germline immunoglobulin gene. In certain other embodiments, the human antibody may display no more than 5, or even no more than 4, 3, 2, or 1 amino acid difference from the amino acid sequence encoded by the germline immunoglobulin gene.

Homologous Antibodies

In yet another embodiment, an antibody of the invention comprises heavy and light chain variable regions comprising amino acid sequences that are homologous to the amino acid sequences of the preferred antibodies described herein, and wherein the antibodies retain the desired functional properties of the anti-PD-L1 antibodies of the invention.

For example, the invention provides an isolated monoclonal antibody, or antigen binding portion thereof, comprising a heavy chain variable region and a light chain variable region, wherein:

- (a) the heavy chain variable region comprises an amino acid sequence that is at least 80% homologous to an amino acid sequence selected from the group consisting of SEQ ID NOs:1, 2, 3, 4, 5, 6, 7, 8, 9, and 10;
- (b) the light chain variable region comprises an amino acid sequence that is at least 80% homologous to an amino acid sequence selected from the group consisting of SEQ ID NOs:11, 12, 13, 14, 15, 16, 17, 18, 19, and 20;
- (c) the antibody binds to human PD-L1 with a K_D of 1×10^{-7} M or less;
- (d) the antibody increases T-cell proliferation in a mixed lymphocyte reaction (MLR) assay;
- (e) the antibody increases interferon- γ production in an MLR assay;
- (f) the antibody increases IL-2 secretion in an MLR assay;
- (g) the antibody stimulates antibody responses; and

(h) reverses the effect of T regulatory cells on T cell effector cells and /or dendritic cells.

In other embodiments, the V_H and/or V_L amino acid sequences may be 85%, 90%, 95%, 96%, 97%, 98% or 99% homologous to the sequences set forth above. An antibody
5 having V_H and V_L regions having high (*i.e.*, 80% or greater) homology to the V_H and V_L regions of the sequences set forth above, can be obtained by mutagenesis (*e.g.*, site-directed or PCR-mediated mutagenesis) of nucleic acid molecules encoding SEQ ID NOs:25, 26, 27, 28, 29, and 30, followed by testing of the encoded altered antibody for retained function (*i.e.*, the functions set forth in (c) through (h) above) using the
10 functional assays described herein.

As used herein, the percent homology between two amino acid sequences is equivalent to the percent identity between the two sequences. The percent identity between the two sequences is a function of the number of identical positions shared by the sequences (*i.e.*, % homology = # of identical positions/total # of positions x 100),
15 taking into account the number of gaps, and the length of each gap, which need to be introduced for optimal alignment of the two sequences. The comparison of sequences and determination of percent identity between two sequences can be accomplished using a mathematical algorithm, as described in the non-limiting examples below.

The percent identity between two amino acid sequences can be determined using
20 the algorithm of E. Meyers and W. Miller (*Comput. Appl. Biosci.*, 4:11-17 (1988)) which has been incorporated into the ALIGN program (version 2.0), using a PAM120 weight residue table, a gap length penalty of 12 and a gap penalty of 4. In addition, the percent identity between two amino acid sequences can be determined using the Needleman and Wunsch (*J. Mol. Biol.* 48:444-453 (1970)) algorithm which has been incorporated into the
25 GAP program in the GCG software package (available at www.gcg.com), using either a Blossum 62 matrix or a PAM250 matrix, and a gap weight of 16, 14, 12, 10, 8, 6, or 4 and a length weight of 1, 2, 3, 4, 5, or 6.

In certain instances, the protein sequences of the present disclosure can be further used as a "query sequence" to perform a search against public databases to, for example,
30 identify related sequences. Such searches can be performed using the XBLAST program (version 2.0) of Altschul, *et al.* (1990) *J. Mol. Biol.* 215:403-10. BLAST protein searches can be performed with the XBLAST program, score = 50, wordlength = 3 to obtain amino acid sequences homologous to the antibody molecules of the invention. To obtain gapped

alignments for comparison purposes, Gapped BLAST can be utilized as described in Altschul *et al.*, (1997) *Nucleic Acids Res.* 25(17):3389-3402. When utilizing BLAST and Gapped BLAST programs, the default parameters of the respective programs (*e.g.*, XBLAST and NBLAST) can be used. See www.ncbi.nlm.nih.gov.

5 Antibodies with Conservative Modifications

 In certain embodiments, an antibody of the invention comprises a heavy chain variable region comprising CDR1, CDR2 and CDR3 sequences and a light chain variable region comprising CDR1, CDR2 and CDR3 sequences, wherein one or more of these CDR sequences comprise specified amino acid sequences based on the preferred
10 antibodies described herein (*e.g.*, 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7 or 13G4), or conservative modifications thereof, and wherein the antibodies retain the desired functional properties of the anti-PD-L1 antibodies of the invention. Accordingly, the invention provides an isolated monoclonal antibody, or antigen binding portion thereof, comprising a heavy chain variable region comprising CDR1, CDR2, and
15 CDR3 sequences and a light chain variable region comprising CDR1, CDR2, and CDR3 sequences, wherein:

- (a) the heavy chain variable region CDR3 sequence comprises an amino acid sequence selected from the group consisting of amino acid sequences of SEQ ID NOs:41, 42, 43, 44, 45, 46, 47, 48, 49, and 50, and conservative modifications thereof;
- 20 (b) the light chain variable region CDR3 sequence comprises an amino acid sequence selected from the group consisting of amino acid sequence of SEQ ID NOs:71, 72, 73, 74, 75, 76, 77, 78, 79, and 80, and conservative modifications thereof;
- (c) the antibody binds to human PD-L1 with a K_D of 1×10^{-7} M or less;
- (d) the antibody increases T-cell proliferation in a mixed lymphocyte reaction
25 (MLR) assay;
- (e) the antibody increases interferon- γ production in an MLR assay;
- (f) the antibody increases IL-2 secretion in an MLR assay
- (g) the antibody stimulates antibody responses; and
- (h) reverses the effect of T regulatory cells on T cell effector cells and /or
30 dendritic cells.

 In a preferred embodiment, the heavy chain variable region CDR2 sequence comprises an amino acid sequence selected from the group consisting of amino acid

sequences of SEQ ID NOs:31, 32, 33, 34, 35, 36, 37, 38, 39, and 40, and conservative modifications thereof; and the light chain variable region CDR2 sequence comprises an amino acid sequence selected from the group consisting of amino acid sequences of SEQ ID NOs:61, 62, 63, 64, 65, 66, 67, 68, 69, and 70, and conservative modifications thereof.

- 5 In another preferred embodiment, the heavy chain variable region CDR1 sequence comprises an amino acid sequence selected from the group consisting of amino acid sequences of SEQ ID NOs:21, 22, 23, 24, 25, 26, 27, 28, 29, and 30, and conservative modifications thereof; and the light chain variable region CDR1 sequence comprises an amino acid sequence selected from the group consisting of amino acid sequences of SEQ
10 ID NOs:51, 52, 53, 54, 55, 56, 57, 58, 59, and 60, and conservative modifications thereof.

As used herein, the term "conservative sequence modifications" is intended to refer to amino acid modifications that do not significantly affect or alter the binding characteristics of the antibody containing the amino acid sequence. Such conservative modifications include amino acid substitutions, additions and deletions. Modifications
15 can be introduced into an antibody of the invention by standard techniques known in the art, such as site-directed mutagenesis and PCR-mediated mutagenesis. Conservative amino acid substitutions are ones in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined in the art. These families include amino acids with
20 basic side chains (*e.g.*, lysine, arginine, histidine), acidic side chains (*e.g.*, aspartic acid, glutamic acid), uncharged polar side chains (*e.g.*, glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine, tryptophan), nonpolar side chains (*e.g.*, alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine), beta-branched side chains (*e.g.*, threonine, valine, isoleucine) and aromatic side chains (*e.g.*, tyrosine, phenylalanine,
25 tryptophan, histidine). Thus, one or more amino acid residues within the CDR regions of an antibody of the invention can be replaced with other amino acid residues from the same side chain family and the altered antibody can be tested for retained function (*i.e.*, the functions set forth in (c) through (h) above) using the functional assays described herein.

30 Antibodies that Bind to the Same Epitope as Anti-PD-L1 Antibodies of the Invention

In another embodiment, the invention provides antibodies that bind to the same epitope on human PD-L1 as any of the PD-L1 monoclonal antibodies of the invention

(i.e., antibodies that have the ability to cross-compete for binding to PD-L1 with any of the monoclonal antibodies of the invention). In preferred embodiments, the reference antibody for cross-competition studies can be the monoclonal antibody 3G10 (having V_H and V_L sequences as shown in SEQ ID NOs:1 and 11, respectively), or the monoclonal antibody 12A4 (having V_H and V_L sequences as shown in SEQ ID NOs:2 and 12, respectively), or the monoclonal antibody 10A5 (having V_H and V_L sequences as shown in SEQ ID NOs:3 and 13, respectively), or the monoclonal antibody 10A5 (having V_H and V_L sequences as shown in SEQ ID NOs:3 and 13, respectively), or the monoclonal antibody 5F8 (having V_H and V_L sequences as shown in SEQ ID NOs:4 and 14, respectively), or the monoclonal antibody 10H10 (having V_H and V_L sequences as shown in SEQ ID NOs:5 and 15, respectively), or the monoclonal antibody 1B12 (having V_H and V_L sequences as shown in SEQ ID NOs:6 and 16, respectively), or the monoclonal antibody 7H1 (having V_H and V_L sequences as shown in SEQ ID NOs:7 and 17, respectively), or the monoclonal antibody 11E6 (having V_H and V_L sequences as shown in SEQ ID NOs:8 and 18, respectively), or the monoclonal antibody 12B7 (having V_H and V_L sequences as shown in SEQ ID NOs:9 and 19, respectively), or the monoclonal antibody 13G4 (having V_H and V_L sequences as shown in SEQ ID NOs:10 and 20, respectively). Such cross-competing antibodies can be identified based on their ability to cross-compete with 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7 or 13G4 in standard PD-L1 binding assays. For example, BIAcore analysis, ELISA assays or flow cytometry may be used to demonstrate cross-competition with the antibodies of the current invention. The ability of a test antibody to inhibit the binding of, for example, 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7 or 13G4, to human PD-L1 demonstrates that the test antibody can compete with 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7 or 13G4 for binding to human PD-L1 and thus binds to the same epitope on human PD-L1 as 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7 or 13G4. In a preferred embodiment, the antibody that binds to the same epitope on human PD-L1 as 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7 or 13G4 is a human monoclonal antibody. Such human monoclonal antibodies can be prepared and isolated as described in the Examples.

Engineered and Modified Antibodies

An antibody of the invention further can be prepared using an antibody having one or more of the V_H and/or V_L sequences disclosed herein as starting material to engineer a modified antibody, which modified antibody may have altered properties from the starting antibody. An antibody can be engineered by modifying one or more residues within one or both variable regions (*i.e.*, V_H and/or V_L), for example within one or more CDR regions and/or within one or more framework regions. Additionally or alternatively, an antibody can be engineered by modifying residues within the constant region(s), for example to alter the effector function(s) of the antibody.

One type of variable region engineering that can be performed is CDR grafting. Antibodies interact with target antigens predominantly through amino acid residues that are located in the six heavy and light chain complementarity determining regions (CDRs). For this reason, the amino acid sequences within CDRs are more diverse between individual antibodies than sequences outside of CDRs. Because CDR sequences are responsible for most antibody-antigen interactions, it is possible to express recombinant antibodies that mimic the properties of specific naturally occurring antibodies by constructing expression vectors that include CDR sequences from the specific naturally occurring antibody grafted onto framework sequences from a different antibody with different properties (see, *e.g.*, Riechmann, L. *et al.* (1998) *Nature* 332:323-327; Jones, P. *et al.* (1986) *Nature* 321:522-525; Queen, C. *et al.* (1989) *Proc. Natl. Acad. Sci. U.S.A.* 86:10029-10033; U.S. Patent No. 5,225,539 to Winter, and U.S. Patent Nos. 5,530,101; 5,585,089; 5,693,762 and 6,180,370 to Queen *et al.*)

Accordingly, another embodiment of the invention pertains to an isolated monoclonal antibody, or antigen binding portion thereof, comprising a heavy chain variable region comprising CDR1, CDR2, and CDR3 sequences comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:21, 22, 23, 24, 25, 26, 27, 28, 29, and 30, SEQ ID NOs:31, 32, 33, 34, 35, 36, 37, 38, 39, and 40, and SEQ ID NOs:41, 42, 43, 44, 45, 46, 47, 48, 49, and 50, respectively, and a light chain variable region comprising CDR1, CDR2, and CDR3 sequences comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:51, 52, 53, 54, 55, 56, 57, 58, 59, and 60, SEQ ID NOs:61, 62, 63, 64, 65, 66, 67, 68, 69, and 70, and SEQ ID NOs:71, 72, 73, 74, 75, 76, 77, 78, 79, and 80, respectively. Thus, such antibodies contain the V_H and V_L CDR sequences of monoclonal antibodies 3G10, 12A4, 10A5,

5F8, 10H10, 1B12, 7H1, 11E6, 12B7 or 13G4 yet may contain different framework sequences from these antibodies.

Such framework sequences can be obtained from public DNA databases or published references that include germline antibody gene sequences. For example, 5 germline DNA sequences for human heavy and light chain variable region genes can be found in the "VBase" human germline sequence database (available on the Internet at www.mrc-cpe.cam.ac.uk/vbase), as well as in Kabat, E. A., *et al.* (1991) Sequences of Proteins of Immunological Interest, Fifth Edition, U.S. Department of Health and Human Services, NIH Publication No. 91-3242; Tomlinson, I. M., *et al.* (1992) "The Repertoire of Human Germline V_H Sequences Reveals about Fifty Groups of V_H Segments with Different Hypervariable Loops" *J. Mol. Biol.* 227:776-798; and Cox, J. P. L. *et al.* (1994) "A Directory of Human Germ-line V_H Segments Reveals a Strong Bias in their Usage" *Eur. J. Immunol.* 24:827-836; the contents of each of which are expressly incorporated herein by reference.

15 Antibody protein sequences are compared against a compiled protein sequence database using one of the sequence similarity searching methods called the Gapped BLAST (Altschul *et al.* (1997) *Nucleic Acids Research* 25:3389-3402), which is well known to those skilled in the art. BLAST is a heuristic algorithm in that a statistically significant alignment between the antibody sequence and the database sequence is likely 20 to contain high-scoring segment pairs (HSP) of aligned words. Segment pairs whose scores cannot be improved by extension or trimming is called a hit. Briefly, the nucleotide sequences of VBASE origin (vbase.mrc-cpe.cam.ac.uk/vbase1/list2.php) are translated and the region between and including FR1 through FR3 framework region is retained. The database sequences have an average length of 98 residues. Duplicate 25 sequences which are exact matches over the entire length of the protein are removed. A BLAST search for proteins using the program blastp with default, standard parameters except the low complexity filter which is turned off and the substitution matrix of BLOSUM62, filters for top 5 hits yielding sequence matches. The nucleotide sequences are translated in all six frames and the frame with no stop codons in the matching segment 30 of the database sequence is considered the potential hit. This is in turn confirmed using the BLAST program tblastx. This translates the antibody sequence in all six frames and compares those translations to the VBASE nucleotide sequences dynamically translated in all six frames.

The identities are exact amino acid matches between the antibody sequence and the protein database over the entire length of the sequence. The positives (identities + substitution match) are not identical but amino acid substitutions guided by the BLOSUM62 substitution matrix. If the antibody sequence matches two of the database sequences with same identity, the hit with most positives would be decided to be the matching sequence hit.

Preferred framework sequences for use in the antibodies of the invention are those that are structurally similar to the framework sequences used by selected antibodies of the invention, *e.g.*, similar to the V_H 1-18 framework sequences (SEQ ID NO:101) and/or the V_H 1-69 framework sequences (SEQ ID NO:102) and/or the V_H 1-3 framework sequences (SEQ ID NO:103) and/or the V_H 3-9 framework sequences (SEQ ID NO:104) and/or the V_K L6 framework sequences (SEQ ID NO:105) and/or the V_K L15 framework sequences (SEQ ID NO:106) and/or the V_K A27 framework sequences (SEQ ID NO:107) and/or the V_K L18 framework sequences (SEQ ID NO:107) used by preferred monoclonal antibodies of the invention. The V_H CDR1, CDR2, and CDR3 sequences, and the V_K CDR1, CDR2, and CDR3 sequences, can be grafted onto framework regions that have the identical sequence as that found in the germline immunoglobulin gene from which the framework sequence derive, or the CDR sequences can be grafted onto framework regions that contain one or more mutations as compared to the germline sequences. For example, it has been found that in certain instances it is beneficial to mutate residues within the framework regions to maintain or enhance the antigen binding ability of the antibody (see *e.g.*, U.S. Patent Nos. 5,530,101; 5,585,089; 5,693,762 and 6,180,370 to Queen *et al*).

Another type of variable region modification is to mutate amino acid residues within the V_H and/or V_K CDR1, CDR2 and/or CDR3 regions to thereby improve one or more binding properties (*e.g.*, affinity) of the antibody of interest. Site-directed mutagenesis or PCR-mediated mutagenesis can be performed to introduce the mutation(s) and the effect on antibody binding, or other functional property of interest, can be evaluated in *in vitro* or *in vivo* assays as described herein and provided in the Examples. Preferably conservative modifications (as discussed above) are introduced. The mutations may be amino acid substitutions, additions or deletions, but are preferably substitutions. Moreover, typically no more than one, two, three, four or five residues within a CDR region are altered.

Accordingly, in another embodiment, the invention provides isolated anti-PD-L1 monoclonal antibodies, or antigen binding portions thereof, comprising a heavy chain variable region comprising: (a) a V_H CDR1 region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:21, 22, 23, 24, 25, 26, 27, 28, 29, and 30, or an amino acid sequence having one, two, three, four or five amino acid substitutions, deletions or additions as compared to SEQ ID NOs:21, 22, 23, 24, 25, 26, 27, 28, 29, and 30; (b) a V_H CDR2 region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:31, 32, 33, 34, 35, 36, 37, 38, 39, and 40, or an amino acid sequence having one, two, three, four or five amino acid substitutions, deletions or additions as compared to SEQ ID NOs:31, 32, 33, 34, 35, 36, 37, 38, 39, and 40; (c) a V_H CDR3 region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:41, 42, 43, 44, 45, 46, 47, 48, 49, and 50, or an amino acid sequence having one, two, three, four or five amino acid substitutions, deletions or additions as compared to SEQ ID NOs:41, 42, 43, 44, 45, 46, 47, 48, 49, and 50; (d) a V_K CDR1 region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:51, 52, 53, 54, 55, 56, 57, 58, 59, and 60, or an amino acid sequence having one, two, three, four or five amino acid substitutions, deletions or additions as compared to SEQ ID NOs:51, 52, 53, 54, 55, 56, 57, 58, 59, and 60; (e) a V_K CDR2 region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:61, 62, 63, 64, 65, 66, 67, 68, 69, and 70, or an amino acid sequence having one, two, three, four or five amino acid substitutions, deletions or additions as compared to SEQ ID NOs:61, 62, 63, 64, 65, 66, 67, 68, 69, and 70; and (f) a V_K CDR3 region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:71, 72, 73, 74, 75, 76, 77, 78, 79, and 80, or an amino acid sequence having one, two, three, four or five amino acid substitutions, deletions or additions as compared to SEQ ID NOs:71, 72, 73, 74, 75, 76, 77, 78, 79, and 80.

Engineered antibodies of the invention include those in which modifications have been made to framework residues within V_H and/or V_K, *e.g.* to improve the properties of the antibody. Typically such framework modifications are made to decrease the immunogenicity of the antibody. For example, one approach is to "backmutate" one or more framework residues to the corresponding germline sequence. More specifically, an antibody that has undergone somatic mutation may contain framework residues that differ from the germline sequence from which the antibody is derived. Such residues can be

identified by comparing the antibody framework sequences to the germline sequences from which the antibody is derived. For example, as described below, a number of amino acid changes in the framework regions of the anti-PD-L1 antibodies 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7 and 13G4 that differ from the parent germline sequence. To return the framework region sequences to their germline configuration, the somatic mutations can be "backmutated" to the germline sequence by, for example, site-directed mutagenesis or PCR-mediated mutagenesis. The alignment of the V_H region for 3G10 against the parent germline V_H 1-18 sequence is shown in Figure 11. The alignment of the V_H region for 12A4 against the parent germline V_H 1-69 sequence is shown in Figure 12. The alignment of the V_H region for 10A5 against the parent germline V_H 1-3 sequence is shown in Figure 13. The alignment of the V_H region for 5F8 against the parent germline V_H 1-69 sequence is shown in Figure 14. The alignment of the V_H region for 10H10 against the parent germline V_H 3-9 sequence is shown in Figure 15. The alignment of the V_H region for 1B12 against the parent germline V_H 1-69 sequence is shown in Figure 16. The alignment of the V_H region for 7H1 against the parent germline V_H 1-69 sequence is shown in Figure 17. The alignment of the V_H region for 11E6 against the parent germline V_H 1-69 sequence is shown in Figure 18. The alignment of the V_H region for 12B7 against the parent germline V_H 1-69 sequence is shown in Figure 19. The alignment of the V_H region for 13G4 against the parent germline V_H 3-9 sequence is shown in Figure 20.

For example, for 3G10, amino acid residue #79 (within FR3) of V_H is a valine whereas this residue in the corresponding V_H 1-18 germline sequence is an alanine. To return the framework region sequences to their germline configuration, the somatic mutations can be "backmutated" to the germline sequence by, for example, site-directed mutagenesis or PCR-mediated mutagenesis (e.g., residue #79 (residue #13 of FR3) of the V_H of 3G10 can be "backmutated" from valine to alanine).

As another example, for 12A4, amino acid residue #24 (within FR1) of V_H is a threonine whereas this residue in the corresponding V_H 1-69 germline sequence is an alanine. To return the framework region sequences to their germline configuration, for example, residue #24 of the V_H of 12A4 can be "backmutated" from threonine to alanine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

As another example, for 12A4, amino acid residue #27 (within FR1) of V_H is an aspartic acid whereas this residue in the corresponding V_H 1-69 germline sequence is a

glycine. To return the framework region sequences to their germline configuration, for example, residue #27 of the V_H of 12A4 can be "backmutated" from aspartic acid to glycine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

5 As another example, for 12A4, amino acid residue #95 (within FR3) of V_H is a phenylalanine whereas this residue in the corresponding V_H 1-69 germline sequence is a tyrosine. To return the framework region sequences to their germline configuration, for example, residue #95 (residue #29 of FR3) of the V_H of 12A4 can be "backmutated" from phenylalanine to tyrosine. Such "backmutated" antibodies are also intended to be
10 encompassed by the invention.

 As another example, for 5F8, amino acid residue #24 (within FR1) is a valine whereas this residue in the corresponding V_H 1-69 germline sequence is an alanine. To return the framework region sequences to their germline configuration, for example, residue #24 of the V_H of 5F8 can be "backmutated" from valine to alanine. Such
15 "backmutated" antibodies are also intended to be encompassed by the invention.

 As another example, for 5F8, amino acid residue #28 (within FR1) is an isoleucine whereas this residue in the corresponding V_H 1-69 germline sequence is an threonine. To return the framework region sequences to their germline configuration, for example, residue #28 of the V_H of 5F8 can be "backmutated" from isoleucine to
20 threonine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

 As another example, for 10H10, amino acid residue #24 (within FR1) is a valine whereas this residue in the corresponding V_H 3-9 germline sequence is an alanine. To return the framework region sequences to their germline configuration, for example, residue #24 of the V_H of 10H10 can be "backmutated" from valine to alanine. Such
25 "backmutated" antibodies are also intended to be encompassed by the invention.

 As another example, for 10H10, an amino acid can be inserted following amino acid residue #97 (within FR3). This amino acid is a valine. To return the framework region sequences to their germline configuration, for example, the inserted amino acid following residue #97 of the V_H of 10H10 can be "backmutated" to delete this valine.
30 Such "backmutated" antibodies are also intended to be encompassed by the invention.

 As another example, for 1B12, amino acid residue #24 (within FR1) is a threonine whereas this residue in the corresponding V_H 1-69 germline sequence is an alanine. To

return the framework region sequences to their germline configuration, for example, residue #24 of the V_H of 1B12 can be "backmutated" from threonine to alanine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

5 As another example, for 1B12, amino acid residue #27 (within FR1) is an aspartic acid whereas this residue in the corresponding V_H 1-69 germline sequence is a glycine. To return the framework region sequences to their germline configuration, for example, residue #27 of the V_H of 1B12 can be "backmutated" from aspartic acid to glycine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

10 As another example, for 1B12, amino acid residue #95 (within FR3) is a phenylalanine whereas this residue in the corresponding V_H 1-69 germline sequence is a tyrosine. To return the framework region sequences to their germline configuration, for example, residue #95 (residue #29 of FR3) of the V_H of 1B12 can be "backmutated" from phenylalanine to tyrosine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

15 As another example, for 7H1, amino acid residue #24 (within FR1) is a threonine whereas this residue in the corresponding V_H 1-69 germline sequence is an alanine. To return the framework region sequences to their germline configuration, for example, residue #24 of the V_H of 7H1 can be "backmutated" from threonine to alanine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

20 As another example, for 7H1, amino acid residue #77 (within FR3) is a threonine whereas this residue in the corresponding V_H 1-69 germline sequence is a serine. To return the framework region sequences to their germline configuration, for example, residue #72 (residue #11 of FR3) of the V_H of 7H1 can be "backmutated" from threonine to serine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

25 As another example, for 11E6, amino acid residue #78 (within FR3) is an alanine whereas this residue in the corresponding V_H 1-69 germline sequence is a threonine. To return the framework region sequences to their germline configuration, for example, residue #78 (residue 12 of FR3) of the V_H of 11E6 can be "backmutated" from alanine to threonine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

30 As another example, for 12B7, amino acid residue #13 (within FR1) is a glutamic acid whereas this residue in the corresponding V_H 1-69 germline sequence is a lysine. To

return the framework region sequences to their germline configuration, for example, residue #13 of the V_H of 12B7 can be "backmutated" glutamic acid to lysine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

5 As another example, for 12B7, amino acid residue #30 (within FR1) is an asparagine whereas this residue in the corresponding V_H 1-69 germline sequence is an serine. To return the framework region sequences to their germline configuration, for example, residue #30 of the V_H of 12B7 can be "backmutated" from asparagine to serine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

10 As another example, for 12B7, amino acid residue #77 (within FR3) is an asparagine whereas this residue in the corresponding V_H 1-69 germline sequence is an serine. To return the framework region sequences to their germline configuration, for example, residue #377 (residue 11 of FR3) of the V_H of 12B7 can be "backmutated" from asparagine to serine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

15 As another example, for 12B7, amino acid residue #82 (within FR3) is an aspartic acid whereas this residue in the corresponding V_H 1-69 germline sequence is a glutamic acid. To return the framework region sequences to their germline configuration, for example, residue #82 (residue #16 of FR3) of the V_H of 12B7 can be "backmutated" from aspartic acid to glutamic acid. Such "backmutated" antibodies are also intended to be encompassed by the invention.

20 As another example, for 13G4, amino acid residue #27 (within FR1) is an isoleucine whereas this residue in the corresponding V_H 1-69 germline sequence is an phenylalanine. To return the framework region sequences to their germline configuration, for example, residue #27 of the V_H of 12B7 can be "backmutated" from isoleucine to phenylalanine. Such "backmutated" antibodies are also intended to be encompassed by the invention.

30 Another type of framework modification involves mutating one or more residues within the framework region, or even within one or more CDR regions, to remove T cell epitopes to thereby reduce the potential immunogenicity of the antibody. This approach is also referred to as "deimmunization" and is described in further detail in U.S. Patent Publication No. 20030153043 by Carr *et al.*

In addition or alternative to modifications made within the framework or CDR regions, antibodies of the invention may be engineered to include modifications within

the Fc region, typically to alter one or more functional properties of the antibody, such as serum half-life, complement fixation, Fc receptor binding, and/or antigen-dependent cellular cytotoxicity. Furthermore, an antibody of the invention may be chemically modified (e.g., one or more chemical moieties can be attached to the antibody) or be
5 modified to alter its glycosylation, again to alter one or more functional properties of the antibody. Each of these embodiments is described in further detail below. The numbering of residues in the Fc region is that of the EU index of Kabat.

In one embodiment, the hinge region of CH1 is modified such that the number of cysteine residues in the hinge region is altered, e.g., increased or decreased. This
10 approach is described further in U.S. Patent No. 5,677,425 by Bodmer *et al.* The number of cysteine residues in the hinge region of CH1 is altered to, for example, facilitate assembly of the light and heavy chains or to increase or decrease the stability of the antibody.

In another embodiment, the Fc hinge region of an antibody is mutated to decrease
15 the biological half life of the antibody. More specifically, one or more amino acid mutations are introduced into the CH2-CH3 domain interface region of the Fc-hinge fragment such that the antibody has impaired Staphylococcal protein A (SpA) binding relative to native Fc-hinge domain SpA binding. This approach is described in further detail in U.S. Patent No. 6,165,745 by Ward *et al.*

In another embodiment, the antibody is modified to increase its biological half
20 life. Various approaches are possible. For example, one or more of the following mutations can be introduced: T252L, T254S, T256F, as described in U.S. Patent No. 6,277,375 to Ward. Alternatively, to increase the biological half life, the antibody can be altered within the CH1 or CL region to contain a salvage receptor binding epitope taken
25 from two loops of a CH2 domain of an Fc region of an IgG, as described in U.S. Patent Nos. 5,869,046 and 6,121,022 by Presta *et al.*

In yet other embodiments, the Fc region is altered by replacing at least one amino acid residue with a different amino acid residue to alter the effector function(s) of the antibody. For example, one or more amino acids selected from amino acid residues 234,
30 235, 236, 237, 297, 318, 320 and 322 can be replaced with a different amino acid residue such that the antibody has an altered affinity for an effector ligand but retains the antigen-binding ability of the parent antibody. The effector ligand to which affinity is altered can be, for example, an Fc receptor or the C1 component of complement. This approach is

described in further detail in U.S. Patent Nos. 5,624,821 and 5,648,260, both by Winter *et al.*

In another example, one or more amino acids selected from amino acid residues 329, 331 and 322 can be replaced with a different amino acid residue such that the antibody has altered C1q binding and/or reduced or abolished complement dependent cytotoxicity (CDC). This approach is described in further detail in U.S. Patent Nos. 6,194,551 by Idusogie *et al.*

In another example, one or more amino acid residues within amino acid positions 231 and 239 are altered to thereby alter the ability of the antibody to fix complement. This approach is described further in PCT Publication WO 94/29351 by Bodmer *et al.*

In yet another example, the Fc region is modified to increase the ability of the antibody to mediate antibody dependent cellular cytotoxicity (ADCC) and/or to increase the affinity of the antibody for an Fcγ receptor by modifying one or more amino acids at the following positions: 238, 239, 248, 249, 252, 254, 255, 256, 258, 265, 267, 268, 269, 270, 272, 276, 278, 280, 283, 285, 286, 289, 290, 292, 293, 294, 295, 296, 298, 301, 303, 305, 307, 309, 312, 315, 320, 322, 324, 326, 327, 329, 330, 331, 333, 334, 335, 337, 338, 340, 360, 373, 376, 378, 382, 388, 389, 398, 414, 416, 419, 430, 434, 435, 437, 438 or 439. This approach is described further in PCT Publication WO 00/42072 by Presta. Moreover, the binding sites on human IgG1 for FcγRI, FcγRII, FcγRIII and FcRn have been mapped and variants with improved binding have been described (see Shields, R.L. *et al.* (2001) *J. Biol. Chem.* 276:6591-6604). Specific mutations at positions 256, 290, 298, 333, 334 and 339 were shown to improve binding to FcγRIII. Additionally, the following combination mutants were shown to improve FcγRIII binding: T256A/S298A, S298A/E333A, S298A/K224A and S298A/E333A/K334A.

In still another embodiment, the glycosylation of an antibody is modified. For example, an aglycosylated antibody can be made (*i.e.*, the antibody lacks glycosylation). Glycosylation can be altered to, for example, increase the affinity of the antibody for antigen. Such carbohydrate modifications can be accomplished by, for example, altering one or more sites of glycosylation within the antibody sequence. For example, one or more amino acid substitutions can be made that result in elimination of one or more variable region framework glycosylation sites to thereby eliminate glycosylation at that site. Such aglycosylation may increase the affinity of the antibody for antigen. Such an

approach is described in further detail in U.S. Patent Nos. 5,714,350 and 6,350,861 by Co *et al.*

In certain other embodiments, an antibody can be made that has an altered type of glycosylation, such as a hypofucosylated antibody having reduced amounts of fucosyl
5 residues or an antibody having increased bisecting GlcNac structures. Such altered glycosylation patterns have been demonstrated to increase the ADCC ability of antibodies. Such carbohydrate modifications can be accomplished by, for example, expressing the antibody in a host cell with altered glycosylation machinery. Cells with altered glycosylation machinery have been described in the art and can be used as host
10 cells in which to express recombinant antibodies of the invention to thereby produce an antibody with altered glycosylation. For example, the cell lines Ms704, Ms705, and Ms709 lack the fucosyltransferase gene, FUT8 (alpha (1,6) fucosyltransferase), such that antibodies expressed in the Ms704, Ms705, and Ms709 cell lines lack fucose on their carbohydrates. The Ms704, Ms705, and Ms709 FUT8^{-/-} cell lines were created by the
15 targeted disruption of the FUT8 gene in CHO/DG44 cells using two replacement vectors (see U.S. Patent Publication No. 20040110704 by Yamane *et al.* and Yamane-Ohnuki *et al.* (2004) *Biotechnol Bioeng* 87:614-22). As another example, EP 1,176,195 by Hanai *et al.* describes a cell line with a functionally disrupted FUT8 gene, which encodes a fucosyl transferase, such that antibodies expressed in such a cell line exhibit hypofucosylation by
20 reducing or eliminating the alpha 1,6 bond-related enzyme. Hanai *et al.* also describe cell lines which have a low enzyme activity for adding fucose to the N-acetylglucosamine that binds to the Fc region of the antibody or does not have the enzyme activity, for example the rat myeloma cell line YB2/0 (ATCC CRL 1662). PCT Publication WO 03/035835 by Presta describes a variant CHO cell line, Lec13 cells, with reduced ability
25 to attach fucose to Asn(297)-linked carbohydrates, also resulting in hypofucosylation of antibodies expressed in that host cell (see also Shields, R.L. *et al.* (2002) *J. Biol. Chem.* 277:26733-26740). PCT Publication WO 99/54342 by Umana *et al.* describes cell lines engineered to express glycoprotein-modifying glycosyl transferases (e.g., beta(1,4)-N-acetylglucosaminyltransferase III (GnTIII)) such that antibodies expressed in the
30 engineered cell lines exhibit increased bisecting GlcNac structures which results in increased ADCC activity of the antibodies (see also Umana *et al.* (1999) *Nat. Biotech.* 17:176-180). Alternatively, the fucose residues of the antibody may be cleaved off using

a fucosidase enzyme. For example, the fucosidase alpha-L-fucosidase removes fucosyl residues from antibodies (Tarentino, A.L. *et al.* (1975) *Biochem.* 14:5516-23).

Another modification of the antibodies herein that is contemplated by the invention is pegylation. An antibody can be pegylated to, for example, increase the biological (*e.g.*, serum) half life of the antibody. To pegylate an antibody, the antibody, or fragment thereof, typically is reacted with polyethylene glycol (PEG), such as a reactive ester or aldehyde derivative of PEG, under conditions in which one or more PEG groups become attached to the antibody or antibody fragment. Preferably, the pegylation is carried out via an acylation reaction or an alkylation reaction with a reactive PEG molecule (or an analogous reactive water-soluble polymer). As used herein, the term "polyethylene glycol" is intended to encompass any of the forms of PEG that have been used to derivatize other proteins, such as mono (C1-C10) alkoxy- or aryloxy-polyethylene glycol or polyethylene glycol-maleimide. In certain embodiments, the antibody to be pegylated is an aglycosylated antibody. Methods for pegylating proteins are known in the art and can be applied to the antibodies of the invention. See for example, EP 0 154 316 by Nishimura *et al.* and EP 0 401 384 by Ishikawa *et al.*

Methods of Engineering Antibodies

As discussed above, the anti-PD-L1 antibodies having V_H and V_K sequences disclosed herein can be used to create new anti-PD-L1 antibodies by modifying the V_H and/or V_K sequences, or the constant region(s) attached thereto. Thus, in another aspect of the invention, the structural features of an anti-PD-L1 antibody of the invention, *e.g.* 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, or 13G4,, are used to create structurally related anti-PD-L1 antibodies that retain at least one functional property of the antibodies of the invention, such as binding to human PD-L1. For example, one or more CDR regions of 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, or 13G4 or mutations thereof, can be combined recombinantly with known framework regions and/or other CDRs to create additional, recombinantly-engineered, anti-PD-L1 antibodies of the invention, as discussed above. Other types of modifications include those described in the previous section. The starting material for the engineering method is one or more of the V_H and/or V_K sequences provided herein, or one or more CDR regions thereof. To create the engineered antibody, it is not necessary to actually prepare (*i.e.*, express as a protein) an antibody having one or more of the V_H and/or V_K sequences

provided herein, or one or more CDR regions thereof. Rather, the information contained in the sequence(s) is used as the starting material to create a "second generation" sequence(s) derived from the original sequence(s) and then the "second generation" sequence(s) is prepared and expressed as a protein.

5 Accordingly, in another embodiment, the invention provides a method for preparing an anti-PD-L1 antibody comprising:

 (a) providing: (i) a heavy chain variable region antibody sequence comprising a CDR1 sequence selected from the group consisting of SEQ ID NOs:21, 22, 23, 24, 25, 26, 27, 28, 29, and 30, a CDR2 sequence selected from the group consisting of SEQ ID
10 NOs:31, 32, 33, 34, 35, 36, 37, 38, 39, and 40, and/or a CDR3 sequence selected from the group consisting of SEQ ID NOs:41, 42, 43, 44, 45, 46, 47, 48, 49, and 50; and/or (ii) a light chain variable region antibody sequence comprising a CDR1 sequence selected from the group consisting of SEQ ID NOs:51, 52, 53, 54, 55, 56, 57, 58, 59, and 60, a CDR2 sequence selected from the group consisting of SEQ ID NOs:61, 62, 63, 64, 65, 66, 67,
15 68, 69, and 70, and/or a CDR3 sequence selected from the group consisting of SEQ ID NOs:71, 72, 73, 74, 75, 76, 77, 78, 79, and 80;

 (b) altering at least one amino acid residue within the heavy chain variable region antibody sequence and/or the light chain variable region antibody sequence to create at least one altered antibody sequence; and

20 (c) expressing the altered antibody sequence as a protein.

 Standard molecular biology techniques can be used to prepare and express the altered antibody sequence.

 Preferably, the antibody encoded by the altered antibody sequence(s) is one that retains one, some or all of the functional properties of the anti-PD-L1 antibodies

25 described herein, which functional properties include, but are not limited to:

 (i) binds to human PD-L1 with a K_D of 1×10^{-7} M or less;

 (ii) increases T-cell proliferation in a mixed lymphocyte reaction (MLR) assay;

 (iii) increases interferon- γ production in an MLR assay;

30 (iv) increases IL-2 secretion in an MLR assay;

 (v) stimulates antibody responses; and/or

 (vi) reverses the effect of T regulatory cells on T cell effector cells and /or dendritic cells.

The functional properties of the altered antibodies can be assessed using standard assays available in the art and/or described herein, such as those set forth in the Examples (e.g., flow cytometry, binding assays).

In certain embodiments of the methods of engineering antibodies of the invention, mutations can be introduced randomly or selectively along all or part of an anti-PD-L1 antibody coding sequence and the resulting modified anti-PD-L1 antibodies can be screened for binding activity and/or other functional properties as described herein. Mutational methods have been described in the art. For example, PCT Publication WO 02/092780 by Short describes methods for creating and screening antibody mutations using saturation mutagenesis, synthetic ligation assembly, or a combination thereof. Alternatively, PCT Publication WO 03/074679 by Lazar *et al.* describes methods of using computational screening methods to optimize physiochemical properties of antibodies.

Nucleic Acid Molecules Encoding Antibodies of the Disclosure

Another aspect of the disclosure pertains to nucleic acid molecules that encode the antibodies of the invention. The nucleic acids may be present in whole cells, in a cell lysate, or in a partially purified or substantially pure form. A nucleic acid is "isolated" or "rendered substantially pure" when purified away from other cellular components or other contaminants, e.g., other cellular nucleic acids or proteins, by standard techniques, including alkaline/SDS treatment, CsCl banding, column chromatography, agarose gel electrophoresis and others well known in the art. See, F. Ausubel, *et al.*, ed. (1987) Current Protocols in Molecular Biology, Greene Publishing and Wiley Interscience, New York. A nucleic acid of the invention can be, for example, DNA or RNA and may or may not contain intronic sequences. In a preferred embodiment, the nucleic acid is a cDNA molecule.

Nucleic acids of the invention can be obtained using standard molecular biology techniques. For antibodies expressed by hybridomas (e.g., hybridomas prepared from transgenic mice carrying human immunoglobulin genes as described further below), cDNAs encoding the light and heavy chains of the antibody made by the hybridoma can be obtained by standard PCR amplification or cDNA cloning techniques. For antibodies obtained from an immunoglobulin gene library (e.g., using phage display techniques), nucleic acid encoding the antibody can be recovered from the library.

Preferred nucleic acids molecules of the invention are those encoding the VH and VL sequences of the 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4, monoclonal antibodies. DNA sequences encoding the VH sequences of 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4, are shown in SEQ ID
5 NOs:81, 82, 83, 84, 85, 86, 87, 88, 89 and 90, respectively. DNA sequences encoding the VL sequences of 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4, are shown in SEQ ID NOs:91, 92, 93, 94, 95, 96, 97, 98, 99 and 100, respectively.

Once DNA fragments encoding VH and VL segments are obtained, these DNA fragments can be further manipulated by standard recombinant DNA techniques, for
10 example to convert the variable region genes to full-length antibody chain genes, to Fab fragment genes or to a scFv gene. In these manipulations, a VL- or VH-encoding DNA fragment is operatively linked to another DNA fragment encoding another protein, such as an antibody constant region or a flexible linker. The term "operatively linked," as used in this context, is intended to mean that the two DNA fragments are joined such that the
15 amino acid sequences encoded by the two DNA fragments remain in-frame.

The isolated DNA encoding the VH region can be converted to a full-length heavy chain gene by operatively linking the VH-encoding DNA to another DNA molecule encoding heavy chain constant regions (CH1, CH2 and CH3). The sequences of human heavy chain constant region genes are known in the art (see *e.g.*, Kabat, E. A., *et al.*
20 (1991) Sequences of Proteins of Immunological Interest, Fifth Edition, U.S. Department of Health and Human Services, NIH Publication No. 91-3242) and DNA fragments encompassing these regions can be obtained by standard PCR amplification. The heavy chain constant region can be an IgG1, IgG2, IgG3, IgG4, IgA, IgE, IgM or IgD constant region, but most preferably is an IgG1 or IgG4 constant region. For a Fab fragment
25 heavy chain gene, the VH-encoding DNA can be operatively linked to another DNA molecule encoding only the heavy chain CH1 constant region.

The isolated DNA encoding the VL region can be converted to a full-length light chain gene (as well as a Fab light chain gene) by operatively linking the VL-encoding DNA to another DNA molecule encoding the light chain constant region, CL. The
30 sequences of human light chain constant region genes are known in the art (see *e.g.*, Kabat, E. A., *et al.* (1991) Sequences of Proteins of Immunological Interest, Fifth Edition, U.S. Department of Health and Human Services, NIH Publication No. 91-3242) and DNA fragments encompassing these regions can be obtained by standard PCR amplification.

The light chain constant region can be a kappa or lambda constant region, but most preferably is a kappa constant region.

To create a scFv gene, the VH- and VL-encoding DNA fragments are operatively linked to another fragment encoding a flexible linker, *e.g.*, encoding the amino acid sequence (Gly₄-Ser)₃, such that the VH and VL sequences can be expressed as a contiguous single-chain protein, with the VL and VH regions joined by the flexible linker (see *e.g.*, Bird *et al.* (1988) *Science* 242:423-426; Huston *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:5879-5883; McCafferty *et al.*, (1990) *Nature* 348:552-554).

Production of Monoclonal Antibodies of the Invention

Monoclonal antibodies (mAbs) of the present invention can be produced by a variety of techniques, including conventional monoclonal antibody methodology *e.g.*, the standard somatic cell hybridization technique of Kohler and Milstein (1975) *Nature* 256:495. Although somatic cell hybridization procedures are preferred, in principle, other techniques for producing monoclonal antibody can be employed *e.g.*, viral or oncogenic transformation of B lymphocytes.

The preferred animal system for preparing hybridomas is the murine system. Hybridoma production in the mouse is a very well-established procedure. Immunization protocols and techniques for isolation of immunized splenocytes for fusion are known in the art. Fusion partners (*e.g.*, murine myeloma cells) and fusion procedures are also known.

Chimeric or humanized antibodies of the present invention can be prepared based on the sequence of a murine monoclonal antibody prepared as described above. DNA encoding the heavy and light chain immunoglobulins can be obtained from the murine hybridoma of interest and engineered to contain non-murine (*e.g.*, human) immunoglobulin sequences using standard molecular biology techniques. For example, to create a chimeric antibody, the murine variable regions can be linked to human constant regions using methods known in the art (see *e.g.*, U.S. Patent No. 4,816,567 to Cabilly *et al.*). To create a humanized antibody, the murine CDR regions can be inserted into a human framework using methods known in the art (see *e.g.*, U.S. Patent No. 5,225,539 to Winter, and U.S. Patent Nos. 5,530,101; 5,585,089; 5,693,762 and 6,180,370 to Queen *et al.*).

In a preferred embodiment, the antibodies of the invention are human monoclonal antibodies. Such human monoclonal antibodies directed against PD-L1 can be generated using transgenic or transchromosomal mice carrying parts of the human immune system rather than the mouse system. These transgenic and transchromosomal mice include mice referred to herein as HuMAb mice and KM miceTM, respectively, and are collectively referred to herein as "human Ig mice."

The HuMAb mouse® (Medarex, Inc.) contains human immunoglobulin gene miniloci that encode unrearranged human heavy (μ and γ) and κ light chain immunoglobulin sequences, together with targeted mutations that inactivate the endogenous μ and κ chain loci (see e.g., Lonberg, *et al.* (1994) *Nature* 368(6474): 856-859). Accordingly, the mice exhibit reduced expression of mouse IgM or κ , and in response to immunization, the introduced human heavy and light chain transgenes undergo class switching and somatic mutation to generate high affinity human IgG κ monoclonal (Lonberg, N. *et al.* (1994), *supra*; reviewed in Lonberg, N. (1994) *Handbook of Experimental Pharmacology* 113:49-101; Lonberg, N. and Huszar, D. (1995) *Intern. Rev. Immunol.* 13: 65-93, and Harding, F. and Lonberg, N. (1995) *Ann. N.Y. Acad. Sci.* 764:536-546). The preparation and use of HuMAb mice, and the genomic modifications carried by such mice, is further described in Taylor, L. *et al.* (1992) *Nucleic Acids Research* 20:6287-6295; Chen, J. *et al.* (1993) *International Immunology* 5: 647-656; Tuaillon *et al.* (1993) *Proc. Natl. Acad. Sci. USA* 90:3720-3724; Choi *et al.* (1993) *Nature Genetics* 4:117-123; Chen, J. *et al.* (1993) *EMBO J.* 12: 821-830; Tuaillon *et al.* (1994) *J. Immunol.* 152:2912-2920; Taylor, L. *et al.* (1994) *International Immunology* 6: 579-591; and Fishwild, D. *et al.* (1996) *Nature Biotechnology* 14: 845-851, the contents of all of which are hereby specifically incorporated by reference in their entirety. See further, U.S. Patent Nos. 5,545,806; 5,569,825; 5,625,126; 5,633,425; 5,789,650; 5,877,397; 5,661,016; 5,814,318; 5,874,299; and 5,770,429; all to Lonberg and Kay; U.S. Patent No. 5,545,807 to Surani *et al.*; PCT Publication Nos. WO 92/03918, WO 93/12227, WO 94/25585, WO 97/13852, WO 98/24884 and WO 99/45962, all to Lonberg and Kay; and PCT Publication No. WO 01/14424 to Korman *et al.*

In another embodiment, human antibodies of the invention can be raised using a mouse that carries human immunoglobulin sequences on transgenes and transchromosomes, such as a mouse that carries a human heavy chain transgene and a

human light chain transchromosome. Such mice, referred to herein as "KM miceTM," are described in detail in PCT Publication WO 02/43478 to Ishida *et al.*

Still further, alternative transgenic animal systems expressing human immunoglobulin genes are available in the art and can be used to raise anti-PD-L1 antibodies of the invention. For example, an alternative transgenic system referred to as the Xenomouse (Abgenix, Inc.) can be used; such mice are described in, for example, U.S. Patent Nos. 5,939,598; 6,075,181; 6,114,598; 6,150,584 and 6,162,963 to Kucherlapati *et al.*

Moreover, alternative transchromosomic animal systems expressing human immunoglobulin genes are available in the art and can be used to raise anti-PD-L1 antibodies of the invention. For example, mice carrying both a human heavy chain transchromosome and a human light chain transchromosome, referred to as "TC mice" can be used; such mice are described in Tomizuka *et al.* (2000) *Proc. Natl. Acad. Sci. USA* 97:722-727. Furthermore, cows carrying human heavy and light chain transchromosomes have been described in the art (Kuroiwa *et al.* (2002) *Nature Biotechnology* 20:889-894) and can be used to raise anti-PD-L1 antibodies of the invention.

Human monoclonal antibodies of the invention can also be prepared using phage display methods for screening libraries of human immunoglobulin genes. Such phage display methods for isolating human antibodies are established in the art. See for example: U.S. Patent Nos. 5,223,409; 5,403,484; and 5,571,698 to Ladner *et al.*; U.S. Patent Nos. 5,427,908 and 5,580,717 to Dower *et al.*; U.S. Patent Nos. 5,969,108 and 6,172,197 to McCafferty *et al.*; and U.S. Patent Nos. 5,885,793; 6,521,404; 6,544,731; 6,555,313; 6,582,915 and 6,593,081 to Griffiths *et al.*

Human monoclonal antibodies of the invention can also be prepared using SCID mice into which human immune cells have been reconstituted such that a human antibody response can be generated upon immunization. Such mice are described in, for example, U.S. Patent Nos. 5,476,996 and 5,698,767 to Wilson *et al.*

Immunization of Human Ig Mice

When human Ig mice are used to raise human antibodies of the invention, such mice can be immunized with a purified or enriched preparation of PD-L1 antigen and/or recombinant PD-L1, or an PD-L1 fusion protein, as described by Lonberg, N. *et al.* (1994) *Nature* 368(6474): 856-859; Fishwild, D. *et al.* (1996) *Nature Biotechnology* 14:

845-851; and PCT Publication WO 98/24884 and WO 01/14424. Preferably, the mice will be 6-16 weeks of age upon the first infusion. For example, a purified or recombinant preparation (5-50 µg) of PD-L1 antigen can be used to immunize the human Ig mice intraperitoneally.

5 Detailed procedures to generate fully human monoclonal antibodies to PD-L1 are described in Example 1 below. Cumulative experience with various antigens has shown that the transgenic mice respond when initially immunized intraperitoneally (IP) with antigen in complete Freund's adjuvant, followed by every other week IP immunizations (up to a total of 6) with antigen in incomplete Freund's adjuvant. However, adjuvants
10 other than Freund's are also found to be effective. In addition, whole cells in the absence of adjuvant are found to be highly immunogenic. The immune response can be monitored over the course of the immunization protocol with plasma samples being obtained by retroorbital bleeds. The plasma can be screened by ELISA (as described below), and mice with sufficient titers of anti-PD-L1 human immunoglobulin can be used for fusions. Mice
15 can be boosted intravenously with antigen 3 days before sacrifice and removal of the spleen. It is expected that 2-3 fusions for each immunization may need to be performed. Between 6 and 24 mice are typically immunized for each antigen. Usually both HCo7 and HCo12 strains are used. In addition, both HCo7 and HCo12 transgene can be bred together into a single mouse having two different human heavy chain transgenes
20 (HCo7/HCo12). Alternatively or additionally, the KM mouseTM strain can be used, as described in Example 1.

Generation of Hybridomas Producing Human Monoclonal Antibodies of the Disclosure

To generate hybridomas producing human monoclonal antibodies of the invention, splenocytes and/or lymph node cells from immunized mice can be isolated and
25 fused to an appropriate immortalized cell line, such as a mouse myeloma cell line. The resulting hybridomas can be screened for the production of antigen-specific antibodies. For example, single cell suspensions of splenic lymphocytes from immunized mice can be fused to one-sixth the number of P3X63-Ag8.653 nonsecreting mouse myeloma cells (ATCC, CRL 1580) with 50% PEG. Cells are plated at approximately 2×10^5 in flat
30 bottom microtiter plate, followed by a two week incubation in selective medium containing 20% fetal Clone Serum, 18% "653" conditioned media, 5% origen (IGEN), 4 mM L-glutamine, 1 mM sodium pyruvate, 5mM HEPES, 0.055 mM 2-mercaptoethanol,

50 units/ml penicillin, 50 mg/ml streptomycin, 50 mg/ml gentamycin and 1X HAT (Sigma; the HAT is added 24 hours after the fusion). After approximately two weeks, cells can be cultured in medium in which the HAT is replaced with HT. Individual wells can then be screened by ELISA for human monoclonal IgM and IgG antibodies. Once
5 extensive hybridoma growth occurs, medium can be observed usually after 10-14 days. The antibody secreting hybridomas can be re-plated, screened again, and if still positive for human IgG, the monoclonal antibodies can be subcloned at least twice by limiting dilution. The stable subclones can then be cultured *in vitro* to generate small amounts of antibody in tissue culture medium for characterization.

10 To purify human monoclonal antibodies, selected hybridomas can be grown in two-liter spinner-flasks for monoclonal antibody purification. Supernatants can be filtered and concentrated before affinity chromatography with protein A-sepharose (Pharmacia, Piscataway, N.J.). Eluted IgG can be checked by gel electrophoresis and high performance liquid chromatography to ensure purity. The buffer solution can be
15 exchanged into PBS, and the concentration can be determined by OD280 using 1.43 extinction coefficient. The monoclonal antibodies can be aliquoted and stored at -80° C.

Generation of Transfectomas Producing Monoclonal Antibodies of the Disclosure

Antibodies of the invention also can be produced in a host cell transfectoma using, for example, a combination of recombinant DNA techniques and gene transfection
20 methods as is well known in the art (*e.g.*, Morrison, S. (1985) Science 229:1202).

For example, to express the antibodies, or antibody fragments thereof, DNAs encoding partial or full-length light and heavy chains, can be obtained by standard molecular biology techniques (*e.g.*, PCR amplification or cDNA cloning using a hybridoma that expresses the antibody of interest) and the DNAs can be inserted into
25 expression vectors such that the genes are operatively linked to transcriptional and translational control sequences. In this context, the term "operatively linked" is intended to mean that an antibody gene is ligated into a vector such that transcriptional and translational control sequences within the vector serve their intended function of regulating the transcription and translation of the antibody gene. The expression vector
30 and expression control sequences are chosen to be compatible with the expression host cell used. The antibody light chain gene and the antibody heavy chain gene can be inserted into separate vector or, more typically, both genes are inserted into the same

expression vector. The antibody genes are inserted into the expression vector by standard methods (e.g., ligation of complementary restriction sites on the antibody gene fragment and vector, or blunt end ligation if no restriction sites are present). The light and heavy chain variable regions of the antibodies described herein can be used to create full-length antibody genes of any antibody isotype by inserting them into expression vectors already encoding heavy chain constant and light chain constant regions of the desired isotype such that the V_H segment is operatively linked to the C_H segment(s) within the vector and the V_K segment is operatively linked to the C_L segment within the vector. Additionally or alternatively, the recombinant expression vector can encode a signal peptide that facilitates secretion of the antibody chain from a host cell. The antibody chain gene can be cloned into the vector such that the signal peptide is linked in-frame to the amino terminus of the antibody chain gene. The signal peptide can be an immunoglobulin signal peptide or a heterologous signal peptide (i.e., a signal peptide from a non-immunoglobulin protein).

In addition to the antibody chain genes, the recombinant expression vectors of the invention carry regulatory sequences that control the expression of the antibody chain genes in a host cell. The term "regulatory sequence" is intended to include promoters, enhancers and other expression control elements (e.g., polyadenylation signals) that control the transcription or translation of the antibody chain genes. Such regulatory sequences are described, for example, in Goeddel (Gene Expression Technology. Methods in Enzymology 185, Academic Press, San Diego, CA (1990)). It will be appreciated by those skilled in the art that the design of the expression vector, including the selection of regulatory sequences, may depend on such factors as the choice of the host cell to be transformed, the level of expression of protein desired, etc. Preferred regulatory sequences for mammalian host cell expression include viral elements that direct high levels of protein expression in mammalian cells, such as promoters and/or enhancers derived from cytomegalovirus (CMV), Simian Virus 40 (SV40), adenovirus, (e.g., the adenovirus major late promoter (AdMLP) and polyoma. Alternatively, nonviral regulatory sequences may be used, such as the ubiquitin promoter or β -globin promoter. Still further, regulatory elements composed of sequences from different sources, such as the SR α promoter system, which contains sequences from the SV40 early promoter and the long terminal repeat of human T cell leukemia virus type 1 (Takebe, Y. *et al.* (1988) *Mol. Cell. Biol.* 8:466-472).

In addition to the antibody chain genes and regulatory sequences, the recombinant expression vectors of the invention may carry additional sequences, such as sequences that regulate replication of the vector in host cells (*e.g.*, origins of replication) and selectable marker genes. The selectable marker gene facilitates selection of host cells into
5 which the vector has been introduced (see, *e.g.*, U.S. Pat. Nos. 4,399,216, 4,634,665 and 5,179,017, all by Axel *et al.*). For example, typically the selectable marker gene confers resistance to drugs, such as G418, hygromycin or methotrexate, on a host cell into which the vector has been introduced. Preferred selectable marker genes include the dihydrofolate reductase (DHFR) gene (for use in dhfr- host cells with methotrexate
10 selection/amplification) and the neo gene (for G418 selection).

For expression of the light and heavy chains, the expression vector(s) encoding the heavy and light chains is transfected into a host cell by standard techniques. The various forms of the term "transfection" are intended to encompass a wide variety of techniques commonly used for the introduction of exogenous DNA into a prokaryotic or
15 eukaryotic host cell, *e.g.*, electroporation, calcium-phosphate precipitation, DEAE-dextran transfection and the like. Although it is theoretically possible to express the antibodies of the invention in either prokaryotic or eukaryotic host cells, expression of antibodies in eukaryotic cells, and most preferably mammalian host cells, is the most preferred because such eukaryotic cells, and in particular mammalian cells, are more
20 likely than prokaryotic cells to assemble and secrete a properly folded and immunologically active antibody. Prokaryotic expression of antibody genes has been reported to be ineffective for production of high yields of active antibody (Boss, M. A. and Wood, C. R. (1985) *Immunology Today* 6:12-13).

Preferred mammalian host cells for expressing the recombinant antibodies of the
25 invention include Chinese Hamster Ovary (CHO cells) (including dhfr- CHO cells, described in Urlaub and Chasin, (1980) *Proc. Natl. Acad. Sci. USA* 77:4216-4220, used with a DHFR selectable marker, *e.g.*, as described in R. J. Kaufman and P. A. Sharp (1982) *Mol. Biol.* 159:601-621), NSO myeloma cells, COS cells and SP2 cells. In particular, for use with NSO myeloma cells, another preferred expression system is the
30 GS gene expression system disclosed in WO 87/04462, WO 89/01036 and EP 338,841. When recombinant expression vectors encoding antibody genes are introduced into mammalian host cells, the antibodies are produced by culturing the host cells for a period of time sufficient to allow for expression of the antibody in the host cells or, more

preferably, secretion of the antibody into the culture medium in which the host cells are grown. Antibodies can be recovered from the culture medium using standard protein purification methods.

Characterization of Antibody Binding to Antigen

5 Antibodies of the invention can be tested for binding to PD-L1 by, for example, standard ELISA. Briefly, microtiter plates are coated with purified PD-L1 at 0.25 µg/ml in PBS, and then blocked with 5% bovine serum albumin in PBS. Dilutions of antibody (e.g., dilutions of plasma from PD-L1-immunized mice) are added to each well and
10 incubated for 1-2 hours at 37°C. The plates are washed with PBS/Tween and then incubated with secondary reagent (e.g., for human antibodies, a goat-anti-human IgG Fc-specific polyclonal reagent) conjugated to alkaline phosphatase for 1 hour at 37°C. After washing, the plates are developed with pNPP substrate (1 mg/ml), and analyzed at OD of 405-650. Preferably, mice which develop the highest titers will be used for fusions.

 An ELISA assay as described above can also be used to screen for hybridomas
15 that show positive reactivity with PD-L1 immunogen. Hybridomas that bind with high avidity to PD-L1 are subcloned and further characterized. One clone from each hybridoma, which retains the reactivity of the parent cells (by ELISA), can be chosen for making a 5-10 vial cell bank stored at -140 °C, and for antibody purification.

 To purify anti-PD-L1 antibodies, selected hybridomas can be grown in two-liter
20 spinner-flasks for monoclonal antibody purification. Supernatants can be filtered and concentrated before affinity chromatography with protein A-sepharose (Pharmacia, Piscataway, NJ). Eluted IgG can be checked by gel electrophoresis and high performance liquid chromatography to ensure purity. The buffer solution can be exchanged into PBS, and the concentration can be determined by OD₂₈₀ using 1.43 extinction coefficient. The
25 monoclonal antibodies can be aliquoted and stored at -80 °C.

 To determine if the selected anti-PD-L1 monoclonal antibodies bind to unique epitopes, each antibody can be biotinylated using commercially available reagents (Pierce, Rockford, IL). Competition studies using unlabeled monoclonal antibodies and biotinylated monoclonal antibodies can be performed using PD-L1 coated-ELISA plates
30 as described above. Biotinylated mAb binding can be detected with a strep-avidin-alkaline phosphatase probe.

To determine the isotype of purified antibodies, isotype ELISAs can be performed using reagents specific for antibodies of a particular isotype. For example, to determine the isotype of a human monoclonal antibody, wells of microtiter plates can be coated with 1 µg/ml of anti-human immunoglobulin overnight at 4° C. After blocking with 1% BSA, the plates are reacted with 1 µg /ml or less of test monoclonal antibodies or purified isotype controls, at ambient temperature for one to two hours. The wells can then be reacted with either human IgG1 or human IgM-specific alkaline phosphatase-conjugated probes. Plates are developed and analyzed as described above.

Anti-PD-L1 human IgGs can be further tested for reactivity with PD-L1 antigen by Western blotting. Briefly, PD-L1 can be prepared and subjected to sodium dodecyl sulfate polyacrylamide gel electrophoresis. After electrophoresis, the separated antigens are transferred to nitrocellulose membranes, blocked with 10% fetal calf serum, and probed with the monoclonal antibodies to be tested. Human IgG binding can be detected using anti-human IgG alkaline phosphatase and developed with BCIP/NBT substrate tablets (Sigma Chem. Co., St. Louis, Mo.).

Antibody Physical Properties

The antibodies of the present invention may be further characterized by the various physical properties of the anti-PD-L1 antibodies. Various assays may be used to detect and/or differentiate different classes of antibodies based on these physical properties.

In some embodiments, antibodies of the present invention may contain one or more glycosylation sites in either the light or heavy chain variable region. The presence of one or more glycosylation sites in the variable region may result in increased immunogenicity of the antibody or an alteration of the pK of the antibody due to altered antigen binding (Marshall *et al* (1972) *Annu Rev Biochem* 41:673-702; Gala FA and Morrison SL (2004) *J Immunol* 172:5489-94; Wallick *et al* (1988) *J Exp Med* 168:1099-109; Spiro RG (2002) *Glycobiology* 12:43R-56R; Parekh *et al* (1985) *Nature* 316:452-7; Mimura *et al.* (2000) *Mol Immunol* 37:697-706). Glycosylation has been known to occur at motifs containing an N-X-S/T sequence. Variable region glycosylation may be tested using a Glycoblott assay, which cleaves the antibody to produce a Fab, and then tests for glycosylation using an assay that measures periodate oxidation and Schiff base formation. Alternatively, variable region glycosylation may be tested using Dionex light

chromatography (Dionex-LC), which cleaves saccharides from a Fab into monosaccharides and analyzes the individual saccharide content. In some instances, it is preferred to have an anti- PD-L1 antibody that does not contain variable region glycosylation. This can be achieved either by selecting antibodies that do not contain the glycosylation motif in the variable region or by mutating residues within the glycosylation motif using standard techniques well known in the art.

In a preferred embodiment, the antibodies of the present invention do not contain asparagine isomerism sites. A deamidation or isoaspartic acid effect may occur on N-G or D-G sequences, respectively. The deamidation or isoaspartic acid effect results in the creation of isoaspartic acid which decreases the stability of an antibody by creating a kinked structure off a side chain carboxy terminus rather than the main chain. The creation of isoaspartic acid can be measured using an iso-quant assay, which uses a reverse-phase HPLC to test for isoaspartic acid.

Each antibody will have a unique isoelectric point (pI), but generally antibodies will fall in the pH range of between 6 and 9.5. The pI for an IgG1 antibody typically falls within the pH range of 7-9.5 and the pI for an IgG4 antibody typically falls within the pH range of 6-8. Antibodies may have a pI that is outside this range. Although the effects are generally unknown, there is speculation that antibodies with a pI outside the normal range may have some unfolding and instability under *in vivo* conditions. The isoelectric point may be tested using a capillary isoelectric focusing assay, which creates a pH gradient and may utilize laser focusing for increased accuracy (Janjini *et al* (2002) *Electrophoresis* 23:1605-11; Ma *et al.* (2001) *Chromatographia* 53:875-89; Hunt *et al* (1998) *J Chromatogr A* 800:355-67). In some instances, it is preferred to have an anti-PD-L1 antibody that contains a pI value that falls in the normal range. This can be achieved either by selecting antibodies with a pI in the normal range, or by mutating charged surface residues using standard techniques well known in the art.

Each antibody will have a melting temperature that is indicative of thermal stability (Krishnamurthy R and Manning MC (2002) *Curr Pharm Biotechnol* 3:361-71). A higher thermal stability indicates greater overall antibody stability *in vivo*. The melting point of an antibody may be measure using techniques such as differential scanning calorimetry (Chen *et al* (2003) *Pharm Res* 20:1952-60; Ghirlando *et al* (1999) *Immunol Lett* 68:47-52). T_{M1} indicates the temperature of the initial unfolding of the antibody. T_{M2} indicates the temperature of complete unfolding of the antibody. Generally, it is

preferred that the T_{M1} of an antibody of the present invention is greater than 60°C, preferably greater than 65°C, even more preferably greater than 70°C. Alternatively, the thermal stability of an antibody may be measure using circular dichroism (Murray *et al.* (2002) *J. Chromatogr Sci* 40:343-9). The thermal stability of anti-PD-L1 antibodies

5 disclosed herein is summarized in Table 1.

Table 1.

mAb	Tm1 (°C)	Tm2 (°C)
3G10	70	75
5F8	72	74
11E6	64	73
1B12	69	72
12A4	68	72
10A5		71
12B7		70
13G4	66	69
10H10		69

In a preferred embodiment, antibodies are selected that do not rapidly degrade. Fragmentation of an anti- PD-L1 antibody may be measured using capillary electrophoresis (CE) and MALDI-MS, as is well understood in the art (Alexander AJ and
10 Hughes DE (1995) *Anal Chem* 67:3626-32).

In another preferred embodiment, antibodies are selected that have minimal aggregation effects. Aggregation may lead to triggering of an unwanted immune response and/or altered or unfavorable pharmacokinetic properties. Generally, antibodies are acceptable with aggregation of 25% or less, preferably 20% or less, even more
15 preferably 15% or less, even more preferably 10% or less and even more preferably 5% or less. Aggregation may be measured by several techniques well known in the art, including size-exclusion column (SEC) high performance liquid chromatography (HPLC), and light scattering to identify monomers, dimers, trimers or multimers.

Immunoconjugates

In another aspect, the present invention features an anti-PD-L1 antibody, or a fragment thereof, conjugated to a therapeutic moiety, such as a cytotoxin, a drug (e.g., an immunosuppressant) or a radiotoxin. Such conjugates are referred to herein as

5 “immunoconjugates”. Immunoconjugates that include one or more cytotoxins are referred to as “immunotoxins.” A cytotoxin or cytotoxic agent includes any agent that is detrimental to (e.g., kills) cells. Examples include taxol, cytochalasin B, gramicidin D, ethidium bromide, emetine, mitomycin, etoposide, tenoposide, vincristine, vinblastine, colchicin, doxorubicin, daunorubicin, dihydroxy anthracin dione, mitoxantrone, 10 mithramycin, actinomycin D, 1-dehydrotestosterone, glucocorticoids, procaine, tetracaine, lidocaine, propranolol, and puromycin and analogs or homologs thereof. Therapeutic agents also include, for example, antimetabolites (e.g., methotrexate, 6-mercaptopurine, 6-thioguanine, cytarabine, 5-fluorouracil decarbazine), alkylating agents (e.g., mechlorethamine, thioepa chlorambucil, melphalan, carmustine (BSNU) and 15 lomustine (CCNU), cyclophosphamide, busulfan, dibromomannitol, streptozotocin, mitomycin C, and cis-dichlorodiamine platinum (II) (DDP) cisplatin), anthracyclines (e.g., daunorubicin (formerly daunomycin) and doxorubicin), antibiotics (e.g., dactinomycin (formerly actinomycin), bleomycin, mithramycin, and anthramycin (AMC)), and anti-mitotic agents (e.g., vincristine and vinblastine).

20 Other preferred examples of therapeutic cytotoxins that can be conjugated to an antibody of the invention include duocarmycins, calicheamicins, maytansines and auristatins, and derivatives thereof. An example of a calicheamicin antibody conjugate is commercially available (Mylotarg™; Wyeth-Ayerst).

Cytotoxins can be conjugated to antibodies of the invention using linker 25 technology available in the art. Examples of linker types that have been used to conjugate a cytotoxin to an antibody include, but are not limited to, hydrazones, thioethers, esters, disulfides and peptide-containing linkers. A linker can be chosen that is, for example, susceptible to cleavage by low pH within the lysosomal compartment or susceptible to cleavage by proteases, such as proteases preferentially expressed in tumor tissue such as 30 cathepsins (e.g., cathepsins B, C, D).

For further discussion of types of cytotoxins, linkers and methods for conjugating therapeutic agents to antibodies, see also Saito, G. *et al.* (2003) *Adv. Drug Deliv. Rev.* 55:199-215; Trail, P.A. *et al.* (2003) *Cancer Immunol. Immunother.* 52:328-337; Payne,

G. (2003) *Cancer Cell* 3:207-212; Allen, T.M. (2002) *Nat. Rev. Cancer* 2:750-763; Pastan, I. and Kreitman, R. J. (2002) *Curr. Opin. Investig. Drugs* 3:1089-1091; Senter, P.D. and Springer, C.J. (2001) *Adv. Drug Deliv. Rev.* 53:247-264.

Antibodies of the present invention also can be conjugated to a radioactive isotope to generate cytotoxic radiopharmaceuticals, also referred to as radioimmunoconjugates. Examples of radioactive isotopes that can be conjugated to antibodies for use diagnostically or therapeutically include, but are not limited to, iodine¹³¹, indium¹¹¹, yttrium⁹⁰ and lutetium¹⁷⁷. Method for preparing radioimmunoconjugates are established in the art. Examples of radioimmunoconjugates are commercially available, including ZevalinTM (IDEC Pharmaceuticals) and BexxarTM (Corixa Pharmaceuticals), and similar methods can be used to prepare radioimmunoconjugates using the antibodies of the invention.

The antibody conjugates of the invention can be used to modify a given biological response, and the drug moiety is not to be construed as limited to classical chemical therapeutic agents. For example, the drug moiety may be a protein or polypeptide possessing a desired biological activity. Such proteins may include, for example, an enzymatically active toxin, or active fragment thereof, such as abrin, ricin A, pseudomonas exotoxin, or diphtheria toxin; a protein such as tumor necrosis factor or interferon- γ ; or, biological response modifiers such as, for example, lymphokines, interleukin-1 ("IL-1"), interleukin-2 ("IL-2"), interleukin-6 ("IL-6"), granulocyte macrophage colony stimulating factor ("GM-CSF"), granulocyte colony stimulating factor ("G-CSF"), or other growth factors.

Techniques for conjugating such therapeutic moiety to antibodies are well known, see, e.g., Arnon *et al.*, "Monoclonal Antibodies For Immunotargeting Of Drugs In Cancer Therapy," in *Monoclonal Antibodies And Cancer Therapy*, Reisfeld *et al.* (eds.), pp. 243-56 (Alan R. Liss, Inc. 1985); Hellstrom *et al.*, "Antibodies For Drug Delivery," in *Controlled Drug Delivery* (2nd Ed.), Robinson *et al.* (eds.), pp. 623-53 (Marcel Dekker, Inc. 1987); Thorpe, "Antibody Carriers Of Cytotoxic Agents In Cancer Therapy: A Review," in *Monoclonal Antibodies '84: Biological And Clinical Applications*, Pinchera *et al.* (eds.), pp. 475-506 (1985); "Analysis, Results, And Future Prospective Of The Therapeutic Use Of Radiolabeled Antibody In Cancer Therapy," in *Monoclonal Antibodies For Cancer Detection And Therapy*, Baldwin *et al.* (eds.), pp. 303-16

(Academic Press 1985), and Thorpe *et al.*, "The Preparation And Cytotoxic Properties Of Antibody-Toxin Conjugates," Immunol. Rev., 62:119-58 (1982).

Bispecific Molecules

In another aspect, the present invention features bispecific molecules comprising
5 an anti-PD-L1 antibody, or a fragment thereof, of the invention. An antibody of the invention, or antigen-binding portions thereof, can be derivatized or linked to another functional molecule, *e.g.*, another peptide or protein (*e.g.*, another antibody or ligand for a receptor) to generate a bispecific molecule that binds to at least two different binding sites or target molecules. The antibody of the invention may in fact be derivatized or linked to
10 more than one other functional molecule to generate multispecific molecules that bind to more than two different binding sites and/or target molecules; such multispecific molecules are also intended to be encompassed by the term "bispecific molecule" as used herein. To create a bispecific molecule of the invention, an antibody of the invention can be functionally linked (*e.g.*, by chemical coupling, genetic fusion, noncovalent
15 association or otherwise) to one or more other binding molecules, such as another antibody, antibody fragment, peptide or binding mimetic, such that a bispecific molecule results.

Accordingly, the present invention includes bispecific molecules comprising at least one first binding specificity for PD-L1 and a second binding specificity for a second
20 target epitope. In a particular embodiment of the invention, the second target epitope is an Fc receptor, *e.g.*, human Fc γ RI (CD64) or a human Fc α receptor (CD89). Therefore, the invention includes bispecific molecules capable of binding both to Fc γ R or Fc α R expressing effector cells (*e.g.*, monocytes, macrophages or polymorphonuclear cells (PMNs)), and to target cells expressing PD-L1. These bispecific molecules target PD-L1
25 expressing cells to effector cell and trigger Fc receptor-mediated effector cell activities, such as phagocytosis of an PD-L1 expressing cells, antibody dependent cell-mediated cytotoxicity (ADCC), cytokine release, or generation of superoxide anion.

In an embodiment of the invention in which the bispecific molecule is multispecific, the molecule can further include a third binding specificity, in addition to
30 an anti-Fc binding specificity and an anti-PD-L1 binding specificity. In one embodiment, the third binding specificity is an anti-enhancement factor (EF) portion, *e.g.*, a molecule which binds to a surface protein involved in cytotoxic activity and thereby increases the

immune response against the target cell. The "anti-enhancement factor portion" can be an antibody, functional antibody fragment or a ligand that binds to a given molecule, *e.g.*, an antigen or a receptor, and thereby results in an enhancement of the effect of the binding determinants for the F_C receptor or target cell antigen. The "anti-enhancement factor portion" can bind an F_C receptor or a target cell antigen. Alternatively, the anti-enhancement factor portion can bind to an entity that is different from the entity to which the first and second binding specificities bind. For example, the anti-enhancement factor portion can bind a cytotoxic T-cell (*e.g.* via CD2, CD3, CD8, CD28, CD4, CD40, ICAM-1 or other immune cell that results in an increased immune response against the target cell).

In one embodiment, the bispecific molecules of the invention comprise as a binding specificity at least one antibody, or an antibody fragment thereof, including, *e.g.*, an Fab, Fab', F(ab')₂, Fv, or a single chain Fv. The antibody may also be a light chain or heavy chain dimer, or any minimal fragment thereof such as a Fv or a single chain construct as described in Ladner *et al.* U.S. Patent No. 4,946,778, the contents of which is expressly incorporated by reference.

In one embodiment, the binding specificity for an Fcγ receptor is provided by a monoclonal antibody, the binding of which is not blocked by human immunoglobulin G (IgG). As used herein, the term "IgG receptor" refers to any of the eight γ-chain genes located on chromosome 1. These genes encode a total of twelve transmembrane or soluble receptor isoforms which are grouped into three Fcγ receptor classes: FcγRI (CD64), FcγRII (CD32), and FcγRIII (CD16). In one preferred embodiment, the Fcγ receptor is a human high affinity FcγRI. The human FcγRI is a 72 kDa molecule, which shows high affinity for monomeric IgG (10⁸ - 10⁹ M⁻¹).

The production and characterization of certain preferred anti-Fcγ monoclonal antibodies are described by Fanger *et al.* in PCT Publication WO 88/00052 and in U.S. Patent No. 4,954,617, the teachings of which are fully incorporated by reference herein. These antibodies bind to an epitope of FcγRI, FcγRII or FcγRIII at a site which is distinct from the Fcγ binding site of the receptor and, thus, their binding is not blocked substantially by physiological levels of IgG. Specific anti-FcγRI antibodies useful in this invention are mAb 22, mAb 32, mAb 44, mAb 62 and mAb 197. The hybridoma producing mAb 32 is available from the American Type Culture Collection, ATCC

Accession No. HB9469. In other embodiments, the anti-Fc γ receptor antibody is a humanized form of monoclonal antibody 22 (H22). The production and characterization of the H22 antibody is described in Graziano, R.F. *et al.* (1995) *J. Immunol* **155** (10): 4996-5002 and PCT Publication WO 94/10332. The H22 antibody producing cell line
5 was deposited at the American Type Culture Collection under the designation HA022CL1 and has the accession no. CRL 11177.

In still other preferred embodiments, the binding specificity for an Fc receptor is provided by an antibody that binds to a human IgA receptor, *e.g.*, an Fc α receptor (Fc α RI (CD89)), the binding of which is preferably not blocked by human
10 immunoglobulin A (IgA). The term "IgA receptor" is intended to include the gene product of one α -gene (Fc α RI) located on chromosome 19. This gene is known to encode several alternatively spliced transmembrane isoforms of 55 to 110 kDa. Fc α RI (CD89) is constitutively expressed on monocytes/macrophages, eosinophilic and neutrophilic granulocytes, but not on non-effector cell populations. Fc α RI has medium
15 affinity ($\approx 5 \times 10^7$ M $^{-1}$) for both IgA1 and IgA2, which is increased upon exposure to cytokines such as G-CSF or GM-CSF (Morton, H.C. *et al.* (1996) *Critical Reviews in Immunology* **16**:423-440). Four Fc α RI-specific monoclonal antibodies, identified as A3, A59, A62 and A77, which bind Fc α RI outside the IgA ligand binding domain, have been described (Monteiro, R.C. *et al.* (1992) *J. Immunol.* **148**:1764).

20 Fc α RI and Fc γ RI are preferred trigger receptors for use in the bispecific molecules of the invention because they are (1) expressed primarily on immune effector cells, *e.g.*, monocytes, PMNs, macrophages and dendritic cells; (2) expressed at high levels (*e.g.*, 5,000-100,000 per cell); (3) mediators of cytotoxic activities (*e.g.*, ADCC, phagocytosis); (4) mediate enhanced antigen presentation of antigens, including self-
25 antigens, targeted to them.

While human monoclonal antibodies are preferred, other antibodies which can be employed in the bispecific molecules of the invention are murine, chimeric and humanized monoclonal antibodies.

The bispecific molecules of the present invention can be prepared by conjugating
30 the constituent binding specificities, *e.g.*, the anti-FcR and anti-PD-L1 binding specificities, using methods known in the art. For example, each binding specificity of the bispecific molecule can be generated separately and then conjugated to one another. When the binding specificities are proteins or peptides, a variety of coupling or cross-

linking agents can be used for covalent conjugation. Examples of cross-linking agents include protein A, carbodiimide, N-succinimidyl-S-acetyl-thioacetate (SATA), 5,5'-dithiobis(2-nitrobenzoic acid) (DTNB), o-phenylenedimaleimide (oPDM), N-succinimidyl-3-(2-pyridyldithio)propionate (SPDP), and sulfosuccinimidyl 4-(N-maleimidomethyl) cyclohexane-1-carboxylate (sulfo-SMCC) (see *e.g.*, Karpovsky *et al.* (1984) *J. Exp. Med.* 160:1686; Liu, MA *et al.* (1985) *Proc. Natl. Acad. Sci. USA* 82:8648). Other methods include those described in Paulus (1985) Behring Ins. Mitt. No. 78, 118-132; Brennan *et al.* (1985) *Science* 229:81-83, and Glennie *et al.* (1987) *J. Immunol.* 139: 2367-2375). Preferred conjugating agents are SATA and sulfo-SMCC, both available from Pierce Chemical Co. (Rockford, IL).

When the binding specificities are antibodies, they can be conjugated via sulfhydryl bonding of the C-terminus hinge regions of the two heavy chains. In a particularly preferred embodiment, the hinge region is modified to contain an odd number of sulfhydryl residues, preferably one, prior to conjugation.

Alternatively, both binding specificities can be encoded in the same vector and expressed and assembled in the same host cell. This method is particularly useful where the bispecific molecule is a mAb x mAb, mAb x Fab, Fab x F(ab')₂ or ligand x Fab fusion protein. A bispecific molecule of the invention can be a single chain molecule comprising one single chain antibody and a binding determinant, or a single chain bispecific molecule comprising two binding determinants. Bispecific molecules may comprise at least two single chain molecules. Methods for preparing bispecific molecules are described for example in U.S. Patent Number 5,260,203; U.S. Patent Number 5,455,030; U.S. Patent Number 4,881,175; U.S. Patent Number 5,132,405; U.S. Patent Number 5,091,513; U.S. Patent Number 5,476,786; U.S. Patent Number 5,013,653; U.S. Patent Number 5,258,498; and U.S. Patent Number 5,482,858.

Binding of the bispecific molecules to their specific targets can be confirmed by, for example, enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA), FACS analysis, bioassay (*e.g.*, growth inhibition), or Western Blot assay. Each of these assays generally detects the presence of protein-antibody complexes of particular interest by employing a labeled reagent (*e.g.*, an antibody) specific for the complex of interest. For example, the FcR-antibody complexes can be detected using *e.g.*, an enzyme-linked antibody or antibody fragment which recognizes and specifically binds to the antibody-FcR complexes. Alternatively, the complexes can be detected using any of a variety of

other immunoassays. For example, the antibody can be radioactively labeled and used in a radioimmunoassay (RIA) (see, for example, Weintraub, B., Principles of Radioimmunoassays, Seventh Training Course on Radioligand Assay Techniques, The Endocrine Society, March, 1986, which is incorporated by reference herein). The
5 radioactive isotope can be detected by such means as the use of a γ counter or a scintillation counter or by autoradiography.

Pharmaceutical Compositions

In another aspect, the present invention provides a composition, *e.g.*, a
pharmaceutical composition, containing one or a combination of monoclonal antibodies,
10 or antigen-binding portion(s) thereof, of the present invention, formulated together with a pharmaceutically acceptable carrier. Such compositions may include one or a combination of (*e.g.*, two or more different) antibodies, or immunoconjugates or bispecific molecules of the invention. For example, a pharmaceutical composition of the invention can comprise a combination of antibodies (or immunoconjugates or bispecifics)
15 that bind to different epitopes on the target antigen or that have complementary activities.

Pharmaceutical compositions of the invention also can be administered in combination therapy, *i.e.*, combined with other agents. For example, the combination therapy can include an anti-PD-L1 antibody of the present invention combined with at least one other anti-inflammatory or immunosuppressant agent. Examples of therapeutic
20 agents that can be used in combination therapy are described in greater detail below in the section on uses of the antibodies of the invention.

As used herein, "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like that are physiologically compatible. Preferably,
25 the carrier is suitable for intravenous, intramuscular, subcutaneous, parenteral, spinal or epidermal administration (*e.g.*, by injection or infusion). Depending on the route of administration, the active compound, *i.e.*, antibody, immunoconjugate, or bispecific molecule, may be coated in a material to protect the compound from the action of acids and other natural conditions that may inactivate the compound.

30 The pharmaceutical compounds of the invention may include one or more pharmaceutically acceptable salts. A "pharmaceutically acceptable salt" refers to a salt that retains the desired biological activity of the parent compound and does not impart

any undesired toxicological effects (see e.g., Berge, S.M., *et al.* (1977) *J. Pharm. Sci.* 66:1-19). Examples of such salts include acid addition salts and base addition salts. Acid addition salts include those derived from nontoxic inorganic acids, such as hydrochloric, nitric, phosphoric, sulfuric, hydrobromic, hydroiodic, phosphorous and the like, as well as
5 from nontoxic organic acids such as aliphatic mono- and dicarboxylic acids, phenyl-substituted alkanolic acids, hydroxy alkanolic acids, aromatic acids, aliphatic and aromatic sulfonic acids and the like. Base addition salts include those derived from alkaline earth metals, such as sodium, potassium, magnesium, calcium and the like, as well as from nontoxic organic amines, such as N,N'-dibenzylethylenediamine, N-methylglucamine,
10 chloroprocaine, choline, diethanolamine, ethylenediamine, procaine and the like.

A pharmaceutical composition of the invention also may include a pharmaceutically acceptable anti-oxidant. Examples of pharmaceutically acceptable antioxidants include: (1) water soluble antioxidants, such as ascorbic acid, cysteine hydrochloride, sodium bisulfate, sodium metabisulfite, sodium sulfite and the like; (2) oil-
15 soluble antioxidants, such as ascorbyl palmitate, butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), lecithin, propyl gallate, alpha-tocopherol, and the like; and (3) metal chelating agents, such as citric acid, ethylenediamine tetraacetic acid (EDTA), sorbitol, tartaric acid, phosphoric acid, and the like.

Examples of suitable aqueous and nonaqueous carriers that may be employed in
20 the pharmaceutical compositions of the invention include water, ethanol, polyols (such as glycerol, propylene glycol, polyethylene glycol, and the like), and suitable mixtures thereof, vegetable oils, such as olive oil, and injectable organic esters, such as ethyl oleate. Proper fluidity can be maintained, for example, by the use of coating materials, such as lecithin, by the maintenance of the required particle size in the case of
25 dispersions, and by the use of surfactants.

These compositions may also contain adjuvants such as preservatives, wetting agents, emulsifying agents and dispersing agents. Prevention of presence of microorganisms may be ensured both by sterilization procedures, *supra*, and by the inclusion of various antibacterial and antifungal agents, for example, paraben,
30 chlorobutanol, phenol sorbic acid, and the like. It may also be desirable to include isotonic agents, such as sugars, sodium chloride, and the like into the compositions. In addition, prolonged absorption of the injectable pharmaceutical form may be brought

about by the inclusion of agents which delay absorption such as aluminum monostearate and gelatin.

Pharmaceutically acceptable carriers include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. The use of such media and agents for pharmaceutically active substances is known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, use thereof in the pharmaceutical compositions of the invention is contemplated. Supplementary active compounds can also be incorporated into the compositions.

Therapeutic compositions typically must be sterile and stable under the conditions of manufacture and storage. The composition can be formulated as a solution, microemulsion, liposome, or other ordered structure suitable to high drug concentration. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as mannitol, sorbitol, or sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent that delays absorption, for example, monostearate salts and gelatin.

Sterile injectable solutions can be prepared by incorporating the active compound in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by sterilization microfiltration. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle that contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying (lyophilization) that yield a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

The amount of active ingredient which can be combined with a carrier material to produce a single dosage form will vary depending upon the subject being treated, and the particular mode of administration. The amount of active ingredient which can be

combined with a carrier material to produce a single dosage form will generally be that amount of the composition which produces a therapeutic effect. Generally, out of one hundred per cent, this amount will range from about 0.01 per cent to about ninety-nine percent of active ingredient, preferably from about 0.1 per cent to about 70 per cent, most preferably from about 1 per cent to about 30 per cent of active ingredient in combination with a pharmaceutically acceptable carrier.

Dosage regimens are adjusted to provide the optimum desired response (e.g., a therapeutic response). For example, a single bolus may be administered, several divided doses may be administered over time or the dose may be proportionally reduced or increased as indicated by the exigencies of the therapeutic situation. It is especially advantageous to formulate parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the subjects to be treated; each unit contains a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on (a) the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and (b) the limitations inherent in the art of compounding such an active compound for the treatment of sensitivity in individuals.

For administration of the antibody, the dosage ranges from about 0.0001 to 100 mg/kg, and more usually 0.01 to 5 mg/kg, of the host body weight. For example dosages can be 0.3 mg/kg body weight, 1 mg/kg body weight, 3 mg/kg body weight, 5 mg/kg body weight or 10 mg/kg body weight or within the range of 1-10 mg/kg. An exemplary treatment regime entails administration once per week, once every two weeks, once every three weeks, once every four weeks, once a month, once every 3 months or once every three to 6 months. Preferred dosage regimens for an anti-PD-L1 antibody of the invention include 1 mg/kg body weight or 3 mg/kg body weight via intravenous administration, with the antibody being given using one of the following dosing schedules: (i) every four weeks for six dosages, then every three months; (ii) every three weeks; (iii) 3 mg/kg body weight once followed by 1 mg/kg body weight every three weeks.

In some methods, two or more monoclonal antibodies with different binding specificities are administered simultaneously, in which case the dosage of each antibody

administered falls within the ranges indicated. Antibody is usually administered on multiple occasions. Intervals between single dosages can be, for example, weekly, monthly, every three months or yearly. Intervals can also be irregular as indicated by measuring blood levels of antibody to the target antigen in the patient. In some methods, dosage is adjusted to achieve a plasma antibody concentration of about 1-1000 $\mu\text{g}/\text{ml}$ and in some methods about 25-300 $\mu\text{g}/\text{ml}$.

Alternatively, antibody can be administered as a sustained release formulation, in which case less frequent administration is required. Dosage and frequency vary depending on the half-life of the antibody in the patient. In general, human antibodies show the longest half life, followed by humanized antibodies, chimeric antibodies, and nonhuman antibodies. The dosage and frequency of administration can vary depending on whether the treatment is prophylactic or therapeutic. In prophylactic applications, a relatively low dosage is administered at relatively infrequent intervals over a long period of time. Some patients continue to receive treatment for the rest of their lives. In therapeutic applications, a relatively high dosage at relatively short intervals is sometimes required until progression of the disease is reduced or terminated, and preferably until the patient shows partial or complete amelioration of symptoms of disease. Thereafter, the patient can be administered a prophylactic regime.

Actual dosage levels of the active ingredients in the pharmaceutical compositions of the present invention may be varied so as to obtain an amount of the active ingredient which is effective to achieve the desired therapeutic response for a particular patient, composition, and mode of administration, without being toxic to the patient. The selected dosage level will depend upon a variety of pharmacokinetic factors including the activity of the particular compositions of the present invention employed, or the ester, salt or amide thereof, the route of administration, the time of administration, the rate of excretion of the particular compound being employed, the duration of the treatment, other drugs, compounds and/or materials used in combination with the particular compositions employed, the age, sex, weight, condition, general health and prior medical history of the patient being treated, and like factors well known in the medical arts.

A "therapeutically effective dosage" of an anti-PD-L1 antibody of the invention preferably results in a decrease in severity of disease symptoms, an increase in frequency and duration of disease symptom-free periods, or a prevention of impairment or disability due to the disease affliction. For example, for the treatment of PD-L1+

tumors, a "therapeutically effective dosage" preferably inhibits cell growth or tumor growth by at least about 20%, more preferably by at least about 40%, even more preferably by at least about 60%, and still more preferably by at least about 80% relative to untreated subjects. The ability of a compound to inhibit tumor growth can be
5 evaluated in an animal model system predictive of efficacy in human tumors. Alternatively, this property of a composition can be evaluated by examining the ability of the compound to inhibit, such inhibition *in vitro* by assays known to the skilled practitioner. A therapeutically effective amount of a therapeutic compound can decrease tumor size, or otherwise ameliorate symptoms in a subject. One of ordinary
10 skill in the art would be able to determine such amounts based on such factors as the subject's size, the severity of the subject's symptoms, and the particular composition or route of administration selected.

A composition of the present invention can be administered via one or more routes of administration using one or more of a variety of methods known in the art. As
15 will be appreciated by the skilled artisan, the route and/or mode of administration will vary depending upon the desired results. Preferred routes of administration for antibodies of the invention include intravenous, intramuscular, intradermal, intraperitoneal, subcutaneous, spinal or other parenteral routes of administration, for example by injection or infusion. The phrase "parenteral administration" as used herein means modes of
20 administration other than enteral and topical administration, usually by injection, and includes, without limitation, intravenous, intramuscular, intraarterial, intrathecal, intracapsular, intraorbital, intracardiac, intradermal, intraperitoneal, transtracheal, subcutaneous, subcuticular, intraarticular, subcapsular, subarachnoid, intraspinal, epidural and intrasternal injection and infusion.

25 Alternatively, an antibody of the invention can be administered via a non-parenteral route, such as a topical, epidermal or mucosal route of administration, for example, intranasally, orally, vaginally, rectally, sublingually or topically.

The active compounds can be prepared with carriers that will protect the compound against rapid release, such as a controlled release formulation, including
30 implants, transdermal patches, and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Many methods for the preparation of such formulations are patented or generally known to those skilled in the

art. See, e.g., *Sustained and Controlled Release Drug Delivery Systems*, J.R. Robinson, ed., Marcel Dekker, Inc., New York, 1978.

Therapeutic compositions can be administered with medical devices known in the art. For example, in a preferred embodiment, a therapeutic composition of the invention
5 can be administered with a needleless hypodermic injection device, such as the devices disclosed in U.S. Patent Nos. 5,399,163; 5,383,851; 5,312,335; 5,064,413; 4,941,880; 4,790,824; or 4,596,556. Examples of well-known implants and modules useful in the present invention include: U.S. Patent No. 4,487,603, which discloses an implantable micro-infusion pump for dispensing medication at a controlled rate; U.S. Patent
10 No. 4,486,194, which discloses a therapeutic device for administering medicants through the skin; U.S. Patent No. 4,447,233, which discloses a medication infusion pump for delivering medication at a precise infusion rate; U.S. Patent No. 4,447,224, which discloses a variable flow implantable infusion apparatus for continuous drug delivery; U.S. Patent No. 4,439,196, which discloses an osmotic drug delivery system having
15 multi-chamber compartments; and U.S. Patent No. 4,475,196, which discloses an osmotic drug delivery system. These patents are incorporated herein by reference. Many other such implants, delivery systems, and modules are known to those skilled in the art.

In certain embodiments, the human monoclonal antibodies of the invention can be formulated to ensure proper distribution *in vivo*. For example, the blood-brain barrier
20 (BBB) excludes many highly hydrophilic compounds. To ensure that the therapeutic compounds of the invention cross the BBB (if desired), they can be formulated, for example, in liposomes. For methods of manufacturing liposomes, see, e.g., U.S. Patents 4,522,811; 5,374,548; and 5,399,331. The liposomes may comprise one or more moieties which are selectively transported into specific cells or organs, thus enhance targeted drug
25 delivery (see, e.g., V.V. Ranade (1989) *J. Clin. Pharmacol.* 29:685). Exemplary targeting moieties include folate or biotin (see, e.g., U.S. Patent 5,416,016 to Low *et al.*); mannosides (Umezawa *et al.*, (1988) *Biochem. Biophys. Res. Commun.* 153:1038); antibodies (P.G. Bloeman *et al.* (1995) *FEBS Lett.* 357:140; M. Owais *et al.* (1995) *Antimicrob. Agents Chemoth.* 39:180); surfactant protein A receptor (Briscoe *et al.*
30 (1995) *Am. J. Physiol.* 1233:134); p120 (Schreier *et al.* (1994) *J. Biol. Chem.* 269:9090); see also K. Keinänen; M.L. Laukkanen (1994) *FEBS Lett.* 346:123; J.J. Killian; I.J. Fidler (1994) *Immunomethods* 4:273.

Uses and Methods of the Invention

The antibodies, antibody compositions and methods of the present invention have numerous *in vitro* and *in vivo* utilities involving, for example, detection of PD-L1 or enhancement of immune response by blockade of PD-L1. In a preferred embodiment, the antibodies of the present invention are human antibodies. For example, these molecules can be administered to cells in culture, *in vitro* or *ex vivo*, or to human subjects, *e.g.*, *in vivo*, to enhance immunity in a variety of situations. Accordingly, in one aspect, the invention provides a method of modifying an immune response in a subject comprising administering to the subject the antibody, or antigen-binding portion thereof, of the invention such that the immune response in the subject is modified. Preferably, the response is enhanced, stimulated or up-regulated.

As used herein, the term "subject" is intended to include human and non-human animals. Non-human animals includes all vertebrates, *e.g.*, mammals and non-mammals, such as non-human primates, sheep, dogs, cats, cows, horses, chickens, amphibians, and reptiles, although mammals are preferred, such as non-human primates, sheep, dogs, cats, cows and horses. Preferred subjects include human patients in need of enhancement of an immune response. The methods are particularly suitable for treating human patients having a disorder that can be treated by augmenting the T-cell mediated immune response. In a particular embodiment, the methods are particularly suitable for treatment of cancer cells *in vivo*. To achieve antigen-specific enhancement of immunity, the anti-PD-L1 antibodies can be administered together with an antigen of interest. When antibodies to PD-L1 are administered together with another agent, the two can be administered in either order or simultaneously.

The invention further provides methods for detecting the presence of human PD-L1 antigen in a sample, or measuring the amount of human PD-L1 antigen, comprising contacting the sample, and a control sample, with a human monoclonal antibody, or an antigen binding portion thereof, which specifically binds to human PD-L1, under conditions that allow for formation of a complex between the antibody or portion thereof and human PD-L1. The formation of a complex is then detected, wherein a difference complex formation between the sample compared to the control sample is indicative the presence of human PD-L1 antigen in the sample.

Cancer

Blockade of PD-L1 by antibodies can enhance the immune response to cancerous cells in the patient. PD-L1 is not expressed in normal human cells, but is abundant in a variety of human cancers (Dong *et al.* (2002) *Nat Med* 8:787-9). The interaction
5 between PD-1 and PD-L1 results in a decrease in tumor infiltrating lymphocytes, a decrease in T-cell receptor mediated proliferation, and immune evasion by the cancerous cells (Dong *et al.* (2003) *J Mol Med* 81:281-7; Blank *et al.* (2004) *Cancer Immunol. Immunother.* [epub]; Konishi *et al.* (2004) *Clin. Cancer Res.* 10:5094-100). Immune
10 suppression can be reversed by inhibiting the local interaction of PD-L1 to PD-1 and the effect is additive when the interaction of PD-L2 to PD-1 is blocked as well (Iwai *et al.* (2002) *PNAS* 99:12293-7; Brown *et al.* (2003) *J. Immunol.* 170:1257-66). An anti-PD-L1 antibody may be used alone to inhibit the growth of cancerous tumors. Alternatively, an anti-PD-L1 antibody may be used in conjunction with other immunogenic agents, standard cancer treatments, or other antibodies, as described below.

15 Accordingly, in one embodiment, the invention provides a method of inhibiting growth of tumor cells in a subject, comprising administering to the subject a therapeutically effective amount of an anti-PD-L1 antibody, or antigen-binding portion thereof. Preferably, the antibody is a human anti-PD-L1 antibody (such as any of the human anti-human PD-L1 antibodies described herein). Additionally or alternatively, the
20 antibody may be a chimeric or humanized anti-PD-L1 antibody.

Preferred cancers whose growth may be inhibited using the antibodies of the invention include cancers typically responsive to immunotherapy. Non-limiting examples of preferred cancers for treatment include melanoma (e.g., metastatic malignant melanoma), renal cancer, prostate cancer, breast cancer, colon cancer and lung cancer.
25 Examples of other cancers that may be treated using the methods of the invention include bone cancer, pancreatic cancer, skin cancer, cancer of the head or neck, cutaneous or intraocular malignant melanoma, uterine cancer, ovarian cancer, rectal cancer, cancer of the anal region, stomach cancer, testicular cancer, uterine cancer, carcinoma of the fallopian tubes, carcinoma of the endometrium, carcinoma of the cervix, carcinoma of the
30 vagina, carcinoma of the vulva, Hodgkin's Disease, non-Hodgkin's lymphoma, cancer of the esophagus, cancer of the small intestine, cancer of the endocrine system, cancer of the thyroid gland, cancer of the parathyroid gland, cancer of the adrenal gland, sarcoma of soft tissue, cancer of the urethra, cancer of the penis, chronic or acute leukemias including

acute myeloid leukemia, chronic myeloid leukemia, acute lymphoblastic leukemia, chronic lymphocytic leukemia, solid tumors of childhood, lymphocytic lymphoma, cancer of the bladder, cancer of the kidney or ureter, carcinoma of the renal pelvis, neoplasm of the central nervous system (CNS), primary CNS lymphoma, tumor angiogenesis, spinal axis tumor, brain stem glioma, pituitary adenoma, Kaposi's sarcoma, epidermoid cancer, squamous cell cancer, T-cell lymphoma, environmentally induced cancers including those induced by asbestos, and combinations of said cancers. The present invention is also useful for treatment of metastatic cancers, especially metastatic cancers that express PD-L1 (Iwai *et al.* (2005) *Int. Immunol.* 17:133-144).

10 Optionally, antibodies to PD-L1 can be combined with an immunogenic agent, such as cancerous cells, purified tumor antigens (including recombinant proteins, peptides, and carbohydrate molecules), cells, and cells transfected with genes encoding immune stimulating cytokines (He *et al.* (2004) *J. Immunol.* 173:4919-28). Non-limiting examples of tumor vaccines that can be used include peptides of melanoma antigens, such as peptides of gp100, MAGE antigens, Trp-2, MART1 and/or tyrosinase, or tumor cells
15 transfected to express the cytokine GM-CSF (discussed further below).

 In humans, some tumors have been shown to be immunogenic such as melanomas. It is anticipated that by raising the threshold of T cell activation by PD-L1 blockade, we may expect to activate tumor responses in the host.

20 PD-L1 blockade is likely to be most effective when combined with a vaccination protocol. Many experimental strategies for vaccination against tumors have been devised (see Rosenberg, S., 2000, Development of Cancer Vaccines, ASCO Educational Book Spring: 60-62; Logothetis, C., 2000, ASCO Educational Book Spring: 300-302; Khayat, D. 2000, ASCO Educational Book Spring: 414-428; Foon, K. 2000, ASCO Educational
25 Book Spring: 730-738; see also Restifo, N. and Sznol, M., Cancer Vaccines, Ch. 61, pp. 3023-3043 in DeVita, V. *et al.* (eds.), 1997, Cancer: Principles and Practice of Oncology. Fifth Edition). In one of these strategies, a vaccine is prepared using autologous or allogeneic tumor cells. These cellular vaccines have been shown to be most effective when the tumor cells are transduced to express GM-CSF. GM-CSF has been shown to be
30 a potent activator of antigen presentation for tumor vaccination (Dranoff *et al.* (1993) *Proc. Natl. Acad. Sci U.S.A.* 90: 3539-43).

 The study of gene expression and large scale gene expression patterns in various tumors has led to the definition of so-called tumor specific antigens (Rosenberg, SA

(1999) *Immunity* 10: 281-7). In many cases, these tumor specific antigens are differentiation antigens expressed in the tumors and in the cell from which the tumor arose, for example melanocyte antigens gp100, MAGE antigens, and Trp-2. More importantly, many of these antigens can be shown to be the targets of tumor specific T cells found in the host. PD-L1 blockade may be used in conjunction with a collection of recombinant proteins and/or peptides expressed in a tumor in order to generate an immune response to these proteins. These proteins are normally viewed by the immune system as self antigens and are therefore tolerant to them. The tumor antigen may also include the protein telomerase, which is required for the synthesis of telomeres of chromosomes and which is expressed in more than 85% of human cancers and in only a limited number of somatic tissues (Kim, N *et al.* (1994) *Science* 266: 2011-2013). (These somatic tissues may be protected from immune attack by various means). Tumor antigen may also be "neo-antigens" expressed in cancer cells because of somatic mutations that alter protein sequence or create fusion proteins between two unrelated sequences (i.e. bcr-abl in the Philadelphia chromosome), or idiotype from B cell tumors.

Other tumor vaccines may include the proteins from viruses implicated in human cancers such as Human Papilloma Viruses (HPV), Hepatitis Viruses (HBV and HCV) and Kaposi's Herpes Sarcoma Virus (KHSV). Another form of tumor specific antigen which may be used in conjunction with PD-L1 blockade is purified heat shock proteins (HSP) isolated from the tumor tissue itself. These heat shock proteins contain fragments of proteins from the tumor cells and these HSPs are highly efficient at delivery to antigen presenting cells for eliciting tumor immunity (Suot, R & Srivastava, P (1995) *Science* 269:1585-1588; Tamura, Y. *et al.* (1997) *Science* 278:117-120).

Dendritic cells (DC) are potent antigen presenting cells that can be used to prime antigen-specific responses. DC's can be produced *ex vivo* and loaded with various protein and peptide antigens as well as tumor cell extracts (Nestle, F. *et al.* (1998) *Nature Medicine* 4: 328-332). DCs may also be transduced by genetic means to express these tumor antigens as well. DCs have also been fused directly to tumor cells for the purposes of immunization (Kugler, A. *et al.* (2000) *Nature Medicine* 6:332-336). As a method of vaccination, DC immunization may be effectively combined with PD-L1 blockade to activate more potent anti-tumor responses.

PD-L1 blockade may also be combined with standard cancer treatments. PD-L1 blockade may be effectively combined with chemotherapeutic regimes. In these

instances, it may be possible to reduce the dose of chemotherapeutic reagent administered (Mokyr, M. *et al.* (1998) *Cancer Research* 58: 5301-5304). An example of such a combination is an anti-PD-L1 antibody in combination with decarbazine for the treatment of melanoma. Another example of such a combination is an anti-PD-L1 antibody in
5 combination with interleukin-2 (IL-2) for the treatment of melanoma. The scientific rationale behind the combined use of PD-L1 blockade and chemotherapy is that cell death, that is a consequence of the cytotoxic action of most chemotherapeutic compounds, should result in increased levels of tumor antigen in the antigen presentation pathway. Other combination therapies that may result in synergy with PD-L1 blockade through cell
10 death are radiation, surgery, and hormone deprivation. Each of these protocols creates a source of tumor antigen in the host. Angiogenesis inhibitors may also be combined with PD-L1 blockade. Inhibition of angiogenesis leads to tumor cell death which may feed tumor antigen into host antigen presentation pathways.

PD-L1 blocking antibodies can also be used in combination with bispecific
15 antibodies that target Fc alpha or Fc gamma receptor-expressing effectors cells to tumor cells (see, *e.g.*, U.S. Pat. Nos. 5,922,845 and 5,837,243). Bispecific antibodies can be used to target two separate antigens. For example anti-Fc receptor/anti tumor antigen (*e.g.*, Her-2/neu) bispecific antibodies have been used to target macrophages to sites of tumor. This targeting may more effectively activate tumor specific responses. The T cell arm of these
20 responses would be augmented by the use of PD-L1 blockade. Alternatively, antigen may be delivered directly to DCs by the use of bispecific antibodies which bind to tumor antigen and a dendritic cell specific cell surface marker.

Tumors evade host immune surveillance by a large variety of mechanisms. Many of these mechanisms may be overcome by the inactivation of proteins which are
25 expressed by the tumors and which are immunosuppressive. These include among others TGF-beta (Kehrl, J. *et al.* (1986) *J. Exp. Med.* 163: 1037-1050), IL-10 (Howard, M. & O'Garra, A. (1992) *Immunology Today* 13: 198-200), and Fas ligand (Hahne, M. *et al.* (1996) *Science* 274: 1363-1365). Antibodies to each of these entities may be used in combination with anti-PD-L1 to counteract the effects of the immunosuppressive agent
30 and favor tumor immune responses by the host.

Other antibodies which may be used to activate host immune responsiveness can be used in combination with anti-PD-L1. These include molecules on the surface of dendritic cells which activate DC function and antigen presentation. Anti-CD40

antibodies are able to substitute effectively for T cell helper activity (Ridge, J. *et al.* (1998) *Nature* 393: 474-478) and can be used in conjunction with PD-L1 antibodies (Ito, N. *et al.* (2000) *Immunobiology* 201 (5) 527-40). Activating antibodies to T cell costimulatory molecules such as OX-40 (Weinberg, A. *et al.* (2000) *Immunol* 164: 2160-2169), 4-1BB (Melero, I. *et al.* (1997) *Nature Medicine* 3: 682-685 (1997), and ICOS (Hutloff, A. *et al.* (1999) *Nature* 397: 262-266) as well as antibodies which block the activity of negative costimulatory molecules such as CTLA-4 (e.g., US Patent No. 5,811,097) or BTLA (Watanabe, N. *et al.* (2003) *Nat Immunol* 4:670-9), B7-H4 (Sica, GL *et al.* (2003) *Immunity* 18:849-61) may also provide for increased levels of T cell activation.

Bone marrow transplantation is currently being used to treat a variety of tumors of hematopoietic origin. While graft versus host disease is a consequence of this treatment, therapeutic benefit may be obtained from graft vs. tumor responses. PD-L1 blockade can be used to increase the effectiveness of the donor engrafted tumor specific T cells.

There are also several experimental treatment protocols that involve *ex vivo* activation and expansion of antigen specific T cells and adoptive transfer of these cells into recipients in order to antigen-specific T cells against tumor (Greenberg, R. & Riddell, S. (1999) *Science* 285: 546-51). These methods may also be used to activate T cell responses to infectious agents such as CMV. *Ex vivo* activation in the presence of anti-PD-L1 antibodies may be expected to increase the frequency and activity of the adoptively transferred T cells.

Infectious Diseases

Other methods of the invention are used to treat patients that have been exposed to particular toxins or pathogens. Accordingly, another aspect of the invention provides a method of treating an infectious disease in a subject comprising administering to the subject an anti-PD-L1 antibody, or antigen-binding portion thereof, such that the subject is treated for the infectious disease. Preferably, the antibody is a human anti-human PD-L1 antibody (such as any of the human anti-PD-L1 antibodies described herein). Additionally or alternatively, the antibody can be a chimeric or humanized antibody.

Similar to its application to tumors as discussed above, antibody mediated PD-L1 blockade can be used alone, or as an adjuvant, in combination with vaccines, to stimulate the immune response to pathogens, toxins, and self-antigens. Examples of pathogens for which this therapeutic approach may be particularly useful, include pathogens for which

there is currently no effective vaccine, or pathogens for which conventional vaccines are less than completely effective. These include, but are not limited to HIV, Hepatitis (A, B, & C), Influenza, Herpes, Giardia, Malaria, Leishmania, Staphylococcus aureus, Pseudomonas Aeruginosa. PD-L1 blockade is particularly useful against established
5 infections by agents such as HIV that present altered antigens over the course of the infections. These novel epitopes are recognized as foreign at the time of anti-human PD-L1 administration, thus provoking a strong T cell response that is not dampened by negative signals through PD-L1.

Some examples of pathogenic viruses causing infections treatable by methods of
10 the invention include HIV, hepatitis (A, B, or C), herpes virus (*e.g.*, VZV, HSV-1, HAV-6, HSV-II, and CMV, Epstein Barr virus), adenovirus, influenza virus, flaviviruses, echovirus, rhinovirus, coxsackie virus, cornovirus, respiratory syncytial virus, mumps virus, rotavirus, measles virus, rubella virus, parvovirus, vaccinia virus, HTLV virus, dengue virus, papillomavirus, molluscum virus, poliovirus, rabies virus, JC virus and
15 arboviral encephalitis virus.

Some examples of pathogenic bacteria causing infections treatable by methods of the invention include chlamydia, rickettsial bacteria, mycobacteria, staphylococci, streptococci, pneumonococci, meningococci and conococci, klebsiella, proteus, serratia, pseudomonas, legionella, diphtheria, salmonella, bacilli, cholera, tetanus, botulism,
20 anthrax, plague, leptospirosis, and Lyme's disease bacteria.

Some examples of pathogenic fungi causing infections treatable by methods of the invention include Candida (*albicans*, *krusei*, *glabrata*, *tropicalis*, etc.), Cryptococcus neoformans, Aspergillus (*fumigatus*, *niger*, etc.), Genus Mucorales (*mucor*, *absidia*, *rhizopus*), Sporothrix *schenkii*, Blastomyces *dermatitidis*, Paracoccidioides *brasiliensis*,
25 Coccidioides *immitis* and Histoplasma *capsulatum*.

Some examples of pathogenic parasites causing infections treatable by methods of the invention include Entamoeba *histolytica*, Balantidium *coli*, Naegleria *fowleri*, Acanthamoeba *sp.*, Giardia *lamblia*, Cryptosporidium *sp.*, Pneumocystis *carinii*, Plasmodium *vivax*, Babesia *microti*, Trypanosoma *brucei*, Trypanosoma *cruzi*,
30 Leishmania *donovani*, Toxoplasma *gondi*, Nippostrongylus *brasiliensis*.

In all of the above methods, PD-L1 blockade can be combined with other forms of immunotherapy such as cytokine treatment (*e.g.*, interferons, GM-CSF, G-CSF, IL-2), or bispecific antibody therapy, which provides for enhanced presentation of tumor antigens

(see, e.g., Holliger (1993) *Proc. Natl. Acad. Sci. USA* 90:6444-6448; Poljak (1994) *Structure* 2:1121-1123).

Autoimmune reactions

Anti-PD-L1 antibodies may provoke and amplify autoimmune responses. Indeed, induction of anti-tumor responses using tumor cell and peptide vaccines reveals that many anti-tumor responses involve anti-self reactivities (depigmentation observed in anti-CTLA-4 + GM-CSF-modified B16 melanoma in van Elsas *et al.* *supra*; depigmentation in Trp-2 vaccinated mice (Overwijk, W. *et al.* (1999) *Proc. Natl. Acad. Sci. U.S.A.* 96: 2982-2987); autoimmune prostatitis evoked by TRAMP tumor cell vaccines (Hurwitz, A. (2000) *supra*), melanoma peptide antigen vaccination and vitiligo observed in human clinical trials (Rosenberg, SA and White, DE (1996) *J. Immunother Emphasis Tumor Immunol* 19 (1): 81-4).

Therefore, it is possible to consider using anti-PD-L1 blockade in conjunction with various self proteins in order to devise vaccination protocols to efficiently generate immune responses against these self proteins for disease treatment. For example, Alzheimer's disease involves inappropriate accumulation of A β peptide in amyloid deposits in the brain; antibody responses against amyloid are able to clear these amyloid deposits (Schenk *et al.*, (1999) *Nature* 400: 173-177).

Other self proteins may also be used as targets such as IgE for the treatment of allergy and asthma, and TNF α for rheumatoid arthritis. Finally, antibody responses to various hormones may be induced by the use of anti-PD-L1 antibody. Neutralizing antibody responses to reproductive hormones may be used for contraception. Neutralizing antibody response to hormones and other soluble factors that are required for the growth of particular tumors may also be considered as possible vaccination targets.

Analogous methods as described above for the use of anti-PD-L1 antibody can be used for induction of therapeutic autoimmune responses to treat patients having an inappropriate accumulation of other self-antigens, such as amyloid deposits, including A β in Alzheimer's disease, cytokines such as TNF α , and IgE.

Vaccines

Anti-PD-L1 antibodies may be used to stimulate antigen-specific immune responses by coadministration of an anti-PD-L1 antibody with an antigen of interest (e.g., a vaccine). Accordingly, in another aspect the invention provides a method of enhancing

an immune response to an antigen in a subject, comprising administering to the subject: (i) the antigen; and (ii) an anti-PD-L1 antibody, or antigen-binding portion thereof, such that an immune response to the antigen in the subject is enhanced. Preferably, the antibody is a human anti-human PD-L1 antibody (such as any of the human anti-PD-L1 antibodies described herein). Additionally or alternatively, the antibody can be a chimeric or humanized antibody. The antigen can be, for example, a tumor antigen, a viral antigen, a bacterial antigen or an antigen from a pathogen. Non-limiting examples of such antigens include those discussed in the sections above, such as the tumor antigens (or tumor vaccines) discussed above, or antigens from the viruses, bacteria or other pathogens described above.

Anti-PD-L1 antibodies may also be used to abrogate secondary effects associated with diseases such as T cell suppressed wasting disease with colitis (Kanai *et al.* (2003) *J. Immunol.* 171:4156-63). Accordingly, in another aspect the invention provides a method of abrogating leukocyte infiltration, decreasing production of IFN- γ , IL-2, and IFN- α by T cells. Preferably, the antibody is a human anti-human PD-L1 antibody (such as any of the human anti-PD-L1 antibodies described herein). Additionally or alternatively, the antibody can be a chimeric or humanized antibody.

Anti-PD-L1 antibodies may also be used to treat diseases such as chronic inflammatory diseases, such as lichen planus, a T-cell mediated chronic inflammatory mucocutaneous disease (Youngnak-Piboonratanakit *et al.* (2004) *Immunol Letters* 94:215-22). Accordingly, in another aspect the invention provides a method of abrogating chronic inflammatory disease by T cells. Preferably, the antibody is a human anti-human PD-L1 antibody (such as any of the human anti-PD-L1 antibodies described herein). Additionally or alternatively, the antibody can be a chimeric or humanized antibody.

Suitable routes of administering the antibody compositions (*e.g.*, human monoclonal antibodies, multispecific and bispecific molecules and immunoconjugates) of the invention *in vivo* and *in vitro* are well known in the art and can be selected by those of ordinary skill. For example, the antibody compositions can be administered by injection (*e.g.*, intravenous or subcutaneous). Suitable dosages of the molecules used will depend on the age and weight of the subject and the concentration and/or formulation of the antibody composition.

As previously described, human anti-PD-L1 antibodies of the invention can be co-administered with one or other more therapeutic agents, *e.g.*, a cytotoxic agent, a radiotoxic agent or an immunosuppressive agent. The antibody can be linked to the agent (as an immunocomplex) or can be administered separate from the agent. In the latter case
5 (separate administration), the antibody can be administered before, after or concurrently with the agent or can be co-administered with other known therapies, *e.g.*, an anti-cancer therapy, *e.g.*, radiation. Such therapeutic agents include, among others, anti-neoplastic agents such as doxorubicin (adriamycin), cisplatin bleomycin sulfate, carmustine, chlorambucil, and cyclophosphamide hydroxyurea which, by themselves, are only
10 effective at levels which are toxic or subtoxic to a patient. Cisplatin is intravenously administered as a 100 mg/ dose once every four weeks and adriamycin is intravenously administered as a 60-75 mg/ml dose once every 21 days. Co-administration of the human anti-PD-L1 antibodies, or antigen binding fragments thereof, of the present invention with chemotherapeutic agents provides two anti-cancer agents which operate via different
15 mechanisms which yield a cytotoxic effect to human tumor cells. Such co-administration can solve problems due to development of resistance to drugs or a change in the antigenicity of the tumor cells which would render them unreactive with the antibody.

Also within the scope of the present invention are kits comprising the antibody compositions of the invention (*e.g.*, human antibodies, bispecific or multispecific
20 molecules, or immunoconjugates) and instructions for use. The kit can further contain a least one additional reagent, or one or more additional human antibodies of the invention (*e.g.*, a human antibody having a complementary activity which binds to an epitope in PD-L1 antigen distinct from the first human antibody). Kits typically include a label indicating the intended use of the contents of the kit. The term label includes any writing,
25 or recorded material supplied on or with the kit, or which otherwise accompanies the kit.

The present invention is further illustrated by the following examples which should not be construed as further limiting. The contents of all figures and all references, patents and published patent applications cited throughout this application are expressly
30 incorporated herein by reference.

Examples

Example 1: Generation of Human Monoclonal Antibodies Against PD-L1

Antigen

Immunization protocols utilized as antigen both (i) a recombinant fusion protein comprising the extracellular portion of PD-L1, and (ii) membrane bound full-length PD-L1. Both antigens were generated by recombinant transfection methods in a CHO cell line.

Transgenic mice (KM-Mouse® colony)

Fully human monoclonal antibodies to PD-L1 were prepared using the KM strain of transgenic transchromosomic mice, which expresses human antibody genes. In this mouse strain, the endogenous mouse kappa light chain gene has been homozygously disrupted as described in Chen *et al.* (1993) *EMBO J.* 12:811-820 and the endogenous mouse heavy chain gene has been homozygously disrupted as described in Example 1 of PCT Publication WO 01/09187. Furthermore, this mouse strain carries a human kappa light chain transgene, KCo5, as described in Fishwild *et al.* (1996) *Nature Biotechnology* 14:845-851, and a SC20 transchromosome as described in PCT Publication WO 02/43478.

KM-Mouse® Immunizations

To generate fully human monoclonal antibodies to PD-L1, a cohort of mice of the KM-Mouse® strain were immunized with purified recombinant PD-L1-Ig and PD-L1-transfected CHO cells as antigen. General immunization schemes for HuMab mice are described in Lonberg, N. *et al.* (1994) *Nature* 368(6474): 856-859; Fishwild, D. *et al.* (1996) *Nature Biotechnology* 14: 845-851 and PCT Publication WO 98/24884. The mice were 6-16 weeks of age upon the first infusion of antigen. A purified recombinant preparation (5-50 µg) of PD-L1-Ig antigen and 5 -10x10⁶ cells were used to immunize the HuMab mice intraperitoneally (IP), subcutaneously (Sc) or via footpad injection.

Transgenic mice were immunized twice with antigen in complete Freund's adjuvant or Ribi adjuvant IP, followed by 3-21 days IP (up to a total of 11 immunizations) with the antigen in incomplete Freund's or Ribi adjuvant. The immune response was monitored by retroorbital bleeds. The plasma was screened by ELISA (as described below), and mice with sufficient titers of anti-PD-L1 human immunoglobulin

were used for fusions. Mice were boosted intravenously with antigen 3 days before sacrifice and removal of the spleen. Typically, 10-35 fusions for each antigen were performed. Several dozen mice were immunized for each antigen.

Selection of KM-Mouse[®] Producing Anti-PD-L1 Antibodies:

- 5 To select HuMab mice producing antibodies that bound PD-L1, sera from immunized mice were tested by ELISA as described by Fishwild, D. *et al.* (1996). Briefly, microtiter plates were coated with purified recombinant PD-L1 fusion protein from transfected CHO cells at 1-2 µg/ml in PBS, 100 µl/wells incubated 4 °C overnight then blocked with 200 µl/well of 5% fetal bovine serum in PBS/Tween (0.05%).
- 10 Dilutions of sera from PD-L1-immunized mice were added to each well and incubated for 1-2 hours at ambient temperature. The plates were washed with PBS/Tween and then incubated with a goat-anti-human IgG polyclonal antibody conjugated with horseradish peroxidase (HRP) for 1 hour at room temperature. After washing, the plates were developed with ABTS substrate (Sigma, A-1888, 0.22 mg/ml) and analyzed by
- 15 spectrophotometer at OD 415-495. Mice that developed the highest titers of anti-PD-L1 antibodies were used for fusions. Fusions were performed as described below and hybridoma supernatants were tested for anti-PD-L1 activity by ELISA.

Generation of Hybridomas Producing Human Monoclonal Antibodies to PD-L1:

- 20 The mouse splenocytes, isolated from a KM mouse, were fused with PEG to a mouse myeloma cell line based upon standard protocols. The resulting hybridomas were then screened for the production of antigen-specific antibodies. Single cell suspensions of splenocytes from immunized mice were fused to one-fourth the number of SP2/0 nonsecreting mouse myeloma cells (ATCC, CRL 1581) with 50% PEG (Sigma). Cells were plated at approximately 1×10^5 /well in flat bottom microtiter plate, followed by
- 25 about two week incubation in selective medium containing 10% fetal bovine serum, 10% P388D1 (ATCC, CRL TIB-63) conditioned medium, 3-5% origen (IGEN) in DMEM (Mediatech, CRL 10013, with high glucose, L-glutamine and sodium pyruvate) plus 5 mM HEPES, 0.055 mM 2-mercaptoethanol, 50 mg/ml gentamycin and 1x HAT (Sigma, CRL P-7185). After 1-2 weeks, cells were cultured in medium in which the HAT was
- 30 replaced with HT. Individual wells were then screened by ELISA (described above) for human anti-PD-L1 monoclonal IgG antibodies. Once extensive hybridoma growth occurred, medium was monitored usually after 10-14 days. The antibody-secreting hybridomas were re-plated, screened again and, if still positive for human IgG, anti-PD-

L1 monoclonal antibodies were subcloned at least twice by limiting dilution. The stable subclones were then cultured *in vitro* to generate small amounts of antibody in tissue culture medium for further characterization.

Hybridoma clones 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and
5 13G4 were selected for further analysis.

Example 2: Structural Characterization of Human Monoclonal Antibodies 3G10, 12A4, and 10A5

The cDNA sequences encoding the heavy and light chain variable regions of the 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4 monoclonal
10 antibodies were obtained from the 3G10, 12A4, 10A5, 5F8, 10H10, 1B12, 7H1, 11E6, 12B7, and 13G4 hybridomas, respectively, using standard PCR techniques and were sequenced using standard DNA sequencing techniques.

The nucleotide and amino acid sequences of the heavy chain variable region of 3G10 are shown in Figure 1A and in SEQ ID NO:81 and 1, respectively.

15 The nucleotide and amino acid sequences of the light chain variable region of 3G10 are shown in Figure 1B and in SEQ ID NO:91 and 11, respectively.

Comparison of the 3G10 heavy chain immunoglobulin sequence to the known human germline immunoglobulin heavy chain sequences demonstrated that the 3G10 heavy chain utilizes a VH segment from human germline VH 1-18, an undetermined D
20 segment, and a JH segment from human germline JH 6b. The alignment of the 3G10 VH sequence to the germline VH 1-18 sequence is shown in Figure 11. Further analysis of the 3G10 VH sequence using the Kabat system of CDR region determination led to the delineation of the heavy chain CDR1, CDR2 and CD3 regions as shown in Figures 1A and 11, and in SEQ ID NOs:21, 31 and 41, respectively.

25 Comparison of the 3G10 light chain immunoglobulin sequence to the known human germline immunoglobulin light chain sequences demonstrated that the 3G10 light chain utilizes a VL segment from human germline VK L6 and a JK segment from human germline JK 1. The alignment of the 3G10 VL sequence to the germline VK L6 sequence is shown in Figure 21. Further analysis of the 3G10 VL sequence using the Kabat system
30 of CDR region determination led to the delineation of the light chain CDR1, CDR2 and CD3 regions as shown in Figures 1B and 21, and in SEQ ID NOs:51, 61 and 71, respectively.

The nucleotide and amino acid sequences of the heavy chain variable region of 12A4 are shown in Figure 2A and in SEQ ID NO:82 and 2, respectively.

The nucleotide and amino acid sequences of the light chain variable region of 12A4 are shown in Figure 2B and in SEQ ID NO:92 and 12, respectively.

5 Comparison of the 12A4 heavy chain immunoglobulin sequence to the known human germline immunoglobulin heavy chain sequences demonstrated that the 12A4 heavy chain utilizes a VH segment from human germline VH 1-69, a D segment from human germline 3-10, and a JH segment from human germline JH 6b. The alignment of the 12A4 VH sequence to the germline VH 1-69 sequence is shown in Figure 12. Further
10 analysis of the 12A4 VH sequence using the Kabat system of CDR region determination led to the delineation of the heavy chain CDR1, CDR2 and CD3 regions as shown in Figures 2A and 12, and in SEQ ID NOs:22, 32 and 42, respectively.

 Comparison of the 12A4 light chain immunoglobulin sequence to the known human germline immunoglobulin light chain sequences demonstrated that the 12A4 light
15 chain utilizes a VL segment from human germline VK L6 and a JK segment from human germline JK 1. The alignment of the 12A4 VL sequence to the germline VK L6 sequence is shown in Figure 22. Further analysis of the 12A4 VL sequence using the Kabat system of CDR region determination led to the delineation of the light chain CDR1, CDR2 and CD3 regions as shown in Figures 2B and 22, and in SEQ ID NOs:52, 62 and
20 72, respectively.

The nucleotide and amino acid sequences of the heavy chain variable region of 10A5 are shown in Figure 3A and in SEQ ID NO:83 and 3, respectively.

The nucleotide and amino acid sequences of the light chain variable region of 10A5 are shown in Figure 3B and in SEQ ID NO:93 and 13, respectively.

25 Comparison of the 10A5 heavy chain immunoglobulin sequence to the known human germline immunoglobulin heavy chain sequences demonstrated that the 10A5 heavy chain utilizes a VH segment from human germline VH 1-3, a D segment from human germline 5-5, and a JH segment from human germline JH 4b. The alignment of the 10A5 VH sequence to the germline VH 1-3 sequence is shown in Figure 13. Further
30 analysis of the 10A5 VH sequence using the Kabat system of CDR region determination led to the delineation of the heavy chain CDR1, CDR2 and CD3 regions as shown in Figures 3A and 13, and in SEQ ID NOs:23, 33, and 43, respectively.

Comparison of the 10A5 light chain immunoglobulin sequence to the known human germline immunoglobulin light chain sequences demonstrated that the 10A5 light chain utilizes a VL segment from human germline VK L15 and a JK segment from human germline JK 2. The alignment of the 10A5 VL sequence to the germline VK L15 sequence is shown in Figure 23. Further analysis of the 10A5 VL sequence using the Kabat system of CDR region determination led to the delineation of the light chain CDR1, CDR2 and CD3 regions as shown in Figures 3B and 23, and in SEQ ID NOs:53, 63, and 73, respectively.

The nucleotide and amino acid sequences of the heavy chain variable region of 5F8 are shown in Figure 4A and in SEQ ID NO:84 and 4, respectively.

The nucleotide and amino acid sequences of the light chain variable region of 5F8 are shown in Figure 4B and in SEQ ID NO:94 and 14, respectively.

Comparison of the 5F8 heavy chain immunoglobulin sequence to the known human germline immunoglobulin heavy chain sequences demonstrated that the 5F8 heavy chain utilizes a VH segment from human germline VH 1-69, a D segment from human germline 6-13, and a JH segment from human germline JH 4b. The alignment of the 5F8 VH sequence to the germline VH 1-69 sequence is shown in Figure 14. Further analysis of the 5F8 VH sequence using the Kabat system of CDR region determination led to the delineation of the heavy chain CDR1, CDR2 and CD3 regions as shown in Figures 4A and 14, and in SEQ ID NOs:24, 34, and 44, respectively.

Comparison of the 5F8 light chain immunoglobulin sequence to the known human germline immunoglobulin light chain sequences demonstrated that the 5F8 light chain utilizes a VL segment from human germline VK A27 and a JK segment from human germline JK 1. The alignment of the 5F8 VL sequence to the germline VK A27 sequence is shown in Figure 24. Further analysis of the 5F8 VL sequence using the Kabat system of CDR region determination led to the delineation of the light chain CDR1, CDR2 and CD3 regions as shown in Figures 4B and 24, and in SEQ ID NOs:54, 64, and 74, respectively.

The nucleotide and amino acid sequences of the heavy chain variable region of 10H10 are shown in Figure 5A and in SEQ ID NO:85 and 5, respectively.

The nucleotide and amino acid sequences of the light chain variable region of 10H10 are shown in Figure 5B and in SEQ ID NO:95 and 15, respectively.

Comparison of the 10H10 heavy chain immunoglobulin sequence to the known human germline immunoglobulin heavy chain sequences demonstrated that the 10H10 heavy chain utilizes a VH segment from human germline VH 3-9, a D segment from human germline 4-17, and a JH segment from human germline JH 4b. The alignment of the 10H10 VH sequence to the germline VH 3-9 sequence is shown in Figure 15. Further analysis of the 10H10 VH sequence using the Kabat system of CDR region determination led to the delineation of the heavy chain CDR1, CDR2 and CD3 regions as shown in Figures 5A and 15, and in SEQ ID NOs:25, 35, and 45, respectively.

Comparison of the 10H10 light chain immunoglobulin sequence to the known human germline immunoglobulin light chain sequences demonstrated that the 10H10 light chain utilizes a VL segment from human germline VK L15 and a JK segment from human germline JK 2. The alignment of the 10H10 VL sequence to the germline VK L15 sequence is shown in Figure 25. Further analysis of the 10H10 VL sequence using the Kabat system of CDR region determination led to the delineation of the light chain CDR1, CDR2 and CD3 regions as shown in Figures 5B and 25, and in SEQ ID NOs:55, 65, and 75, respectively.

The nucleotide and amino acid sequences of the heavy chain variable region of 1B12 are shown in Figure 6A and in SEQ ID NO:86 and 6, respectively.

The nucleotide and amino acid sequences of the light chain variable region of 1B12 are shown in Figure 6B and in SEQ ID NO:96 and 16, respectively.

Comparison of the 1B12 heavy chain immunoglobulin sequence to the known human germline immunoglobulin heavy chain sequences demonstrated that the 1B12 heavy chain utilizes a VH segment from human germline VH 1-69, a D segment from human germline 3-10, and a JH segment from human germline JH 6b. The alignment of the 1B12 VH sequence to the germline VH 1-69 sequence is shown in Figure 16. Further analysis of the 1B12 VH sequence using the Kabat system of CDR region determination led to the delineation of the heavy chain CDR1, CDR2 and CD3 regions as shown in Figures 6A and 16, and in SEQ ID NOs:26, 36, and 46, respectively.

Comparison of the 1B12 light chain immunoglobulin sequence to the known human germline immunoglobulin light chain sequences demonstrated that the 1B12 light chain utilizes a VL segment from human germline VK L6 and a JK segment from human germline JK 1. The alignment of the 1B12 VL sequence to the germline VK L6 sequence is shown in Figure 26. Further analysis of the 1B12 VL sequence using the Kabat system

of CDR region determination led to the delineation of the light chain CDR1, CDR2 and CD3 regions as shown in Figures 6B and 26; and in SEQ ID NOs:56, 66, and 76, respectively.

5 The nucleotide and amino acid sequences of the heavy chain variable region of 7H1 are shown in Figure 7A and in SEQ ID NO:87 and 7, respectively.

The nucleotide and amino acid sequences of the light chain variable region of 7H1 are shown in Figure 7B and in SEQ ID NO:97 and 17, respectively.

Comparison of the 7H1 heavy chain immunoglobulin sequence to the known human germline immunoglobulin heavy chain sequences demonstrated that the 7H1
10 heavy chain utilizes a VH segment from human germline VH 1-69, a D segment from human germline 3-10, and a JH segment from human germline JH 6b. The alignment of the 7H1 VH sequence to the germline VH 1-69 sequence is shown in Figure 17. Further analysis of the 7H1 VH sequence using the Kabat system of CDR region determination led to the delineation of the heavy chain CDR1, CDR2 and CD3 regions as shown in
15 Figures 7A and 17, and in SEQ ID NOs:27, 37, and 47, respectively.

Comparison of the 7H1 light chain immunoglobulin sequence to the known human germline immunoglobulin light chain sequences demonstrated that the 7H1 light chain utilizes a VL segment from human germline VK L6 and a JK segment from human germline JK 1. The alignment of the 7H1 VL sequence to the germline VK L6 sequence
20 is shown in Figure 27. Further analysis of the 7H1 VL sequence using the Kabat system of CDR region determination led to the delineation of the light chain CDR1, CDR2 and CD3 regions as shown in Figures 7B and 27, and in SEQ ID NOs:57, 67, and 77, respectively.

25 The nucleotide and amino acid sequences of the heavy chain variable region of 11E6 are shown in Figure 4A and in SEQ ID NO:84 and 4, respectively.

The nucleotide and amino acid sequences of the light chain variable region of 11E6 are shown in Figure 4B and in SEQ ID NO:94 and 14, respectively.

Comparison of the 11E6 heavy chain immunoglobulin sequence to the known human germline immunoglobulin heavy chain sequences demonstrated that the 11E6
30 heavy chain utilizes a VH segment from human germline VH 1-69, a D segment from human germline 6-19, and a JH segment from human germline JH 6c. The alignment of the 11E6 VH sequence to the germline VH 1-69 sequence is shown in Figure 18. Further analysis of the 11E6 VH sequence using the Kabat system of CDR region determination

led to the delineation of the heavy chain CDR1, CDR2 and CD3 regions as shown in Figures 8A and 18, and in SEQ ID NOs:28, 38, and 48, respectively.

Comparison of the 11E6 light chain immunoglobulin sequence to the known human germline immunoglobulin light chain sequences demonstrated that the 11E6 light chain utilizes a VL segment from human germline VK A27 and a JK segment from human germline JK 4. The alignment of the 11E6 VL sequence to the germline VK A27 sequence is shown in Figure 27. Further analysis of the 11E6 VL sequence using the Kabat system of CDR region determination led to the delineation of the light chain CDR1, CDR2 and CD3 regions as shown in Figures 8B and 28, and in SEQ ID NOs:58, 68, and 78, respectively. In addition, a second related clone included the VK sequence as shown in SEQ ID NO:109. This antibody is denoted herein as 11E6a.

The nucleotide and amino acid sequences of the heavy chain variable region of 12B7 are shown in Figure 9A and in SEQ ID NO:89 and 9, respectively.

The nucleotide and amino acid sequences of the light chain variable region of 12B7 are shown in Figure 9B and in SEQ ID NO:99 and 19, respectively.

Comparison of the 12B7 heavy chain immunoglobulin sequence to the known human germline immunoglobulin heavy chain sequences demonstrated that the 12B7 heavy chain utilizes a VH segment from human germline VH 1-69, a D segment from human germline 3-10, and a JH segment from human germline JH 6b. The alignment of the 12B7 VH sequence to the germline VH 1-69 sequence is shown in Figure 19. Further analysis of the 12B7 VH sequence using the Kabat system of CDR region determination led to the delineation of the heavy chain CDR1, CDR2 and CD3 regions as shown in Figures 9A and 19, and in SEQ ID NOs:29, 39, and 49, respectively.

Comparison of the 12B7 light chain immunoglobulin sequence to the known human germline immunoglobulin light chain sequences demonstrated that the 12B7 light chain utilizes a VL segment from human germline VK L6 and a JK segment from human germline JK 5. The alignment of the 12B7 VL sequence to the germline VK L6 sequence is shown in Figure 29. Further analysis of the 12B7 VL sequence using the Kabat system of CDR region determination led to the delineation of the light chain CDR1, CDR2 and CD3 regions as shown in Figures 9B and 29, and in SEQ ID NOs:59, 69, and 79, respectively.

The nucleotide and amino acid sequences of the heavy chain variable region of 13G4 are shown in Figure 10A and in SEQ ID NO:90 and 10, respectively.

The nucleotide and amino acid sequences of the light chain variable region of 13G4 are shown in Figure 10B and in SEQ ID NO:100 and 20, respectively.

Comparison of the 13G4 heavy chain immunoglobulin sequence to the known human germline immunoglobulin heavy chain sequences demonstrated that the 13G4 heavy chain utilizes a VH segment from human germline VH 3-9, a D segment from human germline 3-9, and a JH segment from human germline JH 4b. The alignment of the 13G4 VH sequence to the germline VH 3-9 sequence is shown in Figure 20. Further analysis of the 13G4 VH sequence using the Kabat system of CDR region determination led to the delineation of the heavy chain CDR1, CDR2 and CD3 regions as shown in Figures 10A and 20, and in SEQ ID NOs:30, 40, and 50, respectively.

Comparison of the 13G4 light chain immunoglobulin sequence to the known human germline immunoglobulin light chain sequences demonstrated that the 13G4 light chain utilizes a VL segment from human germline VK L18 and a JK segment from human germline JK 3. The alignment of the 13G4 VL sequence to the germline VK L18 sequence is shown in Figure 30. Further analysis of the 13G4 VL sequence using the Kabat system of CDR region determination led to the delineation of the light chain CDR1, CDR2 and CD3 regions as shown in Figures 10B and 30, and in SEQ ID NOs:60, 70, and 80, respectively.

Example 3: Characterization of Binding Specificity and Binding Kinetics of Anti-PD-L1 Human Monoclonal Antibodies

In this example, binding affinity and binding kinetics of anti-PD-L1 antibodies were examined by Biacore analysis. Binding specificity, and cross-competition were examined by flow cytometry.

Binding affinity and kinetics

Anti-PD-L1 antibodies were characterized for affinities and binding kinetics by Biacore analysis (Biacore AB, Uppsala, Sweden). Purified recombinant human PD-L1 fusion protein was covalently linked to a CM5 chip (carboxy methyl dextran coated chip) via primary amines, using standard amine coupling chemistry and kit provided by Biacore, to a density of 562 RUs. Binding was measured by flowing the antibodies in HBS EP buffer (provided by Biacore AB) at a concentration of 133 nM at a flow rate of 50 µl/min. The antigen-antibody association kinetics was followed for 1 minute and the dissociation kinetics was followed for 1 minute. The association and dissociation curves

were fit to a 1:1 Langmuir binding model using BIAevaluation software (Biacore AB). To minimize the effects of avidity in the estimation of the binding constants, only the initial segment of data corresponding to association and dissociation phases were used for fitting. The K_D , k_{on} and k_{off} values that were determined are shown in Table 2.

5

Table 2. Biacore binding data for PD-L1 human monoclonal antibodies.

Sample #	Sample ID	Affinity $K_D \times 10^{-9}$ (M)	On rate $k_{on} \times 10^5$ (1/Ms)	Off rate $k_{off} \times 10^{-4}$ 1/s
1	3G10	3.39	5.25	17.8
3	10A5	1.45	2.58	3.72

10 Additional binding data obtained by equilibrium binding method and analyzed on GraphPad Prism is shown in Table 3..

Table 3. Biacore equilibrium binding data for PD-L1 human monoclonal antibodies.

Clone ID	K_D (nM)	K_D (nM)
	37 C	25 C
12A4	1.94	0.76
7H1	2.15	nd
1B12	1.38	0.61
12B7	0.83	0.53
10A5	2.41	0.57
10H10	5.93	5.48
13G4	1.87	3.3
11E6	0.53	2.9
5F8	2.17	0.75

15 Binding specificity by flow cytometry

Chinese hamster ovary (CHO) cell lines that express recombinant human PD-L1 at the cell surface were developed and used to determine the specificity of PD-L1 human monoclonal antibodies by flow cytometry. CHO cells were transfected with expression plasmids containing full length cDNA encoding transmembrane forms of PD-L1.

20 Binding of the 3G10, 10A5, and 12A4 anti-PD-L1 human monoclonal antibodies was assessed by incubating the transfected cells with the anti-PD-L1 human monoclonal antibody. The cells were washed and binding was detected with a FITC-labeled anti-human IgG Ab. Flow cytometric analyses were performed using a FACScan flow cytometry (Becton Dickinson, San Jose, CA). The binding was compared to the parent
25 CHO cell line. The results are shown in Figures 32A (HuMAb 3G10), 32B (HuMAb

10A5) and 32C (HuMAb 12A4). Binding was also tested using varying concentrations of an anti-PD-L1 antibody. The results are shown in Figure 33. The anti-PD-L1 human monoclonal antibodies 3G10, 10A5, and 12A4 bound to the CHO cells transfected with PD-L1 in a concentration dependent manner. These data demonstrate that the anti-PD-L1 human monoclonal antibodies specifically bind to cell surface PD-L1.

Binding specificity by ELISA

The specificity of the anti-PD-L1 monoclonal antibodies was determined using a standard ELISA assay for binding to a human PD-L1 fusion to an immunoglobulin Fc region.

10 An Fc-fusion protein of human PD-L1 was tested for binding against the anti-PD-L1 human monoclonal antibodies 3G10, 12A4, and 10A5. Standard ELISA procedures were performed. The anti-PD-L1 human monoclonal antibodies were added at different concentrations. Goat-anti-human IgG (kappa chain-specific) polyclonal antibody conjugated with horseradish peroxidase (HRP) was used as secondary antibody. The results are shown in Figure 34. Each of the anti-PD-L1 human monoclonal antibodies 3G10, 12A4, and 10A5 bound with high specificity to PD-L1.

Example 4: Characterization of anti-PD-L1 antibody binding to PD-L1 expressed on the cell surface of human and monkey T cells

20 Anti-PD-L1 antibodies were tested by flow cytometry for binding to activated human or cynomolgus monkey T cells expressing PD-L1 on their surface.

Human or monkey T cells were activated by anti-CD3 antibody to induce PD-L1 expression prior to binding with a human anti-PD-L1 monoclonal antibody. Binding of the 3G10, 1B12, 13G4, and 12A4 anti-PD-L1 human monoclonal antibodies was assessed by incubating the activated cells with serial dilutions of the anti-PD-L1 human monoclonal antibodies. An isotype control antibody was used as a negative control. The cells were washed and binding was detected with a FITC-labeled anti-human Ig-kappa light chain Ab. Flow cytometric analyses were performed using a FACScalibur flow cytometer (Becton Dickinson, San Jose, CA). The results are shown in Figure 35 and 36. The anti-PD-L1 monoclonal antibodies 3G10, 1B12, 13G4, and 12A4 bound to activated human and monkey T cells. These data demonstrate that the anti-PD-L1 human monoclonal antibodies bind to human and cynomolgus monkey cell surface PD-L1.

Example 5: Characterization of anti-PD-L1 antibody binding to PD-L1 expressed on the cell surface of human T cells

Anti-PD-L1 antibodies were tested for binding to activated human T cells expressing PD-L1 on their cell surface by flow cytometry.

- 5 Human T cells were activated by anti-CD3 antibody to induce PD-L1 expression on T cells prior to binding with a human anti-PD-L1 monoclonal antibody. Binding of the 3G10, 10A5 and 12A4 anti-PD-L1 human monoclonal antibodies was assessed by incubating the activated T cells with the anti-PD-L1 human monoclonal antibodies at a concentration of 20 µg/ml. An isotype control antibody was used as a negative control.
- 10 The cells were washed and binding was detected with a FITC-labeled anti-human IgG Ab. Flow cytometric analyses were performed using a FACScalibur flow cytometry (Becton Dickinson, San Jose, CA). The results are shown in Figures 37A (HuMAb 3G10), 37B (HuMAb 10A5) and 37C (HuMAb 12A4). The anti-PD-L1 monoclonal antibodies 3G10, 10A5, and 12A4 bound to activated human T cells (bold line), as shown
- 15 in histogram plots compared to control (light line). These data demonstrate that the anti-PD-L1 human monoclonal antibodies bind to human cell surface PD-L1.

Example 6: Binding specificity by flow cytometry

- The ES-2 human ovarian carcinoma cell line that expresses human PD-L1 at the cell surface was used to determine the specificity of PD-L1 human monoclonal antibodies
- 20 by flow cytometry. ES-2 cells were treated overnight with 500 IU/mL of recombinant hIFN-γ to increase PD-L1 expression over the basal level. Binding of the 12A4, 1B12, 3G10, 10A5, 12B7, 13G4, 11E6, and 5F8 anti-PD-L1 human monoclonal antibodies was assessed by incubating the induced cells with serial dilutions of the anti-PD-L1 human monoclonal antibody. The cells were washed and binding was detected with a PE-labeled
- 25 anti-human IgG Ab. Flow cytometric analyses were performed using a FACScalibur flow cytometer (Becton Dickinson, San Jose, CA). The binding was compared to isotype control antibody. The results are shown in Figures 38. The anti-PD-L1 human monoclonal antibodies 12A4, 1B12, 3G10, 10A5, 12B7, 13G4, 11E6, and 5F8 bound to the hIFN-γ-induced ES-2 cells in a concentration dependent manner. These data
- 30 demonstrate that the anti-PD-L1 human monoclonal antibodies specifically bind to cell surface PD-L1.

Example 7: Effect of human anti-PD-L1 antibodies on cell proliferation and cytokine production in a Mixed Lymphocyte Reaction

A mixed lymphocyte reaction was employed to demonstrate the effect of blocking the PD-L1/PD-1 pathway to lymphocyte effector cells. T cells in the assay were tested for proliferation, IFN- γ secretion and IL-2 secretion in the presence or absence of an anti-PD-L1 human monoclonal antibody.

Human CD4⁺ T-cells were purified from PBMC using a CD4⁺ positive selection kit (Dyna1 Biotech). Dendritic cells were derived from purified monocytes cultured with 1000 U/ml of IL-4 and 500 U/ml of GM-CSF (R&D Biosystems) for seven days.

- 10 Monocytes were prepared using a monocyte negative selection kit (Miltenyi Biotech). Each culture contained 10^5 purified T-cells and 10^4 allogeneic dendritic cells in a total volume of 200 μ l. Anti-PD-L1 monoclonal antibody 10A5, 12A4, or 3G10 was added to each culture at different antibody concentrations. Either no antibody or an isotype control antibody was used as a negative control. The cells were cultured for 5 days at 37°C.
- 15 After day 5, 100 μ l of medium was taken from each culture for cytokine measurement. The levels of IFN- γ and IL-2 were measured using OptEIA ELISA kits (BD Biosciences). The cells were labeled with 3 H-thymidine, cultured for another 18 hours, and analyzed for cell proliferation. The results are shown in Figures 39A (T cell proliferation), 39B (IFN- γ secretion using HuMAb 10A5), 39C (IFN- γ secretion using HuMAb 12A4 or 3G10) and 39D (IL-2 secretion). The anti-PD-L1 human monoclonal antibody 10A5 promotes T-cell proliferation, IFN- γ secretion and IL-2 secretion in a concentration dependent manner. The anti-PD-L1 human monoclonal antibodies 12A4 and 3G10 also showed an increase in IFN- γ secretion. In contrast, cultures containing the control antibody did not show an increase in T cell proliferation, IFN- γ or IL-2 secretion.

- 25 In a separate experiment, an allogeneic mixed lymphocyte reaction (MLR) was employed to demonstrate the effect of blocking the PD-L1/PD-1 pathway in lymphocyte effector cells. T cells in the assay were tested for proliferation and IFN- γ secretion in the presence or absence of an anti-PD-L1 human monoclonal antibody or isotype control antibody.

- 30 Human CD4⁺ T-cells were purified from PBMC using a CD4⁺ negative selection kit (Miltenyi). Monocytes were prepared using a monocyte negative selection kit (Miltenyi Biotech). Dendritic cells were derived from purified monocytes cultured with 1000 U/ml of IL-4 and 500 U/ml of GM-CSF (R&D Biosystems) for seven days. Each MLR culture

contained 10^5 purified T-cells and 10^4 allogeneic dendritic cells in a total volume of 200 μ l. Anti-PD-L1 monoclonal antibody 12A4, 11B6, 3G10, 13G4, 1B12, 10A5, and 12B7 were added to each culture at different antibody concentrations. Either no antibody or an isotype control antibody was used as a negative control. The cells were cultured for 5 days at 37°C. On day 5, 50 μ l of medium was taken from each culture for cytokine measurement and replaced with an equal volume of culture medium containing 1 μ Ci of 3 H-thymidine. The cells were cultured for another 18 hours, harvested, and analyzed for cell proliferation. The levels of IFN- γ in the culture fluid were measured using an OptEIA hIFN- γ ELISA kit (BD Biosciences). The results are shown in Figure 40. The anti-PD-L1 human monoclonal antibodies promote T-cell proliferation and IFN- γ secretion in a concentration-dependent manner. In contrast, cultures containing the control antibody did not show an increase in T cell proliferation or IFN- γ secretion.

Example 8: Effect of human anti-PD-L1 antibody on function of T regulatory cells

T regulatory cells (CD4+, CD25+) are lymphocytes that suppress the immune response. The effect of the addition of T regulatory cells on proliferation and IFN- γ secretion in the allogeneic dendritic cell and T cell MLR in the presence or absence of an anti-PD-L1 human monoclonal antibody was tested.

T regulatory cells were purified from PBMC using a CD4+CD25+ regulatory T cell isolation kit (Miltenyi Biotec). T regulatory cells were added into a mixed lymphocyte reaction (see above) containing purified CD4+CD25- T cells and allogeneic dendritic cells in a 2:1 ratio of CD4+CD25- to T regulatory cells. Anti-PD-L1 monoclonal antibody 10A5 was added to each culture at a concentration of 10 μ g/ml. Either no antibody or an isotype control antibody was used as a negative control. The cells were cultured for 5 days at 37°C at which time the supernatants were analyzed for IFN- γ secretion using a Beadlyte cytokine detection system (Upstate). The cells were labeled with 3 H-thymidine, cultured for another 18 hours, and analyzed for cell proliferation. The results are shown in Figures 41A (T cell proliferation) and 41B (IFN- γ secretion). The addition of anti-PD-L1 human monoclonal antibody 10A5 promotes both T cell proliferation and IFN- γ secretion in cell cultures of allogeneic dendritic cells, T cells and T regulatory cells, indicating that anti-PD-L1 antibodies can reverse the effect of T regulatory cells in the allogeneic DC – T cell -MLR

In a separate experiment, human anti-PD-L1 antibodies 12A4 and 13G4, and a control antibody 1D12, were tested in the MLR assay with T regulatory cells. The results are shown in Figures 42 (T cell proliferation) and 43 (IFN- γ secretion). The addition of anti-PD-L1 human monoclonal antibodies 12A4 or 13G4 partially reverses the suppression of both T cell proliferation and IFN- γ secretion in cell cultures of allogeneic dendritic cells and T cells containing T-regulatory cells, indicating that anti-PD-L1 antibodies may have an effect on T-regulatory cells.

Example 9: Effect of anti-PD-1 antibodies on cytokine secretion by viral antigen-stimulated PBMC cells from a positive CMV responsive donor

CMV antigen-responsive human PBMC (Astarte Biologics, Redmond, WA) were cultured at 2×10^5 cells/well in flat bottom TC-treated 96 well plates, in the presence of 0.5 μ g/ml CMV lysate (Astarte Biologics) +/- titrated anti-PD-L1 antibodies. AIM-V medium (Invitrogen) supplemented with heat-inactivated FBS (10% final) was used at a total volume of 200 μ l/well. The cells were cultured for 4 days at 37°C, 5%CO₂ at which time culture supernatant was harvested for determination of secreted interferon- γ by ELISA (OptEIA hIFN- γ ELISA kit - BD Biosciences). The results are shown in Figure 44. The anti-PD-L1 human monoclonal antibodies promote IFN- γ secretion by CMV-specific T-cells in a dose-dependent manner. The most robust response was generated by antibodies 13G4, 1B12, and 12A4 compared to isotype control. These results shows that anti-PD-L1 HuMAbs can stimulate IFN- γ release in a memory T cell response from PBMC cells previously stimulated against an antigen.

Example 10: Blocking of PD-L1 ligand binding to PD-1 by human anti-PD-L1 antibodies

Anti-PD-L1 human monoclonal antibodies were tested for the ability to block binding of the ligand PD-L1 to PD-1 expressed on transfected CHO cells by using a cell cytometry assay.

PD-1 expressing CHO cells were suspended in FACS buffer (PBS with 4% fetal calf serum). Various concentrations of the anti-PD-L1 HuMAbs 3G10, 10A5 or 12A4 was added to the cell suspension tubes at 4°C for 30 minutes, followed by addition FITC-labeled PD-L1 fused to an immunoglobulin Fc-region. Flow cytometric analyses were performed using a FACScalibur flow cytometer (Becton Dickinson, San Jose, CA). The

results are depicted in Figure 45. The anti-PD-L1 monoclonal antibodies 3G10, 10A5, and 12A4 blocked binding of PD-L1 to CHO cells transfected with human PD-1, as measured by the mean fluorescent intensity (MFI) of staining. These data demonstrate that the anti-PD-L1 HuMAbs block binding of PD-L1 ligand to cell surface PD-1.

5 **Example 11: Inhibition of the binding of soluble PD-1 to cell-surface PD-L1 by human anti-PD-L1 antibodies.**

Anti-PD-L1 human monoclonal antibodies were tested for the ability to block binding of a soluble dimeric version of the PD-1 receptor (PD-1-hFc) to PD-L1 expressed on hIFN- γ -induced ES-2 human ovarian carcinoma cells using a flow cytometry assay.

10 The blocking was compared to isotype control antibody.

ES-2 cells were induced overnight with 500 IU/mL of hIFN- γ to upregulate hPD-L1 cell surface expression. Induced cells were suspended in FACS buffer. Serial dilutions of the anti-PD-L1 HuMAbs 12A4, 1B12, 3G10, 10A5, 12B7, 13G4, 11E6, and 5F8 were added to the cell suspension tubes at 4°C for 30 minutes, followed by two washes to remove unbound antibody. Next PD-1-hFc protein was added at a constant 2 ug/mL to all wells at 4°C for 30 minutes, followed by two washes to remove unbound PD-1-hFc. Next bound PD-1-Fc was detected on the ES-2 cells by addition of biotinylated-non-blocking anti-PD-1 HuMab 26D5, which binds to PD-1 when bound to PD-L1, at 4°C for 30 minutes, followed by two washes to remove unbound antibody. Finally, bound 26D5 antibody was detected by addition of streptavidin-PE conjugate at 4°C for 30 minutes, followed by two washes to remove unbound conjugate. Flow cytometric analysis was performed using a FACScalibur flow cytometer (Becton Dickinson, San Jose, CA). The results are depicted in Figure 46. The anti-PD-L1 monoclonal antibodies 12A4, 1B12, 3G10, 10A5, 12B7, 13G4, 11E6, and 5F8 blocked binding of PD-1 to ES-2 cells that express human PD-L1, as measured by the geometric mean fluorescent intensity (GMFI) of staining. These data demonstrate that the anti-PD-L1 HuMAbs block binding of soluble PD-1 receptor to cell surface PD-L1.

Example 12: Treatment of *in vivo* tumor model using anti-PD-L1 antibodies

Mice implanted with a cancerous tumor are treated *in vivo* with anti-PD-L1 antibodies to examine the *in vivo* effect of the antibodies on tumor growth. For the tumor studies, female AJ mice between 6-8 weeks of age (Harlan Laboratories) are randomized

by weight into 6 groups. The mice are implanted subcutaneously in the right flank with 2×10^6 SA1/N fibrosarcoma cells dissolved in 200 μ l of DMEM media on day 0. The mice are treated with PBS vehicle, or anti-PD-L1 antibodies at 10 mg/kg. The animals are dosed by intraperitoneal injection with approximately 200 μ l of PBS containing antibody or vehicle on days 1, 4, 8 and 11. Each group contains 10 animals and the groups consist of: (i) a vehicle group, (ii) control mouse IgG, and (iii) an anti-PD-L1 antibody. The mice are monitored twice weekly for tumor growth for approximately 6 weeks. Using an electronic caliper, the tumors are measured three dimensionally (height x width x length) and tumor volume is calculated. Mice are euthanized when the tumors reached tumor end point (1500 mm³) or show greater than 15% weight loss.

Example 13: In Vivo Efficacy of Combination Therapy (anti-CTLA-4 and anti-PD-L1 Antibodies) on Tumor Establishment and Growth

MC38 colorectal cancer cells (available from Dr. N. Restifo, National Cancer Institute, Bethesda, MD; or Jeffrey Schlom, National Institutes of Health, Bethesda, MD) were implanted in C57BL/6 mice (2×10^6 cells/mouse) and selected for treatment when tumors reached a size of 100-200 mm³). On day 0 (*i.e.*, the first day of treatment), each of four groups of 10 mice each was injected intraperitoneally (IP) with one of the following: (1) 10 mg/kg mouse IgG and 10 mg/kg of rat IgG (control), (2) 10 mg/kg anti-CTLA-4 monoclonal antibody 9D9 (mouse anti-mouse CTLA-4, obtained from J. Allison, Memorial Sloan-Kettering Cancer Center, New York, NY) and 10 mg/kg rat IgG, (3) anti-PD-L1 monoclonal antibody MIH5 (rat anti-mouse PD-L1, eBioscience) and 10 mg/kg mouse IgG, or (4) 10 mg/kg anti-CTLA-4 antibody 9D9 and 10 mg/kg anti-PD-L1 antibody MIH5. Antibody injections were then further administered on days 3 and, 6. Using an electronic caliper, the tumors were measured three dimensionally (height x width x length) and tumor volume was calculated. Mice were euthanized when the tumors reached a designated tumor end-point. The results are shown in Figure 47.

This study indicates that, in the MC38 murine tumor model, anti-PD-L1 antibody treatment alone has a modest effect on tumor growth resulting in a delay of tumor growth while anti-CTLA-4 has little effect in this model. However, the combination treatment of CTLA-4 antibody and PD-L1 antibody has a significantly greater effect on tumor growth and results in tumor free mice.

Example 14: Immunohistochemistry Using Anti-PD-L1 Antibodies

To assess the tissue binding profiles of HuMab anti-PD-L1, unmodified 12A4, 13G4, 3G10 and 12B7 were examined in a panel of normal (non-neoplastic) human tissues, including spleen, tonsil, cerebrum, cerebellum, heart, liver, lung, kidney, pancreas, pituitary, skin, and small intestine, as well as lung carcinoma tissues (1 sample/each). ES-2 cells were used as positive control. Hu-IgG₁ and Hu-IgG₄ were used as isotype control antibodies.

Snap frozen and OCT embedded normal and tumor tissues were purchased from Cooperative Human Tissue Network (Philadelphia, PA) or National Disease Research Institute (Philadelphia, PA). Cryostat sections at 5 μ m were fixed with acetone for 10 min at room temperature, and stored at -80 °C until use. A Medarex developed immunohistochemistry protocol was performed using unmodified HuMab anti-PD-L1 by pre-complex of the primary antibodies (12A4, 13G4, 3G10 and 12B7) and secondary antibody (FITC conjugated Fab fragment of goat anti-Hu-IgG, Jackson ImmunoResearch Laboratories, West Grove, PA) before applying onto the sections. Briefly, 1 μ g/ml or 5 μ g/ml of the un-conjugated primary antibodies were mixed with 3 fold excess of secondary antibody respectively and incubated for 30 min at room temperature, and then excess human gamma globulin was added for another 30 min to block the unbound secondary antibody. In parallel, isotype control antibodies Hu-IgG₁ or Hu-IgG₄ were pre-complexed in the same manner. Slides were washed with PBS (Sigma, St. Louis, MO) twice, and then incubated with peroxidase block supplied in Dako EnVision+System (Dako, Carpinteria, CA) for 10 minutes. After two washes with PBS, slides were incubated with Dako protein block to block the non-specific binding sites. Subsequently, the pre-complex of primary antibodies or isotype controls were applied onto sections and incubated for 1 hr. Following three washes with PBS, slides were incubated with mouse anti-FITC antibody (20 μ g/ml, Sigma) for 30 min. After another three washes with PBS, the slides were incubated with the peroxidase-conjugated anti-mouse IgG polymer supplied in the Dako EnVision+System for 30 min. Finally, slides were washed as above and reacted with DAB substrate-chromogen solution supplied in the Dako EnVision+System for 6 min. Slides were then washed with deionized water, counterstained with Mayer's hematoxylin (Dako), dehydrated, cleared and coverslipped with Permount (Fischer Scientific, Fair Lawn, NJ) following routine histological procedure.

Weak to moderate staining was observed in ES-2 cells, as well as in tumor cells of lung carcinoma tissues. In tonsil sections, strong staining was seen in crypt epithelium that is heavily infiltrated by lymphoid cells, but not in the mucous stratified squamous epithelial cells. Moderate staining was seen in some cells in the inter-follicular region, and very weak staining was seen in scattered large cells (dendritic reticulum-like cells) in the germinal center. In lung, weak staining was found in alveoli macrophages. The staining patterns in tonsil and lung tissues were similarly seen in immunohistochemistry sections using commercial anti-PD-L1 mAb (eBiosciences, San Diego, CA). There was overall less intense staining by HuMabs, especially for the staining in the germinal centers. In spleen, diffuse weak immunoreactivity in red pulp was slightly above the background staining. In addition, weak to moderate staining was displayed in Kupffer-like cells in liver and scattered cells in Peyer's patch, as well as in scattered macrophage-like cells and fibroblasts mainly in focal region of the muscularis externa of small intestine.

In cerebellum, cerebrum, heart, kidney, pancreas, pituitary and skin tissues, no meaningful staining was observed when stained with all four anti-PD-L1 HuMabs. No evident difference in staining was noted among these four antibodies except 12B7 and/or 3G10 displayed slightly stronger staining in liver and ES-2 cells.

PD-L1 Antibody Summary

SEQ ID NO:	SEQUENCE	SEQ ID NO:	SEQUENCE
1	VH a.a. 3G10	26	VH CDR1 a.a. 1B12
2	VH a.a. 12A4	27	VH CDR1 a.a. 7H1
3	VH a.a. 10A5	28	VH CDR1 a.a. 11E6
4	VH a.a. 5F8	29	VH CDR1 a.a. 12B7
5	VH a.a. 10H10	30	VH CDR1 a.a. 13G4
6	VH a.a. 1B12		
7	VH a.a. 7H1	31	VH CDR2 a.a. 3G10
8	VH a.a. 11E6	32	VH CDR2 a.a. 12A4
9	VH a.a. 12B7	33	VH CDR2 a.a. 10A5
10	VH a.a. 13G4	34	VH CDR2 a.a. 5F8
		35	VH CDR2 a.a. 10H10
11	VK a.a. 3G10	36	VH CDR2 a.a. 1B12
12	VK a.a. 12A4	37	VH CDR2 a.a. 7H1
13	VK a.a. 10A5	38	VH CDR2 a.a. 11E6
14	VK a.a. 5F8	39	VH CDR2 a.a. 12B7
15	VK a.a. 10H10	40	VH CDR2 a.a. 13G4
16	VK a.a. 1B12		
17	VK a.a. 7H1	41	VH CDR3 a.a. 3G10
18	VK a.a. 11E6	42	VH CDR3 a.a. 12A4
19	VK a.a. 12B7	43	VH CDR3 a.a. 10A5
20	VK a.a. 13G4	44	VH CDR3 a.a. 5F8
		45	VH CDR3 a.a. 10H10
21	VH CDR1 a.a. 3G10	46	VH CDR3 a.a. 1B12
22	VH CDR1 a.a. 12A4	47	VH CDR3 a.a. 7H1
23	VH CDR1 a.a. 10A5	48	VH CDR3 a.a. 11E6
24	VH CDR1 a.a. 5F8	49	VH CDR3 a.a. 12B7
25	VH CDR1 a.a. 10H10	50	VH CDR3 a.a. 13G4

51	VK CDR1 a.a. 3G10	79	VK CDR3 a.a. 12B7
52	VK CDR1 a.a. 12A4	80	VK CDR3 a.a. 13G4
53	VK CDR1 a.a. 10A5		
54	VK CDR1 a.a. 5F8	81	VH n.t. 3G10
55	VK CDR1 a.a. 10H10	82	VH n.t. 12A4
56	VK CDR1 a.a. 1B12	83	VH n.t. 10A5
57	VK CDR1 a.a. 7H1	84	VH n.t. 5F8
58	VK CDR1 a.a. 11E6	85	VH n.t. 10H10
59	VK CDR1 a.a. 12B7	86	VH n.t. 1B12
60	VK CDR1 a.a. 13G4	87	VH n.t. 7H1
		88	VH n.t. 11E6
61	VK CDR2 a.a. 3G10	89	VH n.t. 12B7
62	VK CDR2 a.a. 12A4	90	VH n.t. 13G4
63	VK CDR2 a.a. 10A5		
64	VK CDR2 a.a. 5F8	91	VK n.t. 3G10
65	VK CDR2 a.a. 10H10	92	VK n.t. 12A4
66	VK CDR2 a.a. 1B12	93	VK n.t. 10A5
67	VK CDR2 a.a. 7H1	94	VK n.t. 5F8
68	VK CDR2 a.a. 11E6	95	VK n.t. 10H10
69	VK CDR2 a.a. 12B7	96	VK n.t. 1B12
70	VK CDR2 a.a. 13G4	97	VK n.t. 7H1
		98	VK n.t. 11E6
71	VK CDR3 a.a. 3G10	99	VK n.t. 12B7
72	VK CDR3 a.a. 12A4	100	VK n.t. 13G4
73	VK CDR3 a.a. 10A5		
74	VK CDR3 a.a. 5F8	101	VH 1-18 germline a.a.
75	VK CDR3 a.a. 10H10	102	VH 1-69 germline a.a.
76	VK CDR3 a.a. 1B12	103	VH 1-3 germline a.a.
77	VK CDR3 a.a. 7H1	104	VH 3-9 germline a.a.
78	VK CDR3 a.a. 11E6		

105	VK L6 germline a.a.		
106	VK L15 germline a.a.		
107	VK A27 germline a.a.		
108	VK L18 germline a.a.		
109	VK a.a. 11E6a		

Claims

1. A monoclonal antibody, or an antigen-binding portion thereof, which cross-competes for binding to human Programmed Death Ligand 1 (PD-L1) with a reference antibody or reference antigen-binding portion thereof comprising:
 - (a) a heavy chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:1 and a light chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:11;
 - (b) a heavy chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:2 and a light chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:12;
 - (c) a heavy chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:6 and a light chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:16;
 - (d) a heavy chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:7 and a light chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:17;
 - (e) a heavy chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:8 and a light chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:18;
 - (f) a heavy chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:9 and a light chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:19; or
 - (g) a heavy chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:10 and a light chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:20.
2. The monoclonal antibody or antigen-binding portion thereof of claim 1, wherein the reference antibody or reference antigen-binding portion thereof comprises a heavy chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:2 and a light chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:12.
3. The monoclonal antibody or an antigen-binding portion thereof of claim 1, comprising:
 - (a) a heavy chain variable region derived from a human VH 1-18 gene; and
 - (b) a light chain variable region derived from a human Vk L6 gene;wherein the antibody specifically binds to PD-L1.
4. The monoclonal antibody or an antigen-binding portion thereof of claim 1, comprising:
 - (a) a heavy chain variable region derived from a human VH 1-69 gene; and
 - (b) a light chain variable region derived from a human Vk L6 gene;wherein the antibody specifically binds to PD-L1.

5. The monoclonal antibody or an antigen-binding portion thereof of claim 1, comprising:
 - (a) a heavy chain variable region derived from a human VH 1-3 gene; and
 - (b) a light chain variable region derived from a human Vk L15 gene;wherein the antibody specifically binds to PD-L1.
6. The monoclonal antibody, or an antigen-binding portion thereof of claim 1, comprising:
 - (a) a heavy chain variable region CDR1 comprising amino acids having the sequence set forth in SEQ ID NO:27; (b) a heavy chain variable region CDR2 comprising amino acids having the sequence set forth in SEQ ID NO:37; (c) a heavy chain variable region CDR3 comprising amino acids having the sequence set forth in SEQ ID NO:47; (d) a light chain variable region CDR1 comprising amino acids having the sequence set forth in SEQ ID NO:57; (e) a light chain variable region CDR2 comprising amino acids having the sequence set forth in SEQ ID NO:67; and (f) a light chain variable region CDR3 comprising amino acids having the sequence set forth in SEQ ID NO:77;wherein the antibody, or antigen-binding portion thereof, specifically binds to Programmed Death Ligand 1 (PD-L1).
7. The monoclonal antibody, or an antigen binding portion thereof of claim 1 comprising:
 - (a) a heavy chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:7, and (b) a light chain variable region comprising amino acids having the sequence set forth in SEQ ID NO:17;wherein the antibody or antigen-binding portion thereof specifically binds to Programmed Death Ligand 1 (PD-L1).
8. The antibody or antigen-binding portion thereof of any one of claims 1-7, which exhibits one or more properties selected from:
 - (a) increases T-cell proliferation in a mixed lymphocyte reaction (MLR) assay;
 - (b) increases interferon- γ production in an MLR assay; and
 - (c) increases IL-2 secretion in an MLR assay.
9. An immunoconjugate comprising the antibody, or antigen-binding portion thereof, of any one of claims 1-8, linked to a therapeutic agent.
10. The immunoconjugate of claim 9, wherein the therapeutic agent is a cytotoxin or a radioactive isotope.
11. A bispecific molecule comprising the antibody, or antigen-binding portion thereof, of any one of claims 1-8, linked to a second functional moiety having a different binding specificity than said antibody, or antigen-binding portion thereof.

12. A composition comprising (a) the monoclonal antibody, or antigen-binding portion thereof, of any one of claims 1-8; (b) the immunoconjugate of claim 9 or 10; or (c) the bispecific molecule of claim 11, and a pharmaceutically acceptable carrier.
13. An isolated nucleic acid encoding the antibody, or antigen-binding portion thereof, of any one of claims 1-8.
14. An expression vector comprising the nucleic acid of claim 13.
15. A host cell comprising the expression vector of claim 14.
16. A transgenic mouse comprising human immunoglobulin heavy and light chain transgenes, wherein the mouse expresses the antibody of any one of claims 1-8.
17. A hybridoma prepared from the mouse of claim 16, wherein the hybridoma produces said antibody, or antigen-binding portion thereof.
18. Use of the monoclonal antibody, or antigen-binding portion thereof, of any one of claims 1-8 for the preparation of a medicament for modulating an immune response in a subject.
19. Use of the monoclonal antibody, or antigen-binding portion thereof, of any one of claims 1-8 for the preparation of a medicament for inhibiting growth of tumor cells in a subject.
20. The use of claim 19, wherein the tumor cells are of a cancer selected from melanoma, renal cancer, prostate cancer, breast cancer, colon cancer and lung cancer.
21. The use of claim 19, wherein the tumor cells are of a cancer of the bladder.
22. The use of claim 19 wherein the tumor cells are of a cancer selected from the list consisting of bone cancer, pancreatic cancer, skin cancer, cancer of the head or neck, cutaneous or intraocular malignant melanoma, uterine cancer, ovarian cancer, rectal cancer, cancer of the anal region, stomach cancer, testicular cancer, uterine cancer, carcinoma of the fallopian tubes, carcinoma of the endometrium, carcinoma of the cervix, carcinoma of the vagina, carcinoma of the vulva, Hodgkin's Disease, non-Hodgkin's lymphoma, cancer of the esophagus, cancer of the small intestine, cancer of the endocrine system, cancer of the thyroid gland, cancer of the parathyroid gland, cancer of the adrenal gland, sarcoma of soft tissue, cancer of the urethra, cancer of the penis, chronic or acute leukemias including acute myeloid leukemia, chronic myeloid leukemia, acute lymphoblastic leukemia, chronic lymphocytic leukemia, solid tumors of childhood, lymphocytic lymphoma, cancer of the kidney or ureter, carcinoma of the renal pelvis, neoplasm of the central nervous system (CNS), primary CNS lymphoma, tumor angiogenesis, spinal axis tumor, brain stem glioma, pituitary adenoma, Kaposi's sarcoma, epidermoid cancer,

squamous cell cancer, T-cell lymphoma, environmentally induced cancers including those induced by asbestos, and combinations of said cancers.

23. Use of the monoclonal antibody, or antigen-binding portion thereof, of any one of claims 1-8 for the preparation of a medicament for treating an infectious disease in a subject.

24. The use of claim 23, wherein the infectious disease is

(a) a disease selected from the group consisting of Influenza, Herpes, Giardia, Malaria, and Leishmania;

(b) a pathogenic infection by a virus selected from the group consisting of human immunodeficiency virus (HIV), Hepatitis virus, herpes virus, adenovirus, influenza virus, flaviviruses, echovirus, rhinovirus, coxsackie virus, cornovirus, respiratory syncytial virus, mumps virus, rotavirus, measles virus, rubella virus, parvovirus, vaccinia virus, HTLV virus, dengue virus, papillomavirus, molluscum virus, poliovirus, rabies virus, JC virus and arboviral encephalitis virus;

(c) a pathogenic infection by a bacterium selected from the group consisting of chlamydia, rickettsial bacteria, mycobacteria, staphylococci, streptococci, pneumococci, meningococci and conococci, klebsiella, proteus, serratia, pseudomonas, legionella, diphtheria, salmonella, bacilli, cholera, tetanus, botulism, anthrax, plague, leptospirosis, and Lyme's disease bacteria;

(d) a pathogenic infection by a fungus selected from the group consisting of Candida, Cryptococcus neoformans, Aspergillus, Mucorales, Sporothrix schenckii, Blastomyces dermatitidis, Paracoccidioides brasiliensis, Coccidioides immitis and Histoplasma capsulatum; and

(e) a pathogenic infection by a parasite selected from the group consisting of Entamoeba histolytica, Balantidium coli, Naegleria fowleri, Acanthamoeba sp., Giardia lamblia, Cryptosporidium sp., Pneumocystis carinii, Plasmodium vivax, Babesia microti, Trypanosoma brucei, Trypanosoma cruzi, Leishmania donovani, Toxoplasma gondi, Nippostrongylus brasiliensis.

25. The use of claim 24, wherein:

(a) the Hepatitis virus is selected from Hepatitis A, Hepatitis B, Hepatitis C, and any combination thereof,

(b) the herpes virus is selected from VZV, HSV-1, HAV-6, HSV-II, CMV, Epstein Barr virus, and any combination thereof,

(c) the Candida fungus is selected from Candida albicans, Candida krusei, Candida glabrata, Candida tropicalis, and any combination thereof,

(d) the Aspergillus fungus is selected from the group consisting of Aspergillus fumigatus, Aspergillus niger, and any combination thereof, and/or

(e) the Mucorales fungus is selected from Mucor, Absidia, Rhizopus, and any combination thereof.

26. Use of the antibody, or antigen-binding portion thereof, of any one of claims 1-8 for the preparation of a medicament for enhancing an immune response to the antigen in a subject.
27. The use of claim 26, wherein the antigen is a tumor antigen, a viral antigen, a bacterial antigen or an antigen from a pathogen.
28. Use of the monoclonal antibody, or antigen-binding portion thereof, of any one of claims 1-8 for the preparation of a medicament for treating or preventing an inflammatory disease in a subject.
29. The use of claim 28, wherein the inflammatory disease is lichen planus (LP).