BLOOD COAGULATION PROTEIN CONJUGATES

[0001] This application claims benefit to U.S. Provisional Application Serial No. 61/347,136, filed May 21, 2010, and U.S. Provisional Application Serial No. 61/228,828 filed July 27, 2009, all of which are incorporated herein by reference in their entirety.

FIELD OF THE INVENTION

[0002] The present invention relates to materials and methods for conjugating a water soluble polymer to a blood coagulation protein.

BACKGROUND OF THE INVENTION

[0003] Therapeutic polypeptides such as blood coagulation proteins including Factor IX (FIX), Factor VIII (FVIII), Factor VIIa (FVIIa), Von Willebrand Factor (VWF), Factor FV (FV), Factor X (FX), Factor XI (FXI), Factor XII (FXII), thrombin (FII), protein C, protein S, tPA, PAI-1, tissue factor (TF) and ADAMTS 13 protease are rapidly degraded by proteolytic enzymes and neutralized by antibodies. This reduces their half-life and circulation time, thereby limiting their therapeutic effectiveness. Relatively high doses and frequent administration are necessary to reach and sustain the desired therapeutic or prophylactic effect of these coagulation proteins. As a consequence, adequate dose regulation is difficult to obtain and the need of frequent intravenous administrations imposes restrictions on the patient's way of living.

[0004] PEGylation of polypeptide drugs protects them in circulation and improves their pharmacodynamic and pharmacokinetic profiles (Harris and Chess, Nat Rev Drug Discov. 2003;2:214-21). The PEGylation process attaches repeating units of ethylene glycol (polyethylene glycol (PEG)) to a polypeptide drug. PEG molecules have a large hydrodynamic volume (5–10 times the size of globular proteins), are highly water soluble and hydrated, non-toxic, non-immunogenic and rapidly cleared from the body. PEGylation of molecules can lead to increased resistance of drugs to enzymatic degradation, increased half-life *in vivo*, reduced dosing frequency, decreased immunogenicity, increased physical and thermal stability, increased solubility, increased liquid stability, and reduced aggregation.

The first PEGylated drugs were approved by the FDA in the early 1990s. Since then, the FDA has approved several PEGylated drugs for oral, injectable, and topical administration.

[0005] Polysialic acid (PSA), also referred to as colominic acid (CA), is a naturally occurring polysaccharide. It is a homopolymer of N-acetylneuraminic acid with α(2→8) ketosidic linkage and contains vicinal diol groups at its non-reducing end. It is negatively charged and a natural constituent of the human body. It can easily be produced from bacteria in large quantities and with pre-determined physical characteristics (US Patent No. 5,846,951). Because the bacterially-produced PSA is chemically and immunologically identical to PSA produced in the human body, bacterial PSA is non-immunogenic, even when coupled to proteins. Unlike some polymers, PSA acid is biodegradable. Covalent coupling of colominic acid to catalase and asparaginase has been shown to increase enzyme stability in the presence of proteolytic enzymes or blood plasma. Comparative studies *in vivo* with polysialylated and unmodified asparaginase revealed that polysialylation increased the half-life of the enzyme (Fernandes and Gregoriadis, Int Biochimica Biophysica Acta 1341:26-34, 1997).

[0006] The preparation of conjugates by forming a covalent linkage between the water soluble polymer and the therapeutic protein can be carried out by a variety of chemical methods. For example, coupling of PEG-derivatives to peptides or proteins is reviewed by Roberts et al. (Adv Drug Deliv Rev 2002;54:459-76). One approach for coupling water soluble polymers to therapeutic proteins is the conjugation of the polymers via the carbohydrate moieties of the protein. Vicinal hydroxyl (OH) groups of carbohydrates in proteins can be easily oxidized with sodium periodate (NaIO4) to form active aldehyde groups (Rothfus et Smith, J Biol Chem 1963; 238:1402-10; van Lenten et Ashwell, J Biol Chem 1971;246:1889-94). Subsequently the polymer can be coupled to the aldehyde groups of the carbohydrate by use of reagents containing, for example, an active hydrazide group (Wilchek M and Bayer EA, Methods Enzymol 1987;138:429-42). A more recent technology is the use of reagents containing aminooxy groups which react with aldehydes to form oxime linkages (WO 96/40662, WO2008/025856).

[0007] Additional examples describing conjugation of a water soluble polymer to a therapeutic protein are described in WO 06/071801 which teaches the oxidation of carbohydrate moieties in Von Willebrand factor and subsequent coupling to PEG using hydrazide chemistry; US Publication No. 2009/0076237 which teaches the oxidation of rFVIII and subsequent coupling to PEG and other water soluble polymers (e.g. PSA, HES,

dextran) using hydrazide chemistry; WO 2008/025856 which teaches oxidation of different coagulation factors, e.g. rFIX, FVIII and FVIIa and subsequent coupling to e.g., PEG, using aminooxy chemistry by forming an oxime linkage; and US Patent No. 5,621,039 which teaches the oxidation of FIX and subsequent coupling to PEG using hydrazide chemistry.

[0008] Recently, an improved method was described comprising mild periodate oxidation of sialic acids to generate aldehydes followed by reaction with an aminooxy group containing reagent in the presence of catalytic amounts of aniline (Dirksen A et Dawson PE, Bioconjugate Chem. 2008;19,2543-8; and Zeng Y et al., Nature Methods 2009;6:207-9). The aniline catalysis dramatically accelerates the oxime ligation, allowing the use of very low concentrations of the reagent.

[0009] Notwithstanding the methods available of conjugating water soluble polymers to therapeutic proteins, there remains a need to develop materials and methods for conjugating water soluble polymers to proteins that improves the protein's pharmacodynamic and/or pharmacokinetic properties while minimizing the costs associated with the various reagents.

SUMMARY

[0010] The present disclosure provides materials and methods for conjugating polymers to proteins that improves the protein's pharmacodynamic and/or pharmacokinetic properties while minimizing the costs associated with the various reagents.

[0010a] In one aspect, there is provided a modified blood coagulation protein comprising:

(a) a blood coagulation protein selected from the group consisting of

(i) a blood coagulation protein with Factor IX (FIX) biological activity; and

(ii) a blood coagulation protein with Factor VIII (FVIII) biological activity; and

(b) at least one polysialic acid (PSA) containing an active aminooxy group bound to said blood coagulation protein of (a) at one or more oxidized carbohydrate moieties;

wherein said modified blood coagulation protein has an oxime linkage between said one or more oxidized carbohydrate moieties and the active aminooxy group on the PSA;

wherein said modified blood coagulation protein has a specific activity of at least 50% relative to an unmodified blood coagulation protein.

[0010b] In one aspect, there is provided a method of conjugating a polysialic acid (PSA) containing an active aminooxy group to an oxidized carbohydrate moiety of a blood coagulation protein, comprising contacting said oxidized carbohydrate moiety with at least one activated PSA under conditions that allow conjugation;

said blood coagulation protein selected from the group consisting of
(i) a blood coagulation protein with FIX biological activity; and
(ii) a blood coagulation protein with FVIII biological activity;

said carbohydrate moiety oxidized by incubation with a buffer comprising an oxidizing agent selected from the group consisting of sodium periodate ($NaIO_4$), lead tetraacetate ($Pb(OAc)_4$) and potassium perruthenate (KRuO4); wherein an oxime linkage is formed between the oxidized carbohydrate moiety and the active aminooxy group on the PSA;

said modified blood coagulation protein having a specific activity of at least 50% relative to an unmodified blood coagulation protein.

[0011] In one embodiment of the disclosure, there is provided a method of conjugating a water soluble polymer to an oxidized carbohydrate moiety of a blood coagulation protein comprising contacting the oxidized carbohydrate moiety with an activated water soluble polymer under conditions that allow conjugation; the blood coagulation protein selected from the group consisting of Factor IX (FIX), Factor VIII (FVIII), Factor VIIa (FVIIa) , Von Willebrand Factor (VWF), Factor FV (FV), Factor X (FX), Factor XI (FXI), Factor XII (FXII), thrombin (FII), protein C, protein S, tPA, PAI-1, tissue factor (TF) and ADAMTS 13 protease or a biologically active fragment, derivative or variant thereof; the water soluble polymer containing an active aminooxy group and is selected from the group consisting of polyethylene glycol (PEG), branched PEG, polysialic acid (PSA), carbohydrate, polysaccharides, pullulane, chitosan, hyaluronic acid, chondroitin sulfate, dermatan sulfate, starch, dextran, carboxymethyl-dextran, polyalkylene oxide (PAO), polyalkylene glycol (PAG), polypropylene glycol (PPG), polyoxazoline, polyacryloylmorpholine, polyvinyl alcohol (PVA), polycarboxylate, polyvinylpyrrolidone, polyphosphazene, polyoxazoline,

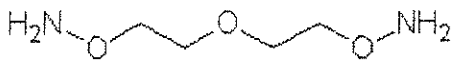
polyethylene-co-maleic acid anhydride, polystyrene-co-maleic acid anhydride, poly(1-hydroxymethylethylene hydroxymethylformal) (PHF), 2-methacryloyloxy-2'-ethyltrimethylammoniumphosphate (MPC); and the carbohydrate moiety oxidized by incubation with a buffer comprising an oxidizing agent selected from the group consisting of sodium periodate (NaIO4), lead tetraacetate (Pb(OAc)4) and potassium perruthenate (KRuO4); wherein an oxime linkage is formed between the oxidized carbohydrate moiety and the active aminooxy group on the water soluble polymer.

[0012] In another embodiment of the disclosure, the water soluble polymer according to the aforementioned method is PSA. In a related embodiment, the PSA is comprised of about 5-500 or 10 – 300 sialic acid units. In still another embodiment, the blood coagulation protein according to the aforementioned method is FIX. In another embodiment, the blood coagulation protein according to the aforementioned method is FVIIa. In still another embodiment, the blood coagulation protein according to the aforementioned method is FVIII. In yet another embodiment, the aforementioned method is provided wherein the oxidizing agent is sodium periodate (NaIO4). In another embodiment, the oxidized carbohydrate moiety of the blood coagulation protein according to the aforementioned method is located in the activation peptide of the blood coagulation protein.

[0013] In yet another embodiment of the disclosure, the aforementioned method is provided wherein the PSA is prepared by reacting an activated aminooxy linker with oxidized PSA;

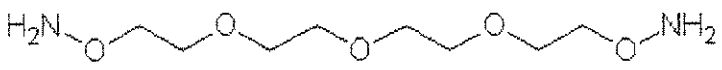
wherein the aminooxy linker is selected from the group consisting of:

a 3-oxa-pentane-1,5-dioxyamine linker of the formula:



and

a 3,6,9-trioxa-undecane-1,11-dioxyamine linker of the formula:



[0014] wherein the PSA is oxidized by incubation with a oxidizing agent to form a terminal aldehyde group at the non-reducing end of the PSA. In still another embodiment, the aforementioned method is provided wherein the activated aminooxy linker comprises 1-50 ethylene glycol units.

In still another embodiment, an aforementioned method is provided wherein the aminooxy linker is 3-oxa-pentane-1,5-dioxyamine. In a related embodiment, the oxidizing agent is $NaIO_4$..

[0015] In another embodiment of the disclosure, the aforementioned method is provided wherein the contacting of the oxidized carbohydrate moiety with the activated water soluble polymer occurs in a buffer comprising a nucleophilic catalyst selected from the group consisting of aniline and aniline derivatives.

[0016] In yet another embodiment of the disclosure, an aforementioned method is provided further comprising the step of reducing an oxime linkage in the conjugated blood coagulation protein by incubating the conjugated blood coagulation protein in a buffer comprising a reducing compound selected from the group consisting of sodium cyanoborohydride (NaCNBH3) and ascorbic acid (vitamin C). In a related embodiment the reducing compound is sodium cyanoborohydride (NaCNBH3).

[0017] In another embodiment of the disclosure, a modified blood coagulation protein produced by an aforementioned method is provided.

[0018] In still another embodiment of the disclosure, a modified FIX is provided comprising a FIX molecule or a biologically active fragment, derivative or variant thereof; and at least one aminooxy PSA bound to the FIX molecule, wherein said aminooxy PSA is attached to the FIX via one or more carbohydrate moieties.

[0019] In another embodiment of the disclosure, a modified FVIIa is provided comprising a FVIIa molecule or a biologically active fragment, derivative or variant thereof; and at least one aminooxy PSA bound to the FVIIa molecule, wherein said aminooxy PSA is attached to the FVIIa via one or more carbohydrate moieties.

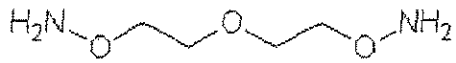
[0020] In still another embodiment of the disclosure, a modified FVIII is provided comprising a FVIII molecule or a biologically active fragment, derivative or variant thereof; and at least one aminooxy PSA bound to the FVIII molecule, wherein said aminooxy PSA is attached to the FVIII via one or more carbohydrate moieties.

[0021] In still another embodiment of the disclosure, a modified FIX is provided comprising a FIX molecule or a biologically active fragment, derivative or variant thereof; and at least one aminooxy PEG bound to the FIX molecule, wherein said aminooxy PEG is attached to the FIX via one or more carbohydrate moieties.

[0022] In another embodiment of the disclosure, a modified FVIIa is provided comprising a FVIIa molecule or a biologically active fragment, derivative or variant thereof; and at least one aminooxy PEG bound to the FVIIa molecule, wherein said aminooxy PEG is attached to the FVIIa via one or more carbohydrate moieties.

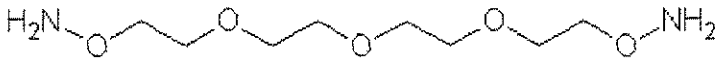
[0023] In still another embodiment of the disclosure, a modified FVIII is provided comprising a FVIII molecule or a biologically active fragment, derivative or variant thereof; and at least one aminooxy PEG bound to the FVIII molecule, wherein said aminooxy PEG is attached to the FVIII via one or more carbohydrate moieties.

[0024] In yet another embodiment, a water soluble polymer is provided comprising an active aminooxy linker; said water soluble polymer selected from the group consisting of polyethylene glycol (PEG), branched PEG, polysialic acid (PSA), carbohydrate, polysaccharides, pullulane, chitosan, hyaluronic acid, chondroitin sulfate, dermatan sulfate, starch, dextran, carboxymethyl-dextran, polyalkylene oxide (PAO), polyalkylene glycol (PAG), polypropylene glycol (PPG), polyoxazoline, poly acryloylmorpholine, polyvinyl alcohol (PVA), polycarboxylate, polyvinylpyrrolidone, polyphosphazene, polyoxazoline, polyethylene-co-maleic acid anhydride, polystyrene-co-maleic acid anhydride, poly(1-hydroxymethylethylene hydroxymethylformal) (PHF), 2-methacryloyloxy-2'-ethyltrimethylammoniumphosphate (MPC); said active aminooxy linker is selected from the group consisting of: a 3-oxa-pentane-1,5-dioxyamine linker of the formula:



and

a 3,6,9-trioxa-undecane-1,11-dioxyamine linker of the formula:



[0025] In still another embodiment, the aforementioned method is provided wherein activated aminooxy linker comprises 1-50 ethylene glycol units.

FIGURES

[0026] Figure 1 shows the primary structure of coagulation Factor IX.

[0027] Figure 2 shows the coupling of oxidized rFIX to aminooxy-PSA.

[0028] Figure 3 shows the synthesis of the water soluble di-aminoxy linkers 3-oxa-pentane-1,5-dioxyamine and 3,6,9-trioxa-undecane-1,11-dioxyamine.

[0029] Figure 4 shows the preparation of aminooxy-PSA.

[0030] Figure 5 shows the analytical characterization of the PSA-rFIX conjugate employing SDS-PAGE and Coomassie staining.

[0031] Figure 6 shows the analytical characterization of the PSA-rFIX conjugate employing detection with anti-FIX and anti-PSA antibodies.

[0032] Figure 7 shows activity of native rFIX and PSA-rFIX conjugate relative to time post infusions.

[0033] Figure 8 shows PSA-rFVIII and Advate levels relative to time post infusion.

DETAILED DESCRIPTION OF THE INVENTION

[0034] The pharmacological and immunological properties of therapeutic proteins can be improved by chemical modification and conjugation with polymeric compounds such as polyethylene glycol (PEG), branched PEG, polysialic acid (PSA), carbohydrate, polysaccharides, pullulane, chitosan, hyaluronic acid, chondroitin sulfate, dermatan sulfate, starch, dextran, carboxymethyl-dextran, polyalkylene oxide (PAO), polyalkylene glycol (PAG), polypropylene glycol (PPG), polyoxazoline, polyacryloylmorpholine, polyvinyl alcohol (PVA), polycarboxylate, polyvinylpyrrolidone, polyphosphazene, polyoxazoline, polyethylene-co-maleic acid anhydride, polystyrene-co-maleic acid anhydride, poly(1-hydroxymethylethylene hydroxymethylformal) (PHF), 2-methacryloyloxy-2'-ethyltrimethylammoniumphosphate (MPC). The properties of the resulting conjugates generally strongly depend on the structure and the size of the polymer. Thus, polymers with a defined and narrow size distribution are usually preferred in the art. Synthetic polymers like PEG can be manufactured easily with a narrow size distribution, while PSA can be purified in such a manner that results in a final PSA preparation with a narrow size distribution. In addition PEGylation reagents with defined polymer chains and narrow size distribution are on the market and commercially available for a reasonable price.

[0035] The addition of a soluble polymer, such as through polysialylation is one approach to improve the properties of a blood coagulation protein such as FIX, as well as other

coagulation proteins (e.g., VWF, FVIIa (see, e.g., US 2008/0221032A1, incorporated herein by reference) and FVIII).

BLOOD COAGULATION PROTEINS

[0036] As described herein, blood coagulation proteins including, but not limited to, Factor IX (FIX), Factor VIII (FVIII), Factor VIIa (FVIIa), Von Willebrand Factor (VWF), Factor FV (FV), Factor X (FX), Factor XI, Factor XII (FXII), thrombin (FII), protein C, protein S, tPA, PAI-1, tissue factor (TF) and ADAMTS 13 protease are contemplated by the invention. As used herein, the term "blood coagulation protein" refers to any Factor IX (FIX), Factor VIII (FVIII), Factor VIIa (FVIIa), Von Willebrand Factor (VWF), Factor FV (FV), Factor X (FX), Factor XII (FXII), thrombin (FII), protein C, protein S, tPA, PAI-1, tissue factor (TF) and ADAMTS 13 protease which exhibits biological activity that is associated with that particular native blood coagulation protein.

[0037] The blood coagulation cascade is divided into three distinct segments: the intrinsic, extrinsic, and common pathways (Schenone et al., Curr Opin Hematol. 2004;11:272-7). The cascade involves a series of serine protease enzymes (zymogens) and protein cofactors. When required, an inactive zymogen precursor is converted into the active form, which consequently converts the next enzyme in the cascade.

[0038] The intrinsic pathway requires the clotting factors VIII, IX, X, XI, and XII. Initiation of the intrinsic pathway occurs when prekallikrein, high-molecular-weight kininogen, factor XI (FXI) and factor XII (FXII) are exposed to a negatively charged surface. Also required are calcium ions and phospholipids secreted from platelets.

[0039] The extrinsic pathway is initiated when the vascular lumen of blood vessels is damaged. The membrane glycoprotein tissue factor is exposed and then binds to circulating factor VII (FVII) and to small preexisting amounts of its activated form FVIIa. This binding facilitates full conversion of FVII to FVIIa and subsequently, in the presence of calcium and phospholipids, the conversion of factor IX (FIX) to factor IXa (FIXa) and factor X (FX) to factor Xa (FXa). The association of FVIIa with tissue factor enhances the proteolytic activity by bringing the binding sites of FVII for the substrate (FIX and FX) into closer proximity and by inducing a conformational change, which enhances the enzymatic activity of FVIIa.

[0040] The activation of FX is the common point of the two pathways. Along with phospholipid and calcium, factors Va (FVa) and Xa convert prothrombin to thrombin (prothrombinase complex), which then cleaves fibrinogen to form fibrin monomers. The

monomers polymerize to form fibrin strands. Factor XIIIa (FXIIIa) covalently bonds these strands to one another to form a rigid mesh.

[0041] Conversion of FVII to FVIIa is also catalyzed by a number of proteases, including thrombin, FIXa, FXa, factor XIa (FXIa), and factor XIIa (FXIIa). For inhibition of the early phase of the cascade, tissue factor pathway inhibitor targets FVIIa/tissue factor/FXa product complex.

A. Polypeptides

[0042] In one aspect, the starting material of the present invention is a blood coagulation protein, which can be derived from human plasma, or produced by recombinant engineering techniques, as described in patents US Patent No. 4,757,006; US Patent No. 5,733,873; US Patent No. 5,198,349; US Patent No. 5,250,421; US Patent No. 5,919,766; and EP 306 968. As described herein, the term blood coagulation protein refers to any blood coagulation protein molecule which exhibits biological activity that is associated with the native blood coagulation protein. In one embodiment of the invention, the blood coagulation protein molecule is a full-length blood coagulation protein.

[0043] Blood coagulation protein molecules contemplated include full-length proteins, precursors of full length proteins, biologically active subunits or fragments of full length proteins, as well as biologically active derivatives and variants of any of these forms of blood coagulation proteins. Thus, blood coagulation protein include those that (1) have an amino acid sequence that has greater than about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98% or about 99% or greater amino acid sequence identity, over a region of at least about 25, about 50, about 100, about 200, about 300, about 400, or more amino acids, to a polypeptide encoded by a referenced nucleic acid or an amino acid sequence described herein; and/or (2) specifically bind to antibodies, e.g., polyclonal or monoclonal antibodies, generated against an immunogen comprising a referenced amino acid sequence as described herein, an immunogenic fragment thereof, and/or a conservatively modified variant thereof.

[0044] According to the present invention, the term "recombinant blood coagulation protein" includes any blood coagulation protein obtained via recombinant DNA technology. In certain embodiments, the term encompasses proteins as described herein.

[0045] As used herein, "endogenous blood coagulation protein" includes a blood coagulation protein which originates from the mammal intended to receive treatment. The term also includes blood coagulation protein transcribed from a transgene or any other foreign DNA present in said mammal. As used herein, "exogenous blood coagulation protein " includes a blood coagulation protein which does not originate from the mammal intended to receive treatment.

[0046] As used herein, "plasma-derived blood coagulation protein " or "plasmatic" includes all forms of the protein found in blood obtained from a mammal having the property participating in the coagulation pathway.

[0047] As used herein "biologically active derivative" or "biologically active variant" includes any derivative or variant of a molecule having substantially the same functional and/or biological properties of said molecule, such as binding properties, and/or the same structural basis, such as a peptidic backbone or a basic polymeric unit.

[0048] An "analog," "variant" or "derivative" is a compound substantially similar in structure and having the same biological activity, albeit in certain instances to a differing degree, to a naturally-occurring molecule. For example, a polypeptide variant refers to a polypeptide sharing substantially similar structure and having the same biological activity as a reference polypeptide. Variants or analogs differ in the composition of their amino acid sequences compared to the naturally-occurring polypeptide from which the analog is derived, based on one or more mutations involving (i) deletion of one or more amino acid residues at one or more termini of the polypeptide and/or one or more internal regions of the naturally-occurring polypeptide sequence (e.g., fragments), (ii) insertion or addition of one or more amino acids at one or more termini (typically an "addition" or "fusion") of the polypeptide and/or one or more internal regions (typically an "insertion") of the naturally-occurring polypeptide sequence or (iii) substitution of one or more amino acids for other amino acids in the naturally-occurring polypeptide sequence. By way of example, a "derivative" refers to a polypeptide sharing the same or substantially similar structure as a reference polypeptide that has been modified, e.g., chemically.

[0049] Variant or analog polypeptides include insertion variants, wherein one or more amino acid residues are added to a blood coagulation protein amino acid sequence of the invention. Insertions may be located at either or both termini of the protein, and/or may be positioned within internal regions of the blood coagulation protein amino acid sequence.

Insertion variants, with additional residues at either or both termini, include for example, fusion proteins and proteins including amino acid tags or other amino acid labels. In one aspect, the blood coagulation protein molecule optionally contains an N-terminal Met, especially when the molecule is expressed recombinantly in a bacterial cell such as E. coli.

[0050] In deletion variants, one or more amino acid residues in a blood coagulation protein polypeptide as described herein are removed. Deletions can be effected at one or both termini of the blood coagulation protein polypeptide, and/or with removal of one or more residues within the blood coagulation protein amino acid sequence. Deletion variants, therefore, include fragments of a blood coagulation protein polypeptide sequence.

[0051] In substitution variants, one or more amino acid residues of a blood coagulation protein polypeptide are removed and replaced with alternative residues. In one aspect, the *substitutions are conservative in nature and conservative substitutions of this type are well* known in the art. Alternatively, the invention embraces substitutions that are also non-conservative. Exemplary conservative substitutions are described in Lehninger, [Biochemistry, 2nd Edition; Worth Publishers, Inc., New York (1975), pp.71-77] and are set out immediately below.

CONSERVATIVE SUBSTITUTIONS

SIDE CHAIN CHARACTERISTIC	AMINO ACID
Non-polar (hydrophobic):	
A. Aliphatic	A L I V P
B. Aromatic	F W
C. Sulfur-containing	M
D. Borderline	G
Uncharged-polar:	
A. Hydroxyl	S T Y
B. Amides	N Q
C. Sulfhydryl	C
D. Borderline	G
Positively charged (basic)	K R H
Negatively charged (acidic)	D E

[0052] Alternatively, exemplary conservative substitutions are set out immediately below.

CONSERVATIVE SUBSTITUTIONS II

ORIGINAL RESIDUE	EXEMPLARY SUBSTITUTION
Ala (A)	Val, Leu, Ile
Arg (R)	Lys, Gln, Asn
Asn (N)	Gln, His, Lys, Arg
Asp (D)	Glu
Cys (C)	Ser
Gln (Q)	Asn
Glu (E)	Asp
His (H)	Asn, Gln, Lys, Arg
Ile (I)	Leu, Val, Met, Ala, Phe,
Leu (L)	Ile, Val, Met, Ala, Phe
Lys (K)	Arg, Gln, Asn
Met (M)	Leu, Phe, Ile
Phe (F)	Leu, Val, Ile, Ala
Pro (P)	Gly
Ser (S)	Thr
Thr (T)	Ser
Trp (W)	Tyr
Tyr (Y)	Trp, Phe, Thr, Ser
Val (V)	Ile, Leu, Met, Phe, Ala

B. Polynucleotides

[0053] Nucleic acids encoding a blood coagulation protein of the invention include, for example and without limitation, genes, pre-mRNAs, mRNAs, cDNAs, polymorphic variants, alleles, synthetic and naturally-occurring mutants.

[0054] Polynucleotides encoding a blood coagulation protein of the invention also include, without limitation, those that (1) specifically hybridize under stringent hybridization conditions to a nucleic acid encoding a referenced amino acid sequence as described herein, and conservatively modified variants thereof; (2) have a nucleic acid sequence that has greater than about 95%, about 96%, about 97%, about 98%, about 99%, or higher nucleotide

sequence identity, over a region of at least about 25, about 50, about 100, about 150, about 200, about 250, about 500, about 1000, or more nucleotides (up to the full length sequence of 1218 nucleotides of the mature protein), to a reference nucleic acid sequence as described herein. Exemplary "stringent hybridization" conditions include hybridization at 42oC in 50% formamide, 5X SSC, 20 mM Na•PO4, pH 6.8; and washing in 1X SSC at 55oC for 30 minutes. It is understood that variation in these exemplary conditions can be made based on the length and GC nucleotide content of the sequences to be hybridized. Formulas standard in the art are appropriate for determining appropriate hybridization conditions. See Sambrook et al., Molecular Cloning: A Laboratory Manual (Second ed., Cold Spring Harbor Laboratory Press, 1989) §§ 9.47-9.51.

[0055] A "naturally-occurring" polynucleotide or polypeptide sequence is typically from a mammal including, but not limited to, primate, e.g., human; rodent, e.g., rat, mouse, hamster; cow, pig, horse, sheep, or any mammal. The nucleic acids and proteins of the invention can be recombinant molecules (e.g., heterologous and encoding the wild type sequence or a variant thereof, or non-naturally occurring).

[0056] In certain embodiments of the invention, the aforementioned polypeptides and polynucleotides are exemplified by the following blood coagulation proteins.

Factor VIIa

[0057] FVII (also known as stable factor or proconvertin) is a vitamin K–dependent serine protease glycoprotein with a pivotal role in hemostasis and coagulation (Eigenbrot, Curr Protein Pept Sci. 2002;3:287-99).

[0058] FVII is synthesized in the liver and secreted as a single-chain glycoprotein of 48 kD. FVII shares with all vitamin K–dependent serine protease glycoproteins a similar protein domain structure consisting of an amino-terminal gamma-carboxyglutamic acid (Gla) domain with 9-12 residues responsible for the interaction of the protein with lipid membranes, a carboxy-terminal serine protease domain (catalytic domain), and two epidermal growth factor–like domains containing a calcium ion binding site that mediates interaction with tissue factor. Gamma-glutamyl carboxylase catalyzes carboxylation of Gla residues in the amino-terminal portion of the molecule. The carboxylase is dependent on a reduced form of vitamin K for its action, which is oxidized to the epoxide form. Vitamin K epoxide reductase is required to convert the epoxide form of vitamin K back to the reduced form.

[0059] The major proportion of FVII circulates in plasma in zymogen form, and activation of this form results in cleavage of the peptide bond between arginine 152 and isoleucine 153. The resulting activated FVIIa consists of a NH_2-derived light chain (20 kD) and a COOH terminal–derived heavy chain (30 kD) linked via a single disulfide bond (Cys 135 to Cys 262). The light chain contains the membrane-binding Gla domain, while the heavy chain contains the catalytic domain.

[0060] The plasma concentration of FVII determined by genetic and environmental factors is about 0.5 mg/mL (Pinotti et al., Blood. 2000;95:3423-8). Different FVII genotypes can result in several-fold differences in mean FVII levels. Plasma FVII levels are elevated during pregnancy in healthy females and also increase with age and are higher in females and in persons with hypertriglyceridemia. FVII has the shortest half-life of all procoagulant factors (3-6 h). The mean plasma concentration of FVIIa is 3.6 ng/mL in healthy individuals and the circulating half-life of FVIIa is relatively long (2.5 h) compared with other coagulation factors.

[0061] Hereditary FVII deficiency is a rare autosomal recessive bleeding disorder with a prevalence estimated to be 1 case per 500,000 persons in the general population (Acharya et al., J Thromb Haemost. 2004;2248-56). Acquired FVII deficiency from inhibitors is also very rare. Cases have also been reported with the deficiency occurring in association with drugs such as cephalosporins, penicillins, and oral anticoagulants. Furthermore, acquired FVII deficiency has been reported to occur spontaneously or with other conditions, such as myeloma, sepsis, aplastic anemia, with interleukin-2 and antithymocyte globulin therapy.

[0062] Reference polynucleotide and polypeptide sequences include, e.g., GenBank Accession Nos. J02933 for the genomic sequence, M13232 for the cDNA (Hagen et al. PNAS 1986; 83: 2412-6), and P08709 for the polypeptide sequence (references incorporated herein in their entireties). A variety of polymorphisms of FVII have been described, for example see Sabater-Lleal et al. (Hum Genet. 2006; 118:741-51) (reference incorporated herein in its entirety).

Factor IX

[0063] FIX is a vitamin K-dependent plasma protein that participates in the intrinsic pathway of blood coagulation by converting FX to its active form in the presence of calcium ions, phospholipids and FVIIIa. The predominant catalytic capability of FIX is as a serine protease with specificity for a particular arginine-isoleucine bond within FX. Activation of

FIX occurs by FXIa which causes excision of the activation peptide from FIX to produce an activated FIX molecule comprising two chains held by one or more disulphide bonds. Defects in FIX are the cause of recessive X-linked hemophilia B.

[0064] Hemophilia A and B are inherited diseases characterized by deficiencies in FVIII and FIX polypeptides, respectively. The underlying cause of the deficiencies is frequently the result of mutations in FVIII and FIX genes, both of which are located on the X chromosome. Traditional therapy for hemophilias often involves intravenous administration of pooled plasma or semi-purified coagulation proteins from normal individuals. These preparations can be contaminated by pathogenic agents or viruses, such as infectious prions, HIV, parvovirus, hepatitis A, and hepatitis C. Hence, there is an urgent need for therapeutic agents that do not require the use of human serum.

[0065] The level of the decrease in FIX activity is directly proportional to the severity of hemophilia B. The current treatment of hemophilia B consists of the replacement of the missing protein by plasma-derived or recombinant FIX (so-called FIX substitution or replacement treatment or therapy).

[0066] Polynucleotide and polypeptide sequences of FIX can be found for example in the UniProtKB/Swiss-Prot Accession No. P00740, US Pat. No. 6,531,298 and in Figure 1.

Factor VIII

[0067] Coagulation factor VIII (FVIII) circulates in plasma at a very low concentration and is bound non-covalently to Von Willebrand factor (VWF). During hemostasis, FVIII is separated from VWF and acts as a cofactor for activated factor IX (FIXa)-mediated FX activation by enhancing the rate of activation in the presence of calcium and phospholipids or cellular membranes.

[0068] FVIII is synthesized as a single-chain precursor of approximately 270-330 kD with the domain structure A1-A2-B-A3-C1-C2. When purified from plasma (e.g., "plasma-derived" or "plasmatic"), FVIII is composed of a heavy chain (A1-A2-B) and a light chain (A3-C1-C2). The molecular mass of the light chain is 80 kD whereas, due to proteolysis within the B domain, the heavy chain is in the range of 90-220 kD.

[0069] FVIII is also synthesized as a recombinant protein for therapeutic use in bleeding disorders. Various in vitro assays have been devised to determine the potential efficacy of recombinant FVIII (rFVIII) as a therapeutic medicine. These assays mimic the in vivo effects of endogenous FVIII. In vitro thrombin treatment of FVIII results in a rapid increase

and subsequent decrease in its procoagulant activity, as measured by in vitro assays. This activation and inactivation coincides with specific limited proteolysis both in the heavy and the light chains, which alter the availability of different binding epitopes in FVIII, e.g. allowing FVIII to dissociate from VWF and bind to a phospholipid surface or altering the binding ability to certain monoclonal antibodies.

[0070] The lack or dysfunction of FVIII is associated with the most frequent bleeding disorder, hemophilia A. The treatment of choice for the management of hemophilia A is replacement therapy with plasma derived or rFVIII concentrates. Patients with severe haemophilia A with FVIII levels below 1%, are generally on prophylactic therapy with the aim of keeping FVIII above 1% between doses. Taking into account the average half-lives of the various FVIII products in the circulation, this result can usually be achieved by giving FVIII two to three times a week.

[0071] Reference polynucleotide and polypeptide sequences include, e.g., UniProtKB/Swiss-Prot P00451 (FA8_HUMAN); Gitschier J et al., Characterization of the human Factor VIII gene, Nature, 312(5992): 326-30 (1984); Vehar GH et al., Structure of human Factor VIII, Nature, 312(5992):337-42 (1984); Thompson AR. Structure and Function of the Factor VIII gene and protein, Semin Thromb Hemost, 2003:29;11-29 (2002).

Von Willebrand Factor

[0072] Von Willebrand factor (VWF) is a glycoprotein circulating in plasma as a series of multimers ranging in size from about 500 to 20,000 kD. Multimeric forms of VWF are composed of 250 kD polypeptide subunits linked together by disulfide bonds. VWF mediates initial platelet adhesion to the sub-endothelium of the damaged vessel wall. Only the larger multimers exhibit hemostatic activity. It is assumed that endothelial cells secrete large polymeric forms of VWF and those forms of VWF which have a low molecular weight (low molecular weight VWF) arise from proteolytic cleavage. The multimers having large molecular masses are stored in the Weibel-Pallade bodies of endothelial cells and liberated upon stimulation.

[0073] VWF is synthesized by endothelial cells and megakaryocytes as prepro-VWF that consists to a large extent of repeated domains. Upon cleavage of the signal peptide, pro-VWF dimerizes through disulfide linkages at its C-terminal region. The dimers serve as protomers for multimerization, which is governed by disulfide linkages between the free end

termini. The assembly to multimers is followed by the proteolytic removal of the propeptide sequence (Leyte et al., Biochem. J. 274 (1991), 257-261).

[0074] The primary translation product predicted from the cloned cDNA of VWF is a 2813-residue precursor polypeptide (prepro-VWF). The prepro-VWF consists of a 22 amino acid signal peptide and a 741 amino acid propeptide, with the mature VWF comprising 2050 amino acids (Ruggeri Z.A., and Ware, J., FASEB J., 308-316 (1993).

[0075] Defects in VWF are causal to Von Willebrand disease (VWD), which is characterized by a more or less pronounced bleeding phenotype. VWD type 3 is the most severe form in which VWF is completely missing, and VWD type 1 relates to a quantitative loss of VWF and its phenotype can be very mild. VWD type 2 relates to qualitative defects of VWF and can be as severe as VWD type 3. VWD type 2 has many sub forms, some being associated with the loss or the decrease of high molecular weight multimers. Von Willebrand disease type 2a (VWD-2A) is characterized by a loss of both intermediate and large multimers. VWD-2B is characterized by a loss of highest-molecular-weight multimers. Other diseases and disorders related to VWF are known in the art.

[0076] The polynucleotide and amino acid sequences of prepro-VWF are available at GenBank Accession Nos. NM_000552 and NP_000543, respectively.

[0077] Other blood coagulation proteins according to the present invention are described in the art, e.g. Mann KG, Thromb Haemost, 1999;82:165-74.

C. Production of blood coagulation proteins

[0078] Production of a blood coagulation protein includes any method known in the art for (i) the production of recombinant DNA by genetic engineering, (ii) introducing recombinant DNA into prokaryotic or eukaryotic cells by, for example and without limitation, transfection, electroporation or microinjection, (iii) cultivating said transformed cells, (iv) expressing blood coagulation protein, e.g. constitutively or upon induction, and (v) isolating said blood coagulation protein, e.g. from the culture medium or by harvesting the transformed cells, in order to obtain purified blood coagulation protein.

[0079] In other aspects, the blood coagulation protein is produced by expression in a suitable prokaryotic or eukaryotic host system characterized by producing a pharmacologically acceptable blood coagulation protein molecule. Examples of eukaryotic cells are mammalian cells, such as CHO, COS, HEK 293, BHK, SK-Hep, and HepG2.

[0080] A wide variety of vectors are used for the preparation of the blood coagulation protein and are selected from eukaryotic and prokaryotic expression vectors. Examples of vectors for prokaryotic expression include plasmids such as, and without limitation, pRSET, pET, and pBAD, wherein the promoters used in prokaryotic expression vectors include one or more of, and without limitation, lac, trc, trp, recA, or araBAD. Examples of vectors for eukaryotic expression include: (i) for expression in yeast, vectors such as, and without limitation, pAO, pPIC, pYES, or pMET, using promoters such as, and without limitation, AOX1, GAP, GAL1, or AUG1; (ii) for expression in insect cells, vectors such as and without limitation, pMT, pAc5, pIB, pMIB, or pBAC, using promoters such as and without limitation PH, p10, MT, Ac5, OpIE2, gp64, or polh, and (iii) for expression in mammalian cells, vectors such as and without limitation pSVL, pCMV, pRc/RSV, pcDNA3, or pBPV, and vectors derived from, in one aspect, viral systems such as and without limitation vaccinia virus, adeno-associated viruses, herpes viruses, or retroviruses, using promoters such as and without limitation CMV, SV40, EF-1, UbC, RSV, ADV, BPV, and β-actin.

D. Administration

[0081] In one embodiment a conjugated blood coagulation protein of the present invention may be administered by injection, such as intravenous, intramuscular, or intraperitoneal injection.

[0082] To administer compositions comprising a conjugated blood coagulation protein of the present invention to human or test animals, in one aspect, the compositions comprise one or more pharmaceutically acceptable carriers. The terms "pharmaceutically" or "pharmacologically acceptable" refer to molecular entities and compositions that are stable, inhibit protein degradation such as aggregation and cleavage products, and in addition do not produce allergic, or other adverse reactions when administered using routes well-known in the art, as described below. "Pharmaceutically acceptable carriers" include any and all clinically useful solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents and the like, including those agents disclosed above.

[0083] As used herein, "effective amount" includes a dose suitable for treating a mammal having a bleeding disorder as described herein.

[0084] The compositions may be administered orally, topically, transdermally, parenterally, by inhalation spray, vaginally, rectally, or by intracranial injection. The term parenteral as used herein includes subcutaneous injections, intravenous, intramuscular,

intracisternal injection, or infusion techniques. Administration by intravenous, intradermal, intramuscular, intramammary, intraperitoneal, intrathecal, retrobulbar, intrapulmonary injection and or surgical implantation at a particular site is contemplated as well. Generally, compositions are essentially free of pyrogens, as well as other impurities that could be harmful to the recipient.

[0085] Single or multiple administrations of the compositions can be carried out with the dose levels and pattern being selected by the treating physician. For the prevention or treatment of disease, the appropriate dosage will depend on the type of disease to be treated, as described above, the severity and course of the disease, whether drug is administered for preventive or therapeutic purposes, previous therapy, the patient's clinical history and response to the drug, and the discretion of the attending physician.

[0086] The present invention also relates to a pharmaceutical composition comprising an effective amount of a conjugated blood coagulation protein as defined herein. The pharmaceutical composition may further comprise a pharmaceutically acceptable carrier, diluent, salt, buffer, or excipient. The pharmaceutical composition can be used for treating the above-defined bleeding disorders. The pharmaceutical composition of the invention may be a solution or a lyophilized product. Solutions of the pharmaceutical composition may be subjected to any suitable lyophilization process.

[0087] As an additional aspect, the invention includes kits which comprise a composition of the invention packaged in a manner which facilitates its use for administration to subjects. In one embodiment, such a kit includes a compound or composition described herein (e.g., a composition comprising a conjugated blood coagulation protein), packaged in a container such as a sealed bottle or vessel, with a label affixed to the container or included in the package that describes use of the compound or composition in practicing the method. In one embodiment, the kit contains a first container having a composition comprising a conjugated blood coagulation protein and a second container having a physiologically acceptable reconstitution solution for the composition in the first container. In one aspect, the compound or composition is packaged in a unit dosage form. The kit may further include a device suitable for administering the composition according to a specific route of administration. Preferably, the kit contains a label that describes use of the therapeutic protein or peptide composition.

WATER SOLUBLE POLYMERS

[0088] In one aspect, a blood coagulation protein derivative (i.e., a conjugated blood coagulation protein) molecule provided is bound to a water-soluble polymer including, but not limited to, polyethylene glycol (PEG), branched PEG, polysialic acid (PSA), carbohydrate, polysaccharides, pullulane, chitosan, hyaluronic acid, chondroitin sulfate, dermatan sulfate, starch, dextran, carboxymethyl-dextran, polyalkylene oxide (PAO), polyalkylene glycol (PAG), polypropylene glycol (PPG) polyoxazoline, poly acryloylmorpholine, polyvinyl alcohol (PVA), polycarboxylate, polyvinylpyrrolidone, polyphosphazene, polyoxazoline, polyethylene-co-maleic acid anhydride, polystyrene-co-maleic acid anhydride, poly(1-hydroxymethylethylene hydroxymethylformal) (PHF), 2-methacryloyloxy-2'-ethyltrimethylammoniumphosphate (MPC). In one embodiment of the invention, the water soluble polymer is consisting of sialic acid molecule having a molecular weight range of 350 to 120,000, 500 to 100,000,1000 to 80,000, 1500 to 60,000, 2,000 to 45,000 Da, 3,000 to 35,000 Da, and 5,000 to 25,000 Da. The coupling of the water soluble polymer can be carried out by direct coupling to the protein or via linker molecules. One example of a chemical linker is MBPH (4-[4-N-Maleimidophenyl]butyric acid hydrazide) containing a carbohydrate-selective hydrazide and a sulfhydryl-reactive maleimide group (Chamow et al., J Biol Chem 1992;267:15916-22). Other exemplary and preferred linkers are described below.

In one embodiment, the derivative retains the full functional activity of native therapeutic blood coagulation protein products, and provides an extended half-life in vivo, as compared to native therapeutic blood coagulation protein products. In another embodiment, the derivative retains at least 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44. 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56,57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 110, 120, 130, 140, or 150 percent (%) biological activity relative to native blood coagulation protein. In a related aspect, the biological activities of the derivative and native blood coagulation protein are determined by the ratios of chromogenic activity to blood coagulation factor antigen value (blood coagulation factor:Chr: blood coagulation factor:Ag). In still another embodiment of the invention, the half-life of the construct is decreased or increased 0.5, 0.6, 0.7, 0.8, 0.9, 1.0, 1.1, 1.2, 1.3, 1.4, 1.5, 2, 3, 4, 5, 6, 7, 8, 9, or 10-fold relative to the in vivo half-life of native blood coagulation protein.

A. Sialic acid and PSA

[0089] As used herein, "sialic acid moieties" includes sialic acid monomers or polymers ("polysaccharides") which are soluble in an aqueous solution or suspension and have little or no negative impact, such as side effects, to mammals upon administration of the PSA-blood coagulation protein conjugate in a pharmaceutically effective amount. The polymers are characterized, in one aspect, as having 1, 2, 3, 4, 5, 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, 300, 400, or 500 sialic acid units. In certain aspects, different sialic acid units are combined in a chain.

[0090] In one embodiment of the invention, the sialic acid portion of the polysaccharide compound is highly hydrophilic, and in another embodiment the entire compound is highly hydrophilic. Hydrophilicity is conferred primarily by the pendant carboxyl groups of the sialic acid units, as well as the hydroxyl groups. The saccharide unit may contain other functional groups, such as, amine, hydroxyl or sulphate groups, or combinations thereof. These groups may be present on naturally-occurring saccharide compounds, or introduced into derivative polysaccharide compounds.

[0091] The naturally occurring polymer PSA is available as a polydisperse preparation showing a broad size distribution (e.g. Sigma C-5762) and high polydispersity (PD). Because the polysaccharides are usually produced in bacteria carrying the inherent risk of copurifying endotoxins, the purification of long sialic acid polymer chains may raise the probability of increased endotoxin content. Short PSA molecules with 1-4 sialic acid units can also be synthetically prepared (Kang SH et al., Chem Commun. 2000;227-8; Ress DK and Linhardt RJ, Current Organic Synthesis. 2004;1:31-46), thus minimizing the risk of high endotoxin levels. However PSA preparations with a narrow size distribution and low polydispersity, which are also endotoxin-free, can now be manufactured. Polysaccharide compounds of particular use for the invention are, in one aspect, those produced by bacteria. Some of these naturally-occurring polysaccharides are known as glycolipids. In one embodiment, the polysaccharide compounds are substantially free of terminal galactose units.

B. Polyethylene glycol (PEG) and Pegylation

[0092] In certain aspects, blood coagulation factor, e.g., FVIII, FVIIa, FIX, or other blood coagulation factor molecules are conjugated to a water soluble polymer by any of a variety of chemical methods (Roberts JM et al., Advan Drug Delivery Rev 2002;54:459-76). For example, in one embodiment FVIII, FVIIa, or FIX is modified by the conjugation of PEG to

free amino groups of the protein using N-hydroxysuccinimide (NHS) esters. In another embodiment the water soluble polymer, for example PEG, is coupled to free SH groups using maleimide chemistry or the coupling of PEG hydrazides or PEG amines to carbohydrate moieties of the FVIII, FVIIa, or FIX after prior oxidation.

[0093] The conjugation is in one aspect performed by direct coupling (or coupling via linker systems) of the water soluble polymer to blood coagulation factor, e.g., FVIII, FVIIa, or FIX, under formation of stable bonds. In addition degradable, releasable or hydrolysable linker systems are used in certain aspects the present invention (Tsubery et al. J Biol Chem 2004;279:38118-24 / Greenwald et al., J Med Chem 1999;42:3657-67 / Zhao et al., Bioconj Chem 2006;17:341-51 / WO2006/138572A2 / US7259224B2 / US7060259B2).

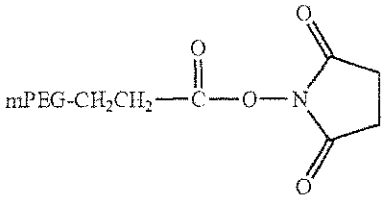
[0094] In one embodiment of the invention, a blood coagulation factor, e.g., FVIII, FVIIa, or FIX, is modified via lysine residues by use of polyethylene glycol derivatives containing an active N-hydroxysuccinimide ester (NHS) such as succinimidyl succinate, succinimidyl glutarate or succinimidyl propionate. These derivatives react with the lysine residues of FVIII, FVIIa, or FIX under mild conditions by forming a stable amide bond. In one embodiment of the invention, the chain length of the PEG derivative is 5,000 Da. Other PEG derivatives with chain lengths of 500 to 2,000 Da, 2,000 to 5,000 Da, greater than 5,000 up to 10,000 Da or greater than 10,000 up to 20,000 Da, or greater than 20,000 up to 150,000 Da are used in various embodiments, including linear and branched structures.

[0095] Alternative methods for the PEGylation of amino groups are, without limitation, the chemical conjugation with PEG carbonates by forming urethane bonds, or the reaction with aldehydes or ketones by reductive amination forming secondary amide bonds.

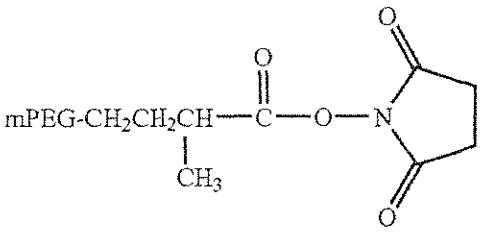
[0096] In one embodiment of the present invention a blood coagulation factor, e.g., FVIII, FVIIa, FIX, or other blood coagulation factor, molecule is chemically modified using PEG derivatives that are commercially available. These PEG derivatives in alternative aspects have a linear or branched structures. Examples of PEG-derivatives containing NHS groups are listed below.

[0097] The following PEG derivatives are non-limiting examples of those commercially available from Nektar Therapeutics (Huntsville, Ala.; see www.nektar.com/PEG reagent catalog; Nektar Advanced PEGylation, price list 2005-2006):

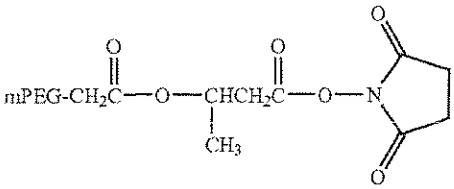
mPEG-Succinimidyl propionate (mPEG-SPA)



mPEG-Succinimidyl α-methylbutanoate (mPEG-SMB)

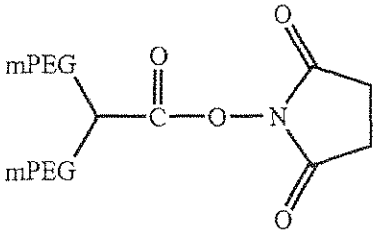


mPEG-CM-HBA-NHS (CM=carboxymethyl; HBA=Hydroxy butyric acid)



Structure of a Branched PEG-derivative (Nektar Therapeutics):

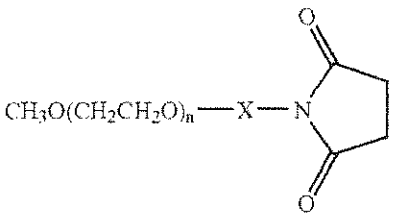
Branched PEG N-Hydroxysuccinimide (mPEG2-NHS)



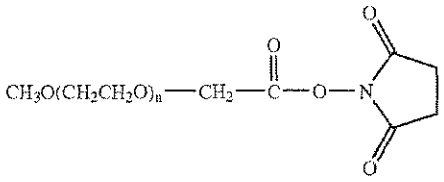
[0098] This reagent with branched structure is described in more detail by Kozlowski et al. (BioDrugs 2001;5:419-29).

[0099] Other non-limiting examples of PEG derivatives are commercially available from NOF Corporation (Tokyo, Japan; see www.nof.co.jp/english: Catalogue 2005)

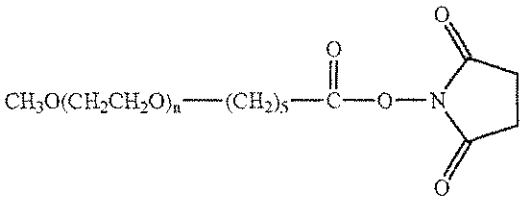
General Structure of Linear PEG-derivatives (NOF Corp.):



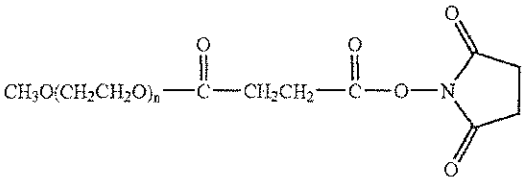
X=carboxymethyl



X=carboxypentyl

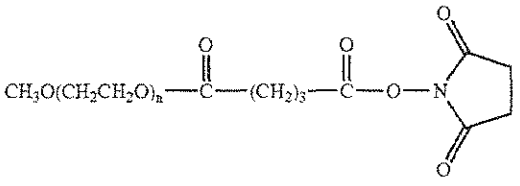


x=succinate



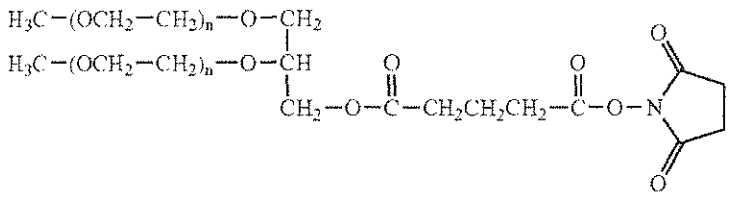
mPEG Succinimidyl succinate

x=glutarate

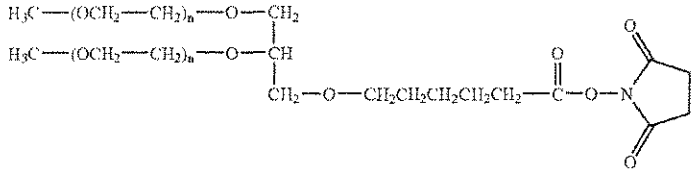


mPEG Succinimidyl glutarate

Structures of Branched PEG-derivatives (NOF Corp.): 2,3-Bis(methylpolyoxyethylene-oxy)-1-(1,5-dioxo-5-succinimidyloxy, pentyloxy)propane



2,3-Bis(methylpolyoxyethylene-oxy)-1-(succinimidyl carboxypentyloxy)propane



[00100] These propane derivatives show a glycerol backbone with a 1,2 substitution pattern. In the present invention branched PEG derivatives based on glycerol structures with 1,3 substitution or other branched structures described in US2003/0143596A1 are also contemplated.

[00101] PEG derivatives with degradable (for example, hydrolysable linkers) as described by Tsubery et al. (J Biol Chem 2004;279:38118-24) and Shechter et al. (WO04089280A3) are also contemplated.

[00102] Surprisingly, the PEGylated FVIII, FVIIa, FIX, or other blood coagulation factor of this invention exhibits functional activity, combined with an extended half-life *in vivo*. In addition the PEGylated rFVIII, FVIIa, FIX, or other blood coagulation factor seems to be more resistant against thrombin inactivation.

C. Methods of attachment

[00103] A blood coagulation protein may be covalently linked to the polysaccharide compounds by any of various techniques known to those of skill in the art. In various aspects of the invention, sialic acid moieties are bound to a blood coagulation protein, e.g., FIX, FVIII, FVIIa or VWF, for example by the method described in US Patent No. 4,356,170, which is herein incorporated by reference.

[00104] Other techniques for coupling PSA to polypeptides are also known and contemplated by the invention. For example, US Publication No. 2007/0282096 describes conjugating an amine or hydrazide derivative of, e.g., PSA, to proteins. In addition, US

Publication No. 2007/0191597 describes PSA derivatives containing an aldehyde group for reaction with substrates (e.g., proteins) at the reducing end. These references are incorporated by reference in their entireties.

[00105] Various methods are disclosed at column 7, line 15, through column 8, line 5 of U.S. Patent No. 5,846,951 (incorporated by reference in its entirety). Exemplary techniques include linkage through a peptide bond between a carboxyl group on one of either the blood coagulation protein or polysaccharide and an amine group of the blood coagulation protein or polysaccharide, or an ester linkage between a carboxyl group of the blood coagulation protein or polysaccharide and a hydroxyl group of the blood coagulation protein or polysaccharide. Another linkage by which the blood coagulation protein is covalently bonded to the polysaccharide compound is via a Schiff base, between a free amino group on the blood coagulation protein being reacted with an aldehyde group formed at the non-reducing end of the polysaccharide by periodate oxidation (Jennings HJ and Lugowski C, J Immunol. 1981;127:1011-8; Fernandes AI and Gregoriadis G, Biochim Biophys Acta. 1997;1341;26-34). The generated Schiff base is in one aspect stabilized by specific reduction with NaCNBH3 to form a secondary amine. An alternative approach is the generation of terminal free amino groups in the PSA by reductive amination with NH4Cl after prior oxidation. Bifunctional reagents can be used for linking two amino or two hydroxyl groups. For example, PSA containing an amino group is coupled to amino groups of the protein with reagents like BS3 (Bis(sulfosuccinimidyl)suberate / Pierce, Rockford, IL). In addition heterobifunctional cross linking reagents like Sulfo-EMCS (N-ε-Maleimidocaproyloxy) sulfosuccinimide ester / Pierce) is used for instance to link amine and thiol groups.

[00106] In another approach, a PSA hydrazide is prepared and coupled to the carbohydrate moiety of the protein after prior oxidation and generation of aldehyde functions.

[00107] As described above, a free amine group of the therapeutic protein reacts with the 1-carboxyl group of the sialic acid residue to form a peptidyl bond or an ester linkage is formed between the 1-carboxylic acid group and a hydroxyl or other suitable active group on a blood coagulation protein. Alternatively, a carboxyl group forms a peptide linkage with deacetylated 5-amino group, or an aldehyde group of a molecule of a blood coagulation protein forms a Schiff base with the N-deacetylated 5-amino group of a sialic acid residue.

[00108] Alternatively, the polysaccharide compound is associated in a non-covalent manner with a blood coagulation protein. For example, the polysaccharide compound and

the pharmaceutically active compound are in one aspect linked via hydrophobic interactions. Other non-covalent associations include electrostatic interactions, with oppositely charged ions attracting each other.

[00109] In various embodiments, the blood coagulation protein is linked to or associated with the polysaccharide compound in stoichiometric amounts (e.g., 1:1, 1:2, 1:3, 1:4, 1:5, 1:6, 1:7, 1:7, 1:8, 1:9, or 1:10, etc.). In various embodiments, 1-6, 7-12 or 13-20 polysaccharides are linked to the blood coagulation protein. In still other embodiments, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or more polysaccharides are linked to the blood coagulation protein.

[00110] In various embodiments, the blood coagulation protein is modified to introduce glycosylation sites (i.e., sites other than the native glycosylation sites). Such modification may be accomplished using standard molecular biological techniques known in the art. Moreover, the blood coagulation protein, prior to conjugation to a water soluble polymer via one or more carbohydrate moieties, may be glycosylated in vivo or in vitro. These glycosylated sites can serve as targets for conjugation of the proteins with water soluble polymers (US Patent Application No. 20090028822, US Patent Application No. 2009/0093399, US Patent Application No. 2009/0081188, US Patent Application No. 2007/0254836, US Patent Application No. 2006/0111279, and DeFrees S. et al., Glycobiology, 2006, 16, 9, 833-43).

D. Aminooxy linkage

[00111] In one embodiment of the invention, the reaction of hydroxylamine or hydroxylamine derivatives with aldehydes (e.g., on a carbohydrate moiety following oxidation by sodium periodate) to form an oxime group is applied to the preparation of conjugates of blood coagulation protein. For example, a glycoprotein (e.g., a blood coagulation protein according to the present invention) is first oxidized with a oxidizing agent such as sodium periodate ($NaIO_4$) (Rothfus JA et Smith EL., J Biol Chem 1963, 238, 1402-10; and Van Lenten L and Ashwell G., J Biol Chem 1971, 246, 1889-94). The periodate oxidation of glycoproteins is based on the classical Malaprade reaction described in 1928, the oxidation of vicinal diols with periodate to form an active aldehyde group (Malaprade L., Analytical application, Bull Soc Chim France, 1928, 43, 683-96). Additional examples for such an oxidizing agent are lead tetraacetate ($Pb(OAc)_4$), manganese acetate ($MnO(Ac)_3$), cobalt acetate ($Co(OAc)_2$), thallium acetate (TlOAc), cerium sulfate ($Ce(SO_4)_2$) (US

4,367,309) or potassium perruthenate ($KRuO_4$) (Marko et al., J Am Chem Soc 1997,119, 12661-2). By "oxidizing agent" a mild oxidizing compound which is capable of oxidizing vicinal diols in carbohydrates, thereby generating active aldehyde groups under physiological reaction conditions is meant.

[00112] The second step is the coupling of the polymer containing an aminooxy group to the oxidized carbohydrate moiety to form an oxime linkage. In one embodiment of the invention, this step can be carried out in the presence of catalytic amounts of the nucleophilic catalyst aniline or aniline derivatives (Dirksen A et Dawson PE, Bioconjugate Chem. 2008; Zeng Y et al., Nature Methods 2009;6:207-9). The aniline catalysis dramatically accelerates the oxime ligation allowing the use of very low concentrations of the reagents. In another embodiment of the invention the oxime linkage is stabilized by reduction with NaCNBH3 to form an alkoxyamine linkage (Figure 2).

[00113] In one embodiment of the invention, the reaction steps to conjugate a water soluble polymer to a blood coagulation protein are carried out separately and sequentially (i.e., starting materials (e.g., blood coagulation protein, water soluble polymer, etc), reagents (e.g., oxidizing agents, aniline, etc) and reaction products (e.g., oxidized carbohydrate on a blood coagulation protein, activated aminooxy water soluble polymer, etc) are separated between individual reaction steps).

[00114] Additional information on aminooxy technology can be found in the following references, each of which is incorporated in their entireties: EP 1681303A1 (HASylated erythropoietin); WO 2005/014024 (conjugates of a polymer and a protein linked by an oxime linking group); WO96/40662 (aminooxy-containing linker compounds and their application in conjugates); WO 2008/025856 (Modified proteins); Peri F et al., Tetrahedron 1998, 54, 12269-78; Kubler-Kielb J et. Pozsgay V., J Org Chem 2005, 70, 6887-90; Lees A et al., Vaccine 2006, 24(6), 716-29; and Heredia KL et al., Macromoecules 2007, 40(14), 4772-9.

[00115] In various embodiments of the invention, the water soluble polymer which is linked according to the aminooxy technology described herein to an oxidized carbohydrate moiety of a blood coagulation protein (e.g., FVIII, FVIIa, or FIX) include, but are not limited to polyethylene glycol (PEG), branched PEG, polysialic acid (PSA), carbohydrate, polysaccharides, pullulane, chitosan, hyaluronic acid, chondroitin sulfate, dermatan sulfate, starch, dextran, carboxymethyl-dextran, polyalkylene oxide (PAO), polyalkylene glycol (PAG), polypropylene glycol (PPG) polyoxazoline, poly acryloylmorpholine, polyvinyl

alcohol (PVA), polycarboxylate, polyvinylpyrrolidone, polyphosphazene, polyoxazoline, polyethylene-co-maleic acid anhydride, polystyrene-co-maleic acid anhydride, poly(1-hydroxymethylethylene hydroxymethylformal) (PHF), 2-methacryloyloxy-2'-ethyltrimethylammoniumphosphate (MPC).

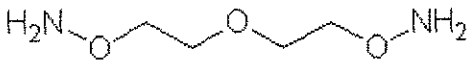
[00116] The following examples are not intended to be limiting but only exemplary of specific embodiments of the invention.

EXAMPLES

Example 1

Preparation of the homobifunctional linker $NH_2[OCH_2CH_2]_2ONH_2$

[00117] The homobifunctional linker $NH_2[OCH_2CH_2]_2ONH_2$

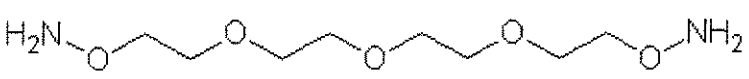


(3-oxa-pentane-1,5-dioxyamine) containing two active aminooxy groups was synthesized according to Boturyn et al. (Tetrahedron 1997;53:5485-92) in a two step organic reaction employing a modified Gabriel-Synthesis of primary amines (Figure 3). In the first step, one molecule of 2,2-chlorodiethylether was reacted with two molecules of Endo-N-hydroxy-5-norbornene-2,3-dicarboximide in dimethylformamide (DMF). The desired homobifunctional product was prepared from the resulting intermediate by hydrazinolysis in ethanol.

Example 2

Preparation of the homobifunctional linker $NH_2[OCH_2CH_2]_4ONH_2$

[00118] The homobifunctional linker $NH_2[OCH_2CH_2]_4ONH_2$



(3,6,9-trioxa-undecane-1,11-dioxyamine) containing two active aminooxy groups was synthesized according to Boturyn et al. (Tetrahedron 1997;53:5485-92) in a two step organic reaction employing a modified Gabriel-Synthesis of primary amines (Figure 3). In the first step one molecule of Bis-(2-(2-chlorethoxy)-ethyl)-ether was reacted with two molecules of

Endo-N-hydroxy-5-norbornene-2,3-dicarboximide in DMF. The desired homobifunctional product was prepared from the resulting intermediate by hydrazinolysis in ethanol.

Example 3

Preparation of aminooxy-PSA

[00119] 500mg of oxidized PSA (MW=18.8kD) obtained from the Serum Institute of India (Pune, India) was dissolved in 8ml 50mM sodium acetate buffer, pH 5.5. Next, 100mg 3-oxa-pentane-1,5-dioxyamine was added. After shaking for 2hrs at room temperature, 44 mg sodium cyanoborohydride was added. After shaking for another 4hrs at 4°C, the reaction mix was loaded into a Slide-A-Lyzer (Pierce, Rockford, IL) dialysis cassette (3.5 kD membrane, regenerated cellulose) and dialyzed against PBS pH 7.2 for 4 days. The product was frozen at -80°C. The preparation of the aminooxy-PSA according to this procedure is illustrated in Figure 4.

Alternative procedure for preparation of aminooxy PSA

[00120] 1000 mg of oxidized PSA (MW = 20 kD) obtained from the Serum Institute of India (Pune, India) was dissolved in 16 ml 50mM phospate buffer pH 6.0. Then 170 mg 3-oxa-pentane-1,5-dioxyamine was given to the reaction mixture. After shaking for 2 hrs at RT 78.5 mg sodium cyanoborohydride was added and the reaction was performed for 18 hours over night. The reaction mixture was then subjected to a ultrafiltration/diafiltration procedure (UF/DF) using a membrane with a 5 kD cut-off made of regenerated cellulose (Millipore).

Example 4

Coupling of aminooxy-PSA to rFIX and purification of the conjugate

[00121] To 12.6mg rFIX, dissolved in 6.3ml 50mM sodium acetate buffer, pH 6.0, 289µl of an aqueous sodium periodate solution (10mM) was added. The mixture was shaken in the dark for 1h at 4°C and quenched for 15min at room temperature by the addition of 6.5µl 1M glycerol. Low molecular weight contaminates were removed by ultrafiltration/diafiltration (UF/DF) employing Vivaspin (Sartorius, Goettingen, Germany) concentrators (30kD membrane, regenerated cellulose) . Next, 43mg aminooxy-PSA was added to the UF/DF retentate and the mixture was shaken for 18hrs at 4°C. The excess PSA reagent was removed

by hydrophobic interaction chromatography (HIC). The conductivity of the cooled reaction mixture was raised to 180mS/cm and loaded onto a 5ml HiTrap Butyl FF (GE Healthcare, Fairfield, CT) HIC column (1.6 x 2.5cm), pre-equilibrated with 50mM HEPES, 3M sodium chloride, 6.7mM calcium chloride, 0.01% Tween 80, pH 6.9. The conjugate was eluted within 2.4 column volumes (CV) with 50mM HEPES, 6.7mM calcium chloride, 0.005% Tween 80, pH 7.4 at a flow rate of 5ml/min. The preparation was analytically characterized by measuring total protein (BCA) and FIX chromogenic activity. For the PSA-rFIX conjugate a specific activity of 80.2 IU/mg protein was determined (56.4 % in comparision to native rFIX). The results are summarized in Table 1.

Table 1

Item	BCA [mg/ml]	FIX:Chrom [IU/ml]	Specific Activity [IU FIX:Chrom/mg BCA	Specific Activity [%]
rFIX	8.58	1221	142.3	100
PSA-rFIX	1.15	92.2	80.2	56.4

[00122] The analytical characterization of the PSA-rFIX conjugate by SDS-PAGE with Coomassie staining is illustrated in Figure 5. An SDS-PAGE followed by Western blot employing anti-FIX and anti-PSA antibodies is shown in Figure 6.

Example 5

Coupling of aminooxy-PSA to rFIX in the presence of aniline as nucleophilic catalyst

[00123] To 3.0mg rFIX, dissolved in 1.4ml 50mM sodium acetate buffer, pH 6.0, 14.1µl of an aqueous sodium periodate solution (10mM) was added. The mixture was shaken in the dark for 1h at 4°C and quenched for 15min at room temperature by the addition of 1.5µl 1M glycerol. Low molecular weight contaminates were removed by means of size exclusion chromatography (SEC) employing PD-10 desalting columns (GE Healthcare, Fairfield, CT). 1.2mg oxidized rFIX, dissolved in 1.33ml 50mM sodium acetate buffer, pH 6.0 was mixed with 70µl of aniline (200mM aqueous stock solution) and shaken for 45min at room temperature. Next, 4.0mg aminooxy-PSA was added and the mixture was shaken for 2hrs at room temperature and another 16hrs at 4°C. Samples were drawn after 1h, after 2hrs and at the end of the reaction after 18hrs. Next, excess PSA reagent and free rFIX were removed by

means of HIC. The conductivity of the cooled reaction mixture was raised to 180mS/cm and loaded onto a 5ml HiTrap Butyl FF (GE Healthcare, Fairfield, CT) HIC column (1.6 x 2.5cm), pre-equilibrated with 50mM HEPES, 3M sodium chloride, 6.7mM calcium chloride, 0.01% Tween 80, pH 6.9. The conjugate was eluted with a linear gradient to 50mM HEPES, 6.7mM calcium chloride, 0.005% Tween 80, pH 7.4 in 20CV with at a flow rate of 5ml/min.

Example 6

Coupling of aminooxy-PSA to rFIX and reduction with $NaCNBH_3$

[00124] To 10.5mg rFIX, dissolved in 5.25ml 50mM sodium acetate buffer, pH 6.0, 53µl of an aqueous sodium periodate solution (10mM) was added. The mixture was shaken in the dark for 1h at 4°C and quenched for 15min at room temperature by the addition of 5.3µl 1M glycerol. Low molecular weight contaminates were removed by means of UF/DF employing Vivaspin (Sartorius, Goettingen, Germany) concentrators (30 kD membrane, regenerated cellulose). Next, 35.9mg aminooxy-PSA was added to the UF/DF retentate and the mixture was shaken for 2hrs at room temperature. Then 53µl of a aqueous sodium cyanoborohydride solution (5M) was added and the reaction was allowed to proceed for another 16 hrs. Then the excess PSA reagent was removed by means of HIC. The conductivity of the cooled reaction mixture was raised to 180mS/cm and loaded onto a 5ml HiTrap Butyl FF HIC (GE Healthcare, Fairfield, CT) column (1.6 x 2.5 cm), pre-equilibrated with 50mM HEPES, 3M sodium chloride, 6.7mM calcium chloride, 0.01% Tween 80, pH 6.9. The conjugate was eluted within 2.4CV with 50mM HEPES, 6.7mM calcium chloride, 0.005% Tween 80, pH 7.4 at a flow rate of 5 ml/min.

Example 7

Coupling of aminooxy-PSA (linker: $NH_2[OCH_2CH_2]_4ONH_2$) to rFIX and purification of the conjugate

[00125] To 5.6mg rFIX, dissolved in 2.8ml 50mM sodium acetate buffer, pH 6.0, 102 µl of an aqueous solution of sodium periodate (10mM) was added. The mixture was shaken in the dark for 1h at 4°C and quenched for 15min at room temperature by the addition of 2.9 µl of 1M glycerol. Low molecular weight contaminates were removed by means of UF/DF employing Vivaspin (Sartorius, Goettingen, Germany) concentrators (30kD membrane,

regenerated cellulose). Then 19mg aminooxy-PSA was added to the UF/DF retentate and the mixture was shaken for 18hrs at 4°C. The excess PSA reagent was removed by means of HIC. The conductivity of the cooled reaction mixture was raised to 180mS/cm and loaded onto a 5ml HiTrap Butyl FF (GE Healthcare, Fairfield, CT) HIC column (1.6 x 2.5cm), pre-equilibrated with 50mM HEPES, 3M sodium chloride, 6.7mM calcium chloride, 0.01% Tween 80, pH 6.9. The conjugate was eluted within 2.4CV with 50mM HEPES, 6.7mM calcium chloride, 0.005% Tween 80, pH 7.4 at a flow rate of 5ml/min.

Example 8

Coupling of aminooxy-PSA to rFVIII

[00126] To 11mg rFVIII, dissolved in 11ml Hepes buffer pH 6 (50mM Hepes, 5mM $CaCl_2$, 150mM NaCl, 0.01% Tween) 57µl 10mM sodium periodate was added. The mixture was shaken in the dark for 30min at 4°C and quenched for 30min at 4°C by the addition of 107µl of an aqueous1M glycerol solution. Then 19.8mg aminooxy-PSA (18.8kD) was added and the mixture was shaken over night at 4°C. The ionic strength was increased by adding a buffer containing 8M ammonium acetate (8M ammonium acetate, 50mM Hepes, 5mM $CaCl_2$, 350mM NaCl, 0.01% Tween 80, pH 6.9) to get a final concentration of 2.5M ammonium acetate. Next, the reaction mixture was loaded on a HiTrap Butyl FF (GE Healthcare, Fairfield, CT) column which was equilibrated with equilibration buffer (2.5M ammonium acetate, 50mM Hepes, 5mM $CaCl_2$, 350mM NaCl, 0.01% Tween 80, pH 6.9). The product was eluted with elution buffer (50mM Hepes, 5mM $CaCl_2$, 0.01% Tween 80, pH 7.4), and the eluate was concentrated by centrifugal filtration using Vivaspin (Sartorius, Goettingen, Germany) devices with 30,000 MWCO.

Example 9

PK studies in hemophilic mice

[00127] FIX-deficient mice were injected with either rFIX or PSA-rFIX (prepared according to Example 4) in formulation buffer (10mM histidine, 260mM glycine, 29mM sucrose, 0.005% Tween 80, pH 6.8) in a volume dose of 10 ml/kg bodyweight. Groups of 6 mice were sacrificed 5 minutes, 3 hours, 9, 16, 24 and 48 hours after substance injection and

blood was collected by heart puncture. Citrated plasma was prepared and stored frozen until analysis of FIX activity.

[00128] FIX activity was determined with a chromogenic FIX assay (Biophen FIX assay, Hyphen Biomed, Neuville-sur-Oise, France) and elimination curves were constructed (Figure 7). Actual FIX activity doses were 123IU FIX/kg for PSA-rFIX and 143IU FIX/kg for rFIX. Pharmacokinetic parameters were calculated with program R (The R Foundation for Statistical Computing, 2008). *In vivo* recovery was 13% for rFIX and 29% for PSA-rFIX. Dose adjusted AUC for PSA-rFIX increased 6.4-fold relative to rFIX, terminal half life increased by a factor of 1.2 and MRT was 1.7-times longer for PSA-rFIX compared to rFIX (Table 2).

Table 2

Item	In vivo recovery [%]	AUC [(IU/ml)/(IU/kg)]	Increase factor	Terminal HL [h]	Increase factor	MRT [h]	Increase factor
rFIX	13	0.0100	=1	8.0	=1	7.3	=1
PSA-rFIX	29	0.0650	6.4x	9.6	1.2x	12.3	1.7x

Example 10

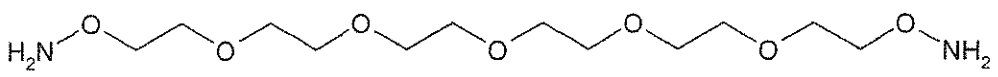
Polysialylation of blood coagulation proteins

[00129] Polysialylation as described herein may be extended to other coagulation proteins. For example, in various aspects of the invention, the above polysialylation as described in Examples 5, 6 and 9 with aminooxy-PSA is repeated with coagulation proteins such as FVIII, FVIIa and VWF.

Example 11

Preparation of the homobifunctional linker $NH_2[OCH_2CH_2]_6ONH_2$

[00130] The homobifunctional linker $NH_2[OCH_2CH_2]_6ONH_2$



(3,6,9,12,15-penatoxa-heptadecane-1,17-dioxyamine) containing two active aminooxy groups was synthesized according to Boturyn et al. (Tetrahedron 1997;53:5485-92) in a two step

organic reaction employing a modified Gabriel-Synthesis of primary amines. In the first step one molecule of hexaethylene glycol dichloride was reacted with two molecules of Endo-N-hydroxy-5-norbornene-2,3-dicarboximide in DMF. The desired homobifunctional product was prepared from the resulting intermediate by hydrazinolysis in ethanol.

Example 12

Polysialylation of rFIX employing a maleimido/aminooxy linker system

[00131] A. Preparation of the modification reagent

[00132] An Aminooxy-PSA reagent is prepared by use of a maleimido/aminooxy linker system (Toyokuni et al., Bioconjugate Chem 2003; 14, 1253-9). PSA-SH (20kD) containing a free terminal SH – group is prepared using a two step procedure: a) Preparation of PSA-NH_2 by reductive amination of oxidized PSA with NH_4Cl according to WO05016973A1 and b) introduction of a sulfhydryl group by reaction of the terminal primary amino group with 2-iminothiolane (Traut's reagent / Pierce, Rockford, IL) as described in US7645860. PSA-SH is coupled to the maleimido-group of the linker at pH 7.5 in PBS - buffer using a 10 fold molar excess of the linker and a PSA-SH concentration of 50 mg/ml. The reaction mixture is incubated for 2 hours under gentle shaking at room temperature. Then the excess linker reagent is removed and the aminooxy-PSA is buffer exchanged into oxidation buffer (50mM sodium phosphate, pH 6.0) by diafiltration. The buffer is exchanged 25 times employing a Pellicon XL5kD regenerated cellulose membrane (Millipore, Billerica, MA).

[00133] B. Modification of rFIX after prior oxidation with $NaIO_4$

[00134] rFIX is oxidized in 50 mM sodium phosphate buffer, pH 6.0 employing 100 µM sodium periodate in the buffer. The mixture was shaken in the dark for 1h at 4°C and quenched for 15 min at room temperature by the addition of glycerol to a final concentration of 5mM. Low molecular weight contaminates were removed by means of size exclusion chromatography (SEC) employing PD-10 desalting columns (GE Healthcare, Fairfield, CT). Oxidized rFIX is then spiked with aniline to obtain a final concentration of 10mM and mixed with the aminooxy-PSA reagent to achieve a 5 fold molar excess of PSA. The reaction mixture was incubated for 2 hours under gentle shaking in the dark at room temperature.

[00135] C. Purification of the conjugates

[00136] The excess of PSA reagent and free rFIX is removed by means of HIC. The conductivity of the reaction mixture is raised to 180 mS/cm and loaded onto a column filled with 48 ml Butyl - Sepharose FF (GE Healthcare, Fairfield, CT) pre-equilibrated with 50 mM Hepes, 3 M sodium chloride, 6.7 mM calcium chloride, 0.01% Tween 80, pH 6.9. Subsequently the conjugate is eluted with a linear gradient of 60% elution buffer (50mM Hepes, 6.7mM calcium chloride, pH 7.4) in 40 CV. Finally the PSA-rFIX containing fractions are collected and subjected to UF/DF by use of a 30 kD membrane made of regenerated cellulose (Millipore). The preparation is analytically characterized by measuring total protein (BCA) and FIX chromogenic activity. For the PSA-rFIX conjugates prepared with both variants a specific activity of > 50 % in comparison to native rFIX is determined.

Example 13

Preparation of aminooxy-PSA reagent

[00137] An Aminooxy - PSA reagent was prepared according to Example 3. The final product was diafiltrated against buffer, pH 7.2 (50 mM Hepes) using a 5 kD membrane (regenerated cellulose, Millipore), frozen at -80°C and lyophilized. After lyophilization the reagent was dissolved in the appropriate volume of water and used for preparation of PSA-protein conjugates via carbohydrate modification.

Example 14

Pharmacokinetics of polysialylated rFVIII in a FVIII deficient knock out mouse model

[00138] A PSA-FVIII conjugate was prepared according Example 8. The conjugate showed a specific activity of 6237 IU/mg (FVIII activity determined by the chromogenic assay; total protein determined by the Bradford assay) and had a polysialylation degree of 6.7 (mole PSA per mole FVIII) as measured by the Resorcinol assay (Svennerholm L, Biochim Biophys Acta 1957; 24: 604-11).

[00139] FVIII deficient mice described in detail by Bi et al. (Nat Genet 1995;10:119-21) were used as a model of severe human hemophilia A. Groups of 6 mice received a bolus injection (200 IU FVIII/kg) via the tail vein with either PSA-rFVIII prepared according to Example 8 or native rFVIII (ADVATE, Baxter Healthcare Corporation) in a dose of 200 IU FVIII/kg bodyweight. Citrate plasma by heart puncture after anesthesia was prepared from

the respective groups 5 minutes, 3, 6, 9, 16, 24, 32 and 42 hours after injection. FVIII activity levels were measured in plasma samples by use of the chromogenic assay. The results of this experiment are summarized in Table 3 and illustrated in Figure 8. All calculations were performed with R version 2.10.1 (A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. http://www.R-project.org.). As a result the mean residence time (MRT) increased from 5.4 h (Advate control) to 11.1 h for the PSA-rFVIII conjugate.

Table 3:

Item	In vivo recovery IVR %	AUC 0-24 (IU/ml.h)/IU/kg	Terminal half-life (h)	Mean residence time MRT (h)	Clearance CL (ml/h/kg)
PSA-rFVIII	71	0.161	7.2	11.1	6.0
rFVIII control (Advate)	58	0.054	4.4	5.4	17.1

Example 15

Detailed synthesis of the aminooxy-PSA reagent

[00140] 3-oxa-pentane-1,5 dioxyamine was synthesized according to Botyryn et al (Tetrahedron 1997; 53:5485-92) in a two step organic synthesis as outlined in Example 1.

[00141] Step 1:

[00142] To a solution of Endo-N-hydroxy-5-norbonene-2,3- dicarboxiimide (59.0g; 1.00eq) in 700 ml anhydrous N,N-dimetylformamide anhydrous K_2CO_3 (45.51g; 1.00eq) and 2,2-dichlorodiethylether (15.84 ml; 0.41eq) were added. The reaction mixture was stirred for 22 h at 50°C. The mixture was evaporated to dryness under reduced pressure. The residue was suspended in 2 L dichloromethane and extracted two times with saturated aqueous NaCl-solution (each 1 L). The Dichloromethane layer was dried over Na_2SO_4 and then evaporated to dryness under reduced pressure and dried in high vacuum to give 64.5 g of 3-oxapentane-1,5-dioxy-endo-2',3'-dicarboxydiimidenorbornene as a white-yellow solid (intermediate 1).

[00143] Step2:

[00144] To a solution of intermediate 1 (64.25 g; 1.00eq) in 800 ml anhydrous Ethanol, 31.0 ml Hydrazine hydrate (4.26eq) were added. The reaction mixture was then refluxed for 2hrs. The mixture was concentrated to the half of the starting volume by evaporating the solvent under reduced pressure. The occurring precipitate was filtered off. The remaining

ethanol layer was evaporated to dryness under reduced pressure. The residue containing the crude product 3-oxa-pentane -1,5-dioxyamine was dried in vacuum to yield 46.3g. The crude product was further purified by column chromatography (Silicagel 60; isocratic elution with Dichloromethane/Methanol mixture, 9+1) to yield 11.7 g of the pure final product 3-oxa-pentane -1,5-dioxyamine.

Example 16

Polysialylation of rFIX using PSA hydrazide

[00145] rFIX is polysialylated by use of a PSA hydrazide reagent, which was prepared by reaction of oxidized PSA with adipic acid dihydrazide (ADH).

Step 1: Preparation of PSA hydrazide

[00146] 500mg of oxidized PSA (MW = 20 kD) obtained from the Serum Institute of India (Pune, India) was dissolved in 8ml 50mM sodium acetate buffer, pH 5.5. 100mg adipic acid dihydrazide (ADH) was then added. The solution was gently shaken for 2 hrs. 44mg sodium cyanoborohydride were then added. After the reaction was incubated for an additional 4hrs at 4°C, the reaction mix was loaded into a Slide-A-Lyzer (Pierce, Rockford, IL) dialysis cassette (3.5 kD membrane, regenerated cellulose) and dialyzed against PBS pH 7.2 for 4 days. The product was frozen at -80°C.

Step 2: Reaction of PSA hydrazide with rFIX and purification of the conjugate

[00147] rFIX is polysialylated by use of a PSA hydrazide reagent as described in Step 1. rFIX (concentration 1 mg/ml) is oxidized with $NaIO_4$ (concentration: 80µM) for 1h at 4°C in the dark under gentle shaking. The reaction is stopped by addition of glycerol and the oxidized FIX is subjected to UF/DF by use of a 30 kD membrane made of regenerated cellulose (Vivaspin). The oxidized rFIX is then polysialylated at pH 6.5 using a 200-fold molar excess of reagent and a protein concentration of 1 mg/ml. rFIX and the polysialyation reagent are incubated for 2 hours under gentle shaking in the dark at room temperature. Finally, the PSA-rFIX conjugate is purified by HIC. The conductivity of the reaction mixture is raised to 130 mS/cm by adding a buffer containing ammonium acetate (50mM Hepes, 350mM NaCl, 5mM Calcium chloride, 8M ammonium acetate, 0.01% Tween 80, pH 6.9) and loaded onto a HiTrap Butyl FF column (5 ml, GE Healthcare, Fairfield, CT) pre-

equilibrated with 50mM Hepes, 2.5M ammonium acetate, 350mM sodium chloride, 5mM calcium chloride, 0.01% Tween 80, pH 6.9. Subsequently, the conjugate is eluted with 50mM Hepes, 5mM calcium chloride, 0.01% Tween 80, pH 7.4. Finally the PSA-rFIX containing fractions are collected and subjected to UF/DF by use of a 30 kD membrane made of regenerated cellulose (Vivaspin). For the PEG-rFIX conjugate, a specific activity of > 50% in comparison to native rFIX is determined (chromogenic assay).

Example 17

Polysialylation of rFIX using PSA hydrazide in the presence of aniline as a nucleophilic catalyst

[00148] 123mg rFIX are dissolved in 60ml phosphate buffer (50mM $NaPO_4$, pH 6.5) buffer. Then 1.2ml of an aqueous sodium periodate solution (10mM) is added and the mixture is incubated for 1h in the dark at 4°C under gentle stirring. Subsequently the reaction is quenched for 15min at RT by the addition of 600µl of 1M aqueous glycerol solution. The mixture is subsequently subjected to UF/DF employing a Pellicon XL Ultracel 30kD membrane.

[00149] The UF/D F retentate (63.4ml), containing oxidized rFIX, is further diluted with 59.6ml phosphate buffer (50mM $NaPO_4$, pH 6.0) and mixed with 6.5ml of an aqueous aniline solution (200mM) and incubated for 30 min at RT. Then 12.3ml of the PSA-hydrazide reagent (prepared according Example 16) is added to give a 5 fold molar reagent excess. This mixture is incubated for 2 h at RT in the dark under gentle stirring.

[00150] The excess of the PSA-hydrazide reagent and free rFIX is removed by means of HIC. The conductivity of the reaction mixture is raised to 180 mS/cm and loaded onto a column filled with 48 ml Butyl - Sepharose FF (GE Healthcare, Fairfield, CT) pre-equilibrated with 50mM Hepes, 3M sodium chloride, 6.7 mM calcium chloride, 0.01% Tween 80, pH 6.9. Subsequently the conjugate is eluted with 50mM Hepes, 5mM calcium chloride, 0.01% Tween 80, pH 7.4. Finally the PSA-rFIX containing fractions are collected and subjected to UF/DF by use of a 30 kD membrane made of regenerated cellulose (Millipore). The preparation is analytically characterized by measuring total protein (BCA) and FIX chromogenic activity. For the PSA-rFIX conjugate a specific activity of > 50% in comparison to native rFIX is determined.

Example 18

Polysialylation of rFIX and purification using a two step procedure

[00151] 140mg rFIX was dissolved in 62ml phosphate buffer (50mM $NaPO_4$,pH 6.0) buffer. Then 1.92ml of an aqueous sodium periodate solution (10mM) were added and the mixture was incubated for 1h in the dark at 4°C under gentle stirring and quenched for 15min at RT by the addition of 64µl of an 1M aqueous glycerol solution. Subsequently the mixture was subjected to UF/DF employing a Pellicon XL Ultracel 30kD membrane.

[00152] The UF/DF retentate (69.4ml), containing oxidized rFIX, was further diluted with 73.8ml phosphate buffer (50mM $NaPO_4$, pH 6.0), mixed with 8.2ml of an aqueous aniline solution (200mM) and incubated for 30 min at RT. Then 12.3ml of the aminooxy reagent (prepared according to Example 3) were added to give a 2.5 fold molar reagent excess. This mixture was incubated for 2.5h at RT in the dark under gentle stirring.

[00153] The free rFIX is removed by means of anion exchange chromatography (AIEC). The reaction mixture is diluted with 20ml Buffer A (50mM Hepes, 5mM $CaCl_2$, pH 7.5) and loaded onto a Q-Sepharose FF 26/10 column (GE Healthcare, Fairfield, CT) pre-equilibrated with Buffer A. Then the column is eluted with Buffer B (50mM Hepes, 1M NaCl, 5mM $CaCl_2$, pH 7.5). Free rFIX elutes at a conductivity between 12 - 25mS/cm and the conjugate between 27 - 45mS/cm. The conductivity of the conjugate containing fractions are subsequently raised to 190mS/cm by addition of Buffer C (50mM Hepes, 5M NaCl, 5mM $CaCl_2$, pH 6.9) and loaded onto a Butyl Sepharose FF 26/10 column (GE Healthcare, Fairfield, CT) pre-equilibrated with Buffer D (50mM Hepes, 3M NaCl, 5mM $CaCl_2$, pH 6.9). Free PSA-reagent is washed out within 5CV Buffer D. Subsequently the conjugate is eluted with 100% Buffer E (50mM Hepes, 5mM $CaCl_2$, pH 7.4). The conjugate containing fractions are concentrated by UF/DF using a 10 kD membrane made of regenerated cellulose (88cm^2, cut-off 10kD / Millipore). The final diafiltration step is performed against histidine buffer, pH 7.2 containing 150mM NaCl and 5mM $CaCl_2$. The preparation is analytically characterized by measuring total protein (BCA) and FIX chromogenic activity. For the PSA-rFIX conjugate a specific activity of > 50% in comparison to native rFIX is determined.

Example 19

Coupling of aminooxy-PSA to rFVIIa and purification of the conjugate

[00154] A solution of 10 mg rFVIIa in 5 ml reaction buffer (50 mM Hepes, 150 mM sodium chloride, 5 mM calcium chloride, pH 6.0) is mixed with an aqueous solution of $NaIO_4$ (final concentration: 100µM) and incubated for 1h at 4°C under gentle stirring in the dark and quenched by the addition of an aqueous solution of cysteine (final concentration: 1 mM) for 15 min. The reaction mixture is subsequently subjected to UF/DF. To the retentate (10 ml) a 30 fold molar excess of Aminooxy reagent (prepared according to Example 1) is added. The coupling reaction is performed for 2 hours at room temperature in the dark under gentle shaking. The excess of aminooxy reagent is removed by HIC. The conductivity of the reaction mixture is raised to 130 mS/cm by adding a buffer containing ammonium acetate (50 mM Hepes, 350 mM NaCl, 5 mM Calcium chloride, 8 M ammonium acetate, 0.01% Tween 80, pH 6.9) and loaded onto a HiTrap Butyl FF column (5 ml, GE Healthcare, Fairfield, CT) pre-equilibrated with 50 mM Hepes, 2.5 M ammonium acetate, 350 mM sodium chloride, 5 mM calcium chloride, 0.01% Tween 80, pH 6.9. Subsequently the conjugate is eluted with 50 mM Hepes, 5 mM calcium chloride, 0.01% Tween 80, pH 7.4 by a linear gradient of 100% elution buffer in 20 CV. Finally the PSA-rFVIIa containing fractions are collected and subjected to UF/DF by use of a 30 kD membrane made of regenerated cellulose (Vivaspin). The preparation is analytically characterized by measuring total protein (BCA) and FVIIa chromogenic activity (Staclot assay, Diagnostica Stago, Asnières, France) and shows a specific activity of > 20% compared to the rFVIIa starting material.

Example 20

Coupling of aminooxy-PSA to rFVIIa in the presence of aniline as nucleophilic catalyst

[00155] To 3.0mg rFVIIa, dissolved in 1.4ml 50mM sodium acetate buffer, pH 6.0, 14.1µl of an aqueous sodium periodate solution (10mM) is added. The mixture is shaken in the dark for 1h at 4°C and quenched for 15min at room temperature by the addition of 1.5µl 1M glycerol. Low molecular weight contaminates are removed by means of size exclusion chromatography (SEC) employing PD-10 desalting columns (GE Healthcare, Fairfield, CT). 3mg oxidized rFVIIa, dissolved in 3ml 50mM sodium acetate buffer, pH 6.0 is mixed with aniline (a nucleophilic catalyst, final concentration: 10mM) and shaken for 30 min at room temperature. Next, aminooxy-PSA is added to give a 5 fold molar excess and the mixture is shaken for 2hrs at room temperature. Subsequently the excess PSA reagent and free rFIX are removed by means of HIC. The conductivity of the cooled reaction mixture is raised to

180mS/cm and loaded onto a 5ml HiTrap Butyl FF (GE Healthcare, Fairfield, CT) HIC column (1.6 x 2.5cm), pre-equilibrated with 50mM Hepes, 3M sodium chloride, 6.7mM calcium chloride, 0.01% Tween 80, pH 6.9. The conjugate is eluted with a linear gradient to 50mM HEPES, 6.7mM calcium chloride, 0.005% Tween 80, pH 7.4 in 20 CV with at a flow rate of 5ml/min.

Example 21

Preparation of an Aminooxy-PEG reagent

[00156] A branched PEG-aldehyde (MW 40 kD) is used for coupling to the diaminooxy linker, which is prepared as described in Example 1. This PEG-aldehyde reagent is available from NOF (NOF Corp., Tokyo, Japan). 500 mg of PEG-aldehyde is dissolved in 8 ml 50 mM sodium acetate buffer, pH 5.5. Then 100 mg 3-oxa-pentane-1,5-dioxyamine is added. After shaking for 2 hrs at room temperature, 44 mg sodium cyanoborohydride is added. After shaking for another 4 hrs at 4°C, the reaction mix is loaded into a Slide-A-Lyzer (Pierce, Rockford, IL) dialysis cassette (3.5 kD membrane, regenerated cellulose) and dialyzed against PBS pH 7.2 for 4 days. The product is frozen at -80°C.

Example 22

PEGylation of rFIX with an aminooxy PEG-reagent

[00157] rFIX is PEGylated by use of a linear 20 kD PEGylation reagent containing an aminooxy group. An example of this type of reagent is the Sunbright ® CA series from NOF (NOF Corp., Tokyo, Japan). rFIX is oxidized at a protein concentration of 2mg/ml with $NaIO_4$ (final: concentration: 100 µM) for 1 hour under gentle shaking in the dark at 4°C in reaction buffer (50 mM Hepes, 150 mM sodium chloride, 5 mM calcium chloride, pH 6.0) and quenched by the addition of an aqueous solution of glycerol (final concentration: 1 mM) for 15 min. The reaction mixture is subsequently subjected to UF/DF. To the retentate a 3 fold molar excess of Aminooxy reagent and aniline (a nucleophilic catalyst, final concentration: 10mM) are added. The coupling reaction is performed for 2 hours at room temperature in the dark under gentle shaking. Finally the PEG-rFIX conjugate is purified by ion-exchange chromatography on Q-Sepharose FF. 1.5 mg protein/ml gel is loaded on the column pre equilibrated with 50 mM Tris, pH 8.0. The conjugate is eluted with 50 mM Tris

and 1 M sodium chloride, pH 8.0 in 20 CV and is then subjected to UF/DF using a 30 kD membrane. The preparation is analytically characterized by measuring total protein (BCA) and FIX chromogenic activity. For the PEG-rFIX conjugate a specific activity of > 75% in comparison to native rFIX is determined.

Example 23

PEGylation of rFVIII with an aminooxy PEG-reagent

[00158] rFVIII is PEGylated by use of a linear 20 kD PEGylation reagent containing an aminooxy group. An example of this type of reagent is the Sunbright ® CA series from NOF (NOF Corp., Tokyo, Japan). rFVIII is oxidized at a protein concentration of 1mg/ml with $NaIO_4$ (final: concentration: 100 µM) for 1 hour under gentle shaking in the dark at 4°C in reaction buffer (50 mM Hepes, 150 mM sodium chloride, 5 mM calcium chloride, pH 6.0) and quenched by the addition of an aqueous solution of cysteine (final concentration: 1 mM) for 15 min. The reaction mixture is subsequently subjected to UF/DF. To the retentate a 20 fold molar excess of Aminooxy reagent and aniline (a nucleophilic catalyst, final concentration: 10mM) are added. The coupling reaction is performed for 2 hours at room temperature in the dark under gentle shaking. Finally the PEG-rFVIII conjugate is purified by ion-exchange chromatography on Q-Sepharose FF. 1.5 mg protein/ml gel is loaded on the column pre equilibrated with 50 mM Hepes buffer, pH 7.4 containing 5 mM $CaCl_2$. The conjugate is eluted with 50 mM Hepes buffer containing 5 mM $CaCl_2$ and 500 mM sodium chloride, pH 7.4 and is then subjected to UF/DF using a 30 kD membrane. The analytical characterization of the conjugate by FVIII chromogenic assay and determination of total protein (BCA assay) shows a specific activity of > 60% compared to the rFVIII starting material.

Example 24

PEGylation of rFVIIa with an aminooxy PEG-reagent

[00159] rFVIIa is PEGylated by use of a linear 20 kD PEGylation reagent containing an aminooxy group. An example of this type of reagent is the Sunbright ® CA series from NOF (NOF Corp., Tokyo, Japan). rFVIIa is oxidized at a protein concentration of 2mg/ml with $NaIO_4$ (final: concentration: 100 µM) for 1 hour under gentle shaking in the dark at 4°C in

reaction buffer (50 mM Hepes, 150 mM sodium chloride, 5 mM calcium chloride, pH 6.0) and quenched by the addition of an aqueous solution of glycerol (final concentration: 1 mM) for 15 min. The reaction mixture is subsequently subjected to UF/DF. To the retentate a 5 fold molar excess of Aminooxy reagent and aniline (a nucleophilic catalyst, final concentration: 10mM) are added. The coupling reaction is performed for 2 hours at room temperature in the dark under gentle shaking. Finally the PEG-rFVIIa conjugate is purified by ion-exchange chromatography on Q-Sepharose FF. 1.5 mg protein/ml gel is loaded on the column pre equilibrated with 20mM Hepes buffer containing 1 mM $CaCl_2$, pH 7.4. The conjugate is eluted with 20mM Hepes buffer containing 1 mM $CaCl_2$ and 500 mM sodium chloride, pH 7.4 and is then subjected to UF/DF using a 30 kD membrane. The analytical characterization of the conjugate by measuring FVIIa activity (Staclot assay, Diagnostica Stago, Asnières, France) and total protein (BCA assay) shows a specific activity of > 25% compared to the rFVIIa starting material.

Example 25

PEGylation of rFIX with an PEG-hydrazide reagent

[00160] rFIX is PEGylated by use of a linear 20 kD PEGylation reagent containing a hydrazide group. An example of this type of reagent is the Sunbright ® HZ series from NOF (NOF Corp., Tokyo, Japan). rFIX is oxidized at a protein concentration of 2mg/ml with $NaIO_4$ (final: concentration: 100 µM) for 1 hour under gentle shaking in the dark at 4°C in reaction buffer (50 mM Hepes, 150 mM sodium chloride, 5 mM calcium chloride, pH 6.0) and quenched by the addition of an aqueous solution of glycerol (final concentration: 1 mM) for 15 min. The reaction mixture is subsequently subjected to UF/DF. To the retentate a 50 fold molar excess of Hydrazide reagent and aniline (a nucleophilic catalyst, final concentration: 10mM) are added. The coupling reaction is performed for 2 hours at room temperature in the dark under gentle shaking. Finally the PEG-rFIX conjugate is purified by ion-exchange chromatography on Q-Sepharose FF. The reaction mixture is loaded onto the column (1.5 mg protein/ml gel), which is preequilibrated with 50 mM Tris - buffer, pH 8.0. The conjugate is eluted with 20 CV Tris – buffer, pH 8.0 (50 mM Tris, 1 M NaCl) and is then subjected to UF/DF using a 30 kD membrane. The preparation is analytically characterized by measuring total protein (BCA) and FIX chromogenic activity. For the PEG-rFIX

conjugate a specific activity of > 50% in comparison to native rFIX is determined (chromogenic assay).

Example 26

Polysialylation of rFVIII in the presence of 2mM aniline

[00161] rFVIII is transferred into reaction buffer (50 mM Hepes, 350 mM sodium chloride, 5 mM calcium chloride, 0.01% Tween 80, pH 6), diluted to a protein concentration of 1 mg/ml and oxidized with $NaIO_4$ (final: concentration: 100 µM) for 1 hour under gentle shaking in the dark at 4°C in reaction buffer (50 mM Hepes, 150 mM sodium chloride, 5 mM calcium chloride, pH 6.0) and quenched by the addition of an aqueous solution of cysteine (final concentration: 1 mM) for 15 min. The reaction mixture is subsequently subjected to UF/DF. To the retentate a 20 fold molar excess of Aminooxy reagent and aniline (a nucleophilic catalyst, final concentration: 2mM) are added. The coupling reaction is performed for 2 hours at room temperature in the dark under gentle shaking. The excess of aminooxy reagent is removed by means of HIC. The conductivity of the reaction mixture is raised to 130 mS/cm by adding a buffer containing ammonium acetate (50 mM Hepes, 350 mM sodium chloride, 5 mM calcium chloride, 8 M ammonium acetate, 0.01% Tween 80, pH 6.9) and loaded onto a column filled with 53 ml Butyl - Sepharose FF (GE Healthcare, Fairfield, CT) pre-equilibrated with 50 mM Hepes, 2.5 M ammonium acetate, 350 mM sodium chloride, 5 mM calcium chloride, 0.01% Tween 80, pH 6.9. Subsequently the conjugate is eluted with 50 mM Hepes, 5 mM calcium chloride, 0.01% Tween 80, pH 7.4. Finally the PSA-rFIX containing fractions are collected and subjected to UF/DF by use of a 30 kD membrane made of regenerated cellulose (Millipore, Billerica, MA). The preparation is analytically characterized by measuring total protein (BCA) and FVIII chromogenic activity. For the PSA-rFVIII conjugate a specific activity of 80% in comparison to native rFVIII is determined.

Example 27

Polysialylation of rFVIII in the presence of 10mM aniline

[00162] rFVIII is transferred into reaction buffer (50 mM Hepes, 350 mM sodium chloride, 5 mM calcium chloride, 0.01% Tween 80, pH 6), diluted to a protein concentration

of 1 mg/ml and oxidized with $NaIO_4$ (final: concentration: 100 µM) for 1 hour under gentle shaking in the dark at 4°C in reaction buffer (50 mM Hepes, 150 mM sodium chloride, 5 mM calcium chloride, pH 6.0) and quenched by the addition of an aqueous solution of cysteine (final concentration: 1 mM) for 15 min. The reaction mixture is subsequently subjected to UF/DF. To the retentate a 20 fold molar excess of Aminooxy reagent and aniline (a nucleophilic catalyst, final concentration: 10mM) are added. The coupling reaction is performed for 2 hours at room temperature in the dark under gentle shaking. The excess of aminooxy reagent is removed by means of HIC. The conductivity of the reaction mixture is raised to 130 mS/cm by adding a buffer containing ammonium acetate (50 mM Hepes, 350 mM sodium chloride, 5 mM calcium chloride, 8 M ammonium acetate, 0.01% Tween 80, pH 6.9) and loaded onto a column filled with 53 ml Butyl - Sepharose FF (GE Healthcare, Fairfield, CT) pre-equilibrated with 50 mM Hepes, 2.5 M ammonium acetate, 350 mM sodium chloride, 5 mM calcium chloride, 0.01% Tween 80, pH 6.9. Subsequently the conjugate is eluted with 50 mM Hepes, 5 mM calcium chloride, 0.01% Tween 80, pH 7.4. Finally the PSA-rFIX containing fractions are collected and subjected to UF/DF by use of a 30 kD membrane made of regenerated cellulose (Millipore, Billerica, MA). The preparation is analytically characterized by measuring total protein (BCA) and FVIII chromogenic activity. For the PSA-rFVIII conjugate a specific activity of 80% in comparison to native rFVIII is determined.

Example 28

PEGylation of a blood coagulation protein using branched PEG

[00163] PEGylation of a blood coagulation proteins (such as FIX, FVIII and FVIIa as described in Examples 22-25) may be extended to a branched or linear PEGylation reagent as described in Example 21, which is made of an aldehyde and a suitable linker containing an active aminooxy group.

Claims:

1. A modified blood coagulation protein comprising:

(a) a blood coagulation protein selected from the group consisting of

(i) a blood coagulation protein with Factor IX (FIX) biological activity; and

(ii) a blood coagulation protein with Factor VIII (FVIII) biological activity; and

(b) at least one polysialic acid (PSA) containing an active aminooxy group bound to said blood coagulation protein of (a) at one or more oxidized carbohydrate moieties;

wherein said modified blood coagulation protein has an oxime linkage between said one or more oxidized carbohydrate moieties and the active aminooxy group on the PSA;

wherein said modified blood coagulation protein has a specific activity of at least 50% relative to an unmodified blood coagulation protein.

2. The modified blood coagulation protein of claim 1 wherein said blood coagulation protein is a blood coagulation protein with FIX biological activity, and wherein said modified blood coagulation protein has a specific activity of at least 60% relative to an unmodified blood coagulation protein.

3. The modified blood coagulation protein of claim 1 wherein said blood coagulation protein is a blood coagulation protein with FVIII biological activity, and wherein said modified blood coagulation protein has a specific activity of at least 70% relative to an unmodified blood coagulation protein.

4. A method of conjugating a polysialic acid (PSA) containing an active aminooxy group to an oxidized carbohydrate moiety of a blood coagulation protein, comprising contacting said oxidized carbohydrate moiety with at least one activated PSA under conditions that allow conjugation;

said blood coagulation protein selected from the group consisting of

(i) a blood coagulation protein with FIX biological activity; and

(ii) a blood coagulation protein with FVIII biological activity;

said carbohydrate moiety oxidized by incubation with a buffer comprising an oxidizing agent selected from the group consisting of sodium periodate ($NaIO_4$), lead tetraacetate ($Pb(OAc)_4$) and potassium perruthenate (KRuO4); wherein an oxime linkage is formed between the oxidized carbohydrate moiety and the active aminooxy group on the PSA;

said modified blood coagulation protein having a specific activity of at least 50% relative to an unmodified blood coagulation protein.

5. The method of claim 4 wherein said blood coagulation protein is a blood coagulation protein with FIX biological activity, and wherein said modified blood coagulation protein has a specific activity of at least 60% relative to an unmodified blood coagulation protein.

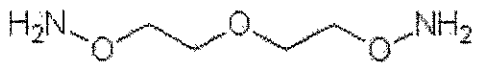
6. The method of claim 4 wherein said blood coagulation protein is a blood coagulation protein with FVIII biological activity, and wherein said modified blood coagulation protein has a specific activity of at least 70% relative to an unmodified blood coagulation protein.

7. The method according claim 4 wherein the PSA is comprised of about 10 – 300 sialic acid units.

8. The method according to claim 4 wherein the PSA is prepared by reacting an activated aminooxy linker with oxidized PSA;

wherein the aminooxy linker is selected from the group consisting of:

(a) a 3-oxa-pentane-1,5-dioxyamine linker of the formula:



and

(b) a 3,6,9-trioxa-undecane-1,11-dioxyamine linker of the formula:



wherein the PSA is oxidized by incubation with a oxidizing agent to form a terminal aldehyde group at the non-reducing end of the PSA.

9. The method according to claim 8 wherein the aminooxy linker is 3-oxa-pentane-1,5-dioxyamine.

BLOOD COAGULATION PROTEIN CONJUGATES

Abstract

The invention relates to materials and methods of conjugating a water soluble polymer to an oxidized carbohydrate moiety of a blood coagulation protein comprising contacting the oxidized carbohydrate moiety with an activated water soluble polymer under conditions that allow conjugation. More specifically, the present invention relates to the aforementioned materials and methods wherein the water soluble polymer contains an active aminooxy group and wherein an oxime linkage is formed between the oxidized carbohydrate moiety and the active aminooxy group on the water soluble polymer. In one embodiment of the invention the conjugation is carried out in the presence of the nucleophilic catalyst aniline. In addition the generated oxime linkage can be stabilized by reduction with $NaCNBH_3$ to form an alkoxyamine linkage.

No suitable figure.