HYDROPHOBIC PREPARATIONS CONTAINING MEDIUM CHAIN MONOGLYCERIDES

The present invention relates to preparations of substances in hydrophobic solvents in which they would not normally be soluble and to processes for obtaining these preparations. In particular, the invention relates to preparations of hydrophilic species in mixtures of medium chain monoglycerides (MCMs) and diglycerides.

The invention in particular applies to hydrophilic macromolecules which would not normally be soluble in oils or other hydrophobic solvents.

For many applications, e.g in the pharmaceutical sciences, in food technology or the cosmetics industry, work with proteins and similar macromolecules presents problems because their hydrophilicity and high degree of polarity limit the extent to which they can interact with or incorporate into lipid phases. Many natural systems employ lipidic barriers (eg skin, cell membranes) to prevent access of hydrophilic molecules to internal compartments; the ability to disperse proteins in lipidic vehicles would open up a new route to introduction of these macromolecules into biological systems, whereby the lipid medium containing the protein can integrate with the hydrophobic constituents of barriers, instead of being excluded by them.

We have previously disclosed, in WO-A-9513795, WO-A-9617593 and W0-A-9617594, methods for preparing hydrophobic preparations where a hydrophilic species is solubilised in a hydrophobic phase in which it would not normally be soluble. In particular, these methods are suitable for solubilising proteins.

Although the above-described preparations provide simple and efficient methods for solubilising macomolecules such as proteins, we have now found that the macromolecule delivery properties of the preparations can be improved by the use of a

particular oil phase and, optionally, particular amphiphiles. This is particularly advantageous when the macromolecule to be solubilised is a protein. eg a pharmaceutically active protein, since the preparations disclosed herein provide not only enhanced uptake of the therapeutic macromolecule but also good dose repeatability.

Thus, in a first aspect, the present invention provides a hydrophobic preparation comprising:

(i) an oil phase comprising one or more medium chain monoglycerides: and

(ii) at least one amphiphile;

(iii) a hydrophilic species solubilised or otherwise dispersed in the mixture of glycerides;

wherein the hydrophilic species is one that is not normally soluble in the one or more monoglycerides.

In the context of the present invention "hydrophobic preparation" is a preparation in which the hydrophilic species is not present in aqueous phase. Such a hydrophobic preparation is particularly suitable for use in orally delivering a hydrophilic macromolecule such as a protein.

The prior art does contain a number of examples of the use of medium chain monoglycerides as permeation enhancers in the intestine (Sekine *et al*, *J.Pharmacobiodyn.*, 7:856-63 (1984); Higaki *et al*, *J.Pharmacobiodyn.*, 9:532-9 (1986); Unowsky *et al*, *Chemotherapy*, 34:272-6 (1988); Watanabe *et al*, *J.Pharm.Sci.*, 77:847-9 (1988); Yeh *et al*, *Pharm.Res.*, 11:1148-54 (1994);

Constinides *et al*, *Pharm.Res.*, 11:1385-90 (1994)). However, in every case the formulations disclosed are ones where the active principle/drug is solubilised in an aqueous phase. In fact until the methods disclosed in, *inter alia*, WO-A-9513795 preparations where a hydrophilic species was truly and readily solubilised in a hydrophobic phase, with retention of biological activity, were not available.

Preparations in accordance with the invention will generally have no bulk aqueous phase and may have no free water molecules.

In a preferred embodiment the oil phase i) will comprise a mixture of medium chain mono- and diglycerides. Suitably, medium chain glycerides useful in the present invention have chain lengths of 8 to 10 carbon atoms, for example, they can comprise straight chain saturated fatty acids. In another embodiment the oil phase i) may comprise one or more medium chain monoglycerides together with at least one other component such as oleic acid, glycerol mono-oleate or gelucires. For both these embodiments the essential component will be the medium chain monoglyceride(s). Whether medium chain monoglyceride(s) are used alone or as mixtures of glycerides or the like, the oil component used should be such that the amount of monoglyceride(s) present should be maximised while ensuring that the oil component remains liquid at a temperature of 45°C or lower. In particular, monoglyceride(s) can make up 40-90% of the total amount of oil present, preferably 60-70%. An example of a suitable mixture of glycerides is Akoline MCM™ which contains both medium chain mono- and diglycerides, available from Karlshamns Sweden AB, S/374 82 Karlshamn, Sweden.

Preferably, the ratio of amphiphile:macromolecule is in the range 1:1 to 20:1 by weight and more preferably in the range 2:1 to 8:1 by weight.

Examples of suitable amphiphiles include phospholipids such as phosphatidyl choline, phosphatidic acid, phosphatidyl glycerol, phosphatidyl ethanolamine and lyso-

derivatives of these, octyl glucoside and other glycolipids, tocopherol succinate and cholesterol hemisuccinate. Other suitable amphiphiles include phosphatidyl serine, sodium docusate and hydroxypropyl cellulose. More than one amphiphile may be used.

In one preferred embodiment the amphiphile used is a bile salt. In the present invention it should be understood that the terms bile salt and bile acid are used interchangeably because whether the salt or its conjugate acid is present will depend on the pH of the surrounding environment.

Bile salts are naturally occurring surfactants. They are a group of compounds with a common "backbone" structure based on cholanic acid found in all mammals and higher vegetables. Bile salts may be mono-, di- or tri-hydroxylated; they always contain a 3α-hydroxyl group whereas the other hydroxyl groups. most commonly found at C_6, C_7 or C_{12}, may be positioned either above (β) or below (α) the plane of the molecule.

Within the class of compounds described as bile salts are included amphiphilic polyhydric sterols bearing carboxyl groups as part of the primary side chain. The most common examples of these in mammals result from cholesterol metabolism and are found in the bile and, in derivatised form, throughout the intestine.

In the context of this specification, the term may also apply to synthetic analogues of naturally occurring bile salts which display similar biological effects, or to microbially derived molecules such as fusidic acid and its derivatives.

The bile salt (or salts) may be either unconjugated or conjugated. The term "unconjugated" refers to a bile salt in which the primary side chain has a single carboxyl group which is at the terminal position and which is unsubstituted. Examples of unconjugated bile salts include cholate, ursodeoxycholate, chenodeoxycholate and

deoxycholate. A conjugated bile salt is one in which the primary side chain has a carboxyl group which is substituted. Often the substituent will be an amino acid derivative which is linked via its nitrogen atom to the carboxyl group of the bile salt. Examples of conjugated bile salts include taurocholate, glycocholate, taurodeoxycholate and glycodeoxycholate.

Thus. in the present invention, examples of suitable bile salts include cholate, deoxycholate, chenodeoxycholate and ursodeoxycholate, with ursodeoxycholate being particularly preferred. Other bile salts which may be employed include taurocholate, taurodeoxycholate, tauroursodeoxycholate, taurochenodeoxycholate, glycholate, glycodeoxycholate, glycoursodeoxycholate, glycochenodeoxycholate, lithocholate, taurolithocholate and, glycolithocholate.

In the present invention the term "hydrophilic species" relates to any species which is generally soluble in aqueous solvents but insoluble in hydrophobic solvents. The range of hydrophilic species of use in the present invention is diverse but hydrophilic macromolecules represent an example of a species which may be used.

A wide variety of macromolecules is suitable for use in the present invention. In general, the macromolecular compound will be hydrophilic or will at least have hydrophilic regions since there is usually little difficulty in solubilising a hydrophobic macromolecule in oily solutions. Examples of suitable macromolecules include proteins and glycoproteins, oligo and polynucleic acids, for example DNA, eg plasmid DNA. and RNA, as well as DNA and/or RNA analogues, polysaccharides such as heparin (particularly low molecular weight heparin) and supramolecular assemblies of any of these including, in some cases, whole cells or organelles. It may also be convenient to co-solubilise a small molecule such as a vitamin in association with a macromolecule, particularly a polysaccharide such as a cyclodextrin. Small molecules such as vitamin B12 may also be chemically conjugated with macromolecules and may

thus be included in the compositions.

Examples of particular proteins which may be successfully solubilised by the method of the present invention include insulin, calcitonin, haemoglobin, cytochrome C, horseradish peroxidase, aprotinin, mushroom tyrosinase, erythropoietin, somatotropin, growth hormone, growth hormone releasing factor, galanin. urokinase. Factor IX, tissue plasminogen activator, superoxide dismutase, catalase, peroxidase, ferritin, interferon. Factor VIII and fragments thereof (all of the above proteins can be from any suitable source). Other proteins include soy bean trypsin inhibitor, GLP1, other blood coagulation factors, somatostatin, hirudin, and LHRH and analogues and fragments of all of them.

Mixtures of one or more of these or other proteins may be solubilised by the invention.

It seems that there is no upper limit of molecular weight for the macromolecular compound since dextran having a molecular weight of about 1,000,000 can easily be solubilised by the process of the present invention.

In addition to macromolecules, the process of the present invention is of use in solubilising smaller organic molecules as well as or instead of macromolecules. Examples of small organic molecules include glucose, carboxyfluorescin and many pharmaceutical agents. for example anti-cancer agents, but, of course. the process could equally be applied to other small organic molecules, for example vitamins or pharmaceutically or biologically active agents. In addition, compounds such as calcium chloride and sodium phosphate can also be solubilised using this process. Indeed, the present invention would be particularly advantageous for pharmaceutically and biologically active agents since the use of non aqueous solutions may enable the route by which the molecule enters the body to be varied, for example to increase bioavailability.

Small organic molecules which may be incorporated into macromolecule-containing preparations of the invention include stabilising agents such as polyglycerols, PEGs and glycerol (particularly in the case of insulin or, possibly, other proteins), chelating agents, such as citric acid, EDTA and EGTA, and antioxidants such as ascorbate.

Another type of species which may be included in the hydrophobic compositions of the invention is an inorganic material such as a small inorganic molecule or a colloidal substance, for example a colloidal metal. The process of the present invention enables some of the properties of a colloidal metal such as colloidal gold, palladium, platinum or rhodium, to be retained even in hydrophobic solvents in which the particles would, under normal circumstances, aggregate. This could be particularly useful for catalysis of reactions carried out in organic solvents.

The hydrophobic preparations of the invention may also optionally comprise further components. Examples of these include antioxidants, metal chelating agents, buffering agents and dispersion agents. Examples of suitable dispersion agents include surface active agents such as the Tween, Span and Brij classes of agent, as well as polyoxyethylated castor oil derivatives, and other POE-containing surfactants.

The hydrophobic preparations of the present invention can be prepared using a method comprising:

(i) associating the hydrophilic species with the amphiphile in a liquid medium such that, in the liquid medium, there is no chemical interaction between the amphiphile and the hydrophilic species;

(ii) removing the liquid medium to leave an array of amphiphile molecules with their hydrophilic head groups orientated towards the hydrophilic species;

and

(iii) providing a mixture of medium chain mono- and diglycerides around the hydrophilic species/amphiphile array.

Such methods are disclosed in WO-A-9513795. In particular, the hydrophobic preparations of the present invention can be prepared by methods disclosed in PCT patent application No. PCT/GB97/00749 which comprise:

(i) associating the hydrophilic species with the amphiphile in the presence of a hydrophobic phase; and

(ii) removing any hydrophilic solvent which is present;

wherein the hydrophilic solvent removal step is carried out under conditions which maintain the hydrophobic phase in a solid state.

Preferably, when using this method, the hydrophilic species and the amphiphile are first dissolved in a hydrophilic solvent, eg an aqueous solvent, often water alone, and this solution is then brought into association with the glyceride mixture. The hydrophilic solvent removal step is conveniently achieved by lyophilisation, such that it is carried out at temperatures which will ensure that the glyceride mixture is maintained in the solid state until all the water has been removed. Under certain circumstances, the oil may become liquid during lyophilisation, as a result of local rises in temperature in parts of the solid block (usually at the surface and edges) where all the hydrophilic solvent has already been removed. Here the cooling effect deriving from sublimation of hydrophilic solvent no longer exists, and in those areas the oil will melt. This situation will lead to the production of a satisfactory end-product providing that the oil is allowed to drain away from the remainder of the solid block as soon as it

appears (if not, then accumulating oil will form a layer which prevents further removal of hydrophilic solvent).

Alternatively, the temperature during lyophilisation can be maintained such that the oil remains solid even after the hydrophilic solvent has been driven off. In this case following lyophilisation, the temperature of the preparation is elevated to produce the single phase preparation. This can often simply be achieved by bringing the lyophilised preparation up to room temperature which in turn will cause the glyceride mixture to return to the liquid state. Other methods for removal of hydrophilic solvent may also be employed, eg spray drying.

The hydrophobic preparations of the present invention are extremely versatile and have many applications. They may either be used alone or they may be combined with an aqueous phase to form an emulsion or similar two phase composition which forms a second aspect of the invention.

In this aspect of the invention there is provided a two phase composition comprising a hydrophilic phase and a hydrophobic phase, the hydrophobic phase comprising a hydrophobic preparation of the invention which is obtainable by the methods described above.

Generally, in this type of composition, the hydrophobic phase will be dispersed in the hydrophilic phase.

As mentioned herein the hydrophobic preparations of the invention have particular advantages in that the hydrophilic species is readily taken up, eg into the bloodstream following oral administration. They are therefore particularly suitable for the oral delivery of proteins for example. However, the skilled man will appreciate that the preparations of the invention will also provide advantages for other routes of

administration, eg topical or vaginal. Thus, in a third aspect the present invention provides a pharmaceutical formulation comprising a hydrophobic preparation of the invention. Pharmaceutical formulations within the scope of the invention include capsules, tablets and other presentations. In addition, in a fourth aspect the present invention provides the use of a hydrophobic preparation of the invention in the preparation of a medicament for oral delivery of a hydrophilic species, for instance a hydrophilic macromolecule such as a protein. The invention can also be used to modify the immune response, whether by stimulation or suppression.

Preferred features of each aspect of the invention are equally applicable to each other aspect *mutatis mutandis*.

The invention will now be described with reference to the following examples which should not be construed as in any way limiting the invention.

EXAMPLE 1: PREPARATION OF FORMULATION CONTAINING CALCITONIN/PC COMPLEX

(i) Preparation of Phospholipid Dispersion

1 Weigh out 1.6g of SOYA PHOSPHATIDYL CHOLINE in a boiling tube with a ground glass stopper and add DISTILLED WATER to give a final weight of 8g. Flush with nitrogen, stopper well and seal with parafilm and mix gently on orbital shaker until all the solid has dispersed.

2 Transfer to a glass round-bottomed sonicator vessel.

3 Clamp the sonicator vessel into the Sonics 4 Materials Vibra Cell VC X 600 Sonicator fitted with a 1 inch diameter probe and immerse the probe until its base is

sufficiently below the meniscus of the liquid. Use a strip of cling film to form a sleeve between the probe and the top of the tube and purge the air space above the liquid with nitrogen. Immerse the sonicator vessel in an ice slurry bath.

4 Sonicate the lipid suspension at an amplitude of 50%, in 1 second bursts interspersed with 4 second cooling intervals, until an opalescent dispersion is formed (normally 4 minutes total sonication time).

5 Transfer the dispersion to a plastic conical centrifuge tube and centrifuge for 15 minutes at 1200 g. Separate the supernatant from any pellet that is present.

6 Dilute two-fold in distilled water to give a final concentration of 100mg/ml of phospholipid.

(ii) Preparation of Protein Solution

7 Weigh out 20mg of APROTININ in a 2ml glass screw-capped vial and add 1ml of DISTILLED WATER. Screw cap tightly and mix gently until all the solid is dissolved.

8 Weigh out 16mg of SALMON CALCITONIN in a ml glass screw-capped vial and add 0.8ml of DISTILLED WATER. Screw cap tightly and mix gently until all the solid is dissolved.

9 Weigh out 50mg of ASCORBIC ACID and 5mg of CITRIC ACID in a ml glass screw-capped vial and add 1ml of DISTILLED WATER. Screw cap tightly and mix gently until all the solid is dissolved.

10 To each of three glass screw-capped 10ml vials dispense 2ml of phospholipid

dispersion (200mg solid) from step 6, 250µl of aprotinin solution (5mg of solid) from step 7, 250µl of calcitonin solution (5mg of solid) from step 8, and 200µl of ascorbic acid/citric acid solution (10mg and 1mg of solid respectively) from step 9 Mix contents of each vial gently.

(iii) Lyophilisation of Aqueous Phase

11 Freeze the contents of each vial rapidly in liquid nitrogen.

12 Lyophilise overnight at a condenser temperature of less than -40°C and a vacuum of less than 0.1 mBar.

(iv) Preparation of Oil Phase

13 Weigh out 0.8g of POLYSORBATE 80 and 7.2g of AKOLINE MCM into a B8 glass screw-capped vial. Flush vial with nitrogen, screw cap tightly and seal with parafilm and mix gently on roller mixer until a homogenous solution has been obtained.

(v) Solubilisation in Oil Phase

14 After samples have dried down fully in the lyophiliser, add to each vial 1.779g of oil phase from step 14. Flush each vial with nitrogen, screw caps tightly and seal with parafilm.

15 Allow solids to dissolve by mixing on roller mixer at room temperature for two hours, followed by shaking at 37°C until a clear solution has formed.

16 Store at +4°C until required for use.

(vi) Administration *in vivo*

17 Warm each vial to 37°C in a water bath in order to melt the oil and form a clear solution.

18 To each vial add 4ml of warm PBS, vortex for 30 seconds and administer 1.2ml to one pig i.j. as described above.

EXAMPLE 2: PREPARATION OF FORMULATION CONTAINING INSULIN/CHENODEOXYCHOLATE COMPLEX

(i) Preparation of Chenodeoxycholate Solution

1 Weigh out 6.0g of SODIUM CHENO DEOXY CHOLATE in a glass conical flask and add DISTILLED WATER to give a final weight of 60g. Flush with nitrogen, stopper well and seal with parafilm and mix gently on orbital shaker until all the solid has dissolved.

(ii) Preparation of Protein Solution

2 Weigh out 120mg of APROTININ in a 2ml glass screw-capped vial and add 6.0ml of DISTILLED WATER. Screw cap tightly and mix gently until all the solid is dissolved.

3 Weigh out three separate lots of 106.5mg of INSULIN into (A) a 100ml conical flask, (B) a 25ml conical flask and (C) a 10ml glass screw-capped vials. To flask (A) add 30ml of chenodeoxycholate solution (3g solid) from step 3.1.1. To flask (B) add 15ml of chenodeoxycholate solution (1.5g solid), and to vial (C) add 7.5 ml (0.75g

solid). Cover flasks and vial with parafilm and mix gently on orbital shaker at 37°C until all the solid is dissolved.

4 Weigh out 54mg of CITRIC ACID in a 2ml glass screw-capped vial and add 6.0ml of DISTILLED WATER. Screw cap tightly and mix gently until all the solid is dissolved.

5 To each of twelve glass screw-capped 7ml vials dispense 2ml of insulin/chenodeoxycholate solution (A) from step 3, 100µl of aprotinin solution from step 2, and 100µl of citric acid solution from step 4. Mix contents of each vial gently.

6 To each of twelve glass screw-capped 7ml vials dispense 1ml of insulin/chenodeoxycholate solution (B) from step 3, 100µl of aprotinin solution from step 2, and 100µl of citric acid solution from step 4. Mix contents of each vial gently.

7 To each of twelve glass screw-capped 7ml vials dispense 0.5ml of insulin/chenodeoxycholate solution (C) from step 3, 100µl of aprotinin solution from step 2, and 100µl of citric acid solution from step 4. Mix contents of each vial gently.

(iii) Lyophilisation of Aqueous Phase

8 Freeze the contents of each vial rapidly in liquid nitrogen and lyophilise overnight at a condenser temperature of less than -40°C and a vacuum of less than 0.1 mBar.

(iv) Preparation of Oil Phase

9 Weigh out 6.0g of POLYSORBATE 80 and 54g of AKOLINE MCM into a 100ml glass bottle. Flush vial with nitrogen, screw cap tightly, seal with parafilm and

mix gently on roller mixer until a homogenous solution has been obtained.

(v) Solubilisation in Oleic Acid

10 After samples have dried down fully in the lyophiliser, add to each vial of preparation (A) 0.79g of oil phase from step 10. Flush each vial with nitrogen, screw caps tightly and seal with parafilm.

11 After samples have dried down fully in the lyophiliser, add to each vial of preparation (B) 0.89g of oil phase from step10. Flush each vial with nitrogen, screw caps tightly and seal with parafilm.

12 After samples have dried down fully in the lyophiliser, add to each vial of preparation (C) 0.94g of oil phase from step 10. Flush each vial with nitrogen, screw caps tightly and seal with parafilm.

13 Allow solids to dissolve by mixing on roller mixer at room temperature for two hours, followed by shaking at 37°C until a clear solution has formed and store at +4°C until required for use.

(vi) Administration *in vivo*

14 Warm each vial to 37°C in a water bath in order to melt the oil and form a clear solution.

15 To each vial add 2ml of warm PBS, vortex for 30 seconds and administer the contents of one whole vial to one pig i.j. as described above.

EXAMPLE 3: PREPARATION OF FORMULATION CONTAINING INSULIN/PC COMPLEX

(i) Preparation of Phospholipid Dispersion

1 Weigh out 2g of SOYA PHOSPHATIDYL CHOLINE in a boiling tube with a ground glass stopper and add DISTILLED WATER to give a final weight of 8g. Flush with nitrogen, stopper well and seal with parafilm and mix gently on orbital shaker until all the solid has dispersed.

2 Transfer to a glass round-bottomed sonicator vessel.

3 Clamp the sonicator vessel into the Sonics 4 Materials VibraCell VC x 60 ultrasonicator fitted with a 1 inch diameter probe and immerse the probe until its base is 1 cm below the meniscus of the liquid. Use a strip of cling film to form a sleeve between the probe and the top of the tube and purge the air space above the liquid with nitrogen. Immerse the sonicator vessel in an ice slurry bath.

4 Sonicate the lipid suspension at an amplitude of 50%, in 1 second bursts interspersed with 4 second cooling intervals, until an opalescent dispersion is formed (normally 4 minutes total sonication time).

5 Transfer the dispersion to a plastic conical centrifuge tube and centrifuge for 15 minutes at 1200 g. Separate the supernatant from any pellet that is present.

(ii) Preparation of Protein Solution

6 Weigh out 30mg of APROTININ in a 2ml glass screw-capped vial and add 1.5ml of DISTILLED WATER. Screw cap tightly and mix gently until all the solid is

dissolved.

7 Weigh out 110mg of INSULIN into a 25ml conical flask and add all 15ml of DISTILLED WATER to which 150µl of GLACIAL ACETIC ACID has been added. Cover the flask with parafilm and mix gently on orbital shaker at 37°C until all the solid is dissolved.

8 Weigh out 10mg of SODIUM CITRATE in a 2ml glasss screw-capped vial and add 1.5ml of DISTILLED WATER. Screw cap tightly and mix gently until all the solid is dissolved.

9 To each of twelve glass screw-capped 7ml vials dispense 1ml of insulin solution (7.33mg solid, 200iu) from step 7, 100µl of sodium citrate (0.67mg solid) from step 8, 100µl of aprotinin solution (2mg solid) from step 6 and 400µl of phospholipid dispersion (100mg solid) from step 5. Mix contents of each vial gently.

(iii) Preparation of Oil Phase

10 Weigh out 1.5g of POLYSORBATE 80 and 13.5g of AKOLINE MCM into a 20ml glass bottle. Flush vial with nitrogen, screw cap tightly and seal with parafilm and mix gently on roller mixer until a homogenous solution has been obtained.

(iv) Lyophilisation of Aqueous Phase

11 To each vial in step 9 add 1ml of akoline/polysorbate 80 mixture from step 10 using a large volume positive displacement dispenser.

12 Vortex each vial rapidly for ten seconds, and then freeze immediately in liquid nitrogen.

13 Lyophilise for two nights at a condenser temperature of less than -40°C and a vacuum of less than 0.1 mBar.

(v) Preparation of Solution in Oil

14 Remove each of the vials from the lyophiliser, flush with nitrogen, cap tightly and seal with parafilm. Incubate with gentle shaking at 37°C until a clear solution is obtained and store at +4°C until required for use.

(vi) Administration *in vivo*

15 Warm each vial to 37°C in a water bath in order to melt the oil and form a clear solution.

16 To each vial add 2ml of warm PBS, vortex for 30 seconds and administer the contents of one whole vial to one pig i.j. as described above.

EXAMPLE 4: PREPARATION OF FORMULATION CONTAINING INSULIN/URSODEOXYHOLATE COMPLEX

(i) Preparation of Urso Deoxycholate Solution

1 Weigh out 525mg of SODIUM URSO DEOXY CHOLATE in a glass conical flask and add DISTILLED WATER to give a final weight of 15g. Flush with nitrogen, stopper well and seal with parafilm and mix gently on orbital shaker until all the solid has dissolved.

(ii) Preparation of Protein Solution

2 Weigh out 30mg of APROTININ in a 2ml glass screw-capped vial and add 1.5ml of DISTILLED WATER. Screw cap tightly and mix gently until all the solid is dissolved.

3 Weigh out 110mg of INSULIN into a 25ml conical flask and add all 15ml of ursodeoxycholate solution (525g solid) from step 1. Cover the flask with parafilm and mix gently on orbital shaker at 37°C until all the solid is dissolved.

4 Weigh out 13.5mg of SODIUM CITRATE in a 2ml glass screw-capped vial and add 1.5ml of DISTILLED WATER. Screw cap tightly and mix gently until all the solid is dissolved.

5 To each of twelve glass screw-capped 7ml vials dispense 1ml of insulin/chenodeoxycholate solution (7.3mg insulin/ 35mg bile salt) from step 3, 100µl of sodium citrate (0.9mg solid) from step 4, and 100µl of aprotinin solution (2mg solid) from step 2. Mix contents of each vial gently.

(iii) Preparation of Oil Phase

6 Weigh out 1.5g of POLYSORBATE 80 and 13.5g of AKOLINE MCM into a 20ml glass bottle. Flush vial with nitrogen, screw cap tightly and seal with parafilm and mix gently on roller mixer until a homogenous solution has been obtained.

(iv) Lyophilisation of Aqueous Phase

7 To each vial in step 5 add 1ml of akoline/polysorbate 80 mixture from step 6 using a large volume positive displacement dispenser.

8 Vortex each vial rapidly for ten seconds. and then freeze immediately in liquid

nitrogen.

9 Lyophilise for two nights at a condenser temperature of less than -40°C and a vacuum of less than 0.1 mBar.

(v) Preparation of Solution in Oil

10 Remove each of the vials from the lyophiliser, flush with nitrogen, cap tightly and seal with parafilm. Incubate for ten minutes at 37°C until a clear solution is obtained.

11 Store at +4°C until required for use.

(vi) Administration *in vivo*

12 Warm each vial to 37°C in a water bath in order to melt the oil and form a clear solution.

13 To each vial add 2ml of warm PBS, vortex for 30 seconds and administer the contents of one whole vial to one pig i.j. as described above.

EXAMPLE 5: FORMULATIONS FOR INTESTINAL DELIVERY OF CALCITONIN

Formulations were prepared as follows:

1) ***Mixture of Salmon Calcitonin with Akoline MCM.***

As in Example 1, except that (a) the phospholipid dispersion was omitted (b)

the protein-containing lyophilate was not dissolved in oil phase. (c) prior to administration to animals the protein lyophilate was dissolved in 4ml phosphate-buffered saline and (d) the resulting solution was added to 2ml of the oil phase (Akoline MCM/Tween 80) and dispersed by vortexing.

2) ***Mixture of Salmon Calcitonin with Glycerol Di-Oleate***

As for formulation (1) except that glycerol di-oleate was employed in place of Akoline MCM.

3) ***Mixture Salmon Calcitonin with Oleic Acid.***

As for formulation (1) except that oleic acid was employed in phase of Akoline MCM.

4) ***Oil Containing Salmon Calcitonin/PC complex in Akoline MCM.***

As described in example 1.

5) ***Oil Containing Salmon Calcitonin/Chenodeoxycholate complex in Akoline MCM.***

Formulation identical in composition to that in formulation (4) of this Example, except that sodium chenodeoxycholate was employed instead of PC at a concentration of 100 mg/g in the oil phase.

6) ***Oil Containing Salmon Calcitonin/Chenodeoxycholate complex in Akoline MCM.***

Formulation identical in composition to that in formulation (4) of this Example, except that aprotinin was omitted, and sodium chenodeoxycholate was employed instead of PC at a concentration of 17.5 mg/g in the oil phase.

7) ***Oil Containing Salmon Calcitonin/Ursodeoxycholate complex in Akoline MCM.***

Formulation identical in composition to that in (4) above, except that aprotinin was omitted, and sodium ursodeoxycholate was employed instead of PC at a concentration of 17.5 mg/g in the oil phase.

8) ***Oil Containing Salmon Calcitonin/Tocopherol succinate complex in Akoline***

Formulation identical in composition to that in (4) above, except that aprotinin was omitted, and alpha-tocopherol succinate was employed instead of PC at a concentration of 25 mg/g in the oil phase.

9) ***Oil Containing Salmon Calcitonin/Sodium docusate complex in Akoline MCM***

Formulation identical in composition to that in (4)above, except that aprotinin was omitted, and sodium docusate was employed instead of PC at a concentration of 25 mg/g in the oil phase.

EXAMPLE 6: FORMULATIONS FOR INTESTINAL DELIVERY OF INSULIN

Formulations were prepared as follows:

10) ***Mixture of Insulin with Akoline MCM.***

As for formulation (1) except that 7.3mg of insulin (200iu), 2mg of aprotinin, and 1 ml of Akoline/Tween 80 oil phase employed.

11) ***Oil Containing Insulin/PC Complex in Akoline MCM.***

As in Example 2.

12) ***Oil Containing Insulin/ChenodeoxycholateComplex in Akoline MCM at High Bile Salt Concentration.***

As in Example 3.

13) ***Oil Containing Insulin/ChenodeoxycholateComplex in Akoline MCM at Medium Bile Salt Concentration.***

As in Example 3.

14) ***Oil Containing Insulin/ChenodeoxycholateComplex in Akoline MCM at Low Bile Salt Concentration.***

As in Example 3.

15) ***Oil Containing Insulin/Ursodeoxycholate Complex in Akoline MCM (with aprotinin).***

As in Example 4.

16) ***Oil Containing Insulin/Ursodeoxycholate complex in Akoline MCM (without aprotinin).***

Exactly as in Example 4 except that aprotinin omitted from the preparation.

17) ***Solution of Insulin/Urso in Phosphate-Buffered Saline.***

Lyophilised solution of insulin prepared as in step (3) of Example 4. Redissolved in Phosphate-Buffered Saline (7.3mg insulin/2ml) before administration.

EXAMPLE 7: ANIMAL STUDIES

ANIMAL STUDIES

Preparation of Animal Model

The animal model employed for testing oral formulations of calcitonin is the juvenile pig. The pig is selected because it has similar weight to man and the structure, function and size of the small intestine are similar to the human small intestine. In order to eliminate concerns relating to differences between human and porcine stomachs, the animals were surgically manipulated so that material could be introduced directly into the jejunum via an in-dwelling cannula. This also reduces the uncertainty usually associated with oral administration as to time of arrival of the dose in the intestine, and leads to improvement in the quality of the statistics obtained.

At least 10 days before the first dosing, a fine plastic cannula was surgically inserted into the jejunum of the pig and then brought out under the skin onto the back of the pig so that the test materials can be injected into the jejunum without distressing the

animal. An indwelling catheter which was also brought out through the back skin was inserted into the aorta via the medium saphenous artery so that repeated blood samples could be obtained. A second catheter was also introduced in a carotid artery.

Administration of Formulations

Pigs were tested while fully conscious in the fasting state and peptide formulations were administered via the oral route after three baseline blood samples were taken. A single experiment usually occupied a period of 8 to 9 hours and the surgically-prepared pigs participated in tests up to three times per week over a four week period. On completion of the tests the pigs were killed and the intestines examined for macroscopic and microscopic changes.

During the study water was provided ad lib and the pigs fed at 8:00 and 15:30. Any food remaining after the 15:30 feed was removed at 16:30. On the days of treatment, the 08:00 food was withheld and 3/4 of the total daily requirement was given after the last blood sample was taken.

On the days of treatment, the dosing solutions were given by instillation via the in-dwelling cannula into the jejunum. The cannula was flushed through before dosing with 1ml warm (25-30°C) sterile phosphate-buffered saline, and flushed through after dosing with 5 ml of the same material at the same temperature. Calcitonin incorporated in the lipidic delivery system was given as a dispersion in which 2ml of oil was vortexed in 4ml of warm phosphate buffered saline, and 1.3ml of the resultant dispersion was administered to each animal. Each animal received 5000iu of calcitonin. Insulin formulations were also given as dispersions in which 1ml of oil containing 200iu insulin was dispersed in 2ml of warm phosphate saline before all three millilitres were administered i.j. to a single animal.

Sample Collection in Animals

The catheters used for collection of blood were flushed once daily with heparinised saline (500iu/ml). On sampling days a less concentrated heparin solution (50iu/ml) was used in the catheters between sampling.

At 09:00 on the study days a blood sample not exceeding 5ml was taken, after withdrawing 2ml of blood to remove any residual anticoagulant from the catheter. The catheter was then flushed with 50iu/ml heparin solution to prevent clotting in the cannula. The animal was then dosed according to the treatment schedule. Blood sampling and dosing was staggered to allow a number of animals to be handled at the same time. Blood samples, totalling 4ml at each time point, were taken at 0.25., 0.5., 1.0., 1.5., 2.0., 3.0., 4.0., and 6.0. hours after dosing as for the 09:00 (time zero) samples.

Method of Assay

The blood samples were handled as follows:-

After the discard, the first 2ml was taken into a plastic heparinised container. The samples were then stored at +4°C for up to 30 minutes before they were centrifuged in a refrigerated centrifuge at 3000 rpm for 20 minutes. The resultant plasma was then divided and transferred into two suitable containers and frozen to -20°C prior to colorimetric assay of plasma calcium or glucaose concentrations using a Kone autoanalyser. One sample was assayed and the other retained.

RESULTS

Tables 1 and 2 below present the results of the animal studies in terms of AUC of fall in either calcium or glucose levels, as well as peak fall in calcium or glucose level.

Table 1: Results for Calcitonin formulations as described in Example 5

Formulation details Calcitonin administered at a dose of 5000 iu per animal	AUC of Ca fall (4 hours)	Peak fall in Ca mmol/L
1. sCT/aprotinin+Akoline/tween 80 mixture in PBS	-2.19 ± 0.97	-0.74 ± 0.31
2. sCT/aprotinin+GDO/Tween 80 mixture in PBS	-0.6 ± 0.39	-0.33 ± 0.26
3. sCT/aprotinin+Oleic acid/Tween 80 mixture in PBS	-0.5 ± 0.39	-0.44 ± 0.04
4. sCT/aprotinin||PC||Akoline|| Tween 80	-0.82 ± 0.89	-0.39 ± 0.10
5. sCT/Aprotinin||Cheno||Akoline|| Tween 80	-1.85 ± 1.48	-0.63 ± 0.06
6. sCT/aprotinin||Cheno||oleic acid|| Tween 80	-0.63 ± 0.38	-0.24 ± 0.18
7. sCT||Cheno||Akoline||Tween 80	-1.73 ± 1.20	-0.60 ± 0.36
8. sCT||Urso||Akoline||Tween 80	-2.26 ± 1.31	-0.73 ± 0.44
9. sCT||αTS||Akoline||Tween 80	-2.22 ± 1.04	-0.73 ± 0.32
10.sCT||Docusate||Akoline||Tween 80	-1.55 ± 0.83	-0.54 ± 0.25

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Table 2: Results for Insulin formulations as described in Example 6

Formulation details Insulin administered at a dose of 200iu/animal	AUC of fall in glucose (4hr)	Peak fall in glucose mmol/L
11. Insulin/aprotinin+Akoline/Tween 80 mixture in PBS	-3.78 ± 2.65	-2.34 ± 1.79
12. Insulin/aprotinin||PC||Akoline||Tween 80	-2.84 ± 2.97	-1.45 ± 1.54
13. Insulin/aprotinin||Cheno(high)||Akoline||Tween 80	-2.02 ± 1.44	-1.94 ± 1.17
14. Insulin/aprotinin||Cheno(med)||Akoline||Tween 80	-2.93 ± 1.58	-1.96 ± 1.17
15. Insulin/aprotinin||Cheno(low)||Akoline||Tween 80	-2.98 ± 2.88	-2.18 ± 1.68
16. Insulin/aprotinin||Urso(low)||Akoline||Tween 80	-3.66 ± 2.17	-1.85 ± 1.29
17. Insulin alone||Urso(low)||Akoline||Tween 80	-5.69 ± 2.32	-2.75 ± 0.59
18. Insulin/Urso in PBS	-2.09 ± 1.23	-1.03 ± 0.93
19. Negative control (non Insulin)	-1.58 ± 1.03	N/A
11-18.	-1.69	-
17-18.	-3.60	

In Table 2, formulating insulin as a hydrophobic solution in medium-chain monoglyceride (17) gives a greater efficacy than a simple two-phase mixture of components (11). This is more noticeable in the AUC of fall of glucose, rather than the peak glucose fall, indicating that a longer duration of action is an important factor in improvement of efficacy. Subtraction of the AUC value of group (18) from (11) and (17) gives the contribution made by the delivery vehicle over and above the effect of insulin administered in free form. As can be seen, the insulin formulated as a solution in medium-chain monoglyceride gives over double the enhancement afforded by a simple mixture of the components.



Claims

1. A hydrophobic preparation comprising:

(i) an oil phase comprising one or more medium chain monoglycerides;

(ii) at least one amphiphile; and

(iii) a hydrophilic species solubilised or otherwise dispersed in the one or more glycerides;

wherein the hydrophilic species is one that is not normally soluble in the one or more glycerides.

2. A hydrophobic preparation as claimed in claim 1 wherein the oil phase comprises a mixture of medium chain mono- and diglycerides.

3. A hydrophobic preparation as claimed in claim 2 wherein the medium chain glycerides have chain lengths of 8 to 10 carbon atoms.

4. A hydrophobic preparation as claimed in claim 2 or claim 3 wherein the oil phase further comprises an additional compound such as oleic acid, glycerol mono-oleate or gelucires.

5. A hydrophobic preparation as claimed in claim 4 wherein the oil phase comprises 40-90% monoglycerides.

6. A hydrophobic preparation as claimed in claim 5 wherein the oil phase comprises 60-70% monoglycerides.

7. A hydrophobic preparation as claimed in any one of claims 1 to 6 wherein the oil phase comprises Akoline MCM.

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8. A hydrophobic preparation as claimed in any one of claims 1 to 7 wherein the ratio of amphiphile: hydrophilic species is in the range 1:1 to 20:1 by weight.

9. A hydrophobic preparation as claimed in claim 8 wherein the ratio of amphiphile: hydrophilic species 1, 2:1 to 8:1 by weight.

10. A hydrophobic preparation as claimed in any one of claims 1 to 9 wherein the amphiphile is phosphatidyl choline, phosphatidic acid, phosphatidyl glycerol, phosphatidyl ethanolamine, phosphatidyl serine or lyso derivatives thereof, octyl glucoside or other glycolipids, tocopherol succinate and cholesterol hemisuccinate, sodium docusate or hydroxypropyl cellulose.

11. A hydrophobic preparation as claimed in any one of claims 1 to 9 wherein the amphiphile is a bile salt.

12. A hydrophobic preparation as claimed in claim 11 wherein the amphiphile is a bile salt comprising cholate, deoxycholate, chenodeoxycholate, ursodeoxycholate, taurocholate, taurodeoxycholate, tauroursodeoxycholate, taurochenodeoxycholate, glycholate, glycodeoxycholate, glycoursodeoxycholate, glycochenodeoxycholate, lithocholate, taurolithocholate or glycolithocholate.

13. A hydrophobic preparation as claimed in any one of claims 1 to 12 wherein the hydrophilic species is a hydrophilic macromolecule.

14. A hydrophobic preparation as claimed in claim 13 wherein the hydrophilic macromolecule comprises proteins, glycoproteins, oligo and/or polynucleic acids, for example DNA or RNA, and analogues thereof, polysaccharides such as heparin, or supramolecular assemblies thereof.

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15. A hydrophobic preparation as claimed in claim 13 or claim 14 wherein a small molecule is co-solubilised with the hydrophilic macromolecule.

16. A hydrophobic preparation as claimed in claim 15 wherein the small molecule is a polysaccharide such as cyclodextrins or vitamin B12.

17. A hydrophobic preparation as claimed in any one of claims 13 to 16 wherein the macromolecule is a protein.

18. A hydrophobic preparation as claimed in claim 17 wherein the protein is insulin, calcitonin, haemoglobin, soy bean trypsin inhibitor, aprotinin, GLP1, erythropoietin, somatotropin, growth hormone, growth hormone releasing factor, galanin, urokinase, blood factors such as Factor VIII and Factor IX, tissue plasminogen activator, superoxide dismutase, catalase, peroxidase, interferon, somatostatin, hirudin, LHRU or an analogue or fragment thereof.

19. A hydrophobic preparation as claimed in any one of claims 1 to 12 wherein the hydrophilic species is a small organic molecule a small inorganic molecule or a colloidal substance.

20. A hydrophobic preparation as claimed in claim 19 wherein the hydrophilic species is glucose, carboxyfluorescin, an anti-cancer agent, vitamins, calcium chloride, sodium phosphate or a colloidal metal such as colloidal gold, palladium, platinum or rhodium.

21. A hydrophobic preparation as claimed in any one of claims 1 to 20 which further comprises one or more further components selected from antioxidants, metal chelating agents, buffering agents and dispersion agents.

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22. An emulsion preparation comprising a hydrophobic preparation as defined in any one of claims 1 to 21.

23. A pharmaceutical formulation comprising a hydrophobic preparation as defined in any one of claims 1 to 21.

24. The use of a hydrophobic preparation as defined in any one of claims 1 to 21 in the preparation of a medicament for oral delivery of a hydrophilic species.

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