

DISPERSION AND DETACHMENT OF CELL AGGREGATES

CROSS-REFERENCE TO RELATED PATENT APPLICATIONS

This application claims priority to U.S.A. Priority Application 61/491,756, filed 05/31/2011, and is incorporated herein by reference in its entirety.

FIELD OF THE INVENTION

The present invention relates to compositions and methods for dispersing or detaching a single cell organism from a surface or from other cells or single cell organisms, especially where the organism is in a biofilm or *in vivo*.

BACKGROUND OF THE INVENTION

Microorganisms can live and proliferate as individual cells swimming freely in the environment (e.g., plankton), or they can grow as highly organized, multicellular communities encased in a self-produced polymeric matrix in close association with surfaces and interfaces. The latter microbial lifestyle is referred to as biofilms. Biofilm formation represents an ancient, protected mode of growth that allows microbial survival in hostile environments and allows microorganisms to disperse and colonize new niches [Hall-Stoodley et al., Nat Rev Microbiol. (2004) 2(2):95-108]. The composition of biofilms is complex and variable among different microbial species and even within the same species under different environmental conditions. Nonetheless, biofilm formation represents the normal lifestyle of microorganism in the environment and all microbes can make biofilms. Previous studies revealed that bacterial biofilm formation progresses through multiple developmental stages differing in protein profiles [Sauer et al., J Bacteriol. (2002) 184(4): 1140-54], beginning with attachment to surface, followed by the immigration and division to form microcolonies and finally maturation involving expression of matrix polymers. Bacteria within each biofilm stage display phenotypes and possess properties that are markedly different from those of the same group growing planktonically [Sauer et al., J Bacteriol. (2004) 186(21):7312-26]. Biofilms are a major cause of systemic infections (e.g., nosocomial infections) in humans.

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In the body, biofilms can be associated with tissues (e.g., inner ears, teeth, gums, lungs, heart valves and the urogenital tract) and can be a major source of systemic infections. An estimated 65 % of bacterial infections in humans are biofilm in nature. Additionally, after forming biofilms, microorganisms tend to change their characteristics, sometimes drastically, such that doses of antibiotics which normally kill the organisms in suspended cultures are completely ineffective against the same microorganisms when the organisms are in attached or conglomerate biofilm form. See U.S. Pat. No. 7,189,351, incorporated by reference in its entirety.

One of the principal concerns with respect to products that are introduced into the body (e.g., contact lenses, central venous catheters, mechanical heart valves and pacemakers) or provide a pathway into the body is microbial infection and invariably biofilm formation. As these infections are difficult to treat with antibiotics, removal of the device is often necessitated, which is traumatic to the patient and increases the medical cost. PCT Application No. WO 06/006172 discloses the use of anti-amyloid agents, such as aromatic compounds, for inhibiting formation or disintegrating a pre existing biofilm. The application discloses that compounds preventing amyloid fibril formation in Alzheimers can act against fibril formation in biofilms, and concludes that amino acids having an aromatic arm are effective against biofilms. However, the analysis was limited to full length sequences.

Biofilms can cause, amongst a wide range of negative effects, accelerated corrosion in industrial systems, oil souring and biofouling. Bacterial aggregation can occur in agriculture

[Monier et al., Applied and Environmental Microbiology, 70(1): 346-355 (2004); Biofilms in the food and beverage industries, Edited by P M Fratamico, B A Annous and N W Guenther, USDA ARS, USA, Woodhead Publishing Series in Food Science, Technology and Nutrition No. 181, Chapter 20, pages 517-535] and in water systems [Carlson et al., Zentralbl Bakteriell Orig B, 161(3): 233-247 (1975)]. Biofouling may be caused by the adhesion of organisms to any surface in a marine or freshwater environment, including cooling towers, water pipes and filters in cooling or desalinization installations, irrigation and power stations, and membranes, such as those used in wastewater and desalinization systems. Biofouling also occurs in aquaculture systems in fish farms. Furthermore the commercial shipping fleets of the world consume approximately 300 million tons of fuel annually. Without antifouling measures, that fuel consumption would increase by as much as 40%, equivalent to an extra 120 million tonnes of fuel annually. The economic cost of this was estimated as about \$7.5 billion in 2000; a more recent estimate is \$30 billion. Generally, biofilms are very difficult to eliminate since microbes growing within are highly organized and can withstand hostile environments, such as high temperatures and anti-microbial agents (e.g., antibiotics).

Since marine-aquatic plants and animals are continuously exposed to a large diversity and abundance of potentially harmful microorganisms in the form of biofilm, and it is known that marine life produce anti-microbial peptides, it is possible that broad spectrum natural factors that interfere with biofilm formation may also be present in marine life.

U.S. Publication No. 20070098745 discloses means of preventing biofilm formation by the use of reef fish microflora. This invention describes anti-biofilm substances derived from bacteria isolated from the epithelial mucosal surfaces of healthy coral reef fish (e.g., *Sparisoma nigrum* and *Lutjanus purpureus*). The bacterial isolates produce signals or toxins that prevent biofilm formation.

Cell clustering is not limited to microbial biofilms, but can exist *in vivo*. Alzheimer's Disease, for example, involves neuron clusters (i.e., neuritic plaques) in the brain [Tiraboschi et al., J. Neurology, 62(11): 1984-1989 (2004)]. In the body, bacterial aggregation can occur orally [Duffau et al., 16 September 2005 RAI Congress, #0299; Liljemark et al., Infect. Immun., 31(3): 935-941 (1981)]; in sepsis [Reid et al., Current Microbiology, 20(3): 185-190 (1990)]; diarrhea [Bieber et al., Science, 280(5372): 2114-2118 (1998)]; in nosocomial

infections [Bortz et al., Bulletin of Mathematical Biology, Volume 70, Number 3, 745-768] ; in relation to drug efficacy [Kraal et al., J Dent Res 58(11): 2125-2131 (1979)]; in relation to peritoneal dialysis [Reid et al., Peritoneal Dialysis International, 10: 21-24 (1990)]; lung diseases [Sanchez et al., PLoS Pathog 6(8)]; and Crohn's disease [Isenmann et al., Digestive Diseases and Sciences, 47(2): 462-468 (2002)].

Cell clustering can also occur among white blood cells *in vivo*. For example, white blood cells can aggregate in whole blood as the result of cigarette smoking and lead to microvascular occlusion and damage [Hill et al., J. R. Soc. Med., 86(3):139-140 (1993)]. White blood cell aggregation can also occur in vascular disease [Belch et al., Thrombosis Research, 48(6):631-639 (1987)]. Macrophage-lymphocyte clustering is correlated to rheumatoid arthritis [Webb et al., Macrophage-lymphocyte clustering in rheumatoid arthritis, Ann. rheum. Dis. (1975), 34, 38] Additionally, Sun et al. state, "Both platelet aggregation and white blood cell aggregation are involved in pathological processes such as thrombosis, atherosclerosis and chronic inflammation. People in older age groups are likely to suffer from cardiovascular diseases and may have increased white cell and platelet aggregation which could contribute to this increased risk." [Sun et al., A study of whole blood platelet and white cell aggregation using a laser flow aggregometer, Platelets (2003) Mar 14(2):103-8.] Furthermore, adhesion and aggregation of white blood cells are involved in vascular diseases and thrombosis [Belch et al., Whole blood white cell aggregation: a novel technique, Thrombosis Research, 48; 631-639 (1987)].

Cell clustering also occurs in restenosis, which can develop as the result of implanted medical stents [Dangas et al., Circulation, 105:2586 (2005)]. Such clustering can lead to the occlusion of a blood vessel and dramatically reduced blood flow. One of the symptoms of the second stage of restenosis, which tends to occur 3–6 months after surgery, is platelet aggregation at the site of the injury [Michael Kirchengast*, Klaus Munter. Endothelin and restenosis. Cardiovascular Research 39 (1998) 550–555] and residual plaque burden outside the stent [Prati et al., In-Stent Neointimal Proliferation Correlates With the Amount of Residual Plaque Burden Outside the Stent. An Intravascular Ultrasound Study, Circulation, (1999) 99:1011-1014.], both phenomena being the main causes of in-stent neointimal proliferation. Patri et al concludes with the following: "Late in-stent neointimal proliferation

has a direct correlation with the amount of residual plaque burden after coronary stent implantation, supporting the hypothesis that plaque removal before stent implantation may reduce restenosis."

SUMMARY OF THE INVENTION

The present invention provides a peptide consisting of amino acids X^1 - X^2 - X^3 - X^4 - X^5 - X^6 - X^7 - X^8 - X^9 - X^{10} - X^{11} - X^{12} , wherein X^1 is S, N, I, V, R, K, Q or L; X^2 is V, I, A, N, L or Q; X^3 is P; X^4 is Y, F or W; X^5 is D, N, Q or E; X^6 is Y, F, R, W, V, H, L, K or I; X^7 is N, S, G, D, H, E, Q or I; X^8 is W, L, F, M, S, T, R, A, G, V, P, Y, I or K; X^9 is Y, N, F, K, L, R, I, V, W or Q; X^{10} is S, K, N, T, E, R, L, Q, I, V, D, or K; X^{11} is N, E, D, Q, S, A or I; and X^{12} is W, R, V, L, I, K, F or E, wherein the peptide is not SVPYDYNWYSNW. The present invention also provides a peptide consisting of amino acids X^1 - X^2 - X^3 - X^4 - X^5 - X^6 - X^7 - X^8 - X^9 - X^{10} - X^{11} - X^{12} , wherein X^1 is S, N, T, K, R, H, E, I, Q or D; X^2 is V, I, L, Y, G, F or W; X^3 is H, N, Q, E, D or S; X^4 is S, P, A or T; X^5 is F, W or Y; X^6 is D, N, E or Q; X^7 is Y, F or W; X^8 is D, G or E; X^9 is W, F or Y; X^{10} is Y, F or W; X^{11} is N or Q; and X^{12} is V, I or L, wherein the peptide is not SVHSFDYDWYNV.

In some embodiments, the peptide is SVPFDYNLYSNW; SAPYNFNFYSNW; NIPFNFSLNKR; SVPYQYNWYSNW; SVPWEYNFYSNW; RPYDRGMIVNV; KVPYDWDSVINL; QLPYDVHTYNDW; LAPYDHNRYTQW; SNPYDLEAYENW; SVPYDYQGYRNI; SVPYDYNVYLNK; IQPYDKNYFQNF; VVPYDINIKDNW; SVPYDYNPYSNW; SVPYDYNKLKNW; SVPYDYNWRSSW; SVPYDYNWWSAW; SVPYDYNWQSNW; ELSSFNFDWYNV; RYSSFDYDWYNV; NVHSFDYDWYNV; RVESFNFDWYNV; RVESFDYDWYNI; RINSFDYDWYNV; TVNSFDYDWYNV; KVNSFDYDWYNV; TVHSFDYDWYNV; SVHSWDYDWYNV; SVHSYDFDWYNV; TLQAFNYEWYQL; KYETFEYGWYNI; HGDSFQYEWYNL; SVHSFDWDWYNV; SVHSFDYDYNNV; SVHSFDYDFYNNV; SVHSFDYDWFNNV; SVHSFDYDWWNV; IFNPFDYDWYNV; QWHSFDYDWYNV or DVHPFDYDWYNV.

In some embodiments, the peptide is cyclic peptide. In other embodiments, the peptide is soluble. In some embodiments, the peptide is attached to a linker. In some

embodiments, the linker is polyethylene glycol or palmitic acid. In other embodiments, the peptide is synthetic.

The present invention also provides a composition comprising a protein or peptide, wherein said composition is capable of detaching a single cell organism from a surface or from other single cell organisms. In some embodiments, the peptide consists of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, I, V, R, K, Q or L; X^2 is V, I, A, N, L or Q; X^3 is P; X^4 is Y, F or W; X^5 is D, N, Q or E; X^6 is Y, F, R, W, V, H, L, K or I; X^7 is N, S, G, D, H, E, Q or I; X^8 is W, L, F, M, S, T, R, A, G, V, P, Y, I or K; X^9 is Y, N, F, K, L, R, I, V, W or Q; X^{10} is S, K, N, T, E, R, L, Q, I, V, D, or K; X^{11} is N, E, D, Q, S, A or I; and X^{12} is W, R, V, L, I, K, F or E, wherein the peptide is not SVPYDYNWYSNW. In other embodiments, the peptide consists of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, T, K, R, H, E, I, Q or D; X^2 is V, I, L, Y, G, F or W; X^3 is H, N, Q, E, D or S; X^4 is S, P, A or T; X^5 is F, W or Y; X^6 is D, N, E or Q; X^7 is Y, F or W; X^8 is D, G or E; X^9 is W, F or Y; X^{10} is Y, F or W; X^{11} is N or Q; and X^{12} is V, I or L, wherein the peptide is not SVHSFDYDWYNV.

In specific embodiments, the peptide is SVPFDYNLYSNW; SAPYNFNFYSNW; NIPFNFSLNKER; SVPYQYNWYSNW; SVPWEYNFYSNW; RIPPYDRGMIVNV; KVPYDWDSVINL; QLPYDVHTYNDW; LAPYDHNRYTQW; SNPYDLEAYENW; SVPYDYQGYRNI; SVPYDYNVYLNK; IQPYDKNYFQNF; VVPYDINIKDNW; SVPYDYNPYSNW; SVPYDYNKLKNW; SVPYDYNWRSSW; SVPYDYNWWSAW; SVPYDYNWQSNW; ELSSFNFDWYNV; RYSSFDYDWYNV; NVHSFDYDWYNV; RVESFNFDWYNV; RVESFDYDWYNI; RINSFDYDWYNV; TVNSFDYDWYNV; KVNSFDYDWYNV; TVHSFDYDWYNV; SVHSWDYDWYNV; SVHSYDFDWYNV; TLQAFNYEWYQL; KYETFEYGWYNI; HGDSFQYEWYNL; SVHSFDWDWYNV; SVHSFDYDYNNV; SVHSFDYDFYNNV; SVHSFDYDWFNNV; SVHSFDYDWWNNV; IFNPFYDWYNV; QWHSFDYDWYNV or DVHPFDYDWYNV.

In some embodiments, peptide is a cyclic peptide. In some embodiments, the peptide is soluble. In other embodiments, the peptide is attached to a linker. In some embodiments, the linker is polyethylene glycol or palmitic acid. In some embodiments, the peptide is

synthetic. In some embodiments, the organism is in a biofilm. In some embodiments, the organism is an aquatic microorganism. In some embodiments, the organisms are attached in a cluster or aggregate. In some embodiments, the composition is capable of breaking or dispersing said cluster or aggregate. In some embodiments, the detaching affects the ability of said organism to produce polysaccharide matrix. In some embodiments, the surface is a selected from the group comprising a fabric, a fiber, a foam, a film, a concrete, a masonry, a glass, a metal and a plastic.

The present invention also provides a method of detaching a single cell organism from a surface or from other single cell organisms, comprising contacting said organism with a composition comprising a protein or peptide. In some embodiments, the peptide consists of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, I, V, R, K, Q or L; X^2 is V, I, A, N, L or Q; X^3 is P; X^4 is Y, F or W; X^5 is D, N, Q or E; X^6 is Y, F, R, W, V, H, L, K or I; X^7 is N, S, G, D, H, E, Q or I; X^8 is W, L, F, M, S, T, R, A, G, V, P, Y, I or K; X^9 is Y, N, F, K, L, R, I, V, W or Q; X^{10} is S, K, N, T, E, R, L, Q, I, V, D, or K; X^{11} is N, E, D, Q, S, A or I; and X^{12} is W, R, V, L, I, K, F or E, wherein the peptide is not SVPYDYNWYSNW. In other embodiments, the peptide consists of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, T, K, R, H, E, I, Q or D; X^2 is V, I, L, Y, G, F or W; X^3 is H, N, Q, E, D or S; X^4 is S, P, A or T; X^5 is F, W or Y; X^6 is D, N, E or Q; X^7 is Y, F or W; X^8 is D, G or E; X^9 is W, F or Y; X^{10} is Y, F or W; X^{11} is N or Q; and X^{12} is V, I or L, wherein the peptide is not SVHSFDYDWYNV.

In specific embodiments, the peptide is SVPFDYNLYSNW; SAPYNFNFYSNW; NIPFNFSLNKER; SVPYQYNWYSNW; SVPWEYNFYSNW; RIPPYDRGMIVNV; KVPYDWDSVINL; QLPYDVHTYNDW; LAPYDHNRYTQW; SNPYDLEAYENW; SVPYDYQGYRNI; SVPYDYNVYLNK; IQPYDKNYFQNF; VVPYDINIKDNW; SVPYDYNPYSNW; SVPYDYNKLKNW; SVPYDYNWRSSW; SVPYDYNWWSAW; SVPYDYNWQSNW; ELSSFNFDWYNV; RYSSFDYDWYNV; NVHSFDYDWYNV; RVESFNFDWYNV; RVESFDYDWYNI; RINSFDYDWYNV; TVNSFDYDWYNV; KVNSFDYDWYNV; TVHSFDYDWYNV; SVHSWDYDWYNV; SVHSYDFDYNV; TLQAFNYEWYQL; KYETFEYGWYNI; HGDSFQYEWYNL; SVHSFDWDWYNV;

SVHSFDYDYNNV; SVHSFDYDFYNNV; SVHSFDYDWFNNV; SVHSFDYDWWNNV;
IFNPFDYDWYNNV; QWHSFDYDWYNNV or DVHPFDYDWYNNV.

In some embodiments, the peptide is a cyclic peptide. In some embodiments, the peptide is soluble. In other embodiments, the peptide is attached to a linker. In some embodiments, the linker is polyethylene glycol or palmitic acid. In some embodiments, the peptide is synthetic. In some embodiments, the surface is selected from the group comprising a fabric, a fiber, a foam, a film, a concrete, a masonry, a glass, a metal and a plastic. In some embodiments, the organism is in a biofilm. In some embodiments, the organism is an aquatic microorganism. In some embodiments, the organisms are attached in a cluster or aggregate. In some embodiments, composition breaks or disperses said cluster or aggregate. In some embodiments, the composition prevents said organism from producing polysaccharide matrix. In some embodiments, the surface is selected from the group comprising a fabric, a fiber, a foam, a film, a concrete, a masonry, a glass, a metal and a plastic.

The present invention also provides a pharmaceutical composition comprising a protein or peptide, wherein the composition is capable of detaching a single cell organism from a surface or from other single cell organisms, and a pharmaceutically acceptable carrier or diluent. In some embodiments, the pharmaceutical composition comprises a peptide consisting of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, I, V, R, K, Q or L; X^2 is V, I, A, N, L or Q; X^3 is P; X^4 is Y, F or W; X^5 is D, N, Q or E; X^6 is Y, F, R, W, V, H, L, K or I; X^7 is N, S, G, D, H, E, Q or I; X^8 is W, L, F, M, S, T, R, A, G, V, P, Y, I or K; X^9 is Y, N, F, K, L, R, I, V, W or Q; X^{10} is S, K, N, T, E, R, L, Q, I, V, D, or K; X^{11} is N, E, D, Q, S, A or I; and X^{12} is W, R, V, L, I, K, F or E, wherein the peptide is not SVPYDYNWYSNW. In other embodiments, the peptide consists of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, T, K, R, H, E, I, Q or D; X^2 is V, I, L, Y, G, F or W; X^3 is H, N, Q, E, D or S; X^4 is S, P, A or T; X^5 is F, W or Y; X^6 is D, N, E or Q; X^7 is Y, F or W; X^8 is D, G or E; X^9 is W, F or Y; X^{10} is Y, F or W; X^{11} is N or Q; and X^{12} is V, I or L, wherein the peptide is not SVHSFDYDWYNNV.

In specific embodiments, the peptide is SVPFDYNLYSNW; SAPYNFNFYSNW; NIPFNFSLNKER; SVPYQYNWYSNW; SVPWEYNFYSNW; RIPPYDRGMIVNV; KVPYDWDSVINL; QLPYDVHTYNDW; LAPYDHNRYTQW; SNPYDLEAYENW; SVPYDYQGYRNI; SVPYDYNVYLNK; IQPYDKNYFQNF; VVPYDINIKDNW; SVPYDYNPYSNW; SVPYDYNKLKNW; SVPYDYNWRSSW; SVPYDYNWWSAW; SVPYDYNWQSNW; ELSSFNFDWYNV; RYSSFDYDWYNV; NVHSFDYDWYNV; RVESFNFDWYNV; RVESFDYDWYNI; RINSFDYDWYNV; TVNSFDYDWYNV; KVNSFDYDWYNV; TVHSFDYDWYNV; SVHSWDYDWYNV; SVHSYDFDYNV; TLQAFNYEWYQL; KYETFEYGWYNI; HGDSFQYEWYNL; SVHSFDYDWYNV; SVHSFDYDYNNV; SVHSFDYDFYNNV; SVHSFDYDWFNV; SVHSFDYDWWNV; IFNPFYDWYNV; QWHSFDYDWYNV or DVHPFDYDWYNV.

The present invention also provides a method of preventing or treating a pathogen infection in a subject in need thereof, the method comprising administering to the subject a therapeutically effective amount of a pharmaceutical composition comprising a protein or peptide, wherein said composition is capable of detaching a single cell organism from a surface or from other single cell organisms. In some embodiments, the pharmaceutical composition comprises a peptide consisting of amino acids X^1 - X^2 - X^3 - X^4 - X^5 - X^6 - X^7 - X^8 - X^9 - X^{10} - X^{11} - X^{12} , wherein X^1 is S, N, I, V, R, K, Q or L; X^2 is V, I, A, N, L or Q; X^3 is P; X^4 is Y, F or W; X^5 is D, N, Q or E; X^6 is Y, F, R, W, V, H, L, K or I; X^7 is N, S, G, D, H, E, Q or I; X^8 is W, L, F, M, S, T, R, A, G, V, P, Y, I or K; X^9 is Y, N, F, K, L, R, I, V, W or Q; X^{10} is S, K, N, T, E, R, L, Q, I, V, D, or K; X^{11} is N, E, D, Q, S, A or I; and X^{12} is W, R, V, L, I, K, F or E, wherein the peptide is not SVPYDYNWYSNW. In other embodiments, the peptide consists of amino acids X^1 - X^2 - X^3 - X^4 - X^5 - X^6 - X^7 - X^8 - X^9 - X^{10} - X^{11} - X^{12} , wherein X^1 is S, N, T, K, R, H, E, I, Q or D; X^2 is V, I, L, Y, G, F or W; X^3 is H, N, Q, E, D or S; X^4 is S, P, A or T; X^5 is F, W or Y; X^6 is D, N, E or Q; X^7 is Y, F or W; X^8 is D, G or E; X^9 is W, F or Y; X^{10} is Y, F or W; X^{11} is N or Q; and X^{12} is V, I or L, wherein the peptide is not SVHSFDYDWYNV.

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SVPYDYNPYSNW; SVPYDYNKLKNW; SVPYDYNWRSSW; SVPYDYNWWSAW;
 SVPYDYNWQSNW; ELSSFNFDWYNV; RYSSFDYDWYNV; NVHSFDYDWYNV;
 RVESFNYDWYNV; RVESFDFDWYNI; RINSFDYDWYNV; TVNSFDYDWYNV;
 KVNSFDYDWYNV; TVHSFDYDWYNV; SVHSWDYDWYNV; SVHSYDFDWYNV;
 TLQAFNYEWYQL; KYETFEYGWYNI; HGDSFQYEWYNL; SVHSFDWDWYNV;
 SVHSFDYDYNNV; SVHSFDYDFYNNV; SVHSFDYDWFNNV; SVHSFDYDWWNNV;
 IFNPFYDWYNV; QWHSFDYDWYNV or DVHPFDYDWYNV.

The present invention also provides a method of increasing the effectiveness of a pharmaceutical composition, the method comprising administering a composition comprising a protein or peptide, wherein the composition is capable of detaching a microorganism from a surface or from other microorganisms to a subject in need of the pharmaceutical composition. In some embodiments, the pharmaceutical composition comprises a peptide consisting of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, I, V, R, K, Q or L; X^2 is V, I, A, N, L or Q; X^3 is P; X^4 is Y, F or W; X^5 is D, N, Q or E; X^6 is Y, F, R, W, V, H, L, K or I; X^7 is N, S, G, D, H, E, Q or I; X^8 is W, L, F, M, S, T, R, A, G, V, P, Y, I or K; X^9 is Y, N, F, K, L, R, I, V, W or Q; X^{10} is S, K, N, T, E, R, L, Q, I, V, D, or K; X^{11} is N, E, D, Q, S, A or I; and X^{12} is W, R, V, L, I, K, F or E, wherein the peptide is not SVPYDYNWYSNW. In other embodiments, the peptide consists of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, T, K, R, H, E, I, Q or D; X^2 is V, I, L, Y, G, F or W; X^3 is H, N, Q, E, D or S; X^4 is S, P, A or T; X^5 is F, W or Y; X^6 is D, N, E or Q; X^7 is Y, F or W; X^8 is D, G or E; X^9 is W, F or Y; X^{10} is Y, F or W; X^{11} is N or Q; and X^{12} is V, I or L, wherein the peptide is not SVHSFDYDWYNV.

In specific embodiments, the peptide is SVPFDYNLYSNW; SAPYNFNFYSNW;
 NIPFNFLNKR; SVPYQYNWYSNW; SVPWEYNFYSNW; RPYDRGMIVNV;
 KVPYDWDSVINL; QLPYDVHTYNDW; LAPYDHNRYTQW; SNPYDLEAYENW;
 SVPYDYQGYRNI; SVPYDYNVYLNK; IQPYDKNYFQNF; VVPYDINIKDNW;
 SVPYDYNPYSNW; SVPYDYNKLKNW; SVPYDYNWRSSW; SVPYDYNWWSAW;
 SVPYDYNWQSNW; ELSSFNFDWYNV; RYSSFDYDWYNV; NVHSFDYDWYNV;
 RVESFNYDWYNV; RVESFDFDWYNI; RINSFDYDWYNV; TVNSFDYDWYNV;
 KVNSFDYDWYNV; TVHSFDYDWYNV; SVHSWDYDWYNV; SVHSYDFDWYNV;

TLQAFNYEWYQL; KYETFEYGWYNI; HGDSFQYEWYNL; SVHSFDWDWYNV;
SVHSFDYDYNNV; SVHSFDYDFYNV; SVHSFDYDWFNV; SVHSFDYDWWNV;
IFNPFYDWYNV; QWHSFDYDWYNV or DVHPFDYDWYNV.

In some embodiments, the pharmaceutical composition is an antibiotic.

The present invention also provides a method of identifying an anti-biofilm composition, the method comprising: (a) contacting said biofilm with a plurality of compositions, each composition comprising a protein or peptide; (b) assaying the ability of said biofilm to resist anti-biofilm activity, wherein said anti-biofilm activity comprises detaching said biofilm from a surface or breaking up said biofilm; and (c) identifying from said plurality of compositions at least one composition having said anti-biofilm activity above a predetermined threshold, thereby identifying the anti-biofilm composition. In some embodiments, the composition comprises a peptide consisting of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, I, V, R, K, Q or L; X^2 is V, I, A, N, L or Q; X^3 is P; X^4 is Y, F or W; X^5 is D, N, Q or E; X^6 is Y, F, R, W, V, H, L, K or I; X^7 is N, S, G, D, H, E, Q or I; X^8 is W, L, F, M, S, T, R, A, G, V, P, Y, I or K; X^9 is Y, N, F, K, L, R, I, V, W or Q; X^{10} is S, K, N, T, E, R, L, Q, I, V, D, or K; X^{11} is N, E, D, Q, S, A or I; and X^{12} is W, R, V, L, I, K, F or E, wherein the peptide is not SVPYDYNWYSNW. In other embodiments, the peptide consists of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, T, K, R, H, E, I, Q or D; X^2 is V, I, L, Y, G, F or W; X^3 is H, N, Q, E, D or S; X^4 is S, P, A or T; X^5 is F, W or Y; X^6 is D, N, E or Q; X^7 is Y, F or W; X^8 is D, G or E; X^9 is W, F or Y; X^{10} is Y, F or W; X^{11} is N or Q; and X^{12} is V, I or L, wherein the peptide is not SVHSFDYDWYNV.

In specific embodiments, the peptide is SVPFDYNLYSNW; SAPYNFNFYSNW;
NIPFNFLNKR; SVPYQYNWYSNW; SVPWEYNFYSNW; RIPPYDRGMIVNV;
KVPYDWDSVINL; QLPYDVHTYNDW; LAPYDHNRYTQW; SNPYDLEAYENW;
SVPYDYQGYRNI; SVPYDYNVYLNK; IQPYDKNYFQNF; VVPYDINIKDNW;
SVPYDYNPYSNW; SVPYDYNKLKNW; SVPYDYNWRSSW; SVPYDYNWWSAW;
SVPYDYNWQSNW; ELSSFNFDWYNV; RYSSFYDWYNV; NVHSFDYDWYNV;
RVESFNFDWYNV; RVESFDFDWYNI; RINSFDYDWYNV; TVNSFDYDWYNV;

KVNSFDYDWYNNV; TVHSFDYDWYNNV; SVHSWDYDWYNNV; SVHSYDFDWYNNV;
 TLQAFNYEWYQL; KYETFEYGWYNI; HGDSFQYEWYNL; SVHSFDWDWYNNV;
 SVHSFDYDYNNV; SVHSFDYDFYNNV; SVHSFDYDWFNNV; SVHSFDYDWWNNV;
 IFNPFDYDWYNNV; QWHSFDYDWYNNV or DVHPFDYDWYNNV.

The present invention also provides a medical device comprising a composition comprising a protein or peptide, wherein the composition is capable of detaching a single cell organism from a surface or from other single cell organisms. In some embodiments, the medical device comprises a peptide consisting of amino acids X^1 - X^2 - X^3 - X^4 - X^5 - X^6 - X^7 - X^8 - X^9 - X^{10} - X^{11} - X^{12} , wherein X^1 is S, N, I, V, R, K, Q or L; X^2 is V, I, A, N, L or Q; X^3 is P; X^4 is Y, F or W; X^5 is D, N, Q or E; X^6 is Y, F, R, W, V, H, L, K or I; X^7 is N, S, G, D, H, E, Q or I; X^8 is W, L, F, M, S, T, R, A, G, V, P, Y, I or K; X^9 is Y, N, F, K, L, R, I, V, W or Q; X^{10} is S, K, N, T, E, R, L, Q, I, V, D, or K; X^{11} is N, E, D, Q, S, A or I; and X^{12} is W, R, V, L, I, K, F or E, wherein the peptide is not SVPYDYNWYSNW. In other embodiments, the peptide consists of amino acids X^1 - X^2 - X^3 - X^4 - X^5 - X^6 - X^7 - X^8 - X^9 - X^{10} - X^{11} - X^{12} , wherein X^1 is S, N, T, K, R, H, E, I, Q or D; X^2 is V, I, L, Y, G, F or W; X^3 is H, N, Q, E, D or S; X^4 is S, P, A or T; X^5 is F, W or Y; X^6 is D, N, E or Q; X^7 is Y, F or W; X^8 is D, G or E; X^9 is W, F or Y; X^{10} is Y, F or W; X^{11} is N or Q; and X^{12} is V, I or L, wherein the peptide is not SVHSFDYDWYNNV.

In specific embodiments, the peptide is SVPFDYNLYSNW; SAPYNFNFYSNW;
 NIPFNFSLNKER; SVPYQYNWYSNW; SVPWEYNFYSNW; RIPPYDRGMIVNV;
 KVPYDWDSVINL; QLPYDVHTYNDW; LAPYDHNRYTQW; SNPYDLEAYENW;
 SVPYDYQGYRNI; SVPYDYNVYLNK; IQPYDKNYFQNF; VVPYDINIKDNW;
 SVPYDYNPYSNW; SVPYDYNKLKNW; SVPYDYNWRSSW; SVPYDYNWWSAW;
 SVPYDYNWQSNW; ELSSFNFDWYNNV; RYSSFDYDWYNNV; NVHSFDYDWYNNV;
 RVESFNFDWYNNV; RVESFDYDWYNNV; RINSFDYDWYNNV; TVNSFDYDWYNNV;
 KVNSFDYDWYNNV; TVHSFDYDWYNNV; SVHSWDYDWYNNV; SVHSYDFDWYNNV;
 TLQAFNYEWYQL; KYETFEYGWYNI; HGDSFQYEWYNL; SVHSFDWDWYNNV;
 SVHSFDYDYNNV; SVHSFDYDFYNNV; SVHSFDYDWFNNV; SVHSFDYDWWNNV;
 IFNPFDYDWYNNV; QWHSFDYDWYNNV or DVHPFDYDWYNNV.

The present invention also provides a method of dispersing a biofilm or detaching biofilm formation from a surface, the method comprising treating water with or coating said surface with a composition comprising a protein or peptide, wherein the composition is capable of detaching a single cell organism from a surface or from other single cell organisms. In some embodiments, the method comprises a peptide consisting of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, I, V, R, K, Q or L; X^2 is V, I, A, N, L or Q; X^3 is P; X^4 is Y, F or W; X^5 is D, N, Q or E; X^6 is Y, F, R, W, V, H, L, K or I; X^7 is N, S, G, D, H, E, Q or I; X^8 is W, L, F, M, S, T, R, A, G, V, P, Y, I or K; X^9 is Y, N, F, K, L, R, I, V, W or Q; X^{10} is S, K, N, T, E, R, L, Q, I, V, D, or K; X^{11} is N, E, D, Q, S, A or I; and X^{12} is W, R, V, L, I, K, F or E, wherein the peptide is not SVPYDYNWYSNW. In other embodiments, the peptide consists of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, T, K, R, H, E, I, Q or D; X^2 is V, I, L, Y, G, F or W; X^3 is H, N, Q, E, D or S; X^4 is S, P, A or T; X^5 is F, W or Y; X^6 is D, N, E or Q; X^7 is Y, F or W; X^8 is D, G or E; X^9 is W, F or Y; X^{10} is Y, F or W; X^{11} is N or Q; and X^{12} is V, I or L, wherein the peptide is not SVHSFDYDWYNV.

In specific embodiments, the peptide is SVPFDYNLYSNW; SAPYNFNFYSNW; NIPFNFLNKER; SVPYQYNWYSNW; SVPWEYNFYSNW; RIPPYDRGMIVNV; KVPYDWDSVINL; QLPYDVHTYNDW; LAPYDHNRYTQW; SNPYDLEAYENW; SVPYDYQGYRNI; SVPYDYNVYLNK; IQPYDKNYFQNF; VVPYDINIKDNW; SVPYDYNPYSNW; SVPYDYNKLKNW; SVPYDYNWRSSW; SVPYDYNWWSAW; SVPYDYNWQSNW; ELSSFNFDWYNV; RYSSFDYDWYNV; NVHSFDYDWYNV; RVESFNFDWYNV; RVESFDYDWYNI; RINSFDYDWYNV; TVNSFDYDWYNV; KVNSFDYDWYNV; TVHSFDYDWYNV; SVHSWDYDWYNV; SVHSYDFDWYNV; TLQAFNYEWYQL; KYETFEYGWYNI; HGDSFQYEWYNL; SVHSFDWDWYNV; SVHSFDYDYNNV; SVHSFDYDFYNNV; SVHSFDYDWFNNV; SVHSFDYDWWNNV; IFNPFYDWYNV; QWHSFDYDWYNV or DVHPFDYDWYNV.

The present invention also provides a method of treating a disease, comprising administering a peptide consisting of amino acids $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, I, V, R, K, Q or L; X^2 is V, I, A, N, L or Q; X^3 is P; X^4 is Y, F or W; X^5 is D, N, Q or E; X^6 is Y, F, R, W, V, H, L, K or I; X^7 is N, S, G, D, H, E, Q or I; X^8 is W,

L, F, M S, T, R, A, G, V, P, Y, I or K; X⁹ is Y, N, F, K, L, R, I, V, W or Q; X¹⁰ is S, K, N, T, E, R, L, Q, I, V, D, or K; X¹¹ is N, E, D, Q, S, A or I; and X¹² is W, R, V, L, I, K, F or E, wherein the peptide is not SVPYDYNWYSNW. In other embodiments, the peptide consists of amino acids X¹-X²-X³-X⁴-X⁵-X⁶-X⁷-X⁸-X⁹-X¹⁰-X¹¹-X¹², wherein X¹ is S, N, T, K, R, H, E, I, Q or D; X² is V, I, L, Y, G, F or W; X³ is H, N, Q, E, D or S; X⁴ is S, P, A or T; X⁵ is F, W or Y; X⁶ is D, N, E or Q; X⁷ is Y, F or W; X⁸ is D, G or E; X⁹ is W, F or Y; X¹⁰ is Y, F or W; X¹¹ is N or Q; and X¹² is V, I or L, wherein the peptide is not SVHSFDYDWYNV.

In specific embodiments, the peptide is SVPFDYNLYSNW; SAPYNFNFYSNW; NIPFNFSLNKER; SVPYQYNWYSNW; SVPWEYNFYSNW; RPYDRGMIVNV; KVPYDWDSVINL; QLPYDVHTYNDW; LAPYDHNRYTQW; SNPYDLEAYENW; SVPYDYQGYRNI; SVPYDYNVYLNK; IQPYDKNYFQNF; VVPYDINIKDNW; SVPYDYNPYSNW; SVPYDYNKLKNW; SVPYDYNWRSSW; SVPYDYNWWSAW; SVPYDYNWQSNW; ELSSFNFDWYNV; RYSSFDYDWYNV; NVHSFDYDWYNV; RVESFNFDWYNV; RVESFDYDWYNI; RINSFDYDWYNV; TVNSFDYDWYNV; KVNSFDYDWYNV; TVHSFDYDWYNV; SVHSWDYDWYNV; SVHSYDFDYNV; TLQAFNYEWYQL; KYETFEYGWYNI; HGDSFQYEWYNL; SVHSFDYDWYNV; SVHSFDYDYNNV; SVHSFDYDFYNV; SVHSFDYDWFNV; SVHSFDYDWWNV; IFNPFYDWYNV; QWHSFDYDWYNV or DVHPFDYDWYNV.

In some embodiments, the disease is autoimmune, inflammatory or degenerative disease. In some embodiments, the disease is Alzheimer's Disease.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows the detachment of *Pseudomonas aeruginosa* biofilm from a well in a 96-well culture plate after incubating the microorganism overnight to create the biofilm and subsequently incubating the biofilm with Eqt2Z-cyc peptide overnight.

FIG. 2A shows the detachment of *Pseudomonas aeruginosa* biofilm from a well after incubating the microorganism for 2 hours to create the biofilm and subsequently incubating the biofilm with different concentrations of Eqt2Z-cyc peptide and Abac10 negative control overnight.

FIG. 2B shows the detachment of *Pseudomonas aeruginosa* biofilm from a well after incubating the microorganism for 2 hours to create the biofilm and subsequently incubating the biofilm with different concentrations of Eqt2Z-cyc peptide and Abac10 negative control for 24 hours.

FIG. 3 shows the detachment of *Pseudomonas aeruginosa* biofilm from a well after incubating the microorganism for 2 hours to create the biofilm and subsequently incubating the biofilm with grZ14s-nvcyc peptide overnight.

FIG. 4 shows the detachment of *Pseudomonas aeruginosa* biofilm from a well after incubating the microorganism for 24 hours to create the biofilm and subsequently incubating the biofilm with Eqt2-cyc peptide for 24 hours.

FIG. 5 shows the detachment of *Pseudomonas aeruginosa* biofilm from a well after incubating the microorganism for 24 hours to create the biofilm and subsequently incubating the biofilm with grZ14s-nvcyc peptide for 24 hours.

FIG. 6 shows the detachment of *Pseudomonas aeruginosa* biofilm from a well after incubating the microorganism for 24 hours to create the biofilm and subsequently incubating the biofilm with Physco-cyc peptide overnight.

FIG. 7 shows the enhancement of Imipenem activity with grZ14s-nvCyc.

FIG. 8 shows the enhancement of Ampicillin activity with grZ14s-nvCyc.

FIG. 9 shows the enhancement of Vancomycin activity with grZ14s-nvCyc.

FIG. 10 shows the enhancement of Amphotericin activity with grZ14s-nvCyc.

FIG. 11 shows the enhancement of Fluconazole activity with grZ14s-nvCyc

FIG. 12 shows the enhancement of Kanamycin activity with grZ14s-nvCyc

FIG. 13A shows the prevention of *Pseudomonas aeruginosa* adherence by various peptides.

FIG. 13B shows the detachment of *Pseudomonas aeruginosa* adherence by various peptides.

FIG. 14A shows the prevention of *Staphylococcus aureus* adherence by various peptides.

FIG. 14B shows the detachment of *Staphylococcus aureus* adherence by various peptides.

FIG. 15A shows the prevention of *Candida albicans* adherence by various peptides.

FIG. 15B shows the detachment of *Candida albicans* adherence by various peptides.

FIG. 16A shows the prevention of *Escherichia coli* adherence by various peptides.

FIG. 16B shows the detachment of *Escherichia coli* adherence by various peptides.

FIG. 17 shows the prevention of adhesion of *Pseudomonas aeruginosa* by various modifications of Eqt2Z-Cyc.

FIG. 18 shows Congo Red staining of Eqt2Z-Cyc incubated with *Pseudomonas aeruginosa*.

FIG. 19 shows Congo Red staining of grZ14s-nvCyc incubated with *Pseudomonas aeruginosa*.

FIG. 20 shows Trypan Blue staining of grZ14s-nvCyc incubated with *Pseudomonas aeruginosa*.

FIG. 21 shows Congo Red staining of grZ14s-nvCyc incubated with *Candida albicans*.

FIG. 22 shows Congo Red staining of Eqt2Z-Cyc incubated with *Candida albicans*.

FIG. 23 shows Congo Red staining of grZ14s-nvCyc incubated with *Staphylococcus aureus*.

DETAIL DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention relates to compositions and methods comprising a protein or a peptide which has one or more effects relating to detaching a single cell organism from a surface or other single cell organisms, especially where the organism is in a biofilm.

Generally, the nomenclature used herein and the laboratory procedures utilized in the present invention include molecular, biochemical, microbiological and recombinant DNA techniques. Such techniques are thoroughly explained in the literature. See, for example, "Molecular Cloning: A laboratory Manual" Sambrook et al., (1989); "Current Protocols in Molecular Biology" Volumes I-III Ausubel, R. M., ed. (1994); Ausubel et al., "Current Protocols in Molecular Biology", John Wiley and Sons, Baltimore, Maryland (1989); Perbal, "A Practical Guide to Molecular Cloning", John Wiley & Sons, New York (1988); Watson et al., "Recombinant DNA", Scientific American Books, New York; Birren et al. (eds) "Genome Analysis: A Laboratory Manual Series", Vols. 1-4, Cold Spring Harbor Laboratory Press, New York (1998); methodologies as set forth in U.S. Pat. Nos. 4,666,828; 4,683,202; 4,801,531; 5,192,659 and 5,272,057; "Cell Biology: A Laboratory Handbook", Volumes I-III Cellis, J. E., ed. (1994); "Current Protocols in Immunology" Volumes I-III Coligan J. E., ed. (1994); Stites et al. (eds), "Basic and Clinical Immunology" (8th Edition), Appleton & Lange, Norwalk, CT (1994); Mishell and Shiigi (eds), "Selected Methods in Cellular Immunology", W. H. Freeman and Co., New York (1980); available immunoassays are extensively described in the patent and scientific literature, see, for example, U.S. Pat. Nos. 3,791,932; 3,839,153; 3,850,752; 3,850,578; 3,853,987; 3,867,517; 3,879,262; 3,901,654; 3,935,074; 3,984,533; 3,996,345; 4,034,074; 4,098,876; 4,879,219; 5,011,771 and 5,281,521; "Oligonucleotide Synthesis" Gait, M. J., ed. (1984); "Nucleic Acid Hybridization" Hames, B. D., and Higgins S. J., eds. (1985); "Transcription and Translation" Hames, B. D., and Higgins S. J., Eds. (1984); "Animal Cell Culture" Freshney, R. I., ed. (1986); "Immobilized Cells and Enzymes" IRL Press, (1986); "A Practical Guide to Molecular Cloning" Perbal, B., (1984) and "Methods in Enzymology" Vol. 1-317, Academic Press; "PCR Protocols: A Guide To Methods And Applications", Academic Press, San Diego, CA (1990); Marshak et al., "Strategies for Protein Purification and Characterization - A Laboratory Course Manual" CSHL Press (1996); all of which are incorporated by reference as if fully set forth herein.

Other general references are provided throughout this document. The procedures therein are provided for the convenience of the reader. All the information contained therein is incorporated herein by reference.

DEFINITIONS

As used herein, the term "isolated" refers to a composition that has been removed from its in vivo location. Preferably the isolated compositions of the present invention are substantially free from other substances (e.g., other proteins that do not comprise anti-adhesive effects) that are present in their in vivo location (i.e., purified or semi-purified). Isolated proteins and peptides may optionally be synthetic or obtained from natural sources, including optionally by being expressed in vivo using genetic engineering techniques.

As used herein the phrase "single cell organism" refers to a unicellular organism also termed a microorganism or a microbe. The single cell organism of the present invention can be a eukaryotic single cell organism (e.g., protozoa or fungi for example yeast) or a prokaryotic single cell organism (e.g., bacteria or archaea). The single cell organisms of the present invention may be in any cellular environment, such as for example, in a biofilm, as isolated cells or as a cell suspension.

The term "Gram-positive bacteria" as used herein refers to bacteria characterized by having as part of their cell wall structure peptidoglycan as well as polysaccharides and/or teichoic acids and are characterized by their blue-violet color reaction in the Gram-staining procedure. Representative Gram-positive bacteria include: *Actinomyces* spp., *Bacillus anthracis*, *Bifidobacterium* spp., *Clostridium botulinum*, *Clostridium perfringens*, *Clostridium* spp., *Clostridium tetani*, *Corynebacterium diphtheriae*, *Corynebacterium jeikeium*, *Enterococcus faecalis*, *Enterococcus faecium*, *Erysipelothrix rhusiopathiae*, *Eubacterium* spp., *Gardnerella vaginalis*, *Gemella morbillorum*, *Leuconostoc* spp., *Mycobacterium abscessus*, *Mycobacterium avium* complex, *Mycobacterium chelonae*, *Mycobacterium fortuitum*, *Mycobacterium haemophilum*, *Mycobacterium kansasii*, *Mycobacterium leprae*, *Mycobacterium marinum*, *Mycobacterium scrofulaceum*, *Mycobacterium smegmatis*, *Mycobacterium terrae*, *Mycobacterium tuberculosis*, *Mycobacterium ulcerans*, *Nocardia* spp., *Peptococcus niger*, *Peptostreptococcus* spp., *Propionibacterium* spp., *Sarcina lutea*,

Staphylococcus aureus, *Staphylococcus auricularis*, *Staphylococcus capitis*, *Staphylococcus cohnii*, *Staphylococcus epidermidis*, *Staphylococcus haemolyticus*, *Staphylococcus hominis*, *Staphylococcus lugdunensis*, *Staphylococcus saccharolyticus*, *Staphylococcus saprophyticus*, *Staphylococcus schleiferi*, *Staphylococcus similans*, *Staphylococcus warneri*, *Staphylococcus xylosum*, *Streptococcus agalactiae* (group B streptococcus), *Streptococcus anginosus*, *Streptococcus bovis*, *Streptococcus canis*, *Streptococcus equi*, *Streptococcus milleri*, *Streptococcus mitior*, *Streptococcus mutans*, *Streptococcus pneumoniae*, *Streptococcus pyogenes* (group A streptococcus), *Streptococcus salivarius*, *Streptococcus sanguis*.

The term "Gram-negative bacteria" as used herein refer to bacteria characterized by the presence of a double membrane surrounding each bacterial cell. Representative Gram-negative bacteria include *Acinetobacter calcoaceticus*, *Acinetobacter baumannii*, *Actinobacillus actinomycetemcomitans*, *Aeromonas hydrophila*, *Alcaligenes xylosoxidans*, *Bacteroides*, *Bacteroides fragilis*, *Bartonella bacilliformis*, *Bordetella* spp., *Borrelia burgdorferi*, *Branhamella catarrhalis*, *Brucella* spp., *Campylobacter* spp., *Chlamydia pneumoniae*, *Chlamydia psittaci*, *Chlamydia trachomatis*, *Chromobacterium violaceum*, *Citrobacter* spp., *Eikenella corrodens*, *Enterobacter aerogenes*, *Escherichia coli*, *Flavobacterium meningosepticum*, *Fusobacterium* spp., *Haemophilus influenzae*, *Haemophilus* spp., *Helicobacter pylori*, *Klebsiella pneumoniae*, *Klebsiella* spp., *Legionella* spp., *Leptospira* spp., *Moraxella catarrhalis*, *Morganella morganii*, *Mycoplasma pneumoniae*, *Neisseria gonorrhoeae*, *Neisseria meningitidis*, *Pasteurella multocida*, *Plesiomonas shigelloides*, *Prevotella* spp., *Proteus* spp., *Providencia rettgeri*, *Pseudomonas aeruginosa*, *Pseudomonas* spp., *Rickettsia prowazekii*, *Rickettsia rickettsii*, *Rochalimaea* spp., *Salmonella* spp., *Salmonella typhi*, *Serratia marcescens*, *Shigella* spp., *Shigella sonnei*, *Treponema carateum*, *Treponema pallidum*, *Treponema pallidum endemicum*, *Treponema pertenue*, *Veillonella* spp., *Vibrio cholerae*, *Vibrio vulnificus*, *Yersinia enterocolitica*, *Yersinia pestis*.

The term "fungi" as used herein refers to the heterotrophic organisms characterized by the presence of a chitinous cell wall, and in the majority of species, filamentous growth as multicellular hyphae. Representative fungi whose adhesion may be prevented according to the method of the present invention include *Candida albicans*, *Saccharomyces cerevisiae*, *Candida glabrata*, *Candida parapsilosis* and *Candida dubliniensis*.

As used herein the term "biofilm" refers to an extracellular matrix in which single cell organisms can be dispersed and/or form colonies. The biofilm typically is made of polysaccharides and other macromolecules.

As used herein, the term "detach" refers to removing a single cell organism, *in vitro* or *in vivo*, from a surface to which the cell is adhered (e.g., by reducing the rate of growth on a surface) or removing the cell from other single cell organisms to which they are adhered. Preferably, the compositions of the present invention detach cells from adherence by as much as 10 %, more preferably by 20 %, more preferably by 30 %, more preferably by 40 %, more preferably by 50 %, more preferably by 60 %, more preferably by 70 %, more preferably by 80 %, more preferably by 90 % and most preferably by 100% as measured by an adhesion assay. Exemplary detachment assays are described herein below and in the Examples section that follows.

"Detachment" of a biofilm occurs when a single or cluster of cell organisms in the biofilm detaches from a surface and "dispersion" of a biofilm occurs when single cell organisms in a biofilm detach from each other.

As used herein the term "contacting" refers to the positioning of the compositions of the present invention such that they are in direct or indirect contact with the adhesive single cell organisms in such a way that the active agent comprised within is able to detach of cells therefrom. Thus, the present invention contemplates both applying the compositions of the present invention to a desirable surface and/or directly to the adhesive cells. Contacting the compositions with a surface can be effected using any method known in the art including spraying, spreading, wetting, immersing, dipping, painting, ultrasonic welding, welding, bonding or adhering. The compositions of the present invention may be attached as monolayers or multiple layers.

A "peptidomimetic organic moiety" can optionally be substituted for amino acid residues in a peptide according to the present invention both as conservative and as non-conservative substitutions. These moieties are also termed "non-natural amino acids" and may optionally replace amino acid residues, amino acids or act as spacer groups within the peptides in lieu of deleted amino acids. The peptidomimetic organic moieties optionally and

preferably have steric, electronic or configurational properties similar to the replaced amino acid and such peptidomimetics are used to replace amino acids in the essential positions, and are considered conservative substitutions. However such similarities are not necessarily required. The only restriction on the use of peptidomimetics is that the composition at least substantially retains its physiological activity as compared to the native peptide according to the present invention.

As used herein the term "chemical modification", when referring to a peptide according to the present invention, refers to a peptide where at least one of its amino acid residues is modified either by natural processes, such as processing or other post-translational modifications, or by chemical modification techniques which are well known in the art. Examples of the numerous known modifications typically include, but are not limited to: acetylation, acylation, amidation, ADP-ribosylation, glycosylation, GPI anchor formation, covalent attachment of a lipid or lipid derivative, methylation, myristylation, pegylation, prenylation, phosphorylation, ubiquitination, or any similar process.

As used herein the term "medical device" refers to any implant, instrument, apparatus, implement, machine, device or any other similar or related object (including any component or accessory), which is intended for use in the diagnosis, treatment, cure or prevention of disease or other conditions. Such medical device is intended for use in man or other animals and is anticipated to affect the structure or any function of the body. Such medical device does not achieve its primary intended purposes through chemical action and is not dependent upon being metabolized for the achievement of its primary intended purposes.

As used herein the term "implant" refers to any object intended for placement in a human body that is not a living tissue. The implant may be temporary or permanent. An implant can be an article comprising artificial components, such as catheters or pacemakers. Implants can also include naturally derived objects that have been processed so that their living tissues have been devitalized. As an example, bone grafts that have been processed so that their living cells are removed (acellularized), but so that their shape is retained to serve as a template for ingrowth of bone from a host. As another example, naturally occurring coral can be processed to yield hydroxyapatite preparations that can be applied to the body

for certain orthopedic and dental therapies. The present invention therefore envisions coating medical devices with the compositions of the present invention to prevent microorganism adherence thereto so as to reduce/eliminate any possible cell aggregation and biofilm formation known to occur following implantation. Device-related infections usually result from the introduction of microorganisms, primarily bacteria, during the device insertion or implantation procedure, or from attachment of blood-borne organisms to the newly inserted device and their subsequent propagation on its surface. Coating the medical device with the compositions of the present invention will therefore inhibit biofilm formation of one or more microbial species, will prevent medical device related infections, and consequently will reduce the need of antibiotic treatment or removal of the medical device from the subject.

As used herein the term "anti-biofouling agents" refers to the compounds used to protect underwater surfaces from attaching single cell organisms. These single cell organisms include microorganism such as bacteria and fungi.

Hereinafter, the phrases "physiologically acceptable carrier" and "pharmaceutically acceptable carrier," which may be used interchangeably, refer to a carrier or a diluent that does not cause significant irritation to an organism and does not abrogate the biological activity and properties of the administered compound. An adjuvant is included under these phrases.

As used herein the term "excipient" refers to an inert substance added to a pharmaceutical composition to further facilitate administration of an active ingredient.

As used herein, the phrase "sessile aquatic organism" refers to an aquatic organism which is not freely moving for at least some a part of its life cycle. Aquatic sessile organisms are usually permanently attached to a solid substrate of some kind, such as to a rock or the hull of a ship due to physical anchorage to the substrate, or for any other reason (e.g., stone fish). Exemplary sessile organisms include, but are not limited to, sessile cnidarians such as corals, sea anemones (e.g., *Actinia equine* and *Aiptasia pulchella*), sea pens, aquatic sessile larva (e.g., jellyfish larva), tube dwelling anemones and hydroids (e.g., *Chlorohydra viridissima* and *Hydra vulgaris*). Exemplary fish that may be used according to this aspect of

the present invention are preferably those dwelling in shallow waters or those that hide at the bottom layer of the ocean, sometimes in holes or caves. Such fish include eel and catfish.

As used herein the term "subject in need thereof" refers to a mammal, preferably a human subject.

As used herein the term "treating" refers to curing, reversing, attenuating, alleviating, minimizing, suppressing or halting the deleterious effects of a pathogen infection.

As used herein the phrase "pathogen infection" refers to any medical condition which is caused by a pathogenic organism. Examples of pathogen infections include, but are not limited to, chronic infectious diseases, subacute infectious diseases, acute infectious diseases, viral diseases, bacterial diseases, protozoan diseases, parasitic diseases, fungal diseases, mycoplasma diseases, archaea diseases and prion diseases. Pathogen infection can be caused by an organism capable of growing in or on a biofilm. Examples of pathogen infections caused by microbial biofilms include native valve endocarditis (NVE), otitis media (OM), chronic bacterial prostatitis, cystic fibrosis (CF) and periodontitis. Additional pathogen infections that are not specifically attributed to biofilms include, but are not limited to urinary infections, female genital tract infections and pneumonia. Infections due to implantation of medical devices include vascular catheter infections, arterial prosthetic infections, infections of prosthetic heart valves, prosthetic joint infections, infections of central nervous system shunts, orthopedic implant infections, pacemaker and defibrillator infections, hemodialysis and peritoneal dialysis infections, ocular infections, urinary tract infections, infections of the female genital tract, infections associated with endotracheal intubation and tracheostomy and dental infections.

As used herein the phrase "pathogenic organism" refers to any single cell organism which is capable of causing disease, especially a living microorganism such as a bacteria or fungi. Preferably the pathogenic organism is capable of growing in or on a biofilm. Many common pathogenic organisms exist in mammals (e.g. humans) as biofilms and cause disease. These include, but are not limited to, *Mannheimia haemolytica* and *Pasteurella multocida* (causing pneumonia), *Fusobacterium necrophorum* (causing liver abscess), *Staphylococcus aureus* and *Pseudomonas aeruginosa* (causing wound infections), *Escherichia*

coli and Salmonella spp (causing enteritis), Staphylococcus aureus and Staphylococcus epidermidis (causing OM), and Streptococci sp., Staphylococci sp., Candida, and Aspergillus sp. (causing NVE).

APPLICATIONS

The present invention has many applications. One application is to use a composition comprising the protein or peptide to disperse a biofilm or detach a biofilm from a surface. Another application is to use the composition to combat systemic infections (e.g., nosocomial infections) in patient caused by microbial biofilms *in vivo*. Another application is to use the composition to disperse or detach a biofilm present in other fields, including the food, agriculture, pharmaceutical, paint, water, shipping and engineering industries. Another application is to use the composition to disperse or detach a biofilm where coaggregation – i.e., aggregation of more than one species that create a single cluster – occurs. The foregoing applications are not limiting and other applications are appropriate in which a composition can be used to detach a microorganism from a surface or from other microorganisms. Another application is to use the composition to treat disease, including autoimmune, inflammatory diseases and degenerative diseases such as Alzheimer's Disease. These diseases are caused by cell aggregation or clustering and the use of the inventive peptides to prevent or dissociate cell aggregates can lead to alleviation of the disease. Another application is to use the composition to treat restenosis, including restenosis caused by implanted medical stents. Another application is to use the composition to treat blood cell clustering, including white blood cells.

Exemplary bacterial cells that can be detached from surfaces and from each other according to the method of the present invention include gram positive bacteria and gram negative bacteria.

Exemplary surfaces upon which single cell organisms can attach and which are contemplated by the present invention include fabrics, fibers, foams, films, concretes, masonries, glass, metals, plastics, polymers, and like.

Exemplary devices whose surfaces are susceptible to biofilm formation and which are contemplated by the present invention include, but are not limited to, vessel hulls, automobile surfaces, air plane surfaces, membranes, filters, and industrial equipment. The surface may also be comprised in medical devices, instruments, and implants.

Examples of such medical devices, instruments, and implants include any object that is capable of being implanted temporarily or permanently into a mammalian organism, such as a human. Representative medical devices, instruments, and implants that may be used according to the present invention include, for example, central venous catheters, urinary catheters, endotracheal tubes, mechanical heart valves, pacemakers, vascular grafts, stents and prosthetic joints.

Medical devices that may be coated according to the teachings of the present invention include, but are not limited to, artificial blood vessels, catheters and other devices for the removal or delivery of fluids to patients, artificial hearts, artificial kidneys, orthopedic pins, prosthetic joints, plates and implants; catheters and other tubes (including urological and biliary tubes, endotracheal tubes, peripherally insertable central venous catheters, dialysis catheters, long term tunneled central venous catheters, peripheral venous catheters, short term central venous catheters, arterial catheters, pulmonary catheters, Swan-Ganz catheters, urinary catheters, peritoneal catheters), urinary devices (including long term urinary devices, tissue bonding urinary devices, artificial urinary sphincters, urinary dilators), shunts (including ventricular or arterio-venous shunts); prostheses (including breast implants, penile prostheses, vascular grafting prostheses, aneurysm repair devices, mechanical heart valves, artificial joints, artificial larynxes, otological implants), anastomotic devices, vascular catheter ports, vascular stents, clamps, embolic devices, wound drain tubes, ocular lenses, dental implants, hydrocephalus shunts, pacemakers and implantable defibrillators, needleless connectors, voice prostheses and the like. Another possible application of the compositions of the present invention is the coating of surfaces found in the medical and dental environment. Such surfaces include the inner and outer aspects of various instruments and devices, whether disposable or intended for repeated uses. Such surfaces include the entire spectrum of articles adapted for medical use, including without limitation, scalpels, needles,

scissors and other devices used in invasive surgical, therapeutic or diagnostic procedures; blood filters. Other examples will be readily apparent to practitioners in these arts.

Surfaces found in the medical environment also include the inner and outer aspects of pieces of medical equipment, medical gear worn or carried by personnel in the health care setting. Such surfaces can include surfaces intended as biological barriers to infectious organisms in medical settings, such as gloves, aprons and face shields. Commonly used materials for biological barriers are thermoplastic or polymeric materials such as polyethylene, dacron, nylon, polyesters, polytetrafluoroethylene, polyurethane, latex, silicone and vinyl. Other surfaces can include counter tops and fixtures in areas used for medical procedures or for preparing medical apparatus, tubes and canisters used in respiratory treatments, including the administration of oxygen, of solubilized drugs in nebulizers and of anesthetic agents. Other such surfaces can include handles and cables for medical or dental equipment not intended to be sterile. Additionally, such surfaces can include those non-sterile external surfaces of tubes and other apparatus found in areas where blood or body fluids or other hazardous biomaterials are commonly encountered. The compositions of the present invention can be used on the surface of or within these medical devices to provide long term protection against colonization by single cell organisms and reduce the incidence of device-related infections. These compositions can also be incorporated in combination with an anti-microbial agent (e.g., antibiotic agent) into coatings for medical devices. Such a combination will sufficiently kill or inhibit the initial colonizing bacteria and prevent device-related infections as long as the substance is presented in an inhibitory concentration at the device-microbe interface.

The compositions of the present invention can be directly incorporated into the polymeric matrix of the medical device at the polymer synthesis stage or at the device manufacture stage. The compositions can also be covalently attached to the medical device polymer. These and many other methods of coating medical devices are evident to one of ordinary skill in the art.

Additional surfaces that can be treated according to the teachings of the present invention include the inner and outer aspects of those articles involved in water purification,

water storage and water delivery, and those articles involved in food processing. Thus the present invention envisions coating a solid surface of a food or beverage container to extend the shelf life of its contents.

Surfaces related to health can also include the inner and outer aspects of those household articles involved in providing for nutrition, sanitation or disease prevention. Thus, the compositions of the present invention can be used for removal of disease-causing microorganisms from external surfaces. These can include, for example food processing equipment for home use, materials for infant care, tampons, soap, detergents, health and skincare products, household cleaners and toilet bowls.

The surface may be also be laboratory articles including, but not limited to, microscopic slide, a culturing hood, a Petri dish or any other suitable type of tissue culture vessel or container known in the art.

The inventors of this application also envision the use of the compositions of the present invention as anti-Biofouling agents.

Underwater surfaces include any water immersed surface, including ships'/boats's hulls (i.e., the body or frame of a ship or boat), submergence vehicles, navigational aids, screens, nets, constructions, floating or emplaced offshore platforms (e.g., docks), buoys, signaling equipment and articles which come into contact with sea water or salty water. Other underwater surfaces include structures exposed to sea water including pilings, marine markers, undersea conveyances like cabling and pipes, fishing nets, bulkheads, cooling towers, and any device or structure that operates submerged.

The compositions of the present invention can be incorporated into marine coatings to limit undesirable marine biofouling. Thus, the anti-biofouling agents of the present invention can be formulated so as not to contain toxic materials (such as heavy metals), and still retain their efficacy. The anti-biofouling paint of the present invention may further contain binders(s), pigment(s), solvent(s) and additive(s).

Examples of solvents that may be used include aromatic hydrocarbons such as xylene and toluene; aliphatic hydrocarbons such as hexane and heptane, esters such as ethyl acetate

and butyl acetate; amides such as N-methylpyrrolidone and N,N-dimethylformamide; alcohols such as isopropyl alcohol and butyl alcohol; ethers such as dioxane, THF and diethyl ether; and ketones such as methyl ethyl ketone, methyl isobutyl ketone and methyl isoamyl ketone. The solvents may be used alone or in combination thereof.

Examples of binders that may be used include alkyd resin, acrylic or vinyl emulsions, polyurethane resins, epoxy resins, silicone based resins, acrylic resins, inorganic silicate based resins, vinyl resins, particularly a vinyl chloride/vinyl acetate copolymer, and rosin. Examples of pigments that may be used include titanium dioxide, cuprous oxide, iron oxide, talc, aluminum flakes, mica flakes, ferric oxide, cuprous thiocyanate, zinc oxide, cupric acetate meta-arsenate, zinc chromate, zinc dimethyl dithiocarbamate, zinc ethylene bis(dithiocarbamate) and zinc diethyl dithiocarbamate.

Examples of additives that may be incorporated into the coating composition include dehumidifiers, wetting/dispersing agents, anti-settling agents, anti-skinning agents, drying/curing agents, anti-marring agents and additives ordinarily employed in coating compositions as stabilizers and anti-foaming agents. Additionally, any antibiotic which is relatively insoluble in seawater can be used with an anti-biofouling marine paint.

Methods of preparing marine anti-biofouling paints are explained in detail in U.S. Pat. No. 4,678,512; U.S. Pat. No. 4,286,988; U.S. Pat. No. 4,675,051; U.S. Pat. No. 4,865,909; and U.S. Pat. No. 5,143,545.

The compositions of the present invention may also be used for providing antibacterial properties in cosmetics, to prevent spoiling of the product.

The compositions may further be used to provide an antibacterial effect to the mouth, teeth and gums, such as by incorporation in a toothpaste, mouthwash, or chewing gum. Taken together the present teachings portray a wide range of novel anti-adhesive agents isolated from organisms such as aquatic organisms and moss. The broad spectrum of the detachment effects of these anti-adhesive agents (e.g., removing adhesion of gram positive and gram negative bacteria) together with their ability to effect the initial, vulnerable stages of microbial biofilm formation, makes these agents prime candidates as anti-biofilm agents.

Moreover, the anti-adhesive agents described herein are clonable enabling modifications and mass production thereof. In addition their stability (i.e., resistance to environmental conditions) makes these agents suitable for a diverse array of applications.

It will be appreciated that treatment of infectious diseases according to the present invention may be combined with other treatment methods known in the art (i.e., combination therapy). These include, but are not limited to, antimicrobial agents such as penicillins, cephalosporins, carbapenems, aminoglycosides, macrolides, lincomycins, tetracyclines, chloramphenicol, and griseofulvin. Suitable routes of administration may, for example, include oral, rectal, transmucosal, especially transnasal, intestinal, or parenteral delivery, including intramuscular, subcutaneous, and intramedullary injections, as well as intrathecal, direct intraventricular, intravenous, intraperitoneal, intranasal, or intraocular injections.

Alternately, one may administer the pharmaceutical composition in a local rather than systemic manner, for example, via injection of the pharmaceutical composition directly into a tissue region of a patient.

Pharmaceutical compositions of the present invention may be manufactured by processes well known in the art, e.g., by means of conventional mixing, dissolving, granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping, or lyophilizing processes. Pharmaceutical compositions for use in accordance with the present invention thus may be formulated in conventional manner using one or more physiologically acceptable carriers comprising excipients and auxiliaries, which facilitate processing of the active ingredients into preparations that can be used pharmaceutically. Proper formulation is dependent upon the route of administration chosen.

For injection, the active ingredients of the pharmaceutical composition may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hank's solution, Ringer's solution, or physiological salt buffer. For transmucosal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art.

For topical administration, the compositions of the present invention may be formulated as a gel, a cream, a wash, a rinse or a spray.

For oral administration, the pharmaceutical composition can be formulated readily by combining the active compounds with pharmaceutically acceptable carriers well known in the art. Such carriers enable the pharmaceutical composition to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions, and the like, for oral ingestion by a patient. Pharmacological preparations for oral use can be made using a solid excipient, optionally grinding the resulting mixture, and processing the mixture of granules, after adding suitable auxiliaries as desired, to obtain tablets or dragee cores. Suitable excipients are, in particular, fillers such as sugars, including lactose, sucrose, mannitol, or sorbitol; cellulose preparations such as, for example, maize starch, wheat starch, rice starch, potato starch, gelatin, gum tragacanth, methyl cellulose, hydroxypropylmethyl-cellulose, and sodium carbomethylcellulose; and/or physiologically acceptable polymers such as polyvinylpyrrolidone (PVP). If desired, disintegrating agents, such as cross-linked polyvinyl pyrrolidone, agar, or alginic acid or a salt thereof, such as sodium alginate, may be added.

Dragee cores are provided with suitable coatings. For this purpose, concentrated sugar solutions may be used which may optionally contain gum arabic, talc, polyvinyl pyrrolidone, carbopol gel, polyethylene glycol, titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dyestuffs or pigments may be added to the tablets or dragee coatings for identification or to characterize different combinations of active compound doses. Pharmaceutical compositions that can be used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a plasticizer, such as glycerol or sorbitol. The push-fit capsules may contain the active ingredients in admixture with filler such as lactose, binders such as starches, lubricants such as talc or magnesium stearate, and, optionally, stabilizers. In soft capsules, the active ingredients may be dissolved or suspended in suitable liquids, such as fatty oils, liquid paraffin, or liquid polyethylene glycols. In addition, stabilizers may be added. All formulations for oral administration should be in dosages suitable for the chosen route of administration.

For buccal administration, the compositions may take the form of tablets or lozenges formulated in conventional manner.

For administration by nasal inhalation, the active ingredients for use according to the present invention are conveniently delivered in the form of an aerosol spray presentation from a pressurized pack or a nebulizer with the use of a suitable propellant, e.g., dichlorodifluoromethane, trichlorofluoromethane, dichloro-tetrafluoroethane, or carbon dioxide. In the case of a pressurized aerosol, the dosage may be determined by providing a valve to deliver a metered amount. Capsules and cartridges of, for example, gelatin for use in a dispenser may be formulated containing a powder mix of the compound and a suitable powder base, such as lactose or starch.

The pharmaceutical composition described herein may be formulated for parenteral administration, e.g., by bolus injection or continuous infusion. Formulations for injection may be presented in unit dosage form, e.g., in ampoules or in multidose containers with, optionally, an added preservative. The compositions may be suspensions, solutions, or emulsions in oily or aqueous vehicles, and may contain formulatory agents such as suspending, stabilizing, and/or dispersing agents.

Pharmaceutical compositions for parenteral administration include aqueous solutions of the active preparation in water-soluble form. Additionally, suspensions of the active ingredients may be prepared as appropriate oily or water-based injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters such as ethyl oleate, triglycerides, or liposomes. Aqueous injection suspensions may contain substances that increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran.

Optionally, the suspension may also contain suitable stabilizers or agents that increase the solubility of the active ingredients, to allow for the preparation of highly concentrated solutions. Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle, e.g., a sterile, pyrogen-free, water-based solution, before use.

The pharmaceutical composition of the present invention may also be formulated in rectal compositions such as suppositories or retention enemas, using, for example, conventional suppository bases such as cocoa butter or other glycerides. Pharmaceutical compositions suitable for use in the context of the present invention include compositions wherein the active ingredients are contained in an amount effective to achieve the intended purpose. More specifically, a "therapeutically effective amount" means an amount of active ingredients (e.g., an aquatic organism composition) effective to prevent, alleviate, or ameliorate symptoms of a pathogenic infection (e.g., fever) or prolong the survival of the subject being treated. Determination of a therapeutically effective amount is well within the capability of those skilled in the art, especially in light of the detailed disclosure provided herein.

For any preparation used in the methods of the invention, the dosage or the therapeutically effective amount can be estimated initially from in vitro and cell culture assays. For example, a dose can be formulated in animal models to achieve a desired concentration or titer. Such information can be used to more accurately determine useful doses in humans.

Toxicity and therapeutic efficacy of the active ingredients described herein can be determined by standard pharmaceutical procedures in vitro, in cell cultures or experimental animals. The data obtained from these in vitro and cell culture assays and animal studies can be used in formulating a range of dosage for use in human. The dosage may vary depending upon the dosage form employed and the route of administration utilized. The exact formulation, route of administration, and dosage can be chosen by the individual physician in view of the patient's condition. (See, e.g., Fingl, E. et al. (1975), "The Pharmacological Basis of Therapeutics," Ch. 1, p.1.) Dosage amount and administration intervals may be adjusted individually to provide sufficient plasma or brain levels of the active ingredient to induce or suppress the biological effect (i.e., minimally effective concentration, MEC). The MEC will vary for each preparation, but can be estimated from in vitro data. Dosages necessary to achieve the MEC will depend on individual characteristics and route of administration. Detection assays can be used to determine plasma concentrations.

Depending on the severity and responsiveness of the condition to be treated, dosing can be of a single or a plurality of administrations, with course of treatment lasting from several days to several weeks, or until cure is effected or diminution of the disease state is achieved.

The amount of a composition to be administered will, of course, be dependent on the subject being treated, the severity of the affliction, the manner of administration, the judgment of the prescribing physician, etc.

Compositions of the present invention may, if desired, be presented in a pack or dispenser device, such as an FDA-approved kit, which may contain one or more unit dosage forms containing the active ingredient. The pack may, for example, comprise metal or plastic foil, such as a blister pack. The pack or dispenser device may be accompanied by instructions for administration. The pack or dispenser device may also be accompanied by a notice in a form prescribed by a governmental agency regulating the manufacture, use, or sale of pharmaceuticals, which notice is reflective of approval by the agency of the form of the compositions for human or veterinary administration. Such notice, for example, may include labeling approved by the U.S. Food and Drug Administration for prescription drugs or of an approved product insert. Compositions comprising a preparation of the invention formulated in a ' pharmaceutically acceptable carrier may also be prepared, placed in an appropriate container, and labeled for treatment of an indicated condition, as further detailed above.

Additional objects, advantages, and novel features of the present invention will become apparent to one ordinarily skilled in the art upon examination of the following examples, which are not intended to be limiting. Additionally, each of the various embodiments and aspects of the present invention as delineated hereinabove and as claimed in the claims section below finds experimental support in the following examples.

PROTEINS AND PEPTIDES

The protein or peptide in the compositions can be obtained from different sources. Natural proteins and peptides can be found in animals, including humans. Also, marine and fresh water plants and organisms, including soft bodied water invertebrates, fish and moss produce several factors that can prevent microbial colonization on their body surface since

they lack specific immunity and are surrounded by broad spectrum species of microbial organisms. The most "sensitive" organisms are invertebrates belong to the phylum cnidaria that include the sea anemones, corals, jellyfish, hydroids, medusae, and sea fans. Such soft bodied organism, which lack physical protection such as scales or shells, use proteins and secondary metabolites to protect themselves from the microbial environment surrounding them.

Additionally, it has been previously reported that marine organisms (e.g., sponges) produce secondary metabolites that exhibit antibacterial and antifungal activities [Amade et al., supra]. Moreover, sea anemones (e.g., *Actinia equina*) have been shown to produce toxic, pore forming peptides (i.e., equinatoxins), which lyse and kill eukaryotic cells similarly to other small antimicrobial peptides [Anderluh et al., supra].

Although it is known in the art that the full length sequences of various proteins are related to their cytolytic function, the specific peptides responsible for the cytolytic effect have not been previously identified.

Whilst reducing the present invention to practice, the present inventors discovered that human proteins and aquatic sessile organisms comprise anti-biofilm properties. As is shown in the Examples section which follows, the present inventors have shown that proteins and peptides found in humans and sessile cnidarians (e.g., sea anemones) can cause microorganisms to detach from surfaces and from each other (Figures 1-6). These proteins and peptides were not bactericidal and did not affect bacterial growth.

Taken together the present teachings portray a wide range of novel anti-adhesive agents derived from human and aquatic sessile organisms, in particular from sessile cnidarians. The broad spectrum of the anti-biofilm effects of these agents makes these agents prime candidates as anti-biofilm agents. Moreover, the anti-adhesive agents described herein are clonable enabling modifications and mass production thereof. In addition their stability (i.e., resistance to environmental conditions) makes these agents suitable for a diverse array of applications.

The present inventors have demonstrated that several active fractions obtained from sea anemones using liquid chromatography separations show a high level of prevention of microbial adherence to abiotic surfaces. The sea anemone includes 46 families that can be found in water sources around the world. Most sea anemones are sessile, with a specialized foot used to anchor them in soft substrates, or attach themselves to rocks and corals. The anti-adhesive activity was demonstrated with several species of sea anemone belonging to different genera: *Actinia equine*, *Aiptasia* and *Ammonia*. The N terminus region of anemone cytotoxin has been shown to be involved in the cytotoxic effect [Ref: Kristan K, Podlessek Z, Hojnik V, Gutierrez-Aguirre I, Guncar G, Turk D, Gonzalez-Manas JM, Lakey JH, Macek P, Anderluh G (2004): Pore formation by equinatoxin, an eukaryotic pore-forming toxin, requires a flexible N-terminal region and a stable beta sandwich. *J Biol Chem*. 279(45):46509-46517]. A protein having some resemblance to the C terminus region of anemone cytotoxin superfamily (pfam06369) (a superfamily cluster is a set of conserved domain models, from one or more source databases, that generate overlapping annotation on the same protein sequences. These models are assumed to represent evolutionarily related domains and may be redundant with each other), which region is not involved in cytotoxicity, has also been identified in fish by the present inventors. This protein has a highly conserved region, with unknown function, which is also a Trp-rich domain, and may be important for binding of the protein to lipid membrane. The present inventors have also found this region in the moss *Physcomitrella patens*. The present inventors therefore hypothesized that this region provides a peptide which is highly effective in dispersing biofilms or detaching biofilms from surfaces, while being devoid of cytotoxic activity.

Based on bioinformatics analysis, it is believed that the protein has changed in upper organisms (including in humans), at the highly conserved region, to FDYDWY, that can be found in proteins, represented in the GenBank as GPCR 137b like, with size range from 128aa- 400aa. In humans, this peptide is part of the G protein-coupled receptor 137B (GENE ID: 7107 GPR137B) located at 269-274. Based on UniProtKB/Swiss-Prot entry 060478, the region, which starts at 259 and ends at 292, is an extracellular region, which supports the theory that this peptide is the active part of the protein. Bioinformatics analysis of the ancient sea organism *Ciona intestinalis* identified a 368 amino acid protein, similar to the G protein-coupled receptor 137ba [GeneBank Accession number XP_002125109]. The region similar to

the anti adhesive peptide is SPLRCSELSSFNFDWYNVSDQADLVN. Based on this information, and the fact that *Ciona intestinalis* is also exposed to a large diversity of microorganisms, the peptide FNFDWY is also highly effective in dispersing biofilms or detaching biofilms from surfaces, while being devoid of cytotoxic activity.

The protein or peptide may be natural and isolated from any animal. Preferably, the animal is a vertebrate, such as, for example, a fish, an amphibian (including a frog, a toad, a newt or a salamander), a bird, a reptile (such as a crocodile, a lizard, a snake, a turtle, a tortoise or a terrapin) or a mammal (including a human).

According to some embodiments, the peptide comprises part of a sequence comprising up to about 30, up to about 40, or up to about 50 amino acids.

The peptides of the present invention may optionally comprise at least two sequences, connected by a linker of some type, such that the N-terminal of a first peptide sequence is connected to the C-terminal of the linker, and the C-terminal of a second peptide sequence is connected to the N-terminal of the linker.

The peptides of the present invention can be cyclized (i.e., in cyclic form) and are indicated in this application as such with the term "cyc."

In one embodiment, each peptide is modified with a cysteine at the C-terminal and a cysteine at the N-terminal, wherein the C- and N-terminals are S-S bridged. In specific embodiments, one or more of the peptides are modified with a cysteine at the C-terminal and a cysteine at the N-terminal, wherein the C- and N-terminals are S-S bridged.

Exemplary peptides contain a domain which comprises at least one peptide and which is effective against adhesion of a single cell organism to a surface or to other single cell organisms. More preferably, the domain is included as part of a protein. Optionally and most preferably, the domain exhibits anti-adhesive behavior, for example for the detachment and/or treatment of a biofilm, but does not exhibit cytotoxic behavior.

The peptides may optionally be altered so as to form non-peptide analogs, including but not limited to replacing one or more bonds with less labile bonds, cyclization and the like.

Additionally or alternatively, a peptide may optionally be converted to a small molecule through computer modeling, as described for example in PCT Application No. WO/2007/147098, hereby incorporated by reference as if fully set forth herein.

Peptidomimetics may optionally be used to inhibit degradation of the peptides by enzymatic or other degradative processes. The peptidomimetics can optionally and preferably be produced by organic synthetic techniques. Non-limiting examples of suitable peptidomimetics include D amino acids of the corresponding L amino acids, tetrazol (Zabrocki et al., J. Am. Chem. Soc. 110:5875 5880 (1988)); isosteres of amide bonds (Jones et al., Tetrahedron Lett. 29: 3853 3856 (1988)); LL 3 amino 2 propenidone 6 carboxylic acid (LL Acp) (Kemp et al., J. Org. Chem. 50:5834 5838 (1985)). Similar analogs are shown in Kemp et al., Tetrahedron Lett. 29:5081 5082 (1988) as well as Kemp et al., Tetrahedron Lett. 29:5057 5060 (1988), Kemp et al., Tetrahedron Lett. 29:4935 4938 (1988) and Kemp et al., J. Org. Chem. 54:109 115 (1987). Other suitable but exemplary peptidomimetics are shown in Nagai and Sato, Tetrahedron Lett. 26:647 650 (1985); Di Maio et al., J. Chem. Soc. Perkin Trans., 1687 (1985); Kahn et al., Tetrahedron Lett. 30:2317 (1989); Olson et al., J. Am. Chem. Soc. 112:323 333 (1990); Garvey et al., J. Org. Chem. 56:436 (1990). Further suitable exemplary peptidomimetics include hydroxy 1,2,3,4 tetrahydroisoquinoline 3 carboxylate (Miyake et al., J. Takeda Res. Labs 43:53 76 (1989)); 1,2,3,4 tetrahydro- isoquinoline 3 carboxylate (Kazmierski et al., J. Am. Chem. Soc. 113:2275 2283 (1991)); histidine isoquinolone carboxylic acid (HIC) (Zechel et al., Int. J. Pep. Protein Res. 43 (1991)); (2S, 3S) methyl phenylalanine, (2S, 3R) methyl phenylalanine, (2R, 3S) methyl phenylalanine and (2R, 3R) methyl phenylalanine (Kazmierski and Hruby, Tetrahedron Lett. (1991)).

Exemplary, illustrative but non-limiting non-natural amino acids include beta-amino acids (beta3 and beta2), homo-amino acids, cyclic amino acids, aromatic amino acids, Pro and Pyr derivatives, 3-substituted Alanine derivatives, Glycine derivatives, ring-substituted Phe and Tyr Derivatives, linear core amino acids or diamino acids. They are available from a variety of suppliers, such as Sigma- Aldrich (USA) for example. In the present invention any part of a peptide may optionally be chemically modified, i.e., changed by addition of functional groups. The modification may optionally be performed during synthesis of the molecule if a chemical synthetic process is followed, for example by adding a chemically

modified amino acid. However, chemical modification of an amino acid when it is already present in the molecule ("in situ" modification) is also possible.

The amino acid of any of the sequence regions of the molecule can optionally be modified according to any one of the following exemplary types of modification (in the peptide conceptually viewed as "chemically modified"). Non-limiting exemplary types of modification include carboxymethylation, acylation, phosphorylation, glycosylation or fatty acylation. Ether bonds can optionally be used to join the serine or threonine hydroxyl to the hydroxyl of a sugar. Amide bonds can optionally be used to join the glutamate or aspartate carboxyl groups to an amino group on a sugar (Garg and Jeanloz, *Advances in Carbohydrate Chemistry and Biochemistry*, Vol. 43, Academic Press (1985); Kunz, *Ang. Chem. Int. Ed. English* 26:294-308 (1987)). Acetal and ketal bonds can also optionally be formed between amino acids and carbohydrates. Fatty acid acyl derivatives can optionally be made, for example, by acylation of a free amino group (e.g., lysine) (Toth et al., *Peptides: Chemistry, Structure and Biology*, Rivier and Marshal, eds., ESCOM Publ., Leiden, 1078-1079 (1990)).

The compositions of the present invention may also be expressed in-vivo using genetic engineering techniques (e.g., using transgenic aquatic sessile organisms).

The compositions of the present invention may be devoid of cytotoxic or cytostatic activity – e.g., they are not bactericidal or bacteristatic.

The present inventors have characterized and isolated a natural peptide comprising a sequence selected from the group consisting of YDYNWY, YDYNLY, FDYNFY, FDYNLY, WDYNLY, FDYNWY, YDWNLY, YDWHLY and YDYSFY having effective anti-cell aggregate (e.g., anti-biofilm) properties. For example, the peptide may comprise at least one of the following sequences: LFSVPYDYNWYSNWW, LFSVPYDYNWYSNWW, FSVPYDYNLYSNWW, MFSVPYDYNLYSNWV, MFSVPFDYNFYSNWW, LFSVPFDYNFYSNWW, MFSVPFDYNLYSNWW, MFSVPFDYNLYTNWW, MWSVPFDYNLYSNWW, MFSVPFDYNLYKNWL, LFSVPFDYNLYSNWW, LFSIPFDYNLYSNWW, MFSVPWDYNLYKNWL, MFSVPWDYNLYKNWF, MFSVPFFDYNWYSNWW, MASIPYDWNLYQSWA, MASIPYDWNLYSAWA or MASIPYDWHLYNAWA, or combinations thereof.

The present inventors have characterized and isolated a natural peptide comprising the sequence selected from the group consisting of: FDYDWY, FNFDWY and FDFDWY, having effective anti-cell aggregate (e.g., anti-biofilm) properties. For example, the peptide may comprise at least one of the following sequences: SFDYDWY, SFDYDWYN, HSFDYDWYN, HSFDYDWYNV, VHSFDYDWYNV, VHSFDYDWYNVS, SVHSFDYDWYNVS, SVHSFDYDWYNVSD, KSVHSFDYDWYNVSD, KSVHSFDYDWYNVSDQ, NKSVDYDWYNVSDQ, NKSVDYDWYNVSDQA, QNKSVHSFDYDWYNVSDQA, QNKSVHSFDYDWYNVSDQAD, SQNKSVHSFDYDWYNVSDQAD, SQNKSVHSFDYDWYNVSDQADL, FSQNKSVHSFDYDWYNVSDQADL, FSQNKSVHSFDYDWYNVSDQADLK, SFSQNKSVHSFDYDWYNVSDQADLK, SFSQNKSVHSFDYDWYNVSDQADLKN, CSFSQNKSVHSFDYDWYNVSDQADLKN or CSFSQNKSVHSFDYDWYNVSDQADLKNC, or combinations thereof.

The present inventors have characterized and isolated a natural peptide comprising a sequence selected from the group consisting of SVPYDYNWYSNW, SFSQNKSVHSFDYDWYNVSDQADLKN and SVHSFDYDWYNV.

An exemplary protein or peptide agent derived from an aquatic organism may be used to detach a single cell organism from a surface or from other single cell organisms is equinatoxin. Equinatoxins (i.e., equinatoxins 1, 2 and 3) have pore forming toxins found in sea anemones (e.g., *Actinia equina*). Equinatoxins may be comprised in sea anemone cells or may be isolated therefrom. Any equinatoxin may be used according to the teachings of the present invention for detaching microorganisms from a surface or from each other. One example is the Equinatoxin-2 precursor (GenBank accession identifier >gi|48428895|sp|P61914.1|ACTP2_ACTEQ (Equinatoxin II) (EqT II) (EqTII)):

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MSRLIIVFIVVTMICSATALPSKKIIDEDEDEKRSADVAGAVIDGASLS
FDILKTVLEALGNVKKRIAVGVDNESGKTWTALNTYFRSGTSDIVLPH
KVPHGKALLYNGQKDRGPVATGAVGVLAYLMSDGNTLAVLFSVPYD
YNWYSNWWNVRIYKGRRADQRMYEELYNNLSPFRGDNGWHTRNL
GYGLKSRGFMNSSGHAILEIHVSKA
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Another exemplary protein derived from a sea anemone is *Stichodactyla helianthus* (GenBank accession identifier >gi|2815496|sp|P07845.2|ACTP2_STOHE (Sticholysin II) (StnII) (Cytolysin St II) (Cytolysin III) (Cytotoxin)):

ALAGTIIAGASLTFQVLDKVLEELGKVSARKIAVGIDNESGGTWTALNA
YFRSGTTDVILPEFVPNTKALLYSGRKDTGPVATGAVAAAFAYYMSSG
NTLGVMFSPFDYNWYSNWWVDVKIYSGKRRADQGMIEDLYYGNPY
RGDNGWHEKNLGYGLRMKGIMTSAGEAKMQIKISR

An exemplary protein derived from bony fish is *Danio rerio* (GenBank accession identifier >gi|125821212|ref|XP_001342650.1|PREDICTED (Danio rerio)):

MTESAEVAANVSSRRHATVEITNLNNYCFLNPKVYLENGETSNPPQ
PTVRPLKTEVCTFSKSAAHATGSVGVLTIDLFERRRNDYTETLAIMFS
VPWDYNLYKNWFAVGIYPKGKECDQALYKEMYYQKNQHGFVREEA
NGSGINFEGKNLDIRATMCPMGRAIVKVEVWDKLLSPMAQMDC

Another exemplary protein derived from bony fish is *Tetraodon nigroviridis* (GenBank accession identifier >gi|47218822|emb|CAG02807.1|unnamed protein product (Tetraodon nigroviridis)):

MESAEVAADVSRRSVTIEISNLTKNYCLINPRVYLESGETYNPPQPT
VRPLMTEVCTFSKSSGIPTGSVGVLTIDYELLERRSTMLPETLAIMFSVPY
DYSFYNNWFAVGIYETGTCNEGLYKQMYNEKKQAEHGFVREKANG
SGINYVGGNLDIRATMNPLGKAIMKVEVWDAFFPFSE

An exemplary protein derived from moss is *Physcomitrella patens* (GenBank accession identifier >gi|16806023|ref|XP_001782104.1|predicted protein (Physcomitrella patens subsp. patens)):

MVVHLIAMGLRYSETIMKTARMAEAIIPAAELSIKTLQNIVEGITGVDR
KIAIGFKNLTDYTLENLGVYFNSGSSDRSIAYKINAQEALLFSARKSDH
TARGTVGTFSYYIQDEDKTVHVMWSVPFDYNLYSNWWNIAVVDGRQ

PPDSNVHDNLYNGSGGMPYPNKPQYINNEQKGFHLFGSMTNNGQA
TIEVELKKA

An exemplary protein derived from birds is *Gallus gallus* (GenBank accession identifier >gi|118129726|ref|XP_001231839.1|PREDICTED: hypothetical protein isoform (*Gallus gallus*)):

MPPKEKKENDKPCNDNCQPKPQGKGVESLMKNIDVCRSVGLEIINRTR
TVTLTDFRSYCFSGKIVTTLPFEIGPDSKGICIFAKTPYSLRGSGVTVC
KADTFFLAITFSNPYDYILYKIEFALEIFTEPNHLGNLGDVFSKMMKSK
PYCGSSSLFQRAVLESEHETLEVSKGSIRVQAKMSNNRKAILKVQVEDM
DPPPYSKGM

An exemplary protein derived from a platypus is *Ornithorhynchus anatinus* (GenBank accession identifier >gi|149638239|ref|XP_001512702.1|PREDICTED: integral membrane protein GPR137B-like (*Ornithorhynchus anatinus*)):

MEGSPPGRPPGNDLSPPTLSPAVPPYVKLGLTSVYTAFYSLLFVFVYAQLWL
LHHRHRRLSYQTVFLFLCLLWAALRTVLFSFYFRDFLAANKLGPFQFWLLYC
CPVCLQFFTLTLMNLYFSQVIFKAKSKFSPELLKYRLALYLASLVVSLVFLV
NLCAVLVKTGTWERKVVVSVRVAINDTLFLCAVSLSVCLYKISKMSLANI
YLESKGSSVCQVTAIGVTVILLYASRACYNLFTLSFSRHHGSSFDYDWNVSDQ
ADLKSQGLDAGYVVFGVVLFWELLPTSLVVYFFRVRNPTKDPTNPRGVP
AFSPRSYFFDNPRRYDSDDDLAWNVA PQGFQGSFAPDYDYGQPSSTGHI
GSLQQDSDLNKGKPSHA

An example of an applicable peptide derived from equinatoxin is SVPYDYNWYSNW, in either soluble form or conjugated to polyethylene glycol (PEG) and palmitic acid and capable of adhering to a surface. The soluble and conjugated form of the peptide in cyclic form and having a cysteine at each of the N- and C-terminal ends are as follows:

Eqt2ZCyc (soluble aquatic peptide): CSVPYDYNWYSNWC

Eqt2ZCyc-3PEG-Pal (adhesive aquatic peptide): Pal--(miniPEG)3-
CSVPYDYNWYSNWC

Peptides based on a subset of equinatoxin sequences and variants thereof are also applicable. The chart below demonstrates specific 12-mer peptide embodiments, with applicable amino acids for each of the 12 positions. Bolded amino acids correspond to amino acids not naturally occurring at the corresponding positions, but which provide for peptides falling within the scope of the present invention. In some embodiments, the peptide is not SVPYDYNWYSNW.

		1	2	3	4	5	6	7	8	9	10	11	12	
Equinatoxin		S	V	P	Y	D	Y	N	W	Y	S	N	W	
		N	I		F	N	F	S	L	N	K	E	R	
		I	A		W	Q	R	G	F	F	N	D	V	
		V	N			E	W	D	M	K	T	Q	L	
		R	L				V	H	S	L	E	S	I	
		K	Q				H	E	T	R	R	A	K	
		Q					L	Q	R	I	L	I	F	
		L					K	I	A	V	Q		E	
							I		G	W	I			
									V	Q	V			
									P		D			
									Y		K			
									I					
									K					

An exemplary peptide agent may be derived from a human. For example, a peptide derived from GPCR137b (GPR137B (TM7SF1)) may be used to detach a single cell organism from a surface or from other single cell organisms. GPCR137b can be found in human kidney, heart, brain, and placenta tissue. [Spangenberg et al., Genomics. 1998 Mar 1;48(2):178-85; Bjarnadottir et al., Genomics, 88(3): 263-273 (2006)]. Peptides SFSQNKSVHSFDYDWYNVSDQADLKN and SVHSFDYDWYNV, as cyclic peptides with cysteine attached to both the C- and N-terminals of the peptides in either soluble form or conjugated to polyethylene glycol and palmitic acid and capable of adhering to a surface, are two examples of applicable peptides derived from human GPCR137b:

grZ28Cyc (soluble long human peptide):
CSFSQNKSVHSFDYDWYNVSDQADLKNC

grZ28Cyc-3PEG-Pal (adhesive long human peptide): Palmitoyl-(miniPEG)3-CSFSQNKSVHSFDYDWYNVSDQADLKNC

grZ14s-nvCyc (soluble short human peptide): CSVHSFDYDWYNVC

grZ14s-nvCyc-3PEG-Pal (adhesive short human peptide): Pal--(miniPEG)3-CSVHSFDYDWYNVC

Peptides based on a subset of GPCR137b sequences and variants thereof are also applicable. The chart below demonstrates specific 12-mer peptide embodiments, with applicable amino acids for each of the 12 positions. Bolded amino acids correspond to amino acids not naturally occurring at the corresponding positions, but which provide for peptides falling within the scope of the present invention. In some embodiments, the peptide is not SVHSFDYDWYNV.

	1	2	3	4	5	6	7	8	9	10	11	12	
GPCR137B	S	V	H	S	F	D	Y	D	W	Y	N	V	
	N	I	N	P	W	N	F	G	F	F	Q	I	
	T	L	Q	A	Y	E	W	E	Y	W		L	
	K	Y	E	T		Q							
	R	G	D										
	H	F	S										
	E	W											
	I												
	Q												
	D												

An exemplary peptide derived from *Physcomitrella patens* (moss) is SVVPFDYNLYSNW. This same sequence can also be found in *Selaginella moellendorffii* (Genbank Accession No: XP_002963283), a type of plant, and in the following sea anemones: *Phyllodiscus semoni* (Genbank Accession No: BAI70365); *Heteractis crispa* (Genbank Accession No: AAW47930); and *ActinERIA villosa* (Genbank Accession No: BAD74019). The cyclicized form of the peptide is as follows:

Physco-Cyc: CSVVPFDYNLYSNWC

EXAMPLES

Example 1: Detaching Overnight Biofilm With Overnight Eqt2Z-cyc Incubation

A biofilm was created by incubating *Pseudomonas aeruginosa* (ATCC 27853) overnight in wells at 37 °C, allowing the microorganism to attach to the well surface. Eqt2Z-cyc peptide [Custom Peptide Synthesis; Peptron, Korea] at different concentrations was subsequently added to the wells and allowed to incubate overnight. The P.a. control was *Pseudomonas aeruginosa* alone. Figure 1 demonstrates that Eqt2Z-cyc peptides caused far greater microorganism detachment than in the P.a. control.

Example 2: Detaching 2-hour Biofilm With Overnight Eqt2Z-cyc Incubation

A biofilm was created by culturing *Pseudomonas aeruginosa* (ATCC 27853) for 2 hours in a well at 37 °C. Eqt2Z-cyc peptide at different concentrations was subsequently added to the well and allowed to incubate overnight. The Abac10-cyc peptide served as a negative control. The P.a. control was *Pseudomonas aeruginosa* alone. Figure 2A demonstrates that Eqt2Z-cyc peptides caused far greater microorganism detachment than in the P.a. control or negative control.

Example 3: Detaching 2-hour Biofilm With 24-hour Eqt2Z-cyc Incubation

A biofilm was created by culturing *Pseudomonas aeruginosa* (ATCC 27853) for 2 hours in a well at 37 °C. Eqt2Z-cyc peptide at different concentrations was subsequently added to the well and allowed to incubate for 24 hours. The Abac10-cyc peptide served as a negative control. The P.a. control was *Pseudomonas aeruginosa* alone. Figure 2B demonstrates that Eqt2Z-cyc peptides caused far greater microorganism detachment than in the P.a. control or negative control. Staining was performed with crystal violet at OD 595 nm.

Example 4: Detaching 2-hour Biofilm With 24-hour grZ14s-nvCyc Incubation

A biofilm was created by culturing *Pseudomonas aeruginosa* (ATCC 27853) for 2 hours in a well at 37 °C. grZ14s-nvCyc peptide [Custom Peptide Synthesis; Peptron, Korea] at different concentrations was subsequently added to the well and allowed to incubate for 24 hours. The Abac10-cyc peptide served as a negative control. The P.a. control was

Pseudomonas aeruginosa alone. Figure 3 demonstrates that grZ14s-nvCyc peptides caused far greater microorganism detachment than in the P.a. control or negative control. Staining was performed with crystal violet at OD 595 nm.

Example 5: Detaching 24-hour Biofilm With 24-hour Eqt2Z-cyc Incubation

A biofilm was created by culturing *Pseudomonas aeruginosa* (ATCC 27853) for 24 hours in a well at 37 °C. Eqt2Z-cyc peptide at different concentrations was subsequently added to the well and allowed to incubate for 24 hours. The Abac10-cyc peptide served as a negative control. The P.a. control was *Pseudomonas aeruginosa* alone. Figure 4 demonstrates that Eqt2Z-cyc peptides caused far greater microorganism detachment than in the P.a. control or negative control. Staining was performed with crystal violet at OD 595 nm.

Example 6: Detaching 24-hour Biofilm With 24-hour grZ14s-nvCyc

A biofilm was created by culturing *Pseudomonas aeruginosa* (ATCC 27853) for 24 hours in a well at 37 °C. grZ14s-nvCyc peptide at different concentrations was subsequently added to the well and allowed to incubate for 24 hours. The Abac10-cyc peptide served as a negative control. The P.a. control was *Pseudomonas aeruginosa* alone. Figure 5 demonstrates that grZ14s-nvCyc peptides caused far greater microorganism detachment than in the P.a. control or negative control. Staining was performed with crystal violet at OD 595 nm.

Example 7: Detaching 24-hour Biofilm With Overnight PhyscoZ-Cyc

A biofilm was created by culturing *Pseudomonas aeruginosa* (ATCC 27853) for 24 hours in a well at 37 °C. PhyscoZ-Cyc peptide [Custom Peptide Synthesis; Peptron, Korea] at different concentrations was subsequently added to the well and allowed to incubate for 24 hours. The P.a. control was *Pseudomonas aeruginosa* alone. Figure 6 demonstrates that PhyscoZ-Cyc peptides caused far greater microorganism detachment than in the P.a. control. Staining was performed with crystal violet at OD 595 nm.

Example 8: grZ14s-nvCyc enhances antibiotic activity of Imipenem and Ampicillin

The ability of grZ14s-nvCyc to enhance the activity of Imipenem or Ampicillin was performed by first incubating the wells in a 96-well plate with *Pseudomonas aeruginosa* for 24 hours at 37 °C and 50 rpm shaking. The plate was washed 2 times with phosphate buffer solution. The wells were then filled with either the antibiotic alone or the antibiotic with grZ14s-nvCyc (100 ng/ml). The wells were then incubated for 48 hours (24 hours for Ampicillin) at 37 °C and 50 rpm shaking. 0.3% triton X-100 and 0.45% EDTA were then added to the wells, bringing the wells to a final concentration of 0.075% triton X-100 and 0.1125% EDTA. The plates were then sonicated in a bath-sonicator for 12 minutes. The wells were then serially diluted in PBS. The solutions were then seeded on a blood plate agar and incubated at 37 °C for 24 hours.

The results are presented in Figures 7 and 8. As Figure 7 demonstrates, the number of live bacteria was much lower in wells with grZ14s-nvCyc and Imipenem than in wells with Imipenem alone, thereby demonstrating that grZ14s-nvCyc enhances the activity of the antibiotic. Figure 8 demonstrates that the number of live bacterial is much lower in wells with grZ14s-nvCyc and Ampicillin than in wells with Ampicillin alone, thereby demonstrating that grZ14s-nvCyc enhances the activity of the antibiotic.

Example 9: grZ14s-nvCyc enhances antibiotic activity of Vancomycin

The ability of grZ14s-nvCyc to enhance the activity of Vancomycin was performed by first incubating the wells in a 96-well plate with *Staphylococcus aureus* for 24 hours at 37 °C and 50 rpm shaking. The plate was washed 2 times with phosphate buffer solution. The wells were then filled with either the antibiotic alone or the antibiotic with grZ14s-nvCyc (100 ng/ml). The wells were then incubated for 24 hours at 37 °C and 50 rpm shaking. 0.3% triton X-100 and 0.45% EDTA were then added to the wells, bringing the wells to a final concentration of 0.075% triton X-100 and 0.1125% EDTA. The plates were then sonicated in a bath-sonicator for 12 minutes. The wells were then serially diluted in PBS. The solutions were then seeded on a blood plate agar and incubated at 37 °C for 24 hours.

The results are presented in Figure 9. As the demonstrates, the number of live bacteria was much lower in wells with grZ14s-nvCyc and Vancomycin than in wells with

Vancomycin alone, thereby demonstrating that grZ14s-nvCyc enhances the activity of the antibiotic.

Example 10: grZ14s-nvCyc enhances antifungal activity of Amphotericin

The ability of grZ14s-nvCyc to enhance the activity of Amphotericin was performed by first incubating the wells in a 96-well plate with *Candida albicans* for 24 hours at 37 °C and 50 rpm shaking. The plate was washed 2 times with phosphate buffer solution. The wells were then filled with either the antifungal alone or the antifungal with grZ14s-nvCyc (100 ng/ml). The wells were then incubated for 24 hours at 37 °C and 50 rpm shaking. The plates were again washed 2 times with PBS. BacTiter-Glo Microbial Cell Viability Assay (Promega, USA) was used following the manufacturer protocol and luminescence read.

The results are presented in Figure 10. As the demonstrates, the number of live fungus was much lower in wells with grZ14s-nvCyc and Amphotericin than in wells with Amphotericin alone, thereby demonstrating that grZ14s-nvCyc enhances the activity of Amphotericin.

Example 11: grZ14s-nvCyc enhances antifungal activity of Fluconazole

The ability of grZ14s-nvCyc to enhance the activity of Fluconazole was performed by first incubating the wells in a 96-well plate with *Candida albicans* for 24 hours at 37 °C and 50 rpm shaking. The plate was washed 2 times with phosphate buffer solution. The wells were then filled with either the antifungal alone or the antifungal with grZ14s-nvCyc (100 ng/ml). The wells were then incubated for 24 hours at 37 °C and 50 rpm shaking. The plates were again washed 2 times with PBS. BacTiter-Glo Microbial Cell Viability Assay (Promega, USA) was used following the manufacturer protocol and luminescence read.

The results are presented in Figure 11. As the demonstrates, the number of live fungus was much lower in wells with grZ14s-nvCyc and Fluconazole than in wells with Fluconazole alone, thereby demonstrating that grZ14s-nvCyc enhances the activity of Fluconazole.

Example 12: grZ14s-nvCyc enhances antibiotic activity of Kanamycin

The ability of grZ14s-nvCyc to enhance the activity of Kanamycin was performed by first incubating the wells in a 96-well plate with *Pseudomonas aeruginosa* for 24 hours at 37 °C and 50 rpm shaking. The plate was washed 2 times with phosphate buffer solution. The wells were then filled with either Kanamycin alone or Kanamycin with grZ14s-nvCyc (100 ng/ml). The wells were then incubated for 24 hours at 37 °C and 50 rpm shaking. The plates were again washed 2 times with PBS. BacTiter-Glo Microbial Cell Viability Assay (Promega, USA) was used following the manufacturer protocol and luminescence read.

The results are presented in Figure 12. As the demonstrates, the number of live bacteria was much lower in wells with grZ14s-nvCyc and Kanamycin than in wells with Kanamycin alone, thereby demonstrating that grZ14s-nvCyc enhances the activity of Kanamycin.

Example 13: Preventing adhesion and detachment of microorganisms with various peptides

The peptides depicted in Table 1 (each with a cysteine attached to the N- and C-terminus of the peptides) were synthesized using solid-phase methods and were purified to 90% (Peptron, Inc.; Taejeon, South Korea). The peptides were dissolved in 20 µl dimethyl sulfoxide and diluted in double distilled water to a concentration of 10 mg/ml. Additional dilutions were performed in phosphate buffered saline (PBS).

The ability of these peptides to prevent microbial adhesion to a surface or detach microbial adhesion from a surface were measured against the following microbial strains: *Pseudomonas aeruginosa* (ATCC27853), *Staphylococcus aureus* (ATCC25923), *Candida albicans* (ATCC14053) and *Acinetobacter baumannii* (clinical isolate). Specifically, peptides diluted to concentrations of 100 ng/ml were incubated with the foregoing microbes for 24 hours.

Biofilms of the foregoing microorganisms were grown in a 96-well found-bottom polystyrene plate. Specifically, 180 µl of overnight cultures were added to the wells with 20 µl of each peptide diluted in PBS, either simultaneously to measure prevention or after a period of time to measure detachment from 4 hours up to 24 hours post incubation. After 24

hours of incubation at 37 °C, each well was washed with water and stained with 250 µl of crystal violet solution. The crystal violet solution was then removed by thoroughly washing the wells with water. To quantify the number of bacteria cells attached to the wells, the crystal violet was solubilized in 250 µl of 1% sodium dodecyl sulfate and the absorbance was measured at 595 nm. As Table 1 demonstrates, the peptides decreased the amount of microbial attachment to the wells (Table 1: "Biofilm prevention") and promoted microbial detachment (Table 1: "Detachment"), thereby preventing the formation of biofilm by the microorganisms and causing them to detach where they have already adhered to a surface.

TABLE 1. Biofilm prevention and detachment by various peptides

Peptide: SelagZ-cyc CSVPFDYNLYSNWC	Biofilm prevention (%)	Detachment (%) (100ng/ml)
<i>Pseudomonas aeruginosa</i> ATCC27853	88%(100ng/ml)	67%
<i>Staphylococcus aureus</i> ATCC25923	88%(100ng/ml)	35%
<i>Candida albicans</i> ATCC14053	80%(10ng/ml)	NA
<i>Acinetobacterbaumannii</i>	80%(100ng/ml)	50%
Peptide: PhyscoZ-cyc CSVPFDYNLYSNWC	Biofilm prevention (%)	Detachment (%) (100ng/ml)
<i>Pseudomonas aeruginosa</i> ATCC27853	95%(100ng/ml)	75%
<i>Staphylococcus aureus</i> ATCC25923	65%(100ng/ml)	70%
<i>Candida albicans</i> ATCC14053	62%(10ng/ml)	NA
<i>Acinetobacterbaumannii</i>	NA	50%
Peptide: EchoZ-cyc CSAPYNFNFYSNWC	Biofilm prevention (%)	Detachment (%) (100ng/ml)
<i>Pseudomonas aeruginosa</i> ATCC27853	90%(100ng/ml)	65%
<i>Staphylococcus aureus</i> ATCC25923	85%(100ng/ml)	35%
<i>Candida albicans</i> ATCC14053	80%(10ng/ml)	NA
<i>Acinetobacterbaumannii</i>	80%(100ng/ml)	60%

Peptide: XenoZ-S-cyc CSRYSSFDYDWYNVC	Biofilm prevention (%)	Detachment (%) (100ng/ml)
<i>Pseudomonas aeruginosa</i> ATCC27853	57%(10ng/ml)	30%
<i>Staphylococcus aureus</i> ATCC25923	65%(100ng/ml)	NA
<i>Candida albicans</i> ATCC14053	75%(100ng/ml)	50%
<i>Acinetobacterbaumannii</i>	95%(100ng/ml)	49%

Peptide: CionaZ-S-cyc CSELSSFNFDWYNVC	Biofilm prevention (%)	Detachment (%) (100ng/ml)
<i>Pseudomonas aeruginosa</i> ATCC27853	75%(100ng/ml)	78%
<i>Staphylococcus aureus</i> ATCC25923	90%(100ng/ml)	NA
<i>Candida albicans</i> ATCC14053	68%(100ng/ml)	42%
<i>Acinetobacterbaumannii</i>	52%(100ng/ml)	80%

Peptide: CanisZ-cyc CNVHSFDYDWYNVC	Biofilm prevention (%)	Detachment (%) (100ng/ml)
<i>Pseudomonas aeruginosa</i> ATCC27853	95%(100ng/ml)	72%
<i>Staphylococcus aureus</i> ATCC25923	75%(100ng/ml)	NA
<i>Candida albicans</i> ATCC14053	80%(100ng/ml)	45%
<i>Acinetobacterbaumannii</i>	60%(10ng/ml)	73%

Peptide: NilotiZ-cyc CRVESFNFDWYNVC	Biofilm prevention (%)	Detachment (%) (100ng/ml)
<i>Pseudomonas aeruginosa</i> ATCC27853	90%(100ng/ml)	40%
<i>Staphylococcus aureus</i> ATCC25923	63%(100ng/ml)	65%
<i>Candida albicans</i> ATCC14053	30%(100ng/ml)	65%

<i>Acinetobacterbaumannii</i>	87%(100ng/ml)	NA
Peptide: SalmoZ-cyc CRVESFDFDWYNIC	Biofilm prevention (%)	Detachment (%) (100ng/ml)
<i>Pseudomonas aeruginosa</i> ATCC27853	90%(10ng/ml)	80%
<i>Staphylococcus aureus</i> ATCC25923	80%(100ng/ml)	75%
<i>Candida albicans</i> ATCC14053	55%(100ng/ml)	73%
<i>Acinetobacterbaumannii</i>	95%(100ng/ml)	NA
Peptide: TetraoZ-cyc CRINSFDYDWYNVC	Biofilm prevention (%)	Detachment (%) (100ng/ml)
<i>Pseudomonas aeruginosa</i> ATCC27853	45%(100ng/ml)	NA
<i>Staphylococcus aureus</i> ATCC25923	52%(100ng/ml)	43%
<i>Candida albicans</i> ATCC14053	25%(100ng/ml)	55%
<i>Acinetobacterbaumannii</i>	70%(10ng/ml)	NA
Peptide: AnoliZ-cyc CTVNSFDYDWYNVC	Biofilm prevention (%)	Detachment (%) (100ng/ml)
<i>Pseudomonas aeruginosa</i> ATCC27853	95%(100ng/ml)	90%
<i>Staphylococcus aureus</i> ATCC25923	75%(100ng/ml)	72%
<i>Candida albicans</i> ATCC14053	55%(100ng/ml)	72%
<i>Acinetobacterbaumannii</i>	92%(10ng/ml)	NA
Peptide: MeleagZ-cyc CKVNSFDYDWYNVC	Biofilm prevention (%)	Detachment (%) (100ng/ml)
<i>Pseudomonas aeruginosa</i> ATCC27853	82%(100ng/ml)	65%
<i>Staphylococcus aureus</i> ATCC25923	73%(100ng/ml)	80%
<i>Candida albicans</i> ATCC14053	30%(100ng/ml)	62%
<i>Acinetobacterbaumannii</i>	95%(100ng/ml)	NA
Peptide: CapraZ-cyc CTVHSFDYDWYNVC	Biofilm prevention (%)	Detachment (%) (100ng/ml)
<i>Pseudomonas aeruginosa</i> ATCC27853	68%(100ng/ml)	65%

<i>Staphylococcus aureus</i> ATCC25923	42%(100ng/ml)	55%
<i>Candida albicans</i> ATCC14053	25%(100ng/ml)	60%
<i>Acinetobacterbaumannii</i>	90%(100ng/ml)	NA

Peptides listed in Table 2, synthesized in the same manner described above for Table 1, have a cysteine that was added to the N- and C-terminal ends of the peptides, as represented by the "C" at either end of the peptides. These peptides also are cyclized, as indicated by the "(C-C)" term at the end of each peptide. EqSyn-Neg- and GRSyn-Neg were added as negative controls.

Table 2. Various peptides

No	Peptide ID	Sequence	No	Peptide ID	Sequence
1	EqSyn 1	CNIPFNFSLNKERC (C-C)	20	GRSyn 1	CSVHSDYDWYNVC (C-C)
2	EqSyn2	CSVPHYQYNWYSNWC (C-C)	21	GRSyn2	CSVHSYDFDWYNVC (C-C)
3	EqSyn 3	CSVPWEYNFYSNWC (C-C)	22	GRSyn 3	CTLQAFNYEWYQLC (C-C)
4	EqSyn 4	CRIPYDRGMIVNVC (C-C)	23	GRSyn 4	CKYETFEYGWYNIC (C-C)
5	EqSyn 5	CKVPYDWDSVINLC (C-C)	24	GRSyn 5	CHGDSFQYEWYNLC (C-C)
6	EqSyn 6	CQLPYDVHTYNDWC (C-C)	25	GRSyn 6	CSVHSFDWDWYNVC (C-C)
7	EqSyn 7	CLAPYDHNRYTQWC (C-C)	26	GRSyn 7	CSVHSFDYDYYNVC (C-C)
8	EqSyn 8	CSNPYDLEAYENWC (C-C)	27	GRSyn 8	CSVHSFDYDFYNVC (C-C)
9	EqSyn 9	CSVPHYDYQGYRNIC (C-C)	28	GRSyn 9	CSVHSFDYDWFNVC (C-C)
10	EqSyn 10	CSVPHYDYNVYLKNC (C-C)	29	GRSyn 10	CSVHSFDYDWWNVC (C-C)
11	EqSyn 11	CIQPYDKNYFQNFC (C-C)	30	GRSyn 11	CIFNPFDYDWYNVC (C-C)
12	EqSyn 12	CVVPYDINIKDNWC (C-C)	31	GRSyn 12	CQWHSFDYDWYNVC (C-C)
13	EqSyn 13	CSVPHYDYNPYSNWC (C-C)	32	GRSyn 13	CDVHPFDYDWYNVC (C-C)
14	EqSyn 14	CSVPHYDYNKLKNWC (C-C)	33	GRSyn-Neg	CSVHSGDYDGGNVC (C-C)
15	EqSyn 15	CSVPHYDYNWRSSWC (C-C)	34	grZ14s-nvCyc	CSVHSFDYDWYNVC (C-C)
16	EqSyn 16	CSVPHYDYNWWSAWC (C-C)			
17	EqSyn 17	CSVPHYDYNWQSNWC (C-C)			
18	EqSyn-Neg	CSVGYDYNWYSNWC (C-C)			
19	Eq2Z-cyc	CSVPHYDYNWYSNWC (C-C)			

The ability of these peptides to (1) prevent microbial adhesion to a surface and to (2) detach microbes already adhered to a surface were measured against the following microbial strains: *Pseudomonas aeruginosa* (ATCC27853), *Staphylococcus aureus* (ATCC25923), *Candida albicans* (ATCC14053) and/or *Escherichia coli*. The resulting percentage of prevention or detachment, relative to control wells in which the microbes was allowed to culture in the absence of peptides, are indicated in Figures 13 to 16. The figures demonstrate that these peptides are able to prevent microbial adhesion to a surface and detach microbes already adhered to a surface.

Example 14: Preventing adhesion of *Pseudomonas aeruginosa* with peptides modified from Eqt2Z-cyc

The Eqt2Z-cyc peptide was modified by inserting synthetic amino acid analogs in different positions in the natural sequence. Table 3 depicts the modifications performed on Eqt2Z-cyc. The modified peptides were incubated with *Pseudomonas aeruginosa*. A biofilm was created by culturing the microorganism for 24 hours in a well at 37 °C, modified Eqt2Z-cyc-mod peptide was subsequently added to the well and allowed to incubate for 24 hours. Figure 17 depicts the results of the incubation, in which the peptides prevented adhesion of the microorganism when compared with incubation of the microorganism without any peptide.

Table 3. Modifications of Eqt2Z-cyc

Eqt2ZCyc-mod1	(D-form Cys)-SVPYDYNWYSNWC (C-C)
Eqt2ZCyc-mod2	CSVYDYNWYSNWC-(D-form Cys) (C-C)
Eqt2ZCyc-mod3	(D-form Cys)-SVPYDYNWYSNWC-(D-form Cys) (C-C)
Eqt2ZCyc-mod4	homoCys-SVPYDYNWYSNWC (C-C)
Eqt2ZCyc-mod5	CSVYDYNWYSNWC-homoCys (C-C)
Eqt2ZCyc-mod7	CS-(D-form V)-PYDYNWYSNWC (C-C)
Eqt2ZCyc-mod8	CSV-(D-form P)-YDYNWYSNWC (C-C)

Example 15: Peptides contribute to detachment of microorganisms by affecting polysaccharide matrix production

In one embodiment, peptides can cause microorganisms to detach from a surface by affecting the microorganisms' production of polysaccharide matrix. The effect of Eqt2Z-Cyc and grZ14s-nvCYC on the polysaccharide matrix production of *Pseudomonas aeruginosa*, *Candida albicans* and *Staphylococcus aureus* were evaluated by measuring Congo Red and/or Trypan Blue after the peptides were incubated with the microorganisms. Congo Red and Trypan Blue are dyes that bind to the polysaccharide matrix via exopolysaccharide fibrils. A reduction in absorbance of these dyes corresponds with a reduction in polysaccharide matrix production.

Specifically, Congo Red and Trypan blue binding assays were performed by growing *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Candida albicans* in specific growth media (LB, TSB+0.25% glucose, RPMI 1640, respectively) simultaneously with the peptides until they reached a density of $OD_{600nm} = 0.25$. The cells were then pelleted, the supernatant removed and the cells were resuspended in TPM buffer (contains 10.0 mM Tris/HCl (pH 8.0), 1 mM KH_2PO_4 and 8.0 mM $MgSO_4$) to a density of $OD_{600nm} = 0.25$. Aliquots of the cell suspensions were mixed with stock solutions of Congo Red ($150 \mu g\ ml^{-1}$) and Trypan Blue ($100 \mu g\ ml^{-1}$). TPM buffer was added to the cell/dye mixtures to give final concentrations of 2.5×10^8 cells ml^{-1} and either $15 \mu g$ Congo Red ml^{-1} or $10 \mu g$ Trypan Blue ml^{-1} . Cell-free samples containing TPM buffer and $15 \mu g$ Congo Red ml^{-1} or $10 \mu g$ Trypan Blue ml^{-1} were used as controls. All samples were vortexed briefly and incubated in a 25 °C dark room for 30 min. Following the incubation, the cells were pelleted, and the supernatants were transferred to cuvettes. The absorbance of each supernatant sample was measured at 490 nm to detect Congo Red or at 585 nm to detect Trypan Blue, and these values were compared to the absorbance of the appropriate control sample. Each test sample and control sample was analysed three times.

The results are depicted in Figures 18-23. As the figures demonstrate, both peptides contributed to a reduced polysaccharide matrix production in each of the microorganisms,

evident by the reduced Congo Red and Trypan Blue absorbance coming from cells incubated with the peptides as compared to cells that were not incubated with the peptides.

1. A cyclic peptide consisting of: a) the amino acid sequence $X^1-X^2-X^3-X^4-X^5-X^6-X^7-X^8-X^9-X^{10}-X^{11}-X^{12}$, wherein X^1 is S, N, T, K, R, H, E, I, Q or D; X^2 is V, I, L, Y, G, F or W; X^3 is H, N, Q, E, D or S; X^4 is S, P, A or T; X^5 is F, W or Y; X^6 is D, N, E or Q; X^7 is Y, F or W; X^8 is D, G or E; X^9 is W, F or Y; X^{10} is Y, F or W; X^{11} is N or Q; and X^{12} is V, I or L; and b) a cyclization linker between X^1 and X^{12} , wherein the peptide is not SVHSFDYDWYNV, and wherein the cyclic peptide has a total length of up to 50 amino acids.
2. The peptide according to claim 1, wherein the amino acid sequence is selected from the group consisting of SVHSWDYDWYNV, SVHSYDFDWYNV, TLQAFNYEWYQL, KYETFEYGWYNI, HGDSFQYEWYNL, SVHSFDWDWYNV, SVHSFDYDYNNV, SVHSFDYDFYNV, SVHSFDYDWFNV, SVHSFDYDWWNV, IFNPFYDWYNV, QWHSFDYDWYNV, and DVHPFDYDWYNV.
3. The cyclic peptide according to claim 1, wherein the cyclization linker comprises two amino acids independently selected from the group consisting of natural or non-natural amino acids.
4. The cyclic peptide according to claim 3, wherein the cyclization linker comprises two cysteines.
5. The cyclic peptide according to claim 4, wherein the cyclization linker consists of two cysteines.
6. A composition comprising a peptide according to claim 1.
7. A method of detaching a single cell organism from a surface or from other single cell organisms, comprising contacting said organism with a composition according to claim 6.
8. A method of dispersing a biofilm in water or detaching biofilm formation from a surface, comprising treating the water with or coating said surface with a composition according to claim 6.