Claims

1. Use of an *in vitro* produced peripheral blood mononuclear cell (PBMC) monolayer or bone-marrow cell monolayer, wherein natural-occurring cell-cell interactions and membrane integrity are maintained during formation of the monolayer, for determining whether the PBMC donor/bone-marrow cell donor suffers from a disease or is predisposed to suffer from a disease or in a diagnostic method for determining whether a subject suffering from or predisposed to a disease will respond or is responsive to treatment with a therapeutic agent, wherein the disease is a hematologic malignancy or a malignancy of myeloid and/or lymphoid tissue.

2. The use of claim 1, wherein the PBMC monolayer or bone-marrow cell monolayer is produced by a method for culturing PBMCs or bone-marrow cells comprising

   (a) isolating PBMCs from a blood sample or bone-marrow cells from bone-marrow; and
   (b) incubating PBMCs or bone-marrow cells at a density of about 100 cells per mm^2 to about 30000 cells per mm^2.

3. The use of claim 2, wherein the PBMCs or bone-marrow cells are incubated at a density of about 2000 cells per mm^2.

4. The use of claims 2 or 3, wherein PBMCs or bone-marrow cells are isolated from non-nucleated cells, wherein the PBMC sample or bone-marrow cell sample to be incubated contains less than about 100 non-nucleated cells per PBMC/bone-marrow cell.

5. The use of any one of claims 2 to 4, wherein the method further comprises a step (c), fixing said PBMCs or bone-marrow cells and/or fixing the formed PBMC monolayer or bone-marrow cell monolayer.

6. The use of claim 5, wherein the method further comprises, subsequent to step (c), a step (d) comprising adding a detectable label to the fixed PBMCs/bone-marrow cells.

7. The use of claim 6, wherein the detectable label is an antibody.

8. The use of any one of claims 2 to 7, wherein the method further comprises a step of adding a viability dye.

9. The use of claim 8, wherein the addition of a viability dye occurs prior to fixation of the formed PBMC monolayer or bone-marrow cell monolayer.

10. The use of any one of claims 2 to 9, wherein subsequent to isolation the PBMCs or bone-marrow cells are maintained/processed/analyzed at about 1g, i.e. 9.81 m/s^2.

11. The use of any one of claims 2 to 10, wherein subsequent to isolation the PBMCs or bone-marrow cells are not subjected to centrifugation or spinning.

12. A method for diagnosing a disease or predisposition to a disease in a PBMC donor/bone-marrow cell donor, comprising contacting the monolayer as defined in any one of claims 1 to 11 with a diagnostic agent, wherein the disease is a hematologic malignancy or a malignancy of myeloid and/or lymphoid tissue.

13. The use of any one of claims 1 to 11 or the method of claim 12, wherein the disease is a myeloproliferative disorder, inflammatory disorder, latent virus infection, cellular growth disorder, cellular chemotaxis disorder, metabolic disorder, or autoimmune disorder.

14. The use or method of claim 13, wherein the disease is leukemia or lymphoma.

~~15. A method for determining whether a subject suffering from or predisposed to a disease will respond or is responsive to treatment with a therapeutic agent comprising:~~

~~(a) isolating PBMCs/bone-marrow cells from a blood sample obtained from said subject;~~

~~(b) incubating said PBMCs/bone-marrow cells at a density of about 100 cells per mm^2 to about 30000 cells per mm^2;~~

~~(c) contacting said PBMCs/bone-marrow cells with said therapeutic agent; and~~

~~(d) assessing the response of the PBMCs/bone-marrow cells to the therapeutic agent,~~

~~wherein a PBMC monolayer or bone-marrow cell monolayer is formed during step (b), wherein natural-occurring cell-cell interactions and membrane integrity are maintained during formation of the monolayer and wherein the disease is a hematologic malignancy or a malignancy of myeloid and/or lymphoid tissue.~~

~~16. The method of claim 15, wherein the PBMCs/bone-marrow cells are incubated at a density of about 2000 cells per mm^2.~~

1. Use of an *in vitro* produced peripheral blood mononuclear cell (PBMC) monolayer or bone-marrow cell monolayer, wherein natural-occurring cell-cell interactions and membrane integrity are maintained during formation of the monolayer, for determining whether the PBMC donor/bone-marrow cell donor suffers from a disease or is predisposed to suffer from a disease or in a diagnostic method for determining whether a subject suffering from or predisposed to a disease will respond or is responsive to treatment with a therapeutic agent, wherein the disease is a hematologic malignancy, a malignancy of myeloid and/or lymphoid tissue, a myeloproliferative disorder, an inflammatory disorder, a latent virus infection, a cellular growth disorder, a cellular chemotaxis disorder, a metabolic disorder, or an autoimmune disorder.

2. The use of claim 1, wherein the PBMC monolayer or bone-marrow cell monolayer is produced by a method for culturing PBMCs or bone-marrow cells comprising

(a) isolating PBMCs from a blood sample or bone-marrow cells from bone-marrow; and

(b) incubating PBMCs or bone-marrow cells at a density of 100 cells per mm^2 to 30000 cells per mm^2.

3. The use of claim 2, wherein the PBMCs or bone-marrow cells are incubated at a density of about 2000 cells per mm^2.

4. The use of claims 2 or 3, wherein PBMCs or bone-marrow cells are isolated from non-nucleated cells, wherein the PBMC sample or bone-marrow cell sample to be incubated contains less than about 100 non-nucleated cells per PBMC/bone-marrow cell.

5. The use of any one of claims 2 to 4, wherein the method further comprises a step (c), fixing said PBMCs or bone-marrow cells and/or fixing the formed PBMC monolayer or bone-marrow cell monolayer.

6. The use of claim 5, wherein the method further comprises, subsequent to step (c), a step (d) comprising adding a detectable label to the fixed PBMCs/bone-marrow cells.

7. The use of claim 6, wherein the detectable label is an antibody.

8. The use of any one of claims 2 to 7, wherein the method further comprises a step of adding a viability dye.

9. The use of claim 8, wherein the addition of a viability dye occurs prior to fixation of the formed PBMC monolayer or bone-marrow cell monolayer.

10. The use of any one of claims 2 to 9, wherein subsequent to isolation the PBMCs or bone-marrow cells are maintained/processed/analyzed at about 1g, i.e. 9.81 m/s^2.

11. The use of any one of claims 2 to 10, wherein subsequent to isolation the PBMCs or bone-marrow cells are not subjected to centrifugation or spinning.

12. A method for diagnosing a disease or predisposition to a disease in a PBMC donor/bone-marrow cell donor, comprising contacting the monolayer as defined in any one of claims 1 to 11 with a diagnostic agent, wherein the disease is a hematologic malignancy, a malignancy of myeloid and/or lymphoid tissue, a myeloproliferative disorder, an inflammatory disorder, a latent virus infection, a cellular growth disorder, a cellular chemotaxis disorder, a metabolic disorder, or an autoimmune disorder.

13. The use or method of any one of claims 1 to 12, wherein the disease is leukemia or lymphoma.

14. A method for determining whether a subject suffering from or predisposed to a disease will respond or is responsive to treatment with a therapeutic agent comprising:

(a) isolating PBMCs/bone-marrow cells from a blood sample obtained from said subject;
(b) incubating said PBMCs/bone-marrow cells at a density of 100 cells per mm^2 to 30000 cells per mm^2;
(c) contacting said PBMCs/bone-marrow cells with said therapeutic agent; and
(d) assessing the response of the PBMCs/bone-marrow cells to the therapeutic agent,

wherein a PBMC monolayer or bone-marrow cell monolayer is formed during step (b), wherein natural-occurring cell-cell interactions and membrane integrity are maintained during formation of the monolayer and wherein the

disease is a hematologic malignancy or a malignancy of myeloid and/or lymphoid tissue.

15. The method of claim 14, wherein the PBMCs/bone-marrow cells are incubated at a density of about 2000 cells per mm^2.