

SPECIFICATION

METHOD FOR NUCLEIC ACID REPLICATION AND NOVEL ARTIFICIAL BASE PAIRS

TECHNICAL FIELD

[0001] The present application claims priority to Japanese Patent Application Nos. 2005-356883 (filed on December 9, 2005) and 2006-189806 (filed on July 10, 2006), the entire contents of which are incorporated herein by reference.

[0002] The present invention relates to a method for nucleic acid replication and novel artificial base pairs.

BACKGROUND ART

[0003] Nucleic acids are amplified and act as catalysts and ligands through self-complementarity of A-T(U) and G-C base pairs. However, unlike 20 different amino acids in natural proteins, natural nucleic acids are composed of nucleotides consisting of only 4 different bases. This number limit restricts the functions of DNA and RNA molecules. Unnatural base pair systems provide a resolution to this problem because they increase the types of nucleic acid bases to allow expansion of genetic information (Non-patent Documents 1-5). Unnatural base pairs are required to have highly specific complementarity which allows site-specific incorporation of special nucleotide analogs into DNA and RNA through polymerase-catalyzed reactions. If this requirement is achieved, current genetic engineering technology, which is limited by the number of naturally-occurring bases, can be replaced with a novel technology using unnatural base pair

systems.

[0004] The first attempt to create unnatural base pairs was made by Benner et al (Non-patent Documents 6-7). They developed some unnatural base pairs, including isoguanine-isocytosine (isoG-isoC) and xanthosine-diaminopyrimidine, based on different hydrogen bonding patterns than those of natural base pairs. Recently, these unnatural base pairs have been applied to PCR amplification (Non-patent Documents 8-9) and sequence analysis (Non-patent Document 10) of DNA fragments containing these base pairs. However, the fidelity is relatively not high and/or complicated procedures are required. In addition to these problems, 2-aminopyrimidine analogs such as isoC and diaminopyrimidine are not recognized as substrates by T7 RNA polymerase. Thus, these base pairs are of limited use.

[0005] Subsequently, Kool et al. synthesized hydrophobic bases having shapes similar to those of natural bases, but lacking the ability to form a hydrogen bond during base pairing (Non-patent Documents 11-12). These hydrophobic bases were selectively recognized by DNA polymerases, suggesting that geometric shape complementarity between pairing bases is more important during replication, rather than hydrogen bonding interaction. Recently, a series of hydrophobic base pairs have been developed by Romesberg et al. and introduced into DNA in a complementary manner by the action of the Klenow fragment of *E. coli*-derived DNA polymerase I (Non-patent Documents 13-15). However, these hydrophobic bases did not conform to shape complementarity during replication, and

non-specific incorporation occurred between hydrophobic bases (Non-patent Document 14). Moreover, there is no report of these base pairs functioning during transcription.

[0006] By combining the ideas of hydrogen bonding pattern and shape complementarity, the inventors of the present invention developed unnatural base pairs between 2-amino-6-(2-thienyl)purine (s) and 2-oxopyridine (y) (Non-patent Documents 16-17) as well as between 2-amino-6-(2-thiazolyl)purine (v) and y (Non-patent Document 18). The bulky substituents at the 6-position of s and v efficiently prevented undesirable base pairing (non-cognate pairing) with a natural base, and a substrate (nucleoside 5'-triphosphate) of y or modified y was introduced in a site-specific manner into RNA opposite s or v in the template by the action of T7 RNA polymerase. This specific transcription is available for practical use as a means for developing functional RNA molecules (Non-patent Documents 19-21), but the selectivity of s-y and v-y base pairings during replication is not notably higher than that during transcription (Non-patent Documents 16 and 18).

[0007] To solve the problems stated above, there is a demand for a novel artificial base pair showing excellent efficiency and selectivity during replication and transcription (for design of functional nucleic acids) or during all of replication, transcription and translation (for design of functional proteins).

[0008] The following documents are listed as reference documents, the entire contents of which are incorporated herein by reference.

Patent Document 1: WO2001/005801

Patent Document 2: WO2004/007713

Patent Document 3: WO2005/026187

Patent Document 4: Japanese Patent Application

No. 2005-226492

Non-patent Document 1: Benner, S. A., Burgstaller, P., Battersby, T. R. & Jurczyk, S. in *The RNA World* (eds Gesteland, R. F., Cech, T. R. & Atkins, J. F.) 163-181 (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1999).

Non-patent Document 2: Henry, A. A. & Romesberg, F. E. Beyond A, C, G and T: augmenting nature's alphabet. *Curr. Opin. Chem. Biol.* 7, 727-733 (2003).

Non-patent Document 3: Moser, M. J. & Prudent, J. R. Enzymatic repair of an expanded genetic information system. *Nucleic Acids Res.* 31, 5048-5053 (2003).

Non-patent Document 4: Bergstrom, D. E. Orthogonal base pairs continue to evolve. *Chem. Biol.* 11, 18-20 (2004).

Non-patent Document 5: Benner, S. A. & Sismour, A. M. Synthetic biology. *Nat. Rev.* 6, 533-543 (2005).

Non-patent Document 6: Piccirilli, J. A., Krauch, T., Moroney, S. E. & Benner, S. A. Enzymatic incorporation of a new base pair into DNA and RNA extends the genetic alphabet. *Nature* 343, 33-37 (1990).

Non-patent Document 7: Switzer, C. Y., Moroney, S. E. & Benner, S. A. Enzymatic recognition of the base pair between isocytidine and isoguanosine. *Biochemistry* 32, 10489-10496 (1993).

Non-patent Document 8: Sismour, A. M. et al. PCR amplification of DNA containing non-standard base pairs by variants of reverse transcriptase from Human Immunodeficiency Virus-1. *Nucleic Acids Res.* 32, 728-735 (2004).

Non-patent Document 9: Johnson, S. C., Sherrill, C. B., Marshall, D. J., Moser, M. J. & Prudent, J. R. A third base pair for the polymerase chain reaction: inserting isoC and isoG. *Nucleic Acids Res.* 32, 1937-1941 (2004).

Non-patent Document 10: Ahle, J. D., Barr, S., Chin, A. M. & Battersby, T. R. Sequence determination of nucleic acids containing 5-methylisocytosine and isoguanine: identification and insight into polymerase replication of the non-natural nucleobases. *Nucleic Acids Res.* 33, 3176-3184 (2005).

Non-patent Document 11: Morales, J. C. & Kool, E. T. Efficient replication between non-hydrogen-bonded nucleoside shape analogs. *Nat. Struct. Biol.* 5, 950-954 (1998).

Non-patent Document 12: Kool, E. T., Morales, J. C. & Guckian, K. M. Mimicking the structure and function of DNA: Insights into DNA stability and replication. *Angew. Chem. Int. Ed.* 39, 990-1009 (2000).

Non-patent Document 13: McMinn, D. L. et al. Efforts toward expansion of the genetic alphabet: DNA polymerase recognition of a highly stable, self-pairing hydrophobic base. *J. Am. Chem. Soc.* 121, 11585-11586 (1999).

Non-patent Document 14: Wu, Y. et al. Efforts toward expansion of the genetic alphabet: optimization of interbase hydrophobic interactions. *J. Am. Chem. Soc.* 122, 7621-7632 (2000).

Non-patent Document 15: Ogawa, A. K. et al. Efforts toward the expansion of the genetic alphabet: Information storage and replication with unnatural hydrophobic base pairs. J. Am. Chem. Soc. 122, 3274-3287 (2000).

Non-patent Document 16: Fujiwara, T., Kimoto, M., Sugiyama, H., Hirao, I. & Yokoyama, S. Synthesis of 6-(2-thienyl)purine nucleoside derivatives that form unnatural base pairs with pyridin-2-one nucleosides. Bioorg. Med. Chem. Lett. 11, 2221-2223 (2001).

Non-patent Document 17: Hirao, I. et al. An unnatural base pair for incorporating amino acid analogs into proteins. Nat. Biotechnol. 20, 177-182 (2002).

Non-patent Document 18: Mitsui, T., Kimoto, M., Harada, Y., Yokoyama, S. & Hirao, I. An efficient unnatural base pair for a base-pair-expanded transcription system. J. Am. Chem. Soc. 127, 8652-8658 (2005).

Non-patent Document 19: Kimoto M. et al. Site-specific incorporation of a photo-crosslinking component into RNA by T7 transcription mediated by unnatural base pairs. Chem. Biol. 11, 47-55 (2004).

Non-patent Document 20: Moriyama, K., Kimoto, M., Mitsui, T., Yokoyama, S. & Hirao, I. Site-specific biotinylation of RNA molecules by transcription using unnatural base pairs. Nucleic Acids Res. 33, e129 (2005).

Non-patent Document 21: Kawai, R. et al. Site-specific fluorescent labeling of RNA molecules by specific transcription using unnatural base pairs. J. Am. Chem. Soc. in press.

Non-patent Document 22: Matray, T.J. & Kool, E.T. A specific partner for abasic damage in DNA. *Nature* 399, 704-708 (1999).

Non-patent Document 23: Doubl  , S., Tabor, S., Long, A. M., Richardson, C. C. & Elenberger, T. Crystal structure of a bacteriophage T7 DNA replication complex at 2.2   resolution. *Nature* 391, 251-258 (1998).

Non-patent Document 24: Kiefer, J. R., Mao, C., Braman, J. C. & Beese, L. S. Visualizing DNA replication in a catalytically active *Bacillus* DNA polymerase crystal. *Nature* 391, 304-307 (1998).

Non-patent Document 25: Morales, J. C. & Kool, E. T. Functional hydrogen-bonding map of the minor groove binding tracks of six DNA polymerases. *Biochemistry* 39, 12979-12988 (2000).

Non-patent Document 26: Mitsui, T. et al. An unnatural hydrophobic base pair with shape complementarity between pyrrole-2-carbaldehyde and 9-methylimidazo[(4,5)-b]pyridine. *J. Am. Chem. Soc.* 125, 5298-5307 (2003).

Non-patent Document 27: Morales, J. C. & Kool, E. T. Minor groove interactions between polymerase and DNA: More essential to replication than Watson-Crick hydrogen bonds? *J. Am. Chem. Soc.* 121, 2323-2324 (1999).

Non-patent Document 28: Hirao, I. et al. A two-unnatural-base-pair system toward the expansion of the genetic code. *J. Am. Chem. Soc.* 126, 13298-13305 (2004).

Non-patent Document 29: Tae, E. L., Wu, Y., Xia, G., Schultz, P. G. & Romesberg, F. E. Efforts toward expansion of

the genetic alphabet: Replication of DNA with three base pairs. J. Am. Chem. Soc. 123,7439-7440 (2001).

Non-patent Document 30: Petruska, J. et al. Comparison between DNA melting thermodynamics and DNA polymerase fidelity. Proc. Natl. Acad. Sci. USA 85, 6252-6256 (1988).

Non-patent Document 31: Goodman, M. F., Creighton, S., Bloom, L. B. & Petruska, J. Biochemical basis of DNA replication fidelity. Crit. Rev. Biochem. Mol. Biol. 28, 83-126 (1993).

Non-patent Document 32: Kimoto, M., Yokoyama, S. & Hirao, I. A quantitative, non-radioactive single-nucleotide insertion assay for analysis of DNA replication fidelity by using an automated DNA sequencer. Biotechnol. Lett. 26, 999-1005 (2004).

Non-patent Document 33: Ohtsuki, T. et al. Unnatural base pair for specific transcription. Proc. Natl. Acad. Sci. USA 98, 4922-4925 (2001).

Non-patent Document 34: Mitsui, T., Kimoto, M., Sato, A., Yokoyama, S. & Hirao, I. An unnatural hydrophobic base, 4-propynylpyrrole-2-carbaldehyde, as an efficient pairing partner of 9-methylimidazo[(4,5)-b]pyridine. Bioorg. Med. Chem. Lett. 13, 4515-4518 (2003).

Non-patent Document 35: Cha, R. S. & Thilly W, G. in PCR Primer (eds Dieffenbach, C. W. & Dveksler, G. S.) 37-51 (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1995).

Non-patent Document 36: Himeno, H., Hasegawa, T., Ueda, T., Watanabe, K. & Shimizu, M. Conversion of aminoacylation

specificity from tRNA^{Tyr} to tRNA^{Ser} in vitro. Nucleic Acids Res. 18, 6815-6819 (1990).

Non-patent Document 37: Bedouelle, H. Recognition of tRNA^{Tyr} by tyrosyl-tRNA synthetase. Biochimie 72, 589-598 (1990).

Non-patent Document 38: Mulder, B. A. et al. Nucleotide modification at the γ -phosphate leads to the improved fidelity of HIV-1 reverse transcriptase. Nucleic Acids Res. 33, 4865-4873 (2005).

Non-patent Document 39: Mitsui, T., Kitamura, A., Kimoto, M., To, T., Sato, A., Hirao, I. & Yokoyama, S. An unnatural hydrophobic base pair with shape complementarity between pyrrole-2-carbaldehyde and 9-methylimidazo[(4,5)-b]pyridine. J. Am. Chem. Soc. 125, 5298-5307 (2003).

Non-patent Document 40: Mitsui, T., Kimoto, M., Sato, A., Yokoyama, S. & Hirao, I. An unnatural hydrophobic base, 4-propynylpyrrole-2-carbaldehyde, as an efficient pairing partner of 9-methylimidazo[(4,5)-b]pyridine. Bioorg. Med. Chem. Lett. 13, 4515-4518 (2003).

Non-patent Document 41: De Roos, K. B. & Salemink, C. A., Deazapurine derivatives. V, A new synthesis of 1- and 3-deaza-adenine and related compound. Recueil. 88, 1263-1274 (1963).

Non-patent Document 42: Rolland, V., Kotera, M. & Lhomme, J. Convenient preparation of 2-deoxy 3,5-di-O-p-toluoyl- α -D-erythro-pentofuranosyl chloride. Synthetic Commun. 27, 3505-3511 (1997).

Non-patent Document 43: Ludwig, J. & Eckstein, F. Rapid and efficient synthesis of 5'-O-(1-thiotriphosphates), 5'-O-

triphosphates and 2',3'-cyclophosphorothioates using 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one. J. Org. Chem. 54, 631-635 (1989).

Non-patent Document 44: Stevens, J. D., Ness, R. K. & Fletcher, Jr, H. G. Syntheses with partially benzylated sugars. XI. Studies on the synthesis of the anomeric 5,6-dimethyl-1-D-ribofuranosylbenzimidazole (Ribazoles). Comparison of the condensation of 2,3,5-tri-O-benzoyl-D-ribofuranosyl bromide and 2,3,5-tri-O-benzoyl-D-ribofuranosyl chloride with 5,6-dimethylbenzimidazole. J. Org. Chem. 33, 1806-1810 (1968).

Non-patent Document 45: Kovacs, T. & Otvos, L. Simple synthesis of 5-vinyl- and 5-ethynyl-2'-deoxyuridine-5'-triphosphates. Tetrahedron Lett. 29, 4525-4528 (1988).

Non-patent Document 46: Ti, G. S., Gaffney, B. L. & Jones, R. A., Transient protection: Efficient One-Flask Syntheses of Protected Deoxynucleosides. J. Am. Chem. Soc. 104, 1316-1319 (1982).

Non-patent Document 47: Stumber, M., Herrmann, C., Wohlgemuth, S., Kalbitzer, H. R., Jahn, W. & Geyer, M. Synthesis, characterization and application of two nucleoside triphosphate analogues, GTP γ NH₂ and GTP γ F. Eur. J. Biochem. 269, 3270-3278 (2002).

Non-patent Document 48: Knorre, D. G., Kurbatov, V. A. & Samukov, V. V. General method for the synthesis of ATP gamma-derivatives. FEBS Lett. 70, 105-108 (1976).

Non-patent Document 49: K. J. Morgan and D. P. Morrey. Nitropyrrole-I, The preparation and properties of 2- and 3-

nitropyrrole, Tetrahedron, 22, 57-62, 1966.

DISCLOSURE OF THE INVENTION

PROBLEMS TO BE SOLVED BY THE INVENTION

[0009] The present invention aims to provide the following embodiments 1-27.

[0010] Embodiment 1: A method for replicating a nucleic acid, wherein a deoxyribonucleoside 5'-triphosphate, in which the hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group, is used as a substrate during replication reaction (preferably in combination with usual substrates).

[0011] Embodiment 2: The method according to embodiment 1, wherein the substituent is an amino group.

[0012] Embodiment 3: The method according to embodiment 1 or 2, wherein a DNA polymerase having exonuclease activity is used during the replication reaction.

[0013] Embodiment 4: The method according to any one of embodiments 1 to 3, wherein the polymerase having exonuclease activity is selected from the group consisting of Klenow fragment, T4 DNA polymerase and thermophilic DNA polymerase (e.g., *Thermococcus litoralis*-derived Vent DNA polymerase), each having 3'→5' exonuclease activity.

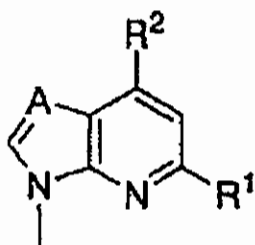
[0014] Embodiment 5: The method according to any one of embodiments 1 to 4, wherein the deoxyribonucleoside 5'-triphosphate used as a substrate has an unnatural base.

[0015] Embodiment 6: The method according to any one of

embodiments 1 to 4, wherein the deoxyribonucleoside 5'-triphosphate used as a substrate has a natural base.

[0016] Embodiment 7: The method according to any one of embodiments 1 to 5, wherein the deoxyribonucleoside 5'-triphosphate used as a substrate has a base represented by the following formula 1:

[0017] [Formula 1]



[0018] [wherein

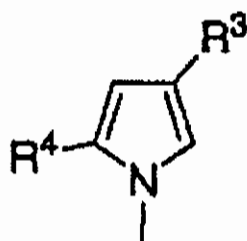
R¹ is hydrogen or an amino group,

R² is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH].

[0019] Embodiment 8: The method according to any one of embodiments 1 to 5, wherein the deoxyribonucleoside 5'-triphosphate used as a substrate has a base represented by the following formula 2:

[0020] [Formula 2]



[0021] [wherein

R^3 is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C_1 - C_3 alkyl group, a substituted or unsubstituted C_2 - C_3 alkenyl group, or a substituted or unsubstituted C_2 - C_3 alkynyl group, and

R^4 is a formyl group or a nitro group].

[0022] Embodiment 9: A deoxyribonucleoside 5'-triphosphate, in which the hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group.

[0023] Embodiment 10: A deoxyribonucleoside 5'-triphosphate having an unnatural base, in which the hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group.

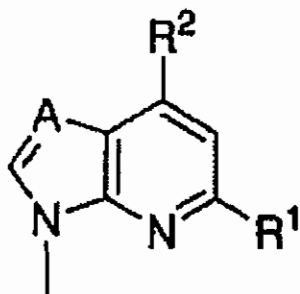
[0024] Embodiment 11: A deoxyribonucleoside 5'-triphosphate having a natural base, in which the hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group.

[0025] Embodiment 12: The use of a deoxyribonucleoside 5'-triphosphate as a substrate in the method for replicating a nucleic acid according to any one of embodiments 1 to 8, in which the hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting

of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group.

[0026] Embodiment 13: A nucleic acid, in which a nucleotide having a base represented by the following formula 1:

[0027] [Formula 3]



[0028] [wherein

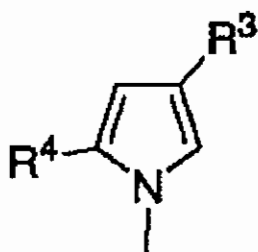
R¹ is hydrogen or an amino group,

R² is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH]

forms a base pair with a nucleotide having a base represented by the following formula 2:

[0029] [Formula 4]



[0030] [wherein

R³ is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C₁-C₃ alkyl group, a substituted or unsubstituted C₂-C₃ alkenyl group, or a substituted or

unsubstituted C₂-C₃ alkynyl group, and

R⁴ is a formyl group or a nitro group].

[0031] Embodiment 14: The nucleic acid according to embodiment 13, wherein the base of formula 1 is selected from the group consisting of:

A1) a 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl group;

A2) a 7-(2-thiazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group;

A3) a 7-(1H-2-imidazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group;

A4) a 5-amino-7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl group;

A5) a 5-amino-7-(2-thiazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group;

A6) a 5-amino-7-(1H-2-imidazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group;

A7) a 4-(2-thienyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group;

A8) a 4-(2-thiazolyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group;

A-9) a 4-(1H-2-imidazolyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group;

A-10) a 6-amino-4-(2-thienyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group;

A-11) a 6-amino-4-(2-thiazolyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group; and

A-12) a 6-amino-4-(1H-2-imidazolyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group.

[0032] Embodiment 15: The nucleic acid according to

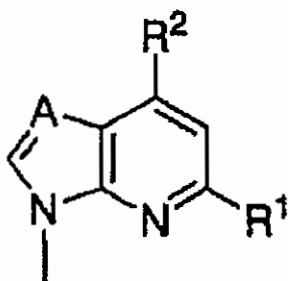
embodiment 13, wherein the base of formula 2 is selected from the group consisting of:

- B1) a 2-formyl-1H-pyrrol-1-yl group;
- B2) a 2-formyl-4-iodo-1H-pyrrol-1-yl group;
- B3) a 2-formyl-4-methyl-1H-pyrrol-1-yl group;
- B4) a 2-formyl-4-(1-propyn-1-yl)-1H-pyrrol-1-yl group;
- B5) a 2-formyl-4-(2-substituted aminovinyl)-1H-pyrrol-1-yl group;
- B6) a 2-formyl-4-(3-substituted amino-1-propyn-1-yl)-1H-pyrrol-1-yl group;
- B7) a 2-nitro-1H-pyrrol-1-yl group;
- B8) a 2-nitro-4-iodo-1H-pyrrol-1-yl group;
- B9) a 2-nitro-4-methyl-1H-pyrrol-1-yl group;
- B10) a 2-nitro-4-(1-propyn-1-yl)-1H-pyrrol-1-yl group;
- B11) a 2-nitro-4-(2-substituted aminovinyl)-1H-pyrrol-1-yl group; and
- B12) a 2-nitro-4-(3-substituted amino-1-propyn-1-yl)-1H-pyrrol-1-yl group.

[0033] Embodiment 16: The nucleic acid according to any one of embodiments 13 to 15, which forms a base pair(s) in the step of transcription, reverse transcription, replication or translation.

[0034] Embodiment 17: A method for preparing a nucleic acid containing a nucleotide having a base represented by the following formula 1:

[0035] [Formula 5]



[0036] [wherein

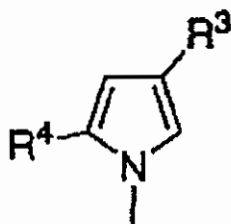
R¹ is hydrogen or an amino group,

R² is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH],

wherein the method comprises effecting transcription, reverse transcription or replication by using, as a template, a nucleic acid containing a nucleotide having a base represented by the following formula 2:

[0037] [Formula 6]



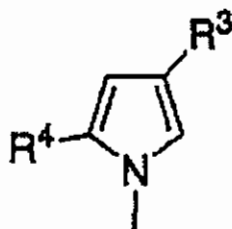
[0038] [wherein

R³ is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C₁-C₃ alkyl group, a substituted or unsubstituted C₂-C₃ alkenyl group, or a substituted or unsubstituted C₂-C₃ alkynyl group, and

R^4 is a formyl group or a nitro group],
whereby the nucleotide having a base of formula 1 is
incorporated at a site complementary to the nucleotide having
a base of formula 2.

[0039] Embodiment 18: A method for preparing a nucleic acid
containing a nucleotide having a base represented by the
following formula 2:

[0040] [Formula 7]



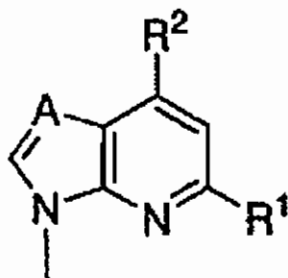
[0041] [wherein

R^3 is a group selected from hydrogen, an iodo group, a
substituted or unsubstituted C_1 - C_3 alkyl group, a substituted
or unsubstituted C_2 - C_3 alkenyl group, or a substituted or
unsubstituted C_2 - C_3 alkynyl group, and

R^4 is a formyl group or a nitro group],

wherein the method comprises effecting transcription,
reverse transcription or replication by using, as a template,
a nucleic acid containing a nucleotide having a base
represented by the following formula 1:

[0042] [Formula 8]



[0043] [wherein

R^1 is hydrogen or an amino group,

R^2 is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH],

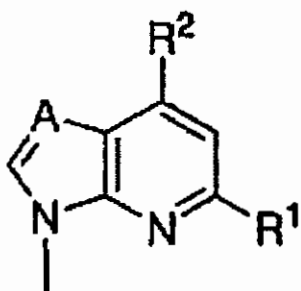
whereby the nucleotide having a base of formula 2 is incorporated at a site complementary to the nucleotide having a base of formula 1.

[0044] Embodiment 19: A nucleic acid containing a nucleotide having a base of formula 1 and/or formula 2, which is prepared by the method according to embodiment 17 or 18.

[0045] Embodiment 20: The nucleic acid according to embodiment 19, which is tRNA, mRNA, antisense DNA or RNA, a ribozyme, an aptamer or siRNA.

[0046] Embodiment 21: A ribonucleoside 5'-triphosphate having a base represented by the following formula 1:

[0047] [Formula 9]



[0048] [wherein

R^1 is hydrogen or an amino group,

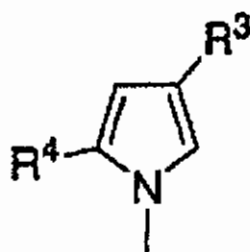
R^2 is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a

substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH].

[0049] Embodiment 22: A ribonucleoside 5'-triphosphate having a base represented by the following formula 2:

[0050] [Formula 10]



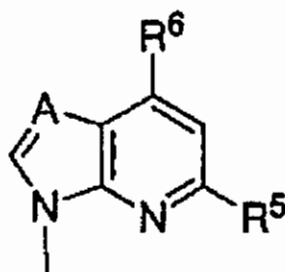
[0051] [wherein

R³ is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C₁-C₃ alkyl group, a substituted or unsubstituted C₂-C₃ alkenyl group, or a substituted or unsubstituted C₂-C₃ alkynyl group, and

R⁴ is a formyl group or a nitro group].

[0052] Embodiment 23: A 5'-O-(4,4'-dimethoxytrityl)-3'-O-(2-cyanoethyl N,N-diisopropylphosphoroamidite) deoxyribonucleoside having a base represented by the following formula 3:

[0053] [Formula 11]



[0054] [wherein

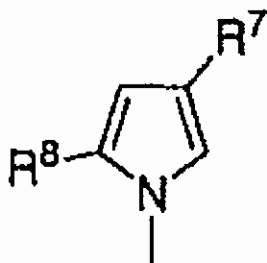
R⁵ is hydrogen or a substituted amino group,

R⁶ is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH].

[0055] Embodiment 24: A 5'-O-(4,4'-dimethoxytrityl)-3'-O-(2-cyanoethyl N,N-diisopropylphosphoroamidite) deoxyribonucleoside having a base represented by the following formula 4:

[0056] [Formula 12]



[0057] [wherein

R⁷ is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C₁-C₃ alkyl group, a substituted or unsubstituted C₂-C₃ alkenyl group, or a substituted or unsubstituted C₂-C₃ alkynyl group, and

R⁸ is a formyl group or a nitro group,

excluding the case where R⁷ is hydrogen or a 1-propynyl group and R⁸ is a formyl group].

[0058] Embodiment 25: A nucleic acid containing a nucleotide having a base represented by formula 2, which is prepared by the method according to embodiment 18, wherein the substituent R³ in formula 2 is a C₁-C₃ alkyl group, a C₂-C₃ alkenyl group or a C₂-C₃ alkynyl group, each being substituted with biotin or a fluorescent molecule.

[0059] Embodiment 26: The ribonucleoside 5'-triphosphate according to embodiment 22, wherein R^3 is a C_1 - C_3 alkyl group, a C_2 - C_3 alkenyl group or a C_2 - C_3 alkynyl group, each being substituted with biotin or a fluorescent molecule.

[0060] Embodiment 27: The 5'-O-(4,4'-dimethoxytrityl)-3'-O-(2-cyanoethyl N,N-diisopropylphosphoroamidite) deoxyribonucleoside according to embodiment 24, wherein R^7 is a C_1 - C_3 alkyl group, a C_2 - C_3 alkenyl group or a C_2 - C_3 alkynyl group, each being substituted with biotin or a fluorescent molecule.

MEANS FOR SOLVING THE PROBLEMS

[0061] As a result of extensive and intensive efforts made to solve the problems stated above, the inventors of the present invention have arrived at the present invention.

[0062] Development of unnatural base pair systems of the present invention

To obtain an artificial base pair showing excellent efficiency and selectivity during reactions of replication and transcription, the inventors of the present invention have studied base pairs constructed by combining several unnatural base pairs which had been developed on their own.

[0063] The inventors of the present invention have found that a combination between 2-amino-6-(2-thienyl)purin-9-yl (s) (Non-patent Documents 17 and 33) and 2-formyl-1H-pyrrol-1-yl (Pa) (Non-patent Document 26) is highly selective and efficient during transcription (Patent Document 4: not yet published). In the present invention, the inventors have further prepared a 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl

group (Ds) from s through CH substitution for one of the two N atoms in the 6-membered ring of the purine ring and replacement of the amino group at the 2-position with hydrogen, i.e., through deaza modification and deamination, and have studied the fidelity of artificial base pairing between Ds and Pa. As a result, the inventors have found that artificial base pairing between Ds and Pa is highly selective and efficient during replication and transcription, regardless of which of them serves as a template or substrate, thereby arriving at the present invention.

[0064] More specifically, it has been found that hydrophobic base pairings between 7-(2-thienyl)-imidazo[4,5-b]pyridine (Ds) and pyrrole-2-carbaldehyde (Pa) as well as between Ds and 4-propynylpyrrole-2-carbaldehyde (Pa') (Figure 1a) are highly selective during *in vitro* replication and transcription (Figure 1c). During replication, the inventors of the present invention have used usual 5'-triphosphate substrates in combination with modified 5'-triphosphate substrates, i.e., 5'- γ -amidotriphosphates (Figure 1b), and have further used DNA polymerases having 3'→5' exonuclease activity to thereby achieve high selectivity which allows PCR amplification of DNA fragments containing a Ds-Pa base pair. Moreover, these unnatural bases have been introduced into RNA in a complementary manner during normal transcription with T7 RNA polymerase.

[0065] More specifically, the inventors of the present invention have designed this Ds-Pa base pair in consideration of the following two ideas: 1) hydrophobic bases whose shape

differs from that of natural bases are used with the aim of improving the selectivity of base pairing (Non-patent Documents 12 and 22); and 2) proton acceptor groups required for interaction with polymerases (Non-patent Documents 23-25) are further provided, including the nitrogen at the 4-position of Ds (corresponding to the 3-position of A and G) and the aldehyde group of Pa (corresponding to the keto group at the 2-position of C and T) (Figure 1a).

[0066] Hydrophobic base pairs have a fatal problem in that non-cognate base pairing efficiently occurs between hydrophobic bases, whose are not complementary in shape (e.g., Ds-Ds pairing) (Non-patent Documents 14 and 29). To test the selectivity of Ds-Pa base pairing during replication, the inventors of the present invention have studied the base pairing capacity between substrate and template in a single nucleotide insertion experiment using the Klenow fragment lacking exonuclease activity (KF exo⁻) (Figure 2a) (Non-patent Documents 30-32).

[0067] The substrates (dDsTP and dPaTP) and template DNA containing Ds or Pa used in the experiment were chemically synthesized (Examples I and II). The results from the experiment indicated that Ds-Pa base pairing and A-T base pairing each showed higher selectivity than the pairing selectivity of the other, non-cognate, base pairing combinations (Figure 2b, Tables 1 and 2).

[0068] Table 1 Experiment of single-nucleotide insertion into template DNA with the Klenow fragment

[0069] [Table 1]

primer temp35N-2		5'-ACTCACTATAGGGAGCTTCT 3'-TATTATGCTGAGTGATATCCCTCGAAGANAGAGCT			
Entry	Template (N)	Nucleoside triphosphate	K_M (μM)	V_{max} (% min ⁻¹)	Efficiency (V_{max}/K_M) ^d
1	Pa	dDsTP	26 (12) ^b	28 (5)	1.1×10^6
2	Pa	dDsTP _N	180 (20)	12 (1)	6.7×10^4
3	Pa	dATP	490 (260)	21 (6)	4.3×10^4
4	Pa	dATP _N	1200 (400)	2.2 (1.3)	1.8×10^3
5	Pa	dGTP	480 (140)	0.42 (0.09)	8.8×10^2
6	Pa	dCTP	n.d. ^c	n.d.	-
7	Pa	dTTP	880 (530)	0.097 (0.025)	1.1×10^2
8	Pa	dPaTP	380 (90)	0.56 (0.09)	1.5×10^3
9	Pa'	dDsTP	24 (2)	21 (6)	8.8×10^5
10	Pa'	dDsTP _N	230 (50)	13 (5)	5.7×10^4
11	Pa'	dATP	570 (240)	21 (11)	3.7×10^4
12	Pa'	dATP _N	800 (400)	2.4 (1.3)	3.0×10^3
13	Pa'	dGTP	800 (170)	0.44 (0.09)	5.5×10^2
14	Pa'	dCTP	n.d.	n.d.	-
15	Pa'	dTTP	1400 (100)	0.14 (0.04)	1.0×10^2
16	Pa'	dPa'TP	190 (90)	7.8 (2.4)	4.1×10^4
17	A	dDsTP	33 (9)	1.2 (0.1)	3.6×10^4
18	G	dDsTP	37 (6)	1.5 (0.3)	4.1×10^4
19	C	dDsTP	47 (26)	1.3 (0.1)	2.8×10^4
20	T	dDsTP	45 (14)	3.9 (0.3)	8.7×10^4
21	A	dDsTP _N	270 (150)	0.52 (0.16)	1.9×10^3
22	G	dDsTP _N	300 (80)	0.58 (0.06)	1.9×10^3
23	C	dDsTP _N	440 (130)	0.64 (0.15)	1.5×10^3
24	T	dDsTP _N	480 (110)	1.7 (0.2)	3.5×10^3
25	T	dATP	0.81 (0.44)	3.3 (1.8)	4.1×10^6
26	T	dATP _N	13 (11)	3.2 (1.7)	2.5×10^5
27	C	dATP	500 (90)	2.3 (0.8)	4.6×10^3
28	C	dATP _N	590 (110)	0.20 (0.04)	3.4×10^2
29	C	dGTP	2.3 (0.1)	16 (4)	7.0×10^6
30	T	dGTP	420 (20)	1.2 (0.1)	2.9×10^3

[0070] a. Assays were carried out at 37°C for 1 to 35 minutes using 5 μM template-primer duplex, 5-50 nM enzyme and 0.3-1500 μM nucleoside 5'-triphosphate in a solution (10 μl) containing 50 mM Tris-HCl (pH 7.5), 10 mM MgCl₂, 1 mM DTT and 0.05 mg/ml bovine serum albumin. Each parameter was

averaged from 3 to 8 data sets.

[0071] b. Standard deviations are given in parentheses.

[0072] c. Minimal inserted products (<2%) were detected after incubation for 20 minutes with 1500 μ M nucleoside 5'-triphosphate and 50 nM enzyme.

[0073] d. The units of this term are % $\text{min}^{-1}\text{M}^{-1}$.

[0074] Table 2 Experiment of single-nucleotide insertion into template DNA with the Klenow fragment

[0075] [Table 2-1]

primer temp35N-1		5'-ACTCACTATAGGGAGGAAGA 3'-TATTATGCTGAGTGATATCCCTCCTTCTNCTCGA			
Entry	Template (N)	Nucleoside triphosphate	K_M (μ M)	V_{max} (% min^{-1})	Efficiency (V_{max}/K_M) ^d
1	Ds	dPaTP	340 (150) ^b	21 (3)	6.2×10^4
2	Ds	dPa'TP	82 (17)	21 (6)	2.6×10^5
3	Ds	dATP	150 (40)	0.36 (0.09)	2.4×10^3
4	Ds	dATP _N	150 (30)	0.048 (0.022)	3.2×10^2
5	Ds	dGTP	n.d. ^c	n.d.	-
6	Ds	dCTP	410 (190)	0.34 (0.05)	8.3×10^2
7	Ds	dTTP	220 (20)	0.41 (0.17)	1.9×10^3
8	Ds	dDsTP	8.0 (3.9)	1.6 (0.1)	2.0×10^5
9	Ds	dDsTP _N	79 (13)	0.78 (0.12)	9.9×10^3
10	A	dPaTP	330 (160)	17 (7)	5.2×10^4
11	G	dPaTP	140 (20)	0.061 (0.006)	4.4×10^2
12	C	dPaTP	n.d.	n.d.	-
13	T	dPaTP	170 (60)	0.053 (0.016)	3.1×10^2
14	A	dPa'TP	110 (40)	20 (6)	1.8×10^5
15	G	dPa'TP	80 (14)	0.13 (0.03)	1.6×10^3
16	C	dPa'TP	80 (34)	0.059 (0.023)	7.4×10^2
17	T	dPa'TP	120 (80)	0.19 (0.07)	1.6×10^3
18	A	dTTP	0.70 (0.40)	2.8 (1.5)	4.0×10^6
19	A	dCTP	1200 (600)	2.2 (0.9)	1.8×10^3
20	G	dCTP	0.24 (0.18)	5.5 (1.7)	2.3×10^2
21	G	dTTP	140 (70)	0.29 (0.12)	2.1×10^3

[0076] a. Assays were carried out at 37°C for 1 to 35 minutes using 5 μ M template-primer duplex, 5-50 nM enzyme and 0.3-1500 μ M nucleoside 5'-triphosphate in a solution (10 μ l)

containing 50 mM Tris-HCl (pH 7.5), 10 mM MgCl₂, 1 mM DTT and 0.05 mg/ml bovine serum albumin. Each parameter was averaged from 3 to 8 data sets.

[0077] b. Standard deviations are given in parentheses.

[0078] c. Minimal inserted products (<2%) were detected after incubation for 20 minutes with 1500 μ M nucleoside 5'-triphosphate and 50 nM enzyme.

[0079] d. The units of this term are % min⁻¹M⁻¹.

However, dDsTP was incorporated opposite Ds in the template with high efficiency ($V_{\max}/K_M = 2.0 \times 10^5$), which was higher than the incorporation efficiency of dPaTP opposite Ds ($V_{\max}/K_M = 6.2 \times 10^4$). This Ds-Ds pairing falls within a problematic base pairing with no shape fitting between hydrophobic bases, and hence will cause deformation in the B-type DNA structure. As a result, dDsTP incorporation opposite Ds in the template will stop the subsequent extension during replication. For example, when Ds-containing template DNA was used to perform primer extension in a solution containing both dPaTP and dDsTP substrates, primer extension with the Klenow fragment having 3'→5' exonuclease activity (KF exo⁺) was inhibited (Figure 2d, Lanes 3 and 4) because dDsTP incorporation opposite Ds in the template would be facilitated with increase in dDsTP (0.5 or 1 molar equivalent of dPaTP).

[0080] To solve this fundamental problem of hydrophobic base pairing during replication, the inventors of the present invention have further used a modified 5'-triphosphate, i.e., 5'- γ -amidotriphosphate of Ds (Figure 1b; denoted as dDsTP_N) as a substrate. The inventors have found that the incorporation

efficiency of dDsTP_N opposite Ds in the template was significantly reduced ($V_{\max}/K_M = 9.9 \times 10^3$). However, a further problem arose in that the incorporation efficiency of dDsTP_N opposite Pa in the template ($V_{\max}/K_M = 6.7 \times 10^4$) was reduced to a level close to the incorporation efficiency of A opposite Pa in the template ($V_{\max}/K_M = 4.3 \times 10^4$). This problem could be solved by using 5'- γ -amidotriphosphate of A (dATP_N). When using dATP_N instead of dATP, the incorporation efficiency of dDsTP_N opposite template Pa was 37-fold higher than that of dATP_N opposite template Pa ($V_{\max}/K_M = 1.8 \times 10^3$), so that the high selectivity of Pa-Ds base pairing could be maintained.

[0081] Thus, the inventors of the present invention have achieved the highly complementary selectivity of unnatural base pairing during replication by using usual 5'-triphosphates, dPaTP, dGTP, dCTP and dTTP in combination with 5'- γ -amidotriphosphates dDsTP_N and dATP_N (Figure 2c).

[0082] In the presence of the modified substrate dDsTP_N, primer extension after Pa incorporation opposite template Ds proceeded without being inhibited (Figure 2d, Lanes 5 and 6). Moreover, primer extension after dDsTP_N incorporation opposite template Pa or after dATP_N incorporation opposite template T also proceeded efficiently (Figure 2e, Lanes 2 and 14). Interestingly, when dATP_N was misincorporated opposite template Pa, the subsequent extension reaction showed significantly reduced efficiency as compared to the extension reaction after misincorporation of dATP opposite template Pa (Figure 2e, Lanes 4 and 5).

[0083] To further improve the selectivity of unnatural base

pairing, the inventors of the present invention have used a DNA polymerase having 3'→5' exonuclease activity in this system. The use of KF exo+ significantly reduced the efficiency of primer extension after formation of undesired A-Pa and Ds-T base pairs, whereby primer extension paused around the unnatural base position (Figure 2d, Lane 1 and Figure 2e, Lanes 9, 10, 17 and 18).

[0084] Thus, the inventors of the present invention have preferably used a DNA polymerase having 3'→5' exonuclease activity in combination with usual 5'-triphosphate substrates and modified 5'-triphosphate substrates to create a specific unnatural base pair system which functions together with A-T and G-C base pairs during replication.

[0085] Unnatural base pairing is formed by specific shape complementarity between bases, and hence lacks hydrogen bonding interaction between bases. During replication, this unnatural base pairing shows particularly high selectivity when combining usual 5'-triphosphates and modified 5'-triphosphates (i.e., 5'-γ-amidotriphosphates) for use as substrates of a DNA polymerase having 3'→5' exonuclease activity (3'→5' exonuclease-proficient DNA polymerase) which allows PCR amplification, preferably on a practical level and with high fidelity. DNA fragments containing unnatural bases can be confirmed for their sequences by dideoxynucleotide chain termination sequencing supplemented with substrates of the unnatural bases. Moreover, the complementarity of unnatural base pairs can mediate incorporation of these bases into RNA during normal T7 transcription.

[0086] The present invention provides a novel nucleic acid amplification system using any one of or a combination of 1) to 3) shown below:

[0087] 1) a deoxyribonucleoside 5'-triphosphate, in which the hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group, is used as a substrate during replication reaction;

2) a DNA polymerase having exonuclease activity is used during replication reaction; and

3) artificial base pairing between a nucleotide having a base of formula 1 described later and a nucleotide having a base of formula 2 is used.

[0088] To enable a better understanding of the present invention, development of the background and of the present invention has been explained above. The scope of the present invention is not limited by the above explanation, but is defined by the claims.

[0089] Method for nucleic acid replication

In one embodiment, the present invention provides a novel method for nucleic acid replication. The method of the present invention is characterized in that a deoxyribonucleoside 5'-triphosphate, in which the hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group, is used as a substrate during replication

reaction.

[0090] When the hydroxyl group of phosphoric acid at the γ -position of a deoxyribonucleoside 5'-triphosphate to be used as a substrate during replication reaction is substituted with such a substituent as listed above, the selectivity of replication reaction is further improved. Because of their improved selectivity, the use of such modified 5'-triphosphates allows the replication reaction involving artificial base pairing to proceed in a substantially available manner, even if the efficiency of substrate incorporation is reduced as compared to unsubstituted substrates.

[0091] The above substituent is preferably an amino group.

[0092] The hydroxyl group of phosphoric acid at the γ -position of deoxyribonucleoside 5'-triphosphates can be modified in any known manner. For example, Example I-3-(15) and (16) described herein later disclose synthesis examples from corresponding nucleosides where the substituent is an amino group. Alternatively, Non-patent Document 47 also discloses synthesis procedures for γ -amidated nucleotides.

[0093] Likewise, in other cases where the substituent is a group other than an amino group, i.e., a methylamino group, a dimethylamino group, a mercapto group or a fluoro group, such modified triphosphates can also be synthesized in a manner known to those skilled in the art.

[0094] Replication reaction is not limited in any way except for using the above substrates, and can be effected in a known manner. Without being limited thereto, for example,

it is preferable to use a DNA polymerase having exonuclease activity for the purpose of avoiding undesired non-specific base pairing during replication reaction. The polymerase having exonuclease activity is selected from the group consisting of the Klenow fragment, T4 DNA polymerases and thermophilic DNA polymerase (e.g., *Thermococcus litoralis*-derived Vent DNA polymerase), each having 3'→5' exonuclease activity.

[0095] As an embodiment of the method of the present invention, a deoxyribonucleoside 5'-triphosphate having an unnatural base may be used as a substrate. In recent years, studies have been conducted to develop base pairs having hydrogen modes different from those of natural base pairs and capable of eliminating base pairing with natural bases by steric hindrance; and hence some artificial base pairs have been reported. Combinations known as artificial base pairs include those based on hydrogen bonding between bases, and those based on the hydrophobicity of bases. In nucleic acids having these unnatural bases, the present invention has achieved increased selectivity and efficiency during transcription, replication and/or translation reaction by modifying the hydroxyl group of phosphoric acid at the γ -position.

[0096] There is no particular limitation on the type of unnatural bases in the present invention. For example, the present invention encompasses the use of deoxyribonucleotides having any known unnatural bases shown below:

[0097] 2-amino-6-dimethylaminopurine (x) and 2-amino-6-

thienylpurine (s) (Non-patent Document 33);

a 2-amino-6-(2-thienyl)-9H-purin-9-yl group (s) and a 2-oxo-(1H)pyridin-3-yl group (y) (Patent Document 1, Non-patent Document 17);

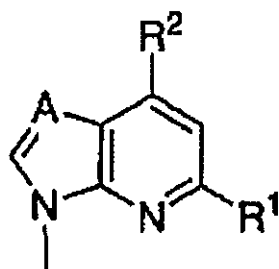
a 2-amino-6-(2-thiazolyl)purin-9-yl group (v) and a 2-oxo-(1H)pyridin-3-yl group (y) (Patent Document 3, Non-patent Document 18); and

pyrrole-2-carbaldehyde (Pa) and 7-methyl-imidazo[4,5-b]pyridine (Q) (Non-patent Documents 26 and 34).

The inventors of the present invention have further invented a novel artificial base pair, and filed a patent application on August 4, 2005 (Japanese Patent Application No. 2005-226492). The above application relates to an artificial base pair between pyrrole-2-carbaldehyde (Pa) and a 2-amino-6-(2-thienyl)-9H-purin-9-yl group (s). The above base pair shows excellent selectivity, particularly during transcription reaction where Pa is used as a template and s is used as a substrate.

[0098] Moreover, the inventors of the present invention provides a further novel artificial base pair in the present invention. This novel artificial base pair may also be used effectively during replication reaction in the present invention. Thus, without being limited thereto, the deoxyribonucleoside 5'-triphosphate used as a substrate has a base represented by the following formula 1:

[0099] [Formula 13]



[0100] [wherein

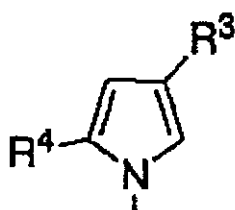
R¹ is hydrogen or an amino group,

R² is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH].

[0101] Alternatively, the deoxyribonucleoside 5'-triphosphate used as a substrate has a base represented by the following formula 2:

[0102] [Formula 14]



[0103] [wherein

R³ is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C₁-C₃ alkyl group, a substituted or unsubstituted C₂-C₃ alkenyl group, or a substituted or unsubstituted C₂-C₃ alkynyl group, and

R⁴ is a formyl group or a nitro group].

[0104] Bases of formulae 1 and 2 each show excellent

selectivity and efficiency during replication reaction, regardless of which of them is used as a substrate or template. Moreover, these bases also show excellent selectivity and efficiency during transcription reaction.

[0105] The novel artificial base pair between bases of formulae 1 and 2 provided in the present invention will be described in detail below in the section "Nucleic acids of the present invention based on artificial base pairing."

[0106] Alternatively, the deoxyribonucleoside 5'-triphosphate used as a substrate may have a natural base. There are 4 types of natural bases known for deoxyribonucleosides, including adenine (A), guanine (G), cytosine (C) and thymine (T). For these natural bases, deoxyribonucleoside 5'-triphosphates whose hydroxyl group of phosphoric acid at the γ -position is substituted may also be used effectively as substrates during replication reaction of nucleic acids, as in the case of unnatural bases.

[0107] Deoxyribonucleoside 5'-triphosphates

The present invention also provides a deoxyribonucleoside 5'-triphosphate, in which the hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group.

[0108] The deoxyribonucleoside 5'-triphosphate of the present invention may have either an unnatural base or a natural base. The terms "unnatural base" and "natural base" are as defined above in the method for nucleic acid

replication.

[0109] The deoxyribonucleoside 5'-triphosphate of the present invention can be used as a substrate in the above method of the present invention for nucleic acid replication.

[0110] Nucleic acids of the present invention based on artificial base pairing

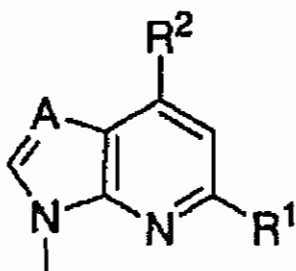
To obtain an artificial base pair showing excellent efficiency and selectivity during all reactions of replication, transcription and translation, the inventors of the present invention have studied base pairs constructed by combining several unnatural base pairs which had been developed on their own. As a result, the inventors have found that a combination between 2-amino-6-(2-thienyl)purin-9-yl (s) (Non-patent Documents 17 and 33) and 2-formyl-1H-pyrrol-1-yl (Pa) (Non-patent Document 26) is highly selective and efficient during transcription (Patent Document 4: not yet published).

[0111] In the present invention, the inventors have further prepared a 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl group (Ds) from s through CH substitution for one of the two N atoms in the 6-membered ring of the purine ring and replacement of the amino group at the 2-position with hydrogen, i.e., through deaza modification and deamination, and have studied the fidelity of artificial base pairing between Ds and Pa. As a result, the inventors have found that artificial base pairing between Ds and Pa is highly selective and efficient during replication and transcription, regardless of which of them serves as a template or substrate, thereby arriving at the

present invention.

[0112] Thus, in one embodiment, the present invention provides a nucleic acid, in which a nucleotide having a base represented by the following formula 1:

[0113] [Formula 15]



[0114] [wherein

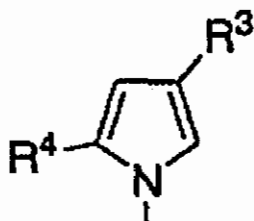
R¹ is hydrogen or an amino group,

R² is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH]

forms a base pair with a nucleotide having a base represented by the following formula 2:

[0115] [Formula 16]



[0116] [wherein

R³ is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C₁-C₃ alkyl group, a substituted

or unsubstituted C₂-C₃ alkenyl group, or a substituted or unsubstituted C₂-C₃ alkynyl group, and

R⁴ is a formyl group or a nitro group].

[0117] As used herein, the term "nucleoside" is intended to mean a glycoside compound formed through glycosidic linking between a nucleic acid base and a reducing group of a sugar. It should be noted that the term "nucleic acid base" is intended to encompass adenine, guanine, cytosine, thymine, uracil, and also derivatives thereof. The type of "derivative" is not limited in any way. Specific examples include bases represented by the above formulae 1 and 2. The term "nucleotide" refers to a compound in which the sugar moiety of the above nucleoside forms an ester with phosphoric acid, more preferably a mono-, di- or tri-phosphate ester. The sugar moiety of such a nucleoside or nucleotide may be ribofuranosyl, 2'-deoxyribofuranosyl, or 2'-substituted ribofuranosyl having a substituent (e.g., halogen) at the 2'-position. Without being limited thereto, in the phosphoric acid moiety, the hydroxyl group of phosphoric acid at the γ -position is desirably substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group. The sugar and phosphoric acid moieties may be in the same form as found in known nucleosides, nucleotides, or derivatives thereof. A ribonucleotide whose sugar moiety is ribofuranosyl can be used as a component of RNA, while a deoxyribonucleotide whose sugar moiety is deoxyribofuranosyl can be used as a component of DNA.

[0118] In bases of formula 1, the thienyl, thiazolyl or imidazolyl group listed as R^2 may be unsubstituted or may be substituted at the 4- and/or 5-position(s) with one or more groups independently selected from the group consisting of a methyl group, an amino group, a nitro group and a hydroxy group.

[0119] Among bases of formula 1 in the present invention, those in which R^2 is a substituted or unsubstituted 2-thienyl group are herein referred to as "Ds" or "Ds analog," depending on the context. Among bases of formula 1 in the present invention, those in which R^2 is a substituted or unsubstituted 2-thiazolyl group are herein referred to as "Dv" or "Dv analog," depending on the context. Among bases of formula 1 in the present invention, those in which R^2 is a substituted or unsubstituted 1H-2-imidazolyl group are herein referred to as "Dm" or "Dm analog," depending on the context.

[0120] With respect to R^1 in formula 1, for example, "Ds" as used herein encompasses both cases where R^1 is hydrogen and an amino group. On the other hand, in some prior art documents, such a base is described in different terminology depending on the embodiment of R^1 , for example, "s" for the case where R^1 is an amino group or "s'" for the case where R^1 is hydrogen.

[0121] A may either be N or CH. In a case where A is N (deaza form), such a base is expressed as, e.g., "Ds", "Dv" or "Dm." In a case where A is CH, such a base is expressed as, e.g., "DDs", "DDv" or "DDm." For example, the term "Ds analog" as used herein includes "DDs." Moreover, "DDs" and

"Ds analog" may also be collectively referred to as "Ds," depending on the context.

[0122] Without being limited thereto, the base of formula 1 is preferably selected from the group consisting of:

- A1) a 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl group (Ds);
- A2) a 7-(2-thiazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group (Dv);
- A3) a 7-(1H-2-imidazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group;
- A4) a 5-amino-7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl group (Ds);
- A5) a 5-amino-7-(2-thiazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group (Dv);
- A6) a 5-amino-7-(1H-2-imidazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group;
- A7) a 4-(2-thienyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group (DDs);
- A8) a 4-(2-thiazolyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group (DDv);
- A-9) a 4-(1H-2-imidazolyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group;
- A-10) a 6-amino-4-(2-thienyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group (DDs);
- A-11) a 6-amino-4-(2-thiazolyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group (DDv); and
- A-12) a 6-amino-4-(1H-2-imidazolyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group.

[0123] Among the above bases, A1 and A4 are members of Ds, while A7 and A10 are members of DDs. A2 and A5 are members of Dv, while A8 and A11 are members of DDv. A3 and A6 are members of "Dm," while A9 and A12 are members of "DDm."

[0124] More preferred is A1) a 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl group (Ds), A2) a 7-(2-thiazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group (Dv), A4) a 5-amino-7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl group (Ds), or A5) a 5-amino-7-(2-thiazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group (Dv). Most preferred is a 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl group (Ds) (Compound 10 or 14 in Example I).

[0125] The base of formula 1 in the present invention and a nucleoside or nucleotide containing the same may be synthesized in a known manner. More specifically, Example I-3 described later discloses procedures for synthesis of nucleoside 5'-triphosphates or 5'- γ -amidotriphosphates of a 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl group (Ds) (Compound 10 or 14 in Example I) from 2-amino-3-nitro-4-chloropyridine (Compound 1 in Example I) (Non-patent Document 41), by way of example. Likewise, Example II-1 discloses procedures for synthesis of nucleoside 5'-triphosphates of 7-(2-thiazolyl)-3H-imidazo[4,5]pyridine (Compound 4 in Example II) (Dv).

[0126] Further, Example II-3 discloses procedures for synthesis of nucleoside 5'-triphosphates of 4-(2-thienyl)-1H-pyrrolo[2,3-b]pyridine (DDs) and 4-(2-thiazolyl)-1H-pyrrolo[2,3-b]pyridine (DDv).

[0127] Nucleosides, nucleotides or 5'- γ -amidotriphosphates having other bases of formula 1 in the present invention may

also be synthesized in the same manner as actually disclosed herein and/or in a manner known to those skilled in the art.

[0128] In bases of formula 2 in the present invention, R^3 is a group selected from hydrogen, iodo group, a substituted or unsubstituted C_1 - C_3 alkyl group, a substituted or unsubstituted C_2 - C_3 alkenyl group, or a substituted or unsubstituted C_2 - C_3 alkynyl group.

[0129] Such an alkyl, alkenyl or alkynyl group may further be substituted with one or more groups independently selected from the group consisting of a lower alkyl group, a halogen group, a hydroxyl group, an amino group, an alkylamino group and an aromatic heterocyclic ring.

[0130] Alternatively, such an alkyl, alkenyl or alkynyl group may further be substituted with biotin or a fluorescent molecule.

[0131] Biotin is also called Coenzyme R and is a member of vitamins B. Biotin is known to specifically bind to and form a complex with avidin (a glycoprotein contained in albumen). Thus, a nucleoside and others having biotin as a substituent will specifically bind to avidin protein. This means that a nucleic acid containing a biotin-labeled nucleoside and others can be attached to and hence immobilized and separated on avidin-bound carriers. If nucleic acids (e.g., aptamers) binding to specific molecules are immobilized, such immobilized nucleic acids can be used for detection and isolation of specific substances or used as diagnostic reagents, by way of example. To introduce biotin into the alkyl, alkenyl or alkynyl group listed as R^3 in the base of

formula 2, biotin may be attached via an amino group, either directly or through a linker.

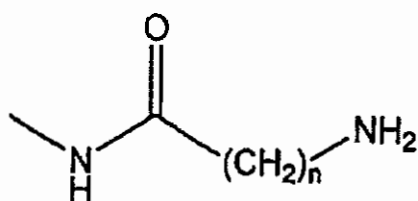
[0132] As a fluorescent molecule, any known molecule may be used and is preferably selected from the group consisting of 5-carboxyfluorescein (5-FAM), 6-carboxyfluorescein (6-FAM), 5-carboxytetramethylrhodamine (5-TAMRA), 6-carboxytetramethylrhodamine (6-TAMRA), 5-dimethylaminonaphthalene-1-sulfonic acid (DANSYL), 5-carboxy-2',4,4',5',7,7'-hexachlorofluorescein (5-HEX), 6-carboxy-2',4,4',5',7,7'-hexachlorofluorescein (6-HEX), 5-carboxy-2',4,7,7'-tetrachlorofluorescein (5-TET), 6-carboxy-2',4,7,7'-tetrachlorofluorescein (6-TET), 5-carboxy-X-rhodamine (5-ROX), 6-carboxy-X-rhodamine (6-ROX), and derivatives thereof. In general, fluorescein and rhodamine are expressed in both open-ring and spiro forms.

[0133] For example, FAM has an absorption peak wavelength of 493 nm and a fluorescence peak wavelength of 522 nm. Likewise, TAMRA has an absorption peak wavelength of 553 nm and a fluorescence peak wavelength of 578 nm. DANSYL has an absorption peak wavelength of 335 nm and a fluorescence peak wavelength of 518 nm. HEX has an absorption peak wavelength of 535 nm and a fluorescence peak wavelength of 556 nm. TET has an absorption peak wavelength of 521 nm and a fluorescence peak wavelength of 536 nm. 5-ROX has an absorption peak wavelength of 567 nm and a fluorescence peak wavelength of 591 nm. 6-ROX has an absorption peak wavelength of 570 nm and a fluorescence peak wavelength of 590 nm. A nucleoside or nucleotide having a base of formula 2 in which R³ is

substituted with a fluorescent molecule allows nucleic acid detection in a manner dependent on the type of fluorescent molecule. Thus, a nucleic acid containing a nucleotide having a base of formula 2 in which the alkyl, alkenyl or alkynyl group listed as R^3 is substituted with a fluorescent molecule can be used as a labeled nucleic acid probe to detect substances interacting with the nucleic acid. Moreover, since these individual fluorescent molecules have fluorescent colors different from each other, they can also be used in multiple staining. To introduce a fluorescent molecule into the alkyl, alkenyl or alkynyl group listed as R^3 in the base of formula 2, the fluorescent molecule may be attached via an amino group, either directly or through a linker.

[0134] It should be noted that in a case where biotin or a fluorescent molecule is attached through a linker to the alkyl, alkenyl or alkynyl group listed as R^3 in the base of formula 2, the type of linker is not limited in any way and may be determined as appropriate by those skilled in the art. Without being limited thereto, the linker is preferably selected from the group consisting of chemical formulae I and II shown below:

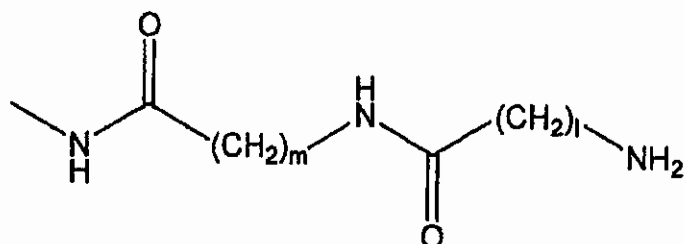
[0135] [Formula 17]



I

[0136] [wherein n is selected from integers of 1 to 5]; and

[0137] [Formula 18]



[0138] [wherein m and l are each independently selected from integers of 1 to 5].

[0139] R^4 is a formyl group or a nitro group. Among bases of formula 2, those in which R^4 is a formyl group are herein referred to as "Pa" or "Pa analog," depending on the context. Those in which R^4 is a nitro group are herein referred to as "Pn" or "Pn analog," depending on the context. "Pa", "Pa analog", "Pn" and "Pn analog" may also be collectively referred to as "Pa."

[0140] Without being limited thereto, the base of formula 2 is preferably selected from the group consisting of:

- B1) a 2-formyl-1H-pyrrol-1-yl group (Pa);
- B2) a 2-formyl-4-iodo-1H-pyrrol-1-yl group;
- B3) a 2-formyl-4-methyl-1H-pyrrol-1-yl group;
- B4) a 2-formyl-4-(1-propyn-1-yl)-1H-pyrrol-1-yl group;
- B5) a 2-formyl-4-(2-substituted aminovinyl)-1H-pyrrol-1-yl group;
- B6) a 2-formyl-4-(3-substituted amino-1-propyn-1-yl)-1H-pyrrol-1-yl group;
- B7) a 2-nitro-1H-pyrrol-1-yl group (Pn);

- B8) a 2-nitro-4-iodo-1H-pyrrol-1-yl group;
B9) a 2-nitro-4-methyl-1H-pyrrol-1-yl group;
B10) a 2-nitro-4-(1-propyn-1-yl)-1H-pyrrol-1-yl group;
B11) a 2-nitro-4-(2-substituted aminovinyl)-1H-pyrrol-1-yl group; and
B12) a 2-nitro-4-(3-substituted amino-1-propyn-1-yl)-1H-pyrrol-1-yl group.

[0141] Among the above bases, B1 to B6 are members of "Pa" or "Pa derivative," while B7 to B12 are members of "Pn" or "Pn derivative."

[0142] More preferred is B1) a 2-formyl-1H-pyrrol-1-yl group (Pa) (Compound 19 in Example I) or B4) a 2-formyl-4-(1-propyn-1-yl)-1H-pyrrol-1-yl group (Compound 20 in Example I).

[0143] A nucleoside or nucleotide having a base of formula 2 in the present invention may be synthesized in a known manner. Taking Pa as an example, starting materials (e.g., pyrrole-2-carbaldehyde) can be purchased from, for example, Aldrich [1003-29-8] or Merck [807574]. Likewise, Pa derivatives may be synthesized by being derived from Pa, in principle. For example, a derivative having propyne introduced at the 4-position of Pa can be found in Bioorg. Med. Chem. Lett., 13, p. 4515-4518 (2003) (Non-patent Document 28).

[0144] In Example I-3-(12) described later, 1-(β -D-ribofuranosyl)pyrrole-2-carbaldehyde (Compound 17) was synthesized from pyrrole-2-carbaldehyde. Likewise, in Example I-3-(13), 4-propynyl-1-(β -D-ribofuranosyl)pyrrole-2-carbaldehyde (Compound 18) was synthesized from 4-propynyl-2-

pyrrolecarbaldehyde (Compound 16) (Non-patent Document 40).

Moreover, their nucleoside 5'-triphosphates and 5'- γ -amidotriphosphates were synthesized (Compounds 19 and 20).

[0145] Further, in Example II-2 described later, nucleoside 5'-triphosphates of 2-nitropyrrole (Compound 1) (Pn) were synthesized from 2-nitropyrrole (Compound 1) (Non-patent Document 49).

[0146] The present invention provides a nucleic acid in which a nucleotide having a base of formula 1 forms a base pair with a nucleotide having a base of formula 2. As used herein, the term "nucleic acid" is intended to mean a molecule of a nucleic acid strand in which more than one nucleotide is linked in the direction of 5'→3'. The nucleic acid of the present invention encompasses single-stranded or double-stranded RNA or DNA. The double-stranded nucleic acid may be DNA/DNA, RNA/RNA, or DNA/RNA. DNA also includes cDNA obtained by reverse transcription using RNA as a template. Alternatively, the nucleic acid may form a triplex, a quadruplex, etc.

[0147] With the aim of further expansion of nucleic acid functions, the inventors of the present invention have attempted to design nucleosides or nucleotides having unnatural bases. Embodiments of newly developed artificial base pairs include a base pair between a nucleotide having a base of formula 1 and a nucleotide having a base of formula 2. Such a nucleotide having a base of formula 1 and such a nucleotide having a base of formula 2 each function as a substrate and as a template with high efficiency and/or high

selectivity in the mechanisms of both replication and transcription.

[0148] Although there is no significant hydrogen bonding interaction between bases of formulae 1 and 2, e.g., between Ds and Pa (non-hydrogen-bonded Ds-Pa base pair), the efficiency and selectivity of Ds-Pa base pairing is as high as that of natural base pairing. This Ds-Pa base pair shows a higher efficiency than the previously developed hydrophilic s-z base pair. The complementary shapes of Ds and Pa are fitted to each other, but their shapes differ from those of natural purines and pyrimidines. This specific stereochemical fitting would eliminate undesired base pairing with natural bases, thereby resulting in high selectivity between Ds and Pa during replication and transcription. In this way, shape complementarity plays an important role in specific base pairing during replication and transcription.

[0149] In the present invention, a nucleotide having a base of formula 1 and a nucleotide having a base of formula 2 are present in two separate nucleic acid strands and can form a duplex through base pairing. Alternatively, these nucleotides may be present in the same single-stranded nucleic acid. In this case, such a single strand may form a loop structure through base pairing.

[0150] In the present invention, such a nucleotide having a base of formula 1 or 2 can be incorporated into nucleic acids such as DNA or RNA through replication, transcription or reverse transcription reaction. Alternatively, such a nucleotide may be incorporated into DNA or RNA through

chemical synthesis, as in the case of nucleosides or nucleotides having natural bases.

[0151] These replication, transcription and reverse transcription reactions may be accomplished according to known techniques. Without being limited thereto, for example, it is possible to use T7 RNA polymerase (Takara or other suppliers) for transcription, Klenow fragment (KF) for replication, and AMV Reverse Transcriptase XL (AMV-RT, Life Science) for reverse transcription.

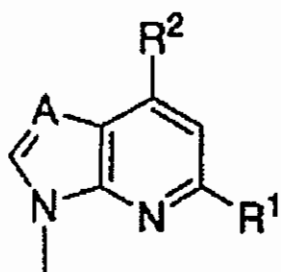
[0152] Without being limited thereto, in one embodiment, the nucleic acid of the present invention forms a base pair(s) in the step of replication, transcription or reverse transcription of the nucleic acid. In a case where the nucleic acid of the present invention forms a base pair(s) in the transcription step, a nucleotide having a base of formula 1 may be a part of DNA, while a nucleotide having a base of formula 2 may be a part of RNA, or alternatively, vice versa.

[0153] The unnatural base pair system of the present invention is summarized in Figure 1c. Effects provided by the system of the present invention during replication are shown in, for example, Figures 2b to 2e, Figures 3b to 3i, and Figures 8 to 20. Effects provided by the system of the present invention during transcription are shown in, for example, Figures 4c to 4e and Figures 5a to 5c.

[0154] Method for nucleic acid preparation

The present invention also provides a method for preparing a nucleic acid containing a nucleotide having a base represented by the following formula 1:

[0155] [Formula 19]



[0156] [wherein

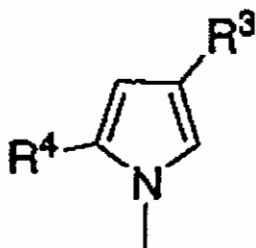
R¹ is hydrogen or an amino group,

R² is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH],

wherein the method comprises effecting transcription, reverse transcription or replication by using, as a template, a nucleic acid containing a nucleotide having a base represented by the following formula 2:

[0157] [Formula 20]



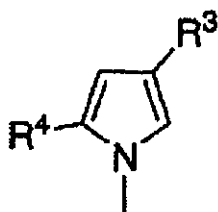
[0158] [wherein

R³ is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C₁-C₃ alkyl group, a substituted or unsubstituted C₂-C₃ alkenyl group, or a substituted or unsubstituted C₂-C₃ alkynyl group, and

R^4 is a formyl group or a nitro group],
whereby the nucleotide having a base of formula 1 is
incorporated at a site complementary to the nucleotide having
a base of formula 2.

[0159] The present invention also provides a method for
preparing a nucleic acid containing a nucleotide having a base
represented by the following formula 2:

[0160] [Formula 21]



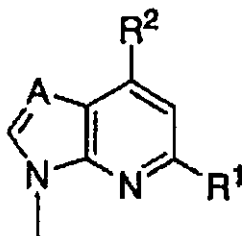
[0161] [wherein

R^3 is a group selected from hydrogen, an iodo group, a
substituted or unsubstituted C_1 - C_3 alkyl group, a substituted
or unsubstituted C_2 - C_3 alkenyl group, or a substituted or
unsubstituted C_2 - C_3 alkynyl group, and

R^4 is a formyl group or a nitro group],

wherein the method comprises effecting transcription,
reverse transcription or replication by using, as a template,
a nucleic acid containing a nucleotide having a base
represented by the following formula 1:

[0162] [Formula 22]



[0163] [wherein

R^1 is hydrogen or an amino group,

R^2 is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH],

whereby the nucleotide having a base of formula 2 is incorporated at a site complementary to the nucleotide having a base of formula 1.

[0164] The nucleotide having a base of formula 1 and the nucleotide having a base of formula 2 are as defined herein above in the section "Nucleic acids of the present invention based on artificial base pairing."

[0165] The present invention further aims to provide such a nucleic acid containing a nucleotide having a base of formula 1 and/or 2, which is prepared by the above method of the present invention.

[0166] The nucleic acid incorporating the nucleotide(s) of the present invention may be used as tRNA, mRNA, antisense DNA or RNA, a ribozyme or an aptamer. The term "antisense DNA or RNA" refers to DNA or RNA capable of inhibiting the expression of a specific gene. It was named to mean that such DNA or RNA is complementary to the full-length or partial sequence of a target gene sequence (sense strand). Antisense DNA or RNA may be used as a tool for artificial regulation of gene expression. Because of containing unnatural bases, such antisense DNA or RNA incorporating the nucleotide(s) of the present invention can be designed to have a different complementarity to a

target when compared to the case of using natural bases only. The term "ribozyme" is a generic name for catalysts composed of RNA. The term "aptamer" refers to an *in vitro*-selected nucleic acid having the ability to bind to a specific molecule such as a protein.

The nucleic acid (DNA or RNA) (e.g., mRNA, synthetic RNA) incorporating the nucleotide(s) of the present invention may also encode all or part of a protein or peptide. The nucleic acid of the present invention may be used, e.g., as a gene fragment or a probe. The present invention also encompasses the following embodiments: partial or complete replacement of native genes by the nucleic acids of the present invention; addition of one or more nucleotides of the present invention to native genes; or combinations thereof.

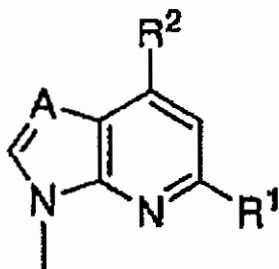
[0167] Furthermore, the nucleic acids of the present invention incorporating nucleotides having unnatural bases may also be used in RNA interference (RNAi). RNA interference is a phenomenon in which double-stranded RNA (dsRNA) induces mRNA degradation in a sequence-specific manner and hence inhibits gene expression. In a typical example of RNA interference, dsRNA is processed by Dicer belonging to the RNaseIII family into siRNA (short interfering RNA) of approximately 21 to 23 bases in length, which has a 3'-terminal overhang of approximately 2 bases. siRNA is associated into an siRNA-protein complex called RISC and induces mRNA degradation in a sequence-specific manner. RNA interference is shown to be a phenomenon conserved among a wide range of organism species including mammals (e.g., human, mouse), nematodes, plants,

drosophila and fungi. The nucleic acids of the present invention incorporating nucleotides having unnatural bases can be used as siRNA in RNA interference or as a part of mRNA to be degraded.

[0168] Ribonucleoside 5'-triphosphates

The present invention further provides a ribonucleoside 5'-triphosphate having a base represented by the following formula 1:

[0169] [Formula 23]



[0170] [wherein

R¹ is hydrogen or an amino group,

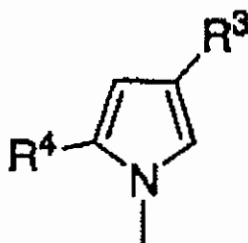
R² is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH].

[0171] The structure of formula 1 including substituents is as defined above for deoxyribonucleotides.

[0172] The present invention furthermore provides a ribonucleoside 5'-triphosphate having a base represented by the following formula 2:

[0173] [Formula 24]



[0174] [wherein

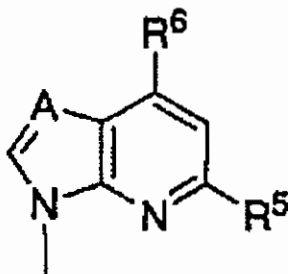
R^3 is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C_1 - C_3 alkyl group, a substituted or unsubstituted C_2 - C_3 alkenyl group, or a substituted or unsubstituted C_2 - C_3 alkynyl group, and

R^4 is a formyl group or a nitro group].

[0175] The structure of formula 2 including substituents is as defined above for deoxyribonucleotides.

[0176] Further, the present invention provides a 5'-O-(4,4'-dimethoxytrityl)-3'-O-(2-cyanoethyl N,N-diisopropylphosphoroamidite) deoxyribonucleoside having a base represented by the following formula 3:

[0177] [Formula 25]



[0178] [wherein

R^5 is hydrogen or a substituted amino group,

R^6 is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a

substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH].

[0179] The substituted amino group listed as R⁵ includes an amino group substituted with a methyl group, an isobutyryl group, a benzoyl group or the like.

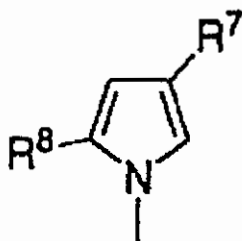
[0180] R⁶ is the same as R² in formula 1.

[0181] As an example of compounds falling within formula 3, Compound 8 was synthesized in Example I.

[0182] Compounds of formula 3 are useful, for example, as starting materials in chemical synthesis of DNA templates for replication and transcription.

[0183] Furthermore, the present invention provides a 5'-O-(4,4'-dimethoxytrityl)-3'-O-(2-cyanoethyl N,N-diisopropylphosphoroamidite) deoxyribonucleoside having a base represented by the following formula 4:

[0184] [Formula 26]



[0185] [wherein

R⁷ is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C₁-C₃ alkyl group, a substituted or unsubstituted C₂-C₃ alkenyl group, or a substituted or unsubstituted C₂-C₃ alkynyl group, and

R⁸ is a formyl group or a nitro group,

excluding the case where R⁷ is hydrogen or a 1-propynyl group and R⁸ is a formyl group].

[0186] R^7 is the same as R^3 in formula 2.

[0187] R^8 is the same as R^4 in formula 2.

[0188] Compounds of formula 4 are useful, for example, as starting materials in chemical synthesis of DNA templates for replication and transcription.

BRIEF DESCRIPTION OF THE DRAWINGS

[0189] [Figure 1a] Figures 1a to 1c show an unnatural base pair system which allows specific replication and transcription. Figure 1a shows an unnatural Ds-Pa base pair, along with natural base pairs.

[Figure 1b] Figures 1a to 1c show an unnatural base pair system which allows specific replication and transcription. Figure 1b shows the structures of unmodified 5'-triphosphate (dNTP) and modified 5'- γ -amidotriphosphate for use as substrates for PCR and primer extension.

[Figure 1c] Figures 1a to 1c show an unnatural base pair system which allows specific replication and transcription. Figure 1c shows an unnatural base pair system which functions in PCR amplification, primer extension, DNA sequencing and T7 transcription.

[Figure 2a] Figures 2a to 2e show single nucleotide insertion and primer extension experiments with an unnatural base pair system based on Ds-Pa base pairing. Figure 2a shows the sequences of template-primer duplexes for use in the single nucleotide insertion experiment.

[Figure 2b] Figures 2a to 2e show single nucleotide insertion and primer extension experiments with an unnatural base pair system based on Ds-Pa base pairing. Figure 2b shows

the incorporation efficiency of each unmodified substrate (dN'TP) opposite each template base by KF exo^- .

[Figure 2c] Figures 2a to 2e show single nucleotide insertion and primer extension experiments with an unnatural base pair system based on Ds-Pa base pairing. Figure 2c shows the incorporation efficiency of each substrate (unmodified substrate dN'TP, N = Pa, G, C and T, and modified substrate dN'TP_N, N = Ds and A) opposite each template base by KF exo^- .

[Figure 2d] Figures 2a to 2e show single nucleotide insertion and primer extension experiments with an unnatural base pair system based on Ds-Pa base pairing. Figure 2d shows the results of primer extension by KF exo^+ for 5 minutes using a Ds-containing template with dPaTP and natural substrates in the presence or absence of dDsTP or dDsTP_N.

[Figure 2e] Figures 2a to 2e show single nucleotide insertion and primer extension experiments with an unnatural base pair system based on Ds-Pa base pairing. Figure 2e shows 3 minute primer extension by KF exo^- or exo^+ using a Pa-containing template or a natural template with a series of substrates ($D_{SN} = \text{dDsTP}_N$ and $A_N = \text{dATP}_N$). Since dGTP is absent from each reaction, the full-length products are mainly 33-mers.

[Figure 3a] Figures 3a to 3i show DNA sequencing and PCR amplification of DNA fragments containing a Ds-Pa base pair. Figure 3a shows the experimental scheme. A double-stranded DNA fragment (150-mer, DNA1) was prepared by primer extension using chemically synthesized DNA fragments (91-mer and 81-mer) containing Ds and Pa.

[Figure 3b] Figures 3a to 3i show DNA sequencing and PCR amplification of DNA fragments containing a Ds-Pa base pair. Figure 3b shows agarose gel analysis of original DNA fragments (0 cycle) and their PCR products after 5 and 10 cycles. DNA1 was amplified by PCR with 0.04 units/ μ l VENT DNA polymerase under the following cycle conditions: 94°C for 0.5 minutes, 45°C for 0.5 minutes, and 65°C for 4 minutes. DNAcont1 composed only of natural bases was amplified by PCR with 0.01 units/ μ l VENT DNA polymerase under the following cycle conditions: 94°C for 0.5 minutes, 45°C for 0.5 minutes, and 72°C for 1 minute.

[Figure 3c] Figures 3a to 3i show DNA sequencing and PCR amplification of DNA fragments containing a Ds-Pa base pair. Figures 3c to 3i show the results of PCR and sequencing performed according to the procedures explained for Figures 3a and 3b. Sequencing in the presence (c-f) or absence (g-i) of dPa'TP performed on the original DNA1 (c, g) and its PCR products after 10 cycles (d, g) and 10+10 cycles (e, h) using the unnatural base pair system or after 10 cycles using natural substrates alone (f).

[0190] Figure 3c shows the results of sequencing with dPa'TP obtained before PCR (0 cycle). The base at position 40 is a base corresponding to a site complementary to Ds in the template, and Pa' is incorporated at this site during sequencing reaction. Thus, peaks of A, G, C and T all disappear only at this site.

[Figure 3d] Figures 3a to 3i show DNA sequencing and PCR amplification of DNA fragments containing a Ds-Pa base pair.

Figures 3c to 3i show the results of PCR and sequencing performed according to the procedures explained for Figures 3a and 3b. Figure 3d shows the results of sequencing with dPa'TP obtained after 10 cycle PCR. The base at position 40 is a base corresponding to a site complementary to Ds in the template, and Pa' is incorporated at this site during sequencing reaction. Thus, peaks of A, G, C and T all disappear only at this site.

[Figure 3e] Figures 3a to 3i show DNA sequencing and PCR amplification of DNA fragments containing a Ds-Pa base pair. Figures 3c to 3i show the results of PCR and sequencing performed according to the procedures explained for Figures 3a and 3b. Figure 3e shows the results of sequencing with dPa'TP obtained after 10+10 cycle PCR. The base at position 40 is a base corresponding to a site complementary to Ds in the template, and Pa' is incorporated at this site during sequencing reaction. Thus, peaks of A, G, C and T all disappear only at this site.

[Figure 3f] Figures 3a to 3i show DNA sequencing and PCR amplification of DNA fragments containing a Ds-Pa base pair. Figures 3c to 3i show the results of PCR and sequencing performed according to the procedures explained for Figures 3a and 3b. Figure 3f shows the results of sequencing with dPa'TP obtained after PCR (10 cycles) with natural substrates alone, without using s and Pa substrates. The Ds-Pa base pair in the template DNA1, which corresponds to the base at position 40, is completely replaced with an A-T base pair (base T at position 40).

[Figure 3g] Figures 3a to 3i show DNA sequencing and PCR amplification of DNA fragments containing a Ds-Pa base pair. Figures 3c to 3i show the results of PCR and sequencing performed according to the procedures explained for Figures 3a and 3b. Figure 3g shows the results of sequencing without dPa'TP obtained before PCR (0 cycle). The base at position 40 is a base corresponding to a site complementary to Ds in the template, but there is no base which forms a specific base pair with Ds, so that the subsequent peaks almost disappear.

[Figure 3h] Figures 3a to 3i show DNA sequencing and PCR amplification of DNA fragments containing a Ds-Pa base pair. Figures 3c to 3i show the results of PCR and sequencing performed according to the procedures explained for Figures 3a and 3b. Figure 3h shows the results of sequencing without dPa'TP obtained after 10 cycle PCR. The base at position 40 is a base corresponding to a site complementary to Ds in the template, but there is no base which forms a specific base pair with Ds, so that the subsequent peaks almost disappear. However, non-specific incorporation (read-through) of bases is also observed. These read-through peaks were slightly increased with an increase in the number of cycles required for PCR amplification.

[Figure 3i] Figures 3a to 3i show DNA sequencing and PCR amplification of DNA fragments containing a Ds-Pa base pair. Figures 3c to 3i show the results of PCR and sequencing performed according to the procedures explained for Figures 3a and 3b. Figure 3i shows the results of sequencing without dPa'TP obtained after 10+10 cycle PCR. The base at position

40 is a base corresponding to a site complementary to Ds in the template, but there is no base which forms a specific base pair with Ds, so that the subsequent peaks almost disappear. However, non-specific incorporation (read-through) of bases is also observed. These read-through peaks were slightly increased with an increase in the number of cycles required for PCR amplification.

[Figure 4a] Figure 4 shows T7 transcription mediated by Ds-Pa base pairing. Figures 4a and 4b show the experimental schemes.

[Figure 4b] Figure 4 shows T7 transcription mediated by Ds-Pa base pairing. Figures 4a and 4b show the experimental schemes.

[Figure 4c] Figure 4 shows T7 transcription mediated by Ds-Pa base pairing. Figure 4c shows gel electrophoresis of transcripts obtained using a template (N = Ds, Pa, Pa', A or C) with natural NTPs (1 mM) in the presence (1 mM) or absence of PaTP, Pa'TP or DsTP. The transcripts were labeled with [γ -³²P]GTP. The yield of each transcript was determined in comparison with the yield of a natural transcript from a template composed only of natural bases (N = A or C), and the transcription efficiency (yield) was averaged from 3 data sets for each case.

[Figure 4d] Figure 4 shows T7 transcription mediated by Ds-Pa base pairing. Figures 4d and 4e show the results of 2D-TLC analysis on labeled ribonucleotide 3'-monophosphates obtained from nuclease digestion of transcripts (17-mer).

[Figure 4e] Figure 4 shows T7 transcription mediated by

Ds-Pa base pairing. Figures 4d and 4e show the results of 2D-TLC analysis on labeled ribonucleotide 3'-monophosphates obtained from nuclease digestion of transcripts (17-mer).

[Figure 5a] Figure 5 shows specific T7 transcription of tRNA molecules containing unnatural anticodons. Figure 5a shows the results of gel electrophoresis on tRNA transcripts obtained using templates (DNA11 to DNA14 and DNAcont4) with natural NTPs (2 or 3 mM) in the presence of Pa'TP (3 mM) or DsTP (2 mM). The transcripts were internally labeled with [α -³²P]GTP. The yield of each transcript was determined in comparison with the yield of a suppressor tRNA_{CUA} transcript from DNAcont4, and the transcription efficiency (yield) was averaged from 3 data sets for each case.

[Figure 5b] Figure 5 shows specific T7 transcription of tRNA molecules containing unnatural anticodons. Figures 5b and 5c show the results of 2D-TLC analysis on labeled ribonucleoside 3'-monophosphates obtained from nuclease digestion of tRNA_{CPA'A} and tRNA_{CUDs} transcripts.

[Figure 5c] Figure 5 shows specific T7 transcription of tRNA molecules containing unnatural anticodons. Figures 5b and 5c show the results of 2D-TLC analysis on labeled ribonucleoside 3'-monophosphates obtained from nuclease digestion of tRNA_{CPA'A} and tRNA_{CUDs} transcripts.

[Figure 5d] Figure 5 shows specific T7 transcription of tRNA molecules containing unnatural anticodons. Figure 5d shows aminoacylation of tRNA transcripts in an *E. coli* cell-free system (RTS-100, Roche). The tRNA transcripts internally labeled with [α -³²P]GTP were incubated in the cell-free system

at 30°C for 30 minutes, and the aminoacylated tRNAs were then analyzed by electrophoresis on a 10% polyacrylamide gel containing 0.2 M Tris-acetate (pH 4.75) and 3 mM EDTA.

[Figure 6] Figure 6 shows the sequences of DNA fragments for use in PCR, sequencing and transcription experiments.

[Figure 7a] Figure 7 shows the sequences of chemically synthesized DNA fragments.

[Figure 7b] Figure 7 shows the sequences of chemically synthesized DNA fragments.

[Figure 8] Figure 8 shows dye terminator sequencing of DNA1 containing 0-10% DNAcont1 in the absence of dPa'TP. The fidelity of Ds-Pa base pairing during PCR can be studied by comparing this peak pattern with that of DNA fragment PCR containing a Ds-Pa base pair.

[Figure 9] Figure 9 shows dye terminator sequencing of PCR products from DNA1 with 300 mM dDSTP_N, dPaTP, dATP, dGTP, dCTP and dTTP. Sequencing in the presence (a-c) or absence (d-f) of dPa'TP performed on the original DNA1 (a, d) and its PCR products after 10 cycles (b, e) or 10+10 cycles (c, f). When using dATP instead of dATP_N to perform PCR, the fidelity was significantly reduced (e, f).

[Figure 10] Figure 10 shows autoradiographs of PCR products obtained with a ³²P-labeled 5'- or 3'-primer. PCR was performed using ³²P-labeled 5'- and non-labeled 3'-primers or ³²P-labeled 3'- and non-labeled 5'-primers in 100 µl buffer solution for Vent DNA polymerase containing DNA1 (a) or DNAcont1 (b). An aliquot (10 µl) from each reaction solution after 1, 3, 5, 10 or 15 PCR cycles was analyzed on a 10%

polyacrylamide-7 M urea gel. The products on the gel were analyzed and quantified with a bio-imaging analyzer. The efficiency per cycle (Y) was calculated with KaleidaGraph using a general curve fit: $N_f = N_0 \times (1 + Y)^n$ [where n is the number of PCR cycles (n = 1, 3, 5 and 10), N_0 is the initial copy number, and N_f is the copy number of a product after n cycles]. N_f was determined from the following equation: $N_f = N_0 + P_0 \times (I_{full}/I_{total})$ [where P_0 is the number of ^{32}P -labeled primer molecules, I_{full} is the intensity of a band corresponding to the full-length product, and I_{total} is the total intensity of bands in the lane]. The Y values thus determined are shown in the table below.

[0191] [Table 2-2]

PCR condition	substrate	elongation step	Vent DNA polymerase (unit/ μl)	Y values 5'-/3'-labeled primer
Unnatural using DNA1(a)	0.3 mM dNTPs and dNTP _{NS}	65°C, 4 min	0.04	0.38/0.29
Conventional using DNAcont1(b)	0.2 mM natural dNTPs	72°C, 1 min	0.01	0.43/0.35

[Figure 11] Figure 11 shows the results of dye terminator sequencing without dPa'TP obtained for PCR products of DNA2 and DNA4 fragments prepared from different samples (a-d) of chemically synthesized fragments containing Ds and Pa. These results indicate that the purity of the chemically synthesized DNA fragments containing a Ds-Pa base pair affects their peak patterns after PCR. This means that the previously determined

fidelity of Ds-Pa base pairing during PCR (99.8%) depends on the purity of chemically synthesized DNA fragments. Thus, the fidelity appears to be much higher than 99.8%

[Figure 12] Figure 12 shows the amplification efficiency of PCR products from DNA1, DNA3 to DNA10 and DNAcont1, as determined by agarose gel electrophoresis. The PCR products were detected on a 4% agarose gel and stained with ethidium bromide. The stained bands were then quantified for their intensity using a Molecular Imager FX Pro system and Quantity One software (Bio-Rad). The amplification efficiency was determined using the following equation: $(\text{Intensity of PCR product})/(\text{Intensity of input DNA for PCR})$. The efficiency was averaged from 3 to 4 data sheets. Standard deviations are given in parentheses.

[Figure 13] Figure 13 shows the results of dye terminator sequencing obtained for PCR products from DNA2 with primer 2.

[Figure 14] Figure 14 shows the results of dye terminator sequencing obtained for PCR products from DNA3 with primer 2.

[Figure 15] Figure 15 shows the results of dye terminator sequencing obtained for PCR products from DNA4 with primer 2.

[Figure 16] Figure 16 shows the results of dye terminator sequencing obtained for PCR products from DNA5 with primer 1.

[Figure 17] Figure 17 shows the results of dye terminator sequencing obtained for PCR products from DNA6 with primer 1.

[Figure 18] Figure 18 shows the results of dye terminator sequencing obtained for PCR products from DNA7 with primer 2.

[Figure 19] Figure 19 shows the results of dye terminator sequencing obtained for PCR products from DNA8 with primer 2.

[Figure 20] Figure 20 shows the results of dye terminator sequencing obtained for PCR products from DNA9 with primer 2.

[Figure 21] Figure 21 shows a scheme for aminoacylation of unnatural anticodon-containing tRNAs in an *E. coli*-derived cell-free translation system.

[Figure 22] Figure 22 shows a synthesis scheme for nucleoside derivatives of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine. Reagents and abbreviations: (a) dichlorobis(triphenylphosphine)palladium, 2-(tributylstanyl)thiophene, DMF; (b) palladium carbon, sodium borohydride, ethanol, ethyl acetate; (c) formic acid; (d) NaH, 2-deoxy-3,5-di-O-p-toluoyl- α -D-erythro-pentofuranosyl chloride, CH₃CN; (e) NH₃, methanol; (f) and (l) 4,4'-dimethoxytrityl chloride, pyridine; (g) 2-cyanoethyl tetraisopropylphosphordiamidite, tetrazole, CH₃CN; (h) and (m) acetic anhydride, pyridine, then dichloroacetic acid, dichloromethane; (i) and (n) 2-chloro-4H-1,3,2,-benzodioxaphosphorin-4-one, dioxane, pyridine, tributylamine, bis(tributylammonium) pyrophosphate, DMF, then I₂/pyridine, water, NH₄OH (for 5'-triphosphate), I₂/pyridine, NH₄OH (for 5'- γ -amidotriphosphate); (j) tetra-O-acetyl- β -D-ribofuranose, chloroacetic acid; (k) ammonia-saturated methanol. Tol: toluoyl; DMT: 4,4'-dimethoxytrityl; Ac: acetyl.

[Figure 23] Figure 23 shows a synthesis scheme for nucleoside derivatives of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde, as well as a synthesis scheme for 6-amino-9-(2-deoxy- β -D-ribofuranosyl)purine 5'- γ -amidotriphosphate. Reagents and abbreviations: (a) NaH,

2-deoxy-3,5-di-O-p-toluoyl- α -D-erythro-pentofuranosyl chloride, CH₃CN; (b) ammonia-saturated methanol; (c) 4,4'-dimethoxytrityl chloride, pyridine; (d) 2-cyanoethyl-N,N'-diisopropylamino chloro phosphoramidite, diisopropylethylamine, THF; (e) proton sponge, POCl₃, trimethyl phosphate, then tributylamine, bis(tributylammonium) pyrophosphate, DMF; (f) NaH, CH₃CN, then 2,3,5-tri-O-benzyl-D-ribofuranosyl chloride; (g) BBr₃, dichloromethane; (h) 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one, dioxane, pyridine, tributylamine, bis(tributylammonium) pyrophosphate, DMF, then I₂/pyridine, water, NH₄OH (for 5'- γ -amidotriphosphate); Tol: toluoyl; DMT: 4,4'-dimethoxytrityl; Ac: acetyl.

[Figure 24a] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24a shows ¹H NMR (270 MHz, DMOSO-d₆) spectra of Compounds 6 and 11.

[Figure 24b] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24b shows ¹H NMR (270 MHz, DMOSO-d₆) spectra of Compounds 6 and 11 (7.0-9.0 ppm).

[Figure 24c] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24c shows 1D NOE spectra of Compound 6 (in DMOSO-d₆).

[Figure 24d] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24d shows 1D NOE spectra of Compound 11 (in DMOSO-d₆).

[Figure 24e] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24e shows a ¹³C NMR (75 MHz, DMOSO-d₆) spectrum of Compound 6.

[Figure 24f] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24f shows a 2D COSY spectrum of Compound 6.

[Figure 24g] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24g shows a 2D NOESY spectrum of Compound 6.

[Figure 24h] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24h shows a 2D HSQC spectrum of Compound 6.

[Figure 24i] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24i shows a 2D HMBC spectrum of Compound 6.

[Figure 24j] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24j shows a 2D HMBC spectrum of Compound 6 (enlarged).

[Figure 24k] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24k shows a ^{13}C NMR (75 MHz, $\text{DMSO}-d_6$) spectrum of Compound 11.

[Figure 24l] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24l shows a 2D COSY spectrum of Compound 11.

[Figure 24m] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24m shows a 2D NOESY spectrum of Compound 11.

[Figure 24n] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24n shows a 2D HSQC spectrum of Compound 11.

[Figure 24o] Figure 24 shows NMR spectra for nucleosides

of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24o shows a 2D HMBC spectrum of Compound 11.

[Figure 24p] Figure 24 shows NMR spectra for nucleosides of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Ds). Figure 24p shows a 2D HMBC spectrum of Compound 11 (enlarged).

[Figure 25a] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25a shows a ^1H NMR (300 MHz, DMOSO-d_6) spectrum of Compound 17.

[Figure 25b] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25b shows a ^{13}C NMR (75 MHz, DMOSO-d_6) spectrum of Compound 17.

[Figure 25c] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25c shows a 2D COSY spectrum of Compound 17.

[Figure 25d] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25d shows a 2D HSQC spectrum of Compound 17.

[Figure 25e] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25e shows a 2D HMBC spectrum of Compound 17.

[Figure 25f] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25f shows a 2D NOESY spectrum of

Compound 17.

[Figure 25g] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25g shows a ^1H NMR (300 MHz, DMOSO-d_6) spectrum of Compound 18.

[Figure 25h] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25h shows a ^{13}C NMR (75 MHz, DMOSO-d_6) spectrum of Compound 18.

[Figure 25i] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25i shows a 2D COSY spectrum of Compound 18.

[Figure 25j] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25j shows a 2D NOESY spectrum of Compound 18.

[Figure 25k] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25k shows a 2D HSQC spectrum of Compound 18.

[Figure 25l] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25l shows a 2D HMBC spectrum of Compound 18.

[Figure 25m] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25m shows 1D NOE spectra of Compound 17

(in DMOSO- d_6).

[Figure 25n] Figure 25 shows NMR spectra for nucleosides of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde. Figure 25n shows 1D NOE spectra of Compound 18 (in DMOSO- d_6).

[Figure 26] Figure 26 shows DEAE Sephadex ion exchange column elution patterns of deoxyadenosine 5'- γ -amidotriphosphate. Open circles indicate the results at 260 nm, while solid circles indicate the results at 280 nm.

[Figure 27] Figure 27 shows ESI-mass spectra of deoxyadenosine 5'- γ -amidotriphosphate.

[Figure 28a] Figure 28a shows a ^1H NMR (270 MHz, D_2O) spectrum of deoxyadenosine 5'- γ -amidotriphosphate.

[Figure 28b] Figure 28b shows a ^1P NMR (109 MHz, D_2O) spectrum of deoxyadenosine 5'- γ -amidotriphosphate.

[Figure 29a] Figure 29a shows ^1H NMR (270 MHz, D_2O) spectra of 7-(2-thienyl)-3-(2-deoxy- β -D-ribofuranosyl)3H-imidazo[4,5-b]pyridine 5'- γ -amidotriphosphate (upper) and 5'-triphosphate (lower).

[Figure 29b] Figure 29b shows ^1P NMR (109 MHz, D_2O) spectra of 7-(2-thienyl)-3-(2-deoxy- β -D-ribofuranosyl)3H-imidazo[4,5-b]pyridine 5'- γ -amidotriphosphate (upper) and 5'-triphosphate (lower).

[Figure 30a] Figure 30a shows a ^1H NMR (300 MHz, D_2O) spectrum of 7-(2-thienyl)-3-(β -D-ribofuranosyl)-3H-imidazo[4,5-b]pyridine 5'-triphosphate (Compound 14).

[Figure 30b] Figure 30b shows a ^1H NMR (270 MHz, D_2O) spectrum of 1-(β -D-ribofuranosyl)pyrrole-2-carbaldehyde

5'-triphosphate (Compound 19).

[Figure 30c] Figure 30c shows a ^1H NMR (270 MHz, D_2O) spectrum of 4-propynyl-1-(β -D-ribofuranosyl)pyrrole-2-carbaldehyde 5'-triphosphate (Compound 20).

[Figure 31a] Figure 31a shows a ^{31}P NMR (109 MHz, D_2O) spectrum of 7-(2-thienyl)-3-(β -D-ribofuranosyl)-3H-imidazo[4,5-b]pyridine 5'-triphosphate (Compound 14).

[Figure 31b] Figure 31b shows a ^{31}P NMR (109 MHz, D_2O) spectrum of 1-(β -D-ribofuranosyl)pyrrole-2-carbaldehyde 5'-triphosphate (Compound 19).

[Figure 31c] Figure 31c shows a ^{31}P NMR (109 MHz, D_2O) spectrum of 4-propynyl-1-(β -D-ribofuranosyl)pyrrole-2-carbaldehyde 5'-triphosphate (Compound 20).

[Figure 32] Figure 32 shows the results of electrospray ionization mass spectrum (ESI-MS) for $\text{C}_{35}\text{H}_{40}\text{O}_{17}\text{P}_2\text{S}$. $\text{C}_{35}\text{H}_{40}\text{O}_{17}\text{P}_2\text{S}$: calcd: 924.17(1-); 461.58(2-), found: 924.02(1-); 461.70(2-)

[Figure 33] Figure 33 shows a synthesis scheme for nucleoside derivatives of 7-(2-thiazolyl)-3H-imidazo[4,5-b]pyridine. Reagents and abbreviations: (a) dichlorobis(triphenylphosphine)palladium, 2-(tributylstanyl)thiazole, DMF; (b) palladium carbon, sodium borohydride, ethanol, ethyl acetate; (c) formic acid; (d) NaH, 2-deoxy-3,5-di-O-p-toluoyl- α -D-erythro-pentofuranosyl chloride, CH_3CN ; (e) ammonia-saturated methanol; (f) 4,4'-dimethoxytrityl chloride, pyridine; (g) 2-cyanoethyl-N,N-diisopropylamino chloro phosphoroamidite, diisopropylethylamine, THF; (h) acetic anhydride, pyridine, then dichloroacetic acid, dichloromethane; (i) 2-chloro-4H-

1,3,2,-benzodioxaphosphorin-4-one, dioxane, pyridine, tri-n-butylamine, bis(tributylammonium) pyrophosphate, I₂/pyridine, water, NH₄OH. Tol: toluoyl; DMT: 4,4'-dimethoxytrityl; Ac: acetyl.

[Figure 34] Figure 34 shows a synthesis scheme for nucleoside derivatives of 2-nitropyrrole (Compound 1). Reagents and abbreviations: (a) NaH, 2-deoxy-3,5-di-O-p-toluoyl- α -D-erythro-pentofuranosyl chloride, CH₃CN; (b) ammonia-saturated methanol; (c) 4,4'-dimethoxytrityl chloride, pyridine; (d) 2-cyanoethyl-N,N'-diisopropylamino chloro phosphoramidite, diisopropylethylamine, THF; (e) acetic anhydride, pyridine, then dichloroacetic acid, dichloromethane; (f) 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one, dioxane, pyridine, tri-n-butylamine, bis(tributylammonium) pyrophosphate, I₂/pyridine, water, NH₄OH. Tol: toluoyl; DMT: 4,4'-dimethoxytrityl; Ac: acetyl.

[Figure 35] Figure 35 shows a synthesis scheme for nucleoside derivatives of 4-(2-thienyl)-1H-pyrrolo[2,3-b]pyridine (DDs) and 4-(2-thienyl)-1H-pyrrolo[2,3-b]pyridine (DDv). Reagents and abbreviations: (a) mCPBA, EtOAc, then methanesulfonyl chloride, DMF; (b) NaI, CH₃COCl, CH₃CN; (c) NaH, 2-deoxy-3,5-di-O-p-toluoyl- α -D-erythro-pentofuranosyl chloride, CH₃CN; (d) dichlorobis(triphenylphosphine)palladium, 2-(tributylstanyl)thiophene or 2-(tributylstanyl)thiazole, DMF; (e) ammonia-saturated methanol; (f) 4,4'-dimethoxytrityl chloride, pyridine; (g) 2-cyanoethyl-N,N'-diisopropylamino chloro phosphoroamidite, diisopropylethylamine, THF; (h) acetic anhydride, pyridine, then dichloroacetic acid,

dichloromethane; (i) 2-chloro-4H-1,3,2,-benzodioxaphosphorin-4-one, dioxane, pyridine, tri-n-butylamine, bis(tributylammonium) pyrophosphate, I₂/pyridine, water, NH₄OH. Tol: toluoyl; DMT: 4,4'-dimethoxytrityl; Ac: acetyl.

[Figure 36] Figure 36 shows a synthesis scheme for 1-(β-D-ribofuranosyl)-4-[(3-biotinamido-1-propynyl)]pyrrole-2-carbaldehyde 5'-triphosphate (Compound 28). Reagents and abbreviations: (a) NaH, CH₃CN, then 2,3,5-tri-O-benzyl-D-ribofuranosyl chloride; (b) BBr₃, dichloromethane; (c) 3-(dichloroacetamido)-1-propyne, tetrakis(triphenylphosphine)palladium, CuI, triethylamine, DMF; (d) 4,4'-dimethyltrityl chloride, pyridine; (e) acetic anhydride, pyridine, then dichloroacetic acid, dichloromethane; (f) 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one, dioxane, pyridine, tri-n-butylamine, bis(tributylammonium) pyrophosphate, DMF, then I₂/pyridine, water, NH₄OH; (g) biotin-N-hydroxysuccinimide, DMF, sodium carbonate buffer (pH 8.6), then NH₄OH. DMTr: 4,4'-dimethoxytrityl; Ac: acetyl.

[Figure 37] Figure 37 shows a ¹H NMR (500 MHz, DMSO-d₆) spectrum for ribonucleoside of 4-iodopyrrole-2-carbaldehyde.

[Figure 38a] Figure 38 shows NMR spectra for 1-(β-D-ribofuranosyl)-4-[(3-dichloroacetamido)-1-propynyl]pyrrole-2-carbaldehyde (Compound 24). Figure 38a shows a ¹H NMR (300 MHz, DMSO-d₆) spectrum of Compound 24.

[Figure 38b] Figure 38 shows NMR spectra for 1-(β-D-ribofuranosyl)-4-[(3-dichloroacetamido)-1-propynyl]pyrrole-2-carbaldehyde (Compound 24). Figure 38b shows a ¹³C NMR

(75 MHz, DMSO-d₆) spectrum of Compound 24.

[Figure 38c] Figure 38 shows NMR spectra for 1-(β-D-ribofuranosyl)-4-[(3-dichloroacetamido)-1-propynyl]pyrrole-2-carbaldehyde (Compound 24). Figure 38c shows a 2D COSY spectrum of Compound 24.

[Figure 38d] Figure 38 shows NMR spectra for 1-(β-D-ribofuranosyl)-4-[(3-dichloroacetamido)-1-propynyl]pyrrole-2-carbaldehyde (Compound 24). Figure 38d shows a 2D NOESY spectrum of Compound 24.

[Figure 38e] Figure 38 shows NMR spectra for 1-(β-D-ribofuranosyl)-4-[(3-dichloroacetamido)-1-propynyl]pyrrole-2-carbaldehyde (Compound 24). Figure 38e shows a 2D HMQC spectrum of Compound 24.

[Figure 38f] Figure 38 shows NMR spectra for 1-(β-D-ribofuranosyl)-4-[(3-dichloroacetamido)-1-propynyl]pyrrole-2-carbaldehyde (Compound 24). Figure 38f shows a 2D HMBC spectrum of Compound 24.

[Figure 38g] Figure 38 shows NMR spectra for 1-(β-D-ribofuranosyl)-4-[(3-dichloroacetamido)-1-propynyl]pyrrole-2-carbaldehyde (Compound 24). Figure 38g shows a 2D HMBC spectrum of Compound 24 (enlarged).

[Figure 39] Figure 39 shows a ¹H NMR (500 MHz, DMSO-d₆) spectrum of 1-[5-O-(4,4'-dimethoxytrityl)-β-D-ribofuranosyl]-4-[(3-dichloroacetamido)-1-propynyl]pyrrole-2-carbaldehyde (Compound 25).

[Figure 40] Figure 40 shows a ¹H NMR (500 MHz, DMSO-d₆) spectrum of 1-(2,3-di-O-acetyl-β-D-ribofuranosyl)-4-[(3-dichloroacetamido)-1-propynyl]pyrrole-2-carbaldehyde

(Compound 26).

[Figure 41] Figure 41 shows DEAE Sephadex ion exchange column elution patterns of 1-(β -D-ribofuranosyl)-4-[(3-amino-1-propynyl)]pyrrole-2-carbaldehyde 5'-triphosphate (Compound 27) and 1-(β -D-ribofuranosyl)-4-[(3-biotinamido-1-propynyl)]pyrrole-2-carbaldehyde 5'-triphosphate (Compound 28).

[Figure 42] Figure 42 shows ESI mass spectra of 1-(β -D-ribofuranosyl)-4-[(3-amino-1-propynyl)]pyrrole-2-carbaldehyde 5'-triphosphate (Compound 27; upper) and 1-(β -D-ribofuranosyl)-4-[(3-biotinamido-1-propynyl)]pyrrole-2-carbaldehyde 5'-triphosphate (Compound 28; lower).

[Figure 43a] Figure 43 shows NMR spectra for 1-(β -D-ribofuranosyl)-4-[(3-biotinamido-1-propynyl)]pyrrole-2-carbaldehyde 5'-triphosphate (Compound 28). Figure 43a shows a ^1H NMR (300 MHz, D_2O) spectrum of Compound 28.

[Figure 43b] Figure 43 shows NMR spectra for 1-(β -D-ribofuranosyl)-4-[(3-biotinamido-1-propynyl)]pyrrole-2-carbaldehyde 5'-triphosphate (Compound 28). Figure 43b shows a 2D COSY spectrum of Compound 28.

[Figure 43c] Figure 43 shows NMR spectra for 1-(β -D-ribofuranosyl)-4-[(3-biotinamido-1-propynyl)]pyrrole-2-carbaldehyde 5'-triphosphate (Compound 28). Figure 43c shows a ^{31}P NMR (109 MHz, D_2O) spectrum of Compound 28.

[Figure 44a] Figure 44a shows the chemical structure of biotinylated PaTP (Bio-PaTP; Compound 28).

[Figure 44b] Figure 44b shows an experimental scheme for transcription and transcript analysis on site-specific

biotinylation of RNA molecules by T7 transcription through Ds-Pa base pairing.

[Figure 44c] Figure 44c is a photograph showing gel electrophoresis of transcripts obtained in the site-specific biotinylation experiment on RNA molecules by T7 transcription through Ds-Pa base pairing. Lanes 1-4 indicate the results with the use of templates obtained by ligation, while Lanes 5-12 indicate the results with the use of templates obtained by PCR amplification. The templates used were DNA6 (Lanes 1, 2, 5, 6, 9 and 10) and DNAcont2 (as a control; Lanes 3, 4, 7, 8, 11 and 12). Transcription reaction was performed in the presence (2 mM) or absence of Bio-PaTP together with natural NTPs (2 mM). The transcripts were labeled with $[\gamma\text{-}^{32}\text{P}]\text{GTP}$.

[Figure 44d] Figure 44d is photographs showing the results of gel shift assay for analysis of biotinylated transcripts. 152-mer RNAs transcribed with Bio-PaTP from DNA6 template obtained by ligation and DNAcont2 template obtained by PCR amplification (20 cycles), as well as 152-mer RNAs transcribed with biotinylated UTP (Bio-UTP) from DNAcont5 and DNAcont6 templates obtained by ligation were mixed with streptavidin. Biotinylated RNA-streptavidin complexes were separated from free RNAs on a 7% polyacrylamide-7 M urea gel, and the percentages (yields) of the complexes were determined from their band intensity.

[Figure 44e] Figure 44e is a photograph showing the results of sequence analysis on 152-mer transcripts containing Bio-Pa or A at position 59. The 5'-terminally ^{32}P -labeled transcripts were partially digested with RNase T1 (T1) or

alkali (AL). A portion of each partially alkali-digested transcript was treated with streptavidin magnetic beads to capture RNA fragments containing Bio-Pa (AL+SA), and the rest was electrophoresed. The digested fragments were each analyzed on a 10% polyacrylamide-7 M urea gel.

[Figure 45] Figure 45 shows a synthesis scheme for 4-position modified nucleoside derivatives of 2-nitropyrrole. Reagents and abbreviations: (a) N-iodosuccinimide, CH_3CN ; (b) propynyl-1-tributyltin, $\text{Pd}(\text{PPh}_3)_2\text{Cl}_2$, DMF or N-(2-propynyl)-dichloroacetamide, $\text{Pd}(\text{PPh}_3)_4$, CuI, triethylamine, DMF; (c) 4,4'-dimethoxytrityl chloride, pyridine; (d) 2-cyanoethyl-N,N-diisopropylamino chloro phosphoramidite, diisopropylethylamine, THF; (e) acetic anhydride, pyridine, then dichloroacetic acid, dichloromethane; (f) 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one, dioxane, pyridine, tri-n-butylamine, bis(tributylammonium) pyrophosphate, $\text{I}_2/\text{pyridine}$, water, NH_4OH . DMTr: 4,4'-dimethoxytrityl; Ac: acetyl.

[Figure 46] Figure 46 shows a synthesis scheme for $\text{NH}_2\text{-hx-dPnTP}$, ROX-hx-dPnTP and FAM-hx-dPnTP . Reagents and abbreviations: (a) CuI, $\text{Pd}[\text{P}(\text{C}_6\text{H}_5)_3]_4$, DMF, triethylamine, room temperature, then N-(2-propynyl)-6-trifluoroacetamidohexanamide; (b) DMTr-Cl, pyridine, room temperature; (c) acetic anhydride, pyridine, room temperature, then dichloroacetic acid, dichloromethane, 0°C ; (d) 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one/dioxane, pyridine, tri-n-butylamine, bis(tri-n-butylammonium) pyrophosphate, DMF, then $\text{I}_2/\text{pyridine}$, water, NH_4OH , room temperature; (e) R-N-hydroxysuccinimidyl ester (R = FAM or ROX)/DMF, 0.1 M NaHCO_3 -

Na₂CO₃ buffer (pH 8.5), room temperature, 8 hours, then NH₄OH.

[Figure 47] Figure 47 shows a synthesis scheme for NH₂-hx-PaTP, FAM-hx-PaTP and TAMRA-hx-PaTP. Reagents and abbreviations: (a) CuI, Pd[P(C₆H₅)₃]₄, DMF, triethylamine, room temperature, then N-(2-propynyl)-6-trifluoroacetamidohexanamide; (b) DMTr-Cl, pyridine, room temperature; (c) acetic anhydride, pyridine, room temperature, then dichloroacetic acid, dichloromethane, 0°C; (d) 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one/dioxane, pyridine, tri-n-butylamine, bis(tri-n-butylammonium) pyrophosphate, DMF, then I₂/pyridine, water, NH₄OH, room temperature; (e) R-N-hydroxysuccinimidyl ester (R = FAM or TAMRA)/DMF, 0.1 M NaHCO₃-Na₂CO₃ buffer (pH 8.5), room temperature, 8 hours, then NH₄OH.

[Figure 48] Figure 48 shows a scheme for incorporation of substrate Pn having an amino group or a fluorescent dye into DNA (55-mer) through replication using Klenow fragment, along with the results obtained.

[Figure 49] Figure 49 shows a scheme for incorporation of substrate Pa having a fluorescent dye into RNA (17-mer) through transcription using T7 RNA polymerase, along with the results obtained.

EXAMPLES

[0192] The present invention will now be further described in the following examples, which are not intended to limit the technical scope of the invention. Based on the detailed description, various modifications and changes will be apparent to those skilled in the art, and such modifications

and changes fall within the technical scope of the invention.

[0193] Example I Chemical synthesis-1

1. General methods and materials

All reagents and solvents were purchased from standard suppliers and were used without further purification. Two-dimensional thin-layer chromatography (2D TLC) was performed using 0.25 mm silica gel 60 plates containing a 254 nm fluorescence indicator (Merck) to monitor transcription reaction. ^1H NMR, ^{13}C NMR and ^{31}P NMR spectra were recorded on JEOL EX270 and BRUKER nuclear magnetic resonance spectrometers (300 MHz and 600 MHz). Nucleoside purification was performed on a Gilson HPLC system equipped with a preparative C18 column (Waters Microbond Sphere, 150 × 19 mm). Triphosphate derivatives were purified with a DEAE-Sephadex A-25 column (300 × 15 mm) and an analytical C18 column (Synchropak RPP, 250 × 4.6 mm, Eichrom Technologies).

[0194] High-resolution mass spectra (HRMS) and electrospray ionization mass spectra (ESI-MS) were recorded on a JEOL HX-110 or JM700 mass spectrometer and a Waters micro mass ZMD4000 equipped with a Waters 2690 LC system, respectively. Fluorescence measurement was accomplished by using a FP-6500 spectral fluorometer (JASCO).

[0195] Pyrrole-2-carbaldehyde (Non-patent Document 39) and 4-propynylpyrrole-2-carbaldehyde (Non-patent Document 40) were synthesized as described in these prior documents.

[0196] 2. Explanation of compound synthesis

Synthesis of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine was accomplished through the reactions shown in Figure 22. More

specifically, 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Compound 4) was synthesized from 2-amino-3-nitro-4-chloropyridine (Compound 1) (Non-patent Document 41) in 3 steps (72%). A deoxyribonucleoside of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (shown as Compound 6) was obtained as a single product from Compound 4 in a yield of 61% by reaction of 1-chloro-2-deoxy-3,5-di-O-toluoyl- α -D-erythro-pentofuranose (Non-patent Document 42) with a sodium salt of Compound 4, followed by deprotection of the toluoyl groups from Compound 5 with methanolic ammonium.

[0197] A ribonucleoside of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (shown as Compound 11) was synthesized by reaction between tetra-O-acetyl- β -D-ribofuranose and 4 at 200°C with a catalytic amount of chloroacetic acid. Compound 11 was obtained in a yield of 29% after deprotection with ammonia-saturated methanol and purification by RP-HPLC.

[0198] The structures of Nucleosides 6 and 11 were confirmed by NMR and high-resolution mass spectroscopy (NMR spectra of Compounds 6 and 11 in the section "1. General methods and materials"). The aromatic proton peaks of Compounds 6 and 11 showed the same chemical shifts. The HMBC and HSQC spectra of Compounds 6 and 11 indicated that an N-glycosidic linkage was formed between sugar C1' and N-3 position of the imidazo[4,5-b]pyridine base moiety. Further, the inventors of the present invention confirmed that the thienyl moiety of Compounds 6 and 11 was attached at the 7-position of the imidazo[4,5-b]pyridine ring, as is seen from their 2D NOESY and 2D HMBC spectra. The anomeric

configurations of Compounds 6 and 11 were confirmed to be β by their 2D NOESY and 1D NOE spectra. The main results of the 1D NOE experiment are that H1' proton irradiation causes enhancements of 2% and 3% in the H4' signal and enhancements of 8% and 9% in the H2 signal in Compounds 6 and 11, respectively. In the differential NOE experiment, H2 proton irradiation caused enhancements of 9% and 10% in the H1' signal and an enhancement of 3% to 5% in the H2' and H3' signals. Thus, based on the NOE experiment, the anomeric configurations of Compounds 6 and 11 were determined to be β . Compound 6 was converted into its amidite form in a standard manner, and the formation of a Tp(6)pT trimer was confirmed by electrospray ionization mass spectrometry (ESI-MS) (see the trimer's mass spectrum in "nucleoside derivatives of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde, as well as 6-amino-9-(2-deoxy- β -D-ribofuranosyl)purine 5'- γ -amidotriphosphate"). Nucleoside 5'-triphosphates shown as Compounds 10 and 14 were synthesized in a standard manner (Non-patent Document 43) for use as substrates in enzymatic reactions.

[0199] Synthesis of nucleoside derivatives of pyrrole-2-carbaldehyde (Compound 15) and 4-propynylpyrrole-2-carbaldehyde (Compound 16) was accomplished by the reactions shown in Figure 23. Synthesis of deoxynucleoside derivatives of Compounds 15 and 16 has been reported (Non-patent Documents 39 and 40). Ribonucleosides of Compounds 15 and 16 were synthesized by reaction of 2,3,5-tri-O-benzyl-D-ribofuranosyl chloride (Non-patent Document 44) with a sodium salt of

Compound 15 or 16, followed by BBr₃ treatment to deprotect the benzyl groups, thereby obtaining ribonucleosides shown as Compounds 17 (yield 15%) and 18 (yield 7%) in 2 steps.

[0200] The structures of Compounds 17 and 18 were confirmed by NMR (see NMR spectra of Compounds 17 and 18 shown in Figure 25) and high-resolution mass spectroscopy. The HMBC and HSQC spectra of Compounds 17 and 18 indicated that an N-glycosidic linkage was formed between sugar and pyrrole base moiety at the C1' carbon. The anomeric configurations of Compounds 17 and 18 were confirmed by the NOE experiment (differential NOE and NOESY spectra).

[0201] The main results of the differential NOESY spectrum experiment are as follows. H1' proton irradiation caused an enhancement of 3% to 4% in the H4' signal. H2' (and/or H3') proton irradiation caused an enhancement of 9% to 10% in the H5 signal. The NOESY spectra of Compounds 17 and 18 showed cross peaks between H1' and H4', between H1' and CHO proton, and between H5 and H2' (and/or H3'). Thus, based on NOE NMR, the anomeric configurations of Compounds 17 and 18 were determined to be β . The ribonucleosides shown as Compounds 17 and 18 were converted into their ribonucleoside 5'-triphosphate form in a standard manner (Non-patent Document 45).

[0202] Nucleoside 5'-triphosphates were synthesized as disclosed in prior art documents (Non-patent Documents 43 and 45). Synthesis of 5'- γ -amidotriphosphates was accomplished in a standard manner (Non-patent Document 43) with modifications. Protected nucleosides shown as Compounds 9 and 21 were

phosphitylated with 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one. The protected nucleoside phosphite derivatives were converted into P^2, P^3 -dioxo- P^1 -5'-nucleosidylcyclotriphosphites by treatment with pyrophosphate. After treatment with iodine/water, the resulting 5'-trimetaphosphates were treated with concentrated aqueous ammonia to give nucleoside 5'- γ -amidotriphosphates.

[0203] Purification of these 5'- γ -amidotriphosphates was performed by anion exchange DEAE Sephadex column chromatography and RP-HPLC. The DEAE column elution patterns and electrospray ionization mass spectra of deoxyadenosine 5'- γ -amidotriphosphate are shown in Figures 26 and 27.

[0204] During 5'- γ -amidotriphosphate synthesis, 5'-triphosphate of deoxyadenosine was also formed, and the 5'- γ -amidotriphosphate and 5'-triphosphate were in the ratio of 4.8:1, as calculated from their fraction absorbance. Compound 22 was eluted relatively faster than deoxyadenosine 5'-phosphate (dATP) and separated by DEAE column purification. The 5'- γ -amidotriphosphate was confirmed for its molecular weight by ESI-MS spectrometry. The difference between Compound 22 and dATP was 1 m/z. After final purification by HPLC, the nucleoside 5'- γ -amidotriphosphates were obtained as triethylammonium salts and confirmed for their structure by NMR spectroscopy (1H and ^{31}P NMR). γ -Phosphate signals from deoxyadenosine 5'- γ -amidotriphosphate and Compound 6 were shifted to lower magnetic field (-0.50 and -0.52 p.p.m), as compared to those of 5'-phosphates of Compounds 6, 11, 17 and 18. This phenomenon was also observed in guanosine 5'- γ -

amidotriphosphate.

[0205] 3. Detailed explanation of synthesis

(1) 2-Amino-3-nitro-4-(2-thienyl)pyridine (Compound 2)

To a solution of 2-amino-3-nitro-4-chloropyridine (Compound 1) (Non-patent Document 41) (1.74 g, 10 mmol) and dichlorobis(triphenylphosphine)palladium (II) (350 mg, 0.50 mmol) in DMF (50 ml), 2-(tributylstanyl)thiophene (3.82 ml, 12 mmol) was added under an argon atmosphere. The resulting mixture was stirred at 100°C for 4 hours. The mixture was poured into water (250 ml) and then extracted with ethyl acetate (250 ml × 3). After drying over Na₂SO₄, the solvent was distilled off under reduced pressure. The residue was applied to flash silica gel chromatography using methylene chloride:ethyl acetate (100:1 to 49:1) as an eluent to give 2.07 g of Compound 2 (R_f 0.30 on methylene chloride:ethyl acetate = 19:1) in a yield of 93%.

[0206] ¹H NMR (270 MHz, CDCl₃) δ

8.17 (d, 1H, J=5.1 Hz),

7.4 (dd, 1H, J=5.0 and 1.1 Hz),

7.12 (dd, 1H, J=3.6 and 1.1 Hz),

7.07 (dd, 1H, J=5.0 and 3.6 Hz),

6.77 (d, 1H, J=5.1 Hz), 5.66 (bs, 2H).

[0207] HRMS (FAB, 3-NBA, matrix)

C₉H₈N₃O₂S (M+1): calcd, 222.0337; found, 222.0337.

[0208] (2) 2,3-Diamino-4-(2-thienyl)pyridine (Compound 3)

To a mixture of Compound 2 (2.06 g, 9.3 mmol) and 466 mg palladium carbon (10% by weight) in ethanol (130 ml) and ethyl acetate (65 ml), 28 ml of 1 M aqueous sodium borohydride was

added at 0°C. The resulting mixture was stirred at 0°C for 1 hour. To the mixture, 43 ml of 5% aqueous ammonium chloride was added. The mixture was filtered through celite. The filtrate was diluted with 500 ml water. After evaporation of ethanol and ethyl acetate, the mixture was extracted with ethyl acetate (250 ml × 3). After drying over Na₂SO₄, the solvent was distilled off. The residue was purified by flash silica gel chromatography using methylene chloride:ethyl acetate (19:1 to 93:7) as an elution solvent to give 1.46 g of Compound 3 (R_f 0.24 on methylene chloride:ethyl acetate = 9:1) in a yield of 82%.

[0209] ¹H NMR (270 MHz, CDCl₃) δ

7.64 (d, 1H, J=5.1 Hz),
7.40 (dd, 1H, J=5.1 and 1.1 Hz),
7.23 (dd, 1H, J=3.5 and 1.1 Hz),
7.14 (dd, 1H, J=5.1 and 3.5 Hz),
6.74 (d, 1H, J=5.1 Hz),
4.26 (bs, 2H), 3.72 (bs, 2H).

[0210] HRMS (FAB, 3-NBA matrix)

C₉H₁₀N₃S (M+1): calcd, 192.0595; found, 192.0588.

[0211] (3) 7-(2-Thienyl)-3H-imidazo[4,5-b]pyridine
(Compound 4)

A solution of Compound 3 (956 mg, 5.0 mmol) in formic acid (15 ml) was refluxed for 12 hours. To the reaction mixture, 24 ml of 28% NH₄OH was added on an ice-cold bath. The resulting precipitate was filtered, washed with H₂O and ethyl ether, and then dried at 60°C for 12 hours to give 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (970 mg, 96%).

[0212] ^1H NMR (300 MHz, DMSO-d_6) δ

13.20 (s, 1H), 8.48 (s, 1H), 8.30 (m, 2H),

7.78 (dd, 1H, $J=1.0$ and 5.1 Hz),

7.54 (d, 1H, $J=5.1$ Hz),

7.25 (dd, 1H, $J=3.8$ and 4.9 Hz).

[0213] ^{13}C NMR (75 MHz, DMSO-d_6) δ

113.12, 128.00, 128.71, 129.24,

130.00, 131.16, 137.60, 143.44,

144.09, 148.66.

[0214] HRMS (FAB, 3-NBA matrix)

$\text{C}_{10}\text{H}_8\text{N}_3\text{S}$ ($M+1$): calcd, 202.0439; found, 202.0444.

[0215] (4) 7-(2-Thienyl)-3-[2-deoxy-3,5-di-O-(toluoyl)- β -D-ribofuranosyl]-3H-imidazo[4,5-b]pyridine (Compound 5)

To a solution of Compound 4 (403 mg, 2.0 mmol) in CH_3CN (32 ml), NaH (96 mg, 2.4 mmol, 60% dispersion in mineral oil) was added. The resulting mixture was stirred at room temperature for 1 hour. To the mixture, 2-deoxy-3,5-di-O-p-toluoyl- α -D-erythro-pentofuranosyl chloride (933 mg, 2.4 mmol) (Non-patent Document 42) was added. After stirring at room temperature for 2.5 hours, the reaction mixture was partitioned between ethyl acetate and water. The organic layer was washed three times with saturated aqueous sodium chloride, dried over Na_2SO_4 , and then evaporated under reduced pressure. The product was purified by silica gel column chromatography (0.5% methanol in CH_2Cl_2) to give Compound 5 (714 mg, 65%).

[0216] ^1H NMR (270 MHz, CDCl_3) δ

8.32 (d, 1H, $J=5.3$ Hz), 8.26 (s, 1H),

8.16 (dd, 1H, J=3.8 and 1.2 Hz),
7.93 (m, 4H), 7.50 (dd, 1H, J=5.1 and 1.2 Hz),
7.47 (d, 1H, J=5.3 Hz), 7.22 (m, 5H),
6.68 (dd, 1H, J=8.6 and 5.8 Hz),
5.82 (m, 1H), 4.69 (m, 3H),
3.18 (ddd, 1H, J=14.2, 8.6 and 6.4 Hz),
2.86 (ddd, 1H, J=14.2, 5.8 and 2.0 Hz),
2.43 (s, 3H), 2.37 (s, 3H).

[0217] HRMS (FAB, 3-NBA matrix)

C₃₁H₂₈N₃O₅S (M+1): calcd, 554.1750; found, 554.1748.

[0218] (5) 7-(2-Thienyl)-3-(2-deoxy-β-D-ribofuranosyl)-3H-imidazo[4,5-b]pyridine (Compound 6)

To 1.33 g (2.40 mmol) of Compound 5, ammonia-saturated methanol (120 ml) was added at 0°C. The solution was stirred at room temperature for 2 hours. After the reaction, the solvent was distilled off and the residue was purified by flash silica gel chromatography using methylene chloride:ethanol (97:3 to 93:7) as an elution solvent to give 717 mg of Compound 6 in a yield of 94%.

[0219] ¹H NMR (300 MHz, DMSO-d₆) δ

8.75 (s, 1H), 8.35 (d, 1H, J=5.1 Hz),
8.30 (d, 1H, J=3.7 Hz), 7.83 (d, 1H, J=5.1 Hz),
7.65 (d, 1H, J=5.1 Hz), 7.28 (t, 1H, J=4.2 Hz),
6.54 (t, 1H, J=6.9 Hz), 5.34 (d, 1H, J=4.1 Hz),
5.11 (t, 1H, J=5.7 Hz), 4.46 (m, 1H),
3.91 (m, 1H), 3.60 (m, 2H),
2.80 (m, 1H), 2.37 (m, 1H).

[0220] ¹³C NMR (75 MHz, DMSO-d₆) δ

147.10, 144.04, 143.62, 137.06,
132.00, 131.00, 129.78, 129.06,
128.07, 113.93, 87.88, 83.72,
70.80, 61.74, 39.40.

[0221] HRMS (FAB, 3-NBA matrix)

C₁₅H₁₆N₃O₃S (M+1): calcd, 318.0912; found, 318.0905.

[0222] UV: λ_{max} , 311 nm;

$\epsilon = 2.04 \times 10^4$ in 25 mM sodium phosphate buffer (pH 6.8).

[0223] (6) 7-(2-Thienyl)-3-[2-deoxy-5-O-(4,4'-dimethoxytrityl)- β -D-ribofuranosyl]-3H-imidazo[4,5-b]pyridine (Compound 7)

A 317 mg (1.0 mmol) portion of Compound 6 was azeotroped three times with dry pyridine. To this, 5.0 ml anhydrous pyridine was added and dimethoxytrityl chloride (356 mg, 1.1 mmol) was further added. The mixture was stirred overnight at room temperature, poured into water (50 ml) and then extracted with methylene chloride (50 ml $\times$ 3). After drying over Na₂SO₄, the solvent was distilled off under reduced pressure. The residue was purified by flash silica gel chromatography using methylene chloride:ethyl acetate (9:1 to 13:7) as an elution solvent to give 550 mg of Compound 7 in a yield of 89%.

[0224] ¹H NMR (270 MHz, CDCl₃) δ

8.29 (d, 1H, J=5.1 Hz), 8.22 (s, 1H),
8.17 (dd, 1H, J=3.8 and 1.1 Hz),
7.49 (dd, 1H, J=5.1 and 1.1 Hz),
7.44 (d, 1H, J=5.1 Hz), 7.37 (m, 2H),
7.27 (m, 5H), 7.20 (m, 3H), 6.78 (m, 4H),
6.57 (dd, 1H, J=6.5 and 6.2 Hz),

4.66 (m, 1H), 4.12 (m, 1H), 3.75 (s, 3H),
3.75 (s, 3H),
3.42 (dd, 1H, J=10.1 and 4.6 Hz),
3.38 (dd, 1H, J=10.1 and 5.4 Hz),
2.86 (m, 1H),
2.56 (ddd, 1H, J=13.8, 6.5 and 4.6 Hz),
2.06 (d, 1H, J=3.5 Hz).

[0225] HRMS (FAB, 3-NBA matrix)

$C_{36}H_{34}N_3O_5S$ (M+1): calcd, 620.2219; found, 620.2230.

[0226] (7) 7-(2-Thienyl)-3-[2-deoxy-5-O-(4,4'-dimethoxytrityl)- β -D-ribofuranosyl]-3H-imidazo[4,5-b]pyridine 2-cyanoethyl-N,N-diisopropylphosphoramidite (Compound 8)

Compound 7 (425 mg, 0.69 mmol) was azeotroped three times with anhydrous pyridine and then three times with anhydrous acetonitrile. This was dissolved in anhydrous acetonitrile (4.6 ml), followed by addition of 2-cyanoethyl tetraisopropylphosphorodiamidite (262 μ l, 0.82 mmol) and a 0.45 M acetonitrile solution of tetrazole (1.68 ml). This mixture was stirred for 1 hour at room temperature. After addition of anhydrous methanol (90 μ l), the mixture was poured into water (50 ml) and extracted with 1% triethylamine (v/v)-containing methylene chloride (50 ml $\times$ 3). After drying over Na_2SO_4 , the solvent was distilled off under reduced pressure. The residue was purified by flash silica gel chromatography using 2% triethylamine (v/v)-containing hexane:ethyl acetate (4:1 to 3:2) as an elution solvent to give 490 mg of Compound 8 in a yield of 87%.

[0227] 1H NMR (270 MHz, DMSO- d_6) δ

8.65 (m, 1H), 8.27 (m, 1H), 8.23 (m, 1H),
7.81 (m, 1H), 7.62 (m, 1H), 7.26 (m, 3H),
7.18 (m, 7H), 6.75 (m, 4H), 6.54 (m, 1H),
4.81 (m, 1H), 4.13 (m, 1H),
3.84-3.45 (m, 10H), 3.21 (m, 3H),
2.82-2.48 (m, 3H) 1.13 (m, 12H).

[0228] ^{31}P NMR (109 MHz, DMSO- d_6) δ

148.76 ppm and 148.14 ppm.

[0229] HRMS (FAB, 3-NBA matrix)

$\text{C}_{45}\text{H}_{51}\text{N}_5\text{O}_6\text{SP}$ (M+1): calcd, 820.3298; found, 820.3325.

[0230] (8) 7-(2-Thienyl)-3-(2-deoxy-3-O-acetyl- β -D-ribofuranosyl)-3H-imidazo[4,5-b]pyridine (Compound 9)

Compound 7 (124 mg, 0.20 mmol) was azeotroped three times with anhydrous pyridine. This was dissolved in anhydrous pyridine (2.0 ml), and 38 μl (0.40 mmol) of acetic anhydride was further added thereto. The resulting mixture was stirred at room temperature for 2 days. The mixture was poured into water (50 ml) and extracted with methylene chloride (50 ml $\times$ 3). After drying over Na_2SO_4 , the solvent was distilled off under reduced pressure. The residue was dissolved in 20 ml anhydrous methylene chloride, and 200 μl dichloroacetic acid was added thereto at 0°C. After stirring at 2°C for 15 minutes, the mixture was poured into 8 ml saturated aqueous sodium bicarbonate, diluted with 42 ml water, and then extracted with methylene chloride (50 ml $\times$ 3). After drying over Na_2SO_4 , the solvent was distilled off under reduced pressure. The residue was purified by flash silica gel chromatography using methylene chloride:ethyl acetate (9:1 to

3:2) as an elution solvent to give 65 mg of Compound 9 in a yield of 88%.

[0231] ^1H NMR (270 MHz, CDCl_3) δ

8.29 (d, 1H, $J=5.4$ Hz),
8.18 (dd, 1H, $J=3.8$ and 1.1 Hz),
8.13 (s, 1H), 7.53 (dd, 1H, $J=5.0$ and 1.1 Hz),
7.50 (d, 1H, $J=5.4$ Hz),
7.20 (dd, 1H, $J=5.0$ and 3.8 Hz),
6.69 (m, 1H), 6.37 (dd, 1H, $J=10.1$ and 5.4 Hz),
5.58 (m, 1H), 4.28 (m, 1H), 3.96 (m, 2H),
3.32 (ddd, 1H, $J=15.9$, 10.1 and 5.9 Hz),
2.41 (m, 1H), 2.12 (s, 3H).

[0232] HRMS (FAB, 3-NBA matrix)

$\text{C}_{17}\text{H}_{18}\text{N}_3\text{O}_4\text{S}$ ($M+1$): calcd, 360.1018; found, 360.0993.

[0233] (9) 7-(2-Thienyl)-3-(β -D-ribofuranosyl)-3H-imidazo[4,5-b]pyridine (Compound 11)

A mixture of Compound 4 (80 mg, 0.4 mmol), tetra-O-acetyl- β -D-ribofuranose (130 mg, 0.4 mmol) and chloroacetic acid (2 mg) was heated at 200°C for 5 minutes. The resulting dark syrup was treated with methanolic ammonia (40 ml) at room temperature for 18 hours. After the solvent was distilled off under reduced pressure, the residue was diluted with 30% aqueous CH_3CN , and the product was purified by reversed-phase HPLC to give Compound 11 (39 mg, 29%, 2 steps).

[0234] ^1H NMR (300 MHz, $\text{DMSO}-d_6$) δ

8.78 (s, 1H), 8.36 (d, 1H, $J=5.2$ Hz),
8.31 (dd, 1H, $J=1.0$ and 3.7 Hz),
7.84 (dd, 1H, $J=0.9$ and 5.1 Hz),

7.66 (d, 1H, J=5.2 Hz),
7.28 (dd, 1H, J=3.7 and 5.0 Hz),
6.08 (d, 1H, J=5.8 Hz), 5.49 (d, 1H, J=6.0 Hz),
5.26 (t, 1H, J=6.5 Hz), 5.20 (d, 1H, J=4.9 Hz),
4.67 (q, 1H, J=5.8 Hz), 4.20 (q, 1H, J=4.9 Hz),
4.00 (m, 1H), 3.71 (m, 1H), 3.59 (m, 1H).

[0235] ^{13}C NMR (75 MHz, DMSO- d_6) δ

147.29, 144.07, 143.91, 137.00,
132.13, 131.08, 129.86, 129.14,
128.10, 114.02, 87.76, 85.57,
73.48, 70.43, 61.43.

[0236] HRMS (FAB, 3-NBA matrix)

$\text{C}_{15}\text{H}_{16}\text{N}_3\text{O}_4\text{S}$ (M+1): calcd, 334.0862; found, 334.0871.

[0237] (10) 7-(2-Thienyl)-3-[5-O-(4,4'-dimethoxytrityl)- β -D-ribofuranosyl]-3H-imidazo[4,5-b]pyridine (Compound 12)

Compound 11 (99 mg, 0.29 mmol) was azeotroped three times with anhydrous pyridine and then dissolved in pyridine (3.0 ml). To this solution, 4,4'-dimethoxytrityl chloride (106 mg, 0.31 mmol) was added, and the resulting mixture was stirred at room temperature for 2 hours. The reaction mixture was poured into 5% aqueous NaHCO_3 and then extracted with ethyl acetate. The organic layer was washed three times with saturated aqueous sodium chloride, dried over Na_2SO_4 , and then evaporated under reduced pressure to remove the solvent. The product was purified by silica gel column chromatography (1% methanol- CH_2Cl_2) to give Compound 12 (131 mg, 71%).

[0238] ^1H NMR (600 MHz, CDCl_3) δ

8.39 (s, 1H), 8.28 (d, 1H, J=5.2 Hz),

8.25 (d, 1H, J=3.4 Hz), 7.52 (t, 2H, H=5.7 Hz),
7.22 (m, 2H), 7.14 (m, 7H), 6.69 (m, 5H),
6.04 (d, 1H, J=6.2 Hz), 4.77 (m, 1H),
4.48 (m, 1H), 4.34 (d, 1H, J=4.6 Hz),
3.70 (s, 6H), 3.46 (dd, 1H, J=3.4 and 10.5 Hz),
3.23 (dd, 1H, J=2.9 and 10.5 Hz).

[0239] HRMS (FAB, 3-NBA matrix)

$C_{36}H_{34}N_3O_6S$ (M+1): calcd, 636.2168; found, 636.2173.

[0240] (11) 7-(2-Thienyl)-3-(2,3-di-O-acetyl- β -D-ribofuranosyl)-3H-imidazo[4,5-b]pyridine (Compound 13)

Compound 12 (120 mg, 0.19 mmol) was azeotroped three times with anhydrous pyridine. This was dissolved in anhydrous pyridine (1.9 ml), and 72 μ l (0.76 mmol) of acetic anhydride was further added thereto. This mixture was stirred at room temperature for 7 hours and poured into 5% $NaHCO_3$ (50 ml) and ethyl acetate (50 ml). The organic layer was washed once with saturated aqueous sodium chloride, dried over Na_2SO_4 , and then evaporated under reduced pressure to remove the solvent. The residue was azeotroped twice with toluene and dissolved in CH_2Cl_2 (19 ml), followed by addition of 190 μ l dichloroacetic acid at 0°C. This reaction mixture was stirred at 0°C for 15 minutes. The mixture was poured into 5% $NaHCO_3$ and extracted with CH_2Cl_2 . The organic layer was washed once with saturated aqueous sodium chloride, dried over Na_2SO_4 , and then evaporated under reduced pressure to remove the solvent. The product was purified by flash silica gel chromatography using a 2% methanol-methylene chloride solution as a solvent to give 77 mg of Compound 13 in a yield of 93%.

[0241] ^1H NMR (300 MHz, DMSO- d_6) δ

8.79 (s, 1H), 8.37 (d, 1H, $J=5.2$ Hz),
8.29 (dd, 1H, $J=1.2$ and 3.7 Hz),
7.84 (dd, 1H, $J=1.1$ and 5.1 Hz),
7.68 (d, 1H, $J=5.2$ Hz),
7.27 (dd, 1H, $J=3.7$ and 5.1 Hz),
6.38 (d, 1H, $J=6.8$ Hz),
6.02 (dd, 1H, $J=5.6$ and 6.6 Hz),
5.54 (m, 2H), 4.26 (m, 1H), 3.71 (m, 2H),
2.14 (s, 3H), 1.98 (s, 3H).

[0242] ^{13}C NMR (75 MHz, DMSO- d_6) δ

169.55, 169.22, 146.98, 144.39,
143.85, 136.74, 132.50, 131.02,
130.11, 129.32, 128.14, 114.36,
85.20, 83.63, 72.34, 71.23, 61.05,
20.44, 20.13.

[0243] HRMS (FAB, 3-NBA matrix)

$\text{C}_{19}\text{H}_{20}\text{N}_3\text{O}_6\text{S}$ ($M+1$): calcd, 418.1073; found, 418.1049.

[0244] (12) 1-(β -D-Ribofuranosyl)pyrrole-2-carbaldehyde
(Compound 17)

To a solution of pyrrole-2-carbaldehyde (330 mg, 3.5 mmol) in CH_3CN (18 ml), NaH (60% oil dispersion, 152 mg, 3.8 mmol) was added and stirred at room temperature for 45 minutes, followed by addition of a solution of 2,3,5-tri-O-benzyl-D-ribofuranosyl chloride (3.1 mmol) (Non-patent Document 44) in CH_3CN (18 ml). The reaction mixture was stirred at room temperature for 4 hours. The product was partitioned between ethyl acetate and water, and the organic

layer was washed three times with saturated aqueous sodium chloride, dried over Na_2SO_4 , and then evaporated under reduced pressure to remove the solvent. The residue was purified by silica gel chromatography (eluted with 20% ethyl acetate in hexane) to give crude 1-(2,3,5-tri-O-benzyl-D-ribofuranosyl)pyrrole-2-carboxyaldehyde (506 mg). After the crude 1-(2,3,5-tri-O-benzyl-D-ribofuranosyl)pyrrole-2-carboxyaldehyde (506 mg, 1.0 mmol) was azeotroped with toluene, CH_2Cl_2 (17 ml) was added to the residue. To this solution, BBr_3 (1 M solution, 3.0 ml) was added at -78°C and stirred for 2.5 hours, followed by addition of a 50% methanol- CH_2Cl_2 solution (25 ml). After the solution was stirred at -78°C for 10 minutes, 28% NH_4OH (0.5 ml) was added. The reaction mixture was further stirred until it reached room temperature. The product was partitioned between CH_2Cl_2 and H_2O , and the aqueous layer was washed three times with CH_2Cl_2 and evaporated under reduced pressure to remove the solvent. The product was purified by reversed-phase C18 HPLC to give 1-(β -D-ribofuranosyl)pyrrole-2-carbaldehyde (108 mg, 15%, 2 steps).

[0245] ^1H NMR (270 MHz, $\text{DMSO}-d_6$) δ
9.54 (s, 1H), 7.74 (s, 1H),
7.06 (dd, 1H, $J=1.6$ and 4.0 Hz),
6.39 (d, 1H, $J=4.3$ Hz),
6.30 (dd, 1H, $J=3.0$ and 4.0 Hz),
5.27 (d, 1H, $J=5.6$ Hz),
5.05 (d, 1H, $J=4.9$ Hz),
5.00 (t, 1H, $J=5.3$ Hz), 4.02 (m, 2H),

3.85 (m, 1H), 3.52 (m, 2H).

[0246] ^{13}C NMR (75 MHz, DMSO- d_6) δ

179.41, 131.68, 128.24, 124.94,

110.23, 89.34, 84.33, 75.64,

69.50, 60.82.

[0247] HRMS (FAB, 3-NBA matrix)

$\text{C}_{10}\text{H}_{14}\text{NO}_5$ (M+1): calcd, 228.0872; found, 228.0863.

[0248] (13) 4-Propynyl-1-(β -D-ribofuranosyl)pyrrole-2-carbaldehyde (Compound 18)

4-Propynyl-1-(β -D-ribofuranosyl)pyrrole-2-carbaldehyde (Compound 18) was synthesized in the same manner as used for Compound 17, starting with 4-propynyl-2-pyrrolecarbaldehyde (Compound 16) (Non-patent Document 40) (266 mg, 2.0 mmol). After purification by RP-HPLC, Compound 18 was obtained (39 mg, 7%, 2 steps).

[0249] ^1H NMR (300 MHz, DMSO- d_6) δ

9.50 (s, 1H), 7.91 (s, 1H),

7.09 (d, 1H, $J=1.8$ Hz), 6.32 (d, 1H, $J=3.6$ Hz),

5.32 (d, 1H, $J=5.5$ Hz), 5.07 (t, 1H, $J=5.2$ Hz),

5.05 (d, 1H, $J=4.2$ Hz), 4.01 (m, 1H),

3.86 (m, 1H),

3.66 (ddd, 1H, $J=3.4$, 5.3 and 11.9 Hz),

3.55 (ddd, 1H, $J=3.6$, 4.9 and 12.1 Hz),

1.97 (s, 3H).

[0250] ^{13}C NMR (75 MHz, DMSO- d_6) δ

179.60, 131.15, 130.43, 126.13,

106.22, 89.62, 85.26, 84.49,

75.86, 73.17, 69.30, 60.53, 3.79.

[0251] HRMS (FAB, 3-NBA matrix)

$C_{13}H_{16}NO_5$ (M+1): calcd, 266.1028; found, 266.1023.

[0252] (14) Synthesis of nucleoside 5'-triphosphates of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine

A protected nucleoside (0.1 mmol, Compound 9 or 13) was azeotroped with pyridine to dryness. The residue was dissolved in a mixed solvent of pyridine (100 μ l) and dioxane (300 μ l). To this, a 1 M dioxane solution of 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one (110 μ l, 0.11 mmol) was added. After 10 minutes, triethylamine (10 μ l) and 0.5 M bis(tributylammonium) pyrophosphate in DMF (300 μ l, 0.15 mmol) were quickly added to the reaction mixture. This mixture was stirred at room temperature for 10 minutes, followed by addition of 1% iodine in pyridine/water (98/2, v/v) (2.0 ml). After 15 minutes, 150 μ l of 5% aqueous NaHSO₃ was added, and 5.0 ml water was further added thereto. After the solution was stirred at room temperature for 30 minutes, 20 ml of concentrated aqueous ammonia was added to cause ammonolysis at room temperature for 2 hours. The solvent was distilled off under reduced pressure, and the product was purified by DEAE Sephadex (A-25) column chromatography (eluted with a linear gradient of 50 mM to 1 M TEAB) and then purified on a C18-HPLC column (eluted with a linear gradient of 0% to 30% CH₃CN in 100 mM triethylammonium acetate) to give the desired nucleoside 5'-triphosphate.

[0253] 7-(2-Thienyl)-3-(2-deoxy- β -ribofuranosyl)-3H-imidazo[4,5-b]pyridine 5'-triphosphate (Compound 10)

¹H NMR (270 MHz, D₂O) δ

8.51 (s, 1H), 8.09 (d, 1H, J=5.3 Hz),
7.78 (d, 1H, J=3.6 Hz), 7.56 (d, 1H, J=4.9 Hz),
7.36 (d, 1H, J=5.3 Hz), 7.12 (t, 1H, J=4.9 Hz),
6.41 (t, 1H, J=7.3 Hz), 4.16 (m, 1H),
4.04 (m, 2H), 3.01 (q, 18H, J=7.3 Hz),
2.72 (m, 1H), 2.46 (m, 1H),
1.09 (t, 27H, J=7.3 Hz).

[0254] ^{31}P NMR (109 MHz, D_2O) δ

-9.94 (d, 1P, J=20.1 Hz),
-10.72 (d, 1P, J=20.1 Hz),
-22.58 (t, 1P, J=20.1 Hz).

[0255] Electrospray ionization mass spectroscopy (ESI-MS)

$\text{C}_{15}\text{H}_{18}\text{N}_3\text{O}_{12}\text{P}_3\text{S}$; calcd, 555.97 (M-H)-; found, 555.69 (M-H)-.

[0256] 7-(2-Thienyl)-3-(β -D-ribofuranosyl)-3H-imidazo[4,5-b]pyridine 5'-triphosphate (Compound 14)

^1H NMR (300 MHz, D_2O) δ

8.74 (s, 1H), 8.32 (d, 1H, J=5.4 Hz),
8.01 (d, 1H, J=3.5 Hz),
7.68 (dd, 1H, J=1.1 and 5.1 Hz),
7.64 (d, 1H, J=5.1 Hz),
7.25 (dd, 1H, J=3.5 and 5.1 Hz),
6.25 (d, 1H, J=6.0 Hz),
4.82 (m, 1H), 4.57 (m, 1H), 4.36 (m, 1H),
4.20 (m, 2H), 3.11 (q, 18H, J=7.3 Hz),
1.19 (t, 27H, J=7.3 Hz).

[0257] ^{31}P NMR (109 MHz, D_2O) δ

-9.80 (d, 1P, J=20.1 Hz),
-11.03 (d, 1P, J=18.9 Hz),

-22.78 (t, 1P, J=20.1 and 18.9 Hz).

[0258] Electrospray ionization mass spectroscopy (ESI-MS)

$C_{15}H_{18}N_3O_{13}P_3S$; calcd, 571.97 (M-H)-; found, 571.74 (M-H)-.

[0259] (15) Synthesis of nucleoside 5'- γ -amidotriphosphate of adenine or 7-(2-thienyl)-imidazo[4,5-b]pyridine (Non-patent Documents 43, 47 and 48)

A protected nucleoside (0.1 mmol, Compound 21 (Non-patent Document 46) or Compound 9) was azeotroped with pyridine to dryness. The residue was dissolved in a mixture of pyridine (100 μ l) and dioxane (300 μ l). To this, a 1 M dioxane solution of 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one (110 μ l, 0.11 mmol) was added. After 10 minutes, triethylamine (10 μ l) and 0.5 M bis(tributylammonium) pyrophosphate in DMF (300 μ l, 0.15 mmol) were quickly added to the reaction mixture and stirred at room temperature for 10 minutes, followed by addition of 1% iodine in pyridine/water (98/2, v/v) (2.0 ml). After 15 minutes, 150 μ l of 5% aqueous $NaHSO_3$ was added. After the solvent was distilled off under reduced pressure, 20 ml of 28% aqueous ammonia was added to the residue to cause ammonolysis at 60°C for 5 hours (for deoxyadenosine 5'- γ -amidotriphosphate) or at room temperature for 2 hours (for 5'- γ -amidotriphosphate of Compound 6). After the solvent was distilled off under reduced pressure, the product was purified by DEAE Sephadex (A-25) column chromatography (eluted with a linear gradient of 50 mM to 1 M TEAB) and then purified on a C18-HPLC column (eluted with a linear gradient of 0% to 30% CH_3CN in 100 mM triethylammonium acetate) to give the desired nucleoside 5'- γ -amidotriphosphate.

[0260] 6-Amino-9-(2-deoxy- β -D-ribofuranosyl)purine 5'- γ -amidotriphosphate (Compound 22)

^1H NMR (270 MHz, D_2O) δ

8.35 (s, 1H), 8.11 (s, 1H),
6.37 (t, 1H, $J=6.9$ Hz), 4.16 (m, 1H),
4.03 (m, 2H), 3.05 (q, 18H, $J=7.3$ Hz),
2.70 (m, 1H), 2.48 (m, 1H),
1.13 (t, 27H, $J=7.3$ Hz).

[0261] ^{31}P NMR (109 MHz, D_2O) δ

-0.50 (d, 1P, $J=19.5$ Hz),
-10.77 (d, 1P, $J=19.5$ Hz),
-22.14 (t, 1P, 20.1 Hz).

[0262] Electrospray ionization mass spectroscopy (ESI-MS)

$\text{C}_{10}\text{H}_{17}\text{N}_6\text{O}_{11}\text{P}_3$; calcd, 489.01 (M-H)-; found, 488.98 (M-H)-.

[0263] 7-(2-Thienyl)-3-(2-deoxy- β -D-ribofuranosyl)-3H-imidazo[4,5-b]pyridine 5'- γ -amidotriphosphate (γ -amidotriphosphate of Compound 10)

^1H NMR (270 MHz, D_2O) δ

8.57 (s, 1H), 8.16 (d, 1H, $J=5.3$ Hz),
7.85 (d, 1H, $J=3.6$ Hz), 7.58 (d, 1H, $J=4.9$ Hz),
7.45 (d, 1H, $J=5.3$ Hz), 7.15 (t, 1H, $J=4.6$ Hz),
6.48 (t, 1H, $J=6.9$ Hz), 4.18 (m, 1H),
4.05 (m, 2H), 3.03 (q, 18H, $J=7.3$ Hz),
2.75 (m, 1H), 2.50 (m, 1H),
1.11 (t, 27H, $J=7.3$ Hz).

[0264] ^{31}P NMR (109 MHz, D_2O) δ

-0.52 (d, 1P, $J=20.1$ Hz),
-10.75 (d, 1P, $J=19.5$ Hz),

-22.14 (t, 1P, J=20.8 Hz).

[0265] Electrospray ionization mass spectroscopy (ESI-MS)

$C_{15}H_{19}N_4O_{11}P_3S$; calcd, 554.99 (M-H)-; found, 555.01 (M-H)-.

[0266] (16) Nucleoside 5'-triphosphates of pyrrole-2-carbaldehyde and 4-propynylpyrrole-2-carbaldehyde (Non-patent Document 45)

To a solution containing 1-(β -D-ribofuranosyl)pyrrole-2-carbaldehyde (0.1 mmol) (Compound 17) or 4-propynyl-1-(β -D-ribofuranosyl)pyrrole-2-carbaldehyde (0.1 mmol) (Compound 18) and a proton sponge (33 mg, 0.15 mmol) in trimethyl phosphate (500 μ l), $POCl_3$ (12 μ l, 0.13 mmol) was added at 0°C and stirred at 0°C for 2 hours. To the reaction mixture, tri-n-butylamine (120 μ l, 0.5 mmol) was added and a 0.5 M DMF solution of bis(tributylammonium) pyrophosphate (1.0 ml, 0.5 mmol) was further added thereto. After 5 minutes, 0.5 M aqueous triethylammonium bicarbonate (TEAB, 500 μ l) was added to stop the reaction. The resulting crude product was purified by DEAE Sephadex (A-25) column chromatography (1.5 cm $\times$ 30 cm, eluted with a linear gradient of 50 mM to 1 M TEAB) and then purified on a C18-HPLC column (Synchropak RPP, Eichrom Technologies, eluted with a gradient of 0% to 30% CH_3CN in 100 mM triethylammonium acetate).

[0267] 1-(β -D-Ribofuranosyl)pyrrole-2-carbaldehyde 5'-triphosphate (Compound 19)

1H NMR (270 MHz, D_2O) δ

9.28 (s, 1H), 7.64 (s, 1H),

7.08 (d, 1H, J=3.9 Hz),

6.45 (d, 1H, J=4.1 Hz), 6.32 (m, 1H),

4.32 (m, 2H), 4.10 (m, 3H),

3.03 (q, 18H, J=7.3 Hz),

1.11 (t, 27H, J=7.3 Hz).

[0268] ^{31}P NMR (109 MHz, D_2O) δ

-10.51 (d, 1P, J=19.5 Hz),

-11.3 (d, 1P, J=20.1 Hz),

-22.91 (t, 1H, J=20.1 Hz).

[0269] Electrospray ionization mass spectroscopy (ESI-MS)

$\text{C}_{10}\text{H}_{16}\text{NO}_{14}\text{P}_3$: calcd, 465.97 (M-H)-; found, 465.85 (M-H)-.

[0270] 4-Propynyl-1-(β -D-ribofuranosyl)pyrrole-2-carbaldehyde 5'-triphosphate (Compound 20)

^1H NMR (270 MHz, D_2O) δ

9.28 (s, 1H), 7.70 (s, 1H), 7.08 (s, 1H),

6.40 (d, 1H, J=4.0 Hz), 4.30 (m, 2H),

4.13 (m, 3H), 3.06 (q, 18H, J=7.3 Hz),

1.86 (s, 3H), 1.14 (t, 27H, J=7.3 Hz).

[0271] ^{31}P NMR (109 MHz, D_2O) δ

-10.10 (d, 1P, J=19.5 Hz),

-11.02 (d, 1P, J=19.5 Hz),

-22.82 (t, 1P, J=20.1 Hz).

[0272] Electrospray ionization mass spectroscopy (ESI-MS)

$\text{C}_{13}\text{H}_{18}\text{NO}_{14}\text{P}_3$: calcd, 503.99 (M-H)-; found, 503.94 (M-H)-.

[0273] Example II Chemical synthesis-2

1. Synthesis of nucleoside derivatives of 7-(2-thiazolyl)-3H-imidazo[4,5-b]pyridine (Compound 4) (Dv) (Figure 33)

The same procedure as shown in Figure 22 for synthesis of nucleoside derivatives of 7-(2-thienyl)-3H-imidazo[4,5-b]pyridine (Compound 4 in Example I) (Ds) was repeated, except

that 2-(tributylstanyl)thiazole was used in Reaction (a) for introducing a thiazolyl group.

[0274] However, with respect to amidite synthesis (g), 2-cyanoethyl-N,N-diisopropylamino chloro phosphoroamidite, diisopropylethylamine and THF were used in this example, although 2-cyanoethyl tetraisopropylphosphorodiamidite and 0.45 M tetrazole in acetonitrile were used in Example I-3(7) for synthesis of 7-(2-thienyl)-3-[2-deoxy-5-O-(4,4'-dimethoxytrityl)- β -D-ribofuranosyl]-3H-imidazo[4,5-b]pyridine 2-cyanoethyl-N,N-diisopropylphosphoroamidite (Compound 8). Although different reagents are used, the same amidite is produced.

[0275] 2. Synthesis of nucleoside derivatives of 2-nitropyrrole (Compound 1) (Pn) (Figure 34)

(1) Synthesis of 1-(2-deoxy- β -D-ribofuranosyl)-2-nitropyrrole (Compound 3)

2-Nitropyrrole (Compound 1) (Non-patent Document 49) (224 mg, 2.0 mmol) was dissolved in acetonitrile (20 ml), followed by addition of NaH (80 mg, 60% oil dispersion, 2.0 mmol). After stirring at room temperature for 30 minutes, 2-deoxy-3,5-di-O-p-toluoyl- α -D-erythro-pentofuranosyl chloride (855 mg, 2.2 mmol) was added and stirred at room temperature for 2 hours. The reaction mixture was washed with ethyl acetate and water, and the organic layer was washed with water and saturated aqueous sodium chloride. The organic layer was dried over anhydrous sodium sulfate, concentrated and then purified on a silica gel column to give Compound 2 (722 mg, 78%). Compound 2 (722 mg) was treated with methanolic ammonia

(50 ml) to deprotect the toluoyl groups at room temperature for 12 hours, and purified on a silica gel column, followed by final HPLC purification to give Compound 3 (291 mg, 82%).

[0276] Compound 3: ^1H NMR (270 MHz, DMSO- d_6) δ
7.76 (bs, 1H), 7.26 (dd, 1H, $J=1.6$ and 3.6 Hz),
6.59 (t, 1H, $J=5.9$ Hz), 6.30 (t, 1H, $J=3.6$ Hz),
5.27 (d, 1H, $J=4.3$ Hz), 5.03 (t, 1H, $J=5.3$ Hz)
4.23 (m, 1H), 3.85 (m, 1H), 3.59 (m, 2H),
2.42 (m, 1H), 2.19 (m, 1H).

[0277] Electrospray ionization mass spectroscopy (ESI-MS)
 $\text{C}_9\text{H}_{11}\text{O}_5\text{N}_2$; calcd, 227.07 (M-H) $^-$; found, 227.03 (M-H) $^-$,
 $\text{C}_9\text{H}_{13}\text{O}_5\text{N}_2$; calcd, 229.08 (M+H) $^+$; found, 229.06 (M+H) $^+$.

[0278] (2) Synthesis of 1-[2-deoxy-5-O-(4,4'-dimethoxytrityl)- β -D-ribofuranosyl]-2-nitropyrrole 2-cyanoethyl-N,N-diisopropylphosphoramidite (Compound 5)

Compound 3 (228 mg, 1.0 mmol) was azeotroped with pyridine, followed by addition of pyridine (10 ml) and 4,4'-dimethoxytrityl chloride (373 mg, 1.1 mmol). After stirring at room temperature for 1 hour, the reaction mixture was washed with ethyl acetate and 5% aqueous NaHCO_3 , and the organic layer was washed with saturated aqueous sodium chloride. After drying over anhydrous sodium sulfate, the organic layer was concentrated and purified on a silica gel column to give Compound 4 (493 mg, 93%).

[0279] Compound 4 (265 mg, 0.5 mmol) was azeotroped with pyridine, followed by addition of THF (2.5 ml) and diisopropylethylamine (131 μl , 0.75 mmol). To this solution, 2-cyanoethyl-N,N-diisopropylamino chloro phosphoramidite

(123 μ l, 0.55 mmol) was added and stirred at room temperature for 1 hour. After addition of methanol (50 μ l), the reaction mixture was diluted with ethyl acetate:water (20:1, v/v) and washed with 5% aqueous NaHCO_3 . The organic layer was washed with saturated aqueous sodium chloride, dried over anhydrous sodium sulfate, concentrated and then purified on a silica gel column to give Compound 5 (315 mg, 86%).

[0280] (3) Synthesis of 1-(2-deoxy- β -D-ribofuranosyl)-2-nitropyrrole 5'-triphosphate (Compound 7)

Compound 4 (159 mg, 0.3 mmol) was azeotroped with pyridine, followed by addition of pyridine (3 ml) and acetic anhydride (57 μ l, 0.6 mmol). The reaction mixture was stirred at room temperature for 12 hours. The reaction mixture was washed with ethyl acetate and 5% NaHCO_3 , and then with 5% aqueous NaHCO_3 . After drying over anhydrous sodium sulfate, the organic layer was concentrated, azeotroped with toluene, and then dissolved in methylene chloride (30 ml). To this reaction mixture, dichloroacetic acid (300 μ l) was added at 0°C and stirred for 15 minutes at 0°C. The reaction mixture was washed with 5% aqueous NaHCO_3 , and the organic layer was washed with 5% NaHCO_3 . After drying over anhydrous sodium sulfate, the organic layer was concentrated and purified on a silica gel column to give Compound 6 (73 mg, 91%).

[0281] Compound 6 (41 mg, 0.15 mmol) was azeotroped with pyridine, followed by addition of pyridine (150 μ l) and dioxane (450 μ l). To this solution, 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one (180 μ l, 1 M in dioxane) was added and stirred for 10 minutes at room temperature. Tri-n-

butylamine (150 μ l) and bis(tributylammonium) pyrophosphate (450 μ l, 0.5 M in DMF) were added to the reaction mixture, followed by stirring for 10 minutes. After I₂/pyridine (3.0 ml, 1% iodine in pyridine/H₂O, 98/2, v/v) was added and stirred for 15 minutes, 5% aqueous NaHSO₃ (225 μ l) was added and the reaction mixture was then concentrated. After H₂O (7.5 ml) was added and stirred at room temperature for 30 minutes, 28% aqueous ammonia (30 ml) was added and stirred at room temperature for 2 hours. The reaction mixture was concentrated, lyophilized and then purified on DEAE Sephadex A-25 (with a linear gradient of 50 mM to 1.0 mM TEAB), followed by final HPLC purification to give Compound 7 of interest.

[0282] Compound 7:

Electrospray ionization mass spectroscopy (ESI-MS)

C₉H₁₄O₁₄N₂P₃; calcd, 466.97 (M-H)⁻; found, 466.70 (M-H)⁻.

[0283] 3. Synthesis of nucleoside derivatives of 4-(thienyl)-1H-pyrrolo[2,3-b]pyridine (DDs) and 4-(thienyl)-1H-pyrrolo[2,3-b]pyridine (DDv) (Figure 35)

(1) Synthesis of 4-iodo-1H-pyrrolo[2,3-b]pyridine (Compound 3)

1H-Pyrrolo[2,3-b]pyridine (Compound 1) (5.3 g, 45 mmol) was dissolved in ethyl acetate (45 ml). To this solution, a solution of meta-chloroperbenzoic acid (14 g, 54 mmol) in ethyl acetate (30 ml) was added dropwise over 1 hour while stirring at 0°C. After dropwise addition, the reaction mixture was stirred at room temperature for 3 hours and then allowed to stand at 0°C. The crystal was filtered, washed with ethyl acetate and then dried under reduced pressure. This was

dissolved in water (30 ml), adjusted to pH 10 with 30% K_2CO_3 , and allowed to stand at room temperature for 1 hour and then at 0°C for 1 hour. The precipitate was then filtered and washed with ether to give N-oxide (3.5 g, 58%). N-oxide (3.0 g, 22 mmol) was dissolved in DMF (16 ml) and heated at 50°C. A solution of methanesulfonyl chloride (4.7 ml, 60 mmol) in DMF (6.4 ml) was added dropwise at 70°C, and this reaction mixture was stirred at 75°C for 2 hours. The reaction mixture was added to ice and then neutralized at 0°C with 10 N NaOH. After stirring at room temperature for 1 hour, the resulting precipitate was filtered, washed with water and then dried at 60°C under reduced pressure to give Compound 2 (2.7 g, 80%).

[0284] Compound 2 (2.7 g, 18 mmol) and NaI (13 g, 88 mmol) were dissolved in acetonitrile (28 ml). To this solution, CH_3COCl (3.5 ml, 50 mmol) was added while stirring at room temperature. The reaction mixture was heated at 85°C for 12 hours. After the reaction mixture was cooled to room temperature, 10% aqueous Na_2CO_3 (28 ml) and 10% aqueous $NaHSO_3$ (28 ml) were added sequentially and stirred at room temperature for 15 minutes. The reaction mixture was washed with ethyl acetate, and the organic layer was further washed with saturated aqueous sodium chloride. After drying over anhydrous sodium sulfate, the organic layer was concentrated and purified on a silica gel column to give 4-iodo-1-N-acetylpyrrolo[2,3-b]pyridine (2.0 g) and 4-iodo-1H-pyrrolo[2,3-b]pyridine (Compound 3) (2.3 g). 4-Iodo-1-N-acetylpyrrolo[2,3-b]pyridine (2.0 g, 7.0 mmol) was dissolved in ethanol (70 ml) and, after addition of 28% sodium methoxide in

methanol (1.4 ml, 7.0 mmol), was then heated under reflux for 1 hour. The reaction mixture was concentrated and then partitioned between ethyl acetate and saturated aqueous ammonium chloride, and the organic layer was washed with saturated aqueous ammonium chloride. After drying over anhydrous sodium sulfate, the organic layer was concentrated and combined with Compound 3 (2.3 g) which had been previously obtained, followed by recrystallization from ethanol to give Compound 3 (4.0 g, 92%).

[0285] (2) Synthesis of 1-[2-deoxy-3,5-di-O-(toluoyl)- β -D-ribofuranosyl]-4-iodo-pyrrolo[2,3-b]pyridine (Compound 4)

To a solution of Compound 3 (950 mg, 3.9 mmol) in acetonitrile (39 ml), NaH (156 mg, 60% oil dispersion, 3.9 mmol) was added. After stirring at room temperature for 1 hour, 2-deoxy-3,5-di-O-p-toluoyl- α -D-erythro-pentofuranosyl chloride (1.8 g, 1.2 equivalents) was added and stirred at room temperature for 1.5 hours. The reaction mixture was washed with ethyl acetate and saturated aqueous ammonium chloride, and the organic layer was partitioned with saturated aqueous ammonium chloride and saturated aqueous sodium chloride. The organic layer was dried over anhydrous sodium sulfate, concentrated and then purified on a silica gel column to give Compound 4 (1.8 g, 77%).

[0286] (3) Synthesis of 1-(2-deoxy- β -D-ribofuranosyl)-4-(2-thienyl)-pyrrolo[2,3-b]pyridine (Compound 7) and 1-(2-deoxy- β -D-ribofuranosyl)-4-(2-thiazolyl)-pyrrolo[2,3-b]pyridine (Compound 8)

To a solution of Compound 4 (715 mg, 1.2 mmol) and

dichlorobis(triphenylphosphine)palladium (42 mg, 0.06 mmol) in DMF (12 ml), 2-(tributylstanyl)thiophene (601 μ l, 1.8 mmol) was added and stirred at 100°C for 1 hour. The reaction mixture was partitioned between ethyl acetate and water, and the organic layer was washed with water and then with saturated aqueous sodium chloride. The organic layer was dried over anhydrous sodium sulfate, concentrated and then purified on a silica gel column to give Compound 5 (586 mg, 88%).

[0287] Compound 5 (580 mg) was treated with ammonia-saturated methanol (50 ml) to deprotect the toluoyl groups at room temperature for 12 hours, and purified on a silica gel column, followed by final HPLC purification to give Compound 7 (304 mg, 91%).

[0288] Compounds 6 and 8 were synthesized in the same manner, except for using 2-(tributylstanyl)thiazole. Their synthesis started with Compound 4 (600 mg, 1.0 mmol) to give Compounds 6 (449 mg, 81%) and 8 (245 mg, 97%).

[0289] Compound 7: ^1H NMR (300 MHz, DMSO- d_6) δ
8.27 (d, 1H, $J=5.1$ Hz), 7.87 (d, 1H, $J=3.8$ Hz),
7.83 (d, 1H, $J=3.6$ Hz), 7.79 (d, 1H, $J=5.1$ Hz),
7.41 (d, 1H, $J=5.0$ Hz),
7.28 (dd, 1H, $J=3.7$ and 5.0 Hz),
6.93 (d, 1H, $J=3.8$ Hz),
6.76 (dd, 1H, $J=6.0$ and 8.2 Hz),
5.28 (d, 1H, $J=4.1$ Hz),
5.02 (t, 1H, $J=5.6$ Hz), 4.39 (m, 1H),
3.85 (m, 1H), 3.56 (m, 2H), 2.57 (m, 1H),

2.26 (m, 1H).

[0290] Compound 8: ¹H NMR (300 MHz, DMSO-d₆) δ

8.38 (d, 1H, J=5.1 Hz), 8.13 (d, 1H, J=3.2 Hz),

8.01 (d, 1H, J=3.2 Hz), 7.96 (d, 1H, J=3.7 Hz),

7.72 (d, 1H, J=5.1 Hz), 7.15 (d, 1H, J=3.7 Hz),

6.79 (dd, 1H, J=6.1 and 8.0 Hz),

5.29 (d, 1H, J=4.0 Hz), 4.50 (t, 1H, J=5.5 Hz),

4.40 (m, 1H), 3.86 (m, 1H), 3.57 (m, 2H),

2.58 (m, 1H), 2.26 (m, 1H).

[0291] (4) Synthesis of 1-[2-deoxy-5-O-(4,4'-dimethoxytrityl)-β-D-ribofuranosyl]-4-(2-thienyl)-pyrrolo[2,3-b]pyridine 2-cyanoethyl-N,N-diisopropylphosphoroamidite (Compound 11) and 1-[2-deoxy-5-O-(4,4'-dimethoxytrityl)-β-D-ribofuranosyl]-4-(2-thiazolyl)-pyrrolo[2,3-b]pyridine 2-cyanoethyl-N,N-diisopropylphosphoroamidite (Compound 12)

Compound 7 (300 mg, 0.9 mmol) was azeotroped with pyridine, followed by addition of pyridine (9 ml) and 4,4'-dimethoxytrityl chloride (386 mg, 1.1 mmol). After stirring at room temperature for 1 hour, the reaction mixture was partitioned between ethyl acetate and 5% aqueous NaHCO₃, and the organic layer was washed with saturated aqueous sodium chloride. After drying over anhydrous sodium sulfate, the organic layer was concentrated and purified on a silica gel column to give Compound 9 (570 mg, 97%).

[0292] Compound 9 (290 mg, 0.47 mmol) was azeotroped with pyridine, followed by addition of THF (2.4 ml) and diisopropylethylamine (123 μl, 0.7 mmol). To this solution, 2-cyanoethyl-N,N-diisopropylamino chloro phosphoroamidite

(115 μ l, 0.52 mmol) was added and stirred at room temperature for 1 hour. After addition of methanol (50 μ l), the reaction mixture was diluted with ethyl acetate:water (20:1, v/v) and partitioned with 5% aqueous NaHCO_3 . The organic layer was washed with saturated aqueous sodium chloride, dried over anhydrous sodium sulfate, concentrated and then purified on a silica gel column to give Compound 11 (345 mg, 90%).

[0293] Starting with Compound 8 (220 mg, 0.7 mmol), the same procedure was repeated to synthesize Compounds 10 (424 mg, 99%) and 12 (227 mg, 86%).

[0294] (5) Synthesis of 1-(2-deoxy- β -D-ribofuranosyl)-4-(2-thienyl)-pyrrolo[2,3-b]pyridine 5'-triphosphate (Compound 15) and 1-(2-deoxy- β -D-ribofuranosyl)-4-(2-thiazolyl)-pyrrolo[2,3-b]pyridine 5'-triphosphate (Compound 16)

Compound 9 (247 mg, 0.4 mmol) was azeotroped with pyridine, followed by addition of pyridine (4 ml) and acetic anhydride (75 μ l, 0.8 mmol). After stirring at room temperature for 12 hours, the reaction mixture was partitioned between ethyl acetate and 5% aqueous NaHCO_3 , and the organic layer was washed with 5% aqueous NaHCO_3 . This organic layer was dried over anhydrous sodium sulfate, concentrated and azeotroped with toluene. The residue was then dissolved in methylene chloride (40 ml). To this reaction mixture, dichloroacetic acid (400 μ l) was added at 0°C and stirred for 15 minutes at 0°C. The reaction mixture was partitioned with 5% aqueous NaHCO_3 , and the organic layer was washed with 5% aqueous NaHCO_3 . After drying over anhydrous sodium sulfate, the organic layer was concentrated and purified on a silica

gel column to give Compound 13 (125 mg, 87%).

[0295] Compound 13 (36 mg, 0.1 mmol) was azeotroped with pyridine, followed by addition of pyridine (100 μ l) and dioxane (300 μ l). To this solution, 2-chloro-4H-1,3,2-benzodioxaphosphin-4-one (110 μ l, 1 M in dioxane) was added and stirred for 10 minutes at room temperature. Tri-n-butylamine (100 μ l) and bis(tributylammonium) pyrophosphate (300 μ l, 0.5 M in DMF) were added to the reaction mixture, followed by stirring for 10 minutes. After I₂/pyridine (2.0 ml, 1% iodine in pyridine/H₂O, 98/2, v/v) was added and stirred for 15 minutes, 5% aqueous NaHSO₃ (150 μ l) was added and the reaction mixture was then concentrated. After H₂O (5 ml) was added and stirred at room temperature for 30 minutes, 28% aqueous ammonia (20 ml) was added and stirred at room temperature for 2 hours. The reaction mixture was concentrated, lyophilized and then purified on DEAE Sephadex A-25 (with a linear gradient of 50 mM to 1.0 mM TEAB), followed by final HPLC purification to give Compound 15.

[0296] Starting with Compound 10 (210 mg, 0.34 mmol), the same procedure was repeated to synthesize Compounds 14 (108 mg, 89%) and 16 (0.1 mmol synthesis scale).

[0297] Compound 15:

Electrospray ionization mass spectroscopy (ESI-MS)

C₁₆H₁₈O₁₂N₂P₃S; calcd, 554.98 (M-H)⁻; found, 554.73 (M-H)⁻.

[0298] Compound 16:

Electrospray ionization mass spectroscopy (ESI-MS)

C₁₅H₁₇O₁₂N₃P₃S; calcd, 555.97 (M-H)⁻; found, 555.82 (M-H)⁻.

[0299] Example III Biological experiments

1. Procedures

In this example, the following procedures were used unless otherwise specified.

[0300] Single nucleotide insertion experiment using KF exo⁻

The single nucleotide insertion experiment was performed as described in documents (Non-patent Documents 30-32). A primer (20-mer) labeled with 6-carboxylfluorescein at the 5'-end was annealed with template DNA (35-mer) by heating at 95°C and slow cooling to 4°C in 100 mM Tris-HCl (pH 7.5) buffer containing 20 mM MgCl₂, 2 mM DTT and 0.1 mg/ml bovine serum albumin. The primer-template duplex solution (10 μM, 5 μl) was mixed with 2 μl enzyme solution of the Klenow fragment lacking exonuclease activity (KF exo⁻, Amersham USB). After the mixture was incubated for 2 minutes, each dNTP solution (3 μl) was added to this solution to start the reaction at 37°C. The amount of the enzyme used (5-50 nM), the reaction time (1-35 minutes) and the gradient concentration of each dNTP (0.3-1500 μM) were adjusted to ensure conditions giving 25% or less products. The reaction was stopped with 10 μl stop solution (95% formamide and 20 mM EDTA), and the mixture was immediately heated at 75°C for 3 minutes. The diluted products were analyzed in an automated ABI377 DNA sequencer equipped with GeneScan software (version 3.0) (Non-patent Document 32). The relative velocity (v_0) was calculated by dividing the extent of reaction by the reaction time, and then normalized to the enzyme concentration (20 nM) for the various enzyme concentrations used. Kinetic parameters (K_M and V_{max}) were obtained from Hanes-Woolf plots of $[dNTP]/v_0$ against $[dNTP]$.

Each parameter was averaged from 3 to 8 data sets.

[0301] Primer extension reaction using KF

A 5'-³²P-labeled primer (23-mer) and template DNA (35-mer) were annealed into duplex form in 20 mM Tris-HCl (pH 7.5) buffer containing 14 mM MgCl₂ and 0.2 mM DTT. The duplex solution (400 nM, 5 µl) was mixed with each 5 x dNTP solution (2 µl) on ice, followed by addition of 3 µl enzyme solution containing KF exo⁻ (1 unit) or KF exo⁺ (1 unit) (TAKARA) to start the reaction. After the reaction solution was incubated at 37°C for 3 or 5 minutes, a dye solution (10 µl) containing 89 mM Tris-borate, 2 mM EDTA, 10 M urea and 0.05% BPB was added to stop the reaction. This solution was immediately heated at 75°C for 3 minutes and then electrophoresed on a 15% polyacrylamide-7 M urea gel. The products on the gel were analyzed with a bio-imaging analyzer (model BAS2500, Fuji).

[0302] DNA sequencing

Dideoxy-cycle sequencing (20 µl) was performed on a PTC-100 Program Thermal Controller (MJ Research) with 8 µl Ready Reaction Mix (BigDye1.1, Applied Biosystems) containing 0.3 pmol template and 4 pmol primer 1 or 2 in the presence or absence of 1 nmol dPa'TP. After 25 cycles of PCR (96°C for 10 seconds, 50°C for 5 seconds, and 60°C for 4 minutes), the remaining dye terminators were removed from the reaction solution using a Centri-SepTM spin column (Applied Biosystems), and this solution was then dried at 55°C under reduced pressure. The residue was resuspended in a formamide solution (4 µl) and analyzed with an ABI 377 DNA sequencer equipped with a 6% polyacrylamide-6 M urea gel. The sequence data were

analyzed with Applied Biosystems PRISM sequencing analysis v3.2 software.

[0303] PCR amplification

A PTC-100 Controller was used to perform the reaction in 20 mM Tris-HCl buffer (pH 8.8) containing 10 mM KCl, 10 mM $(\text{NH}_4)_2\text{SO}_4$, 2 mM MgSO_4 , 0.1% Triton X-100, 0.3 mM each dNTP (N = Pa, G, C and T) and dNTP_N (N = Ds and A), 1 μM each primer 1 or 2, 2.3 nM double-stranded DNA fragment, and 0.04 units/ μl Vent DNA polymerase (New England BioLabs). The PCR cycle was set as follows: 94°C for 0.5 minutes, 45°C for 0.5 minutes, and 65°C for 4 minutes. In control PCR, 0.2 mM each natural dNTP and 0.01 units/ μl Vent DNA polymerase were used, and the step of extension reaction was performed at 72°C for 1 minute. The PCR products were stained with ethidium bromide on a 4% agarose gel, and the stained bands were quantified for their intensity with a Molecular Imager FX Pro system and Quantity One software (Bio-Rad). For sequencing analysis, the PCR products had been purified by gel electrophoresis (7% polyacrylamide-7 M urea gel) or filtration (Microcon YM-30 and Micropure-EZ).

[0304] T7 transcription

Transcription (20 μl) was accomplished in 40 mM Tris-HCl buffer (pH 8.0) containing 24 mM MgCl_2 , 2 mM spermidine, 5 mM DTT, 0.01% Triton X-100, 1-3 mM each natural NTP, 0-3 mM Pa'TP, 0-3 mM DsTP, 10 mM GMP, 2 μM template DNA (for 17-mer transcript synthesis) or 0.5 μM template DNA (for tRNA transcript synthesis), and 2.5 units/ μl T7 RNA polymerase (TAKARA). To study the transcription efficiency, the

transcription was performed in the presence of 2 μCi [γ - ^{32}P]GTP (PerkinElmer, in place of GMP) for 17-mer transcript synthesis or [α - ^{32}P]GTP (Amersham) for tRNA transcript synthesis. After incubation at 37°C for 3 hours (17-mer synthesis) or 6 hours (tRNA synthesis), a urea-containing dye solution was added to stop the reaction. This solution was heated at 75°C for 3 minutes and then electrophoresed on a 15% or 20% (17-mer) or 10% (tRNA) polyacrylamide-7 M urea gel. The products were analyzed with a bio-imaging analyzer. For nucleotide composition analysis in transcripts (Non-patent Documents 17 and 33), the transcripts were internally labeled with 2 μCi [α - ^{32}P]UTP or [α - ^{32}P]ATP (Amersham). After the transcription, the products were digested with 0.75 units of RNase T₂ at 37°C for 2 hours in 10 μl of 15 mM sodium acetate buffer (pH 4.5). For 17-mer analysis, 0.05 A₂₆₀ units of *E. coli* tRNA (Sigma) was added to the digestion reaction solution. The digestion products were analyzed by 2D-TLC (HPTLC plate, 100 x 100 mm, Merck). The developing solvents used for the first and second dimensions were isobutyric acid-ammonia-water (66:1:33 v/v/v) and isopropyl alcohol-HCl-water (70:15:15 v/v/v or 75:15:10 v/v/v for Pa' transcripts), respectively. Spots of the labeled nucleotides on the TLC plate were analyzed with a bio-imaging analyzer.

[0305] Dideoxy-cycle sequencing on PCR-amplified products of DNA2 to DNA9

PCR for each DNA was accomplished by using a PTC-100 controller in Tris-HCl buffer (pH 8.8) containing 10 mM KCl, 10 mM (NH₄)₂SO₄, 2 mM MgSO₄, 0.1% Triton X-100, 0.3 mM each

dNTP (N = Pa, G, C and T) and dNTP_N (N = Ds and A), 1 μ M each primer 1 or 2, 2.3 M double-stranded DNA fragment (DNA2 to DNA9), and 0.04 units/ μ l Vent DNA polymerase (New England BioLabs). The PCR cycle was set as follows: 94°C for 0.5 minutes, 45°C for 0.5 minutes, and 65°C for 4 minutes. For sequencing, the PCR products had been purified by gel electrophoresis (7% polyacrylamide-7 M urea gel) or filtration (Microcon YM-30 and Micropure-EZ). After sequencing with or without dPa'TP, the sequences were determined using an ABI377 DNA sequencer equipped with a 6% polyacrylamide-6 M urea gel.

[0306] The positions of unnatural bases in DNA fragments can be confirmed by comparing both peak patterns obtained from sequencing with and without dPa'TP.

[0307] Aminoacylation of unnatural anticodon-containing tRNAs in *E. coli* cell-free translation system

Aminoacylation of tRNA transcripts was studied in a rapid translation system (RTS-100, Roche) according to the manufacturer's protocol with minor modifications. The tRNA transcripts (0.4 μ M) internally labeled with [α -³²P] were incubated at 30°C for 30 minutes in the system (25 μ l). The tRNAs were extracted with phenol saturated with sodium acetate (pH 4.5) and then analyzed by electrophoresis at 4°C on a 10% polyacrylamide gel in an acidic buffer (pH 4.75) containing 0.2 M Tris-acetate and 3 mM EDTA (Figure 5d, Figure 21).

[0308] 2. Sequencing of DNA fragments containing Ds

The position of Ds-Pa base pairing in DNA fragments was confirmed by dideoxynucleotide chain termination sequencing (Non-patent Document 33) supplemented with a nucleoside

triphosphate derivative of Pa' (dPa'TP) (Non-patent Document 34) (Pa' is 4-propynylpyrrole-2-carbaldehyde which is a modified form of Pa). The efficiency of KF exo^- -mediated incorporation of dPa'TP opposite Ds in the template ($V_{\text{max}}/K_M = 2.2 \times 10^5$) is 3.9-fold higher than that of dPaTP opposite Ds in the template ($V_{\text{max}}/K_M = 5.7 \times 10^4$) (Tables 1 and 2). Thus, sequencing of Ds-containing strands was performed using a Taq DNA polymerase sequencing kit BigDye1.1 (Applied Biosystems) supplemented with dPa'TP. For sequencing and the subsequent experiments, double-stranded DNA fragments containing a Ds-Pa base pair (150-mer and 174-mer, DNA1 to DNA14) were prepared by primer extension or ligation of chemically synthesized DNA fragments (Figure 6).

[0309] In sequencing of DNA1 containing 5'-CDsA/3'-GPAT, addition of 0.05 mM dPa'TP prevented the dye terminators of natural bases from being incorporated opposite Ds in the template. Thus, peaks of A, G, C and T all disappeared only at a site corresponding to Ds in the template (Figure 3c). Namely, the position at which no peak appeared and a gap was observed in this sequencing indicates the unnatural base position.

[0310] Unlike DNA1, some of the other DNAs containing different sequences around the unnatural base showed no clear gap pattern in their sequencing with dPa'TP, (Figures 13-20). However, since the subsequent peaks following the position of template Ds almost disappeared in sequencing without dPa'TP (Figure 3g), the unnatural base position in DNA can be confirmed by comparing both peak patterns obtained from

sequencing with and without dPa'TP (e.g., Figures 3c and 3g).

[0311] 3. PCR amplification of DNA fragments containing Ds-Pa base pair

PCR amplification of DNA1 to DNA14 containing a Ds-Pa base pair (Figure 6) (150-mer or 174-mer, 2.3 nM) was performed using thermophilic DNA polymerase having 3'→5' exonuclease activity (0.04 units/μl, VENT DNA polymerase, New England BioLabs) and a substrate mixture containing dDsTP_N, dPaTP, dATP_N, dGTP, dCTP and dTTP (0.3 mM each). The PCR cycle was set as follows: 94°C for 30 seconds, 45°C for 30 seconds, and 65°C for 4 minutes.

[0312] The inventors of the present invention first tested DNA1 for its amplification selectivity and efficiency (Figure 3a). After 10 cycles of PCR (Figure 3b), the products were analyzed by sequencing with or without dPa'TP (Figures 3d and 3h). Further, additional 10 cycles of PCR (10+10 cycles in total) were performed on a part of the 10 cycle PCR product (Figures 3e and 3i). In sequencing with dPa'TP, the PCR products after 10 cycles and 10+10 cycles resulted in peak patterns similar to those of the original DNA1. In sequencing without dPa'TP (Figures 3g-3i), the read-through peaks following the unnatural base position in DNA1 were slightly increased with an increase in the number of PCR cycles. Thus, the height of read-through peaks can be used to determine the rate of PCR-induced mutation from the Ds-Pa base pair to a natural base pair. The mutation rate was determined from the height of read-through peaks in the PCR products (Figures 3g-3i), in comparison with the height of read-through peaks

obtained by sequencing without dPa'TP for control DNA fragments containing 1-10% A-T base pair instead of Ds-Pa (Figure 8). The mutation rate of the Ds-Pa base pair in DNA1 after 10 cycles and 10+10 cycles was ~ 1% and 3-4%, respectively. By using this procedure, it was also indicated that the use of γ -amidotriphosphate of A increased the selectivity of Ds-Pa base pairing during PCR. In PCR amplification of DNA1 performed using dDsTP_N, dPaTP and natural dNTPs, but not using dATP_N, the mutation rate was increased. After 10 cycles and 10+10 cycles, the mutation rate of the Ds-Pa base pair was ~ 5% and ~ 10%, respectively (Figure 9). Moreover, in 10 cycle PCR in the absence of dDsTP_N and dPaTP, the Ds-Pa base pair in DNA1 was completely replaced with an A-T base pair (Figure 3f).

[0313] The amplification efficiency of DNA1 was evaluated from band intensity after dye staining of the PCR products on a gel or autoradiography of the products obtained with each labeled primer. The results (Figure 3b) indicated that DNA1 was amplified 15-fold after 10 cycles of PCR, while a corresponding natural DNA fragment (DNAcont1) was amplified 37-fold after 10 cycles of PCR. The more accurate efficiency was obtained by autoradiography of PCR products with a 5'- or 3'-³²P-labeled primer (Figure 10). The extension efficiency per cycle (Y) was determined using the equation: $N_f = N_0(1+Y)^n$ where N_f is the final copy number of a product, N_0 is the initial copy number, and n is the number of PCR cycles (Non-patent Document 35). For DNA1, the efficiency (Y) of PCR with 1 to 10 cycles was 0.38 for extension from the 5'-primer and

0.29 for extension from the 3'-primer, and the Y value in PCR amplification of DNAcont1 under usual conditions was 0.43 and 0.35, respectively. Thus, the efficiency per cycle of PCR in the unnatural base pair system is 76-88% of that in the conventional natural base pair systems. Based on the thus obtained amplification efficiency of PCR products and the mutation rate of the Ds-Pa base pair determined by DNA sequencing, the fidelity of the Ds-Pa base pair in DNA1 during PCR (for 2-fold amplification of DNA1) is calculated to be 99.8% or more. However, this fidelity was found to depend on the purity of the chemically synthesized original DNA fragments, rather than the selectivity of Ds-Pa base pairing during PCR amplification (Figure 11). Thus, the selectivity of the unnatural base pair system appears to be considerably higher than 99.8%.

[0314] The inventors of the present invention also studied amplification of the other DNA fragments (DNA2 to DNA14) containing different sequences around the Ds-Pa base pair. After 10 cycles of PCR, each DNA fragment was amplified 16- to 30-fold (Figure 12), and the mutation rate of the Ds-Pa base pair was 1-3% for each DNA (Figures 13-20), except for amplification of DNA10 to DNA14 containing 5'-X(A)_n-3' sequences (where X = Ds or Pa, and n = 2). The amplification efficiency of these DNA fragments after 10 cycles of PCR was less than 5-fold. This low efficiency is caused when substrates of Ds or Pa are consecutively incorporated subsequent to the incorporation of γ -amidotriphosphate of A. Thus, all the sequences containing unnatural bases, except for

5'-DsAA-3' and 5'-PaAA-3' sequences, can be used for DNA amplification.

[0315] 4. T7 transcription mediated by Ds-Pa base pairing

Ds-Pa and Ds-Pa' base pairings complementarily mediated site-specific incorporation of DsTP, PaTP and Pa'TP into RNA by T7 RNA polymerase. Transcription was studied with template DNA (35-mer) containing Ds, Pa or Pa' (Figures 4a and 4b). After 3 hour transcription with ribonucleoside triphosphates of the unnatural bases, the ³²P-labeled transcripts were analyzed on a gel (Figure 4c). The full-length transcripts (17-mer) containing Pa, Pa' and Ds had yields ranging from 28% to 91% (Figure 4c, Lanes 1, 2, 5 and 7), relative to transcripts consisting only of natural bases obtained with template DNA composed of natural bases (Figure 4c, Lane 9). In transcription of template DNAs containing the unnatural bases, the yields of transcripts obtained without adding substrates of the unnatural bases were significantly reduced (Figure 4c, Lanes 3, 6 and 8).

[0316] The high selectivity of Ds-Pa and Ds-Pa' base pairings in T7 transcription was confirmed by nucleotide composition analysis of internally ³²P-labeled transcripts (Non-patent Documents 17 and 33) (Figures 4d and 4e, as well as Table 3).

[0317] Table 3. Nucleotide composition analysis of T7 transcripts

[0318] [Table 3a]

Entry	Template	[α - 32 P] NTP	Natural NTP (mM)	Unnatural NTP (mM)	Composition of nucleotide incorporated as 5' neighbor of A or U ^a				
					Ap	Gp	Cp	Up	Np ^b
1	temp35Ds-3	ATP	1	Pa (1)	1.03 ^c [1] (0.01) ^e	1.97 [2] (0.04)	0.01 [0] (<0.01)	0.02 [0] (<0.01)	0.97 [1] (0.04)
2	temp35Ds-3	ATP	1	Pa (3)	1.01 [1] (0.02)	1.96 [2] (0.03)	0.01 [0] (<0.01)	0.01 [0] (0.01)	1.00 [1] (0.02)
3	temp35A-3	ATP	1	Pa (1)	0.99 [1] (0.01)	1.96 [2] (0.02)	n.d. ^f [0] (-)	1.04 [1] (0.02)	n.d. [0] (-)
4	temp35A-3	ATP	1	Pa (3)	0.99 [1] (0.01)	1.97 [2] (0.02)	n.d. [0] (-)	1.03 [1] (0.03)	n.d. [0] (-)
5	temp35G-3	ATP	1	Pa (1)	1.02 [1] (0.01)	1.96 [2] (0.02)	1.01 [1] (0.01)	0.01 [0] (<0.01)	n.d. [0] (-)
6	temp35G-3	ATP	1	Pa (3)	1.01 [1] (0.03)	1.98 [2] (0.04)	1.00 [1] (0.02)	0.01 [0] (<0.01)	n.d. [0] (-)
7	temp35Ds-3	ATP	1	Pa' (1)	1.03 [1] (0.02)	2.00 [2] (0.04)	0.01 [0] (<0.01)	0.02 [0] (0.01)	0.95 [1] (0.03)
8	temp35Ds-3	ATP	1	Pa' (3)	1.01 [1] (0.02)	1.99 [2] (0.03)	0.01 [0] (0.01)	0.02 [0] (0.01)	0.97 [1] (0.04)
9	temp35A-3	ATP	1	Pa' (1)	0.99 [1] (0.03)	1.98 [2] (0.03)	n.d. [0] (-)	1.03 [1] (0.03)	n.d. [0] (-)
10	temp35A-3	ATP	1	Pa' (3)	0.99 [1] (0.01)	1.99 [2] (0.01)	0.01 [0] (0.01)	1.00 [1] (0.01)	n.d. [0] (-)
11	temp35G-3	ATP	1	Pa' (1)	1.01 [1] (0.01)	1.98 [2] (0.02)	1.00 [1] (0.01)	0.01 [0] (<0.01)	n.d. [0] (-)
12	temp35G-3	ATP	1	Pa' (3)	1.00 [1] (<0.01)	1.98 [2] (0.01)	1.01 [1] (0.01)	0.01 [0] (<0.01)	n.d. [0] (-)
13	temp35Ds-3	ATP	1	none	1.31 [1] (0.02)	2.18 [2] (0.02)	0.15 [0] (<0.01)	0.36 [0] (0.03)	n.d. [-] (-)
14	temp35Pa-4	UTP	1	Ds (1)	0.02 [0] (<0.01)	n.d. [0] (-)	1.99 [2] (0.01)	1.00 [1] (0.01)	0.98 [1] (0.01)
15	temp35Pa-4	UTP	1	Ds (3)	0.01 [0] (<0.01)	n.d. [0] (-)	1.99 [2] (0.03)	1.01 [1] (0.01)	0.99 [1] (0.02)
16	temp35Pa'-4	UTP	1	Ds (1)	0.05 [0] (<0.01)	n.d. [0] (-)	1.98 [2] (0.01)	1.02 [1] (0.01)	0.94 [1] (0.01)
17	temp35Pa'-4	UTP	1	Ds (3)	0.02 [0] (<0.01)	n.d. [0] (-)	1.99 [2] (0.01)	1.02 [1] (0.01)	0.96 [1] (0.01)
18	temp35C-4	UTP	1	Ds (1)	0.01 [0] (<0.01)	0.98 [1] (<0.01)	2.01 [2] (0.01)	1.00 [1] (0.01)	n.d. [0] (-)

[0319] [Table 3b]

19	temp35C-4	UTP	1	Ds (3)	0.01 [0] (<0.01)	0.97 [1] (0.02)	2.03 [2] (0.04)	1.00 [1] (0.02)	n.d. [0] (-)
20	temp35T-4	UTP	1	Ds (1)	0.99 [1] (0.01)	n.d. [0] (-)	1.99 [2] (0.01)	1.02 [1] (0.02)	n.d. [0] (-)
21	temp35T-4	UTP	1	Ds (3)	0.99 [1] (0.01)	n.d. [0] (-)	2.00 [2] (0.01)	1.01 [1] (0.02)	n.d. [0] (-)
22	temp35Pa-4	UTP	1	none	0.91 [1] (0.01)	0.08 [0] (0.01)	1.99 [2] (<0.01)	1.02 [1] (0.01)	n.d. [-] (-)
23	temp35Pa'-4	UTP	1	none	0.97 [1] (0.01)	0.02 [0] (<0.01)	1.99 [2] (0.01)	1.02 [1] (0.01)	n.d. [-] (-)
24	DNA11	ATP	3	Pa' (3)	5.08 [5] (0.02)	5.98 [6] (0.04)	6.88 [7] (0.03)	0.09 [0] (<0.01)	0.97 [1] (0.02)
25	DNA12	ATP	3	Pa' (3)	6.05 [6] (0.06)	5.94 [6] (0.03)	6.96 [7] (0.05)	0.07 [0] (0.01)	0.98 [1] (0.03)
26	DNAcont4	ATP	3	none	6.05 [6] (0.07)	5.99 [6] (0.05)	6.89 [7] (0.02)	1.07 [1] (0.03)	n.d. [-] (-)
27	DNAcont4	ATP	3	Pa' (3)	6.04 [6] (0.10)	6.00 [6] (0.10)	6.90 [7] (0.05)	1.06 [1] (0.03)	n.d. [0] (-)
28	DNA13	ATP	2	Ds (2)	5.04 [5] (0.05)	6.02 [6] (0.08)	6.93 [7] (0.05)	0.05 [0] (0.01)	0.96 [1] (0.03)
29	DNA14	ATP	2	Ds (2)	6.13 [6] (0.04)	5.96 [6] (0.05)	6.93 [7] (0.06)	0.03 [0] (0.01)	0.96 [1] (0.02)
30	DNAcont4	ATP	2	none	6.10 [6] (0.04)	5.98 [6] (0.03)	6.88 [7] (0.03)	1.04 [1] (0.02)	n.d. [-] (-)
31	DNAcont4	ATP	2	Ds (2)	6.10 [6] (0.05)	5.98 [6] (0.04)	6.83 [7] (0.02)	1.09 [1] (0.01)	0.02 [0] (0.01)
32	DNA11	UTP	3	Pa' (3)	2.01 [2] (0.03)	3.98 [4] (0.04)	5.01 [5] (0.04)	4.00 [4] (0.05)	0.01 [0] (<0.01)
33	DNA12	UTP	3	Pa' (3)	2.00 [2] (0.02)	3.94 [4] (0.04)	4.05 [4] (0.02)	4.01 [4] (0.06)	0.01 [0] (<0.01)
34	DNAcont4	UTP	3	none	2.01 [2] (0.03)	3.97 [4] (0.04)	5.03 [5] (0.04)	4.00 [4] (0.06)	n.d. [-] (-)
35	DNAcont4	UTP	3	Pa' (3)	2.02 [2] (0.02)	4.00 [4] (0.09)	5.01 [5] (0.07)	3.97 [4] (0.05)	n.d. [0] (-)
36	DNA13	UTP	2	Ds (2)	2.00 [2] (0.03)	3.97 [4] (0.10)	4.99 [5] (0.07)	4.03 [4] (0.06)	n.d. [0] (-)
37	DNA14	UTP	2	Ds (2)	1.99 [2] (0.02)	3.91 [4] (0.03)	4.02 [4] (0.01)	4.08 [4] (0.04)	0.01 [0] (0.01)
38	DNAcont4	UTP	2	none	2.01 [2] (0.03)	3.98 [4] (0.04)	5.01 [5] (0.02)	4.01 [4] (0.06)	n.d. [-] (-)
39	DNAcont4	UTP	2	Ds (2)	1.99 [2] (0.03)	3.95 [4] (0.04)	5.01 [5] (0.06)	4.05 [4] (0.04)	0.01 [0] (<0.01)

[0320] a: Composition of nucleotides incorporated at the 5'-side of A (Entry Nos. 1-13 and 24-31) or U (Entry Nos. 14-23 and 32-39), as shown in Figures 4 and 5

b: Np = Pap, Pa'p or Dsp

c: The values were determined by using the following equation.

[0321] [Formula 27]

$$\frac{(\text{Radioactivity of each nucleotide})/[\text{Total radioactivity of all nucleotides (3'-monophosphates)}] \times (\text{Total number of nucleotides at 5' neighbor of } [\alpha\text{-}^{32}\text{P}]\text{NTP})}{\text{[0322]}}$$

d: The theoretical number of each nucleotide is shown in square brackets.

[0323] e: Standard deviations are shown in parentheses.

[0324] f: Not detected.

[0325] The internally ^{32}P -labeled transcripts were digested with Rnase T₂, and the resulting labeled nucleoside 3'-monophosphates were analyzed by 2D thin-layer chromatography (2D-TLC). In the case of transcripts obtained with templates containing unnatural bases, each spot corresponding to Ds or Pa appeared on 2D-TLC, and there was no spot corresponding to misincorporation of natural substrates having shapes similar to those of the unnatural bases (e.g., CTP and UTP to PaTP, as well as ATP and GTP to DsTP) (Figure 4d, N = Ds and Figure 4e, N = Pa). In the case of transcripts obtained with templates composed entirely of natural bases, there was no spot corresponding to Pa or Ds (Figure 4d, N = A and G, as well as Figure 4e, N = T and C). The quantified amount of each nucleotide spot on 2D-TLC was extremely close to its theoretical value expected from the product sequence (Table 3, Entry Nos. 1-23), resulting in an estimated selectivity of 95% or more for Ds-Pa or Ds-Pa' base pairing during transcription.

[0326] As an application example, the inventors of the

present invention prepared tRNA molecules (85-mer) containing unnatural anticodons CUPa', CPa'A, CUDs and CDsA through T7 transcription. For this purpose, the sequence of *E. coli* suppressor tRNA^{Tyr} was used.

[0327] The transcription efficiency of PaTP or Pa'TP into 17-mer transcripts was relatively low, but transcription of Pa'-containing longer tRNAs (85-mer) showed high efficiency; the 85-mers had yields ranging from 88% to 93%, relative to tRNA transcripts having a CUA anticodon (Figure 5a). For nucleotide composition analysis, the transcripts were internally labeled with [α -³²P]ATP or [α -³²P]UTP. Since the unnatural nucleotides are each located at the 5'-side of A, 3'-monophosphate of each unnatural nucleotide is labeled only with [α -³²P]ATP. Thus, spots corresponding to the unnatural nucleotides are detected by 2D-TLC only when the transcripts are labeled with [α -³²P]ATP (Figures 5b and 5c). The selectivity of both Pa' incorporation opposite template Ds and Ds incorporation opposite template Pa was 96% or more (Table 3, Entry Nos. 24-39).

[0328] Using the resulting tRNA transcripts, the inventors of the present invention studied aminoacylation in an *E. coli* extract (RTS 100 *E. coli* HY kit, Roche Diagnostics). tRNAs having CPa'A and CDsA anticodons were not aminoacylated, indicating that they avoided recognition by *E. coli*-derived aminoacyl tRNA synthetase (Figure 5d, Lanes 7-10). In contrast, tRNAs having CUPa' and CUDs anticodons were aminoacylated (Figure 5d, Lanes 1-4). The selectivity of aminoacylation of tRNAs having these unnatural anticodons is

in good agreement with the selectivity of recognition between *E. coli* tRNA^{Tyr} and tyrosyl tRNA synthetase (Non-patent Documents 36 and 37). These results confirm site-specific incorporation of unnatural bases into tRNA.

[0329] Example IV Synthesis of biotinylated PaTP

Biotinylated PaTP (i.e., 1-(β -D-ribofuranosyl)-4-[(3-biotinamido-1-propynyl)]pyrrole-2-carbaldehyde 5'-triphosphate; Compound 28, Bio-PaTP) was synthesized according to the scheme shown in Figure 36.

[0330] (1) Synthesis of 1-(β -D-ribofuranosyl)-4-[(3-dichloroacetamido)-1-propynyl]pyrrole-2-carbaldehyde (Compound 24) (Reactions (a)-(c) in the scheme shown in Figure 36):

To a solution containing 2,3,5-tri-O-benzyl-D-ribofuranose (1.0 g, 2.3 mmol) and CCl₄ (344 μ l, 3.6 mmol) in THF (4.6 ml), hexamethylphosphorous triamide (562 μ l, 3.0 mmol) was added at -78°C. This solution was stirred at -78°C for 2 hours and then at room temperature for 30 minutes (Solution A). To 4-iodo-pyrrole-2-carboxyaldehyde (Compound 23) (830 mg, 3.7 mmol) in CH₃CN (25 mL), NaH (60% oil dispersion, 150 mg, 3.7 mmol) was added. The reaction mixture was stirred at room temperature for 30 minutes, followed by addition of 2,3,5-tri-O-benzyl-D-ribofuranosyl chloride in THF (Solution A). The reaction mixture was stirred at room temperature for 12 hours. The product was partitioned between ethyl acetate and water. The organic layer was washed three times with saturated NH₄Cl, dried over Na₂SO₄ and then concentrated under reduced pressure. The product was purified by silica gel column chromatography (eluted with 1% methanol

in dichloromethane) to give 1-(2,3,5-tri-O-benzyl-D-ribofuranosyl)-4-iodopyrrole-2-carbaldehyde.

[0331] To 1-(2,3,5-tri-O-benzyl-D-ribofuranosyl)-4-iodopyrrole-2-carbaldehyde in dichloromethane (15 ml), BBr_3 (1 M, 8.5 ml) was added at -78°C . The reaction mixture was stirred for 2 hours, followed by addition of 50% methanol in CH_2Cl_2 (30 ml). After this solution was stirred at -78°C for 10 minutes, 28% NH_4OH (4 ml) was added and the reaction mixture was stirred until it reached room temperature. The solution was added to CH_2Cl_2 and H_2O . The aqueous layer was partitioned and washed three times with CH_2Cl_2 , and the residue was concentrated under reduced pressure. The product was purified by reversed-phase C18 HPLC to give 1-(β -D-ribofuranosyl)-4-iodopyrrole-2-carbaldehyde (330 mg).

[0332] 1-(β -D-Ribofuranosyl)-4-iodopyrrole-2-carbaldehyde (176 mg, 0.5 mmol, containing α -anomer) was azeotroped with pyridine and toluene. 1-(β -D-Ribofuranosyl)-4-iodopyrrole-2-carbaldehyde (176 mg), tetrakis(triphenylphosphine)palladium (29 mg, 0.025 mmol), CuI (15 mg, 0.08 mmol) and triethylamine (105 μl , 0.75 mmol) were dissolved in DMF (1.8 ml), and a 1 M solution of 3-(dichloroacetamido)-1-propyne (0.75 mmol) in DMF (750 μl) was added thereto. The reaction mixture was stirred at room temperature for 12 hours. The product was partitioned between EtOAc and H_2O , and the organic layer was dried over Na_2SO_4 and concentrated under reduced pressure. The product was purified by silica gel column chromatography (10% methanol in dichloromethane) and RP-HPLC to give Compound 24 as a β -isomer (123 mg, 26%, total yield for 3 steps). The structure

of Compound 24 was confirmed by NMR (Figure 38) and high-resolution mass spectrometry. The HMQC (Figure 38e) and HMBC (Figures 38f and 38g) spectra of Compound 24 indicated that an N-glycosidic linkage was formed between sugar and pyrrole base site at the C1' carbon. Moreover, the cross peaks in the NOESY spectrum of Compound 24 (Figure 38d) were similar to those of Compound 18 (Figure 25j), and Compound 24 showed a cross peak between H1' and H4' protons. Thus, the anomeric configuration of Compound 24 was identified to be β .

[0333] Compound 24:

^1H NMR (300 MHz, DMSO- d_6) δ

9.54 (d, 1H, $J=0.8$ Hz), 9.10 (t, 1H, $J=5.2$ Hz),

8.01 (s, 1H), 7.18 (d, 1H, $J=1.8$ Hz),

6.49 (s, 1H), 6.34 (d, 1H, $J=3.5$ Hz),

5.35 (d, 1H, $J=5.6$ Hz), 5.10 (m, 2H),

4.17 (d, 2H, $J=5.4$ Hz), 4.02 (m, 2H),

3.88 (m, 1H),

3.68 (ddd, 1H, $J=3.4, 5.3, 12.1$ Hz),

3.57 (ddd, 1H, $J=3.5, 5.0, 12.1$ Hz).

^{13}C NMR (75 MHz, DMSO- d_6) δ

180.28, 163.78, 131.88, 131.49,

126.85, 105.23, 90.23, 85.26,

85.04, 76.83, 76.41, 69.75,

67.01, 60.96, 30.20.

HRMS (FAB, 3-NBA matrix)

$\text{C}_{15}\text{H}_{17}\text{N}_2\text{O}_6\text{Cl}_2$ ($M+1$): calcd, 391.0464; found, 391.0462.

(2) Synthesis of 1-(2,3-di-O-acetyl- β -D-ribofuranosyl)-4-[(3-dichloroacetamido)-1-propynyl]pyrrole-2-carbaldehyde (Compound

26) (Reactions (d)-(e) in the scheme shown in Figure 36):

[0334] Compound 24 (118 mg, 0.3 mmol) was azeotroped three times with pyridine. To the residue in pyridine (3.0 ml), 4,4'-dimethoxytrityl chloride (113 mg, 0.33 mmol) was added. The mixture was stirred at room temperature for 1 hour and then added to EtOAc and 5% NaHCO₃. The organic layer was washed with saturated NaCl, dried over Na₂SO₄ and then concentrated under reduced pressure. The product was purified by silica gel chromatography (1% methanol in dichloromethane) to give 197 mg of Compound 25 in a yield of 95%.

[0335] Compound 25 (188 mg, 0.27 mmol) was azeotroped three times with pyridine. To the residue in pyridine (2.7 ml), acetic anhydride (103 μ l, 1.1 mmol) was added. The mixture was stirred at room temperature for 12 hours and then added to EtOAc and 5% NaHCO₃. The organic layer was washed with saturated NaCl, dried over Na₂SO₄ and then concentrated under reduced pressure. To the residue in dichloromethane (27 ml), dichloroacetic acid (270 μ l) was added at 0°C. The mixture was stirred at 0°C for 15 minutes, poured into 5% aqueous sodium bicarbonate, and then extracted with dichloromethane. After drying over Na₂SO₄, the solution was concentrated under reduced pressure. The product was purified by silica gel chromatography (1% methanol in dichloromethane) to give 118 mg of Compound 26 in a yield of 92%.

[0336] Compound 25:

¹H NMR (500 MHz, DMSO-d₆) δ

9.56 (s, 1H), 9.05 (t, 1H, J=5.0 Hz),

7.72 (s, 1H), 7.38-7.20 (m, 10H),

6.87 (d, 4H, J=7.1 Hz), 6.47 (s, 1H),
6.34 (d, 1H, J=3.3 Hz),
5.47 (d, 1H, J=5.3 Hz),
5.13 (d, 1H, J=5.7 Hz),
4.12-4.00 (m, 5H), 3.73 (s, 6H),
3.22 (m, 2H).

HRMS (FAB, 3-NBA matrix)

C₃₆H₃₅N₂O₈Cl₂ (M+1): calcd, 693.1770; found, 693.1721.

Compound 26:

¹H NMR (500 MHz, DMSO-d₆) δ

9.50 (s, 1H), 9.10 (bs, 1H), 8.06 (s, 1H),
7.24 (d, 1H, J=1.3 Hz), 6.64 (d, 1H, J=5.0 Hz),
6.48 (s, 1H), 5.43 (t, 1H, J=5.0 Hz),
5.35 (t, 1H, J=5.2 Hz), 5.32 (t, 1H, J=4.8 Hz),
4.17 (m, 3H), 3.73 (m, 1H), 3.62 (m, 1H),
2.07 (s, 3H), 2.01 (s, 3H).

HRMS (FAB, 3-NBA matrix)

C₁₉H₂₁N₂O₈Cl₂ (M+1): calcd, 475.0675; found, 475.0687.

(3) Synthesis of 1-(β-D-ribofuranosyl)-4-[(3-biotinamido-1-propynyl)]pyrrole-2-carbaldehyde 5'-triphosphate (Compound 28) (Reactions (f)-(g) in the scheme shown in Figure 36):

[0337] Protected nucleoside 26 (47 mg, 0.1 mmol) was dissolved in pyridine and concentrated under reduced pressure. The residue was dissolved in pyridine (100 μl) and dioxane (300 μl), followed by addition of a 1 M dioxane solution of 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one (110 μl, 0.11 mmol). After 10 minutes, tri-n-butylamine (100 μl) and 0.5 M bis(tributylammonium) pyrophosphate in DMF (300 μl, 0.15 mmol)

were quickly added to the reaction mixture. The mixture was stirred at room temperature for 10 minutes, followed by addition of a 1% iodine solution in pyridine/water (98/2, v/v) (2.0 ml). After 15 minutes, a 5% aqueous solution of NaHSO_3 (150 μl) was added. The volatile components were removed by evaporation, and water (5.0 ml) was then added to the residue. The mixture was stirred at room temperature for 30 minutes and then treated with concentrated ammonia (20 ml) at room temperature for 12 hours. The solution was concentrated under reduced pressure, and the product was purified by DEAE Sephadex (A-25) column chromatography (eluted with a linear gradient of 50 mM to 1 M TEAB) and then by C18-HPLC (eluted with a gradient of 0% to 30% CH_3CN in 100 mM triethylammonium acetate) to give Nucleoside 5'-triphosphate 27.

[0338] After lyophilization, Compound 27 in 0.1 M NaHCO_3 - Na_2CO_3 buffer (5 ml, pH 8.6) was reacted with biotin-N-hydroxysuccinimide (900 μl , 0.14 M in DMF) at room temperature for 3 hours. The mixture was treated with 28% NH_4OH (2 ml) for 1 hour. The product was purified by DEAE Sephadex (A-25) column chromatography (eluted with a linear gradient of 50 mM to 1 M TEAB) and then by C18-HPLC (eluted with a gradient of 0% to 30% CH_3CN in 100 mM triethylammonium acetate) to give Nucleoside 5'-triphosphate 28 in a yield of 14% from Compound 26. The structure of Compound 28 was confirmed by ^1H NMR (Figures 43a and 43b), ^{31}P NMR (Figure 43c) and mass spectrometry (Figure 42). Proton signals from the biotin moiety in Compound 28 were identical with those of biotinylated γTTP previously synthesized, and the ^{31}P NMR

spectrum in D₂O showed typical phosphorus signals corresponding to nucleotide 5'-triphosphate.

[0339] Compound 27:

Electrospray ionization-mass spectroscopy (ESI-MS)

C₁₃H₁₉O₁₄N₂P₃: calcd, 519.00 (M-H)⁻; found, 518.98 (M-H)⁻.

Compound 28:

¹H NMR (300 MHz, D₂O) δ

9.36 (d, 1H, J=0.9 Hz), 7.86 (s, 1H),

7.20 (d, 1H, J=1.7 Hz), 6.44 (d, 1H, J=4.1 Hz),

4.39-4.33 (m, 3H), 4.23-4.15 (m, 4H),

4.06 (d, 2H, J=3.7 Hz), 3.18 (m, 1H),

3.12 (q, 24H, J=7.3 Hz),

2.82 (dd, 1H, J=4.9 and 13.1 Hz),

2.60 (d, 1H, J=13.0 Hz),

2.23 (t, 2H, J=7.0 Hz), 1.60 (m, 4H),

1.32 (m, 2H), 1.20 (t, 36H, J=7.3 Hz).

³¹P NMR (107 MHz, D₂O) δ

-8.96 (d, 1P, J=16.5 Hz),

-10.67 (d, 1H, J=20.1 Hz),

-22.36 (t, 1P, J=20.1 Hz).

Electrospray ionization-mass spectroscopy (ESI-MS)

C₂₃H₃₃O₁₆N₄P₃S: calcd, 745.07 (M-H)⁻; found, 745.07 (M-H)⁻.

UV-visible spectrum (in 10 mM sodium phosphate buffer, pH 7.0),

λ_{max} = 258 nm (ε = 1.1 × 10⁴), 308 nm (ε = 9.5 × 10³).

Example V Site-specific biotinylation of RNA through T7 transcription mediated by Ds-Pa artificial base pairing

To study in more detail the selectivity and potential of

Ds-Pa artificial base pairing in both transcription and PCR amplification, site-specific biotinylation was performed on 152-mer RNA molecules.

[0340] Site-specific biotinylation of RNA (152-mer)

Transcription (20 μ l) was performed for 6 hours at 37°C using 3 μ Ci [γ -³²P]GTP, 2 mM each natural NTP, 0-4 mM Bio-PaTP (biotinylated Pa substrate: chemically synthesized as shown in Example IV), and 30 nM template (DNA6 and DNAcont2 (see Figure 6), obtained by both ligation and PCR amplification).

[0341] For control reactions using DNAcont5 and DNAcont6 obtained by ligation (see Figure 6), transcription was performed only with 2 mM ATP, GTP and CTP in the presence or absence of 2 mM Bio-UTP (biotin-16-uridine-5'-triphosphate, Roche Applied Science). The products were analyzed and purified on a 7-10% polyacrylamide-7 M urea gel.

[0342] The biotinylated RNA transcripts were detected by gel shift assay using streptavidin. A mixture (10 μ l) of 2 pmol ³²P-labeled transcript and 100 pmol streptavidin (Promega) was incubated at 20°C for 1 hour in 10 mM Tris-HCl buffer (pH 7.6) containing 50 mM NaCl and 10 mM EDTA. The biotinylated RNA-streptavidin complexes were analyzed by electrophoresis on a 7% polyacrylamide-7 M urea gel. To determine the position where Bio-Pa was introduced, each RNA (152-mer) was labeled with [γ -³²P]ATP (Perkin Elmer) after 5'-dephosphorylation with calf intestinal alkaline phosphatase (Takara). The labeled RNA was partially digested with RNase T1 at 55°C for 12 minutes in 13.3 mM sodium citrate buffer (pH 5.0) containing 4.7 M urea, 0.7 mM EDTA and 0.17 mg/ml *E. coli* tRNA, or with

alkali at 90°C for 15 minutes in 32 mM sodium carbonate buffer (pH 9.1) containing 0.6 mM EDTA. The alkali-digested RNA solution (9 µl) was mixed with 11 µl of 20 mM Tris-HCl buffer (pH 7.6) containing 150 mM NaCl, and the mixture (10 µl) was then incubated with streptavidin magnetic beads (0.4 mg) (New England BioLabs) for 5 minutes at room temperature to capture the biotinylated RNA. An aliquot (5 µl) of the supernatant was analyzed together with other digested samples on a 10% polyacrylamide-7 M urea gel.

[0343] Results

Biotin-linked Pa (Bio-Pa) was introduced into RNA through T7 transcription mediated by Ds-Pa base pairing when using a template obtained by either ligation or PCR amplification. To analyze incorporation of Bio-Pa into RNA molecules (152-mer), DNA6 and DNAcont2 templates (see Figure 6) were used to perform T7 transcription in the presence of natural substrate NTPs (2 mM) and Bio-PaTP (2 or 4 mM). The transcription efficiency through Ds-Pa base pairing was 47% to 85% (Figure 44c, Lanes 2, 6 and 10), relative to the case of using DNAcont2, a DNA fragment containing natural bases alone (Figure 44c, Lanes 3, 7 and 11). In transcription using DNA6 template obtained by either ligation or PCR amplification, the amount of the full-length product was significantly reduced in the absence of Bio-PaTP (Figure 44c, Lanes 1, 5 and 9). These results indicate that very little replacement of the Ds-Pa base pair with a natural base pair occurred during PCR amplification of DNA6.

[0344] The selectivity of Bio-Pa incorporation into RNA was

evaluated by gel shift assay on the biotinylated transcripts using streptavidin. Since binding between streptavidin and biotin is very strong and specific, the percentages of the gel-shifted transcripts would be almost identical with the rate of biotin incorporation through transcription (i.e., yields of the biotinylated transcripts). In transcription using 2 mM Bio-PaTP (equivalent to natural substrates) and DNA6 template obtained by ligation, 90% of the transcripts were biotinylated (Figure 44d, Lane 2), and the incorporation rate was improved to 96% when the Bio-PaTP concentration was increased (4 mM) (Figure 44d, Lane 4). With respect to Bio-PaTP misincorporation opposite natural bases, the yields of the biotinylated transcripts were 9% and 16%, as evaluated in transcription experiments using DNAcont2 as a template in the presence of 2 and 4 mM Bio-PaTP, respectively (Figure 44d, Lanes 6 and 7). These misincorporations were estimated to be only 0.06% (9% for total) and 0.12% (16% for total) per base position in the 152-mer transcripts. In transcription using the template obtained by PCR amplification (20 cycles), the incorporation rate of Bio-Pa was reduced by about 3-4% (Figure 44d, Lanes 9 and 11). This value was in agreement with the mutation rate at the Ds-Pa base pair site (3-4%) estimated by sequence analysis (Figure 31). Interestingly, Bio-Pa misincorporation opposite natural bases was not increased when transcription was performed in the presence of natural and artificial base substrates using DNAcont2 template obtained by PCR amplification (20 cycles) (Figure 44d, Lanes 13 and 14). Thus, misincorporation of Ds and Pa opposite natural bases

would be very low during PCR amplification.

[0345] Moreover, the selectivity of Ds-Pa base pairing during T7 transcription was compared to that of natural A-T(U) base pairing. To evaluate misincorporation of Bio-UTP opposite bases G, C and T in the template strand, control DNA containing only one A (DNAcont5) or no A (DNAcont6) in the coding region of the template strand was synthesized (see Figure 6). Transcription using DNAcont5 or DNAcont6 as a template was performed under the same conditions as used in the Ds-Pa base pair system, except for using 0 or 2 mM Bio-UTP and other natural substrates (2 mM ATP, GTP and CTP). In transcription of DNAcont5, the transcripts were completely biotinylated by incorporation of Bio-U opposite A (Figure 44d, Lane 16). However, in transcription of DNAcont6, 21% of the transcripts were biotinylated by misincorporation of Bio-U opposite bases G, C and T in the template strand (Figure 44d, Lane 18). Thus, the misincorporation rate of Bio-PaTP opposite natural bases (9% and 16% in Figure 44d, Lanes 6 and 7, respectively) was found to be lower than that of biotin-linked UTP (Bio-UTP) opposite natural bases (21%). It should be noted that this Bio-U misincorporation rate (21%) was higher than the misincorporation rate of Bio-Pa opposite natural bases (16%) even under conditions where Bio-PaTP was present in an amount of 2 molar equivalents relative to natural substrates. Thus, during T7 transcription, selective elimination of Bio-Pa misincorporation at the natural base sites in the template strand is higher than that of Bio-U misincorporation opposite bases G, T and C in the template

strand, although Bio-Pa incorporation opposite Ds is slightly less efficient than Bio-U incorporation opposite A.

[0346] To determine the incorporation site of Bio-Pa in the transcripts, the biotinylated transcripts were analyzed for their sequences. If Bio-Pa is precisely introduced into the transcripts opposite Ds in the template, the transcripts will be biotinylated at position 59. The 5'-terminally ^{32}P -labeled transcripts were partially digested with alkali or RNase T1, and the resulting products were analyzed by electrophoresis. In sequence ladders of the transcripts containing Bio-Pa, bands corresponding to fragments larger than 59-mer were shifted (Figure 44e, Lanes 2 and 8). The alkali-digested fragments were further treated with streptavidin to capture the biotinylated fragments, and the rest was electrophoresed, indicating that the fragments larger than 59-mer almost disappeared (Figure 44e, Lanes 2 and 9). These results prove site-specific incorporation of Bio-Pa at position 59 in the transcripts. Further, transcripts from the templates obtained by PCR amplification (20 cycles) (Figure 44e, Lanes 7-12) produced the same patterns as those from the templates obtained by ligation (Figure 44e, Lanes 1-6).

[0347] Thus, site-specific biotinylation of RNA based on this approach is useful for immobilizing RNA molecules without loss of RNA activity. Since the artificial base Pa even modified with a bulky residue such as biotin was efficiently incorporated into RNA, a series of functional 4-modified Pa bases (e.g., fluorophore-linked Pa) can also be incorporated into RNA.

[0348] Example VI Synthesis of 4-position modified nucleoside derivatives of 2-nitropyrrole

4-Position modified nucleoside derivatives of 2-nitropyrrole were synthesized according to the scheme shown in Figure 45.

[0349] (1) Synthesis of 1-(2-deoxy- β -D-ribofuranosyl)-4-iodo-2-nitropyrrole (Compound 1 in Figure 45)

1-(2-Deoxy- β -D-ribofuranosyl)-2-nitropyrrole (700 mg, 3.1 mmol) was dissolved in CH₃CN (12 ml), followed by addition of N-iodosuccinimide (1.38 g, 6.1 mmol). After stirring at room temperature for 12 hours, the reaction mixture was partitioned between ethyl acetate and water, and the organic layer was washed twice with water. After drying over anhydrous sodium sulfate, the organic layer was concentrated and purified on a silica gel column (CH₂Cl₂:MeOH = 50:1, v/v) and by HPLC (25%-40% CH₃CN, 10 minutes, 40%-50% CH₃CN, 5 minutes) to give Compound 1 (607 mg, 56%). Recovered unreacted starting materials: 219 mg (31%).

[0350] Compound 1

¹H NMR (500 MHz, DMSO-d₆): δ

7.90 (d, 1H, J=2.1 Hz), 7.40 (d, 1H, J=2.1 Hz),
6.54 (t, 1H, J=5.6 Hz), 5.27 (d, 1H, J=4.5 Hz),
5.10 (t, 1H, J=5.2 Hz), 4.23 (m, 1H),
3.83 (m, 1H), 3.65 (m, 1H), 3.56 (m, 1H),
2.40 (m, 1H), 2.23 (m, 1H).

HRMS (FAB, 3-NBA matrix)

C₉H₁₂IN₂O₅ (M+1): calcd, 354.9791; found, 354.9784.

(2) Synthesis of 1-(2-deoxy- β -D-ribofuranosyl)-4-propynyl-2-

nitropyrrole (Compound 2)

1-(2-Deoxy- β -D-ribofuranosyl)-4-iodo-2-nitropyrrole (Compound 1) (280 mg, 0.79 mmol) and $\text{Pd}(\text{PPh}_3)_2\text{Cl}_2$ (56 mg, 0.08 mmol) were dissolved in DMF (7.9 ml), followed by addition of tributyl(1-propynyl)tin (481 μl , 1.6 mmol). The reaction mixture was heated at 100°C for 1.5 hours, concentrated and then purified on a silica gel column (CH_2Cl_2 :MeOH, 50:1, v/v) and by HPLC (34%-35% CH_3CN , 13 minutes) to give Compound 2 (125 mg, 60%).

[0351] Compound 2:

^1H NMR (300 MHz, DMSO-d_6): δ

7.92 (d, 1H, $J=2.2$ Hz), 7.27 (d, 1H, $J=2.2$ Hz),

6.55 (t, 1H, $J=5.7$ Hz), 5.28 (d, 1H, $J=4.5$ Hz),

5.11 (t, 1H, $J=5.2$ Hz), 4.24 (m, 1H),

3.85 (m, 1H),

3.67 (ddd, 1H, $J=3.6, 5.3, 12.1$ Hz),

3.57 (m, 1H), 2.43 (m, 1H), 2.23 (m, 1H),

1.99 (s, 3H).

^{13}C NMR (75 MHz, DMSO-d_6): δ

136.21, 129.17, 117.08, 105.29,

88.74, 88.30, 86.75, 72.91, 69.21,

60.83, 42.58, 4.29.

HRMS (FAB, 3-NBA matrix)

$\text{C}_{12}\text{H}_{15}\text{N}_2\text{O}_5$ ($M+1$): calcd, 267.0981; found, 267.0991.

(3) Synthesis of 1-(2-deoxy- β -D-ribofuranosyl)-4-(dichloroacetamido-1-propynyl)-2-nitropyrrole (Compound 3)

1-(2-Deoxy- β -D-ribofuranosyl)-4-iodo-2-nitropyrrole (Compound 1) (354 mg, 1.0 mmol), $\text{Pd}(\text{PPh}_3)_4$ (58 mg, 0.05 mmol),

CuI (30 mg, 0.16 mmol) and triethylamine (209 μ l, 1.5 mmol) were dissolved in DMF (3.5 ml), followed by addition of a 1 M DMF solution of N-(2-propynyl)-dichloroacetamide (1.5 ml, 1.5 mmol). After stirring at room temperature for 12 hours, the reaction mixture was partitioned between ethyl acetate and water, and the organic layer was washed three times with water. After drying over anhydrous sodium sulfate, the organic layer was concentrated and purified on a silica gel column (CH_2Cl_2 :MeOH, 20:1, v/v) and by HPLC (34%-35% CH_3CN , 12 minutes) to give Compound 3 (317 mg, 81%).

[0352] Compound 3: ^1H NMR (300 MHz, $\text{DMSO}-d_6$): δ
9.01 (t, 1H, $J=5.3$ Hz), 7.98 (d, 1H, $J=2.1$ Hz),
7.33 (d, 1H, $J=2.1$ Hz), 6.54 (t, 1H, $J=5.8$ Hz),
6.47 (s, 1H), 5.27 (d, 1H, $J=4.5$ Hz),
5.10 (t, 1H, $J=5.2$ Hz), 4.23 (m, 1H),
4.17 (d, 2H, $J=5.4$ Hz), 3.84 (m, 1H),
3.66 (ddd, 1H, $J=3.5, 5.2, 12.1$ Hz),
3.56 (dt, 1H, $J=4.6, 12.2$ Hz), 2.43 (m, 1H),
2.23 (m, 1H).

Electrospray ionization mass spectroscopy (ESI-MS)

$\text{C}_{14}\text{H}_{15}\text{O}_6\text{N}_3\text{Cl}_2$; calcd, 390.03 (M-H) $^-$; found, 389.85 (M-H) $^-$.

(4) Synthesis of 1-[2-deoxy-5-O-(4,4'-dimethoxytrityl)- β -D-ribofuranosyl]-4-propynyl-2-nitropyrrole 2-cyanoethyl-N,N-diisopropylphosphoramidite (Compound 6)

Compound 2 (200 mg, 0.75 mmol) was azeotroped with pyridine, followed by addition of pyridine (7.5 ml) and 4,4'-dimethoxytrityl chloride (280 mg, 0.83 mmol). After stirring at room temperature for 1 hour, the reaction mixture was

partitioned between ethyl acetate and 5% NaHCO₃, and the organic layer was washed with saturated aqueous sodium chloride. After drying over anhydrous sodium sulfate, the organic layer was concentrated and purified on a silica gel column (CH₂Cl₂:MeOH, 200:1, v/v) to give Compound 4 (365 mg, 86%). Compound 4 (190 mg, 0.33 mmol) was azeotroped with pyridine, followed by addition of THF (1.7 ml) and diisopropylethylamine (87 µl, 1.5 equivalents). To this solution, 2-cyanoethyl-N,N-diisopropylamino chloro phosphoramidite (82 µl, 0.37 mmol) was added and stirred at room temperature for 1 hour. After addition of methanol (50 µl), the reaction mixture was diluted with ethyl acetate:water (20:1, v/v) and partitioned with 5% NaHCO₃. The organic layer was washed with saturated aqueous sodium chloride, dried over anhydrous sodium sulfate, concentrated and then purified on a silica gel column (CH₂Cl₂:hexane, 1:4, v/v, 2% triethylamine) to give Compound 6 (223 mg, 87%).

[0353] Compound 4:

¹H NMR (300 MHz, DMSO-d₆): δ

7.65 (d, 1H, J=2.2 Hz), 7.41-7.23 (m, 10H),
6.89 (d, 4H, J=8.8 Hz), 6.59 (t, 1H, J=5.3 Hz),
5.37 (d, 1H, J=5.0 Hz), 4.30 (m, 1H),
3.97 (m, 1H), 3.75 (s, 6H),
3.21 (d, 2H, J=4.1 Hz), 2.48-2.32 (m, 2H),
1.92 (s, 3H).

HRMS (FAB, 3-NBA matrix)

C₃₃H₃₃N₂O₇ (M+1): calcd, 569.2288; found, 569.2246.

Compound 6:

^1H NMR (300 MHz, CDCl_3): δ

7.62 and 7.55 (d and d, 1H, $J=2.2$ Hz),
7.48-7.44 (m, 2H), 7.39-7.22 (m, 8H),
6.87 (m, 4H), 6.68 (m, 1H), 4.56 (m, 1H),
4.25 (m, 1H), 3.88-3.35 (m, 6H),
3.82(s and s, 6H), 2.85-2.72 (m, 1H),
2.63 (t, 1H, $J=6.4$ Hz), 2.46 (t, 1H, $J=6.4$ Hz),
2.36-2.25 (m, 1H), 1.95 and 1.93(s and s, 3H),
1.20-1.08 (m, 12H).

HRMS (FAB, 3-NBA matrix)

$\text{C}_{42}\text{H}_{50}\text{N}_4\text{O}_8\text{P}$ ($M+1$): calcd, 769.3366; found, 769.3166.

(5) Synthesis of 1-(2-deoxy- β -D-ribofuranosyl)-4-propynyl-2-nitropyrrole 5'-triphosphate (Compound 9)

Compound 4 (160 mg, 0.28 mmol) was azeotroped with pyridine, followed by addition of pyridine (2.8 ml) and acetic anhydride (53 μl , 0.56 mmol). After stirring at room temperature for 12 hours, the reaction mixture was partitioned between ethyl acetate and 5% NaHCO_3 , and then washed with 5% NaHCO_3 . The organic layer was dried over anhydrous sodium sulfate, concentrated, azeotroped with toluene, and then dissolved in methylene chloride (28 ml). To this reaction mixture, dichloroacetic acid (280 μl) was added at 0°C and stirred for 15 minutes at 0°C . The reaction mixture was partitioned with 5% NaHCO_3 , and the organic layer was washed with 5% NaHCO_3 . After drying over anhydrous sodium sulfate, the organic layer was concentrated and purified on a silica gel column to give Compound 7 (78 mg, 90%, 2 steps). Compound 7 (31 mg, 0.1 mmol) was azeotroped with pyridine, followed by

addition of pyridine (100 μ l) and dioxane (300 μ l). To this solution, 2-chloro-4H-1,3,2-benzodioxaphosphorin-4-one (110 μ l, 1 M in dioxane) was added and stirred for 10 minutes at room temperature. Tri-n-butylamine (100 μ l) and bis(tributylammonium) pyrophosphate (300 μ l, 0.5 M in DMF) were added to the reaction mixture, followed by stirring for 10 minutes. After I₂/pyridine (2.0 ml, 1% iodo in pyridine/H₂O, 98/2, v/v) was added and stirred for 15 minutes, 5% NaHSO₃ (150 μ l) was added and the reaction mixture was then concentrated. After H₂O (5.0 ml) was added and stirred at room temperature for 30 minutes, 28% aqueous ammonia (20 ml) was added and stirred at room temperature for 2 hours. The reaction mixture was concentrated, lyophilized and then purified on DEAE Sephadex A-25 (with a linear gradient of 50 mM to 1.0 mM TEAB) to give Compound 9.

[0354] Compound 7:

¹H NMR (300 MHz, DMSO-d₆): δ

7.90 (d, 1H, J=2.1 Hz), 7.30 (d, 1H, J=2.1 Hz),

6.60 (t, 1H, J=6.4 Hz), 5.22 (m, 2H),

4.13 (m, 1H), 3.65 (m, 2H),

2.62 (ddd, 1H, J=3.0, 6.0, 14.3 Hz),

2.43 (m, 1H), 2.08 (s, 3H), 2.00 (s, 3H).

HRMS (FAB, 3-NBA matrix)

C₁₄H₁₇N₂O₆ (M+1): calcd, 309.1087; found, 309.1066.

Compound 9:

Electrospray ionization mass spectroscopy (ESI-MS)

C₁₂H₁₇O₁₄N₂P₃; calcd, 504.98 (M-H)⁻; found, 505.95 (M-H)⁻.

(6) Synthesis of 1-(2-deoxy-3-O-acetyl- β -D-ribofuranosyl)-4-

(dichloroacetamido-1-propynyl)-2-nitropyrrole (Compound 8)

Compound 3 (305 mg, 0.78 mmol) was azeotroped with pyridine, followed by addition of pyridine (7.8 ml) and 4,4'-dimethoxytrityl chloride (291 mg, 0.86 mmol). After stirring at room temperature for 1 hour, the reaction mixture was partitioned between ethyl acetate and 5% NaHCO₃, and the organic layer was washed with saturated aqueous sodium chloride. After drying over anhydrous sodium sulfate, the organic layer was concentrated and purified on a silica gel column (CH₂Cl₂:EtOAc, 9:1, v/v) to give Compound 5 (526 mg, 97%). Compound 5 (515 mg, 0.74 mmol) was azeotroped with pyridine, followed by addition of pyridine (7.4 ml) and acetic anhydride (280 µl, 3.0 mmol). After stirring at room temperature for 12 hours, the reaction mixture was partitioned between ethyl acetate and 5% NaHCO₃, and then washed with 5% NaHCO₃. The organic layer was dried over anhydrous sodium sulfate, concentrated, azeotroped with toluene, and then dissolved in methylene chloride (74 ml). To this reaction mixture, dichloroacetic acid (740 µl) was added at 0°C and stirred for 15 minutes at 0°C. The reaction mixture was partitioned with 5% NaHCO₃, and the organic layer was washed with 5% NaHCO₃. After drying over anhydrous sodium sulfate, the organic layer was concentrated and purified on a silica gel column (CH₂Cl₂:MeOH, 50:1, v/v) to give Compound 8 (289 mg, 90%, 2 steps).

[0355] Compound 5: ¹H NMR (300 MHz, DMSO-d₆): δ
9.07 (t, 1H, J=5.3 Hz), 7.67 (d, 1H, J=2.2 Hz),
7.41-7.21 (m, 10H),

6.89 (dd, 4H, J=1.7, 8.9 Hz),
6.59 (t, 1H, J=5.4 Hz), 6.48 (s, 1H),
5.38 (d, 1H, J=4.9 Hz), 4.29 (m, 1H),
4.12 (d, 2H, J=4.4 Hz), 3.99 (m, 1H),
3.74 (s, 6H), 3.17 (m, 2H),
2.46-2.33 (m, 2H).

HRMS (FAB, 3-NBA matrix)

$C_{35}H_{34}N_3O_8Cl_2$ (M+1): calcd, 694.1723; found, 694.1729.

Compound 8: 1H NMR (300 MHz, DMSO- d_6): δ

9.12 (t, 1H, J=5.3 Hz), 7.96 (d, 1H, J=2.2 Hz),
7.36 (d, 1H, J=2.2 Hz), 6.61 (t, 1H, J=6.3 Hz),
6.49 (s, 1H), 5.23 (m, 2H),
4.18 (d, 2H, J=5.4 Hz), 4.13 (m, 1H),
3.68 (m, 2H),
2.63 (ddd, 1H, J=3.0, 6.0, 14.3 Hz),
2.44 (m, 1H), 2.07 (s, 3H).

HRMS (FAB, 3-NBA matrix)

$C_{16}H_{18}N_3O_7Cl_2$ (M+1): calcd, 434.0522; found, 434.0549.

Example VII Synthesis of NH_2 -hx-dPnTP, ROX-hx-dPnTP and FAM-hx-dPnTP

In this example, NH_2 -hx-dPnTP, ROX-hx-dPnTP and FAM-hx-dPnTP were synthesized as dPnTP derivatives according to the synthesis scheme shown in Figure 46. These derivatives can be introduced into DNA, for example, through replication (PCR or primer extension). The derivative having an amino group can be used for DNA modification after incorporation, while the other derivatives having fluorescent groups can each be adapted for fluorescent labeling of DNA, FRET, etc.

[0356] (1) 1-(2-Deoxy- β -D-ribofuranosyl)-4-[3-(6-trifluoroacetamidohexanamido)-1-propynyl]-2-nitropyrrole (Step (a) in Figure 46)

1-(2-Deoxy- β -D-ribofuranosyl)-4-iodo-2-nitropyrrole (354 mg, 1 mmol, containing α -anomer) was azeotroped twice with anhydrous acetonitrile in a 50 mL flask, followed by addition of copper iodide (31 mg, 160 μ mol) and tetrakis(triphenylphosphine) palladium (0) (58 mg, 50 μ mol). After these materials were dissolved in anhydrous DMF (5 mL), triethylamine (210 μ L, 1.5 mmol) was added while stirring at room temperature and further stirred at room temperature under light-shielding conditions. To this mixture, a solution of N-(2-propynyl)-6-trifluoroacetamidohexanamide (396 mg, 1.5 mmol) in DMF (4 mL) was added dropwise and stirred overnight at room temperature. The reaction mixture was concentrated under reduced pressure, and the resulting crude product was purified by silica gel column chromatography (5-10% CH₃OH in CH₂Cl₂) and C18-HPLC (39-41% CH₃CN in H₂O, 15 minutes) to give the desired product as an amorphous substance (408 mg, yield 83%).

[0357] ¹H NMR (270 MHz, DMSO-d₆) δ

9.38 (brs, 1H), 8.28 (t, 1H, J=5.3 Hz),
7.95 (d, 1H, J=2.0 Hz), 7.30 (d, 1H, J=2.0 Hz),
6.54 (t, 1H, J=5.6 Hz), 5.29 (d, 1H, J=4.0 Hz),
5.20-5.05 (m, 1H), 4.30-4.20 (m, 1H),
4.05 (d, 2H, J=5.3 Hz), 3.83 (q, 1H, J=4.0 Hz),
3.70-3.50 (m, 2H), 3.14 (t, 2H, J=7.3 Hz),
2.50-2.36 (m, 1H), 2.30-2.17 (m, 1H),
2.08 (t, 2H, J=7.3 Hz), 1.58-1.40 (m, 4H),

1.28-1.16 (m, 2H).

HRMS (FAB, 3-NBA matrix)

$C_{20}H_{26}F_3N_4O_7$ (M + 1): calcd, 491.1754; found, 491.1761.

(2) 1-(2-Deoxy-3-O-acetyl- β -D-ribofuranosyl)-4-[3-(6-trifluoroacetamidohexanamido)-1-propynyl]-2-nitropyrrole
(Steps (b) and (c))

1-(2-Deoxy- β -D-ribofuranosyl)-4-[3-(6-trifluoroacetamidohexanamido)-1-propynyl]-2-nitropyrrole (394 mg, 803 μ mol) was azeotroped three times with anhydrous pyridine in a 50 mL flask and dissolved in anhydrous pyridine (4 mL). To this solution, dimethoxytrityl chloride (286 mg, 844 μ mol) was added and stirred at room temperature for 1.5 hours. The reaction mixture was added to ethyl acetate/water, and the aqueous layer was removed. The organic layer was washed with saturated aqueous sodium bicarbonate, dried over magnesium sulfate and then evaporated to remove the solvent. The resulting crude product was purified by silica gel column chromatography (0-0.5% CH_3OH in CH_2Cl_2) to give a tritylated product as an amorphous substance (543 mg). 1-(2-Deoxy-5-O-dimethoxytrityl- β -D-ribofuranosyl)-4-[3-(6-trifluoroacetamidohexanamido)-1-propynyl]-2-nitropyrrole (542 mg, 684 μ mol) was azeotroped three times with anhydrous pyridine in a 30 mL flask and dissolved in anhydrous pyridine (7 mL). To this solution, acetic anhydride (169 μ L, 1.79 mmol) was added and stirred overnight at room temperature. The reaction mixture was diluted with ethyl acetate, and the organic layer was washed with saturated aqueous sodium bicarbonate and saturated aqueous sodium chloride, dried over

magnesium sulfate, and then evaporated to remove the solvent. The resulting crude product was dissolved in anhydrous dichloromethane (68 mL). To this solution, dichloroacetic acid (680 μ L) was added while stirring at 0°C and further stirred for 15 minutes. The reaction mixture was added to saturated aqueous sodium bicarbonate, and the aqueous layer was extracted with dichloromethane. The combined organic layers were dried over magnesium sulfate and then evaporated to remove the solvent. The resulting oil was purified by silica gel column chromatography (2% CH₃OH in CH₂Cl₂) to give the desired product as an amorphous substance (328 mg, 77%, yield for 2 steps).

[0358] ¹H NMR (300 MHz, DMSO-d₆) δ

9.39 (brs, 1H), 8.30 (t, 1H, J=5.4 Hz),
7.94 (d, 1H, J=2.2 Hz), 7.33 (d, 1H, J=2.2 Hz),
6.61 (t, 1H, J=6.3 Hz), 5.30-5.18 (m, 2H),
4.20-4.10 (m, 1H), 4.06 (d, 2H, J=5.4 Hz),
3.75-3.55 (m, 2H), 3.16 (t, 2H, J=7.0 Hz),
2.62 (ddd, 1H, J=3.0, 6.0, 14.2 Hz),
2.50-2.35 (m, 1H), 2.15-2.05 (m, 2H),
2.07 (s, 3H), 1.60-1.40 (m, 4H),
1.30-1.20 (m, 2H).

HRMS (FAB, 3-NBA matrix)

C₂₂H₂₈F₃N₄O₈ (M + 1): calcd, 533.1859; found, 533.1907.

(3) 1-(2-Deoxy- β -D-ribofuranosyl)-4-[3-(6-aminohexanamido)-1-propynyl]-2-nitropyrrole 5'-triphosphate (NH₂-hx-dPnTP) (Step (d))

1-(2-Deoxy-3-O-acetyl- β -D-ribofuranosyl)-4-[3-(6-

trifluoroacetamidohexanamido)-1-propynyl]-2-nitropyrrole (53 mg, 100 μ mol) was azeotroped three times with anhydrous pyridine in a 10 mL flask, and the reaction vessel was then filled with argon gas. To this, anhydrous pyridine (100 μ L) and anhydrous dioxane (300 μ L) were added for dissolution purposes, followed by addition of a 1 M dioxane solution of 2-chloro-4H-1,2,3-dioxaphosphorin-4-one (110 μ L, 110 μ mol). After stirring at room temperature for 10 minutes, tri-n-butylamine (100 μ L) and a 0.5 M DMF solution of bis(tri-n-butylammonium) pyrophosphate (300 μ L) were added and stirred for 10 minutes. A 1% iodine/water/pyridine solution (2 mL) was added and stirred at room temperature for 15 minutes. After addition of 5% aqueous sodium bisulfite (150 μ L), the reaction mixture was concentrated under reduced pressure. The resulting oil was mixed with water (5 mL) and stirred at room temperature for 30 minutes, followed by addition of concentrated aqueous ammonia (20 mL). After stirring for 8 hours, this mixture was purified by DEAE Sephadex A-25 column chromatography (1.5 x 30 cm, linear concentration gradient; 50 mM to 1 M TEAB solution) and C18-HPLC (concentration gradient; x%-xx% acetonitrile in 0.1 M triethylammonium acetate buffer, pH 7.0) to give the desired product.

[0359] ^1H NMR (300 MHz, D_2O) δ

7.79 (s, 1H), 7.31 (s, 1H),
6.68 (t, 1H, $J=5.6$ Hz), 4.60-4.50 (m, 1H),
4.30-4.00 (m, 5H), 3.13 (q, 18H, $J=7.3$ Hz),
2.90 (t, 2H, $J=7.5$ Hz), 2.62-2.52 (m, 1H),
2.46-2.30 (m, 1H), 2.24 (t, 2H, $J=7.0$ Hz),

1.63-1.55 (m, 4H), 1.38-1.16 (m, 29H).

^{31}P NMR (121 MHz, D_2O) δ

-8.26, -10.51, -22.05.

MS (ESI)

$\text{C}_{18}\text{H}_{28}\text{N}_4\text{O}_{15}\text{P}_3$, $[\text{M}-\text{H}]^-$: calcd, 633.08; found, 633.00.

(4) 1-(2-Deoxy- β -D-ribofuranosyl)-4-[3-[6-(fluorescein-5-carboxamido)hexanamido]-1-propynyl]-2-nitropyrrole 5'-triphosphate (FAM-hx-dPnTP) (Step (e))

NH_2 -hx-dPnTP (6 μmol) was dissolved in 0.1 M aqueous sodium bicarbonate (pH 8.5, 0.72 mL), followed by addition of a solution of 5-carboxyfluorescein N-hydroxysuccinimidyl ester (FAM-SE) (3.7 mg, 7.8 μmol) in DMF (500 μL). The mixture was reacted at room temperature for 8 hours with occasional shaking under light-shielding conditions. To this mixture, concentrated aqueous ammonia (0.5 mL) was added and reacted for 5 minutes with occasional shaking. This mixture was lyophilized and then purified by DEAE Sephadex A-25 column chromatography (1.5 x 30 cm, linear concentration gradient; 50 mM to 1 M TEAB solution) and C18-HPLC (concentration gradient; 0%-50% acetonitrile in 0.1 M triethylammonium acetate buffer, pH 7.0) to give the desired product.

[0360] ^1H NMR (300 MHz, D_2O) δ

8.29 (s, 1H), 8.04 (d, 1H, $J=7.5$ Hz),

7.58 (brs, 1H), 7.26 (d, 1H, 7.5 Hz),

7.00-6.60 (m, 7H), 6.39 (brs, 1H),

4.50-4.35 (m, 1H), 4.40-3.90 (m, 5H),

3.50-3.30 (m, 2H), 3.12 (q, 18H, $J=7.3$ Hz),

2.41-2.10 (m, 4H), 1.75-1.50 (m, 4H),

1.45-1.10 (m, 29H).

^{31}P NMR (121 MHz, D_2O) δ

-10.86, -10.86, -23.13.

MS (ESI)

$\text{C}_{39}\text{H}_{38}\text{N}_4\text{O}_{21}\text{P}_3$ $[\text{M}-\text{H}]^-$: calcd, 991.12; found, 990.56

(5) 1-(2-Deoxy- β -D-ribofuranosyl)-4-[3-[6-(rhodamine-X-5-carboxamido)hexanamido]-1-propynyl]-2-nitropyrrole 5'-triphosphate (ROX-hx-dPnTP) (Step (e))

NH_2 -hx-dPnTP (6 μmol) was dissolved in 0.1 M aqueous sodium bicarbonate (pH 8.5, 0.85 mL), followed by addition of a solution of 5-carboxy-X-rhodamine N-hydroxysuccinimidyl ester (ROX-SE) (5 mg, 7.92 μmol) in DMF (1 mL). This mixture was reacted at room temperature for 18 hours with occasional shaking under light-shielding conditions. To this mixture, concentrated aqueous ammonia (0.5 mL) was added and reacted for 5 minutes with occasional shaking. This mixture was lyophilized and then purified by DEAE Sephadex A-25 column chromatography (1.5 x 30 cm, linear concentration gradient; 50 mM to 1 M TEAB solution) and C18-HPLC (concentration gradient; 12.5%-50% acetonitrile in 0.1 M triethylammonium acetate buffer, pH 7.0) to give the desired product.

[0361] ^1H NMR (300 MHz, D_2O) δ

8.28 (s, 1H), 8.03 (d, 1H, $J=8.9$ Hz),

7.47 (s, 1H), 7.19 (d, 1H, $J=7.7$ Hz),

6.68 (brs, 1H), 6.55 (brs, 1H),

6.43 (brs, 1H), 6.09 (brs, 1H),

4.40-4.25 (m, 1H), 4.15-3.70 (m, 5H),

3.60-3.27 (m, 10H), 3.12 (q, 18H, $J=7.3$ Hz),

2.95-2.70 (m, 4H), 2.65-2.40 (m, 4H),
2.40-2.15 (m, 3H), 2.05-1.70 (m, 9H),
1.60-1.50 (m, 4H), 1.50-1.15 (m, 29H).

^{31}P NMR (121 MHz, D_2O) δ
-10.90, -11.59, -23.27.

MS(ESI)

$\text{C}_{51}\text{H}_{56}\text{N}_6\text{O}_{19}\text{P}_3$ $[\text{M}-\text{H}]^-$: calcd, 1149.28; found, 1148.77

Example VIII Synthesis of NH_2 -hx-PaTP, FAM-hx-PaTP and TAMRA-hx-PaTP

In this example, NH_2 -hx-PaTP, FAM-hx-PaTP and TAMRA-hx-PaTP were synthesized as rPaTP derivatives according to the synthesis scheme shown in Figure 47. These derivatives can be introduced into RNA, for example, through transcription. The derivative having an amino group can be used for RNA modification after incorporation, while the other derivatives having fluorescent groups can each be adapted for fluorescent labeling of RNA, FRET, etc.

[0362] (1) 1-(β -D-Ribofuranosyl)-4-[(3-(6-trifluoroacetamidohexanamido)-1-propynyl)pyrrole-2-carbaldehyde (Step (a) in Figure 47)

1-(β -D-Ribofuranosyl)-4-iodopyrrole-2-carbaldehyde (177 mg, 500 μmol , containing α -anomer) was azeotroped twice with anhydrous acetonitrile in a 10 mL flask, followed by addition of copper iodide (15 mg, 80 μmol) and tetrakis(triphenylphosphine) palladium (0) (29 mg, 25 μmol). After these materials were dissolved in anhydrous DMF (2.5 mL), triethylamine (105 μL , 750 μmol) was added while stirring at room temperature and further stirred at room temperature

under light-shielding conditions. To this mixture, a solution of N-(2-propynyl)-6-trifluoroacetamidohexanamide (198 mg, 750 μ mol) in DMF (2 mL) was added dropwise and stirred overnight at room temperature. The reaction mixture was concentrated under reduced pressure, and the resulting crude product was purified by silica gel column chromatography (10% CH₃OH in CH₂Cl₂) and C18-HPLC (22-24% CH₃CN in H₂O, 15 minutes) to give the desired product as an amorphous substance (126 mg, yield 51%).

[0363] ¹H NMR (270 MHz, DMSO-d₆) δ

9.52 (d, 1H, J=0.66 Hz), 9.38 (brs, 1H),
8.27 (t, 1H, J=5.3 Hz), 7.96 (s, 1H),
7.13 (d, 1H, J=1.6 Hz), 6.32 (d, 1H, J=3.6 Hz),
5.42-5.32 (brs, 1H), 5.18-5.05 (m, 2H),
4.08-3.95 (m, 4H), 3.90-3.84 (m, 1H),
3.70-3.50 (m, 2H), 3.20-3.10 (m, 2H),
2.08 (t, 2H, J=7.3 Hz), 1.55-1.40 (m, 4H),
1.30-1.15 (m, 2H).

HRMS (FAB, 3-NBA matrix)

C₂₁H₂₇F₃N₃O₇ (M + 1): calcd, 490.1801; found, 490.1815.

(2) 1-(2,3-Di-O-acetyl- β -D-ribofuranosyl)-4-[(3-(6-trifluoroacetamidohexanamido)-1-propynyl)pyrrole-2-carbaldehyde (Steps (b) and (c))

1-(β -D-Ribofuranosyl)-4-[(3-(6-trifluoroacetamidohexanamido)-1-propynyl)pyrrole-2-carbaldehyde (122 mg, 250 μ mol) was azeotroped three times with anhydrous pyridine in a 10 mL flask and dissolved in anhydrous pyridine (4 mL). To this solution, dimethoxytrityl

chloride (89 mg, 263 μmol) was added and stirred at room temperature for 2.5 hours. The reaction mixture was added to ethyl acetate/water, and the aqueous layer was removed. The organic layer was washed with saturated aqueous sodium bicarbonate, dried over magnesium sulfate and then evaporated to remove the solvent. The resulting crude product was purified by silica gel column chromatography (0-0.5% CH_3OH in CH_2Cl_2) to give a tritylated product as an amorphous substance (150 mg). 1-(5-O-Dimethoxytrityl- β -D-ribofuranosyl)-4-[(3-(6-trifluoroacetamidohexanamido)-1-propynyl)pyrrole-2-carbaldehyde (149 mg, 188 μmol) was azeotroped three times with anhydrous pyridine in a 10 mL flask and dissolved in anhydrous pyridine (2 mL). To this solution, acetic anhydride (53 μL , 565 μmol) was added and stirred overnight at room temperature. The reaction mixture was diluted with ethyl acetate, and the organic layer was washed with saturated aqueous sodium bicarbonate and saturated aqueous sodium chloride, dried over magnesium sulfate, and then evaporated to remove the solvent. The resulting crude product was dissolved in anhydrous dichloromethane (19 mL). To this solution, dichloroacetic acid (280 μL) was added while stirring at 0°C and further stirred for 15 minutes. The reaction mixture was added to saturated aqueous sodium bicarbonate, and the aqueous layer was extracted with dichloromethane. The combined organic layers were dried over magnesium sulfate and then evaporated to remove the solvent. The resulting oil was purified by silica gel column chromatography (1-2.5% CH_3OH in CH_2Cl_2) to give the desired product as an amorphous substance

(90 mg, 63%, yield for 2 steps).

[0364] ^1H NMR (270 MHz, DMSO- d_6) δ

9.49 (s, 1H), 9.39 (brs, 1H),
8.28 (t, 1H, $J=5.3$ Hz), 8.03 (s, 1H),
7.21 (d, 1H, $J=1.6$ Hz), 6.63 (d, 1H, $J=5.3$ Hz),
5.44 (brs, 1H), 5.40-5.28 (m, 3H),
4.20-4.15 (m, 1H), 4.05 (d, 2H, $J=5.3$ Hz),
3.80-3.56 (m, 2H), 3.20-3.10 (m, 2H),
2.14-1.98 (m, 8H), 1.55-1.40 (m, 4H),
1.30-1.15 (m, 2H).

HRMS (FAB, 3-NBA matrix)

$\text{C}_{25}\text{H}_{31}\text{F}_3\text{N}_3\text{O}_9$ ($M + 1$): calcd, 574.2012; found, 574.2061.

(3) 1-(β -D-Ribofuranosyl)-4-[3-(6-aminohexanamido)-1-propynyl]pyrrole-2-carbaldehyde 5'-triphosphate (NH_2 -hx-PaTP)
(Step (d))

1-(2,3-Di-O-acetyl- β -D-ribofuranosyl)-4-[(3-(6-trifluoroacetamidohexanamido)-1-propynyl)pyrrole-2-carbaldehyde (57 mg, 100 μmol) was azeotroped three times with anhydrous pyridine in a 10 mL flask, and the reaction vessel was then filled with argon gas. To this, anhydrous pyridine (100 μL) and anhydrous dioxane (300 μL) were added for dissolution purposes, followed by addition of a 1 M dioxane solution of 2-chloro-4H-1,2,3-dioxaphosphorin-4-one (110 μL , 110 μmol). After stirring at room temperature for 10 minutes, tri-*n*-butylamine (100 μL) and a 0.5 M DMF solution of bis(tri-*n*-butylammonium) pyrophosphate (300 μL) were added and stirred for 10 minutes. A 1% iodine/water/pyridine solution (2 mL) was added and stirred at room temperature for 15 minutes.

After addition of 5% aqueous sodium bisulfite (150 μ L), the reaction mixture was concentrated under reduced pressure. The resulting oil was mixed with water (5 mL) and stirred at room temperature for 30 minutes, followed by addition of concentrated aqueous ammonia (20 mL). After stirring for 8 hours, this mixture was purified by DEAE Sephadex A-25 column chromatography (1.5 x 30 cm, linear concentration gradient; 50 mM to 1 M TEAB solution) and C18-HPLC (concentration gradient; 0%-50% acetonitrile in 0.1 M triethylammonium acetate buffer, pH 7.0) to give the desired product.

[0365] MS (ESI)

$C_{19}H_{29}N_3O_{15}P_3$ [M-H]⁻: calcd, 632.08; found, 632.00

(4) 1-(β -D-Ribofuranosyl)-4-[3-(6-(fluorescein-5-carboxamido)hexanamido)-1-propynyl]pyrrole-2-carbaldehyde 5'-triphosphate (FAM-hx-PaTP) (Step (e))

One-third of NH₂-hx-PaTP synthesized above was dissolved in 0.1 M aqueous sodium bicarbonate (pH 8.5, 2.5 mL), followed by addition of a solution of 5-carboxyfluorescein N-hydroxysuccinimidyl ester (9 mg, 19 μ mol) in DMF (200 μ L). The mixture was reacted at room temperature for 9 hours with occasional shaking under light-shielding conditions. To this mixture, concentrated aqueous ammonia (1 mL) was added and reacted for 1 hour with occasional shaking. This mixture was lyophilized and then purified by DEAE Sephadex A-25 column chromatography (1.5 x 30 cm, linear concentration gradient; 50 mM to 1 M TEAB solution) and C18-HPLC (concentration gradient; 0%-50% acetonitrile in 0.1 M triethylammonium acetate buffer, pH 7.0) to give the desired product.

[0366] ^1H NMR (300 MHz, D_2O) δ

9.07 (s, 1H), 8.20 (s, 1H),
7.93 (d, 1H, $J=7.9$ Hz), 7.62 (brs, 1H),
7.13 (d, 1H, $J=7.9$ Hz), 7.00-6.60 (m, 7H),
6.10 (d, 1H, $J=3.2$ Hz), 4.25-3.80 (m, 7H),
3.40-3.20 (m, 2H), 3.04 (q, 18H, $J=7.3$ Hz),
2.16 (t, 2H, $J=6.4$ Hz), 1.65-1.45 (m, 4H),
1.40-1.00 (m, 29H).

^{31}P NMR (121 MHz, D_2O) δ

-10.90, -11.38, -23.25.

MS (ESI)

$\text{C}_{40}\text{H}_{39}\text{N}_3\text{O}_{21}\text{P}_3$ $[\text{M}-\text{H}]^-$: calcd, 990.13; found, 989.78.

(5) 1-(β -D-Ribofuranosyl)-4-[3-(6-(tetramethylrhodamine-5-carboxamido)hexanamido)-1-propynyl]pyrrole-2-carbaldehyde
5'-triphosphate (TAMRA-hx-PaTP) (Step (e))

One-third of NH_2 -hx-PaTP synthesized above was dissolved in 0.1 M aqueous sodium bicarbonate (pH 8.5, 2.5 mL), followed by addition of a solution of 5-carboxytetramethylrhodamine N-hydroxysuccinimidyl ester (TAMRA-SE) (10 mg, 19 μmol) in DMF (1 mL). This mixture was reacted at room temperature for 9 hours with occasional shaking under light-shielding conditions. To this mixture, concentrated aqueous ammonia (1 mL) was added and reacted for 8 hours with occasional shaking. This mixture was lyophilized and then purified by DEAE Sephadex A-25 column chromatography (1.5 x 30 cm, linear concentration gradient; 50 mM to 1 M TEAB solution) and C18-HPLC (concentration gradient; 12.5%-50% acetonitrile in 0.1 M triethylammonium acetate buffer, pH 7.0) to give the desired

product.

[0367] ^1H NMR (300 MHz, D_2O) δ

9.07 (s, 1H), 8.26 (s, 1H),
8.02 (d, 1H, $J=7.9$ Hz), 7.69 (s, 1H),
7.35 (d, 1H, $J=7.9$ Hz), 7.10 (d, 1H, $J=9.4$ Hz),
7.01 (d, 1H, $J=9.4$ Hz), 6.88-6.70 (m, 3H),
6.39 (d, 2H, $J=9.2$ Hz), 6.06 (d, 1H, $J=3.5$ Hz),
4.20 (t, 1H, $J=4.9$ Hz), 4.10-3.80 (m, 6H),
3.55-3.30 (m, 2H), 3.25-3.00 (m, 30H),
2.25 (t, 2H, $J=6.3$ Hz), 1.75-1.55 (m, 4H),
1.40-1.10 (m, 29H).

^{31}P NMR (121 MHz, D_2O) δ -10.84, -11.56, -23.22.

MS (ESI)

$\text{C}_{44}\text{H}_{49}\text{N}_5\text{O}_{19}\text{P}_3$ $[\text{M}-\text{H}]^-$: calcd, 1044.22; found, 1044.09.

Example IX Incorporation of amino group- or fluorescent dye-linked substrate Pn into DNA (55-mer) through replication using Klenow fragment

In this example, substrates of Pn having an amino group or a fluorescent dye (FAM) attached through a linker (NH_2 -hx-dPnTP and FAM-hx-dPnTP, prepared in Example VII) were used to study the incorporation of these substrates into DNA through replication.

[0368] More specifically, experimental procedures as shown below were used. A solution (10 μl) containing a duplex between template DNA containing Ds at different positions (Template, 55-mer; 1Ds, 2Ds, 3Ds, 4Ds, 5Ds (SEQ ID NOs: 33-37) or a control (SEQ ID NO: 38)) and a primer fluorescently labeled with fluorescein (Primer, 20-mer) was mixed with a

deoxynucleoside triphosphate solution diluted with sterilized water (5 μ l) and Klenow fragment lacking 3'-5' exonuclease activity (2U, 5 μ l) to cause primer extension reaction (20 μ l scale). The reaction was performed using 200 nM Template-Primer, 100 μ M or 200 μ M dNTPs (N = A, G, C, T), 10 μ M NH₂-hx-dPnTP or FAM-hx-dPnTP, and 0.1 μ U/ μ l Klenow fragment (GE Health Care) in 50 mM Tris-HCl (pH 7.5), 10 mM MgCl₂, 1 mM DTT, 0.05 mg/ml BSA. After reaction at 37°C for 5 minutes, 10 M urea (20 μ l) was added to this solution to stop the reaction. The solution was heated at 75°C for 3 minutes, and the reaction products were then analyzed by electrophoresis on a 10% polyacrylamide-7 M urea gel. Bands of the reaction products were detected with a fluorescence image analyzer (Molecular Imager, Bio-Rad, FAM detection mode).

[0369] The results obtained are shown in Figures 48B and 48C. The amount of the full-length product was higher in the presence of NH₂-hx-dPnTP (Figures 48B and 48C) or FAM-hx-dPnTP (Figure 48C) than in the absence of these modified substrates of Pn, indicating that these modified dPnTP substrates were incorporated into DNA opposite Ds in the template DNA.

[0370] Example X Incorporation of fluorescent dye-linked substrate Pa into RNA (17-mer) through transcription using T7 RNA polymerase

In this example, substrates of Pa having a fluorescent dye (FAM or TAMRA) attached through a linker (FAM-hx-PaTP and TAMRA-hx-PaTP, prepared in Example VIII) were used to study the incorporation of these substrates into RNA through transcription.

[0371] More specifically, experimental procedures as shown below were used. In 10 mM Tris-HCl buffer (pH 7.6) containing 10 mM NaCl, template strand DNA (35-mer, 10 μ M) was annealed with its complementary strand DNA of the promoter region (21-mer, 10 μ M). Transcription reaction was performed using 2 μ Ci [γ -³²P]GTP, 1 mM NTPs, 1 mM FAM- or TAMRA-hx-PaTP, 2 μ M template DNA, and 50 units of T7 RNA polymerase (Takara) in 40 mM Tris-HCl buffer (pH 8.0), 24 mM MgCl₂, 2 mM spermidine, 5 mM DTT, 0.01% Triton X-100. After reaction at 37°C for 3 hours, 10 M urea (20 μ L) was added to this solution to stop the reaction. The solution was heated at 75°C for 3 minutes, and the transcripts were then analyzed by electrophoresis on a 20% polyacrylamide-7 M urea gel.

[0372] The results obtained are shown in Figure 49. The band mobility of the transcript (17-mer) on electrophoresis was slow only when template DNA containing Ds was used, indicating that these modified PaTP substrates were introduced into RNA opposite Ds in the template. The transcription efficiency was 14% for FAM-hx-PaTP and 10% for TAMRA-hx-PaTP, when compared to the control experiment with natural substrates alone (Figure 49, rightmost lane indicated as "None").

INDUSTRIAL APPLICABILITY

[0373] The inventors of the present invention have developed an unnatural base pair system ensuring highly practical selectivity during replication and transcription, which enables the creation of new biotechnologies with expanded genetic alphabet. Hydrophobic Ds-Pa base pairing

allows PCR amplification of DNA fragments containing the same. Moreover, Ds-Pa base pairing also allows site-specific incorporation of Ds and Pa into RNA through normal T7 transcription. Thus, this system provides a novel technique for creation of new functional artificial DNAs and RNAs. In addition, modified substrates and hydrophobic base pairs are scientifically important in further elucidation of mechanisms for replication and transcription.

[0374] To avoid undesired base pairings (e.g., Ds-Ds and natural-unnatural base pairings) during replication, the inventors of the present invention have achieved high selectivity of unnatural base pairing by combination of 5'- γ -amidotriphosphates with usual 5'-triphosphates. 5'- γ -Amidotriphosphates are useful as DNA polymerase substrates for recognizing proper complementarity between pairing bases. Recently, 5'- γ -P-aminonaphthalene-5-sulfonate triphosphate, which is another member of 5'- γ -modified triphosphates, has been reported to improve the fidelity of reverse transcription (Non-patent Document 38). These findings suggest that recognition of γ -modified triphosphates by DNA polymerases requires more accurate geometric fitting between pairing bases than that between usual triphosphate substrates. The combination of 5'- γ -amidotriphosphates and polymerases having 3'→5' exonuclease activity can also be applied to other unnatural base pairs.

[0375] The Ds-Pa base pair provides the first case where hydrophobic base pairs are functional during transcription. This suggests that shape complementarity between pairing bases

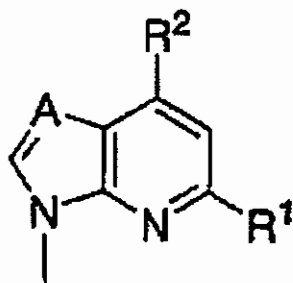
is also important in hydrophobic base pairing during transcription, as already shown for replication.

[0376] The Ds-Pa base pair system is very useful for the purpose of creating novel functional RNA molecules. Since 4-propynylpyrrole-2-carbaldehyde (Pa') is also introduced in a site-specific manner into RNA opposite Ds in the template, a series of 4-position modified Pa base derivatives can also be introduced into RNA. In this case, template DNA can be amplified by PCR of a DNA fragment containing a Ds-Pa base pair. Thus, the Ds-Pa base pair system provides a powerful tool for creating nucleic acids having functional artificial components at desired sites.

CLAIMS

1. A method for replicating a nucleic acid, wherein a deoxyribonucleoside 5'-triphosphate, in which the hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group, is used as a substrate during replication reaction.
2. The method according to claim 1, wherein the substituent is an amino group.
3. The method according to claim 1 or 2, wherein a DNA polymerase having exonuclease activity is used during the replication reaction.
4. The method according to any one of claims 1 to 3, wherein the polymerase having exonuclease activity is selected from the group consisting of the Klenow fragment, T4 DNA polymerase and thermophilic DNA polymerases, each having 3'→5' exonuclease activity.
5. The method according to any one of claims 1 to 4, wherein the deoxyribonucleoside 5'-triphosphate used as a substrate has an unnatural base.
6. The method according to any one of claims 1 to 4, wherein the deoxyribonucleoside 5'-triphosphate used as a substrate has a natural base.
7. The method according to any one of claims 1 to 5, wherein the deoxyribonucleoside 5'-triphosphate used as a substrate has a base represented by the following formula 1:

[Formula 1]



[wherein

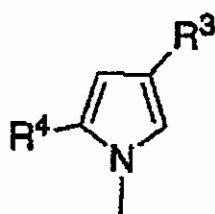
R¹ is hydrogen or an amino group,

R² is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH].

8. The method according to any one of claims 1 to 5, wherein the deoxyribonucleoside 5'-triphosphate used as a substrate has a base represented by the following formula 2:

[Formula 2]



[wherein

R³ is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C₁-C₃ alkyl group, a substituted or unsubstituted C₂-C₃ alkenyl group, or a substituted or unsubstituted C₂-C₃ alkynyl group, and

R⁴ is a formyl group or a nitro group].

9. A deoxyribonucleoside 5'-triphosphate, in which the

hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group.

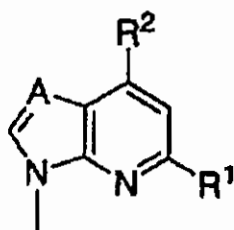
10. A deoxyribonucleoside 5'-triphosphate having an unnatural base, in which the hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group.

11. A deoxyribonucleoside 5'-triphosphate having a natural base, in which the hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group.

12. The use of a deoxyribonucleoside 5'-triphosphate as a substrate in the method for replicating a nucleic acid according to any one of claims 1 to 8, in which the hydroxyl group of phosphoric acid at the γ -position is substituted with a group selected from the group consisting of an amino group, a methylamino group, a dimethylamino group, a mercapto group and a fluoro group.

13. A nucleic acid, in which a nucleotide having a base represented by the following formula 1:

[Formula 3]



[wherein

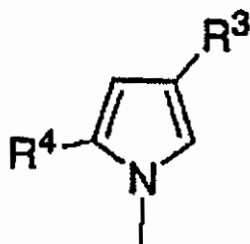
R^1 is hydrogen or an amino group,

R^2 is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH]

forms a base pair with a nucleotide having a base represented by the following formula 2:

[Formula 4]



[wherein

R^3 is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C_1 - C_3 alkyl group, a substituted or unsubstituted C_2 - C_3 alkenyl group, or a substituted or unsubstituted C_2 - C_3 alkynyl group, and

R^4 is a formyl group or a nitro group].

14. The nucleic acid according to claim 13, wherein the base of formula 1 is selected from the group consisting of:

A1) a 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl group;

A2) a 7-(2-thiazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group;

A3) a 7-(1H-2-imidazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group;

- A4) a 5-amino-7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl group;
- A5) a 5-amino-7-(2-thiazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group;
- A6) a 5-amino-7-(1H-2-imidazolyl)-3H-imidazo[4,5-b]pyridin-3-yl group;
- A7) a 4-(2-thienyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group;
- A8) a 4-(2-thiazolyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group;
- A-9) a 4-(1H-2-imidazolyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group;
- A-10) a 6-amino-4-(2-thienyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group;
- A-11) a 6-amino-4-(2-thiazolyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group; and
- A-12) a 6-amino-4-(1H-2-imidazolyl)-1H-pyrrolo[2,3-b]pyridin-1-yl group.

15. The nucleic acid according to claim 13, wherein the base of formula 2 is selected from the group consisting of:

- B1) a 2-formyl-1H-pyrrol-1-yl group;
- B2) a 2-formyl-4-iodo-1H-pyrrol-1-yl group;
- B3) a 2-formyl-4-methyl-1H-pyrrol-1-yl group;
- B4) a 2-formyl-4-(1-propyn-1-yl)-1H-pyrrol-1-yl group;
- B5) a 2-formyl-4-(2-substituted aminovinyl)-1H-pyrrol-1-yl group;
- B6) a 2-formyl-4-(3-substituted amino-1-propyn-1-yl)-1H-pyrrol-1-yl group;

B7) a 2-nitro-1H-pyrrol-1-yl group;

B8) a 2-nitro-4-iodo-1H-pyrrol-1-yl group;

B9) a 2-nitro-4-methyl-1H-pyrrol-1-yl group;

B10) a 2-nitro-4-(1-propyn-1-yl)-1H-pyrrol-1-yl group;

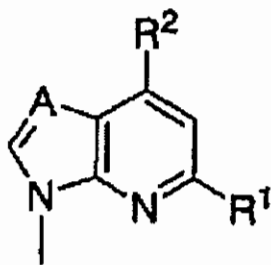
B11) a 2-nitro-4-(2-substituted aminovinyl)-1H-pyrrol-1-yl group; and

B12) a 2-nitro-4-(3-substituted amino-1-propyn-1-yl)-1H-pyrrol-1-yl group.

16. The nucleic acid according to any one of claims 13 to 15, which forms a base pair(s) in the step of transcription, reverse transcription, replication or translation.

17. A method for preparing a nucleic acid containing a nucleotide having a base represented by the following formula 1:

[Formula 5]



[wherein

R^1 is hydrogen or an amino group,

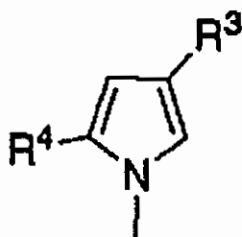
R^2 is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH],

wherein the method comprises effecting transcription, reverse transcription or replication by using, as a template,

a nucleic acid containing a nucleotide having a base represented by the following formula 2:

[Formula 6]



[wherein

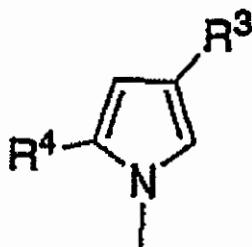
R^3 is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C_1 - C_3 alkyl group, a substituted or unsubstituted C_2 - C_3 alkenyl group, or a substituted or unsubstituted C_2 - C_3 alkynyl group, and

R^4 is a formyl group or a nitro group],

whereby the nucleotide having a base of formula 1 is incorporated at a site complementary to the nucleotide having a base of formula 2.

18. A method for preparing a nucleic acid containing a nucleotide having a base represented by the following formula 2:

[Formula 7]



[wherein

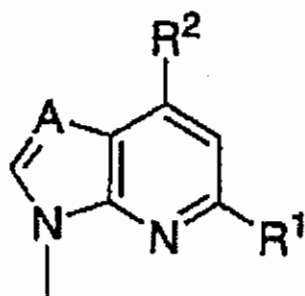
R^3 is a group selected from hydrogen, an iodo group, a

substituted or unsubstituted C₁-C₃ alkyl group, a substituted or unsubstituted C₂-C₃ alkenyl group, or a substituted or unsubstituted C₂-C₃ alkynyl group, and

R⁴ is a formyl group or a nitro group],

wherein the method comprises effecting transcription, reverse transcription or replication by using, as a template, a nucleic acid containing a nucleotide having a base represented by the following formula 1:

[Formula 8]



[wherein

R¹ is hydrogen or an amino group,

R² is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH],

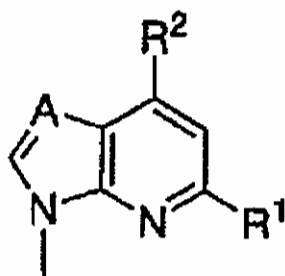
whereby the nucleotide having a base of formula 2 is incorporated at a site complementary to the nucleotide having a base of formula 1.

19. A nucleic acid containing a nucleotide having a base of formula 1 and/or formula 2, which is prepared by the method according to claim 17 or 18.

20. The nucleic acid according to claim 19, which is tRNA, mRNA, antisense DNA or RNA, a ribozyme, an aptamer or siRNA.

21. A ribonucleoside 5'-triphosphate having a base represented by the following formula 1:

[Formula 9]



[wherein

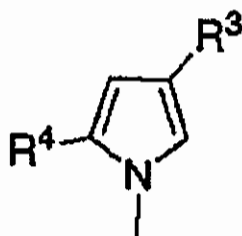
R¹ is hydrogen or an amino group,

R² is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH].

22. A ribonucleoside 5'-triphosphate having a base represented by the following formula 2:

[Formula 10]



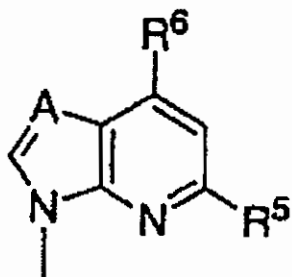
[wherein

R³ is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C₁-C₃ alkyl group, a substituted or unsubstituted C₂-C₃ alkenyl group, or a substituted or unsubstituted C₂-C₃ alkynyl group, and

R⁴ is a formyl group or a nitro group].

23. A 5'-O-(4,4'-dimethoxytrityl)-3'-O-(2-cyanoethyl N,N-diisopropylphosphoroamidite) deoxyribonucleoside having a base represented by the following formula 3:

[Formula 11]



[wherein

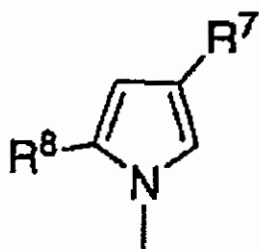
R⁵ is hydrogen or a substituted amino group,

R⁶ is a substituted or unsubstituted 2-thienyl group, a substituted or unsubstituted 2-thiazolyl group, or a substituted or unsubstituted 1H-2-imidazolyl group, and

A is N or CH].

24. A 5'-O-(4,4'-dimethoxytrityl)-3'-O-(2-cyanoethyl N,N-diisopropylphosphoroamidite) deoxyribonucleoside having a base represented by the following formula 4:

[Formula 12]



[wherein

R⁷ is a group selected from hydrogen, an iodo group, a substituted or unsubstituted C₁-C₃ alkyl group, a substituted or unsubstituted C₂-C₃ alkenyl group, or a substituted or

unsubstituted C₂-C₃ alkynyl group, and

R⁸ is a formyl group or a nitro group,

excluding the case where R⁷ is hydrogen or a 1-propynyl group and R⁸ is a formyl group].

25. A nucleic acid containing a nucleotide having a base represented by formula 2, which is prepared by the method according to claim 18, wherein the substituent R³ in formula 2 is a C₁-C₃ alkyl group, a C₂-C₃ alkenyl group or a C₂-C₃ alkynyl group, each being substituted with biotin or a fluorescent molecule.

26. The ribonucleoside 5'-triphosphate according to claim 22, wherein R³ is a C₁-C₃ alkyl group, a C₂-C₃ alkenyl group or a C₂-C₃ alkynyl group, each being substituted with biotin or a fluorescent molecule.

27. The 5'-O-(4,4'-dimethoxytrityl)-3'-O-(2-cyanoethyl N,N-diisopropylphosphoroamidite) deoxyribonucleoside according to claim 24, wherein R⁷ is a C₁-C₃ alkyl group, a C₂-C₃ alkenyl group or a C₂-C₃ alkynyl group, each being substituted with biotin or a fluorescent molecule.

SEQUENCE LISTING

<110> University of Tokyo
 <110> Riken

<120> METHOD FOR NUCLEIC ACID REPLICATION AND NOVEL ARTIFICIAL BASE PAIRS

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 adenine,
 guanine,
 thymine
 cytosine

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 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl,
 adenine,
 guanine,
 thymine
 cytosine

<220>
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 <223> n means any one of the followings:
 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl,
 adenine,
 guanine

 <220>
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 cactnctcgg gattccctat agtgagtcgt attat 35

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 2-formyl-1H-pyrrol-1-yl,
 2-formyl-4-(1-propynyl)-1H-pyrrol-1-yl,
 thymine
 cytosine

<220>
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 2-formyl-1H-pyrrol-1-yl,
 2-formyl-4-(1-propynyl)-1H-pyrrol-1-yl,
 uracil
 cytosine

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 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl,
 adenine,
 guanine

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 gggagcuucc cunugag 17

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 <211> 91
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 <223> nnnnn means any one of the followings:
 cytosine- 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - adenine - guanine - thymine,
 cytosine - thymine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - guanine - thymine,
 cytosine - thymine - adenine - guanine - thymine,

<220>
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 adenine - cytosine - thymine - 2-formyl-1H-pyrrol-1-yl - guanine
 adenine - cytosine - 2-formyl-1H-pyrrol-1-yl - adenine - guanine
 adenine - cytosine - thymine - adenine - guanine

 <220>
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 cggttgcnnn nnatgatggg g 81

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 gaagcc 66

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 <223> nnnnn means any one of the followings:
 cytosine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - adenine - guanine - thymine,
 cytosine - thymine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - guanine - thymine,
 cytosine - 2-formyl-1H-pyrrol-1-yl - adenine - guanine - thymine,
 cytosine - thymine - 2-formyl-1H-pyrrol-1-yl - guanine - thymine,
 cytosine - adenine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - cytosine - thymine,
 cytosine - guanine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - thymine - thymine,
 cytosine - guanine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - thymine - cytosine,
 thymine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - adenine - guanine - thymine,
 cytosine - thymine - adenine - guanine - thymine,
 cytosine - thymine - adenine - cytosine - thymine

<220>
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 a 61

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 <223> nnnnn means any one of the followings:
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 adenine - cytosine - 2-formyl-1H-pyrrol-1-yl - adenine - guanine,
 adenine - cytosine - thymine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - guanine,
 adenine - cytosine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - adenine - guanine,
 adenine - guanine - 2-formyl-1H-pyrrol-1-yl - thymine - guanine,
 adenine - adenine - 2-formyl-1H-pyrrol-1-yl - cytosine - guanine,
 guanine - adenine - 2-formyl-1H-pyrrol-1-yl - cytosine - guanine,
 adenine - cytosine - thymine - 2-formyl-1H-pyrrol-1-yl - adenine,
 adenine - cytosine - thymine - adenine - guanine,
 adenine - guanine - thymine - adenine - guanine

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<210> 21
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 <213> Artificial sequence

<220>
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 <223> nnnnn means any one of the followings:
 cytosine - thymine - 2-formyl-1H-pyrrol-1-yl - adenine - adenine,
 cytosine - 2-formyl-1H-pyrrol-1-yl -adenine - adenine - adenine,
 cytosine - thymine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - adenine - adenine,
 cytosine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl -adenine - adenine - adenine,
 cytosine - thymine - adenine - adenine - adenine

<220>
 <223> Synthesized DNA fragment for PCR, sequencing and transcription

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 tgccgtc 67

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<220>
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 <222> 2
 <223> n means gm

<220>
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 <223> nnnnn means any one of the followings:
 thymine - thymine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - adenine - guanine,
 thymine - thymine - thymine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - guanine,
 thymine - thymine - 2-formyl-1H-pyrrol-1-yl - adenine - guanine,
 thymine - thymine - thymine - 2-formyl-1H-pyrrol-1-yl - guanine,
 thymine - thymine - thymine - adenine - guanine

<220>
 <223> Synthesized DNA fragment for PCR, sequencing and transcription

<400> 22
 nngtgggtggg ggaaggattc gaaccttcga agtctgtgac ggcagannnn nagtctgctc 60

<210> 23
 <211> 25
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Synthesized primer for PCR and sequencing

<400> 23
 gaaattaata cgactcacta taggg 25

<210> 24
 <211> 24
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Synthesized primer for PCR and sequencing

<400> 24
 ttccacacag gaaacagcta tgac 24

<210> 25
 <211> 86
 <212> RNA
 <213> Artificial sequence

<220>
 <221> misc_feature
 <222> 35...37
 <223> nnn means any one of the followings:
 cytosine - uracil - 2-formyl-4-(1-propynyl)-1H-pyrrol-1-yl,
 cytosine - 2-formyl-4-(1-propynyl)-1H-pyrrol-1-yl - adenine,
 cytosine - uracil - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl,
 cytosine - 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl - adenine,
 cytosine - uracil - adenine

<220>
 <223> RNA transcript

<400> 25
 gguggggguuc ccgagcggcc aaaggagca gacunnnaau cugccguca ucgacuucga 60
 agguucgaau ccuuccccca ccacca 86

<210> 26
 <211> 66
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Synthesized DNA fragment for PCR, sequencing and transcription

<400> 26
 gaaattaata cgactcacta tagggccaac cagaagaagg agacagacca agggcaccaa 60
 gaagcc 66

<210> 27
 <211> 47
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Synthesized DNA fragment for PCR, sequencing and transcription
 <400> 27
 ggaccccaac aacaygagca accgccgaac gggcaagacg cagcgcg 47
 <210> 28
 <211> 61
 <212> DNA
 <213> Artificial sequence
 <220>
 <223> Synthesized DNA fragment for PCR, sequencing and transcription
 <400> 28
 ggggaacaca acagcaccaa cagcaccaac aaccgggac aaagcagcaa ccaggagaa 60
 a 61
 <210> 29
 <211> 53
 <212> DNA
 <213> Artificial sequence
 <220>
 <223> Synthesized DNA fragment for PCR, sequencing and transcription
 <400> 29
 ccttggtctg tctccttctt ctggtggcc ctatagttag tcgtattaat ttc 53
 <210> 30
 <211> 50
 <212> DNA
 <213> Artificial sequence
 <220>
 <223> Synthesized DNA fragment for PCR, sequencing and transcription
 <400> 30
 cttgcccggt cggcggttgc tctgttgtt ggggtccggc ttcttggtgc 50
 <210> 31
 <211> 71
 <212> DNA
 <213> Artificial sequence
 <220>
 <223> Synthesized DNA fragment for PCR, sequencing and transcription
 <400> 31
 ttctccctg gttctgtctt tgtccgggt tgttggtgct gttggtgct ttgtgtccc 60
 ccgcgtgcg t 71
 <210> 32
 <211> 20
 <212> DNA
 <213> Artificial sequence
 <220>
 <223> Synthesized DNA fragment for replication
 <400> 32
 gataatacga ctactatag
 <210> 33
 <211> 55
 <212> DNA
 <213> Artificial sequence
 <220>
 <221> misc_feature
 <222> 28
 <223> n means 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl
 <220>
 <223> Synthesized DNA fragment for replication

```

<400> 33
gcagtgaaga attcgcttag atgtggancc attccctata gtgagtcgta ttatc 55

<210> 34
<211> 55
<212> DNA
<213> Artificial sequence

<220>
<221> misc_feature
<222> 25
<223> n means 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl

<220>
<223> Synthesized DNA fragment for replication

<400> 34
gcagtgaaga attcgcttag atgtngatcc attccctata gtgagtcgta ttatc 55

<210> 35
<211> 55
<212> DNA
<213> Artificial sequence

<220>
<221> misc_feature
<222> 21
<223> n means 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl

<220>
<223> Synthesized DNA fragment for replication

<400> 35
gcagtgaaga attcgcttag angtgatcc attccctata gtgagtcgta ttatc 55

<210> 36
<211> 55
<212> DNA
<213> Artificial sequence

<220>
<221> misc_feature
<222> 31
<223> n means 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl

<220>
<223> Synthesized DNA fragment for replication

<400> 36
gcagtgaaga attcgcttag atgtggatcc nttccctata gtgagtcgta ttatc 55

<210> 37
<211> 55
<212> DNA
<213> Artificial sequence

<220>
<221> misc_feature
<222> 34
<223> n means 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl

<220>
<223> Synthesized DNA fragment for replication

<400> 37
gcagtgaaga attcgcttag atgtggatcc attncctata gtgagtcgta ttatc 55

<210> 38
<211> 55
<212> DNA
<213> Artificial sequence

<220>
<223> Synthesized DNA fragment for replication

```

<400> 38
gcagtgaaga attcgcttag atgtggatcc attccctata gtgagtcgta ttatc 55

<210> 39
<211> 21
<212> DNA
<213> Artificial sequence

<220>
<223> Synthesized DNA fragment for transcription

<400> 39
ataatacgac tcactatagg g 21

<210> 40
<211> 35
<212> DNA
<213> Artificial sequence

<220>
<221> misc_feature
<222> 5
<223> n means 7-(2-thienyl)-3H-imidazo[4,5-b]pyridin-3-yl or adenine

<220>
<223> Synthesized DNA fragment for transcription

<400> 40
cactnctcgg gattccctat agtgagtcgt attat 35

<210> 41
<211> 17
<212> RNA
<213> Artificial sequence

<220>
<221> misc_feature
<222> 13
<223> n means any one of the followings:
2-formyl-4-[3-[6-(fluorescein-5-carboxamido)hexanamido]-1-propynyl]-1H-pyrrol-1-yl,
2-formyl-4-[3-[6-(tetramethylrhodamine-5-carboxamido)hexanamido]-1-propynyl]-1H-pyrrol-1-yl,

<400> 41
gggaaucccg agnagug 17