TITLE OF THE INVENTION

Compositions against SARS-coronavirus and uses thereof

FIELD OF THE INVENTION

The invention relates to medicine. In particular the invention relates to compositions comprising binding molecules capable of specifically binding to and neutralizing SARS-coronavirus (SARS-CoV). The compositions are useful in the diagnosis of SARS-CoV and the prophylaxis and/or treatment of a condition resulting from SARS-CoV.

BACKGROUND OF THE INVENTION

Recently a new and in several cases deadly clinical syndrome was observed in the human population, now called severe acute respiratory syndrome (SARS) (Holmes, 2003). The syndrome is caused by a novel coronavirus (Ksiazek *et al.*, 2003), referred to as the SARS-CoV. The genome sequence of SARS-CoV has been determined (Rota *et al.*, 2003; Marra *et al.*, 2003). However, much remains to be learnt about this virus, and means and methods for diagnostics, prophylaxis and/or treatment of the virus and the syndrome are needed. The present invention provides means and methods for use in diagnostics, prevention and/or treatment of SARS-CoV.

DESCRIPTION OF THE FIGURES

Figure 1 shows results from an ELISA, wherein the binding of the single-chain phage antibodies called SC03-014 and SC03-022 to an immobilized UV-inactivated SARS-CoV preparation (left column) or immobilized FBS (right column) was measured. The binding of the control single-chain phage antibody called SC02-006 is also shown. On the y-axis the absorbance at 492 nm is shown.

Figure 2 shows an ELISA binding of IgGs CR03-014, CR03-022, control IgG and no IgG to an inactivated SARS-CoV preparation. On the Y-axis the absorbance at 492 nm is shown.

Figure 3 shows a FACS binding of the scFv phage antibodies SC03-014, SC03-022 and a control scFv phage antibody to the full length S protein expressed on HEK293T cells (left column) or mock transfected HEK293T cells (right column). On the Y-axis the mean fluorescense intensity is shown.

Figure 4 shows an ELISA binding of the antibodies CR03-014, CR03-022 and a control antibody to the S565 fragment (amino acids 1-565 of the S protein of SARS-CoV; left column), S318-510 fragment (amino acids 318-510 of the S protein of SARS-CoV; middle column) and an irrelevant control myc-tagged antigen (right column). On the Y-axis the absorbance at 492 nm is shown.

Figure 5 shows an ELISA binding of dilutions of antibodies CR03-014, CR03-022 and a control antibody to the S565 fragment of the S protein of SARS-CoV. On the Y-axis the absorbance at 492 nm and on the X-axis the amount of antibody in µg/ml is shown.

Figure 6 shows a competition ELISA for binding to the S565 fragment. Figure 6A shows competition between biotinylated antibody CR03-014 without competing antibody or with 1, 5 or 25 µg/ml competing antibody CR03-014, CR03-022 or control antibody. Figure 6B shows competition between biotinylated antibody CR03-022 with or without the competing antibodies as described above. On the Y-axis the % of maximal binding is

shown and on the X-axis the amount of the competing antibody in µg/ml.

Figure 7 shows a sandwich ELISA using anti-S protein antibodies. Immobilized antibodies CR03-014 and CR03-022 were used to capture S protein fragment S318-510. Bound fragment was detected using biotinylated antibody CR03-014, CR03-022 or control antibody. On the Y-axis the absorbance at 492 nm is shown.

Figure 8 shows binding of the monoclonal anti-SARS-antibodies CR03-014 and CR03-022 to the amino acid region of 318-510 of the S protein of the SARS-CoV strain Frankfurt 1 (called WT S318-510) and naturally occuring variants of the WT S318-510 fragment (variant A, mutation K344R; variant B, mutation S353F; variant C, mutation R426G and N437D; variant D, mutation Y436H; variant E, mutation Y442S; variant F, mutation N479S; variant G, mutation K344R, F360S, N479K and T487S; variant H, mutation K344R, F360S, L472P, D480G, and T487S; variant I, mutation K344R and F501Y). The control is an irrelevant myc-His tagged protein. On the Y-axis the absorbance at 492 nm is shown.

Figure 9 shows the comparison of the nucleotide and amino acid sequences of the SARS-CoV wild-type strain (SARS-CoV strain HKU 39849) and escape viruses of antibody CR03-014. Virus-infected cells were harvested 2 days post-infection and total RNA was isolated. cDNA was generated and used for DNA sequencing. Regions containing mutations are shown and the mutations are indicated in bold. Numbers above amino acids indicate amino acids numbers from S protein including signal

peptide. The sequences in Figure 9 are also represented by SEQ ID Nos:118-121.

Figure 10 shows binding of the monoclonal anti-SARS-antibodies CR03-014 and CR03-022 to the amino acid region of 318-510 of the S protein of the SARS-CoV strain Frankfurt 1 (called FRA1 S318-510) and an escape variant of antibody CR03-014 harboring a P462L substitution. On the Y-axis the absorbance at 492 nm is shown.

DESCRIPTION OF THE INVENTION

Herebelow follow definitions of terms as used in the invention

DEFINITIONS

Binding molecule

As used herein the term "binding molecule" refers to an intact immunoglobulin including monoclonal antibodies, such as chimeric, humanised or human monoclonal antibodies, or to an antigen-binding and/or variable domain comprising fragment of an immunoglobulin that competes with the intact immunoglobulin for specific binding to the binding partner of the immunoglobulin, *e.g.* the SARS-CoV. Regardless of structure, the antigen-binding fragment binds with the same antigen that is recognised by the intact immunoglobulin. An antigen-binding fragment can comprise a peptide or polypeptide comprising an amino acid sequence of at least 2 contiguous amino acid residues, at least 5 contiguous amino acid residues, at least 10 contiguous amino acid residues, at least 15 contiguous amino acid residues, at least 20 contiguous amino acid residues, at least 25 contiguous amino acid residues, at least 30 contiguous amino acid residues, at least 35 contiguous

amino acid residues, at least 40 contiguous amino acid residues, at least 50 contiguous amino acid residues, at least 60 contiguous amino residues, at least 70 contiguous amino acid residues, at least contiguous 80 amino acid residues, at least contiguous 90 amino acid residues, at least contiguous 100 amino acid residues, at least contiguous 125 amino acid residues, at least 150 contiguous amino acid residues, at least contiguous 175 amino acid residues, at least 200 contiguous amino acid residues, or at least contiguous 250 amino acid residues of the amino acid sequence of the binding molecule.

The term "binding molecule", as used herein includes all immunoglobulin classes and subclasses known in the art. Depending on the amino acid sequence of the constant domain of their heavy chains, binding molecules can be divided into the five major classes of intact antibodies: IgA, IgD, IgE, IgG, and IgM, and several of these may be further divided into subclasses (isotypes), *e.g.*, IgA1, IgA2, IgG1, IgG2, IgG3 and IgG4.

Antigen-binding fragments include, *inter alia*, Fab, F(ab'), $F(ab')_2$, Fv, dAb, Fd, complementarity determining region (CDR) fragments, single-chain antibodies (scFv), bivalent single-chain antibodies, single-chain phage antibodies, diabodies, triabodies, tetrabodies, (poly)peptides that contain at least a fragment of an immunoglobulin that is sufficient to confer specific antigen binding to the (poly)peptide, etc. The above fragments may be produced synthetically or by enzymatic or chemical cleavage of intact immunoglobulins or they may be genetically engineerd by recombinant DNA techniques. The methods of production are well known in the art and are described, for example, in Antibodies: A Laboratory Manual, Edited by: E. Harlow and D,

Lane (1988), Cold Spring Harbor Laboratory, Cold Spring Harbor, New York, which is incorporated herein by reference. A binding molecule or antigen-binding fragment thereof may have one or more binding sites. If there is more than one binding site, the binding sites may be identical to one another or they may be different.

The binding molecule can be a naked or unconjugated binding molecule but can also be part of an immunoconjugate. A naked or unconjugated binding molecule is intended to refer to a binding molecule that is not conjugated, operatively linked or otherwise physically or functionally associated with an effector moiety or tag, such as *inter alia* a toxic substance, a radioactive substance, a liposome, an enzyme. It will be understood that naked or unconjugated binding molecules do not exclude binding molecules that have been stabilized, multimerized, humanized or in any other way manipulated, other than by the attachment of an effector moiety or tag. Accordingly, all post-translationally modified naked and unconjugated binding molecules are included herewith, including where the modifications are made in the natural binding molecule-producing cell environment, by a recombinant binding molecule-producing cell, and are introduced by the hand of man after initial binding molecule preparation. Of course, the term naked or unconjugated binding molecule does not exclude the ability of the binding molecule to form functional associations with effector cells and/or molecules after administration to the body, as some of such interactions are necessary in order to exert a biological effect.

Biological sample

As used herein, the term "biological sample" encompasses a variety of sample types, including blood and other liquid

samples of biological origin, solid tissue samples such as a biopsy specimen or tissue cultures, or cells derived therefrom and the progeny thereof. The term also includes samples that have been manipulated in any way after their procurement, such as by treatment with reagents, solubilization, or enrichment for certain components, such as proteins or polynucleotides. The term encompasses various kinds of clinical samples obtained from any species, and also includes cells in culture, cell supernatants and cell lysates.

Complementarity determining regions (CDR)

The term "complementarity determining regions" as used herein means sequences within the variable regions of binding molecules, such as immunoglobulins, that usually contribute to a large extent to the antigen binding site which is complementary in shape and charge distribution to the epitope recognised on the antigen. The CDR regions can be specific for linear epitopes, discontinuous epitopes, or conformational epitopes of proteins or protein fragments, either as present on the protein in its native conformation or, in some cases, as present on the proteins as denatured, *e.g.*, by solubilization in SDS. Epitopes may also consist of posttranslational modifications of proteins.

Expression-regulating nucleic acid sequence

The term "expression-regulating nucleic acid sequence" as used herein refers to polynucleotide sequences necessary for and/or affecting the expression of an operably linked coding sequence in a particular host organism. The expression-regulating nucleic acid sequences, such as *inter alia* appropriate transcription initiation, termination, promoter, enhancer sequences; repressor or activator sequences; efficient RNA

processing signals such as splicing and polyadenylation signals; sequences that stabilize cytoplasmic mRNA; sequences that enhance translation efficiency (*e.g.*, ribosome binding sites); sequences that enhance protein stability; and when desired, sequences that enhance protein secretion, can be any nucleic acid sequence showing activity in the host organism of choice and can be derived from genes encoding proteins, which are either homologous or heterologous to the host organism. The identification and employment of expression-regulating sequences is routine to the person skilled in the art.

Functional variant

The term "functional variant", as used herein, refers to a binding molecule that comprises a nucleotide and/or amino acid sequence that is altered by one or more nucleotides and/or amino acids compared to the nucleotide and/or amino acid sequences of the parent binding molecule and that is still capable of competing for binding to the binding partner, *e.g.* SARS-CoV, with the parent binding molecule. In other words, the modifications in the amino acid and/or nucleotide sequence of the parent binding molecule do not significantly affect or alter the binding characteristics of the binding molecule encoded by the nucleotide sequence or containing the amino acid sequence, *i.e.* the binding molecule is still able to recognize and bind its target. The functional variant may have conservative sequence modifications including nucleotide and amino acid substitutions, additions and deletions. These modifications can be introduced by standard techniques known in the art, such as site-directed mutagenesis and random PCR-mediated mutagenesis, and may comprise natural as well as non-natural nucleotides and amino acids.

Conservative amino acid substitutions include the ones in which the amino acid residue is replaced with an amino acid residue having similar structural or chemical properties. Families of amino acid residues having similar side chains have been defined in the art. These families include amino acids with basic side chains (*e.g.*, lysine, arginine, histidine), acidic side chains (*e.g.*, aspartic acid, glutamic acid), uncharged polar side chains (*e.g.*, glycine, asparagine, glutamine, serine, threonine, tyrosine, cystine, tryptophan), nonpolar side chains (*e.g.*, alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine), beta-branched side chains (*e.g.*, threonine, valine, isoleucine) and aromatic side chains (*e.g.*, tyrosine, phenylalanine, tryptophan, histidine). It will be clear to the skilled artisan that other classifications of amino acid residue families than the one used above can also be employed. Furthermore, a variant may have non-conservative amino acid substitutions, *e.g.*, replacement of an amino acid with an amino acid residue having different structural or chemical properties. Similar minor variations may also include amino acid deletions or insertions, or both. Guidance in determining which amino acid residues may be substituted, inserted, or deleted without abolishing immunological activity may be found using computer programs well known in the art.

A mutation in a nucleotide sequence can be a single alteration made at a locus (a point mutation), such as transition or transversion mutations, or alternatively, multiple nucleotides may be inserted, deleted or changed at a single locus. In addition, one or more alterations may be made at any number of loci within a nucleotide sequence. The mutations may be performed by any suitable method known in the art.

Host

The term "host", as used herein, is intended to refer to an organism or a cell into which a vector such as a cloning vector or an expression vector has been introduced. The organism or cell can be prokaryotic or eukaryotic. It should be understood that this term is intended to refer not only to the particular subject organism or cell, but to the progeny of such an organism or cell as well. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent organism or cell, but are still included within the scope of the term "host" as used herein.

Human

The term "human", when applied to binding molecules as defined herein, refers to molecules that are either directly derived from a human or based upon a human sequence. When a binding molecule is derived from or based on a human sequence and subsequently modified, it is still to be considered human as used throughout the specification. In other words, the term human, when applied to binding molecules is intended to include binding molecules having variable and constant regions derived from human germline immunoglobulin sequences based on variable or constant regions either or not occuring in a human or human lymphocyte or in modified form. Thus, the human binding molecules may include amino acid residues not encoded by human germline immunoglobulin sequences, comprise substitutions and/or deletions (*e.g.*, mutations introduced by for instance random or site-specific mutagenesis *in vitro* or by somatic mutation *in vivo*). "Based on" as used herein refers

to the situation that a nucleic acid sequence may be exactly copied from a template, or with minor mutations, such as by error-prone PCR methods, or synthetically made matching the template exactly or with minor modifications. Semisynthetic molecules based on human sequences are also considered to be human as used herein.

Isolated

The term "isolated", when applied to binding molecules as defined herein, refers to binding molecules that are substantially free of other proteins or polypeptides, particularly free of other binding molecules having different antigenic specificities, and are also substantially free of other cellular material and/or chemicals. For example, when the binding molecules are recombinantly produced, they are preferably substantially free of culture medium, and when the binding molecules are produced by chemical synthesis, they are preferably substantially free of chemical precursors or other chemicals, *i.e.*, they are separated from chemical precursors or other chemicals which are involved in the synthesis of the protein. The term "isolated" when applied to nucleic acid molecules encoding binding molecules as defined herein, is intended to refer to nucleic acid molecules in which the nucleotide sequences encoding the binding molecules are free of other nucleotide sequences, particularly nucleotide sequences encoding binding molecules that bind binding partners other than SARS-CoV. Furthermore, the term "isolated" refers to nucleic acid molecules that are substantially separated from other cellular components that naturally accompany the native nucleic acid molecule in its natural host, *e.g.*, ribosomes, polymerases, or genomic sequences with which it is naturally associated. Moreover, "isolated" nucleic

acid molecules, such as a cDNA molecules, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized.

Monoclonal antibody

The term "monoclonal antibody" as used herein refers to a preparation of antibody molecules of single molecular composition. A monoclonal antibody displays a single binding specificity and affinity for a particular epitope. Accordingly, the term "human monoclonal antibody" refers to an antibody displaying a single binding specificity which have variable and constant regions derived from or based on human germline immunoglobulin sequences or derived from completely synthetic sequences. The method of preparing the monoclonal antibody is not relevant.

Nucleic acid molecule

The term "nucleic acid molecule" as used in the present invention refers to a polymeric form of nucleotides and includes both sense and antisense strands of RNA, cDNA, genomic DNA, and synthetic forms and mixed polymers of the above. A nucleotide refers to a ribonucleotide, deoxynucleotide or a modified form of either type of nucleotide. The term also includes single- and double-stranded forms of DNA. In addition, a polynucleotide may include either or both naturally-occurring and modified nucleotides linked together by naturally-occurring and/or non-naturally occurring nucleotide linkages. The nucleic acid molecules may be modified chemically or biochemically or may contain non-natural or derivatized nucleotide bases, as will be readily

appreciated by those of skill in the art. Such modifications include, for example, labels, methylation, substitution of one or more of the naturally occurring nucleotides with an analog, internucleotide modifications such as uncharged linkages (*e.g.*, methyl phosphonates, phosphotriesters, phosphoramidates, carbamates, etc.), charged linkages (*e.g.*, phosphorothioates, phosphorodithioates, etc.), pendent moieties (*e.g.*, polypeptides), intercalators (*e.g.*, acridine, psoralen, etc.), chelators, alkylators, and modified linkages (*e.g.*, alpha anomeric nucleic acids, etc.). The above term is also intended to include any topological conformation, including single-stranded, double-stranded, partially duplexed, triplex, hairpinned, circular and padlocked conformations. Also included are synthetic molecules that mimic polynucleotides in their ability to bind to a designated sequence via hydrogen bonding and other chemical interactions. Such molecules are known in the art and include, for example, those in which peptide linkages substitute for phosphate linkages in the backbone of the molecule. A reference to a nucleic acid sequence encompasses its complement unless otherwise specified. Thus, a reference to a nucleic acid molecule having a particular sequence should be understood to encompass its complementary strand, with its complementary sequence. The complementary strand is also useful, *e.g.*, for antisense therapy, hybridization probes and PCR primers.

Operably linked

The term "operably linked" refers to two or more nucleic acid sequence elements that are usually physically linked and are in a functional relationship with each other. For instance, a promoter is operably linked to a coding sequence if the promoter is able to initiate or regulate the transcription or

expression of a coding sequence, in which case the coding sequence should be understood as being "under the control of" the promoter.

Pharmaceutically acceptable excipient

By "pharmaceutically acceptable excipient" is meant any inert substance that is combined with an active molecule such as a drug, agent, or binding molecule for preparing an agreeable or convenient dosage form. The "pharmaceutically acceptable excipient" is an excipient that is non-toxic to recipients at the dosages and concentrations employed and is compatible with other ingredients of the formulation comprising the drug, agent or binding molecule.

Specifically Binding

The term "specifically binding", as used herein, in reference to the interaction of a binding molecule, *e.g.* an antibody, and its binding partner, *e.g.* an antigen, means that the interaction is dependent upon the presence of a particular structure, *e.g.* an antigenic determinant or epitope, on the binding partner. In other words, the antibody preferentially binds or recognizes the binding partner even when the binding partner is present in a mixture of other molecules or organisms. The binding may be mediated by covalent or non-covalent interactions or a combination of both. In yet other words, the term "specifically binding" means immunospecifically binding to an antigen or a fragment thereof and not immunospecifically binding to other antigens. A binding molecule that immunospecifically binds to an antigen may bind to other peptides or polypeptides with lower affinity as determined by, *e.g.*, radioimmunoassays (RIA), enzyme-linked immunosorbent assays (ELISA), BIACORE, or other assays known

in the art. Binding molecules or fragments thereof that immunospecifically bind to an antigen may be cross-reactive with related antigens. Preferably, binding molecules or fragments thereof that immunospecifically bind to an antigen do not cross-react with other antigens.

Therapeutically effective amount

The term "therapeutically effective amount" refers to an amount of the binding molecule as defined herein that is effective for preventing, ameliorating and/or treating a condition resulting from infection with SARS-CoV.

Treatment

The term "treatment" refers to therapeutic treatment as well as prophylactic or preventative measures to cure or halt or at least retard disease progress. Those in need of treatment include those already inflicted with a condition resulting from infection with SARS-CoV as well as those in which infection with SARS-CoV is to be prevented. Subjects partially or totally recovered form infection with SARS-CoV might also be in need of treatment. Prevention encompasses inhibiting or reducing the spread of SARS-CoV or inhibiting or reducing the onset, development or progression of one or more of the symptoms associated with infection with SARS-CoV.

Vector

The term "vector" denotes a nucleic acid molecule into which a second nucleic acid molecule can be inserted for introduction into a host where it will be replicated, and in some cases expressed. In other words, a vector is capable of transporting a nucleic acid molecule to which it has been linked. Cloning as well as expression vectors are contemplated by the term

"vector", as used herein. Vectors include, but are not limited to, plasmids, cosmids, bacterial artificial chromosomes (BAC) and yeast artificial chromosomes (YAC) and vectors derived from bacteriophages or plant or animal (including human) viruses. Vectors comprise an origin of replication recognised by the proposed host and in case of expression vectors, promoter and other regulatory regions recognised by the host. A vector containing a second nucleic acid molecule is introduced into a cell by transformation, transfection, or by making use of viral entry mechanisms. Certain vectors are capable of autonomous replication in a host into which they are introduced (*e.g.*, vectors having a bacterial origin of replication can replicate in bacteria). Other vectors can be integrated into the genome of a host upon introduction into the host, and thereby are replicated along with the host genome.

SUMMARY OF THE INVENTION

The invention provides compositions of binding molecules capable of specifically binding to SARS-CoV and having SARS-CoV neutralizing activity. In a preferred embodiment, said binding molecules are human binding molecules. The invention further provides for the use of the compositions of the invention in the prophylaxis and/or treatment of a subject having, or at risk of developing, a condition resulting from SARS-CoV. Besides that, the invention pertains to the use of the compositions of the invention in the diagnosis/detection of SARS-CoV.

DETAILED DESCRIPTION OF THE INVENTION

In a first aspect the present invention encompasses compositions comprising at least two binding molecules. In an embodiment the binding molecules are immunoglobulins. Fragments of immunoglobulins still having the desired functionality and/or activity of the complete immunoglobulin are also considered immunoglobulins according to the invention. Preferably, the at least two binding molecules, *i.e.* immunoglobulins, are capable of specifically binding to a coronavirus. Coronaviruses include, but not limited to, avian infectious bronchitis virus, avian infectious laryngotracheitis virus, enteric coronavirus, equine coronavirus, coronavirus Group 1 species such as for instance human coronavirus 229E or human coronavirus NL63, coronavirus Group 2 species such as human coronavirus OC43 or chicken enteric coronavirus, coronavirus Group 3 species, human enteric coronavirus 4408, and SARS-CoV. The compositions can be administered to a mammal to treat, prevent or ameliorate one or more symptoms associated with a coronavirus infection.

In an embodiment the invention relates to synergistic compositions, *i.e.* compositions exhibiting synergistic coronavirus neutralizing activity. In other words, the compositions comprise at least two binding molecules, *i.e.* immunoglobulins, that are capable of specifically binding to a coronavirus and that have coronavirus neutralizing activity, characterized in that the binding molecules act synergistically in neutralizing coronavirus. As used herein, the term "synergistic" means that the combined effect of the binding molecules when used in combination is greater than their additive effects when used individually. In other words, the neutralizing activity of the composition is greater than the sum of the neutralizing activity of each immunoglobulin alone. In an embodiment none of the binding molecules, *i.e.*

immunoglobulins, present in the synergistic coronavirus neutralizing activity exhibiting compositions may have coronavirus neutralizing activity when used as an individual binding molecule. Alternatively, one binding molecule of the at least two binding molecules in the compositions exhibiting synergistic coronavirus neutralizing activity may have coronavirus neutralizing activity when used individually. In a preferred embodiment both of the at least two binding molecules, *i.e.* immunoglobulins, have coronavirus neutralizing activity when used individually. In an embodiment one of the at least two binding molecules in the synergistic coronavirus neutralizing activity exhibiting compositions may bind to a coronavirus and the other binding molecule may bind to a cell associated receptor of the coronavirus. Alternatively, both binding molecules may bind to either the coronavirus or cell associated receptor.

In a preferred embodiment of the invention the coronavirus is a SARS-CoV including animal or human SARS-CoV. Preferably, the SARS-CoV is a human SARS-CoV. In another aspect the invention thus provides compositions comprising at least two binding molecules, *i.e.* immunoglobulins, capable of specifically binding to SARS-CoV and preferably having SARS-CoV neutralizing activity. The compositions preferably exhibit synergistic SARS-CoV neutralizing activity. In other words, the compositions comprise at least two binding molecules, *i.e.* immunoglobulins, having SARS-CoV neutralizing activity, characterized in that the binding molecules act synergistically in neutralizing SARS-CoV. The SARS-CoV neutralizing activity of the composition is greater than the sum of the neutralizing activity of each immunoglobulin alone. In a preferred embodiment of the invention, the binding molecules in the compositions act synergistically in

neutralizing a plurality of SARS-CoV strains (see Table 1 for a list of several known SARS-CoV genome sequences and S protein genes). In another embodiment each of the immunoglobulins in the composition is capable of neutralizing a plurality of (different) SARS-CoV strains, preferably human SARS-CoV strains. In another embodiment at least one of the binding molecules, *i.e.* immunoglobulins, of the compositions of the invention is capable of neutralizing an animal SARS-CoV. The binding molecules in the compositions of the invention may neutralize coronavirus infectivity, such as SARS-CoV infectivity, by several modes of action including, but not limited to, preventing the attachment of the coronavirus to possible receptors on host cells, inhibition of the release of RNA into the cytoplasm of the cell, prevention of RNA transcription or translation, or inhibition or downregulation of coronavirus replication. Furthermore, the binding molecules may act by fixing complement or be capable of assisting in the lysis of enveloped coronavirus. They might also act as opsonins and augment phagocytosis of coronavirus either by promoting its uptake via Fc or C3b receptors or by agglutinating the coronavirus to make it more easily phagocytosed. The binding molecules in the compositions may have similar modes of action or may have different modes of action. In a specific embodiment, the compositions neutralize coronavirus, such as SARS-CoV, infectivity by at least 10%, at least 20%, at least 30%, at least 40%, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, or at least 99% relative to infection of host cells by the coronavirus in the absence of said compositions. Assays for measuring virus neutralizing activity are known to the skilled person. Examples of such assays are described below.

The binding molecules, *i.e.* immunoglobulins, in the compositions of the invention may be capable of specifically binding to a coronavirus, such as SARS-CoV, in activated or inactivated/attenuated form. Methods for inactivating/attenuating viruses are well known in the art and include, but are not limited to, heat inactivation, inactivation by UV irradiation, inactivation by gamma irradiation. The binding molecules may also be capable of specifically binding to one or more fragments of the coronavirus including *inter alia* a preparation of one or more proteins and/or (poly)peptides derived from the coronavirus. Preferably, the fragment at least comprises an antigenic determinant recognised by the binding molecules of the invention. An "antigenic determinant" as used herein is a moiety, such as a coronavirus (such as SARS-CoV), (poly)peptide, protein, glycoprotein, analog or fragment thereof, that is capable of binding to a binding molecule of the invention with sufficiently high affinity to form a detectable antigen-binding molecule complex.

In an embodiment the binding molecules, *i.e.* immunoglobulins, are capable of specifically binding to surface accessible proteins of a coronavirus which include, but are not limited to, inner and outer membrane proteins, proteins adhering to the cell wall, and potential secreted proteins. Relevant proteins of SARS-CoV in that respect are *inter alia* the spike (S) protein, the membrane (matrix) protein, the (small) envelope protein, Orf 3, Orf 4, Orf 7, Orf 8, Orf 9, Orf 10 and Orf 14. The amino acid sequence of proteins and potential proteins of various known strains of coronaviruses, such as SARS-CoV, can be found in the EMBL-database and/or other databases. For instance the complete genome of the SARS coronavirus Urbani can be found in the

EMBL-database under accession number AY278741, the complete genome of the SARS coronavirus HSR 1 can be found under accession number AY323977, the complete genome of the SARS coronavirus Frankfurt 1 can be found under accession number AY291315 and the complete genome of the SARS coronavirus TOR2 can be found under accession number AY274119.

In an embodiment at least one of the binding molecules, *i.e.* immunoglobulins, in the compositions of the invention is capable of specifically binding to the S protein of SARS-CoV. The other binding molecule may bind to a receptor of SARS-CoV present on or associated with target cells. An example of such a receptor is the ACE-2 receptor (see Li *et al.*, 2003). In another embodiment all binding molecules in the compositions of the invention are capable of specifically binding to the S protein of SARS-CoV.

In yet another embodiment at least one of the binding molecules in the compositions of the invention is capable of specifically binding to the extracellular domain of the S protein of SARS-CoV. This domain consists of amino acids 15-1195 of the S protein. In a specific embodiment at least one binding molecule in the compositions of the invention is capable of specifically binding to amino acids 318-510 of the S protein of SARS-CoV. The neutralizing binding molecules, *i.e.* immunoglogulins, in the compositions of the invention may react with overlapping, competing epitopes, but preferably they react with different/distinct, non-competing epitopes of the coronavirus, such as SARS-CoV.

Another aspect of the invention are compositions comprising at least two binding molecules capable of specifically binding to a coronavirus, such as SARS-CoV, wherein the binding molecules are capable of reacting with different, non-competing epitopes of the coronavirus.

Preferably, the coronavirus is a human coronavirus, more preferably the coronavirus is SARS-CoV. Compositions comprising at least two binding molecules wherein each binding molecule binds to a different epitope or site on a virus are more suitable for preventing the escape of resistant variants of the virus compared to compositions comprising at least two binding molecules wherein each binding molecule binds to an overlapping epitope or site on the virus.

In a specific embodiment the different, non-competing epitopes recognised by the binding molecules, *i.e.* immunoglobulins, in the compositions of the invention are located on the S protein of SARS-CoV, particularly the extracellular domain of the S protein, more particularly within amino acids 318-510 of the S protein. In another aspect, at least one of the binding molecues, *i.e.* immunoglobulins, of the compositions of the invention is capable of reacting with amino acids 318-510 of the S protein of a human and an animal SARS-CoV. In another embodiment at least one of the binding molecues, *i.e.* immunoglobulins, of the compositions of the invention reacts with an epitope comprising the amino acid sequence of SEQ ID NO:128. The epitope may consist of 11, 11 to 15, 11 to 20, 11 to 25, 11 to 30, 11 to 35, 11 to 40, 11 to 45 or even more amino acids. In another aspect at least one of the binding molecules, *i.e.* immunoglobulins, in the compositions of the invention is capable of reacting with amino acids 318-510 of the S protein of a SARS-CoV, wherein the amino acid at position 479 is an amino acid other than asparagine, to a similar extent as with amino acids 318-510 of the S protein of a SARS-CoV, wherein the amino acid at position 479 is an asparagine. In other words, substitution of the amino acid at position 479 does not dramatically influence the binding of at least one of the

immunoglobulins in the compositions of the invention to amino acids 318-510 of the S protein of SARS-CoV. "To a similar extent" as defined above means that the binding molecule binds to amino acids 318-510 of the S protein of a SARS-CoV, wherein the amino acid at position 479 is an amino acid other than asparagines, in an amount of at least 60%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, preferably at least 100%, compared to the binding of the binding molecule to amino acids 318-510 of the S protein of a SARS-CoV, wherein the amino acid at position 479 is an asparagine. Binding can be measured by methods well known to a person skilled in the art such as for instance ELISA.

The binding molecules in the compositions according to the invention are preferably human binding molecules, *i.e.* immunoglobulins. They can be intact immunoglobulin molecules such as polyclonal or monoclonal antibodies, in particular human monoclonal antibodies, or the binding molecules can be antigen-binding fragments including, but not limited to, Fab, F(ab'), $F(ab')_2$, Fv, dAb, Fd, complementarity determining region (CDR) fragments, single-chain antibodies (scFv), bivalent single-chain antibodies, single-chain phage antibodies, diabodies, triabodies, tetrabodies, and (poly)peptides that contain at least a fragment of an immunoglobulin that is sufficient to confer specific antigen binding to the SARS-CoV or fragment thereof. The human binding molecules are preferably human monoclonal antibodies. The binding molecules in the compositions can be in non-isolated or isolated form. The compositions may further comprise at least one other therapeutic agent. Preferably, the therapeutic agent is useful in the prophylaxis and/or treatment of a condition resulting from a coronavirus such as for instance SARS-CoV.

Typically, binding molecules can bind to their binding partners, *i.e.* a coronavirus or fragments thereof, with an affinity constant (K_d-value) that is lower than $0.2*10^{-4}$ M, $1.0*10^{-5}$ M, $1.0*10^{-6}$ M, $1.0*10^{-7}$ M, preferably lower than $1.0*10^{-8}$ M, more preferably lower than $1.0*10^{-9}$ M, more preferably lower than $1.0*10^{-10}$ M, even more preferably lower than $1.0*10^{-11}$ M, and in particular lower than $1.0*10^{-12}$ M. The affinity constants can vary for antibody isotypes. For example, affinity binding for an IgM isotype refers to a binding affinity of at least about $1.0*10^{-7}$ M. Affinity constants can for instance be measured using surface plasmon resonance, *i.e.* an optical phenomenon that allows for the analysis of real-time biospecific interactions by detection of alterations in protein concentrations within a biosensor matrix, for example using the BIACORE system (Pharmacia Biosensor AB, Uppsala, Sweden).

The binding molecules may bind to a coronavirus in soluble form such as for instance in a sample or may bind to a coronavirus bound or attached to a carrier or substrate, *e.g.*, microtiter plates, membranes and beads, etc. Carriers or substrates may be made of glass, plastic (*e.g.*, polystyrene), polysaccharides, nylon, nitrocellulose, or teflon, etc. The surface of such supports may be solid or porous and of any convenient shape. Furthermore, the binding molecules may bind to a coronavirus in purified/isolated or non-purified/non-isolated form.

In a preferred embodiment, the binding molecules, i.e. immunoglobulins, of the compositions according to the invention comprise at least a CDR3 region, preferably a heavy chain CDR3 region, comprising the amino acid sequence of SEQ ID NO:1 or SEQ ID NO:2. In another embodiment the binding molecules comprising the heavy chain CDR3 region of SEQ ID

NO:1 further comprise a light chain CDR3 region comprising the amino acid sequence of SEQ ID NO:129. In another embodiment, the binding molecules comprising the heavy chain CDR3 region of SEQ ID NO:1 further comprise a heavy chain CDR1, heavy chain CDR2, light chain CDR1 and light chain CDR2 region comprising the amino acid sequence of SEQ ID NO:130, SEQ ID NO:131, SEQ ID NO:132, and SEQ ID NO:133, respectively. In another embodiment the binding molecules comprising the heavy chain CDR3 region of SEQ ID NO:2 further comprise a light chain CDR3 region comprising the amino acid sequence of SEQ ID NO:134. In another embodiment, the binding molecules comprising the heavy chain CDR3 region of SEQ ID NO:2 further comprise a heavy chain CDR1, heavy chain CDR2, light chain CDR1 and light chain CDR2 region comprising the amino acid sequence of SEQ ID NO:135, SEQ ID NO:136, SEQ ID NO:137, and SEQ ID NO:138, respectively. In yet another embodiment, the binding molecules according to the invention comprise a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:4 or SEQ ID NO:6. In an embodiment the binding molecules, *i.e.* immunoglobulins, in the compositions of the invention comprise at least one CDR region of a binding molecule comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:4 or SEQ ID NO:6. In another embodiment they comprise two, three, four, five or even all six CDR regions.

In a further embodiment, the binding molecules according to the invention comprise a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:4 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:8, or a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:6 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:10.

In a preferred embodiment the binding molecules having coronavirus, such as SARS-CoV, neutralizing activity are administered in IgG1 or IgA format.

In another aspect of the invention the compositions may comprise at least one functional variant of a binding molecule as defined herein. The compositions may also consist of only functional variants of binding molecules as herein described. Molecules are considered to be functional variants of a binding molecule, if the variants are capable of competing for specifically binding to a coronavirus, such as SARS-CoV, or a fragment thereof with the parent binding molecules. In other words, when the functional variants are still capable of binding to the coronavirus, such as SARS-CoV, or a fragment thereof. Preferably, the functional variants are capable of neutralizing coronavirus, such as SARS-CoV, infectivity and should together with the other binding molecule (or other functional variant) or other binding molecules (or other functional variants) form a composition exhibiting synergistic coronavirus, such as SARS-CoV, neutralizing activity. The neutralizing activity of a functional variant may either be higher or be lower compared to the parent binding molecules.

Functional variants include, but are not limited to, derivatives that are substantially similar in primary structural sequence, but which contain *e.g. in vitro* or *in vivo* modifications, chemical and/or biochemical, that are not found in the parent binding molecule. Such modifications are well known to the skilled artisan and include *inter alia* acetylation, acylation, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, glycosylation, methylation, pegylation, proteolytic processing, phosphorylation, and the like.

Alternatively, functional variants can be binding molecules as defined in the present invention comprising an amino acid sequence containing substitutions, insertions, deletions or combinations thereof of one or more amino acids compared to the amino acid sequences of the parent binding molecules. Furthermore, functional variants can comprise truncations of the amino acid sequence at either or both the amino or carboxy termini. Functional variants may have the same or different, either higher or lower, binding affinities compared to the parent binding molecule but are still capable of binding to a coronavirus, such as SARS-CoV, or a fragment thereof and preferably still capable of neutralizing coronavirus, such as SARS-CoV, infectivity. For instance, functional variants may have increased or decreased binding affinities for a coronavirus, such as SARS-CoV, or a fragment thereof compared to the parent binding molecules. Preferably, the amino acid sequences of the variable regions, including, but not limited to, framework regions, hypervariable regions, in particular the CDR3 regions, are modified. Generally, the light chain and the heavy chain variable regions comprise three hypervariable regions, comprising three CDRs, and more conserved regions, the so-called framework regions (FRs). The hypervariable regions comprise amino acid residues from CDRs and amino acid residues from hypervariable loops. Functional variants intended to fall within the scope of the present invention have at least about 50% to about 99%, preferably at least about 60% to about 99%, more preferably at least about 70% to about 99%, even more preferably at least about 80% to about 99%, most preferably at least about 90% to about 99%, in particular at least about 95% to about 99%, and in particluar at least about 97% to about 99% amino acid sequence homology with the parent binding molecules as defined herein. Computer

algorithms such as *inter alia* Gap or Bestfit known to a person skilled in the art can be used to optimally align amino acid sequences to be compared and to define similar or identical amino acid residues. Functional variants can be obtained by altering the parent binding molecules or parts thereof by general molecular biology methods known in the art including, but not limited to, error-prone PCR, oligonucleotide-directed mutagenesis and site-directed mutagenesis.

In yet a further aspect, the invention includes compositions comprising at least one immunoconjugate, *i.e.* a molecule comprising at least one binding molecule or functional variant thereof as defined herein and further comprising at least one tag. Also contemplated in the present invention are compositions consisting of immunoconjugates. The compositions may further comprise another molecule, such as a therapeutic agent or immunoconjugate having a different specificity. In a further embodiment, the immunoconjugates of the invention may comprise more than one tag. These tags can be the same or distinct from each other and can be joined/conjugated non-covalently to the binding molecules. The tag(s) can also be joined/conjugated directly to the binding molecules through covalent bonding. Alternatively, the tag(s) can be joined/conjugated to the binding molecules by means of one or more linking compounds. Techniques for conjugating tags to binding molecules are well known to the skilled artisan.

The tags of the immunoconjugates of the present invention may be therapeutic agents, but preferably they are detectable moieties/agents. Compositions comprising immunoconjugates comprising a detectable agent can be used diagnostically to, for example, assess if a subject has been infected with a coronavirus, such as SARS-CoV, or monitor the development or progression of a coronavirus, such as SARS-CoV, infection as

part of a clinical testing procedure to, *e.g.*, determine the efficacy of a given treatment regimen. However, they may also be used for other detection and/or analytical and/or diagnostic purposes. Detectable moieties/agents include, but are not limited to, enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, radioactive materials, positron emitting metals, and nonradioactive paramagnetic metal ions. The tags used to label the binding molecules for detection and/or analytical and/or diagnostic purposes depend on the specific detection/analysis/diagnosis techniques and/or methods used such as *inter alia* immunohistochemical staining of (tissue) samples, flow cytometric detection, scanning laser cytometric detection, fluorescent immunoassays, enzyme-linked immunosorbent assays (ELISA's), radioimmunoassays (RIA's), bioassays (*e.g.*, neutralisation assays), Western blotting applications, etc. For immunohistochemical staining of tissue samples preferred labels are enzymes that catalyze production and local deposition of a detectable product. Furthermore, the compositions of the invention can also be attached to solid supports, which are particularly useful for *in vitro* immunoassays or purification of a coronavirus or a fragment thereof. Such solid supports might be porous or nonporous, planar or nonplanar. The binding molecules of the present invention or functional fragments thereof can be fused to marker sequences, such as a peptide to facilitate purification. Alternatively, an antibody can be conjugated to a second antibody to form an antibody heteroconjugate. In another aspect the binding molecules of the invention may be conjugated/attached to one or more antigens. Preferably, these antigens are antigens which are recognised by the immune

system of a subject to which the binding molecule-antigen conjugate is administered.

Next to producing immunoconjugates chemically by conjugating, directly or indirectly via for instance a linker, the immunoconjugates can be produced as fusion proteins comprising the binding molecules of the invention and a suitable tag. Fusion proteins can be produced by methods known in the art such as, *e.g.*, recombinantly by constructing nucleic acid molecules comprising nucleotide sequences encoding the binding molecules in frame with nucleotide sequences encoding the suitable tag(s) and then expressing the nucleic acid molecules.

It is another aspect of the present invention to provide a nucleic acid molecule encoding at least a binding molecule or functional fragment thereof present in the compositions according to the invention. Such nucleic acid molecules can be used as intermediates for cloning purposes, *e.g.* in the process of affinity maturation described above. In a preferred embodiment, the nucleic acid molecules are isolated or purified.

The skilled man will appreciate that functional variants of these nucleic acid molecules are also intended to be a part of the present invention. Functional variants are nucleic acid sequences that can be directly translated, using the standard genetic code, to provide an amino acid sequence identical to that translated from the parent nucleic acid molecules.

Preferably, the nucleic acid molecules encode binding molecules comprising a CDR3 region, preferably a heavy chain CDR3 region, comprising the amino acid sequence of SEQ ID NO:1 or SEQ ID NO:2. In another embodiment the nucleic acid molecules of the invention encode binding molecules comprising a light chain CDR3 region comprising the amino acid sequence

of SEQ ID NO:129 or SEQ ID NO:134. In yet another embodiment the nucleic acid molecules of the invention encode binding molecules comprising a heavy chain CDR1 region comprising the amino acid sequence of SEQ ID NO:130 or SEQ ID NO:135; a heavy chain CDR2 region comprising the amino acid sequence of SEQ ID NO:131 or SEQ ID NO:136; a light chain CDR1 region comprising the amino acid sequence of SEQ ID NO:132 or SEQ ID NO:137; and/or a light chain CDR2 region comprising the amino acid sequence of SEQ ID NO:133 or SEQ ID NO:138.

Even more preferably, the nucleic acid molecules encode binding molecules comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:4 or SEQ ID NO:6.

In yet another embodiment, the nucleic acid molecules encode binding molecules comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:4 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:8 or they encode a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:6 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:10.

In a specific embodiment of the invention the nucleic acid molecules encoding the heavy chain variable region of the binding molecules of the invention comprise the nucleotide sequence of SEQ ID NO:3 or SEQ ID NO:5.

In yet another specific embodiment of the present invention, the nucleic acid molecules encoding the light chain variable region of the binding molecules of the invention comprise the nucleotide sequence of SEQ ID NO:7 or SEQ ID NO:9. The nucleic acid molecules of the invention may even contain the nucleotide sequences or parts thereof of the at

least two binding molecules present in the compostions of the invention.

It is another aspect of the invention to provide vectors, *i.e.* nucleic acid constructs, comprising one or more nucleic acid molecules according to the present invention. Vectors can be derived from plasmids such as *inter alia* F, Rl, RP1, Col, pBR322, TOL, Ti, etc; cosmids; phages such as lambda, lambdoid, M13, Mu, P1, P22, Q_{β}, T-even, T-odd, T2, T4, T7, etc; plant viruses; or animal viruses. Vectors can be used for cloning and/or for expression of the binding molecules of the invention and might even be used for gene therapy purposes. Vectors comprising one or more nucleic acid molecules according to the invention operably linked to one or more expression-regulating nucleic acid molecules are also covered by the present invention. The binding molecules present in the compositions of the invention may be expressed on separate vectors but may also be expressed on the same vector. The choice of the vector(s) is dependent on the recombinant procedures followed and the host used. Introduction of vectors in host cells can be effected by *inter alia* calcium phosphate transfection, virus infection, DEAE-dextran mediated transfection, lipofectamin transfection or electroporation. Vectors may be autonomously replicating or may replicate together with the chromosome into which they have been integrated. Preferably, the vectors contain one or more selection markers. The choice of the markers may depend on the host cells of choice, although this is not critical to the invention as is well known to persons skilled in the art. They include, but are not limited to, kanamycin, neomycin, puromycin, hygromycin, zeocin, thymidine kinase gene from Herpes simplex virus (HSV-TK), dihydrofolate reductase gene from mouse (dhfr). Vectors comprising one or more nucleic acid

molecules encoding the binding molecules as described above operably linked to one or more nucleic acid molecules encoding proteins or peptides that can be used to isolate the binding molecules are also covered by the invention. These proteins or peptides include, but are not limited to, glutathione-S-transferase, maltose binding protein, metal-binding polyhistidine, green fluorescent protein, luciferase and beta-galactosidase.

Hosts containing one or more copies of the vectors mentioned above are an additional subject of the present invention. Preferably, the hosts are host cells. Host cells include, but are not limited to, cells of mammalian, plant, insect, fungal or bacterial origin. Bacterial cells include, but are not limited to, cells from Gram positive bacteria such as several species of the genera *Bacillus*, *Streptomyces* and *Staphylococcus* or cells of Gram negative bacteria such as several species of the genera Escherichia, such as *E. coli*, and *Pseudomonas*. In the group of fungal cells preferably yeast cells are used. Expression in yeast can be achieved by using yeast strains such as *inter alia Pichia pastoris*, *Saccharomyces cerevisiae* and *Hansenula polymorpha*. Furthermore, insect cells such as cells from Drosophila and Sf9 can be used as host cells. Besides that, the host cells can be plant cells expression systems using mammalian cells such as Chinese Hamster Ovary (CHO) cells, COS cells, BHK cells or Bowes melanoma cells are preferred in the present invention. Mammalian cells provide expressed proteins with posttranslational modifications that are most similar to natural molecules of mammalian origin. Since the present invention deals with molecules that may have to be administered to humans, a completely human expression system would be particularly preferred. Therefore, even more

preferably, the host cells are human cells. Examples of human cells are *inter alia* HeLa, 911, AT1080, A549, 293 and HEK293T cells. Preferred mammalian cells are human retina cells such as 911 cells or the cell line deposited at the European Collection of Cell Cultures (ECACC), CAMR, Salisbury, Wiltshire SP4 OJG, Great Britain on 29 February 1996 under number 96022940 and marketed under the trademark PER.C6® (PER.C6 is a registered trademark of Crucell Holland B.V.). For the purposes of this application "PER.C6" refers to cells deposited under number 96022940 or ancestors, passages upstream or downstream as well as descendants from ancestors of deposited cells, as well as derivatives of any of the foregoing. In preferred embodiments, the human producer cells comprise at least a functional part of a nucleic acid sequence encoding an adenovirus E1 region in expressible format. In even more preferred embodiments, said host cells are derived from a human retina and immortalised with nucleic acids comprising adenoviral E1 sequences, such as the cell line deposited at the European Collection of Cell Cultures (ECACC), CAMR, Salisbury, Wiltshire SP4 OJG, Great Britain on 29 February 1996 under number 96022940 and marketed under the trademark PER.C6®, and derivatives thereof. Production of recombinant proteins in host cells can be performed according to methods well known in the art. The use of the cells marketed under the trademark PER.C6® as a production platform for proteins of interest has been described in WO 00/63403 the disclosure of which is incorporated herein by reference in its entirety.

Methods of producing binding molecules or functional variants are well known to the skilled artisan. One method comprises the steps of a) culturing a host as defined above under conditions conducive to the expression of the binding

molecules or functional variants, and b) optionally, recovering the expressed binding molecules or functional variants. The expressed binding molecules or functional variants thereof can be recovered from the cell free extract, but preferably they are recovered from the culture medium. Methods to recover proteins, such as binding molecules, from cell free extracts or culture medium are well known to the man skilled in the art. Binding molecules or functional variants thereof as obtainable by the above described method are also a part of the present invention. Alternatively, next to the expression in hosts, such as host cells, the binding molecules or functional variants thereof can be produced synthetically by conventional peptide synthesizers or in cell-free translation systems using RNA nucleic acid derived from DNA molecules according to the invention. Binding molecule or functional variants thereof as obtainable by the above described synthetic production methods or cell-free translation systems are also a part of the present invention. In yet another alternative embodiment, binding molecules according to the present invention, preferably human binding molecules specifically binding to a coronavirus, such as SARS-CoV, or a fragment thereof, may be generated by transgenic non-human mammals, such as for instance transgenic mice or rabbits, that express human immunoglobulin genes. Preferably, the transgenic non-human mammals have a genome comprising a human heavy chain transgene and a human light chain transgene encoding all or a portion of the human binding molecules as described above. The transgenic non-human mammals can be immunized with a purified or enriched preparation of a coronavirus, such as SARS-CoV, or a fragment thereof. Protocols for immunizing non-human mammals are well established in the art. See Using Antibodies: A Laboratory

Manual, Edited by: E. Harlow, D. Lane (1998), Cold Spring Harbor Laboratory, Cold Spring Harbor, New York and Current Protocols in Immunology, Edited by: J.E. Coligan, A.M. Kruisbeek, D.H. Margulies, E.M. Shevach, W. Strober (2001), John Wiley & Sons Inc., New York, the disclosures of which are incorporated herein by reference. Immunization protocols often include multiple immunizations, either with or without adjuvants such as Freund's complete adjuvant and Freund's incomplete adjuvant, but may also include naked DNA immunizations. In another embodiment, the human binding molecules are produced by B cells or plasma cells derived from the transgenic animals or human subjects that have been exposed to SARS-CoV. In yet another embodiment, the human binding molecules are produced by hybridomas which are prepared by fusion of B cells obtained from the above described transgenic non-human mammals or human subjects to immortalized cells. B cells, plasma cells and hybridomas as obtainable from the above described transgenic non-human mammals or human subjects and human binding molecules as obtainable from the above described transgenic non-human mammals or human subjects are also a part of the present invention.

Methods of identifying binding molecules, preferably human binding molecules such as human monoclonal antibodies or fragments thereof, or nucleic acid molecules encoding the binding molecules may comprise the steps of a) contacting a phage library of binding molecules, preferably human binding molecules, with a coronavirus, such as SARS-CoV, or a fragment thereof, b) selecting at least once for a phage binding to the coronavirus or the fragment thereof, and c) separating and recovering the phage binding to the coronavirus or the fragment thereof. The selection step may be performed by

contacting a phage library with a coronavirus which is inactivated. The coronavirus may be isolated or non-isolated, *e.g.* present in serum and/or blood of an infected individual. Alternatively, the selection step may be performed in the presence of a fragment of a coronavirus such as an extracellular part of the coronavirus (such as SARS-CoV), one or more proteins or (poly)peptides derived from a coronavirus, fusion proteins comprising these proteins or (poly)peptides, and the like. Phage display methods for identifying and obtaining binding molecules, *e.g.* antibodies, are by now well-established methods known by the person skilled in the art. They are *e.g.* described in US Patent Number 5,696,108; Burton and Barbas, 1994; de Kruif *et al.*, 1995b; and Phage Display: A Laboratory Manual. Edited by: CF Barbas, DR Burton, JK Scott and GJ Silverman (2001), Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York. All these references are herewith incorporated herein in their entirety. For the construction of phage display libraries, collections of human monoclonal antibody heavy and light chain variable region genes are expressed on the surface of bacteriophage, preferably filamentous bacteriophage, particles, in for example single-chain Fv (scFv) or in Fab format (see de Kruif *et al.*, 1995b). Large libraries of antibody fragment-expressing phages typically contain more than $1.0*10^9$ antibody specificities and may be assembled from the immunoglobulin V regions expressed in the B lymphocytes of non-immunized or immunized individuals. In a specific embodiment the phage library of binding molecules, preferably scFv phage library, is prepared from RNA isolated from cells obtained from a subject that has been vaccinated or exposed to a coronavirus, such as SARS-CoV. RNA can be isolated from *inter alia* bone marrow or peripheral blood, preferably peripheral blood

lymphocytes. The subject can be an animal vaccinated or exposed to a coronavirus, but is preferably a human subject which has been vaccinated or has been exposed to a coronavirus. Preferably, the human subject has recovered from the coronavirus.

Alternatively, phage display libraries may be constructed from immunoglobulin variable regions that have been partially assembled *in vitro* to introduce additional antibody diversity in the library (semi-synthetic libraries). For example, *in vitro* assembled variable regions contain stretches of synthetically produced, randomized or partially randomized DNA in those regions of the molecules that are important for antibody specificity, *e.g.* CDR regions. Coronavirus specific phage antibodies can be selected from libraries by immobilising a coronavirus (in inactivated or active form) or target antigens such as antigens from a coronavirus on a solid phase and subsequently exposing the coronavirus (in inactivated or active form) or target antigens to a phage library to allow binding of phages expressing antibody fragments specific for the solid phase-bound antigen(s). Non-bound phages are removed by washing and bound phages eluted from the solid phase for infection of *Escherichia coli* (*E.coli*) bacteria and subsequent propagation. Multiple rounds of selection and propagation are usually required to sufficiently enrich for phages binding specifically to the coronavirus (in inactivated or active form) or target antigen(s). If desired, before exposing the phage library to a coronavirus (in inactivated or active form) or target antigens the phage library can first be subtracted by exposing the phage library to non-target antigens bound to a solid phase. Phages may also be selected for binding to complex antigens such as complex mixtures of coronavirus proteins or

(poly)peptides or host cells expressing one or more proteins or (poly)peptides of a coronavirus. Antigen specific phage antibodies can be selected from the library by incubating a solid phase with bound thereon a preparation of an inactivated coronavirus with the phage antibody library to let for example the scFv or Fab part of the phage bind to the proteins/polypeptides of the coronavirus preparation. After incubation and several washes to remove unbound and loosely attached phages, the phages that have bound with their scFv or Fab part to the preparation are eluted and used to infect *Escherichia coli* to allow amplification of the new specificity. Generally, one or more selection rounds are required to separate the phages of interest from the large excess of non-binding phages. Alternatively, known proteins or (poly)peptides of the coronavirus can be expressed in host cells and these cells can be used for selection of phage antibodies specific for the proteins or (poly)peptides. A phage display method using these host cells can be extended and improved by subtracting non-relevant binders during screening by addition of an excess of host cells comprising no target molecules or non-target molecules that are similar, but not identical, to the target, and thereby strongly enhance the chance of finding relevant binding molecules (This process is referred to as the Mabstract™ process. Mabstract™ is a pending trademark application of Crucell Holland B.V., see also US Patent Number 6,265,150 which is incorporated herein by reference). An example of a coronavirus against which binding molecules can be found using the above described method of identification is SARS-CoV.

A method of obtaining a binding molecule, preferably a human binding molecule or a nucleic acid molecule encoding such a binding molecule may comprise the steps of a)

performing the above described method of identifying binding molecules, preferably human binding molecules such as human monoclonal antibodies or fragments thereof, or nucleic acid molecules encoding the binding molecules, and b) isolating from the recovered phage the human binding molecule and/or the nucleic acid encoding the human binding molecule. Once a new monoclonal phage antibody has been established or identified with the above mentioned method of identifying binding molecules or nucleic acid molecules encoding the binding molecules, the DNA encoding the scFv or Fab can be isolated from the bacteria or phages and combined with standard molecular biological techniques to make constructs encoding bivalent scFvs or complete human immunoglobulins of a desired specificity (*e.g.* IgG, IgA or IgM). These constructs can be transfected into suitable cell lines and complete human monoclonal antibodies can be produced (see Huls *et al.*, 1999; Boel *et al.*, 2000).

In addition to the at least two binding molecule, the compositions of the invention may comprise *inter alia* stabilising molecules, such as albumin or polyethylene glycol, or salts. Preferably, the salts used are salts that retain the desired biological activity of the binding molecules and do not impart any undesired toxicological effects. Examples of such salts include, but are not limited to, acid addition salts and base addition salts. Acid addition salts include, but are not limited to, those derived from nontoxic inorganic acids, such as hydrochloric, nitric, phosphoric, sulfuric, hydrobromic, hydroiodic, phosphorous and the like, as well as from nontoxic organic acids such as aliphatic mono- and dicarboxylic acids, phenyl-substituted alkanoic acids, hydroxy alkanoic acids, aromatic acids, aliphatic and aromatic sulfonic acids and the like. Base addition salts include, but

are not limited to, those derived from alkaline earth metals, such as sodium, potassium, magnesium, calcium and the like, as well as from nontoxic organic amines, such as N,N'-dibenzylethylenediamine, N-methylglucamine, chloroprocaine, choline, diethanolamine, ethylenediamine, procaine and the like. If necessary, the binding molecules of the invention may be coated in or on a material to protect them from the action of acids or other natural or non-natural conditions that may inactivate the binding molecules.

In yet a further aspect, the invention provides compositions comprising at least two nucleic acid molecule encoding binding molecules as defined in the present invention. The compositions may comprise aqueous solutions such as aqueous solutions containing salts (*e.g.*, NaCl or salsts as described above), detergents (*e.g.*, SDS) and/or other suitable components.

Furthermore, the present invention pertains to pharmaceutical compositions comprising a composition according to the invention. The pharmaceutical composition of the invention further comprises at least one pharmaceutically acceptable excipient.

A pharmaceutical composition according to the invention can further comprise at least one other therapeutic, prophylactic and/or diagnostic agent. Preferably, said further therapeutic and/or prophylactic agents are agents capable of preventing and/or treating an infection and/or a condition resulting from a coronavirus, such as SARS-CoV. Therapeutic and/or prophylactic agents include, but are not limited to, anti-viral agents. Such agents can be binding molecules, small molecules, organic or inorganic compounds, enzymes, polynucleotide sequences etc.

Examples of anti-viral agents are well known to the skilled artisan. Agents that are currently used to treat patients infected with for instance SARS-CoV are interferon-alpha, steroids and potential replicase inhibitors. Furthermore, patients infected with SARS-CoV are currently treated by transfusion of serum produced from blood donated by recovering/recovered SARS patients who have produced antibodies after being exposed to the virus. Agents capable of preventing and/or treating an infection with SARS-CoV or other coronavirus and/or a condition resulting from SARS-CoV or other coronavirus that are in the experimental phase might also be used as other therapeutic and/or prophylactic agents useful in the present invention.

The pharmaceutical compositions of the invention can be tested in suitable animal model systems prior to use in humans. Such animal model systems include, but are not limited to, ferrets, mice, rats, chicken, cows, monkeys, pigs, dogs, rabbits, etc.

Typically, pharmaceutical compositions must be sterile and stable under the conditions of manufacture and storage. The compositions of the present invention can be in powder form for reconstitution in the appropriate pharmaceutically acceptable excipient before or at the time of delivery. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying (lyophilization) that yield a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Alternatively, the compositions of the present invention can be in solution and the appropriate pharmaceutically acceptable excipient can be added and/or mixed before or at

the time of delivery to provide a unit dosage injectable form. Preferably, the pharmaceutically acceptable excipient used in the present invention is suitable to high drug concentration, can maintain proper fluidity and, if necessary, can delay absorption.

The choice of the optimal route of administration of the pharmaceutical compositions will be influenced by several factors including the physico-chemical properties of the active molecules within the compositions, the urgency of the clinical situation and the relationship of the plasma concentrations of the active molecules to the desired therapeutic effect. For instance, if necessary, the compositions of the invention can be prepared with carriers that will protect them against rapid release, such as a controlled release formulation, including implants, transdermal patches, and microencapsulated delivery systems. Biodegradable, biocompatible polymers can *inter alia* be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Furthermore, it may be necessary to coat the compositions with, or co-administer the compositions with, a material or compound that prevents the inactivation of the binding molecules in the compositions. For example, the binding molecules of the compositions may be administered to a subject in an appropriate carrier, for example, liposomes, or a diluent.

The routes of administration can be divided into two main categories, oral and parenteral administration. These two categories include several routes of administration well known to the skilled person. The preferred administration route is intravenous, particularly preferred is intramuscular.

Oral dosage forms can be formulated in several formulations and may contain pharmaceutically acceptable excipients including, but not limited to, inert diluents, granulating and disintegrating agents, binding agents, lubricating agents, preservatives, colouring, flavouring or sweetening agents, vegetable oils, mineral oils, wetting agents, and thickening agents.

The pharmaceutical compositions of the present invention can also be formulated for parenteral administration. Formulations for parenteral administration can be *inter alia* in the form of aqueous or non-aqueous isotonic sterile non-toxic injection or infusion solutions or suspensions. The solutions or suspensions may comprise agents that are non-toxic to recipients at the dosages and concentrations employed. Such agents are well known to the skilled artisan and include 1,3-butanediol, Ringer's solution, Hank's solution, isotonic sodium chloride solution, oils or fatty acids, local anaesthetic agents, preservatives, buffers, viscosity or solubility increasing agents, water-soluble antioxidants, oil-soluble antioxidants, and metal chelating agents.

In a further aspect, the pharmaceutical compositions of the invention can be used as a medicament. So, a method of treatment and/or prevention of a coronavirus infection using the pharmaceutical compositions of the invention is another part of the present invention. The (pharmaceutical) compositions of the invention can *inter alia* be used in the diagnosis, prophylaxis, treatment, or combination thereof, of one or more conditions resulting from a coronavirus. They are suitable for treatment of yet untreated patients suffering from a condition resulting from a coronavirus and patients who have been or are treated from a condition resulting from a

coronavirus. They protect against further infection by a coronavirus and/or will retard the onset or progress of the symptoms associated with a coronavirus. They may even be used in the prophylaxis of conditions resulting from a coronavirus in for instance people exposed to the coronavirus such as hospital personnel taking care of suspected patients. Preferably, the (pharmaceutical) compositions can be used in a method to detect, prevent, and/or treat a human coronavirus, such as SARS-CoV, infection.

The above mentioned compositions and pharmaceutical compositions may be employed in conjunction with other molecules useful in diagnosis, prophylaxis and/or treatment of a coronavirus infection. They can be used *in vitro*, *ex vivo* or *in vivo*. For instance, the pharmaceutical compositions of the invention can be co-administered with a vaccine against a coronavirus, such as SARS-CoV. Alternatively, the vaccine may also be administered before or after administration of the pharmaceutical compositions of the invention. Administration of the pharmaceutical compositions of the invention with a vaccine might be suitable for postexposure prophylaxis and might also decrease possible side effects of a live-attenuated vaccine in immunocompromised recipients.

The binding molecules are typically formulated in the compositions and pharmaceutical compositions of the invention in a therapeutically or diagnostically effective amount. Dosage regimens can be adjusted to provide the optimum desired response (*e.g.*, a therapeutic response). A suitable dosage range may for instance be 0.05-100 mg/kg body weight, preferably 0.1-15 mg/kg body weight.

Typically, the molar ratio of the two binding molecules in the compositions and pharmaceutical compositions of the invention may vary from 1:100 to 100:1, preferably from 1:50

to 50:1, more preferably from 1:25 to 25:1, particularly 1:10 to 10:1, and more particularly 1:5 to 5:1. Furthermore, for example, a single bolus may be administered, several divided doses may be administered over time or the dose may be proportionally reduced or increased as indicated by the exigencies of the therapeutic situation. The molecules and compositions according to the present invention are preferably sterile. Methods to render these molecules and compositions sterile are well known in the art. The other molecules useful in diagnosis, prophylaxis and/or treatment can be administered in a similar dosage regimen as proposed for the binding molecules of the invention. If the other molecules are administered separately, they may be adminstered to a patient prior to (*e.g.*, 2 minutes, 5 minutes, 10 minutes, 15 minutes, 30 minutes, 45 minutes, 60 minutes, 2 hours, 4 hours, 6 hours, 8 hours, 10 hours, 12 hours, 14 hours, 16 hours, 18 hours, 20 hours, 22 hours, 24 hours, 2 days, 3 days, 4 days, 5 days, 7 days, 2 weeks, 4 weeks or 6 weeks before), concomitantly with, or subsequent to (*e.g.*, 2 minutes, 5 minutes, 10 minutes, 15 minutes, 30 minutes, 45 minutes, 60 minutes, 2 hours, 4 hours, 6 hours, 8 hours, 10 hours, 12 hours, 14 hours, 16 hours, 18 hours, 20 hours, 22 hours, 24 hours, 2 days, 3 days, 4 days, 5 days, 7 days, 2 weeks, 4 weeks or 6 weeks after) the administration of one or more of the binding molecules or pharmaceutical compositions of the invention. The exact dosing regimen is usually sorted out during clinical trials in human patients.

Human binding molecules and pharmaceutical compositions comprising the human binding molecules are particularly useful, and often preferred, when to be administered to human beings as *in vivo* therapeutic agents, since recipient immune response to the administered antibody will often be

substantially less than that occasioned by administration of a monoclonal murine, chimeric or humanised binding molecule.

In another aspect, the invention concerns the use of (pharmaceutical) compositions according to the invention in the preparation of a medicament for the diagnosis, prophylaxis, treatment, or combination thereof, of a condition resulting from a coronavirus. Preferably, the coronavirus is a human coronavirus such as SARS-CoV.

Next to that, kits comprising at least one composition according to the invention or at least one pharmaceutical composition according to the invention are also a part of the present invention. Optionally, the above described components of the kits of the invention are packed in suitable containers and labeled for diagnosis, prophylaxis and/or treatment of the indicated conditions. The binding molecules in the (pharmaceutical) compositions may be packaged individually. The above-mentioned components may be stored in unit or multi-dose containers, for example, sealed ampules, vials, bottles, syringes, and test tubes, as an aqueous, preferably sterile, solution or as a lyophilized, preferably sterile, formulation for reconstitution. The containers may be formed from a variety of materials and may have a sterile access port (for example the container may be an intravenous solution bag or a vial having a stopper pierceable by a hypodermic injection needle). The kit may further comprise more containers comprising a pharmaceutically acceptable buffer. It may further include other materials desirable from a commercial and user standpoint, including other buffers, diluents, filters, needles, syringes, culture medium for one or more of the suitable hosts. Associated with the kits can be instructions customarily included in commercial packages of therapeutic, prophylactic or diagnostic products, that contain

information about for example the indications, usage, dosage, manufacture, administration, contraindications and/or warnings concerning the use of such therapeutic, prophylactic or diagnostic products.

The invention further pertains to a method of detecting a SARS-CoV in a sample, wherein the method comprises the steps of a) contacting a sample with a diagnostically effective amount of compositions or pharmaceutical compositions according to the invention, and b) determining whether the compositions or pharmaceutical compositions specifically bind to a molecule of the sample. The sample may be a biological sample including, but not limited to blood, serum, urine, tissue or other biological material from (potentially) infected subjects, or a nonbiological sample such as water, drink, etc. The (potentially) infected subjects may be human subjects, but also animals that are suspected as carriers of a coronavirus, such as SARS-CoV, might be tested for the presence of the coronavirus using the compositions or pharmaceutical compositions. The sample may first be manipulated to make it more suitable for the method of detection. Manipulation mean *inter alia* treating the sample suspected to contain and/or containing the coronavirus in such a way that the coronavirus will disintigrate into antigenic components such as proteins, (poly)peptides or other antigenic fragments. Preferably, the compositions or pharmaceutical compositions are contacted with the sample under conditions which allow the formation of an immunological complex between the binding molecules in the compositions or pharmaceutical compositions and the coronavirus or antigenic components thereof that may be present in the sample. The formation of an immunological complex, if any, indicating the presence of the coronavirus in the sample, is then detected and measured by

suitable means. Such methods include, *inter alia*, homogeneous and heterogeneous binding immunoassays, such as radioimmunoassays (RIA), ELISA, immunofluorescence, immunohistochemistry, FACS, BIACORE and Western blot analyses.

Preferred assay techniques, especially for large-scale clinical screening of patient sera and blood and blood-derived products are ELISA and Western blot techniques. ELISA tests are particularly preferred and well known to persons skilled in the art.

In a further aspect, the invention provides a method of screening a binding molecule or a functional variant of a binding molecule for specific binding to a different, non-overlapping epitope of a coronavirus such as SARS-CoV as the epitope bound by a binding molecule or functional variant of the invention, wherein the method comprises the steps of a) contacting a binding molecule or a functional variant to be screened, a binding molecule or functional variant of the invention and a coronavirus or fragment thereof, b) measure if the binding molecule or functional variant to be screened is capable of competing for specifically binding to the coronavirus or fragment thereof with the binding molecule or functional variant of the invention. If the binding molecule or functional variant to be screened is not capable of competing for specifically binding to the coronavirus or fragment thereof with the binding molecule or functional variant of the invention, it most likely binds to a different, non-overlapping epitope. In a further step it may be determined if the screened binding molecules that bind to a different, non-overlapping epitope compared to the binding molecules of the invention have coronavirus neutralizing activity. In yet a further step it can be determined if the screened binding molecules that bind to a different, non-

overlapping epitope compared to the binding molecules of the invention and have coronavirus neutralizing activity form together with the binding molecules of the invention a composition exhibiting synergistic coronavirus neutralizing activity. Assays to screen for non-competing binding molecules and measure (synergistic) neutralizing activity are well known to the skilled person.

EXAMPLES

To illustrate the invention, the following examples are provided. The examples are not intended to limit the scope of the invention in any way.

Example 1

Construction of a scFv phage display library using peripheral blood lymphocytes of a patient having been exposed to SARS-CoV

Lymphocytes were obtained from a patient recovered from SARS-CoV (see Rickerts *et al.* 2003) and frozen in liquid nitrogen. The lymphocytes were quickly thawed in a 37°C water bath and transferred to wet-ice. The lymphocytes were diluted with cold DMEM culture medium to a final volume of 50 ml in a 50 ml tube and centrifuged for 5 minutes at 350xg. The obtained cell pellet was suspended in 5 ml DMEM and cell density was determined microscopically using trypan-blue exclusion to visualize dead cells. All cells (~$5x10^6$) were spun again for 5 minutes at 350xg, decanted and suspended in residual fluid (DMEM). Total RNA was prepared from these cells using organic phase separation (TRIZOL™) and subsequent ethanol precipitation. The obtained RNA was dissolved in DEPC treated ultrapure water and the concentration was determined by OD 260 nm measurement. Thereafter, the RNA was diluted to a concentration of 100 ng/µl. Next, 1 µg of RNA was converted

into cDNA as follows: To 10 µl total RNA, 13 µl DEPC treated ultrapure water and 1 µl random hexamers (500 ng/µl) were added and the obtained mixture was heated at 65°C for 5 minutes and quickly cooled on wet-ice. Then, 8 µl 5X First-Strand buffer, 2 µl dNTP (10 mM each), 2 µl DTT (0.1 M), 2 µl Rnase-inhibitor (40 U/µl) and 2 µl Superscript™III MMLV reverse transcriptase (200 U/µl) were added to the mixture, incubated at room temperature for 5 minutes and incubated for 1 hour at 50°C. The reaction was terminated by heat inactivation, *i.e.* by incubating the mixture for 15 minutes at 75°C.

The obtained cDNA products were diluted to a final volume of 200 µl with DEPC treated ultrapure water. The OD 260 nm of a 50 times diluted solution (in 10 mM Tris buffer) of the dilution of the obtained cDNA products gave a value of 0.1.

5 to 10 µl of the diluted cDNA products were used as template for PCR amplification of the immunoglobulin gamma heavy chain family and kappa or lambda light chain sequences using specific oligonucleotide primers (see Tables 2-9). PCR reaction mixtures contained, besides the diluted cDNA products, 25 pmol sense primer and 25 pmol anti-sense primer in a final volume of 50 µl of 20 mM Tris-HCl (pH 8.4), 50 mM KCl, 2.5 mM $MgCl_2$, 250 µM dNTPs and 1.25 units Taq polymerase. In a heated-lid thermal cycler having a temperature of 96°C, the mixtures obtained were quickly melted for 2 minutes, followed by 30 amplification cycles of: 30 seconds at 96°C, 30 seconds at 60°C and 60 seconds at 72°C. In a first round amplification, each of nine sense directed primers (see Table 2; covering all families of heavy chain variable regions) was combined with an IgG specific constant region anti-sense primer called HuCIgG 5′-GTC CAC CTT GGT GTT GCT GGG CTT-3′ (SEQ ID NO:87) yielding nine products of about 650 basepairs.

These products were purified on a 2% agarose gel and isolated from the gel using Qiagen gel-extraction columns. 1/10 of each of the isolated products was used in an identical PCR reaction as described above using the same nine sense primers (covering all families of heavy chain variable regions), whereby each sense primer was combined with one of the four J-region specific anti-sense primers (see Table 3). This resulted in 36 products of approximately 350 basepairs. The products obtained were purified on a 2% agarose gel and isolated from the gel using Qiagen gel-extraction columns. In a third round, 1/10 of each of the isolated products was subjected to re-amplification with the same set of primers as in the second round with the proviso that the primers used were extended with restriction sites (see Table 4) to enable directed cloning in the phage display vector pDV-C05 (see SEQ ID NO:88). This resulted again in 36 products. These products were pooled per used (VH) sense primer into nine fractions. In the next step, 2.5 µg of pooled fraction and 100 µg pDV-C05 vector were digested with NcoI and XhoI and purified by gel. Thereafter, a ligation was performed overnight at 16°C as follows. To 500 ng pDV-C05 vector 70 ng pooled fraction was added in a total volume of 50 µl ligation mix containing 50 mM Tris-HCl (pH 7.5), 10 mM $MgCl_2$, 10 mM DTT, 1 mM ATP, 25 µg/ml BSA and 2.5 µl T4 DNA Ligase (400 u/µl). This procedure was followed for each pooled fraction. The ligation mixes were purified by phenol/chloroform, followed by a chloroform extraction and ethanol precipitation, methods well known to the skilled artisan. The DNA obtained was dissolved in 50 µl ultrapure water and per ligation mix two times 2.5 µl aliquots were electroporated into 40 µl of TG1 competent *E. coli* bacteria according to the manufacturer's protocol (Stratagene). Transformants were grown overnight at 37°C in a

total of 27 dishes (three dishes per pooled fraction; dimension of dish: 240 mm x 240 mm) containing 2TY agar supplemented with 50 µg/ml ampicillin and 4.5% glucose. A (sub)library of variable heavy chain regions was obtained by scraping the transformants from the agar plates. This (sub)library was directly used for plasmid DNA preparation using a Qiagen™ kit.

The light chain immunoglobulin sequences were amplified from the same cDNA preparation in a similar three round PCR procedure and identical reaction parameters as described above for the heavy chain regions with the proviso that the primers depicted in Tables 5-9 were used. The first amplification was performed using a set of seventeen light chain variable region sense primers (eleven for the lambda light chain (see Table 5) and six for the kappa light chain (see Table 6)) each combined with an anti-sense primer recognizing the C-kappa called HuCk 5′-ACACTCTCCCCTGTTGAAGCTCTT-3′ (see SEQ ID NO:89) or C-lambda constant region HuCλ2 5′-TGAACATTCTGTAGGGGCCACTG-3′ (see SEQ ID NO:90) or HuCλ7 5′-AGAGCATTCTGCAGGGGCCACTG-3′ (see SEQ ID NO:91) (the HuCλ2 and HuCλ7 anti-sense primers were mixed to equimolarity before use), yielding 17 products of about 600 basepairs. These products were purified on a 2% agarose gel and isolated from the gel using Qiagen gel-extraction columns. 1/10 of each of the isolated products was used in an identical PCR reaction as described above using the same seventeen sense primers, whereby each lambda light chain sense primer was combined with one of the three Jlambda-region specific anti-sense primers (see Table 7) and each kappa light chain sense primer was combined with one of the five Jkappa-region specific anti-sense primers (see Table 8). This resulted in 63 products of approximately 350 basepairs. The products obtained were purified on a 2% agarose gel and isolated from the gel

using Qiagen gel-extraction columns. In a third round, 1/10 of each of the isolated products was subjected to re-amplification with the same set of primers as in the second round with the proviso that the primers used were extended with restriction sites (see Table 9) to enable directed cloning in the heavy chain (sub)library vector. This resulted again in 63 products. These products were pooled to a total of 10 fractions. This number of fractions was chosen to maintain the natural distribution of the different light chain families within the library and to over or under represent certain families. The number of alleles within a family was used to determine the percentage of representation within a library (see Table 10). Next, the fractions were digested with SalI and NotI and ligated in the heavy chain (sub)library vector, which was cut with the same restriction enzymes, using the same ligation procedure and volumes as described above for the heavy chain (sub)library. Ligation purification and subsequent transformation of the resulting definitive library was also performed as described above for the heavy chain (sub)library. The transformants were grown in 30 dishes (three dishes per pooled fraction; dimension of dish: 240 mm x 240 mm) containing 2TY agar supplemented with 50 µg/ml ampicillin and 4.5% glucose. All bacteria were harvested in 2TY culture medium containing 50 µg/ml ampicillin and 4.5% glucose, mixed with glycerol to 15% (v/v) and frozen in 1.5 ml aliquots at -80°C.

Example 2

Selection of phage carrying single-chain Fv fragments specifically recognizing SARS-CoV

Antibody fragments were selected using antibody phage display libraries and technology, essentially as described in

US patent 6,265,150 and in WO 98/15833, both of which are incorporated herein in their entirety. All procedures were performed at room temperature unless stated otherwise. An inactivated SARS-CoV preparation (Frankfurt 1 strain) was prepared as follows. Medium from Vero cells which were infected with SARS-CoV strain Frankfurt 1 was harvested as soon as cyotopathic effect (CPE) was observed. Cell debris was removed by centrifugation of the harvested medium for 15 minutes at 3000 rpm. The obtained supernatant was collected, spun again for 15 minutes at 3000 rpm and transferred to a clean tube. Subsequently, ultracentrifuge tubes were filled with 10 ml sterile 25% glycerol in PBS. 20 ml of the cleared supernatant was gently applied on the glycerol cushion and the tubes were spun for 2 hours at 20,000 rpm at 4°C. The supernatant was discarded and the virus pellets were resuspended in 1 ml TNE buffer (10 mM Tris-HCl pH 7.4, 1 mM EDTA, 200 mM NaCl) and stored at -80°C. Next, the resuspended virus pellets were gamma-irradiated at 45kGy on dry ice. Subsequently, they were tested for the absence of infectivity in cell culture. If absence of infectivity was established, the thus obtained inactivated SARS-CoV preparation was used for selection of single-chain phage antibodies specifically binding to SARS-CoV.

The inactivated virus preparation and heat-inactivated fetal bovine serum (FBS) were coated overnight at 4°C onto the surface of separate Maxisorp™ plastic tubes (Nunc). The tubes were blocked for two hours in 3 ml PBS containing 2% FBS and 2% fat free milk powder (2% PBS-FM). After two hours the FBS-coated tube was emptied and washed three times with PBS. To this tube, 500 µl (approximately 10^{13} cfu) of a phage display library expressing single-chain Fv fragments (scFvs) essentially prepared as described by De Kruif *et al.* (1995a)

and references therein (which are incorporated herein in their entirety), 500 µl 4% PBS-FM and 2 ml 2% PBS-FM were added. The tube was sealed and rotated slowly at room temperature for two hours. Subsequently, the obtained blocked phage library (3 ml) was transferred to a SARS-CoV preparation-coated tube that had been washed three times with PBS. Tween-20 was added to a final concentration of 0.05% and binding was allowed to proceed for two hours on a slowly rotating wheel at room temperature or at 37°C. The tube was emptied and washed ten times with PBS containing 0.05% Tween-20, followed by washing ten times with PBS. 1 ml glycine-HCL (0.05 M, pH 2.2) was added to elute bound phages, and the tube was rotated slowly for ten minutes. For neutralisation purposes, the eluted phages were added to 500 µl 1 M Tris-HCl pH 7.4. To this mixture, 5 ml of exponentially growing XL-1 blue bacterial culture was added. The obtained culture was incubated for thirty minutes at 37°C without shaking. Then, the bacteria were plated on TYE agar plates containing ampicillin, tetracycline and glucose. After overnight incubation of the plates at 37°C, the colonies were scraped from the plates and used to prepare an enriched phage library, essentially as described by De Kruif *et al.* (1995a) and WO 02/103012 (both are incorporated by reference herein). Briefly, scraped bacteria were used to inoculate 2TY medium containing ampicillin, tetracycline and glucose and grown at a temperature of 37°C to an OD 600 nm of ~0.3. CT or VCSM13 helper phages were added and allowed to infect the bacteria after which the medium was changed to 2TY containing ampicillin, tetracycline and kanamycin. Incubation was continued overnight at 30°C. The next day, the bacteria were removed from the 2TY medium by centrifugation after which the phages in the obtained supernatant were precipitated using

polyethylene glycol 6000/NaCl. Finally, the phages were dissolved in a small volume of PBS containing 1% BSA, filter-sterilized and used for a next round of selection. The selection/re-infection procedure was performed two or three times. After each round of selection, individual *E.coli* colonies were used to prepare monoclonal phage antibodies. Essentially, individual colonies were grown to log-phase and infected with VCSM13 helper phages after which phage antibody production was allowed to proceed overnight. Phage antibody containing supernatants were tested in ELISA for binding activity to the SARS-CoV preparation which was coated to 96-well plates. In the above-described selection, the phage antibody called SC03-014 was obtained. ScFvs of the phage antibody SC03-014 were produced as described before in De Kruif *et al.* (1995a and 1995b) and references therein (which are incorporated herein in their entirety). The buffer of the scFvs was adjusted to 1 x PBS.

Additionally, antibody fragments were selected from the immune phage display library expressing single chain Fv fragments (scFvs) (see Example 1 for the construction of this library) essentially as described *supra*. For the selection described below an UV-inactivated SARS-CoV preparation was used. UV-inactivated SARS-CoV (Frankfurt 1 strain) was prepared as follows. Medium from Vero cells which were infected with O.I. moi (multiplicity of infection) SARS-CoV strain Frankfurt 1 was harvested as soon as cytopathic effect (CPE) was observed. Cells were once frozen at -20°C and thawed. Cell debris was removed by centrifugation of the harvested medium for 15 minutes at 3000 rpm. The obtained supernatant was collected, spun again for 15 minutes at 3000 rpm and transferred to a clean tube. Subsequently, ultracentrifuge tubes were filled with 10 ml sterile 25% v/v

glycerol in PBS. 20 ml of the cleared supernatant was gently applied on the glycerol cushion and the tubes were spun for 2 hours at 20,000 rpm at 4°C in a Beckman SW28 rotor. The supernatant was discarded and the virus pellets were resuspended in 1 ml TNE buffer (10 mM Tris-HCl pH 7.4, 1 mM EDTA, 200 mM NaCl) and stored at -80°C. Next, the resuspended virus pellets were UV-irradiated at 4°C for 15 minutes (UV-B radiation 280-350 nm; λmax 306 nm). Subsequently, they were tested for the absence of infectivity in cell culture. If absence of infectivity was established, the thus obtained inactivated SARS-CoV preparations were used for further experiments.

In contrast to the selections described *supra* no pre-subtraction using heat-inactivated fetal bovine serum coated Maxisorp™ tubes (Nunc) was performed. To the SARS-CoV coated tubes, 500 µl (approximately 10^{13} cfu) of the immune phage display library expressing single chain Fv fragments (scFvs), one volume of 4% PBS-FM and Tween-20 to a final concentration of 0.05% was added.

For the immune phage display library selections which consisted of a single selection round only, binding was allowed to proceed at 37°C or room temperature on a slowly rotating wheel at 37°C followed by an incubation of 30 minutes without agitation. The following selections and washes were performed: incubation at 37°C, washing 5 times with PBS containing 0.05% Tween-20 (PBST) and 5 times with PBS; incubation at 37°C, washing 10 times with PBST and 10 times with PBS; incubation at room temperature, washing 10 times with PBST and 10 times with PBS. Bound phages were eluted and processed as described above. Phages derived from individual colonies were tested in ELISA for binding activity to SARS-CoV coated to 96-well plates. In the selections from the immune

phage display library the phage antibody called SC03-022 was obtained.

Example 3

Validation of the SARS-CoV specific single-chain phage antibodies

Selected single-chain phage antibodies that were obtained in the screens described above, were validated in ELISA for specificity, *i.e.* binding to the UV-inactivated SARS-CoV preparation prepared as described *supra*. Additionally, the single-chain phage antibodies were also tested for binding to 10% FBS. For this purpose, the UV-inactivated SARS-CoV preparation or 10% FBS preparation was coated to Maxisorp™ ELISA plates. After coating, the plates were blocked in 2% PBS-FM. The selected single-chain phage antibodies were incubated in an equal volume of 4% PBS-FM to obtain blocked phage antibodies. The plates were emptied, washed three times with PBS, after which the blocked phage antibodies were added. Incubation was allowed to proceed for one hour, the plates were washed in PBS containing 0.05% Tween-20 and bound phage antibodies were detected (using OD 492 nm measurement) using an anti-M13 antibody conjugated to peroxidase. As a control, the procedure was performed simultaneously using no single-chain phage antibody or control single-chain phage antibody directed against thyroglobulin (SC02-006) (see De Kruif *et al.* 1995a and 1995b). Both controls served as a negative control. As shown in Figure 1, the selected phage antibodies called SC03-014 and SC03-022 displayed significant binding to the immobilized UV-inactivated SARS-CoV preparation, while no binding to FBS was observed.

Example 4

Characterization of the scFvs specific for SARS-CoV.

From the selected specific single chain phage antibody (scFv) clones plasmid DNA was obtained and nucleotide sequences were determined according to standard techniques. The nucleotide sequences of the scFvs (including restriction sites for cloning) called SC03-014 and SC03-022 are shown in SEQ ID NO:92 and SEQ ID NO:94, respectively. The amino acid sequences of the scFvs called SC03-014 and SC03-022 are shown in SEQ ID NO:93 and SEQ ID NO:95, respectively. The VH and VL gene identity (see Tomlinson IM, Williams SC, Ignatovitch O, Corbett SJ, Winter G. V-BASE Sequence Directory. Cambridge United Kingdom: MRC Centre for Protein Engineering (1997)) and heavy chain CDR3 compositions of the scFvs specifically binding the SARS-CoV preparation are depicted in Table 11.

Example 5

Construction of fully human immunoglobulin molecules (human monoclonal anti-SARS-CoV antibodies) from the selected anti-SARS-CoV single chain Fvs

Heavy and light chain variable regions of the scFvs called SC03-014 and SC03-022 were PCR-amplified using oligonucleotides to append restriction sites and/or sequences for expression in the IgG expression vectors pSyn-C03-HCγ1 (see SEQ ID No:96) and pSyn-C05-Cκ (see SEQ ID No:97), respectively. The V_L gene of scFv SC03-014 was amplified using oligonucleotides 5K-I acctgtctcgagttttccatggctgacatccagat gacccagtctccatcctcc (SEQ ID NO:98) and sy3K-C gctgggggcggccac ggtccgtttgatctccaccttggtccc (SEQ ID NO:99) and the PCR product cloned into vector pSyn-C05-Cκ. The V_L gene of scFv SC03-022 was amplified using oligonucleotides 5K-J acctgtctcgagt tttccatggctgacatcgtgatgacccagtctccag (SEQ ID NO:100) and sy3K-F gctgggggcggccacggtccgcttgatctccaccttggtccc (SEQ ID NO:101)

and the PCR product cloned into vector pSyn-C05-Cκ. Nucleotide sequences for all constructs were verified according to standard techniques known to the skilled artisan. V_H genes of scFv SC03-014 were amplified using oligonucleotides 5H-B acctgtcttgaattctccatggccgaggtgcagctggtggagtctg (SEQ ID NO:102) and sy3H-A gcccttggtgctagcgctggagacggtcaccagggtgccctggcccc (SEQ ID NO:103). V_H genes of scFv SC03-022 were amplified using oligonucleotide set 5H-H acctgtcttgaattctccatggccgaggtgcag ctggtgcagtctgg (SEQ ID NO:104) and sy3H-C gcccttggtgctagcgct ggagacggtcacggtggtgccctggcccc (SEQ ID NO:105). Thereafter, the PCR products were cloned into vector pSyn-C03-HCγ1 and nucleotide sequences were verified according to standard techniques known to the skilled person in the art.

The resulting expression constructs pgG103-014C03 and pgG103-022C03 encoding the anti-SARS-CoV human IgG1 heavy chains were transiently expressed in combination with the pSyn-C05-VkI (V_L SC03-014) and pgG103-022C05 (V_L SC03-022), respectively in HEK293T or PER.C6® cells and supernatants containing IgG1 antibodies were obtained. The nucleotide sequences of the heavy chains of the antibodies called CR03-014 and CR03-022 are shown in SEQ ID NO:106 and SEQ ID NO:108, respectively. The amino acid sequences of the heavy chains of the antibodies CR03-014 and CR03-022 are shown in SEQ ID NO:107 and SEQ ID NO:109, respectively.

The nucleotide sequences of the light chain of antibodies CR03-014 and CR03-022 is shown in SEQ ID NO:110 and SEQ ID NO:112, respectively. The amino acid sequences of the light chain of antibodies CR03-014 and CR03-022 is shown in SEQ ID NO:111 and SEQ ID NO:113. Subsequently, the recombinant human monoclonal antibodies were purified over protein-A columns and size-exclusion columns using standard purification methods

used generally for immunoglobulins (see for instance WO 00/63403 which is incorporated by reference herein).

Example 6

Screening assay for SARS-CoV neutralizing activity of recombinant human anti-SARS-CoV antibodies

The SARS-CoV neutralization assay was performed on Vero cells (ATCC CCL 81). The SARS-CoV strains used in the neutralization assay were the Frankfurt 1 strain (for the complete genome of this strain see EMBL-database accession # AY291315) (Rickerts *et al.* 2003). Virus stocks of the strains were used in a titer of $4x10^3$ $TCID_{50}$/ml (50% tissue culture infective dose per ml), with the titer calculated according to the method of Spearman and Kaerber which is known to the average skilled person. Recombinant human anti-SARS-CoV antibodies produced as described above were screened by serially 2-fold-dilution of the undiluted material (2.5 mg/ml) in PBS starting from 1:4 (dilution range 1:4 - 1:512). A neutralization titer of $\geq$ 1:4 was regarded as specific evidence of reactivity of the antibodies against SARS-CoV in the screening assay. Convalescent serum from a SARS-patient was used as a positive control for the neutralization assay.

In general, the neutralization assay worked as follows. 25 µl of the respective antibody dilutions were mixed with 25 µl of virus suspension (= approx. 100 $TCID_{50}$/25 µl) and incubated for one hour at 37°C. The suspension was then pipetted in triplicate into 96-well plates. Next, 50 µl of a freshly trypsinized and homogenized suspension of Vero cells (1:3 split of the confluent cell monolayer of a T75-flask), resuspended in DMEM containing 10% w/v FCS and antibiotics, were added. The inoculated cells were cultured for 3-4 days at 37°C and observed daily for the development of cytopathic

effect (CPE). CPE was compared to the positive control (virus inoculated cells) and negative controls (mock-inoculated cells. The complete absence of CPE in an individual cell culture was defined as protection (= 100% titer reduction). The highest antibody/serum dilution giving protection in 66% percent of wells was defined as the neutralizing antibody titer. The experiment was performed three times in triplicate (see Tables 12A, B and C). The IgGs CR03-014, CR03-022, a negative control IgG1 and a positive control serum from a SARS-patient were tested for SARS-CoV neutralizing activity. It is clear from Table 12A, B and C that the IgGs CR03-014 and CR03-022 displayed significant neutralizing activity. The CR03-014 IgG neutralized the Frankfurt 1 strain at titers of 128 (n=1) or 256 (n=2) in the above described assay. The CR03-022 IgG neutralized the Frankfurt 1 strain at titers of 32 (n=1) or 64 (n=2). These titers correspond to final antibody neutralization concentrations in the tissue culture well of 2.4 µg/ml (n=2) and 4.9 µg/ml (n=1) for CR03-014 and 9.8 µg/ml (n=2) and 19.5 µg/ml (n=1) for CR03-022. In the light of these concentrations both neutralizing antibodies may be suitable in the prophylaxis and/or treatment of a condition resulting from a SARS-CoV infection.

Additionally, different SARS-CoV strains were used to evaluate the potency and breadth of protection of the anti-SARS-CoV antibodies. The SARS-CoV strains HKU-36, HKU-39849, HKU-66, HKU-61567, GZ43 and GZ50 were passaged on FRhK-4 cells for two or three times before testing (see Table 13). Strain HKU-61644 was passaged on Vero cells and tested after passage 1 and 15. The SARS-CoV neutralization assay was performed on FRhK-4 cells as follows. A 500 µl stock solution (100 µg/ml) of antibody was prepared in maintenance medium (MM, MEM supplemented with 1% w/v FCS). From this stock solution 2-

fold-serially dilutions were prepared. 220 µl stock solution (100 µg/ml) was added in duplo in a 96-well plate from which 110 µl was taken and mixed with 110 µl MM in each of the nine subsequent wells. 110 µl of the tenth well was discarded. This resulted in ten wells containing 110 µl 0.2-100 µg/ml antibody. 110 µl of the antibody dilution was mixed with 110 µl of the different SARS-CoV isolates at a concentration of 2000 $TCID_{50}$/ml with the titer calculated according to the method of Reed and Muench which is known to the skilled artisan. At this stage, in a 220 µl volume, antibody concentrations varied from 0.1 to 50 µg/ml in the presence of 1000 $TCID_{50}$/ml SARS-CoV. The 96-well plate containing the antibody/virus mixtures was preincubated for 1-2 hours at 37°C. 100 µl of the antibody/virus mixtures were added in quadruplicate to wells from a second 96-well tissue culture plate containing confluent FRhK-4 cells in 100 µl MM and incubated at 37°C. During this final incubation step, 100 $TCID_{50}$ of SARS-CoV was present in the presence of antibody concentrations varying from 0.05 to 25 µg/ml. The cells were cultured at 37°C and observed for the development of CPE at 72 and 96 hours. CPE was compared to a positive control (SARS-CoV inoculated cells) and a negative control (cells incubated with MM only). The antibody neutralization titer was determined as the concentration of antibody which gave 100% protection of the quadruplicate cell cultures. The monoclonal anti-SARS-CoV antibody CR03-014 completely neutralized 100 $TCID_{50}$ of all tested SARS-CoV isolates at a concentration of 12.5 µg/ml (see Table 13). This indicates that antibody CR03-014 is able to neutralize a variety of SARS-CoV isolates.

In an additional experiment the SARS-CoV neutralization assay was performed as described for the Frankfurt 1 strain *supra* to determine synergy between SARS-CoV neutralizing

antibodies CR03-014 and CR03-022. Stock solutions of antibody CR03-014 and CR03-022 of approximately similar potency were mixed in different ratios. To compensate for an estimated 4 times higher potency of CR03-014 compared to CR03-022, the CR03-014 antibody stock solution of 2.5 mg/ml was diluted 4-fold to 625 µg/ml. Subsequently, antibody CR03-014 and CR03-022 were mixed in the following ratios (mixture A: CR03-014 0%, CR03-022 100%; mixture B: CR03-014 10%, CR03-022 90%; mixture C: CR03-014 50%, CR03-022 50%; mixture D: CR03-014 90%, CR03-022 10%; and mixture E: CR03-014 100%, CR03-022 0%). When the antibodies in the mixtures have an additive effect, the mixtures should neutralize SARS-CoV at the same titer as the individual antibodies present in the mixtures. When the antibodies in the mixtures have a synergistic effect, the mixtures should neutralize SARS-CoV at a higher titer as the individual antibodies present in the mixtures. The neutralization assay was performed twice in triplicate wells as described above. The results of both assays were combined. Protection of at least 66% percent of the wells (4 of the 6 wells tested) was defined as the neutralizing antibody titer. The neutralization titers of the separate mixtures are shown in Table 14. From Table 14 can be deducted that the mixtures had the following titers: mixture A, 64; mixture B, 256; mixture C, >1024; mixture D, 256; and mixture E, 16. From this can be concluded that, when both antibodies were tested in combination (mixtures B-D), the neutralizing titers were higher than those for antibody CR03-014 and CR03-022 individually. Together, these data indicate that antibodies CR03-014 and CR03-022 exhibit synergistic SARS-CoV neutralizing activity.

In yet another embodiment the SARS-CoV neutralization assay showing synergy between the anti-SARS-CoV antibodies was

performed on FRhK-4 cells (ATCC CRL-1688) as follows. The SARS-CoV strain called HK-39849 (GenBank accession number AY278491) was used in a titer of $2x10^3$ $TCID_{50}$/ml as calculated according to the method of Reed and Muench known to the average skilled person. The human anti-SARS-CoV antibodies were screened by serially 1.46-fold-dilution in maintenance medium (MM) (1% w/v FCS in MEM with antibiotic) starting at a concentration of 200 µg/ml (dilution range 200 - 6.7 µg/ml) in duplo. Four different compositions were tested: antibody CR03-014 individually, antibody CR03-022 individually, control IgG1 antibody, and antibodies CR03-014 and CR03-022 in combination (start concentration 200 µg/ml of each antibody). 110 µl of virus suspension was mixed with 110 µl of the respective recombinant human anti-SARS-CoV antibody dilution and incubated for one hour at 37°C. 100 µl of this suspension was then pipetted two times in duplicate into 96-well plates containing an 80% confluent monolayer of FRhK-4 cells in 100 µl MM. The FRhK-4 cells were cultured at 37°C and observed after 3-4 days for the development of CPE. CPE was compared to the positive control (virus inoculated cells) and negative controls (mock-inoculated cells or cells incubated with recombinant antibody only). The complete absence of CPE in an individual cell culture was defined as protection (= 100% titer reduction). The concept of the combination index (CI) was used to quantitate synergistic effects as described previously (Chou and Talalay, 1984). According to the concept a combination of agents that produce an additive effect the sum of the ratios of their concentrations in the mixture (cmixt) to the concentrations of agents that individually have the same effect as the mixture (ceffect) is 1. This sum is the CI. When this sum is lower than 1, the agents act in synergy.

For a two-component system, as in the present study, CI is calculated as follows:

$$\frac{C1mixt}{C1eff} + \frac{C2mixt}{C2eff} = 1$$

C1mixt is the concentration of the first component in the mixture which leads to a certain level of inhibition (f), c1effect is that concentration of the first component which alone (in the absence of the second component) will result in the same inhibitory effect as the mixture of the two components, and c2mixt and c2effect are the corresponding concentrations for the second component.

To determine the CI for antibodies CR03-014 and CR03-022 the neutralization assay was performed as described above. Complete neutralization of 100 $TCID_{50}$ of strain HKU39849 was reached at 7.6 µg/ml for CR03-014, 50.0 µg/ml for CR03-022 and 2.4 µg/ml of each antibody when added in combination. This outcome results in a CI of 2.4/7.6 + 2.4/50.0= 0.36. Fifty percent neutralization was achieved at 5.9 µg/ml for CR03-014, 30.2 µg/ml for CR03-022 and 1.7 µg/ml of each antibody when added in combination. This results in a CI of 1.7/5.9 + 1.7/30.2= 0.34. Thus, at both 50 and 100% neutralization CI values lower than 1 were obtained for a mixture of CR03-014 and CR03-022, which indicates that both antibodies act in synergy against SARS-CoV.

Example 7

Binding of anti-SARS antibodies to SARS-CoV, SARS-CoV spike protein and fragments thereof.

An ELISA to detect binding of anti-SARS antibodies to SARS-CoV was performed as follows. Wells of ELISA plates were coated overnight with UV-inactivated SARS-CoV preparation in 50 mM bicarbonate buffer pH 9.6. The wells of the plates were washed three times with PBS containing 0.05% Tween and blocked for 2 hours at 37°C with PBS containing 1% BSA. Next, the antibodies diluted in PBS containing 1% BSA were incubated for 1 hour at room temperature. The wells were washed three times with PBS containing 0.05% Tween and incubated for 1 hour at room temperature using a murine anti-Hu-IgG HRP conjugate. Development was done with O-phenylenediamine substrate, the reaction was stopped by the addition of 1 M H_2SO_4 and the absorbance was measured at 492 nm. As shown in Figure 2 antibodies CR03-014 and CR03-022 were both capable of binding an inactivated SARS-CoV preparation in ELISA in contrast to a negative control IgG which was directed against an irrelevant antigen.

To detect the target of the antibodies CR03-014 and CR03-022 another binding assay was used. Single chain phage antibodies SC03-014 and SC03-022 were analyzed for their ability to bind HEK293T cells that recombinantly express proteins of SARS-CoV. To this end HEK293T cells were transfected with a plasmid carrying a cDNA sequence encoding the spike (S) protein from SARS-CoV strain Frankfurt 1 or with control vector. For flow cytometry analysis, single-chain phage antibodies were first blocked in an equal volume of 4% PBS-M for 15 minutes at 4°C prior to the staining of the transfected HEK293T cells. The blocked phage antibodies were added to mock transfected HEK293T cells and HEK293T cells transfected with the SARS-CoV S protein. The binding of the single chain phage antibodies to the cells was visualized using a biotinylated anti-M13 antibody (Santa Cruz

Biotechnology) followed by streptavidin-phycoerythrin (Caltag). As shown in Figure 3, the single chain phage antibodies SC03-014 and SC03-022 were capable of binding spike transfected HEK293T cells, whereas no binding to mock transfected HEK293T cells was observed. A control single chain phage antibody did neither recognize the spike transfected HEK293T cells nor the mock transfected HEK293T cells. These data suggest that both antibodies are directed against the S protein of SARS-CoV.

To further localize the binding sites of the antibodies within the S protein, an assay was performed wherein the antibodies were analyzed for their ability to bind to portions of the S protein of SARS-CoV. The nucleotide and amino acid sequence of the S protein is shown in SEQ ID NO:114 and SEQ ID NO:115, respectively. DNA coding for the N-terminal 565 amino acids (portion called S565) was cloned as a *Kpn*I-*Bam*HI fragment in pAdapt (Havenga *et al.*, 2001) that was modified by insertion of the polylinker of the vector called pcDNA3.1/myc-His C (Invitrogen) (vector called pAdapt/myc-His C). A fragment corresponding to amino acid residues 318-510 of the S protein (portion called S318-510) was amplified on S gene cDNA using the oligonucleotide primers: *Eco*RIspikeFor318 5′-cctggaattctccatggccaacatcaccaacc-3′ (SEQ ID NO:116) and *Xba*IspikeRev510 5′-gaagggccctctagacacggtggcagg-3′ (SEQ ID NO:117). The resulting fragment was digested with *Eco*RI-*Xba*I and cloned into pHAVT20/myc His A to yield pHAVT20/myc-His A S318-510. In this vector expression of fragment S318-510 fused to the HAVT20 leader sequence was under control of the human, full-length, immediate-early CMV promoter. DNA transfections were performed in HEK293T cells for transient expression using standard techniques. The S protein derived fragments were used directly from culture supernatant or were purified from

culture supernatant using Ni-NTA (Qiagen). An ELISA to evaluate binding of antibodies to the S protein derived fragments was performed as follows. Wells of ELISA plates were coated overnight with 5 µg/ml anti-myc antibody in 50 mM bicarbonate buffer pH 9.6. The wells of the plates were washed three times with PBS containing 0.05% Tween and blocked for 2 hours at 37°C with PBS containing 1% BSA. The wells coated with anti-myc antibody were incubated with the myc-tagged fragments S565 or S318-510 diluted in PBS containing 1% BSA for 1 hour at room temperature. The wells were washed three times with PBS containing 0.05% Tween. Next, the antibodies CR03-014, CR03-022 or control antibody diluted in PBS containing 1% BSA were incubated for 1 hour at room temperature. Detection of bound antibody was performed as described *supra*. As shown in Figure 4, antibodies CR03-014 and CR03-022 were both capable of binding to the S565 and S318-510 fragment, but not to an irrelevant control myc-tagged antigen. A control antibody did not bind any of the fragments.

In order to rank the affinities of the antibodies for binding to the S565 fragment, a titration of IgG concentration was performed (by diluting the antibodies in PBS containing 1% ELK), followed by an ELISA as described above. Titration of the monoclonal antibodies showed that CR03-014 and CR03-022 bound S565 with approximately similar affinities (see Figure 5). To investigate changes of affinities as a possible mechanism of synergy, the K_D for CR03-014 and CR03-022 binding sequentially or simultaneously to recombinant receptor binding domain fragment S318-510 was investigated by means of BIAcore analysis. Surface plasmon resonance analyses were performed at 25°C on a BIAcore3000™. CM5 sensorchips and running buffer HBS-EP were from Biacore AB (Uppsala Sweden). Recombinant S318-510 fragment was immobilized to CM5 chips using an amine

coupling procedure resulting in a response level of approximately 1,000 resonance units (RU). Kinetic analysis was performed to determine the association rate (k_a), dissociation rate (k_d) constants and the affinity (K_D) of the monoclonal antibodies. Therefore, a concentration series of 0.4 to 250 nM IgG was prepared using two-fold dilutions in HBS-EP. Samples were injected in duplicate at a flow rate of 30 µl/min (injection time = 2 min, dissociation time = 5 min). The sensor chip surface was regenerated with a pulse of 5 µl 5 nM NaOH. Biacore evaluation software (BIAevalution, July 2001) was used to fit the association and dissociation curves of all concentrations injected. The individual K_D for CR03-014 and CR03-022 was determined as 16.3 nM and 0.125 nM, respectively, the K_D for the antibodies binding simultaneously as 5.71 nM and for binding of CR03-014 to CR03-022 saturated S318-510 as 14.8 nM. Compared to the dose reduction indices of 3 and 20 for CR03-014 and CR03-022, respectively, neither simultaneous nor sequential binding of the monoclonal antibodies resulted in changes of K_D which could explain their synergistic neutralizing action through cooperative binding.

To further explore the antibody binding sites within the S protein, a competition ELISA on immobilized S318-510 fragment was performed. Captured S318-510 was incubated with non-saturating amounts of biotinylated antibody without competing antibody or in the presence of 1, 5, and 25 µg/ml of competing antibody (antibody CR03-014, CR03-022 or control antibody). Bound biotinylated antibody was detected with streptavidin-conjugated-HRP (BD Pharmingen) and developed as described above. Figure 6A shows that binding of monoclonal antibody CR03-014 was unaffected in the presence of excess unlabeled monoclonal antibody control or antibody CR03-022. Figure 6B shows that binding of antibody CR03-022 was

unaffected in the presence of excess unlabeled monoclonal antibody control or antibody CR03-014. As expected, binding of both biotinylated CR03-014 and CR03-022 was effectively reduced by their unlabeled counterparts (see Figures 6A and B). These results demonstrate that the antibodies CR03-014 and CR03-022 do not compete with each other for binding to the S318-510 fragment and recognize different/distinct, non-competing epitopes.

To confirm these findings, an antibody sandwich ELISA was performed. Antibodies CR03-014 and CR03-022 were coated overnight to microtiter wells at 5 µg/ml in 50 mM bicarbonate buffer pH 9.6. Capture of the S318-510 fragment, binding of biotinylated antibodies and subsequent development of the ELISA reaction was performed as described *supra*. Figure 7 indicates that CR03-014-captured S318-510 could be bound by biotinylated CR03-022, but not by CR03-014. CR03-022-captured S318-510 could only be bound by biotinylated CR03-014 and not CR03-022. This indicates that CR03-014 and CR03-022 are able to bind simultaneously to different epitopes within the S derived fragment S318-510 and furthermore indicates that both antibodies bind to different non-competing epitopes.

Example 8

Construction and evaluation of binding of the monoclonal anti-SARS-CoV antibodies to variant S318-510 fragments.

The diversity within the region 318-510 of the S protein was determined as follows. A list containing more than 146 genomes or genes encoding complete human SARS-CoV or fragments thereof was compiled. In 114 cases, an open reading frame encoding for full-length spike (S) protein was identified. Alignment of the spike amino acid residues 318-510 revealed 30 spike proteins, in which the region 318-510 was not identical

to that of the spike protein of strain Frankfurt 1 (see Genbank accession number AY291315), which was used herein as wild-type. The mutations, strains and Genbank numbers are depicted in Table 15. To investigate if CR03-014 and CR03-022 bound the S protein of all currently known human SARS-CoV isolates, eight recombinant spike 318-510 fragments (variants A-F and variants H and I) harbouring the different amino acid substitutions as shown in Table 15 were generated. In addition, a 318-510 fragment (variant G) corresponding to the sequence of four animal SARS-like CoVs (Genbank accession numbers AY304486 - AY304489; see also Table 1, SARS-CoV SZ3, SZ13, SZ16 and SZ1, respectively) was generated. The four SARS-like CoVs, which were isolated from raccoon dogs and palm civet cats, contain the amino acid substitutions K344R, F360S, N479K and T487S (see Guan *et al.* 2003). To this end, the above substitutions were introduced in the pHAVT20/myc-His A S318-510 vector using the Stratagene's QuikChange II site-directed mutagenesis kit according to the manufacturer's instructions. In case a sequence contained multiple amino acid substitutions, the process of mutagenesis, sequence analysis and confirmation was repeated for every single substitution. To rule out the introduction of additional mutations in the plasmid outside the gene of interest, the mutated (592 bp *Eco*RI-*Xba*I) fragment was recloned in *Eco*RI-*Xba*I cut pHAVT20/myc-His A. The resulting plasmids were transfected into HEK293T cells, and binding of CR03-014 and CR03-022 was evaluated by means of ELISA as described *supra*. As shown in Figure 8, CR03-014 was capable of binding to variants A-E and variants H and I to a similar extent as to the wild-type fragment. Binding of CR03-014 to variant F (N479S substitution) was substantially lower than binding to the other fragments. No binding of CR03-014 to fragment G (K344R,

F360S, N479K and T487S substitutions) was observed. Antibody CR03-022 was capable of binding all variant S318-510 fragments to a similar extent as the wild-type (non-mutated) S318-510 fragment. Together this indicates that residue N479 is involved in binding of CR03-014, either directly by being part of the binding site of CR03-014 or indirectly by being important for the correct conformation of the binding site of CR03-014 within the S protein. Since, antibody CR03-022 is capable of binding to recombinant fragments composed of amino acid residues 318-510 of all human SARS-CoV isolates (as described in Table 15) and in addition is also capable of binding to animal SARS-like CoV, CR03-022 is suitable for treatment and/or protection against SARS-CoV isolates in general.

Particularly suitable for treatment and/or protection against human SARS-CoV isolates is a combination/cocktail comprising both antibodies, CR03-014 and CR03-022, as both antibodies are capable of specifically binding to human SARS-CoV and the antibodies act synergistically in neutralizing human SARS-CoV. In other words, the combination/cocktail of CR03-014 and CR03-022 comprises synergistic human SARS-CoV neutralizing activity. An additional advantage of such a combination/cocktail is its capability of neutralizing human SARS-CoV as well as animal SARS-like CoV.

Example 9

Generation of CR03-014 and CR03-022 escape viruses of SARS-CoV

To further elucidate the epitopes recognized by the human monoclonal antibodies CR03-014 and CR03-022, escape viruses of CR03-014 and CR03-022 were generated. The process for generating escape viruses of CR03-014 is given *infra*. The process for generating escape viruses of CR03-022 was

identical with the proviso that 60 µg/ml antibody instead of 20 µg/ml was used in all respective steps. Serial dilutions (0.5 ml) of SARS-CoV strain HKU 39849 (dilutions ranging from 10^{-1} - 10^{-8}) were incubated with a constant amount (20 µg/ml giving a ~3 log reduction of $TCID_{50}$/ml) of antibody CR03-014 (0.5 ml) for 1 hour at 37°C/5% CO_2 before addition to wells containing FRhK-4 cells. The virus was allowed to attach to the cells for 1 hour at 37°C/5% CO_2, then removed and cells were washed twice with medium. Finally, cells were incubated for 2 days in the presence of selecting antibody CR03-014 at 20 µg/ml in 0.5 ml medium. Then, medium of wells with highest virus dilution showing CPE (cytopathic effect) containing potential escape viruses was harvested and stored at 4°C until further use. Subsequently, virus samples were freeze/thawed once and serial dilutions were prepared in medium. Virus dilutions were added to wells containing FRhK-4 cells and incubated for 1-2 hours at 37°C/5% CO_2 in the presence of CR03-014 at 20 µg/ml. Wells were then overlayed with agarose containing CR03-014 at 20 µg/ml and incubated for 3-5 days at 37°C/5% CO_2. Individual escape virus plaques were picked using a Pasteur pipet, freeze/thawed once, and the escape viruses were amplified on FRhK-4 cells. To analyse the escape viruses the following experiments were performed.

Firstly, to identify possible mutations in the SARS-CoV spike protein of each of the escape viruses the nucleotide sequence of the SARS-CoV spike open reading frame (ORF) was determined. Viral RNA of each of the escape viruses and wild-type SARS-CoV virus was isolated and converted into cDNA by standard RT-PCR. Subsequently, the cDNA was used for nucleotide sequencing of the SARS-CoV spike ORF in order to identify mutations. Figure 9 shows the results of the

sequencing data for the five E014 escape viruses obtained. All escape viruses contained a nucleotide mutation at position 1385 (C to T), which resulted in an amino acid mutation P to L at position 462 in the spike protein. Apparently, P462L resulted in loss of epitope recognition and, subsequently, loss of neutralization of SARS-CoV by CR03-014. This indicates that next to amino acid 479, also amino acid 462 is involved in the binding of CR03-014 to the S protein of SARS-CoV. The results of the sequencing data for the five E022 escape viruses obtained were as follows. Four out of five escape viruses contained a nucleotide mutation at position 2588 (C to T), which resulted in an amino acid mutation T to I at position 863 in the spike protein.

Secondly, the neutralization index (NI) was determined for each of the E014 and E022 escape viruses. A virus was defined as an escape variant, if the neutralization index was <2.5 logs. The process of determining the NI is given below for E014 escape viruses. The process was identical for E022 escape viruses with the proviso that 60 µg/ml instead of 20 µg/ml monoclonal antibody was used in all respective steps. The neutralization index was determined by subtracting the number of infectious virus particles (in $TCID_{50}$/ml) produced in FRhK-4 cell cultures infected with virus plus monoclonal antibody (20 µg/ml) from the number of infectious virus particles (in $TCID_{50}$/ml) produced in FRhK-4 cell cultures infected with virus alone ([log TCID50/ml virus in absence of monoclonal antibody minus log TCID50/ml virus in presence of monoclonal antibody]). An index lower than 2.5 logs was considered as evidence of escape. Therefore, each escape virus and wild-type SARS-CoV (100 $TCID_{50}$) was incubated for 1 hour at 37°C/5% CO_2 with 20 µg/ml of CR03-014 before addition to FRhK-4 cells. The

virus was allowed to attach to the cells for 1 hour at 37°C/5% CO_2 after which the inoculum was removed and cells were washed twice with medium before being replenished with medium containing 20 µg/ml of CR03-014. After a 2 day incubation at 37°C/5% CO_2 the medium was harvested and the $TCID_{50}$/ml of each virus was determined. As shown in Table 16, the concentration of antibody used to determine the NI resulted in a ~3 log reduction of virus titer when performed on the wild-type SARS-CoV virus. Thus, wild type SARS-CoV was neutralized by CR03-014 as judged by the NI of 3.3. In contrast, the NI for each escape virus was <2.5 and thus each of the escape viruses was no longer neutralized by CR03-014. As judged by the NI of 3.3, wild-type SARS-CoV virus was also neutralized by CR03-022 (see Table 17). The NI for each E022 escape virus was >2.5 and thus it was concluded that each of the escape viruses was still neutralized by CR03-022. The amino acid substitution in four of the five E022 escape viruses apparently does not play a role in neutralization of SARS-CoV by CR03-022. It might have been induced non-specifically during the course of the experiment. This agrees with finding by Poon *et al.* (2005) who observed the mutation at position 863 (T to I) when SARS-CoV was passaged multiple times in FRhK-4 cells. The neutralizing epitope of CR03-022 could not be determined by means of generating escape viruses. This may be caused by the functional constraints of the binding region on the S protein. A mutation occurring in this region may be detrimental to the stability of the virus and could therefore not be isolated in the experiments described above.

In order to confirm lack of binding of antibody CR03-014 to the escape variant, a recombinant S318-510 fragment harbouring the P to L substitution at position 462 was

constructed essentially as described *supra*. DNA transfection of the resulting plasmid was performed in HEK293T cells for transient expression and the fragments were directly used from culture supernatant. The ELISA was performed as described *supra*. Briefly, the fragments were captured on anti-myc coated microtiter wells. Subsequently, antibodies CR03-014 and CR03-022 were added and binding of the antibodies was detected using an anti-human IgG HRP-conjugate. As shown in Figure 10, antibody CR03-014 was not able to bind the S318-510 fragment carrying a P to L substitution at position 462. Binding of CR03-022 was not affected by this amino acid substitution. This further indicates that antibody CR03-022 is capable of binding to a different/distinct, non-competing epitope on the S protein and suggests that CR03-022 might be used to compensate potential lack of neutralization of SARS-CoV variants by CR03-014.

Furthermore, cross-neutralization assays were performed on E14 escape viruses. Table 18 clearly shows that CR03-022 neutralized the E14 escape viruses to a similar level as wild-type virus further illustrating that CR03-022 binds to a different epitope compared to CR03-014, which no longer neutralized the E14 escape viruses. This result is in agreement with the ELISA data shown in Figure 10. The reverse experiment could not be performed as escape viruses of CR03-022 could not be generated. From the above can be concluded that the combination of CR03-014 and CR03-022 in a cocktail prevents the escape of neutralization-resistant SARS-CoV variants and is therefore an ideal immunoglobulin preparation for SARS-CoV prophylaxis and therapy.

Example 10

Assessment of potential enhancement of SARS-CoV infection in human macrophages by the human anti-SARS monoclonal antibody CR03-014

It is known that in certain diseases caused by coronaviruses prior immunity or passive antibody increases the severity of the disease. In feline infectious peritonitis, the macrophage is the main target cell for virus replication and anti-viral antibodies enhance replication of the virus in macrophage cultures *in vitro*. The macrophage is also a prominent cell seen in the cell infiltrates of lungs of patients dying of SARS-CoV (Nicholls *et al.* 2003). This has led to concern whether antibody dependent enhancement (ADE) may be relevant in the pathogenesis of SARS-CoV. To investigate this it was tested whether ADE occurred when macrophages were infected with SARS-CoV in the presence of the neutralizing anti-SARS-CoV monoclonal antibody CR03-014, the non-neutralizing anti-SARS-CoV monoclonal antibody CR-03-015, the monoclonal antibody called CR-JA (an monoclonal antibody against rabies which is used herein as a control antibody), convalescent serum from an individual exposed to SARS-CoV and serum from a healthy individual.

Human peripheral blood mononuclear cells (PBMCs) were obtained from leukocyte-rich buffy coats of healthy blood donors (The Hong Kong Red Cross Blood Transfusion Service, Hong Kong). The PBMCs were separated by Ficoll-Paque gradient centrifugation (Pharmacia Biotech, Uppsala, Sweden). To isolate monocytes 2 x 10^7 PBMCs were allowed to adhere onto petridishes (Greiner, Frickenhausen, Germany) for 1 hour in RPMI 1640 medium supplemented with 20 mM HEPES, 2 mM glutamine, 0.6 µg/ml penicillin, and 60 µg/ml streptomycin and 5% heat-inactivated autologous plasma. After washing with

medium, the adherent monocytes were detached by pipeting and re-seeded into 24 well plates at a density of 2×10^5 cells per well in supplemented RPMI 1640 medium.

To check the purity of the monocyte preparations monocytes were also seeded and allowed to adhere onto glass coverslips. The purity of the monocytes on the glass coverslips was confirmed by staining with a CD14 R-phycoerythrin (R-PE)-conjugated mouse anti-human monoclonal antibody (BD Biosciences, San Diego, U.S.A.).

Medium in the monocyte cultures was replaced every 2-3 days and the cells were allowed to differentiate for 14 days *in vitro*. Differentiation of monocytes into macrophages was confirmed by the typical morphology of macrophages. The obtained primary human monocyte-derived macrophages were used in further experiments. Two days prior to the ADE experiments, the supplemented RPMI 1640 medium was exchanged into Macrophage Serum Free medium (SFM) (Invitrogen, Carlsbad, CA, U.S.A.).

To investigate the effect of sub-neutralizing doses of antibody on viral infection in macrophages, 300 µl of serial 10-fold dilutions in MM medium (MEM including 1% FCS and 0.6 µg/ml penicillin, and 60 µg/ml streptomycin) of the monoclonal antibodies CR03-014, CR03-015 and CR-JA, convalescent serum from a SARS-CoV exposed individual and serum from a healthy individual was mixed with 300 µl of SARS-CoV. MM medium mixed with SARS-CoV served as the virus control. The virus/monoclonal antibody mixtures and virus/serum mixtures were incubated for one hour at 37°C. Then, 250 µl of the mixtures was added to duplicate wells containing macrophages. After one hour of virus adsorption at 37°C, the virus inoculum was removed, infected cells were washed with macrophage SFM culture medium and incubated in macrophage SFM medium

supplemented with 0.6 µg/ml penicillin and 60 µg/ml streptomycin. Samples of the culture supernatants were collected at days 0, 1, 2, 3, 5, and 7 post-infection and stored at -70°C for virus titration experiments. SARS-CoV was titrated and the TCID50 determined essentially as described supra.

To detect virus inside the macrophages (due to abortive infection) and to detect potential transcription of SARS-CoV RNA inside the macrophages RNA was isolated from infected macrophages at 3, 6, and 24 hours post-infection using the RNeasy Mini kit (Qiagen) according to the manufacturer's instructions. Reverse transcription with oligo-dT primers was performed by using Superscript II reverse transcriptase (Invitrogen) according to the manufacturer's instructions. Complementary DNA was generated with 10 µl of RNA, and reverse-transcribed by 200 U of Superscript II reverse transcriptase (Invitrogen) in a 20 µl reaction containing 25 ng oligo-dT_{12-18} primer, 10 mM dithiothreitol, and 0.5 mM deoxynucleotide triphosphates. Reactions were incubated at 42°C for 50 minutes, followed by a heat inactivation step (72°C for 15 minutes). The reaction mix was diluted 10 times by the addition of 180 µl buffer AE (Qiagen) and stored at -20°C.

Real time quantitative PCR was performed using FastStart DNA Master SYBR Green I fluorescence reaction (Roche). 5 µl of diluted complementary DNA was amplified in a 20 µl reaction containing 4 mM of $MgCl_2$, 0.5 mM of forward primer (Actin-LF: CCCAAGGCCAACCGCGAGAAGAT (SEQ ID NO:122)), and 0.5 mM of reverse primer (Actin-LR: GTCCCGGCCAGCCAGGTCCAG (SEQ ID NO:123)). Reactions were performed in a LightCycler (Roche) with the following conditions: 10 minutes at 95°C, followed by 40 cycles of 95°C for 0 seconds, 66°C for 5 seconds, and 72°C

for 9 seconds. Plasmids containing the target sequence were used as positive controls. Fluorescence signals from these reactions were captured at the end of the extension step in each cycle (87°C). To determine the specificity of the assay, PCR products were subjected to melting curve analysis at the end of the assay (65 to 95°C; 2°C/second). Reverse transcription with sense (negative strand detection) or anti-sense (positive strand detection) primers to the polymerase gene of SARS-CoV was achieved by using Superscript II reverse transcriptase (Invitrogen) according to manufacturer's instructions. Complementary DNA was generated with 5 µL of RNA, and reverse-transcribed by 200 U of Superscript II reverse transcriptase (Invitrogen) in a 20 µl reaction containing 0.1 µM gene specific primer, 10 mM dithiothreitol, and 0.5 mM deoxynucleotide triphosphates. Reactions were incubated at 42°C for 50 minutes, followed by a heat inactivation step (72°C for 15 minutes). The reaction was diluted 10 times by the addition of 180 µL buffer AE (Qiagen) and stored at -20°C. 2 µl of diluted complementary DNA was amplified in 20 µl containing 3.5 mM of $MgCl_2$, 0.25 µM of forward primer (coro3: 5'-TACACACCTCAGCGTTG-3' (SEQ ID NO:124)), and 0.25 µM of reverse primer (coro4: 5'-CACGAACGTGACGAAT-3' (SEQ ID NO:125)). Reactions were performed in a LightCycler (Roche) with the following conditions: 10 min at 95°C, followed by 50 cycles of 95°C for 10 seconds, 60°C for 5 seconds, and 72°C for 9 seconds. Plasmids containing the target sequence were used as positive controls. Fluorescence signals from these reactions were captured at the end of the extension step in each cycle. To determine the specificity of the assay, PCR products were subjected to melting curve analysis at the end of the assay (65 to 95°C; 0.1°C/seconds).

Data for viral RNA were normalised for RNA levels of β-actin housekeeping gene.

After infection of macrophages with SARS-CoV, the cells were monitored daily under the microscope. Ten days after infection with SARS-CoV no detectable cytopathic effect was detected, nor was the abortive infection of macrophages converted into a productive infection. Pre-incubating of SARS-CoV with different concentrations of the monoclonal antibodies CR03-014, CR03-015, CR-JA, convalescent serum or serum from a healthy individual, prior to infection of macrophages did not change this out-come.

Titration of aliquots of supernatant taken from infected macrophage culture at days 0, 1, 2, 3, 5, and 7 days post-infection on FRhk-4 cells revealed no evidence of significant enhancement of SARS-CoV replication in macrophages by the monoclonal antibodies and sera tested (data not shown).

In addition, the effect of antibodies on the (abortive) infection of macrophages by SARS-CoV and the transcription of SARS-CoV genes within the macrophages was measured on the molecular level. To this end, total RNA was extracted from the infected macrophages at various time points post-infection as described *supra*. Subsequently, total RNA was analyzed for SARS-CoV viral positive strand RNA and viral negative strand RNA transcripts using real-time RT-PCR. The SARS-CoV RNA levels were normalized for the levels of β-actin mRNA. The results show that positive strand SARS-CoV RNA was detected in all macrophage cultures that were incubated with SARS-CoV, which confirms the abortive infection of macrophages by SARS-CoV. The levels of positive strand RNA observed in macrophage cultures infected with SARS-CoV in the presence of anti-SARS-CoV monoclonal antibodies CR03-014 or CR03-015 or convalescent serum were not significantly higher than in macrophage

cultures infected with SARS-CoV in the presence of control monoclonal antibody CR-JA or serum from a healthy individual or in the absence of monoclonal antibody or serum (data not shown).

Furthermore, the presence of negative strand RNA which is indicative for SARS-CoV gene transcriptional activity after infection of macrophages by SARS-CoV was measured. Again no correlation between the levels of negative strand RNA and the presence or absence of anti-SARS-CoV antibodies was observed (data not shown).

Together, the experiments assaying infectious virus yield and virus related RNA levels inside macrophages showed that there was no antibody dependent enhancement of SARS-CoV replication in human macrophages by anti-SARS-CoV monoclonal antibodies CR03-014 and CR03-015 and anti-SARS-CoV antibodies present in convalescent serum from a SARS patient.

Example 11

Identification of epitopes recognized by recombinant human anti-SARS-CoV antibodies by PEPSCAN-ELISA

15-mer linear and looped/cyclic peptides were synthesized from proteins of SARS-CoV and screened using credit-card format mini-PEPSCAN cards (455 peptide formats/card) as described previously (see *inter alia* WO 84/03564, WO 93/09872, Slootstra *et al.* 1996). All peptides were acetylated at the amino terminus. In short, series of overlapping peptides, which were either in linear form or in looped/cyclic form, of the spike protein of SARS-CoV Urbani (the protein-id of the surface spike glycoprotein in the EMBL-database is AAP13441), was produced and tested for binding to the recombinant human anti-SARS-CoV antibodies of the invention by means of PEPSCAN analysis.

Because the Urbani spike protein indicated above was also found in identical or highly homologous form in other SARS-CoV strains, the antigenic peptides found in the analysis method may not only be used for detection of the SARS-CoV strain Urbani and the prevention and/or treatment of a condition resulting from the SARS-CoV strain Urbani, but may also be useful in detecting SARS-CoV in general and preventing and/or treating a condition resulting from SARS-CoV in general. The protein-id of the surface spike glycoprotein of for instance the SARS-CoV strains TOR2, Frankfurt 1 and HSR 1 in the EMBL-database is AAP41037, AAP33697 and AAP72986. The accession number in the EMBL-database of the complete genome of the strains TOR2, Frankfurt 1 and HSR 1 is AY274119, AY291315 and AY323977, respectively. Under these accession numbers the amino acid sequence of the other (potential) proteins of these strains can be found.

In all looped peptides position-2 and position-14 were replaced by a cysteine (acetyl-XCXXXXXXXXXXXCX-minicard). If other cysteines besides the cysteines at position-2 and position-14 were present in a prepared peptide, the other cysteines were replaced by an alanine. The looped peptides were synthesized using standard Fmoc-chemistry and deprotected using trifluoric acid with scavengers. Subsequently, the deprotected peptides were reacted on the cards with an 0.5 mM solution of 1,3-bis(bromomethyl)benzene in ammonium bicarbonate (20 mM, pH 7.9/acetonitril (1:1 (v/v)). The cards were gently shaken in the solution for 30-60 minutes, while completely covered in the solution. Finally, the cards were washed extensively with excess of H2O and sonicated in disrupt-buffer containing 1% SDS/0.1% beta-mercaptoethanol in PBS (pH 7.2) at 70°C for 30 minutes, followed by sonication in H2O for another 45 minutes.

Recombinant human anti-SARS-CoV antibodies CR03-014 and CR03-022 were tested for binding to each linear and looped peptide in a PEPSCAN-based enzyme-linked immuno assay (ELISA). The 455-well creditcard-format polypropylene cards, containing the covalently linked peptides, were incubated with the antibodies (1-10 µg/ml; diluted in blocking solution which contains 5% horse-serum (v/v) and 5% ovalbumin (w/v)) (4°C, overnight). After washing, the peptides were incubated with anti-human antibody peroxidase (dilution 1/1000) (1 hour, 25°C), and subsequently, after washing the peroxidase substrate 2,2'-azino-di-3-ethylbenzthiazoline sulfonate (ABTS) and 2 µl/ml 3% H_2O_2 were added. Controls (for linear and looped) were incubated with anti-human antibody peroxidase only. After 1 hour the colour development was measured. The colour development of the ELISA was quantified with a CCD-camera and an image processing system. The set-up consisted of a CCD-camera and a 55 mm lens (Sony CCD Video Camera XC-77RR, Nikon micro-nikkor 55 mm f/2.8 lens), a camera adaptor (Sony Camera adaptor DC-77RR) and the Image Processing Software package Optimas, version 6.5 (Media Cybernetics, Silver Spring, MD 20910, U.S.A.). Optimas runs on a pentium II computer system.

The recombinant human anti-SARS-CoV-antibodies CR03-014 and CR03-022 were tested for binding to the 15-mer linear and looped/cyclic peptides synthesized as described supra. Relevant binding of a peptide to a recombinant human anti-SARS-CoV antibody was calculated as follows. The average OD-value for each antibody was calculated for the respective proteins (sum of OD-values of all peptides/total number of peptides). Next, the standard deviation of this average was calculated. The standard deviation was multiplied by 2 and the obtained value was added to the average value to obtain the

correction factor. The OD-value of each peptide was then divided by this correction factor. If a value of 0.9 or higher was found, then relevant binding was considered to be present between the specific peptide and the respective antibody. Particularly interesting appear to be domains comprising several reactive peptides, *i.e.* domains comprising consecutive peptides, wherein at least most of the peptides in the domains are reactive with the antibody.

Monoclonal antibody CR03-014 did not appear to react specifically with a peptide or domains comprising several peptides within the SARS-CoV spike protein indicating that CR03-014 may recognize a discontinuous non-linear epitope. Monoclonal antibody CR03-022 reacted with a series of looped peptides in two domains (data not shown). The domains are comprised of amino acid residues 430-449 and 484-503 of the SARS-CoV spike protein and have the amino acid sequences ATSTGNYNYKYRYLRHGKLR (SEQ ID NO:126) and YTTTGIGYQPYRVVVLSFEL (SEQ ID NO:127), respectively. Strikingly, both domains have the motif TXTGXXXXXYR (SEQ ID NO:128) in common, indicating that this motif may be crucial for the binding of antibody CR03-022 to the SARS-CoV spike protein.

Table 1. List of currently known SARS-CoV genome sequence and spike genes.

SARS Isolate	Gene/genome	Genbank	FASTA
SARS coronavirus AS	SARS coronavirus AS, complete genome.	AY427439	37576845
SARS coronavirus BJ01	SARS coronavirus BJ01, complete genome.	AY278488	30275666
SARS coronavirus BJ02	SARS coronavirus BJ02, complete genome.	AY278487	31416292
SARS coronavirus BJ03	SARS coronavirus BJ03, complete genome.	AY278490	31416305
SARS coronavirus BJ04	SARS coronavirus BJ04, complete genome.	AY279354	31416306
SARS coronavirus BJ2232			
SARS coronavirus BJ302			
SARS coronavirus CUHK-AG01	SARS coronavirus CUHK-AG01, complete genome.	AY345986	33114190
SARS coronavirus CUHK-AG02	SARS coronavirus CUHK-AG02, complete genome.	AY345987	33114202
SARS coronavirus CUHK-AG03	SARS coronavirus CUHK-AG03, complete genome.	AY345988	33114214
SARS coronavirus CUHK-L2			
SARS coronavirus CUHK-Su10	SARS coronavirus CUHK-Su10, complete genome.	AY282752	38304867
SARS coronavirus CUHK-W1	SARS coronavirus CUHK-W1, complete genome.	AY276554	30027610
SARS coronavirus cw037			
SARS coronavirus cw049			
SARS coronavirus FRA	SARS coronavirus FRA, complete genome.	AY310120	33578015
SARS coronavirus Frankfurt 1	SARS coronavirus Frankfurt 1, complete genome.	AY291315	31581502
SARS coronavirus GD01	SARS coronavirus GD01, complete genome.	AY278489	31416290
SARS coronavirus GD03T0013	SARS coronavirus GD03T0013 spike glycoprotein gene, complete cds.	AY525636	41764105
SARS coronavirus GD69	SARS coronavirus GD69, complete genome.	AY313906	37960831
SARS coronavirus GZ-A	SARS coronavirus GZ-A, partial genome.	AY394977	37624320
SARS coronavirus GZ-B	SARS coronavirus GZ-B, complete genome.	AY394978	37624321
SARS coronavirus GZ-C	SARS coronevirus GZ-C, complete genome.	AY394979	37624322
SARS coronavirus GZ-D	SARS coronavirus GZ-D, partial genome.	AY394980	37624323
SARS coronavirus GZ02	SARS coronavirus GZ02, complete genome.	AY390556	41323719
SARS coronavirus GZ43	SARS coronavirus GZ43, partial genome.	AY304490	34482141
SARS coronavirus GZ50	SARS coronavirus GZ50, complete genome.	AY304495	34482146
SARS coronavirus GZ60	SARS coronavirus GZ60, partiel genome.	AY304491	34482142
SARS coronavirus HB			
SARS coronavirus HGZ8L1-A	SARS coronavirus HGZ8L1-A, partial genome.	AY394981	37624324
SARS coronavirus HGZ8L1-B	SARS coronavirus HGZ8L1-B, partial genome.	AY394982	37624325
SARS coronavirus HGZ8L2	SARS coronavirus HGZ8L2, complete genome.	AY394993	37624336
SARS coronavirus HKU-36871	SARS coronavirus HKU-36871, partial genome.	AY304492	34482143
SARS coronavirus HKU-39849	SARS coronavirus HKU-39849, complete genome.	AY278491	30023963
SARS coronavirus HKU-65806	SARS coronavirus HKU-65806, partial genome.	AY304493	34482144
SARS coronavirus HKU-66078	SARS coronavirus HKU-66076, partial genome.	AY304494	34482145
SARS coronavirus Hong Kong/03/2003			

SARS coronavirus HPZ-2003			
SARS coronavirus HSR 1	SARS coronavirus HSR 1, complete genome.	AY323977	33115118
SARS coronavirus HSZ-A	SARS coronavirus HSZ-A, partial genome.	AY394984	37624327
SARS coronavirus HSZ-Bb	SARS coronavirus HSZ-Bb, complete genome.	AY394985	37624328
SARS coronavirus HSZ-Bc	SARS coronavirus HSZ-Bc, complete genome.	AY394994	37624337
SARS coronavirus HSZ-Cb	SARS coronavirus HSZ-Cb, complete genome.	AY394986	37624329
SARS coronavirus HSZ-Cc	SARS coronavirus HSZ-Cc, complete genome.	AY394995	37624338
SARS coronavirus HSZ2-A	SARS coronavirus HSZ2-A, complete genome.	AY394983	37624326
SARS coronavirus HZS2-Bb	SARS coronavirus HZS2-Bb, partial genome.	AY395004	37624347
SARS coronavirus HZS2-C	SARS coronavirus HZS2-C, complete genome.	AY394992	37624335
SARS coronavirus HZS2-D	SARS coronavirus HZS2-D, complete genome.	AY394989	37624332
SARS coronavirus HZS2-E	SARS coronavirus HZS2-E, complete genome.	AY394990	37624333
SARS coronavirus HZS2-Fb	SARS coronavirus HZS2-Fb, complete genome.	AY394987	37624330
SARS coronavirus HZS2-Fc	SARS coronavirus HZS2-Fc, complete genome.	AY394991	37624334
SARS coronavirus JMD	SARS coronavirus JMD, partial genome.	AY394988	37624331
SARS coronavirus LC1	SARS coronavirus LC1, complete genome.	AY394998	37624341
SARS coronavirus LC2	SARS coronavirus LC2, complete genome.	AY394999	37624342
SARS coronavirus LC3	SARS coronavirus LC3, complete genome.	AY395000	37624343
SARS coronavirus LC4	SARS coronavirus LC4, complete genome.	AY395001	37624344
SARS coronavirus LC5	SARS coronavirus LC5, complete genome.	AY395002	37624345
SARS coronavirus NS-1	SARS coronavirus NS-1, complete genome.	AY508724	40795744
SARS coronavirus PUMC01	SARS coronavirus PUMC01, complete genome.	AY350750	38231927
SARS coronavirus PUMC02	SARS coronavirus PUMC02, complete genome.	AY357075	38231932
SARS coronavirus PUMC03	SARS coronavirus PUMC03, complete genome.	AY357076	38231937
SARS coronavirus sf098			
SARS coronavirus sf099			
SARS coronavirus ShanghaiQXC1	SARS coronavirus ShanghaiQXC1, complete genome.	AY463059	40457433
SARS coronavirus ShanghaiQXC2	SARS coronavirus ShanghaiQXC2, complete genome.	AY463060	40457448
SARS coronavirus Shanhgai LY	SARS coronavirus Shanhgai LY spike glycoprotein gene, complete cds.	AY322205S3	32454341
SARS coronavirus Sin0409			
SARS coronavirus Sin2500	SARS coronavirus Sin2500, complete genome.	AY283794	30468042
SARS coronavirus Sin2677	SARS coronavirus Sin2677, complete genome.	AY283795	30468043
SARS coronavirus Sin2679	SARS coronavirus Sin2679, complete genome.	AY283796	30468044
SARS coronavirus Sin2748	SARS coronavirus Sin2748, complete genome.	AY283797	30468045
SARS coronavirus Sin2774	SARS coronavirus Sin2774, complete genome.	AY283798	37361915
SARS coronavirus Sin3408	SARS coronavirus Sin3408, complete genome	AY559083	45644998
SARS coronavirus Sin3408L	SARS coronavirus Sin3408L, complete genome	AY559097	45645024
SARS coronavirus Sin3725V	SARS coronavirus Sin3725V, complete genome	AY559087	45645004
SARS coronavirus Sin3765V	SARS coronavirus Sin3765V, complete genome	AY559084	45645000
SARS coronavirus Sin842	SARS coronavirus Sin842, complete genome	AY559081	45644994

SARS coronavirus Sin845	SARS coronavirus Sin845, complete genome	AY559093	45645019
SARS coronavirus Sin846	SARS coronavirus Sin846, complete genome	AY559094	45645021
SARS coronavirus Sin847	SARS coronavirus Sin847, complete genome	AY559095	45645022
SARS coronavirus Sin848	SARS coronavirus Sin848, complete genome	AY559085	45645001
SARS coronavirus Sin849	SARS coronavirus Sin849, complete genome	AY559086	45645003
SARS coronavirus Sin850	SARS coronavirus Sin850, complete genome	AY559096	45645023
SARS coronavirus Sin852	SARS coronavirus Sin852, complete genome	AY559082	45644996
SARS coronavirus Sin_WNV			
SARS coronavirus Sino1-11	SARS coronavirus Sino1-11, complete genome.	AY485277	38505482
SARS coronavirus Sino3-11	SARS coronavirus Sino3-11, complete genome.	AY485278	38505491
SARS coronavirus SinP1	SARS coronavirus SinP1, complete genome	AY559088	45645007
SARS coronavirus SinP2	SARS coronavirus SinP2, complete genome	AY559089	45645010
SARS coronavirus SinP3	SARS coronavirus SinP3, complete genome	AY559090	45645013
SARS coronavirus SinP4	SARS coronavirus SinP4, complete genome	AY559091	45645016
SARS coronavirus SinP5	SARS coronavirus SinP5, complete genome	AY559092	45645017
SARS coronavirus SoD	SARS coronavirus SoD, complete genome.	AY461660	38385714
SARS coronavirus SZ1	SARS coronavirus SZ1, partial genome.	AY304489	34482140
SARS coronavirus SZ13	SARS coronavirus SZ13, partial genome.	AY304487	34482138
SARS coronavirus SZ16	SARS coronavirus SZ16, complete genome.	AY304488	34482139
SARS coronavirus SZ3	SARS coronavirus SZ3, complete genome.	AY304486	34482137
SARS coronavirus Taiwan			
SARS coronavirus Taiwan JC-2003			
SARS coronavirus Taiwan TC1	SARS coronavirus Taiwan TC1, complete genome.	AY338174	32493129
SARS coronavirus Taiwan TC2	SARS coronavirus Taiwan TC2, complete genome.	AY338175	32493130
SARS coronavirus Taiwan TC3	SARS coronavirus Taiwan TC3, complete genome.	AY348314	33188324
SARS coronavirus Tor2	SARS coronavirus TOR2, complete genome.	AY274119	30248028
SARS coronavirus TW			
SARS coronavirus TW-GD1			
SARS coronavirus TW-GD2			
SARS coronavirus TW-GD3			
SARS coronavirus TW-GD4			
SARS coronavirus TW-GD5			
SARS coronavirus TW-HP1			
SARS coronavirus TW-HP2			
SARS coronavirus TW-HP3			
SARS coronavirus TW-HP4			
SARS coronavirus TW-JC2			
SARS coronavirus TW-KC1			
SARS coronavirus TW-KC3			
SARS coronavirus TW-PH1			
SARS coronavirus TW-PH2			

SARS coronavirus TW-YM1			
SARS coronavirus TW-YM2			
SARS coronavirus TW-YM3			
SARS coronavirus TW-YM4			
SARS coronavirus TW1	SARS coronavirus TW1, complete genome.	AY291451	30698326
SARS coronavirus TW10	SARS coronavirus TW10, complete genome.	AY502923	40548873
SARS coronavirus TW11	SARS coronavirus TW11, complete genome.	AY502924	40548885
SARS coronavirus TW2	SARS coronavirus TW2, complete genome.	AY502925	40548897
SARS coronavirus TW3	SARS coronavirus TW3, complete genome.	AY502926	40548909
SARS coronavirus TW4	SARS coronavirus TW4, complete genome.	AY502927	40548921
SARS coronavirus TW5	SARS coronavirus TW5, complete genome.	AY502928	40548933
SARS coronavirus TW6	SARS coronavirus TW6, complete genome.	AY502929	40548945
SARS coronavirus TW7	SARS coronavirus TW7, complete genome.	AY502930	40548957
SARS coronavirus TW8	SARS coronavirus TW8, complete genome.	AY502931	40548969
SARS coronavirus TW9	SARS coronavirus TW9, complete genome.	AY502932	40548981
SARS coronavirus TWC	SARS coronavirus TWC, complete genome.	AY321118	31873092
SARS coronavirus TWC2	SARS coronavirus TWC2, complete genome.	AY382698	33518724
SARS coronavirus TWC3	SARS coronavirus TWC3, complete genome.	AY362699	33518725
SARS coronavirus TWH	SARS coronavirus TWH genomic RNA, complete genome.	AP006557	33411399
SARS coronavirus TWJ	SARS coronavirus TWJ genomic RNA, complete genome.	AP006558	33411414
SARS coronavirus TWK	SARS coronavirus TWK genomic RNA, complete genome.	AP006559	33411429
SARS coronavirus TWS	SARS coronavirus TWS genomic RNA, complete genome.	AP006560	33411444
SARS coronavirus TWY	SARS coronavirus TWY genomic RNA, complete genome.	AP006561	33411459
SARS coronavirus Urbani	SARS coronavirus Urbani, complete genome.	AY278741	30027617
SARS coronavirus Vietnam			
SARS coronavirus WHU	SARS coronavirus WHU, complete genome.	AY394850	40795428
SARS coronavirus xw002			
SARS coronavirus ZJ01	SARS coronavirus ZJ01, complete genome.	AY297028	30910859
SARS coronavirus ZMY 1	SARS coronavirus ZMY 1, complete genome.	AY351680	33304219
SARS coronavirus ZS-A	SARS coronavirus ZS-A, complete genome.	AY394997	37624340
SARS coronavirus ZS-B	SARS coronavirus ZS-B, complete genome.	AY394996	37624339
SARS coronavirus ZS-C	SARS coronavirus ZS-C, complete genome.	AY395003	37624346
	SARS coronavirus, TOR2 complete genome, curated	NC_004718	30271926
	SARS coronavirus ZJ01, partial genome.	AY286320	39980888
	SARS coronavirus BJ302 clone 1 spike glycoprotein gene, complete	AY429072	38016580
	SARS coronavirus BJ302 clone 2 spike glycoprotein gene, complete	AY429073	38016582
	SARS coronavirus BJ302 clone 3 spike glycoprotein gene, complete	AY429074	38016584
	SARS coronavirus BJ302 clone 4 spike glycoprotein gene, complete	AY429075	38016586
	SARS coronavirus BJ302 clone 5 spike glycoprotein gene, complete	AY429076	38016588
	SARS coronavirus BJ302 clone 6 spike glycoprotein gene, complete	AY429077	38016590

	SARS coronavirus BJ302 clone 7 spike glycoprotein gene, complete	AY429078	38016592
	SARS coronavirus BJ302 clone 8 spike glycoprotein gene, complete	AY429079	38016594

Table 2: Human IgG heavy chain variable region primers (sense).

Primer name	Primer nucleotide sequence	SEQ ID NO
HuVH1B/7A	5′-CAGRTGCAGCTGGTG CARTCTGG-3′	SEQ ID NO:11
HuVH1C	5′-SAGGTCCAGCTGGTR CAGTCTGG-3′	SEQ ID NO:12
HuVH2B	5′-SAGGTGCAGCTGGTG GAGTCTGG-3′	SEQ ID NO:13
HuVH3B	5′-SAGGTGCAGCTGGTG GAGTCTGG-3'	SEQ ID NO:14
HuVH3C	5′-GAGGTGCAGCTGGTG GAGWCYGG-3′	SEQ ID NO:15
HuVH4B	5′-CAGGTGCAGCTACAG CAGTGGGG-3′	SEQ ID NO:16
HuVH4C	5′-CAGSTGCAGCTGCAG GAGTCSGG-3′	SEQ ID NO:17
HuVH5B	5′-GARGTGCAGCTGGTG CAGTCTGG-3′	SEQ ID NO:18
HuVH6A	5′-CAGGTACAGCTGCAG CAGTCAGG-3′	SEQ ID NO:19

Table 3: Human IgG heavy chain J-region primers (anti-sense).

Primer name	Primer nucleotide sequence	SEQ ID NO
HuJH1/2	5′-TGAGGAGACGGTGAC CAGGGTGCC-3′	SEQ ID NO:20
HuJH3	5′-TGAAGAGACGGTGAC CATTGTCCC-3′	SEQ ID NO:21
HuJH4/5	5′-TGAGGAGACGGTGAC CAGGGTTCC-3′	SEQ ID NO:22
HuJH6	5′-TGAGGAGACGGTGAC CGTGGTCCC-3′	SEQ ID NO:23

Table 4: Human IgG heavy chain variable region primers extended with SfiI/NcoI restriction sites (sense) and human IgG heavy chain J-region primers extended with XhoI/BstEII restriction sites (anti-sense).

Primer name	Primer nucleotide sequence	SEQ ID NO
HuVH1B/7A-NcoI	5'-GTCCTCGCAACTGCG GCCCAGCCGGCCATGGCC CAGRTGCAGCTGGTGCAR TCTGG-3'	SEQ ID NO:24
HuVH1C-NcoI	5'-GTCCTCGCAACTGCG GCCCAGCCGGCCATGGCC SAGGTCCAGCTGGTRCAG TCTGG-3'	SEQ ID NO:25
HuVH2B-NcoI	5'-GTCCTCGCAACTGCG GCCCAGCCGGCCATGGCC CAGRTCACCTTGAAGGAG TCTGG-3'	SEQ ID NO:26

HuVH3B-NcoI	5'-GTCCTCGCAACTGCGGCC CAGCCGGCCATGGCCSAGGTG CAGCTGGTGGAGTCTGG-3'	SEQ ID NO:27
HuVH3C-NcoI	5'-GTCCTCGCAACTGCG GCCCAGCCGGCCATGGCC GAGGTGCAGCTGGTGGAG WCYGG-3'	SEQ ID NO:28
HuVH4B-NcoI	5'-GTCCTCGCAACTGCG GCCCAGCCGGCCATGGCC CAGGTGCAGCTACAGCAG TGGGG-3'	SEQ ID NO:29
HuVH4C-NcoI	5'-GTCCTCGCAACTGCGGCC CAGCCGGCCATGGCCCAGSTG CAGCTGCAGGAGTCSGG-3'	SEQ ID NO:30
HuVH5B-NcoI	5'-GTCCTCGCAACTGCG GCCCAGCCGGCCATGGCC GARGTGCAGCTGGTGCAG TCTGG-3'	SEQ ID NO:31
HuVH6A-NcoI	5'-GTCCTCGCAACTGCG GCCCAGCCGGCCATGGCC CAGGTACAGCTGCAGCAG TCAGG-3'	SEQ ID NO:32
HuJH1/2-XhoI	5′-GAGTCATTCTCGACTCGA GACGGTGACCAGGGTGCC-3′	SEQ ID NO:33
HuJH3-XhoI	5′-GAGTCATTCTCGACT CGAGACGGTGACCATTGT CCC-3′	SEQ ID NO:34
HuJH4/5-XhoI	5′-GAGTCATTCTCGACT CGAGACGGTGACCAGGGT TCC-3′	SEQ ID NO:35
HuJH6-XhoI	5′-GAGTCATTCTCGACTCGA	SEQ ID NO:36

	GACGGTGACCGTGGTCCC-3′	

Table 5: Human lambda chain variable region primers (sense).

Primer name	Primer nucleotide sequence	SEQ ID NO
HuVλ1A	5′-CAGTCTGTGCTGACT CAGCCACC-3′	SEQ ID NO:37
HuVλ1B	5′-CAGTCTGTGYTGACG CAGCCGCC-3′	SEQ ID NO:38
HuVλ1C	5′-CAGTCTGTCGTGACG CAGCCGCC-3′	SEQ ID NO:39
HuVλ2	5′-CARTCTGCCCTGACT CAGCCT-3′	SEQ ID NO:40
HuVλ3A	5′-TCCTATGWGCTGACT CAGCCACC-3′	SEQ ID NO:41
HuVλ3B	5′-TCTTCTGAGCTGACT CAGGACCC-3′	SEQ ID NO:42
HuVλ4	5′-CACGTTATACTGACT CAACCGCC-3′	SEQ ID NO:43
HuVλ5	5′-CAGGCTGTGCTGACT CAGCCGTC-3′	SEQ ID NO:44
HuVλ6	5′-AATTTTATGCTGACT CAGCCCCA-3′	SEQ ID NO:45
HuVλ7/8	5′-CAGRCTGTGGTGACY CAGGAGCC-3′	SEQ ID NO:46
HuVλ9	5′-CWGCCTGTGCTGACT CAGCCMCC-3′	SEQ ID NO:47

Table 6: Human kappa chain variable region primers (sense).

Primer name	Primer nucleotide sequence	SEQ ID NO
HuVκ1B	5′-GACATCCAGWTGACCC AGTCTCC-3′	SEQ ID NO:48
HuVκ2	5′-GATGTTGTGATGACT CAGTCTCC-3′	SEQ ID NO:49
HuVκ3	5′-GAAATTGTGWTGACR CAGTCTCC-3′	SEQ ID NO:50
HuVκ4	5′-GATATTGTGATGACC CACACTCC-3′	SEQ ID NO:51
HuVκ5	5′-GAAACGACACTCACG CAGTCTCC-3′	SEQ ID NO:52
HuVκ6	5′-GAAATTGTGCTGACTC AGTCTCC-3′	SEQ ID NO:53

Table 7: Human lambda chain J-region primers (anti-sense).

Primer name	Primer nucleotide sequence	SEQ ID NO
HuJλ1	5′-ACCTAGGACGGTGACC TTGGTCCC-3′	SEQ ID NO:54
HuJλ2/3	5′-ACCTAGGACGGTCAG CTTGGTCCC-3′	SEQ ID NO:55
HuJλ4/5	5′-ACYTAAAACGGTGAG CTGGGTCCC-3′	SEQ ID NO:56

Table 8: Human lambda chain J-region primers (anti-sense).

Primer name	Primer nucleotide sequence	SEQ ID NO

HuJκ1	5′-ACGTTTGATTTCCAC CTTGGTCCC-3′	SEQ ID NO:57
HuJκ2	5′-ACGTTTGATCTCCAG CTTGGTCCC-3′	SEQ ID NO:58
HuJκ3	5′-ACGTTTGATATCCAC TTTGGTCCC-3′	SEQ ID NO:59
HuJκ4	5′-ACGTTTGATCTCCAC CTTGGTCCC-3′	SEQ ID NO:60
HuJκ5	5′-ACGTTTAATCTCCAG TCGTGTCCC-3′	SEQ ID NO:61

Table 9: Human kappa chain variable region primers extended with SalI restriction sites (sense), human kappa chain J-region primers extended with NotI restriction sites (anti-sense), human lambda chain variable region primers extended with SalI restriction sites (sense) and human lambda chain J-region primers extended with NotI restriction sites (anti-sense).

Primer name	Primer nucleotide sequence	SEQ ID NO
HuVκ1B-SalI	5′-TGAGCACACAGGTCG ACGGACATCCAGWTGACC CAGTCTCC-3′	SEQ ID NO:62
HuVκ2-SalI	5′-TGAGCACACAGGTCG ACGGATGTTGTGATGACT CAGTCTCC-3′	SEQ ID NO:63
HuVκ3B-SalI	5′-TGAGCACACAGGTCG ACGGAAATTGTGWTGACR CAGTCTCC-3′	SEQ ID NO:64
HuVκ4B-SalI	5′-TGAGCACACAGGTCG	SEQ ID NO:65

	ACGGATATTGTGATGACC CACACTCC-3′	
HuVκ5-SalI	5′-TGAGCACACAGGTCGACG GAAACGACACTCACGCAGTCT CC-3′	SEQ ID NO:66
HuVκ6-SalI	5′-TGAGCACACAGGTCG ACGGAAATTGTGCTGACT CAGTCTCC-3′	SEQ ID NO:67
HuJκ1-NotI	5′-GAGTCATTCTCGACTTGC GGCCGCACGTTTGATTTCCAC CTTGGTCCC-3′	SEQ ID NO:68
HuJκ2-NotI	5′-GAGTCATTCTCGACT TGCGGCCGCACGTTTGAT CTCCAGCTTGGTCCC-3′	SEQ ID NO:69
HuJκ3-NotI	5′-GAGTCATTCTCGACTTGC GGCCGCACGTTTGATATCCAC TTTGGTCCC-3′	SEQ ID NO:70
HuJκ4-NotI	5′-GAGTCATTCTCGACT TGCGGCCGCACGTTTGAT CTCCACCTTGGTCCC-3′	SEQ ID NO:71
HuJκ5-NotI	5′-GAGTCATTCTCGACTTGC GGCCGCACGTTTAATCTCCAG TCGTGTCCC-3′	SEQ ID NO:72
HuVλ1A-SalI	5′-TGAGCACACAGGTCGACG CAGTCTGTGCTGACTCAGCCA CC-3′	SEQ ID NO:73
HuVλ1B-SalI	5′-TGAGCACACAGGTCGACG CAGTCTGTGYTGACGCAGCCG CC-3′	SEQ ID NO:74
HuVλ1C-SalI	5′-TGAGCACACAGGTCGACG CAGTCTGTCGTGACGCAGCCG	SEQ ID NO:75

	CC-3′	
HuVλ2-SalI	5′-TGAGCACACAGGTCGACG CARTCTGCCCTGACTCAGCCT-3′	SEQ ID NO:76
HuVλ3A-SalI	5′-TGAGCACACAGGTCGACG TCCTATGWGCTGACTCAGCCA CC-3′	SEQ ID NO:77
HuVλ3B-SalI	5′-TGAGCACACAGGTCGACG TCTTCTGAGCTGACTCAGGAC CC-3′	SEQ ID NO:78
HuVλ4-SalI	5′-TGAGCACACAGGTCGACG CACGTTATACTGACTCAACCG CC-3′	SEQ ID NO:79
HuVλ5-SalI	5′-TGAGCACACAGGTCGACG CAGGCTGTGCTGACTCAGCCG TC-3′	SEQ ID NO:80
HuVλ6-SalI	5′-TGAGCACACAGGTCGACG AATTTTATGCTGACTCAGCCC CA-3′	SEQ ID NO:81
HuVλ7/8-SalI	5′-TGAGCACACAGGTCGACG CAGRCTGTGGTGACYCAGGAG CC-3′	SEQ ID NO:82
HuVλ9-SalI	5′-TGAGCACACAGGTCGACG CWGCCTGTGCTGACTCAGCCM CC-3′	SEQ ID NO:83
HuJλ1-NotI	5′-GAGTCATTCTCGACTTGC GGCCGCACCTAGGACGGTGAC CTTGGTCCC-3′	SEQ ID NO:84
HuJλ2/3-NotI	5′-GAGTCATTCTCGACTTGC GGCCGCACCTAGGACGGTCAG CTTGGTCCC-3′	SEQ ID NO:85

HuJλ4/5-NotI	5'-GAGTCATTCTCGACTTGC GGCCGCACYTAAAACGGTGAG CTGGGTCCC-3'	SEQ ID NO:86

Table 10: Distribution of the different light chain products over the 10 fractions.

Light chain products	Number of alleles	Fraction number	alleles/fraction
Vk1B/Jk1-5	19	1 and 2	9.5
Vk2/Jk1-5	9	3	9
Vk3B/Jk1-5	7	4	7
Vk4B/Jk1-5	1	5	5
Vk5/Jk1-5	1		
Vk6/Jk1-5	3		
Vλ1A/Jl1-3	5	6	5
Vλ1B/Jl1-3			
Vλ1C/Jl1-3			
Vλ2/Jl1-3	5	7	5
Vλ3A/Jl1-3	9	8	9
Vλ3B/Jl1-3			
Vλ4/Jl1-3	3	9	5
Vλ5/Jl1-3	1		
Vλ6/Jl1-3	1		
Vλ7/8/Jl1-3	3	10	6
Vλ9/Jl1-3	3		

Table 11: Data of the single-chain Fvs capable of binding SARS-CoV.

Name scFv	SEQ ID NO of nucleo-tide sequence	SEQ ID NO of amino acid sequence	HCDR3	VH-germline	VL-germline
SC03-014	92	93	GISPFYFDY (SEQ ID NO:1)	V_H3 DP29 (V_H3-72)	V_kI DPK9 (02/012)
SC03-022	94	95	GSGISTPMDV (SEQ ID NO:2)	V_H5-51 (DP-73)	V_KIV (B3-DPK24)

Table 12A: SARS-CoV neutralization experiment I.

	Antibody	CR03-014			CR03-022			cIgG1			Convalescent serum		
dilution		1	2	3	4	5	6	7	8	9	10	11	12
4 x	A	-	-	-	-	-	-	+	+	+	-	-	-
8 x	B	-	-	-	+	+	+	+	+	+	-	-	-
16 x	C	-	-	-	-	-	-	+	+	+	-	-	-
32 x	D	-	-	-	-	-	+	+	+	+	-	-	-
64 x	E	-	-	-	-	-	-	+	+	+	-	-	-
128 x	F	-	-	-	+	+	+	+	+	+	-	-	-
256 x	G	-	-	-	+	+	+	+	+	+	+	+	+
512 x	H	+	+	+	+	+	+	+	+	+	+	+	+

Table 12B: SARS-CoV neutralization experiment II.

	Antibody	CR03-014			CR03-022			cIgG1			Convalescent serum		
dilution		1	2	3	4	5	6	7	8	9	10	11	12
4 x	A	-	-	-	-	-	+	+	+	+	-	-	-
8 x	B	-	-	-	+	+	-	+	+	+	-	-	-
16 x	C	-	-	-	+	-	-	+	+	+	-	-	-
32 x	D	-	-	-	+	+	-	+	+	+	-	-	-
64 x	E	-	+	-	-	-	-	+	+	+	-	-	-
128 x	F	-	-	+	+	+	+	+	+	+	+	-	-
256 x	G	-	-	-	+	+	+	+	+	+	+	+	+
512 x	H	+	+	+	+	+	+	+	+	+	+	+	+

Table 12C: SARS-CoV neutralization experiment III.

	Antibody	CR03-014			CR03-022			cIgG1			Convalescent serum		
dilution		1	2	3	4	5	6	7	8	9	10	11	12
4 x	A	-	-	-	-	-	-	+	+	+	-	-	-
8 x	B	-	-	-	-	-	-	+	+	+	-	-	-
16 x	C	-	-	-	-	-	-	+	+	+	-	-	-
32 x	D	-	-	-	-	-	-	+	+	+	-	-	-
64 x	E	-	-	-	-	+	+	+	+	+	-	-	-
128 x	F	-	-	-	+	+	+	+	+	+	-	+	-
256 x	G	+	+	+	+	+	+	+	+	+	+	+	+
512 x	H	+	+	+	+	+	+	+	+	+	+	+	+

-: No CPE

+: CPE

Table 13: Concentrations of the monoclonal anti-SARS-CoV antibody CR03-014 giving complete protection against 100 $TCID_{50}$ of the different SARS-CoV isolates indicated in an *in vitro* neutralization assay.

SARS-CoV Strain*	Concentration of CR03-014 (µg/ml) resulting in 100% protection against 100 $TCID_{50}$
36 (3)	12.5
39849 (3)	12.5
66 (2)	12.5
61567 (2)	12.5
61644 (1)	12.5
61644 (15)	12.5
GZ43[1]	12.5
GZ50[1]	12.5

* Between brackets the passage numbers of the respective strains is indicated. [1] For strains GZ43 and GZ50 the passage history is unknown.

Table 14: SARS-CoV neutralization experiment of several mixtures of CR03-014 and CR03-022.

Mix	CR03-014 (625 µg/ml) %	CR03-022 (2.5 mg/ml) %	Wells protected at indicated dilution							Titer
			16x	32x	64x	128x	256x	512x	1024x	

A	0	100	6/6	6/6	5/6	3/6	3/6	0/6	0/6	64
B	10	90	6/6	6/6	6/6	6/6	6/6	2/6	0/6	256
C	50	50	6/6	6/6	6/6	6/6	6/6	6/6	4/6	>1024
D	90	10	ND	6/6	6/6	6/6	4/6	1/6	0/6	256
E	100	0	5/6	2/6	0/6	0/6	0/6	0/6	0/6	16

Table 15: List of SARS-CoV strains having a region 318-510 of the S protein not identical to the region 318-510 of the S protein of SARS-CoV Frankfurt 1 strain.

Mutation	Strain	Genbank
K344R	GZ02	AY390556
	GZ60	AY304491
	JMD	AY394988
	ZS-B	AY394996
	GZ43	AY304490
	HGZ8L1-A	AY394981
	ZS-A	AY394997
	ZS-C	AY395003
K344R F501Y	GD01	AY278489
K344R F360S L472P D480G T487S	GD03T0013	AY525636
S353F	Sin3408	AY559083
	Sin3765V	AY559084
	Sin845	AY559093
	Sin847	AY559095

	Sin849	AY559086
	Sin852	AY559082
	Sin3725V	AY559087
	Sin842	AY559081
	Sin846	AY559094
	Sin848	AY559085
	Sin850	AY559096
R426G N437D	Shanghai LY	AY322205S3
Y436H	GZ-C	AY394979
Y442S	Sino1-11	AY485277
N479S	BJ302 cl. 2	AY429073
	BJ302 cl. 4	AY429075
	BJ302 cl. 6	AY429077
	BJ302 cl. 3	AY429074
	BJ302 cl. 5	AY429076
	BJ302 cl. 8	AY429079

The amino acid substitutions compared to the Frankfurt 1 S protein are indicated in the left column. Strain and GenBank accession number are indicated in second and third column.

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Table 16. Neutralization Index of E014 escape viruses

Escape virus	Log($TCID_{50}$/ml) without mAb	Log($TCID_{50}$/ml) with mAb	NI
E014-C06	7.45	7.53	0
E014-C07	7.07	6.85	0.22
E014-C08	7.83	7.1	0.73
E014-C09	7.53	6.85	0.68
E014-C10	6.77	7.1	0
Wt SARS-	7.23	3.93	3.3

CoV			

Table 17. Neutralization index of putative E022 escape viruses

Virus	Log($TCID_{50}$/ml) without mAb	Log($TCID_{50}$/ml) with mAb	NI	Escape
Wt SARS-CoV	7.3	4	3.3	No
E22-2	7.3	2.5	4.8	No
E22-3	7.45	2.5	4.95	No
E22-5	7.3	2.7	4.6	No
E22-6	7.3	2.95	4.35	No
E22-9	8.1	3.7	4.4	No

Table 18. CR03-022 neutralizing titers for two representative escape viruses of CR03-014

Virus	Titer (μg/ml)[a]	Titer (μg/ml)[b]
Wild-type	35.46	50.00
E14-6	25.00	50.00
E14-7	31.50	50.00

[a] 50% neutralizing titer of CR03-022 for each virus is indicated in μg/ml

[b] 100% neutralizing titer of CR03-022 for each virus is indicated in μg/ml

SEQUENCE LISTING

<110> Crucell Holland B.V.
Ter Meulen, Jan H.
Goudsmit, Jaap
Van den Brink, Edward N.
De Kruif, Cornelis A.

<120> Compositions against SARS-coronavirus and uses thereof

<130> 0114 WO 00 ORD

<160> 138

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gga ggg tcc ctg aga ctc tcc tgt gca gcc tct gga ttc acc ttc agt       96
Gly Gly Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser
            20                  25                  30

gac cac tac atg gac tgg gtc cgc cag gct cca ggg aag ggg ctg gag      144
Asp His Tyr Met Asp Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu
        35                  40                  45

tgg gtt ggc cgt act aga aac aaa gct aac agt tac acc aca gaa tac      192
Trp Val Gly Arg Thr Arg Asn Lys Ala Asn Ser Tyr Thr Thr Glu Tyr
    50                  55                  60

gcc gcg tct gtg aaa ggc aga ttc acc atc tca aga gat gat tca aag      240
Ala Ala Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser Lys
65                  70                  75                  80

aac tca ctg tat ctg caa atg aac agc ctg aaa acc gag gac acg gcc      288
Asn Ser Leu Tyr Leu Gln Met Asn Ser Leu Lys Thr Glu Asp Thr Ala
                85                  90                  95

gtg tat tac tgt gca agg ggg att tcg ccg ttt tac ttt gac tac tgg      336
Val Tyr Tyr Cys Ala Arg Gly Ile Ser Pro Phe Tyr Phe Asp Tyr Trp
            100                 105                 110
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35 40 45

Trp Val Gly Arg Thr Arg Asn Lys Ala Asn Ser Tyr Thr Thr Glu Tyr
50 55 60

Ala Ala Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser Lys
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Asn Ser Leu Tyr Leu Gln Met Asn Ser Leu Lys Thr Glu Asp Thr Ala
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Val Tyr Tyr Cys Ala Arg Gly Ile Ser Pro Phe Tyr Phe Asp Tyr Trp
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1               5                   10                  15

ggg gag tct ctg aag atc tcc tgt aag ggt tct gga tac ggc ttt atc       96
Gly Glu Ser Leu Lys Ile Ser Cys Lys Gly Ser Gly Tyr Gly Phe Ile
            20                  25                  30

acc tac tgg atc ggc tgg gtg cgc cag atg ccc ggg aaa ggc ctg gag      144
Thr Tyr Trp Ile Gly Trp Val Arg Gln Met Pro Gly Lys Gly Leu Glu
        35                  40                  45

tgg atg ggg atc atc tat cct ggt gac tct gaa acc aga tac agc ccg      192
Trp Met Gly Ile Ile Tyr Pro Gly Asp Ser Glu Thr Arg Tyr Ser Pro
    50                  55                  60

tcc ttc caa ggc cag gtc acc atc tca gcc gac aag tcc atc aac acc      240
Ser Phe Gln Gly Gln Val Thr Ile Ser Ala Asp Lys Ser Ile Asn Thr
65                  70                  75                  80

gcc tac ctg cag tgg agc agc ctg aag gcc tcg gac acc gcc ata tat      288
Ala Tyr Leu Gln Trp Ser Ser Leu Lys Ala Ser Asp Thr Ala Ile Tyr
                85                  90                  95

tac tgt gcg ggg ggt tcg ggg att tct acc cct atg gac gtc tgg ggc      336
Tyr Cys Ala Gly Gly Ser Gly Ile Ser Thr Pro Met Asp Val Trp Gly
            100                 105                 110

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Trp Met Gly Ile Ile Tyr Pro Gly Asp Ser Glu Thr Arg Tyr Ser Pro
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Ser Phe Gln Gly Gln Val Thr Ile Ser Ala Asp Lys Ser Ile Asn Thr
65 70 75 80

Ala Tyr Leu Gln Trp Ser Ser Leu Lys Ala Ser Asp Thr Ala Ile Tyr
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Val Thr Ile Thr Cys Arg Ala Ser Gln Ser Ile Ser Ser Tyr Leu Asn
20 25 30

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Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Leu Ile Tyr Ala
35 40 45

gca tcc agt ttg caa agt ggg gtc cca tca agg ttc agt ggc agt gga 192
Ala Ser Ser Leu Gln Ser Gly Val Pro Ser Arg Phe Ser Gly Ser Gly
50 55 60

tct ggg aca gat ttc act ctc acc atc agc agt ctg caa cct gaa gat 240
Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro Glu Asp
65 70 75 80

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Phe Ala Thr Tyr Tyr Cys Gln Gln Ser Tyr Ser Thr Pro Pro Thr Phe
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35 40 45

Ala Ser Ser Leu Gln Ser Gly Val Pro Ser Arg Phe Ser Gly Ser Gly
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Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro Glu Asp
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Glu Arg Ala Thr Ile Asn Cys Lys Ser Ser Gln Ser Val Leu Tyr Ser
            20                  25                  30

tcc atc aat aag aac tac tta gct tgg tac cag cag aaa cca gga cag      144
Ser Ile Asn Lys Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln
        35                  40                  45

cct cct aag ctg ctc att tac tgg gca tct acc cgg gaa tcc ggg gtc      192
Pro Pro Lys Leu Leu Ile Tyr Trp Ala Ser Thr Arg Glu Ser Gly Val
    50                  55                  60

cct gac cga ttc agt ggc agc ggg tct ggg aca gat ttc act ctc acc      240
Pro Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr
65                  70                  75                  80

atc agc agc ctg cag gct gaa gat gtg gca gtt tat tac tgt cag caa      288
Ile Ser Ser Leu Gln Ala Glu Asp Val Ala Val Tyr Tyr Cys Gln Gln
                85                  90                  95

tat tat agt act ccg tac act ttt ggc cag ggg acc aag gtg gaa atc      336
Tyr Tyr Ser Thr Pro Tyr Thr Phe Gly Gln Gly Thr Lys Val Glu Ile
            100                 105                 110

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Pro Pro Lys Leu Leu Ile Tyr Trp Ala Ser Thr Arg Glu Ser Gly Val
50 55 60

Pro Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr
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Ile Ser Ser Leu Gln Ala Glu Asp Val Ala Val Tyr Tyr Cys Gln Gln
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<223> Primer HuJH1/2

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<223> Primer HuJH4/5

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<223> Primer HuVH1B/7A-NcoI

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<223> Primer HuVH2B-NcoI

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<210> 27

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<223> Primer HuVH3B-NcoI

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<223> Primer HuVH3C-NcoI

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<210> 29

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<223> Primer HuVH4B-NcoI

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<223> Primer HuVH5B-NcoI

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<223> Primer HuVH6A-NcoI

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<223> Primer HuJH1/2-XhoI

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<223> Primer HuVlambda1A

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<223> Primer HuVlambda1B

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<223> Primer HuVlambda1C

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<223> Primer HuVkappa3B-SalI

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cct ggg ggg tcc ctg aga ctc tcc tgt gca gcc tct gga ttc acc ttt       96
Pro Gly Gly Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe
            20                  25                  30
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agc gac tac ttg atg aac tgg gtc cgc cag gcg ccc ggg aag ggg ctg      144
Ser Asp Tyr Leu Met Asn Trp Val Arg Gln Ala Pro Gly Lys Gly Leu
        35                  40                  45

gag tgg gtt ggc cgt att aga agc aaa gct aac agt tac gcg aca gca      192
Glu Trp Val Gly Arg Ile Arg Ser Lys Ala Asn Ser Tyr Ala Thr Ala
    50                  55                  60

tat gct gcg tcg gtg aaa ggc agg ttc acc atc tcc aga gat gat tca      240
Tyr Ala Ala Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser
65                  70                  75                  80

aag aac acg gcg tat ctg caa atg aac agc ctg aaa acc gag gac acg      288
Lys Asn Thr Ala Tyr Leu Gln Met Asn Ser Leu Lys Thr Glu Asp Thr
                85                  90                  95

gcc gtg tat tac tgt gct aaa gac ggc agc cgg ttc ccc gcc cgc ttc      336
Ala Val Tyr Tyr Cys Ala Lys Asp Gly Ser Arg Phe Pro Ala Arg Phe
            100                 105                 110

gat tac tgg ggc cag ggc acc ctg gtg acc gtg ctc gag ggt acc gga      384
Asp Tyr Trp Gly Gln Gly Thr Leu Val Thr Val Leu Glu Gly Thr Gly
        115                 120                 125

ggt tcc ggc gga acc ggg tct ggg act ggt acg agc gag ctc acc cag      432
Gly Ser Gly Gly Thr Gly Ser Gly Thr Gly Thr Ser Glu Leu Thr Gln
    130                 135                 140

tct cca tcc tcc ctg tct gca tct gta gga gac aga gtc acc atc act      480
Ser Pro Ser Ser Leu Ser Ala Ser Val Gly Asp Arg Val Thr Ile Thr
145                 150                 155                 160

tgc cgg gca agt cag agc att agc agc tac tta aat tgg tat cag cag      528
Cys Arg Ala Ser Gln Ser Ile Ser Ser Tyr Leu Asn Trp Tyr Gln Gln
                165                 170                 175

aaa cca ggg aaa gcc cct aag ctc ctg atc tat gct gca tcc agt ttg      576
Lys Pro Gly Lys Ala Pro Lys Leu Leu Ile Tyr Ala Ala Ser Ser Leu
            180                 185                 190

caa agt ggg gtc cca tca agg ttc agt ggc agt gga tct ggg aca gat      624
Gln Ser Gly Val Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp
        195                 200                 205

ttc act ctc acc atc agc agt ctg caa cct gaa gat ttt gca act tac      672
Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro Glu Asp Phe Ala Thr Tyr
    210                 215                 220

tac tgt caa cag agt tac agt acc cct cca acg ttc ggc caa ggg acc      720
Tyr Cys Gln Gln Ser Tyr Ser Thr Pro Pro Thr Phe Gly Gln Gly Thr
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Ser Asp Tyr Leu Met Asn Trp Val Arg Gln Ala Pro Gly Lys Gly Leu
        35                  40                  45

Glu Trp Val Gly Arg Ile Arg Ser Lys Ala Asn Ser Tyr Ala Thr Ala
    50                  55                  60

Tyr Ala Ala Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser
65                  70                  75                  80

Lys Asn Thr Ala Tyr Leu Gln Met Asn Ser Leu Lys Thr Glu Asp Thr
                85                  90                  95

Ala Val Tyr Tyr Cys Ala Lys Asp Gly Ser Arg Phe Pro Ala Arg Phe
            100                 105                 110

Asp Tyr Trp Gly Gln Gly Thr Leu Val Thr Val Leu Glu Gly Thr Gly
        115                 120                 125

Gly Ser Gly Gly Thr Gly Ser Gly Thr Gly Thr Ser Glu Leu Thr Gln
    130                 135                 140

Ser Pro Ser Ser Leu Ser Ala Ser Val Gly Asp Arg Val Thr Ile Thr
145                 150                 155                 160

Cys Arg Ala Ser Gln Ser Ile Ser Ser Tyr Leu Asn Trp Tyr Gln Gln
                165                 170                 175

Lys Pro Gly Lys Ala Pro Lys Leu Leu Ile Tyr Ala Ala Ser Ser Leu
            180                 185                 190

Gln Ser Gly Val Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp
        195                 200                 205

Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro Glu Asp Phe Ala Thr Tyr
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        210                 215                 220

    Tyr Cys Gln Gln Ser Tyr Ser Thr Pro Pro Thr Phe Gly Gln Gly Thr
 5  225                 230                 235                 240

    Lys Val Glu Ile Lys Arg Ala Ala Ala
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    1               5                   10                  15
40
    ccg ggg gag tct ctg aag atc tcc tgt aag ggt tct gga tac ggc ttt       96
    Pro Gly Glu Ser Leu Lys Ile Ser Cys Lys Gly Ser Gly Tyr Gly Phe
                20                  25                  30

45  atc acc tac tgg atc ggc tgg gtg cgc cag atg ccc ggg aaa ggc ctg      144
    Ile Thr Tyr Trp Ile Gly Trp Val Arg Gln Met Pro Gly Lys Gly Leu
            35                  40                  45

    gag tgg atg ggg atc atc tat cct ggt gac tct gaa acc aga tac agc      192
50  Glu Trp Met Gly Ile Ile Tyr Pro Gly Asp Ser Glu Thr Arg Tyr Ser
        50                  55                  60

    ccg tcc ttc caa ggc cag gtc acc atc tca gcc gac aag tcc atc aac      240
    Pro Ser Phe Gln Gly Gln Val Thr Ile Ser Ala Asp Lys Ser Ile Asn
55  65                  70                  75                  80

    acc gcc tac ctg cag tgg agc agc ctg aag gcc tcg gac acc gcc ata      288
    Thr Ala Tyr Leu Gln Trp Ser Ser Leu Lys Ala Ser Asp Thr Ala Ile
                85                  90                  95
60
    tat tac tgt gcg ggg ggt tcg ggg att tct acc cct atg gac gtc tgg      336
    Tyr Tyr Cys Ala Gly Gly Ser Gly Ile Ser Thr Pro Met Asp Val Trp
                100                 105                 110
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ggc caa ggg acc acg gtc acc gtc tcg agc ggt acg ggc ggt tca ggc      384
Gly Gln Gly Thr Thr Val Thr Val Ser Ser Gly Thr Gly Gly Ser Gly
        115                 120                 125

gga acc ggc agc ggc act ggc ggg tcg acg gac atc cag ttg acc cag      432
Gly Thr Gly Ser Gly Thr Gly Gly Ser Thr Asp Ile Gln Leu Thr Gln
    130                 135                 140

tct cca gac tcc ctg gct gtg tct ctg ggc gag agg gcc acc atc aac      480
Ser Pro Asp Ser Leu Ala Val Ser Leu Gly Glu Arg Ala Thr Ile Asn
145                 150                 155                 160

tgc aag tcc agc cag agt gtt tta tac agc tcc atc aat aag aac tac      528
Cys Lys Ser Ser Gln Ser Val Leu Tyr Ser Ser Ile Asn Lys Asn Tyr
                165                 170                 175

tta gct tgg tac cag cag aaa cca gga cag cct cct aag ctg ctc att      576
Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln Pro Pro Lys Leu Leu Ile
            180                 185                 190

tac tgg gca tct acc cgg gaa tcc ggg gtc cct gac cga ttc agt ggc      624
Tyr Trp Ala Ser Thr Arg Glu Ser Gly Val Pro Asp Arg Phe Ser Gly
        195                 200                 205

agc ggg tct ggg aca gat ttc act ctc acc atc agc agc ctg cag gct      672
Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Ala
    210                 215                 220

gaa gat gtg gca gtt tat tac tgt cag caa tat tat agt act ccg tac      720
Glu Asp Val Ala Val Tyr Tyr Cys Gln Gln Tyr Tyr Ser Thr Pro Tyr
225                 230                 235                 240

act ttt ggc cag ggg acc aag gtg gaa atc aaa cgt gcg gcc              762
Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Arg Ala Ala
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            20                  25                  30

Ile Thr Tyr Trp Ile Gly Trp Val Arg Gln Met Pro Gly Lys Gly Leu
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Glu Trp Met Gly Ile Ile Tyr Pro Gly Asp Ser Glu Thr Arg Tyr Ser
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Pro Ser Phe Gln Gly Gln Val Thr Ile Ser Ala Asp Lys Ser Ile Asn
65 70 75 80

Thr Ala Tyr Leu Gln Trp Ser Ser Leu Lys Ala Ser Asp Thr Ala Ile
85 90 95

Tyr Tyr Cys Ala Gly Gly Ser Gly Ile Ser Thr Pro Met Asp Val Trp
100 105 110

Gly Gln Gly Thr Thr Val Thr Val Ser Ser Gly Thr Gly Gly Ser Gly
115 120 125

Gly Thr Gly Ser Gly Thr Gly Gly Ser Thr Asp Ile Gln Leu Thr Gln
130 135 140

Ser Pro Asp Ser Leu Ala Val Ser Leu Gly Glu Arg Ala Thr Ile Asn
145 150 155 160

Cys Lys Ser Ser Gln Ser Val Leu Tyr Ser Ser Ile Asn Lys Asn Tyr
165 170 175

Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln Pro Pro Lys Leu Leu Ile
180 185 190

Tyr Trp Ala Ser Thr Arg Glu Ser Gly Val Pro Asp Arg Phe Ser Gly
195 200 205

Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Ala
210 215 220

Glu Asp Val Ala Val Tyr Tyr Cys Gln Gln Tyr Tyr Ser Thr Pro Tyr
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Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Arg Ala Ala
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<220>

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     tcc ctg aga ctc tcc tgt gca gcc tct gga ttc acc ttc agt gac cac      96
 5   Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Asp His
                 20                  25                  30

     tac atg gac tgg gtc cgc cag gct cca ggg aag ggg ctg gag tgg gtt     144
     Tyr Met Asp Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val
10           35                  40                  45

     ggc cgt act aga aac aaa gct aac agt tac acc aca gaa tac gcc gcg     192
     Gly Arg Thr Arg Asn Lys Ala Asn Ser Tyr Thr Thr Glu Tyr Ala Ala
         50                  55                  60
15
     tct gtg aaa ggc aga ttc acc atc tca aga gat gat tca aag aac tca     240
     Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser Lys Asn Ser
     65                  70                  75                  80

20   ctg tat ctg caa atg aac agc ctg aaa acc gag gac acg gcc gtg tat     288
     Leu Tyr Leu Gln Met Asn Ser Leu Lys Thr Glu Asp Thr Ala Val Tyr
                     85                  90                  95

     tac tgt gca agg ggg att tcg ccg ttt tac ttt gac tac tgg ggc cag     336
25   Tyr Cys Ala Arg Gly Ile Ser Pro Phe Tyr Phe Asp Tyr Trp Gly Gln
                100                 105                 110

     ggc acc ctg gtg acc gtc tcc agc gct agc acc aag ggc ccc agc gtg     384
     Gly Thr Leu Val Thr Val Ser Ser Ala Ser Thr Lys Gly Pro Ser Val
30           115                 120                 125

     ttc ccc ctg gcc ccc agc agc aag agc acc agc ggc ggc aca gcc gcc     432
     Phe Pro Leu Ala Pro Ser Ser Lys Ser Thr Ser Gly Gly Thr Ala Ala
         130                 135                 140
35
     ctg ggc tgc ctg gtg aag gac tac ttc ccc gag ccc gtg acc gtg agc     480
     Leu Gly Cys Leu Val Lys Asp Tyr Phe Pro Glu Pro Val Thr Val Ser
     145                 150                 155                 160

40   tgg aac agc ggc gcc ttg acc agc ggc gtg cac acc ttc ccc gcc gtg     528
     Trp Asn Ser Gly Ala Leu Thr Ser Gly Val His Thr Phe Pro Ala Val
                     165                 170                 175

     ctg cag agc agc ggc ctg tac agc ctg agc agc gtg gtg acc gtg ccc     576
45   Leu Gln Ser Ser Gly Leu Tyr Ser Leu Ser Ser Val Val Thr Val Pro
                180                 185                 190

     agc agc agc ctg ggc acc cag acc tac atc tgc aac gtg aac cac aag     624
     Ser Ser Ser Leu Gly Thr Gln Thr Tyr Ile Cys Asn Val Asn His Lys
50           195                 200                 205

     ccc agc aac acc aag gtg gac aaa cgc gtg gag ccc aag agc tgc gac     672
     Pro Ser Asn Thr Lys Val Asp Lys Arg Val Glu Pro Lys Ser Cys Asp
         210                 215                 220
55
     aag acc cac acc tgc ccc ccc tgc cct gcc ccc gag ctg ctg ggc gga     720
     Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly
     225                 230                 235                 240

60   ccc tcc gtg ttc ctg ttc ccc ccc aag ccc aag gac acc ctc atg atc     768
     Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile
                     245                 250                 255
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agc cgg acc ccc gag gtg acc tgc gtg gtg gtg gac gtg agc cac gag      816
Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu
            260                 265                 270

gac ccc gag gtg aag ttc aac tgg tac gtg gac ggc gtg gag gtg cac      864
Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His
        275                 280                 285

aac gcc aag acc aag ccc cgg gag gag cag tac aac agc acc tac cgg      912
Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg
    290                 295                 300

gtg gtg agc gtg ctc acc gtg ctg cac cag gac tgg ctg aac ggc aag      960
Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys
305                 310                 315                 320

gag tac aag tgc aag gtg agc aac aag gcc ctg cct gcc ccc atc gag     1008
Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu
                325                 330                 335

aag acc atc agc aag gcc aag ggc cag ccc cgg gag ccc cag gtg tac     1056
Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr
            340                 345                 350

acc ctg ccc ccc agc cgg gag gag atg acc aag aac cag gtg tcc ctc     1104
Thr Leu Pro Pro Ser Arg Glu Glu Met Thr Lys Asn Gln Val Ser Leu
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acc tgt ctg gtg aag ggc ttc tac ccc agc gac atc gcc gtg gag tgg     1152
Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp
    370                 375                 380

gag agc aac ggc cag ccc gag aac aac tac aag acc acc ccc cct gtg     1200
Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val
385                 390                 395                 400

ctg gac agc gac ggc agc ttc ttc ctg tac agc aag ctc acc gtg gac     1248
Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp
                405                 410                 415

aag agc cgg tgg cag cag ggc aac gtg ttc agc tgc agc gtg atg cac     1296
Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His
            420                 425                 430

gag gcc ctg cac aac cac tac acc cag aag agc ctg agc ctg agc ccc     1344
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<400> 107

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            20                  25                  30

Tyr Met Asp Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val
        35                  40                  45

Gly Arg Thr Arg Asn Lys Ala Asn Ser Tyr Thr Thr Glu Tyr Ala Ala
    50                  55                  60

Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser Lys Asn Ser
65                  70                  75                  80

Leu Tyr Leu Gln Met Asn Ser Leu Lys Thr Glu Asp Thr Ala Val Tyr
                85                  90                  95

Tyr Cys Ala Arg Gly Ile Ser Pro Phe Tyr Phe Asp Tyr Trp Gly Gln
            100                 105                 110

Gly Thr Leu Val Thr Val Ser Ser Ala Ser Thr Lys Gly Pro Ser Val
        115                 120                 125

Phe Pro Leu Ala Pro Ser Ser Lys Ser Thr Ser Gly Gly Thr Ala Ala
    130                 135                 140

Leu Gly Cys Leu Val Lys Asp Tyr Phe Pro Glu Pro Val Thr Val Ser
145                 150                 155                 160

Trp Asn Ser Gly Ala Leu Thr Ser Gly Val His Thr Phe Pro Ala Val
                165                 170                 175

Leu Gln Ser Ser Gly Leu Tyr Ser Leu Ser Ser Val Val Thr Val Pro
            180                 185                 190

Ser Ser Ser Leu Gly Thr Gln Thr Tyr Ile Cys Asn Val Asn His Lys
        195                 200                 205

Pro Ser Asn Thr Lys Val Asp Lys Arg Val Glu Pro Lys Ser Cys Asp
    210                 215                 220

Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly
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225 230 235 240

Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile
245 250 255

Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu
260 265 270

Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His
275 280 285

Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg
290 295 300

Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys
305 310 315 320

Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu
325 330 335

Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr
340 345 350

Thr Leu Pro Pro Ser Arg Glu Glu Met Thr Lys Asn Gln Val Ser Leu
355 360 365

Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp
370 375 380

Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val
385 390 395 400

Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp
405 410 415

Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His
420 425 430

Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro
435 440 445

Gly Lys
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<211> 1347

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<222> (1)..(1347)

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1               5                   10                  15

tct ctg aag atc tcc tgt aag ggt tct gga tac ggc ttt atc acc tac       96
Ser Leu Lys Ile Ser Cys Lys Gly Ser Gly Tyr Gly Phe Ile Thr Tyr
            20                  25                  30

tgg atc ggc tgg gtg cgc cag atg ccc ggg aaa ggc ctg gag tgg atg      144
Trp Ile Gly Trp Val Arg Gln Met Pro Gly Lys Gly Leu Glu Trp Met
        35                  40                  45

ggg atc atc tat cct ggt gac tct gaa acc aga tac agc ccg tcc ttc      192
Gly Ile Ile Tyr Pro Gly Asp Ser Glu Thr Arg Tyr Ser Pro Ser Phe
    50                  55                  60

caa ggc cag gtc acc atc tca gcc gac aag tcc atc aac acc gcc tac      240
Gln Gly Gln Val Thr Ile Ser Ala Asp Lys Ser Ile Asn Thr Ala Tyr
65                  70                  75                  80

ctg cag tgg agc agc ctg aag gcc tcg gac acc gcc ata tat tac tgt      288
Leu Gln Trp Ser Ser Leu Lys Ala Ser Asp Thr Ala Ile Tyr Tyr Cys
                85                  90                  95

gcg ggg ggt tcg ggg att tct acc cct atg gac gtc tgg ggc cag ggc      336
Ala Gly Gly Ser Gly Ile Ser Thr Pro Met Asp Val Trp Gly Gln Gly
            100                 105                 110

acc acc gtg acc gtc tcc agc gct agc acc aag ggc ccc agc gtg ttc      384
Thr Thr Val Thr Val Ser Ser Ala Ser Thr Lys Gly Pro Ser Val Phe
        115                 120                 125

ccc ctg gcc ccc agc agc aag agc acc agc ggc ggc aca gcc gcc ctg      432
Pro Leu Ala Pro Ser Ser Lys Ser Thr Ser Gly Gly Thr Ala Ala Leu
    130                 135                 140

ggc tgc ctg gtg aag gac tac ttc ccc gag ccc gtg acc gtg agc tgg      480
Gly Cys Leu Val Lys Asp Tyr Phe Pro Glu Pro Val Thr Val Ser Trp
145                 150                 155                 160

aac agc ggc gcc ttg acc agc ggc gtg cac acc ttc ccc gcc gtg ctg      528
Asn Ser Gly Ala Leu Thr Ser Gly Val His Thr Phe Pro Ala Val Leu
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                    165                 170                 175

     cag agc agc ggc ctg tac agc ctg agc agc gtg gtg acc gtg ccc agc      576
     Gln Ser Ser Gly Leu Tyr Ser Leu Ser Ser Val Val Thr Val Pro Ser
 5               180                 185                 190

     agc agc ctg ggc acc cag acc tac atc tgc aac gtg aac cac aag ccc      624
     Ser Ser Leu Gly Thr Gln Thr Tyr Ile Cys Asn Val Asn His Lys Pro
             195                 200                 205
10
     agc aac acc aag gtg gac aaa cgc gtg gag ccc aag agc tgc gac aag      672
     Ser Asn Thr Lys Val Asp Lys Arg Val Glu Pro Lys Ser Cys Asp Lys
         210                 215                 220

15   acc cac acc tgc ccc ccc tgc cct gcc ccc gag ctg ctg ggc gga ccc      720
     Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro
     225                 230                 235                 240

     tcc gtg ttc ctg ttc ccc ccc aag ccc aag gac acc ctc atg atc agc      768
20   Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser
                     245                 250                 255

     cgg acc ccc gag gtg acc tgc gtg gtg gtg gac gtg agc cac gag gac      816
     Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp
25               260                 265                 270

     ccc gag gtg aag ttc aac tgg tac gtg gac ggc gtg gag gtg cac aac      864
     Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn
             275                 280                 285
30
     gcc aag acc aag ccc cgg gag gag cag tac aac agc acc tac cgg gtg      912
     Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val
         290                 295                 300

35   gtg agc gtg ctc acc gtg ctg cac cag gac tgg ctg aac ggc aag gag      960
     Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu
     305                 310                 315                 320

     tac aag tgc aag gtg agc aac aag gcc ctg cct gcc ccc atc gag aag     1008
40   Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys
                     325                 330                 335

     acc atc agc aag gcc aag ggc cag ccc cgg gag ccc cag gtg tac acc     1056
     Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr
45               340                 345                 350

     ctg ccc ccc agc cgg gag gag atg acc aag aac cag gtg tcc ctc acc     1104
     Leu Pro Pro Ser Arg Glu Glu Met Thr Lys Asn Gln Val Ser Leu Thr
             355                 360                 365
50
     tgt ctg gtg aag ggc ttc tac ccc agc gac atc gcc gtg gag tgg gag     1152
     Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu
         370                 375                 380

55   agc aac ggc cag ccc gag aac aac tac aag acc acc ccc cct gtg ctg     1200
     Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu
     385                 390                 395                 400

     gac agc gac ggc agc ttc ttc ctg tac agc aag ctc acc gtg gac aag     1248
60   Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys
                     405                 410                 415

     agc cgg tgg cag cag ggc aac gtg ttc agc tgc agc gtg atg cac gag     1296
```

```
     Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu
                 420                 425                 430

     gcc ctg cac aac cac tac acc cag aag agc ctg agc ctg agc ccc ggc     1344
 5   Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly
             435                 440                 445

     aag                                                                1347
     Lys
10


     <210>  109

15   <211>  449

     <212>  PRT

     <213>  Artificial sequence
20


     <220>

25   <223>  IgG heavy chain of CR03-022

     <400>  109

     Glu Val Gln Leu Val Gln Ser Gly Thr Glu Val Lys Lys Pro Gly Glu
30   1               5                   10                  15


     Ser Leu Lys Ile Ser Cys Lys Gly Ser Gly Tyr Gly Phe Ile Thr Tyr
                 20                  25                  30
35

     Trp Ile Gly Trp Val Arg Gln Met Pro Gly Lys Gly Leu Glu Trp Met
             35                  40                  45

40
     Gly Ile Ile Tyr Pro Gly Asp Ser Glu Thr Arg Tyr Ser Pro Ser Phe
         50                  55                  60


45   Gln Gly Gln Val Thr Ile Ser Ala Asp Lys Ser Ile Asn Thr Ala Tyr
     65                  70                  75                  80


     Leu Gln Trp Ser Ser Leu Lys Ala Ser Asp Thr Ala Ile Tyr Tyr Cys
50                   85                  90                  95


     Ala Gly Gly Ser Gly Ile Ser Thr Pro Met Asp Val Trp Gly Gln Gly
                 100                 105                 110
55

     Thr Thr Val Thr Val Ser Ser Ala Ser Thr Lys Gly Pro Ser Val Phe
             115                 120                 125

60
     Pro Leu Ala Pro Ser Ser Lys Ser Thr Ser Gly Gly Thr Ala Ala Leu
         130                 135                 140
```

```
Gly Cys Leu Val Lys Asp Tyr Phe Pro Glu Pro Val Thr Val Ser Trp
145                 150                 155                 160

Asn Ser Gly Ala Leu Thr Ser Gly Val His Thr Phe Pro Ala Val Leu
                165                 170                 175

Gln Ser Ser Gly Leu Tyr Ser Leu Ser Ser Val Val Thr Val Pro Ser
            180                 185                 190

Ser Ser Leu Gly Thr Gln Thr Tyr Ile Cys Asn Val Asn His Lys Pro
        195                 200                 205

Ser Asn Thr Lys Val Asp Lys Arg Val Glu Pro Lys Ser Cys Asp Lys
    210                 215                 220

Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro
225                 230                 235                 240

Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser
                245                 250                 255

Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp
            260                 265                 270

Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn
        275                 280                 285

Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val
    290                 295                 300

Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu
305                 310                 315                 320

Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys
                325                 330                 335

Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr
            340                 345                 350

Leu Pro Pro Ser Arg Glu Glu Met Thr Lys Asn Gln Val Ser Leu Thr
        355                 360                 365

Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu
    370                 375                 380

Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu
385                 390                 395                 400
```

Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys
405 410 415

Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu
420 425 430

Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly
435 440 445

Lys

<210> 110

<211> 642

<212> DNA

<213> Artificial sequence

<220>

<223> IgG light chain of CR03-014

<220>

<221> CDS

<222> (1)..(642)

<223>

<400> 110

gac atc cag atg acc cag tct cca tcc tcc ctg tct gca tct gta gga 48
Asp Ile Gln Met Thr Gln Ser Pro Ser Ser Leu Ser Ala Ser Val Gly
1 5 10 15

gac aga gtc acc atc act tgc cgg gca agt cag agc att agc agc tac 96
Asp Arg Val Thr Ile Thr Cys Arg Ala Ser Gln Ser Ile Ser Ser Tyr
20 25 30

tta aat tgg tat cag cag aaa cca ggg aaa gcc cct aag ctc ctg atc 144
Leu Asn Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Leu Ile
35 40 45

tat gct gca tcc agt ttg caa agt ggg gtc cca tca agg ttc agt ggc 192
Tyr Ala Ala Ser Ser Leu Gln Ser Gly Val Pro Ser Arg Phe Ser Gly
50 55 60

agt gga tct ggg aca gat ttc act ctc acc atc agc agt ctg caa cct 240
Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro
65 70 75 80

```
gaa gat ttt gca act tac tac tgt caa cag agt tac agt acc cct cca      288
Glu Asp Phe Ala Thr Tyr Tyr Cys Gln Gln Ser Tyr Ser Thr Pro Pro
                85                  90                  95

acg ttc ggc caa ggg acc aag gtg gag atc aaa cgg acc gtg gcc gct      336
Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Arg Thr Val Ala Ala
            100                 105                 110

ccc agc gtg ttc atc ttc ccc ccc tcc gac gag cag ctg aag agc ggc      384
Pro Ser Val Phe Ile Phe Pro Pro Ser Asp Glu Gln Leu Lys Ser Gly
        115                 120                 125

acc gcc agc gtg gtg tgc ctg ctg aac aac ttc tac ccc cgg gag gcc      432
Thr Ala Ser Val Val Cys Leu Leu Asn Asn Phe Tyr Pro Arg Glu Ala
    130                 135                 140

aag gtg cag tgg aag gtg gac aac gcc ctg cag agc ggc aac agc cag      480
Lys Val Gln Trp Lys Val Asp Asn Ala Leu Gln Ser Gly Asn Ser Gln
145                 150                 155                 160

gag agc gtg acc gag cag gac agc aag gac tcc acc tac agc ctg agc      528
Glu Ser Val Thr Glu Gln Asp Ser Lys Asp Ser Thr Tyr Ser Leu Ser
                165                 170                 175

agc acc ctc acc ctg agc aag gcc gac tac gag aag cac aag gtg tac      576
Ser Thr Leu Thr Leu Ser Lys Ala Asp Tyr Glu Lys His Lys Val Tyr
            180                 185                 190

gcc tgc gag gtg acc cac cag ggc ctg agc agc ccc gtg acc aag agc      624
Ala Cys Glu Val Thr His Gln Gly Leu Ser Ser Pro Val Thr Lys Ser
        195                 200                 205

ttc aac cgg ggc gag tgt                                             642
Phe Asn Arg Gly Glu Cys
    210
```

<210> 111

<211> 214

<212> PRT

<213> Artificial sequence

<220>

<223> IgG light chain of CR03-014

<400> 111

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Asp Ile Gln Met Thr Gln Ser Pro Ser Ser Leu Ser Ala Ser Val Gly
1               5                   10                  15

Asp Arg Val Thr Ile Thr Cys Arg Ala Ser Gln Ser Ile Ser Ser Tyr
            20                  25                  30

Leu Asn Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Leu Ile
        35                  40                  45
```

Tyr Ala Ala Ser Ser Leu Gln Ser Gly Val Pro Ser Arg Phe Ser Gly
50 55 60

Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro
65 70 75 80

Glu Asp Phe Ala Thr Tyr Tyr Cys Gln Gln Ser Tyr Ser Thr Pro Pro
85 90 95

Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Arg Thr Val Ala Ala
100 105 110

Pro Ser Val Phe Ile Phe Pro Pro Ser Asp Glu Gln Leu Lys Ser Gly
115 120 125

Thr Ala Ser Val Val Cys Leu Leu Asn Asn Phe Tyr Pro Arg Glu Ala
130 135 140

Lys Val Gln Trp Lys Val Asp Asn Ala Leu Gln Ser Gly Asn Ser Gln
145 150 155 160

Glu Ser Val Thr Glu Gln Asp Ser Lys Asp Ser Thr Tyr Ser Leu Ser
165 170 175

Ser Thr Leu Thr Leu Ser Lys Ala Asp Tyr Glu Lys His Lys Val Tyr
180 185 190

Ala Cys Glu Val Thr His Gln Gly Leu Ser Ser Pro Val Thr Lys Ser
195 200 205

Phe Asn Arg Gly Glu Cys
210

<210> 112

<211> 660

<212> DNA

<213> Artificial sequence

<220>

<223> IgG light chain of CR03-022

<220>

<221> CDS

<222> (1)..(660)

<223>

<400> 112

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gac atc gtg atg acc cag tct cca gac tcc ctg gct gtg tct ctg ggc       48
Asp Ile Val Met Thr Gln Ser Pro Asp Ser Leu Ala Val Ser Leu Gly
1               5                   10                  15

gag agg gcc acc atc aac tgc aag tcc agc cag agt gtt tta tac agc       96
Glu Arg Ala Thr Ile Asn Cys Lys Ser Ser Gln Ser Val Leu Tyr Ser
            20                  25                  30

tcc atc aat aag aac tac tta gct tgg tac cag cag aaa cca gga cag      144
Ser Ile Asn Lys Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln
        35                  40                  45

cct cct aag ctg ctc att tac tgg gca tct acc cgg gaa tcc ggg gtc      192
Pro Pro Lys Leu Leu Ile Tyr Trp Ala Ser Thr Arg Glu Ser Gly Val
    50                  55                  60

cct gac cga ttc agt ggc agc ggg tct ggg aca gat ttc act ctc acc      240
Pro Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr
65                  70                  75                  80

atc agc agc ctg cag gct gaa gat gtg gca gtt tat tac tgt cag caa      288
Ile Ser Ser Leu Gln Ala Glu Asp Val Ala Val Tyr Tyr Cys Gln Gln
                85                  90                  95

tat tat agt act ccg tac act ttt ggc cag ggg acc aag gtg gag atc      336
Tyr Tyr Ser Thr Pro Tyr Thr Phe Gly Gln Gly Thr Lys Val Glu Ile
            100                 105                 110

aag cgg acc gtg gcc gct ccc agc gtg ttc atc ttc ccc ccc tcc gac      384
Lys Arg Thr Val Ala Ala Pro Ser Val Phe Ile Phe Pro Pro Ser Asp
        115                 120                 125

gag cag ctg aag agc ggc acc gcc agc gtg gtg tgc ctg ctg aac aac      432
Glu Gln Leu Lys Ser Gly Thr Ala Ser Val Val Cys Leu Leu Asn Asn
    130                 135                 140

ttc tac ccc cgg gag gcc aag gtg cag tgg aag gtg gac aac gcc ctg      480
Phe Tyr Pro Arg Glu Ala Lys Val Gln Trp Lys Val Asp Asn Ala Leu
145                 150                 155                 160

cag agc ggc aac agc cag gag agc gtg acc gag cag gac agc aag gac      528
Gln Ser Gly Asn Ser Gln Glu Ser Val Thr Glu Gln Asp Ser Lys Asp
                165                 170                 175

tcc acc tac agc ctg agc agc acc ctc acc ctg agc aag gcc gac tac      576
Ser Thr Tyr Ser Leu Ser Ser Thr Leu Thr Leu Ser Lys Ala Asp Tyr
            180                 185                 190

gag aag cac aag gtg tac gcc tgc gag gtg acc cac cag ggc ctg agc      624
Glu Lys His Lys Val Tyr Ala Cys Glu Val Thr His Gln Gly Leu Ser
        195                 200                 205

agc ccc gtg acc aag agc ttc aac cgg ggc gag tgt                      660
Ser Pro Val Thr Lys Ser Phe Asn Arg Gly Glu Cys
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210 215 220

<210> 113

<211> 220

<212> PRT

<213> Artificial sequence

<220>

<223> IgG light chain of CR03-022

<400> 113

Asp Ile Val Met Thr Gln Ser Pro Asp Ser Leu Ala Val Ser Leu Gly
1 5 10 15

Glu Arg Ala Thr Ile Asn Cys Lys Ser Ser Gln Ser Val Leu Tyr Ser
20 25 30

Ser Ile Asn Lys Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln
35 40 45

Pro Pro Lys Leu Leu Ile Tyr Trp Ala Ser Thr Arg Glu Ser Gly Val
50 55 60

Pro Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr
65 70 75 80

Ile Ser Ser Leu Gln Ala Glu Asp Val Ala Val Tyr Tyr Cys Gln Gln
85 90 95

Tyr Tyr Ser Thr Pro Tyr Thr Phe Gly Gln Gly Thr Lys Val Glu Ile
100 105 110

Lys Arg Thr Val Ala Ala Pro Ser Val Phe Ile Phe Pro Pro Ser Asp
115 120 125

Glu Gln Leu Lys Ser Gly Thr Ala Ser Val Val Cys Leu Leu Asn Asn
130 135 140

Phe Tyr Pro Arg Glu Ala Lys Val Gln Trp Lys Val Asp Asn Ala Leu
145 150 155 160

Gln Ser Gly Asn Ser Gln Glu Ser Val Thr Glu Gln Asp Ser Lys Asp
165 170 175

Ser Thr Tyr Ser Leu Ser Ser Thr Leu Thr Leu Ser Lys Ala Asp Tyr
180 185 190

Glu Lys His Lys Val Tyr Ala Cys Glu Val Thr His Gln Gly Leu Ser
195 200 205

Ser Pro Val Thr Lys Ser Phe Asn Arg Gly Glu Cys
210 215 220

<210> 114

<211> 3789

<212> DNA

<213> Artificial sequence

<220>

<223> S protein of SARS-CoV strain Frankfurt 1

<220>

<221> CDS

<222> (10)..(3777)

<223>

<400> 114

ggtaccgcc atg ttc atc ttc ctg ctg ttc ctg acc ctg acc agc ggc agc 51
Met Phe Ile Phe Leu Leu Phe Leu Thr Leu Thr Ser Gly Ser
1 5 10

gat ctg gat agg tgc acc acc ttc gac gac gtg cag gcc cct aat tac 99
Asp Leu Asp Arg Cys Thr Thr Phe Asp Asp Val Gln Ala Pro Asn Tyr
15 20 25 30

acc cag cac acc agc tct atg cgg ggc gtg tac tac ccc gac gag atc 147
Thr Gln His Thr Ser Ser Met Arg Gly Val Tyr Tyr Pro Asp Glu Ile
35 40 45

ttc aga agc gac acc ctg tac ctg aca cag gac ctg ttc ctg ccc ttc 195
Phe Arg Ser Asp Thr Leu Tyr Leu Thr Gln Asp Leu Phe Leu Pro Phe
50 55 60

tac agc aac gtg acc ggc ttc cac acc atc aac cac acc ttc ggc aac 243
Tyr Ser Asn Val Thr Gly Phe His Thr Ile Asn His Thr Phe Gly Asn
65 70 75

ccc gtg atc cct ttc aag gac ggc atc tac ttc gcc gcc acc gag aag 291
Pro Val Ile Pro Phe Lys Asp Gly Ile Tyr Phe Ala Ala Thr Glu Lys
80 85 90

agc aat gtg gtg cgg ggc tgg gtg ttc ggc agc acc atg aac aac aag 339
Ser Asn Val Val Arg Gly Trp Val Phe Gly Ser Thr Met Asn Asn Lys

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     95                  100                 105                 110

     agc cag agc gtg atc atc atc aac aat agc acc aac gtg gtg atc agg      387
     Ser Gln Ser Val Ile Ile Ile Asn Asn Ser Thr Asn Val Val Ile Arg
 5                   115                 120                 125

     gcc tgc aac ttc gag ctg tgc gac aac cct ttc ttc gcc gtg tcc aaa      435
     Ala Cys Asn Phe Glu Leu Cys Asp Asn Pro Phe Phe Ala Val Ser Lys
                 130                 135                 140
10
     cct atg ggc acc cag acc cac acc atg atc ttc gac aac gcc ttc aac      483
     Pro Met Gly Thr Gln Thr His Thr Met Ile Phe Asp Asn Ala Phe Asn
         145                 150                 155

15   tgc acc ttc gag tac atc agc gac gcc ttc agc ctg gat gtg agc gag      531
     Cys Thr Phe Glu Tyr Ile Ser Asp Ala Phe Ser Leu Asp Val Ser Glu
     160                 165                 170

     aag agc ggg aac ttc aag cac ctg cgg gag ttc gtg ttc aag aac aag      579
20   Lys Ser Gly Asn Phe Lys His Leu Arg Glu Phe Val Phe Lys Asn Lys
     175                 180                 185                 190

     gac ggc ttc ctg tac gtg tac aag ggc tac cag ccc atc gac gtg gtg      627
     Asp Gly Phe Leu Tyr Val Tyr Lys Gly Tyr Gln Pro Ile Asp Val Val
25                   195                 200                 205

     aga gat ctg ccc agc ggc ttc aac acc ctg aag ccc atc ttc aag ctg      675
     Arg Asp Leu Pro Ser Gly Phe Asn Thr Leu Lys Pro Ile Phe Lys Leu
                 210                 215                 220
30
     ccc ctg ggc atc aac atc acc aac ttc cgg gcc atc ctg acc gcc ttc      723
     Pro Leu Gly Ile Asn Ile Thr Asn Phe Arg Ala Ile Leu Thr Ala Phe
         225                 230                 235

35   agc cct gcc cag gac atc tgg ggc acc agc gcc gct gcc tac ttc gtg      771
     Ser Pro Ala Gln Asp Ile Trp Gly Thr Ser Ala Ala Ala Tyr Phe Val
     240                 245                 250

     ggc tac ctg aag ccc acc acc ttc atg ctg aag tac gac gag aac ggc      819
40   Gly Tyr Leu Lys Pro Thr Thr Phe Met Leu Lys Tyr Asp Glu Asn Gly
     255                 260                 265                 270

     acc atc acc gat gcc gtg gac tgc agc cag aac ccc ctg gcc gag ctg      867
     Thr Ile Thr Asp Ala Val Asp Cys Ser Gln Asn Pro Leu Ala Glu Leu
45                   275                 280                 285

     aag tgc agc gtg aag agc ttc gag atc gac aag ggc atc tac cag acc      915
     Lys Cys Ser Val Lys Ser Phe Glu Ile Asp Lys Gly Ile Tyr Gln Thr
                 290                 295                 300
50
     agc aac ttc aga gtg gtg ccc agc ggc gat gtg gtg agg ttc ccc aac      963
     Ser Asn Phe Arg Val Val Pro Ser Gly Asp Val Val Arg Phe Pro Asn
         305                 310                 315

55   atc acc aac ctg tgc cct ttc ggc gag gtg ttc aac gcc acc aag ttc     1011
     Ile Thr Asn Leu Cys Pro Phe Gly Glu Val Phe Asn Ala Thr Lys Phe
     320                 325                 330

     cct agc gtg tac gcc tgg gag cgg aag aag atc agc aac tgc gtg gcc     1059
60   Pro Ser Val Tyr Ala Trp Glu Arg Lys Lys Ile Ser Asn Cys Val Ala
     335                 340                 345                 350

     gat tac agc gtg ctg tac aac agc acc ttc ttc agc acc ttc aag tgc     1107
```

```
Asp Tyr Ser Val Leu Tyr Asn Ser Thr Phe Phe Ser Thr Phe Lys Cys
                 355                 360                 365

tac ggc gtg agc gcc acc aag ctg aac gac ctg tgc ttc agc aac gtg     1155
Tyr Gly Val Ser Ala Thr Lys Leu Asn Asp Leu Cys Phe Ser Asn Val
             370                 375                 380

tac gcc gac agc ttc gtg gtg aag ggc gac gac gtg aga cag atc gcc     1203
Tyr Ala Asp Ser Phe Val Val Lys Gly Asp Asp Val Arg Gln Ile Ala
         385                 390                 395

cct ggc cag acc ggc gtg atc gcc gac tac aat tac aag ctg ccc gac     1251
Pro Gly Gln Thr Gly Val Ile Ala Asp Tyr Asn Tyr Lys Leu Pro Asp
     400                 405                 410

gac ttc atg ggc tgc gtg ctg gcc tgg aac acc aga aac atc gac gcc     1299
Asp Phe Met Gly Cys Val Leu Ala Trp Asn Thr Arg Asn Ile Asp Ala
415                 420                 425                 430

acc tcc acc ggc aac tac aac tac aag tac cgc tac ctg agg cac ggc     1347
Thr Ser Thr Gly Asn Tyr Asn Tyr Lys Tyr Arg Tyr Leu Arg His Gly
                 435                 440                 445

aag ctg aga ccc ttc gag cgg gac atc agc aac gtg ccc ttc agc cct     1395
Lys Leu Arg Pro Phe Glu Arg Asp Ile Ser Asn Val Pro Phe Ser Pro
             450                 455                 460

gac ggc aag ccc tgc acc ccc cct gcc ctg aac tgc tac tgg ccc ctg     1443
Asp Gly Lys Pro Cys Thr Pro Pro Ala Leu Asn Cys Tyr Trp Pro Leu
         465                 470                 475

aac gac tac ggc ttc tac acc acc acc ggc atc ggc tac cag cct tac     1491
Asn Asp Tyr Gly Phe Tyr Thr Thr Thr Gly Ile Gly Tyr Gln Pro Tyr
     480                 485                 490

aga gtg gtg gtg ctg agc ttc gag ctg ctg aac gcc cct gcc acc gtg     1539
Arg Val Val Val Leu Ser Phe Glu Leu Leu Asn Ala Pro Ala Thr Val
495                 500                 505                 510

tgc ggc ccc aag ctg agc acc gac ctg atc aag aac cag tgc gtg aac     1587
Cys Gly Pro Lys Leu Ser Thr Asp Leu Ile Lys Asn Gln Cys Val Asn
                 515                 520                 525

ttc aac ttc aac ggc ctg acc ggc acc ggc gtg ctg acc cct agc agc     1635
Phe Asn Phe Asn Gly Leu Thr Gly Thr Gly Val Leu Thr Pro Ser Ser
             530                 535                 540

aag agg ttc cag ccc ttc cag cag ttc ggc agg gac gtg agc gat ttc     1683
Lys Arg Phe Gln Pro Phe Gln Gln Phe Gly Arg Asp Val Ser Asp Phe
         545                 550                 555

acc gac agc gtg agg gat cct aag acc agc gag atc ctg gac atc agc     1731
Thr Asp Ser Val Arg Asp Pro Lys Thr Ser Glu Ile Leu Asp Ile Ser
     560                 565                 570

cct tgc agc ttc ggc ggc gtg agc gtg atc acc ccc ggc acc aac gcc     1779
Pro Cys Ser Phe Gly Gly Val Ser Val Ile Thr Pro Gly Thr Asn Ala
575                 580                 585                 590

agc tcc gag gtg gcc gtg ctg tac cag gac gtg aac tgc acc gac gtg     1827
Ser Ser Glu Val Ala Val Leu Tyr Gln Asp Val Asn Cys Thr Asp Val
                 595                 600                 605
```

```
     agc acc gcc atc cac gcc gac cag ctg acc ccc gcc tgg aga atc tac     1875
     Ser Thr Ala Ile His Ala Asp Gln Leu Thr Pro Ala Trp Arg Ile Tyr
                 610                 615                 620

 5   agc acc ggc aac aac gtg ttc cag acc cag gcc ggc tgc ctg atc ggc     1923
     Ser Thr Gly Asn Asn Val Phe Gln Thr Gln Ala Gly Cys Leu Ile Gly
             625                 630                 635

     gcc gag cac gtg gac acc agc tac gag tgc gac atc ccc atc gga gcc     1971
10   Ala Glu His Val Asp Thr Ser Tyr Glu Cys Asp Ile Pro Ile Gly Ala
         640                 645                 650

     ggc atc tgc gcc agc tac cac acc gtg agc ctg ctg aga agc acc agc     2019
     Gly Ile Cys Ala Ser Tyr His Thr Val Ser Leu Leu Arg Ser Thr Ser
15   655                 660                 665                 670

     cag aag agc atc gtg gcc tac acc atg agc ctg ggc gcc gac agc agc     2067
     Gln Lys Ser Ile Val Ala Tyr Thr Met Ser Leu Gly Ala Asp Ser Ser
                     675                 680                 685
20
     atc gcc tac agc aac aac acc atc gcc atc ccc acc aac ttc agc atc     2115
     Ile Ala Tyr Ser Asn Asn Thr Ile Ala Ile Pro Thr Asn Phe Ser Ile
                 690                 695                 700

25   agc atc acc acc gag gtg atg ccc gtg agc atg gcc aag acc agc gtg     2163
     Ser Ile Thr Thr Glu Val Met Pro Val Ser Met Ala Lys Thr Ser Val
             705                 710                 715

     gac tgc aac atg tac atc tgc ggc gac agc acc gag tgc gcc aac ctg     2211
30   Asp Cys Asn Met Tyr Ile Cys Gly Asp Ser Thr Glu Cys Ala Asn Leu
         720                 725                 730

     ctg ctg cag tac ggc agc ttc tgc acc cag ctg aac aga gcc ctg agc     2259
     Leu Leu Gln Tyr Gly Ser Phe Cys Thr Gln Leu Asn Arg Ala Leu Ser
35   735                 740                 745                 750

     ggc atc gcc gcc gag cag gac aga aac acc agg gag gtg ttc gcc cag     2307
     Gly Ile Ala Ala Glu Gln Asp Arg Asn Thr Arg Glu Val Phe Ala Gln
                     755                 760                 765
40
     gtg aag cag atg tat aag acc ccc acc ctg aag tac ttc ggc ggc ttc     2355
     Val Lys Gln Met Tyr Lys Thr Pro Thr Leu Lys Tyr Phe Gly Gly Phe
                 770                 775                 780

45   aac ttc agc cag atc ctg ccc gat cct ctg aag ccc acc aag cgg agc     2403
     Asn Phe Ser Gln Ile Leu Pro Asp Pro Leu Lys Pro Thr Lys Arg Ser
             785                 790                 795

     ttc atc gag gac ctg ctg ttc aac aag gtg acc ctg gcc gac gcc ggc     2451
50   Phe Ile Glu Asp Leu Leu Phe Asn Lys Val Thr Leu Ala Asp Ala Gly
         800                 805                 810

     ttt atg aag cag tac ggc gag tgc ctg ggc gat atc aac gcc agg gac     2499
     Phe Met Lys Gln Tyr Gly Glu Cys Leu Gly Asp Ile Asn Ala Arg Asp
55   815                 820                 825                 830

     ctg atc tgc gcc cag aag ttc aat ggc ctg acc gtg ctg ccc ccc ctg     2547
     Leu Ile Cys Ala Gln Lys Phe Asn Gly Leu Thr Val Leu Pro Pro Leu
                     835                 840                 845
60
     ctg acc gac gac atg atc gcc gcc tac aca gcc gcc ctg gtg agc ggc     2595
     Leu Thr Asp Asp Met Ile Ala Ala Tyr Thr Ala Ala Leu Val Ser Gly
                 850                 855                 860
```

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    acc gcc acc gcc ggc tgg acc ttt ggc gcc gga gcc gcc ctg cag atc     2643
    Thr Ala Thr Ala Gly Trp Thr Phe Gly Ala Gly Ala Ala Leu Gln Ile
            865                 870                 875
5
    ccc ttc gcc atg cag atg gcc tac cgg ttc aat ggc atc ggc gtg acc     2691
    Pro Phe Ala Met Gln Met Ala Tyr Arg Phe Asn Gly Ile Gly Val Thr
        880                 885                 890

10  cag aac gtg ctg tac gag aac cag aag cag atc gcc aac cag ttc aac     2739
    Gln Asn Val Leu Tyr Glu Asn Gln Lys Gln Ile Ala Asn Gln Phe Asn
    895                 900                 905                 910

    aag gcc atc agc cag atc cag gag agc ctg acc acc aca agc acc gcc     2787
15  Lys Ala Ile Ser Gln Ile Gln Glu Ser Leu Thr Thr Thr Ser Thr Ala
                    915                 920                 925

    ctg ggc aag ctg cag gac gtg gtg aac cag aac gcc cag gcc ctg aat     2835
    Leu Gly Lys Leu Gln Asp Val Val Asn Gln Asn Ala Gln Ala Leu Asn
20              930                 935                 940

    acc ctg gtg aag cag ctg agc agc aac ttc ggc gcc atc agc tcc gtg     2883
    Thr Leu Val Lys Gln Leu Ser Ser Asn Phe Gly Ala Ile Ser Ser Val
            945                 950                 955
25
    ctg aac gac atc ctg agc cgg ctg gac aag gtg gag gcc gag gtg cag     2931
    Leu Asn Asp Ile Leu Ser Arg Leu Asp Lys Val Glu Ala Glu Val Gln
        960                 965                 970

30  atc gac aga ctg atc acc ggc aga ctg cag agc ctg cag acc tac gtg     2979
    Ile Asp Arg Leu Ile Thr Gly Arg Leu Gln Ser Leu Gln Thr Tyr Val
    975                 980                 985                 990

    acc cag cag ctg atc aga gcc gcc gag atc  aga gcc agc gcc aac  ctg   3027
35  Thr Gln Gln Leu Ile Arg Ala Ala Glu Ile  Arg Ala Ser Ala Asn  Leu
                    995                 1000                1005

    gcc gcc acc aag  atg agc gag tgc gtg  ctg ggc cag agc aag  aga      3072
    Ala Ala Thr Lys  Met Ser Glu Cys Val  Leu Gly Gln Ser Lys  Arg
40              1010                1015                1020

    gtg gac ttc tgc  ggc aag ggc tac cac  ctg atg agc ttc ccc  cag      3117
    Val Asp Phe Cys  Gly Lys Gly Tyr His  Leu Met Ser Phe Pro  Gln
                1025                1030                1035
45
    gcc gct ccc cac  ggc gtg gtg ttc ctg  cac gtg acc tac gtg  cct      3162
    Ala Ala Pro His  Gly Val Val Phe Leu  His Val Thr Tyr Val  Pro
                1040                1045                1050

50  agc cag gag agg  aat ttc acc acc gcc  cct gcc atc tgc cac  gag      3207
    Ser Gln Glu Arg  Asn Phe Thr Thr Ala  Pro Ala Ile Cys His  Glu
                1055                1060                1065

    ggc aag gcc tac  ttc ccc aga gag ggc  gtg ttc gtg ttc aat  ggc      3252
55  Gly Lys Ala Tyr  Phe Pro Arg Glu Gly  Val Phe Val Phe Asn  Gly
                1070                1075                1080

    acc agc tgg ttc  atc acc cag cgg aac  ttc ttc agc ccc cag  atc      3297
    Thr Ser Trp Phe  Ile Thr Gln Arg Asn  Phe Phe Ser Pro Gln  Ile
60              1085                1090                1095

    atc aca acc gac  aac acc ttc gtg agc  ggc aac tgc gac gtg  gtg      3342
    Ile Thr Thr Asp  Asn Thr Phe Val Ser  Gly Asn Cys Asp Val  Val
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                1100                1105                1110

    atc ggc atc att  aac aat acc gtg tac  gac ccc ctg cag ccc  gag      3387
    Ile Gly Ile Ile  Asn Asn Thr Val Tyr  Asp Pro Leu Gln Pro  Glu
5               1115                1120                1125

    ctg gat agc ttc  aag gag gag ctg gac  aag tac ttc aag aac  cac      3432
    Leu Asp Ser Phe  Lys Glu Glu Leu Asp  Lys Tyr Phe Lys Asn  His
                1130                1135                1140
10
    acc agc ccc gat  gtg gac ttc ggc gac  atc agc ggc atc aat  gcc      3477
    Thr Ser Pro Asp  Val Asp Phe Gly Asp  Ile Ser Gly Ile Asn  Ala
                1145                1150                1155

15  agc gtg gtg aac  atc cag aag gag atc  gac cgg ctg aac gag  gtg      3522
    Ser Val Val Asn  Ile Gln Lys Glu Ile  Asp Arg Leu Asn Glu  Val
                1160                1165                1170

    gcc aag aac ctg  aac gag agc ctg atc  gac ctg cag gag ctg  ggc      3567
20  Ala Lys Asn Leu  Asn Glu Ser Leu Ile  Asp Leu Gln Glu Leu  Gly
                1175                1180                1185

    aag tac gag cag  tac atc aag tgg ccc  tgg tac gtg tgg ctg  ggc      3612
    Lys Tyr Glu Gln  Tyr Ile Lys Trp Pro  Trp Tyr Val Trp Leu  Gly
25              1190                1195                1200

    ttc atc gcc ggc  ctg atc gcc atc gtg  atg gtg acc atc ctg  ctg      3657
    Phe Ile Ala Gly  Leu Ile Ala Ile Val  Met Val Thr Ile Leu  Leu
                1205                1210                1215
30
    tgc tgc atg acc  agc tgc tgc tcc tgc  ctg aag ggc gcc tgc  agc      3702
    Cys Cys Met Thr  Ser Cys Cys Ser Cys  Leu Lys Gly Ala Cys  Ser
                1220                1225                1230

35  tgt ggc agc tgc  tgc aag ttc gac gag  gac gat agc gag ccc  gtg      3747
    Cys Gly Ser Cys  Cys Lys Phe Asp Glu  Asp Asp Ser Glu Pro  Val
                1235                1240                1245

    ctg aag ggc gtg  aag ctg cac tac acc  tga tgaattctcg ag           3789
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Asp Arg Cys Thr Thr Phe Asp Asp Val Gln Ala Pro Asn Tyr Thr Gln
            20                  25                  30

His Thr Ser Ser Met Arg Gly Val Tyr Tyr Pro Asp Glu Ile Phe Arg
        35                  40                  45

Ser Asp Thr Leu Tyr Leu Thr Gln Asp Leu Phe Leu Pro Phe Tyr Ser
    50                  55                  60

Asn Val Thr Gly Phe His Thr Ile Asn His Thr Phe Gly Asn Pro Val
65                  70                  75                  80

Ile Pro Phe Lys Asp Gly Ile Tyr Phe Ala Ala Thr Glu Lys Ser Asn
                85                  90                  95

Val Val Arg Gly Trp Val Phe Gly Ser Thr Met Asn Asn Lys Ser Gln
            100                 105                 110

Ser Val Ile Ile Ile Asn Asn Ser Thr Asn Val Val Ile Arg Ala Cys
        115                 120                 125

Asn Phe Glu Leu Cys Asp Asn Pro Phe Phe Ala Val Ser Lys Pro Met
    130                 135                 140

Gly Thr Gln Thr His Thr Met Ile Phe Asp Asn Ala Phe Asn Cys Thr
145                 150                 155                 160

Phe Glu Tyr Ile Ser Asp Ala Phe Ser Leu Asp Val Ser Glu Lys Ser
                165                 170                 175

Gly Asn Phe Lys His Leu Arg Glu Phe Val Phe Lys Asn Lys Asp Gly
            180                 185                 190

Phe Leu Tyr Val Tyr Lys Gly Tyr Gln Pro Ile Asp Val Val Arg Asp
        195                 200                 205

Leu Pro Ser Gly Phe Asn Thr Leu Lys Pro Ile Phe Lys Leu Pro Leu
    210                 215                 220

Gly Ile Asn Ile Thr Asn Phe Arg Ala Ile Leu Thr Ala Phe Ser Pro
225                 230                 235                 240

Ala Gln Asp Ile Trp Gly Thr Ser Ala Ala Ala Tyr Phe Val Gly Tyr
                245                 250                 255

Leu Lys Pro Thr Thr Phe Met Leu Lys Tyr Asp Glu Asn Gly Thr Ile
            260                 265                 270
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Thr Asp Ala Val Asp Cys Ser Gln Asn Pro Leu Ala Glu Leu Lys Cys
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Ser Val Lys Ser Phe Glu Ile Asp Lys Gly Ile Tyr Gln Thr Ser Asn
    290                 295                 300

Phe Arg Val Val Pro Ser Gly Asp Val Val Arg Phe Pro Asn Ile Thr
305                 310                 315                 320

Asn Leu Cys Pro Phe Gly Glu Val Phe Asn Ala Thr Lys Phe Pro Ser
                325                 330                 335

Val Tyr Ala Trp Glu Arg Lys Lys Ile Ser Asn Cys Val Ala Asp Tyr
            340                 345                 350

Ser Val Leu Tyr Asn Ser Thr Phe Phe Ser Thr Phe Lys Cys Tyr Gly
        355                 360                 365

Val Ser Ala Thr Lys Leu Asn Asp Leu Cys Phe Ser Asn Val Tyr Ala
    370                 375                 380

Asp Ser Phe Val Val Lys Gly Asp Asp Val Arg Gln Ile Ala Pro Gly
385                 390                 395                 400

Gln Thr Gly Val Ile Ala Asp Tyr Asn Tyr Lys Leu Pro Asp Asp Phe
                405                 410                 415

Met Gly Cys Val Leu Ala Trp Asn Thr Arg Asn Ile Asp Ala Thr Ser
            420                 425                 430

Thr Gly Asn Tyr Asn Tyr Lys Tyr Arg Tyr Leu Arg His Gly Lys Leu
        435                 440                 445

Arg Pro Phe Glu Arg Asp Ile Ser Asn Val Pro Phe Ser Pro Asp Gly
    450                 455                 460

Lys Pro Cys Thr Pro Pro Ala Leu Asn Cys Tyr Trp Pro Leu Asn Asp
465                 470                 475                 480

Tyr Gly Phe Tyr Thr Thr Thr Gly Ile Gly Tyr Gln Pro Tyr Arg Val
                485                 490                 495

Val Val Leu Ser Phe Glu Leu Leu Asn Ala Pro Ala Thr Val Cys Gly
            500                 505                 510

Pro Lys Leu Ser Thr Asp Leu Ile Lys Asn Gln Cys Val Asn Phe Asn
        515                 520                 525
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Phe Asn Gly Leu Thr Gly Thr Gly Val Leu Thr Pro Ser Ser Lys Arg
530 535 540

Phe Gln Pro Phe Gln Gln Phe Gly Arg Asp Val Ser Asp Phe Thr Asp
545 550 555 560

Ser Val Arg Asp Pro Lys Thr Ser Glu Ile Leu Asp Ile Ser Pro Cys
565 570 575

Ser Phe Gly Gly Val Ser Val Ile Thr Pro Gly Thr Asn Ala Ser Ser
580 585 590

Glu Val Ala Val Leu Tyr Gln Asp Val Asn Cys Thr Asp Val Ser Thr
595 600 605

Ala Ile His Ala Asp Gln Leu Thr Pro Ala Trp Arg Ile Tyr Ser Thr
610 615 620

Gly Asn Asn Val Phe Gln Thr Gln Ala Gly Cys Leu Ile Gly Ala Glu
625 630 635 640

His Val Asp Thr Ser Tyr Glu Cys Asp Ile Pro Ile Gly Ala Gly Ile
645 650 655

Cys Ala Ser Tyr His Thr Val Ser Leu Leu Arg Ser Thr Ser Gln Lys
660 665 670

Ser Ile Val Ala Tyr Thr Met Ser Leu Gly Ala Asp Ser Ser Ile Ala
675 680 685

Tyr Ser Asn Asn Thr Ile Ala Ile Pro Thr Asn Phe Ser Ile Ser Ile
690 695 700

Thr Thr Glu Val Met Pro Val Ser Met Ala Lys Thr Ser Val Asp Cys
705 710 715 720

Asn Met Tyr Ile Cys Gly Asp Ser Thr Glu Cys Ala Asn Leu Leu Leu
725 730 735

Gln Tyr Gly Ser Phe Cys Thr Gln Leu Asn Arg Ala Leu Ser Gly Ile
740 745 750

Ala Ala Glu Gln Asp Arg Asn Thr Arg Glu Val Phe Ala Gln Val Lys
755 760 765

Gln Met Tyr Lys Thr Pro Thr Leu Lys Tyr Phe Gly Gly Phe Asn Phe

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        770                 775                 780

    Ser Gln Ile Leu Pro Asp Pro Leu Lys Pro Thr Lys Arg Ser Phe Ile
5   785                 790                 795                 800

    Glu Asp Leu Leu Phe Asn Lys Val Thr Leu Ala Asp Ala Gly Phe Met
                805                 810                 815
10
    Lys Gln Tyr Gly Glu Cys Leu Gly Asp Ile Asn Ala Arg Asp Leu Ile
            820                 825                 830

15  Cys Ala Gln Lys Phe Asn Gly Leu Thr Val Leu Pro Pro Leu Leu Thr
        835                 840                 845

20  Asp Asp Met Ile Ala Ala Tyr Thr Ala Ala Leu Val Ser Gly Thr Ala
    850                 855                 860

    Thr Ala Gly Trp Thr Phe Gly Ala Gly Ala Ala Leu Gln Ile Pro Phe
25  865                 870                 875                 880

    Ala Met Gln Met Ala Tyr Arg Phe Asn Gly Ile Gly Val Thr Gln Asn
                885                 890                 895
30
    Val Leu Tyr Glu Asn Gln Lys Gln Ile Ala Asn Gln Phe Asn Lys Ala
            900                 905                 910

35  Ile Ser Gln Ile Gln Glu Ser Leu Thr Thr Thr Ser Thr Ala Leu Gly
        915                 920                 925

40  Lys Leu Gln Asp Val Val Asn Gln Asn Ala Gln Ala Leu Asn Thr Leu
    930                 935                 940

    Val Lys Gln Leu Ser Ser Asn Phe Gly Ala Ile Ser Ser Val Leu Asn
45  945                 950                 955                 960

    Asp Ile Leu Ser Arg Leu Asp Lys Val Glu Ala Glu Val Gln Ile Asp
                965                 970                 975
50
    Arg Leu Ile Thr Gly Arg Leu Gln Ser Leu Gln Thr Tyr Val Thr Gln
            980                 985                 990

55  Gln Leu Ile Arg Ala Ala Glu Ile  Arg Ala Ser Ala Asn  Leu Ala Ala
        995                 1000                1005

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    1010                1015                1020
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Phe Cys  Gly Lys Gly Tyr His  Leu Met Ser Phe Pro  Gln Ala Ala
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Pro His  Gly Val Val Phe Leu  His Val Thr Tyr Val  Pro Ser Gln
    1040                1045                1050

Glu Arg  Asn Phe Thr Thr Ala  Pro Ala Ile Cys His  Glu Gly Lys
    1055                1060                1065

Ala Tyr  Phe Pro Arg Glu Gly  Val Phe Val Phe Asn  Gly Thr Ser
    1070                1075                1080

Trp Phe  Ile Thr Gln Arg Asn  Phe Phe Ser Pro Gln  Ile Ile Thr
    1085                1090                1095

Thr Asp  Asn Thr Phe Val Ser  Gly Asn Cys Asp Val  Val Ile Gly
    1100                1105                1110

Ile Ile  Asn Asn Thr Val Tyr  Asp Pro Leu Gln Pro  Glu Leu Asp
    1115                1120                1125

Ser Phe  Lys Glu Glu Leu Asp  Lys Tyr Phe Lys Asn  His Thr Ser
    1130                1135                1140

Pro Asp  Val Asp Phe Gly Asp  Ile Ser Gly Ile Asn  Ala Ser Val
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Val Asn  Ile Gln Lys Glu Ile  Asp Arg Leu Asn Glu  Val Ala Lys
    1160                1165                1170

Asn Leu  Asn Glu Ser Leu Ile  Asp Leu Gln Glu Leu  Gly Lys Tyr
    1175                1180                1185

Glu Gln  Tyr Ile Lys Trp Pro  Trp Tyr Val Trp Leu  Gly Phe Ile
    1190                1195                1200

Ala Gly  Leu Ile Ala Ile Val  Met Val Thr Ile Leu  Leu Cys Cys
    1205                1210                1215

Met Thr  Ser Cys Cys Ser Cys  Leu Lys Gly Ala Cys  Ser Cys Gly
    1220                1225                1230

Ser Cys  Cys Lys Phe Asp Glu  Asp Asp Ser Glu Pro  Val Leu Lys
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REFERENCES

Boel E, Verlaan S, Poppelier MJ, Westerdaal NA, Van Strijp JA and Logtenberg T (2000), Functional human monoclonal antibodies of all isotypes constructed from phage display library-derived single-chain Fv antibody fragments. J. Immunol. Methods 239:153-166.

Burton DR and Barbas CF (1994), Human antibodies from combinatorial libraries. Adv. Immunol. 57:191-280.

Chou, TC and P Talalay (1984), Quantitative analysis of dose-effect relationships: the combined effects of multiple drugs or enzyme inhibitors. Adv Enzyme Regul. 22:27-55.

De Kruif J, Terstappen L, Boel E and Logtenberg T (1995a), Rapid selection of cell subpopulation-specific human monoclonal antibodies from a synthetic phage antibody library. Proc. Natl. Acad. Sci. USA 92:3938.

De Kruif J, Boel E and Logtenberg T (1995b), Selection and application of human single-chain Fv antibody fragments from a semi-synthetic phage antibody display library with designed CDR3 regions. J. Mol. Biol. 248:97-105.

Emini EA, Nara PL, Schleif WA, Lewis JA, Davide JP, Lee DR, Kessler J, Conley S, Matsushita S, Putney SDJ, Gerety RJ and Eichberg JW (1990), Antibody-mediated *in vitro* neutralization of human immunodeficiency virus type 1 abolishes infectivity for chimpanzees. J. Virol. 64:3674-3678.

Fouchier RA, Kuiken T, Schutten M, van Amerongen G, van Doornum GJ, van den Hoogen BG, Peiris M, Lim W, Stohr K, Osterhaus AD (2003), Koch's postulates fulfilled for SARS virus. Nature 243:240.

Guan Y, Zheng BJ, He YQ, Liu XL, Zhuang ZX, Cheung CL, Luo SW, Li PH, Zhang LJ, Guan YJ, Butt KM, Wong KL, Chan KW, Lim W, Shortridge KF, Yuen KY, Peiris JS, and Poon LL (2003),

Isolation and characterization of viruses related to the SARS coronavirus from animals in southern China. Science 302:276-278.

Havenga MJ, Lemckert AA, Grimbergen JM, Vogels R, Huisman LG, Valerio D, Bout A and Quax PH (2001), Improved adenovirus vectors for infection of cardiovascular tissues. J. Virol. 75:3335-3342.

Holmes KV (2003), SARS coronavirus: a new challenge for prevention and therapy. J. Clin. Invest. 111:1605-1609.

Huls G, Heijnen IJ, Cuomo E, van der Linden J, Boel E, van de Winkel J and Logtenberg T (1999), Antitumor immune effector mechanisms recruited by phage display-derived fully human IgG1 and IgA1 monoclonal antibodies. Cancer Res. 59:5778-5784.

Ksiazek TG, Erdman D, Goldsmith CS, Zaki SR, Peret T, Emery S, Tong S, Urbani C, Comer JA, Lim W, Rollin PE, Dowell SF, Ling AE, Humphrey CD, Shieh WJ, Guarner J, Paddock CD, Rota P, Fields B, DeRisi J, Yang JY, Cox N, Hughes JM, LeDuc JW, Bellini WJ, Anderson LJ (2003), A novel coronavirus associated with severe acute respiratory syndrome. N. Eng. J. Med. 348:1953-1966.

Kuiken T, Fouchier RA, Schutten M, Rimmelzwaan GF, van Amerongen G, van Riel D, Laman JD, de Jong T, van Doornum G, Lim W, Ling AE, Chan PK, Tam JS, Zambon MC, Gopal R, Drosten C, van der Werf S, Escriou N, Manuguerra JC, Stohr K, Peiris JS and Osterhaus AD (2003), Newly discovered coronavirus as the primary cause of severe acute respiratory syndrome. Lancet 362:263-270.

Li W, Moore MJ, Vasilieva N, Sui J, Wong SK, Berne MA, Somasundaran M, Sullivan JL, Luzuriaga K, Greenough TC, Choe H and Farzan M (2003), Angiotensin-converting enzyme 2 is a

functional receptor for the SARS coronavirus. Nature 2003 426:450-454.

Marra MA, Jones SJ, Astell CR, Holt RA, Brooks-Wilson A, Butterfield YS, Khattra J, Asano JK, Barber SA, Chan SY, Cloutier A, Coughlin SM, Freeman D, Girn N, Griffith OL, Leach SR, Mayo M, McDonald H, Montgomery SB, Pandoh PK, Petrescu AS, Robertson AG, Schein JE, Siddiqui A, Smailus DE, Stott JM, Yang GS, Plummer F, Andonov A, Artsob H, Bastien N, Bernard K, Booth TF, Bowness D, Czub M, Drebot M, Fernando L, Flick R, Garbutt M, Gray M, Grolla A, Jones S, Feldmann H, Meyers A, Kabani A, Li Y, Normand S, Stroher U, Tipples GA, Tyler S, Vogrig R, Ward D, Watson B, Brunham RC, Krajden M, Petric M, Skowronski DM, Upton C, Roper RL (2003), The genome sequence of the SARS-associated coronavirus. Science 300, 1399-1404.

Nicholls JM, Poon LL, Lee KC, Ng WF, Lai ST, Leung CY, Chu CM, Hui PK, Mak KL, Lim W, Yan KW, Chan KH, Tsang NC, Guan Y, Yuen KY, Peiris JS (2003), Lung pathology of fatal severe acute respiratory syndrome. Lancet 361:1773-1778.

Poon LLM, Leung CSW, Yuen KH, Guan Y, and Peiris JSM (2005), Recurrent mutations associated with isolation and passage of SARS Coronavirus in cells from non-human primates. J. Med. Virol. 76:435-440.

Rickerts V, Wolf T, Rottmann C, Preiser W, Drosten C, Jakobi V, Leong HN, Brodt HR (2003), Klinik und Behandlung des schweren akuten respiratorischen Syndroms. Dtsch. Med. Wochenschrift 128:1109-1114.

Rota PA, Oberste MS, Monroe SS, Nix WA, Campagnoli R, Icenogle JP, Penaranda S, Bankamp B, Maher K, Chen MH, Tong S, Tamin A, Lowe L, Frace M, DeRisi JL, Chen Q, Wang D, Erdman DD, Peret TC, Burns C, Ksiazek TG, Rollin PE, Sanchez A, Liffick S, Holloway B, Limor J, McCaustland K, Olsen-Rasmussen M, Fouchier R, Gunther S, Osterhaus AD, Drosten C, Pallansch

MA, Anderson LJ, Bellini WJ. (2003), Characterization of a novel coronavirus associated with severe acute respiratory syndrome. Science 300:1394-1399.

Slootstra JW, Puijk WC, Ligtvoet GJ, Langeveld JP, Meloen RH (1996), Structural aspects of antibody-antigen interaction revealed through small random peptide libraries. Mol. Divers. 1:87-96.

Ter Meulen J, Bakker ABH, van den Brink EN, Weverling GJ, Martina BEE, Haagmans BL, Kuiken T, de Kruif J, Preiser W, Spaan W, Gelderblom HR, Goudsmit J, Osterhaus ADME (2004), Human monoclonal antibody as prophylaxis for SARS coronavirus infection in ferrets. The Lancet 363:2139-2141.

Claims

1. A composition comprising at least two immunoglobulins that are capable of specifically binding to a SARS-CoV and have SARS-CoV neutralizing activity,
wherein the first immunoglobulin comprises a heavy chain variable region comprising the amino acid sequence of SEQ ID No: 4 and a light chain variable region comprising the amino acid sequence of SEQ ID NO: 8 or a heavy and light chain variable region having at least 97% sequence homology thereto and still having SARS-CoV binding and neutralizing activity and;
wherein the second immunoglobulin comprises a heavy chain variable region comprising the amino acid sequence of SEQ ID NO: 6 and a light chain variable region comprising the amino acid sequence of SEQ ID NO: 10 or a heavy and light chain variable region having at least 97% sequence homology thereto and still having SARS-CoV binding and neutralizing activity;
characterized in that the relative amounts of the immunoglobulins are such that they exhibit a synergistic neutralizing activity.

2. A composition according to claim 1, **characterized in that** the immunoglobulins are capable of reacting with different, non-competing epitopes of the SARS-CoV.

3. A composition according to claims 1 or 2, **characterized in that** the immunoglobulins are capable of reacting with different, non-competing epitopes of the S protein of SARS-CoV.

4. A composition according to any of the claims 1-3, **characterized in that** the immunoglobulins are capable of reacting with different, non-competing epitopes of amino acids 318-510 of the S protein of SARS-CoV.

5. A composition according to any of the claims 1-4, **characterized in that** at least one of the immunoglobulins reacts with an epitope comprising the amino acid sequence of SEQ ID NO:128.

6. A composition according to any of the claims 1-5, **characterized in that** at least one of the immunoglobulins is capable of reacting with amino acids 318-510 of the S protein of a human and an animal SARS-CoV.

7. A composition according to any of the claims 1-6, **characterized in that** at least one of the immunoglobulins is capable of reacting with amino acids 318-510 of the S protein of a SARS-CoV, wherein amino acid 479 is an amino acid other than asparagine, to a similar extent as with amino acids 318-510 of the S protein of a SARS-CoV, wherein amino acid 479 is an asparagine.

8. A composition according to any of the claims 1-7, **characterized in that** each of the immunoglobulins is capable of neutralizing a plurality of SARS-CoV strains.

9. A composition according to any of the claims 1-8, **characterized in that** the molar ratio of the two immunoglobulins is from 1:100 to 100:1.

10. A composition according to any of the claims 1-9, **characterized in that** a first immunoglobulin comprises at least a heavy chain CDR3 region comprising the amino acid sequence of SEQ ID NO:1 and a second immunoglobulin comprises at least a heavy chain CDR3 region comprising the amino acid sequence of SEQ ID NO:2.

11. A pharmaceutical composition comprising a composition according to any of the claims 1-10, the pharmaceutical composition further comprising a pharmaceutically acceptable excipient.

12. A pharmaceutical composition according to claim 11 for use as a medicament.

13. A pharmaceutical composition according to claim 11 for use in the diagnosis, prophylaxis, treatment, or combination thereof of a condition resulting from a SARS-CoV.

14. Use of a pharmaceutical composition according to claim 11 in the preparation of a medicament for the diagnosis, prophylaxis, treatment, or combination thereof of a condition resulting from a SARS-CoV.