CLAIMS:

1. A method for obtaining an epithelial organoid, the method comprising culturing one or more epithelial ducts, epithelial duct fragments and/or epithelial stem or progenitor cells isolated therefrom in the presence of a medium comprising
(a) a basal medium comprising a carbon source, water, salt, a source of amino acids and nitrogen, and more than one buffering agent, and
(b) Rh insulin, progesterone, putrescine, sodium selenite, human apo-transferrin, corticosterone, D-(+)-galactose, and BSA,
wherein the medium is free of FGF and/or nicotinamide, and wherein the medium is
(i) free of CHIR99021, R-spondin1, EGF, Noggin and LDN193189, wherein the medium supports the organoids for 5 or more passages,
(ii) free of EGF, Noggin and LDN193189, and supplemented with CHIR99021 and/or R-spondin1, wherein the medium supports the organoids for 10 or more passages, or
(iii) free of Noggin and LDN193189, and supplemented with EGF, and with CHIR99021 and/or R-spondin1, wherein the medium supports the organoids for 30 or more passages.

2. The method of claim 1, wherein the medium comprises N-Acetylcysteine.

3. The method of claim 1 or 2, wherein the epithelial stem or progenitor cells are:
(i) human cells, mouse cells or rat cells; and/or
(ii) hepatic epithelial stem cells, pancreatic epithelial stem cells or intestinal epithelial stem cells.

4. The method of any one of claims 1-3, wherein the medium is effective for long-term culture of the epithelial organoid, wherein the long-term culture is about 50 or more passages.

5. The method of any one of claims 1-4, further comprising culturing the one or more epithelial ducts, epithelial duct fragments and/or epithelial stem or progenitor cells:
(i) in contact with an extracellular matrix;
(ii) in contact with reduced support from an extracellular matrix; or
(iii) in suspension.

6. The method of any one of claims 1-5, wherein the epithelial ducts, epithelial duct fragments and/or epithelial stem or progenitor cells isolated therefrom are obtained from uninjured tissue.

7. A method of conducting a drug discovery screen; assaying toxicity; researching embryology, cell lineages, and differentiation pathways; studying gene expression including recombinant gene expression; researching mechanisms involved in injury and repair; researching inflammatory and infectious diseases; studying pathogenic mechanisms of cell transformation and etiology of cancer comprising obtaining an epithelial organoid according to any one of claims 1-6.

8. A method for obtaining an epithelial organoid, the method comprising:
isolating an organ comprising epithelial tissue or a portion thereof;
cutting and disrupting mechanically the isolated organ or the portion thereof to yield one or more pieces of epithelial tissue;
digesting enzymatically the one or more pieces of epithelial tissue to yield epithelial ducts and/or duct fragments;
collecting the digested epithelial ducts and/or duct fragments through a cell strainer;
plating the collected epithelial ducts and/or duct fragments; and
culturing the plated epithelial ducts and/or duct fragments in the presence of a medium comprising (a) a basal medium comprising a carbon source, water, salt, a source of amino acids and nitrogen, and more than one buffering agent, and (b) Rh insulin, progesterone, putrescine, sodium selenite, human apo-transferrin, corticosterone, D-(+)-galactose and BSA, wherein the medium is free of FGF and/or nicotinamide and wherein the medium is

(i) free of CHIR99021, R-spondin1, EGF, Noggin and LDN193189, wherein the medium supports the organoids for 5 or more passages,
(ii) free of EGF, Noggin and LDN193189, and supplemented with CHIR99021 and/or R-spondin1, wherein the medium supports the organoid for 10 or more passages or
(iii) free of Noggin and LDN193189, and supplemented with EGF, and with CHIR99021 and/or R-spondin1, wherein the medium supports the organoids for 30 or more passages, and
wherein the organ comprising epithelial tissue or a portion thereof is removed from a deceased subject using conventional dissection.

9. The method of claim 8, wherein the medium comprises N-Acetylcysteine.

10. The method of claim 8 or 9, the method further comprising maintaining intactness of a ductal architecture during digestion.

11. The method of any one of claims 8-10, the method further comprising treating a surface or implement with a surfactant prior to contact with the one or more pieces of epithelial tissue.

12. The method of any one of claims 8-11, further comprising plating the collected epithelial ducts and/or epithelial duct fragments:
(i) in contact with an extracellular matrix;
(ii) in contact with reduced support from an extracellular matrix; or
(iii) in suspension.

13. The method of claim 5 or 12, wherein the extracellular matrix concentration ranges between 0.1 to 50% (v/v).

14. The method of claim 5 or 12, wherein the suspension comprises an extracellular matrix with a concentration of about 0.1% (v/v) or less.

15. The method of any one of claims 5, 12, 13, and 14, wherein the extracellular matrix comprises Matrigel™.

16. The method of any one of claims 1-6 and 8-15, wherein the method further comprises subjecting the epithelial organoid to maturation and/or differentiation conditions.

17. A medium for obtaining an epithelial organoid, the medium comprising (a) a basal medium comprising a carbon source, water, salt, a source of amino acids and nitrogen, and more than one buffering agent, and (b) Rh insulin, progesterone, putrescine, sodium selenite, human apo-transferrin, corticosterone, D-(+)-galactose, and BSA, wherein the medium is free from FGF and/or nicotinamide, and wherein the medium is
(i) free of CHIR99021, R-spondin1, EGF, Noggin and LDN193189, wherein the medium supports the organoids for 5 or more passages,
(ii) free of EGF, Noggin and LDN193189, and supplemented with CHIR99021 and/or R-spondin1, wherein the medium supports the organoids for 10 or more passages, or
(iii) free of Noggin and LDN193189, and supplemented with an EGF, and with CHIR99021 and/or R-spondin1, wherein the medium supports the organoids for 30 or more passages.

18. The medium of claim 17, further comprising N-Acetylcysteine.

19. The medium of claim 17 or 18, further comprising an extracellular matrix.

20. The method of any one of claims 1 to 16 or the medium of any one of claims 17 to 19, wherein:
(i) a concentration of Rh insulin ranges from 0.01-100 µg/mL;
(ii) a concentration of progesterone ranges from 2 nM to 400 µM;
(iii) a concentration of putrescine ranges from 0.1-10 mM;
(iv) a concentration of sodium selenite ranges from 1 ng/mL to 100 µg/mL;
(v) a concentration of human apo-transferrin ranges from 0.2-200 µg/mL;

(vi) a concentration of corticosterone ranges from 1-500 ng/mL;

(vii) a concentration of D-(+)-galactose ranges from 30 μM to 30 mM; and

(viii) a concentration of BSA ranges from 0.5 μg/mL to 5 mg/mL.