COMPOSITION

TECHNICAL FIELD

The present invention relates to influenza vaccine formulations and vaccination regimes for immunising against various diseases. In particular the invention relates to vaccine formulations comprising an oil-in-water emulsion adjuvant and 3D-MPL, their use in medicine, in particular their use in augmenting immune responses to various antigens, and to methods of preparation, wherein the oil in water emulsion comprises a sterol, a metabolisable oil and an emulsifying agent.

TECHNICAL BACKGROUND

New compositions or vaccines with an improved immunogenicity are always needed. As one strategy, adjuvants have been used to try and improve the immune response raised to any given antigen.

By way of example, influenza vaccines and vaccines against human papilloma virus (HPV) have been developed with adjuvants.

Influenza viruses are one of the most ubiquitous viruses present in the world, affecting both humans and livestock. Influenza results in an economic burden, morbidity and even mortality, which are significant.

The influenza virus is an RNA enveloped virus with a particle size of about 125 nm in diameter. It consists basically of an internal nucleocapsid or core of ribonucleic acid (RNA) associated with nucleoprotein, surrounded by a viral envelope with a lipid bilayer structure and external glycoproteins. The inner layer of the viral envelope is composed predominantly of matrix proteins and the outer layer mostly of host-derived lipid material. Influenza virus comprises two surface antigens, glycoproteins neuraminidase (NA) and haemagglutinin (HA), which appear as spikes, 10 to 12 nm long, at the surface of the particles. It is these surface proteins, particularly the haemagglutinin that determine the antigenic specificity of the influenza subtypes.

These surface antigens progressively, sometimes rapidly, undergo some changes leading to the antigenic variations in influenza. These antigenic changes, called 'drifts' and 'shifts' are unpredictable and may have a dramatic impact from an immunological point of view as they eventually lead to the emergence of new influenza strains and that enable the virus to escape the immune system causing the well known, almost annual, epidemics.

The influenza virus strains to be incorporated into influenza vaccine each season are determined by the World Health Organisation in collaboration with national health authorities and vaccine manufacturers.

HA is the most important antigen in defining the serological specificity of the different influenza strains. This 75-80 kD protein contains numerous antigenic determinants, several of which are in regions that undergo sequence changes in different strains (strain-specific determinants) and others in regions which are common to many HA molecules (common to determinants).

Influenza viruses cause epidemics almost every winter, with infection rates for type A or B virus as high as 40% over a six-week period. Influenza infection results in various disease states, from a sub-clinical infection through mild upper respiratory infection to a severe viral pneumonia. Typical influenza epidemics cause increases in incidence of pneumonia and lower respiratory disease as witnessed by increased rates of hospitalization or mortality. The severity of the disease is primarily determined by the age of the host, his immune status and the site of infection.

Elderly people, 65 years old and over, are especially vulnerable, accounting for 80–90% of all influenza-related deaths in developed countries. Individuals with underlying chronic diseases are also most likely to experience such complications. Young infants also may suffer severe disease. These groups in particular therefore need to be protected. Besides these 'at risk'-groups, the health authorities are also recommending to vaccinate healthy adults who are in contact with elderly persons.

Vaccination plays a critical role in controlling annual influenza epidemics. Currently available influenza vaccines are either inactivated or live attenuated influenza vaccine. Inactivated flu vaccines are composed of three possible forms of antigen preparation: inactivated whole virus, sub-virions where purified virus particles are disrupted with detergents or other reagents to solubilise the lipid envelope (so-called "split" vaccine) or

purified HA and NA (subunit vaccine). These inactivated vaccines are given intramuscularly (i.m.) or intranasaly (i.n.).

Influenza vaccines, of all kinds, are usually trivalent vaccines. They generally contain antigens derived from two influenza A virus strains and one influenza B strain. A standard 0.5 ml injectable dose in most cases contains 15 µg of haemagglutinin antigen component from each strain, as measured by single radial immunodiffusion (SRD) (J.M. Wood et al.: An improved single radial immunodiffusion technique for the assay of influenza haemagglutinin antigen: adaptation for potency determination of inactivated whole virus and subunit vaccines. J. Biol. Stand. 5 (1977) 237-247; J. M. Wood et al., International collaborative study of single radial diffusion and immunoelectrophoresis techniques for the assay of haemagglutinin antigen of influenza virus. J. Biol. Stand. 9 (1981) 317-330).

Influenza vaccines currently available are considered safe in all age groups (De Donato et al. 1999, Vaccine, 17, 3094-3101). However, there is little evidence that current influenza vaccines work in small children under two years of age. Furthermore, reported rates of vaccine efficacy for prevention of typical confirmed influenza illness are 23–72% for the elderly, which are significantly lower than the 60–90% efficacy rates reported for younger adults (Govaert, 1994, J. Am. Med. Assoc., 21, 166-1665; Gross, 1995, Ann Intern. Med. 123, 523-527). The effectiveness of an influenza vaccine has been shown to correlate with serum titres of hemagglutination inhibition (HI) antibodies to the viral strain, and several studies have found that older adults exhibit lower HI titres after influenza immunisation than do younger adults (Murasko, 2002, Experimental gerontology, 37, 427-439).

New vaccines with an improved immunogenicity are therefore still needed. Formulation of vaccine antigen with potent adjuvants is a possible approach for enhancing immune responses to subvirion antigens.

A sub-unit influenza vaccine adjuvanted with the adjuvant MF59, in the form of an oil-in-water emulsion is commercially available, and has demonstrated its ability to induce a higher antibody titer than that obtained with the non-adjuvanted sub-unit vaccine (De Donato et al. 1999, Vaccine, 17, 3094-3101). However, in a later publication, the same vaccine has not demonstrated its improved profile compared to a non-adjuvanted split vaccine (Puig-Barbera et al., 2004, Vaccine 23, 283-289).

Papillomaviruses are small DNA tumour viruses, which are highly species specific. So far, over 100 individual human papillomavirus (HPV) genotypes have been described. HPVs are generally specific either for the skin (e.g. HPV-1 and -2) or mucosal surfaces (e.g. HPV-6 and -11) and usually cause benign tumours (warts) that persist for several months or years. Such benign tumours may be distressing for the individuals concerned but tend not to be life threatening, with a few exceptions.

Some HPVs are also associated with cancers. The strongest positive association between an HPV and human cancer is that which exists between HPV-16 and HPV-18 and cervical carcinoma. Cervical cancer is the most common malignancy in developing countries, with about 500,000 new cases occurring in the world each year. It is now technically feasible to actively combat primary HPV-16 infections, and even established HPV-16-containing cancers, using vaccines. For a review on the prospects for prophylactic and therapeutic vaccination against HPV-16 see Cason J., Clin. Immunother. 1994; 1(4) 293-306 and Hagenesee M.E., Infections in Medicine 1997 14(7) 555-556,559-564.

Although minor variations do occur, all HPVs genomes described have at least eight early genes, E1 to E8 and two late genes L1 and L2. In addition, an upstream regulatory region harbors the regulatory sequences which appear to control most transcriptional events of the HPV genome.

HPV L1 based vaccines are disclosed in WO94/00152, WO94/20137, WO93/02184 and WO94/05792. Such a vaccine can comprise the L1 antigen as a monomer, a capsomer or a virus like particle. Methods for the preparation of VLPs are well known in the art, and include VLP disassembly-reassembly approaches to provide enhanced homogeneity, for example as described in WO9913056 and US6245568. Such particles may additionally comprise L2 proteins. L2 based vaccines are described, for example, in WO93/00436. Other HPV vaccine approaches are based on the early proteins, such as E7 or fusion proteins such as L2-E7.

There is still a need for improved vaccines, such as influenza or HPV vaccines.

STATEMENT OF THE INVENTION

In first aspect of the present invention, there is provided an immunogenic composition comprising:

(a) an antigen,

(b) an oil-in-water emulsion adjuvant; and

(c) 3D MPL.

wherein said oil-in-water emulsion comprises a metabolisable oil, a sterol and an emulsifying agent.

The invention also relates to use of a composition comprising:

(a) an antigen, and

(b) an oil-in-water emulsion adjuvant; and

(c) 3D MPL

wherein said oil-in-water emulsion comprises a metabolisable oil, a sterol and an emulsifying agent, in the manufacture of an immunogenic composition for the prevention of infection and/or disease.

The invention also relates to a method of vaccination comprising delivery of an antigen, an oil in water emulsion adjuvant as defined herein and 3D-MPL.

The invention also relates to a method for the preparation of an immunogenic composition comprising combining an oil in water emulsion as defined herein with an antigen and 3D-MPL.

Other aspects and advantages of the present invention are described further in the following detailed description of the preferred embodiments thereof.

LEGEND TO FIGURES

Figure 1: Oil droplet particle size distribution in SB62 oil-in-water emulsion as measured by PCS. Figure 1A shows SB62 lot 1023 size measurements with the Malvern Zetasizer 3000HS: A = dilution 1/10000 (Rec22 to Rec24) (Analysis in Contin and adapted optical model 1.5/0.01); B = Dilution 1/20000 (Rec28 to Rec30) (Analysis in Contin and adapted optical model 1.5/0.01). Figure 1B shows a schematic illustration of record 22 (upper part) and record 23 (lower part) by intensity.

Figure 2: Schematic illustration of the preparation of MPL bulk.

Figure 3: Schematic illustration of the preparation of AS03+MPL adjuvant.

Figure 4: Explo Flu-001 clinical trial. CD4 T cell response to split influenza antigen (Q1 = first quartile, Q3 = third quartile).

Figure 5: Explo Flu-001 clinical trial. CD8 T cell response to split influenza antigen (Q1 = first quartile, Q3 = third quartile).

Figure 6: Explo Flu-001 clinical trial. Cross-reactive CD4 T-cell response to split influenza virus antigen after vaccination with Fluarix + AS03.

Figure 7: Explo Flu-001 clinical trial. B cell memory response post vaccination.

Figure 8: Explo Flu-002 clinical trial. CD4 T cell response against split influenza antigen following revaccination.

Figure 9: Explo Flu-002 clinical trial. Anti-HI titers following revaccination.

Figure 10: Ferret study I. Temperature monitoring (priming and challenge). Figure 10A is priming, Figure 10B is challenge.

Figure 11 : Ferret study I. Viral shedding.

Figure 12 : Ferret study II. Temperature monitoring (priming and challenge). Figure 12A is priming, Figure 12B is challenge.

Figure 13 : Ferret study II. Viral shedding.

Figure 14 : Ferret study II. HI titers to H3N2 A/Panama (vaccine strain) (Figure 14A) and to H3N2 A/Wyoming (challenge strain) (Figure 14B).

Figure 15 : Mice study. Frequencies of CD4 T cells in C57Bl/6 primed mice using whole inactivated virus as re-stimulating antigen (day 7 post-immunisation).

Figure 16 : Mice study. Frequencies of CD8 T cells in C57Bl/6 primed mice using whole inactivated virus as re-stimulating antigen (day 7 post-immunisation).

Figure 17 : Mice study. Frequencies of CD4 (upper part) and CD8 (lower part) T cells in C57Bl/6 mice primed with heterologous strains, using whole inactivated virus as re-stimulating antigen (day 7 post-immunisation).

Figure 18 : Human clinical trial. B cell memory response post-vaccination of elderly with Fluarix, Fluarix + AS03, Fluarix + AS03+MPL (difference between pre- and post-).

Figure 19 : Ferret study III. Temperature monitoring before and after challenge.

Figure 20: Ferret study III. Viral shedding before and after challenge.

Figure 21 : Ferret study III. HI titers to H3N2 A/Woming (vaccine strain).

Figure 22 : Ferret study III. HI titers to H3N2 A/Panama (challenge strain).

Figure 23 : Human clinical trial. HI titers (GMTs) at days 21, 90 and 180post vaccination (persistence).

Figure 24 : Human clinical trial. CD4 response – all double test - Pool antigen at days 21, 90 and 180 post vaccination (persistence).

Figure 25 : Human clinical trial. HI titers in a revaccination clinical trial with AS03+MPL compared to Fluarix.

Figure 26 : Human clinical trial. CMI for CD4 response - all double test - Pool antigen at days 0 and 21.

Figure 27: Human clinical trial with AS03+MPL at two concentrations. HI titers at days 0 and 21.

Figure 28: Human clinical trial with AS03+MPL at two concentrations. Reactogenicity.

DETAILED DESCRIPTION

The principles of the present invention are demonstrated in respect of a sub-unit or split influenza antigen or with various cancer-associated HPV antigens in the form of VLP, combined with an oil in water emulsion and 3D-MPL.

In one aspect of the invention, the present inventors have discovered that an influenza formulation comprising a sub-unit or split influenza virus or antigenic preparation thereof together with an oil-in-water emulsion adjuvant and 3D-MPL is capable of improving the CD4 T-cell immune response and/or the B cell memory response, against said antigen or antigenic composition in a human compared to that obtained with the un-adjuvanted sub-unit or split virus or split virus antigenic preparation thereof.

Said compositions thus provide improved influenza vaccines.

The claimed formulations may be used to induce anti-influenza CD4-T cell responses capable of detection of influenza epitopes presented by MHC class II molecules. The present Applicant has now found that it is effective to target the cell-mediated immune system in order to increase responsiveness against homologous and drift influenza strains (upon vaccination and infection).

The present inventors have discovered that an influenza formulation comprising a split influenza virus or split virus antigenic preparation thereof together with an oil-in-water emulsion adjuvant as defined herein and 3D MPL is capable of inducing at least a trend for a higher B cell memory response following the first vaccination of a human subject, compared to the un-adjuvanted composition.

The adjuvanted influenza compositions according to the invention have several advantages:

1) An improved immunogenicity: they will allow to restore weak immune response in the elderly peope (over 50 years of age, typically over 65 years of age) to levels seen in young people (antibody and/or T cell responses);
2) An improved cross-protection profile: increased cross-protection against variant (drifted) influenza strains;
3) They will also allow an reduced antigen dosage to be used for a similar response, thus ensuring an increased capacity in case of emergency (pandemics for example).

In another aspect of the invention, the inventors have also discovered that an oil-in-water emulsion adjuvant as defined herein + 3D MPL demonstrates immunogenicity results for both antibody production and B cell memory which are equivalent to, or sometimes greater than, those generated with an adjuvant devoid of the oil-in-water emulsion component.

These findings can be applied to other forms of the same antigens, and to other antigens.

Antigens

Antigens that may be used in the present invention include:

Streptococcal antigens such as from Group A Streptococcus, or Group B Streptococcus, but is most preferably from *Streptococcus pneumoniae*. A protein and/or saccharide antigen is most preferably used. The *Streptococcus pneumoniae* saccharide antigen and/or at least one *Streptococcus pneumoniae* protein antigen(s) is most preferably selected from the group consisting of: pneumolysin, PspA or transmembrane deletion variants thereof, PspC or transmembrane deletion variants thereof, PsaA or transmembrane deletion variants thereof, glyceraldehyde-3-phosphate dehydrogenase, CbpA or transmembrane deletion variants thereof, PhtA, PhtD, PhtB, PhtE, SpsA, LytB, LytC, LytA, Sp125, Sp101, Sp128, Sp130 and Sp133, or immunologically functional equivalent thereof.

Also preferred at least one (2, 3, 4, 5, 6, 7, 8, 9 or 10) *Streptococcus pneumoniae* saccharide antigen(s) and/or *Streptococcus pneumoniae* protein antigen preferably selected from the group of protein antigens listed above.

Certain compositions are described in WO 00/56359 and WO 02/22167 and WO 02/22168 (incorporated by reference herein).

The antigen may comprise capsular saccharide antigens (preferably conjugated to a carrier protein), wherein the saccharides (most preferably polysaccharides) are derived from at least four serotypes of pneumococcus. Preferably the four serotypes include 6B, 14, 19F and 23F. More preferably, at least 7 serotypes are included in the composition, for example those derived from serotypes 4, 6B, 9V, 14, 18C, 19F, and 23F. More preferably still, at least 11 serotypes are included in the composition, for example the composition in one embodiment includes capsular saccharides derived from serotypes 1, 3, 4, 5, 6B, 7F, 9V, 14, 18C, 19F and 23F (preferably conjugated to a carrier protein). In a preferred embodiment of the invention at least 13 saccharide antigens (preferably conjugated to a carrier protein) are included, although further saccharide antigens, for example 23 valent (such as serotypes 1, 2, 3, 4, 5, 6B, 7F, 8, 9N, 9V, 10A, 11A, 12F, 14, 15B, 17F, 18C, 19A, 19F, 20, 22F, 23F and 33F), are also contemplated by the invention.

For elderly vaccination (for instance for the prevention of pneumonia) it is advantageous to include serotypes 8 and 12F (and most preferably 15 and 22 as well) to the 11 valent antigenic composition described above to form a 15 valent composition, whereas for infants or toddlers (where otitis media is of more concern) serotypes 6A and 19A are advantageously included to form a 13 valent composition.

Although the above saccharides are advantageously in their full-length, native polysaccharide form, it should be understood that size-reduced polysaccharides may also be used which are still immunogenic (see for example EP 497524 and 497525) if necessary when coupled to a protein carrier.

For the prevention/amelioration of pneumonia in the elderly (+55 years) population and Otitis media in Infants (up to 18 months) and toddlers (typically 18 months to 5 years), it is a preferred embodiment of the invention to combine a multivalent *Streptococcus pneumonia* saccharide as herein described with a *Streptococcus pneumoniae* protein preferably selected from the group of proteins listed above. A combination of pneumococcal proteins may also be advantageously utilised as described below.

Pneumococcal Proteins

Streptococcus pneumoniae antigens are preferably selected from the group consisting of: a protein from the polyhistidine triad family (Pht), a protein from the Lyt family, a choline binding protein, proteins having an LPXTG motif (where X is any amino acid), proteins having a Type II Signal sequence motif of LXXC (where X is any amino acid), and proteins having a Type I Signal sequence motif. Preferred examples within these

categories (or motifs) are the following proteins (or truncate or immunologically functional equivalent thereof):

The Pht (Poly Histidine Triad) family comprises proteins PhtA, PhtB, PhtD, and PhtE. The family is characterised by a lipidation sequence, two domains separated by a proline-rich region and several histidine triads, possibly involved in metal or nucleoside binding or enzymatic activity, (3-5) coiled-coil regions, a conserved N-terminus and a heterogeneous C terminus. It is present in all strains of pneumococci tested. Homologous proteins have also been found in other Streptococci and *Neisseria*. Preferred members of the family comprise PhtA, PhtB and PhtD. More preferably, it comprises PhtA or PhtD. It is understood, however, that the terms Pht A, B, D, and E refer to proteins having sequences disclosed in the citations below as well as naturally-occurring (and man-made) variants thereof that have a sequence homology that is at least 90% identical to the referenced proteins. Preferably it is at least 95% identical and most preferably it is 97% identical.

With regards to the Pht proteins, PhtA is disclosed in WO 98/18930, and is also referred to Sp36. As noted above, it is a protein from the polyhistidine triad family and has the type II signal motif of LXXC.

PhtD is disclosed in WO 00/37105, and is also referred to Sp036D. As noted above, it also is a protein from the polyhistidine triad family and has the type II LXXC signal motif.

PhtB is disclosed in WO 00/37105, and is also referred to Sp036B. Another member of the PhtB family is the C3-Degrading Polypeptide, as disclosed in WO 00/17370. This protein also is from the polyhistidine triad family and has the type II LXXC signal motif. A preferred immunologically functional equivalent is the protein Sp42 disclosed in WO 98/18930. A PhtB truncate (approximately 79kD) is disclosed in WO99/15675 which is also considered a member of the PhtX family.

PhtE is disclosed in WO00/30299 and is referred to as BVH-3.

SpsA is a Choline binding protein (Cbp) disclosed in WO 98/39450.

The Lyt family is membrane associated proteins associated with cell lysis. The N-terminal domain comprises choline binding domain(s), however the Lyt family does not have all the features found in the choline binding protein family (Cbp) family noted below and thus for the present invention, the Lyt family is considered distinct from the Cbp family. In contrast with the Cbp family, the C-terminal domain contains the catalytic domain of the Lyt protein family. The family comprises LytA, B and C. With regards to the Lyt family, LytA is disclosed in Ronda et al., Eur J Biochem, 164:621-624 (1987). LytB is disclosed in WO 98/18930, and is also referred to as Sp46. LytC is also disclosed in WO 98/18930, and is also referred to as Sp91. A preferred member of that family is LytC.

Another preferred embodiment are Lyt family (particularly LytA) truncates wherein "Lyt" is defined above and "truncates" refers to proteins lacking 50% or more of the Choline binding region. Preferably such proteins lack the entire choline binding region.

Sp125 is an example of a pneumococcal surface protein with the Cell Wall Anchored motif of LPXTG (where X is any amino acid). Any protein within this class of pneumococcal surface protein with this motif has been found to be useful within the context of this invention, and is therefore considered a further protein of the invention. Sp125 itself is disclosed in WO 98/18930, and is also known as ZmpB – a zinc metalloproteinase.

Sp101 is disclosed in WO 98/06734 (where it has the reference # y85993. It is characterised by a Type I signal sequence.

Sp133 is disclosed in WO 98/06734 (where it has the reference # y85992. It is also characterised by a Type I signal sequence.

Sp128 and Sp130 are disclosed in WO 00/76540.

The proteins used in the present invention are preferably selected from the group PhtD, PhtA and PhtE, or a combination of 2 or all 3 of these proteins (i.e. PhtA+D, A+E, D+E or A+D+E).

Further pneumococcal protein antigens that may be included are one or more from the group consisting of: pneumolysin (also referred to as Ply; preferably detoxified by chemical treatment or mutation) [WO 96/05859, WO 90/06951, WO 99/03884], PsaA and transmembrane deletion variants thereof (Berry & Paton, Infect Immun 1996 Dec;64(12):5255-62), PspA and transmembrane deletion variants thereof (US 5804193, WO 92/14488, WO 99/53940), PspC and transmembrane deletion variants thereof (WO 97/09994, WO 99/53940), a member of the Choline binding protein (Cbp) family [e.g. CbpA and transmembrane deletion variants thereof (WO 97/41151; WO 99/51266)], Glyceraldehyde–3-phosphate – dehydrogenase (Infect. Immun. 1996 64:3544), HSP70 (WO 96/40928), PcpA (Sanchez-Beato et al. *FEMS Microbiol Lett* 1998, 164:207-14), M like protein (SB patent application No. EP 0837130), and adhesin 18627 (SB Patent application No. EP 0834568). The present invention also encompasses immunologically functional equivalents or truncates of such proteins (as defined above).

Concerning the Choline Binding Protein family, members of that family were originally identified as pneumococcal proteins that could be purified by choline-affininty chromatography. All of the choline-binding proteins are non-covalently bound to phosphorylcholine moieties of cell wall teichoic acid and membrane-associated lipoteichoic acid. Structurally, they have several regions in common over the entire family, although the exact nature of the proteins (amino acid sequence, length, etc.) can vary. In general, choline binding proteins comprise an N terminal region (N), conserved repeat

regions (R1 and/or R2), a proline rich region (P) and a conserved choline binding region (C), made up of multiple repeats, that comprises approximately one half of the protein. As used in this application, the term "Choline Binding Protein family (Cbp)" is selected from the group consisting of Choline Binding Proteins as identified in WO 97/41151, PbcA, SpsA, PspC, CbpA, CbpD, and CbpG. CbpA is disclosed in WO 97/41151. CbpD and CbpG are disclosed in WO 00/29434. PspC is disclosed in WO 97/09994. PbcA is disclosed in WO 98/21337. Preferably the Choline Binding Proteins are selected from the group consisting of CbpA, PbcA, SpsA and PspC.

If a Cbp is the further protein utilised it may be a Cbp truncate wherein "Cbp" is defined above and "truncate" refers to proteins lacking 50% or more of the Choline binding region (C). Preferably such proteins lack the entire choline binding region. More preferably, the such protein truncates lack (i) the choline binding region and (ii) a portion of the N-terminal half of the protein as well, yet retain at least one repeat region (R1 or R2). More preferably still, the truncate has 2 repeat regions (R1 and R2). Examples of such preferred embodiments are NR1xR2, R1xR2, NR1xR2P and R1xR2P as illustrated in WO99/51266 or WO99/51188, however, other choline binding proteins lacking a similar choline binding region are also contemplated within the scope of this invention.

Cbp truncate-Lyt truncate chimeric proteins (or fusions) may also be used in the composition of the invention. Preferably this comprises NR1xR2 (or R1xR2 or NR1xR2P or R1xR2P) of Cbp and the C-terminal portion (Cterm, i.e., lacking the choline binding domains) of Lyt (e.g., LytCCterm or Sp91Cterm). More preferably Cbp is selected from the group consisting of CbpA, PbcA, SpsA and PspC. More preferably still, it is CbpA. Preferably, Lyt is LytC (also referred to as Sp91).

A PspA or PsaA truncate lacking the choline binding domain (C) and expressed as a fusion protein with Lyt may also be used. Preferably, Lyt is LytC.

In a pneumococcal composition it is possible to combine different pneumococcal proteins of the invention.

Preferably the combination of proteins of the invention are selected from 2 or more (3 or 4) different categories such as proteins having a Type II Signal sequence motif of LXXC (where X is any amino acid, e.g., the polyhistidine triad family (Pht)), choline binding proteins (Cbp), proteins having a Type I Signal sequence motif (e.g., Sp101), proteins having a LPXTG motif (where X is any amino acid, e.g., Sp128, Sp130), toxins (e.g., Ply), etc. Preferred examples within these categories (or motifs) are the proteins mentioned above, or immunologically functional equivalents thereof. Toxin + Pht, toxin + Cbp, Pht + Cbp, and toxin + Pht + Cbp are preferred category combinations.

Preferred beneficial combinations include, but are not limited to, PhtD + NR1xR2, PhtD + NR1xR2-Sp91Cterm chimeric or fusion proteins, PhtD + Ply, PhtD + Sp128, PhtD + PsaA, PhtD + PspA, PhtA + NR1xR2, PhtA + NR1xR2-Sp91Cterm chimeric or fusion proteins, PhtA + Ply, PhtA + Sp128, PhtA + PsaA, PhtA + PspA, NR1xR2 + LytC, NR1xR2 + PspA, NR1xR2 + PsaA, NR1xR2 + Sp128, R1xR2 + LytC, R1xR2 + PspA, R1xR2 + PsaA, R1xR2 + Sp128, R1xR2 + PhtD, R1xR2 + PhtA. Preferably, NR1xR2 (or R1xR2) is from CbpA or PspC. More preferably it is from CbpA.

A particularly preferred combination of pneumococcal proteins comprises Ply (or a truncate or immunologically functional equivalent thereof) + PhtD (or a truncate or immunologically functional equivalent thereof) optionally with NR1xR2 (or R1xR2 or NR1xR2P or R1xR2P). Preferably, NR1xR2 (or R1xR2 or NR1xR2P or R1xR2P) is from CbpA or PspC. More preferably it is from CbpA.

The antigen may be a pneumococcus saccharide conjugate comprising polysaccharide antigens derived from at least four serotypes, preferably at least seven serotypes, more preferably at least eleven serotypes, and at least one, but preferably 2, 3, or 4, *Streptococcus pneumoniae* proteins preferably selected from the group of proteins described above. Preferably one of the proteins is PhtD (or an immunologically functional equivalent thereof) and/or Ply (or an immunologically functional equivalent thereof).

A problem associated with the polysaccharide approach to vaccination, is the fact that polysaccharides *per se* are poor immunogens. To overcome this, saccharides may be conjugated to protein carriers, which provide bystander T-cell help. It is preferred, therefore, that the saccharides utilised in the invention are linked to such a protein carrier. Examples of such carriers which are currently commonly used for the production of saccharide immunogens include the Diphtheria and Tetanus toxoids (DT, DT CRM197 and TT respectively), Keyhole Limpet Haemocyanin (KLH), OMPC from *N. meningitidis*, and the purified protein derivative of Tuberculin (PPD).

A preferred carrier for the pneumococcal saccharide based immunogenic compositions (or vaccines) is protein D from *Haemophilus influenzae* (EP 594610-B), or fragments thereof. Fragments suitable for use include fragments encompassing T-helper epitopes. In particular a protein D fragment will preferably contain the N-terminal 1/3 of the protein. A protein D carrier is useful as a carrier in compositions where multiple pneumococcal saccharide antigens are conjugated. One or more pneumococcal saccharides in a combination may be advantageously conjugated onto protein D.

A further preferred carrier for the pneumococcal saccharide is the pneumococcal protein itself (as defined above in section "Pneumococcal Proteins of the invention").

The saccharide may be linked to the carrier protein by any known method (for example, by Likhite, U.S. Patent 4,372,945 and by Armor *et al.*, U.S. Patent 4,474,757). Preferably, CDAP conjugation is carried out (WO 95/08348).

Preferably the protein:saccharide (weight:weight) ratio of the conjugates is 0.3:1 to 1:1, more preferably 0.6:1 to 0.8:1, and most preferably about 0.7:1.

Particularly preferred compositions of the invention comprise one or more conjugated pneumococcal saccharides, and one or more pneumococcal proteins of the inventionIn addition, pneumococcal saccharides and proteins can be stably stored as bulk components adsorbed onto aluminium phosphate in a liquid form.

Other antigens are suitably derived from HIV-1, (such as gag or fragments thereof such as p24, tat, nef, gp120 or gp160 or fragments of any of these), human herpes viruses, such as gD or derivatives thereof or Immediate Early protein such as ICP27 from HSV1 or HSV2, cytomegalovirus ((esp Human)(such as gB or derivatives thereof), Rotavirus (including live-attenuated viruses), Epstein Barr virus (such as gp350 or derivatives thereof), Varicella Zoster Virus (such as gpI, II and IE63), or from a hepatitis virus such as hepatitis B virus (for example Hepatitis B Surface antigen or a derivative thereof), hepatitis A virus, hepatitis C virus and hepatitis E virus, or from other viral pathogens, such as paramyxoviruses: Respiratory Syncytial virus (such as F, N, M and G proteins or derivatives thereof), parainfluenza virus, measles virus, mumps virus, human papilloma viruses (for example HPV6, 11, 16, 18,), flaviviruses (e.g. Yellow Fever Virus, Dengue Virus, Tick-borne encephalitis virus, Japanese Encephalitis Virus) or Influenza virus (whole live or inactivated virus, split influenza virus, grown in eggs or MDCK cells, or whole flu virosomes (as described by R. Gluck, Vaccine, 1992, 10, 915-920) or purified or recombinant proteins thereof, such as HA, NP, NA, or M proteins, or combinations thereof), or derived from bacterial pathogens such as *Neisseria spp,* including *N. gonorrhea* and *N. meningitidis* (for example capsular saccharides and conjugates thereof, transferrin-binding proteins, lactoferrin binding proteins, PilC, adhesins); *S. pyogenes* (for example M proteins or fragments thereof, C5A protease, lipoteichoic acids), *S. agalactiae, S. mutans; H. ducreyi; Moraxella spp, including M catarrhalis, also known as Branhamella* catarrhalis (for example high and low molecular weight adhesins and invasins*); Bordetella spp, including B. pertussis* (for example pertactin, pertussis toxin or derivatives thereof, filamenteous hemagglutinin, adenylate cyclase, fimbriae), *B. parapertussis and B. bronchiseptica; Mycobacterium spp., including M. tuberculosis* (for example ESAT6, Antigen 85A, -B or –C), *M. bovis, M. leprae, M. avium, M. paratuberculosis, M. smegmatis; Legionella spp,* including *L. pneumophila; Escherichia spp,* including

enterotoxic E. coli (for example colonization factors, heat-labile toxin or derivatives thereof, heat-stable toxin or derivatives thereof), enterohemorragic *E. coli,* enteropathogenic *E. coli* (for example shiga toxin-like toxin or derivatives thereof); *Vibrio spp,* including *V. cholera* (for example cholera toxin or derivatives thereof*); Shigella spp,* including *S. sonnei, S. dysenteriae, S. flexnerii; Yersinia spp,* including *Y.* enterocolitica (for example a Yop protein) , *Y. pestis, Y. pseudotuberculosis; Campylobacter spp,* including *C. jejuni* (for example toxins, adhesins and invasins) and *C. coli; Salmonella spp,* including *S. typhi, S. paratyphi, S. choleraesuis, S. enteritidis; Listeria spp.,* including *L. monocytogenes; Helicobacter spp,* including *H.* pylori (for example urease, catalase, vacuolating toxin); *Pseudomonas spp,* including *P. aeruginosa; Staphylococcus spp.,* including *S. aureus, S. epidermidis; Enterococcus spp.,* including *E. faecalis, E. faecium; Clostridium spp.,* including *C.* tetani (for example tetanus toxin and derivative thereof), *C. botulinum* (for example botulinum toxin and derivative thereof*), C. difficile* (for example clostridium toxins A or B and derivatives thereof); *Bacillus spp.,* including *B. anthracis* (for example botulinum toxin and derivatives thereof*); Corynebacterium spp.,* including *C. diphtheriae* (for example diphtheria toxin and derivatives thereof); *Borrelia spp.,* including *B. burgdorferi* (for example OspA, OspC, DbpA, DbpB), *B. garinii* (for example OspA, OspC, DbpA, DbpB), *B. afzelii* (for example OspA, OspC, DbpA, DbpB*), B. andersonii* (for example OspA, OspC, DbpA, DbpB), *B. hermsii; Ehrlichia spp.,* including *E. equi* and the agent of the Human Granulocytic Ehrlichiosis*; Rickettsia spp,* including *R. rickettsii; Chlamydia spp., including C. trachomatis* (for example MOMP, heparin-binding proteins), *C. pneumoniae* (for example MOMP, heparin-binding proteins*), C. psittaci; Leptospira spp.,* including *L. interrogans; Treponema spp.,* including *T. pallidum* (for example the rare outer membrane proteins*), T. denticola, T. hyodysenteriae;* or derived from parasites such as *Plasmodium spp.,* including *P. falciparum; Toxoplasma spp.,* including *T. gondii (for example SAG2, SAG3, Tg34); Entamoeba spp.,* including *E. histolytica; Babesia spp.,* including *B. microti; Trypanosoma spp.,* including *T. cruzi; Giardia spp.,* including *G. lamblia; Leshmania spp.,* including *L. major; Pneumocystis spp.,* including *P. carinii; Trichomonas spp.,* including *T. vaginalis; Schisostoma spp.,* including *S. mansoni,* or derived from yeast such as *Candida spp.,* including *C. albicans; Cryptococcus spp.,* including *C. neoformans.*

Other preferred specific antigens for *M. tuberculosis* are for example Tb Ra12, Tb H9, Tb Ra35, Tb38-1, Erd 14, DPV, MTI, MSL, mTTC2 and hTCC1 (WO 99/51748). Proteins for *M. tuberculosis* also include fusion proteins and variants thereof where at least two, preferably three polypeptides of *M. tuberculosis* are fused into a larger protein. Preferred fusions include Ra12-TbH9-Ra35, Erd14-DPV-MTI, DPV-MTI-MSL, Erd14-DPV-MTI-

MSL-mTCC2, Erd14-DPV-MTI-MSL, DPV-MTI-MSL-mTCC2, TbH9-DPV-MTI (WO 99/51748).

Most preferred antigens for Chlamydia include for example the High Molecular Weight Protein (HWMP) (WO 99/17741), ORF3 (EP 366 412), and putative membrane proteins (Pmps). Other Chlamydia antigens of the composition can be selected from the group described in WO 99/28475.

Preferred bacterial compositions comprise antigens derived from *Haemophilus spp., including H. influenzae type B* (for example PRP and conjugates thereof), *non typeable H. influenzae,* for example OMP26, high molecular weight adhesins, P5, P6, protein D and lipoprotein D, and fimbrin and fimbrin derived peptides (US 5,843,464) or multiple copy varients or fusion proteins thereof.

Derivatives of Hepatitis B Surface antigen are well known in the art and include, inter alia, those PreS1, PreS2 S antigens set forth described in European Patent applications EP-A-414 374; EP-A-0304 578, and EP 198-474. In one preferred aspect the vaccine formulation of the invention comprises the HIV-1 antigen, gp120, especially when expressed in CHO cells. In a further embodiment, the composition of the invention comprises gD2t as hereinabove defined.

In a preferred embodiment of the present invention compositions contain an antigen derived from the Human Papilloma Virus (HPV) considered to be responsible for genital warts *(*HPV 6 or HPV 11 and others*)*, and the HPV viruses responsible for cervical cancer (HPV16, HPV18 and others*)*.

Particularly preferred forms of genital wart prophylactic, or therapeutic, compositions comprise L1 particles or capsomers, and fusion protein*s* comprising one or more antigens selected from the HPV proteins E1, E2, E5 E6, E7, L1, and L2.

The most preferred forms of fusion protein are: L2E7 as disclosed in WO 96/26277, and proteinD(1/3)-E7 disclosed in GB 9717953.5 (PCT/EP98/05285).

A preferred HPV cervical infection or cancer, prophylaxis or therapeutic compositions may comprise HPV 16 or 18 antigens. For example, L1 or L2 antigen monomers, or L1 or L2 antigens presented together as a virus like particle (VLP) or the L1 alone protein presented alone in a VLP or caposmer structure. Such antigens, virus like particles and capsomer are per se known. See for example WO94/00152, WO94/20137, WO94/05792, and WO93/02184.

Additional early proteins may be included alone or as fusion proteins such as E7, E2 or preferably E5 for example; particularly preferred embodiments of this includes a VLP comprising L1E7 fusion proteins (WO 96/11272).

Particularly preferred HPV 16 antigens comprise the early proteins E6 or E7 in fusion with a protein D carrier to form Protein D - E6 or E7 fusions from HPV 16, or combinations thereof; or combinations of E6 or E7 with L2 (WO 96/26277).

Alternatively the HPV 16 or 18 early proteins E6 and E7, may be presented in a single molecule, preferably a Protein D- E6/E7 fusion. Such a composition may optionally contain either or both E6 and E7 proteins from HPV 18, preferably in the form of a Protein D - E6 or Protein D - E7 fusion protein or Protein D E6/E7 fusion protein.

The composition of the present invention may additionally comprise antigens from other HPV strains, preferably from strains HPV 31 or 33.

Compositions of the present invention further comprise antigens derived from parasites that cause Malaria. For example, preferred antigens from *Plasmodia falciparum* include circumsporozoite protein (CS protein), RTS,S MSP1, MSP3, LSA1, LSA3, AMA1 and TRAP. RTS is a hybrid protein comprising substantially all the C-terminal portion of the circumsporozoite (CS) protein of *P.falciparum* linked via four amino acids of the preS2 portion of Hepatitis B surface antigen to the surface (S) antigen of hepatitis B virus. Its full structure is disclosed in the International Patent Application No. PCT/EP92/02591, published under Number WO 93/10152 claiming priority from UK patent application No.9124390.7. When expressed in yeast RTS is produced as a lipoprotein particle, and when it is co-expressed with the S antigen from HBV it produces a mixed particle known as RTS,S. TRAP antigens are described in the International Patent Application No. PCT/GB89/00895, published under WO 90/01496. A preferred embodiment of the present invention is a Malaria vaccine wherein the antigenic preparation comprises a combination of the RTS,S and TRAP antigens. Other plasmodia antigens that are likely candidates to be components of a multistage Malaria vaccine are *P. faciparum* MSP1, AMA1, MSP3, EBA, GLURP, RAP1, RAP2, Sequestrin, PfEMP1, Pf332, LSA1, LSA3, STARP, SALSA, PfEXP1, Pfs25, Pfs28, PFS27/25, Pfs16, Pfs48/45, Pfs230 and their analogues in Plasmodium spp. . One embodiment of the present invention is a composition comprising RTS, S or CS protein or a fragment thereof such as the CS portion of RTS, S in combination with one or more further malarial antigens which may be selected for example from the group consisting of MPS1, MSP3, AMA1, LSA1 or LSA3.

The compositions may also contain an anti-tumour antigen and be useful for the immunotherapeutic treatment of cancers. For example, the antigen may be a tumour rejection antigens such as those for prostrate, breast, colorectal, lung, pancreatic, renal or melanoma cancers. Exemplary antigens include MAGE 1 , 3 and MAGE 4 or other MAGE antigens such as disclosed in WO99/40188, PRAME, BAGE, Lage (also known as NY Eos 1) SAGE and HAGE (WO 99/53061) or GAGE (Robbins and Kawakami, 1996,

Current Opinions in Immunology 8, pps 628-636; Van den Eynde et al., International Journal of Clinical & Laboratory Research (submitted 1997); Correale et al. (1997), Journal of the National Cancer Institute 89, p293. Indeed these antigens are expressed in a wide range of tumour types such as melanoma, lung carcinoma, sarcoma and bladder carcinoma.

MAGE antigens for use in the present invention may be expressed as a fusion protein with an expression enhancer or an Immunological fusion partner. In particular, the Mage protein may be fused to Protein D from Heamophilus infuenzae B or a lipidated derivative thereof.. In particular, the fusion partner may comprise the first 1/3 of Protein D. Such constructs are disclosed in Wo99/40188.

Other tumour-specific antigens include, but are not restricted to KSA (GA733) tumour-specific gangliosides such as GM 2, and GM3 or conjugates thereof to carrier proteins; or said antigen may be a self peptide hormone such as whole length Gonadotrophin hormone releasing hormone (GnRH, WO 95/20600), a short 10 amino acid long peptide, useful in the treatment of many cancers, or in immunocastration.

In a preferred embodiment prostate antigens are utilised, such as Prostate specific antigen (PSA), PAP, PSCA (PNAS 95(4) 1735 –1740 1998), PSMA or antigen known as Prostase.

Prostase is a prostate-specific serine protease (trypsin-like), 254 amino acid-long, with a conserved serine protease catalytic triad H-D-S and a amino-terminal pre-propeptide sequence, indicating a potential secretory function (P. Nelson, Lu Gan, C. Ferguson, P. Moss, R. Gelinas, L. Hood & K. Wand, "Molecular cloning and characterisation of prostase, an androgen-regulated serine protease with prostate restricted expression, *In* Proc. Natl. Acad. Sci. USA (1999) 96, 3114-3119). A putative glycosylation site has been described. The predicted structure is very similar to other known serine proteases, showing that the mature polypeptide folds into a single domain. The mature protein is 224 amino acids-long, with one A2 epitope shown to be naturally processed.

Prostase nucleotide sequence and deduced polypeptide sequence and homologs are disclosed in Ferguson, et al. (Proc. Natl. Acad. Sci. USA 1999, 96, 3114-3119) and in International Patent Applications No. WO 98/12302 (and also the corresponding granted patent US 5,955,306), WO 98/20117 (and also the corresponding granted patents US 5,840,871 and US 5,786,148) (prostate-specific kallikrein) and WO 00/04149 (P703P).

The present invention provides compositions comprising prostase protein fusions based on prostase protein and fragments and homologues thereof ("derivatives"). Such derivatives are suitable for use in therapeutic vaccine formulations which are suitable for the treatment of a prostate tumours. Typically the fragment will contain at least 20,

preferably 50, more preferably 100 contiguous amino acids as disclosed in the above referenced patent and patent applications.

A further preferred prostate antigen is known as P501S, sequence ID no 113 of Wo98/37814. Immunogenic fragments and portions thereof comprising at least 20, preferably 50, more preferably 100 contiguous amino acids as disclosed in the above referenced patent application. See for example PS108 (WO 98/50567).

Other prostate specific antigens are known from Wo98/37418, and WO/004149. Another is STEAP PNAS 96 14523 14528 7 –12 1999.

Other tumour associated antigens useful in the context of the present invention include: Plu –1 J Biol. Chem 274 (22) 15633 –15645, 1999, HASH –1, HasH-2, Cripto (Salomon et al Bioessays 199, 21 61 –70,US patent 5654140) Criptin US patent 5 981 215, . Additionally, antigens particularly relevant for therapy of cancer also comprise tyrosinase and survivin.

Mucin dervied peptides such as Muc1 see for example US 5744,144 US 5827, 666 WO 8805054, US 4,963,484. Specifically contemplated are Muc 1 derived peptides that comprise at least one repeat unit of the the Muc 1 peptide, preferably at least two such repeats and which is recognised by the SM3 antibody (US 6 054 438). Other mucin derived peptides include peptide from Muc 5.

The antigen of the invention may be a breast cancer antigens such as her 2/ Neu, mammaglobin (US patent 5668267) or those disclosed in WO/00 52165, WO99/33869, WO99/19479, WO 98/45328. Her 2 neu antigens are disclosed inter alia, in US patent 5,801,005. Preferably the Her 2 neu comprises the entire extracellular domain (comprising approximately amino acid 1 –645) or fragments thereof and at least an immunogenic portion of or the entire intracellular domain approximately the C terminal 580 amino acids . In particular, the intracellular portion should comprise the phosphorylation domain or fragments thereof. Such constructs are disclosed in WO00/44899. A particularly preferred construct is known as ECD PD a second is known as ECD PD See Wo/00/44899. The her 2 neu as used herein can be derived from rat, mouse or human .

The compositions may contain antigens associated with tumour-support mechanisms (e.g. angiogenesis, tumour invasion) for example tie 2, VEGF.

It is foreseen that compositions of the present invention may use antigens derived from *Borrelia sp.*. For example, antigens may include nucleic acid, pathogen derived antigen or antigenic preparations, recombinantly produced protein or peptides, and chimeric fusion proteins. In particular the antigen is OspA. The OspA may be a full mature protein in a lipidated form virtue of the host cell (E.Coli) termed (Lipo-OspA) or a non-lipidated

derivative. Such non-lipidated derivatives include the non-lipidated NS1-OspA fusion protein which has the first 81 N-terminal amino acids of the non-structural protein (NS1) of the influenza virus, and the complete OspA protein, and another, MDP-OspA is a non-lipidated form of OspA carrying 3 additional N-terminal amino acids.

Compositions of the present invention may be used for the prophylaxis or therapy of allergy. Such vaccines would comprise allergen specific (for example Der p1) and allergen non-specific antigens (for example peptides derived from human IgE, including but not restricted to the stanworth decapeptide (EP 0 477 231 B1)).

Compositions of the present invention may also be used for the prophylaxis or therapy of chronic disorders others than allergy, cancer or infectious diseases. Such chronic disorders are diseases such as atherosclerosis, and Alzheimer.

The compositions of the present invention are particularly suited for the immunotherapeutic treatment of diseases, such as chronic conditions and cancers, but also for the therapy of persistent infections. Accordingly the compositions of the present invention are particularly suitable for the immunotherapy of infectious diseases, such as Tuberculosis (TB), AIDS and Hepatitis B (HepB) virus infections.

Also, in the context of AIDS, there is provided a method of treatment of an individual susceptible to or suffering from AIDS. The method comprising the administration of a vaccine of the present invention to the individual, thereby reducing the amount of CD4+ T-cell decline caused by subsequent HIV infection, or slowing or halting the CD4+ T-cell decline in an individual already infected with HIV.

Other antigens include bacterial (preferably capsular) saccharides other than (or in addition to) those pneumococcal antigens described above. Polysaccharide antigens are conveniently stored in liquid bulk adsorbed onto aluminium phosphate – it is therefore straightforward to generate vaccine compositions of the invention by admixing said liquid bulk with the oil emulsions of the invention extemporaneously. Preferably the other bacterial saccharides are selected from a group consisting of: *N. meningitidis* serogroup A capsular saccharide (MenA), *N. meningitidis* serogroup C capsular saccharide (MenC), *N. meningitidis* serogroup Y capsular saccharide (MenY), *N. meningitidis* serogroup W-135 capsular saccharide (MenW), Group B Streptococcus group I capsular saccharide, Group B Streptococcus group II capsular saccharide, Group B Streptococcus group III capsular saccharide, Group B Streptococcus group IV capsular saccharide, Group B Streptococcus group V capsular saccharide, *Staphylococcus aureus* type 5 capsular saccharide, *Staphylococcus aureus* type 8 capsular saccharide, Vi saccharide from *Salmonella typhi*, *N. meningitidis* LPS, *M. catarrhalis* LPS, and *H. influenzae* LPS. By LPS it is meant either

native lipo-polysaccharide (or lipo-oligosaccharide), or lipo-polysaccharide where the lipid A portion has been detoxified by any of a number of known methods (see for example WO 97/18837 or WO 98/33923), or any molecule comprising the O-polysaccharide derived from said LPS. By *N. meningitidis* LPS it is meant one or more of the 12 known immunotypes (L1, L2, L3, L4, L5, L6, L7, L8, L9, L10, L11 or L12).

Particularly preferred combinations are compositions comprising: 1) conjugated Hib, conjugated MenA and conjugated MenC; 2) conjugated Hib, conjugated MenY and conjugated MenC; 3) conjugated Hib and conjugated MenC; and 4) conjugated MenA, conjugated MenC, conjugated MenY and conjugated MenW-135. The amount of PS in each of the above conjugates may be 5 or 10 □g each per 0.5 mL human dose. Preferably Hib, MenA, MenC, MenW-135 and MenY are TT conjugates.

A problem associated with the polysaccharide approach to vaccination, is the fact that polysaccharides *per se* are poor immunogens. To overcome this, saccharides of the invention may be conjugated to protein carriers, which provide bystander T-cell help. It is preferred, therefore, that the saccharides utilised in the invention are linked to such a protein carrier. Examples of such carriers which are currently commonly used for the production of saccharide immunogens include the Diphtheria and Tetanus toxoids (DT, DT CRM197 and TT respectively), Keyhole Limpet Haemocyanin (KLH), protein D from *Haemophilus influenzae* (EP 594610-B), OMPC from *N. meningitidis*, and the purified protein derivative of Tuberculin (PPD).

The saccharide may be linked to the carrier protein by any known method (for example, by Likhite, U.S. Patent 4,372,945 and by Armor *et al.*, U.S. Patent 4,474,757). Preferably, CDAP conjugation is carried out (WO 95/08348).

Preferably the protein:saccharide (weight:weight) ratio of the conjugates is 0.3:1 to 1:1, more preferably 0.6:1 to 0.8:1, and most preferably about 0.7:1.

Combinations of antigens which provide protection against pneumococcus and a different pathogen are included in the present invention. Many Paediatric vaccines are now given as a combination vaccine so as to reduce the number of injections a child has to receive. Thus for Paediatric vaccines other antigens from other pathogens may be formulated with the pneumococcal vaccines of the invention. For example the vaccines of the invention can be formulated with (or administered separately but at the same time) the well known 'trivalent' combination vaccine comprising Diphtheria toxoid (DT), tetanus toxoid (TT), and pertussis components [typically detoxified Pertussis toxoid (PT) and filamentous haemagglutinin (FHA) with optional pertactin (PRN) and/or agglutinin 1+2], for example the marketed vaccine INFANRIX-DTPa™ (SmithKlineBeecham Biologicals) which

contains DT, TT, PT, FHA and PRN antigens, or with a whole cell pertussis component for example as marketed by SmithKlineBeecham Biologicals s.a., as Tritanrix™. The combined vaccine may also comprise other antigen, such as Hepatitis B surface antigen (HBsAg), Polio virus antigens (for instance inactivated trivalent polio virus – IPV), *Moraxella catarrhalis* outer membrane proteins, non-typeable *Haemophilus influenzae* proteins, *N.meningitidis* B outer membrane proteins.

Examples of preferred *Moraxella catarrhalis* protein antigens which can be included in a combination vaccine (especially for the prevention of otitis media) are: OMP106 [WO 97/41731 (Antex) & WO 96/34960 (PMC)]; OMP21; LbpA &/or LbpB [WO 98/55606 (PMC)]; TbpA &/or TbpB [WO 97/13785 & WO 97/32980 (PMC)]; CopB [Helminen ME, *et al.* (1993) Infect. Immun. 61:2003-2010]; UspA1 and/or UspA2 [WO 93/03761 (University of Texas)]; OmpCD; HasR (PCT/EP99/03824); PilQ (PCT/EP99/03823); OMP85 (PCT/EP00/01468); lipo06 (GB 9917977.2); lipo10 (GB 9918208.1); lipo11 (GB 9918302.2); lipo18 (GB 9918038.2); P6 (PCT/EP99/03038); D15 (PCT/EP99/03822); OmplA1 (PCT/EP99/06781); Hly3 (PCT/EP99/03257); and OmpE. Examples of non-typeable *Haemophilus influenzae* antigens which can be included in a combination vaccine (especially for the prevention of otitis media) include: Fimbrin protein [(US 5766608 - Ohio State Research Foundation)] and fusions comprising peptides therefrom [eg LB1(f) peptide fusions; US 5843464 (OSU) or WO 99/64067]; OMP26 [WO 97/01638 (Cortecs)]; P6 [EP 281673 (State University of New York)]; TbpA and/or TbpB; Hia; Hsf; Hin47; Hif; Hmw1; Hmw2; Hmw3; Hmw4; Hap; D15 (WO 94/12641); protein D (EP 594610); P2; and P5 (WO 94/26304).

Other combinations contemplated are the pneumococcal saccharide & protein of the invention in combination with viral antigens, for example, from influenza (attenuated, split, or subunit [e.g., surface glycoproteins neuraminidase (NA) and haemagglutinin (HA). See, e.g., Chaloupka I. et al, Eur. Journal Clin. Microbiol. Infect. Dis. 1996, 15:121-127], RSV (e.g., F and G antigens or F/G fusions, see, eg, Schmidt A. C. et al, J Virol, May 2001, p4594 – 4603), PIV3 (e.g., HN and F proteins, see Schmidt et al. supra), Varicella (e.g., attenuated, glycoproteins I-V, etc.), and any (or all) component(s) of MMR (measles, mumps, rubella).

A preferred Paediatric combination vaccine contemplated by the present invention for global treatment or prevention of otitis media comprises: one or more *Streptococcus pneumoniae* saccharide antigen(s) (preferably conjugated to protein D), one or more pneumococcal proteins (preferably those described above), and one or more surface-exposed antigen from *Moraxella catarrhalis* and/or non-typeable *Haemophilus influenzae.* Protein D can advantageously be used as a protein carrier for the pneumococcal

saccharides (as mentioned above), and because it is in itself an immunogen capable of producing B-cell mediated protection against non-typeable *H. influenzae* (ntHi). The *Moraxella catarrhalis* or non-typeable *Haemophilus influenzae* antigens can be included in the vaccine in a sub-unit form, or may be added as antigens present on the surface of outer membrane vesicles (blebs) made from the bacteria.

Preferred antigens

As mentioned above, in one aspect the invention relates to use of a composition comprising:

(a) an antigen, and

(d) an oil-in-water emulsion adjuvant; and

(e) 3D MPL

wherein said oil-in-water emulsion comprises a metabolisable oil, a sterol and an emulsifying agent, in the manufacture of an immunogenic composition for the prevention of infection and/or disease.

The composition of the invention is thus used for infections and/or diseases which are capable of being prevented or ameliorated by that composition, and suitably in which the antigen is derived from or associated with a pathogen (such as a bacteria or virus) which is associated with the disease.

Antigens from influenza virus A and B, HPV antigens, RSV A and B, SARS, streptococcus, VZV, rhinovirus, parainfluenza virus are preferred for use in the present invention, such as split influenza , VZV gE, VZV IE63, and PhtD from Streptococcus pneumonia. However, any suitable antigen may be used.

In one embodiment the composition used in the invention does not comprise an influenza subunit antigen with the MF59™ adjuvant.

For all aspects of the invention it is preferred that antigens comprise a CD4 T cell epitope, or a B cell epitope or suitably both.

Influenza viral strains and antigens

An influenza virus or antigenic preparation thereof for use according to the present invention may be a split influenza virus or split virus antigenic preparation thereof. In an alternative embodiment the influenza preparation may contain another type of inactivated influenza antigen, such as inactivated whole virus or purified HA and NA (subunit vaccine), or an influenza virosome. In a still further embodiment, the influenza virus may be a live attenuated influenza preparation.

A split influenza virus or split virus antigenic preparation thereof for use according to the present invention is suitably an inactivated virus preparation where virus particles are disrupted with detergents or other reagents to solubilise the lipid envelope. Split virus or split virus antigenic preparations thereof are suitably prepared by fragmentation of whole influenza virus, either infectious or inactivated, with solubilising concentrations of organic solvents or detergents and subsequent removal of all or the majority of the solubilising agent and some or most of the viral lipid material. By split virus antigenic preparation thereof is meant a split virus preparation which may have undergone some degree of purification compared to the split virus whilst retaining most of the antigenic properties of the split virus components. For example, when produced in eggs, the split virus may be depleted from egg-contaminating proteins, or when produced in cell culture, the split virus may be depleted from host cell contaminants. A split virus antigenic preparation may comprise split virus antigenic components of more than one viral strain. Vaccines containing split virus (called 'influenza split vaccine') or split virus antigenic preparations generally contain residual matrix protein and nucleoprotein and sometimes lipid, as well as the membrane envelope proteins. Such split virus vaccines will usually contain most or all of the virus structural proteins although not necessarily in the same proportions as they occur in the whole virus.

In another embodiment, the influenza virus preparation is in the form of a purified sub-unit influenza. Sub-unit influenza vaccines generally contain the two major envelope proteins, HA and NA, and may have an additional advantage over whole virion vaccines as they are generally less reactogenic, particularly in young vaccinees. Sub-unit vaccines can be produced either recombinantly or purified from disrupted viral particles.

In another embodiment, the influenza virus preparation is in the form of a virosome. Virosomes are spherical, unilamellar vesicles which retain the functional viral envelope glycoproteins HA and NA in authentic conformation, intercalated in the virosomes' phospholipids bilayer membrane.

Said influenza virus or antigenic preparation thereof may be egg-derived or tissue-culture derived.

For example, the influenza virus antigen or antigenic preparations thereof according to the invention may be derived from the conventional embryonated egg method, by growing influenza virus in eggs and purifying the harvested allantoic fluid. Eggs can be accumulated in large numbers at short notice. Alternatively, they may be derived from any of the new generation methods using tissue culture to grow the virus or express recombinant influenza virus surface antigens. Suitable cell substrates for growing the virus include for example dog kidney cells such as MDCK or cells from a clone of MDCK, MDCK-like cells, monkey kidney cells such as AGMK cells including Vero cells, suitable pig cell lines, or any other mammalian cell type suitable for the production of influenza virus for vaccine purposes. Suitable cell substrates also include human cells e.g. MRC-5 cells. Suitable cell substrates are not limited to cell lines; for example primary cells such as chicken embryo fibroblasts and avian cell lines are also included.

The influenza virus antigen or antigenic preparation thereof may be produced by any of a number of commercially applicable processes, for example the split flu process described in patent no. DD 300 833 and DD 211 444, incorporated herein by reference. Traditionally split flu was produced using a solvent/detergent treatment, such as tri-*n*-butyl phosphate, or diethylether in combination with Tween™ (known as "Tween-ether" splitting) and this process is still used in some production facilities. Other splitting agents now employed include detergents or proteolytic enzymes or bile salts, for example sodium deoxycholate as described in patent no. DD 155 875, incorporated herein by reference. Detergents that can be used as splitting agents include cationic detergents e.g. cetyl trimethyl ammonium bromide (CTAB), other ionic detergents e.g. laurylsulfate, taurodeoxycholate, or non-ionic detergents such as the ones described above including Triton X-100 (for example in a process described in Lina et al, 2000, Biologicals 28, 95-103) and Triton N-101, or combinations of any two or more detergents.

The preparation process for a split vaccine may include a number of different filtration and/or other separation steps such as ultracentrifugation, ultrafiltration, zonal centrifugation and chromatography (e.g. ion exchange) steps in a variety of combinations, and optionally an inactivation step eg with heat, formaldehyde or β-propiolactone or U.V. which may be carried out before or after splitting. The splitting process may be carried

out as a batch, continuous or semi-continuous process. A preferred splitting and purification process for a split immunogenic composition is described in WO 02/097072.

Preferred split flu vaccine antigen preparations according to the invention comprise a residual amount of Tween 80 and/or Triton X-100 remaining from the production process, although these may be added or their concentrations adjusted after preparation of the split antigen. Preferably both Tween 80 and Triton X-100 are present. The preferred ranges for the final concentrations of these non-ionic surfactants in the vaccine dose are:
Tween 80: 0.01 to 1%, more preferably about 0.1% (v/v)
Triton X-100: 0.001 to 0.1 (% w/v), more preferably 0.005 to 0.02% (w/v).

In a specific embodiment, the final concentration for Tween 80 ranges from 0.045%-0.09% w/v. In another specific embodiment, the antigen is provided as a 2 fold concentrated mixture, which has a Tween 80 concentration ranging from 0.045%-0.2% (w/v) and has to be diluted two times upon final formulation with the adjuvanted (or the buffer in the control formulation).

In another specific embodiment, the final concentration for Triton X-100 ranges from 0.005%-0.017% w/v. In another specific embodiment, the antigen is provided as a 2 fold concentrated mixture, which has a Triton X-100 concentration ranging from 0.005%-0.034% (w/v) and has to be diluted two times upon final formulation with the adjuvanted (or the buffer in the control formulation).

Preferably the influenza preparation is prepared in the presence of low level of thiomersal, or preferably in the absence of thiomersal. Preferably the resulting influenza preparation is stable in the absence of organomercurial preservatives, in particular the preparation contains no residual thiomersal. In particular the influenza virus preparation comprises a haemagglutinin antigen stabilised in the absence of thiomersal, or at low levels of thiomersal (generally 5 μg/ml or less). Specifically the stabilization of B influenza strain is performed by a derivative of alpha tocopherol, such as alpha tocopherol succinate (also known as vitamin E succinate, i.e. VES). Such preparations and methods to prepare them are disclosed in WO 02/097072.

A preferred composition contains three inactivated split virion antigens prepared from the WHO recommended strains of the appropriate influenza season.

Preferably the influenza virus or antigenic preparation thereof and the oil-in-water emulsion adjuvant are contained in the same container. It is referred to as 'one vial approach'. Preferably the vial is a pre-filled syringe. In an alternative embodiment, the influenza virus or antigenic preparation thereof and the oil-in-water emulsion adjuvant are contained in separate containers or vials and admixed shortly before or upon administration into the subject. It is referred to as 'two vials approach'. By way of example, when the vaccine is a 2 components vaccine for a total dose volume of 0.7 ml, the concentrated antigens (for example the concentrated trivalent inactivated split virion antigens) are presented in one vial (335 µl) (antigen container) and a pre-filled syringe contains the adjuvant (360 µl) (adjuvant container). At the time of injection, the content of the vial containing the concentrated trivalent inactivated split virion antigens is removed from the vial by using the syringe containing the adjuvant followed by gentle mixing of the syringe. Prior to injection, the used needle is replaced by an intramuscular needle and the volume is corrected to 530 µl. One dose of the reconstituted adjuvanted influenza vaccine candidate corresponds to 530 µl.

Preferred compositions of the invention comprise antigens having CD4 T cell epitopes and optionally B cell epitopes.

HPV antigens

In another aspect of the present invention compositions contain an antigen derived from the Human Papilloma Virus (HPV), for example from a virus considered to be responsible for genital warts (HPV 6 or HPV 11 and others), or the HPV viruses responsible for cervical cancer (HPV16, HPV18 and others). In one aspect prophylactic or therapeutic compositions comprise HPV 16 or 18 antigens. Infection by HPV 16 and HPV 18 is related to development of cancer.

Combinations of antigens from different HPV genotypes may be employed in the invention, such as a combination of HPV 16 and HPV 18 antigens, suitably in the form of VLP. Antigens from additional VLP types that may be included with 16 and/or 18 include antigens from other known cancer-causing types such as HPV 31, 45, 33, 58 and 52.

In one aspect the HPV antigens are L1 or L2 antigen monomers. In one aspect the invention relates to a combination of HPV L1 and L2 antigens from the same genotype presented together as a capsomer or virus like particle (VLP). In one aspect the HPV

antigen is an L1 protein (absent an L2 antigen) in the form of a VLP or capsomer structure. Such antigens, virus like particles and capsomer are *per se* known. See, for example, WO94/00152, WO94/20137, WO94/05792, and WO93/02184.

In one aspect a truncated L1 protein may be used in the invention, for example as disclosed in WO 96/11272. Preferably a C-terminal truncation of L1 is used, for example a 34 amino acid C-terminal truncation of HPV 16, or an equivalent truncation from other HPV type.

In one aspect of the invention a composition comprises a combination of HPV virus like particles or capsomers from HPV 16 and HPV 18, together with HPV 31 and/or HPV 45.

In one aspect of the invention a composition comprises a combination of HPV virus like particles or capsomers from HPV 16 and HPV 18, together with HPV 33 and/or HPV 58.

In one aspect of the invention a composition comprises a combination of HPV virus like particles or capsomers from HPV 16 and HPV 18, together with VLPs or capsomers from one or more cancer causing HPV types, such as one, two, three, four or all of HPV 31, 33, 45, 52, and 58.

The invention thus relates in one aspect to a composition comprising virus like particles from HPV 16, 18, 31 and 45 in combination with an adjuvant comprising 3D MPL and an oil in water emulsion as described herein.

In one aspect of the invention a composition comprises a mixture of HPV 16, 18, 31, 33, 45, 52, and 58 L1-only virus like particles or capsomers. L1 or L2 proteins may be provided in the form of fusion proteins.

Particularly suitable forms of genital wart prophylactic, or therapeutic, compositions comprise L1 particles or capsomers, and fusion proteins comprising one or more antigens selected from the HPV 6 and HPV 11 proteins, for example E6, E7, L1, and L2.

HPV antigens from cancer types may be combined with antigens from genital warts types, such as HPV 16 and/or 18 with HPV 6 and/or 11. For example, a composition comprising HPV 16, 18, 6 and 11 is contemplated. In one aspect of the invention a combination of HPV 16, 18, 31, 33, 45, 52, and 58 L1-only virus like particle or capsomers may be used in combination with virus like particles or capsomers from HPV 6 and/or HPV 11.

In one aspect early proteins such as E7, E2 or E5 for example may be included alone, in combinations, or may be fusion proteins; an embodiment of this includes a VLP comprising L1E7 fusion proteins (WO 96/11272).

In one aspect the fusion protein is L2E7 as disclosed in WO 96/26277, or proteinD (1/3)-E7 disclosed in GB 9717953.5 (PCT/EP98/05285).
In one aspect HPV 16 antigens comprise the early proteins E6 or E7 in fusion with a protein D carrier to form Protein D - E6 or E7 fusions from HPV 16, or combinations thereof; or combinations of E6 or E7 with L2 (WO 96/26277).

Alternatively the HPV 16 or 18 early proteins E6 and E7, may be presented in a single molecule, such as a Protein D- E6/E7 fusion. Such a composition may optionally contain either or both E6 and E7 proteins from HPV 18, such as in the form of a Protein D - E6 or Protein D - E7 fusion protein or Protein D E6/E7 fusion protein.

Oil-in-water emulsion adjuvant component

The adjuvant composition of the invention contains an oil-in-water emulsion adjuvant, preferably said emulsion comprises a metabolisable oil in an amount of 0.5% to 20% of the total volume, and having oil droplets of which at least 70% by intensity have diameters of less than 1 μm.

In order for any oil in water composition to be suitable for human administration, the oil phase of the emulsion system has to comprise a metabolisable oil. The meaning of the term metabolisable oil is well known in the art. Metabolisable can be defined as 'being capable of being transformed by metabolism' (Dorland's Illustrated Medical Dictionary, W.B. Sanders Company, 25th edition (1974)). The oil may be any vegetable oil, fish oil, animal oil or synthetic oil, which is not toxic to the recipient and is capable of being transformed by metabolism. Nuts, seeds, and grains are common sources of vegetable oils. Synthetic oils are also part of this invention and can include commercially available oils such as NEOBEE® and others. A particularly suitable metabolisable oil is squalene. Squalene (2,6,10,15,19,23-Hexamethyl-2,6,10,14,18,22-tetracosahexaene) is an unsaturated oil which is found in large quantities in shark-liver oil, and in lower quantities in olive oil, wheat germ oil, rice bran oil, and yeast, and is a particularly preferred oil for use in this invention. Squalene is a metabolisable oil by virtue of the fact that it is an intermediate in the biosynthesis of cholesterol (Merck index, 10th Edition, entry no.8619).

Oil in water emulsions *per se* are well known in the art, and have been suggested to be useful as adjuvant compositions (EP 399843; WO 95/17210).

Suitably the metabolisable oil is present in an amount of 0.5% to 20% (final concentration) of the total volume of the immunogenic composition, preferably an amount of 1.0% to 10% of the total volume, preferably in an amount of 2.0% to 6.0% of the total volume.

In a specific embodiment, the metabolisable oil is present in a final amount of about 0.5%, 1%, 3.5% or 5% of the total volume of the immunogenic composition. In another specific embodiment, the metabolisable oil is present in a final amount of 0.5%, 1%, 3.57% or 5% of the total volume of the immunogenic composition.

Preferably the oil-in-water emulsion systems of the present invention have a small oil droplet size in the sub-micron range. Suitably the droplet sizes will be in the range 120 to 750 nm, more preferably sizes from 120 to 600 nm in diameter. Most preferably the oil-in water emulsion contains oil droplets of which at least 70% by intensity are less than 500 nm in diameter, more preferably at least 80% by intensity are less than 300 nm in diameter, more preferably at least 90% by intensity are in the range of 120 to 200 nm in diameter.

The oil droplet size, i.e. diameter, according to the present invention is given by intensity. There are several ways of determining the diameter of the oil droplet size by intensity. Intensity is measured by use of a sizing instrument, suitably by dynamic light scattering such as the Malvern Zetasizer 4000 or preferably the Malvern Zetasizer 3000HS. A detailed procedure is given in Example II.2. A first possibility is to determine the z average diameter ZAD by dynamic light scattering (PCS-Photon correlation spectroscopy); this method additionally give the polydispersity index (PDI), and both the ZAD and PDI are calculated with the cumulants algorithm. These values do not require the knowledge of the particle refractive index. A second mean is to calculate the diameter of the oil droplet by determining the whole particle size distribution by another algorithm, either the Contin, or NNLS, or the automatic "Malvern" one (the default algorithm provided for by the sizing instrument). Most of the time, as the particle refractive index of a complex composition is unknown, only the intensity distribution is taken into consideration, and if necessary the intensity mean originating from this distribution.

The oil in water emulsion comprises a sterol. Sterols are well known in the art, for example cholesterol is well known and is, for example, disclosed in the Merck Index, 11th Edn., page 341, as a naturally occurring sterol found in animal fat. Other suitable sterols

include β-sitosterol, stigmasterol, ergosterol, alpha-tocopherol and ergocalciferol. Said sterol is suitably present in an amount of 0.01% to 20% (w/v) of the total volume of the immunogenic composition, preferably at an amount of 0.1% to 5% (w/v). Preferably, when the sterol is cholesterol, it is present in an amount of between 0.02% and 0.2% (w/v) of the total volume of the immunogenic composition, more preferably at an amount of 0.02% (w/v) in a 0.5 ml vaccine dose volume, or 0.07% (w/v) in 0.5 ml vaccine dose volume or 0.1% (w/v) in 0.7 ml vaccine dose volume.

Suitably the sterol is alpha-tocopherol or a derivative thereof such as alpha-tocopherol succinate (also known as vitamin E succinate). Preferably alpha-tocopherol is present in an amount of between 0.2% and 5.0% (v/v) of the total volume of the immunogenic composition, more preferably at an amount of 2.5% (v/v) in a 0.5 ml vaccine dose volume, or 0.5% (v/v) in 0.5 ml vaccine dose volume or 1.7-1.9% (v/v), preferably 1.8% in 0.7 ml vaccine dose volume. By way of clarification, concentrations given in v/v can be converted into concentration in w/v by applying the following conversion factor: a 5% (v/v) alpha-tocopherol concentration is equivalent to a 4.8% (w/v) alpha-tocopherol concentration.

The oil in water emulsion further comprise an emulsifying agent. The emulsifying agent may be present at an amount of 0.01 to 5.0% by weight of the immunogenic composition (w/w), preferably present at an amount of 0.1 to 2.0% by weight (w/w). Preferred concentration are 0.5 to 1.5% by weight (w/w) of the total composition.

The emulsifying agent may suitably be polyoxyethylene sorbitan monooleate (Tween 80). In a specific embodiment, a 0.5 ml vaccine dose volume contains 1% (w/w) Tween 80, and a 0.7 ml vaccine dose volume contains 0.7% (w/w) Tween 80. In another specific embodiment the concentration of Tween 80 is 0.2% (w/w).

The oil in water emulsion adjuvant may be utilised with other adjuvants or immuno-stimulants and therefore an important embodiment of the invention is an oil in water formulation comprising squalene or another metabolisable oil, alpha tocopherol, and tween 80. The oil in water emulsion may also contain span 85 and/or Lecithin. Typically the oil in water will comprise from 2 to 10% squalene of the total volume of the immunogenic composition, from 2 to 10% alpha tocopherol and from 0.3 to 3% Tween 80, and may be produced according to the procedure described in WO 95/17210. Preferably the ratio of squalene: alpha tocopherol is equal or less than 1 as this provides a more

stable emulsion. Span 85 (polyoxyethylene sorbitan trioleate) may also be present, for example at a level of 1%.

3D-MPL

The composition comprise an additional adjuvant, 3 de-O-acylated monophosphoryl lipid A (3D-MPL). 3D MPL is a TRL-4 ligand adjuvant, a non-toxic derivative of lipid A.

3D-MPL is sold under the trademark MPL® by Corixa corporation and is referred throughout the document as MPL. It primarily promotes CD4+ T cell responses with an IFN-γ (Th1) phenotype. It can be produced according to the methods disclosed in GB 2 220 211 A. Chemically it is a mixture of 3-deacylated monophosphoryl lipid A with 3, 4, 5 or 6 acylated chains. Preferably in the compositions of the present invention small particle 3 D- MPL is used. Small particle 3D -MPL has a particle size such that it may be sterile-filtered through a 0.22 µm filter. Such preparations are described in WO 94/21292 and in Example II.

3D-MPL can be used, for example, at an amount of 1 to 100 µg (w/v) per composition dose, preferably in an amount of 10 to 50 µg (w/v) per composition dose. A suitable amount of 3D-MPL is for example any of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50 µg (w/v) per composition dose. More preferably, 3D-MPL amount ranges from 25 to 75 µg (w/v) per composition dose. Usually a composition dose will be ranging from about 0.5 ml to about 1 ml. A typical vaccine dose are 0.5 ml, 0.6 ml, 0.7 ml, 0.8 ml, 0.9 ml or 1 ml. In a preferred embodiment, a final concentration of 50 µg of 3D-MPL is contained per ml of vaccine composition, or 25 µg per 0.5 ml vaccine dose. In other preferred embodiments, a final concentration of 35.7 µg or 71.4 µg of 3D-MPL is contained per ml of vaccine composition. Specifically, a 0.5 ml vaccine dose volume contains 25 µg or 50 µg of 3D-MPL per dose.

The dose of MPL is suitably able to enhance an immune response to an antigen in a human. In particular a suitable MPL amount is that which improves the immunological potential of the composition compared to the unadjuvanted composition, or compared to the composition adjuvanted with another MPL amount, whilst being acceptable from a reactogenicity profile.

3D MPL is a TRL-4 ligand, a non-toxic derivative of lipid A. The present invention contemplates use of other suitable TLR-4 ligands in place of 3D-MPL, including lipopolysaccharide (LPS) and derivatives, MDP (muramyl dipeptide) and F protein of RSV. Non-toxic derivatives of lipid A, particularly monophosphoryl lipid A, are also contemplated.

Synthetic derivatives of lipid A are known, some being described as TLR-4 agonists, which might be suitable for use in the present invention and include, but are not limited to:

OM174 (2-deoxy-6-o-[2-deoxy-2-[(R)-3-dodecanoyloxytetra-decanoylamino]-4-o-phosphono-β-D-glucopyranosyl]-2-[(R)-3-hydroxytetradecanoylamino]-α-D-glucopyranosyldihydrogenphosphate), (WO 95/14026)

OM 294 DP (3S, 9 R) –3--[(R)-dodecanoyloxytetradecanoylamino]-4-oxo-5-aza-9(R)-[(R)-3-hydroxytetradecanoylamino]decan-1,10-diol,1,10-bis(dihydrogenophosphate) (WO99/64301 and WO 00/0462)

OM 197 MP-Ac DP (3S-, 9R) -3-[(R) -dodecanoyloxytetradecanoylamino]-4-oxo-5-aza-9-[(R)-3-hydroxytetradecanoylamino]decan-1,10-diol,1 -dihydrogenophosphate 10-(6-aminohexanoate) (WO 01/46127)

Immunogenic properties of the immunogenic composition used for the first vaccination of the present invention

The composition of the invention suitably induces an improved CD4 T-cell immune response against at least one of the component antigen(s) or antigenic composition compared to the CD4 T-cell immune response obtained with the corresponding composition which is un-adjuvanted, i.e. does not contain any exogeneous adjuvant (herein also referred to as 'plain composition') or at least lacks one of both components (either 3D-MPL or the oil-in-water emulsion adjuvant) of the adjuvant composition.

By 'improved CD4 T-cell immune response is meant that a higher CD4 response is obtained in a human patient after administration of the adjuvanted immunogenic composition than that obtained after administration of the same composition without adjuvant or lacking one of the two components of the adjuvant composition. For example, a higher CD4 T-cell response is obtained in a human patient upon administration of an

immunogenic composition comprising a split influenza virus or split virus antigenic preparation thereof together with an oil-in-water emulsion adjuvant as herein defined, compared to the response induced after administration of an immunogenic composition comprising a split influenza virus or split virus antigenic preparation thereof. Such formulation will advantageously be used to induce anti-influenza CD4-T cell response capable of detection of influenza epitopes presented by MHC class II molecules.

For influenza, preferably said immunological response induced by an adjuvanted split influenza composition of the present invention is higher than the immunological response induced by any other un-adjuvanted influenza conventional vaccine, such as sub-unit influenza vaccine or whole influenza virus vaccine.

In particular, but not exclusively, said 'improved CD4 T-cell immune response' is suitably obtained in an immunologically "unprimed" patient, i.e. a patient who is seronegative to the antigen. This seronegativity may be the result of said patient having never faced such an antigen (for example, not having been infected with a virus or bacteria containing said antigen - a so-called 'naive' patient) or, alternatively, having failed to respond to said antigen once encountered.

An improved CD4 T-cell immune response may be assessed by measuring the number of cells producing any of the following cytokines:

- cells producing at least two different cytokines (CD40L, IL-2, IFNγ, TNFα)
- cells producing at least CD40L and another cytokine (IL-2, TNFα, IFNγ)
- cells producing at least IL-2 and another cytokine (CD40L, TNFα, IFNγ)
- cells producing at least IFNγ and another cytokine (IL-2, TNFα, CD40L)
- cells producing at least TNFα and another cytokine (IL-2, CD40L, IFNγ)

There will be improved CD4 T-cell immune response when cells producing any of the above cytokines will be in a higher amount following administration of the adjuvanted composition compared to the administration of the un-adjuvanted composition. Typically at least one, preferably two of the five conditions mentioned herein above will be fulfilled. In a particular embodiment, the cells producing all four cytokines will be present at a higher amount in the adjuvanted group compared to the un-adjuvanted group.

An improved CD4 T-cell immune response conferred by an adjuvanted composition of the present invention may be ideally obtained after one single administration. The single dose approach will be extremely relevant for example in a rapidly evolving outbreak situation. In certain circumstances, especially for the elderly population, or in the case of young children (below 9 years of age) who are vaccinated for the first time against a disease eg influenza, it may be beneficial to administer two doses of the same composition for that season. The second dose of said same composition (still considered as 'composition for first vaccination') may be administered during the on-going primary immune response and is adequately spaced. Typically the second dose of the composition is given a few weeks, or about one month, e.g. 2 weeks, 3 weeks, 4 weeks, 5 weeks, or 6 weeks after the first dose, to help prime the immune system in unresponsive or poorly responsive individuals.

Thus, the present invention also relates to use of a composition comprising:

(a) an antigen, and

(f) an oil-in-water emulsion adjuvant; and

(g) 3D MPL

wherein said oil-in-water emulsion comprises a metabolisable oil, a sterol and an emulsifying agent, in the manufacture of an immunogenic composition for inducing an improved CD4 T-cell response against said antigen.

The invention also relates to a method of inducing an improved CD4 T-cell response against an antigen comprising delivery of a composition comprising:

(a) an antigen, and

(b) an oil-in-water emulsion adjuvant as herein defined; and

(c) 3D-MPL.

In a specific embodiment, the administration of said immunogenic composition alternatively or additionally induces an improved B-memory cell response in patients administered with the adjuvanted immunogenic composition compared to the B-memory cell response induced in individuals immunized with the un-adjuvanted composition or with a composition lacking one of the two components of the adjuvant composition. An improved B-memory cell response is intended to mean an increased frequency of peripheral blood B lymphocytes capable of differentiation into antibody-secreting plasma cells upon antigen encounter as measured by stimulation of in-vitro differentiation (see Example sections, e.g. methods of Elispot B cells memory).

In a still further specific embodiment, the vaccination with the composition for the first vaccination, adjuvanted, has no measurable impact on the CD8 response.

Preferably said improved CD4 T-cell immune response is obtained in an immunocompromised subject such as an elderly individual, typically at least 50, 55, 60 or 65 years of age or above, or an adult younger than 55 years of age with a high risk medical condition ('high risk' adult), or a child under the age of two.

The Applicants have surprisingly found that an oil-in-water emulsion adjuvant comprising a metabolisable oil, a sterol and an emulsifying agent, and 3D MPL is effective in promoting T cell responses in an immuno-compromised human population. As the Applicants have demonstrated for influenza, the administration of a single dose of the immunogenic composition for first vaccination, as described in the invention is capable of providing better sero-protection, as assessed by the correlates of protection for influenza vaccines, following re-vaccination against influenza in a human elderly population, than does the vaccination with an un-adjuvanted split vaccine. The claimed adjuvanted formulation has also been able to induce an improved CD4 T-cell immune response against influenza virus compared to that obtained with the un-adjuvanted formulation. This finding can be associated with an increased responsiveness upon vaccination or infection vis-à-vis influenza antigenic exposure. Furthermore, this may also be associated with a cross-responsiveness, i.e. a higher ability to respond against variant influenza strains. This improved response may be especially beneficial in an immuno-compromised human population such as the elderly population (eg 50, 55, 60, 65 years of age and above) and in particular the high risk elderly population. This may result in reducing the overall morbidity and mortality rate and preventing emergency admissions to hospital for pneumonia and other influenza-like illness. This may also be of benefit to the infant population (below 5 years, preferably below 2 years of age). Furthermore it allows to induce a CD4 T cell response which is more persistent in time, e.g. still present one year after the first vaccination, compared to the response induced with the un-adjuvanted formulation.

The Inventors have also been capable of demonstrating that the claimed adjuvanted composition was able to not only induce but also maintain protective levels of antibodies against all three strains present in the vaccine, in more individuals than those obtained with the un-advanted composition (see Table 43 for example).

Thus, in still another embodiment, the claimed composition is capable of ensuring a persistent immune response against influenza related disease. In particular, by persistence it is meant an HI antibody immune response which is capable of meeting regulatory criteria after at least three months, preferably after at least 6 months after the vaccination. In particular, the claimed composition is able to induce protective levels of antibodies in >70% of individuals, suitably in >80% of individuals or suitably in >90% of individuals for at least one influenza strain, preferably for all strains present in the vaccine, after at least three months. In a specific aspect, protective levels of antibodies of >90% are obtained at least 6 months post-vaccination against at least one, suitably two, or all strains present in the vaccine composition.

These findings with influenza are applicable to other diseases and antigens as it has also been demonstrated for HPV antigens (see Example XIII).

Thus, the present invention also relates to the use of the composition of the invention in an immunocompromised human individual or population such as high risk adults or elderly, and in the manufacture of an immunogenic composition for vaccination of a human immuno-compromised individual or population, such as a high risk adult or a elderly population.

We have determined that use of an antigen with an oil in water emulsion as herein defined can generate a cross reactive response against a variant antigen.

Preferably the CD4 T-cell immune response, such as the improved CD4 T-cell immune response obtained in an unprimed subject, involves the induction of a cross-reactive CD4 T helper response. In particular, the amount of cross-reactive CD4 T cells is increased. By 'cross-reactive' CD4 response is meant CD4 T-cell targeting shared epitopes between influenza strains.

For influenza, for example, available influenza vaccines are usually effective only against infecting strains of influenza virus that have haemagglutinins of similar antigenic characteristics. When the infecting (circulating) influenza virus has undergone minor changes (such as a point mutation or an accumulation of point mutations resulting in amino acid changes in the for example) in the surface glycoproteins in particular haemagglutinin (antigenic drift variant virus strain) the vaccine may still provide some protection, although it may only provide limited protection as the newly created variants

may escape immunity induced by prior influenza infection or vaccination. Antigenic drift is responsible for annual epidemics that occur during interpandemic periods (Wiley & Skehel, 1987, Ann. Rev. Biochem. 56, 365-394). The induction of cross-reactive CD4 T cells provides an additional advantage to the composition of the invention, in that it may provide also cross-protection, in other words protection against heterologous infections, i.e. infections caused by a circulating influenza strain which is a variant (e.g. a drift) of the influenza strain contained in the immunogenic composition. This may be advantageous when the circulating strain is difficult to propagate in eggs or to produce in tissue culture, rendering the use of a drifted strain a working alternative. This may also be advantageous when the subject received a first and a second vaccination several months or a year apart, and the influenza strain in the immunogenic composition used for a second immunization is a drift variant strain of the strain used in the composition used for the first vaccination.

Preclinical data given in Example 3 for example show the ability of the composition of the invention to protect against heterotypic influenza infection and disease as assessed by body temperature readouts. The same conclusion holds true for the clinical trials data obtained in revaccination studies.

In another aspect of the present invention, there is provided the use of the composition of the invention for protection against infections or disease caused by a pathogen which is a variant of the pathogen from which the antigen in the first composition is derived. Also provided issue of the composition of the invention for protection against infection or disease caused by a pathogen which comprises an antigen which is a variant of that in the composition of the invention.

Variant pathogens and/or antigens suitably have antigens with common CD4 T cell epitopes and/or B cell epitopes with the first pathogen or antigen, but which are not identical.

Detection of cross-reactive CD4 T-cells (eg following vaccination with Influenza vaccine)

Following classical trivalent Influenza vaccine administration (3 weeks), there is a substantial increase in the frequency of peripheral blood CD4 T-cells responding to antigenic strain preparation (whole virus or split antigen) that is homologous to the one present in the vaccine (H3N2: A/Panama/2007/99, H1N1: A/ New Caledonia/20/99, B:

B/Shangdong/7/97) (see Example III). A comparable increase in frequency can be seen if peripheral blood CD4 T-cells are restimulated with influenza strains classified as drifted strains (H3N2: A/Sydney/5/97, H1N1: A/Beijing/262/95, B: B/Yamanashi/166/98).

In contrast, if peripheral blood CD4 T-cells are restimulated with influenza strains classified as shift strains (H2N2: A/Singapore/1/57, H9N2: A/Hongkong/1073/99) by expert in the field, there is no observable increase following vaccination.

CD4 T-cells that are able to recognize both homologous and drifted Influenza strains have been named in the present document "cross-reactive".

CD4 T-cell epitopes shared by different Influenza strains have been identified in human (Gelder C et al. 1998, Int Immunol. 10(2):211-22; Gelder CM et al. 1996 J Virol. 70(7):4787-90; Gelder CM et al. 1995 J Virol. 1995 69(12):7497-506).

In a specific embodiment, the adjuvanted composition may offer the additional benefit of providing better protection against circulating strains which have undergone a major change (such as gene recombination for example, between two different species) in the haemagglutinin (antigenic shift) against which currently available vaccines have no efficacy.

Other Adjuvants

The composition may comprise additional adjuvants, suitably such as TRL-4 ligand adjuvants or a non-toxic derivative of lipid A. Suitable TLR-4 ligands are lipopolysaccharide (LPS) and derivatives, MDP (muramyl dipeptide) and F protein of RSV.

Synthetic derivatives of lipid A are known, some being described as TLR-4 agonists, and include, but are not limited to:

OM174 (2-deoxy-6-o-[2-deoxy-2-[(R)-3-dodecanoyloxytetra-decanoylamino]-4-o-phosphono-β-D-glucopyranosyl]-2-[(R)-3-hydroxytetradecanoylamino]-α-D-glucopyranosyldihydrogenphosphate), (WO 95/14026)

OM 294 DP (3S, 9 R) –3--[(R)-dodecanoyloxytetradecanoylamino]-4-oxo-5-aza-9(R)-[(R)-3-hydroxytetradecanoylamino]decan-1,10-diol,1,10-bis(dihydrogenophosphate) (WO99/64301 and WO 00/0462)

OM 197 MP-Ac DP (3S-, 9R) -3-[(R) -dodecanoyloxytetradecanoylamino]-4-oxo-5-aza-9-[(R)-3-hydroxytetradecanoylamino]decan-1,10-diol,1 -dihydrogenophosphate 10-(6-aminohexanoate) (WO 01/46127)

Other suitable TLR-4 ligands are, for example, lipopolysaccharide and its derivatives, muramyl dipeptide (MDP) or F protein of respiratory syncitial virus.

Another suitable immunostimulant for use in the present invention is Quil A and its derivatives. Quil A is a saponin preparation isolated from the South American tree *Quilaja Saponaria Molina* and was first described by Dalsgaard *et al.* in 1974 ("Saponin adjuvants", Archiv. für die gesamte Virusforschung, Vol. 44, Springer Verlag, Berlin, p243-254) to have adjuvant activity. Purified fragments of Quil A have been isolated by HPLC which retain adjuvant activity without the toxicity associated with Quil A (EP 0 362 278), for example QS7 and QS21 (also known as QA7 and QA21). QS-21 is a natural saponin derived from the bark of *Quillaja saponaria* Molina, which induces CD8+ cytotoxic T cells (CTLs), Th1 cells and a predominant IgG2a antibody response and is a preferred saponin in the context of the present invention.

Particular formulations of QS21 have been described which are particularly preferred, these formulations further comprise a sterol (WO96/33739). The saponins forming part of the present invention may be in the form of an oil in water emulsion (WO 95/17210).

Revaccination

An aspect of the present invention provides the use of an antigen in the manufacture of an immunogenic composition for revaccination of humans previously vaccinated with the antigen or fragment or variant thereof with 3D MPL and an oil-in-water emulsion adjuvant as herein defined.

Typically revaccination is made at least 6 months after the first vaccination(s), preferably 8 to 14 months after, more preferably at around 10 to 12 months after.

Preferably, there is provided the use of:
(1) an antigen and
(2) an oil-in-water emulsion adjuvant
in the manufacture of an immunogenic composition for revaccination of humans previously vaccinated with the antigen, or fragment or variant thereof, 3D MPL and an oil-in-water emulsion adjuvant as herein defined.

Preferably, there is provided the use of:
(1) an antigen and
(2) an oil-in-water emulsion adjuvant, and
(3) 3D MPL
in the manufacture of an immunogenic composition for revaccination of humans previously vaccinated with the antigen, or fragment or variant thereof, 3D MPL and an oil-in-water emulsion adjuvant as herein defined.

In a preferred embodiment, the invention provides for the use of:
(a) an antigen, and
(b) an oil-in-water emulsion adjuvant, and
(c) 3D MPL
in the manufacture of an immunogenic composition for revaccination of humans previously vaccinated with said antigen or fragment or variant thereof.

In a specific embodiment, the composition for revaccination suitably shares common CD4 T-cell epitopes with the composition used for the first vaccination. In this respect it is a considered a variant of the antigen used in first vaccination.

The immunogenic composition for re-vaccination (the boosting composition) may contain the same type of antigen preparation – eg subunit/split/whole inactivated virus - as the immonogenic composition used for the first vaccination. Alternatively the boosting composition may contain another type of antigen preparation, For example, for influenza the first vaccination may be with a split preparation and the booster vaccination with another inactivated influenza antigen, such as inactivated whole virus or purified HA and NA (subunit vaccine). The booster composition may be adjuvanted or un-adjuvanted.

For influenza the un-adjuvanted booster composition may be Fluarix™/α-Rix® given intramuscularly. The formulation contains three inactivated split virion antigens prepared from the WHO recommended strains of the appropriate influenza season.

A variant may be an antigen which shares common CD4 T-cell epitopes (generally considered as antigenic determinants recognized and bound by the T-cell receptor) with the antigenic composition used for the first vaccination, but which is not identical to that antigenic composition.

A variant may be an antigen which shares common B-cell epitopes (generally considered as antigenic determinants recognized and bound by the B-cell receptor) with the antigenic composition used for the first vaccination, but which is not identical to that antigenic composition.

T cell and B cell epitopes may be predicted using techniques well known in the art or inferred from immune responses using techniques as described herein.

Said oil-in-water emulsion adjuvant preferably comprises at least one metabolisable oil in an amount of 0.5% to 20% of the total volume, and has oil droplets of which at least 70% by intensity have diameters of less than 1 μm.

In a specific embodiment for influenza, the immunogenic composition for revaccination (also called herein below the 'boosting composition') contains a split influenza virus or split virus antigenic preparation thereof which shares a common CD4 T-cell epitope with the split influenza virus or split virus antigenic preparation thereof used for the first vaccination.

Generally, for all antigens, a 'common CD4 T cell epitope is intended to mean peptides/sequences/epitopes from different antigens which can be recognised by the same CD4 cell (see examples of described epitopes in: Gelder C et al. 1998, Int Immunol. 10(2):211-22; Gelder CM et al. 1996 J Virol. 70(7):4787-90; Gelder CM et al. 1995 J Virol. 1995 69(12):7497-506).

In the context of influenza, a preferred embodiment, the influenza strain may be associated with a pandemic outbreak or have the potential to be associated with a pandemic outbreak. In particular, when the vaccine is a multivalent vaccine such as a

bivalent or a trivalent vaccine, at least one strain is associated with a pandemic outbreak or has the potential to be associated with a pandemic outbreak. Suitable strains are, but not limited to: H5N1, H9N2, H7N7, H2N2 and H1N1.

Typically a booster composition, where used, is given at the next season, e.g. approximately one year after the first immunogenic composition. The booster composition may also be given every subsequent year (third, fourth, fifth vaccination and so forth). The boosting composition may be the same as the first composition. Suitably, the boosting composition contains an strain or antigenic preparation therefrom which is a variant of the strain or antigen used for the first vaccination.

The influenza antigen or antigenic composition used in revaccination preferably comprises an adjuvant or an oil-in-water emulsion, suitably as described above. The adjuvant may be an oil-in-water emulsion adjuvant as herein above described, which is preferred, optionally containing an additional adjuvant such as TLR-4 ligand such as 3D-MPL or a saponin, or may be another suitable adjuvant such as alum for example.

For all antigens of the invention re-vaccination suitably induces any, preferably two or all, of the following: (i) an improved CD4 response against the influenza virus or antigenic preparation thereof, or (ii) an improved B cell memory response or (iii) an improved humoral response, compared to the equivalent response induced after a first vaccination with the un-adjuvanted split influenza virus or split virus antigenic preparation thereof. Preferably the immunological responses induced after re-vaccination with the adjuvanted split influenza virus or split virus antigenic preparation thereof as herein defined, are higher than the corresponding response induced after the re-vaccination with the un-adjuvanted composition. Preferably the immunological responses induced after re-vaccination with an un-adjuvanted, preferably split, influenza virus are higher in the population first vaccinated with the adjuvanted split influenza composition than the corresponding response in the population first vaccinated with the un-adjuvanted split influenza composition.

In another aspect of the present invention, there is provided the use of:

(a) an antigen from a first viral or bacterial strain and

(b) an oil-in-water emulsion adjuvant

(c) 3D MPL

in the manufacture of an immunogenic composition for protection against infections or disease caused by a viral or bacterial strain which is a variant of said first strain.

For example, for influenza, the adjuvanted composition of the invention is capable of inducing a better protection against drifted strain (the influenza strain from the next influenza season) compared to the protection conferred by the control vaccine.

Influenza viral strains and antigens thereof

For compositions comprising a split influenza virus or split virus antigenic preparation thereof the composition is suitably monovalent or multivalent such as bivalent or trivalent or quadrivalent. Preferably the split influenza virus or split virus antigenic preparation thereof is trivalent or quadrivalent, having an antigen from three different influenza strains.

Optionally at least one strain is associated with a pandemic outbreak or has the potential to be associated with a pandemic outbreak.

By way of background, during inter-pandemic periods, influenza viruses circulate that are related to those from the preceding epidemic. The viruses spread among people with varying levels of immunity from infections earlier in life. Such circulation, over a period of usually 2-3 years, promotes the selection of new strains that have changed enough to cause an epidemic again among the general population; this process is termed 'antigenic drift'. 'Drift variants' may have different impacts in different communities, regions, countries or continents in any one year, although over several years their overall impact is often similar. In other words, an influenza pandemics occurs when a new influenza virus appears against which the human population has no immunity. Typical influenza epidemics cause increases in incidence of pneumonia and lower respiratory disease as witnessed by increased rates of hospitalisation or mortality. The elderly or those with underlying chronic diseases are most likely to experience such complications, but young infants also may suffer severe disease.

At unpredictable intervals, novel influenza viruses emerge with a key surface antigen, the haemagglutinin, of a totally different subtype from strains circulating the season before. Here, the resulting antigens can vary from 20% to 50% from the corresponding protein of strains that were previously circulating in humans. This can result in virus escaping 'herd immunity' and establishing pandemics. This phenomenon is called 'antigenic shift'. It is

thought that at least in the past pandemics have occurred when an influenza virus from a different species, such as an avian or a porcine influenza virus, has crossed the species barrier. If such viruses have the potential to spread from person to person, they may spread worldwide within a few months to a year, resulting in a pandemic. For example, in 1957 (Asian Flu pandemic), viruses of the H2N2 subtype replaced H1N1 viruses that had been circulating in the human population since at least 1918 when the virus was first isolated. The H2 HA and N2 NA underwent antigenic drift between 1957 and 1968 until the HA was replaced in 1968 (Hong-Kong Flu pandemic) by the emergence of the H3N2 influenza subtype, after which the N2 NA continued to drift along with the H3 HA (Nakajima et al., 1991, Epidemiol. Infect. 106, 383-395).

The features of an influenza virus strain that give it the potential to cause a pandemic outbreak are: it contains a new haemagglutinin compared to the haemagglutinin in the currently circulating strains, which may or not be accompanied by a change in neuraminidase subtype; it is capable of being transmitted horizontally in the human population; and it is pathogenic for humans. A new haemagglutinin may be one which has not been evident in the human population for an extended period of time, probably a number of decades, such as H2. Or it may be a haemagglutinin that has not been circulating in the human population before, for example H5, H9, H7 or H6 which are found in birds. In either case the majority, or at least a large proportion of, or even the entire population has not previously encountered the antigen and is immunologically naïve to it.

Certain parties are generally at an increased risk of becoming infected with influenza in a pandemic situation. The elderly, the chronically ill and small children are particularly susceptible but many young and apparently healthy people are also at risk. For H2 influenza, the part of the population born after 1968 is at an increased risk. It is important for these groups to be protected effectively as soon as possible and in a simple way.

Another group of people who are at increased risk are travelers. People travel more today than ever before and the regions where most new viruses emerge, China and South East Asia, have become popular travel destinations in recent years. This change in travel patterns enables new viruses to reach around the globe in a matter of weeks rather than months or years.

Thus for these groups of people there is a particular need for vaccination to protect against influenza in a pandemic situation or a potential pandemic situation. Suitable strains are, but not limited to: H5N1, H9N2, H7N7, H2N2 and H1N1.

Optionally the composition may contain more than three valencies, for example two non pandemic strains plus a pandemic strain. Alternatively the composition may contain three pandemic strains.

In a further embodiment the invention relates to a vaccination regime in which the first vaccination is made with a split influenza composition containing at least one influenza strain that could potentially cause a pandemic outbreak and the re-vaccination is made with a circulating strain, either a pandemic strain or a classical strain.

CD4 epitope in HA

This antigenic drift mainly resides in epitope regions of the viral surface proteins haemagglutinin (HA) and neuraminidase (NA). It is known that any difference in CD4 and B cell epitopes between different influenza strains, being used by the virus to evade the adaptive response of the host immune system, will play a major role in influenza vaccination and is.

CD4 T-cell epitopes shared by different Influenza strains have been identified in human (see for example: Gelder C et al. 1998, Int Immunol. 10(2):211-22; Gelder CM et al. 1996 J Virol. 70(7):4787-90; and Gelder CM et al. 1995 J Virol. 1995 69(12):7497-506).

In a specific embodiment, the re-vaccination is made by using a booster composition which contain an influenza virus or antigenic preparation thereof which shares common CD4 T-cell epitopes with the split influenza virus antigen or split virus antigenic preparation thereof used for the first vaccination.

The invention thus relates to the use of the immunogenic composition comprising a split influenza virus or split virus antigenic preparation thereof, an oil-in-water emulsion adjuvant as herein defined and 3D MPL in the manufacture of a first vaccination-component of a multi-dose vaccine, the multi-dose vaccine further comprising, as a boosting dose, an influenza virus or antigenic preparation thereof which shares common

CD4 T-cell epitopes with the split influenza virus antigen or split virus antigenic preparation thereof of the dose given at the first vaccination.

Vaccination

The composition of the invention may be administered by any suitable delivery route, such as intradermal, mucosal e.g. intranasal, oral, intramuscular or subcutaneous. Other delivery routes are well known in the art.

The intramuscular delivery route is preferred for the adjuvanted influenza composition.

Intradermal delivery is another suitable route. Any suitable device may be used for intradermal delivery, for example short needle devices such as those described in US 4,886,499, US5,190,521, US 5,328,483, US 5,527,288, US 4,270,537, US 5,015,235, US 5,141,496, US 5,417,662. Intradermal vaccines may also be administered by devices which limit the effective penetration length of a needle into the skin, such as those described in WO99/34850 and EP1092444, incorporated herein by reference, and functional equivalents thereof. Also suitable are jet injection devices which deliver liquid vaccines to the dermis via a liquid jet injector or via a needle which pierces the stratum corneum and produces a jet which reaches the dermis. Jet injection devices are described for example in US 5,480,381, US 5,599,302, US 5,334,144, US 5,993,412, US 5,649,912, US 5,569,189, US 5,704,911, US 5,383,851, US 5,893,397, US 5,466,220, US 5,339,163, US 5,312,335, US 5,503,627, US 5,064,413, US 5,520, 639, US 4,596,556US 4,790,824, US 4,941,880, US 4,940,460, WO 97/37705 and WO 97/13537. Also suitable are ballistic powder/particle delivery devices which use compressed gas to accelerate vaccine in powder form through the outer layers of the skin to the dermis. Additionally, conventional syringes may be used in the classical mantoux method of intradermal administration.

Another suitable administration route is the subcutaneous route. Any suitable device may be used for subcutaneous delivery, for example classical needle. Preferably, a needle-free jet injector service is used, such as that published in WO 01/05453, WO 01/05452, WO 01/05451, WO 01/32243, WO 01/41840, WO 01/41839, WO 01/47585, WO 01/56637, WO 01/58512, WO 01/64269, WO 01/78810, WO 01/91835, WO 01/97884, WO 02/09796, WO 02/34317. More preferably said device is pre-filled with the liquid vaccine formulation.

Alternatively the vaccine is administered intranasally. Typically, the vaccine is administered locally to the nasopharyngeal area, preferably without being inhaled into the lungs. It is desirable to use an intranasal delivery device which delivers the vaccine formulation to the nasopharyngeal area, without or substantially without it entering the lungs.

Preferred devices for intranasal administration of the vaccines according to the invention are spray devices. Suitable commercially available nasal spray devices include Accuspray™ (Becton Dickinson). Nebulisers produce a very fine spray which can be easily inhaled into the lungs and therefore does not efficiently reach the nasal mucosa. Nebulisers are therefore not preferred.

Preferred spray devices for intranasal use are devices for which the performance of the device is not dependent upon the pressure applied by the user. These devices are known as pressure threshold devices. Liquid is released from the nozzle only when a threshold pressure is applied. These devices make it easier to achieve a spray with a regular droplet size. Pressure threshold devices suitable for use with the present invention are known in the art and are described for example in WO 91/13281 and EP 311 863 B and EP 516 636, incorporated herein by reference. Such devices are commercially available from Pfeiffer GmbH and are also described in Bommer, R. Pharmaceutical Technology Europe, Sept 1999.

Preferred intranasal devices produce droplets (measured using water as the liquid) in the range 1 to 200 μm, preferably 10 to 120 μm. Below 10 μm there is a risk of inhalation, therefore it is desirable to have no more than about 5% of droplets below 10 μm. Droplets above 120 μm do not spread as well as smaller droplets, so it is desirable to have no more than about 5% of droplets exceeding 120 μm.

Bi-dose delivery is a further preferred feature of an intranasal delivery system for use with the vaccines according to the invention. Bi-dose devices contain two sub-doses of a single vaccine dose, one sub-dose for administration to each nostril. Generally, the two sub-doses are present in a single chamber and the construction of the device allows the efficient delivery of a single sub-dose at a time. Alternatively, a monodose device may be used for administering the vaccines according to the invention.

Alternatively, the epidermal or transdermal vaccination route is also contempletd in the present invention.

In a specific aspect of the present invention, the adjuvanted immunogenic composition for the first administration may be given intramuscularly, and the booster composition, either adjuvanted or not, may be administered through a different route, for example intradermal, subcutaneous or intranasal. In another specific embodiment, the composition for the first administration may contain a standard HA content of 15 µg per influenza strain, and the booster composition may contain a low dose of HA, i.e. below 15 µg, and depending on the administration route, may be given in a smaller volume.

Populations to vaccinate

The target population is preferably a human population or individual.

The target population to vaccinate may be immuno-compromised human. Immuno-compromised humans generally are less well able to respond to an antigen, in particular to an influenza antigen, in comparison to healthy adults.

Preferably the target population is a population which is unprimed against influenza, either being naïve (such as vis à vis a pandemic strain), or having failed to respond previously to infection or vaccination. Preferably the target population is elderly persons suitably aged 55 years and over, younger high-risk adults (i.e. between 18 and 54 years of age) such as people working in health institutions, or those young adults with a risk factor such as cardiovascular and pulmonary disease, or diabetes. Another target population is all children 6 months of age and over, especially children 6-23 months of age who experience a relatively high influenza-related hospitalization rate. Preferably the target population is elderly above 65 years of age.

Vaccination regimes, dosing and additional efficacy criteria

The amount of saccharide (or conjugate thereof) antigen in each vaccine dose is selected as an amount which induces an immunoprotective response without significant, adverse side effects in typical vaccines. Such amount will vary depending upon which specific immunogen is employed and how it is presented. Generally, it is expected that

each dose will comprise 0.1-100 μg of saccharide, preferably 0.1-50 μg, preferably 0.1-10 μg, of which 1 to 5 μg is the most preferable range.

The content of protein antigens in the vaccine will typically be in the range 1-100μg, preferably 5-50μg, most typically in the range 5 - 25μg.

Optimal amounts of components for a particular vaccine can be ascertained by standard studies involving observation of appropriate immune responses in subjects. Following an initial vaccination, subjects may receive one or several booster immunisations adequately spaced.

Vaccine preparation is generally described in Vaccine Design ("The subunit and adjuvant approach" (eds Powell M.F. & Newman M.J.) (1995) Plenum Press New York). Encapsulation within liposomes is described by Fullerton, US Patent 4,235,877.

Influenza antigen dose

For influenza, suitably the immunogenic compositions according to the present invention are a standard 0.5 ml injectable dose in most cases, and contains 15 μg of haemagglutinin antigen component from the or each influenza strain, as measured by single radial immunodiffusion (SRD) (J.M. Wood et al.: J. Biol. Stand. 5 (1977) 237-247; J. M. Wood et al., J. Biol. Stand. 9 (1981) 317-330). Suitably the vaccine dose volume will be between 0.5 ml and 1 ml, in particular a standard 0.5 ml, or 0.7 ml vaccine dose volume. Slight adaptation of the dose volume will be made routinely depending on the HA concentration in the original bulk sample.

Suitably said immunogenic composition contains a low dose of HA antigen – e.g any of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13 or 14 μg of HA per influenza strain. A suitable low dose of HA is between 1 to 7.5 μg of HA per influenza strain, suitably between 3.5 to 5 μg such as 3.75 μg of HA per influenza strain, typically about 5 μg of HA per influenza strain.

Advantageously, a vaccine dose according to the invention, in particular a low dose vaccine, may be provided in a smaller volume than the conventional injected split flu vaccines, which are generally around 0.5, 0.7 or 1 ml per dose. The low volume doses according to the invention are preferably below 500 μl, more preferably below 300 μl and most preferably not more than about 200 μl or less per dose.

Thus, a preferred low volume vaccine dose according to one aspect of the invention is a dose with a low antigen dose in a low volume, e.g. about 15 μg or about 7.5 μg HA or about 3.0 μg HA (per strain) in a volume of about 200 μl.

The influenza medicament of the invention preferably meets certain international criteria for vaccines.

Standards are applied internationally to measure the efficacy of influenza vaccines. The European Union official criteria for an effective vaccine against influenza are set out in the Table 1 below. Theoretically, to meet the European Union requirements, an influenza vaccine has to meet only one of the criteria in the table, for all strains of influenza included in the vaccine. The compositions of the present invention suitably meet at least one such criteria.

However in practice, at least two or all three of the criteria will need to be met for all strains, particularly for a new vaccine such as a new vaccine for delivery via a different route. Under some circumstances two criteria may be sufficient. For example, it may be acceptable for two of the three criteria to be met by all strains while the third criterion is met by some but not all strains (e.g. two out of three strains). The requirements are different for adult populations (18-60 years) and elderly populations (>60 years).

Table 1

	18 - 60 years	> 60 years
Seroconversion rate*	>40%	>30%
Conversion factor**	>2.5	>2.0
Protection rate***	>70%	>60%

* Seroconversion rate is defined as the percentage of vaccinees who have at least a 4-fold increase in serum haemagglutinin inhibition (HI) titres after vaccination, for each vaccine strain.

** Conversion factor is defined as the fold increase in serum HI geometric mean titres (GMTs) after vaccination, for each vaccine strain.

*** Protection rate is defined as the percentage of vaccinees with a serum HI titre equal to or greater than 1:40 after vaccination (for each vaccine strain) and is normally accepted as indicating protection.

In a further aspect the invention provides a method of designing a vaccine for diseases known to be cured or treated through a CD4+ T cell activation, comprising

1) selecting an antigen containing CD4+ epitopes, and

2) combining said antigen with an oil-in-water emulsion adjuvant as defined herein above with 3D MPL , wherein said vaccine upon administration in said mammal is capable of inducing an enhanced CD4 T cell response in said mammal.

The teaching of all references in the present application, including patent applications and granted patents, are herein fully incorporated by reference.

For the avoidance of doubt the terms 'comprising', 'comprise' and 'comprises' herein is intended by the inventors to be optionally substitutable with the terms 'consisting of', 'consist of', and 'consists of', respectively, in every instance.

The invention will be further described by reference to the following, non-limiting, examples:

Example I describes immunological read-out methods used in mice, ferret and human studies.

Example II describes the preparation and characterization of the oil in water emulsion and adjuvant formulations used in the studies exemplified.

Example III describes a clinical trial in an elderly population aged over 65 years with a vaccine containing a split influenza antigen preparation and AS03 adjuvant

Example IV describes a second clinical trial – revaccination trial - in an elderly population aged over 65 years with a vaccine containing a split influenza antigen preparation and AS03 adjuvant.

Example V shows a pre-clinical evaluation of adjuvanted and un-adjuvanted influenza vaccines in ferrets (study I and study II). The temperature monitoring, viral shedding and CD4 T-cell response were measured.

Example VI shows a pre-clinical evaluation of adjuvanted and un-adjuvanted influenza vaccines in C57Bl/6 naïve and primed mice.

Example VII shows a pre-clinical evaluation of adjuvanted and un-adjuvanted split and sub-unit influenza vaccines in C57Bl/6 mice primed with heterologous strains.

Example VIII describes a clinical trial in an elderly population aged over 65 years with a vaccine containing a split influenza antigen preparation containing AS03 adjuvant, AS03+MPL adjuvant, or no exogeneous adjuvant.

Example IX shows a pre-clinical evaluation of adjuvanted and un-adjuvanted influenza vaccines in ferrets (study III). The temperature monitoring, viral shedding and HI titers were measured.

Example X shows a clinical trial in an elderly population aged over 65 years with a vaccine containing a split influenza antigen preparation containing AS03 with or without MPL adjuvant: immunogenicity persistence data at day 90 and day 180.

Example XI shows a clinical trial in an elderly population aged over 65 years with a vaccine containing a split influenza antigen preparation containing AS03 with MPL adjuvant.

Example XII shows a clinical trial in an elderly population aged over 65 years with a vaccine containing a split influenza antigen preparation containing AS03 with MPL adjuvant at two concentrations.

Example XIII shows a pre-clinical evaluation of two adjuvanted HPV vaccines in mice. Antibody and B cell memory responses were measured.

Example I – Immunological Read-out Methods

I.1. Mice methods

I.1.1. Hemagglutination Inhibition Test

Test procedure

Anti-Hemagglutinin antibody titers to the three influenza virus strains were determined using the hemagglutination inhibition test (HI). The principle of the HI test is based on the ability of specific anti-Influenza antibodies to inhibit hemagglutination of chicken red blood cells (RBC) by influenza virus hemagglutinin (HA). Heat inactivated sera were previously treated by Kaolin and chicken RBC to remove non-specific inhibitors. After pretreatment, two-fold dilutions of sera were incubated with 4 hemagglutination units of each influenza strain. Chicken red blood cells were then added and the inhibition of agglutination was scored. The titers were expressed as the reciprocal of the highest dilution of serum that completely inhibited hemagglutination. As the first dilution of sera was 1:20, an undetectable level was scored as a titer equal to 10.

Statistical analysis

Statistical analysis were performed on post vaccination HI titers using UNISTAT .The protocol applied for analysis of variance can be briefly described as follow:

- Log transformation of data
- Shapiro-Wilk test on each population (group) in order to verify the normality of groups distribution
- Cochran test in order to verify the homogenicity of variance between the different populations (groups)
- Two-way Analysis of variance performed on groups
- Tukey HSD test for multiple comparisons

I.1.2. Intracellular cytokine staining

This technique allows a quantification of antigen specific T lymphocytes on the basis of cytokine production: effector T cells and/or effector-memory T cells produce IFN-γ and/or central memory T cells produce IL-2. PBMCs are harvested at day 7 post-immunization.

Lymphoid cells are re-stimulated in vitro in the presence of secretion inhibitor (Brefeldine). These cells are then processed by conventional immunofluorescent procedure using fluorescent antibodies (CD4, CD8, IFN-γ and IL-2). Results are expressed as a frequency of cytokine positive cell within CD4/CD8 T cells. Intracellular staining of cytokines of T cells was performed on PBMC 7 days after the second immunization. Blood was collected from mice and pooled in heparinated medium RPMI+ Add. For blood, RPMI + Add-diluted PBL suspensions were layered onto a Lympholyte–Mammal gradient according to the recommended protocol (centrifuge 20 min at 2500 rpm and R.T.). The mononuclear cells at the interface were removed, washed 2x in RPMI + Add and PBMCs suspensions were adjusted to 2 x 10^6 cells/ml in RPMI 5% fetal calf serum.

In vitro antigen stimulation of PBMCs was carried out at a final concentration of 1 x 10^7 cells/ml (tube FACS) with Whole FI (1µgHA/strain) and then incubated 2 hrs at 37°C with the addition of anti-CD28 and anti-CD49d (1 µg/ml for both).

Following the antigen restimulation step, PBMC are incubated overnight at 37°C in presence of Brefeldin (1µg/ml) at 37°C to inhibit cytokine secretion.

IFN-γ /IL-2/CD4/CD8 staining was performed as follows: Cell suspensions were washed, resuspended in 50µl of PBS 1% FCS containing 2% Fc blocking reagent (1/50; 2.4G2). After 10 min incubation at 4°C, 50 µl of a mixture of anti-CD4-PE (2/50) and anti-CD8 perCp (3/50) was added and incubated 30 min at 4°C. After a washing in PBS 1% FCS, cells were permeabilized by resuspending in 200 µl of Cytofix-Cytoperm (Kit BD) and incubated 20 min at 4°C. Cells were then washed with Perm Wash (Kit BD) and resuspended with 50 µl of a mix of anti- IFN-γ APC (1/50) + anti-IL-2 FITC (1/50) diluted in Perm Wash. After an incubation min 2 h max overnight at 4°C, cells were washed with Perm Wash and resuspended in PBS 1% FCS + 1% paraformaldéhyde. Sample analysis was performed by FACS. Live cells were gated (FSC/SSC) and acquisition was performed on ~ 20,000 events (lymphocytes) or 35,000 events on CD4+T cells. The percentages of IFN-γ + or IL2+ were calculated on CD4+ and CD8+ gated populations.

I.2. Ferrets methods

I.2.1. Hemagglutination Inhibition Test (HI)

Test procedure.

Anti-Hemagglutinin antibody titers to the three influenza virus strains were determined using the hemagglutination inhibition test (HI). The principle of the HI test is based on the ability of specific anti-Influenza antibodies to inhibit hemagglutination of chicken red blood cells (RBC) by influenza virus hemagglutinin (HA). Sera were first treated with a 25% neuraminidase solution (RDE) and were heat-inactivated to remove non-specific inhibitors. After pre-treatment, two-fold dilutions of sera were incubated with 4 hemagglutination units of each influenza strain. Chicken red blood cells were then added and the inhibition of agglutination was scored. The titers were expressed as the reciprocal of the highest dilution of serum that completely inhibited hemagglutination. As the first dilution of sera was 1:10, an undetectable level was scored as a titer equal to 5.

Statistical analysis.

Statistical analysis were performed on HI titers (Day 41, before challenge) using UNISTAT. The protocol applied for analysis of variance can be briefly described as followed:

- Log transformation of data.
- Shapiro-wilk test on each population (group) in ordetr to verify the normality of groups distribution.

- Cochran test in order to verify the homogenicity of variance between the different populations (groups).
- Test for interaction of one-way ANOVA.
- Tuckey-HSD Test for multiple comparisons.

I.2.2. Body temperature monitoring

Individual temperatures were monitored during the challenge period with the transmitters and by the telemetry recording. All implants were checked and refurbished and a new calibration was performed by DSI (Data Sciences International, Centaurusweg 123, 5015 TC Tilburg, The Netherlands) before placement in the intraperitoneal cavity. All animals were individually housed in single cage during these measurements.
Temperatures were recorded every 15 minutes 4 days before challenge until 7 days Post-challenge.

I.2.3. Nasal washes

The nasal washes were performed by administration of 5 ml of PBS in both nostrils in awoke animals. The inoculum was collected in a Petri dish and placed into sample containers on dry ice.

Viral titration in nasal washes

All nasal samples were first sterile filtered through Spin X filters (Costar) to remove any bacterial contamination. 50 µl of serial ten-fold dilutions of nasal washes were transferred to microtiter plates containing 50 µl of medium (10 wells/dilution). 100µl of MDCK cells (2.4×10^5 cells/ml) were then added to each well and incubated at 35°C for 5-7days.

After 5-7 days of incubation, the culture medium is gently removed and 100 µl of a 1/20 WST-1 containing medium is added and incubated for another 18 hrs.

The intensity of the yellow formazan dye produced upon reduction of WST-1 by viable cells is proportional to the number of viable cells present in the well at the end of the viral titration assay and is quantified by measuring the absorbance of each well at the appropriate wavelength (450 nanometers). The cut-off is defined as the OD average of uninfected control cells – 0.3 OD (0.3 OD correspond to +/- 3 StDev of OD of uninfected control cells). A positive score is defined when OD is < cut-off and in contrast a negative score is defined when OD is > cut-off. Viral shedding titers were determined by "Reed and Muench" and expressed as Log TCID50/ml.

I.3. Assays for assessing the immune response in humans

I.3.1. Hemagglutination Inhibition Assay

The immune response was determined by measuring HI antibodies using the method described by the WHO Collaborating Centre for influenza, Centres for Disease Control, Atlanta, USA (1991).

Antibody titre measurements were conducted on thawed frozen serum samples with a standardised and comprehensively validated micromethod using 4 hemagglutination-inhibiting units (4 HIU) of the appropriate antigens and a 0.5% fowl erythrocyte suspension. Non-specific serum inhibitors were removed by heat treatment and receptor-destroying enzyme.

The sera obtained were evaluated for HI antibody levels. Starting with an initial dilution of 1:10, a dilution series (by a factor of 2) was prepared up to an end dilution of 1:20480. The titration end-point was taken as the highest dilution step that showed complete inhibition (100%) of hemagglutination. All assays were performed in duplicate.

I.3.2. Neuraminidase Inhibition Assay

The assay was performed in fetuin-coated microtitre plates. A 2-fold dilution series of the antiserum was prepared and mixed with a standardised amount of influenza A H3N2, H1N1 or influenza B virus. The test was based on the biological activity of the neuraminidase which enzymatically releases neuraminic acid from fetuin. After cleavage of the terminal neuraminic acid ß-D-glactose-N-acetyl-galactosamin was unmasked. Horseradish peroxidase (HRP)-labelled peanut agglutinin from Arachis hypogaea, which binds specifically to the galactose structures, was added to the wells. The amount of bound agglutinin can be detected and quantified in a substrate reaction with tetra-methylbenzidine (TMB) The highest antibody dilution that still inhibits the viral neuraminidase activity by at least 50% was indicated is the NI titre.

I.3.3. Neutralising Antibody Assay

Neutralising antibody measurements were conducted on thawed frozen serum samples. Virus neutralisation by antibodies contained in the serum was determined in a microneutralization assay. The sera were used without further treatment in the assay.

Each serum was tested in triplicate. A standardised amount of virus was mixed with serial dilutions of serum and incubated to allow binding of the antibodies to the virus. A cell suspension, containing a defined amount of MDCK cells was then added to the mixture of

virus and antiserum and incubated at 33°C. After the incubation period, virus replication was visualised by hemagglutination of chicken red blood cells. The 50% neutralisation titre of a serum was calculated by the method of Reed and Muench.

I.3.4. Cell-mediated Immunity was evaluated by Cytokine Flow Cytometry (CFC)

Peripheral blood antigen-specific CD4 and CD8 T cells can be restimulated *in vitro* to produce IL-2, CD40L, TNF-alpha and IFN if incubated with their corresponding antigen. Consequently, antigen-specific CD4 and CD8 T cells can be enumerated by flow cytometry following conventional immunofluorescence labelling of cellular phenotype as well as intracellular cytokines production. In the present study, Influenza vaccine antigen as well as peptides derived from specific influenza protein were used as antigen to restimulate Influenza-specific T cells. Results were expressed as a frequency of cytokine(s)-positive CD4 or CD8 T cell within the CD4 or CD8 T cell sub-population.

I.3.5. Statistical Methods

I.3.5.1. Primary endpoints

- Percentage, intensity and relationship to vaccination of solicited local and general signs and symptoms during a 7 day follow-up period (i.e. day of vaccination and 6 subsequent days) after vaccination and overall.
- Percentage, intensity and relationship to vaccination of unsolicited local and general signs and symptoms during a 21 day follow-up period (i.e. day of vaccination and 20 subsequent days) after vaccination and overall.

- Occurrence of serious adverse events during the entire study.

I.3.5.2. Secondary endpoints

For the humoral immune response:

Observed variables:

- At days 0 and 21: serum hemagglutination-inhibition (HI) and NI antibody titres, tested separately against each of the three influenza virus strains represented in the vaccine (anti-H1N1, anti-H3N2 & anti-B-antibodies).
- At days 0 and 21: neutralising antibody titres, tested separately against each of the three influenza virus strains represented in the vaccine

Derived variables (with 95% confidence intervals):

• Geometric mean titres (GMTs) of serum HI antibodies with 95% confidence intervals (95% CI) pre and post-vaccination

• Seroconversion rates* with 95% CI at day 21

• Conversion factors** with 95% CI at day 21

• Seroprotection rates*** with 95% CI at day 21

• Serum NI antibody GMTs' (with 95% confidence intervals) at all timepoints.

* Seroconversion rate defined as the percentage of vaccinees who have at least a 4-fold increase in serum HI titres on day 21 compared to day 0, for each vaccine strain.

**Conversion factor defined as the fold increase in serum HI GMTs on day 21 compared to day 0, for each vaccine strain.

***Protection rate defined as the percentage of vaccinees with a serum HI titre =40 after vaccination (for each vaccine strain) that usually is accepted as indicating protection.

For the cell mediated immune (CMI) response

Observed variable

At days 0 and 21: frequency of cytokine-positive CD4/CD8 cells per 10^6 in different tests. Each test quantifies the response of CD4/CD8 T cell to:

• Peptide Influenza (pf) antigen (the precise nature and origin of these antigens needs to be given/explained

• Split Influenza (sf) antigen

• Whole Influenza (wf) antigen.

Derived variables:

• cells producing at least two different cytokines (CD40L, IL-2, IFNγ, TNFα)

• cells producing at least CD40L and another cytokine (IL-2, TNFα, IFNγ)

• cells producing at least IL-2 and another cytokine (CD40L, TNFα, IFNγ)

• cells producing at least IFNγ and another cytokine (IL-2, TNFα, CD40L)

• cells producing at least TNFα and another cytokine (IL-2, CD40L, IFNγ)

I.3.5.3. Analysis of immunogenicity

The immunogenicity analysis was based on the total vaccinated cohort. For each treatment group, the following parameters (with 95% confidence intervals) were calculated:

• Geometric mean titres (GMTs) of HI and NI antibody titres at days 0 and 21

- Geometric mean titres (GMTs) of neutralising antibody titres at days 0 and 21.
- Conversion factors at day 21.
- Seroconversion rates (SC) at day 21 defined as the percentage of vaccinees that have at least a 4-fold increase in serum HI titres on day 21 compared to day 0.
- Protection rates at day 21 defined as the percentage of vaccinees with a serum HI titre =1:40.
- The frequency of CD4/CD8 T-lymphocytes secreting in response was summarised (descriptive statistics) for each vaccination group, at each timepoint (Day 0, Day 21) and for each antigen (Peptide influenza (pf), split influenza (sf) and whole influenza (wf)).
- Descriptive statistics in individual difference between timepoint (Post-Pre) responses fore each vaccination group and each antigen (pf, sf, and wf) at each 5 different tests.
- A non-parametric test (Kruskall-Wallis test) was used to compare the location differences between the 3 groups and the statistical p-value was calculated for each antigen at each 5 different tests. All significance tests were two-tailed. P-values less than or equal to 0.05 were considered as statistically significant.

Example II – Preparation and characterization of the oil in water emulsion and adjuvant formulations

Unless otherwise stated, the oil/water emulsion used in the subsequent examples is composed an organic phase made of 2 oils (alpha-tocopherol and squalene), and an aqueous phase of PBS containing Tween 80 as emulsifying agent. Unless otherwise stated, the oil in water emulsion adjuvant formulations used in the subsequent examples were made comprising the following oil in water emulsion component (final concentrations given): 2.5% squalene (v/v), 2.5% alpha-tocopherol (v/v), 0.9% polyoxyethylene sorbitan monooleate (v/v) (Tween 80), see WO 95/17210. This emulsion, termed AS03 in the subsequent examples, was prepared as followed as a two-fold concentrate.

II.1. Preparation of emulsion SB62

II.1.1. Lab-scale preparation

Tween 80 is dissolved in phosphate buffered saline (PBS) to give a 2% solution in the PBS. To provide 100ml two-fold concentrate emulsion 5g of DL alpha tocopherol and 5ml of squalene are vortexed to mix thoroughly. 90ml of PBS/Tween solution is added and

mixed thoroughly. The resulting emulsion is then passed through a syringe and finally microfluidised by using an M110S microfluidics machine. The resulting oil droplets have a size of approximately 120-180 nm (expressed as Z average measured by PCS).
The other adjuvants/antigen components are added to the emulsion in simple admixture.

II.1.2. Scaled-up preparation

The preparation of the SB62 emulsion is made by mixing under strong agitation of an oil phase composed of hydrophobic components (α-tocopherol and squalene) and an aqueous phase containing the water soluble components (Tween 80 and PBS mod (modified), pH 6.8). While stirring, the oil phase (1/10 total volume) is transferred to the aqueous phase (9/10 total volume), and the mixture is stirred for 15 minutes at room temperature. The resulting mixture then subjected to shear, impact and cavitation forces in the interaction chamber of a microfluidizer (15000 PSI – 8 cycles) to produce submicron droplets (distribution between 100 and 200 nm). The resulting pH is between 6.8 ± 0.1. The SB62 emulsion is then sterilised by filtration through a 0.22 µm membrane and the sterile bulk emulsion is stored refrigerated in Cupac containers at 2 to 8°C. Sterile inert gas (nitrogen or argon) is flushed into the dead volume of the SB62 emulsion final bulk container for at least 15 seconds.

The final composition of the SB62 emulsion is as follows :
Tween 80: 1.8 % (v/v) 19.4 mg/ml; Squalene: 5 % (v/v) 42.8 mg/ml; α-tocopherol: 5 % (v/v) 47.5 mg/ml; PBS-mod: NaCl 121 mM, KCl 2.38 mM, Na2HPO4 7.14 mM, KH2PO4 1.3 mM; pH 6.8 ± 0.1.

II.2. Measure of oil droplet size dynamic light scattering

II.2.1. Introduction

The size of the diameter of the oil droplets is determined according to the following procedure and under the following experimental conditions. The droplet size measure is given as an intensity measure and expressed as z average measured by PCS.

II.2.2. Sample preparation

Size measurements have been performed on the oil-in-water emulsion adjuvant: SB62 prepared following the scaled-up method, AS03 and AS03+MPL (50 µg/ml), the last two being prepared just before use. The composition of the samples is given below (see section II.2.4). Samples were diluted 4000x – 8000x in PBS 7.4.

As a control, PL-Nanocal Particle size standards 100 nm (cat n° 6011-1015) was diluted in 10 mM NaCl.

II.2.3. Malvern Zetasizer 3000HS size measurements

All size measurements were performed with both Malvern Zetasizer 3000HS.
Samples were measured into a plastic cuvette for Malvern analysis at a suitable dilution (usually at a dilution of 4000x to 20000x depending on the sample concentration), and with two optical models:

- either real particle refractive index of 0 and imaginary one of 0.
- or real particle refractive index of 1.5 and imaginary one of 0.01 (the adapted optical model for the emulsion, according to the values found in literature).

The technical conditions were:

- laser wavelength: 532 nm (Zeta3000HS).
- laser power: 50 mW (Zeta3000HS).
- scattered light detected at 90° (Zeta3000HS).
- temperature: 25°C,
- duration: automatic determination by the soft,
- number: 3 consecutive measurements,
- z-average diameter: by cumulants analysis
- size distribution: by the Contin or the Automatic method.

The Automatic Malvern algorithm uses a combination of cumulants, Contin and non negative least squares (NNLS) algorithms.
The intensity distribution may be converted into volume distribution thanks to the Mie theory.

II.2.4. Results (see Table 2)

Cumulants analysis (Z average diameter):

Table 2

Sample	Dilution	Record	Count rate	ZAD	Polydispersity
SB62	5000	1 2 3 **average**	7987 7520 6586 **7364**	153 153 152 **153**	0.06 0.06 0.07 **0.06**
SB62 (Example IV)	8000	1	8640	151	0.03

		2	8656	151	0.00
		3	8634	150	0.00
		average	**8643**	**151**	**0.01**
SB62+MPL 25µg (*)	8000	1	8720	154	0.03
		2	8659	151	0.03
		3	8710	152	0.02
		average	**8697**	**152**	**0.02**

(*) Prepared as follows : Water for injection, PBS 10x concentrated , 250µl of SB62 emulsion and 25µg of MPL are mixed together to reach a final volume of 280µl.

The z-average diameter (ZAD) size is weighed by the amount of light scattered by each size of particles in the sample. This value is related to a monomodal analysis of the sample and is mainly used for reproducibility purposes.

The count rate (CR) is a measure of scattered light: it corresponds to thousands of photons per second.

The polydispersity (Poly) index is the width of the distribution. This is a dimensionless measure of the distribution broadness.

Contin and automatic analysis:

Two other SB62 preparations (2 fold concentrated AS03) have been made and assessed according to the procedure explained above with the following minor modifications:

Samples were measured into a plastic cuvette for Malvern analysis, at two dilutions determined to obtain an optimal count rate values: 10000x and 20000x for the Zetasizer 3000HS, the same optical models as used in the above example.

Results are shown in Table 3.

Table 3

		IR		Analysis in Contin (mean in nm)		Analysis in Automatic (mean in nm)	
SB62	**Dilution**	**Real**	**Imaginary**	**Intensity**	**Volume**	**Intensity**	**Volume**
1022	1/10000	0	0	149	167	150	-
		1.5	0.01	158	139	155	143
	1/20000	0	0	159	200	155	196
		1.5	0.01	161	141	147	-
1023	1/10000	0	0	158	198	155	-
		1.5	0.01	161	140	150	144
	1/20000	0	0	154	185	151	182
		1.5	0.01	160	133	154	-

"-" when the obtained values were not coherent.

A schematic representation of these results is shown in Figure 1 for formulation 1023. As can be seen, the great majority of the particles (e.g. at least 80%) have a diameter of less than 300 nm by intensity.

II.2.5. Overall Conclusion

SB62 formulation was measured at different dilutions with the Malvern Zetasizer 3000HS and two optical models. The particle size ZAD (i.e. intensity mean by cumulant analysis) of the formulations assessed above was around 150-155 nm.

When using the cumulants algorithm, we observed no influence of the dilution on the ZAD and polydispersity.

II.3. Preparation of AS03 comprising MPL

II.3.1. Preparation of MPL liquid suspension

The MPL (as used throughout the document it is an abbreviation for 3D-MPL, i.e. 3-O-deacylated monophosphoryl lipid A) liquid bulk is prepared from MPL® lyophilized powder. MPL liquid bulk is a stable concentrated (around 1 mg/ml) aqueous dispersion of the raw material, which is ready-to-use for vaccine or adjuvant formulation. A schematic representation of the preparation process is given in Figure 2.

For a maximum batch size of 12g, MPL liquid bulk preparation is carried over in sterile glass containers. The dispersion of MPL consists of the following steps:

- suspend the MPL powder in water for injection
- desaggregate any big aggregates by heating (thermal treatment)
- reduce the particle size between 100 nm and 200 nm by microfluidization
- prefilter the preparation on a Sartoclean Pre-filter unit, 0.8/0.65µm
- sterile filter the preparation at room temperature (Sartobran P unit, 0.22 µm)

MPL powder is lyophilized by microfluidisation resulting in a stable colloidal aqueous dispersion (MPL particle size smaller than 200 nm). The MPL lyophilized powder is dispersed in water for injection in order to obtain a coarse 10 mg/ml suspension. The suspension then undergoes a thermal treatment under stirring. After cooling to room temperature, the microfluidization process is started in order to decrease the particle size. Microfluidization is conducted using Microfluidics apparatus M110EH, by continuously circulating the dispersion through a microfluidization interaction chamber, at a defined pressure for a minimum amount of passages (number of cycles: n_{min}). The

microfluidization duration, representing the number of cycles, is calculated on basis of the measured flow rate and the dispersion volume. On a given equipment at a given pressure, the resulting flow rate may vary from one interaction chamber to another, and throughout the lifecycle of a particular interaction chamber. In the present example the interaction chamber used is of the type F20Y Microfluidics. As the microfluidization efficiency is linked to the couple pressure – flow rate, the processing time may vary from one batch to another. The time required for 1 cycle is calculated on basis of the flow rate. The flow rate to be considered is the flow rate measured with water for injection just before introduction of MPL into the apparatus. One cycle is defined as the time (in minutes) needed for the total volume of MPL to pass once through the apparatus. The time needed to obtain n cycles is calculated as follows:

$$\frac{n \times \text{quantity of MPL to treat (ml)}}{\text{flow rate (ml/min)}}$$

The number of cycles is thus adapted accordingly. Minimum amount of cycles to perform (n_{min}) are described for the preferred equipment and interaction chambers used. The total amount of cycles to run is determined by the result of a particle size measurement performed after n_{min} cycles. A particle size limit (d_{lim}) is defined, based on historical data. The measurement is realized by photon correlation spectroscopy (PCS) technique, and d_{lim} is expressed as an unimodal result ($Z_{average}$). Under this limit, the microfluidization can be stopped after n_{min} cycles. Above this limit, microfluidization is continued until satisfactory size reduction is obtained, for maximum another 50 cycles.

If the filtration does not take place immediately after microfluidization, the dispersed MPL is stored at +2 to +8°C awaiting transfer to the filtration area.
After microfluidization, the dispersion is diluted with water for injection, and sterile filtered through a 0.22µm filter under laminal flow. The final MPL concentration is 1 mg/ml (0.80-1.20 mg/ml).

II.3.2. Preparation of AS03+MPL adjuvanted vaccine: 1 vial approach

To the AS03 adjuvant formulation, MPL is added at a final concentration of between 10 and 50 µg per vaccine dose.

PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a SB62 mixture containing Tween, Triton X-100 and VES (vitamin E succinate, i.e. alpha-tocopherol succinate) is added to water for injection. The quantities take into account the detergent present in the influenza strains so as to reach a target final concentration of 750µg/ml

Tween 80, 110µg/ml Triton X-100 and 100µg/ml VES. After 5 min stirring, 15µg of each influenza strain of interest (for example strain H1N1, H3N2 and B in a classical tri-valent vaccine) are added. After 15 min stirring, 250µl of SB62 emulsion is added and then 25µg or 50µg of MPL.

A schematic representation of the preparation process is given in Figure 3. The final composition of AS03 comprising MPL per human dose is given the Table 4.

Table 4

Ingredients		Concentration	Per human dose	
Name	Component		Quantity	Other
SB62	Squalene (solution 43 mg/ml) Tocopherol (solution 48 mg/ml) Tween 80 (solution 20 mg/ml)	781 µl/ml	250 µl 10.68 mg 11.86 mg 4.85 mg	
MPL**	(solution 1 mg/ml)	78 µg/ml or 156 µg/ml	25 µg or 50 µg	
PBS mod*	NaCl KCl Na2HPO4 KH2PO4	137 mM 2.7 mM 8.1 mM 1.47 mM	2.56 mg 0.064 mg 0.368 mg 0.064 mg	
Water for injection				Ad 320 µl
pH				6.8 +/- 0.1

*PBS mod 10x concentrated pH 6.8 = KH2PO4, Na2HPO4, NaCl, KCl-HCl
**MPL is either 25 µg or 50 µg per dose

II.3.3. Preparation of AS03+MPL adjuvanted vaccine: 2 vials approach

The same formulation can be prepared from a 2 vials approach by mixing 2 fold concentrated antigen or antigenic preparation with the AS03 (SB62 250µl) or the AS03+MPL (SB62 250µl + 25µg or 50 µg MPL) adjuvant. In this instance it is proceeded as follows. The manufacturing of the AS25-adjuvanted influenza vaccine consists of three main steps:

1) Formulation of the trivalent final bulk (2 x concentrated) without adjuvant and filling in the antigen container

2) Preparation of the AS03+MPL adjuvant

3) Extemporaneous reconstitution of the AS03+MPL adjuvanted split virus vaccine.

1) Formulation of the trivalent final bulk without adjuvant and filling in the antigen

container

The volumes of the three monovalent bulks are based on the HA content measured in each monovalent bulk prior to the formulation and on a target volume of 1100 ml. Concentrated phosphate buffered saline and a pre-mixture of Tween 80, Triton X-100 and α-tocopheryl hydrogen succinate are diluted in water for injection. The three concentrated monobulks (A/New Caledonia, A/New York, B/Jiangsu) are then successively diluted in the resulting phosphate buffered saline / Tween 80 - Triton X-100 - α-tocopheryl hydrogen succinate solution (pH 7.4, 137 mM NaCl, 2.7 mM KCl, 8.1 mM Na2HPO4, 1.47 mM KH2PO4, 990 µg/ml Tween 80, 150 µg/ml Triton X-100 and 130 µg/ml α-tocopheryl hydrogen succinate) in order to have a final concentration of 39.47 µg HA of A strains (H1N1, H3N2) per ml of trivalent final bulk (15 µg HA/ A strain /380 µl trivalent final bulk) and 46 µg HA of B strain (17.5 µg HA/ B strain /380 µl trivalent final bulk). Between addition of each monovalent bulk, the mixture is stirred for 10-30 minutes at room temperature. After addition of the last monovalent bulk and 15-30 minutes of stirring, the pH is checked and adjusted to 7.2 ± 0.2 with HCl or NaOH.

The trivalent final bulk of antigens is aseptically filled into 3-ml sterile Type I (Ph. Eur.) glass vials. Each vial contains a volume of 470 µl (380 µl + 90 µl overfill).

2) Preparation of AS03/MPL adjuvant bulk and filling in the adjuvant container.

The adjuvant *AS03/MPL* is prepared by mixing of two components: SB62 emulsion (method in section II.1.2) and MPL (method in section II.3.1). One-fold concentrated PBS mod (prepared by diluting 10 x concentrated PBS mod in water for injection) with SB62 bulk and MPL liquid bulk at 1 mg/ml. MPL concentration will be determined so as to reach a final content of between 10 to 50 µg, suitably around 25µg per final human vaccine dose. The mixture is stirred for 5-30 minutes at room temperature, and the pH is adjusted to 6.8 ± 0.1 with NAOH (0.05 or 0.5 M)/ HCl (0.03 M or 0.3 M). After another stirring for 5-30 minutes at room temperature the mixture is sterilised by filtration through a 0.22 µm membrane. Sterile inert gas (nitrogen) flushing is performed to produce inert head space in the filled containers during minimum 1 minute. The sterile AS03+MPL adjuvant is stored at +2-8°C until aseptical filling into 1.25-ml sterile Type I (Ph. Eur.) glass syringes. Each syringe contains a volume overage of 80 µl (320 µl + 80 µl overfill).

At the time of injection, the content of the prefilled syringe containing the adjuvant is injected into the vial that contains the concentrated trivalent inactivated split virion antigens. After mixing the content is withdrawn into the syringe and the needle is

replaced by an intramuscular needle. One dose of the reconstituted the AS25-adjuvanted influenza candidate vaccine corresponds to 0.7 mL.

II.4. Preparation of immunogenic compositions comprising an influenza antigen and optionally MPL in an oil in water emulsion formulation

To the SB62 emulsion of II.1 an equal volume of twice concentrated split influenza antigen (Fluarix™) (15 µg HA per strain) was added and mixed. This was combined, when appropriate, with 50 µg/ml of MPL to give the final formulation.

Example III – Clinical trial in an elderly population aged over 65 years with a vaccine containing a split influenza antigen preparation and AS03 adjuvant (Explo-Flu-001)

A phase I, open, randomised study was conducted in an elderly population aged over 65 years in 2003 in order to evaluate the reactogenicity and the immunogenicity of GlaxoSmithKline Biologicals influenza candidate vaccine containing the adjuvant AS03. The humoral immune response (i.e. anti-hemagglutinin, neutralising and anti-neuraminidase antibody titres) and cell mediated immune response (CD4 and/or CD8 T cell responses) was measured 21 days after intramuscular administration of one dose of an AS03 adjuvanted vaccine or a WV vaccine. Fluarix™ was used as reference.

III.1. Study design

Three groups of subjects in parallel received the following vaccine intramuscularly:

- one group of 50 subjects receiving one dose of the reconstituted and adjuvanted SV influenza vaccine (FluAS03)
- one group of 50 subjects receiving one dose of whole virus influenza vaccine (FluWVV)
- one group of 50 subjects receiving one dose of Fluarix™ (Fluarix) = control

Vaccination schedule: one injection of influenza vaccine at day 0, blood sample collection, read-out analysis at day 21 (HI antibody determination, NI antibody determination, determination of neutralising antibodies, and CMI analysis) and study conclusion.

The standard trivalent split influenza vaccine - Fluarix™ used in this study, is a commercial vaccine from the year 2003 developed and manufactured by GlaxoSmithKline Biologicals.

III.2. Vaccine composition and administration (Table 5)

III.2.1. Vaccine preparation

AS03 adjuvanted influenza vaccine

The AS03-adjuvanted influenza vaccine candidate is a 2 components vaccine consisting of a concentrated trivalent inactivated split virion antigens presented in a type I glass vial (335 µl) (antigen container) and of a pre-filled type I glass syringe containing the SB62 emulsion (335 µl) (adjuvant container). At the time of injection, the content of the antigen container is removed from the with the help of the SB62 emulsion pre-filled syringe, followed by gently mixing of the syringe. Mixing of the SB62 emulsion with the vaccine antigens reconstitute the AS03 adjuvant. Prior to injection, the used needle is replaced by an intramuscular needle and the volume is corrected to 500 µl.

One dose of the reconstituted AS03-adjuvanted influenza vaccine corresponds to 0.5 ml, contains 15 µg HA of each influenza virus strain as in the registered Fluarix™/α-Rix® vaccine and contains 10.68 mg squalene, 11.86 mg DL-alpha tocopherol, and 4.85 mg polysorbate 80 (Tween 80).

Preparation

The manufacturing of the AS03-adjuvanted influenza vaccine consists of three main steps:

1) Formulation of the trivalent final bulk without adjuvant and filling in the antigen container.

The volumes of the three monovalent bulks are based on the HA content measured in each monovalent bulk prior to the formulation and on a target volume of 800 ml.

Concentrated phosphate buffered saline and a pre-mixture of Tween 80, Triton X-100 and α-tocopheryl hydrogen succinate are diluted in water for injection. The three concentrated monobulks (strain A/New Caledonia -, strain A/Panama - and strain B/Shangdong -) are then successively diluted in the resulting phosphate buffered saline / Tween 80 - Triton X-100 - α-tocopheryl hydrogen succinate solution (pH 7.4, 137 mM NaCl, 2.7 mM KCl, 8.1

mM Na_2HPO_4, 1.47 mM KH_2PO_4, 1500 µg/ml Tween 80, 220 µg/ml Triton X-100 and 200 µg/ml α-tocopheryl hydrogen succinate) in order to have a final concentration of 60 µg HA of A strains per ml of trivalent final bulk (15 µg HA/ A strain /250 µl trivalent final bulk) and 70 µg HA of B strain (17.5 µg HA/ B strain /250 µl trivalent final bulk). Between addition of each monovalent bulk, the mixture is stirred for 10 minutes at room temperature. After addition of the last monovalent bulk and 15 minutes of stirring, the pH is checked and adjusted to 7.2 ± 0.1 with HCl or NaOH.

The trivalent final bulk of antigens is aseptically filled into 3-ml sterile Type I glass vials. Each vial contains a 34 % volume overage (335 µl total volume).

2) Preparation of the SB62 emulsion sterile bulk and filling in the adjuvant container.

- Aqueous phase: while stirring, 902 ml of Tween 80 is mixed with 44105 ml of PBS-mod buffer (pH = 6.8 after adjustment with HCl).
- Oil phase: while stirring, 2550 ml of squalene is added to 2550 ml of α-tocopherol.
- Mixing of the aqueous and oil phases: while stirring, 5000 ml of oil phase (1/10 total volume) is transferred to 45007 ml of aqueous phase (9/10 total volume). The mixture is stirred for 15 minutes at room temperature.
- Emulsification: the resulting mixture is subjected to shear, impact and cavitation forces in the interaction chamber of a microfluidizer (15000 PSI – 8 cycles) to produce submicron droplets (distribution between 100 and 200 nm). The resulting pH is between 6.8 ± 0.1.
- Sterile filtration: the SB62 emulsion is sterilised by filtration through a 0.22 µm membrane and the sterile bulk emulsion is stored refrigerated in Cupac containers at 2 to 8 °C. Sterile inert gas (nitrogen or argon) is flushed into the dead volume of the SB62 emulsion final bulk container for at least 15 seconds.

All quantities of ingredients given are for the preparation of 50 L of emulsion and are given in volumes. In practice, amounts are weighed taking into account the densities of the ingredients. Density of PBS is considered equal to 1.

The final composition of the SB62 emulsion is as follows :

Table 5

Tween 80:	1.8 % (v/v)	19.4 mg/ml
Squalene:	5 % (v/v)	42.8 mg/ml
alpha-tocopherol:	5 % (v/v)	47.5 mg/ml

PBS-mod:		
NaCl		121 mM
KCl		2.38 mM
Na_2HPO_4		7.14 mM
KH_2PO_4		1.3 mM
pH		6.8 ± 0.1

The sterile SB62 bulk emulsion is then aseptically filled into 1.25-ml sterile Type I glass syringes. Each syringe contains a 34 % volume overage (335 µl total volume).

3) Extemporaneous reconstitution of the AS03 adjuvanted split virus vaccine.
At the time of injection, the content of the vial containing the concentrated trivalent inactivated split virion antigens is removed from the vial with the help the syringe containing the SB62 emulsion followed by gently mixing of the syringe. Mixing of the SB62 emulsion with the vaccine antigens reconstitutes the AS03 adjuvant.

III.2.2. Vaccine composition (Table 6) and administration

Table 6

Vaccine	Formulation	Group
Fluarix ™	HA from 3 influenza strains (total HA = 45 µg) • A/New Caledonia/20/99 (IVR-116):15 µg • A/Panama/2007/99 (RESVIR-17): 15 µg • B/Shangdong/7/97: 15 µg Thiomersal content: 5 µg In pre-filled syringes of 0.5 ml	Fluarix
WVV	HA from 3 influenza strains (total HA = 45 µg) • A/New Caledonia/20/99 (IVR-116):15 µg • A/Panama/2007/99 (RESVIR-17): 15 µg • B/Shangdong/7/97: 15 µg Thiomersal content: 5 µg In vials of 0.5 ml	FluWVV
Fluarix + AS03	HA from 3 influenza strains (total HA = 45 µg) • A/New Caledonia/20/99 (IVR-116):15 µg • A/Panama/2007/99 (RESVIR-17): 15 µg • B/Shangdong/7/97: 15 µg Thiomersal content: 5 µg In vial of 0.335 ml (2 times concentrated)	Flu-AS03

	+ syringe (0.335ml) containing oil-in-water SB62 emulsion (scaled-up preparation)	

The vaccines were administered intramuscularly in the deltoid region of the non-dominant arm. The vaccinees were observed closely for at least 30 minutes, with appropriate medical treatment readily available in case of a rare anaphylactic reaction following the administration of vaccine.

III.3. Study population results

A total of 148 subjects were enrolled in this study: 49 subjects in the FluAS03 group, 49 subjects in the Fluarix group and 50 subjects in the FluWVV group. The mean age of the total vaccinated cohort at the time of vaccination was 71.8 years with a standard deviation of 6.0 years. The mean age and gender distribution of the subjects across the three vaccine groups was similar.

III.4. Safety conclusions

The administration of the influenza vaccine adjuvanted with AS03 was safe and clinically well tolerated in the study population, i.e. elderly people aged over 65 years.

III.5. Immunogenicity results

Analysis of immunogenicity was performed on the total vaccinated cohort.

III.5.1. Humoral immune response

In order to evaluate the humoral immune response induced by the AS03 adjuvanted vaccine, the following parameters (with 95% confidence intervals) were calculated for each treatment group:

- Geometric mean titres (GMTs) of HI and NI antibody titres at days 0 and 21
- Geometric mean titres (GMTs) of neutralising antibody titres at days 0 and 21.
- Seroconversion rates (SC) at day 21 defined as the percentage of vaccinees that have at least a 4-fold increase in serum HI titres on day 21 compared to day 0.
- Conversion factors at day 21 defined as the fold increase in serum HI GMTs on day 21 compared to day 0, for each vaccine strain.
- Protection rates at day 21 defined as the percentage of vaccinees with a serum HI

titre =1:40.

III.5.1.1 Anti-hemagglutinin antibody response

a) HI Geometric mean titres (GMT)

The GMTs for HI antibodies with 95% CI are shown in Table 7 (GMT for anti-HI antibody). Pre -vaccination GMTs of antibodies for all vaccine strains were within the same range in the three groups. After vaccination, anti-haemagglutinin antibody levels increased significantly. Post vaccination, there was a trend for higher GMTs of HI antibody for all three vaccine strains in the FluAS03 and Fluarix groups although there was some overlap of 95% CI between the Fluarix group and the FluWVV group.

Table 7

Antibody	Group	Timing	N	GMT		
				Value	95% CI	
					LL	UL
A/New Caledonia	FluAS03	Pre	49	25.6	17.3	37.9
	Fuarix	PI(day21)	49	317.7	219.1	460.7
	FluWVV	Pre	49	26.3	18.1	38.4
		PI(day21)	49	358.5	244.2	526.4
		Pre	50	19.7	13.6	28.6
		PI(day21)	50	138.2	90.3	211.7
A/Panama	FluAS03	Pre	49	52.3	35.4	77.4
	Fuarix	PI(day21)	49	366.1	264.5	506.6
	FluWVV	Pre	49	40.9	28.1	59.5
		PI(day21)	49	296.0	205.4	426.6
		Pre	50	25.8	18.0	37.1
		PI(day21)	50	165.6	116.0	236.5
B/ Shangdong	FluAS03	Pre	49	27.5	19.0	39.8
	Fuarix	PI(day21)	49	317.7	226.9	444.9
	FluWVV	Pre	49	26.0	17.2	39.2
		PI(day21)	49	270.0	187.0	389.7
		Pre	50	32.0	20.8	49.3
		PI(day21)	50	195.6	135.2	282.9

N = number of subjects with available results
95% CI = 95% confidence interval; LL = Lower Limit; UL = Upper Limit
MIN/MAX = Minimum/Maximum
PRE = Prevaccination at Day 0
PI(D21) = Post-vaccination at Day 21)

b) Conversion factors of anti-HI antibody titres, seroprotection rates and seroconversion rates (correlates for protection in human)

Results are presented in Table 8.

The *conversion factors* represent the fold increase in serum HI GMTs for each vaccine strain on day 21 compared to day 0. The conversion factor varies from 6.1 to 13.6 according to the virus strain and the vaccine. This conversion factor is largely superior to the 2.0 fold increase in GMT required by the European Authorities.

The *seroprotection rates* represent the proportion of subjects with a serum HI titre ≥40 on day 21. At the outset of the study, half of the subjects (range 34.0% -69.4%) in all groups had protective levels of antibodies for all strains At day 21, the seroprotection rates in the three groups ranged from 88.0% to 100% for the different virus strains. In terms of protection, this means that more than 88% of the subjects had a serum HI titre ≥40 after vaccination and were deemed to be protected against the three strains. This rate is largely superior to the seroprotection rate of 60% required in the ≥60 years old population, by the European Authorities.

The *seroconversion rates* represent the proportion of subjects with at least a four-fold increase in serum HI titres on day 21 as compared to day 0. Overall response rates for the three strains were essentially equal in the three groups. To be deemed effective and according to European Union, a vaccine should induce a seroconversion rate greater than 30% in the =60 years old population. In this study, the seroconversion rate was greater than 50% for the three groups.

Table 8

			Seroprotection rate	**Seroconversion rate**	**Conversion factor**
EU standard (>60 years)			**> 60%**	**> 30%**	**> 2.0**
Strains	**Group**	**N**	**% [95%CI]**	**% [95%CI]**	**GMR [95%CI]**
A/ New Caledonia	Flu AS03	49	98.0 [89.1-99.9]	69.4 [54.6-81.7]	12.4 [7.3-21.0]
	Fluarix	49	98.0 [89.1-99.9]	69.4[54.6-81.7]	13.6 [8.0-23.2]
	Flu WVV	50	88.0 [75.7-95.5]	52.0 [37.4-66.3]	7.0 [4.0-12.2]
A/ Panama	Flu AS03	49	100.0 [92.7-100.0]	55.1 [40.2-69.3]	7.0 [4.2-11.6]
	Fluarix	49	91.8 [80.4-97.7]	65.3 [50.4-78.3]	7.2 [4.7-11.3]
	Flu WVV	50	90.0 [78.2-96.7]	56.0 [41.3-70.0]	6.4 [3.9-10.4]
B/ shangdong	Flu AS03	49	100.0 [92.7-100.0]	73.5 [58.9-85.1]	11.6 [7.2-18.6]
	Fluarix	49	95.9 [86.0-99.5]	69.4 [54.6-81.7]	10.4 [6.5-16.5]
	Flu WVV	50	90.0 [78.2-96.7]	50.0 [35.5-64.5]	6.1 [3.6-10.3]

N = total number of subjects

In conclusion:

- Post vaccination, there was a trend for higher GMTs of HI antibody for all three vaccine strains in the FluAS03 and Fluarix groups although there was some overlap of 95% CI between the Fluarix group and the FluWVV group.
- The conversion factor varies from 6.1 to 13.6 according to the virus strain and the vaccine. This conversion factor is largely superior to the 2.0 fold increase in GMT required by the European Authorities.
- At day 21, the seroprotection rates in the three groups ranged from 88.0% to 100% for the different virus strains. This rate is largely superior to the seroprotection rate of 60% required in the ≥ 60 years old population, by the European Authorities.
- In this study, the seroconversion rate was greater than 50% for the three groups. Overall response rates for the three strains were essentially equal in the three groups.

III.5.1.2 Neutralising antibody titers

In order to better characterise the immune response to influenza vaccination in the elderly, the serum antibody responses to the neutralising antigens was assessed. Results are shown in Table 9 (Seroprotection rates and geometric mean titres (GMT) for anti-neutralising antibody titres) and Table 10 (Seroconversion rates for anti-neutralising at post vaccination day 21 (fold-increase = 4)).

Titres of neutralising antibody against the three influenza strains were measured in pre- and post-immunisation sera. The following parameters were determined:

• Geometric mean titres (GMTs) of serum neutralising antibodies with 95% confidence intervals (95% CI) pre and post-vaccination

• Seroconversion rates with 95% CI at day 21, defined as the percentage of vaccinees with at least a 4-fold increase in HI titres on day 21 compared to day 0, for each vaccine strain.

Table 9

Antibody	Group	Timing	N	>= 18 1/DIL				GMT		
						95% CI		Value	95% CI	
				n	%	LL	UL		LL	UL
A/NEW_CALEDONIA	1	PRE	49	46	93.9	83.1	98.7	106.6	77.6	146.6
		PI(D21)	49	49	100.0	92.7	100.0	870.2	608.5	1244.3
	2	PRE	49	48	98.0	89.1	99.9	115.6	89.4	149.5
		PI(D21)	49	49	100.0	92.7	100.0	955.8	649.5	1406.5
	3	PRE	50	46	92.0	80.8	97.8	87.7	63.6	120.8

		PI(D21)	50	50	100.0	92.9	100.0	375.4	271.2	519.6
A/PANAMA	1	PRE	49	49	100.0	92.7	100.0	724.7	558.0	941.1
		PI(D21)	49	49	100.0	92.7	100.0	2012.8	1438.4	2816.5
	2	PRE	49	49	100.0	92.7	100.0	727.8	556.1	952.6
		PI(D21)	49	49	100.0	92.7	100.0	1597.7	1128.8	2261.5
	3	PRE	50	50	100.0	92.9	100.0	512.0	409.3	640.6
		PI(D21)	50	50	100.0	92.9	100.0	977.8	738.2	1295.0
B/SHANGDONG	1	PRE	49	29	59.2	44.2	73.0	25.6	18.8	35.0
		PI(D21)	49	48	98.0	89.1	99.9	222.5	148.1	334.2
	2	PRE	49	27	55.1	40.2	69.3	29.3	20.1	42.7
		PI(D21)	49	49	100.0	92.7	100.0	190.4	127.6	284.3
B/SHANGDONG	3	PRE	50	31	62.0	47.2	75.3	33.4	23.1	48.4
		PI(D21)	50	46	92.0	80.8	97.8	117.8	82.6	168.0

Group 1: Flu vaccine mix Adjuvant 2x conc Flu vac
Group 2: Flu vaccine Flu vaccine
Group 3: Flu vaccine Flu WVV vaccine
N = number of subjects with available results
n/% = number/percentage of subjects with titre within the specified range
95% CI = 95% confidence interval; LL = Lower Limit; UL = Upper Limit
PRE = Pre-vaccination at Day 0
PI(D21) = Post-vaccination at Day 21)

Table 10

Antibody	**Group**	**N**	**Responders**			
			n	**%**	**95% CI**	
					LL	**UL**
A/New Caledonia	1	49	29	59.2	44.2	73.0
	2	49	30	61.2	46.2	74.8
	3	50	21	42.0	28.2	56.8
A/Panama	1	49	12	24.5	13.3	38.9
	2	49	9	18.4	8.8	32.0
	3	50	9	18.0	8.6	31.4
B/Shangdong	1	49	29	59.2	44.2	73.0
	2	49	26	53.1	38.3	67.5
	3	50	19	38.0	24.7	52.8

Group 1: Flu vaccine (DFLU58A16) mix Adjuvant (D621024A8) 2x conc Flu vac
Group 2: Flu vaccine (18854B9) Flu vaccine
Group 3: Flu vaccine (DFLU59A2) Flu WVV vaccine
N = number of subjects with both pre and post vaccination result available.
n = number of responders
% = Proportion of responders (n/N x 100).
95% CI = exact 95% confidence interval; LL = lower limit, UL = upper limit

The main findings are:

- For the three vaccines, at day 21, a seroprotection rate of 100 % is obtained for both A strains. For the B strain, the seroprotection rates in the three groups ranged from 92% to 100%.
- Post vaccination, there was a significant increase of GMT for all strains, in the three

groups. However, there was a trend for higher GMTs of neutralising antibody for all three vaccine strains in the FluAS03 and Fluarix groups than in the FluWVV although there was some overlap of 95% CI between the Fluarix group and the FluWVV group.

- For the seroconversion rates, overall response rates for the three strains were essentially equal in the three groups.

In all groups, the results were consistent with those obtained from the analysis performed for anti –hemagglutinin antibodies.

III.5.1.3 Nauraminidase (NA) antibody titers

In order to better characterise the immune response to influenza vaccination in the elderly population, the serum antibody responses to the neuraminidase antigens was assessed. Similarly to the HI antibody titre, the following endpoints were determined:

- GMT (taking the anti-log of the mean of the log titre transformations)
- Seroconversion rate defined as the percentage of vaccinees with at least a 4-fold increase in HI titres on day 21 compared to day 0, for each vaccine strain.

The GMTs and seroconversion rates for NI antibodies with 95% CI are shown in Table 11 (anti-NA antibody GMT) and Table 12 (Seroconversion rates of NA at post –vaccination (day 21) (4-fold-increase)).

Table 11

Antibody	Group	Timing	**N**	**GMT**	**95%CI**	
					LL	**UL**
A/New Caledonia	FluAS03	PRE	49	77.8	61.8	97.9
		PI(D21)	48	270.0	212.9	342.3
	Fluarix	PRE	49	77.8	64.6	93.6
		PI(D21)	49	249.1	190.0	326.5
	FluWVV	PRE	50	66.8	53.8	83.0
		PI(D21)	50	159.2	122.8	206.4
A/Panama	FluAS03	PRE	49	33.3	28.5	48.7
		PI(D21)	48	156.8	124.8	196.9
	Fluarix	PRE	49	34.2	25.6	45.8
		PI(D21)	49	133.7	100.9	177.3
	FluWVV	PRE	50	24.6	18.7	32.4
		PI(D21)	49	78.9	59.4	104.7
B/Shangdong	FluAS03	PRE	49	46.7	36.5	59.9
		PI(D21)	49	204.2	156.4	266.7
	Fluarix	PRE	49	46.1	35.3	60.1
		PI(D21)	49	133.7	100.9	177.3
	FluWVV	PRE	50	48.6	36.4	64.7
		PI(D21)	49	128.2	101.7	161.6

FluAS03: Flu vaccine (DFLU58A16) mix with AS03 Adjuvant (D621024A8)

Fluarix: Flu vaccine (18854B9)
FluWVV: Flu WVV vaccine (DFLU59A2)
PRE= Pre-vaccination, PI(D21)= Day 21 post vaccination
95%CI, LL, and UL = 95% confidence interval, lower and upper limit

Table 12

Antibody	Group	N	Responders			
			n	%	95% CI	
					LL	UL
A/New Caledonia	FluAS03	48	25	52.1	37.2	66.7
	Fluarix	49	24	49.0	34.4	63.7
	FluWVV	49	18	36.7	23.4	51.7
A/Panama	FluAS03	48	27	56.3	41.2	70.5
	Fluarix	49	23	46.9	32.5	61.7
	FluWVV	49	21	42.9	28.8	57.8
B/Shangdong	FluAS03	48	26	54.2	39.2	68.6
	Fluarix	49	23	46.9	32.5	61.7
	FluWVV	49	16	32.7	19.9	47.5

FluAS03: Flu vaccine (DFLU58A16) mix with AS03 Adjuvant (D621024A8), Fluarix: Flu vaccine (18854B9), FluWVV: Flu WVV vaccine (DFLU59A2)
N = number of subjects with both pre and post vaccination result available, n = number of responders.
% = Proportion of responders (n/N x 100).
95% CI = exact 95% confidence interval; LL = lower limit, UL = upper limit

The main findings are:

- Higher value of the GMT and seroconversion rates were observed for hemagglutinin than for neuraminidase.
- Pre -vaccination GMTs of antibodies for all vaccine strains were within the same range in the three groups. After vaccination, anti-neuraminidase antibody levels increased significantly. As for the HI antibody titres, post vaccination, there was a trend for higher GMTs of HI antibody for all three vaccine strains in the FluAS03 and Fluarix groups although there was some overlap of 95% CI between the Fluarix group and the FluWVV group.
- Regarding the seroconversion rates, overall response rates for the three strains were essentially equal in the three groups and for the three strains.

Our results show that healthy elderly vaccinated in this study against influenza developed good antibody responses to neuraminidase antigens whatever the influenza vaccine. However, the response to the neuraminidase antigen is lower than the response to the hemagglutinin antigen.

III.5.2. Cellular immune response

Peripheral blood antigen-specific CD4 and CD8 T cells can be restimulated in vitro to produce IL-2, CD40L, TNF-alpha and IFNγ if incubated with their corresponding antigen. Consequently, antigen-specific CD4 and CD8 T cells can be enumerated by flow cytometry following conventional immunofluorescence labelling of cellular phenotype as well as intracellular cytokines production. In the present study, Influenza vaccine antigen as well as peptides derived from specific influenza protein were used as antigen to restimulate Influenza-specific T cells. Results are presented for the CD4 and CD8 T-cell response in Tables 13 to 18.

Table 13 Antigen specific CD4 'T-cell responses expressed into cells producing at least two different cytokines: Descriptive Statistics on PRE and POST for CD40L/IL2/TNF-α/IFN-γ (Total vaccinated cohort)

Secretion	Antigen	Gr	Time point	N	Mean	SD	Min
CD40L/IL2/IFNγ/ TNFα in CD4	Peptide Influenza	1	Day 0	44	33.50	139.026	1.00
		1	Day 21	45	58.40	132.664	1.00
		2	Day 0	42	92.10	368.790	1.00
		2	Day 21	44	88.36	272.528	1.00
		3	Day 0	45	80.13	284.316	1.00
		3	Day 21	47	91.40	382.967	1.00
	Split Influenza	1	Day 0	47	1901.66	1596.203	102.00
		1	Day 21	48	6163.75	4265.900	773.00
		2	Day 0	45	2151.04	2622.594	265.00
		2	Day 21	49	4150.73	3712.469	328.00
		3	Day 0	48	1678.44	916.329	142.00
		3	Day 21	50	3374.60	1920.194	449.00
	Whole Influenza	1	Day 0	48	3134.33	2568.369	507.00
		1	Day 21	47	9332.04	6875.403	1482.00
		2	Day 0	47	3050.85	2654.936	486.00
		2	Day 21	49	6760.31	6788.258	1852.00
		3	Day 0	48	2955.33	2019.233	473.00
		3	Day 21	50	5661.40	4530.321	635.00

Secretion	Antigen	Gr	Time point	Q1	Median	Q3	Max	Kruskall -Wallis test (P-value)
CD40L/ IL2/ IFNγ/TNFα in CD4	Peptide Influenza	1	Day 0	1.00	1.00	4.00	915.00	
		1	Day 21	1.00	1.00	56.00	733.00	
		2	Day 0	1.00	1.00	54.00	2393.00	0.7631
		2	Day 21	1.00	1.00	69.50	1740.00	
		3	Day 0	1.00	1.00	65.00	1908.00	
		3	Day 21	1.00	1.00	63.00	2615.00	
	Split Influenza	1	Day 0	957.00	1560.00	2408.00	9514.00	
		1	Day 21	3468.00	4908.00	7624.00	21324.00	
		2	Day 0	930.00	1381.00	2274.00	16289.00	**0.0002**
		2	Day 21	2247.00	3036.00	4744.00	21924.00	
		3	Day 0	1086.00	1502.00	2189.00	3899.00	

		3	Day 21	2312.00	3040.00	4437.00	10431.00	
	Whole Influenza	1	Day 0	1730.00	2298.50	3876.00	15066.00	
		1	Day 21	4091.00	6523.00	14045.00	29251.00	
		2	Day 0	1190.00	2031.00	4161.00	11994.00	**0.0040**
		2	Day 21	3573.00	4621.00	7234.00	40173.00	
		3	Day 0	1421.50	2668.50	3411.50	10578.00	
		3	Day 21	2459.00	4315.00	7303.00	22053.00	

Group 1: FluAS03: Flu vaccine Fluarix™ mixed with AS03 Adjuvant
Group 2: Fluarix: Flu vaccine Fluarix™
Group 3: FluWVV: Flu WVV vaccine
SD = Standard Deviation; Min, Max = Minimum, Maximum
Q1 = First quartile; Q3 = Third quartile
N= number of subjects with available results
P-value: Kruskall –Wallis Test (Non-parametric procedure) to test location difference (Wilcoxon rank-sum test) between the 3 groups at Day 21.

Table 14 Antigen-specific CD4 T-cell responses expressed into cells producing at least two different cytokines: Descriptive Statistics on difference between PRE and POST ('Total vaccinated cohort)

Secretion	Antigen	Group	N	Mean	SD	Min
CD40L/IFN-γ/TNF-α in CD4	Peptide Influenza	1	44	9.57	159.363	-860.00
		2	42	-40.98	386.998	-2392.00
		3	45	-50.73	256.596	-1664.00
	Split Influenza	1	47	4307.02	4468.828	-8161.00
		2	45	1982.93	3802.332	-14318.0
		3	48	1555.90	1596.216	-526.00
	Whole Influenza	1	47	6197.98	7220.765	-11763.0
		2	47	3791.34	5820.894	-2128.00
		3	48	2535.98	3966.345	-4766.00
CD40L/IFN-γ/TNF-α in CD8	Peptide Influenza	1	42	-15.95	215.710	-451.00
		2	41	50.83	264.370	-614.00
		3	44	-52.11	243.811	-684.00
	Split Influenza	1	42	134.71	426.699	-603.00
		2	44	-65.05	822.036	-4938.00
		3	45	2.49	330.700	-1094.00
	Whole Influenza	1	39	189.38	1394.153	-2641.00
		2	44	-479.75	1790.094	-9455.00
		3	44	-243.73	719.269	-1892.00

Secretion	Antigen	Group	Q1	Median	Q3	Max	P-value
CD40L/IFN-γ/TNF-α in CD4	Peptide Influenza	1	0.00	0.00	37.50	430.00	0.0765
		2	-15.00	0.00	26.00	514.00	
		3	-37.00	0.00	0.00	212.00	
	Split Influenza	1	1888.00	3396.00	6634.00	19555.00	**<0.0001**
		2	699.00	1490.00	2573.00	15169.00	
		3	466.00	1183.50	2186.50	7851.00	
	Whole Influenza	1	2170.00	4009.00	11681.00	25570.00	**0.0003**
		2	1246.00	2382.00	3992.00	33801.00	

		3	503.00	1382.50	3300.50	19337.00	
CD40L/IFN-γ/TNF-α in CD8	Peptide Influenza	1	-106.00	0.00	81.00	655.00	0.0932
		2	-58.00	13.00	202.00	703.00	
		3	-160.50	0.00	53.00	567.00	
	Split Influenza	1	-122.00	35.50	221.00	1387.00	0.2121
		2	-64.50	0.00	160.50	1252.00	
		3	-99.00	0.00	76.00	1060.00	
	Whole Influenza	1	-420.00	49.00	591.00	5045.00	0.0851
		2	-1016.00	-263.50	180.00	3743.00	
		3	-651.00	-86.50	180.00	1011.00	

Table 15 **Antigen-specific CD4 T-cell responses expressed into cells producing at least CD40L and another cytokine: Descriptive Statistics on difference between PRE and POST ('Total vaccinated cohort)**

Secretion	Antigen	Group	N	Mean	SD	Min
CD40L in CD4	Peptide Influenza	1	44	10.09	153.007	-815.00
		2	42	-29.40	316.983	-1921.00
		3	45	-43.73	251.146	-1629.00
	Split Influenza	1	46	4266.20	4470.807	-8093.00
		2	45	2026.42	3511.508	-11482.0
		3	47	1512.34	1576.133	-494.00
	Whole Influenza	1	47	6071.96	7118.132	-11691.0
		2	47	3764.64	5740.762	-2114.00
		3	48	2544.27	3959.879	-4390.00
CD40L in CD8	Peptide Influenza	1	44	-19.41	81.675	-370.00
		2	41	-3.98	100.998	-399.00
		3	45	-5.56	64.666	-181.00
	Split Influenza	1	43	39.53	190.122	-438.00
		2	44	27.61	91.173	-155.00
		3	45	30.18	191.326	-291.00
	Whole Influenza	1	41	-91.24	617.077	-1779.00
		2	44	-115.91	588.424	-2583.00
		3	45	-150.89	367.300	-1239.00

Secretion	Antigen	Group	Q1	Median	Q3	Max	P-value
CD40L in CD4	Peptide Influenza	1	0.00	0.00	36.50	428.00	0.1233
		2	-8.00	0.00	27.00	494.00	
		3	-35.00	0.00	3.00	230.00	
	Split Influenza	1	1799.00	3156.50	6647.00	19480.00	**<0.0001**
		2	783.00	1485.00	2546.00	15021.00	
		3	469.00	1107.00	2035.00	7687.00	
	Whole Influenza	1	2109.00	4048.00	11472.00	25448.00	**0.0004**
		2	1212.00	2509.00	3957.00	33428.00	

		3	523.00	1392.00	3261.50	19478.00	
CD40L in CD8	Peptide Influenza	1	-2.00	0.00	0.50	100.00	0.9721
		2	-28.00	0.00	24.00	231.00	
		3	-13.00	0.00	3.00	176.00	
	Split Influenza	1	-35.00	0.00	140.00	608.00	0.6175
		2	-18.50	0.00	77.50	326.00	
		3	-9.00	0.00	28.00	1188.00	
	Whole Influenza	1	-142.00	-8.00	175.00	2087.00	0.3178
		2	-195.50	-34.50	150.00	1258.00	
		3	-270.00	-103.00	88.00	588.00	

Table 16 **Antigen-specific CD4 T-cell responses expressed into cells producing at least IFNγ and another cytokine: Descriptive Statistics on difference between PRE and POST ('Total vaccinated cohort)**

Secretion	Antigen	Group	N	N missing	Mean	SD	Min
IFNγ in CD4	Peptide Influenza	1	44	5	7.50	64.539	-171.00
		2	42	7	-30.67	277.984	-1766.00
		3	45	5	-27.91	103.403	-639.00
	Split Influenza	1	46	3	2712.87	2905.629	-4394.00
		2	45	4	1148.56	2526.536	-10586.0
		3	47	3	871.00	1016.251	-764.00
	Whole Influenza	1	47	2	4240.09	4811.891	-8272.00
		2	47	2	2445.38	4030.694	-3018.00
		3	48	2	1535.48	2456.915	-3670.00
IFNγ in CD8	Peptide Influenza	1	44	5	7.75	146.412	-226.00
		2	41	8	10.68	176.026	-420.00
		3	44	6	-49.80	217.214	-699.00
	Split Influenza	1	43	6	138.58	365.565	-470.00
		2	44	5	-112.82	793.746	-4919.00
		3	44	6	29.91	238.157	-708.00
	Whole Influenza	1	41	8	6.66	1642.577	-5610.00
		2	44	5	-471.55	1792.348	-9586.00
		3	44	6	-189.05	685.291	-1879.00

Secretion	Antigen	Group	Q1	Median	Q3	Max	P-value
	Peptide Influenza		-9.50	0.00	7.50	265.00	0.1541
		2	-5.00	0.00	24.00	222.00	
		3	-20.00	0.00	0.00	51.00	

IFNγ in CD4	Split Influenza	1	1273.00	1644.00	4057.00	13296.00	**<0.0001**
		2	405.00	931.00	1757.00	9426.00	
		3	283.00	624.00	1114.00	5031.00	
	Whole Influenza	1	1610.00	2693.00	7437.00	17489.00	**<0.0001**
		2	723.00	1487.00	2983.00	21594.00	
		3	232.50	810.00	2218.50	11319.00	
IFNγ in CD8	Peptide Influenza	1	-52.50	0.00	40.00	615.00	0.3322
		2	-1.00	0.00	72.00	610.00	
		3	-172.00	0.00	90.50	424.00	
	Split Influenza	1	-46.00	42.00	294.00	1549.00	0.1257
		2	-62.00	0.00	74.00	1028.00	
		3	-59.50	26.50	123.00	643.00	
	Whole Influenza	1	-385.00	131.00	450.00	5068.00	0.1179
		2	-955.50	-221.00	177.00	3492.00	
		3	-476.50	-36.50	198.00	1299.00	

Table 17 **Antigen-specific CD4 T-cell responses expressed into cells producing at least IL2 and another cytokine: Descriptive Statistics on difference between PRE and POST ('Total vaccinated cohort)**

Secretion	Antigen	Group	N	Mean	SD	Min
IL2 in CD4	Peptide Influenza	1	44	2.82	118.164	-595.00
		2	42	0.90	84.255	-167.00
		3	45	-28.62	191.709	-1222.00
	Split Influenza	1	46	3456.15	3853.960	-7009.00
		2	45	1738.29	2406.045	-451.00
		3	47	1210.02	1361.705	-634.00
	Whole Influenza	1	47	4839.02	5978.277	-9178.00
		2	47	2891.00	4493.387	-1370.00
		3	48	2042.50	3123.912	-3179.00
IL2 in CD8	Peptide Influenza	1	42	-30.60	219.777	-630.00
		2	41	38.85	210.715	-674.00
		3	45	-44.80	197.026	-526.00
	Split Influenza	1	41	54.85	250.817	-336.00
		2	44	-2.36	423.957	-2272.00
		3	45	-26.07	244.870	-1004.00
	Whole Influenza	1	39	56.21	406.262	-704.00
		2	44	-151.02	822.384	-4304.00
		3	45	-63.56	359.699	-1036.00

Secretion	Antigen	Group	Q1	Median	Q3	Max	P-value

	Peptide Influenza	1	-1.50	0.00	31.50	324.00	0.0806
		2	-34.00	0.00	2.00	362.00	
IL2 in CD4		3	-19.00	0.00	0.00	253.00	
	Split Influenza	1	1309.00	2598.50	5926.00	16988.00	**<0.0001**
		2	453.00	1113.00	2049.00	12273.00	
		3	331.00	806.00	1596.00	6474.00	
	Whole Influenza	1	1516.00	3341.00	8955.00	21032.00	**0.0006**
		2	995.00	1942.00	3007.00	26358.00	
		3	371.50	1083.50	2624.50	14057.00	
	Peptide Influenza	1	-111.00	0.00	103.00	412.00	0.1684
		2	-41.00	0.00	138.00	542.00	
IL2 in CD8		3	-150.00	-34.00	71.00	447.00	
	Split Influenza	1	-76.00	26.00	133.00	803.00	0.2311
		2	-78.50	0.00	121.50	1064.00	
		3	-93.00	-1.00	30.00	705.00	
	Whole Influenza	1	-167.00	63.00	261.00	1302.00	0.4586
		2	-444.50	-4.00	199.00	1398.00	
		3	-198.00	9.00	131.00	838.00	

Table 18 **Antigen-specific CD4 T-cell responses expressed into cells producing at least TNFα and another cytokine: Descriptive Statistics on difference between PRE and POST ('Total vaccinated cohort)**

Secretion	Antigen	Group	N	Mean	SD	Min
	Peptide Influenza	1	44	9.48	92.992	-466.00
		2	42	-47.71	367.624	-2333.00
TNF-α in CD4		3	45	-37.38	179.147	-1169.00
	Split Influenza	1	46	2343.11	2596.177	-4450.00
		2	45	703.87	2973.241	-14260.0
		3	47	732.00	740.001	-611.00
	Whole Influenza	1	47	3103.74	4248.997	-5146.00
		2	47	1658.38	3639.959	-1393.00
		3	48	1010.15	1689.394	-1482.00
	Peptide Influenza	1	42	11.71	201.031	-453.00
		2	41	37.46	245.241	-612.00
TNF-α in CD8		3	44	-42.95	210.185	-645.00
	Split Influenza	1	41	138.54	362.601	-329.00
		2	44	-70.27	790.309	-4741.00
		3	44	-39.75	348.803	-1044.00
	Whole Influenza	1	39	279.59	1048.352	-1184.00
		2	44	-280.70	1562.095	-9070.00
		3	44	-71.57	492.135	-1574.00

Secretion	Antigen	Group	Q1	Median	Q3	Max	P-value

TNF-α in CD4	Peptide Influenza	1	-1.50	0.00	39.00	239.00	0.1836
		2	-4.00	0.00	12.00	277.00	
		3	-26.00	0.00	5.00	53.00	
	Split Influenza	1	862.00	1466.50	3931.00	9267.00	**<0.0001**
		2	251.00	698.00	1229.00	12275.00	
		3	191.00	540.00	1010.00	3288.00	
	Whole Influenza	1	868.00	1607.00	5266.00	17199.00	**0.0008**
		2	367.00	871.00	1584.00	23540.00	
		3	175.00	592.00	1385.50	8760.00	
TNF-α in CD8	Peptide Influenza	1	-80.00	0.50	70.00	772.00	0.2759
		2	-81.00	0.00	155.00	791.00	
		3	-179.00	0.00	39.50	566.00	
	Split Influenza	1	-23.00	60.00	178.00	1468.00	0.0790
		2	-107.00	0.00	158.00	1286.00	
		3	-185.00	0.00	78.50	1021.00	
	Whole Influenza	1	-250.00	108.00	399.00	4601.00	0.1482
		2	-392.00	-56.50	205.00	3258.00	
		3	-233.50	-54.00	160.00	1543.00	

Results were also expressed as a frequency of cytokine(s)-positive CD4 or CD8 T cell within the CD4 or CD8 T cell sub-population and presented in Figure 4 and Figure 5.

In a similar analysis, the cross-reactive CD4 T-cells response was evaluated using influenza antigen from drifted strains (A/H1N1/Beijing/262/95 (H1N1d), A/H3N2/Sydney/5/97 (H3N2d), B/Yamanashi/166/98 (Bd)) or shift strains (A/Singapore/1/57 (H2N2), A/Hongkong/1073/99 (H9N2)). Results expressed as a frequency of cytokine(s)-positive CD4 T cells are presented in Figure 6.

The main findings are:

• Vaccination with Fluarix or Whole virus slightly boosts the CD4 T-cell response. Vaccination with Flu AS03 induces a strong CD4 T-cell response (Figure 4), and this is statistically significant. The same conclusion is made after In Vitro stimulation with the split antigen or Whole virus, and this with all cytokines investigated (IL-2, IFNγ, TNFα, and CD40L).

• Most individuals have a CD8 T-cell response against the whole flu, however the vaccination has no measurable impact on the CD8 T-cell response (i.e. Pre = post), whatever the group studied (Figure 5).

Vaccination with Fluarix only induces low levels of cross-reactive CD4 T-cell response (Figure 6). Vaccination with FluAS03 induces a strong CD4 T-cell response against drifted influenza strains and this is statistically significant (Figure 6). A little response was detected against shift strains.

III.5.3. B-cells ELISPOT MEMORY

III.5.3.1 Objective

In order to better characterise the CMI response induced by the AS03-adjuvanted influenza vaccine, the B-cells Elispot memory response induced to differentiate into plasma cells in vitro using influenza vaccine strains or anti-human immunoglobulin was evaluate in order to enumerate anti-influenza or IgG secreting plasma. The results are described in Table 19 and Table 20 and in Figure 7.

A subset of **22** first subjects having received one dose of FluAS03 vaccine and **21** first subjects having received one dose of Fluarix vaccine was selected to evaluate the impact of vaccination on influenza-specific memory B-cells using the B-cell memory Elispot technology. The following endpoints were determined

- At days 0 and 21: Influenza-specific memory B-cells have been measured by B-cell Elispot in all subjects. Results have been expressed as a frequency of Influenza specific- antibody forming cells per million (10^6) of antibody forming cells.
- Difference between post (day 21) and pre (day 0) vaccination is also expressed as a frequency of Influenza specific- antibody forming cells per million (10^6) of antibody forming cells.

III.5.3.2 Statistical methods

Descriptive statistics for each vaccination group at days 0 and day 21 expressed as a frequency of Influenza specific- antibody forming cells per million (10^6) of antibody forming cells. Descriptive statistics in individual difference between day 21 and day 0 (Post-Pre) as a frequency of Influenza specific- antibody forming cells per million (10^6) of antibody forming cells.

A Wilcoxon test was used to compare the location of difference between the two groups and the statistical p-value was calculated for each of 3 strains (A/New Caledonia, A/Panama and B/Shangdong).

III.5.3.3 Results

There is a tendency in favour of the influenza adjuvanted AS03 vaccine compared to

Fluarix group. For A/New Caledonia strain, there is a statistical significant difference (p-value = 0.021) in favour of FluAS03 compared to Fluarix. No statistical difference between the two groups was observed for A/Panama and B/Shangdong strains.

Table 19 B-cells Memory: descriptive statistics on pre (Day 0) and post (Day 21) and inferential statistics of post (Day 21) frequency of antigen-plasma within a 10^6 of IgG-producing plasma cells (subset of subjects)

STRAIN	Group	Time-point	N	Mean	SD	Min
A/NEW CALEDONIA	1	Day 0	22	9751.58	6630.335	0.00
	1	Day 21	22	22001.65	11308.261	3981.90
	2	Day 0	21	9193.61	4339.421	1300.81
	2	Day 21	21	12263.08	7285.698	789.47
A/PANAMA	1	Day 0	22	4329.17	2923.497	0.00
	1	Day 21	22	18066.69	14604.842	714.29
	2	Day 0	21	4860.41	3392.373	0.00
	2	Day 21	21	13872.95	12052.163	0.00
B/SHANDONG	1	Day 0	22	3722.80	2347.315	0.00
	1	Day 21	22	15949.60	12385.965	0.00
	2	Day 0	21	3030.39	2206.589	640.57
	2	Day 21	21	9714.03	5656.805	0.00

STRAIN	Gr	Time-point	Q1	Median	Q3	Max	P-value (Wilcoxon test)
A/NEW CALEDONIA	1	Day 0	4117.65	9606.46	13430.66	25570.78	
	1	Day 21	11052.63	20450.55	30234.74	40526.32	
	2	Day 0	6363.64	9686.41	11698.11	19164.84	**0.0056**
	2	Day 21	7741.05	9545.45	17069.60	32000.00	
A/PANAMA	1	Day 0	2275.45	4003.02	5764.55	10842.49	
	1	Day 21	9347.37	13176.41	21471.39	54789.92	
	2	Day 0	2222.22	4545.45	7495.74	11698.11	0.1814
	2	Day 21	6231.88	10147.06	20540.54	52188.84	
B/SHANDONG	1	Day 0	2058.82	2956.78	5972.22	7832.17	
	1	Day 21	6860.47	12796.90	22947.37	48947.37	
	2	Day 0	1290.32	2113.82	4770.02	7783.25	0.1483
	2	Day 21	6590.91	9009.01	12774.87	21201.72	

Group 1: Flu vaccine Fluarix™ + AS03 oil-in-water emulsion adjuvant
Group 2: Flu vaccine Fluarix™
SD = Standard Deviation
Min, Max = Minimum, Maximum
Q1 = First quartile
Q3 = Third quartile
N= number of subjects with available results
P-value: Wilcoxon Test (Non-parametric procedure) to test location difference (Wilcoxon rank-sum test) between the 2 groups at Day 21.

Table 20

B cells Memory: Descriptive and inferential statistics on *difference* between POST (Day 21) and PRE (Day 0) frequency of antigen-specific plasma within a 10 6 of IgG-producing plasma cells (subset of subjects)

STRAIN	Group	N	Mean	SD	Min
A/NEW CALEDONIA	1	22	12250.07	12875.755	-4365.08
	2	21	3069.46	7309.731	-10043.4
A/PANAMA	1	22	13737.52	13677.942	-188.29
	2	21	9012.54	11489.012	-1551.05
B/SHANDONG	1	22	12226.81	12243.895	-2222.22
	2	21	6683.64	6240.312	-2113.82

STRAIN	Gr	Q1	Median	Q3	Max	P-value (Wilcoxon test)
A/NEW CALEDONIA	1	2418.07	6776.65	26036.01	35059.98	**0.0210**
	2	-1762.54	1694.51	6850.19	18579.97	
A/PANAMA	1	4551.30	11039.04	16614.85	49881.94	0.1449
	2	1522.85	6480.96	9214.67	47812.47	
B/SHANDONG	1	1788.75	9322.70	18907.05	42134.18	0.1895
	2	2117.44	5384.41	9897.27	19801.28	

Group 1: Flu vaccine Fluarix™ + AS03 oil-in-water emulsion adjuvant
Group 2: Flu vaccine Fluarix™
SD = Standard Deviation
Min, Max = Minimum, Maximum
Q1 = First quartile
Q3 = Third quartile
N= number of subjects with available results
P-value: Wilcoxon Test (Non-parametric procedure) to test location difference (Wilcoxon rank-sum test) between the 2 groups at Day 21.

III.6. Overall conclusions

III.6.1. Reactogenicity and safety results

While influenza immunisation significantly reduces the risk of pneumonia and associated deaths, vaccination of elderly only affords 23-72% protection against influenza disease. Formulation of vaccine antigen with potent adjuvants is an attractive approach for enhancing immune responses to subunit antigens. This study was designed to evaluate (1) the safety and reactogenicity in healthy elderly of an influenza vaccine adjuvanted with oil in water emulsion, i.e. AS03, (2) the antibody and cell-mediated immune responses. Reactogenicity data show that the influenza vaccine adjuvanted with AS03 induced more local and general symptoms than the two other vaccines. However regarding unsolicited

adverse events, no difference was observed between the three vaccines. From these results, it can be concluded that the reactogenicity and safety profile of the candidate vaccines is satisfactory and clinically acceptable.

III.6.2. Immunogenicity results

Regarding the immune response, the three vaccines exceeded the requirements of the European authorities for annual registration of split virion influenza vaccines ("Note for Guidance on Harmonisation of Requirements for influenza Vaccines" for the immunological assessment of the annual strain changes -CPMP/BWP/214/96). The three influenza vaccines tested in this study were immunogenic in the healthy elderly, who developed a good antibody response to influenza hemagglutinin and neutralising antigens (Table 21).

Table 21

Variable	EU standard for antibody response	Results
Conversion factor	>2.0	> 6.1
Seroconversion rate	>30%	> 50%
Protection rate	>60%	> 88%

Regarding cell-mediated immunity (CMI) response, the influenza vaccine adjuvanted with AS03 induced a significantly stronger CD4 response (included drifted strains) than the two other vaccines (Fluarix and whole influenza virus vaccine). However, vaccination has no measurable impact on the CD8 response.

Regarding the B cell memory response, there is a tendency in favour of the influenza adjuvanted vaccine compared to the un-adjuvanted vaccine.

Example IV – Clinical trial in an elderly population aged over 65 years with a vaccine containing a split influenza antigen preparation and AS03 adjuvant - Explo-Flu-002

A phase I/II, open, controlled study has been conducted in order to evaluate the reactogenicity and the immunogenicity of the GlaxoSmithKline Biologicals influenza candidate vaccine containing the adjuvant AS03, in an elderly population aged over 65 years and previously vaccinated in 2003 with the candidate vaccine in the Explo-Flu-001

clinical trial. For immunogenicity and safety evaluations, Fluarix™ vaccine (known as α-rix™ in Belgium) has been used as reference.

IV.1. Objective

The humoral immune response (i.e. anti-hemagglutinin antibody titres) and cell mediated immune response (CD4 and/or CD8 T cell responses) and B memory cell response were measured 21 days after intramuscular administration of one dose of an AS03 adjuvanted vaccine. Fluarix™ was used as reference.

The objectives were:

1) to determine if AS03 adjuvanted Flu (40 subjects) versus *Fluarix* (18 subjects) confirm his strongest immunostimulating activity on CD4- and/or CD8-mediated immunity of individuals vaccinated with influenza antigens;

2) to investigate, using a longitudinal analysis, the influence of AS03 adjuvanted on the immune response in prevaccination 2004 (so response one year after the first vaccination in 2003).

IV.2. Study design, vaccine composition and end-points

- 40 subjects aged > 65 years who have previously received one dose of the AS03 adjuvanted influenza vaccine during the Explo-Flu-001 clinical trial in 2003 (FluAS03)
- one control group of about 20 subjects aged > 65 years who have previously received one dose of Fluarix™ during the Explo-Flu-001 clinical trial in 2003 (Fluarix)

IV.2.1. Vaccine composition

The vaccine composition is similar to that used for the study Explo-Flu-001 except for the influenza strains included in the vaccine (year 2004 vaccine). The strains are as follows:

- A/New Caledonia/20/99 (IVR-116) (H1N1) = A/New Caledonia/ (HINI) – like strain
- A/Wyoming/3/2003 (X-147) (H3N2) = A/Fujian (H3N2) – like strain
- B/Jiangsu/10/2003 = B/Shanghai – like strain

IV.2.2. Immunogenicity (HI) end-points

- GMTs (taking the anti-log of the mean of the log titre transformations)
- Conversion factors (the fold increase in serum HI GMTs on day 21 compared to day 0)
- Seroconversion rate (the percentage of vaccinees with at least a four-fold increases in HI titers on day 21 compared to day 0, for each vaccine strain)

- Protection rate (the percentage of vaccinees with a serum HI ≥1: 40 at day 21)

IV.2.3. CMI-Endpoints

Observed variable:

At days 0 and 21: frequency of cytokine-positive CD4/CD8 cells per 10^6 into 4 different cytokines. Each test quantifies the response of CD4/CD8 T cell to:

- Pool of the 3 following antigens
- New Caledonia antigen
- Wyoming antigen
- Jiangsu antigen.

Derived variables:

Antigen-specific CD4 and CD8-T-cell response expressed into the 5 different tests (cytokines):

1. cells producing at least two different cytokines (CD40L, IL-2, IFNγ, TNFα)
2. cells producing at least CD40L and another cytokine (IL-2, TNFα, IFNγ)
3. cells producing at least IL-2 and another cytokine (CD40L, TNFα, IFNγ)
4. cells producing at least IFNγ and another cytokine (IL-2, TNFα, CD40L)
5. cells producing at least TNFα and another cytokine (IL-2, CD40L, IFNγ)

IV.2.4. CMI Analysis

The first CMI analysis was based on the Total Vaccinated cohort (N=40 subjects for FluAS03 group and N=18 subjects for Fluarix group).

A longitudinal analysis was based on the Kinetic cohort of the Explo-Flu-001 (split protein) and Explo-Flu-002 (pool flu antigen) studies:

- Pre: N=36 subjects for FluAS03 group and N=15 for Fluarix group.
- Post-Pre: N=34 subjects for FluAS03 group and N=15 for Fluarix group.

(a) The frequency of CD4/CD8 T-lymphocytes secreting in response was summarised by descriptive statistics for each antigen, for each cytokine, for each vaccine group and at each timepoint (pre- and post-vaccination).

(b) Descriptive statistics in individual difference between timepoints (Post-Pre) responses were tabulated for each antigen, for each cytokine and for each vaccine group.

(c) For the timepoints post and (post-pre) vaccination, non-parametric Wilcoxon's test was used to compare the location differences between the two vaccine groups and to calculate the statistical p-value regarding the 4 different cytokines on:

- CD4 T-cell response to New Caledonia, Wyoming, Jiangsu and the pool of the 3 strains.
- CD8 T-cell response to New Caledonia, Wyoming, Jiangsu and the pool of the 3 strains.

(d) Non-parametric test (Wilcoxon-test) was also used:

- To investigate the kinetic of the immune response at Pre (Day 0) in term of frequency of specific CD4 between Explo-Flu-001 and Explo-Flu-002 in each vaccine group
- To investigate the kinetic of the immune response at Pre (Day 0) in term of frequency of specific CD4 between the 2 vaccine groups in each of the study Explo-Flu-001 and Explo-Flu-002
- To investigate the kinetic of the immune response in term of differences (Post-Pre) of frequency of specific CD4 between Explo-Flu-001 and Explo-Flu-002 in each vaccine group.
- To investigate the kinetic of the immune response in term of differences (Post-Pre) of frequency of specific CD4 between the 2 vaccine groups in each of the study Explo-Flu-001 and Explo-Flu-002

All significance tests were two-tailed. P-values less than or equal to 0.05 were considered as statistically significant.

IV.3. Results

Results were expressed as a frequency of cytokine(s)-positive CD4 or CD8 T cell within the CD4 or CD8 T cell sub-population.

IV.3.1. Antigen specific CD4 T- lymphocytes

The frequency of antigen-specific CD4 T-lymphocytes secreting in response was summarised by descriptive statistics for each antigen, for each cytokine, for each vaccine group and at each timepoint (pre- and post-vaccination).

Descriptive statistics in individual difference between time points (Post-Pre) in CD4 T-lymphocytes responses for each antigen at each 5 different cytokines and for each vaccine group are shown in Table 22.

Table 22 **Descriptive Statistics on difference between Post-vaccination (at Day 21) and Prevaccination (at Day 0) for the antigen-specific CD4 T-lymphocyte responses (Total vaccinated cohort)**

Antigen	Cytokine	Vaccine Group	N	Mean	SD	Min	Q1	Median	Q3	Max
Pool Flu	All double	Fluarix	18	1268.67	1051.744	197.00	724.00	863.00	1561.00	4676.00
		Flu AS03	36	1781.31	1484.860	-2379.00	929.50	1664.50	2821.00	4669.00
	CD40L	Fluarix	18	1260.11	1054.487	243.00	721.00	849.00	1602.00	4743.00
		Flu AS03	36	1711.56	1433.113	-2359.00	838.00	1576.00	2759.50	4575.00
	IFNγ	Fluarix	18	762.94	813.884	-12.00	294.00	496.00	1061.00	3564.00
		Flu AS03	36	1179.92	881.255	-817.00	692.50	1180.50	1865.50	2831.00
	IL2	Fluarix	18	1019.06	917.905	-258.00	544.00	702.00	1174.00	3850.00
		Flu AS03	36	1423.33	1359.471	-2702.00	651.00	1260.00	2200.50	4342.00
	TNFα	Fluarix	18	803.39	915.838	32.00	231.00	533.00	936.00	3892.00
		Flu AS03	36	1078.28	1029.122	-1816.00	446.00	983.00	1836.00	3310.00
A/New Caledonia	All double	Fluarix	18	481.44	381.534	-241.00	282.00	448.50	598.00	1412.00
		Flu AS03	36	812.78	749.192	-828.00	215.50	911.50	1274.50	3206.00
	CD40L	Fluarix	18	450.78	360.378	-239.00	291.00	447.00	580.00	1248.00
		Flu AS03	36	783.75	711.608	-760.00	242.00	808.00	1161.00	3050.00
	IFNγ	Fluarix	18	316.28	279.662	-165.00	175.00	259.00	387.00	1111.00
		Flu AS03	36	438.22	420.770	-685.00	125.00	393.00	733.50	1557.00
	IL2	Fluarix	18	326.06	290.792	-294.00	193.00	330.00	488.00	834.00
		Flu AS03	36	634.72	616.478	-557.00	179.50	678.50	952.00	2602.00
	TNFα	Fluarix	18	316.44	372.492	-140.00	50.00	278.00	542.00	1449.00
		Flu AS03	36	449.17	591.796	-916.00	100.50	343.50	848.00	2452.00
A/Wyoming	All double	Fluarix	18	609.56	559.396	-176.00	257.00	510.50	957.00	1998.00
		Flu AS03	36	766.61	579.191	-568.00	316.00	864.50	1221.00	1662.00
	CD40L	Fluarix	18	616.33	550.853	-176.00	274.00	488.00	939.00	2017.00
		Flu AS03	36	728.61	570.316	-670.00	260.00	789.50	1216.00	1675.00
	IFNγ	Fluarix	18	407.06	424.758	-311.00	129.00	370.50	723.00	1372.00
		Flu AS03	36	526.72	443.938	-770.00	219.00	556.50	776.00	1342.00
	IL2	Fluarix	18	495.83	503.805	-187.00	88.00	540.50	801.00	1841.00
		Flu AS03	36	572.89	533.728	-789.00	220.00	602.00	882.50	1512.00
	TNFα	Fluarix	18	424.56	485.591	-260.00	110.00	359.50	461.00	1718.00
		Flu AS03	36	550.58	538.461	-765.00	269.50	543.50	905.50	1678.00
B/Jiangsu	All double	Fluarix	18	698.44	793.119	-306.00	233.00	433.00	961.00	2822.00
		Flu	36	861.42	688.852	-223.00	339.00	745.00	1325.50	2284.00

		AS03								
	CD40L	Fluarix	18	678.39	777.259	-206.00	227.00	401.50	962.00	2878.00
		Flu AS03	36	825.89	674.879	-223.00	305.00	722.00	1282.00	2337.00
	IFNγ	Fluarix	18	431.72	489.912	-95.00	191.00	272.50	382.00	1712.00
		Flu AS03	36	615.94	473.543	-286.00	288.50	501.50	897.50	1740.00
	IL2	Fluarix	18	552.50	666.853	-234.00	155.00	278.50	833.00	2386.00
		Flu AS03	36	696.19	622.931	-359.00	207.50	540.50	1146.50	2182.00
	TNFα	Fluarix	18	441.39	695.792	-338.00	97.00	269.50	564.00	2440.00
		Flu AS03	36	500.03	448.636	-166.00	107.50	436.00	745.00	1626.00

SD = Standard Deviation
Min, Max = Minimum, Maximum
Q1 = First quartile
Q3 = Third quartile
N= number of subjects tested with available results

Vaccine-induced CD4 T-cells are shown to be able to persist at least for one year since there is an observable difference in prevaccination levels of CD4 T-cell responses between individuals vaccinated with Fluarix has compared to those vaccinated with Fluarix/AS03 the year before. The results are also shown in Figure 8, showing the CD4 T-cell response to split Flu antigen before and after revaccination. D0 corresponds to 12 months after first year vaccination and thus indicates persistence.

Comparing the difference in the frequency of antigen-specific CD4 T-lymphocytes between the 2 groups by Wilcoxon test at post-vaccination, almost all p-values were less than 0.05 and were considered as statistically significant (see Table 23) in favour of the FluAS03 group.

Table 23 Inferential statistics: p-values from Wilcoxon rank-sum test between the two vaccine groups at Day 21 for antigen-specific CD4 T-lymphocyte responses (Total vaccinated cohort)

Cytokine	P-value			
	Pool	New Caledonia	Wyoming	Jiangsu
All double	0.0014	0.0023	0.0286	0.0133
CD40L	0.0016	0.0014	0.0427	0.0155
INFγ	0.0006	0.0366	0.0400	0.0041
IL2	0.0037	0.0024	0.0584	0.0162
TNFα	0.0031	0.0103	0.0918	0.0114

P-value: Wilcoxon Test (Non-parametric procedure) to test location difference (Wilcoxon rank-sum test) between the 2 groups at Day 21.

Comparing the difference of the individual difference (Post-Pre) in the frequency of antigen-specific CD4-T-lymphocytes responses between the 2 groups by Wilcoxon test, p-values less than 0.05 and considered as statistically significant occurred for the following antigen-cytokine combinations: pool flu–all double, pool flu–IFNγ and Jiangsu-IFNγ in favour of the FluAS03 group (Table 24).

Table 24 **Inferential statistics: p-values calculated by Wilcoxon rank-sum test between the different groups on the difference between Post-vaccination (at Day 21) and Prevaccination (at 0) for the antigen-specific CD4 T-lymphocyte responses (Total vaccinated cohort)**

Cytokine	P-value			
	Pool	New Caledonia	Wyoming	Jiangsu
All double	0.0435	0.1124	0.2189	0.3085
CD40L	0.0638	0.0781	0.2831	0.2872
INFγ	0.0290	0.3589	0.2553	0.0435
IL2	0.1024	0.0563	0.3986	0.0435
TNFα	0.0693	0.4090	0.1232	0.3129

P-value: Wilcoxon Test (Non-parametric procedure) to test location difference (Wilcoxon rank-sum test) between the 2 groups.

IV.3.2. Antigen specific CD8 T- lymphocytes

The frequency of antigen-specific CD8 T-lymphocytes secreting in response was summarised by descriptive statistics for each antigen, for each cytokine, for each vaccine group and at each timepoint (pre- and post-vaccination), similarly to the procedure followed in respect of CD4 T cell response.

Comparing the difference in the frequency of antigen-specific CD8 T-lymphocytes between the 2 groups by Wilcoxon test at post-vaccination, all p-values were higher than 0.05 and were not considered as statistically significant. Comparing the difference of the individual difference (Post-Pre) in the frequency of antigen-specific CD8-T-lymphocytes responses between the 2 groups by Wilcoxon test, all p-values were higher than 0.05 and were not considered as statistically significant.

IV.3.3. Kinetic analysis : immune response at Prevaccination (one year after the first vaccination in 2003)

The frequency of antigen-specific CD4 T-lymphocytes secreting in response at prevaccination was summarised by descriptive statistics for each cytokine and for each vaccine group and for each of the two studies in Table 25, for each of the two studies

study and for each vaccine group in Table 27. Inferential statistics are given in Table 26 and Table 28.

Table 25 Descriptive Statistics on prevaccination (Day 0) for the specific CD4 T-lymphocytes response vaccination (Kinetic)

Cyto-kine	Group	Study	N	Mean	SD	Min	Q1	Median	Q3	Max
All double	Flu AS03	EXPLO 001	36	2000.86	1783.474	102.00	911.50	1461.50	2791.00	9514.00
		EXPLO 002	36	2028.28	1427.000	55.00	1190.50	1647.50	2575.00	7214.00
	Fluarix	EXPLO 001	15	2152.87	2162.463	747.00	930.00	1354.00	2101.00	7868.00
		EXPLO 002	15	1587.07	2123.841	192.00	468.00	735.00	1578.00	8536.00
CD40L	Flu AS03	EXPLO 001	35	1946.66	1771.102	120.00	837.00	1340.00	2819.00	9462.00
		EXPLO 002	35	1992.20	1440.721	77.00	1125.00	1590.00	2587.00	7286.00
	Fluarix	EXPLO 001	15	2094.93	2076.632	745.00	902.00	1340.00	2077.00	7385.00
		EXPLO 002	15	1561.73	2097.201	34.00	475.00	672.00	1579.00	8428.00
INFγ	Flu AS03	EXPLO 001	35	1068.63	1030.745	91.00	448.00	790.00	1503.00	5425.00
		EXPLO 002	35	1259.23	890.590	312.00	725.00	984.00	1354.00	4146.00
	Fluarix	EXPLO 001	15	1248.07	1452.459	320.00	388.00	778.00	1227.00	5431.00
		EXPLO 002	15	974.80	1394.044	52.00	252.00	337.00	1057.00	5576.00
IL2	Flu AS03	EXPLO 001	35	1690.20	1524.689	37.00	688.00	1211.00	2416.00	8235.00
		EXPLO 002	35	1883.60	1361.337	14.00	1068.00	1413.00	2370.00	6891.00
	Fluarix	EXPLO 001	15	1888.40	2085.857	568.00	715.00	1136.00	1770.00	7403.00
		EXPLO 002	15	1493.93	2037.139	58.00	444.00	755.00	1485.00	8193.00
TNFα	Flu AS03	EXPLO 001	35	1174.74	1119.633	55.00	466.00	795.00	1720.00	5415.00
		EXPLO 002	35	1545.40	1159.490	135.00	831.00	1203.00	1857.00	5354.00
	Fluarix	EXPLO 001	15	1444.20	1946.211	201.00	520.00	688.00	1254.00	7213.00
		EXPLO 002	15	1304.73	1759.716	144.00	316.00	824.00	1171.00	7056.00

SD = Standard Deviation
Min, Max = Minimum, Maximum
Q1 = First quartile
Q3 = Third quartile
N= number of subjects tested with available results

Comparing the difference in the frequency of antigen-specific CD4 T-lymphocytes between the 2 studies by Wilcoxon test for each vaccine group, p-values less than 0.05 and considered as statistically significant (in favour of Explo-Flu-002) occurred only for FluAS03 group and with TNFα cytokine (see Table 26).

Table 26 Inferential statistics: p-values from Wilcoxon rank-sum test between the different studies at Day 0 for antigen-specific CD4 T-lymphocyte responses (Kinetic)

Cytokine	Group	p-value
ALL	FluAS03	0.5209
DOUBLE	Fluarix	0.0712

CD40L	FluAS03	0.4957
	Fluarix	0.0744
INFγ	FluAS03	0.0896
	Fluarix	0.1103
IL2	FluAS03	0.1903
	Fluarix	0.1647
TNFα	FluAS03	0.0427
	Fluarix	0.5476

P-value: Wilcoxon Test (Non-parametric procedure) to test location difference (Wilcoxon rank-sum test) between the 2 groups at Day 21.

Table 27 Descriptive Statistics on Prevaccination (Day 0) for the specific CD4 T-lymphocytes response vaccination (Kinetic)

Cyto-kine	Study	Group	N	Mean	SD	Min	Q1	Median	Q3	Max
All double	EXPLO 001	Flu AS03	36	2000.86	1783.474	102.00	911.50	1461.50	2791.00	9514.00
		Fluarix	15	2152.87	2162.463	747.00	930.00	1354.00	2101.00	7868.00
	EXPLO 002	Flu AS03	36	2028.28	1427.000	55.00	1190.50	1647.50	2575.00	7214.00
		Fluarix	15	1587.07	2123.841	192.00	468.00	735.00	1578.00	8536.00
CD40L	EXPLO 001	Flu AS03	35	1946.66	1771.102	120.00	837.00	1340.00	2819.00	9462.00
		Fluarix	15	2094.93	2076.632	745.00	902.00	1340.00	2077.00	7385.00
	EXPLO 002	Flu AS03	35	1992.20	1440.721	77.00	1125.00	1590.00	2587.00	7286.00
		Fluarix	15	1561.73	2097.201	34.00	475.00	672.00	1579.00	8428.00
INFγ	EXPLO 001	Flu AS03	35	1068.63	1030.745	91.00	448.00	790.00	1503.00	5425.00
		Fluarix	15	1248.07	1452.459	320.00	388.00	778.00	1227.00	5431.00
	EXPLO 002	Flu AS03	35	1259.23	890.590	312.00	725.00	984.00	1354.00	4146.00
		Fluarix	15	974.80	1394.044	52.00	252.00	337.00	1057.00	5576.00
IL2	EXPLO 001	Flu AS03	35	1690.20	1524.689	37.00	688.00	1211.00	2416.00	8235.00
		Fluarix	15	1888.40	2085.857	568.00	715.00	1136.00	1770.00	7403.00
	EXPLO 002	Flu AS03	35	1883.60	1361.337	14.00	1068.00	1413.00	2370.00	6891.00
		Fluarix	15	1493.93	2037.139	58.00	444.00	755.00	1485.00	8193.00
TNFα	EXPLO 001	Flu AS03	35	1174.74	1119.633	55.00	466.00	795.00	1720.00	5415.00
		Fluarix	15	1444.20	1946.211	201.00	520.00	688.00	1254.00	7213.00
	EXPLO 002	Flu AS03	35	1545.40	1159.490	135.00	831.00	1203.00	1857.00	5354.00
		Fluarix	15	1304.73	1759.716	144.00	316.00	824.00	1171.00	7056.00

SD = Standard Deviation
Min, Max = Minimum, Maximum
Q1 = First quartile
Q3 = Third quartile
N= number of subjects tested with available results

Comparing the difference in the frequency of antigen-specific CD4 T-lymphocytes between the 2 vaccine groups by Wilcoxon test for each study, all p-values for Explo-Flu-

002 were less than 0.05 and were considered as statistically significant (in favour of FluAS03) (see Table 28).

Table 28 Inferential statistics: p-values from Wilcoxon rank-sum test between the different groups at Day 21 for antigen-specific CD4 T-lymphocyte responses (Kinetic)

Cytokine	Study	p-value
ALL DOUBLE	Explo Flu 001	0.9423
	Explo Flu 002	0.0300
CD40L	Explo Flu 001	0.8989
	Explo Flu 002	0.0361
INFγ	Explo Flu 001	0.8738
	Explo Flu 002	0.0121
IL2	Explo Flu 001	0.9747
	Explo Flu 002	0.0216
TNFα	Explo Flu 001	0.9916
	Explo Flu 002	0.0514

P-value: Wilcoxon Test (Non-parametric procedure) to test location difference (Wilcoxon rank-sum test) between the 2 groups at Day 21.

IV.3.4. Kinetic analysis : immune response at Post minus Prevaccination

The frequency of antigen-specific CD4 T-lymphocytes secreting in response at (post-pre) timepoint was summarised by descriptive statistics for each cytokine and for each vaccine group and for each study in Table 29, for each study and for each vaccine group in Table 31. Inferential statistics are given in Table 30 and Table 32.

Table 29 Descriptive Statistics on the difference between Post-vaccination (Day 21) and Prevaccination (Day 0) for the specific CD4 T-lymphocytes response vaccination (Kinetic)

Cyto-kine	Group	Study	N	Mean	SD	Min	Q1	Median	Q3	Max
All double	Flu AS03	EXPLO 001	34	4837.56	4476.129	-609.00	1888.00	3483.50	8148.00	19555.00
		EXPLO 002	34	1737.79	1450.177	-2379.00	936.00	1664.50	2743.00	4669.00
	Fluarix	EXPLO 001	15	3103.53	3726.645	436.00	800.00	2283.00	3226.00	15169.00
		EXPLO 002	15	1369.00	1127.784	197.00	725.00	869.00	1808.00	4676.00
CD40L	Flu AS03	EXPLO 001	33	4819.06	4489.788	-718.00	1799.00	3479.00	8288.00	19480.00
		EXPLO 002	33	1694.73	1431.082	-2359.00	921.00	1659.00	2662.00	4575.00
	Fluarix	EXPLO 001	15	3090.00	3684.759	477.00	822.00	2189.00	3208.00	15021.00
		EXPLO 002	15	1360.93	1131.051	243.00	725.00	860.00	1687.00	4743.00
IFNγ	Flu AS03	EXPLO 001	33	3127.09	2974.067	-453.00	1325.00	1721.00	5162.00	13296.00
		EXPLO 002	33	1167.85	893.363	-817.00	633.00	1207.00	1803.00	2831.00
	Fluarix	EXPLO 001	15	1660.13	1834.023	-84.00	480.00	1386.00	2284.00	7120.00
		EXPLO 002	15	851.87	859.585	148.00	294.00	501.00	1222.00	3564.00
IL2	Flu AS03	EXPLO 001	33	3950.18	3878.538	-358.00	1309.00	2780.00	6635.00	16988.00
		EXPLO 002	33	1404.67	1355.665	-2702.00	719.00	1341.00	2109.00	4342.00

	Fluarix	EXPLO 001	15	2413.87	3027.392	263.00	674.00	1672.00	2425.00	12273.00
		EXPLO 002	15	1117.80	975.934	-258.00	575.00	714.00	1618.00	3850.00
TNFα	Flu AS03	EXPLO 001	33	2627.36	2574.458	-825.00	862.00	1475.00	4764.00	9267.00
		EXPLO 002	33	1072.36	1044.140	-1816.00	447.00	1000.00	1752.00	3310.00
	Fluarix	EXPLO 001	15	1460.53	3115.174	-1586.00	251.00	813.00	1314.00	12275.00
		EXPLO 002	15	904.67	974.958	32.00	338.00	752.00	965.00	3892.00

SD = Standard Deviation
Min, Max = Minimum, Maximum
Q1 = First quartile
Q3 = Third quartile
N= number of subjects tested with available results

Comparing the difference in the frequency of antigen-specific CD4 T-lymphocytes between the 2 studies by Wilcoxon test for each vaccine group, all p-values for FluAS03 group were less than 0.05 and were considered as statistically significant (in favour of Explo-Flu-001) (see Table 30).

Table 30 Inferential statistics on the difference between Post-vaccination (Day 21) and Prevaccination (Day 0): p-values from Wilcoxon rank-sum test between the different studies at Day 21 for antigen-specific CD4 T-lymphocyte responses (Kinetic)

Cytokine	Group	p-value
ALL DOUBLE	FluAS03	0.0005
	Fluarix	0.1300
CD40L	FluAS03	0.0007
	Fluarix	0.0890
INFγ	FluAS03	0.0012
	Fluarix	0.1103
IL2	FluAS03	0.0025
	Fluarix	0.1409
TNFα	FluAS03	0.0327
	Fluarix	0.6936

P-value: Wilcoxon Test (Non-parametric procedure) to test location difference (Wilcoxon rank-sum test) between the 2 groups at Day 21.

Table 31 Descriptive Statistics on the difference between Post-vaccination (Day 21) and Prevaccination (Day 0) for the specific CD4 T-lymphocytes response vaccination (Kinetic)

Cyto-kine	Study	Group	N	Mean	SD	Min	Q1	Median	Q3	Max
All double	EXPLO 001	Flu AS03	34	4837.56	4476.129	-609.00	1888.00	3483.50	8148.00	19555.00
		Fluarix	15	3103.53	3726.645	436.00	800.00	2283.00	3226.00	15169.00
	EXPLO 002	Flu AS03	34	1737.79	1450.177	-2379.00	936.00	1664.50	2743.00	4669.00
		Fluarix	15	1369.00	1127.784	197.00	725.00	869.00	1808.00	4676.00
CD40L	EXPLO 001	Flu AS03	33	4819.06	4489.788	-718.00	1799.00	3479.00	8288.00	19480.00
		Fluarix	15	3090.00	3684.759	477.00	822.00	2189.00	3208.00	15021.00

	EXPLO 002	Flu AS03	33	1694.73	1431.082	-2359.00	921.00	1659.00	2662.00	4575.00
		Fluarix	15	1360.93	1131.051	243.00	725.00	860.00	1687.00	4743.00
IFNγ	EXPLO 001	Flu AS03	33	3127.09	2974.067	-453.00	1325.00	1721.00	5162.00	13296.00
		Fluarix	15	1660.13	1834.023	-84.00	480.00	1386.00	2284.00	7120.00
	EXPLO 002	Flu AS03	33	1167.85	893.363	-817.00	633.00	1207.00	1803.00	2831.00
		Fluarix	15	851.87	859.585	148.00	294.00	501.00	1222.00	3564.00
IL2	EXPLO 001	Flu AS03	33	3950.18	3878.538	-358.00	1309.00	2780.00	6635.00	16988.00
		Fluarix	15	2413.87	3027.392	263.00	674.00	1672.00	2425.00	12273.00
	EXPLO 002	Flu AS03	33	1404.67	1355.665	-2702.00	719.00	1341.00	2109.00	4342.00
		Fluarix	15	1117.80	975.934	-258.00	575.00	714.00	1618.00	3850.00
TFNα	EXPLO 001	Flu AS03	33	2627.36	2574.458	-825.00	862.00	1475.00	4764.00	9267.00
		Fluarix	15	1460.53	3115.174	-1586.00	251.00	813.00	1314.00	12275.00
	EXPLO 002	Flu AS03	33	1072.36	1044.140	-1816.00	447.00	1000.00	1752.00	3310.00
		Fluarix	15	904.67	974.958	32.00	338.00	752.00	965.00	3892.00

SD = Standard Deviation
Min, Max = Minimum, Maximum
Q1 = First quartile
Q3 = Third quartile
N= number of subjects tested with available results

Comparing the difference in the frequency of antigen-specific CD4 T-lymphocytes between the 2 vaccine groups by Wilcoxon test for each study, p-value was less than 0.05 only for Explo-Flu-001 and was considered as statistically significant (in favour of FluAS03) (see Table 32).

Table 32 Inferential statistics: p-values from Wilcoxon rank-sum test between the different groups at Day 21 for antigen-specific CD4 T-lymphocyte responses (Kinetic)

Cytokine	Study	p-value
ALL DOUBLE	Explo Flu 001	0.0827
	Explo Flu 002	0.0992
CD40L	Explo Flu 001	0.0931
	Explo Flu 002	0.1391
INFγ	Explo Flu 001	0.0543
	Explo Flu 002	0.1068
IL2	Explo Flu 001	0.0847
	Explo Flu 002	0.2254
TNFα	Explo Flu 001	0.0375
	Explo Flu 002	0.2009

P-value: Wilcoxon Test (Non-parametric procedure) to test location difference (Wilcoxon rank-sum test) between the 2 groups at Day 21.

IV.4. HI titers

Results are shown in Figure 9 and in Tables 33 to 36.

Table 33: Geometric Mean Titers (GMT) and seropositivity rates of anti-HI titers (GMTs calculated on vaccinated subjects)

Antibody	*Group*	Timing	N	S+	%	95% CI		GMT	95%CI	
						L.L.	U.L.		L.L.	U.L.
New Caledonia	Fluarix	PRE	18	17	94.4	72.6	99.9	63.5	38.1	105.9
		PI(D21)	18	18	100	81.5	100	131.9	77.1	225.6
	FluAS03	PRE	40	39	97.5	86.8	99.9	70.3	50.5	97.7
		PI(D21)	40	40	100	91.3	100	218.6	158.2	302.0
A/Fujian	Fluarix	PRE	18	18	100	81.5	100	95.0	51.0	176.9
		PI(D21)	18	18	100	81.5	100	498.3	272.1	912.7
	FluAS03	PRE	40	40	100	91.3	100	94.3	71.4	124.6
		PI(D21)	40	40	100	91.3	100	735.1	564.4	957.5
B/Shanghai	Fluarix	PRE	18	16	88.9	65.3	98.6	23.3	15.2	35.8
		PI(D21)	18	17	94.4	72.6	99.9	139.8	64.0	305.0
	FluAS03	PRE	40	38	95.0	83.1	99.4	58.6	43.9	78.1
		PI(D21)	40	40	100	91.3	100	364.4	269.7	492.4

PRE= Prevaccination,
PI(D21)= day 21 post vaccination
95%CI, LL, and UL = 95% confidence interval, lower and upper limit
S+= number of seropositive subjects

Table 34: Conversion factor of anti-HI titers (All vaccinated subjects)

Group	A/N-Caledonia		A/Fujian		B/Shanghai	
	N	GMR [95 % CI]	N	GMR [95 % CI]	N	GMR [95 % CI]
Fluarix	18	2.1 [1.4;3.2]	18	5.2 [3.0;9.3]	18	6.0 [3.5;10.2]
FluAS03	40	3.1 [2.4;4.0]	40	7.8 [5.6;10.9]	40	6.2 [4.7;8.2]

N = total number of subjects
GMR = Geometric Mean Ratio (antilog of the mean log day 21/day 0 titers ratios)
95%CI = 95% confidence interval

Table 35: Seroprotection rates of anti-HI titers (All vaccinated subjects)

Antibody	Group	Timing	N	>=40			
				n	%	95 % CI	
A/New Caledonia	Fluarix	PRE	18	14	77.8	52.4	93.6
		PI(D21)	18	16	88.9	65.3	98.6
	FluAS03	PRE	40	32	80	64.4	90.9
		PI(D21)	40	39	97.5	86.8	99.9
A/Fujian	Fluarix	PRE	18	14	77.8	52.4	93.6
		PI(D21)	18	18	100	81.5	100
	FluAS03	PRE	40	36	90	76.3	97.2
		PI(D21)	40	40	100	91.2	100
B/Shanghai	Fluarix	PRE	18	6	33.3	13.3	59.0

		PI(D21)	18	14	77.8	52.4	93.6
	FluAS03	PRE	40	34	85	70.2	94.3
		PI(D21)	40	40	100	91.2	100

PRE= Prevaccination,
PI(D21)= day 21 post vaccination
N = number of subjects with available results.
n = number of subjects with titres within the specified range.
% = percentage of subjects with titres within the specified range

Table 36: Seroconversion rates at PI day 21(fold-increase = 4) (All vaccinated subjects)

Antibody	**Vaccine Group**	**N**	**Responders**			
			n	**%**	**95% CI**	
					LL	**UL**
A/New Caledonia	Fluarix	18	3	16.7	3.6	41.5
	FluAS03	40	19	47.5	31.5	63.9
A/Fujian	Fluarix	18	13	72.2	46.5	90.3
	FluAS03	40	34	85.0	70.2	94.3
B/Shanghai	Fluarix	18	12	66.7	41.0	86.7
	FluAS03	40	31	77.5	61.5	89.2

N = number of subjects with both pre and post vaccination result available.
n = number of responders.
% = Proportion of responders (n/N x 100).
95% CI = exact 95% confidence interval; LL = lower limit, UL = upper limit

IV.5. Overall conclusions

From this clinical study it is confirmed that the adjuvanted vaccine Flu-AS03 is superior to the equivalent unadjuvated vaccine Fluarix in terms of frequency of influenza specific CD4 T cells, and also in terms of persistence of the immune response elicited by the first Flu-AS03 vaccination (primo-vaccination in Explo Flu 001) until D0 of the revaccination study (Explo Flu 002 i.e. +/- 1 year later). Furthermore this response is capable to recognise drifted influenza strains present in the new vaccine and to recognise the strains of the 2004 influenza vaccine.

In contrast to first year vaccination, upon revaccination individuals previously vaccinated with the adjuvanted Fluarix™ showed increased HI titer responsiveness as compared to those vaccinated with un-adjuvanted Fluarix™. There is an observable trend for 1.5- to 2-fold increase in HI titer directed against H1N1 and H3N2 strains and a demonstrated statistical increase in HI titer directed against B strain.

Example V – Pre-clinical evaluation of adjuvanted and unadjuvanted influenza vaccines in ferrets

FIRST STUDY - Efficacy of new formulations AS03 and AS03+MPL

V.1. Rationale and objectives

Influenza infection in the ferret model closely mimics human influenza, with regards both to the sensitivity to infection and the clinical response.

The ferret is extremely sensitive to infection with both influenza A and B viruses without prior adaptation of viral strains. Therefore, it provides an excellent model system for studies of protection conferred by administered influenza vaccines.

This study investigated the efficacy of various Trivalent Split vaccines, adjuvanted or not, to reduce disease symptoms (body temperature) and viral shedding in nasal secretions of ferrets challenged with homologous strains.

The objective of this experiment was to demonstrate the efficacy of an adjuvanted influenza vaccine compared to the plain (un-adjuvanted) vaccine.

The end-points were:

1) primary end-point: Reduction of viral shedding in nasal washes after homologous challenge:

2) secondary end-points: Analysis of the humoral reponse by IHA and monitoring of the temperature around the priming and the challenge.

V.2. Experimental design

V.2.1. Treatment/group (Table 37)

Female ferrets (*Mustela putorius furo*) (6 ferrets/group) aged 14-20 weeks were obtained from MISAY Consultancy (Hampshire, UK). Ferrets were primed on day 0 with heterosubtypic strain H1N1 A/Stockholm/24/90 (4 Log $TCID_{50}$/ml). On day 21, ferrets were injected intramuscularly with a full human dose (500 µg vaccine dose, 15 µg HA/strain) of a combination of H1N1 A/New Caledonia/20/99, H3N2 A/Panama/2007/99 and B/Shangdong/7/97. Ferrets were then challenged on day 41 by intranasal route with an homotypic strain H3N2 A/Panama/2007/99 (4.51 Log $TCID_{50}$/ml).

Table 37

Group	Antigen(s) + dosage	Formulation + dosage	Comments (schedule/route /challenge)	Other treatments
1	Trivalent	Full HD: 15µg	IM; Day 21	Priming H1N1

	Plain	HA/strain		(A/Stockolm/24 /90) Day 0
2	Trivalent AS03	Full HD: 15µg HA/strain	IM; Day 21	Priming H1N1 (A/Stockolm/24/90) Day 0
3	Trivalent AS03+MPL	Full HD: 15µg HA/strain	IM; Day 21	Priming H1N1 (A/Stockolm/24 /90) Day 0
4	PBS		IM; Day 21	Priming H1N1 (A/Stockolm/24 /90) Day 0

V.2.2. Preparation of the vaccine formulations

Formulation 1: Trivalent plain (un-adjuvanted) formulation (500µl):

PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a mixture containing Tween 80, Triton X-100 and VES (quantities taking into account the detergents present in the strains) are added to water for injection. The detergents quantities reached are the following: 750µg Tween 80, 110µg Triton X-100 and 100µg VES per 1 ml. After 5 min stirring, 15µg of each strain H1N1, H3N2 and 17.5µg of B strain are added in sequence with 10 min stirring between each addition. The formulation is stirred for 15 minutes at room temperature and stored at 4°C if not administered directly.

Formulation 2: Trivalent split influenza adjuvanted with AS03 (500µl):

PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a mixtutre containing Tween 80, Triton X-100 and VES (quantities taking into account the detergents present in the strains) is added to water for injection. The detergents quantities reached are the following: 750µg Tween 80, 110µg Triton X-100 and 100µg VES per 1 ml. After 5 min stirring, 15µg of each strain H1N1, H3N2 and 17.5µg of B strain are added with 10 min stirring between each addition. After 15 min stirring, 250µl of SB62 emulsion (prepared as in taught in Example II.1) is added. The formulation is stirred for 15 minutes at room temperature and stored at 4°C if not administered directly.

Formulation 3: Trivalent split influenza adjuvanted with AS03+MPL

PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a mixtutre containing Tween 80, Triton X-100 and VES (quantities taking into account the detergents present in the strains) is added to water for injection. The detergents quantities reached are the following: 750µg Tween 80, 110µg Triton X-100 and 100µg VES per 1 ml. After 5 min stirring, 15µg of each strain H1N1, H3N2 and 17.5µg of B strain are added with 10 min stirring between each addition. After 15 min stirring, 250µl of SB62 emulsion

(prepared as in taught in Example II.1) is added. The mixture is stirred again for 15 min just prior addition of 25µg of MPL from a suspension prepared as detailed in Example II.3.1. The formulation is stirred for 15 minutes at room temperature and stored at 4°C if not administered directly.

Remark: In each formulation, PBS 10 fold concentrated is added to reach isotonicity and is 1 fold concentrated in the final volume. H2O volume is calculated to reach the targeted volume.

V.2.3. Read-outs (Table 38)

Table 38

Readout	Timepoint	Sample-type	I/P	Analysis method
Viral shedding	D-1 to D+7 Post priming D-1 to D+5 Post challenge	Nasal washes	In	Titration
T° monitoring	D-1 to D+3 Post priming D-2 to D+3 Post challenge	Implant in peritoneal cavity	In	Telemetry
IHA	Pre, Post priming, Post imm, Post challenge	Serum	In	IHA

In= Individual / Po= Pool

V.3. Results

A schematic representation of the results is given in Figure 10 and Figure 11.

V.3.1. Temperature monitoring

Individual temperature were monitored with the transmitters and by the telemetry recording (according to the procedure detailed under I.2.2). All implants were checked and refurbished and a new calibration was performed by DSI before placement in the intraperitoneal cavity. All animals were individually housed in single cage during these measurements.

Temperature were monitored from 3 days Pre-challenge until 5 days Post challenge every 15 minutes and an average has been calculated by mid-day. Results from baseline to baseline body temperature are shown in Figures 10A (results from -1 to +3 days are shown) and 10B (results from -2 to +3 days are shown).

Post-challenge, a peak of body temperature only observed after immunization with trivalent split plain or PBS. No peak observed after immunization with trivalent split adjuvanted with AS03 or AS03+MPL.

V.3.2. Viral shedding (Figure 11)

Viral titration of nasal washes was performed on 6 animals per group.

The nasal washes were performed by administration of 5 ml of PBS in both nostrils in awake animals. The inoculation was collected in a Petri dish and placed into sample containers at -80°C (dry ice).

All nasal samples were first sterile filtered through Spin X filters (Costar) to remove any bacterial contamination. 50 µl of serial ten-fold dilutions of nasal washes were transferred to microtiter plates containing 50 µl of medium (10 wells/dilution). 100µl of MDCK cells (2.4×10^5 cells/ml) were then added to each well and incubated at 35°C until cell confluence is reached for the control cells, e.g. for 5-7days. After 6-7 days of incubation, the culture medium is gently removed and 100 µl of a 1/20 WST-1 containing medium is added and incubated for another 18 hrs.

The intensity of the yellow formazan dye produced upon reduction of WST-1 by viable cells is proportional to the number of viable cells present in the well at the end of the viral titration assay and is quantified by measuring the absorbance of each well at the appropriate wavelength (450 nanometers). The cut-off is defined as the OD average of uninfected control cells – 0.3 OD (0.3 OD correspond to +/- 3 StDev of OD of uninfected control cells). A positive score is defined when OD is < cut-off and in contrast a negative score is defined when OD is > cut-off. Viral shedding titers were determined by "Reed and Muench" and expressed as Log TCID50/ml.

Lower viral shedding was observed Post-challenge with Trivalent Split adjuvanted with AS03 or AS03+MPL compared to Trivalent Split Plain or PBS. The protective effect was slightly better with AS03 compared to AS03+MPL (see Day 2 Post-challenge). Statistical significance could not be determined due to the low number of animals per group.

V.3.3. Conclusion of the experiment

Higher humoral responses (HI titers) were observed with Trivalent Split adjuvanted with AS03 or AS03+MPL compared to the Trivalent Split Plain for all 3 strains (at least 2-fold for 2 out of 3 strains, i.e. H3N2 and B strains).

AS03 and AS03+MPL formulations showed added benefit in terms of protective efficacy in ferrets (lower viral shedding and temperature) (Figures 10 and 11).

Post-challenge, no boost of the humoral response was observed after immunization with Trivalent Split adjuvanted with AS03 or AS03+MPL.

SECOND STUDY - Heterotypic Challenge study in ferrets: demonstration of efficacy of new formulation tested

V.4. Rationale and objectives

This study investigated the efficacy of various Trivalent Split vaccines, adjuvanted or not, by their ability to reduce disease symptoms (body temperature) and their effects on viral shedding in nasal secretions of immunized ferrets after a heterologuous challenge.

V.5. Experimental design

Female ferrets (*Mustela putorius furo*) (6 ferrets/group) aged 14-20 weeks were obtained from MISAY Consultancy (Hampshire, UK). Four groups were tested:

* Fluarix
* Trivalent Split AS03
* Trivalent Split AS03+MPL
* PBS

Ferrets were primed on day 0 with heterosubtypic strain H1N1 A/Stockholm/24/90 (4 Log $TCID_{50}$/ml). On day 21, ferrets were injected intramuscularly with a full human dose (500 µg vaccine dose, 15 µg HA/strain) of a combination of H1N1 A/New Caledonia/20/99, H3N2 A/Panama/2007/99 and B/Shangdong/7/97 (17.5 µg HA). Ferrets were then challenged on day 43 by intranasal route with an heterosubtypic strain H3N2 A/Wyoming/3/2003 (4.51 Log $TCID_{50}$/ml).

V.6. Results

A schematic representation of the results is given in Figure 12 and in Figure 13.

V.6.1. Temperature monitoring

Individual temperature were monitored with the transmitters and by the telemetry recording. All implants were checked and refurbished and a new calibration was performed by DSI before placement in the intraperitoneal cavity. All animals were individually housed in single cage during these measurements.

The results (Figure 12) show that:
- A high variability from one group to another was observed around the priming. The baseline seemed to be higher before priming than after priming.
- Despite the high variability in the body temperature, a peak was only observed Post-challenge in ferrets immunized with PBS (6/6 ferrets), Trivalent Split Plain (5/6 ferrets) and Trivalent Split adjuvanted with AS03 (2/6 ferrets). No peak was observed after immunization with trivalent split adjuvanted with AS03+MPL (0/6 ferrets).
- AS03 seemed to be less efficient than AS03+MPL against heterologous strains in terms of fever prevention. We cannot conclude the possibility that difference between adjuvant is due to different level in pre-challenge antibody levels.

V.6.2. Viral shedding (Figure 13)

The nasal washes were performed by administration of 5 ml of PBS in both nostrils in awake animals. The inoculation was collected in a Petri dish and placed into sample containers at –80°C (dry ice).

All nasal samples were first sterile filtered through Spin X filters (Costar) to remove any bacterial contamination. 50 µl of serial ten-fold dilutions of nasal washes were transferred to microtiter plates containing 50 µl of medium (10 wells/dilution). 100µl of MDCK cells (2.4×10^5 cells/ml) were then added to each well and incubated at 35°C until cell confluence is reached for the control cells, e.g. for 5-7days. After 6-7 days of incubation, the culture medium is gently removed and 100 µl of a 1/20 WST-1 containing medium is added and incubated for another 18 hrs.

The intensity of the yellow formazan dye produced upon reduction of WST-1 by viable cells is proportional to the number of viable cells present in the well at the end of the viral titration assay and is quantified by measuring the absorbance of each well at the appropriate wavelength (450 nanometers). The cut-off is defined as the OD average of uninfected control cells – 0.3 OD (0.3 OD corresponds to +/- 3 St Dev of OD of uninfected control cells). A positive score is defined when OD is < cut-off and in contrast a negative score is defined when OD is > cut-off. Viral shedding titers were determined by "Reed and Muench" and expressed as Log TCID50/ml.

Viral shedding after priming

Viral shedding was measured for 12 ferrets from Day 1 Pre-priming- to Day 7 Post-priming. Results are expressed in pool.

The viral clearance was observed on Day 7 Post-priming in all ferrets.

Viral shedding after challenge

Viral shedding was measured for 6 ferrets/group from Day 1 Pre-challenge to Day 7 Post-challenge.

Two days Post-challenge, statistically significant lower viral titers were observed in ferrets immunized with Trivalent Split adjuvanted with AS03 and AS03+MPL compared to ferrets immunized with Trivalent Split Plain and PBS (difference of 1.25/1.22 log and 1.67/1.64 log with adjuvanted groups AS03/ AS03+MPL compared to the Plain vaccine, respectively).

On Day 50, no virus was detected in nasal washes.

V.6.3. Hemagglutination Inhibition Test (HI titers) (Figures 14A and B)

Serum samples were collected 1 day before priming, 21 days Post-priming, 22 days post-immunization and 14 days post-challenge.

Anti-Hemagglutinin antibody titers to the H3N2 influenza virus (vaccine and challenge strains) were determined using the hemagglutination inhibition test (HI). The principle of the HI test is based on the ability of specific anti-Influenza antibodies to inhibit hemagglutination of chicken red blood cells (RBC) by influenza virus hemagglutinin (HA). Sera were first treated with a 25% neuraminidase solution (RDE) and were heat-inactivated to remove non-specific inhibitors. After pre-treatment, two-fold dilutions of sera were incubated with 4 hemagglutination units of each influenza strain. Chicken red blood cells were then added and the inhibition of agglutination was scored. The titers were expressed as the reciprocal of the highest dilution of serum that completely inhibited hemagglutination. As the first dilution of sera was 1:10, an undetectable level was scored as a titer equal to 5.

Results:

Results are shown in Figures 14A and 14B. After immunization with H3N2 A/Panama, higher humoral responses (HI titers) were observed in ferrets immunized with the trivalent split vaccine adjuvanted with AS03 or AS03+MPL, as compared to the humoral response observed after immunization of ferrets with the un-adjuvanted (plain) trivalent split vaccine (Fluarix™).

Similar HI titers were observed in ferrets immunized with H3N2 A/Panama adjuvanted with AS03 or AS03+MPL.

Cross-reactive HI titers to the heterologous strain A/Wyoming H3N2 was only observed after immunization with A/Panama H3N2 strain containing vaccine adjuvanted with AS03 or AS03+MPL (not observed after immunization with Trivalent Split Plain).

A boost of A/Wyoming-specific HI titers was observed in ferrets immunized with the heterologous strain A/Panama H3N2 and challenged with A/Wyoming H3N2. As expected and contrary to the homologous challenge, the heterologous challenge resulted in an increase of A/Panama-specific HI titers in ferrets immunized with A/Panama H3N2 adjuvanted with AS03 and AS03+MPL.

V.6.4. Conclusion of this experiment

As expected, a boost of anti-H3N2 HI titers was observed after heterologous challenge compared to the situation after homologous challenge (no boost).

However, similar protection (viral shedding) was observed after heterologous and homologous challenge.

Example VI – Pre-clinical evaluation of adjuvanted and unadjuvanted influenza vaccines in C57Bl/6 primed mice

VI.1. Experimental design and objective

Significant higher CD4 T cell responses were observed, in Explo-Flu-001 clinical study (see Example III), for Trivalent Flu Split AS03 compared to Fluarix Plain (un-adjuvanted). No difference was observed for both CD8 T cell and humoral responses between these two groups.

The purpose was to select readouts to induce in mice similar CMI responses than observed in humans. Particularly, the purpose was to show higher CMI responses in mice by using Split AS03 or split AS03+MPL compared to Split plain.

VI.1.1. Treatment/group

Female C57Bl/6 mice (15 mice/group) aged 6-8 weeks were obtained from Harlan Horst, Netherland. The groups tested were:

- Trivalent Split Plain
- Trivalent Split AS03
- Trivalent Split AS03+MPL

- PBS

Mice were primed on day 0 with heterosubtypic strains (5 µg HA whole inactivated H1N1 A/Johnannesburg/82/96, H3N2 A/Sydney/5/97, B/Harbin/7/94). On day 28, mice were injected intramuscularly with 1,5 µg HA Trivalent split (A/New Caledonia/20/99, A/Panama/2007/99, B/Shangdong/7/97) plain or adjuvanted (see groups below).

VI.1.2. Preparation of the vaccine formulations

In each formulation, PBS 10 fold concentrated is added to reach isotonicity and is 1 fold concentrated in the final volume. H_2O volume is calculated to reach the targeted volume.

Split trivalent plain (un-adjuvanted):

Formulation 1 (for 500 µl): PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a mixture containing Tween 80, Triton X-100 and VES (quantities taking into account the detergents present in the strains) are added to water for injection. The detergents quantities reached are the following: 750µg Tween 80, 110µg Triton X-100 and 100µg VES per 1 ml After 5 min stirring, 15µg of each strain H1N1, H3N2 and B are added with 10 min stirring between each addition. The formulation is stirred for 15 minutes at room temperature and stored at 4°C if not administered directly.

Split trivalent adjuvanted with the oil-in-water emulsion adjuvant AS03:

PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a mixture containing Tween 80, Triton X-100 and VES (quantities taking into account the detergents present in the strains) is added to water for injection. The detergents quantities reached are the following: 750µg Tween 80, 110µg Triton X-100 and 100µg VES per 1 ml. After 5 min stirring, 15µg of each strain H1N1, H3N2 and B are added with 10 min stirring between each addition. After 15 min stirring, 250µl of SB62 emulsion (prepared as taught in Example II.1) is added. The formulation is stirred for 15 minutes at room temperature and stored at 4°C if not administered directly.

Split trivalent adjuvanted with AS03+MPL:

PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a mixture containing Tween 80, Triton X-100 and VES (quantities taking into account the detergents present in the strains) is added to water for injection. The detergents quantities reached are the following: 750µg Tween 80, 110µg Triton X-100 and 100µg VES per 1 ml After 5 min stirring, 15µg of each strain H1N1, H3N2 and B are added with 10 min stirring between each addition. After 15 min stirring, 250µl of SB62 emulsion (prepared as taught in Example II.1) is added. The mixture is stirred again for 15 min just prior addition of 25µg of MPL. The formulation is stirred for 15 minutes at room temperature and stored at 4°C if not administered directly.

VI.1.3. Read-outs

CMI analysis (ICS: CD4/CD8, IL-2/IFNg staining)

PBMCs from primed mice were harvested 7 days post-immunization. They were tested in pools/group.

VI.2. Results

Conditions that showed higher frequencies of CD4 and CD8+ T cells, as well as lower background, were determined by using C57Bl/6 primed mice and whole inactivated virus 1 µg/ml as re-stimulating antigen. Results are shown in Figure 15 (CD4 T-cell responses) and in Figure 16 (CD8 T-cell response).

With these conditions, it was possible to induce:

- Higher CD4 T cell responses for Split AS03 compared to Split Plain, as observed in humans.
- Higher CD4 T cell responses for Split AS03+MPL compared to Split Plain.
- Similar CD8 T cell responses between Split Plain and Split AS03, as observed in humans.
- Trend for higher CD8 T cell responses for AS03+MPL compared to Split AS03 or Split Plain

Example VII – Pre-clinical evaluation of adjuvanted and unadjuvanted split and sub-unit influenza vaccines in C57Bl/6 mice primed with heterologous strains

VII.1. Experimental design and objective

Significant higher CD4 T cell responses were observed, in Explo-Flu-001 clinical study (see Example III), for Trivalent Flu Split AS03 compared to Fluarix Plain (un-adjuvanted). No difference was observed for both CD8 T cell and humoral responses between these two groups.

An animal model reproducing similar immune profiles than observed in humans was developed by using C57Bl/6 mice primed with heterologous strains. For ICS (intracellular cytokine staining), the re-stimulation is performed with an inactivated whole virus.

The purpose was to compare the CMI response induced by a GlaxoSmithKline commercially available split vaccine (Fluarix™) versus a subunit vaccine (Chiron's vaccine

Fluad™) as well as the CMI response obtained with these vaccines adjuvanted with AS03, or AS03+MPL or another oil-in-water emulsion adjuvant (OW).

VII.1.1. Treatment/group

Female C57Bl/6 mice (24 mice/group) aged 6-8 weeks were obtained from Harlan Horst, Netherland. Mice were primed intranasally on day 0 with heterosubtypic strains (5 µg HA whole formaldehyde inactivated H1N1 A/Johnannesburg/82/96, H3N2 A/Sydney/5/97, B/Harbin/7/94). On day 29, mice were injected intramuscularly with 1,5 µg HA Trivalent split (A/New Caledonia/20/99, A/Wyoming/3/2003, B/Jiangsu/10/2003) plain or adjuvanted (see groups in Table 39 below).

Table 39

Gr	Antigen / Formulation	Other treatment
1	Trivalent split* / Plain (un-adjuvanted) = Fluarix™	Heterologous priming D0
2	Trivalent split* / OW	Heterologous priming D0
3	Trivalent split* / AS03	Heterologous priming D0
4	Trivalent split* / AS03+MPL (2.5 µg per dose)	Heterologous priming D0
5	Gripguard (= Fluad ™) = sub-unit in an oil-in-water emulsion	Heterologous priming D0
6	Aggripal ™ (sub-unit) / AS03	Heterologous priming D0
7	Aggripal ™ (sub-unit) / AS03+MPL (2.5 µg per dose)	Heterologous priming D0
8	Aggripal ™ (sub-unit) / OW**	Heterologous priming D0
9	Aggripal ™ (sub-unit)	Heterologous priming D0
10	PBS	Heterologous priming D0

* Fluarix ™

**OW produced as explained in the section below

VII.1.2. Preparation of the vaccine formulations

Preparation of OW

An oil-in-water emulsion called OW is prepared following the recipe published in the instruction booklet contained in Chiron Behring FluAd vaccine.

Water for injection, 36.67mg of Citric acid and 627.4mg of Na Citrate.2H2O are mixed together and the volume is adjusted to 200ml. 470mg of Tween 80 is mixed with 94.47ml of this buffer and this mixture is called "solution A". The oil mixture is prepared by mixing 3.9g of squalene and 470 mg of Span 85 under magnetic stirring. Solution A is then added to the oil mixture and the final volume obtained is 100 ml. The mixture is then first passed trough a 18Gx 1 ½ needle and is then put in the M110S microfluidiser (from Microfluidics) in two samples to reduce the size of the oil dropplets. When a particle size

around 150nm is obtained for each, the 2 samples are pooled and filtrated on 0.2 µm filter. A z average mean of 143 nm with a polydispersity of 0.10 is obtained for the pooled sample at T0 and of 145 nm with a polydispersity of 0.06 after 4 months storage at 4°C. This size is obtained using the Zetasizer 3000HS (from Malvern), under the following technical conditions:

- laser wavelength: 532 nm (Zeta3000HS).
- laser power: 50 mW (Zeta3000HS).
- scattered light detected at 90° (Zeta3000HS).
- temperature: 25°C,
- duration: automatic determination by the soft,
- number: 3 consecutive measurements,
- z-average diameter: by cumulants analysis

Formulation for group 1 (for 1 ml):

PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a mixture containing Tween 80, Triton X-100 and VES (quantities taking into account the detergents present in the strains) to reach a final concentration of 375 µg/ml Tween 80, 55 µg/ml Triton X-100 and 50 µg/ml VES, are added to water for injection. After 5 min stirring, 15µg of each strain H1N1, H3N2 and B are added with 10 min stirring between each addition. The formulation is stirred for 15 minutes and stored at 4°C if not administered directly.

Formulation for group 2 (for 1 ml):

PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a mixture containing Tween 80, Triton X-100 and VES (quantities taking into account the detergents present in the strains) to reach a final concentration of 375 µg/ml Tween 80, 55 µg/ml Triton X-100 and 50 µg/ml VES, is added to water for injection. After 5 min stirring, 15µg of each strain H1N1, H3N2 and B are added with 10 min stirring between each addition. After 15 min stirring, 250µl of OW emulsion is added. The formulation is stirred for 15 minutes and stored at 4°C if not administered directly.

Formulation for group 3: for 1 ml:

PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a mixture containing Tween 80, Triton X-100 and VES (quantities taking into account the detergents present in the strains) to reach a final concentration of 375 µg/ml Tween 80, 55 µg/ml Triton X-100 and 50 µg/ml VES, is added to water for injection. After 5 min stirring, 15µg of each strain H1N1, H3N2 and B are added with 10 min stirring between each addition.

After 15 min stirring, 250µl of SB62 emulsion is added. The formulation is stirred for 15 minutes and stored at 4°C if not administered directly.

Formulation for group 4: for 1 ml:

PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a mixture containing Tween 80, Triton X-100 and VES (quantities taking into account the detergents present in the strains) to reach a final concentration of 375 µg/ml Tween 80, 55 µg/ml Triton X-100 and 50 µg/ml VES, is added to water for injection. After 5 min stirring, 15µg of each strain H1N1, H3N2 and B are added with 10 min stirring between each addition. After 15 min stirring, 250µl of SB62 emulsion is added. The mixture is stirred again for 15 min just prior addition of 25µg of MPL. The formulation is stirred for 15 minutes and stored at 4°C if not administered directly.

Formulation for group 5: for 1 ml:

Equal volume of PBS and FluAd™/Gripguard™ (commercial vaccine) vaccine are mixed. The formulation is stirred for 15 minutes and stored at 4°C if not administered directly.

Formulation for group 6: for 1 ml:

250µl of PBS mod pH 7.4 are added to a 500µl dose of Aggripal™ (commercial vaccine). After 15min stirring, 250µl of SB62 is added (prepared according to the methodoly detailed for the scaled-up production). The formulation is stirred for 15 minutes and stored at 4°C if not administered directly.

Formulation for group 7: for 1 ml:

PBS mod pH 7.4 (to reach a final volume of 1ml) is added to a 500 µl dose of Aggripal™ (commercial vaccine). After 15min stirring, 250µl of SB62 is added (prepared according to the methodoly detailed for the scaled-up production). 25µg of MPL are then added. The formulation is stirred for 15 minutes and stored at 4°C if not administered directly.

Formulation for group 8: for 1 ml:

250µl of PBS mod pH 7.4 are added to a 500µl dose of Aggripal. After 15min stirring, 250µl of OW as prepared for group 2 is added and the formulation is stirred 15min and stored at 4°C if not administered directly.

Formulation for group 9: for 1 ml:

Equal volume of PBS mod pH 7.4 and Aggripal are mixed. The formulation is stirred for 15 minutes and stored at 4°C if not administered directly.

VII.1.3. Read-outs (Table 40)

CMI (ICS): 7 Days Post-immunization.

IHA/neutralization assay: 21 Days Post-immunization.

Table 40

Read-out	Timepoint	Sample type	I/P	Analysis method
ICS (CD4,CD8, IL-2, IFN-γ)	D35	PBLs	Po	FACS analysis
Humoral response	D14, D44	Sera	In	IHA, neutra

In= Individual / Po= Pool

CMI analysis (ICS: CD4/CD8; IL-2/IFN-gamma staining)

PBMCs from 24 mice/group were harvested 7 days post-immunization and tested in pools/group.

VII.2. Results

VII.2.1. Humoral immunity

Haemagglutination inhibition activity against the 3 vaccine strains was detected in sera from 24 animals per group at Day 14 after intranasal heterologous priming and at Day 16 Post-immunization.

For the 3 strains and for all groups, a boost of HI titers was observed after immunization.

- For a same adjuvant and for the 3 strains, similar HI titers were induced by the subunit vaccine and the Split vaccine.
- Similar HI titers were observed for Fluad compared to Aggripal OW for the 3 strains
- No difference was observed between Fluarix and Aggripal for H1N1 and B strains.
- For the 3 strains, statistically significant higher HI titers were observed when the Flu vaccine (Split or subunit) was adjuvanted with AS03 with or without MPL compared to the plain Flu vaccine.
- HI titers were statistically significant higher for the Flu vaccine (Split or subunit) adjuvanted with OW compared to the Flu vaccine plain only for the A/Wyoming strain.

VII.2.2. Cell-mediated immune response (ICS at day 7 post immunization)

CD4 T cell responses – Figure 17 upper part

PBMCs from 24 mice per group were harvested at Day 7 Post-immunization and tested in one pool/group. Inactivated trivalent whole viruses (1µg/ml) were used as re-stimulating antigen. Results are shown in Figure 17 upper part.

In terms of Flu whole virus-specific CD4+ T cells expressing IL-2, IFN-γ or both cytokines (Figure 17 upper part):

1. GSK adjuvants showed the same trend as previously observed (Example VI): AS03+MPL was superior to AS03 which was in turn superior to the result obtained with the plain vaccine. This trend was observed both for the split or the subunit vaccine.
2. Whatever the formulation (Plain, AS03 or AS03+MPL), the split vaccine induced a higher CD4+ T cell responses than the subunit vaccine.
3. Fluad (subunit + oil-in-water emulsion OW – see preparation section) seemed to induce similar frequencies than Fluarix Plain.
4. Formulations Trivalent Split/AS03 or Trivalent Split/AS03+MPL induced higher CD4+T cell responses than the formulation subunit/oil-in-water emulsion OW.

CD8 T cell responses – Figure 17 lower part

PBMCs from 24 mice per group were harvested at Day 7 Post-immunization and tested in one pool/group. Inactivated trivalent whole viruses (1µg/ml) were used as re-stimulating antigen.

In terms of Flu whole virus-specific CD8+ T cells expressing IL-2, IFN-γ or both cytokines (Figure 17 lower part):

• The cut-off of this experiment was relatively high due to the high background observed for the PBS negative control group.

• However higher specific CD8 T cell responses were observed for mice immunized with Trivalent Split/AS03+MPL compared to other vaccine formulations.

VII.3. Summary of results and conclusions

The following results were obtained:

1) Flu-specific CD4+ T cells obtained by ICS at Day 7 post –immunization showed:

1. Similar responses were obtained for Fluad compared to Fluarix.
2. The adjuvanted formulation induced higher immune response compared to the un-adjuvanted vaccine, both for the split influenza vaccine (as observed in humans) and for the subunit (Aggripal) vaccine (not assessed in humans). The oil-in-water emulsion adjuvant AS03 supplemented with MPL (groups 4 and 9) gave higher responses than the oil-in-water emulsion adjuvant AS03 (groups 3 and 8).
3. There is a trend of a higher CD4 responses with Split/AS03+MPL compared to Split/AS03 (Figure 17).
4. The responses induced by the split vaccine were superior to the responses obtained with the subunit vaccine (compare groups 1 to 4 and groups 5 to 9).

5. The split vaccine, whether adjuvanted with AS03 with or without MPL (groups 3 and 4) showed higher CD4+ T cell responses than the sub-unit vaccine, either Fluad (group 5) or Aggripal + OW (group 7).

2) Flu-specific CD8+ T cells obtained by ICS at Day 7 post –immunization showed no differences are observed between Split/AS3 and Split Plain (as observed in humans). There was a trend for a higher CD8+ T cell response by using Split/AS03+MPL compared to Split/AS03 or Split Plain.

3) For a same adjuvant and for the 3 strains, similar HI titers were induced by the subunit vaccine and the split vaccine. For the 3 strains, statistically significant higher titers were observed when the Flu vaccine (subunit or split) was adjuvanted with AS03 or AS03+MPL compared to the Flu vaccine plain (Flu vaccine OW > Flu vaccine Plain only for the A/Wyoming strain).

Example VIII – Clinical trial in an elderly population aged over 65 years with a vaccine containing a split influenza antigen preparation and AS03 with or without MPL adjuvant

VIII.1. Study design

A phase I, open, randomised, controlled study in an elderly population aged over 65 years (≥ 65 years-old) in order to evaluate the reactogenicity and the immunogenicity of GlaxoSmithKline Biologicals influenza candidate vaccines containing the adjuvant AS03 or AS03+MPL, administered intramuscularly as compared to Fluarix™ vaccine (known as α-Rix™ in Belgium).

Three parallel groups were assessed:

- one group of 50 subjects receiving one dose of the reconstituted and AS03 adjuvanted SV influenza vaccine (Flu AS03)
- one group of 50 subjects receiving one dose of the reconstituted and Flu AS03+MPL adjuvanted SV influenza vaccine (Flu AS03+MPL)
- one control group of 50 subjects receiving one dose of Fluarix™ (Fluarix)

VIII.2. Vaccine composition and administration

The strains used in the three vaccines were the ones that had been recommended by the WHO for the 2004-2005 Northern Hemisphere season, i.e. A/New Caledonia/20/99 (H1N1), A/New California/3/2003 (H3N2) and B/Jiangsu/10/2003. Like Fluarix™/α-Rix™, the commercially available vaccine used as a comparator, the adjuvanted vaccines (AS03, or AS03+MPL) contain 15 µg haemagglutinin (HA) of each influenza virus strain per dose.

The adjuvanted influenza candidate vaccines are 2 component vaccines consisting of a concentrated trivalent inactivated split virion antigens presented in a type I glass vial and of a pre-filled type I glass syringe containing the adjuvant (AS03 or AS03+MPL). They have been prepared as detailed in Example II. The three inactivated split virion antigens (monovalent bulks) used in formulation of the adjuvanted influenza candidate vaccines, are exactly the same as the active ingredients used in formulation of the commercial Fluarix™/α-Rix.

AS03 adjuvanted vaccine:

The AS03-adjuvanted influenza candidate vaccine is a 2 components vaccine consisting of a concentrated trivalent inactivated split virion antigens presented in a type I glass vial (335 µl) (antigen container) and of a pre-filled type I glass syringe containing the SB62 emulsion (335 µl) (adjuvant container). Description and composition of the AS03 candidate vaccine is explained in Example III.

AS03+MPL adjuvanted vaccine:

Briefly, the AS03+MPL-adjuvanted influenza candidate vaccine is a 2 components vaccine consisting of a concentrated trivalent inactivated split virion antigens presented in a type I glass vial (335 µl) (antigen container) and of a pre-filled type I glass syringe containing the AS03+MPL adjuvant (360 µl) (adjuvant container). At the time of injection, the content of the antigen container is removed from the vial by using the syringe containing the AS03+MPL adjuvant, followed by gently mixing of the syringe. Prior to injection, the used needle is replaced by an intramuscular needle and the volume is corrected to 530 µl. One dose of the reconstituted the AS03+MPL -adjuvanted influenza candidate vaccine corresponds to 530 µl. To obtain the 15 µg HA for each influenza strain at reconstitution of the AS03+MPL adjuvanted vaccine, the inactivated split virion antigen are concentrated two-fold in the antigen container (i.e. 60 µg HA/ml) as compared to Fluarix™ (i.e. 30 µg HA/ml).

The composition of one dose of the reconstituted adjuvanted influenza vaccine is identical to that reported in Table 45 (see Example XI) except for the influenza strains. Both vaccines were given intramuscularly.

VIII.3. CMI Objective, end-points and results

The CMI objectives were to determine which immunogenic composition between the formulation adjuvanted with AS03, or AS03+MPL versus the composition without any adjuvant has the strongest immunostimulating activity on CD4- and CD8- mediated immunity of individuals vaccinated with influenza antigens.

VIII.3.1. CMI end points and results

Observed variable

At days 0 and 21: frequency of cytokine-positive CD4/CD8 cells per 10^6 into 5 different cytokines. Each test quantifies the response of CD4/CD8 T cell to:

- Pool of the 3 following antigens
- New Caledonia antigen
- Wyoming antigen
- Jiangsu antigen.

Derived variables:

Antigen-specific CD4and CD8-T-cell response expressed into the 5 different tests:

(a) cells producing at least two different cytokines (CD40L, IL-2, IFNγ, TNFα)

(b) cells producing at least CD40L and another cytokine (IL-2, TNFα, IFNγ)

(c) cells producing at least IL-2 and another cytokine (CD40L, TNFα, IFNγ)

(d) cells producing at least IFNγ and another cytokine (IL-2, TNFα, CD40L)

(e) cells producing at least TNFα and another cytokine (IL-2, CD40L, IFNγ)

Analysis of the CMI response:

The CMI analysis was based on the Total vaccinated cohort.

(a) For each treatment group, the frequency of CD4/CD8 T-lymphocytes secreting in response was determined for each vaccination group, at each timepoint (Day 0, Day 21) and for each antigen: New Caledonia, Wyoming and Jiangsu and the pooled of the 3 different strains.

(b) Descriptive statistics in individual difference between timepoint (POST-PRE) responses for each vaccination group and each antigen at each 5 different cytokines.

(c) Comparison of the 3 groups regarding the 5 different cytokines on:

- CD4 T-cell response to New Caledonia, Wyoming, Jiangsu and the pool of the 3 strains

- CD8 T-cell response to New Caledonia, Wyoming, Jiangsu and the pool of the 3 strains

(d) A non-parametric test (Kruskall-Wallis test) was used to compare the location differences between the 3 groups and the statistical p-value was calculated for each antigen at each 5 different cytokines.

(e) A Wilcoxon test were use to test pairwise comparison of 2 groups respectively between Flu AS03+MPL versus Fluarix, Flu AS03+MPL versus Flu AS03 and Flu AS03 versus Fluarix

(f) All significance tests were two-tailed. P-values less than or equal to 0.05 were considered as statistically significant.

VIII.3.2. CMI results

Results were expressed as a frequency of cytokine(s)-positive CD4 or CD8 T cell within the CD4 or CD8 T cell sub-population.

Frequency of antigen specific CD4 T- lymphocytes

(a) The frequency of antigen-specific CD4 T-lymphocytes secreting in response was determined for each vaccination group, at each time point (Day 0, Day 21) and for each antigen (Pool, New Caledonia, Wyoming and Jiangsu), similarly to that performed in Example III.

(b) Comparing the difference in the frequency of antigen-specific CD4 T-lymphocytes between the 3 groups by Kruskall-Wallis test, all p-values were less than 0.05 and were considered as statistically significant.

(c) Comparing the difference in the frequency of antigen-specific CD4 T-lymphocytes between Flu AS03+MPL and *Fluarix* groups by the Wilcoxon test, all p-values were less than 0.05 and were considered as statistically significant.

(d) Comparing the difference in the frequency antigen-specific of CD4 T-lymphocytes between Flu AS03 and Fluarix groups by the Wilcoxon test, all p-values were less than 0.05 and were considered as statistically significant.

(e) Comparing the difference in the frequency of antigen-specific CD4 T-lymphocytes between Flu AS03 and Flu AS03+MPL groups by the Wilcoxon test, all p-values were more than 0.05 and were considered as no statistically significant.

Individual difference between time point (post-pre) in CD4 T-lymphocytes

(a) Descriptive statistics in individual difference between time point (POST-PRE) in CD4 T-lymphocytes responses was calculated for each vaccination group and for each antigen at each 5 different cytokines, similarly to what has been done in Example III.

(b) Comparing the individual difference POST-PRE in the antigen-specific CD4-T-lymphocytes responses between the 3 groups by Kruskall-Wallis test, all p-values were less than to 0.001 and were considered as highly statistically significant.

(c) Comparing the individual difference POST-PRE in the antigen-specific CD4-T-lymphocytes responses between Flu AS03+MPL and Fluarix using Wilcoxon test, all p-values were less than to 0.05 and were considered as statistically significant.

(d) Comparing the individual difference POST-PRE in the antigen-specific CD4-T-lymphocytes responses between Flu AS03 and Fluarix using Wilcoxon test, all p-values were less than to 0.001 and were considered as highly statistically significant.

(e) Comparing the individual difference POST-PRE in the antigen-specific CD4-T-lymphocytes responses between Flu AS03+MPL and Flu AS03 using Wilcoxon test, all p-values were more than 0.05 and were considered as no statistically significant.

VIII.4. B cell memory response Objective, end-points and Results

The objective of the study was to investigate whether the frequency of memory B cell specific to Flu Antigen are significantly induced upon one intramuscular vaccination with the Flu candidate vaccine containing the Adjuvant AS03+MPL or AS03, as compared to Fluarix in elderly population. The frequency of memory B cell has been assessed by B cell Elispot assay.

VIII.4.1. B cell memory response end-points

The end points are:

(a) At days 0, 21: cells generated in vitro cultivated memory B-cells measured by B-cell ELISPOST in all subjects in term of frequency of specific- antigen plasma within a million (10^6) of IgG producing plasma cells.

(b) Difference between post (day 21) and pre (day 0) vaccination are also expressed as a frequency of Influenza specific- antibody forming cells per million (10^6) of antibody forming cells.

VIII.4.2. B cell memory response results

The frequency of Influenza-specific antibody forming cells per million (10^6) of antibody forming cells were determined. The results showed that the frequency of memory B cell

specific to Flu antigen between Flu AS03+MPL and Fluarix groups by the Wilcoxon test was significantly ($p<0.05$) higher for B/Jiangsu strain, whilst not for the other two strains (A strains New Caledonia and Wyoming).

The individual difference between time point (post-pre) in memory B cell specific to Flu antigen was also determined. The results showed that individual difference between time point (post-pre) in the frequency of memory B cell specific to Flu antigen between Flu AS03+MPL and Fluarix groups by the by the Kruskall-Wallis test was significantly ($p<0.05$) higher for B/Jiangsu strain, whilst not for the other two strains (A strains New Caledonia and Wyoming).

The results are shown in Figure 18.

Example IX – Pre-clinical evaluation of adjuvanted and unadjuvanted influenza vaccines in ferrets (study III)

IX.1. Rationale and objectives

This study compared GSK commercial influenza trivalent split vaccine, either un-adjuvanted (Fluarix™) or adjuvanted with AS03+MPL, with two other commercially available sub-unit vaccines:

- Fluad™, Chiron's adjuvanted subunit vaccine (the adjuvant is Chiron's MF59 adjuvant),
- Agrippal™, Chiron un-adjuvanted commercial sub-unit vaccine, which was in the present study adjuvanted with AS03 adjuvant.

The objective of this experiment was to evaluate the ability of these vaccines to reduce disease symptoms (body temperature and viral shedding) in nasal secretions of ferrets challenged with heterologous strains.

The end-points were:

1) Primary end-point: reduction of viral shedding in nasal washes after heterologous challenge:

2) Secondary end-points: analysis of the humoral response by IHA and monitoring of the temperature around the priming and the heterologous challenge.

IX.2. Experimental design

IX.2.1. Treatment/group

Female ferrets (*Mustela putorius furo*) aged 14-20 weeks were obtained from MISAY Consultancy (Hampshire, UK). Ferrets were primed intranasally on day 0 with the heterosubtypic strain H1N1 A/Stockholm/24/90 (4 Log $TCID_{50}$/ml). On day 21, ferrets were injected intramuscularly with a full human dose (1ml vaccine dose, 15 µg HA/strain) of a combination of H1N1 A/New Caledonia/20/99, H3N2 A/Wyoming/3/2003 and B/Jiangsu/10/2003. Ferrets were then challenged on day 42 by intranasal route with a heterotypic strain H3N2 A/Panama/2007/99 (4.51 Log $TCID_{50}$/ml). The groups (6 ferrets/group) are illustrated in Table 41. The read-out that were performed are detailed in Table 42.

Table 41

Group	Antigen(s) + dosage	Formulation + dosage	Comments (ex: schedule /route/challenge)	Other treatments
1	Trivalent plain (Fluarix™)	Full HD: 15µg HA/strain	IM; Day 21	Priming H1N1 (A/Stockolm/24 /90) Day 0
2	Trivalent AS03+MPL	Full HD: 15µg HA/strain	IM; Day 21	Priming H1N1 (A/Stockolm/24/90) Day 0
3	Fluad™	Full HD: 15µg HA/strain	IM; Day 21	Priming H1N1 (A/Stockolm/24 /90) Day 0
4	Agrippal™ AS03	Full HD: 15µg HA/strain	IM; Day 21	Priming H1N1 (A/Stockolm/24 /90) Day 0

IX.2.2. Preparation of the vaccine formulations

Split trivalent plain (un-adjuvanted): formulation for 1 ml:

PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a mixture containing Tween 80, Triton X-100 and VES (quantities taking into account the detergents present in the strains) are added to water for injection. The detergents quantities reached are the following: 375µg Tween 80, 55µg Triton X-100 and 50µg VES per 1 ml. After 5 min stirring, 15µg of each strain H1N1, H3N2 and 17.5µg of B strain are added with 10 min stirring between each addition. The formulation is stirred for 15 minutes at room temperature and stored at 4°C if not administered directly.

Split trivalent adjuvanted with AS03+MPL: formulation for 1 ml:

PBS 10 fold concentrated (pH 7.4 when one fold concentrated) as well as a mixture containing Tween 80, Triton X-100 and VES (quantities taking into account the detergents present in the strains) is added to water for injection. The detergents quantities reached

are the following: 375µg Tween 80, 55µg Triton X-100 and 50µg VES per 1 ml. After 5 min stirring, 15µg of each strain H1N1, H3N2 and B are added with 10 min stirring between each addition. After 15 min stirring, 250µl of SB62 emulsion (prepared as detailed in Example II.1) is added. The mixture is stirred again for 15 minutes just prior addition of 25 µg of MPL. The formulation is stirred for 15 minutes at room temperature and stored at 4°C if not administered directly.

FluAd™ formulation: formulation for 1 ml:

A 2 fold dilution of FluAd™ vaccine is made in PBS buffer pH 7.4.

Agrippal™ AS03 formulation: formulation for 1 ml:

250µl of PBS buffer pH 7.4 is added to one dose of Aggripal™. After mixing, 250µl of SB62 emulsion (prepared as detailed in Example II.1) is added. The mixture is stirred at room temperature.

IX.2.2. Read-outs

Table 42

Readout	Timepoint	Sample-type	I/Po	Analysis method
Viral shedding	D-3 to D+7 Post priming D+1 to D+5 Post challenge	Nasal washes	In	Titration
T° monitoring	D-3 to D+4 Post priming D-2 to D+4 Post challenge	Implant in peritoneal cavity	In	Telemetry
IHA	Pre, Post priming, Post imm, Post challenge	Serum	In	IHA

In= Individual / Po= Pool

IX.3. Results (Figures 19 to 22)

IX.3.1. Temperature monitoring

Individual temperatures were monitored with the transmitters and by the telemetry recording. All implants were checked and refurbished and a new calibration was performed by DSI before placement in the intraperitoneal cavity. All animals were individually housed in single cage during these measurements. Temperature was monitored from 2 days Pre-challenge until 4 days Post challenge every 15 minutes and an average temperature calculated by mid-day. Results are shown in Figure 19.

Results:

Post-challenge, a peak of body temperature was observed after immunization of ferrets with the un-adjuvanted (plain) trivalent split (Fluarix™) or the sub-unit vaccine Fluad™ (which contains MF59 oil-in-water emulsion). No peak was observed after immunization of ferrets with the trivalent split vaccine adjuvanted neither with AS03+MPL nor with sub-unit Agrippal™ adjuvanted with AS03. In conclusion, an added value of the AS03-containing vaccines in the prevention of body temperature rise after challenge was shown for both the split and sub-unit tested vaccines, by contrast to the inability of the MF59 - containing vaccines to prevent this temperature rise in ferrets after challenge.

IX.3.2. Viral shedding

Viral titration of nasal washes was performed on 6 animals per group. The nasal washes were performed by the administration of 5 ml of PBS in both nostrils in awake animals. The inoculation was collected in a Petri dish and placed into sample containers on dry ice (-80°C).

All nasal samples were first sterile filtered through Spin X filters (Costar) to remove any bacterial contamination. 50 µl of serial ten-fold dilutions of nasal washes were transferred to microtiter plates containing 50 µl of medium (10 wells/dilution). 100µl of MDCK cells (2.4×10^5 cells/ml) were then added to each well and incubated at 35°C for 5-7days. After 5-7 days of incubation, the culture medium is gently removed and 100 µl of a 1/20 WST-1 containing medium is added and incubated for another 18 hrs.

The intensity of the yellow formazan dye produced upon reduction of WST-1 by viable cells is proportional to the number of viable cells present in the well at the end of the viral titration assay and is quantified by measuring the absorbance of each well at the appropriate wavelength (450 nanometers). The cut-off is defined as the OD average of uninfected control cells – 0.3 OD (0.3 OD corresponds to +/- 3 St Dev of OD of uninfected control cells). A positive score is defined when OD is < cut-off and in contrast a negative score is defined when OD is > cut-off. Viral shedding titers were determined by "Reed and Muench" and expressed as Log TCID50/ml.

Results:

Results are shown in Figure 20. Lower viral shedding was observed post-challenge with the trivalent split vaccine adjuvanted with AS03+MPL, or with the Agrippal™ sub-unit vaccine adjuvanted with AS03, as compared to the very low viral shedding reduction observed after immunization of ferrets with the un-adjuvanted (plain) trivalent split vaccine (Fluarix™) or with Fluad™ sub-unit vaccine.

Similarly to what was discussed in respect of body temperature rise, an added value of the AS03-containing vaccines was observed compared to the MF59-containing vaccines.

IX.3.3. HI titers

Anti-Hemagglutinin antibody titers to the H3N2 influenza virus strains were determined using the hemagglutination inhibition test (HI). The principle of the HI test is based on the ability of specific anti-Influenza antibodies to inhibit hemagglutination of chicken red blood cells (RBC) by influenza virus hemagglutinin (HA). Sera were first treated with a 25% neuraminidase solution (RDE) and were heat-inactivated to remove non-specific inhibitors. After pre-treatment, two-fold dilutions of sera were incubated with 4 hemagglutination units of each influenza strain. Chicken red blood cells were then added and the inhibition of agglutination was scored. The titers were expressed as the reciprocal of the highest dilution of serum that completely inhibited hemagglutination. As the first dilution of sera was 1:10, an undetectable level was scored as a titer equal to 5.

Results:

After immunization with H3N2 A/Wyoming, higher humoral responses (HI titers) were observed in ferrets immunized with the trivalent split vaccine adjuvanted with AS03+MPL or with the Agrippal™ sub-unit vaccine adjuvanted with AS03, as compared to the humoral response observed after immunization of ferrets with the un-adjuvanted (plain) trivalent split vaccine (Fluarix™) or with Fluad™ sub-unit vaccine (Figure 21).

After immunization with H3N2 A/Wyoming, higher humoral responses (HI titers) were also observed against the drift strain H3N2 A/Panama, used as the challenge strain, in ferrets immunized with Trivalent Split adjuvanted with AS03+MPL or Agrippal™ adjuvanted with AS03 compared to ferrets immunized with Trivalent Split Plain or Fluad (Figure 22).

This cross-reaction observed with our adjuvant (AS03 or AS03+MPL) against a heterologous strain correlated with the protection observed in ferrets immunized with the trivalent split vaccine adjuvanted with AS03+MPL or with the Agrippal™ sub-unit vaccine adjuvanted with AS03, and then challenged with this heterologous strain. This cross-reactivity to heterologous strain induced by AS03-containing vaccines was not induced by the MF59's adjuvanted vaccines (FluAd™).

Example X – Clinical trial in an elderly population aged over 65 years with a vaccine containing a split influenza antigen preparation and AS03 with or without MPL adjuvant: Immunogenicity persistence data at day 90 and 180

X.1. Study design

A phase I, open, randomised, controlled study in an elderly population aged over 65 years (≥ 65 years-old) in order to evaluate the reactogenicity and the immunogenicity of GlaxoSmithKline Biologicals influenza candidate vaccines containing the adjuvant AS03 or AS03+MPL, administered intramuscularly as compared to Fluarix™ vaccine (known as α-Rix™ in Belgium). This study follows that reported in Example VIII.

Three parallel groups were assessed:

- one group of 50 subjects receiving one dose of the reconstituted and AS03 adjuvanted SV influenza vaccine (Flu AS03)
- one group of 50 subjects receiving one dose of the reconstituted and Flu AS03+MPL adjuvanted SV influenza vaccine (Flu AS03+MPL)
- one control group of 50 subjects receiving one dose of Fluarix™ (Fluarix)

X.2. Immunogenicity results

X.2.1. Humoral immune response endpoints and results

In order to evaluate the humoral immune response induced by the AS03 and AS03+MPL adjuvanted vaccines and its persistence, the following parameters were calculated for each treatment group.

At Days 0, 21, 90 and 180: serum haemagglutination-inhibition (HI) antibody titres, tested separately against each of the three influenza virus strains represented in the vaccine (anti-H1N1, anti-H3N2 & anti-B-antibodies).

- Serum HI antibody GMTs' with 95% CI at Days 0, 21, 90 and 180
- Seroconversion rates with 95% CI at Days 21, 90 and 180
- Conversion factors with 95% CI at Day 21
- Seroprotection rates with 95% CI at Days 0, 21, 90 and 180

Results

The GMTs for HI antibodies with 95% CI are shown in Figure 23. Pre-vaccination GMTs of antibodies for all 3 vaccine-strains were within the same range in the 3 groups. After

vaccinations, anti-haemagglutinin antibody levels increased significantly. Post-vaccination GMTs of antibodies for the 3 vaccine strains remained however within the same ranges for all vaccines. On Day 21, a slight tendency in favour of the 2 adjuvanted vaccines compared to Fluarix was noted for the A/New Caledonia and the B/Jiangsu strains and among the two adjuvanted vaccines, the higher GMTs were observed with FLU AS03 for the A/Wyoming and B/Jiangsu strains.

The same trends were observed at Day 90. On Day 180, GMTs of antibodies for the 3 vaccine strains were within the same ranges for the 3 vaccines.

All influenza vaccines fulfilled the requirements of the European authorities for annual registration of influenza inactivated vaccines ["Note for Guidance on Harmonisation of Requirements for Influenza Vaccines for the immunological assessment of the annual strain changes" (CPMP/BWP/214/96)] in subjects aged over 60 years.

Three months (90 days) and 6 months (180 days) after vaccination, the seroprotection rates were still higher than the minimum rate of 60% required by the European Authorities whatever the study group considered. On Day 90, the minimum seroconversion rate of 30% required by the European Authorities was still achieved for all vaccines strains in the 3 vaccine groups except with Fluarix for the A/New Caledonia strain. On Day 180, it was still achieved for the A/Wyoming and B/Jiangsu strains with the 3 vaccines but not for the A/New Caledonia strain (Table 43 and Table 44).

Table 43 Seroprotection rates as the percentage of vaccinees with a serum haemagglutination inhibition titre superior or equal to 1:40 (ATP cohort for immunogenicity)

				≥ 1:40		95% CI	
Antibody	**Group**	**Timing**	**N**	**n**	**%**	**LL**	**UL**
A/New Caledonia	Flu AS03+ MPL	PRE	50	28	56.0	41.3	70.0
		PI(D21)	50	46	92.0	80.8	97.8
		PI(D90)	50	43	86.0	73.3	94.2
		PI(D180)	50	39	78.0	64.0	88.5
	Fluarix	PRE	50	26	52.0	37.4	66.3
		PI(D21)	50	46	92.0	80.8	97.8
		PI(D90)	50	38	76.0	61.8	86.9
		PI(D180)	50	34	68.0	53.3	80.5
	FluAS03	PRE	49	28	57.1	42.2	71.2
		PI(D21)	49	48	98.0	89.1	99.9
		PI(D90)	49	45	91.8	80.4	97.7
		PI(D180)	49	38	77.6	63.4	88.2
A/Wyoming	Flu AS03+ MPL	PRE	50	33	66.0	51.2	78.8
		PI(D21)	50	47	94.0	83.5	98.7
		PI(D90)	50	46	92.0	80.8	97.8
		PI(D180)	50	45	90.0	78.2	96.7

				≥ 1:40		95% CI	
Antibody	**Group**	**Timing**	**N**	**n**	**%**	**LL**	**UL**
	Fluarix	PRE	50	32	64.0	49.2	77.1
		PI(D21)	50	50	100	92.9	100.0
		PI(D90)	50	49	98.0	89.4	99.9
		PI(D180)	50	50	100	92.9	100.0
	FluAS03	PRE	49	34	69.4	54.6	81.7
		PI(D21)	49	48	98.0	89.1	99.9
		PI(D90)	49	46	93.9	83.1	98.7
		PI(D180)	49	47	95.9	86.0	99.5
B/Jiangsu	Flu AS03+ MPL	PRE	50	19	38.0	24.7	52.8
		PI(D21)	50	50	100	92.9	100.0
		PI(D90)	50	47	94.0	83.5	98.7
		PI(D180)	50	46	92.0	80.8	97.8
	Fluarix	PRE	50	17	34.0	21.2	48.8
		PI(D21)	50	48	96.0	86.3	99.5
		PI(D90)	50	47	94.0	83.5	98.7
		PI(D180)	50	47	94.0	83.5	98.7
	FluAS03	PRE	49	25	51.0	36.3	65.6
		PI(D21)	49	49	100	92.7	100.0
		PI(D90)	49	47	95.9	86.0	99.5
		PI(D180)	49	46	93.9	83.1	98.7

N = number of subjects with available results
n/% = number/percentage of subjects with titre within the specified range
PRE = pre-vaccination titre
PI (D21) = post-vaccination blood sampling at Day 21
PI (D90) = post-vaccination blood sampling at Day 90
PI (D180) = post-vaccination blood sampling at Day 180

Table 44 Seroconversion rate for haemagglutination inhibition (HI) antibody titres defined as the percentage of vaccinees who have at least a 4-fold increase in serum HI titre at each post-vaccination time point compared to Day 0 (ATP cohort for immunogenicity)

Vaccine strain	Timing	Group	N	4-fold			
				n	%	95%CI	
						LL	UL
A/NEW CALEDONIA	Day 21	Flu AS03+ MPL	50	30	60.0	45.2	73.6
		Fluarix	50	25	50.0	35.5	64.5
		Flu AS03	49	31	63.3	48.3	76.6
	Day 90	Flu AS03+ MPL	50	19	38.0	24.7	52.8
		Fluarix	50	14	28.0	16.2	42.5
		Flu AS03	49	17	34.7	21.7	49.6
	Day 180	Flu AS03+ MPL	50	12	24.0	13.1	38.2
		Fluarix	50	11	22.0	11.5	36.0
		Flu AS03	49	10	20.4	10.2	34.3
A/WYOMING	Day 21	Flu AS03+ MPL	50	46	92.0	80.8	97.8
		Fluarix	50	38	76.0	61.8	86.9
		Flu AS03	49	40	81.6	68.0	91.2
	Day 90	Flu AS03+ MPL	50	33	66.0	51.2	78.8
		Fluarix	50	33	66.0	51.2	78.8
		Flu AS03	49	31	63.3	48.3	76.6
	Day 180	Flu AS03+ MPL	50	27	54.0	39.3	68.2
		Fluarix	50	23	46.0	31.8	60.7
		Flu AS03	49	26	53.1	38.3	67.5
B/JIANGSU	Day 21	Flu AS03+ MPL	50	44	88.0	75.7	95.5
		Fluarix	50	38	76.0	61.8	86.9
		Flu AS03	49	43	87.8	75.2	95.4
	Day 90	Flu AS03+ MPL	50	37	74.0	59.7	85.4
		Fluarix	50	36	72.0	57.5	83.8
		Flu AS03	49	37	75.5	61.1	86.7
	Day 180	Flu AS03+ MPL	50	32	64.0	49.2	77.1
		Fluarix	50	29	58.0	43.2	71.8
		Flu AS03	49	31	63.3	48.3	76.6

N = number of subjects with both pre- and post-vaccination results available
n/% = number/percentage of subjects with at least a 4-fold increase
95% CI = exact 95% confidence interval; LL = lower limit, UL = upper limit

X.2.2. CMI response endpoints and results

In order to evaluate the cellular immune response induced by the adjuvanted vaccines and its persistence, the following parameters were calculated for each treatment group:
At each time point (Days 0, 21, 90 and 180): frequency of cytokine-positive CD4/CD8 cells per 10^6 in different tests (New Caledonia, Wyoming and Jiangsu antigens considered separately as well as pooled at Days 0 and 21; New Caledonia, Wyoming, Jiangsu and New York antigens considered separately as well as pooled at Days 90 and 180)

- **All double:** cells producing at least two different cytokines (CD40L, IFN-γ, IL-2, TNF-α).

- **CD40L:** cells producing at least CD40L and another cytokine (IFN-γ, IL-2, TNF-α).
- **IFN-γ:** cells producing at least IFN-γ and another cytokine (CD40L, IL-2, TNF-α).
- **IL-2:** cells producing at least IL-2 and another cytokine (CD40L, IFN-γ, TNF-α).
- **TNF-α:** cells producing at least TNF-α and another cytokine (CD40L, IFN-γ, IL-2).

Results

The main findings were (Figure 24):

(a) Twenty-one days after the vaccination, the frequency of cytokine-positive CD4 T cells (IL-2, CD40L, TNF-α and IFN-γ) was significantly higher in the two adjuvanted vaccine groups compared to the Fluarix group. No significant difference was however detected between the two adjuvants.

(b) All statistical differences between adjuvanted vaccines and Fluarix were maintained up to Day 90 and Day 180 with the following exceptions at Day 180:
   - No statistically significant difference was found between FluAS03/MPL and Fluarix for all double, CD40L, IFN-γ and IL2 (Wyoming strain only) and for all double, CD40L and TNF-α (New York strain only)
   - No statistically significant difference was found between FluAS03 and Fluarix for IL2 (Jiangsu strain only)

(c) The absence of statistically significant difference between the two adjuvanted vaccines was confirmed up to Day 90 and Day 180.

(d) The difference between pre and post-vaccination (Day 21) in CD4 T-lymphocytes responses for all cytokines investigated (IL-2, CD40L, TNF-α and IFN-γ) was significantly higher with the two adjuvanted vaccines compared to Fluarix™. No significant difference was however detected between both adjuvants.

(e) The vaccination had no measurable impact on the CD8 response whatever the treatment group.

Example XI - Clinical trial in an elderly population aged over 65 years with a vaccine containing a split influenza antigen preparation and AS03 with MPL adjuvant

XI.1. Study design and objectives

A phase I/II, open, controlled study was conducted in order to evaluate the reactogenicity and the immunogenicity of GlaxoSmithKline Biologicals influenza candidate vaccine containing the AS03+MPL adjuvant in an elderly population aged over 65 years (> 65 years-old) previously vaccinated in 2004 with the same candidate vaccine. For immunogenicity and safety evaluations, Fluarix™ (known as α-Rix™ in Belgium) vaccine was used as reference.

Two parallel groups were assessed:

- One group of about 50 subjects who had previously received one dose of the reconstituted adjuvanted influenza vaccine during the previous clinical trial
- One control group (Fluarix) of about 50 subjects who had previously received one dose of Fluarix™ during the previous clinical trial

One objective of this study was to evaluate the humoral immune response (anti-haemagglutinin and anti-MPL titres) of the revaccination with the adjuvanted influenza vaccine Flu AS03+MPL administered about one year after administration of the first dose. For comparison purposes, subjects who had already received Fluarix™ in the previous trial received a dose of commercial vaccine and formed the control group of this trial.

XI.2. Vaccine composition and administration

The strains used in the three vaccines were the ones that had been recommended by the WHO for the 2005-2006 Northern Hemisphere season, i.e. A/New Caledonia/20/99 (H1N1), A/New California/7/2004 (H3N2) and B/Jiangsu/10/2003. Like Fluarix™/α-Rix™, the commercially available vaccine used as a comparator, the (AS03+MPL – adjuvanted vaccine, hereinafter in short "the adjuvanted vaccine") contains 15 µg haemagglutinin (HA) of each influenza virus strain per dose.

The adjuvanted influenza candidate vaccine is a 2 component vaccine consisting of a concentrated trivalent inactivated split virion antigens presented in a type I glass vial and of a pre-filled type I glass syringe containing the AS03+MPL adjuvant. It has been prepared according the method detailed in Example II.

At the time of injection, the content of the prefilled syringe containing the adjuvant is injected into the vial that contains the concentrated trivalent inactivated split virion antigens. After mixing the content is withdrawn into the syringe and the needle is replaced by an intramuscular needle. One dose of the reconstituted the adjuvanted influenza

candidate vaccine corresponds to 0.7 mL. The adjuvanted influenza candidate vaccine is a preservative-free vaccine.

The composition of one dose of the reconstituted adjuvanted influenza vaccine is given in Table 45. Both vaccines were given intramuscularly.

Table 45 Composition of the reconstituted vaccine adjuvanted (AS03+MPL) influenza candidate vaccine

Component	Quantity per dose
Inactivated split virions	
- A/New Caledonia/20/99 (H1N1)	15 µg HA
- A/New California/7/2004 (H3N2)	15 µg HA
- B/Jiangsu/10/2003	15 µg HA
Adjuvant	
SB62 emulsion	
- (squalene)	10.68 mg
- (DL-alpha-tocopherol)	11.86 mg
- (polysorbate 80 – Tween 80)	4.85 mg
MPL	25 µg

XI.3. Immunogenicity results

XI.3.1. Anti-HA Humoral immune response endpoints and results

Observed variables:

At days 0 and 21: serum haemagglutination-inhibition (HI) antibody titres, tested separately against each of the three influenza virus strains represented in the vaccine (anti-H1N1, anti-H3N2 & anti-B-antibodies).

Derived variables (with 95% confidence intervals):

(f) Geometric mean titres (GMTs) of serum HI antibodies with 95% confidence intervals (95% CI) pre and post-vaccination

(g) Seroconversion rates* with 95% CI at day 21

(h) Seroconversion factors** with 95% CI at day 21

(i) Seroprotection rates*** with 95% CI at day 21

* Seroconversion rate defined as the percentage of vaccinees with either a pre-vaccination HI titre <1:10 and a post-vaccination titre ≥ 1:40, or a pre-vaccination titre ≥ 1:10 and a minimum 4-fold increase at post-vaccination titre, for each vaccine strain.

**Seroconversion factor defined as the fold increase in serum HI GMTs on day 21 compared to day 0, for each vaccine strain.

***Protection rate defined as the percentage of vaccinees with a serum HI titre ≥40 after vaccination (for each vaccine strain) that usually is accepted as indicating protection.

Results

As expected, the vast majority of subjects were already seropositive for the three strains in both groups before vaccination. Pre-vaccination GMTs for all 3 vaccine strains were within the same range in the 2 groups. There was a trend for higher GMTs at post-vaccination for all 3 vaccine strains in the Flu AS03+MPL group compared to the Fluarix group, although 95% CI were overlapping (Figure 25).

The two influenza vaccines fulfilled the requirements of the European authorities for annual registration of influenza inactivated vaccines ["Note for Guidance on Harmonisation of Requirements for Influenza Vaccines for the immunological assessment of the annual strain changes" (CPMP/BWP/214/96)] in subjects aged over 60 years (Table 46).

Table 46 Seroprotection rates seroconversion rates and conversion factors at day 21 (ATP cohort for immunogenicity)

Strains	Group	N	Seroprotection rate (HI titre ≥ 40) %	Seroconversion rate (≥ 4-fold increase) [95% CI] %	Seroconversion factor [95% CI] %
EU standard (>60			>60%	>30%	>2.0
A/New Caledonia	Flu+MPL-AS03	38	89.5 [75.20-97.06]	31.6 [17.5-48.7]	3.1[2.2-4.4]
	Fluarix	45	82.2 [67.95-92.00]	31.1 [18.2-46.6]	2.5 [1.8-3.5]
A/New York (H3N2)	Flu+MPL-AS03	38	92.1 [78.62-98.34]	78.9 [62.7-90.4]	8.8 [6.1-12.5]
	Fluarix	45	95.6 [84.85-99.46]	68.9 [53.4-818]	6.0 [4.4-8.3]
B/Jiangsu (B)	Flu+MPL-AS03	38	100 [90.75-100]	57.9 [40.8-73.7]	5.1 [3.7-7.0]
	Fluarix	45	100 [92.13-100]	37.8 [23.8-53.5]	3.1 [2.4-4.0]

N= total number of subject; %= Percentage of subjects with titre at day 21 within the specified range; CI = confidence interval

Example XII - Clinical trial in an elderly population aged over 65 years with a vaccine containing a split influenza antigen preparation adjuvanted with AS03 and MPL at two different concentrations

XII.1. Study design and objectives

An open, randomized phase I/II study to demonstrate the non inferiority in term of cellular mediated immune response of GlaxoSmithKline Biologicals influenza candidate vaccines containing various adjuvants administered in elderly population (aged 65 years and older) as compared to Fluarix™ (known as α-Rix™ in Belgium) administered in adults (18-40 years)

Four parallel groups were assigned:

(a) 75 adults (aged 18-40 years) in one control group receiving one dose of Fluarix™ (Fluarix group)

(b) 200 elderly subjects (aged 65 years and older) randomized 3:3:2 into three groups:

- one group with 75 subjects receiving influenza vaccine adjuvanted with AS03+MPL (concentration 1 – 25 µg)
- One group with 75 subjects receiving influenza vaccine adjuvanted with AS03+MPL (concentration 2 – 50 µg)
- Reference Flu group with 50 subjects receiving one dose of Fluarix™

Primary Objective

The primary objective is to demonstrate the non inferiority 21 days post-vaccination of the influenza adjuvanted vaccines administered in elderly subjects (aged 65 years and older) as compared to Fluarix™ administered in adults (aged 18-40 years) in terms of frequency of influenza-specific CD4 T-lymphocytes producing at least two different cytokines (CD40L, IL-2, TNF-α, IFN-γ).

Secondary objectives

The secondary objectives are

(a) To evaluate the safety and reactogenicity of vaccination with candidate influenza vaccines adjuvanted during 21 days following the intramuscular administration of the vaccine in elderly subjects (aged 65 years and older). Fluarix™ is used as reference.

(b) To evaluate the humoral immune response (anti-haemagglutinin titre) 21, 90 and 180 days after vaccination with influenza candidate vaccines adjuvanted. Fluarix™ is used as reference.

Tertiary Objective

The tertiary objective is to evaluate the cell mediated immune response (production of IFN-γ, IL-2, CD40L, and TNF-α and memory B- cell response) 21, 90 and 180 days after vaccination with adjuvanted influenza-vaccines. Fluarix™ is used as reference.

XII.2. Vaccine composition and administration

The influenza vaccine adjuvanted with AS03+MPL(25 µg per dose) system is also used in study illustrated in Example XI. The influenza vaccine adjuvanted with AS03+MPL(50 µg per dose) system is of identical composition except that the concentration of MPL is

doubled. The process is the same as the one described in Example VIII for the influenza vaccine adjuvanted with AS03+MPL, with as only difference that the concentration of MPL is doubled.

Control: full dose of Fluarix™ by IM administration.

Four scheduled visits per subject: at days 0, 21, 90 and 180 with blood sample collected at each visit to evaluate immunogenicity.

Vaccination schedule: one injection of influenza vaccine at day 0

XII.3. Immunogenicity results

XII.3.1. CMI endpoints and results

Evaluation of the primary endpoint.

At day 21: CMI response in all subjects in terms of frequency of influenza-specific CD4 T-lymphocyte per 10^6 in tests producing at least two different cytokines (IL-2, IFN-γ, TNF-α and CD40L)

For evaluation of CMI response, frequency of influenza-specific CD4 are analysed as follows:

The GM ratio in term of influenza-specific CD4 frequency between groups vaccinated with adjuvanted vaccines and Flu YNG is obtained using an ANCOVA model on the logarithm-transformed titres. The ANCOVA model includes the vaccine group as fixed effect and the pre-vaccination log-transformed titre as regressor. The GM ratio and their 98.75% CI are derived as exponential-transformation of the corresponding group contrast in the model. The 98.75% CI for the adjusted GM is obtained by exponential-transformation of the 98.75% CI for the group least square mean of the above ANCOVA model.

Results - Inferential analysis (Table 47)

The adjusted GM and GM ratios (with their 98.75% CI) of influenza-specific CD4 T-lymphocyte producing at least two cytokines (IL-2, IFN-γ, TNF-α and CD40L) at day 21, after *in vitro* restimulation with "pooled antigens II", are presented in Table 47. For each adjuvanted influenza vaccine, the upper limit of two-sided 98.75% CI of GM ratio is far below the clinical limit of 2.0. This shows the non-inferiority of both adjuvanted influenza vaccines administered to elderly subjects compared to the Fluarix™ vaccine administered in adults aged between 18 and 40 years in term of post-vaccination frequency of influenza-specific CD4.

Table 47 Adjusted GM ratio of influenza-specific CD4 producing at least two cytokines, Day 21 (ATP cohort for immunogenicity)

				Adjusted GM ratio (Flu YNG / AS03+MPL (conc. 1)		
Flu YNG		AS03+MPL (conc. 1)			98.8% CI	
N	Adjusted GM	N	Adjusted GM	Value	LL	UL
70	1995.3	72	2430.0	0.82	0.65	1.04

				Adjusted GM ratio (Flu YNG / AS03+MPL (conc. 2)		
Flu YNG		AS03+MPL (conc. 2)			98.8% CI	
N	Adjusted GM	N	Adjusted GM	Value	LL	UL
70	1979.4	72	2603.8	0.76	0.59	0.98

Adjusted GM = geometric mean antibody adjusted for baseline titre; N = Number of subjects with both pre- and post-vaccination results available; 98.8% CI = 98.8% confidence interval for the adjusted GM ratio (Ancova model: adjustment for baseline); LL = lower limit, UL = upper limit

Results - Descriptive analysis (Figure 26)

The main findings were:

- Before vaccination the CMI response if higher in young adults than in elderly
- After vaccination,
  - there was a booster effect of the influenza vaccine on the CMI response in young adults (18-40 years)
  - CMI response in the elderly having received adjuvanted influenza vaccine is comparable to the CMI response of young adults.
- The difference between pre and post-vaccination in CD4 T-lymphocytes responses for all cytokines investigated (IL-2, CD40L, TNF-α and IFN-γ) was significantly higher with the adjuvanted vaccines compared to Fluarix™ (18-40 years) for all tests excepted for IFNγ when we compare Fluarix (18-40 years) and Flu/AS03+MPL (conc. 1).

It should be noted that the in vitro stimulation was performed with the Flu strains (i) B/Jiangsu, (ii) A/H3N2/New-York and (iii) A/H3N2/Wyoming instead of A/H1N1/New-Caledonia included in the vaccine. However, preliminary data including the A/H1N1/New Caledonia vaccine strain from subsets of subjects indicate that the results will be similar.

Results - Evaluation of the tertiary end-point (Table 48)

In order to evaluate the tertiary end point, the frequency of influenza-specific CD4/CD8 T-lymphocytes and memory B-cells were measured at days 0, 21, 90 and 180.

The frequency of influenza-specific cytokine-positive CD4/CD8 T-lymphocytes was summarised (descriptive statistics) for each vaccination group at days 0 and 21, for each antigen.

A Non-parametric test (Wilcoxon test) was used to compare the location of difference between the two groups (*influenza adjuvanted vaccine* versus *Fluarix™*) and the statistical p-value is calculated for each antigen at each different test.

Descriptive statistics in individual difference between day 21/day 0 (Post-/Pre-vaccination) responses is calculated for each vaccination group and each antigen at each different test.

A Non-parametric test (Wilcoxon test) is used to compare the individual difference Post-/Pre-vaccination) and the statistical p-value will be calculated for each antigen at each different test.

The p-values from Wilcoxon test used to compare the difference in the frequency of influenza-specific CD4 T-lymphocytes are presented in Table 48.

Table 48 Inferential statistics: p-values from Kruskal-Wallis Tests for CD4 T cells at each time point (ATP Cohort for immunogenicity)

	p-value							
	Group1 and Flu ELD		Group2 and Flu ELD		Group1 and Flu YNG		Group2 and Flu YNG	
	day 0	day 21	day 0	day 21	day 0	day 21	day 0	day 21
ALL DOUBLES	0.4380	0.0003	0.4380	0.0003	0.0000	0.9014	0.0005	0.4889
CD4OL	0.3194	0.0002	0.3194	0.0002	0.0000	0.9841	0.0003	0.5412
IFNγ	0.5450	0.0004	0.5450	0.0004	0.0000	0.5397	0.0001	0.7895
IL2	0.3701	0.0008	0.3701	0.0008	0.0003	0.8557	0.0022	0.4766
TFNα	0.3716	0.0004	0.3716	0.0004	0.0000	0.8730	0.0013	0.2114

Group 1: Influenza vaccine adjuvanted with AS03+MPL (**conc. 1**)
Group 2: Influenza vaccine adjuvanted with AS03+MPL (**conc. 2**)

The main conclusions are:

(a) Pre-vaccination GM frequencies of influenza-specific CD4 were similar in all groups of elderly subjects but superior in the adults aged between 18 and 40 years.

(b) Post-vaccination (day 21) frequency of influenza-specific CD4 T lymphocytes was similar in elderly subjects vaccinated with adjuvanted vaccines and in adults aged between 18 and 40 years vaccinated with Fluarix™.

(c) In elderly subjects, post-vaccination (day 21) frequency of influenza-specific CD4 T lymphocytes was significantly higher after vaccination with adjuvanted vaccines than with Fluarix™.

(d) Pre-vaccination and post vaccination GM frequency of influenza-specific CD8 T cell was essentially similar in all groups.

Results - Evaluation of the Humoral immune response endpoints

Observed variables:

At days 0, 21, 90 and 180: serum haemagglutination-inhibition (HI) antibody titres, tested separately against each of the three influenza virus strains represented in the vaccine (anti-H1N1, anti-H3N2 & anti-B-antibodies).

The cut-off value for HI antibody against all vaccine antigens was defined by the laboratory before the analysis (and equals 1:10). A seronegative subject is a subject whose antibody titre is below the cut-off value. A seropositive subject is a subject whose antibody titre is greater than or equal to the cut-off value. Antibody titre below the cut-off of the assay is given an arbitrary value of half the cut-off.

Based on the HI antibody titres, the following parameters are calculated:

(j) Geometric mean titres (GMTs) of HI antibody at days 0 and 21, calculated by taking the anti-log of the mean of the log titre transformations.

(k) Seroconversion factors (SF) at day 21 defined as the fold increase in serum HI GMTs on day 21 compared to day 0.

(l) Seroconversion rates (SC) at day 21 defined as the percentage of vaccinees with either a pre-vaccination HI titre <1:10 and a post-vaccination titre ≥ 1:40, or a pre-vaccination titre ≥ 1:10 and a minimum 4-fold increase at post-vaccination titre.

(m) Seroprotection rates (SPR) at day 21 defined as the percentage of vaccinees with a serum HI titre ≥1:40.

The 95% CI for GM is obtained within each group separately. The 95% CI for the mean of log-transformed titre is first obtained assuming that log-transformed titres are normally distributed with unknown variance. The 95% CI for the GM is then obtained by exponential-transformation of the 95% CI for the mean of log-transformed titre.

Missing serological result for a particular antibody measurement is not replaced. Therefore a subject without serological result at a given time point do not contribute to the analysis of the assay for that time point.

Humoral immune response results (Figure 27 and Table 49)

Pre-vaccination GMTs of HI antibodies for all 3 vaccine strains were within the same range in the 4 treatment groups. After vaccination, there is clear impact of the 2 adjuvants which increase the humoral response in elderly, compared to standard Fluarix in the same population.

GMTs are

- significantly higher for H1N1 for AS03+MPL (conc. 2),
- significantly higher for H3N2 and for B for both adjuvants,

Twenty one days after vaccination, the subjects of Fluarix (18-40 years) had a higher HI response for New Caledonia and B/Jangsu strains..

As shown in Table 49, the adjuvanted influenza vaccines exceeded the requirements of the European authorities for annual registration of split virion influenza vaccines ["Note for Guidance on Harmonization of Requirements for Influenza Vaccines for the immunological assessment of the annual strain changes" (CPMP/BWP/214/96)] in subjects aged over 60 years.

After vaccination, there was a statistically difference in terms of **seroprotection rates** of HI antibodies between Fluarix (≥65 years) group and

- Flu AS03+MPL (conc 2) for A/New Caledonia strain

For each vaccine strain, the seroprotection rates for the 2 influenza adjuvanted vaccine groups are in the same range compared to Fluarix (18-40 years) group.

There was a statistically difference in terms of **seroconversion rates** of HI antibodies between Fluarix (≥65 years) group and

- Flu AS03+MPL (**conc 2)** for A/New Caledonia strain
- Flu AS03+MPL (**conc 1)** for B/Jiangsu strain

For each vaccine strain, the seroconversion rates for the 2 influenza adjuvanted vaccine groups are in the same range compared to Fluarix (18-40 years) group excepted for New Caledonia strain.

Table 49 Seroprotection rates seroconversion rates and conversion factors at day 21 (ATP cohort for immunogenicity)

Strains	Group	N	Seroprotection rate (HI titre ≥ 40) %	Seroconversion rate (≥ 4-fold increase) [95% CI]	Conversion factor [95% CI] %
EU standard (>60 years)			>60%	>30%	>2.0
EU standard (< 60 years)			> 70%	> 40%	> 2.5
A/New Caledonia (H1N1)	Flu Yng	75	100 [95.20-100]	77.3 [66.2-86.2]	35.1 ^21.9-56.4]
	Flu Elderly	49	71.4 [56.74-83.42]	30.6 [18.3-45.4]	3.7 [2.4-5.7]
	FluAS03+MPL (conc. 1)	75	90.5 [81.48-96.11]	55.4 [43.4-67.0]	6.4 [4.5-9.0]
	FluAS03+MPL (conc. 2)	75	98.7 [92.79-99.97]	74.7 [63.3-84.0]	9.2 [6.4-13.3]
A/New York (H3N2)	Flu Yng	75	93.3 [85.12-97.80]	76.0 [64.7-85.1]	9.2 [7.1-11.8]
	Flu Elderly	49	81.6 [67.98-91.24]	69.4 [54.6-81.7]	8.2 [5.7-11.8]
	FluAS03+MPL (conc. 1)	75	94.6 [86.73-98.51]	90.5 [81.5-96.1]	19.2 [14.6-25.3]

	FluAS03+MPL (conc. 2)	75	93.3 [85.12-97.80]	82.7 [72.2-90.4]	15.0 [11.2-20.2]
B/Jiangsu (B)	Flu Yng	75	100 [95.20-100]	80.0 [69.2-88.4]	13.9 [10.1-19.1]
	Flu Elderly	49	93.9 [83.13-98.72]	81.3 [70.7-89.4]	4.3 [3.0-6.1]
	FluAS03+MPL (conc. 1)	75	95.9 [88.61-99.16]	73.0 [61.4-82.6]	8.5 [6.5-11.2]
	FluAS03+MPL (conc. 2)	75	98.7 [92.79-99.97]	66.7 [54.8-77.1]	7.6 [5.6-10.2]

N= total number of subject; %= Percentage of subjects with titre at day 21 within the specified range; CI = confidence interval

XII.3.2. Immunogenicity conclusions

(a) Pre-vaccination frequency of influenza-specific CD4 was significantly inferior in elderly adults compared to adults aged between 18 and 40 years. After vaccination with Fluarix™, post-vaccination frequency (day 21) remained inferior in elderly adults compared to younger ones. On the contrary, the non-inferiority in term of frequency of post-vaccination frequency of influenza-specific CD4 after vaccination with adjuvanted vaccines of elderly subjects was demonstrated compared to vaccination with Fluarix™ in adults aged between 18 and 40 years.

(b) Regarding the humoral immune response in term of HI antibody response, all influenza vaccines fulfilled the requirements of the European authorities for annual registration of influenza inactivated vaccines ["Note for Guidance on Harmonisation of Requirements for Influenza Vaccines for the immunological assessment of the annual strain changes" (CPMP/BWP/214/96)]. In elderly adults, adjuvanted vaccines mediated at least a trend for a higher humoral immune response to influenza haemagglutinin than Fluarix™. Significant difference between the humoral immune response against each vaccine strain mediated in elderly subjects by adjuvanted vaccines compared to Fluarix™ are summarised in Table 50. Compared to adults aged between 18 and 40 years vaccinated with Fluarix™, elderly subjects vaccinated with the adjuvanted vaccines showed a trend for higher post-vaccination GMTs and seroconversion factor at day 21 against the A/New York strain.

Table 50 Significant difference in humoral immune response between adjuvanted vaccines and Fluarix in elderly subjects

	Post-vacc GMT	Seroconversion Factor	Seroprotection rate	Seroconversion Rate
Flu AS03+MPL (conc. 1)	A/New York B/Jiangsu	A/New York	-	B/Jiangsu
Flu AS03+MPL (conc. 2)	A/New York B/Jiangsu A/New Caledonia	A/New Caledonia	A/New Caledonia	A/New Caledonia

XII.4. Reactogenicity results

XII.4.1. Recording of Adverse events (AE)

Solicited symptoms (see Table 51) occurring during a 7-day follow-up period (day of vaccination and 6 subsequent days) were recorded. Unsolicited symptoms occurring during a 21-day follow-up period (day of vaccination and 20+3 subsequent days) were also recorded. Intensity of the following AEs was assessed as described in Table 52.

Table 51 Solicited local/general adverse events

Solicited local AEs	Solicited general AEs
Pain at the injection site Redness at the injection site Swelling at the injection site Haematoma	Fatigue Fever Headache Muscle ache Shivering Joint pain in the arm of the injection Joint pain at other locations

N.B. Temperature was recorded in the evening. Should additional temperature measurements performed at other times of day, the highest temperature was recorded.

Table 52 Intensity scales for solicited symptoms in adults

Adverse Event	Intensity grade	Parameter
Pain at injection site	0	Absent
	1	on touch
	2	when limb is moved
	3	prevents normal activity
Redness at injection site		Record greatest surface diameter in mm
Swelling at injection site		Record greatest surface diameter in mm
Haematoma at injection site		Record greatest surface diameter in mm
Fever*		Record temperature in °C / °F
Headache	0	Absent
	1	is easily tolerated
	2	interferes with normal activity
	3	prevents normal activity
Fatigue	0	Absent
	1	is easily tolerated
	2	interferes with normal activity
	3	prevents normal activity
Joint pain at the injection site and other locations	0	Absent
	1	is easily tolerated
	2	interferes with normal activity
	3	prevents normal activity
Muscle ache	0	Absent
	1	is easily tolerated

	2	interferes with normal activity
	3	prevents normal activity
Shivering	0	Absent
	1	is easily tolerated
	2	interferes with normal activity
	3	prevents normal activity

*Fever is defined as axillary temperature ≥37.5°C (99.5°F)

The maximum intensity of local injection site redness/swelling is scored as follows:
0 is 0 mm; 1 is > 0 - ≤ 20 mm; 2 is > 20 - ≤ 50 mm; 3 is > 50 mm.
The maximum intensity of fever is scored as follows:
1 is >37.5 - ≤38.0°C; 2 is >38.0 - ≤39.0°C; 3 is >39.0

The investigator makes an assessment of intensity for all other AEs, i.e. unsolicited symptoms, including SAEs reported during the study. The assessment is based on the investigator's clinical judgement. The intensity of each AE recorded is assigned to one of the following categories:
1 (mild) = An AE which is easily tolerated by the subject, causing minimal discomfort and not interfering with everyday activities;
2 (moderate) = An AE which is sufficiently discomforting to interfere with normal everyday activities;
3 (severe) = An AE which prevents normal, everyday activities (In adults/ adolescents, such an AE would, for example, prevent attendance at work/ school and would necessitate the administration of corrective therapy).

XII.4.2. Recording of Adverse events (AE)

The reactogenicity observed in elderly subjects with adjuvanted vaccines, in terms of both local and general symptoms, was found to be higher than with Fluarix™ in the same population. However, it was shown to be similar to the level seen in the adult population. The incidence and the intensity of symptoms was increased after vaccination with adjuvanted vaccines compared to the reactogenity seen in elderly subjects with Fluarix™ (Figure 28). In all cases, symptoms resolved rapidly.
Grade 3 symptoms showed a trend to be higher in the group who received the vaccine adjuvanted with the highest MPL concentration compared to the group who received the adjuvanted vaccine wherein the MPL is at a lower concentration. In all cases, symptoms however resolved rapidly.

Example XIII - Preclinical studies in mice using HPV vaccine adjuvanted with AS03+MPL

XIII.1. Introduction

BALB/C mice were injected with an adjuvanted mixture containing 2.5 µg of each of 4 different antigens, HPV 16 L1, HPV 18 L1, HPV 31 L1, and HPV 45 L1, in the form of virus like particles. Each L1 protein was a C terminal truncate removing, in the case of HPV 16, 34 amino acids (or the equivalent region in the other sequences).

HPV proteins were expressed and purified in baculovirus expression systems, for example as described in WO 03/077942.

The tetravalent VLP combination was combined with one of 2 different adjuvants.

The adjuvants tested were:

1 A mixture of aluminium hydroxide and 3-D MPL (termed AS04).

2 A mixture of AS03 + 3D-MPL, prepared essentially as Example 2.

The adjuvant AS04 was used in a vaccine comprising HPV 16 and HPV 18 L1-only virus like particles, tested in phase II clinical trials as described in The Lancet, vol 364, issue 947, 13 November 2004, 1757-1765. It thus provides a good basis for comparison with AS03+ 3D MPL. This adjuvant can be made as described in, for example, WO 01/17751 and WO 00/23105.

An analysis of antibody titres was carried out for each component of the vaccine. In addition, an analysis of B memory cells specific for HPV 16 and HPV 18 was made within the total population of IgG molecules.

XIII.2. Material and Methods

XIII.2.1. Animal model

Two groups of BALB/c mice (n=10) were immunised intramuscularly in one leg (days 0 and 28) with 2.5 µg of HPV-16/18/31/45 L1, formulated with AS04 ($Al(OH)_3$ 50µg + MPL 5µg) or a mixture of AS03 + 3D-MPL, prepared essentially as Example 2, containing 50µl emulsion + MPL 5µg.

XIII.2.2. Anti-HPV-16/18/31/45 L1 serology: Ig

Quantitation of anti-HPV-16/18/31/45 L1 antibody was performed by ELISA using HPV-16 L1 (lot E16L1P093), HPV-18 L1 (lot E18L1P079), HPV-31 L1 (lot EA31L1P329) and HPV-45 L1 (lot EA45L1P328) as coating antigen. HPV-16/18/31/45 L1 and antibody solutions were used at 50µl per well; only the saturation solution was used at 100µl per well. HPV-16/18/31/45 L1 were diluted at a final concentration of 0.5 µg/ml in PBS and were adsorbed overnight at 4°C to the wells of 96 wells microtiter plates (Maxisorb Immuno-plate, Nunc, Denmark). After removal of coating solution, the plates were then incubated for 1hr at 37°C with PBS containing 1% bovine serum albumin (saturation buffer). Two-fold dilutions of mice sera in the dilution buffer (saturation buffer + 0.1% Tween20) were added to the coated plates after removal of saturation solution and incubated for 1 hr 30 min at 37°C. The plates were washed four times with PBS 0.1% Tween 20 and biotin-conjugated anti-mouse Ig 1/1000 (Dako) diluted in saturation buffer, was added to each well and incubated for 1 hr 30 min at 37°C. After a washing step, avidin-horseradish peroxydase complex (Dako, UK) diluted 1/3000 in saturation buffer was added for an additional 30 min at 37°C. Plates were washed four times as above and incubated for 20 min at room temperature with a solution of o-phenylenediamine (Sigma MO, USA) 0.04% H_2O_2 0.03% in 0.1% Tween 20 0.05M citrate buffer pH4.5. The reaction was stopped with the addition of H_2SO_4 2N and the plates were read at 490/630 nm.

ELISA titer calculation

The optical densities (OD's) were measured using a microplate reader connnected to a computer. Data were captured with the SoftMaxPro software. In order to titrate each sample, a standard is included on each plate. A four parameters logistic log function is used to calculate the standard curve. Antibody concentrations were calculated at each dilution of the test sample by interpolation of the standard curves.

The antibody titers were obtained by averaging the values from all dilutions that fall within the working range (20-80 % OD) of the standard curve. ELISA titers are expressed in EU/ml.

XIII.2.3. B memory cell ELISPOT

Thirty-three or seventy-five days after the second immunization, mice were sacrificed; spleen cells were separated by a lymphoprep gradient. PBMCs were then resuspended in RPMI 1640 medium (Gibco) containing additives (sodium pyruvate 1mM, MEM non-essential amino acids, Pen/Strep, Glutamine and β-2 mercaptoethanol), 5% fetal calf serum, 50U/ml rhIL-2 (eBioscience) and 3µg/ml CpG (phosphothioated CpG ODN-7909 - 5'-TCG TCG TTT TGT CGT TTT GTC GTT-3' – SEQ ID NO.1). Other CpG sequences

are also suitable for use in this B memory evaluation method. Cells were cultured five days at a final concentration of 10^6 cells/ml, in 5 ml per flat-bottomed 6 wells. After an activation step with ethanol, nitrocellulose plates (Multiscreen-IP; Millipore) were coated with 10µg/ml of HPV-16/18 L1 or goat anti-mouse Ig (GAM; Sigma) diluted 1/200 in PBS. After a saturation step with complete medium, 100µl of 2.10^6 cells/ml were added to HPV-16/18 L1 coated plates and 100µl of 10^6 and 5.10^5 cells/ml were added to GAM plates. After an incubation time of 2 hrs at 37°c, plates were stored overnight at 4°C. Plates were washed four times with PBS 0.1% Tween 20 and anti-mouse Ig Biot diluted 1/200 in PBS 1% BSA 5% FCS (dilution buffer) was distributed to plates and incubated for 2 hrs at 37°C. After a washing step, Extravidin HRP (Sigma) diluted 1/550 in dilution buffer was added for an additional 1 hr at 37°C. Plates were washed as above and incubated for 10 min at room temperature with a solution of AEC (Sigma). Reaction was stopped by rinsing plates gently under tap water. After drying, plates were read with an automated ELISPOT image analysis system (Zeiss KS400).

The percentage of B memory cells specific for HPV-16/18 L1 corresponds to the ratio of HPV-16/18 L1 positive spots compared to the total IgG spots.

XIII.3. Serological results (Tables 53-55)

Table 53

	Anti-VLPs titers (EU/ml) at day 14 post II			
	HPV16	**HPV18**	**HPV31***	**HPV45**
VLPs 2.5µg / AS04	528441	512290	28671	42224
VLPs 2.5µg / AS03+ 3D MPL	824773	460800	77232	65716

Table 54

	Anti-VLPs titers (EU/ml) at day 75 post II			
	HPV16	**HPV18**	**HPV31**	**HPV45***
VLPs 2.5µg / AS04	230425	241983	15846	9812
VLPs 2.5µg / AS03+ 3D MPL	415657	292199	29054	32230

* statistical difference between groups

Table 55 – B cell memory

	Frequency of HPV specific memory B cells in total IgG memory B cells at day 33 post II	
	HPV16	**HPV18**
VLPs 2.5µg / AS04	0.4%	0.2%

VLPs 2.5µg / AS03+ 3D MPL	0.7%	0.4%
	Frequency of HPV specific memory B cells in total IgG memory B cells at day 75 post II	
	HPV16	HPV18
VLPs 2.5µg / AS04	2.0%	0.6%
VLPs 2.5µg / AS03+ 3D MPL	3.8%	0.5%

XIII.4. Conclusions

In the animal model system tested the AS03+3D-MPL adjuvant demonstrates immunogenicity results for both antibody production and B cell memory which are equivalent to, or sometimes greater than, those generated with AS04, depending upon the HPV type being assessed.

CLAIMS

1. An immunogenic composition comprising:

(a) an antigen or antigenic composition,

(b) an oil-in-water emulsion adjuvant; and

(c) 3D MPL.

wherein said oil-in-water emulsion comprises a metabolisable oil, a sterol and an emulsifying agent.

2. An immunogenic composition according to claim 1 wherein said metabolisable oil is present in an amount of 0.5% to 20% of the total volume of said immunogenic composition.

3. An immunogenic composition according to claim 1 or claim 2 wherein said metabolisable oil is present in an amount of 1.0% to 10% of the total volume of said immunogenic composition.

4. An immunogenic composition according to any of claims 1 to 3 wherein said metabolisable oil is present in an amount of 2.0% to 6.0% of the total volume of said immunogenic composition.

5. An immunogenic composition as claimed in any of claims 1 to 4 wherein said sterol is alpha-tocopherol.

6. An immunogenic composition as claimed in any of claims 1 to 5 wherein said sterol is present in an amount of 1.0% to 20% of the total volume of said immunogenic composition.

7. An immunogenic composition as claimed in claim 6 wherein said alpha-tocopherol is present in an amount of 1.0% to 5.0% of the total volume of said immunogenic composition.

8. An immunogenic composition as claimed in any of claims 1 to 7 wherein said emulsifying agent is Tween 80.

9. An immunogenic composition as claimed in any of claims 1 to 8 wherein said emulsifying agent is present at an amount of 0.01 to 5.0% by weight (w/w) of said immunogenic composition.

10. An immunogenic composition as claimed in claim 9 wherein said emulsifying agent is present at an amount of 0.1 to 2.0% by weight (w/w) of said immunogenic composition.

11. An immunogenic composition as claimed in any of claims 1 to 10, wherein 3D-MPL is present at an amount of 10 to 50 μg (w/v) per composition dose.

12. An immunogenic composition as claimed in claim 11, wherein 3D-MPL is present at an amount of 25 μg (w/v) per composition dose.

13. An immunogenic composition as claimed in claims 11 to 12, wherein said composition dose is about 0.5 ml, between 0.5 and 1 ml or about 1 ml.

14. Use of a composition comprising:
(a) an antigen or antigenic composition
(b) an oil-in-water emulsion adjuvant; and
(c) 3D MPL
in the manufacture of an immunogenic composition as defined in any of claims 1 to 14 for the prevention of infection and/or disease associated with exposure to said antigen or a pathogen comprising said antigen or variant thereof.

15. A method of vaccination comprising delivery of an antigen or antigenic composition, 3D MPL and an oil-in-water emulsion adjuvant as defined in any of claims 1 to 10 to an individual or population in need thereof.

16. Use or method according to claims 14 or 15 for inducing at least one of i) an improved CD4 T-cell response, ii) an improved B cell memomry response, iii) an improved antibody response against said antigen.

17. Use or method according to any of claims 14 to 16 in an immunocompromised human individual or population.

18. Use or method according to any of claims 14 to 17 for protection against infection or disease caused by a pathogen which is a variant of the pathogen from which the antigen in the immunogenic composition is derived.

19. Use or method according to any of claims claims 14 to 18 for protection against infections or disease caused by a pathogen which comprises an antigen which is a variant of that antigen in the immunogenic composition.

20. Use or method according to claim 18 or 19 wherein protection is assessed by body temperature of the infected individual or population.

21. A composition, use or method according to any of claims 1 to 20 wherein the immunogenic composition comprises an antigen with a CD4 T cell epitope.

22. A composition, use or method according to any of claims 1 to 21 wherein the immunogenic composition comprises an antigen with a B cell epitope.

23. Use of an antigen in the manufacture of an immunogenic composition for revaccination of individuals previously vaccinated with the antigen or antigenic composition, or a fragment or variant thereof, 3D MPL and an oil-in-water emulsion adjuvant.

24. Use according to claim 23 wherein the antigen for revaccination shares common CD4 T-cell epitopes with an antigen or antigenic composition used for a previous vaccination.

25. Use according to claims 23 or 24 wherein the antigen or antigenic composition for revaccination is adjuvanted.

26. Use according to claim 25 wherein the adjuvant is an oil in water emulsion as defined in any of claims 1 to 10.

27. Use according to claim 25 or 26 wherein said adjuvant further comprises 3D-MPL.

28. A method for the preparation of an immunogenic composition comprising combining an oil in water emulsion as defined herein with an antigen or antigenic composition and 3D-MPL.

29. An immunogenic composition according to any of claims 1 to 14 in combination with a pharmaceutically acceptable carrier.

30. A composition comprising:

(a) an antigen

(b) an oil-in-water emulsion adjuvant as defined herein; and

(c) 3D MPL,

for use in medicine.

31. Composition, use or method according to any of claims 1 to 30 wherein the antigen or antigenic preparation is selected from the list consisting of: influenza virus, HPV.

32. Composition, use or method according to claim 31 wherein said influenza antigen is selected from the list consisting of: a split influenza virus, a whole influenza virus, a sub-unit influenza virus, an influenza virosome, and antigenic preparation thereof.

33. Composition, use or method according to claim 31 wherein said HPV antigen is associated with cancer or with genital warts.

34. Composition, use or method according to claim 33 wherein said cancer-associated HPV is HPV type 16 and/or HPV type 18.

35. Composition, use or method according to claim 34 wherein one or more additional antigens from cancer-causing HPV types are used with HPV 16 and/or 18 antigens, the antigens being selected from the following HPV types: HPV 31, 45, 33, 58 and 52.

36. Composition, use or method according to claim 34 or 35 wherein the antigens are in the form of virus like particles.