Nucleic acid comprising or coding for a histone stem-loop and a poly(A) sequence or a polyadenylation signal for increasing the expression of an encoded tumour antigen

The present invention relates to a nucleic acid sequence, comprising or coding for a coding region, encoding at least one peptide or protein comprising a tumour antigen or a fragment, variant or derivative thereof, at least one histone stem-loop and a poly(A) sequence or a polyadenylation signal. Furthermore the present invention provides the use of the nucleic acid for increasing the expression of said encoded peptide or protein. It also discloses its use for the preparation of a pharmaceutical composition, especially a vaccine, e.g. for use in the treatment of cancer or tumour diseases. The present invention further describes a method for increasing the expression of a peptide or protein comprising a tumour antigen or a fragment, variant or derivative thereof, using the nucleic acid comprising or coding for a histone stem-loop and a poly(A) sequence or a polyadenylation signal.

Apart from cardiovascular diseases and infectious diseases, the occurrence of tumours and cancer diseases is one of the most frequent causes of death in modern society and is in most cases associated with considerable costs in terms of therapy and subsequent rehabilitation measures. The treatment of tumours and cancer diseases is greatly dependent, for example, on the type of tumour that occurs, on the age, the distribution of cancer cells in the patient to be treated, etc. Cancer therapy is nowadays conventionally carried out by the use of radiation therapy or chemotherapy in addition to invasive operations. However, such conventional therapies typically place extraordinary stress on the immune system and can be used in some cases to only a limited extent. In addition, most of these conventional therapies require long intervals between the individual treatments to allow for regeneration of the immune system.

Therefore, supplementary strategies have been investigated in recent years in addition to such "conventional treatments" to avoid or at least reduce the impact on the immune system

by such treatments. One such supplementary treatment in particular includes gene therapeutic approaches or genetic vaccination, which already have been found to be highly promising for treatment or for supporting such conventional therapies.

Gene therapy and genetic vaccination are methods of molecular medicine which already have been proven in the therapy and prevention of diseases and generally exhibit a considerable effect on daily medical practice, in particular on the treatment of diseases as mentioned above. Both methods, gene therapy and genetic vaccination, are based on the introduction of nucleic acids into the patient's cells or tissue and subsequent processing of the information coded for by the nucleic acid that has been introduced into the cells or tissue, that is to say the (protein) expression of the desired polypeptides.

In gene therapy approaches, typically DNA is used even though RNA is also known in recent developments. Importantly, in all these gene therapy approaches mRNA functions as messenger for the sequence information of the encoded protein, irrespectively if DNA, viral RNA or mRNA is used.

In general RNA is considered an unstable molecule: RNases are ubiquitous and notoriously difficult to inactivate. Furthermore, RNA is also chemically more labile than DNA. Thus, it is perhaps surprising that the "default state" of an mRNA in a eukaryotic cell is characterized by a relative stability and specific signals are required to accelerate the decay of individual mRNAs. The main reason for this finding appears to be that mRNA decay within cells is catalyzed almost exclusively by exonucleases. However, the ends of eukaryotic mRNAs are protected against these enzymes by specific terminal structures and their associated proteins: a m7GpppN CAP at the 5′ end and typically a poly(A) sequence at the 3′ end. Removal of these two terminal modifications is thus considered rate limiting for mRNA decay. Although a stabilizing element has been characterized in the 3′ UTR of the alpha-globin mRNA, RNA sequences affecting turnover of eukaryotic mRNAs typically act as a promoter of decay usually by accelerating deadenylation (reviewed in Meyer, S., C. Temme, *et al.* (2004), Crit Rev Biochem Mol Biol 39(4): 197-216.).

As mentioned above, the 5′ ends of eukaryotic mRNAs are typically modified posttranscriptionally to carry a methylated CAP structure, e.g. m7GpppN. Aside from roles

in RNA splicing, stabilization, and transport, the CAP structure significantly enhances the recruitment of the 40S ribosomal subunit to the 5′ end of the mRNA during translation initiation. The latter function requires recognition of the CAP structure by the eukaryotic initiation factor complex eIF4F. The poly(A) sequence additionally stimulates translation via increased 40S subunit recruitment to mRNAs, an effect that requires the intervention of poly(A) binding protein (PABP). PABP, in turn, was recently demonstrated to interact physically with eIF4G, which is part of the CAP-bound eIF4F complex. Thus, a closed loop model of translation initiation on capped, polyadenylated mRNAs was postulated (Michel, Y. M., D. Poncet, *et al.* (2000), J Biol Chem 275(41): 32268-76.).

Nearly all eukaryotic mRNAs end with such a poly(A) sequence that is added to their 3′ end by the ubiquitous cleavage/polyadenylation machinery. The presence of a poly(A) sequence at the 3′ end is one of the most recognizable features of eukaryotic mRNAs. After cleavage, most pre-mRNAs, with the exception of replication-dependent histone transcripts, acquire a polyadenylated tail. In this context, 3′ end processing is a nuclear co-transcriptional process that promotes transport of mRNAs from the nucleus to the cytoplasm and affects the stability and the translation of mRNAs. Formation of this 3′ end occurs in a two step reaction directed by the cleavage/polyadenylation machinery and depends on the presence of two sequence elements in mRNA precursors (pre-mRNAs); a highly conserved hexanucleotide AAUAAA (polyadenylation signal) and a downstream G/U-rich sequence. In a first step, pre-mRNAs are cleaved between these two elements. In a second step tightly coupled to the first step the newly formed 3′ end is extended by addition of a poly(A) sequence consisting of 200–250 adenylates which affects subsequently all aspects of mRNA metabolism, including mRNA export, stability and translation (Dominski, Z. and W. F. Marzluff (2007), Gene 396(2): 373-90.).

The only known exception to this rule are the replication-dependent histone mRNAs which end with a histone stem–loop instead of a poly(A) sequence. Exemplary histone stem-loop sequences are described in Lopez *et al.* (Dávila López, M., & Samuelsson, T. (2008), RNA (New York, N.Y.), 14(1), 1-10. doi:10.1261/rna.782308.).

The stem-loops in histone pre-mRNAs are typically followed by a purine-rich sequence known as the histone downstream element (HDE). These pre-mRNAs are processed in the

nucleus by a single endonucleolytic cleavage approximately 5 nucleotides downstream of the stem-loop, catalyzed by the U7 snRNP through base pairing of the U7 snRNA with the HDE. The 3'-UTR sequence comprising the histone stem-loop structure and the histone downstream element (HDE) (binding site of the U7 snRNP) were usually termed as histone 3'-processing signal (see e.g. Chodchoy, N., N. B. Pandey, et al. (1991). Mol Cell Biol 11(1): 497-509.).

Due to the requirement to package newly synthesized DNA into chromatin, histone synthesis is regulated in concert with the cell cycle. Increased synthesis of histone proteins during S phase is achieved by transcriptional activation of histone genes as well as posttranscriptional regulation of histone mRNA levels. It could be shown that the histone stem-loop is essential for all posttranscriptional steps of histone expression regulation. It is necessary for efficient processing, export of the mRNA into the cytoplasm, loading onto polyribosomes, and regulation of mRNA stability.

In the above context, a 32 kDa protein was identified, which is associated with the histone stem-loop at the 3'-end of the histone messages in both the nucleus and the cytoplasm. The expression level of this stem-loop binding protein (SLBP) is cell-cycle regulated and is highest during S-phase when histone mRNA levels are increased. SLBP is necessary for efficient 3'-end processing of histone pre-mRNA by the U7 snRNP. After completion of processing, SLBP remains associated with the stem–loop at the end of mature histone mRNAs and stimulates their translation into histone proteins in the cytoplasm. (Dominski, Z. and W. F. Marzluff (2007), Gene 396(2): 373-90). Interestingly, the RNA binding domain of SLBP is conserved throughout metazoa and protozoa (Dávila López, M., & Samuelsson, T. (2008), RNA (New York, N.Y.), 14(1), 1-10. doi:10.1261/rna.782308) and it could be shown that its binding to the histone stem-loop sequence is dependent on the stem-loop structure and that the minimum binding site contains at least 3 nucleotides 5' and 2 nucleotides 3' of the stem-loop (Pandey, N. B., *et al.* (1994), *Molecular and Cellular Biology, 14*(3), 1709-1720 and Williams, A. S., & Marzluff, W. F., (1995), *Nucleic Acids Research, 23*(4), 654-662.).

Even though histone genes are generally classified as either "replication-dependent", giving rise to mRNA ending in a histone stem-loop, or "replacement-type", giving rise to mRNA

bearing a poly(A)-tail instead, naturally occurring mRNAs containing both a histone stem-loop and poly(A) or oligo(A) 3' thereof have been identified in some very rare cases. Sanchez *et al.* examined the effect of naturally occurring oligo(A) tails appended 3' of the histone stem-loop of histone mRNA during Xenopus oogenesis using Luciferase as a reporter protein and found that the oligo(A) tail is an active part of the translation repression mechanism that silences histone mRNA during oogenesis and its removal is part of the mechanism that activates translation of histone mRNAs (Sanchez, R. and W. F. Marzluff (2004), Mol Cell Biol 24(6): 2513-25).

Furthermore, the requirements for regulation of replication dependent histones at the level of pre-mRNA processing and mRNA stability have been investigated using artificial constructs coding for the marker protein alpha Globin, taking advantage of the fact that the globin gene contains introns as opposed to the intron-less histone genes. For this purpose constructs were generated in which the alpha globin coding sequence was followed by a histone stem-loop signal (histone stem-loop followed by the histone downstream element) and a polyadenylation signal (Whitelaw, E., *et al.* (1986). Nucleic Acids Research, 14(17), 7059-7070.; Pandey, N. B., & Marzluff, W. F. (1987). Molecular and Cellular Biology, 7(12), 4557-4559.; Pandey, N. B., *et al.* (1990). Nucleic Acids Research, 18(11), 3161-3170).

In another approach Lüscher *et al.* investigated the cell-cycle dependent regulation of a recombinant histone H4 gene. Constructs were generated in which the H4 coding sequence was followed by a histone stem-loop signal and a polyadenylation signal, the two processing signals incidentally separated by a galactokinase coding sequence (Lüscher, B. *et al.*, (1985). Proc. Natl. Acad. Sci. USA, 82(13), 4389-4393).

Additionally, Stauber *et al.* identified the minimal sequence required to confer cell-cycle regulation on histone H4 mRNA levels. For these investigations constructs were used, comprising a coding sequence for the selection marker Xanthine:guanine phosphoribosyl transferase (GPT) preceding a histone stem-loop signal followed by a polyadenylation signal (Stauber, C. *et al.*, (1986). EMBO J, 5(12), 3297-3303).

Examining histone pre-mRNA processing Wagner *et al.* identified factors required for cleavage of histone pre-mRNAs using a reporter construct placing EGFP between a histone stem-loop signal and a polyadenylation signal, such that EGFP was expressed only in case histone pre-mRNA processing was disrupted (Wagner, E. J. *et al.,* (2007). Mol Cell 28(4), 692-9).

To be noted, translation of polyadenylated mRNA usually requires the 3' poly(A) sequence to be brought into proximity of the 5' CAP. This is mediated through protein–protein interaction between the poly(A) binding protein and eukaryotic initiation factor eIF4G. With respect to replication-dependent histone mRNAs, an analogous mechanism has been uncovered. In this context, Gallie *et al.* show that the histone stem-loop is functionally similar to a poly(A) sequence in that it enhances translational efficiency and is co-dependent on a 5'-CAP in order to establish an efficient level of translation. They showed that the histone stem-loop is sufficient and necessary to increase the translation of a reporter mRNA in transfected Chinese hamster ovary cells but must be positioned at the 3'-terminus in order to function optimally. Therefore, similar to the poly(A) tail on other mRNAs, the 3' end of these histone mRNAs appears to be essential for translation *in vivo* and is functionally analogous to a poly(A) tail (Gallie, D. R., Lewis, N. J., & Marzluff, W. F. (1996), Nucleic Acids Research, 24(10), 1954-1962).

Additionally, it could be shown that SLBP is bound to the cytoplasmic histone mRNA and is required for its translation. Even though SLBP does not interact directly with eIF4G, the domain required for translation of histone mRNA interacts with the recently identified protein SLIP1. In a further step, SLIP1 interacts with eIF4G and allows to circularize histone mRNA and to support efficient translation of histone mRNA by a mechanism similar to the translation of polyadenylated mRNAs.

As mentioned above, gene therapy approaches normally use DNA to transfer the coding information into the cell which is then transcribed into mRNA, carrying the naturally occurring elements of an mRNA, particularly the 5'-CAP structure and the 3' poly(A) sequence to ensure expression of the encoded therapeutic or antigenic protein.

However, in many cases expression systems based on the introduction of such nucleic acids into the patient's cells or tissue and the subsequent expression of the desired polypeptides coded for by these nucleic acids do not exhibit the desired, or even the required, level of expression which may allow for an efficient therapy, irrespective as to whether DNA or RNA is used.

In the prior art, different attempts have hitherto been made to increase the yield of the expression of an encoded protein, in particular by use of improved expression systems, both *in vitro* and/or *in vivo*. Methods for increasing expression described generally in the prior art are conventionally based on the use of expression vectors or cassettes containing specific promoters and corresponding regulation elements. As these expression vectors or cassettes are typically limited to particular cell systems, these expression systems have to be adapted for use in different cell systems. Such adapted expression vectors or cassettes are then usually transfected into the cells and typically treated in dependence of the specific cell line. Therefore, preference is given primarily to those nucleic acid molecules which are able to express the encoded proteins in a target cell by systems inherent in the cell, independent of promoters and regulation elements which are specific for particular cell types. In this context, there can be distinguished between mRNA stabilizing elements and elements which increase translation efficiency of the mRNA.

mRNAs which are optimized in their coding sequence and which are in general suitable for such a purpose are described in application WO 02/098443 (CureVac GmbH). For example, WO 02/098443 describes mRNAs that are stabilised in general form and optimised for translation in their coding regions. WO 02/098443 further discloses a method for determining sequence modifications. WO 02/098443 additionally describes possibilities for substituting adenine and uracil nucleotides in mRNA sequences in order to increase the guanine/cytosine (G/C) content of the sequences. According to WO 02/098443, such substitutions and adaptations for increasing the G/C content can be used for gene therapeutic applications but also genetic vaccines in the treatment of cancer or infectious diseases. In this context, WO 02/098443 generally mentions sequences as a base sequence for such modifications, in which the modified mRNA codes for at least one biologically active peptide or polypeptide, which is translated in the patient to be treated, for example, either not at all or inadequately or with faults. Alternatively, WO 02/098443 proposes

mRNAs coding for antigens e.g. tumour antigens or viral antigens as a base sequence for such modifications.

In a further approach to increase the expression of an encoded protein the application WO 2007/036366 describes the positive effect of long poly(A) sequences (particularly longer than 120 bp) and the combination of at least two 3' untranslated regions of the beta globin gene on mRNA stability and translational activity.

However, even though all these latter prior art documents already try to provide quite efficient tools for gene therapy approaches and additionally improved mRNA stability and translational activity, there still remains the problem of a generally lower stability of RNA-based applications versus DNA vaccines and DNA based gene therapeutic approaches. Accordingly, there still exists a need in the art to provide improved tools for gene therapy approaches and genetic vaccination or as a supplementary therapy for conventional treatments as discussed above, which allow for better provision of encoded proteins *in vivo*, e.g. via a further improved mRNA stability and/or translational activity, preferably for gene therapy and genetic vaccination.

Furthermore despite of all progress in the art, efficient expression of an encoded peptide or protein in cell-free systems, cells or organisms (recombinant expression) is still a challenging problem.

The object underlying the present invention is, therefore, to provide additional and/or alternative methods to increase expression of an encoded protein, preferably via further stabilization of the mRNA and/or an increase of the translational efficiency of such an mRNA with respect to such nucleic acids known from the prior art for the use in genetic vaccination in the therapeutic or prophylactic treatment of cancer or tumour diseases.

This object is solved by the subject matter of the attached claims. Particularly, the object underlying the present invention is solved according to a first aspect by an inventive nucleic acid sequence comprising or coding for

a) a coding region, encoding at least one peptide or protein which comprises a tumour antigen or a fragment, variant or derivative thereof;

b) at least one histone stem-loop, and

c) a poly(A) sequence or a polyadenylation signal,

preferably for increasing the expression of said encoded peptide or protein.

Alternatively, any appropriate stem loop sequence other than a histone stem loop sequence (derived from histone genes, in particular histone genes of the families H1, H2A, H2B, H3 and H4)) may be employed by the present invention in all of its aspects and embodiments.

In this context it is particularly preferred that the inventive nucleic acid according to the first aspect of the present invention is produced at least partially by DNA or RNA synthesis, preferably as described herein or is an isolated nucleic acid.

The present invention is based on the surprising finding of the present inventors, that the combination of a poly(A) sequence or polyadenylation signal and at least one histone stem-loop, even though both representing alternative mechanisms in nature, acts synergistically as this combination increases the protein expression manifold above the level observed with either of the individual elements. The synergistic effect of the combination of poly(A) and at least one histone stem-loop is seen irrespective of the order of poly(A) and histone stem-loop and irrespective of the length of the poly(A) sequence.

Therefore it is particularly preferred that the inventive nucleic acid sequence comprises or codes for a) a coding region, encoding at least one peptide or protein which comprises a tumour antigen or a fragment, variant or derivative thereof; b) at least one histone stem-loop, and c) a poly(A) sequence or polyadenylation sequence; preferably for increasing the expression level of said encoded peptide or protein. In some embodiments, it may be preferred if the encoded protein is not a histone protein, in particular no histone protein of the H4, H3, H2A and/or H2B histone family or a fragment, derivative or variant thereof retaining histone(-like) function), namely forming a nucleosome. Also, the encoded protein typically does not correspond to a histone linker protein of the H1 histone family. The inventive nucleic acid molecule does typically not contain any regulatory signals (5' and/or, particularly, 3' of a mouse histone gene, in particular not of a mouse histone gene H2A and, further, most preferably not of the mouse histone gene H2A614. In particular, it does not contain a histone stem loop and/or a histone stem loop processing signal from a mouse

histone gene, in particular not of mouse histone gene H2A und, most preferably not of mouse histone gene H2A614.

Also, the inventive nucleic acid typically does not provide a reporter protein (e.g. Luciferase, GFP, EGFP, β-Galactosidase, particularly EGFP) galactokinase (galK) and/or marker or selection protein (e.g. alpha-Globin, galactokinase and Xanthine:Guanine phosphoribosyl transferase (GPT)) or a bacterial reporter protein, e.g. chloramphenicol acetyl transferase (CAT) or other bacterial antibiotics resistance proteins, e.g. derived from the bacterial *neo* gene in its element (a).

A reporter, marker or selection protein is typically understood not to be a tumour antigen according to the invention. A reporter, marker or selection protein or its underlying gene is commonly used as a research tool in bacteria, cell culture, animals or plants. They confer on organisms (preferably heterologously) expressing them an easily identifiable property, which may be measured or which allows for selection. Specifically, marker or selection proteins exhibit a selectable function. Typically, such selection, marker or reporter proteins do not naturally occur in humans or other mammals, but are derived from other organisms, in particular from bacteria or plants. Accordingly, proteins with selection, marker or reporter function originating from species other than mammals, in particular other than humans, are preferably excluded from being understood as a protein having the property to act as a "tumour antigen" according to the present invention. In particular, a selection, marker or reporter protein allows to identify transformed cells by in vitro assays based e.g. on fluorescence or other spectroscopic techniques and resistance towards antibiotics. Selection, reporter or marker genes awarding such properties to transformed cells are therefore typically not understood to be a protein acting as a tumour antigen according to the invention in vivo.

In any case, reporter, marker or selection proteins do usually not exert any tumour antigenic properties and, therefore, do not exert an immunological effect which therapeutically allows to treat tumour diseases. If any single reporter, marker or selection protein should nevertheless do so (in addition to its reporter, selection or marker function), such a reporter, marker or selection protein is preferably not understood to be a "tumour antigen" within the meaning of the present invention.

In contrast, a protein or peptide acting as a tumour antigen (in particular excluding histone genes of the families H1, H2A, H2B, H3 and H4) according to the present invention does typically not exhibit a selection, marker or reporter function. If any single "tumour antigen" nevertheless should do so (in addition to its tumour antigenic function), such a tumour antigen is preferably not understood to be a "selection, marker or reporter protein" within the meaning of the present invention.

It is most preferably understood that a protein acting as tumour antigen according to the invention is derived from mammals, in particular humans, in particular from mammalian tumours, and does not qualify as selection, marker or reporter protein. In particular, such tumour antigens are derived from mammalian, in particular from human tumours. These tumour antigenic proteins are understood to be antigenic, as they are meant to treat the subject by triggering the subject's immune response such that the subject's immune system is enabled to combat the subject's tumor cells by TH1 and/or TH2 immune responses. Accordingly, such antigenic tumour proteins are typically mammalian, in particular human proteins characterizing the subject's cancer type.

Accordingly, it is preferred that the coding region (a) encoding at least one peptide or protein is heterologous to at least (b) the at least one histone stem loop, or more broadly, to any appropriate stem loop. In other words, "heterologous" in the context of the present invention means that the at least one stem loop sequence does not naturally occur as a (regulatory) sequence (e.g. at the 3'UTR) of the specific gene, which encodes the (tumour antigenic) protein or peptide of element (a) of the inventive nucleic acid. Accordingly, the (histone) stem loop of the inventive nucleic acid is derived preferably from the 3' UTR of a gene other than the one comprising the coding region of element (a) of the inventive nucleic acid. E.g., the coding region of element (a) will not encode a histone protein or a fragment, variant or derivative thereof (retaining the function of a histone protein), if the inventive nucleic is heterologous, but will encode any other peptide or sequence (of the same or another species) which exerts a biological function, preferably tumour antigenic function other than a histone(-like) function, e.g. will encode a protein (exerting a tumour antigenic function, e.g. in terms of vaccinating against mammalian, in particular human tumours thereby triggering a immunological reaction against the subject's tumour cells, which preferably express the tumour antigen encoded by the inventive nucleic acid.

In this context it is particularly preferred that the inventive nucleic acid comprises or codes for in 5'- to 3'-direction:

a) a coding region, encoding at least one peptide or protein which comprises a tumour antigen or a fragment, variant or derivative thereof;

b) *at least one histone stem-loop, optionally without a histone downstream element* (HDE) 3' to the histone stem-loop

c) a poly(A) sequence or a polyadenylation signal.

The term "histone downstream element (HDE) refers to a purine-rich polynucleotide stretch of about 15 to 20 nucleotides 3' of naturally occurring histone stem-loops, which represents the binding site for the U7 snRNA involved in processing of histone pre-mRNA into mature histone mRNA. For example in sea urchins the HDE is CAAGAAAGA (Dominski, Z. and W. F. Marzluff (2007), Gene 396(2): 373-90).

Furthermore it is preferable that the inventive nucleic acid according to the first aspect of the present invention does not comprise an intron.

In another particular preferred embodiment, the inventive nucleic acid sequence according to the first aspect of the present invention comprises or codes for from 5' to 3':

a) a coding region, preferably encoding at least one peptide or protein which comprises a tumour antigen or a fragment, variant or derivative thereof;

c) a poly(A) sequence; and

b) at least one histone stem-loop.

The inventive nucleic acid sequence according to the first embodiment of the present invention comprise any suitable nucleic acid, selected e.g. from any (single-stranded or double-stranded) DNA, preferably, without being limited thereto, e.g. genomic DNA, plasmid DNA, single-stranded DNA molecules, double-stranded DNA molecules, or may be selected e.g. from any PNA (peptide nucleic acid) or may be selected e.g. from any (single-stranded or double-stranded) RNA, preferably a messenger RNA (mRNA); etc. The inventive nucleic acid sequence may also comprise a viral RNA (vRNA). However, the

inventive nucleic acid sequence may not be a viral RNA or may not contain a viral RNA. More specifically, the inventive nucleic acid sequence may not contain viral sequence elements, e.g. viral enhancers or viral promotors (e.g. no inactivated viral promoter or sequence elements, more specifically not inactivated by replacement strategies), or other viral sequence elements, or viral or retroviral nucleic acid sequences. More specifically, the inventive nucleic acid sequence may not be a retroviral or viral vector or a modified retroviral or viral vector.

In any case, the inventive nucleic acid sequence may or may not contain an enhancer and/or promoter sequence, which may be modified or not or which may be activated or not. The enhancer and or promoter may be plant expressible or not expressible, and/or in eukaryotes expressible or not expressible and/or in prokaryotes expressible or not expressible. The inventive nucleic acid sequence may contain a sequence encoding a (self-splicing) ribozyme or not.

In specific embodiments the inventive nucleic acid sequence may be or may comprise a self-replicating RNA (replicon).

Preferably, the inventive nucleic acid sequence is a plasmid DNA, or an RNA, particularly an mRNA.

In particular embodiments of the first aspect of the present invention, the inventive nucleic acid is a nucleic acid sequence comprised in a nucleic acid suitable for *in vitro* transcription, particularly in an appropriate *in vitro* transcription vector (e.g. a plasmid or a linear nucleic acid sequence comprising specific promoters for *in vitro* transcription such as T3, T7 or Sp6 promoters).

In further particular preferred embodiments of the first aspect of the present invention, the inventive nucleic acid is comprised in a nucleic acid suitable for transcription and/or translation in an expression system (e.g. in an expression vector or plasmid), particularly a prokaryotic (e.g. bacteria like *E. coli*) or eukaryotic (e.g. mammalian cells like CHO cells, yeast cells or insect cells or whole organisms like plants or animals) expression system.

The term "expression system" means a system (cell culture or whole organisms) which is suitable for production of peptides, proteins or RNA particularly mRNA (recombinant expression).

The inventive nucleic acid sequence according to the first aspect of the present invention comprises or codes for at least one histone stem-loop. In the context of the present invention, such a histone stem-loop , in general (irrespective of whether it is a histone stem loop or not),is typically derived from histone genes and comprises an intramolecular base pairing of two neighbored entirely or partially reverse complementary sequences, thereby forming a stem-loop. A stem-loop can occur in single-stranded DNA or, more commonly, in RNA. The structure is also known as a hairpin or hairpin loop and usually consists of a stem and a (terminal) loop within a consecutive sequence, wherein the stem is formed by two neighbored entirely or partially reverse complementary sequences separated by a short sequence as sort of spacer, which builds the loop of the stem-loop structure. The two neighbored entirely or partially reverse complementary sequences may be defined as e.g. stem loop elements stem1 and stem2. The stem loop is formed when these two neighbored entirely or partially reverse complementary sequences, e.g. stem loop elements stem1 and stem2, form base-pairs with each other, leading to a double stranded nucleic acid sequence stretch comprising an unpaired loop at its terminal ending formed by the short sequence located between stem loop elements stem1 and stem2 on the consecutive sequence. The unpaired loop thereby typically represents a region of the nucleic acid which is not capable of base pairing with either of these stem loop elements. The resulting lollipop-shaped structure is a key building block of many RNA secondary structures. The formation of a stem-loop structure is thus dependent on the stability of the resulting stem and loop regions, wherein the first prerequisite is typically the presence of a sequence that can fold back on itself to form a paired double strand. The stability of paired stem loop elements is determined by the length, the number of mismatches or bulges it contains (a small number of mismatches is typically tolerable, especially in a long double stranded stretch), and the base composition of the paired region. In the context of the present invention, a loop length of 3 to 15 bases is conceivable, while a more preferred loop length is 3-10 bases, more preferably 3 to 8, 3 to 7, 3 to 6 or even more preferably 4 to 5 bases, and most preferably 4 bases. The stem sequence forming the double stranded structure typically has a length of between 5 to 10 bases, more preferably, between 5 to 8 bases.

In the context of the present invention, a histone stem-loop is typically derived from histone genes (e.g. genes from the histone families H1, H2A, H2B, H3, H4) and comprises an intramolecular base pairing of two neighbored entirely or partially reverse complementary sequences, thereby forming a stem-loop. Typically, a histone 3' UTR stem-loop is an RNA element involved in nucleocytoplasmic transport of the histone mRNAs, and in the regulation of stability and of translation efficiency in the cytoplasm. The mRNAs of metazoan histone genes lack polyadenylation and a poly-A tail, instead 3' end processing occurs at a site between this highly conserved stem-loop and a purine rich region around 20 nucleotides downstream (the histone downstream element, or HDE). The histone stem-loop is bound by a 31 kDa stem-loop binding protein (SLBP - also termed the histone hairpin binding protein, or HBP). Such histone stem-loop structures are preferably employed by the present invention in combination with other sequence elements and structures, which do not occur naturally (which means in untransformed living organisms/cells) in histone genes, but are combined – according to the invention – to provide an artificial, heterologous nucleic acid. Accordingly, the present invention is particularly based on the finding that an artificial (non-native) combination of a histone stem-loop structure with other heterologous sequence elements, which do not occur in histone genes or metazoan histone genes and are isolated from operational and/or regulatory sequence regions (influencing transcription and/or translation) of genes coding for proteins other than histones, provide advantageous effects. Accordingly, one aspect of the invention provides the combination of a histone stem-loop structure with a poly(A) sequence or a sequence representing a polyadenylation signal (3'-terminal of a coding region), which does not occur in metazoan histone genes. According to another preferred aspect of the invention, a combination of a histone stem-loop structure with a coding region coding for a tumour antigenic protein, which does, preferably not occur in metazoan histone genes, is provided herewith (coding region and histone stem loop sequence are heterologous). It is preferred, if such tumour antigenic proteins occur in mammalian, preferably humans, when suffering from tumour diseases. In a still further preferred embodiment, all the elements (a), (b) and (c) of the inventive nucleic acid are heterologous to each other and are combined artificially from three different sources, e.g. the (a) tumour antigenic protein coding region from a human gene, (b) the histone stem loop from the untranslated region of a metazoan, e.g. mammalian, non-human or human, histone gene and (c) the poly(A) sequence or the polyadenylation signal from e.g.

an untranslated region of a gene other than a histone gene and other than the untranslated region of the tumour antigen coding region according to element (a) of such an inventive nucleic acid.

A histone stem loop is, therefore, a stem-loop structure as described herein, which, if preferably functionally defined, exhibits/retains the property of binding to its natural binding partner, the stem-loop binding protein (SLBP - also termed the histone hairpin binding protein, or HBP).

According to the present invention the histone stem loop sequence according to component (b) of claim 1 may not derived from a mouse histone protein. More specifically, the histone stem loop sequence may not be derived from mouse histone gene H2A614. Also, the nucleic acid of the invention may neither contain a mouse histone stem loop sequence nor contain mouse histone gene H2A614. Further, the inventive nucleic acid sequence may not contain a stem-loop processing signal, more specifically, a mouse histone processing signal and, most specifically, may not contain mouse stem loop processing signal H2kA614, even if the inventive nucleic acid sequence may contain at least one mammalian histone gene. However, the at least one mammalian histone gene may not be Seq. ID No. 7 of WO 01/12824.

According to one preferred embodiment of the first inventive aspect, the inventive nucleic acid sequence comprises or codes for at least one histone stem-loop sequence, preferably according to at least one of the following formulae (I) or (II):

formula (I) (stem-loop sequence without stem bordering elements):

$$\underbrace{[N_{0-2}GN_{3-5}]}_{\text{stem1}}\ \underbrace{[N_{0-4}(U/T)N_{0-4}]}_{\text{loop}}\ \underbrace{[N_{3-5}CN_{0-2}]}_{\text{stem2}}$$

formula (II) (stem-loop sequence with stem bordering elements):

$$\underbrace{N_{1-6}}_{\substack{\text{stem1}\\\text{bordering element}}}\underbrace{[N_{0-2}GN_{3-5}]}_{\text{stem1}}\ \underbrace{[N_{0-4}(U/T)N_{0-4}]}_{\text{loop}}\ \underbrace{[N_{3-5}CN_{0-2}]}_{\text{stem2}}\underbrace{N_{1-6}}_{\substack{\text{stem2}\\\text{bordering element}}}$$

wherein:

stem1 or stem2 bordering elements N_{1-6} is a consecutive sequence of 1 to 6, preferably of 2 to 6, more preferably of 2 to 5, even more preferably of 3 to 5, most preferably of 4 to 5 or 5 N, wherein each N is independently from another selected from a nucleotide selected from A, U, T, G and C, or a nucleotide analogue thereof;

stem1 $[N_{0-2}GN_{3-5}]$ is reverse complementary or partially reverse complementary with element stem2, and is a consecutive sequence between of 5 to 7 nucleotides;

wherein N_{0-2} is a consecutive sequence of 0 to 2, preferably of 0 to 1, more preferably of 1 N, wherein each N is independently from another

	selected from a nucleotide selected from A, U, T, G and C or a nucleotide analogue thereof;
	wherein N_{3-5} is a consecutive sequence of 3 to 5, preferably of 4 to 5, more preferably of 4 N, wherein each N is independently from another selected from a nucleotide selected from A, U, T, G and C or a nucleotide analogue thereof, and
	wherein G is guanosine or an analogue thereof, and may be optionally replaced by a cytidine or an analogue thereof, provided that its complementary nucleotide cytidine in stem2 is replaced by guanosine;
loop sequence $[N_{0-4}(U/T)N_{0-4}]$	is located between elements stem1 and stem2, and is a consecutive sequence of 3 to 5 nucleotides, more preferably of 4 nucleotides;
	wherein each N_{0-4} is independent from another a consecutive sequence of 0 to 4, preferably of 1 to 3, more preferably of 1 to 2 N, wherein each N is independently from another selected from a nucleotide selected from A, U, T, G and C or a nucleotide analogue thereof; and
	wherein U/T represents uridine, or optionally thymidine;
stem2 $[N_{3-5}CN_{0-2}]$	is reverse complementary or partially reverse complementary with element stem1, and is a consecutive sequence between of 5 to 7 nucleotides;

wherein N_{3-5} is a consecutive sequence of 3 to 5, preferably of 4 to 5, more preferably of 4 N, wherein each N is independently from another selected from a nucleotide selected from A, U, T, G and C or a nucleotide analogue thereof;

wherein N_{0-2} is a consecutive sequence of 0 to 2, preferably of 0 to 1, more preferably of 1 N, wherein each N is independently from another selected from a nucleotide selected from A, U, T, G or C or a nucleotide analogue thereof; and

wherein C is cytidine or an analogue thereof, and may be optionally replaced by a guanosine or an analogue thereof provided that its complementary nucleotide guanosine in stem1 is replaced by cytidine;

wherein

stem1 and stem2 are capable of base pairing with each other forming a reverse complementary sequence, wherein base pairing may occur between stem1 and stem2, e.g. by Watson-Crick base pairing of nucleotides A and U/T or G and C or by non-Watson-Crick base pairing e.g. wobble base pairing, reverse Watson-Crick base pairing, Hoogsteen base pairing, reverse Hoogsteen base pairing or are capable of base pairing with each other forming a partially reverse complementary sequence, wherein an incomplete base pairing may occur between stem1 and stem2, on the basis that one or more bases in one stem do not have a complementary base in the reverse complementary sequence of the other stem.

In the above context, a wobble base pairing is typically a non-Watson-Crick base pairing between two nucleotides. The four main wobble base pairs in the present context, which

may be used, are guanosine-uridine, inosine-uridine, inosine-adenosine, inosine-cytidine (G-U/T, I-U/T, I-A and I-C) and adenosine-cytidine (A-C).

Accordingly, in the context of the present invention, a wobble base is a base, which forms a wobble base pair with a further base as described above. Therefore non-Watson-Crick base pairing, e.g. wobble base pairing, may occur in the stem of the histone stem-loop structure according to the present invention.

In the above context a partially reverse complementary sequence comprises maximally 2, preferably only one mismatch in the stem-structure of the stem-loop sequence formed by base pairing of stem1 and stem2. In other words, stem1 and stem2 are preferably capable of (full) base pairing with each other throughout the entire sequence of stem1 and stem2 (100% of possible correct Watson-Crick or non-Watson-Crick base pairings), thereby forming a reverse complementary sequence, wherein each base has its correct Watson-Crick or non-Watson-Crick base pendant as a complementary binding partner. Alternatively, stem1 and stem2 are preferably capable of partial base pairing with each other throughout the entire sequence of stem1 and stem2, wherein at least about 70%, 75%, 80%, 85%, 90%, or 95% of the 100% possible correct Watson-Crick or non-Watson-Crick base pairings are occupied with the correct Watson-Crick or non-Watson-Crick base pairings and at most about 30%, 25%, 20%, 15%, 10%, or 5% of the remaining bases are unpaired.

According to a preferred embodiment of the first inventive aspect, the at least one histone stem-loop sequence (with stem bordering elements) of the inventive nucleic acid sequence as defined herein comprises a length of about 15 to about 45 nucleotides, preferably a length of about 15 to about 40 nucleotides, preferably a length of about 15 to about 35 nucleotides, preferably a length of about 15 to about 30 nucleotides and even more preferably a length of about 20 to about 30 and most preferably a length of about 24 to about 28 nucleotides.

According to a further preferred embodiment of the first inventive aspect, the at least one histone stem-loop sequence (without stem bordering elements) of the inventive nucleic acid sequence as defined herein comprises a length of about 10 to about 30 nucleotides,

preferably a length of about 10 to about 20 nucleotides, preferably a length of about 12 to about 20 nucleotides, preferably a length of about 14 to about 20 nucleotides and even more preferably a length of about 16 to about 17 and most preferably a length of about 16 nucleotides.

According to a further preferred embodiment of the first inventive aspect, the inventive nucleic acid sequence according to the first aspect of the present invention may comprise or code for at least one histone stem-loop sequence according to at least one of the following specific formulae (Ia) or (IIa):

formula (Ia) (stem-loop sequence without stem bordering elements):

$$\underbrace{[N_{0-1}GN_{3-5}]}_{\text{stem1}}\ \underbrace{[N_{1-3}(U/T)N_{0-2}]}_{\text{loop}}\ \underbrace{[N_{3-5}CN_{0-1}]}_{\text{stem2}}$$

formula (IIa) (stem-loop sequence with stem bordering elements):

$$\underbrace{N_{2-5}}_{\substack{\text{stem1}\\ \text{bordering element}}}\ \underbrace{[N_{0-1}GN_{3-5}]}_{\text{stem1}}\ \underbrace{[N_{1-3}(U/T)N_{0-2}]}_{\text{loop}}\ \underbrace{[N_{3-5}CN_{0-1}]}_{\text{stem2}}\ \underbrace{N_{2-5}}_{\substack{\text{stem2}\\ \text{bordering element}}}$$

wherein:

N, C, G, T and U are as defined above.

According to a further more particularly preferred embodiment of the first aspect, the inventive nucleic acid sequence may comprise or code for at least one histone stem-loop sequence according to at least one of the following specific formulae (Ib) or (IIb):

formula (Ib) (stem-loop sequence without stem bordering elements):

$$\underbrace{[N_1GN_4]}_{\text{stem1}}\ \underbrace{[N_2(U/T)N_1]}_{\text{loop}}\ \underbrace{[N_4CN_1]}_{\text{stem2}}$$

formula (IIb) (stem-loop sequence with stem bordering elements):

$$\underbrace{N_{4-5}}_{\text{stem1 bordering element}}\ \underbrace{[N_1GN_4]}_{\text{stem1}}\ \underbrace{[N_2(U/T)N_1]}_{\text{loop}}\ \underbrace{[N_4CN_1]}_{\text{stem2}}\ \underbrace{N_{4-5}}_{\text{stem2 bordering element}}$$

wherein:

N, C, G, T and U are as defined above.

According to an even more preferred embodiment of the first inventive aspect, the inventive nucleic acid sequence according to the first aspect of the present invention may comprise or code for at least one histone stem-loop sequence according to at least one of the following specific formulae (Ic) to (Ih) or (IIc) to (IIh), shown alternatively in its stem-loop structure and as a linear sequence representing histone stem-loop sequences as generated according to Example 1:

formula (Ic): (metazoan and protozoan histone stem-loop consensus sequence without stem bordering elements):

N U
N N
N-N
N-N
N-N
N-N
G-C
N-N (stem-loop structure)

NGNNNNNNUNNNNNCN
(linear sequence) (SEQ ID NO: 1)

formula (IIc): (metazoan and protozoan histone stem-loop consensus sequence with stem bordering elements):

N U
N N
N-N
N-N
N-N
N-N
G-C
N*N*NNNN-NNNN*N*N* (stem-loop structure)

N*N*NNNNGNNNNNNUNNNNNCNNNN*N*N*
(linear sequence) (SEQ ID NO: 2)

formula (Id): (without stem bordering elements)

N U
N N
N-N
N-N
N-N
N-N
C-G
N-N (stem-loop structure)

NCNNNNNNUNNNNNGN
(linear sequence) (SEQ ID NO: 3)

formula (IId): (with stem bordering elements)

N U
N N
N-N
N-N
N-N
N-N
C-G
N*N*NNNN-NNNN*N*N* (stem-loop structure)

N*N*NNNNCNNNNNNUNNNNNGNNNN*N*N*
(linear sequence) (SEQ ID NO: 4)

formula (Ie): (protozoan histone stem-loop consensus sequence without stem bordering elements)

N U
N N
N-N
N-N
N-N
N-N
G-C
D-H (stem-loop structure)

DGNNNNNNUNNNNNCH
(linear sequence) (SEQ ID NO: 5)

formula (IIe): (protozoan histone stem-loop consensus sequence with stem bordering elements)

N U
N N
N-N
N-N
N-N
N-N
G-C
N*N*NNND-HNNN*N*N* (stem-loop structure)

N*N*NNNDGNNNNNNUNNNNNCHNNN*N*N*
(linear sequence) (SEQ ID NO: 6)

formula (If): (metazoan histone stem-loop consensus sequence without stem bordering elements)

N U
N N
Y-V
Y-N
B-D
N-N
G-C
N-N (stem-loop structure)

NGNBYYNNUNVNDNCN
(linear sequence) (SEQ ID NO: 7)

formula (IIf): (metazoan histone stem-loop consensus sequence with stem bordering elements)

N U
N N
Y-V
Y-N
B-D
N-N
G-C
N*N*NNNN-NNNN*N*N* (stem-loop structure)

N*N*NNNNGNBYYNNUNVNDNCNNNN*N*N*
(linear sequence) (SEQ ID NO: 8)

formula (Ig): (vertebrate histone stem-loop consensus sequence without stem bordering elements)

N U
D H
Y-A
Y-B
Y-R
H-D
G-C
N-N (stem-loop structure)

NGHYYYDNUHABRDCN
(linear sequence) (SEQ ID NO: 9)

formula (IIg): (vertebrate histone stem-loop consensus sequence with stem bordering elements)

N U
D H
Y-A
Y-B
Y-R
H-D
G-C
N*N*HNNN-NNNN*N*H* (stem-loop structure)

N*N*HNNNGHYYYDNUHABRDCNNNN*N*H*
(linear sequence) (SEQ ID NO: 10)

formula (Ih): (human histone stem-loop consensus sequence (Homo sapiens) without stem bordering elements)

Y U
D H
U-A
C-S
Y-R
H-R
G-C
D-C (stem-loop structure)

DGHYCUDYUHASRRCC
(linear sequence) (SEQ ID NO: 11)

formula (IIh): (human histone stem-loop consensus sequence (Homo sapiens) with stem bordering elements)

Y U
D H
U-A
C-S
Y-R
H-R
G-C
N*H*AAHD-CVHB*N*H* (stem loop structure)

N*H*AAHDGHYCUDYUHASRRCCVHB*N*H*
(linear sequence) (SEQ ID NO: 12)

wherein in each of above formulae (Ic) to (Ih) or (IIc) to (IIh):

N, C, G, A, T and U are as defined above;

each U may be replaced by T;

each (highly) conserved G or C in the stem elements 1 and 2 may be replaced by its complementary nucleotide base C or G, provided that its complementary nucleotide in the corresponding stem is replaced by its complementary nucleotide in parallel; and/or

G, A, T, U, C, R, Y, M, K, S, W, H, B, V, D, and N are nucleotide bases as defined in the following Table:

abbreviation	Nucleotide bases	remark
G	G	Guanine
A	A	Adenine
T	T	Thymine
U	U	Uracile
C	C	Cytosine
R	G or A	Purine
Y	T/U or C	Pyrimidine
M	A or C	Amino
K	G or T/U	Keto
S	G or C	Strong (3H bonds)
W	A or T/U	Weak (2H bonds)
H	A or C or T/U	Not G
B	G or T/U or C	Not A
V	G or C or A	Not T/U
D	G or A or T/U	Not C
N	G or C or T/U or A	Any base
*	Present or not	Base may be present or not

In this context it is particularly preferred that the histone stem-loop sequence according to at least one of the formulae (I) or (Ia) to (Ih) or (II) or (IIa) to (IIh) of the present invention is selected from a naturally occurring histone stem loop sequence, more particularly preferred from protozoan or metazoan histone stem-loop sequences, and even more particularly preferred from vertebrate and mostly preferred from mammalian histone stem-loop sequences especially from human histone stem-loop sequences.

According to a particularly preferred embodiment of the first aspect, the histone stem-loop sequence according to at least one of the specific formulae (I) or (Ia) to (Ih) or (II) or (IIa) to (IIh) of the present invention is a histone stem-loop sequence comprising at each nucleotide position the most frequently occurring nucleotide, or either the most frequently or the second-most frequently occurring nucleotide of naturally occurring histone stem-loop sequences in metazoa and protozoa (Fig. 1), protozoa (Fig. 2), metazoa (Fig. 3), vertebrates (Fig. 4) and humans (Fig. 5) as shown in figure 1-5. In this context it is particularly preferred that at least 80%, preferably at least 85%, or most preferably at least 90% of all nucleotides correspond to the most frequently occurring nucleotide of naturally occurring histone stem-loop sequences.

In a further particular embodiment of the first aspect, the histone stem-loop sequence according to at least one of the specific formulae (I) or (Ia) to (Ih) of the present invention is selected from following histone stem-loop sequences (without stem-bordering elements) representing histone stem-loop sequences as generated according to Example 1:

VGYYYYHHTHRVVRCB (SEQ ID NO: 13 according to formula (Ic))
SGYYYTTYTMARRRCS (SEQ ID NO: 14 according to formula (Ic))
SGYYCTTTTMAGRRCS (SEQ ID NO: 15 according to formula (Ic))

DGNNNBNNTHVNNNCH (SEQ ID NO: 16 according to formula (Ie))
RGNNNYHBTHRDNNCY (SEQ ID NO: 17 according to formula (Ie))
RGNDBYHYTHRDHNCY (SEQ ID NO: 18 according to formula (Ie))

VGYYYTYHTHRVRRCB (SEQ ID NO: 19 according to formula (If))
SGYYCTTYTMAGRRCS (SEQ ID NO: 20 according to formula (If))
SGYYCTTTTMAGRRCS (SEQ ID NO: 21 according to formula (If))

GGYYCTTYTHAGRRCC (SEQ ID NO: 22 according to formula (Ig))
GGCYCTTYTMAGRGCC (SEQ ID NO: 23 according to formula (Ig))
GGCTCTTTTMAGRGCC (SEQ ID NO: 24 according to formula (Ig))

DGHYCTDYTHASRRCC (SEQ ID NO: 25 according to formula (Ih))
GGCYCTTTTHAGRGCC (SEQ ID NO: 26 according to formula (Ih))
GGCYCTTTTMAGRGCC (SEQ ID NO: 27 according to formula (Ih))

Furthermore in this context following histone stem-loop sequences (with stem bordering elements) as generated according to Example 1 according to one of specific formulae (II) or (IIa) to (IIh) are particularly preferred:

H*H*HHVVGYYYYHHTHRVVRCBVHH*N*N* (SEQ ID NO: 28 according to formula (IIc))
M*H*MHMSGYYYTTYTMARRRCSMCH*H*H* (SEQ ID NO: 29 according to formula (IIc))
M*M*MMMSGYYCTTTTMAGRRCSACH*M*H* (SEQ ID NO: 30 according to formula (IIc))

N*N*NNNDGNNNBNNTHVNNNCHNHN*N*N* (SEQ ID NO: 31 according to formula (IIe))
N*N*HHNRGNNNYHBTHRDNNCYDHH*N*N* (SEQ ID NO: 32 according to formula (IIe))
N*H*HHVRGNDBYHYTHRDHNCYRHH*H*H* (SEQ ID NO: 33 according to formula (IIe))

H*H*MHMVGYYYTYHTHRVRRCBVMH*H*N* (SEQ ID NO: 34 according to formula (IIf))
M*M*MMMSGYYCTTYTMAGRRCSMCH*H*H* (SEQ ID NO: 35 according to formula (IIf))
M*M*MMMSGYYCTTTTMAGRRCSACH*M*H* (SEQ ID NO: 36 according to formula (IIf))

H*H*MAMGGYYCTTYTHAGRRCCVHN*N*M* (SEQ ID NO: 37 according to formula (IIg))
H*H*AAMGGCYCTTYTMAGRGCCVCH*H*M* (SEQ ID NO: 38 according to formula (IIg))
M*M*AAMGGCTCTTTTMAGRGCCMCY*M*M* (SEQ ID NO: 39 according to formula (IIg))

N*H*AAHDGHYCTDYTHASRRCCVHB*N*H* (SEQ ID NO: 40 according to formula (IIh))
H*H*AAMGGCYCTTTTHAGRGCCVMY*N*M* (SEQ ID NO: 41 according to formula (IIh))
H*M*AAAGGCYCTTTTMAGRGCCRMY*H*M* (SEQ ID NO: 42 according to formula (IIh))

According to a further preferred embodiment of the first inventive aspect, the inventive nucleic acid sequence comprises or codes for at least one histone stem-loop sequence showing at least about 80%, preferably at least about 85%, more preferably at least about 90%, or even more preferably at least about 95%, sequence identity with the not to 100% conserved nucleotides in the histone stem-loop sequences according to at least one of specific formulae (I) or (Ia) to (Ih) or (II) or (IIa) to (IIh) or with a naturally occurring histone stem-loop sequence.

In a preferred embodiment, the histone stem loop sequence does not contain the loop sequence 5'-UUUC-3'. More specifically, the histone stem loop does not contain the stem1 sequence 5'-GGCUCU-3' and/or the stem2 sequence 5'-AGAGCC-3', respectively. In another preferred embodiment, the stem loop sequence does not contain the loop sequence 5'-CCUGCCC-3' or the loop sequence 5'-UGAAU-3'. More specifically, the stem loop does not contain the stem1 sequence 5'-CCUGAGC-3' or does not contain the stem1 sequence 5'-ACCUUUCUCCA-3' and/or the stem2 sequence 5'-GCUCAGG-3' or 5'-UGGAGAAAGGU-3', respectively. Also, as far as the invention is not limited to histone stem loop sequences specifically, stem loop sequences are preferably not derived from a mammalian insulin

receptor 3'-untranslated region. Also, preferably, the inventive nucleic acid may not contain histone stem loop processing signals, in particular not those derived from mouse histone gene H2A614 gene (H2kA614).

The inventive nucleic acid sequence according to the first aspect of the present invention may optionally comprise or code for a poly(A) sequence. When present, such a poly(A) sequence comprises a sequence of about 25 to about 400 adenosine nucleotides, preferably a sequence of about 30 or, more preferably, of about 50 to about 400 adenosine nucleotides, more preferably a sequence of about 50 to about 300 adenosine nucleotides, even more preferably a sequence of about 50 to about 250 adenosine nucleotides, most preferably a sequence of about 60 to about 250 adenosine nucleotides. In this context the term "about" refers to a deviation of ± 10% of the value(s) it is attached to. Accordingly, the poly(A) sequence contains at least 25 or more than 25 , more preferably, at least 30, more preferably at least 50 adenosine nucleotides. Therefore, such a poly (A) sequence does typically not contain less than 20 adenosine nucleotides. More particularly, it does not contain 10 and/or less than 10 adenosine nucleotides.

Preferably, the nucleic acid according of the present invention does not contain one or two or at least one or all but one or all of the components of the group consisting of: a sequence encoding a ribozyme (preferably a self-splicing ribozyme), a viral nucleic acid sequence, a histone stem-loop processing signal, in particular a histone-stem loop processing sequence derived from mouse histone H2A614 gene, a Neo gene, an inactivated promoter sequence and an inactivated enhancer sequence. Even more preferably, the nucleic acid according to the invention does not contain a ribozyme, preferably a self-splicing ribozyme, and one of the group consisting of: a Neo gene, an inactivated promoter sequence, an inactivated enhancer sequence, a histone stem-loop processing signal, in particular a histone-stem loop processing sequence derived from mouse histone H2A614 gene. Accordingly, the nucleic acid may in a preferred mode neither contain a ribozyme, preferably a self-splicing ribozyme, nor a Neo gene or, alternatively, neither a ribozyme, preferably a self-splicing ribozyme, nor any resistance gene (e.g. usually applied for selection). In another preferred mode, the nucleic acid of the invention may neither contain a ribozyme, preferably a self-

splicing ribozyme nor a histone stem-loop processing signal, in particular a histone-stem loop processing sequence derived from mouse histone H2A614 gene

Alternatively, according to the first aspect of the present invention, the inventive nucleic sequence optionally comprises a polyadenylation signal which is defined herein as a signal which conveys polyadenylation to a (transcribed) mRNA by specific protein factors (e.g. cleavage and polyadenylation specificity factor (CPSF), cleavage stimulation factor (CstF), cleavage factors I and II (CF I and CF II), poly(A) polymerase (PAP)). In this context a consensus polyadenylation signal is preferred comprising the NN(U/T)ANA consensus sequence. In a particular preferred aspect the polyadenylation signal comprises one of the following sequences: AA(U/T)AAA or A(U/T)(U/T)AAA (wherein uridine is usually present in RNA and thymidine is usually present in DNA). In some embodiments, the polyadenylation signal used in the inventive nucleic acid does not correspond to the U3 snRNA, U5, the polyadenylation processing signal from human gene G-CSF, or the SV40 polyadenylation signal sequences. In particular, the above polyadenylation signals are not combined with any antibiotics resistance gene (or any other reporter, marker or selection gene), in particular not with the resistance *neo* gene (neomycin phosphotransferase) (as the gene of the coding region according to element (a) of the inventive nucleic acid in such an inventive nucleic acid. And any of the above polyadenylation signals are preferably not combined with the histone stem loop or the histone stem loop processing signal from mouse histone gene H2A614 in an inventive nucleic acid.

The inventive nucleic acid sequence according to the first aspect of the present invention may furthermore encode a protein or a peptide, which comprises a tumour antigen or a fragment, variant or derivative thereof.

Tumour antigens are preferably located on the surface of the (tumour) cell characterizing a mammalian, in particular human tumour (in e.g. systemic or solid tumour diseases). Tumour antigens may also be selected from proteins, which are overexpressed in tumour cells compared to a normal cell. Furthermore, tumour antigens also includes antigens expressed in cells which are (were) not themselves (or originally not themselves) degenerated but are associated with the supposed tumour. Antigens which are connected with tumour-supplying

vessels or (re)formation thereof, in particular those antigens which are associated with neovascularization, e.g. growth factors, such as VEGF, bFGF etc., are also included herein. Antigens connected with a tumour furthermore include antigens from cells or tissues, typically embedding the tumour. Further, some substances (usually proteins or peptides) are expressed in patients suffering (knowingly or not-knowingly) from a cancer disease and they occur in increased concentrations in the body fluids of said patients. These substances are also referred to as "tumour antigens", however they are not antigens in the stringent meaning of an immune response inducing substance. The class of tumour antigens can be divided further into tumour-specific antigens (TSAs) and tumour-associated-antigens (TAAs). TSAs can only be presented by tumour cells and never by normal "healthy" cells. They typically result from a tumour specific mutation. TAAs, which are more common, are usually presented by both tumour and healthy cells. These antigens are recognized and the antigen-presenting cell can be destroyed by cytotoxic T cells. Additionally, tumour antigens can also occur on the surface of the tumour in the form of, e.g., a mutated receptor. In this case, they can be recognized by antibodies.

Further, tumour associated antigens may be classified as tissue-specific antigens, also called melanocyte-specific antigens, cancer-testis antigens and tumour-specific antigens. Cancer-testis antigens are typically understood to be peptides or proteins of germ-line associated genes which may be activated in a wide variety of tumours. Human cancer-testis antigens may be further subdivided into antigens which are encoded on the X chromosome, so-called CT-X antigens, and those antigens which are not encoded on the X chromosome, the so-called (non-X CT antigens). Cancer-testis antigens which are encoded on the X-chromosome comprises, for example, the family of melanoma antigen genes, the so-called MAGE-family. The genes of the MAGE-family may be characterised by a shared MAGE homology domain (MHD). Each of these antigens, i.e. melanocyte-specific antigens, cancer-testis antigens and tumour-specific antigens, may elicit autologous cellular and humoral immune response. Accordingly, the tumour antigen encoded by the inventive nucleic acid sequence is preferably a melanocyte-specific antigen, a cancer-testis antigen or a tumour-specific antigens, preferably it may be a CT-X antigen, a non-X CT-antigens, a binding partner for a CT-X antigen or a binding partner for a non-X CT-antigen or a tumour-specific antigen , more preferably a CT-X antigen, a binding partner for a non-X CT-antigen or a tumour-specific antigen.

Particular preferred tumour antigens are selected from the list consisting of 5T4, 707-AP, 9D7, AFP, AlbZIP HPG1, alpha-5-beta-1-integrin, alpha-5-beta-6-integrin, alpha-actinin-4/m, alpha-methylacyl-coenzyme A racemase, ART-4, ARTC1/m, B7H4, BAGE-1, BCL-2, bcr/abl, beta-catenin/m, BING-4, BRCA1/m, BRCA2/m, CA 15-3/CA 27-29, CA 19-9, CA72-4, CA125, calreticulin, CAMEL, CASP-8/m, cathepsin B, cathepsin L, CD19, CD20, CD22, CD25, CDE30, CD33, CD4, CD52, CD55, CD56, CD80, CDC27/m, CDK4/m, CDKN2A/m, CEA, CLCA2, CML28, CML66, COA-1/m, coactosin-like protein, collage XXIII, COX-2, CT-9/BRD6, Cten, cyclin B1, cyclin D1, cyp-B, CYPB1, DAM-10, DAM-6, DEK-CAN, EFTUD2/m, EGFR, ELF2/m, EMMPRIN, EpCam, EphA2, EphA3, ErbB3, ETV6-AML1, EZH2, FGF-5, FN, Frau-1, G250, GAGE-1, GAGE-2, GAGE-3, GAGE-4, GAGE-5, GAGE-6, GAGE7b, GAGE-8, GDEP, GnT-V, gp100, GPC3, GPNMB/m, HAGE, HAST-2, hepsin, Her2/neu, HERV-K-MEL, HLA-A*0201-R17I, HLA-A11/m, HLA-A2/m, HNE, homeobox NKX3.1, HOM-TES-14/SCP-1, HOM-TES-85, HPV-E6, HPV-E7, HSP70-2M, HST-2, hTERT, iCE, IGF-1R, IL-13Ra2, IL-2R, IL-5, immature laminin receptor, kallikrein-2, kallikrein-4, Ki67, KIAA0205, KIAA0205/m, KK-LC-1, K-Ras/m, LAGE-A1, LDLR-FUT, MAGE-A1, MAGE-A2, MAGE-A3, MAGE-A4, MAGE-A6, MAGE-A9, MAGE-A10, MAGE-A12, MAGE-B1, MAGE-B2, MAGE-B3, MAGE-B4, MAGE-B5, MAGE-B6, MAGE-B10, MAGE-B16, MAGE-B17, MAGE-C1, MAGE-C2, MAGE-C3, MAGE-D1, MAGE-D2, MAGE-D4, MAGE-E1, MAGE-E2, MAGE-F1, MAGE-H1, MAGEL2, mammaglobin A, MART-1/melan-A, MART-2, MART-2/m, matrix protein 22, MC1R, M-CSF, ME1/m, mesothelin, MG50/PXDN, MMP11, MN/CA IX-antigen, MRP-3, MUC-1, MUC-2, MUM-1/m, MUM-2/m, MUM-3/m, myosin class I/m, NA88-A, N-acetylglucosaminyltransferase-V, Neo-PAP, Neo-PAP/m, NFYC/m, NGEP, NMP22, NPM/ALK, N-Ras/m, NSE, NY-ESO-B, NY-ESO-1, OA1, OFA-iLRP, OGT, OGT/m, OS-9, OS-9/m, osteocalcin, osteopontin, p15, p190 minor bcr-abl, p53, p53/m, PAGE-4, PAI-1, PAI-2, PAP, PART-1, PATE, PDEF, Pim-1-Kinase, Pin-1, Pml/PARalpha, POTE, PRAME, PRDX5/m, prostein, proteinase-3, PSA, PSCA, PSGR, PSM, PSMA, PTPRK/m, RAGE-1, RBAF600/m, RHAMM/CD168, RU1, RU2, S-100, SAGE, SART-1, SART-2, SART-3, SCC, SIRT2/m, Sp17, SSX-1, SSX-2/HOM-MEL-40, SSX-4, STAMP-1, STEAP-1, survivin, survivin-2B, SYT-SSX-1, SYT-SSX-2, TA-90, TAG-72, TARP, TEL-AML1, TGFbeta, TGFbetaRII, TGM-4, TPI/m, TRAG-3, TRG, TRP-1, TRP-2/6b, TRP/INT2, TRP-p8, tyrosinase, UPA, VEGFR1, VEGFR-2/FLK-1, and WT1. Such tumour antigens preferably may be selected from the group consisting of p53, CA125, EGFR, Her2/neu, hTERT, PAP, MAGE-

A1, MAGE-A3, Mesothelin, MUC-1, GP100, MART-1, Tyrosinase, PSA, PSCA, PSMA, STEAP-1, VEGF, VEGFR1, VEGFR2, Ras, CEA or WT1, and more preferably from PAP, MAGE-A3, WT1, and MUC-1. Such tumour antigens preferably may be selected from the group consisting of MAGE-A1 (e.g. MAGE-A1 according to accession number M77481), MAGE-A2, MAGE-A3, MAGE-A6 (e.g. MAGE-A6 according to accession number NM_005363), MAGE-C1, MAGE-C2, melan-A (e.g. melan-A according to accession number NM_005511), GP100 (e.g. GP100 according to accession number M77348), tyrosinase (e.g. tyrosinase according to accession number NM_000372), surviving (e.g. survivin according to accession number AF077350), CEA (e.g. CEA according to accession number NM_004363), Her-2/neu (e.g. Her-2/neu according to accession number M11730), WT1 (e.g. WT1 according to accession number NM_000378), PRAME (e.g. PRAME according to accession number NM_006115), EGFRI (epidermal growth factor receptor 1) (e.g. EGFRI (epidermal growth factor receptor 1) according to accession number AF288738), MUC1, mucin-1 (e.g. mucin-1 according to accession number NM_002456), SEC61G (e.g. SEC61G according to accession number NM_014302), hTERT (e.g. hTERT accession number NM_198253), 5T4 (e.g. 5T4 according to accession number NM_006670), TRP-2 (e.g. TRP-2 according to accession number NM_001922), STEAP1, PCA, PSA, PSMA, etc.

Furthermore tumour antigens also may encompass idiotypic antigens associated with a cancer or tumour disease, particularly lymphoma or a lymphoma associated disease, wherein said idiotypic antigen is an immunoglobulin idiotype of a lymphoid blood cell or a T cell receptor idiotype of a lymphoid blood cell.

Tumour antigenic proteins for the treatment of cancer or tumour diseases, are typically proteins of mammalian origin, preferably of human origin. Their selection for treatment of the subject depends on the tumour type to be treated and the expression profile of the individual tumour. A human suffering from prostate cancer, is e.g. preferably treated by a tumour antigen, which is typically expressed (or overexpressed) in prostate carcinoma or specifically overexpressed in the subject to be treated, e.g. any of PSMA, PSCA, and/or PSA.

The coding region of the inventive nucleic acid according to the first aspect of the present invention may occur as a mono-, di-, or even multicistronic nucleic acid, i.e. a nucleic acid which carries the coding sequences of one, two or more proteins or peptides. Such coding

sequences in di-, or even multicistronic nucleic acids may be separated by at least one internal ribosome entry site (IRES) sequence, e.g. as described herein or by signal peptides which induce the cleavage of the resulting polypeptide which comprises several proteins or peptides.

According to the first aspect of the present invention, the inventive nucleic acid sequence comprises a coding region, encoding a peptide or protein which comprises a tumour antigen or a fragment, variant or derivative thereof. Preferably, the encoded tumour antigen is no histone protein. In the context of the present invention such a histone protein is typically a strongly alkaline protein found in eukaryotic cell nuclei, which package and order the DNA into structural units called nucleosomes. Histone proteins are the chief protein components of chromatin, act as spools around which DNA winds, and play a role in gene regulation. Without histones, the unwound DNA in chromosomes would be very long (a length to width ratio of more than 10 million to one in human DNA). For example, each human cell has about 1.8 meters of DNA, but wound on the histones it has about 90 millimeters of chromatin, which, when duplicated and condensed during mitosis, result in about 120 micrometers of chromosomes. More preferably, in the context of the present invention such a histone protein is typically defined as a highly conserved protein selected from one of the following five major classes of histones: H1/H5, H2A, H2B, H3, and H4", preferably selected from mammalian histone, more preferably from human histones or histone proteins. Such histones or histone proteins are typically organised into two super-classes defined as core histones, comprising histones H2A, H2B, H3 and H4, and linker histones, comprising histones H1 and H5.

In this context, linker histones, preferably excluded from the scope of protection of the pending invention, preferably mammalian linker histones, more preferably human linker histones, are typically selected from H1, including H1F, particularly including H1F0, H1FNT, H1FOO, H1FX, and H1H1, particularly including HIST1H1A, HIST1H1B, HIST1H1C, HIST1H1D, HIST1H1E, HIST1H1T; and

Furthermore, core histones, preferably excluded from the scope of protection of the pending invention, preferably mammalian core histones, more preferably human core histones, are typically selected from H2A, including H2AF, particularly including H2AFB1, H2AFB2,

H2AFB3, H2AFJ, H2AFV, H2AFX, H2AFY, H2AFY2, H2AFZ, and H2A1, particularly including HIST1H2AA, HIST1H2AB, HIST1H2AC, HIST1H2AD, HIST1H2AE, HIST1H2AG, HIST1H2AI, HIST1H2AJ, HIST1H2AK, HIST1H2AL, HIST1H2AM, and H2A2, particularly including HIST2H2AA3, HIST2H2AC; H2B, including H2BF, particularly including H2BFM, H2BFO, H2BFS, H2BFWT H2B1, particularly including HIST1H2BA, HIST1H2BB, HIST1H2BC, HIST1H2BD, HIST1H2BE, HIST1H2BF, HIST1H2BG, HIST1H2BH, HIST1H2BI, HIST1H2BJ, HIST1H2BK, HIST1H2BL, HIST1H2BM, HIST1H2BN, HIST1H2BO, and H2B2, particularly including HIST2H2BE; H3, including H3A1, particularly including HIST1H3A, HIST1H3B, HIST1H3C, HIST1H3D, HIST1H3E, HIST1H3F, HIST1H3G, HIST1H3H, HIST1H3I, HIST1H3J, and H3A2, particularly including HIST2H3C, and H3A3, particularly including HIST3H3; H4, including H41, particularly including HIST1H4A, HIST1H4B, HIST1H4C, HIST1H4D, HIST1H4E, HIST1H4F, HIST1H4G, HIST1H4H, HIST1H4I, HIST1H4J, HIST1H4K, HIST1H4L, and H44, particularly including HIST4H4, and H5.

According to the first aspect of the present invention, the inventive nucleic acid sequence comprises a coding region, encoding a peptide or protein which comprises a tumour antigen or a fragment, variant or derivative thereof. Preferably, the encoded tumour antigen is no reporter protein (e.g. Luciferase, Green Fluorescent Protein (GFP), Enhanced Green Fluorescent Protein (EGFP), β-Galactosidase) and no marker or selection protein (e.g. alpha-Globin, Galactokinase and Xanthine:guanine phosphoribosyl transferase (GPT)). Preferably, the nucleic acid sequence of the invention does not contain a (bacterial) antibiotics resistance gene, in particular not a *neo* gene sequence (Neomycin resistance gene) or CAT gene sequence (chloramphenicol acetyl transferase, chloramphenicol resistance gene).

The inventive nucleic acid as define above, comprises or codes for a) a coding region, encoding a peptide or protein which comprises a tumour antigen or a fragment, variant or derivative thereof; b) at least one histone stem-loop, and c) a poly(A) sequence or polyadenylation signal; preferably for increasing the expression of said encoded peptide or protein, wherein the encoded peptide or protein is preferably no histone protein, no reporter protein and/or no marker or selection protein, as defined above. The elements b) to c) of the inventive nucleic acid may occur in the inventive nucleic acid in any order, i.e. the elements a), b) and c) may occur in the order a), b) and c) or a), c) and b) from 5′ to 3′

direction in the inventive nucleic acid sequence, wherein further elements as described herein, may also be contained, such as a 5'-CAP structure, a poly(C) sequence, stabilization sequences, IRES sequences, etc. Each of the elements a) to c) of the inventive nucleic acid, particularly a) in di- or multicistronic constructs and/or each of the elements b) and c), more preferably element b) may also be repeated at least once, preferably twice or more in the inventive nucleic acid. As an example, the inventive nucleic acid may show its sequence elements a), b) and optionally c) in e.g. the following order:

5' – coding region – histone stem-loop – poly(A) sequence – 3'; or

5' – coding region – histone stem-loop – polyadenylation signal – 3'; or

5' – coding region – poly(A) sequence – histone stem-loop – 3'; or

5' – coding region – polyadenylation signal– histone stem-loop – 3'; or

5' – coding region – coding region - histone stem-loop – polyadenylation signal - 3'; or

5' – coding region – histone stem-loop – histone stem-loop – poly(A) sequence – 3'; or

5' – coding region – histone stem-loop – histone stem-loop – polyadenylation signal– 3';etc.

In this context it is particularly preferred that the inventive nucleic acid sequence comprises or codes for a) a coding region, encoding a peptide or protein which comprises a tumour antigen or fragment, variant or derivative thereof; b) at least one histone stem-loop, and c) a poly(A) sequence or polyadenylation sequence; preferably for increasing the expression level of said encoded peptide or protein, wherein the encoded protein is preferably no histone protein, no reporter protein (e.g. Luciferase, GFP, EGFP, β-Galactosidase, particularly EGFP) and/or no marker or selection protein (e.g. alpha-Globin, Galactokinase and Xanthine:Guanine phosphoribosyl transferase (GPT)).

In a further preferred embodiment of the first aspect the inventive nucleic acid sequence as defined herein may also occur in the form of a modified nucleic acid.

In this context, the inventive nucleic acid sequence as defined herein may be modified to provide a "stabilized nucleic acid", preferably a stabilized RNA, more preferably an RNA that is essentially resistant to *in vivo* degradation (e.g. by an exo- or endo-nuclease). A stabilized nucleic acid may e.g. be obtained by modification of the G/C content of the coding region of the inventive nucleic acid sequence, by introduction of nucleotide

analogues (e.g. nucleotides with backbone modifications, sugar modifications or base modifications) or by introduction of stabilization sequences in the 3'- and/or 5'-untranslated region of the inventive nucleic acid sequence.

As mentioned above, the inventive nucleic acid sequence as defined herein may contain nucleotide analogues/modifications e.g. backbone modifications, sugar modifications or base modifications. A backbone modification in connection with the present invention is a modification in which phosphates of the backbone of the nucleotides contained in inventive nucleic acid sequence as defined herein are chemically modified. A sugar modification in connection with the present invention is a chemical modification of the sugar of the nucleotides of the inventive nucleic acid sequence as defined herein. Furthermore, a base modification in connection with the present invention is a chemical modification of the base moiety of the nucleotides of the nucleic acid molecule of the inventive nucleic acid sequence. In this context nucleotide analogues or modifications are preferably selected from nucleotide analogues which are applicable for transcription and/or translation.

In a particular preferred embodiment of the first aspect of the present invention the herein defined nucleotide analogues/modifications are selected from base modifications which additionally increase the expression of the encoded protein and which are preferably selected from 2-amino-6-chloropurineriboside-5'-triphosphate, 2-aminoadenosine-5'-triphosphate, 2-thiocytidine-5'-triphosphate, 2-thiouridine-5'-triphosphate, 4-thiouridine-5'-triphosphate, 5-aminoallylcytidine-5'-triphosphate, 5-aminoallyluridine-5'-triphosphate, 5-bromocytidine-5'-triphosphate, 5-bromouridine-5'-triphosphate, 5-iodocytidine-5'-triphosphate, 5-iodouridine-5'-triphosphate, 5-methylcytidine-5'-triphosphate, 5-methyluridine-5'-triphosphate, 6-azacytidine-5'-triphosphate, 6-azauridine-5'-triphosphate, 6-chloropurineriboside-5'-triphosphate, 7-deazaadenosine-5'-triphosphate, 7-deazaguanosine-5'-triphosphate, 8-azaadenosine-5'-triphosphate, 8-azidoadenosine-5'-triphosphate, benzimidazole-riboside-5'-triphosphate, N1-methyladenosine-5'-triphosphate, N1-methylguanosine-5'-triphosphate, N6-methyladenosine-5'-triphosphate, O6-methylguanosine-5'-triphosphate, pseudouridine-5'-triphosphate, or puromycin-5'-triphosphate, xanthosine-5'-triphosphate. Particular preference is given to nucleotides for base modifications selected from the group of base-modified nucleotides consisting of 5-

methylcytidine-5'-triphosphate, 7-deazaguanosine-5'-triphosphate, 5-bromocytidine-5'-triphosphate, and pseudouridine-5'-triphosphate.

According to a further embodiment, the inventive nucleic acid sequence as defined herein can contain a lipid modification. Such a lipid-modified nucleic acid typically comprises a nucleic acid as defined herein. Such a lipid-modified nucleic acid molecule of the inventive nucleic acid sequence as defined herein typically further comprises at least one linker covalently linked with that nucleic acid molecule, and at least one lipid covalently linked with the respective linker. Alternatively, the lipid-modified nucleic acid molecule comprises at least one nucleic acid molecule as defined herein and at least one (bifunctional) lipid covalently linked (without a linker) with that nucleic acid molecule. According to a third alternative, the lipid-modified nucleic acid molecule comprises a nucleic acid molecule as defined herein, at least one linker covalently linked with that nucleic acid molecule, and at least one lipid covalently linked with the respective linker, and also at least one (bifunctional) lipid covalently linked (without a linker) with that nucleic acid molecule. In this context it is particularly preferred that the lipid modification is present at the terminal ends of a linear inventive nucleic acid sequence.

According to another preferred embodiment of the first aspect of the invention, the inventive nucleic acid sequence as defined herein, particularly if provided as an (m)RNA, can therefore be stabilized against degradation by RNases by the addition of a so-called "5' CAP" structure.

According to a further preferred embodiment of the first aspect of the invention, the inventive nucleic acid sequence as defined herein can be modified by a sequence of at least 10 cytidines, preferably at least 20 cytidines, more preferably at least 30 cytidines (so-called "poly(C) sequence"). Particularly, the inventive nucleic acid sequence may contain or code for a poly(C) sequence of typically about 10 to 200 cytidine nucleotides, preferably about 10 to 100 cytidine nucleotides, more preferably about 10 to 70 cytidine nucleotides or even more preferably about 20 to 50 or even 20 to 30 cytidine nucleotides. This poly(C) sequence is preferably located 3' of the coding region comprised in the inventive nucleic acid according to the first aspect of the present invention.

In a particularly preferred embodiment of the present invention, the G/C content of the coding region, encoding at least one peptide or protein which comprises a tumour antigen or a fragment, variant or derivative thereof of the inventive nucleic acid sequence as defined herein, is modified, particularly increased, compared to the G/C content of its particular wild type coding region, i.e. the unmodified coding region. The encoded amino acid sequence of the coding region is preferably not modified compared to the coded amino acid sequence of the particular wild type coding region.

The modification of the G/C-content of the coding region of the inventive nucleic acid sequence as defined herein is based on the fact that the sequence of any mRNA region to be translated is important for efficient translation of that mRNA. Thus, the composition and the sequence of various nucleotides are important. In particular, mRNA sequences having an increased G (guanosine)/C (cytosine) content are more stable than mRNA sequences having an increased A (adenosine)/U (uracil) content. According to the invention, the codons of the coding region are therefore varied compared to its wild type coding region, while retaining the translated amino acid sequence, such that they include an increased amount of G/C nucleotides. In respect to the fact that several codons code for one and the same amino acid (so-called degeneration of the genetic code), the most favourable codons for the stability can be determined (so-called alternative codon usage).

Depending on the amino acid to be encoded by the coding region of the inventive nucleic acid sequence as defined herein, there are various possibilities for modification of the nucleic acid sequence, e.g. the coding region, compared to its wild type coding region. In the case of amino acids which are encoded by codons which contain exclusively G or C nucleotides, no modification of the codon is necessary. Thus, the codons for Pro (CCC or CCG), Arg (CGC or CGG), Ala (GCC or GCG) and Gly (GGC or GGG) require no modification, since no A or U is present.

In contrast, codons which contain A and/or U nucleotides can be modified by substitution of other codons which code for the same amino acids but contain no A and/or U. Examples of these are:

the codons for Pro can be modified from CCU or CCA to CCC or CCG;

the codons for Arg can be modified from CGU or CGA or AGA or AGG to CGC or CGG;
the codons for Ala can be modified from GCU or GCA to GCC or GCG;
the codons for Gly can be modified from GGU or GGA to GGC or GGG.

In other cases, although A or U nucleotides cannot be eliminated from the codons, it is however possible to decrease the A and U content by using codons which contain a lower content of A and/or U nucleotides. Examples of these are:

the codons for Phe can be modified from UUU to UUC;
the codons for Leu can be modified from UUA, UUG, CUU or CUA to CUC or CUG;
the codons for Ser can be modified from UCU or UCA or AGU to UCC, UCG or AGC;
the codon for Tyr can be modified from UAU to UAC;
the codon for Cys can be modified from UGU to UGC;
the codon for His can be modified from CAU to CAC;
the codon for Gln can be modified from CAA to CAG;
the codons for Ile can be modified from AUU or AUA to AUC;
the codons for Thr can be modified from ACU or ACA to ACC or ACG;
the codon for Asn can be modified from AAU to AAC;
the codon for Lys can be modified from AAA to AAG;
the codons for Val can be modified from GUU or GUA to GUC or GUG;
the codon for Asp can be modified from GAU to GAC;
the codon for Glu can be modified from GAA to GAG;
the stop codon UAA can be modified to UAG or UGA.

In the case of the codons for Met (AUG) and Trp (UGG), on the other hand, there is no possibility of sequence modification.

The substitutions listed above can be used either individually or in all possible combinations to increase the G/C content of the coding region of the inventive nucleic acid sequence as defined herein, compared to its particular wild type coding region (i.e. the original sequence). Thus, for example, all codons for Thr occurring in the wild type sequence can be modified to ACC (or ACG).

In the above context, mRNA codons are shown. Therefore uridine present in an mRNA may also be present as thymidine in the respective DNA coding for the particular mRNA.

Preferably, the G/C content of the coding region of the inventive nucleic acid sequence as defined herein is increased by at least 7%, more preferably by at least 15%, particularly preferably by at least 20%, compared to the G/C content of the wild type coding region. According to a specific embodiment at least 5%, 10%, 20%, 30%, 40%, 50%, 60%, more preferably at least 70 %, even more preferably at least 80% and most preferably at least 90%, 95% or even 100% of the substitutable codons in the coding region encoding at least one peptide or protein which comprises a tumour antigen or a fragment, variant or derivative thereof are substituted, thereby increasing the G/C content of said coding region.

In this context, it is particularly preferable to increase the G/C content of the coding region of the inventive nucleic acid sequence as defined herein, to the maximum (i.e. 100% of the substitutable codons), compared to the wild type coding region.

According to the invention, a further preferred modification of the coding region encoding at least one peptide or protein which comprises a tumour antigen or a fragment, variant or derivative thereof of the inventive nucleic acid sequence as defined herein, is based on the finding that the translation efficiency is also determined by a different frequency in the occurrence of tRNAs in cells. Thus, if so-called "rare codons" are present in the coding region of the inventive nucleic acid sequence as defined herein, to an increased extent, the corresponding modified nucleic acid sequence is translated to a significantly poorer degree than in the case where codons coding for relatively "frequent" tRNAs are present.

In this context the coding region of the inventive nucleic acid sequence is preferably modified compared to the corresponding wild type coding region such that at least one codon of the wild type sequence which codes for a tRNA which is relatively rare in the cell is exchanged for a codon which codes for a tRNA which is relatively frequent in the cell and carries the same amino acid as the relatively rare tRNA. By this modification, the coding region of the inventive nucleic acid sequence as defined herein, is modified such that codons for which frequently occurring tRNAs are available are inserted. In other words, according to the invention, by this modification all codons of the wild type coding region

which code for a tRNA which is relatively rare in the cell can in each case be exchanged for a codon which codes for a tRNA which is relatively frequent in the cell and which, in each case, carries the same amino acid as the relatively rare tRNA.

Which tRNAs occur relatively frequently in the cell and which, in contrast, occur relatively rarely is known to a person skilled in the art; cf. e.g. Akashi, Curr. Opin. Genet. Dev. 2001, 11(6): 660-666. The codons which use for the particular amino acid the tRNA which occurs the most frequently, e.g. the Gly codon, which uses the tRNA which occurs the most frequently in the (human) cell, are particularly preferred.

According to the invention, it is particularly preferable to link the sequential G/C content which is increased, in particular maximized, in the coding region of the inventive nucleic acid sequence as defined herein, with the "frequent" codons without modifying the amino acid sequence of the peptide or protein encoded by the coding region of the nucleic acid sequence. This preferred embodiment allows provision of a particularly efficiently translated and stabilized (modified) inventive nucleic acid sequence as defined herein.

According to another preferred embodiment of the first aspect of the invention, the inventive nucleic acid sequence as defined herein, preferably has additionally at least one 5' and/or 3' stabilizing sequence. These stabilizing sequences in the 5' and/or 3' untranslated regions have the effect of increasing the half-life of the nucleic acid, particularly of the mRNA in the cytosol. These stabilizing sequences can have 100% sequence identity to naturally occurring sequences which occur in viruses, bacteria and eukaryotes, but can also be partly or completely synthetic. The untranslated sequences (UTR) of the (alpha-)globin gene, e.g. from *Homo sapiens* or *Xenopus laevis* may be mentioned as an example of stabilizing sequences which can be used in the present invention for a stabilized nucleic acid. Another example of a stabilizing sequence has the general formula (C/U)CCAN$_x$CCC(U/A)Py$_x$UC(C/U)CC (SEQ ID NO: 55), which is contained in the 3'-UTRs of the very stable RNAs which code for (alpha-)globin, type(I)-collagen, 15-lipoxygenase or for tyrosine hydroxylase (cf. Holcik *et al.*, Proc. Natl. Acad. Sci. USA 1997, 94: 2410 to 2414). Such stabilizing sequences can of course be used individually or in combination with one another and also in combination with other stabilizing sequences known to a person skilled in the art. In this context it is particularly preferred that the 3'

UTR sequence of the alpha globin gene is located 3' of the coding region encoding at least one peptide or protein which comprises a tumour antigen or a fragment, variant or derivative thereof comprised in the inventive nucleic acid sequence according to the first aspect of the present invention.

Substitutions, additions or eliminations of bases are preferably carried out with the inventive nucleic acid sequence as defined herein, using a DNA matrix for preparation of the nucleic acid sequence by techniques of the well-known site directed mutagenesis or with an oligonucleotide ligation strategy (see e.g. Maniatis *et al.*, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, 3rd ed., Cold Spring Harbor, NY, 2001).

Any of the above modifications may be applied to the inventive nucleic acid sequence as defined herein and further to any nucleic acid as used in the context of the present invention and may be, if suitable or necessary, be combined with each other in any combination, provided, these combinations of modifications do not interfere with each other in the respective nucleic acid. A person skilled in the art will be able to take his choice accordingly.

Nucleic acid sequences used according to the present invention as defined herein may be prepared using any method known in the art, including synthetic methods such as e.g. solid phase synthesis, as well as *in vitro* methods, such as *in vitro* transcription reactions or *in vivo* reactions, such as *in vivo* propagation of DNA plasmids in bacteria.

In such a process, for preparation of the inventive nucleic acid sequence as defined herein, especially if the nucleic acid is in the form of an mRNA, a corresponding DNA molecule may be transcribed *in vitro*. This DNA matrix preferably comprises a suitable promoter, e.g. a T7 or SP6 promoter, for *in vitro* transcription, which is followed by the desired nucleotide sequence for the nucleic acid molecule, e.g. mRNA, to be prepared and a termination signal for *in vitro* transcription. The DNA molecule, which forms the matrix of the at least one RNA of interest, may be prepared by fermentative proliferation and subsequent isolation as part of a plasmid which can be replicated in bacteria. Plasmids which may be mentioned as suitable for the present invention are e.g. the plasmids pT7Ts (GenBank accession number U26404; Lai *et al.*, Development 1995, 121: 2349 to 2360), pGEM®

series, e.g. pGEM®-1 (GenBank accession number X65300; from Promega) and pSP64 (GenBank accession number X65327); cf. also Mezei and Storts, Purification of PCR Products, in: Griffin and Griffin (ed.), PCR Technology: Current Innovation, CRC Press, Boca Raton, FL, 2001.

The inventive nucleic acid sequence as defined herein as well as proteins or peptides as encoded by this nucleic acid sequence may comprise fragments or variants of those sequences. Such fragments or variants may typically comprise a sequence having a sequence identity with one of the above mentioned nucleic acids, or with one of the proteins or peptides or sequences, if encoded by the inventive nucleic acid sequence, of at least 5%, 10%, 20%, 30%, 40%, 50%, 60%, preferably at least 70%, more preferably at least 80%, equally more preferably at least 85%, even more preferably at least 90% and most preferably at least 95% or even 97%, 98% or 99%, to the entire wild type sequence, either on nucleic acid level or on amino acid level.

"Fragments" of proteins or peptides in the context of the present invention (e.g. as encoded by the inventive nucleic acid sequence as defined herein) may comprise a sequence of a protein or peptide as defined herein, which is, with regard to its amino acid sequence (or its encoded nucleic acid molecule), N-terminally, C-terminally and/or intrasequentially truncated/shortened compared to the amino acid sequence of the original (native) protein (or its encoded nucleic acid molecule). Such truncation may thus occur either on the amino acid level or correspondingly on the nucleic acid level. A sequence identity with respect to such a fragment as defined herein may therefore preferably refer to the entire protein or peptide as defined herein or to the entire (coding) nucleic acid molecule of such a protein or peptide. Likewise, "fragments" of nucleic acids in the context of the present invention may comprise a sequence of a nucleic acid as defined herein, which is, with regard to its nucleic acid molecule 5'-, 3'- and/or intrasequentially truncated/shortened compared to the nucleic acid molecule of the original (native) nucleic acid molecule. A sequence identity with respect to such a fragment as defined herein may therefore preferably refer to the entire nucleic acid as defined herein and the preferred sequence identity level typically is as indicated herein. Fragments have the same biological function or specific activity or at least retain an activity of the natural full length protein of at least 50%, more preferably at least 70%, even more preferably at least 90% (measured in an appropriate functional assay, e.g.

an assay assessing the antigenic property by a appropriate assay system which e.g. measures an immunological reaction, e.g. expression and/or secretion of an appropriate cytokine (as an indicator of the immune reaction)) as compared to the full-length native peptide or protein, e.g. its specific antigenic or therapeutic property. Accordingly, in a preferred embodiment, the "fragment" is a portion of the full-length (naturally occurring) tumour antigenic protein, which exerts tumour antigenic properties on the immune system as indicated herein..

Fragments of proteins or peptides in the context of the present invention (e.g. as encoded by the inventive nucleic acid sequence as defined herein) may furthermore comprise a sequence of a protein or peptide as defined herein, which has a length of about 6 to about 20 or even more amino acids, e.g. fragments as processed and presented by MHC class I molecules, preferably having a length of about 8 to about 10 amino acids, e.g. 8, 9, or 10, (or even 6, 7, 11, or 12 amino acids), or fragments as processed and presented by MHC class II molecules, preferably having a length of about 13 or more amino acids, e.g. 13, 14, 15, 16, 17, 18, 19, 20 or even more amino acids, wherein these fragments may be selected from any part of the amino acid sequence. These fragments are typically recognized by T-cells in form of a complex consisting of the peptide fragment and an MHC molecule, i.e. the fragments are typically not recognized in their native form. Fragments of proteins or peptides as defined herein may comprise at least one epitope of those proteins or peptides. Furthermore, also domains of a protein, like the extracellular domain, the intracellular domain or the transmembrane domain and shortened or truncated versions of a protein may be understood to comprise a fragment of a protein.

Fragments of proteins or peptides as defined herein (e.g. as encoded by the inventive nucleic acid sequence as defined herein) may also comprise epitopes of those proteins or peptides. T cell epitopes or parts of the proteins in the context of the present invention may comprise fragments preferably having a length of about 6 to about 20 or even more amino acids, e.g. fragments as processed and presented by MHC class I molecules, preferably having a length of about 8 to about 10 amino acids, e.g. 8, 9, or 10, (or even 11, or 12 amino acids), or fragments as processed and presented by MHC class II molecules, preferably having a length of about 13 or more amino acids, e.g. 13, 14, 15, 16, 17, 18, 19, 20 or even more amino acids, wherein these fragments may be selected from any part of the

amino acid sequence. These fragments are typically recognized by T cells in form of a complex consisting of the peptide fragment and an MHC molecule, i.e. the fragments are typically not recognized in their native form.

B cell epitopes are typically fragments located on the outer surface of (native) protein or peptide antigens as defined herein, preferably having 5 to 15 amino acids, more preferably having 5 to 12 amino acids, even more preferably having 6 to 9 amino acids, which may be recognized by antibodies, i.e. in their native form.

Such epitopes of proteins or peptides may furthermore be selected from any of the herein mentioned variants of such proteins or peptides. In this context antigenic determinants can be conformational or discontinuous epitopes which are composed of segments of the proteins or peptides as defined herein that are discontinuous in the amino acid sequence of the proteins or peptides as defined herein but are brought together in the three-dimensional structure or continuous or linear epitopes which are composed of a single polypeptide chain.

"Variants" of proteins or peptides as defined in the context of the present invention may be encoded by the inventive nucleic acid sequence as defined herein. Thereby, a protein or peptide may be generated, having an amino acid sequence which differs from the original sequence in one or more (2, 3, 4, 5, 6, 7 or more) mutation(s), such as one or more substituted, inserted and/or deleted amino acid(s). The preferred level of sequence identity of "variants" in view of the full-length natural protein sequence typically is as indicated herein. Preferably, variants have the same biological function or specific activity or at least retain an activity of the natural full length protein of at least 50%, more preferably at least 70%, even more preferably at least 90% (measured in an appropriate functional assay, e.g. by an assay assessing the immunological reaction towards the tumour antigen by the secretion and/or expression of one or more cytokines) compared to the full-length native peptide or protein, e.g. its specific antigenic property. Accordingly, in a preferred embodiment, the "variant" is a variant of a tumour antigenic protein, which exerts tumour antigenic properties to the extent as indicated herein.

"Variants" of proteins or peptides as defined in the context of the present invention (e.g. as encoded by a nucleic acid as defined herein) may comprise conservative amino acid substitution(s) compared to their native, i.e. non-mutated physiological, sequence. Those amino acid sequences as well as their encoding nucleotide sequences in particular fall under the term variants as defined herein. Substitutions in which amino acids, which originate from the same class, are exchanged for one another are called conservative substitutions. In particular, these are amino acids having aliphatic side chains, positively or negatively charged side chains, aromatic groups in the side chains or amino acids, the side chains of which can enter into hydrogen bridges, e.g. side chains which have a hydroxyl function. This means that e.g. an amino acid having a polar side chain is replaced by another amino acid having a likewise polar side chain, or, for example, an amino acid characterized by a hydrophobic side chain is substituted by another amino acid having a likewise hydrophobic side chain (e.g. serine (threonine) by threonine (serine) or leucine (isoleucine) by isoleucine (leucine)). Insertions and substitutions are possible, in particular, at those sequence positions which cause no modification to the three-dimensional structure or do not affect the binding region. Modifications to a three-dimensional structure by insertion(s) or deletion(s) can easily be determined e.g. using CD spectra (circular dichroism spectra) (Urry, 1985, Absorption, Circular Dichroism and ORD of Polypeptides, in: Modern Physical Methods in Biochemistry, Neuberger *et al.* (ed.), Elsevier, Amsterdam).

Furthermore, variants of proteins or peptides as defined herein, which may be encoded by the inventive nucleic acid sequence as defined herein, may also comprise those sequences, wherein nucleotides of the nucleic acid are exchanged according to the degeneration of the genetic code, without leading to an alteration of the respective amino acid sequence of the protein or peptide, i.e. the amino acid sequence or at least part thereof may not differ from the original sequence in one or more mutation(s) within the above meaning.

In order to determine the percentage to which two sequences are identical, e.g. nucleic acid sequences or amino acid sequences as defined herein, preferably the amino acid sequences encoded by the inventive nucleic acid sequence as defined herein or the amino acid sequences themselves, the sequences can be aligned in order to be subsequently compared to one another. Therefore, e.g. a position of a first sequence may be compared with the corresponding position of the second sequence. If a position in the first sequence is

occupied by the same component as is the case at a position in the second sequence, the two sequences are identical at this position. If this is not the case, the sequences differ at this position. If insertions occur in the second sequence in comparison to the first sequence, gaps can be inserted into the first sequence to allow a further alignment. If deletions occur in the second sequence in comparison to the first sequence, gaps can be inserted into the second sequence to allow a further alignment. The percentage to which two sequences are identical is then a function of the number of identical positions divided by the total number of positions including those positions which are only occupied in one sequence. The percentage to which two sequences are identical can be determined using a mathematical algorithm. A preferred, but not limiting, example of a mathematical algorithm which can be used is the algorithm of Karlin *et al.* (1993), PNAS USA, 90:5873-5877 or Altschul *et al.* (1997), Nucleic Acids Res., 25:3389-3402. Such an algorithm is integrated in the BLAST program. Sequences which are identical to the sequences of the present invention to a certain extent can be identified by this program.

The inventive nucleic acid sequence as defined herein may encode derivatives of a peptide or protein. Such a derivative of a peptide or protein is a molecule that is derived from another molecule, such as said peptide or protein. A "derivative" typically contains the full-length sequence of the natural peptide or protein and additional sequence features, e.g. at the N- or at the C-terminus, which may exhibit an additional function to the natural full-length peptide/protein. Again such derivatives have the same biological function or specific activity or at least retain an activity of the natural full length protein of at least 50%, more preferably at least 70%, even more preferably at least 90% (measured in an appropriate functional assay, see above, e.g. as expressed by cytokine expression and/or secretion in an immunological reaction) as compared to the full-length native peptide or protein, e.g. its specific therapeutic property. Thereby, a "derivative" of a peptide or protein also encompasses (chimeric) fusion peptides/proteins comprising a peptide or protein used in the present invention or a natural full-length protein (or variant/fragment thereof) fused to a distinct peptide/protein awarding e.g. two or more biological functions to the fusion peptide/protein. For example, the fusion comprises a label, such as, for example, an epitope, e.g., a FLAG epitope or a V5 epitope or an HA epitope. For example, the epitope is a FLAG epitope. Such a tag is useful for, for example, purifying the fusion protein.

In this context, a "variant" of a protein or peptide may have at least 70%, 75%, 80%, 85%, 90%, 95%, 98% or 99% amino acid identity over a stretch of 10, 20, 30, 50, 75 or 100 amino acids of such protein or peptide. Analogously, a "variant" of a nucleic acid sequence, or particularly, a fragment, may have at least 70%, 75%, 80%, 85%, 90%, 95%, 98% or 99% nucleotide identity over a stretch of 10, 20, 30, 50, 75 or 100 nucleotide of such nucleic acid sequence; typically, however, referring to the naturally occurring full-length sequences. In case of "fragments" typically, sequence identity is determined for the fragment over length (of the fragment) on the portion of the full-length protein (reflecting the same length as the fragment), which exhibits the highest level of sequence identity.

In a further preferred embodiment of the first aspect of the present invention the inventive nucleic acid sequence is associated with a vehicle, transfection or complexation agent for increasing the transfection efficiency and/or the immunostimulatory properties of the inventive nucleic acid sequence. Particularly preferred agents in this context suitable for increasing the transfection efficiency are cationic or polycationic compounds, including protamine, nucleoline, spermine or spermidine, or other cationic peptides or proteins, such as poly-L-lysine (PLL), poly-arginine, basic polypeptides, cell penetrating peptides (CPPs), including HIV-binding peptides, HIV-1 Tat (HIV), Tat-derived peptides, Penetratin, VP22 derived or analog peptides, HSV VP22 (Herpes simplex), MAP, KALA or protein transduction domains (PTDs), PpT620, prolin-rich peptides, arginine-rich peptides, lysine-rich peptides, MPG-peptide(s), Pep-1, L-oligomers, Calcitonin peptide(s), Antennapedia-derived peptides (particularly from *Drosophila antennapedia*), pAntp, pIsl, FGF, Lactoferrin, Transportan, Buforin-2, Bac715-24, SynB, SynB(1), pVEC, hCT-derived peptides, SAP, or histones. Additionally, preferred cationic or polycationic proteins or peptides may be selected from the following proteins or peptides having the following total formula: $(Arg)_l;(Lys)_m;(His)_n;(Orn)_o;(Xaa)_x$, wherein l + m + n +o + x = 8-15, and l, m, n or o independently of each other may be any number selected from 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14 or 15, provided that the overall content of Arg, Lys, His and Orn represents at least 50% of all amino acids of the oligopeptide; and Xaa may be any amino acid selected from native (= naturally occurring) or non-native amino acids except of Arg, Lys, His or Orn; and x may be any number selected from 0, 1, 2, 3 or 4, provided, that the overall content of Xaa does not exceed 50 % of all amino acids of the oligopeptide.

Particularly preferred cationic peptides in this context are e.g. Arg_7, Arg_8, Arg_9, H_3R_9, R_9H_3, $H_3R_9H_3$, $YSSR_9SSY$, $(RKH)_4$, $Y(RKH)_2R$, etc. Further preferred cationic or polycationic compounds, which can be used as transfection agent may include cationic polysaccharides, for example chitosan, polybrene, cationic polymers, e.g. polyethyleneimine (PEI), cationic lipids, e.g. DOTMA: [1-(2,3-sioleyloxy)propyl)]-N,N,N-trimethylammonium chloride, DMRIE, di-C14-amidine, DOTIM, SAINT, DC-Chol, BGTC, CTAP, DOPC, DODAP, DOPE: Dioleyl phosphatidylethanol-amine, DOSPA, DODAB, DOIC, DMEPC, DOGS: Dioctadecylamidoglicylspermin, DIMRI: Dimyristo-oxypropyl dimethyl hydroxyethyl ammonium bromide, DOTAP: dioleoyloxy-3-(trimethylammonio)propane, DC-6-14: O,O-ditetradecanoyl-N-(α-trimethylammonioacetyl)diethanolamine chloride, CLIP1: rac-[(2,3-dioctadecyloxypropyl)(2-hydroxyethyl)]-dimethylammonium chloride, CLIP6: rac-[2(2,3-dihexadecyloxypropyl-oxymethyloxy)ethyl]trimethylammonium, CLIP9: rac-[2(2,3-dihexadecyloxypropyl-oxysuccinyloxy)ethyl]-trimethylammonium, oligofectamine, or cationic or polycationic polymers, e.g. modified polyaminoacids, such as β-aminoacid-polymers or reversed polyamides, etc., modified polyethylenes, such as PVP (poly(N-ethyl-4-vinylpyridinium bromide)), etc., modified acrylates, such as pDMAEMA (poly(dimethylaminoethyl methylacrylate)), etc., modified Amidoamines such as pAMAM (poly(amidoamine)), etc., modified polybetaaminoester (PBAE), such as diamine end modified 1,4 butanediol diacrylate-co-5-amino-1-pentanol polymers, etc., dendrimers, such as polypropylamine dendrimers or pAMAM based dendrimers, etc., polyimine(s), such as PEI: poly(ethyleneimine), poly(propyleneimine), etc., polyallylamine, sugar backbone based polymers, such as cyclodextrin based polymers, dextran based polymers, chitosan, etc., silan backbone based polymers, such as PMOXA-PDMS copolymers, etc., blockpolymers consisting of a combination of one or more cationic blocks (e.g. selected from a cationic polymer as mentioned above) and of one or more hydrophilic or hydrophobic blocks (e.g polyethyleneglycole); etc.

In this context it is particularly preferred that the inventive nucleic acid is complexed at least partially with a cationic or polycationic compound, preferably cationic proteins or peptides. Partially means that only a part of the inventive nucleic acid is complexed with a cationic or polycationic compound and that the rest of the inventive nucleic acid is in uncomplexed form ("free"). Preferably the ratio of complexed nucleic acid to: free nucleic acid is selected from a range. of about 5:1 (w/w) to about 1:10 (w/w), more preferably from

a range of about 4:1 (w/w) to about 1:8 (w/w), even more preferably from a range of about 3:1 (w/w) to about 1:5 (w/w) or 1:3 (w/w), and most preferably the ratio of complexed nucleic acid to free nucleic acid is selected from a ratio of about 1:1 (w/w).

It is preferred that the nucleic acid sequence of the invention is provided in either naked form or complexed, e.g. by polycationic compounds of whatever chemical structure, preferably polycationic (poly)peptides or synthetic polycationic compounds. Preferably, the nucleic acid sequence is not provided together with a packaging cell.

In a further aspect the invention provides for a composition or kit or kit of parts comprising a plurality or more than one, preferably 2 to 10, more preferably 2 to 5, most preferably 2 to 4 of the inventive nucleic acid sequences as defined herein. These inventive compositions comprise more than one inventive nucleic acid sequences, preferably encoding different peptides or proteins which comprise preferably different tumour antigens or fragments, variants or derivatives thereof.

In a preferred embodiment the inventive composition or kit or kit of parts comprising a plurality (which means typically more than 1, 2, 3, 4, 5, 6 or more than 10 nucleic acids, e.g. 2 to 10, preferably 2 to 5 nucleic acids) of inventive nucleic acid sequences, particularly for use in the treatment of prostate cancer (PCa) comprises at least:

a) an inventive nucleic acid encoding at least one peptide or protein, wherein said encoded peptide or protein comprises the tumour antigen PSA, or a fragment, variant or derivative thereof; and
b) an inventive nucleic acid encoding at least one peptide or protein, wherein said encoded peptide or protein comprises the tumour antigen PSMA, or a fragment, variant or derivative thereof; and
c) an inventive nucleic acid encoding at least one peptide or protein, wherein said encoded peptide or protein comprises the tumour antigen PSCA, or a fragment, variant or derivative thereof; and
d) an inventive nucleic acid encoding at least one peptide or protein, wherein said encoded peptide or protein comprises the tumour antigen STEAP-1, or a fragment, variant or derivative thereof.

In a further preferred embodiment the inventive composition or kit or kit of parts comprising a plurality (which means typically more than 1, 2, 3, 4, 5, 6 or more than 10 nucleic acids, e.g. 2 to 10, preferably 2 to 5 nucleic acids) of inventive nucleic acid sequences, particularly for use in the treatment of non-small lung cancer (NSCLC) comprises at least:

a) a nucleic acid sequence comprising or coding for
   i. a coding region, encoding at least one peptide or protein which comprises the tumour antigen NY-ESO-1, or a fragment, variant or derivative thereof,
   ii. at least one histone stem-loop, and
   iii. a poly(A) sequence or a polyadenylation signal;
b) an inventive nucleic acid encoding at least one peptide or protein, wherein said encoded peptide or protein comprises the tumour antigen 5T4, or a fragment, variant or derivative thereof; and
c) an inventive nucleic acid encoding at least one peptide or protein, wherein said encoded peptide or protein comprises the tumour antigen Survivin, or a fragment, variant or derivative thereof.

Furthermore in an alternative, the inventive composition or kit or kit of parts comprising a plurality (which means typically more than 1, 2, 3, 4, 5, 6 or more than 10 nucleic acids, e.g. 2 to 10, preferably 2 to 5 nucleic acids) of inventive nucleic acid sequences, particularly for use in the treatment of non-small lung cancer (NSCLC) comprises at least:

a) a nucleic acid sequence comprising or coding for
   i. a coding region, encoding at least one peptide or protein which comprises the tumour antigen NY-ESO-1, or a fragment, variant or derivative thereof,
   ii. at least one histone stem-loop, and
   iii. a poly(A) sequence or a polyadenylation signal;
b) an inventive nucleic acid encoding at least one peptide or protein, wherein said encoded peptide or protein comprises the tumour antigen 5T4, or a fragment, variant or derivative thereof; and

c) an inventive nucleic acid encoding at least one peptide or protein, wherein said encoded peptide or protein comprises the tumour antigen Survivin, or a fragment, variant or derivative thereof; and

d) an inventive nucleic acid encoding at least one peptide or protein, wherein said encoded peptide or protein comprises the tumour antigen MAGE-C1, or a fragment, variant or derivative thereof; and

e) an inventive nucleic acid encoding at least one peptide or protein, wherein said encoded peptide or protein comprises the tumour antigen MAGE-C2, or a fragment, variant or derivative thereof.

According to a further aspect, the present invention also provides a method for increasing the expression of an encoded peptide or protein comprising the steps, e.g. a) providing the inventive nucleic acid sequence as defined herein or the inventive composition comprising a plurality (which means typically more than 1, 2, 3, 4, 5, 6 or more than 10 nucleic acids, e.g. 2 to 10, preferably 2 to 5 nucleic acids) of inventive nucleic acid sequences as defined herein, b) applying or administering the inventive nucleic acid sequence as defined herein or the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein to an expression system, e.g. to a cell-free expression system, a cell (e.g. an expression host cell or a somatic cell), a tissue or an organism. The method may be applied for laboratory, for research, for diagnostic, for commercial production of peptides or proteins and/or for therapeutic purposes. In this context, typically after preparing the inventive nucleic acid sequence as defined herein or of the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein, it is typically applied or administered to a cell-free expression system, a cell (e.g. an expression host cell or a somatic cell), a tissue or an organism, e.g. in naked or complexed form or as a pharmaceutical composition or vaccine as described herein, preferably via transfection or by using any of the administration modes as described herein. The method may be carried out *in vitro*, *in vivo* or *ex vivo*. The method may furthermore be carried out in the context of the treatment of a specific disease, particularly in the treatment of cancer or tumour diseases, preferably as defined herein.

In this context *in vitro* is defined herein as transfection or transduction of the inventive nucleic acid as defined herein or of the inventive composition comprising a plurality of

inventive nucleic acid sequences as defined herein into cells in culture outside of an organism; *in vivo* is defined herein as transfection or transduction of the inventive nucleic acid or of the inventive composition comprising a plurality of inventive nucleic acid sequences into cells by application of the inventive nucleic acid or of the inventive composition to the whole organism or individual and *ex vivo* is defined herein as transfection or transduction of the inventive nucleic acid or of the inventive composition comprising a plurality of inventive nucleic acid sequences (which means typically more than 1, 2, 3, 4, 5, 6 or more than 10 nucleic acids, e.g. 2 to 10, preferably 2 to 5 nucleic acids) into cells outside of an organism or individual and subsequent application of the transfected cells to the organism or individual.

Likewise, according to another aspect, the present invention also provides the use of the inventive nucleic acid sequence as defined herein or of the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein, preferably for diagnostic or therapeutic purposes, for increasing the expression of an encoded peptide or protein, e.g. by applying or administering the inventive nucleic acid sequence as defined herein or of the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein, e.g. to a cell-free expression system, a cell (e.g. an expression host cell or a somatic cell), a tissue or an organism. The use may be applied for laboratory, for research, for diagnostic for commercial production of peptides or proteins and/or for therapeutic purposes. In this context, typically after preparing the inventive nucleic acid sequence as defined herein or of the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein, it is typically applied or administered to a cell-free expression system, a cell (e.g. an expression host cell or a somatic cell), a tissue or an organism, preferably in naked form or complexed form, or as a pharmaceutical composition or vaccine as described herein, preferably via transfection or by using any of the administration modes as described herein. The use may be carried out *in vitro, in vivo* or *ex vivo*. The use may furthermore be carried out in the context of the treatment of a specific disease, particularly in the treatment of cancer or tumour diseases, preferably as defined herein.

In yet another aspect the present invention also relates to an inventive expression system comprising an inventive nucleic acid sequence or expression vector or plasmid according to

the first aspect of the present invention. In this context the expression system may be a cell-free expression system (e.g. an *in vitro* transcription/translation system), a cellular expression system (e.g. mammalian cells like CHO cells, insect cells, yeast cells, bacterial cells like *E. coli*) or organisms used for expression of peptides or proteins (e.g. plants or animals like cows).

Additionally, according to another aspect, the present invention also relates to the use of the inventive nucleic acid as defined herein or of the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein for the preparation of a pharmaceutical composition for increasing the expression of an encoded peptide or protein, e.g. for treating a cancer or tumour disease, preferably as defined herein, e.g. applying or administering the inventive nucleic acid as defined herein or of the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein to a cell (e.g. an expression host cell or a somatic cell), a tissue or an organism, preferably in naked form or complexed form or as a pharmaceutical composition or vaccine as described herein, more preferably using any of the administration modes as described herein.

Accordingly, in a particular preferred aspect, the present invention also provides a pharmaceutical composition, comprising an inventive nucleic acid as defined herein or an inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein and optionally a pharmaceutically acceptable carrier and/or vehicle.

As a first ingredient, the inventive pharmaceutical composition comprises at least one inventive nucleic acid as defined herein.

As a second ingredient the inventive pharmaceutical composition may optional comprise at least one additional pharmaceutically active component. A pharmaceutically active component in this connection is a compound that has a therapeutic effect to heal, ameliorate or prevent a particular indication or disease as mentioned herein, preferably cancer or tumour diseases. Such compounds include, without implying any limitation, peptides or proteins, preferably as defined herein, nucleic acids, preferably as defined herein, (therapeutically active) low molecular weight organic or inorganic compounds (molecular weight less than 5000, preferably less than 1000), sugars, antigens or antibodies,

preferably as defined herein, therapeutic agents already known in the prior art, antigenic cells, antigenic cellular fragments, cellular fractions; cell wall components (e.g. polysaccharides), modified, attenuated or de-activated (e.g. chemically or by irradiation) pathogens (virus, bacteria etc.), adjuvants, preferably as defined herein, etc.

Furthermore, the inventive pharmaceutical composition may comprise a pharmaceutically acceptable carrier and/or vehicle. In the context of the present invention, a pharmaceutically acceptable carrier typically includes the liquid or non-liquid basis of the inventive pharmaceutical composition. If the inventive pharmaceutical composition is provided in liquid form, the carrier will typically be pyrogen-free water; isotonic saline or buffered (aqueous) solutions, e.g phosphate, citrate etc. buffered solutions. The injection buffer may be hypertonic, isotonic or hypotonic with reference to the specific reference medium, i.e. the buffer may have a higher, identical or lower salt content with reference to the specific reference medium, wherein preferably such concentrations of the afore mentioned salts may be used, which do not lead to damage of cells due to osmosis or other concentration effects. Reference media are e.g. liquids occurring in "*in vivo*" methods, such as blood, lymph, cytosolic liquids, or other body liquids, or e.g. liquids, which may be used as reference media in "*in vitro*" methods, such as common buffers or liquids. Such common buffers or liquids are known to a skilled person. Ringer-Lactate solution is particularly preferred as a liquid basis.

However, one or more compatible solid or liquid fillers or diluents or encapsulating compounds may be used as well for the inventive pharmaceutical composition, which are suitable for administration to a patient to be treated. The term "compatible" as used here means that these constituents of the inventive pharmaceutical composition are capable of being mixed with the inventive nucleic acid as defined herein in such a manner that no interaction occurs which would substantially reduce the pharmaceutical effectiveness of the inventive pharmaceutical composition under typical use conditions.

According to a specific embodiment, the inventive pharmaceutical composition may comprise an adjuvant. In this context, an adjuvant may be understood as any compound, which is suitable to initiate or increase an immune response of the innate immune system, i.e. a non-specific immune response. With other words, when administered, the inventive

pharmaceutical composition preferably elicits an innate immune response due to the adjuvant, optionally contained therein. Preferably, such an adjuvant may be selected from an adjuvant known to a skilled person and suitable for the present case, i.e. supporting the induction of an innate immune response in a mammal, e.g. an adjuvant protein as defined above or an adjuvant as defined in the following.

Particularly preferred as adjuvants suitable for depot and delivery are cationic or polycationic compounds as defined above for the inventive nucleic acid sequence as vehicle, transfection or complexation agent.

The inventive pharmaceutical composition can additionally contain one or more auxiliary substances in order to increase its immunogenicity or immunostimulatory capacity, if desired. A synergistic action of the inventive nucleic acid sequence as defined herein and of an auxiliary substance, which may be optionally contained in the inventive pharmaceutical composition, is preferably achieved thereby. Depending on the various types of auxiliary substances, various mechanisms can come into consideration in this respect. For example, compounds that permit the maturation of dendritic cells (DCs), for example lipopolysaccharides, TNF-alpha or CD40 ligand, form a first class of suitable auxiliary substances. In general, it is possible to use as auxiliary substance any agent that influences the immune system in the manner of a "danger signal" (LPS, GP96, etc.) or cytokines, such as GM-CFS, which allow an immune response to be enhanced and/or influenced in a targeted manner. Particularly preferred auxiliary substances are cytokines, such as monokines, lymphokines, interleukins or chemokines, that further promote the innate immune response, such as IL-1, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-12, IL-13, IL-14, IL-15, IL-16, IL-17, IL-18, IL-19, IL-20, IL-21, IL-22, IL-23, IL-24, IL-25, IL-26, IL-27, IL-28, IL-29, IL-30, IL-31, IL-32, IL-33, IFN-alpha, IFN-beta, IFN-gamma, GM-CSF, G-CSF, M-CSF, LT-beta or TNF-alpha, growth factors, such as hGH.

Further additives which may be included in the inventive pharmaceutical composition are emulsifiers, such as, for example, Tween®; wetting agents, such as, for example, sodium lauryl sulfate; colouring agents; taste-imparting agents, pharmaceutical carriers; tablet-forming agents; stabilizers; antioxidants; preservatives.

The inventive pharmaceutical composition can also additionally contain any further compound, which is known to be immunostimulating due to its binding affinity (as ligands) to human Toll-like receptors TLR1, TLR2, TLR3, TLR4, TLR5, TLR6, TLR7, TLR8, TLR9, TLR10, or due to its binding affinity (as ligands) to murine Toll-like receptors TLR1, TLR2, TLR3, TLR4, TLR5, TLR6, TLR7, TLR8, TLR9, TLR10, TLR11, TLR12 or TLR13.

The inventive pharmaceutical composition may be administered orally, parenterally, by inhalation spray, topically, rectally, nasally, buccally, vaginally or via an implanted reservoir. The term parenteral as used herein includes subcutaneous, intravenous, intramuscular, intra-articular, intra-synovial, intrasternal, intrathecal, intrahepatic, intralesional, intracranial, transdermal, intradermal, intrapulmonal, intraperitoneal, intracardial, intraarterial, and sublingual injection or infusion techniques.

Preferably, the inventive pharmaceutical composition may be administered by parenteral injection, more preferably by subcutaneous, intravenous, intramuscular, intra-articular, intra-synovial, intrasternal, intrathecal, intrahepatic, intralesional, intracranial, transdermal, intradermal, intrapulmonal, intraperitoneal, intracardial, intraarterial, and sublingual injection or via infusion techniques. Particularly preferred is intradermal and intramuscular injection. Sterile injectable forms of the inventive pharmaceutical compositions may be aqueous or oleaginous suspension. These suspensions may be formulated according to techniques known in the art using suitable dispersing or wetting agents and suspending agents.

The inventive pharmaceutical composition as defined herein may also be administered orally in any orally acceptable dosage form including, but not limited to, capsules, tablets, aqueous suspensions or solutions.

The inventive pharmaceutical composition may also be administered topically, especially when the target of treatment includes areas or organs readily accessible by topical application, e.g. including diseases of the skin or of any other accessible epithelial tissue. Suitable topical formulations are readily prepared for each of these areas or organs. For topical applications, the inventive pharmaceutical composition may be formulated in a

suitable ointment, containing the inventive nucleic acid as defined herein suspended or dissolved in one or more carriers.

The inventive pharmaceutical composition typically comprises a "safe and effective amount" of the components of the inventive pharmaceutical composition, particularly of the inventive nucleic acid sequence(s) as defined herein. As used herein, a "safe and effective amount" means an amount of the inventive nucleic acid sequence(s) as defined herein as such that is sufficient to significantly induce a positive modification of a disease or disorder as defined herein. At the same time, however, a "safe and effective amount" is small enough to avoid serious side-effects and to permit a sensible relationship between advantage and risk. The determination of these limits typically lies within the scope of sensible medical judgment.

The inventive pharmaceutical composition may be used for human and also for veterinary medical purposes, preferably for human medical purposes, as a pharmaceutical composition in general or as a vaccine.

According to another particularly preferred aspect, the inventive pharmaceutical composition (or the inventive nucleic acid sequence as defined herein or the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein) may be provided or used as a vaccine. Typically, such a vaccine is as defined above for pharmaceutical compositions. Additionally, such a vaccine typically contains the inventive nucleic acid as defined herein or the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein.

The inventive vaccine may also comprise a pharmaceutically acceptable carrier, adjuvant, and/or vehicle as defined herein for the inventive pharmaceutical composition. In the specific context of the inventive vaccine, the choice of a pharmaceutically acceptable carrier is determined in principle by the manner in which the inventive vaccine is administered. The inventive vaccine can be administered, for example, systemically or locally. Routes for systemic administration in general include, for example, transdermal, oral, parenteral routes, including subcutaneous, intravenous, intramuscular, intraarterial, intradermal and intraperitoneal injections and/or intranasal administration routes. Routes for

local administration in general include, for example, topical administration routes but also intradermal, transdermal, subcutaneous, or intramuscular injections or intralesional, intracranial, intrapulmonal, intracardial, and sublingual injections. More preferably, vaccines may be administered by an intradermal, subcutaneous, or intramuscular route. Inventive vaccines are therefore preferably formulated in liquid (or sometimes in solid) form.

The inventive vaccine can additionally contain one or more auxiliary substances in order to increase its immunogenicity or immunostimulatory capacity, if desired. Particularly preferred are adjuvants as auxiliary substances or additives as defined for the pharmaceutical composition.

The present invention furthermore provides several applications and uses of the inventive nucleic acid sequence as defined herein, of the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein, of the inventive pharmaceutical composition, of the inventive vaccine, all comprising the inventive nucleic acid sequence as defined herein or of kits comprising same.

According to one specific aspect, the present invention is directed to the first medical use of the inventive nucleic acid sequence as defined herein or of the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein as a medicament, preferably as a vaccine particularly in the treatment of cancer or tumour diseases.

According to another aspect, the present invention is directed to the second medical use of the inventive nucleic acid sequence as defined herein or of the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein, for the treatment of cancer and tumour diseases as defined herein, preferably to the use of the inventive nucleic acid sequence as defined herein, of the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein, of a pharmaceutical composition or vaccine comprising same or of kits comprising same for the preparation of a medicament for the prophylaxis, treatment and/or amelioration of cancer or tumour diseases as defined herein. Preferably, the pharmaceutical composition or a vaccine is used or to be administered to a patient in need thereof for this purpose.

Preferably, diseases as mentioned herein are selected from cancer or tumour diseases which preferably include e.g. Acute lymphoblastic leukemia, Acute myeloid leukemia, Adrenocortical carcinoma, AIDS-related cancers, AIDS-related lymphoma, Anal cancer, Appendix cancer, Astrocytoma, Basal cell carcinoma, Bile duct cancer, Bladder cancer, Bone cancer, Osteosarcoma/Malignant fibrous histiocytoma, Brainstem glioma, Brain tumor, cerebellar astrocytoma, cerebral astrocytoma/malignant glioma, ependymoma, medulloblastoma, supratentorial primitive neuroectodermal tumors, visual pathway and hypothalamic glioma, Breast cancer, Bronchial adenomas/carcinoids, Burkitt lymphoma, childhood Carcinoid tumor, gastrointestinal Carcinoid tumor, Carcinoma of unknown primary, primary Central nervous system lymphoma, childhood Cerebellar astrocytoma, childhood Cerebral astrocytoma/Malignant glioma, Cervical cancer, Childhood cancers, Chronic lymphocytic leukemia, Chronic myelogenous leukemia, Chronic myeloproliferative disorders, Colon Cancer, Cutaneous T-cell lymphoma, Desmoplastic small round cell tumor, Endometrial cancer, Ependymoma, Esophageal cancer, Ewing's sarcoma in the Ewing family of tumors, Childhood Extracranial germ cell tumor, Extragonadal Germ cell tumor, Extrahepatic bile duct cancer, Intraocular melanoma, Retinoblastoma, Gallbladder cancer, Gastric (Stomach) cancer, Gastrointestinal Carcinoid Tumor, Gastrointestinal stromal tumor (GIST), extracranial, extragonadal, or ovarian Germ cell tumor, Gestational trophoblastic tumor, Glioma of the brain stem, Childhood Cerebral Astrocytoma, Childhood Visual Pathway and Hypothalamic Glioma, Gastric carcinoid, Hairy cell leukemia, Head and neck cancer, Heart cancer, Hepatocellular (liver) cancer, Hodgkin lymphoma, Hypopharyngeal cancer, childhood Hypothalamic and visual pathway glioma, Intraocular Melanoma, Islet Cell Carcinoma (Endocrine Pancreas), Kaposi sarcoma, Kidney cancer (renal cell cancer), Laryngeal Cancer, Leukemias, acute lymphoblastic Leukemia, acute myeloid Leukemia, chronic lymphocytic Leukemia, chronic myelogenous Leukemia, hairy cell Leukemia, Lip and Oral Cavity Cancer, Liposarcoma, Liver Cancer, Non-Small Cell Lung Cancer, Small Cell Lung Cancer, Lymphomas, AIDS-related Lymphoma, Burkitt Lymphoma, cutaneous T-Cell Lymphoma, Hodgkin Lymphoma, Non-Hodgkin Lymphomas, Primary Central Nervous System Lymphoma, Waldenström Macroglobulinemia, Malignant Fibrous Histiocytoma of Bone/Osteosarcoma, Childhood Medulloblastoma, Melanoma, Intraocular (Eye) Melanoma, Merkel Cell Carcinoma, Adult Malignant Mesothelioma, Childhood Mesothelioma, Metastatic Squamous Neck Cancer with Occult Primary, Mouth

Cancer, Childhood Multiple Endocrine Neoplasia Syndrome, Multiple Myeloma/Plasma Cell Neoplasm, Mycosis Fungoides, Myelodysplastic Syndromes, Myelodysplastic/Myeloproliferative Diseases, Chronic Myelogenous Leukemia, Adult Acute Myeloid Leukemia, Childhood Acute Myeloid Leukemia, Multiple Myeloma (Cancer of the Bone-Marrow), Chronic Myeloproliferative Disorders, Nasal cavity and paranasal sinus cancer, Nasopharyngeal carcinoma, Neuroblastoma, Oral Cancer, Oropharyngeal cancer, Osteosarcoma/malignant fibrous histiocytoma of bone, Ovarian cancer, Ovarian epithelial cancer (Surface epithelial-stromal tumor), Ovarian germ cell tumor, Ovarian low malignant potential tumor, Pancreatic cancer, islet cell Pancreatic cancer, Paranasal sinus and nasal cavity cancer, Parathyroid cancer, Penile cancer, Pharyngeal cancer, Pheochromocytoma, Pineal astrocytoma, Pineal germinoma, childhood Pineoblastoma and supratentorial primitive neuroectodermal tumors, Pituitary adenoma, Plasma cell neoplasia/Multiple myeloma, Pleuropulmonary blastoma, Primary central nervous system lymphoma, Prostate cancer, Rectal cancer, Renal cell carcinoma (kidney cancer), Cancer of the Renal pelvis and ureter, Retinoblastoma, childhood Rhabdomyosarcoma, Salivary gland cancer, Sarcoma of the Ewing family of tumors, Kaposi Sarcoma, soft tissue Sarcoma, uterine Sarcoma, Sézary syndrome, Skin cancer (nonmelanoma), Skin cancer (melanoma), Merkel cell Skin carcinoma, Small intestine cancer, Squamous cell carcinoma, metastatic Squamous neck cancer with occult primary, childhood Supratentorial primitive neuroectodermal tumor, Testicular cancer, Throat cancer, childhood Thymoma, Thymoma and Thymic carcinoma, Thyroid cancer, childhood Thyroid cancer, Transitional cell cancer of the renal pelvis and ureter, gestational Trophoblastic tumor, Urethral cancer, endometrial Uterine cancer, Uterine sarcoma, Vaginal cancer, childhood Visual pathway and hypothalamic glioma, Vulvar cancer, Waldenström macroglobulinemia, and childhood Wilms tumor (kidney cancer).

In a further preferred aspect, the inventive nucleic acid sequence as defined herein or the inventive composition comprising a plurality of inventive nucleic acid sequences as defined herein may be used for the preparation of a pharmaceutical composition or a vaccine, particularly for purposes as defined herein.

The inventive pharmaceutical composition or vaccine may furthermore be used for the treatment of a disease or a disorder, preferably of cancer or tumour diseases as defined herein.

According to a final aspect, the present invention also provides kits, particularly kits of parts. Such kits, particularly kits of parts, typically comprise as components alone or in combination with further components as defined herein at least one inventive nucleic acid sequence as defined herein, the inventive pharmaceutical composition or vaccine comprising the inventive nucleic acid sequence. The at least one inventive nucleic acid sequence as defined herein, is optionally in combination with further components as defined herein, whereby the at least one nucleic acid of the invention is provided separately (first part of the kit) from at least one other part of the kit comprising one or more other components. The inventive pharmaceutical composition and/or the inventive vaccine may e.g. occur in one or different parts of the kit. As an example, e.g. at least one part of the kit may comprise at least one inventive nucleic acid sequence as defined herein, and at least one further part of the kit at least one other component as defined herein, e.g. at least one other part of the kit may comprise at least one pharmaceutical composition or vaccine or a part thereof, e.g. at least one part of the kit may comprise the inventive nucleic acid sequence as defined herein, at least one further part of the kit at least one other component as defined herein, at least one further part of the kit at least one component of the inventive pharmaceutical composition or vaccine or the inventive pharmaceutical composition or vaccine as a whole, and at least one further part of the kit e.g. at least one pharmaceutical carrier or vehicle, etc. In case the kit or kit of parts comprises a plurality of inventive nucleic acid sequences, one component of the kit can comprise only one, several or all inventive nucleic acid sequences comprised in the kit. In an alternative embodiment each/every inventive nucleic acid sequence may be comprised in a different /separate component of the kit such that each component forms a part of the kit. Also, more than one nucleic acid may be comprised in a first component as part of the kit, whereas one or more other (second, third etc.) components (providing one or more other parts of the kit) may either contain one or more than one inventive nucleic acids, which may be identical or partially identical or different from the first component. The kit or kit of parts may furthermore contain technical instructions with information on the administration and dosage of the

inventive nucleic acid sequence, the inventive pharmaceutical composition or the inventive vaccine or of any of its components or parts, e.g. if the kit is prepared as a kit of parts.

Taken together, the invention provides a nucleic acid sequence comprising or coding for

a) a coding region, encoding at least one peptide or protein;

b) at least one histone stem-loop, and

c) a poly(A) sequence or a polyadenylation signal;

wherein said peptide or protein comprises a tumour antigen a fragment, variant or derivative of said tumour antigen, preferably, wherein the tumour antigen is a melanocyte-specific antigen, a cancer-testis antigen or a tumour-specific antigen, preferably a CT-X antigen, a non-X CT-antigen, a binding partner for a CT-X antigen or a binding partner for a non-X CT-antigen or a tumour-specific antigen, more preferably a CT-X antigen, a binding partner for a non-X CT-antigen or a tumour-specific antigen or a fragment, variant or derivative of said tumour antigen.

In a further preferred embodiment, the invention relates to a composition comprising at least one type of nucleic acid sequence comprising or coding for

a) a coding region, encoding at least one peptide or protein;

b) at least one histone stem-loop, and

c) a poly(A) sequence or a polyadenylation signal;

wherein said peptide or protein comprises a tumour antigen a fragment, variant or derivative of said tumour antigen, preferably, wherein the tumour antigen is a melanocyte-specific antigen, a cancer-testis antigen or a tumour-specific antigen, preferably a CT-X antigen, a non-X CT-antigen, a binding partner for a CT-X antigen or a binding partner for a non-X CT-antigen or a tumour-specific antigen, more preferably a CT-X antigen, a binding partner for a non-X CT-antigen or a tumour-specific antigen or a fragment, variant or derivative of said tumour antigen; and wherein each of the nucleic acid sequences encodes a different peptide or protein.

The composition may comprise further an pharmaceutically acceptable carrier and/or pharmaceutically acceptable adjuvants as defined herein. The composition may be used as a vaccine or for treatment of a disease associated with cancer or tumour.

In a further preferred embodiment, the invention provides a composition comprising at least two, preferably two or more, more preferably a plurality of nucleic acid sequences sequence (which means typically more than 1, 2, 3, 4, 5, 6 or more than 10 nucleic acids, e.g. 2 to 10, preferably 2 to 5 nucleic acids) comprising or coding for

a) a coding region, encoding at least one peptide or protein;

b) at least one histone stem-loop, and

c) a poly(A) sequence or a polyadenylation signal;

wherein said peptide or protein comprises a tumour antigen a fragment, variant or derivative of said tumour antigen, preferably, wherein the tumour antigen is a melanocyte-specific antigen, a cancer-testis antigen or a tumour-specific antigen, preferably a CT-X antigen, a non-X CT-antigen, a binding partner for a CT-X antigen or a binding partner for a non-X CT-antigen or a tumour-specific antigen, more preferably a CT-X antigen, a binding partner for a non-X CT-antigen or a tumour-specific antigen or a fragment, variant or derivative of said tumour antigen; and wherein each of the nucleic acid sequences encodes a different peptide or protein.

The composition may comprise further an pharmaceutically acceptable carrier and/or pharmaceutically acceptable adjuvants as defined herein. The composition may be used as a vaccine or for treatment of a disease associated with cancer or tumour.

In a further preferred embodiment, the invention provides a composition comprising at least two, preferably two or more, more preferably a plurality (which means typically more than 1, 2, 3, 4, 5, 6 or more than 10 nucleic acids, e.g. 2 to 10, preferably 2 to 5 nucleic acids) of nucleic acid sequences sequence comprising or coding for

a) a coding region, encoding at least one peptide or protein;

b) at least one histone stem-loop, and

c) a poly(A) sequence or a polyadenylation signal;

wherein said peptide or protein comprises a tumour antigen a fragment, variant or derivative of said tumour antigen, preferably, wherein the tumour antigen is a melanocyte-specific antigen, a cancer-testis antigen or a tumour-specific antigen, preferably a CT-X antigen, a non-X CT-antigen, a binding partner for a CT-X antigen or a binding partner for a non-X CT-antigen or a tumour-specific antigen, more preferably a CT-X antigen, a binding partner for a non-X CT-antigen or a tumour-specific antigen or a fragment, variant or derivative of said tumour antigen; and

wherein each of the nucleic acid sequences encodes a different peptide or protein; and preferably wherein each type of nucleic acid sequence encodes for a different peptide or protein, preferably for a different tumour antigen.

The composition may comprise further an pharmaceutically acceptable carrier and/or pharmaceutically acceptable adjuvants as defined herein. The composition may be used as a vaccine or for treatment of a disease associated with cancer or tumour.

In a further preferred embodiment, the invention provides a composition comprising at least two, preferably two or more, more preferably a plurality of nucleic acid sequences sequence (which means typically more than 1, 2, 3, 4, 5, 6 or more than 10 nucleic acids, e.g. 2 to 10, preferably 2 to 5 nucleic acids) comprising or coding for

a) a coding region, encoding at least one peptide or protein;
b) at least one histone stem-loop, and
c) a poly(A) sequence or a polyadenylation signal;

wherein said peptide or protein comprises a tumour antigen a fragment, variant or derivative of said tumour antigen, preferably, wherein the tumour antigen is a melanocyte-specific antigen, a cancer-testis antigen or a tumour-specific antigen, preferably a CT-X antigen, a non-X CT-antigen, a binding partner for a CT-X antigen or a binding partner for a non-X CT-antigen or a tumour-specific antigen, more preferably a CT-X antigen, a binding partner for a non-X CT-antigen or a tumour-specific antigen or a fragment, variant or derivative of said tumour antigen; and wherein each of the nucleic acid sequences encodes a different peptide or protein; and preferably wherein each type of nucleic acid sequence encodes for a different peptide or protein, preferably for a different tumour antigen, more preferably, wherein one type of the contained nucleic acid sequences encodes for PSA, PSMA, PSCA, STEAP-1, NY-ESO-1, 5T4, Survivin, MAGE-C1, or MAGE-C2.

The composition may comprise further an pharmaceutically acceptable carrier and/or pharmaceutically acceptable adjuvants as defined herein. The composition may be used as a vaccine or for treatment of a disease associated with cancer or tumour.

In some embodiments, it may be preferred, provided that the composition contains only one type of nucleic acid sequence, if the nucleic acid sequence does not encode for NY-ESO1, provided that the composition contains only one type of nucleic acid sequence.

In a further preferred embodiment, the invention provides a composition comprising at least two nucleic acid sequences sequence comprising or coding for

a) a coding region, encoding at least one peptide or protein;

b) at least one histone stem-loop, and

c) a poly(A) sequence or a polyadenylation signal;

wherein said peptide or protein comprises a tumour antigen a fragment, variant or derivative of said tumour antigen, preferably, wherein the tumour antigen is a melanocyte-specific antigen, a cancer-testis antigen or a tumour-specific antigen, preferably a CT-X antigen, a non-X CT-antigen, a binding partner for a CT-X antigen or a binding partner for a non-X CT-antigen or a tumour-specific antigen, more preferably a CT-X antigen, a binding partner for a non-X CT-antigen or a tumour-specific antigen or a fragment, variant or derivative of said tumour antigen; and wherein each of the nucleic acid sequences encodes a different peptide or protein; and wherein at least one of the nucleic acid sequences encodes for 5T4, 707-AP, 9D7, AFP, AlbZIP HPG1, alpha-5-beta-1-integrin, alpha-5-beta-6-integrin, alpha-actinin-4/m, alpha-methylacyl-coenzyme A racemase, ART-4, ARTC1/m, B7H4, BAGE-1, BCL-2, bcr/abl, beta-catenin/m, BING-4, BRCA1/m, BRCA2/m, CA 15-3/CA 27-29, CA 19-9, CA72-4, CA125, calreticulin, CAMEL, CASP-8/m, cathepsin B, cathepsin L, CD19, CD20, CD22, CD25, CDE30, CD33, CD4, CD52, CD55, CD56, CD80, CDC27/m, CDK4/m, CDKN2A/m, CEA, CLCA2, CML28, CML66, COA-1/m, coactosin-like protein, collage XXIII, COX-2, CT-9/BRD6, Cten, cyclin B1, cyclin D1, cyp-B, CYPB1, DAM-10, DAM-6, DEK-CAN, EFTUD2/m, EGFR, ELF2/m, EMMPRIN, EpCam, EphA2, EphA3, ErbB3, ETV6-AML1, EZH2, FGF-5, FN, Frau-1, G250, GAGE-1, GAGE-2, GAGE-3, GAGE-4, GAGE-5, GAGE-6, GAGE7b, GAGE-8, GDEP, GnT-V, gp100, GPC3, GPNMB/m, HAGE, HAST-2, hepsin, Her2/neu, HERV-K-MEL, HLA-A*0201-R17I, HLA-A11/m, HLA-A2/m, HNE, homeobox NKX3.1, HOM-TES-14/SCP-1, HOM-TES-85, HPV-E6, HPV-E7, HSP70-2M, HST-2, hTERT, iCE, IGF-1R, IL-13Ra2, IL-2R, IL-5, immature laminin receptor, kallikrein-2, kallikrein-4, Ki67, KIAA0205, KIAA0205/m, KK-LC-1, K-Ras/m, LAGE-A1, LDLR-FUT, MAGE-A1, MAGE-A2, MAGE-A3, MAGE-A4, MAGE-A6, MAGE-A9, MAGE-A10, MAGE-A12, MAGE-B1, MAGE-B2, MAGE-B3, MAGE-B4, MAGE-B5, MAGE-B6, MAGE-B10, MAGE-B16, MAGE-B17, MAGE-C1, MAGE-C2, MAGE-C3, MAGE-

D1, MAGE-D2, MAGE-D4, MAGE-E1, MAGE-E2, MAGE-F1, MAGE-H1, MAGEL2, mammaglobin A, MART-1/melan-A, MART-2, MART-2/m, matrix protein 22, MC1R, M-CSF, ME1/m, mesothelin, MG50/PXDN, MMP11, MN/CA IX-antigen, MRP-3, MUC-1, MUC-2, MUM-1/m, MUM-2/m, MUM-3/m, myosin class I/m, NA88-A, N-acetylglucosaminyltransferase-V, Neo-PAP, Neo-PAP/m, NFYC/m, NGEP, NMP22, NPM/ALK, N-Ras/m, NSE, NY-ESO-B, OA1, OFA-iLRP, OGT, OGT/m, OS-9, OS-9/m, osteocalcin, osteopontin, p15, p190 minor bcr-abl, p53, p53/m, PAGE-4, PAI-1, PAI-2, PAP, PART-1, PATE, PDEF, Pim-1-Kinase, Pin-1, Pml/PARalpha, POTE, PRAME, PRDX5/m, prostein, proteinase-3, PSA, PSCA, PSGR, PSM, PSMA, PTPRK/m, RAGE-1, RBAF600/m, RHAMM/CD168, RU1, RU2, S-100, SAGE, SART-1, SART-2, SART-3, SCC, SIRT2/m, Sp17, SSX-1, SSX-2/HOM-MEL-40, SSX-4, STAMP-1, STEAP-1, survivin, survivin-2B, SYT-SSX-1, SYT-SSX-2, TA-90, TAG-72, TARP, TEL-AML1, TGFbeta, TGFbetaRII, TGM-4, TPI/m, TRAG-3, TRG, TRP-1, TRP-2/6b, TRP/INT2, TRP-p8, tyrosinase, UPA, VEGFR1, VEGFR-2/FLK-1, WT1 and a immunoglobulin idiotype of a lymphoid blood cell or a T cell receptor idiotype of a lymphoid blood cell, or a fragment, variant or derivative of said tumour antigen; preferably survivin or a homologue thereof, an antigen from the MAGE-family or a binding partner thereof or a fragment, variant or derivative of said tumour antigen.

The composition may comprise further an pharmaceutically acceptable carrier and/or pharmaceutically acceptable adjuvants as defined herein. The composition may be used as a vaccine or for treatment of a disease associated with cancer or tumour.

In the present invention, if not otherwise indicated, different features of alternatives and embodiments may be combined with each other. Furthermore, the term "comprising" shall not be construed as meaning "consisting of", if not specifically mentioned. However, in the context of the present invention, term "comprising" may be substituted with the term "consisting of", where applicable.

Figures:

The following Figures are intended to illustrate the invention further and shall not be construed to limit the present invention thereto.

Figure 1: shows the histone stem-loop consensus sequence generated from metazoan and protozoan stem loop sequences (as reported by Dávila López, M., & Samuelsson, T. (2008), RNA (New York, N.Y.), 14(1), 1-10. doi:10.1261/rna.782308). 4001 histone stem-loop sequences from metazoa and protozoa were aligned and the quantity of the occurring nucleotides is indicated for every position in the stem-loop sequence. The generated consensus sequence representing all nucleotides present in the sequences analyzed is given using the single-letter nucleotide code. In addition to the consensus sequence, sequences are shown representing at least 99%, 95% and 90% of the nucleotides present in the sequences analyzed.

Figure 2: shows the histone stem-loop consensus sequence generated from protozoan stem loop sequences (as reported by Dávila López, M., & Samuelsson, T. (2008), RNA (New York, N.Y.), 14(1), 1-10. doi:10.1261/rna.782308). 131 histone stem-loop sequences from protozoa were aligned and the quantity of the occurring nucleotides is indicated for every position in the stem-loop sequence. The generated consensus sequence representing all nucleotides present in the sequences analyzed is given using the single-letter nucleotide code. In addition to the consensus sequence, sequences are shown representing at least 99%, 95% and 90% of the nucleotides present in the sequences analyzed.

Figure 3: shows the histone stem-loop consensus sequence generated from metazoan stem loop sequences (as reported by Dávila López, M., & Samuelsson, T. (2008), RNA (New York, N.Y.), 14(1), 1-10. doi:10.1261/rna.782308). 3870 histone stem-loop sequences from metazoa were aligned and the quantity of the occurring nucleotides is indicated for every position in the stem-loop

sequence. The generated consensus sequence representing all nucleotides present in the sequences analyzed is given using the single-letter nucleotide code. In addition to the consensus sequence, sequences are shown representing at least 99%, 95% and 90% of the nucleotides present in the sequences analyzed.

Figure 4: shows the histone stem-loop consensus sequence generated from vertebrate stem loop sequences (as reported by Dávila López, M., & Samuelsson, T. (2008), RNA (New York, N.Y.), 14(1), 1-10. doi:10.1261/rna.782308). 1333 histone stem-loop sequences from vertebrates were aligned and the quantity of the occurring nucleotides is indicated for every position in the stem-loop sequence. The generated consensus sequence representing all nucleotides present in the sequences analyzed is given using the single-letter nucleotide code. In addition to the consensus sequence, sequences are shown representing at least 99%, 95% and 90% of the nucleotides present in the sequences analyzed.

Figure 5: shows the histone stem-loop consensus sequence generated from human (Homo sapiens) stem loop sequences (as reported by Dávila López, M., & Samuelsson, T. (2008), RNA (New York, N.Y.), 14(1), 1-10. doi:10.1261/rna.782308). 84 histone stem-loop sequences from humans were aligned and the quantity of the occurring nucleotides is indicated for every position in the stem-loop sequence. The generated consensus sequence representing all nucleotides present in the sequences analyzed is given using the single-letter nucleotide code. In addition to the consensus sequence, sequences are shown representing at least 99%, 95% and 90% of the nucleotides present in the sequences analyzed.

Figures 6 to 21: show mRNAs from *in vitro* transcription.

Given are the designation and the sequence of mRNAs obtained by *in vitro* transcription. The following abbreviations are used:

ppLuc (GC): GC-enriched mRNA sequence coding for *Photinus pyralis* luciferase

ag:	3' untranslated region (UTR) of the alpha globin gene
A64:	poly(A)-sequence with 64 adenylates
A120:	poly(A)-sequence wit 120 adenylates
histoneSL:	histone stem-loop
aCPSL:	stem loop which has been selected from a library for its specific binding of the αCP-2KL protein
PolioCL:	5' clover leaf from Polio virus genomic RNA
G30:	poly(G) sequence with 30 guanylates
U30:	poly(U) sequence with 30 uridylates
SL:	unspecific/artificial stem-loop
N32:	unspecific sequence of 32 nucleotides
NY-ESO-1 (G/C):	GC-enriched mRNA sequence coding for the human tumour antigen NY-ESO-1
Survivin(G/C):	GC-enriched mRNA sequence coding for the human tumour antigen Survivin
MAGE-C1(G/C):	GC-enriched mRNA sequence coding for the human tumour antigen MAGE-C1

Within the sequences, the following elements are highlighted: coding region (ORF) (capital letters), ag (bold), histoneSL (underlined), further distinct sequences tested (italic).

Figure 6: shows the mRNA sequence of ppLuc(GC) – ag (SEQ ID NO: 43).
By linearization of the original vector at the restriction site immediately following the alpha-globin 3'-UTR (ag), mRNA is obtained lacking a poly(A) sequence.

Figure 7: shows the mRNA sequence of ppLuc(GC) – ag – A64 (SEQ ID NO: 44).
By linearization of the original vector at the restriction site immediately following the A64 poly(A)-sequence, mRNA is obtained ending with an A64 poly(A) sequence.

Figure 8: shows the mRNA sequence of ppLuc(GC) – ag – histoneSL (SEQ ID NO: 45).
The A64 poly(A) sequence was replaced by a histoneSL. The histone stem-loop sequence used in the examples was obtained from Cakmakci *et al.* (2008). Molecular and Cellular Biology, 28(3), 1182-1194.

Figure 9: shows the mRNA sequence of ppLuc(GC) – ag – A64 – histoneSL (SEQ ID NO: 46).
The histoneSL was appended 3' of A64 poly(A).

Figure 10: shows the mRNA sequence of ppLuc(GC) – ag – A120 (SEQ ID NO: 47). The A64 poly(A) sequence was replaced by an A120 poly(A) sequence.

Figure 11: shows the mRNA sequence of ppLuc(GC) – ag – A64 – ag (SEQ ID NO: 48). A second alpha-globin 3'-UTR was appended 3' of A64 poly(A).

Figure 12: shows the mRNA sequence of ppLuc(GC) – ag – A64 – aCPSL (SEQ ID NO: 49).
A stem loop was appended 3' of A64 poly(A). The stem loop has been selected from a library for its specific binding of the αCP-2KL protein (Thisted *et al.*, (2001), The Journal of Biological Chemistry, 276(20), 17484-17496). αCP-2KL is an isoform of αCP-2, the most strongly expressed αCP protein (alpha-globin mRNA poly(C) binding protein) (Makeyev *et al.*, (2000), Genomics, 67(3), 301-316), a group of RNA binding proteins, which bind to the alpha-globin 3'-UTR (Chkheidze *et al.*, (1999), Molecular and Cellular Biology, 19(7), 4572-4581).

Figure 13: shows the mRNA sequence of ppLuc(GC) – ag – A64 – PolioCL (SEQ ID NO: 50).
The 5' clover leaf from Polio virus genomic RNA was appended 3' of A64 poly(A).

Figure 14: shows the mRNA sequence of ppLuc(GC) – ag – A64 – G30 (SEQ ID NO: 51)
A stretch of 30 guanylates was appended 3' of A64 poly(A).

Figure 15: shows the mRNA sequence of ppLuc(GC) – ag – A64 – U30 (SEQ ID NO: 52)
A stretch of 30 uridylates was appended 3' of A64 poly(A).

Figure 16: shows the mRNA sequence of ppLuc(GC) – ag – A64 – SL (SEQ ID NO: 53)
A stem loop was appended 3' of A64 poly(A). The upper part of the stem and the loop were taken from (Babendure *et al.*, (2006), RNA (New York, N.Y.),

12(5), 851-861). The stem loop consists of a 17 base pair long, CG-rich stem and a 6 base long loop.

Figure 17: shows ppLuc(GC) – ag – A64 – N32 (SEQ ID NO: 54)
By linearization of the original vector at an alternative restriction site, mRNA is obtained with 32 additional nucleotides following poly(A).

Figure 18: shows the mRNA sequence of NY-ESO-1(GC) – ag – A64 –C30 (SEQ ID NO: 55)

Figure 19: shows the mRNA sequence of NY-ESO-1(GC) – ag – A64 –C30 - histoneSL (SEQ ID NO: 56)

Figure 20: shows the mRNA sequence of Survivin(GC) – ag – A64 –C30 - histoneSL (SEQ ID NO: 57)

Figure 21: shows the mRNA sequence of MAGE-C1(GC) – ag – A64 – C30 - histoneSL (SEQ ID NO: 58)

Figure 22: shows that the combination of poly(A) and histoneSL increases protein expression from mRNA in a synergistic manner.
The effect of poly(A) sequence, histoneSL, and the combination of poly(A) and histoneSL on luciferase expression from mRNA was examined. Therefore different mRNAs were electroporated into HeLa cells. Luciferase levels were measured at 6, 24, and 48 hours after transfection. Little luciferase is expressed from mRNA having neither poly(A) sequence nor histoneSL. Both a poly(A) sequence or the histoneSL increase the luciferase level. Strikingly however, the combination of poly(A) and histoneSL further strongly increases the luciferase level, manifold above the level observed with either of the individual elements, thus acting synergistically. Data are graphed as mean RLU ± SD (relative light units ± standard deviation) for triplicate transfections. Specific RLU are summarized in Example 14.2.

Figure 23: shows that the combination of poly(A) and histoneSL increases protein expression from mRNA irrespective of their order.

The effect of poly(A) sequence, histoneSL, the combination of poly(A) and histoneSL, and their order on luciferase expression from mRNA was examined. Therefore different mRNAs were lipofected into HeLa cells. Luciferase levels were measured at 6, 24, and 48 hours after the start of transfection. Both an A64 poly(A) sequence or the histoneSL give rise to comparable luciferase levels. Increasing the length of the poly(A) sequence from A64 to A120 or to A300 increases the luciferase level moderately. In contrast, the combination of poly(A) and histoneSL increases the luciferase level much further than lengthening of the poly(A) sequence. The combination of poly(A) and histoneSL acts synergistically as it increases the luciferase level manifold above the level observed with either of the individual elements. The synergistic effect of the combination of poly(A) and histoneSL is seen irrespective of the order of poly(A) and histoneSL and irrespective of the length of poly(A) with A64-histoneSL or histoneSL-A250 mRNA. Data are graphed as mean RLU ± SD for triplicate transfections. Specific RLU are summarized in Example 14.3.

Figure 24: shows that the rise in protein expression by the combination of poly(A) and histoneSL is specific.

The effect of combining poly(A) and histoneSL or poly(A) and alternative sequences on luciferase expression from mRNA was examined. Therefore different mRNAs were electroporated into HeLa cells. Luciferase levels were measured at 6, 24, and 48 hours after transfection. Both a poly(A) sequence or the histoneSL give rise to comparable luciferase levels. The combination of poly(A) and histoneSL strongly increases the luciferase level, manifold above the level observed with either of the individual elements, thus acting synergistically. In contrast, combining poly(A) with any of the other sequences is without effect on the luciferase level compared to mRNA containing only a poly(A) sequence. Thus, the combination of poly(A) and histoneSL acts specifically and synergistically. Data are graphed as mean

RLU ± SD for triplicate transfections. Specific RLU are summarized in Example 14.4.

Figure 25: shows that the combination of poly(A) and histoneSL increases protein expression from mRNA in a synergistic manner in vivo.
The effect of poly(A) sequence, histoneSL, and the combination of poly(A) and histoneSL on luciferase expression from mRNA in vivo was examined. Therefore different mRNAs were injected intradermally into mice. Mice were sacrificed 16 hours after injection and Luciferase levels at the injection sites were measured. Luciferase is expressed from mRNA having either a histoneSL or a poly(A) sequence. Strikingly however, the combination of poly(A) and histoneSL strongly increases the luciferase level, manifold above the level observed with either of the individual elements, thus acting synergistically. Data are graphed as mean RLU ± SEM (relative light units ± standard error of mean). Specific RLU are summarized in Example 14.5.

Figure 26: shows that the combination of poly(A) and histoneSL increases NY-ESO-1 protein expression from mRNA.
The effect of poly(A) sequence and the combination of poly(A) and histoneSL on NY-ESO-1 expression from mRNA was examined. Therefore different mRNAs were electroporated into HeLa cells. NY-ESO-1 levels were measured at 24 hours after transfection by flow cytometry. NY-ESO-1 is expressed from mRNA having only a poly(A) sequence. Strikingly however, the combination of poly(A) and histoneSL strongly increases the NY-ESO-1 level, manifold above the level observed with only a poly(A) sequence. Data are graphed as counts against fluorescence intensity. Median fluorescence intensities (MFI) are summarized in Example 14.6.

Figure 27: shows that the combination of poly(A) and histoneSL increases the level of antibodies elicited by vaccination with mRNA.
The effect of poly(A) sequence and the combination of poly(A) and histoneSL on the induction of anti NY-ESO-1 antibodies elicited by vaccination with mRNA was examined. Therefore C57BL/6 mice were vaccinated

intradermally with different mRNAs complexed with protamine. The level of NY-ESO-1-specific antibodies in vaccinated and control mice was analyzed by ELISA with serial dilutions of sera. Anti NY-ESO-1 IgG2a[b] is induced by mRNA having only a poly(A) sequence. Strikingly however, the combination of poly(A) and histoneSL strongly increases the anti NY-ESO-1 IgG2a[b] level, manifold above the level observed with only a poly(A) sequence. Data are graphed as mean endpoint titers. Mean endpoint titers are summarized in Example 14.7.

Examples:

The following Examples are intended to illustrate the invention further and shall not be construed to limit the present invention thereto.

1. Generation of histone-stem-loop consensus sequences

Prior to the experiments, histone stem-loop consensus sequences were determined on the basis of metazoan and protozoan histone stem-loop sequences. Sequences were taken from the supplement provided by Lopez *et al.* (Dávila López, M., & Samuelsson, T. (2008), RNA (New York, N.Y.), 14(1), 1-10. doi:10.1261/rna.782308), who identified a large number of natural histone stem-loop sequences by searching genomic sequences and expressed sequence tags. First, all sequences from metazoa and protozoa (4001 sequences), or all sequences from protozoa (131 sequences) or alternatively from metazoa (3870 sequences), or from vertebrates (1333 sequences) or from humans (84 sequences) were grouped and aligned. Then, the quantity of the occurring nucleotides was determined for every position. Based on the tables thus obtained, consensus sequences for the 5 different groups of sequences were generated representing all nucleotides present in the sequences analyzed. In addition, more restrictive consensus sequences were also obtained, increasingly emphasizing conserved nucleotides

2. Preparation of DNA-templates

A vector for *in vitro* transcription was constructed containing a T7 promoter followed by a GC-enriched sequence coding for *Photinus pyralis* luciferase (ppLuc(GC)), the center part of the 3' untranslated region (UTR) of alpha-globin (ag), and a poly(A) sequence. The poly(A) sequence was immediately followed by a restriction site used for linearization of the vector before *in vitro* transcription in order to obtain mRNA ending in an A64 poly(A) sequence. mRNA obtained from this vector accordingly by *in vitro* transcription is designated as „ppLuc(GC) – ag – A64".

Linearization of this vector at alternative restriction sites before *in vitro* transcription allowed to obtain mRNA either extended by additional nucleotides 3' of A64 or

lacking A64. In addition, the original vector was modified to include alternative sequences. In summary, the following mRNAs were obtained from these vectors by *in vitro* transcription (mRNA sequences are given in Figures 6 to 17):

ppLuc(GC) – ag	(SEQ ID NO: 43)
ppLuc(GC) – ag – A64	(SEQ ID NO: 44)
ppLuc(GC) – ag – histoneSL	(SEQ ID NO: 45)
ppLuc(GC) – ag – A64 – histoneSL	(SEQ ID NO: 46)
ppLuc(GC) – ag – A120	(SEQ ID NO: 47)
ppLuc(GC) – ag – A64 – ag	(SEQ ID NO: 48)
ppLuc(GC) – ag – A64 – aCPSL	(SEQ ID NO: 49)
ppLuc(GC) – ag – A64 – PolioCL	(SEQ ID NO: 50)
ppLuc(GC) – ag – A64 – G30	(SEQ ID NO: 51)
ppLuc(GC) – ag – A64 – U30	(SEQ ID NO: 52)
ppLuc(GC) – ag – A64 – SL	(SEQ ID NO: 53)
ppLuc(GC) – ag – A64 – N32	(SEQ ID NO: 54)

Furthermore DNA plasmid sequences coding for the tumour antigens NY-ESO-1, Survivin and MAGE-C1 were prepared accordingly as described above.

In summary, the following mRNAs were obtained from these vectors by *in vitro* transcription (mRNA sequences are given in Figures 18 to 21):

NY-ESO-1(GC) - ag – A62 – C30	(SEQ ID NO: 55)
NY-ESO-1(GC) - ag – A62 – C30 – histoneSL	(SEQ ID NO: 56)
Survivin(GC) – ag – A62 – C30 – histoneSL	(SEQ ID NO: 57)
MAGE-C1(GC) – ag – A64 – C30 – histoneSL	(SEQ ID NO: 58)

3. In vitro transcription

The DNA-template according to Example 2 was linearized and transcribed *in vitro* using T7-Polymerase. The DNA-template was then digested by DNase-treatment. All mRNA-transcripts contained a 5'-CAP structure obtained by adding an excess of N7-Methyl-Guanosine-5'-Triphosphate-5'-Guanosine to the transcription reaction. mRNA thus obtained was purified and resuspended in water.

4. Enzymatic adenylation of mRNA

Two mRNAs were enzymatically adenylated:

ppLuc(GC) – ag – A64 and ppLuc(GC) – ag – histoneSL.

To this end, RNA was incubated with *E. coli* Poly(A)-polymerase and ATP (Poly(A) Polymerase Tailing Kit, Epicentre, Madison, USA) following the manufacturer's instructions. mRNA with extended poly(A) sequence was purified and resuspended in water. The length of the poly(A) sequence was determined via agarose gel electrophoresis. Starting mRNAs were extended by approximately 250 adenylates, the mRNAs obtained are designated as
ppLuc(GC) – ag – A300 and ppLuc(GC) – ag – histoneSL – A250, respectively.

5. Luciferase expression by mRNA electroporation

HeLa cells were trypsinized and washed in opti-MEM. $1x10^5$ cells in 200 µl of opti-MEM each were electroporated with 0.5 µg of ppLuc-encoding mRNA. As a control, mRNA not coding for ppLuc was electroporated separately. Electroporated cells were seeded in 24-well plates in 1 ml of RPMI 1640 medium. 6, 24, or 48 hours after transfection, medium was aspirated and cells were lysed in 200 µl of lysis buffer (25 mM Tris, pH 7.5 (HCl), 2 mM EDTA, 10% glycerol, 1% Triton X-100, 2 mM DTT, 1 mM PMSF). Lysates were stored at -20°C until ppLuc activity was measured.

6. Luciferase expression by mRNA lipofection

HeLa cells were seeded in 96 well plates at a density of $2x10^4$ cells per well. The following day, cells were washed in opti-MEM and then transfected with 0.25 µg of Lipofectin-complexed ppLuc-encoding mRNA in 150 µl of opti-MEM. As a control, mRNA not coding for ppLuc was lipofected separately. In some wells, opti-MEM was aspirated and cells were lysed in 200 µl of lysis buffer 6 hours after the start of transfection. In the remaining wells, opti-MEM was exchanged for RPMI 1640 medium at that time. In these wells, medium was aspirated and cells were lysed in 200 µl of lysis buffer 24 or 48 hours after the start of transfection. Lysates were stored at -20°C until ppLuc activity was measured.

7. Luciferase measurement

ppLuc activity was measured as relative light units (RLU) in a BioTek SynergyHT plate reader at 5 seconds measuring time using 50 µl of lysate and 200 µl of luciferin

buffer (25 mM Glycylglycin, pH 7.8 (NaOH), 15 mM $MgSO_4$, 2 mM ATP, 75 µM luciferin). Specific RLU were calculated by subtracting RLU of the control RNA from total RLU.

8. Luciferase expression by intradermal mRNA injection (Luciferase expression *in vivo*)

Mice were anaesthetized with a mixture of Rompun and Ketavet. Each ppLuc-encoding mRNA was injected intradermally (0.5 µg of mRNA in 50 µl per injection). As a control, mRNA not coding for ppLuc was injected separately. 16 hours after injection, mice were sacrificed and tissue collected. Tissue samples were flash frozen in liquid nitrogen and lysed in a tissue lyser (Qiagen) in 800 µl of lysis buffer (25 mM Tris, pH 7.5 (HCl), 2 mM EDTA, 10% glycerol, 1% Triton X-100, 2 mM DTT, 1 mM PMSF). Subsequently samples were centrifuged at 13500 rpm at 4°C for 10 minutes. Lysates were stored at -80°C until ppLuc activity was measured (see 7. luciferase measurement).

9. NY-ESO-1 expression by mRNA electroporation

HeLa cells were trypsinized and washed in opti-MEM. $2x10^5$ cells in 200 µl of opti-MEM were electroporated with 10 µg of NY-ESO-1-encoding mRNA. Cells from three electroporations were combined and seeded in a 6-well plate in 2 ml of RPMI 1640 medium. 24 hours after transfection, cells were harvested and transferred into a 96 well V-bottom plate (2 wells per mRNA). Cells were washed with phosphate buffered saline (PBS) and permeabilized with 200 µl per well of Cytofix/Cytoperm (Becton Dickinson (BD)). After 15 minutes, cells were washed with PermWash (BD). Then, cells were incubated for 1 hour at room temperature with either mouse anti-NY-ESO-1 IgG1 or an isotype control (20 µg/ml). Cells were washed twice with PermWash again. Next, cells were incubated for 1 hour at 4°C with a 1:500 dilution of Alexa-647 coupled goat-anti-mouse IgG. Finally, cells were washed twice with PermWash. Cells were resuspended in 200 µl of buffer (PBS, 2% FCS, 2 mM EDTA, 0.01% sodium azide). NY-ESO-1 expression was quantified by flow cytometry as median fluorescence intensity (MFI).

10. Induction of anti NY-ESO-1 antibodies by vaccination with mRNA

C57BL/6 mice were vaccinated intradermally with NY-ESO-1-encoding mRNA complexed with protamine (5 times in 14 days). Control mice were treated with buffer. The level of NY-ESO-1-specific antibodies in vaccinated and control mice was analyzed 8 days after the last vaccination by ELISA: 96 well ELISA plates (Nunc) were coated with 100 µl per well of 10 µg/ml recombinant NY-ESO-1 protein for 16 hours at 4°C. Plates were washed two times with wash buffer (PBS, 0.05% Tween-20). To block unspecific binding, plates were then incubated for 2 hours at 37°C with blocking buffer (PBS, 0.05% Tween-20, 1% BSA). After blocking, 100 µl per well of serially diluted mouse sera were added and incubated for 4 hours at room temperature. Plates were then washed three times with wash buffer. Next, 100 µl per well of biotinylated rat anti-mouse IgG2a[b] detection antibody (BD Biosciences) diluted 1:600 in blocking buffer was allowed to bind for 1 hour at room temperature. Plates were washed again three times with wash buffer, followed by incubation for 30 minutes at room temperature with 100 µl per well of horseradish peroxidase-coupled streptavidin. After four washes with wash buffer, 100 µl per well of 3,3',5,5'-tetramethylbenzidine (Thermo Scientific) was added. Upon the resulting change in color 100 µl per well of 20% sulfuric acid was added. Absorbance was measured at 405 nm.

11. Induction of anti Survivin antibodies by vaccination with mRNA

C57BL/6 mice were vaccinated intradermally with Survivin-encoding mRNA complexed with protamine (5 times in 14 days). Control mice were treated with buffer. The level of Survivin-specific antibodies in vaccinated and control mice was analyzed 8 days after the last vaccination by ELISA: 96 well ELISA plates (Nunc) were coated with 100 µl per well of 10 µg/ml recombinant Survivin protein for 16 hours at 4°C. Plates were washed two times with wash buffer (PBS, 0.05% Tween-20). To block unspecific binding, plates were then incubated for 2 hours at 37°C with blocking buffer (PBS, 0.05% Tween-20, 1% BSA). After blocking, 100 µl per well of serially diluted mouse sera were added and incubated for 4 hours at room temperature. Plates were then washed three times with wash buffer. Next, 100 µl per well of biotinylated rat anti-mouse IgG2a[b] detection antibody (BD Biosciences)

diluted 1:600 in blocking buffer was allowed to bind for 1 hour at room temperature. Plates were washed again three times with wash buffer, followed by incubation for 30 minutes at room temperature with 100 µl per well of horseradish peroxidase-coupled streptavidin. After four washes with wash buffer, 100 µl per well of 3,3',5,5'-tetramethylbenzidine (Thermo Scientific) was added. Upon the resulting change in color 100 µl per well of 20% sulfuric acid was added. Absorbance was measured at 405 nm.

12. Induction of anti MAGE-C1 antibodies by vaccination with mRNA

C57BL/6 mice were vaccinated intradermally with MAGE-C1-encoding mRNA complexed with protamine (5 times in 14 days). Control mice were treated with buffer. The level of MAGE-C1-specific antibodies in vaccinated and control mice was analyzed 8 days after the last vaccination by ELISA: 96 well ELISA plates (Nunc) were coated with 100 µl per well of 10 µg/ml recombinant MAGE-C1 protein for 16 hours at 4°C. Plates were washed two times with wash buffer (PBS, 0.05% Tween-20). To block unspecific binding, plates were then incubated for 2 hours at 37°C with blocking buffer (PBS, 0.05% Tween-20, 1% BSA). After blocking, 100 µl per well of serially diluted mouse sera were added and incubated for 4 hours at room temperature. Plates were then washed three times with wash buffer. Next, 100 µl per well of biotinylated rat anti-mouse IgG2a[b] detection antibody (BD Biosciences) diluted 1:600 in blocking buffer was allowed to bind for 1 hour at room temperature. Plates were washed again three times with wash buffer, followed by incubation for 30 minutes at room temperature with 100 µl per well of horseradish peroxidase-coupled streptavidin. After four washes with wash buffer, 100 µl per well of 3,3',5,5'-tetramethylbenzidine (Thermo Scientific) was added. Upon the resulting change in color 100 µl per well of 20% sulfuric acid was added. Absorbance was measured at 405 nm.

13. <u>Detection of an antigen-specific cellular immune response (T cell immune response) by ELISPOT:</u>

C57BL/6 mice are vaccinated intradermally with MAGE-C1 encoding mRNA complexed with protamine (5 times in 14 days). Control mice are treated with

buffer. 1 week after the last vaccination mice are sacrificed, the spleens are removed and the splenocytes are isolated. The splenocytes are restimulated for 7 days in the presence of peptides from the above antigen (peptide library) or coincubated with dendritic cells generated from bone marrow cells of native syngeneic mice, which are electroporated with mRNA coding for the antigen. To determine an antigen-specific cellular immune response INFgamma secretion was measured after re-stimulation. For detection of INFgamma a coat multiscreen plate (Millipore) is incubated overnight with coating buffer 0.1 M carbonate-bicarbonate buffer pH 9.6, 10.59 g/l Na2CO3, 8.4g/l NaHCO3) comprising antibody against INFγ (BD Pharmingen, Heidelberg, Germany). Stimulators and effector cells are incubated together in the plate in the ratio of 1:20 for 24h. The plate is washed with 1×PBS and incubated with a biotin-coupled secondary antibody. After washing with 1×PBS/0.05% Tween-20 the substrate (5-Bromo-4-Cloro-3-Indolyl Phosphate/Nitro Blue Tetrazolium Liquid Substrate System from Sigma Aldrich, Taufkirchen, Germany) is added to the plate and the conversion of the substrate could be detected visually.

14. RESULTS

14.1 Histone stem-loop sequences:

In order to characterize histone stem-loop sequences, sequences from metazoa and protozoa (4001 sequences), or from protozoa (131 sequences) or alternatively from metazoa (3870 sequences), or from vertebrates (1333 sequences) or from humans (84 sequences) were grouped and aligned. Then, the quantity of the occurring nucleotides was determined for every position. Based on the tables thus obtained, consensus sequences for the 5 different groups of sequences were generated representing all nucleotides present in the sequences analyzed. Within the consensus sequence of metazoa and protozoa combined, 3 nucleotides are conserved, a T/U in the loop and a G and a C in the stem, forming a base pair. Structurally, typically a 6 base-pair stem and a loop of 4 nucleotides is formed. However, deviating structures are common: Of 84 human histone stem-loops, two contain a stem of only 5 nucleotides comprising 4 base-pairs and one mismatch. Another human histone stem-loop contains a stem of only 5 base-pairs. Four more

human histone stem-loops contain a 6 nucleotide long stem, but include one mismatch at three different positions, respectively. Furthermore, four human histone stem-loops contain one wobble base-pair at two different positions, respectively. Concerning the loop, a length of 4 nucleotides seems not to be strictly required, as a loop of 5 nucleotides has been identified in *D. discoideum*.

In addition to the consensus sequences representing all nucleotides present in the sequences analyzed, more restrictive consensus sequences were also obtained, increasingly emphasizing conserved nucleotides. In summary, the following sequences were obtained:

(Cons): represents all nucleotides present
(99%): represents at least 99% of all nucleotides present
(95%): represents at least 95% of all nucleotides present
(90%): represents at least 90% of all nucleotides present

The results of the analysis of histone stem-loop sequences are summarized in the following Tables 1 to 5 (see also Fig. 1 to 5):

Table 1: Metazoan and protozoan histone stem-loop consensus sequence: (based on an alignment of 4001 metazoan and protozoan histone stem-loop sequences) (see also Fig. 1)

						<	<	<	<	<	<	•	•	•	•	>	>	>	>	>	>					
# A	2224	1586	3075	2872	1284	184	0	13	12	9	1	47	59	0	675	3818	195	1596	523	0	14	3727	61	771	2012	2499
# T	172	188	47	205	19	6	0	569	1620	199	3947	3830	3704	4001	182	1	21	15	11	0	179	8	64	557	201	690
# C	1557	2211	875	918	2675	270	0	3394	2342	3783	51	119	227	0	3140	7	50	31	16	4001	3543	154	3870	2636	1744	674
# G	25	16	4	6	23	3541	4001	25	27	10	2	5	11	0	4	175	3735	2359	3451	0	265	112	4	37	43	138
Cons	N*	N*	N	N	N	N	G	N	N	N	N	N	N	T	N	N	N	N	N	C	N	N	N	N*	N*	N*
99%	H*	H*	H	H	V	V	G	Y	Y	Y	Y	H	H	T	H	R	V	V	R	C	B	V	H	H*	N*	N*
95%	M*	H*	M	H	M	S	G	Y	Y	Y	T	T	Y	T	M	A	R	R	R	C	S	M	C	H*	H*	H*
90%	M*	M*	M	M	M	S	G	Y	Y	C	T	T	T	T	M	A	G	R	R	C	S	A	C	H*	M*	H*

Table 2: Protozoan histone stem-loop consensus sequence: (based on an alignment of 131 protozoan histone stem-loop sequences) (see also Fig. 2)

						<	<	<	<	<	<	•	•	•	•	>	>	>	>	>	>					
# A	52	32	71	82	76	13	0	12	12	9	1	46	3	0	75	82	53	79	20	0	4	94	17	35	74	56
# T	20	32	37	21	8	3	0	21	85	58	86	70	65	131	28	1	17	13	10	0	15	7	31	32	20	28
# C	45	59	20	25	38	0	0	86	8	54	42	13	58	0	27	2	6	31	10	131	112	5	82	58	30	40
# G	14	8	3	3	9	115	131	12	26	10	2	2	5	0	1	46	55	8	91	0	0	25	1	6	7	7
Cons	N*	N*	N	N	N	D	G	N	N	N	N	N	N	T	N	N	N	N	N	C	H	N	N	N*	N*	N*
99%	N*	N*	N	N	N	D	G	N	N	N	B	N	N	T	H	V	N	N	N	C	H	N	H	N*	N*	N*
95%	N*	N*	H	H	N	R	G	N	N	N	Y	H	B	T	H	R	D	N	N	C	Y	D	H	H*	N*	N*
90%	N*	H*	H	H	V	R	G	N	D	B	Y	H	Y	T	H	R	D	H	N	C	Y	R	H	H*	H*	H*

Table 3: Metazoan histone stem-loop consensus sequence: (based on an alignment of 3870 (including 1333 vertebrate sequences) metazoan histone stem-loop sequences) (see also Fig. 3)

						<	<	<	<	<	<	•	•	•	•	>	>	>	>	>	>					
# A	2172	1554	3004	2790	1208	171	0	1	0	0	0	1	56	0	600	3736	142	1517	503	0	10	3633	44	736	1938	2443
# T	152	156	10	184	11	3	0	548	1535	141	3861	3760	3639	3870	154	0	4	2	1	0	164	1	33	525	181	662
# C	1512	2152	855	893	2637	270	0	3308	2334	3729	9	106	169	0	3113	5	44	0	6	3870	3431	149	3788	2578	1714	634
# G	11	8	1	3	14	3426	3870	13	1	0	0	3	6	0	3	129	3680	2351	3360	0	265	87	3	31	36	131
Cons	N*	N*	N	N	N	N	G	N	B	Y	Y	N	N	T	N	V	N	D	N	C	N	N	N	N*	N*	N*
99%	H*	H*	M	H	M	V	G	Y	Y	Y	T	Y	H	T	H	R	V	R	R	C	B	V	M	H*	H*	N*
95%	M*	M*	M	M	M	S	G	Y	Y	C	T	T	Y	T	M	A	G	R	R	C	S	M	C	H*	H*	H*
90%	M*	M*	M	M	M	S	G	Y	Y	C	T	T	T	T	M	A	G	R	R	C	S	A	C	H*	M*	H*

Table 4: Vertebrate histone stem-loop consensus sequence: (based on an alignment of 1333 vertebrate histone stem-loop sequences) (see also Fig. 4)

						<	<	<	<	<	<	•	•	•	•	>	>	>	>	>	>					
# A	661	146	1315	1323	920	8	0	1	0	0	0	1	4	0	441	1333	0	1199	21	0	1	1126	26	81	380	960
# T	63	121	2	2	6	2	0	39	1217	2	1331	1329	1207	1333	30	0	1	0	1	0	2	1	22	91	91	12
# C	601	1062	16	6	403	1	0	1293	116	1331	2	0	121	0	862	0	2	0	0	1333	1328	128	1284	1143	834	361
# G	8	4	0	2	4	1322	1333	0	0	0	0	3	1	0	0	0	1330	134	1311	0	2	78	1	18	28	0
Cons	N*	N*	H	N	N	N	G	H	Y	Y	Y	D	N	T	H	A	B	R	D	C	N	N	N	N*	N*	H*
99%	H*	H*	M	A	M	G	G	Y	Y	C	T	T	Y	T	H	A	G	R	R	C	C	V	H	N*	N*	M*
95%	H*	H*	A	A	M	G	G	C	Y	C	T	T	Y	T	M	A	G	R	G	C	C	V	C	H*	H*	M*
90%	M*	M*	A	A	M	G	G	C	T	C	T	T	T	T	M	A	G	R	G	C	C	M	C	Y*	M*	M*

Table 5: *Homo sapiens* histone stem-loop consensus sequence: (based on an alignment of 84 human histone stem-loop sequences) (see also Fig. 5)

						<	<	<	<	<	<	•	•	•	•	>	>	>	>	>	>					
# A	10	17	84	84	76	1	0	1	0	0	0	1	0	0	12	84	0	65	3	0	0	69	5	0	10	64
# T	8	6	0	0	2	2	0	1	67	0	84	80	81	84	5	0	0	0	0	0	0	0	4	25	24	3
# C	62	61	0	0	6	0	0	82	17	84	0	0	3	0	67	0	1	0	0	84	84	5	75	57	44	17
# G	4	0	0	0	0	81	84	0	0	0	0	3	0	0	0	0	83	19	81	0	0	10	0	2	6	0
Cons	N*	H*	A	A	H	D	G	H	Y	C	T	D	Y	T	H	A	S	R	R	C	C	V	H	B*	N*	H*
99%	N*	H*	A	A	H	D	G	H	Y	C	T	D	Y	T	H	A	S	R	R	C	C	V	H	B*	N*	H*
95%	H*	H*	A	A	M	G	G	C	Y	C	T	T	T	T	H	A	G	R	G	C	C	V	M	Y*	N*	M*
90%	H*	M*	A	A	A	G	G	C	Y	C	T	T	T	T	M	A	G	R	G	C	C	R	M	Y*	H*	M*

Wherein the used abbreviations were defined as followed:

abbreviation	Nucleotide bases	remark
G	G	Guanine
A	A	Adenine
T	T	Thymine
U	U	Uracile
C	C	Cytosine
R	G or A	Purine
Y	T/U or C	Pyrimidine
M	A or C	Amino
K	G or T/U	Keto
S	G or C	Strong (3H bonds)
W	A or T/U	Weak (2H bonds)
H	A or C or T/U	Not G
B	G or T/U or C	Not A
V	G or C or A	Not T/U
D	G or A or T/U	Not C
N *	G or C or T/U or A present or not	Any base Base may be present or not

14.2 **The combination of poly(A) and histoneSL increases protein expression from mRNA in a synergistic manner.**

To investigate the effect of the combination of poly(A) and histoneSL on protein expression from mRNA, mRNAs with different sequences 3′ of the alpha-globin 3′-UTR were synthesized: mRNAs either ended just 3′ of the 3′-UTR, thus lacking both poly(A) sequence and histoneSL, or contained either an A64 poly(A) sequence or a histoneSL instead, or both A64 poly(A) and histoneSL 3′ of the 3′-UTR. Luciferase-encoding mRNAs or control mRNA were electroporated into HeLa cells. Luciferase levels were measured at 6, 24, and 48 hours after transfection (see following Table 6 and Figure 22).

Table 6:

mRNA	RLU at 6 hours	RLU at 24 hours	RLU at 48 hours
ppLuc(GC)-ag-A64-histoneSL	466553	375169	70735
ppLuc(GC)-ag-histoneSL	50947	3022	84
ppLuc(GC)-ag-A64	10471	19529	4364
ppLuc(GC)-ag	997	217	42

Little luciferase was expressed from mRNA having neither poly(A) sequence nor histoneSL. Both a poly(A) sequence or the histoneSL increased the luciferase level to a similar extent. Either mRNA gave rise to a luciferase level much higher than did mRNA lacking both poly(A) and histoneSL. Strikingly however, the combination of poly(A) and histoneSL further strongly increased the luciferase level, manifold above the level observed with either of the individual elements. The magnitude of the rise in luciferase level due to combining poly(A) and histoneSL in the same mRNA demonstrates that they are acting synergistically.

The synergy between poly(A) and histoneSL was quantified by dividing the signal from poly(A)-histoneSL mRNA (+/+) by the sum of the signals from histoneSL mRNA (-/+) plus poly(A) mRNA (+/-) (see following Table 7).

Table 7:

	A64	histoneSL	RLU at 6 hours	RLU at 24 hours	RLU at 48 hours
	+	+	466553	375169	70735
	—	+	50947	3022	84
	+	—	10471	19529	4364
Synergy			7.6	16.6	15.9

The factor thus calculated specifies how much higher the luciferase level from mRNA combining poly(A) and histoneSL is than would be expected if the effects of poly(A) and histoneSL were purely additive. The luciferase level from mRNA combining poly(A) and histoneSL was up to 16.6 times higher than if their effects were purely additive. This result confirms that the combination of poly(A) and histoneSL effects a markedly synergistic increase in protein expression.

14.3 **The combination of poly(A) and histoneSL increases protein expression from mRNA irrespective of their order.**

The effect of the combination of poly(A) and histoneSL might depend on the length of the poly(A) sequence and the order of poly(A) and histoneSL. Thus, mRNAs with increasing poly(A) sequence length and mRNA with poly(A) and histoneSL in reversed order were synthesized: Two mRNAs contained 3′ of the 3′-UTR either an A120 or an A300 poly(A) sequence. One further mRNA contained 3′ of the 3′-UTR

first a histoneSL followed by an A250 poly(A) sequence. Luciferase-encoding mRNAs or control mRNA were lipofected into HeLa cells. Luciferase levels were measured at 6, 24, and 48 hours after the start of transfection (see following Table 8 and Figure 23).

Table 8:

mRNA	RLU at 6 hours	RLU at 24 hours	RLU at 48 hours
ppLuc(GC)-ag-histoneSL-A250	98472	734222	146479
ppLuc(GC)-ag-A64-histoneSL	123674	317343	89579
ppLuc(GC)-ag-histoneSL	7291	4565	916
ppLuc(GC)-ag-A300	4357	38560	11829
ppLuc(GC)-ag-A120	4371	45929	10142
ppLuc(GC)-ag-A64	1928	26781	537

Both an A64 poly(A) sequence or the histoneSL gave rise to comparable luciferase levels. In agreement with the previous experiment did the combination of A64 and histoneSL strongly increase the luciferase level, manifold above the level observed with either of the individual elements. The magnitude of the rise in luciferase level due to combining poly(A) and histoneSL in the same mRNA demonstrates that they are acting synergistically. The synergy between A64 and histoneSL was quantified as before based on the luciferase levels of A64-histoneSL, A64, and histoneSL mRNA (see following Table 9). The luciferase level from mRNA combining A64 and histoneSL was up to 61.7 times higher than if the effects of poly(A) and histoneSL were purely additive.

Table 9:

	A64	histoneSL	RLU at 6 hours	RLU at 24 hours	RLU at 48 hours
	+	+	123674	317343	89579
	—	+	7291	4565	916
	+	—	1928	26781	537
Synergy			13.4	10.1	61.7

In contrast, increasing the length of the poly(A) sequence from A64 to A120 or to A300 increased the luciferase level only moderately (see Table 8 and Figure 19).

mRNA with the longest poly(A) sequence, A300, was also compared to mRNA in which a poly(A) sequence of similar length was combined with the histoneSL, histoneSL-A250. In addition to having a long poly(A) sequence, the order of histoneSL and poly(A) is reversed in this mRNA relative to A64-histoneSL mRNA. The combination of A250 and histoneSL strongly increased the luciferase level, manifold above the level observed with either histoneSL or A300. Again, the synergy between A250 and histoneSL was quantified as before comparing RLU from histoneSL-A250 mRNA to RLU from A300 mRNA plus histoneSL mRNA (see following Table 10). The luciferase level from mRNA combining A250 and histoneSL was up to 17.0 times higher than if the effects of poly(A) and histoneSL were purely additive.

Table 10:

	histoneSL	A250/A300	RLU at 6 hours	RLU at 24 hours	RLU at 48 hours
	+	+	98472	734222	146479
	+	—	7291	4565	916
	—	+	4357	38560	11829
Synergy			8.5	17.0	11.5

In summary, a highly synergistic effect of the combination of histoneSL and poly(A) on protein expression from mRNA has been demonstrated for substantially different lengths of poly(A) and irrespective of the order of poly(A) and histoneSL.

14.4 **The rise in protein expression by the combination of poly(A) and histoneSL is specific**

To investigate whether the effect of the combination of poly(A) and histoneSL on protein expression from mRNA is specific, mRNAs with alternative sequences in combination with poly(A) were synthesized: These mRNAs contained 3′ of A64 one of seven distinct sequences, respectively. Luciferase-encoding mRNAs or control mRNA were electroporated into HeLa cells. Luciferase levels were measured at 6, 24, and 48 hours after transfection (see following Table 11 and Figure 24).

Table 11:

mRNA	RLU at 6 hours	RLU at 24 hours	RLU at 48 hours
ppLuc(GC)-ag-A64-N32	33501	38979	2641
ppLuc(GC)-ag-A64-SL	28176	20364	874
ppLuc(GC)-ag-A64-U30	41632	54676	3408
ppLuc(GC)-ag-A64-G30	46763	49210	3382
ppLuc(GC)-ag-A64-PolioCL	46428	26090	1655
ppLuc(GC)-ag-A64-aCPSL	34176	53090	3338
ppLuc(GC)-ag-A64-ag	18534	18194	989
ppLuc(GC)-ag-A64-histoneSL	282677	437543	69292
ppLuc(GC)-ag-histoneSL	27597	3171	0
ppLuc(GC)-ag-A64	14339	48414	9357

Both a poly(A) sequence or the histoneSL gave rise to comparable luciferase levels. Again, the combination of poly(A) and histoneSL strongly increased the luciferase level, manifold above the level observed with either of the individual elements, thus acting synergistically. In contrast, combining poly(A) with any of the alternative sequences was without effect on the luciferase level compared to mRNA containing only a poly(A) sequence. Thus, the combination of poly(A) and histoneSL increases protein expression from mRNA in a synergistic manner, and this effect is specific.

14.5 **The combination of poly(A) and histoneSL increases protein expression from mRNA in a synergistic manner in vivo.**

To investigate the effect of the combination of poly(A) and histoneSL on protein expression from mRNA in vivo, Luciferase-encoding mRNAs with different sequences 3′ of the alpha-globin 3′-UTR or control mRNA were injected intradermally into mice: mRNAs contained either an A64 poly(A) sequence or a histoneSL instead, or both A64 poly(A) and histoneSL 3′ of the 3′-UTR. Luciferase levels were measured at 16 hours after injection (see following Table 12 and Figure 25).

Table 12:

mRNA	RLU at 16 hours
ppLuc(GC)-ag-A64-histoneSL	38081
ppLuc(GC)-ag-histoneSL	137
ppLuc(GC)-ag-A64	4607

Luciferase was expressed from mRNA having either a histoneSL or a poly(A) sequence. Strikingly however, the combination of poly(A) and histoneSL further strongly increased the luciferase level, manifold above the level observed with either of the individual elements. The magnitude of the rise in luciferase level due to combining poly(A) and histoneSL in the same mRNA demonstrates that they are acting synergistically.

The synergy between poly(A) and histoneSL was quantified by dividing the signal from poly(A)-histoneSL mRNA (+/+) by the sum of the signals from histoneSL mRNA (-/+) plus poly(A) mRNA (+/-) (see following Table 13).

Table 13:

	A64	histoneSL	RLU at 16 hours
	+	+	38081
	—	+	137
	+	—	4607
Synergy			8.0

The factor thus calculated specifies how much higher the luciferase level from mRNA combining poly(A) and histoneSL is than would be expected if the effects of poly(A) and histoneSL were purely additive. The luciferase level from mRNA combining poly(A) and histoneSL was 8 times higher than if their effects were purely additive. This result confirms that the combination of poly(A) and histoneSL effects a markedly synergistic increase in protein expression in vivo.

14.6 The combination of poly(A) and histoneSL increases NY-ESO-1 protein expression from mRNA.

To investigate the effect of the combination of poly(A) and histoneSL on protein expression from mRNA, NY-ESO-1-encoding mRNAs with different sequences 3′ of the alpha-globin 3′-UTR were synthesized: mRNAs contained either an A64 poly(A) sequence or both A64 poly(A) and histoneSL 3′ of the 3′-UTR. NY-ESO-1-encoding mRNAs were electroporated into HeLa cells. NY-ESO-1 levels were measured at 24 hours after transfection by flow cytometry (see following Table 14 and Figure 26).

Table 14:

	MFI at 24 hours	
mRNA	anti-NY-ESO-1	isotype control
NY-ESO-1(GC)-ag-A64-histoneSL	15600	1831
NY-ESO-1(GC)-ag-A64	1294	849

NY-ESO-1 was expressed from mRNA having only a poly(A) sequence. Strikingly however, the combination of poly(A) and histoneSL strongly increased the NY-ESO-1 level, manifold above the level observed with only a poly(A) sequence.

14.7 The combination of poly(A) and histoneSL increases the level of antibodies elicited by vaccination with mRNA.

To investigate the effect of the combination of poly(A) and histoneSL on the induction of antibodies elicited by vaccination with mRNA, C57BL/6 mice were vaccinated intradermally with protamine-complexed, NY-ESO-1-encoding mRNAs with different sequences 3′ of the alpha-globin 3′-UTR. mRNAs contained either an A64 poly(A) sequence or both A64 poly(A) and histoneSL 3′ of the 3′-UTR. The level of NY-ESO-1-specific antibodies in vaccinated and control mice was analyzed by ELISA with serial dilutions of sera (see following Table 15 and Figure 27).

Table 15:

mRNA	mean IgG2a[b] endpoint titer
NY-ESO-1(GC)-ag-A64-histoneSL	763
NY-ESO-1(GC)-ag-A64	20

Anti NY-ESO-1 IgG2a[b] was induced by mRNA having only a poly(A) sequence. Strikingly however, the combination of poly(A) and histoneSL strongly increased the anti NY-ESO-1 IgG2a[b] level, manifold above the level observed with only a poly(A) sequence.

5

Claims

1. Nucleic acid sequence comprising or coding in 5' to 3'-direction for

i) - a coding region, encoding at least one peptide or protein;
- at least one histone stem-loop, and
- a poly(A) sequence;

or

ii) - a coding region, encoding at least one peptide or protein;
- a poly(A) sequence; and
- at least one histone stem-loop;

wherein the at least one histone stem-loop in i) or ii) is selected from following formulae (I) or (II):

formula (I) (stem-loop sequence without stem bordering elements):

$$\underbrace{[N_{0-2}GN_{3-5}]}_{\text{stem1}}\ \underbrace{[N_{0-4}(U/T)N_{0-4}]}_{\text{loop}}\ \underbrace{[N_{3-5}CN_{0-2}]}_{\text{stem2}}$$

formula (II) (stem-loop sequence with stem bordering elements):

$$\underbrace{N_{1-6}}_{\substack{\text{stem1}\\ \text{bordering element}}}\ \underbrace{[N_{0-2}GN_{3-5}]}_{\text{stem1}}\ \underbrace{[N_{0-4}(U/T)N_{0-4}]}_{\text{loop}}\ \underbrace{[N_{3-5}CN_{0-2}]}_{\text{stem2}}\ \underbrace{N_{1-6}}_{\substack{\text{stem2}\\ \text{bordering element}}}$$

wherein:

stem1 or stem2 bordering elements N_{1-6} is a consecutive sequence of 1 to 6, preferably of 2 to 6, more preferably of 2 to 5, even more preferably of 3 to 5, most preferably of 4 to 5 or 5 N, wherein each N is

independently from another selected from a nucleotide selected from A, U, T, G and C, or a nucleotide analogue thereof;

stem1 $[N_{0-2}GN_{3-5}]$ is reverse complementary or partially reverse complementary with element stem2, and is a consecutive sequence between of 5 to 7 nucleotides;

wherein N_{0-2} is a consecutive sequence of 0 to 2, preferably of 0 to 1, more preferably of 1 N, wherein each N is independently from another selected from a nucleotide selected from A, U, T, G and C or a nucleotide analogue thereof;

wherein N_{3-5} is a consecutive sequence of 3 to 5, preferably of 4 to 5, more preferably of 4 N, wherein each N is independently from another selected from a nucleotide selected from A, U, T, G and C or a nucleotide analogue thereof, and

wherein G is guanosine or an analogue thereof, and may be optionally replaced by a cytidine or an analogue thereof, provided that its complementary nucleotide cytidine in stem2 is replaced by guanosine;

loop sequence $[N_{0-4}(U/T)N_{0-4}]$ is located between elements stem1 and stem2, and is a consecutive sequence of 3 to 5 nucleotides, more preferably of 4 nucleotides;

wherein each N_{0-4} is independent from another a consecutive sequence of 0 to 4, preferably of 1 to 3, more preferably of 1 to 2 N, wherein each N is independently from another selected from a nucleotide selected from A, U, T, G and C or a nucleotide analogue thereof; and

wherein U/T represents uridine, or optionally thymidine;

stem2 $[N_{3-5}CN_{0-2}]$ is reverse complementary or partially reverse complementary with element stem1, and is a consecutive sequence between of 5 to 7 nucleotides;

wherein N_{3-5} is a consecutive sequence of 3 to 5, preferably of 4 to 5, more preferably of 4 N, wherein each N is independently from another selected from a nucleotide selected from A, U, T, G and C or a nucleotide analogue thereof;

wherein N_{0-2} is a consecutive sequence of 0 to 2, preferably of 0 to 1, more preferably of 1 N, wherein each N is independently from another selected from a nucleotide selected from A, U, T, G and C or a nucleotide analogue thereof; and

wherein C is cytidine or an analogue thereof, and may be optionally replaced by a guanosine or an analogue thereof provided that its complementary nucleotide guanosine in stem1 is replaced by cytidine;

wherein

stem1 and stem2 are capable of base pairing with each other

forming a reverse complementary sequence, wherein base pairing may occur between stem1 and stem2, or

forming a partially reverse complementary sequence, wherein an incomplete base pairing may occur between stem1 and stem2;

wherein the at least one histone stem-loop binds to stem-loop binding protein (SLBP); and

wherein said peptide or protein comprises a tumour antigen,

or

a fragment of said tumour antigen, wherein the fragment has a length of at least six amino acid residues, and

wherein the tumour antigen is selected from the list of:

5T4, 707-AP, 9D7, AFP, AlbZIP HPG1, alpha-5-beta-1-integrin, alpha-5-beta-6-integrin, alpha-actinin-4/m, alpha-methylacyl-coenzyme A racemase, ART-4, ARTC1/m, B7H4, BAGE-1, BCL-2, bcr/abl, beta-catenin/m, BING-4, BRCA1/m, BRCA2/m, CA 15-3/CA 27-29, CA 19-9, CA72-4, CA125, calreticulin, CAMEL, CASP-8/m, cathepsin B, cathepsin L, CD19, CD20, CD22, CD25, CDE30, CD33, CD4, CD52, CD55, CD56, CD80, CDC27/m, CDK4/m, CDKN2A/m, CEA, CLCA2, CML28, CML66, COA-1/m, coactosin-like protein, collage XXIII, COX-2, CT-9/BRD6, Cten, cyclin B1, cyclin D1, cyp-B, CYPB1, DAM-10, DAM-6, DEK-CAN, EFTUD2/m, EGFR, ELF2/m, EMMPRIN, EpCam, EphA2, EphA3, ErbB3, ETV6-AML1, EZH2, FGF-5, FN, Frau-1, G250, GAGE-1, GAGE-2, GAGE-3, GAGE-4, GAGE-5, GAGE-6, GAGE7b, GAGE-8, GDEP, GnT-V, gp100, GPC3, GPNMB/m, HAGE, HAST-2, hepsin, Her2/neu, HERV-K-MEL, HLA-A*0201-R17I, HLA-A11/m, HLA-A2/m, HNE, homeobox NKX3.1, HOM-TES-14/SCP-1, HOM-TES-85, HPV-E6, HPV-E7, HSP70-2M, HST-2, hTERT, iCE, IGF-1R, IL-13Ra2, IL-2R, IL-5, immature laminin receptor, kallikrein-2, kallikrein-4, Ki67, KIAA0205, KIAA0205/m, KK-LC-1, K-Ras/m, LAGE-A1, LDLR-FUT, MAGE-A1, MAGE-A2, MAGE-A3, MAGE-A4, MAGE-A6, MAGE-A9, MAGE-A10, MAGE-A12, MAGE-B1, MAGE-B2, MAGE-B3, MAGE-B4, MAGE-B5, MAGE-B6, MAGE-B10, MAGE-B16, MAGE-B17, MAGE-C1, MAGE-C2, MAGE-C3, MAGE-D1, MAGE-D2, MAGE-D4, MAGE-E1, MAGE-E2, MAGE-F1, MAGE-H1, MAGEL2, mammaglobin A, MART-1/melan-A, MART-2, MART-2/m, matrix protein 22, MC1R, M-CSF, ME1/m, mesothelin, MG50/PXDN, MMP11, MN/CA IX-antigen, MRP-3, MUC-1, MUC-2, MUM-1/m, MUM-2/m, MUM-3/m, myosin class I/m, NA88-A, N-acetylglucosaminyltransferase-V, Neo-PAP, Neo-PAP/m, NFYC/m, NGEP, NMP22, NPM/ALK, N-Ras/m, NSE, NY-ESO-B, OA1, OFA-iLRP, OGT, OGT/m, OS-9, OS-9/m, osteocalcin, osteopontin, p15, p190 minor bcr-abl, p53, p53/m, PAGE-4, PAI-1, PAI-2, PAP, PART-1, PATE, PDEF, Pim-1-Kinase, Pin-1, Pml/PARalpha, POTE, PRAME, PRDX5/m, prostein, proteinase-3, PSA, PSCA, PSGR, PSM, PSMA, PTPRK/m, RAGE-1, RBAF600/m, RHAMM/CD168, RU1, RU2, S-100, SAGE, SART-1, SART-2, SART-3, SCC, SIRT2/m, Sp17, SSX-1, SSX-2/HOM-MEL-40, SSX-4, STAMP-1, STEAP-1, survivin, survivin-2B, SYT-SSX-1, SYT-SSX-2, TA-90, TAG-72, TARP, TEL-AML1, TGFbeta, TGFbetaRII, TGM-4, TPI/m, TRAG-3, TRG, TRP-1, TRP-2/6b, TRP/INT2, TRP-p8, tyrosinase, UPA, VEGFR1, VEGFR-2/FLK-1, WT1 and a immunoglobulin idiotype of a lymphoid blood cell or a T cell receptor idiotype of a lymphoid blood cell.

2. The nucleic acid sequence according to claim 1, wherein the histone stem loop is a metazoan histone stem loop sequence.

3. The nucleic acid sequence according to claim 1 or 2, wherein the tumour antigen is selected from survivin or an antigen from the MAGE-family.

4. The nucleic acid sequence according to any of claims 1 to 3, wherein the at least one histone stem loop is heterologous to the coding region encoding the at least one peptide or protein, preferably, wherein the coding region does not encode a histone protein or fragment thereof having histone or histone-like function, wherein the fragment has a length of at least six amino acid residues.

5. The nucleic acid sequence of any of claims 1 to 4, wherein its coding region does not encode a reporter protein or a marker or selection protein, wherein the reporter protein, marker or selection protein is not a tumour antigen.

6. The nucleic acid sequence according to any of claims 1 to 5, wherein the nucleic acid does not contain one or two or at least one or all but one or all of the components of the group consisting of: a sequence encoding a ribozyme (preferably a self-splicing ribozyme), a histone stem-loop processing signal, in particular a histone-stem loop processing sequence derived from mouse histone H2A614 gene and a Neo gene.

7. The nucleic acid sequence according to any of claims 1 to 5, wherein the nucleic acid does not contain a mouse histone stem-loop sequence.

8. Nucleic acid sequence according to any of claims 1 to 7, wherein the nucleic acid is an RNA, preferably an mRNA.

9. The nucleic acid sequence according to any of claims 1 to 8, wherein the at least one histone stem-loop is selected from at least one of following formulae (Ia) or (IIa):

$$\underbrace{[N_{0-1}GN_{3-5}]}_{\text{stem1}}\ \underbrace{[N_{1-3}(U/T)N_{0-2}]}_{\text{loop}}\ \underbrace{[N_{3-5}CN_{0-1}]}_{\text{stem2}}$$

formula (Ia) (stem-loop sequence without stem bordering elements)

$$\underbrace{N_{2-5}}_{\text{stem1}}\ \underbrace{[N_{0-1}GN_{3-5}]}_{\text{stem1}}\ \underbrace{[N_{1-3}(U/T)N_{0-2}]}_{\text{loop}}\ \underbrace{[N_{3-5}CN_{0-1}]}_{\text{stem2}}\ \underbrace{N_{2-5}}_{\text{stem2}}$$

bordering element — bordering element

formula (IIa) (stem-loop sequence with stem bordering elements)

10. The nucleic acid sequence according to any of claims 1 to 9, wherein the poly(A) sequence comprises a sequence of about 25 to about 400 adenosine nucleotides, preferably a sequence of about 50 to about 400 adenosine nucleotides, more preferably a sequence of about 50 to about 300 adenosine nucleotides, even more preferably a sequence of about 50 to about 250 adenosine nucleotides, most preferably a sequence of about 60 to about 250 adenosine nucleotides.

11. Nucleic acid sequence according to any of claims 1 to 10, wherein the nucleic acid sequence is a modified nucleic acid, in particular a stabilized nucleic acid.

12. A composition comprising at least one type of nucleic acid sequences according to any of claims 1 to 11.

13. The composition according to claim 12, wherein the composition comprises at least two types of nucleic acid sequences wherein each type of nucleic acid sequence encodes for a different peptide or protein, preferably for a different tumour antigen.

14. The composition according to claim 12 or 13, wherein one type of the contained nucleic acid sequences encodes for PSA, PSMA, PSCA, STEAP-1, NY-ESO-1, 5T4, Survivin, MAGE-C1, or MAGE-C2.

15. The composition according to any of claims 12 to 14, wherein the nucleic acid sequence does not encode for NY-ESO1, provided that the composition contains only one type of nucleic acid sequence.

16. The composition according to any of claims 12 to 15 comprising at least:

a) a nucleic acid sequence of any of claims 1 to 11 wherein said encoded peptide or protein comprises the tumour antigen PSA, or a fragment thereof, wherein the fragment has a length of at least six amino acid residues; and

b) a nucleic acid sequence of any of claims 1 to 11 wherein said encoded peptide or protein comprises the tumour antigen PSMA, or a fragment thereof, wherein the fragment has a length of at least six amino acid residues; and

c) a nucleic acid sequence of any of claims 1 to 11 wherein said encoded peptide or protein comprises the tumour antigen PSCA, or a fragment thereof, wherein the fragment has a length of at least six amino acid residues; and

d) a nucleic acid sequence of any of claims 1 to 11 wherein said encoded peptide or protein comprises the tumour antigen STEAP-1, or a fragment thereof, wherein the fragment has a length of at least six amino acid residues.

17. The composition according to any of claims 12 to 15 comprising at least:

a) a nucleic acid sequence comprising or coding for

i. a coding region, encoding at least one peptide or protein which comprises the tumour antigen NY-ESO-1, or a fragment thereof, wherein the fragment has a length of at least six amino acid residues,

ii. at least one histone stem-loop, and

iii. a poly(A) sequence or a polyadenylation signal;

b) a nucleic acid sequence of any of claims 1 to 11 wherein said encoded peptide or protein comprises the tumour antigen 5T4, or a fragment thereof, wherein the fragment has a length of at least six amino acid residues; and

c) a nucleic acid sequence of any of claims 1 to 11 wherein said encoded peptide or protein comprises the tumour antigen Survivin, or a fragment thereof, wherein the fragment has a length of at least six amino acid residues.

18. The composition according to any of claims 12 to 15 further comprising at least:

a) a nucleic acid sequence of any of claims 1 to 11 wherein said encoded peptide or protein comprises the tumour antigen MAGE-C1, or a fragment thereof, wherein the fragment has a length of at least six amino acid residues; and

b) a nucleic acid sequence of any of claims 1 to 11 wherein said encoded peptide or protein comprises the tumour antigen MAGE-C2, or a fragment thereof, wherein the fragment has a length of at least six amino acid residues.

19. Pharmaceutical composition comprising a nucleic acid sequence as defined according to any of claims 1 to 11, or a composition as defined according to any of claims 12 to 18 and optionally a pharmaceutically acceptable carrier.

20. Use of a nucleic acid sequence as defined according to any of claims 1 to 11, or a composition as defined according to any of claims 12 to 18 for increasing the expression of said encoded peptide or protein in vitro.

21. An in vitro method for increasing the expression of an encoded peptide or protein comprising the steps:

a) providing the nucleic acid sequence as defined according to any of claims 1 to 12 or the composition as defined according to any of claims 12 to 18,

b) applying or administering the nucleic acid sequence or the composition to a cell-free expression system, a cell or a tissue.