



METHOD OF ISOLATING *STREPTOCOCCUS SOBRINUS*
AND SELECTIVE MEDIUM THEREFOR

5 Technical Field

The present invention relates to a method of selectively isolating *Streptococcus sobrinus* out of dental caries-causing bacteria (mutans streptococci) inhabiting in the oral cavity, and a medium which allows *Streptococcus sobrinus* alone to selectively grow.

10

Background Art

Heretofore, it has been said that the bacteria which cause dental caries in human is *Streptococcus mutans* out of streptococci (oral streptococci) inhabiting in the oral cavity. However, it is considered
15 nowadays that dental caries is caused by a group of bacteria generically called as mutans streptococci which is constituted by bacterial species distinguished from each other in the different serotypes *a* to *h* (Hamada, S. and H. D. Slade, 1980, Microbiological Review 44:331-384; Hirasawa, M. et al., 1980, Infect. Immun. 27:1003-1011; and Loesche, W. J., 1986,
20 Microbiological Review 50:353-380). Oral streptococci classified into mutans streptococci include *Streptococcus cricetus*, *Streptococcus rattus*, *Streptococcus mutans*, *Streptococcus sobrinus* and *Streptococcus downei*.

Further, the recognition is recently being accepted that *Streptococcus mutans* and *Streptococcus sobrinus* out of the bacteria
25 classified into mutans streptococci play particularly important roles in dental caries. Particularly, in Europe, the significance of *Streptococcus*



sobrinus attracts attention, and there are reports that caries risk in the case where both of *Streptococcus mutans* and *Streptococcus sobrinus* inhabit in the oral cavity becomes higher than that in the case of *Streptococcus mutans* alone (Babaahmady, K. G. et al., 1998, Caries Res. 32:51-58; Kohler, B. and S. Bjarnason, 1987, Community Dent. Oral Epidemiol. 15:332-335; and Lindquist, S. and C. G. Emilson, 1991, Caries Res. 25:146-152). However, the distribution of *Streptococcus mutans* and *Streptococcus sobrinus* in dental caries, and the relative significance thereof to dental caries have not been clarified.

In Japan, it has been said that mainly found out the clinical samples of dental caries (carious cavity, dental plaque and saliva) is *Streptococcus mutans*. However, it is now discussed by the researchers how many *Streptococcus sobrinus* inhabit the oral cavity, and, whether or not the presence of *Streptococcus sobrinus* is really important for caries risk.

Mitis Salivarius Agar (MS, manufactured by Difco) is conventionally used as a selective medium for streptococci in the oral cavity. A medium prepared by the addition of sucrose and an antibiotic bacitracin to the Mitis Salivarius Agar is used as a selective medium for mutans streptococci (MSB medium). However, it is known that in the MSB medium, the recovery of *Streptococcus sobrinus* is lower than that of *Streptococcus mutans*. Further, *Streptococcus mutans* and *Streptococcus sobrinus* are closely resemble to each other in their colony morphology and so on, therefore, unless a researcher is conversant with the colony morphology of these species, under the circumstances, he or she cannot distinguish these species inhabiting on the same plate medium from each

other by observation with the naked eye.

Some methods for detection and identification of *Streptococcus mutans* and *Streptococcus sobrinus* individually were reported. In these methods, these two kinds of bacteria are separated out of the oral streptococci including these two kinds of bacteria by means of a selective medium capable of substantially selecting these two kinds of bacteria alone, followed by biochemical or serological tests to distinguish these two species from each other (Gold, O. G. et al., 1973, Archs. Oral Biol. 18:1357-1364; Hirasawa, M. et al., 1980, Infect. Immun. 27:697-699; Kimmel, L. and N. Tinanoff, 1991, Oral Microbiol. Immunol. 6:275-279; Linke, H. A. B., 1977, J. Clin. Microbiol. 5:604-609; Schaeken, M. J. M. et al., 1986, J. Dent. Res. 65:906-908; Takada, K. et al., 1984, Infect. Immun. 45:464-469; and Wada, W. G. et al., 1986, J. Med. Microbiol. 22:319-323), or by a DNA probe method to distinguish them (Igarashi, T. et al., 1996, Oral Microbiol. Immunol. 5:294-298; and Shiroza, T. et al., 1998, Oral Microbiol. Immunol. 13:11-16). Further, some selective media for *Streptococcus mutans* have been reported (Gold, O. G. et al., 1973, Archs. Oral Biol. 18:1357-1364; Kimmel, L. and N. Tinanoff, 1991, Oral Microbiol. Immunol. 6:275-279; Linke, H. A. B., 1977, J. Clin. Microbiol. 5:604-609; Schaeken, M. J. M. et al., 1986, J. Dent. Res. 65:906-908; and Wada, W. G. et al., 1986, J. Med. Microbiol. 22:319-323). However, up to the present date, no selective culture technique capable of isolating *Streptococcus sobrinus* alone has been reported. Furthermore, the difference in the susceptibility to antibiotics between the different serotypes of mutans streptococci has been reported (Little, W. A. et al., 1979, Antimicrob. Agents Chemother. 15:440-443). However, no antibiotic which has the possibility to select

Streptococcus sobrinus alone has not yet found.

Disclosure of the Invention

The object of the present invention is to develop a method of
5 selectively isolating *Streptococcus sobrinus* alone out of the population of
oral streptococci including mutans streptococci, and a selective medium for
selecting *Streptococcus sobrinus*.

In order to attain the above-mentioned object, the present inventors
focused the susceptibilities of mutans streptococci to cell wall inhibitors,
10 and tested the susceptibilities thereof to various types of antibiotics. As
the results, the difference between *Streptococcus mutans* and *Streptococcus
sobrinus* in their susceptibility to monobactam antibiotics was found, and
the present invention was accomplished by the finding.

Namely, the present invention is to provide a method of
15 substantially isolating *Streptococcus sobrinus* alone by the addition of a
monobactam antibiotic to a medium on which oral streptococci including
mutans streptococci can grow, and a selective medium for *Streptococcus
sobrinus* prepared by the addition of a monobactam antibiotic to a medium
on which oral streptococci including mutans streptococci can grow.

20 In the present invention, it is preferred that the medium on which
only the oral streptococci can grow is Mitis Salivarius Agar (MS medium;
manufactured by Difco).

In the present invention, as the monobactam antibiotic, aztreonam
(AZT) or carumonam (CRMN) is preferred, and aztreonam (AZT) is
25 particularly preferred. Here, the monobactam antibiotic is preferably
added in a concentration of from 0.05 to 0.5 g/L.

The selective medium for *Streptococcus sobrinus* of the present invention preferably contains fosfomycin and/or bacitracin.

Further, it is particularly preferred that the selective medium for *Streptococcus sobrinus* of the present invention has the following
5 composition:

	Mitis Salivarius Medium	90 g/L
	Aztreonam	0.05 - 0.5 g/L
	Fosfomycin	0.05 - 0.5 g/L
	Bacitracin	5 - 50 units/L
10	Sodium Chloride	5 - 40 g/L

Best Mode for Carrying Out the Invention

The method and selective medium of the present invention are characterized in that a monobactam antibiotic is added to a medium on
15 which oral streptococci including mutans streptococci can grow.

In the background of the present invention, there is the fact that distinction of *Streptococcus mutans* and *Streptococcus sobrinus* on the medium on which the two kinds of bacteria inhabit together requires a lot of skill, since the two kinds of bacteria which attract the researchers
20 attention as the major dental caries-causing bacteria among oral streptococci classified into the mutans streptococci, are extremely resemble to each other in their colony morphology and so on as is mentioned above. Here, mutans streptococci other than *Streptococcus mutans* and *Streptococcus sobrinus* have respective identifiable features in their colony
25 morphologies and so on without requiring the particular skill, therefore, even if these bacteria inhabit together with *Streptococcus mutans* or

Streptococcus sobrinus on the same medium, these two kinds of bacteria can easily be differentiated from the other bacteria.

In view of the above-mentioned background, in the present invention, the present inventors narrowed the focus to clearly differentiate
5 *Streptococcus mutans* and *Streptococcus sobrinus*, searched for antibiotics to which significant difference between the two kinds of bacteria in their susceptibilities was observed, and prepared a selective medium for *Streptococcus sobrinus*, on which *Streptococcus sobrinus* alone could substantially grow.

10 Now, test conditions such as materials and methods used in the tests carried out in the research to accomplish the present invention will be described.

<Materials and Methods>

Bacterial Strains

15 All of the bacterial strains used in the research are shown in Table 1 described below.

Table 1

Strain	Serotype
<Mutans streptococci>	
<i>Streptococcus cricetus</i>	
OMZ-61	<i>a</i>
E49	<i>a</i>
<i>Streptococcus rattus</i>	
BHT	<i>b</i>
FA1	<i>b</i>
<i>Streptococcus mutans</i>	
PS14	<i>c</i>
Ingbritt	<i>c</i>
JC2	<i>c</i>
NCTC10449	<i>c</i>
LM7	<i>e</i>
NUM-Sme1	<i>e</i>
OMZ175	<i>f</i>
NUM-Smf1	<i>f</i>
<i>Streptococcus sobrinus</i>	
OMZ176	<i>d</i>
B13	<i>d</i>
NUM-Ssd1	<i>d</i>
NUM-Ssd2	<i>d</i>
NIDR6715	<i>g</i>
OMZ65	<i>g</i>
NUM-Ssg1	<i>g</i>
<i>Streptococcus downei</i>	
NCTC11391	<i>h</i>
<Oral Streptococci>	
<i>Streptococcus salivarius</i>	
ATCC9222	
HHT	
<i>Streptococcus mitis</i>	
NCTC3168	
ATCC903	
<i>Streptococcus sanguis</i>	
ATCC10556	
<i>Streptococcus oralis</i>	
ATCC10557	
<i>Streptococcus gordonii</i>	
ATCC10558	
Challis	
<i>Streptococcus anginosus</i>	
ATCC9895	
NCTC10709	

Out of the above-described bacterial strains, the strains indicated by NUM-Sme, NUM-Smf, NUM-Ssd and NUM-Ssg were strains isolated from clinical samples, and identified by the combination of a colony morphological method and a serological method (Hamada, S. and H. D. Slade, 1980, Microbiological Review 44:331-384; Hirasawa, M. et al., 1980, Infect. Immun. 27:697-699; and Takada, K. et al., 1984, Infect. Immun. 45:464-469).

Out of the other bacterial strains, the strains to which "ATCC" is attached were purchased from American Type Culture Collection, the strains to which "NCTC" is attached were purchased from National Collection Type Culture, and the other strains are those which have been preserved before in the laboratory to which the present inventors belong.

All the bacterial strains were grown and maintained anaerobically at 37°C on Brain Heart Infusion (BHI broth, manufactured by Difco Laboratories, Detroit, MI) agar plates supplemented with 5% (v/v) horse blood.

Susceptibility Test to Antibiotics

The susceptibility test was carried out in accordance with microbroth dilution method.

In order to develop a selective medium for *Streptococcus sobrinus*, the preliminary study for selection of antibiotics was carried out by a disk susceptibility test (Sensi-Disc; manufactured by Becton Dickinson Co., USA) (no data given herein), and on the basis of the result of the preliminary study, the antibiotics used in the main test were determined. The antibiotics used in the main test are described below.

Cephaloridine (manufactured by Sigma Chemical Co., MO. USA)

Cefazoline (manufactured by Wako Pure Chemical Industries Ltd., Tokyo, Japan)

Cloxacillin (manufactured by Sigma)

Imipenem (manufactured by Banyu Pharmaceutical Co., Ltd., Tokyo, Japan)

Aztreonam (AZT) (manufactured by ICN Biomedicals Inc., OH, USA)

Carumonam (CAM) (manufactured by Takeda Chemical Industries Ltd., Osaka, Japan)

10 Cephalexin (manufactured by Sigma)

Benzylpenicillin (Wako Pure Chemical Industries Ltd.)

Cefoxitin (manufactured by Sigma)

The above-described antibiotics were dissolved in an appropriate solution, respectively, and the respective antibiotic solutions were divided into serial two-fold dilutions in a 96-well microtiter plates containing BHI broth. The bacteria were precultured in BHI broth, respectively, and inoculated into the wells containing the antibiotic solution to make a final concentration of the bacteria to be 1×10^4 cells / well, respectively. The plates were anaerobically incubated at a temperature of 37°C for 48 hours. After mixing the broth in each well by the use of a pipette, the absorbance at 550 nm in each well was measured with a microplate reader (manufactured by Corona Electric Co., Ibaraki, Japan), to determine bacterial growth. The antibiotic concentration at which no bacterial growth was observed was defined as the minimal inhibitory concentration (MIC).

Aliquots of 10 μ l of the broth in the well, which no bacterial growth

was observed were inoculated onto Mitis Salivarius Agar (MS medium; manufactured by Difco) plates, respectively, and anaerobically incubated at a temperature of 37°C for 48 hours. The recovery of the inoculated bacteria was calculated to estimate which the mechanism of growth inhibition was bactericidal or bacteriostatic.

Results

Results of the susceptibility tests of *Streptococcus mutans* and *Streptococcus sobrinus* to β -lactam antibiotics are shown in Table 2 below.

Table 2
Susceptibilities of *Streptococcus mutans* and *Streptococcus sobrinus*
to β -lactam antibiotics

Antibiotic	MIC(μ g/mL)			
	<i>Streptococcus mutans</i>		<i>Streptococcus sobrinus</i>	
	JC2(c) ^a	10449(c)	OMZ176(d)	B-13(d)
Cephaloridine	0.025	0.025	0.025	0.025
Cefazoline	0.25	0.25	0.25	0.50
Cloxacillin	0.63	0.63	1.25	1.25
Imipenem	0.125	0.125	0.031	0.016
Aztreonam	125	125	2000	2000
Carumonam	125	125	2000	2000
Cephalexin	1.25	1.25	1.25	1.25
Benzylpenicillin	0.1	0.1	0.05	0.05
Cefoxitin	3.13	3.13	12.5	3.13

^a Serotype given in parentheses.

From the results shown in Table 2, as to cepharoridine, cefazoline and cephalixin, no significant difference between *Streptococcus mutans* and *Streptococcus sobrinus* in their susceptibilities were observed.

As to cloxacillin, the susceptibilities of *Streptococcus mutans* were twice as high.

As to imipenem, the susceptibilities of *Streptococcus sobrinus* were approximately from 4 to 8 times as high as those of *Streptococcus mutans*.

As to benzylpenicillin, contrary to cloxacillin, the susceptibilities of *Streptococcus sobrinus* were twice as high.

5 As to cefoxitin, although only OMZ176 strain out of *Streptococcus sobrinus* indicated the susceptibility thereto approximately 4 times as high as those of the other bacterial strains, no significant difference between *Streptococcus mutans* and *Streptococcus sobrinus* in their susceptibilities was observed.

10 As to aztreonam and carumonam classified into monobactam antibiotics, the MICs of the both antibiotics to *Streptococcus mutans* were 125 µg/mL, respectively, while the MICs of these antibiotics to *Streptococcus sobrinus* were 2000 µg/mL, respectively, it becoming clear that the susceptibilities of *Streptococcus mutans* to these antibiotics were
15 16-fold as high as those of *Streptococcus sobrinus*. These observations indicate that the monobactam antibiotics are useful agents for separation of *Streptococcus sobrinus* from *Streptococcus mutans*.

It has been reported that imipenem, aztreonam and carumonam targeted the penicillin-binding protein-3 (PBP-3) with respect to *E. coli*
20 (Georgopapadakou, N. H. et al., 1982, Antimicrob. Agents Chemother. 21:950-956; and Imada, A. et al., 1985, Antimicrob. Agents Chemother. 27:821-827), while with respect to *Streptococcus mutans* and *Streptococcus sobrinus*, imipenem and the monobactam antibiotics did not show the same mode of action.

25 Here, both aztreonam and carumonam had the mode of action of bactericidal to *Streptococcus mutans* and *Streptococcus sobrinus*.

Although in the above-described susceptibility tests to the antibiotics, only the two kinds of bacterial strains classified into the serotype *c* out of *Streptococcus mutans*, and only the two kinds of bacterial strains classified into the serotype *d* out of *Streptococcus sobrinus* were used, the susceptibilities of bacterial strains classified into the various serotypes to the monobactam antibiotics were studied. The results are shown in Table 3 below.

Table 3

Susceptibilities of bacterial strains classified into various serotypes of *Streptococcus mutans* and *Streptococcus sobrinus* to monobactam antibiotics

Strain	Serotype	MIC($\mu\text{g/mL}$)	
		Aztreonam	Carumonam
<i>Streptococcus mutans</i>			
PS14	<i>c</i>	125	125
Ingbritt	<i>c</i>	125	125
LM7	<i>e</i>	125	125
NUM-Sme1	<i>e</i>	125	125
OMZ175	<i>f</i>	250	250
NUM-Smf1	<i>f</i>	250	250
<i>Streptococcus sobrinus</i>			
NUM-Ssd1	<i>d</i>	2000	2000
NUM-Ssd2	<i>d</i>	2000	2000
NIDR6715	<i>g</i>	2000	2000
OMZ65	<i>g</i>	2000	1000

From the results shown in Table 3, the bacterial strains classified into the serotypes *c* and *e* of *Streptococcus mutans* had MICs of 125 $\mu\text{g/mL}$, respectively, in the same as the results shown in Table 2. The bacterial strains classified into the serotype *f* of *Streptococcus mutans* had MICs of 250 $\mu\text{g/mL}$, respectively, and had the susceptibilities to aztreonam and

carumonam lower than the bacterial strains classified into the serotypes *c* and *e*.

The bacterial strains classified into the serotypes *d* and *g* of *Streptococcus sobrinus* had MICs in the same as the results shown in Table 2, respectively, except that OMZ65 strain classified into the serotype *g* had an MIC to carumonam of 1000 µg/mL, it having the susceptibility twice as high as the other bacterial strains.

Also, both aztreonam and carumonam had the mode of action of bactericidal to the bacterial strains of *Streptococcus mutans* and *Streptococcus sobrinus* as shown in Table 3.

From the results shown in Tables 2 and 3, it became clear that the susceptibilities of *Streptococcus sobrinus* to the monobactam antibiotics were significantly lower than those of *Streptococcus mutans*. Therefore, it is understood that *Streptococcus sobrinus* can grow on a medium containing the monobactam antibiotics in such a concentration that *Streptococcus mutans* can not grow, and such a medium allow *Streptococcus sobrinus* alone to grow selectively. Namely, the present inventors have found that by the use of the monobactam antibiotics, *Streptococcus sobrinus* alone can selectively be isolated.

Examples

Now, the present invention will be specifically described with reference to examples. On the basis of the above-mentioned findings, the present inventors produced a trial model of a selective medium for *Streptococcus sobrinus* and evaluated it.

Preparation of Selective Medium

The selective medium for *Streptococcus sobrinus* having the

composition shown in Table 4 below (hereinafter this medium will be referred to as MS-SOB medium.) was prepared.

Table 4

5 Composition of Selective Medium for *Streptococcus sobrinus*
(MS-SOB medium)

Component	
Mitis Salivarius Medium	90 g/L
Aztreonam	0.2 g/L
Fosfomycin	0.02 g/L
Bacitracin	20 units/L
Sodium Chloride	20 g/L

Mitis Salivarius medium (MS medium) in Table 4, manufactured by Difco, which is generally used for isolation of not only mutans streptococci
10 but also the other streptococci, has the following composition.

	Bacto Tryptose	10 g/L
	Bacto Proteose Peptone No. 3	5 g/L
	Bacto Proteose Peptone	5 g/L
	Bacto Dextrose	1 g/L
15	Bacto Saccharose	50 g/L
	Dipotassium Phosphate	4 g/L
	Trypan Blue	0.075 g/L
	Bacto Crystal Violet	0.0008 g/L
	Bacto Agar	15 g/L

20 However, the base medium used for the selective medium for *Streptococcus sobrinus* of the present invention is not limited to the MS medium, and any media may be used on which only oral streptococci including mutans streptococci can grow.

Although aztreonam is used in the MS-SOB medium as the monobactam antibiotic, the monobactam antibiotic used in the present invention is not limited to aztreonam, and the antibiotics classified into the monobactam antibiotics, for instance, carumonam may be used. Further, the monobactam antibiotics available commercially in Japan at present are only two kinds of aztreonam and carumonam but the monobactam antibiotics used in the present invention are not limited to these two antibiotics. The concentration of the monobactam antibiotic are determined from the results shown in Table 2 and 3 within such a range that *Streptococcus sobrinus* can grow and the other oral streptococci including mutans streptococci can significantly be controlled. The concentration of the monobactam antibiotic is usually within a range of from 0.05 to 0.5 g/L, preferably from 0.1 to 0.3 g/L and particularly preferably 0.2 g/L.

Fosfomycin and bacitracin used here were purchased from Sigma. The mechanisms of action of fosfomycin and bacitracin are inhibitions of pyruvate N-acetylglucosamine-3-o-enol transferase and pyrophosphatase by binding to membrane phospholipids, respectively. Namely, the mechanisms of action thereof are to block from synthesizing cell wall peptide glycan. By the combined use of the monobactam antibiotics with fosfomycin and bacitracin which have the modes of action different from the monobactam antibiotics, it was expected that synergistic effect could be obtained. However, in the present invention, agents such as antibiotics to be added to the medium other than the monobactam antibiotic are not limited to fosfomycin and bacitracin. Fosfomycin is preferably used in a concentration of from 0.05 to 0.5 g/L, and bacitracin is preferably used in a

concentration of from 5 to 50 units/L.

Sodium chloride is added to the medium for the purpose of adjusting osmotic pressure and the like, and the concentration thereof in the medium may optionally be determined on the basis of the relationship
5 of the type, concentrations and so on of the other components, and is preferably within a range of from 5 to 40 g/L.

To the selective medium for *Streptococcus sobrinus* of the present invention, additional components capable of increasing the selectivity of *Streptococcus sobrinus* other than the components described in Table 4
10 above may be added.

The MS medium to which sodium chloride was added was sterilized and the medium was cooled to a temperature of 50 °C. Then, the above-mentioned three kinds of antibiotics were added to the medium to prepare the MS-SOB medium.

15 Performance Evaluation Test of Selectivity to *Streptococcus sobrinus*

By the use of the MS-SOB medium and the MS medium selected as a medium on which oral streptococci including mutans streptococci can grow, the recoveries of *Streptococcus sobrinus* were obtained, to evaluate the performance of selectivity of the MS-SOB medium to *Streptococcus*
20 *sobrinus*.

The bacterial strains indicated in Table 5 below were incubated anaerobically at 37 °C over night in advance. Ten-fold dilution of the culture solution was prepared by the addition of 0.9 mL of Tris-HCl buffer (0.05 M, pH 7.2) thereto, aliquots of 0.1 mL of the dilution were plated on
25 the test media (MS-SOB and MS), respectively. The plates were incubated anaerobically at a temperature of 37 °C for 3 days, and then, colony forming

units (CFU/mL) were counted. The results are shown in Table 5.

Table 5

Recoveries of Oral Streptococci and Mutans Streptococci
on MS Medium and MS-SOB Medium

Strain	Serotype	Recovery (CFU%)	
		MS medium	MS-SOB medium
<i>Streptococcus cricetus</i>			
OMZ-61	<i>a</i>	1.3×10^7	1.8×10^5 (13.5)
E49	<i>a</i>	1.0×10^7	0 (0)
<i>Streptococcus rattus</i>			
BHT	<i>b</i>	1.1×10^8	0 (0)
FA1	<i>b</i>	2.4×10^8	0 (0)
<i>Streptococcus mutans</i>			
JC2	<i>c</i>	2.9×10^8	0 (0)
NCTC10449	<i>c</i>	1.9×10^8	0 (0)
LM7	<i>e</i>	2.4×10^8	0 (0)
NUM-Sme1	<i>e</i>	3.0×10^8	0 (0)
OMZ175	<i>f</i>	1.4×10^8	0 (0)
NUM-Smf1	<i>f</i>	6.3×10^7	0 (0)
<i>Streptococcus sobrinus</i>			
OMZ176	<i>d</i>	1.0×10^7	7.0×10^6 (70.2)
B13	<i>d</i>	5.8×10^7	4.7×10^7 (81.3)
NUM-Ssd1	<i>d</i>	1.4×10^7	1.2×10^7 (84.8)
NIDR6715	<i>g</i>	1.5×10^7	1.1×10^7 (70.7)
OMZ65	<i>g</i>	2.5×10^7	1.5×10^7 (60.8)
NUM-Ssg1	<i>g</i>	1.8×10^8	1.4×10^8 (76.6)
<i>Streptococcus downei</i>			
NCTC11391	<i>h</i>	6.5×10^8	3.9×10^8 (60.2)
<i>Streptococcus salivarius</i>			
ATCC9222		2.0×10^8	0 (0)
HHT		2.3×10^8	0 (0)
<i>Streptococcus mitis</i>			
NCTC3168		9.6×10^7	0 (0)
ATCC903		1.2×10^8	0 (0)
<i>Streptococcus sanguis</i>			
ATCC10556		4.5×10^8	0 (0)
<i>Streptococcus oralis</i>			
ATCC10557		2.4×10^8	0 (0)
<i>Streptococcus gordonii</i>			
ATCC10558		9.4×10^7	0 (0)
Challis		1.1×10^8	0 (0)
<i>Streptococcus anginosus</i>			
ATCC9895		8.0×10^7	2.0×10^3 (0.0025)
NCTC10709		3.4×10^8	1.0×10^3 (0.0003)

As is apparent from Table 5 above, all the bacterial strains used for the test grew well on the MS medium at a density of from 10^7 to 10^8 CFU/mL. On the other hand, the bacterial strains able to grow on the MS-SOB medium were limited only to *Streptococcus cricetus* OMZ-61 strain (recovery: 13.5%) out of the serotype *a*, all of the bacterial strains classified into the serotypes *d* and *g* of *Streptococcus sobrinus* (recovery: 74.1 % on the average), *Streptococcus downei* NCTC11391 strain of the serotype *h* (recovery: 60.2 %), and *Streptococcus anginosus* ATCC9895 strain (recovery: 0.0025 %) and NCTC10709 strain (recovery: 0.0003 %).

Out of mutans streptococci able to grow on the MS-SOB medium, *Streptococcus cricetus* OMZ-61 strain of the serotype *a* is a rare bacterial strain detected in the oral cavity. Although *Streptococcus downei* NCTC11391 strain of the serotype *h* was found at a high recovery of 60.2 %, this bacterial strain has not been detected from human oral cavity up until now. Further, these bacterial strains can be distinguished from *Streptococcus sobrinus* in their colony morphological features. Therefore, it is vanishingly impossible to confuse these bacterial strains and *Streptococcus sobrinus*.

Streptococcus anginosus ATCC9895 strain and NCTC10709 strain belonging to the oral streptococci are extremely low in their recoveries, and they have features significantly different from *Streptococcus sobrinus* in their colony morphology. Therefore, even when these bacterial strains inhabit together with *Streptococcus sobrinus*, a person who is not conversant with their colony morphologies can certainly distinguished between *Streptococcus sobrinus* and these bacterial strains.

Out of the tested mutans streptococci, *Streptococcus mutans* which

are hardly distinguished from *Streptococcus sobrinus* based on their colony morphologies can not grow on the MS-SOB medium at all, therefore, there is no possibility to confuse *Streptococcus mutans* and *Streptococcus sobrinus*.

From the above-mentioned facts, it is understood that by the use of the MS-SOB medium, *Streptococcus sobrinus* can be isolated selectively.

Confirmation of Selectivity of MS-SOB Medium using Clinical Samples

The selectivity of the MS-SOB medium was confirmed with respect to clinical samples obtained from the dental caries area in human oral cavity.

Sampling area of dental caries was wiped with a cotton roll, and saliva was removed from the sampling area with air blast for a short period of time. Then, sample was taken out from the area of dental caries with a sterilized dental probe. Each sample was taken into a sterilized micro-centrifuging tube containing 1 mL of 50 mM Tris-HCl buffer (pH 7.2). After ultrasonic dispersion, by the use of the same buffer, ten-fold serial dilutions of each sample were prepared, and appropriate dilutions were plated triplicate on the MS plate medium and the MS-SOB plate medium. These plate media were incubated anaerobically at a temperature of 37°C for 3 days, and then, recoveries (CFU%) were calculated. The samples containing bacteria in the total number of 1×10^6 to 10^8 on the average were used for clinical analysis.

Five colonies per a test subject, which seemed to be *Streptococcus sobrinus* in the light of their colony morphology, were subcultured to confirm the presence of *Streptococcus sobrinus*.

The pure culture strain from each separated strain was identified

by 1) the colony morphology; 2) the fermentation pattern using their rapid ID 32 STREP (manufactured by bio Metrieux) or Streptogram (manufactured Wako Pure Chemical Industries Ltd.) ; and 3) agar gel immunodiffusion method using rabbit antiserum generated against the
5 control strain prepared in our laboratory as mentioned above (Hirasawa, M. et al., 1980, Infect. Immun. 27:697-699; and Igarashi, T. et al., 1996, Oral Microbiol. Immunol. 5:294-298).

The clinical samples were divided into two groups of Group I having the ratio of *Streptococcus sobrinus* in all the streptococci of less than 1 %, and Group II having the ratio of not less than 1 %, on the basis of the
10 number of existing *Streptococcus sobrinus*. All the samples contained the streptococci in the number of 10^6 to 10^7 on the average on the MS plate medium.

By the use of the above-mentioned clinical samples, the recoveries
15 of *Streptococcus sobrinus* were calculated. The results are shown in Table 6 below.

Table 6

Recoveries of *Streptococcus sobrinus* on MS medium and MS-SOB medium

from human dental caries samples

Group	Percentage of <i>Streptococcus sobrinus</i> in all bacteria	Number of samples	MS-SOB medium			MS medium ^a	
			<i>Streptococcus sobrinus</i>	Other bacteria	<i>Streptococcus sobrinus</i>	<i>Streptococcus sobrinus</i>	Other bacteria
I	Less than 1%	10	7.6×10^3 , ^b (4×10 to 6.9×10^4)	1.9×10 (0 to 7.0×10)	N.D. ^c	6.2×10^6 (5.0×10^4 to 3.0×10^7)	
II	Not less than 1%	8	5.4×10^6 , ^a (1.8×10^5 to 3.6×10^7)	0	5.6×10^6 (1.0×10^5 to 3.7×10^7)	2.3×10^7 (6.5×10^5 to 6.6×10^7)	

5 ^a Mean number of CFU per mL on the triplicate plates containing 0.1 mL of a 10^{-4} or 10^{-5} dilution range

^b Mean number of CFU per mL on the triplicate plates containing 0.1 mL of an undiluted sample or a 10^{-1} dilution

^c N.D.: Not detectable, plates too crowded.

 Number given in parenthesis indicates a range of detection.

As is apparent from the results of Table 6 above, in Group I, no *Streptococcus sobrinus* was detected on the crowded MS plate medium. On the other hand, on the MS-SOB medium, even less than 1% of *Streptococcus sobrinus* inhabited among the bacteria of the total number of from 10^6 to 10^7 could be detected. When the MS-SOB medium was used, the bacteria other than *Streptococcus sobrinus* were almost totally inhibited. The bacteria other than *Streptococcus sobrinus*, which grew on the MS-SOB medium, formed a small number of colonies or pinpoint colonies.

As for the samples of Group II, the pure strains of *Streptococcus sobrinus* were obtained on the MS-SOB medium, and the other bacteria (contaminants) did not grow on the MS-SOB medium. On the MS medium, *Streptococcus sobrinus* and the other bacteria inhabited together in almost the same quantity. It was difficult to find out *Streptococcus sobrinus* from the sample on the MS plate medium of Group II, which was contaminated by the other bacteria, as was expected.

From the results of Table 6 above, it is understood that by the use of the MS-SOB medium, *Streptococcus sobrinus* can selectively be isolated from clinical dental caries samples.

Study on Bacterial Growth on MS-SOB Medium with respect to Clinical Dental Caries Sample

Sixty human clinical dental caries samples were incubated on the MS-SOB media, and the results of incubation are shown in Table 7 below. Here, all of these 60 samples contained bacteria in the number of 1×10^7 on the average when measured on the MS plate media.

Table 7

Isolation of *Streptococcus sobrinus* on MS-SOB medium
from 60 human clinical dental caries samples

Growth ^a	Number of Samples
<i>Streptococcus sobrinus</i> pure culture	14
<i>Streptococcus sobrinus</i> plus other bacteria	4
Other bacteria only	
>500	0
10 to 500	10
<10	9
No bacterial growth	23

- 5 ^a Mean number of CFU per mL on the triplicate plates containing 0.1 mL of an undiluted sample.

In 14 samples out of the 60 samples, *Streptococcus sobrinus* alone grew, and in 23 samples, no growth of any bacteria was observed.

- 10 In 4 samples, growth of *Streptococcus sobrinus* was mainly observed with the small number of colonies (number of colonies: 70 at the maximum) of the other bacteria (contaminants). These other bacteria could morphologically be distinguished from *Streptococcus sobrinus*.

- 15 Although growth of the other bacteria alone was observed in 19 samples, these bacteria formed their colonies in the number of less than 5×10^2 CFU/mL at the maximum. These other bacteria were, of course, morphologically distinguishable from *Streptococcus sobrinus*.

Industrial Applicability

- 20 According to the present invention, a method of substantially isolating *Streptococcus sobrinus* alone selectively and a selective medium for *Streptococcus sobrinus*.

According to the present invention, there is no necessity to

morphologically distinguish between *Streptococcus mutans* and *Streptococcus sobrinus*, which are closely resemble to each other in their morphologies so that the distinction of these species requires a lot of skill. Therefore, no confusion of these bacterial species could be brought about.

5 Further, no confusion of *Streptococcus sobrinus* with oral streptococci including the mutans streptococci other than *Streptococcus sobrinus* could be brought about, too.

By the use of the selective medium of the present invention, *Streptococcus sobrinus* alone can selectively be isolated among oral
10 streptococci including mutans streptococci and can be incubated, thereby it being possible to clarify the exact aspect of the presence of *Streptococcus sobrinus* in clinical dental caries samples.

If the aspect of the presence of *Streptococcus sobrinus* can be exactly gotten hold of, it becomes possible to clarify the significance and
15 role of *Streptococcus sobrinus* in caries risk, which has been discussed in the art.

Furthermore, in the future, the distribution of *Streptococcus sobrinus* in oral cavity will be investigated and be able to be used for diagnosis of the caries risk of a patient.

CLAIMS

1. A method of substantially isolating *Streptococcus sobrinus* alone by the addition of a monobactam antibiotic to a medium on which only oral streptococci including mutans streptococci can grow.
2. A selective medium for *Streptococcus sobrinus* prepared by the addition of a monobactam antibiotic to a medium on which only oral streptococci including mutans streptococci can grow.
3. The selective medium for *Streptococcus sobrinus* according to Claim 2, wherein the medium on which only oral streptococci can grow is Mitis Salivarius Agar (MS medium, manufactured by Difco).
4. The selective medium for *Streptococcus sobrinus* according to Claim 2 or 3, wherein the monobactam antibiotic is aztreonam (AZT) or carumonam (CRMN).
5. The selective medium for *Streptococcus sobrinus* according to Claim 4, wherein the monobactam antibiotic is aztreonam (AZT).
6. The selective medium for *Streptococcus sobrinus* according to any one of Claims 2 to 5, wherein the selective medium contains the monobactam antibiotic in a concentration of from 0.05 to 0.5 g/L.
7. The selective medium for *Streptococcus sobrinus* according to any one of Claims 2 to 6, wherein the selective medium contains fosfomycin and/or bacitracin.
8. The selective medium for *Streptococcus sobrinus* according to any one of Claims 2 to 7, wherein the selective medium has the composition below:

	Mitis Salivarius Medium	90 g/L
	Aztreonam	0.05 - 0.5 g/L
	Fosfomycin	0.05 - 0.5 g/L
	Bacitracin	5 - 50 units/L
5	Sodium Chloride	5 - 40 g/L.