

DESCRIPTION

Title of Invention: METHOD FOR PRODUCING COMPLEX

Technical Field

[0001]

The present invention relates to a method for producing a conjugate.

Background Art

[0002]

For the purpose of use in reagents and diagnostics for detection of target molecules, or use in pharmaceuticals for treatment of diseases, studies have been conducted on the labeling rate of a conjugate in which a radioactive metal is coordinated to a ligand compound that is conjugated with an antibody.

[0003]

Patent Literature 1 discloses that radioactive zirconium and DTPA, which is a kind of ligand compound, are reacted in acidic physiological saline to form a Zr complex.

[0004]

Patent Literature 2 discloses that radioactive zirconium and DOTA, which is a kind of ligand compound, are heated and reacted in a neutral HEPES buffer solution to form a Zr complex.

[0005]

Patent Literature 3 discloses that radioactive actinium and rituximab conjugated with DOTA, which is a kind of ligand compound, are reacted in a basic Tris buffer solution to form an Ac complex.

[0006]

Patent Literature 4 discloses that radioactive actinium and HuM-195, which is a recombinant IgG1 antibody conjugated with DOTA as a kind of ligand compound, are reacted in an acetate buffer solution containing gentisic acid at pH 5.5 to 7.0 to form an Ac complex.

[0007]

Non Patent Literature 1 describes a method in which radioactive zirconium and DOTA as a ligand compound are reacted in a buffer solution to form a Zr complex.

Citation List

Patent Literature

[0008]

Patent Literature 1: US 2014/147381 A

Patent Literature 2: US 2019/038785 A

Patent Literature 3: US 2015/0157742 A

Patent Literature 4: US 2012/0220754 A

Non Patent Literature

[0009]

Non Patent Literature 1: Pandya et al, Chem Sci. 2017; 8 (3): 2309-14

Summary of Invention

[0010]

However, under the conditions disclosed in Patent Literature 1 and 2 and Non Patent Literature 1, complexation does not proceed well between a ligand compound

and a radioisotope, and a sufficient labeling rate cannot be achieved by the radioisotope for an antibody conjugated with the ligand compound (hereinafter, referred to as a ligand-conjugated antibody). Therefore, reaction conditions have been desired under which a high labeling rate can be stably achieved by a radioisotope for a ligand-conjugated antibody. Therefore, an object of the present invention is to provide a method for producing a conjugate that includes a radioisotope, a ligand compound, and an antibody and is capable of imparting a high labeling rate by the radioisotope to a ligand-conjugated antibody.

[0011]

As a result of intensive studies to solve the above problem, the present inventors have found that if a radioactive metal nuclide and an antibody conjugated with a ligand compound are reacted in a reaction solution not containing a pH buffer but containing water and having a pH of 2 or more and 10 or less, a high labeling rate can be imparted by the radioisotope to the ligand-conjugated antibody.

That is, the present invention provides a method for producing a conjugate of a radioactive metal nuclide and a ligand-conjugated antibody, the method including a step of reacting a radioactive metal nuclide with a ligand-conjugated antibody in a reaction solution containing water to coordinate the radioactive metal nuclide to the ligand compound, wherein the reaction solution does not contain a pH buffer and has a pH of 2 or more and 10 or less.

[0012]

According to the present invention, it is possible to provide a method for producing a conjugate that includes a radioisotope, a ligand compound, and an antibody and is capable of imparting a high labeling rate by the radioisotope to a ligand-conjugated antibody.

Brief Description of Drawing

[0013]

Fig. 1 is a graph showing the antigen-binding activity of conjugates produced in Example 1 and Comparative Example 1 to SK-OV-3 tumor sections and MDA-MB-231 tumor sections.

Description of Embodiments

[0014]

Hereinafter, the present invention will be described on the basis of preferred embodiments of the present invention.

The method for producing a conjugate of the present invention includes a step of reacting a radioactive metal nuclide with a ligand-conjugated antibody in a reaction solution containing water to coordinate the radioactive metal nuclide to the ligand compound. The step is performed in a state in which the reaction solution contains water but does not contain a pH buffer, and has a pH of 2 or more and 10 or less. Through the step, the radioisotope is bound to the ligand compound conjugated with the antibody, by a combination of bonds such as a covalent bond and an ionic bond in addition to a coordinate bond, and thus a conjugate is obtained in which the antibody is directly labeled.

[0015]

From the viewpoint of enhancing the labeling rate, the radioisotope used in the step is preferably used in the form of a compound that can ionize in water, and more preferably used in the form of a radioisotope ion (hereinafter, these forms are also collectively referred to as a “radioisotope source”). As the radioisotope source, for

example, a radioisotope ion-containing liquid can be used in which a radioisotope ion is dissolved or dispersed in a solvent mainly containing water.

[0016]

The nuclide of the radioisotope is preferably at least one selected from ^{68}Ga , ^{89}Zr , ^{90}Y , ^{111}In , ^{177}Lu , and ^{225}Ac , and particularly preferably at least one of ^{89}Zr or ^{225}Ac from the viewpoint of high labeling rate and easy labeling.

[0017]

^{89}Zr is a β^+ ray decay nuclide and is an electron capture decay nuclide. ^{89}Zr can be produced, for example, by a nuclear reaction of $^{89}\text{Y}(\text{p}, \text{n})^{89}\text{Zr}$ using a cyclotron. Specifically, a solution obtained by dissolving a ^{89}Y target after proton irradiation with an acid is passed through a column cartridge or the like carrying a collector capable of adsorbing ^{89}Zr . Thereafter, the column cartridge is washed with a solvent such as water, then an oxalic acid aqueous solution is passed through the column cartridge, and thus ^{89}Zr ions can be eluted and collected as a solution.

[0018]

^{225}Ac can be generated by any of milking from a Th-229 generator, Th-232 spallation, Ra-226 neutron addition (a method in which a nuclear reaction of $^{226}\text{Ra}(\text{n}, 2\text{n})^{225}\text{Ra}$ and β -decay of ^{225}Ra are performed), Ra-226 photonuclear reaction (a method in which a nuclear reaction of $^{226}\text{Ra}(\gamma, \text{n})^{225}\text{Ra}$ and β -decay of ^{225}Ra are performed), or Ra-226 nuclear transformation (nuclear reaction of $^{226}\text{Ra}(\text{p}, 2\text{n})^{225}\text{Ac}$ using a cyclotron). In the case of using Ra-226 nuclear transformation, the generated ^{225}Ac can be purified by, for example, separation and purification from a ^{226}Ra target, and purified ^{225}Ac can be obtained by dissolving a ^{226}Ra target containing ^{225}Ac with an acid or the like, adding an alkali to the solution to precipitate a salt containing ^{225}Ac , and separating and purifying the salt.

[0019]

The ligand compound used in the step is not particularly limited as long as it is a compound that can be coordinated with a radioisotope, and examples of the ligand compound include the following organic compounds and compounds including a structure derived from a compound described below.

[0020]

CB-TE2A (1,4,8,11-tetraazabicyclo[6.6.2]hexadecane-4,11-diacetic acid)

CDTA (cyclohexane-trans-1,2-diamine tetra-acetic acid)

CDTPA (4-cyano-4-[[[(dodecylthio)thioxomethyl]thio]-pentanoic acid)

[0021]

DOTA (1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid)

DOTMA ((1R,4R,7R,10R)- $\alpha,\alpha',\alpha'',\alpha'''$ -tetramethyl-1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid)

DOTAM (1,4,7,10-tetrakis(carbamoylmethyl)-1,4,7,10-tetraazacyclododecane)

DOTA-GA (α -(2-carboxyethyl)-1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid)

DOTP (((1,4,7,10-tetraazacyclododecane-1,4,7,10-tetrayl)tetrakis(methylene))tetrakisphosphonic acid)

DOTMP (1,4,7,10-tetraazacyclododecane-1,4,7,10-tetrakis(methylenephosphonic acid))

DOTA-4AMP (1,4,7,10-tetraazacyclododecane-1,4,7,10-tetrakis(acetamidomethylenephosphonic acid))

DO2P (tetraazacyclododecane dimethanephosphonic acid)

[0022]

Deferoxamine (DFO)

DTPA (N,N-bis[2-[bis(carboxymethyl)amino]ethyl]-glycine)

DTPA-BMA (5,8-bis(carboxymethyl)-11-[2-(methylamino)-2-oxoethyl]-3-oxo-2,5,8,11-tetraazatridecan-13-oic acid)

EDTA (2,2',2'',2'''-(ethane-1,2-diylbis(azanetriyl))tetraacetic acid)

NOTA (1,4,7-triazacyclononane-1,4,7-triacetic acid)

OTP (1,4,7-triazacyclononane-1,4,7-triyltris(methylenephosphonic acid)

[0023]

TETPA (1,4,8,11-tetraazacyclotetradecane-1,4,8,11-tetrapropionic acid)

TETA (1,4,8,11-tetraazacyclotetradecane-N,N',N'',N'''-tetraacetic acid)

TTHA (3,6,9,12-tetrakis(carboxymethyl)-3,6,9,12-tetraazatetradecanedioic acid)

HEHA (1,2,7,10,13-hexaazacyclooctadecane-1,4,7,10,13,16-hexaacetic acid)

1,2-HOPO (N,N',N'',N'''-tetra(1,2-dihydro-1-hydroxy-2-oxopyridine-6-carbonyl)-1,5,10,14-tetraazatetradecane)

PEPA (1,4,7,10,13-pentaazacyclopentadecane-N,N',N'',N''',N''''-penta-acetic acid)

[0024]

H4octapa (N,N'-bis(6-carboxy-2-pyridylmethyl)-ethylenediamine-N,N'-diacetic acid)

H2bispa2 (6,6'-({9-hydroxy-1,5-bis(methoxycarbonyl)-2,4-di(pyridine-2-yl)-3,7-diazabicyclo[3.3.1]nonane-3,7-diyl} bis(-methylene))dipicolinic acid)

H2dedpa (1,2-[{6-(carboxy)-pyridin-2-yl}-methylamino]ethane)

H2macropa (6-(1,4,10,13-tetraoxa-7,16-diazacyclooctadecan-N,N'-methyl)picolinic acid)

H5decapa (N,N''-bis(6-carboxy-2-pyridylmethyl)-diethylenetriamine-
N,N',N''-triacetic acid)

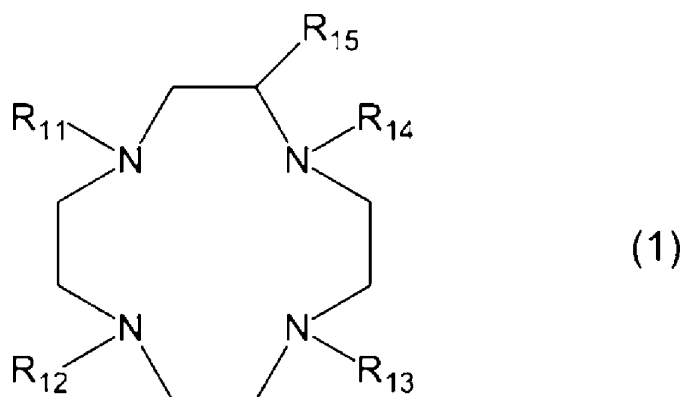
H6phospa (N,N'-(methylenephosphonate)-N,N'-[6-
(methoxycarbonyl)pyridin-2-yl]-methyl-1,2-diaminoethane)

HP-D03A (hydroxypropyltetraazacyclododecanetriacetic acid)

[0025]

Among these, the ligand compound used in the step is preferably an organic compound having a structure represented by Formula (1) described below.

[0026]



[0027]

In Formula (1), R_{11} , R_{13} , and R_{14} each independently represent a group including $-(CH_2)_pCOOH$, $-(CH_2)_pC_5H_4N$, $-(CH_2)_pPO_3H_2$, or $-(CH_2)_pCONH_2$. Each p is independently an integer of 0 or more and 3 or less.

[0028]

In Formula (1), one of R_{12} or R_{15} represents a hydrogen atom, a carboxyl group, or a carboxyalkyl group having 2 or 3 carbon atoms, the other represents a substituent that is to be conjugated with the antibody, R_{15} is a hydrogen atom when R_{12} is a substituent that is to be conjugated with the antibody, and R_{15} is a substituent that is to be conjugated with the antibody when R_{12} is not a substituent that is to be conjugated

with the antibody.

[0029]

The ligand compound used in the step is more preferably 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid (DOTA) or a compound including a structure derived from a derivative of DOTA, and specifically, the ligand compound is still more preferably one compound described below or a compound including a structure derived from a compound described below. The ligand compound used in the step is preferably water-soluble.

[0030]

DOTA (1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid)

DOTMA ((1R,4R,7R,10R)- $\alpha,\alpha',\alpha'',\alpha'''$ -tetramethyl-1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid)

DOTAM (1,4,7,10-tetrakis(carbamoylmethyl)-1,4,7,10-tetraazacyclododecane)

DOTA-GA (α -(2-carboxyethyl)-1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid)

DOTP (((1,4,7,10-tetraazacyclododecane-1,4,7,10-tetrayl)tetrakis(methylene))tetraphosphonic acid)

DOTMP (1,4,7,10-tetraazacyclododecane-1,4,7,10-tetrakis(methylenephosphonic acid))

DOTA-4AMP (1,4,7,10-tetraazacyclododecane-1,4,7,10-tetrakis(acetamidomethylenephosphonic acid))

DO2P (tetraazacyclododecane dimethanephosphonic acid)

[0031]

The reaction solution in the step contains water but does not contain a pH

buffer and preferably does not contain an organic solvent.

[0032]

As the water, water commonly used in the present technical field can be adopted, and for example, distilled water or ion-exchanged water can be used.

[0033]

The pH buffer exhibits a pH buffering action in a predetermined pH range when dissolved in water. In the present invention, it has been found that if the reaction solution does not contain a pH buffer and if a radioactive metal nuclide and an antibody conjugated with the ligand compound are reacted in the reaction solution, a high labeling rate can be imparted by the radioisotope to the antibody.

[0034]

The pH buffer that is not contained is a buffer such as sodium acetate, ammonium acetate, phosphoric acid, phosphate buffered physiological saline, a trishydroxymethylaminomethane buffer solution (Tris buffer solution), a 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid buffer solution (HEPES buffer solution), or a tetramethylammonium acetate buffer solution.

The phrase “not containing a buffer” means that a buffer is not intentionally contained in the reaction solution but may be unavoidably mixed in the reaction solution.

[0035]

Examples of the organic solvent that is not contained include water-soluble organic solvents and water-insoluble organic solvents.

Examples of the water-soluble organic solvents include protic polar solvents such as methanol and ethanol, and aprotic polar solvents such as acetonitrile, N,N-dimethylformamide, tetrahydrofuran, dimethyl sulfoxide, and acetone.

Examples of the water-insoluble organic solvents include hexane, toluene, and ethyl acetate.

If such an organic solvent is not contained, a step of separating an organic solvent, such as a solid phase extraction method, is not to be performed, and therefore a high labeling rate can be stably achieved while radiolysis due to concentration is avoided.

The phrase “not containing an organic solvent” means that an organic solvent is not intentionally contained in the reaction solution but may be unavoidably mixed in the reaction solution.

[0036]

In the present invention, the reaction solution has a pH of 2 or more and 10 or less. Specifically, in the case of using, as the radioisotope, ^{89}Zr as a radioactive metal nuclide, the pH is preferably 2 or more and 4 or less, and more preferably 3. In the case of using ^{225}Ac as a radioactive metal nuclide, the pH is preferably 4 or more and 10 or less, and more preferably 6 or more and 9 or less.

If the pH of the reaction solution is set within the above range, when a radioactive metal nuclide and an antibody conjugated with the ligand compound are reacted in the reaction solution, a high labeling rate can be imparted by the radioisotope to the antibody.

[0037]

The pH of the reaction solution is adjusted to be within the above pH range before starting the reaction. Specifically, an inorganic acid such as hydrochloric acid or a metal hydroxide such as sodium hydroxide is added to water to adjust the pH of the reaction solution.

When a radioactive metal nuclide and a ligand-conjugated antibody are

reacted in the reaction solution, the reaction solution may contain the radioactive metal nuclide in advance and then contain the ligand-conjugated antibody for the reaction, or the reaction solution may contain the radioactive metal nuclide and the ligand-conjugated antibody at the same time. Alternatively, the reaction solution may contain the ligand-conjugated antibody in advance and then contain the radioactive metal nuclide.

[0038]

Next, the reaction pressure is set to, for example, atmospheric pressure, and heating is performed as necessary. By heating, a further improvement in labeling efficiency can be achieved in a short production time. Specifically, the temperature is preferably 30°C or higher and 80°C or lower. The temperature is more preferably 50°C or higher and 80°C or lower in the case of using, as the radioisotope, ^{89}Zr as a radioactive metal nuclide, and more preferably 30°C or higher and 70°C or lower in the case of using ^{225}Ac as a radioactive metal nuclide. Examples of the method of heating include a method of applying heat from the outside of the reaction system using, for example, a block heater, an air heater, a water bath, an oil bath, a microwave, or a mantle heater. A block heater or an air heater is particularly preferably used for heating from the viewpoint of being able to be incorporated into an automatic synthesizer.

The reaction time is preferably 10 minutes or more and 150 minutes or less, and more preferably 15 minutes or more and 120 minutes or less on condition that the reaction temperature is as described above.

[0039]

The amount of the reaction solution in the step may be appropriately changed according to the production scale, and is practically 0.01 mL or more and 100 mL or less at the start of the step from the viewpoint of practicality in the production step.

[0040]

From the viewpoint of enhancing the labeling rate with the target radioisotope, the concentration of the antibody in the reaction solution is preferably 1 $\mu\text{mol/L}$ or more and 2000 $\mu\text{mol/L}$ or less, more preferably 5 $\mu\text{mol/L}$ or more and 100 $\mu\text{mol/L}$ or less, and still more preferably 10 $\mu\text{mol/L}$ or more and 50 $\mu\text{mol/L}$ or less at the start of the step.

[0041]

In the present invention, as necessary, to the reaction solution, at least one can be added that is selected from the group of saccharides consisting of sucrose, trehalose, mannitol, maltose, lactose, arabinose, xylose, sorbitol, fructose, glucose, mannose, melezitose, and nystose, and is preferably selected from the group of saccharides consisting of glucose, xylose, sucrose, trehalose, mannitol, and sorbitol. Thus, it is possible to suppress aggregation of the antibody and, consequently, the conjugate including the radioisotope, the ligand compound, and the antibody.

The amount of these aggregation inhibitors added is, for example, preferably 10 mmol/L or more and 2000 mmol/L or less, and more preferably 100 mmol/L or more and 1000 mmol/L or less.

[0042]

In the finally obtained conjugate, the ligand compound may be connected to the antibody via a linker.

Examples of the antibody include chimeric antibodies, humanized antibodies, and human antibodies, and the antibody is preferably a human IgG, and more preferably human IgG1, IgG2, or IgG4.

Examples of the linker include substituted or unsubstituted alkyl groups, substituted or unsubstituted heteroalkyl groups, polyethylene glycol (PEG) groups,

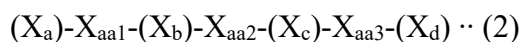
peptides, sugar chains, a disulfide group, and combinations thereof.

The ligand compound preferably is conjugated with the antibody site-specifically, and more preferably at the Fc region, via the linker. In this case, as the linker, a linker can be used that includes a peptide having Formula (2) described below and including 13 or more and 17 or less amino acid residues (hereinafter, also referred to as “antibody-conjugated peptide”) and is formed by a crosslinking reaction between the antibody-conjugated peptide modified with a crosslinking agent and the antibody.

[0043]

For Formula (2), the description will be given assuming that the left side of the amino acid sequence on the paper surface indicates the N-terminal side, and the right side of the amino acid sequence on the paper surface indicates the C-terminal side. In a case where the ligand compound is connected to the antibody via the antibody-conjugated peptide as a linker, the position at which the ligand compound and the antibody-conjugated peptide are linked is not particularly limited, and for example, the ligand compound can be linked directly or indirectly to the N-terminal or the C-terminal, preferably the N-terminal of the antibody-conjugated peptide. The C-terminal of the antibody-conjugated peptide may be modified by amidation or the like in order to, for example, improve the stability.

[0044]



In Formula (2), X_a , X_b , X_c , and X_d represent consecutive Xs of which number is a, consecutive Xs of which number is b, consecutive Xs of which number is c, and consecutive Xs of which number is d, respectively, X represents an amino acid residue having a side chain having neither a thiol group nor a haloacetyl group, a, b, c, and d are each independently an integer of 1 or more and 5 or less and satisfy $a + b + c + d \leq 14$,

X_{aa1} and X_{aa3} each independently represent an amino acid residue derived from an amino acid having a side chain having a thiol group, or one of X_{aa1} or X_{aa3} represents an amino acid residue derived from an amino acid having a side chain having a thiol group and the other represents an amino acid residue derived from an amino acid having a side chain having a haloacetyl group, X_{aa1} and X_{aa3} are linked to each other, and X_{aa2} is a lysine residue, an arginine residue, a cysteine residue, an aspartic acid residue, a glutamic acid residue, a 2-aminosuberic acid residue, or a diaminopropionic acid residue, and is modified with the crosslinking agent.

[0045]

Examples of the amino acid residue that can be included in X in Formula (2) include amino acid residues derived from an amino acid such as glycine, alanine, phenylalanine, proline, asparagine, aspartic acid, glutamic acid, arginine, histidine, serine, threonine, tyrosine, or methionine, and X may be an amino acid residue including the same type of amino acids or may be an amino acid residue including different types of amino acids.

[0046]

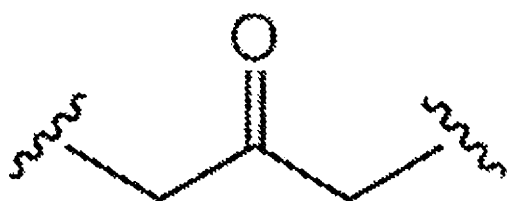
In Formula (2), a, b, c, and d are not particularly limited as long as they are a number within the above-described range, but from the viewpoint of the binding stability between the peptide and the antibody, a is preferably an integer of 1 or more and 3 or less, b is preferably an integer of 1 or more and 3 or less, c is preferably an integer of 3 or more and 5 or less, and d is preferably an integer of 1 or more and 3 or less under the condition of $a + b + c + d \leq 14$.

[0047]

X_{aa1} and X_{aa3} are amino acid residues derived from an amino acid having a side chain having a thiol group, and the amino acids may be the same or different.

Examples of the amino acid having a side chain having a thiol group include cysteine and homocysteine. Such amino acid residues are preferably bound by a disulfide bond, or the sulfide groups are preferably bound via a linker having Formula (3) described below. In Formula (3), the broken line part represents a binding moiety with a sulfide group.

[0048]



(3)

[0049]

Instead of the above-described combination, combination of X_{aa1} and X_{aa3} may be such that one of X_{aa1} or X_{aa3} is an amino acid residue derived from an amino acid having a side chain having a thiol group and the other is an amino acid residue derived from an amino acid having a side chain having a haloacetyl group. These amino acid residues are bound to each other via a thioether bond. The haloacetyl group has a terminal substituted with a halogen such as iodine, and reacts with the thiol group in the side chain of the other amino acid residue to eliminate the halogen, and thus a thioether bond is formed.

[0050]

Specific examples of the amino acid sequence of the antibody-conjugated peptide having Formula (2) include peptides described in WO 2016/186206 A, WO 2017/217347 A, and WO 2018/230257 A, and these can also be used.

[0051]

Among these, the amino acid sequence of the antibody-conjugated peptide is

preferably any one of the following sequences (1) to (14), and more preferably the following sequence (1), (2), (13), or (14). In the following amino acid sequences (1) to (14), (X_{aa2}) represents a lysine residue, a cysteine residue, an aspartic acid residue, a glutamic acid residue, 2-aminosuberic acid, or diaminopropionic acid, and both (X_{aa1}) and (X_{aa3}) represent a homocysteine residue. In the following amino acid sequences (1) to (14), amino acids other than (X_{aa1}), (X_{aa2}), and (X_{aa3}) are represented by one-letter codes.

[0052]

- (1) DCAYH(X_{aa2})GELVWCT (SEQ ID NO: 9)
- (2) GPDCAYH(X_{aa2})GELVWCTFH (SEQ ID NO: 10)
- (3) RCAYH(X_{aa2})GELVWCS (SEQ ID NO: 11)
- (4) GPRCAYH(X_{aa2})GELVWCSFH (SEQ ID NO: 12)
- (5) SPDCAYH(X_{aa2})GELVWCTFH (SEQ ID NO: 13)
- (6) GDDCAYH(X_{aa2})GELVWCTFH (SEQ ID NO: 14)
- (7) GPSCAYH(X_{aa2})GELVWCTFH (SEQ ID NO: 15)
- (8) GPDCAYH(X_{aa2})GELVWCSFH (SEQ ID NO: 16)
- (9) GPDCAYH(X_{aa2})GELVWCTHH (SEQ ID NO: 17)
- (10) GPDCAYH(X_{aa2})GELVWCTFY (SEQ ID NO: 18)
- (11) SPDCAYH(X_{aa2})GELVWCTFY (SEQ ID NO: 19)
- (12) SDDCAYH(X_{aa2})GELVWCTFY (SEQ ID NO: 20)
- (13) RGNCAHY(X_{aa2})GQLVWCTYH (SEQ ID NO: 21)
- (14) G(X_{aa1})DCAYH(X_{aa2})GELVWCT(X_{aa3})H (SEQ ID NO: 22)

[0053]

The above embodiments of the present invention include the following technical ideas.

[1] A method for producing a conjugate of a radioactive metal nuclide and an antibody, the method including:

a step of reacting a radioactive metal nuclide with an antibody conjugated with a ligand compound in a reaction solution containing water to coordinate the radioactive metal nuclide to the ligand compound, wherein

the reaction solution does not contain a pH buffer and has a pH of 2 or more and 10 or less.

[2] The method according to [1], wherein the radioactive metal nuclide is at least one selected from ^{68}Ga , ^{89}Zr , ^{90}Y , ^{111}In , ^{177}Lu , and ^{225}Ac .

[3] The method according to [1] or [2], wherein the radioactive metal nuclide is ^{89}Zr , and the reaction solution has a pH of 2 or more and 4 or less.

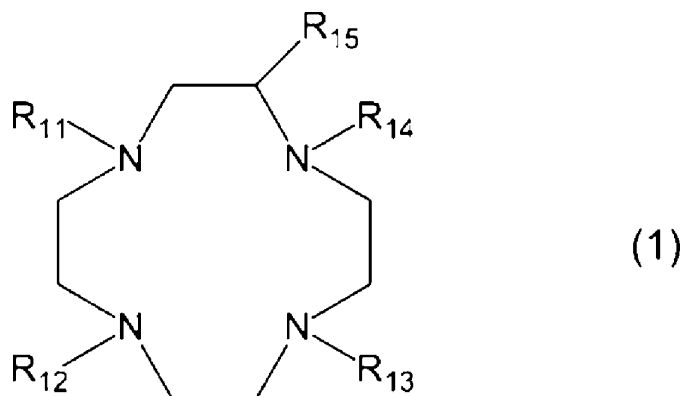
[4] The method according to any one of [1] to [3], wherein the radioactive metal nuclide is ^{225}Ac , and the reaction solution has a pH of 5 or more and 10 or less.

[5] The method according to any one of [1] to [4], wherein a concentration of the antibody in the reaction solution is 1 $\mu\text{mol/L}$ or more and 2000 $\mu\text{mol/L}$ or less.

[6] The method according to any one of [1] to [5], wherein the reaction solution is heated to 30°C or higher and 80°C or lower to coordinate the radioactive metal nuclide to the ligand compound.

[7] The method according to any one of [1] to [6], wherein the reaction solution contains at least one aggregation inhibitor selected from the group of saccharides, the group consisting of sucrose, trehalose, mannitol, maltose, lactose, arabinose, xylose, sorbitol, fructose, glucose, mannose, melezitose, and nystose.

[8] The method according to any one of [1] to [7], wherein the ligand compound has Formula (1) described below:

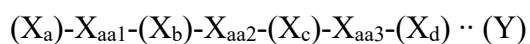


wherein R_{11} , R_{13} , and R_{14} each independently represent a group including - $(CH_2)_pCOOH$, $-(CH_2)_pC_5H_4N$, $-(CH_2)_pPO_3H_2$, $-(CH_2)_pCONH_2$, or - $(CHCOOH)(CH_2)_pCOOH$, one of R_{12} or R_{15} represents a hydrogen atom, a carboxyl group, or a carboxyalkyl group having 2 or 3 carbon atoms, another of R_{12} or R_{15} represents a substituent that is to be conjugated with the antibody, p is an integer of 0 or more and 3 or less, R_{15} is a hydrogen atom when R_{12} is a substituent that is to be conjugated with the antibody, and R_{15} is a substituent that is to be conjugated with the antibody when R_{12} is not a substituent that is to be conjugated with the antibody.

[9] The method according to any one of [1] to [8], wherein an Fc region of the antibody is site-specifically conjugated with the ligand compound via a linker.

[10] The method according to any one of [1] to [9], wherein the antibody is a human IgG antibody, and the linker includes a peptide including 13 or more and 17 or less amino acid residues and is formed by a crosslinking reaction between the peptide modified with a crosslinking agent and the antibody,

the peptide having Formula (Y) below:



wherein X_a , X_b , X_c , and X_d represent consecutive Xs of which number is a, consecutive Xs of which number is b, consecutive Xs of which number is c, and consecutive Xs of which number is d, respectively, X represents an amino acid residue

having a side chain having neither a thiol group nor a haloacetyl group, a, b, c, and d are each independently an integer of 1 or more and 5 or less and satisfy $a + b + c + d \leq 14$, X_{aa1} and X_{aa3} each independently represent an amino acid residue derived from an amino acid having a side chain having a thiol group, or one of X_{aa1} or X_{aa3} represents an amino acid residue derived from an amino acid having a side chain having a thiol group and another of X_{aa1} or X_{aa3} represents an amino acid residue derived from an amino acid having a side chain having a haloacetyl group, X_{aa1} and X_{aa3} are linked to each other, and X_{aa2} is a lysine residue, an arginine residue, a cysteine residue, an aspartic acid residue, a glutamic acid residue, a 2-aminosuberic acid residue, or a diaminopropionic acid residue, and is modified with the crosslinking agent.

Examples

[0054]

Hereinafter, the present invention will be described in more detail with reference to Examples. However, the scope of the present invention is not limited to Examples.

[0055]

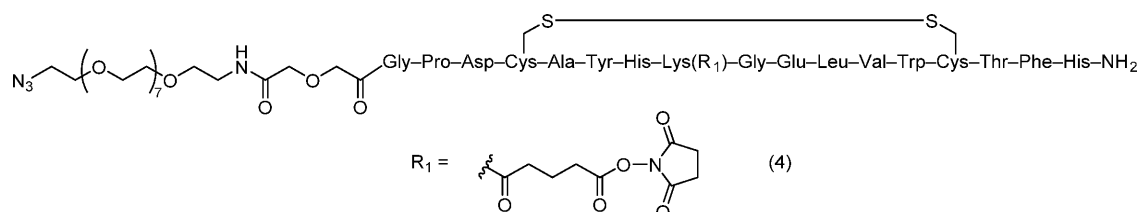
[Example 1] Production of ^{89}Zr -Pertuzumab Conjugate

(1) Synthesis of Peptide-Conjugated Pertuzumab

A peptide having Formula (4) described below and including 17 amino acid residues was obtained by the method described in WO 2017/217347 A. The amino acid sequence of this peptide is the same as the sequence of SEQ ID NO. 2 described in paragraph [0038] of WO 2017/217347 A in a case where X_{aa1} is a lysine residue, and the side chain terminal amino group of the lysine residue is conjugated with a structure of R_1 in Formula (4). A disulfide bond is formed between two cysteine residues, and an

azido group is bound to the N-terminal of the peptide via a linker structure having diglycolic acid and eight PEG groups.

[0056]



In Formula (4), Gly represents glycine, Pro represents proline, Asp represents aspartic acid, Cys represents cysteine, Ala represents alanine, Tyr represents tyrosine, His represents histidine, Lys represents lysine, Glu represents glutamic acid, Leu represents leucine, Val represents valine, Trp represents tryptophan, Phe represents phenylalanine, and Thr represents threonine.

[0057]

Next, 1 mL of a pertuzumab solution (containing 30 mg of pertuzumab (trade name: Perjeta, produced by Roche)) was added into a centrifuge tube into which 6.5 mL of a 20 mmol/L acetic acid-sodium acetate buffer solution (pH: 6.0) was added. Then, 140 μ L of a dimethyl sulfoxide (DMSO) solution of the peptide including 17 amino acid residues (peptide concentration: 2.4 mmol/L) was added, then pipetting was immediately performed, and the mixture was allowed to stand at room temperature and reacted for 60 minutes to obtain a solution containing peptide-conjugated pertuzumab.

[0058]

Next, the solution containing peptide-conjugated pertuzumab obtained as described above was diluted with 22.5 mL of a 20 mmol/L acetic acid-sodium acetate buffer solution (pH: 6.0), the diluted solution was passed through an IgG-BP column (a column obtained by immobilizing 5632 nmol of a peptide having a sequence

represented by GSGGSGPDCAHYHRGELVWCTFH on a HiTrap NHS-activated HP column (manufactured by Cytiva) having a volume of 5 mL, in which peptide cysteine residues are crosslinked by a disulfide bond), and thus the antibody was retained. A 0.10 mol/L acetic acid-sodium acetate buffer solution (pH: 5.7) containing 0.15 mol/L sodium chloride was passed through the column to remove the peptide-conjugated antibody in which one molecule of pertuzumab is conjugated with two molecules of the peptide (hereinafter, referred to as “divalent peptide-conjugated pertuzumab”), and then a 0.1 mol/L glycine-hydrochloric acid buffer solution (pH: 3.5) was passed through the column to collect a fraction containing peptide-conjugated pertuzumab in which one molecule of pertuzumab is conjugated with one molecule of the peptide (hereinafter, referred to as “monovalent peptide-conjugated pertuzumab”).

[0059]

The fraction containing the monovalent peptide-conjugated pertuzumab was poured into an ultrafiltration filter (Amicon Ultra-15, manufactured by Merck KGaA) into which 10 mL of a pertuzumab formulation buffer solution (aqueous solution containing sucrose at a concentration of 41 g/L, L-histidine at a concentration of 3.1 g/L, and glacial acetic acid at a concentration of 657 mg/L) was added in advance, and the solution was centrifuged with a centrifuge (centrifugation conditions: 25°C, 3000 x g, 20 min). After the centrifugation, the filtrate was discarded, a pertuzumab formulation buffer solution was added again so that the total amount of the solution was 15 mL, and the second centrifugation was performed under the same conditions.

[0060]

Next, a pertuzumab formulation buffer solution was added to the solution obtained as described above to obtain a peptide-conjugated pertuzumab solution in which the concentration of the monovalent peptide-conjugated pertuzumab was adjusted

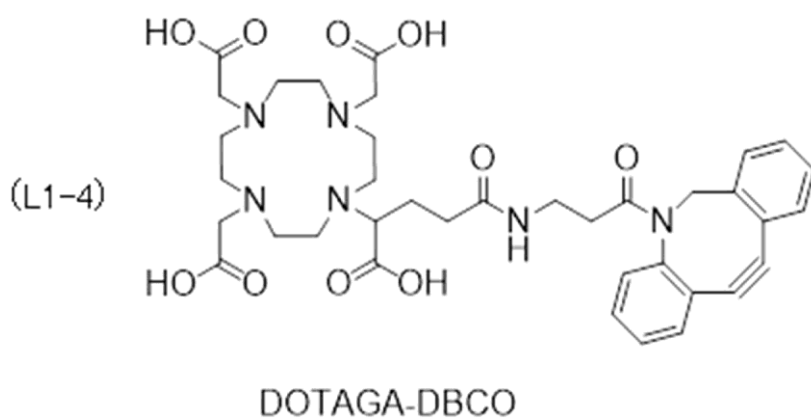
to 15 mg/mL (hereinafter, sometimes referred to as “solution A”). At this time, the concentration of the monovalent peptide-conjugated pertuzumab in the solution A was measured using a UV-Vis spectrophotometer (NanoDrop-2000, manufactured by Thermo Scientific Inc.) under the measurement conditions of a measurement wavelength of 280 nm, a measurement optical path length of 1 mm, and an absorbance coefficient 0.1% (280 nm) of 1.37 (an absorbance coefficient of a 1 mg/mL IgG solution at 280 nm).

[0061]

(2) Synthesis of Ligand-Conjugated Pertuzumab

DOTAGA-DBCO shown in Formula (L1-4) below was produced on the basis of the method described in Bernhard et al. DOTAGA-Anhydride: A Valuable Building Block for the Preparation of DOTA-Like Chelating Agents Chem. Eur. J. 2012, 18, 7834-7841.

[0062]



Water was added to the DOTAGA-DBCO to prepare a DOTAGA-DBCO-containing solution having a concentration of 4 mg/mL (hereinafter, sometimes referred

to as “solution B”).

[0063]

Next, 0.6 mL of the solution A (containing 9 mg and 0.06 μ mol of the monovalent peptide-conjugated pertuzumab) was added into a microtube. Then, 0.33 mL of the solution B was added and pipetting was immediately performed. The microtube was allowed to stand on a block heater set at 37°C for 1 hour, and the monovalent peptide-conjugated pertuzumab synthesized in (1) above and the DOTAGA-DBCO were subjected to a click reaction.

[0064]

Next, the reaction mixture liquid obtained as described above was diluted with a pertuzumab formulation buffer solution so that the volume of the diluted liquid was 10 mL, the diluted liquid was passed through a protein A column (1 mL MabSelect SuRe, manufactured by Cytiva), and thus the peptide-conjugated antibody was retained. A 0.02 mol/L histidine buffer solution (pH: 6.0) containing 0.15 mol/L sodium chloride was passed through the column, unreacted DOTAGA-DBCO was washed, and then a 0.1 mol/L glycine-hydrochloric acid buffer solution (pH: 3.5) was passed through the column to collect a fraction containing the ligand-conjugated pertuzumab.

[0065]

The total amount of the collected solution obtained above was added into an ultrafiltration filter (Amicon Ultra-15, manufactured by Merck KGaA) into which 10 mL of water for injection was added in advance, and the solution was centrifuged with a centrifuge (centrifugation conditions: 25°C, 3000 x g, 20 min). After the centrifugation, the filtrate was discarded, 13.5 mL of water for injection was added again, and centrifugation was performed under the same conditions. This centrifugation was performed twice in total, and finally, the centrifuged residue was diluted with water for

injection to obtain a ligand-conjugated pertuzumab solution in which the concentration of the monovalent ligand-conjugated pertuzumab obtained by the reaction between the monovalent peptide-conjugated antibody and DOTAGA-DBCO was 15 mg/mL (hereinafter, sometimes referred to as “solution C”).

[0066]

(3) Labeling Reaction

^{89}Zr obtained by a nuclear reaction of $^{89}\text{Y}(\text{p}, \text{n})^{89}\text{Zr}$ and prepared into a $^{89}\text{Zr}^{4+}$ ion aqueous solution (manufactured by Nihon Medi-Physics Co., Ltd.) was added into a vial, and the solvent was distilled off.

[0067]

Into the vial, 0.1 mol/L hydrochloric acid was added, pipetting was performed to wash the vial well mainly at the bottom surface, and the liquid was dispensed into a microtube so that the radioactivity was 19.8 MBq. Into the microtube, 0.1 mol/L hydrochloric acid and water for injection were added, and then a 1 mol/L sodium hydroxide aqueous solution was added to obtain 545.6 μL of a $^{89}\text{Zr}^{4+}$ ion solution having a pH of 3.0 (radioactivity concentration in the reaction solution: 36.3 MBq/mL). The pH was measured with a pH test paper.

[0068]

To the $^{89}\text{Zr}^{4+}$ ion solution, 60 μL of the solution C (containing 6 nmol of the monovalent ligand-conjugated pertuzumab) was added to set the concentration of the monovalent ligand-conjugated pertuzumab in the reaction solution to 10 $\mu\text{mol/L}$, and the reaction solution was stirred with a vortex mixer. Next, the reaction solution was heated at 70°C for 30 minutes using a block heater to coordinate $^{89}\text{Zr}^{4+}$ ions to the ligand of the ligand-conjugated pertuzumab, and thus a ^{89}Zr -pertuzumab conjugate was synthesized.

The reaction mixture was extracted in an amount of 0.5 μL and separated by thin layer chromatography (iTLC-SG, manufactured by Agilent Technologies, Inc., developing solvent: mixed liquid of EDTA aqueous solution (0.1 mol/L) and hydrochloric acid (1 mol/L) (volume ratio 1 : 1)). The developed thin layer chromatogram was introduced into a TLC analyzer (GITA Star, manufactured by M&S Instruments Inc.), and the radioactivity count of the total ^{89}Zr including unreacted ^{89}Zr and the radioactivity count of the ^{89}Zr -pertuzumab conjugate in the reaction mixture were measured. The percentage of the radioactivity count of the ^{89}Zr -pertuzumab conjugate to the total ^{89}Zr radioactivity count was 99.3%.

[0069]

[Example 2] Production of ^{89}Zr -Trastuzumab Conjugate

(1) Synthesis of Peptide-Conjugated Trastuzumab

In a centrifuge tube into which 25 mL of a 20 mmol/L acetic acid-sodium acetate buffer solution (pH: 6.0) was added, 50 mg of trastuzumab (trade name: Herceptin, produced by Roche) weighed out was dissolved to obtain a 2 mg/mL solution. Then, 233 μL of a DMSO solution of the peptide including 17 amino acid residues (concentration: 2.4 mmol/L), which was the same kind as in Example 1 (1), was added, then pipetting was immediately performed, and the mixture was allowed to stand at room temperature and reacted for 60 minutes to obtain a solution containing peptide-conjugated trastuzumab.

[0070]

Next, the solution containing the peptide-conjugated trastuzumab obtained as described above was diluted with 25 mL of a 20 mmol/L acetic acid-sodium acetate buffer solution (pH: 6.0) and added to an IgG-BP column (the same as used in Example 1 (1)). A 0.10 mol/L acetic acid-sodium acetate buffer solution (pH: 5.7) containing

0.15 mol/L sodium chloride was passed through the column to remove the peptide-conjugated antibody in which one molecule of trastuzumab is conjugated with two molecules of the peptide (hereinafter, referred to as “divalent peptide-conjugated trastuzumab”), and then a 0.10 mol/L acetic acid-sodium acetate buffer solution (pH: 3.5) containing 0.15 mol/L sodium chloride was passed through the column to collect a fraction containing the peptide-conjugated antibody in which one molecule of trastuzumab is conjugated with one molecule of the peptide (hereinafter, referred to as “monovalent peptide-conjugated trastuzumab”).

[0071]

The fraction containing the monovalent peptide-conjugated trastuzumab was poured into an ultrafiltration filter (Amicon Ultra-15, manufactured by Merck KGaA) into which a trastuzumab formulation buffer solution (19 g/L trehalose hydrate, 470 mg/L L-histidine hydrochloride, 300 mg/L L-histidine) was added in advance, and the solution was centrifuged with a centrifuge (centrifugation conditions: 20°C, 2330 x g, 20 min). After the centrifugation, the filtrate was discarded, a trastuzumab formulation buffer solution was added again so that the total amount of the solution was 15 mL, and the second centrifugation was performed under the same conditions. The addition of a trastuzumab formulation buffer solution, the extraction of the filtrate, and the centrifugation were performed three times in total.

[0072]

Next, a trastuzumab formulation buffer solution was added to the solution obtained as described above to obtain a peptide-conjugated trastuzumab solution in which the concentration of the monovalent peptide-conjugated trastuzumab was adjusted to 15 mg/mL (hereinafter, sometimes referred to as “solution D”). At this time, the concentration of the monovalent peptide-conjugated trastuzumab in the solution D

was measured as described above using a UV-Vis spectrophotometer (NanoDrop-2000, manufactured by Thermo Scientific Inc.).

[0073]

(2) Synthesis of Ligand-Conjugated Trastuzumab

Into a microtube, 0.6 mL of the solution D (containing 9 mg and 0.06 μmol of the monovalent peptide-conjugated trastuzumab) was added. Then, 0.33 mL of the solution B prepared in the same manner as in Example 1 (2) was added, and pipetting was immediately performed. The microtube was allowed to stand on a block heater set at 37°C for 1 hour, and the peptide-conjugated trastuzumab synthesized in (1) above and DOTAGA-DBCO were subjected to a click reaction.

[0074]

Next, the reaction mixture liquid obtained as described above was diluted with a trastuzumab formulation buffer solution so that the volume of the diluted liquid was 10 mL, the diluted liquid was passed through a protein A column (1 mL MabSelect SuRe, manufactured by Cytiva), and thus the antibody was retained. A 0.02 mol/L histidine buffer solution (pH: 6.0) containing 0.15 mol/L sodium chloride was passed through the column, unreacted DOTAGA-DBCO was washed, and then a 0.1 mol/L glycine-hydrochloric acid buffer solution (pH: 3.5) was passed through the column to collect a fraction containing the ligand-conjugated trastuzumab.

[0075]

The total amount of the collected solution obtained above was added into an ultrafiltration filter (Amicon Ultra-15, manufactured by Merck KGaA) into which 10 mL of water for injection was added in advance, and the solution was centrifuged with a centrifuge (centrifugation conditions: 25°C, 3000 x g, 20 min). After the centrifugation, the filtrate was discarded, 13.5 mL of water for injection was added again, and

centrifugation was performed under the same conditions. This centrifugation was performed 4 times in total, and finally, the centrifuged residue was diluted with water for injection to obtain a ligand-conjugated trastuzumab solution in which the concentration of the monovalent ligand-conjugated trastuzumab obtained by the reaction between the monovalent peptide-conjugated antibody and DOTAGA-DBCO was 15 mg/mL (hereinafter, sometimes referred to as “solution E”).

[0076]

(3) Labeling Reaction

$^{89}\text{Zr}^{4+}$ ions were coordinated to the ligand of the ligand-conjugated trastuzumab to synthesize a ^{89}Zr -trastuzumab conjugate in the same manner as in Example 1 (3) except that 545.6 μL of a $^{89}\text{Zr}^{4+}$ ion solution containing $^{89}\text{Zr}^{4+}$ ions at 36.3 MBq/mL and having a pH of 3.0 was used and the solution C was changed to the solution E. As a result of analyzing the reaction mixture by a thin layer chromatography method in the same manner as in Example 1 (3), the percentage of the radioactivity count of the ^{89}Zr -trastuzumab conjugate to the total ^{89}Zr radioactivity count was 99.5%.

[0077]

[Example 3] Production of ^{225}Ac -Pertuzumab Conjugate

(1) Synthesis of Ligand-Conjugated Pertuzumab

In accordance with the same procedure as in Examples 1 (1) and 1 (2), ligand-conjugated pertuzumab was synthesized to obtain a solution C.

[0078]

(2) Labeling Reaction

Into a vial containing $^{225}\text{Ac}(\text{NO}_3)_3$ purchased from Oak Ridge National Laboratory, 0.2 mol/L hydrochloric acid was added, pipetting was performed to wash the vial well mainly at the bottom surface, and the liquid was dispensed into a

microtube so that the radioactivity was 0.332 MBq. Into the microtube, 0.2 mol/L hydrochloric acid and water for injection were added, and then a 1 mol/L sodium hydroxide aqueous solution was added to obtain 276 μL of an $^{225}\text{Ac}^{3+}$ ion solution having a pH of 7.0 (radioactivity concentration in the reaction solution: 1.2 MBq/mL). The pH was measured with a pH test paper.

[0079]

To the $^{225}\text{Ac}^{3+}$ ion solution, 30 μL of the solution C (containing 3 nmol of the monovalent ligand-conjugated pertuzumab) was added to set the concentration of the monovalent ligand-conjugated pertuzumab in the reaction solution to 9.8 $\mu\text{mol/L}$, and the reaction liquid was stirred with a vortex mixer. Next, the reaction liquid was heated at 40°C for 30 minutes using a block heater to coordinate $^{225}\text{Ac}^{3+}$ ions to the ligand of the ligand-conjugated pertuzumab, and thus an ^{225}Ac -pertuzumab conjugate was synthesized.

The reaction mixture was extracted in an amount of 2 μL and separated by thin layer chromatography (iTLC-SG, manufactured by Agilent Technologies, Inc., developing solvent: mixed liquid of EDTA aqueous solution (0.1 mol/L) and hydrochloric acid (1 mol/L) (volume ratio 1 : 1)). The developed thin layer chromatogram was introduced into a TLC analyzer (GITA Star, manufactured by M&S Instruments Inc.), and the radioactivity count of the total ^{225}Ac including unreacted ^{225}Ac and the radioactivity count of the ^{225}Ac -pertuzumab conjugate in the reaction mixture were measured. The percentage of the radioactivity count of the ^{225}Ac -pertuzumab conjugate to the total ^{225}Ac radioactivity count was 91.0%.

[0080]

[Example 4] Production of ^{225}Ac -Trastuzumab Conjugate

(1) Synthesis of Ligand-Conjugated Trastuzumab

In accordance with the same procedure as in Examples 2 (1) and 2 (2), ligand-conjugated trastuzumab was synthesized to obtain a solution E.

[0081]

(2) Labeling Reaction

^{225}Ac ions were coordinated to the ligand of the ligand-conjugated trastuzumab to synthesize an ^{225}Ac -trastuzumab conjugate in the same manner as in Example 3 (2) except that 276 μL of an $^{225}\text{Ac}^{3+}$ ion solution having a pH of 7.0 (radioactivity concentration in the reaction solution: 0.98 MBq/mL) was used and the solution C was changed to the solution E. As a result of analyzing the reaction mixture by a thin layer chromatography method in the same manner as in Example 3 (2), the percentage of the radioactivity count of the ^{225}Ac -trastuzumab conjugate to the total ^{225}Ac radioactivity count was 83.6%.

[0082]

[Example 5] Production of ^{225}Ac -Pertuzumab Conjugate under Stabilizer-Added Condition

(1) Synthesis of Ligand-Conjugated Pertuzumab

In accordance with the same procedure as in Examples 1 (1) and 1 (2), ligand-conjugated pertuzumab was synthesized to obtain a solution C.

[0083]

(2) Labeling Reaction

^{225}Ac ions were coordinated to the ligand of the ligand-conjugated pertuzumab to synthesize an ^{225}Ac -pertuzumab conjugate in the same manner as in Example 3 (2) except that 186 μL of an $^{225}\text{Ac}^{3+}$ ion solution having a pH of 7.0 (radioactivity concentration in the reaction solution: 1.5 MBq/mL) was used and 90 μL of a 500 mmol/L sucrose aqueous solution was added to the reaction solution.

As a result of analyzing the reaction mixture by a thin layer chromatography method in the same manner as in Example 3 (2), the percentage of the radioactivity count of the ^{225}Ac -pertuzumab conjugate to the total ^{225}Ac radioactivity count was 91.0%.

[0084]

[Example 6] Production of ^{225}Ac -Trastuzumab Conjugate under Acidic Condition

(1) Synthesis of Ligand-Conjugated Trastuzumab

In accordance with the same procedure as in Examples 2 (1) and 2 (2), ligand-conjugated trastuzumab was synthesized to obtain a solution (E).

[0085]

(2) Labeling Reaction

Into a vial containing $^{225}\text{Ac}(\text{NO}_3)_3$ purchased from Oak Ridge National Laboratory, 0.2 mol/L hydrochloric acid was added, pipetting was performed to wash the vial well mainly at the bottom surface, and the liquid was dispensed into a microtube so that the radioactivity was 0.301 MBq. Into the microtube, 0.2 mol/L hydrochloric acid and water for injection were added, and then a 1 mol/L sodium hydroxide aqueous solution was added to obtain 272 μL of an $^{225}\text{Ac}^{3+}$ ion solution having a pH of 4.0 (radioactivity concentration in the reaction solution: 1.1 MBq/mL). The pH was measured with a pH test paper.

An ^{225}Ac -trastuzumab conjugate was synthesized in the same manner as in Example 4 (2) except that the above-described $^{225}\text{Ac}^{3+}$ ion solution was used as an $^{225}\text{Ac}^{3+}$ ion solution. As a result of analyzing the reaction mixture by a thin layer chromatography method, the percentage of the radioactivity count of the ^{225}Ac -trastuzumab conjugate to the total ^{225}Ac radioactivity count was 93.0%.

[0086]

[Comparative Example 1] Production of ^{89}Zr -Pertuzumab Conjugate with Two-Step Method

(1) Production of ^{89}Zr -labeled DOTAGA-DBCO

To 20 μL of a 0.1 mol/L hydrochloric acid containing $^{89}\text{Zr}^{4+}$ ions having a radioactivity of 19.9 MBq, 20 μL of an aqueous solution (containing 6 nmol of a ligand) containing DOTAGA-DBCO (concentration: 0.3 mmol/L) and sodium acetate (concentration: 0.156 mol/L) and 20 μL of an aqueous solution containing gentisic acid (concentration: 0.15 mol/L) and sodium acetate (concentration: 0.156 mol/L) were added, and the mixture was reacted at 70°C for 60 minutes to coordinate DOTAGA-DBCO to $^{89}\text{Zr}^{4+}$. The obtained reaction mixture of ^{89}Zr -labeled DOTAGA-DBCO was extracted in an amount of 0.5 μL and separated by thin layer chromatography (iTLC-SG, manufactured by Agilent Technologies, Inc., developing solvent: mixed liquid of water for injection and acetonitrile (volume ratio 1 : 1)). The developed thin layer chromatogram was introduced into a TLC analyzer (GITA Star, manufactured by M&S Instruments Inc.), and the radioactivity count of the total ^{89}Zr including unreacted ^{89}Zr and the radioactivity count of ^{89}Zr -labeled DOTAGA-DBCO in the reaction mixture were measured. The percentage of the radioactivity count of ^{89}Zr -labeled DOTAGA-DBCO to the total ^{89}Zr radioactivity count was 98.7%.

[0087]

(2) Peptide-Conjugated Pertuzumab

In accordance with the same procedure as in Example 1 (1), peptide-conjugated pertuzumab was synthesized.

[0088]

(3) Production of ^{89}Zr -Pertuzumab Conjugate

To 90 μL of the ^{89}Zr -labeled DOTAGA-DBCO solution obtained above, 0.06 mL of the solution A (containing 6 nmol of the monovalent antibody) was added and reacted at 37°C for 90 minutes, and the antibody and ^{89}Zr -labeled DOTAGA-DBCO were bound by a click reaction to obtain a ^{89}Zr -pertuzumab conjugate by a production method including two steps of a step of labeling a ligand with ^{89}Zr and a step of a click reaction of a ^{89}Zr ligand. As a result of measuring the radioactivity count of the total ^{89}Zr including unreacted ^{89}Zr and the radioactivity count of the ^{89}Zr -pertuzumab conjugate in the obtained reaction mixture of the ^{89}Zr -pertuzumab conjugate in the same manner as in Example 1 (3), the percentage of the radioactivity count of the ^{89}Zr -pertuzumab conjugate to the total ^{89}Zr radioactivity count was 82.4%.

[0089]

[Comparative Example 2] Production of ^{89}Zr -Trastuzumab Conjugate with Two-Step Method

(1) Production of ^{89}Zr -labeled DOTAGA-DBCO

In accordance with the same procedure as in Comparative Example 1 (1), ^{89}Zr -labeled DOTAGA-DBCO was produced using $^{89}\text{Zr}^{4+}$ ions having a radioactivity of 19.8 MBq. The obtained reaction mixture of ^{89}Zr -labeled DOTAGA-DBCO was extracted in an amount of 0.5 μL and separated by thin layer chromatography (iTLC-SG, manufactured by Agilent Technologies, Inc., developing solvent: mixed liquid of water for injection and acetonitrile (volume ratio 1 : 1)). The developed thin layer chromatogram was introduced into a TLC analyzer (GITA Star, manufactured by M&S Instruments Inc.), and the radioactivity count of the total ^{89}Zr including unreacted ^{89}Zr and the radioactivity count of ^{89}Zr -labeled DOTAGA-DBCO in the reaction mixture were measured. The percentage of the radioactivity count of ^{89}Zr -labeled DOTAGA-DBCO to the total ^{89}Zr radioactivity count was 98.6%.

[0090]

(2) Peptide-Conjugated Trastuzumab

In accordance with the same procedure as in Example 2 (1), peptide-conjugated trastuzumab was synthesized.

[0091]

(3) Production of ^{89}Zr -Trastuzumab Conjugate

To 90 μL of the ^{89}Zr -labeled DOTAGA-DBCO solution obtained above, 0.06 mL of the solution D (containing 6 nmol of the monovalent antibody) was added and reacted at 37°C for 90 minutes, and the antibody and ^{89}Zr -labeled DOTAGA-DBCO were bound to obtain a ^{89}Zr -trastuzumab conjugate. As a result of measuring the radioactivity count of the total ^{89}Zr including unreacted ^{89}Zr and the radioactivity count of the ^{89}Zr -trastuzumab conjugate in the obtained reaction mixture of the ^{89}Zr -trastuzumab conjugate in the same manner as in Example 1 (3), the percentage of the radioactivity count of the ^{89}Zr -trastuzumab conjugate to the total ^{89}Zr radioactivity count was 81.7%.

[0092]

[Comparative Example 3] Production of ^{225}Ac -Pertuzumab Conjugate with Two-Step Method

(1) Production of ^{225}Ac -labeled DOTAGA-DBCO

To 20 μL of a 0.2 mol/L hydrochloric acid containing $^{225}\text{Ac}^{3+}$ ions having a radioactivity of 0.6 MBq, 40 μL of an aqueous solution containing DOTAGA-DBCO (concentration: 0.3 mmol/L) and sodium acetate (concentration: 0.156 mol/L) and 20 μL of sodium acetate aqueous solution (concentration: 0.156 mol/L) were added, and the mixture was reacted at 70°C for 60 minutes to coordinate $^{225}\text{Ac}^{3+}$ to DOTAGA-DBCO. The obtained reaction mixture of ^{225}Ac -labeled DOTAGA-DBCO was extracted in an

amount of 2 μ L and separated by thin layer chromatography (iTLC-SG, manufactured by Agilent Technologies, Inc., developing solvent: mixed liquid of water for injection and acetonitrile (volume ratio 1 : 1)). The developed thin layer chromatogram was introduced into a TLC analyzer (GITA Star, manufactured by M&S Instruments Inc.), and the radioactivity count of the total ^{225}Ac including unreacted ^{225}Ac and the radioactivity count of ^{225}Ac -labeled DOTAGA-DBCO in the reaction mixture were measured. The percentage of the radioactivity count of ^{225}Ac -labeled DOTAGA-DBCO to the total ^{225}Ac radioactivity count was 98.9%.

[0093]

(2) Peptide-Conjugated Pertuzumab

In accordance with the same procedure as in Example 1 (1), peptide-conjugated pertuzumab was synthesized.

[0094]

(3) Production of ^{225}Ac -Pertuzumab Conjugate

To 40 μ L of the ^{225}Ac -labeled DOTAGA-DBCO solution obtained above, 0.06 mL of the solution A (containing 6 nmol of the monovalent antibody) was added and reacted at 37°C for 90 minutes, and the antibody and ^{225}Ac -labeled DOTAGA-DBCO were bound to obtain an ^{225}Ac -pertuzumab conjugate. The obtained reaction mixture of the ^{225}Ac -pertuzumab conjugate was extracted in an amount of 2 μ L and separated by thin layer chromatography (iTLC-SG, manufactured by Agilent Technologies, Inc., developing solvent: mixed liquid of EDTA aqueous solution (0.1 mol/L) and acetonitrile (volume ratio 1 : 1)). The developed thin layer chromatogram was introduced into a TLC analyzer (GITA Star, manufactured by M&S Instruments Inc.), and the radioactivity count of the total ^{225}Ac including unreacted ^{225}Ac and the radioactivity count of the ^{225}Ac -pertuzumab conjugate in the reaction mixture were measured. The

percentage of the radioactivity count of the ^{225}Ac -pertuzumab conjugate to the total ^{225}Ac radioactivity count was 67.3%.

[0095]

[Comparative Example 4] Production of ^{225}Ac -Trastuzumab Conjugate with Two-Step Method

(1) Production of ^{225}Ac -labeled DOTAGA-DBCO

In accordance with the same procedure as in Comparative Example 3 (1), ^{225}Ac -labeled DOTAGA-DBCO was produced. The obtained reaction mixture of ^{225}Ac -labeled DOTAGA-DBCO was extracted in an amount of 2 μL and separated by thin layer chromatography (iTLC-SG, manufactured by Agilent Technologies, Inc., developing solvent: mixed liquid of water for injection and acetonitrile (volume ratio 1 : 1)). The developed thin layer chromatogram was introduced into a TLC analyzer (GITA Star, manufactured by M&S Instruments Inc.), and the radioactivity count of the total ^{225}Ac including unreacted ^{225}Ac and the radioactivity count of ^{225}Ac -labeled DOTAGA-DBCO in the reaction mixture were measured. The percentage of the radioactivity count of ^{225}Ac -labeled DOTAGA-DBCO to the total ^{225}Ac radioactivity count was 98.8%.

[0096]

(2) Peptide-Conjugated Trastuzumab

In accordance with the same procedure as in Example 2 (1), peptide-conjugated trastuzumab was synthesized.

[0097]

(3) Production of ^{225}Ac -Trastuzumab Conjugate

To 40 μL of the ^{225}Ac -labeled DOTAGA-DBCO solution obtained above, 0.06 mL of the solution D (containing 6 nmol of the monovalent antibody) was added and

reacted at 37°C for 90 minutes, and the antibody and ^{225}Ac -labeled DOTAGA-DBCO were bound to obtain an ^{225}Ac -trastuzumab conjugate. The obtained reaction mixture of the ^{225}Ac -trastuzumab conjugate was extracted in an amount of 2 μL and separated by thin layer chromatography (iTLC-SG, manufactured by Agilent Technologies, Inc., developing solvent: mixed liquid of EDTA aqueous solution (0.1 mol/L) and acetonitrile (volume ratio 1 : 1)). The developed thin layer chromatogram was introduced into a TLC analyzer (GITA Star, manufactured by M&S Instruments Inc.), and the radioactivity count of the total ^{225}Ac including unreacted ^{225}Ac and the radioactivity count of the ^{225}Ac -trastuzumab conjugate in the reaction mixture were measured. The percentage of the radioactivity count of the ^{225}Ac -trastuzumab conjugate to the total ^{225}Ac radioactivity count was 67.2%.

[0098]

In all of Examples 1 to 6 in which antibody labeling was performed in one step, a high labeling rate was exhibited. Meanwhile, in Comparative Examples 1 to 4 in which labeling was performed in two steps, the labeling rate was low. Thus, it has been shown that the production method with one step is superior to the production method with two steps.

[0099]

[Comparative Example 5] Production of ^{225}Ac -Trastuzumab Conjugate with One-Step Method (Condition Setting with Reference to US 2015/0157742 A)

(1) Ligand-Conjugated Trastuzumab

In accordance with the same procedure as in Examples 2 (1) and 2 (2), ligand-conjugated trastuzumab was synthesized. The synthesized ligand-conjugated trastuzumab was added into an ultrafiltration filter (Amicon Ultra-15, manufactured by Merck KGaA) into which 10 mL of a 0.15 mol/L sodium chloride-50 mmol/L sodium

acetate buffer solution (pH: 7.2) was added in advance, and the solution was centrifuged with a centrifuge (centrifugation conditions: 25°C, 3000 x g, 20 min). After the centrifugation, the filtrate was discarded, 13.5 mL of a 0.15 mol/L sodium chloride-50 mmol/L sodium acetate buffer solution (pH: 7.2) was added again, and centrifugation was performed under the same conditions. This centrifugation was performed 4 times in total, and finally, the centrifuged residue was diluted with a 0.15 mol/L sodium chloride-50 mmol/L sodium acetate buffer solution (pH: 7.2) to obtain a ligand-conjugated antibody solution having a monovalent antibody concentration of 5 mg/mL (hereinafter, sometimes referred to as “solution F”).

[0100]

(2) Production of ^{225}Ac -Trastuzumab Conjugate

With Reference to the conditions disclosed in the prior art documents, an ^{225}Ac -trastuzumab conjugate was produced using the ligand-conjugated trastuzumab of (1) as follows.

Into a vial containing $^{225}\text{Ac}(\text{NO}_3)_3$ purchased from Oak Ridge National Laboratory, 0.2 mol/L hydrochloric acid was added, pipetting was performed to wash the vial well mainly at the bottom surface, and the liquid was dispensed into a microtube so that the radioactivity was 0.288 MBq (the liquid amount was 5 μL). Into the microtube, 5 μL of water for injection and 500 μL of a 100 mmol/L Tris buffer solution (pH: 9.0) were added to obtain 510 μL of an $^{225}\text{Ac}^{3+}$ ion solution having a pH of 9.0 (radioactivity concentration in the reaction solution: 0.56 MBq/mL). The pH was measured with a pH test paper.

To the $^{225}\text{Ac}^{3+}$ ion solution, 20 μL of the solution F was added, the mixture was heated at 40°C for 15 minutes to synthesize an ^{225}Ac -trastuzumab conjugate, and the reaction mixture was analyzed by a thin layer chromatography method in the same

manner as in Example 3 (2). Thus, the percentage of the radioactivity count of the ^{225}Ac -trastuzumab conjugate to the total ^{225}Ac radioactivity count was measured. The result was 10.0%.

[0101]

[Comparative Example 6] Production of ^{225}Ac -Trastuzumab Conjugate with One-Step Method (US 2012/0220754 A)

(1) Ligand-Conjugated Trastuzumab

In accordance with the same procedure as in Examples 2 (1) and 2 (2), ligand-conjugated trastuzumab was synthesized. The synthesized ligand-conjugated trastuzumab was added into an ultrafiltration filter (Amicon Ultra-15, manufactured by Merck KGaA) into which 10 mL of physiological saline was added in advance, and the solution was centrifuged with a centrifuge (centrifugation conditions: 25°C, 3000 x g, 20 min). After the centrifugation, the filtrate was discarded, 13.5 mL of physiological saline was added again, and centrifugation was performed under the same conditions. This centrifugation was performed 4 times in total, and finally, the centrifuged residue was diluted with physiological saline to obtain a ligand-conjugated antibody solution having a monovalent antibody concentration of 10 mg/mL (hereinafter, sometimes referred to as “solution G”).

[0102]

(2) Production of ^{225}Ac -Trastuzumab Conjugate

With Reference to the conditions disclosed in the prior art documents, an ^{225}Ac -trastuzumab conjugate was produced using the ligand-conjugated trastuzumab of (1) as follows.

Into a vial containing $^{225}\text{Ac}(\text{NO}_3)_3$ purchased from Oak Ridge National Laboratory, 0.2 mol/L hydrochloric acid was added, pipetting was performed to wash

the vial well mainly at the bottom surface, and the liquid was dispensed into a microtube so that the radioactivity was 0.272 MBq (the liquid amount was 75 μL). Into the microtube, 225 μL of water for injection, 100 μL of a 3 mol/L sodium acetate buffer solution, and 20 μL of a saturated gentisic acid solution were added to obtain 420 μL of an $^{225}\text{Ac}^{3+}$ ion solution (radioactivity concentration in the reaction solution: 0.65 MBq/mL). The pH was measured with a pH test paper and confirmed to be in the range of 5.5 to 7.0.

To the $^{225}\text{Ac}^{3+}$ ion solution, 100 μL of the solution G was added, the mixture was heated at 37°C for 90 minutes to synthesize an ^{225}Ac -trastuzumab conjugate, and the reaction mixture was analyzed by a thin layer chromatography method in the same manner as in Example 3 (2). Thus, the percentage of the radioactivity count of the ^{225}Ac -trastuzumab conjugate to the total ^{225}Ac radioactivity count was measured. The result was 17.9%.

[0103]

In all of Examples 3 to 6 in which ^{225}Ac labeling was performed in one step, a high labeling rate was exhibited. Meanwhile, in Comparative Examples 5 to 6 in which labeling was performed under the conditions set on the basis of the conditions in the prior art documents, the labeling rate was low. Thus, it has been shown that the production method with one step of the present invention is superior to the production method according to the prior art documents.

[0104]

[Example 7] Production of ^{89}Zr -Trastuzumab Conjugate under Aggregation Inhibitor-Added Condition

(1) Synthesis of Ligand-Conjugated Trastuzumab

In accordance with the same procedure as in Examples 2 (1) and 2 (2), ligand-

conjugated trastuzumab was synthesized to obtain a solution E.

[0105]

(2) Labeling Reaction

^{89}Zr obtained by a nuclear reaction of $^{89}\text{Y}(\text{p}, \text{n})^{89}\text{Zr}$ and prepared into a $^{89}\text{Zr}^{4+}$ ion aqueous solution (manufactured by Nihon Medi-Physics Co., Ltd.) was added into a vial, and the solvent was distilled off.

[0106]

Into the vial, 814 μL of 0.1 mol/L hydrochloric acid was added, pipetting was performed to wash the vial well mainly at the bottom surface, and a solution containing $^{89}\text{Zr}^{4+}$ ions at 1000 MBq/mL was obtained. Into a microtube, 20 μL of this solution was dispensed, 40 μL of 0.1 mol/L hydrochloric acid, 300 μL of an aggregation inhibitor that is water for injection or an aqueous solution of sorbitol, xylose, glucose, trehalose, or sucrose adjusted to 300 mmol/L, and 210 μL of water for injection were added, and then a 1 mol/L sodium hydroxide aqueous solution was added to obtain 570 μL of a $^{89}\text{Zr}^{4+}$ ion solution having a pH of 3.0 (radioactivity concentration in the reaction solution: 35 MBq/mL). The pH was measured with a pH test paper.

[0107]

To the $^{89}\text{Zr}^{4+}$ ion solution, 30 μL of the solution E (containing 3 nmol of the monovalent ligand-conjugated trastuzumab) was added to set the concentration of the monovalent ligand-conjugated trastuzumab in the reaction solution to 5 $\mu\text{mol/L}$, and the reaction solution was stirred with a vortex mixer. Next, the reaction solution was heated at 70°C for 30 minutes using a block heater to coordinate $^{89}\text{Zr}^{4+}$ ions to the ligand of the ligand-conjugated trastuzumab, and thus a ^{89}Zr -trastuzumab conjugate was synthesized.

[0108]

(Evaluation)

In size exclusion chromatography (SEC) analysis, a chromatogram was acquired using an RI detector, and the labeling rate was evaluated. Furthermore, in the SEC analysis, a chromatogram was acquired using a UV detector, and the proportion of aggregates was evaluated to verify the effect of stabilizing the antibody by the aggregation inhibitor. The analysis was performed under the following conditions using an e2695 separation module manufactured by Waters Corporation as a liquid chromatography apparatus, a 2489 UV/Vis detector manufactured by Waters Corporation as a UV detector, and a Gabi Star detector manufactured by Raytest as an RI detector. The area percentage of the peak area value of the ^{89}Zr trastuzumab conjugate to the total peak area value in the chromatogram obtained using an RI detector was regarded as the labeling rate. In the chromatogram obtained using a UV detector, a peak detected other than the peak of the ^{89}Zr trastuzumab conjugate (hereinafter, sometimes referred to as the “main peak”) was regarded as a peak of an aggregate.

[0109]

Table 1 shows the labeling rates of each component and Table 2 shows the proportions of each component after the end of production (0-day point) and after storage for 14 days. Because of the lower concentration of the ligand-conjugated antibody, the labeling rate was lower than in Example 2, but in all cases, the labeling rate was higher than 50%. Thus, even in the case of adding an aggregation inhibitor, the labeling rate was not lower by 20% or more than in the case of not adding an aggregation inhibitor. After the storage for 14 days, formation of aggregates was suppressed in the case of adding an aggregation inhibitor as compared with the case of not adding an aggregation inhibitor.

[0110]

[HPLC conditions]

Column: TOSOH TSKgel guardcolumn SWXL (6 mm × 4 cm), TOSOH TSKgel G3000SWXL (5 μm, 7.8 x 30 cm)

Column temperature: constant temperature around 25°C

Mobile phase: 0.2 mol/L arginine hydrochloride-containing 0.1 mol/L sodium dihydrogen phosphate buffer solution (pH: 6.8)

Flow rate: 1.0 mL per minute

Area measurement range: 15 minutes

Detection wavelength (UV): 280 nm

Detection range (RI): 50 to 803 keV

[0111]

[Table 1]

Aggregation inhibitor	Measurement date	Labeling rate (%)
-	0-Day point	71.5
	14-Day point	63.6
Sorbitol	0-Day point	52.5
	14-Day point	57.4
Xylose	0-Day point	73.7
	14-Day point	71.4
Glucose	0-Day point	54.3
	14-Day point	65.1
Trehalose	0-Day point	81.7
	14-Day point	74.6
Sucrose	0-Day point	85.3
	14-Day point	72.7

[0112]

[Table 2]

Aggregation inhibitor	Measurement date	Proportion of main peak (%)	Proportion of peak of aggregates (%)
-	0-Day point	97.1	2.9
	14-Day point	89.0	11.0
Sorbitol	0-Day point	97.1	2.9
	14-Day point	94.5	5.5
Xylose	0-Day point	95.7	4.3
	14-Day point	95.6	4.4
Glucose	0-Day point	96.2	3.8
	14-Day point	95.6	4.4
Trehalose	0-Day point	96.3	3.7
	14-Day point	98.3	1.7
Sucrose	0-Day point	95.0	5.0
	14-Day point	92.4	7.6

[0113]

[Evaluation of Antigen-Binding Activity]

The antigen-binding activity was confirmed by in vitro autoradiography (ARG). Five $\times 10^6$ cells of SK-OV-3 cells, which are HER2-positive human ovarian cancer cell lines, and 1×10^7 cells of MDA-MB-231 cells, which are HER2-negative human breast cancer cell lines, purchased from American Type Culture Collection (ATCC) were administered subcutaneously in the flank of female BALB/c nu/nu mice (available from Charles River Laboratories Japan, Inc.) to prepare tumor-bearing mice. Then, the SK-OV-3 tumor and the MDA-MB-231 tumor were extracted and embedded in Tissue-Tek O.C.T. Compound (manufactured by Sakura Finetek Japan Co., Ltd.) to prepare frozen sections. The radioconjugates obtained in Example 1 and Comparative Example 1 were each added to 1% bovine serum albumin-containing PBS so that the

radioactivity concentration was 5 kBq/mL, and the SK-OV-3 tumor section and the MDA-MB-231 tumor section were immersed. Each section was contacted with an imaging plate and then read by a scanner type image analyzer (Typhoon FLA 7000, manufactured by GE Healthcare Japan), and the amount of radioactivity bound to the section was evaluated. Fig. 1 shows the results. The vertical axis of the graph represents a value obtained as follows. The count value in an ROI set in the used tumor section was divided by the area of the ROI, and the quotient was corrected by a value obtained by dividing the count value of a standard radiation source by the area of the ROI. The resulting value was represented by the vertical axis. The horizontal axis represents the cell type of the tumor section used for evaluation. The graph shows the average $\pm$ standard deviation in the ROIs set in each sample. In the radioconjugates produced in accordance with the descriptions of Example 1 and Comparative Example 1, the binding activity to HER2 was confirmed. The radioconjugates produced in accordance with Example 1 and Comparative Example 1 bound only to SK-OV-3 tumor sections, and the HER2-selective binding activity was confirmed.

[0114]

The present application is based on Japanese Patent Application No. 2022-057723 filed in Japan (filing date: March 30, 2022), the contents of which are all incorporated herein.

CLAIMS

[Claim 1]

A method for producing a conjugate of a radioactive metal nuclide and an antibody, the method comprising:

a step of reacting a radioactive metal nuclide with an antibody conjugated with a ligand compound in a reaction solution containing water to coordinate the radioactive metal nuclide to the ligand compound, wherein

the reaction solution does not contain a pH buffer and has a pH of 2 or more and 10 or less.

[Claim 2]

The method according to claim 1, wherein the radioactive metal nuclide is at least one selected from ^{68}Ga , ^{89}Zr , ^{90}Y , ^{111}In , ^{177}Lu , and ^{225}Ac .

[Claim 3]

The method according to claim 2, wherein the radioactive metal nuclide is ^{89}Zr , and the reaction solution has a pH of 2 or more and 4 or less.

[Claim 4]

The method according to claim 2, wherein the radioactive metal nuclide is ^{225}Ac , and the reaction solution has a pH of 5 or more and 10 or less.

[Claim 5]

The method according to claim 1 or 2, wherein a concentration of the antibody in the reaction solution is 1 $\mu\text{mol/L}$ or more and 2000 $\mu\text{mol/L}$ or less.

[Claim 6]

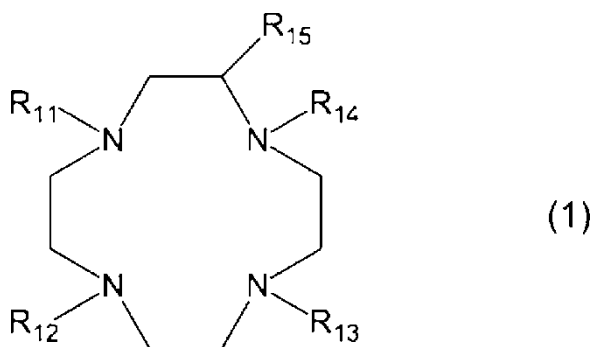
The method according to claim 1 or 2, wherein the reaction solution is heated to 30°C or higher and 80°C or lower to coordinate the radioactive metal nuclide to the ligand compound.

[Claim 7]

The method according to claim 1 or 2, wherein the reaction solution contains at least one aggregation inhibitor selected from the group of saccharides, the group consisting of sucrose, trehalose, mannitol, maltose, lactose, arabinose, xylose, sorbitol, fructose, glucose, mannose, melezitose, and nystose.

[Claim 8]

The method according to claim 1 or 2, wherein the ligand compound has Formula (1) described below:



wherein R_{11} , R_{13} , and R_{14} each independently represent a group including $-(CH_2)_pCOOH$, $-(CH_2)_pC_5H_4N$, $-(CH_2)_pPO_3H_2$, $-(CH_2)_pCONH_2$, or $-(CHCOOH)(CH_2)_pCOOH$, one of R_{12} or R_{15} represents a hydrogen atom, a carboxyl group, or a carboxyalkyl group having 2 or 3 carbon atoms, another of R_{12} or R_{15} represents a substituent that is to be conjugated with the antibody, p is an integer of 0 or

more and 3 or less, R_{15} is a hydrogen atom when R_{12} is a substituent that is to be conjugated with the antibody, and R_{15} is a substituent that is to be conjugated with the antibody when R_{12} is not a substituent that is to be conjugated with the antibody.

[Claim 9]

The method according to claim 1 or 2, wherein an Fc region of the antibody is site-specifically conjugated with the ligand compound via a linker.

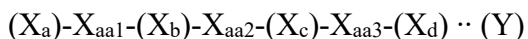
[Claim 10]

The method according to claim 9, wherein

the antibody is a human IgG antibody, and

the linker includes a peptide including 13 or more and 17 or less amino acid residues and is formed by a crosslinking reaction between the peptide conjugated with a crosslinking agent and the antibody,

the peptide having Formula (Y) below:



wherein X_a , X_b , X_c , and X_d represent consecutive Xs of which number is a, consecutive Xs of which number is b, consecutive Xs of which number is c, and consecutive Xs of which number is d, respectively, X represents an amino acid residue having a side chain having neither a thiol group nor a haloacetyl group, a, b, c, and d are each independently an integer of 1 or more and 5 or less and satisfy $a + b + c + d \leq 14$, X_{aa1} and X_{aa3} each independently represent an amino acid residue derived from an amino acid having a side chain having a thiol group, or one of X_{aa1} or X_{aa3} represents an amino acid residue derived from an amino acid having a side chain having a thiol group and another of X_{aa1} or X_{aa3} represents an amino acid residue derived from an amino acid

having a side chain having a haloacetyl group, X_{aa1} and X_{aa3} are linked to each other, and X_{aa2} is a lysine residue, an arginine residue, a cysteine residue, an aspartic acid residue, a glutamic acid residue, a 2-aminosuberic acid residue, or a diaminopropionic acid residue, and is modified with the crosslinking agent.