DESIGNER OSTEOGENIC PROTEINS

FIELD OF THE INVENTION

This application relates to the field of osteogenic proteins, methods of making improved osteogenic proteins, and methods of treating patients with osteogenic proteins.

BACKGROUND OF THE INVENTION

The cystine knot cytokine superfamily is divided into subfamilies, which include, the transforming growth factor β (TGFβ) proteins, the glycoprotein hormones, the platelet-derived growth factor-like (PDGF-like) proteins, nerve growth factors (NGF), and the differential screening-selected gene aberrative in neuroblastoma (DAN) family (e.g., cerberus). In turn, the TGFβ superfamily comprises approximately 43 members, subdivided into three subfamilies: the TGFβs, the activins and the bone morphogenetic/growth differentiation factor proteins (BMP/GDF).

The TGF-β superfamily members contain the canonical cystine knot topology. That is, cystine knots are the result of an unusual arrangement of six cysteine residues. The knot consists of bonds between cysteines 1-4, cysteines 2-5, and the intervening sequence forming a ring, through which the disulfide bond between cysteines 3-6 passes. The active forms of these proteins are homodimers or heterodimers. In each case the monomer topology is stabilized by the cysteine knot and additional cysteines contribute to additional intrachain bonds and/or mediate dimerization with another protein unit. See Kingsley, 1994, Genes Dev. 8:133-146; Lander et al, 2001, Nature 409:860-921.

BMP/GDFs are the most numerous members of the TGF-β protein superfamily. The BMP/GDF subfamily includes, but is not limited to, BMP2, BMP3 (osteogenin), BMP3b (GDF-10), BMP4 (BMP2b), BMP5, BMP6, BMP7 (osteogenic protein-1 or OP1), BMP8 (OP2), BMP8B (OP3), BMP9 (GDF2), BMP10, BMP11 (GDF11), BMP12 (GDF7), BMP13 (GDF6, CDMP2), BMP15 (GDF9), BMP16, GDF1, GDF3, GDF5 (CDMP1; MP52), and GDF8 (myostatin). BMPs are sometimes referred to as Osteogenic Protein (OPs), Growth Differentiation Factors (GDFs), or Cartilage-Derived Morphogenetic Proteins (CDMPs). BMPs are also present in other animal species. Furthermore, there is some allelic variation in BMP sequences among different members of the human population.

BMPs are naturally expressed as pro-proteins comprising a long pro-domain, one or more cleavage sites, and a mature domain. This pro-protein is then processed by the cellular machinery to yield a dimeric mature BMP molecule. The pro-domain is believed to aid in the correct folding and processing of BMPs. Furthermore, in some but not all BMPs, the pro-domain may noncovalently bind the mature domain and may act as a chaperone, as well as an inhibitor (e.g., Thies et al., Growth Factors 18:251-9 (2001)).

BMP signal transduction is initiated when a BMP dimer binds two type I and two type II serine/threonine kinase receptors. Type I receptors include, but are not limited to, ALK-1 (Activin receptor-Like Kinase 1), ALK-2 (also called ActRIa or ActRI), ALK-3 (also called BMPRIa), and ALK-6

(also called BMPRIb). Type II receptors include, but are not limited to, ActRIIa (also called ActRII), ActRIIb, and BMPRII. The human genome contains 12 members of the receptor serine/threonine kinase family, including 7 type I and 5 type II receptors, all of which are involved in TGF-β signaling (Manning et al., Science 298:1912-34 (2002)), the disclosures of which are hereby incorporated by reference). Thus, there are 12 receptors and 43 superfamily members, suggesting that at least some TGF-β superfamily members bind the same receptor(s). Following BMP binding, the type II receptors phosphorylate the type I receptors, the type I receptors phosphorylate members of the Smad family of transcription factors, and the Smads translocate to the nucleus and activate the expression of a number of genes.

BMPs are among the most numerous members of TGF-β superfamily, and control a diverse set of cellular and developmental processes, such as embryonic pattern formation and tissue specification as well as promoting wound healing and repair processes in adult tissues. BMPs were initially isolated by their ability to induce bone and cartilage formation. BMP signaling is inducible upon bone fracture and related tissue injury, leading to bone regeneration and repair. BMP molecules which have altered affinity for their receptors would have improved biological activity relative to the native proteins. Such BMPs include proteins with increased in vivo activity and may provide potential improved therapeutics for, among other things, tissue regeneration, repair, and the like, by providing greater or altered activity at lower protein levels thereby providing improved protein therapeutics.

SUMMARY OF THE INVENTION

The invention includes a designer BMP protein comprising at least one mutation in at least one type I or type II receptor binding domain, wherein the mutation confers altered binding to the type I or type II BMP receptor compared with the binding to the type I or type II receptor by a corresponding wild type BMP.

In one aspect, the protein is selected from the group consisting of BMP2, BMP4, BMP5, BMP6, BMP7, BMP8 and BMP9.

In another aspect, the protein comprises at least one mutation within: the type II binding domain A; the type II binding domain B; the type I binding domain; and any combination thereof.

The invention also includes a designer osteogenic protein comprising an amino acid sequence comprising at least one mutation in at least one type I or type II receptor binding domain, wherein the mutation confers altered binding to the type I or type II BMP receptor compared with the binding to the type I or type II receptor by wild type BMP2.

In one aspect, the mutation is a mutation within the type II binding domain A wherein said mutation is at least one mutation selected from the group consisting of a mutation at V33, P36, H39, and F41 with respect to the sequence of SEQ ID NO:1.

In another aspect, the is a mutation within the type II binding domain A wherein said mutation is at least one mutation selected from the group consisting of V33I, P36K, P36R, H39A, and F41N with respect to SEQ ID NO:1.

In yet another aspect, the mutation is a mutation within the type II binding domain B wherein said mutation is at least one mutation selected from the group consisting of a mutation at E83, S85, M89, L92, E94, E96, K97, and V99 with respect to the sequence of SEQ ID NO:1.

In a further aspect, the mutation is a mutation within the type II binding domain B wherein said mutation is at least one mutation selected from the group consisting of E83K, S85N, M89V, L92F, E94D, E96S, K97N, and V99I with respect to of SEQ ID NO:1.

In another aspect, the mutation is a mutation within the type I binding domain wherein said mutation is at least one mutation selected from the group consisting of a mutation at H44, P48, A52, D53, L55, S57, N68, S69, V70, an insertion of a single amino acid after N71, S72, K73, I74, A77, and V80 with respect to the sequence of SEQ ID NO:1.

In yet another aspect, the mutation is a mutation within the type I binding domain wherein said mutation is at least one mutation selected from the group consisting of H44D, P48S, A52N, D53A, L55M, S57A, N68H, S69L, V70M, insertion of P after N71, S72E, K73Y, I74V, A77P, and V80A with respect to the sequence of SEQ ID NO:1.

In a further aspect, the protein comprises a mutation at each of amino acids H44, P48, A52, D53, L55, S57, N68, S69, V70, insertion of a single amino acid after N71, S72, K73, I74, A77, and V80 with respect to the sequence of SEQ ID NO:1.

In another aspect, the protein comprises a mutation at each of amino acids H44, P48, A52, D53, L55, S57, N68, S69, V70, insertion of a single amino acid after N71, S72, K73, I74, A77, and V80 with respect to the sequence of SEQ ID NO:1 wherein the mutations are H44D, P48S, A52N, D53A, L55M, S57A, N68H, S69L, V70M, insertion of a P after N71, S72E, K73Y, I74V, A77P, and V80A.

In yet another aspect, the protein comprises a mutation at each of amino acids V33, P36, H39, S85, M89, L92, E94, E96, K97, and V99 with respect to the sequence of SEQ ID NO:1.

In another aspect, the protein comprises a mutation at each of amino acids V33, P36, H39, S85, M89, L92, E94, E96, K97, and V99 with respect to the sequence of SEQ ID NO:1, wherein the mutations are V33I, P36K, H39A, S85N, M89, L92F, E94D, E96S, K97N, and V99I.

In a further aspect, the protein comprises a mutation at each of amino acids V33, P36, H39, H44, P48, A52, D53, L55, S57, N68, S69, V70, insertion of a single amino acid after N71, S72, K73, I74, A77, and V80, S85, M89, L92, E94, E96, K97, and V99 with respect to the sequence of SEQ ID NO:1.

In yet another aspect, the protein comprises a mutation at each of amino acids V33, P36, H39, H44, P48, A52, D53, L55, S57, N68, S69, V70, insertion of a single amino acid after N71, S72, K73, I74, A77, and V80, S85, M89, L92, E94, E96, K97, and V99 with respect to the sequence of SEQ ID NO:1 wherein the mutations are V33I, P36K, H39A, H44D, P48S, A52N, D53A, L55M, S57A, N68H, S69L, V70M, insertion of a P after N71, S72E, K73Y, I74V, A77P, and V80A, S85N, M89, L92F, E94D, E96S, K97N, and V99I.

In yet another aspect, the protein comprises a mutation at each of amino acids V33, P36, H39, H44, P48, A52, D53, L55, S57, N68, S69, V70, insertion of a single amino acid after N71, S72, K73, I74,

A77, and V80, S85, M89, L92, E94, E96, K97, and V99 with respect to the sequence of SEQ ID NO:1 wherein the mutations are V33I, P36R, H39A, H44D, P48S, A52N, D53A, L55M, S57A, N68H, S69L, V70M, insertion of a P after N71, S72E, K73Y, I74V, A77P, and V80A, S85N, M89, L92F, E94D, E96S, K97N, and V99I.

In another aspect, the protein binds: the ALK2 receptor with a K_D not greater than about 2 nM; the ALK3 receptor with a K_D not greater than about 2 nM; the ALK6 receptor with a K_D not greater than about 1 nM; the ActRIIA receptor with a K_D not greater than about 2 nM; the ActRIIB receptor with a K_D not greater than about 0.5 nM; and the BMPRIIA receptor with a K_D not greater than about 3.5 nM.

In one aspect, the protein further comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid mutations not located within the type I or the type II binding regions.

The invention includes a designer osteogenic protein comprising the amino acid sequence of any one of SEQ ID NOs:8-73.

The invention includes a designer osteogenic protein comprising the amino acid sequence of SEQ ID NO:12.

The invention includes a designer osteogenic protein comprising the amino acid sequence of SEQ ID NO:14.

The invention includes a designer osteogenic protein comprising the amino acid sequence of SEQ ID NO:36.

The invention includes a designer osteogenic protein comprising the amino acid sequence of SEQ ID NO:37.

The invention includes method of producing a designer BMP protein comprising at least one mutation in at least one type I or type II receptor binding domain, wherein the mutation confers altered binding to the type I or type II BMP receptor compared with the binding to the type I or type II receptor by a corresponding wild type BMP. The method comprises introducing a nucleic acid encoding the protein into a host cell, culturing the cell under conditions where the protein is produced, and purifying the protein.

In one aspect, the nucleic acid comprises a sequence selected from the nucleic acid sequence of any one of SEQ ID NOs:74-139.

The invention includes a designer BMP6 protein comprising an amino acid sequence comprising at least one mutation in at least one type I or type II receptor binding domain, wherein the mutation confers altered binding to the type I or type II BMP receptor compared with the binding to the type I or type II receptor by wild type BMP6.

In one aspect, the mutation is a mutation within the type II binding domain A wherein said mutation is at least one mutation selected from the group consisting of a mutation at I57, K60, G61, A63, N65, Y66, and D68 with respect to the sequence of SEQ ID NO:4.

In another aspect, the mutation is a mutation within the type II binding domain B wherein said mutation is at least one mutation selected from the group consisting of K108, N110, A111, V114, F117, D119, N120, S121, N122, V123, and I124 with respect to the sequence of SEQ ID NO:4.

In yet another aspect, the mutation is a mutation within the type I binding domain wherein said mutation is at least one mutation selected from the group consisting of a mutation at S72, N76, A77, H78, M79, N80, A81, N83, V87, T89, H92, L93, M94, N95, P96, E97, Y98, V99, and P100 with respect to the sequence of SEQ ID NO:4.

In another aspect, the mutation is a mutation at each of amino acid residues I57, K60, G61, A63, N65, Y66, and D68 with respect to the sequence of SEQ ID NO:4.

In a further aspect, the mutation is a mutation at each of amino acid residues K108, N110, A111, V114, F117, D119, N120, S121, N122, V123, and I124 with respect to the amino acid sequence of SEQ ID NO:4.

In yet another aspect, the mutation is a mutation at each of amino acid residues S72, N76, A77, H78, M79, N80, A81, N83, V87, T89, H92, L93, M94, N95, P96, E97, Y98, V99, or P100 with respect to the amino acid sequence of SEQ ID NO:4.

In another aspect, the designer BMP6 protein comprising an amino acid sequence comprising at least one mutation in at least one type I or type II receptor binding domain, wherein the mutation confers altered binding to the type I or type II BMP receptor compared with the binding to the type I or type II receptor by wild type BMP6 further comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid mutations not located within the type I or the type II binding domains.

The invention includes an isolated nucleic acid molecule comprising a nucleotide sequence encoding an amino acid sequence selected from the group consisting of the sequence of SEQ ID NOs:8 to 73.

In one aspect, the nucleic acid encodes a protein comprising an amino acid sequence selected from the group consisting of the sequence of SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:36 and SEQ ID NO:37.$$

The invention includes an isolated nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of SEQ ID NOs:74 to 139.

In one aspect, the nucleic acid comprises a nucleotide sequence selected from the group consisting of the sequence of SEQ ID NO:78, SEQ ID NO:80, SEQ ID NO:102, and SEQ ID NO:103.

The invention includes a method of producing the designer BMP6 protein comprising an amino acid sequence comprising at least one mutation in at least one type I or type II receptor binding domain, wherein the mutation confers altered binding to the type I or type II BMP receptor compared with the binding to the type I or type II receptor by wild type BMP6. The method comprises introducing a nucleic acid encoding said protein into a host cell, culturing said cell under conditions where said protein is produced, and purifying said protein.

The invention includes a method of treating a bone disease associated with bone loss in a patient in need thereof. The method comprises administering a therapeutically effective amount of a designer BMP protein comprising at least one mutation in at least one type I or type II receptor binding domain, wherein the mutation confers altered binding to the type I or type II BMP receptor compared with the binding to the type I or type II receptor by a corresponding wild type BMP protein to the patient, thereby treating bone disease in the patient.

The invention includes a method of treating fibrosis in a patient in need thereof. The method comprises administering a therapeutically effective amount of a designer BMP protein comprising at least one mutation in at least one type I or type II receptor binding domain, wherein the mutation confers altered binding to the type I or type II BMP receptor compared with the binding to the type I or type II receptor by a corresponding wild type BMP to the patient, thereby treating fibrosis.

The invention includes a method of inducing bone formation in a tissue. The method comprises contacting the tissue with a designer BMP protein comprising at least one mutation in at least one type I or type II receptor binding domain, wherein the mutation confers altered binding to the type I or type II BMP receptor compared with the binding to the type I or type II receptor by a corresponding wild type BMP, thereby inducing bone formation in said tissue.

BRIEF DESCRIPTION OF THE DRAWINGS

For the purpose of illustrating the invention, there are depicted in the drawings certain embodiments of the invention. However, the invention is not limited to the precise arrangements and instrumentalities of the embodiments depicted in the drawings.

Figure 1, comprising panels A-C, is a diagram showing the alignment of various wild type and designer BMP amino acid sequences and indicating (by being within a box) the regions of these proteins potentially involved in type I and type II receptor interactions. Figure 1A shows the amino acid sequence alignment of wild type BMP2, BMP4, BMP5, BMP6, BMP7, BMP8 and BMP9. Figure 1B shows the amino acid sequence alignment of various designer BMPs where the corresponding wild type BMP is BMP2. Figure 1C shows the amino acid sequence alignment of various designer BMP6 molecules where the corresponding wild type BMP is BMP6.

Figure 2 is an illustration of a structural model showing a wild type BMP2 homodimer binding to two type I and two type II BMP receptors.

Figure 3, comprising panels A and B, is an diagram of a structural model showing the position of the histidine doorstop (H54) in human BMP2 produced in Chinese Hamster Ovary (CHO) (Figure 3A) and E. coli cells (Figure 3B).

Figure 4, comprising panels A and B, is a diagram illustrating the location of the glycan tether and potential histidine (His) doorstop. Figure 4A shows the glycan tether (N-linked glycan at N56) and histidine 54, in the non-doorstop orientation, as well as the interaction of the glycan tether with R16 all in CHO-produced BMP2. Figure 4B shows the glycan tether (N-linked glycan at N80) and the histidine in

the non-doorstop configuration at H78 in BMP6, as well as the R39 corresponding to R16 in BMP2. The sequence alignment of BMP2 (11-KSSCKRHP) and BMP6 (35-KTACRKHE) showing the corresponding amino acids between BMP2 and BMP6 is shown along the top of the figure.

Figure 5, comprising panels A-D, shows various steps in the process for purification of BMPs and designer BMPs. Figure 5A shows a chromatogram showing gradient elution of BMPs using a cellufine sulfate column. Figure 5B is an image of a Coomassie stained SDS-PAGE (non-reduced on the left and reduced on the right side) gel containing samples of fractions from the cellufine sulfate column step. Figure 5C shows a chromatogram showing the profile from preparative reversed phase purification step. Figure 5D is an image of a Coomassie stained SDS-PAGE (non-reduced on the left and reduced on the right) gel of BMP containing samples of the fractions obtained by the preparative reversed phase purification step.

Figure 6, comprising panels A-D, show images of Coomassie-stained SDS-PAGE protein gels showing purified BMP2 wild type and various mutants as indicated along the top of each gel image. The gels were run under either non-reducing (Figures 6A and 6B) and reducing (Figures 6C and 6D) conditions.

Figure 7 shows alkaline phosphatase assay results in C2C12 pre-myoblasts comparing the osteogenic activity of wild type BMP2 and BMP2/6 heterodimer to the various designer BMPs as indicated in the graph legend.

Figure 8 shows the results of a C2C12 BMP-Response Element luciferase (BRE-luciferase) assay indicative of Smad activity showing stronger signaling by BMPE compared to BMP2 and equivalent signaling to BMP2/6.

Figure 9, comprising panels A and B, shows the ectopic bone formation mediated by various BMPs. Figure 9A is a graph showing the amount of ectopic bone (calculated as milligrams of hydroxyapatite; mg HA) as determined by μCT analysis for each limb which was implanted with the indicated BMP (BMP2, BMPE, and BMP2/6) at the dose indicated (0.1 or 0.5 μg). Figure 9B is a graph showing the amount of ectopic bone (calculated as milligrams of hydroxyapatite) as determined by μCT analysis for each limb which was implanted with the indicated BMP (BMP2, BMPG, BMPA, and BMPF) at the dose indicated (0.1 or 0.5 μg). The data presented are from 2 separate experiments.

Figure 10, comprising panels A-D, shows images of radiographs showing the results of a non-human primate (NHP) fibula osteotomy model at 4 and 8 weeks. Radiographs are shown of the fibulas of 7 representative NHPs that received BMPE and BMPG, respectively, at 0.5 mg/ml (250 μg total BMP delivered/limb). Each NHP received WT BMP2 at the same dose in the contralateral limb. Figures 10A and 10B show the radiographs for the NHPs indicated at the top of each diagram showing the effects of BMPE compared with BMP2 wild type at 4 weeks and 8 weeks, respectively. Figures 10C and 10D show the radiographs for the NHPs indicated at the top of each diagram showing the effects of BMPG compared with BMP2 wild type at 4 weeks and 8 weeks, respectively.

Figure 11 is a graph showing the bone volume of the limbs treated with BMP-E versus contralateral limbs treated with BMP-2.

Figure 12 is a graph showing results of an alkaline phosphatase assay in C2C12 pre-myoblasts comparing the osteogenic activity of wild type BMP2 and BMP-GER, BMP-GEP, and BMP2/6 heterodimer.

Figure 13 is a graph showing the amount of ectopic bone (calculated as milligrams of hydroxyapatite) as determined by μCT analysis for each limb which was implanted with the indicated BMP (BMP-2, BMP-2/6, BMP-E, BMP-GER, and BMP-6) at the dose indicated (0.05 or 0.25μg).

Figure 14 is a graph showing the amount of ectopic bone (calculated as milligrams of hydroxyapatite) as determined by μCT analysis for each limb which was implanted with the indicated BMP (BMP-2, BMP-2/6, BMP-E, BMP-GER, and BMP-6) at the dose indicated (0.05 or 0.25μg). These are the results from an experiment separate from that shown in Figure 13.

Figure 15, comprising panels A and B, shows images of radiographs and μCT images showing the results of a non-human primate (NHP) fibula wedge osteotomy model at 5 and 10 weeks. Figure 15A shows images of 5-week radiographs obtained in a NHP fibula wedge osteotomy model. Figure 15A shows images of the fibulas of 4 representative NHPs which received BMP-GER in one limb and WT BMP-2 in the contralateral limb at 0.5 mg/ml (250 μG total BMP delivered/limb) at 5 weeks. Figure 15B shows uCT images of the same limbs at 10 weeks showing the large calluses of the BMP-GER treated limbs compared with the BMP2-treated contralateral limbs for each animal.

Figure 16, comprising panels A-C, shows graphs illustrating the strength (Figure 16A), stiffness (Figure 16B), and callus bone volume (Figure 16C) of the BMP-GER treated limbs versus the BMP-2 treated contralateral limbs.

Figure 17, comprising panels A-C, shows radiographic images of the healing over time of 3 non-human primate's (NHP) fibulas treated with BMP-GER at 0.5 mg/ml and BMP-2 in the contra lateral limb at 1.5mg/ml using a calcium phosphate based cement as a carrier following the wedge defect model. Figure 17A, upper panel, shows results for NHP number 1 left arm treated with 0.5 mg/ml GER as follows: panels 1 and 2 show LAT (lateral) and AP (anterior-posterior) images, respectively, at the initial time point; panels 3 and 4 show LAT and AP images, respectively, at 2 weeks; panels 5 and 6 show LAT and AP images, respectively, at 4 weeks; panels 7 and 8 show LAT and AP images, respectively, at 6 weeks; panels 9 and 10 show LAT and AP images, respectively, at 7 weeks; panels 11 and 12 show LAT and AP images, respectively, at 8 weeks; Figure 17A, lower panel, shows results for NHP number 1 right arm treated with 1.5 mg/ml BMP-2 as follows: panels 1 and 2 show LAT (lateral) and AP (anterior-posterior) images, respectively, at the initial time point; panels 3 and 4 show LAT and AP images, respectively, at 2 weeks; panels 5 and 6 show LAT and AP images, respectively, at 4 weeks; panels 7 and 8 show LAT and AP images, respectively, at 6 weeks; panels 9 and 10 show LAT and AP images, respectively, at 7 weeks; panels 11 and 12 show LAT and AP images, respectively, at 8 weeks; Figure 17B shows the

radiographic the results for NHP number 2 as described for NHP #1 in Figure 17A; and Figure 17C sets out the results for NHP number 3 as described for NHP #1 in Figure 17A.

Figure 18 is a diagram of a structural model showing representations and comparison of the crystal structures BMP-E and BMP-6 WT. The differences in the length of the glycan resolved is highlighted showing that the glycan for BMPE that is resolved is much longer than that for BMP6. This indicates that the BMPE glycan is more conformationally constrained than that of BMP6 such that more of the glycan can be rendered in this model. The histidine doorstop residues for both BMPE and BMP6 are shown in similar non-doorstop configurations. Also, the arginine glycan "tether" stabilizing the BMPE glycan is shown by dotted lines representing the interactions of the arginine with the glycan.

Figure 19 is a closer view of the histidine doorstop and arginine tether of the BMPE and BMP6 comparison shown in Figure 18. This image shows the similar conformation of the H54 histidine residue of BMPE and the equivalent histidine of BMP6 both in the non-doorstop position. The image also shows the R16 tethering (via interactions of the BMPE glycan such that the glycan is more rigid and therefore more is rendered by the model compared to the more "floppy" and less constrained glycan of BMP6 such that less of the BMP6 glycan is visualized in this model. The diagram of this model also shows the similar positioning of asparagine N56 of BMPE showing N-linked attachment of the glycan and the equivalent and similarly positioned asparagine of BMP6. The diagram also illustrates the potential additional glycan tethering interaction of BMPE E110 shown by dotted lines between the amino acid residue and the distal end of the glycan. The differences in the length of the glycan resolved is highlighted showing that less of the darker BMP6 glycan can be resolved compared with the lighter shaded longer glycan rendered for BMPE indicating that the BMPE glycan is more conformationally constrained and thus more is rendered upon structural analysis.

Figure 20 is a graph showing the results of an alkaline phoshatase assay using C2C12 pre-myoblasts comparing the osteogenic activity of BMP-2, BMPE and BMP-6 with their Endo-H treated deglycosylated (Degly.) counterparts.

Figure 21 is a diagram illustrating the structural model of BMPE showing the location of the glycan tether at R16 and illustrating the stabilizing interactions between the arginine (R16) and glutamic acid (E110 corresponding to E109 of BMP2) residues. The diagram shows that R16 and E110 both form multiple hydrogen bonds with the third (β-mannose) and fourth (α-mannose) glycan moieties. The diagram also shows the position of H54 potential "doorstop" and asparagine 56 (N56) which provides the N-linked attachment site of the glycan.

Figure 22 is a graph showing the results of an alkaline phoshatase assay using C2C12 pre-myoblasts comparing the osteogenic activity of BMP-E, with BMP-E-NR, BMP-GER and BMP-GER-NR in the presence of increasing doses of Noggin - a natural inhibitor of BMP-2. The data demonstrate that BMP-GER-NR comprising sequences derived from activin was not inhibited by Noggin even at high concentrations but that BMP-GER was sensitive to Noggin inhibition. Thus, addition of sequences derived from activin caused BMP-GER to become Noggin resistant (NR). These results demonstrate that

at least in this in vitro assay, BMP-GER and BMPE, which are Noggin sensitive, become Noggin resistant (NR) upon replacement of the C-terminal region of the protein with sequences derived from activin.

Figures 23 is a graph showing the bone score as determined by immunohistochemistry (IHC) for rat ectopic implants treated with the indicated BMP at the specified dose. The data show that the bone forming activity of BMP-GER was greatly decreased when the C-terminal sequence of the molecule was replaced with a sequence derived from activin (NR). Thus, the data demonstrate that BMP-GER-NR was much less active than BMP-GER in vivo.

Figures 24 is a graph showing the bone score as determined by immunohistochemistry (IHC) for rat ectopic implants treated with the indicated BMP at the specified dose. The data show that the bone forming activity of BMP-E was greatly decreased, indeed, it was completely abrogated, when the C-terminal sequence of the molecule was replaced with a sequence derived from activin (NR).

DETAILED DESCRIPTION OF THE INVENTION

This invention relates a “designer” bone morphogenetic protein, referred to herein as “designer BMP,” “designer osteogenic protein” and “designer protein.” The designer BMPs of the invention may correspond to the amino acid sequences of wild type unmodified BMP, such as, but not limited to, BMP2, BMP4, BMP5, BMP6, BMP7, BMP8, and BMP9. In particular embodiments, the designer BMPs show altered binding to a type I and/or type II BMP receptor when compared to its corresponding wild type BMP. In further embodiments, the designer BMP may be modified to have altered half-life, immunogenicity, or any pharmacokinetic/pharmacodynamic (PK/PD) parameter when compared to its corresponding BMP.

Definitions

Unless otherwise defined herein, scientific and technical terms used in connection with the present invention shall have the meanings that are commonly understood by those of ordinary skill in the art. Further, unless otherwise required by context, singular terms shall include pluralities and plural terms shall include the singular. Generally, nomenclatures used in connection with, and techniques of, cell and tissue culture, molecular biology, immunology, microbiology, genetics and protein and nucleic acid chemistry and hybridization described herein are those well known and commonly used in the art.

The methods and techniques of the present invention are generally performed according to methods well known in the art and as described in various general and more specific references that are cited and discussed throughout the present specification unless otherwise indicated. Such references include, *e.g.*, Sambrook and Russell, *Molecular Cloning, A Laboratory Approach*, Cold Spring Harbor Press, Cold Spring Harbor, NY (2001), Ausubel et al., *Current Protocols in Molecular Biology*, John Wiley & Sons, NY (2002), and Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1990), which are incorporated herein by reference. Enzymatic reactions and purification techniques are performed according to manufacturer's specifications, as commonly accomplished in the art or as described herein. The nomenclatures used in connection with,

and the laboratory procedures and techniques of, analytical chemistry, synthetic organic chemistry, and medicinal and pharmaceutical chemistry described herein are those well known and commonly used in the art. Standard techniques are used for chemical syntheses, chemical analyses, pharmaceutical preparation, formulation, and delivery, and treatment of patients.

As used herein, each of the following terms has the meaning associated with it in this section.

The articles “a” and “an” are used herein to refer to one or to more than one (*i.e.*, to at least one) of the grammatical object of the article. By way of example, “an element” means one element or more than one element.

In this application, the use of "or" means "and/or" unless stated otherwise.

Conventional notation is used herein to portray polypeptide sequences: the left-hand end of a polypeptide sequence is the amino-terminus; the right-hand end of a polypeptide sequence is the carboxyl-terminus. As used herein, the twenty conventional amino acids and their abbreviations follow conventional usage. See *Immunology--A Synthesis* (2nd Edition, E. S. Golub and D. R. Gren, Eds., Sinauer Associates, Sunderland, Mass. (1991)), which is incorporated herein by reference. As used herein, amino acids are represented by the full name thereof, by the three letter code corresponding thereto, or by the one-letter code corresponding thereto, as indicated as follows:

Full Name	Three-Letter Code	One-Letter Code
Aspartic Acid	Asp	D
Glutamic Acid	Glu	E
Lysine	Lys	K
Arginine	Arg	R
Histidine	His	H
Tyrosine	Tyr	Y
Cysteine	Cys	C
Asparagine	Asn	N
Glutamine	Gln	Q
Serine	Ser	S
Threonine	Thr	T
Glycine	Gly	G
Alanine	Ala	A
Valine	Val	V
Leucine	Leu	L
Isoleucine	Ile	I
Methionine	Met	M
Proline	Pro	P
Phenylalanine	Phe	F
Tryptophan	Trp	W

A "conservative amino acid substitution" is one in which an amino acid residue is substituted by another amino acid residue having a side chain R group with similar chemical properties (e.g., charge or hydrophobicity). In general, a conservative amino acid substitution will not substantially change the functional properties of a protein. In cases where two or more amino acid sequences differ from each other by conservative substitutions, the percent sequence identity or degree of similarity may be adjusted upwards to correct for the conservative nature of the substitution. Means for making this adjustment are well-known to those of skill in the art. See, e.g., Pearson, *Methods Mol. Biol.* 243:307-31 (1994).

Examples of groups of amino acids that have side chains with similar chemical properties include 1) aliphatic side chains: glycine, alanine, valine, leucine, and isoleucine; 2) aliphatic-hydroxyl side chains: serine and threonine; 3) amide-containing side chains: asparagine and glutamine; 4) aromatic side chains: phenylalanine, tyrosine, and tryptophan; 5) basic side chains: lysine, arginine, and histidine; 6) acidic side chains: aspartic acid and glutamic acid; and 7) sulfur-containing side chains: cysteine and methionine. Preferred conservative amino acids substitution groups are: valine-leucine-isoleucine, phenylalanine-tyrosine, lysine-arginine, alanine-valine, glutamate-aspartate, and asparagine-glutamine.

Alternatively, a conservative replacement is any change having a positive value in the PAM250 log-likelihood matrix disclosed in Gonnet et al., *Science* 256:1443-1445 (1992), herein incorporated by reference. A "moderately conservative" replacement is any change having a nonnegative value in the PAM250 log-likelihood matrix.

Preferred amino acid substitutions are those which: (1) reduce susceptibility to proteolysis, (2) reduce susceptibility to oxidation, (3) alter binding affinity for forming protein complexes, and (4) confer or modify other physicochemical or functional properties of such analogs. Analogs comprising substitutions, deletions, and/or insertions can include various muteins of a sequence other than the specified peptide sequence. For example, single or multiple amino acid substitutions (preferably conservative amino acid substitutions) may be made in the specified sequence (preferably in the portion of the polypeptide outside the domain(s) forming intermolecular contacts, *e.g.*, outside of the CDRs or the type I or type II receptor binding sites). A conservative amino acid substitution should not substantially change the structural characteristics of the parent sequence (e.g., a replacement amino acid should not tend to break a helix that occurs in the parent sequence, or disrupt other types of secondary structure that characterizes the parent sequence). Examples of art-recognized polypeptide secondary and tertiary structures are described in *Proteins, Structures and Molecular Principles* (Creighton, Ed., W. H. Freeman and Company, New York (1984)); *Introduction to Protein Structure* (C. Branden and J. Tooze, eds., Garland Publishing, New York, N.Y. (1991)); and Thornton et al., *Nature* 354:105 (1991), which are each incorporated herein by reference.

The terms "polynucleotide", "nucleotide sequence", "nucleic acid", "nucleic acid molecule", "nucleic acid sequence", and "oligonucleotide" refer to a series of nucleotide bases (also called "nucleotides") in DNA and RNA, and mean any chain of two or more nucleotides. The polynucleotides can be chimeric mixtures or derivatives or modified versions thereof, single-stranded or double-stranded. The

oligonucleotide can be modified at the base moiety, sugar moiety, or phosphate backbone, for example, to improve stability of the molecule, its hybridization parameters, etc. A nucleotide sequence typically carries genetic information, including the information used by cellular machinery to make proteins and enzymes. These terms include double- or single-stranded genomic and cDNA, RNA, any synthetic and genetically manipulated polynucleotide, and both sense and antisense polynucleotides. This also includes nucleic acids containing modified bases, for example, thio-uracil, thio-guanine, and fluoro-uracil, or containing carbohydrate, or lipids.

In the context of a nucleotide sequence, the term “substantially identical” is used herein to refer to a first nucleic acid sequence that contains a sufficient or minimum number of nucleotides that are identical to aligned nucleotides in a second nucleic acid sequence such that the first and second nucleotide sequences encode a polypeptide having common functional activity, or encode a common structural polypeptide domain or a common functional polypeptide activity. For example, nucleotide sequences having at least about 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% identity to a reference sequence.

By "designer BMP nucleic acids," and grammatical equivalents herein is meant nucleic acids that encode designer BMPs.

The terms “protein” and “polypeptide” are used interchangeably herein. These terms refer to a sequential chain of amino acids linked together via peptide bonds. The terms include one or more proteins that function as a discrete unit. If a single polypeptide is the discrete functioning unit and does not require permanent or temporary physical association with other polypeptides in order to form the discrete functioning unit, the terms "polypeptide" and "protein" may be used interchangeably. If the discrete functional unit is comprised of multiple polypeptides that physically associate with one another, the term "protein" as used herein refers to the multiple polypeptides that are physically coupled and function together as the discrete unit. A protein to be expressed according to the present invention can be a protein therapeutic. A protein therapeutic is a protein that has a biological effect on a region in the body on which it acts or on a region of the body on which it remotely acts via intermediates. Examples of protein therapeutics are discussed in more detail below.

“Designer BMP,” as the term is used herein, relates to a BMP protein comprising at least one amino acid mutation compared to a corresponding wild type BMP without the mutation, wherein the designer BMP has detectably altered binding for at least a type I receptor and/or at least one type II receptor compared with the binding of the corresponding wild type BMP for the type I and/or type II receptor.

By "corresponding wild type protein" it is meant the wild type version of the designer BMP prior to the introduction of any mutations. For example, if the designer BMP is a designer BMP2, the corresponding wild-type BMP is wild-type BMP2. Thus, in one embodiment, design of a designer BMP can, but need not, begin with a wild type BMP sequence wherein mutations (e.g., amino acid substitutions, deletions and/or insertion) are introduced into the wild type sequence. Therefore, the

designer BMP can correspond with a wild type BMP, and the locations of the mutations can be said, for instance, to correspond with, be relative to and/or be respective with the amino acid sequence of the wild type corresponding or “reference” BMP sequence.

The proteins of the present invention include fragments, derivatives, analogs, or variants of the polypeptides described herein, and any combination thereof. The terms “fragment,” “variant,” “derivative” and “analog” when referring to proteins of the present invention include any proteins which retain at least some of the functional properties of the protein from which it was derived.

By the term “fragment" as used herein refers to a polypeptide and is defined as any discrete portion of a given polypeptide that is unique to or characteristic of that polypeptide. The term as used herein also refers to any discrete portion of a given polypeptide that retains at least a fraction of the activity of the full-length polypeptide. In certain embodiments, the fraction of activity retained is at least 10% of the activity of the full-length polypeptide. In certain embodiments, the fraction of activity retained is at least 20%, 30%, 40%, 50%, 60%, 70%, 80% or 90% of the activity of the full-length polypeptide. In certain embodiments, the fraction of activity retained is at least 95%, 96%, 97%, 98% or 99% of the activity of the full-length polypeptide. In certain embodiments, the fraction of activity retained is 100% or more of the activity of the full-length polypeptide. Alternatively or additionally, the term as used herein also refers to any portion of a given polypeptide that includes at least an established sequence element found in the full-length polypeptide. In some embodiments, the sequence element spans at least about 4-5, 10, 15, 20, 25, 30, 35, 40, 45, 50 or more amino acids of the full-length polypeptide. Fragments of proteins of the present invention include proteolytic fragments, as well as deletion fragments.

Variants of the proteins of the present invention include fragments as described above, and also polypeptides with altered amino acid sequences due to amino acid substitutions, deletions, or insertions. Variants may occur naturally or be non-naturally occurring. Non-naturally occurring variants may be produced using art-known mutagenesis techniques. Variant proteins may comprise conservative or non-conservative amino acid substitutions, deletions or additions.

The proteins of the invention include proteins having one or more residues chemically derivatized by reaction of a functional side group. Also included as proteins of the invention are polypeptides that contain one or more naturally occurring amino acid derivatives of the twenty standard amino acids. For example, 4-hydroxyproline may be substituted for proline; 5-hydroxylysine may be substituted for lysine; 3-methylhistidine may be substituted for histidine; homoserine may be substituted for serine; and ornithine may be substituted for lysine.

"Recombinantly expressed polypeptide" and "recombinant polypeptide" as used herein refer to a polypeptide expressed from a host cell that has been manipulated to express that polypeptide. In certain embodiments, the host cell is a mammalian cell. In certain embodiments, this manipulation may comprise one or more genetic modifications. For example, the host cells may be genetically modified by the introduction of one or more heterologous genes encoding the polypeptide to be expressed. The heterologous recombinantly expressed polypeptide can be identical or similar to polypeptides that are

normally expressed in the host cell. The heterologous recombinantly expressed polypeptide can also be foreign to the host cell, e.g. heterologous to polypeptides normally expressed in the host cell. In certain embodiments, the heterologous recombinantly expressed polypeptide is chimeric. For example, portions of a polypeptide may contain amino acid sequences that are identical or similar to polypeptides normally expressed in the host cell, while other portions contain amino acid sequences that are foreign to the host cell. Additionally or alternatively, a polypeptide may contain amino acid sequences from two or more different polypeptides that are both normally expressed in the host cell. Furthermore, a polypeptide may contain amino acid sequences from two or more polypeptides that are both foreign to the host cell. In some embodiments, the host cell is genetically modified by the activation or upregulation of one or more endogenous genes.

Calculations of homology or sequence identity between sequences (the terms are used interchangeably herein) are performed as follows. To determine the percent identity of two amino acid sequences, or of two nucleic acid sequences, the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in one or both of a first and a second amino acid or nucleic acid sequence for optimal alignment and non-homologous sequences can be disregarded for comparison purposes). In a typical embodiment, the length of a reference sequence aligned for comparison purposes is at least 30%, at least 40%, at least 50% or 60%, or at least 70%, 80%, 90%, or 100% of the length of the reference sequence. The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the molecules are identical at that position (as used herein amino acid or nucleic acid “identity” is equivalent to amino acid or nucleic acid “homology”).

To determine the percent identity of two amino acid sequences or of two nucleic acids, the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in the sequence of a first amino acid or nucleic acid sequence for optimal alignment with a second amino acid or nucleic acid sequence). The percent identity between the two sequences is a function of the number of identical positions shared by the sequences (i.e., % homology=# of identical positions/total # of positions X 100). The determination of percent homology between two sequences can be accomplished using a mathematical algorithm. A preferred, non-limiting example of a mathematical algorithm utilized for the comparison of two sequences is the algorithm of Karlin et al., *Proc Natl Acad Sci* U S A 87:2264-8 (1990), modified as in Karlin et al., *Proc Natl Acad Sci* U S A 90:5873-7 (1993). Such an algorithm is incorporated into the NBLAST and XBLAST programs of Altschul et al., *J Mol Biol* 215:403-10 (1990). BLAST nucleotide searches can be performed with the NBLAST program, score=100, wordlength=12.

BLAST protein searches can be performed with the XBLAST program, score=50, wordlength=3. To obtain gapped alignments for comparison purposes, Gapped BLAST can be utilized as described in Altschul et al., *Nucleic Acids Res* 25:3389-402 (1997). When utilizing BLAST and Gapped BLAST programs, the default parameters of the respective programs (e.g., XBLAST and NBLAST) can be used.

The percent identity between the two sequences is a function of the number of identical positions shared by the sequences, taking into account the number of gaps, and the length of each gap, which need to be introduced for optimal alignment of the two sequences.

The comparison of sequences and determination of percent identity between two sequences can be accomplished using a mathematical algorithm. In one embodiment, the percent identity between two amino acid sequences is determined using the Needleman-Wunsch algorithm (Needleman et al., J Mol Biol 48:443-53 (1970)) which has been incorporated into the GAP program in the GCG software package (available on at gcg.com), using either a Blossum 62 matrix or a PAM250 matrix, and a gap weight of 16, 14, 12, 10, 8, 6, or 4 and a length weight of 1, 2, 3, 4, 5, or 6. In yet another embodiment, the percent identity between two nucleotide sequences is determined using the GAP program in the GCG software package (available on the internet at gcg.com), using a NWSgapdna.CMP matrix and a gap weight of 40, 50, 60, 70, or 80 and a length weight of 1, 2, 3, 4, 5, or 6. One typical set of parameters (and the one that should be used unless otherwise specified) are a Blossum 62 scoring matrix with a gap penalty of 12, a gap extend penalty of 4, and a frameshift gap penalty of 5.

The percent identity between two amino acid or nucleotide sequences can be determined using the algorithm of E. Myers and W. Miller (Myers et al., *Comput Appl Biosci* 4:11-7 (1988)) which has been incorporated into the ALIGN program (version 2.0), using a PAM120 weight residue table, a gap length penalty of 12 and a gap penalty of 4.

"Instructional material," as that term is used herein, includes a publication, a recording, a diagram, or any other medium of expression which can be used to communicate the usefulness of the compound, combination, and/or composition of the invention in the kit for affecting, alleviating or treating the various diseases or disorders recited herein. Optionally, or alternately, the instructional material can describe one or more methods of alleviating the diseases or disorders in a cell, a tissue, or a mammal, including as disclosed elsewhere herein.

The instructional material of the kit may, for example, be affixed to a container that contains the compound and/or composition of the invention or be shipped together with a container which contains the compound and/or composition. Alternatively, the instructional material may be shipped separately from the container with the intention that the recipient uses the instructional material and the compound cooperatively.

Except when noted, the terms "patient" or "subject" are used interchangeably and refer to mammals such as human patients and non-human primates, as well as veterinary subjects such as rabbits, rats, and mice, and other animals. Preferably, patient refers to a human.

"Effective amount", or "therapeutically effective amount," as the terms are used interchangeably herein, is an amount that when administered to a tissue or a mammal, preferably a human, mediates a detectable therapeutic response compared to the response detected in the absence of the compound. A therapeutic response, such as, but not limited to, inhibition of and/or decreased fibrosis, increased bone

mass or bone density, and the like, can be readily assessed by a plethora of art-recognized methods, including, *e.g.*, such methods as disclosed herein.

The skilled artisan would understand that the effective amount of the compound or composition administered herein varies and can be readily determined based on a number of factors such as the disease or condition being treated, the stage of the disease, the age and health and physical condition of the mammal being treated, the severity of the disease, the particular compound being administered, and the like

As used herein, to "treat" means reducing the frequency with which symptoms of a disease (*e.g.*, decreased bone density, fracture, fibrosis, and the like) are experienced by a patient. The term includes the administration of the compounds or agents of the present invention to prevent or delay the onset of the symptoms, complications, or biochemical indicia of a disease, alleviating the symptoms or arresting or inhibiting further development of the disease, condition, or disorder. Treatment may be prophylactic (to prevent or delay the onset of the disease, or to prevent the manifestation of clinical or subclinical symptoms thereof) or therapeutic suppression or alleviation of symptoms after the manifestation of the disease.

By the phrase "specifically binds," as used herein, is meant a compound, e.g., a protein, a nucleic acid, an antibody, and the like, which recognizes and binds a specific molecule, but does not substantially recognize or bind other molecules in a sample. For instance, an BMP protein, an antibody or a peptide inhibitor which recognizes and binds a cognate receptor (e.g., a BMP type I or type II receptor, an antibody that binds with its cognate antigen, and the like) in a sample, but does not substantially recognize or bind other molecules in the sample. Thus, under designated assay conditions, the specified binding moiety (e.g., a BMP or a receptor binding fragment thereof) binds preferentially to a particular target molecule and does not bind in a significant amount to other components present in a test sample. A variety of assay formats may be used to select an antibody that specifically binds a molecule of interest. For example, solid-phase ELISA immunoassay, immunoprecipitation, BIAcore, FACS, Octet, and Western blot analysis are among many assays that may be used to identify a BMP that specifically reacts with a BMP receptor. Typically, a specific or selective reaction will be at least twice background signal or noise, more preferably, at least five-fold greater than background signal or noise, and more typically, more than 10 times background, even more specifically, a BMP is said to "specifically bind" a BMP receptor when the equilibrium dissociation constant (K_D) is ≤ 100 μM, more preferably ≤ 10 μM, even more preferably ≤ 1 μM, yet more preferably ≤ 100 nM and most preferably ≤ 10 nM.

The term "K_D" refers to the equilibrium dissociation constant of a particular ligand-receptor interaction.

"Binding affinity" generally refers to the strength of the sum total of noncovalent interactions between a binding site of a molecule (e.g., a BMP ligand) and its binding partner (e.g., a BMP type I or type II receptor). Unless indicated otherwise, as used herein, "binding affinity" refers to intrinsic binding affinity which reflects a 1:1 interaction between members of a binding pair (e.g., BMP and its cognate

receptor). The affinity of a molecule X for its partner Y can generally be represented by the dissociation constant (Kd).

Affinity can be measured by common methods known in the art, including those described herein. Low-affinity BMPs generally bind a receptor slowly and tend to dissociate readily, whereas high-affinity BMPs generally bind a receptor faster and tend to remain bound longer. A variety of methods of measuring binding affinity are known in the art, any of which can be used for purposes of the present invention. Specific illustrative embodiments are described elsewhere herein.

The term "k_{on}", as used herein is intended to refer to the association or on rate constant, or specific reaction rate, of the forward, or complex-forming, reaction, measured in units: M^{-1} sec^{-1}.

The term "k_{off}", as used herein, is intended to refer to the dissociation or off rate constant, or specific reaction rate, for dissociation of an antibody from the antibody/antigen complex, measured in units: sec^{-1}.

The term "K_d", as used herein, is intended to refer to the dissociation constant of a particular antibody-antigen interaction. It is calculated by the formula:

$k_{off} / k_{on} = K_d$

The term “altered binding” as used herein means the designer BMP comprises a different binding specificity for at least a type I receptor and/or a type II receptor when compared with the binding of a corresponding wild type BMP to the same type I and/or type II receptor. The designer BMP may bind with greater or lesser affinity with the receptor compared to the binding of the wild type BMP to that receptor. For instance, if the wild type BMP bound a certain type I receptor with a certain binding affinity, the corresponding designer BMP binds that receptor with greater or lesser affinity compared with the wild type BMP. It may even be that the designer BMP will specifically bind a receptor that the wild type BMP did not detectably bind and vice-a-versa where the designer BMP will no longer detectably bind a receptor that the wild type BMP binds. Thus, altered binding encompasses any detectable change in binding by a designer BMP to a type I or type II receptor compared with the binding of that receptor by the corresponding wild type BMP. It may be that the designer BMP has a greater or lesser k_{on} value compared with the k_{on} value for a corresponding wild type BMP and/or the designer BMP has a greater or lesser k_{off} value compared with the k_{off} value of the corresponding wild type BMP such that the Kd of the designer BMP is greater or lesser than the Kd of a corresponding wild type BMP for the same BMP receptor. Thus, any difference in a binding characteristic and/or affinity value between a designer BMP and a corresponding wild type BMP are encompassed by the term “altered binding” as used herein.

The term "surface plasmon resonance", as used herein, refers to an optical phenomenon that allows for the analysis of real-time biospecific interactions by detection of alterations in protein concentrations within a biosensor matrix, for example using the BIAcore system (Pharmacia Biosensor AB, Uppsala, Sweden, and Piscataway, N.J.). For further descriptions, see, e.g., Johnsson, et al., *Ann. Biol. Clin.* 51: 19-26 (1993); Johnsson, et al., *Biotechniques* 11: 620-627 (1991); Johnsson, et al., *J. Mol. Recognit.* 8: 125-131 (1995); and Johnnson, et al., *Anal. Biochem.* 198: 268-277 (1991).

As used herein, "substantially pure" means an object species is the predominant species present (i.e., on a molar basis it is more abundant than any other individual species in the composition), and preferably a substantially purified fraction is a composition wherein the object species (e.g., a designer BMP) comprises at least about 50 percent (on a molar basis) of all macromolecular species present. Generally, a substantially pure composition will comprise more than about 80 percent of all macromolecular species present in the composition, more preferably more than about 85%, 90%, 95%, and 99%. Most preferably, the object species is purified to essential homogeneity (contaminant species cannot be detected in the composition by conventional detection methods) wherein the composition consists essentially of a single macromolecular species.

Description

Bone Morphogenetic Proteins (BMPs)

As stated previously elsewhere herein, BMPs are members of the TGF-β protein superfamily all of which are characterized by six-conserved cysteine residues (Lander et al, (2001) Nature, 409:860-921. The BMP/GDF subfamily includes, but is not limited to, BMP2 , BMP3 (osteogenin) (see, e.g., US Patent No. 6,177,406), BMP3b (GDF-10) (see, e.g., US Patent No. 6,204,047), BMP4 (BMP2b) (see, e.g., US Patent No. 6,245,889), BMP5 (see, e.g., US Patent No. 5,543,394), BMP6 (see, e.g., US Patent No. 6,613,744), BMP7 (osteogenic protein-1 or OP1) (see, e.g., US Patent No. 5,141,905), BMP8 (OP2) (see, e.g., US Patent No. 5,688,678), BMP8B (OP3) (see, e.g., US Patent No. 5,854,071), BMP9 (GDF2) (see, e.g., US Patent No. 6,287,816), BMP10 (see, e.g., US Patent No. 5,703,043), BMP11 (GDF11) (see, e.g., US Patent No. 6,437,111), BMP12 (GDF7) (see, e.g., US Patent No. 6,027,919), BMP13 (GDF6, CDMP2) (see, e.g., US Patent No. 6,027,919), BMP15 (GDF9) (see, e.g., US Patent No. 6,034,229), BMP16 (see, e.g., US Patent No. 6,331,612), GDF1 (see, e.g., US Application No. 2004/0039162), GDF3 (see, e.g., US Patent No. 6,025,475), GDF5 (CDMP1; MP52) (see, e.g., US Patent No. 5,994,094), and GDF8 (myostatin) (see, e.g., US Patent No. 5,827,733).

BMPs specifically bind their cognate receptors, which include Type I receptors: ALK-I, ALK-2 (also called ActRIa or ActRI), ALK-3 (also called BMPRIa), and ALK-6 (also called BMPRIb); and Type II receptors: ActRIIa (also called ActRII), ActRIIb, and BMPRII. The BMP-receptor binding interactions have been studied extensively, and the binding specificities of each wild type BMP for each type I and/or type II receptor is generally known in the art and are shown in Table 1. See, e.g., Nickel et al., *Cytokine Growth Factor Rev* 20:367-77 (2009); Heinecke et al., *BMC Biol* 7:59 (2009).

TABLE 1

	ALK 1	ALK 2	ALK 3	ALK 6	ACTIIA	ACTIIB	BMPRII
BMP-2	No Binding	No Binding	++++	++++	++	+++	++
BMP-4	No Binding	No Binding	++++	++++	++	++	++
BMP-6	No Binding	No Binding	++	++	++++	++++	++++

	ALK 1	ALK 2	ALK 3	ALK 6	ACTIIA	ACTIIB	BMPRII
BMP-7	No Binding	No Binding	++	++	++++	++++	++++
BMP-9	+++++	No Binding	No Binding	No Binding	++	+++	++++

Designer Bone Morphogenetic Proteins with Improved Osteogenic Activity

This application is based, in part on the understanding that each BMP dimer binds to four BMP receptors two type I receptors and two type II receptors. The specificities of each BMP for each receptor are known in the art as shown above in Table 1. Also, the receptor binding regions of various BMPs that mediate binding of the BMP for each receptor have been mapped and are shown in Table 2. For instance, it is well established that wild type BMP2 and BMP4 bind type I BMP receptors Alk-3 and ALK-6 with high affinity and bind type II BMP receptors with lower affinity. On the other hand, wild type BMP6 and BMP7 are known to have bind type II receptors ActrIIA, ActrIIB, and BMPRII with high affinity but bind type I receptors with lower affinity than they do to type II. It is believed that the differing cellular responses from the approximately forty-three TGFβ superfamily members signaling through interaction with approximately twelve receptors is believed to be due to each ligand utilizing a specific repertoire of receptors with which it binds with differing affinities. The type I and II binding domains are described in Table 2.

TABLE 2

BMP	Type II domain A amino acids	Type I domain amino acids	Type II domain B amino acids
BMP2 (SEQ ID NO 1)	31-44	48-76	83-100
BMP4 (SEQ ID NO:2)	33-46	50-78	85-102
BMP5 (SEQ ID NO:3)	54-67	71-100	107-120
BMP6 (SEQ ID NO:4)	55-69	73-102	108-126
BMP7 (SEQ ID NO:5)	55-69	73-102	108-126
BMP8 (SEQ ID NO:6)	55-69	73-102	108-126
BMP9 (SEQ ID NO:7)	25-39	42-71	78-96

Rational amino acid substitution to alter receptor binding of designer BMPs

In one embodiment, the invention comprises introducing an amino acid mutation in at least one receptor binding site thereby providing altered binding to type I and type II BMP receptors by designer BMPs compared to the binding of the corresponding wild type BMP to those receptors. That is, it is well known in the art that wild type BMP2 shows a relatively high affinity for type I receptors, while wild type BMP6 shows a high affinity for type II receptors. It is further known in the art that heterodimers of wild type BMP2 and BMP6 bind to both type I and type II receptors with relatively high affinity each BMP apparently providing the higher affinity binding site for each receptor. See Table 3, below. The BMP2/6

heterodimers are known to be more active that BMP2 or BMP6 alone or as homodimers, in both *in vitro* and *in vivo* bone formation assays. Table 3 shows an example of BMP2 and BMP6 binding affinities to type I and II receptors.

TABLE 3

	Type I		Type II	
Ligand	ALK3 K_D (nM)	ALK6 K_D (nM)	ActRIIA K_D (nM)	ActRIIB K_D (nM)
BMP2	0.69	0.17	141	42
BMP6	150	102	0.73	2.0
BMP2/6	1.67	0.43	2.56	1.15

Accordingly, it is an object of the invention to provide designer BMPs with improved binding to type I and/or type II receptors. As shown in Figure 1A and Table 2, each BMP comprises three binding sites that contribute to receptor binding. From N- to C-terminus, each BMP comprises a type II receptor binding site A, a type I receptor binding site, and a second type II receptor binding site B. Although an exemplary alignment of wild type BMP2, BMP4, BMP5, BMP6, BMP7, BMP8, and BMP9 is illustrated in Figure 1, the skilled artisan will appreciate that there are well-known alignments providing the relative positioning of various amino acids among the members of the TGBβ superfamily. Such alignments are provided, among others, in International Publication Nos. WO 2009/086131 (e.g., Figures 15-17, Figure 31A), WO 2008/051526 (Figures 9-12), WO 2005/118636 (Figure 6), WO 2005/118635, WO 2005/113585 (Figure 3), WO 2001/92298 (Figure 6A-6C), Kirsch et al., *EMBO J.* 19 :3314-3324 (2000) (Figure 1), US Patent Application Publication No., 2007/0293425 (Figure 6), Groppe et al., *Nature* 420 :636-642 (2002), Nickel et al., *J. Bone Joint Surg. Am.* 83:7-14 (2001), and Weber et al., *BMC Structural Biol.* 7 :6 (2007). Thus, using protein sequence alignment algorithms and tools well-known in the art, including the alignments of the amino acid sequences of the various TGFβ superfamily members, as well as the disclosure provided herein, the corresponding amino acid in one BMP/GDF protein relative to the amino acid at any position in another BMP/GDF protein can be determined. In one embodiment, the corresponding amino acid residues in BMP-2, BMP-4, BMP-5, BMP-6, BMP-7, BMP-8 and BMP-9 are shown (see, e.g., Figure 1A).

In some embodiments of the invention, the designer BMP comprises mutations in a type I binding domain or a type II binding domain, wherein the mutations confer altered binding to a type I or type II BMP receptor. In some embodiments, the designer BMP comprises one or more mutations in both a type I binding domain and a first (binding domain A) or second (binding domain B) type II binding domain. In other embodiments, the designer BMP comprises one or more mutations in both type II binding domains. In other embodiments, the designer BMP comprises one or more mutations in the first type II binding

domain, in the second type II binding domain, and in the type I binding domain. In some embodiments, the designer BMP comprises one or more mutations in the type I binding domain.

In some embodiments, the mutations improve binding to a type I receptor. In other embodiments, the mutations improve binding to a type II receptor. In other embodiments, the mutations decrease binding to a type I or type II receptors. In some embodiments, the mutations create or destroy a glycan tether as more fully set forth below. In some embodiments, the mutations create or destroy a His doorstop as more fully set forth below.

Because BMPs are so well characterized and understood in the art, it would be understood, once provided with the disclosure provided herein, the location of possible mutations that can be made that do not further affect the activity of the designer BMPs would be understood. Accordingly, the designer BMPs of the invention encompass variant BMPs which differ from a corresponding wild type or designer BMP in that it contains additional insertions, deletions, or substitutions which do not affect the receptor binding affinity of the variant BMPs. In some non-limiting embodiments, those of skill in the art would understand that the cysteines involved in cysteine knot formation and amino acids involved in receptor interactions should not be mutated or should be changed with conservative substitutions, while other amino acids may be more freely substituted, inserted, or deleted without adversely affecting biological activity of the designer BMP.

It should be noted that unless otherwise stated, all positional numbering of designed or modified BMPs is based on the sequences of the mature native BMPs. Designer BMPs are characterized by the predetermined nature of the variation, a feature that sets them apart from naturally occurring allelic or interspecies variation of the BMP sequence. Variants of designer BMPs must retain at least 50% of the activity of the corresponding wild type or designer BMP activity in one or more cell types, as determined using an appropriate assay described below. Variants that retain at least 75%, 80%, 85%, 90% or 95% of wild type activity are more preferred, and variants that are more active than wild type are especially preferred. A designer BMP may contain insertions, deletions, and/or substitutions at the N- terminus, C-terminus, or internally. In a preferred embodiment, designed or modified BMPs have at least 1 residue that differs from the most similar human BMP sequence, with at least 2, 3, 4, 5, 6, 7, 8, 9, 10 or more different residues being more preferred.

Designer BMPs of the invention maintain at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% identity with the corresponding wild-type BMP protein sequence.

Designer BMPs of the invention may maintain at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% identity with the conserved cysteine domain of the C-terminal region of the corresponding wild-type BMP protein sequence.

Designer BMPs may contain further modifications, for instance mutations that alter additional protein properties such as stability or immunogenicity or which enable or prevent posttranslational modifications such as PEGylation or glycosylation. Designer BMPs may be subjected to co- or post-translational modifications, including but not limited to synthetic derivatization of one or more side chains or termini, glycosylation, PEGylation, circular permutation, cyclization, fusion to proteins or protein domains, and addition of peptide tags or labels.

Due to the degeneracy of the genetic code, an extremely large number of nucleic acids may be made, all of which encode the designer BMPs of the present invention, by simply modifying the sequence of one or more codons in a way that does not change the amino acid sequence of the designer BMP. The designer BMPs of the invention do not comprise these sequences set forth in WO2008/051526 or WO2009/086131.

As described above, BMPs are naturally expressed as pro-proteins comprising a long pro-domain, one or more cleavage sites, and a mature domain. This pro-protein is then processed by the cellular machinery to yield a dimeric mature BMP molecule. In a preferred embodiment, the designer BMPs of the invention are produced in a similar manner. The pro-domain is believed to aid in the correct folding and processing of BMPs. Furthermore, in some but not all BMPs, the pro-domain may noncovalently bind the mature domain and may act as a chaperone, as well as an inhibitor (e.g., Thies et al. (2001) Growth Factors, 18:251-259). Preferably, the modified BMPs of the invention are produced and/or administered therapeutically in this form. Alternatively, BMPs may be produced in other forms, including, but not limited to, where the mature domain is produced directly or refolded from inclusion bodies, or comprises full-length intact pro protein. The designer BMPs of the invention will be useful in these and other forms.

In particular embodiments, the designer BMP of the invention comprises a backbone BMP, i.e., the wild type BMP, to which the designer BMP corresponds. In particular embodiments, this backbone BMP may be a wild type BMP2, BMP4, BMP5, BMP6, BMP7, BMP8, or BMP9 backbone.

In some embodiments of the invention, the designer BMP comprises at least one mutation in a type I binding domain and/or a type II binding domain, wherein the mutation confers altered binding to a type I or type II BMP receptor compared with the binding of a corresponding wild type BMP not comprising the mutation. In some embodiments, the designer BMP comprises at least one mutation in both a type I binding domain and at least one mutation in a type II binding domain. In other embodiments, the designer BMP comprises at least one mutation within the type II binding domain A and the type II binding domain B. In other embodiments, the designer BMP comprises at least one mutation in type II binding domain A, type II binding domain B, and a type I binding domain.

In certain embodiments, the mutation may comprise an amino or nucleic acid substitution, deletion and/or insertion. In a preferred embodiment, the mutation comprises an amino acid substitution.

In some embodiments, the backbone BMP is a wild type BMP and the mutations are one or more of the mutations listed in Tables 4 to 6. The designer BMP may contain any combination and any number of mutations listed in these tables.

In some embodiments, the backbone BMP is a wild type BMP and the mutations are one or more of the mutations listed in Tables 4 to 6. The designer BMP may contain a permutation and any and all of the mutations listed in these tables or disclosed elsewhere herein.

TABLE 4

Type I Binding Domain Mutations							
BMP2	BMP4	BMP5	BMP6	BMP7	BMP8	BMP9	Possible mutations
P48	P50	S71	S72	A72	S72	F42	F, S, N, A, P
F49	F51	F72	F73	F73	F73	F43	Y
A52	A54	N75	N76	N76	D76	A46	N, A
D53	D55	A76	A77	S77	S77	D47	A, E, D
H54	H56	H77	H78	Y78	C78	D48	D, C
L55	L57	M78	M79	M79	M79	V49	M, V, L
N56	N58	N79	N80	N80	N80	T50	T, N
S57	S59	A80	A81	A81	A82	P51	A P
N59	N61	N82	N83	N83	N83	K53	K, N
V63	V65	V86	V87	V87	L87	V57	I, V, L
T65	T67	T88	T89	T89	S89	T59	A, T, S
N68	N70	H91	H92	H92	H92	H62	H, N
S69	S71	L92	L93	F93	L93	L63	L, S, F
V70	V72	M93	M94	I94	M94	K64	M, K, I, V
N71	N73	F94	N95	N95	M95	F65	F, N, M
		P95	P96	P96	P96	P66	INSERT S, P; DELETE P
S72	S74	D96	E97	E97	D97	T67	Q, T, E, D
K73	S75	H97	Y98	T98	A98	K68	Y, H, T, A, K
I74	I76	V98	V99	V99	V99	V69	A, V, I
P75	P77	P99	P100	P100	P100	G70	S, G

TABLE 5

Type II Binding Domain A Mutations							
BMP2	BMP4	BMP5	BMP6	BMP7	BMP8	BMP9	Possible mutations
V33	V35	I56	I57	I57	I57	I27	I, V

P36	P38	E59	K60	E60	Q60	K30	K, R, P, E, Q
G37	G39	G60	G61	G61	G61	E31	G, E
H39	Q41	A62	A63	A63	S63	E33	A, E, S, Q
F41	F43	F64	N65	Y65	Y65	Y35	N, Y, F
Y42	Y44	Y65	Y66	Y66	Y66	E36	Y, E
H44	H46	D67	D68	E68	E68	K38	H, D, K, R, E

TABLE 6

Type II Binding Domain B Mutations							
BMP2	BMP4	BMP5	BMP6	BMP7	BMP8	BMP9	Possible mutations
E83	E85	K107	K108	Q108	K108	K78	Q, K, E
S85	S87	N109	N110	N110	S110	S80	N, S
A86	A88	A110	A111	A111	A111	P81	P, A
M89	M91	V113	V114	V114	V114	V84	M, V
L92	L94	F116	F117	F117	Y117	K87	F, K, L, Y
E94	E96	D118	D119	D119	D118	D89	D, E
N95	Y97	S119	N120	S120	S119	M90	M, N, S
E96	D98	S120	S121	S121	S120	G91	S, G, D
K97	K99	N121	N122	N122	N121, N122	V92	N, V, K
V98	V100	V122	V123	V123	V123	P93	P, V
V99	V101	I123	I124	I124	I124	T94	T, I, V

In some embodiments, the mutations improve binding to a type I receptor. In other embodiments improve binding to a type II receptor. In other embodiments, the mutations decrease binding to a type I or type II receptors.

Tables 4-6 above provide a non-limiting compilation of example mutations of the present invention where the position of the mutation is provided relative to the corresponding wild type BMP amino acid sequence. Thus, in some embodiments, the designer BMP comprises the following preferred combinations of mutations.

In certain embodiments, the corresponding wild type BMP to the designer BMP is BMP2. Further, the at least one mutation within the type II receptor binding domain A is a mutation selected from the group consisting of V33, P36, G37, H39, F41, Y42 AND H44.

In other embodiments, the designer BMP comprises at least one mutation within the type II receptor binding domain A and further comprises at least one additional mutation within a type I receptor binding domain. The mutation within the type I receptor binding domain is at least one mutation at P48,

F49, A52, D53, H54, L55, N56, S57, N59, V63, T65, N68, S69, V70, N71, S72, K73, I74, and P75 with respect to the sequence of SEQ ID NO:1.

In yet further embodiments, the designer BMP comprises at least one mutation within a type II receptor binding domain A, at least one mutation within the type I receptor binding domain, and further comprises at least one additional mutation within a type IIB receptor binding domain. The mutation within the type II receptor binding domain B is at least one mutation at E83, S85, A86, M89, L92, E94, N95, E96, K97, V98, and V99 with respect to the sequence of SEQ ID NO:1.

In some embodiments, the designer BMP comprises mutations at each of amino acids H44, P48, A52, D53, L55, S57, N68, S69, V70, insertion of P after N71, S72, K73, I74, A77, and V80 with respect to the sequence of SEQ ID NO:1.

In one embodiment, the designer BMP comprises the following mutations: H44D, P48S, A52N, D53A, L55M, S57A, N68H, S69L, V70M, insertion of a P after N71, S72E, K73Y, I74V, A77P, and V80A with respect to the sequence of SEQ ID NO:1.

In some embodiments the designer BMP comprises mutations at each of amino acids V33, P36, H39, S85, M89, L92, E94, E96, K97, and V99 with respect to the sequence of SEQ ID NO:1.

In some embodiments, the designer BMP comprises mutations at each of amino acids V33I, P36K, H39A, S85N, M89, L92F, E94D, E96S, K97N, and V99I with respect to the sequence of SEQ ID NO:1.

In other embodiments, the designer BMP comprises the following mutations: V33I, P36K, H39A, H44D, P48S, A52N, L54M, S56M, N68H, V70M, S72E, K73E, insertion of a Y after K73, I74V, 77AP, S85N, M89V, L92F, E94D, E96S, K97N, and V99I with respect to the sequence of SEQ ID NO:1.

In yet other embodiments, the designer BMP comprises the following mutations: V33I, P36R, H39A, H44D, P48S, A52N, L54M, S56M, N68H, V70M, S72E, K73E, insertion of a Y after K73, I74V, 77AP, S85N, M89V, L92F, E94D, E96S, K97N, and V99I with respect to the sequence of SEQ ID NO:1.

In certain embodiments, the corresponding wild type BMP to the designer BMP is BMP4. In certain embodiments, the at least one mutation within the type II receptor binding domain A is at V35, P38, G39, Q41, F43, Y44, and H46 of SEQ ID NO:2.

In other embodiments, the designer BMP4 comprises at least one mutation within the type II receptor binding domain A and further comprises at least one additional mutation within a type I receptor binding domain. The mutation within the type I receptor binding domain is at least one mutation at P50, A54, D55, H56, L57, N58, S59, N61, V65, T67, N70, S71, V72, N73, S74, S75, I76, and P77 of SEQ ID NO:2.

In yet further embodiments, the designer BMP4 comprises at least one mutation within a type II receptor binding domain A, at least one mutation within the type I receptor binding domain, and further comprises at least one additional mutation within a type IIB receptor binding domain. The mutation within the type II receptor binding domain B is at least one mutation at E85, S87, A88, M91, L94, E96, K97, V98 and V99 of SEQ ID NO:2.

In certain embodiments, the corresponding wild type BMP to the designer BMP is BMP5. In certain embodiments, the mutation within the type II receptor binding domain A is at least one mutation at I56, E59, G60, A62, F64, Y65, or D67 of SEQ ID NO:3.

In other embodiments, the designer BMP comprises at least one mutation within the type II receptor binding domain A and further comprises at least one additional mutation within a type I receptor binding domain. The mutation within the type I receptor binding domain is at least one mutation at S71, F72, N75, A76, H77, M78, N79, A80, N82, V86, T88, H91, L92, M93, F94, P95, D96, H97, V98, or P99 of SEQ ID NO:3.

In yet further embodiments, the designer BMP comprises at least one mutation within a type II receptor binding domain A, at least one mutation within the type I receptor binding domain, and further comprises at least one additional mutation within a type IIB receptor binding domain. The mutation within the type II receptor binding domain B is at least one mutation at K107, N109, A110, V113, F116, D118, S119, S120, N121, V122, or I123 of SEQ ID NO:3.

In certain embodiments, the corresponding wild type BMP to the designer BMP is BMP6. In certain embodiments, the mutation within the type II receptor binding domain A is at least one mutation at I57, K60, G61, A63, N65, Y66, or D68 of SEQ ID NO:4.

In other embodiments, the designer BMP6 comprises at least one mutation within the type II receptor binding domain A and further comprises at least one additional mutation within a type I receptor binding domain. The mutation within the type I receptor binding domain is at least one mutation at S72, N76, A77, H78, M79, N80, A81, N83, V87, T89, H92, L93, M94, N95, P96, E97, Y98, V99, or P100 of SEQ ID NO:4.

In yet further embodiments, the designer BMP6 comprises at least one mutation within a type II receptor binding domain A, at least one mutation within the type I receptor binding domain, and further comprises at least one additional mutation within a type IIB receptor binding domain. The mutation within the type II receptor binding domain B is at least one mutation at K108, N110, A111, V114, F117, D119, N120, S121, N122, V123, or I124 of SEQ ID NO:4.

In certain embodiments, the corresponding wild type BMP to the designer BMP is BMP7. In certain embodiments, the mutation within the type II receptor binding domain A is at least one mutation at I57, E60, G61, A63, Y65, Y66, or E68 of SEQ ID NO:5.

In other embodiments, the designer BMP7 comprises at least one mutation within the type II receptor binding domain A and further comprises at least one additional mutation within a type I receptor binding domain. The mutation within the type I receptor binding domain is at least one mutation at A72, F73, N76, S77, Y78, M79, N80, A81, N83, V87, T89, H92, F93, I94, N95, P96, E97, T98, V99, or P100 of SEQ ID NO:5.

In yet further embodiments, the designer BMP7 comprises at least one mutation within a type II receptor binding domain A, at least one mutation within the type I receptor binding domain, and further comprises at least one additional mutation within a type IIB receptor binding domain. The mutation within

the type II receptor binding domain B is at least one mutation at Q108, N110, A111, V114, F117, D119, S120, S121, N122, V123, or I124 of SEQ ID NO:5.

In certain embodiments, the corresponding wild type BMP to the designer BMP is BMP8. In certain embodiments, the mutation within the type II receptor binding domain A is at least one mutation at I57, Q60, G61, S63, Y65, Y66, or E68 of SEQ ID NO:6.

In other embodiments, the designer BMP8 comprises at least one mutation within the type II receptor binding domain A and further comprises at least one additional mutation within a type I receptor binding domain. The mutation within the type I receptor binding domain is at least one mutation at S72, F73, D76, S77, C78, M79, N80, A82, N83, L87, S89, H92, L93, M94, M95, P96, D97, A98, V99, or P100 of SEQ ID NO:6.

In yet further embodiments, the designer BMP8 comprises at least one mutation within a type II receptor binding domain A, at least one mutation within the type I receptor binding domain, and further comprises at least one additional mutation within a type IIB receptor binding domain. The mutation within the type II receptor binding domain B is at least one mutation at K108, S110, A111, V114, Y117, D118, S119, S120, N121, N122, V123, or I124 or SEQ ID NO:6

In certain embodiments, the mutation within the type II receptor binding domain A is at least one mutation at I27, K30, E31, E33, Y35, or E36 of SEQ ID NO:7.

In other embodiments, the designer BMP9 comprises at least one mutation within the type II receptor binding domain A and further comprises at least one additional mutation within a type I receptor binding domain. The mutation within the type I receptor binding domain is at least one mutation at F42, F43, A46, D47, D48, V49, T50, P51, K53, V57, T59, H62, L63, K64, F65, P66, T67, K68, V69, or G70 of SEQ ID NO:7.

In yet further embodiments, the designer BMP9 comprises at least one mutation within a type II receptor binding domain A, at least one mutation within the type I receptor binding domain, and further comprises at least one additional mutation within a type IIB receptor binding domain. The mutation within the type II receptor binding domain B is at least one mutation at K78, S80, P81, V84, K87, D89, M90, G91, V92, P93, or T94 of SEQ ID NO:7.

Exemplary amino acid sequences of designer BMPs are set forth in Table 7, below. Table 7 shows the name and sequence of the designed molecules.

TABLE 7

NAME	SEQUENCE	SEQ ID NO
BMP-A	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYAANYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	8
BMP-B	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTKLNAISVLYFDDNSNVILKNYQDMVVEGCGCR	9
BMP-C	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYAANYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTKLNAISVLYFDDNSNVILKNYQDMVVEGCGCR	10
BMP-D	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCHGECPFPLNAHMNATNHAIVQTLVH LMN-SKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	11

NAME	SEQUENCE	SEQ ID NO
BMP-E	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCAPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	12
BMP-F	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYAAFYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	13
BMP-G	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYAAFYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELNAISVLYFDDNSNVILKNYQDMVVEGCGCR	14
BMP-H	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYHAFYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELNAISVLYFDENSNVVLKKYQDMVVRGCGCR	15
BMP-I	QAKHKQRKRLKSSCKRHELYVSFQDLGWQDWIIAPKGYAANYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELNAISVLYFDDNSNVILKKYRNMVVRACGCR	16
BMP-J	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYHAFYCDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCAPTELNAISVLYFDENSNVVLKKYQDMVVRGCGCR	17
BMP-K	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCHGECPFPLADHLNSTKHAIVQTLVN SVNSKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	18
BMP-T	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCHGECPFPLADHLNSTTHAIVQTLVN SVNSKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	19
BMP-AP	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPPGYAANYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	20
BMP-AR	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPRGYAANYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	21
BMP-AK	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYAANYCHGECPFPLADHLNSTKHAIVQTLVN SVNSKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	22
BMP-AT	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYAANYCHGECPFPLADHLNSTTHAIVQTLVN SVNSKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	23
BMP-DP	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCHGECPFPLNAHMNATNHAIVQTLVH LMNPSKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	24
BMP-E9	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCKGGCFFPLADDVTPTKHAIVQTLVH LKFPTKVGKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	25
BMP-E10	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCRGVCNYPLAEHLTPTKHAIIQALVH LKNSQKASKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	26
BMP-EK	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCDGECSFPLNAHMNATKHAIVQTLVH LMNPEYVPKPCCAPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	27
BMP-ET	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCDGECSFPLNAHMNATTHAIVQTLVH LMNPEYVPKPCCAPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	28
BMP-R	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPRGYHAFYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	29
BMP-G5	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCEGLCEFPLRSHLEPTNHAVIQTLMN SMDPESTPPTCCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	30
BMP-ER	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPRGYHAFYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	31
BMP-GP	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPPGYAAFYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELNAISVLYFDDNSNVILKNYQDMVVEGCGCR	32
BMP-GR	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPRGYAAFYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELNAISVLYFDDNSNVILKNYQDMVVEGCGCR	33
BMP-GK	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYAAFYCHGECPFPLADHLNSTKHAIVQTLVN SVNSKIPKACCVPTELNAISVLYFDDNSNVILKNYQDMVVEGCGCR	34
BMP-GT	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYAAFYCHGECPFPLADHLNSTTHAIVQTLVN SVNSKIPKACCVPTELNAISVLYFDDNSNVILKNYQDMVVEGCGCR	35
BMP-GE	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYAAFYCDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCAPTELNAISVLYFDDNSNVILKNYQDMVVEGCGCR	36
BMP-GER	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPRGYAAFYCDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCAPTELNAISVLYFDDNSNVILKNYQDMVVEGCGCR	37
BMP-JP	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPPGYHAFYCDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCAPTELNAISVLYFDENSNVVLKKYQDMVVRGCGCR	38
BMP-JR	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPRGYHAFYCDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCAPTELNAISVLYFDENSNVVLKKYQDMVVRGCGCR	39
BMP-JK	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYHAFYCDGECSFPLNAHMNATKHAIVQTLVH LMNPEYVPKPCCAPTELNAISVLYFDENSNVVLKKYQDMVVRGCGCR	40
BMP-JT	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKGYHAFYCDGECSFPLNAHMNATTHAIVQTLVH LMNPEYVPKPCCAPTELNAISVLYFDENSNVVLKKYQDMVVRGCGCR	41
BMP-A9	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKEYEAYECHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	42

NAME	SEQUENCE	SEQ ID NO
BMP-B9	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELSPISVLYKDDMGVPTLKNYQDMVVEGCGCR	43
BMP-E9B	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCAPTELSPISVLYKDDMGVPTLKNYQDMVVEGCGCR.	44
BMP-G9	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPKEYEAYECHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTELSPISVLYKDDMGVPTLKNYQDMVVEGCGCR	45
BMP929	QAKHKQRKRLKSSCQKTSLRVNFEDIGWDSWIIAPKEYEAYECHGECPFPLADHLNSTNHAIVQTLVN SVNSKIPKACCVPTKLSPISVLYKDDMGVPTLKYHYEGMSVAECGCR	46
BMP969	QAKHKQRKRLKSSCQKTSLRVNFEDIGWDSWIIAPKEYEAYECDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCVPTKLSPISVLYKDDMGVPTLKYHYEGMSVAECGCR	47
BMPQAK no SAGA	QAKHKQRKRLKSSCQKTSLRVNFEDIGWDSWIIAPKEYEAYECKGGCFFPLADDVTPTKHAIVQTLVH LKFPTKVGKACCVPTKLSPISVLYKDDMGVPTLKYHYEGMSVAECGCR	48
BMP-QAKSAGAC	QAKHKQRKRLKSSSAGAGSHCQKTSLRVNFEDIGWDSWIIAPKEYEAYECKGGCFFPLADDVTPTKHA IVQTLVHLKFPTKVGKACCVPTKLSPISVLYKDDMGVPTLKYHYEGMSVAECGCR	49
BMP-GEP	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPPGYAAFYCDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCAPTELNAISVLYFDDNSNVILKNYQDMVVEGCGCR	50
BMP6-SA	VSSASDYNSSELKTACRKHELYVSFQDLGWQDWIIAPKGYAANYCDGECSFPLNAAMNATNHAIVQTL VHLMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRACGCH	51
BMP6-SL	VSSASDYNSSELKTACRKHELYVSFQDLGWQDWIIAPKGYAANYCDGECSFPLNAHLNATNHAIVQTL VHLMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRACGCH	52
BMP6A	SASSRRRQQSRNRSTQSQDVARVSSASDYNSSELKTACRKHELYVSFQDLGWQDWIIAPKGYAANYCD GECSFPLADHLNSTNHAIVQTLVNSVNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRAC GCH.	53
BMP6B	SASSRRRQQSRNRSTQSQDVARVSSASDYNSSELKTACRKHELYVSFQDLGWQDWIIAPKGYAANYCH GECPFPLADHLNSTNHAIVQTLVNSVNSKIPKACCVPTKLNAISVLYFDDNSNVILKKYRNMVVRACG CH.	54
BMP6C	SASSRRRQQSRNRSTQSQDVARVSSASDYNSSELKTACRKHELYVSFQDLGWQDWIVAPPGYHAFYCD GECSFPLNAHMNATNHAIVQTLVHLMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRAC GCH.	55
BMP6D	SASSRRRQQSRNRSTQSQDVARVSSASDYNSSELKTACRKHELYVSFQDLGWQDWIIAPKGYAANYCD GECSFPLNAHMNATNHAIVQTLVHLMNPEYVPKPCCAPELSAISMLYLDENEKVVLKKYRNMVVRACG CH.	56
BMP6 ADHL	VSSASDYNSSELKTACRKHELYVSFQDLGWQDWIIAPKGYAANYCDGECSFPLADHLNATNHAIVQTL VHLMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRACGCH	57
BMP6-RK-KR	VSSASDYNSSELKTACKRHELYVSFQDLGWQDWIIAPKGYAANYCDGECSFPLNAHMNATNHAIVQTL VHLMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRACGCH	58
BMP6 RK-KR ADHL long	VSSASDYNSSELKTACKRHELYVSFQDLGWQDWIIAPKGYAANYCDGECSFPLADHLNATNHAIVQTL VHLMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRACGCH	59
BMP6A RK-KR	SASSRRRQQSRNRSTQSQDVARVSSASDYNSSELKTACKRHELYVSFQDLGWQDWIIAPKGYAANYCD GECSFPLADHLNSTNHAIVQTLVNSVNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRAC GCH	60
BMP6 ADHL long	SASSRRRQQSRNRSTQSQDVARVSSASDYNSSELKTACRKHELYVSFQDLGWQDWIIAPKGYAANYCD GECSFPLADHLNATNHAIVQTLVHLMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRAC GCH.	61
BMP6 RK-KR ADHL	SASSRRRQQSRNRSTQSQDVARVSSASDYNSSELKTACKRHELYVSFQDLGWQDWIIAPKGYAANYCD GECSFPLADHLNATNHAIVQTLVHLMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRAC GCH.	62
BMP6 RK-KR long	SASSRRRQQSRNRSTQSQDVARVSSASDYNSSELKTACKRHELYVSFQDLGWQDWIIAPKGYAANYCD GECSFPLNAHMNATNHAIVQTLVHLMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRAC GCH	63
BMP6B-RK-KR	SASSRRRQQSRNRSTQSQDVARVSSASDYNSSELKTACKRHELYVSFQDLGWQDWIIAPKGYAANYCH GECPFPLADHLNSTNHAIVQTLVNSVNSKIPKACCVPTKLNAISVLYFDDNSNVILKKYRNMVVRACG	64

NAME	SEQUENCE	SEQ ID NO
	CH	
BMP9E2	REKRSAGAGSHCQKTSLRVNFEDIGWDSWIIAPKEYEAYECHGECPFPLADHLNSTNHAIVQTLVNSV NSKIPKACCVPTKLSPISVLYKDDMGVPTLKYHYEGMSVAECGCR	65
BMP9E6	SAGAGSHCQKTSLRVNFEDIGWDSWIIAPKEYEAYECDGECSFPLNAHMNATNHAIVQTLVHLMNPEY VPKPCCAPTKLSPISVLYKDDMGVPTLKYHYEGMSVAECGCR	66
BMP6-Short	VSSASDYNSSELKTACRKHELYVSFQDLGWQDWIIAPKGYAANYCDGECSFPLNAHMNATNHAIVQTL VHLMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRACGCH	67
BMP6-SA	VSSASDYNSSELKTACRKHELYVSFQDLGWQDWIIAPKGYAANYCDGECSFPLNAAMNATNHAIVQTL VHLMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRACGCH	68
BMP6-SL	VSSASDYNSSELKTACRKHELYVSFQDLGWQDWIIAPKGYAANYCDGECSFPLNAHLNATNHAIVQTL VHLMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRACGCH	69
BMP-E-NR	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCAPTELSAISMLYLDENEKVVLKNYQDMVVEGCGCR	70
BMP-GER-NR	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPRGYAAFYCDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCAPTKLRPMSMLYYDDGQNIIKKDIQNMIVEECGCS	71
BMP-E-NR-6	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIVAPPGYHAFYCDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRACGCH	72
BMP-GER-NR-6	QAKHKQRKRLKSSCKRHPLYVDFSDVGWNDWIIAPRGYAAFYCDGECSFPLNAHMNATNHAIVQTLVH LMNPEYVPKPCCAPTKLNAISVLYFDDNSNVILKKYRNMVVRACGCH	73

Although the above listed designer BMPs comprise embodiments of the invention, the invention is not limited in any way to any specific molecules. Instead, the invention encompasses any designer BMP comprising altered receptor binding where the designer BMP comprises at least one mutation within a type II receptor binding domain A, even more preferably, the designer BMP comprises at least one further mutation within a type I receptor binding domain, most preferably, the designer BMP comprises yet another at least one further mutation within a type II receptor binding domain B.

In other embodiments, the designer BMP of the present invention comprises an amino acid sequence at least about 70%, 75%, 80%, 85%, 87%, 90%, 92%, 95%, 96%, 97%, 98%, 99% or identical to one of the sequences described above. In another embodiment, the designer BMP comprises an amino acid sequence at least about 70%, 75%, 80%, 85%, 87%, 90%, 92%, 95%, 96%, 97%, 98%, 99% or identical to the sequence of SEQ ID NOs:8-73.

In yet another embodiment, the designer BMP comprises an amino acid sequence as set forth in any one of SEQ ID NOs:8-73. In another embodiment, the amino acid sequence of the designer BMP consists of one of the sequences of SEQ ID NOs:8-73.

Further, in one embodiment, the designer BMP comprises an amino acid sequence at least about 70%, 75%, 80%, 85%, 87%, 90%, 92%, 95%, 96%, 97%, 98%, 99% or identical to the sequence of SEQ ID NO:12. In another embodiment, the amino acid sequence is the sequence of SEQ ID NO:12. In yet another embodiment, the designer BMP is BMPE.

In an additional embodiment, the designer BMP comprises an amino acid sequence at least about 70%, 75%, 80%, 85%, 87%, 90%, 92%, 95%, 96%, 97%, 98%, 99% or identical to the sequence of

SEQ ID NO:14. In another embodiment, the amino acid sequence is the sequence of SEQ ID NO:14. In yet another embodiment, the designer BMP is BMPG.

In another embodiment, the designer BMP comprises an amino acid sequence at least about 70%, 75%, 80%, 85%, 87%, 90%, 92%, 95%, 96%, 97%, 98%, 99% or identical to the sequence of SEQ ID NO:36. In another embodiment, the amino acid sequence is the sequence of SEQ ID NO:36. In yet another embodiment, the designer BMP is BMPGE.

In another embodiment, the designer BMP comprises an amino acid sequence at least about 70%, 75%, 80%, 85%, 87%, 90%, 92%, 95%, 96%, 97%, 98%, 99% or identical to the sequence of SEQ ID NO:37. In another embodiment, the amino acid sequence is the sequence of SEQ ID NO:37. In yet another embodiment, the designer BMP is BMPGER.

A designer BMP of the invention may comprise a fragment of any one of the sequences described above. In an embodiment, a designer BMP fragment may comprise a fragment of at least an uninterrupted 20, 22, 24, 25, 26, 27, 28, 30, 32, 33, 34, 35, 36, 37, 38, 40, 41, 43, 44, 45, 47, 50, 53, 54, 56, 58, 60, 62, 66, 68, 70, 71, 74, 77, 80, 83, 85, 88, 90, 91, 93, 95, 97, 99, 100, 102, 105, 108, 110, 112, 115, 117, 119, 120, 121, 122, or 125 amino acid sequence from the sequence of any one of the sequences of SEQ ID NOs:8-73.

It is well known in the art that BMPs are often heterogeneous with respect to the amino and/or carboxyl termini of the protein. That is, the present invention comprises a designer BMP comprising an amino acid deletion/truncation at the amino and/or carboxyl terminus comprising a deletion of at least 10 amino acid residues, preferably, 9 amino acid residues, even more preferably, 8 amino acid residues, yet more preferably, 7 amino acid residues, preferably 6 amino acid residues, even more preferably, 5 amino acid residues, preferably 4 amino acid residues, more preferably 3 amino acid residues, even more preferably 2 amino acid residues, and most preferably 1 amino acid reside from the C and or N terminus of the designer BMP.

In another embodiment, the invention comprises a designer BMP protein comprising an amino acid sequence of any one of the sequences of SEQ ID NO:8-73 and further comprising a deletion/truncation from the amino and/or carboxyl termini of the protein. In another embodiment, the invention comprises a designer BMP protein derived from a BMP protein comprising an amino acid sequence of any of the sequences of SEQ ID NOs:8-73, wherein the protein comprises an amino acid deletion/truncation at the amino and/or carboxyl terminus comprising a deletion of at least 10 amino acid residues, preferably, 9 amino acid residues, even more preferably, 8 amino acid residues, yet more preferably, 7 amino acid residues, preferably 6 amino acid residues, even more preferably, 5 amino acid residues, preferably 4 amino acid residues, more preferably 3 amino acid residues, even more preferably 2 amino acid residues, and most preferably 1 amino acid reside from the C and or N terminus of the designer BMP protein amino acid sequence.

Structural design of BMPs with altered receptor affinity mediated by glycosylation

The data disclosed herein demonstrate that BMP2 homodimers produced in E. coli (referred to herein as "E. coli BMP2"), which are not glycosylated, are less active than glycosylated BMP2 produced in mammalian cells, such as CHO cells (referred to herein as "CHO BMP2"). In addition, data disclosed herein further demonstrate that E. coli produced BMP6 homodimers are essentially non-functional compared with BMP6 homodimers produced in mammalian cell culture.

The data disclosed herein demonstrate that there are significant variations in the crystal structure of E. coli BMP2 compared with CHO BMP2 in the type I receptor binding region.

In one embodiment, the designer BMP comprises an altered conformation mediated by glycosylation thereby affecting a binding motif that, in turn, mediates altered binding to a type I receptor. This is based on the present discovery that in mammalian (e.g., CHO) cell produced wild type BMP2, D53 points towards the receptor interface while the H54 points away from the receptor. This is in contrast to E. coli-produced BMP2 where the D53 residue points away from the receptor interface and the H54 residue lines up toward the receptor, stacking against a proline reside as illustrated in Figure 3, apparently acting as a "doorstop." In addition, the data disclosed herein demonstrate for the first time that CHO-produced BMP6, which is fully glycosylated and active, also comprises a histidine residue pointing toward the incoming receptor, i.e., a histidine "doorstop."

Without wishing to be bound by any particular theory, the data disclosed herein suggest, for the first time, that moving a "doorstop" residue away from the receptor interface, can mediate increased binding between the BMP ligand and its receptor. The data further demonstrate that the doorstop residue may be either mutated itself to remove the doorstop or other residues may be mutated to shift the position of the doorstop residue. Further, the data disclosed herein further demonstrate that other residues may be mutated to provide a "glycan tether" which then, in turn, can orient a glycan such that the tethering of the glycan will reorient the doorstop residue.

Therefore, in some embodiments, a designer BMP can be produced by incorporating at least one amino acid mutation that affects the glycan tether and/or removes a histidine doorstop structure thereby providing a designer BMP with altered receptor binding.

In summary, in some embodiments, the designer BMPs of the invention may comprise at least one mutation in the type I and/or type II binding domains of BMPs that confer altered type I and/or type II receptor binding. In one embodiment, the BMP sequence is engineered to alter the receptor affinity of BMPs in order to alter and improve the receptor binding and/or osteogenic activity of the engineered or "designer" BMP. In one embodiment, this engineering involves identifying the residues involved in type I and type II receptor binding and replacing them to create designer BMP molecules that show, among other things, higher affinity to both type I and type II receptors than the parental BMP from which the designer is derived.

In other embodiments, the designer BMPs of the invention comprise mutations that create a new arginine "glycan tether" or destroy an existing one to reshape the type I receptor binding domain. That is, the mutation to an arginine in the position two residues C-terminal from the first cysteine, equivalent to

R16 of BMP2, appears to cause the glycan chain to be "tethered" onto the BMP surface and consequently alter the conformation of the pre-helical loop region compared with the wild type BMP that lacks the mutation. In other embodiments, the designer BMP of the invention may comprise at least one mutation that alters, creates or destroys (abolishes) the "doorstop" residue that blocks type I receptor from further engagement with BMP. That is, the mutation of H54 in the designer BMP, or a corresponding equivalent residue thereof, that is oriented in such a way that it impedes or increases interaction of the designer BMP with a type I receptor.

In some embodiments, the amino acid mutation affects the conformation of the designer BMP such that the mutation mediates the creation and or abolishment of an arginine "glycan tether" otherwise present in the corresponding wild type BMP. In some embodiments, the mutation mediates an altered conformation which creates or removes/abolishes a histidine doorstop conformation in the designer BMP where such doorstop conformation is either not present or active, respectively, in the corresponding wild type BMP.

Therefore, the skilled artisan, once armed with the teachings provided herein, would appreciate that the presence or absence of an arginine "glycan tether" and/or a histidine "doorstop" in a TGFβ superfamily member may be assessed using any method known in the art for the structural analysis of proteins, including, but not limited to, the methods exemplified herein. Once the presence of a "doorstop" residue has been identified, then at least one mutation can be introduced into the molecule to reorient the histidine away from the receptor binding interface. Alternatively, a mutation can be introduced that will create or enhance a "glycan tether" such that the inhibitory effect of the histidine "doorstop", if present, is decreased or, more preferably, eliminated.

In one embodiment, where the TGFβ superfamily member is BMP2, the mutation that removes the histidine doorstop is substitution of another amino acid for H54. In some embodiments, the H54 is replaced with alanine, glycine, serine, or threonine.

Although the present invention discloses such "doorstop"-removing mutations for BMP2, the skilled artisan would understand, based on the knowledge in the art, how to identify corresponding mutations for other TGFβ superfamily members and readily produce mutants lacking a "doorstop," i.e., removing or reorienting a residue that would otherwise interfere with receptor binding by facing or projecting into the binding interface. The effects of the mutation on protein conformation can be determined using any art-recognized method for the structural analysis of proteins such as, but not limited to, those disclosed herein. Alternatively, mutations that can remove the doorstop and increase ligand binding to the type I receptor can be identified *in silico* using computer modeling methods available in the art. Therefore, the present invention encompasses the design of TGFβ superfamily members having improved binding with the type I receptor in that they lack a histidine "doorstop" residue that would otherwise be present in the receptor interface.

The present invention further provides the skilled artisan with the understanding of how to identify mutations for other TGFβ family members that would generate or destroy the arginine glycan tether.

Mutations that add the arginine glycan tether to a protein lacking the tether are contemplated by the instant invention. Therefore, the present invention encompasses the design of TGFβ superfamily members having improved binding with the type I receptor in that they contain an arginine glycan tether that alters the conformation of the type I receptor binding domain.

In some embodiments, the removal of the histidine doorstop thereby removing the requirement of a glycan tether, provides a designer BMP that can be produced without glycosylation while maintaining biological activity. For example, designer BMPs may be produced in cells with glycosylation activity that differs from mammalian cells or is not present, such as bacterial cells, yeast cells, insect cells, or slime mold cells. In particular embodiments, the designer BMPs may be produced in E. coli and maintain biological activity.

Thus, in some embodiments, the invention provides methods for designing and producing BMPs that can be produced in cells either lacking glycosylation or comprising altered glycosylation such that an altered glycan is produced which differs from that produced by a mammalian cell. That is, the present invention encompasses methods for introducing a mutation that removes a doorstop residue that would otherwise impair or inhibit receptor binding. The skilled artisan would understand once provided with the teachings of the invention that a doorstop residue that impinges upon the receptor-ligand interface may be mutated to entirely remove the residue or other mutations can be introduced such that the residue is oriented away from the interface. Such other mutations include, but are not limited to, providing a glycan tether that will alter the conformation of a glycan and thereby alter the conformation of the ligand such that the doorstop residue is orientated away from the binding interface.

Nucleic Acids Encoding Designer BMPs

The invention also includes nucleic acids encoding designer the BMPs described herein. Nucleic acids encoding the designer BMPs described herein can be prepared according to a wide plethora of methods known in the art.

In one, nucleic acids encoding designer BMPs are prepared by total gene synthesis, or by site-directed mutagenesis of a nucleic acid encoding wild type or modified BMPs. Methods including template-directed ligation, recursive PCR, cassette mutagenesis, site-directed mutagenesis or other techniques that are well known in the art may be utilized (see for example Strizhov et al., *Proc. Natl. Acad. Sci. USA* 93:15012-15017 (1996); Prodromou and Perl, *Prot. Eng.* 5: 827-829 (1992); Jayaraman and Puccini, *Biotechniques* 12: 392-398 (1992); and Chalmers et al., *Biotechniques* 30: 249-252 (2001)).

Thus, embodiments of the present invention can comprise nucleic acid molecules that encode the designer BMPs of the present invention. In certain embodiments, the invention provides a nucleic acid molecule that encodes for one of the amino acid sequences of SEQ ID NOs:8 to 66.

In other embodiments, the nucleic acid molecule encodes a designer BMP protein that comprises an amino acid sequence at least 70%, 75%, 80%, 85%, 87%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identical to the amino acid sequence of SEQ ID NO:12. In some embodiments, the nucleic

acid molecule encodes a designer BMP protein that comprises the amino acid sequence of SEQ ID NO:12. In another embodiment, the nucleic acid molecule encodes the amino acid sequence of BMPE as set forth in Table 8.

In other embodiments, the nucleic acid molecule encodes a designer BMP protein that comprises an amino acid sequence at least 70%, 75%, 80%, 85%, 87%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identical to the amino acid sequence of SEQ ID NO:14. In some embodiments, the nucleic acid molecule encodes a designer BMP protein that comprises the amino acid sequence of SEQ ID NO:14. In another embodiment, the nucleic acid molecule encodes the amino acid sequence of BMPG as set forth in Table 8.

In other embodiments, the nucleic acid molecule encodes a designer BMP protein that comprises an amino acid sequence at least 70%, 75%, 80%, 85%, 87%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identical to the amino acid sequence of SEQ ID NO:36. In some embodiments, the nucleic acid molecule encodes a designer BMP protein that comprises the amino acid sequence of SEQ ID NO:36. In another embodiment, the nucleic acid molecule encodes the amino acid sequence of BMPGE as set forth in Table 8.

In other embodiments, the nucleic acid molecule encodes a designer BMP protein that comprises an amino acid sequence at least 70%, 75%, 80%, 85%, 87%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identical to the amino acid sequence of SEQ ID NO:37. In some embodiments, the nucleic acid molecule encodes a designer BMP protein that comprises the amino acid sequence of SEQ ID NO:37. In another embodiment, the nucleic acid molecule encodes the amino acid sequence of BMPGER as set forth in Table 8.

Exemplary nucleotide sequences encoding designer BMPs are set forth in Table 8, below. Table 8 shows the name of the protein encoded and the nucleotide sequence encoding that protein. In general, the mature protein coding sequence begins at nucleotide 847 of the sequences listed below.

TABLE 8

NAME	SEQUENCE	SEQ ID NO
BMP-A	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTATTGCACCCA AGGGCTATGCTGCCAATTACTGCCACGGAGAATGCCCTTTTCCTCTGGCTGATCATCTGAACTCCACT AATCATGCCATTGTTCAGACGTTGGTCAACTCTGTTAACTCTAAGATTCCTAAGGCATGCTGTGTCCC GACAGAACTCAGTGCTATCTCGATGCTGTACCTTGACGAGAATGAAAAGGTTGTATTAAAGAACTATC AGGACATGGTTGTGGAGGGTTGTGGGTGTCGCTGA	74

NAME	SEQUENCE	SEQ ID NO
BMP-B	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTGTGGCTCCCC CGGGGTATCACGCCTTTTACTGCCACGGAGAATGCCCTTTTCCTCTGGCTGATCATCTGAACTCCACT AATCATGCCATTGTTCAGACGTTGGTCAACTCTGTTAACTCTAAGATTCCTAAGGCATGCTGTGTCCC GACAAAGCTAAATGCCATCTCGGTTCTTTACTTTGATGACAACTCCAATGTCATTTTAAAGAACTATC AGGACATGGTTGTGGAGGGTTGTGGGTGTCGCTGA	75
BMP-C	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTATTGCACCCA AGGGCTATGCTGCCAATTACTGCCACGGAGAATGCCCTTTTCCTCTGGCTGATCATCTGAACTCCACT AATCATGCCATTGTTCAGACGTTGGTCAACTCTGTTAACTCTAAGATTCCTAAGGCATGCTGTGTCCC GACAAAGCTAAATGCCATCTCGGTTCTTTACTTTGATGACAACTCCAATGTCATTTTAAAGAACTATC AGGACATGGTTGTGGAGGGTTGTGGGTGTCGCTGA	76
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BMP-G	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTATTGCACCCA AGGGCTATGCTGCCTTTTACTGCCACGGAGAATGCCCTTTTCCTCTGGCTGATCATCTGAACTCCACT AATCATGCCATTGTTCAGACGTTGGTCAACTCTGTTAACTCTAAGATTCCTAAGGCATGCTGTGTCCC GACAGAACTCAGTGCTATCTCGATGCTGTACCTTGACGAGAATGAAAAGGTTGTATTAAAGAACTATC AGGACATGGTTGTGGAGGGTTGTGGGTGTCGCTGA	80
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BMP-K	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTGTGGCTCCCC CGGGGTATCACGCCTTTTACTGCCACGGAGAATGCCCTTTTCCTCTGGCTGATCATCTGAACTCCACT AAACATGCCATTGTTCAGACGTTGGTCAACTCTGTTAACTCTAAGATTCCTAAGGCATGCTGTGTCCC GACAGAACTCAGTGCTATCTCGATGCTGTACCTTGACGAGAATGAAAAGGTTGTATTAAAGAACTATC AGGACATGGTTGTGGAGGGTTGTGGGTGTCGCTAG	84
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BMP-AK	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTATTGCACCCA AGGGCTATGCTGCCAATTACTGCCACGGAGAATGCCCTTTTCCTCTGGCTGATCATCTGAACTCCACT AAACATGCCATTGTTCAGACGTTGGTCAACTCTGTTAACTCTAAGATTCCTAAGGCATGCTGTGTCCC GACAGAACTCAGTGCTATCTCGATGCTGTACCTTGACGAGAATGAAAAGGTTGTATTAAAGAACTATC AGGACATGGTTGTGGAGGGTTGTGGGTGTCGCTGA	88
BMP-AT	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTATTGCACCCA AGGGCTATGCTGCCAATTACTGCCACGGAGAATGCCCTTTTCCTCTGGCTGATCATCTGAACTCCACT ACTCATGCCATTGTTCAGACGTTGGTCAACTCTGTTAACTCTAAGATTCCTAAGGCATGCTGTGTCCC GACAGAACTCAGTGCTATCTCGATGCTGTACCTTGACGAGAATGAAAAGGTTGTATTAAAGAACTATC AGGACATGGTTGTGGAGGGTTGTGGGTGTCGCTGA	89
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BMP-E10	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTGTGGCTCCCC CGGGGTATCACGCCTTTTACTGCCGTGGTGTTTGTAACTACCCCCTGGCAGAGCATCTCACACCCACA AAGCATGCAATTATCCAGGCCTTGGTCCACCTCAAGAATTCCCAGAAAGCTTCCAAAGCCTGCTGTGT CCCGACAGAACTCAGTGCTATCTCGATGCTGTACCTTGACGAGAATGAAAAGGTTGTATTAAAGAACT ATCAGGACATGGTTGTGGAGGGTTGTGGGTGTCGCTAG	92
BMP-EK	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTGTGGCTCCCC CGGGGTATCACGCCTTTTACTGCGATGGAGAATGCTCCTTCCCACTCAACGCACACATGAATGCAACC AAACACGCGATTGTGCAGACCTTGGTTCACCTTATGAACCCCGAGTATGTCCCCAAACCGTGCTGTGC	93

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BMP-R	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTGTGGCTCCCA GGGGGTATCACGCCTTTTACTGCCACGGAGAATGCCCTTTTCCTCTGGCTGATCATCTGAACTCCACT AATCATGCCATTGTTCAGACGTTGGTCAACTCTGTTAACTCTAAGATTCCTAAGGCATGCTGTGTCCC GACAGAACTCAGTGCTATCTCGATGCTGTACCTTGACGAGAATGAAAAGGTTGTATTAAAGAACTATC AGGACATGGTTGTGGAGGGTTGTGGGTGTCGCTAG	95
BMP-G5	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTGTGGCTCCCC CGGGGTATCACGCCTTTTACTGCGAGGGGCTGTGCGAGTTCCCATTGCGCTCCCACCTGGAGCCCACG AATCATGCAGTCATCCAGACCCTGATGAACTCCATGGACCCCGAGTCCACACCACCCACCTGCTGTGT CCCGACAGAACTCAGTGCTATCTCGATGCTGTACCTTGACGAGAATGAAAAGGTTGTATTAAAGAACT ATCAGGACATGGTTGTGGAGGGTTGTGGGTGTCGCTAG	96
BMP-ER	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA	97

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BMP-GR	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTATTGCACCCA GGGGCTATGCTGCCTTTTACTGCCACGGAGAATGCCCTTTTCCTCTGGCTGATCATCTGAACTCCACT AATCATGCCATTGTTCAGACGTTGGTCAACTCTGTTAACTCTAAGATTCCTAAGGCATGCTGTGTCCC GACAGAACTCAATGCCATCTCGGTTCTTTACTTTGATGACAACTCCAATGTCATTTTAAAGAACTATC AGGACATGGTTGTGGAGGGTTGTGGGTGTCGCTGA	99
BMP-GK	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTATTGCACCCA AGGGCTATGCTGCCTTTTACTGCCACGGAGAATGCCCTTTTCCTCTGGCTGATCATCTGAACTCCACT AAACATGCCATTGTTCAGACGTTGGTCAACTCTGTTAACTCTAAGATTCCTAAGGCATGCTGTGTCCC GACAGAACTCAATGCCATCTCGGTTCTTTACTTTGATGACAACTCCAATGTCATTTTAAAGAACTATC AGGACATGGTTGTGGAGGGTTGTGGGTGTCGCTGA	100
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BMP-GER	ATGGTGGCTGGCACCAGATGTCTGCTGGCCCTGCTGCTGCCCCAGGTGCTGCTGGGCGGAGCTGCTGG ACTGGTGCCCGAGCTGGGCAGAAGAAAGTTCGCCGCTGCCTCCTCTGGCCGGCCTTCCAGCCAGCCTT CCGACGAGGTGCTGTCCGAGTTCGAGCTGCGGCTGCTGTCCATGTTCGGCCTGAAGCAGCGGCCCACC CCTTCTAGGGACGCCGTGGTGCCCCCCTACATGCTGGACCTGTACCGGCGGCACTCCGGCCAGCCTGG ATCTCCTGCCCCCGACCACAGACTGGAAAGAGCCGCCTCCCGGGCCAACACCGTGCGGTCTTTCCACC ACGAGGAATCCCTGGAAGAACTGCCCGAGACATCCGGCAAGACCACCCGGCGGTTCTTTTTCAACCTG TCATCCATCCCCACCGAAGAGTTCATCACCTCCGCCGAGCTGCAGGTGTTCCGCGAGCAGATGCAGGA CGCCCTGGGCAACAACTCCTCCTTCCACCACCGGATCAACATCTACGAGATCATCAAGCCCGCCACCG CCAACTCCAAGTTCCCCGTGACCCGGCTGCTGGACACCCGGCTGGTGAACCAGAACGCCTCCAGATGG GAGTCCTTCGACGTGACCCCTGCCGTGATGAGATGGACCGCCCAGGGCCACGCCAACCACGGCTTTGT GGTGGAAGTGGCCCACCTGGAAGAGAAGCAGGGCGTGTCCAAGCGGCACGTGCGGATCTCTCGGTCCC TGCACCAGGACGAGCACAGCTGGTCCCAGATCCGGCCCCTGCTGGTGACATTCGGCCACGATGGCAAG GGCCACCCCCTGCACAAGAGAGAGAAGCGGCAGGCCAAGCACAAGCAGCGGAAGCGGCTGAAGTCCTC CTGCAAGCGGCACCCCCTGTACGTGGACTTCTCCGACGTGGGCTGGAACGACTGGATYRTKGCYCCCA GGGGSTAYSMCGCCTTYTACTGCGACGGCGAGTGCTCCTTCCCCCTGAACGCCCACATGAACGCCACC AACCACGCCATCGTGCAGACCCTGGTGCACCTGATGAACCCCGAGTACGTGCCCAAGCCTTGTTGCGC CCCCACCGAGCTGAACGCCATCTCCGTGCTGTACTTCGACGACAACTCCAACGTGATCCTGAAGAACT ACCAGGACATGGTGGTCGAAGGCTGCGGCTGTAGATGA	103
BMP-JP	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTATTGCTCCCC CGGGGTATCACGCCTTTTACTGCGATGGAGAATGCTCCTTCCCACTCAACGCACACATGAATGCAACC AACCACGCGATTGTGCAGACCTTGGTTCACCTTATGAACCCCGAGTATGTCCCCAAACCGTGCTGTGC GCCGACAGAACTCAATGCTATCTCGGTTCTGTACTTTGACGAGAATTCCAATGTTGTATTAAAGAAAT ATCAGGACATGGTTGTGAGAGGTTGTGGGTGTCGCTGA	104
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BMP-JT	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTATTGCTCCCA AGGGGTATCACGCCTTTTACTGCGATGGAGAATGCTCCTTCCCACTCAACGCACACATGAATGCAACC ACCCACGCGATTGTGCAGACCTTGGTTCACCTTATGAACCCCGAGTATGTCCCCAAACCGTGCTGTGC GCCGACAGAACTCAATGCTATCTCGGTTCTGTACTTTGACGAGAATTCCAATGTTGTATTAAAGAAAT ATCAGGACATGGTTGTGAGAGGTTGTGGGTGTCGCTGA	107
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BMP-E9B	ATGGTGGCTGGCACCAGATGTCTGCTGGCCCTGCTGCTGCCCCAGGTGCTGCTGGGCGGAGCTGCTGG ACTGGTGCCCGAGCTGGGCAGAAGAAAGTTCGCCGCTGCCTCCTCTGGCCGGCCTTCCAGCCAGCCTT CCGACGAGGTGCTGTCCGAGTTCGAGCTGCGGCTGCTGTCCATGTTCGGCCTGAAGCAGCGGCCCACC CCTTCTAGGGACGCCGTGGTGCCCCCCTACATGCTGGACCTGTACCGGCGGCACTCCGGACAGCCTGG ATCTCCTGCCCCCGACCACAGACTGGAAAGAGCCGCCTCCCGGGCCAACACCGTGCGGTCTTTCCACC ACGAGGAATCCCTGGAAGAACTGCCCGAGACATCCGGCAAGACCACCCGGCGGTTCTTTTTCAACCTG TCCTCCATCCCCACCGAAGAGTTCATCACCTCCGCCGAGCTGCAGGTGTTCCGCGAGCAGATGCAGGA CGCCCTGGGCAACAACTCCTCCTTCCACCATCGGATCAACATCTACGAGATCATCAAGCCCGCCACCG CCAACTCCAAGTTCCCCGTGACCCGGCTGCTGGACACCCGGCTGGTGAACCAGAACGCCTCCAGATGG GAGTCCTTCGACGTGACCCCTGCCGTGATGAGATGGACCGCCCAGGGCCACGCCAACCACGGCTTTGT GGTGGAAGTGGCCCACCTGGAAGAGAAGCAGGGCGTGTCCAAGCGGCACGTGCGGATCTCTCGGTCCC TGCACCAGGACGAGCACAGCTGGTCCCAGATCCGGCCCCTGCTGGTGACATTCGGCCACGATGGCAAG GGCCACCCCCTGCACAAGAGAGAGAAGCGGCAGGCCAAGCACAAGCAGCGGAAGCGGCTGAAGTCCTC CTGCAAGCGGCACCCCCTGTACGTGGACTTCTCCGACGTGGGCTGGAACGACTGGATCGTGGCCCCTC CCGGCTACCACGCCTTCTACTGCGACGGCGAGTGCTCCTTCCCCCTGAACGCCCACATGAACGCCACC AACCACGCCATCGTGCAGACCCTGGTGCACCTGATGAACCCCGAGTACGTGCCCAAGCCCTGCTGCGC CCCCACCGAGCTGTCCCCCATCTCCGTGCTGTACAAGGACGACATGGGCGTGCCCACCCTGAAGAACT ACCAGGACATGGTGGTCGAAGGCTGCGGCTGTCGGTGA	110
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BMP-QAK no SAGA	ATGTGTCCTGGCGCTCTGTGGGTGGCCCTGCCTCTGCTGTCTCTGCTGGCCGGCAGCCTGCAGGGCAA GCCTCTGCAGTCCTGGGGCAGAGGCTCCGCTGGCGGCAATGCTCACAGCCCTCTGGGAGTGCCTGGCG GCGGACTGCCCGAGCACACCTTCAACCTGAAGATGTTCCTGGAAAACGTGAAGGTGGACTTCCTGCGG TCCCTGAACCTGTCCGGCGTGCCCAGCCAGGACAAGACCCGGGTGGAACCCCCCCAGTACATGATCGA CCTGTACAACCGGTACACCTCCGACAAGTCCACCACCCCCGCCTCCAACATCGTGCGGTCCTTCAGCA TGGAAGATGCCATCTCCATTACCGCCACCGAGGACTTCCCATTTCAGAAGCACATCCTGCTGTTCAAC ATCTCCATCCCCCGGCACGAGCAGATCACCAGAGCCGAGCTGCGGCTGTACGTGTCCTGCCAGAACCA CGTGGACCCCTCCCACGACCTGAAGGGCTCCGTGGTGATCTACGACGTGCTGGACGGCACCGACGCCT GGGACTCCGCTACCGAGACAAAGACCTTCCTGGTGTCCCAGGATATCCAGGACGAGGGCTGGGAGACA CTGGAAGTGTCCTCCGCCGTGAAGAGATGGGTGCGATCCGACTCCACCAAGTCCAAGAACAAGCTGGA AGTGACCGTGGAATCCCACCGGAAGGGCTGCGACACCCTGGACATCTCCGTGCCCCCTGGCTCCCGGA ACCTGCCCTTCTTCGTGGTGTTCTCCAACGACCACTCCTCCGGCACCAAAGAGACACGGCTGGAACTG AGAGAGATGATCTCCCACGAGCAGGAATCCGTCCTGAAGAAGCTGTCCAAGGACGGCTCCACCGAGGC CGGCGAGTCCTCTCACGAAGAGGACACCGACGGCCACGTGGCAGCTGGCTCTACCCTGGCCAGACGGA AGCGGCAGGCCAAGCACAAGCAGCGGAAGCGGCTGAAGTCCAGCTGCCAGAAAACCTCCCTGAGAGTG AACTTCGAGGACATCGGCTGGGACAGCTGGATCATTGCCCCCAAAGAGTACGAGGCCTACGAGTGCAA GGGCGGCTGCTTCTTCCCCCTGGCCGACGACGTGACCCCCACCGAAGCACGCCATCGTGCAGACCCTGG TGCACCTGAAGTTCCCCACCAAAGTGGGCAAGGCCTGCTGCGTGCCCACCAAGCTGTCCCCCATCAGC GTGCTGTACAAGGACGACATGGGCGTGCCAACCCTGAAGTACCACTACGAGGGCATGTCCGTGGCCGA GTGTGGCTGCCGGTGA	114
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BMP6-SA	ATGCCGGGGCTGGGGCGGAGGGCGCAGTGGCTGTGCTGGTGGTGGGGGCTGCTGTGCAGCTGCTGCGG GCCCCCGCCGCTGCGGCCGCCCTTGCCCGCTGCCGCGGCCGCCGCCGCCGGGGGGCAGCTGCTGGGGG ACGGCGGGAGCCCCGGCCGCACGGAGCAGCCGCCGCCGTCGCCGCAGTCCTCCTCGGGCTTCCTGTAC CGGCGGCTCAAGACGCAGGAGAAGCGGGAGATGCAGAAGGAGATCTTGTCGGTGCTGGGGCTCCCGCA CCGGCCCCGGCCCCTGCACGGCCTCCAACAGCCGCAGCCCCCGGCGCTCCGGCAGCAGGAGGAGCAGC AGCAGCAGCAGCAGCTGCCTCGCGGAGAGCCCCCTCCCGGGCGACTGAAGTCCGCGCCCCTCTTCATG CTGGATCTGTACAACGCCCTGTCCGCCGACAACGACGAGGACGGGGCGTCGGAGGGGGAGAGGCAGCA GTCCTGGCCCCACGAAGCAGCCAGCTCGTCCCAGCGTCGGCAGCCGCCCCCGGGCGCCGCGCACCCGC TCAACCGCAAGAGCCTTCTGGCCCCCGGATCTGGCAGCGGCGGCGCGTCCCCACTGACCAGCGCGCAG GACAGCGCCTTCCTCAACGACGCGGACATGGTCATGAGCTTTGTGAACCTGGTGGAGTACGACAAGGA GTTCTCCCCTCGTCAGCGACACCACAAAGAGTTCAAGTTCAACTTATCCCAGATTCCTGAGGGTGAGG	134

NAME	SEQUENCE	SEQ ID NO
	TGGTGACGGCTGCAGAATTCCGCATCTACAAGGACTGTGTTATGGGGAGTTTTAAAAACCAAACTTTT CTTATCAGCATTTATCAAGTCTTACAGGAGCATCAGCACAGAGACTCTGACCTGTTTTTGTTGGACAC CCGTGTAGTATGGGCCTCAGAAGAAGGCTGGCTGGAATTTGACATCACGGCCACTAGCAATCTGTGGG TTGTGACTCCACAGCATAACATGGGGCTTCAGCTGAGCGTGGTGACAAGGGATGGAGTCCACGTCCAC CCCCGAGCCGCAGGCCTGGTGGGCAGAGACGGCCCTTACGATAAGCAGCCCTTCATGGTGGCTTTCTT CAAAGTGAGTGAGGTCCACGTGCGCACCACCAGGTCAGCCTCCAGCCGGCGCCGACAACAGAGTCGTA ATCGCTCTACCCAGTCCCAGGACGTGGCGCGGGTCTCCAGTGCTTCAGATTACAACAGCAGTGAATTG AAAACAGCCTGCAGGAAGCATGAGCTGTATGTGAGTTTCCAAGACCTGGGATGGCAGGACTGGATCAT TGCACCCAAGGGCTATGCTGCCAATTACTGTGATGGAGAATGCTCCTTCCCACTCAACGCAGCCATGA ATGCAACCAACCACGCGATTGTGCAGACCTTGGTTCACCTTATGAACCCCGAGTATGTCCCCAAACCG TGCTGTGCGCCAACTAAGCTAAATGCCATCTCGGTTCTTTACTTTGATGACAACTCCAATGTCATTCT GAAAAAATACAGGAATATGGTTGTAAGAGCTTGTGGATGCCACTAA	
MBP6-SL	ATGCCGGGGCTGGGGCGGAGGGCGCAGTGGCTGTGCTGGTGGTGGGGGCTGCTGTGCAGCTGCTGCGG GCCCCCGCCGCTGCGGCCGCCCTTGCCCGCTGCCGCGGCCGCCGCCGCCGGGGGGCAGCTGCTGGGGG ACGGCGGGAGCCCCGGCCGCACGGAGCAGCCGCCGCCGTCGCCGCAGTCCTCCTCGGGCTTCCTGTAC CGGCGGCTCAAGACGCAGGAGAAGCGGGAGATGCAGAAGGAGATCTTGTCGGTGCTGGGGCTCCCGCA CCGGCCCCGGCCCCTGCACGGCCTCCAACAGCCGCAGCCCCCGGCGCTCCGGCAGCAGGAGGAGCAGC AGCAGCAGCAGCAGCTGCCTCGCGGAGAGCCCCCTCCCGGGCGACTGAAGTCCGCGCCCCTCTTCATG CTGGATCTGTACAACGCCCTGTCCGCCGACAACGACGAGGACGGGGCGTCGGAGGGGGAGAGGCAGCA GTCCTGGCCCCACGAAGCAGCCAGCTCGTCCCAGCGTCGGCAGCCGCCCCGGGCGCCGCGCACCCGC TCAACCGCAAGAGCCTTCTGGCCCCCGGATCTGGCAGCGGCGGCGCGTCCCCACTGACCAGCGCGCAG GACAGCGCCTTCCTCAACGACGCGGACATGGTCATGAGCTTTGTGAACCTGGTGGAGTACGACAAGGA GTTCTCCCCTCGTCAGCGACACCACAAAGAGTTCAAGTTCAACTTATCCCAGATTCCTGAGGGTGAGG TGGTGACGGCTGCAGAATTCCGCATCTACAAGGACTGTGTTATGGGGAGTTTTAAAAACCAAACTTTT CTTATCAGCATTTATCAAGTCTTACAGGAGCATCAGCACAGAGACTCTGACCTGTTTTTGTTGGACAC CCGTGTAGTATGGGCCTCAGAAGAAGGCTGGCTGGAATTTGACATCACGGCCACTAGCAATCTGTGGG TTGTGACTCCACAGCATAACATGGGGCTTCAGCTGAGCGTGGTGACAAGGGATGGAGTCCACGTCCAC CCCCGAGCCGCAGGCCTGGTGGGCAGAGACGGCCCTTACGATAAGCAGCCCTTCATGGTGGCTTTCTT CAAAGTGAGTGAGGTCCACGTGCGCACCACCAGGTCAGCCTCCAGCCGGCGCCGACAACAGAGTCGTA ATCGCTCTACCCAGTCCCAGGACGTGGCGCGGGTCTCCAGTGCTTCAGATTACAACAGCAGTGAATTG AAAACAGCCTGCAGGAAGCATGAGCTGTATGTGAGTTTCCAAGACCTGGGATGGCAGGACTGGATCAT TGCACCCAAGGGCTATGCTGCCAATTACTGTGATGGAGAATGCTCCTTCCCACTCAACGCACACCTCA ATGCAACCAACCACGCGATTGTGCAGACCTTGGTTCACCTTATGAACCCCGAGTATGTCCCCAAACCG TGCTGTGCGCCAACTAAGCTAAATGCCATCTCGGTTCTTTACTTTGATGACAACTCCAATGTCATTCT GAAAAAATACAGGAATATGGTTGTAAGAGCTTGTGGATGCCACTAA	135
BMP-E-NR	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTGTGGCTCCCC CGGGGTATCACGCCTTTTACTGCGATGGAGAATGCTCCTTCCCACTCAACGCACACATGAATGCAACC AACCACGCGATTGTGCAGACCTTGGTTCACCTTATGAACCCCGAGTATGTCCCCAAACCGTGCTGTGC GCCCACCAAGCTGAGACCCATGTCCATGTTGTACTATGATGATGGTCAAAACATCATCAAAAAGGACA TTCAGAACATGATCGTGGAGGAGTGTGGGTGCTCATAG	136
BMP-GER-NR	ATGGTGGCTGGCACCAGATGTCTGCTGGCCCTGCTGCTGCCCCAGGTGCTGCTGGGCGGAGCTGCTGG ACTGGTGCCCGAGCTGGGCAGAAGAAAGTTCGCCGCTGCCTCCTCTGGCCGGCCTTCCAGCCAGCCTT CCGACGAGGTGCTGTCCGAGTTCGAGCTGCGGCTGCTGTCCATGTTCGGCCTGAAGCAGCGGCCCACC CCTTCTAGGGACGCCGTGGTGCCCCCCTACATGCTGGACCTGTACCGGCGGCACTCCGGCCAGCCTGG ATCTCCTGCCCCCGACCACAGACTGGAAAGAGCCGCCTCCCGGGCCAACACCGTGCGGTCTTTCCACC ACGAGGAATCCCTGGAAGAACTGCCCGAGACATCCGGCAAGACCACCCGGCGGTTCTTTTTCAACCTG TCATCCATCCCCACCGAAGAGTTCATCACCTCCGCCGAGCTGCAGGTGTTCCGCGAGCAGATGCAGGA CGCCCTGGGCAACAACTCCTCCTTCCACCACCGGATCAACATCTACGAGATCATCAAGCCCGCCACCG CCAACTCCAAGTTCCCCGTGACCCGGCTGCTGGACACCCGGCTGGTGAACCAGAACGCCTCCAGATGG GAGTCCTTCGACGTGACCCCTGCCGTGATGAGATGGACCGCCCAGGGCCACGCCAACCACGGCTTTGT GGTGGAAGTGGCCCACCTGGAAGAGAAGCAGGGCGTGTCCAAGCGGCACGTGCGGATCTCTCGGTCCC TGCACCAGGACGAGCACAGCTGGTCCCAGATCCGGCCCCTGCTGGTGACATTCGGCCACGATGGCAAG GGCCACCCCCTGCACAAGAGAGAGAAGCGGCAGGCCAAGCACAAGCAGCGGAAGCGGCTGAAGTCCTC CTGCAAGCGGCACCCCCTGTACGTGGACTTCTCCGACGTGGGCTGGAACGACTGGATCATTGCCCCCA GGGGCTACGCCGCCTTCTACTGCGACGGCGAGTGCTCCTTCCCCCTGAACGCCCACATGAACGCCACC	137

NAME	SEQUENCE	SEQ ID NO
	AACCACGCCATCGTGCAGACCCTGGTGCACCTGATGAACCCCGAGTACGTGCCCAAGCCTTGTTGCGC CCCCACCAAGCTGAGACCCATGTCCATGTTGTACTATGATGATGGTCAAAACATCATCAAAAAGGACA TTCAGAACATGATCGTGGAGGAGTGTGGGTGCTCATAG	
BMP-E-NR-6	ATGGTGGCCGGGACCCGCTGTCTTCTAGCGTTGCTGCTTCCCCAGGTCCTCCTGGGCGGCGCGGCTGG CCTCGTTCCGGAGCTGGGCCGCAGGAAGTTCGCGGCGGCGTCGTCGGGCCGCCCCTCATCCCAGCCCT CTGACGAGGTCCTGAGCGAGTTCGAGTTGCGGCTGCTCAGCATGTTCGGCCTGAAACAGAGACCCACC CCCAGCAGGGACGCCGTGGTGCCCCCCCTACATGCTAGACCTGTATCGCAGGCACTCAGGTCAGCCGGG CTCACCCGCCCCAGACCACCGGTTGGAGAGGGCAGCCAGCCGAGCCAACACTGTGCGCAGCTTCCACC ATGAAGAATCTTTGGAAGAACTACCAGAAACGAGTGGGAAAACAACCCGGAGATTCTTCTTTAATTTA AGTTCTATCCCCACGGAGGAGTTTATCACCTCAGCAGAGCTTCAGGTTTTCCGAGAACAGATGCAAGA TGCTTTAGGAAACAATAGCAGTTTCCATCACCGAATTAATATTTATGAAATCATAAAACCTGCAACAG CCAACTCGAAATTCCCCGTGACCAGACTTTTGGACACCAGGTTGGTGAATCAGAATGCAAGCAGGTGG GAAAGTTTTGATGTCACCCCCGCTGTGATGCGGTGGACTGCACAGGGACACGCCAACCATGGATTCGT GGTGGAAGTGGCCCACTTGGAGGAGAAACAAGGTGTCTCCAAGAGACATGTTAGGATAAGCAGGTCTT TGCACCAAGATGAACACAGCTGGTCACAGATAAGGCCATTGCTAGTAACTTTTGGCCATGATGGAAAA GGGCATCCTCTCCACAAAAGAGAAAAACGTCAAGCCAAACACAAACAGCGGAAACGCCTTAAGTCCAG CTGTAAGAGACACCCTTTGTACGTGGACTTCAGTGACGTGGGGTGGAATGACTGGATTGTGGCTCCCC CGGGGTATCACGCCTTTTACTGCGATGGAGAATGCTCCTTCCCACTCAACGCACACATGAATGCAACC AACCACGCGATTGTGCAGACCTTGGTTCACCTTATGAACCCCGAGTATGTCCCCAAACCGTGCTGTGC GCCAACTAAGCTAAATGCCATCTCGGTTCTTTACTTTGATGACAACTCCAATGTCATTCTGAAAAAAT ACAGGAATATGGTTGTAAGAGCTTGTGGATGCCACTAA	138
BMP-GER-NR-6	ATGGTGGCTGGCACCAGATGTCTGCTGGCCCTGCTGCTGCCCCAGGTGCTGCTGGGCGGAGCTGCTGG ACTGGTGCCCGAGCTGGGCAGAAGAAAGTTCGCCGCTGCCTCCTCTGGCCGGCCTTCCAGCCAGCCTT CCGACGAGGTGCTGTCCGAGTTCGAGCTGCGGCTGCTGTCCATGTTCGGCCTGAAGCAGCGGCCCACC CCTTCTAGGGACGCCGTGGTGCCCCCCCTACATGCTGGACCTGTACCGGCGGCACTCCGGCCAGCCTGG ATCTCCTGCCCCCGACCACAGACTGGAAAGAGCCGCCTCCCGGGCCAACACCGTGCGGTCTTTCCACC ACGAGGAATCCCTGGAAGAACTGCCCGAGACATCCGGCAAGACCACCCGGCGGTTCTTTTTCAACCTG TCATCCATCCCCACCGAAGAGTTCATCACCTCCGCCGAGCTGCAGGTGTTCCGCGAGCAGATGCAGGA CGCCCTGGGCAACAACTCCTCCTTCCACCACCGGATCAACATCTACGAGATCATCAAGCCCGCCACCG CCAACTCCAAGTTCCCCGTGACCCGGCTGCTGGACACCCGGCTGGTGAACCAGAACGCCTCCAGATGG GAGTCCTTCGACGTGACCCCTGCCGTGATGAGATGGACCGCCCAGGGCCACGCCAACCACGGCTTTGT GGTGGAAGTGGCCCACCTGGAAGAGAAGCAGGGCGTGTCCAAGCGGCACGTGCGGATCTCTCGGTCCC TGCACCAGGACGAGCACAGCTGGTCCCAGATCCGGCCCCTGCTGGTGACATTCGGCCACGATGGCAAG GGCCACCCCCTGCACAAGAGAGAGAAGCGGCAGGCCAAGCACAAGCAGCGGAAGCGGCTGAAGTCCTC CTGCAAGCGGCACCCCCTGTACGTGGACTTCTCCGACGTGGGCTGGAACGACTGGATCATTGCCCCCA GGGGCTACGCCGCCTTCTACTGCGACGGCGAGTGCTCCTTCCCCCTGAACGCCCACATGAACGCCACC AACCACGCCATCGTGCAGACCCTGGTGCACCTGATGAACCCCGAGTACGTGCCCAAGCCTTGTTGCGC CCCAACTAAGCTAAATGCCATCTCGGTTCTTTACTTTGATGACAACTCCAATGTCATTCTGAAAAAAT ACAGGAATATGGTTGTAAGAGCTTGTGGATGCCACTAA	139

In other embodiments, the nucleic acid molecule encoding a designer BMP comprises a nucleic acid sequence at least 40%, 50%, 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identical to one of the nucleic acid sequences set forth in SEQ ID NOs:74-139 or a fragment thereof. In other embodiments, the nucleic acid molecule encoding a designer BMP comprises a nucleic acid sequence at least 40%, 50%, 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identical to one of the nucleic acid sequences set forth in Table 8 or a fragment thereof. In another embodiment, the nucleic acid molecule encoding a designer BMP comprises the nucleic acid sequence of any sequence set forth in SEQ ID NOs:74-139. In yet another embodiment, the nucleic acid molecule consists of the nucleic acid sequence of any one of the nucleic acid sequences of SEQ ID NOs:74-139.

In another embodiment, the nucleic acid molecule encoding a designer BMP comprises a nucleic acid sequence at least 40%, 50%, 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identical to the nucleic acid sequence of SEQ ID NO:78, or a fragment thereof. In another embodiment, the nucleic acid molecule encoding a designer BMP comprises the nucleic acid

sequence of SEQ ID NO:78. In yet another embodiment, the nucleic acid molecule consists of the nucleic acid sequence of SEQ ID NO:78 encoding BMPE.

In another embodiment, the nucleic acid molecule encoding a designer BMP comprises a nucleic acid sequence at least 40%, 50%, 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identical to the nucleic acid sequence of SEQ ID NO:80, or a fragment thereof. In another embodiment, the nucleic acid molecule encoding a designer BMP comprises the nucleic acid sequence of SEQ ID NO:80. In yet another embodiment, the nucleic acid molecule consists of the nucleic acid sequence of SEQ ID NO:80 encoding BMPG.

In another embodiment, the nucleic acid molecule encoding a designer BMP comprises a nucleic acid sequence at least 40%, 50%, 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identical to the nucleic acid sequence of SEQ ID NO:102, or a fragment thereof. In another embodiment, the nucleic acid molecule encoding a designer BMP comprises the nucleic acid sequence of SEQ ID NO:102. In yet another embodiment, the nucleic acid molecule consists of the nucleic acid sequence of SEQ ID NO:102 encoding BMPGE.

In another embodiment, the nucleic acid molecule encoding a designer BMP comprises a nucleic acid sequence at least 40%, 50%, 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identical to the nucleic acid sequence of SEQ ID NO:103, or a fragment thereof. In another embodiment, the nucleic acid molecule encoding a designer BMP comprises the nucleic acid sequence of SEQ ID NO:103. In yet another embodiment, the nucleic acid molecule consists of the nucleic acid sequence of SEQ ID NO:103 encoding BMPGER.

Methods of Producing Designer BMPs

BMPs are naturally expressed as pro-proteins comprising a long prodomain, one or more cleavage sites, and a mature domain. This pro-protein is then processed by the cellular machinery to yield a, typically, dimeric mature BMP molecule. In some embodiments, the designer BMPs are produced in a similar manner. The prodomain is believed to play a role in the folding and processing of BMPs. Furthermore, in some BMPs, the prodomain may noncovalently bind to the mature protein and act as a solubility enhancer, chaperone, or inhibitor. In some embodiments, BMPs may be produced as mature domains produced directly from or refolded from inclusion bodies. In other embodiments, the BMPs are produced via chemical synthesis or any other known method for protein production.

In one embodiment, the designer BMP is producing using chemical synthesis methods such as, but not limited to, synthetic methods well-known in the art.

In some embodiments, nucleic acids encoding designer BMPs are prepared by total gene synthesis or by site directed mutagenesis of a nucleic acid encoding a wild type, designer, or variant BMP. Methods include template directed ligation, PCR, cassette mutagenesis, site-directed mutagenesis, restriction enzyme digestion and ligation, or other techniques that are well known in the art (see, e.g., Prodromou et al., Protein Eng 5:827-9 (1992); Jayaraman et al., Biotechniques 12:392-8

(1992); Chalmers et al., Biotechniques 30:249-52 (2001); and Sambrook and Russell, In: Molecular Cloning, A Laboratory Approach, Cold Spring Harbor Press, Cold Spring Harbor, NY (2001)).

In some embodiments, an expression vector that comprises a gene encoding a designer BMP is prepared. Numerous types of appropriate expression vectors and suitable regulatory sequences for a variety of host cells are known in the art. The expression vectors may contain transcriptional and translational regulatory sequences including by not limited to promoter sequences, ribosomal binding sites, transcriptional terminator signals, polyadenylation signals, and enhancer or activator sequences. In some embodiments, the regulatory sequences include a promoter and transcriptional start and stop sequences. In addition, the expression vector may comprise additional elements, such as two replication systems to allow it to be maintained in two organisms. The expression vectors may be extrachromasomal vectors or vectors that integrate into a host cell's genome. In some embodiments, the expression vector contains at least one sequence homologous to a host cell's genome to promote integration into the genome. Constructs for integrating vectors are well known in the art. In some embodiments, the expression vector comprises a selectable marker gene to allow the selection of a stably transformed host cell. Selection marker genes are well known in the art and will vary with the host cell used.

The expression vector may include a secretory leader sequence or signal peptide sequence that provides for secretion of the designer BMP from the host cell. Suitable secretory leader sequences and signal peptides are known in the art.

Nucleic acids encoding designer BMPs may be introduced into host cells either alone or in combination with an expression vector so that the designer BMP is expressed from the nucleic acid. The method of introduction is largely dictated by the host cell type. Exemplary methods of transfection/transformation include $CaPO_4$ precipitation, liposome fusion, electroporation, viral infection, dextran-mediated transfection, polybrene-mediated transfection, protoplast fusion, direct microinjection, and other methods known in the art. Nucleic acids encoding designer BMPs may stable integrate into the host cell genome or may exist transiently or stably in the cytoplasm.

Appropriate host cells for expressing designer BMPs include any cell suitable for expressing wild type or native BMPs, including, but not limited to yeast, bacteria, archaebacteria, fungi, insect, and animal cells. In some embodiments the host cell is Saccharomyces cerevisiae or Escheria coli. In some embodiments, the host cell is a mammalian cell such as 293 (e.g., 293-T and 293-EBNA), BHK, CHO (e.g., CHOK1 and DG44), COS, Jurkat, NIH3T3, or C2C12 cells. Other suitable cells may be found in the ATCC catalog. Designer BMPs may be produced in more complex organisms, including but not limited to plants and animals. In one embodiment, the cells may be additionally genetically engineered, i.e., to contain exogenous nucleic acids other than the expression vector comprising the designer BMP nucleic acid.

In some embodiments, designer BMPs are produced by culturing a host cell transformed with an expression vector containing a nucleic acid encoding a designer BMP under the appropriate conditions to induce or cause expression of the designer BMP. The conditions appropriate for designer BMP

expression are the same conditions known to be appropriate for expressing native or wild type BMPs. These conditions will vary with the choice of expression vector and host cell, and may be easily ascertained by one skilled in the art through routine experimentation.

In some embodiments, the designer BMPs may be purified or isolated after expression. Standard purification methods include electrophoretic, molecular, immunological, and chromatographic techniques, including ion exchange, hydrophobic, affinity, and reverse-phase HPLC chromatography, and chromatofocusing. General guidance in suitable purification techniques may be found in Scopes, In: Protein Purification, Springer-Verlag, NY, 3rd Ed. (1994). The degree of purification necessary will vary depending on the desired use, and in some instances no purification will be necessary.

Purification from bacterial cells may result in the expression of BMPs in inclusion bodies and a subsequent step of refolding in a CHAPS/High salt system. Purification from mammalian cells may involve a two-step purification via Cellufine-Sulfate and Reversed Phase chromatography columns.

In some embodiments, the designer BMPs may be modified covalently or non-covalently. Covalent modifications may be introduced to a protein by reacting targeted amino acid residues of the protein with an organic derivatizing agent capable of reacting with selected side chains or terminal residues. Optimal sites for modification can be chosen using a variety of criteria, including but not limited to visual inspection, structural analysis, sequence analysis, and molecular simulation.

In some embodiments, designer BMPs may be labeled with at least one element, isotope, or chemical compound. The label may be an isotopic label, such as a radioactive or heavy isotope. In some embodiments, the label may be an immune label such as an antibody or antigen. In some embodiments, the label may be a colored or fluorescent label, such as fluorescein. In some embodiments, the label may be biotin, a tag (e.g., FLAG, Myc, His).

The designer BMPs may be derivatized with bifunctional agents to crosslink a designer BMP to a support matrix or surface for use in purifying antibodies or proteins that bind to the proteins or to detect binding in screening assays. Commonly used crosslinking agents include but are not limited to 1, ,1-bis(diazoacetyl)-2- phenylethane, glutaraldehyde, N-hydroxysuccinimide esters, for example, esters with 4- azidosalicylic acid, homobifunctional imidoesters, including disuccinimidyl esters such as 3,3'-dithiobis(succinimidylpropionate), bifunctional maleimides such as bis-N- maleimido-l,8-octane. Other modifications include deamidation of glutaminyl and asparaginyl residues to the corresponding glutamyl and aspartyl residues, respectively, hydroxylation of praline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the amino groups of lysine, arginine, and histidine side chains (T.E. Creighton, Proteins: Structure and Molecular Properties, W. H. Freeman & Co., San Francisco, pp. 79-86 (1983)), acetylation of the N-terminal amine, and amidation of any C- terminal carboxyl group. Such derivatization may improve the solubility, absorption, transport across the blood brain barrier, serum half-life, and the like. Modifications of designer BMPs may alternatively eliminate or attenuate any possible undesirable side effect of the protein. Moieties capable of mediating such effects

are disclosed, for example, in Remington's Pharmaceutical Sciences, 16th ed., Mack Publishing Co., Easton, PA (1980).

Another type of covalent modification of designer BMPs comprises linking the protein to one of a variety of nonproteinaceous polymers, e.g., polyethylene glycol ("PEG"), polypropylene glycol, or polyoxyalkylenes, in the manner set forth in U.S. Patent Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192 or 4,179,337. A variety of coupling chemistries may be used to achieve PEG attachment, as is well known in the art.

In another embodiment, the designer BMP comprises linking the protein via a CovX-body linker to a CovX-body antibody such as, but not limited to, the CovX-bodies described in US Patent No. 5,733,757, and US Patent Publication No. US 2009/0098130. Such CovX-bodies may exhibit improved characteristics, including, but not limited to, improved stability and extended serum half-life.

Methods of Assaying Receptor Binding Activity of Designer BMPs

The receptor binding activity of designer BMPs may be assessed using any methods used for assessing the activity of wild type BMPs.

The affinity of designer BMPs for one or more BMP receptors can be determined by receptor binding assays. For example, affinities for ALK-2, ALK-3, ALK-6, ActRII, ActRIIb, or BMPRII can be determined. Suitable binding assays include, but are not limited to ELISA, fluorescence anisotropy and intensity, scintillation proximity assays (SPA), Biacore (Pearce et al., *Biochemistry* 38:81-89 (1999)), DELFIA assays, and AlphaScreen™ (commercially available from PerkinElmer; Bosse R., Illy C, and Chelsky D (2002)).

In some embodiments, Biacore or surface plasmon resonance assays are used. See, for example, McDonnell, *Curr. Opin. Chem. Biol.* 5:572- 577 (2001). Biacore experiments have been used previously to characterize binding of TGF-β isoforms to their receptors (De Crescenzo et al., J. Biol. Chem., 276: 29632-29643 (2001); De Crescenzo et al., J. Mol. Biol. 328: 1173-1183) (2003).

In other embodiments, a plate-based Direct Binding Assay is used to determine the affinity of one or more modified BMPs for one or more BMP receptors. This method is a modified sandwich ELISA in which BMP is captured using an anti-BMP monoclonal antibody and then detected using a BMP receptor-Fc fusion protein.

In other embodiments, AlphaScreen™ assays (Bosse R. et al., Principles of AlphaScreen™, PerkinElmer Literature Application Note Ref #4069, http://lifesciences.perkinelmer.com/ Notes/S4069-0802.pdf (2002)) can be used to characterize receptor and inhibitor binding. Fluorescence assays may also be used to characterize receptor and inhibitor binding. For example, either BMP2 or a BMP2 receptor or inhibitor may be labeled with a fluorescent dye (for examples of suitable dyes, see the Molecular Probes catalog). Additionally, scintillation proximity assays (SPA) can be used to determine receptor binding affinity. For example, BMP receptor-Fc fusions may be bound to protein A coated SPA beads or flash-plate and treated with S35-labeled BMP; the binding event results in production of light.

In a particular embodiment, the KD of a specific BMP mutant to a Type I or Type II receptor can be determined by using receptor extracellular domain fusions to a human IgG-Fc. The receptor can be bound to an octet sensor using anti-human-IgG-Fc sensors and the BMP can bind the receptor extracellular domain in solution to determine Kon and Koff rates. The Octet systems utilize proprietary BioLayer Interferometry (BLI) to enable real-time, label-free analysis of biomolecular interactions and to provide information on affinity, kinetics and concentration. As proteins bind the Octet sensor the light passing through the sensor has a wavelength shift that can be measured with a spectrophotometer. The rate of the shift is measured as the analyte binds the sensor and when it loses binding.

Methods of Assaying Osteogenic Activity of designer BMP

The osteogenic activity of designer BMPs may be assessed using any methods used for assessing the activity of wild type BMPs.

BMPs promote the growth and differentiation of a number of types of cells. Differentiation may be monitored using, for example, luminescence reporters for alkaline phosphatase or calorimetric reagents such as Alcian Blue or PNPP (Asahina et al. (1996) Exp. Cell Res,, 222:38-47; Inada et al. (1996) Biochem. Biophys. Res. Commun.. 222:317- 322; Jortikka et al. (1998) Life ScL 62:2359-2368; Cheng et al. (2003) J. Bone Joint Surgery 95A:1544-1552).

The rat limb bud cartilage differentiation assay may also be used to monitor activity in primary cells. In alternative embodiments, reporter gene or kinase assays may be used. Since BMPs activate the JAK-STAT signal transduction pathway, a BMP responsive cell line containing a STAT-responsive reporter such as GFP or luciferase may be used (Kusanagi et al. (2000) Mol Biol. Cell., 11 :555-565). For example, BMP activity in kidney cells may be determined using cell-based assays; see for example Wang and Hirschberg (2004) J. Biol. Chem., 279:23200- 23206.

Osteogenic activity may be measured in cell based assays such as alkaline phosphatase, BRE-luciferase, or Alizarin red mineralization, all of which are described in Isaacs et al., *Mol. Endocrinol.* 24:1469-1477 (2010).

Osteogenic activity may also be measured in vivo, via rat ectopic bone assays or mammalian bone growth models. In some embodiments, osteogenic activity is measured in non-human primate models. These models are described in Isaacs et al., *Mol. Endocrinol.* 24:1469-1477 (2010).

Methods for evaluating bone mass and quality are known in the art and include, but are not limited to X-ray diffraction; DXA; DEQCT; pQCT, chemical analysis, density fractionation, histophotometry, histomorphometry, and histochemical analysis as described, for example, in Lane et al., *J. Bone Min. Res.* 18:2105-2115 (2003). One assay for determining cortical bone density is the MicroCT assay. Following pQCT measurement, the microCT evaluation can be performed, for example, using a Scanco mCT40 (Scanco Medical AG) on a femur.

Any known or later developed in vitro or in vivo method for assessing bone growth/density/strength may be used to assess the osteogenic activity of the designer BMPs of the invention.

Pharmaceutical Compositions

Designer BMPs of the present invention may be formulated for administration to a mammal, preferably a human in need thereof as part of a pharmaceutical composition. The composition can be administered by any suitable means, e.g., parenterally, orally or locally. Where the designed BMPs is to be administered locally, as by injection, to a desired tissue site, or systemically, such as by intravenous, subcutaneous, intramuscular, intraorbital, ophthalmic, intraventricular, intracranial, intracapsular, intraspinal, intracistemal, intraperitoneal, buccal, rectal, vaginal, intranasal or aerosol administration, the composition preferably comprises an aqueous solution. The solution preferably is physiologically acceptable, such that administration thereof to a mammal does not adversely affect the mammal's normal electrolyte and fluid volume balance. The aqueous solution thus can comprise, e.g., normal physiologic saline (0.9% NaCl, 0.15M), pH 7-7.4.

Useful solutions for oral or parenteral systemic administration can be prepared by any of the methods well known in the pharmaceutical arts, described, for example, in "Remington's Pharmaceutical Sciences" (Gennaro, A., ed., Mack Pub., 1990, the disclosure of which is incorporated herein by reference). Formulations can include, for example, polyalkylene glycols such as polyethylene glycol, oils of vegetable origin, hydrogenated naphthalenes, and the like. Formulations for direct administration, in particular, can include glycerol and other compositions of high viscosity.

Biocompatible, preferably bioresorbable polymers, including, for example, hyaluronic acid, collagen, tricalcium phosphate, polybutyrate, polylactide, polyglycolide and lactide/glycolide copolymers, may be useful excipients to control the release of the designer BMPs in vivo. Other potentially useful parenteral delivery systems for the present designer BMPs can include ethylene-vinyl acetate copolymer particles, osmotic pumps, implantable infusion systems, and liposomes. Formulations for inhalation administration can contain as excipients, for example, lactose, or can be aqueous solutions containing, for example, polyoxyethylene-9-lauryl ether, glycocholate or deoxycholate, or oily solutions for administration in the form of nasal drops or as a gel to be applied intranasally.

Alternatively, the designer BMPs of the invention, including designer BMP2 and BMP6, identified as described herein may be administered orally. For example, liquid formulations of designer BMPs can be prepared according to standard practices such as those described in "Remington's Pharmaceutical Sciences" (supra). Such liquid formulations can then be added to a beverage or another food supplement for administration. Oral administration can also be achieved using aerosols of these liquid formulations. Alternatively, solid formulations prepared using art-recognized emulsifiers can be fabricated into tablets, capsules or lozenges suitable for oral administration.

Optionally, the designer BMPs can be formulated in compositions comprising means for enhancing uptake of the protein by a desired tissue. For example, tetracycline and diphosphonates (bisphosphonates) are known to bind to bone mineral, particularly at zones of bone remodeling, when they are provided systemically in a mammal. Accordingly, such components can be used to enhance delivery of the present designer BMPs to bone tissue. Alternatively, an antibody or portion thereof that binds specifically to an accessible substance specifically associated with the desired target tissue, such as a cell surface antigen, also can be used. If desired, such specific targeting molecules can be covalently bound to the present designer BMP, e.g. , by chemical crosslinking or by using standard genetic engineering techniques to create, for example, an acid labile bond such as an Asp-Pro linkage. Useful targeting molecules can be designed, for example, according to the teachings of US Patent No. 5,091,513.

It is contemplated also that some of the designer BMPs may exhibit the highest levels of activity in vivo when combined with carrier matrices, i.e., insoluble polymer matrices. See for example, US Patent No.5,266,683 the disclosure of which is incorporated by reference herein. Currently preferred carrier matrices are xenogenic, allogenic or autogenic in nature. It is contemplated, however, that synthetic materials comprising polylactic acid, polyglycolic acid, polybutyric acid, derivatives and copolymers thereof may also be used to generate suitable carrier matrices. Preferred synthetic and naturally derived matrix materials, their preparation, methods for formulating them with the designer BMPs of the invention, and methods of administration are well known in the art and so are not discussed in detailed herein. See for example, US Patent No.5,266,683.

In certain embodiments, the designer BMPs can be administered to the mammal in need thereof either alone or in combination with another substance known to have a beneficial effect on tissue morphogenesis. Examples of such substances (herein, cofactors) include substances that promote tissue repair and regeneration and/or inhibit inflammation or fibrosis. Examples of useful cofactors for stimulating bone tissue growth in osteoporotic individuals, for example, include but are not limited to, vitamin D3, calcitonin, prostaglandins, parathyroid hormone, dexamethasone, estrogen and IGF-I or IGF-II. Useful cofactors for nerve tissue repair and regeneration can include nerve growth factors. Other useful cofactors include symptom-alleviating cofactors, including antiseptics, antibiotics, antiviral and antifungal agents, analgesics and anesthetics.

Designer BMPs are preferably formulated into pharmaceutical compositions by admixture with pharmaceutically acceptable, nontoxic excipients and carriers. As noted above, such compositions can be prepared for systemic, e.g., parenteral, administration, particularly in the form of liquid solutions or suspensions; for oral administration, particularly in the form of tablets or capsules; or intranasally, particularly in the form of powders, nasal drops or aerosols. Where adhesion to a tissue surface is desired, the composition can comprise a fibrinogen-thrombin dispersant or other bioadhesive such as is disclosed, for example, in PCT US91/09275, the disclosure of which is incorporated herein by reference. The composition then can be painted, sprayed or otherwise applied to the desired tissue surface.

When administered, the pharmaceutical composition of this invention is typically delivered in a pyrogen-free, physiologically acceptable form. Further, the composition may desirably be encapsulated or injected in a viscous form for delivery to the site of bone cartilage or tissue damage. Local administration may be suitable for wound healing and tissue repair. Preferably for bone and/or cartilage formation, the composition includes a matrix capable of delivering BMP protein to the site of bone and/or cartilage damage, providing a structure for the developing bone and cartilage and optimally capable of being resorbed into the body. Such matrices may be formed of materials presently in use for other implanted medical applications.

The choice of matrix material is based on biocompatibility, biodegradability, mechanical properties, cosmetic appearance and interface properties. The particular application of the designer BMP compositions will define the appropriate formulation. Potential matrices for the compositions may be biodegradable and chemically defined calcium sulfate, tricalciumphosphate, hydroxyapatite, polylactic acid and polyanhydrides. Other potential materials are biodegradable and biologically well defined, such as bone or dermal collagen. Further matrices are comprised of pure proteins or extracellular matrix components. Other potential matrices are nonbiodegradable and chemically defined, such as sintered hydroxyapatite, bioglass, aluminates, or other ceramics. Matrices may be comprised of combinations of any of the above-mentioned types of material, such as polylactic acid and hydroxyapatite or collagen and tricalciumphosphate. The bioceramics may be altered in composition, such as in calcium-aluminate-phosphate and processing to alter pore size, particle size, particle shape, and biodegradability.

The dosage regimen will be determined by the attending physician considering various factors which modify the action of the designer BMP protein. These factors include, without limitation, the amount of bone weight desired to be formed, the site of bone damage, the condition of the damaged bone, the size of a wound, type of damaged tissue, the patient's age, sex, and diet, the severity of any infection, time of administration and other clinical factors. The dosage may vary with the type of matrix used in the reconstitution. The addition of other known growth factors, such as IGF I (insulin like growth factor I), to the final composition, may also effect the dosage. Progress can be monitored by periodic assessment of bone growth and/or repair. One method of assessing bone growth or repair is by x-ray imaging and/or CT scanning, among many art-recognized methods.

The compositions can be formulated for parenteral or oral administration to humans or other mammals in therapeutically effective amounts, e.g., amounts which provide appropriate concentrations of the designer BMPs to target tissue for a time sufficient to induce the desired effect. Preferably, the present compositions alleviate or mitigate the mammal's need for a morphogen-associated biological response, such as maintenance of tissue-specific function or restoration of tissue-specific phenotype to senescent tissues (e.g., osteopenic bone tissue) or the inhibition or reversal of a fibrotic response in a tissue.

As will be appreciated by those skilled in the art, the concentration of the compounds described in a therapeutic composition will vary depending upon a number of factors, including the dosage of the drug

to be administered, the chemical characteristics (e.g., hydrophobicity) of the compounds employed, and the route of administration. The preferred dosage of drug to be administered also is likely to depend on such variables as the type and extent of a disease, tissue loss or defect, the overall health status of the particular patient, the relative biological efficacy of the compound selected, the formulation of the compound, the presence and types of excipients in the formulation, and the route of administration.

In general terms, the compounds of this invention may be provided in an aqueous physiological buffer solution containing about 0.1 to 10% w/v compound for parenteral administration. Typical doses ranges are from about 10 ng/kg to about 1 g/kg of body weight per day; with a preferred dose range being from about 0.1 mg/kg to 100 mg/kg of body weight.

Therapeutic Uses

Designer BMPs may be used for any indication that wild type BMPs are useful for or for any method in which a TGFβ superfamily member can be used. Designer BMPs are capable of inducing the developmental cascade of bone and cartilage morphogenesis and to induce or mediate Smad signaling pathways. Designer BMPs induce greater bone augmentation and repair, including, but not limited to, production of greater bone mass, bone stiffness and bone density that corresponding wild type BMP. Accordingly, designer BMPs may be used to induce bone formation in a tissue. Also, designer BMPs may be used to induce proliferation of bone and cartilage in a variety of locations in the body. For example, designer BMPs may be used to repair joints such as knee, elbow, ankle, and finger. For example, designer BMPs may be useful for regenerating cartilage in patients suffering from arthritis or other cartilage degenerating diseases. Further, designer BMPs are indicated for treating tears in cartilage due to injury. In addition, designer BMPs are useful for inducing bone growth in patients. For example, designer BMPs are indicated for use in treating patients suffering from bone fractures or breaks, osteoporosis, or patients in need of spinal fusion or for repair of the spine, vertebrae or the like.

In another embodiment, the invention includes a method of bone augmentation and/or repair. Thus, the invention encompasses administering a therapeutically effective amount of a designer BMP to a site where it mediates detectable bone augmentation or repair.

In another embodiment, the invention includes a method of inducing or increasing Smad expression. The method comprises contacting a cell comprising Smad mediated expression pathway with a designer BMP of the invention.

Designer BMPs are capable of inducing the developmental cascade of bone morphogenesis and tissue morphogenesis for a variety of tissues in mammals different from bone or bone cartilage. This morphogenic activity includes the ability to induce proliferation and differentiation of progenitor cells, and the ability to support and maintain the differentiated phenotype through the progression of events that results in the formation of bone, cartilage, non-mineralized skeletal or connective tissues, and other adult tissues.

For example, designer BMPs may be used for treatment to prevent loss of and/or increase bone mass in metabolic bone diseases. General methods for treatment to prevent loss of and/or increase bone mass in metabolic bone diseases using osteogenic proteins are disclosed in U.S. Patent No. 5,674,844, the disclosures of which are hereby incorporated by reference. Designer BMPs may also be administered to replace or repair bone or cartilage at injury sites such as bone breaks, bone fractures, and cartilage tears. Designer BMPs of the present invention may be used for periodontal tissue regeneration. General methods for periodontal tissue regeneration using osteogenic proteins are disclosed in U.S. Patent No. 5,733,878, the disclosures of which are hereby incorporated by reference.

Designer BMPs may be used for liver regeneration. General methods for liver regeneration using osteogenic proteins are disclosed in U.S. Patent No. 5,849,686, the disclosures of which are hereby incorporated by reference. Designer BMPs may be used for treatment of chronic renal failure. General methods for treatment of chronic renal failure using osteogenic proteins are disclosed in U.S. Patent No. 6,861,404, the disclosures of which are hereby incorporated by reference. Designer BMPs may be used for enhancing functional recovery following central nervous system ischemia or trauma. General methods for enhancing functional recovery following central nervous system ischemia or trauma using osteogenic proteins are disclosed in U.S. Patent No. 6,407,060, the disclosures of which are hereby incorporated by reference.

Designer BMPs may be used for inducing dendritic growth. General methods for inducing dendritic growth using osteogenic proteins are disclosed in U.S. Patent No. 6,949,505, the disclosures of which are hereby incorporated by reference.

Designer BMPs may be used for inducing neural cell adhesion. General methods for inducing neural cell adhesion using osteogenic proteins are disclosed in U.S. Patent No. 6,800,603, the disclosures of which are hereby incorporated by reference.

Designer BMPs may be used for treatment and prevention of Parkinson's disease. General methods for treatment and prevention of Parkinson's disease using osteogenic proteins are disclosed in U.S. Patent No. 6,506,729, the disclosures of which are hereby incorporated by reference.

It is within skills of an ordinary artisan to modify the general methods using the modified BMPs of the present invention for various therapeutic uses described above. Exemplary embodiments of therapeutic applications of the modified BMPs of the present invention are further described below.

Designer BMPs may be used to repair diseased or damaged mammalian tissue. The tissue to be repaired is preferably assessed, and excess necrotic or interfering scar tissue removed as needed, by surgical, chemical, ablating or other methods known in the medical arts. The designer BMPs then may be provided directly to the tissue locus as part of a sterile, biocompatible composition, either by surgical implantation or injection. Alternatively, a sterile, biocompatible composition containing modified BMP-stimulated progenitor cells may be provided to the tissue locus. The existing tissue at the locus, whether diseased or damaged, provides the appropriate matrix to allow the proliferation and tissue-specific differentiation of progenitor cells. In addition, a damaged or diseased tissue locus, particularly one that

has been further assaulted by surgical means, provides a morphogenically permissive environment. For some tissues, it is envisioned that systemic provision of the modified BMPs will be sufficient.

Designer BMPs may be used to prevent or substantially inhibit scar tissue formation following an injury. If a designer BMP is provided to a newly injured tissue locus, it can induce tissue morphogenesis at the locus, preventing the aggregation of migrating fibroblasts into non-differentiated connective tissue. The designer BMP preferably is provided as a sterile pharmaceutical preparation injected into the tissue locus within five hours of the injury.

For example, the designer BMPs may be used for protein-induced morphogenesis of substantially injured liver tissue following a partial hepatectomy. Variations on this general protocol may be used for other tissues. The general method involves excising an essentially nonregenerating portion of a tissue and providing the modified BMP, preferably as a soluble pharmaceutical preparation to the excised tissue locus, closing the wound and examining the site at a future date. Like bone, liver has a potential to regenerate upon injury during post-fetal life.

As another example, designer BMPs can also be used to induce dentinogenesis. To date, the unpredictable response of dental pulp tissue to injury is a basic clinical problem in dentistry. Using standard dental surgical procedures, small areas (e.g., 2 mm) of dental pulps can be surgically exposed by removing the enamel and dentin immediately above the pulp (by drilling) of sample teeth, performing a partial amputation of the coronal pulp tissue, inducing hemostasis, application of the pulp treatment, and sealing and filling the cavity by standard procedures.

The designer BMPs of the invention may be used to treat fibrosis. The fibrosis may be located in various parts of the body and can be of a particular kind, for example, the fibrosis may be located: in the kidney, for example, fibrosis as observed in glomerulonenephritis, diabetic nephropathy, allograft rejection, and HIV nephropathy; in the liver, for example, cirrhosis, and veno-occlusive disease; in the lung, for example, idiopathic fibrosis (and autoimmune fibrosis); in the skin, for example, systemic sclerosis, keloids, scars, and eosinophilia-myalgia syndrome; in the central nervous system, for example, intraocular fibrosis; in the cardiovascular system, for example, vascular restenosis; in the nose, for example, nasal polyposis; in bone or bone marrow; in an endocrine organ; and in the gastrointestinal system.

In one embodiment, a designer BMP having the binding characteristics of BMP7, or useful modification thereof (extended half life, increase binding affinity for a same or different receptor compared with wild type BMP7, resistance to inhibition by a BMP7 antagonist, such as, but not limited to, Noggin, and the like) may be useful to treat, ameliorate or reverse fibrosis. That is, as reviewed recently in Weiskirchen et al., 2009, Frontiers in Biosci. 14:4992-5012, $TGF\beta_1$ mediates a cascade leading to increased fibrosis, including, but not limited to, epithelial-to-mesenchymal transition. The fibrosis-inducing effects of $TGF\beta_1$ may be inhibited or reversed by BMP7. See also Loureiro et al., 2010, Nephrol. Dial. Transplant. 25:1098-1108. Further, certain fribotic conditions may also be treated or ameliorated by administration of BMP4 (see Pegorier et al., 2010, Resp. Res. 11:85). Therefore, the invention

encompasses a designer BMP either based on a BMP7 framework and/or incorporating the type I and type II mutations disclosed elsewhere herein, to alter receptor binding and provide a potential useful therapeutic for treatment of fibrosis in a patient in need thereof.

A fibrotic disorder may be induced by a number of causes including: chemotherapy, for example, pulmonary fibrosis resulting from bleomycin, chlorambucil, cyclophsphamide, methotrexate, mustine, or procarbazine treatment; radiation exposure whether accidental or purposeful as in radiation therapy, for example, interstitial lung disease (ILD) resulting from radiation; environmental or industrial factors or pollutants such as chemicals, fumes, metals, vapors, gases, etc., for example, ILD resulting from asbestos or coal dust; a drug or a combination of drugs, for example, antibiotics (e.g. penicillins, sulfonamides, etc.), cardiovascular drugs (e.g., hydralazine, beta blockers, etc.), CNS drugs (phenytoin, chlorpromazine, etc.) anti-inflammatory drugs (e.g., gold salts, phenylbutazone, etc.), etc. can cause ILD; an immune reaction disorder, for example, chronic graft-versus-host disease with dermal fibrosis,; disease states such as aspiration pneumonia which is a known cause of ILD, and parasite induced fibrosis; and wounds, for example, blunt trauma, surgical incisions, battlefield wounds, etc., as in penetrating injuries of the CNS.

In a particular embodiment, designer BMPs with improved binding to type I receptor ALK2, such as BMPE, may be used to treat diseases related to ALK2.

Kits

The invention includes various kits which comprise a therapeutically effective amount of a designer BMP of the invention, along with an applicator and instructional materials which describe use of the designer BMP to perform the methods of the invention. Although exemplary kits are described below, the contents of other useful kits will be apparent to the skilled artisan in light of the present disclosure. Each of these kits is included within the invention.

The invention includes a kit for treatment to prevent loss of and/or increase bone mass in a metabolic bone disease in a patient in need thereof. The kit includes a designer BMP of the invention. The kit further comprises an applicator, including, but not limited to, a syringe, a bone cement mixing device, and the like, for administration of the components of the kit to a patient. Further, the kit comprises an instructional material setting forth the pertinent information for the use of the kit to treat or prevent bone mass and/or increase bone mass in the patient.

More preferably, the kit comprises at least one designer BMP selected from an antibody having an amino acid sequence selected from the amino acid sequence of SEQ ID NOs:8-73, even more preferably, the designer BMP comprises the amino acid sequence of SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:36 and SEQ ID NO:37. Preferably, the designer BMP is BMPE, BMPG, BMPGE and BMPGER.

The kit can comprise any number of additional therapeutic agents for treatment to prevent bone loss and/or increase bone mass. Such agents are set forth previously and include therapeutic compounds, cytokines, vitamins, other members of the TGFβ superfamily, among many others.

The invention also relates to an article of manufacture (*e.g.*, dosage form adapted for i.v. or oral administration) comprising a designer BMP in the amount effective to prevent bone loss and/or increase bone mass (*e.g.*, more than 10 mg/kg, at least 15 mg/kg, or 15 mg/kg). In certain embodiments, the article of manufacture comprises a container or containers comprising a designer BMP and a label and/or instructions for use to treat or prevent bone loss and/or increase bone mass.

The invention also includes a kit to treat or prevent fibrosis in a tissue or organ in a patient in need thereof. The kit includes a designer BMP of the invention. The kit further comprises an applicator, including, but not limited to, a syringe or device for delivering the protein, a mixing device, and the like, for administration of the components of the kit to a patient. Further, the kit comprises an instructional material setting forth the pertinent information for the use of the kit to treat or prevent fibrosis in the patient.

More preferably, the kit comprises at least one designer BMP selected from a protein having an amino acid sequence selected from the amino acid sequence of SEQ ID NOs:8-73, even more preferably, the designer BMP comprises the amino acid sequence of SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:36 and SEQ ID NO:37. Preferably, the designer BMP is BMPE, BMPG, BMPGE or BMPGER.

The kit can comprise any number of additional therapeutic agents for treatment to prevent bone loss and/or increase bone mass or treat or prevent fibrosis. Such agents are set forth previously and include therapeutic compounds, cytokines, vitamins, other members of the TGFβ superfamily, among many others.

The invention also relates to an article of manufacture (*e.g.*, dosage form adapted for i.v. or oral administration) comprising a designer BMP in the amount effective to prevent bone loss and/or increase bone mass or to treat or prevent fibrosis (*e.g.*, more than 1 mg/kg, at least 10 mg/kg, at least 15 mg/kg, or 15 mg/kg). In certain embodiments, the article of manufacture comprises a container or containers comprising a designer BMP and a label and/or instructions for use to treat or prevent bone loss and/or increase bone mass or to treat or prevent fibrosis.

The invention is further described in detail by reference to the following experimental examples. These examples are provided for purposes of illustration only, and are not intended to be limiting unless otherwise specified. Thus, the invention should in no way be construed as being limited to the following examples, but rather, should be construed to encompass any and all variations which become evident as a result of the teaching provided herein.

EXAMPLES

EXAMPLE 1

Production and Purification of designer BMP proteins

Production using Mammalian Cell Culture

Recombinant host CHO cells producing and secreting wild type and designer BMPs were generated using standard recombinant DNA procedures. Conditioned medium was generated from adherent cell cultures. Briefly, CHO cells were seeded in medium containing 10% dFBS and allowed to grow to near confluence for 3-4 days. After this growth phase, growth medium was discarded and the cells were rinsed once with PBS-CMF and subsequently switched to a serum-free medium supplemented with 200 ug/ml dextran sulfate, 2mM sodium butyrate, and 10mM HEPES. Cells were then cultured for 7 days at a temperature of 31°C. Conditioned medium was harvested and clarified by using sterilizing 0.2 uM filtration. Conditioned medium was stored at -20°C until purification.

Purification of designer BMPs

In order to purify the novel designer BMP molecules from CHO cell conditioned media the BMPs were captured by two steps of conventional chromatography and the results are shown in Figure 5, comprising panels A-D. Only the details of the purification of BMPE are shown herein since all of the other novel designer BMPs were purified in an essentially similar manner.

CHO conditioned medium (CHO CM) (pH adjusted to 8.0 with 1.0 M Tris, pH 8.0) was loaded onto a Cellufine Sulfate column (65ml, 2.6 x 12.3cm) that was equilibrated with 20mM MES pH 8.0. The column was washed with 10 column volumes (CV) of 20 mM Tris, pH 8.0, 10 CV 50mM MES pH 5.6 and 10 CV of Buffer A (6.0 M Urea, 50mM MES, pH 5.6). The BMPs were eluted with a linear 0-1.0 M NaCl gradient over 5 CV (Buffer B= 6.0 M Urea, 50 mM MES, 1.0 M NaCl, pH 5.6). Upon application of a sodium chloride gradient, a broad peak between conductivities of 30 and 45 mS/cm characteristic of BMP2 was observed (Figure 5A). Fractions were analyzed by Coomassie stained SDS-PAGE gels and BMP containing fractions were pooled. BMPs in fractions were identified as reducible dimers on SDS-PAGE Non-Reduced gels (left panel of Figure 5B). The BMP pools from the Cellufine Sulfate chromatography step were further purified by preparative Reverse Phase HPLC on a 10 x 250mm Vydac 15 µm C8 Column (Solvent A= 0.1% TFA, Solvent B= 90% acetonitrile, 0.1% TFA), with BMP eluting with approximately 32% acetonitrile. A tracing of the Reversed Phase chromatography step is shown in Figure 5C. The protein was concentrated and acetonitrile was removed using a speedvac and the concentrate was formulated into MFR-169 buffer via dialysis. The purified BMPs were characterized by SDS-PAGE, A280 and LAL Assay (endotoxins). A photograph of an Non-Reduced SDS-PAGE gel (left side of Figure 5D) and a Reduced SDS-PAGE gel (right side of Figure 5D) showing the same gel fractions (F13 through F18) is shown. A total of 16 BMP designer proteins were purified to essentially the same levels of purity and expression/purification yields ranging from 0.3-1.4 mg/L CM and the results are shown in Figure 6 showing photographsa (Figure 6). Briefly, wild type BMP2 (WT) and designer BMPs

BMPSE, BMPE, E109R, BMPD, BMP S85R, BMP SNE, BMPB, and BMP-EN are shown in photographs of a non-reduced gel (Figure 6A) and a reduced SDS-PAGE (Figure 6B), and designer BMPs ai (variant of BMPA), aii (variant of BMPA), c (BMPC), hi (variant of BMPI), hii, i, f, and g are shown in photographs of a non-reduced SDS-PAGE (Figure 6C) and reduced SDS-PAGE (Figure 6D).

EXAMPLE 2

Osteogenic Activity of designer BMPs demonstrated using In Vitro and In Vivo Assays

Alkaline phosphatase assay

Approximately 8000 C2C12 cells/well in a 96-well plate were treated with the indicated BMP ant the dose indicated. Twenty-four hours post-treatment, the plates were processed to measure alkaline phosphatase which is an art-recognized assay for osteogenic activity. The culture medium was removed, and the plates were washed twice with calcium/magnesium-free PBS. 50 µl of 4-Methylumbelliferyl phosphate (4-MUP Liquid Alkaline Phosphatase Substrate; Sigma cat. # M3168) was added to each well, and the plates were incubated in the dark at 37°C for 15 minutes. Fluorescence was measured on a Victor luminometer (settings: excitation at 355nM; emission at 460nM; CW lamp energy at 1120), 1 second per well. After the reading was complete, 50 µl of 2x protein assay lysis buffer (200 mM Tris-HCl, pH 9.8/0.4% Triton X-100) was added to each well and the protein concentration was determined using the BCA Protein assay (Pierce) following the manufacturer's microplate procedure. The alkaline phosphatase measurements were then normalized to the total protein concentration (i.e., fluorometric units per microgram of protein). As shown by the graph in Figure 7, C2C12 muscle pre-myoblast cells treated with multiple designer BMP molecules showed significantly increased Alkaline Phosphatase activity, as a marker of osteoblast differentiation, compared to treatment with wild type BMP2 (heavy line with small circles). Designer BMPs exhibiting increased AP activity compared with WT BMP2 included designers BMPA, BMPF, BMPG, and BMPE. Surprisingly, designer BMPE demonstrated equivalent activity to that of the wild type BMP2/6 heterodimer (heavy line with squares), which is known to bind both the type I receptors of BMP2 and type II receptors of BMP6 with high affinity. Designer BMPE is the result of introduction of the low affinity type I binding region of BMP6 into BMP2. The extremely high activity of the designer BMPE molecule was extremely surprising since it was predicted that BMPE would have low affinity binding to both type I and type II receptors. Interestingly, the other designer BMP molecules, designer BMPA, designer BMPF, and designer BMPG, have regions of wild type BMP6 that bind the type II (high affinity) receptors of BMP6 which have been introduced into BMP2 (see Figure 1B), and these designer BMPs showed increased activity compared to BMP2, but not as high as that of wild type BMP2/6 heterodimer (Figure 7).

BRE-luciferase assay

C2C12 cells stably expressing the BMP-response-element luciferase reporter (element is from the Id1 promoter) were plated at 8000 cells/well of a 96 well and treated with the indicated BMP and

dose. 48 hours post treatment, the cells were lysed and luciferase activity was read using the Promega Dual-Glo assay kit.

The data disclosed herein demonstrated that not only was the activity of BMPE equivalent to that of BMP-2/6 in an alkaline phosphatase assay, it was also equivalent in a BRE-luciferase assay in C2C12 cells as shown in Figure 8. Further, BMPE demonstrated approximately 10-20 fold greater activity in the BRE-luciferase assay compared with wild type BMP-2 (Figure 8). Thus, the results observed in the BRE-luciferase (BRE-luc) assay correlated strongly with those obtained in the Alkaline Phosphatase (Alk-phos) activity assay in this same cell type (compare Figure 7 and Figure 8). Results from both the Alk-phos and BRE-luc assays are also shown in Table 10 for wild type BMP2 and the indicated designer BMPs.

Without wishing to be bound by any particular theory, these data suggest that the addition of ALK-2 as a high affinity receptor for BMPE could be the reason for its increased osteogenic activity. This is because an ALK-2 mutation has been found to cause fibrodysplasia ossificans progressiva (FOP), a disease where young children develop inappropriate ectopic bone formation. Thus, mutation of ALK-2 binding is associated with increased osteogenesis and may be correlated to the increased osteogenic activity of BMPE. Thus, BMPE is a new class of BMP molecule with high affinity for the type I receptors ALK-2, 3, and 6.

Alizarin Red Staining for cell mineralization

C2C12 cells were plated in 6-well tissue culture plates at a density of 4 x 10^4 cells/cm^2 and incubated overnight at 37°C inside a 5% CO_2/95% humidified air incubator. After the recovery period, the culture medium was replaced with freshly prepared osteogenic differentiation medium: Growth Medium containing 50ug/ml L-ascorbic acid phosphate (L-Ascorbic Acid Phosphate Magnesium Salt n-Hydrate; WAKO Pure Chemical Industries; Cat. No. 013-12061); β-glycerol phosphate (β-Glycerol phosphate Disodium salt, 10mM Pentahydrate; Fluka BioChemica Cat. No. 50020); and 100 nM Menadione sodium bisulfite (Vitamin K3; Sigma Cat. No. M2518). The indicated BMP was added to the appropriate wells at the desired concentration. The plates were incubated at 37°C for approximately 15 days, with medium replacement every 2 to 3 days. The cells were stained with the Alizarin Red stain following the standard published protocols.

As shown in Table 9, below, designer BMPE induced mineralization of C3H10T-1/2 mouse mesenchymal stem cells to a far greater extent than corresponding wild type BMP2 as indicated by alizarin red staining. That is, as more fully discussed below, at doses where wild type BMP-2 was unable to induce mineralization of the C3H10T-1/2 cells (5, 25, 50, and 100 ng/ml) BMPE homodimer induced strong mineralization similar to that of the BMP-2/6 heterodimer all as shown in Table 9. Thus, the alizarin red staining assay results further correlate the results obtained in the Alk-phos and BRE-luc assays as disclosed previously herein.

TABLE 9

Treatment	BMP2	BMP2/6	BMPE
5 ng/ml	-	-	-
25 ng/ml	-	++	+
50 ng/ml	-	++++	++
100 ng/ml	-	++++	+++

Rat intramuscular ectopic bone assay

To determine whether the stronger osteogenic activity observed in vitro by the designer BMPs corresponded to similar increased activity in vivo, rat ectopic bone formation assays were performed. Briefly, an ACS (absorbable collagen sponge) impregnated with the indicated total amount of designer BMP in 160 microliters of buffer was implanted into the hamstring of 8 week old male Long Evans rats. More specifically, three 8 mm biopsy punched ACS discs were sutured together with non-resorbable silk sutures. The sponges were wetted with 160 microliters of the BMP solution containing the amount of BMP indicated in the chart in Figure 9 (i.e., 0.1 µg or 0.5 µg). The wetted sponges were equilibrated at room temperature for 20 minutes. The sponges were then surgically placed into the hamstrings of each rat bilaterally. Each BMP (wild type and designer molecules) was placed into both limbs of 4 rats. Two weeks post implantation, the animals were sacrificed and the hamstrings were dissected, placed in 10% formalin and scanned by µCT (Scanco Inc.) to determine the amount of ectopic bone present. The amount of hydroxyapatite in milligrams (mg HA) present in the limbs of the treated animals is shown in Figure 9. Figure 9A shows the results for BMP2, BMPE and BMP2/6 heterodimer. Figure 9B shows the results for BMP2, BMPG, BMPA, and BMPF. For each of the designer BMPs, ectopic bone was formed at doses at which wild type BMP2 was unable to form a detectable bone mass. In a head-to-head comparison of wild type BMP2 with designer BMPE was able to induce ectopic bone to the same extent as wild type BMP2/6 heterodimer, closely matching the results obtained in the in vitro experiments disclosed previously. Designer BMPs BMPG, BMPA, and BMPF also demonstrated significantly higher ectopic bone formation compared to wild type BMP2 (Figure 9B). Results from this assay are shown in Figure 9 and also presented in Table 10.

TABLE 10

Name	Alk-phos	BRE-luc	Rat ectopic bone formation
BMP2WT	++	++	++
BMPA	++++	++++	++++
BMPB	++	++	++
BMPC	++	++	++
BMPD	++	++	++
BMPE	++++++	++++++	++++++
BMPF	++++	++++	++++
BMPG	+++++	+++++	+++++
BMPH	++	++	++

BMPI	+++	+++	+++
BMPJ	+	+	+
BMPD-P	++	++	++
BMP6-short	++++	++++	++++

EXAMPLE 3

BMP Receptor Binding

To further elucidate the mechanism of increased osteogenic activity of the designer BMPs, binding kinetic analysis of each of the designer BMPs with a panel of BMP receptors was performed using the Octet system (ForteBio, Menlo Park, CA). The Octet QK analysis was performed at degrees in TBS with 0.1% Tween-20. Samples were agitated at 1000 rpm. Anti-Human IgG Octet tips were saturated with 10 ug/mL of each receptor-human-IgG1-Fc fusion protein for 20min, which typically resulted in capture levels of the receptor that are saturated within a row of eight tips. Each BMP was prepared as a sevenfold serial dilution (typically 200-3 nM in singlicate) plus buffer blanks. Each Receptor/BMP binding pair was run at least in duplicate. Association was monitored for 10 min and dissociation was followed for 30 into buffer alone. Kinetic parameters (kon and koff) and affinities (KD) were calculated using the Octet Data Analysis software 6.0 using a partial binding 1:1 model following manufacturer's instructions.

The data set forth in Table 12 show that wild type BMP2 and BMP6 proteins each demonstrated the expected high affinity binding to type I (ALK-3 and ALK-6) and type II receptors (ActRIIA, ActRIIB, and BMPRII), respectively. The wild type BMP2/6 heterodimer exhibited high affinity binding to both groups of type I and type II receptors, as did designer BMPG, which the type II binding domains A and B of BMP2 have been replaced by the domains of wild type BMP6. Designer BMPE showed similar affinity as wild type BMP2 for the type II receptors as expected since no mutations were made in the type II binding regions. Unexpectedly, designer BMPE maintained high affinity binding for the type I receptors ALK-3 and ALK-6 with the type I binding domain of BMP6 which has been substituted in place for that of BMP2, while also unexpectedly binding the type I receptor ALK-2 with a KD of 2 nm. Thus, BMPE surprisingly gained a very high affinity binding for ALK-2 not observed in either WT BMP2 or WT BMP6.

TABLE 12

Receptor	BMP-2 (nM)	BMP-6 (nM)	BMP-2/6 (nM)	BMPE	BMPG
ALK2	>1000	>1000	250	2	<1000
ALK3	1	11	2	3	1
ALK6	1	20	0.5	1	1
ACTR IIA	53	3	2.5	40	2
ACTR IIB	8	0.5	1	6	0.5
BMPR IIA	62	4	3	82	4

As shown in Table 13, combining the mutations of BMPG and BMPE, comprising either proline or arginine at amino acid residue 36 (P36R) relative to the amino acid sequence of wild type BMP2 as set forth in SEQ ID NO:1, to produce BMP-GEP (also referred to as BMPGE P36) and BMP-GER (also referred to as BMPGE P36R), respectively, produced designer BMPs which demonstrated high affinity, low nM KDs, for all type I and Type II BMP receptors including ALK-2.

TABLE 13

Receptor	BMP-2	BMP-6	BMP-2/6	BMP-E	BMP-G	BMP-GER
ALK2	>1000	700	250	2	>1000	2
ALK3	1	11	2	3	1	2
ALK6	1	20	0.5	1	1	1
ACTRIIA	53	3	2.5	40	2	2
ATRIIB	8	0.5	1	6	0.5	0.5
BMPRIIA	62	4	3	82	4	3.5

Thus, the data disclosed herein demonstrate novel designer BMPs, such as, but not limited to, BMP-GER and BMP-GEP, which combine the attributes of BMP-G and BMP-E such that these novel designer BMPs demonstrate high affinity binding to a wide repertoire of both type I and type II receptors, including, but not limited to, ALK2, ALK3, ALK6, ActRIIA, ActRIIB and BMPRIIA. The data further demonstrated that replacing the proline at residue number 36 of the amino acid sequence of WT BMP2 (SEQ ID NO:1) to arginine produced a designer BMP that was as effective as an otherwise identical BMP where the amino acid was not replaced. These novel osteogenic BMPs as exemplified by BMP-GER, provide high levels of biologic activity thus allowing lower dosing and, in some cases, more rapid osteogenic response, strongly suggesting that these molecules would provide highly effective therapeutics.

EXAMPLE 4

In Vivo Osteogenic Activity in Non-human primates

NHP fibula osteotomy model

To further assess the potential therapeutic potential of the novel designer BMPs of the invention, the activity of designer BMPE and BMPG was compared to that of wild type BMP2 in an NHP (non-human primate) fibula osteotomy model.

A mid-diaphyseal osteotomy of the fibula was performed bilaterally with the 1-mm blade of an oscillating saw in adult male Cynomolgus monkeys (*Macaca fascicularis*) with a mean body weight (and standard deviation) of 7.5 ± 0.2 kg and an age range of seven to ten years. A small intramedullary Kirschner wire was added to the previously described fibular osteotomy model to maintain alignment of the proximal and distal bone ends for more uniform torsional biomechanical testing. The two major advantages of this model are the ability to utilize a bilateral study design as a result of the low morbidity of the procedure and the ability to remove a 6 to 8-cm segment of the fibula containing the osteotomy site for subsequent biomechanical and histological evaluation without having to sacrifice the animal. A 500 μL solution of 0.5 mg/ml of either wild type or designer BMP was added to a 30mm X 15mm ACS sponge. The sponge was wrapped around the defect following surgery. An approximately 2mm fracture of the fibula of each limb of a skeletally mature NHP was wrapped in an ACS sponge comprising either a designer BMP molecule at 0.5 mg/ml dose (250 μg total delivered) or the same amount of wild type BMP2 in the contralateral limb. Thus, each animal received wild type BMP in one limb and a designer BMP in the contralateral limb.

In this model, designer BMPE and BMPG were chosen since each represents a different class of designer molecule; designer BMPG shows high affinity for both type I and type II receptors while BMPE binds the type I receptor ALK-2 with high affinity in addition to binding type I receptors ALK-3 and ALK-6 with high affinity. Radiographs were obtained every 2 weeks to compare the healing of the limbs treated with the designer BMP molecule compared with the healing of the contralateral limb treated with wild type BMP2 in each animal. As shown in Figures 10A-10C, the data, which include seven animals from each group, demonstrated that the callus formed earlier and more robustly in the limbs treated with each designer BMP (BMPE shown in Figure 10A and BMPG shown in Figure 10B-10C) molecule compared to that with bone formation observed in the limb treated with wild type BMP2.

Tables 14 and 15, below, set forth data providing quantitative assessments of the difference in bone mass and bone volume observed between limbs treated with wild type BMP2 and limbs treated with designer BMPE. As shown in figure 11, BMPE administration resulted in an average of a 33% increase in bone volume (mm^3) when compared with bone volume increase in wild type BMP2 treated limbs. This μCT analysis included all the native bone where there was callus, accordingly, BMP-E was much more robust than BMP-2 in the same animals.

TABLE 14

Bone Mass (mg HA)

NHP	Left - BMPE	Right – WT BMP2	% increase vs. R
5304	721.2298	609.3317	18%
5604	561.4103	489.706	15%
8104	511.4216	313.4301	63%
9804	524.7777	474.0646	11%
16204	714.6123	536.7611	33%
17504	431.5738	406.1264	6%
22506	625.7583	466.0707	34%
		average	26%
		std dev	20%
		std error	7.40%
		paired t-test	p=.0040

TABLE 15

Bone volume (mm3)

NHP	Left - BMPE	Right – WT BMP2	% increase vs. R
5304	897.4342	720.0308	25%
5604	632.8525	564.9525	12%
8104	583.9513	336.0737	74%
9804	573.0165	507.0014	13%
16204	852.5689	551.2446	55%
17504	514.226	482.9475	6%
22506	766.8873	528.5033	45%
		average	33%
		std dev	25%
		std error	9.60%
		paired t-test	p=.0070

Replacement of P36 relative to wild type BMP2 with arginine did not affect activity of BMPGE

Proline at position 36 relative to the amino acid sequence of wild type BMP2 as set forth in SEQ ID NO:1 is purportedly important in conferring Noggin resistance and providing increased osteogenic activity to wild type BMP2 (see, e.g., WO 2009/086131). Therefore, to assess the effect of replacing P36 with a non-conserved amino acid substitution on the novel activity of BMPGE, P36 of BMPGEP was mutated to argininine to produce BMPGER and osteogenic activity of the two designer molecules was assessed in vitro. The data disclosed herein in Figure 12 demonstrate that replacing P36 with arginine (P36R) did not affect the binding affinity of the novel BMP-GE designer BMPs and both BMPGEP and BMPGER were as active as BMP2/6 heterodimer.

BMP-GER has in vivo activity comparable to BMP2/6 heterodimer

As shown in figures 13 and 14, rat ectopic experiments show that BMP-GER is as potent as BMP-2/6 at driving the formation of ectopic bone at the very low dose of 0.25 ug total BMP when all

molecules are delivered on an ACS sponge. Figure 13 shows that only BMP-2/6 and BMP-GER, but not BMPE or BMPG, were significantly more active than BMP-2 at this low dose when the milligrams of HA formed in the ectopic were quantified by μCT analysis.

The same samples were demineralized and scored for bone formation (Bone Score) by histology and these results are shown in Figure 14. By this method of scoring, at the low dose of 0.25ug delivered BMP-2 has no bone formation, and BMP-GER and 2/6 had the highest score. BMP-G and BMP-E were also significantly more potent than BMP-2 but not as active as BMP-GER.

Comparison of BMP-GER with BMP-2 in In Vivo models of osteogenesis and tissue repair

Figures 15 and 16 show the results of a severe NHP fibula ostectomy model comparing the activity of BMP-2 and BMP-GER. In this model a wedge with and approximate width of 4-6 mm was removed for each fibula of the NHP and put back in place and held with a titanium pin. The defect was then wrapped with an ACS sponge containing 250 ug total BMP at a dose of 0.5mg/ml. In each NHP BMP-2 was placed in one limb and BMP-GER was placed in the contralateral limb. Figure 15A shows photographs of radiographs taken at 5 weeks showing the defect in 4 of the 6 animals. The BMP-GER limbs showed significantly more robust bone formation than those with BMP-2. Figure 15B (bottom panel of the figure) shows μCT images of the fibulas of the same 4 animals following their sacrifice at week 10. As can be seen, the amount of bone formed is much more robust in the BMP-GER limbs than in the contralateral limbs treated with BMP2.

Figure 16A-C shows the analysis of these limbs comparing the strength, stiffness, and callus bone volume comparing the BMP-2 and BMP-GER treated limbs from each animal. On average the BMP-GER treated limbs required 21% more torque to break (Figure 16A), were 24% more stiff (Figure 16B), and the calluses were on average 55% larger (Figure 16C) than the contra lateral BMP-2 treated limb. All of these comparisons had a p value of less than .01 by pairwise analysis. These data show that BMP-GER induced fracture repair and bone formation significantly earlier and more robustly than BMP-2 in the same animal.

BMP-GER induced bone formation in an NHP model equivalently to BMP-2 at a 3 fold lower dose.

To further assess the effectiveness of BMPE bone formation in NHP, the ability of BMPE to induce osteogenesis in a wedge defect assay was compared to that of BMP2. Figure 17A-C shows radiographs of the bone formation following the wedge defect model in three non-human primates where 1.5mg/ml of BMP-2 was used in one limb and only 0.5 mg/ml of BMP-GER was used in the other limb using a calcium phosphate cement based carrier. Radiographically, the healing and bone formation were equivalent for each of the animals whether the treatment was with the high dose of BMP-2 or the lower dose of BMP-GER. Thus, even at one-third the dose, BMPE was equivalent to BMP2 in inducing bone formation, demonstrating the greatly increased activity of this designer BMP compared with wild type BMP2.

EXAMPLE 5

BMP Structural Analysis

Crystallization BMP-2 and BMP-6

Purified, fully-glycosylated wild type BMP2/6 heterodimer, wild type BMP2/2 homodimer, and wild type BMP6/6 homodimer, each produced in mammalian cells, were concentrated to 6-10 mg/ml in 10mM sodium acetate (pH 3.5), and crystallization attempts were performed using a "mosquito" automated robotic setup at 18°C (TTP LabTech Inc., Cambridge, MA). Initial crystallization hits were obtained for each dimer and the conditions were subsequently optimized to acquire crystals of good diffraction quality.

Crystals of wild type BMP2/6, BMP2/2 and BMP6/6 were transiently cryoprotected and frozen in liquid nitrogen prior to X-ray diffraction data collection at the synchrotron sources (ID beamline of Advanced Photon Source SER-CAT). Data were processed and scaled using programs Mosflm/Scala to deduce correct crystal lattice type and to integrate/scale data. The resolution and unit cell parameters are listed as follows: BMP2/6 belongs to the space group of $P4_32_12$ with two copies of the heterodimer per asymmetric unit; it diffracted to 2.8Å in one direction and 3.0Å in the other two, with a unit cell of a=b=105.23Å, c=188.73Å, α=β=γ=90°. BMP2/2 belongs to the space group of $P3_1$ with two copies of the homodimer per asymmetric unit; it diffracted to 2.7Å with a unit cell of a=b=62.74Å, c=126.35Å, α=β=90°, γ=120.° BMP6/6 belongs to the space group of $P3_121$ with one copy of the homodimer per asymmetric unit; it diffracted to 2.6Å with a unit cell of a=b=97.40Å, c=85.64Å, α=β=90°, γ=120°. Due to anisotropic diffracting nature of BMP2/6 crystals, the data was ellipsoidally truncated and anisotropically scaled to preserve contribution of high-resolution data.

The structures of CHO BMP2/6, BMP2/2, and BMP6/6 were determined by molecular replacement method with program Phaser, using *E coli* BMP2 (PDB accession: 1REW) and *E. coli* BMP6 (PDB accession: 2R52) as search models. After correct molecular replacement solutions were obtained and space groups confirmed, Phaser-calculated electron density maps were used to evaluate the quality of the search models, and regions in question (especially areas involving type I and type II receptor binding) were stripped from the original model for rebuilding in order to avoid model bias.

The structural models went through rigid-body refinement, followed by simulated annealing, positional and temperature factor refinement. Stripped areas were rebuilt using omit maps, and the processes were repeated along with TLS refinement until the refinement stabilized. The final refinement statistics are as follows: For BMP2/6, Rw/Rf = 0.2231/0.2775, rmsd bonds = 0.008, rmsd angles = 1.545; For BMP2/2, Rw/Rf = 0.2114/0.2659, rmsd bonds = 0.005, rmsd angles = 0.982; For BMP6/6, Rw/Rf = 0.2170/0.2510, rmsd bonds = 0.006, rmsd angles = 1.182. All three structures are in very good geometry based on Procheck results.

The CHO BMP2/6 crystal structure revealed extensive glycosylation. In particular, the prehelical loop of CHO-produced BMP2, which is an important binding motif for type I receptors, is different from the

corresponding region of E. coli-produced and refolded BMP2. In the presence of glycosylation, the CHO BMP2 loop has a uniquely "loopy" conformation when compared to the same region in bacterially refolded BMP2, which is more helical (Keller et al., *Nat Struct Mol Biol* 11:481-488 (2004)). The data demonstrated that the D53 of CHO-produced BMP2 points towards the receptor interface, while the H54 points away from the receptor as shown in Figure 3A. In E. coli BMP2, the D53 points away from the receptor and the H54 lines up toward the receptor (referred to herein as a "histidine doorstop"), stacking against a proline residue (P45) on the BMP2 type I receptor Alk3 as shown in Figure 3B (H54 is alternatively labeled H336). Without wishing to be bound by any particular theory, this stacking could prevent the type I receptor from fully binding to E. coli refolded BMP2, explaining the reduced binding activity of E. coli BMP2 when compared with CHO BMP2. This structural feature is illustrated in Figure 3A-B. In this figure, histidine 54 (H54) is numbered as H336, asparagine 56 (N56) is labeled N338, and P45 of ALK3 is shown in darker gray.

As illustrated in Figure 4, fully glycosylated CHO BMP6 also has this "doorstop" histidine residue pointing into the receptor binding site. This doorstop His structural motif is a common structural feature among BMPs (excluding CHO BMP2) (see, e.g., Keller et al., *Nat Struct Mol Biol* 11:481-8 (2004); *Kotzsch et al., EMBO J* 28:937-47 (2009). Without wishing to be bound by any particular theory, it may be that a specific glycan of CHO BMP2 is linked though extensive hydrogen bonding with arginine 16 ("the glycan tether" also designated as R298). This glycan tether is illustrated in Figure 4A and its interaction with the glycan is depicted using dotted lines between the glycan and this tether R298 which is also referred to herein as R16. Thus, without wishing to be bound by any particular theory, the glycan tether may serve to stabilize the conformation of the pre-helical loop of the BMP2 molecule such that the histidine doorstop, if otherwise present, is instead oriented away from the type I receptor interface thereby allowing the ligand to contact the receptor to a greater extent than in the presence of the histidine doorstop. In other words, the re-orientation of the histidine doorstop as observed in CHO BMP2 is most likely to be the consequence of glycan tethering. Without wishing to be bound by any particular theory, the data disclosed herein suggest that where the histidine doorstop is present, removal of the doorstop in the absence of glycosylation (i.e., by introducing a mutation that changes the orientation of the His away from the receptor interface) increases binding of the BMP ligand with the type I receptor.

Designer BMPE, which contains a low affinity type II binding domain of BMP2 and a low affinity type I binding domain similar to that of BMP6, shows (1) increased osteogenic activity in both in vitro and in vivo assays; and (2) has an unexpected gain of function to bind Alk2, a type I receptor, despite the presence of a low affinity type I receptor binding domain. Without wishing to be bound by any particular theory, it may be that this surprising discovery is mediated by multiple hydrogen bonds formed between the glycan moieties and the R16 (the "glycan tether") in the type I receptor-binding domain of BMPE. This tethering interaction may mediate a structural rearrangement at the pre-helical region of the BMPE molecule that presents a proper binding surface for Alk2 by positing H54 (the "doorstop") away from the interface thereby allowing closer interaction between the BMP and the receptor. In contrast, as illustrated

in Figure 4B, BMP6, which also has a low affinity type I binding domain similar to that of BMPE, does not bind Alk2 because its "glycan tether" (R413) which would be needed to tether its glycan moieties, is shifted in location when compared to the BMPE tether (R298/R16). Thus, in BMP6, the glycan is not tethered and the doorstop (H454) is not positioned away from the ligand-receptor interface. The "glycan tether" appears to be a phenomenon unique to wild type glycosylated BMP2 (as exemplified by BMP2 produced in CHO cells), and structural remodeling of the prehelical loop of BMPs by introducing (or removing) "glycan tether" can now be used, for the first time, to modulate type I receptor-binding ability of other BMPs. Therefore, one skilled in the art, now armed with the teachings provided herein, would understand how to mutate the BMP in order to position the doorstop away from the receptor interface by introducing mutations that swing the H54 away or by affecting the glycan tether so that tethering mediates the shift in H54 and would further appreciate that these teachings can be used to design a BMP with increased (or decreased if mutations are introduced to swing H54 into the doorstop position) binding to its receptor or to create designer BMPs with gain-of-function mutations such that they bind to novel receptors that they did previously bind. As more fully set forth below, the present invention demonstrates how to use this novel doorstop/tether design method to design improved osteogenic proteins. Thus, the present invention provides a novel method for rational design of improved osteogenic proteins comprising altered receptor binding.

In order to more fully understand what drives the BMP-E and BMP-GER binding to ALK-2, and to further elucidate this novel mechanism of affecting receptor binding using the doorstop/glycan tether, the crystal structure of BMP-E was solved and compared to that of BMP-2 and BMP-6. The key structural findings are shown in Figures 18 and 19. As illustrated in Figure 18 BMP-E maintains the ordered sugar of BMP-2 while maintaining the central helical structure of BMP-6. The structure shown in Figure 18 demonstrates that BMP-E, and presumably BMP-GER, is different from both BMP-2 and BMP-6 in the critical region of type I receptor binding. Figure 19 is a blow-up comparing the area surrounding the potential His doorstop of BMPE (light gray) and BMP6 (dark gray). The diagram demonstrates the similarity of the alignment of the histidine and asparagine in both molecules and also shows the difference in glycan positioning and demonstrating the tethering of the BMPE glycan by R16 (the tether) which also causes a more rigid conformation of the glycan such that a longer glycan is rendered for BMPE by the analysis compared with the shorter glycan rendered for BMP6 (in dark gray).

In order to determine if the glycan of BMP-E is driving the interaction with ALK-2 and its higher activity, BMP-2, BMP-6, and BMP-E were treated with Endo H to clip the sugar down to two GlcNac units. The binding affinity of BMP-E for AIK-2 decreased to 400 nM whereas it's affinity for ALK-3 and ALK-6 were still in the 3-6 nM range showing the intact carbohydrate is extremely important for this interaction. The activity of this deglycosylated mutant also decreased significantly. As shown in Figure 20, in this experiment the Endo H treated deglycosylated BMP-E activity shifts to the right and is almost equivalent to BMP-6 WT. The EC-50 shifts from 3nM to approximately 50 nM. These data show that the carbohydrate of BMP-E is essential for its activity, and this should translate to BMP-GER since it has the

exact same region of BMP-6 substituted into BMP-2 with only the finger domains differing. Since the carbohydrate is essential for increased receptor binding and osteogenic activity, these results indicate indicates that production of BMP-E or BMP-GER in E. coli, or any other system lacking glycosylation, would not produce a BMP with activity superior to BMP-2 WT.

Crystallization BMP-E and BMP-GER

Purified, fully-glycosylated BMP-E, was concentrated to 8.7 mg/ml in 25mM sodium acetate (pH 3.5), and crystallization attempts were performed using a "mosquito" automated robotic setup at 18°C (TTP LabTech Inc., Cambridge, MA). Initial crystallization hits were obtained for each dimer and the conditions were subsequently optimized to acquire crystals of good diffraction quality.

Crystals of BMP-E were transiently cryoprotected and frozen in liquid nitrogen prior to X-ray diffraction data collection at the synchrotron sources (ID beamline of Advanced Photon Source SER-CAT). Data were processed and scaled using programs Mosflm/Scala in the CCP4 package to deduce correct crystal lattice type and to integrate/scale data. The resolution and unit cell parameters are listed as follows: BMPE belongs to the space group of $P4_32_12$ with two copies of the BMPE in each asymmetric unit; it diffracted to 2.7Å, with a unit cell of a=b=67.78Å, c=148.01Å, α=β=γ=90°.

The structure of BMPE was determined by molecular replacement method with program Phaser, using fully glycosylated CHO BMP2 and BMP6, both determined at Pfizer, as search models. After correct molecular replacement solutions were obtained and space groups confirmed, Phaser-calculated electron density maps were used to evaluate the quality of the search models, and regions in question (especially areas around type I receptor binding and glycosylation) were stripped from the original model for rebuilding in order to avoid model bias.

The structural model of BMPE went through rigid-body refinement, followed by simulated annealing, positional and temperature factor refinement using program Phenix. Stripped areas were rebuilt using omit maps, and the processes were repeated along with TLS refinement until the refinement stabilized. The final refinement statistics are: Rw/Rf = 0.2252/0.2840, rmsd bonds = 0.006, rmsd angles = 0.935. The structure is in very good geometry based on Procheck results.

BMPE, a designer molecule wherein residues 44-80 of BMP2 replaced by the corresponding region from BMP6, maintains the overall framework of BMP2 while possessing the Type I receptor-binding segment of BMP6. As shown in Figure 21, the crystal structure revealed that the grafted segment still retains a similar conformation as in BMP6, forming a small helix in the pre-helical loop within which the "doorstop" H54 points toward the receptor. However, without wishing to be bound by any particular theory, it may be that due to the presence of "glycan tethers" at R16 and E110* (E109 of BMP-2), both of which form multiple hydrogen bonds with the third and forth glycan moieties (β-mannose and α-mannose, respectively), the extended glycosylation chain is attached to the protein surface, exactly as seen in CHO BMP2. The tethering of glycan chain also dislocated the pre-helical loop by about 2Å in reference to the overall framework. Without wishing to be bound by any particular theory, it may be that the surprising discovery that the BMP6-like pre-helical loop combined with the BMP2-like glycosylation present a

binding epitope for the Alk2 receptor, which does not normally interact with either BMP2 or BMP6. Deglycosylation renders BMPE incapable of binding to Alk2, which underscores the importance of glycosylation in mediating Alk2 recognition for BMPE.

EXAMPLE 6

Noggin Resistance

In order to investigate if resistance to the secreted BMP inhibitor Noggin would increase the activity of BMP-GER or BMP-E, these potential therapeutic molecules were further modified to potentially increase their resistance to Noggin. Recently, it was demonstrated that in E. coli-produced proteins, incorporation of a C-terminal portion of activin-A into wild type BMP2 increased resistance to Noggin inhibition. See WO 2010/099219 at, e.g., Figures 15 and 16. Therefore, to determine whether the novel designer proteins disclosed herein could be improved even further by incorporation of activin-A sequences, the Noggin resistance (NR) amino acid sequences were substituted into BMP-E (SEQ ID NO:12) and BMP-GER (SEQ ID NO:37) to produce BMP-E-NR (SEQ ID NO:70) and BMP-GER-NR (SEQ ID NO:71). As shown in Figure 22 BMP-E-NR and BMP-GER-NR have equivalent in vitro activity in an Alkaline phosphate activity assay compared with BMP-E and BMP-GER and are completely resistant to Noggin while BMP-E and BMP-GER are sensitive to Noggin.

To understand the potential basis for the Noggin resistance demonstrated in vitro by BMPE-NR and BMP-GER-NR, the binding affinity of these molecules for the type II activin receptor ActRIIB was assessed. As shown in Table 16, below, activin-A is unable to bind Noggin but the Noggin resistant BMP-E-NR and BMP-GER-NR bind Noggin, but not as strongly as BMP-2, BMP-E, or BMP-GER. These data also show that the Noggin resistant BMPs bind the type II BMP receptor ActRIIB with extremely high affinity that is even higher than that of BMP-GER. Without wishing to be bound by any particular theory, these data suggests that BMP-GER-NR and BMP-E-NR are resistant to Noggin due to their much higher affinity for the BMP type II receptors than that of Noggin and are therefore able to bind BMP receptors even in the presence of high amounts of Noggin.

TABLE 16

	ActRIIb affinity (nM)
BMP-E	9.00
BMP-E-NR	0.50
BMP-GER	2.00
BMP-GER-NR	0.07
	Noggin Affinity
BMP-E	1.00
BMP-E-NR	6.00
BMP-GER	4.00

BMP-GER-NR	7.50
	ActRIIb on rate (K on) with equal molar ration of noggin*
BMP-E	no binding
BMP-E-NR	no binding
BMP-GER	1.00E+03
BMP-GER-NR	1.00E+06
*little to no change in on rate with up to 10 fold molar excess of Noggin	

Although the BMP-E and BMP-GER molecules comprising the Noggin resistant portions of activin-A demonstrated Noggin resistance in vitro, these results did not correlate to improved in vivo activity. That is, when the osteogenic activity of these BMP-E-NR and BMP-GER-NR was compared with that of BMP-E and BMP-GER in a rat ectopic assay, the NR molecules were much less potent. This data is shown in Figures 23 and 24. More specifically, the Bone Score for BMP-GER and BMP-GER-NR was compared and, at all concentrations tested (0.125 µg, 0.25 µg, 0.5 µg, and 1.0 µg), BMP-GER greatly outperformed BMP-GER–NR as shown in Figure 23. Similarly, Figure 24 demonstrates that BMP-E produced a much higher Bone Score compared with BMP-E-NR in this in vivo assay. Thus, for both BMP-E and BMP-GER the purportedly Noggin resistant versions were much less potent in vivo than their NR (Noggin resistant) counterparts, and in the case of BMP-E, almost all in vivo activity was lost due to incorporation of sequences of activin-A (see Figure 24 comparing BMP-E-NR with BMP-E).

These data demonstrate that incorporation of sequences potentially conferring Noggin resistance, while increasing binding for certain type II receptors (e.g., ActRIIB), did not increase in vivo osteogenic activity of the designer BMP.

Further, although the addition of Noggin did not improve the osteogenic activity of the designer BMPs in vivo, indeed, it appeared to decrease their in vivo activity, the novel designer BMPs of the invention demonstrate greatly increased osteogenic characteristics compared with wild type BMP and provide potential novel therapeutics for a wide variety of applications even without demonstrating Noggin resistance in vitro. Therefore, the designer BMPs of the invention provide remarkable novel potential therapeutics demonstrating a greatly improved clinical profile for, among other uses, bone augmentation and repair.

The disclosures of each and every patent, patent application, and publication cited herein are hereby incorporated herein by reference in their entirety.

While the invention has been disclosed with reference to specific embodiments, it is apparent that other embodiments and variations of this invention may be devised by others skilled in the art without departing from the true spirit and scope of the invention. The appended claims are intended to be construed to include all such embodiments and equivalent variations.

CLAIMS

1. A designer BMP protein comprising at least one mutation in at least one type I or type II receptor binding domain, wherein the mutation confers altered binding to the type I or type II BMP receptor compared with the binding to the type I or type II receptor by a corresponding wild type BMP.

2. The protein of claim 1, wherein the protein is selected from the group consisting of BMP2, BMP4, BMP5, BMP6, BMP7, BMP8 and BMP9.

3. The protein of claim 1, comprising at least one mutation within:
   a. the type II binding domain A;
   b. the type II binding domain B;
   c. the type I binding domain; and
   d. any combination of the above.

4. A designer osteogenic protein comprising an amino acid sequence comprising at least one mutation in at least one type I or type II receptor binding domain, wherein the mutation confers altered binding to the type I or type II BMP receptor compared with the binding to the type I or type II receptor by wild type BMP2.

5. The protein of claim 4, wherein the mutation is selected from the group consisting of:

   a. a mutation within the type II binding domain A wherein said mutation is at least one mutation selected from the group consisting of a mutation at V33, P36, H39, and F41 with respect to the sequence of SEQ ID NO:1;
   b. a mutation within the type II binding domain A wherein said mutation is at least one mutation selected from the group consisting of V33I, P36K, P36R, H39A, and F41N with respect to SEQ ID NO:1;
   c. a mutation within the type II binding domain B wherein said mutation is at least one mutation selected from the group consisting of a mutation at E83, S85, M89, L92, E94, E96, K97, and V99 with respect to the sequence of SEQ ID NO:1;
   d. a mutation within the type II binding domain B wherein said mutation is at least one mutation selected from the group consisting of E83K, S85N, M89V, L92F, E94D, E96S, K97N, and V99I with respect to of SEQ ID NO:1;
   e. a mutation within the type I binding domain wherein said mutation is at least one mutation selected from the group consisting of a mutation at H44, P48, A52, D53,

L55, S57, N68, S69, V70, an insertion of a single amino acid after N71, S72, K73, I74, A77, and V80 with respect to the sequence of SEQ ID NO:1; and

f. a mutation within the type I binding domain wherein said mutation is at least one mutation selected from the group consisting of H44D, P48S, A52N, D53A, L55M, S57A, N68H, S69L, V70M, insertion of P after N71, S72E, K73Y, I74V, A77P, and V80A with respect to the sequence of SEQ ID NO:1.

6. The protein of claim 4, wherein said protein comprises mutations selected from the group consisting of:

a. a mutation at each of amino acids H44, P48, A52, D53, L55, S57, N68, S69, V70, insertion of a single amino acid after N71, S72, K73, I74, A77, and V80 with respect to the sequence of SEQ ID NO:1;

b. the protein of (a) wherein the mutations are H44D, P48S, A52N, D53A, L55M, S57A, N68H, S69L, V70M, insertion of a P after N71, S72E, K73Y, I74V, A77P, and V80A;

c. a mutation at each of amino acids V33, P36, H39, S85, M89, L92, E94, E96, K97, and V99 with respect to the sequence of SEQ ID NO:1;

d. the protein of (c) wherein the mutations are V33I, P36K, H39A, S85N, M89, L92F, E94D, E96S, K97N, and V99I;

e. a mutation at each of amino acids V33, P36, H39, H44, P48, A52, D53, L55, S57, N68, S69, V70, insertion of a single amino acid after N71, S72, K73, I74, A77, and V80, S85, M89, L92, E94, E96, K97, and V99 with respect to the sequence of SEQ ID NO:1; and

f. the protein of (e) wherein the mutations are V33I, P36K, H39A, H44D, P48S, A52N, D53A, L55M, S57A, N68H, S69L, V70M, insertion of a P after N71, S72E, K73Y, I74V, A77P, and V80A, S85N, M89, L92F, E94D, E96S, K97N, and V99I; and

g. the protein of (e) wherein the mutations are V33I, P36R, H39A, H44D, P48S, A52N, D53A, L55M, S57A, N68H, S69L, V70M, insertion of a P after N71, S72E, K73Y, I74V, A77P, and V80A, S85N, M89, L92F, E94D, E96S, K97N, and V99I.

7. The protein of claim 1, wherein the protein binds the ALK2 receptor with a K_D not greater than about 2 nM, the protein binds the ALK3 receptor with a K_D not greater than about 2 nM, the protein binds the ALK6 receptor with a K_D not greater than about 1 nM, the protein binds the ActRIIA receptor with a K_D not greater than about 2 nM, the protein binds the ActRIIB receptor with a K_D not greater than about 0.5 nM, and protein binds the BMPRIIA receptor with a K_D not greater than about 3.5 nM.

8. The protein of claim 1, further comprising 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid mutations not located within the type I or the type II binding regions.

9. A designer osteogenic protein selected from a protein comprising the amino acid sequence of any one of SEQ ID NOs:8-73.

10. A designer osteogenic protein comprising the amino acid sequence of SEQ ID NO:12.

11. A designer osteogenic protein comprising the amino acid sequence of SEQ ID NO:14.

12. A designer osteogenic protein comprising the amino acid sequence of SEQ ID NO:36.

13. A designer osteogenic protein comprising the amino acid sequence of SEQ ID NO:37.

14. A method of producing the protein of claim 1, the method comprising introducing a nucleic acid encoding the protein into a host cell, culturing the cell under conditions where the protein is produced, and purifying the protein.

15. The method of claim 14, wherein said nucleic acid comprises a sequence selected from the group consisting of the nucleic acid sequence of any one of SEQ ID NOs:74-139.

16. A designer BMP6 protein comprising an amino acid sequence comprising at least one mutation in at least one type I or type II receptor binding domain, wherein the mutation confers altered binding to the type I or type II BMP receptor compared with the binding to the type I or type II receptor by wild type BMP6.

17. The protein of claim 16, wherein the mutation is selected from the group consisting of:

a. a mutation within the type II binding domain A wherein said mutation is at least one mutation selected from the group consisting of a mutation at I57, K60, G61, A63, N65, Y66, and D68 with respect to the sequence of SEQ ID NO:4;
b. a mutation within the type II binding domain B wherein said mutation is at least one mutation selected from the group consisting of K108, N110, A111, V114, F117, D119, N120, S121, N122, V123, and I124 with respect to the sequence of SEQ ID NO:4;
c. a mutation within the type I binding domain wherein said mutation is at least one mutation selected from the group consisting of a mutation at S72, N76, A77, H78, M79, N80, A81,

N83, V87, T89, H92, L93, M94, N95, P96, E97, Y98, V99, and P100 with respect to the sequence of SEQ ID NO:4;

d. a mutation at each of amino acid residues I57, K60, G61, A63, N65, Y66, and D68 with respect to the sequence of SEQ ID NO:4;

e. a mutation at each of amino acid residues K108, N110, A111, V114, F117, D119, N120, S121, N122, V123, and I124 with respect to the amino acid sequence of SEQ ID NO:4; and

f. a mutation at each of amino acid residues S72, N76, A77, H78, M79, N80, A81, N83, V87, T89, H92, L93, M94, N95, P96, E97, Y98, V99, or P100 with respect to the amino acid sequence of SEQ ID NO:4.

18. The protein of claim 16, further comprising 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acid mutations not located within the type I or the type II binding domains.

19. An isolated nucleic acid molecule comprising a nucleotide sequence encoding an amino acid sequence selected from the group consisting of SEQ ID NOs:8 to 73.

20. The nucleic acid of claim 19, said nucleic acid encoding a protein comprising an amino acid sequence selected from the group consisting of the sequence of SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:36 and SEQ ID NO:37.

21. An isolated nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of SEQ ID NOs:74 to 139.

22. The nucleic acid of claim 21, said nucleic acid comprising a nucleotide sequence selected from the group consisting of the sequence of SEQ ID NO:78, SEQ ID NO:80, SEQ ID NO:102, and SEQ ID NO:103.

23. A method of producing the protein of claim 16, the method comprising introducing a nucleic acid encoding said protein into a host cell, culturing said cell under conditions where said protein is produced, and purifying said protein.

24. A method of treating a bone disease associated with bone loss in a patient in need thereof, the method comprising administering an effective amount of a designer osteogenic protein of claim 1 to said patient, thereby treating bone disease in said patient.

25. A method of treating fibrosis in a patient in need thereof, the method comprising administering an effective amount of a designer osteogenic protein of claim 1 to said patient, thereby treating fibrosis.

26. A method of inducing bone formation in a tissue, said method comprising contacting said tissue with a designer osteogenic protein of claim 1, thereby inducing bone formation in said tissue.