

PROBE COMBINATION FOR DETECTION OF CANCER

CROSS-REFERENCE TO RELATED APPLICATION

[0001] The present disclosure claims priority to U.S. provisional patent application No. 62/456087, filed on February 7, 2017, the entirety of which are incorporated herein by reference.

FIELD OF THE INVENTION

[0002] The present disclosure relates to a probe combination, and more particularly, to a combination of sequence-specific probes for detection of cancer.

BACKGROUND OF THE INVENTION

[0003] *Hepadnaviridae* is a family of viruses that has been reported to associate with pathogenesis of hepatitis, hepatocellular carcinoma (HCC) and cirrhosis. Hepatitis B virus (HBV) is among the most common members of the hepadnavirus family and is a small DNA virus that can be classified into genotypes A to J. While most adults infected with HBV can recover, about 5-10% of HBV infected patients are unable to clear the virus and become chronically infected. Those with chronic HBV infection are at high risk of developing HCC as HBV is capable of integrating into host genome and causing genetic and epigenetic alterations in hepatocytes.

[0004] A few methods for detection of HBV integration have been reported. For example, Jiang in “The effects of hepatitis B virus integration into the genomes of hepatocellular carcinoma patients” (*Genome Res.* (2012) 22, 593-601) and Sung in “Genome-wide survey of recurrent HBV integration in hepatocellular carcinoma” (*Nature Genetics* (2012) 44, 765-769) disclosed to utilize whole genome sequencing to detect HBV integration in HCC liver samples. However, efficiency of these direct sequencing methods was poor. As reported by Jiang, as much as 25-35 million 75-bp reads on average were generated for each data set, and the typical numbers of HBV and junctions reads for Jiang’s data sets were 6 million and only 400 reads, respectively. Further, there has yet been any direct sequencing based studies that can detect HBV integration from circulating tumor DNA (ctDNA) samples.

[0005] Later on, Li in “HIVID: An efficient method to detect HBV integration using low coverage sequencing” (*Genomics* (2013) 102:4, 338-344)” and Zhao in “Genomic and oncogenic preference of HBV integration in hepatocellular carcinoma” (*Nature Communications* (2016) 7:12992) disclosed the use of sequence-capture probes designed according to sequences the HBV genome for detection of HBV integration. However, neither Li nor Zhao provided a clear idea regarding the design rationale of their probes. Furthermore, efficiency of the probes reported by Li and Zhao was poor. In both Li and Zhao, the average human ratio was as high as 83.7% and the average HBV alignment ratio and average integration rate were as low as 0.08% and 0.01% respectively, suggesting that the probes were still inefficient and ineffective in detecting HBV integration.

BRIEF SUMMARY OF THE INVENTION

[0006] The present invention provides a panel of probe combinations and the analytic methodology to be used therewith that are highly sensitive and efficient in capture viral DNA and viral-host junctions.

[0007] An embodiment of the present invention provides a probe combination for detecting cancer. The probe combination includes one or more sets of partial hepatitis B virus (HBV) targeting probes. When sequences of each of the sets of partial HBV targeting probes are aligned, an overall sequence of the aligned partial HBV targeting probes matches a reference sequence of a direct repeat (DR) region of a genome of a HBV genotype. In the aligned set of partial HBV targeting probes, each of the partial HBV targeting probes overlap with one or two adjacent partial HBV targeting probes by a portion of a length of the partial HBV targeting probes.

[0008] In a preferred embodiment, the HBV genotype includes genotype A, genotype B, genotype C, genotype D, genotype E, genotype F, genotype G, genotype H, genotype I and genotype J.

[0009] In a preferred embodiment, the reference sequence of the DR region includes SEQ ID NOs. 3-32.

[0010] In a preferred embodiment, the probe combination includes or further includes one or more sets of full HBV targeting probes. When sequences of each of the sets of full

HBV targeting probes are aligned, an overall sequence of the aligned full HBV targeting probes matches a reference sequence of the genome of the HBV genotype. In the aligned set of full HBV targeting probes, each of the full HBV targeting probes overlap with one or two adjacent full HBV targeting probes by a portion of a length of one of the full HBV targeting probes.

[0011] In a preferred embodiment, the probe combination further includes one or more sets of hotspot gene targeting probes. When sequences of the each of the sets of hotspot gene targeting probes are aligned, an overall sequence of the aligned hotspot gene targeting probes matches a reference sequence of a cancer hotspot gene. In the aligned set of hotspot gene targeting probes, each of the hotspot gene targeting probes overlap with one or two adjacent hotspot gene targeting probes by a portion of a length of the hotspot gene targeting probes.

[0012] In a preferred embodiment, the cancer hotspot gene includes CTNNB1, TERT, and TP53 genes.

[0013] In a preferred embodiment, the reference sequence of the cancer hotspot gene comprises SEQ ID NOs. 33-41.

[0014] In a preferred embodiment, the probe combination further includes one or more sets of exogenous gene targeting probes. When sequences of the exogenous gene targeting probes are aligned, an overall sequence of the aligned set of exogenous gene targeting probes matches a reference sequence of an exogenous gene. In the aligned set of exogenous gene targeting probes, each of the exogenous gene targeting probes overlap with one or two adjacent exogenous gene targeting probes by a portion of a length of the exogenous gene targeting probes.

[0015] In a preferred embodiment, the exogenous gene originates a lambda phage.

[0016] In a preferred embodiment, the reference sequence of the exogenous gene comprises SEQ ID NOs. 42-54.

[0017] In a preferred embodiment, the probe combination further includes one or more sets of endogenous gene targeting probes. When sequences of the endogenous gene targeting probes are aligned, a sequence of the aligned set of endogenous gene targeting probes matches a reference sequence of an endogenous gene. In the aligned set of endogenous gene targeting probes, each of the endogenous gene targeting probes overlap

with one or two adjacent endogenous gene targeting probes by a portion of a length of the endogenous gene targeting probes.

[0018] In a preferred embodiment, the endogenous gene includes GAPDH and GdX genes.

[0019] In a preferred embodiment, the reference sequence of the endogenous gene comprises SEQ ID NO. 55 and SEQ ID NO. 56.

[0020] Preferably, the cancer detected by the probe combination of the various embodiment includes hepatocellular carcinoma.

[0021] Preferably, the probe combination of the various embodiments is used for capturing target nucleotide fragments having viral-host junctions from DNA obtained for a specimen of a subject infected with HBV.

[0022] Preferably, the DNA obtained from the specimen includes genomic DNA and circulating tumor DNA (ctDNA) of the subject.

[0023] Preferably, the specimen comprises biological fluid and liver tissues.

[0024] In sum, the present invention according to the aforementioned embodiments provides a powerful and versatile tool for detection of viral infection and viral infection induced cancer. The embodiments of the present invention can be applied to detect presence of various types of DNA viruses and viral integration. The probe combination designed according to the embodiments ensures optimal viral/host sequence coverage and considers genetic stability, and is thus demonstrated to be highly sensitive, efficient, and reliable.

BRIEF DESCRIPTION OF THE DRAWINGS

[0025] The accompanying drawings illustrate one or more embodiments of the present invention and, together with the written description, explain the principles of the present invention. Wherever possible, the same reference numbers are used throughout the drawings to refer to the same or like elements of an embodiment.

[0026] FIG. 1 is a flowchart depicting the steps of obtaining a reference sequence in accordance with an embodiment of the present invention;

[0027] FIGs. 2A and 2B are schematic illustrations showing the design concept of the probes in accordance with an embodiment of the present invention;

[0028] FIG. 3 is an experimental result showing the selective hybridization of the probes designed in accordance with an embodiment of the present invention;

[0029] FIGs. 4A and 4B are experimental results showing the specificity of the probes designed in accordance with an embodiment of the present invention;

[0030] FIG. 5A is a heat map converted bar chart showing the next generation sequencing (NGS) results at various genetic regions in a paired tumor genomic DNA (gDNA) sample by using a probe combination in accordance with an embodiment of the present invention;

[0031] FIG. 5B is a heat map converted bar charts showing the NGS results at various genetic regions in a paired plasma circulating tumor DNA (ctDNA) sample by using the probe combination in accordance with an embodiment of the present invention;

[0032] FIG. 6 is a heat map converted bar charts showing the NGS results at various genetic regions in a tumor gDNA sample by using a probe combination in accordance with an embodiment of the present invention; and

[0033] FIG. 7 is an experimental result showing the NGS statistics of DNA samples hybridized by a probe combination in accordance with an embodiment of the present invention.

[0034] In accordance with common practice, the various described features are not drawn to scale and are drawn to emphasize features relevant to the present disclosure. Like reference characters denote like elements throughout the figures and text.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

[0035] The present invention will now be described more fully hereinafter with reference to the accompanying drawings illustrating various exemplary embodiments of the invention. The present invention may, however, be embodied in many different forms and should not be construed as limited to the embodiments set forth herein. Rather, these embodiments are provided so that this disclosure will be thorough and complete, and will fully convey the scope of the disclosure to those skilled in the art. Like reference numerals refer to like elements throughout.

[0036] The terminology used herein is for the purpose of describing particular embodiments only and is not intended to be limiting of the disclosure. As used herein, the

singular forms “a”, “an” and “the” are intended to include the plural forms as well, unless the context clearly indicates otherwise. It will be further understood that the terms “comprises” and/or “comprising,” or “includes” and/or “including” or “has” and/or “having” when used herein, specify the presence of stated features, regions, integers, steps, operations, elements, and/or components, but do not preclude the presence or addition of one or more other features, regions, integers, steps, operations, elements, components, and/or groups thereof.

[0037] It will be understood that the terms “and/or” and “at least one” include any and all combinations of one or more of the associated listed items. It will also be understood that, although the terms first, second, third etc. may be used herein to describe various elements, components, regions, parts and/or sections, these elements, components, regions, parts and/or sections should not be limited by these terms. These terms are only used to distinguish one element, component, region, part or section from another element, component, region, layer or section. Thus, a first element, component, region, part or section discussed below could be termed a second element, component, region, layer or section without departing from the teachings of the present disclosure.

[0038] Unless otherwise defined, all terms (including technical and scientific terms) used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this disclosure belongs. It will be further understood that terms, such as those defined in commonly used dictionaries, should be interpreted as having a meaning that is consistent with their meaning in the context of the relevant art and the present disclosure, and will not be interpreted in an idealized or overly formal sense unless expressly so defined herein.

[0039] An aspect of the present invention provides a probe combination that includes one or more sets of sequence targeting probes. The probes may include single stranded oligonucleotides and polynucleotides, such as single stranded deoxyribonucleic acids (ssDNA), ribonucleic acids (RNA), and artificial nucleotides. The probe combination may be used for detection of viral infection or viral infection induced cancer, especially those caused by or associated with DNA virus. In some embodiments, the probe combination may be used for detecting infection with hepatitis B virus (HBV), human papillomavirus (HPV), Epstein-Barr virus (EBV), herpes virus 8 (HHV-8), human T-

lymphotropic virus (HTLV), Merkel cell polyomavirus (MCV), or other DNA virus. In other embodiments, the probe combination may be used for detection for hepatocellular carcinoma, liver cancer, cervical cancer, penile cancer, anal cancer, vaginal cancer, vulvar cancer, oral cancer, oropharyngeal cancer, nasopharyngeal cancer, head and neck cancer, lymphoma, primary effusion lymphoma, stomach cancer, Kaposi sarcoma, Merkel cell carcinoma, or other cancer associated with infection with the DNA viruses.

[0040] According to an embodiment of the present invention, the probe combination includes one or more sets of full viral sequence targeting probes. When sequences of each of the sets of full viral sequence targeting probes are aligned, an overall sequence of the aligned set of full viral sequence targeting probes matches a reference sequence of a genome of a genotype of a target virus. The target virus may include various genotypes of the aforementioned DNA viruses. For example, in cases where HBV is the target virus, the genotype thereof may include genotype A, genotype B, genotype C, genotype D, genotype E, genotype F, genotype G, genotype H, genotype I and genotype J. The reference sequence of the viral genome may be retrieved from the NCBI GenBank or calculated from sequences obtained from clinical specimens. For example, reference sequences for HBV genotype A may be retrieved from NCBI GenBank (<https://www.ncbi.nlm.nih.gov/genbank/>) Accession No. AP007263, HE974383 or HE974381; reference sequences for HBV genotype B may be retrieved from GenBank Accession No. AB981581, AB602818, or AB554017; reference sequences for HBV genotype C may be retrieved from GenBank Accession No. LC360507, AB644287 or AB113879; reference sequences for HBV genotype D may be retrieved from GenBank Accession No. HE815465, HE974382 or AB554024; reference sequences for HBV genotype E may be retrieved from GenBank Accession No. HE974380, HE974384, AP007262; reference sequences for HBV genotype F may be retrieved from GenBank Accession No. DQ823095, AB036909 or AB036920; reference sequences for HBV genotype G may be retrieved from GenBank Accession No. AB625342, HE981176 or GU563559; reference sequences for HBV genotype H may be retrieved from GenBank Accession No. AB298362, AB846650, AB516395; reference sequences for HBV genotype I may be retrieved from GenBank Accession No. EU833891, KF214680 or KU950741; and reference sequences for HBV genotype J may be retrieved from

GenBank Accession No. AB486012.

[0041] In an exemplary embodiment, the probe combination includes two sets of full HBV targeting probes. When sequences of one set of the full HBV targeting probes are aligned, an overall sequence of the aligned full HBV targeting probes matches a reference sequence of a genome of HBV genotype B (SEQ ID NO. 1). Likewise, when sequences of the other sets of the full HBV targeting probes are aligned, an overall sequence of the aligned full HBV targeting probes matches a reference sequence of a genome of HBV genotype C (SEQ ID NO. 2). In the exemplary embodiment, the reference sequence of the HBV genome is obtained as shown in FIG. 1. As in Step S1, DNA were extracted from specimens, such as biological fluid (e.g., blood, lymph, urine, sweat, saliva, tears, or intestinal fluid) and tissues (e.g., liver tissues), of patients with chronic HBV infection. The extracted DNA contains genomic DNA (gDNA) and/or circulating tumor DNA (ctDNA) of the patients. As in Step S2, the extracted DNA are sequenced and classified according to known sequences of various HBV genotypes as retrieved from the NCBI GenBank. In the exemplary embodiment, HBV genotype B and HBV genotype C were chosen for their prevalence among hepatocellular carcinoma (HCC) patients in Taiwan; however, embodiments of the present invention are not limited only to genotypes B and C of HBV, but encompass various genotypes of all DNA viruses that can cause persistent infection. As in Step S3, sequence of each of the chosen genotypes was aligned and calculated by the Clustal algorithm to obtain a consensus sequence of the virus genotype according to the major allele (i.e., the most common nucleotide) at each position of the sequence. Finally, the consensus sequence is used as a reference sequence for the virus genotype. In the exemplary embodiment, the reference sequences of the HBV genome include consensus sequences of HBV genotype B and genotype C, each covering all of the 3191 base pairs (bp) of the HBV genotype B or C genome.

[0042] In the embodiment, the viral sequence targeting probes are so designed that when sequences of the full viral sequence targeting probes are aligned, each of the full viral sequence targeting probes overlap with the immediately adjacent full viral sequence targeting probes by a portion of the length of the full viral sequence targeting probe. In the exemplary embodiment as illustrated in FIG. 2A, each of the full HBV targeting probes overlap with one or two immediately adjacent full HBV targeting probes by a

portion of the length of the full HBV targeting probe. The portion of sequence overlapping may vary and is preferably 50% (denoted 2X tiling density) or 75% (denoted 4X tiling density), as illustrated in FIG. 2B. In an exemplary embodiment in which the probes are designed to extend 120 bp in length with a 2X tiling density, each of the probes would overlap with the immediate adjacent probe for 60 bp. Likewise, if the probes are 120 bp in length with a 4X tiling density, each of the probes would overlap with the immediate adjacent probe for 90 bp.

[0043] Furthermore, structure of the viral genome may also be taken into consideration when designing the probes. In the exemplary embodiment, considering the HBV genome is circular in nature, the last probe of the full HBV targeting probes that extends beyond the terminal 3191 position of the reference sequence of the HBV genome is designed to continue at the start (i.e., position 1) of the reference sequence. For example, a probe having a length of 120 bp and starting at position 3121 of the reference sequence of the HBV genome would consist of a 71-bp region corresponding to positions 3121-3191, followed by a 49-bp region corresponding to positions 1-49.

[0044] It is to be understood that the embodiments of the present invention do not limit the lengths of the probes; the lengths of the probes may be designed according to cost, capture efficiency, sensitivity, specificity, or other specific concerns. In some embodiments, the possible number or amount N of the probes for any given reference sequence may be calculated according to Equation (1).

$$N = \sum_{P=\min}^{\max} L - P + 1 \quad \dots(1)$$

[0045] In Equation 1, L represents the length of the reference sequence, and P represents the length of the probes, which may range from a minimum length (denoted min) to a maximum length (denoted max). For example, a total of 220,597 probes, ranging from 50 bp to 120 bp, can be designed for the 3191-bp-long reference sequence of the HBV genotype B or C genome.

[0046] According to an embodiment of the present invention, the probe combination includes one or more sets of partial viral sequence targeting probes. When sequences of the partial viral sequence targeting probes are aligned, an overall sequence of the aligned set of partial viral sequence targeting probes matches a reference sequence of a

characteristic region on the genome of the target virus. In the aligned set of partial viral sequence targeting probes, each of the partial viral sequence targeting probes overlap with the immediately adjacent partial viral sequence targeting probes by a portion of the length of the partial viral sequence targeting probe. In some embodiments, the characteristic region may include a region between direct repeat 1 (DR1) and direct repeat 2 (DR2) on the HBV genome. In other embodiments, the characteristic region may be the region between DR1 and DR2 plus two elongated regions extending from two ends of the region to reach a predetermined length. For example, in defining a 960-bp-long reference sequence for a direct repeat (DR) region, assuming that DR1 and DR2 are located at positions 360-370 and 594-604 on a viral genome, the reference sequence of the DR region may be defined as the region between DR1 and DR2 with further elongation of 360 bp from two ends of the region. Consequently, reference sequence for a DR region on the HBV genotype A genome may be SEQ ID NOs. 3-5; reference sequence for a DR region on the HBV genotype B genome may be SEQ ID NOs. 6-9; reference sequence for a DR region on the HBV genotype C genome may be SEQ ID NOs. 10-13; reference sequence for a DR region on the HBV genotype D genome may be SEQ ID NOs. 14-16; reference sequence for a DR region on the HBV genotype E genome may be SEQ ID NOs. 17-19; reference sequence for a DR region on the HBV genotype F genome may be SEQ ID Nos. 20-22; reference sequence for a DR region on the HBV genotype G genome may be SEQ ID NOs. 23-25; reference sequence for a DR region on the HBV genotype H genome may be SEQ ID NOs. 26-28; reference sequence for a DR region on the HBV genotype genome I may be SEQ ID NOs. 29-31; and reference sequence for a DR region on the HBV genotype J genome may be SEQ ID NO. 32.

[0047] In an exemplary embodiment, the probe combination may include two sets of partial HBV targeting probes. When sequences of one set of the partial HBV targeting probes are aligned, an overall sequence of the aligned partial HBV targeting probes matches a reference sequence of the direct repeat (DR) region of the genome of HBV genotype B (SEQ ID NO. 9) or the DR region of the HBV genotype C genome (SEQ ID NO. 13). The DR region may be defined as positions 1190-2234, positions 1231-2190 or other characteristic range on the HBV genome. Similar to the aforementioned, each of the

partial HBV targeting probes overlap with one or two immediately adjacent partial HBV targeting probes by a portion of the length of the partial HBV targeting probe. The portion of sequence overlapping may be, but is not limited to, 50% (i.e., 2X tiling density) or 75% (i.e., 4X tiling density).

[0048] The possible number of the partial HBV targeting probes for the reference sequence of the DR region (SEQ ID NOs. 9, 13) may be calculated according to the aforementioned Equation (1). For example, a total of 62,196 probes, ranging from 50 bp to 120 bp, can be designed for the 960-bp-long reference sequence of the DR region of the HBV genome.

[0049] According to an embodiment of the present invention, the probe combination includes a set of the full viral sequence probes and a set of the partial viral sequence probes. The full and partial viral sequence probes are combined to enhance sequence coverage over the reference sequences of the viral genome. In the exemplary embodiment, the partial HBV targeting probes are designed to cover between the full HBV targeting probes at the DR region. For example, assuming that the full HBV targeting probes are 120 bp in length and start at positions 1, 61, and 121 (2X tiling density), the partial HBV targeting probes having 2X tiling would start at 31, 91, and 151. In other words, the DR region would be covered by two sets of probes (i.e., the full HBV targeting probes and the partial HBV targeting probes) with a 4X tiling density (i.e., each chain overlaps with 75% of its immediate adjacent chain).

[0050] According to an embodiment of the present invention, the probe combination further includes one or more sets of hotspot gene targeting probes. When sequences of each of the set of hotspot gene targeting probes are aligned, an overall sequence of the aligned set of hotspot gene targeting probes matches a reference sequence of a cancer hotspot gene. In the aligned set of hotspot gene targeting probes, each of the hotspot gene targeting probes overlap with the immediately adjacent hotspot gene targeting probes by a portion of the length of the hotspot gene targeting probe. The portion of sequence overlapping may be, but is not limited to, 50% (i.e., 2X tiling density) or 75% (i.e., 4X tiling density).

[0051] The reference sequence of the cancer hotspot gene is retrievable from the NCBI gene database. The cancer hotspot gene may include, but is not limited to, at least one of

the following genes, as identified by Entrez Gene IDs according to the NCBI gene database (www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=gene): ABL1 (Entrez Gene ID: 25), ABL2 (Entrez Gene ID: 27), ACSL3 (Entrez Gene ID: 2181), AF15Q14 (Entrez Gene ID: 57082), AF1Q (Entrez Gene ID: 10962), AF3p21 (Entrez Gene ID: 51517), AF5q31 (Entrez Gene ID: 27125), AKAP9 (Entrez Gene ID: 10142), AKT1 (Entrez Gene ID: 207), AKT2 (Entrez Gene ID: 208), ALDH2 (Entrez Gene ID: 217), ALK (Entrez Gene ID: 238), ALO17 (Entrez Gene ID: 57674), APC (Entrez Gene ID: 11789), ARHGEF12 (Entrez Gene ID: 23365), ARHH (Entrez Gene ID: 399), ARID1A (Entrez Gene ID: 8289), ARID2 (Entrez Gene ID: 196528), ARNT (Entrez Gene ID: 405), ASPSCR1 (Entrez Gene ID: 79058), ASXL1 (Entrez Gene ID: 171023), ATF1 (Entrez Gene ID: 466), ATIC (Entrez Gene ID: 471), ATM (Entrez Gene ID: 472), ATRX (Entrez Gene ID: 546), BAP1 (Entrez Gene ID: 8314), BCL10 (Entrez Gene ID: 8915), BCL11A (Entrez Gene ID: 53335), BCL11B (Entrez Gene ID: 64919), BCL2 (Entrez Gene ID: 596), BCL3 (Entrez Gene ID: 602), BCL5 (Entrez Gene ID: 603), BCL6 (Entrez Gene ID: 604), BCL7A (Entrez Gene ID: 605), BCL9 (Entrez Gene ID: 607), BCOR (Entrez Gene ID: 54880), BCR (Entrez Gene ID: 613), BHD (Entrez Gene ID: 50947), BIRC3 (Entrez Gene ID: 330), BLM (Entrez Gene ID: 641), BMPRIA (Entrez Gene ID: 12166), BRAF (Entrez Gene ID: 673), BRCA1 (Entrez Gene ID: 672), BRCA2 (Entrez Gene ID: 675), BRD3 (Entrez Gene ID: 8019), BRD4 (Entrez Gene ID: 23476), BRIP1 (Entrez Gene ID: 83990), BTG1 (Entrez Gene ID: 694), BUB1B (Entrez Gene ID: 701), C15orf55 (Entrez Gene ID: 144535), C16orf75 (Entrez Gene ID: 387882), CANT1 (Entrez Gene ID: 124583), CARD11 (Entrez Gene ID: 84433), CARs (Entrez Gene ID: 833), CBFB (Entrez Gene ID: 865), CBL (Entrez Gene ID: 867), CBLB (Entrez Gene ID: 868), CBLC (Entrez Gene ID: 23624), CCNB1IP1 (Entrez Gene ID: 57820), CCND1 (Entrez Gene ID: 595), CCND2 (Entrez Gene ID: 894), CCND3 (Entrez Gene ID: 896), CCNE1 (Entrez Gene ID: 898), CD273 (Entrez Gene ID: 80380), CD274 (Entrez Gene ID: 29126), CD74 (Entrez Gene ID: 972), CD79A (Entrez Gene ID: 973), CD79B (Entrez Gene ID: 974), CDH1 (Entrez Gene ID: 999), CDH11 (Entrez Gene ID: 1009), CDK12 (Entrez Gene ID: 51755), CDK4 (Entrez Gene ID: 1019), CDK6 (Entrez Gene ID: 1021), CDKN2A (Entrez Gene ID: 1029), CDKN2C (Entrez Gene ID: 1031), CDX2 (Entrez Gene ID: 1045), CEBPA (Entrez Gene ID: 1050), CEP1 (Entrez Gene ID:

11064), CHCHD7 (Entrez Gene ID: 79145), CHEK2 (Entrez Gene ID: 11200), CHIC2 (Entrez Gene ID: 26511), CHN1 (Entrez Gene ID: 1123), CIC (Entrez Gene ID: 23152), CIITA (Entrez Gene ID: 4261), CLTC (Entrez Gene ID: 1213), CLTCL1 (Entrez Gene ID: 8218), CMKOR1 (Entrez Gene ID: 57007), CoL1A1 (Entrez Gene ID: 1277), CPBP (Entrez Gene ID: 1316), COX6C (Entrez Gene ID: 1345), CREB1 (Entrez Gene ID: 1385), CREB3L1 (Entrez Gene ID: 90993), CREB3L2 (Entrez Gene ID: 64764), CREBBP (Entrez Gene ID: 1387), CRLF2 (Entrez Gene ID: 64109), CRT3 (Entrez Gene ID: 64784), CTNNA1 (catenin beta 1; Entrez Gene ID: 1499), CYLD (Entrez Gene ID: 1540), D10S170 (Entrez Gene ID: 8030), DAXX (Entrez Gene ID: 1616), DDB2 (Entrez Gene ID: 1643), DDX10 (Entrez Gene ID: 1662), DDX5 (Entrez Gene ID: 1655), DDX6 (Entrez Gene ID: 1656), DEK (Entrez Gene ID: 7913), DICER1 (Entrez Gene ID: 23405), DNMT3A (Entrez Gene ID: 1788), DUX4 (Entrez Gene ID: 100288687), EBF1 (Entrez Gene ID: 1879), EGFR (Entrez Gene ID: 1956), EIF4A2 (Entrez Gene ID: 1974), ELF4 (Entrez Gene ID: 2000), ELK4 (Entrez Gene ID: 2005), ELKS (Entrez Gene ID: 23085), ELL (Entrez Gene ID: 8178), ELN (Entrez Gene ID: 2006), EML4 (Entrez Gene ID: 27436), EP300 (Entrez Gene ID: 2033), EPS15 (Entrez Gene ID: 2060), ERBB2 (Entrez Gene ID: 2064), ERCC2 (Entrez Gene ID: 2068), ERCC3 (Entrez Gene ID: 2071), ERCC4 (Entrez Gene ID: 2072), ERCC5 (Entrez Gene ID: 2073), ERG (Entrez Gene ID: 2078), ETV1 (Entrez Gene ID: 2115), ETV4 (Entrez Gene ID: 2118), ETV5 (Entrez Gene ID: 2119), ETV6 (Entrez Gene ID: 2120), EVI1 (Entrez Gene ID: 2122), EWsR1 (Entrez Gene ID: 2130), EXT1 (Entrez Gene ID: 2131), EXT2 (Entrez Gene ID: 2132), EZH2 (Entrez Gene ID: 2146), FACL6 (Entrez Gene ID: 23305), FAM22A (Entrez Gene ID: 728118), FAM22B (Entrez Gene ID: 729262), FAM46C (Entrez Gene ID: 54855), FANCA (Entrez Gene ID: 2175), FANCC (Entrez Gene ID: 2176), FANCD2 (Entrez Gene ID: 2177), FANCE (Entrez Gene ID: 2178), FANCF (Entrez Gene ID: 2188), FANCG (Entrez Gene ID: 2189), FBXO11 (Entrez Gene ID: 80204), FBXW7 (Entrez Gene ID: 55294), FCGR2B (Entrez Gene ID: 2213), FEV (Entrez Gene ID: 54738), FGFR1 (Entrez Gene ID: 2260), FGFR1OP (Entrez Gene ID: 11116), FGFR2 (Entrez Gene ID: 2263), FGFR3 (Entrez Gene ID: 2261), FH (Entrez Gene ID: 2271), FHIT (Entrez Gene ID: 2272), FIP1L1 (Entrez Gene ID: 81608), FLI1 (Entrez Gene ID: 2313), FLT3 (Entrez Gene ID: 2322), FNBP1 (Entrez Gene ID: 23048), FOXL2 (Entrez

Gene ID: 668), FOXO1 (Entrez Gene ID: 2308), FOXO3A (Entrez Gene ID: 2309), FOXP1 (Entrez Gene ID: 27086), FSTL3 (Entrez Gene ID: 10272), FUBP1 (Entrez Gene ID: 8880), FUS (Entrez Gene ID: 2521), FVT1 (Entrez Gene ID: 2531), GAS7 (Entrez Gene ID: 8522), GATA1 (Entrez Gene ID: 2623), GATA2 (Entrez Gene ID: 2624), GATA3 (Entrez Gene ID: 2625), GMPS (Entrez Gene ID: 8833), GNA11 (Entrez Gene ID: 2767), GNAQ (Entrez Gene ID: 2776), GNAS (Entrez Gene ID: 2778), GOLGA5 (Entrez Gene ID: 9950), GOPC (Entrez Gene ID: 57120), GPC3 (Entrez Gene ID: 2719), GPHN (Entrez Gene ID: 10243), GRAF (Entrez Gene ID: 23092), HCMOGT-1 (Entrez Gene ID: 92521), HEAB (Entrez Gene ID: 10978), HERPUD1 (Entrez Gene ID: 9709), HEY1 (Entrez Gene ID: 23462), HIP1 (Entrez Gene ID: 3092), HIST1H4I (Entrez Gene ID: 8294), HLF (Entrez Gene ID: 3131), HLXB9 (Entrez Gene ID: 3110), HMGA1 (Entrez Gene ID: 3159), HMGA2 (Entrez Gene ID: 8091), HNRNPA2B1 (Entrez Gene ID: 3181), HOOK3 (Entrez Gene ID: 84376), HOXA11 (Entrez Gene ID: 3207), HOXA13 (Entrez Gene ID: 3209), HOXA9 (Entrez Gene ID: 3205), HOXC11 (Entrez Gene ID: 3227), HOXC13 (Entrez Gene ID: 3229), HOXD11 (Entrez Gene ID: 3237), HOXD13 (Entrez Gene ID: 3239), HRAS (Entrez Gene ID: 3265), HRPT2 (Entrez Gene ID: 79577), HSPCA (Entrez Gene ID: 3320), HSPCB (Entrez Gene ID: 3326), IDH1 (Entrez Gene ID: 3417), IDH2 (Entrez Gene ID: 3418), IGH@ (Entrez Gene ID: 3492), IGK@ (Entrez Gene ID: 50802), IGL@ (Entrez Gene ID: 3535), IKZF1 (Entrez Gene ID: 10320), IL2 (Entrez Gene ID: 3558), IL21R (Entrez Gene ID: 50615), IL6ST (Entrez Gene ID: 3572), IL7R (Entrez Gene ID: 3575), IRF4 (Entrez Gene ID: 3662), IRTA1 (Entrez Gene ID: 83417), ITK (Entrez Gene ID: 3702), JAK1 (Entrez Gene ID: 3716), JAK2 (Entrez Gene ID: 3717), JAK3 (Entrez Gene ID: 3718), JAZF1 (Entrez Gene ID: 221895), JUN (Entrez Gene ID: 3725), KDR (Entrez Gene ID: 3791), KIAA1549 (Entrez Gene ID: 57670), KIT (Entrez Gene ID: 3815), KLK2 (Entrez Gene ID: 3817), KRAS (Entrez Gene ID: 3845), KTN1 (Entrez Gene ID: 3895), LAF4 (Entrez Gene ID: 3899), LASP1 (Entrez Gene ID: 3927), LCK (Entrez Gene ID: 3932), LCP1 (Entrez Gene ID: 3936), LCX (Entrez Gene ID: 80312), LHFP (Entrez Gene ID: 10186), LIFR (Entrez Gene ID: 3977), LMO1 (Entrez Gene ID: 4004), LMO2 (Entrez Gene ID: 4005), LPP (Entrez Gene ID: 4026), LYL1 (Entrez Gene ID: 4066), MADH4 (Entrez Gene ID: 4089), MAF (Entrez Gene ID: 4094), MAFB (Entrez Gene ID: 9935), MALT1 (Entrez

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6917), TCF1 (Entrez Gene ID: 6927), TCF12 (Entrez Gene ID: 6938), TCF3 (Entrez Gene ID: 6929), TCF7L2 (Entrez Gene ID: 6934), TCL1A (Entrez Gene ID: 8115), TCL6 (Entrez Gene ID: 27004), TET2 (Entrez Gene ID: 54790), TERT (telomerase reverse transcriptase; Entrez Gene ID: 7015), TFE3 (Entrez Gene ID: 7030), TFEB (Entrez Gene ID: 7942), TFG (Entrez Gene ID: 10342), TFPT (Entrez Gene ID: 29844), TFRC (Entrez Gene ID: 7037), THRAP3 (Entrez Gene ID: 9967), TIF1 (Entrez Gene ID: 8805), TLX1 (Entrez Gene ID: 3195), TLX3 (Entrez Gene ID: 30012), TMPRSS2 (Entrez Gene ID: 7113), TNFAIP3 (Entrez Gene ID: 7128), TNFRSF14 (Entrez Gene ID: 8764), TNFRSF17 (Entrez Gene ID: 608), TNFRSF6 (Entrez Gene ID: 355), TOP1 (Entrez Gene ID: 7150), TP53 (tumor protein p53; Entrez Gene ID: 7157), TPM3 (Entrez Gene ID: 7170), TPM4 (Entrez Gene ID: 7171), TPR (Entrez Gene ID: 7175), TRA@ (Entrez Gene ID: 6955), TRB@ (Entrez Gene ID: 6957), TRD@ (Entrez Gene ID: 6964), TRIM27 (Entrez Gene ID: 5987), TRIM33 (Entrez Gene ID: 51592), TRIP11 (Entrez Gene ID: 9321), TSC1 (Entrez Gene ID: 7248), TSC2 (Entrez Gene ID: 7249), TSHR (Entrez Gene ID: 7253), TTL (Entrez Gene ID: 150465), U2AF1 (Entrez Gene ID: 7307), USP6 (Entrez Gene ID: 9098), VHL (Entrez Gene ID: 7428), WAS (Entrez Gene ID: 7454), WHSC1 (Entrez Gene ID: 7468), WHSC1L1 (Entrez Gene ID: 54904), WIF1 (Entrez Gene ID: 11197), WRN (Entrez Gene ID: 7486), WT1 (Entrez Gene ID: 7490), WTX (Entrez Gene ID: 139285), XPA (Entrez Gene ID: 7507), XPC (Entrez Gene ID: 7508), XPO1 (Entrez Gene ID: 7514), YWHAE (Entrez Gene ID: 7531), ZNF145 (Entrez Gene ID: 7704), ZNF198 (Entrez Gene ID: 7750), ZNF278 (Entrez Gene ID: 23598), ZNF331 (Entrez Gene ID: 55422), ZNF384 (Entrez Gene ID: 171017), ZNF521 (Entrez Gene ID: 25925), ZNF9 (Entrez Gene ID: 7555), and ZRSR2 (Entrez Gene ID: 8233).

[0052] In the embodiment, considering that the terminal regions (e.g., first 60 bp and last 60 bp) of the reference sequences of the selected cancer hotspot genes would only be covered by a single probe and might result in lower capture efficiency as compared with the non-terminal regions which would be covered by 2 probes in case of a 2X tiling, the reference sequences of the cancer hotspot genes may be elongated beyond both ends of the sequences. For example, exon 3 of the CTNNB1 gene is 228 bp in length; elongation of 75 bp at two ends of the sequence results in a 378-bp-long reference sequence of

CTNNB1 exon 3 (SEQ ID NO. 33). Other reference sequences of the cancer hotspot genes may also be designed in a similar fashion. In addition, if the elongated region(s) of an exon overlaps with an adjacent exon or the elongated regions thereof, the two elongated reference sequences may be integrated into a single reference sequence covering both exons and all elongated regions.

[0053] The possible number of the hotspot gene targeting probes for the cancer hotspot genes may be calculated according to the aforementioned Equation (1). For example, a total of 20,874 probes that range from 50 bp to 120 bp can be designed for the 378-bp-long reference sequence of CTNNB1 exon 3 (SEQ ID NO. 33). Likewise, a total of 41,819 probes ranging 50-120 bp may be designed for a 673-bp-long reference sequence of a TERT promoter (SEQ ID NO. 34). A total of 49,345 probes ranging 50-120 bp may be designed for a 779-bp-long reference sequence of TP53 exons 2/3/4 (SEQ ID NO. 35). A total of 31,524 probes ranging 50-120 bp may be designed for a 528-bp-long reference sequence of TP53 exons 5/6 (SEQ ID NO. 36). A total of 12,496 probes ranging 50-120 bp may be designed for a 260-bp-long reference sequence of TP53 exon 7 (SEQ ID NO. 37). A total of 26,199 probes ranging 50-120 bp may be designed for a 453-bp-long reference sequence of TP53 exons 8/9 (SEQ ID NO. 39). A total of 12,283 probes ranging 50-120 bp may be designed for a 257-bp-long reference sequence of TP53 exon 10 (SEQ ID NO. 40). A total of 10,508 probes ranging 50-120 bp may be designed for a 232-bp-long reference sequence of TP53 exon 11 (SEQ ID NO. 41).

[0054] According to an embodiment of the present invention, the probe combination further includes one or more sets of exogenous gene targeting probes for negative control and quantitation. When sequences of one of the sets of exogenous gene targeting probes are aligned, an overall sequence of the aligned set exogenous gene targeting probes matches a reference sequence of an exogenous gene. In the aligned set of exogenous gene targeting probes, each of the exogenous gene targeting probes overlap with the immediately adjacent exogenous gene targeting probes by a portion of the length of the exogenous gene targeting probe. The portion of sequence overlapping may be, but is not limited to, 50% (i.e., 2X tiling density) or 75% (i.e., 4X tiling density).

[0055] The reference sequence of the exogenous gene is retrievable from the NCBI gene database. The exogenous gene may originate from lambda phage, *E.coli*, yeast, ϕ X174,

or other common microorganism. The possible number of the exogenous gene targeting probes for the exogenous genes may be calculated according to the aforementioned Equation (1). For example, a total of 478,682 probes that range from 50 bp to 120 bp can be designed for the 48502-bp-long reference sequence of lambda phage genome (GenBank Accession No. NC_001416).

[0056] In the embodiment, an external source of nucleotide fragments (e.g., spike-in DNA) corresponding to the sequences of the exogenous gene targeting probes is required. In other words, since the human genome, regardless of its hepatitis B or HCC status, does not contain genomic regions similar to the sequences of the exogenous gene targeting probes, the exogenous gene targeting probes theoretically would not capture any nucleotide fragments from genomic (gDNA) or circulating tumor DNA (ctDNA) of human samples if no nucleotide fragments corresponding to the sequences of the exogenous gene targeting probes are added externally during the detection process. As all nucleotide fragments captured by the exogenous gene targeting probes are theoretically the externally added nucleotide fragments, quantity and quality of the externally added nucleotide fragments can be manipulated, thus providing a reliable mean for absolute quantitation.

[0057] In an exemplary embodiment, four 120-bp regions on the lambda phage genome (SEQ ID NOs 42-45) were chosen for designing the lambda targeting probes according to the following selection criteria: a) no homology with human or HBV genome; b) unique among the lambda phage genome; c) GC content within a predefined range; d) no long monomer sequence (e.g., AAAAA); and/or e) no significant secondary structure as predicted by primer3, netprimer, and other primer design algorithms. As exemplified in Table 1, the full HBV targeting probes, the partial HBV targeting probes, the hotspot gene targeting probes, and the exogenous gene targeting probes may be used in combination to capture target nucleotide fragments that contains HBV DNA with or without viral-host junctions.

Reference	Tiling	Copy	Amount	Length Covered	SEQ ID
HBV Genome	2X	2N	108	3191	1, 2
HBV DR Region	2X	2N	32	960	9, 13
CTNNB1 exon 3	2X	1N	6	378	33
TERT promoter	2X	1N	11	673	34
TP53 exons 2-11	2X	1N	39	2492	35-38, 40-41

Lambda	1X	1N	4	480	43-46
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Table 1

[0058] In another exemplary embodiment as depicted in Table 2, additional lambda targeting probes may be designed to cover elongated regions downstream of one of the four 120-bp regions (SEQ ID NOs 46-49) at a 2X or 4X tiling density. Additional sets (or copies) of the lambda targeting probes may also be used to simulate the two copies (2N) of the HBV targeting probes (one for genotype B and the other for genotype C) and one copy (1N) of the hotspot gene targeting probes, therefore resulting in a combination of 2X/1N, 2X/2N, 4X/1N, and 4X/2N lambda targeting probes corresponding to the elongated regions on the lambda genome.

Reference	Tiling	Copy	Amount	Length Covered	SEQ ID
HBV Genome	2X	2N	106	3191	1, 2
HBV DR Region	2X	2N	30	960	9, 13
CTNNB1 exon 3	2X	1N	6	378	33
TERT promoter	2X	1N	11	673	34
TP53 exons 2-11	2X	1N	38	2509	35-37, 39-41
Lambda	2X	1N	3	240	46
	2X	2N	6	240	47
	4X	1N	3	180	48
	4X	2N	6	180	49
	1X	1N	5	600	50-54
GAPDH	2X	1N	3	240	55
GdX	2X	1N	3	240	56

Table 2

[0059] Further, GC content has been reported to affect sequencing coverage, exhibiting approximately 3 fold difference among samples with low GC ratio (GC=0.3, coverage=0.6X), optimal GC ratio (GC=0.48, coverage=1.3X), and high GC ratio (0.7, coverage=0.4X). Therefore, as depicted in Table 2, additional sets of lambda targeting probes may also be designed to internally control the GC content of the probes. Five 120-bp regions on the lambda phage genome (SEQ ID NOs 50-54) were chosen according to the following selection criteria: a) no homology with human or HBV genome; b) unique among the lambda phage genome; c) GC content within a predefined range; d) no long monomer sequence (e.g., AAAAA); and e) no significant secondary structure as predicted by primer3, netprimer, and other primer design algorithms. Consequently, five 120-bp-long regions having GC contents of 0.3, 0.4, 0.5, 0.6, and 0.68 are selected for designing the five additional sets of lambda targeting probes (1X/1N).

[0060] According to an embodiment of the present invention, the probe combination further includes one or more sets of endogenous gene targeting probes for positive internal control and relative quantitation. When sequences of one of the sets of endogenous gene targeting probes are aligned, an overall sequence of the aligned set of endogenous gene targeting probes matches a reference sequence of an endogenous gene. In the aligned set of endogenous gene targeting probes, each of the endogenous gene targeting probes overlap with the immediately adjacent endogenous gene targeting probes by a portion of the length of the endogenous gene targeting probe. The portion of sequence overlapping may be, but is not limited to, 50% (i.e., 2X tiling density) or 75% (i.e., 4X tiling density).

[0061] The reference sequence of the endogenous gene is retrievable from the NCBI gene database. In the embodiment, the endogenous gene is intrinsic of the human genome and may include, but is not limited to, at least one of the following genes, as identified by Entrez Gene IDs according to the NCBI gene database (www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=gene): GAPDH (glyceraldehyde-3-phosphate dehydrogenase; Entrez Gene ID: 2597), UBL4A (ubiquitin like 4A; GdX; Entrez Gene ID: 8266), HPRT1 (Entrez Gene ID: 3251), TBP (Entrez Gene ID: 6908), B2M (Entrez Gene ID: 567), RPL13A (Entrez Gene ID: 23521), RN18S1 (Entrez Gene ID: 100008588), C1orf43 (Entrez Gene ID: 25912), CHMP2A (Entrez Gene ID: 27243), EMC7 (Entrez Gene ID: 56851), GPI (Entrez Gene ID: 2821), PSMB2 (Entrez Gene ID: 5690), PSMB4 (Entrez Gene ID: 5692), RAB7A (Entrez Gene ID: 7879), REEP5 (Entrez Gene ID: 7905), SNRPD3 (Entrez Gene ID: 6634), VCP (Entrez Gene ID: 7415), VPS29 (Entrez Gene ID: 51699), ACTB (Entrez Gene ID: 60), PPIA (Entrez Gene ID: 5478), GUSB (Entrez Gene ID: 2990), HSP90AB1 (Entrez Gene ID: 3326), RPLP0 (Entrez Gene ID: 6175), TFRC (Entrez Gene ID: 7037), UBC (Entrez Gene ID: 7316).

[0062] The possible number of the endogenous gene targeting probes for the endogenous genes may be calculated according to the aforementioned Equation (1). For example, a total of 113,458 probes that range from 50 bp to 120 bp can be designed for the 1682-bp-long reference sequence of GAPDH gene. Likewise, a total of 248,429 probes that range from 50 bp to 120 bp can be designed for the 3583-bp-long reference sequence of GdX gene.

[0063] In the embodiment, the endogenous genes are chosen to enhance reliability of sequence detection; that is, the endogenous genes are adopted for being common housekeeping genes that are stably expressed and have not been found to vary in tumors. It can be understood that quantification according to detection of only the cancer hotspot genes could be unreliable as structural variation of the cancer hotspot genes, such as CTNNB1, TP53, TERT, and other cancer related genes listed above, have been reported in tumor samples and that their copy numbers may change during tumorigenesis due to deletion, duplication, or other structural variations. Therefore, in an exemplary embodiment as depicted in Table 2, probes targeting a 240-bp region on the GAPDH gene (SEQ ID NO. 55) at a 2X tiling density and probes targeting a 240-region of the GdX gene (SEQ ID NO. 56) at a 2X tiling are included in the probe combination as internal control. The 240-bp regions on the GAPDH and GdX genes do not have homology to HBV genome, long monomers, significant secondary structure, or GC content out of a predefined range. Additional advantage of adopting the GdX gene is that GdX can also be used to identify gender of the test subject.

[0064] In some embodiments of the present invention, the probes are each labeled with a marker molecule to facilitate detection and quantitation. The marker molecule may include, but are not limited to, biotin, fluorescent protein, luminescent protein, antibody, radioactive compounds, or any combination thereof.

[0065] Another aspect of the present invention provides a method for detecting infection with DNA virus (e.g., HBV, HPV, EBV, HHV-8, HTLV, MCV, or other DNA viruses) or viral infection associated cancer (e.g., hepatocellular carcinoma, cervical cancer, nasopharyngeal cancer, lymphoma, Merkel cell carcinoma, or other cancers associated with infection with the DNA viruses). In an embodiment, the method includes the steps of: extracting nucleic acids from a specimen of a subject; amplifying the nucleic acids; hybridizing the nucleic acids with the probe combination according to the various embodiments mentioned above to capture target nucleotide fragments; sequencing the target nucleotide fragments; and analyzing the target nucleotide fragments.

[0066] In the embodiment, the nucleic acids may include viral nucleic acids, host genome nucleic acids, and nucleic acids with viral-host junction, and may be DNA, RNA, or polynucleotides. Extraction of the nucleic acids may be performed by precipitation,

chromatography and/or magnetic bead capturing. The specimen from which the nucleic acids are extracted may be biological fluid (e.g., blood, sweat, saliva, tears, urine, lymph, or interstitial fluid) or tissues (e.g., liver tissue). Amplification of the extracted nucleic acids may be performed by DNA cloning, polymerase chain reaction (PCR), reverse transcription PCR (RT-PCR), nested-PCR, quantitative (qPCR) and/or digital PCR. The target nucleotide fragments may be captured by the probe combination via hybridization (e.g., southern blot hybridization, *in situ* hybridization, or northern blot hybridization) and/or lockdown (e.g., bead-based method or chip-based method). The captured target nucleic acids may be sequenced by NGS (e.g., massively parallel sequencing, single molecule sequencing, or NanoString). Maxam-Gilbert sequencing, Sanger sequencing, pyrosequencing, and/or DNA microarray.

[0067] In an alternative embodiment, the amplification step and the hybridization step may be reversed. In other words, the method according to another embodiment of the present invention includes the steps of: extracting nucleic acids from a specimen of a subject; hybridizing the nucleic acids with the probe combination according to the various embodiments of the present invention to capture target nucleotide fragments; amplifying the captured target nucleotide fragments; sequencing the target nucleotide fragments; and analyzing the target nucleotide fragments.

[0068] Analysis and quantitation of the target nucleic acids captured by the probe combination may be performed as follows. Raw reads (RR) are generated directly from the NGS sequencing instrument. Low quality reads among the raw reads are excluded to obtain high quality reads (HQR). The HQRs are compressed into unique reads (UR); in other words, HQRs having completely identical sequences are collapsed into a single unique read, while the information regarding the copy numbers (redundancy) thereof is retained. Finally, URs with low redundancies are excluded to result in high redundancy unique reads (HRUR). Further, the total number of reads included in the high redundancy unique reads (RiHRUR) can be calculated by the retained redundancy information during the compression process.

[0069] In an embodiment, the bioinformatics analytic methodology adopted in analyzing the NGS data set is summarized in Table 3.

Analysis	Preferred Embodiment	Prior Art
Junction Detection	Direct identification of HBV-	Direct identification of HBV-human

	human junction	junction
Mapping Method	BLAST	SOAP2 (for initial mapping) BWA (for junction detection)
Pair-end Reads	Viewed as individual reads	Merge whenever possible (overlap >5 bp and mismatch rate <0.2)
Low Quality Read	A read whose bases with quality value ≤ 5 occupying 50% of the read length	A read whose bases with quality value ≤ 5 occupying 50% of the read length
Redundancy	Reads with redundancy <5 excluded	Duplicated reads removed
Mapping Criteria	BLAST e-value < 10^{-10} for both HBV and human (~20-40 bp)	30 bp for both HBV and human
Quantification	Total number of reads	Total number of unique reads; Normalized total number of paired- reads; Merge neighboring junctions (20 bp)

Table 3

[0070] Table 3 also compares the methodology of the embodiment with that reported by Zhao. As shown in Table 3, some major differences between the two include: Zhao merges junctions of close vicinity, whereas the methodology of the present embodiment merges junctions based on sequence similarity. Also, Zhao removes duplicated reads, considering only unique junctions, whereas the methodology of the present embodiment excludes reads having redundancies of less than 5, retains redundancy information, and quantifies junctions based on the total number of reads for single unique junction.

[0071] Validation of Probe Specificity

[0072] Probes targeting TP53 exons 2-11 (SEQ ID NOs 35-38, 40-41) designed according to an embodiment of the present invention were hybridized with HCC tumor genomic DNA (gDNA), non-tumor gDNA, and ctDNA of an HCC patient and quantified by qPCR. MicroRNA miR-122 that is conserved among vertebrates and highly expressed in the liver was also quantified as a negative control.

Sample Type	Post-Hybridization Retention Ratio		Fold Difference
	TP53	miR-122	
HCC Tumor gDNA	3.40%	0.01%	274.37
Non-Tumor gDNA	4.61%	0.02%	268.72
ctDNA	2.65%	0.26%	10.26

Table 4

[0073] As demonstrated in Table 4, the significantly higher post-hybridization retentions of TP53 over miR-122 in all of the three sample types indicated that TP53 fragments were successfully hybridized, captured, and recovered by the TP53 targeting probes; in

contrast, miR-122 fragments were washed off during the procedure as the TP53 targeting probes have no specificity to miR-122. Table 4 also shows that the amounts of TP53 fragments captured by the TP53 targeting probes from the genomic DNA were over 250 folds higher than that of miR-122 fragments, and that TP53 fragments captured from ctDNA were over 10 times more concentrated than that miR-122 fragments. The results demonstrated that the TP53 targeting probes is sequence specific and can selectively capture TP53 gene fragments from DNA samples.

[0074] Meanwhile, as shown in Table 5, a total of 26 HCC tumor gDNA samples are enriched by the probe combination listed above in Table 1 and sequenced by next generation sequencing (NGS) for analysis of presence of HBV genome, HBV-human junction (denoted "Junction") and cancer hotspot genes (including CTNNB1, TERT, and TP53). The HBV-human junction is indicative of HBV integration into human genome. The host genome ratio in Table 5 is the calculated length ratios of the captured sequences over the human genome. As demonstrated in Table 5, the significant differences between the calculated host genome ratios and the observed NGS read ratios indicated successful enrichment of the HBV genome, cancer hotspot genes and HBV-human junction by the probe combination.

	NGS Dataset Read Ratio	Host Genome Ratio
HBV	35.01%	<0.0001% (32.15 kb)
Junction	0.72%	<0.0001% (3.00 kb)
CTNNB1/TERT/TP53	33.57%	<0.0001% (3.56 kb)

Table 5

[0075] It is to be understood that the estimated 3kb junction length in the host genome was calculated by estimating that a single junction would have a detection range of 150 bp. Therefore, a single integration event, which results in two junctions, would be represented by 300 bp of junction regions. By using a rough estimate of 10 detectable junctions per patient, the estimated junction length of an individual patient was hence set at 3 kb (i.e., 300 bp $\times$ 10). The length of integrated HBV (excluding free-form non-integrated HBV) was then estimated at 32.15 kb (i.e., 3.215 kb $\times$ 10). The estimation of the junction and HBV ratio in the human genome presented herein is very crude and most likely an over-estimate, which would result in an under-estimation of the enrichment efficiency of junctions and HBV.

[0076] Referring now to FIG. 3. Junction targeting probes designed according to the embodiments of the present invention are demonstrated to selectively capture DNA fragments having specific viral-host junctions. HCC tumor gDNA samples were collected from tumor (denoted T) and non-tumor (denoted N) tissues of three HCC patients (denoted pt3, pt11 and pt15), and were hybridized with probe 1 (denoted pt3 junction) and probe 2 (denoted pt11 junction) that were designed according to sequences of HBV-human junctions in pt3 and pt11, respectively. Western blot analysis shown in FIG. 3 indicated that probe 1 selectively hybridized with tumor gDNA of patient pt3 and probe 2 selectively hybridized with tumor gDNA of patient pt11. NTC refers to “no template control” and was used as a negative control for the experiment; whereas PBGD (i.e., porphobilinogen deaminase gene) and miR-122 were used as positive controls.

[0077] Referring now to FIGs. 4A and 4B. Probe 1 (denoted pt3 junction) was then used to detect presence of HBV-human junction in genomic DNA and serum DNA of patient pt3. As shown in FIG. 4A, a pt3 specific HBV-human junction was observed in tumor gDNA and pre-operation (pre-OP) and post-operation (post-OP) serum DNA of patient pt3. Such junction was not observed in patient pt11 or in non-HCC HBV positive patient (denoted “Normal”). Similarly, as shown in FIG. 4B, when probe 2 (denoted pt11 junction) was used to detect presence of HBV-human junction in genomic DNA and serum DNA of patient pt11, a pt11 specific HBV-human junction was observed in tumor gDNA and pre-operation (pre-OP) serum DNA of patient pt11. Such junction was not observed in patient pt15 or in non-HCC HBV positive patient (denoted “Normal”).

[0078] Validation of Capture Efficiency

[0079] The probe combination listed above in Table 2 was used to analyze DNA fragments in a pair of tumor gDNA and plasma ctDNA samples (i.e., DNA samples from a single HCC patient) for determining the capture efficiency of the probe combination in different sample types. As shown in the NGS statistics in Table 6, tumor gDNA was 10-18 times higher in full HBV, partial HBV, and HBV-human junction reads than plasma ctDNA, demonstrating a higher capture efficiency of the probe combination in tumor gDNA samples. In addition, 8 of the 10 junction types identified in the tumor gDNA sample were with significant read numbers (>947), indicating a junction recovery rate of 75%.

	Sample Type	
	Tumor gDNA	Plasma ctDNA
Raw Reads (RR)	3,933,540	2,674,212
High Quality Reads (HQR)	3,919,958	2,658,014
Unique Reads (UR)	1,271,268	2,400,413
High Redundancy Unique Reads (HRUR)	23,245	3,809
Reads included in HRUR (RiHRUR)	2,341,219	140,452
Partial HBV	72,346	7,514
Full HBV	355,838	20,130
Unique Junction Reads	512	32
Total Junction Reads	16,242	1,184
Junction Type	10	28
CTNNB1 exon 3	46,177	6,779
TERT	19,851	17,355
TP53 exon 2/3/4	43,050	17,272
TP53 exon 5/6	42,423	11,495
TP53 exon 7	14,701	3,903
TP53 exon 8/9	26,181	5,447
TP53 exon 10	13,403	6,111
TP53 exon 11	8,772	1,980
TP53 total	148,530	46,208

Table 6

[0080] FIG. 5A and FIG. 5B are heat map converted bar charts showing the predicted ratio of sequenced NGS reads captured by each of the probes. Meanwhile, FIG. 6 shows the NGS results of tumor gDNA sample of another HCC patient using the same probe combination (as in Table 2).

[0081] Referring now to FIG. 7. In addition to detection of HBV DNA and HBV integration into human genome, the probe combination according to the embodiments of the present invention can also detect cancer associated gene mutations. As shown in FIG. 8, NGS analysis of DNA samples from male and female HCC patients with genotype B HBV infection revealed different numbers of unique HBV-human junctions detected (denoted “Junction Type”), unique junctions detected in the TERT or MLL4 region (denoted “TERT/MLL4 integration”), and known cancer hotspot mutations in the TERT promoter or CTNNB1 exon 3 region (denoted “TERT/CTNNB1 mutations”) among the patients.

[0082] Benefits and Advantages

[0083] The probe combination and analytic methodology according to the embodiments of the present invention exhibits significantly superior sensitivity and efficiency over the

prior art. As compared with the results reported by Li as shown in Table 7, the target nucleotide fragments captured by the probe combination of a preferred embodiment of the present invention (as in Table 1) has significantly higher HBV ratio, higher junction reads, and lower human ratio.

	Preferred Embodiment	Prior Art
Probes	70% HBV targeting 28% human targeting	100% HBV targeting
HBV-only Reads	37.68%	0.08%
Partial HBV Reads	5.61%	n/a
Junction Reads	0.78%	0.01%
Human Reads	29.75%	83.70%

Table 7

[0084] In addition, analysis of the NGS data set reported by Zhao using the bioinformatics analytic methodology for identifying viral-host junctions according to the embodiments of the present invention also showed that 97.5% of the reads of Zhao were human, with only 1.49% HBV, 1.43% partial HBV and 1% junction, reconfirming the poor efficiency of existing HBV capture probes in enrichment of HBV fragments and HBV-human junctions. Furthermore, while Zhao only reported to identify 157 junctions, analysis of Zhao's NGS data set by the analytic methodology of the embodiments of the present invention reveal 469 junctions and recover nearly 80% of Zhao's junctions. These results demonstrate that the analytic methodology of the embodiments of the present invention are highly sensitive in detection of viral integration and can identify significantly more viral-host junctions than the existing art.

[0085] Further, as compared with the direct sequencing approaches of Jiang and Sung, the probe combination and analytic methodology according to the embodiments of the present invention generate only about 5 million 150-bp reads in a typical NGS data set (i.e., 80% less in read number or 60% less in total nucleotides than Jiang); yet, as many as 307,101 HBV reads and 69,198 junction reads can be identified from the 5 million reads. The results also demonstrate that the embodiments of the present invention are not only sensitive but also highly efficient in identification of viral integration.

[0086] In sum, the present invention according to the aforementioned embodiments provides a powerful and versatile tool for detection of viral infection and viral infection induced cancer. The embodiments of the present invention can be applied to detect presence of various types of DNA viruses and viral integration. The probe combination

designed according to the embodiments ensures optimal viral/host sequence coverage and considers genetic stability, and is thus demonstrated to be highly sensitive, efficient, and reliable.

[0087] Previous descriptions are only embodiments of the present disclosure and are not intended to limit the scope of the present disclosure. Many variations and modifications according to the claims and specification of the disclosure are still within the scope of the claimed disclosure. In addition, each of the embodiments and claims does not have to achieve all the advantages or characteristics disclosed. Moreover, the abstract and the title only serve to facilitate searching patent documents and are not intended in any way to limit the scope of the claimed disclosure.

CLAIMS

WHAT IS CLAIMED IS:

1. A probe combination for detecting cancer, comprising:
 - one or more sets of partial hepatitis B virus (HBV) targeting probes,
 - wherein when sequences of each of the sets of partial HBV targeting probes are aligned, an overall sequence of the aligned set of partial HBV targeting probes matches a reference sequence of a direct repeat (DR) region on a genome of a HBV genotype, and
 - in the aligned set of partial HBV targeting probes, each of the partial HBV targeting probes overlap with one or two adjacent partial HBV targeting probes by a portion of a length of the partial HBV targeting probe.
2. The probe combination according to claim 1, wherein the HBV genotype comprises genotype A, genotype B, genotype C, genotype D, genotype E, genotype F, genotype G, genotype H, genotype I and genotype J.
3. The probe combination according to claim 1, wherein the reference sequence of the DR region comprises SEQ ID NOs. 3-32.
4. The probe combination according to claim 1, further comprising:
 - one or more sets of full HBV targeting probes,
 - wherein when sequences of each of the sets of full HBV targeting probes are aligned, an overall sequence of the aligned set of full HBV targeting probes matches a reference sequence of the genome of the HBV genotype, and
 - in the aligned set of full HBV targeting probes, each of the full HBV targeting probes overlap with one or two adjacent full HBV targeting probes by a portion of a length of the full HBV targeting probe.
5. The probe combination according to claim 4, further comprising:
 - one or more sets of hotspot gene targeting probes,
 - wherein when sequences of each of the sets of hotspot gene targeting probes are aligned, an overall sequence of the aligned set of hotspot gene targeting probes matches a reference sequence of a cancer hotspot gene, and
 - in the aligned set of hotspot gene targeting probes, each of the hotspot gene targeting probes overlap with one or two adjacent hotspot gene targeting probes by a

- portion of a length of the hotspot gene targeting probe.
6. The probe combination according to claim 5, wherein the cancer hotspot gene comprises CTNNB1 gene, TERT gene, and TP53 gene.
 7. The probe combination according to claim 5, wherein the reference sequence of the cancer hotspot gene comprises SEQ ID NOs. 33-41.
 8. The probe combination according to claim 5, further comprising:
 - one or more sets of exogenous gene targeting probes,
 - wherein when sequences of each of the sets of exogenous gene targeting probes are aligned, an overall sequence of the aligned set of exogenous gene targeting probes matches a reference sequence of an exogenous gene, and
 - in the aligned set of exogenous gene targeting probes, each of the exogenous gene targeting probes overlap with one or two adjacent exogenous gene targeting probes by a portion of a length of the exogenous gene targeting probe.
 9. The probe combination according to claim 8, wherein the exogenous gene originates a lambda phage.
 10. The probe combination according to claim 8, wherein the reference sequence of the exogenous gene comprises SEQ ID NOs. 42-54.
 11. The probe combination according to claim 8, further comprising:
 - one or more set of endogenous gene targeting probes,
 - wherein when sequences of each set of endogenous gene targeting probes are aligned, an overall sequence of the aligned set of endogenous gene targeting probes matches a reference sequence of an endogenous gene, and
 - in the aligned set of endogenous gene targeting probes, each of the endogenous gene targeting probes overlap with one or two adjacent endogenous gene targeting probes by a portion of a length of the endogenous gene targeting probe.
 12. The probe combination according to claim 11, wherein the endogenous gene comprises GAPDH gene and GdX gene.
 13. The probe combination according to claim 11, wherein the reference sequence of the endogenous gene comprises SEQ ID NO. 55 and SEQ ID NO. 56.
 14. The probe combination according to claim 1, wherein the cancer comprises hepatocellular carcinoma.

15. The probe combination according to claim 1, used for capturing target nucleotide fragments having viral-host junctions from DNA obtained for a specimen of a subject infected with HBV.
16. The probe combination according to claim 15, wherein the DNA comprises genomic DNA and circulating tumor DNA of the subject.
17. The probe combination according to claim 15, wherein the specimen comprises biological fluid and liver tissues.
18. A probe combination for detecting cancer, comprising:
 - one or more sets of full HBV targeting probes,
 - wherein when sequences of each of the sets of full HBV targeting probes are aligned, an overall sequence of the aligned set of full HBV targeting probes matches a reference sequence of a genome of a HBV genotype, and
 - in the aligned set of full HBV targeting probes, each of the full HBV targeting probes overlap with one or two adjacent full HBV targeting probes by a portion of a length of the full HBV targeting probe.
19. The probe combination according to claim 18, wherein the HBV genotype comprises genotype A, genotype B, genotype C, genotype D, genotype E, genotype F, genotype G, genotype H, genotype I and genotype J.
20. The probe combination according to claim 18, further comprising:
 - one or more sets of partial hepatitis B virus (HBV) targeting probes,
 - wherein when sequences of each of the sets of partial HBV targeting probes are aligned, an overall sequence of the aligned set of partial HBV targeting probes matches a reference sequence of a direct repeat (DR) region on the genome of the HBV genotype, and
 - in the aligned set of partial HBV targeting probes, each of the partial HBV targeting probes overlap with one or two adjacent partial HBV targeting probes by a portion of a length of the partial HBV targeting probe.
21. The probe combination according to claim 20, the reference sequence of the DR region comprises SEQ ID NOs. 3-32.
22. The probe combination according to claim 20, further comprising:
 - one or more sets of hotspot gene targeting probes,

wherein when sequences of each of the sets of hotspot gene targeting probes are aligned, an overall sequence of the aligned set of hotspot gene targeting probes matches a reference sequence of a cancer hotspot gene, and

in the aligned set of hotspot gene targeting probes, each of the hotspot gene targeting probes overlap with one or two adjacent hotspot gene targeting probes by a portion of a length of the hotspot gene targeting probe.

23. The probe combination according to claim 22, wherein the cancer hotspot gene comprises CTNNB1 gene, TERT gene, and TP53 gene.

24. The probe combination according to claim 22, wherein the reference sequence of the cancer hotspot gene comprises SEQ ID NOs. 33-41.

25. The probe combination according to claim 22, further comprising:

one or more sets of exogenous gene targeting probes,

wherein when sequences of each of the sets of exogenous gene targeting probes are aligned, an overall sequence of the aligned set of exogenous gene targeting probes matches a reference sequence of an exogenous gene, and

in the aligned set of exogenous gene targeting probes, each of the exogenous gene targeting probes overlap with one or two adjacent exogenous gene targeting probes by a portion of a length of the exogenous gene targeting probe.

26. The probe combination according to claim 25, wherein the exogenous gene originates a lambda phage.

27. The probe combination according to claim 25, wherein the reference sequence of the exogenous gene comprises SEQ ID NOs. 42-54.

28. The probe combination according to claim 25, further comprising:

one or more set of endogenous gene targeting probes,

wherein when sequences of each set of endogenous gene targeting probes are aligned, an overall sequence of the aligned set of endogenous gene targeting probes matches a reference sequence of an endogenous gene, and

in the aligned set of endogenous gene targeting probes, each of the endogenous gene targeting probes overlap with one or two adjacent endogenous gene targeting probes by a portion of a length of the endogenous gene targeting probe.

29. The probe combination according to claim 26, wherein the endogenous gene

comprises GAPDH gene and GdX gene.

30. The probe combination according to claim 26, wherein the reference sequence of the endogenous gene comprises SEQ ID NO. 55 and SEQ ID NO. 56.
31. The probe combination according to claim 18, wherein the cancer comprises hepatocellular carcinoma.
32. The probe combination according to claim 18, used for capturing target nucleotide fragments having viral-host junctions from DNA obtained for a specimen of a subject infected with HBV.
33. The probe combination according to claim 32, wherein the DNA comprises genomic DNA and circulating tumor DNA of the subject.
34. The probe combination according to claim 32, wherein the specimen comprises biological fluid and liver tissues.