Description

Title of Invention: MPHOSPH1 PEPTIDES AND VACCINES INCLUDING THE SAME

Technical Field

[0001] The present invention relates to the field of biological science, more specifically to the field of cancer therapy. In particular, the present invention relates to novel peptides that are

effective as cancer vaccines, as well as drugs for either or both of treating and preventing tumors.

[0002] Priority

The present application claims the benefit of U.S. Provisional Application No. 61/522,991, filed on August 12, 2011, the entire contents of which are incorporated by reference herein.

Background Art

[0003] Cytotoxic T lymphocytes (CTLs) have been shown to recognize epitope peptides derived from tumor-associated antigens (TAAs) found on major the histocompatibility complex (MHC) class I molecule, and then kill the tumor cells. Since the discovery of the melanoma antigen (MAGE) family, many other TAAs have been discovered through immunological approaches (NPL 1, Boon T, Int J Cancer 1993 May 8, 54(2): 177-80; NPL 2, Boon T & van der Bruggen P, J Exp Med 1996 Mar 1, 183(3): 725-9). Some of these TAAs are currently undergoing clinical development as immunotherapeutic targets.

[0004] Favorable TAAs are indispensable for the proliferation and survival of cancer cells. The use of such TAAs as targets for immunotherapy may minimize the well-described risk of immune escape of cancer cells attributable to deletion, mutation, or down-regulation of TAAs as a consequence of therapeutically driven immune selection. Accordingly, the identification of new TAAs capable of inducing potent and specific anti-tumor immune responses, warrants further development and thus the clinical application of peptide vaccination strategies for various types of cancer is ongoing (NPL 3, Harris CC, J Natl Cancer Inst 1996 Oct 16, 88(20): 1442-55; NPL 4, Butterfield LH et al., Cancer Res 1999 Jul 1, 59(13): 3134-42; NPL 5, Vissers JL et al., Cancer Res 1999 Nov 1, 59(21): 5554-9; NPL 6, van der Burg SH et al., J Immunol 1996 May 1, 156(9): 3308-14; NPL 7, Tanaka F et al., Cancer Res 1997 Oct 15, 57(20): 4465-8; NPL 8, Fujie T et al., Int J Cancer 1999 Jan 18, 80(2): 169-72; NPL 9, Kikuchi M et al., Int J Cancer 1999 May 5, 81(3): 459-66; NPL 10, Oiso M et al., Int J Cancer 1999 May 5, 81(3): 387-94). To date, several clinical trials using these tumor-associated

antigen derived peptides have been reported. Unfortunately, many of the current cancer vaccine trial have shown only a low objective response rate (NPL 11, Belli F et al., J Clin Oncol 2002 Oct 15, 20(20): 4169-80; NPL 12, Coulie PG et al., Immunol Rev 2002 Oct, 188: 33-42; NPL 13, Rosenberg SA et al., Nat Med 2004 Sep, 10(9): 909-15). Accordingly, there remains a need in the art for new TAAs as immunotherapeutic targets.

[0005] MPHOSPH1 (M-phase phosphoprotein 1; GenBank Accession No. NM_016195 and NP_057279, SEQ ID NO: 125 and 126), was identified as one of the proteins specifically phosphorylated at the G2/M transition and characterized as a plus-end-directed kinesin related protein (NPL 14, Abaza A et al., J Biol Chem 2003, 278: 27844-52.). More particularly, MPHOSPH1 has been reported to be a plus-end-directed molecular motor that plays a crucial role in cytokinesis, and accumulates in the midzone of the spindle during anaphase to telophase in HeLa cells (NPL 14, Abaza A et al., J Biol Chem 2003, 278: 27844-52; NPL 15, Kamimoto T et al., J Biol Chem 2001, 276: 37520-8). In the course of gene expression profile analyses using a genome-wide cDNA microarray containing 23,040 genes, MPHOSPH1 was identified as a novel molecule up-regulated in bladder cancer (NPL 16, Kanehira M et al., Cancer Res. 2007 Apr 1;67(7):3276-85.; PTL 1, WO2006/085684). Furthermore, through northern blot analysis, expression of the MPHOSPH1 gene products were found to be limited to the testis and absent from the normal vital organs.

[0006] Some peptide fragments derived from MPHOSPH1 having cytotoxic T lymphocyte (CTL) inducibility were previously identified (PTL 2, WO2008/047473). These peptide fragments demonstrated the ability to induce CTLs against cells stimulated with the cognate peptide fragments. However, previous studies failed to confirm whether the peptide fragments had the ability to induce CTLs targeting tumor cells expressing the MPHOSPH1 gene and HLA-A2 antigen.

Citation List

Patent Literature

[0007]

[PTL 1] WO2006/085684

[PTL 2] WO2008/047473

Non Patent Literature

[0008]

[NPL 1] Boon T, Int J Cancer 1993 May 8, 54(2): 177-80

[NPL 2] Boon T & van der Bruggen P, J Exp Med 1996 Mar 1, 183(3): 725-9

[NPL 3] Harris CC, J Natl Cancer Inst 1996 Oct 16, 88(20): 1442-55

[NPL 4] Butterfield LH et al., Cancer Res 1999 Jul 1, 59(13): 3134-42

[NPL 5] Vissers JL et al., Cancer Res 1999 Nov 1, 59(21): 5554-9

[NPL 6] van der Burg SH et al., J Immunol 1996 May 1, 156(9): 3308-14

[NPL 7] Tanaka F et al., Cancer Res 1997 Oct 15, 57(20): 4465-8
[NPL 8] Fujie T et al., Int J Cancer 1999 Jan 18, 80(2): 169-72
[NPL 9] Kikuchi M et al., Int J Cancer 1999 May 5, 81(3): 459-66
[NPL 10] Oiso M et al., Int J Cancer 1999 May 5, 81(3): 387-94
[NPL 11] Belli F et al., J Clin Oncol 2002 Oct 15, 20(20): 4169-80
[NPL 12] Coulie PG et al., Immunol Rev 2002 Oct, 188: 33-42
[NPL 13] Rosenberg SA et al., Nat Med 2004 Sep, 10(9): 909-15
[NPL 14] Abaza A et al., J Biol Chem 2003, 278: 27844-52.
[NPL 15] Kamimoto T et al., J Biol Chem 2001, 276: 37520-8
[NPL 16] Kanehira M et al., Cancer Res. 2007 Apr 1; 67(7):3276-85.

Summary of Invention

[0009] The present invention is based, at least in part, on the discovery of novel peptides that may serve as suitable targets of immunotherapy. Because TAAs are generally perceived by the immune system as "self" and therefore often have no immunogenicity, the discovery of appropriate targets is of extremely importance. As noted above, MPHOSPH1 (for example, SEQ ID NO: 126 encoded by the gene of GenBank Accession No. NM_016195 (SEQ ID NO: 125)) has been identified as up-regulated in cancers, examples of which include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, chronic myeloid leukemia (CML), colorectal cancer, gastric cancer, non-small cell lung cancer (NSCLC), lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor. Thus, the present invention focuses on MPHOSPH1 as a candidate target of cancer/tumor immunotherapy, more particularly novel MPHOSPH1 epitope peptides that may serve as suitable immunotherapeutic targets.

[0010] To that end, the present invention is directed, at least in part, to the identification of specific epitope peptides that possess the ability to induce CTLs specific to MPHOSPH1 among peptides derived from MPHOSPH1. As discussed in greater detail below, peripheral blood mononuclear cells (PBMCs) obtained from a healthy donor were stimulated using HLA-A*0201 binding candidate peptides derived from MPHOSPH1. CTL lines were then established with specific cytotoxicity against the HLA-A2 positive target cells pulsed with each of candidate peptides. The results herein demonstrate that these peptides are HLA-A2 restricted epitope peptides that may induce potent and specific immune responses against cells expressing MPHOSPH1. These results further indicate that MPHOSPH1 is strongly immunogenic and the epitopes thereof are effective targets for caner/tumor immunotherapy.

[0011] Accordingly, it is an object of the present invention to provide isolated peptides that bind HLA antigen and induce CTLs, wherein the peptides include an immunologically

active fragment of MPHOSPH1 (SEQ ID NO: 126). Such peptides can be used to induce CTLs in vitro or ex vivo, or to be administered directly to a subject so as to induce in vivo immune responses against cancers, examples of which include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor.

[0012] The peptides of the present invention are generally less than 15, 14, 13, 12, 11, or 10 amino acids in length. Preferred peptides of the present invention are nonapeptides or decapeptides. Particularly preferred peptides have an amino acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120, since those peptides demonstrated to bind to the HLA-A2 antigen and induce CTL.

[0013] Thus, in some embodiments, the peptides of the present invention are peptides of less than 15, 14, 13, 12, 11, or 10 amino acids in length that have an amino acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120. In typical embodiments, the peptides of the present invention are nonapeptides or decapeptides having an amino acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120. Furthermore, as demonstrated herein, a peptide having the amino acid sequence of SEQ ID NO: 120 was confirmed to induce CTLs targeting tumor cells expressing the MPHOSPH1 and HLA-A2 antigen. Thus, in preferred embodiments, the peptides of the present invention are peptides having an amino acid sequence of SEQ ID NO: 120.

[0014] When contacted with antigen presenting cells (APCs) in vitro, ex vivo or in vivo, the peptides of the present invention will bind with HLA-A2 antigens on APCs and be presented on APCs as complexes with HLA-A2 antigens. Alternatively, the peptides of the present invention may be taken into by APCs, processed to fragments composed of an amino acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 in APCs, and presented on APCs as complexes with HLA-A2 antigens. Consequently, CTLs specific to such peptides are induced and such CTLs are considered to be as elements of the present invention.

[0015] The present invention also contemplates modified peptides having an amino acid sequence in which one, two or several amino acids are substituted, deleted, inserted and/or added in the amino acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120, so long as the modified peptides retain the CTL inducibility equivalent to that of the original unmodified peptide. To that end, the present invention provides an isolated peptide of less than 15, 14, 13, 12, 11, or 10 amino acids in length, which has CTL inducibility and comprises the amino acid sequence selected from the group consisting of:

(i) an amino acid sequence that 1, 2, or several amino acid(s) are substituted in the

amino acid sequence selected from the group consisting of SEQ ID NOs: 5, 14 and 64, and

(ii) the amino acid sequence of (i), wherein the amino acid sequence has one or both of the following characteristics:

(a) the second amino acid from the N-terminus of said SEQ ID NO is selected from the group consisting of leucine and methionine; and

(b) the C-terminal amino acid of said SEQ ID NO is selected from the group consisting of valine and leucine.

[0016] Moreover, the present invention also provides an isolated peptide of less than 15, 14, 13, 12, or 11 amino acids in length, which has CTL inducibility and comprises the amino acid sequence selected from the group consisting of:

(i') an amino acid sequence that 1, 2, or several amino acid(s) are substituted in the amino acid sequence selected from the group consisting of SEQ ID NOs: 73, 77, 79, 97, 103 and 120, and

(ii') the amino acid sequence of (i'), wherein the amino acid sequence has one or both of the following characteristics:

(a) the second amino acid from the N-terminus of said SEQ ID NO is selected from the group consisting of leucine and methionine; and

(b) the C-terminal amino acid of said SEQ ID NO is selected from the group consisting of valine and leucine.

[0017] As demonstrated herein, such peptides may bind with HLA-A2 antigens on APCs and be presented on APCs as complexes with HLA-A2 antigens. Alternatively, such peptides may be taken into by APCs, processed to fragments composed of an amino acid sequence selected from among (i), (ii), (i'), and (ii') in APCs, and presented on APCs as complexes with HLA-A2 antigens, when those peptides are contacted with APCs. Consequently, CTLs specific to such peptides are induced and such CTLs are considered to be as elements of the present invention.

The present invention further encompasses isolated polynucleotides that encode any of the peptides of the present invention. These polynucleotides can be used to induce or prepare APCs having CTL inducibility. Like above-described peptides of the present invention, such APCs can be administered to a subject for inducing immune responses against cancers.

[0018] When administered to a subject, the peptides of the present invention are presented on the surface of APCs so as to induce CTLs targeting the respective peptides. Therefore, one object of the present invention is to provide agents or compositions that include one or more peptide(s) or polynucleotide(s) provided by the present invention for inducing either or both of APCs and CTLs. Such agents or compositions can be also used for one or more purpose(s) selected from among the treatment of cancer, the

prophylaxis of cancer, and the prevention of postoperative recurrence of cancer. Examples of targetted cancers include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor. Thus, it is yet another object of the present invention to provide pharmaceutical agents or compositions for either or both of the treatment of cancer and the prophylaxis of cancer, such pharmaceutical agents or compositions formulated to include one or more peptides or polynucleotides of the present invention. Instead of or in addition to the peptides or polynucleotides of the present invention, the pharmaceutical agents or compositions of the present invention may include as active ingredients APCs or exosomes that present any of the peptides of the present invention.

[0019] The peptides or polynucleotides of the present invention may be used to induce APCs that present on the surface a complex of an HLA antigen and a present peptide, for example, by contacting APCs with the peptide of the present invention or introducing a polynucleotide encoding a peptide of the present invention into APCs. Such APCs have the ability of inducing CTLs that specifically recognize cells that presents target peptides on their surface and find use in cancer immunotherapy. Accordingly, the present invention encompasses the methods for inducing APCs having CTL inducibility as well as the APCs obtained by such methods. In addition, the present invention also encompasses the agents or compositions for use in inducing APCs, such agents or compositions including any peptides or polynucleotides of the present invention.

[0020] It is further object of the present invention to provide a method for inducing CTLs, such method including the step of co-culturing CD8 positive T cells with APCs or exosomes presenting the peptide of the present invention on its surface or the step of introducing a polynucleotide encoding both of T cell receptor (TCR) subunits or polynucleotides encoding each of TCR subunits, wherein the TCR can binds to a complex of the peptide of the present invention and HLA antigen presented on cell surface. CTLs obtained by such methods can find use in either or both of the treatment of cancer and the prevention of cancer. Examples of cancers include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor.

[0021] Yet another object of the present invention is to provide isolated APCs that present on the surface a complex of an HLA antigen and a peptide of the present invention. The present invention further provides isolated CTLs that target peptides of the present invention. These APCs and CTLs find utility in the context of cancer immunotherapy.

It is yet another object of the present invention to provide methods for inducing an

immune response against a cancer in a subject in need thereof, such methods including the step of administering an agent or composition that include at least one component selected from among the peptides of the present invention, polynucleotides encoding such peptides, exosomes or APCs presenting such peptides and CTLs that recognize cells presenting such peptides on their surface.

[0022] The applicability of the present invention extends to any of a number of the diseases relating to or arising from MPHOSPH1 over-expression, such as cancer, examples of which include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor.

[0023] More specifically, the present invention provides followings:

[1] An isolated peptide of the following (a) or (b):

(a) a peptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120;

(b) a peptide comprising an amino acid sequence that 1, 2, or several amino acid(s) are substituted, deleted, inserted, and/or added in the amino acid sequence selected from the group consisting of SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120, and wherein the peptide has cytotoxic T lymphocyte (CTL) inducibility,

[2] The isolated peptide of [1], wherein the peptide has one or both of the following characteristics:

(a) the second amino acid from the N-terminus of the amino acid sequence selected from the group consisting of SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 is selected from the group consisting of leucine and methionine; and

(b) the C-terminal amino acid of the amino acid sequence selected from the group consisting of SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 is selected from the group consisting of valine and leucine,

[3] The isolated peptide of [1] or [2], wherein the peptide is a nonapeptide or decapeptide,

[4] An isolated polynucleotide encoding the peptide of any one of [1] to [3],

[5] A composition for inducing a CTL, wherein the composition comprises one or more peptide(s) of any one of [1] to [3], or one or more polynucleotide(s) of [4],

[6] A composition for inducing an APC having CTL inducibility, wherein the composition comprises one or more peptide(s) of any one of [1] to [3], or one or more polynucleotide(s) of [4],

[7] A pharmaceutical composition comprising at least one active ingredient selected from the group consisting of:

(a) one or more peptides of any one of [1] to [3];

(b) one or more polynucleotides encoding the peptide of any one of [1] to [3];

(c) one or more APCs or exosomes that present a complex of the peptide of any one of [1] to [3] and an HLA antigen on their surface; and

(d) one or more CTLs that recognize a cell presenting a complex of the peptide of any one of [1] to [3] and an HLA antigen on its surface,

[8] The pharmaceutical composition of [7] for use in either or both of the treatment and the prophylaxis of cancer, or inducing an immune response against cancer in a subject,

[9] The pharmaceutical composition of [7] or [8], wherein the pharmaceutical composition is formulated for administration to a subject whose HLA antigen is HLA-A2,

[10] A method for inducing an antigen-presenting cell (APC) having CTL inducibility, said method comprising a step selected from the group consisting of:

(a) contacting an APC with the peptide of any one of [1] to [3] in vitro, ex vivo or in vivo, and

(b) introducing a polynucleotide encoding the peptide of any one of [1] to [3] into an APC,

[11] A method for inducing a CTL, said method comprising a step selected from the group consisting of:

(a) co-culturing a CD8 positive T cell with an APC that presents on its surface a complex of an HLA antigen and the peptide of any one of [1] to [3];

(b) co-culturing a CD8 positive T cell with an exosome that presents on its surface a complex of an HLA antigen and the peptide of any one of [1] to [3]; and

(c) introducing a polynucleotide encoding both of T cell receptor (TCR) subunits or polynucleotides encoding each of TCR subunits into a CD8 positive T cell, wherein the TCR can bind to a complex of an HLA antigen and the peptide of any one of [1] to [3] presented on a cell surface,

[12] An isolated APC that presents on its surface a complex of an HLA antigen and the peptide of any one of [1] to [3],

[13] The APC of [12], which is induced by the method of [10],

[14] An isolated CTL that recognizes a cell presenting on its surface a complex of an HLA antigen and the peptide of any one of [1] to [3],

[15] The CTL of [14], wherein said CTL is induced by the method of [11],

[16] A method of either or both of the treatment and prophylaxis of cancer in a subject, wherein the method comprises the step of administering to the subject (a) pharmaceutically effective amount(s) of:

(a) one or more peptides of any one of [1] to [3];

(b) one or more polynucleotides encoding the peptide of any one of [1] to [3];

(c) one or more APCs or exosomes that present a complex of the peptide of any one of [1] to [3] and an HLA antigen on their surface; or

(d) one or more CTLs that recognize a cell presenting a complex of the peptide of any

one of [1] to [3] and an HLA antigen on its surface,

[17] A method of inducing an immune response against cancer in a subject in need thereof, said method comprising the step of administering to the subject a composition comprising a peptide of any one of [1] to [3] or a polynucleotide encoding the peptide,

[18] An antibody or immunologically active fragment thereof against the peptide of any one of [1] to [3],

[19] A vector comprising a nucleotide sequence encoding the peptide of any one of [1] to [3],

[20] A host cell transformed or transfected with the vector of [19], and

[21] A diagnostic kit comprising the peptide of any one of [1] to [3], the polynucleotide of [4] or the antibody of [18].

[0024] Objects and features of the invention will become more fully apparent when the following detailed description is read in conjunction with the accompanying figures and examples. It is to be understood that both the foregoing summary of the present invention and the following detailed description are of exemplified embodiments, and not restrictive of the present invention or other alternate embodiments of the present invention.

In particular, while the invention is described herein with reference to a number of specific embodiments, it will be appreciated that the description is illustrative of the invention and is not constructed as limiting of the invention. Various modifications and applications may occur to those who are skilled in the art, without departing from the spirit and the scope of the invention, as described by the appended claims. Likewise, other objects, features, benefits and advantages of the present invention will be apparent from this summary and certain embodiments described below, and will be readily apparent to those skilled in the art. Such objects, features, benefits and advantages will be apparent from the above in conjunction with the accompanying examples, data, figures and all reasonable inferences to be drawn there from, alone or with consideration of the references incorporated herein.

Brief Description of Drawings

[0025] Various aspects and applications of the present invention will become apparent to the skilled artisan upon consideration of the brief description of the figures and the detailed description of the present invention and its preferred embodiments that follow.

[0026] [fig.1]Figure 1 is composed of a series of photographs, (a) to (j), depicting the results of IFN-gamma ELISPOT assays on CTLs that were induced with peptides derived from MPHOSPH1. The CTLs in well number #7 stimulated with MPHOSPH1-A02-9-850 (SEQ ID NO: 5) (a), in #5 stimulated with MPHOSPH1-A02-9-129 (SEQ ID NO: 14) (b), in #5 stimulated with

MPHOSPH1-A02-9-846 (SEQ ID NO: 64) (c), in #2 stimulated with MPHOSPH1-A02-10-460 (SEQ ID NO: 73) (d), in #1 stimulated with MPHOSPH1-A02-10-770 (SEQ ID NO: 77) (e), in #1 stimulated with MPHOSPH1-A02-10-407 (SEQ ID NO: 79) (f), in #4 stimulated with MPHOSPH1-A02-10-923 (SEQ ID NO: 97) (g), in #5 stimulated with MPHOSPH1-A02-10-1484 (SEQ ID NO: 103) (h) and in #8 stimulated with MPHOSPH1-A02-10-282 (SEQ ID NO: 120) (i) showed potent IFN-gamma production as compared with the control, respectively. The square on the well of these pictures indicates that the cells from corresponding well were expanded to establish CTL lines. In contrast, as typical case of negative data, specific IFN-gamma production from the CTL stimulated with MPHOSPH1-A02-9-575 (SEQ ID NO: 1) (j) was not shown. In the figures, "+" indicates the IFN-gamma production against target cells pulsed with the appropriate peptide, and "-" indicates the IFN-gamma production against target cells not pulsed with any peptides.

[0027] [fig.2a-f]Figure 2a-f is composed of a series of line graphs, (a) to (f), depicting the results of an IFN-gamma ELISA assay demonstrating the IFN-gamma production of CTL lines stimulated with MPHOSPH1-A02-9-850 (SEQ ID NO: 5) (a), MPHOSPH1-A02-9-129 (SEQ ID NO: 14) (b), MPHOSPH1-A02-9-846 (SEQ ID NO: 64) (c), MPHOSPH1-A02-10-460 (SEQ ID NO: 73) (d), MPHOSPH1-A02-10-770 (SEQ ID NO: 77) (e), and MPHOSPH1-A02-10-407 (SEQ ID NO: 79) (f). The quantity of IFN-gamma which CTL lines produced was measured by IFN-gamma enzyme-linked immunosorbent assay (ELISA). The results demonstrate that CTL lines established by stimulation with each peptide show potent IFN-gamma production as compared with the control. In the figures, "+" indicates the IFN-gamma production against target cells pulsed with the appropriate peptide, and "-" indicates the IFN-gamma production against target cells not pulsed with any peptides. R/S ratio indicates the ratio of the number of responder cells (CTL line) and stimulator cells.

[0028] [fig.2g-i]Figure 2g-i is composed of a series of line graphs, (g) to (i), depicting the results of an IFN-gamma ELISA assay demonstrating the IFN-gamma production of CTL lines stimulated with MPHOSPH1-A02-10-923 (SEQ ID NO: 97) (g), MPHOSPH1-A02-10-1484 (SEQ ID NO: 103) (h) and MPHOSPH1-A02-10-282 (SEQ ID NO: 120) (i). The quantity of IFN-gamma which CTL lines produced was measured by IFN-gamma enzyme-linked immunosorbent assay (ELISA). The results demonstrate that CTL lines established by stimulation with each peptide show potent IFN-gamma production as compared with the control. In the figures, "+" indicates the IFN-gamma production against target cells pulsed with the appropriate peptide, and "-" indicates the IFN-gamma production against target cells not pulsed with any peptides. R/S ratio indicates the ratio of the number of responder cells (CTL line) and stimulator

cells.

[0029] [fig.3]Figure 3 is composed of a series of line graphs, (a) to (e), depicting the IFN-gamma production of the CTL clones established by limiting dilution from the CTL lines stimulated with MPHOSPH1-A02-9-850 (SEQ ID NO: 5) (a), MPHOSPH1-A02-9-846 (SEQ ID NO: 64) (b), MPHOSPH1-A02-10-460 (SEQ ID NO: 73) (c), MPHOSPH1-A02-10-770 (SEQ ID NO: 77) (d) and MPHOSPH1-A02-10-282 (SEQ ID NO: 120) (e). The results demonstrate that the CTL clones established by stimulation with each peptide show potent IFN-gamma production as compared with the control. In the figure, "+" indicates the IFN-gamma production against target cells pulsed with the appropriate peptide and "-" indicates the IFN-gamma production against target cells not pulsed with any peptides. R/S ratio indicates the ratio of the number of responder cells (CTL clone) and stimulator cells.

[0030] [fig.4]Figure 4 is a line graph depicting specific CTL activity against the tumor cell lines. J82 cells which express both MPHOSPH1 and HLA-A*0201, HT1376 cells which express MPHOSPH1 but not HLA-A*0201 and T2 cells which express HLA-A*0201 but not MPHOSPH1 were used as stimulator cells. The CTL clone established with MPHOSPH1-A02-10-282 (SEQ ID NO: 120) showed specific CTL activity against J82 cells. On the other hand, no significant specific CTL activity was detected against HT1376 and T2 cells. R/S ratio indicates the ratio of the number of the responder cells (CTL clone) and the stimulator cells.

[0031] [fig.5]Figure 5 is a line graph depicting cytotoxic activity of CTL against the tumor cell lines. UMUC-3 cells which express both MPHOSPH1 and HLA-A*0201, MKN45 cells which express MPHOSPH1 but not HLA-A*0201 and T2 that expressed HLA-A*0201 but not MPHOSPH1 were used as target cells. The CTL clone established with MPHOSPH1-A02-10-282 (SEQ ID NO: 120) showed potent cytotoxic activity against UMUC-3 cells. On the other hand, no significant specific CTL activity was detected against MKN45 and T2 cells. E/T ratio indicates the ratio of the number of the effecter cells (CTL clone) and the target cells.

[0032] [fig.6]Figure 6 is a line graph depicting cytotoxic activity of CTL against the target cells that express MPHOSPH1 and HLA-A*0206. COS7 cells transfected with HLA-A*0206 or the full length MPHOSPH1 gene were prepared as the controls. The CTL line established with MPHOSPH1-A02-10-282 (SEQ ID NO: 120) showed specific CTL activity against COS7 cells transfected with both MPHOSPH1 and HLA-A*0206 (black lozenge). On the other hand, no significant specific CTL activity was detected against target cells expressing either HLA-A*0206 (triangle) or MPHOSPH1 (circle).

Description of Embodiments

[0033] Although any methods and materials similar or equivalent to those described herein

can be used in the practice or testing of embodiments of the present invention, the preferred methods, devices, and materials are now described. However, before the present materials and methods are described, it should be understood that these descriptions are merely illustrative only and not intended to be limiting. It should also be understood that the present invention is not limited to the particular sizes, shapes, dimensions, materials, methodologies, protocols, etc. described herein, as these may vary in accordance with routine experimentation and/or optimization. Furthermore, the terminology used in the description is for the purpose of describing the particular versions or embodiments only, and is not intended to limit the scope of the present invention that will be limited only by the appended claims.

[0034] The disclosure of each publication, patent or patent application mentioned in this specification is specifically incorporated by reference herein in its entirety. However, nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue or prior invention.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which the present invention belongs. In case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

[0035] I. Definitions:

The words "a", "an", and "the" as used herein mean "at least one" unless otherwise specifically indicated.

The terms "isolated" and "purified" used in relation with a substance (e.g., peptide, antibody, polynucleotide, etc.) indicates that the substance is substantially free from at least one substance that may else be included in the natural source. Thus, an isolated or purified peptide refers to peptide that are substantially free of cellular material such as carbohydrate, lipid, or other contaminating proteins from the cell or tissue source from which the peptide is derived, or substantially free of chemical precursors or other chemicals when chemically synthesized. The term "substantially free of cellular material" includes preparations of a peptide in which the peptide is separated from cellular components of the cells from which it is isolated or recombinantly produced. Thus, a peptide that is substantially free of cellular material includes preparations of polypeptide having less than about 30%, 20%, 10%, or 5% (by dry weight) of heterologous protein (also referred to herein as a "contaminating protein"). When the peptide is recombinantly produced, it is also preferably substantially free of culture medium, which includes preparations of peptide with culture medium less than about 20%, 10%, or 5% of the volume of the peptide preparation. When the peptide is produced by chemical synthesis, it is preferably substantially free of chemical

precursors or other chemicals, which includes preparations of peptide with chemical precursors or other chemicals involved in the synthesis of the peptide less than about 30%, 20%, 10%, 5% (by dry weight) of the volume of the peptide preparation. That a particular peptide preparation contains an isolated or purified peptide can be shown, for example, by the appearance of a single band following sodium dodecyl sulfate (SDS)-polyacrylamide gel electrophoresis of the protein preparation and Coomassie Brilliant Blue staining or the like of the gel. In a preferred embodiment, peptides and polynucleotides of the present invention are isolated or purified.

[0036] The terms "polypeptide", "peptide" and "protein" are used interchangeably herein to refer to a polymer of amino acid residues. The terms apply to amino acid polymers in which one or more amino acid residue(s) may be modified residue(s), or non-naturally occurring residue(s), such as artificial chemical mimetic(s) of corresponding naturally occurring amino acid(s), as well as to naturally occurring amino acid polymers.

The term "oligopeptide" sometimes used in the present specification is used to refer to peptides of the present invention which are 20 residues or fewer, typically 15 residues or fewer in length and is typically composed of between about 8 and about 11 residues, often 9 or 10 residues. The latter are referred to herein as "nonapeptides" and "decapeptides", respectively.

[0037] The term "amino acid" as used herein refers to naturally occurring and synthetic amino acids, as well as amino acid analogs and amino acid mimetics that similarly function to the naturally occurring amino acids. Amino acid may be either L-amino acids or D-amino acids. Naturally occurring amino acids are those encoded by the genetic code, as well as those modified after translation in cells (e.g., hydroxyproline, gamma-carboxyglutamate, and O-phosphoserine). The phrase "amino acid analog" refers to compounds that have the same basic chemical structure (an alpha carbon bound to a hydrogen, a carboxy group, an amino group, and an R group) as a naturally occurring amino acid but have one or more modified R group(s) or modified backbones (e.g., homoserine, norleucine, methionine, sulfoxide, methionine methyl sulfonium). The phrase "amino acid mimetic" refers to chemical compounds that have different structures but similar functions to general amino acids.

[0038] Amino acids may be referred to herein by their commonly known three letter symbols or the one-letter symbols recommended by the IUPAC-IUB Biochemical Nomenclature Commission.

The terms "gene", "polynucleotide" and "nucleic acid" are used interchangeably herein and, unless otherwise specifically indicated are similarly to the amino acids referred to by their commonly accepted single-letter codes.

[0039] The terms "agent" and "composition" are used interchangeably herein to refer to a product that includes the specified ingredients in the specified amounts, as well as any

product that results, directly or indirectly, from combination of the specified ingredients in the specified amounts. Such terms, when used in relation to the modifier "pharmaceutical" (as in "pharmaceutical agent" and "pharmaceutical composition") are intended to encompass a product that includes the active ingredient(s), and any inert ingredient(s) that make up the carrier, as well as any product that results, directly or indirectly, from combination, complexation or aggregation of any two or more of the ingredients, or from dissociation of one or more of the ingredients, or from other types of reactions or interactions of one or more of the ingredients. Accordingly, in the context of the present invention, the terms "pharmaceutical agent" and "pharmaceutical composition" refer to any product made by admixing a molecule or compound of the present invention and a pharmaceutically or physiologically acceptable carrier.

[0040] The phrase "pharmaceutically acceptable carrier" or "physiologically acceptable carrier", as used herein, means a pharmaceutically or physiologically acceptable material, composition, substance or vehicle, including but not limited to, a liquid or solid filler, diluent, excipient, solvent or encapsulating material.

The pharmaceutical agents or compositions of the present invention find particular use as vaccines. In the context of the present invention, the phrase "vaccine" (also referred to as an "immunogenic composition") refers to an agent or composition that has the function to induce anti-tumor immunity upon inoculation into animals.

[0041] The term "active ingredient" herein refers to a substance in an agent or composition that is biologically or physiologically active. Particularly, in the context of pharmaceutical agent or composition, the term "active ingredient" refers to a substance that shows an objective pharmacological effect. For example, in case of pharmaceutical agents or compositions for use in the treatment or prevention of cancer, active ingredients in the agents or compositions may lead to at least one biological or physiologically action on cancer cells and/or tissues directly or indirectly. Preferably, such action may include reducing or inhibiting cancer cell growth, damaging or killing cancer cells and/or tissues, and so on. Typically, indirect effect of active ingredients is inductions of CTLs recognizing or killing cancer cells. Before being formulated, the "active ingredient" may also be referred to as "bulk", "drug substance" or "technical product".

[0042] Unless otherwise defined, the term "cancer" refers to the cancers that over-express MPHOSPH1 gene, examples of which include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor.

Unless otherwise defined, the terms "cytotoxic T lymphocyte", "cytotoxic T cell" and "CTL" are used interchangeably herein and unless otherwise specifically indicated,

refer to a sub-group of T lymphocytes that are capable of recognizing non-self cells (e.g., tumor/cancer cells, virus-infected cells) and inducing the death of such cells.

[0043] Unless otherwise defined, the term "HLA-A2", as used herein, representatively refers to the subtypes, examples of which include, but are not limited to, HLA-A*0201, HLA-A*0202, HLA-A*0203, HLA-A*0204, HLA-A*0205, HLA-A*0206, HLA-A*0207, HLA-A*0210, HLA-A*0211, HLA-A*0213, HLA-A*0216, HLA-A*0218, HLA-A*0219, HLA-A*0228 and HLA-A*0250.

[0044] Unless otherwise defined, the term "kit" as used herein, is used in reference to a combination of reagents and other materials. It is contemplated herein that the kit may include microarray, chip, marker, and so on. It is not intended that the term "kit" be limited to a particular combination of reagents and/or materials.

As used herein, in the context of a subject or patient, the phrase "subject's (or patient's) HLA antigen is HLA-A2" refers to that the subject or patient homozygously or heterozygously possess HLA-A2 antigen gene as the MHC (major histocompatibility complex) Class I molecule, and HLA-A2 antigen is expressed in cells of the subject or patient as an HLA antigen.

[0045] To the extent that the methods and compositions of the present invention find utility in the context of the "treatment" of cancer, a treatment is deemed "efficacious" if it leads to clinical benefit such as a decrease in size, prevalence, or metastatic potential of the cancer in a subject, retarding progression of cancer, alleviation of a clinical symptom of cancer, prolongation of survival time, suppression of postoperative recurrence and so on. When the treatment is applied prophylactically, "efficacious" means that it retards or prevents cancers from forming or prevents or alleviates a clinical symptom of cancer. Efficaciousness is determined in association with any known method for diagnosing or treating the particular tumor type.

[0046] To the extent that the methods and compositions of the present invention find utility in the context of the "prevention" and "prophylaxis" of cancer, such terms are interchangeably used herein to refer to any activity that reduces the burden of mortality or morbidity from disease. Prevention and prophylaxis can occur "at primary, secondary and tertiary prevention levels." While primary prevention and prophylaxis avoid the development of a disease, secondary and tertiary levels of prevention and prophylaxis encompass activities aimed at the prevention and prophylaxis of the progression of a disease and the emergence of symptoms as well as reducing the negative impact of an already established disease by restoring function and reducing disease-related complications. Alternatively, prevention and prophylaxis can include a wide range of prophylactic therapies aimed at alleviating the severity of the particular disorder, e.g. reducing the proliferation and metastasis of tumors.

[0047] In the context of the present invention, the treatment and/or prophylaxis of cancer

and/or the prevention of post operative recurrence thereof, include any activities that lead to, for example, the following events, such as the inhibition of the growth of cancerous cells, the involution or regression of a tumor, the induction of remission, the suppression of occurrence of cancer, the tumor regression, the reduction or inhibition of metastasis, the suppression of postoperative recurrence of cancer, and prolongation of survival time. Effective treatment and/or the prophylaxis of cancer decreases mortality and improves the prognosis of individuals having cancer, decreases the levels of tumor markers in the blood, and alleviates detectable symptoms accompanying cancer. For example, reduction or improvement of symptoms constitutes effectively treating and/or the prophylaxis include 10%, 20%, 30% or more reduction, or stable disease.

[0048] In the context of the present invention, the term "antibody" refers to immunoglobulins and fragments thereof that are specifically reactive to a designated protein or peptide thereof. An antibody can include human antibodies, primatized antibodies, chimeric antibodies, bispecific antibodies, humanized antibodies, antibodies fused to other proteins or radiolabels, and antibody fragments. Furthermore, an antibody herein is used in the broadest sense and specifically covers intact monoclonal antibodies, polyclonal antibodies, multi-specific antibodies (e.g. bispecific antibodies) formed from at least two intact antibodies, and antibody fragments so long as they exhibit the desired biological activity. An "antibody" indicates all classes (e.g. IgA, IgD, IgE, IgG and IgM).

[0049] II. Peptides:

Peptides of the present invention described in detail below may be referred to as "MPHOSPH1 peptide(s)" or " MPHOSPH1 polypeptide(s)".

To demonstrate that peptides derived from MPHOSPH1 function as an antigen recognized by CTLs, peptides derived from MPHOSPH1 (SEQ ID NO: 126) were analyzed to determine whether they were antigen epitopes restricted by HLA-A2 which are commonly encountered HLA alleles (Date Y et al., Tissue Antigens 47: 93-101, 1996; Kondo A et al., J Immunol 155: 4307-12, 1995; Kubo RT et al., J Immunol 152: 3913-24, 1994).

Candidates of HLA-A2 binding peptides derived from MPHOSPH1 were identified based on their binding affinities to HLA-A2.

[0050] Moreover, after in vitro stimulation of T-cells by dendritic cells (DCs) pulsed (loaded) with these peptides, CTLs were successfully established by stimulating the DCs with each of the following peptides:

MPHOSPH1-A2-9-850 (SEQ ID NO: 5),

MPHOSPH1-A2-9-129 (SEQ ID NO: 14),

MPHOSPH1-A2-9-846 (SEQ ID NO: 64),

MPHOSPH1- A2-10-460 (SEQ ID NO: 73),
MPHOSPH1-A2-10- 770 (SEQ ID NO: 77),
MPHOSPH1-A2-10- 407 (SEQ ID NO: 79),
MPHOSPH1-A2-10- 923 (SEQ ID NO: 97),
MPHOSPH1- A2-10-1484 (SEQ ID NO: 103) and
MPHOSPH1- A2-10-282 (SEQ ID NO: 120).

[0051] These established CTLs showed potent specific CTL activity against target cells pulsed with respective peptides. These results demonstrate that MPHOSPH1 is an antigen recognized by CTLs and that the peptides tested are epitope peptides of MPHOSPH1 restricted by HLA-A2; and therefore, the peptides may be effective as target antigens for cytotoxicity by CTLs. Furthermore, MPHOSPH1-A2-10-282 (SEQ ID NO: 120) induced CTLs having potent cytotoxic activity against cancer cells expressing both MPHOSPH1 and HLA-A2 antigen as the MHC Class I molecule. This result suggests that the MPHOSPH1-A2-10-282 peptide occurs naturally in vivo to be presented on cancer cells expressing MPHOSPH1 by HLA-A2 antigen (e.g., HLA-A*0201 or HLA-A*0206). According to the findings, a peptide comprising the amino acid sequence selected from the group consisting of SEQ ID NO: 5, 14, 64, 73, 77, 79, 97, 103 and 120, or derivatives, mutants, variants or modified peptides thereof - are useful in the context of immunological therapy for treating a cancer that expresses MPHOSPH1 and HLA-A2 antigen. In certain embodiments of the present invention, peptide composed of an amino acid sequence disclosed herein can be used for immunological therapy of cancer. Examples of cancers to be treated include, for example, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor. However, the peptides of the present invention can be applied to any cancers, so long as they express MPHOSPH1 and HLA-A2 antigen.

[0052] Since the MPHOSPH1 gene is over-expressed in cancer cells such as bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor, but is not expressed in most normal organs, it is a good target for immunotherapy. Thus, the present invention provides nonapeptides (peptides composed of nine amino acid residues) and decapeptides (peptides composed of ten amino acid residues) of CTL-recognized epitopes from MPHOSPH1. Alternatively, the present invention provides isolated peptides which bind to HLA antigens and induce cytotoxic T lymphocytes (CTLs), wherein the peptide is composed of an immunologically active fragment of MPHOSPH1 (SEQ ID NO: 126). More specifically, in some embodiments, the present invention provides peptides comprising an amino acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120. In preferred

embodiments, the peptides of the present invention are peptides that comprise an amino acid sequence of SEQ ID NO: 120.

[0053] Generally, software programs now available, for example, on the Internet, such as those described in Parker KC et al., J Immunol 1994 Jan 1, 152(1): 163-75 and Nielsen M et al., Protein Sci 2003; 12: 1007-17 can be used to calculate the binding affinities between various peptides and HLA antigens in silico. Binding affinity with HLA antigens can be measured as described, for example, in Parker KC et al., J Immunol 1994 Jan 1, 152(1): 163-75, Kuzushima K et al., Blood 2001, 98(6): 1872-81, Larsen MV et al. BMC Bioinformatics. 2007 Oct 31; 8: 424, Buus S et al. Tissue Antigens., 62:378-84, 2003, Nielsen M et al., Protein Sci 2003; 12: 1007-17, and Nielsen M et al. PLoS ONE 2007; 2: e796, which are summarized in, e.g., Lafuente EM et al., Current Pharmaceutical Design, 2009, 15, 3209-3220. Methods for determining binding affinity are described, for example, in the Journal of Immunological Methods (1995, 185: 181-190) and Protein Science (2000, 9: 1838-1846). Therefore, one can utilize such software programs to select those fragments derived from MPHOSPH1 that have high binding affinity with HLA antigens. Accordingly, the present invention encompasses peptides composed of any fragments derived from MPHOSPH1, which would be determined to bind with HLA antigens by such known programs. Furthermore, such peptides may include the peptide composed of the full length sequence of MPHOSPH1.

[0054] The peptides of the present invention, particularly the nonapeptides and decapeptides of the present invention, may be flanked with additional amino acid residues so long as the peptides retain their CTL inducibility. The particular additional amino acid residues may be composed of any kind of amino acids so long as they do not impair the CTL inducibility of the original peptide. Thus, the present invention encompasses peptides having CTL inducibility, wherein the peptides include an amino acid sequence derived from MPHOSPH1. Such peptides are, for example, less than about 40 amino acids, often less than about 20 amino acids, usually less than about 15 amino acids.

[0055] It is generally known that modifications of one, two, several or more amino acids in a peptide do not influence the function of the peptide, or in some cases even enhance the desired function of the original peptide. In fact, modified peptides (i.e., peptides composed of an amino acid sequence modified by substituting, inserting, deleting and/or adding one, two or several amino acid residues to an original reference sequence) have been known to retain the biological activity of the original peptide (Mark et al., Proc Natl Acad Sci USA 1984, 81: 5662-6; Zoller and Smith, Nucleic Acids Res 1982, 10: 6487-500; Dalbadie-McFarland et al., Proc Natl Acad Sci USA 1982, 79: 6409-13). Thus, in one embodiment of the present invention, the peptide having CTL inducibility of the present invention may be composed of a peptide having an amino

acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120, in which one, two or several amino acids are added, deleted, inserted and/or substituted. In another embodiment, the peptides of the present invention may be peptides comprising an amino acid sequence in which one, two, or several amino acid(s) are substituted, deleted, inserted, and/or added in the amino acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120, provided the modified peptide retain the CTL inducibility of the original peptide. In preferred embodiments, the peptide of the present invention may be peptides comprising an amino acid sequence in which one, two, or several amino acid(s) are substituted, deleted, inserted, and/or added in the amino acid sequence of SEQ ID NO: 120, provided the modified peptide retain the CTL inducibility of the original peptide.

[0056] Those of skill in the art will recognize that individual modifications (i.e., additions, insertions, deletions and/or substitutions) to an amino acid sequence that alter a single amino acid or a small percentage of the overall amino acid sequence tend to result in the conservation of the properties of the original amino acid side-chain; it is thus referred to as "conservative substitution" or "conservative modification", wherein the alteration of a protein results in a protein with similar functions. Conservative substitution tables providing functionally similar amino acids are well known in the art. Examples of properties of amino acid side chains are hydrophobic amino acids (A, I, L, M, F, P, W, Y, V), hydrophilic amino acids (R, D, N, C, E, Q, G, H, K, S, T), and side chains having the following functional groups or characteristics in common: an aliphatic side-chain (G, A, V, L, I, P); a hydroxyl group containing side-chain (S, T, Y); a sulfur atom containing side-chain (C, M); a carboxylic acid and amide containing side-chain (D, N, E, Q); a base containing side-chain (R, K, H); and an aromatic group containing side-chain (H, F, Y, W). In addition, the following eight groups each contain amino acids that are conservative substitutions for one another:

1) Alanine (A), Glycine (G);
2) Aspartic acid (D), Glutamic acid (E);
3) Asparagine (N), Glutamine (Q);
4) Arginine (R), Lysine (K);
5) Isoleucine (I), Leucine (L), Methionine (M), Valine (V);
6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W);
7) Serine (S), Threonine (T); and
8) Cysteine (C), Methionine (M) (see, e.g., Creighton, Proteins 1984).

Such conservatively modified peptides are also considered to be peptides of the present invention. However, the peptide of the present invention is not restricted thereto and may include non-conservative modifications, so long as the resulting modified peptide retains the CTL inducibility of the original unmodified peptide. Fur-

thermore, the modified peptides should not exclude CTL inducible peptides derived from polymorphic variants, interspecies homologues, or alleles of MPHOSPH1.

[0057] Amino acid residues may be inserted, substituted, deleted and/or added to the peptides of the present invention or, alternatively, amino acid residues may be deleted therefrom to achieve a higher binding affinity. To retain the requisite CTL inducibility, one preferably modifies (inserts, deletes, adds and/or substitutes) only a small number (for example, 1, 2 or several) or a small percentage of amino acids. Herein, the term "several" means 5 or fewer amino acids, for example, 4 or 3 or fewer. The percentage of amino acids to be modified may be, for example, 20% or less, preferably 15% or less, more preferably 10% or less, even more preferably 1 to 5%.

[0058] When used in the context of cancer immunotherapy, the peptides of the present invention may be presented on the surface of a cell or exosome as a complex with an HLA antigen. Therefore, it is preferable to select peptides that not only induce CTLs but also possess high binding affinity to the HLA antigen. To that end, the peptides can be modified by substitution, insertion, deletion and/or addition of amino acid residues to yield a modified peptide having improved binding affinity. In addition to peptides that are naturally displayed, since the regularity of the sequences of peptides displayed by binding to HLA antigens has already been known (J Immunol 1994, 152: 3913; Immunogenetics 1995, 41: 178; J Immunol 1994, 155: 4307), modifications based on such regularity may be introduced into the immunogenic peptides of the present invention.

[0059] For example, peptides exhibiting high HLA-A2 binding affinity tend to have the second amino acid from the N-terminus substituted with leucine or methionine. Likewise, peptides in which the C-terminal amino acid is substituted with valine or leucine can also be favorably used. Thus, peptides having an amino acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 in which the second amino acid from the N-terminus of the amino acid sequence of the SEQ ID NO is substituted with leucine or methionine, and/or wherein the C-terminus of the amino acid sequence of the SEQ ID NO is substituted with valine or leucine are contemplated by the present invention. In another embodiment, the present invention encompasses peptides having an amino acid sequence in which the second amino acid from the N-terminus of the amino acid sequence selected from among of the SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 is substituted with leucine or methionine, and/or the C-terminus of the amino acid sequence of the SEQ ID NO is substituted with valine or leucine. In preferred embodiments, the peptides of the present invention may comprise an amino acid sequence in which the second amino acid from the N-terminus of the amino acid sequence of SEQ ID NO: 120 is substituted with leucine or methionine, and/or the C-terminus of the amino acid sequence of the SEQ ID NO is substituted

with valine or leucine.

[0060] In one embodiment, the present invention provides the peptides having CTL inducibility, wherein the peptides have general formula selected from the group consisting of (1) to (9) as follows:

(1) -corresponding to MPHOSPH1-A2-9-850 (SEQ ID NO: 5)-
F [X1] L T I E N E [X2],

(2) -corresponding to MPHOSPH1-A2-9-129 (SEQ ID NO: 14)-
F [X1] G C I M Q P [X2],

(3) -corresponding to MPHOSPH1-A2-9-846 (SEQ ID NO: 64)-
G [X1] R A F L L T [X2],

(4) -corresponding to MPHOSPH1- A2-10-460 (SEQ ID NO: 73)-
Y [X1] A Y D E T L N [X2],

(5) -corresponding to MPHOSPH1-A2-10- 770 (SEQ ID NO: 77)-
K [X1] I C N E T V E [X2]

(6) -corresponding to MPHOSPH1-A2-10- 407 (SEQ ID NO: 79)-
L [X1] T L G K C I N [X2]

(7) -corresponding to MPHOSPH1-A2-10- 923 (SEQ ID NO: 97)-
K [X1] S N E I E T A [X2]

(8) -corresponding to MPHOSPH1- A2-10-1484 (SEQ ID NO: 103)-
Q [X1] V A A L E I Q [X2], and

(9) -corresponding to MPHOSPH1- A2-10-282 (SEQ ID NO: 120)-
Y [X1] Y D L F V P V [X2].

[0061] In the general formula (1)-(9), [X1] is leucine or methionine, and [X2] is valine or leucine. In a particularly preferred embodiment of the present invention, the general formula may be (9), which corresponds to SEQ ID NO: 120. The present invention further provides isolated peptide represented by the general formula (1)-(9) defined above, to which one, two, or several amino acids are added at either or both of N-terminus and C-terminus thereof. In an alternative embodiment, the present invention provides isolated peptides represented by the general formula (1)-(9) from which one, two, or several amino acid residues are deleted at either or both of N-terminus and C-terminus thereof. The present invention also provides isolated peptide represented by the general formula (1)-(9), to which one, two, or several amino acids are inserted or deleted at anywhere of the sequence.

[0062] Substitutions may be introduced not only at the terminal amino acids but also at the positions of potential T cell receptor (TCR) recognition sites of peptides. Several studies have demonstrated that a peptide with amino acid substitutions may have equal to or better function than that of the original, for example, CAP1, $p53_{(264\text{-}272)}$, Her-2/$neu_{(369\text{-}377)}$ or $gp100_{(209\text{-}217)}$ (Zaremba et al. Cancer Res. 57, 4570-4577, 1997, T. K.

Hoffmann et al. J Immunol. (2002) Feb 1;168(3):1338-47., S. O. Dionne et al. Cancer Immunol immunother. (2003) 52: 199-206 and S. O. Dionne et al. Cancer Immunology, Immunotherapy (2004) 53, 307-314).

The present invention also contemplates the addition of one, two or several amino acids may also be added to either or both of the N and C-terminus of the peptides of the present invention. Such modified peptides retaining CTL inducibility are also included in the present invention.

[0063] However, when the peptide sequence is identical to a portion of the amino acid sequence of an endogenous or exogenous protein having a different function, negative side effects such as autoimmune disorders or allergic symptoms against specific substances may be induced. Therefore, it may be desirable to perform homology searches using available databases to avoid situations in which the sequence of the peptide matches the amino acid sequence of another protein. When it becomes clear from the homology searches that no peptide identical to or having a 1 or 2 amino acid difference with respect to the objective peptide exists in nature, the objective peptide may be modified in order to increase its binding affinity with HLA antigens, and/or increase its CTL inducibility without any danger of such side effects.

[0064] Although peptides having high binding affinity to the HLA antigens as described above are expected to be highly effective, the candidate peptides, which are selected according to the presence of high binding affinity as an indicator, are further examined for the presence of CTL inducibility. Herein, the phrase "CTL inducibility" indicates the ability of a peptide to induce a CTL when presented on an antigen-presenting cell (APC). Further, "CTL inducibility" includes the ability of a peptide to induce CTL activation, CTL proliferation, promote lysis of target cells by CTL, and to increase IFN-gamma production by CTL.

[0065] Confirmation of CTL inducibility is accomplished by inducing APCs carrying human MHC antigens (for example, B-lymphocytes, macrophages, and dendritic cells (DCs)), or more specifically DCs derived from human peripheral blood mononuclear leukocytes, and after stimulation of APCs with a test peptide, mixing APCs with CD8-positive cells to induce CTLs, and then measuring the IFN-gamma against the target cells produced and released by CTLs. As the reaction system, transgenic animals that have been produced to express a human HLA antigen (for example, those described in BenMohamed L, Krishnan R, Longmate J, Auge C, Low L, Primus J, Diamond DJ, Hum Immunol 2000 Aug, 61(8): 764-79, Related Articles, Books, Linkout Induction of CTL response by a minimal epitope vaccine in HLA A*0201/DR1 transgenic mice: dependent on MHC (HLA) class II restricted T(H) response) can be used. Alternatively, the target cells may be radiolabeled with ^{51}Cr and such, and cytotoxic activity of CTLs may be calculated from radioactivity released

from the target cells. Alternatively, it may be examined by measuring IFN-gamma produced and released by CTLs in the presence of cells that carry immobilized peptides, and visualizing the inhibition zone on the media using anti-IFN-gamma monoclonal antibodies.

[0066] As a result of examining the CTL inducibility of the peptides as described above, it was discovered that nonapeptides and decapeptides selected from among those peptides having the amino acid sequence indicated by SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 showed CTL inducibility as well as high binding affinity to an HLA antigen. Thus, these peptides are exemplified as preferred embodiments of the present invention.

[0067] Furthermore, homology analysis results demonstrated that such peptides do not share significant homology with peptides derived from any other known human gene products. This lowers the possibility of unknown or undesired immune responses when used for immunotherapy. Therefore, also from this aspect, these peptides are useful for eliciting immunity against MPHOSPH1 in cancer patients. Thus, peptides having an amino acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120, are encompassed by the present invention.

[0068] In addition to modification as discussed above, the peptides of the present invention may be linked to other peptides, so long as the resulting linked peptide retains the CTL inducibility of the original peptide. Examples of suitable "other" peptides include: the peptides of the present invention or the CTL inducible peptides derived from other TAAs. The peptide of the present invention can be linked to a "other" peptide directly or indirectly via a linker. The linkers between the peptides are well known in the art, for example, AAY (P. M. Daftarian et al., J Trans Med 2007, 5:26), AAA, NKRK (R. P. M. Sutmuller et al., J Immunol. 2000, 165: 7308-7315) or K (S. Ota et al., Can Res. 62, 1471-1476, K. S. Kawamura et al., J Immunol. 2002, 168: 5709-5715).

[0069] For example, peptides derived from non-MPHOSPH1 tumor-associated antigens also can be used to increase immune response via HLA class I and/or class II. It is well-known in the art that cancer cells express more than one tumor associated gene. Some CTL inducible peptides derived from such TAAs have been isolated (for example, WO2008/047473, WO2010/047062, WO2008/102557, WO2009/025116). Accordingly, examples of "other" peptides that is linked to the peptide of the present invention include, but are not limited to, CTL inducible peptides derived from TAAs other than MHPOSPH1. In the present invention, "other" peptides may not be only MHC Class I restricted peptides but also MHC Class II restricted peptide. One of ordinary skill in the art can prepare polypeptides including one or more MPHOSPH1 peptides and one or more of the non-MPHOSPH1 peptides, or nucleic acids encoding such polypeptides, using conventional molecular biology procedures.

[0070] The above-described linked peptides are referred to herein as "polytopes", i.e., groups of two or more potentially immunogenic or immune response stimulating peptides that can be joined together in various arrangements (e.g., concatenated, overlapping). The polytope (or nucleic acid encoding the polytope) can be administered in accordance with standard immunization protocols, e.g., to animals, to test the effectiveness of the polytope in stimulating, enhancing and/or provoking an immune response.

[0071] The peptides can be joined together directly or via the use of flanking sequences to form polytopes, and the use of polytopes as vaccines is well known in the art (see, e.g., Thomson et al., Proc. Natl. Acad. Sci USA 92(13):5845-5849, 1995; Gilbert et al., Nature Biotechnol. 15(12):1280-1284, 1997; Thomson et al., J Immunol. 157(2):822-826, 1996; Tarn et al., J Exp. Med. 171(l):299-306, 1990). Polytopes containing various numbers and combinations of epitopes can be prepared and tested for recognition by CTLs and for efficacy in increasing an immune response.

[0072] The peptides of the present invention may be further linked to other substances, so long as they retain the CTL inducibility. Illustrative examples of such "other" substances include, but are not limited to, peptides, lipids, sugar and sugar chains, acetyl groups, natural and synthetic polymers, etc. The peptides may contain modifications such as glycosylation, side chain oxidation, or phosphorylation, so long as the modifications do not destroy the biological activity of the peptides as described herein. These kinds of modifications may be performed to confer additional functions (e.g., targeting function, and delivery function) or to stabilize the polypeptide.

[0073] For example, to increase the in vivo stability of a polypeptide, it is known in the art to introduce D-amino acids, amino acid mimetics or unnatural amino acids; this concept may also be adopted for the present polypeptides. The stability of a polypeptide may be assayed in a number of ways. For instance, peptidases and various biological media, such as human plasma and serum, can be used to test stability (see, e.g., Verhoef et al., Eur J Drug Metab Pharmacokin 1986, 11: 291-302).

When the peptides of the present invention include a cysteine residue, the peptides tend to form dimers via a disulfide bond between SH groups of the cysteine residues. Therefore, dimers of the peptides of the present invention are also included in the peptides of the present invention.

[0074] Moreover, as noted above, among the modified peptides that are substituted, deleted, inserted and/or added by one, two or several amino acid residues, those having same or higher activity as compared to original peptides can be screened for or selected. The present invention, therefore, also provides the method of screening for or selecting modified peptides having same or higher activity as compared to originals. For example, the method may include steps of:

a: modifying (i.e., substituting, deleting, inserting or adding) at least one amino acid residue of a peptide of the present invention,

b: determining the activity of the peptide modified in step (a), and

c: selecting the peptide having same or higher activity as compared to the original peptide.

Herein, the activity may include MHC binding activity and APC or CTL inducibility. Preferably, the activity of the peptide is CTL inducibility.

[0075] III. Preparation of MPHOSPH1 Peptides:

The peptides of the present invention may be prepared using well known techniques. For example, the peptides of the present invention may be prepared synthetically, by recombinant DNA technology or chemical synthesis. The peptides of the present invention may be synthesized individually or as longer polypeptides including two or more peptides. The peptides may be isolated, i.e., purified or isolated substantially free from other naturally occurring host cell proteins and fragments thereof, or any other chemical substances.

[0076] The peptides of the present invention may contain modifications, such as glycosylation, side chain oxidation, or phosphorylation, provided such modifications do not destroy the biological activity of the original peptides. Other illustrative modifications include incorporation of D-amino acids or other amino acid mimetics that may be used, for example, to increase the serum half-life of the peptides.

[0077] A peptide of the present invention may be obtained through chemical synthesis based on the selected amino acid sequence. For example, conventional peptide synthesis methods that may be adopted for the synthesis include:

(i) Peptide Synthesis, Interscience, New York, 1966;

(ii) The Proteins, Vol. 2, Academic Press, New York, 1976;

(iii) Peptide Synthesis (in Japanese), Maruzen Co., 1975;

(iv) Basics and Experiment of Peptide Synthesis (in Japanese), Maruzen Co., 1985;

(v) Development of Pharmaceuticals (second volume) (in Japanese), Vol. 14 (peptide synthesis), Hirokawa, 1991;

(vi) WO99/67288; and

(vii) Barany G. & Merrifield R.B., Peptides Vol. 2, "Solid Phase Peptide Synthesis", Academic Press, New York, 1980, 100-118.

[0078] Alternatively, the peptides of the present invention may be obtained adopting any known genetic engineering methods for producing peptides (e.g., Morrison J, J Bacteriology 1977, 132: 349-51; Clark-Curtiss & Curtiss, Methods in Enzymology (eds. Wu et al.) 1983, 101: 347-62). For example, first, a suitable vector harboring a polynucleotide encoding the objective peptide in an expressible form (e.g., downstream of a regulatory sequence corresponding to a promoter sequence) is prepared and

transformed into a suitable host cell. Such vectors and host cells are also provided by the present invention. The host cell is then cultured to produce the peptide of interest. The peptide may also be produced in vitro adopting an in vitro translation system.

[0079] IV. Polynucleotides:

The present invention provides polynucleotides that encode any of the aforementioned peptides of the present invention. The polynucleotides of the present invention may include polynucleotides derived from the natural occurring MPHOSPH1 gene (for example, GenBank Accession No. NM_001031702 (SEQ ID NO: 125)) or those having a conservatively modified nucleotide sequences thereof. Herein, the phrase "conservatively modified nucleotide sequence" refers to sequences which encode identical or essentially identical amino acid sequences. Due to the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given protein. For instance, the codons GCA, GCC, GCG, and GCU all encode the amino acid alanine. Thus, at every position where an alanine is specified by a codon, the codon may be altered to any of the corresponding codons described without altering the encoded polypeptide. Such nucleic acid variations, referred to in the art as "silent variations," represent one species of conservatively modified variant. Every nucleic acid sequence described herein as encoding a peptide also describes every possible silent variation of the nucleic acid. One of skill in the art will readily recognize that each codon in a nucleic acid (except AUG, which is ordinarily the only codon for methionine, and TGG, which is ordinarily the only codon for tryptophan) may be modified to yield a functionally identical molecule. Accordingly, each disclosed peptide-encoding nucleotide sequence represents an implicit disclosure of the silent variations associated therewith.

[0080] The polynucleotide of the present invention may be composed of DNA, RNA, and derivatives thereof. As is well known in the art, a DNA molecule is suitably composed of bases such as the naturally occurring bases A, T, C, and G, and T is replaced by U in an RNA. One of skill in the art will recognize that non-naturally occurring bases be included in polynucleotides, as well.

[0081] The polynucleotide of the present invention may encode multiple peptides of the present invention with or without intervening amino acid sequences. For example, the intervening amino acid sequence may provide a cleavage site (e.g., enzyme recognition sequence) of the polynucleotide or the translated peptides. Furthermore, the polynucleotide of the present invention may include any additional sequences to the coding sequence encoding the peptide of the present invention. For example, the polynucleotide of the present invention may be a recombinant polynucleotide that includes regulatory sequences required for the expression of the peptide or may be an expression vector (plasmid) with marker genes and such. In general, such recombinant

polynucleotides may be prepared by the manipulation of polynucleotides through conventional recombinant techniques using, for example, polymerases and endonucleases.

[0082] Both recombinant and chemical synthesis techniques may be used to produce the polynucleotides of the present invention. For example, the polynucleotide of the present invention may be produced by insertion of the polynucleotide having the coding sequence of the peptide of the present invention into an appropriate vector, which may be expressed when transfected into a competent cell. Alternatively, the polynucleotide of the present invention may be amplified using PCR techniques or replicated in a suitable host (see, e.g., Sambrook et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York, 1989). Alternatively, the polynucleotide of the present invention may be synthesized using the solid phase techniques, as described in Beaucage SL & Iyer RP, Tetrahedron 1992, 48: 2223-311; Matthes et al., EMBO J 1984, 3: 801-5.

[0083] V. Exosomes:

The present invention further provides intracellular vesicles called exosomes, which present complexes formed between the peptides of the present invention and HLA antigens on their surface. Exosomes may be prepared, for example by using the methods detailed in Japanese Patent Application Kohyo Publications No. Hei 11-510507 and WO99/03499, and may be prepared using APCs obtained from patients who are subject to treatment and/or prevention. The exosomes of the present invention may be inoculated as vaccines, similarly to the peptides of the present invention.

[0084] The type of HLA antigens included in the complexes must match that of the subject requiring treatment and/or prevention. For example, for Japanese, HLA-A2, particularly HLA-A*0201 and HLA-A*0206 are often appropriate. The use of HLA-A2 type that is highly expressed among the Japanese and Caucasian is favorable for obtaining effective results, and subtypes such as HLA-A*0201 and HLA-A*0206 find use. Typically, in the clinic, the type of HLA antigen of the patient requiring treatment is investigated in advance, which enables appropriate selection of peptides having high levels of binding affinity to this antigen, or having CTL inducibility by antigen presentation. Furthermore, in order to obtain peptides showing high binding affinity and CTL inducibility, substitution, deletion, or addition of 1, 2, or several amino acids may be performed based on the amino acid sequence of the naturally occurring MPHOSPH1 partial peptide.

[0085] When using the HLA-A2 type HLA antigen for the exosomes of the present invention, the peptides having an amino acid sequence of any one of SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 have particular utility. In some embodiments, the exosomes of the present invention are exosomes that present a complex of the peptide of the present invention and HLA-A2 antigen on their surface. Typical examples of the

HLA-A2 antigen contained in such complexes include, but are not limited to, HLA-A*0201 and HLA-A*0206.

[0086] VI. Antigen-Presenting Cells (APCs)

The present invention also provides isolated APCs that present complexes formed with HLA antigens and the peptides of the present invention on their surface. The APCs may be derived from patients who are subject to treatment and/or prevention, and may be administered as vaccines by themselves or in combination with other drugs including the peptides, exosomes, or CTLs of the present invention.

[0087] The APCs are not limited to a particular kind of cells and include dendritic cells (DCs), Langerhans cells, macrophages, B cells, and activated T cells, which are known to present proteinaceous antigens on their cell surface so as to be recognized by lymphocytes. Since DCs are representative APCs having the strongest CTL inducing activity among APCs, DCs are suitable for the APCs of the present invention.

[0088] For example, the APCs of the present invention may be obtained by inducing DCs from peripheral blood monocytes and then contacting (stimulating) them with the peptides of the present invention in vitro, ex vivo or in vivo. When the peptides of the present invention are administered to a subject, APCs that present the peptides of the present invention are induced in the body of the subject. Therefore, the APCs of the present invention may be obtained by collecting the APCs from the subject after administering the peptides of the present invention to the subject. Alternatively, the APCs of the present invention may be obtained by contacting APCs collected from a subject with the peptide of the present invention.

[0089] The APCs of the present invention may be administered to a subject for inducing immune response against cancer in the subject by themselves or in combination with other drugs including the peptides, exosomes or CTLs of the present invention. For example, the ex vivo administration may include steps of:

a: collecting APCs from a first subject,

b: contacting the APCs of step a, with the peptide of the present invention, and

c: administering the APCs of step b to a second subject.

[0090] The first subject and the second subject may be the same individual, or may be different individuals. The APCs obtained by step b may be administered as a vaccine for treating and/or preventing cancer, examples of which include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor.

The present invention also provides a method or process for manufacturing a pharmaceutical composition for inducing APCs, wherein the method includes the step of admixing or formulating the peptide of the present invention with a pharmaceutically

acceptable carrier.

[0091] According to an aspect of the present invention, the APCs of the present invention have CTL inducibility. In the context of the APCs, the phrase "having CTL inducibility" refers to showing higher CTL inducibility than those of APCs contacted with no peptides. Such APCs having CTL inducibility may be prepared by a method which includes the step of transferring a polynucleotide encoding the peptide of the present invention to APCs in vitro as well as the method mentioned above. The introduced genes may be in the form of DNA or RNA. Examples of methods for introduction include, without particular limitations, various methods conventionally performed in this field, such as lipofection, electroporation, or calcium phosphate method may be used. More specifically, it may be performed as described in Cancer Res 1996, 56: 5672-7; J Immunol 1998, 161: 5607-13; J Exp Med 1996, 184: 465-72; Published Japanese Translation of International Publication No. 2000-509281. By transferring the gene into APCs, the gene undergoes transcription, translation, and such in the cell, and then the obtained protein is processed by MHC Class I or Class II, and proceeds through a presentation pathway to present partial peptides.

In some embodiments, the APCs of the present invention are APCs that present complexes of HLA-A2 antigen and the peptide of the present invention on their surface. Typical examples of the HLA-A2 antigen contained in such complexes include, but are not limited to, HLA-A*0201 and HLA-A*0206.

[0092] VII. Cytotoxic T Lymphocytes (CTLs):

A CTL induced against any of the peptides of the present invention strengthens the immune response targeting cancer cells in vivo and thus may be used as vaccines similar to the peptides. Thus, the present invention provides isolated CTLs that are specifically induced or activated by any of the present peptides of the present invention.

[0093] Such CTLs may be obtained by (1) administering the peptide(s) of the present invention to a subject,,(2) contacting (stimulating) subject-derived APCs, and CD8 positive T cells, or peripheral blood mononuclear leukocytes in vitro with the peptide(s) of the present invention, (3) contacting CD8 positive T cells or peripheral blood mononuclear leukocytes in vitro with the APCs or exosomes presenting a complex of an HLA antigen and the peptide on their surface or (4) introducing a polynucleotide encoding both of T cell receptor (TCR) subunits or polynucleotides encoding each of TCR subunits, wherein the TCR can bind a complex of the peptide of the present invention and HLA-A2 antigen on a cell surface. Such APCs or exosomes to be used in preparation of CTLs may be prepared by the methods described above. Details of the method of (4) is described bellow in section "VIII. T Cell Receptor (TCR)".

[0094] The CTLs of the present invention may be derived from patients who are subject to treatment and/or prevention, and may be administered by themselves or in combination with other drugs including the peptides, APCs or exosomes of the present invention for the purpose of regulating effects. The obtained CTLs act specifically against target cells presenting the peptides of the present invention, for example, the same peptides used for induction. The target cells may be cells that endogenously express MPHOSPH1, such as cancer cells, or cells that are transfected with the MPHOSPH1 gene; and cells that present a peptide of the present invention on the cell surface due to stimulation by the peptide may also serve as targets of activated CTL attack.

[0095] In some embodiments, the CTLs of the present invention recognize cells presenting complexes of HLA-A2 antigen and the peptide of the present invention. In the context of the CTL, the phrase "recognize a cell" refers to binding a complex of HLA-A2 antigen and the peptide of the present invention on the cell surface via its TCR and showing specific cytotoxic activity against the cell. Herein, "specific cytotoxic activity" refers to showing cytotoxic activity against the cell presenting a complex of HLA-A2 antigen and the peptide of the present invention but not other cells. Typical examples of the HLA-A2 antigen contained in such complex include, but are not limited to, HLA-A*0201 and HLA-A*0206.

[0096] VIII. T Cell Receptor (TCR):

The present invention also provides for compositions that include a polynucleotide encoding both of TCR subunits or polynucleotides encoding each of TCR subunits, wherein the TCR can bind to a complex of HLA-A2 antigen and the peptide of the present invention on a cell surface, and methods of using the same. Such TCR subunits have the ability to form TCRs that confer specificity to T cells against tumor cells expressing MPHOSPH1. By using the known methods in the art, the polynucleotides encoding each of alpha- and beta- chains of the TCR of the CTL induced with the peptide of the present invention can be identified (WO2007/032255 and Morgan et al., J Immunol, 171, 3288 (2003)). For example, the PCR method can be preferably used. The PCR primers for the analysis can be, for example, 5'-R primer (5'-gtctaccaggcattcgcttcat-3') (SEQ ID NO: 127) as a 5' side primer and 3-TRa-C primer (5'-tcagctggaccacagccgcagcgt-3') (SEQ ID NO: 128) specific to TCR alpha-chain C region, 3-TRb-C1 primer (5'-tcagaaatcctttctcttgac-3') (SEQ ID NO: 129) specific to TCR beta-chain C1 region or 3-TRbeta-C2 primer (5'- ctagcctctggaatc-ctttctctt-3') (SEQ ID NO: 130) specific to TCR beta-chain C2 region as 3' side primers, but not limited thereto. The derivative TCRs can bind target cells presenting the peptide of the present invention with high avidity, and optionally mediate efficient killing of target cells presenting the peptide of the present invention in vivo and in vitro.

[0097] The polynucleotide encoding both of the TCR subunits or polynucleotides encoding each of the TCR subunits may be incorporated into suitable vectors, e.g., retroviral vectors. These vectors are well known in the art. The polynucleotides or the vectors including them usefully may be transferred into a T cell (e.g., CD8 positive T cell), for example, a T cell from a patient. Advantageously, the present invention provides an off-the-shelf composition allowing rapid modification of a patient's own T cells (or those of another mammal) to rapidly and easily produce modified T cells having excellent cancer cell killing properties.

[0098] Specific TCRs against the peptides of the present invention should be capable of specifically recognizing a complex of the peptide of the present invention and HLA antigen, giving a T cell specific activity against a target cell presenting a complex of the peptide of the present invention and HLA antigen when the TCR is presented on the surface of the T cell. The requisite activity can be confirmed by any known methods that CTLs prepared by introducing the polypeptide(s) encoding such TCR subunits can be specifically recognize such target cells. Preferred examples of such methods include, for example, HLA multimer staining analysis using HLA molecules and the peptides of the present invention, and ELISPOT assay. By performing the ELISPOT assay, it can be confirmed that CTLs prepared by the above methods can specifically recognize the target cells, and that signals generated by such recognition can be transmitted intracellularly. Furthermore, it can be also confirmed by known methods that CTLs prepared by the above methods have specific cytotoxic activity against the target cells. Examples of such methods include, for example, Cr release assay using cells expressing both of HLA-A2 antigen and MPHOSPH1.

[0099] In one aspect, the present invention provides CTLs which are prepared by transduction with the polynucleotide encoding both of the TCR subunits or polynucleotides encoding each of the TCR subunits, wherein the TCR can bind to a complex of the peptide having an amino acid sequence selected from the group consisting of SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 and HLA-A2 antigen on a cell surface.

[0100] The transduced CTLs are capable of homing to cancer cells in vivo, and may be expanded by well known culturing methods in vitro (e.g., Kawakami et al., J Immunol., 142, 3452-3461 (1989)). The CTLs of the present invention may be used to form an immunogenic composition useful in either or both of the treatment and the prevention of cancer in a patient in need of therapy or protection (WO2006/031221).

[0101] IX. Pharmaceutical Compositions:

Since MPHOSPH1 expression is specifically elevated in cancer, examples of which include, but are not necessarily limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC,

lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor, as compared with normal tissues, the peptides or polynucleotides of the present invention may be used for either or both of the treatment and the prophylaxis of cancer. Thus, the present invention provides a pharmaceutical agent or composition formulated for either or both of the treatment and prophylaxis of cancer, such agent or composition including one or more peptides, or polynucleotides of the present invention as active ingredients. Alternatively, any of the foregoing exosomes or APCs that present a complex of the peptide of the present invention and HLA antigen may be used as active ingredients for pharmaceutical agents or compositions. In addition, the aforementioned CTLs that can recognize a cell presenting a complex of the peptide of the present invention and HLA antigen may also be used as active ingredients for pharmaceutical agents or compositions of the present invention.

[0102] Accordingly, the present invention provides agents or compositions that include at least one active ingredient selected from among:

(a) one or more peptides of the present invention;

(b) one or more polynucleotides encoding such a peptide as disclosed herein in an expressible form;

(c) one or more APCs or exosomes of the present invention; and

(d) one or more CTLs of the present invention.

[0103] The pharmaceutical agents or compositions of the present invention find use as a vaccine. In the context of the present invention, the phrase "vaccine" (also referred to as an immunogenic composition) refers to a composition that has the function to improve, enhance and/or induce anti-tumor immunity upon inoculation into animals. In other words, the present invention provides the pharmaceutical agents or compositions of the present invention for inducing an immune response against cancer in a subject.

[0104] The pharmaceutical agents or compositions of the present invention can be used for either or both of the treatment and the prevention of cancer in a subject. Examples of such subjects to which the pharmaceutical agents or compositions may be applied include humans, as well as other mammal including, but not limited to, mouse, rat, guinea-pig, rabbit, cat, dog, sheep, goat, pig, cattle, horse, monkey, baboon, and chimpanzee, particularly a commercially important animal or a domesticated animal. In some embodiments, the pharmaceutical agents or compositions of the present invention can be formulated for the administration to a subject whose HLA antigen is HLA-A2.

[0105] In another embodiment, the present invention also provides the use of an active ingredient in the manufacture of a pharmaceutical agent or composition formulated for either or both of the treatment and the prevention of cancer or tumor, including the post-operative recurrence thereof, such active ingredient selected from among:

(a) a peptide of the present invention;

(b) a polynucleotide encoding such a peptide as disclosed herein in an expressible form;

(c) an APC or an exosome presenting a peptide of the present invention on its surface; and

(d) a cytotoxic T cell of the present invention.

[0106] Alternatively, the present invention further provides an active ingredient for use in either or both of the treatment and prevention of a cancer or tumor, such active ingredient selected from among:

(a) a peptide of the present invention;

(b) a polynucleotide encoding such a peptide as disclosed herein in an expressible form;

(c) an APC or an exosome presenting a peptide of the present invention on its surface; and

(d) a cytotoxic T cell of the present invention.

[0107] Alternatively, the present invention further provides a method or process for manufacturing a pharmaceutical composition or agent for either or both of the treatment and prevention of a cancer or tumor, wherein the method or process includes the step of formulating a pharmaceutically or physiologically acceptable carrier with an active ingredient selected from among:

(a) a peptide of the present invention;

(b) a polynucleotide encoding such a peptide as disclosed herein in an expressible form;

(c) an APC or an exosome of the present invention; and

(d) a cytotoxic T cell of the present invention.

[0108] In another embodiment, the present invention also provides a method or process for manufacturing a pharmaceutical composition or agent for either or both of the treatment and prevention of a cancer or tumor, wherein the method or process includes the steps of admixing an active ingredient with a pharmaceutically or physiologically acceptable carrier, wherein the active ingredient is selected from among:

(a) a peptide of the present invention;

(b) a polynucleotide encoding such a peptide as disclosed herein in an expressible form;

(c) an APC or an exosome of the present invention; and

(d) a cytotoxic T cell of the present invention.

[0109] In another embodiment, the present invention also provides the method for either or both of the treatment and prevention of cancer or tumor, wherein the method comprises the step of administering to a subject at least one active ingredient selected from

among:

(a) a peptide of the present invention;

(b) a polynucleotide encoding such a peptide as disclosed herein in an expressible form;

(c) an APC or an exosome of the present invention; and

(d) a cytotoxic T cell of the present invention.

[0110] According to the present invention, peptides having an amino acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 have been found to be HLA-A2 restricted epitope peptides and thus serve as candidates that may induce specific immune response against cancer expressing HLA-A2 and MPHOSPH1 in a subject. Therefore, the pharmaceutical agents or compositions of the present invention that include any of these peptides, with the amino acid sequence of SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 or 120, are particularly suited for the administration to subjects whose HLA antigen is HLA-A2. Particularly preferred example of these peptides is the peptide having an amino acid sequence of SEQ ID NO: 120 that was confirmed to have the ability to induce CTLs targeting cancer cells expressing HLA-A2 antigen and MPHOSPH1. Accordingly, in preferred embodiments, the pharmaceutical agents or compositions of the present invention will include the peptide having an amino acid sequence of SEQ ID NO: 120 or a modified version thereof, or include a polynucleotide encoding such peptide. The same applies to pharmaceutical agents or compositions that include polynucleotides encoding any of these peptides (i.e., the polynucleotides of the present invention).

[0111] Cancers to be treated by the pharmaceutical agents or compositions of the present invention include any cancer in which MPHOSPH1 is expressed (e.g., is over-expressed), examples of which include, but not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor.

[0112] The pharmaceutical agents or compositions of the present invention may contain in addition to the aforementioned active ingredients, such as other peptides that have the ability to induce CTLs against cancerous cells, other polynucleotides encoding the other peptides, other cells that present the other peptides, and the like. Examples of such "other" peptides having the ability to induce CTLs against cancerous cells include, but are not limited to, cancer specific antigens (e.g., identified TAAs).

[0113] If necessary, the pharmaceutical agents or compositions of the present invention may optionally include other therapeutic substances as an additional active ingredient, so long as the substance does not inhibit the antitumoral effect of the active ingredient of the present invention (i.e., the peptide, polynucleotide, exosome, APC, CTL of the

present invention). For example, formulations may include anti-inflammatory substances, pain killers, chemotherapeutics, and the like. In addition to other therapeutic substances in the medicament itself, the medicaments of the present invention may also be administered sequentially or concurrently with the one or more other pharmacologic agents or compositions. The amounts of medicament and pharmacologic agent or composition depend, for example, on what type of pharmacologic agent(s) or composition(s) is/are used, the disease being treated, and the scheduling and routes of administration.

Those of skill in the art will readily recognize that, in addition to the ingredients particularly mentioned herein, the pharmaceutical agents or compositions of the present invention may include other substances conventional in the art having regard to the type of formulation in question (e.g., fillers, binders, diluents, excipients, etc.).

[0114] In one embodiment of the present invention, the pharmaceutical agents or compositions of the present invention may be packaged in articles of manufacture, e.g., as kits containing materials useful for treating the pathological conditions of the disease to be treated, e.g., cancer. The article of manufacture may include a container of any of the present pharmaceutical agents or compositions with a label. Suitable containers include bottles, vials, and test tubes. The containers may be formed from a variety of materials, such as glass or plastic. The label on the container should indicate the agent or composition is used for treatment or prevention of one or more conditions of the disease. The label may also indicate directions for administration and so on.

[0115] In addition to the container described above, a kit including a pharmaceutical agent or composition of the present invention may optionally further include a second container housing a pharmaceutically-acceptable diluent. It may further include other materials desirable from a commercial and user standpoint, including other buffers, diluents, filters, needles, syringes, and package inserts with instructions for use.

The pharmaceutical agents or compositions of the present invention can, if desired, be packaged in a pack or dispenser device that can contain one or more unit dosage forms containing the active ingredient. The pack can, for example, include metal or plastic foil, such as a blister pack. The pack or dispenser device can be accompanied by instructions for administration.

[0116] (1) Pharmaceutical compositions containing the peptides as the active ingredient:

The peptides of the present invention can be administered directly as a pharmaceutical agents or composition, or if necessary, that may be formulated by conventional formulation methods. In the latter case, in addition to the peptides of the present invention, carriers, excipients, and such that are ordinarily used for drugs can be included as appropriate without particular limitations. Examples of such carriers include, but are not limited to, sterilized water, physiological saline, phosphate buffer,

culture fluid and such. Furthermore, the pharmaceutical substances, agents or compositions can contain as necessary, stabilizers, suspensions, preservatives, surfactants and such. The pharmaceutical agents or compositions of the present invention can be used for anticancer purposes.

[0117] The peptides of the present invention can be prepared in combination, which includes two or more of peptides of the present invention, to induce CTL in vivo. The peptides can be in a cocktail or can be conjugated to each other using standard techniques. For example, the peptides can be chemically linked or expressed as a single fusion polypeptide sequence that may have one or several amino acid(s) as a linker (e.g., Lysine linker: K. S. Kawamura et al. J. Immunol. 2002, 168: 5709-5715). The peptides in the combination can be the same or different. By administering the peptides of the present invention, the peptides are presented in high density by the HLA antigens on APCs, then CTLs that specifically react toward the complex formed between the displayed peptide and the HLA antigen are induced. Alternatively, APCs (e.g., DCs) may be removed from a subject and then stimulated by the peptides of the present invention to obtain APCs that present any of the peptides of the present invention on their cell surface. These APCs can be re-administered to the subject to induce CTLs in the subjects, and as a result, aggressiveness towards the tumor-associated endothelium can be increased.

[0118] The pharmaceutical agents or compositions of the present invention, that include any of the peptides of the present invention as active ingredients, can also include an adjuvant so that cellular immunity will be established effectively. Alternatively, the pharmaceutical agent or composition of the present invention can be administered with other active ingredients or can be administered by formulation into granules. An adjuvant refers to any compound, substance or composition that enhances the immune response against the protein when administered together (or successively) with the protein having immunological activity. An adjuvant that can be applied includes those described in the literature (Clin Microbiol Rev 1994, 7: 277-89). Exemplary adjuvants include aluminum phosphate, aluminum hydroxide, alum, cholera toxin, salmonella toxin, Incomplete Freund's adjuvant (IFA), Complete Freund's adjuvant (CFA), IS-COMatrix, GM-CSF, CpG, O/W emulsion, and such, but are not limited thereto.

Furthermore, liposome formulations, granular formulations in which the peptide is bound to few-micrometers diameter beads, and formulations in which a lipid is bound to the peptide may be conveniently used.

[0119] In another embodiment of the present invention, the peptides of the present invention may also be administered in the form of a pharmaceutically acceptable salt. Preferable examples of the salts include salts with an alkali metal, salts with a metal, salts with an organic base, salts with an amine, salts with an organic acid (acetic acid, formic acid,

propionic acid, fumaric acid, maleic acid, succinic acid, tartaric acid, citric acid, malic acid, oxalic acid, benzoic acid, methanesulfonic acid and so on) and salts with an inorganic acid (hydrochloric acid, phosphoric acid, hydrobromic acid, sulfuric acid and so on). As used herein, the phrase "pharmaceutically acceptable salt" refers to those salts that retain the biological effectiveness and properties of the compound and that are obtained by reaction with inorganic or organic acids or bases.

[0120] In some embodiments, the pharmaceutical agents or compositions of the present invention include a component which primes CTL. Lipids have been identified as substances capable of priming CTL in vivo against viral antigens. For example, palmitic acid residues can be attached to the epsilon- and alpha-amino groups of a lysine residue and then linked to a peptide of the present invention. The lipidated peptide can then be administered either directly in a micelle or particle, incorporated into a liposome, or emulsified in an adjuvant. As another example of lipid priming of CTL responses, E. coli lipoproteins, such as tripalmitoyl-S-glycerylcysteinyl-seryl-serine (P3CSS) can be used to prime CTL when covalently attached to an appropriate peptide (see, e.g., Deres et al., Nature 1989, 342: 561-4).

[0121] Examples of suitable methods of administration include, but are not necessarily limited to, oral, intradermal, subcutaneous, intramuscular, intraosseous, peritoneal, and intravenous injection, or such, and systemic administration or local administration to the vicinity of the targeted sites (i.e., direct injection). The administration can be performed by single administration or boosted by multiple administrations. The dose of the peptides of the present invention can be adjusted appropriately according to the disease to be treated, age of the patient, weight, method of administration, and such, and is ordinarily 0.001 mg to 1,000 mg, for example, 0.01 mg to 100 mg, for example, 0.1 mg to 10 mg, and can be administered once in a few days to few months. One skilled in the art readily determine suitable and optimal dosages.

[0122] (2) Pharmaceutical compositions containing polynucleotides as active ingredient:

The pharmaceutical agents or compositions of the present invention can also include polynucleotides encoding the peptide(s) disclosed herein in an expressible form. Herein, the phrase "in an expressible form" means that the polynucleotide, when introduced into a cell, will be expressed in vivo as a polypeptide that induces anti-tumor immunity. In an illustrative embodiment, the nucleic acid sequence of the polynucleotide of interest includes regulatory elements necessary for expression of the polynucleotide. The polynucleotide(s) can be equipped so to achieve stable insertion into the genome of the target cell (see, e.g., Thomas KR & Capecchi MR, Cell 1987, 51: 503-12 for a description of homologous recombination cassette vectors. See also, e.g., Wolff et al., Science 1990, 247: 1465-8; U.S. Patent Nos. 5,580,859; 5,589,466; 5,804,566; 5,739,118; 5,736,524; 5,679,647; and WO 98/04720). Examples of DNA-

based delivery technologies include "naked DNA", facilitated (bupivacaine, polymers, peptide-mediated) delivery, cationic lipid complexes, and particle-mediated ("gene gun") or pressure-mediated delivery (see, e.g., U.S. Patent No. 5,922,687).

[0123] The peptides of the present invention can also be expressed by viral or bacterial vectors. Examples of expression vectors include attenuated viral hosts, such as vaccinia or fowlpox. This approach involves the use of vaccinia virus, e.g., as a vector to express nucleotide sequences that encode the peptide. Upon introduction into a host, the recombinant vaccinia virus expresses the immunogenic peptide, and thereby elicits an immune response. Vaccinia vectors and methods useful in immunization protocols are described in, e.g., U.S. Patent No. 4,722,848. Another vector is BCG (Bacille Calmette Guerin). BCG vectors are described in Stover et al., Nature 1991, 351: 456-60. A wide variety of other vectors useful for therapeutic administration or immunization, e.g., adeno and adeno-associated virus vectors, retroviral vectors, Salmonella typhi vectors, detoxified anthrax toxin vectors, and the like, will be apparent. See, e.g., Shata et al., Mol Med Today 2000, 6: 66-71; Shedlock et al., J Leukoc Biol 2000, 68: 793-806; Hipp et al., In Vivo 2000, 14: 571-85.

[0124] Delivery of a polynucleotide into a patient can be either direct, in which case the patient is directly exposed to a polynucleotide-carrying vector, or indirect, in which case, cells are first transformed with the polynucleotide of interest in vitro, then the cells are transplanted into the patient. Theses two approaches are known, respectively, as in vivo and ex vivo gene therapies.

[0125] For general reviews of the methods of gene therapy, see Goldspiel et al., Clinical Pharmacy 1993, 12: 488-505; Wu and Wu, Biotherapy 1991, 3: 87-95; Tolstoshev, Ann Rev Pharmacol Toxicol 1993, 33: 573-96; Mulligan, Science 1993, 260: 926-32; Morgan & Anderson, Ann Rev Biochem 1993, 62: 191-217; Trends in Biotechnology 1993, 11(5): 155-215). Methods commonly known in the art of recombinant DNA technology that are applicable to the present invention are described by Ausubel et al., in Current Protocols in Molecular Biology, John Wiley & Sons, NY, 1993; and by Krieger, in Gene Transfer and Expression, A Laboratory Manual, Stockton Press, NY, 1990.

[0126] Like administration of peptides, administration of polynucleotides may be performed by oral, intradermal, subcutaneous, intravenous, intramuscular, intraosseous, and/or peritoneal injection, or such, and via systemic administration or local administration to the vicinity of the targeted sites finds use. The administration can be performed by single administration or boosted by multiple administrations. The dose of the polynucleotide in the suitable carrier or cells transformed with the polynucleotide encoding the peptides of the present invention can be adjusted appropriately according to the disease to be treated, age of the patient, weight, method of administration, and such,

and is ordinarily 0.001 mg to 1000 mg, for example, 0.01 mg to 100 mg, for example, 0.1 mg to 10 mg, and can be administered once every a few days to once every few months. One skilled in the art can appropriately select the suitable dose.

[0127] X. Methods using Peptides, Exosomes, APCs and CTLs:

The peptides and polynucleotides of the present invention can be used for preparing or inducing APCs and CTLs. The exosomes and APCs of the present invention can be also used for inducing CTLs. The peptides, polynucleotides, exosomes and APCs can be used in combination with any other compounds so long as the additional compounds do not inhibit CTL inducibility. Thus, any of the aforementioned pharmaceutical agents or compositions of the present invention can be used for inducing CTLs. In addition thereto, those including the peptides and polynucleotides can be also used for inducing APCs as explained below.

[0128] (1) Methods of inducing antigen-presenting cells (APCs):

The present invention provides methods of inducing APCs with CTL inducibility using the peptides or polynucleotides of the present invention.

The methods of the present invention include the step of contacting APCs with the peptides of the present invention in vitro, ex vivo or in vivo. For example, the method including the step of contacting APCs with the peptides of the present invention ex vivo can include steps of:

a: collecting APCs from a subject:, and

b: contacting the APCs of step a with the peptide of the present invention.

[0129] The APCs are not limited to a particular kind of cells and include DCs, Langerhans cells, macrophages, B cells, and activated T cells, which are known to present proteinaceous antigens on their cell surface so as to be recognized by lymphocytes. Preferably, DCs can be used since they have the strongest CTL inducibility among APCs. Any one of peptide of the present invention can be used by itself or in combination with other peptides of the present invention or CTL inducible peptides derived from TAAs other than MPHOSPH1.

[0130] On the other hand, when the peptides of the present invention are administered to a subject, the APCs are contacted with the peptides in vivo, and consequently, the APCs with CTL inducibility are induced in the body of the subject. Thus, the method of the present invention includes administering the peptides of the present invention to a subject to induce APCs with CTL inducibility in the body of the subject. Similarly, when the polynucleotides of the present invention are administered to a subject in an expressible form, the peptides of the present invention are expressed and contacted with APCs in vivo, and consequently, APCs with CTL inducibility are induced in the body of the subject. Thus, the method of the present invention may also include administering the polynucleotides of the present invention to a subject to induce APCs with

CTL inducibility in the body of the subject. The phrase "expressible form" is described above in section "IX. Pharmaceutical Compositions, (2) Pharmaceutical compositions containing polynucleotides as the active ingredient".

[0131] Furthermore, the method of the present invention may include introducing the polynucleotide of the present invention into an APCs to induce APCs with CTL inducibility. For example, the method can include steps of:

a: collecting APCs from a subject:, and

b: introducing a polynucleotide encoding the peptide of the present invention into an APC.

Step b can be performed as described above in section "VI. Antigen-presenting cells".

[0132] Alternatively, the present invention provides a method for preparing an antigen-presenting cell (APC) which specifically induces CTL activity against MPHOSPH1, wherein the method can include one of the following steps:

(a) contacting an APC with a peptide of the present invention in vitro, ex vivo or in vivo; and

(b) introducing a polynucleotide encoding a peptide of the present invention into an APC.

Alternatively, the present invention provides methods for inducing an APC having CTL inducibility, wherein the methods include the step selected from among:

(a) contacting an APC with the peptide of the present invention;

(b) introducing the polynucleotide encoding the peptide of the present invention into an APC.

[0133] The methods of the present invention can be carried out in vitro, ex vivo or in vivo. Preferably, the methods of the present invention can be carried out in vitro or ex vivo. APCs used for induction of APCs having CTL inducibility can be preferably APCs expressing HLA-A2 antigen. Such APCs can be prepared by the methods well-known in the arts from peripheral blood mononuclear cells (PBMCs) obtained from a subject whose HLA antigen is HLA-A2. The APCs induced by the method of the present invention can be APCs that present a complex of the peptide of the present invention and HLA antigen (HLA-A2 antigen) in their surface. When APCs induced by the method of the present invention are administered to a subject in order to induce immune responses against cancer in the subject, the subject is preferably the same one from whom APCs are derived. However, the subject may be a different one from the APC donor so long as the subject has the same HLA type with the APC donor.

In another embodiment, the present invention provide agents or compositions for use in inducing an APC having CTL inducibility, and such agents or compositions include one or more peptides or polynucleotides of the present invention.

[0134] In another embodiment, the present invention provides for the use of the peptide of the present invention or the polynucleotide encoding the peptide in the manufacture of an agent or composition formulated for inducing APCs.

Alternatively, the present invention further provides the peptide of the present invention or the polypeptide encoding the peptide for use in inducing an APC having CTL inducibility.

[0135] (2) Methods of inducing CTLs:

The present invention also provides methods for inducing CTLs using the peptides, polynucleotides, exosomes or APCs of the present invention.

The present invention also provides methods for inducing CTLs using a polynucleotide encoding both of TCR subunits or polynucleotides encoding each of TCR subunits, wherein the TCR can recognize (bind to) a complex of the peptide of the present invention and HLA antigen presented on a cell surface. Preferably, the methods for inducing CTLs may include at least one step selected from among:

a) contacting a CD8 positive T cell with an antigen-presenting cell and/or an exosome that presents on its surface a complex of an HLA antigen and a peptide of the preset invention; and

b) introducing a polynucleotide encoding both of TCR subunits or polynucleotides encoding each of TCR subunits into a CD8 positive T cell, wherein the TCR can recognize (bind to) a complex of a peptide of the present invention and an HLA antigen presented on a cell surface.

[0136] When the peptides, the polynucleotides, APCs, or exosomes of the present invention are administered to a subject, CTLs are induced in the body of the subject, and the immune response targeting the cancer cells expressing MPHOSPH1 is enhanced. Thus, the methods of the present invention can include the step of administering the peptides, the polynucleotides, the APCs or exosomes of the present invention to a subject.

[0137] Alternatively, CTLs can be also induced by using them ex vivo or in vitro, and after inducing CTL, the activated CTLs can be returned to the subject. For example, the method can include steps of:

a: collecting APCs from a subject;

b: contacting the APCs of step a, with the peptide; and

c: co-culturing the APCs of step b with CD8 positive T cells.

[0138] The APCs to be co-cultured with the CD8 positive T cells in above step c can also be prepared by transferring a polynucleotide of the present invention into APCs as described above in section "VI. Antigen-Presenting Cells", though the present invention is not limited thereto, and thus encompasses any APCs that effectively present on their surface a complex of an HLA antigen and a peptide of the present invention.

[0139] One may optionally utilize exosomes that presents on its surface a complex of an HLA antigen and the peptide of the present invention instead of the afore-mentioned APCs. Namely, the present invention can include the step of co-culturing exosomes presenting on their surface a complex of an HLA antigen and the peptide of the present invention. Such exosomes can be prepared by the methods described above in section "V. Exosomes".

APCs or exosomes used for induction of CTLs can be preferably APCs or exosomes that present on their surface a complex of the peptide of the present invention and HLA-A2 antigen.

[0140] Furthermore, a CTL can be induced by introducing a polynucleotide encoding both of the TCR subunits or polynucleotides encoding each of the TCR subunits into a CD8 positive T cell, wherein the TCR can bind to a complex of the peptide of the present invention and HLA antigen presented on a cell surface. Such transduction can be performed as described above in section "VIII. T Cell Receptor (TCR)".

[0141] The methods of the present invention can be carried out in vitro, ex vivo or in vivo. Preferably, the methods of the present invention can be carried out in vitro or ex vivo. CD8 positive T cells used for induction of CTLs can be prepared by well-known methods in the art from PBMCs obtained from a subject. In preferred embodiments, a donor for CD8 positive T cells can be a subject whose HLA antigen is HLA-A2. The CTLs induced by the methods of the present invention can be CTLs that can recognize cells presenting a complex of the peptide of the present invention and an HLA antigen on its surface. When CTLs induced by the method of the present invention are administered to a subject in order to induce immune responses against cancer in the subject, the subject is preferably the same one from whom CD8 positive T cells are derived. However, the subject may be a different one from the CD8 positive T cell donor so long as the subject has the same HLA type with the CD8 positive T cell donor.

[0142] In addition, the present invention provides a method or process for manufacturing a pharmaceutical agent or composition for inducing CTLs, wherein the method or process includes the step of admixing or formulating the peptide of the present invention with a pharmaceutically acceptable carrier.

In another embodiment, the present invention provides an agent or composition for inducing CTL, wherein the agent or composition comprises one or more peptide(s), one or more polynucleotide(s), or one or more APCs or exosomes of the present invention.

In another embodiment, the present invention provides the use of the peptide, polynucleotide, or APC or exosome of the present invention in the manufacture of an agent or composition formulated for inducing a CTL.

Alternatively, the present invention further provides the peptide, polynucleotide, or APC or exosome of the present invention for use in inducing a CTL.

[0143] (3) Methods of inducing immune response:

Moreover, the present invention provides methods of inducing an immune response against diseases related to MPHOSPH1. Diseases contemplated include cancer, examples of which include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor.

[0144] The methods of the present invention may include the step of administering to a subject agent(s) or composition(s) containing any of the peptides of the present invention or polynucleotides encoding them. The method of the present invention may also contemplate the administration of exosomes or APCs presenting any of the peptides of the present invention. For details, see the item of "IX. Pharmaceutical Compositions", particularly the part describing the use of the pharmaceutical agents and compositions of the present invention as vaccines. In addition, the exosomes and APCs that can be employed for the present methods for inducing immune response are described in detail under the items of "V. Exosomes", "VI. Antigen-Presenting Cells (APCs)", and (1) and (2) of "X. Methods using Peptides, Exosomes, APCs and CTLs", supra.

In preferred embodiments, the subjects treated by the method of the present invention can be subjects whose HLA antigen is HLA-A2.

[0145] The present invention also provides a method or process for manufacturing a pharmaceutical agent or composition inducing immune response, wherein the method may include the step of admixing or formulating a polypeptide or polynucleotide of the present invention with a pharmaceutically acceptable carrier.

Alternatively, the method of the present invention may include the step of administrating a vaccine or a pharmaceutical composition of the present invention that contains:

(a) a peptide of the present invention;

(b) a nucleic acid (polynucleotide) encoding such a peptide as disclosed herein in an expressible form;

(c) an APC or an exosome presenting a peptide of the present invention on its surface; or

(d) a CTL of the present invention.

[0146] In the context of the present invention, a cancer expressing MPHOSPH1 can be treated with these active ingredients. Examples of such cancer include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate

cancer, renal cancer and soft tissue tumor. Accordingly, prior to the administration of the vaccines or pharmaceutical compositions including the active ingredients, it is preferable to confirm whether the expression level of MPHOSPH1 in the cells or tissues to be treated is enhanced compared with normal cells of the same organ. Thus, in one embodiment, the present invention provides a method for treating cancer expressing MPHOSPH1, which method may include the steps of:

i) determining the expression level of MPHOSPH1 in cells or tissue(s) obtained from a subject with the cancer to be treated;

ii) comparing the expression level of MPHOSPH1 with normal control; and

iii) administrating at least one component selected from among steps (a) to (d) described above to a subject with cancer over-expressing MPHOSPH1 compared with normal control.

[0147] Alternatively, the present invention also provides a vaccine or pharmaceutical composition that includes at least one component selected from among (a) to (d) described above, for use in administrating to a subject having cancer over-expressing MPHOSPH1. In other words, the present invention further provides a method for identifying a subject to be treated with the MPHOSPH1 polypeptide of the present invention, such method including the step of determining an expression level of MPHOSPH1 in subject-derived cells or tissue(s), wherein an increase of the level compared to a normal control level of the gene indicates that the subject may have cancer which may be treated with the MPHOSPH1 polypeptide of the present invention. The method of identifying a subject to be treated cancer of the present invention are described in more detail below.

[0148] Any subject-derived cell or tissue can be used for the determination of MPHOSPH1 expression so long as it includes the objective transcription or translation product of MPHOSPH1. Examples of suitable samples include, but are not limited to, bodily tissues and fluids, such as blood, sputum and urine. Preferably, the subject-derived cell or tissue sample contains a cell population including an epithelial cell, more preferably a cancerous epithelial cell or an epithelial cell derived from tissue suspected to be cancerous. Further, if necessary, the cell may be purified from the obtained bodily tissues and fluids, and then used as the subjected-derived sample.

A subject to be treated by the present method is preferably a mammal. Illustrative mammals include, but are not limited to, e.g., human, non-human primate, mouse, rat, dog, cat, horse, and cow.

[0149] According to the present invention, the expression level of MPHOSPH1 in cells or tissues obtained from a subject may be determined. The expression level can be determined at the transcription (nucleic acid) product level, using methods known in the art. For example, the mRNA of MPHOSPH1 may be quantified using probes by hy-

bridization methods (e.g., Northern hybridization). The detection may be carried out on a chip, an array or as such. The use of an array may be preferable for detecting the expression level of MPHOSPH1. Those skilled in the art can prepare such probes utilizing the sequence information of MPHOSPH1. For example, the cDNA of MPHOSPH1 may be used as the probes. If necessary, the probes may be labeled with a suitable label, such as dyes, fluorescent substances and isotopes, and the expression level of the gene may be detected as the intensity of the hybridized labels.

[0150] Furthermore, the transcription product of MPHOSPH1 (e.g., SEQ ID NO: 125) may be quantified using primers by amplification-based detection methods (e.g., RT-PCR). Such primers may be prepared based on the available sequence information of the gene.

Specifically, a probe or primer used for the present method hybridizes under stringent, moderately stringent, or low stringent conditions to the mRNA of MPHOSPH1. As used herein, the phrase "stringent (hybridization) conditions" refers to conditions under which a probe or primer will hybridize to its target sequence, but not to other sequences. Stringent conditions are sequence-dependent and will be different under different circumstances. Specific hybridization of longer sequences is observed at higher temperatures than shorter sequences. Generally, the temperature of a stringent condition is selected to be about 5 degrees C lower than the thermal melting point (Tm) for a specific sequence at a defined ionic strength and pH. The Tm is the temperature (under a defined ionic strength, pH and nucleic acid concentration) at which 50% of the probes complementary to their target sequence hybridize to the target sequence at equilibrium. Since the target sequences are generally present at excess, at Tm, 50% of the probes are occupied at equilibrium. Typically, stringent conditions will be those in which the salt concentration is less than about 1.0 M sodium ion, typically about 0.01 to 1.0 M sodium ion (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30 degrees C for short probes or primers (e.g., 10 to 50 nucleotides) and at least about 60 degrees C for longer probes or primers. Stringent conditions may also be achieved with the addition of destabilizing substances, such as formamide.

[0151] A probe or primer of the present invention is typically a substantially purified oligonucleotide. The oligonucleotide typically includes a region of nucleotide sequence that hybridizes under stringent conditions to at least about 2000, 1000, 500, 400, 350, 300, 250, 200, 150, 100, 50, or 25, consecutive sense strand nucleotide sequence of a nucleic acid including a MPHOSPH1, or an anti sense strand nucleotide sequence of a nucleic acid including a MPHOSPH1, or of a naturally occurring mutant of these sequences. In particular, for example, in a preferred embodiment, an oligonucleotide having 5-50 in length can be used as a primer for amplifying the genes, to be detected.

More preferably, mRNA or cDNA of a MPHOSPH1 gene can be detected with oligonucleotide probe or primer of a specific size, generally 15- 30b in length. The size may range from at least 10 nucleotides, at least 12 nucleotides, at least 15 nucleotides, at least 20 nucleotides, at least 25 nucleotides, at least 30 nucleotides and the probes and primers may range in size from 5-10 nucleotides, 10-15 nucleotides, 15-20 nucleotides, 20-25 nucleotides and 25-30 nucleotides. In preferred embodiments, length of the oligonucleotide probe or primer can be selected from 15-25. Assay procedures, devices, or reagents for the detection of gene by using such oligonucleotide probe or primer are well known (e.g. oligonucleotide microarray or PCR). In these assays, probes or primers can also include tag or linker sequences. Further, probes or primers can be modified with detectable label or affinity ligand to be captured. Alternatively, in hybridization based detection procedures, a polynucleotide having a few hundreds (e.g., about 100-200) bases to a few kilo (e.g., about 1000-2000) bases in length can also be used for a probe (e.g., northern blotting assay or cDNA microarray analysis).

[0152] Alternatively, the translation product may be detected for the diagnosis of the present invention. For example, the quantity of MPHOSPH1 protein (SEQ ID NO: 126) or the immunologically fragment thereof may be determined. Methods for determining the quantity of the protein as the translation product include immunoassay methods that use an antibody specifically recognizing the protein. The antibody may be monoclonal or polyclonal. Furthermore, any fragment or modification (e.g., chimeric antibody, scFv, Fab, $F(ab')_2$, Fv, etc.) of the antibody may be used for the detection, so long as the fragment or modified antibody retains the binding ability to the MPHOSPH1 protein. Such antibodies against the peptides of the present invention and the fragments thereof are also provided by the present invention. Methods to prepare these kinds of antibodies for the detection of proteins are well known in the art, and any method may be employed in the present invention to prepare such antibodies and equivalents thereof.

[0153] As another method to detect the expression level of MPHOSPH1 gene based on its translation product, the intensity of staining may be measured via immunohistochemical analysis using an antibody against the MPHOSPH1 protein. Namely, in this measurement, strong staining indicates increased presence/level of the protein and, at the same time, high expression level of MPHOSPH1 gene.

The expression level of a target gene, e.g., the MPHOSPH1 gene, in cancer cells can be determined to be increased if the level increases from the control level (e.g., the level in normal cells) of the target gene by, for example, 10%, 25%, or 50%; or increases to more than 1.1 fold, more than 1.5 fold, more than 2.0 fold, more than 5.0 fold, more than 10.0 fold, or more.

[0154] The control level may be determined at the same time as the cancer cells using a

sample(s) previously collected and stored from a subject(s) whose disease state(s) (cancerous or non-cancerous) is/are known. In addition, normal cells obtained from non-cancerous regions of an organ that has the cancer to be treated may be used as normal control. Alternatively, the control level may be determined by a statistical method based on the results obtained by analyzing previously determined expression level(s) of MPHOSPH1 gene in samples from subjects whose disease states are known. Furthermore, the control level can be derived from a database of expression patterns from previously tested cells. Moreover, according to an aspect of the present invention, the expression level of MPHOSPH1 gene in a biological sample may be compared to multiple control levels, determined from multiple reference samples. It is preferred to use a control level determined from a reference sample derived from a tissue type similar to that of the subject-derived biological sample. Moreover, it is preferred to use the standard value of the expression levels of MPHOSPH1 gene in a population with a known disease state. The standard value may be obtained by any method known in the art. For example, a range of mean +/- 2 S.D. or mean +/- 3 S.D. may be used as the standard value.

[0155] In the context of the present invention, a control level determined from a biological sample that is known to be non-cancerous is referred to as a "normal control level". On the other hand, if the control level is determined from a cancerous biological sample, it is referred to as a "cancerous control level". Difference between a sample expression level and a control level can be normalized to the expression level of control nucleic acids, e.g., housekeeping genes, whose expression levels are known not to differ depending on the cancerous or non-cancerous state of the cell. Exemplary control genes include, but are not limited to, beta-actin, glyceraldehyde 3 phosphate dehydrogenase, and ribosomal protein P1.

When the expression level of MPHOSPH1 gene is increased as compared to the normal control level, or is similar/equivalent to the cancerous control level, the subject may be diagnosed with cancer to be treated.

[0156] The present invention also provides a method of (i) diagnosing whether a subject suspected to have cancer to be treated, and/or (ii) selecting a subject for cancer treatment, which method may include the steps of:

a) determining the expression level of MPHOSPH1 in cells or tissue(s) obtained from a subject who is suspected to have the cancer to be treated;

b) comparing the expression level of MPHOSPH1 with a normal control level;

c) diagnosing the subject as having the cancer to be treated, if the expression level of MPHOSPH1 is increased as compared to the normal control level; and

d) selecting the subject for cancer treatment, if the subject is diagnosed as having the cancer to be treated, in step c).

[0157] Alternatively, such a method may include the steps of:

a) determining the expression level of MPHOSPH1 in cells or tissue(s) obtained from a subject who is suspected to have the cancer to be treated;

b) comparing the expression level of MPHOSPH1 with a cancerous control level;

c) diagnosing the subject as having the cancer to be treated, if the expression level of MPHOSPH1 is similar or equivalent to the cancerous control level; and

d) selecting the subject for cancer treatment, if the subject is diagnosed as having the cancer to be treated, in step c).

[0158] The present invention also provides a diagnostic kit for diagnosing or determining a subject who is or is suspected to be suffering from or at risk for developing cancer that can be treated with the MPHOSPH1 polypeptide of the present invention, which may also find use in assessing and/or monitoring the efficacy or applicability of a cancer immunotherapy. Preferably, the cancer includes, but is not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor. More particularly, the kit preferably may include at least one reagent for detecting the expression of the MPHOSPH1 gene in a subject-derived cell, which reagent may be selected from the group of:

(a) a reagent for detecting an mRNA of the MPHOSPH1 gene;

(b) a reagent for detecting the MPHOSPH1 protein or the immunologically fragment thereof; and

(c) a reagent for detecting the biological activity of the MPHOSPH1 protein.

[0159] Examples of reagents suitable for the detection of mRNA of the MPHOSPH1 gene may include nucleic acids that specifically bind to or identify the MPHOSPH1 mRNA, such as oligonucleotides that have a complementary sequence to a portion of the MPHOSPH1 mRNA. These kinds of oligonucleotides are exemplified by primers and probes that are specific to the MPHOSPH1 mRNA. These kinds of oligonucleotides may be prepared based on methods well known in the art. If needed, the reagent for detecting the MPHOSPH1 mRNA may be immobilized on a solid matrix. Moreover, more than one reagent for detecting the MPHOSPH1 mRNA may be included in the kit.

[0160] On the other hand, examples of reagents suitable for the detection of the MPHOSPH1 protein or the immunologically fragment thereof may include antibodies to the MPHOSPH1 protein or the immunologically fragment thereof. The antibody may be monoclonal or polyclonal. Furthermore, any fragment or modification (e.g., chimeric antibody, scFv, Fab, $F(ab')_2$, Fv, etc.) of the antibody may be used as the reagent, so long as the fragment or modified antibody retains the binding ability to the MPHOSPH1 protein or the immunologically fragment thereof. Methods to prepare

these kinds of antibodies for the detection of proteins are well known in the art, and any method may be employed in the present invention to prepare such antibodies and equivalents thereof. Furthermore, the antibody may be labeled with signal generating molecules via direct linkage or an indirect labeling technique. Labels and methods for labeling antibodies and detecting the binding of the antibodies to their targets are well known in the art, and any labels and methods may be employed for the present invention. Moreover, more than one reagent for detecting the MPHOSPH1 protein may be included in the kit.

[0161] The kit may contain more than one of the aforementioned reagents. The kit can further include a solid matrix and reagent for binding a probe against a MPHOSPH1 gene or antibody against a MPHOSPH1 peptide, a medium and container for culturing cells, positive and negative control reagents, and a secondary antibody for detecting an antibody against a MPHOSPH1 peptide. For example, tissue samples obtained from subjects without cancer or suffering from cancer, may serve as useful control reagents. A kit of the present invention may further include other materials desirable from a commercial and user standpoint, including buffers, diluents, filters, needles, syringes, and package inserts (e.g., written, tape, CD-ROM, etc.) with instructions for use. These reagents and such may be retained in a container with a label. Suitable containers may include bottles, vials, and test tubes. The containers may be formed from a variety of materials, such as glass or plastic.

[0162] In an embodiment of the present invention, when the reagent is a probe against the MPHOSPH1 mRNA, the reagent may be immobilized on a solid matrix, such as a porous strip, to form at least one detection site. The measurement or detection region of the porous strip may include a plurality of sites, each containing a nucleic acid (probe). A test strip may also contain sites for negative and/or positive controls. Alternatively, control sites may be located on a strip separated from the test strip. Optionally, the different detection sites may contain different amounts of immobilized nucleic acids, i.e., a higher amount in the first detection site and lesser amounts in subsequent sites. Upon the addition of a test sample, the number of sites displaying a detectable signal provides a quantitative indication of the amount of MPHOSPH1 mRNA present in the sample. The detection sites may be configured in any suitably detectable shape and are typically in the shape of a bar or dot spanning the width of a test strip.

[0163] The kit of the present invention may further include a positive control sample or MPHOSPH1 standard sample. The positive control sample of the present invention may be prepared by collecting MPHOSPH1 positive samples and then assaying their MPHOSPH1 levels. Alternatively, a purified MPHOSPH1 protein or polynucleotide may be added to cells that do not express MPHOSPH1 to form the positive sample or

the MPHOSPH1 standard sample. In the context of the present invention, purified MPHOSPH1 may be a recombinant protein. The MPHOSPH1 level of the positive control sample is, for example, more than the cut off value.

[0164] In one embodiment, the present invention further provides a diagnostic kit including, a protein or a partial protein thereof specifically recognized by the antibody of the present invention or the fragment thereof.

[0165] Examples of partial peptides of the present invention include polypeptides composed of at least 8, preferably 15, and more preferably 20 contiguous amino acids in the amino acid sequence of a protein of the present invention. Cancer can be diagnosed by detecting an antibody in a sample (e.g., blood, tissue) using a protein or a peptide (polypeptide) of the present invention. The method for preparing the protein of the present invention and peptides are as described above.

[0166] The methods for diagnosing cancer of the present invention can be performed by determining the difference between the amount of anti-MPHOSPH1 antibody and that in the corresponding control sample as describe above. The subject is suspected to be suffering from cancer, if cells or tissues of the subject contain antibodies against the expression products (MPHOSPH1) of the gene and the quantity of the anti-MPHOSPH1 antibody is determined to be more than the cut off value in level compared to that in normal control.

[0167] In another embodiment, a diagnostic kit of the present invention may include the peptide of the present invention and an HLA molecule binding thereto. The method for detecting antigen specific CTLs using antigenic peptides and HLA molecules has already been established (for example, Altman JD et al., Science. 1996, 274(5284): 94-6). Thus, the complex of the peptide of the present invention and the HLA molecule can be applied to the detection method to detect tumor antigen specific CTLs, thereby enabling earlier detection, recurrence and/or metastasis of cancer. Further, it can be employed for the selection of subjects applicable with the pharmaceuticals including the peptide of the present invention as an active ingredient, or the assessment of the treatment effect of the pharmaceuticals.

[0168] Particularly, according to the known method (see, for example, Altman JD et al., Science. 1996, 274(5284): 94-6), the oligomer complex, such as tetramer, of the radiolabeled HLA molecule and the peptide of the present invention can be prepared. With using the complex, the diagnosis can be done, for example, by quantifying the antigen-peptide specific CTLs in the peripheral blood lymphocytes derived from the subject suspected to be suffering from cancer.

[0169] The present invention further provides methods and diagnostic agents for evaluating immunological response of subject by using peptide epitopes as described herein. In one embodiment of the invention, HLA-A2 restricted peptides as described herein may

be used as reagents for evaluating or predicting an immune response of a subject. The immune response to be evaluated may be induced by contacting an immunogen with immunocompetent cells in vitro or in vivo. In certain embodiments, any substances or compositions that may result in the production of antigen specific CTLs that recognize and bind to the peptide epitope(s) may be employed as the reagent. The peptide reagents may need not to be used as the immunogen. Assay systems that are used for such an analysis include relatively recent technical developments such as tetramers, staining for intracellular lymphokines and interferon release assays, or ELISPOT assays. In preferred embodiments, the immunocompetent cells for evaluating an immunological response, may be selected from among peripheral blood, peripheral blood lymphocyte (PBL), and peripheral blood mononuclear cell (PBMC). Methods for collecting or isolating such immunocompetent cells are well known in the arts. In an alternate preferred embodiment, the immunocompetent cells to be contacted with peptide reagent include antigen presenting cells such as dendritic cells.

[0170] For example, peptides of the present invention may be used in tetramer staining assays to assess peripheral blood mononuclear cells for the presence of antigen-specific CTLs following exposure to a tumor cell antigen or an immunogen. The HLA tetrameric complex may be used to directly visualize antigen specific CTLs (see, e.g., Ogg et al., Science 279: 2103-2106, 1998; and Altman et al, Science 174 : 94-96, 1996) and determine the frequency of the antigen-specific CTL population in a sample of peripheral blood mononuclear cells. A tetramer reagent using a peptide of the invention may be generated as described below.

[0171] A peptide that binds to an HLA molecule is refolded in the presence of the corresponding HLA heavy chain and beta 2-microglobulin to generate a trimolecular complex. In the complex, carboxyl terminal of the heavy chain is biotinylated at a site that was previously engineered into the protein. Then, streptavidin is added to the complex to form tetramer composed of the trimolecular complex and streptavidin. By means of fluorescently labeled streptavidin, the tetramer can be used to stain antigen specific cells. The cells can then be identified, for example, by flow cytometry. Such an analysis may be used for diagnostic or prognostic purposes. Cells identified by the procedure can also be used for therapeutic purposes.

[0172] The present invention also provides reagents to evaluate immune recall responses (see, e.g., Bertoni et al, J. Clin. Invest. 100: 503-513, 1997 and Penna et al., J Exp. Med. 174: 1565-1570, 1991) including peptides of the present invention. For example, patient PBMC samples obtained from individuals with a cancer to be treated can be analyzed for the presence of antigen-specific CTLs using specific peptides. A blood sample containing mononuclear cells can be evaluated by cultivating the PBMCs and stimulating the cells with a peptide of the invention. After an appropriate cultivation

period, the expanded cell population can be analyzed, for example, for CTL activity.

[0173] The peptides may also be used as reagents to evaluate the efficacy of a vaccine. PBMCs obtained from a patient vaccinated with an immunogen may be analyzed using, for example, either of the methods described above. The patient is HLA typed, and peptide epitope reagents that recognize the allele specific molecules present in the patient are selected for the analysis. The immunogenicity of the vaccine may be indicated by the presence of epitope-specific CTLs in the PBMC sample. The peptides of the invention may also be used to make antibodies, using techniques well known in the art (see, e.g., CURRENT PROTOCOLS IN IMMUNOLOGY, Wiley/Greene, NY; and Antibodies A Laboratory Manual, Harlow and Lane, Cold Spring Harbor Laboratory Press, 1989), which may find use as reagents to diagnose, detect or monitor cancer. Such antibodies may include those that recognize a peptide in the context of an HLA molecule, i.e., antibodies that bind to a peptide-MHC complex.

[0174] The peptides and compositions of the present invention have a number of additional uses, some of which are described herein. For instance, the present invention provides a method for diagnosing or detecting a disorder characterized by quantity of a MPHOSPH1 immunogenic polypeptide. These methods involve determining quantity of a MPHOSPH1 peptide, or a complex of a MPHOSPH1 peptide and an HLA class I molecule in a biological sample. The expression of a peptide or complex of peptide and HLA class I molecule can be determined or detected by assaying with a binding partner for the peptide or complex. In an preferred embodiment, a binding partner for the peptide or complex may be an antibody recognizes and specifically bind to the peptide. The expression of MPHOSPH1 in a biological sample, such as a tumor biopsy, can also be tested by standard PCR amplification protocols using MPHOSPH1 primers. An example of tumor expression is presented herein and further disclosure of exemplary conditions and primers for MPHOSPH1 amplification can be found in WO2003/27322, the contents of which are incorporated by reference herein.

[0175] Preferably, the diagnostic methods involve contacting a biological sample isolated from a subject with an agent specific for the MPHOSPH1 peptide to detect the presence of the MPHOSPH1 peptide in the biological sample. As used herein, "contacting" means placing the biological sample in sufficient proximity to the agent and under the appropriate conditions of, e.g., concentration, temperature, time, ionic strength, to allow the specific interaction between the agent and MPHOSPH1 peptide that are present in the biological sample. In general, the conditions for contacting the agent with the biological sample are conditions known by those of ordinary skill in the art to facilitate a specific interaction between a molecule and its cognate (e.g., a protein and its receptor cognate, an antibody and its protein antigen cognate, a nucleic acid and its complementary sequence cognate) in a biological sample. Optimal conditions for

facilitating a specific interaction between a molecule and its cognate are described in U. S. Patent No. 5,108,921, issued to Low et al., the contents of which are incorporated by reference herein.

[0176] The diagnostic method of the present invention can be performed in either or both of in vivo and in vitro. Accordingly, biological sample can be located in vivo or in vitro in the present invention. For example, the biological sample can be a tissue in vivo and the agent specific for the MPHOSPH1 immunogenic polypeptide can be used to detect the presence of such molecules in the tissue. Alternatively, the biological sample can be collected or isolated in vitro (e.g., a blood sample, tumor biopsy, tissue extract). In a particularly preferred embodiment, the biological sample can be a cell-containing sample, more preferably a sample containing tumor cells collected from a subject to be diagnosed or treated.

[0177] Alternatively, the diagnosis can be done, by a method which allows direct quantification of antigen-specific T cells by staining with Fluorescein-labeled HLA multimeric complexes (e.g., Altman, J. D. et al., 1996, Science 274 : 94; Altman, J. D. et al., 1993, Proc. Natl. Acad. Sci. USA 90 : 10330). Staining for intracellular lymphokines, and interferon-gamma release assays or ELISPOT assays also has been provided. Multimer staining, intracellular lymphokine staining and ELISPOT assays all appear to be at least 10-fold more sensitive than more conventional assays (Murali-Krishna, K. et al., 1998, Immunity 8: 177; Lalvani, A. et al., 1997, J. Exp. Med. 186: 859; Dunbar, P. R. et al., 1998, Curr. Biol. 8: 413). Pentamers (e.g., US 2004-209295A), dextramers (e.g., WO 02/072631), and streptamers (e.g., Nature medicine 6. 631-637 (2002)) may also be used.

[0178] Accordingly, in some embodiments, the present invention provides a method for diagnosing or evaluating an immunological response of a subject administered at least one of the MPHOSPH1 peptides of the present invention, the method including the steps of:

(a) contacting an immunogen with immunocompetent cells under the condition suitable for induction of CTL specific to the immunogen;

(b) detecting or determining induction level of the CTL induced in step (a); and

(c) correlating the immunological response of the subject with the CTL induction level.

[0179] In the context of the present invention, the immunogen preferably includes at least one of (a) a MPHOSPH1 peptide selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 and (b) peptides having such amino acid sequences in which such amino acid sequences have been modified with 1, 2 or more amino acid substitution(s). In the meantime, conditions suitable of induction of immunogen specific CTL are well known in the art. For example, immunocompetent cells may be cultured in vitro under

the presence of immunogen(s) to induce immunogen specific CTL. In order to induce immunogen specific CTLs, any stimulating factors may be added to the cell culture. For example, IL-2 is preferable stimulating factors for the CTL induction.

[0180] In some embodiments, the step of monitoring or evaluating immunological response of a subject to be treated with peptide cancer therapy may be performed before, during and/or after the treatment. In general, during a protocol of cancer therapy, immunogenic peptides are administered repeatedly to a subject to be treated. For example, immunogenic peptides may be administered every week for 3-10 weeks. Accordingly, the immunological response of the subject can be evaluated or monitored during the cancer therapy protocol. Alternatively, the step of evaluation or monitoring of immunological response to the cancer therapy may at the completion of the therapy protocol.

[0181] According to the present invention, enhanced induction of immunogen specific CTL as compared with a control indicates that the subject to be evaluated or diagnosed immunologically responded to the immunogen(s) that has/ have been administered. Suitable controls for evaluating the immunological response may include, for example, a CTL induction level when the immunocompetent cells are contacted with no peptide, or control peptide(s) having amino acid sequences other than any MPHOSPH1 peptides. (e.g. random amino acid sequence). In a preferred embodiment, the immunological response of the subject is evaluated in a sequence specific manner, by comparison with an immunological response between each immunogen administered to the subject. In particular, even when a mixture of some kinds of MPHOSPH1 peptides is administered to the subject, immunological response might vary depending on the peptides. In that case, by comparison of the immunological response between each peptide, peptides to which the subject show higher response can be identified.

[0182] XI. Antibodies:

The present invention further provides antibodies that bind to peptides of the present invention. Preferred antibodies specifically bind to a peptide of the present invention and will not bind (or will bind weakly) to other peptide. Alternatively, antibodies may bind to peptides of the invention as well as the homologs thereof. Antibodies against peptides of the invention can find use in cancer diagnostic and prognostic assays, as well as imaging methodologies. Similarly, such antibodies can find use in the treatment, diagnosis, and/or prognosis of other cancers, to the extent MPHOSPH1 is also expressed or over-expressed in a cancer patient. Moreover, intracellularly expressed antibodies (e.g., single chain antibodies) may therapeutically find use in treating cancers in which the expression of MPHOSPH1 is involved, examples of which include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC,

lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor.

[0183] The present invention also provides various immunological assays for the detection and/or quantification of an MPHOSPH1 protein (SEQ ID NO: 126) or a fragment thereof, including a polypeptide composed of amino acid sequences selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120. Such assays may include one or more anti-MPHOSPH1 antibodies capable of recognizing and binding a MPHOSPH1 protein or fragments thereof, as appropriate. In the context of the present invention, anti-MPHOSPH1 antibodies binding to an MPHOSPH1 polypeptide will preferably recognize a polypeptide composed of amino acid sequences selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 to the exclusion of other peptides. The binding specificity of antibody can be confirmed by means of an inhibition test. That is, when the binding between an antibody to be analyzed and full-length of MPHOSPH1 polypeptide is inhibited under presence of any fragment polypeptides having an amino acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120, the antibody is deemed to specifically bind to the fragment. In the context of the present invention, such immunological assays are performed within various immunological assay formats well known in the art, including but not limited to, various types of radio-immunoassays, immunochromatograph technique, enzyme-linked immunosorbent assays (ELISA), enzyme-linked immunofluorescent assays (ELIFA), and the like.

[0184] Related immunological but non-antibody assays of the invention may also include T cell immunogenicity assays (inhibitory or stimulatory) as well as MHC binding assays. In addition, immunological imaging methods capable of detecting cancers expressing MPHOSPH1 are also provided by the invention, including, but not limited to, radioscintigraphic imaging methods using labeled antibodies of the present invention. Such assays can clinically find use in the detection, monitoring, and prognosis of MPHOSPH1 expressing cancers, examples of which include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor.

[0185] The present invention also provides antibodies that bind to a peptide of the present invention. An antibody of the present invention can be used in any form, such as monoclonal or polyclonal antibodies, and include antiserum obtained by immunizing an animal such as a rabbit with the peptide of the invention, all classes of polyclonal and monoclonal antibodies, human antibodies and humanized antibodies produced by genetic recombination.

[0186] A peptide of the present invention used as an antigen to obtain an antibody may be derived from any animal species, but preferably is derived from a mammal such as a

human, mouse, or rat, more preferably from a human. A human-derived peptide may be obtained from the nucleotide or amino acid sequences disclosed herein.

[0187] According to the present invention, complete and partial peptides of a protein may serve as an immunization antigen. Examples of suitable partial peptides include, for example, the amino (N)-terminal or carboxy (C)-terminal fragment of a peptide of the present invention.

Herein, an antibody is defined as a protein that reacts with either the full length or a fragment of a MPHOSPH1 peptide. In a preferred embodiment, an antibody of the present invention will recognize fragment peptides of MPHOSPH1 that have an amino acid sequence selected from among SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120. Methods for synthesizing oligopeptide are well known in the arts. After the synthesis, peptides may be optionally purified prior to use as immunogen. In the present invention, the oligopeptide (e.g., 9- or 10mer) may be conjugated or linked with carriers to enhance the immunogenicity. Keyhole-limpet hemocyanin (KLH) is well known as the carrier. Method for conjugating KLH and peptide are also well known in the arts.

[0188] Alternatively, a gene encoding a peptide of the present invention or fragment thereof may be inserted into a known expression vector, which is then used to transform a host cell as described herein. The desired peptide or fragment thereof may be recovered from the outside or inside of host cells by any standard method, and may subsequently be used as an antigen. Alternatively, whole cells expressing the peptide or their lysates or a chemically synthesized peptide may be used as the antigen.

[0189] Any mammalian animal may be immunized with the antigen, though preferably the compatibility with parental cells used for cell fusion is taken into account. In general, animals of Rodentia, Lagomorpha or Primate family may be used. Animals of the family Rodentia include, for example, mouse, rat and hamster. Animals of the family Lagomorpha include, for example, rabbit. Animals of the Primate family include, for example, a monkey of Catarrhini (old world monkey) such as Macaca fascicularis, rhesus monkey, sacred baboon and chimpanzees.

[0190] Methods for immunizing animals with antigens are known in the art. Intraperitoneal injection or subcutaneous injection of antigens is a standard method for immunization of mammals. More specifically, antigens may be diluted and suspended in an appropriate amount of phosphate buffered saline (PBS), physiological saline, etc. If desired, the antigen suspension may be mixed with an appropriate amount of a standard adjuvant, such as Freund's complete adjuvant, made into emulsion and then administered to mammalian animals. Preferably, it is followed by several administrations of antigen mixed with an appropriately amount of Freund's incomplete adjuvant every 4 to 21 days. An appropriate carrier may also be used for immunization.

After immunization as above, serum may be examined by a standard method for an increase in the amount of desired antibodies.

[0191] Polyclonal antibodies against the peptides of the present invention may be prepared by collecting blood from the immunized mammal examined for the increase of desired antibodies in the serum, and by separating serum from the blood by any conventional method. Polyclonal antibodies include serum containing the polyclonal antibodies, as well as the fraction containing the polyclonal antibodies may be isolated from the serum. Immunoglobulin G or M can be prepared from a fraction which recognizes only the peptide of the present invention using, for example, an affinity column coupled with the peptide of the present invention, and further purifying this fraction using protein A or protein G column.

[0192] To prepare monoclonal antibodies for use in the context of the present invention, immune cells are collected from the mammal immunized with the antigen and checked for the increased level of desired antibodies in the serum as described above, and are subjected to cell fusion. The immune cells used for cell fusion may preferably be obtained from spleen. Other preferred parental cells to be fused with the above immunocyte include, for example, myeloma cells of mammalians, and more preferably myeloma cells having an acquired property for the selection of fused cells by drugs.

The above immunocyte and myeloma cells can be fused according to known methods, for example, the method of Milstein et al. (Galfre and Milstein, Methods Enzymol 73: 3-46 (1981)).

[0193] Resulting hybridomas obtained by cell fusion may be selected by cultivating them in a standard selection medium, such as HAT medium (hypoxanthine, aminopterin and thymidine containing medium). The cell culture is typically continued in the HAT medium for several days to several weeks, the time being sufficient to allow all the other cells, with the exception of the desired hybridoma (non-fused cells), to die. Then, the standard limiting dilution may be performed to screen and clone a hybridoma cell producing the desired antibody.

[0194] In addition to the above method, wherein a non-human animal is immunized with an antigen for preparing hybridoma, human lymphocytes such as those infected by EB virus may be immunized with a peptide, peptide expressing cells or their lysates in vitro. Then, the immunized lymphocytes may be fused with human-derived myeloma cells that are capable of indefinitely dividing, such as U266, to yield a hybridoma producing a desired human antibody that is able to bind to the peptide can be obtained (Unexamined Published Japanese Patent Application No. Sho 63-17688).

[0195] The obtained hybridomas may then subsequently be transplanted into the abdominal cavity of a mouse and the ascites extracted. The obtained monoclonal antibodies can be purified by, for example, ammonium sulfate precipitation, a protein A or protein G

column, DEAE ion exchange chromatography or an affinity column to which the peptide of the present invention is coupled. An antibody of the present invention can be used not only for purification and detection of a peptide of the present invention, but also as a candidate for agonists and antagonists of a peptide of the present invention.

[0196] Alternatively, an immune cell, such as an immunized lymphocyte, producing antibodies may be immortalized by an oncogene and used for preparing monoclonal antibodies.

Monoclonal antibodies thus obtained can be also recombinantly prepared using genetic engineering techniques (see, for example, Borrebaeck and Larrick, Therapeutic Monoclonal Antibodies, published in the United Kingdom by MacMillan Publishers LTD (1990)). For example, a DNA encoding an antibody may be cloned from an immune cell, such as a hybridoma or an immunized lymphocyte producing the antibody, inserted into an appropriate vector, and introduced into host cells to prepare a recombinant antibody. The present invention also provides recombinant antibodies prepared as described above.

[0197] An antibody of the present invention may be a fragment of an antibody or modified antibody, so long as it binds to one or more of the peptides of the invention. For instance, the antibody fragment may be Fab, $F(ab')_2$, Fv or single chain Fv (scFv), in which Fv fragments from H and L chains are ligated by an appropriate linker (Huston et al., Proc Natl Acad Sci USA 85: 5879-83 (1988)). More specifically, an antibody fragment may be generated by treating an antibody with an enzyme, such as papain or pepsin. Alternatively, a gene encoding the antibody fragment may be constructed, inserted into an expression vector and expressed in an appropriate host cell (see, for example, Co et al., J Immunol 152: 2968-76 (1994); Better and Horwitz, Methods Enzymol 178: 476-96 (1989); Pluckthun and Skerra, Methods Enzymol 178: 497-515 (1989); Lamoyi, Methods Enzymol 121: 652-63 (1986); Rousseaux et al., Methods Enzymol 121: 663-9 (1986); Bird and Walker, Trends Biotechnol 9: 132-7 (1991)).

[0198] An antibody may be modified by conjugation with a variety of molecules, such as polyethylene glycol (PEG). The present invention provides for such modified antibodies. The modified antibody can be obtained by chemically modifying an antibody. These modification methods are conventional in the field.

[0199] Alternatively, an antibody of the present invention may be obtained as a chimeric antibody, between a variable region derived from nonhuman antibody and the constant region derived from human antibody, or as a humanized antibody, including the complementarity determining region (CDR) derived from nonhuman antibody, the frame work region (FR) and the constant region derived from human antibody. Such antibodies can be prepared according to known technology. Humanization can be performed by substituting rodent CDRs or CDR sequences for the corresponding

sequences of a human antibody (see, e.g., Verhoeyen et al., Science 239:1534-1536 (1988)). Accordingly, such humanized antibodies are chimeric antibodies, wherein substantially less than an intact human variable domain has been substituted by the corresponding sequence from a non-human species.

[0200] Fully human antibodies including human variable regions in addition to human framework and constant regions can also be used. Such antibodies can be produced using various techniques known in the art. For example, in vitro methods involve use of recombinant libraries of human antibody fragments displayed on bacteriophage (e.g., Hoogenboom & Winter, J. Mol. Biol. 227:381 (1991). Similarly, human antibodies can be made by introducing of human immunoglobulin loci into transgenic animals, e.g., mice in which the endogenous immunoglobulin genes have been partially or completely inactivated. This approach is described, e.g., in U.S. Patent Nos. 6,150,584, 5,545,807; 5,545,806; 5,569,825; 5,625,126; 5,633,425; 5,661,016.

[0201] Antibodies obtained as above may be purified to homogeneity. For example, the separation and purification of the antibody can be performed according to the separation and purification methods used for general proteins. For example, the antibody may be separated and isolated by the appropriately selected and combined use of column chromatographies, such as affinity chromatography, filter, ultrafiltration, salting-out, dialysis, SDS polyacrylamide gel electrophoresis and isoelectric focusing (Antibodies: A Laboratory Manual. Ed Harlow and David Lane, Cold Spring Harbor Laboratory (1988)), but are not limited thereto. A protein A column and protein G column can be used as the affinity column. Exemplary protein A columns to be used include, for example, Hyper D, POROS and Sepharose F.F. (Pharmacia).

[0202] Examples of suitable chromatography techniques, with the exception of affinity chromatography includes, for example, ion-exchange chromatography, hydrophobic chromatography, gel filtration, reverse phase chromatography, adsorption chromatography and the like (Strategies for Protein Purification and Characterization: A Laboratory Course Manual. Ed Daniel R. Marshak et al., Cold Spring Harbor Laboratory Press (1996)). The chromatographic procedures can be carried out by liquid-phase chromatography, such as HPLC and FPLC.

[0203] For example, measurement of absorbance, enzyme-linked immunosorbent assay (ELISA), enzyme immunoassay (EIA), radioimmunoassay (RIA) and/or immunofluorescence may be used to measure the antigen binding activity of the antibody of the invention. In ELISA, the antibody of the present invention is immobilized on a plate, a peptide of the invention is applied to the plate, and then a sample containing a desired antibody, such as culture supernatant of antibody producing cells or purified antibodies, is applied. Then, a secondary antibody that recognizes the primary antibody and is labeled with an enzyme, such as alkaline phosphatase, is applied, and the plate is

incubated. Next, after washing, an enzyme substrate, such as p-nitrophenyl phosphate, is added to the plate, and the absorbance is measured to evaluate the antigen binding activity of the sample. A fragment of the peptide, such as a C-terminal or N-terminal fragment, may be used as the antigen to evaluate the binding activity of the antibody. BIAcore (Pharmacia) may be used to evaluate the activity of the antibody according to the present invention.

[0204] The above methods allow for the detection or measurement of the peptide of the invention, by exposing the antibody of the invention to a sample assumed to contain the peptide of the invention, and detecting or measuring the immune complex formed by the antibody and the peptide.

Because the method of detection or measurement of the peptide according to the invention can specifically detect or measure a peptide, the method can find use in a variety of experiments in which the peptide is used.

[0205] XII. Vectors and Host Cells:

The present invention also provides vectors encoding a peptide of the present invention and host cells into which a polynucleotide encoding a peptide of the present invention is introduced. A vector of the present invention finds utility as a polynucleotide carrier, especially a DNA, of the present invention in host cell, to express the peptide of the present invention, or to administer the polynucleotide of the present invention for gene therapy.

[0206] When E. coli is selected as the host cell and the vector is amplified and produced in a large amount in E. coli (e.g., JM109, DH5 alpha, HB101 or XL1Blue), the vector should have an "ori" suitable for amplification in E. coli and a marker gene suitable for selected transformed E. coli (e.g., a drug-resistance gene selected by a drug such as ampicillin, tetracycline, kanamycin, chloramphenicol or the like). For example, M13-series vectors, pUC-series vectors, pBR322, pBluescript, pCR-Script, etc., can be used. In addition, pGEM-T, pDIRECT and pT7 can also be used for subcloning and extracting cDNA as well as the vectors described above. When a vector is used to produce the protein of the present invention, an expression vector can find use. For example, an expression vector to be expressed in E. coli should have the above characteristics to be amplified in E. coli. When E. coli, such as JM109, DH5 alpha, HB101 or XL1 Blue, are used as a host cell, the vector should have a promoter, for example, lacZ promoter (Ward et al., Nature 341: 544-6 (1989); FASEB J 6: 2422-7 (1992)), araB promoter (Better et al., Science 240: 1041-3 (1988)), T7 promoter or the like, that can efficiently express the desired gene in E. coli. In that respect, pGEX-5X-1 (Pharmacia), "QIAexpress system" (Qiagen), pEGFP and pET (in this case, the host is preferably BL21 which expresses T7 RNA polymerase), for example, can be used instead of the above vectors. Additionally, the vector may also contain a signal sequence for peptide

secretion. An exemplary signal sequence that directs the peptide to be secreted to the periplasm of the E. coli is the pelB signal sequence (Lei et al., J Bacteriol 169: 4379 (1987)). Means for introducing of the vectors into the target host cells include, for example, the calcium chloride method, and the electroporation method.

[0207] In addition to E. coli, for example, expression vectors derived from mammals (for example, pcDNA3 (Invitrogen) and pEGF-BOS (Nucleic Acids Res 18(17): 5322 (1990)), pEF, pCDM8), expression vectors derived from insect cells (for example, "Bac-to-BAC baculovirus expression system" (GIBCO BRL), pBacPAK8), expression vectors derived from plants (e.g., pMH1, pMH2), expression vectors derived from animal viruses (e.g., pHSV, pMV, pAdexLcw), expression vectors derived from retroviruses (e.g., pZIpneo), expression vector derived from yeast (e.g., "Pichia Expression Kit" (Invitrogen), pNV11, SP-Q01) and expression vectors derived from Bacillus subtilis (e.g., pPL608, pKTH50) can be used for producing the polypeptide of the present invention.

[0208] In order to express the vector in animal cells, such as CHO, COS or NIH3T3 cells, the vector should carry a promoter necessary for expression in such cells, for example, the SV40 promoter (Mulligan et al., Nature 277: 108 (1979)), the MMLV-LTR promoter, the EF1 alpha promoter (Mizushima et al., Nucleic Acids Res 18: 5322 (1990)), the CMV promoter and the like, and preferably a marker gene for selecting transformants (for example, a drug resistance gene selected by a drug (e.g., neomycin, G418)). Examples of known vectors with these characteristics include, for example, pMAM, pDR2, pBK-RSV, pBK-CMV, pOPRSV and pOP13.

[0209] Hereinafter, the present invention is described in more detail with reference to the Examples. However, while the following materials, methods and examples may serve to assist one of ordinary skill in making and using certain embodiments of the present invention, there are only intended to illustrate aspects of the present invention and thus in no way to limit the scope of the present invention. As one of ordinary skill in the art will readily recognize, methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention.

Examples

[0210] Materials and Methods

Cell lines

T2, HLA-A*0201-positive B-lymphoblastoid cell line, HLA-A*0206-positive B-lymphoblastoid cell line, HT1376, J82, COS7 and UM-UC3 were purchased from ATCC. MKN-45 was purchased from JCRB.

[0211] Candidate selection of peptides derived from MPHOSPH1

9-mer and 10-mer peptides derived from MPHOSPH1 that bind to HLA-A*0201

molecule were predicted using binding prediction software "BIMAS" (http://www-bimas.cit.nih.gov/molbio/hla_bind) (Parker et al., J Immunol 1994, 152(1): 163-75; Kuzushima et al., Blood 2001, 98(6): 1872-81). These peptides were synthesized by Biosynthesis (Lewisville, Texas) according to a standard solid phase synthesis method and purified by reversed phase high performance liquid chromatography (HPLC). The purity (>90%) and the identity of the peptides were determined by analytical HPLC and mass spectrometry analysis, respectively. Peptides were dissolved in dimethylsulfoxide at 20 mg/ml and stored at -80 degrees C.

[0212] In vitro CTL Induction

Monocyte-derived dendritic cells (DCs) were used as antigen-presenting cells to induce cytotoxic T lymphocyte (CTL) that responses against peptides presented on human leukocyte antigen (HLA). DCs were generated in vitro as described elsewhere (Nakahara S et al., Cancer Res 2003 Jul 15, 63(14): 4112-8). Specifically, peripheral blood mononuclear cells isolated from a normal volunteer (HLA-A*0201 positive) by Ficoll-Paque plus (Pharmacia) solution were separated by adherence to a plastic tissue culture dish (Becton Dickinson) so as to enrich them as the monocyte fraction. The monocyte-enriched population was cultured in the presence of 1000 U/ml of granulocyte-macrophage colony-stimulating factor (R&D System) and 1000 U/ml of interleukin (IL)-4 (R&D System) in AIM-V Medium (Invitrogen) containing 2% heat-inactivated autologous serum (AS). After 7 days of culture, the cytokine-induced DCs were pulsed with 20 microgram/ml of each of the synthesized peptides in the presence of 3 microgram/ml of beta 2-microglobulin for 3 hr at 37 degrees C in AIM-V Medium. The generated cells appeared to express DC-associated molecules, such as CD80, CD83, CD86 and HLA class II, on their cell surfaces (data not shown). These peptide-pulsed DCs were then inactivated by X ray-irradiated (20 Gy) and mixed at a 1:20 ratio with autologous $CD8^+$ T cells, obtained by positive selection with CD8 Positive Isolation Kit (Dynal). These cultures were set up in 48-well plates (Corning); each well contained 1.5 X 10^4 peptide-pulsed DCs, 3 X 10^5 $CD8^+$ T cells and 10 ng/ml of IL-7 (R&D System) in 0.5 ml of AIM-V/2% AS medium. Three days later, these cultures were supplemented with IL-2 (CHIRON) to a final concentration of 20 IU/ml. On day 7 and 14, the T cells were further stimulated with the autologous peptide-pulsed DCs. The DCs were prepared each time by the same way described above. CTL was tested against peptide-pulsed T2 cells after the 3rd round of peptide stimulation on day 21 (Tanaka H et al., Br J Cancer 2001 Jan 5, 84(1): 94-9; Umano Y et al., Br J Cancer 2001 Apr 20, 84(8): 1052-7; Uchida N et al., Clin Cancer Res 2004 Dec 15, 10(24): 8577-86; Suda T et al., Cancer Sci 2006 May, 97(5): 411-9; Watanabe T et al., Cancer Sci 2005 Aug, 96(8): 498-506).

[0213] CTL Expansion Procedure

CTLs were expanded in culture using the method similar to the one described by Riddell et al. (Walter EA et al., N Engl J Med 1995 Oct 19, 333(16): 1038-44; Riddell SR et al., Nat Med 1996 Feb, 2(2): 216-23). A total of 5 X 10^4 CTLs were suspended in 25 ml of AIM-V/5% AS medium with 2 kinds of human B-lymphoblastoid cell lines, inactivated by Mitomycin C, in the presence of 40 ng/ml of anti-CD3 monoclonal antibody (Pharmingen). One day after initiating the cultures, 120 IU/ml of IL-2 were added to the cultures. The cultures were fed with fresh AIM-V/5% AS medium containing 30 IU/ml of IL-2 on days 5, 8 and 11 (Tanaka H et al., Br J Cancer 2001 Jan 5, 84(1): 94-9; Umano Y et al., Br J Cancer 2001 Apr 20, 84(8): 1052-7; Uchida N et al., Clin Cancer Res 2004 Dec 15, 10(24): 8577-86; Suda T et al., Cancer Sci 2006 May, 97(5): 411-9; Watanabe T et al., Cancer Sci 2005 Aug, 96(8): 498-506).

[0214] Establishment of CTL clones

The dilutions were made to have 0.3, 1, and 3 CTLs/well in 96 round-bottomed micro titer plate (Nalge Nunc International). CTLs were cultured with 1 X 10^4 cells/ well of 2 kinds of human B-lymphoblastoid cell lines, 30ng/ml of anti-CD3 antibody, and 125 U/ml of IL-2 in a total of 150 microliter/well of AIM-V Medium containing 5%AS. 50 microliter/well of IL-2 were added to the medium 10 days later to reach a final concentration of 125 U/ml IL-2. CTL activity was tested on the 14th day, and CTL clones were expanded using the same method as described above (Uchida N et al., Clin Cancer Res 2004 Dec 15, 10(24): 8577-86; Suda T et al., Cancer Sci 2006 May, 97(5): 411-9; Watanabe T et al., Cancer Sci 2005 Aug, 96(8): 498-506).

[0215] Specific CTL activity

To examine specific CTL activity, interferon(IFN)-gamma enzyme-linked immunospot (ELISPOT) assay and IFN-gamma enzyme-linked immunosorbent assay (ELISA)were performed. Peptide-pulsed T2 (1 x 10^4/well) was prepared as stimulator cells. Cultured cells in 48-well plate, CTL lines and CTL clones were used as responder cells. IFN-gamma ELISPOT assay and IFN-gamma ELISA assay were performed under manufacture procedure.

[0216] CTL ability to recognize the target cell line that endogenously expressed MPHOSPH1 and HLA-A*0201

The CTL clone was examined for its ability to recognize the target cell that endogenously expressed MPHOSPH1 and HLA-A*0201. Established CTL clone was cultured with target cell lines (5 X 10^4/well) for two overnight. After incubation, IFN-gamma in the culture media was measured by ELISA. IFN-gamma ELISA was performed under the manufacturer's procedure.

[0217] Cytotoxic activity

The CTL clones were examined for their ability to kill the tumor cells endogenously expressing MPHOSPH1 and HLA-A*0201. Target cells (tumor cell lines) were labeled

with 100 micro-Ci of $Na_2{}^{51}CrO_4$ (Perkin Elmer) for 1 hr in CO_2 incubator. Peptide-pulsed target cells were prepared by incubating the cells with 20 microgram/ml of the peptide for 16 hrs before labeling. Target cells labeled with ^{51}Cr were rinsed and mixed with CTL clones in a final volume of 200 microliter in 96-well round-bottom microtiter plates. The plates were centrifuged (4 minutes at 800 rpm) to increase cell-to-cell contact and placed into CO_2 incubator. After 4 hrs of incubation, 50 microliter of the supernatant was collected from each well and the radioactivity was determined with a gamma counter (PerkinElmer). The percentage of specific cytotoxicity was determined by calculating the percentage of specific ^{51}Cr-release by the following formula: {(cpm of the test sample release - cpm of the spontaneous release)/ (cpm of the maximum release - cpm of the spontaneous release)} X 100. Spontaneous release was determined by incubating the target cells alone, in the absence of effector cells. The maximum release was obtained by incubating the targets with 1N HCl. All measurements were done in duplicate.

[0218] Establishment of the cells forcibly expressing either or both of the target gene and HLA-A0206

The cDNA encoding an open reading frame of target genes or HLA-A*0206 was amplified by PCR. The PCR-amplified product was cloned into expression vector. The plasmids were transfected into COS7, which is the target gene and HLA-A*0206-null cell line, using lipofectamine 2000 (Invitrogen) according to the manufacturer's procedure. After 2 days from transfection, the transfected cells were harvested with versene (Invitrogen) and used as the stimulator cells (5 X 10^4 cells/ well) for CTL activity assay.

[0219] CTL ability to recognize the target cell line that endogenously expressed MPHOSPH1 and HLA-A*0206

The CTL clone were examined for its ability to recognize the target cell that endogenously expressed MPHOSPH1 and HLA-A*0206. Established CTL line and clone were cultured with target cell lines (5 X 10^4/well) for two overnight. After incubation, IFN-gamma in the culture media was measured by ELISA. IFN-gamma ELISA was performed under the manufacturer's procedure.

[0220] Results

Enhanced MPHOSPH1 expression in cancers

The wide gene expression profile data obtained from various cancers using cDNA-microarray revealed that MPHOSPH1 (GenBank Accession No. NM_016195; SEQ ID No: 125) expression was specifically elevated in cancer tissues as compared with corresponding normal tissue. MPHOSPH1 expression was validly elevated in 30 out of 31 Bladder cancer, 8 out of 36 Breast cancer, 18 out of 18 Cervical cancer, 5 out of 17 Cholangiocellular carcinoma, 25 out of 31 CML, 6 out of 11 Colorectal cancer, 6 out

of 14 Gastric cancer, 5 out of 5 NSCLC, 7 out of 7 Lymphoma, 6 out of 10 Osteosarcoma, 7 out of 22 Prostate cancer, 10 out of 18 Renal cancer and 15 out of 21 Soft tissue tumor (Table 1).

[0221] [Table 1]

Ratio of cases observed up-regulation of MPHOSPH1 in cancerous tissue as compared with normal corresponding tissue.

Cancer/Tumor	Ratio
Bladder cancer	30/31
Breast cancer	8/36
Cervical cancer	18/18
Cholangiocellular carcinoma	5/17
CML	25/31
Colorectal cancer	6/11
Gastric cancer	6/14
NSCLC	5/5
Lymphoma	7/7
Osteosarcoma	6/10
Prostate cancer	7/22
Renal cancer	10/18
Soft tissue tumor	15/21

[0222] Prediction of HLA-A02 binding peptides derived from MPHOSPH1

Tables 2a and 2b show the HLA-A02 binding 9mer and 10mer peptides of MPHOSPH1 in the order of high binding affinity. A total of 47 peptides having potential HLA-A02 binding ability were selected and examined to determine the epitope peptides.

[0223]

[Table 2a]

HLA-A02 binding 9mer peptides derived from MPHOSPH1

Start Position	amino acid sequence	score	SEQ ID NO
575	KLLDLIEDL	1278.3	1
282	YIYDLFVPV	1096.6	2
298	KMLRLSQDV	650.5	3
218	ALLRQIKEV	591.9	4
850	FLLTIENEL	363.6	5
1108	ALSELTQGV	285.2	6
331	KLGIKHQSV	243.4	7
1689	TLQKFGDFL	218.8	8
1251	KLTDAKKQI	149.7	9
638	RLAIFKDLV	129.5	10
1467	QLTEKDSDL	87.6	11
1195	NLQDMKHLL	87.6	12
270	SVWVSFFEI	83.5	13
129	FQGCIMQPV	74.6	14
839	VLQENNEGL	73.0	15
1094	TLDVQIQHV	64.0	16
1019	AIWEECKEI	48.8	17
1696	FLQHSPSIL	40.3	18
528	DLMEDEDLV	38.8	19
406	SLLTLGKCI	38.6	20
1400	KLTNLQDEL	36.6	21
170	GILPRTLNV	35.4	22
171	ILPRTLNVL	34.2	23
786	KICSERKRV	33.5	24
880	SLSEKKNLT	30.6	25
944	LMHTKIDEL	29.6	26
1422	WLEEKMMLI	29.0	27
466	TLNVLKFSA	28.8	28
1539	KLQTEPLST	26.1	29
132	CIMQPVKDL	25.0	30
1260	KQVQKEVSV	24.7	31
1184	KLKEEITQL	24.7	32
888	TLSKEVQQI	24.0	33
280	NEYIYDLFV	23.8	34

552	LLDEDLDKT	23.4	35
461	LAYDETLNV	21.5	36
980	NLPNTQLDL	21.4	37
409	TLGKCINVL	20.1	38
175	TLNVLFDSL	19.9	39
923	KLSNEIETA	19.6	40
1389	KEHENNTDV	19.4	41
987	DLLGNDYLV	19.3	42
920	KIMKLSNEI	18.6	43
1703	ILQSKAKKI	17.7	44
512	ILNVKRATI	17.7	45
1124	KELETILET	17.7	46
453	IVNISQCYL	17.5	47
771	LICNETVEV	16.3	48
623	TLLQEREIL	15.9	49
560	TLEENKAFI	15.1	50
1415	YNADRKKWL	14.5	51
307	KGYSFIKDL	13.7	52
133	IMQPVKDLL	12.9	53
1594	KMAVKHPGC	12.6	54
365	SEMSRVIRV	11.5	55
1191	QLTNNLQDM	11.4	56
871	QIVHFQQEL	11.2	57
245	NISEFEESI	11.0	58
484	TLNSSQEKL	10.5	59
764	SLIINNKLI	10.4	60
587	LINEKKEKL	10.0	61
263	MANSIKFSV	9.525	62
1354	VLEAKLEEV	8.528	63
846	GLRAFLLTI	6.93	64
83	ILDSQTVVL	5.956	65
1562	VLDSCEVST	5.067	66
15	YVFSADPIA	3.033	67
1741	YTSEISSPI	2.733	68
959	SQISNIDLL	2.441	69
82	HILDSQTVV	2.022	70

[0224]

[Table 2b]

HLA-A02 binding 10mer peptides derived from MPHOSPH1

Start Position	amino acid sequence	score	SEQ ID NO
1274	KLLRiKINEL	636.3	71
551	KLLDeDLDKT	445.9	72
460	YLAYdETLNV	319.9	73
943	KLMHtKIDEL	311.8	74
262	NMANsIKFSV	291.3	75
178	VLFDsLQERL	269.9	76
770	KLICnETVEV	243.4	77
34	KLDLsHEFSL	173.5	78
407	LLTLgKCINV	118.2	79
1714	TMSSsKLSNV	115.5	80
1353	QVLEaKLEEV	104.0	81
880	SLSEkKNLTL	87.6	82
235	TLYGsLTNSL	68.4	83
1019	AIWEeCKEIV	65.4	84
552	LLDEdLDKTL	59.6	85
1093	VTLDvQIQHV	57.3	86
559	KTLEeNKAFI	42.3	87
1332	KIIEdMRMTL	42.2	88
152	GLTNsGKTYT	41.0	89
830	NIAEiEDIRV	39.2	90
586	KLINeKKEKL	36.6	91
182	SLQErLYTKM	30.6	92
1043	QQIEkLQAEV	28.9	93
870	KQIVhFQQEL	28.8	94
1318	QQYErACKDL	28.4	95
452	MIVNiSQCYL	27.5	96
923	KLSNeIETAT	26.1	97
1257	KQIKqVQKEV	24.7	98
980	NLPNtQLDLL	24.1	99
985	QLDLIGNDYL	23.0	100
1427	MMLItQAKEA	22.6	101
1523	QIMDiKPKRI	21.8	102
1484	QLVAaLEIQL	21.4	103
466	TLNVIKFSAI	19.8	104

511	KILNvKRATI	18.6	105
1340	TLEEqEQTQV	18.3	106
372	RVSElSLCDL	17.6	107
1561	VVLDsCEVST	16.8	108
309	YSFIkDLQWI	14.7	109
353	SIFTvKILQI	12.2	110
1094	TLDVqIQHVV	11.4	111
1688	GTLQkFGDFL	11.2	112
311	FIKDlQWIQV	10.7	113
1079	TLIQqLKEEL	10.5	114
1128	TILEtQKVEC	10.4	115
1487	AALEiQLKAL	10.4	116
170	GILPrTLNVL	10.2	117
503	SLDSNSNSKI	4.173	118
1107	RALSELTQGV	3.574	119
282	YIYDLFVPVS	2.216	120
160	YTFQGTEENI	1.208	121
174	RTLNVLFDSL	1.022	122
82	HILDSQTVVL	0.621	123
128	FFQGCIMQPV	0.511	124

Start position indicates the number of amino acid residue from the N-terminus of MPHOSPH1. Binding score is derived from "BIMAS".

[0225] CTL induction with the predicted peptides from MPHOSPH1 restricted with HLA-A*0201

CTLs for those peptides derived from MPHOSPH1 were generated according to the protocols as described in "Materials and Methods". Peptide specific CTL activity was detected by IFN-gamma ELISPOT assay (Figure 1). The well number #7 stimulated with MPHOSPH1-A02-9-850 (SEQ ID NO: 5) (a), #5 stimulated with MPHOSPH1-A02-9-129 (SEQ ID NO: 14) (b), #5 stimulated with MPHOSPH1-A02-9-846 (SEQ ID NO: 64) (c), #2 stimulated with MPHOSPH1-A02-10-460 (SEQ ID NO: 73) (d), #1 stimulated with MPHOSPH1-A02-10-770 (SEQ ID NO: 77) (e), #1 stimulated with MPHOSPH1-A02-10-407 (SEQ ID NO: 79) (f), #4 stimulated with MPHOSPH1-A02-10-923 (SEQ ID NO: 97) (g), #5 stimulated with MPHOSPH1-A02-10-1484 (SEQ ID NO: 103) (h) and #8 stimulated with MPHOSPH1-A02-10-282 (SEQ ID NO: 120) (i) demonstrated potent IFN-gamma production as compared to the control wells. On the other hand, no specific CTL activity was detected by stimulation with other peptides shown in Table 2 a and b,

despite those peptides had possible binding activity with HLA-A*0201. As a typical case of negative data, no specific IFN-gamma production was observed from the CTL stimulated with MPHOSPH1-A02-9-575 (SEQ ID NO: 1) (j). Taken together, these results suggest that 10 selected peptides derived from MPHOSPH1 could induce potent CTLs.

[0226] Establishment of CTL line and clone against MPHOSPH1 derived peptide

The cells in the well number #7 stimulated with MPHOSPH1-A02-9-850 (SEQ ID NO: 5) (a), #5 stimulated with MPHOSPH1-A02-9-129 (SEQ ID NO: 14) (b), #5 stimulated with MPHOSPH1-A02-9-846 (SEQ ID NO: 64) (c), #2 stimulated with MPHOSPH1-A02-10-460 (SEQ ID NO: 73) (d), #1 stimulated with MPHOSPH1-A02-10-770 (SEQ ID NO: 77) (e), #1 stimulated with MPHOSPH1-A02-10-407 (SEQ ID NO: 79) (f), #4 stimulated with MPHOSPH1-A02-10-923 (SEQ ID NO: 97) (g), #5 stimulated with MPHOSPH1-A02-10-1484 (SEQ ID NO: 103) (h) and #8 stimulated with MPHOSPH1-A02-10-282 (SEQ ID NO: 120) (i), which showed peptide specific CTL activity in IFN-gamma ELISPOT assay were expanded and established the CTL lines (Figure 2). CTL activity of these CTL lines was measured by IFN-gamma ELISA. CTL lines demonstrated potent IFN-gamma production against T2 cells pulsed with the corresponding peptide as compared to T2 cells without peptide pulse. Furthermore, the CTL clones were established by limiting dilution from the CTL lines as described in "Materials and Methods", and IFN-gamma production from the CTL clones against T2 cells pulsed with corresponding peptide was measured by IFN-gamma ELISA. Potent IFN-gamma production was observed from the CTL clones stimulated with MPHOSPH1-A02-9-850 (SEQ ID NO: 5) (a), MPHOSPH1-A02-9-846 (SEQ ID NO: 64) (b), MPHOSPH1-A02-10-460 (SEQ ID NO: 73) (c), MPHOSPH1-A02-10-770 (SEQ ID NO: 77) (d) and MPHOSPH1-A02-10-282 (SEQ ID NO: 120) (e) (Figure 3).

[0227] Specific CTL activity against target cells expressing MPHOSPH1 and HLA-A*0201

The established CTL clone was examined for the ability to recognize target cells that express MPHOSPH1 and HLA-A*0201 molecule. The CTL clone stimulated with MPHOSPH1-A02-10-282 (SEQ ID NO: 120) showed potent CTL activity against J82 cells which both express MPHOSPH1 and HLA-A*0201. On the other hand, no significant specific CTL activity was detected against HT1376 cells which express MPHOSPH1 but not HLA-A*0201 and T2 cells which express HLA-A*0201 but not MPHOSPH1 (Figure 4). Thus, this data clearly demonstrates that MPHOSPH1-A02-10-282 (SEQ ID NO: 120) peptide was endogenously processed and expressed on the target cells with HLA-A*0201 molecule and was recognized by the CTLs.

[0228] Cytotoxic activity against tumor cell line expressing MPHOSPH1 and HLA-A*0201

The established CTL clones were examined for their ability to recognize and kill tumor cell lines that expressed MPHOSPH1 and HLA-A*0201. The CTL clone stimulated with MPHOSPH1-A02-10-282 (SEQ ID NO: 120) showed potent cytotoxic activity against UMUC-3 cells which both express MPHOSPH1 and HLA-A*0201. On the other hand, no significant cytotoxic activity was detected with both CTL clones against MKN45 cells which express MPHOSPH1 but not HLA-A*0201 and T2 cells which express HLA-A*0201 but not MPHOSPH1 (Fgure5). These results indicate that MPHOSPH1-A02-10-282 (SEQ ID NO: 120) peptide derived from MPHOSPH1 may be available to apply the cancer vaccines for patients with MPHOSPH1 expressing tumors.

[0229] Specific CTL activity against target cells expressing MPHOSPH1 and HLA-A*0206

The established CTL line raised against MPHOSPH1-A02-10-282 (SEQ ID NO: 120) peptide was examined for the ability to recognize target cells that express MPHOSPH1 and HLA-A*0206 molecule. COS7 cells transfected with both the full length of MPHOSPH1 and HLA-A*0206 gene (a specific model for the target cells that express MPHOSPH1 and HLA-A*0206 gene) were prepared as a stimulator cells, and COS7 cells transfected with either full length of MPHOSPH1 or HLA-A*0206 were used as the controls. In Figure 6, the CTL clone stimulated with MPHOSPH1-A02-10-282 (SEQ ID NO: 120) showed potent CTL activity against COS7 cells expressing both MPHOSPH1 and HLA- A*0206. On the other hand, no significant specific CTL activity was detected against the controls. Thus, these data clearly demonstrate that MPHOSPH1-A02-10-282 (SEQ ID NO: 120) peptide is endogenously processed and expressed on the target cells with HLA-A*0206 molecule and was recognized by the CTLs.

[0230] Homology analysis of antigen peptides

The CTLs stimulated with MPHOSPH1-A02-9-850 (SEQ ID NO: 5), MPHOSPH1-A02-9-129 (SEQ ID NO: 14), MPHOSPH1-A02-9-846 (SEQ ID NO: 64), MPHOSPH1-A02-10-460 (SEQ ID NO: 73), MPHOSPH1-A02-10-770 (SEQ ID NO: 77), MPHOSPH1-A02-10-407 (SEQ ID NO: 79), MPHOSPH1-A02-10-923 (SEQ ID NO: 97), MPHOSPH1-A02-10-1484 (SEQ ID NO: 103) and MPHOSPH1-A02-10-282 (SEQ ID NO: 120) showed significant and specific CTL activity. This result may be due to the fact that the sequence of MPHOSPH1-A02-9-850 (SEQ ID NO: 5), MPHOSPH1-A02-9-129 (SEQ ID NO: 14), MPHOSPH1-A02-9-846 (SEQ ID NO: 64), MPHOSPH1-A02-10-460 (SEQ ID NO: 73), MPHOSPH1-A02-10-770 (SEQ ID NO: 77), MPHOSPH1-A02-10-407 (SEQ ID NO: 79), MPHOSPH1-A02-10-923 (SEQ ID NO: 97), MPHOSPH1-A02-10-1484 (SEQ ID NO: 103) and MPHOSPH1-A02-10-282 (SEQ ID NO: 120) are homologous to peptide derived from other molecules that are known to sensitize the human immune

system. To exclude this possibility, homology analyses were performed for these peptide sequences using as queries the BLAST algorithm (http://www.ncbi.nlm.nih.gov/blast/blast.cgi) which revealed no sequence with significant homology. The results of homology analyses indicate that the sequence of MPHOSPH1-A02-9-850 (SEQ ID NO: 5), MPHOSPH1-A02-9-129 (SEQ ID NO: 14), MPHOSPH1-A02-9-846 (SEQ ID NO: 64), MPHOSPH1-A02-10-460 (SEQ ID NO: 73), MPHOSPH1-A02-10-770 (SEQ ID NO: 77), MPHOSPH1-A02-10-407 (SEQ ID NO: 79), MPHOSPH1-A02-10-923 (SEQ ID NO: 97), MPHOSPH1-A02-10-1484 (SEQ ID NO: 103) and MPHOSPH1-A02-10-282 (SEQ ID NO: 120) are unique and thus, there is little possibility, to our best knowledge, that this molecules raise unintended immunologic response to some unrelated molecule.

[0231] In conclusion, the novel HLA-A*0201 epitope peptides derived from MPHOSPH1 identified herein may find utility in the field of cancer immunotherapy.

Industrial Applicability

[0232] The present invention provides new TAAs, particularly those derived from MPHOSPH1, that may induce potent and specific anti-tumor immune responses and thus have applicability to a wide variety of cancer types. Such TAAs can find use as peptide vaccines against diseases associated with MPHOSPH1, e.g., cancer, more particularly, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, gastric cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, renal cancer and soft tissue tumor.

[0233] While the present invention is herein described in detail and with reference to specific embodiments thereof, it is to be understood that the foregoing description is exemplary and explanatory in nature and is intended to illustrate the present invention and its preferred embodiments. Through routine experimentation, one skilled in the art will readily recognize that various changes and modifications can be made therein without departing from the spirit and scope of the present invention, the metes and bounds of which are defined by the appended claims.

Claims

[Claim 1] An isolated peptide of the following (a) or (b):
(a) a peptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120;
(b) a peptide comprising an amino acid sequence that 1, 2, or several amino acid(s) are substituted, deleted, inserted, and/or added in the amino acid sequence selected from the group consisting of SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120, and wherein the peptide has cytotoxic T lymphocyte (CTL) inducibility.

[Claim 2] The isolated peptide of claim 1, wherein the peptide has one or both of the following characteristics:
(a) the second amino acid from the N-terminus of the amino acid sequence selected from the group consisting of SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 is selected from the group consisting of leucine and methionine; and
(b) the C-terminal amino acid of the amino acid sequence selected from the group consisting of SEQ ID NOs: 5, 14, 64, 73, 77, 79, 97, 103 and 120 is selected from the group consisting of valine and leucine.

[Claim 3] The isolated peptide of claim 1 or 2, wherein the peptide is a non-apeptide or decapeptide.

[Claim 4] An isolated polynucleotide encoding the peptide of any one of claim 1 to 3.

[Claim 5] A composition for inducing a CTL, wherein the composition comprises one or more peptide(s) of any one of claims 1 to 3, or one or more polynucleotide(s) of claim 4,

[Claim 6] A composition for inducing an APC having CTL inducibility, wherein the composition comprises one or more peptide(s) of any one of claims 1 to 3, or one or more polynucleotide(s) of claim 4.

[Claim 7] A pharmaceutical composition comprising at least one active ingredient selected from the group consisting of:
(a) one or more peptides of any one of claims 1 to 3;
(b) one or more polynucleotides encoding the peptide of any one of claims 1 to 3;
(c) one or more APCs or exosomes that present a complex of the peptide of any one of claims 1 to 3 and an HLA antigen on their surface; and

(d) one or more CTLs that recognize a cell presenting a complex of the peptide of any one of claims 1 to 3 and an HLA antigen on its surface.

[Claim 8] The pharmaceutical composition of claim 7 for use in either or both of the treatment and the prophylaxis of cancer, or inducing an immune response against cancer in a subject.

[Claim 9] The pharmaceutical composition of claim 7 or 8, wherein the pharmaceutical composition is formulated for administration to a subject whose HLA antigen is HLA-A2.

[Claim 10] A method for inducing an antigen-presenting cell (APC) having CTL inducibility, said method comprising a step selected from the group consisting of:

(a) contacting an APC with the peptide of any one of claims 1 to 3 in vitro, ex vivo or in vivo, and

(b) introducing a polynucleotide encoding the peptide of any one of claims 1 to 3 into an APC.

[Claim 11] A method for inducing a CTL, said method comprising a step selected from the group consisting of:

(a) co-culturing a CD8 positive T cell with an APC that presents on its surface a complex of an HLA antigen and the peptide of any one of claims 1 to 3;

(b) co-culturing a CD8 positive T cell with an exosome that presents on its surface a complex of an HLA antigen and the peptide of any one of claim 1 to 3; and

(c) introducing a polynucleotide encoding both of T cell receptor (TCR) subunits or polynucleotides encoding each of TCR subunits into a CD8 positive T cell, wherein the TCR can bind to a complex of an HLA antigen and the peptide of any one of claims 1 to 3 presented on a cell surface.

[Claim 12] An isolated APC that presents on its surface a complex of an HLA antigen and the peptide of any one of claims 1 to 3.

[Claim 13] The APC of claim 12, which is induced by the method of claim 10.

[Claim 14] An isolated CTL that recognizes a cell presenting on its surface a complex of an HLA antigen and the peptide of any one of claims 1 to 3.

[Claim 15] The CTL of claim 14, wherein said CTL is induced by the method of claim 11.

[Claim 16] A method of either or both of the treatment and prophylaxis of cancer in a subject, wherein the method comprises the step of administering to the subject (a) pharmaceutically effective amount(s) of:

(a) one or more peptides of any one of claims 1 to 3;
(b) one or more polynucleotides encoding the peptide of any one of claims 1 to 3;
(c) one or more APCs or exosomes that present a complex of the peptide of any one of claims 1 to 3 and an HLA antigen on their surface; or
(d) one or more CTLs that recognize a cell presenting a complex of the peptide of any one of claims 1 to 3 and an HLA antigen on its surface.

[Claim 17] A method of inducing an immune response against cancer in a subject in need thereof, said method comprising the step of administering to the subject a composition comprising a peptide of any one of claims 1 to 3 or a polynucleotide encoding the peptide.

[Claim 18] An antibody or immunologically active fragment thereof against the peptide of any one of claims 1 to 3.

[Claim 19] A vector comprising a nucleotide sequence encoding the peptide of any one of claims 1 to 3.

[Claim 20] A host cell transformed or transfected with the vector of claim 19.

[Claim 21] A diagnostic kit comprising the peptide of any one of claims 1 to 3, the polynucleotide of claim 4 or the antibody of claim 18.