

RNAI CONSTRUCTS FOR INHIBITING PNPLA3 EXPRESSION AND METHODS OF USE THEREOF

RELATED APPLICATIONS

[0001] This application claims priority to U.S. Provisional Patent Application No. 62/597,841, filed December 12, 2017, the contents of which are incorporated herein by reference.

SEQUENCE LISTING

[0002] The present application contains a Sequence Listing which has been submitted electronically in ASCII format and is hereby incorporated by reference in its entirety. Said ASCII copy, created on December 12, 2018, is named A-2219-WO-PCT_SL.txt and is 708,961 bytes in size. The information in the electronic format of the sequence listing is incorporated herein by reference in its entirety.

FIELD OF THE INVENTION

[0003] The present invention relates to compositions and methods for modulating liver expression of patatin-like phospholipase domain-containing 3 (PNPLA3). In particular, the present invention relates to nucleic acid-based therapeutics for reducing PNPLA3 expression via RNA interference and methods of using such nucleic acid-based therapeutics to treat or prevent liver disease, such as nonalcoholic fatty liver disease (NAFLD).

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BACKGROUND OF THE INVENTION

[0005] Comprising a spectrum of hepatic pathologies, nonalcoholic fatty liver disease (NAFLD) is the most common chronic liver disease in the world, the prevalence of which doubled in the last 20 years and now is estimated to affect approximately 20% of the world population (Sattar et al. (2014) BMJ 349:g4596; Loomba and Sanyal (2013) Nature Reviews

Gastroenterology & hepatology 10(11):686-690; Kim and Kim (2017) Clin Gastroenterol Hepatol 15(4):474-485; Petta et al. (2016) Dig Liver Dis 48(3):333-342). NAFLD begins with the accumulation of triglyceride in the liver and is defined by the presence of cytoplasmic lipid droplets in more than 5% of hepatocytes in an individual 1) without a history of significant alcohol consumption and 2) in which the diagnosis of other types of liver disease have been excluded (Zhu et al (2016) World J Gastroenterol 22(36):8226-33; Rinella (2015) JAMA 313(22):2263-73; Yki-Jarvinen (2016) Diabetologia 59(6):1104-11). In some individuals the accumulation of ectopic fat in the liver, called steatosis, triggers inflammation and hepatocellular injury leading to a more advanced stage of disease called, nonalcoholic steatohepatitis (NASH) (Rinella, supra). As of 2015, 75-100 million Americans are predicted to have NAFLD; NASH accounting for approximately 10-30% of NAFLD diagnoses (Rinella, supra; Younossi et al (2016) Hepatology 64(5):1577-1586).

[0006] Patatin-like phospholipase domain-containing 3 (PNPLA3), formerly known as adiponutrin (ADPN) and calcium-independent phospholipase A2-epsilon (iPLA(2) ϵ), is a type II transmembrane protein (Wilson et al (2006) J Lipid Res 47(9):1940-9; Jenkins et al (2004) J Biol Chem 279(47):48968-75). Initially identified in adipose cells as a membrane-associated, adipose-enriched protein induced during adipogenesis in mice, it is now well characterized to be expressed in other tissues, including the liver (Wilson et al, supra; Baulande et al (2001) J Biol Chem 276(36):33336-44; Moldes et al. (2006) Eur J Endocrinol 155(3):461-8; Faraj et al. (2006) J Endocrinol 191(2):427-35; Liu et al (2004) J Clin Endocrinol Metab 89(6):2684-9; Lake et al (2005) J Lipid Res 46(11):2477-87). In cell-free biochemical systems, recombinant PNPLA3 protein can exhibit either triacylglycerol lipase or transacylation activity (Jenkins et al., supra; Kumari et al (2012) Cell Metab 15(5):691-702; He et al (2010) J Biol Chem 285(9):6706-15). In hepatocytes, PNPLA3 is expressed on the endoplasmic reticulum and lipid membranes and predominantly exhibits triacylglycerol hydrolase activity (He et al., supra; Huang et al (2010) Proc Natl Acad Sci USA 107(17):7892-7; Ruhanen et al (2014) J Lipid Res 55(4):739-46; Pingitore et al. (2014) Biochim Biophys Acta 1841(4):574-80). Although lacking a secretory signal, data indicates PNPLA3 is secreted and can be found in human plasma as disulfide-bond dependent multimers (Winberg et al. (2014) Biochem Biophys Res Commun 446(4):1114-9). Accordingly,

novel therapeutics targeting PNPLA3 function represents a novel approach to reducing PNPLA3 levels and treating hepatologic diseases, such as nonalcoholic fatty liver disease.

SUMMARY

[0007] The present disclosure is based, in part, on the design and generation of RNAi constructs that target the PNPLA3 gene and reduce expression of PNPLA3 in liver cells. The sequence specific inhibition of PNPLA3 expression is useful for treating or preventing conditions associated with PNPLA3 expression, such as liver-related diseases, such as, for example, simple fatty liver (steatosis), nonalcoholic steatohepatitis (NASH), cirrhosis (irreversible, advanced scarring of the liver), or PNPLA3 related obesity. Accordingly, in one example, the present disclosure provides an RNAi construct comprising a sense strand and an antisense strand, wherein the antisense strand comprises a region having a sequence that is complementary to a PNPLA3 mRNA sequence. In certain examples, the antisense strand comprises a region having at least 15 contiguous nucleotides from an antisense sequence listed in Table 1 or Table 2. In some examples, the RNAi of the present disclosure selectively inhibits PNPLA3-rs738409, PNPLA3-rs738408, and/or PNPLA3-rs738409-rs738408 minor alleles over the reference allele which does not contain these changes.

[0008] In some examples, the sense strand of the RNAi constructs described herein comprises a sequence that is sufficiently complementary to the sequence of the antisense strand to form a duplex region of about 15 to about 30 base pairs in length. In these and other examples, the sense and antisense strands each are about 15 to about 30 nucleotides in length. In some examples, the RNAi constructs comprise at least one blunt end. In other examples, the RNAi constructs comprise at least one nucleotide overhang. Such nucleotide overhangs may comprise at least 1 to 6 unpaired nucleotides and can be located at the 3' end of the sense strand, the 3' end of the antisense strand, or the 3' end of both the sense and antisense strand. In certain examples, the RNAi constructs comprise an overhang of two unpaired nucleotides at the 3' end of the sense strand and the 3' end of the antisense strand. In other examples, the RNAi constructs comprise an overhang of two unpaired nucleotides at the 3' end of the antisense strand and a blunt end of the 3' end of the sense strand/5' end of the antisense strand.

[0009] The RNAi constructs of the disclosure may comprise one or more modified nucleotides, including nucleotides having modifications to the ribose ring, nucleobase, or phosphodiester backbone. In some examples, the RNAi constructs comprise one or more 2'-modified nucleotides. Such 2'-modified nucleotides can include 2'-fluoro modified nucleotides, 2'-O-methyl modified nucleotides, 2'-O-methoxyethyl modified nucleotides, 2'-O-allyl modified nucleotides, bicyclic nucleic acids (BNA), glycol nucleic acids (GNAs), inverted bases (e.g. inverted adenosine) or combinations thereof. In one particular example, the RNAi constructs comprise one or more 2'-fluoro modified nucleotides, 2'-O-methyl modified nucleotides, or combinations thereof. In some examples, all of the nucleotides in the sense and antisense strand of the RNAi construct are modified nucleotides.

[0010] In some examples, the RNAi constructs comprise at least one backbone modification, such as a modified internucleotide or internucleoside linkage. In certain examples, the RNAi constructs described herein comprise at least one phosphorothioate internucleotide linkage. In particular examples, the phosphorothioate internucleotide linkages may be positioned at the 3' or 5' ends of the sense and/or antisense strands.

[0011] In some examples, the antisense strand and/or the sense strand of the RNAi constructs of the disclosure may comprise or consist of a sequence from the antisense and sense sequences listed in Tables 1 or 2. In certain examples, the RNAi construct may be any one of the duplex compounds listed in any one of Tables 1 to 2.

[0011a] In one aspect, the present disclosure refers to an RNAi construct comprising a sense strand and an antisense strand, wherein the antisense strand consists of a nucleic acid sequence of SEQ ID NO: 648, 734, or 2, and wherein the RNAi construct inhibits the expression of Patatin-Like Phospholipase Domain Containing 3 (PNPLA3). In another aspect, the present disclosure refers to a pharmaceutical composition comprising the RNAi construct as disclosed herein and a pharmaceutically acceptable carrier, excipient, or diluent.

BRIEF DESCRIPTION OF THE FIGURES

[0012] Figure 1A-D show screening of five siRNA molecules for both dose-dependent mRNA knockdown and functional durability in vivo.

[0013] Figure 2A-G shows effect of PNPLA3 siRNA molecules in vivo in mice, liver weight, confirmation of human PNPLA3 expression, hepatic triglyceride content, serum TIMP1 levels, and histological indication of steatosis or inflammation.

[0014] Figure 3A-G shows effect of PNPLA3 siRNA molecules in vivo, liver weight, confirmation of human PNPLA3 expression, hepatic triglyceride content, serum TIMP1 levels, and histological indication of steatosis or inflammation.

[0015] Figure 4A-D shows the ability of a PNPLA3^{rs738409-rs738408} – specific siRNA molecule to rescue disease-associated phenotypes due to overexpression of PNPLA3^{rs738409-rs738408}, hepatic triglyceride content, serum TIMP1 levels, and histological indication of steatosis or inflammation.

[0016] Figure 5A-L shows the ability of a PNPLA3^{rs738409-rs738408} – specific siRNA molecule to prevent the development of early fibrosis.

DETAILED DESCRIPTION OF THE INVENTION

[0017] The present invention is directed to compositions and methods for regulating the expression of the Patatin-Like Phospholipase Domain Containing 3 (PNPLA3) gene. In some embodiments, the gene may be within a cell or subject, such as a mammal (e.g. a human). In some embodiments, compositions of the invention comprise RNAi constructs that target a PNPLA3 mRNA and reduce PNPLA3 expression in a cell or mammal. Such RNAi constructs are useful for treating or preventing various forms of liver-related diseases, such as, for example, simple fatty liver (steatosis), nonalcoholic steatohepatitis (NASH), cirrhosis (irreversible, advanced scarring of the liver), or PNPLA3 related obesity.

[0018] In 2008, a genome wide association study (GWAS) exploring nonsynonymous sequence variations, or single nucleotide polymorphisms (SNPs), associated with NAFLD identified a variant in PNPLA3, (rs738409[G], encoding I148M; which can be referred to as PNPLA3-rs738409, PNPLA3-ma or PNPLA3-minor allele), as significantly associated with hepatic fat content. Since this initial report, subsequent GWAS confirm PNPLA3 rs738409 as the major genetic determinant of NAFLD, significantly associated with 1) increased levels of the serum biomarker for liver damage, alanine transaminase (ALT), 2) NAFLD incidence, progression and severity, 3) both obese and lean individuals, and 4) the only known SNP shown to be significantly associated with all stages of NAFLD: steatosis, NASH, cirrhosis and hepatic cell carcinoma. The consensus among numerous GWAS indicate the association of PNPLA3 rs738409 with NAFLD is independent of age, gender, ethnicity, metabolic syndrome, body mass index, insulin resistance, and serum lipids. Furthermore, statistical analyses from multiple sources estimate approximately 50% of NAFLD patients carry the PNPLA3 rs738409 mutation. Patients can be homozygous or heterozygous for the PNPLA3 rs738409 mutation. Additionally, it has

been discovered that patients having the PNPLA3 rs738409 mutation often also carry an rs738408 mutation 3 base pairs away (Tian et al (2010) Nature Genetics 42:21-23). Thus, a patient can have a PNPLA3-rs738409 minor allele, PNPLA3-rs738408 minor allele or PNPLA3-rs738409-rs738408 double minor allele mutation (PNPLA3-dma).

[0019] Investigators have developed mouse models for exploring PNPLA3 function in vivo. To date, no detectable metabolic phenotype has been identified as the result of *Pnpla3*-deficiency or *Pnpla3* over-expression. In contrast, expression of *Pnpla3*^{I148M} in both transgenic mice and knock-in mice, led to increased hepatic triglyceride levels akin to NAFLD. Thus, combined, the in vivo mouse model data points to expression of the mutant *Pnpla3*^{I148M} protein, and not over-expression of the wild type protein, as the driver of the disease phenotype. These findings, in addition to the high frequency of the minor allele in NAFLD-affected individuals and prevailing association with the disease, underline PNPLA3 rs738409 as a prime therapeutic target for NAFLD.

[0020] RNA interference (RNAi) is the process of introducing exogenous RNA into a cell leading to specific degradation of the mRNA encoding the targeted protein with a resultant decrease in protein expression. Advances in both the RNAi technology and hepatic delivery and growing positive outcomes with other RNAi-based therapies, suggest RNAi as a compelling means to therapeutically treat NAFLD by directly targeting PNPLA3I148M. Numerous GWAS indicate a dose-dependent effect of PNPLA3 rs738409 on NAFLD incidence, progression and severity (GWAS); the odds ratio tending to be double, if not more, for homozygote carriers versus heterozygote carriers, yet still at least two-fold more for heterozygotes versus wild type individuals. Thus, silencing PNPLA3 employing allelic discrimination specificity could be both a potential means to reduce hepatic triglyceride in PNPLA3I148M carriers, but also present a scenario in which heterozygotes may gain benefit without silencing of the wild type allele. Along these lines, we identified SNP-specific short interfering RNAs (siRNA) to PNPLA3I148M and demonstrate proof of concept in vitro. Using both hepatoma cell lines, Hep3B (homozygous for the reference allele, PNPLA3I148I) and HEPG2 (homozygous for the minor allele, PNPLA3I148M), we identified siRNA sequences capable of inhibiting specifically PNPLA3I148M gene expression. The inhibitory effect of these sequences were confirmed by

screening on Chinese hamster ovary (CHO) cells over-expressing either PNPLA3I148I or PNPLA3I148M. Using adeno-associated virus (AAV) to overexpress human PNPLA3I148M in vivo, we then demonstrated treatment with minor allele-specific SNPs not only specifically reduced human PNPLA3I148M expression in mice, but also significantly reversed hepatic triglyceride accumulation induced by over-expression of human PNPLA3I148M.

[0021] As used herein, the term "RNAi construct" refers to an agent comprising a RNA molecule that is capable of downregulating expression of a target gene (e.g. PNPLA3) via a RNA interference mechanism when introduced into a cell. RNA interference is the process by which a nucleic acid molecule induces the cleavage and degradation of a target RNA molecule (e.g. messenger RNA or mRNA molecule) in a sequence-specific manner, e.g. through a RNA induced silencing complex (RISC) pathway. In some embodiments, the RNAi construct comprises a double-stranded RNA molecule comprising two antiparallel strands of contiguous nucleotides that are sufficiently complementary to each other to hybridize to form a duplex region. "Hybridize" or "hybridization" refers to the pairing of complementary polynucleotides, typically via hydrogen bonding (e.g. Watson-Crick, Hoogsteen or reversed Hoogsteen hydrogen bonding) between complementary bases in the two polynucleotides. The strand comprising a region having a sequence that is substantially complementary to a target sequence (e.g. target mRNA) is referred to as the "antisense strand." The "sense strand" refers to the strand that includes a region that is substantially complementary to a region of the antisense strand. In some embodiments, the sense strand may comprise a region that has a sequence that is substantially identical to the target sequence.

[0022] In some embodiments, the invention is an RNAi directed to PNPLA3. In some embodiments, the invention is an RNAi that binds at the PNPLA3 rs738409 site. In some embodiments, the invention is an RNAi that binds at the PNPLA3 rs738408 site. In some embodiments, the invention is an RNAi that binds at both the PNPLA3 rs738409 rs738408 sites. In some embodiments, the invention is an RNAi that preferentially binds PNPLA3 rs738409 over the native PNPLA3 sequence (PNPLA3-ref). In some embodiments, the invention is an RNAi that preferentially binds PNPLA3 rs738408 over the PNPLA3-ref sequence. In some embodiments, the invention is an RNAi that preferentially binds PNPLA3-dma over PNPLA3-ma. In some

embodiments, the invention in an RNAi molecule that contains any of the sequences found in Table 1 or 2.

[0023] A double-stranded RNA molecule may include chemical modifications to ribonucleotides, including modifications to the ribose sugar, base, or backbone components of the ribonucleotides, such as those described herein or known in the art. Any such modifications, as used in a double-stranded RNA molecule (e.g. siRNA, shRNA, or the like), are encompassed by the term "double-stranded RNA" for the purposes of this disclosure.

[0024] As used herein, a first sequence is "complementary" to a second sequence if a polynucleotide comprising the first sequence can hybridize to a polynucleotide comprising the second sequence to form a duplex region under certain conditions, such as physiological conditions. Other such conditions can include moderate or stringent hybridization conditions, which are known to those of skill in the art. A first sequence is considered to be fully complementary (100% complementary) to a second sequence if a polynucleotide comprising the first sequence base pairs with a polynucleotide comprising the second sequence over the entire length of one or both nucleotide sequences without any mismatches. A sequence is "substantially complementary" to a target sequence if the sequence is at least about 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or 100% complementary to a target sequence. Percent complementarity can be calculated by dividing the number of bases in a first sequence that are complementary to bases at corresponding positions in a second or target sequence by the total length of the first sequence. A sequence may also be said to be substantially complementary to another sequence if there are no more than 5, 4, 3, 2, or 1 mismatches over a 30 base pair duplex region when the two sequences are hybridized. Generally, if any nucleotide overhangs, as defined herein, are present, the sequence of such overhangs is not considered in determining the degree of complementarity between two sequences. By way of example, a sense strand of 21 nucleotides in length and an antisense strand of 21 nucleotides in length that hybridize to form a 19 base pair duplex region with a 2 nucleotide overhang at the 3' end of each strand would be considered to be fully complementary as the term is used herein.

[0025] In some embodiments, a region of the antisense strand comprises a sequence that is fully complementary to a region of the target RNA sequence (e.g. PNPLA3 mRNA). In such

embodiments, the sense strand may comprise a sequence that is fully complementary to the sequence of the antisense strand. In other such embodiments, the sense strand may comprise a sequence that is substantially complementary to the sequence of the antisense strand, e.g. having 1, 2, 3, 4, or 5 mismatches in the duplex region formed by the sense and antisense strands. In certain embodiments, it is preferred that any mismatches occur within the terminal regions (e.g. within 6, 5, 4, 3, 2, or 1 nucleotides of the 5' and/or 3' ends of the strands). In one embodiment, any mismatches in the duplex region formed from the sense and antisense strands occur within 6, 5, 4, 3, 2, or 1 nucleotides of the 5' end of the antisense strand.

[0026] In certain embodiments, the sense strand and antisense strand of the double-stranded RNA may be two separate molecules that hybridize to form a duplex region, but are otherwise unconnected. Such double-stranded RNA molecules formed from two separate strands are referred to as "small interfering RNAs" or "short interfering RNAs" (siRNAs). Thus, in some embodiments, the RNAi constructs of the invention comprise a siRNA.

[0027] Where the two substantially complementary strands of a dsRNA are comprised by separate RNA molecules, those molecules need not, but can be covalently connected. Where the two strands are connected covalently by means other than an uninterrupted chain of nucleotides between the 3'-end of one strand and the 5'-end of the respective other strand forming the duplex structure, the connecting structure is referred to as a "linker." The RNA strands may have the same or a different number of nucleotides. The maximum number of base pairs in the duplex is the number of nucleotides in the shortest strand of the dsRNA minus any overhangs that are present in the duplex. In addition to the duplex structure, an RNAi may comprise one or more nucleotide overhangs.

[0028] In other embodiments, the sense strand and the antisense strand that hybridize to form a duplex region may be part of a single RNA molecule, i.e. the sense and antisense strands are part of a self-complementary region of a single RNA molecule. In such cases, a single RNA molecule comprises a duplex region (also referred to as a stem region) and a loop region. The 3' end of the sense strand is connected to the 5' end of the antisense strand by a contiguous sequence of unpaired nucleotides, which will form the loop region. The loop region is typically of a sufficient length to allow the RNA molecule to fold back on itself such that the antisense strand can base pair with the

sense strand to form the duplex or stem region. The loop region can comprise from about 3 to about 25, from about 5 to about 15, or from about 8 to about 12 unpaired nucleotides. Such RNA molecules with at least partially self-complementary regions are referred to as "short hairpin RNAs" (shRNAs). In some embodiments, the loop region can comprise at least 1, 2, 3, 4, 5, 10, 20, or 25 unpaired nucleotides. In some embodiments, the loop region can have 10, 9, 8, 7, 6, 5, 4, 3, 2, or fewer unpaired nucleotides. In certain embodiments, the RNAi constructs of the invention comprise a shRNA. The length of a single, at least partially self-complementary RNA molecule can be from about 35 nucleotides to about 100 nucleotides, from about 45 nucleotides to about 85 nucleotides, or from about 50 to about 60 nucleotides and comprise a duplex region and loop region each having the lengths recited herein.

[0029] In some embodiments, the RNAi constructs of the invention comprise a sense strand and an antisense strand, wherein the antisense strand comprises a region having a sequence that is substantially or fully complementary to a PNPLA3 messenger RNA (mRNA) sequence. As used herein, a "PNPLA3 mRNA sequence" refers to any messenger RNA sequence, including splice variants, encoding a PNPLA3 protein, including PNPLA3 protein variants or isoforms from any species (e.g. mouse, rat, non-human primate, human). PNPLA3 protein is also known as adiponutrin (ADPN) and calcium-independent phospholipase A2-epsilon (iPLA(2)).

[0030] A PNPLA3 mRNA sequence also includes the transcript sequence expressed as its complementary DNA (cDNA) sequence. A cDNA sequence refers to the sequence of an mRNA transcript expressed as DNA bases (e.g. guanine, adenine, thymine, and cytosine) rather than RNA bases (e.g. guanine, adenine, uracil, and cytosine). Thus, the antisense strand of the RNAi constructs of the invention may comprise a region having a sequence that is substantially or fully complementary to a target PNPLA3 mRNA sequence or PNPLA3 cDNA sequence. A PNPLA3 mRNA or cDNA sequence can include, but is not limited to, any PNPLA3 mRNA or cDNA sequence such as can be derived from the NCBI Reference sequence NM_025225.2.

[0031] A region of the antisense strand can be substantially complementary or fully complementary to at least 15 consecutive nucleotides of the PNPLA3 mRNA sequence. In some embodiments, the target region of the PNPLA3 mRNA sequence to which the antisense strand comprises a region of complementarity can range from about 15 to about 30 consecutive

nucleotides, from about 16 to about 28 consecutive nucleotides, from about 18 to about 26 consecutive nucleotides, from about 17 to about 24 consecutive nucleotides, from about 19 to about 25 consecutive nucleotides, from about 19 to about 23 consecutive nucleotides, or from about 19 to about 21 consecutive nucleotides. In certain embodiments, the region of the antisense strand comprising a sequence that is substantially or fully complementary to a PNPLA3 mRNA sequence may, in some embodiments, comprise at least 15 contiguous nucleotides from an antisense sequence listed in Table 1 or Table 2. In other embodiments, the antisense sequence comprises at least 16, at least 17, at least 18, or at least 19 contiguous nucleotides from an antisense sequence listed in Table 1 or Table 2. In some embodiments, the sense and/or antisense sequence comprises at least 15 nucleotides from a sequence listed in Table 1 or 2 with no more than 1, 2, or 3 nucleotide mismatches.

[0032] The sense strand of the RNAi construct typically comprises a sequence that is sufficiently complementary to the sequence of the antisense strand such that the two strands hybridize under physiological conditions to form a duplex region. A "duplex region" refers to the region in two complementary or substantially complementary polynucleotides that form base pairs with one another, either by Watson-Crick base pairing or other hydrogen bonding interaction, to create a duplex between the two polynucleotides. The duplex region of the RNAi construct should be of sufficient length to allow the RNAi construct to enter the RNA interference pathway, e.g. by engaging the Dicer enzyme and/or the RISC complex. For instance, in some embodiments, the duplex region is about 15 to about 30 base pairs in length. Other lengths for the duplex region within this range are also suitable, such as about 15 to about 28 base pairs, about 15 to about 26 base pairs, about 15 to about 24 base pairs, about 15 to about 22 base pairs, about 17 to about 28 base pairs, about 17 to about 26 base pairs, about 17 to about 24 base pairs, about 17 to about 23 base pairs, about 17 to about 21 base pairs, about 19 to about 25 base pairs, about 19 to about 23 base pairs, or about 19 to about 21 base pairs. In one embodiment, the duplex region is about 17 to about 24 base pairs in length. In another embodiment, the duplex region is about 19 to about 21 base pairs in length.

[0033] In some embodiments, an RNAi agent of the invention contains a duplex region of about 24 to about 30 nucleotides that interacts with a target RNA sequence, e.g., an PNPLA3 target

mRNA sequence, to direct the cleavage of the target RNA. Without wishing to be bound by theory, long double stranded RNA introduced into cells can be broken down into siRNA by a Type III endonuclease known as Dicer (Sharp et al. (2001) *Genes Dev.* 15:485). Dicer, a ribonuclease-III-like enzyme, processes the dsRNA into 19-23 base pair short interfering RNAs with characteristic two base 3' overhangs (Bernstein, et al., (2001) *Nature* 409:363). The siRNAs are then incorporated into an RNA-induced silencing complex (RISC) where one or more helicases unwind the siRNA duplex, enabling the complementary antisense strand to guide target recognition (Nykanen, et al., (2001) *Cell* 107:309). Upon binding to the appropriate target mRNA, one or more endonucleases within the RISC cleave the target to induce silencing (Elbashir, et al., (2001) *Genes Dev.* 15: 188).

[0034] For embodiments in which the sense strand and antisense strand are two separate molecules (e.g. RNAi construct comprises a siRNA), the sense strand and antisense strand need not be the same length as the length of the duplex region. For instance, one or both strands may be longer than the duplex region and have one or more unpaired nucleotides or mismatches flanking the duplex region. Thus, in some embodiments, the RNAi construct comprises at least one nucleotide overhang. As used herein, a "nucleotide overhang" refers to the unpaired nucleotide or nucleotides that extend beyond the duplex region at the terminal ends of the strands. Nucleotide overhangs are typically created when the 3' end of one strand extends beyond the 5' end of the other strand or when the 5' end of one strand extends beyond the 3' end of the other strand. The length of a nucleotide overhang is generally between 1 and 6 nucleotides, 1 and 5 nucleotides, 1 and 4 nucleotides, 1 and 3 nucleotides, 2 and 6 nucleotides, 2 and 5 nucleotides, or 2 and 4 nucleotides. In some embodiments, the nucleotide overhang comprises 1, 2, 3, 4, 5, or 6 nucleotides. In one particular embodiment, the nucleotide overhang comprises 1 to 4 nucleotides. In certain embodiments, the nucleotide overhang comprises 2 nucleotides. The nucleotides in the overhang can be ribonucleotides, deoxyribonucleotides, or modified nucleotides as described herein. In some embodiments, the overhang comprises a 5'-uridine-uridine-3' (5'-UU-3') dinucleotide. In such embodiments, the UU dinucleotide may comprise ribonucleotides or modified nucleotides, e.g. 2'-modified nucleotides. In other embodiments, the overhang comprises a 5'-deoxythymidine-deoxythymidine-3' (5'-dTdT-3') dinucleotide.

[0035] The nucleotide overhang can be at the 5' end or 3' end of one or both strands. For example, in one embodiment, the RNAi construct comprises a nucleotide overhang at the 5' end and the 3' end of the antisense strand. In another embodiment, the RNAi construct comprises a nucleotide overhang at the 5' end and the 3' end of the sense strand. In some embodiments, the RNAi construct comprises a nucleotide overhang at the 5' end of the sense strand and the 5' end of the antisense strand. In other embodiments, the RNAi construct comprises a nucleotide overhang at the 3' end of the sense strand and the 3' end of the antisense strand.

[0036] The RNAi constructs may comprise a single nucleotide overhang at one end of the double-stranded RNA molecule and a blunt end at the other. A "blunt end" means that the sense strand and antisense strand are fully base-paired at the end of the molecule and there are no unpaired nucleotides that extend beyond the duplex region. In some embodiments, the RNAi construct comprises a nucleotide overhang at the 3' end of the sense strand and a blunt end at the 5' end of the sense strand and 3' end of the antisense strand. In other embodiments, the RNAi construct comprises a nucleotide overhang at the 3' end of the antisense strand and a blunt end at the 5' end of the antisense strand and the 3' end of the sense strand. In certain embodiments, the RNAi construct comprises a blunt end at both ends of the double-stranded RNA molecule. In such embodiments, the sense strand and antisense strand have the same length and the duplex region is the same length as the sense and antisense strands (i.e. the molecule is double-stranded over its entire length).

[0037] The sense strand and antisense strand can each independently be about 15 to about 30 nucleotides in length, about 18 to about 28 nucleotides in length, about 19 to about 27 nucleotides in length, about 19 to about 25 nucleotides in length, about 19 to about 23 nucleotides in length, about 21 to about 25 nucleotides in length, or about 21 to about 23 nucleotides in length. In certain embodiments, the sense strand and antisense strand are each about 18, about 19, about 20, about 21, about 22, about 23, about 24, or about 25 nucleotides in length. In some embodiments, the sense strand and antisense strand have the same length but form a duplex region that is shorter than the strands such that the RNAi construct has two nucleotide overhangs. For instance, in one embodiment, the RNAi construct comprises (i) a sense strand and an antisense strand that are each 21 nucleotides in length, (ii) a duplex region that is 19 base pairs in length, and (iii) nucleotide

overhangs of 2 unpaired nucleotides at both the 3' end of the sense strand and the 3' end of the antisense strand. In another embodiment, the RNAi construct comprises (i) a sense strand and an antisense strand that are each 23 nucleotides in length, (ii) a duplex region that is 21 base pairs in length, and (iii) nucleotide overhangs of 2 unpaired nucleotides at both the 3' end of the sense strand and the 3' end of the antisense strand. In other embodiments, the sense strand and antisense strand have the same length and form a duplex region over their entire length such that there are no nucleotide overhangs on either end of the double-stranded molecule. In one such embodiment, the RNAi construct is blunt ended and comprises (i) a sense strand and an antisense strand, each of which is 21 nucleotides in length, and (ii) a duplex region that is 21 base pairs in length. In another such embodiment, the RNAi construct is blunt ended and comprises (i) a sense strand and an antisense strand, each of which is 23 nucleotides in length, and (ii) a duplex region that is 23 base pairs in length.

[0038] In other embodiments, the sense strand or the antisense strand is longer than the other strand and the two strands form a duplex region having a length equal to that of the shorter strand such that the RNAi construct comprises at least one nucleotide overhang. For example, in one embodiment, the RNAi construct comprises (i) a sense strand that is 19 nucleotides in length, (ii) an antisense strand that is 21 nucleotides in length, (iii) a duplex region of 19 base pairs in length, and (iv) a single nucleotide overhang of 2 unpaired nucleotides at the 3' end of the antisense strand. In another embodiment, the RNAi construct comprises (i) a sense strand that is 21 nucleotides in length, (ii) an antisense strand that is 23 nucleotides in length, (iii) a duplex region of 21 base pairs in length, and (iv) a single nucleotide overhang of 2 unpaired nucleotides at the 3' end of the antisense strand.

[0039] The antisense strand of the RNAi constructs of the invention can comprise the sequence of any one of the antisense sequences listed in Table 1 or Table 2 or the sequence of nucleotides 1-19 of any of these antisense sequences. Each of the antisense sequences listed in Tables 1 and 6 comprises a sequence of 19 consecutive nucleotides (first 19 nucleotides counting from the 5' end) that is complementary to a PNPLA3 mRNA sequence plus a two nucleotide overhang sequence. Thus, in some embodiments, the antisense strand comprises a sequence of nucleotides 1-19 of any one of SEQ ID NOs: 1-166 or 167-332.

Modified nucleotides

[0040] The RNAi constructs of the invention may comprise one or more modified nucleotides. A "modified nucleotide" refers to a nucleotide that has one or more chemical modifications to the nucleoside, nucleobase, pentose ring, or phosphate group. As used herein, modified nucleotides do not encompass ribonucleotides containing adenosine monophosphate, guanosine monophosphate, uridine monophosphate, and cytidine monophosphate, and deoxyribonucleotides containing deoxyadenosine monophosphate, deoxyguanosine monophosphate, deoxythymidine monophosphate, and deoxycytidine monophosphate. However, the RNAi constructs may comprise combinations of modified nucleotides, ribonucleotides, and deoxyribonucleotides. Incorporation of modified nucleotides into one or both strands of double-stranded RNA molecules can improve the in vivo stability of the RNA molecules, e.g., by reducing the molecules' susceptibility to nucleases and other degradation processes. The potency of RNAi constructs for reducing expression of the target gene can also be enhanced by incorporation of modified nucleotides.

[0041] In certain embodiments, the modified nucleotides have a modification of the ribose sugar. These sugar modifications can include modifications at the 2' and/or 5' position of the pentose ring as well as bicyclic sugar modifications. A 2'-modified nucleotide refers to a nucleotide having a pentose ring with a substituent at the 2' position other than H or OH. Such 2' modifications include, but are not limited to, 2'-O-alkyl (e.g. O-C1-C10 or O-C1-C10 substituted alkyl), 2'-O-allyl (O-CH₂CH=CH₂), 2'-C-allyl, 2'-fluoro, 2'-O-methyl (OCH₃), 2'-O-methoxyethyl (O-(CH₂)₂OCH₃), 2'-OCF₃, 2'-O(CH₂)₂SCH₃, 2'-O-aminoalkyl, 2'-amino (e.g. NH₂), 2'-O-ethylamine, and 2'-azido. Modifications at the 5' position of the pentose ring include, but are not limited to, 5'-methyl (R or S), 5'-vinyl, and 5'-methoxy.

[0042] A "bicyclic sugar modification" refers to a modification of the pentose ring where a bridge connects two atoms of the ring to form a second ring resulting in a bicyclic sugar structure. In some embodiments the bicyclic sugar modification comprises a bridge between the 4' and 2' carbons of the pentose ring. Nucleotides comprising a sugar moiety with a bicyclic sugar modification are referred to herein as bicyclic nucleic acids or BNAs. Exemplary bicyclic sugar modifications include, but are not limited to, □-L-Methyleneoxy (4'-CH₂-O-2') bicyclic nucleic

acid (BNA); $\square$ -D-Methyleneoxy (4'-CH₂-O-2') BNA (also referred to as a locked nucleic acid or LNA); Ethyleneoxy (4'-(CH₂)₂-O-2') BNA; Aminooxy (4'-CH₂-O-N(R)-2')BNA; Oxyamino (4'-CH₂-N(R)-O-2') BNA; Methyl(methyleneoxy) (4'-CH(CH₃)-O-2') BNA (also referred to as constrained ethyl or cEt); methylene-thio (4'-CH₂-S-2') BNA; methylene-amino (4'-CH₂-N(R)-2') BNA; methyl carbocyclic (4'-CH₂-CH(CH₃)-2') BNA; propylene carbocyclic (4'-(CH₂)₃-2') BNA; and Methoxy(ethyleneoxy) (4'-CH(CH₂OMe)-O-2')BNA (also referred to as constrained MOE or cMOE). These and other sugar-modified nucleotides that can be incorporated into the RNAi constructs of the invention are described in U.S. Patent No. 9,181,551, U.S. Patent Publication No. 2016/0122761, and Deleavey and Damha, Chemistry and Biology, Vol. 19: 937-954, 2012, all of which are hereby incorporated by reference in their entireties.

[0043] In some embodiments, the RNAi constructs comprise one or more 2'-fluoro modified nucleotides, 2'-O-methyl modified nucleotides, 2'-O-methoxyethyl modified nucleotides, 2'-O-allyl modified nucleotides, bicyclic nucleic acids (BNAs), or combinations thereof. In certain embodiments, the RNAi constructs comprise one or more 2'-fluoro modified nucleotides, 2'-O-methyl modified nucleotides, 2'-O-methoxyethyl modified nucleotides, or combinations thereof. In one particular embodiment, the RNAi constructs comprise one or more 2'-fluoro modified nucleotides, 2'-O-methyl modified nucleotides or combinations thereof.

[0044] Both the sense and antisense strands of the RNAi constructs can comprise one or multiple modified nucleotides. For instance, in some embodiments, the sense strand comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more modified nucleotides. In certain embodiments, all nucleotides in the sense strand are modified nucleotides. In some embodiments, the antisense strand comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more modified nucleotides. In other embodiments, all nucleotides in the antisense strand are modified nucleotides. In certain other embodiments, all nucleotides in the sense strand and all nucleotides in the antisense strand are modified nucleotides. In these and other embodiments, the modified nucleotides can be 2'-fluoro modified nucleotides, 2'-O-methyl modified nucleotides, or combinations thereof.

[0045] In some embodiments, all pyrimidine nucleotides preceding an adenosine nucleotide in the sense strand, antisense strand, or both strands are modified nucleotides. For example, where the sequence 5'-CA-3' or 5'-UA-3' appears in either strand, the cytidine and uridine nucleotides are

modified nucleotides, preferably 2'-O-methyl modified nucleotides. In certain embodiments, all pyrimidine nucleotides in the sense strand are modified nucleotides (e.g. 2'-O-methyl modified nucleotides), and the 5' nucleotide in all occurrences of the sequence 5'-CA-3' or 5'-UA-3' in the antisense strand are modified nucleotides (e.g. 2'-O-methyl modified nucleotides). In other embodiments, all nucleotides in the duplex region are modified nucleotides. In such embodiments, the modified nucleotides are preferably 2'-O-methyl modified nucleotides, 2'-fluoro modified nucleotides or combinations thereof.

[0046] In embodiments in which the RNAi construct comprises a nucleotide overhang, the nucleotides in the overhang can be ribonucleotides, deoxyribonucleotides, or modified nucleotides. In one embodiment, the nucleotides in the overhang are deoxyribonucleotides, e.g., deoxythymidine. In another embodiment, the nucleotides in the overhang are modified nucleotides. For instance, in some embodiments, the nucleotides in the overhang are 2'-O-methyl modified nucleotides, 2'-fluoro modified nucleotides, 2'-methoxyethyl modified nucleotides, or combinations thereof.

[0047] The RNAi constructs of the invention may also comprise one or more modified internucleotide linkages. As used herein, the term "modified internucleotide linkage" refers to an internucleotide linkage other than the natural 3' to 5' phosphodiester linkage. In some embodiments, the modified internucleotide linkage is a phosphorous-containing internucleotide linkage, such as a phosphotriester, aminoalkyl phosphotriester, an alkylphosphonate (e.g. methylphosphonate, 3'-alkylene phosphonate), a phosphinate, a phosphoramidate (e.g. 3'-aminophosphoramidate and aminoalkylphosphoramidate), a phosphorothioate (P=S), a chiralphosphorothioate, a phosphorodithioate, a thionophosphoramidate, a thionoalkylphosphonate, a thionoalkylphosphotriester, and a boranophosphate. In one embodiment, a modified internucleotide linkage is a 2' to 5' phosphodiester linkage. In other embodiments, the modified internucleotide linkage is a non-phosphorous-containing internucleotide linkage and thus can be referred to as a modified internucleoside linkage. Such non-phosphorous-containing linkages include, but are not limited to, morpholino linkages (formed in part from the sugar portion of a nucleoside); siloxane linkages (-O-Si(H)₂-O-); sulfide, sulfoxide and sulfone linkages; formacetyl and thioformacetyl linkages; alkene containing backbones;

sulfamate backbones; methylenemethylimino ($-\text{CH}_2\text{-N}(\text{CH}_3)\text{-O-CH}_2\text{-}$) and methylenehydrazino linkages; sulfonate and sulfonamide linkages; amide linkages; and others having mixed N, O, S and CH_2 component parts. In one embodiment, the modified internucleoside linkage is a peptide-based linkage (e.g. aminoethylglycine) to create a peptide nucleic acid or PNA, such as those described in U.S. Patent Nos. 5,539,082; 5,714,331; and 5,719,262. Other suitable modified internucleotide and internucleoside linkages that may be employed in the RNAi constructs of the invention are described in U.S. Patent No. 6,693,187, U.S. Patent No. 9,181,551, U.S. Patent Publication No. 2016/0122761, and Deleavey and Damha, Chemistry and Biology, Vol. 19: 937-954, 2012, all of which are hereby incorporated by reference in their entireties.

[0048] In certain embodiments, the RNAi constructs comprise one or more phosphorothioate internucleotide linkages. The phosphorothioate internucleotide linkages may be present in the sense strand, antisense strand, or both strands of the RNAi constructs. For instance, in some embodiments, the sense strand comprises 1, 2, 3, 4, 5, 6, 7, 8, or more phosphorothioate internucleotide linkages. In other embodiments, the antisense strand comprises 1, 2, 3, 4, 5, 6, 7, 8, or more phosphorothioate internucleotide linkages. In still other embodiments, both strands comprise 1, 2, 3, 4, 5, 6, 7, 8, or more phosphorothioate internucleotide linkages. The RNAi constructs can comprise one or more phosphorothioate internucleotide linkages at the 3'-end, the 5'-end, or both the 3'- and 5'-ends of the sense strand, the antisense strand, or both strands. For instance, in certain embodiments, the RNAi construct comprises about 1 to about 6 or more (e.g., about 1, 2, 3, 4, 5, 6 or more) consecutive phosphorothioate internucleotide linkages at the 3'-end of the sense strand, the antisense strand, or both strands. In other embodiments, the RNAi construct comprises about 1 to about 6 or more (e.g., about 1, 2, 3, 4, 5, 6 or more) consecutive phosphorothioate internucleotide linkages at the 5'-end of the sense strand, the antisense strand, or both strands. In one embodiment, the RNAi construct comprises a single phosphorothioate internucleotide linkage at the 3' end of the sense strand and a single phosphorothioate internucleotide linkage at the 3' end of the antisense strand. In another embodiment, the RNAi construct comprises two consecutive phosphorothioate internucleotide linkages at the 3' end of the antisense strand (i.e. a phosphorothioate internucleotide linkage at the first and second internucleotide linkages at the 3' end of the antisense strand). In another embodiment, the RNAi construct comprises two consecutive phosphorothioate internucleotide linkages at both the 3' and

5' ends of the antisense strand. In yet another embodiment, the RNAi construct comprises two consecutive phosphorothioate internucleotide linkages at both the 3' and 5' ends of the antisense strand and two consecutive phosphorothioate internucleotide linkages at the 5' end of the sense strand. In still another embodiment, the RNAi construct comprises two consecutive phosphorothioate internucleotide linkages at both the 3' and 5' ends of the antisense strand and two consecutive phosphorothioate internucleotide linkages at both the 3' and 5' ends of the sense strand (i.e. a phosphorothioate internucleotide linkage at the first and second internucleotide linkages at both the 5' and 3' ends of the antisense strand and a phosphorothioate internucleotide linkage at the first and second internucleotide linkages at both the 5' and 3' ends of the sense strand). In any of the embodiments in which one or both strands comprises one or more phosphorothioate internucleotide linkages, the remaining internucleotide linkages within the strands can be the natural 3' to 5' phosphodiester linkages. For instance, in some embodiments, each internucleotide linkage of the sense and antisense strands is selected from phosphodiester and phosphorothioate, wherein at least one internucleotide linkage is a phosphorothioate.

[0049] In embodiments in which the RNAi construct comprises a nucleotide overhang, two or more of the unpaired nucleotides in the overhang can be connected by a phosphorothioate internucleotide linkage. In certain embodiments, all the unpaired nucleotides in a nucleotide overhang at the 3' end of the antisense strand and/or the sense strand are connected by phosphorothioate internucleotide linkages. In other embodiments, all the unpaired nucleotides in a nucleotide overhang at the 5' end of the antisense strand and/or the sense strand are connected by phosphorothioate internucleotide linkages. In still other embodiments, all the unpaired nucleotides in any nucleotide overhang are connected by phosphorothioate internucleotide linkages.

[0050] In certain embodiments, the modified nucleotides incorporated into one or both of the strands of the RNAi constructs of the invention have a modification of the nucleobase (also referred to herein as "base"). A "modified nucleobase" or "modified base" refers to a base other than the naturally occurring purine bases adenine (A) and guanine (G) and pyrimidine bases thymine (T), cytosine (C), and uracil (U). Modified nucleobases can be synthetic or naturally occurring modifications and include, but are not limited to, universal bases, 5-methylcytosine (5-

me-C), 5-hydroxymethyl cytosine, xanthine (X), hypoxanthine (I), 2-aminoadenine, 6-methyladenine, 6-methylguanine, and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 2-thiouracil, 2-thiothymine and 2-thiocytosine, 5-halouracil and cytosine, 5-propynyl uracil and cytosine, 6-azo uracil, cytosine and thymine, 5-uracil (pseudouracil), 4-thiouracil, 8-halo, 8-amino, 8-thiol, 8-thioalkyl, 8-hydroxyl and other 8-substituted adenines and guanines, 5-halo, particularly 5-bromo, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine and 7-methyladenine, 8-azaguanine and 8-azaadenine, 7-deazaguanine and 7-daazaadenine and 3-deazaguanine and 3-deazaadenine.

[0051] In some embodiments, the modified base is a universal base. A "universal base" refers to a base analog that indiscriminately forms base pairs with all of the natural bases in RNA and DNA without altering the double helical structure of the resulting duplex region. Universal bases are known to those of skill in the art and include, but are not limited to, inosine, C-phenyl, C-naphthyl and other aromatic derivatives, azole carboxamides, and nitroazole derivatives, such as 3-nitropyrrole, 4-nitroindole, 5-nitroindole, and 6-nitroindole.

[0052] Other suitable modified bases that can be incorporated into the RNAi constructs of the invention include those described in Herdewijn, *Antisense Nucleic Acid Drug Dev.*, Vol. 10:297-310, 2000 and Peacock et al., *J. Org. Chem.*, Vol. 76: 7295-7300, 2011, both of which are hereby incorporated by reference in their entireties. The skilled person is well aware that guanine, cytosine, adenine, thymine, and uracil may be replaced by other nucleobases, such as the modified nucleobases described above, without substantially altering the base pairing properties of a polynucleotide comprising a nucleotide bearing such replacement nucleobase.

[0053] In some embodiments of the RNAi constructs of the invention, the 5' end of the sense strand, antisense strand, or both the antisense and sense strands comprises a phosphate moiety. As used herein, the term "phosphate moiety" refers to a terminal phosphate group that includes unmodified phosphates ($-\text{O}-\text{P}=\text{O}(\text{OH})\text{OH}$) as well as modified phosphates. Modified phosphates include phosphates in which one or more of the O and OH groups is replaced with H, O, S, N(R) or alkyl where R is H, an amino protecting group or unsubstituted or substituted alkyl. Exemplary phosphate moieties include, but are not limited to, 5'-monophosphate; 5'-diphosphate; 5'-triphosphate; 5'-guanosine cap (7-methylated or non-methylated); 5'-adenosine cap or any other

modified or unmodified nucleotide cap structure; 5'-monothiophosphate (phosphorothioate); 5'-monodithiophosphate (phosphorodithioate); 5'-alpha-thiotriphosphate; 5'-gamma-thiotriphosphate, 5'-phosphoramidates; 5'-vinylphosphates; 5'-alkylphosphonates (e.g., alkyl=methyl, ethyl, isopropyl, propyl, etc.); and 5'-alkyletherphosphonates (e.g., alkylether=methoxymethyl, ethoxymethyl, etc.).

[0054] The modified nucleotides that can be incorporated into the RNAi constructs of the invention may have more than one chemical modification described herein. For instance, the modified nucleotide may have a modification to the ribose sugar as well as a modification to the nucleobase. By way of example, a modified nucleotide may comprise a 2' sugar modification (e.g. 2'-fluoro or 2'-methyl) and comprise a modified base (e.g. 5-methyl cytosine or pseudouracil). In other embodiments, the modified nucleotide may comprise a sugar modification in combination with a modification to the 5' phosphate that would create a modified internucleotide or internucleoside linkage when the modified nucleotide was incorporated into a polynucleotide. For instance, in some embodiments, the modified nucleotide may comprise a sugar modification, such as a 2'-fluoro modification, a 2'-O-methyl modification, or a bicyclic sugar modification, as well as a 5' phosphorothioate group. Accordingly, in some embodiments, one or both strands of the RNAi constructs of the invention comprise a combination of 2' modified nucleotides or BNAs and phosphorothioate internucleotide linkages. In certain embodiments, both the sense and antisense strands of the RNAi constructs of the invention comprise a combination of 2'-fluoro modified nucleotides, 2'-O-methyl modified nucleotides, and phosphorothioate internucleotide linkages. Exemplary RNAi constructs comprising modified nucleotides and internucleotide linkages are shown in Table 2.

Function of RNAi constructs

[0055] Preferably, the RNAi constructs of the invention reduce or inhibit the expression of PNPLA3 in cells, particularly liver cells. Accordingly, in one embodiment, the present invention provides a method of reducing PNPLA3 expression in a cell by contacting the cell with any RNAi construct described herein. The cell may be in vitro or in vivo. PNPLA3 expression can be assessed by measuring the amount or level of PNPLA3 mRNA, PNPLA3 protein, or another biomarker linked to PNPLA3 expression. The reduction of PNPLA3 expression in cells or animals treated

with an RNAi construct of the invention can be determined relative to the PNPLA3 expression in cells or animals not treated with the RNAi construct or treated with a control RNAi construct. For instance, in some embodiments, reduction of PNPLA3 expression is assessed by (a) measuring the amount or level of PNPLA3 mRNA in liver cells treated with a RNAi construct of the invention, (b) measuring the amount or level of PNPLA3 mRNA in liver cells treated with a control RNAi construct (e.g., RNAi agent directed to a RNA molecule not expressed in liver cells or a RNAi construct having a nonsense or scrambled sequence) or no construct, and (c) comparing the measured PNPLA3 mRNA levels from treated cells in (a) to the measured PNPLA3 mRNA levels from control cells in (b). The PNPLA3 mRNA levels in the treated cells and controls cells can be normalized to RNA levels for a control gene (e.g. 18S ribosomal RNA) prior to comparison. PNPLA3 mRNA levels can be measured by a variety of methods, including Northern blot analysis, nuclease protection assays, fluorescence in situ hybridization (FISH), reverse-transcriptase (RT)-PCR, real-time RT-PCR, quantitative PCR, and the like.

[0056] In other embodiments, reduction of PNPLA3 expression is assessed by (a) measuring the amount or level of PNPLA3 protein in liver cells treated with a RNAi construct of the invention, (b) measuring the amount or level of PNPLA3 protein in liver cells treated with a control RNAi construct (e.g. RNAi agent directed to a RNA molecule not expressed in liver cells or a RNAi construct having a nonsense or scrambled sequence) or no construct, and (c) comparing the measured PNPLA3 protein levels from treated cells in (a) to the measured PNPLA3 protein levels from control cells in (b). Methods of measuring PNPLA3 protein levels are known to those of skill in the art, and include Western Blots, immunoassays (e.g. ELISA), and flow cytometry. An exemplary immunoassay-based method for assessing PNPLA3 protein expression is described in Example 2. Example 3 describes an exemplary method for measuring PNPLA3 mRNA using RNA FISH. Any method capable of measuring PNPLA3 mRNA or protein can be used to assess the efficacy of the RNAi constructs of the invention.

[0057] In some embodiments, the methods to assess PNPLA3 expression levels are performed in vitro in cells that natively express PNPLA3 (e.g. liver cells) or cells that have been engineered to express PNPLA3. In certain embodiments, the methods are performed in vitro in liver cells. Suitable liver cells include, but are not limited to, primary hepatocytes (e.g. human, non-human

primate, or rodent hepatocytes), HepAD38 cells, HuH-6 cells, HuH-7 cells, HuH-5-2 cells, BNLCL2 cells, Hep3B cells, or HepG2 cells. In one embodiment, the liver cells are Hep3B cells. In another embodiment, the liver cells are HepG2 cells.

[0058] In other embodiments, the methods to assess PNPLA3 expression levels are performed in vivo. The RNAi constructs and any control RNAi constructs can be administered to an animal (e.g. rodent or non-human primate) and PNPLA3 mRNA or protein levels assessed in liver tissue harvested from the animal following treatment. Alternatively or additionally, a biomarker or functional phenotype associated with PNPLA3 expression can be assessed in the treated animals.

[0059] In certain embodiments, expression of PNPLA3 is reduced in liver cells by at least 10%, at least 15%, at least 20%, at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, or at least 50% by an RNAi construct of the invention. In some embodiments, expression of PNPLA3 is reduced in liver cells by at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, or at least 85% by an RNAi construct of the invention. In other embodiments, the expression of PNPLA3 is reduced in liver cells by about 90% or more, e.g., 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more by an RNAi construct of the invention. The percent reduction of PNPLA3 expression can be measured by any of the methods described herein as well as others known in the art. For instance, in certain embodiments, the RNAi constructs of the invention inhibit at least 45% of PNPLA3 expression at 5 nM in Hep3B cells (contains wild type PNPLA3) in vitro. In related embodiments, the RNAi constructs of the invention inhibit at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, or at least 75% of PNPLA3 expression at 5 nM in Hep3B cells in vitro. In other embodiments, the RNAi constructs of the invention inhibit at least 80%, at least 85%, at least 90%, at least 92%, at least 94%, at least 96%, or at least 98% of PNPLA3 expression at 5 nM in Hep3B cells in vitro. In certain embodiments, the RNAi constructs of the invention inhibit at least 45% of PNPLA3 expression at 5 nM in HepG2 cells (contains the PNPLA3-rs738409-rs738408 double minor allele) in vitro. In related embodiments, the RNAi constructs of the invention inhibit at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, or at least 75% of PNPLA3 expression at 5 nM in HepG2 cells in vitro. In other embodiments, the RNAi constructs of the invention inhibit at least 80%, at least 85%, at least 90%, at least 92%, at least 94%, at least 96%, or at least 98% of PNPLA3 expression at 5 nM in HepG2 cells in vitro. In certain embodiments, the RNAi constructs of the invention inhibit at least 45%

of PNPLA3 expression at 5 nM in CHO transfected cells expressing human PNPLA3 I148I or I148M cells in vitro. In related embodiments, the RNAi constructs of the invention inhibit at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, or at least 75% of PNPLA3 expression at 5 nM in CHO transfected cells expressing human PNPLA3 I148I or I148M in vitro. In other embodiments, the RNAi constructs of the invention inhibit at least 80%, at least 85%, at least 90%, at least 92%, at least 94%, at least 96%, or at least 98% of PNPLA3 expression at 5 nM in CHO transfected cells expressing human PNPLA3 I148I or I148M in vitro. Reduction of PNPLA3 can be measured using a variety of techniques including RNA FISH or droplet digital PCR, as described in Examples 2 and 3.

[0060] In some embodiments, an IC₅₀ value is calculated to assess the potency of an RNAi construct of the invention for inhibiting PNPLA3 expression in liver cells. An "IC₅₀ value" is the dose/concentration required to achieve 50% inhibition of a biological or biochemical function. The IC₅₀ value of any particular substance or antagonist can be determined by constructing a dose-response curve and examining the effect of different concentrations of the substance or antagonist on expression levels or functional activity in any assay. IC₅₀ values can be calculated for a given antagonist or substance by determining the concentration needed to inhibit half of the maximum biological response or native expression levels. Thus, the IC₅₀ value for any RNAi construct can be calculated by determining the concentration of the RNAi construct needed to inhibit half of the native PNPLA3 expression level in liver cells (e.g. PNPLA3 expression level in control liver cells) in any assay, such as the immunoassay or RNA FISH assay or droplet digital PCR assays described in the Examples. The RNAi constructs of the invention may inhibit PNPLA3 expression in liver cells (e.g. Hep3B cells) with an IC₅₀ of less than about 20 nM. For example, the RNAi constructs inhibit PNPLA3 expression in liver cells with an IC₅₀ of about 0.001 nM to about 20 nM, about 0.001 nM to about 10 nM, about 0.001 nM to about 5 nM, about 0.001 nM to about 1 nM, about 0.1 nM to about 10 nM, about 0.1 nM to about 5 nM, or about 0.1 nM to about 1 nM. In certain embodiments, the RNAi construct inhibits PNPLA3 expression in liver cells (e.g. Hep3B cells) with an IC₅₀ of about 1 nM to about 10 nM. The RNAi constructs of the invention may inhibit PNPLA3 expression in liver cells (e.g. HepG2 cells) with an IC₅₀ of less than about 20 nM. For example, the RNAi constructs inhibit PNPLA3 expression in liver cells with an IC₅₀ of about 0.001 nM to about 20 nM, about 0.001 nM to about 10 nM, about 0.001 nM to about 5 nM, about

0.001 nM to about 1 nM, about 0.1 nM to about 10 nM, about 0.1 nM to about 5 nM, or about 0.1 nM to about 1 nM. In certain embodiments, the RNAi construct inhibits PNPLA3 expression in liver cells (e.g. HepG2 cells) with an IC₅₀ of about 1 nM to about 10 nM. The RNAi constructs of the invention may inhibit PNPLA3 expression in liver cells (e.g. CHO transfected cells expressing human PNPLA3 I148I or I148M) with an IC₅₀ of less than about 20 nM. For example, the RNAi constructs inhibit PNPLA3 expression in liver cells with an IC₅₀ of about 0.001 nM to about 20 nM, about 0.001 nM to about 10 nM, about 0.001 nM to about 5 nM, about 0.001 nM to about 1 nM, about 0.1 nM to about 10 nM, about 0.1 nM to about 5 nM, or about 0.1 nM to about 1 nM. In certain embodiments, the RNAi construct inhibits PNPLA3 expression in liver cells (e.g. CHO transfected cells expressing human PNPLA3 I148I or I148M) with an IC₅₀ of about 1 nM to about 10 nM.

[0061] The RNAi constructs of the invention can readily be made using techniques known in the art, for example, using conventional nucleic acid solid phase synthesis. The polynucleotides of the RNAi constructs can be assembled on a suitable nucleic acid synthesizer utilizing standard nucleotide or nucleoside precursors (e.g. phosphoramidites). Automated nucleic acid synthesizers are sold commercially by several vendors, including DNA/RNA synthesizers from Applied Biosystems (Foster City, CA), MerMade synthesizers from BioAutomation (Irving, TX), and OligoPilot synthesizers from GE Healthcare Life Sciences (Pittsburgh, PA).

[0062] The 2' silyl protecting group can be used in conjunction with acid labile dimethoxytrityl (DMT) at the 5' position of ribonucleosides to synthesize oligonucleotides via phosphoramidite chemistry. Final deprotection conditions are known not to significantly degrade RNA products. All syntheses can be conducted in any automated or manual synthesizer on large, medium, or small scale. The syntheses may also be carried out in multiple well plates, columns, or glass slides.

[0063] The 2'-O-silyl group can be removed via exposure to fluoride ions, which can include any source of fluoride ion, e.g., those salts containing fluoride ion paired with inorganic counterions, e.g., cesium fluoride and potassium fluoride or those salts containing fluoride ion paired with an organic counterion, e.g., a tetraalkylammonium fluoride. A crown ether catalyst can be utilized in combination with the inorganic fluoride in the deprotection reaction. Preferred

fluoride ion source are tetrabutylammonium fluoride or aminohydrofluorides (e.g., combining aqueous HF with triethylamine in a dipolar aprotic solvent, e.g., dimethylformamide).

[0064] The choice of protecting groups for use on the phosphite triesters and phosphotriesters can alter the stability of the triesters towards fluoride. Methyl protection of the phosphotriester or phosphitetriester can stabilize the linkage against fluoride ions and improve process yields.

[0065] Since ribonucleosides have a reactive 2' hydroxyl substituent, it can be desirable to protect the reactive 2' position in RNA with a protecting group that is orthogonal to a 5'-O-dimethoxytrityl protecting group, e.g., one stable to treatment with acid. Silyl protecting groups meet this criterion and can be readily removed in a final fluoride deprotection step that can result in minimal RNA degradation.

[0066] Tetrazole catalysts can be used in the standard phosphoramidite coupling reaction. Preferred catalysts include, e.g., tetrazole, S-ethyl-tetrazole, benzylthiotetrazole, p-nitrophenyltetrazole.

[0067] As can be appreciated by the skilled artisan, further methods of synthesizing the RNAi constructs described herein will be evident to those of ordinary skill in the art. Additionally, the various synthetic steps may be performed in an alternate sequence or order to give the desired compounds. Other synthetic chemistry transformations, protecting groups (e.g., for hydroxyl, amino, etc. present on the bases) and protecting group methodologies (protection and deprotection) useful in synthesizing the RNAi constructs described herein are known in the art and include, for example, those such as described in R. Larock, *Comprehensive Organic Transformations*, VCH Publishers (1989); T. W. Greene and P. G. M. Wuts, *Protective Groups in Organic Synthesis*, 2d. Ed., John Wiley and Sons (1991); L. Fieser and M. Fieser, *Fieser and Fieser's Reagents for Organic Synthesis*, John Wiley and Sons (1994); and L. Paquette, ed., *Encyclopedia of Reagents for Organic Synthesis*, John Wiley and Sons (1995), and subsequent editions thereof. Custom synthesis of RNAi agents is also available from several commercial vendors, including Dharmacon, Inc. (Lafayette, CO), AxoLabs GmbH (Kulmbach, Germany), and Ambion, Inc. (Foster City, CA).

[0068] The RNAi constructs of the invention may comprise a ligand. As used herein, a "ligand" refers to any compound or molecule that is capable of interacting with another compound

or molecule, directly or indirectly. The interaction of a ligand with another compound or molecule may elicit a biological response (e.g. initiate a signal transduction cascade, induce receptor mediated endocytosis) or may just be a physical association. The ligand can modify one or more properties of the double-stranded RNA molecule to which is attached, such as the pharmacodynamic, pharmacokinetic, binding, absorption, cellular distribution, cellular uptake, charge and/or clearance properties of the RNA molecule.

[0069] The ligand may comprise a serum protein (e.g., human serum albumin, low-density lipoprotein, globulin), a cholesterol moiety, a vitamin (biotin, vitamin E, vitamin B12), a folate moiety, a steroid, a bile acid (e.g. cholic acid), a fatty acid (e.g., palmitic acid, myristic acid), a carbohydrate (e.g., a dextran, pullulan, chitin, chitosan, inulin, cyclodextrin or hyaluronic acid), a glycoside, a phospholipid, or antibody or binding fragment thereof (e.g. antibody or binding fragment that targets the RNAi construct to a specific cell type, such as liver). Other examples of ligands include dyes, intercalating agents (e.g. acridines), cross-linkers (e.g. psoralene, mitomycin C), porphyrins (TPPC4, texaphyrin, Sapphyrin), polycyclic aromatic hydrocarbons (e.g., phenazine, dihydrophenazine), artificial endonucleases (e.g. EDTA), lipophilic molecules, e.g., adamantane acetic acid, 1-pyrene butyric acid, dihydrotestosterone, 1,3-BisO(hexadecyl)glycerol, geranyloxyhexyl group, hexadecylglycerol, borneol, menthol, 1,3-propanediol, heptadecyl group, 03-(oleoyl)lithocholic acid, 03-(oleoyl)cholenic acid, dimethoxytrityl, or phenoxazine), peptides (e.g., antennapedia peptide, Tat peptide, RGD peptides), alkylating agents, polymers, such as polyethylene glycol (PEG) (e.g., PEG-40K), poly amino acids, and polyamines (e.g. spermine, spermidine).

[0070] In certain embodiments, the ligands have endosomolytic properties. The endosomolytic ligands promote the lysis of the endosome and/or transport of the RNAi construct of the invention, or its components, from the endosome to the cytoplasm of the cell. The endosomolytic ligand may be a polycationic peptide or peptidomimetic which shows pH dependent membrane activity and fusogenicity. In one embodiment, the endosomolytic ligand assumes its active conformation at endosomal pH. The "active" conformation is that conformation in which the endosomolytic ligand promotes lysis of the endosome and/or transport of the RNAi construct of the invention, or its components, from the endosome to the cytoplasm of the cell.

Exemplary endosomolytic ligands include the GALA peptide (Subbarao et al., *Biochemistry*, Vol. 26: 2964-2972, 1987), the EALA peptide (Vogel et al., *J. Am. Chem. Soc.*, Vol. 118: 1581-1586, 1996), and their derivatives (Turk et al., *Biochem. Biophys. Acta*, Vol. 1559: 56-68, 2002). In one embodiment, the endosomolytic component may contain a chemical group (e.g., an amino acid) which will undergo a change in charge or protonation in response to a change in pH. The endosomolytic component may be linear or branched.

[0071] In some embodiments, the ligand comprises a lipid or other hydrophobic molecule. In one embodiment, the ligand comprises a cholesterol moiety or other steroid. Cholesterol conjugated oligonucleotides have been reported to be more active than their unconjugated counterparts (Manoharan, *Antisense Nucleic Acid Drug Development*, Vol. 12: 103-228, 2002). Ligands comprising cholesterol moieties and other lipids for conjugation to nucleic acid molecules have also been described in U.S. Patent Nos. 7,851,615; 7,745,608; and 7,833,992, all of which are hereby incorporated by reference in their entireties. In another embodiment, the ligand comprises a folate moiety. Polynucleotides conjugated to folate moieties can be taken up by cells via a receptor-mediated endocytosis pathway. Such folate-polynucleotide conjugates are described in U.S. Patent No. 8,188,247, which is hereby incorporated by reference in its entirety.

[0072] Given that PNPLA3 is expressed in liver cells (e.g. hepatocytes), in certain embodiments, it is desirable to specifically deliver the RNAi construct to those liver cells. In some embodiments, RNAi constructs can be specifically targeted to the liver by employing ligands that bind to or interact with proteins expressed on the surface of liver cells. For example, in certain embodiments, the ligands may comprise antigen binding proteins (e.g. antibodies or binding fragments thereof (e.g. Fab, scFv)) that specifically bind to a receptor expressed on hepatocytes.

[0073] In certain embodiments, the ligand comprises a carbohydrate. A "carbohydrate" refers to a compound made up of one or more monosaccharide units having at least 6 carbon atoms (which can be linear, branched or cyclic) with an oxygen, nitrogen or sulfur atom bonded to each carbon atom. Carbohydrates include, but are not limited to, the sugars (e.g., monosaccharides, disaccharides, trisaccharides, tetrasaccharides, and oligosaccharides containing from about 4, 5, 6, 7, 8, or 9 monosaccharide units), and polysaccharides, such as starches, glycogen, cellulose and polysaccharide gums. In some embodiments, the carbohydrate incorporated into the ligand is a

monosaccharide selected from a pentose, hexose, or heptose and di- and tri-saccharides including such monosaccharide units. In other embodiments, the carbohydrate incorporated into the ligand is an amino sugar, such as galactosamine, glucosamine, N-acetylgalactosamine, and N-acetylglucosamine.

[0074] In some embodiments, the ligand comprises a hexose or hexosamine. The hexose may be selected from glucose, galactose, mannose, fucose, or fructose. The hexosamine may be selected from fructosamine, galactosamine, glucosamine, or mannosamine. In certain embodiments, the ligand comprises glucose, galactose, galactosamine, or glucosamine. In one embodiment, the ligand comprises glucose, glucosamine, or N-acetylglucosamine. In another embodiment, the ligand comprises galactose, galactosamine, or N-acetyl-galactosamine. In particular embodiments, the ligand comprises N-acetyl-galactosamine. Ligands comprising glucose, galactose, and N-acetyl-galactosamine (GalNAc) are particularly effective in targeting compounds to liver cells. See, e.g., D'Souza and Devarajan, J. Control Release, Vol. 203: 126-139, 2015. Examples of GalNAc- or galactose-containing ligands that can be incorporated into the RNAi constructs of the invention are described in U.S. Patent Nos. 7,491,805; 8,106,022; and 8,877,917; U.S. Patent Publication No. 20030130186; and WIPO Publication No. WO2013166155, all of which are hereby incorporated by reference in their entireties.

[0075] In certain embodiments, the ligand comprises a multivalent carbohydrate moiety. As used herein, a "multivalent carbohydrate moiety" refers to a moiety comprising two or more carbohydrate units capable of independently binding or interacting with other molecules. For example, a multivalent carbohydrate moiety comprises two or more binding domains comprised of carbohydrates that can bind to two or more different molecules or two or more different sites on the same molecule. The valency of the carbohydrate moiety denotes the number of individual binding domains within the carbohydrate moiety. For instance, the terms "monovalent," "bivalent," "trivalent," and "tetravalent" with reference to the carbohydrate moiety refer to carbohydrate moieties with one, two, three, and four binding domains, respectively. The multivalent carbohydrate moiety may comprise a multivalent lactose moiety, a multivalent galactose moiety, a multivalent glucose moiety, a multivalent N-acetyl-galactosamine moiety, a multivalent N-acetyl-glucosamine moiety, a multivalent mannose moiety, or a multivalent fucose moiety. In

some embodiments, the ligand comprises a multivalent galactose moiety. In other embodiments, the ligand comprises a multivalent N-acetyl-galactosamine moiety. In these and other embodiments, the multivalent carbohydrate moiety is bivalent, trivalent, or tetravalent. In such embodiments, the multivalent carbohydrate moiety can be bi-antennary or tri-antennary. In one particular embodiment, the multivalent N-acetyl-galactosamine moiety is trivalent or tetravalent. In another particular embodiment, the multivalent galactose moiety is trivalent or tetravalent. Exemplary trivalent and tetravalent GalNAc-containing ligands for incorporation into the RNAi constructs of the invention are described in detail below.

[0076] The ligand can be attached or conjugated to the RNA molecule of the RNAi construct directly or indirectly. For instance, in some embodiments, the ligand is covalently attached directly to the sense or antisense strand of the RNAi construct. In other embodiments, the ligand is covalently attached via a linker to the sense or antisense strand of the RNAi construct. The ligand can be attached to nucleobases, sugar moieties, or internucleotide linkages of polynucleotides (e.g. sense strand or antisense strand) of the RNAi constructs of the invention. Conjugation or attachment to purine nucleobases or derivatives thereof can occur at any position including, endocyclic and exocyclic atoms. In certain embodiments, the 2-, 6-, 7-, or 8-positions of a purine nucleobase are attached to a ligand. Conjugation or attachment to pyrimidine nucleobases or derivatives thereof can also occur at any position. In some embodiments, the 2-, 5-, and 6-positions of a pyrimidine nucleobase can be attached to a ligand. Conjugation or attachment to sugar moieties of nucleotides can occur at any carbon atom. Example carbon atoms of a sugar moiety that can be attached to a ligand include the 2', 3', and 5' carbon atoms. The 1' position can also be attached to a ligand, such as in an a basic residue. Internucleotide linkages can also support ligand attachments. For phosphorus-containing linkages (e.g., phosphodiester, phosphorothioate, phosphorodithiotate, phosphoroamidate, and the like), the ligand can be attached directly to the phosphorus atom or to an O, N, or S atom bound to the phosphorus atom. For amine- or amide-containing internucleoside linkages (e.g., PNA), the ligand can be attached to the nitrogen atom of the amine or amide or to an adjacent carbon atom.

[0077] In certain embodiments, the ligand may be attached to the 3' or 5' end of either the sense or antisense strand. In certain embodiments, the ligand is covalently attached to the 5' end of the

sense strand. In other embodiments, the ligand is covalently attached to the 3' end of the sense strand. For example, in some embodiments, the ligand is attached to the 3'-terminal nucleotide of the sense strand. In certain such embodiments, the ligand is attached at the 3'-position of the 3'-terminal nucleotide of the sense strand. In alternative embodiments, the ligand is attached near the 3' end of the sense strand, but before one or more terminal nucleotides (i.e. before 1, 2, 3, or 4 terminal nucleotides). In some embodiments, the ligand is attached at the 2'-position of the sugar of the 3'-terminal nucleotide of the sense strand.

[0078] In certain embodiments, the ligand is attached to the sense or antisense strand via a linker. A "linker" is an atom or group of atoms that covalently joins a ligand to a polynucleotide component of the RNAi construct. The linker may be from about 1 to about 30 atoms in length, from about 2 to about 28 atoms in length, from about 3 to about 26 atoms in length, from about 4 to about 24 atoms in length, from about 6 to about 20 atoms in length, from about 7 to about 20 atoms in length, from about 8 to about 20 atoms in length, from about 8 to about 18 atoms in length, from about 10 to about 18 atoms in length, and from about 12 to about 18 atoms in length. In some embodiments, the linker may comprise a bifunctional linking moiety, which generally comprises an alkyl moiety with two functional groups. One of the functional groups is selected to bind to the compound of interest (e.g. sense or antisense strand of the RNAi construct) and the other is selected to bind essentially any selected group, such as a ligand as described herein. In certain embodiments, the linker comprises a chain structure or an oligomer of repeating units, such as ethylene glycol or amino acid units. Examples of functional groups that are typically employed in a bifunctional linking moiety include, but are not limited to, electrophiles for reacting with nucleophilic groups and nucleophiles for reacting with electrophilic groups. In some embodiments, bifunctional linking moieties include amino, hydroxyl, carboxylic acid, thiol, unsaturations (e.g., double or triple bonds), and the like.

[0079] Linkers that may be used to attach a ligand to the sense or antisense strand in the RNAi constructs of the invention include, but are not limited to, pyrrolidine, 8-amino-3,6-di oxaoctanoic acid, succinimidy1 4-(N-maleimidomethyl)cyclohexane-1-carboxylate, 6-aminohexanoic acid, substituted C1-C10 alkyl, substituted or unsubstituted C2-C10 alkenyl or substituted or unsubstituted C2-C10 alkynyl. Preferred substituent groups for such linkers include, but are not

limited to, hydroxyl, amino, alkoxy, carboxy, benzyl, phenyl, nitro, thiol, thioalkoxy, halogen, alkyl, aryl, alkenyl and alkynyl.

[0080] In certain embodiments, the linkers are cleavable. A cleavable linker is one which is sufficiently stable outside the cell, but which upon entry into a target cell is cleaved to release the two parts the linker is holding together. In some embodiments, the cleavable linker is cleaved at least 10 times, 20 times, 30 times, 40 times, 50 times, 60 times, 70 times, 80 times, 90 times, or more, or at least 100 times faster in the target cell or under a first reference condition (which can, e.g., be selected to mimic or represent intracellular conditions) than in the blood of a subject, or under a second reference condition (which can, e.g., be selected to mimic or represent conditions found in the blood or serum).

[0081] Cleavable linkers are susceptible to cleavage agents, e.g., pH, redox potential or the presence of degradative molecules. Generally, cleavage agents are more prevalent or found at higher levels or activities inside cells than in serum or blood. Examples of such degradative agents include: redox agents which are selected for particular substrates or which have no substrate specificity, including, e.g., oxidative or reductive enzymes or reductive agents such as mercaptans, present in cells, that can degrade a redox cleavable linker by reduction; esterases; endosomes or agents that can create an acidic environment, e.g., those that result in a pH of five or lower; enzymes that can hydrolyze or degrade an acid cleavable linker by acting as a general acid, peptidases (which can be substrate specific), and phosphatases.

[0082] A cleavable linker may comprise a moiety that is susceptible to pH. The pH of human serum is 7.4, while the average intracellular pH is slightly lower, ranging from about 7.1-7.3. Endosomes have a more acidic pH, in the range of 5.5-6.0, and lysosomes have an even more acidic pH at around 5.0. Some linkers will have a cleavable group that is cleaved at a preferred pH, thereby releasing the RNA molecule from the ligand inside the cell, or into the desired compartment of the cell.

[0083] A linker can include a cleavable group that is cleavable by a particular enzyme. The type of cleavable group incorporated into a linker can depend on the cell to be targeted. For example, liver-targeting ligands can be linked to RNA molecules through a linker that includes an ester group. Liver cells are rich in esterases, and therefore the linker will be cleaved more

efficiently in liver cells than in cell types that are not esterase-rich. Other types of cells rich in esterases include cells of the lung, renal cortex, and testis. Linkers that contain peptide bonds can be used when targeting cells rich in peptidases, such as liver cells and synoviocytes.

[0084] In general, the suitability of a candidate cleavable linker can be evaluated by testing the ability of a degradative agent (or condition) to cleave the candidate linker. It will also be desirable to also test the candidate cleavable linker for the ability to resist cleavage in the blood or when in contact with other non-target tissue. Thus, one can determine the relative susceptibility to cleavage between a first and a second condition, where the first is selected to be indicative of cleavage in a target cell and the second is selected to be indicative of cleavage in other tissues or biological fluids, e.g., blood or serum. The evaluations can be carried out in cell free systems, in cells, in cell culture, in organ or tissue culture, or in whole animals. It may be useful to make initial evaluations in cell-free or culture conditions and to confirm by further evaluations in whole animals. In some embodiments, useful candidate linkers are cleaved at least 2, 4, 10, 20, 50, 70, or 100 times faster in the cell (or under in vitro conditions selected to mimic intracellular conditions) as compared to blood or serum (or under in vitro conditions selected to mimic extracellular conditions).

[0085] In other embodiments, redox cleavable linkers are utilized. Redox cleavable linkers are cleaved upon reduction or oxidation. An example of reductively cleavable group is a disulfide linking group (-S-S-). To determine if a candidate cleavable linker is a suitable "reductively cleavable linker," or for example is suitable for use with a particular RNAi construct and particular ligand, one can use one or more methods described herein. For example, a candidate linker can be evaluated by incubation with dithiothreitol (DTT), or other reducing agent known in the art, which mimics the rate of cleavage that would be observed in a cell, e.g., a target cell. The candidate linkers can also be evaluated under conditions which are selected to mimic blood or serum conditions. In a specific embodiment, candidate linkers are cleaved by at most 10% in the blood. In other embodiments, useful candidate linkers are degraded at least 2, 4, 10, 20, 50, 70, or 100 times faster in the cell (or under in vitro conditions selected to mimic intracellular conditions) as compared to blood (or under in vitro conditions selected to mimic extracellular conditions).

[0086] In yet other embodiments, phosphate-based cleavable linkers are cleaved by agents that degrade or hydrolyze the phosphate group. An example of an agent that hydrolyzes phosphate groups in cells are enzymes, such as phosphatases in cells. Examples of phosphate-based cleavable groups are -O-P(O)(ORk)-O-, -O-P(S)(ORk)-O-, -O-P(S)(SRk)-O-, -S-P(O)(ORk)-O-, -O-P(O)(ORk)-S-, -S-P(O)(ORk)-S-, -O-P(S)(ORk)-S-, -S-P(S)(ORk)-O-, -O-P(O)(Rk)-O-, -O-P(S)(Rk)-O-, -S-P(O)(Rk)-O-, -S-P(S)(Rk)-O-, -S-P(O)(Rk)-S-, -O-P(S)(Rk)-S-. Specific embodiments include -O-P(O)(OH)-O-, -O-P(S)(OH)-O-, -O-P(S)(SH)-O-, -S-P(O)(OH)-O-, -O-P(O)(OH)-S-, -S-P(O)(OH)-S-, -O-P(S)(OH)-S-, -S-P(S)(OH)-O-, -O-P(O)(H)-O-, -O-P(S)(H)-O-, -S-P(O)(H)-O-, -S-P(S)(H)-O-, -S-P(O)(H)-S-, -O-P(S)(H)-S-. Another specific embodiment is -O-P(O)(OH)-O-. These candidate linkers can be evaluated using methods analogous to those described above.

[0087] In other embodiments, the linkers may comprise acid cleavable groups, which are groups that are cleaved under acidic conditions. In some embodiments, acid cleavable groups are cleaved in an acidic environment with a pH of about 6.5 or lower (e.g., about 6.0, 5.5, 5.0, or lower), or by agents, such as enzymes that can act as a general acid. In a cell, specific low pH organelles, such as endosomes and lysosomes, can provide a cleaving environment for acid cleavable groups. Examples of acid cleavable linking groups include, but are not limited to, hydrazones, esters, and esters of amino acids. Acid cleavable groups can have the general formula -C=NN-, C(O)O, or -OC(O). A specific embodiment is when the carbon attached to the oxygen of the ester (the alkoxy group) is an aryl group, substituted alkyl group, or tertiaryalkyl group such as dimethyl, pentyl or t-butyl. These candidates can be evaluated using methods analogous to those described above.

[0088] In other embodiments, the linkers may comprise ester-based cleavable groups, which are cleaved by enzymes, such as esterases and amidases in cells. Examples of ester-based cleavable groups include, but are not limited to, esters of alkylene, alkenylene and alkynylene groups. Ester cleavable groups have the general formula -C(O)O-, or -OC(O) -. These candidate linkers can be evaluated using methods analogous to those described above.

[0089] In further embodiments, the linkers may comprise peptide-based cleavable groups, which are cleaved by enzymes, such as peptidases and proteases in cells. Peptide-based cleavable

groups are peptide bonds formed between amino acids to yield oligopeptides (e.g., dipeptides, tripeptides etc.) and polypeptides. Peptide-based cleavable groups do not include the amide group (-C(O)NH-). The amide group can be formed between any alkylene, alkenylene or alkynylene. A peptide bond is a special type of amide bond formed between amino acids to yield peptides and proteins. The peptide based cleavage group is generally limited to the peptide bond(i.e., the amide bond) formed between amino acids yielding peptides and proteins and does not include the entire amide functional group. Peptide-based cleavable linking groups have the general formula -NHCHRA-C(O)NHCHRB-C(O)-, where RA and RB are the R groups of the two adjacent amino acids. These candidates can be evaluated using methods analogous to those described above.

[0090] Other types of linkers suitable for attaching ligands to the sense or antisense strands in the RNAi constructs of the invention are known in the art and can include the linkers described in U.S. Patent Nos. 7,723,509; 8,017,762; 8,828,956; 8,877,917; and 9,181,551, all of which are hereby incorporated by reference in their entireties.

[0091] In certain embodiments, the ligand covalently attached to the sense or antisense strand of the RNAi constructs of the invention comprises a GalNAc moiety, e.g, a multivalent GalNAc moiety. In some embodiments, the multivalent GalNAc moiety is a trivalent GalNAc moiety and is attached to the 3' end of the sense strand. In other embodiments, the multivalent GalNAc moiety is a trivalent GalNAc moiety and is attached to the 5' end of the sense strand. In yet other embodiments, the multivalent GalNAc moiety is a tetravalent GalNAc moiety and is attached to the 3' end of the sense strand. In still other embodiments, the multivalent GalNAc moiety is a tetravalent GalNAc moiety and is attached to the 5' end of the sense strand.

[0092] In some embodiments, the RNAi constructs of the invention may be delivered to a cell or tissue of interest by administering a vector that encodes and controls the intracellular expression of the RNAi construct. A "vector" (also referred to herein as an "expression vector") is a composition of matter which can be used to deliver a nucleic acid of interest to the interior of a cell. Numerous vectors are known in the art including, but not limited to, linear polynucleotides, polynucleotides associated with ionic or amphiphilic compounds, plasmids, and viruses. Thus, the term "vector" includes an autonomously replicating plasmid or a virus. Examples of viral vectors

include, but are not limited to, adenoviral vectors, adeno-associated viral vectors, retroviral vectors, and the like. A vector can be replicated in a living cell, or it can be made synthetically.

[0093] Generally, a vector for expressing an RNAi construct of the invention will comprise one or more promoters operably linked to sequences encoding the RNAi construct. The phrase "operably linked" or "under transcriptional control" as used herein means that the promoter is in the correct location and orientation in relation to a polynucleotide sequence to control the initiation of transcription by RNA polymerase and expression of the polynucleotide sequence. A "promoter" refers to a sequence recognized by the synthetic machinery of the cell, or introduced synthetic machinery, required to initiate the specific transcription of a gene sequence. Suitable promoters include, but are not limited to, RNA pol I, pol II, HI or U6 RNA pol III, and viral promoters (e.g. human cytomegalovirus (CMV) immediate early gene promoter, the SV40 early promoter, and the Rous sarcoma virus long terminal repeat). In some embodiments, a HI or U6RNA pol III promoter is preferred. The promoter can be a tissue-specific or inducible promoter. Of particular interest are liver-specific promoters, such as promoter sequences from human alpha¹-antitrypsin gene, albumin gene, hemopexin gene, and hepatic lipase gene. Inducible promoters include promoters regulated by ecdysone, estrogen, progesterone, tetracycline, and isopropyl-PD1-thiogalactopyranoside (IPTG).

[0094] In some embodiments in which the RNAi construct comprises a siRNA, the two separate strands (sense and antisense strand) can be expressed from a single vector or two separate vectors. For example, in one embodiment, the sequence encoding the sense strand is operably linked to a promoter on a first vector and the sequence encoding the antisense strand is operably linked to a promoter on a second vector. In such an embodiment, the first and second vectors are co-introduced, e.g., by infection or transfection, into a target cell, such that the sense and antisense strands, once transcribed, will hybridize intracellularly to form the siRNA molecule. In another embodiment, the sense and antisense strands are transcribed from two separate promoters located in a single vector. In some such embodiments, the sequence encoding the sense strand is operably linked to a first promoter and the sequence encoding the antisense strand is operably linked to a second promoter, wherein the first and second promoters are located in a single vector. In one embodiment, the vector comprises a first promoter operably linked to a sequence encoding the

siRNA molecule, and a second promoter operably linked to the same sequence in the opposite direction, such that transcription of the sequence from the first promoter results in the synthesis of the sense strand of the siRNA molecule and transcription of the sequence from the second promoter results in synthesis of the antisense strand of the siRNA molecule.

[0095] In other embodiments in which the RNAi construct comprises a shRNA, a sequence encoding the single, at least partially self-complementary RNA molecule is operably linked to a promoter to produce a single transcript. In some embodiments, the sequence encoding the shRNA comprises an inverted repeat joined by a linker polynucleotide sequence to produce the stem and loop structure of the shRNA following transcription.

[0096] In some embodiments, the vector encoding an RNAi construct of the invention is a viral vector. Various viral vector systems that are suitable to express the RNAi constructs described herein include, but are not limited to, adenoviral vectors, retroviral vectors (e.g., lentiviral vectors, moloney murine leukemia virus), adeno-associated viral vectors; herpes simplex viral vectors; SV 40 vectors; polyoma viral vectors; papilloma viral vectors; picornaviral vectors; and pox viral vectors (e.g. vaccinia virus). In certain embodiments, the viral vector is a retroviral vector (e.g. lentiviral vector).

[0097] Various vectors suitable for use in the invention, methods for inserting nucleic acid sequences encoding siRNA or shRNA molecules into vectors, and methods of delivering the vectors to the cells of interest are within the skill of those in the art. See, e.g., Dornburg, Gene Therap., Vol. 2: 301-310, 1995; Eglitis, Biotechniques, Vol. 6: 608-614, 1988; Miller, HumGene Therap., Vol. 1: 5-14, 1990; Anderson, Nature, Vol. 392: 25-30, 1998; Robinson D A et al., Nat. Genet., Vol. 33: 401-406, 2003; Brummelkamp et al., Science, Vol. 296: 550-553, 2002; Brummelkamp et al., Cancer Cell, Vol. 2: 243-247, 2002; Lee et al., Nat Biotechnol, Vol. 20:500-505, 2002; Miyagishi et al., Nat Biotechnol, Vol. 20: 497-500, 2002; Paddison et al., GenesDev, Vol. 16: 948-958, 2002; Paul et al., Nat Biotechnol, Vol. 20: 505-508, 2002; Sui et al., Proc Natl Acad Sci USA, Vol. 99: 5515-5520, 2002; and Yu et al., Proc Natl Acad Sci USA, Vol. 99:6047-6052, 2002, all of which are hereby incorporated by reference in their entireties.

[0098] The present invention also includes pharmaceutical compositions and formulations comprising the RNAi constructs described herein and pharmaceutically acceptable carriers,

excipients, or diluents. Such compositions and formulations are useful for reducing expression of PNPLA3 in a subject in need thereof. Where clinical applications are contemplated, pharmaceutical compositions and formulations will be prepared in a form appropriate for the intended application. Generally, this will entail preparing compositions that are essentially free of pyrogens, as well as other impurities that could be harmful to humans or animals.

[0099] The phrases "pharmaceutically acceptable" or "pharmacologically acceptable" refer to molecular entities and compositions that do not produce adverse, allergic, or other untoward reactions when administered to an animal or a human. As used herein, "pharmaceutically acceptable carrier, excipient, or diluent" includes solvents, buffers, solutions, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents and the like acceptable for use in formulating pharmaceuticals, such as pharmaceuticals suitable for administration to humans. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the RNAi constructs of the present invention, its use in therapeutic compositions is contemplated. Supplementary active ingredients also can be incorporated into the compositions, provided they do not inactivate the vectors or RNAi constructs of the compositions.

[0100] Compositions and methods for the formulation of pharmaceutical compositions depend on a number of criteria, including, but not limited to, route of administration, type and extent of disease or disorder to be treated, or dose to be administered. In some embodiments, the pharmaceutical compositions are formulated based on the intended route of delivery. For instance, in certain embodiments, the pharmaceutical compositions are formulated for parenteral delivery. Parenteral forms of delivery include intravenous, intraarterial, subcutaneous, intrathecal, intraperitoneal or intramuscular injection or infusion. In one embodiment, the pharmaceutical composition is formulated for intravenous delivery. In such an embodiment, the pharmaceutical composition may include a lipid-based delivery vehicle. In another embodiment, the pharmaceutical composition is formulated for subcutaneous delivery. In such an embodiment, the pharmaceutical composition may include a targeting ligand (e.g. GalNAc containing ligands described herein).

[0101] In some embodiments, the pharmaceutical compositions comprise an effective amount of an RNAi construct described herein. An "effective amount" is an amount sufficient to produce a beneficial or desired clinical result. In some embodiments, an effective amount is an amount sufficient to reduce PNPLA3 expression in hepatocytes of a subject. In some embodiments, an effective amount may be an amount sufficient to only partially reduce PNPLA3 expression, for example, to a level comparable to expression of the wild-type PNPLA3 allele in human heterozygotes. Human heterozygous carriers of loss of function PNPLA3 variant alleles were reported to have lower serum levels of non-HDL cholesterol and a lower risk of coronary artery disease and myocardial infarction as compared to non-carriers (Nioi et al., New England Journal of Medicine, Vol. 374(22):2131-2141, 2016). Thus, without being bound by theory, it is believed that partial reduction of PNPLA3 expression may be sufficient to achieve the beneficial reduction of serum non-HDL cholesterol and reduction of risk of coronary artery disease and myocardial infarction.

[0102] An effective amount of an RNAi construct of the invention may be from about 0.01mg/kg body weight to about 100 mg/kg body weight, about 0.05 mg/kg body weight to about 75mg/kg body weight, about 0.1 mg/kg body weight to about 50 mg/kg body weight, about 1mg/kg to about 30 mg/kg body weight, about 2.5 mg/kg of body weight to about 20 mg/kg bodyweight, or about 5 mg/kg body weight to about 15 mg/kg body weight. In certain embodiments, a single effective dose of an RNAi construct of the invention may be about 0.1 mg/kg, about 0.5mg/kg, about 1 mg/kg, about 2 mg/kg, about 3 mg/kg, about 4 mg/kg, about 5 mg/kg, about 6mg/kg, about 7 mg/kg, about 8 mg/kg, about 9 mg/kg, or about 10 mg/kg. The pharmaceutical composition comprising an effective amount of RNAi construct can be administered weekly, biweekly, monthly, quarterly, or biannually. The precise determination of what would be considered an effective amount and frequency of administration may be based on several factors, including a patient's size, age, and general condition, type of disorder to be treated (e.g. myocardial infarction, heart failure, coronary artery disease, hypercholesterolemia), particular RNAi construct employed, and route of administration. Estimates of effective dosages and in vivo half-lives for any particular RNAi construct of the invention can be ascertained using conventional methods and/or testing in appropriate animal models.

[0103] Administration of the pharmaceutical compositions of the present invention may be via any common route so long as the target tissue is available via that route. Such routes include, but are not limited to, parenteral (e.g., subcutaneous, intramuscular, intraperitoneal or intravenous), oral, nasal, buccal, intradermal, transdermal, and sublingual routes, or by direct injection into liver tissue or delivery through the hepatic portal vein. In some embodiments, the pharmaceutical composition is administered parenterally. For instance, in certain embodiments, the pharmaceutical composition is administered intravenously. In other embodiments, the pharmaceutical composition is administered subcutaneously.

[0104] Colloidal dispersion systems, such as macromolecule complexes, nanocapsules, microspheres, beads, and lipid-based systems, including oil-in-water emulsions, micelles, mixed micelles, and liposomes, may be used as delivery vehicles for the RNAi constructs of the invention or vectors encoding such constructs. Commercially available fat emulsions that are suitable for delivering the nucleic acids of the invention include Intralipid®, Liposyn®, Liposyn®II, Liposyn®III, Nutrilipid, and other similar lipid emulsions. A preferred colloidal system for use as a delivery vehicle in vivo is a liposome (i.e., an artificial membrane vesicle). The RNAi constructs of the invention may be encapsulated within liposomes or may form complexes thereto, in particular to cationic liposomes. Alternatively, RNAi constructs of the invention may be complexed to lipids, in particular to cationic lipids. Suitable lipids and liposomes include neutral (e.g., dioleoylphosphatidyl ethanolamine (DOPE), dimyristoylphosphatidyl choline (DMPC), and dipalmitoyl phosphatidylcholine (DPPC)), distearoylphosphatidyl choline), negative (e.g., dimyristoylphosphatidyl glycerol (DMPG)), and cationic (e.g., dioleoyltetramethylaminopropyl (DOTAP) and dioleoylphosphatidyl ethanolamine (DOTMA)). The preparation and use of such colloidal dispersion systems is well known in the art. Exemplary formulations are also disclosed in U.S. Pat. No. 5,981,505; U.S. Pat. No. 6,217,900; U.S. Pat. No. 6,383,512; U.S. Pat. No. 5,783,565; U.S. Pat. No. 7,202,227; U.S. Pat. No. 6,379,965; U.S. Pat. No. 6,127,170; U.S. Pat. No. 5,837,533; U.S. Pat. No. 6,747,014; and W003/093449.

[0105] In some embodiments, the RNAi constructs of the invention are fully encapsulated in a lipid formulation, e.g., to form a SPLP, pSPLP, SNALP, or other nucleic acid-lipid particle. As used herein, the term "SNALP" refers to a stable nucleic acid-lipid particle, including SPLP. As

used herein, the term "SPLP" refers to a nucleic acid-lipid particle comprising plasmid DNA encapsulated within a lipid vesicle. SNALPs and SPLPs typically contain a cationic lipid, a noncationic lipid, and a lipid that prevents aggregation of the particle (e.g., a PEG-lipid conjugate). SNALPs and SPLPs are exceptionally useful for systemic applications, as they exhibit extended circulation lifetimes following intravenous injection and accumulate at distal sites (e.g., sites physically separated from the administration site). SPLPs include "pSPLP," which include an encapsulated condensing agent-nucleic acid complex as set forth in PCT Publication No. WO00/03683. The nucleic acid-lipid particles typically have a mean diameter of about 50 nm to about 150 nm, about 60 nm to about 130 nm, about 70 nm to about 110 nm, or about 70 nm to about 90 nm, and are substantially nontoxic. In addition, the nucleic acids when present in the nucleic acid-lipid particles are resistant in aqueous solution to degradation with a nuclease. Nucleic acid-lipid particles and their method of preparation are disclosed in, e.g., U.S. Patent Nos. 5,976,567; 5,981,501; 6,534,484; 6,586,410; 6,815,432; and PCT Publication No. WO96/40964.

[0106] The pharmaceutical compositions suitable for injectable use include, for example, sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. Generally, these preparations are sterile and fluid to the extent that easy injectability exists. Preparations should be stable under the conditions of manufacture and storage and should be preserved against the contaminating action of microorganisms, such as bacteria and fungi. Appropriate solvents or dispersion media may contain, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and vegetable oils. The proper fluidity can be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. The prevention of the action of microorganisms can be brought about by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

[0107] Sterile injectable solutions may be prepared by incorporating the active compounds in an appropriate amount into a solvent along with any other ingredients (for example as enumerated above) as desired, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the various sterilized active ingredients into a sterile vehicle which contains the basic dispersion medium and the desired other ingredients, e.g., as enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation include vacuum-drying and freeze-drying techniques which yield a powder of the active ingredient(s) plus any additional desired ingredient from a previously sterile-filtered solution thereof

[0108] The compositions of the present invention generally may be formulated in a neutral or salt form. Pharmaceutically-acceptable salts include, for example, acid addition salts (formed with free amino groups) derived from inorganic acids (e.g., hydrochloric or phosphoric acids), or from organic acids (e.g., acetic, oxalic, tartaric, mandelic, and the like). Salts formed with the free carboxyl groups can also be derived from inorganic bases (e.g., sodium, potassium, ammonium, calcium, or ferric hydroxides) or from organic bases (e.g., isopropylamine, trimethylamine, histidine, procaine and the like).

[0109] For parenteral administration in an aqueous solution, for example, the solution generally is suitably buffered and the liquid diluent first rendered isotonic for example with sufficient saline or glucose. Such aqueous solutions may be used, for example, for intravenous, intramuscular, subcutaneous and intraperitoneal administration. Preferably, sterile aqueous media are employed as is known to those of skill in the art, particularly in light of the present disclosure. By way of illustration, a single dose may be dissolved in 1 ml of isotonic NaCl solution and either added to 1000 ml of hypodermoclysis fluid or injected at the proposed site of infusion, (see for example, "Remington's Pharmaceutical Sciences" 15th Edition, pages 1035-1038 and 1570-1580). For human administration, preparations should meet sterility, pyrogenicity, general safety and purity standards as required by FDA standards. In certain embodiments, a pharmaceutical composition of the invention comprises or consists of a sterile saline solution and an RNAi construct described herein. In other embodiments, a pharmaceutical composition of the invention comprises or consists of an RNAi construct described herein and sterile water (e.g. water for

injection, WFI). In still other embodiments, a pharmaceutical composition of the invention comprises or consists of an RNAi construct described herein and phosphate-buffered saline (PBS).

[0110] In some embodiments, the pharmaceutical compositions of the invention are packaged with or stored within a device for administration. Devices for injectable formulations include, but are not limited to, injection ports, pre-filled syringes, auto injectors, injection pumps, on-body injectors, and injection pens. Devices for aerosolized or powder formulations include, but are not limited to, inhalers, insufflators, aspirators, and the like. Thus, the present invention includes administration devices comprising a pharmaceutical composition of the invention for treating or preventing one or more of the disorders described herein.

Methods for inhibiting PNPLA3 expression

[0111] The present invention also provides methods of inhibiting expression of a PNPLA3 gene in a cell. The methods include contacting a cell with an RNAi agent, e.g., double stranded RNAi agent, in an amount effective to inhibit expression of PNPLA3 in the cell, thereby inhibiting expression of PNPLA3 in the cell. Contacting of a cell with an RNAi agent, e.g., a double stranded RNAi agent, may be done in vitro or in vivo. Contacting a cell in vivo with the RNAi agent includes contacting a cell or group of cells within a subject, e.g., a human subject, with the RNAi agent. Combinations of in vitro and in vivo methods of contacting a cell are also possible.

[0112] The present invention provides methods for reducing or inhibiting expression of PNPLA3 in a subject in need thereof as well as methods of treating or preventing conditions, diseases, or disorders associated with PNPLA3 expression or activity. A "condition, disease, or disorder associated with PNPLA3 expression" refers to conditions, diseases, or disorders in which PNPLA3 expression levels are altered or where elevated expression levels of PNPLA3 are associated with an increased risk of developing the condition, disease or disorder.

[0113] Contacting a cell may be direct or indirect, as discussed above. Furthermore, contacting a cell may be accomplished via a targeting ligand, including any ligand described herein or known in the art. In preferred embodiments, the targeting ligand is a carbohydrate moiety, e.g., a GalNAc3 ligand, or any other ligand that directs the RNAi agent to a site of interest.

[0114] In one embodiment, contacting a cell with an RNAi includes "introducing" or "delivering the RNAi into the cell" by facilitating or effecting uptake or absorption into the cell. Absorption or uptake of an RNAi can occur through unaided diffusive or active cellular processes, or by auxiliary agents or devices. Introducing an RNAi into a cell may be in vitro and/or in vivo. For example, for in vivo introduction, RNAi can be injected into a tissue site or administered systemically. In vitro introduction into a cell includes methods known in the art such as electroporation and lipofection. Further approaches are described herein below and/or are known in the art.

[0115] The term "inhibiting," as used herein, is used interchangeably with "reducing," "silencing," "downregulating", "suppressing", and other similar terms, and includes any level of inhibition.

[0116] The phrase "inhibiting expression of a PNPLA3" is intended to refer to inhibition of expression of any PNPLA3 gene (such as, e.g., a mouse PNPLA3 gene, a rat PNPLA3 gene, a monkey PNPLA3 gene, or a human PNPLA3 gene) as well as variants or mutants of a PNPLA3 gene. Thus, the PNPLA3 gene may be a wild-type PNPLA3 gene, a mutant PNPLA3 gene (such as a mutant PNPLA3 gene giving rise to amyloid deposition), or a transgenic PNPLA3 gene in the context of a genetically manipulated cell, group of cells, or organism.

[0117] "Inhibiting expression of a PNPLA3 gene" includes any level of inhibition of a PNPLA3 gene, e.g., at least partial suppression of the expression of a PNPLA3 gene. The expression of the PNPLA3 gene may be assessed based on the level, or the change in the level, of any variable associated with PNPLA3 gene expression, e.g., PNPLA3 mRNA level, PNPLA3 protein level, or the number or extent of amyloid deposits. This level may be assessed in an individual cell or in a group of cells, including, for example, a sample derived from a subject.

[0118] Inhibition may be assessed by a decrease in an absolute or relative level of one or more variables that are associated with PNPLA3 expression compared with a control level. The control level may be any type of control level that is utilized in the art, e.g., a pre-dose baseline level, or a level determined from a similar subject, cell, or sample that is untreated or treated with a control (such as, e.g., buffer only control or inactive agent control). In some embodiments of the methods of the invention, expression of a PNPLA3 gene is inhibited by at least about 5%, at least about

10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 55%, at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, or at least about 99%.

[0119] Inhibition of the expression of a PNPLA3 gene may be manifested by a reduction of the amount of mRNA expressed by a first cell or group of cells (such cells may be present, for example, in a sample derived from a subject) in which a PNPLA3 gene is transcribed and which has or have been treated (e.g., by contacting the cell or cells with an RNAi agent of the invention, or by administering an RNAi agent of the invention to a subject in which the cells are or were present) such that the expression of a PNPLA3 gene is inhibited, as compared to a second cell or group of cells substantially identical to the first cell or group of cells but which has not or have not been so treated (control cell(s)). In preferred embodiments, the inhibition is assessed by expressing the level of mRNA in treated cells as a percentage of the level of mRNA in control cells, using the following formula:

$$\frac{(\text{mRNA in control cells}) - (\text{mRNA in treated cells})}{(\text{mRNA in control cells})} \cdot 100\%$$

[0120] Alternatively, inhibition of the expression of a PNPLA3 gene may be assessed in terms of a reduction of a parameter that is functionally linked to PNPLA3 gene expression, e.g., PNPLA3 protein expression or Hedgehog pathway protein activities. PNPLA3 gene silencing may be determined in any cell expressing PNPLA3, either constitutively or by genomic engineering, and by any assay known in the art.

[0121] Inhibition of the expression of a PNPLA3 protein may be manifested by a reduction in the level of the PNPLA3 protein that is expressed by a cell or group of cells (e.g., the level of protein expressed in a sample derived from a subject). As explained above, for the assessment of

mRNA suppression, the inhibition of protein expression levels in a treated cell or group of cells may similarly be expressed as a percentage of the level of protein in a control cell or group of cells.

[0122] A control cell or group of cells that may be used to assess the inhibition of the expression of a PNPLA3 gene includes a cell or group of cells that has not yet been contacted with an RNAi agent of the invention. For example, the control cell or group of cells may be derived from an individual subject (e.g., a human or animal subject) prior to treatment of the subject with an RNAi agent.

[0123] The level of PNPLA3 mRNA that is expressed by a cell or group of cells, or the level of circulating PNPLA3 mRNA, may be determined using any method known in the art for assessing mRNA expression. In one embodiment, the level of expression of PNPLA3 in a sample is determined by detecting a transcribed polynucleotide, or portion thereof, e.g., mRNA of the PNPLA3 gene. RNA may be extracted from cells using RNA extraction techniques including, for example, using acid phenol/guanidine isothiocyanate extraction (RNAzol B; Biogenesis), RNeasy RNA preparation kits (Qiagen) or PAXgene (PreAnalytix, Switzerland). Typical assay formats utilizing ribonucleic acid hybridization include nuclear run-on assays, RT-PCR, RNase protection assays (Melton et al., Nuc. Acids Res. 12:7035), Northern blotting, in situ hybridization, and microarray analysis. Circulating PNPLA3 mRNA may be detected using methods the described in PCT/US2012/043584, the entire contents of which are hereby incorporated herein by reference.

[0124] In one embodiment, the level of expression of PNPLA3 is determined using a nucleic acid probe. The term "probe", as used herein, refers to any molecule that is capable of selectively binding to a specific PNPLA3. Probes can be synthesized by one of skill in the art, or derived from appropriate biological preparations. Probes may be specifically designed to be labeled. Examples of molecules that can be utilized as probes include, but are not limited to, RNA, DNA, proteins, antibodies, and organic molecules.

[0125] Isolated mRNA can be used in hybridization or amplification assays that include, but are not limited to, Southern or Northern analyses, polymerase chain reaction (PCR) analyses and probe arrays. One method for the determination of mRNA levels involves contacting the isolated mRNA with a nucleic acid molecule (probe) that can hybridize to PNPLA3 mRNA. In one embodiment, the mRNA is immobilized on a solid surface and contacted with a probe, for example

by running the isolated mRNA on an agarose gel and transferring the mRNA from the gel to a membrane, such as nitrocellulose. In an alternative embodiment, the probe(s) are immobilized on a solid surface and the mRNA is contacted with the probe(s), for example, in an Affymetrix gene chip array. A skilled artisan can readily adapt known mRNA detection methods for use in determining the level of PNPLA3 mRNA.

[0126] An alternative method for determining the level of expression of PNPLA3 in a sample involves the process of nucleic acid amplification and/or reverse transcriptase (to prepare cDNA) of for example mRNA in the sample, e.g., by RT-PCR (the experimental embodiment set forth in Mullis, 1987, U.S. Pat. No. 4,683,202), ligase chain reaction (Barany (1991) Proc. Natl. Acad. Sci. USA 88: 189-193), self sustained sequence replication (Guatelli et al. (1990) Proc. Natl. Acad. Sci. USA 87: 1874-1878), transcriptional amplification system (Kwoh et al. (1989) Proc. Natl. Acad. Sci. USA 86: 1173-1177), Q-Beta Replicase (Lizardi et al. (1988) Bio/Technology 6: 1197), rolling circle replication (Lizardi et al., U.S. Pat. No. 5,854,033) or any other nucleic acid amplification method, followed by the detection of the amplified molecules using techniques well known to those of skill in the art. These detection schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers. In particular aspects of the invention, the level of expression of PNPLA3 is determined by quantitative fluorogenic RT-PCR (i.e., the TaqMan™ System). The expression levels of PNPLA3 mRNA may be monitored using a membrane blot (such as used in hybridization analysis such as Northern, Southern, dot, and the like), or microwells, sample tubes, gels, beads or fibers (or any solid support comprising bound nucleic acids). See U.S. Pat. Nos. 5,770,722, 5,874,219, 5,744,305, 5,677,195 and 5,445,934, which are incorporated herein by reference. The determination of PNPLA3 expression level may also comprise using nucleic acid probes in solution.

[0127] In preferred embodiments, the level of mRNA expression is assessed using branched DNA (bDNA) assays or real time PCR (qPCR). The use of these methods is described and exemplified in the Examples presented herein.

[0128] The level of PNPLA3 protein expression may be determined using any method known in the art for the measurement of protein levels. Such methods include, for example, electrophoresis, capillary electrophoresis, high performance liquid chromatography (HPLC), thin

layer chromatography (TLC), hyperdiffusion chromatography, fluid or gel precipitin reactions, absorption spectroscopy, a colorimetric assays, spectrophotometric assays, flow cytometry, immunodiffusion (single or double), Immunoelectrophoresis, Western blotting, radioimmunoassay (RIA), enzyme-linked immunosorbent assays (ELISAs), immunofluorescent assays, electrochemiluminescence assays, and the like.

[0129] In some embodiments, the efficacy of the methods of the invention can be monitored by detecting or monitoring a reduction in a symptom of a PNPLA3 disease, such as reduction in edema swelling of the extremities, face, larynx, upper respiratory tract, abdomen, trunk, and genitals, prodrome; laryngeal swelling; nonpruritic rash; nausea; vomiting; or abdominal pain. These symptoms may be assessed in vitro or in vivo using any method known in the art.

[0130] In some embodiments of the methods of the invention, the RNAi agent is administered to a subject such that the RNAi agent is delivered to a specific site within the subject. The inhibition of expression of PNPLA3 may be assessed using measurements of the level or change in the level of PNPLA3 mRNA or PNPLA3 protein in a sample derived from fluid or tissue from the specific site within the subject. In preferred embodiments, the site is selected from the group consisting of liver, choroid plexus, retina, and pancreas. The site may also be a subsection or subgroup of cells from any one of the aforementioned sites. The site may also include cells that express a particular type of receptor.

Methods of treating or preventing PNPLA3-Associated Diseases

[0131] The present invention provides therapeutic and prophylactic methods which include administering to a subject with a PNPLA3 -associated disease, disorder, and/or condition, or prone to developing, a PNPLA3- associated disease, disorder, and/or condition, compositions comprising an RNAi agent, or pharmaceutical compositions comprising an RNAi agent, or vectors comprising an RNAi of the invention. Non-limiting examples of PNPLA3- associated diseases include, for example, fatty liver (steatosis), nonalcoholic steatohepatitis (NASH), cirrhosis of the liver, accumulation of fat in the liver, inflammation of the liver, hepatocellular necrosis, liver fibrosis, obesity, or nonalcoholic fatty liver disease (NAFLD). In one embodiment, the PNPLA3-associated disease is NAFLD. In another embodiment, the PNPLA3 -associated disease is NASH. In another embodiment, the PNPLA3 -associated disease is fatty liver (steatosis). In another embodiment, the

PNPLA3-associated disease is insulin resistance. In another embodiment, the PNPLA3-associated disease is not insulin resistance.

[0132] In certain embodiments, the present invention provides a method for reducing the expression of PNPLA3 in a patient in need thereof comprising administering to the patient any of the RNAi constructs described herein. The term "patient," as used herein, refers to a mammal, including humans, and can be used interchangeably with the term "subject." Preferably, the expression level of PNPLA3 in hepatocytes in the patient is reduced following administration of the RNAi construct as compared to the PNPLA3 expression level in a patient not receiving the RNAi construct.

[0133] The methods of the invention are useful for treating a subject having a PNPLA3-associated disease, e.g., a subject that would benefit from reduction in PNPLA3 gene expression and/or PNPLA3 protein production. In one aspect, the present invention provides methods of reducing the level of Patatin-Like Phospholipase Domain Containing 3 (PNPLA3) gene expression in a subject having nonalcoholic fatty liver disease (NAFLD). In another aspect, the present invention provides methods of reducing the level of PNPLA3 protein in a subject with NAFLD. The present invention also provides methods of reducing the level of activity of the hedgehog pathway in a subject with NAFLD.

[0134] In another aspect, the present invention provides methods of treating a subject having an NAFLD. In one aspect, the present invention provides methods of treating a subject having a PNPLA3-associated disease, e.g., fatty liver (steatosis), nonalcoholic steatohepatitis (NASH), cirrhosis of the liver, accumulation of fat in the liver, inflammation of the liver, hepatocellular necrosis, liver fibrosis, obesity, or nonalcoholic fatty liver disease (NAFLD). The treatment methods (and uses) of the invention include administering to the subject, e.g., a human, a therapeutically effective amount of an RNAi agent of the invention targeting a PNPLA3 gene or a pharmaceutical composition comprising an RNAi agent of the invention targeting a PNPLA3 gene or a vector of the invention comprising an RNAi agent targeting a PNPLA3 gene.

[0135] In one aspect, the invention provides methods of preventing at least one symptom in a subject having NAFLD, e.g., the presence of elevated hedgehog signaling pathways, fatigue, weakness, weight loss, loss of appetite, nausea, abdominal pain, spider-like blood vessels, yellowing

of the skin and eyes (jaundice), itching, fluid build up and swelling of the legs (edema), abdomen swelling (ascites), and mental confusion. The methods include administering to the subject a therapeutically effective amount of the RNAi agent, e.g. dsRNA, pharmaceutical compositions, or vectors of the invention, thereby preventing at least one symptom in the subject having a disorder that would benefit from reduction in PNPLA3 gene expression.

[0136] In another aspect, the present invention provides uses of a therapeutically effective amount of an RNAi agent of the invention for treating a subject, e.g., a subject that would benefit from a reduction and/or inhibition of PNPLA3 gene expression. In a further aspect, the present invention provides uses of an RNAi agent, e.g., a dsRNA, of the invention targeting an PNPLA3 gene or pharmaceutical composition comprising an RNAi agent targeting an PNPLA3 gene in the manufacture of a medicament for treating a subject, e.g., a subject that would benefit from a reduction and/or inhibition of PNPLA3 gene expression and/or PNPLA3 protein production, such as a subject having a disorder that would benefit from reduction in PNPLA3 gene expression, e.g., a PNPLA3- associated disease.

[0137] In another aspect, the invention provides uses of an RNAi, e.g., a dsRNA, of the invention for preventing at least one symptom in a subject suffering from a disorder that would benefit from a reduction and/or inhibition of PNPLA3 gene expression and/or PNPLA3 protein production.

[0138] In a further aspect, the present invention provides uses of an RNAi agent of the invention in the manufacture of a medicament for preventing at least one symptom in a subject suffering from a disorder that would benefit from a reduction and/or inhibition of PNPLA3 gene expression and/or PNPLA3 protein production, such as a PNPLA3-associated disease.

[0139] In one embodiment, an RNAi agent targeting PNPLA3 is administered to a subject having a PNPLA3-associated disease, e.g., nonalcoholic fatty liver disease (NAFLD), such that the expression of a PNPLA3 gene, e.g., in a cell, tissue, blood or other tissue or fluid of the subject are reduced by at least about 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 62%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%,

73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91 %, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or at least about 99% or more when the dsRNA agent is administered to the subject.

[0140] The methods and uses of the invention include administering a composition described herein such that expression of the target PNPLA3 gene is decreased, such as for about 1, 2, 3, 4 5, 6, 7, 8, 12, 16, 18, 24, 28, 32, 36, 40, 44, 48, 52, 56, 60, 64, 68, 72, 76, or about 80 hours. In one embodiment, expression of the target PNPLA3 gene is decreased for an extended duration, e.g., at least about two, three, four, five, six, seven days or more, e.g., about one week, two weeks, three weeks, or about four weeks or longer.

[0141] Administration of the dsRNA according to the methods and uses of the invention may result in a reduction of the severity, signs, symptoms, and/or markers of such diseases or disorders in a patient with a PNPLA3- associated disease, e.g., nonalcoholic fatty liver disease (NAFLD). By "reduction" in this context is meant a statistically significant decrease in such level. The reduction can be, for example, at least about 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, or about 100%. Efficacy of treatment or prevention of disease can be assessed, for example by measuring disease progression, disease remission, symptom severity, reduction in pain, quality of life, dose of a medication required to sustain a treatment effect, level of a disease marker or any other measurable parameter appropriate for a given disease being treated or targeted for prevention. It is well within the ability of one skilled in the art to monitor efficacy of treatment or prevention by measuring any one of such parameters, or any combination of parameters. For example, efficacy of treatment of NAFLD may be assessed, for example, by periodic monitoring of NAFLD symptoms, liver fat levels, or expression of downstream genes. Comparison of the later readings with the initial readings provide a physician an indication of whether the treatment is effective. It is well within the ability of one skilled in the art to monitor efficacy of treatment or prevention by measuring any one of such parameters, or any combination of parameters. In connection with the administration of an RNAi targeting PNPLA3 or pharmaceutical composition thereof, "effective against" an PNPLA3 - associated disease indicates that administration in a clinically appropriate manner results in a beneficial effect for at least a statistically significant fraction of patients, such as improvement of

symptoms, a cure, a reduction in disease, extension of life, improvement in quality of life, or other effect generally recognized as positive by medical doctors familiar with treating NAFLD and/or an PNPLA3 -associated disease and the related causes.

[0142] A treatment or preventive effect is evident when there is a statistically significant improvement in one or more parameters of disease status, or by a failure to worsen or to develop symptoms where they would otherwise be anticipated. As an example, a favorable change of at least 10% in a measurable parameter of disease, and preferably at least 20%, 30%, 40%, 50% or more can be indicative of effective treatment. Efficacy for a given RNAi drug or formulation of that drug can also be judged using an experimental animal model for the given disease as known in the art. When using an experimental animal model, efficacy of treatment is evidenced when a statistically significant reduction in a marker or symptom is observed.

[0143] Subjects can be administered a therapeutic amount of RNAi, such as about 0.01 mg/kg, 0.02 mg/kg, 0.03 mg/kg, 0.04 mg/kg, 0.05 mg/kg, 0.1 mg/kg, 0.15 mg/kg, 0.2 mg/kg, 0.25 mg/kg, 0.3 mg/kg, 0.35 mg/kg, 0.4 mg/kg, 0.45 mg/kg, 0.5 mg/kg, 0.55 mg/kg, 0.6 mg/kg, 0.65 mg/kg, 0.7 mg/kg, 0.75 mg/kg, 0.8 mg/kg, 0.85 mg/kg, 0.9 mg/kg, 0.95 mg/kg, 1.0 mg/kg, 1.1 mg/kg, 1.2 mg/kg, 1.3 mg/kg, 1.4 mg/kg, 1.5 mg/kg, 1.6 mg/kg, 1.7 mg/kg, 1.8 mg/kg, 1.9 mg/kg, 2.0 mg/kg, 2.1 mg/kg, 2.2 mg/kg, 2.3 mg/kg, 2.4 mg/kg, 2.5 mg/kg dsRNA, 2.6 mg/kg dsRNA, 2.7 mg/kg dsRNA, 2.8 mg/kg dsRNA, 2.9 mg/kg dsRNA, 3.0 mg/kg dsRNA, 3.1 mg/kg dsRNA, 3.2 mg/kg dsRNA, 3.3 mg/kg dsRNA, 3.4 mg/kg dsRNA, 3.5 mg/kg dsRNA, 3.6 mg/kg dsRNA, 3.7 mg/kg dsRNA, 3.8 mg/kg dsRNA, 3.9 mg/kg dsRNA, 4.0 mg/kg dsRNA, 4.1 mg/kg dsRNA, 4.2 mg/kg dsRNA, 4.3 mg/kg dsRNA, 4.4 mg/kg dsRNA, 4.5 mg/kg dsRNA, 4.6 mg/kg dsRNA, 4.7 mg/kg dsRNA, 4.8 mg/kg dsRNA, 4.9 mg/kg dsRNA, 5.0 mg/kg dsRNA, 5.1 mg/kg dsRNA, 5.2 mg/kg dsRNA, 5.3 mg/kg dsRNA, 5.4 mg/kg dsRNA, 5.5 mg/kg dsRNA, 5.6 mg/kg dsRNA, 5.7 mg/kg dsRNA, 5.8 mg/kg dsRNA, 5.9 mg/kg dsRNA, 6.0 mg/kg dsRNA, 6.1 mg/kg dsRNA, 6.2 mg/kg dsRNA, 6.3 mg/kg dsRNA, 6.4 mg/kg dsRNA, 6.5 mg/kg dsRNA, 6.6 mg/kg dsRNA, 6.7 mg/kg dsRNA, 6.8 mg/kg dsRNA, 6.9 mg/kg dsRNA, 7.0 mg/kg dsRNA, 7.1 mg/kg dsRNA, 7.2 mg/kg dsRNA, 7.3 mg/kg dsRNA, 7.4 mg/kg dsRNA, 7.5 mg/kg dsRNA, 7.6 mg/kg dsRNA, 7.7 mg/kg dsRNA, 7.8 mg/kg dsRNA, 7.9 mg/kg dsRNA, 8.0 mg/kg dsRNA, 8.1 mg/kg dsRNA, 8.2 mg/kg dsRNA, 8.3 mg/kg dsRNA, 8.4 mg/kg dsRNA, 8.5 mg/kg dsRNA, 8.6 mg/kg dsRNA, 8.7 mg/kg

dsRNA, 8.8 mg/kg dsRNA, 8.9 mg/kg dsRNA, 9.0 mg/kg dsRNA, 9.1 mg/kg dsRNA, 9.2 mg/kg dsRNA, 9.3 mg/kg dsRNA, 9.4 mg/kg dsRNA, 9.5 mg/kg dsRNA, 9.6 mg/kg dsRNA, 9.7 mg/kg dsRNA, 9.8 mg/kg dsRNA, 9.9 mg/kg dsRNA, 9.0 mg/kg dsRNA, 10 mg/kg dsRNA, 15 mg/kg dsRNA, 20 mg/kg dsRNA, 25 mg/kg dsRNA, 30 mg/kg dsRNA, 35 mg/kg dsRNA, 40 mg/kg dsRNA, 45 mg/kg dsRNA, or about 50 mg/kg dsRNA. In one embodiment, subjects can be administered 0.5 mg/kg of the dsRNA. Values and ranges intermediate to the recited values are also intended to be part of this invention.

[0144] Administration of the RNAi can reduce the presence of PNPLA3 protein levels, e.g. , in a cell, tissue, blood, urine or other compartment of the patient by at least about 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31 %, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or at least about 99% or more.

[0145] Before administration of a full dose of the RNAi, patients can be administered a smaller dose, such as a 5% infusion, and monitored for adverse effects, such as an allergic reaction. In another example, the patient can be monitored for unwanted immunostimulatory effects, such as increased cytokine (e.g. , TNF-alpha or INF-alpha) levels.

[0146] Owing to the inhibitory effects on PNPLA3 expression, a composition according to the invention or a pharmaceutical composition prepared therefrom can enhance the quality of life.

[0147] An RNAi of the invention may be administered in "naked" form, where the modified or unmodified RNAi agent is directly suspended in aqueous or suitable buffer solvent, as a "free RNAi." A free RNAi is administered in the absence of a pharmaceutical composition. The free RNAi may be in a suitable buffer solution. The buffer solution may comprise acetate, citrate, prolamine, carbonate, or phosphate, or any combination thereof. In one embodiment, the buffer solution is phosphate buffered saline (PBS). The pH and osmolality of the buffer solution containing the RNAi can be adjusted such that it is suitable for administering to a subject.

[0148] Alternatively, an RNAi of the invention may be administered as a pharmaceutical composition, such as a dsRNA liposomal formulation.

[0149] Subjects that would benefit from a reduction and/or inhibition of PNPLA3 gene expression are those having nonalcoholic fatty liver disease (NAFLD) and/or an PNPLA3-associated disease or disorder as described herein.

[0150] Treatment of a subject that would benefit from a reduction and/or inhibition of PNPLA3 gene expression includes therapeutic and prophylactic treatment.

[0151] The invention further provides methods and uses of an RNAi agent or a pharmaceutical composition thereof for treating a subject that would benefit from reduction and/or inhibition of PNPLA3 gene expression, e.g., a subject having a PNPLA3-associated disease, in combination with other pharmaceuticals and/or other therapeutic methods, e.g., with known pharmaceuticals and/or known therapeutic methods, such as, for example, those which are currently employed for treating these disorders.

[0152] For example, in certain embodiments, an RNAi targeting a PNPLA3 gene is administered in combination with, e.g., an agent useful in treating an PNPLA3-associated disease as described elsewhere herein. For example, additional therapeutics and therapeutic methods suitable for treating a subject that would benefit from reduction in PNPLA3 expression, e.g., a subject having a PNPLA3-associated disease, include an RNAi agent targeting a different portion of the PNPLA3 gene, a therapeutic agent, and/or procedures for treating a PNPLA3-associated disease or a combination of any of the foregoing.

[0153] In certain embodiments, a first RNAi agent targeting a PNPLA3 gene is administered in combination with a second RNAi agent targeting a different portion of the PNPLA3 gene. For example, the first RNAi agent comprises a first sense strand and a first antisense strand forming a double stranded region, wherein substantially all of the nucleotides of said first sense strand and substantially all of the nucleotides of the first antisense strand are modified nucleotides, wherein said first sense strand is conjugated to a ligand attached at the 3'-terminus, and wherein the ligand is one or more GalNAc derivatives attached through a bivalent or trivalent branched linker; and the second RNAi agent comprises a second sense strand and a second antisense strand forming a

double stranded region, wherein substantially all of the nucleotides of the second sense strand and substantially all of the nucleotides of the second antisense strand are modified nucleotides, wherein the second sense strand is conjugated to a ligand attached at the 3'-terminus, and wherein the ligand is one or more GalNAc derivatives attached through a bivalent or trivalent branched linker.

[0154] In one embodiment, all of the nucleotides of the first and second sense strand and/or all of the nucleotides of the first and second antisense strand comprise a modification.

[0155] In one embodiment, the at least one of the modified nucleotides is selected from the group consisting of a 3'-terminal deoxy-thymine (dT) nucleotide, a 2'-O-methyl modified nucleotide, a 2'-fluoro modified nucleotide, a 2'-deoxy-modified nucleotide, a locked nucleotide, an unlocked nucleotide, a conformationally restricted nucleotide, a constrained ethyl nucleotide, an abasic nucleotide, a 2'-amino-modified nucleotide, a 2'-O-allyl-modified nucleotide, 2'-C-alkyl-modified nucleotide, 2'-hydroxyl-modified nucleotide, a 2'-methoxyethyl modified nucleotide, a 2'-O-alkyl-modified nucleotide, a morpholino nucleotide, a phosphoramidate, a non-natural base comprising nucleotide, a tetrahydropyran modified nucleotide, a 1,5-anhydrohexitol modified nucleotide, a cyclohexenyl modified nucleotide, a nucleotide comprising a phosphorothioate group, a nucleotide comprising a methylphosphonate group, a nucleotide comprising a 5'-phosphate, and a nucleotide comprising a 5'-phosphate mimic.

[0156] In certain embodiments, a first RNAi agent targeting a PNPLA3 gene is administered in combination with a second RNAi agent targeting a gene that is different from the PNPLA3 gene. For example, the RNAi agent targeting the PNPLA3 gene may be administered in combination with an RNAi agent targeting the SCAP gene. The first RNAi agent targeting a PNPLA3 gene and the second RNAi agent targeting a gene different from the PNPLA3 gene, e.g., the SCAP gene, may be administered as parts of the same pharmaceutical composition. Alternatively, the first RNAi agent targeting a PNPLA3 gene and the second RNAi agent targeting a gene different from the PNPLA3 gene, e.g., the SCAP gene, may be administered as parts of different pharmaceutical compositions.

[0157] The RNAi agent and an additional therapeutic agent and/or treatment may be administered at the same time and/or in the same combination, e.g., parenterally, or the additional

therapeutic agent can be administered as part of a separate composition or at separate times and/or by another method known in the art or described herein.

[0158] The present invention also provides methods of using an RNAi agent of the invention and/or a composition containing an RNAi agent of the invention to reduce and/or inhibit PNPLA3 expression in a cell. In other aspects, the present invention provides an RNAi of the invention and/or a composition comprising an RNAi of the invention for use in reducing and/or inhibiting PNPLA3 gene expression in a cell. In yet other aspects, use of an RNAi of the invention and/or a composition comprising an RNAi of the invention for the manufacture of a medicament for reducing and/or inhibiting PNPLA3 gene expression in a cell are provided. In still other aspects, the the present invention provides an RNAi of the invention and/or a composition comprising an RNAi of the invention for use in reducing and/or inhibiting PNPLA3 protein production in a cell. In yet other aspects, use of an RNAi of the invention and/or a composition comprising an RNAi of the invention for the manufacture of a medicament for reducing and/or inhibiting PNPLA3 protein production in a cell are provided. The methods and uses include contacting the cell with an RNAi, e.g., a dsRNA, of the invention and maintaining the cell for a time sufficient to obtain degradation of the mRNA transcript of an PNPLA3 gene, thereby inhibiting expression of the PNPLA3 gene or inhibiting PNPLA3 protein production in the cell.

[0159] Reduction in gene expression can be assessed by any methods known in the art. For example, a reduction in the expression of PNPLA3 may be determined by determining the mRNA expression level of PNPLA3 using methods routine to one of ordinary skill in the art, e.g., Northern blotting, qRT-PCR, by determining the protein level of PNPLA3 using methods routine to one of ordinary skill in the art, such as Western blotting, immunological techniques, flow cytometry methods, ELISA, and/or by determining a biological activity of PNPLA3.

[0160] In the methods and uses of the invention the cell may be contacted in vitro or in vivo, i.e., the cell may be within a subject.

[0161] A cell suitable for treatment using the methods of the invention may be any cell that expresses an PNPLA3 gene, e.g., a cell from a subject having NAFLD or a cell comprising an expression vector comprising a PNPLA3 gene or portion of a PNPLA3 gene. A cell suitable for use in the methods and uses of the invention may be a mammalian cell, e.g., a primate cell (such

as a human cell or a non-human primate cell, e.g., a monkey cell or a chimpanzee cell), a non-primate cell (such as a cow cell, a pig cell, a camel cell, a llama cell, a horse cell, a goat cell, a rabbit cell, a sheep cell, a hamster, a guinea pig cell, a cat cell, a dog cell, a rat cell, a mouse cell, a lion cell, a tiger cell, a bear cell, or a buffalo cell), a bird cell (e.g., a duck cell or a goose cell), or a whale cell. In one embodiment, the cell is a human cell.

[0162] PNPLA3 gene expression may be inhibited in the cell by at least about 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or about 100%.

[0163] PNPLA3 protein production may be inhibited in the cell by at least about 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or about 100%.

[0164] The in vivo methods and uses of the invention may include administering to a subject a composition containing an RNAi, where the RNAi includes a nucleotide sequence that is complementary to at least a part of an RNA transcript of the PNPLA3 gene of the mammal to be treated. When the organism to be treated is a human, the composition can be administered by any means known in the art including, but not limited to subcutaneous, intravenous, oral, intraperitoneal, or parenteral routes, including intracranial (e.g., intraventricular, intraparenchymal and intrathecal), intramuscular, transdermal, airway (aerosol), nasal, rectal, and topical (including buccal and sublingual) administration. In certain embodiments, the compositions are administered by subcutaneous or intravenous infusion or injection. In one embodiment, the compositions are administered by subcutaneous injection.

[0165] In some embodiments, the administration is via a depot injection. A depot injection may release the RNAi in a consistent way over a prolonged time period. Thus, a depot injection may reduce the frequency of dosing needed to obtain a desired effect, e.g., a desired inhibition of PNPLA3, or a therapeutic or prophylactic effect. A depot injection may also provide more consistent serum concentrations. Depot injections may include subcutaneous injections or intramuscular injections. In preferred embodiments, the depot injection is a subcutaneous injection.

[0166] In some embodiments, the administration is via a pump. The pump may be an external pump or a surgically implanted pump. In certain embodiments, the pump is a subcutaneously implanted osmotic pump. In other embodiments, the pump is an infusion pump. An infusion pump may be used for intravenous, subcutaneous, arterial, or epidural infusions. In preferred embodiments, the infusion pump is a subcutaneous infusion pump. In other embodiments, the pump is a surgically implanted pump that delivers the RNAi to the subject.

[0167] The mode of administration may be chosen based upon whether local or systemic treatment is desired and based upon the area to be treated. The route and site of administration may be chosen to enhance targeting.

[0168] In one aspect, the present invention also provides methods for inhibiting the expression of an PNPLA3 gene in a mammal, e.g., a human. The present invention also provides a composition comprising an RNAi, e.g., a dsRNA, that targets an PNPLA3 gene in a cell of a mammal for use in inhibiting expression of the PNPLA3 gene in the mammal. In another aspect, the present invention provides use of an RNAi, e.g., a dsRNA, that targets an PNPLA3 gene in a cell of a mammal in the manufacture of a medicament for inhibiting expression of the PNPLA3 gene in the mammal.

[0169] The methods and uses include administering to the mammal, e.g., a human, a composition comprising an RNAi, e.g., a dsRNA, that targets an PNPLA3 gene in a cell of the mammal and maintaining the mammal for a time sufficient to obtain degradation of the mRNA transcript of the PNPLA3 gene, thereby inhibiting expression of the PNPLA3 gene in the mammal.

[0170] Reduction in gene expression can be assessed in peripheral blood sample of the RNAi-administered subject by any methods known in the art, e.g. qRT-PCR, described herein. Reduction in protein production can be assessed by any methods known in the art and by methods, e.g., ELISA or Western blotting, described herein. In one embodiment, a tissue sample serves as the tissue material for monitoring the reduction in PNPLA3 gene and/or protein expression. In another embodiment, a blood sample serves as the tissue material for monitoring the reduction in PNPLA3 gene and/or protein expression.

[0171] In one embodiment, verification of RISC mediated cleavage of target in vivo following administration of RNAi agent is done by performing 5'-RACE or modifications of the protocol as known in the art (Lasham A et al., (2010) Nucleic Acid Res., 38 (3) p-e19) (Zimmermann et al. (2006) Nature 441: 111-4).

[0172] It is understood that all ribonucleic acid sequences disclosed herein can be converted to deoxyribonucleic acid sequences by substituting a thymine base for a uracil base in the sequence. Likewise, all deoxyribonucleic acid sequences disclosed herein can be converted to ribonucleic acid sequences by substituting a uracil base for a thymine base in the sequence. Deoxyribonucleic acid sequences, ribonucleic acid sequences, and sequences containing mixtures of deoxyribonucleotides and ribonucleotides of all sequences disclosed herein are included in the invention.

[0173] Additionally, any nucleic acid sequences disclosed herein may be modified with any combination of chemical modifications. One of skill in the art will readily appreciate that such designation as "RNA" or "DNA" to describe modified polynucleotides is, in certain instances, arbitrary. For example, a polynucleotide comprising a nucleotide having a 2'-OH substituent on the ribose sugar and a thymine base could be described as a DNA molecule having a modified sugar (2'-OH for the natural 2'-H of DNA) or as an RNA molecule having a modified base (thymine (methylated uracil) for natural uracil of RNA).

[0174] Accordingly, nucleic acid sequences provided herein, including, but not limited to those in the sequence listing, are intended to encompass nucleic acids containing any combination of natural or modified RNA and/or DNA, including, but not limited to such nucleic acids having modified nucleobases. By way of a further example and without limitation, a polynucleotide

having the sequence "ATCGATCG" encompasses any polynucleotides having such a sequence, whether modified or unmodified, including, but not limited to, such compounds comprising RNA bases, such as those having sequence "AUCGAUCG" and those having some DNA bases and some RNA bases such as "AUCGATCG" and polynucleotides having other modified bases, such as "ATmeCGAUCG," wherein meC indicates a cytosine base comprising a methyl group at the 5-position.

[0175] The following examples, including the experiments conducted and the results achieved, are provided for illustrative purposes only and are not to be construed as limiting the scope of the appended claims.

INCORPORATION BY REFERENCE

[0176] All publications, patents, and patent applications mentioned in this specification are herein incorporated by reference to the same extent as if each individual publication, patent, or patent application was specifically and individually indicated to be incorporated by reference. However, the citation of a reference herein should not be construed as an acknowledgement that such reference is prior art to the present invention. To the extent that any of the definitions or terms provided in the references incorporated by reference differ from the terms and discussion provided herein, the present terms and definitions control.

EQUIVALENTS

[0177] The foregoing written specification is considered to be sufficient to enable one skilled in the art to practice the invention. The foregoing description and examples detail certain preferred embodiments of the invention and describe the best mode contemplated by the inventors. It will be appreciated, however, that no matter how detailed the foregoing may appear in text, the invention may be practiced in many ways and the invention should be construed in accordance with the appended claims and any equivalents thereof.

[0178] The following examples, including the experiments conducted and results achieved, are provided for illustrative purposes only and are not to be construed as limiting the present invention.

EXAMPLE 1: Selection, Design and Synthesis of Modified PNPLA3 siRNA molecules

[0001] The identification and selection of optimal sequences for therapeutic siRNA molecules targeting patatin-like phospholipase domain-containing 3 (PNPLA3) were identified using bioinformatics analysis of a human PNPLA3 transcript (NM_025225.2). Table 1 shows sequences identified as having therapeutic properties. Throughout the various sequences, {INVAB} is an inverted A basic, {INVDA} is an inverted deoxythymidine, GNA is a glycol nucleic acid, dT is deoxythymidine and dC is deoxycytosine.

Table 1. siRNA sequences directed to PNPLA3

Duplex No.	Sense sequence (5'-3')	SEQ ID NO: (sense)	Antisense sequence (5'-3')	SEQ ID NO: (antisense)
D-1000	GGGCAAUAAAGUACCUGCUUU	1	AGCAGGUACUUUAUUGCCCUU	2
D-1001	CGGCCAAUGUCCACCAGCUUU	3	AGCUGGUGGACAUUGGCCGUU	4
D-1002	GGUCCAGCCUGAACUUCUUUU	5	AAGAAGUUCAGGCUGGACCUU	6
D-1003	GCUUCAUCCCCUUCACAGUU	7	CUGUAGAAGGGGAUGAAGCUU	8
D-1004	GCGGCUUCCUGGGCUUCUAUU	9	UAGAAGCCCAGGAAGCCGCUU	10
D-1005	GCCUCUGAGCUGAGUUGGUUU	11	ACCAACUCAGCUCAGAGGCUU	12
D-1006	GUGACAACGUACCCUUCAUUU	13	AUGAAGGGUACGUUGUCACUU	14
D-1007	CCCGCCUCCAGGUCCCAAUU	15	UUUGGGACCUGGAGGCGGGUU	16
D-1008	CUUCAUCCCCUUCACAGUUU	17	ACUGUAGAAGGGGAUGAAGUU	18
D-1009	GGUAUGUUCUGCUUCAUGUU	19	CAUGAAGCAGGAACAUACCUU	20
D-1010	GUAUGUUCUGCUUCAUGCUU	21	GCAUGAAGCAGGAACAUACUU	22
D-1011	UAUGUUCUGCUUCAUGCCUU	23	GGCAUGAAGCAGGAACAUUU	24
D-1012	AUGUUCUGCUUCAUGCCCUU	25	GGGCAUGAAGCAGGAACAUUU	26

D-1013	UGUCCUGCUUCAUGCCCUUU	27	AGGGCAUGAAGCAGGAACAUU	28
D-1014	GUUCCUGCUUCAUGCCCUUUU	29	AAGGGCAUGAAGCAGGAACUU	30
D-1015	UUCUGCUUCAUGCCCUUCUU	31	GAAGGGCAUGAAGCAGGAAUU	32
D-1016	UCCUGCUUCAUGCCCUUCUUU	33	AGAAGGGCAUGAAGCAGGAUU	34
D-1017	CCUGCUUCAUGCCCUUCUAUU	35	UAGAAGGGCAUGAAGCAGGUU	36
D-1018	CUGCUUCAUGCCCUUCUACUU	37	GUAGAAGGGCAUGAAGCAGUU	38
D-1019	UGCUCUUGCCCUUCUACAUU	39	UGUAGAAGGGCAUGAAGCAUU	40
D-1020	GCUCUUGCCCUUCUACAGUU	41	CUGUAGAAGGGCAUGAAGCUU	42
D-1021	CUUCUUGCCCUUCUACAGUUU	43	ACUGUAGAAGGGCAUGAAGUU	44
D-1022	UUCUUGCCCUUCUACAGUGUU	45	CACUGUAGAAGGGCAUGAAUU	46
D-1023	UCAUGCCCUUCUACAGUGGUU	47	CCACUGUAGAAGGGCAUGAUU	48
D-1024	CAUGCCCUUCUACAGUGGCUU	49	GCCACUGUAGAAGGGCAUGUU	50
D-1025	AUGCCCUUCUACAGUGGCCUU	51	GGCCACUGUAGAAGGGCAUUU	52
D-1026	UGCCCUUCUACAGUGGCCUUU	53	AGGCCACUGUAGAAGGGCAUU	54
D-1027	GCCCUUCUACAGUGGCCUUUU	55	AAGGCCACUGUAGAAGGGCUU	56
D-1028	GGUAUGUUCUGCUUCAUCUU	57	GAUGAAGCAGGAACAUACCUU	58
D-1029	GUAUGUUCUGCUUCAUCCUU	59	GGAUGAAGCAGGAACAUACUU	60
D-1030	UAUGUUCUGCUUCAUCCUUU	61	GGGAUGAAGCAGGAACAUUU	62
D-1031	AUGUUCUGCUUCAUCCCUUU	63	GGGAUGAAGCAGGAACAUUU	64
D-1032	UGUCCUGCUUCAUCCCUUUU	65	AGGGGAUGAAGCAGGAACAUU	66
D-1033	GUUCCUGCUUCAUCCCUUUU	67	AAGGGGAUGAAGCAGGAACUU	68
D-1034	UUCUGCUUCAUCCCUUCUU	69	GAAGGGGAUGAAGCAGGAAUU	70
D-1035	UCCUGCUUCAUCCCUUCUUU	71	AGAAGGGGAUGAAGCAGGAUU	72
D-1036	CCUGCUUCAUCCCUUCUAUU	73	UAGAAGGGGAUGAAGCAGGUU	74
D-1037	CUGCUUCAUCCCUUCUACUU	75	GUAGAAGGGGAUGAAGCAGUU	76
D-1038	UGCUCUUAUCCCUUCUACAUU	77	UGUAGAAGGGGAUGAAGCAUU	78

D-1039	UUCAUCCCCUUCUACAGUGUU	79	CACUGUAGAAGGGGAUGAAUU	80
D-1040	UCAUCCCCUUCUACAGUGGUU	81	CCACUGUAGAAGGGGAUGAUU	82
D-1041	CAUCCCCUUCUACAGUGGCUU	83	GCCACUGUAGAAGGGGAUGUU	84
D-1042	UCCCCUUCUACAGUGGCCUUU	85	AGGCCACUGUAGAAGGGGAUU	86
D-1043	GAUCAGGACCCGAGCCGAUUU	87	AUCGGCUCGGGUCCUGAUCUU	88
D-1044	UGGGCUUCUACCACGUCGUUU	89	ACGACGUGGUAGAAGCCAUU	90
D-1045	GAGCGAGCACGCCCCGCAUUU	91	AUGCGGGGCGUGCUCGCUCUU	92
D-1046	UGCACUGCGUCGGGCGUCCUU	93	AGGACGCGCGACGCAGUGCAUU	94
D-1047	UGGAGCAGACUCUGCAGGUUU	95	ACCUGCAGAGUCUGCUCCAUU	96
D-1048	UGCAGGUCCUCUCAGAUCUUU	97	AGAUCUGAGAGGACCUGCAUU	98
D-1049	CCCGGCCAAUGUCCACCAUUU	99	AUGGUGGACAUUGGCCGGGUU	100
D-1050	UUCUACAGUGGCCUUAUCUUU	101	AGAUAAAGGCCACUGUAGAAUU	102
D-1051	UCUACAGUGGCCUUAUCCUUU	103	AGGAUAAGGCCACUGUAGAUU	104
D-1052	CUUCCUUCAGAGGCUGUCUUU	105	AGCACGCCUCUGAAGGAAGUU	106
D-1053	UUCUUCAGAGGCUGCGAUU	107	UCGCACGCCUCUGAAGGAUU	108
D-1054	GCGUGCGAUUAUGUGGAUGUUU	109	ACAUCCACAUAUCGCACGCUU	110
D-1055	CGUGCGAUUAUGUGGAUGGAUU	111	UCCAUCCACAUAUCGCACGUU	112
D-1056	UGGAUGGAGGAGUGAGUGAUU	113	UCACUCACUCCUCCAUCAUU	114
D-1057	ACGUACCCUUCAUUGAUGUUU	115	ACAUCAAUGAAGGGUACGUUU	116
D-1058	UGGACAUCACCAAGCUCAUUU	117	AUGAGCUUGGUGAUGUCCAUU	118
D-1059	CACCGCGUCUCAGCAUCUUU	119	AGAUGCUGAGACGCAGGUGUU	120
D-1060	ACCUGCGUCUCAGCAUCCUUU	121	AGGAUGCUGAGACGCAGGUUU	122
D-1061	CCAGAGACUGGUGACAUGUUU	123	ACAUGUACACAGUCUCUGGUU	124
D-1062	AUGGCUUCCAGAUUAUGCCUUU	125	AGGCAUAUCUGGAAGCCAUUU	126
D-1063	CCGCCUCCAGGUCCAAAUUU	127	AUUUGGGACCUGGAGGCGGUU	128
D-1064	UACCUGCGUGUGCUGAGGUUU	129	ACCUCAGCACCAGCAGGUUU	130

D-1065	ACCUGCUGGUGCUGAGGGUUU	131	ACCCUCAGCACCAGCAGGUUU	132
D-1066	CUCUCCACCUUCCAGUUUU	133	AACUGGGAAGGUGGAGAGUU	134
D-1067	UUUUUACCUAACUAAAAUUU	135	AUUUUAGUUAGGUGAAAAUU	136
D-1068	CGGCCAAUGUCCACCAGCUUU	137	AGCUGGUGGACAUUGGCCGUU	138
D-1069	GGUCCAGCCUGAACUUCUUUU	139	AAGAAGUUCAGGCUGGACCUU	140
D-1070	GCGGCUUCCUGGGCUUCUAUU	141	UAGAAGCCCAGGAAGCCGCUU	142
D-1071	GUGACAACGUACCCUUCAUUU	143	AUGAAGGGUACGUUGUCACUU	144
D-1072	GGUAUGUUCUGCUUCAUGUU	145	CAUGAAGCAGGAACAUACCUU	146
D-1073	GUAUGUUCUGCUUCAUGCUU	147	GCAUGAAGCAGGAACAUACUU	148
D-1074	UGUUCUGCUUCAUGCCCUUU	149	AGGGCAUGAAGCAGGAACAUU	150
D-1075	GUUCCUGCUUCAUGCCCUUUU	151	AAGGGCAUGAAGCAGGAACUU	152
D-1076	CCUGCUUCAUGCCCUUCUAUU	153	UAGAAGGGCAUGAAGCAGGUU	154
D-1077	GCUUCAUGCCCUUCUACAGUU	155	CUGUAGAAGGGCAUGAAGCUU	156
D-1078	CUUCAUGCCCUUCUACAGUUU	157	ACUGUAGAAGGGCAUGAAGUU	158
D-1079	UUCAUGCCCUUCUACAGUGUU	159	CACUGUAGAAGGGCAUGAAUU	160
D-1080	AUGGCUUCCAGAUAGCCUUU	161	AGGCAUAUCUGGAAGCCAUUU	162
D-1081	AUGCCCUUCUACAGUGGCCUU	163	GGCCACUGUAGAAGGGCAUUU	164
D-1082	GCUUCAUGCCCUUCUACAUUU	165	AUGUAGAAGGGCAUGAAGCUU	166
D-1083	GGAAAGACUGUUCAAAAUUU	333	UUUUUGGAACAGUCUUUCCUU	334
D-1084	GGUAUGUUCUGCUUCAUGUU	335	CAUGAAGCAGGAACAUACCUU	336
D-1085	GUAUGUUCUGCUUCAUGCUU	337	GCAUGAAGCAGGAACAUACUU	338
D-1086	UGUUCUGCUUCAUGCCCUUU	339	AGGGCAUGAAGCAGGAACAUU	340
D-1087	GCUUCAUGCCCUUCUACAGUU	341	CUGUAGAAGGGCAUGAAGCUU	342
D-1088	CUUCAUGCCCUUCUACAGUUU	343	ACUGUAGAAGGGCAUGAAGUU	344
D-1089	GCGGCUUCCUGGGCUUCUAUU	345	UAGAAGCCCAGGAAGCCGCUU	346
D-1090	GUUCCUGCUUCAUGCCCUUUU	347	AAGGGCAUGAAGCAGGAACUU	348

D-1091	AUGGCUUCCAGAU AUGCCUUU	349	AGGCAUAUCUGGAAGCCAUUU	350
D-1092	CCUGCUUCAUGCCCUUCAUU	351	UAGAAGGGCAUGAAGCAGGUU	352
D-1093	UUCAUGCCCUUCUACAGUUUU	353	AACUGUAGAAGGGCAUGAAUU	354
D-1094	UUCAUGCCCUUCUACAGUGUU	355	CACUGUAGAAGGGCAUGAAUU	356
D-1095	GCUUCAUGCCCUUCUACAUUU	357	AUGUAGAAGGGCAUGAAGCUU	358
D-1096	GGUCCAGCCUGAACUUCUUUU	359	AAGAAGUUCAGGCGGACCUU	360
D-1097	GCGGCUUCCUGGGCUUCAUU	361	UAGAAGCCCAGGAAGCCGCUU	362
D-1098	GCGGCUUCCUGGGCUUCAUU	363	UAGAAGCCCAGGAAGCCGCUU	364
D-1099	GUUCCUGCUUCAUGCCCUUUU	365	AAGGGCAUGAAGCAGGAACUU	366
D-1100	GUUCCUGCUUCAUGCCCUUUU	367	AAGGGCAUGAAGCAGGAACUU	368
D-1101	CCUGCUUCAUGCCCUUCAUU	369	UAGAAGGGCAUGAAGCAGGUU	370
D-1102	CCUGCUUCAUGCCCUUCAUU	371	UAGAAGGGCAUGAAGCAGGUU	372
D-1103	AUGCCCUUCUACAGUGGCCUU	373	GGCCACUGUAGAAGGGCAUUU	374
D-1104	AUGGCUUCCAGAU AUGCCUUU	375	AGGCAUAUCUGGAAGCCAUUU	376
D-1105	GUGACAACGUACCCUUCAUUU	377	AUGAAGGGUACGUUGUCACUU	378
D-1106	GUAUGUCCUGCUUCAUGCUU	379	GCAUGAAGCAGGAACAUACUU	380
D-1107	GUAUGUCCUGCUUCAUGCUU	381	GCAUGAAGCAGGAACAUACUU	382
D-1108	GUAUGUCCUGCUUCAUGCUU	383	GCAUGAAGCAGGAACAUACUU	384
D-1109	GUAUGUCCUGCUUCAUGCCU	385	AGGCAUGAAGCAGGAACAUACUU	386
D-1110	UGGUAUGUCCUGCUUCAUGU	387	GCAUGAAGCAGGAACAUACCUU	388
D-1111	GUAUGUCCUGCUUCAUGU	389	GCAUGAAGCAGGAACAUACUU	390
D-1112	GUAUGUCCUGCUUCAUGC { INVAB }	391	GCAUGAAGCAGGAACAUACUU	392
D-1113	GCUUCAUGCCCUUCUACAUUU	393	AUGUAGAAGGGCAUGAAGCUU	394
D-1114	GCUUCAUGCCCUUCUACAUUU	395	AUGUAGAAGGGCAUGAAGCUU	396
D-1115	GCUUCAUGCCCUUCUACAUUU	397	AUGUAGAAGGGCAUGAAGCUU	398
D-1116	GCUUCAUGCCCUUCUACAUUU	399	AUGUAGAAGGGCAUGAAGCUU	400

D-1117	GCUUCAUGCCCUUCUACAUUU	401	AUGUAGAAGGGCAUGAAGCUU	402
D-1118	GCUUCAUGCCCUUCUACAUUU	403	AUGUAGAAGGGCAUGAAGCUU	404
D-1119	GCUUCAUGCCCUUCUACAUUU	405	AUGUAGAAGGGCAUGAAGCUU	406
D-1120	GCUUCAUGCCCUUCUACAUUU	407	AUGUAGAAGGGCAUGAAGCUU	408
D-1121	GCUUCAUGCCCUUCUACAUUU	409	AUGUAGAAGGGCAUGAAGCUU	410
D-1122	GCUUCAUGCCCUUCUACAUUU	411	AUGUAGAAGGGCAUGAAGCUU	412
D-1123	GCUUCAUGCCCUUCUACAUUU	413	AUGUAGAAGGGCAUGAAGCUU	414
D-1124	GCUUCAUGCCCUUCUACAUUU	415	AUGUAGAAGGGCAUGAAGCUU	416
D-1125	GCUUCAUGCCCUUCUACAUUU	417	AUGUAGAAGGGCAUGAAGCUU	418
D-1126	GCUUCAUGCCCUUCUACAUUU	419	AUGUAGAAGGGCAUGAAGCUU	420
D-1127	GCUUCAUGCCCUUCUACAUUU	421	AUGUAGAAGGGCAUGAAGCUU	422
D-1128	GCUUCAUGCCCUUCUACAUUU	423	AUGUAGAAGGGCAUGAAGCUU	424
D-1129	GCUUCAUG [DC] CCUUCUACAUUU	425	AUGUAGAAGGGCAUGAAGCUU	426
D-1130	GCUUCAUGCC [DC] UUCUACAUUU	427	AUGUAGAAGGGCAUGAAGCUU	428
D-1131	GCUUCAUGC [DC] CUUCUACAUUU	429	AUGUAGAAGGGCAUGAAGCUU	430
D-1132	GCUUCAUGCCCUUCUACAUUU	431	AUGUAGAAGGGCAUGAAGCUU	432
D-1133	GCUUCAUGCCCUUCUACAUUU	433	AUGUAGAAGGGCAUGAAGCUU	434
D-1134	GCUUCAUGCCCUUCUACAUUU	435	AUGUAGAAGGGCAUGAAGCUU	436
D-1135	GCUUCAUGCCCUUCUACAUUU	437	AUGUAGAAGGGCAUGAAGCUU	438
D-1136	GCUUCAUGCCCUUCUACAUUU	439	AUGUAGAAGGGCAUGAAGCUU	440
D-1137	GCUUCAUGCCCUUCUACAGUU	441	AACUGUAGAAGGGCAUGAAGCUU	442
D-1138	CUGCUUCAUGCCCUUCUACAU	443	AUGUAGAAGGGCAUGAAGCAGUU	444
D-1139	GCUUCAUGCCCUUCUACAU	445	AUGUAGAAGGGCAUGAAGCUU	446
D-1140	GCUUCAUGCCCUUCUACAU { INVAB }	447	AUGUAGAAGGGCAUGAAGCUU	448
D-1141	GCUUCAUGCCCUUCUACAUU { INVAB }	449	AUGUAGAAGGGCAUGAAGCUU	450
D-1142	GCUUCAUGCCCUUCUACAUUU	451	AUGUAGAAGGGCAUGAAGCUU	452

D-1143	GCUUCAUGCCCUUCUACAUUU	453	AUGUAGAAGGGCAUGAAGCUU	454
D-1144	GCUUCAUGCCCUUCUACAUUU	455	AUGUAGAAGGGCAUGAAGCUU	456
D-1145	GCUUCAUGCCCUUCUACAUUU	457	AUGUAGAAGGGCAUGAAGCUU	458
D-1146	GCUUCAUGCCCUUCUACAUUU	459	AUGUAGAAGGGCAUGAAGCUU	460
D-1147	GCUUCAUGCCCUUCUACAUUU	461	AUGUAGAAGGGCAUGAAGCUU	462
D-1148	GCUUCAUGCCCUUCUACAUUU	463	AUGUAGAAGGGCAUGAAGCUU	464
D-1149	GCUUCAUGCCCUUCUACAUUU	465	AUGUAGAAGGGCAUGAAGCUU	466
D-1150	GCUUCAUGCCCUUCUACAUUU	467	AUGUAGAAGGGCAUGAAGCUU	468
D-1151	GUAUGUUCUGCUUCAUGCUU{ INVAB }	469	GCAUGAAGCAGGAACAUACUU	470
D-1152	GGUAUGUUCUGCUUCAUUUU	471	AAUGAAGCAGGAACAUACUU	472
D-1153	GUAUGUUCUGCUUCAUGUUU	473	ACAUGAAGCAGGAACAUACUU	474
D-1154	CGGCCAAUGUCCACCAGCUUU	475	AGCUGGUGGACAUUGGCCGUU	476
D-1155	UGGAGCAGACUCUGCAGGUUU	477	ACCUGCAGAGUCUGCUCCAUU	478
D-1156	ACGUACCCUUCUAUGAUGUUU	479	ACAUCAAUGAAGGGUACGUUU	480
D-1157	CCAGAGACUGGUGACAUGUUU	481	ACAUGUCACCAGUCUCUGGUU	482
D-1158	[INVAB] GCUUCAUGCCCUUCUACAUUU	483	AUGUAGAAAGGCAUGAAGCUU	484
D-1159	[INVAB] GCUUCAUGCCCUUCUACAUUU	485	AAAUGUAGAAAGGCAUGAAGCUU	486
D-1160	[INVAB] GCUUCAUGCCCUUCUACAUUU	487	AAAUGUAGAAAGGCAUGAAGCUU	488
D-1161	UGCUUCAUGCCCUUCUACAUU	489	UGUAGAAAGGCAUGAAGCAUU	490
D-1162	UAUGUUCUGCUUCAUGCUUU	491	AGCAUGAAGCAGGAACAUUUU	492
D-1163	UUCCUGCUUCAUGCCUUUUUU	493	AAAAGGCAUGAAGCAGGAAUU	494
D-1164	UCAUGCCCUUCUACAGUGUUU	495	ACACUGUAGAAAGGCAUGAUU	496
D-1165	CAUGCCCUUCUACAGUGGUUU	497	ACCACUGUAGAAAGGCAUGUU	498
D-1166	AUGCCUUUCUACAGUGGCUUU	499	AGCCACUGUAGAAAGGCAUUU	500
D-1167	GGUAUGUUCUGCUUCAUAUU	501	UAUGAAGCAGGAACAUACUU	502
D-1168	GUAUGUUCUGCUUCAUGAUU	503	UCAUGAAGCAGGAACAUACUU	504

D-1169	UAUGUUCUGCUUCAUGCAUU	505	UGCAUGAAGCAGGAACAUAUU	506
D-1170	UUCUGCUUCAUGCCUUUAUU	507	UAAAGGCAUGAAGCAGGAAUU	508
D-1171	CUGCUUCAUGCCUUUCUAAUU	509	UUAGAAAGGCAUGAAGCAGUU	510
D-1172	GCUUCAUGCCUUUCUACAAUU	511	UUGUAGAAAGGCAUGAAGCUU	512
D-1173	UUCAUGCCUUUCUACAGUAUU	513	UACUGUAGAAAGGCAUGAAUU	514
D-1174	UCAUGCCUUUCUACAGUGAUU	515	UCACUGUAGAAAGGCAUGAUU	516
D-1175	CAUGCCUUUCUACAGUGGAUU	517	UCCACUGUAGAAAGGCAUGUU	518
D-1176	AUGCCUUUCUACAGUGGCAUU	519	UGCCACUGUAGAAAGGCAUUU	520
D-1177	ACGUACCCUUAUUGAUGAUU	521	UCAUCAUGAAGGGUACGUUU	522
D-1178	CCAGAGACUGGUGACAUGAUU	523	UCAUGUCACCAGUCUCUGGUU	524
D-1179	AUGGCUUCAGAUUUGCCAUU	525	UGGCAUAUCUGGAAGCCAUUU	526
D-1180	GUUCCUGCUUCAUGCCUUUUU	527	AAAGGCAUGAAGCAGGAACUU	528
D-1181	CCUGCUUCAUGCCUUUCUAUU	529	UAGAAAGGCAUGAAGCAGGUU	530
D-1182	GCUUCAUGCCUUUCUACAUUU	531	AUGUAGAAAGGCAUGAAGCUU	532
D-1183	CUUCAUGCCUUUCUACAGUUU	533	ACUGUAGAAAGGCAUGAAGUU	534
D-1184	UUCAUGCCUUUCUACAGUUUU	535	AACUGUAGAAAGGCAUGAAUU	536
D-1185	GCUUCAUCCUUUCUACAUUU	537	AUGUAGAAAGGGAUGAAGCUU	538
D-1186	[INVAB] GCUUCAUGCCUUUCUACAUUU	539	AUGUAGAAGGGCAUGAAGCUU	540
D-1187	[INVAB] GCUUCAUGCCUUUCUACAUUU	541	AAAUGUAGAAGGGCAUGAAGCUU	542
D-1188	[INVAB] GCUUCAUGCCUUUCUACAUUU	543	AAAUGUAGAAGGGCAUGAAGCUU	544
D-1189	[INVAB] GCGGCUUCCUGGGCUUCUAUU	545	UAGAAGCCCAGGAAGCCGCUU	546
D-1190	[INVAB] CUGCGGCUUCCUGGGCUUCUA	547	UAGAAGCCCAGGAAGCCGCAGUU	548
D-1191	CUGCGGCUUCCUGGGCUUCU{ INVAB }	549	UAGAAGCCCAGGAAGCCGCAGUU	550
D-1192	[INVAB] CUGCGGCUUCCUGGGCUUCUA	551	UAGAAGCCCAGGAAGCCGCAGUU	552
D-1193	[INVAB] GCGGCUUCCUGGGCUUCUAUU	553	UAGAAGCCCAGGAAGCCGCUU	554
D-1194	CUGCGGCUUCCUGGGCUUCU{ INVAB }	555	UAGAAGCCCAGGAAGCCGCAGUU	556

D-1195	[INVAB] CUGCGGCUUCCUGGGCUUCUA	557	UAGAAGCCCAGGAAGCCGCAGUU	558
D-1196	[INVAB] GCGGCUUCCUGGGCUUCUAUU	559	UAGAAGCCCAGGAAGCCGCUU	560
D-1197	CUGCGGCUUCCUGGGCUUCU{ INVAB }	561	UAGAAGCCCAGGAAGCCGCAGUU	562
D-1198	[INVAB] CUGCGGCUUCCUGGGCUUCUA	563	UAGAAGCCCAGGAAGCCGCAGUU	564
D-1199	[INVAB] GCGGCUUCCUGGGCUUCUAUU	565	UAGAAGCCCAGGAAGCCGCUU	566
D-1200	[INVAB] CUGCGGCUUCCUGGGCUUCUA	567	UAGAAGCCCAGGAAGCCGCAGUU	568
D-1201	[INVAB] CUGCGGCUUCCUGGGCUUCUA	569	UAGAAGCCCAGGAAGCCGCAGUU	570
D-1202	[INVAB] CUGCGGCUUCCUGGGCUUCUA	571	UAGAAGCCCAGGAAGCCGCAGUU	572
D-1203	[INVAB] AUGGCUUCCAGAU AUGCCUUU	573	AGGCAUAUCUGGAAGCCAUUU	574
D-1204	[INVAB] ACAUGGCUUCCAGAU AUGCCU	575	AGGCAUAUCUGGAAGCCAUUUU	576
D-1205	ACAUGGCUUCCAGAU AUGCC{ INVAB }	577	AGGCAUAUCUGGAAGCCAUUUU	578
D-1206	[INVAB] ACAUGGCUUCCAGAU AUGCCU	579	AGGCAUAUCUGGAAGCCAUUUU	580
D-1207	[INVAB] AUGGCUUCCAGAU AUGCCUUU	581	AGGCAUAUCUGGAAGCCAUUU	582
D-1208	ACAUGGCUUCCAGAU AUGCC{ INVAB }	583	AGGCAUAUCUGGAAGCCAUUUU	584
D-1209	[INVAB] ACAUGGCUUCCAGAU AUGCCU	585	AGGCAUAUCUGGAAGCCAUUUU	586
D-1210	[INVAB] AUGGCUUCCAGAU AUGCCUUU	587	AGGCAUAUCUGGAAGCCAUUU	588
D-1211	ACAUGGCUUCCAGAU AUGCC{ INVAB }	589	AGGCAUAUCUGGAAGCCAUUUU	590
D-1212	[INVAB] ACAUGGCUUCCAGAU AUGCCU	591	AGGCAUAUCUGGAAGCCAUUUU	592
D-1213	[INVAB] AUGGCUUCCAGAU AUGCCUUU	593	AGGCAUAUCUGGAAGCCAUUU	594
D-1214	ACAUGGCUUCCAGAU AUGCC{ INVAB }	595	AGGCAUAUCUGGAAGCCAUUUU	596
D-1215	[INVAB] ACAUGGCUUCCAGAU AUGCCU	597	AGGCAUAUCUGGAAGCCAUUUU	598
D-1216	[INVAB] ACAUGGCUUCCAGAU AUGCCU	599	AGGCAUAUCUGGAAGCCAUUUU	600
D-1217	[INVAB] ACAUGGCUUCCAGAU AUGCCU	601	AGGCAUAUCUGGAAGCCAUUUU	602
D-1218	[INVAB] ACAUGGCUUCCAGAU AUGCCU	603	AGGCAUAUCUGGAAGCCAUUUU	604
D-1219	[INVAB] ACGUACCCUUAUUGAUGUUU	605	ACAUCAAUGAAGGGUACGUUU	606
D-1220	[INVAB] CAACGUACCCUUAUUGAUGU	607	ACAUCAAUGAAGGGUACGUUGU	608

D-1221	[INVAB] CAACGUACCCUUCAUUGAUGU	609	ACAUCAAUGAAGGGUACGUUGUU	610
D-1222	[INVAB] ACGUACCCUUCAUUGAUGUUU	611	ACAUCAAUGAAGGGUACGUUU	612
D-1223	CAACGUACCCUUCAUUGAUG{ INVAB }	613	ACAUCAAUGAAGGGUACGUUGUU	614
D-1224	[INVAB] ACGUACCCUUCAUUGAUGUUU	615	ACAUCAAUGAAGGGUACGUUU	616
D-1225	CAACGUACCCUUCAUUGAUG{ INVAB }	617	ACAUCAAUGAAGGGUACGUUGUU	618
D-1226	[INVAB] CAACGUACCCUUCAUUGAUGU	619	ACAUCAAUGAAGGGUACGUUGUU	620
D-1227	[INVAB] ACGUACCCUUCAUUGAUGUUU	621	ACAUCAAUGAAGGGUACGUUU	622
D-1228	CAACGUACCCUUCAUUGAUG{ INVAB }	623	ACAUCAAUGAAGGGUACGUUGUU	624
D-1229	[INVAB] CAACGUACCCUUCAUUGAUGU	625	ACAUCAAUGAAGGGUACGUUGUU	626
D-1230	[INVAB] CAACGUACCCUUCAUUGAUGU	627	ACAUCAAUGAAGGGUACGUUGUU	628
D-1231	[INVAB] CAACGUACCCUUCAUUGAUGU	629	ACAUCAAUGAAGGGUACGUUGUU	630
D-1232	[INVAB] CAACGUACCCUUCAUUGAUGU	631	ACAUCAAUGAAGGGUACGUUGUU	632
D-1233	CUGCUUCAUGCCUUUCUACAU	633	AUGUAGAAAGGCAUGAAGCAGUU	634
D-1234	CUGCUUCAUGCCUUUCUACAU	635	AUGUAGAAAGGCAUGAAGCAGUU	636
D-1235	[INVAB] GCUUCAUGCCUUUCUACAUUU	637	AUGUAGAAAGGCAUGAAGCUU	638
D-1236	[INVAB] GCUUCAUGCCUUUCUACAUUU	639	AUGUAGAAAGGCAUGAAGCUU	640
D-1237	CUGCUUCAUGCCUUUCUACAU	641	AUGUAGAAAGGCAUGAAGCAGUU	642
D-1238	CUGCUUCAUGCCUUUCUACAU	643	AUGUAGAAAGGCAUGAAGCAGUU	644
D-1239	[INVAB] CUGCUUCAUGCCUUUCUACAU	645	AUGUAGAAAGGCAUGAAGCAGUU	646
D-1240	CUGCUUCAUGCCUUUCUACA{ INVAB }	647	AUGUAGAAAGGCAUGAAGCAGUU	648
D-1241	[INVAB] CUGCUUCAUGCCUUUCUACAU	649	AUGUAGAAAGGCAUGAAGCAGUU	650
D-1242	[INVAB] CUGCUUCAUGC [DC] UUUUCUACAU	651	AUGUAGAAAGGCAUGAAGCAGUU	652
D-1243	CUGCUUCAUGCCUUUCUACAU	653	AUGUAGAAAGGCAUGAAGCAGUU	654
D-1244	[INVAB] CUGCUUCAUGCCUUUCUACAU	655	AUGUAGAAAGGCAUGAAGCAGUU	656
D-1245	CUGCUUCAUGCCUUUCUACAU	657	AUGUAGAAAGGCAUGAAGCAGUU	658
D-1246	CUGCUUCAUGCCUUUCUACA{ INVAB }	659	AUGUAGAAAGGCAUGAAGCAGUU	660

D-1247	[INVAB] CUGCUUCAUGCCUUUCUACAU	661	AUGUAGAAAGGCAUGAAGCAGUU	662
D-1248	CUGCUUCAUGCCUUUCUACAU	663	AUGUAGAAAGGCAUGAAGCAGUU	664
D-1249	CUGCUUCAUGCCUUUCUACA{ INVAB }	665	AUGUAGAAAGGCAUGAAGCAGUU	666
D-1250	[INVAB] CUGCUUCAUGCCUUUCUACAU	667	AUGUAGAAAGGCAUGAAGCAGUU	668
D-1251	CUGCUUCAUGCCUUUCUACA{ INVAB }	669	AUGUAGAAAGGCAUGAAGCAGUU	670
D-1252	[INVAB] CUGCUUCAUGCCUUUCUACAU	671	AUGUAGAAAGGCAUGAAGCAGUU	672
D-1253	[INVAB] GCUUCAUGCCUUUCUACAUUU	673	AUGUAGAAAGGCAUGAAGCUU	674
D-1254	[INVAB] GCUUCAUGCCUUUCUACAUUU	675	AUGUAGAAAGGCAUGAAGCUU	676
D-1255	[INVAB] GCUUCAUGCCUUUCUACAUUU	677	AUGUAGAAAGGCAUGAAGCUU	678
D-1256	[INVAB] GCUUCAUGCCUUUCUACAUUU	679	AUGUAGAAAGGCAUGAAGCUU	680
D-1257	[INVAB] GCUUCAUGCCUUUCUACAUUU	681	AUGUAGAAAGGCAUGAAGCUU	682
D-1258	CUGCUUCAUGCCCUUCUACAU	683	AUGUAGAAAGGCAUGAAGCAGUU	684
D-1259	[INVAB] GCUUCAUGCCCUUCUACAUUU	685	AUGUAGAAAGGCAUGAAGCUU	686
D-1260	[INVAB] GCUUCAUGCCCUUCUACAUUU	687	AUGUAGAAAGGCAUGAAGCUU	688
D-1261	[INVAB] GCUUCAUGCCCUUCUACAUUU	689	AUGUAGAAAGGCAUGAAGCUU	690
D-1262	CUGCUUCAUGCCCUUCUACA{ INVAB }	691	AUGUAGAAAGGCAUGAAGCAGUU	692
D-1263	[INVAB] CUGCUUCAUGCCCUUCUACAU	693	AUGUAGAAAGGCAUGAAGCAGUU	694
D-1264	[INVAB] GCUUCAUGCCCUUCUACAUUU	695	AUGUAGAAAGGCAUGAAGCUU	696
D-1265	AUGUCCUGCUUCAUGCCUUU	697	AGGCAUGAAGCAGGAACAUU	698
D-1266	UGUCCUGCUUCAUGCCUUU	699	AAGGCAUGAAGCAGGAACAUU	700
D-1267	UGCCUUUCUACAGUGGCCUUU	701	AGGCCACUGUAGAAAGGCAUU	702
D-1268	CGUACUUCGUCUUGUAUGUU	703	CAUACAAGGACGAAGUACGUU	704
D-1269	[INVAB] GCUUCAUGC [DC] CUUCUACAUUU	705	AUGUAGAAAGGCAUGAAGCUU	706
D-1270	[INVAB] GCUUCAUGCCCUUCUACAUUU	707	AUGUAGAAAGGCAUGAAGCUU	708
D-1271	[INVAB] GCUUCAUGCCCUUCUACAUUU	709	AUGUAGAAAGGCAUGAAGCUU	710
D-1272	[INVAB] GCUUCAUGCCCUUCUACAUUU	711	AUGUAGAAAGGCAUGAAGCUU	712

D-1273	[INVAB] GCUUCAUGCCCUUCUACAUUU	713	AUGUAGAAGGGCAUGAAGCUU	714
D-1274	[INVAB] GCUUCAUGCCCUUCUACAUUU	715	AUGUAGAAGGGCAUGAAGCUU	716
D-1275	[INVAB] CCUGCUUCAUGCCUUCUUAUU	717	UAGAAAGGCAUGAAGCAGGUU	718
D-1276	[INVAB] CCUGCUUCAUGCCUUCUUAUU	719	UAGAAAGGCAUGAAGCAGGUU	720
D-1277	UCCUGCUUCAUGCCUUCU{ INVAB }	721	UAGAAAGGCAUGAAGCAGGAAUU	722
D-1278	[INVAB] CCUGCUUCAUGCCUUCUUAUU	723	UAGAAAGGCAUGAAGCAGGUU	724
D-1279	UCCUGCUUCAUGCCUUCU{ INVAB }	725	UAGAAAGGCAUGAAGCAGGAAUU	726
D-1280	[INVAB] AUGCCUUCUACAGUGGCUUU	727	AGCCACUGUAGAAAGGCAUUU	728
D-1281	[INVAB] AUGCCUUCUACAGUGGCUUU	729	AGCCACUGUAGAAAGGCAUUU	730
D-1282	[INVAB] AUGCCUUCUACAGUGGCUUU	731	AGCCACUGUAGAAAGGCAUUU	732
D-1283	UCAUGCCUUCUACAGUGGC{ INVAB }	733	AGCCACUGUAGAAAGGCAUGAUU	734
D-1284	[INVAB] AUGCCUUCUACAGUGGCUUU	735	AGCCACUGUAGAAAGGCAUUU	736
D-1285	[INVAB] UCAUGCCUUCUACAGUGGCU	737	AGCCACUGUAGAAAGGCAUGAUU	738
D-1286	[INVAB] UGCUUCAUGCCUUCUACAGU	739	ACUGUAGAAAGGCAUGAAGCAUU	740
D-1287	[INVAB] CUUCAUGCCUUCUACAGUUU	741	ACUGUAGAAAGGCAUGAAGUU	742
D-1289	UGCUCUUGCCUUCUACAG{ INVAB }	743	ACUGUAGAAAGGCAUGAAGCAUU	744
D-1290	[INVAB] CUUCAUGCCUUCUACAGUUU	745	ACUGUAGAAAGGCAUGAAGUU	746
D-1291	UGCUCUUGCCUUCUACAG{ INVAB }	747	ACUGUAGAAAGGCAUGAAGCAUU	748
D-1292	[INVAB] UGCUUCAUGCCUUCUACAGU	749	ACUGUAGAAAGGCAUGAAGCAUU	750
D-1293	[INVAB] UGCUUCAUGCCUUCUACAGU	751	ACUGUAGAAAGGCAUGAAGCAUU	752
D-1294	[INVAB] UGCUUCAUGCCUUCUACAGU	753	ACUGUAGAAAGGCAUGAAGCAUU	754
D-1295	[INVAB] UGCUUCAUGCCUUCUACAGU	755	ACUGUAGAAAGGCAUGAAGCAUU	756
D-1296	[INVAB] GUCCUGCUUCAUGCCUUUUU	757	AAAGGCAUGAAGCAGGAACUU	758
D-1297	[INVAB] CCUGCUUCAUGCCUUCUUAUU	759	UAGAAAGGCAUGAAGCAGGUU	760
D-1298	[INVAB] CUUCAUGCCUUCUACAGUUU	761	ACUGUAGAAAGGCAUGAAGUU	762
D-1299	[INVAB] UCCUGCUUCAUGCCUUCUA	763	UAGAAAGGCAUGAAGCAGGAAUU	764

D-1300	UCCUGCUUCAUGCCUUUCU{ INVAB }	765	UAGAAAGGCAUGAAGCAGGAAUU	766
D-1301	[INVAB] UCCUGCUUCAUGCCUUUCUA	767	UAGAAAGGCAUGAAGCAGGAAUU	768
D-1302	UCCUGCUUCAUGCCUUUCU{ INVAB }	769	UAGAAAGGCAUGAAGCAGGAAUU	770
D-1303	[INVAB] UCCUGCUUCAUGCCUUUCUA	771	UAGAAAGGCAUGAAGCAGGAAUU	772
D-1304	[INVAB] UCCUGCUUCAUGCCUUUCUA	773	UAGAAAGGCAUGAAGCAGGAAUU	774
D-1305	[INVAB] UCCUGCUUCAUGCCUUUCUA	775	UAGAAAGGCAUGAAGCAGGAAUU	776
D-1306	[INVAB] UCCUGCUUCAUGCCUUUCUA	777	UAGAAAGGCAUGAAGCAGGAAUU	778
D-1307	[INVAB] UCCUGCUUCAUGCCUUUCUA	779	UAGAAAGGCAUGAAGCAGGAAUU	780
D-1308	[INVAB] UCCUGCUUCAUGCCUUUCUA	781	UAGAAAGGCAUGAAGCAGGAAUU	782
D-1309	[INVAB] UCAUGCCUUUCUACAGUGGCU	783	AGCCACUGUAGAAAGGCAUGAUU	784
D-1310	UCAUGCCUUUCUACAGUGGC{ INVAB }	785	AGCCACUGUAGAAAGGCAUGAUU	786
D-1311	[INVAB] UCAUGCCUUUCUACAGUGGCU	787	AGCCACUGUAGAAAGGCAUGAUU	788
D-1312	UCAUGCCUUUCUACAGUGGC{ INVAB }	789	AGCCACUGUAGAAAGGCAUGAUU	790
D-1313	[INVAB] UCAUGCCUUUCUACAGUGGCU	791	AGCCACUGUAGAAAGGCAUGAUU	792
D-1314	[INVAB] UCAUGCCUUUCUACAGUGGCU	793	AGCCACUGUAGAAAGGCAUGAUU	794
D-1315	UCAUGCCUUUCUACAGUGGC{ INVAB }	795	AGCCACUGUAGAAAGGCAUGAUU	796
D-1316	[INVAB] UCAUGCCUUUCUACAGUGGCU	797	AGCCACUGUAGAAAGGCAUGAUU	798
D-1317	[INVAB] UCAUGCCUUUCUACAGUGGCU	799	AGCCACUGUAGAAAGGCAUGAUU	800
D-1318	[INVAB] UCAUGCCUUUCUACAGUGGCU	801	AGCCACUGUAGAAAGGCAUGAUU	802
D-1319	[INVAB] UGCUUCAUGCCUUUCUACAGU	803	ACUGUAGAAAGGCAUGAAGCAUU	804
D-1320	UGCUUCAUGCCUUUCUACAG{ INVAB }	805	ACUGUAGAAAGGCAUGAAGCAUU	806
D-1321	UGCUUCAUGCCUUUCUACAG{ INVAB }	807	ACUGUAGAAAGGCAUGAAGCAUU	808
D-1322	[INVAB] UGCUUCAUGCCUUUCUACAGU	809	ACUGUAGAAAGGCAUGAAGCAUU	810
D-1323	[INVAB] CUUCAUGCCUUUCUACAGUUU	811	ACUGUAGAAAGGCAUGAAGUUU	812
D-1324	[INVAB] UGCUUCAUGCCUUUCUACAGU	813	ACUGUAGAAAGGCAUGAAGCAUU	814
D-1325	[INVAB] GCUUCAUGCCUUUCUACAUUU	815	AUGUAGAAAGGCAUGAAGCUU	816

D-1326	[INVAB] CAUGCCUUCUACAGUGGUUU	817	ACCACUGUAGAAAGGCAUGUU	818
D-1327	[INVAB] CUGCUUCAUGCCUUCUAAUU	819	UUAGAAAGGCAUGAAGCAGUU	820
D-1328	[INVAB] GCUUCAUGCCUUCUACAAUU	821	UUGUAGAAAGGCAUGAAGCUU	822
D-1329	[INVAB] UUCAUGCCUUCUACAGUAUU	823	UACUGUAGAAAGGCAUGAAUU	824
D-1330	[INVAB] GUUCCUGCUUCAUGCCUUUU	825	AAAGGCAUGAAGCAGGAACUU	826
D-1331	AUGUCCUGCUUCAUGCCUU{ INVAB }	827	AAAGGCAUGAAGCAGGAACAUUU	828
D-1332	[INVAB] AUGUCCUGCUUCAUGCCUUU	829	AAAGGCAUGAAGCAGGAACAUUU	830
D-1333	[INVAB] GUUCCUGCUUCAUGCCUUUU	831	AAAGGCAUGAAGCAGGAACUU	832
D-1334	AUGUCCUGCUUCAUGCCUU{ INVAB }	833	AAAGGCAUGAAGCAGGAACAUUU	834
D-1335	[INVAB] AUGUCCUGCUUCAUGCCUUU	835	AAAGGCAUGAAGCAGGAACAUUU	836
D-1336	[INVAB] GUUCCUGCUUCAUGCCUUUU	837	AAAGGCAUGAAGCAGGAACUU	838
D-1337	AUGUCCUGCUUCAUGCCUU{ INVAB }	839	AAAGGCAUGAAGCAGGAACAUUU	840
D-1338	[INVAB] AUGUCCUGCUUCAUGCCUUU	841	AAAGGCAUGAAGCAGGAACAUUU	842
D-1339	[INVAB] AUGUCCUGCUUCAUGCCUUU	843	AAAGGCAUGAAGCAGGAACAUUU	844
D-1340	[INVAB] AUGUCCUGCUUCAUGCCUUU	845	AAAGGCAUGAAGCAGGAACAUUU	846
D-1341	[INVAB] GCUUCAUGCCUUCUACAGUA	847	UACUGUAGAAAGGCAUGAAGCUU	848
D-1342	[INVAB] GCUUCAUGCCUUCUACAGUA	849	UACUGUAGAAAGGCAUGAAGCUU	850
D-1343	[INVAB] UUCAUGCCUUCUACAGUAUU	851	UACUGUAGAAAGGCAUGAAUU	852
D-1344	[INVAB] GCUUCAUGCCUUCUACAGUA	853	UACUGUAGAAAGGCAUGAAGCUU	854
D-1345	GCUUCAUGCCUUCUACAGU{ INVAB }	855	UACUGUAGAAAGGCAUGAAGCUU	856
D-1346	[INVAB] GCGGCUUCCUGGGCUUCUAUU	857	UAGAAGCCCAGGAAGCCGCUU	858
D-1347	[INVAB] AUGGCUUCCAGAUUAUGCCUU	859	AGGCAUAUCUGGAAGCCAUUU	860
D-1348	[INVAB] ACAUGGCUUCCAGAUUAUGCCU	861	AGGCAUAUCUGGAAGCCAUUUU	862
D-1349	[INVAB] ACAUGGCUUCCAGAUUAUGCCU	863	AGGCAUAUCUGGAAGCCAUUUU	864
D-1350	[INVAB] CAACGUACCCUUCAUUGAUGU	865	ACAUCAAUGAAGGGUACGUUGUU	866
D-1351	[INVAB] CAACGUACCCUUCAUUGAUGU	867	ACAUCAAUGAAGGGUACGUUGUU	868

D-1352	[INVAB] ACGUACCCUUCAUUGAUGUUU	869	ACAUCAAUGAAGGGUACGUUU	870
D-1353	CUGCUUCAUGCCUUUCUACA{ INVAB }	871	AUGUAGAAAGGCAUGAAGCAGUU	872
D-1354	[INVAB] CUGCUUCAUGCCUUUCUACAU	873	AUGUAGAAAGGCAUGAAGCAGUU	874
D-1355	[INVAB] GCUUCAUGCCUUUCUACAUUU	875	AUGUAGAAAGGCAUGAAGCUU	876
D-1356	CUGCUUCAUGCCUUUCUACA{ INVAB }	877	AUGUAGAAAGGCAUGAAGCAGUU	878
D-1357	[INVAB] CUGCUUCAUGCCUUUCUACAU	879	AUGUAGAAAGGCAUGAAGCAGUU	880
D-1358	CUGCUUCAUGCCUUUCUACA{ INVAB }	881	AUGUAGAAAGGCAUGAAGCAGUU	882
D-1359	[INVAB] CUGCUUCAUGCCUUUCUACA{ INVA B }	883	AUGUAGAAAGGCAUGAAGCAGUU	884
D-1360	[INVAB] GCUUCAUGCCUUUCUACAUUU	885	AUGUAGAAAGGCAUGAAGCUU	886
D-1361	[INVAB] CUUCAUGCCUUUCUACAGUUU	887	ACUGUAGAAAGGCAUGAAGUU	888
D-1362	UGCUCUUGCCUUUCUACAG{ INVAB }	889	ACUGUAGAAAGGCAUGAAGCAUU	890
D-1363	[INVAB] CUGCUUCAUGCCUUUCUACAU	891	AUGUAGAAAGGCAUGAAGCAGUU	892
D-1364	CUGCUUCAUGCCUUUCUACA{ INVAB }	893	AUGUAGAAAGGCAUGAAGCAGUU	894
D-1365	CUGCUUCAUGCCUUUCUACA{ INVAB }	895	AUGUAGAAAGGCAUGAAGCAGUU	896
D-1366	[INVAB] CUUCAUCCCUUUCUACAGUUU	897	ACUGUAGAAAGGGAUGAAGUU	898
D-1367	[INVAB] GCUUCAUCCCUUUCUACAUUU	899	AUGUAGAAAGGGAUGAAGCUU	900
D-1368	UGCUCUAUCCCUUUCUACAG{ INVAB }	901	ACUGUAGAAAGGGAUGAAGCAUU	902
D-1369	CUGCUUCAUCCCUUUCUACA{ INVAB }	903	AUGUAGAAAGGGAUGAAGCAGUU	904
D-1370	[INVAB] ACAUUGCUCUUUCACCUGAUU	905	UCAGGUGAAAGAGCAAUGUUU	906
D-1371	CUGCUUCAUGCCUUUCUACA{ INVAB }	907	AUGUAGAAAGGCAUGAAGCAGUU	908
D-1372	CUGCUUCAUGCCUUUCUACA{ INVAB }	909	AUGUAGAAAGGCAUGAAGCAGUU	910
D-1373	CUGCUUCAUGCCUUUCUACA{ INVAB }	911	AUGUAGAAAGGCAUGAAGCAGUU	912
D-1374	CUGCUUCAUGCCUUUCUACA{ INVAB }	913	AUGUAGAAAGGCAUGAAGCAGUU	914
D-1375	CUGCUUCAUGCCUUUCUACA{ INVAB }	915	AUGUAGAAAGGCAUGAAGCAGUU	916
D-1376	CUGCUUCAUGCCUUUCUACA{ INVAB }	917	AUGUAGAAAGGCAUGAAGCAGUU	918

D-1377	CUGCUUCAUGCCUUUCUACA{ INVAB }	919	AUGUAGAAAGGCAUGAAGCAGUU	920
D-1378	CUGCUUCAUGCCUUUCUACA{ INVAB }	921	AUGUAGAAAGGCAUGAAGCAGUU	922
D-1379	CUGCUUCAUGCCUUUCUACA{ INVAB }	923	AUGUAGAAAGGCAUGAAGCAGUU	924
D-1380	CUGCUUCAUGCCUUUCUACA{ INVAB }	925	AUGUAGAAAGGCAUGAAGCAGUU	926
D-1381	CUGCUUCAUGCCUUUCUACA{ INVAB }	927	AUGUAGAAAGGCAUGAAGCAGUU	928
D-1381	[INVAB] CUUCAUGCC [DT] UUCUACAGUUU	929	ACUGUAGAAAGGCAUGAAGUU	930
D-1382	[INVAB] CUUCAUGCCUUUCUACAGUUU	931	ACUGUAGAAAGGCAUGAAGUU	932
D-1383	[INVAB] CUUCAUGCCUUUCUACAGUUU	933	ACUGUAGAAAGGCAUGAAGUU	934
D-1384	[INVAB] CUUCAUGCCUUUCUACAGUUU	935	ACUGUAGAAAGGCAUGAAGUU	936
D-1385	[INVAB] CUUCAUGCCUUUCUACAGUUU	937	ACUGUAGAAAGGCAUGAAGUU	938
D-1386	[INVAB] CUUCAUGC [DC] UUCUACAGUUU	939	ACUGUAGAAAGGCAUGAAGUU	940
D-1387	[INVAB] CUUCAUGCCU [DT] UCUACAGUUU	941	ACUGUAGAAAGGCAUGAAGUU	942
D-1388	[INVAB] GCUUCAUGGGAUUCUACAUUU	943	AUGUAGAAUCCCAUGAAGCUU	944
D-1389	[INVAB] CUUCAUGCGAAUCUACAGUUU	945	ACUGUAGAUUCGCAUGAAGUU	946
D-1390	CUGCUUCAUGCCUUUCUACA{ INVAB }	947	UUGUAGAAAGGCAUGAAGCAGUU	948
D-1391	[INVAB] CUGCUUCAUGCCUUUCUACAA	949	UUGUAGAAAGGCAUGAAGCAGUU	950
D-1392	CUGCUUCAUGCCUUUCUACA{ INVDA }	951	UUGUAGAAAGGCAUGAAGCAGUU	952
D-1393	CUGCUUCAUGGGAUUCUACA{ INVAB }	953	AUGUAGAAUCCCAUGAAGCAGUU	954
D-1394	[INVAB] CUGCUUCAUGGGAUUCUACAU	955	AUGUAGAAUCCCAUGAAGCAGUU	956
D-1395	[INVAB] CUUCAUGCCUUUCUACAGUUU	957	ACUGUAGAAAGGCAUGAAGUU	958
D-1396	[INVAB] CUUCAUGCCUUUCUACAGUUU	959	ACUGUAGAAAGGCAUGAAGUU	960
D-1397	[INVAB] GCUUCAUGCCUUUCUACAUUU	961	AUGUAGAAAGGCAUGAAGCUU	962
D-1398	[INVAB] UGCUUCAUGCCUUUCUACAG{ INVAB }	963	ACUGUAGAAAGGCAUGAAGCAUU	964
D-1399	[INVAB] CUUCAUGCCUUUCUACAGUUU	965	ACUGUAGAAAGGCAUGAAGUU	966
D-1400	UGCUUCAUGCCUUUCUACAG{ INVAB }	967	ACUGUAGAAAGGCAUGAAGCAUU	968

D-1401	UGC UU CAUGCCUUUCUACAG{ INVAB }	969	ACUGUAGAAAGGCAUGAAGCAUU	970
D-1402	UGC UU CAUGCCUUUCUACAG{ INVAB }	971	ACUGUAGAAAGGCAUGAAGCAUU	972
D-1403	[INVAB] CUUCAUGCCUUUCUACAGUUU	973	ACUGUAGAAAGGCAUGAAGUU	974
D-1404	UGC UU CAUGCCUUUCUACAG{ INVAB }	975	ACUGUAGAAAGGCAUGAAGCAUU	976
D-1405	AUGCCUUUCUACAGUGGCUU{ INVAB }	977	AGCCACUGUAGAAAGGCAUGAUU	978
D-1406	UCAUGCCUUUCUACAGUGGC{ INVAB }	979	AGCCACUGUAGAAAGGCAUGAUU	980
D-1407	[INVAB] AUGCCUUUCUACAGUGGCUUU	981	AGCCACUGUAGAAAGGCAUUU	982
D-1408	[INVAB] AUGCCUUUCUACAGUGGCUUU	983	AGCCACUGUAGAAAGGCAUUU	984
D-1409	[INVAB] UCAUGCCUUUCUACAGUGGCU	985	AGCCACUGUAGAAAGGCAUGAUU	986
D-1410	UCAUGCCUUUCUACAGUGGC{ INVAB }	987	AGCCACUGUAGAAAGGCAUGAUU	988
D-1411	[INVAB] AUGCCUUUCUACAGUGGCUUU	989	AGCCACUGUAGAAAGGCAUUU	990
D-1412	UCAUGCCUUUCUACAGUGGC{ INVAB }	991	AGCCACUGUAGAAAGGCAUGAUU	992
D-1413	UCAUGCCUUUCUACAGUGGC{ INVAB }	993	AGCCACUGUAGAAAGGCAUGAUU	994
D-1414	[INVAB] UCAUGCCUUUCUACAGUGGC{ INVAB }	995	AGCCACUGUAGAAAGGCAUGAUU	996
D-1415	[INVAB] AUGCCUUUCUACAGUGGCAUU	997	UGCCACUGUAGAAAGGCAUUU	998
D-1416	UCAUGCCUUUCUACAGUGGC{ INVAB }	999	UGCCACUGUAGAAAGGCAUGAUU	1000
D-1417	[INVAB] UCAUGCCUUUCUACAGUGGCA	1001	UGCCACUGUAGAAAGGCAUGAUU	1002
D-1418	UCAUGCCUUUCUACAGUGGC{ INVDA }	1003	UGCCACUGUAGAAAGGCAUGAUU	1004
D-1419	CUGCUUCAUGCCUUUCUACA{ INVAB }	1005	AUGUAGAAAGGCAUGAAGCAGUU	1006
D-1420	CUGCUUCAUGCCUUUCUACA{ INVAB }	1007	UUGUAGAAAGGCAUGAAGCAGUU	1008
D-1421	CUGCUUCAUGCCUUUCUACA{ INVDA }	1009	UUGUAGAAAGGCAUGAAGCAGUU	1010
D-1422	UCAUGCCUUUCUACAGUGGC{ INVAB }	1011	UGCCACUGUAGAAAGGCAUGAUU	1012
D-1423	UCAUGCCUUUCUACAGUGGC{ INVDA }	1013	UGCCACUGUAGAAAGGCAUGAUU	1014
D-1424	[INVAB] CUUCAUGCCUUUCUACAGUUU	1015	ACUGUAGAAAGGCAUGAAGUU	1016
D-1425	CUGCUUCAUGCCUUUCUACA{ INVAB }	1017	AUGUA [AB] AAAGGCAUGAAGCAGUU	1018

D-1426	CUGCUUCAUGCCUUUCUACA{ INVAB }	1019	AUGUA [AB] AAAGGCAUGAAGCAGUU	1020
D-1427	UGCUCUUGCCUUUCUACAG{ INVAB }	1021	ACUGU [AB] GAAAGGCAUGAAGCAUU	1022
D-1428	UGCUCUUGCCUUUCUACAG{ INVAB }	1023	ACUGU [AB] GAAAGGCAUGAAGCAUU	1024
D-1429	UCAUGCCUUUCUACAGUGGC{ INVAB }	1025	AGCCA [AB] UGUAGAAAGGCAUGAUU	1026
D-1430	UCAUGCCUUUCUACAGUGGC{ INVAB }	1027	AGCCA [AB] UGUAGAAAGGCAUGAUU	1028
D-1431	CUGCUUCAUGCCUUUCUACA{ INVAB }	1029	AU [GNA-G] UAGAAAGGCAUGAAGCAGUU	1030
D-1432	CUGCUUCAUGCCUUUCUACA{ INVAB }	1031	AUG [GNA-U] AGAAAGGCAUGAAGCAGUU	1032
D-1433	CUGCUUCAUGCCUUUCUACA{ INVAB }	1033	AUGU [GNA-A] GAAAGGCAUGAAGCAGUU	1034
D-1434	CUGCUUCAUGCCUUUCUACA{ INVAB }	1035	AUGUA [GNA-G] AAAGGCAUGAAGCAGUU	1036
D-1435	CUGCUUCAUGCCUUUCUACA{ INVAB }	1037	AUGUAG [GNA-A] AAGGCAUGAAGCAGUU	1038
D-1436	CUGCUUCAUGCCUUUCUACA{ INVAB }	1039	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1040
D-1437	CUGCUUCAUGCCUUUCUACA{ INVAB }	1041	AUGUAGAAAGGCAUGAAGCAGUU	1042
D-1438	[INVAB] CUGCUUCAUGCCUUUCUACAU	1043	AUGUAGAAAGGCAUGAAGCAGUU	1044
D-1439	[INVAB] CUGCUUCAUGCCUUUCUACA{ INVAB }	1045	AUGUAGAAAGGCAUGAAGCAGUU	1046
D-1440	CUGCUUCAUGCCUUUCUACA{ INVAB }	1047	AUGUAGAAAGGCAUGAAGCAGUU	1048
D-1441	CUGCUUCAUGCCUUUCUACA{ INVAB }	1049	AUGUAGAAAGGCAUGAAGCAGUU	1050
D-1442	[INVAB] CUGCUUCAUGCCUUUCUACAU	1051	AUGUAGAAAGGCAUGAAGCAGUU	1052
D-1443	CUGCUUCAUGCCUUUCUACA{ INVAB }	1053	AUGUAGAAAGGCAUGAAGCAGUU	1054
D-1444	CUGCUUCAUGCCUUUCUACA{ INVAB }	1055	AUGUAGAAAGGCAUGAAGCAGUU	1056
D-1445	[INVAB] CUGCUUCAUGCCUUUCUACAU	1057	AUGUAGAAAGGCAUGAAGCAGUU	1058
D-1446	[INVAB] CUGCUUCAUGCCUUUCUACA{ INVAB }	1059	AUGUAGAAAGGCAUGAAGCAGUU	1060
D-1447	CUGCUUCAUGCCUUUCUACA{ INVAB }	1061	A [AB] GUAGAAAGGCAUGAAGCAGUU	1062
D-1448	CUGCUUCAUGCCUUUCUACA{ INVAB }	1063	AU [AB] UAGAAAGGCAUGAAGCAGUU	1064
D-1449	CUGCUUCAUGCCUUUCUACA{ INVAB }	1065	AUG [AB] AGAAAGGCAUGAAGCAGUU	1066
D-1450	CUGCUUCAUGCCUUUCUACA{ INVAB }	1067	AUGU [AB] GAAAGGCAUGAAGCAGUU	1068

D-1451	CUGCUUCAUGCCUUUCUACA{ INVAB }	1069	AUGUAG [AB] AAGGCAUGAAGCAGUU	1070
D-1452	CUGCUUCAUGCCUUUCUACA{ INVAB }	1071	AUGUAGA [AB] AGGCAUGAAGCAGUU	1072
D-1453	CAACGUACCCUUCUAUGAUG{ INVAB }	1073	ACAUCAAUGAAGGGUACGUUGUU	1074
D-1454	CAACGUACCCUUCUAUGAUG{ INVAB }	1075	ACAUCAAUGAAGGGUACGUUGUU	1076
D-1455	ACAUGGCUUCCAGAUUGCC{ INVAB }	1077	AGGCAUAUCUGGAAGCCAUGUUU	1078
D-1456	ACAUGGCUUCCAGAUUGCC{ INVAB }	1079	AGGCAUAUCUGGAAGCCAUGUUU	1080
D-1457	CUGCUUCAUGCCUUUCUACA{ INVAB }	1081	AUGUAGAAAGGCAUGAAGCAGUU	1082
D-1458	CUGCUUCAUGCCUUUCUACA{ INVAB }	1083	AUGUAGAAAGGCAUGAAGCAGUU	1084
D-1459	CUGCUUCAUGCCUUUCUACA{ INVAB }	1085	AUGUAGAAAGGCAUGAAGCAGUU	1086
D-1460	CUGCGGCUUCCUGGGCUUCU{ INVAB }	1087	UAGAAGCCCAGGAAGCCGCAGUU	1088
D-1461	CUGCGGCUUCCUGGGCUUCU{ INVAB }	1089	UAGAAGCCCAGGAAGCCGCAGUU	1090
D-1462	UGCUCUUGCCUUUCUACAG{ INVAB }	1091	ACUGUAGAAAGGCAUGAAGCAUU	1092
D-1463	[INVAB] UGCUCUUGCCUUUCUACAGU	1093	ACUGUAGAAAGGCAUGAAGCAUU	1094
D-1464	[INVAB] UGCUCUUGCCUUUCUACAG{ INVA B }	1095	ACUGUAGAAAGGCAUGAAGCAUU	1096
D-1465	UGCUCUUGCCUUUCUACAG{ INVAB }	1097	ACUGUAGAAAGGCAUGAAGCAUU	1098
D-1466	UGCUCUUGCCUUUCUACAG{ INVAB }	1099	ACUGUAGAAAGGCAUGAAGCAUU	1100
D-1467	UGCUCUUGCCUUUCUACAG{ INVAB }	1101	ACUGUAGAAAGGCAUGAAGCAUU	1102
D-1468	CUGCUUCAUGCCUUUCUACAU	1103	AUGUAGAAAGGCAUGAAGCAGUU	1104
D-1469	CUGCUUCAUGCCUUUCUACAU	1105	AUGUAGAAAGGCAUGAAGCAGUU	1106
D-1470	CUGCUUCAUGCCUUUCUACAU	1107	AUGUAGAAAGGCAUGAAGCAGUU	1108
D-1471	UGCUCUUGCCUUUCUACAG{ INVAB }	1109	UCUGUAGAAAGGCAUGAAGCAUU	1110
D-1472	UGCUCUUGCCUUUCUACAG{ INVDA }	1111	UCUGUAGAAAGGCAUGAAGCAUU	1112
D-1473	CUGCUUCAUGCCUUUCUACA{ INVDT }	1113	AUGUAGAAAGGCAUGAAGCAGUU	1114
D-1474	CUGCUUCAUGCCUUUCUACA{ INVAB }	1115	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1116
D-1475	CUGCUUCAUGCCUUUCUACA{ INVAB }	1117	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1118

D-1476	CUGCUUCAUGCCUUUCUACA{ INVAB }	1119	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1120
D-1477	CUGCUUCAUGCCUUUCUACA{ INVAB }	1121	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1122
D-1478	CUGCUUCAUGCCUUUCUACA{ INVAB }	1123	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1124
D-1479	CUGCUUCAUGCCUUUCUACA{ INVAB }	1125	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1126
D-1480	CUGCUUCAUGCCUUUCUACA{ INVAB }	1127	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1128
D-1481	CUGCUUCAUGCCUUUCUACA{ INVAB }	1129	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1130
D-1482	CUGCUUCAUGCCUUUCUACA{ INVAB }	1131	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1132
D-1483	CUGCUUCAUGCCUUUCUACA{ INVAB }	1133	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1134
D-1484	CUGCUUCAUGCCUUUCUACA{ INVAB }	1135	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1136
D-1485	CUGCUUCAUGCCUUUCUACA{ INVAB }	1137	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1138
D-1486	CUGCUUCAUGCCUUUCUACA{ INVAB }	1139	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1140
D-1487	CUGCUUCAUGCCUUUCUACA{ INVAB }	1141	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1142
D-1488	CUGCUUCAUGCCUUUCUACA{ INVAB }	1143	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1144
D-1489	CUGCUUCAUGCCUUUCUACA{ INVAB }	1145	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1146
D-1490	CUGCUUCAUGCCUUUCUACA{ INVAB }	1147	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1148
D-1491	CUGCUUCAUGCCUUUCUACA{ INVAB }	1149	AUGUAGA [GNA-A] A [DG] GCAUGAAGCAGUU	1150
D-1492	CUGCUUCAUGCCUUUCUACA{ INVAB }	1151	AUGUAGA [GNA-A] [DA] GGCAUGAAGCAGUU	1152
D-1493	CUGCUUCAUGCCUUUCUACA{ INVAB }	1153	AUGUAGA [GNA-A] AG [DG] CAUGAAGCAGUU	1154
D-1494	CUGCUUCAUGCCUUUCUACA{ INVAB }	1155	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1156
D-1495	CUGCUUCAUGCCUUUCUACA{ INVAB }	1157	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1158
D-1496	CUGCUUCAUGCCUUUCUACA{ INVAB }	1159	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1160
D-1497	CUGCUUCAUGCCUUUCUACA{ INVAB }	1161	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1162
D-1498	CUGCUUCAUGCCUUUCUACA{ INVAB }	1163	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1164
D-1499	CUGCUUCAUGCCUUUCUACA{ INVAB }	1165	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1166
D-1500	CUGCUUCAUGCCUUUCUACA{ INVAB }	1167	AUGUAGAAAGGCAUGAAGCAGUU	1168
D-1501	UGCUUCAUGCCUUUCUACAG{ INVDA }	1169	U [GNA-C] UGUAGAAAGGCAUGAAGCAUU	1170

D-1502	UGCUCUAGCCUUUCUACAG{ INVDA }	1171	UC [GNA-U] GUAGAAAGGCAUGAAGCAUU	1172
D-1503	UGCUCUAGCCUUUCUACAG{ INVDA }	1173	UCUG [GNA-U] AGAAAGGCAUGAAGCAUU	1174
D-1504	UGCUCUAGCCUUUCUACAG{ INVDA }	1175	UCUGU [GNA-A] GAAAGGCAUGAAGCAUU	1176
D-1505	UGCUCUAGCCUUUCUACAG{ INVDA }	1177	UCUGUAG [GNA-A] AAGGCAUGAAGCAUU	1178
D-1506	UGCUCUAGCCUUUCUACAG{ INVDA }	1179	U [AB] UGUAGAAAGGCAUGAAGCAUU	1180
D-1507	UGCUCUAGCCUUUCUACAG{ INVDA }	1181	UC [AB] GUAGAAAGGCAUGAAGCAUU	1182
D-1508	UGCUCUAGCCUUUCUACAG{ INVDA }	1183	UCU [AB] UAGAAAGGCAUGAAGCAUU	1184
D-1509	UGCUCUAGCCUUUCUACAG{ INVDA }	1185	UCUG [AB] AGAAAGGCAUGAAGCAUU	1186
D-1510	UGCUCUAGCCUUUCUACAG{ INVDA }	1187	UCUGU [AB] GAAAGGCAUGAAGCAUU	1188
D-1511	UGCUCUAGCCUUUCUACAG{ INVDA }	1189	UCUGUA [AB] AAAGGCAUGAAGCAUU	1190
D-1512	UGCUCUAGCCUUUCUACAG{ INVDA }	1191	UCUGUAG [AB] AAGGCAUGAAGCAUU	1192
D-1513	UCAUGCCUUUCUACAGUGGC{ INVDT }	1193	AGCCACUGUAGAAAGGCAUGAUU	1194
D-1514	UCAUGCCUUUCUACAGUGGC{ INVAB }	1195	UGCCACUGUAGAAAGGCAUGAUU	1196
D-1515	UCAUGCCUUUCUACAGUGGC{ INVDA }	1197	UGCCACUGUAGAAAGGCAUGAUU	1198
D-1516	UCCUGCUUAGCCUUUCUA{ INVDT }	1199	AUAGAAAGGCAUGAAGCAGGAUU	1200
D-1517	UCCUGCUUAGCCUUUCUA{ INVAB }	1201	UUAGAAAGGCAUGAAGCAGGAUU	1202
D-1518	UCCUGCUUAGCCUUUCUA{ INVDA }	1203	UUAGAAAGGCAUGAAGCAGGAUU	1204
D-1519	UAUGUCCUGCUUAGCCU{ INVDT }	1205	AAGGCAUGAAGCAGGAACAUUU	1206
D-1520	UAUGUCCUGCUUAGCCU{ INVAB }	1207	UAGGCAUGAAGCAGGAACAUUU	1208
D-1521	UAUGUCCUGCUUAGCCU{ INVDA }	1209	UAGGCAUGAAGCAGGAACAUUU	1210
D-1522	UCCUGCUUAGCCUUUCUA{ INVAB }	1211	AUAGAAAGGCAUGAAGCAGGAUU	1212
D-1523	UAUGUCCUGCUUAGCCU{ INVAB }	1213	AAGGCAUGAAGCAGGAACAUUU	1214
D-1524	UCAUGCCUUUCUACAGUGGC{ INVDT }	1215	AGCCACUGUAGAAAGGCAUGAUU	1216
D-1525	UCCUGCUUAGCCUUUCUA{ INVDT }	1217	AUAGAAAGGCAUGAAGCAGGAUU	1218
D-1526	UCCUGCUUAGCCUUUCUA{ INVDA }	1219	UUAGAAAGGCAUGAAGCAGGAUU	1220
D-1527	UAUGUCCUGCUUAGCCU{ INVDA }	1221	UAGGCAUGAAGCAGGAACAUUU	1222

D-1528	UGCUCUCAUGCCUUUCUACAG{ INVDA }	1223	UCUGUA [GNA-G] AAAGGCAUGAAGCAUU	1224
D-1529	CUGCUUCAUGCCUUUCUACA{ INVAB }	1225	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1226
D-1530	CUGCUUCAUGCCUUUCUACA{ INVAB }	1227	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1228
D-1531	CUGCUUCAUGCCUUUCUACA{ INVAB }	1229	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1230
D-1532	CUGCUUCAUGCCUUUCUACA{ INVAB }	1231	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1232
D-1533	CUGCUUCAUGCCUUUCUACA{ INVAB }	1233	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1234
D-1534	CUGCUUCAUGCCUUUCUACA{ INVAB }	1235	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1236
D-1535	CUGCUUCAUGCCUUUCUACA{ INVAB }	1237	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1238
D-1536	CUGCUUCAUGCCUUUCUACA{ INVAB }	1239	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1240
D-1537	CUGCUUCAUGCCUUUCUACA{ INVAB }	1241	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1242
D-1538	CUGCUUCAUGCCU [LNA-T] UCUACA{ INVAB }	1243	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1244
D-1539	CUGCUUCAUGCC [LNA-T] UUCUACA{ INVAB }	1245	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1246
D-1540	CUGCUUCAUGCCUU [LNA-T] CUACA{ INVAB }	1247	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1248
D-1541	CUGCUUCAUGCCUUUC [LNA-T] ACA{ INVAB }	1249	AUGU [GNA-A] GAAAGGCAUGAAGCAGUU	1250
D-1542	CUGCUUCAUGCCUUUCU [LNA-A] CA{ INVAB }	1251	AUG [GNA-U] AGAAAGGCAUGAAGCAGUU	1252
D-1543	CUGCUUCAUGCCUUUCUAC [LNA-A] { INVAB }	1253	A [GNA-U] GUAGAAAGGCAUGAAGCAGUU	1254
D-1544	CUGCUUCAUGCCUU [LNA-T] CUACA{ INVAB }	1255	AUGUAG [AB] AAGGCAUGAAGCAGUU	1256
D-1545	CUGCUUCAUGCCUUUC [LNA-T] ACA{ INVAB }	1257	AUGU [AB] GAAAGGCAUGAAGCAGUU	1258
D-1546	CUGCUUCAUGCCUUUCU [LNA-A] CA{ INVAB }	1259	AUG [AB] AGAAAGGCAUGAAGCAGUU	1260
D-1547	CUGCUUCAUGCCUUUCUACA{ INVAB }	1261	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1262
D-1548	CUGCUUCAUGCCUUUCUACA{ INVAB }	1263	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1264

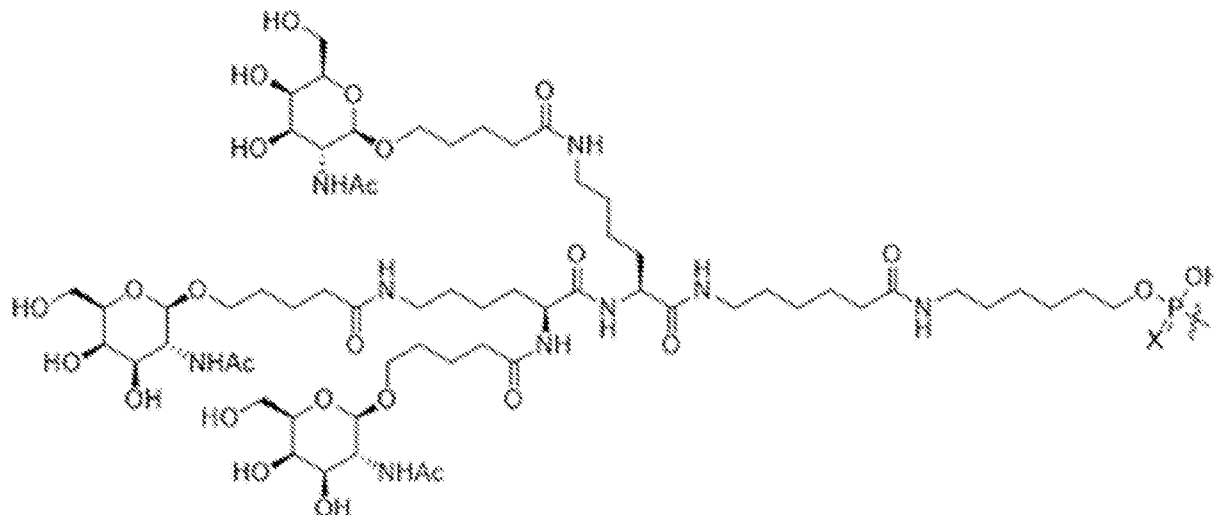
D-1549	CUGCUUCAUGCCU [LNA-T] UCUACA { INVAB }	1265	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1266
D-1550	CUGCUUCAUGCCU [LNA-T] UCUACA { INVAB }	1267	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1268
D-1551	CUGCUUCAUGCCU [LNA-T] UCUACA { INVAB }	1269	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1270
D-1552	CUGCUUCAUGCCU [LNA-T] UCUACA { INVAB }	1271	AUGUAGA [GNA-A] AGGCAUGAAGCAGUU	1272
D-1553	UGCUCUUGCCUUUCU [LNA-A] CAG { INVDA }	1273	UCUG [AB] AGAAAGGCAUGAAGCAUU	1274
D-1554	UGCUCUUGCCUUUC [LNA-T] ACAG { INVDA }	1275	UCUGU [AB] GAAAGGCAUGAAGCAUU	1276
D-1555	UGCUCUUGCCUU [LNA-T] CUACAG { INVDA }	1277	UCUGUAG [AB] AAGGCAUGAAGCAUU	1278
D-1556	UGCUCUUGCCUUUCUAC [LNA-A] G { INVDA }	1279	UC [GNA-U] GUAGAAAGGCAUGAAGCAUU	1280
D-1557	UGCUCUUGCCUUUCU [LNA-A] CAG { INVDA }	1281	UCUG [GNA-U] AGAAAGGCAUGAAGCAUU	1282
D-1558	UGCUCUUGCCUUUC [LNA-T] ACAG { INVDA }	1283	UCUGU [GNA-A] GAAAGGCAUGAAGCAUU	1284
D-1559	UGCUCUUGCCUUUCUACA [LNA-G] { INVDA }	1285	U [AB] UGUAGAAAGGCAUGAAGCAUU	1286
D-1560	UGCUCUUGCCUUUCUAC [LNA-A] G { INVDA }	1287	UC [AB] GUAGAAAGGCAUGAAGCAUU	1288
D-1561	UCAUGCCUUUCUACA [LNA-G] UGGC { INVAB }	1289	AGCCA [AB] UGUAGAAAGGCAUGAUU	1290
D-1562	UCAUGCCUUUCUACAG [LNA-T] GGC { INVAB }	1291	AGCCA [AB] UGUAGAAAGGCAUGAUU	1292
D-1563	UCAUGCCUUUCUACAGUGGC { INVAB }	1293	AGCCA [AB] UGUAGAAAGGCAUGAUU	1294
D-1564	GGUAUGUCCUGCUUCAUUUU	2257	AAUGAAGCAGGAACAUACCUU	2258
D-1565	GGUAUGUCCUGCUUCAUAUU	2259	UAUGAAGCAGGAACAUACCUU	2260
D-1566	GUAUGUCCUGCUUCAUGUUU	2261	ACAUGAAGCAGGAACAUACCUU	2262
D-1567	UAUGUCCUGCUUCAUGCAUU	2263	UGCAUGAAGCAGGAACAUUU	2264
D-1568	AUGUCCUGCUUCAUGCCUUU	2265	AGGCAUGAAGCAGGAACAUUU	2266

D-1569	UGUCCUGCUUCAUGCCUUUU	2267	AAGGCAUGAAGCAGGAACAUU	2268
D-1570	GUUCCUGCUUCAUGCCUUUUU	2269	AAAGGCAUGAAGCAGGAACUU	2270
D-1571	UUCCUGCUUCAUGCCUUUUUU	2271	AAAAGGCAUGAAGCAGGAUUU	2272
D-1572	UUCCUGCUUCAUGCCUUUAUU	2273	UAAAGGCAUGAAGCAGGAUUU	2274
D-1573	UCCUGCUUCAUGCCUUUCUUU	2275	AGAAAGGCAUGAAGCAGGAUU	2276
D-1574	CCUGCUUCAUGCCUUUCUAUU	2277	UAGAAAGGCAUGAAGCAGGUU	2278
D-1575	CUGCUUCAUGCCUUUCUAUUU	2279	AUAGAAAGGCAUGAAGCAGUU	2280
D-1576	CUGCUUCAUGCCUUUCUAAUU	2281	UUAGAAAGGCAUGAAGCAGUU	2282
D-1577	UGCUUCAUGCCUUUCUACAUU	2283	UGUAGAAAGGCAUGAAGCAUU	2284
D-1578	GCUUCAUGCCUUUCUACAUUU	2285	AUGUAGAAAGGCAUGAAGCUU	2286
D-1579	GCUUCAUGCCUUUCUACAAUU	2287	UUGUAGAAAGGCAUGAAGCUU	2288
D-1580	CUUCAUGCCUUUCUACAGUUU	2289	ACUGUAGAAAGGCAUGAAGUU	2290
D-1581	UUCAUGCCUUUCUACAGUUUU	2291	AACUGUAGAAAGGCAUGAAUU	2292
D-1582	UUCAUGCCUUUCUACAGUAUU	2293	UACUGUAGAAAGGCAUGAAUU	2294
D-1583	UCAUGCCUUUCUACAGUGUUU	2295	ACACUGUAGAAAGGCAUGAUU	2296
D-1584	UCAUGCCUUUCUACAGUGAUU	2297	UCACUGUAGAAAGGCAUGAUU	2298
D-1585	CAUGCCUUUCUACAGUGGUUU	2299	ACCACUGUAGAAAGGCAUGUU	2300
D-1586	CAUGCCUUUCUACAGUGGAUU	2301	UCCACUGUAGAAAGGCAUGUU	2302
D-1587	AUGCCUUUCUACAGUGGCUUU	2303	AGCCACUGUAGAAAGGCAUUU	2304
D-1588	AUGCCUUUCUACAGUGGCAUU	2305	UGCCACUGUAGAAAGGCAUUU	2306
D-1589	UGCCUUUCUACAGUGGCCUUU	2307	AGGCCACUGUAGAAAGGCAUU	2308
D-1590	GCCUUUCUACAGUGGCCUUUU	2309	AAGGCCACUGUAGAAAGGCUU	2310
D-1591	GGUAUGUUCUGCUUCAUCUU	2311	GAUGAAGCAGGAACAUACUU	2312
D-1592	GUUUGUUCUGCUUCAUCUUU	2313	GGAUGAAGCAGGAACAUACUU	2314
D-1593	UAUGUUCUGCUUCAUCCUUU	2315	GGGAUGAAGCAGGAACAUUUU	2316
D-1594	AUGUUCUGCUUCAUCCCUUU	2317	GGGGAUGAAGCAGGAACAUUU	2318

D-1595	UGUCCUGCUUCAUCCCCUUU	2319	AGGGGAUGAAGCAGGAACAUU	2320
D-1596	GUUCCUGCUUCAUCCCCUUU	2321	AAGGGGAUGAAGCAGGAACUU	2322
D-1597	UUCUGCUUCAUCCCCUUCUU	2323	GAAGGGGAUGAAGCAGGAAUU	2324
D-1598	UCCUGCUUCAUCCCCUUCUU	2325	AGAAGGGGAUGAAGCAGGAUU	2326
D-1599	CCUGCUUCAUCCCCUUCUAUU	2327	UAGAAGGGGAUGAAGCAGGUU	2328
D-1600	CUGCUUCAUCCCCUUCUACUU	2329	GUAGAAGGGGAUGAAGCAGUU	2330
D-1601	UGCUCUAUCCCCUUCUACAUU	2331	UGUAGAAGGGGAUGAAGCAUU	2332
D-1602	GCUUCAUCCCCUUCUACAGUU	2333	CUGUAGAAGGGGAUGAAGCUU	2334
D-1603	CUUCAUCCCCUUCUACAGUUU	2335	ACUGUAGAAGGGGAUGAAGUU	2336
D-1604	UUCAUCCCCUUCUACAGUGUU	2337	CACUGUAGAAGGGGAUGAAUU	2338
D-1605	UCAUCCCCUUCUACAGUGGUU	2339	CCACUGUAGAAGGGGAUGAUU	2340
D-1606	CAUCCCCUUCUACAGUGGCUU	2341	GCCACUGUAGAAGGGGAUGUU	2342
D-1607	AUCCCCUUCUACAGUGGCCUU	2343	GGCCACUGUAGAAGGGGAUUU	2344
D-1608	UCCCCUUCUACAGUGGCCUUU	2345	AGGCCACUGUAGAAGGGGAUU	2346
D-1609	CCCCUUCUACAGUGGCCUUU	2347	AAGGCCACUGUAGAAGGGGUU	2348

[0002] To improve the potency and in vivo stability of PNPLA3 siRNA sequences, chemical modifications were incorporated into PNPLA3 siRNA molecules. Specifically, 2'-O-methyl and 2'-fluoro modifications of the ribose sugar were incorporated at specific positions within the PNPLA3 siRNAs. Phosphorothioate internucleotide linkages were also incorporated at the terminal ends of the antisense and/ or sense sequences. Table 2 below depicts the modifications in the sense and antisense sequences for each of the modified PNPLA3 siRNAs. The nucleotide sequences in Table X are listed according to the following notations: A, U, G, and C = corresponding ribonucleotide; dT = deoxythymidine; a, u, g, and c = corresponding 2'-O-methyl ribonucleotide; Af, Uf, Gf, and Cf = corresponding 2'-deoxy-2'-fluoro ("2'-fluoro") ribonucleotide. Insertion of an "s" in the sequence indicates that the two adjacent nucleotides are connected by a phosphorothiodiester group (e.g. a phosphorothioate internucleotide linkage). Unless indicated otherwise, all other nucleotides are connected by 3'-5' phosphodiester groups. Each of the siRNA

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Duplex No.	Sense sequence (5'-3')	SEQ ID NO: (sense)	Antisense sequence (5'-3')	SEQ ID NO: (antisense)
D-2000	GfsgsGfcAfaUfaAfAfGfuAfcCfuGfcUf susUf	167	asGfscAfgGfuAfcuuUfaUfuGfcCfcsUfs u	168
D-2001	CfsgsGfcCfaAfuGfUfCfcAfcCfaGfcUf susUf	169	asGfscUfgGfuGfgacAfuUfgGfcCfcsUfs u	170
D-2002	GfsgsUfcCfaGfcCfUfGfaAfcUfuCfuUf susUf	171	asAfsGfaGfuUfcagGfcUfgGfaCfcsUfs u	172
D-2003	GfscsUfuCfaUfcCfCfCfuUfcUfaCfaGf susUf	173	csUfsgUfaGfaAfgggGfaUfgAfaGfcsUfs u	174
D-2004	GfscsGfgCfuUfcCfUfGfgGfcUfuCfuAf susUf	175	usAfsGfaGfcCfcagGfaAfgCfcGfcsUfs u	176
D-2005	GfscsCfuCfuGfaGfcCfUfgAfgUfuGfgUf susUf	177	asCfscAfaCfuCfagCufcAfgAfgGfcsUfs u	178
D-2006	GfsusGfaCfaAfcGfUfAfcCfcUfuCfaUf susUf	179	asUfsgAfaGfgGfuacGfuUfgUfcAfcUfs u	180
D-2007	CfscsCfGfcUfcCfaAfGfgUfcCfcAfaAf susUf	181	usUfsuGfgGfaCfcugGfaGfgCfGfgfsUfs u	182
D-2008	CfsusUfcAfuCfcCfCfUfuCfuAfcAfgUf susUf	183	asCfsuGfuAfgAfggGfgAfuGfaAfgsUfs u	184
D-2009	GfsgsUfaUfgUfuCfcCfUfgCfuUfcAfuGf susUf	185	csAfsuGfaAfgCfaggAfaCfaUfaCfcsUfs u	186
D-2010	GfsusAfuGfuUfcCfUfGfcUfuCfaUfgCf susUf	187	gsCfsaUfgAfaGfcagGfaAfcAfuAfcUfs u	188
D-2011	UfsasUfgUfuCfcUfGfcUfuCfuAfuGfcCf susUf	189	gsGfscAfuGfaAfgcaGfgAfaCfaUfasUfs u	190
D-2012	AfsusGfuUfcCfuGfcCfUfuCfaUfgCfcCf susUf	191	gsGfsgCfaUfgAfgagCfGfaAfcAfuUfs u	192
D-2013	UfsgsUfuCfcUfgCfUfUfcAfuGfcCfcUf susUf	193	asGfsgGfcAfuGfaagCfaGfgAfaCfasUfs u	194

D-2014	GfsusUfcCfuGfcUfUfCfaUfgCfcCfuUf susUf	195	asAfsGfgCfaUfgaaGfcAfgGfaAfcUfs u	196
D-2015	UfsusCfcUfgCfuUfCfAfugfcCfcUfuCf susUf	197	gsAfsaGfgGfcAfugaAfgCfaGfgAfasUfs u	198
D-2016	UfscsCfuGfcUfuCfAfUfgCfcCfuUfCf susUf	199	asGfsaAfgGfgCfaugAfaGfcAfgGfasUfs u	200
D-2017	CfscsUfgCfuUfCfAfUfgCfcCfuUfuAf susUf	201	usAfsGfaAfgGfgcaUgfaAfgCfaGfgsUfs u	202
D-2018	CfsusGfcUfuCfaUfGfcCfcCfuUfCfCf susUf	203	gsUfsaGfaAfgGfgcaUfgAfaGfcAfgsUfs u	204
D-2019	UfsgsCfuUfCfAfugfcCfcUfuCfuAfcAf susUf	205	usGfsuAfgAfaGfggcaUgfaAfgCfasUfs u	206
D-2020	GfscsUfuCfaUfgCfcCfuUfCfCfaGf susUf	207	csUfsgUfaGfaAfgggCfaUfgAfaGfcsUfs u	208
D-2021	CfsusUfCfAfugfcCfcUfuCfuAfcAfgUf susUf	209	asCfsuGfuAfgAfggGfcAfuGfaAfgsUfs u	210
D-2022	UfsusCfaUfgCfcCfUfUfCfCfaGfuGf susUf	211	csAfsCufgUfaGfaagGfgCfaUfgAfasUfs u	212
D-2023	UfscsAfugfcCfcUfUfCfuAfcAfgUfgGf susUf	213	csCfsaCfuGfuAfgaaGfgGfcAfuGfasUfs u	214
D-2024	CfsasUfgCfcCfuUfCfUfaCfaGfuGfgCf susUf	215	gsCfscAfcUfgUfagaAfgGfgCfaUfgsUfs u	216
D-2025	AfsusGfcCfcUfuCfUfAfcAfgUfgGfcCf susUf	217	gsGfscCfaCfuGfuagAfaGfgGfcAfuUfs u	218
D-2026	UfsgsCfcCfuUfCfUfAfcAfgUfgGfcCfCf susUf	219	asGfsgCfcAfcUfguaGfaAfgGfgCfasUfs u	220
D-2027	GfscsCfcUfuCfuAfcAfgUfgGfcCfuUf susUf	221	asAfsGfCfcCfaCfuguAfgAfaGfgGfcsUfs u	222
D-2028	GfsgsUfaUfgUfuCfCfUfgCfuUfCfuCf susUf	223	gsAfsuGfaAfgCfaggAfaCfaUfaCfcsUfs u	224
D-2029	GfsusAfugfuUfCfUfGfcUfuCfaUfCfCf susUf	225	gsGfsaUfgAfaGfcagGfaAfcAfuAfcUfs u	226
D-2030	UfsasUfgUfuCfCfUfGfcCfuUfCfCfcCf susUf	227	gsGfsgAfuGfaAfgcaGfgAfaCfaUfasUfs u	228
D-2031	AfsusGfuUfCfCfuGfcCfuUfCfCfcCfCf susUf	229	gsGfsgGfaUfgAfgagCfGfaAfcAfuUfs u	230
D-2032	UfsgsUfuCfCfUfgCfUfUfCfCfCfcCfCf susUf	231	asGfsgGfgAfuGfaagCfaGfgAfaCfasUfs u	232
D-2033	GfsusUfCfCfuGfcUfUfCfCfCfcCfuUf susUf	233	asAfsGfgGfaUfgaaGfcAfgGfaAfcUfs u	234
D-2034	UfsusCfcUfgCfuUfCfAfugfcCfcUfuCf susUf	235	gsAfsaGfgGfgAfugaAfgCfaGfgAfasUfs u	236
D-2035	UfscsCfuGfcUfuCfAfUfCfcCfcCfuUfCf susUf	237	asGfsaAfgGfgGfaugAfaGfcAfgGfasUfs u	238
D-2036	CfscsUfgCfuUfCfAfUfCfcCfcUfuCfuAf susUf	239	usAfsGfaAfgGfggaUgfaAfgCfaGfgsUfs u	240
D-2037	CfsusGfcUfuCfaUfCfCfcCfuUfCfCfCf susUf	241	gsUfsaGfaAfgGfggaUfgAfaGfcAfgsUfs u	242
D-2038	UfsgsCfuUfCfAfugfcCfcUfuCfuAfcAf susUf	243	usGfsuAfgAfaGfgggAfuGfaAfgCfasUfs u	244
D-2039	UfsusCfaUfCfCfcCfUfUfCfCfaGfuGf susUf	245	csAfsCufgUfaGfaagGfgGfaUfgAfasUfs u	246
D-2040	UfscsAfugfcCfcUfUfCfuAfcAfgUfgGf susUf	247	csCfsaCfuGfuAfgaaGfgGfgAfuGfasUfs u	248
D-2041	CfsasUfCfCfcCfuUfCfUfaCfaGfuGfgCf susUf	249	gsCfscAfcUfgUfagaAfgGfgGfaUfgsUfs u	250
D-2042	UfscsCfcCfuUfCfUfAfcAfgUfgGfcCfCf susUf	251	asGfsgCfcAfcUfguaGfaAfgGfgGfasUfs u	252
D-2043	GfsasUfCfAfgGfaCfCfGfgAfgCfcGfaUf susUf	253	asUfscGfgCfuCfgggUfCfuGfaUfcsUfs u	254
D-2044	UfsgsGfgCfuUfCfUfAfcCfcAfcGfuCfGf susUf	255	asCfsgAfcGfuGfguaGfaAfgCfcCfasUfs u	256
D-2045	GfsasGfcGfaGfcAfcGfcCfcCfGfCfaUf susUf	257	asUfsgCfGfgGfgGfcGfcUfCfGfcUfcsUfs u	258
D-2046	UfsgsCfaCfuGfcGfUfCfGfgGfcGfuCfcUf susUf	259	asGfsgAfcGfcCfGacGfcAfgUfgCfasUfs u	260

D-2047	UfsgsGfaGfcAfgAfCfUfcUfgCfaGfgUf susUf	261	asCfscUfgCfaGfaguCfuGfcUfcCfasUfs u	262
D-2048	UfsgsCfaGfgUfcCfUfcCfuCfaGfaUfcUf susUf	263	asGfSaUfcUfgAfgagGfaCfcUfgCfasUfs u	264
D-2049	CfscsCfGfgCfaAfUfGfUfcCfaUfcCfaUf susUf	265	asUfsgGfuGfgAfcauUfgGfcCfGfgfsUfs u	266
D-2050	UfsusCfuAfcAfgUfGfGfcCfuUfaUfcUf susUf	267	asGfSaUfaAfgGfccaCfuGfuAfgAfasUfs u	268
D-2051	UfscsUfaCfaGfuGfGfCfcUfuAfuCfcUf susUf	269	asGfsgAfuAfaGfgccAfcUfgUfaGfasUfs u	270
D-2052	CfsusUfcCfuUfcAfgAfgGfcGfuGfcUf susUf	271	asGfscAfcGfcCfucUgfaAfgGfaAfgsUfs u	272
D-2053	UfscsCfcUfuCfaGfAfgGfgCfGfGfGfAf susUf	273	usCfsgCfaCfGfcUfcUfgAfaGfgAfasUfs u	274
D-2054	GfscsGfuGfcGfaUfAfuUfgUfgGfaUfgUf susUf	275	asCfSaUfcCfaCfaUaUfcGfcAfcGfcsUfs u	276
D-2055	CfsgsUfgCfGfaUfAfuGfGfGfGfGfGfAf susUf	277	usCfscAfuCfcAfcUaUfuCfGfCfaCfGfsUfs u	278
D-2056	UfsgsGfaUfgGfaGfGfAfgUfgAfgUfgAf susUf	279	usCfSaCfuCfaCfuUfcCfaUfcCfasUfs u	280
D-2057	AfscsGfuAfcCfcUfUfcCfaUfuGfaUfgUf susUf	281	asCfSaUfcAfaUfgaaGfgGfuAfcGfusUfs u	282
D-2058	UfsgsGfaCfaUfcAfcCfCfaAfgCfuCfaUf susUf	283	asUfsgAfgCfuUfgguGfaUfgUfcCfasUfs u	284
D-2059	CfsasCfcUfgCfGfUfcUfcAfgCfaUfcUf susUf	285	asGfSaUfgCfuGfagaCfGfCfaGfgUfgsUfs u	286
D-2060	AfscsCfuGfcGfuCfUfcCfaGfcAfuCfcUf susUf	287	asGfsgAfuGfcUfgagAfcGfcAfgGfusUfs u	288
D-2061	CfscsAfgAfgAfcUfGfGfuGfaCfaUfgUf susUf	289	asCfSaUfgUfcAfcUaGfuCfuCfuGfgsUfs u	290
D-2062	AfsusGfgCfuUfcCfAfgGfaUfaUfgCfcUf susUf	291	asGfsgCfaUfaUfcugGfaAfgCfcAfasUfs u	292
D-2063	CfscsGfcCfuCfcAfgGfGfuCfcCfaAfaUf susUf	293	asUfsuUfgGfgAfcUaGfgAfgGfcGfgsUfs u	294
D-2064	UfsasCfcUfgCfuGfGfUfgCfuGfaGfgUf susUf	295	asCfscUfcAfgCfaccAfgCfaGfgUfasUfs u	296
D-2065	AfscsCfuGfcUfgGfUfgGfcUfgAfgGfgUf susUf	297	asCfscCfuCfaGfcacCfaGfcAfgGfusUfs u	298
D-2066	CfsusCfuCfcAfcCfUfUfuCfcCfaGfuUf susUf	299	asAfcUfgGfgAfaagGfuGfgAfgAfgsUfs u	300
D-2067	UfsusUfuUfcAfcCfUfAfaCfuAfaAfaUf susUf	301	asUfsuUfuAfgUfuagGfuGfaAfaAfasUfs u	302
D-2068	CfgGfcCfaAfuGfUfcCfcAfcCfaGfcUfsu sUf	303	asGfscUfgGfuGfgacAfuUfgGfcCfGfsUfs u	304
D-2069	GfgUfcCfaGfcCfUfgGfaAfcUfuCfuUfsu sUf	305	asAfgAfaGfuUfcagGfcUfgGfaCfcsUfs u	306
D-2070	GfcGfgCfuUfcCfUfgGfgGfcUfuCfuAfsu sUf	307	usAfgAfaGfcCfcagGfaAfgCfcGfcsUfs u	308
D-2071	GfuGfaCfaAfcGfUfAfcCfcUfuCfaUfsu sUf	309	asUfsgAfaGfgGfuacGfuUfgUfcAfcUfs u	310
D-2072	GfgUfaUfgUfuCfcUfgCfuUfcAfuGfsu sUf	311	csAfsuGfaAfgCfaggAfaCfaUfaCfcsUfs u	312
D-2073	GfuAfuGfuUfcCfUfgGfcUfuCfaUfgCfsu sUf	313	gsCfSaUfgAfaGfcagGfaAfcAfuAfcUfs u	314
D-2074	UfgUfuCfcUfgCfUfUfcAfuGfcCfcUfsu sUf	315	asGfsgGfcAfuGfaagCfaGfgAfaCfasUfs u	316
D-2075	GfuUfcCfuGfcUfUfcCfaUfgCfcCfuUfsu sUf	317	asAfgGfgCfaUfgaaGfcAfgGfaAfcUfs u	318
D-2076	CfcUfgCfuUfcAfuGfcCfcUfuCfuAfsu sUf	319	usAfgAfaGfgGfcUaGfaAfgCfaGfgsUfs u	320
D-2077	GfcUfuCfaUfgCfcCfuUfcUfaCfaGfsu sUf	321	csUfsgUfaGfaAfgggCfaUfgAfaGfcsUfs u	322
D-2078	CfuUfcAfuGfcCfcUfuCfuAfuAfgUfsu sUf	323	asCfSuGfuAfgAfgagGfcAfuGfaAfgsUfs u	324
D-2079	UfuCfaUfgCfcCfUfUfcUfaCfaGfuGfsu sUf	325	csAfcUfgUfaGfaagGfgCfaUfgAfasUfs u	326

D-2080	AfuGfgCfuUfcCfAfGfaUfaUfgCfcUfsu sUf	327	asGfsgCfaUfaUfcugGfaAfgCfcAfusUfs u	328
D-2081	AfuGfcCfcUfuCfUfAfCfAgUfgGfcCfsu sUf	329	gsGfscCfaCfuGfuagAfaGfgGfcAfusUfs u	330
D-2082	GfcUfuCfaUfgCfcCfCfuUfcUfaCfaUfsu sUf	331	asUfsgUfaGfaAfgggCfaUfgAfaGfcsUfs u	332
D-2083	{ Phosphate } GfsgsAfaAfgAfcUfGfUfu CfcAfaAfaAfsusUf	1295	{ Phosphate } usUfsuUfuGfgAfacaGfuCf uUfuCfcsUfsu	1296
D-2084	{ GalNac3K2AhxC6 } ggsuauGfuCfCfUf Gfcuucagsusu	1297	{ Phosphate } csAfsuGfaAfgGfcaggAfaCf auaccsusu	1298
D-2085	{ GalNac3K2AhxC6 } guauguUfcCfUfGfC fuucagcsusu	1299	{ Phosphate } gsCfsaUfgAfAfgcagGfaAf cauacsusu	1300
D-2086	{ GalNac3K2AhxC6 } uguuccUfgCfUfUfC faugcccsusu	1301	{ Phosphate } asGfsgGfcAfUfgaagCfaGf gaacasusu	1302
D-2087	{ GalNac3K2AhxC6 } gcuucaUfgCfCfCfU fucuacagsusu	1303	{ Phosphate } csUfsgUfaGfAfagggCfaUf gaagcsusu	1304
D-2088	{ GalNac3K2AhxC6 } cuucauGfcCfCfUfU fcuacagsusu	1305	{ Phosphate } asCfsuGfuAfGfaaggGfcAf ugaagsusu	1306
D-2089	{ GalNac3K2AhxC6 } gcgguUfcCfUfGfG fgcuucasusu	1307	{ Phosphate } usAfsuGfaGfCfcagGfaAf gccgcsusu	1308
D-2090	{ GalNac3K2AhxC6 } guuccuGfcUfUfCfA fugccuusu	1309	{ Phosphate } asAfsuGfgCfAfugaaGfcAf ggaacsusu	1310
D-2091	{ GalNac3K2AhxC6 } AfuGfgCfuUfcCfAf GfaUfaUfgCfcUfsusUf	1311	{ Phosphate } asGfsgCfaUfaUfcugGfaAf gCfcAfusUfsu	1312
D-2092	{ GalNac3K2AhxC6 } ccugcuUfcAfUfGfC fccuucasusu	1313	{ Phosphate } usAfsuGfaGfGfgcauGfaAf gcaggsusu	1314
D-2093	{ GalNac3K2AhxC6 } uucaugCfcCfUfUfC fuacaguusu	1315	{ Phosphate } asAfsuUfgUfAfagaagGfgCf augaasusu	1316
D-2094	{ GalNac3K2AhxC6 } uucaugCfcCfUfUfC fuacaggsusu	1317	{ Phosphate } csAfsuUfgUfAfagaagGfgCf augaasusu	1318
D-2095	{ GalNac3K2AhxC6 } gcuucaUfgCfCfCfU fucuacagsusu	1319	{ Phosphate } asUfsgUfaGfAfagggCfaUf gaagcsusu	1320
D-2096	{ GalNac3K2AhxC6 } gguccaGfcCfUfGfA facuucasusu	1321	{ Phosphate } asAfsuGfaGfUfucagGfcUf ggaccsusu	1322
D-2097	{ GalNac3K2AhxC6 } GfcGfgCfuUfcCfUf GfGfgCfuUfcCfuAfsusUf	1323	{ Phosphate } usAfsuGfaGfCfcagGfaAf gCfcGfcsUfsu	1324
D-2098	{ GalNac3K2AhxC6 } GfcGfgCfuUfcCfuG fGfgCfuUfcCfuAfsusUf	1325	{ Phosphate } usAfsuGfaGfCfcagGfaAf fgCfcGfcsUfsu	1326
D-2099	{ GalNac3K2AhxC6 } GfuUfcCfuGfCfUfUf CfAfugCfcCfuUfsusUf	1327	{ Phosphate } asAfsuGfgCfAfugaaGfcAf gGfaAfcUfsu	1328
D-2100	{ GalNac3K2AhxC6 } GfuUfcCfuGfCfUfUf fAfugCfcCfuUfsusUf	1329	{ Phosphate } asAfsuGfgCfAfugAfaGfCfA fgGfaAfcUfsu	1330
D-2101	{ GalNac3K2AhxC6 } CfcUfgCfuUfcAfUf GfCfcCfuUfcCfuAfsusUf	1331	{ Phosphate } usAfsuGfaGfGfgcauGfaAf gCfaGfgUfsu	1332
D-2102	{ GalNac3K2AhxC6 } CfcUfgCfuUfcAfUfG fCfcCfuUfcCfuAfsusUf	1333	{ Phosphate } usAfsuGfaGfGfgCfAfugAfaA fgCfaGfgUfsu	1334
D-2103	{ GalNac3K2AhxC6 } augcccUfcCfUfAfC faguggcsusu	1335	{ Phosphate } gsGfscCfaCfUfguagAfaGf ggcausu	1336
D-2104	{ GalNac3K2AhxC6 } auggcuUfcCfAfGfA fuaugccusu	1337	{ Phosphate } asGfsgCfaUfAfucugGfaAf gccausu	1338
D-2105	{ GalNac3K2AhxC6 } gugacaAfcGfUfAfC fccuucasusu	1339	{ Phosphate } asUfsgAfaGfGfguacGfuUf gucacsusu	1340
D-2106	{ GalNac3K2AhxC6 } guauguUfcCfUfGfC fuucagcsusu	1341	{ Phosphate } gsCfsaUfgAfAfgcagGfaAf fcauacsusu	1342
D-2107	{ GalNac3K2AhxC6 } guauguUfcCfUfGfCf uucagcsusu	1343	{ Phosphate } gsCfsaUfgAfAfgcagGfaAf cauacsusu	1344
D-2108	{ GalNac3K2AhxC6 } guauguUfcCfUfGfCf uucagcsusu	1345	{ Phosphate } gsCfsaUfgAfAfgcagGfaAf fcauacsusu	1346
D-2109	{ GalNac3K2AhxC6 } guauguUfcCfUfGfC fuucagcsusu	1347	{ Phosphate } asGfsgCfaUfAfAfgcagGf aAfcuacsusu	1348
D-2110	{ GalNac3K2AhxC6 } uggauGfuUfcCfCfU fgcuucasusu	1349	{ Phosphate } gsCfsaUfgAfaGfCfaggaAf cAfuaccusu	1350
D-2111	{ GalNac3K2AhxC6 } guauguUfcCfUfGfC fuucagcsusu	1351	{ Phosphate } gsCfsaUfgAfAfgcagGfaAf cauacsusu	1352
D-2112	{ GalNac3K2AhxC6 } guauguUfcCfUfGfC fuucagcs { invAb }	1353	{ Phosphate } gsCfsaUfgAfAfgcagGfaAf cauacsusu	1354

D-2113	{GalNAc3K2AhxC6}GfcUfuCfaUfgCfCfCfUfucUfaCfaUfsusUf	1355	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1356
D-2114	{GalNAc3K2AhxC6}GfcUfuCfaUfgCfCfCfUfucUfaCfaUfsusUf	1357	{Phosphate}asUfsgUfaGfAfagGfgCfaUfGfAfaGfcsUfsu	1358
D-2115	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacaususu	1359	{Phosphate}asusguaGfAfagggCfaUfgaagcsusu	1360
D-2116	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucUfaCfaUfsusUf	1361	{Phosphate}asusguaGfAfagggCfaUfgaagcsusu	1362
D-2117	{GalNAc3K2AhxC6}GfcUfuCfaUfgCfCfCfUfucacaususu	1363	{Phosphate}asusguaGfAfagggCfaUfgaagcsusu	1364
D-2118	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacaususu	1365	{Phosphate}asusguaGfAfagggCfaUfgAfaGfcsUfsu	1366
D-2119	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacaususu	1367	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1368
D-2120	{GalNAc3K2AhxC6}GfcUfuCfaUfgCfCfCfUfucUfaCfaUfsusUf	1369	{Phosphate}asusguaGfAfagggCfaUfgaagcsusu	1370
D-2121	{GalNAc3K2AhxC6}GfcUfuCfaUfgCfCfCfUfucacaususu	1371	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1372
D-2122	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucUfaCfaUfsusUf	1373	{Phosphate}asusguaGfAfagggCfaUfgAfaGfcsUfsu	1374
D-2123	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucUfaCfaUfsusUf	1375	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1376
D-2124	{GalNAc3K2AhxC6}GfcUfuCfaUfgCfCfCfUfucacaususu	1377	{Phosphate}asusguaGfAfagggCfaUfgAfaGfcsUfsu	1378
D-2125	{GalNAc3K2AhxC6}GfcUfuCfaUfgCfCfCfUfucacaususu	1379	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1380
D-2126	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucUfaCfaUfsusUf	1381	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1382
D-2127	{GalNAc3K2AhxC6}GfcUfuCfaUfgCfCfCfUfucUfaCfaUfsusUf	1383	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1384
D-2128	{GalNAc3K2AhxC6}GfcUfuCfaUfgCfCfCfUfucUfaCfaUfsusUf	1385	{Phosphate}asusguaGfAfagggCfaUfgAfaGfcsUfsu	1386
D-2129	{GalNAc3K2AhxC6}gcuucaUfg[dC]CfCfuucacaususu	1387	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1388
D-2130	{GalNAc3K2AhxC6}gcuucaUfgCfCf[dC]Ufucacaususu	1389	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1390
D-2131	{GalNAc3K2AhxC6}gcuucaUfgCf[dC]CfUfucacaususu	1391	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1392
D-2132	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacaususu	1393	{Phosphate}asUfsgUfaGfAfagGfgCfaUfGfAfaGfcsUfsu	1394
D-2133	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacaususu	1395	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1396
D-2134	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacaususu	1397	{Phosphate}asUfsgUfaGfAfagGfgCfaUfGfAfaGfcsUfsu	1398
D-2135	{GalNAc3K2AhxC6}GfscsUfuCfaUfgCfCfUfucUfaCfaUfsusUf	1399	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1400
D-2136	{GalNAc3K2AhxC6}GfscsUfuCfaUfgCfCfCfUfucUfaCfaUfsusUf	1401	{Phosphate}asUfsgUfaGfAfagGfgCfaUfGfAfaGfcsUfsu	1402
D-2137	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacagsusu	1403	{Phosphate}asUfscUfgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1404
D-2138	{GalNAc3K2AhxC6}cugcuuUfaUfgGfCfCfucacaususu	1405	{Phosphate}asUfsgUfaGfAfagGfgCfaUfGfAfaGfcsUfsu	1406
D-2139	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacaususu	1407	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1408
D-2140	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacaus{invAb}	1409	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1410
D-2141	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacuuus{invAb}	1411	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1412
D-2142	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacaususu	1413	{Phosphate}AfsusGfuAfgAfagggCfaUfGfAfaGfcsUfsu	1414
D-2143	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacaususu	1415	{Phosphate}AfsusGfuAfgAfagggCfaUfGfAfaGfcsUfsu	1416
D-2144	{GalNAc3K2AhxC6}gcuucaugCfCfCfUfucacaususu	1417	{Phosphate}asUfsgUfaGfAfagggCfaUfGfAfaGfcsUfsu	1418
D-2145	{GalNAc3K2AhxC6}gcuucaUfgCfCfCfUfucacaususu	1419	{Phosphate}asUfsgUfaGfAfagggCfaugGfAfaGfcsUfsu	1420

D-2146	{GalNac3K2AhxC6}gcuucaUfgCfCfCfU fucuacaususu	1421	{Phosphate}asusgUfaGfAfagggCfaUfg aagcsusu	1422
D-2147	{GalNac3K2AhxC6}gcuucaUfgCfCfCfU fucuacaususu	1423	{Phosphate}asUfsgUfagAfagggCfaUfg aagcsusu	1424
D-2148	{GalNac3K2AhxC6}gcuucaUfgCfCfCfU fucuacaususu	1425	{Phosphate}asusgUfagAfagggCfaUfga agcsusu	1426
D-2149	{GalNac3K2AhxC6}gcuucaUfgCfCfCfU fucuacaususu	1427	{Phosphate}asUfsguagAfagggCfaUfga agcsusu	1428
D-2150	{GalNac3K2AhxC6}gcuucaUfgCfCfCfU fucuacaususu	1429	{Phosphate}asusguagAfagggCfaUfgaa gcsusu	1430
D-2151	{GalNac3K2AhxC6}guauguUfcCfUfGfC fuucaugcusu{invAb}	1431	{Phosphate}gsCfsaUfgAfAfgcagGfaAf cauacsusu	1432
D-2152	{GalNac3K2AhxC6}gguaugUfuCfCfUfG fucuauususu	1433	{Phosphate}aAfsuGfaAfGfcaggAfaCfa uaccsusu	1434
D-2153	{GalNac3K2AhxC6}guauguUfcCfUfGfC fuucaugususu	1435	{Phosphate}asCfsaUfgAfAfgcagGfaAf cauacsusu	1436
D-2154	{GalNac3K2AhxC6}cggccAfuGfUfCfC faccagcsusu	1437	{Phosphate}asGfscUfgGfUfggacAfuUf ggccgsusu	1438
D-2155	{GalNac3K2AhxC6}uggagcAfgAfCfUfC fugcaggususu	1439	{Phosphate}asCfscUfgCfAfgaguCfuGf uccasusu	1440
D-2156	{GalNac3K2AhxC6}acguacCfcUfUfCfA fuugaugususu	1441	{Phosphate}aCfsaUfcAfAfugaaGfgGfu acgususu	1442
D-2157	{GalNac3K2AhxC6}ccagagAfcUfGfGfU fgacaugususu	1443	{Phosphate}asCfsaUfgUfCfaccagGfuCf ucuggsusu	1444
D-2158	{GalNac3K2AhxC6}[invAb]gcuucaUfg CfCfUfUfucuacaususu	1445	{Phosphate}asUfsgUfaGfAfaaggCfaUf gaagcsusu	1446
D-2159	{GalNac3K2AhxC6}[invAb]gcuucaUfg CfCfUfUfucuacaususu	1447	{Phosphate}asAfsaUfgUfaGfAfaaggCf aUfgaagcsusu	1448
D-2160	{GalNac3K2AhxC6}[invAb]gcuucaUfg CfCfUfUfCfuaacaususu	1449	{Phosphate}asAfsaUfgUfAfgaaaGfgCf aUfgaagcsusu	1450
D-2161	{GalNac3K2AhxC6}ugcuucaAfuGfCfCfU fuucacaususu	1451	{Phosphate}usGfscAfuGfAfaggaCfuGf aagcasusu	1452
D-2162	{GalNac3K2AhxC6}uaguuCfcUfGfCfU fucaugcsusu	1453	{Phosphate}asGfscAfuGfAfagcaGfgAf acauasusu	1454
D-2163	{GalNac3K2AhxC6}uuccugCfuUfCfAfU fgccuuuasusu	1455	{Phosphate}asAfsaAfgGfCfaugaAfgCf aggaasusu	1456
D-2164	{GalNac3K2AhxC6}ucaugcCfuUfCfCfU facagugususu	1457	{Phosphate}asCfsaCfuGfUfagaaAfgGf caugasusu	1458
D-2165	{GalNac3K2AhxC6}caugccUfuUfCfUfA fcaguggususu	1459	{Phosphate}asCfscAfcUfGfuagaAfaGf gcaugsusu	1460
D-2166	{GalNac3K2AhxC6}augccuUfuCfUfAfc faguggcsusu	1461	{Phosphate}asGfscCfaCfUfguagAfaAf ggcaususu	1462
D-2167	{GalNac3K2AhxC6}gguaugUfuCfCfUfG fucuauasusu	1463	{Phosphate}usAfsuGfaAfGfcaggAfaCf auaccsusu	1464
D-2168	{GalNac3K2AhxC6}guauguUfcCfUfGfC fuucaugasusu	1465	{Phosphate}usCfsaUfgAfAfgcagGfaAf cauacsusu	1466
D-2169	{GalNac3K2AhxC6}uaguuCfcUfGfCfU fucaugcasusu	1467	{Phosphate}usGfscAfuGfAfagcaGfgAf acauasusu	1468
D-2170	{GalNac3K2AhxC6}uuccugCfuUfCfAfU fgccuuuasusu	1469	{Phosphate}usAfsaAfgGfCfaugaAfgCf aggaasusu	1470
D-2171	{GalNac3K2AhxC6}cugcuuCfaUfGfCfC fuucuaasusu	1471	{Phosphate}usUfsgUfaGfAfaaggCfaUf agcagsusu	1472
D-2172	{GalNac3K2AhxC6}gcuucaUfgCfCfUfU fucuacaasusu	1473	{Phosphate}usUfsgUfaGfAfaaggCfaUf gaagcsusu	1474
D-2173	{GalNac3K2AhxC6}uucaugCfcUfUfUfC fuacaguasusu	1475	{Phosphate}usAfsaUfgUfAfgaaaGfgCf augaasusu	1476
D-2174	{GalNac3K2AhxC6}ucaugcCfuUfUfCfU facagugasusu	1477	{Phosphate}usCfsaCfuGfUfagaaAfgGf caugasusu	1478
D-2175	{GalNac3K2AhxC6}caugccUfuUfCfUfA fcaguggasusu	1479	{Phosphate}usCfscAfcUfGfuagaAfaGf gcaugsusu	1480
D-2176	{GalNac3K2AhxC6}augccuUfuCfUfAfc faguggcasusu	1481	{Phosphate}usGfscCfaCfUfguagAfaAf ggcaususu	1482
D-2177	{GalNac3K2AhxC6}acguacCfcUfUfCfA fuugaugasusu	1483	{Phosphate}usCfsaUfcAfAfugaaGfgGf uacgususu	1484
D-2178	{GalNac3K2AhxC6}ccagagAfcUfGfGfU fgacaugasusu	1485	{Phosphate}usCfsaUfgUfCfaccagGfuCf ucuggsusu	1486

D-2179	{GalNac3K2AhxC6}auggcuUfcCfAfGfAfuauGCCAsusu	1487	{Phosphate}usGfsgCfaUfAfucugGfaAfGCCAsusu	1488
D-2180	{GalNac3K2AhxC6}guuccuGfcUfUfCfaFugcauususu	1489	{Phosphate}asAfsaGfgCfAfugaaGfcAfGgaCsusu	1490
D-2181	{GalNac3K2AhxC6}ccugcuUfcAfUfGfCfcuuucuasusu	1491	{Phosphate}usAfsgAfaAfGfgcauGfaAfGcaggsusu	1492
D-2182	{GalNac3K2AhxC6}gcuucaUfgCfCfUfUfucuaCausu	1493	{Phosphate}asUfsgUfaGfAfaaggCfaUfGaagCsusu	1494
D-2183	{GalNac3K2AhxC6}cuucauGfcCfUfUfUfouacagususu	1495	{Phosphate}asCfsuGfuAfGfGfaagGfcAfugaagsusu	1496
D-2184	{GalNac3K2AhxC6}uucAugCfcUfUfUfCfuacaguususu	1497	{Phosphate}asAfscUfgUfAfGaaGfgCfaugaasusu	1498
D-2185	{GalNac3K2AhxC6}gcuucaUfcCfCfUfUfucuaCausu	1499	{Phosphate}asUfsgUfaGfAfaaggGfaUfGaagCsusu	1500
D-2186	{GalNac3K2AhxC6}[invAb]gcuucaUfgCfCfCfUfucuaCausu	1501	{Phosphate}asUfsgUfaGfAfaaggCfaUfGaagCsusu	1502
D-2187	{GalNac3K2AhxC6}[invAb]gcuucaUfgCfCfCfUfucuaCausu	1503	{Phosphate}asAfsaUfgUfaGfAfagggCfaUfGaagCsusu	1504
D-2188	{GalNac3K2AhxC6}[invAb]gcuucaUfgCfcCfUfUfCfuacaususu	1505	{Phosphate}asAfsaUfgUfAfgaagGfgCfaUfGaagCsusu	1506
D-2189	{GalNac3K2AhxC6}[invAb]gcggcuUfcCfUfGfGfgcuucuasusu	1507	{Phosphate}usAfsgAfaGfCfccagGfaAfGCCCsusu	1508
D-2190	{GalNac3K2AhxC6}[invAb]CfuGfcGfgCfuUfcCfuGfGfgCfuCfsusAf	1509	{Phosphate}usAfsgAfaGfCfccAfgGfaAfgCfcGfcAfgsUfsu	1510
D-2191	{GalNac3K2AhxC6}cugcgGcFuUfCfCfUfgggcuuc{invAb}	1511	{Phosphate}usAfsgAfaGfcCfCfaggaAfGfcGcgagsusu	1512
D-2192	{GalNac3K2AhxC6}[invAb]cugcgGcFuUfCfCfUfgggcuucsusa	1513	{Phosphate}usAfsgAfaGfcCfCfaggaAfGfcGcgagsusu	1514
D-2193	{GalNac3K2AhxC6}[invAb]gcggcuUfcCfUfGfGfgCfuCfuAfsusUf	1515	{Phosphate}usAfsgAfaGfCfccagGfaAfGCCCsusu	1516
D-2194	{GalNac3K2AhxC6}cugcgGcFuUfCfCfUfggGfcUfuCfus{invAb}	1517	{Phosphate}usAfsgAfaGfcCfCfaggaAfGfcGcgagsusu	1518
D-2195	{GalNac3K2AhxC6}[invAb]cugcgGcFuUfCfCfUfggGfcUfuCfsusAf	1519	{Phosphate}usAfsgAfaGfcCfCfaggaAfGfcGcgagsusu	1520
D-2196	{GalNac3K2AhxC6}[invAb]GfcGfgCfuUfcCfuGfGfgCfuCfuAfsusUf	1521	{Phosphate}usAfsgAfaGfCfccAfgGfaAfgCfcGfcsUfsu	1522
D-2197	{GalNac3K2AhxC6}CfuGfcGfgCfuUfcCfUfggGfcUfuCfus{invAb}	1523	{Phosphate}usAfsgAfaGfcCfCfagGfaAfgCfcGfcAfgsUfsu	1524
D-2198	{GalNac3K2AhxC6}[invAb]CfuGfcGfgCfuUfcCfUfggGfcUfuCfsusAf	1525	{Phosphate}usAfsgAfaGfcCfCfagGfaAfgCfcGfcAfgsUfsu	1526
D-2199	{GalNac3K2AhxC6}[invAb]GfcGfgCfuUfcCfUfGfGfgcuucuasusu	1527	{Phosphate}usAfsgAfaGfCfccagGfaAfGfcGfcsUfsu	1528
D-2200	{GalNac3K2AhxC6}[invAb]CfuGfcGfgCfuUfcCfCfUfgggcuucsusa	1529	{Phosphate}usAfsgAfaGfcCfCfaggaAfGfcGfcAfgsUfsu	1530
D-2201	{GalNac3K2AhxC6}[invAb]cugcgGcUfcCfUfGfGfgCfuCfusAf	1531	{Phosphate}usAfsgAfaGfCfccagGfaAfGCCCsusu	1532
D-2202	{GalNac3K2AhxC6}[invAb]CfuGfcGfgCfuUfcCfUfGfGfgcuucsusa	1533	{Phosphate}usAfsgAfaGfCfccagGfaAfGfcGfcAfgsUfsu	1534
D-2203	{GalNac3K2AhxC6}[invAb]auggcuUfcCfAfGfAfuauGCCAsusu	1535	{Phosphate}asGfsgCfaUfAfucugGfaAfGCCAsusu	1536
D-2204	{GalNac3K2AhxC6}[invAb]AfcAfuGfgCfuUfcCfaGfAfuUfgCfscsUf	1537	{Phosphate}asGfsgCfaUfAfucUfgGfaAfgCfcAfuGfusUfsu	1538
D-2205	{GalNac3K2AhxC6}acauggCfuUfcCfCfAfgauGCCs{invAb}	1539	{Phosphate}asGfsgCfaUfaUfCfuggaAfGcCcaugususu	1540
D-2206	{GalNac3K2AhxC6}[invAb]acauggCfuUfcCfCfAfgauaGccscsu	1541	{Phosphate}asGfsgCfaUfaUfCfuggaAfGcCcaugususu	1542
D-2207	{GalNac3K2AhxC6}[invAb]auggcuUfcCfAfGfAfuUfgCfCfUfsusUf	1543	{Phosphate}asGfsgCfaUfAfucugGfaAfGCCAsusu	1544
D-2208	{GalNac3K2AhxC6}acauggCfuUfcCfCfAfgaUfaUfgCfcs{invAb}	1545	{Phosphate}asGfsgCfaUfaUfCfuggaAfGcCcaugususu	1546
D-2209	{GalNac3K2AhxC6}[invAb]acauggCfuUfcCfCfAfgaUfaUfgCfscsUf	1547	{Phosphate}asGfsgCfaUfaUfCfuggaAfGcCcaugususu	1548
D-2210	{GalNac3K2AhxC6}[invAb]AfuGfgCfuUfcCfaGfAfuUfgCfcUfsusUf	1549	{Phosphate}asGfsgCfaUfAfucUfgGfaAfgCfcAfusUfsu	1550
D-2211	{GalNac3K2AhxC6}AfcAfuGfgCfuUfcCfAfgaUfaUfgCfcs{invAb}	1551	{Phosphate}asGfsgCfaUfaUfCfugGfaAfgCfcAfuGfusUfsu	1552

D-2212	{GalNac3K2AhxC6}[invAb]AfcAfuGfgCfuUfcCfAfgaUfaUfgCfscsUf	1553	{Phosphate}asGfsgCfaUfaUfCfugGfaAfgCfcAfuGfusUfsu	1554
D-2213	{GalNac3K2AhxC6}[invAb]AfuGfgCfuUfcCfAfgaUfaUfgCfscsUf	1555	{Phosphate}asGfsgCfaUfaUfCfugGfaAfgCfcAfuGfusUfsu	1556
D-2214	{GalNac3K2AhxC6}AfcAfuGfgCfuUfcCfAfgaUfaUfgCfscsUf	1557	{Phosphate}asGfsgCfaUfaUfCfugGfaAfgCfcAfuGfusUfsu	1558
D-2215	{GalNac3K2AhxC6}[invAb]AfcAfuGfgCfuUfcCfAfgaUfaUfgCfscsUf	1559	{Phosphate}asGfsgCfaUfaUfCfugGfaAfgCfcAfuGfusUfsu	1560
D-2216	{GalNac3K2AhxC6}[invAb]acauggeUfcCfAfgaUfaUfgCfscsUf	1561	{Phosphate}asGfsgCfaUfaUfCfugGfaAfgCfcAfuGfusUfsu	1562
D-2217	{GalNac3K2AhxC6}[invAb]acauggeUfcCfAfgaUfaUfgCfscsUf	1563	{Phosphate}asGfsgCfaUfaUfCfugGfaAfgCfcAfuGfusUfsu	1564
D-2218	{GalNac3K2AhxC6}[invAb]AfcAfuGfgCfuUfcCfAfgaUfaUfgCfscsUf	1565	{Phosphate}asGfsgCfaUfaUfCfugGfaAfgCfcAfuGfusUfsu	1566
D-2219	{GalNac3K2AhxC6}[invAb]acguacCfcUfUfcCfAfuugaugususu	1567	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1568
D-2220	{GalNac3K2AhxC6}[invAb]CfaAfcGfuAfcCfcUfuCfAfuugaugususu	1569	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1570
D-2221	{GalNac3K2AhxC6}[invAb]caacguAfcCfcUfUfcCfAfuugaugususu	1571	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1572
D-2222	{GalNac3K2AhxC6}[invAb]acguacCfcUfUfcCfAfuugaugususu	1573	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1574
D-2223	{GalNac3K2AhxC6}caacguAfcCfcUfUfcCfAfuugaugususu	1575	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1576
D-2224	{GalNac3K2AhxC6}[invAb]AfcGfuAfcCfcUfUfcCfAfuugaugususu	1577	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1578
D-2225	{GalNac3K2AhxC6}CfaAfcGfuAfcCfcUfUfcCfAfuugaugususu	1579	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1580
D-2226	{GalNac3K2AhxC6}[invAb]CfaAfcGfuAfcCfcUfUfcCfAfuugaugususu	1581	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1582
D-2227	{GalNac3K2AhxC6}[invAb]AfcGfuAfcCfcUfUfcCfAfuugaugususu	1583	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1584
D-2228	{GalNac3K2AhxC6}CfaAfcGfuAfcCfcUfUfcCfAfuugaugususu	1585	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1586
D-2229	{GalNac3K2AhxC6}[invAb]CfaAfcGfuAfcCfcUfUfcCfAfuugaugususu	1587	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1588
D-2230	{GalNac3K2AhxC6}[invAb]caacguacCfcUfUfcCfAfuugaugususu	1589	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1590
D-2231	{GalNac3K2AhxC6}[invAb]caacguacCfcUfUfcCfAfuugaugususu	1591	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1592
D-2232	{GalNac3K2AhxC6}[invAb]CfaAfcGfuAfcCfcUfUfcCfAfuugaugususu	1593	{Phosphate}asCfsaUfcAfaUfGfaagGfuAfcGfuUfgsUfsu	1594
D-2233	{GalNac3K2AhxC6}cugcuucaUfgCfcUfUfucuaacsasu	1595	{Phosphate}asUfsgUfaGfAfaagGfaUfgAfaGfcsUfsu	1596
D-2234	{GalNac3K2AhxC6}cugcuucaUfgCfcUfUfucuaacsasu	1597	{Phosphate}asUfsgUfaGfAfaagGfaUfgAfaGfcsUfsu	1598
D-2235	{GalNac3K2AhxC6}[invAb]GfcUfuCfaUfgCfcUfUfucuaacsasu	1599	{Phosphate}asUfsgUfaGfAfaagGfaUfgAfaGfcsUfsu	1600
D-2236	{GalNac3K2AhxC6}[invAb]GfcUfuCfaUfgCfcUfUfucuaacsasu	1601	{Phosphate}asUfsgUfaGfAfaagGfaUfgAfaGfcsUfsu	1602
D-2237	{GalNac3K2AhxC6}CfuGfcUfuCfaUfgCfcUfUfucuaacsasu	1603	{Phosphate}asUfsgUfaGfAfaagGfaUfgAfaGfcsUfsu	1604
D-2238	{GalNac3K2AhxC6}CfuGfcUfuCfaUfgCfcUfUfucuaacsasu	1605	{Phosphate}asUfsgUfaGfAfaagGfaUfgAfaGfcsUfsu	1606
D-2239	{GalNac3K2AhxC6}[invAb]CfuGfcUfuCfaUfgCfcUfUfucuaacsasu	1607	{Phosphate}asUfsgUfaGfAfaagGfaUfgAfaGfcsUfsu	1608
D-2240	{GalNac3K2AhxC6}CfuGfcUfuCfaUfgCfcUfUfucuaacsasu	1609	{Phosphate}asUfsgUfaGfAfaagGfaUfgAfaGfcsUfsu	1610
D-2241	{GalNac3K2AhxC6}[invAb]CfuGfcUfuCfaUfgCfcUfUfucuaacsasu	1611	{Phosphate}asUfsgUfaGfAfaagGfaUfgAfaGfcsUfsu	1612
D-2242	{GalNac3K2AhxC6}[invAb]cugcuucaUfgCfcUfUfucuaacsasu	1613	{Phosphate}asUfsgUfaGfAfaagGfaUfgAfaGfcsUfsu	1614
D-2243	{GalNac3K2AhxC6}cugcuucaUfgCfcUfUfucuaacsasu	1615	{Phosphate}asUfsgUfaGfAfaagGfaUfgAfaGfcsUfsu	1616
D-2244	{GalNac3K2AhxC6}[invAb]cugcuucaUfgCfcUfUfucuaacsasu	1617	{Phosphate}asUfsgUfaGfAfaagGfaUfgAfaGfcsUfsu	1618

D-2245	{GalNAc3K2AhxC6}cugcuucaUfgCfcUfUfucucacsasu	1619	{Phosphate}asUfsgUfaGfAfaaggCfaUfgaagcagsusu	1620
D-2246	{GalNAc3K2AhxC6}cugcuucaUfgCfcUfUfucucacsasu{invAb}	1621	{Phosphate}asUfsgUfaGfAfaaggCfaUfgaagcagsusu	1622
D-2247	{GalNAc3K2AhxC6}[invAb]cugcuucaUfgCfcUfUfucucacsasu	1623	{Phosphate}asUfsgUfaGfAfaaggCfaUfgaagcagsusu	1624
D-2248	{GalNAc3K2AhxC6}cugcuucaUfgCfcUfUfucucacsasu	1625	{Phosphate}asUfsgUfagAfaaggCfaUfgaagcagsusu	1626
D-2249	{GalNAc3K2AhxC6}cugcuucaUfgCfcUfUfucucacsasu{invAb}	1627	{Phosphate}asUfsgUfagAfaaggCfaUfgaagcagsusu	1628
D-2250	{GalNAc3K2AhxC6}[invAb]cugcuucaUfgCfcUfUfucucacsasu	1629	{Phosphate}asUfsgUfagAfaaggCfaUfgaagcagsusu	1630
D-2251	{GalNAc3K2AhxC6}cugcuucaUfgCfcUfUfucucacsasu{invAb}	1631	{Phosphate}asUfsguagAfaaggCfaUfgaagcagsusu	1632
D-2252	{GalNAc3K2AhxC6}[invAb]cugcuucaUfgCfcUfUfucucacsasu	1633	{Phosphate}asUfsguagAfaaggCfaUfgaagcagsusu	1634
D-2253	{GalNAc3K2AhxC6}[invAb]GfcUfuCfaUfgCfcUfUfucUfaCfaUfsusUf	1635	{Phosphate}asUfsgUfagAfaaGfgCfaUfgAfaGfcsUfsu	1636
D-2254	{GalNAc3K2AhxC6}[invAb]GfcUfuCfaUfgCfcUfUfucUfaCfaUfsusUf	1637	{Phosphate}asUfsgUfagAfaaggCfaUfgAfaGfcsUfsu	1638
D-2255	{GalNAc3K2AhxC6}[invAb]GfcUfuCfaUfgCfcUfUfucUfaCfaUfsusUf	1639	{Phosphate}asUfsgUfagAfaaggCfaUfgaagcsusu	1640
D-2256	{GalNAc3K2AhxC6}[invAb]gcuucaUfgCfcUfUfucUfaCfaUfsusUf	1641	{Phosphate}asUfsgUfagAfaaGfgCfaUfgaagcsusu	1642
D-2257	{GalNAc3K2AhxC6}[invAb]gcuucaUfgCfcUfUfucUfaCfaUfsusUf	1643	{Phosphate}asUfsgUfagAfaaGfgCfaUfgaagcsusu	1644
D-2258	{GalNAc3K2AhxC6}cugcuucaUfgCfcUfUfucucacsasu	1645	{Phosphate}asUfsgUfaGfAfaaggCfaUfgaagcagsusu	1646
D-2259	{GalNAc3K2AhxC6}[invAb]gcuucaUfgCfcUfUfucUfaCfaUfsusUf	1647	{Phosphate}asUfsgUfaGfAfaaggCfaUfgaagcsusu	1648
D-2260	{GalNAc3K2AhxC6}[invAb]GfcUfuCfaUfgCfcUfUfucUfaCfaUfsusUf	1649	{Phosphate}asUfsgUfaGfAfaaggCfaUfgAfaGfcsUfsu	1650
D-2261	{GalNAc3K2AhxC6}[invAb]GfcUfuCfaUfgCfcUfUfucUfaCfaUfsusUf	1651	{Phosphate}asUfsgUfaGfAfaaggCfaUfgAfaGfcsUfsu	1652
D-2262	{GalNAc3K2AhxC6}cugcuucaUfgCfcUfUfucucacsasu{invAb}	1653	{Phosphate}asUfsgUfaGfAfaaggCfaUfgaagcagsusu	1654
D-2263	{GalNAc3K2AhxC6}[invAb]cugcuucaUfgCfcUfUfucucacsasu	1655	{Phosphate}asUfsgUfaGfAfaaggCfaUfgaagcagsusu	1656
D-2264	{GalNAc3K2AhxC6}[invAb]GfcUfuCfaUfgCfcUfUfucUfaCfaUfsusUf	1657	{Phosphate}asUfsgUfaGfAfaaggCfaUfgaagcsusu	1658
D-2265	{GalNAc3K2AhxC6}auguucCfuGfCfUfUfcaugccususu	1659	{Phosphate}asGfsgCfaUfgGfaagCfGfcaacaususu	1660
D-2266	{GalNAc3K2AhxC6}uguuccUfgCfUfUfCfaugccususu	1661	{Phosphate}asAfsgGfCfAfUfgaagCfaGfcaacaususu	1662
D-2267	{GalNAc3K2AhxC6}ugccuuUfCfUfAfCfAfgugccususu	1663	{Phosphate}asGfsgCfCfAfCfuguaGfaAfaggcasusu	1664
D-2268	oguacuUfCfGfUfCfCfuuguaugsusu	1665	{Phosphate}csAfsuAfcAfAfggacGfaAfguagcsusu	1666
D-2269	{GalNAc3K2AhxC6}[invAb]gcuucaUfgCf[dC]CfUfucUfaCfaUfsusUf	1667	{Phosphate}asUfsgUfaGfAfaaggCfaUfgaagcsusu	1668
D-2270	{GalNAc3K2AhxC6}[invAb]gcuucaUfgCfCfCfUfucUfaCfaUfsusUf	1669	{Phosphate}asUfsgUfaGfAfaaggCfaUfgaagcsusu	1670
D-2271	{GalNAc3K2AhxC6}[invAb]gcuucaUfgCfCfCfUfucUfaCfaUfsusUf	1671	{Phosphate}asUfsgUfaGfAfaaggCfaUfgaagcsusu	1672
D-2272	{GalNAc3K2AhxC6}[invAb]gcuucaUfgCfCfCfUfucUfaCfaUfsusUf	1673	{Phosphate}asUfsgUfagAfaaggCfaUfgaagcsusu	1674
D-2273	{GalNAc3K2AhxC6}[invAb]gcuucaUfgCfCfCfUfucUfaCfaUfsusUf	1675	{Phosphate}asUfsguagAfaaggCfaUfgaagcsusu	1676
D-2274	{GalNAc3K2AhxC6}[invAb]GfcUfuCfaUfgCfcUfUfucUfaCfaUfsusUf	1677	{Phosphate}asUfsgUfaGfAfaaggCfaUfgaagcsusu	1678
D-2275	{sGalNAc3K2AhxC6}[invAb]ccugcuUfcaUfGfCfCfUfCfuAfsusUf	1679	{Phosphate}usAfsgAfaAfGfgcaUfGfaAfgcagsusu	1680
D-2276	{sGalNAc3K2AhxC6}[invAb]CfcUfgCfUfCfAfuGfCfCfUfCfuAfsusUf	1681	{Phosphate}usAfsgAfaAfGfgcaUfGfaAfgCfAfgGfUfsu	1682
D-2277	{sGalNAc3K2AhxC6}UfuCfcUfgCfUfCfAfuGfCfCfUfCfuAfsusUf	1683	{Phosphate}usAfsgAfaAfGfgCfCfAfuGfaAfgCfAfgGfAfasUfsu	1684

D-2278	{sGalNac3K2AhxC6}[invAb]CfcUfgCf uUfcAfUfGfCfcuuucuasusu	1685	{Phosphate}usAfsGfAfaAfGfgcauGfaAf gCfaGfgsUfsu	1686
D-2279	{sGalNac3K2AhxC6}UfuCfcUfgCfuUfC fAfUfgccuuucus{invAb}	1687	{Phosphate}usAfsGfAfaAfGfCfAugaAf gCfaGfgAfasUfsu	1688
D-2280	{sGalNac3K2AhxC6}[invAb]augccuUf uCfUfAfCfaguggcususu	1689	{Phosphate}asGfscCfaCfUfguagAfaAf ggcaususu	1690
D-2281	{sGalNac3K2AhxC6}[invAb]augccuUf uCfUfAfCfagUfgGfcUfsusUf	1691	{Phosphate}asGfscCfaCfUfguagAfaAf ggcaususu	1692
D-2282	{sGalNac3K2AhxC6}[invAb]AfUGfcCf uUfuCfuAfCfagUfgGfcUfsusUf	1693	{Phosphate}asGfscCfaCfUfguAfgAfaA fgGfcAfusUfsu	1694
D-2283	{sGalNac3K2AhxC6}UfcAfUGfcCfuUfu CfUfacAfUGfgfcs{invAb}	1695	{Phosphate}asGfscCfaCfuGfUfagAfaA fgGfcAfUGfasUfsu	1696
D-2284	{sGalNac3K2AhxC6}[invAb]AfUGfcCf uUfuCfuAfCfaguggcususu	1697	{Phosphate}asGfscCfaCfUfguagAfaAf ggfcAfusUfsu	1698
D-2285	{sGalNac3K2AhxC6}[invAb]UfcAfUGf cCfuUfUfCfUfacaguggscsu	1699	{Phosphate}asGfscCfaCfuGfUfagaaAf ggfcAfUGfasUfsu	1700
D-2286	{sGalNac3K2AhxC6}[invAb]ugcuucAf uGfCfCfuUfucacagsu	1701	{Phosphate}asCfsuGfuAfgAfaAggCfAf uGfaagcasusu	1702
D-2287	{sGalNac3K2AhxC6}[invAb]cuucAuGf cCfUfUfUfcuAfcAfUGfUfsusUf	1703	{Phosphate}asCfsuGfuAfgAfaagGfcAf ugaagsusu	1704
D-2288	{sGalNac3K2AhxC6}UfgCfuUfcAfUGfc CfUfuCfuAfcAfgs{invAb}	1705	{Phosphate}asCfsuGfuAfgAfaAgGfcA fuGfaAfgCfasUfsu	1706
D-2289	{sGalNac3K2AhxC6}[invAb]CfuUfcAf uGfcCfUfUfUfcuacagususu	1707	{Phosphate}asCfsuGfuAfgAfaagGfcAf uGfaAfgsUfsu	1708
D-2290	{sGalNac3K2AhxC6}UfgCfuUfcAfUGfc fCfUfucacags{invAb}	1709	{Phosphate}asCfsuGfuAfgAfaAggCfAf uGfaAfgCfasUfsu	1710
D-2291	{sGalNac3K2AhxC6}[invAb]UfgCfuUf cAfUGfCfCfUfucacagsu	1711	{Phosphate}asCfsuGfuAfgAfaAggCfAf uGfaAfgCfasUfsu	1712
D-2292	{sGalNac3K2AhxC6}[invAb]ugcuucAu GfcCfUfUfUfcuacagsu	1713	{Phosphate}asCfsuGfuAfgAfaagGfcAf ugaagcasusu	1714
D-2293	{sGalNac3K2AhxC6}[invAb]ugcuucAu GfcCfUfUfUfcuAfcAfgsUf	1715	{Phosphate}asCfsuGfuAfgAfaagGfcAf ugaagcasusu	1716
D-2294	{sGalNac3K2AhxC6}[invAb]UfgCfuUf cAfUGfcCfUfUfUfcuacagsu	1717	{Phosphate}asCfsuGfuAfgAfaagGfcAf uGfaAfgCfasUfsu	1718
D-2295	{sGalNac3K2AhxC6}[invAb]guuccuGf cUfUfCfUfugccuuususu	1719	{Phosphate}asAfsaGfgCfAfugaaGfcAf ggaacsusu	1720
D-2296	{sGalNac3K2AhxC6}[invAb]ccugcuUf cAfUfGfCfcuuucuasusu	1721	{Phosphate}usAfsGfAfaAfGfgcauGfaAf gcaggssusu	1722
D-2297	{sGalNac3K2AhxC6}[invAb]cuucAuGf cCfUfUfUfcuacagususu	1723	{Phosphate}asCfsuGfuAfgAfaagGfcAf ugaagsusu	1724
D-2298	{sGalNac3K2AhxC6}[invAb]UfuCfcUf gCfuUfcAfUGfCfcuUfuCfsusAf	1725	{Phosphate}usAfsGfAfaAfGfgcAfUGfaA fgCfaGfgAfasUfsu	1726
D-2299	{sGalNac3K2AhxC6}uuccugCfuUfCfAf Ufgccuuucus{invAb}	1727	{Phosphate}usAfsGfAfaAfGfCfAugaAf gCfaggaasusu	1728
D-2300	{sGalNac3K2AhxC6}[invAb]uuccugCf uUfCfAfUfgccuuucsusa	1729	{Phosphate}usAfsGfAfaAfGfCfAugaAf gCfaggaasusu	1730
D-2301	{sGalNac3K2AhxC6}uuccugCfuUfCfAf UfgcCfuUfuCfus{invAb}	1731	{Phosphate}usAfsGfAfaAfGfCfAugaAf gCfaggaasusu	1732
D-2302	{sGalNac3K2AhxC6}[invAb]uuccugCf uUfCfAfUfgcCfuUfuCfsusAf	1733	{Phosphate}usAfsGfAfaAfGfCfAugaAf gCfaggaasusu	1734
D-2303	{sGalNac3K2AhxC6}[invAb]UfuCfcUf gCfuUfcAfUfgcCfuUfuCfsusAf	1735	{Phosphate}usAfsGfAfaAfGfCfauGfaA fgCfaGfgAfasUfsu	1736
D-2304	{sGalNac3K2AhxC6}[invAb]UfuCfcUf gCfuUfCfAfUfgccuuucsusa	1737	{Phosphate}usAfsGfAfaAfGfCfAugaAf gCfaGfgAfasUfsu	1738
D-2305	{sGalNac3K2AhxC6}[invAb]uuccugcu UfcAfUfGfCfcuuucsusa	1739	{Phosphate}usAfsGfAfaAfGfgcauGfaAf gcaggaasusu	1740
D-2306	{sGalNac3K2AhxC6}[invAb]uuccugcu UfcAfUfGfCfcuUfuCfusAf	1741	{Phosphate}usAfsGfAfaAfGfgcauGfaAf gcaggaasusu	1742
D-2307	{sGalNac3K2AhxC6}[invAb]UfuCfcUf gCfuUfcAfUfGfCfcuuucsusa	1743	{Phosphate}usAfsGfAfaAfGfgcauGfaAf gCfaGfgAfasUfsu	1744
D-2308	{sGalNac3K2AhxC6}[invAb]UfcAfUGf cCfuUfuCfuAfcCfagUfgGfscsUf	1745	{Phosphate}asGfscCfaCfUfguAfgAfaA fgGfcAfUGfasUfsu	1746
D-2309	{sGalNac3K2AhxC6}ucaugcCfuUfUfCf Ufacaguggcs{invAb}	1747	{Phosphate}asGfscCfaCfuGfUfagaaAf ggfcaugasusu	1748
D-2310	{sGalNac3K2AhxC6}[invAb]ucaugcCf uUfUfCfUfUfacaguggscsu	1749	{Phosphate}asGfscCfaCfuGfUfagaaAf ggfcaugasusu	1750

D-2311	{sGalNac3K2AhxC6}ucaugcCfuUfUfCfUfacAfgUfgGfcs{invAb}	1751	{Phosphate}asGfscCfaCfuGfUfagaaAfgGfcaugasusu	1752
D-2312	{sGalNac3K2AhxC6}[invAb]ucaugcCfuUfUfCfUfacAfgUfgGfcsUf	1753	{Phosphate}asGfscCfaCfuGfUfagaaAfgGfcaugasusu	1754
D-2313	{sGalNac3K2AhxC6}[invAb]UfcAfuGfcCfuUfUfUfacAfgUfgGfcsUf	1755	{Phosphate}asGfscCfaCfuGfUfagAfaAfgGfcAfuGfasUfsu	1756
D-2314	{sGalNac3K2AhxC6}UfcAfuGfcCfuUfUfCfUfacaguggcs{invAb}	1757	{Phosphate}asGfscCfaCfuGfUfagaaAfgGfcAfuGfasUfsu	1758
D-2315	{sGalNac3K2AhxC6}[invAb]ucaugccuUfuCfuUfAfcfaguggscsu	1759	{Phosphate}asGfscCfaCfuGfugagAfaAfggcaugasusu	1760
D-2316	{sGalNac3K2AhxC6}[invAb]ucaugccuUfuCfuUfAfcfagUfgGfcsUf	1761	{Phosphate}asGfscCfaCfuGfugagAfaAfggcaugasusu	1762
D-2317	{sGalNac3K2AhxC6}[invAb]UfcAfuGfcCfuUfUfUfAfcfaguggscsu	1763	{Phosphate}asGfscCfaCfuGfugagAfaAfgGfcAfuGfasUfsu	1764
D-2318	{sGalNac3K2AhxC6}[invAb]UfgCfuUfcAfuGfcCfuUfUfcuAfcAfsUf	1765	{Phosphate}asCfsuGfuAfgAfaAfgGfcAfuGfaAfgCfasUfsu	1766
D-2319	{sGalNac3K2AhxC6}ugcuucAfuGfcCfCfUfuucacags{invAb}	1767	{Phosphate}asCfsuGfuAfgAfaAfggcAfugfaagcasusu	1768
D-2320	{sGalNac3K2AhxC6}ugcuucAfuGfcCfUfuCfuAfcAfs{invAb}	1769	{Phosphate}asCfsuGfuAfgAfaAfggcAfugfaagcasusu	1770
D-2321	{sGalNac3K2AhxC6}[invAb]ugcuucAfuGfcCfUfuCfuAfcAfsUf	1771	{Phosphate}asCfsuGfuAfgAfaAfggcAfugfaagcasusu	1772
D-2322	{sGalNac3K2AhxC6}[invAb]CfuUfcAfuGfcCfuUfUfcuAfcAfgUfsUf	1773	{Phosphate}asCfsuGfuAfgAfaAfgGfcAfuGfaAfgsUfsu	1774
D-2323	{sGalNac3K2AhxC6}[invAb]UfgCfuUfcAfuGfcCfUfuCfuAfcAfsUf	1775	{Phosphate}asCfsuGfuAfgAfaAfgGfcAfuGfaAfgCfasUfsu	1776
D-2324	{sGalNac3K2AhxC6}[invAb]gcuucaUfgCfCfUfUfucuaucasusu	1777	{Phosphate}asUfsgUfaGfAfaagGcfaUfgaagcsusu	1778
D-2325	{sGalNac3K2AhxC6}[invAb]caugccUfUfCfUfAfcaguggsususu	1779	{Phosphate}asCfscAfcUfGfuagaAfaGfgcaugsusu	1780
D-2326	{sGalNac3K2AhxC6}[invAb]cugcuCfaUfGfcCfUfuucuaasusu	1781	{Phosphate}usUfsgAfaAfaagGcfaUfgaagcasusu	1782
D-2327	{sGalNac3K2AhxC6}[invAb]gcuucaUfgCfCfUfUfucuaasusu	1783	{Phosphate}usUfsgUfaGfAfaagGcfaUfgaagcsusu	1784
D-2328	{sGalNac3K2AhxC6}[invAb]uucaugCfUfUfUfCfuacaguasusu	1785	{Phosphate}usAfsCfUfUfAfgaaGfgCfaugaasusu	1786
D-2329	{sGalNac3K2AhxC6}[invAb]guuccuGfcUfUfCfAfuGfcCfuUfUfsUf	1787	{Phosphate}asAfsaGfgCfaUfGfaagCfaUfgaagcsusu	1788
D-2330	{sGalNac3K2AhxC6}auguucCfuGfcCfUfUfcaUfgCfUfUf{invAb}	1789	{Phosphate}asAfsaGfgCfaUfGfaagCfaUfgaagcsusu	1790
D-2331	{sGalNac3K2AhxC6}[invAb]auguucCfuGfcCfUfUfcaUfgCfUfUf{invAb}	1791	{Phosphate}asAfsaGfgCfaUfGfaagCfaUfgaagcsusu	1792
D-2332	{sGalNac3K2AhxC6}[invAb]GfuUfcCfuGfcUfUfCfAfuGfcCfuUfUfsUf	1793	{Phosphate}asAfsaGfgCfaUfGfaagCfaUfgaagcsusu	1794
D-2333	{sGalNac3K2AhxC6}AfuGfuUfcCfuGfcUfUfcaUfgCfUfUf{invAb}	1795	{Phosphate}asAfsaGfgCfaUfGfaagCfaUfgaagcsusu	1796
D-2334	{sGalNac3K2AhxC6}[invAb]AfuGfuUfcCfuGfcUfUfcaUfgCfUfUf{invAb}	1797	{Phosphate}asAfsaGfgCfaUfGfaagCfaUfgaagcsusu	1798
D-2335	{sGalNac3K2AhxC6}[invAb]GfuUfcCfuGfcUfUfCfAfuGfcCfuUfUfsUf	1799	{Phosphate}asAfsaGfgCfaUfGfaagCfaUfgaagcsusu	1800
D-2336	{sGalNac3K2AhxC6}AfuGfuUfcCfuGfcUfUfCfAfuGfcCfuUfUfsUf	1801	{Phosphate}asAfsaGfgCfaUfGfaagCfaUfgaagcsusu	1802
D-2337	{sGalNac3K2AhxC6}[invAb]AfuGfuUfcCfuGfcCfuUfUfcaugccususu	1803	{Phosphate}asAfsaGfgCfaUfGfaagCfaUfgaagcsusu	1804
D-2338	{sGalNac3K2AhxC6}[invAb]auguuccuGfcUfUfCfAfuGfcCfuUfUfsUf	1805	{Phosphate}asAfsaGfgCfaUfGfaagCfaUfgaagcsusu	1806
D-2339	{sGalNac3K2AhxC6}[invAb]auguuccuGfcUfUfCfAfuGfcCfuUfUfsUf	1807	{Phosphate}asAfsaGfgCfaUfGfaagCfaUfgaagcsusu	1808
D-2340	{sGalNac3K2AhxC6}[invAb]GfcUfUfCfaUfGfcCfuUfUfCfaGfsUf	1809	{Phosphate}usAfsCfUfUfAfgaAfaGfgCfaUfGfaagCfaUfUfsUf	1810
D-2341	{sGalNac3K2AhxC6}[invAb]gcuucaUfgCfCfUfUfucuaagcsusa	1811	{Phosphate}usAfsCfUfUfAfgaAfaagGcfaUfgaagcsusu	1812
D-2342	{sGalNac3K2AhxC6}[invAb]uucaugCfUfUfUfCfuaCfaGfuAfsUf	1813	{Phosphate}usAfsCfUfUfAfgaaGfgCfaugaasusu	1814
D-2343	{sGalNac3K2AhxC6}[invAb]gcuucaUfgCfCfUfUfucUfaCfaGfsUf	1815	{Phosphate}usAfsCfUfUfAfgaAfaagGcfaUfgaagcsusu	1816

D-2344	{sGalNAc3K2AhxC6}GfcUfuCfaUfgCfcfUfuUfucucacagus{invAb}	1817	{Phosphate}usAfscUfgUfaGfAfaaggCfaUfgAfaGfcsUfsu	1818
D-2345	{sGalNAc3K2AhxC6}[invAb]GfcGfgCfuUfcCfUfGfGfgcuucuasusu	1819	{Phosphate}usAfsgAfaGfCfccagGfaAfGfcGfcsUfsu	1820
D-2346	{sGalNAc3K2AhxC6}[invAb]auggcuUfcCfAfGfAfuauugccususu	1821	{Phosphate}asGfsgCfaUfaUfcugGfaAfGCCaususu	1822
D-2347	{sGalNAc3K2AhxC6}[invAb]acauggCfuUfcCfCfAfgaUfaUfgCfscsUf	1823	{Phosphate}asGfsgCfaUfaUfcugGfaAfGfcCaugususu	1824
D-2348	{sGalNAc3K2AhxC6}[invAb]AfcAfuGfgCfuUfcCfAfGfAfuauugccsusu	1825	{Phosphate}asGfsgCfaUfaUfcugGfaAfGfcCfuGfusUfsu	1826
D-2349	{sGalNAc3K2AhxC6}[invAb]caacguAfCfcCfUfuUfcuugausgsu	1827	{Phosphate}asCfsaUfcAfaUfGfaaggGfaUfcguugsusu	1828
D-2350	{sGalNAc3K2AhxC6}[invAb]caacguacCfcUfuUfcAfuUgfaUfgsUf	1829	{Phosphate}asCfsaUfcAfaUfugaaGfgGfaUcgugsusu	1830
D-2351	{sGalNAc3K2AhxC6}[invAb]acguacCfcUfuUfcAfuugausgsusu	1831	{Phosphate}asCfsaUfcAfaUfugaaGfgGfaUcgususu	1832
D-2352	{sGalNAc3K2AhxC6}cugcuucaUfgCfcfUfuUfucucac{invAb}	1833	{Phosphate}asUfsgUfaGfAfaaggCfaUfGaagcagsusu	1834
D-2353	{sGalNAc3K2AhxC6}[invAb]cugcuucaUfgCfcfUfuUfucucacsasu	1835	{Phosphate}asUfsgUfaGfAfaaggCfaUfGaagcagsusu	1836
D-2354	{sGalNAc3K2AhxC6}[invAb]gcuucaUfgCfcfUfuUfucUfaCfaUfsusUf	1837	{Phosphate}asUfsgUfaGfAfaaggCfaUfGaagcsusu	1838
D-2355	{sGalNAc3K2AhxC6}cugcuucaUfgCfcfUfuUfucUfaCfas{invAb}	1839	{Phosphate}asUfsgUfaGfAfaaggCfaUfGaagcagsusu	1840
D-2356	{sGalNAc3K2AhxC6}[invAb]cugcuucaUfgCfcfUfuUfucUfaCfasUf	1841	{Phosphate}asUfsgUfaGfAfaaggCfaUfGaagcagsusu	1842
D-2357	{sGalNAc3K2AhxC6}cugcuucaUfgCfcfUfuUfucucac{invAb}	1843	{Phosphate}asUfsguagAfaaggCfaUfGaagcagsusu	1844
D-2358	{sGalNAc3K2AhxC6}[invAb]cugcuucaUfgCfcfUfuUfucucac{invAb}	1845	{Phosphate}asUfsgUfaGfAfaaggCfaUfGaagcagsusu	1846
D-2359	{sGalNAc3K2AhxC6}[invAb]gcuucaUfgCfcfUfuUfucucacuasusu	1847	{Phosphate}asUfsgUfaGfAfaaggCfaUfGaagcsusu	1848
D-2360	{sGalNAc3K2AhxC6}[invAb]CfuUfcAfUgGfcCfUfuUfucucacagususu	1849	{Phosphate}asCfsuGfuAfGfaaagGfcAfUgaagsusu	1850
D-2361	{sGalNAc3K2AhxC6}ugcuucaUfgCfcfUfuUfucucacs{invAb}	1851	{Phosphate}asCfsuGfuAfGfaaagGfcAfUgaagcasusu	1852
D-2362	{sGalNAc3K2AhxC6}[invAb]cugcuucaUfgCfcfUfuUfucucacsasu	1853	{Phosphate}asUfsgUfaGfAfaaGfgCfaUfGaagcagsusu	1854
D-2363	{sGalNAc3K2AhxC6}cugcuucaUfgCfcfUfuUfucucacs{invAb}	1855	{Phosphate}asUfsgUfaGfAfaaggcaUfgaagcagsusu	1856
D-2364	{sGalNAc3K2AhxC6}cugcuucaUfgCfcfUfuUfucucacs{invAb}	1857	{Phosphate}asUfsgUfaGfAfaaGfgCfaUfGaagcagsusu	1858
D-2365	{sGalNAc3K2AhxC6}[invAb]cuucaUfcCfUfuUfucucacagususu	1859	{Phosphate}asCfsuGfuAfGfaaagGfgAfUgaagsusu	1860
D-2366	{sGalNAc3K2AhxC6}[invAb]gcuucaUfcCfCfUfuUfucucacuasusu	1861	{Phosphate}asUfsgUfaGfAfaaggGfaUfGaagcsusu	1862
D-2367	{sGalNAc3K2AhxC6}ugcuucaUfcCfUfuUfucucacs{invAb}	1863	{Phosphate}asCfsuGfuAfGfaaagGfgAfUgaagcasusu	1864
D-2368	{sGalNAc3K2AhxC6}cugcuucaUfcCfcfUfuUfucucacs{invAb}	1865	{Phosphate}asUfsgUfaGfAfaaggGfaUfGaagcagsusu	1866
D-2369	{sGalNAc3K2AhxC6}[invAb]acaugCfuCfUfuUfucaccugasusu	1867	{Phosphate}usCfsaGfgUfGfaaagAfgCfaaugususu	1868
D-2370	{sGalNAc3K2AhxC6}cugcuucaUfgCfcfUfuUfucucacs{invAb}	1869	asUfsgUfaGfAfaaggCfaUfGaagcagsusu	1870
D-2371	{sGalNAc3K2AhxC6}cugcuucaUfgCfcfUfuUfucucacs{invAb}	1871	asUfsgUfaGfAfaaggCfaUfGaagcagsusu	1872
D-2372	{sGalNAc3K2AhxC6}cugcuucaUfgCfcfUfuUfucucacs{invAb}	1873	asUfsgUfaGfAfaaggCfaUfGaagcagsusu	1874
D-2373	{sGalNAc3K2AhxC6}cugcuucaUfgCfcfUfuUfucucacs{invAb}	1875	asUfsgUfaGfAfaaggCfaUfGaagcagsusu	1876
D-2374	{sGalNAc3K2AhxC6}cugcuucaUfgCfcfUfuUfucucacs{invAb}	1877	asUfsgUfaGfAfaaggCfaUfGaagcagsusu	1878
D-2375	{sGalNAc3K2AhxC6}cugcuucaUfgCfcfUfuUfucucacs{invAb}	1879	asUfsgUfaGfAfaaggCfaUfGaagcagsusu	1880
D-2376	{sGalNAc3K2AhxC6}cugcuucaUfgCfcfUfuUfucucacs{invAb}	1881	asUfsgUfaGfAfaaggCfaUfGaagcagsusu	1882

D-2377	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucuascsas{invAb}	1883	asUfsgUfaGfAfaaggCfaUfgaagcagsusu	1884
D-2378	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucuascsas{invAb}	1885	asUfsgUfaGfAfaaggCfaUfgaagcagsusu	1886
D-2379	{sGalNAc3K2AhxC6}csugcuucaUfgCfCfUfUfucuascsas{invAb}	1887	asUfsgUfaGfAfaaggCfaUfgaagcagsusu	1888
D-2380	{sGalNAc3K2AhxC6}csugcuucaUfgCfCfUfUfucuascsas{invAb}	1889	asUfsgUfaGfAfaaggCfaUfgaagcagsusu	1890
D-2381	{sGalNAc3K2AhxC6}[invAb]cuucauGfcCf[dT]UfUfucacagususu	1891	{Phosphate}asCfsuGfuAfGfaaagGfcAfugaagsusu	1892
D-2382	{sGalNAc3K2AhxC6}[invAb]cuucauGfcCfUfUfUfucacagususu	1893	{Phosphate}asCfsuGfuAfGfaaagGfcAfugaagsusu	1894
D-2383	{sGalNAc3K2AhxC6}[invAb]cuucauGfcCfUfUfUfucacagususu	1895	{Phosphate}asCfsuGfuAfGfaaagGfcAfugaagsusu	1896
D-2384	{sGalNAc3K2AhxC6}[invAb]cuucauGfcCfUfUfUfucacagususu	1897	{Phosphate}asCfsuGfuAfGfaaagGfcAfugaagsusu	1898
D-2385	{sGalNAc3K2AhxC6}[invAb]cuucauGfcCfUfUfUfucacagususu	1899	{Phosphate}asCfsuguaGfaaagGfcAfugaagsusu	1900
D-2386	{sGalNAc3K2AhxC6}[invAb]cuucauGfc[dC]UfUfUfucacagususu	1901	{Phosphate}asCfsuGfuAfGfaaagGfcAfugaagsusu	1902
D-2387	{sGalNAc3K2AhxC6}[invAb]cuucauGfcCfUf[dT]UfUfucacagususu	1903	{Phosphate}asCfsuGfuAfGfaaagGfcAfugaagsusu	1904
D-2388	{sGalNAc3K2AhxC6}[invAb]gcuucaUfgGfGfAfUfucacaususu	1905	{Phosphate}asUfsgUfaGfAfauccCfaUfgaagcsusu	1906
D-2389	{sGalNAc3K2AhxC6}[invAb]cuucauGfcGfAfAfUfucacagususu	1907	{Phosphate}asCfsuGfuAfGfaaagGfcAfugaagsusu	1908
D-2390	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucuascsas{invAb}	1909	{Phosphate}usUfsgUfaGfAfaaggCfaUfgaagcagsusu	1910
D-2391	{sGalNAc3K2AhxC6}[invAb]cugcuucaUfgCfCfUfUfucuascsasa	1911	{Phosphate}usUfsgUfaGfAfaaggCfaUfgaagcagsusu	1912
D-2392	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucuascsas{invDA}	1913	{Phosphate}usUfsgUfaGfAfaaggCfaUfgaagcagsusu	1914
D-2393	{sGalNAc3K2AhxC6}cugcuucaUfgGfGfAfUfucuascsas{invAb}	1915	{Phosphate}asUfsgUfaGfAfauccCfaUfgaagcagsusu	1916
D-2394	{sGalNAc3K2AhxC6}[invAb]cugcuucaUfgGfGfAfUfucuascsasu	1917	{Phosphate}asUfsgUfaGfAfauccCfaUfgaagcagsusu	1918
D-2395	{sGalNAc3K2AhxC6}[invAb]CfuUfcAfugGfcCfUfUfUfucAfcAfgUfsusUf	1919	{Phosphate}asCfsuGfuAfGfaaagGfcAfugaagsusu	1920
D-2396	{sGalNAc3K2AhxC6}[invAb]cuucauGfcCfUfUfUfucAfcAfgUfsusUf	1921	{Phosphate}asCfsuGfuAfGfaaagGfcAfugaagsusu	1922
D-2397	{sGalNAc3K2AhxC6}[invAb]gcuucaUfgCfCfUfUfUfucacaususu	1923	asUfsgUfaGfAfaaggCfaUfgaagcsusu	1924
D-2398	{sGalNAc3K2AhxC6}[invAb]ugcuucaUfgCfCfUfUfUfucacags{invAb}	1925	asCfsuGfuAfGfaaagGfcAfugaagcasusu	1926
D-2399	{sGalNAc3K2AhxC6}[invAb]cuucauGfcCfUfUfUfucacagususu	1927	{Phosphate}asCfsuGfuAfGfaaagGfcAfugaagsusu	1928
D-2400	{sGalNAc3K2AhxC6}ugcuucaUfgCfCfUfUfUfucAfcAfg{invAb}	1929	asCfsuGfuAfGfaaagGfcAfugaagcasusu	1930
D-2401	{sGalNAc3K2AhxC6}ugcuucaUfgCfCfUfUfUfucacags{invAb}	1931	asCfsuGfuAfGfaaagGfcAfugaagcasusu	1932
D-2402	{sGalNAc3K2AhxC6}ugcuucaUfgCfCfUfUfUfucacags{invAb}	1933	asCfsuguaGfaaagGfcAfugaagcasusu	1934
D-2403	{sGalNAc3K2AhxC6}[invAb]cuucauGfcCfUfUfUfucacagususu	1935	asCfsuGfuAfGfaaagGfcAfugaagsusu	1936
D-2404	{sGalNAc3K2AhxC6}ugcuucaUfgCfCfUfUfUfucacags{invAb}	1937	asCfsuGfuAfGfaaagGfcAfugaagcasusu	1938
D-2405	{GalNAc3K2AhxC6}augccuuCfuAfCfAfgGfcUfsusUf{invAb}	1939	{Phosphate}asGfscCfaCfUfguAfgAfaAfgGfcAfuGfasUfsu	1940
D-2406	{sGalNAc3K2AhxC6}ucaugccuUfuCfUfAfCfaguggcs{invAb}	1941	asGfscCfaCfUfguagAfaAfggcaugasusu	1942
D-2407	{sGalNAc3K2AhxC6}[invAb]augccuUfuCfUfAfCfaguggcsusu	1943	asGfscCfaCfUfguagAfaAfggcaugasusu	1944
D-2408	{sGalNAc3K2AhxC6}[invAb]augccuUfuCfUfAfCfagUfgGfcUfsusUf	1945	asGfscCfaCfUfguagAfaAfggcaugasusu	1946
D-2409	{sGalNAc3K2AhxC6}[invAb]ucaugccuUfuCfUfAfCfaguggcsusu	1947	asGfscCfaCfUfguagAfaAfggcaugasusu	1948

D-2410	{sGalNac3K2AhxC6}ucaugccuUfuCfUfAfCfagUfgGfcs{invAb}	1949	asGfscCfaCfUfguagAfaAfggcaugasusu	1950
D-2411	{sGalNac3K2AhxC6}[invAb]augccuUfuCfUfAfCfaguggcususu	1951	asGfscCfaCfUfguagAfaAfggcaususu	1952
D-2412	{sGalNac3K2AhxC6}ucaugccuUfuCfUfAfCfaguggcs{invAb}	1953	asGfscCfaCfUfguagAfaAfggcaugasusu	1954
D-2413	{sGalNac3K2AhxC6}ucaugccuUfuCfUfAfCfaguggcs{invAb}	1955	asGfscCfaCfUfguagAfaAfggcaugasusu	1956
D-2414	{sGalNac3K2AhxC6}[invAb]ucaugccuUfuCfUfAfCfaguggcs{invAb}	1957	asGfscCfaCfUfguagAfaAfggcaugasusu	1958
D-2415	{sGalNac3K2AhxC6}[invAb]augccuUfuCfUfAfCfaguggcasusu	1959	usGfscCfaCfUfguagAfaAfggcaususu	1960
D-2416	{sGalNac3K2AhxC6}ucaugccuUfuCfUfAfCfaguggcs{invAb}	1961	usGfscCfaCfUfguagAfaAfggcaugasusu	1962
D-2417	{sGalNac3K2AhxC6}[invAb]ucaugccuUfuCfUfAfCfaguggcsa	1963	usGfscCfaCfUfguagAfaAfggcaugasusu	1964
D-2418	{sGalNac3K2AhxC6}ucaugccuUfuCfUfAfCfaguggcs{invDA}	1965	usGfscCfaCfUfguagAfaAfggcaugasusu	1966
D-2419	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invAb}	1967	asUfsguagAfaaggCfaUfgaagcagsusu	1968
D-2420	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invAb}	1969	usUfsguagAfaaggCfaUfgaagcagsusu	1970
D-2421	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invDA}	1971	usUfsguagAfaaggCfaUfgaagcagsusu	1972
D-2422	{sGalNac3K2AhxC6}ucaugccuUfuCfUfAfCfaguggcs{invAb}	1973	usGfscCfaCfUfguagAfaAfggcaugasusu	1974
D-2423	{sGalNac3K2AhxC6}ucaugccuUfuCfUfAfCfaguggcs{invDA}	1975	usGfscCfaCfUfguagAfaAfggcaugasusu	1976
D-2424	{sGalNac3K2AhxC6}[invAb]cuucaUfgCfCfUfUfucucacas{invAb}	1977	{Phosphate}asCfsuGfuAfGfaaAfgGfcAfugaagsusu	1978
D-2425	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invAb}	1979	asUfsgUfa[Ab]AfaaggCfaUfgaagcagsusu	1980
D-2426	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invAb}	1981	asUfsgua[Ab]AfaaggCfaUfgaagcagsusu	1982
D-2427	{sGalNac3K2AhxC6}ugcuucaUfgCfCfUfUfucucacs{invAb}	1983	asCfsuGfu[Ab]GfaaagGfcAfugaagcasusu	1984
D-2428	{sGalNac3K2AhxC6}ugcuucaUfgCfCfUfUfucucacs{invAb}	1985	asCfsugu[Ab]GfaaagGfcAfugaagcasusu	1986
D-2429	{sGalNac3K2AhxC6}ucaugccuUfuCfUfAfCfaguggcs{invAb}	1987	asGfscCfa[Ab]UfguagAfaAfggcaugasusu	1988
D-2430	{sGalNac3K2AhxC6}ucaugccuUfuCfUfAfCfaguggcs{invAb}	1989	asGfscCfa[Ab]UfguagAfaAfggcaugasusu	1990
D-2431	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invAb}	1991	asUfs[GNA-G]uagAfaaggCfaUfgaagcagsusu	1992
D-2432	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invAb}	1993	asUfsg[GNA-U]agAfaaggCfaUfgaagcagsusu	1994
D-2433	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invAb}	1995	asUfsgu[GNA-A]gAfaaggCfaUfgaagcagsusu	1996
D-2434	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invAb}	1997	asUfsgua[GNA-G]AfaaggCfaUfgaagcagsusu	1998
D-2435	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invAb}	1999	asUfsguag[GNA-A]aaggCfaUfgaagcagsusu	2000
D-2436	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invAb}	2001	asUfsguagAf[GNA-A]aggCfaUfgaagcagsusu	2002
D-2437	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invAb}	2003	asUfsgUfagAfaaggCfaUfgaagcagsusu	2004
D-2438	{sGalNac3K2AhxC6}[invAb]cugcuucaUfgCfCfUfUfucucacasu	2005	asUfsguagAfaaggCfaUfgaagcagsusu	2006
D-2439	{sGalNac3K2AhxC6}[invAb]cugcuucaUfgCfCfUfUfucucacas{invAb}	2007	asUfsguagAfaaggCfaUfgaagcagsusu	2008
D-2440	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucucacas{invAb}	2009	asUfsguagAfaaGfgCfaUfgaagcagsusu	2010
D-2441	{sGalNac3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2011	asUfsguagAfaaggCfaUfgaagcagsusu	2012
D-2442	{sGalNac3K2AhxC6}[invAb]cugcuucaUfgCfCfUfUfucUfaCfasUf	2013	asUfsguagAfaaggCfaUfgaagcagsusu	2014

D-2443	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2015	asUfsguagAfaaGfgCfaUfgaagcagsusu	2016
D-2444	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2017	asUfsgUfaGfAfaaGfgCfaUfgaagcagsusu	2018
D-2445	{sGalNAc3K2AhxC6}[invAb]cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2019	asUfsgUfaGfAfaaGfgCfaUfgaagcagsusu	2020
D-2446	{sGalNAc3K2AhxC6}[invAb]cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2021	asUfsgUfaGfAfaaGfgCfaUfgaagcagsusu	2022
D-2447	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2023	as[Ab]guagAfaaggCfaUfgaagcagsusu	2024
D-2448	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2025	asUfs[Ab]uagAfaaggCfaUfgaagcagsusu	2026
D-2449	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2027	asUfsg[Ab]agAfaaggCfaUfgaagcagsusu	2028
D-2450	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2029	asUfsgu[Ab]gAfaaggCfaUfgaagcagsusu	2030
D-2451	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2031	asUfsguag[Ab]aaggCfaUfgaagcagsusu	2032
D-2452	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2033	asUfsguagAf[Ab]aggCfaUfgaagcagsusu	2034
D-2453	{sGalNAc3K2AhxC6}caacguacCfcUfUfCfAfuugaugs{invAb}	2035	asCfsaucaAfugaaGfgGfuacguugsusu	2036
D-2454	{sGalNAc3K2AhxC6}caacguacCfcUfUfCfAfuugaugs{invAb}	2037	asCfsaUfcAfAfugaaGfgGfuacguugsusu	2038
D-2455	{sGalNAc3K2AhxC6}acauggcuUfcCfAfGfAfuugaugs{invAb}	2039	asGfsgcaUfucugGfaAfGCCaugususu	2040
D-2456	{sGalNAc3K2AhxC6}acauggcuUfcCfAfGfAfuugaugs{invAb}	2041	asGfsgCfaUfAfucugGfaAfGCCaugususu	2042
D-2457	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2043	asUfsguaGfAfaaggCfaUfgaagcagsusu	2044
D-2458	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2045	asUfsguagAfaaggCfaUfgaagcagsusu	2046
D-2459	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2047	asUfsguagAfaaggCfaUfgaagcagsusu	2048
D-2460	{sGalNAc3K2AhxC6}cugcgguUfcCfUfGfGfgcuucus{invAb}	2049	usAfsgAfaGfCfccagGfaAfGCCcagsusu	2050
D-2461	{sGalNAc3K2AhxC6}cugcgguUfcCfUfGfGfgcuucus{invAb}	2051	usAfsgaagCfccagGfaAfGCCcagsusu	2052
D-2462	{sGalNAc3K2AhxC6}ugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2053	asCfsuGfuaGfaaagGfcAfugaagcasusu	2054
D-2463	{sGalNAc3K2AhxC6}[invAb]ugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2055	asCfsuGfuaGfaaagGfcAfugaagcasusu	2056
D-2464	{sGalNAc3K2AhxC6}[invAb]ugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2057	asCfsuGfuaGfaaagGfcAfugaagcasusu	2058
D-2465	{sGalNAc3K2AhxC6}ugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2059	asCfsuGfuaGfaaagGfcAfugaagcasusu	2060
D-2466	{sGalNAc3K2AhxC6}ugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2061	asCfsuGfuaGfaaagGfcAfugaagcasusu	2062
D-2467	{sGalNAc3K2AhxC6}ugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2063	asCfsuGfuAfGfaaagGfcAfugAfgCfasUfsu	2064
D-2468	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2065	asUfsguaGfaaAfggcaUfgAfagcagsusu	2066
D-2469	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2067	asUfsguaGfaaAfggcaUfgAfagcagsusu	2068
D-2470	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2069	asUfsguaGfaaAfggcaUfgAfagcagsusu	2070
D-2471	{sGalNAc3K2AhxC6}ugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2071	usCfsuguaGfaaagGfcAfugaagcasusu	2072
D-2472	{sGalNAc3K2AhxC6}ugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2073	usCfsuguaGfaaagGfcAfugaagcasusu	2074
D-2473	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2075	asUfsguagAfaaggCfaUfgaagcagsusu	2076
D-2474	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2077	asUfsguagAf[sGNA-A]aggCfaUfgaagcagsusu	2078
D-2475	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucUfaCfas{invAb}	2079	asUfsguagAfs[GNA-A]aggCfaUfgaagcagsusu	2080

D-2476	{sGalNAc3K2AhxC6}cugcuucaUfgCfCfUfUfucucacac{invAb}	2081	asUfsguagAfs[sGNA-A]aggCfaUfgaagcagsusu	2082
D-2477	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2083	asUfsguagAf[GNA-A]aggCfaUfgaagcagsusu	2084
D-2478	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2085	asUfsguagAf[GNA-A]AfggCfaUfgaagcagsusu	2086
D-2479	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2087	asUfsguaga[GNA-A]aggCfaUfgaagcagsusu	2088
D-2480	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2089	asUfsguagAf[GNA-A]aggCfaUfgaagcagsusu	2090
D-2481	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2091	asUfsguaga[GNA-A]aggCfaUfgaagcagsusu	2092
D-2482	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2093	asUfsguaga[GNA-A]AfggCfaUfgaagcagsusu	2094
D-2483	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2095	asUfsguagAf[GNA-A]AfggCfaUfgaagcagsusu	2096
D-2484	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2097	asUfsguaga[GNA-A]AfggCfaUfgaagcagsusu	2098
D-2485	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2099	asUfsgUfaGfAf[GNA-A]aggCfaUfgaagcagsusu	2100
D-2486	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2101	asUfsgUfagAf[GNA-A]aGfgCfaUfgaagcagsusu	2102
D-2487	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2103	asUfsgUfagAf[GNA-A]aggCfaUfgaagcagsusu	2104
D-2488	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2105	asUfsgUfaGfAf[GNA-A]aGfgCfaUfgaagcagsusu	2106
D-2489	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2107	asUfsguagAf[GNA-A]aGfgCfaUfgaagcagsusu	2108
D-2490	csusgcuucaUfgCfCfUfUfucUfaCfas{invAb}	2109	asUfsguagAf[GNA-A]aGfgCfaUfgaagcagsusu	2110
D-2491	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2111	asUfsguagAf[GNA-A]a[dG]gCfaUfgaagcagsusu	2112
D-2492	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2113	asUfsguagAf[GNA-A][dA]ggCfaUfgaagcagsusu	2114
D-2493	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2115	asUfsguagAf[GNA-A]ag[dG]CfaUfgaagcagsusu	2116
D-2494	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2117	asUfsguaGfAf[GNA-A]aggCfaUfgaagcagsusu	2118
D-2495	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2119	asUfsguaGfa[GNA-A]aggCfaUfgaagcagsusu	2120
D-2496	csusgcuucaUfgCfCfUfUfucucacac{invAb}	2121	asUfsguaGfa[GNA-A]AfggCfaUfgaagcagsusu	2122
D-2497	csusgcuuCaUfgGfCfcuuucacac{invAb}	2123	asUfsguaGfa[GNA-A]aggcaUfgafagcagsusu	2124
D-2498	csusgcuuCaUfgGfCfcuuucacac{invAb}	2125	asUfsguaGfa[GNA-A]AfggcaUfgafagcagsusu	2126
D-2499	csusgcuuCaUfgCfCfcuuucacac{invAb}	2127	asUfsguaga[GNA-A]aggCfaUfgafagcagsusu	2128
D-2500	csusgcuuCaUfgCfCfcuuucacac{invAb}	2129	asUfsguagaaaggCfaUfgafagcagsusu	2130
D-2501	usgscuucauGfcCfUfUfucucacags{invDA}	2131	us[sGNA-C]uguaGfaaagGfcAfugaagcasusu	2132
D-2502	usgscuucauGfcCfUfUfucucacags{invDA}	2133	usCfs[GNA-U]guaGfaaagGfcAfugaagcasusu	2134
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D-2504	usgscuucauGfcCfUfUfucucacags{invDA}	2137	usCfsugu[GNA-A]GfaaagGfcAfugaagcasusu	2138
D-2505	usgscuucauGfcCfUfUfucucacags{invDA}	2139	usCfsuguaGf[GNA-A]aagGfcAfugaagcasusu	2140
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D-2512	usgscuucauGfcCfUfUfUfcuacags{invDA}	2153	usCfsuguaGf[Ab]aagGfcAfugaagcasusu	2154
D-2513	uscсаugccuUfuCfUfAfCfagugges{invDT}	2155	asGfscacUfguagAfaAfggcaugasusu	2156
D-2514	uscсаugccuUfuCfUfAfCfagugges{invAb}	2157	usGfscacUfguagAfaAfggcaugasusu	2158
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D-2517	uscсugccuuCfaUfGfCfCfuucuas{invAb}	2163	usUfsagaaAfggcaUfgAfagcaggasusu	2164
D-2518	uscсugccuuCfaUfGfCfCfuucuas{invDA}	2165	usUfsagaaAfggcaUfgAfagcaggasusu	2166
D-2519	usasuguuccUfgCfUfUfCfaugccus{invDT}	2167	asAfsggcaUfgaagCfaGfgaacauasusu	2168
D-2520	usasuguuccUfgCfUfUfCfaugccus{invAb}	2169	usAfsggcaUfgaagCfaGfgaacauasusu	2170
D-2521	usasuguuccUfgCfUfUfCfaugccus{invDA}	2171	usAfsggcaUfgaagCfaGfgaacauasusu	2172
D-2522	{sGalNAc3K2AhxC6}uccugccuuCfaUfGfCfCfuucuas{invAb}	2173	asUfsagaaAfggcaUfgAfagcaggasusu	2174
D-2523	{sGalNAc3K2AhxC6}uauuguuccUfgCfUfUfCfaugccus{invAb}	2175	asAfsggcaUfgaagCfaGfgaacauasusu	2176
D-2524	{sGalNAc3K2AhxC6}ucaugccuUfuCfUfAfCfagugges{invDT}	2177	asGfscacUfguagAfaAfggcaugasusu	2178
D-2525	{sGalNAc3K2AhxC6}uccugccuuCfaUfGfCfCfuucuas{invDT}	2179	asUfsagaaAfggcaUfgAfagcaggasusu	2180
D-2526	{sGalNAc3K2AhxC6}uccugccuuCfaUfGfCfCfuucuas{invDA}	2181	usUfsagaaAfggcaUfgAfagcaggasusu	2182
D-2527	{sGalNAc3K2AhxC6}uauuguuccUfgCfUfUfCfaugccus{invDA}	2183	usAfsggcaUfgaagCfaGfgaacauasusu	2184
D-2528	usgscuucauGfcCfUfUfUfcuacags{invDA}	2185	usCfsugua[GNA-G]aaagGfcAfugaagcasusu	2186
D-2529	cugcuucaUfgCfCfUfUfsucuacacs{invAb}	2187	asUfsquagAf[GNA-A]aggCfaUfgaagcagsusu	2188
D-2530	cugcuucaUfgCfCfUfUfsucuacacs{invAb}	2189	asUfsquagAf[GNA-A]aggCfaUfgaagcagsusu	2190
D-2531	cugcuucaUfgCfCfUfUfsucuacacs{invAb}	2191	asUfsquagAf[GNA-A]aggCfaUfgaagcagsusu	2192
D-2532	cugcuucaUfgCfCfUfUfsucuacacs{invAb}	2193	asUfsquagAfs[GNA-A]aggCfaUfgaagcagsusu	2194
D-2533	cugcuucaUfgCfCfUfUfsucuacacs{invAb}	2195	asUfsquagAfs[GNA-A]aggCfaUfgaagcagsusu	2196
D-2534	cugcuucaUfgCfCfUfUfsucuacacs{invAb}	2197	asUfsquagAf[sGNA-A]aggCfaUfgaagcagsusu	2198
D-2535	cugcuucaUfgCfCfUfUfsucuacacs{invAb}	2199	asUfsquagAf[sGNA-A]aggCfaUfgaagcagsusu	2200
D-2536	cugcuucaUfgCfCfUfUfsucuacacs{invAb}	2201	asUfsquagAfs[sGNA-A]aggCfaUfgaagcagsusu	2202
D-2537	cugcuucaUfgCfCfUfUfsucuacacs{invAb}	2203	asUfsquagAfs[sGNA-A]aggCfaUfgaagcagsusu	2204
D-2538	cugcuucaUfgCfCfUf[LNA-T]ucuacacs{invAb}	2205	asUfsquagAf[GNA-A]aggCfaUfgaagcagsusu	2206
D-2539	cugcuucaUfgCfCf[LNA-T]Ufucuacacs{invAb}	2207	asUfsquagAf[GNA-A]aggCfaUfgaagcagsusu	2208
D-2540	cugcuucaUfgCfCfUfUf[LNA-T]cuacacs{invAb}	2209	asUfsquagAf[GNA-A]aggCfaUfgaagcagsusu	2210
D-2541	cugcuucaUfgCfCfUfUfuc[LNA-T]acacs{invAb}	2211	asUfsqu[GNA-A]gAfaaggCfaUfgaagcagsusu	2212

D-2542	cugcuucaUfgCfCfUfUfuc [LNA-A] cas {invAb}	2213	asUfsg [GNA-U] agAfaaggCfaUfgaagcagsusu	2214
D-2543	cugcuucaUfgCfCfUfUfucac [sLNA-A] {invAb}	2215	as [sGNA-U] guagAfaaggCfaUfgaagcagsusu	2216
D-2544	cugcuucaUfgCfCfUfUf [LNA-T] cuacas {invAb}	2217	asUfsguag [Ab] aaggCfaUfgaagcagsusu	2218
D-2545	cugcuucaUfgCfCfUfUfuc [LNA-T] acas {invAb}	2219	asUfsgu [Ab] gAfaaggCfaUfgaagcagsusu	2220
D-2546	cugcuucaUfgCfCfUfUfuc [LNA-A] cas {invAb}	2221	asUfsg [Ab] agAfaaggCfaUfgaagcagsusu	2222
D-2547	cugcuucaUfgCfCfUfUfucUfaCfas {invAb}	2223	asUfsgUfagAf [GNA-A] aggCfaUfgaagcagsusu	2224
D-2548	cugcuucaUfgCfCfUfUfucUfaCfas {invAb}	2225	asUfsguagAf [GNA-A] aggCfaUfgaagcagsusu	2226
D-2549	cugcuucaUfgCfCfUfs [LNA-T] ucuacas {invAb}	2227	asUfsguagAf [GNA-A] aggCfaUfgaagcagsusu	2228
D-2550	cugcuucaUfgCfCfUfs [sLNA-T] ucuacas {invAb}	2229	asUfsguagAf [GNA-A] aggCfaUfgaagcagsusu	2230
D-2551	cugcuucaUfgCfCfUf [LNA-T] ucuacas {invAb}	2231	asUfsguagAf [sGNA-A] aggCfaUfgaagcagsusu	2232
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D-2553	usgsuucauGfcCfUfUfUfcu [LNA-A] cags {invDA}	2235	usCfsug [Ab] aGfaaagGfcAfugaagcasusu	2236
D-2554	usgsuucauGfcCfUfUfUfc [LNA-T] acags {invDA}	2237	usCfsugu [Ab] GfaaagGfcAfugaagcasusu	2238
D-2555	usgsuucauGfcCfUfUfUf [LNA-T] cuacags {invDA}	2239	usCfsuguaGf [Ab] aagGfcAfugaagcasusu	2240
D-2556	usgsuucauGfcCfUfUfUfcuac [LNA-A] gs {invDA}	2241	usCfs [GNA-U] guaGfaaagGfcAfugaagcasusu	2242
D-2557	usgsuucauGfcCfUfUfUfcu [LNA-A] cags {invDA}	2243	usCfsug [GNA-U] aGfaaagGfcAfugaagcasusu	2244
D-2558	usgsuucauGfcCfUfUfUfc [LNA-T] acags {invDA}	2245	usCfsugu [GNA-A] GfaaagGfcAfugaagcasusu	2246
D-2559	usgsuucauGfcCfUfUfUfcuaca [sLNA-G] {invDA}	2247	us [Ab] uguaGfaaagGfcAfugaagcasusu	2248
D-2560	usgsuucauGfcCfUfUfUfcuac [LNA-A] gs {invDA}	2249	usCfs [Ab] guaGfaaagGfcAfugaagcasusu	2250
D-2561	ucaugccuUfuCfUfAfCfa [LNA-G] uggs {invAb}	2251	asGfscoc [Ab] UfguagAfaAfggcaugasusu	2252
D-2562	ucaugccuUfuCfUfAfCfag [LNA-T] ggs {invAb}	2253	asGfscoc [Ab] UfguagAfaAfggcaugasusu	2254
D-2563	ucaugccuUfuCfUfAfCfasgsggs {invAb}	2255	asGfscoc [Ab] UfguagAfaAfggcaugasusu	2256
D-2564	{GalNAc3K2AhxC6} GfgUfaUfgUfuCfCfUfGfcuUfcAfuUfsusUf	2349	{Phosphate} asAfsuGfaAfGfcaggAfaCfaUfaCfcsUfsu	2350
D-2565	{GalNAc3K2AhxC6} GfgUfaUfgUfuCfCfUfGfcuUfcAfuAfsusUf	2351	{Phosphate} usAfsuGfaAfGfcaggAfaCfaUfaCfcsUfsu	2352
D-2566	{GalNAc3K2AhxC6} GfuAfuGfuUfcCfUfGfCfuUfcAfuUfgUfsusUf	2353	{Phosphate} asCfsaUfgAfAfgcagGfaAfcafuAfcUfsu	2354
D-2567	{GalNAc3K2AhxC6} UfaUfgUfuCfCfUfGfCfUfucAfuGfcAfsusUf	2355	{Phosphate} usGfscAfuGfAfagcaGfgAfaCfaUfasUfsu	2356
D-2568	{GalNAc3K2AhxC6} AfuGfuUfcCfuGfCfUfUfcaUfgCfCfUfsusUf	2357	{Phosphate} asGfsgCfaUfGfaagCafGfaAfcAfsUfsu	2358
D-2569	{GalNAc3K2AhxC6} UfgUfuCfCfUfgCfUfUfCfauGfcCfuUfsusUf	2359	{Phosphate} asAfsGfCfUfgaagCfaGfgAfaCfasUfsu	2360
D-2570	{GalNAc3K2AhxC6} GfuUfcCfuGfCfUfUfCfAfugCfcUfuUfsusUf	2361	{Phosphate} asAfsaGfgCfAfugaaGfcAfgGfaAfcUfsu	2362
D-2571	{GalNAc3K2AhxC6} UfuCfCfUfgCfuUfCfAfUfgCfuUfuUfsusUf	2363	{Phosphate} asAfsaAfgGfCfaugaAfgCfaGfgAfasUfsu	2364
D-2572	{GalNAc3K2AhxC6} UfuCfCfUfgCfuUfCfAfUfgCfuUfuAfsusUf	2365	{Phosphate} usAfsaAfgGfCfaugaAfgCfaGfgAfasUfsu	2366
D-2573	{GalNAc3K2AhxC6} UfcCfuGfcUfuCfAfUfGfCfuUfuUfsusUf	2367	{Phosphate} asGfsaAfaGfGfcaugaAfaGfcaAfgGfasUfsu	2368
D-2574	{GalNAc3K2AhxC6} CfcUfgCfuUfcAfuUfGfCfuUfuCfuAfsusUf	2369	{Phosphate} usAfsGfaAfGfgcauGfaAfgCfaGfgsUfsu	2370

D-2575	{GalNac3K2AhxC6}CfuGfcUfuCfaUfGfCfCfuUfuCfaUfAfsusUf	2371	{Phosphate}asUfsaGfaAfAfggcaUfgAfagGfcAfgsUfsu	2372
D-2576	{GalNac3K2AhxC6}CfuGfcUfuCfaUfGfCfCfuUfuCfaUfAfsusUf	2373	{Phosphate}usUfsaGfaAfAfggcaUfgAfagGfcAfgsUfsu	2374
D-2577	{GalNac3K2AhxC6}UfgCfuUfuCfaUfGfCfCfUfuUfuCfaUfAfsusUf	2375	{Phosphate}usGfsuAfgAfAfaaggCfaUfgAfgCfasUfsu	2376
D-2578	{GalNac3K2AhxC6}GfcUfuCfaUfGfCfCfUfUfuUfuCfaUfAfsusUf	2377	{Phosphate}asUfsgUfaGfAfaaggCfaUfgAfaGfcsUfsu	2378
D-2579	{GalNac3K2AhxC6}GfcUfuCfaUfGfCfCfUfUfuUfuCfaUfAfsusUf	2379	{Phosphate}usUfsgUfaGfAfaaggCfaUfgAfaGfcsUfsu	2380
D-2580	{GalNac3K2AhxC6}CfuUfuCfaUfGfCfCfUfUfuUfuCfaUfAfsusUf	2381	{Phosphate}asCfsuGfuAfgGfaaaggGfcAfuGfaAfgsUfsu	2382
D-2581	{GalNac3K2AhxC6}UfuCfaUfGfCfCfUfUfUfCfuUfuCfaUfAfsusUf	2383	{Phosphate}asAfscUfgUfAfgaaaGfgCfaUfgAfafUfsu	2384
D-2582	{GalNac3K2AhxC6}UfuCfaUfGfCfCfUfUfUfCfuUfuCfaUfAfsusUf	2385	{Phosphate}usAfscUfgUfAfgaaaGfgCfaUfgAfafUfsu	2386
D-2583	{GalNac3K2AhxC6}UfuCfaUfGfCfCfUfUfUfCfuUfuCfaUfAfsusUf	2387	{Phosphate}asCfsaCfuGfUfagaaAfgGfcAfuGfasUfsu	2388
D-2584	{GalNac3K2AhxC6}UfuCfaUfGfCfCfUfUfUfCfuUfuCfaUfAfsusUf	2389	{Phosphate}usCfsaCfuGfUfagaaAfgGfcAfuGfasUfsu	2390
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D-2588	{GalNac3K2AhxC6}AfuGfcCfuUfuCfUfAfaGfuGfgUfUfsusUf	2397	{Phosphate}usGfscCfaCfUfguagAfaAfgGfcAfuUfsu	2398
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D-2592	{GalNac3K2AhxC6}GfuAfuGfuUfuCfUfGfCfuUfuCfaUfAfsusUf	2405	{Phosphate}gsGfsaUfgAfAfgcagGfaAfaUfaAfcUfsu	2406
D-2593	{GalNac3K2AhxC6}UfaUfgUfuCfCfUfGfCfUfuUfuCfaUfAfsusUf	2407	{Phosphate}gsGfsgAfuGfAfaagGfaAfaUfaUfsu	2408
D-2594	{GalNac3K2AhxC6}AfuGfuUfuCfuGfCfUfUfCfaUfuCfCfUfsusUf	2409	{Phosphate}gsGfsgGfaUfgfaagGfaAfaUfaUfsu	2410
D-2595	{GalNac3K2AhxC6}UfgUfuCfCfUfgCfUfUfCfaUfuCfCfUfsusUf	2411	{Phosphate}asGfsgGfgAfuGfaagGfaAfaUfaUfsu	2412
D-2596	{GalNac3K2AhxC6}GfuUfuCfuGfCfUfUfCfaUfuCfCfUfsusUf	2413	{Phosphate}asAfsGfgGfAfuGfaagGfaAfaUfaUfsu	2414
D-2597	{GalNac3K2AhxC6}UfuCfCfUfgCfuUfCfAfuUfuCfuCfUfsusUf	2415	{Phosphate}gsAfsaGfgGfgfaugaAfgCfaGfgAfafUfsu	2416
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D-2600	{GalNac3K2AhxC6}CfuGfcUfuCfaUfCfCfCfCfuUfuCfaUfAfsusUf	2421	{Phosphate}gsUfsaGfaAfgfgggaUfgAfaGfcAfgsUfsu	2422
D-2601	{GalNac3K2AhxC6}UfgCfuUfuCfaUfCfCfCfCfuUfuCfaUfAfsusUf	2423	{Phosphate}usGfsuAfgAfAfggggAfuGfaAfgCfasUfsu	2424
D-2602	{GalNac3K2AhxC6}GfcUfuCfaUfuCfCfCfCfuUfuCfaUfAfsusUf	2425	{Phosphate}csUfsgUfaGfAfaaggGfaUfgAfaGfcsUfsu	2426
D-2603	{GalNac3K2AhxC6}CfuUfuCfaUfuCfCfCfCfuUfuCfaUfAfsusUf	2427	{Phosphate}asCfsuGfuAfgfaaggGfgAfuGfaAfgsUfsu	2428
D-2604	{GalNac3K2AhxC6}UfuCfaUfuCfCfCfCfuUfuCfaUfAfsusUf	2429	{Phosphate}csAfsUfgUfAfgaagGfgGfaUfgAfafUfsu	2430
D-2605	{GalNac3K2AhxC6}UfuCfaUfuCfCfCfCfuUfuCfaUfAfsusUf	2431	{Phosphate}csCfsaCfuGfUfagaaGfgGfgAfuGfasUfsu	2432
D-2606	{GalNac3K2AhxC6}CfaUfuCfCfCfuUfCfUfAfaGfuGfgCfUfsusUf	2433	{Phosphate}gsCfscAfcUfGfuagaAfgGfgGfaUfgsUfsu	2434
D-2607	{GalNac3K2AhxC6}AfuCfCfCfuUfCfUfAfaGfuGfgCfUfsusUf	2435	{Phosphate}gsGfscCfaCfUfguagAfaGfgGfgAfuUfsu	2436

D-2608	{GalNAc3K2AhxC6}UfcCfcCfuUfcUfAfCfAfGuGfgCfcUfsusUf	2437	{Phosphate}asGfsgCfcAfCfuguaGfaAfGfgGfasUfsu	2438
D-2609	{GalNAc3K2AhxC6}CfcCfcUfuCfuAfCfAfGfugGfcCfuUfsusUf	2439	{Phosphate}asAfsgGfcCfAfcuguAfgAfGfgGfgsUfsu	2440

EXAMPLE 2: Efficacy of select PNPLA3 siRNA molecules in RNA FISH assay

[0003] A panel of fully chemically modified siRNA, including siRNA spanning the rs738409 and/or the rs738408 SNP in PNPLA3, were prepared and tested for potency and selectivity of mRNA knockdown in vitro. Each siRNA duplex consisted of two strands, the sense or 'passenger' strand and the antisense or 'guide' strand. The strands are 21 or 23 nucleotides in length with 19 complementary base pairs. In some instances, there are two base pair 3' overhangs. The siRNA were prepared with substitution of the natural 2'-OH in the ribose of each nucleotide with either a 2'-OMe or 2'-F group. Optionally, phosphodiester internucleotide linkages at one or both strands were replaced with phosphorothioates to reduce degradation by exonucleases.

[0004] The efficacy of each of the siRNA molecules in reducing PNPLA3 expression was assessed using a 384-well format in vitro siRNA transfection assay followed by a fluorescent in situ hybridization targeting ribonucleic acid molecules (RNA FISH) assay to determine IC₅₀ and maximum activity values. This assay was performed on human hepatocellular carcinoma cell line Hep3B cells (ATCC HB-8064) and Chinese hamster ovary (CHO) cells expressing human PNPLA3 I148I. Human hepatocellular carcinoma HepB3 was maintained in EMEM media (ATCC 30-2003) supplemented with 10% fetal bovine serum and 1% antibiotic/antimycotic at 37°C and 5% CO₂. CHO cells expressing human PNPLA3 I148I was maintained in media containing 50% CD-CHO (Life Technologies), 50% Ex-Cell CHO 5 Medium (Sigma), 8mM L-Glutamine, 1 x HT, 1% antibiotic/antimycotic, and 10µg/mL Puromycin at 37°C and 5% CO₂.

[0005] For Hep3B cell assays, transfection complexes of the siRNA molecules and the Lipofectamine RNAiMAX transfection reagent (Life Technologies) in EMEM media (ATCC 30-2003) were prepared in 384-well plates (PerkinElmer), at 10µg per well, in accordance with manufacturer's recommendations. For CHO human cell assays, transfection complexes of the siRNA molecules and the Lipofectamine RNAiMAX transfection reagent in F12K media (Mediatech) were prepared in 384-well plates, at 10µg per well, in accordance with manufacturer's recommendations. Cells were diluted to 67,000 cells/ml in antibiotic/antimycotic-free media and

30µl added to each well, a final density of 2000 cells/well in 40µl media. After a 20 minute incubation at room temperature, plates were transferred to a 37°C and 5% CO₂ incubator. Hep3B cell transfection assays were incubated for 72 hours and CHO human PNPLA3 I148I transfection assays were incubated for 48 hours.

[0184] At harvest, the cells were fixed in an 8% formaldehyde fixative solution (Thermo Scientific) for 15 minutes at room temperature. The plates were then subjected to dehydration with sequential 50%, 70%, and 100% ethanol washes. Plates were then sealed and stored at -20°C.

[0185] The RNA FISH assay was performed using the Affymetrix QuantiGene® View RNA HC Screening Assay kit (QVP0011), the Affymetrix View HC Signal Amplification Kit 3-plex (QVP0213), and Affymetrix gene specific probes: PNPLA3 Human 0.33mL View RNA Type 6 (650 label) VA6-20279-01 and PPIB Human 0.44mL View RNA Type 1 (488 label) VA1-10148-01.

[0186] Plates were first rehydrated with sequential 100%, 70%, and 50% ethanol washes. Cells were then washed with PBS, and then permeabilized and protease-digested according to the kit instructions. The target Working Probe Sets were prepared according to the manufacturer's protocol, added to the wells, and incubated for 3 hours at 40°C. The manufacturer's protocol was followed for the sequential hybridizations with the Working Probe Sets, the Working PreAmps, the Working Amps, and the Working LPs. Last, nuclei counterstains were applied (Hoechst 33342 and Cell Mask Blue; Molecular Probes). Plates were incubated for 30 minutes at room temperature, washed with PBS, overlaid with 80µl of PBS, and then the plates were sealed for imaging.

[0187] All plates were imaged on an Opera Phenix High Content Screening System (PerkinElmer), using the UV Channel for Hoechst 33342 and Cell Mask Blue, the 488 Channel for Type1 probes, and the 647 Channel for Type6 probes.

[0188] RNA FISH data was analyzed using Columbus software and images were generated using Genedata Screener. The results of the assay for CHO transfected PNPLA3 I148I are shown in Table 3. The results of the assay for CHO transfected PNPLA3 I148M are shown in Table 4.

PNPLA3 knockdown provides a percentage of knockdown compared to control. Negative values indicate a decrease in PNPLA3 levels.

Table 3. RNA FISH assay on CHO transfected PNPLA3 I148I

Duplex No.	IC50 (μ M)	PNPLA3 knockdown (%)
D-2001	.0589	-33.9
D-2002	.0158	-67.2
D-2003	.0427	-43.4
D-2004	.00835	-63.5
D-2006	.0177	-77.8
D-2008	.125	-10.7
D-2009	.00769	-45.5
D-2010	.00558	-80
D-2011	.035	-3
D-2012	> 0.5	-3.7
D-2013	.036	-6.4
D-2014	.0122	-58.2
D-2015	> 0.5	8
D-2016	> 0.5	8
D-2017	.0153	-73.2
D-2018	.00386	-31.5
D-2019	.125	
D-2020	> 0.5	
D-2021	> 0.5	8
D-2022	> 0.167	-6.8
D-2023	.0257	-36.9
D-2024	> 0.5	4
D-2025	> 0.5	-2.5
D-2026	.022	-35.1
D-2027	.00172	-16.5
D-2028	> 0.5	10
D-2029	.0106	-56.2
D-2032	.00205	-52.9
D-2033	.0107	-61.7
D-2034	> 0.5	6
D-2035	> 0.5	-3.8
D-2036	.00665	-55.9
D-2037	> 0.5	4

D-2038	> 0.5	10
D-2039	.0116	-23.9
D-2040	> 0.5	-25.4
D-2041	> 0.5	9
D-2042	.00959	-25.5
D-2043	> 0.5	9
D-2044	.00552	-29
D-2045	> 0.5	9
D-2046	> 0.5	9
D-2047	> 0.5	-5.9
D-2048	.00618	-56.3
D-2049	> 0.5	12
D-2050	> 0.5	10
D-2051	> 0.5	-17.2
D-2052	> 0.5	-8.3
D-2053	> 0.5	11
D-2054	> 0.5	-14.9
D-2055	> 0.5	-10.6
D-2056	> 0.5	10
D-2057	.00485	-59.2
D-2058	.014	-53
D-2059	> 0.5	-4.9
D-2060	> 0.5	18
D-2061	.00795	-44.8
D-2062	.000668	-74.6
D-2063	> 0.5	-21.8
D-2064	> 0.5	9
D-2065	> 0.5	-10.5
D-2066	.0412	-42.2
D-2067	> 0.5	10

Table 4. RNA FISH assay on CHO transfected PNPLA3 I148M

Duplex No.	IC50 (μ M)	PNPLA3 knockdown (%)
D-2000	.0316	-29.2
D-2001	.0131	-81.8
D-2002	.00216	-90.5
D-2003	.022	-50.4

D-2004	.00429	-88
D-2005	> 0.5	15
D-2006	.00301	-89.2
D-2007	> 0.5	6
D-2009	.00274	-86.9
D-2010	.00203	-93.3
D-2011	.000694	-11.9
D-2012	> 0.5	-18
D-2013	.011	-66.4
D-2014	.0057	-84.3
D-2015	> 0.5	-13.5
D-2016	> 0.5	-12.4
D-2017	.00448	-89.4
D-2018	.0104	-36.2
D-2019	.0302	-7.7
D-2020	.01	-78.9
D-2021	.00435	-83.5
D-2022	.00628	-88.6
D-2023	.0143	-44.3
D-2024	> 0.5	11
D-2025	.00355	-58.2
D-2026	.000867	-39.4
D-2027	> 0.5	32
D-2028	.00205	-89.9
D-2029	.0019	-94
D-2030	> 0.5	-9.4
D-2031	> 0.5	4

RNA FISH was also run on a hepatic cell line containing the double mutant PNPLA3-rs738408-rs738409, as well as a control wild-type cell line, Hep3B. Hep3B and HepG2 cells (purchased from ATCC) were cultured in minimal essential medium (MEM from Corning for Hep3B and EMEM from ATCC for HepG2) supplemented with 10% fetal bovine serum (FBS, Sigma) and 1% penicillin-streptomycin (P-S, Corning). The siRNA transfection was performed as follows: 1 μ L of test siRNAs and 4 μ L of plain MEM or EMEM, depending on the cell line, were added to PDL-coated CellCarrier-384 Ultra assay plates (PerkinElmer) by BioMek FX (Beckman Coulter). 5 μ L of Lipofectamine RNAiMAX (Thermo Fisher Scientific), pre-diluted in plain MEM or

EMEM (specifically 0.035 μ L RNAiMAX in 5 μ L MEM for Hep3B and 0.06 μ L of RNAiMAX in 5 μ L EMEM for HepG2), was then dispensed into the assay plates by Multidrop Combi Reagent Dispenser (Thermo Fisher Scientific). After 20 mins incubation of the siRNA/RNAiMAX mixture at room temperature (RT), 30 μ L of either Hep3B or HepG2 cells (2000 cells per well) in MEM or EMEM supplemented with 10% FBS and 1% P-S were added to the transfection complex using Multidrop Combi Reagent Dispenser. The assay plates were incubated for 20 mins at RT prior to being placed in an incubator. Cells were then incubated for 72 hrs at 37 °C and 5% CO₂. ViewRNA ISH Cell Assay was performed following manufacture's protocol (Thermo Fisher Scientific) using an in-house assembled automated FISH assay platform for liquid handling. In brief, cells were fixed in 4% formaldehyde (Thermo Fisher Scientific) for 15 mins at RT, permeabilized with detergent for 3 mins at RT and then treated with protease solution for 10 mins at RT. Incubation of target-specific probe pairs (Thermo Fisher Scientific) was done for 3 hrs, while for Preamplifiers, Amplifiers and Label Probes (Thermo Fisher Scientific) were for 1 hr each. All hybridization steps were carried out at 40 °C in Cytomat 2 C-LIN automated incubator (Thermo Fisher Scientific). After hybridization reactions, cells were stained for 30 mins with Hoechst and CellMask Blue (Thermo Fisher Scientific) and then imaged on Opera Phenix (PerkinElmer). The images were analyzed using Columbus Image Data Storage and Analysis System (PerkinElmer) to obtain mean spot counts per cell. The spot counts were normalized using the high (containing phosphate-buffered saline, Corning) and low (without target probe pairs) control wells. The normalized values against the total siRNA concentrations were plotted and the data were fit to a four-parameter sigmoidal model in Genedata Screener (Genedata) to obtain IC₅₀ and maximum activity. The results of the assay for HepG2 cells is shown in Table 5 and the results of the assay for Hep3B cells is shown in Table 6. PNPLA3 knockdown provides a percentage of knockdown compared to control. Negative values indicate a decrease in PNPLA3 levels. In instances where a duplex was run more than once, the average IC₅₀ is shown, with a standard deviation.

Table 5. RNA FISH assay on hepatic HepG2 cells

Duplex No.	IC ₅₀ (μM)	PNPLA3 knockdown (%)
D-2001	.0132	-70.5
D-2003	.0458	-46.9
D-2004	.0049	-74.2
D-2006	.00283	-69.6
D-2009	.00448	-62.2
D-2010	.00206	-52
D-2013	.00319	-71.7
D-2014	.00164	-67.4
D-2017	.00222	-60.9
D-2018	.00717	-52.7
D-2020	.00664	-68.3
D-2021	.00559	-61.5
D-2022	.004	-30.5
D-2023	> 0.5	-18
D-2025	.0041	-61.9
D-2026	.00937	-34.6
D-2564	0.009	-64.021
D-2565	0.00609	-70.965
D-2566	0.00255	-52.65
D-2567	0.00584	-56.603
D-2568	0.0157	-63.002
D-2569	0.00327	-64.898
D-2570	0.00144	-55.424
D-2571	> 0.1	-18.661
D-2572	0.00557	-25.668
D-2573	0.0115	-55.329
D-2574	0.00289	-69.84
D-2575	0.00378	-69.491
D-2576	0.00527	-64.841
D-2577	0.00511	-46.449
D-2578	0.0026	-67.821
D-2579	0.00402	-67.057
D-2580	0.00119	-64.422
D-2581	0.00915	-73.008
D-2582	0.000823	-77.053
D-2583	0.00851	-66.555
D-2584	0.00513	-54.442

D-2585	0.0154	-67.707
D-2586	0.00701	-69.624
D-2587	0.00732	-66.627
D-2588	0.00226	-70.854
D-2589	0.00837	-31.221
D-2590	0.0249	-41.857
D-2591	0.009	-64.021
D-2592	0.00609	-70.965
D-2593	0.00255	-52.65
D-2594	0.00584	-56.603
D-2595	0.0157	-63.002
D-2596	0.00327	-64.898
D-2597	0.00144	-55.424
D-2598	> 0.1	-18.661
D-2599	0.00557	-25.668
D-2600	0.0115	-55.329
D-2601	0.00289	-69.84
D-2602	0.00378	-69.491
D-2603	0.00527	-64.841
D-2604	0.00511	-46.449
D-2605	0.0026	-67.821
D-2606	0.00402	-67.057
D-2607	0.00119	-64.422
D-2608	0.00915	-73.008
D-2609	0.000823	-77.053
D-2591	0.00851	-66.555
D-2592	0.00513	-54.442
D-2593	0.0154	-67.707
D-2594	0.00701	-69.624
D-2595	0.00732	-66.627
D-2596	0.00226	-70.854
D-2597	0.00837	-31.221
D-2598	0.0249	-41.857
D-2426	85.098	-14.902
D-2444	31.575 +/- 6.79	-68.425 +/- 6.79
D-2454	53.215	-46.785
D-2473	29.35	-70.65

Table 6. RNA FISH assay on hepatic Hep3B cells

Duplex No.	IC ₅₀ (μM)	PNPLA3 knockdown (%)
D-2001	.00842	-37
D-2003	.0158	-32.1
D-2004	.00266	-32.4
D-2006	.00948	-54.1
D-2009	.00228	-29.5
D-2010	.00219	-37.2
D-2013	.00524	-31.5
D-2014	.00148	-37.6
D-2017	.00333	-37.6
D-2018	.00315	-21.3
D-2020	> 0.5	6
D-2021	> 0.5	-1.6
D-2022	.00272	-30.9
D-2023	> 0.5	24
D-2025	.0101	-30.3
D-2026	.00551	-23
D-2564	0.01	-63.199
D-2565	0.00938	-58.392
D-2566	0.00343	-61.484
D-2567	0.0175	-53.489
D-2568	> 0.1	-18.367
D-2569	0.0195	-62.568
D-2570	0.0127	-77.141
D-2571	> 0.1	-15.922
D-2572	> 0.1	-12.434
D-2573	> 0.1	-14.649
D-2574	0.0215	-52.515
D-2575	0.0203	-53.175
D-2576	0.018	-48.137
D-2577	> 0.1	-16.105
D-2578	> 0.1	-21.309
D-2579	> 0.1	-17.510
D-2580	> 0.1	-24.616
D-2581	> 0.1	-13.987
D-2582	0.0574	-30.543

D-2583	> 0.1	-23.990
D-2584	> 0.1	-6.715
D-2585	> 0.1	-17.518
D-2586	> 0.1	-24.518
D-2587	0.0391	-58.478
D-2588	0.0218	-56.609
D-2589	> 0.1	-17.418
D-2590	> 0.1	-21.161
D-2591	0.0167	-59.366
D-2592	0.0104	-61.548
D-2593	Undefined	-61.879
D-2594	0.0211	-43.856
D-2595	0.0272	-63.020
D-2596	> 0.1	-10.278
D-2597	0.0546	-31.743
D-2598	Undefined	-47.517
D-2599	0.00489	-70.825
D-2600	> 0.1	-8.522
D-2601	0.0364	-31.836
D-2602	0.00577	-65.062
D-2603	0.01	-58.287
D-2604	0.00353	-40.649
D-2605	0.0113	-50.691
D-2606	> 0.1	-5.097
D-2607	0.0261	-49.898
D-2608	> 0.1	-23.747
D-2609	> 0.1	-23.804
D-2426	> 0.1	-15.207
D-2454	0.00187	-51.735
D-2473	> 0.1	-21.333

EXAMPLE 3: Droplet digital PCR assay of siRNA for PNPLA3-rs738409 and PNPLA3-rs738409-rs738408

[0189] Following the manufacturers protocol, thawed human primary hepatocyte cells (Xenotech/Sekisui donor lot#HC3-38) in OptiThaw media (Xenotech cat#K8000), cells were centrifuged and post media aspiration, resuspended in OptiPlate hepatocyte media (Xenotech cat#K8200) and plated into 96 well collagen coated plates (Greiner cat#655950). Following a 2-

4 hour incubation period, media was removed and replaced with OptiCulture hepatocyte media (Xenotech cat#K8300). 2-4 hours post addition of OptiCulture media, delivered GalNAc conjugated siRNAs to cells via free uptake (no transfection reagent). Cells were incubated 24-72 hours at 37°C and 5% CO₂. Cells were then lysed with Qiagen RLT buffer (79216) +1% 2-mercaptoethanol (Sigma, M-3148), and lysates were stored at -20°C. RNA was purified using a Qiagen QIAcube HT instrument (9001793) and a Qiagen RNeasy 96 QIAcube HT Kit (74171) according to manufacturer's instructions. Samples were analyzed using a QIAxpert system (9002340). cDNA was synthesized from RNA samples using the Applied Biosystems High Capacity cDNA Reverse Transcription kit (4368813), reactions were assembled according to manufacturer's instructions, input RNA concentration varied by sample. Reverse transcription was carried out on a BioRad tetrad thermal cycler (model# PTC-0240G) under the following conditions: 25°C 10 minutes, 37°C 120 minutes, 85°C 5 minutes followed by (an optional) 4°C infinite hold.

[0190] Droplet digital PCR (ddPCR) was performed using BioRad's QX200 AutoDG droplet digital PCR system according to manufacturer's instructions. Reactions were assembled into an Eppendorf clear 96 well PCR plate (951020303) using BioRad ddPCR Supermix for Probes (1863010), and fluorescently labeled qPCR assays for PNPLA3 (IDT Hs.PT.58.21464637, primer to probe ratio 3.6:1 and TBP (IDT Hs.PT.53a.20105486, primer to probe ratio 3.6:1) and RNase free water (Ambion, AM9937). Final primer/probe concentration is 900nM/250nM respectively, input cDNA concentration varied among wells. Droplets were formed using a BioRad Auto DG droplet generator (1864101) set up with manufacturer recommended consumables (BioRad DG32 cartridges 1864108, BioRad tips 1864121, Eppendorf blue 96 well PCR plate 951020362, BioRad droplet generation oil for probes 1864110 and a BioRad droplet plate assembly). Droplets were amplified on a BioRad C1000 touch thermal cycler (1851197) using the following conditions: enzyme activation 95°C 10 minutes, denaturation 94°C 30 seconds followed by annealing/extension 60°C for one minute, 40 cycles using a 2°C/second ramp rate, enzyme deactivation 98°C 10 minutes followed by (an optional) 4°C infinite hold. Samples were then read on a BioRad QX200 Droplet Reader measuring FAM/HEX signal that correlates to PNPLA3 or TBP concentration. Data was analyzed using BioRad's QuantaSoft software package. Samples are gated by channel (fluorescent label) to determine the concentration per sample. Each sample

is then expressed as the ratio of the concentration of the gene of interest (PNPLA3)/concentration of the housekeeping gene (TBP) to control for differences in sample loading. Data is then imported into Genedata Screener, where each test siRNA is normalized to the median of the neutral control wells (buffer only). IC50 values are reported in Table 7.

Table 7. ddPCR assay on primary hepatocyte cells

Duplex No.	IC50 (μ M)	% PNPLA3 knockdown
D-2068	.0339	-49.628
D-2069	.00408	-52.997
D-2070	.00433	-42.193
D-2072	.00884	-53.16
D-2073	> 2.0	-7.435
D-2078	.0044	-43.123
D-2084	0.00499	-38.791
D-2085	0.00539	-64.312
D-2086	> 2.0	-14.938
D-2087	> 2.0	-25.465
D-2088	0.207	-34.944
D-2089	0.0107	-38.791
D-2090	0.0218	-38.977
D-2091	0.0508	-41.209
D-2092	0.00192	-44
D-2093	0.00634	-30.233
D-2094	> 2.0	4.93
D-2095	0.00181	-59.814
D-2096	0.0181	-52.807
D-2099	0.00549	-39.296
D-2100	0.0142	-55.281
D-2158	0.0681	-48.649
D-2159	0.0325	-36.036
D-2160	> 0.667	-13.514
D-2161	> 2.0	-24.229
D-2162	0.0726	-28.634
D-2163	> 0.667	-16.3
D-2164	> 2.0	-15.418
D-2165	0.00644	-26.872
D-2166	0.00192	-30.045

D-2167	> 0.667	-6.726
D-2168	> 2.0	-15.418
D-2169	> 2.0	-13.004
D-2170	> 2.0	-9.417
D-2171	0.00505	-44.395
D-2172	0.003	-55.336
D-2173	0.00598	-46.188
D-2174	> 2.0	-9.009
D-2175	0.017	-27.928
D-2176	0.00452	-35.426
D-2177	> 2.0	4.5
D-2178	> 2.0	1.8
D-2179	> 2.0	-6.306
D-2180	0.00546	-40.969
D-2181	0.00152	-43.119
D-2182	0.00317	-54.128
D-2183	0.00948	-58.079
D-2184	0.0109	-50.459
D-2185	> 2.0	-7.339
D-2186	0.0021	-48.624
D-2187	> 2.0	-11.009
D-2188	> 2.0	1.32
D-2191	0.0984	-66.923
D-2192	0.124	-64.231
D-2193	0.138	-60.606
D-2194	0.0478	-54.182
D-2195	0.0801	-47.515
D-2199	0.0517	-62.973
D-2201	0.0517	-62.973
D-2202	0.0165	-72.404
D-2203	0.00946	-49.459
D-2204	0.0241	-58.545
D-2205	0.0382	-45.576
D-2206	0.0222	-50.946
D-2209	0.0867	-46.622
D-2212	0.358	-60
D-2216	0.0826	-58.942
D-2218	0.0242	-63.462
D-2220	0.0113	-69.333
D-2221	0.138	-55.541

D-2224	0.0198	-66.486
D-2225	0.245	-68.077
D-2228	0.155	-44.606
D-2229	0.0651	-38.909
D-2231	0.0512	-56.892
D-2232	0.0678	-67.981
D-2233	0.00802	-57.182
D-2234	0.00473	-55.947
D-2235	0.00816	-62.115
D-2236	0.00245	-51.542
D-2237	0.00495	-60
D-2238	0.00561	-63.017
D-2239	0.00453	-55.537
D-2240	0.00584	-56.116
D-2241	0.00755	-54.76
D-2242	0.0137	-56.332
D-2243	0.00329	-57.118
D-2244	0.0127	-56.909
D-2245	0.00697	-58.364
D-2246	0.00713	-56.828
D-2247	0.00875	-57.797
D-2248	0.0098	-58.59
D-2249	0.00603	-57.759
D-2250	0.0105	-62.155
D-2251	0.00521	-59.914
D-2252	0.00988	-58.678
D-2253	0.00481	-57.118
D-2254	0.00721	-56.332
D-2255	0.00788	-52.838
D-2256	0.00831	-55.455
D-2257	0.00503	-54.545
D-2258	0.00626	-54.545
D-2259	0.00401	-55.947
D-2260	0.00379	-52.423
D-2261	0.00151	-54.31
D-2262	0.00292	-53.448
D-2263	0.00607	-59.483
D-2264	0.00703	-59.504
D-2265	> 4.0	-20.524
D-2266	0.0129	-32.727

D-2267	0.107	-27.273
D-2275	0.00359	-45.701
D-2276	0.00416	-43.891
D-2277	0.00218	-49.14
D-2278	0.00743	-42.986
D-2279	0.0116	-53.846
D-2280	0.00347	-36.652
D-2281	0.00449	-43.891
D-2282	0.0134	-37.557
D-2283	0.00864	-34.842
D-2284	0.00738	-49.558
D-2285	0.0202	-38.053
D-2286	0.00543	-45.487
D-2287	0.00934	-47.611
D-2288	0.00652	-55.575
D-2289	0.0259	-61.593
D-2290	0.00549	-53.805
D-2291	0.00476	-51.062
D-2292	0.0105	-42.584
D-2293	0.0059	-45.455
D-2294	0.0117	-45.646
D-2295	0.0109	-52.823
D-2296	0.01246 +/- 0.015	-59.847 +/- 15.2
D-2297	0.00828	-44.444
D-2298	0.0279	-41.346
D-2299	0.00529	-55.926
D-2300	0.0195	-50.423
D-2301	0.00838	-35.577
D-2302	0.00832	-40.865
D-2303	0.00371	-40.096
D-2304	0.00563	-38.365
D-2305	0.00639	-40.385
D-2306	0.00669	-41.25
D-2307	0.00212	-38.462
D-2308	0.00573	-37.736
D-2309	0.0645	-33.962
D-2310	0.00232	-33.019
D-2311	0.00181	-31.132
D-2312	0.0447	-45.283
D-2313	0.00655	-42.453

D-2314	0.00613	-41.509
D-2315	0.00941	-50
D-2316	0.0218	-41.784
D-2317	0.0142	-42.723
D-2318	0.0182	-31.455
D-2319	> 4.0	-23.005
D-2320	0.0228	-37.089
D-2321	0.00809	-45.352
D-2322	0.0165	-44.601
D-2323	0.0184	-38.373
D-2324	0.00766	-41.627
D-2325	0.00815	-46.507
D-2326	0.0168	-48.325
D-2327	0.00663	-62.254
D-2328	0.00716	-39.367
D-2329	0.044	-52.036
D-2330	0.00282	-65.701
D-2331	0.00411	-53.991
D-2332	0.0127	-51.222
D-2333	0.012	-48.357
D-2334	0.019	-42.593
D-2335	0.00448	-47.418
D-2336	0.00944	-37.327
D-2337	0.00514	-37.327
D-2338	0.154	-38.249
D-2339	0.0089	-40.092
D-2340	0.0169	-48.148
D-2341	0.00274	-46.296
D-2342	0.0225	-42.723
D-2343	0.00222 +/- 0.00136	-43.218 +/- 4.81
D-2344	0.0136	-56.561
D-2345	0.0222	-50.226
D-2346	0.0273	-55.385
D-2347	0.0164	-41.784
D-2348	0.0314	-60.282
D-2349	0.0103	-65.1
D-2350	0.0427	-63.9

Example 4: Efficacy screening of select PNPLA3 siRNA molecules in a humanized mouse model

[0191] Associated adenovirus (AAV; serotype AAV8 or AAV7; endotoxin-free, prepared internally by Amgen) diluted in phosphate buffered saline (Thermo Fisher Scientific, 14190-136) to 4×10^{11} up to 1×10^{12} viral particles per animal, was injected intravenously into the tail vein of C57BL/6NCrl male mice (Charles River Laboratories Inc.) to drive expression of either human *PNPLA3*^{WT} (*PNPLA3-WT*), *PNPLA3*^{rs738409} (*PNPLA3-I148M*), or *PNPLA3*^{rs738409-rs738408} (*PNPLA3-I148MDM*) in the liver. Mice were generally 10-12 weeks of age and an n=4-6 animals were included per group. Every round of screening included at least two vehicle-treated control groups: AAV-empty vector and AAV-*PNPLA3*^{WT} or *PNPLA3*^{rs738409} and *PNPLA3*^{rs738409-rs738408} treated with vehicle. All siRNAs were tested against AAV-*PNPLA3*^{WT}, *PNPLA3*^{rs738409}, and/or *PNPLA3*^{rs738409-rs738408}. Two weeks after AAV injection, mice were treated with a single dose of siRNA D-2324 (0.5mM), via subcutaneous injection, at 0.5, 1.0, 3.0 or 5.0 milligrams per kilogram of animal, diluted in phosphate buffered saline (Thermo Fisher Scientific, 14190-136). At 8, 15, 22, 28 or 42 days post-siRNA injection, livers were collected from the animals, snap frozen in liquid nitrogen, processed for purified RNA using a QIAcube HT instrument (Qiagen, 9001793) and RNeasy 96 QIAcube HT kits (Qiagen, 74171) according to manufacturer's instructions. Samples were analyzed using a QIAxpert system (Qiagen, 9002340). RNA was treated with RQ1 RNase-Free DNase (Promega, M6101) and prepared for Real-Time qPCR using the TaqManTM RNA-to-CTTM 1-Step kit (Applied Biosystems, 4392653). Real-Time qPCR was run on a QuantStudio Real-Time PCR machine. Results are based on gene expression of human *PNPLA3* as normalized to mouse *Gapdh* (TaqManTM assays from Invitrogen, hs00228747_m1 and 4352932E, respectively), and presented as the relative knockdown of human *PNPLA3* mRNA expression compared to vehicle-treated control animals. Endogenous mouse *Pnpla3* expression was determined for comparison (Invitrogen, Mm00504420_m1).

[0192] For hepatic triglyceride content analysis, approximately 0.05-0.1 milligrams of frozen liver from an animal was homogenized in one milliliter of isopropanol. After one hour of incubation on ice, samples were spun at 10,000 rpm in a microfuge, and supernatants transferred to a clean deep-well 96-well plate. Triglyceride content was determined using the colorimetric Infinity Triglyceride Reagent (Thermo Fisher Scientific, TR22421) and Triglyceride Standard (Pointe Scientific, T7531-STD) according to manufacturer's instructions. Results are presented as milligrams of triglyceride per milligrams of tissue.

[0193] All animal experiments described herein were approved by the Institutional Animal Care and Use Committee (IACUC) of Amgen and cared for in accordance to the *Guide for the Care and Use of Laboratory Animals*, 8th Edition (National Research Council (U.S.). Committee for the Update of the Guide for the Care and Use of Laboratory Animals., Institute for Laboratory Animal Research (U.S.), and National Academies Press (U.S.) (2011) Guide for the care and use of laboratory animals. 8th Ed., National Academies Press, Washington, D.C. Mice were single-housed in an air-conditioned room at 22±2°C with a twelve-hour light; twelve-hour darkness cycle (0600-1800 hours). Animals had *ad libitum* access to a regular chow diet (Envigo, 2920X) and to water (reverse osmosis-purified) via automatic watering system, unless otherwise indicated. At termination, blood was collected by cardiac puncture under deep anesthesia, and then, following Association for Assessment and Accreditation of Laboratory Animal Care (AAALAC) guidelines, euthanized by a secondary physical method. **Figure 1A-D.** An example of five siRNA molecules screened for both dose-dependent mRNA knockdown and functional durability in vivo. Mice expressing human *PNPLA3*^{rs738409-rs738408} were treated with siRNA two-weeks after intravenous AAV injections. N=6 mice per group; data represented as the average and the standard error of the mean. (A) siRNAs were injected at 0.5, 1.0, 3.0, or 5.0 milligrams per kilogram of body weight subcutaneously into the abdomen of the mouse. Four weeks after siRNA treatment, mice were sacrificed, and the livers were collected and processed for gene expression analysis. The data represents the average relative knockdown of human *PNPLA3*^{rs738409-rs738408} in each group set to the vehicle-treated control group. (B) Livers from the same four-week treatment group were also processed for triglyceride content to access functional efficacy. The data represents the average milligrams of triglyceride per gram of tissue processed. (C) siRNAs were injected at 1.0 and 3.0 milligrams per kilogram of body weight subcutaneously into the abdomen of a parallel cohort of animals. Mice were harvested six weeks after siRNA treatment to compare durability of the siRNA molecules in vivo. The livers were collected and processed for gene expression analysis. The data represents the average relative knockdown of human *PNPLA3*^{rs738409-rs738408} in each group set to the vehicle-treated control group. (D) Livers from the same six-week treatment group were also processed for triglyceride content to access functional efficacy over time. The data represents the average milligrams of triglyceride per gram of tissue processed.

[0194] Data for relative knockdown is shown in Tables 8-12, showing relative knockdown at day 8, 15, 22, 28, and 42, respectively and the various doses. PNPLA3 knockdown is a percentage, with negative values indicating a decrease in PNPLA3 levels.

Table 8. Day 8 PNPLA3 knockdown assay

Duplex number	AAV vector	AAV particles/ animal	Dose administered (mg/kg)	PNPLA3 knockdown (%)
D-2092	PNPLA3-I148M	1.00E+12	3	-91.51
D-2092	PNPLA3-I148M	1.00E+12	5	-92.64
D-2095	PNPLA3-I148M	1.00E+12	3	-79.21
D-2095	PNPLA3-I148M	1.00E+12	5	-86.38
D-2068	PNPLA3-I148M	4.00E+11	5	-34.15
D-2069	PNPLA3-I148M	4.00E+11	5	-82.74
D-2070	PNPLA3-I148M	4.00E+11	5	-79.64
D-2071	PNPLA3-I148M	4.00E+11	5	-48.80
D-2071	PNPLA3-WT	4.00E+11	5	-59.43
D-2075	PNPLA3-I148M	4.00E+11	5	-79.10
D-2075	PNPLA3-WT	4.00E+11	5	-60.60
D-2079	PNPLA3-I148M	4.00E+11	5	-50.08
D-2072	PNPLA3-I148M	4.00E+11	5	-44.40
D-2072	PNPLA3-WT	4.00E+11	5	-43.75
D-2077	PNPLA3-I148M	4.00E+11	5	-28.46
D-2076	PNPLA3-I148M	4.00E+11	5	-83.25
D-2078	PNPLA3-I148M	4.00E+11	5	-49.10
D-2078	PNPLA3-WT	4.00E+11	5	-3.45
D-2073	PNPLA3-I148M	4.00E+11	5	-39.42
D-2074	PNPLA3-I148M	4.00E+11	5	-46.68
D-2074	PNPLA3-WT	4.00E+11	5	-5.11
D-2084	PNPLA3-I148M	4.00E+11	5	-87.49
D-2084	PNPLA3-I148M	8.00E+11	5	-88.75
D-2084	PNPLA3-I148M	8.00E+11	1	-16.49
D-2084	PNPLA3-WT	8.00E+11	1	-15.42
D-2085	PNPLA3-I148M	4.00E+11	5	-77.14
D-2085	PNPLA3-I148M	8.00E+11	5	-84.42
D-2085	PNPLA3-I148M	8.00E+11	1	-25.19
D-2085	PNPLA3-WT	8.00E+11	1	-21.14

D-2086	PNPLA3-I148M	4.00E+11	5	-64.54
D-2086	PNPLA3-I148M	4.00E+11	5	-52.95
D-2087	PNPLA3-I148M	4.00E+11	5	-20.15
D-2088	PNPLA3-I148M	4.00E+11	5	-47.18
D-2088	PNPLA3-I148M	8.00E+11	5	-66.96
D-2089	PNPLA3-I148M	8.00E+11	5	-85.47
D-2089	PNPLA3-WT	8.00E+11	1	-21.01
D-2089	PNPLA3-I148M	8.00E+11	1	-34.21
D-2089	PNPLA3-I148M	1.00E+12	5	-90.55
D-2090	PNPLA3-I148M	8.00E+11	5	-89.58
D-2090	PNPLA3-I148M	8.00E+11	1	-50.13
D-2090	PNPLA3-WT	8.00E+11	1	8.76
D-2091	PNPLA3-WT	8.00E+11	5	-35.70
D-2092	PNPLA3-I148M	8.00E+11	5	-92.34
D-2092	PNPLA3-I148M	8.00E+11	1	-51.35
D-2092	PNPLA3-WT	8.00E+11	1	-42.88
D-2093	PNPLA3-I148M	8.00E+11	5	-83.87
D-2094	PNPLA3-I148M	8.00E+11	5	-70.12
D-2095	PNPLA3-I148M	8.00E+11	1	-29.95
D-2095	PNPLA3-WT	8.00E+11	1	67.40
D-2095	PNPLA3-I148M	8.00E+11	5	-85.42
D-2096	PNPLA3-WT	8.00E+11	5	-90.44
D-2081	PNPLA3-I148M	8.00E+11	5	6.25
D-2081	PNPLA3-I148M	8.00E+11	5	-11.81
D-2097	PNPLA3-I148M	8.00E+11	5	-87.61
D-2098	PNPLA3-I148M	8.00E+11	5	-79.84
D-2099	PNPLA3-I148M	8.00E+11	5	-84.36
D-2100	PNPLA3-I148M	8.00E+11	5	-79.67
D-2101	PNPLA3-I148M	8.00E+11	5	-89.64
D-2102	PNPLA3-I148M	8.00E+11	5	-61.49
D-2103	PNPLA3-I148M	8.00E+11	5	-19.65
D-2104	PNPLA3-I148M	8.00E+11	5	-79.70
D-2104	PNPLA3-I148M	1.00E+12	5	-82.87
D-2105	PNPLA3-I148M	8.00E+11	5	-84.49
D-2105	PNPLA3-I148M	1.00E+12	5	-87.71
D-2152	PNPLA3-I148M	1.00E+12	5	-39.35
D-2153	PNPLA3-I148M	1.00E+12	5	-79.04
D-2154	PNPLA3-I148M	1.00E+12	5	-66.72
D-2155	PNPLA3-I148M	1.00E+12	5	-44.68

D-2156	PNPLA3-I148M	1.00E+12	5	-84.72
D-2157	PNPLA3-I148M	1.00E+12	5	-17.25
D-2280	PNPLA3-I148M DM	1.00E+12	5	-99.70
D-2280	PNPLA3 WT	1.00E+12	5	-51.13
D-2295	PNPLA3-WT	1.00E+12	5	-35.90
D-2295	PNPLA3-I148M DM	1.00E+12	5	-94.68
D-2296	PNPLA3-WT	1.00E+12	5	-23.24
D-2296	PNPLA3-I148M DM	1.00E+12	5	-92.78
D-2297	PNPLA3-WT	1.00E+12	5	43.71
D-2297	PNPLA3-I148M DM	1.00E+12	5	-94.59
D-2324	PNPLA3-WT	1.00E+12	5	6.53
D-2324	PNPLA3-I148M DM	1.00E+12	5	-97.39
D-2326	PNPLA3-WT	1.00E+12	5	-8.25
D-2326	PNPLA3-I148M DM	1.00E+12	5	-77.82
D-2328	PNPLA3-WT	1.00E+12	5	-2.12
D-2328	PNPLA3-I148M DM	1.00E+12	5	-92.49

Table 9. Day 15 PNPLA3 knockdown assay

Duplex number	AAV vector	AAV particles/ animal	Dose administered (mg/kg)	PNPLA3 knockdown (%)
D-2092	PNPLA3-I148M	1.00E+12	3	-87.58
D-2092	PNPLA3-I148M	1.00E+12	5	-93.96
D-2095	PNPLA3-I148M	1.00E+12	3	-81.73
D-2095	PNPLA3-I148M	1.00E+12	5	-72.99
D-2131	PNPLA3-I148M	1.00E+12	5	-66.02
D-2186	PNPLA3-I148M	1.00E+12	5	-87.67
D-2089	PNPLA3-I148M	1.00E+12	5	-95.01
D-2104	PNPLA3-I148M	1.00E+12	5	-76.01
D-2105	PNPLA3-I148M	1.00E+12	5	-72.67
D-2123	PNPLA3-I148M	1.00E+12	5	-56.93
D-2128	PNPLA3-I148M	1.00E+12	5	-37.26
D-2138	PNPLA3-I148M	1.00E+12	5	-62.16
D-2149	PNPLA3-I148M	1.00E+12	5	-72.34
D-2156	PNPLA3-I148M	1.00E+12	5	-75.81
D-2259	PNPLA3-I148M	1.00E+12	5	-79.15
D-2260	PNPLA3-I148M	1.00E+12	5	-69.97
D-2261	PNPLA3-I148M	1.00E+12	5	-50.40

D-2262	PNPLA3-I148M	1.00E+12	5	-84.36
D-2263	PNPLA3-I148M	1.00E+12	5	-77.08
D-2264	PNPLA3-I148M	1.00E+12	5	-40.86
D-2269	PNPLA3-I148M	1.00E+12	5	-37.55
D-2270	PNPLA3-I148M	1.00E+12	5	-77.42
D-2271	PNPLA3-I148M	1.00E+12	5	-26.87
D-2272	PNPLA3-I148M	1.00E+12	5	-56.05
D-2280	PNPLA3-I148M DM	1.00E+12	3	-86.30
D-2287	PNPLA3-I148M DM	1.00E+12	3	-94.73
D-2289	PNPLA3-I148M DM	1.00E+12	3	-93.48
D-2292	PNPLA3-I148M DM	1.00E+12	3	-82.48
D-2297	PNPLA3-I148M DM	1.00E+12	3	-72.61
D-2322	PNPLA3-I148M DM	1.00E+12	3	-32.92
D-2324	PNPLA3-I148M DM	1.00E+12	3	-87.56
D-2327	PNPLA3-I148M DM	1.00E+12	3	-86.70
D-2345	PNPLA3-WT	1.00E+12	5	-83.48
D-2346	PNPLA3-WT	1.00E+12	5	-75.93
D-2347	PNPLA3-WT	1.00E+12	5	-22.83
D-2348	PNPLA3-WT	1.00E+12	5	-20.09
D-2349	PNPLA3-WT	1.00E+12	5	-67.70
D-2350	PNPLA3-WT	1.00E+12	5	-57.51
D-2351	PNPLA3-WT	1.00E+12	5	-56.11
D-2352	PNPLA3-I148M DM	1.00E+12	3	-92.21
D-2353	PNPLA3-I148M DM	1.00E+12	3	-89.55
D-2354	PNPLA3-I148M DM	1.00E+12	3	-90.21
D-2358	PNPLA3-I148M DM	1.00E+12	3	-95.10
D-2359	PNPLA3-I148M DM	1.00E+12	3	-91.17
D-2360	PNPLA3-I148M DM	1.00E+12	3	-63.98
D-2361	PNPLA3-I148M DM	1.00E+12	3	-92.47
D-2362	PNPLA3-I148M DM	1.00E+12	3	-95.10
D-2364	PNPLA3-I148M DM	1.00E+12	3	-95.31
D-2370	PNPLA3-I148M DM	1.00E+12	3	-95.99
D-2370	PNPLA3 WT	1.00E+12	3	11.88
D-2371	PNPLA3-I148M DM	1.00E+12	3	-91.14
D-2372	PNPLA3-I148M DM	1.00E+12	3	-93.71
D-2373	PNPLA3-I148M DM	1.00E+12	3	-73.80
D-2374	PNPLA3-I148M DM	1.00E+12	3	-75.98
D-2375	PNPLA3-I148M DM	1.00E+12	3	-96.68
D-2376	PNPLA3-I148M DM	1.00E+12	3	-96.78
D-2377	PNPLA3-I148M DM	1.00E+12	3	-96.88

D-2378	PNPLA3-I148M DM	1.00E+12	3	-97.60
D-2379	PNPLA3-I148M DM	1.00E+12	3	-94.73
D-2380	PNPLA3-I148M DM	1.00E+12	3	-94.73
D-2381	PNPLA3-I148M DM	1.00E+12	3	-76.24
D-2382	PNPLA3-I148M DM	1.00E+12	3	-80.33
D-2383	PNPLA3-I148M DM	1.00E+12	3	-71.98
D-2384	PNPLA3-I148M DM	1.00E+12	3	-87.24
D-2385	PNPLA3-I148M DM	1.00E+12	3	-78.77
D-2386	PNPLA3-I148M DM	1.00E+12	3	-70.58
D-2387	PNPLA3-I148M DM	1.00E+12	3	-67.09
D-2390	PNPLA3-I148M DM	1.00E+12	3	-92.97
D-2391	PNPLA3-I148M DM	1.00E+12	3	-94.10
D-2392	PNPLA3-I148M DM	1.00E+12	3	-92.14
D-2395	PNPLA3-I148M DM	1.00E+12	3	-85.63
D-2396	PNPLA3-I148M DM	1.00E+12	3	-94.63
D-2397	PNPLA3-I148M DM	1.00E+12	3	-92.90
D-2398	PNPLA3-I148M DM	1.00E+12	3	-95.48
D-2399	PNPLA3-I148M DM	1.00E+12	3	-93.40
D-2400	PNPLA3-I148M DM	1.00E+12	3	-97.55
D-2401	PNPLA3-I148M DM	1.00E+12	3	-96.98
D-2402	PNPLA3-I148M DM	1.00E+12	3	-97.25
D-2403	PNPLA3 WT	1.00E+12	3	36.90
D-2404	PNPLA3-I148M DM	1.00E+12	3	-95.08
D-2405	PNPLA3-I148M DM	1.00E+12	3	-96.93
D-2406	PNPLA3 WT	1.00E+12	3	28.80
D-2395	PNPLA3-I148M DM	1.00E+12	3	-17.25
D-2396	PNPLA3-I148M DM	1.00E+12	3	-93.70
D-2413	PNPLA3-I148M DM	1.00E+12	3	-94.80
D-2415	PNPLA3-I148M DM	1.00E+12	3	-90.95
D-2418	PNPLA3-I148M DM	1.00E+12	3	-88.45
D-2419	PNPLA3 WT	1.00E+12	3	43.72
D-2453	PNPLA3 WT	1.00E+12	3	-81.33
D-2454	PNPLA3 WT	1.00E+12	3	-83.38
D-2455	PNPLA3 WT	1.00E+12	3	-68.85
D-2456	PNPLA3 WT	1.00E+12	3	-89.03
D-2460	PNPLA3 WT	1.00E+12	3	-68.65
D-2461	PNPLA3 WT	1.00E+12	3	-47.85

Table 10. Day 22 PNPLA3 knockdown assay

Duplex number	AAV vector	AAV particles/ animal	Dose administered (mg/kg)	PNPLA3 knockdown (%)
D-2092	PNPLA3-I148M	1.00E+12	3	-74.61
D-2092	PNPLA3-I148M	1.00E+12	5	-85.78
D-2095	PNPLA3-I148M	1.00E+12	3	-27.78
D-2095	PNPLA3-I148M	1.00E+12	5	-33.60
D-2324	PNPLA3-I148M DM	1.00E+12	3	-56.68
D-2324	PNPLA3-I148M DM	1.00E+12	5	-88.02
D-2089	PNPLA3-I148M	1.00E+12	5	-80.86
D-2104	PNPLA3-I148M	1.00E+12	5	-65.61
D-2105	PNPLA3-I148M	1.00E+12	5	-39.67
D-2280	PNPLA3-I148M DM	1.00E+12	5	-84.12
D-2297	PNPLA3-I148M DM	1.00E+12	3	-58.01
D-2297	PNPLA3-I148M DM	1.00E+12	5	-76.43
D-2352	PNPLA3-I148M DM	1.00E+12	5	-92.74
D-2353	PNPLA3-I148M DM	1.00E+12	5	-84.54
D-2354	PNPLA3-I148M DM	1.00E+12	5	-89.06
D-2355	PNPLA3-I148M DM	1.00E+12	5	-95.53
D-2356	PNPLA3-I148M DM	1.00E+12	5	-94.99
D-2357	PNPLA3-I148M DM	1.00E+12	5	-97.37

Table 11. Day 28 PNPLA3 knockdown assay

Duplex number	AAV vector	AAV particles/ animal	Dose administered (mg/kg)	PNPLA3 knockdown (%)
D-2092	PNPLA3-I148M	1.00E+12	3	-72.57
D-2092	PNPLA3-I148M	1.00E+12	5	-82.74
D-2095	PNPLA3-I148M	1.00E+12	3	-32.93
D-2095	PNPLA3-I148M	1.00E+12	5	-55.31
D-2089	PNPLA3-I148M	1.00E+12	5	-63.49
D-2104	PNPLA3-I148M	1.00E+12	5	-44.39
D-2105	PNPLA3-I148M	1.00E+12	5	-39.80
D-2370	PNPLA3-I148M DM	1.00E+12	3	-89.28
D-2400	PNPLA3-I148M DM	1.00E+12	3	-88.23

D-2401	PNPLA3-I148M DM	1.00E+12	3	-91.95
D-2402	PNPLA3-I148M DM	1.00E+12	0.5	-50.65
D-2402	PNPLA3-I148M DM	1.00E+12	1	-69.37
D-2402	PNPLA3-I148M DM	1.00E+12	3	-96.43
D-2402	PNPLA3-I148M DM	1.00E+12	3	-85.44
D-2402	PNPLA3-I148M DM	1.00E+12	3	-90.52
D-2404	PNPLA3-I148M DM	1.00E+12	3	-95.67
D-2419	PNPLA3-I148M DM	1.00E+12	0.5	-68.83
D-2419	PNPLA3-I148M DM	1.00E+12	1	-76.88
D-2419	PNPLA3-I148M DM	1.00E+12	3	-97.53
D-2419	PNPLA3-I148M DM	1.00E+12	3	-93.95
D-2419	PNPLA3-I148M DM	1.00E+12	3	-89.66
D-2419	PNPLA3-I148M DM	1.00E+12	3	-93.25
D-2420	PNPLA3-I148M DM	1.00E+12	3	-95.93
D-2421	PNPLA3-I148M DM	1.00E+12	0.5	-50.01
D-2421	PNPLA3-I148M DM	1.00E+12	1	-66.30
D-2421	PNPLA3-I148M DM	1.00E+12	3	-94.99
D-2421	PNPLA3-I148M DM	1.00E+12	3	-80.05
D-2421	PNPLA3 WT	1.00E+12	3	-36.65
D-2421	PNPLA3-I148M DM	1.00E+12	3	-91.08
D-2425	PNPLA3-I148M DM	1.00E+12	3	-16.07
D-2426	PNPLA3-I148M DM	1.00E+12	3	-37.54
D-2427	PNPLA3-I148M DM	1.00E+12	3	-25.19

D-2428	PNPLA3-I148M DM	1.00E+12	3	-16.71
D-2437	PNPLA3-I148M DM	1.00E+12	3	-93.78
D-2438	PNPLA3-I148M DM	1.00E+12	3	-90.63
D-2439	PNPLA3-I148M DM	1.00E+12	3	-88.10
D-2440	PNPLA3-I148M DM	1.00E+12	3	-95.25
D-2441	PNPLA3-I148M DM	1.00E+12	3	-90.13
D-2442	PNPLA3-I148M DM	1.00E+12	3	-57.24
D-2443	PNPLA3-I148M DM	1.00E+12	3	-95.43
D-2444	PNPLA3-I148M DM	1.00E+12	3	-90.58
D-2445	PNPLA3-I148M DM	1.00E+12	3	-84.55
D-2446	PNPLA3-I148M DM	1.00E+12	3	-81.50
D-2427	PNPLA3-I148M DM	1.00E+12	3	-90.09
D-2462	PNPLA3-I148M DM	1.00E+12	3	-89.20
D-2463	PNPLA3-I148M DM	1.00E+12	3	-41.25
D-2464	PNPLA3-I148M DM	1.00E+12	3	-60.53
D-2465	PNPLA3-I148M DM	1.00E+12	3	-91.35
D-2466	PNPLA3-I148M DM	1.00E+12	3	-93.68
D-2467	PNPLA3-I148M DM	1.00E+12	3	-85.15
D-2472	PNPLA3-I148M DM	1.00E+12	0.5	-46.58
D-2472	PNPLA3-I148M DM	1.00E+12	1	-74.47
D-2472	PNPLA3-I148M DM	1.00E+12	3	-88.79
D-2472	PNPLA3 WT	1.00E+12	3	-23.16
D-2472	PNPLA3-I148M DM	1.00E+12	3	-90.54

D-2473	PNPLA3-I148M DM	1.00E+12	0.5	-57.95
D-2473	PNPLA3-I148M DM	1.00E+12	1	-71.96
D-2473	PNPLA3-I148M DM	1.00E+12	3	-91.70
D-2473	PNPLA3-I148M DM	1.00E+12	3	-85.94
D-2473	PNPLA3 WT	1.00E+12	3	-18.70

Table 12. Day 42 PNPLA3 knockdown assay

Duplex number	AAV vector	AAV particles/ animal	Dose administered (mg/kg)	PNPLA3 knockdown (%)
D-2402	PNPLA3-I148M DM	1.00E+12	1	-57.74
D-2402	PNPLA3-I148M DM	1.00E+12	3	-83.75
D-2419	PNPLA3-I148M DM	1.00E+12	1	-71.07
D-2419	PNPLA3-I148M DM	1.00E+12	3	-70.8
D-2421	PNPLA3-I148M DM	1.00E+12	1	-62.21
D-2421	PNPLA3-I148M DM	1.00E+12	3	-80.12
D-2472	PNPLA3-I148M DM	1.00E+12	1	-60.54
D-2472	PNPLA3-I148M DM	1.00E+12	3	-74.77
D-2473	PNPLA3-I148M DM	1.00E+12	1	-54.55
D-2473	PNPLA3-I148M DM	1.00E+12	3	-81.13

Example 5: Prevention and rescue of NAFLD by siRNA molecules in a humanized *PNPLA3*^{rs738409-rs738408} mouse model

[0195] The ‘American Lifestyle-Induced Obesity Syndrome’, or ALIOS mouse model for NAFLD/NASH is developed by feeding mice with a diet high in trans-fats (45% of the total

amount of fat) and sugar (Tetri 2008). For these studies, eight to ten-week old C57BL/6NCrl male mice (Charles River Laboratories Inc.) were injected with AAV-empty vector or AAV8-*PNPLA3^{rs738409-rs738408}*, as described previously. At the time of AAV injection, mice were either maintained on a normal chow diet or placed on the ALIOS diet (Envigo, TD.06303) complete with drinking water composed of 55% fructose and 45% glucose (Sigma, F0127 and G7021, respectively) until harvest. In previous experiments, it was established that over-expression of *PNPLA3^{rs738409-rs738408}*, in this context, both accelerates and worsens NAFLD phenotypes (data not shown).

[0196] Two weeks after AAV injection and diet initiation, mice were treated with a single dose of siRNA D-2324 (0.5mM), via subcutaneous injection, at 5.0 milligrams per kilogram of animal, diluted in phosphate buffered saline (Thermo Fisher Scientific, 14190-136) or a vehicle control. Dosing was repeated biweekly until harvest. At the time of harvest, body weights were collected, followed by serum via cardiac puncture under isoflurane anesthesia, followed by liver weights. The median lobe was fixed via 10% neutral buffered formalin, followed by paraffin processing and embedding. The remainder of the liver was snap frozen for content and gene expression analysis, as described previously.

[0197] Snap frozen liver tissue was processed for RNA and gene expression analysis, as described previously. Results are shown as both the raw Ct value and relative mRNA expression of the indicated gene normalized to mouse *Gapdh*. (TaqMan™ assays from Invitrogen: human *PNPLA3*, hs00228747_m1; mouse *Pnpla3*, Mm00504420_m1; mouse *Gapdh*, 4352932E).

[0198] Formalin-fixed tissues were processed for Hematoxylin and Eosin staining (Dako, CS70030-2, CS70130-2, respectively) according to the manufacturer's instructions. Scoring for steatosis and inflammation was performed by a board-certified pathologist.

[0199] Serum analysis included TIMP1, a biomarker associated with NASH and NASH-related fibrosis (Youssani 2011). The TIMP1 ELISA (R&D Systems, MTM100) was performed according to the manufacturer's instructions.

[0200] **Figure 2A-G.** To evaluate the ability of a *PNPLA3^{rs738409-rs738408}* – specific siRNA molecule D-2324 to prevent the development of phenotypes associated with NAFLD and overexpression of *PNPLA3^{rs738409-rs738408}*, mice received AAV8-empty vector (EV), or AAV8-*PNPLA3^{rs738409-rs738408}*, or vehicle, and were maintained on a regular chow diet or transitioned to

the ALIOS diet. Two weeks after AAV injections, mice were treated with siRNA or vehicle, every other week for six weeks; a total of three injection rounds. Mice were harvested at the eight-week time point. Results are presented as the group average and standard error, N=8 per group. Asterisks represent statistical significance to the AAV8-*PNPLA3*^{rs738409-rs738408} cohort, generated by One-way ANOVA employing Dunnett's multiple comparisons test. (A) The ratio of liver weight (grams) to body weight (grams) at the time of harvest. Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, **=0.0018, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, ****<0.0001. (B) Confirmation of human *PNPLA3* mRNA expression and silencing in the liver by qPCR. (left) Raw Ct value and (right) relative fold mRNA expression normalized to mouse *Gapdh*; comparing the ALIOS-fed *PNPLA3*^{rs738409-rs738408} + vehicle and *PNPLA3*^{rs738409-rs738408} + siRNA groups. (C) Analysis of mouse *Pnpla3* mRNA expression in liver by qPCR indicates endogenous *Pnpla3* is not significantly altered by AAV-mediated over-expression or siRNA silencing. (left) Raw Ct value and (right) relative fold mRNA expression normalized to mouse *Gapdh*; comparing the chow-fed no-AAV group to the ALIOS-fed *PNPLA3*^{rs738409-rs738408} + vehicle and *PNPLA3*^{rs738409-rs738408} + siRNA groups. (D) Hepatic triglyceride content presented as milligrams of triglyceride per gram of liver tissue. Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, *=0.0393, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, **=0.0063. (E) Serum TIMP1 presented as picograms per milliliter of serum. Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, ****<0.0001, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, ****<0.0001. (F) Histological indication of steatosis based on H&E staining, scored as: Within normal limits (0), minimal (1), mild (2), moderate (3), and severe (4). Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, no significance, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, **=0.0012. (G) Histological indication of inflammation based on H&E staining, scored as: Within normal limits (0), minimal (1), mild (2), moderate (3), and severe (4). Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, ****<0.0001, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, ****<0.0001.

[0201] **Figure 3A-G.** To evaluate the ability of a *PNPLA3*^{rs738409-rs738408} – specific siRNA molecule to prevent further progression of *PNPLA3*^{rs738409-rs738408}-mediated disease after disease onset, mice received AAV8-empty vector (EV), or AAV8-*PNPLA3*^{rs738409-rs738408}, or vehicle, and were maintained on a regular chow diet or transitioned to the ALIOS diet. Eight weeks after AAV

injections and diet changes, mice were treated with siRNA or vehicle, every other week for eight more weeks; a total of four injection rounds. Mice were harvested at the sixteen-week time point. Although no change was observed in steatosis, if siRNA treatment began after disease induction, several other disease-associated end points were significantly reduced. Results are presented as averages and standard error, N=8 per group. Asterisks represent statistical significance to the AAV8-*PNPLA3*^{rs738409-rs738408} cohort, generated by One-way ANOVA employing Dunnett's multiple comparisons test. (A) The ratio of liver weight (grams) to body weight (grams) at the time of harvest. Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, not significant, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, ***=0.0006. (B) Confirmation of human *PNPLA3* mRNA expression and silencing in the liver by qPCR. (left) Raw Ct value and (right) relative fold mRNA expression normalized to mouse *Gapdh*; comparing the ALIOS-fed *PNPLA3*^{rs738409-rs738408} + vehicle and *PNPLA3*^{rs738409-rs738408} + siRNA groups. (C) Analysis of mouse *Pnpla3* mRNA expression in liver by qPCR indicates endogenous *Pnpla3* is not significantly altered by AAV-mediated over-expression or siRNA silencing. (left) Raw Ct value and (right) relative fold mRNA expression normalized to mouse *Gapdh*; comparing the chow-fed no-AAV group to the ALIOS-fed *PNPLA3*^{rs738409-rs738408} + vehicle and *PNPLA3*^{rs738409-rs738408} + siRNA groups. (D) Hepatic triglyceride content presented as milligrams of triglyceride per gram of liver tissue. Adjusted P values: AAV-EV + vehicle, not significant, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, *=0.0403. (E) Serum TIMP1 presented as picograms per milliliter of serum. Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, **=0.0027, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, **=0.002. (F) Histological indication of steatosis based on H&E staining, scored as: Within normal limits (0), minimal (1), mild (2), moderate (3), and severe (4). Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, not significant, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, not significant. (G) Histological indication of inflammation based on H&E staining, scored as: Within normal limits (0), minimal (1), mild (2), moderate (3), and severe (4). Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, not significant, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, **=0.0068.

[0202] **Figure 4A-D.** To evaluate the ability of a *PNPLA3*^{rs738409-rs738408} – specific siRNA molecule to rescue disease-associated phenotypes due to overexpression of *PNPLA3*^{rs738409-rs738408}, liver and serum from ALIOS-fed eight-week AAV8-*PNPLA3*^{rs738409-rs738408} -vehicle treated mice

were compared to livers and serum from ALIOS-fed sixteen-week vehicle- or siRNA-treated AAV8-*PNPLA3^{rs738409-rs738408}* mice. Although no change was observed in steatosis at this time point with siRNA treatment, hepatic triglyceride, serum TIMP1, and inflammation were all statistically lower at sixteen weeks compared to vehicle controls at eight weeks. Results are presented as averages and standard error, N=8 per group. Asterisks represent statistical significance to the eight-week AAV8-*PNPLA3^{rs738409-rs738408}* vehicle-treated cohort, generated by One-way ANOVA employing Dunnett's multiple comparisons test. (A) Hepatic triglyceride content presented as milligrams of triglyceride per gram of liver tissue. Adjusted P values: 16WK AAV- *PNPLA3^{rs738409-rs738408}* + vehicle, not significant; 16WK AAV- *PNPLA3^{rs738409-rs738408}* + siRNA, **=0.0011. (B) Serum Timp1 presented as picograms per milliliter of serum. Adjusted P values: 16WK AAV- *PNPLA3^{rs738409-rs738408}* + vehicle, not significant; 16WK AAV- *PNPLA3^{rs738409-rs738408}* + siRNA, *=0.0134. (C) Histological indication of steatosis based on H&E staining, scored as: Within normal limits (0), minimal (1), mild (2), moderate (3), and severe (4). Adjusted P values: 16WK AAV- *PNPLA3^{rs738409-rs738408}* + vehicle, not significant; 16WK AAV- *PNPLA3^{rs738409-rs738408}* + siRNA, not significant. (D) Histological indication of inflammation based on H&E staining, scored as: Within normal limits (0), minimal (1), mild (2), moderate (3), and severe (4). Adjusted P values: 16WK AAV- *PNPLA3^{rs738409-rs738408}* + vehicle, not significant; 16WK AAV- *PNPLA3^{rs738409-rs738408}* + siRNA, *=0.0112.

Example 6: Prevention of liver fibrosis by siRNA molecules in a humanized *PNPLA3^{rs738409-rs738408}* mouse model

[0203] The “AMLN” diet, developed by Amylin Pharmaceuticals (Clapper 2013), is a modified version of the ALIOS diet. The feed includes a ten-fold increase in cholesterol (2%) and additional sucrose. Mice placed on the “AMLN” diet develop mild to moderate fibrosis after 20-30 weeks (Clapper, Mells and Kristiansen papers). For this study, eight to ten-week old C57BL/6NCrl male mice (Charles River Laboratories Inc.) were injected with AAV-empty vector or AAV- *PNPLA3^{rs738409-rs738408}*, as described above, to accelerate disease onset. At the time of AAV injection, mice were continued on a normal chow diet or placed on the Envigo diet, TD.170748, complete with drinking water composed of 55% fructose and 45% glucose (Sigma, F0127 and G7021, respectively) until harvest.

- [0204] Two weeks post-AAV injection and diet initiation, mice were treated with a single dose of siRNA D-2324 (0.5mM), via subcutaneous injection, at 5.0 milligrams per kilogram of animal, diluted in phosphate buffered saline (Thermo Fisher Scientific, 14190-136) or a vehicle control. Dosing was repeated biweekly until harvest. At the time of harvest, body weights were collected, followed by serum via cardiac puncture under isoflurane anesthesia, followed by liver weights. The median lobe was fixed via 10% neutral buffered formalin, followed by paraffin processing and embedding. The remainder of the liver was snap frozen for gene expression analysis.
- [0205] Snap frozen liver tissue was processed for RNA and gene expression analysis, as described previously. Results are shown as both the raw Ct value and relative mRNA expression of the indicated gene normalized to mouse *Gapdh*. (TaqMan™ assays from Invitrogen: human *PNPLA3*, hs00228747_m1; mouse *Pnpla3*, Mm00504420_m1; mouse *Colla1*, Mm00801666_g1; mouse *Col3a1*, Mm01254471_g1; *Col4a1*, Mm01210125_m1; mouse *Gapdh*, 4352932E). *Colla1*, *Col3a1* and *Col4a1* are extracellular matrix markers associated with hepatic stellate cell activation and liver fibrosis (Baiocchi 2016).
- [0206] Formalin-fixed tissues were processed for Hematoxylin, Eosin, and Masson's Trichrome staining (Dako, CS70030-2, CS70130-2, AR17311-2, respectively) according to the manufacturer's instructions. Anti-Smooth Muscle Actin staining was performed without antigen retrieval and using a DAKO autostainer. Slides were processed using Peroxidized 1 and Sniper (Biocare, PX968 and BS966, respectively) and stained with monoclonal anti-Actin, alpha-Smooth Muscle antibody (Sigma, F3777), followed by Rabbit anti-FITC (Invitrogen, 711900), Envision-Rabbit HRP polymer (Dako, K4003), DAB+ (Dako, K3468), and Hematoxylin. Scoring for the amount of steatosis, inflammation, oval cell/ bile duct hyperplasia, and aSMA-positive cells was performed by a board-certified pathologist.
- [0207] Serum was analyzed for mouse TIMP1 (R&D Systems, MTM100) and Mouse Cytokeratin 18-M30 (Cusabio, CSB-E14265m) according to manufacturer's instructions. In addition to TIMP1, Cytokine 18-M30 has been identified as a potential biomarker for NAFLD/NASH, including early fibrosis (Neuman 2014 and Yang 2015). **Figure 5A-L.** To evaluate the ability of a *PNPLA3*^{rs738409-rs738408} – specific siRNA molecule to prevent the development of early fibrosis, mice received AAV8-empty vector (EV), or AAV8-*PNPLA3*^{rs738409-rs738408}, or vehicle, and were maintained on a regular chow diet or transitioned to the AMLN diet.

Two weeks after AAV injections, mice were treated with siRNA, D-2324, or vehicle, every other week for ten more weeks; a total of six injection rounds. Mice were harvested at the ten-week time point. Results are presented as averages and standard error, Chow-fed no AAV + vehicle and AMLN-fed AAV8-*PNPLA3*^{rs738409-rs738408} + vehicle, N=8 per group; AMLN-fed AAV8-*PNPLA3*^{rs738409-rs738408} + vehicle and AAV8-*PNPLA3*^{rs738409-rs738408} + siRNA, N=12 per group. Asterisks represent statistical significance to the AAV8-*PNPLA3*^{rs738409-rs738408} – vehicle-treated cohort, generated by One-way ANOVA employing Dunnett's multiple comparisons test. (A) The ratio of liver weight (grams) to body weight (grams) at the time of harvest. Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, not significant, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, ****<0.0001. (B) Confirmation of human *PNPLA3* mRNA expression and silencing in the liver by qPCR. (left) Raw Ct value and (right) relative fold mRNA expression normalized to mouse *Gapdh*; comparing the *PNPLA3*^{rs738409-rs738408} + vehicle and *PNPLA3*^{rs738409-rs738408} + siRNA groups. (C) Analysis of mouse *Pnpla3* mRNA expression in liver by qPCR indicates endogenous *Pnpla3* is not significantly altered by AAV-mediated over-expression or siRNA silencing. (left) Raw Ct value and (right) relative fold mRNA expression normalized to mouse *Gapdh*; comparing the chow-fed no-AAV group to the AMLN-fed *PNPLA3*^{rs738409-rs738408} + vehicle and *PNPLA3*^{rs738409-rs738408} + siRNA groups. (D) Serum Timp1 presented as picograms per milliliter of serum. Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, ****<0.0001, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, ****<0.0001. (E) Serum CK18m30 presented as picograms per milliliter of serum. Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, ****<0.0001, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, ****<0.0001. (F) Histological indication of inflammation based on H&E staining, scored as: Within normal limits (0), minimal (1), mild (2), moderate (3), and severe (4). Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, *=0.0108, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, ****<0.0001. (G) Histological indication of oval cell/bile duct hyperplasia based on H&E staining, scored as: Within normal limits (0), minimal (1), mild (2), moderate (3), and severe (4). Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, not significant, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, **=0.0081. (H) Immunohistochemical staining for anti-Smooth Muscle Actin, scored as: Within normal limits (0), minimal (1), mild (2), moderate (3), and severe (4). Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle,

*=0.0101, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, ***=0.0002. (I) Masson's Trichrome staining for fibrosis, scored as: Within normal limits (0), minimal (1), mild (2), moderate (3), and severe (4). Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, not significant, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, not significant. (J) Mouse *Coll1a1* mRNA expression in liver by qPCR. Relative fold mRNA expression normalized to mouse *Gapdh*. Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, ****<0.0001, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, ****<0.0001. (K) Mouse *Col3a1* mRNA expression in liver by qPCR. Relative fold mRNA expression normalized to mouse *Gapdh*. Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, ****<0.0001, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, ****<0.0001. (L) Mouse *Col4a1* mRNA expression in liver by qPCR. Relative fold mRNA expression normalized to mouse *Gapdh*. Adjusted P values: no AAV + vehicle group, ****<0.0001, AAV-EV + vehicle, ***<0.0005, AAV- *PNPLA3*^{rs738409-rs738408} + siRNA, **<0.0041.

What is Claimed

1. An RNAi construct comprising a sense strand and an antisense strand, wherein the antisense strand consists of a nucleic acid sequence of SEQ ID NO: 648, 734, or 2, and wherein the RNAi construct inhibits the expression of Patatin-Like Phospholipase Domain Containing 3 (PNPLA3).
2. The RNAi construct of claim 1, wherein the antisense strand comprises a region that is complementary to a PNPLA3 mRNA sequence.
3. The RNAi construct any of the above claims, wherein the sense strand consists of a nucleic acid sequence of SEQ ID NO: 647, 733, or 1.
4. The RNAi construct any of the above claims, wherein the construct preferentially inhibits a PNPLA3-rs738409 minor allele.
5. The RNAi construct of claim 4, wherein the construct has at least 10% greater inhibition of a PNPLA3-rs738409 minor allele as compared to the major allele.
6. The RNAi construct of any of the above claims, wherein the construct preferentially inhibits a PNPLA3-rs738408 minor allele.
7. The RNAi construct of claim 6, wherein the construct has at least 10% greater inhibition of a PNPLA3-rs738409-rs738408 double minor allele as compared to the major allele.
8. The RNAi construct any of the above claims, wherein the sense strand comprises a sequence that is sufficiently complementary to the sequence of the antisense strand to form a duplex region of 15 to 30 base pairs in length.
9. The RNAi construct of claim 8, wherein the duplex region is 19 base pairs in length.
10. The RNAi construct of any one of claims 1 to 9, wherein the RNAi construct comprises at least one blunt end.
11. The RNAi construct of any one of claims 1 to 9, wherein the RNAi construct comprises at least one nucleotide overhang of 1 to 4 unpaired nucleotides.
12. The RNAi construct of claim 11, wherein the nucleotide overhang has 2 unpaired nucleotides.

13. The RNAi construct of any one of claims 11 to 12, wherein the nucleotide overhang comprises a 5'-UU-3' dinucleotide or a 5'-dTdT-3' dinucleotide.
14. The RNAi construct of any one of claims 1 to 13, wherein the RNAi construct comprises at least one modified nucleotide, wherein the modified nucleotide is a 2'-modified nucleotide.
15. The RNAi construct of claim 14, wherein the modified nucleotide is a 2'-fluoro modified nucleotide, a 2'-O-methyl modified nucleotide, a 2'-O-methoxyethyl modified nucleotide, a 2'-O-allyl modified nucleotide, a bicyclic nucleic acid (BNA), a glycol nucleic acid, an inverted base or combinations thereof.
16. The RNAi construct of claim 15, wherein the modified nucleotide is a 2'-O-methyl modified nucleotide, a 2'-O-methoxyethyl modified nucleotide, a 2'-fluoro modified nucleotide, or combinations thereof.
17. The RNAi construct of claim 14, wherein all of the nucleotides in the sense and antisense strands are modified nucleotides, wherein the modified nucleotide is a 2'-modified nucleotide.
18. The RNAi construct of claim 17, wherein the modified nucleotides are 2'-O-methylmodified nucleotides, 2'-fluoro modified nucleotides, or combinations thereof.
19. The RNAi construct of any one of claims 1 to 18, wherein the RNAi construct comprises at least one phosphorothioate internucleotide linkage.
20. The RNAi construct of claim 19, wherein the RNAi construct comprises two consecutive phosphorothioate internucleotide linkages at the 3' end of the antisense strand.
21. The RNAi construct of claim 19, wherein the RNAi construct comprises two consecutive phosphorothioate internucleotide linkages at both the 3' and 5' ends of the antisense strand and two consecutive phosphorothioate internucleotide linkages at the 5' end of the sense strand.
22. The RNAi construct of any one of claims 1 to 21, wherein the RNAi construct consists of a sense and antisense strand having SEQ ID Nos: 647 and 648; 733 and 734; 1 and 2; 1609 and 1610; 1695 and 1696; and 167 and 168.
23. The RNAi construct of any one of claims 1 to 22, wherein the RNAi construct reduces the expression level of PNPLA3 in liver cells following incubation with the RNAi construct as

compared to the PNPLA3 expression level in liver cells that have been incubated with a control RNAi construct.

24. The RNAi construct of claim 23, wherein the liver cells are Hep3B or HepG2 cells.
25. The RNAi construct of any one of claims 1 to 24, wherein the RNAi construct inhibits at least 10% of PNPLA3 expression at 5 nM in Hep3B cells *in vitro*.
26. The RNAi construct of any one of claims 1 to 24, wherein the RNAi construct inhibits at least 10% of PNPLA3 expression at 5 nM in HepG2 cells *in vitro*.
27. The RNAi construct of any one of claims 1 to 24, wherein the RNAi construct inhibits PNPLA3 expression in Hep3B cells with an IC₅₀ of less than 1 nM.
28. The RNAi construct of any one of claims 1 to 24, wherein the RNAi construct inhibits PNPLA3 expression in HepG2 cells with an IC₅₀ of less than 1 nM.
29. A pharmaceutical composition comprising the RNAi construct of any one of claims 1 to 28 and a pharmaceutically acceptable carrier, excipient, or diluent.