DESCRIPTION

T CELL-REDIRECTED ANTIGEN-BINDING MOLECULE FOR CELLS HAVING IMMUNOSUPPRESSION FUNCTION

Technical Field

The present invention relates to antigen-binding molecules that enable treatment of cancer by damaging cells having an immune response-suppressing function; pharmaceutical compositions for treating various carcinomas comprising the antigen-binding molecule; or methods for treating cancer using the pharmaceutical compositions.

Background Art

Antibodies are drawing attention as pharmaceuticals since they are highly stable in plasma and have few side effects. In particular, a number of IgG-type antibody pharmaceuticals are available on the market and many antibody pharmaceuticals are currently being developed (Non-patent Documents 1 and 2).

As cancer therapeutic agents using antibody pharmaceuticals, Rituxan against the CD20 antigen, Cetuximab against the EGFR antigen, Herceptin against the HER2 antigen, and such have been approved so far (Non-patent Document 3). These antibody molecules bind to antigens expressed on cancer cells, and exhibit cytotoxic activity against cancer cells through ADCC and such.

In contrast to such antibody pharmaceuticals that exhibit cytotoxic activities directly against cancer cells, Ipilimumab which is an IgG1-type antibody against CTLA4 was recently confirmed to be effective against metastatic melanoma, and received FDA approval in 2011. In solid cancer microenvironments, effector T cells that recognize tumors are known to be suppressed by various factors in the cancer microenvironment (Non-patent Document 4). Among them, CTLA4 which is an immune checkpoint molecule is known to have the function of suppressing T cell activation, and is strongly expressed in activated effector T cells and regulatory T cells. In cancer microenvironments, tumor-recognizing effector T cells are considered to be suppressed by CTLA4. Ipilimumab is a neutralizing antibody against CTLA4, and it is considered to exhibit antitumor effects by inhibiting the suppression of effector T cell-activation and activating the tumor-recognizing effector T cells (Non-patent Document 5). Similarly, nivolumab which is a neutralizing antibody against the immune checkpoint molecule PD1 has been recently reported to be also highly effective against metastatic melanoma by the activation of tumor-recognizing effector T cells that are suppressed by PD1 (Non-patent

Document 6). Furthermore, not only regulatory T cells but also exhausted T cells are known to be present in the cancer microenvironment, and the exhausted T cells are known to not only lack cytotoxic activity (antitumor activity) but also to contribute on immunosuppression in tumors (Non-patent Document 7).

Thus, it was thought that antibody pharmaceuticals against immune checkpoint molecules exert antitumor effects by inhibiting T-cell-suppressing effects of immune checkpoint molecules. However, it has been recently reported that for exhibiting antitumor effects of anti-CTLA4 antibodies such as ipilimumab, not only the neutralizing effects on CTLA4, but the antibody-dependent cytotoxic activity (ADCC activity) against CTLA4-expressing cells is also important (Non-patent Documents 8 and 9). Regulatory T cells and exhausted T cells in the tumor microenvironment are known to strongly express CTLA4, and anti-CTLA4 antibodies having ADCC activity are thought to be able to kill the regulatory T cells and exhausted T cells. In Non-patent Documents 8 and 9, while anti-CTLA4 antibodies having ADCC activity could eliminate regulatory T cells in tumors and exert strong antitumor effects in mice, anti-CTLA4 antibodies whose ADCC activity has been removed by amino acid substitutions could not eliminate regulatory T cells in tumors and could not exert antitumor effects.

Thus, it has been found that ADCC activity plays an important role in antitumor effects of anti-CTLA4 antibodies, and that the killing of regulatory T cells by ADCC activity is an important mechanism of action.

On the other hand, ADCC activity of an antibody is exhibited by binding of its Fc region to an Fcγ receptor. For example, substituting (modifying) amino acids of the Fc region of an IgG1 antibody to increase binding to the Fcγ receptor has been reported to enable enhancement of ADCC activity (Non-Patent Document 10). In fact, an anti-CD19 antibody with enhanced ADCC activity due to modification of the Fc region exerted stronger antitumor effects than natural anti-CD19 antibodies (Non-patent Document 10).

Accordingly, if ADCC activity of anti-CTLA4 antibodies can be enhanced by modifying their Fc-region, the anti-CTLA4 antibodies can eliminate regulatory T cells in tumors more powerfully, and may exhibit stronger antitumor effects.

Prior Art Documents

[Non-patent Documents]

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Summary of the Invention

[Problems to be solved by the Invention]

The present invention was achieved in view of the above circumstances. An objective of the present invention is to provide antigen-binding molecules that exert superior antitumor effects by damaging cells having immune response-suppressing functions, and pharmaceutical compositions comprising the antigen-binding molecule as an active ingredient, or methods for treating cancer using the pharmaceutical compositions.

[Means for Solving the Problems]

The present inventors discovered that antigen-binding molecules comprising a domain that binds to a molecule expressed on the surface of cells having immune response-suppressing functions and a domain that binds to a T-cell receptor (TCR) complex, damage cells having immune response-suppressing functions and exert more superior antitumor activity than

conventional antigen-binding molecules that bind to molecules expressed on the surfaces of regulatory T cells and exhausted T cells and also have ADCC activity. Furthermore, the present inventors discovered pharmaceutical compositions that can treat various carcinomas by having the antigen-binding molecule as an active ingredient.

More specifically, the present invention provides the following:

[1] an antigen-binding molecule that inhibits immune response-suppressing activity of a cell having an immune response-suppressing function, comprising:

(i) a domain that binds to a molecule expressed on the surface of a cell having an immune response-suppressing function; and

(ii) a T cell receptor complex-binding domain;

[2] the antigen-binding molecule of [1], wherein the cell having immune response-suppressing function is a regulatory T cell or an exhausted T cell;

[3] the antigen-binding molecule of [1], wherein the molecule expressed on the surface of a cell having an immune response-suppressing function is any molecule selected from among CTLA4, PD1, TIM3, LAG3, CD244 (2B4), CD160, GARP, OX40, CD137 (4-1BB), CD25, VISTA, BTLA, TNFR25, CD57, KLRG1, CCR2, CCR5, CCR6, CD39, CD73, CD4, CD18, CD49b, CD1d, CD5, CD21, TIM1, CD19, CD20, CD23, CD24, CD38, CD93, IgM, B220(CD45R), CD317, PD-L1, CD11b, Ly6G, ICAM-1, FAP, PDGFR, Podoplanin, and TIGIT;

[4] the antigen-binding molecule of [3], wherein the molecule expressed on the surface of a cell having an immune response-suppressing function is any molecule selected from among CTLA4, TIM3, LAG3, CD137 (4-1BB), CD25, CCR5, CCR6, CD38, TIGIT, and OX40;

[5] the antigen-binding molecule of any one of [1] to [4], wherein the T cell receptor complex-binding domain is a T cell receptor-binding domain;

[6] the antigen-binding molecule of any one of [1] to [4], wherein the T cell receptor complex-binding domain is a CD3-binding domain;

[7] the antigen-binding molecule of any one of [1] to [6], which further comprises an FcRn-binding domain;

[8] the antigen-binding molecule of [7], wherein the FcRn-binding domain is an Fc region of an antibody;

[9] the antigen-binding molecule of [8], wherein the FcRn-binding domain is an Fc region of an antibody having reduced Fcγ receptor-binding activity;

[10] the antigen-binding molecule of any one of [1] to [9], which is a multispecific antibody;

[11] a pharmaceutical composition comprising the antigen-binding molecule of any one of [1] to [10] as an active ingredient;

[12] a pharmaceutical composition for inhibiting an immune response-suppressing activity, comprising the antigen-binding molecule of any one of [1] to [10] as an active ingredient;

[13] the pharmaceutical composition of [12], wherein the inhibition of an immune response-suppressing activity is inhibition caused by a T cell;
[14] the pharmaceutical composition of [13], wherein the inhibition caused by a T cell is inhibition resulting from cytotoxic activity of the T cell;
[15] the pharmaceutical composition of any one of [12] to [14], wherein the immune response-suppressing activity is an activity by a regulatory T cell or an exhausted T cell;
[16] the pharmaceutical composition of any one of [12] to [14], wherein the immune response-suppressing activity is an activity of a cell expressing any molecule selected from among CTLA4, PD1, TIM3, LAG3, CD244 (2B4), CD160, GARP, OX40, CD137 (4-1BB), CD25, VISTA, BTLA, TNFR25, CD57, KLRG1, CCR2, CCR5, CCR6, CD39, CD73, CD4, CD18, CD49b, CD1d, CD5, CD21, TIM1, CD19, CD20, CD23, CD24, CD38, CD93, IgM, B220(CD45R), CD317, PD-L1, CD11b, Ly6G, ICAM-1, FAP, PDGFR, Podoplanin, and TIGIT;
[17] the pharmaceutical composition of any one of [11] to [16], which is a therapeutic agent for cancer;
[18] the pharmaceutical composition of claim 17, which is a therapeutic agent for any cancer selected from among ovarian cancer, gastric cancer, esophageal cancer, pancreatic cancer, renal cell carcinoma, hepatocellular carcinoma, breast cancer, malignant melanoma, non-small-cell lung cancer, cervical cancer, glioblastoma, prostate cancer, neuroblastoma, chronic lymphocytic leukemia, papillary thyroid cancer, colorectal cancer, Hodgkin's lymphoma, and endometriosis;
[19] a method for inducing cytotoxicity, suppressing cell proliferation, activating immunity against a cancer cell or a cancer cell-comprising tumor tissue , or treating or preventing cancer, which comprises the step of administering the antigen-binding molecule of any one of [1] to [10] or the pharmaceutical composition of any one of [11] to [18];
[20] the antigen-binding molecule of any one of [1] to [10] or the pharmaceutical composition of any one of [11] to [18] for use in inducing cytotoxicity, suppressing cell proliferation, activating immunity against a cancer cell or a cancer cell-comprising tumor tissue, or treating or preventing cancer;
[21] use of the antigen-binding molecule of any one of [1] to [10] in the production of the pharmaceutical composition of any one of [11] to [18]; and
[22] a method for producing the pharmaceutical composition of any one of [11] to [18], which comprises the step of using the antigen-binding molecule of any one of [1] to [10].

Furthermore, the present invention relates to methods for treating or preventing cancer, or for inhibiting suppression of an immune response, which comprise administering an antigen-binding molecule of the present invention or a pharmaceutical composition of the present invention to patients in need of treatment. The present invention also relates to a kit for use in the methods of the present invention, which comprises an antigen-binding molecule of the

present invention. The present invention also relates to the use of an antigen-binding molecule of the present invention in producing a pharmaceutical composition for inhibiting immune response-suppression. Furthermore, the present invention relates to antigen-binding molecules of the present invention or pharmaceutical compositions of the present invention for use in the methods of the present invention.

Brief Description of the Drawings

Fig. 1 presents a schematic diagram showing cytotoxic activity against cancer cells by bispecific antibodies that target a cancer antigen expressed on cancer cells and CD3 expressed on T cells.

Fig. 2-1 presents a schematic diagram showing crosslinking between T cells caused by crosslinking between CD3s on different T cells *via* an anti-CD3 antibody.

Fig. 2-2 presents a schematic diagram showing crosslinking between CD3s on the same T cell *via* an anti-CD3 antibody.

Fig. 3 presents a schematic diagram showing that an anti-CTLA4/CD3 bispecific antibody causes crosslinking between CTLA4 and CD3 on the same regulatory T cell, and therefore, does not cause intercellular crosslinking between an effector T cell and a regulatory T cell. This schematic diagram shows a regulatory T cell as the target cell and CTLA4 as the antigen, but the target cell and antigen are not limited thereto.

Fig. 4 presents a schematic diagram showing intercellular crosslinking caused by crosslinking between CD3 on an effector T cell and CTLA4 on a regulatory T cell *via* an anti-CTLA4/CD3 bispecific antibody. This schematic diagram shows a regulatory T cell as the target cell and CTLA4 as the antigen, but the target cell and antigen are not limited thereto.

Fig. 5 presents a graph showing the ADCC activity of the anti-mouse CTLA4 antibody hUH02hUL01-mFa55 on mouse CTLA4-expressing cells.

Fig. 6 presents a graph showing the cytotoxic activity of the anti-mouse CTLA4/anti-mouse CD3 bispecific antibody (hUH02UL01/2C11-F760) on mouse CTLA4-expressing cells.

Fig. 7-1 presents a schematic diagram showing crosslinking between a mouse CTLA4-bound bead and a mouse CD3-bound bead by an anti-mouse CTLA4/anti-mouse CD3 bispecific antibody.

Fig. 7-2 presents a graph showing the experiment results of crosslinking between mouse CTLA4-bound beads and mouse CD3-bound beads *via* anti-mouse CTLA4/anti-mouse CD3 bispecific antibodies (N = 3). The graph shows the mean values and standard deviations for each of the measurement results.

Fig. 7-3 presents a graph showing the experiment results of crosslinking between mouse

CTLA4/mouse CD3-bound beads and mouse CD3-bound beads by anti-mouse CTLA4/anti-mouse CD3 bispecific antibodies. The graph shows the mean values and standard deviations for each of the measurement results.

Fig. 7-4 presents a schematic diagram showing crosslinking between a mouse CTLA4/mouse CD3-bound bead and a mouse CD3-bound bead by an anti-mouse CTLA4/anti-mouse CD3 bispecific antibody.

Fig. 8 presents a graph showing the *in vivo* antitumor effects obtained by intratumoral (i.t.) administration of an anti-mouse CTLA4 antibody (hUH02hUL01-mFa55, ADCC-enhanced antibody) and an anti-mouse CTLA4/anti-mouse CD3 bispecific antibody (hUH02UL01/2C11-F760, a T cell-redirecting antibody) on mouse colorectal cancer cell line CT26.WT (n = 2 for each group; the plot shows the tumor volume of each individual).

Fig. 9 presents a graph showing the *in vivo* antitumor effects obtained by intratumoral (i.t.) administration and intravenous (i.v.) administration of an anti-mouse CTLA4/anti-mouse CD3 bispecific antibody (hUH02UL01/2C11-F760, a T cell-redirecting antibody) on mouse colorectal cancer cell line CT26.WT (n = 5 for each group; the plot shows the mean tumor volume + standard deviation of each group).

Fig. 10-1 presents graphs showing the results of analyzing CD4-positive cells based on the expression of CD25 and CD45RA, after a seven-day reaction of PBMC derived from a healthy person with a control antibody (control) and an anti-human CTLA4/anti-human CD3 bispecific antibody (TRAB).

Fig. 10-2 presents graphs showing the proportion of regulatory T cells (Treg) in CD4-positive T cells, calculated based on the expression of CD25 and CD45RA, after a seven-day reaction of PBMC derived from a healthy person with a control antibody (control) and an anti-human CTLA4/anti-human CD3 bispecific antibody (TRAB).

Fig. 10-3 presents graphs showing the ratio of effector T cells (Teff) to regulatory T cells (Treg) (Teff/Treg) calculated based on the expression of CD25 and CD45RA, after a seven-day reaction of PBMC derived from a healthy person with a control antibody (control) and an anti-human CTLA4/anti-human CD3 bispecific antibody (TRAB).

Fig. 11-1 presents graphs showing the results of analyzing CD4-positive cells based on the expression of FoxP3 and CD45RA, after a seven-day reaction of PBMC derived from a healthy person with a control antibody (control) and an anti-human CTLA4/anti-human CD3 bispecific antibody (TRAB).

Fig. 11-2 presents graphs showing the proportion of regulatory T cells (Treg) in CD4-positive T cells, calculated based on the expression of FoxP3 and CD45RA, after a seven-day reaction of PBMC derived from a healthy person with a control antibody (control) and an anti-human CTLA4/anti-human CD3 bispecific antibody (TRAB).

Fig. 11-3 presents graphs showing the ratio of effector T cells (Teff) to regulatory T cells (Treg) (Teff/Treg) calculated based on the expression of FoxP3 and CD45RA, after a seven-day reaction of PBMC derived from a healthy person with a control antibody (control) and an anti-human CTLA4/anti-human CD3 bispecific antibody (TRAB).

Fig. 12 presents a diagram showing the relationship between the amino acid residues constituting the Fc regions of IgG1, IgG2, IgG3, and IgG4, and Kabat's EU numbering (herein, it is also called the EU INDEX).

Fig. 13 presents graphs showing the results of analyzing CD4-positive cells based on the expression of CD25 and CD45RA, after a four- or six-day reaction of PBMC derived from a healthy person with an anti-human LAG3/anti-human CD3 bispecific antibody (TRAB).

Fig. 14 presents graphs showing the proportion of regulatory T cells (Treg) in CD4-positive T cells, calculated based on the expression of CD25 and CD45RA, after a four- or six-day reaction of PBMC derived from a healthy person with an anti-human LAG3/anti-human CD3 bispecific antibody (TRAB).

Fig. 15 presents graphs showing the ratio of effector T cells (Teff) to regulatory T cells (Treg) (Teff/Treg) in CD4-positive T cells, calculated based on the expression of CD25 and CD45RA, after a four- or six-day reaction of PBMC derived from a healthy person with an anti-human LAG3/anti-human CD3 bispecific antibody (TRAB).

Fig. 16 presents graphs showing the results of analyzing CD4-positive cells based on the expression of CD25 and CD45RA, after a seven-day reaction of PBMC derived from a healthy person with an anti-human OX40/anti-human CD3 bispecific antibody (TRAB).

Fig. 17 presents graphs showing the proportion of regulatory T cells (Treg) in CD4-positive T cells, calculated based on the expression of CD25 and CD45RA, after a seven day reaction of PBMC derived from a healthy person with an anti-human OX40/anti-human CD3 bispecific antibody (TRAB).

Fig. 18 presents graphs showing the ratio of effector T cells (Teff) to regulatory T cells (Treg) (Teff/Treg) in CD4-positive T cells, calculated based on the expression of CD25 and CD45RA, after a seven day reaction of PBMC derived from a healthy person with an anti-human OX40/anti-human CD3 bispecific antibody (TRAB).

Mode for Carrying Out the Invention

The following definitions are provided to facilitate understanding of the invention described herein.

Antigen-binding molecules

In the present invention, "antigen-binding molecules" are not particularly limited as

long as they are molecules that comprise a "binding domain" of the present invention, and they may further comprise a peptide or protein having a length of about five amino acids or more. The peptide and protein are not limited to those derived from a living organism, and for example, they may be a polypeptide produced from an artificially designed sequence. They may also be any of a naturally-occurring polypeptide, synthetic polypeptide, recombinant polypeptide, and such.

A favorable example of an antigen-binding molecule of the present invention is an antigen-binding molecule that comprises an FcRn-binding domain contained in an antibody Fc region. As a method for extending the blood half-life of a protein administered to a living body, the method of adding an FcRn-binding domain of an antibody to the protein of interest and utilizing the function of FcRn-mediated recycling is well known.

In the present invention, the "FcRn-binding domain" is not particularly limited as long as it has binding activity to FcRn, and it may be a domain that directly or indirectly binds to FcRn. Examples of the domain that directly binds to FcRn include antibody variable regions, Fab and antibody Fc regions whose antigens are FcRn, fragments thereof, albumin, albumin domain 3, human serum albumin (HSA), transferrin and such. Furthermore, an example of the domain that indirectly binds to FcRn includes a domain that has binding activity toward the aforementioned domain that directly binds to FcRn. A preferred embodiment of the present invention includes antibody Fc regions or fragments containing an FcRn-binding region of an Fc region. Herein, for example, an Fc region derived from a naturally-occurring IgG may be used as the "Fc region". A naturally-occurring IgG means a polypeptide that comprises the same amino acid sequence as an IgG found in nature, and belongs to a class of antibodies substantially encoded by immunoglobulin gamma genes. A naturally-occurring human IgG means, for example, a naturally-occurring human IgG1, a naturally-occurring human IgG2, a naturally-occurring human IgG3, or a naturally-occurring human IgG4. Naturally-occurring IgGs also include mutants and such that naturally generate therefrom. A plurality of allotype sequences that result from genetic polymorphism have been described in Sequences of Proteins of Immunological Interest, NIH Publication No. 91-3242 for the human IgG1, human IgG2, human IgG3, and human IgG4 antibody constant region, and any of the sequences may be used in the present invention. In particular, the amino acid sequence of positions 356 to 358 according to EU numbering may be DEL or EEM for the human IgG1 sequence.

Existing antibody Fc regions are, for example, IgA1, IgA2, IgD, IgE, IgG1, IgG2, IgG3, IgG4, and IgM-type Fc regions. For example, an Fc region derived from a naturally-occurring human IgG antibody can be used as the antibody Fc region of the present invention. Fc regions derived from a constant region of a naturally-occurring IgG, or more specifically, a constant region derived from a naturally-occurring human IgG1 (SEQ ID NO: 1), a constant region

derived from a naturally-occurring human IgG2 (SEQ ID NO: 2), a constant region derived from a naturally-occurring human IgG3 (SEQ ID NO: 3), and a constant region derived from a naturally-occurring human IgG4 (SEQ ID NO: 4), can be used as an Fc region of the present invention. Mutants and such that naturally generate therefrom are also included in the naturally-occurring IgG constant regions.

Such antibody Fc regions can be suitably obtained, for example, by partial digestion of antibodies such as monoclonal antibodies using a protease such as pepsin, then adsorption of the resulting fragments onto a protein A column or a protein G column, and subsequent elution using an appropriate elution buffer and such. The protease is not particularly limited as long as it can digest an antibody such as a monoclonal antibody by appropriately establishing the enzyme reaction conditions such as pH, and examples include pepsin and ficin.

The isotype of an antibody is determined by the structure of the constant region. The constant region of isotypes IgG1, IgG2, IgG3, and IgG4 is called Cγ1, Cγ2, Cγ3, and Cγ4, respectively. The amino acid sequences of polypeptides constituting the Fc regions of human Cγ1, Cγ2, Cγ3, and Cγ4 are exemplified in SEQ ID NOs: 5, 6, 7, and 8. The relationship between amino acid residues constituting each of these amino acid sequences and Kabat's EU numbering (herein, also referred to as EU INDEX) is shown in Fig. 12.

An Fc region refers to a region that excludes $F(ab')_2$ which contains two light chains and two heavy chains containing part of the constant region between the CH1 domain and the CH2 domain such that the disulfide bonds between the chains are formed between the two heavy chains. Fc regions forming the antigen-binding molecules disclosed herein can be obtained suitably by partially digesting the IgG1, IgG2, IgG3, or IgG4 monoclonal antibodies or the like using a protease such as pepsin, and then re-eluting fractions adsorbed to the protein A column. The protease is not particularly limited as long as it can digest a full-length antibody in a restrictive manner to produce $F(ab')_2$ by appropriately establishing the enzyme reaction conditions such as pH. Such proteases include, for example, pepsin and ficin.

A domain with decreased Fcγ receptor-binding activity is particularly preferred as the FcRn-binding domain of the present invention. Here, an Fcγ receptor (herein, also denoted as Fcγ receptor, FcγR, or FcgR) refers to a receptor that can bind to the Fc region of IgG1, IgG2, IgG3, or IgG4, and includes all members belonging to the family of proteins substantially encoded by Fcγ receptor genes. In humans, this family includes, but is not limited to, FcγRI (CD64) including isoforms FcγRIa, FcγRIb, and FcγRIc; FcγRII (CD32) including isoforms FcγRIIa (including allotypes H131 (type H) and R131 (type R), FcγRIIb (including FcγRIIb-1 and FcγRIIb-2), and FcγRIIc; and FcγRIII (CD16) including isoforms FcγRIIIa (including allotypes V158 and F158) and FcγRIIIb (including allotypes FcγRIIIb-NA1 and FcγRIIIb-NA2); as well as any undiscovered human FcγRs, and FcγR isoforms or allotypes. FcγRs include, but

are not limited to, those derived from humans, mice, rats, rabbits, and monkeys, and may be derived from any organism. Mouse FcγRs include, but are not limited to, FcγRI (CD64), FcγRII (CD32), FcγRIII (CD16), and FcγRIII-2 (CD16-2), as well as any undiscovered mouse FcγRs, and FcγR isoforms or allotypes. Suitable examples of such Fcγ receptors include human FcγRI (CD64), FcγRIIa (CD32), FcγRIIb (CD32), FcγRIIIa (CD16) and/or FcγRIIIb (CD16).

Activating receptors which carry an immunoreceptor tyrosine-based activation motif (ITAM) and inhibitory receptors which carry an immunoreceptor tyrosine-based inhibitory motif (ITIM) are present among FcγRs. FcγRs are categorized into activating FcγRs: FcγRI, FcγRIIa R, FcγRIIa H, FcγRIIIa, and FcγRIIIb, and inhibitory FcγR: FcγRIIb.

The polynucleotide sequence and amino acid sequence of FcγRI are shown in NM_000566.3 and NP_000557.1, respectively; the polynucleotide sequence and amino acid sequence of FcγRIIa are shown in BC020823.1 and AAH20823.1, respectively; the polynucleotide sequence and amino acid sequence of FcγRIIb are shown in BC146678.1 and AAI46679.1, respectively; the polynucleotide sequence and amino acid sequence of FcγRIIIa are shown in BC033678.1 and AAH33678.1, respectively; and the polynucleotide sequence and amino acid sequence of FcγRIIIb are shown in BC128562.1 and AAI28563.1, respectively (RefSeq accession number). There are two types of gene polymorphisms for FcγRIIa, where the amino acid at position 131 of FcγRIIa is substituted into histidine (type H) or arginine (type R) (J. Exp. Med, 172, 19-25, 1990). Furthermore, there are two types of gene polymorphisms for FcγRIIb, where the amino acid at position 232 of FcγRIIb is substituted with isoleucine (type I) or threonine (type T) (Arthritis. Rheum. 46: 1242-1254 (2002)). In addition, there are two types of gene polymorphisms for FcγRIIIa, where the amino acid at position 158 of FcγRIIIa is substituted with valine (type V) or phenylalanine (type F) (J. Clin. Invest. 100(5): 1059-1070 (1997)). There are also two types of gene polymorphisms for FcγRIIIb, which are type NA1 and type NA2 (J. Clin. Invest. 85: 1287-1295 (1990)).

Whether the binding activity to an Fcγ receptor is decreased can be confirmed by well-known methods such as FACS, ELISA format, screening by Amplified Luminescent Proximity Homogeneous Assay (ALPHA), surface plasmon resonance (SPR)-based BIACORE method, and others (Proc. Natl. Acad. Sci. USA (2006) 103(11), 4005-4010).

ALPHA screening is performed with ALPHA technology which uses two beads, a donor and an acceptor bead, based on the following principle. Luminescent signals are detected only when molecules bound to donor beads interact biologically with molecules bound to the acceptor beads, and the two beads are in close proximity to each other. The laser-excited photosensitizer within the donor beads converts ambient oxygen to excited-state singlet oxygen. Singlet oxygen is dispersed around the donor beads; and when it reaches the adjacent acceptor beads, a chemiluminescent reaction is induced within the beads, and light is ultimately emitted. When

molecules bound to the donor beads do not interact with molecules bound to the acceptor beads, the chemiluminescent reaction does not take place because singlet oxygen produced by the donor beads does not reach the acceptor beads.

For example, when an antigen-binding molecule contains an antibody Fc region as the FcRn-binding domain, an antigen-binding molecule having a wild-type Fc region and an antigen-binding molecule having a mutant Fc region produced by adding amino acid mutations to change the binding to an Fcγ receptor are prepared, a biotinylated antigen-binding molecule is bound to the donor beads, and an Fcγ receptor tagged with glutathione S transferase (GST) is bond to the acceptor beads. In the presence of an antigen-binding molecule having a mutant Fc region, the antigen-binding molecule having a wild-type Fc region interacts with the Fcγ receptor and produces 520-620 nm signals. When the antigen-binding molecule having a mutant Fc region is untagged, it competes with the antigen-binding molecule having a wild-type Fc region for interaction with the Fcγ receptor. The relative binding affinity can be determined by quantifying the decrease in fluorescence observed as a result of the competition. Biotinylation of antigen-binding molecules using Sulfo-NHS-biotin and such is well known. As a method for tagging an Fcγ receptor with GST, the method of expressing the Fcγ receptor and GST in a cell carrying a vector that can express a fusion gene produced by fusing a polynucleotide encoding the Fcγ receptor in frame with a GST-encoding polynucleotide, and purifying it using a glutathione column can be appropriately adopted. The obtained signals are suitably analyzed, for example, by fitting them into a one-site competition model that utilizes a non-linear regression analysis with software such as GRAPHPAD PRISM (GraphPad, San Diego).

One of the substances (ligand) observed for interaction is immobilized onto a gold thin film on a sensor chip, and by shining light from the reverse side of the sensor chip so that total reflection takes place at the interface between the gold thin film and glass, a portion with reduced reflection intensity is formed in part of the reflected light (SPR signal). The other substance (analyte) observed for interaction is made to flow over the sensor chip surface; and when the ligand binds to the analyte, the mass of the immobilized ligand molecule increases and the refractive index of the solvent on the sensor chip surface changes. The position of the SPR signal shifts as a result of this change in the refractive index (reversely, the signal position returns if this binding dissociates). The Biacore system shows the amount of shift mentioned above, or more specifically the time variable of mass, by plotting the change in mass on the sensor chip surface on the vertical axis as the measurement data (sensorgram). Kinetic parameters such as association rate constant (ka) and dissociation rate constant (kd) are determined from the curve in the sensorgram, and the affinity (KD) is determined from the ratio of these constants. In the BIACORE method, a method for measuring inhibition is also suitably used. An example of the method for measuring inhibition is described in Proc. Natl. Acad. Sci

USA (2006) 103 (11): 4005-4010.

Herein, "decreased Fcγ receptor-binding activity" means that, for example, based on the above-described analytical method, the binding activity of the test antigen-binding molecule is 50% or less, preferably 45% or less, 40% or less, 35% or less, 30% or less, 20% or less, 15% or less, or particularly preferably 10% or less, 9% or less, 8% or less, 7% or less, 6% or less, 5% or less, 4% or less, 3% or less, 2% or less, or 1% or less as compared to the binding activity of the control antigen-binding molecule containing an Fc region.

For the control antigen-binding molecule, antigen-binding molecules having, for example, a domain comprising an Fc region of a monoclonal IgG1, IgG2, IgG3, or IgG4 antibody may be suitably used. The structures of the Fc regions are shown in SEQ ID NO: 1 (A is added to the N terminus of RefSeq Accession No. AAC82527.1), SEQ ID NO: 2 (A is added to the N terminus of RefSeq Accession No. AAB59393.1), SEQ ID NO: 3 (A is added to the N terminus of RefSeq Accession No. CAA27268.1), and SEQ ID NO: 4 (A is added to the N terminus of RefSeq Accession No. AAB59394.1). Further, when an antigen-binding molecule containing a mutant of an Fc region of a particular antibody isotype is used as the test substance, the effect of a mutation possessed by the mutant on the Fcγ receptor-binding activity is tested by using as a control an antigen-binding molecule having an Fc region of an antibody of that particular isotype. In this way, antigen-binding molecules containing an Fc region mutant whose binding activity toward the Fcγ receptor verified to be decreased are suitably produced.

Examples of such mutants include mutants with a 231A-238S deletion (WO 2009/011941), or C226S, C229S, P238S, (C220S) (J. Rheumatol (2007) 34, 11), C226S, C229S (Hum. Antibod. Hybridomas (1990) 1(1), 47-54), C226S, C229S, E233P, L234V, or L235A (Blood (2007) 109, 1185-1192) mutants, where the amino acids are specified by EU numbering.

That is, suitable examples include antigen-binding molecules having an Fc region in which any of the amino acids at positions 220, 226, 229, 231, 232, 233, 234, 235, 236, 237, 238, 239, 240, 264, 265, 266, 267, 269, 270, 295, 296, 297, 298, 299, 300, 325, 327, 328, 329, 330, 331, and 332 specified according to EU numbering has been substituted in the amino acids constituting the Fc region of an antibody of a specific isotype. The isotype of the antibody from which the Fc region originates is not particularly limited, and the Fc region derived from an IgG1, IgG2, IgG3, or IgG4 monoclonal antibody can be used appropriately, and the Fc region derived from a naturally-occurring human IgG1 antibody is suitably used.

For example, an antigen-binding molecule having an Fc region that comprises any substitution specified below based on EU numbering from among amino acids constituting the IgG1 antibody Fc region (wherein the number indicates the position of the amino acid residue specified according to EU numbering, the one-letter amino acid code positioned before the number indicates the amino acid residue before the substitution, and the one-letter amino acid

code positioned after the number indicates the amino acid residue after the substitution):
(a) L234F, L235E, P331S
(b) C226S, C229S, P238S
(c) C226S, C229S
(d) C226S, C229S, E233P, L234V, L235A;
or an Fc region lacking the amino acid sequence of positions 231 to 238 from among amino acids constituting the IgG1 antibody Fc region may be appropriately used.

Furthermore, antigen-binding molecules having an Fc region that comprises any substitution specified below based on EU numbering from among amino acids constituting the IgG2 antibody Fc region (wherein the number indicates the position of the amino acid residue specified according to EU numbering, the one-letter amino acid code positioned before the number indicates the amino acid residue before the substitution, and the one-letter amino acid code positioned after the number indicates the amino acid residue after the substitution):
(e) H268Q, V309L, A330S, P331S
(f) V234A
(g) G237A
(h) V234A, G237A
(i) A235E, G237A
(j) V234A, A235E, G237A
may be appropriately used.

Furthermore, antigen-binding molecules having an Fc region that comprises any substitution specified below based on EU numbering from among amino acids constituting the IgG3 antibody Fc region (wherein the number indicates the position of the amino acid residue specified according to EU numbering, the one-letter amino acid code positioned before the number indicates the amino acid residue before the substitution, and the one-letter amino acid code positioned after the number indicates the amino acid residue after the substitution):
(k) F241A
(l) D265A
(m) V264A
may be appropriately used.

Furthermore, antigen-binding molecules having an Fc region that comprises any substitution specified below based on EU numbering from among amino acids constituting the IgG4 antibody Fc region (wherein the number indicates the position of the amino acid residue specified according to EU numbering, the one-letter amino acid code positioned before the number indicates the amino acid residue before the substitution, and the one-letter amino acid code positioned after the number indicates the amino acid residue after the substitution):

(n) L235A, G237A, E318A
(o) L235E
(p) F234A, L235A
may be appropriately used.

Other preferred examples include antigen-binding molecules having an Fc region in which any of the amino acids at positions 233, 234, 235, 236, 237, 327, 330, and 331 specified according to EU numbering in the amino acids constituting the Fc region of a naturally-occurring human IgG1 antibody is substituted with amino acids of corresponding EU numbering in the corresponding IgG2 or IgG4.

Other preferred examples suitably include antigen-binding molecules having an Fc region in which any one or more of the amino acids at positions 234, 235, and 297 specified according to EU numbering in the amino acids constituting the Fc region of a naturally-occurring human IgG1 antibody are substituted by other amino acids. The type of amino acid present after substitution is not particularly limited, and an antigen-binding molecule having an Fc region in which any one or more of the amino acids at positions 234, 235, and 297 are substituted with alanine is particularly preferred.

Other preferred examples suitably include antigen-binding molecules having an Fc region in which the amino acid at position 265 specified according to EU numbering in the amino acids constituting an IgG1 antibody Fc region is substituted by another amino acid. The type of amino acid present after substitution is not particularly limited, and an antigen-binding molecule having an Fc region in which the amino acid at position 265 is substituted with alanine is particularly preferred.

"A domain that binds to a molecule expressed on the surface of a cell having immune response-suppressing function" and "a T cell receptor complex-binding domain" (hereinafter, these binding domains are collectively referred to as antigen-binding domains) which are included in antigen-binding molecules of the present invention mean regions that specifically bind to all or a portion of their respective antigens. An example of such binding domain is a region comprising the antigen-binding region of an antibody. When the molecular weight of the antigen is large, the antigen-binding region of the antibody can bind only to a specific portion of the antigen. This specific portion is called an epitope. One or more of antibody variable domains may provide an antigen-binding domain. Preferably, an antigen-binding domain comprises an antibody light chain variable region (VL) and an antibody heavy chain variable region (VH). Suitable examples of such antigen-binding domains include "single chain Fv (scFv)", "single chain antibody", "Fv", "single chain Fv2 (scFv2)", "Fab", "$F(ab')_2$", or such.

Here, "a cell having immune response-suppressing function" is not particularly limited as long as it has a function of suppressing an immune response, and examples include regulatory

T cells (Treg), exhausted T cells, myeloma-derived stromal cells (MDSC), tumor-associated macrophages (TAM), induced regulatory T cells (Tr1), tumor-associated dendritic cells (TADC), tumor-associated neutrophils (TAN), cancer-associated fibroblasts (CAF), regulatory B cells (Breg), and such. In particular, regulatory T cells and exhausted T cells are preferable as the cells of interest. Specific examples of the molecules expressed on the surface of such cells having immune response-suppressing functions include CTLA4, PD1, TIM3, LAG-3, CD244 (2B4), CD160, GARP, OX40, CD137 (4-1BB), CD25, VISTA, VISATA, BTLA, TNFR25, CD57, KLRG1, CCR2, CCR5, CCR6, CD39, CD73, CD4, CD18, CD49b, CD1d, CD5, CD21, TIM1, CD19, CD20, CD23, CD24, CD38, CD93, IgM, B220(CD45R), CD317, PD-L1, CD11b, Ly6G, ICAM-1, FAP, PDGFR, Podoplanin, TIGIT, and such. Among these molecules, examples of favorable molecules for targets of the binding domains of the present invention include CTLA4, TIM3, LAG3, CD137 (4-1BB), CD25, CCR5, CCR6, CD38, and TIGIT, which are cell surface molecules specifically expressed in cell fractions ($CD4^+$, $CD25^{high}$, and $CD45RA^-$) that have been reported to have high immune response-suppressing functions. Examples of favorable molecules for targets of the binding domains of the present invention include CTLA4, LAG3, and OX40 in particular.

In the present invention, “regulatory T cell” means a type of T cell in charge of inhibitory regulation of an immune response. This cell plays an important role in the negative regulation mechanism for suppressing excessive immune responses and in homeostasis of immunity, and is classified into two types of regulatory T cells (Tregs) which express CD4 or CD8. CD4 Tregs are classified into endogenous Treg cells (natural Tregs or nTregs) which constitutively express CD25 and FoxP3, and adaptive or inducible Tregs (iTregs) which have low self-recognition ability and which are differentiated from naive CD4-positive T cells. Existing iTregs include $Foxp3^+$ Treg and $Fop3^-$ Treg, and $Fop3^-$ Treg is called a Type I Treg (Tr1). $CD4^+CD25^+LAG3^+$ Treg has been identified as a Treg having properties very similar to Tr1 (Proc Natl Acad Sci USA.2009). Furthermore, CD127 expression is known to be decreased in Treg cells, and the fraction containing $CD127^{lo}CD25^{hi\text{-}int}$ (population showing low CD127 expression and high to intermediate CD25 expression) includes all of the Foxp3-positive Treg population. CD8 Tregs can also be separated into endogenous Tregs and inducible Tregs. The former are classified as $CD8^+CD122^+$ Treg, and the latter are classified as Qa-1a-restricted $CD8^+$ Treg. Tregs are known to express regulatory molecules, and have increased expression levels of CTLA4, PD1, TIM3, LAG3, and the like, and such molecules are favorable as the molecules to which the binding domains of the antigen-binding molecules of the present invention bind.

Furthermore, in the present invention, “exhausted T cell” means a T cell whose cytokine production function and effector function have been markedly weakened by continuous stimulation by antigens, and proliferation ability and long-term survival ability have become low.

These exhausted T cells produce regulatory receptors such as PD1 and regulatory cytokines; therefore, they not only become dysfunctional, but also act suppressive towards immune responses. In exhausted T cells, expression of PD1 is mainly increased (Nature 439, 682-687 (2006)). In addition to PD1, expression of molecules such as LAG-3, CD244 (2B4), CD160, TIM-3, and CTLA-4 are also increased (Nature Immunology Volume: 12, Pages: 492-499 Year published: (2011)). These molecules are favorable as the molecules to which the binding domains of the antigen-binding molecules of the present invention bind.

Furthermore, the "T cell-receptor complex" may be a T cell receptor itself, or an adaptor molecule constituting a T cell-receptor complex together with a T cell receptor. CD3 is suitable as an adaptor molecule.

For the T cell receptor, an epitope to which the T cell receptor binding domain binds may be a variable region or a constant region, but an epitope present in the constant region is preferred. Examples of the constant region sequence include the T cell receptor α chain of RefSeq Accession No. CAA26636.1 (SEQ ID NO: 9), the T cell receptor β chain of RefSeq Accession No. C25777 (SEQ ID NO: 10), the T cell receptor γ1 chain of RefSeq Accession No. A26659 (SEQ ID NO: 11), the T cell receptor γ2 chain of RefSeq Accession No. AAB63312.1 (SEQ ID NO: 12), and the T cell receptor δ chain of RefSeq Accession No. AAA61033.1 (SEQ ID NO: 13).

In the present invention, when the "CD3-binding domain" is used as the T cell receptor complex-binding domain, the CD3-binding domain may be provided by one or more antibody variable domains. Preferably, the CD3-binding domain includes a light chain variable region (VL) and a heavy chain variable region (VH) of the CD3 antibody. Suitable examples of such CD3-binding domains include "single chain Fv (scFv)", "single chain antibody", "Fv", "single chain Fv 2 (scFv2)", "Fab", "$F(ab')_2$", and such.

The CD3-binding domain of the present invention may be those that bind to any epitope as long as the epitope exists in the γ-chain, δ-chain, or ε-chain sequence constituting human CD3. In the present invention, preferably, a CD3-binding domain that comprises a light chain variable region (VL) of a CD3 antibody and a heavy chain variable region (VH) of a CD3 antibody, and which binds to an epitope present in the extracellular region of the ε chain of the human CD3 complex, is suitably used. For such CD3-binding domain, a CD3-binding domain comprising the light chain variable region (VL) and heavy chain variable region (VH) of the OKT3 antibody (Proc. Natl. Acad. Sci. USA (1980) 77, 4914-4917) or various known CD3 antibodies is suitably used. A CD3-binding domain derived from a CD3 antibody that has the desired properties and is obtained by immunizing a desired animal with the γ-chain, δ-chain, or ε-chain constituting the human CD3 by the above-mentioned method may be appropriately used. Human antibodies and appropriately humanized antibodies as described below may be suitably used as the CD3

antibody that serves as the origin for the CD3-binding domain. For the structure of the CD3-constituting γ-chain, δ-chain, or ε-chain, their polynucleotide sequences are shown in SEQ ID NOs: 14 (NM_000073.2), 16 (NM_000732.4), and 18 (NM_000733.3), and their polypeptide sequences are shown in SEQ ID NOs: 15 (NP_000064.1), 17 (NP_000723.1), and 19 (NP_000724.1) (the RefSeq accession number is shown in parentheses).

A preferred embodiment of the "antigen-binding molecule" of the present invention includes an antibody comprising an antibody variable region of the present invention.

Antibody

Herein, an "antibody" refers to a naturally occurring immunoglobulin or an immunoglobulin produced by partial or complete synthesis. Antibodies can be isolated from natural sources such as naturally-occurring plasma and serum, or culture supernatants of antibody-producing hybridoma cells. Alternatively, antibodies can be partially or completely synthesized using techniques such as genetic recombination. Suitable examples of the antibodies include antibodies of an immunoglobulin isotype or subclass of such isotype. Known human immunoglobulins include those of the following nine classes (isotypes): IgG1, IgG2, IgG3, IgG4, IgA1, IgA2, IgD, IgE, and IgM. Of these isotypes, antibodies of the present invention include IgG1, IgG2, IgG3, and IgG4.

Methods for producing antibodies having the desired binding activity are known to those skilled in the art, and the antibodies may be obtained as polyclonal or monoclonal antibodies. Monoclonal antibodies derived from mammals may be suitably produced as the antibodies of the present invention. Such mammalian-derived monoclonal antibodies include antibodies produced by hybridomas and antibodies produced by host cells transformed with an expression vector carrying an antibody gene by genetic engineering techniques.

There is no particular limitation on the mammal to be immunized for obtaining antibodies. It is preferable to select the mammal by considering its compatibility with the parent cells to be used in cell fusion for hybridoma production. In general, rabbits, monkeys, and rodents such as mice, rats, and hamsters are suitably used.

The above animals are immunized with a sensitizing antigen by known methods. Generally performed immunization methods include, for example, intraperitoneal or subcutaneous injection of a sensitizing antigen into mammals. Specifically, a sensitizing antigen is appropriately diluted with Phosphate-Buffered Saline (PBS), physiological saline, or the like. If desired, a conventional adjuvant such as Freund's complete adjuvant is mixed with the antigen, and the mixture is emulsified. Then, the sensitizing antigen is administered to a mammal several times at 4- to 21-day intervals. Appropriate carriers may be used in immunization with the sensitizing antigen. In particular, when a low-molecular-weight partial

peptide is used as the sensitizing antigen, it is sometimes desirable to couple the sensitizing antigen peptide to a carrier protein such as albumin or keyhole limpet hemocyanin for immunization.

Alternatively, hybridomas producing a desired antibody can be prepared using DNA immunization as mentioned below. DNA immunization is an immunization method that confers immunostimulation by expressing a sensitizing antigen in an animal immunized as a result of administering a vector DNA constructed to allow expression of an antigen protein-encoding gene in the animal. As compared to conventional immunization methods in which a protein antigen is administered to animals to be immunized, DNA immunization is expected to be superior in that:

- immunostimulation can be provided while retaining the structure of a membrane protein; and
- there is no need to purify the antigen for immunization.

In order to prepare a monoclonal antibody of the present invention using DNA immunization, first, a DNA expressing an antigen protein is administered to an animal to be immunized. The antigen protein-encoding DNA can be synthesized by known methods such as PCR. The obtained DNA is inserted into an appropriate expression vector, and then this is administered to an animal to be immunized. Preferably used expression vectors include, for example, commercially-available expression vectors such as pcDNA3.1. Vectors can be administered to an organism using conventional methods. For example, DNA immunization is performed by using a gene gun to introduce expression vector-coated gold particles into cells in the body of an animal to be immunized.

After immunizing a mammal as described above, an increase in the titer of an antigen-binding antibody is confirmed in the serum. Then, immune cells are collected from the mammal, and then subjected to cell fusion. In particular, splenocytes are preferably used as immune cells.

A mammalian myeloma cell is used as a cell to be fused with the above-mentioned immune cells. The myeloma cells preferably comprise a suitable selection marker for screening. A selection marker confers characteristics to cells for their survival (or death) under a specific culture condition. Hypoxanthine-guanine phosphoribosyltransferase deficiency (hereinafter abbreviated as HGPRT deficiency) and thymidine kinase deficiency (hereinafter abbreviated as TK deficiency) are known as selection markers. Cells with HGPRT or TK deficiency have hypoxanthine-aminopterin-thymidine sensitivity (hereinafter abbreviated as HAT sensitivity). HAT-sensitive cells cannot synthesize DNA in a HAT selection medium, and are thus killed. However, when the cells are fused with normal cells, they can continue DNA synthesis using the salvage pathway of the normal cells, and therefore they can grow even in the HAT selection medium.

HGPRT-deficient and TK-deficient cells can be selected in a medium containing 6-thioguanine, 8-azaguanine (hereinafter abbreviated as 8AG), or 5'-bromodeoxyuridine. Normal cells are killed because they incorporate these pyrimidine analogs into their DNA. Meanwhile, cells that are deficient in these enzymes can survive in the selection medium, since they cannot incorporate these pyrimidine analogs. In addition, a selection marker referred to as G418 resistance provided by the neomycin-resistant gene confers resistance to 2-deoxystreptamine antibiotics (gentamycin analogs). Various types of myeloma cells that are suitable for cell fusion are known.

For example, myeloma cells including the following cells can be preferably used:
P3(P3x63Ag8.653) (J. Immunol. (1979) 123 (4), 1548-1550);
P3x63Ag8U.1 (Current Topics in Microbiology and Immunology (1978)81, 1-7);
NS-1 (C. Eur. J. Immunol. (1976)6 (7), 511-519);
MPC-11 (Cell (1976) 8 (3), 405-415);
SP2/0 (Nature (1978) 276 (5685), 269-270);
FO (J. Immunol. Methods (1980) 35 (1-2), 1-21);
S194/5.XX0.BU.1 (J. Exp. Med. (1978) 148 (1), 313-323);
R210 (Nature (1979) 277 (5692), 131-133), etc.

Cell fusions between the immunocytes and myeloma cells are essentially carried out using known methods, for example, a method by Kohler and Milstein *et al.* (Methods Enzymol. (1981) 73: 3-46).

More specifically, cell fusion can be carried out, for example, in a conventional culture medium in the presence of a cell fusion-promoting agent. The fusion-promoting agents include, for example, polyethylene glycol (PEG) and Sendai virus (HVJ). If required, an auxiliary substance such as dimethyl sulfoxide is also added to improve fusion efficiency.

The ratio of immunocytes to myeloma cells may be arbitrarily set, preferably, for example, one myeloma cell for every one to ten immunocytes. Culture media to be used for cell fusions include, for example, media that are suitable for the growth of myeloma cell lines, such as RPMI1640 medium and MEM medium, and other conventional culture medium used for this type of cell culture. In addition, serum supplements such as fetal calf serum (FCS) may be preferably added to the culture medium.

For cell fusion, predetermined amounts of the above immune cells and myeloma cells are mixed well in the above culture medium. Then, a PEG solution (for example, the average molecular weight is about 1,000 to 6,000) prewarmed to about 37°C is added thereto at a concentration of generally 30% to 60% (w/v). The mixed solution is gently mixed to produce desired fusion cells (hybridomas). Then, an appropriate culture medium mentioned above is gradually added to the cells, and this is repeatedly centrifuged to remove the supernatant. Thus,

cell fusion agents and such which are unfavorable to hybridoma growth can be removed.

The hybridomas thus obtained can be selected by culture using a conventional selective medium, for example, HAT medium (a culture medium containing hypoxanthine, aminopterin, and thymidine). Culture is continued in the above medium using the HAT medium for a period of time sufficient to kill cells other than the desired hybridomas (non-fused cells). Typically, the period is several days to several weeks. Then, hybridomas producing the desired antibody are screened and singly cloned by conventional limiting dilution methods.

The hybridomas thus obtained can be selected using a selection medium based on the selection marker possessed by the myeloma used for cell fusion. For example, HGPRT- or TK-deficient cells can be selected by culture using the HAT medium (a culture medium containing hypoxanthine, aminopterin, and thymidine). Specifically, when HAT-sensitive myeloma cells are used for cell fusion, cells successfully fused with normal cells can selectively proliferate in the HAT medium. Culture is continued in the above medium using the HAT medium for a period of time sufficient to kill cells other than the desired hybridomas (non-fused cells). Specifically, desired hybridomas can be selected by culture for generally several days to several weeks. Then, hybridomas producing the desired antibody are screened and singly cloned by conventional limiting dilution methods.

Screening and single cloning of desired antibodies can be suitably performed by screening methods based on known antigen-antibody reaction. For example, a desired antibody can be selected by screening using fluorescence activated cell sorting (FACS). FACS is a system that enables measurement of the binding of an antibody to cell surface by analyzing cells contacted with a fluorescent antibody using laser beam, and measuring the fluorescence emitted from individual cells.

To screen for hybridomas that produce a monoclonal antibody of the present invention by FACS, cells that express the antigen bound by the produced antibody are first prepared. Preferred cells used for screening are mammalian cells that are forced to express the antigen. By using mammalian cells that are used as the host cell but have not been transformed as a control, the activity of an antibody to bind to the cell-surface antigen can be selectively detected. Specifically, hybridomas producing a desired monoclonal antibody can be obtained by selecting hybridomas that produce an antibody which binds to cells forced to express the antigen but not to the host cell.

Alternatively, cells expressing the antigen of interest are immobilized and the activity of an antibody to bind to the antigen-expressing cells can be assessed based on the principle of ELISA. For example, antigen-expressing cells are immobilized to the wells of an ELISA plate. Culture supernatants of hybridomas are contacted with the immobilized cells in the wells, and antibodies that bind to the immobilized cells are detected. When the monoclonal antibodies are

derived from mouse, antibodies bound to the cells can be detected using an anti-mouse immunoglobulin antibody. Hybridomas producing a desired antibody having the antigen-binding ability are selected by the above screening, and they can be cloned by a limiting dilution method or the like.

Monoclonal antibody-producing hybridomas thus prepared can be passaged in a conventional culture medium. The hybridomas can be stored in liquid nitrogen for a long period.

The above hybridomas are cultured by a conventional method, and desired monoclonal antibodies can be obtained from the culture supernatants. Alternatively, the hybridomas are administered to and grown in compatible mammals, and monoclonal antibodies can be obtained from the ascites. The former method is suitable for obtaining antibodies with high purity.

Antibodies that are encoded by antibody genes cloned from antibody-producing cells such as the above hybridomas can also be preferably used. A cloned antibody gene is inserted into an appropriate vector, and this is introduced into a host to express the antibody encoded by the gene. Methods for isolating antibody genes, inserting the genes into vectors, and transforming host cells have already been established, for example, by Vandamme *et al*. (Eur. J. Biochem. (1990) 192(3), 767-775). Methods for producing recombinant antibodies are also known as described below.

Generally, to obtain a cDNA encoding the antibody variable region (V region), total RNA is first extracted from hybridomas. For example, the following methods can be used as methods for extracting mRNAs from cells:

- the guanidine ultracentrifugation method (Biochemistry (1979) 18(24), 5294-5299), and
- the AGPC method (Anal. Biochem. (1987) 162(1), 156-159).

Extracted mRNAs can be purified using the mRNA Purification Kit (GE Healthcare Bioscience) or such. Alternatively, kits for extracting total mRNA directly from cells, such as the QuickPrep mRNA Purification Kit (GE Healthcare Bioscience), are also commercially available. mRNAs can be prepared from hybridomas using such kits. cDNAs encoding the antibody V region can be synthesized from the prepared mRNAs using a reverse transcriptase. cDNAs can be synthesized using the AMV Reverse Transcriptase First-strand cDNA Synthesis Kit (Seikagaku Corporation) or such. Furthermore, the SMART RACE cDNA amplification kit (Clontech) and the PCR-based 5'-RACE method (Proc. Natl. Acad. Sci. USA (1988) 85(23), 8998-9002; Nucleic Acids Res. (1989) 17(8), 2919-2932) can be appropriately used to synthesize and amplify cDNAs. In such a cDNA synthesis process, appropriate restriction enzyme sites described below may be introduced into both ends of a cDNA.

The cDNA fragment of interest is purified from the resulting PCR product, and then this is ligated to a vector DNA. A recombinant vector is thus constructed, and introduced into *E.*

coli or such. After colony selection, the desired recombinant vector can be prepared from the colony-forming *E. coli*. Then, whether the recombinant vector has the cDNA nucleotide sequence of interest is tested by a known method such as the dideoxy nucleotide chain termination method.

The 5'-RACE method which uses primers to amplify the variable region gene is conveniently used for isolating the gene encoding the variable region. First, a 5'-RACE cDNA library is constructed by cDNA synthesis using RNAs extracted from hybridoma cells as a template. A commercially available kit such as the SMART RACE cDNA amplification kit is appropriately used to synthesize the 5'-RACE cDNA library.

The antibody gene is amplified by PCR using the prepared 5'-RACE cDNA library as a template. Primers for amplifying the mouse antibody gene can be designed based on known antibody gene sequences. The nucleotide sequences of the primers vary depending on the immunoglobulin subclass. Therefore, it is preferable that the subclass is determined in advance using a commercially available kit such as the Iso Strip mouse monoclonal antibody isotyping kit (Roche Diagnostics).

Specifically, for example, primers that allow amplification of genes encoding γ1, γ2a, γ2b, and γ3 heavy chains and κ and λ light chains are used to isolate mouse IgG-encoding genes. In general, a primer that anneals to a constant region site close to the variable region is used as a 3'-side primer to amplify an IgG variable region gene. Meanwhile, a primer attached to a 5' RACE cDNA library construction kit is used as a 5'-side primer.

Immunoglobulins composed of a combination of heavy and light chains may be reshaped using the thus amplified PCR products. A desired antibody can be selected by screening using the antigen-binding activity of a reshaped immunoglobulin as an indicator. The screening can be carried out, for example, by the following steps:

(1) contacting a desired antigen-expressing cell with an antibody comprising the V region encoded by a cDNA obtained from a hybridoma;

(2) detecting the binding of the antibody to the antigen-expressing cell; and

(3) selecting an antibody that binds to the antigen-expressing cell.

Methods for detecting the binding of an antibody to the antigen-expressing cells are known. Specifically, the binding of an antibody to the antigen-expressing cells can be detected by the above-described techniques such as FACS. Fixed samples of the antigen-expressing cells may be appropriately used to assess the binding activity of an antibody.

For antibody screening methods that use the binding activity as an indicator, panning methods that use phage vectors can also be used suitably. Screening methods using phage vectors are advantageous when the antibody genes are obtained from a polyclonal antibody-expressing cell population as heavy-chain and light-chain subclass libraries. Genes

encoding the heavy-chain and light-chain variable regions can be linked by an appropriate linker sequence to form a single-chain Fv (scFv). Phages expressing scFv on their surface can be produced by inserting a scFv-encoding gene into a phage vector. The phages are contacted with an antigen of interest. Then, a DNA encoding scFv having the binding activity of interest can be isolated by collecting phages bound to the antigen. This process can be repeated as necessary to enrich scFv having the binding activity of interest.

After isolation of the cDNA encoding the V region of the antibody of interest, the cDNA is digested with restriction enzymes that recognize the restriction sites introduced into both ends of the cDNA. Preferred restriction enzymes recognize and cleave a nucleotide sequence that occurs in the nucleotide sequence of the antibody gene at a low frequency. Furthermore, a restriction site for an enzyme that produces a sticky end is preferably introduced into a vector to insert a single-copy digested fragment in the correct orientation. The cDNA encoding the V region of the antibody is digested as described above, and this is inserted into an appropriate expression vector to construct an antibody expression vector. In this case, if a gene encoding the antibody constant region (C region) and a gene encoding the above V region are fused in-frame, a chimeric antibody is obtained. Herein, a “chimeric antibody” means that the origin of the constant region is different from that of the variable region. Thus, in addition to mouse/human heterochimeric antibodies, human/human allochimeric antibodies are included in the chimeric antibodies of the present invention. A chimeric antibody expression vector can be constructed by inserting the above V region gene into an expression vector that already has the constant region. Specifically, for example, a recognition sequence for a restriction enzyme that excises the above V region gene can be appropriately placed on the 5’ side of an expression vector carrying a DNA that encodes a desired antibody constant region (C region). A chimeric antibody expression vector is constructed by fusing in-frame two genes digested with the same combination of restriction enzymes.

To produce a monoclonal antibody, antibody genes are inserted into an expression vector so that the genes are expressed under the control of an expression regulatory region. The expression regulatory region for antibody expression includes, for example, enhancers and promoters. Furthermore, an appropriate signal sequence may be attached to the amino terminus so that the expressed antibody is secreted to the outside of cells. The signal sequence is cleaved from the carboxyl terminus of the expressed polypeptide, and the resulting antibody can be secreted to the outside of cells. Then, appropriate host cells are transformed with the expression vector, and recombinant cells expressing the antibody-encoding DNA can be obtained.

DNAs encoding the antibody heavy chain (H chain) and light chain (L chain) are separately inserted into different expression vectors to express the antibody gene. An antibody

molecule having the H and L chains can be expressed by co-transfecting the same host cell with vectors inserted with the H chain and L chain. Alternatively, host cells can be transformed with a single expression vector into which DNAs encoding the H and L chains are inserted (see WO 94/11523).

There are many known combinations of host cells and expression vectors for antibody preparation by introducing isolated antibody genes into appropriate hosts. All these expression systems are applicable to isolation of the domains that bind to a molecule expressed on the surface of a cell having immune response-suppressing function of the present invention and T cell receptor complex-binding domain.

Appropriate eukaryotic cells used as host cells include animal cells, plant cells, and fungal cells. Specifically, the animal cells include, for example, the following cells.
(1) mammalian cells: CHO, COS, myeloma, baby hamster kidney (BHK), HeLa, Vero, or such;
(2) amphibian cells: Xenopus oocytes, or such; and
(3) insect cells: sf9, sf21, Tn5, or such.

In addition, as a plant cell, an antibody gene expression system using cells derived from the *Nicotiana* genus such as *Nicotiana tabacum* is known. Callus cultured cells can be appropriately used to transform plant cells.

Furthermore, the following cells can be used as fungal cells:
yeasts: the *Saccharomyces* genus such as *Saccharomyces cerevisiae*, and the *Pichia* genus such as *Pichia pastoris*; and
filamentous fungi: the *Aspergillus* genus such as *Aspergillus niger*.

Furthermore, antibody gene expression systems that utilize prokaryotic cells are also known. For example, when using bacterial cells, *E. coli* cells, *Bacillus subtilis* cells, and such can suitably be utilized in the present invention. Expression vectors carrying the antibody genes of interest are introduced into these cells by transfection. The transfected cells are cultured *in vitro*, and the desired antibody can be prepared from the culture of transformed cells.

In addition to the above-described host cells, transgenic animals can also be used to produce a recombinant antibody. That is, the antibody can be obtained from an animal into which the gene encoding the antibody of interest is introduced. For example, the antibody gene can be constructed as a fusion gene by inserting in frame into a gene that encodes a protein produced specifically in milk. Goat β-casein or such can be used, for example, as the protein secreted in milk. DNA fragments containing the fused gene inserted with the antibody gene is injected into a goat embryo, and then this embryo is introduced into a female goat. Desired antibodies can be obtained as a protein fused with the milk protein from milk produced by the transgenic goat born from the embryo-recipient goat (or progeny thereof). In addition, to increase the volume of milk containing the desired antibody produced by the transgenic goat,

hormones can be administered to the transgenic goat as necessary (Bio/Technology (1994) 12 (7), 699-702).

When an antigen-binding molecule described herein is administered to human, an antigen-binding domain derived from a genetically recombinant antibody that has been artificially modified to reduce the heterologous antigenicity against human and such, can be appropriately used as the various binding domains in the molecule when domains comprising an antibody variable region are used. Such genetically recombinant antibodies include, for example, humanized antibodies. These modified antibodies are appropriately produced by known methods.

An antibody variable region used to produce the various binding domains of antigen-binding molecules described herein is generally formed by three complementarity-determining regions (CDRs) that are separated by four framework regions (FRs). CDR is a region that substantially determines the binding specificity of an antibody. The amino acid sequences of CDRs are highly diverse. On the other hand, the FR-forming amino acid sequences often have high identity even among antibodies with different binding specificities. Therefore, generally, the binding specificity of a certain antibody can be introduced into another antibody by CDR grafting.

A humanized antibody is also called a reshaped human antibody. Specifically, humanized antibodies prepared by grafting the CDR of a non-human animal antibody such as a mouse antibody to a human antibody and such are known. Common genetic engineering techniques for obtaining humanized antibodies are also known. Specifically, for example, overlap extension PCR is known as a method for grafting a mouse antibody CDR to a human FR. In overlap extension PCR, a nucleotide sequence encoding a mouse antibody CDR to be grafted is added to primers for synthesizing a human antibody FR. Primers are prepared for each of the four FRs. It is generally considered that when grafting a mouse CDR to a human FR, selecting a human FR that has high identity to a mouse FR is advantageous for maintaining the CDR function. That is, it is generally preferable to use a human FR comprising an amino acid sequence which has high identity to the amino acid sequence of the FR adjacent to the mouse CDR to be grafted.

Nucleotide sequences to be ligated are designed so that they will be connected to each other in frame. Human FRs are individually synthesized using the respective primers. As a result, products in which the mouse CDR-encoding DNA is attached to the individual FR-encoding DNAs are obtained. Nucleotide sequences encoding the mouse CDR of each product are designed so that they overlap with each other. Then, complementary strand synthesis reaction is conducted to anneal the overlapping CDR regions of the products synthesized using a human antibody gene as template. Human FRs are ligated *via* the mouse

CDR sequences by this reaction.

The full length V region gene, in which three CDRs and four FRs are ultimately ligated, is amplified using primers that anneal to its 5'- or 3'-end, which are added with suitable restriction enzyme recognition sequences. An expression vector for humanized antibody can be produced by inserting the DNA obtained as described above and a DNA that encodes a human antibody C region into an expression vector so that they will ligate in frame. After the recombinant vector is transfected into a host to establish recombinant cells, the recombinant cells are cultured, and the DNA encoding the humanized antibody is expressed to produce the humanized antibody in the cell culture (see, European Patent Publication No. EP 239400 and International Patent Publication No. WO 1996/002576).

By qualitatively or quantitatively measuring and evaluating the antigen-binding activity of the humanized antibody produced as described above, one can suitably select human antibody FRs that allow CDRs to form a favorable antigen-binding site when ligated through the CDRs. Amino acid residues in FRs may be substituted as necessary, so that the CDRs of a reshaped human antibody form an appropriate antigen-binding site. For example, amino acid sequence mutations can be introduced into FRs by applying the PCR method used for grafting a mouse CDR into a human FR. More specifically, partial nucleotide sequence mutations can be introduced into primers that anneal to the FR. Nucleotide sequence mutations are introduced into the FRs synthesized by using such primers. Mutant FR sequences having the desired characteristics can be selected by measuring and evaluating the activity of the amino acid-substituted mutant antibody to bind to the antigen by the above-mentioned method (Sato, K. *et al.*, Cancer Res. (1993) 53: 851-856).

Alternatively, desired human antibodies can be obtained by immunizing transgenic animals having the entire repertoire of human antibody genes (see WO 1993/012227; WO 1992/003918; WO 1994/002602; WO 1994/025585; WO 1996/034096; WO 1996/033735) by DNA immunization.

Furthermore, techniques for preparing human antibodies by panning using human antibody libraries are also known. For example, the V region of a human antibody is expressed as a single-chain antibody (scFv) on phage surface by the phage display method. Phages expressing a scFv that binds to the antigen can be selected. The DNA sequence encoding the human antibody V region that binds to the antigen can be determined by analyzing the genes of selected phages. The DNA sequence of the scFv that binds to the antigen is determined. An expression vector is prepared by fusing the V region sequence in frame with the C region sequence of a desired human antibody, and inserting this into an appropriate expression vector. The expression vector is introduced into cells appropriate for expression such as those described above. The human antibody can be produced by expressing the human antibody-encoding gene

in the cells. These methods are already known (see WO 1992/001047; WO 1992/020791; WO 1993/006213; WO 1993/011236; WO 1993/019172; WO 1995/001438; WO 1995/015388).

In addition to the phage display method, techniques that use a cell-free translation system, techniques for displaying antigen-binding molecules on the surface of viruses or cells, and techniques that use emulsions are also known as techniques for obtaining human antibodies by panning using human antibody libraries. For example, the ribosome display method where a complex is formed between the translated protein and mRNA *via* the ribosome by removing the stop codon and such, the cDNA display method or the mRNA display method where a genetic sequence and the translated protein are covalently linked using a compound such as puromycin, the CIS display method where a complex is formed between the gene and the translated protein using a nucleic acid-binding protein, or such may be used as techniques of using a cell-free translation system. For the technique of presenting antigen-binding molecules on the surface of cells or viruses, besides the phage display method, the *E. coli* display method, Gram-positive bacteria display method, yeast display method, mammalian cell display method, virus display method, and such may be used. As a technique that uses emulsions, the *in vitro* virus display method which involves incorporating genes and translation-related molecules into an emulsion, and such may be used. These methods are already publicly known (Nat Biotechnol. 2000 Dec;18(12):1287-92; Nucleic Acids Res. 2006;34(19): e127; Proc Natl Acad Sci U S A. 2004 Mar 2;101(9):2806-10; Proc Natl Acad Sci U S A. 2004 Jun 22;101(25):9193-8; Protein Eng Des Sel. 2008 Apr;21(4):247-55; Proc Natl Acad Sci U S A. 2000 Sep 26;97(20):10701-5; MAbs. 2010 Sep-Oct;2(5):508-18; and Methods Mol Biol. 2012, 911:183-98).

In the present invention, "specific" means a condition where one of the molecules involved in specific binding does not show any significant binding to molecules other than a single or a number of binding partner molecules. Furthermore, "specific" is also used when an antigen-binding domain is specific to a particular epitope among multiple epitopes contained in an antigen. When an epitope bound by an antigen-binding domain is contained in multiple different antigens, antigen-binding molecules containing the antigen-binding domain can bind to various antigens that have the epitope.

"Epitope" means an antigenic determinant in an antigen, and refers to an antigen site to which various binding domains in antigen-binding molecules disclosed herein bind. Thus, for example, an epitope can be defined according to its structure. Alternatively, the epitope may be defined according to the antigen-binding activity of an antigen-binding molecule that recognizes the epitope. When the antigen is a peptide or polypeptide, the epitope can be specified by the amino acid residues that form the epitope. Alternatively, when the epitope is a sugar chain, the epitope can be specified by its specific sugar chain structure.

A linear epitope is an epitope that contains an epitope whose primary amino acid

sequence is recognized. Such a linear epitope typically contains at least three and most commonly at least five, for example, about 8 to 10 or 6 to 20 amino acids in its specific sequence.

In contrast to the linear epitope, "conformational epitope" is an epitope in which the primary amino acid sequence containing the epitope is not the only determinant of the recognized epitope (for example, the primary amino acid sequence of a conformational epitope is not necessarily recognized by an epitope-defining antibody). Conformational epitopes may contain a greater number of amino acids compared to linear epitopes. A conformational epitope-recognizing antibody recognizes the three-dimensional structure of a peptide or protein. For example, when a protein molecule folds and forms a three-dimensional structure, amino acids and/or polypeptide main chains that form a conformational epitope become aligned, and the epitope is made recognizable by the antibody. Methods for determining epitope conformations include, for example, X ray crystallography, two-dimensional nuclear magnetic resonance spectroscopy, site-specific spin labeling, and electron paramagnetic resonance spectroscopy, but are not limited thereto. See, for example, Epitope Mapping Protocols in Methods in Molecular Biology (1996), Vol. 66, Morris (ed.).

Examples of a method for assessing the binding of an epitope in a molecule expressed on the surface of a cell having immune response-suppressing function by a test antigen-binding molecule are shown below. According to the examples below, methods for assessing the binding of an epitope in a target antigen by another binding domain can also be appropriately conducted.

For example, when CTLA4 is selected as the molecule expressed on the surface of a cell having immune response-suppressing function, whether a test antigen-binding molecule that comprises an antigen-binding domain for the molecule recognizes a linear epitope in the antigen molecule can be confirmed for example as mentioned below. For example, a linear peptide comprising an amino acid sequence forming the extracellular domain of CTLA4 is synthesized for the above purpose. The peptide can be synthesized chemically, or obtained by genetic engineering techniques using a region in a cDNA of CTLA4 encoding the amino acid sequence that corresponds to the extracellular domain. Then, a test antigen-binding molecule containing an antigen-binding domain for CTLA4 is assessed for its binding activity towards a linear peptide comprising the extracellular domain-constituting amino acid sequence. For example, an immobilized linear peptide can be used as an antigen to evaluate the binding activity of the antigen-binding molecule towards the peptide by ELISA. Alternatively, the binding activity towards a linear peptide can be assessed based on the level at which the linear peptide inhibits binding of the antigen-binding molecule to CTLA4-expressing cells. The binding activity of the antigen-binding molecule towards the linear peptide can be demonstrated by these tests.

Whether the above-mentioned test antigen-binding molecule containing an antigen-binding domain towards an antigen recognizes a conformational epitope can be confirmed as below. For example, the above-mentioned antigen-binding molecule that comprises an antigen-binding domain for CTLA4 strongly binds to CTLA4-expressing cells upon contact, but does not substantially bind to an immobilized linear peptide comprising an amino acid sequence forming the extracellular domain of CTLA4. Herein, "does not substantially bind" means that the binding activity is 80% or less, generally 50% or less, preferably 30% or less, and particularly preferably 15% or less compared to the binding activity to antigen-expressing cells.

Methods for assaying the binding activity of a test antigen-binding molecule comprising an antigen-binding domain to antigen-expressing cells include, for example, the methods described in Antibodies A Laboratory Manual (Ed Harlow, David Lane, Cold Spring Harbor Laboratory (1988) 359-420). Specifically, the assessment can be performed based on the principle of ELISA or fluorescence activated cell sorting (FACS) using antigen-expressing cells as antigen.

In the ELISA format, the binding activity of a test antigen-binding molecule comprising an antigen-binding domain towards antigen-expressing cells can be assessed quantitatively by comparing the levels of signals generated by enzymatic reaction. Specifically, a test antigen-binding molecule is added to an ELISA plate onto which antigen-expressing cells are immobilized. Then, the test antigen-binding molecule bound to the cells is detected using an enzyme-labeled antibody that recognizes the test antigen-binding molecule. Alternatively, when FACS is used, a dilution series of a test antigen-binding molecule is prepared, and the antibody-binding titer for antigen-expressing cells can be determined to compare the binding activity of the test antigen-binding molecule towards antigen-expressing cells.

The binding of a test antigen-binding molecule to an antigen expressed on the surface of cells suspended in buffer or the like can be detected using a flow cytometer. Known flow cytometers include, for example, the following devices:

FACSCanto™ II
FACSAria™
FACSArray™
FACSVantage™ SE
FACSCalibur™ (all are trade names of BD Biosciences)
EPICS ALTRA HyPerSort
Cytomics FC 500
EPICS XL-MCL ADC EPICS XL ADC
Cell Lab Quanta/Cell Lab Quanta SC (all are trade names of Beckman Coulter).

Suitable methods for assaying the binding activity of the above-mentioned test antigen-binding molecule comprising an antigen-binding domain towards an antigen include, for example, the method below. First, antigen-expressing cells are reacted with a test antigen-binding molecule, and then this is stained with an FITC-labeled secondary antibody that recognizes the antigen-binding molecule. The test antigen-binding molecule is appropriately diluted with a suitable buffer to prepare the antigen-binding molecule at a desired concentration. For example, the molecule can be used at a concentration within the range of 10 μg/ml to 10 ng/ml. Then, the fluorescence intensity and cell count are determined using FACSCalibur (BD). The fluorescence intensity obtained by analysis using the CELL QUEST Software (BD), i.e., the Geometric Mean value, reflects the quantity of antibody bound to the cells. That is, the binding activity of a test antigen-binding molecule, which is represented by the quantity of the test antigen-binding molecule bound, can be measured by determining the Geometric Mean value.

Whether a test antigen-binding molecule comprising an antigen-binding domain of the present invention shares a common epitope with another antigen-binding molecule can be assessed based on competition between the two molecules for the same epitope. The competition between antigen-binding molecules can be detected by a cross-blocking assay or the like. For example, the competitive ELISA assay is a preferred cross-blocking assay.

Specifically, in a cross-blocking assay, the antigen coating the wells of a microtiter plate is pre-incubated in the presence or absence of a candidate competitor antigen-binding molecule, and then a test antigen-binding molecule is added thereto. The quantity of test antigen-binding molecule bound to the antigen in the wells indirectly correlates with the binding ability of a candidate competitor antigen-binding molecule that competes for the binding to the same epitope. That is, the greater the affinity of the competitor antigen-binding molecule for the same epitope, the lower the binding activity of the test antigen-binding molecule towards the antigen-coated wells.

The quantity of the test antigen-binding molecule bound to the wells *via* the antigen can be readily determined by labeling the antigen-binding molecule in advance. For example, a biotin-labeled antigen-binding molecule can be measured using an avidin/peroxidase conjugate and appropriate substrate. In particular, a cross-blocking assay that uses enzyme labels such as peroxidase is called “competitive ELISA assay”. The antigen-binding molecule can also be labeled with other labeling substances that enable detection or measurement. Specifically, radiolabels, fluorescent labels, and such are known.

When the candidate competitor antigen-binding molecule can block the binding of a test antigen-binding molecule comprising an antigen-binding domain by at least 20%, preferably at least 20 to 50%, and more preferably at least 50% compared to the binding activity in a control experiment conducted in the absence of the competitor antigen-binding molecule, the test

antigen-binding molecule is determined to substantially bind to the same epitope bound by the competitor antigen-binding molecule, or to compete for binding to the same epitope.

When the structure of an epitope bound by a test antigen-binding molecule comprising an antigen-binding domain of the present invention is already identified, whether the test and control antigen-binding molecules share a common epitope can be assessed by comparing the binding activities of the two antigen-binding molecules towards a peptide prepared by introducing amino acid mutations into the peptide forming the epitope.

As a method for measuring such binding activities, for example, the binding activities of test and control antigen-binding molecules towards a linear peptide into which a mutation is introduced are measured by comparison in the above ELISA format. Besides the ELISA methods, the binding activity towards the mutant peptide bound to a column can be determined by passing the test and control antigen-binding molecules through the column, and then quantifying the antigen-binding molecule eluted in the eluate. Methods for adsorbing a mutant peptide to a column, for example, in the form of a GST fusion peptide, are known.

Alternatively, when the identified epitope is a conformational epitope, whether test and control antigen-binding molecules share a common epitope can be assessed by the following method. First, cells expressing an antigen targeted by an antigen-binding domain and cells expressing an antigen having an epitope introduced with a mutation are prepared. The test and control antigen-binding molecules are added to a cell suspension prepared by suspending these cells in an appropriate buffer such as PBS. Then, the cell suspension is appropriately washed with a buffer, and an FITC-labeled antibody that can recognize the test and control antigen-binding molecules is added thereto. The fluorescence intensity and number of cells stained with the labeled antibody are determined using FACSCalibur (BD). The test and control antigen-binding molecules are appropriately diluted using a suitable buffer, and used at desired concentrations. For example, they may be used at a concentration within the range of 10 μg/ml to 10 ng/ml. The fluorescence intensity determined by analysis using the CELL QUEST Software (BD), i.e., the Geometric Mean value, reflects the quantity of the labeled antibody bound to the cells. That is, the binding activities of the test and control antigen-binding molecules, which are represented by the quantity of the labeled antibody bound, can be measured by determining the Geometric Mean value.

An “antigen-binding molecule” of the present invention comprises both heavy and light chains which form an “antibody variable region” of this invention within a single polypeptide chain; however, it may be an antibody fragment lacking a constant region. Examples of such antibody fragments include a diabody (Db), an scFv, a single-chain antibody, an $sc(Fv)_2$, and an $sc(Fab')_2$.

Db is a dimer composed of two polypeptide chains (Holliger P *et al.*, Proc. Natl. Acad.

Sci. USA 90: 6444-6448 (1993); EP404,097; W093/11161). In each polypeptide chain, an L-chain variable region (VL) and an H-chain variable region (VH) are linked by a linker short enough so that these two regions on the same chain cannot associate with each other, for example, a linker of about five residues.

Because the linker between VL and VH is too short for formation of a single chain variable region fragment, VL and VH encoded on the same polypeptide chain dimerize to form two antigen-binding sites.

Furthermore, herein, the terms "scFv", "single-chain antibody", and "sc(Fv)$_2$" all refer to an antibody fragment of a single polypeptide chain that contains variable regions derived from the heavy and light chains, but not the constant region. In general, a single-chain antibody also contains a polypeptide linker between the VH and VL domains, which enables formation of a desired structure that is thought to allow antigen binding. The single-chain antibody is discussed in detail by Pluckthun in "The Pharmacology of Monoclonal Antibodies, Vol. 113, Rosenburg and Moore, eds., Springer-Verlag, New York, 269-315 (1994)". See also International Patent Publication WO 1988/001649; US Patent Nos. 4,946,778 and 5,260,203. In a particular embodiment, the single-chain antibody can be bispecific and/or humanized.

scFv is an antigen-binding domain in which VH and VL forming Fv are linked together by a peptide linker (Proc. Natl. Acad. Sci. U.S.A. (1988) 85(16), 5879-5883). VH and VL can be retained in close proximity by the peptide linker.

sc(Fv)$_2$ is a single-chain antibody in which four variable regions of two VL and two VH are linked by linkers such as peptide linkers to form a single chain (J Immunol. Methods (1999) 231(1-2), 177-189). The two VH and two VL may be derived from different monoclonal antibodies. Such sc(Fv)$_2$ preferably includes, for example, a bispecific sc(Fv)$_2$ that recognizes two types of epitopes present in a single antigen as disclosed in the Journal of Immunology (1994) 152(11), 5368-5374. sc(Fv)$_2$ can be produced by methods known to those skilled in the art. For example, sc(Fv)$_2$ can be produced by linking scFv by a linker such as a peptide linker.

Herein, the form of an antigen-binding domain forming an sc(Fv)$_2$ include an antibody in which the two VH units and two VL units are arranged in the order of VH, VL, VH, and VL ([VH]-linker-[VL]-linker-[VH]-linker-[VL]) beginning from the N terminus of a single-chain polypeptide. The order of the two VH units and two VL units is not limited to the above form, and they may be arranged in any order. Example order of the form is listed below.

[VL]-linker-[VH]-linker-[VH]-linker-[VL]
[VH]-linker-[VL]-linker-[VL]-linker-[VH]
[VH]-linker-[VH]-linker-[VL]-linker-[VL]
[VL]-linker-[VL]-linker-[VH]-linker-[VH]
[VL]-linker-[VH]-linker-[VL]-linker-[VH]

The molecular form of $sc(Fv)_2$ is also described in detail in WO2006/132352. According to these descriptions, those skilled in the art can appropriately prepare desired $sc(Fv)_2$ to produce the antigen-binding molecules disclosed herein.

Herein, the term "variable fragment (Fv)" refers to the minimum unit of an antibody-derived antigen-binding domain composed of a pair of the antibody light chain variable region (VL) and antibody heavy chain variable region (VH). In 1988, Skerra and Pluckthun found that homogeneous and active antibodies can be prepared from the *E. coli* periplasm fraction by inserting an antibody gene downstream of a bacterial signal sequence and inducing expression of the gene in *E. coli* (Science (1988) 240(4855), 1038-1041). In the Fv prepared from the periplasm fraction, VH associates with VL in a manner so as to bind to an antigen.

Furthermore, the antigen-binding molecule of the present invention may be conjugated with a carrier polymer such as PEG or an organic compound such as an anticancer agent. Alternatively, a glycosylation sequence can be inserted to suitably add a sugar chain for the purpose of producing a desired effect.

The linkers to be used for linking the variable regions of an antibody comprise arbitrary peptide linkers that can be introduced by genetic engineering, synthetic linkers, and linkers disclosed in, for example, Protein Engineering, 9(3), 299-305, 1996. However, peptide linkers are preferred in the present invention. The length of the peptide linkers is not particularly limited, and can be suitably selected by those skilled in the art according to the purpose. The length is preferably five amino acids or more (without particular limitation, the upper limit is generally 30 amino acids or less, preferably 20 amino acids or less), and particularly preferably 15 amino acids. When $sc(Fv)_2$ contains three peptide linkers, their length may be all the same or different.

For example, such peptide linkers include:

Ser
Gly·Ser
Gly·Gly·Ser
Ser·Gly·Gly
Gly·Gly·Gly·Ser (SEQ ID NO: 20)
Ser·Gly·Gly·Gly (SEQ ID NO: 21)
Gly·Gly·Gly·Gly·Ser (SEQ ID NO: 22)
Ser·Gly·Gly·Gly·Gly (SEQ ID NO: 23)
Gly·Gly·Gly·Gly·Gly·Ser (SEQ ID NO: 24)
Ser·Gly·Gly·Gly·Gly·Gly (SEQ ID NO: 25)
Gly·Gly·Gly·Gly·Gly·Gly·Ser (SEQ ID NO: 26)
Ser·Gly·Gly·Gly·Gly·Gly·Gly (SEQ ID NO: 27)

(Gly·Gly·Gly·Gly·Ser (SEQ ID NO: 22))n
(Ser·Gly·Gly·Gly·Gly (SEQ ID NO: 23))n
where n is an integer of 1 or larger. The length or sequences of peptide linkers can be selected accordingly by those skilled in the art depending on the purpose.

Synthetic linkers (chemical crosslinking agents) is routinely used to crosslink peptides, and for example:
N-hydroxy succinimide (NHS),
disuccinimidyl suberate (DSS),
bis(sulfosuccinimidyl) suberate (BS3),
dithiobis(succinimidyl propionate) (DSP),
dithiobis(sulfosuccinimidyl propionate) (DTSSP),
ethylene glycol bis(succinimidyl succinate) (EGS),
ethylene glycol bis(sulfosuccinimidyl succinate) (sulfo-EGS),
disuccinimidyl tartrate (DST), disulfosuccinimidyl tartrate (sulfo-DST),
bis[2-(succinimidoxycarbonyloxy)ethyl] sulfone (BSOCOES),
and bis[2-(sulfosuccinimidoxycarbonyloxy)ethyl] sulfone (sulfo-BSOCOES). These crosslinking agents are commercially available.

In general, three linkers are required to link four antibody variable regions together. The linkers to be used may be of the same type or different types.

Furthermore, "Fab" is composed of a single light chain, and a CH1 domain and variable region from a single heavy chain. The heavy chain of Fab molecule cannot form disulfide bonds with another heavy chain molecule.

"$F(ab')_2$" or "Fab'" is produced by treating an immunoglobulin (monoclonal antibody) with a protease such as pepsin and papain, and refers to an antibody fragment generated by digesting an immunoglobulin (monoclonal antibody) at near the disulfide bonds present between the hinge regions in each of the two H chains. For example, papain cleaves IgG upstream of the disulfide bonds present between the hinge regions in each of the two H chains to generate two homologous antibody fragments, in which an L chain comprising VL (L-chain variable region) and CL (L-chain constant region) is linked to an H-chain fragment comprising VH (H-chain variable region) and CHγ1 (γ1 region in an H-chain constant region) *via* a disulfide bond at their C-terminal regions. Each of these two homologous antibody fragments is called Fab'.

"$F(ab')_2$" contains two light chains and two heavy chains comprising the constant region of a CH1 domain and a portion of a CH2 domain so that disulfide bonds are formed between the two heavy chains. The $F(ab')_2$ constituting an antigen-binding molecule disclosed herein can be preferably obtained as below. A full-length monoclonal antibody or such comprising a

desired antigen-binding domain is partially digested with a protease such as pepsin, and then Fc fragments are removed by adsorption onto a Protein A column. The protease is not particularly limited, as long as it can digest the full-length antibody in a restrictive manner to produce $F(ab')_2$ under an appropriately established enzyme reaction condition such as pH. Such proteases include, for example, pepsin and ficin.

A preferred embodiment of the "antigen-binding molecule" of the present invention includes a multispecific antibody. When using an Fc region with decreased Fcγ receptor-binding activity as the Fc region of a multispecific antibody, an Fc region derived from a multispecific antibody may also be used appropriately. For the multispecific antibodies of the present invention, in particular, bispecific antibodies are preferred.

For association of multispecific antibodies, one can apply the technique of introducing charge repulsion at the interface of the second constant region of the antibody H chain (CH2) or the third constant region of the H chain (CH3) to suppress undesired associations between H chains (WO2006/106905).

In the technique of suppressing unintended association between H chains by introducing charge repulsion at the interface of CH2 or CH3, examples of the amino acid residues that are contacted at the interface of other constant regions of the H chain include the region facing the residue at position 356 (EU numbering), the residue at position 439 (EU numbering), the residue at position 357 (EU numbering), the residue at position 370 (EU numbering), the residue at position 399 (EU numbering), and the residue at position 409 (EU numbering) in the CH3 region.

More specifically, for example, for an antibody comprising two types of H chain CH3 regions, the antibody can be made so that one to three pairs of amino acid residues selected from the amino acid residue pairs shown below in (1) to (3) in the first H chain CH3 region have the same charge: (1) amino acid residues at positions 356 and 439 (EU numbering) which are amino acid residues contained in the H chain CH3 region; (2) amino acid residues at positions 357 and 370 (EU numbering) which are amino acid residues contained in the H chain CH3 region; and (3) amino acid residues at positions 399 and 409 (EU numbering) which are amino acid residues contained in the H chain CH3 region.

Furthermore, the antibody can be made so that one to three pairs of amino acid residues corresponding to the amino acid residue pairs shown above in (1) to (3) having the same type of charge in the first H chain CH3 region, which are amino acid residue pairs selected from the amino acid residue pairs shown above in (1) to (3) in the second H chain CH3 region which differs from the first H chain CH3 region, have a charge opposite to the corresponding amino acid residues in the aforementioned first H chain CH3 region.

The respective amino acid residues of (1) to (3) mentioned above are positioned close to

each other when associated. For the desired H chain CH3 region or H chain constant region, those skilled in the art can find sites corresponding to the above-mentioned amino acid residues of (1) to (3) by homology modeling and such using commercially available software, and amino acid residues of these sites can be subjected to modifications as appropriate.

In the above-mentioned antibodies, "amino acid residues having a charge" are preferably selected, for example, from amino acid residues contained in either one of groups (a) and (b) below:

(a) glutamic acid (E) and aspartic acid (D); and

(b) lysine (K), arginine (R), and histidine (H).

Regarding the above-mentioned antibodies, "having the same type of charge" means, for example, that two or more amino acid residues all have amino acid residues included in either one of the above-mentioned groups (a) and (b). The phrase "having the opposite charge" means that, for example, when at least one of the two or more amino acid residues has an amino acid residue included in either one of the above-mentioned groups (a) and (b), the remaining amino acid residue(s) will have an amino acid residue included in the other group.

In a preferred embodiment of the above-mentioned antibody, the first H chain CH3 region and the second H chain CH3 region may be cross-linked by a disulfide bond.

In the present invention, the amino acid residue to be subjected to alteration is not limited to an amino acid residue of the constant region or variable region of the antibody described above. With regard to polypeptide mutants or heteromultimers, those skilled in the art can find amino acid residues that form the interface through homology modeling and such using commercially available software, and can subject the amino acid residues at those sites to alterations so that association is regulated.

Other known techniques can also be used for the association of multispecific antibodies of the present invention. Polypeptides with different amino acids having an Fc region can be efficiently associated with each other by substituting an amino acid side chain present in one of the H chain variable regions of the antibody with a larger side chain (knob), and substituting an amino acid side chain present in the corresponding variable region of the other H chain with a smaller side chain (hole), to allow placement of the knob within the hole (WO1996/027011; Ridgway JB *et al.*, Protein Engineering (1996) 9, 617-621; Merchant AM *et al.* Nature Biotechnology (1998) 16, 677-681; and US20130336973).

In addition, other known techniques can also be used to form multispecific antibodies of the present invention. Association of polypeptides having different sequences can be induced efficiently by complementary association of CH3s, using a strand-exchange engineered CH3 domain produced by changing part of CH3 in one of the H chains of an antibody into its corresponding IgA-derived sequence, and introducing into the complementary portion of the

CH3 in the other H chain its corresponding IgA-derived sequence (Protein Engineering Design & Selection, 23; 195-202, 2010). This known technique can also be used to efficiently form multispecific antibodies of interest.

In addition, the following techniques and such may be used for the formation of multispecific antibodies: techniques for antibody production using association of antibody CH1 and CL, and association of VH and VL as described in WO 2011/028952, WO2014/018572, and Nat Biotechnol. 2014 Feb;32(2):191-8; techniques for producing bispecific antibodies using separately prepared monoclonal antibodies in combination (Fab Arm Exchange) as described in WO2008/119353 and WO2011/131746; techniques for regulating association between antibody heavy chain CH3s as described in WO2012/058768 and WO2013/063702; techniques for producing bispecific antibodies composed of two types of light chains and one type of heavy chain as described in WO2012/023053; techniques for producing bispecific antibodies using two bacterial cell strains that individually express one of the chains of an antibody comprising a single H chain and a single L chain as described by Christoph *et al.* (Nature Biotechnology Vol. 31, p 753-758 (2013)).

An embodiment of multispecific antibody formation includes methods for obtaining bispecific antibodies by mixing two types of monoclonal antibodies in the presence of a reducing agent to cleave the disulfide bonds in the core hinge region, followed by re-association for heterodimerization (FAE) as described above. Meanwhile, introduction of electrostatic interactions at the interacting interface of the CH3 region (WO2006/106905) can induce even more efficient heterodimerization during the re-association (WO2015/046467). In FAE using naturally-occurring IgG, re-association takes place randomly; and thus theoretically, bispecific antibodies can only be obtained at 50% efficiency; however, in this method, bispecific antibodies can be produced in high yield.

Alternatively, even when a multispecific antibody of interest cannot be formed efficiently, a multispecific antibody of the present invention can be obtained by separating and purifying the multispecific antibody of interest from the produced antibodies. For example, a method has been reported that enables purification of two types of homologous forms and the heterologous antibody of interest by ion exchange chromatography, by conferring a difference in the isoelectric points by introducing amino acid substitutions into the variable regions of the two types of H chains (WO2007114325). To date, as a method for purifying heterologous forms, a method using Protein A to purify a heterodimerized antibody comprising a mouse IgG2a H chain that binds to Protein A and a rat IgG2b H chain that does not bind to Protein A has been reported (WO98050431 and WO95033844). Furthermore, the heterodimerized antibody per se can be purified efficiently using a Protein A column by changing the interaction between each of the H chains and Protein A, by using H chains in which amino acid residues at the IgG-Protein A

binding site, positions 435 and 436 (EU numbering), are substituted with amino acids that yield a different binding strength to Protein A such as Tyr, His, or such.

Alternatively, a common L chain that can confer binding ability to a plurality of different H chains can be obtained and used as the common L chain of a multispecific antibody. Efficient expression of a multispecific IgG can be achieved by introducing the genes of such a common L chain and a plurality of different H chains into cells and expressing the IgG (Nature Biotechnology (1998) 16, 677-681). A method for selecting a common L chain that shows strong binding ability to any different H chains can also be used when selecting a common H chain (WO 2004/065611).

Furthermore, an Fc region whose C-terminal heterogeneity has been improved can be appropriately used as an Fc region of the present invention. More specifically, Fc regions lacking glycine at position 446 and lysine at position 447, as specified by EU numbering, in the amino acid sequences of two polypeptides constituting an Fc region derived from IgG1, IgG2, IgG3, or IgG4, are provided.

A plurality, such as two or more, of these techniques can be used in combination. Furthermore, these techniques can be appropriately and separately applied to the two H chains to be associated. Furthermore, these techniques can be used in combination with the above-mentioned Fc region of which Fcγ receptor-binding activity has been decreased. Furthermore, an antigen-binding molecule of the present invention may be a molecule produced separately based on an antigen-binding molecule subjected to the above-described modifications so as to have the same amino acid sequence.

An antigen-binding molecule of the present invention can be any molecule as long as it comprises:

(i) a domain that binds to a molecule expressed on the surface of a cell having immune response-suppressing function; and

(ii) a T cell receptor complex-binding domain, mentioned above, and its structure is not limited.

By comprising these two binding domains, the antigen-binding molecule can activate immune responses through inhibition of the immune response-suppressing effect by the molecule-expressing cell described in (i), and induce excellent cytotoxicity on cancer cells or cancer cell-comprising tumor tissues. The binding domains described in (i) and (ii) of the present invention can be appropriately selected from among the above-described molecules expressed on the surface of cells having immune response-suppressing function or antigens belonging to the T cell receptor complex, respectively. These binding domains can be directly linked by a peptide bond or *via* a linker.

The antigen-binding molecules of the present invention may further comprise an FcRn-binding domain. When using the Fc region of the above-described antibody as the

FcRn-binding domain, an Fc region with decreased Fcγ receptor-binding activity is preferred. Reducing the Fcγ receptor-binding activity enables suppression of side effects produced by systemic immune activation, such as cytokine release, caused by crosslinking between Fcγ receptor-expressing cells and T cell receptor complex-expressing cells.

Antigen-binding molecules of the present invention can be produced using the known methods described above.

For example, when (i) $F(ab')_2$ is used as the domain that binds to a molecule expressed on the surface of a cell having immune response-suppressing functions, (ii) $F(ab')_2$ is used as a T cell receptor complex-binding domain, and (iii) a domain comprising an Fc region with decreased Fcγ receptor-binding activity is used as the FcRn-binding domain, and when the antigen-binding domains described in (i) and (ii) and the Fc region-comprising domain described in (iii) are directly linked by peptide bonds, the linked polypeptides form an antibody structure. Such antibodies can be produced by purification from the above-described hybridoma culture medium, and also by purifying antibodies from the culture medium of desired host cells that stably carry polynucleotides encoding polypeptides constituting the antibody.

In addition to the linkers exemplified above, linkers with peptide tags such as His tag, HA tag, myc tag, and FLAG tag may also be suitably used as the linkers to be employed when connecting each of the domains *via* linkers. Furthermore, hydrogen bonding, disulfide bonding, covalent bonding, ionic interaction, or the property of mutual binding as a result of combination thereof may be suitably used. For example, the affinity between antibody CH1 and CL may be used, and Fc regions derived from the above-described multispecific antibodies may also be used for heterologous Fc region association.

The present invention also relates to polynucleotides encoding the antigen-binding molecules of the present invention, and they can be incorporated into arbitrary expression vectors. Suitable hosts can be transformed with the expression vectors to produce cells that express the antigen-binding molecules. Antigen-binding molecules encoded by the polynucleotides can be obtained by culturing cells that express the antigen-binding molecules, and collecting expression products from the culture supernatant. That is, the present invention relates to vectors comprising a polynucleotide that encodes an antigen-binding molecule of the present invention, cells carrying such a vector, and methods for producing antigen-binding molecules, which comprise culturing the cells and collecting antigen-binding molecules from the culture supernatant. These can be obtained by techniques similar to those for recombinant antibodies mentioned above.

Pharmaceutical compositions

From another perspective, the present invention provides pharmaceutical compositions

comprising the above-described antigen-binding molecule as an active ingredient. Furthermore, the present invention relates to pharmaceutical compositions that inhibit immune response-suppressing activity (agents that inhibit immune response-suppressing activity), immune response activating agents, agents that induce cytotoxicity, cell proliferation-suppressing agents (cell proliferation inhibitors), and anticancer agents, which comprise the antigen-binding molecule as an active ingredient (hereinafter, pharmaceutical compositions and such). Pharmaceutical compositions of the present invention can be used as agents for treating cancer or agents for preventing cancer. The agents that inhibit immune response-suppressing activity, immune response activating agents, therapeutic agents that induce cytotoxicity, cell proliferation-suppressing agents, and anticancer agents of the present invention are preferably administered to subjects suffering from cancer, or subjects who may relapse.

Furthermore, in the present invention, agents that inhibit immune response-suppressing activity, immune response activating agents, therapeutic agents that induce cytotoxicity, cell proliferation suppressing agents, and anticancer agents, which comprise the above-described antigen-binding molecule as an active ingredient can be presented as methods for inhibiting immune response-suppressing activity, methods for activating immune response, methods for inducing cytotoxicity, methods for suppressing cell proliferation, methods for activating immunity against cancer cells or cancer cell-comprising tumor tissues, or as methods for preventing or treating cancer, which comprise the step of administering the antigen-binding molecule to a subject; or presented as use of the antigen-binding molecules in producing pharmaceutical compositions for inhibiting immune response-suppressing activity, pharmaceutical compositions for activating immune response, pharmaceutical compositions for inducing cytotoxicity, cell proliferation-suppressing agents, and anticancer agents; or alternatively, presented as antigen-binding molecules for use in inhibiting immune response-suppressing activity, activating immune response, inducing cytotoxicity, suppressing cell proliferation, activating immunity against cancer cells or cancer cell-comprising tumor tissues, or treating or preventing cancer.

In the present invention, “comprising the antigen-binding molecule as an active ingredient” means containing the antigen-binding molecule as a major active component, and does not limit the content of the antigen-binding molecule.

Furthermore, pharmaceutical compositions or such of the present invention can be used by combining multiple types of antigen-binding molecules as necessary. For example, by using a cocktail of a plurality of antigen-binding molecules of the present invention that bind to the same antigen, one can enhance the effect on cells expressing the antigen.

If necessary, the antigen-binding molecules of the present invention may be encapsulated in microcapsules (microcapsules made from hydroxymethylcellulose, gelatin,

poly[methylmethacrylate], and the like), and made into components of colloidal drug delivery systems (liposomes, albumin microspheres, microemulsions, nano-particles, and nano-capsules) (for example, see "Remington's Pharmaceutical Science 16th edition", Oslo Ed. (1980)). Moreover, methods for preparing agents as sustained-release agents are known, and these can be applied to the antigen-binding molecules of the present invention (J. Biomed. Mater. Res. (1981) 15, 267-277; Chemtech. (1982) 12, 98-105; US Patent No. 3773719; European Patent Application (EP) Nos. EP58481 and EP133988; Biopolymers (1983) 22, 547-556).

The pharmaceutical compositions, immune response activating agents, cytotoxicity inducing agents, cell proliferation-suppressing agents (cell proliferation inhibitors), or anticancer agents of the present invention may be administered either orally or parenterally to patients. Parental administration is preferred. Specifically, such administration methods include injection, nasal administration, transpulmonary administration, and percutaneous administration. Injections include, for example, intravenous injections, intramuscular injections, intraperitoneal injections, and subcutaneous injections. For example, pharmaceutical compositions, therapeutic agents for inducing cellular cytotoxicity, cell proliferation-suppressing agents, or anticancer agents of the present invention can be administered locally or systemically by injection. Furthermore, appropriate administration methods can be selected according to the patient's age and symptoms. The administered dose can be selected, for example, from the range of 0.0001 mg to 1,000 mg per kg of body weight for each administration. Alternatively, the dose can be selected, for example, from the range of 0.001 mg/body to 100,000 mg/body per patient. However, the dose of a pharmaceutical composition of the present invention is not limited to these doses.

The pharmaceutical compositions of the present invention can be formulated according to conventional methods (for example, Remington's Pharmaceutical Science, latest edition, Mark Publishing Company, Easton, U.S.A.), and may also contain pharmaceutically acceptable carriers and additives. Examples include, but are not limited to, surfactants, excipients, coloring agents, flavoring agents, preservatives, stabilizers, buffers, suspension agents, isotonic agents, binders, disintegrants, lubricants, fluidity promoting agents, and corrigents, and other commonly used carriers can be suitably used. Specific examples of the carriers include light anhydrous silicic acid, lactose, crystalline cellulose, mannitol, starch, carmellose calcium, carmellose sodium, hydroxypropyl cellulose, hydroxypropyl methylcellulose, polyvinylacetal diethylaminoacetate, polyvinylpyrrolidone, gelatin, medium-chain triglyceride, polyoxyethylene hardened castor oil 60, saccharose, carboxymethyl cellulose, corn starch, inorganic salt, and such.

Furthermore, the present invention provides methods for inducing damage to cancer cells or cancer cell-comprising tumor tissues, or methods for suppressing proliferation of cancer

cells or cancer cell-comprising tumor tissues, by contacting the cells having a function of suppressing a certain immune response with antigen-binding molecules of the present invention that bind to the molecules expressed on the surface of these cells and damaging the cells. Cancer cells or cancer cell-comprising tumor tissues which become targets of the antigen-binding molecules of the present invention are not particularly limited, but they are preferably carcinomas in which cells having immune response-suppressing activity are involved in cancer progression, or carcinomas in which the number of regulatory T cells or exhausted T cells in the tumor correlates with the prognosis. Reported examples of such carcinomas include ovarian cancer (Non-patent Documents 11 and 12), gastric cancer (Non-patent Documents 13 and 14), esophageal cancer (Non-patent Document 14), pancreatic cancer (Non-patent Document 15), renal cell carcinoma (Non-patent Document 16), hepatocellular carcinoma (Non-patent Document 17), breast cancer (Non-patent Documents 18 and 19), malignant melanoma (Non-patent Document 20), non-small-cell lung cancer (Non-patent Document 21), cervical cancer (Non-patent Document 22), glioblastoma (Non-patent Document 23), prostate cancer (Non-patent Document 24), neuroblastoma (Non-patent Document 25), chronic lymphocytic leukemia (Non-patent Document 26), papillary thyroid cancer (Non-patent Document 27), colorectal cancer (Non-patent Document 28), and B-cell non-Hodgkin's lymphoma (Non-patent Documents 29 and 30), and they are suitable examples of carcinomas for the present invention. The term "cancer" as used herein means not only epithelial malignant tumors such as ovarian cancer or gastric cancer, but also non-epithelial malignant tumors including hematopoietic organ cancers such as chronic lymphocytic leukemia and Hodgkin's lymphoma.

In the present invention, "contact" is carried out, for example, by adding an antigen-binding molecule of the present invention that binds to an antigen to a solution of the cells expressing the target antigen cultured *in vitro*. In this case, a form suitable for use of the added antigen-binding molecule may be a solution, or a solid or such obtained by freeze-drying, and the like. When added as an aqueous solution, an aqueous solution containing purely the antigen-binding molecule of the present invention alone may be used, or a solution containing surfactants, excipients, coloring agents, perfumes, preservatives, stabilizers, buffers, suspending agents, isotonization agents, binders, disintegrants, lubricants, fluidity promoting agents, flavoring agents, and such described above may be used. The concentration used for the addition is not particularly limited, but a suitable final concentration in the culture solution is preferably in the range of 1 pg/ml to 1 g/ml, more preferably 1 ng/ml to 1 mg/ml, and even more preferably 1 μg/ml to 1 mg/ml.

Furthermore, in another embodiment, "contact" of the present invention is carried out, for example, by administering, to a non-human animal with a cancer cell line of interest transplanted into their bodies, an antigen-binding molecule of the present invention that binds to

a molecule expressed on the surface of a cell having the function of suppressing the immune response of the non-human animal; or by transplanting a cancer cell line of interest into the body of a non-human animal expressing human-derived cells having immune response-suppressing function, and then administering to the animal a molecule of the present invention that binds to a molecule expressed on the surface of the human-derived cells. The method of administration may be oral or parenteral, and parenteral administration is particularly preferred. Specific examples of the administration method include administration by injection, transnasal administration, transpulmonary administration, and transdermal administration. Examples of administration by injection include intravenous injection, intramuscular injection, intraperitoneal injection, and subcutaneous injection. A pharmaceutical composition, or a pharmaceutical composition for inhibiting immune response-suppressing activity, a pharmaceutical composition for activating an immune response, or a pharmaceutical composition for inducing cytotoxicity, a cell proliferation-suppressing agent (cell proliferation inhibitor), or an anticancer agent of the present invention can be administered systemically or locally, for example, through administration by injection. The method of administration can be selected appropriately according to the age and symptoms of the test animal. When administered as an aqueous solution, an aqueous solution containing purely the antigen-binding molecule of the present invention alone may be used, or a solution containing surfactants, excipients, coloring agents, perfumes, preservatives, stabilizers, buffers, suspending agents, isotonization agents, binders, disintegrants, lubricants, fluidity promoting agents, flavoring agents, and such described above may be used. The dose can be selected, for example, in the range from 0.0001 mg to 1000 mg per kilogram body weight for a single administration. Alternatively, for example, the dose may be selected in the range of 0.001 mg/body to 100000 mg/body per patient. However, the amount of the antigen-binding molecule of the present invention administered is not limited to these doses.

The following method is suitably used as a method for evaluating or measuring cytotoxicity induced in the targeted cancer cells or cancer cell-comprising tumor tissues, as a result of contacting the targeted cells or tissues with an antigen-binding molecule of the present invention. Examples of a method for evaluating or measuring the cytotoxic activity *in vitro* include methods for measuring cytotoxic T cell activity, and such. Whether an antigen-binding molecule of the present invention has T cell cytotoxicity can be measured by known methods (for example, Current protocols in Immunology, Chapter 7. Immunologic studies in humans, Editor, John E. Coligan *et al.*, John Wiley & Sons, Inc., (1993) and the like). For activity measurements, an antigen-binding molecule which binds to an antigen different from the antigen bound by the antigen-binding domain of the present invention and is an antigen not expressed in the cells used for the examination can be used as a control in the same manner as the

antigen-binding molecule of the present invention, and the activity can be determined to be present when the antigen-binding molecule of the present invention shows a stronger cytotoxic activity than the cytotoxic activity of the antigen-binding molecule used as a control.

To evaluate or measure cytotoxic activity *in vivo*, for example, cancer cells or cancer cell-comprising tumor tissues which are targeted by an antigen-binding molecule of the present invention are intradermally or subcutaneously transplanted to a non-human test animal, and then a test antigen-binding molecule is intravenously or intraperitoneally administered daily or with interval of few days, starting from the day of transplantation or the following day. Tumor size is measured daily and the differences in the change of tumor size can be defined as the cytotoxic activity. In a similar manner to the *in vitro* evaluation, a control antigen-binding molecule is administered, and an antigen-binding molecule of the present invention can be determined as exhibiting cytotoxic activity based on the finding that the tumor size in the group subjected to administration of an antigen-binding molecule of the present invention is significantly smaller than the tumor size in the group subjected to administration of the control antigen-binding molecule.

As a method for evaluating or measuring the suppressive effect on proliferation of cells expressing an antigen that is bound by a domain that constitutes an antigen-binding molecule of the present invention, which domain binds to a molecule expressed on the surface of a cell having immune response-suppressing function, by contact with the antigen-binding molecule, a method of measuring the uptake of isotope-labeled thymidine into cells or the MTT method may be suitably used. As a method for evaluating or measuring the cell proliferation-suppressing activity *in vivo*, the same method as that described above for evaluating or measuring cytotoxic activity *in vivo* may be suitably used.

The present invention also provides kits for use in the methods of the present invention, which comprise an antigen-binding molecule of the present invention or an antigen-binding molecule produced by a production method of the present invention. Additionally, the kit may include in its package, a pharmaceutically acceptable carrier, solvent, and instructions describing the method of use.

The present invention also relates to an antigen-binding molecule of the present invention or an antigen-binding molecule produced by a production method of the present invention for use in a method of the present invention.

Those skilled in the art will naturally understand that optional combinations of one or more of the embodiments described herein are included in the present invention, as long as they are not technically inconsistent based on common technical knowledge of those skilled in the art.

All prior art references cited herein are incorporated by reference into this description.

Examples

[Example 1] Concept of T cell-redirecting antibody targeting a cell surface marker of a regulatory T cell

(1-1) Antitumor effects of anti-CTLA4 antibodies through elimination of regulatory T cells

As described above in Background Art, Ipilimumab had been considered to inhibit CTLA4 expressed on the surface of effector T cells from suppressing effector T-cell activation, and thereby exhibit antitumor effects However, recently, antibody-dependent cellular cytotoxic activity (ADCC activity) against CTLA4-expressing T cells was also reported to be important, and elimination of regulatory T cells in tumors and ADCC activity have been found to be important mechanisms of action for the antitumor effects of anti-CTLA4 antibodies.

On the other hand, ADCC activity by an IgG1 antibody induces cytotoxic activity through binding of the antibody constant region to FcγR of NK cells or macrophages, and antibodies having a constant region that have been modified so as to enhance such binding are known to induce stronger cytotoxic activities and demonstrate antitumor effects.

As mentioned above, binding to regulatory T cells or exhausted T cells in a cancer microenvironment, and elimination of regulatory T cells or exhausted T cells by cytotoxic activity were found to exert strong antitumor effects. Therefore, if regulatory T cells or exhausted T cells can be eliminated more powerfully, or more specifically, if stronger cytotoxic activity can be exhibited, stronger antitumor effects can be expected to be exerted.

(1-2) T cell-redirecting antibody

Enhancement of the aforementioned ADCC activity, increase of retention in blood, improvement of antigen-binding activity, and reduction of immunogenicity risk have been performed as techniques for improving antibodies. Generally, antibodies recognize and bind a single epitope of an antigen, therefore even when such techniques for improvement are applied to the antibodies, only one type of antigen becomes the target. As molecules that inhibit multiple targets, antibodies that bind to two or more types of antigens by one molecule (referred to as bispecific antibodies) have been studied. Since bispecific antibodies interact with two or more types of antigens, they have not only the effect of neutralizing two or more types of antigens with one molecule, but also the effect of enhancing antitumor activities by crosslinking cells having cytotoxic activity with cancer cells.

Blinatumomab, which is a BiTE molecule, and Catumaxomab are known as bispecific antibodies that recognize a protein expressed on T cells (CD3ε or TCR) and a protein expressed on cancer cells (a cancer antigen). These molecules can bind to a cancer antigen and the CD3ε chain expressed on a T cell with each of their two antigen-binding domains (scFv or Fab), and form intercellular crosslinks between the T cells and the cancer antigen-expressing cells (Fig. 1).

This way, such T cell-redirecting antibodies can use T cells as effector cells to induce strong cytotoxic activity against cancer antigen-expressing cells.

Meanwhile, antibodies that use T cells as effector cells and induce strong cytotoxic activity against T cells have not been reported so far. For example, since CD3ε is a standard T cell marker, an IgG antibody (not having FcgR-binding activity) which binds to CD3εs using both arms (two Fabs) may be able to cause T cells to induce strong cytotoxic activity against T cells by forming intercellular crosslink between a CD3ε-expressing T cell which will become an effector cell and a CD3ε-expressing T cell, as shown in Fig. 2-1, but this hardly occurs in reality. That is because an IgG antibody that binds to CD3εs using both arms (two Fabs) strongly binds to CD3εs expressed on the same cell due to avidity *via* bivalent binding, intercellular crosslink is not formed between CD3ε-expressing T cells (Fig. 2-2).

Since CD3 is a standard T cell marker and it is expressed in both T cells which will become the effector cells and T cells which will become the target (for example, regulatory T cells and exhausted T cells), T cell-redirecting antibodies had been considered to be not able to exert cytotoxic activity against target T cells. More specifically, a T cell-redirecting antibody against a T cell expressing antigen X, a surface marker of a specific T cell population (bispecific antibody against CD3ε and antigen X) strongly binds to the target T cell (for example, a regulatory T cell and an exhausted T cell) with avidity *via* bivalent binding, since CD3ε and antigen X are expressed on the target T cell. Therefore, intercellular crosslinking was considered not to take place with T cells which will become effector cells (Fig. 3). In fact, there have been no reports of T cell-redirecting antibodies against T cell antigens.

Therefore, T cell-redirecting antibodies against CTLA4 expressed on regulatory T cells and exhausted T cells (bispecific antibodies against CD3ε and CTLA4), as shown in Fig. 4, are considered not to be able to induce strong cytotoxic activity against regulatory T cells and exhausted T cells because both CD3ε and CTLA4 are expressed on regulatory T cells and exhausted T cells, which are target cells, and the T cells which will become effector cells are not crosslinked with regulatory T cells and exhausted T cells. So far, there have been no reports on T cell-redirecting antibodies against regulatory T cells or exhausted T cells.

More specifically, it was unknown whether T cell-redirecting antibodies against CTLA4 (anti-CTLA4/anti-CD3ε bispecific antibodies) can actually damage regulatory T cells by inducing intercellular crosslinking and demonstrate antitumor effects *in vivo*. Therefore, we actually produced bispecific antibodies against CTLA4 and CD3 and tested whether they can achieve effects *in vivo* in mice and *in vitro* in humans.

Against standard cancer antigens, T cell-redirecting antibodies have been known to have stronger antitumor effects than antibodies having NK cell-utilizing ADCC activity; however, it had been unknown whether T cell-redirecting antibodies against CTLA4 show stronger

antitumor effects than anti-CTLA4 antibodies with enhanced ADCC activity.

[Example 2] Preparation and assessment of ADCC activity-enhanced antibodies targeting regulatory T cells

(2-1) Expression and purification of an ADCC activity-enhanced antibody that binds specifically to mouse CTLA4 (hUH02hUL01-mFa55)

Genes encoding the variable regions of the anti-mouse CTLA4 antibody hUH02hUL01 (the heavy chain variable region UH02 is SEQ ID NO: 28, and the light chain variable region UL01 is SEQ ID NO: 29) were each inserted into mouse IgG2a/kappa plasmids for expression in animals. Here, constant regions that have been modified so as to enhance binding to mouse FcγR were used (the heavy chain constant region mFa55 is SEQ ID NO: 30, and the light chain constant region mk1 is SEQ ID NO: 31).

Antibodies were expressed by the method described below. Cells of human embryonic kidney cell-derived FreeStyle 293-F strain (Invitrogen) were suspended in FreeStyle 293 Expression Medium (Invitrogen) at a cell density of 1.33×10^6 cells/mL, and seeded into each well of a 6-well plate at 3 mL/well. The prepared plasmids were introduced into the cells by a lipofection method. The cells were cultured for four days in a CO_2 incubator (37°C, 8% CO_2, 90 rpm). From the culture supernatants, antibodies were purified using rProtein A Sepharose™ Fast Flow (Amersham Biosciences) by a method known to those skilled in the art. Absorbance at 280 nm of the purified antibody solutions was measured using a spectrophotometer. Concentrations of the purified antibodies were calculated from the determined values using an extinction coefficient calculated by the PACE method (Protein Science (1995) 4: 2411-2423).

(2-2) Assessment of binding of the anti-mouse CTLA4 antibody (hUH02UL01-mFa55) to various mouse FcgRs

Anti-mouse CTLA4 antibody hUH02hUL01-mFa55 and control antibody hUH02hUL01-mIgG2a (the heavy chain variable region UH02 is SEQ ID NO: 28, the light chain variable region UL01 is SEQ ID NO: 29, the heavy chain constant region mIgG2a is SEQ ID NO: 32, and the light chain constant region mk1 is SEQ ID NO: 31) purified and prepared by the method of Example 2-1 were analyzed for their antigen-antibody reactions with various mouse FcgRs (mFcgRI, II, III, and IV) using Biacore T200 (GE Healthcare). The running buffer used was 20 mmol/L ACES, 150 mmol/L NaCl, 0.05% (w/v) Tween20 at pH7.4, and measurements were taken at 25°C. Protein A/G was immobilized onto a Sensor Chip CM7 by amine coupling. hUH02hUL01-mFa55 was captured onto the sensorchip, and then FcgR was allowed to interact as an analyte for 120 seconds, and change in the bound amount was observed. The running buffer was used for dilution of hUH02hUL01-mFa55. The measurement results were analyzed by curve fitting using the Biacore T200 Evaluation Software (GE Healthcare) to

calculate the association rate constants ka (1/Ms) and the dissociation rate constants kd (1/s). From those values, the dissociation constants K_D (M) were determined. The measurement results are shown in Table 1.

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Table 1

No.	Name of Fc	mFcgRI			mFcgRII			mFcgRIII			mFcgRIV		
		1:1 binding RI=0			1:1 binding RI=0			1:1 binding RI=0			1:1 binding RI=0		
		ka	kd	KD	ka	kd	KD	ka	kd	KD	ka	kd	KD
		(1/Ms)	(1/s)	(M)	(1/Ms)	(1/s)	(M)	(1/Ms)	(1/s)	(M)	(1/Ms)	(1/s)	(M)
1	mIgG2a	7.8E+05	4.7E-03	6.1E-09	1.3E+06	1.1E+00	7.8E-07	2.0E+06	5.1E-01	2.5E-07	1.2E+06	1.6E-02	1.3E-08
2	mFa55	1.2E+06	3.1E-03	2.5E-09	2.1E+06	7.6E-01	3.7E-07	2.4E+06	6.2E-01	2.6E-07	1.5E+06	2.8E-03	1.9E-09

(2-3) Evaluation of an anti-mouse CTLA4 antibody (hUH02UL01-mFa55) as an ADCC activity-enhanced antibody

Whether anti-mouse CTLA4 antibody hUH02hUL01-mFa55 purified and prepared by the method of Example 2-1 exerts ADCC activity on mouse CTLA4-expressing cells (mouse CTLA4-expressing cells were produced by a method known to those skilled in the art by introducing the full length mouse CTLA4 gene into CHO cells) was examined according to the method of Reference Example 1. The measurement results show ADCC activity in an antibody concentration-dependent manner (Fig. 5).

[Example 3] Preparation and assessment of bispecificity which recognizes surface antigens of regulatory T cells and effector T cells.

(3-1) Expression and purification of bispecific antibodies that specifically bind to mouse CTLA4 and mouse CD3

Genes encoding the variable regions of the anti-mouse CTLA4 antibody hUH02hUL01 (the heavy chain variable region UH02 is SEQ ID NO: 28, and the light chain variable region UL01 is SEQ ID NO: 29) were each inserted into human IgG1/kappa plasmids for expression in animals. Here, constant regions that have been modified so as to reduce binding to Fcγ receptors and to produce heterologous association of two heavy chains were used (the heavy chain constant region F760nN17 is SEQ ID NO: 33, and the light chain constant region k0 is SEQ ID NO: 34).

Genes encoding the variable regions of the anti-mouse CD3 antibody 2C11 (the heavy chain variable region is SEQ ID NO: 35, and the light chain variable region is SEQ ID NO: 36) were each inserted into human IgG1/kappa plasmids for expression in animals. Here, constant regions that have been modified so as to reduce binding to Fcγ receptors and to produce heterologous association of two heavy chains were used (the heavy chain constant region F760nP17 is SEQ ID NO: 37, and the light chain constant region k0 is SEQ ID NO: 34).

Each of hUH02hUL01-F760nN17 and 2C11-F760nP17 was expressed and purified by the method shown in Example 2. Each of the purified homologous forms was mixed by a method known to those skilled in the art that uses differences in the charges of the constant regions (Proc. Natl. Acad. Sci., 110, 5145-5150, 2013) to produce the bispecific antibody of interest (hUH02UL01/2C11-F760).

(3-2) Evaluation of a bispecific antibody that specifically binds to mouse CTLA4 and mouse CD3 for the antigen (mCTLA4 and mCD3)-binding properties

Anti-mouse CTLA4/anti-mouse CD3 bispecific antibodies (hUH02UL01/2C11-F760) purified and prepared by the method of Example 3-1 were analyzed for their antigen-antibody

reactions with each antigen (mCTLA4 and mCD3) using Biacore T200 (GE Healthcare). The running buffer used was HBS-EP+ at pH7.4, and measurements were taken at 37°C. Protein A/G was immobilized onto a Sensor Chip CM4 by amine coupling. hUH02UL01/2C11-F760 was captured onto the sensor chip, and then the antigen (mouse CTLA4 or mouse CD3) was allowed to interact as an analyte (for 120 seconds for mouse CTLA4 and for 90 seconds for mouse CD3), and changes in the bound amount were observed. The running buffer was used for dilution of hUH02UL01/2C11-F760. The measurement results were analyzed by curve fitting using the Biacore T200 Evaluation Software (GE Healthcare) to calculate the association rate constants ka (1/Ms) and the dissociation rate constants kd (1/s). From those values, the dissociation constants K_D (M) were determined. The results are shown in Table 2.

Table 2

No	Analyte	ka (1/Ms)	kd (1/s)	KD (M)
1	Mouse CTLA4	4.30E+05	5.30E-04	1.23E-09
2	Mouse CD3	6.16E+04	7.12E-02	1.16E-06

(3-2) Evaluation of cytotoxic activity by an anti-mouse CTLA4/anti-mouse CD3 bispecific antibody (hUH02UL01/2C11-F760)

Whether anti-mouse CTLA4/anti-mouse CD3 bispecific antibody (hUH02UL01/2C11-F760) purified and prepared by the method of Example 3-1 exerts cytotoxic activity on mouse CTLA4-expressing cell lines was examined according to the method of Reference Example 2. The measurement results show cytotoxic activity in an antibody concentration-dependent manner (Fig. 6).

[Example 4] Assessment of crosslinking between beads to which CD3 and CTLA4 have been immobilized and CD3-bound beads by an anti-mouse CTLA4/anti-mouse CD3 bispecific antibody (hUH02UL01/2C11-F760)

Whether an anti-CTLA4/anti-CD3 bispecific antibody recognizes surface antigens of a regulatory T cell (expressing CTLA4 and CD3) and an effector T cell (expressing CD3) and forms a crosslink between the two cells was verified by a physicochemical experiment.

First, the present inventors investigated, using Alpha technology from Perkin Elmer Inc., construction of a system that can evaluate crosslinking between mouse CD3-immobilized beads and mouse CTLA4-immobilized beads using the anti-mouse CTLA4/anti-mouse CD3 bispecific antibody (hUH02UL01/2C11-F760) purified and prepared by the method of Example 3-1. More specifically, 100 nmol/L of biotinylated mouse CTLA4; 0, 20, 100, or 500 nmol/L of

hUH02UL01/2C11-F760; 50 μg/mL of AlphaScreen (registered trademark) Streptavidin-coated Donor Beads (PerkinElmer); and mouse CD3-acceptor beads prepared by conjugating 50 μg/mL of mouse CD3 to AlphaScreen (registered trademark) Unconjugated Acceptor Beads (PerkinElmer) were used. Under this condition, it was thought that biotinylated mouse CTLA4 bound to AlphaScreen (registered trademark) Streptavidin-coated Donor Beads and mouse CD3-acceptor beads may be crosslinked by hUH02UL01/2C11-F760 as shown in Fig. 7-1, and chemiluminescence may be emitted. Alpha 384 (PerkinElmer) was used as the plate, and the measurements were taken using Envision. All experiments were performed three times. As a result, hUH02UL01/2C11-F760 concentration-dependent crosslinking between beads was observed as shown in Fig. 7-2.

Next, 100 nmol/L of biotinylated mouse CTLA4 and 10 nmol/L of biotinylated mouse CD3 were added to AlphaScreen (registered trademark) Streptavidin-coated Donor Beads (PerkinElmer) to produce donor beads that mimic regulatory T cells expressing CTLA4 and CD3. Whether addition of 100 nmol/L of hUH02UL01/2C11-F760 can cause crosslinking between donor beads and acceptor beads in the presence of 50 μg/mL mouse CD3-acceptor beads (acceptor beads mimicking effector T cells) was examined. The condition in which hUH02UL01/2C11-F760 is not added to 100 nmol/L of biotinylated mouse CTLA4; 50 μg/mL of AlphaScreen (registered trademark) Streptavidin-coated Donor Beads (PerkinElmer); and 50 μg/mL of mouse CD3-acceptor beads was used as the control in which crosslinking between each of the beads are not caused. All experiments were performed three times. The results shown in Fig. 7-3 were obtained from the experiments, and it was confirmed that even under the condition where biotinylated mouse CD3 and mouse CTLA4 may be present on the same bead, the anti-mouse CTLA4/anti-mouse CD3 bispecific antibody crosslinks the donor bead and acceptor bead as shown in Fig. 7-4.

The condition where biotinylated mouse CD3 and mouse CTLA4 are bound on the donor beads was considered to be mimicking regulatory T cells (expressing CTLA4 and CD3), and mouse CD3-conjugated acceptor beads was considered to be mimicking effector T cells (expressing CD3). It was confirmed that the anti-mouse CTLA4/anti-mouse CD3 bispecific antibody crosslinks donor beads with acceptor beads under this condition as well, which suggests that even for regulatory T cells and effector T cells, an anti-CTLA4/anti-CD3 bispecific antibody may be able to form crosslinking between the two cells in a similar manner.

[Example 5] *In vivo* drug efficacy evaluation using an anti-mouse CTLA4/anti-mouse CD3 bispecific antibody (hUH02UL01/2C11-F760) (intratumoral administration)

Whether the anti-mouse CTLA4 antibody hUH02hUL01-mFa55 purified and prepared by the method of Example 2-1 and the anti-mouse CTLA4/anti-mouse CD3 bispecific antibody

(hUH02UL01/2C11-F760) purified and prepared by the method of Example 3-1 shows *in vivo* drug efficacy against a mouse colorectal cancer cell line was verified. 1 x 10^6 mouse colorectal cancer cell line CT26.WT cells (ATCC) were subcutaneously transplanted into the right abdomen of BALB/c mice (Japan Charles River) to establish solid tumor. Ten days after the transplantation, hUH02hUL01-mFa55 was administered at a dose of 200 μg/mouse and hUH02UL01/2C11-F760 was administered at a dose of 100 μg/mouse, intratumorally (i.t.) (n = 2 for each group). The results elucidated that hUH02UL01/2C11-F760 shows stronger antitumor effects in comparison to hUH02hUL01-mFa55, and shows remarkable antitumor effects *in vivo* (Fig. 8). More specifically, the results suggested that hUH02UL01/2C11-F760 recognizes the surface antigens of regulatory T cells (expressing CTLA4 and CD3) and effector T cells (expressing CD3), and causes crosslinking between the two cells *in vivo*.

[Example 6] *In vivo* drug efficacy evaluation of an anti-mouse CTLA4/anti-mouse CD3 bispecific antibody (hUH02UL01/2C11-F760) (comparison between intratumoral administration and intravenous administration)

The anti-mouse CTLA4/anti-mouse CD3 bispecific antibody (hUH02UL01/2C11-F760) purified and prepared by the method of Example 3-1 was assessed on whether it shows drug efficacy on the mouse colorectal cancer cell line CT26.WT-transplanted model described in Example 5 even when administered intravenously (i.v.). 1 x 10^6 CT26.WT cells (ATCC) were subcutaneously transplanted into the right abdomen of BALB/c mice (Japan Charles River) to establish solid tumor. Eight days after the transplantation, hUH02UL01/2C11-F760 was administered at a dose of 100 μg/mouse intratumorally (i.t.) or intravenously (i.v.) (n = 5 for each group). The results elucidated that hUH02UL01/2C11-F760 shows equivalent antitumor effects in both intratumoral and intravenous administration, and shows antitumor effects *in vivo* regardless of whether it is administered locally or systemically (Fig. 9).

[Example 7] *In vitro* drug efficacy evaluation of an anti-human CTLA4/anti-human CD3 bispecific antibody

(7-1) Expression and purification of a bispecific antibody that specifically binds to human CTLA4 and human CD3

Genes encoding the variable regions of the anti-human CTLA4 antibody MDX10-F760nN17 (the heavy chain variable region MDX10H is SEQ ID NO: 38, and the light chain variable region MDX10L is SEQ ID NO: 39) were each inserted into human IgG1/kappa plasmids for expression in animals. Here, constant regions that have been modified so as to reduce binding to Fcγ receptors and to produce heterologous association of two heavy chains were used (the heavy chain constant region F760nN17 is SEQ ID NO: 33, and the light chain

constant region k0 is SEQ ID NO: 34).

Genes encoding the variable regions of the anti-human CD3 antibody TR01H113-F760mG3P17 (the heavy chain variable region TR01H113 is SEQ ID NO: 40, and the light chain variable region L0011 is SEQ ID NO: 41) were each inserted into human IgG1/kappa plasmids for expression in animals. Here, constant regions that have been modified so as to reduce binding to Fcγ receptors and to produce heterologous association of two heavy chains were used (the heavy chain constant region F760nG3P17 is SEQ ID NO: 42, and the light chain constant region k0 is SEQ ID NO: 34).

Each of MDX10-F760nN17 and TR01H113-F760nG3P17 was expressed and purified by the method shown in Example 2. Each of the purified homologous forms was mixed in the combination shown in Table 3, and the bispecific antibody of interest was produced by a method known to those skilled in the art (WO2015/046467).

Table 3

No	Name of clone	Antibody 1	Antibody 2
1	MDX10//TR01H113	MDX10-F760nN17	TR01H113-F760nG3P17

In addition, there are other techniques for forming bispecific antibodies. Examples include methods for antibody production using association of antibody CH1 and CL, and association of VH and VL as described in WO 2011/028952, WO2014/018572, and Nat Biotechnol. 2014 Feb;32(2):191-8; methods using association of CH1 and CL or VH and VL, which are described in Proc Natl Acad Sci U S A. 2011 Jul 5;108(27):11187-92, WO2009/080251, WO2009/080252, and WO2009/080253; methods for regulating association between antibody heavy chain CH3s, which are described in WO2012/058768 and WO2013/063702; methods that utilize charge regulation of CH1 and CL, which are described in WO2006/106905; and methods that utilize charge regulation of VH and VL, which are described in WO2013/065708. The bispecific antibody of interest can be produced by applying the above-mentioned technologies to an anti-human CTLA4 antibody (the heavy chain variable region MDX10H is SEQ ID NO: 38, and the light chain variable region MDX10L is SEQ ID NO: 39) and an anti-human CD3 antibody (the heavy chain variable region TR01H113 is SEQ ID NO: 40, and the light chain variable region L0011 is SEQ ID NO: 41).

(7-2) *In vitro* cytotoxic activity of an anti-human CTLA4/anti-human CD3 bispecific antibody on regulatory T cells

Blood was collected using heparin from two healthy donors. Each blood sample was

diluted with HBSS (GIBCO) containing 5% FBS (Moregate BioTech), and then layered onto Ficoll-Paque Plus (GE healthcare). This was centrifuged at 400 x g for 30 minutes to separate the peripheral blood monocyte (PBMC) fraction. The obtained PBMCs were seeded into a 96-well round-bottom plate (Corning) at 5 x 10^5 cells/well using RPMI 1640 (Nacalai Tesque) medium containing 10% FBS, and 100 Units/mL penicillin - 100 µg/mL Streptomycin (GIBCO).

The control antibody (the anti-KLH human IgG1 heavy chain variable region IC17H is SEQ ID NO: 43, the light chain variable region IC17L is SEQ ID NO: 44, the heavy chain constant region hIgG1d is SEQ ID NO: 45, and the light chain constant region k0 is SEQ ID NO: 34) or MDX10//TR01H113 was diluted with the medium to each produce a final concentration of 0.1 µg/mL, 1 µg/mL, or10 µg/mL, and added to the wells. The cells were cultured for seven days in a CO_2 incubator set at 37°C and 5% CO_2.

Seven days later, the cells were transferred to a V-bottom plate (Corning), and centrifuged at 400 x g for five minutes to remove the supernatant. The cells were resuspended in 100 µL of FcR blocking reagent (Miltenyi Biotec) diluted ten times with PBS containing 1% FBS and 2 mM EDTA (Sigma) (FACS buffer). After incubating at room temperature for ten minutes, 2.5 µL of PerCP-Cy5.5 Mouse Anti-Human CD4 (BD Pharmingen), 5 µL of PE Mouse Anti-Human CD25 (BD Pharmingen), and 2.5 µL of PE-Cy7 Mouse Anti-Human CD45RA (BD Pharmingen) were added to each well. After incubating at 4°C for one hour, 100 µL of FACS buffer was added. Centrifugation was performed at 400 x g for five minutes to remove the supernatant.

Based on the protocol of Intracellular Fixation and Permeabilization buffer set (eBioscience), Human FoxP3 buffer A was added 100 µL at a time, this was incubated at room temperature for ten minutes in the dark. Subsequently, centrifugation was performed at 400 x g for five minutes to remove the supernatant. Permeabilization buffer was added 100 µL at a time, this was incubated at room temperature for 30 minutes in the dark. Next, 100 µL of FACS buffer was added, centrifugation was performed at 400 x g for five minutes to remove the supernatant. This washing procedure was performed one more time.

The cells were resuspended in 100 µL of FACS buffer, Alexa Fluor488 Anti-Human FoxP3 (BioLegend) was added 5 µL at a time, this was incubated at room temperature for 30 minutes in the dark. 100 µL of FACS buffer was added, and centrifugation was performed at 400 x g for five minutes to remove the supernatant. This washing procedure was performed one more time. The cells were resuspended in 200 µL of FACS buffer, and analyzed on a FACS CantoII flow cytometer (BD).

Expression analyses were performed using the FACSDiva Software (BD). CD4-positive cells were gated from the cell population subjected to analysis and the expression of CD25 and CD45RA was analyzed. The $CD25^{high}$ $CD45RA^-$ fraction and the $CD25^-$

$CD45RA^+$ fraction were regarded as the regulatory T cells (Treg) and effector T cells (Teff), respectively. Furthermore, the $FoxP3^{high}CD45RA^-$ fraction and the $FoxP3^-CD45RA^+$ fraction were regarded as Treg and Teff, respectively. The Teff/Treg ratio was calculated from the proportion of Treg and Teff present in CD4-positive cells.

The results of analyzing CD4-positive cells based on the expression of CD25 and CD45RA are shown (Fig. 10-1). In Donor 1, treatment with MDX10//TR01H113 at 1 μg/mL and 10 μg/mL showed decrease in Treg. In Donor 2, treatment at 1 μg/mL showed a decreasing trend in Treg, and treatment at 10 μg/mL showed marked decrease in Treg (Fig. 10-2). In both donors, treatment with MDX10//TR01H113 at 1 μg/mL and 10 μg/mL increased the Teff/Treg ratios (Fig. 10-3).

Furthermore, results of analyzing Treg based on the expression of FoxP3 and CD45RA are shown in Fig. 11-1. Similarly to the analyses using CD25 and CD45RA, treatment with MDX10//TR01H113 decreased Treg and increased the Teff/Treg ratios (Figs. 11-2 and 11-3).

According to these examinations, TRAB (a bispecific antibody against CTLA4 and CD3) which showed stronger antitumor effects with regard to cytotoxic activity against regulatory T cells expressing CTLA4 *in vivo* in mice, also showed strong cytotoxic activity against regulatory T cells *in vitro* in humans; therefore, TRAB is expected to demonstrate strong antitumor effects towards cancer patients.

[Example 8] Analysis of surface molecules expressed in cell fractions ($CD4^+$, $CD25^{high}$, $CD45RA^-$) that have been reported to have high immune response-suppressing function

Among the cells having immune response-suppressing functions, regulatory T cells (Treg) in CD4-positive T cells calculated based on CD25 and CD45RA expression, that is the $CD4^+$ $CD25^{high}$ $CD45RA^-$ cell fraction, have been reported to have high immune response-suppressing functions (Immunity, 2009, 30 (6), 899-911). Based on this information, a gene which encodes a cell surface molecule showing significantly high expression in the $CD25^{high}$ $CD45RA^-$ cell fraction among the CD4-positive cells was identified using RNA-seq. As a result, among CTLA4, PD1, TIM3, LAG3, CD244 (2B4), CD160, GARP, OX40, CD137 (4-1BB), CD25, VISTA, BTLA, TNFR25, CD57, KLRG1, CCR2, CCR5, CCR6, CD39, CD73, CD4, CD18, CD49b, CD1d, CD5, CD21, TIM1, CD19, CD20, CD23, CD24, CD38, CD93, IgM, B220(CD45R), CD317, PD-L1, CD11b, Ly6G, ICAM-1, FAP, PDGFR, Podoplanin, and TIGIT, which are molecules expressed on the surface of cells having immune response-suppressing function, nine molecules which are CTLA4, TIM3, LAG3, CD137 (4-1BB), CD25, CCR5, CCR6, CD38, and TIGIT were found to be cell surface molecules highly expressed specifically in the cell fractions ($CD4^+$, $CD25^{high}$, $CD45RA^-$) that have been reported to have high immune response-suppressing functions.

[Example 9] *In vitro* drug efficacy evaluation of an anti-human LAG3/anti-human CD3 bispecific antibody

(9-1) Expression and purification of a bispecific antibody that specifically binds to human LAG3 and human CD3

Genes encoding the variable regions of the anti-human LAG3 antibody 25F7-F760nN17 (the heavy chain variable region 25F7H is SEQ ID NO: 46, and the light chain variable region 25F7L is SEQ ID NO: 47) were each inserted into human IgG1/kappa plasmids for expression in animals. Here, constant regions that have been modified so as to reduce binding to Fcγ receptors and to produce heterologous association of two heavy chains were used (the heavy chain constant region F760nN17 is SEQ ID NO: 33, and the light chain constant region k0 is SEQ ID NO: 34).

Genes encoding the variable regions of the anti-human CD3 antibody TR01H113-F760nG3P17 (the heavy chain variable region TR01H113 is SEQ ID NO: 40, and the light chain variable region L0011 is SEQ ID NO: 41) were each inserted into human IgG1/kappa plasmids for expression in animals. Here, constant regions that have been modified so as to reduce binding to Fcγ receptors and to produce heterologous association of two heavy chains were used (the heavy chain constant region F760nG3P17 is SEQ ID NO: 42, and the light chain constant region k0 is SEQ ID NO: 34).

Each of 25F7-F760nN17 and TR01H113-F760nG3P17 was expressed and purified by the method shown in Example 2. Each of the purified homologous forms was mixed in the combination shown in Table 4, and the bispecific antibody of interest was produced by a method known to those skilled in the art (WO2015/046467).

Table 4

No	Name of clone	Antibody 1	Antibody 2
1	25F7//TR01H113	25F7-F760nN17	TR01H113-F760nG3P17

(9-2) *In vitro* cytotoxic activity of an anti-human LAG3/anti-human CD3 bispecific antibody on regulatory T cells

Blood was collected using heparin from healthy donors. Each blood sample was diluted with PBS and then layered together with Ficoll-Paque Plus (GE healthcare) in a Leucosep tube (greiner bio-one). This was centrifuged at 1000 x g for 10 minutes to separate the peripheral blood monocyte (PBMC) fraction. The obtained PBMCs were seeded into a 96-well round bottom plate (Corning) at 1 x 10^6 cells/well using RPMI 1640 (Nacalai Tesque)

medium containing 10% FBS, and 100 Units/mL penicillin - 100 μg/mL Streptomycin (GIBCO).

TRAB (25F7//TR01H113) was diluted with the medium at a final concentration of 1 μg/mL or 10 μg/mL, and added to the wells. The cells were cultured for four or six days in a CO_2 incubator set at 37°C and 5% CO_2.

Four or six days later, the cells were transferred to tubes for FACS analysis, and centrifuged at 400 x g for five minutes to remove the supernatant. Cell WASH (BD Biosciences) containing 0.2% BSA (Wako) was prepared, and this was used as the FACS Buffer. For complete removal of medium components, washing was performed by adding 2 mL of FACS Buffer to the cells from which the supernatant was removed and performing the centrifugation again at 400 x g for five minutes to remove the supernatant.

FcR blocking reagent (Miltenyi Biotec) diluted ten-fold with the FACS Buffer, to which 1/1000 volume of eFluor780 (eBioscience) for staining dead cells was added, was prepared and used as the Staining Buffer. Solution produced by adding 5 μL of PerCP Mouse Anti-Human CD4 (BD Pharmingen), 2.5 μL of PE-Cy™7 Mouse Anti-Human CD45RA (BD Pharmingen), and 5 μL of PE Mouse Anti-Human CD25 to 50 μL of the Staining Buffer was placed into each tube. After incubation at 4°C for one hour, 2 mL of FACS buffer was added, and the centrifugation was performed at 400 x g for five minutes to remove the supernatant. Then, as a washing procedure, an additional 2 mL of FACS buffer was added, and the centrifugation was performed at 400 x g for five minutes to remove the supernatant. The cells were resuspended in 400 μL of FACS buffer and analyzed on a FACSVerse™ flow cytometer (BD).

Expression analysis was carried out using the FACSDiva Software (BD). CD4-positive cells were gated from the cell population subjected for analysis, from which dead cells had been removed, and the expression of CD25 and CD45RA was analyzed. The $CD25^{high}$ $CD45RA^{-}$ fraction and the $CD25^{-}$ $CD45RA^{+}$ fraction were regarded as regulatory T cells (Treg) and effector T cells (Teff), respectively. The Teff/Treg ratio was calculated from the proportion of Treg and Teff present in CD4-positive cells.

The results of analyzing CD4-positive cells based on the expression of CD25 and CD45RA are shown (Fig. 13). Treatment with TRAB (25F7//TR01H113) at 1 μg/mL and 10 μg/mL showed decrease in Treg in a TRAB antibody dose-dependent manner, both at four and six days after the treatment (Fig. 14). Treatment with TRAB (25F7//TR01H113) at 1 μg/mL and 10 μg/mL increased the Teff/Treg ratios (Fig. 15).

[Example 10] *In vitro* drug efficacy evaluation of an anti-human OX40/anti-human CD3 bispecific antibody

(10-1) Expression and purification of a bispecific antibody that specifically binds to human OX40 and human CD3

Genes encoding the variable regions of the anti-human OX40 antibody 12H3-F760nN17 (the heavy chain variable region 12H3VH is SEQ ID NO: 48, and the light chain variable region 12H3VL is SEQ ID NO: 49) were each inserted into human IgG1/kappa plasmids for expression in animals. Here, constant regions that have been modified so as to reduce binding to Fcγ receptors and to produce heterologous association of two heavy chains were used (the heavy chain constant region F760nN17 is SEQ ID NO: 33, and the light chain constant region k0 is SEQ ID NO: 34).

Genes encoding the variable regions of the anti-human CD3 antibody TR01H113-F760nG3P17 (the heavy chain variable region TR01H113 is SEQ ID NO: 40, and the light chain variable region L0011 is SEQ ID NO: 41) were each inserted into human IgG1/kappa plasmids for expression in animals. Here, constant regions that have been modified so as to reduce binding to Fcγ receptors and to produce heterologous association of two heavy chains were used (the heavy chain constant region F760nG3P17 is SEQ ID NO: 42, and the light chain constant region k0 is SEQ ID NO: 34).

Each of 12H3-F760nN17 and TR01H113-F760nG3P17 was expressed and purified by the method shown in Example 2. Each of the purified homologous forms was mixed in the combination shown in Table 5, and the bispecific antibody of interest was produced by a method known to those skilled in the art (WO2015/046467).

Table 5

No	Name of clone	Antibody 1	Antibody 2
1	12H3//TR01H113	12H3-F760nN17	TR01H113-F760nG3P17

(10-2) *In vitro* cytotoxic activity of an anti-human OX40/anti-human CD3 bispecific antibody on regulatory T cells

Blood was collected using heparin from two healthy donors. Each blood sample was diluted with PBS and then layered together with Ficoll-Paque Plus (GE healthcare) in a Leucosep tube (greiner bio-one). The centrifugation was performed at 1000 x g for 10 minutes to separate the peripheral blood monocyte (PBMC) fraction. The obtained PBMCs were seeded into a 96-well round bottom plate (Corning) at 1 x 10^6 cells/well using RPMI 1640 (Nacalai Tesque) medium containing 10% FBS, and 100 Units/mL penicillin - 100 μg/mL Streptomycin (GIBCO).

TRAB (12H3//TR01H113) was diluted with the medium at a final concentration of 1 μg/mL or 10 μg/mL, and was added to the wells. The cells were cultured for seven days in a CO_2 incubator set at 37°C and 5% CO_2.

Seven days later, the cells were transferred to tubes for FACS analysis, and centrifuged at 400 x g for five minutes to remove the supernatant. Cell WASH (BD Biosciences) containing 0.2% BSA (Wako) was prepared and used as the FACS Buffer. For complete removal of medium components, washing was performed by adding 2 mL of FACS Buffer to the cells from which the supernatant was removed and performing the centrifugation again at 400 x g for five minutes to remove the supernatant.

FcR blocking reagent (Miltenyi Biotec) diluted ten-fold with the FACS Buffer, to which 1/1000 volume of eFluor780 (eBioscience) for staining dead cells was added, was prepared and used as the Staining Buffer. Solution produced by adding 5 μL of PerCP Mouse Anti-Human CD4 (BD Pharmingen), 2.5 μL of PE-Cy^{TM}7 Mouse Anti-Human CD45RA (BD Pharmingen), and 5 μL of PE Mouse Anti-Human CD25 to 50 μL of the Staining Buffer was placed into each tube. After incubation at 4°C for one hour, 2 mL of FACS Buffer was added, and the centrifugation was performed at 400 x g for five minutes to remove the supernatant. Then, as a washing procedure, 2 mL of FACS buffer was further added, and the centrifugation was performed at 400 x g for five minutes to remove the supernatant. The cells were resuspended in 400 μL of FACS buffer and analyzed on a FACSVerse^{TM} flow cytometer (BD).

Expression analysis was carried out using the FACSDiva Software (BD). CD4-positive cells were gated from the cell population subjected to analysis, from which dead cells had been removed, and the expression of CD25 and CD45RA was analyzed. The CD25^{high} CD45RA^{-} fraction and the CD25^{-} CD45RA^{+} fraction were regarded as regulatory T cells (Treg) and effector T cells (Teff), respectively. The Teff/Treg ratio was calculated from the proportion of Treg and Teff present in CD4-positive cells.

The results of analyzing CD4-positive cells based on the expression of CD25 and CD45RA are shown (Fig. 16). Treatment of PBMCs derived from both donors with TRAB (12H3//TR01H113) at 1 μg/mL and 10 μg/mL showed decrease in Treg in a TRAB antibody dose-dependent manner (Fig. 17). Treatment with TRAB (12H3//TR01H113) at 1 μg/mL and 10 μg/mL increased the Teff/Treg ratios (Fig. 18).

[Reference Example 1] ADCC activity of a test antibody using mouse FcgR4-expressing human NK cell line NK-92 as the effector cells

Regarding anti-mouse CTLA4 antibodies, antibody concentration-dependent ADCC activities of test antibodies were measured, using the mouse FcgR4-expressing human NK cell line NK-92 (hereinafter referred to as mFcgR4-NK92) as effector cells by following the method described below.

(1) Preparation of mFcgR4-NK92 solution

After washing mFcgR4-NK92 with RPMI-1640 (nacalai tesque) containing 10% FBS

(hereinafter referred to as 10% FBS/RPMI), the cells were suspended in 10% FBS/RPMI at a cell density of 4 x 10^5 cells/ml. This cell suspension solution was used as the mFcgR4-NK92 solution in the subsequent experiments.

(2) Preparation of target cells

To 2 x 10^6 CHO/mouse CTLA4 cells which are CHO cells forced to express mouse CTLA4, 3.7 MBq of Cr-51 was added. The Cr-51-added cells were incubated in a 5% carbon dioxide gas incubator at 37°C for one hour, then washed three times with 10% FBS/RPMI, and then suspended in 10% FBS/RPMI at a cell density of 2 x 10^5 cells/ml. The cell suspension was used as the target cells in the subsequent experiments.

(3) Chromium-release Assay (ADCC activity)

The ADCC activities were evaluated from the specific chromium release rate according to the chromium release method. First, antibody solutions prepared at each concentration (0, 0.04, 0.4, 4, and 40 μg/ml) were added to a 96-well U-bottomed plate at 50 μl per well. Next, the target cells prepared in (2) were seeded at 50 μl per well (1 x 10^4 cells/well). Furthermore, 10% FBS/RPMI was added at 50 μl per well, and the plate was allowed to stand at room temperature for 15 minutes. The mFcgR4-NK92 solution prepared in (1) was added at 50 μl per well (2 x 10^4 cells/well), the plate was left to stand in a 5% carbon dioxide gas incubator at 37°C for four hours, and resultant was centrifuged. The radioactivity of 100 μl of culture supernatant in each well of the plate was measured using a gamma counter. The specific chromium release rate was determined based on the following equation.

Chromium release rate (%) = (A-C) x 100 / (B-C)

In this equation, "A" represents the mean value of radioactivity (cpm) of 100 μl of culture supernatant in each well; "B" represents the mean value of radioactivity (cpm) of 100 μl of culture supernatant in a well where 50 μl of a 4% NP-40 aqueous solution (Nonidet P-40, Nacalai Tesque) and 100 μl of 10% FBS/RPMI had been added to the target cells; and "C" represents the mean value of radioactivity (cpm) of 100 μl of culture supernatant in a well where 150 μl of 10% FBS/RPMI had been added to the target cells. The examinations were performed in duplicate and the mean value for the specific chromium release rate (%) of the test antibody was calculated.

[Reference Example 2] Measurement of cytotoxic activity of a test antibody using mouse splenocytes as effector cells

Regarding the anti-mouse CTLA4/anti-mouse CD3 bispecific antibody (hUH02UL01/2C11-F760), antibody concentration-dependent cytotoxic activity of the test antibody was measured using mouse splenocytes as effector cells, and by following the method described below.

(1) Preparation of mouse splenocyte solution

Ten mL of 10% FBS/RPMI was added to the spleen excised from a BALB/c mouse. The spleen was sliced into small pieces and passed through a cell strainer. After centrifugation (at 2,150 rpm for ten minutes at room temperature), a hemolysis procedure was performed using a Mouse Erythrocyte Lysing Kit (R&D Systems). After washing once with 10% FBS/RPMI, the cells were suspended in 10% FBS/RPMI at a cell density of 6×10^6 cells/ml. The cell suspension was used as the mouse splenocyte solution in the subsequent experiments.

(2) Cytotoxic activity evaluation assay

Cytotoxic activity was evaluated by cell proliferation inhibition rate using the xCELLigence Real-Time Cell Analyzer (Roche Diagnostics K.K.). CHO/mCTLA4 which is CHO forced to express mouse CTLA4 was used as the target cells. The cells were suspended in CHO-S-SFM II (Life technologies) containing 10% FBS, and were seeded in aliquots of 100 µl into an E-Plate 96 plate (Roche Diagnostics K.K.) at 5×10^3 cells/well. Measurement of living cells was started using the xCELLigence Real-Time Cell Analyzer. On the following day, the plate was removed from the xCELLigence Real-Time Cell Analyzer, and 50 µl of the respective antibodies prepared at each concentration (0.04, 0.4, 4, or 40 µg/ml) were added to the plate. After allowing this to stand at room temperature for 15 minutes, 50 µl of the mouse splenocyte solution prepared in (1) was added (3×10^5 cells/well). By setting the plate into the xCELLigence Real-Time Cell Analyzer again, measurement of living cells was started. The reaction was carried out in a 5% carbon dioxide gas incubator at 37°C, and from the Cell Index value obtained 72 hours after addition of the mouse splenocyte solution, the cell proliferation inhibition rate (%) was determined using the following equation. The Cell Index value used in the calculation was a normalized value where the Cell Index value immediately before antibody addition was defined as 1.

Cell proliferation inhibition rate (%) = (A-B) x 100/(A-1)

"A" represents the mean value of the Cell Index values in wells without antibody addition (containing only the target cells and human PBMC), and "B" represents the mean value of the Cell Index values in each well. The examinations were performed in duplicate.

Industrial Applicability

The present invention provides novel antigen-binding molecules or pharmaceutical compositions having excellent antitumor activity. Pharmaceutical compositions comprising the antigen-binding molecules of the present invention as active ingredients inhibit immune response-suppressing functions, and thereby activate cytotoxic effects and enable treatment and prevention of various cancers.

CLAIMS

1. An antigen-binding molecule that inhibits immune response-suppressing activity of a cell having an immune response-suppressing function, comprising:
(i) a domain that binds to a molecule expressed on the surface of a cell having an immune response-suppressing function; and
(ii) a T cell receptor complex-binding domain.

2. The antigen-binding molecule of claim 1, wherein the cell having immune response-suppressing function is a regulatory T cell or an exhausted T cell.

3. The antigen-binding molecule of claim 1, wherein the molecule expressed on the surface of a cell having an immune response-suppressing function is any molecule selected from among CTLA4, PD1, TIM3, LAG3, CD244 (2B4), CD160, GARP, OX40, CD137 (4-1BB), CD25, VISTA, BTLA, TNFR25, CD57, KLRG1, CCR2, CCR5, CCR6, CD39, CD73, CD4, CD18, CD49b, CD1d, CD5, CD21, TIM1, CD19, CD20, CD23, CD24, CD38, CD93, IgM, B220(CD45R), CD317, PD-L1, CD11b, Ly6G, ICAM-1, FAP, PDGFR, Podoplanin, and TIGIT.

4. The antigen-binding molecule of claim 3, wherein the molecule expressed on the surface of a cell having an immune response-suppressing function is any molecule selected from among CTLA4, TIM3, LAG3, CD137 (4-1BB), CD25, CCR5, CCR6, CD38, TIGIT, and OX40.

5. The antigen-binding molecule of any one of claims 1 to 4, wherein the T cell receptor complex-binding domain is a T cell receptor-binding domain.

6. The antigen-binding molecule of any one of claims 1 to 4, wherein the T cell receptor complex-binding domain is a CD3-binding domain.

7. The antigen-binding molecule of any one of claims 1 to 6, which further comprises an FcRn-binding domain.

8. The antigen-binding molecule of claim 7, wherein the FcRn-binding domain is an Fc region of an antibody.

9. The antigen-binding molecule of claim 8, wherein the FcRn-binding domain is an Fc region of an antibody having reduced Fcγ receptor-binding activity.

10. The antigen-binding molecule of any one of claims 1 to 9, which is a multispecific antibody.

11. A pharmaceutical composition comprising the antigen-binding molecule of any one of claims 1 to 10 as an active ingredient.

12. A pharmaceutical composition for inhibiting an immune response-suppressing activity, comprising the antigen-binding molecule of any one of claims 1 to 10 as an active ingredient.

13. The pharmaceutical composition of claim 12, wherein the inhibition of an immune response-suppressing activity is inhibition caused by a T cell.

14. The pharmaceutical composition of claim 13, wherein the inhibition caused by a T cell is inhibition resulting from cytotoxic activity of the T cell.

15. The pharmaceutical composition of any one of claims 12 to 14, wherein the immune response-suppressing activity is an activity by a regulatory T cell or an exhausted T cell.

16. The pharmaceutical composition of any one of claims 12 to 14, wherein the immune response-suppressing activity is an activity of a cell expressing any molecule selected from among CTLA4, PD1, TIM3, LAG3, CD244 (2B4), CD160, GARP, OX40, CD137 (4-1BB), CD25, VISTA, BTLA, TNFR25, CD57, KLRG1, CCR2, CCR5, CCR6, CD39, CD73, CD4, CD18, CD49b, CD1d, CD5, CD21, TIM1, CD19, CD20, CD23, CD24, CD38, CD93, IgM, B220(CD45R), CD317, PD-L1, CD11b, Ly6G, ICAM-1, FAP, PDGFR, Podoplanin, and TIGIT.

17. The pharmaceutical composition of any one of claims 11 to 16, which is a therapeutic agent for cancer.

18. The pharmaceutical composition of claim 17, which is a therapeutic agent for any cancer selected from among ovarian cancer, gastric cancer, esophageal cancer, pancreatic cancer, renal cell carcinoma, hepatocellular carcinoma, breast cancer, malignant melanoma, non-small-cell lung cancer, cervical cancer, glioblastoma, prostate cancer, neuroblastoma, chronic lymphocytic leukemia, papillary thyroid cancer, colorectal cancer, Hodgkin’s lymphoma, and endometriosis.

SEQUENCE LISTING

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OSAKA UNIVERSITY

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Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val
130 135 140

Asp Val Ser Gln Glu Asp Pro Glu Val Gln Phe Asn Trp Tyr Val Asp
145 150 155 160

Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Phe
165 170 175

Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp
180 185 190

Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Gly Leu
195 200 205

Pro Ser Ser Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg
210 215 220

```
Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Gln Glu Glu Met Thr Lys
225                 230                 235                 240

Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp
                245                 250                 255

Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys
            260                 265                 270

Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser
        275                 280                 285

Arg Leu Thr Val Asp Lys Ser Arg Trp Gln Glu Gly Asn Val Phe Ser
    290                 295                 300

Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser
305                 310                 315                 320

Leu Ser Leu Ser Leu Gly Lys
                325
```

<210> 9
<211> 141
<212> PRT
<213> Homo sapiens

<400> 9

```
Asp Ile Gln Asn Pro Asp Pro Ala Val Tyr Gln Leu Arg Asp Ser Lys
```

1 5 10 15

Ser Ser Asp Lys Ser Val Cys Leu Phe Thr Asp Phe Asp Ser Gln Thr
20 25 30

Asn Val Ser Gln Ser Lys Asp Ser Asp Val Tyr Ile Thr Asp Lys Thr
35 40 45

Val Leu Asp Met Arg Ser Met Asp Phe Lys Ser Asn Ser Ala Val Ala
50 55 60

Trp Ser Asn Lys Ser Asp Phe Ala Cys Ala Asn Ala Phe Asn Asn Ser
65 70 75 80

Ile Ile Pro Glu Asp Thr Phe Phe Pro Ser Pro Glu Ser Ser Cys Asp
85 90 95

Val Lys Leu Val Glu Lys Ser Phe Glu Thr Asp Thr Asn Leu Asn Phe
100 105 110

Gln Asn Leu Ser Val Ile Gly Phe Arg Ile Leu Leu Leu Lys Val Ala
115 120 125

Gly Phe Asn Leu Leu Met Thr Leu Arg Leu Trp Ser Ser
130 135 140

<210> 10

<211> 179
<212> PRT
<213> Homo sapiens

<400> 10

```
Glu Asp Leu Lys Asn Val Phe Pro Pro Glu Val Ala Val Phe Glu Pro
1               5                  10                  15

Ser Glu Ala Glu Ile Ser His Thr Gln Lys Ala Thr Leu Val Cys Leu
            20                  25                  30

Ala Thr Gly Phe Tyr Pro Asp His Val Glu Leu Ser Trp Trp Val Asn
        35                  40                  45

Gly Lys Glu Val His Ser Gly Val Ser Thr Asp Pro Gln Pro Leu Lys
    50                  55                  60

Glu Gln Pro Ala Leu Asn Asp Ser Arg Tyr Cys Leu Ser Ser Arg Leu
65                  70                  75                  80

Arg Val Ser Ala Thr Phe Trp Gln Asn Pro Arg Asn His Phe Arg Cys
                85                  90                  95

Gln Val Gln Phe Tyr Gly Leu Ser Glu Asn Asp Glu Trp Thr Gln Asp
            100                 105                 110

Arg Ala Lys Pro Val Thr Gln Ile Val Ser Ala Glu Ala Trp Gly Arg
        115                 120                 125
```

Ala Asp Cys Gly Phe Thr Ser Glu Ser Tyr Gln Gln Gly Val Leu Ser
130 135 140

Ala Thr Ile Leu Tyr Glu Ile Leu Leu Gly Lys Ala Thr Leu Tyr Ala
145 150 155 160

Val Leu Val Ser Ala Leu Val Leu Met Ala Met Val Lys Arg Lys Asp
165 170 175

Ser Arg Gly

<210> 11
<211> 173
<212> PRT
<213> Homo sapiens

<400> 11

Asp Lys Gln Leu Asp Ala Asp Val Ser Pro Lys Pro Thr Ile Phe Leu
1 5 10 15

Pro Ser Ile Ala Glu Thr Lys Leu Gln Lys Ala Gly Thr Tyr Leu Cys
20 25 30

Leu Leu Glu Lys Phe Phe Pro Asp Val Ile Lys Ile His Trp Gln Glu
35 40 45

```
Lys Lys Ser Asn Thr Ile Leu Gly Ser Gln Glu Gly Asn Thr Met Lys
    50                  55                  60

Thr Asn Asp Thr Tyr Met Lys Phe Ser Trp Leu Thr Val Pro Glu Lys
65                  70                  75                  80

Ser Leu Asp Lys Glu His Arg Cys Ile Val Arg His Glu Asn Asn Lys
                85                  90                  95

Asn Gly Val Asp Gln Glu Ile Ile Phe Pro Pro Ile Lys Thr Asp Val
            100                 105                 110

Ile Thr Met Asp Pro Lys Asp Asn Cys Ser Lys Asp Ala Asn Asp Thr
        115                 120                 125

Leu Leu Leu Gln Leu Thr Asn Thr Ser Ala Tyr Tyr Met Tyr Leu Leu
    130                 135                 140

Leu Leu Leu Lys Ser Val Val Tyr Phe Ala Ile Ile Thr Cys Cys Leu
145                 150                 155                 160

Leu Arg Arg Thr Ala Phe Cys Cys Asn Gly Glu Lys Ser
                165                 170
```

<210> 12
<211> 204
<212> PRT

<213> Homo sapiens

<400> 12

```
Lys Gln Leu Asp Ala Asp Val Ser Pro Lys Pro Thr Ile Phe Leu Pro
1               5                  10                  15

Ser Ile Ala Glu Thr Lys Leu Gln Lys Ala Gly Thr Tyr Leu Cys Leu
            20                  25                  30

Leu Glu Lys Phe Phe Pro Asp Ile Ile Lys Ile His Trp Gln Glu Lys
        35                  40                  45

Lys Ser Asn Thr Ile Leu Gly Ser Gln Glu Gly Asn Thr Met Lys Thr
    50                  55                  60

Asn Asp Thr Tyr Met Lys Phe Ser Trp Leu Thr Val Pro Glu Glu Ser
65                  70                  75                  80

Leu Asp Lys Glu His Arg Cys Ile Val Arg His Glu Asn Asn Lys Asn
                85                  90                  95

Gly Ile Asp Gln Glu Ile Ile Phe Pro Pro Ile Lys Thr Asp Val Thr
            100                 105                 110

Thr Val Asp Pro Lys Asp Ser Tyr Ser Lys Asp Ala Asn Asp Val Thr
        115                 120                 125
```

Thr Val Asp Pro Lys Tyr Asn Tyr Ser Lys Asp Ala Asn Asp Val Ile
130 135 140

Thr Met Asp Pro Lys Asp Asn Trp Ser Lys Asp Ala Asn Asp Thr Leu
145 150 155 160

Leu Leu Gln Leu Thr Asn Thr Ser Ala Tyr Tyr Met Tyr Leu Leu Leu
165 170 175

Leu Leu Lys Ser Val Val Tyr Phe Ala Ile Ile Thr Cys Cys Leu Leu
180 185 190

Gly Arg Thr Ala Phe Cys Cys Asn Gly Glu Lys Ser
195 200

<210> 13
<211> 177
<212> PRT
<213> Homo sapiens

<400> 13

Pro Ser Tyr Thr Gly Gly Tyr Ala Asp Lys Leu Ile Phe Gly Lys Gly
1 5 10 15

Thr Arg Val Thr Val Glu Pro Arg Ser Gln Pro His Thr Lys Pro Ser
20 25 30

Val Phe Val Met Lys Asn Gly Thr Asn Val Ala Cys Leu Val Lys Glu

35 40 45

Phe Tyr Pro Lys Asp Ile Arg Ile Asn Leu Val Ser Ser Lys Lys Ile
50 55 60

Thr Glu Phe Asp Pro Ala Ile Val Ile Ser Pro Ser Gly Lys Tyr Asn
65 70 75 80

Ala Val Lys Leu Gly Lys Tyr Glu Asp Ser Asn Ser Val Thr Cys Ser
85 90 95

Val Gln His Asp Asn Lys Thr Val His Ser Thr Asp Phe Glu Val Lys
100 105 110

Thr Asp Ser Thr Asp His Val Lys Pro Lys Glu Thr Glu Asn Thr Lys
115 120 125

Gln Pro Ser Lys Ser Cys His Lys Pro Lys Ala Ile Val His Thr Glu
130 135 140

Lys Val Asn Met Met Ser Leu Thr Val Leu Gly Leu Arg Met Leu Phe
145 150 155 160

Ala Lys Thr Val Ala Val Asn Phe Leu Leu Thr Ala Lys Leu Phe Phe
165 170 175

Leu

<210> 14
<211> 549
<212> DNA
<213> Homo sapiens

<400> 14
atggaacagg ggaagggcct ggctgtcctc atcctggcta tcattcttct tcaaggtact 60

ttggcccagt caatcaaagg aaaccacttg gttaaggtgt atgactatca agaagatggt 120

tcggtacttc tgacttgtga tgcagaagcc aaaaatatca catggtttaa agatgggaag 180

atgatcggct tcctaactga agataaaaaa aaatggaatc tgggaagtaa tgccaaggac 240

cctcgaggga tgtatcagtg taaaggatca cagaacaagt caaaaccact ccaagtgtat 300

tacagaatgt gtcagaactg cattgaacta aatgcagcca ccatatctgg ctttctcttt 360

gctgaaatcg tcagcatttt cgtccttgct gttggggtct acttcattgc tggacaggat 420

ggagttcgcc agtcgagagc ttcagacaag cagactctgt tgcccaatga ccagctctac 480

cagcccctca aggatcgaga agatgaccag tacagccacc ttcaaggaaa ccagttgagg 540

aggaattga 549

<210> 15
<211> 182
<212> PRT
<213> Homo sapiens

<400> 15

```
Met Glu Gln Gly Lys Gly Leu Ala Val Leu Ile Leu Ala Ile Ile Leu
1               5                  10                  15

Leu Gln Gly Thr Leu Ala Gln Ser Ile Lys Gly Asn His Leu Val Lys
            20                  25                  30

Val Tyr Asp Tyr Gln Glu Asp Gly Ser Val Leu Leu Thr Cys Asp Ala
        35                  40                  45

Glu Ala Lys Asn Ile Thr Trp Phe Lys Asp Gly Lys Met Ile Gly Phe
    50                  55                  60

Leu Thr Glu Asp Lys Lys Lys Trp Asn Leu Gly Ser Asn Ala Lys Asp
65                  70                  75                  80

Pro Arg Gly Met Tyr Gln Cys Lys Gly Ser Gln Asn Lys Ser Lys Pro
                85                  90                  95

Leu Gln Val Tyr Tyr Arg Met Cys Gln Asn Cys Ile Glu Leu Asn Ala
            100                 105                 110

Ala Thr Ile Ser Gly Phe Leu Phe Ala Glu Ile Val Ser Ile Phe Val
        115                 120                 125

Leu Ala Val Gly Val Tyr Phe Ile Ala Gly Gln Asp Gly Val Arg Gln
    130                 135                 140
```

Ser Arg Ala Ser Asp Lys Gln Thr Leu Leu Pro Asn Asp Gln Leu Tyr
145 150 155 160

Gln Pro Leu Lys Asp Arg Glu Asp Asp Gln Tyr Ser His Leu Gln Gly
165 170 175

Asn Gln Leu Arg Arg Asn
180

<210> 16
<211> 516
<212> DNA
<213> Homo sapiens

<400> 16
atggaacata gcacgtttct ctctggcctg gtactggcta cccttctctc gcaagtgagc 60

cccttcaaga tacctataga ggaacttgag gacagagtgt ttgtgaattg caataccagc 120

atcacatggg tagagggaac ggtgggaaca ctgctctcag acattacaag actggacctg 180

ggaaaacgca tcctggaccc acgaggaata tataggtgta atgggacaga tatatacaag 240

gacaaagaat ctaccgtgca agttcattat cgaatgtgcc agagctgtgt ggagctggat 300

ccagccaccg tggctggcat cattgtcact gatgtcattg ccactctgct ccttgctttg 360

ggagtcttct gctttgctgg acatgagact ggaaggctgt ctggggctgc cgacacacaa 420

gctctgttga ggaatgacca ggtctatcag cccctccgag atcgagatga tgctcagtac 480

agccaccttg gaggaaactg ggctcggaac aagtga 516

<210> 17
<211> 171
<212> PRT
<213> Homo sapiens

<400> 17

```
Met Glu His Ser Thr Phe Leu Ser Gly Leu Val Leu Ala Thr Leu Leu
1               5                  10                  15

Ser Gln Val Ser Pro Phe Lys Ile Pro Ile Glu Glu Leu Glu Asp Arg
            20                  25                  30

Val Phe Val Asn Cys Asn Thr Ser Ile Thr Trp Val Glu Gly Thr Val
        35                  40                  45

Gly Thr Leu Leu Ser Asp Ile Thr Arg Leu Asp Leu Gly Lys Arg Ile
    50                  55                  60

Leu Asp Pro Arg Gly Ile Tyr Arg Cys Asn Gly Thr Asp Ile Tyr Lys
65                  70                  75                  80

Asp Lys Glu Ser Thr Val Gln Val His Tyr Arg Met Cys Gln Ser Cys
                85                  90                  95

Val Glu Leu Asp Pro Ala Thr Val Ala Gly Ile Ile Val Thr Asp Val
            100                 105                 110
```

Ile Ala Thr Leu Leu Leu Ala Leu Gly Val Phe Cys Phe Ala Gly His
115 120 125

Glu Thr Gly Arg Leu Ser Gly Ala Ala Asp Thr Gln Ala Leu Leu Arg
130 135 140

Asn Asp Gln Val Tyr Gln Pro Leu Arg Asp Arg Asp Asp Ala Gln Tyr
145 150 155 160

Ser His Leu Gly Gly Asn Trp Ala Arg Asn Lys
165 170

<210> 18
<211> 624
<212> DNA
<213> Homo sapiens

<400> 18
atgcagtcgg gcactcactg gagagttctg ggcctctgcc tcttatcagt tggcgtttgg 60

gggcaagatg gtaatgaaga aatgggtggt attacacaga caccatataa agtctccatc 120

tctggaacca cagtaatatt gacatgccct cagtatcctg gatctgaaat actatggcaa 180

cacaatgata aaaacatagg cggtgatgag gatgataaaa acataggcag tgatgaggat 240

cacctgtcac tgaaggaatt ttcagaattg gagcaaagtg gttattatgt ctgctacccc 300

agaggaagca aaccagaaga tgcgaacttt tatctctacc tgagggcaag agtgtgtgag 360

aactgcatgg agatggatgt gatgtcggtg gccacaattg tcatagtgga catctgcatc 420

actgggggct tgctgctgct ggtttactac tggagcaaga atagaaaggc caaggccaag 480

cctgtgacac gaggagcggg tgctggcggc aggcaaaggg gacaaaacaa ggagaggcca 540

ccacctgttc ccaacccaga ctatgagccc atccggaaag gccagcggga cctgtattct 600

ggcctgaatc agagacgcat ctga 624

<210> 19
<211> 207
<212> PRT
<213> Homo sapiens

<400> 19

Met Gln Ser Gly Thr His Trp Arg Val Leu Gly Leu Cys Leu Leu Ser
1 5 10 15

Val Gly Val Trp Gly Gln Asp Gly Asn Glu Glu Met Gly Gly Ile Thr
20 25 30

Gln Thr Pro Tyr Lys Val Ser Ile Ser Gly Thr Thr Val Ile Leu Thr
35 40 45

Cys Pro Gln Tyr Pro Gly Ser Glu Ile Leu Trp Gln His Asn Asp Lys
50 55 60

Asn Ile Gly Gly Asp Glu Asp Asp Lys Asn Ile Gly Ser Asp Glu Asp
65 70 75 80

```
His Leu Ser Leu Lys Glu Phe Ser Glu Leu Glu Gln Ser Gly Tyr Tyr
                85                  90                  95

Val Cys Tyr Pro Arg Gly Ser Lys Pro Glu Asp Ala Asn Phe Tyr Leu
            100                 105                 110

Tyr Leu Arg Ala Arg Val Cys Glu Asn Cys Met Glu Met Asp Val Met
        115                 120                 125

Ser Val Ala Thr Ile Val Ile Val Asp Ile Cys Ile Thr Gly Gly Leu
    130                 135                 140

Leu Leu Leu Val Tyr Tyr Trp Ser Lys Asn Arg Lys Ala Lys Ala Lys
145                 150                 155                 160

Pro Val Thr Arg Gly Ala Gly Ala Gly Gly Arg Gln Arg Gly Gln Asn
                165                 170                 175

Lys Glu Arg Pro Pro Pro Val Pro Asn Pro Asp Tyr Glu Pro Ile Arg
            180                 185                 190

Lys Gly Gln Arg Asp Leu Tyr Ser Gly Leu Asn Gln Arg Arg Ile
        195                 200                 205
```

<210> 20
<211> 4
<212> PRT

<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 20

Gly Gly Gly Ser
1

<210> 21
<211> 4
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 21

Ser Gly Gly Gly
1

<210> 22
<211> 5
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 22

Gly Gly Gly Gly Ser

1 5

<210> 23
<211> 5
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 23

Ser Gly Gly Gly Gly
1 5

<210> 24
<211> 6
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 24

Gly Gly Gly Gly Gly Ser
1 5

<210> 25
<211> 6
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 25

Ser Gly Gly Gly Gly Gly
1 5

<210> 26
<211> 7
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 26

Gly Gly Gly Gly Gly Gly Ser
1 5

<210> 27
<211> 7
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 27

Ser Gly Gly Gly Gly Gly Gly
1 5

<210> 28
<211> 116
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 28

```
Gln Val Gln Leu Gln Glu Ser Gly Pro Gly Leu Val Lys Pro Ser Glu
1               5                  10                  15

Thr Leu Ser Leu Thr Cys Thr Val Ser Gly Tyr Ser Ile Ser Ser Gly
            20                  25                  30

Tyr Gly Trp Asn Trp Ile Arg Gln Pro Pro Gly Lys Gly Leu Glu Trp
        35                  40                  45

Ile Gly Phe Ile Tyr Tyr Glu Gly Ser Thr Tyr Tyr Asn Pro Ser Ile
    50                  55                  60

Lys Ser Arg Ile Ser Ile Thr Arg Asp Thr Ser Lys Asn Gln Phe Phe
65                  70                  75                  80

Leu Gln Val Asn Ser Val Thr Thr Glu Asp Thr Ala Thr Tyr Tyr Cys
                85                  90                  95

Ala Arg Gln Thr Gly Tyr Phe Asp Tyr Trp Gly Gln Gly Thr Leu Val
```

100 105 110

Thr Val Ser Ser
115

<210> 29
<211> 113
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 29

Asp Ile Val Met Thr Gln Ser Pro Asp Ser Leu Ala Val Ser Leu Gly
1 5 10 15

Glu Arg Ala Thr Ile Asn Cys Lys Ser Ser Gln Ser Leu Phe Asn Ser
20 25 30

Asn Ala Lys Thr Asn Tyr Leu Asn Trp Tyr Gln Gln Lys Pro Gly Gln
35 40 45

Pro Pro Lys Leu Leu Ile Tyr Tyr Ala Ser Thr Arg His Thr Gly Val
50 55 60

Pro Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr
65 70 75 80

Ile Ser Ser Leu Gln Ala Glu Asp Val Ala Val Tyr Tyr Cys Gln Gln
85 90 95

Trp Tyr Asp Tyr Pro Tyr Thr Phe Gly Ala Gly Thr Lys Val Glu Ile
100 105 110

Lys

<210> 30
<211> 330
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 30

Ala Lys Thr Thr Ala Pro Ser Val Tyr Pro Leu Ala Pro Val Cys Gly
1 5 10 15

Asp Thr Thr Gly Ser Ser Val Thr Leu Gly Cys Leu Val Lys Gly Tyr
20 25 30

Phe Pro Glu Pro Val Thr Leu Thr Trp Asn Ser Gly Ser Leu Ser Ser
35 40 45

Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Asp Leu Tyr Thr Leu

50 55 60

Ser Ser Ser Val Thr Val Thr Ser Ser Thr Trp Pro Ser Gln Ser Ile
65 70 75 80

Thr Cys Asn Val Ala His Pro Ala Ser Ser Thr Lys Val Asp Lys Lys
85 90 95

Ile Glu Pro Arg Gly Pro Thr Ile Lys Pro Cys Pro Pro Cys Lys Cys
100 105 110

Pro Ala Pro Asn Leu Leu Gly Gly Pro Asp Val Phe Ile Phe Pro Pro
115 120 125

Lys Ile Lys Asp Val Leu Met Ile Ser Leu Ser Pro Ile Val Thr Cys
130 135 140

Val Val Val Asp Val Ser Glu Asp Asp Pro Asp Val Gln Ile Ser Trp
145 150 155 160

Phe Val Asn Asn Val Glu Val His Thr Ala Gln Thr Gln Thr His Arg
165 170 175

Glu Asp Tyr Asn Ser Thr Leu Arg Val Val Ser Ala Leu Pro Ile Gln
180 185 190

His Gln Asp Trp Met Ser Gly Lys Glu Phe Lys Cys Lys Val Asn Asn

195 200 205

Lys Asp Leu Pro Ala Pro Ile Glu Arg Thr Ile Ser Lys Pro Lys Gly
210 215 220

Ser Val Arg Ala Pro Gln Val Tyr Val Leu Pro Pro Pro Glu Glu Glu
225 230 235 240

Met Thr Lys Lys Gln Val Thr Leu Thr Cys Met Val Thr Asp Phe Met
245 250 255

Pro Glu Asp Ile Tyr Val Glu Trp Thr Asn Asn Gly Lys Thr Glu Leu
260 265 270

Asn Tyr Lys Asn Thr Glu Pro Val Leu Asp Ser Asp Gly Ser Tyr Phe
275 280 285

Met Tyr Ser Lys Leu Arg Val Glu Lys Lys Asn Trp Val Glu Arg Asn
290 295 300

Ser Tyr Ser Cys Ser Val Val His Glu Gly Leu His Asn His His Thr
305 310 315 320

Thr Lys Ser Phe Ser Arg Thr Pro Gly Lys
325 330

<210> 31

<211> 107
<212> PRT
<213> Mus musculus

<400> 31

Arg Ala Asp Ala Ala Pro Thr Val Ser Ile Phe Pro Pro Ser Ser Glu
1 5 10 15

Gln Leu Thr Ser Gly Gly Ala Ser Val Val Cys Phe Leu Asn Asn Phe
20 25 30

Tyr Pro Lys Asp Ile Asn Val Lys Trp Lys Ile Asp Gly Ser Glu Arg
35 40 45

Gln Asn Gly Val Leu Asn Ser Trp Thr Asp Gln Asp Ser Lys Asp Ser
50 55 60

Thr Tyr Ser Met Ser Ser Thr Leu Thr Leu Thr Lys Asp Glu Tyr Glu
65 70 75 80

Arg His Asn Ser Tyr Thr Cys Glu Ala Thr His Lys Thr Ser Thr Ser
85 90 95

Pro Ile Val Lys Ser Phe Asn Arg Asn Glu Cys
100 105

<210> 32
<211> 330

<212> PRT
<213> Mus musculus

<400> 32

Ala Lys Thr Thr Ala Pro Ser Val Tyr Pro Leu Ala Pro Val Cys Gly
1 5 10 15

Asp Thr Thr Gly Ser Ser Val Thr Leu Gly Cys Leu Val Lys Gly Tyr
20 25 30

Phe Pro Glu Pro Val Thr Leu Thr Trp Asn Ser Gly Ser Leu Ser Ser
35 40 45

Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Asp Leu Tyr Thr Leu
50 55 60

Ser Ser Ser Val Thr Val Thr Ser Ser Thr Trp Pro Ser Gln Ser Ile
65 70 75 80

Thr Cys Asn Val Ala His Pro Ala Ser Ser Thr Lys Val Asp Lys Lys
85 90 95

Ile Glu Pro Arg Gly Pro Thr Ile Lys Pro Cys Pro Pro Cys Lys Cys
100 105 110

Pro Ala Pro Asn Leu Leu Gly Gly Pro Ser Val Phe Ile Phe Pro Pro
115 120 125

Lys Ile Lys Asp Val Leu Met Ile Ser Leu Ser Pro Ile Val Thr Cys
130 135 140

Val Val Val Asp Val Ser Glu Asp Asp Pro Asp Val Gln Ile Ser Trp
145 150 155 160

Phe Val Asn Asn Val Glu Val His Thr Ala Gln Thr Gln Thr His Arg
165 170 175

Glu Asp Tyr Asn Ser Thr Leu Arg Val Val Ser Ala Leu Pro Ile Gln
180 185 190

His Gln Asp Trp Met Ser Gly Lys Glu Phe Lys Cys Lys Val Asn Asn
195 200 205

Lys Asp Leu Pro Ala Pro Ile Glu Arg Thr Ile Ser Lys Pro Lys Gly
210 215 220

Ser Val Arg Ala Pro Gln Val Tyr Val Leu Pro Pro Pro Glu Glu Glu
225 230 235 240

Met Thr Lys Lys Gln Val Thr Leu Thr Cys Met Val Thr Asp Phe Met
245 250 255

Pro Glu Asp Ile Tyr Val Glu Trp Thr Asn Asn Gly Lys Thr Glu Leu
260 265 270

Asn Tyr Lys Asn Thr Glu Pro Val Leu Asp Ser Asp Gly Ser Tyr Phe
275 280 285

Met Tyr Ser Lys Leu Arg Val Glu Lys Lys Asn Trp Val Glu Arg Asn
290 295 300

Ser Tyr Ser Cys Ser Val Val His Glu Gly Leu His Asn His His Thr
305 310 315 320

Thr Lys Ser Phe Ser Arg Thr Pro Gly Lys
325 330

<210> 33
<211> 328
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 33

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys
1 5 10 15

Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr
20 25 30

Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser

35 40 45

Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser
50 55 60

Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr
65 70 75 80

Tyr Ile Cys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys
85 90 95

Lys Val Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys
100 105 110

Pro Ala Pro Glu Leu Arg Gly Gly Pro Lys Val Phe Leu Phe Pro Pro
115 120 125

Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys
130 135 140

Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp
145 150 155 160

Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu
165 170 175

Glu Gln Tyr Ala Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu

180 185 190

His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn
195 200 205

Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly
210 215 220

Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu
225 230 235 240

Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr
245 250 255

Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn
260 265 270

Asn Tyr Lys Thr Thr Pro Pro Tyr Leu Asp Ser Asp Gly Ser Phe Phe
275 280 285

Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn
290 295 300

Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr
305 310 315 320

Gln Glu Ser Leu Ser Leu Ser Pro

325

<210> 34
<211> 107
<212> PRT
<213> Homo sapiens

<400> 34

Arg Thr Val Ala Ala Pro Ser Val Phe Ile Phe Pro Pro Ser Asp Glu
1 5 10 15

Gln Leu Lys Ser Gly Thr Ala Ser Val Val Cys Leu Leu Asn Asn Phe
20 25 30

Tyr Pro Arg Glu Ala Lys Val Gln Trp Lys Val Asp Asn Ala Leu Gln
35 40 45

Ser Gly Asn Ser Gln Glu Ser Val Thr Glu Gln Asp Ser Lys Asp Ser
50 55 60

Thr Tyr Ser Leu Ser Ser Thr Leu Thr Leu Ser Lys Ala Asp Tyr Glu
65 70 75 80

Lys His Lys Val Tyr Ala Cys Glu Val Thr His Gln Gly Leu Ser Ser
85 90 95

Pro Val Thr Lys Ser Phe Asn Arg Gly Glu Cys
100 105

<210> 35
<211> 116
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 35

Glu Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Lys
1 5 10 15

Ser Leu Lys Leu Ser Cys Glu Ala Ser Gly Phe Thr Phe Ser Gly Tyr
20 25 30

Gly Met His Trp Val Arg Gln Ala Pro Gly Arg Gly Leu Glu Ser Val
35 40 45

Ala Tyr Ile Thr Ser Ser Ser Ile Asn Ile Lys Tyr Ala Asp Ala Val
50 55 60

Lys Gly Arg Phe Thr Val Ser Arg Asp Asn Ala Lys Asn Leu Leu Phe
65 70 75 80

Leu Gln Met Asn Ile Leu Lys Ser Glu Asp Thr Ala Met Tyr Tyr Cys
85 90 95

Ala Arg Phe Asp Trp Asp Lys Asn Tyr Trp Gly Gln Gly Thr Met Val
100 105 110

Thr Val Ser Ser
115

<210> 36
<211> 107
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 36

Asp Ile Gln Met Thr Gln Ser Pro Ser Ser Leu Pro Ala Ser Leu Gly
1 5 10 15

Asp Arg Val Thr Ile Asn Cys Gln Ala Ser Gln Asp Ile Ser Asn Tyr
20 25 30

Leu Asn Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Leu Ile
35 40 45

Tyr Tyr Thr Asn Lys Leu Ala Asp Gly Val Pro Ser Arg Phe Ser Gly
50 55 60

Ser Gly Ser Gly Arg Asp Ser Ser Phe Thr Ile Ser Ser Leu Glu Ser
65 70 75 80

Glu Asp Ile Gly Ser Tyr Tyr Cys Gln Gln Tyr Tyr Asn Tyr Pro Trp
85 90 95

Thr Phe Gly Pro Gly Thr Lys Leu Glu Ile Lys
100 105

<210> 37
<211> 328
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 37

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys
1 5 10 15

Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr
20 25 30

Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser
35 40 45

Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser
50 55 60

Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr
65 70 75 80

Tyr Ile Cys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys
85 90 95

Lys Val Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys
100 105 110

Pro Ala Pro Glu Leu Arg Gly Gly Pro Lys Val Phe Leu Phe Pro Pro
115 120 125

Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys
130 135 140

Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp
145 150 155 160

Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu
165 170 175

Glu Gln Tyr Ala Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu
180 185 190

His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn
195 200 205

Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly
210 215 220

Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Lys Glu
225 230 235 240

Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr
245 250 255

Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn
260 265 270

Asn Tyr Lys Thr Thr Pro Pro Tyr Leu Asp Ser Asp Gly Ser Phe Phe
275 280 285

Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn
290 295 300

Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr
305 310 315 320

Gln Lys Ser Leu Ser Leu Ser Pro
325

<210> 38
<211> 118
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 38

Gln Val Gln Leu Val Glu Ser Gly Gly Gly Val Val Gln Pro Gly Arg
1 5 10 15

Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Ser Tyr
20 25 30

Thr Met His Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val
35 40 45

Thr Phe Ile Ser Tyr Asp Gly Asn Asn Lys Tyr Tyr Ala Asp Ser Val
50 55 60

Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr
65 70 75 80

Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Ile Tyr Tyr Cys
85 90 95

Ala Arg Thr Gly Trp Leu Gly Pro Phe Asp Tyr Trp Gly Gln Gly Thr
100 105 110

Leu Val Thr Val Ser Ser
115

<210> 39
<211> 110
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 39

Glu Ile Val Leu Thr Gln Ser Pro Gly Thr Leu Ser Leu Ser Pro Gly
1 5 10 15

Glu Arg Ala Thr Leu Ser Cys Arg Ala Ser Gln Ser Val Gly Ser Ser
20 25 30

Tyr Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln Ala Pro Arg Leu Leu
35 40 45

Ile Tyr Gly Ala Phe Ser Arg Ala Thr Gly Ile Pro Asp Arg Phe Ser
50 55 60

Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Arg Leu Glu
65 70 75 80

Pro Glu Asp Phe Ala Val Tyr Tyr Cys Gln Gln Tyr Gly Ser Ser Pro
85 90 95

Trp Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Ser Ser
100 105 110

<210> 40
<211> 122
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 40

Gln Val Gln Leu Val Glu Ser Gly Gly Gly Val Val Gln Pro Gly Gly
1 5 10 15

Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Asn Ala
20 25 30

Trp Met His Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val
35 40 45

Ala Gln Ile Lys Asp Lys Ser Gln Asn Tyr Ala Thr Tyr Val Ala Glu
50 55 60

Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Ala Asp Ser Lys Asn Ser
65 70 75 80

Ile Tyr Leu Gln Met Asn Ser Leu Lys Thr Glu Asp Thr Ala Val Tyr
85 90 95

Tyr Cys Arg Tyr Val His Tyr Ala Ala Gly Tyr Gly Val Asp Ile Trp
100 105 110

Gly Gln Gly Thr Thr Val Thr Val Ser Ser
115 120

<210> 41
<211> 112
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 41

Asp Ile Val Met Thr Gln Ser Pro Leu Ser Leu Pro Val Thr Pro Gly
1 5 10 15

Glu Pro Ala Ser Ile Ser Cys Arg Ser Ser Gln Pro Leu Val His Ser
20 25 30

Asn Arg Asn Thr Tyr Leu His Trp Tyr Gln Gln Lys Pro Gly Gln Ala
35 40 45

Pro Arg Leu Leu Ile Tyr Lys Val Ser Asn Arg Phe Ser Gly Val Pro
50 55 60

Asp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile
65 70 75 80

Ser Arg Val Glu Ala Glu Asp Val Gly Val Tyr Tyr Cys Gly Gln Gly
85 90 95

Thr Gln Val Pro Tyr Thr Phe Gly Gln Gly Thr Lys Leu Glu Ile Lys
100 105 110

<210> 42
<211> 328
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 42

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys
1 5 10 15

Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr
20 25 30

Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser
35 40 45

Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser
50 55 60

Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr
65 70 75 80

Tyr Ile Cys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys
85 90 95

Lys Val Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys
100 105 110

Pro Ala Pro Glu Leu Arg Gly Gly Pro Lys Val Phe Leu Phe Pro Pro
115 120 125

Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys
130 135 140

Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp
145 150 155 160

Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu
165 170 175

Glu Gln Tyr Ala Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu
180 185 190

His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn
195 200 205

Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly
210 215 220

Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Lys Glu
225 230 235 240

Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr
245 250 255

Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn
260 265 270

Asn Tyr Lys Thr Thr Pro Pro Tyr Leu Asp Ser Asp Gly Ser Phe Phe
275 280 285

Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn
290 295 300

Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn Arg Tyr Thr
305 310 315 320

Gln Lys Ser Leu Ser Leu Ser Pro
325

<210> 43
<211> 116

<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 43

Gln Val Gln Leu Gln Gln Ser Gly Pro Gln Leu Val Arg Pro Gly Ala
1 5 10 15

Ser Val Lys Ile Ser Cys Lys Ala Ser Gly Tyr Ser Phe Thr Ser Tyr
20 25 30

Trp Met His Trp Val Asn Gln Arg Pro Gly Gln Gly Leu Glu Trp Ile
35 40 45

Gly Met Ile Asp Pro Ser Tyr Ser Glu Thr Arg Leu Asn Gln Lys Phe
50 55 60

Lys Asp Lys Ala Thr Leu Thr Val Asp Lys Ser Ser Ser Thr Ala Tyr
65 70 75 80

Met Gln Leu Ser Ser Pro Thr Ser Glu Asp Ser Ala Val Tyr Tyr Cys
85 90 95

Ala Leu Tyr Gly Asn Tyr Phe Asp Tyr Trp Gly Gln Gly Thr Thr Leu
100 105 110

Thr Val Ser Ser
115

<210> 44
<211> 107
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 44

Asp Ile Gln Met Thr Gln Ser Ser Ser Ser Phe Ser Val Ser Leu Gly
1 5 10 15

Asp Arg Val Thr Ile Thr Cys Lys Ala Ser Glu Asp Ile Tyr Asn Arg
20 25 30

Leu Ala Trp Tyr Gln Gln Lys Pro Gly Asn Ala Pro Arg Leu Leu Ile
35 40 45

Ser Gly Ala Thr Ser Leu Glu Thr Gly Val Pro Ser Arg Phe Ser Gly
50 55 60

Ser Gly Ser Gly Lys Asp Tyr Thr Leu Ser Ile Thr Ser Leu Gln Thr
65 70 75 80

Glu Asp Val Ala Thr Tyr Tyr Cys Gln Gln Tyr Trp Ser Thr Pro Tyr
85 90 95

Thr Phe Gly Gly Gly Thr Lys Leu Glu Val Lys
100 105

<210> 45
<211> 328
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 45

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys
1 5 10 15

Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr
20 25 30

Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser
35 40 45

Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser
50 55 60

Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr
65 70 75 80

Tyr Ile Cys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys
85 90 95

Lys Val Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys
100 105 110

Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro
115 120 125

Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys
130 135 140

Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp
145 150 155 160

Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu
165 170 175

Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu
180 185 190

His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn
195 200 205

Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly
210 215 220

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Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu
225                 230                 235                 240

Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr
                245                 250                 255

Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn
            260                 265                 270

Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe
        275                 280                 285

Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn
    290                 295                 300

Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr
305                 310                 315                 320

Gln Lys Ser Leu Ser Leu Ser Pro
                325
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<210> 46
<211> 120
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 46

Gln Val Gln Leu Gln Gln Trp Gly Ala Gly Leu Leu Lys Pro Ser Glu
1 5 10 15

Thr Leu Ser Leu Thr Cys Ala Val Tyr Gly Gly Ser Phe Ser Asp Tyr
20 25 30

Tyr Trp Asn Trp Ile Arg Gln Pro Pro Gly Lys Gly Leu Glu Trp Ile
35 40 45

Gly Glu Ile Asn His Asn Gly Asn Thr Asn Ser Asn Pro Ser Leu Lys
50 55 60

Ser Arg Val Thr Leu Ser Leu Asp Thr Ser Lys Asn Gln Phe Ser Leu
65 70 75 80

Lys Leu Arg Ser Val Thr Ala Ala Asp Thr Ala Val Tyr Tyr Cys Ala
85 90 95

Phe Gly Tyr Ser Asp Tyr Glu Tyr Asn Trp Phe Asp Pro Trp Gly Gln
100 105 110

Gly Thr Leu Val Thr Val Ser Ser
115 120

<210> 47
<211> 107

<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 47

Glu Ile Val Leu Thr Gln Ser Pro Ala Thr Leu Ser Leu Ser Pro Gly
1 5 10 15

Glu Arg Ala Thr Leu Ser Cys Arg Ala Ser Gln Ser Ile Ser Ser Tyr
20 25 30

Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln Ala Pro Arg Leu Leu Ile
35 40 45

Tyr Asp Ala Ser Asn Arg Ala Thr Gly Ile Pro Ala Arg Phe Ser Gly
50 55 60

Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Glu Pro
65 70 75 80

Glu Asp Phe Ala Val Tyr Tyr Cys Gln Gln Arg Ser Asn Trp Pro Leu
85 90 95

Thr Phe Gly Gln Gly Thr Asn Leu Glu Ile Lys
100 105

<210> 48
<211> 121
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 48

Glu Val Gln Leu Gln Gln Ser Gly Pro Glu Leu Val Lys Pro Gly Ala
1 5 10 15

Ser Val Lys Ile Ser Cys Lys Thr Ser Gly Tyr Thr Phe Lys Asp Tyr
20 25 30

Thr Met His Trp Val Lys Gln Ser His Gly Lys Ser Leu Glu Trp Ile
35 40 45

Gly Gly Ile Tyr Pro Asn Asn Gly Gly Ser Thr Tyr Asn Gln Asn Phe
50 55 60

Lys Asp Lys Ala Thr Leu Thr Val Asp Lys Ser Ser Ser Thr Ala Tyr
65 70 75 80

Met Glu Phe Arg Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Tyr Cys
85 90 95

Ala Arg Met Gly Tyr His Gly Pro His Leu Asp Phe Asp Val Trp Gly
100 105 110

Ala Gly Thr Thr Val Thr Val Ser Pro
115 120

<210> 49
<211> 107
<212> PRT
<213> Artificial

<220>
<223> an artificially synthesized sequence

<400> 49

Asp Ile Val Met Thr Gln Ser His Lys Phe Met Ser Thr Ser Leu Gly
1 5 10 15

Asp Arg Val Ser Ile Thr Cys Lys Ala Ser Gln Asp Val Gly Ala Ala
20 25 30

Val Ala Trp Tyr Gln Gln Lys Pro Gly Gln Ser Pro Lys Leu Leu Ile
35 40 45

Tyr Trp Ala Ser Thr Arg His Thr Gly Val Pro Asp Arg Phe Thr Gly
50 55 60

Gly Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Asn Val Gln Ser
65 70 75 80

Glu Asp Leu Thr Asp Tyr Phe Cys Gln Gln Tyr Ile Asn Tyr Pro Leu
85 90 95

Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys
100 105