

Co-regulatory Sequence Based on Tetracycline and Cumate

TECHNICAL FIELD

The present disclosure relates to a tetracycline-based and Cumate-dependent nucleic acid sequence for regulating the transcription of a nucleic acid fragment of interest. Specifically, the present disclosure relates to a nucleic acid sequence for regulating the transcription of a nucleic acid fragment of interest, wherein the nucleic acid sequence comprises at least two copies of TetO-operator sequences (Tet operator, TetO) binding to a reverse tetracycline controlled transactivator (rtTA), one copy of a minimal promoter sequence containing the TATA box sequence, and at least one copy of a CuO-operator sequence (Cumate operator, CuO) binding to the CymR transcription repressor. Furthermore, there is a spliceable intron sequence between the 3' end downstream of the aforementioned nucleic acid sequence and the 5' end upstream of the regulated nucleic acid fragment of interest, which can be used to further improve the transcription activity after induction without induction leakage. Still further, the present disclosure also relates to a vector, viruse, cell, transgenic plant or animal containing the above regulatory sequence.

BACKGROUND

The regulation of gene expression can be achieved by regulating several steps in gene expression, including transcription, RNA splicing, translation, and post-translational modification, wherein the regulation of specific gene(s) or nucleic acid fragment(s) of interest in the transcription step is the most extensive and effective way for gene expression regulation. An inducible expression system that controls the transcription and expression of specific gene(s) and nucleic acid fragment(s) in cells, plants or animals is turned on or off through exogenous factor(s), and it plays a key role in the fields of basic biological research, biopharmaceuticals, gene therapy, cell therapy, improvement of transgenic plants or animals. Generally, the design of an inducible expression system is composed of two parts: 1) a specific nucleic acid sequence upstream of the 5' end of the regulated nucleic acid fragment of interest, which can generally be combined with a transcription activator or repressor to regulate the transcription of the nucleic acid fragment of interest; an example of such a specific nucleic acid sequence (hereinafter, a response element) is a nucleic acid sequence containing an operator; 2) a single or multiple regulator(s), which can bind to or be separated from the above specific nucleic acid sequence

under the control of an external factor(s) and generally has a function of transcriptional activation or repression, and which is usually an expressed protein. The external factor(s) can be an environmental factor, such as temperature, light, etc.; or a certain component, such as metabolites, hormones, metal ions, or artificial compounds involved in life activities.

"Tetracycline inducible system (or Tet System)" is currently one of the most widely used inducible expression systems in scientific research and business (see, for example, Gossen, M. and H. Bujard (1992). "Tight control of gene expression in mammalian cells by tetracycline-responsive promoters." Proc Natl Acad Sci USA **89**(12): 5547-5551., US patent No. 5,888,981, 5,814,618, 6,004,941, 5,814,618, 7,541,446, WO96/01313 and WO00/75347 (incorporated herein by reference)). It is mainly composed of a tetracycline-dependent regulator and a response element containing a TetO operator, and regulates the transcription of a nucleic acid fragment of interest linked to the response element containing a TetO operator in a tetracycline-inducible way. The response element containing a TetO operator can bind to a transcription activator in the absence of tetracycline, and after binding, initiate the transcription of the downstream regulated nucleic acid fragment of interest; it can also be placed downstream of a constitutively active promoter and upstream of the regulated nucleic acid fragment of interest, and can bind to a tetracycline-dependent repressor in the absence of tetracycline to inhibit the transcription of the regulated nucleic acid fragment of interest at downstream. Traditionally, the tetracycline-dependent regulator binds to the TetO operator in the absence of tetracycline, and can be separated from the TetO operator in the presence of tetracycline. For example, an activator designed based on this principle can initiate the transcription of the regulated nucleic acid fragment of interest by removing tetracycline in the environment (US Patent Nos. 5,464,758, 6,914,124, 5,789,156, 6,271,348, WO96/01313 and WO00/75347). Later, a trans-tetracycline-dependent regulator was developed, which only binds to the TetO operator in the presence of tetracycline. The trans-tetracycline-dependent activator can activate the transcription of the regulated nucleic acid fragment of interest by adding tetracycline to the environment (US Patent Nos. 5,654,168, 6,136,954, 5,789,156, 6,271,348, 6,087,166, 6,271,341, 7,541,446, WO96/01313 and WO00/75347); for example, the transcription of the regulated nucleic acid fragment of interest can be activated by adding tetracycline to the medium, or the transcription and expression of the regulated gene of interest introduced by genetic engineering can be activated by injecting tetracycline into the transgenic animal.

The transcription activity of the regulated nucleic acid fragment under induction conditions and the leaky transcription activity of the regulated nucleic acid fragment under non-induction

conditions are two important indicators for evaluating an inducible expression system. A good inducible expression system needs to have a higher induced transcription activity and a lower leaky transcription activity. The tetracycline inducible system has been optimized by many laboratories since the first report, and its performance has been continuously improved. The main optimizations include the optimization of the tetracycline-dependent regulator and the response element containing a TetO operator. The optimization of the tetracycline-dependent regulator mainly focuses on (1) reducing the leaky transcription activity under non-induction conditions while maintaining the induced transcription activity as much as possible, and increasing the ratio of induced/leaky transcription activity; (2) optimizing the specificity and affinity of the regulator and tetracycline and a derivative thereof as inducers, to achieve a maximum transcription activity at a lower concentration of tetracycline and a derivative thereof, and at the same time to reduce the affinity of the regulator to the response element in the absence of the inducer. (See Zhou, X., et al. (2006). "Optimization of the Tet-On system for regulated gene expression through viral evolution." Gene Ther **13**(19): 1382-1390., US Patent No. 8,383,364, 7,541,446). The optimization of the response element containing a TetO operator mainly focuses on (1) optimizing the copy number and linkage sequence of the TetO operator; enhancing the stability of multiple copies of a continuous TetO operator sequence, especially in viral vectors, while improving the performance of the response element; (2) under the condition that does not significantly affect the transcription activity after induction, reducing the non-induction leaky transcription activity of the minimal promoter and increasing the ratio of induced/leaky transcription activity (see Loew, R., et al. (2010). "Improved Tet-responsive promoters with minimized background expression." BMC Biotechnol **10**: 81., US Patent No. 9,181,556). The commercial tetracycline inducible systems currently widely used are mainly those from Clontech Laboratories, Inc., such as Tet-On® Advanced Inducible Gene Expression Systems (document number: PT3898-1 (102312), wherein the tetracycline-dependent transactivator rtTA and the TetO Operator-containing response element TRE (a nucleic acid sequence containing multiple copies of continuous TetO operator sequences and a minimal promoter sequence, Tet Response Element, hereinafter referred to as TRE) are hereinafter referred to as rtTA_{adv} and TRE_{adv}), Tet-Off® Advanced Inducible Gene Expression Systems (document number: PT3945-1 (101612)) and Tet-On® 3G Inducible Expression Systems (document number: PT5148-1 (010814), wherein the tetracycline-dependent regulator rtTA and the TetO operator-containing response element TRE are hereinafter referred to as rtTA_{3G} and TRE_{3G} respectively).

Cumate Inducible System is developed from the p-cym operon of *Pseudomonas putida*, and

is composed of a CuO operator downstream of the TATA box of the constitutively active promoter and a repressor CymR protein which is Cumate/a derivative thereof-dependent and can bind to the CuO operator. In the absence of Cumate, the CymR protein binds to the CuO operator to inhibit the transcription of the downstream regulated nucleic acid fragment of interest; when the CymR protein binds to Cumate, the affinity of CymR protein with the CuO operator decreases and thus they are separated from each other, and the transcription of the downstream regulated nucleic acid fragment of interest is no longer inhibited. For example, the CuO operator is placed between the regulated nucleic acid fragment of interest and the constitutively active promoter sequence, and the CymR repressor is constitutively and actively expressed in the cell. When there is no inducer (Cumate or a derivative thereof) in the medium, the binding of the expressed CymR protein to the CuO operator inhibits the transcription of the regulated nucleic acid fragment of interest; when the inducer is added to the medium, the CymR protein that binds to Cumate or its derivative is separated from the CuO operator, and the transcription of the regulated nucleic acid fragment of interest is no longer inhibited (see WO02088346A2, WO2006037215A1 (incorporated herein by reference)).

The optimization of Cumate inducible system mainly focuses on: (1) optimizing the shortest effective CuO operator sequence; (2) screening for inducers of Cumate and a derivative thereof that are soluble in water and have similar functions to p-cumate; (3) constructing an inducible system similar to Tet-Off, which consists of a CymR activator (such as constructing a fusion protein of CymR and HSV VP-16 transcription activation-domain) and a response element containing multiple copies of continuous CuO operators (the sequence contains multiple repeats of the CuO sequence and a downstream minimal promoter). In the absence of Cumate inducer, the binding of the CymR activator with the CuO operator activates the transcription of the regulated nucleic acid fragment of interest; in the presence of Cumate inducer, the CymR activator and the CuO operator are separated from each other, and the transcription of the regulated nucleic acid fragment of interest stops. (4) constructing an inducible system similar to Tet-On, which consists of a trans-CymR activator and a response element containing multiple copies of continuous CuO operators (which is obtained by mutation and screening, and allowing the trans-CymR to bind to the CuO operator in the presence of a Cumate inducer and to be separated from the CuO operator in the absence of a Cumate inducer, and constructing a fusion protein of trans-CymR and HSV VP-16 transcription activation-domain as the trans-CymR activator). In the presence of a Cumate inducer, the trans CymR activator binds to the response element to activate the transcription of the regulated nucleic acid fragment of interest; in the

absence of Cumate inducer, the trans CymR activator is separated from the response element, and the transcription of the regulated nucleic acid fragment of interest stops (see Mullick, A., et al. (2006). "The cumate gene-switch: a system for regulated expression in mammalian cells." BMC Biotechnol 6: 43.; WO02088346A2, WO2006037215A1). The currently widely used Cumate inducible system is mainly the SparQ Cumate Switch system from SBI System Biosciences (document number: 1-090810).

In the current development and improvement of various inducible expression systems, the main optimizations are focused on reducing the basic leaky transcription activity of the regulated nucleic acid fragment of interest under non-induction conditions, and the transcription activity under induction conditions is not the focus of optimization. As a result, the transcription activity of the nucleic acid fragment of interest regulated by the inducible expression system is lower than the transcription activity under the control of a common constitutively active promoter. In the development of biotechnology, it is equally important to improve the absolute transcription activity of the nucleic acid fragment of interest after induction and to control its leaky transcription activity under non-induction conditions. For example, in the field of biopharmaceuticals, the production of a cytotoxic recombinant protein or virus vector requires not only to increase the expression level of the protein encoded by the nucleic acid fragment of interest after induction, but also to control the leaked expression level of the nucleic acid fragment of interest under non induction conditions in order to ensure the culture state and stability of the producer cell line. Therefore, an inducible expression system that can not only increase the transcription activity of the nucleic acid fragment of interest under induction conditions, but also control the leaky transcription activity of the nucleic acid fragment of interest under non-induction conditions can widely promote the research, development and industrialization of biotechnology in many fields.

SUMMARY

The present disclosure relates to a tetracycline and Cumate-dependent nucleic acid sequence capable of regulating the transcription of a nucleic acid fragment of interest. Specifically, the nucleic acid sequence comprises at least two copies of TetO-operator sequences (Tet operator, TetO) binding to a reverse tetracycline/derivatives thereof controlled transactivator (rtTA), one copy of a minimal promoter sequence containing a TATA box sequence, and at least one copy of CuO-operator sequence (Cumate operator, CuO) binding to a CymR transcription repressor. Furthermore, a spliceable intron sequence between the 3' end downstream of the aforementioned

nucleic acid sequence and the 5' end upstream of the regulated nucleic acid fragment of interest can be used to further improve the transcription activity after induction without induction leakage. Still further, the present disclosure also relates to a vector, virus, cell, transgenic plant or animal containing the above regulatory sequences for regulating gene expression.

The term "nucleic acid fragment of interest" may generally refer to a deoxyribonucleic acid fragment (DNA fragment), and according to the application purpose, may refer to a gene, such as a nucleic acid sequence encoding a protein; it can be a fragment of ribonucleic acid (RNA), such as a RNA fragment of the whole or part genome of an RNA virus, microRNA (miRNA), small interfering RNA (siRNA), long non-coding RNAs (LncRNA), guide RNA for CRISPR gene editing system (gRNA), transfer RNA (tRNA), ribosomal RNA (rRNA), an aptamer, a ribonucleic acid fragment that can bind to another nucleic acid, a ribonucleic acid fragment that can bind to a specific protein, or any ribonucleic acid fragment; it can also be a combination of the above one or more nucleic acid sequence(s).

The term "vector" is a nucleic acid-containing molecule that is usually used as a vehicle for artificially transporting a foreign genetic material, such as the above-mentioned nucleic acid fragment of interest, into another cell, where it is replicated and/or expressed. Functionally, all vectors can be used to clone and carry a foreign nucleic acid fragment of interest, and there are expression vectors specifically designed for transcription of a nucleic acid fragment and expression of a protein. Plasmids, viral vectors, cosmids and artificial chromosomes are the four main types of vectors. Among these, the most commonly used vector is plasmids. All engineered plasmid vectors contain an origin of replication for replication in bacteria, a multiple cloning site for insertion of a nucleic acid fragment of interest, and a marker gene for selection of positive strains. The viral vector is another commonly used vector, and is commonly used to deliver a genetic material, such as a nucleic acid fragment of interest, into cells. This process can be carried out in vivo or in cell culture (in vitro). The nucleic acid fragment of interest can be efficiently introduced into the infected target cell based on various molecular mechanisms evolved by the virus itself, such as protection of the genetic material, selection of host cells based on the recipient, delivery of genetic material into the host cell, replication and/or expression in the host cell, modulation of host cell growth, metabolism, reproduction, replication and defense mechanisms, as well as suppression and/or escape of the immune system in higher animals. In addition to being used in molecular biology research, viral vectors are also commonly used in gene therapy, cell therapy, immunotherapy, and vaccine development.

Generally, an inducible expression system will also increase the leaky transcription activity

under non-induction conditions when enhancing the induced transcription activity of the nucleic acid fragment of interest; and the commonly used strategies to control the leaky transcription activity under non-induction conditions, such as reducing the affinity of the transactivator with the operator in the absence of an inducer by point mutation or reducing the basic activity of the minimal promoter downstream of the operator, will affect the maximum transcription activity after induction. This makes most of the single inducible expression systems with the purpose of balancing the leaky transcription activity under non-induction conditions have reduced transcription activation activity for the nucleic acid fragment of interest under induction conditions. Designing a complex inducible expression system containing multiple regulatory components is an effective direction to solve the above problem. In the complex inducible expression system realized by the present disclosure, on the one hand, the maximum transcription activity after induction of the combination of the activator and the corresponding response element in a first inducible expression system is optimized, and on the other hand, the leaky transcription activity of the complex inducible expression system is controlled by using the combination of a repressor and the corresponding response element in a second inducible expression system; for example, the Tet-On inducible expression system is used as the first inducible expression system, and the Cumate inducible expression system is used as the second inducible expression system. The complex response element sequence of the above-mentioned complex inducible expression system contains at least two copies of TetO operator sequences (Tet operator) binding to a reverse tetracycline/derivative thereof controlled transactivator (rtTA), one copy of a minimal promoter sequence containing a TATA box sequence, and at least one copy of a CuO operator sequence (CuO operator) binding to a CymR transcription repressor.

The inventors have found that the induced transcription activity and the non-induced leaky transcription activity of the Tet-On and Cumate complex inducible expression system are affected by the following factors: (1) multiple copies of TetO operators and its linkage nucleic acid sequence; (2) the minimal promoter sequence in the TRE response element; (3) the relative position of the CuO operator and the TATA box in the minimal promoter. The control of the TRE_{3G} response element in the third-generation Tet-On system is more stringent than that of the TRE_{adv} response element in the second-generation Tet-On system, and has a lower non-induced leaky transcription activity; the closer the distance between the CuO operator and the TATA box, the stricter the control of leaky transcription activity, but the effect on the induced transcription activity becomes greater; the farther the distance between the CuO operator and the TATA box, the weaker the control of leaky transcription activity, but the effect on the induced transcription

activity becomes smaller. Based on the above findings, the nucleic acid sequence of the present disclosure for regulating the transcription of the nucleic acid fragment of interest contains: at least 2 copies (for example, 2, 3, 4, 5, 6, 7 or more, preferably 7) of a TetO-operator sequence binding to the reverse tetracycline/a derivative thereof-controlled transactivator rtTA, preferably, the TetO-operator sequence is shown in SEQ ID NO: 24; a minimal promoter sequence containing a TATA box sequence, preferably the minimal promoter sequence is shown in SEQ ID NO: 25 or SEQ ID NO: 26, more preferably the minimal promoter sequence is shown in SEQ ID NO: 25; and a CuO-operator sequence binding to a transcription repressor CymR regulated by cumate, preferably the CuO operator sequence is shown in SEQ ID NO:27. Preferably, at least one CuO operator sequence is located downstream of the above-mentioned promoter TATA box, preferably 10bp to 100bp downstream of the TATA box, such as 14bp, 30bp, 50bp and 100bp; further preferably, there is only one CuO sequence located at 30bp to 50bp downstream of the TATA box; still further, there is only one CuO sequence located at 30bp or 50bp downstream of the TATA box. Preferably, the nucleic acid sequence of the present disclosure for regulating the transcription of the nucleic acid fragment of interest is set forth in SEQ ID NO: 23, SEQ ID NO: 28, SEQ ID NO: 29 or SEQ ID NO: 30, more preferably, set forth in SEQ ID NO:23 or SEQ ID NO:28.

Introns downstream of the promoter in the expression plasmid can generally increase the stability of messenger RNA (mRNA) and increase the efficiency of mRNA transport out of the nucleus (see Akef, A., et al. (2015). "Splicing promotes the nuclear export of beta-globin mRNA by overcoming nuclear retention elements." RNA **21**(11): 1908-1920) and increase the expression of a nucleic acid fragment of interest. The inventors have found that linking a spliceable intron downstream of the 3' end of the response element in a single inducible expression system can also increase the expression of the regulated nucleic acid fragment of interest after induction, but at the same time it will significantly increase the non-induced leaky expression of the nucleic acid fragment of interest. Linking a spliceable intron sequence between the downstream of the 3' end of the tetracycline and Cumate-dependent complex response element sequence and the upstream of the 5' end of the regulated nucleic acid fragment of interest can significantly increase the induced expression level without significantly increasing the non-induced leaky expression level. Based on this, in the present disclosure, preferably, a spliceable intron fragment is linked between the downstream of the 3' end of the tetracycline and Cumate-dependent complex response element sequence and the upstream of the 5' end of the nucleic acid fragment of interest. There is no specific limitation on the selection of the above-

mentioned intron, and those skilled in the art will understand that any sequence capable of performing RNA splicing in mammalian cells can achieve the above functions. Intron sequences that may be selected include, but are not limited to, introns on commonly used cloning vectors, such as a rabbit β -globulin intron, a hybrid intron derived from human β -globulin and immunoglobulin heavy chain intron, EF-1 α intron A, SV40 intron, a hybrid intron derived from adenovirus and immunoglobulin heavy chain intron, a modified human cytomegalovirus intron, a hybrid intron derived from chicken β -actin (CBA) and mouse microvirus (MMV) intron, a chimera derived from chicken β -actin and rabbit β -globulin intron, and a mP1 intron; or any intron of any gene of any eukaryote; or an artificial intron sequence designed based on the intron splicing rules. The intron sequence used in the examples of the present disclosure may be set forth in SEQ ID NO: 31.

In the above-mentioned Tet-On and Cumate complex inducible expression system, in the absence of tetracycline or a derivative thereof and a Cumate inducer (Cumate or a functional analogue), the rtTA transactivator cannot bind to the complex response element, and the binding of the CymR repressor and the CuO operator in the complex response element further inhibits the leaky transcription activity of the regulated nucleic acid fragment of interest caused by the non-specific binding of rtTA transactivator or the basic activity of minimal promoter; in the presence of Cumate inducer alone, neither the rtTA transactivator nor the CymR repressor binding to the Cumate inducer can bind to the complex response element, the non-specific binding of rtTA transactivator or the basic activity of minimal promoter results in little transcription of the regulated nucleic acid fragment of interest; in the presence of tetracycline or a derivative thereof alone, the binding of the rtTA transactivator and the TetO operator in the complex response element activates the transcription of the regulated nucleic acid fragment of interest, but the CymR repressor also binds to the CuO operator in the complex response element, which leads to a limited transcription of the regulated nucleic acid fragment of interest; in the presence of both tetracycline or a derivative thereof and a Cumate inducer, the rtTA transactivator binds to the TetO operator in the complex response element, and the CymR repressor binding to the Cumate inducer is separated from the CuO operator in complex response element, thereby the most active transcription of the regulated nucleic acid fragment of interest is obtained. The nucleic acid coding sequence of the rtTA transactivator is preferably human codon-optimized SEQ ID NO: 18; CymR is preferably human codon-optimized SEQ ID NO: 15.

The above-mentioned Tet-On and Cumate complex inducible expression system can be applied in basic biological scientific research, biopharmaceuticals, gene therapy, cell therapy,

improvement of transgenic plants or animals, including but not limited to controlling the turn-on or -off of transcription of protein-coding nucleic acid sequences in cells, animals or plants; controlling the turn-on and -off of transcription of non-protein-coding nucleic acid sequences, such as a RNA fragment of the whole or part genome of an RNA virus, microRNA (miRNA), small interfering RNA (siRNA), long non-coding RNAs (LncRNA), guide RNA for CRISPR gene editing system (gRNA), transfer RNA (tRNA), ribosomal RNA (rRNA), an aptamer, a ribonucleic acid fragment that can bind to another nucleic acid, a ribonucleic acid fragment that can bind to a specific protein, or any ribonucleic acid fragment; it can also be a combination of the above one or more nucleic acid sequence(s). In one aspect, the above-mentioned Tet-On and Cumate complex inducible expression system can be used to express and produce a virus vector in the field of gene therapy, cell therapy or immunotherapy; to express and produce a cytotoxic or unstable protein; to express and produce an attenuated vaccine or viral like partical (VLP) vaccine in the field of vaccines; to control the turn-on or turn-off of transcription and expression of a nucleic acid fragment of interest in transgenic animals or plants.

In the present disclosure, tetracycline and a derivative thereof for the Tet-On inducible expression system comprise compounds similar to tetracycline in structure, and it can bind to the tetracycline-dependent transactivator rtTA as described herein with an association constant K_a of at least 10^{-6} M; preferably, 10^{-9} M or more. The derivative of tetracycline may be selected from, for example, doxycycline (Dox), anhydrotetracycline (Atc), chlorotetracycline, oxytetracycline, and deoxytetracycline.

In the present disclosure, the Cumate functional analog that binds to the repressor CymR and can be used in the Cumate inducible expression system, can be selected from, for example, p-ethylbenzoic acid, p-propylbenzoic acid, p-isopropylbenzoic acid, p-isobutylbenzoic acid, p-tert-butylbenzoic acid, p-n-dimethylaminobenzoic acid, p-n-ethylaminobenzoic acid, and other Cumate functional analogs such as those described in U.S. patent No. 7,745,592.

The host cell into which the vector containing the co-regulatory sequence of tetracycline and Cumate of the present disclosure can be introduced can be, in principle, any cell, such as bacteria, fungi, animal cells or plant cells. On the other hand, the co-regulatory sequence of tetracycline and Cumate of the present disclosure can also be used in virus vectors. For example, the nucleic acid fragment contained in a prepared virus vector may contain the co-regulatory sequence of tetracycline and Cumate of the present disclosure, which may be present as a deoxyribonucleic acid in DNA virus vectors (such as adenovirus, adeno-associated virus), and may be present as a ribonucleic acid in RNA virus vectors (such as retrovirus).

As will be appreciated by those skilled in the art, there is no particular limitation on the nucleic acid fragment of interest whose transcription is controlled by the regulatory nucleic acid sequence of the present disclosure, as long as its transcription may be controlled by the regulatory nucleic acid sequence of the present disclosure.

In one aspect, the disclosure provides the following:

Item 1. A nucleic acid sequence, comprising at least 2 copies of TetO-operator sequences capable of binding to a transactivator rtTA regulatable by tetracycline or a derivative thereof, and 1 copy of a minimal promoter sequence containing a TATA box sequence, and at least 1 copy of a CuO-operator sequence capable of binding to a transcription repressor CymR regulatable by cumate, wherein the CuO-operator sequence is downstream of the 3' end of the TATA box sequence, and is 10 bp to 100 bp apart from the TATA box.

Item 2. The nucleic acid sequence of item 1, wherein the CuO-operator sequence is 30 bp to 50 bp apart from the TATA box.

Item 3. The nucleic acid sequence of item 1, wherein the CuO-operator sequence is 50 bp apart from the TATA box.

Item 4. The nucleic acid sequence of item 1, wherein the TetO-operator sequence is set forth in SEQ ID NO: 24, and/or the minimal promoter sequence is set forth in SEQ ID NO: 25 or SEQ ID NO: 26, and/or the CuO-operator sequence is set forth in SEQ ID NO:27.

Item 5. The nucleic acid sequence of item 4, wherein the nucleic acid sequence is set forth in SEQ ID NO:23, SEQ ID NO:28, SEQ ID NO:29 or SEQ ID NO:30.

Item 6. The nucleic acid sequence of any one of items 1 to 5, further comprising a spliceable intron sequence at the 3' end thereof.

Item 7. A vector comprising the nucleic acid sequence of any one of items 1 to 6.

Item 8. The vector of item 7, wherein the vector is an expression vector comprising a nucleic acid fragment of interest downstream of the 3' end of the nucleic acid sequence of any one of items 1 to 6, and the transcription of the nucleic acid fragment of interest is controlled by the nucleic acid sequence of any one of items 1 to 6.

Item 9. A host cell comprising the nucleic acid sequence of any one of items 1 to 6 or the vector of item 7 or 8.

Item 10. A method for inducing the expression of a nucleic acid fragment of interest in a host cell, comprising the following steps:

(1) introducing the vector of item 8 and a coding sequence of rtTA and a coding sequence of CymR into the host cell;

(2) expressing rtTA and CymR in the host cell subjected to step (1); and
(3) providing tetracycline or a derivative thereof and cumate or a functional analog thereof for the host cell subjected to step (2).

Item 11. The method of item 10, wherein the rtTA is rtTA_{adv} or rtTA_{3G}.

Item 12. The method of item 10, wherein the coding sequence of rtTA is set forth in SEQ ID NO: 18.

Item 13. The method of item 10, wherein the coding sequence of CymR is set forth in SEQ ID NO: 15.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the effect of the position and quantity of the CuO operator in the complex response element in Example 2 on the induced expression level and non-induced leaky expression level of the nucleic acid fragment of interest. Figure 1A shows respective RLU values of the Luciferase detection experiment corresponding to adding only DOX inducer and adding both DOX and Cuamte inducer; Figure 1B shows the ratio of the RLU value of adding Cumate and without adding Cuamte inducer.

Figure 2 shows the effects of the single regulatory response element, the complex regulatory response element, and the introns linked downstream of the response element in Example 3 on the induced expression level and the non-induced leaky expression level of the nucleic acid fragment of interest. Figure 1A shows the RLU value of Luciferase detection experiment with/without DOX and Cumate inducers; Figure 1B shows the ratio of RLU value with/without DOX and Cumate inducers.

Figure 3 shows the effects of different introns in Example 4 on the induced expression level and non-induced leaky expression level of TRE_{3G}CuO complex response element. Figure 1A shows the RLU value of Luciferase detection experiment with/without DOX and Cumate inducers; Figure 1B shows the ratio of RLU value with/without DOX and Cumate inducers.

Figure 4 shows the respective induced transcription activities of the nucleic acid fragment of interest regulated by TRE_{3G}CuO and TRE_{3G}CuO-BGI in Example 5 under different combinations of induction. The ordinate shows the RLU value of the Luciferase detection experiment with different inducers; the 4-color histogram shows the four combinations of inducers; the numbers on the horizontal line represent the ratio of RLU value under the induction condition and the non-induction condition (white).

DETAILED DESCRIPTION OF THE DISCLOSURE

The following examples are provided to illustrate the technical solutions of the present disclosure and should not be construed as limiting the scope and spirit of the present disclosure.

Example 1: methods for constructing plasmids

Molecular cloning techniques used in the following examples, such as PCR amplification of DNA fragments, restriction enzyme digestion of DNA fragments, gel recovery of DNA fragments, T4 DNA ligase ligation of two or more DNA fragments, transformation of ligation-competent cells, plasmid miniprep and identification, are all well known in the art. The following reagents are involved in the examples below: PCR enzyme (Thermo, F-530S); restriction enzyme (NEB); T4 DNA ligase (Invitrogen, 15224041); DNA fragment gel recovery kit (Omega, D2500-02); plasmid mini kit (TIANGEN, DP 105-03); competent cells (XL-10 Gold, Hu Nanfenghui Biotech Co., Ltd., JZ 011); the nucleic acid sequences set forth in SEQ ID NO: 1 to SEQ ID NO: 22 were synthesized by GenScript and used in the construction of the plasmids of the present disclosure, and the plasmid sequencing and identification were performed by Invitrogen. Table 1 shows the primers information for plasmid construction; Table 2 shows the element composition of the sequences SEQ ID NO: 1 to SEQ ID NO: 31; Table 3 is a description of the functional elements in the plasmids; Table 4 shows the numbers and corresponding names of the plasmids constructed according to the present disclosure. The sequence information of elements adopted by each plasmid involved in the following examples is an example to carry out the present disclosure, and those skilled in the art can expect that the effect of the present disclosure can be achieved by replacing the sequence of element in the plasmid used in the examples below with other sequences of element having similar biological functions, including but not limited to backbone sequences (such as replication origin, resistance genes, etc.), restriction site sequences, transposon repeat sequences, response element sequences of inducible system, insulator sequences, promoter sequences, intron sequences, polyadenylation signal (PolyA) sequences, different codon-optimized gene sequences, mutants of the above sequences of functional elements and gene sequences, and the cloning positions, cloning sequences and cloning directions of the sequences of functional elements and gene sequences. The specific methods for constructing plasmids are as follows:

1. Construction of plasmid 18BF007: the synthetic sequence SEQ ID NO: 2 (2900bp) was digested with NotI and AsiSI, and ligated to NotI and AsiSI restriction sites of plasmid 18BF003 (SEQ ID NO: 1), respectively, to construct plasmid 18BF007.

2. Construction of plasmid 18BF011: the 18BF007 plasmid was digested by MluI and

SphI; and a fragment of 1730bp was recovered by gel and ligated to MluI and SphI restriction sites of plasmid 18BF003, to construct a plasmid 18BF011.

3. Construction of plasmid 18BF210: the synthetic sequence SEQ ID NO: 3 (1208bp) was digested with SpeI and AgeI, and ligated to AvrII and AgeI restriction sites of plasmid 18BF011, respectively, to construct a plasmid 18BF210.

4. Construction of plasmids 18BF211, 18BF212, 18BF213, 18BF214, 18BF215, 18BF216, 18BF217 and 18BF218: the synthetic sequences SEQ ID NO: 4 (908bp), SEQ ID NO:5 (880bp), SEQ ID NO:6 (890bp) and SEQ ID NO: 7 (845bp) were digested with MluI and ClaI, and ligated to MluI and ClaI restriction sites of plasmid 18BF210, respectively, to construct plasmids 18BF212, 18BF211, 18BF214 and 18BF213, respectively. Plasmids 18BF211, 18BF212, 18BF213 and 18BF214 were digested with BstBI, and fragments of 3932bp (18BF211), 3960bp (18BF212), 3897bp (18BF213) and 3942bp (18BF214) were recovered by gel and ligated with T4 ligase to construct plasmids 18BF215, 18BF216, 18BF217 and 18BF218, respectively.

5. Construction of plasmids 18BF229, 18BF232, 18BF233, 18BF234, 18BF235, 18BF236, 18BF237, 18BF240 and 18BF241: the Luciferase gene fragment (1728bp) was PCR-amplified by using pGL3-Basic (Promega, E1751) as a template and using Luc-F (SEQ ID NO: 32) and Luc-R (SEQ ID NO: 33) as primers, then was digested with BamHI and XhoI, and ligated to BamHI and XhoI restriction sites of plasmids 18BF210, 18BF217, 18BF218, 18BF215, 18BF216, 18BF211, 18BF212, 18BF213 and 18BF214, respectively, to construct plasmids 18BF229, 18BF232, 18BF233, 18BF234, 18BF235, 18BF236, 18BF237, 18BF240 and 18BF241.

6. Construction of plasmids 18BF251, 18BF252, 18BF253, 18BF254, 18BF255, 18BF256 and 18BF257: the synthetic sequences SEQ ID NO:8 (414bp), SEQ ID NO:9 (414bp), SEQ ID NO:10 (415bp), SEQ ID NO:11 (472bp), SEQ ID NO:12 (588bp), SEQ ID NO:13 (704bp) and SEQ ID NO:22 (376bp) were digested with MluI and ClaI, and ligated to MluI and ClaI restriction sites of plasmid 18BF235, respectively, to construct plasmids 18BF251, 18BF252, 18BF253, 18BF254, 18BF255, 18BF256 and 18BF257, respectively.

7. Construction of plasmids 18BF261, 18BF262, 18BF263 and 18BF264: a BGI (C&R) intron sequence (1036bp) was PCR-amplified by using SEQ ID NO: 17 as a template and using BGI (C&R)-F (SEQ ID NO:34) and BGI (C&R)-R (SEQ ID NO:35) as primers; an Intron (EF-1a) intron sequence (962bp) was PCR-amplified by using pEF1alpha-IRES-AcGFP1 (Clontech) as a template and using Intron(EF-1a)-F(SEQ ID NO:36) and Intron(EF-1a)-R(SEQ ID NO: 37)

as primers; an Intron (pSI) intron sequence (152bp) was PCR-amplified by using pSI (Promega#E1721) plasmid as a template and using Intron (pSI)-F (SEQ ID NO: 38) and Intron (pSI)-R (SEQ ID NO: 39) as primers; and the above three PCR products were respectively digested with ClaI and BamHI, and then ligated to ClaI and BamHI restriction sites of plasmid 18BF235, thereby to construct plasmids 18BF261, 18BF263 and 18BF264. The synthetic SEQ ID NO: 14 (210bp) was digested with ClaI and BamHI, and ligated to ClaI and BamHI restriction sites of plasmid 18BF235, to construct a plasmid 18BF262.

8. Construction of plasmids 19BF075 and 19BF074: the synthetic sequences SEQ ID NO: 15 (633bp) and SEQ ID NO: 16: (1496bp) were digested with the ClaI and XhoI and the SpeI and AgeI, respectively, and ligated in sequence to the ClaI and XhoI restriction sites and the AvrII and AgeI restriction sites of the plasmid 18BF007 to construct a plasmid 19BF073. The synthetic sequence SEQ ID NO: 17 (1979bp) was digested by MluI and AgeI, and ligated to MluI and AgeI restriction sites of the 18BF007 plasmid to replace a CMV-BGI-MCS-pA sequence, and thereby constructing a plasmid 18BF008. The synthetic sequences SEQ ID NO: 18 (768bp) and SEQ ID NO: 19 (765bp) were respectively digested by ClaI and XhoI, and then respectively ligated to the ClaI and XhoI restriction sites of plasmid 18BF008, to construct plasmids 18BF085 and 18BF084 respectively. The plasmid 19BF073 was digested with SpeI and AgeI, and a fragment of 3821bp was recovered by gel and respectively ligated to AvrII and AgeI restriction sites of plasmids 18BF085 and 18BF084 to construct a plasmids 19BF075 and 19BF074 respectively.

9. Construction of plasmid 18BF019: the synthetic sequences SEQ ID NO: 21 (1044bp) and SEQ ID NO: 20 (1320bp) were respectively digested by the BamHI and XhoI and the XhoI and BglII, and ligated to BamHI and BglII restriction sites of plasmid 18BF011 to construct a plasmid 18BF019.

10. Construction of plasmids 19BF229, 19BF235 and 19BF237: plasmids 18BF229, 18BF235 and 18BF237 were digested with MluI and AgeI, and fragments of 4605bp (18BF229), 3847bp (18BF235) and 4341bp (18BF237) was recovered by gel and respectively ligated to MluI and AgeI restriction sites of plasmid 18BF007, to construct plasmids 19BF229, 19BF235 and 19BF237 respectively.

Table 1. Information for Primers

SEQ ID NO:	Primer Name	Primer Sequence (5'-3')
32	Luc-F	TCAGGATCCATCTGCGATCTAAGTAAGCTTG
33	Luc-R	TCAACTCGAGCTAGAATTACACGGCGATC
34	BGI(C&R)-F	GTCAATCGATGGAGTCGCTGCGCGCTG
35	BGI(C&R)-R	GTCGGATCCCTGTAGGAAAAAGAAGAAGG

36	Intron(EF-1a)-F	GTCAATCGATGTAAGTGCCGTGTGTG
37	Intron(EF-1a)-R	GTCGGATCCCTGAAATGGAAGAAAAAACT
38	Intron(pSI)-F	GTCAATCGATGTAAGTATCAAGGTTACAAG
39	Intron(pSI)-R	GTCGGATCCCTGTGGAGAGAAAGGC

Table 2. Description of the appendix sequence elements

SEQ ID NO	Description of sequence elements
SEQ ID NO:1	18BF003_pma-MCS plasmid sequence (1893bp)
SEQ ID NO:2	NotI-IR/DR-HS4I-CMV-BGI-MCS-hGHPA-HS4I-IR/DR-AsiSI (2900bp)
SEQ ID NO:3	SpeI-SV40p-EGFP-SV40pA-AgeI (1208bp)
SEQ ID NO:4	MluI-TRE _{3G} CuO-BGI-ClaI (908bp)
SEQ ID NO:5	MluI-TRE _{3G} -BGI-ClaI (880bp)
SEQ ID NO:6	MluI-TRE _{adv} CuO ₅₂ -BGI-ClaI (890bp)
SEQ ID NO:7	MluI-TRE _{adv} -BGI-ClaI (845bp)
SEQ ID NO:8	MluI-TRE _{3G} CuO ₁₄ -ClaI (414bp)
SEQ ID NO:9	MluI-TRE _{3G} CuO ₃₀ -ClaI (414bp)
SEQ ID NO:10	MluI-TRE _{3G} CuO ₁₀₀ -ClaI (415bp)
SEQ ID NO:11	MluI-TRE _{3G} CuO _{2x} -ClaI (472bp)
SEQ ID NO:12	MluI-TRE _{3G} CuO _{4x} -ClaI(588bp)
SEQ ID NO:13	MluI-TRE _{3G} CuO _{6x} -ClaI (704bp)
SEQ ID NO:14	CalI-Intron(mP1)-BamHI(210bp)
SEQ ID NO:15	ClaI-optiCymR-XhoI (633bp)
SEQ ID NO:16	SpeI-SV40p- optiHygroR-SV40pA-AgeI(1496bp)
SEQ ID NO:17	MluI-CAGGS-BGI(C&R)-MCS-SV40pA-AgeI (1979bp)
SEQ ID NO:18	ClaI-optirtTA _{3G} -XhoI (768bp)
SEQ ID NO:19	ClaI-rtTA _{adv} -XhoI (765bp)
SEQ ID NO:20	XhoI-ires-ECFP-BglII(1320bp)
SEQ ID NO:21	BamHI-optiSB-XhoI (1044bp)
SEQ ID NO:22	MluI-TRE _{adv} CuO ₃₂ -BGI-ClaI (376bp)
SEQ ID NO:23	TRE _{3G} CuO ₅₀ response element sequence
SEQ ID NO:24	TetO operator sequence
SEQ ID NO:25	minimal promoter sequence #1
SEQ ID NO:26	minimal promoter sequence #2
SEQ ID NO:27	CuO operator sequence
SEQ ID NO:28	TRE _{3G} CuO ₃₀ response element sequence
SEQ ID NO:29	TRE _{adv} CuO ₃₂ response element sequence
SEQ ID NO:30	TRE _{adv} CuO ₅₂ response element sequence
SEQ ID NO:31	sequence of human β -globulin intron (BGI)

Table 3. Description of functional elements of plasmids

Element name	Description of functions
IR/DR(L/R)	inverted repeat (IR) and direct repeat (DR) of SB transposon system
HS4I	4 core isolator of chicken beta-globulin highly sensitive position
HygroR	codon optimized sequence encoding hygromycin resistance gene
Sleeping Beauty (optiSB)	codon-optimized gene sequence encoding Sleeping Beauty (SB) transposase (SEQ ID NO: 21)
BGI	human β -globulin intron (SEQ ID NO:31)
CMV	strong expression promoter of human cytomegalovirus
MCS	multiple cloning site of restriction enzyme
CAGGS	chimeric promoter of cytomegalovirus promoter enhancer part and chicken β -actin

	promoter
BGI (C&R)	chimeric intron of chicken β -actin and rabbit β -globulin
Intron (mP1)	Mouse Prm1 gene intron (7-202bp in SEQ ID NO: 14, which is consistent with 942-1137bp in Accession number of FJ411376)
Intron (EF-1a)	EF-1 α intron of pEF1alpha-IRES-AcGFP1(Clontech) plasmid
Intron (pSI)	chimera intron of human beta globin and immunoglobulin heavy chain gene in pSI (Promega#E1721) plasmid
TRE _{3G} CuO/ TRE _{3G} CuO ₅₀	response element of Tet-On (based on 7xTetO sequence in TRE _{3G}) and Cumate complex inducible expression system designed in the present disclosure, wherein the CuO sequence is spaced 50bp behind the TATA box (SEQ ID NO: 23)
TRE _{3G} CuO ₁₄	response element of Tet-On (based on 7xTetO sequence in TRE _{3G}) and Cumate complex inducible expression system designed in the present disclosure, wherein the CuO sequence is spaced 14bp behind the TATA box (10-317bp in SEQ ID NO: 8)
TRE _{3G} CuO ₃₀	response element of Tet-On (based on 7xTetO sequence in TRE _{3G}) and Cumate complex inducible expression system designed in the present disclosure, wherein the CuO sequence is spaced 30bp behind the TATA box (SEQ ID NO: 28)
TRE _{3G} CuO ₁₀₀	response element of Tet-On (based on 7xTetO sequence in TRE _{3G}) and Cumate complex inducible expression system designed in the present disclosure, wherein the CuO sequence is spaced 100bp behind the TATA box (10-403bp in SEQ ID NO: 10)
TRE _{3G} CuO _{2x}	response element of Tet-On (based on 7xTetO sequence in TRE _{3G}) and Cumate complex inducible expression system designed in the present disclosure, wherein the two repeated CuO sequence is spaced 50bp behind the TATA box (10-411 bp in SEQ ID NO: 11)
TRE _{3G} CuO _{4x}	response element of Tet-On (based on 7xTetO sequence in TRE _{3G}) and Cumate complex inducible expression system designed in the present disclosure, wherein the 4 repeated CuO sequence is spaced 50bp behind the TATA box (10-527 bp in SEQ ID NO: 12)
TRE _{3G} CuO _{6x}	response element of Tet-On (based on 7xTetO sequence in TRE _{3G}) and Cumate complex inducible expression system designed in the present disclosure, wherein the 6 repeated CuO sequence is spaced 50bp behind the TATA box (10-643 bp in SEQ ID NO: 13)
TRE _{adv} CuO/TRE- advCuO ₅₂	response element of Tet-On (based on 7xTetO sequence in TRE _{adv}) and Cumate complex inducible expression system designed in the present disclosure, wherein the CuO sequence is spaced 52bp behind the TATA box (SEQ ID NO: 30)
TRE _{adv} CuO ₃₂	response element of Tet-On (based on 7xTetO sequence in TRE _{adv}) and Cumate complex inducible expression system designed in the present disclosure, wherein the CuO sequence is spaced 32 bp behind the TATA box (SEQ ID NO: 29)
TRE _{3G} /TRE _{adv}	response element of Tet-On inducible expression system: TRE _{3G} is the third generation of response element/TRE _{adv} is the second generation of response element (SEQ ID NO: 5/SEQ ID NO: 7)
SV40p	Simian vacuolar virus 40 promoter
IRES	ribosome entry site
EGFP	gene sequence encoding green fluorescent protein
ECFP	Gene sequence encoding cyan fluorescent protein
Luciferase (Luc)	Gene sequence encoding luciferase
polyA (hGHpA/SV40pA)	polyadenylation sequence of transcription terminator (hGHpA human growth factor terminator/SV40pA simian vacuolar virus 40 terminator)
optiCymR	codon-optimized coding sequence for repressor CymR protein of Cumate-induced expression system (13-621bp in SEQ ID NO: 15)
rtTA _{3G}	codon-optimized gene sequence encoding the third generation of rtTA _{3G} transactivation element of Tet-On regulatory system (13-756bp in SEQ ID NO: 18)
rtTA _{adv}	gene sequence encoding the second generation of rtTA _{adv} transactivation element of Tet-On regulatory system (13-756bp in SEQ ID NO: 19)

Table 4. Plasmid numbers and names

Plasmid Number	Plasmid Name
18BF003	pma-MCS

18BF007	pmaSBT3-2xHS4I-CMV-BGI-MCS
18BF011	pmaCMV-BGI-MCS
18BF210	pmaCMV-BGI-MCS-SV40p-EGFP
18BF211	pmaTRE _{3G} -BGI-MCS-SV40p-EGFP
18BF212	pmaTRE _{3G} CuO ₅₀ -BGI-MCS-SV40p-EGFP
18BF213	pmaTRE _{adv} -BGI-MCS-SV40p-EGFP
18BF214	pmaTRE _{adv} CuO ₅₂ -BGI-MCS-SV40p-EGFP
18BF215	pmaTRE _{3G} -MCS-SV40p-EGFP
18BF216	pmaTRE _{3G} CuO ₅₀ -MCS-SV40p-EGFP
18BF217	pmaTRE _{adv} -MCS-SV40p-EGFP
18BF218	pmaTRE _{adv} CuO ₅₂ -MCS-SV40p-EGFP
18BF229	pmaCMV-BGI-Luciferase-SV40p-EGFP
18BF232	pmaTRE _{adv} -Luciferase-SV40p-EGFP
18BF233	pmaTRE _{adv} CuO ₅₂ -Luciferase-SV40p-EGFP
18BF234	pmaTRE _{3G} -Luciferase-SV40p-EGFP
18BF235	pmaTRE _{3G} CuO ₅₀ -Luciferase-SV40p-EGFP
18BF236	pmaTRE _{3G} -BGI-Luciferase-SV40p-EGFP
18BF237	pmaTRE _{3G} CuO ₅₀ -BGI-Luciferase-SV40p-EGFP
18BF240	pmaTRE _{adv} -BGI-Luciferase-SV40p-EGFP
18BF241	pmaTRE _{adv} CuO ₅₂ -BGI-Luciferase-SV40p-EGFP
18BF251	pmaTRE _{3G} CuO ₁₄ -Luciferase-SV40p-EGFP
18BF252	pmaTRE _{3G} CuO ₃₀ -Luciferase-SV40p-EGFP
18BF253	pmaTRE _{3G} CuO ₁₀₀ -Luciferase-SV40p-EGFP
18BF254	pmaTRE _{3G} CuO _{2x} -Luciferase-SV40p-EGFP
18BF255	pmaTRE _{3G} CuO _{4x} -Luciferase-SV40p-EGFP
18BF256	pmaTRE _{3G} CuO _{6x} -Luciferase-SV40p-EGFP
18BF257	pmaTRE _{adv} CuO ₃₂ -Luciferase-SV40p-EGFP
18BF261	pmaTRE _{3G} CuO-BGI(C&R)-Luciferase-SV40p-EGFP
18BF262	pmaTRE _{3G} CuO-Intron(mP1)-Luciferase-SV40p-EGFP
18BF263	pmaTRE _{3G} CuO-Intron(EF-1a)-Luciferase-SV40p-EGFP
18BF264	pmaTRE _{3G} CuO-Intron(pSI)-Luciferase-SV40p-EGFP
19BF073	pmaSBT3-2xHS4I-CMV-BGI-optiCymR-HygroR
18BF008	pmaSBT3-2xHS4I-CAGGS-BGI (C&R)-MCS
18BF085	pmaSBT3-2xHS4I-CAGGS-BGI (C&R)-optirtTA _{3G}
18BF084	pmaSBT3-2xHS4I-CAGGS-BGI (C&R)-rtTA _{adv}
19BF075	pmaSBT3-2xHS4I-CAGGS-BGI (C&R)-optirtTA _{3G} -CMV-BGI-optiCymR-HygroR
19BF074	pmaSBT3-2xHS4I-CAGGS-BGI (C&R)-rtTA _{adv} -CMV-BGI-optiCymR-HygroR
18BF019	pmaCMV-BGI-optiSB-IRES-ECFP
19BF229	pmaSBT3-2xHS4I-CMV-BGI-Luciferase-SV40p-EGFP
19BF235	pmaSBT3-2xHS4I-TRE _{3G} CUO-Luciferase-SV40p-EGFP
19BF237	pmaSBT3-2xHS4I-TRE _{3G} CUO-BGI-Luciferase-SV40p-EGFP

Example 2: The effect of the position and number of CuO operator on the induced expression level and non-induced leaky expression level

The experiment described in the present Example is to study and verify the effect of the position and copy number of the CuO operator in the Tet-On and Cumate complex response element on the induced expression level and non-induced leaky expression level of the complex response element, and to optimize and confirm the optimal position and copy number of the CuO operator. In the present Example, based on the 7xTetO sequence in the TRE_{3G} response element and the minimal promoter sequence #1 (SEQ ID NO: 25), the response elements TRE_{3G}CuO₁₄

(10-317bp in SEQ ID NO: 8), TRE_{3G}CuO₃₀ (SEQ ID NO: 28), TRE_{3G}CuO₅₀ (SEQ ID NO: 23) and TRE_{3G}CuO₁₀₀ (10-403bp in SEQ ID NO: 10) were designed by linking a CuO operator sequence at 10 bp to 100 bp downstream of the TATA box (at 14 bp, 30 bp, 50 bp and 100 bp downstream of the TATA box, respectively), and a Luciferase reporter gene sequence was linked downstream of the 3' end of the above response elements, to construct plasmids 18BF251, 18BF252, 18BF235 and 18BF253. Further, in order to study the effect of multiple copies of CuO operators, 2, 4 and 6 copies of CuO operator sequences were respectively inserted at 50bp downstream of the TATA box, to design the response elements TRE_{3G}CuO_{2x} (10-411bp in SEQ ID NO: 11), TRE_{3G}CuO_{4x} (10-527bp in SEQ ID NO: 12) and TRE_{3G}CuO_{6x} (10-643bp in SEQ ID NO: 13), and then a Luciferase reporter gene sequence was linked downstream of 3' end of the above response elements to construct plasmids 18BF254, 18BF255 and 18BF256. In 293T-rtTA_{3G}-CymR cells stably expressing rtTA_{3G} and CymR genes, the above plasmids were transiently transfected, and the optimal position and number of CuO operator were validated by measuring the luciferase fluorescence value of samples added with both DOX and Cumate inducers and that of the control added with the DOX inducer only. The specific experimental methods were as follows:

1. Construction of 293T-rtTA_{adv}-CymR and 293T-rtTA_{3G}-CymR cells with SB transposon system

293T cells were seeded at 1.5E+06 cells per 60mm culture dish, and cultured in DMEM (Sigam, D6429) complete medium supplemented with 10% FBS (ExCell, 11H116) at 37°C and 5% CO₂. After 24 hours of culture, transfection was carried out according to the PEI method. During transfection, 500μL of transfection reagent which contained 5.5ug of total plasmid was added to each 60mm culture dish, and the mass ratio of total plasmid to PEI MAX (Polysciences, 24765-1) was 1:4. The transfection was carried out according to the PEI method, wherein the amount of total plasmid was 5.5μg. The transfection was performed with plasmids 19BF074:18BF019 at a molar ratio of 10:1 to obtain 293T-rtTA_{adv}-CymR cells; and the transfection was performed with plasmids 19BF075:18BF019 at a molar ratio of 10:1 to obtain 293T-rtTA_{3G}-CymR cells. The plasmid and PEI MAX were mixed uniformly, and then put into a culture dish after standing for 15 minutes. 3 hours after transfection, the medium was changed to a complete DMEM medium, and the transfection operation was completed. After 24 hours of transfection, the cells were digested with trypsin and all seeded in a 100mm culture dish (Corning, 430167), and 200μg/ml of hygromycin (Sangon Biotech A600230-0001) drug screening was performed for at least three passages. After the growth of cells under the pressure

of drug screening was consistent with that of the original 293T cells, the following experiments were performed.

2. Detection of the performance of each response element under both DOX and Cumate induction or under DOX induction alone by Luciferase fluorescence intensity assay

The 293T-rtTA_{3G}-CymR cells were seeded in a 96-well plate (Corning 3916) at 2.5E+04 cells per well, and the medium was 100 microliters of DMEM complete medium. After 24 hours of culture, the following plasmids were transfected according to the PEI method: 18BF234 (TRE_{3G}), 18BF251 (TRE_{3G}CuO₁₄), 18BF252 (TRE_{3G}CuO₃₀), 18BF235 (TRE_{3G}CuO₅₀), 18BF253 (TRE_{3G}CuO₁₀₀), 18BF254 (TRE_{3G}CuO_{2x}), 18BF255 (TRE_{3G}CuO_{4x}) and 18BF256 (TRE_{3G}CuO_{6x}); 10μL of transfection reagent was added into each well during transfection, which contains 0.3ug of total plasmids including 0.01ug of the above 8 kinds of plasmids to be tested and 0.29 ug of 18BF003 empty plasmid. The mass ratio of total plasmid to PEI MAX (Polysciences, 24765-1) was 1:4, and each plasmid was transfected into 6 wells. After 3 hours of transfection, the complete DMEM medium was changed; and the inducer 1ug/ml DOX (doxycycline hydrochloride, Sangon Biotech (Shanghai), A600889) and 200ug/ml Cumate (Aladdin, I107765) were added into 3 wells ; and 1ug/ml DOX alone was added into the remaining 3 wells. After 24 hours of transfection, relative light unit RLU of each well was detected using a Steady-Glo[®] Luciferase Assay System (Promega, E2610) kit according to the instruction (Promega, FB037), wherein the detection instrument was a fluorescence microplate reader (Perkin Elmer Victor V).

The results were shown in Figure 1: the induced expression level of response element in the Tet-On inducible expression system was affected by the CuO operator sequence; the closer the distance between the CuO operator and the TATA box of TRE response element was, the lower the expression level after induction was. However, as the distance between the CuO operator and the TATA box increases, the leaky expression level of non-Cumate induction became higher. Based on the results of the expression level after induction, the leaky expression level of non-Cumate induction and the ratio of Cumate induced/leaky expression level, it was determined that the optimal distance between the CuO and the TATA box was 30bp to 50bp, and the induced expression level was 3.06E+06 RLU and 4.53E+06 RLU, which was respectively 44.5% and 66.0% of the induced expression level of TRE_{3G} response element; and the ratio of induced/leaky expression level based on Cumate inducible expression system was 4.21 times and 2.81 times that of TRE_{3G} response element, respectively. The expression level after induction was further decreased by increasing the copy number of CuO operator to 2, 4 or 6, and the ratio

of Cumate induced/leaky expression level was not increased. Based on the above results, it is the optimal condition to insert the CuO operator and insert only one copy at a distance of 30bp to 50bp from the TATA box of the TRE response element.

Based on the 7xTetO sequence in the TRE_{adv} response element and the minimal promoter sequence #2 (SEQ ID NO: 26), the response elements TRE_{adv}CuO₃₂ (SEQ ID NO:29) and TRE_{adv}CuO₅₂ (SEQ ID NO:30) were designed by linking a CuO operator at 32 bp and 52 bp downstream of the TATA box, and a Luciferase reporter gene sequence was linked downstream of the 3' end of the above response elements, to construct plasmids 18BF257 and 18BF233. By the same method as above, the detected induced expression levels were 3.26+06 RLU and 4.88E+06 RLU, respectively, which were 37.8% and 53.4% of the induced expression level for TRE_{adv} response element; the ratio of induced/leaky expression level based on Cuamte inducible expression system was 4.97 times and 3.23 times that for the TRE_{adv} response element, respectively. In the subsequent examples, experiments were performed with response elements TRE_{adv}CuO₅₂ and TRE_{3G}CuO₅₀, and they were marked as TRE_{adv}CuO and TRE_{3G}CuO response elements, respectively.

Example 3: The effect of single regulation/complex regulation and introns on the induced expression level and non-induced leaky expression level

In Tet-On inducible expression system, the induced transcription activity and the leaky transcription activity were comprehensively affected by the TetO operator linking sequence of the TRE response element and the minimal promoter sequence, as well as different mutants of the transactivator rtTA. In addition, the transport and stability of messenger ribonucleic acid may be enhanced by linking a spliceable intron sequence downstream of the 3' end of the response element and upstream of the 5' end of the regulated nucleic acid fragment of interest, but the induced transcription activity and leaky transcription activity of response element of the inducible expression system may also be affected. Based on this, the optimal design scheme of complex response element was determined in this Example based on the induced expression level and the ratio of induced/leaky expression level of Luciferase gene under the combination of the following conditions: (1) Tet-On and Cumate complex response elements TRE_{adv}CuO and TRE_{3G}CuO designed based on the TRE_{adv} and TRE_{3G} response element sequences; (2) whether an intron was linked; (3) being regulated by the transactivator rtTA_{adv} or rtTA_{3G}. A total of 8 design schemes were compared in this Example: TRE_{adv} (18BF232), TRE_{adv}CuO (18BF233), TRE_{3G} (18BF234), TRE_{3G}CuO (18BF235), TRE_{adv}-BGI (18BF240, wherein the response

element was TRE_{adv}, and a human β -globulin intron was linked between the 3' end of the response element and the 5' end of the Luciferase gene), TRE_{adv}CuO-BGI (18BF241, wherein the response element was TRE_{adv}CuO, and a human β -globulin intron was linked between the 3' end of the response element and the 5' end of the Luciferase gene), TRE_{3G}-BGI (18BF236, wherein the response element was TRE_{3G}, and a human β -globulin intron was linked between the 3' end of the response element and the 5' end of the Luciferase gene) and TRE_{3G}CuO-BGI (18BF237, wherein the response element was TRE_{3G}CuO, and a human β -globulin intron was linked between the 3' end of the response element and the 5' end of the Luciferase gene). The promoter CMV-BGI (18BF229, wherein the promoter was CMV, and a human β -globulin intron was linked between the 3' end of the CMV promoter and the 5' end of the Luciferase gene) was used as a positive control. In 293T-rtTA_{adv}-CymR cells stably expressing rtTA_{adv} and CymR genes and 293T-rtTA_{3G}-CymR cells stably expressing rtTA_{3G} and CymR genes, the above plasmids were transiently transfected, and the optimal combination of complex response elements was verified by measuring the luciferase fluorescence value of samples added with both DOX and Cumate inducers and that of the control added with no inducer. The specific experimental methods were as follows:

The 293T-rtTA_{adv}-CymR and 293T-rtTA_{3G}-CymR cells constructed in Example 2 were seeded into a 96-well plate (Corning 3916) at 2.5E+04 cells per well, and the medium was 100 microliters of DMEM complete medium. After 24 hours of culture, the above 9 plasmids were transfected according to the PEI method. 10 μ L of transfection reagent, which contains 0.3 μ g of total plasmids including 0.01 μ g of the above 9 plasmids to be tested and 0.29 μ g of 18BF003 empty plasmid, was added into each well during transfection. The mass ratio of total plasmid to PEI MAX (Polysciences, 24765-1) was 1:4, and each plasmid was transfected into 6 wells for 293T-rtTA_{adv}-CymR and 293T-rtTA_{3G}-CymR cells respectively. After 3 hours of transfection, the DMEM complete medium was replaced; the inducer 1 μ g/ml DOX and 200 μ g/ml Cumate were added into 3 wells for each cell; and the same amount of medium was added into the remaining 3 wells as a control. After 24 hours of transfection, relative light unit RLU of each well was detected using a Steady-Glo[®] Luciferase Assay System (Promega, E2610) kit according to the instructions (Promega, FB037), wherein the detection instrument was a fluorescence microplate reader (Perkin Elmer Victor V).

The experimental results were shown in Figure 2: (1) Compared with the TRE single regulatory response element, the TRECuO complex response element significantly increases the ratio of induced/leaky expression level by one to two orders of magnitude. Compared with the

TRE_{adv}CuO complex response element, the TRE_{3G}CuO complex response element can better control the leaky expression. In 293T-rtTA_{adv}-CymR cells, the induced/leaky expression ratios of TRE_{3G}CuO and TRE_{adv}CuO were 1565 and 1345 times, respectively, which showed an increase of 16.4%; in 293T-rtTA_{3G}-CymR cells, the induced/leaky expression ratios of TRE_{3G}CuO and TRE_{adv}CuO were respectively 2635 and 1915 times, which showed an increase of 37.6%. However, compared with the TRE single regulatory response element, the TRECuO complex response element significantly decreased the induced expression level. In 293T-rtTA_{adv}-CymR cells, the induced expression levels of TRE_{3G}CuO and TRE_{adv}CuO were 50.4% and 36.3% of those of the corresponding TRE_{3G} and TRE_{adv} single regulatory response elements, respectively; in 293T-rtTA_{3G}-CymR cells, the induced expression levels of TRE_{3G}CuO and TRE_{adv}CuO were 61.8% and 40.3% of those of the single regulatory response element, respectively. (2) In terms of induced expression level, the TRE_{3G}CuO complex response element was more sensitive to the downstream intron than the TRE_{adv}CuO complex response element, and the induced expression level in the presence of the intron BGI was significantly increased. In 293T-rtTA_{adv}-CymR cells, the induced expression levels of TRE_{3G}CuO and TRE_{3G}CuO-BGI were 3.4E+06 RLU and 10.6E+06 RLU, respectively, which were 50.4% and 157.9% of the induced expression level of TRE_{3G}, and linking an intron can increase the induced expression level of TRE_{3G}CuO complex response element by 2.13 times; in 293T-rtTA_{3G}-CymR cells, the induced expression levels of TRE_{3G}CuO and TRE_{3G}CuO-BGI were 4.24E+06 and 11.3E+06, which were 61.8% and 165.0% of the induced expression level of TRE_{3G}, respectively, and linking an intron can increase the induced expression level of TRE_{3G}CuO complex response element by 1.67 times. (3) Regarding the induced expression level, there was no significant difference between the transactivator rtTA_{3G} and rtTA_{adv} in the 8 design schemes, but rtTA_{3G} had a significant advantage regarding the ratio of induced/leaky expression levels. The average ratio of induced/leaky expression level of rtTA_{3G} in 8 design schemes was 1110 times, which was 1.68 times that of rtTA_{adv} (the average ratio of induced/leaky expression level of rtTA_{adv} was 661 times).

Based on the above results, the TRE_{3G}CuO complex response element has a better ability to control the leaky expression as compared to TRE_{adv} and TRE_{3G} and TRE_{adv}CuO complex response elements; with linking the intron BGI downstream, the TRE_{3G}CuO complex response element can maintain the control of the leaky expression while greatly improving the induced expression level. Under the regulation of rtTA_{3G}, the induced expression level of TRE_{3G}CuO-BGI was 80.6% of that of TRE_{adv}, 80.5% of that of TRE_{adv}-BGI, 199.9% of that of TRE_{adv}CuO, 147.0% of that of TRE_{adv}CuO-BGI, 165.0% of that of TRE_{3G}, 81.5% of that of TRE_{3G}-BGI and

267.2% of that of TRE_{3G}CuO, reaching 84.7% of that of the constitutive promoter CMV-BGI; the ratio of induced/non-induced expression level of TRE_{3G}CuO-BGI was 24.5 times that of TRE_{adv}, 43.4 times that of TRE_{adv}-BGI, 1.2 times that of TRE_{adv}CuO, 1.6 times that of TRE_{adv}CuO-BGI, 5.9 times that of TRE_{3G}, 8.9 times that of TRE_{3G}-BGI and 0.8 times that of TRE_{3G}CuO.

Example 4: The effect of different introns on the induced expression level and non-induced leaky expression level of TRE_{3G}CuO complex response element

In Example 3, the linkage of the intron BGI significantly increased the induced transcription activity of the TRE_{3G}CuO complex induction response element and the expression level of the target gene of Luciferase. It was verified in this Example whether other introns had similar effects. In this example, the introns in the four commonly used plasmid vectors were cloned between the 3'downstream of the TRE_{3G}CuO complex response element and the 5'upstream of the luciferase reporter gene, to construct plasmids 18BF261, 18BF262, 18BF263 and 18BF264 respectively containing BGI (C&R), Intron (mP1), Intron (EF-1a) and Intron (pSI) introns, and the specific methods for constructing a plasmid was described in Example 1. In 293T-rtTA_{3G}-CymR cells stably expressing rtTA_{3G} and CymR genes, the above plasmids and TRE_{3G}CuO (18BF235), TRE_{3G}CuO-BGI (18BF237) and CMV-BGI (18BF229) plasmids were transiently transfected, respectively, and the effect of each intron on the induced expression and non-induced leaky expression of the TRE_{3G}CuO complex response element was verified by measuring the luciferase fluorescence values of samples added with both DOX and Cumate inducers and that of the control added with no inducer. The specific experimental methods were as follows:

The 293T-rtTA_{3G}-CymR cells were seeded in a 96-well plate (Corning 3916) at 2.5E+04 cells per well, and the medium was 100 microliters of DMEM complete medium. After 24 hours of culture, the above 7 plasmids were respectively transfected according to the PEI method. 10μL of transfection reagent, which contains 0.3ug of total plasmids including 0.01ug of the above 7 plasmids to be tested and 0.29 μg of 18BF003 empty plasmid, was added into each well during transfection. The mass ratio of total plasmid to PEI MAX (Polysciences, 24765-1) was 1:4, and each plasmid was transfected into 6 wells. After 3 hours of transfection, the DMEM complete medium was replaced; the inducer 1ug/ml DOX and 200ug/ml Cumate were added into 3 wells; and the same amount of medium was added into the remaining 3 wells. After 24 hours of transfection, relative light unit RLU of each well was detected using a Steady-Glo[®] Luciferase

Assay System (Promega, E2610) kit according to the instructions (Promega, FB037), wherein the detection instrument was a fluorescence microplate reader (Perkin Elmer Victor V).

The results were shown in Figure 3: there is no significant difference in the induced expression level and the ratio of induced/leaky expression level between plasmids 18BF261, 18BF262, 18BF263 and 18BF264 containing four other introns and the TRE_{3G}CuO-BGI (18BF237), all of which can achieve the effect of TRE_{3G}CuO-BGI as described in Example 3. In the intron, a 5'splice site, a 3'splice site, and a splice branch point are required for splicing. This Example demonstrates that spliceable introns which meet the above conditions can achieve the experimental effects as described in the present disclosure. Therefore, the effects as described in the present disclosure should not be limited by specific intron sequences, and any sequence capable of RNA splicing in mammalian cells can achieve the above functions. Intron sequences that may be selected include, but are not limited to, introns in common cloning vectors, such as a rabbit β -globulin intron, a hybrid intron derived from human β -globulin and immunoglobulin heavy chain intron, EF-1 α intron A, SV40 intron, a hybrid intron derived from adenovirus and immunoglobulin heavy chain intron, a modified human cytomegalovirus intron, a hybrid intron derived from chicken β -actin (CBA) and mouse microvirus (MMV) intron, a chimera derived from chicken β -actin and rabbit β -globulin intron, and a mP1 intron; or any intron of any gene of any eukaryote; or an artificial intron sequence designed based on the intron splicing rules.

Example 5: Study of induced transcription activity of the TRE_{3G}CuO complex response element under different induction combinations

In this Example, stable cell lines, under complex regulation of Tet-ON and Cumate, TRE_{3G}CuO or TRE_{3G}CuO-intron, and with Luciferase as the reporter gene, were constructed. Then, based on these cell lines, induced transcription activities of the reporter gene under the following 4 induction conditions were detected: without inducer, Cumate only, DOX only or both DOX and Cumate inducers. In this Example, the Luciferase reporter gene was taken as an example, and in principle, the method of the present disclosure was not affected by the nucleic acid fragment of interest. The specific experimental methods were as follows:

1. Construction of a Luciferase stable cell line regulated by DOX and Cumate by using SB transposon system:

293T cells were seeded at 1.5E+06 cells per 60mm culture dish, and cultured in DMEM (Sigam, D6429) complete medium supplemented with 10% FBS (ExCell, 11H116) at 37°C and 5% CO₂. After 24 hours of culture, transfection was carried out according to the PEI method.

During transfection, 500 μ L of transfection reagent which contained 5.5 μ g of total plasmid was added to each 60mm culture dish, and the mass ratio of total plasmid to PEI MAX (Polysciences, 24765-1) was 1:4; and thereby 3 Luciferase stable cell lines were constructed. The transfection plasmids were transfected at the 19BF075:19BF229:18BF019 molar ratio of 5:5:1 to obtain 293T(T&C)-CMV-BGI-Luc cells; the transfection plasmids were transfected at the 19BF075:19BF235:18BF019 molar ratio of 5:5:1 to obtain 293T(T&C)-TRE_{3G}CuO-Luc cells; the transfection plasmids were transfected at the 19BF075:19BF237:18BF019 molar ratio of 5:5:1 to obtain 293T(T&C)-TRE_{3G}CuO-BGI-Luc cells. The plasmid and PEI MAX were mixed uniformly, and then put into a culture dish after standing for 15 minutes. 3 hours after transfection, the medium was changed to a complete DMEM medium, and the transfection operation was completed. After 24 hours of transfection, the cells were digested with trypsin and all seeded in a 100mm culture dish (Corning, 430167), and 200 μ g/ml of hygromycin (Sangon Biotech A600230-0001) drug screening was performed for at least three passages. After the growth of cells under the pressure of drug screening was consistent with that of the original 293T cells, the above 3 cells were diluted to 1 cell per well by a limiting dilution method and seeded in a 96-well plate; when the cells grew to about 50% of the bottom area of the well, the wells with strong EGFP green fluorescence intensity and uniform luminescence were selected under the fluorescence microscope, and 3 independent clones of each cell were selected for the following experiments.

2. Induction of Luciferase expression by using different combinations of DOX and Cumate:

For the above 3 cells, 3 independent clones for each cell were seeded into a 96-well plate (Corning 3916) at 2.5E+04 cells per well, with 8 wells per clone, and the medium was 100 microliters of DMEM complete medium. After 24 hours of culture, the medium was replaced and the following reagents were added into the duplicate wells of the 8 wells for each cell: (1) the same amount of medium, (2) Cumate at a final concentration of 200 μ g/ml, (3) DOX at a final concentration of 1 μ g/ml, or (4) DOX at a final concentration of 1 μ g/ml and Cumate at a final concentration of 200 μ g/ml. After 24 hours of further culture, relative light unit RLU of each well was detected using a Steady-Glo® Luciferase Assay System (Promega, E2610) kit according to the instruction (Promega, FB037), wherein the detection instrument was a fluorescence microplate reader (Perkin Elmer Victor V).

The experimental results were shown in Figure 4: under the condition of only adding Cumate, the induced transcription activities of TRE_{3G}CuO and TRE_{3G}CuO-BGI were 1.36E+04

RLU and 4.16×10^4 RLU, which were respectively 4.31 times and 3.27 times higher than those obtained by no induction. Under the condition of only adding DOX, the induced transcription activities of TRE_{3G}CuO and TRE_{3G}CuO-BGI were 2.14×10^5 RLU and 5.34×10^5 RLU, which were respectively 67.66 times and 42.07 times higher than those obtained by no induction, and were 15.71 times and 12.86 times higher than those obtained under the induction condition of only adding Cumate. Under the condition of adding both DOX and Cumate, the induced transcription activities of TRE_{3G}CuO and TRE_{3G}CuO-BGI were 7.54×10^6 RLU and 1.29×10^7 RLU, which were respectively 2381.05 times and 1011.81 times higher than those obtained by no induction; were 552.99 times and 309.27 times higher than those obtained under the induction condition of only adding Cumate; were 35.19 times and 24.05 times higher than those obtained under the induction condition of only adding DOX. Based on the above results, TRE_{3G}CuO and TRE_{3G}CuO-BGI complex response elements can regulate the induced transcription activity of the nucleic acid fragment of interest at various levels according to different combinations of inducers, with ranges of 4.31 times to 2381.05 times and 3.27 times to 1011.81 times, respectively; when both DOX and Cumate inducers were added, the maximum induced transcription activities can reach 47.84% and 81.54% of that of CMV promoter, respectively. With further optimization of the concentration of DOX and/or Cumate inducer, the transcription activity of the nucleic acid fragment of interest can be further finely regulated at different transcription activity levels.

I/We claim:

1. A nucleic acid sequence, comprising at least 2 copies of TetO-operator sequences capable of binding to a transactivator rtTA regulatable by tetracycline or a derivative thereof, and 1 copy of a minimal promoter sequence containing a TATA box sequence, and at least 1 copy of a CuO-operator sequence capable of binding to a transcription repressor CymR regulatable by cumate, wherein the CuO-operator sequence is downstream of the 3' end of the TATA box sequence, and is 10 bp to 100 bp apart from the TATA box.

2. The nucleic acid sequence according to claim 1, wherein the CuO-operator sequence is 30 bp to 50 bp apart from the TATA box.

3. The nucleic acid sequence according to claim 1, wherein the CuO-operator sequence is 50 bp apart from the TATA box.

4. The nucleic acid sequence according to claim 1, wherein the TetO-operator sequence is set forth in SEQ ID NO: 24, and/or the minimal promoter sequence is set forth in SEQ ID NO: 25 or SEQ ID NO: 26, and/or the CuO-operator sequence is set forth in SEQ ID NO:27.

5. The nucleic acid sequence according to claim 4, wherein the nucleic acid sequence is set forth in SEQ ID NO:23, SEQ ID NO:28, SEQ ID NO:29 or SEQ ID NO:30.

6. The nucleic acid sequence according to any one of claims 1 to 5, further comprising a spliceable intron sequence at the 3' end thereof.

7. A vector comprising the nucleic acid sequence according to any one of claims 1 to 6.

8. The vector according to claim 7, wherein the vector is an expression vector comprising a nucleic acid fragment of interest downstream of the 3' end of the nucleic acid sequence according to any one of claims 1 to 6, and the transcription of the nucleic acid fragment of interest is controlled by the nucleic acid sequence according to any one of claims 1 to 6.

9. A host cell comprising the nucleic acid sequence according to any one of claims 1 to 6 or the vector according to claim 7 or 8.

10. A method for inducing the expression of a nucleic acid fragment of interest in a host cell, comprising the following steps:

- (1) introducing the vector according to claim 8 and a coding sequence of rtTA and a coding sequence of CymR into the host cell;
- (2) expressing rtTA and CymR in the host cell subjected to step (1); and
- (3) providing tetracycline or a derivative thereof and cumate or a functional analog thereof for the host cell subjected to step (2).

11. The method according to claim 10, wherein the rtTA is rtTA_{adv} or rtTA_{3G}.

12. The method according to claim 10, wherein the coding sequence of rtTA is set forth in SEQ ID NO: 18.

13. The method according to claim 10, wherein the coding sequence of CymR is set forth in SEQ ID NO: 15.