

GENE EXPRESSION PROFILE ALGORITHM AND TEST FOR DETERMINING PROGNOSIS OF PROSTATE CANCER

RELATED APPLICATIONS

[0001] This application claims the benefit of priority of U.S. Provisional Application Nos. 61/593,106, filed January 31, 2012; 61/672,679, filed July 17, 2012; and 61/713,734, filed October 15, 2012, all of which are hereby incorporated by reference in their entirety.

TECHNICAL FIELD

[0002] The present disclosure relates to molecular diagnostic assays that provide information concerning gene expression profiles to determine prognostic information for cancer patients. Specifically, the present disclosure provides an algorithm comprising genes, or co-expressed genes, the expression levels of which may be used to determine the likelihood that a prostate cancer patient will experience a positive or a negative clinical outcome.

INTRODUCTION

[0003] The introduction of prostate-specific antigen (PSA) screening in 1987 has led to the diagnosis and aggressive treatment of many cases of indolent prostate cancer that would never have become clinically significant or caused death. The reason for this is that the natural history of prostate cancer is unusual among malignancies in that the majority of cases are indolent and even if untreated would not progress during the course of a man's life to cause suffering or death. While approximately half of men develop invasive prostate cancer during their lifetimes (as detected by autopsy studies) (B. Halpert et al, *Cancer* 16: 737-742 (1963); B. Holund, *Scand J Urol Nephrol* 14: 29-35 (1980); S. Lundberg et al., *Scand J Urol Nephrol* 4: 93-97 (1970); M. Yin et al., *J Urol* 179: 892-895 (2008)), only 17% will be diagnosed with prostate cancer and only 3% will die as a result of prostate cancer. Cancer Facts and Figures. Atlanta, GA: American Cancer Society (2010); JE Damber et al., *Lancet* 371: 1710-1721 (2008).

[0004] However, currently, over 90% of men who are diagnosed with prostate cancer, even low-risk prostate cancer, are treated with either immediate radical prostatectomy or definitive radiation therapy. MR Cooperberg et al., *J Clin Oncol* 28: 1117-1123 (2010); MR Cooperberg et al., *J Clin Oncol* 23: 8146-8151 (2005). Surgery and radiation therapy reduce the risk of recurrence and death from prostate cancer (AV D'Amico et al., *Jama* 280: 969-974 (1998); M Han et al., *Urol Clin North Am* 28: 555-565 (2001); WU Shipley et al., *Jama*

281: 1598-1604 (1999); AJ Stephenson et al., *J Clin Oncol* 27: 4300-4305 (2009)), however estimates of the number of men that must be treated to prevent one death from prostate cancer range from 12 to 100. A Bill-Axelsson et al., *J Natl Cancer Inst* 100: 1144-1154 (2008); J Hugosson et al., *Lancet Oncol* 11: 725-732 (2010); LH Klotz et al., *Can J Urol* 13 Suppl 1: 48-55 (2006); S Loeb et al., *J Clin Oncol* 29: 464-467 (2011); FH Schroder et al., *N Engl J Med* 360: 1320-1328 (2009). This over-treatment of prostate cancer comes at a cost of money and toxicity. For example, the majority of men who undergo radical prostatectomy suffer incontinence and impotence as a result of the procedure (MS Litwin et al., *Cancer* 109: 2239-2247 (2007); MG Sanda et al., *N Engl J Med* 358: 1250-1261 (2008), and as many as 25% of men regret their choice of treatment for prostate cancer. FR Schroeck et al., *Eur Urol* 54: 785-793 (2008).

[0005] One of the reasons for the over-treatment of prostate cancer is the lack of adequate prognostic tools to distinguish men who need immediate definitive therapy from those who are appropriate candidates to defer immediate therapy and undergo active surveillance instead. For example, of men who appear to have low-risk disease based on the results of clinical staging, pre-treatment PSA, and biopsy Gleason score, and have been managed with active surveillance on protocols, 30-40 % experience disease progression (diagnosed by rising PSA, an increased Gleason score on repeat biopsy, or clinical progression) over the first few years of follow-up, and some of them may have lost the opportunity for curative therapy. HB Carter et al., *J Urol* 178: 2359-2364 and discussion 2364-2355 (2007); MA Dall'Era et al., *Cancer* 112: 2664-2670 (2008); L Klotz et al., *J Clin Oncol* 28: 126-131 (2010). Also, of men who appear to be candidates for active surveillance, but who undergo immediate prostatectomy anyway, 30-40% are found at surgery to have higher risk disease than expected as defined by having high-grade (Gleason score of 3+4 or higher) or non-organ-confined disease (extracapsular extension (ECE) or seminal vesicle involvement (SVI)). SL et al., *J Urol* 181: 1628-1633 and discussion 1633-1624 (2009); CR Griffin et al., *J Urol* 178: 860-863 (2007); PW Mufarrij et al., *J Urol* 181: 607-608 (2009).

[0006] Estimates of recurrence risk and treatment decisions in prostate cancer are currently based primarily on PSA levels and/or clinical tumor stage. Although clinical tumor stage has been demonstrated to have a significant association with outcome, sufficient to be included in pathology reports, the College of American Pathologists Consensus Statement noted that variations in approach to the acquisition, interpretation, reporting, and analysis of this information exist. C. Compton, et al., *Arch Pathol Lab Med* 124:979-992 (2000). As a

consequence, existing pathologic staging methods have been criticized as lacking reproducibility and therefore may provide imprecise estimates of individual patient risk.

SUMMARY

[0007] This application discloses molecular assays that involve measurement of expression level(s) of one or more genes or gene subsets from a biological sample obtained from a prostate cancer patient, and analysis of the measured expression levels to provide information concerning the likelihood of a clinical outcome. For example, the likelihood of a clinical outcome may be described in terms of a quantitative score based on clinical or biochemical recurrence-free interval, overall survival, prostate cancer-specific survival, upstaging/upgrading from biopsy to radical prostatectomy, or presence of high grade or non-organ confined disease at radical prostatectomy.

[0008] In addition, this application discloses molecular assays that involve measurement of expression level(s) of one or more genes or gene subsets from a biological sample obtained to identify a risk classification for a prostate cancer patient. For example, patients may be stratified using expression level(s) of one or more genes, positively or negatively, with positive clinical outcome of prostate cancer, or with a prognostic factor. In an exemplary embodiment, the prognostic factor is Gleason score.

[0009] The present invention provides a method of predicting the likelihood of a clinical outcome for a patient with prostate cancer comprising determination of a level of one or more RNA transcripts, or an expression product thereof, in a biological sample containing tumor cells obtained from the patient, wherein the RNA transcript, or its expression product, is selected from the 81 genes shown in Fig. 1 and listed in Tables 1A and 1B. The method comprises assigning the one or more RNA transcripts, or an expression product thereof, to one or more gene groups selected from a cellular organization gene group, basal epithelia gene group, a stress response gene group, an androgen gene group, a stromal response gene group, and a proliferation gene group. The method further comprises calculating a quantitative score for the patient by weighting the level of the one or more RNA transcripts or an expression product thereof, by their contribution to a clinical outcome and predicting the likelihood of a clinical outcome for the patient based on the quantitative score. In an embodiment of the invention, an increase in the quantitative score correlates with an increased likelihood of a negative clinical outcome.

[0010] In a particular embodiment, the one or more RNA transcripts, or an expression product thereof, is selected from BIN1, IGF1, C7, GSN, DES, TGFB1I1, TPM2, VCL,

FLNC, ITGA7, COL6A1, PPP1R12A, GSTM1, GSTM2, PAGE4, PPAP2B, SRD5A2, PRKCA, IGFBP6, GPM6B, OLFML3, HLF, CYP3A5, KRT15, KRT5, LAMB3, SDC1, DUSP1, EGFR1, FOS, JUN, EGR3, GADD45B, ZFP36, FAM13C, KLK2, ASPN, SFRP4, BGN, THBS2, INHBA, COL1A1, COL3A1, COL1A2, SPARC, COL8A1, COL4A1, FN1, FAP, COL5A2, CDC20, TPX2, UBE2T, MYBL2, and CDKN2C. BIN1, IGF1, C7, GSN, DES, TGFB111, TPM2, VCL, FLNC, ITGA7, COL6A1, PPP1R12A, GSTM1, GSTM2, PAGE4, PPAP2B, SRD5A2, PRKCA, IGFBP6, GPM6B, OLFML3, and HLF are assigned to the cellular organization gene group. CYP3A5, KRT15, KRT5, LAMB3, and SDC1 are assigned to the basal epithelial gene group. DUSP1, EGFR1, FOS, JUN, EGR3, GADD45B, and ZFP36 are assigned to the stress response gene group. FAM13C, KLK2, AZGP1, and SRD5A2 are assigned to the androgen gene group. ASPN, SFRP4, BGN, THBS2, INHBA, COL1A1, COL3A1, COL1A2, SPARC, COL8A1, COL4A1, FN1, FAP and COL5A2 are assigned to the stromal response gene group. CDC20, TPX2, UBE2T, MYBL2, and CDKN2C are assigned to the proliferation gene group. The method may further comprise determining the level of at least one RNA transcript, or an expression product thereof, selected from STAT5B, NFAT5, AZGP1, ANPEP, IGFBP2, SLC22A3, ERG, AR, SRD5A2, GSTM1, and GSTM2.

[0011] In an embodiment of the invention, the level of one or more RNA transcripts, or an expression product thereof, from each of the stromal response gene group and the cellular organization gene group are determined. In another embodiment, the level of one or more RNA transcripts, or expression products thereof, from each of the stromal response gene group and PSA gene group are determined. Additionally, the level of one or more RNA transcripts, or expression products thereof, from the cellular organization gene group and/or proliferation gene group may be determined. In this embodiment, gene(s) to be assayed from the stromal response gene group may be selected from ASPN, BGN, COL1A1, SPARC, FN1, COL3A1, COL4A1, INHBA, THBS2, and SFRP4; gene(s) to be assayed from the androgen gene group may be selected from FAM13C and KLK2; gene(s) to be assayed from the cellular organization gene group may be selected from FLNC, GSN, GSTM2, IGFBP6, PPAP2B, PPP1R12A, BIN1, VCL, IGF1, TPM2, C7, and GSTM1; and gene(s) to be assayed from the proliferation gene group may be selected from TPX2, CDC20, and MYBL2.

[0012] In a particular embodiment, the RNA transcripts, or their expression products, are selected from BGN, COL1A1, SFRP4, FLNC, GSN, TPM2, TPX2, FAM13C, KLK2, AZGP1, GSTM2, and SRD5A2. BGN, COL1A1, and SFRP4 are assigned to the stromal response gene group; FLNC, GSN, and TPM2 are assigned to the cellular organization gene

group; and FAM13C and KLK2 are assigned to the androgen gene group. The level of the RNA transcripts, or their expression products, comprising at least one of the gene groups selected from the stromal response gene group, cellular organization gene group, and androgen gene group, may be determined for the method of the invention. In any of the embodiments, the androgen gene group may further comprise AZGP1 and SRD5A2.

[0013] In addition, the level of any one of the gene combinations shown in Table 4 may be determined. For instance, the RS0 model in Table 4 comprises determining the levels of the RNA transcripts, or gene expression products thereof, of ASPN, BGN, COL1A1, SPARC, FLNC, GSN, GSTM2, IGFBP6, PPAP2B, PPP1R12A, TPX2, CDC20, MYBL2, FAM13C, KLK2, STAT5B, and NFAT5. Furthermore, any one of the algorithms shown in Table 4 may be used to calculate the quantitative score for the patient.

BRIEF DESCRIPTION OF THE DRAWING

[0014] Figure 1 is a dendrogram depicting the association of the 81 genes selected from the gene identification study.

[0015] Figures 2A-2E are scatter plots showing the comparison of normalized gene expression (Cp) for matched samples from each patient where the x-axis is the gene expression from the primary Gleason pattern RP sample (PGP) and the y-axis is the gene expression from the biopsy (BX) sample. Fig. 2A: All ECM (stromal response) genes; Fig. 2B: All migration (cellular organization) genes; Fig. 2C: All proliferation genes; Fig. 2D: PSA (androgen) genes; Fig. 2E: other genes from the 81 gene list that do not fall within any of these four gene groups.

[0016] Figures 3A-3D are range plots of gene expression of individual genes within each gene group in the biopsy (BX) and PGP RP samples. Fig. 3A: All ECM (stromal response) genes; Fig. 3B: All migration (cellular organization) genes; Fig. 3C: All proliferation genes; Fig. 3D: other genes from the 81 gene list that do not fall within any of the gene groups.

[0017] Figure 4 is a schematic illustration of the clique-stack method used to identify co-expressed genes.

[0018] Figure 5 shows examples of cliques and stacks. Fig. 5(a) is an example of a graph that is not a clique; Fig. 5(b) is an example of a clique; Fig. 5(c) is an example of a clique but is not a maximal clique.

[0019] Figure 6 is a graph showing two maximal cliques: 1-2-3-4-5 and 1-2-3-4-6.

[0020] Figure 7 schematically illustrates stacking of two maximal cliques.

[0021] Figure 8 is a graph showing that RS27 and CAPRA risk groups predict freedom from high-grade or non-organ-confined disease.

[0022] Figure 9 is a graph showing that RS27 and AUA risk groups predict freedom from high-grade or non-organ-confined disease.

[0023] Figure 10 is a graph showing time to clinical recurrence of PTEN low and PTEN normal patients from the gene identification study.

[0024] Figure 11 is a graph showing time to clinical recurrence of patients from the gene identification study stratified into PTEN low/normal and TMPRSS-ERG negative/positive.

DEFINITIONS

[0025] Unless defined otherwise, technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Singleton et al., Dictionary of Microbiology and Molecular Biology 2nd ed., J. Wiley & Sons (New York, NY 1994), and March, Advanced Organic Chemistry Reactions, Mechanisms and Structure 4th ed., John Wiley & Sons (New York, NY 1992), provide one skilled in the art with a general guide to many of the terms used in the present application.

[0026] One skilled in the art will recognize many methods and materials similar or equivalent to those described herein, which could be used in the practice of the present invention.

Indeed, the present invention is in no way limited to the methods and materials described herein. For purposes of the invention, the following terms are defined below.

[0027] The terms “tumor” and “lesion” as used herein, refer to all neoplastic cell growth and proliferation, whether malignant or benign, and all pre-cancerous and cancerous cells and tissues. Those skilled in the art will realize that a tumor tissue sample may comprise multiple biological elements, such as one or more cancer cells, partial or fragmented cells, tumors in various stages, surrounding histologically normal-appearing tissue, and/or macro or micro-dissected tissue.

[0028] The terms “cancer” and “cancerous” refer to or describe the physiological condition in mammals that is typically characterized by unregulated cell growth. Examples of cancer in the present disclosure include cancer of the urogenital tract, such as prostate cancer.

[0029] As used herein, the term “prostate cancer” is used in the broadest sense and refers to all stages and all forms of cancer arising from the tissue of the prostate gland.

[0030] Staging of the cancer assists a physician in assessing how far the disease has progressed and to plan a treatment for the patient. Staging may be done clinically (clinical staging) by physical examination, blood tests, or response to radiation therapy, and/or

pathologically (pathologic staging) based on surgery, such as radical prostatectomy. According to the tumor, node, metastasis (TNM) staging system of the American Joint Committee on Cancer (AJCC), AJCC Cancer Staging Manual (7th Ed., 2010), the various stages of prostate cancer are defined as follows: Tumor: T1: clinically inapparent tumor not palpable or visible by imaging, T1a: tumor incidental histological finding in 5% or less of tissue resected, T1b: tumor incidental histological finding in more than 5% of tissue resected, T1c: tumor identified by needle biopsy; T2: tumor confined within prostate, T2a: tumor involves one half of one lobe or less, T2b: tumor involves more than half of one lobe, but not both lobes, T2c: tumor involves both lobes; T3: tumor extends through the prostatic capsule, T3a: extracapsular extension (unilateral or bilateral), T3b: tumor invades seminal vesicle(s); T4: tumor is fixed or invades adjacent structures other than seminal vesicles (bladder neck, external sphincter, rectum, levator muscles, or pelvic wall). Generally, a clinical T (cT) stage is T1 or T2 and pathologic T (pT) stage is T2 or higher. Node: NO: no regional lymph node metastasis; N1: metastasis in regional lymph nodes. Metastasis: M0: no distant metastasis; M1: distant metastasis present.

[0031] The Gleason Grading system is used to help evaluate the prognosis of men with prostate cancer. Together with other parameters, it is incorporated into a strategy of prostate cancer staging, which predicts prognosis and helps guide therapy. A Gleason “score” or “grade” is given to prostate cancer based upon its microscopic appearance. Tumors with a low Gleason score typically grow slowly enough that they may not pose a significant threat to the patients in their lifetimes. These patients are monitored (“watchful waiting” or “active surveillance”) over time. Cancers with a higher Gleason score are more aggressive and have a worse prognosis, and these patients are generally treated with surgery (e.g., radical prostatectomy) and, in some cases, therapy (e.g., radiation, hormone, ultrasound, chemotherapy). Gleason scores (or sums) comprise grades of the two most common tumor patterns. These patterns are referred to as Gleason patterns 1-5, with pattern 1 being the most well-differentiated. Most have a mixture of patterns. To obtain a Gleason score or grade, the dominant pattern is added to the second most prevalent pattern to obtain a number between 2 and 10. The Gleason Grades include: G1: well differentiated (slight anaplasia) (Gleason 2-4); G2: moderately differentiated (moderate anaplasia) (Gleason 5-6); G3-4: poorly differentiated/undifferentiated (marked anaplasia) (Gleason 7-10).

[0032] Stage groupings: Stage I: T1a N0 M0 G1; Stage II: (T1a N0 M0 G2-4) or (T1b, c, T1, T2, N0 M0 Any G); Stage III: T3 N0 M0 Any G; Stage IV: (T4 N0 M0 Any G) or (Any T N1 M0 Any G) or (Any T Any N M1 Any G).

[0033] The term “upgrading” as used herein refers to an increase in Gleason grade determined from biopsy to Gleason grade determined from radical prostatectomy (RP). For example, upgrading includes a change in Gleason grade from 3+3 or 3+4 on biopsy to 3+4 or greater on RP. “Significant upgrading” or “upgrade2” as used herein, refers to a change in Gleason grade from 3+3 or 3+4 determined from biopsy to 4+3 or greater, or seminal vesical involvement (SVI), or extracapsular involvement (ECE) as determined from RP.

[0034] The term “high grade” as used herein refers to Gleason score of $\geq 3+4$ or $\geq 4+3$ on RP. The term “low grade” as used herein refers to a Gleason score of 3+3 on RP. In a particular embodiment, “high grade” disease refers to Gleason score of at least major pattern 4, minor pattern 5, or tertiary pattern 5.

[0035] The term “upstaging” as used herein refers to an increase in tumor stage from biopsy to tumor stage at RP. For example, upstaging is a change in tumor stage from clinical T1 or T2 stage at biopsy to pathologic T3 stage at RP.

[0036] The term “non organ-confined disease” as used herein refers to having pathologic stage T3 disease at RP. The term “organ-confined” as used herein refers to pathologic stage pT2 at RP.

[0037] The term “adverse pathology” as used herein refers to a high grade disease as defined above, or non organ-confined disease as defined above. In a particular embodiment, “adverse pathology” refers to prostate cancer with a Gleason score of $\geq 3+4$ or $\geq 4+3$ or pathologic stage T3.

[0038] In another embodiment, the term “high-grade or non-organ-confined disease” refers to prostate cancer with a Gleason score of at least major pattern 4, minor pattern 5, or tertiary pattern 5, or pathologic stage T3.

[0039] As used herein, the terms “active surveillance” and “watchful waiting” mean closely monitoring a patient’s condition without giving any treatment until symptoms appear or change. For example, in prostate cancer, watchful waiting is usually used in older men with other medical problems and early-stage disease.

[0040] As used herein, the term “surgery” applies to surgical methods undertaken for removal of cancerous tissue, including pelvic lymphadenectomy, radical prostatectomy, transurethral resection of the prostate (TURP), excision, dissection, and tumor biopsy/removal. The tumor tissue or sections used for gene expression analysis may have been obtained from any of these methods.

[0041] As used herein, the term “biological sample containing cancer cells” refers to a sample comprising tumor material obtained from a cancer patient. The term encompasses

tumor tissue samples, for example, tissue obtained by radical prostatectomy and tissue obtained by biopsy, such as for example, a core biopsy or a fine needle biopsy. The biological sample may be fresh, frozen, or a fixed, wax-embedded tissue sample, such as a formalin-fixed, paraffin-embedded tissue sample. A biological sample also encompasses bodily fluids containing cancer cells, such as blood, plasma, serum, urine, and the like. Additionally, the term “biological sample containing cancer cells” encompasses a sample comprising tumor cells obtained from sites other than the primary tumor, e.g., circulating tumor cells. The term also encompasses cells that are the progeny of the patient’s tumor cells, e.g. cell culture samples derived from primary tumor cells or circulating tumor cells. The term further encompasses samples that may comprise protein or nucleic acid material shed from tumor cells *in vivo*, e.g., bone marrow, blood, plasma, serum, and the like. The term also encompasses samples that have been enriched for tumor cells or otherwise manipulated after their procurement and samples comprising polynucleotides and/or polypeptides that are obtained from a patient’s tumor material.

[0042] Prognostic factors are those variables related to the natural history of cancer that influence the recurrence rates and outcome of patients once they have developed cancer. Clinical parameters that have been associated with a worse prognosis include, for example, increased tumor stage, high PSA level at presentation, and high Gleason grade or pattern. Prognostic factors are frequently used to categorize patients into subgroups with different baseline relapse risks.

[0043] The term “prognosis” is used herein to refer to the likelihood that a cancer patient will have a cancer-attributable death or progression, including recurrence, metastatic spread, and drug resistance, of a neoplastic disease, such as prostate cancer. For example, a “good prognosis” would include long term survival without recurrence and a “bad prognosis” would include cancer recurrence.

[0044] A “positive clinical outcome” can be assessed using any endpoint indicating a benefit to the patient, including, without limitation, (1) inhibition, to some extent, of tumor growth, including slowing down and complete growth arrest; (2) reduction in the number of tumor cells; (3) reduction in tumor size; (4) inhibition (i.e., reduction, slowing down, or complete stopping) of tumor cell infiltration into adjacent peripheral organs and/or tissues; (5) inhibition of metastasis; (6) enhancement of anti-tumor immune response, possibly resulting in regression or rejection of the tumor; (7) relief, to some extent, of one or more symptoms associated with the tumor; (8) increase in the duration of survival following treatment; and/or (9) decreased mortality at a given point of time following treatment. Positive clinical

outcome can also be considered in the context of an individual's outcome relative to an outcome of a population of patients having a comparable clinical diagnosis, and can be assessed using various endpoints such as an increase in the duration of Recurrence-Free Interval (RFI), an increase in survival time (Overall Survival (OS)) or prostate cancer-specific survival time (Prostate Cancer-Specific Survival (PCSS)) in a population, no upstaging or upgrading in tumor stage or Gleason grade between biopsy and radical prostatectomy, presence of 3+3 grade and organ-confined disease at radical prostatectomy, and the like.

[0045] The term "risk classification" means a grouping of subjects by the level of risk (or likelihood) that the subject will experience a particular negative clinical outcome. A subject may be classified into a risk group or classified at a level of risk based on the methods of the present disclosure, e.g. high, medium, or low risk. A "risk group" is a group of subjects or individuals with a similar level of risk for a particular clinical outcome.

[0046] The term "long-term" survival is used herein to refer to survival for a particular time period, e.g., for at least 5 years, or for at least 10 years.

[0047] The term "recurrence" is used herein to refer to local or distant recurrence (i.e., metastasis) of cancer. For example, prostate cancer can recur locally in the tissue next to the prostate or in the seminal vesicles. The cancer may also affect the surrounding lymph nodes in the pelvis or lymph nodes outside this area. Prostate cancer can also spread to tissues next to the prostate, such as pelvic muscles, bones, or other organs. Recurrence can be determined by clinical recurrence detected by, for example, imaging study or biopsy, or biochemical recurrence detected by, for example, sustained follow-up prostate-specific antigen (PSA) levels ≥ 0.4 ng/mL or the initiation of salvage therapy as a result of a rising PSA level.

[0048] The term "clinical recurrence-free interval (cRFI)" is used herein as time from surgery to first clinical recurrence or death due to clinical recurrence of prostate cancer. If follow-up ended without occurrence of clinical recurrence, or other primary cancers or death occurred prior to clinical recurrence, time to cRFI is considered censored; when this occurs, the only information known is that up through the censoring time, clinical recurrence has not occurred in this subject. Biochemical recurrences are ignored for the purposes of calculating cRFI.

[0049] The term "biochemical recurrence-free interval (bRFI)" is used herein to mean the time from surgery to first biochemical recurrence of prostate cancer. If clinical recurrence occurred before biochemical recurrence, follow-up ended without occurrence of bRFI, or other primary cancers or death occurred prior to biochemical recurrence, time to biochemical recurrence is considered censored at the first of these.

[0050] The term “Overall Survival (OS)” is used herein to refer to the time from surgery to death from any cause. If the subject was still alive at the time of last follow-up, survival time is considered censored at the time of last follow-up. Biochemical recurrence and clinical recurrence are ignored for the purposes of calculating OS.

[0051] The term “Prostate Cancer-Specific Survival (PCSS)” is used herein to describe the time from surgery to death from prostate cancer. If the patient did not die of prostate cancer before end of followup, or died due to other causes, PCSS is considered censored at this time. Clinical recurrence and biochemical recurrence are ignored for the purposes of calculating PCSS.

[0052] In practice, the calculation of the time-to-event measures listed above may vary from study to study depending on the definition of events to be considered censored.

[0053] As used herein, the term “expression level” as applied to a gene refers to the normalized level of a gene product, e.g. the normalized value determined for the RNA level of a gene or for the polypeptide level of a gene.

[0054] The term “gene product” or “expression product” are used herein to refer to the RNA (ribonucleic acid) transcription products (transcripts) of the gene, including mRNA, and the polypeptide translation products of such RNA transcripts. A gene product can be, for example, an unspliced RNA, an mRNA, a splice variant mRNA, a microRNA, a fragmented RNA, a polypeptide, a post-translationally modified polypeptide, a splice variant polypeptide, etc.

[0055] The term “RNA transcript” as used herein refers to the RNA transcription products of a gene, including, for example, mRNA, an unspliced RNA, a splice variant mRNA, a microRNA, and a fragmented RNA.

[0056] Unless indicated otherwise, each gene name used herein corresponds to the Official Symbol assigned to the gene and provided by Entrez Gene (URL: www.ncbi.nlm.nih.gov/sites/entrez) as of the filing date of this application.

[0057] The term “microarray” refers to an ordered arrangement of hybridizable array elements, e.g. oligonucleotide or polynucleotide probes, on a substrate.

[0058] The term “polynucleotide” generally refers to any polyribonucleotide or polydeoxribonucleotide, which may be unmodified RNA or DNA or modified RNA or DNA. Thus, for instance, polynucleotides as defined herein include, without limitation, single- and double-stranded DNA, DNA including single- and double-stranded regions, single- and double-stranded RNA, and RNA including single- and double-stranded regions, hybrid molecules comprising DNA and RNA that may be single-stranded or, more typically, double-

stranded or include single- and double-stranded regions. In addition, the term “polynucleotide” as used herein refers to triple-stranded regions comprising RNA or DNA or both RNA and DNA. The strands in such regions may be from the same molecule or from different molecules. The regions may include all of one or more of the molecules, but more typically involve only a region of some of the molecules. One of the molecules of a triple-helical region often is an oligonucleotide. The term “polynucleotide” specifically includes cDNAs. The term includes DNAs (including cDNAs) and RNAs that contain one or more modified bases. Thus, DNAs or RNAs with backbones modified for stability or for other reasons, are “polynucleotides” as that term is intended herein. Moreover, DNAs or RNAs comprising unusual bases, such as inosine, or modified bases, such as tritiated bases, are included within the term “polynucleotides” as defined herein. In general, the term “polynucleotide” embraces all chemically, enzymatically and/or metabolically modified forms of unmodified polynucleotides, as well as the chemical forms of DNA and RNA characteristic of viruses and cells, including simple and complex cells.

[0059] The term “oligonucleotide” refers to a relatively short polynucleotide, including, without limitation, single-stranded deoxyribonucleotides, single- or double-stranded ribonucleotides, RNA/DNA hybrids and double-stranded DNAs. Oligonucleotides, such as single-stranded DNA probe oligonucleotides, are often synthesized by chemical methods, for example using automated oligonucleotide synthesizers that are commercially available. However, oligonucleotides can be made by a variety of other methods, including in vitro recombinant DNA-mediated techniques and by expression of DNAs in cells and organisms.

[0060] The term “Ct” as used herein refers to threshold cycle, the cycle number in quantitative polymerase chain reaction (qPCR) at which the fluorescence generated within a reaction well exceeds the defined threshold, i.e. the point during the reaction at which a sufficient number of amplicons have accumulated to meet the defined threshold.

[0061] The term “Cp” as used herein refers to “crossing point.” The Cp value is calculated by determining the second derivatives of entire qPCR amplification curves and their maximum value. The Cp value represents the cycle at which the increase of fluorescence is highest and where the logarithmic phase of a PCR begins.

[0062] The terms “threshold” or “thresholding” refer to a procedure used to account for non-linear relationships between gene expression measurements and clinical response as well as to further reduce variation in reported patient scores. When thresholding is applied, all measurements below or above a threshold are set to that threshold value. A non-linear relationship between gene expression and outcome could be examined using smoothers or

cubic splines to model gene expression on recurrence free interval using Cox PH regression or on adverse pathology status using logistic regression. D. Cox, Journal of the Royal Statistical Society, Series B 34:187-220 (1972). Variation in reported patient scores could be examined as a function of variability in gene expression at the limit of quantitation and/or detection for a particular gene.

[0063] As used herein, the term “amplicon,” refers to pieces of DNA that have been synthesized using amplification techniques, such as polymerase chain reactions (PCR) and ligase chain reactions.

[0064] “Stringency” of hybridization reactions is readily determinable by one of ordinary skill in the art, and generally is an empirical calculation dependent upon probe length, washing temperature, and salt concentration. In general, longer probes require higher temperatures for proper annealing, while shorter probes need lower temperatures. Hybridization generally depends on the ability of denatured DNA to re-anneal when complementary strands are present in an environment below their melting temperature. The higher the degree of desired homology between the probe and hybridizable sequence, the higher the relative temperature which can be used. As a result, it follows that higher relative temperatures would tend to make the reaction conditions more stringent, while lower temperatures less so. For additional details and explanation of stringency of hybridization reactions, see Ausubel et al., Current Protocols in Molecular Biology (Wiley Interscience Publishers, 1995).

[0065] “Stringent conditions” or “high stringency conditions”, as defined herein, typically: (1) employ low ionic strength and high temperature for washing, for example 0.015 M sodium chloride/0.0015 M sodium citrate/0.1% sodium dodecyl sulfate at 50°C; (2) employ during hybridization a denaturing agent, such as formamide, for example, 50% (v/v) formamide with 0.1% bovine serum albumin/0.1% Ficoll/0.1% polyvinylpyrrolidone/50mM sodium phosphate buffer at pH 6.5 with 750 mM sodium chloride, 75 mM sodium citrate at 42°C; or (3) employ 50% formamide, 5 x SSC (0.75 M NaCl, 0.075 M sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5 x Denhardt's solution, sonicated salmon sperm DNA (50 µg/ml), 0.1% SDS, and 10% dextran sulfate at 42°C, with washes at 42°C in 0.2 x SSC (sodium chloride/sodium citrate) and 50% formamide, followed by a high-stringency wash consisting of 0.1 x SSC containing EDTA at 55°C.

[0066] “Moderately stringent conditions” may be identified as described by Sambrook et al., Molecular Cloning: A Laboratory Manual, New York: Cold Spring Harbor Press, 1989, and include the use of washing solution and hybridization conditions (e.g., temperature, ionic

strength and %SDS) less stringent than those described above. An example of moderately stringent conditions is overnight incubation at 37°C in a solution comprising: 20% formamide, 5 x SSC (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5 x Denhardt's solution, 10% dextran sulfate, and 20 mg/ml denatured sheared salmon sperm DNA, followed by washing the filters in 1 x SSC at about 37-50°C. The skilled artisan will recognize how to adjust the temperature, ionic strength, etc. as necessary to accommodate factors such as probe length and the like.

[0067] The terms “splicing” and “RNA splicing” are used interchangeably and refer to RNA processing that removes introns and joins exons to produce mature mRNA with continuous coding sequence that moves into the cytoplasm of an eukaryotic cell.

[0068] As used herein, the term “TMPRSS fusion” and “TMPRSS2 fusion” are used interchangeably and refer to a fusion of the androgen-driven TMPRSS2 gene with the ERG oncogene, which has been demonstrated to have a significant association with prostate cancer. S. Perner, et al., *Urology* 46(7):754-760 (2007); S.A. Narod, et al., *Br J Cancer* 99(6):847-851 (2008). As used herein, positive TMPRSS fusion status indicates that the TMPRSS fusion is present in a tissue sample, whereas negative TMPRSS fusion status indicates that the TMPRSS fusion is not present in a tissue sample. Experts skilled in the art will recognize that there are numerous ways to determine TMPRSS fusion status, such as real-time, quantitative PCR or high-throughput sequencing. See, e.g., K. Mertz, et al., *Neoplasia* 9(3):200-206 (2007); C. Maher, *Nature* 458(7234):97-101 (2009).

[0069] The terms “correlated” and “associated” are used interchangeably herein to refer to the association between two measurements (or measured entities). The disclosure provides genes or gene subsets, the expression levels of which are associated with clinical outcome. For example, the increased expression level of a gene may be positively correlated (positively associated) with a good or positive clinical outcome. Such a positive correlation may be demonstrated statistically in various ways, e.g. by a cancer recurrence hazard ratio less than one or by a cancer upgrading or upstaging odds ratio of less than one. In another example, the increased expression level of a gene may be negatively correlated (negatively associated) with a good or positive clinical outcome. In that case, for example, the patient may experience a cancer recurrence or upgrading/upstaging of the cancer, and this may be demonstrated statistically in various ways, e.g., a hazard ratio greater than 1 or an odds ratio greater than one. “Correlation” is also used herein to refer to the strength of association between the expression levels of two different genes, such that the expression level of a first gene can be substituted with an expression level of a second gene in a given algorithm if their

expression levels are highly correlated. Such “correlated expression” of two genes that are substitutable in an algorithm are usually gene expression levels that are positively correlated with one another, e.g., if increased expression of a first gene is positively correlated with an outcome (e.g., increased likelihood of good clinical outcome), then the second gene that is co-expressed and exhibits correlated expression with the first gene is also positively correlated with the same outcome.

[0070] The terms “co-express” and “co-expressed”, as used herein, refer to a statistical correlation between the amounts of different transcript sequences across a population of different patients. Pairwise co-expression may be calculated by various methods known in the art, e.g., by calculating Pearson correlation coefficients or Spearman correlation coefficients. Co-expressed gene cliques may also be identified by seeding and stacking the maximal clique enumeration (MCE) described in Example 4 herein. An analysis of co-expression may be calculated using normalized expression data. Genes within the same gene subset are also considered to be co-expressed.

[0071] A “computer-based system” refers to a system of hardware, software, and data storage medium used to analyze information. The minimum hardware of a patient computer-based system comprises a central processing unit (CPU), and hardware for data input, data output (e.g., display), and data storage. An ordinarily skilled artisan can readily appreciate that any currently available computer-based systems and/or components thereof are suitable for use in connection with the methods of the present disclosure. The data storage medium may comprise any manufacture comprising a recording of the present information as described above, or a memory access device that can access such a manufacture.

[0072] To “record” data, programming or other information on a computer readable medium refers to a process for storing information, using any such methods as known in the art. Any convenient data storage structure may be chosen, based on the means used to access the stored information. A variety of data processor programs and formats can be used for storage, e.g. word processing text file, database format, etc.

[0073] A “processor” or “computing means” references any hardware and/or software combination that will perform the functions required of it. For example, a suitable processor may be a programmable digital microprocessor such as available in the form of an electronic controller, mainframe, server or personal computer (desktop or portable). Where the processor is programmable, suitable programming can be communicated from a remote location to the processor, or previously saved in a computer program product (such as a portable or fixed computer readable storage medium, whether magnetic, optical or solid state

device based). For example, a magnetic medium or optical disk may carry the programming, and can be read by a suitable reader communicating with each processor at its corresponding station.

ALGORITHM-BASED METHODS AND GENE SUBSETS

[0074] The present invention provides an algorithm-based molecular diagnostic assay for predicting a clinical outcome for a patient with prostate cancer. The expression level of one or more genes may be used alone or arranged into functional gene subsets to calculate a quantitative score that can be used to predict the likelihood of a clinical outcome. The algorithm-based assay and associated information provided by the practice of the methods of the present invention facilitate optimal treatment decision-making in prostate cancer. For example, such a clinical tool would enable physicians to identify patients who have a low likelihood of having an aggressive cancer and therefore would not need RP, or who have a high likelihood of having an aggressive cancer and therefore would need RP.

[0075] As used herein, a “quantitative score” is an arithmetically or mathematically calculated numerical value for aiding in simplifying or disclosing or informing the analysis of more complex quantitative information, such as the correlation of certain expression levels of the disclosed genes or gene subsets to a likelihood of a clinical outcome of a prostate cancer patient. A quantitative score may be determined by the application of a specific algorithm. The algorithm used to calculate the quantitative score in the methods disclosed herein may group the expression level values of genes. The grouping of genes may be performed at least in part based on knowledge of the relative contribution of the genes according to physiologic functions or component cellular characteristics, such as in the groups discussed herein. A quantitative score may be determined for a gene group (“gene group score”). The formation of groups, in addition, can facilitate the mathematical weighting of the contribution of various expression levels of genes or gene subsets to the quantitative score. The weighting of a gene or gene group representing a physiological process or component cellular characteristic can reflect the contribution of that process or characteristic to the pathology of the cancer and clinical outcome, such as recurrence or upgrading/upstaging of the cancer. The present invention provides a number of algorithms for calculating the quantitative scores, for example, as set forth in Table 4. In an embodiment of the invention, an increase in the quantitative score indicates an increased likelihood of a negative clinical outcome.

[0076] In an embodiment, a quantitative score is a “recurrence score,” which indicates the likelihood of a cancer recurrence, upgrading or upstaging of a cancer, adverse pathology, non-organ-confined disease, high-grade disease, and/or highgrade or non-organ-confined

disease. An increase in the recurrence score may correlate with an increase in the likelihood of cancer recurrence, upgrading or upstaging of a cancer, adverse pathology, non-organ-confined disease, high-grade disease, and/or highgrade or non-organ-confined disease.

[0077] The gene subsets of the present invention include an ECM gene group, migration gene group, androgen gene group, proliferation gene group, epithelia gene group, and stress gene group.

[0078] The gene subsets referred to herein as the “ECM gene group,” “stromal gene group,” and “stromal response gene group” are used interchangeably and include genes that are synthesized predominantly by stromal cells and are involved in stromal response and genes that co-express with the genes of the ECM gene group. “Stromal cells” are referred to herein as connective tissue cells that make up the support structure of biological tissues. Stromal cells include fibroblasts, immune cells, pericytes, endothelial cells, and inflammatory cells. “Stromal response” refers to a desmoplastic response of the host tissues at the site of a primary tumor or invasion. See, e.g., E. Rubin, J. Farber, Pathology, 985-986 (end Ed. 1994). The ECM gene group includes, for example, ASPN, SFRP4, BGN, THBS2, INHBA, COL1A1, COL3A1, COL1A2, SPARC, COL8A1, COL4A1, FN1, FAP, and COL5A2, and co-expressed genes thereof. Exemplary co-expressed genes include the genes and/or gene cliques shown in Table 8.

[0079] The gene subsets referred to herein as the “migration gene group” or “migration regulation gene group” or “cytoskeletal gene group” or “cellular organization gene group” are used interchangeably and include genes and co-expressed genes that are part of a dynamic microfilament network of actin and accessory proteins and that provide intracellular support to cells, generate the physical forces for cell movement and cell division, as well as facilitate intracellular transport of vesicles and cellular organelle. The migration gene group includes, for example, BIN1, IGF1, C7, GSN, DES, TGFB111, TPM2, VCL, FLNC, ITGA7, COL6A1, PPP1R12A, GSTM1, GSTM2, PAGE4, PPAP2B, SRD5A2, PRKCA, IGFBP6, GPM6B, OLFML3, and HLF, and co-expressed genes thereof. Exemplary co-expressed genes and/or gene cliques are provided in Table 9.

[0080] The gene subset referred to herein as the “androgen gene group,” “PSA gene group,” and “PSA regulation gene group” are used interchangeably and include genes that are members of the kallikrein family of serine proteases (e.g. kallikrein 3 [PSA]), and genes that co-express with genes of the androgen gene group. The androgen gene group includes, for example, FAM13C and KLK2, and co-expressed genes thereof. The androgen gene group may further comprise AZGP1 and SRD5A2, and co-expressed genes thereof.

[0081] The gene subsets referred to herein as the “proliferation gene group” and “cell cycle gene group” are used interchangeably and include genes that are involved with cell cycle functions and genes that co-express with genes of the proliferation gene group. “Cell cycle functions” as used herein refers to cell proliferation and cell cycle control, e.g., checkpoint/G1 to S phase transition. The proliferation gene group thus includes, for example, CDC20, TPX2, UBE2T, MYBL2, and CDKN2C, and co-expressed genes thereof.

Exemplary co-expressed genes and/or gene cliques are provided in Table 10.

[0082] The gene subsets referred to herein as the “epithelia gene group” and “basal epithelia gene group” are used interchangeably and include genes that are expressed during the differentiation of a polarized epithelium and that provide intracellular structural integrity to facilitate physical interactions with neighboring epithelial cells, and genes that co-express with genes of the epithelia gene group. The epithelia gene group includes, for example, CYP3A5, KRT15, KRT5, LAMB3, and SDC1 and co-expressed genes thereof.

[0083] The gene subset referred to herein as the “stress gene group,” “stress response gene group,” and “early response gene group” are used interchangeably and includes genes and co-expressed genes that are transcription factors and DNA-binding proteins activated rapidly and transiently in response to cellular stress and other extracellular signals. These factors, in turn, regulate the transcription of a diverse range of genes. The stress gene group includes, for example, DUSP1, EGR1, FOS, JUN, EGR3, GADD45B, and ZFP36, and co-expressed genes thereof. Exemplary co-expressed genes and/or gene cliques are provided in Table 11.

[0084] Expression levels of other genes and their co-expressed genes may be used with one more of the above gene subsets to predict a likelihood of a clinical outcome of a prostate cancer patient. For example, the expression level of one or more genes selected from the 81 genes of Figure 1 or Table 1A or 1B that do not fall within any of the disclosed gene subsets may be used with one or more of the disclosed gene subsets. In an embodiment of the invention, one or more of STAT5B, NFAT5, AZGP1, ANPEP, IGFBP2, SLC22A3, ERG, AR, SRD5A2, GSTM1, and GSTM2 may be used in one or more gene subsets described above to predict a likelihood of a clinical outcome.

[0085] The present invention also provides methods to determine a threshold expression level for a particular gene. A threshold expression level may be calculated for a specific gene. A threshold expression level for a gene may be based on a normalized expression level. In one example, a C_p threshold expression level may be calculated by assessing functional forms using logistic regression or Cox proportional hazards regression.

[0086] The present invention further provides methods to determine genes that co-express with particular genes identified by, e.g., quantitative RT-PCR (qRT-PCR), as validated biomarkers relevant to a particular type of cancer. The co-expressed genes are themselves useful biomarkers. The co-expressed genes may be substituted for the genes with which they co-express. The methods can include identifying gene cliques from microarray data, normalizing the microarray data, computing a pairwise Spearman correlation matrix for the array probes, filtering out significant co-expressed probes across different studies, building a graph, mapping the probe to genes, and generating a gene clique report. An exemplary method for identifying co-expressed genes is described in Example 3 below, and co-expressed genes identified using this method are provided in Tables 8-11. The expression levels of one or more genes of a gene clique may be used to calculate the likelihood that a patient with prostate cancer will experience a positive clinical outcome, such as a reduced likelihood of a cancer recurrence.

[0087] Any one or more combinations of gene groups may be assayed in the method of the present invention. For example, a stromal response gene group may be assayed, alone or in combination, with a cellular organization gene group, a proliferation gene group, and/or an androgen gene group. In addition, any number of genes within each gene group may be assayed.

[0088] In a specific embodiment of the invention, a method for predicting a clinical outcome for a patient with prostate cancer comprises measuring an expression level of at least one gene from a stromal response gene group, or a co-expressed gene thereof, and at least one gene from a cellular organization gene group, or a co-expressed gene thereof. In another embodiment, the expression level of at least two genes from a stromal response gene group, or a co-expressed gene thereof, and at least two genes from a cellular organization gene group, or a co-expressed gene thereof, are measured. In yet another embodiment, the expression levels of at least three genes are measured from each of the stromal response gene group and the cellular organization gene group. In a further embodiment, the expression levels of at least four genes are measured from each of the stromal response gene group and the cellular organization gene group. In another embodiment, the expression levels of at least five genes are measured from each of the stromal response gene group and the cellular organization gene group. In yet a further embodiment, the expression levels of at least six genes are measured from each of the stromal response gene group and the cellular organization gene group.

[0089] In another specific embodiment, the expression level of at least one gene from the stromal response gene group, or a co-expressed gene thereof, may be measured in addition to the expression level of at least one gene from an androgen gene group, or a co-expressed gene thereof. In a particular embodiment, the expression levels of at least three genes, or co-expressed genes thereof, from the stromal response gene group, and the expression level of at least one gene, or co-expressed gene thereof, from the androgen gene group may be measured.

[0090] In a further embodiment, the expression level of at least one gene each from the stromal response gene group, the androgen gene group, and the cellular organization gene group, or co-expressed genes thereof, may be measured. In a particular embodiment, the level of at least three genes from the stromal response gene group, at least one gene from the androgen gene group, and at least three genes from the cellular organization gene group may be measured. In another embodiment, the expression level of at least one gene each from the stromal response gene group, the androgen gene group, and the proliferation gene group, or co-expressed genes thereof, may be measured. In a particular embodiment, the level of at least three genes from the stromal response gene group, at least one gene from the androgen gene group, and at least one gene from the proliferation gene group may be measured. In either of these combinations, at least two genes from the androgen gene group may also be measured. In any of the combinations, at least four genes from the androgen gene group may also be measured.

[0091] In another embodiment, the expression level of at least one gene each from the stromal response gene group, the androgen gene group, the cellular organization gene group, and the proliferation gene group, or co-expressed genes thereof, may be measured. In a particular embodiment, the level of at least three genes from the stromal response gene group, at least three genes from the cellular organization gene group, at least one gene from the proliferation gene group, and at least two genes from the androgen gene group may be measured. In any of the embodiments, at least four genes from the androgen gene group may be measured.

[0092] Additionally, expression levels of one or more genes that do not fall within the gene subsets described herein may be measured with any of the combinations of the gene subsets described herein. Alternatively, any gene that falls within a gene subset may be analyzed separately from the gene subset, or in another gene subset. For example, the expression levels of at least one, at least two, at least three, or at least 4 genes may be measured in addition to the gene subsets described herein. In an embodiment of the invention, the

additional gene(s) are selected from STAT5B, NFAT5, AZGP1, ANPEP, IGFBP2, SLC22A3, ERG, AR, SRD5A2, GSTM1, and GSTM2.

[0093] In a specific embodiment, the method of the invention comprises measuring the expression levels of the specific combinations of genes and gene subsets shown in Table 4. In a further embodiment, gene group score(s) and quantitative score(s) are calculated according to the algorithm(s) shown in Table 4.

[0094] Various technological approaches for determination of expression levels of the disclosed genes are set forth in this specification, including, without limitation, RT-PCR, microarrays, high-throughput sequencing, serial analysis of gene expression (SAGE) and Digital Gene Expression (DGE), which will be discussed in detail below. In particular aspects, the expression level of each gene may be determined in relation to various features of the expression products of the gene including exons, introns, protein epitopes and protein activity.

[0095] The expression product that is assayed can be, for example, RNA or a polypeptide. The expression product may be fragmented. For example, the assay may use primers that are complementary to target sequences of an expression product and could thus measure full transcripts as well as those fragmented expression products containing the target sequence. Further information is provided in Table A.

[0096] The RNA expression product may be assayed directly or by detection of a cDNA product resulting from a PCR-based amplification method, e.g., quantitative reverse transcription polymerase chain reaction (qRT-PCR). (See e.g., U.S. Patent No. 7,587,279). Polypeptide expression product may be assayed using immunohistochemistry (IHC) by proteomics techniques. Further, both RNA and polypeptide expression products may also be assayed using microarrays.

METHODS OF ASSAYING EXPRESSION LEVELS OF A GENE PRODUCT

[0097] Methods of gene expression profiling include methods based on hybridization analysis of polynucleotides, methods based on sequencing of polynucleotides, and proteomics- based methods. Exemplary methods known in the art for the quantification of RNA expression in a sample include northern blotting and in situ hybridization (Parker & Barnes, Methods in Molecular Biology 106:247-283 (1999)); RNase protection assays (Hod, Biotechniques 13:852-854 (1992)); and PCR-based methods, such as reverse transcription PCR (RT-PCR) (Weis et al., Trends in Genetics 8:263-264 (1992)). Antibodies may be employed that can recognize sequence-specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. Representative methods

for sequencing-based gene expression analysis include Serial Analysis of Gene Expression (SAGE), and gene expression analysis by massively parallel signature sequencing (MPSS). Other methods known in the art may be used.

Reverse Transcription PCR (RT-PCR)

[0098] Typically, mRNA is isolated from a test sample. The starting material is typically total RNA isolated from a human tumor, usually from a primary tumor. Optionally, normal tissues from the same patient can be used as an internal control. Such normal tissue can be histologically-appearing normal tissue adjacent to a tumor. mRNA can be extracted from a tissue sample, e.g., from a sample that is fresh, frozen (e.g. fresh frozen), or paraffin-embedded and fixed (e.g. formalin-fixed).

[0099] General methods for mRNA extraction are well known in the art and are disclosed in standard textbooks of molecular biology, including Ausubel et al., *Current Protocols of Molecular Biology*, John Wiley and Sons (1997). Methods for RNA extraction from paraffin embedded tissues are disclosed, for example, in Rupp and Locker, *Lab Invest.* 56:A67 (1987), and De Andrés et al., *BioTechniques* 18:42044 (1995). In particular, RNA isolation can be performed using a purification kit, buffer set and protease from commercial manufacturers, such as Qiagen, according to the manufacturer's instructions. For example, total RNA from cells in culture can be isolated using Qiagen RNeasy mini-columns. Other commercially available RNA isolation kits include MasterPure™ Complete DNA and RNA Purification Kit (EPICENTRE®, Madison, WI), and Paraffin Block RNA Isolation Kit (Ambion, Inc.). Total RNA from tissue samples can be isolated using RNA Stat-60 (Tel-Test). RNA prepared from tumor can be isolated, for example, by cesium chloride density gradient centrifugation.

[00100] The sample containing the RNA is then subjected to reverse transcription to produce cDNA from the RNA template, followed by exponential amplification in a PCR reaction. The two most commonly used reverse transcriptases are avilo myeloblastosis virus reverse transcriptase (AMV-RT) and Moloney murine leukemia virus reverse transcriptase (MMLV-RT). The reverse transcription step is typically primed using specific primers, random hexamers, or oligo-dT primers, depending on the circumstances and the goal of expression profiling. For example, extracted RNA can be reverse-transcribed using a GeneAmp RNA PCR kit (Perkin Elmer, CA, USA), following the manufacturer's instructions. The derived cDNA can then be used as a template in the subsequent PCR reaction.

[00101] PCR-based methods use a thermostable DNA-dependent DNA polymerase, such as a Taq DNA polymerase. For example, TaqMan® PCR typically utilizes the 5'-nuclease activity of Taq or Tth polymerase to hydrolyze a hybridization probe bound to its target amplicon, but any enzyme with equivalent 5' nuclease activity can be used. Two oligonucleotide primers are used to generate an amplicon typical of a PCR reaction product. A third oligonucleotide, or probe, can be designed to facilitate detection of a nucleotide sequence of the amplicon located between the hybridization sites the two PCR primers. The probe can be detectably labeled, e.g., with a reporter dye, and can further be provided with both a fluorescent dye, and a quencher fluorescent dye, as in a Taqman® probe configuration. Where a Taqman® probe is used, during the amplification reaction, the Taq DNA polymerase enzyme cleaves the probe in a template-dependent manner. The resultant probe fragments disassociate in solution, and signal from the released reporter dye is free from the quenching effect of the second fluorophore. One molecule of reporter dye is liberated for each new molecule synthesized, and detection of the unquenched reporter dye provides the basis for quantitative interpretation of the data.

[00102] TaqMan® RT-PCR can be performed using commercially available equipment, such as, for example, high-throughput platforms such as the ABI PRISM 7700 Sequence Detection System® (Perkin-Elmer-Applied Biosystems, Foster City, CA, USA), or Lightcycler (Roche Molecular Biochemicals, Mannheim, Germany). In a preferred embodiment, the procedure is run on a LightCycler® 480 (Roche Diagnostics) real-time PCR system, which is a microwell plate-based cycler platform.

[00103] 5'-Nuclease assay data are commonly initially expressed as a threshold cycle ("C_t"). Fluorescence values are recorded during every cycle and represent the amount of product amplified to that point in the amplification reaction. The threshold cycle (C_t) is generally described as the point when the fluorescent signal is first recorded as statistically significant. Alternatively, data may be expressed as a crossing point ("C_p"). The C_p value is calculated by determining the second derivatives of entire qPCR amplification curves and their maximum value. The C_p value represents the cycle at which the increase of fluorescence is highest and where the logarithmic phase of a PCR begins.

[00104] To minimize errors and the effect of sample-to-sample variation, RT-PCR is usually performed using an internal standard. The ideal internal standard gene (also referred to as a reference gene) is expressed at a quite constant level among cancerous and non-cancerous tissue of the same origin (i.e., a level that is not significantly different among normal and cancerous tissues), and is not significantly affected by the experimental treatment (i.e., does

not exhibit a significant difference in expression level in the relevant tissue as a result of exposure to chemotherapy), and expressed at a quite constant level among the same tissue taken from different patients. For example, reference genes useful in the methods disclosed herein should not exhibit significantly different expression levels in cancerous prostate as compared to normal prostate tissue. Exemplary reference genes used for normalization comprise one or more of the following genes: AAMP, ARF1, ATP5E, CLTC, GPS1, and PGK1. Gene expression measurements can be normalized relative to the mean of one or more (e.g., 2, 3, 4, 5, or more) reference genes. Reference-normalized expression measurements can range from 2 to 15, where a one unit increase generally reflects a 2-fold increase in RNA quantity.

[00105] Real time PCR is compatible both with quantitative competitive PCR, where an internal competitor for each target sequence is used for normalization, and with quantitative comparative PCR using a normalization gene contained within the sample, or a housekeeping gene for RT-PCR. For further details see, e.g. Held et al., *Genome Research* 6:986-994 (1996).

[00106] The steps of a representative protocol for use in the methods of the present disclosure use fixed, paraffin-embedded tissues as the RNA source. For example, mRNA isolation, purification, primer extension and amplification can be performed according to methods available in the art. (see, e.g., Godfrey et al. *J. Molec. Diagnostics* 2: 84-91 (2000); Specht et al., *Am. J. Pathol.* 158: 419-29 (2001)). Briefly, a representative process starts with cutting about 10 μ m thick sections of paraffin-embedded tumor tissue samples. The RNA is then extracted, and protein and DNA depleted from the RNA-containing sample. After analysis of the RNA concentration, RNA is reverse transcribed using gene-specific primers followed by RT-PCR to provide for cDNA amplification products.

Design of Intron-Based PCR Primers and Probes

[00107] PCR primers and probes can be designed based upon exon or intron sequences present in the mRNA transcript of the gene of interest. Primer/probe design can be performed using publicly available software, such as the DNA BLAT software developed by Kent, W.J., *Genome Res.* 12(4):656-64 (2002), or by the BLAST software including its variations.

[00108] Where necessary or desired, repetitive sequences of the target sequence can be masked to mitigate non-specific signals. Exemplary tools to accomplish this include the Repeat Masker program available on-line through the Baylor College of Medicine, which screens DNA sequences against a library of repetitive elements and returns a query sequence

in which the repetitive elements are masked. The masked intron sequences can then be used to design primer and probe sequences using any commercially or otherwise publicly available primer/probe design packages, such as Primer Express (Applied Biosystems); MGB assay-by-design (Applied Biosystems); Primer3 (Steve Rozen and Helen J. Skaletsky (2000) Primer3 on the WWW for general users and for biologist programmers. See S. Rrawetz, S. Misener, *Bioinformatics Methods and Protocols: Methods in Molecular Biology*, pp. 365-386 (Humana Press).

[00109] Other factors that can influence PCR primer design include primer length, melting temperature (T_m), and G/C content, specificity, complementary primer sequences, and 3' end sequence. In general, optimal PCR primers are generally 17-30 bases in length, and contain about 20-80%, such as, for example, about 50-60% G+C bases, and exhibit T_m 's between 50 and 80 °C, e.g. about 50 to 70 °C.

[00110] For further guidelines for PCR primer and probe design see, e.g. Dieffenbach, CW. et al, "General Concepts for PCR Primer Design" in: *PCR Primer, A Laboratory Manual*, Cold Spring Harbor Laboratory Press, New York, 1995, pp. 133-155; Innis and Gelfand, "Optimization of PCRs" in: *PCR Protocols, A Guide to Methods and Applications*, CRC Press, London, 1994, pp. 5-11; and Plasterer, T.N. *Primerselect: Primer and probe design. Methods Mol. Biol.* 70:520-527 (1997), the entire disclosures of which are hereby expressly incorporated by reference.

[00111] Table A provides further information concerning the primer, probe, and amplicon sequences associated with the Examples disclosed herein.

MassARRAY® System

[00112] In MassARRAY-based methods, such as the exemplary method developed by Sequenom, Inc. (San Diego, CA) following the isolation of RNA and reverse transcription, the obtained cDNA is spiked with a synthetic DNA molecule (competitor), which matches the targeted cDNA region in all positions, except a single base, and serves as an internal standard. The cDNA/competitor mixture is PCR amplified and is subjected to a post-PCR shrimp alkaline phosphatase (SAP) enzyme treatment, which results in the dephosphorylation of the remaining nucleotides. After inactivation of the alkaline phosphatase, the PCR products from the competitor and cDNA are subjected to primer extension, which generates distinct mass signals for the competitor- and cDNA-derives PCR products. After purification, these products are dispensed on a chip array, which is pre-loaded with components needed for analysis with matrix- assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF MS) analysis. The cDNA present in the reaction is then quantified by

analyzing the ratios of the peak areas in the mass spectrum generated. For further details see, e.g. Ding and Cantor, *Proc. Natl. Acad. Sci. USA* 100:3059-3064 (2003).

Other PCR-based Methods

[00113] Further PCR-based techniques that can find use in the methods disclosed herein include, for example, BeadArray® technology (Illumina, San Diego, CA; Oliphant et al., *Discovery of Markers for Disease* (Supplement to *Biotechniques*), June 2002; Ferguson et al., *Analytical Chemistry* 72:5618 (2000)); BeadsArray for Detection of Gene Expression® (BADGE), using the commercially available Luminex100 LabMAP® system and multiple color-coded microspheres (Luminex Corp., Austin, TX) in a rapid assay for gene expression (Yang et al., *Genome Res.* 11:1888-1898 (2001)); and high coverage expression profiling (HiCEP) analysis (Fukumura et al., *Nucl. Acids. Res.* 31(16) e94 (2003).

Microarrays

[00114] Expression levels of a gene or microArray of interest can also be assessed using the microarray technique. In this method, polynucleotide sequences of interest (including cDNAs and oligonucleotides) are arrayed on a substrate. The arrayed sequences are then contacted under conditions suitable for specific hybridization with detectably labeled cDNA generated from RNA of a test sample. As in the RT-PCR method, the source of RNA typically is total RNA isolated from a tumor sample, and optionally from normal tissue of the same patient as an internal control or cell lines. RNA can be extracted, for example, from frozen or archived paraffin-embedded and fixed (e.g. formalin-fixed) tissue samples.

[00115] For example, PCR amplified inserts of cDNA clones of a gene to be assayed are applied to a substrate in a dense array. Usually at least 10,000 nucleotide sequences are applied to the substrate. For example, the microarrayed genes, immobilized on the microchip at 10,000 elements each, are suitable for hybridization under stringent conditions. Fluorescently labeled cDNA probes may be generated through incorporation of fluorescent nucleotides by reverse transcription of RNA extracted from tissues of interest. Labeled cDNA probes applied to the chip hybridize with specificity to each spot of DNA on the array. After washing under stringent conditions to remove non-specifically bound probes, the chip is scanned by confocal laser microscopy or by another detection method, such as a CCD camera. Quantitation of hybridization of each arrayed element allows for assessment of corresponding RNA abundance.

[00116] With dual color fluorescence, separately labeled cDNA probes generated from two sources of RNA are hybridized pair wise to the array. The relative abundance of the transcripts from the two sources corresponding to each specified gene is thus determined

simultaneously. The miniaturized scale of the hybridization affords a convenient and rapid evaluation of the expression pattern for large numbers of genes. Such methods have been shown to have the sensitivity required to detect rare transcripts, which are expressed at a few copies per cell, and to reproducibly detect at least approximately two-fold differences in the expression levels (Schena et al., *Proc. Natl. Acad. Sci. USA* 93(2):106-149 (1996)). Microarray analysis can be performed by commercially available equipment, following manufacturer's protocols, such as by using the Affymetrix GenChip® technology, or Incyte's microarray technology.

Serial Analysis of Gene Expression (SAGE)

[00117] Serial analysis of gene expression (SAGE) is a method that allows the simultaneous and quantitative analysis of a large number of gene transcripts, without the need of providing an individual hybridization probe for each transcript. First, a short sequence tag (about 10-14 bp) is generated that contains sufficient information to uniquely identify a transcript, provided that the tag is obtained from a unique position within each transcript. Then, many transcripts are linked together to form long serial molecules, that can be sequenced, revealing the identity of the multiple tags simultaneously. The expression pattern of any population of transcripts can be quantitatively evaluated by determining the abundance of individual tags, and identifying the gene corresponding to each tag. For more details see, e.g. Velculescu et al., *Science* 270:484-487 (1995); and Velculescu et al., *Cell* 88:243-51 (1997).

Gene Expression Analysis by Nucleic Acid Sequencing

[00118] Nucleic acid sequencing technologies are suitable methods for analysis of gene expression. The principle underlying these methods is that the number of times a cDNA sequence is detected in a sample is directly related to the relative expression of the RNA corresponding to that sequence. These methods are sometimes referred to by the term Digital Gene Expression (DGE) to reflect the discrete numeric property of the resulting data. Early methods applying this principle were Serial Analysis of Gene Expression (SAGE) and Massively Parallel Signature Sequencing (MPSS). See, e.g., S. Brenner, et al., *Nature Biotechnology* 18(6):630-634 (2000). More recently, the advent of "next-generation" sequencing technologies has made DGE simpler, higher throughput, and more affordable. As a result, more laboratories are able to utilize DGE to screen the expression of more genes in more individual patient samples than previously possible. See, e.g., J. Marioni, *Genome Research* 18(9):1509-1517 (2008); R. Morin, *Genome Research* 18(4):610-621 (2008); A. Mortazavi, *Nature Methods* 5(7):621-628 (2008); N. Cloonan, *Nature Methods* 5(7):613-619 (2008).

Isolating RNA from Body Fluids

[00119] Methods of isolating RNA for expression analysis from blood, plasma and serum (see, e.g., K. Enders, et al., Clin Chem 48,1647-53 (2002) (and references cited therein) and from urine (see, e.g., R. Boom, et al., J Clin Microbiol. 28, 495-503 (1990) and references cited therein) have been described.

Immunohistochemistry

[00120] Immunohistochemistry methods are also suitable for detecting the expression levels of genes and applied to the method disclosed herein. Antibodies (e.g., monoclonal antibodies) that specifically bind a gene product of a gene of interest can be used in such methods. The antibodies can be detected by direct labeling of the antibodies themselves, for example, with radioactive labels, fluorescent labels, hapten' labels such as, biotin, or an enzyme such as horse radish peroxidase or alkaline phosphatase. Alternatively, unlabeled primary antibody can be used in conjunction with a labeled secondary antibody specific for the primary antibody. Immunohistochemistry protocols and kits are well known in the art and are commercially available.

Proteomics

[00121] The term "proteome" is defined as the totality of the proteins present in a sample (e.g. tissue, organism, or cell culture) at a certain point of time. Proteomics includes, among other things, study of the global changes of protein expression in a sample (also referred to as "expression proteomics"). Proteomics typically includes the following steps: (1) separation of individual proteins in a sample by 2-D gel electrophoresis (2-D PAGE); (2) identification of the individual proteins recovered from the gel, e.g. by mass spectrometry or N- terminal sequencing, and (3) analysis of the data using bioinformatics.

General Description of the mRNA Isolation, Purification and Amplification

[00122] The steps of a representative protocol for profiling gene expression using fixed, paraffin-embedded tissues as the RNA source, including mRNA isolation, purification, primer extension and amplification are provided in various published journal articles. (See, e.g., T.E. Godfrey, et al., J. Molec. Diagnostics 2: 84-91 (2000); K. Specht et al., Am. J. Pathol. 158: 419-29 (2001), M. Cronin, et al., Am J Pathol 164:35-42 (2004)). Briefly, a representative process starts with cutting a tissue sample section (e.g. about 10 μ m thick sections of a paraffin-embedded tumor tissue sample). The RNA is then extracted, and protein and DNA are removed. After analysis of the RNA concentration, RNA repair is performed if desired. The sample can then be subjected to analysis, e.g., by reverse transcribed using gene specific promoters followed by RT-PCR.

STATISTICAL ANALYSIS OF EXPRESSION LEVELS IN IDENTIFICATION OF GENES

[00123] One skilled in the art will recognize that there are many statistical methods that may be used to determine whether there is a significant relationship between a clinical outcome of interest (e.g., recurrence) and expression levels of a marker gene as described here. In an exemplary embodiment, the present invention includes three studies. The first study is a stratified cohort sampling design (a form of case-control sampling) using tissue and data from prostate cancer patients. Selection of specimens was stratified by clinical T-stage (T1, T2), year of surgery (<1993 , ≥ 1993), and prostatectomy Gleason Score (low/intermediate, high). All patients with clinical recurrence were selected and a stratified random sample of patients who did not experience a clinical recurrence was selected. For each patient, up to two enriched tumor specimens and one normal-appearing tissue sample were assayed. The second study used a subset of 70 patients from the first study from whom matched prostate biopsy tumor tissue was assayed. The third study includes all patients (170 evaluable patients) who had surgery for their prostate cancer between 1999 and 2010 at the Cleveland Clinic (CC) and had Low or Intermediate risk (by AUA) clinically localized prostate cancer who might have been reasonable candidates for active surveillance but who underwent RP at CC within 6 months of the diagnosis of prostate cancer by biopsy. Biopsy tumor tissue from these patients was assayed.

[00124] All hypothesis tests were reported using two-sided p-values. To investigate if there is a significant relationship of outcomes (eg. clinical recurrence-free interval (cRFI), biochemical recurrence-free interval (bRFI), prostate cancer-specific survival (PCSS), overall survival (OS)) with individual genes, and demographic or clinical covariates), Cox Proportional Hazards (PH) models using maximum weighted pseudo partial-likelihood estimators were used and p-values from Wald tests of the null hypothesis that the hazard ratio (HR) is one are reported. To investigate if there is a significant relationship between individual genes and Gleason pattern of a particular sample, ordinal logistic regression models using maximum weighted pseudolikelihood methods were used and p-values from Wald tests of the null hypothesis that the odds ratio (OR) is one are reported. To investigate if there is a significant relationship between individual genes and upgrading and/or upstaging or adverse pathology at RP, logistic regression models using maximum weighted pseudolikelihood methods were used and p-values from Wald tests of the null hypothesis that the odds ratio (OR) is one are reported.

COEXPRESSION ANALYSIS

[00125] In an exemplary embodiment, the joint correlation of gene expression levels among prostate cancer specimens under study may be assessed. For this purpose, the correlation structures among genes and specimens may be examined through hierarchical cluster methods. This information may be used to confirm that genes that are known to be highly correlated in prostate cancer specimens cluster together as expected. Only genes exhibiting a nominally significant (unadjusted $p < 0.05$) relationship with cRFI in the univariate Cox PH regression analysis are included in these analyses.

[00126] One skilled in the art will recognize that many co-expression analysis methods now known or later developed will fall within the scope and spirit of the present invention. These methods may incorporate, for example, correlation coefficients, co-expression network analysis, clique analysis, etc., and may be based on expression data from RT-PCR, microarrays, sequencing, and other similar technologies. For example, gene expression clusters can be identified using pair-wise analysis of correlation based on Pearson or Spearman correlation coefficients. (See, e.g., Pearson K. and Lee A., *Biometrika* 2, 357 (1902); C. Spearman, *Amer. J. Psychol* 15:72-101 (1904); J. Myers, A. Well, *Research Design and Statistical Analysis*, p. 508 (2nd Ed., 2003).) An exemplary method for identifying co-expressed genes is described in Example 3 below.

NORMALIZATION OF EXPRESSION LEVELS

[00127] The expression data used in the methods disclosed herein can be normalized. Normalization refers to a process to correct for (normalize away), for example, differences in the amount of RNA assayed and variability in the quality of the RNA used, to remove unwanted sources of systematic variation in Ct or Cp measurements, and the like. With respect to RT-PCR experiments involving archived fixed paraffin embedded tissue samples, sources of systematic variation are known to include the degree of RNA degradation relative to the age of the patient sample and the type of fixative used to store the sample. Other sources of systematic variation are attributable to laboratory processing conditions.

[00128] Assays can provide for normalization by incorporating the expression of certain normalizing genes, which do not significantly differ in expression levels under the relevant conditions. Exemplary normalization genes disclosed herein include housekeeping genes. (See, e.g., E. Eisenberg, et al., *Trends in Genetics* 19(7):362-365 (2003).) Normalization can be based on the mean or median signal (Ct or Cp) of all of the assayed genes or a large subset thereof (global normalization approach). In general, the normalizing genes, also referred to as reference genes, are typically genes that are known not to exhibit meaningfully different

expression in prostate cancer as compared to non-cancerous prostate tissue, and track with various sample and process conditions, thus provide for normalizing away extraneous effects.

[00129] In exemplary embodiments, one or more of the following genes are used as references by which the mRNA expression data is normalized: AAMP, ARF1, ATP5E, CLTC, GPS1, and PGK1. The calibrated weighted average C_T or C_p measurements for each of the prognostic and predictive genes may be normalized relative to the mean of five or more reference genes.

[00130] Those skilled in the art will recognize that normalization may be achieved in numerous ways, and the techniques described above are intended only to be exemplary, not exhaustive.

STANDARDIZATION OF EXPRESSION LEVELS

[00131] The expression data used in the methods disclosed herein can be standardized. Standardization refers to a process to effectively put all the genes on a comparable scale. This is performed because some genes will exhibit more variation (a broader range of expression) than others. Standardization is performed by dividing each expression value by its standard deviation across all samples for that gene. Hazard ratios are then interpreted as the proportional change in the hazard for the clinical endpoint (clinical recurrence, biological recurrence, death due to prostate cancer, or death due to any cause) per 1 standard deviation increase in expression.

KITS OF THE INVENTION

[00132] The materials for use in the methods of the present invention are suited for preparation of kits produced in accordance with well-known procedures. The present disclosure thus provides kits comprising agents, which may include gene-specific or gene-selective probes and/or primers, for quantifying the expression of the disclosed genes for predicting prognostic outcome or response to treatment. Such kits may optionally contain reagents for the extraction of RNA from tumor samples, in particular fixed paraffin-embedded tissue samples and/or reagents for RNA amplification. In addition, the kits may optionally comprise the reagent(s) with an identifying description or label or instructions relating to their use in the methods of the present invention. The kits may comprise containers (including microliter plates suitable for use in an automated implementation of the method), each with one or more of the various materials or reagents (typically in concentrated form) utilized in the methods, including, for example, chromatographic columns, pre-fabricated microarrays, buffers, the appropriate nucleotide triphosphates (e.g., dATP, dCTP, dGTP and dTTP; or rATP, rCTP, rGTP and UTP), reverse transcriptase, DNA polymerase, RNA

polymerase, and one or more probes and primers of the present invention (e.g., appropriate length poly(T) or random primers linked to a promoter reactive with the RNA polymerase). Mathematical algorithms used to estimate or quantify prognostic or predictive information are also properly potential components of kits.

REPORTS

[00133] The methods of this invention, when practiced for commercial diagnostic purposes, generally produce a report or summary of information obtained from the herein-described methods. For example, a report may include information concerning expression levels of one or more genes, classification of the tumor or the patient's risk of recurrence, the patient's likely prognosis or risk classification, clinical and pathologic factors, and/or other information. The methods and reports of this invention can further include storing the report in a database. The method can create a record in a database for the subject and populate the record with data. The report may be a paper report, an auditory report, or an electronic record. The report may be displayed and/or stored on a computing device (e.g., handheld device, desktop computer, smart device, website, etc.). It is contemplated that the report is provided to a physician and/or the patient. The receiving of the report can further include establishing a network connection to a server computer that includes the data and report and requesting the data and report from the server computer.

COMPUTER PROGRAM

[00134] The values from the assays described above, such as expression data, can be calculated and stored manually. Alternatively, the above-described steps can be completely or partially performed by a computer program product. The present invention thus provides a computer program product including a computer readable storage medium having a computer program stored on it. The program can, when read by a computer, execute relevant calculations based on values obtained from analysis of one or more biological samples from an individual (e.g., gene expression levels, normalization, standardization, thresholding, and conversion of values from assays to a score and/or text or graphical depiction of tumor stage and related information). The computer program product has stored therein a computer program for performing the calculation.

[00135] The present disclosure provides systems for executing the program described above, which system generally includes: a) a central computing environment; b) an input device, operatively connected to the computing environment, to receive patient data, wherein the patient data can include, for example, expression level or other value obtained from an assay using a biological sample from the patient, or microarray data, as described in detail above;

c) an output device, connected to the computing environment, to provide information to a user (e.g., medical personnel); and d) an algorithm executed by the central computing environment (e.g., a processor), where the algorithm is executed based on the data received by the input device, and wherein the algorithm calculates an expression score, thresholding, or other functions described herein. The methods provided by the present invention may also be automated in whole or in part.

[00136] Having described the invention, the same will be more readily understood through reference to the following Examples, which are provided by way of illustration, and are not intended to limit the invention in any way.

EXAMPLES

EXAMPLE 1: SELECTION OF 81 GENES FOR ALGORITHM DEVELOPMENT

[00137] A gene identification study to identify genes associated with clinical recurrence, biochemical recurrence and/or death from prostate cancer is described in U.S. Provisional Application Nos. 61/368,217, filed July 27, 2010; 61/414,310, filed November 16, 2010; and 61/485,536, filed May 12, 2011, and in U.S. Pub. No. 20120028264, filed July 25, 2011, and published February 2, 2012 (all of which are hereby incorporated by reference). RT-PCR analysis was used to determine RNA expression levels for 732 genes and reference genes in prostate cancer tissue and surrounding normal appearing tissue (NAT) in patients with early-stage prostate cancer treated with radical prostatectomy. Genes significantly associated ($p < 0.05$) with clinical recurrence-free interval (cRFI), biochemical recurrence-free interval (bRFI), prostate cancer-specific survival (PCSS), and upgrading/upstaging were determined.

[00138] From the genes that were identified as being associated with outcome, 81 genes were selected for subsequent algorithm development. The primers, probes, and amplicon sequences of the 81 genes (and 5 reference genes) are listed in Table A. The genes selected were among the most prognostic with respect to cRFI and other properties and shown in Tables 1A-1B. Other properties considered were: 1) Strongest genes with respect to the regression to the mean corrected standardized hazard ratio for the association of gene expression and cRFI in the primary Gleason pattern tumor; 2) Consistency in association (hazard ratio) with cRFI using the highest Gleason pattern tumor; 3) Associated with prostate-cancer specific survival (PCSS); 4) Strong hazard ratio after adjustment for The University of San Francisco Cancer of the Prostate Risk Assessment (CAPRA) (Cooperberg et al., J. Urol. 173:1983-1942, 2005); 5) Statistically significant odds ratio for the association between gene expression and surgical Gleason pattern of the tumor; 6) Large overall

variability with greater between-patient variability than within-patient variability preferable; and 7) Highly expressed.

[00139] The true discovery rate degree of association (TDRDA) method (Crager, *Stat Med*, 2010 Jan 15;29(1):33-45.) was used in the analysis of gene expression and cRFI and results are shown in Table 1A. The true discovery rate is the counterpart to the false discovery rate. Univariate Cox PH regression models were fit and the TDRDA method was used to correct estimated standardized hazard ratios for regression to the mean (RM) and assess false discovery rates for identification of genes with absolute standardized hazard ratio of at least a specified level. The false discovery rates were controlled at 10%. The TDRDA method identifies sets of genes among which a specified proportion are expected to have an absolute association (here, the absolute standardized hazard ratio) of a specified degree or more. This leads to a gene ranking method that uses the maximum lower bound (MLB) degree of association for which each gene belongs to a TDRDA set. Estimates of each gene's actual degree of association with approximate correction for "selection bias" due to regression to the mean can be derived using simple bivariate normal theory and Efron and Tibshirani's empirical Bayes approach. Efron, *Annals of Applied Statistics* 2:197-223 (2008); Efron and Tibshirani. *Genetic Epidemiology* 23: 70-86. Table 1A shows the RM-corrected estimate of the standardized hazard ratio and the MLB for each gene using either the primary Gleason pattern (PGP) or highest Gleason pattern (HGP) sample gene expression. Genes marked with a direction of association of -1 are associated with a reduced likelihood of clinical recurrence, while those marked with a direction of association of 1 are associated with an increased likelihood of clinical recurrence.

[00140] Within patient and between patient variance components were estimated using a mixed model treating the patient effect as random. The overall mean and standard deviation of normalized gene expression as well as within- and between-patient components of variance are shown in Table 1A.

[00141] Univariate Cox PH regression models using maximum weighted partial pseudolikelihood estimation were used to estimate the association between gene expression and prostate cancer specific-survival (PCSS). The standardized hazard ratio (HR), p-value and q-value using Storey's FDR method are reported in Table 1B. Storey, *Journal of the Royal Statistical Society, Series B* 64:479-498 (2002). The q-value can be interpreted as the empirical Bayes posterior probability given the data that the gene identified is a false discovery, that is, the probability that it has no association with clinical recurrence.

[00142] Univariate ordinal logistic regression models were used to estimate the association between gene expression and the Gleason pattern of the primary Gleason pattern tumor (3, 4, 5). The standardized odds ratio (OR), p-value and q-value using Storey's FDR method are reported in Table 1B.

[00143] Figure 1 shows an example of a dendrogram depicting the association of the 81 genes. The y-axis corresponds to the average distance between clusters measured as 1-Pearson r. The smaller the number (distance measure), the more highly correlated the genes. The amalgamation method is weighted pair-group average. Genes that were co-expressed were identified from the dendrogram and are grouped into gene groups. Based on Figure 1, the genes from the Gene Identification study were formed into the following gene groups or subsets:

[00144] Cellular organization gene group (BIN1; IGF1; C7; GSN; DES; TGFB111; TPM2; VCL; FLNC; ITGA7; COL6A1; PPP1R12A; GSTM1; GSTM2; PAGE4; PPAP2B; SRD5A2; PRKCA; IGFBP6; GPM6B; OLFML3; HLF)

[00145] Basal epithelia gene group (CYP3A5; KRT15; KRT5; LAMB3; SDC1)

[00146] Stress response gene group (DUSP1; EGR1; FOS; JUN; EGR3; GADD45B; ZFP36)

[00147] Androgen gene group (FAM13C; KLK2; AZGP1; SRD5A2)

[00148] Stromal gene group (ASPN; SFRP4; BGN; THBS2; INHBA; COL1A1; COL3A1; COL1A2; SPARC; COL8A1; COL4A1; FN1; FAP; COL5A2)

[00149] Proliferation gene group (CDC20; TPX2; UBE2T; MYBL2; CDKN2C)

Table 1A.

GENE	Mean normalized cp	SD normalized CP	Between- patient variance	Within- patient variance	Total variance	Association with cR in PGP sample			Association with cR in HGP sample		
						Direction of Association	Absolute RM Corrected HR	MLB	Direction of Association	Absolute RM Corrected HR	MLB
ARF1	11.656805	0.2475574	0.04408	0.01646	0.0613399	-1	1.0849132		-1	1.2008335	1.0397705
ATP5E	10.896515	0.2667133	0.05141	0.01979	0.0711992	1	1.2599111	1.0908967	1	1.3877428	1.165325
CLTC	10.597008	0.1780634	0.01554	0.01619	0.0317257	1	1.0896793	1.002002	1	1.2293343	1.0554846
GPS1	9.2927019	0.2169116	0.03181	0.01578	0.0470897	1	1.0064191		-1	1.1089053	
PGK1	8.3680642	0.2655009	0.04957	0.02099	0.0705517	1	1.1174157	1.0151131	1	1.2149168	1.060752
ASPN	5.4846081	1.1701993	0.4981	0.8719	1.3699944	1	1.7716283	1.4276075	1	1.7114564	1.4007391
BGN	11.299746	0.7357491	0.3259	0.2159	0.5417276	1	1.6133066	1.3771057	1	1.7312777	1.4007391
COL1A1	11.325411	0.8840402	0.4748	0.3073	0.7821127	1	1.6162028	1.3284329	1	1.7982985	1.4304656
COL1A2	10.093055	0.8232037	0.4321	0.2461	0.6781941	1	1.1379748	1.0222438	1	1.3449362	1.138553
COL3A1	11.007109	0.7944239	0.352	0.2795	0.6315424	1	1.5695255	1.2969301	1	1.7133767	1.4007391
COL4A1	7.8408647	0.6731713	0.2993	0.2142	0.4534598	1	1.3397169	1.14225	1	1.4292283	1.1996142
COL5A2	5.2708574	0.9571692	0.4	0.5166	0.9166661	1	1.1715343	1.0408108	1	1.1827568	1.0408108
F2R	7.0775127	1.0110657	0.5529	0.47	1.022934	1	1.5019815	1.2636445	1	1.4888813	1.2361479
FAP	5.0493366	1.1898915	0.6577	0.759	1.4166546	1	1.3007869	1.1162781	1	1.3882726	1.1641602
FN1	9.5176438	0.7224014	0.3059	0.2163	0.5222401	1	1.0505668		1	1.1524617	1.0253151
INHBA	5.8059993	1.2653019	0.9629	0.6392	1.6021763	1	1.896185	1.4658693	1	2.1859455	1.7177237
SFRP4	7.8229007	1.2053184	0.7997	0.6541	1.4537934	1	1.5382715	1.2763443	1	1.5693525	1.2989301
SPARC	10.544556	0.7978856	0.4311	0.206	0.6371517	1	1.3683799	1.1711662	1	1.6187451	1.3324242
TM62	4.7779897	1.0825934	0.7111	0.4608	1.172864	1	1.5523887	1.2904616	1	1.6829249	1.2785056
BIN1	7.8741434	0.840604	0.4445	0.2627	0.7071728	-1	1.5631385	1.2930451	-1	1.3294226	1.1185129
C7	8.4896479	1.1063704	0.6917	0.3377	1.2793358	-1	1.5658393	1.2687092	-1	1.4724865	1.220382
COL6A1	7.3615421	0.848837	0.3381	0.3828	0.7209474	-1	1.5439152	1.2687092	-1	1.2634411	1.0745553
DES	11.967287	0.896286	0.4101	0.3938	0.8038418	-1	1.9087183	1.2386227	-1	1.4032825	1.0961648

GENE	Mean normalized cp	SD normalized Cp	Between- patient variance	Within- patient variance	Total variance	Association with cR in PGP sample			Association with cR in HGP sample		
						Direction of Association	Absolute RM Corrected HR	MLB	Direction of Association	Absolute RM Corrected HR	MLB
FLNC	8.6795128	1.0679528	0.572	0.5692	1.1412391	-1	1.2696693	1.0650268	-1	1.2353942	1.0491707
GPMB8	7.9403089	0.9453416	0.4441	0.4501	0.8942266	-1	1.4471085	1.3058273	-1	1.4931412	1.2386227
GSN	8.8308175	0.7789199	0.3756	0.2316	0.6071864	-1	1.6223835	1.3073471	-1	1.3639212	1.136553
GSTM1	6.4175396	1.2720365	1.0519	0.5675	1.6193374	-1	1.5226009	1.2568853	-1	1.5822193	1.2969301
GSTM2	7.2950478	1.0014875	0.5069	0.4967	1.0036507	-1	1.6694102	1.3284329	-1	1.4815895	1.220182
HLF	5.0774106	0.9618962	0.4516	0.4741	0.9257242	-1	1.6223351	1.2958338	-1	1.5817618	1.2831718
IGF1	7.6180418	1.1441945	0.7925	0.5177	1.3101729	-1	1.4737764	1.2165269	-1	1.6861196	1.359341
IGFBP6	7.0089783	1.0818262	0.6302	0.5405	1.1707038	-1	1.5534588	1.257342	-1	1.1860594	1.0273678
ITGA7	7.3299653	0.8913845	0.4034	0.3916	0.7950725	-1	1.5556326	1.2636445	-1	1.3587711	1.1308844
ORFMLE3	8.1932023	0.8189012	0.4	0.7711	0.6710997	-1	1.5254982	1.2649088	-1	1.3364894	1.1363694
PAGE4	7.406255	1.4889881	1.3023	0.9164	2.2187195	-1	1.6316984	1.2969301	-1	1.5178657	1.2435871
PPAP2B	8.8879191	0.7884847	0.3841	0.238	0.6221573	-1	1.5664582	1.2648088	-1	1.4703629	1.2088335
PPP1R12A	9.369152	0.5056735	0.1361	0.1198	0.2558759	-1	1.4273047	1.1711662	-1	1.3719705	1.1297541
PRKCA	7.4654299	0.7936779	0.2884	0.3319	0.6302916	1	1.4490244	1.3360207	-1	1.2221961	1.0544846
SRD5A2	5.7878904	1.2691925	0.8776	0.7343	1.6119317	-1	1.8236528	1.4276075	-1	1.723879	1.4007391
VCL	8.9766979	0.720767	0.2667	0.3524	0.5191126	-1	1.526093	1.3421441	-1	1.4080433	1.1525766
TGFBI1	8.2191469	0.7816114	0.3143	0.297	0.6113098	-1	1.4793989	1.2104595	-1	1.3608249	1.1229959
TPM2	11.83198	0.8626062	0.4138	0.3322	0.7445062	1	1.5738312	1.2687092	1	1.4785656	1.2116785
TPX2	4.552336	1.0856094	0.4888	0.6903	1.1791549	1	1.6454619	1.3444702	1	1.886703	1.4879005
CDC20	3.5608774	0.7971167	0.1978	0.4378	0.6358436	1	1.431589	1.231445	1	1.568584	1.3271052
CDKN2C	3.5214932	0.6898842	0.137	0.3391	0.476113	1	1.4751885	1.257342	1	1.6210275	1.3485096
MYBL2	3.7683784	0.961084	0.4061	0.5569	0.9629897	1	1.543274	1.2969301	1	1.5563089	1.3086551
UBE2T	3.5015369	0.7220453	0.1927	0.3289	0.5215967	1	1.5156185	1.272521	1	1.2186754	1.0618365
CYP3A5	4.554983	1.3465744	0.5786	1.3085	1.7871462	-1	1.861987	1.6176075	-1	1.8689598	1.4304656
KRT15	8.1899409	1.8188542	1.3339	1.956	3.3098961	-1	1.4985779	1.2423441	-1	1.7380356	1.4007391
KRT5	7.046586	1.528426	0.8988	1.4983	2.3371203	-1	1.4658131	1.2149963	-1	1.6610573	1.3702593

GENE	Mean normalized cp	SD normalized CP	Between-patient variance	Within-patient variance	Total variance	Association with cR in PGP sample			Association with cR in HGP sample		
						Direction of Association	Absolute RM Corrected HR	MLB	Direction of Association	Absolute RM Corrected HR	MLB
LAMB3	6.3566958	1.3451305	0.6358	1.1744	1.8101739	-1	1.435434	1.1996142	-1	1.5003356	1.2287532
EGR1	12.425851	1.0521413	0.7737	0.3243	1.1079529	-1	1.5840317	1.2687092	-1	1.5773643	1.2801751
FOS	12.383619	1.1226481	0.8969	0.3746	1.2614295	-1	1.580311	1.2687092	-1	1.655218	1.3310925
GADD45B	9.8760029	1.3001535	0.893	0.5485	1.4414667	-1	1.5331503	1.2460767	-1	1.490302	1.220132
JUN	11.184249	1.027684	0.8099	0.2473	1.0571304	-1	1.5816073	1.2649088	-1	1.5652799	1.2649088
ZFP36	12.472841	1.1025855	0.8095	0.4072	1.2167102	-1	1.5189178	1.2423441	-1	1.580749	1.2840254
DUSP1	11.51936	0.8297195	0.439	0.25	0.6889744	-1	1.3986201	1.1502738	-1	1.40278	1.1525766
EGR3	9.7545461	1.3366451	1.2104	0.5778	1.7881414	-1	1.5114362	1.2349124	-1	1.4927024	1.226348
FAM13C	7.4923611	0.9318455	0.58	0.2891	0.8690619	-1	1.678403	1.3716303	-1	1.7981441	1.4520843
KLF2	14.718412	0.6677811	0.3087	0.1376	0.4463134	-1	1.4443314	1.1936311	-1	1.4966995	1.2361479
AIDH1A2	5.8436909	1.0515294	0.4984	0.6079	1.1063379	-1	1.5394232	1.2649088	-1	1.817606	1.4304656
A2GFP1	9.3508493	1.4900096	1.6397	0.5625	2.2223058	-1	1.6322418	1.329762	-1	1.4723607	1.2388619
ANPEP	7.1080338	2.3433136	3.865	1.6309	5.4959682	-1	1.5469141	1.257342	-1	1.7787926	1.4035433
AR	8.3681098	0.9094853	0.1623	0.09684	0.2597757	-1	1.0683368		-1	1.1146402	1.004008
BMP6	4.7641658	1.1079887	0.5639	0.6645	1.2283377	-1	1.5010706	1.2649088	-1	1.7140049	1.4007391
CD276	8.9879201	0.8574321	0.168	0.1429	0.3108408	-1	1.453727	1.2386629	-1	1.6372299	1.3525612
CD44	7.5506865	1.1157404	0.8611	0.3849	1.2459543	-1	1.6840393	1.3444702	-1	1.5715077	1.3008267
COL8A1	7.1807327	0.8924731	0.329	0.4566	0.9856651	-1	1.4787026	1.251071	-1	1.6302371	1.4337573
CSF1	5.604685	0.806399	0.2878	0.3678	0.6506271	-1	1.4727627	1.2092496	-1	1.3423853	1.1162781
SRC	7.4585136	0.6881318	0.2723	0.1745	0.446735	-1	1.4782469	1.2435871	-1	1.5104712	1.2687092
CSRP1	12.967068	0.8434745	0.267	0.4448	0.711785	-1	1.2323596	1.0607752	-1	1.2456215	1.0639623
DPP4	6.5096496	1.228945	0.9313	0.5802	1.5114514	-1	1.5818046	1.2930451	-1	1.538425	1.2789396
TNFRSF10B	5.8731042	0.836448	0.3338	0.3663	0.7000558	-1	1.5759506	1.2687092	-1	1.4449471	1.2032184
ERG	7.2194906	2.3910775	4.1237	1.0805	5.2541704	-1	1.0907212	1.0030045	-1	1.0538367	
FAM107A	4.9863267	1.128037	0.6041	0.5691	1.2732255	-1	1.7326232	1.3539145	-1	1.5773305	1.2801791
IGFBP2	9.855496	0.7862553	0.4734	0.1454	0.6187898	-1	1.6388294	1.2982277	-1	1.4976178	1.2287532

GENE	Mean normalized cp	SD normalized CP	Between-patient variance	Within-patient variance	Total variance	Association with cR in PGP sample			Association with cR in HGP sample		
						Direction of Association	Absolute RIM Corrected HR	MLB	Direction of Association	Absolute RIM Corrected HR	MLB
CADM1	7.5914375	0.7603875	0.3308	0.2478	0.5785959	-1	1.6923754	1.3539145	-1	1.7428282	1.4035433
IL6ST	10.355178	0.5503714	0.1851	0.1187	0.3038017	-1	1.5709544	1.2687082	-1	1.470333	1.2277744
LGALS3	8.5926121	0.7862707	0.3442	0.2743	0.6185665	-1	1.5048215	1.2411024	-1	1.3574436	1.1320159
SMAD4	8.902039	0.4085988	0.1825	0.08456	0.1670791	-1	1.683624	1.325762	-1	1.6100555	1.3112751
NFAT5	9.2271297	0.5003181	0.1647	0.08587	0.2505217	-1	1.7299501	1.3539145	-1	1.5858292	1.2995265
SDCL	7.2405046	0.9438894	0.4431	0.4573	0.9004048	1	1.1870437	1.0502204	1	1.0140388	
SHMT2	7.3818144	0.5716985	0.1376	0.1894	0.3270065	1	1.5185392	1.2687092	1	1.494364	1.2448313
SIC22A3	8.8366385	1.3865065	1.3178	0.6112	1.9240432	-1	1.6100215	1.2930951	-1	1.6531341	1.3271052
STAT5B	7.443638	0.479107	0.09118	0.1385	0.2296576	-1	1.4932136	1.2435871	-1	1.4376605	1.1948253
MMMP11	4.0874635	1.1790667	0.6512	0.7396	1.3908598	1	1.4849754	1.257342	1	1.3586068	1.1514246
TUBB2A	8.3247821	0.9300317	0.553	0.3126	0.8656511	-1	1.4310473	1.1699956	-1	1.4520775	1.1817543

Table 1B.

GENE	Association with PCSS Endpoint			Association with cRFI, CAPRA Adjusted			Association with primary Gleason pattern		
	Std. HR	Wald p-value	Storey q-value	Std. HR	Wald p-value	Storey q-value	Std. OR	Wald p-value	Storey q-value
ARF1	0.9976193	0.9892903	0.5842039	1.13407667	0.83255536	0.4298654	0.982273	0.907845	0.3887901
ATP5E	1.7881176	0.0115817	0.0404889	2.78746082	0.07918786	0.0876892	1.3546007	0.0347604	0.0377781
CLTC	1.025701	0.6673167	0.5533326	1.95459035	0.43611036	0.2800117	3.293925	0.4450274	0.2418238
GPS1	0.849428	0.3590472	0.3461617	1.04907052	0.93720193	0.4574281	0.8771353	0.1863038	0.1358186
PGX1	0.9801478	0.9057919	0.5657924	2.40228939	0.08542284	0.0920345	0.9775945	0.8343649	0.3866666
ASPN	3.0547166	1.85E-08	3.97E-06	1.98680704	3.75E-07	8.71E-06	2.6178187	3.32E-07	2.89E-06
8GN	2.6395973	2.34E-06	0.000144	2.65751131	3.29E-07	8.71E-06	2.5773215	1.19E-07	1.34E-06
COL1A1	2.5740243	1.16E-07	0.0000124	2.45783157	3.46E-09	3.73E-07	2.2304742	2.40E-06	0.0000138
COL1A2	1.6067045	0.0108776	0.0393063	1.30233373	0.14922155	0.1339808	1.1226733	0.4115421	0.2324622

GENE	Association with PCS5 Endpoint			Association with cRF1, CAPRA Adjusted			Association with primary Gleason pattern		
	Std. HR	Wald p-value	Storey q-value	Std. HR	Wald p-value	Storey q-value	Std. OR	Wald p-value	Storey q-value
COL3A1	2.3815758	7.22E-06	0.0002217	2.52327548	6.28E-08	3.29E-06	1.9034844	0.0001218	0.0003527
COL4A1	1.9704368	0.0008215	0.006465	2.14028481	0.00036078	0.0014267	1.0893045	0.5143164	0.2634536
COL5A2	1.9382474	0.0018079	0.0111059	1.20272948	0.23650362	0.1864588	1.1234816	0.4230095	0.2350912
F2R	2.1687429	0.000107	0.00016437	1.62198106	0.00161154	0.0046735	2.3718853	0.00000433	0.0000227
FAP	1.9932031	0.0015781	0.0098344	1.37558042	0.00581828	0.0124219	2.3624961	2.06E-07	2.16E-06
FN1	1.5366589	0.0342031	0.0667138	1.48470831	0.06004749	0.0733063	0.9406955	0.6830807	0.3149529
INHBA	3.0596839	1.07E-07	0.0000124	1.98607554	4.28E-09	3.73E-07	2.5487503	5.65E-06	0.0000273
SFRP4	2.3636087	0.0000248	0.0005927	1.67750397	1.87E-05	0.001515	2.6895394	6.50E-11	1.80E-09
SPARC	2.249132	0.0000652	0.0012192	1.81223384	0.00166372	0.0047457	1.4031045	0.0236369	0.0279596
THBS2	2.5780475	2.97E-07	0.0000256	1.89140939	7.54E-07	0.0000146	1.8704603	0.0000261	0.0000987
BN1	0.6582912	0.0008269	0.006465	0.53215394	1.35E-07	4.02E-06	0.4346961	2.11E-09	4.27E-08
C7	0.5305767	4.96E-06	0.0001778	0.61660718	1.43E-05	0.0001297	0.3687542	3.26E-12	1.24E-10
COL6A1	0.6814495	0.0146421	0.0470118	0.59821729	8.17E-05	0.0004585	0.4431674	2.65E-07	2.52E-06
DES	0.730098	0.0632254	0.1095273	0.61333977	0.00103514	0.0033666	0.3442483	2.78E-08	3.57E-07
FLNC	0.741509	0.0356714	0.0847442	0.85002893	0.23258502	0.1843726	0.3263553	4.74E-09	8.68E-08
GPM6B	0.6683566	0.0048803	0.0233566	0.60320026	0.00006445	0.0003935	0.4814972	2.01E-06	0.0000117
GSN	0.6464018	0.0057152	0.0257736	0.45512167	1.29E-07	4.02E-06	0.4635566	3.08E-07	2.84E-06
GSTM1	0.6720834	0.0063399	0.0269917	0.61970734	4.44E-06	0.000058	0.4449932	1.22E-09	2.65E-08
GSTM2	0.514483	0.0000907	0.0015002	0.52721579	4.50E-06	0.000058	0.2939022	3.94E-13	2.00E-11
HLF	0.5813615	0.0004971	0.0043504	0.50960012	3.14E-06	0.0000338	0.4179279	8.18E-09	1.31E-07
IGF1	0.6118674	0.0001721	0.0022429	0.62158322	7.65E-06	0.0000807	0.3470943	2.24E-13	1.36E-11
IGFBP6	0.5776972	0.003161	0.0172052	0.60163498	7.96E-05	0.0004543	0.4536368	1.44E-08	2.18E-07
ITGA7	0.6760378	0.0331569	0.0810327	0.54167205	2.10E-05	0.0001661	0.3682462	4.07E-10	1.03E-08
OLEML3	0.6460637	0.0011279	0.0008036	0.57873289	0.00001453	0.0001237	0.4154584	1.86E-08	2.64E-07
PAGE4	0.5182669	5.75E-06	0.0001903	0.66287751	2.46E-06	0.0000357	0.2677212	2.80E-17	8.51E-15
PPAP2B	0.5680087	0.0005371	0.0058913	0.45475585	6.81E-06	0.0000765	0.4140322	4.85E-09	8.68E-08

GENE	Association with PCS5 Endpoint			Association with cRF1, CAPRA Adjusted			Association with primary Gleason pattern		
	Std. HR	Wald p-value	Storey q-value	Std. HR	Wald p-value	Storey q-value	Std. OR	Wald p-value	Storey q-value
PPP1R12A	0.6937407	0.0165382	0.0496415	0.46377868	0.00149833	0.0044262	0.4793933	0.0000198	0.0000772
PRKCA	0.6323113	0.0014455	0.0092768	0.52602482	6.68E-05	0.0004008	0.3682007	6.36E-09	1.07E-07
SRD5A2	0.4878954	2.93E-06	0.0001573	0.53197502	1.48E-09	2.58E-07	0.2948852	1.63E-14	1.45E-12
VCL	0.6936282	0.4810739	0.3983524	0.48920691	0.00010426	0.0005415	0.4393103	7.90E-06	0.0002869
TGFB11	0.6700491	0.0291804	0.0749233	0.58638267	0.00350838	0.0089058	0.3711508	1.96E-08	2.64E-07
TPM2	0.6225376	0.0050533	0.0236188	0.55889574	0.00020286	0.0008936	0.4008574	1.04E-09	2.44E-08
TPX2	2.07392	0.0416277	0.0942101	1.80670715	7.56E-08	3.29E-06	2.1153062	4.98E-06	0.000248
CDC30	1.7300441	0.0000725	0.0012988	1.94642748	4.81E-06	0.0000588	1.6858343	0.000045	0.0001519
CDKN2C	1.99305	0.0125796	0.0432737	2.2135602	1.45E-05	0.0001297	1.6707388	0.00039	0.0009879
MYBL2	1.7322773	0.012916	0.0440786	1.64874559	6.39E-06	0.0000745	1.4091306	0.0068668	0.0098934
UBE2T	1.898363	0.0847937	0.1462198	1.87982555	0.00012433	0.0005929	1.594593	0.0006065	0.0014404
CYP3A5	0.5067698	0.0003008	0.0032333	0.58312246	1.39E-07	4.02E-06	0.5204925	9.40E-06	0.0000426
KRT15	0.6841526	0.0052343	0.0242015	0.7687095	7.50E-05	0.000435	0.5603937	0.0000173	0.0000713
KRT5	0.6729884	0.0041853	0.0216827	0.7364908	0.0002245	0.0009645	0.6403713	0.000719	0.0016558
LAMB3	0.7403354	0.0266386	0.0724973	0.76722056	0.00366663	0.0089858	0.730357	0.0177696	0.0219488
EGH1	0.4902253	0.0001344	0.0034238	0.80204599	1.58E-05	0.0001377	0.5936819	0.0007056	0.0016501
FOS	0.5555161	0.0045741	0.0223508	0.6374611	5.34E-05	0.0003316	0.5788589	0.0002155	0.0005851
GADD45B	0.5541679	0.0021788	0.0124919	0.64836478	3.81E-05	0.0002707	0.6176176	0.002403	0.0043482
JUN	0.505934	0.0013437	0.0088891	0.54410296	1.78E-05	0.00015	0.4795161	3.99E-07	3.37E-06
ZFP36	0.5757824	0.001287	0.0083714	0.68520845	0.00023482	0.0009966	0.6470867	0.0043499	0.0068874
DUSP1	0.6603212	0.0498518	0.1055975	0.63670824	0.00347613	0.0086407	0.586205	0.0007564	0.0017289
EGR3	0.5613678	0.0009351	0.0071603	0.71313117	0.0008831	0.002955	0.7186642	0.0260364	0.0000853
FAM13C	0.5260925	4.01E-09	1.72E-06	0.52541845	6.04E-10	2.10E-07	0.3709836	5.73E-11	1.74E-09
KLK2	0.5808923	0.0001686	0.0022425	0.56994638	0.00194411	0.0033687	0.5968229	0.0002289	0.0006158
A1DH1A2	0.5608861	0.0000177	0.0004751	0.65303146	0.00010806	0.000545	0.7822456	4.34E-07	3.57E-06
A2GP1	0.6167537	3.96E-06	0.0001778	0.6708779	4.39E-07	9.13E-06	0.5150783	1.17E-06	7.43E-05

GENE	Association with PCS5 Endpoint			Association with cRFI, CAPRA Adjusted			Association with primary Gleason pattern		
	Std. HR	Wald p-value	Storey q-value	Std. HR	Wald p-value	Storey q-value	Std. OR	Wald p-value	Storey q-value
ANPEP	0.5313085	0.0008229	0.006465	0.80509148	0.00011244	0.000559	0.5761885	0.0081251	0.0114354
AR	0.9479643	0.7435374	0.510526	0.77328075	0.35486409	0.244458	0.9337631	0.6073904	0.295812
BMP6	1.4900185	0.0210638	0.060383	1.44999312	0.00201534	0.0054792	2.3713254	6.85E-07	4.86E-06
CD276	1.6844232	0.0049028	0.0233566	2.18398768	0.00057532	0.0020641	2.1955358	2.75E-06	0.0000153
CD44	0.6866428	0.0155666	0.0485038	0.5502693	1.06E-07	4.02E-06	0.7793905	0.0682278	0.0654298
COL8A1	2.2449967	0.0000271	0.0006141	1.91011656	4.90E-05	0.0003316	1.8755907	5.13E-06	0.0000421
CSF1	0.6749873	0.0193771	0.0561984	0.44321491	5.01E-07	0.0000103	0.9573718	0.7643438	0.3461448
SRC	0.6670294	0.0040925	0.0212478	0.47330013	1.67E-06	0.0000307	0.766355	0.0813478	0.0758976
CSRP1	0.7112339	0.0067019	0.0277098	0.89239013	0.2633928	0.1994666	0.4248705	0.0058729	0.0088825
DPP4	0.5441442	1.12E-06	0.00008	0.68915455	0.00013991	0.0006492	0.4140282	1.92E-06	0.0000114
TNFRSF10B	0.6852925	0.0143692	0.0468086	0.53054603	4.04E-05	0.0002757	0.7430912	0.0304132	0.0339912
ERG	1.0785349	0.6794217	0.4928667	1.12737455	0.0349341	0.0498809	0.8943963	0.4148417	0.2324627
FAM107A	0.540565	0.0000605	0.0011827	0.57090059	6.82E-08	3.29E-06	0.3476335	1.99E-08	2.64E-07
IGFBP2	0.6977969	0.0532357	0.1095273	0.45025927	6.42E-06	0.0000745	0.6063083	0.0001586	0.0004465
CADM1	0.6456383	0.0150546	0.0472518	0.40819615	3.14E-08	2.19E-06	0.5598139	0.0001184	0.000346
IL6ST	0.5740032	0.0003647	0.0036466	0.33440325	2.36E-06	0.0000357	0.5467964	0.0040541	0.0065356
LGALS3	0.6782394	0.0071303	0.0283525	0.53406803	5.18E-05	0.0003278	0.5903729	0.0030449	0.0052894
SMAD4	0.5277628	4.87E-06	0.0001778	0.24376793	2.03E-06	0.0000036	0.3346323	1.85E-06	0.0000112
NFAT5	0.5361732	0.0000856	0.0014722	0.20926313	3.51E-07	8.71E-06	0.5518236	0.0000356	0.000126
SOL1	1.7087025	0.007387	0.0288525	1.43080815	0.02325931	0.0359744	1.8597668	0.0010445	0.002768
SHMT2	1.9491131	0.0031065	0.0171257	1.94514573	0.00591315	0.0124714	1.6896076	0.0074605	0.0105488
SLC22A3	0.5188636	0.000117	0.001706	0.65464628	7.32E-06	0.0000296	0.2293355	1.91E-14	1.45E-12
STAT5B	0.7002104	0.0396042	0.0914258	0.44673718	0.00045662	0.0017462	0.5417213	0.0000465	0.0001553
MMP11	1.8391119	0.0001041	0.0018437	1.62300343	1.23E-05	0.0001222	2.3250232	7.87E-07	5.44E-06
TUBB2A	0.6134538	0.0026235	0.0148438	0.56476388	1.81E-05	0.00015	0.9566513	0.6308242	0.2461448

EXAMPLE 2: ALGORITHM DEVELOPMENT BASED ON DATA FROM A COMPANION STUDY

[00150] The Cleveland Clinic ("CC") Companion study consists of three patient cohorts and separate analyses for each cohort as described in Table 2. The first cohort (Table 2) includes men with low to high risk (based on AUA criteria) prostate cancer from Gene ID study 09-002 who underwent RP at CC between 1987 and 2004 and had diagnostic biopsy tissue available at CC. Cohorts 2 and 3 include men with clinically localized Low and Intermediate Risk (based on AUA criteria) prostate cancer, respectively, who might have been reasonable candidates for active surveillance but who underwent radical prostatectomy (RP) within 6 months of the diagnosis of prostate cancer by biopsy. The main objective of Cohort 1 was to compare the molecular profile from biopsy tissue with that from radical prostatectomy tissue. The main objective of Cohorts 2 and 3 was to develop a multigene predictor of upgrading/upstaging at RP using biopsy tissue in low to intermediate risk patients at diagnosis.

[00151] Matched biopsy samples were obtained for a subset of the patients (70 patients) from the gene identification study. Gene expression of the 81 selected genes and the 5 reference genes (ARF1, ATP5E, CLTC, GPS1, PGK1) were compared in the RP specimens and the biopsy tissue obtained from these 70 patients.

[00152] The 81 genes were evaluated in Cohorts 2 and 3 for association with upgrading and upstaging. The association between these 81 genes and upgrading and upstaging in Cohorts 2 and 3 are shown in Table 3. P values and standardized odds ratio are provided.

[00153] In this context, "upgrade" refers to an increase in Gleason grade from 3+3 or 3+4 at the time of biopsy to greater than or equal to 3+4 at the time of RP. "Upgrade2" refers to an increase in Gleason grade from 3+3 or 3+4 at the time of biopsy to greater than or equal to 4+3 at the time of RP.

TABLE 2

Cohort #	Cohort Description	# of Patients	Objectives
1	<p>Subset of patients from Gene ID study 09-002 who underwent RP at CC between 1987 and 2004 and had diagnostic biopsy tissue available at CC.</p> <p>Patients from the original stratified cohort sample with available biopsy tissue blocks</p>	70	<ul style="list-style-type: none"> • Comparison of gene expression from biopsy sample with gene expression from RP specimen (Co-Primary Objective) • Explore association of risk of recurrence after RP with gene expression from biopsy sample and gene expression from RP sample • Explore association of risk of recurrence after RP with gene expression from RP samples
2	<p>Low Risk Patients from CC database of patients who were biopsied, and then underwent RP at CC between 1999 and 2010</p> <p>All patients in database who meet minimum tumor tissue criteria</p>	92	<ul style="list-style-type: none"> • Association between gene expression from biopsy sample and likelihood of upgrading/upstaging in tissue obtained at prostatectomy • (Co-Primary Objective)
3	<p>Intermediate Risk Patients from CC database of patients who were biopsied, and then underwent RP at CC between 1999 and 2010</p> <p>All patients in database who meet minimum tumor tissue criteria</p>	75	<ul style="list-style-type: none"> • Association between gene expression from biopsy sample and likelihood of upgrading/upstaging in tissue obtained at prostatectomy

[00154] Several different models were explored to compare expression between the RP and biopsy specimens. Genes were chosen based on consistency of expression between the RP and biopsy specimens. Figures 2A-2E are the scatter plots showing the comparison of normalized gene expression (Cp) for matched samples from each patient where the x-axis is the normalized gene expression from the PGP RP sample (PGP) and the y-axis is the normalized gene expression from the biopsy sample (BX). Figures 3A-3D show range plots of gene expression of individual genes within each gene group in the biopsy (BX) and PGP RP samples.

[00155] After evaluating the concordance of gene expression in biopsy and RP samples, the following algorithms (RS models) shown in Table 4 were developed where the weights are determined using non-standardized, but normalized data. Some genes, such as SRD5A2 and GSTM2, which fall within the cellular organization gene group, were also evaluated separately and independent coefficients were assigned (see the “other” category in Table 4). In other instances, GSTM1 and GSMT2 were grouped as an oxidative “stress” group and a

coefficient was assigned to this “stress” group (see RS20 and RS22 models). Other genes, such as AZGP1 and SLC22A3, which did not fall within any of the gene groups, were also included in certain algorithms (see the “other” category in Table 4). Furthermore, the androgen gene group was established to include FAM13C, KLK2, AZGP1, and SRD5A2. Some genes such as BGN, SPARC, FLNC, GSN, TPX2 and SRD5A2 were thresholded before being evaluated in models. For example, normalized expression values below 4.5 were set to 4.5 for TPX2 and normalized expression values below 5.5 were set to 5.5 for SRD5A2.

Table 3: Association between the 81 genes and Upgrading and Upstaging in Cohorts 2/3

Gene	N	p-value UpGrade	Std OR Upgrade	95% CI Upgrade	p-value Upgrade2	Std OR Upgrade2	95% CI Upgrade2	p-value Upstage	Std OR Upstage	95% CI Upstage
ALDH1A2	167	0.501	1.11	(0.82,1.52)	0.932	1.02	(0.70,1.47)	0.388	0.86	(0.61,1.22)
ANPEP	167	0.054	1.36	(0.99,1.87)	0.933	0.98	(0.68,1.42)	0.003	0.58	(0.40,0.83)
AR	167	0.136	1.27	(0.93,1.74)	0.245	0.81	(0.56,1.16)	0.005	0.60	(0.42,0.86)
ARF1	167	0.914	0.98	(0.72,1.34)	0.051	1.45	(1.00,2.11)	0.371	1.17	(0.83,1.66)
ASPN	167	0.382	1.15	(0.84,1.56)	0.040	1.60	(1.02,2.51)	0.069	1.46	(0.97,2.19)
ATP5E	167	0.106	1.30	(0.95,1.77)	0.499	0.88	(0.61,1.27)	0.572	0.90	(0.64,1.28)
AZGP1	167	0.192	1.23	(0.90,1.68)	0.190	0.79	(0.55,1.13)	0.005	0.59	(0.41,0.85)
BGN	167	0.568	0.91	(0.67,1.25)	0.001	2.15	(1.39,3.33)	0.020	1.56	(1.07,2.28)
BIN1	167	0.568	1.09	(0.80,1.49)	0.634	0.92	(0.64,1.32)	0.104	0.75	(0.54,1.06)
BMP6	167	0.509	0.90	(0.66,1.23)	0.015	1.59	(1.09,2.30)	0.650	1.08	(0.77,1.54)
C7	167	0.677	1.07	(0.78,1.46)	0.013	1.66	(1.11,2.47)	0.223	0.80	(0.56,1.14)
CADM1	167	0.082	0.74	(0.52,1.04)	0.235	0.81	(0.57,1.15)	0.039	0.69	(0.48,0.98)
CD276	167	0.454	0.89	(0.65,1.21)	0.362	0.84	(0.58,1.22)	0.214	1.25	(0.88,1.78)
CD44	167	0.122	1.28	(0.94,1.75)	0.305	1.23	(0.83,1.81)	0.876	0.97	(0.69,1.38)
CDC20	166	0.567	1.10	(0.80,1.50)	0.298	1.21	(0.84,1.75)	0.279	1.21	(0.86,1.71)
CDKN2C	152	0.494	0.89	(0.64,1.24)	0.908	0.98	(0.67,1.43)	0.834	1.04	(0.72,1.49)
CLTC	167	0.102	0.76	(0.55,1.06)	0.300	0.82	(0.57,1.19)	0.264	0.82	(0.58,1.16)
COL1A1	167	0.732	1.06	(0.77,1.44)	0.000	3.04	(1.93,4.79)	0.006	1.65	(1.15,2.36)
COL1A2	167	0.574	0.91	(0.67,1.25)	0.017	1.65	(1.09,2.50)	0.521	0.89	(0.63,1.26)
COL3A1	167	0.719	0.94	(0.69,1.29)	0.000	2.98	(1.88,4.71)	0.020	1.53	(1.07,2.20)
COL4A1	167	0.682	0.94	(0.69,1.28)	0.000	2.12	(1.39,3.22)	0.762	0.95	(0.67,1.35)
COL5A2	167	0.499	1.11	(0.82,1.52)	0.009	1.81	(1.16,2.83)	0.516	0.89	(0.63,1.26)
COL6A1	167	0.878	0.98	(0.72,1.33)	0.001	2.14	(1.37,3.34)	0.883	1.03	(0.72,1.46)
COL8A1	165	0.415	0.88	(0.64,1.20)	0.000	3.24	(1.88,5.61)	0.044	1.51	(1.01,2.25)
CSF1	167	0.879	1.02	(0.75,1.40)	0.187	1.31	(0.88,1.96)	0.110	0.76	(0.54,1.07)

Gene	N	p-value UpGrade	Std OR UpGrade	95% CI UpGrade	p-value UpGrade2	Std OR UpGrade2	95% CI UpGrade2	p-value Upstage	Std OR Upstage	95% CI Upstage
C5RP1	165	0.258	1.20	(0.87,1.65)	0.226	1.26	(0.87,1.82)	0.641	0.92	(0.65,1.31)
CYP3A5	167	0.989	1.00	(0.73,1.36)	0.188	1.28	(0.88,1.87)	0.937	1.01	(0.71,1.44)
DES	167	0.776	1.05	(0.77,1.43)	0.088	1.40	(0.95,2.05)	0.242	0.81	(0.57,1.15)
DPP4	167	0.479	0.89	(0.65,1.22)	0.005	0.60	(0.42,0.85)	0.000	0.51	(0.36,0.74)
DUSP1	167	0.295	0.84	(0.61,1.16)	0.262	0.82	(0.58,1.16)	0.427	0.87	(0.62,1.22)
EGR1	167	0.685	0.94	(0.69,1.28)	0.217	1.27	(0.87,1.85)	0.370	1.18	(0.83,1.68)
EGR3	166	0.025	0.69	(0.50,0.95)	0.539	0.89	(0.62,1.29)	0.735	1.06	(0.75,1.51)
ERG	166	0.002	0.58	(0.42,0.81)	0.000	0.42	(0.28,0.64)	0.768	1.05	(0.74,1.50)
F2R	160	0.324	0.85	(0.62,1.17)	0.009	1.77	(1.16,2.70)	0.000	2.39	(1.52,3.76)
FAM107A	143	0.832	1.04	(0.74,1.45)	0.088	1.42	(0.95,2.11)	0.687	1.08	(0.74,1.58)
FAM13C	167	0.546	1.10	(0.81,1.50)	0.041	0.68	(0.47,0.98)	0.003	0.58	(0.40,0.83)
FAP	167	0.540	0.91	(0.67,1.24)	0.093	1.37	(0.95,1.97)	0.001	1.85	(1.28,2.68)
FLNC	167	0.963	1.01	(0.74,1.37)	0.254	1.26	(0.85,1.87)	0.030	0.68	(0.48,0.96)
FN1	167	0.530	0.91	(0.66,1.23)	0.005	1.73	(1.18,2.53)	0.364	1.17	(0.83,1.66)
FOS	167	0.649	0.93	(0.68,1.27)	0.071	1.38	(0.97,1.97)	0.015	1.53	(1.09,2.16)
GADD45B	167	0.978	1.00	(0.73,1.36)	0.105	1.38	(0.94,2.04)	0.876	0.97	(0.69,1.38)
GPM6B	159	0.944	0.99	(0.72,1.36)	0.002	1.95	(1.27,2.97)	0.266	0.81	(0.57,1.17)
GPS1	167	0.404	1.14	(0.84,1.56)	0.609	0.91	(0.62,1.32)	0.125	1.31	(0.93,1.86)
GSN	167	0.272	0.84	(0.61,1.15)	0.309	0.83	(0.57,1.19)	0.027	0.67	(0.47,0.96)
GSTM1	167	0.178	1.24	(0.91,1.69)	0.762	0.95	(0.66,1.36)	0.000	0.50	(0.34,0.72)
GSTM2	167	0.145	1.26	(0.92,1.73)	0.053	1.48	(1.00,2.20)	0.654	0.92	(0.65,1.31)
HLF	167	0.979	1.00	(0.73,1.36)	0.602	1.11	(0.76,1.62)	0.030	0.69	(0.49,0.96)
IGF1	167	0.313	1.17	(0.86,1.60)	0.878	0.97	(0.67,1.40)	0.146	0.77	(0.55,1.09)
IGFBP2	167	0.253	1.20	(0.88,1.64)	0.493	0.88	(0.61,1.27)	0.051	0.70	(0.49,1.00)
IGFBP6	167	0.336	0.86	(0.62,1.17)	0.510	1.14	(0.78,1.66)	0.204	0.80	(0.57,1.13)
IL6ST	167	0.774	1.05	(0.77,1.43)	0.541	1.12	(0.77,1.63)	0.235	0.81	(0.57,1.15)
INHBA	167	0.104	1.30	(0.95,1.78)	0.002	1.89	(1.26,2.84)	0.077	1.38	(0.97,1.97)

Gene	N	p-value UpGrade	Std OR UpGrade	95% CI UpGrade	p-value UpGrade2	Std OR UpGrade2	95% CI UpGrade2	p-value Upstage	Std OR Upstage	95% CI Upstage
ITGA7	167	0.990	1.00	(0.73,1.36)	0.780	1.05	(0.73,1.53)	0.470	0.88	(0.62,1.25)
JUN	167	0.586	1.09	(0.80,1.48)	0.538	0.89	(0.62,1.28)	0.259	0.82	(0.59,1.15)
KLK2	167	0.267	0.84	(0.61,1.15)	0.003	0.56	(0.38,0.82)	0.007	0.61	(0.42,0.87)
KRT15	167	0.500	0.90	(0.65,1.23)	0.738	0.94	(0.65,1.35)	0.987	1.00	(0.71,1.42)
KRT5	152	0.834	0.97	(0.70,1.34)	0.632	1.10	(0.74,1.53)	0.908	0.98	(0.68,1.40)
LAMB3	167	0.090	1.31	(0.96,1.79)	0.013	1.73	(1.12,2.68)	0.132	1.33	(0.92,1.94)
LGALS3	166	0.345	1.16	(0.85,1.59)	0.405	1.18	(0.80,1.72)	0.208	0.80	(0.57,1.13)
MMP11	167	0.715	1.06	(0.78,1.45)	0.080	1.37	(0.96,1.96)	0.257	1.22	(0.87,1.71)
MYBL2	167	0.235	1.21	(0.88,1.67)	0.868	1.03	(0.71,1.49)	0.266	1.21	(0.86,1.70)
NFAT5	167	0.514	0.90	(0.66,1.23)	0.058	0.70	(0.48,1.01)	0.530	0.89	(0.63,1.27)
OLFML3	167	0.448	0.89	(0.65,1.21)	0.056	1.50	(0.99,2.28)	0.129	0.77	(0.54,1.08)
PAGE4	167	0.914	0.98	(0.72,1.34)	0.211	0.80	(0.56,1.14)	0.005	0.61	(0.43,0.86)
PGK1	167	0.138	0.78	(0.56,1.08)	0.666	0.92	(0.64,1.33)	0.292	0.83	(0.59,1.17)
PPAP2B	167	0.952	0.99	(0.73,1.35)	0.989	1.00	(0.69,1.44)	0.221	0.80	(0.56,1.14)
PPP1R12A	167	0.547	0.91	(0.66,1.24)	0.563	0.90	(0.63,1.29)	0.001	0.55	(0.38,0.79)
PRKCA	167	0.337	1.17	(0.85,1.59)	0.141	1.35	(0.90,2.03)	0.029	0.67	(0.46,0.96)
SDC1	167	0.064	1.36	(0.98,1.87)	0.013	1.83	(1.14,2.96)	0.037	1.58	(1.03,2.42)
SFRP4	166	0.986	1.00	(0.73,1.37)	0.047	1.47	(1.01,2.15)	0.031	1.49	(1.04,2.14)
SHMT2	167	0.133	0.78	(0.56,1.08)	0.147	0.77	(0.53,1.10)	0.715	0.94	(0.66,1.33)
SLC22A3	167	0.828	1.03	(0.76,1.41)	0.044	0.69	(0.48,0.99)	0.050	0.71	(0.50,1.00)
SMAD4	167	0.165	1.25	(0.91,1.71)	0.333	0.83	(0.58,1.21)	0.021	0.65	(0.45,0.94)
SPARC	167	0.810	0.96	(0.71,1.31)	0.000	2.15	(1.40,3.30)	0.154	1.30	(0.91,1.86)
SRC	167	0.083	1.34	(0.96,1.86)	0.750	1.06	(0.72,1.56)	0.550	0.90	(0.64,1.26)
SRD5A2	167	0.862	0.97	(0.71,1.33)	0.122	0.75	(0.53,1.08)	0.010	0.63	(0.45,0.90)
STAT5B	167	0.298	0.84	(0.62,1.16)	0.515	0.89	(0.62,1.27)	0.016	0.65	(0.46,0.92)
TGFB1I1	167	0.985	1.00	(0.74,1.37)	0.066	1.45	(0.98,2.14)	0.131	0.76	(0.54,1.08)
THBS2	167	0.415	1.14	(0.83,1.56)	0.001	1.91	(1.30,2.80)	0.288	1.21	(0.85,1.70)

Gene	N	p-value UpGrade	Std OR Upgrade	95% CI Upgrade	p-value Upgrade2	Std OR Upgrade2	95% CI Upgrade2	p-value Upstage	Std OR Upstage	95% CI Upstage
TNFRSF10B	167	0.214	1.22	(0.89,1.66)	0.805	0.95	(0.66,1.38)	0.118	0.76	(0.54,1.07)
TPM2	167	0.996	1.00	(0.73,1.36)	0.527	1.13	(0.78,1.64)	0.094	0.74	(0.52,1.05)
TPX2	167	0.017	1.48	(1.07,2.04)	0.002	1.89	(1.26,2.83)	0.001	1.91	(1.30,2.80)
TUBB2A	167	0.941	0.99	(0.73,1.35)	0.182	0.78	(0.54,1.12)	0.111	0.75	(0.53,1.07)
UBE2T	167	0.095	1.36	(0.95,1.96)	0.009	1.58	(1.12,2.23)	0.084	1.33	(0.96,1.84)
VCL	167	0.954	0.99	(0.73,1.35)	0.165	1.31	(0.90,1.91)	0.265	0.82	(0.57,1.16)
ZFP36	167	0.685	1.07	(0.78,1.45)	0.784	0.95	(0.66,1.37)	0.610	0.91	(0.64,1.29)

Table 4

RS Model	ECM (Stromal Response)	Migration (Cellular Organization)	Prolif.	Androgen (PSA)	Other	Algorithm
RS0	(ASPN+ BGN+COL1A1+SPARC)/4	(FLNC+ GSN+GSTM2+ IGFBP6+PPAP2B+PPP1R12A)/6	(TPX2+ CDC20+ MYBL2)/3	(FAM13C+ KLK2)/2	STAT5B, NFAT5	1.05*ECM-0.58*Migration-0.30*PSA+0.08*Prolif-0.16*STAT5B-0.23*NFAT5
RS1	(BGN+ COL1A1+FN1+SPARC)/4	(FLNC+GSN+GSTM2+PPAP2B+ PPP1R12A)/6	.	(FAM13C+ KLK2)/2	STAT5B, NFAT5	1.15*ECM-0.72*Migration-0.56*PSA-0.45*STAT5B-0.56*NFAT5
RS2	(BGN+ COL1A1+FN1 +SPARC)/4	(BIN1+FLNC+GSN+GSTM2+ PPAP2B+PPP1R12A+ VCL)/7	.	(FAM13C+ KLK2)/2	STAT5B, NFAT5	1.16*ECM-0.75*Migration-0.57*PSA-0.47*STAT5B-0.50*NFAT5
RS3	(BGN+ COL1A1 +COL3A1+ COL4A1+FN1 +SPARC)/6	(FLNC+ GSN+GSTM2+PPAP2B+ PPP1R12A)/5	.	(FAM13C+ KLK2)/2	STAT5B, NFAT5	1.18*ECM-0.75*Migration-0.56*PSA-0.40*STAT5B-0.48*NFAT5
RS4	(BGN +COL1A1+ COL3A1+ COL4A1 +FN1+SPARC)/6	(BIN1+FLNC+ GSN+GSTM2+ PPAP2B+PP1R12A +VCL)/7	.	(FAM13C+ KLK2)/2		1.18*ECM-0.76*Migration-0.58*PSA-0.43*STAT5B-0.43*NFAT5
RS5	(COL4A1 (thresholded) + INHBA + SPARC + THBS2)/4	(BIN1 + IGFBP2 (thresholded) + VCL)/3	.	KLK2	AZGP1, ANPEP, IGFBP2 (thresholded)	1.20*ECM-0.91*Migration-0.29*KLK2-0.14*AZGP1+0.05*ANPEP-0.56*IGFBP2
RS6	(BGN+COL3A1+INHBA+ SPARC)/4	Migratn1: (FLNC+GSN+TPM2)/3 Migratn2: (GSTM2+PPAP2B)/2	TPX2	(FAM13C+ KLK2)/2	AZGP1, SLC22A3	1.09*ECM-0.44*Migration1-0.23*Migration2-0.36*PSA+0.15*TPX2-0.16*AZGP1-0.08*SLC22A3
RS7	(BGN+COL3A1+INHBA+ SPARC)/4	Migratn1: (FLNC+GSN+TPM2)/3 Migratn2: (GSTM2+PPAP2B)/2	.	(FAM13C+ KLK2)/2	AZGP1, SLC22A3	1.16*ECM-0.53*Migration1-0.24*Migration2-0.42*PSA-0.14*AZGP1-0.08*SLC22A3
RS8	(BGN+COL3A1+ SPARC)/3	Migratn1: (FLNC+GSN+TPM2)/3 Migratn2: (GSTM2+PPAP2B)/2	.	KLK2	AZGP1, SLC22A3	1.37*ECM-0.56*Migration1-0.49*Migration2-0.52*KLK2-0.16*AZGP1-0.00*SLC22A3
RS9	(BGN (thresholded)+COL3A1 +	Migratn1: (FLNC (thresholded) +GSN (thresholded) +TPM2)/3	.	(FAM13C+ KLK2)/2	AZGP1, SLC22A3	1.28*ECM-1.11*Migration1-0.00*Migration2-0.34*PSA-

RS Model	ECM (Stromal Response)	Migration (Cellular Organization)	Prolif.	Androgen (PSA)	Other	Algorithm
	INHBA + SPARC (thresholded) //4	Migratn2: (GSTM2+PPAP2B)/2				0.16*AZGP1-0.08*SLC22A3
RS10	(BGN+COL3A1+INHBA+SPARC)/4	(FLNC+GSN+GSTM2+PPAP2B+TPM2)/5	TPX2	(FAM13C+ KLK2)/2	AZGP1, SLC22A3	1.09*ECM-0.68*Migration-0.37*PSA+ 0.16*TPX2-0.16*AZGP1-0.08*SLC22A3
RS11	(BGN (thresholded)+COL3A1+ INHBA+ SPARC(thresholded))/4	(FLNC(thresholded) + GSN(thresholded)+GSTM2 +PPAP2B +TPM2)/5		(FAM13C+ KLK2)/2	AZGP1, SLC22A3	1.19*ECM -0.96*Migration-0.39*PSA-0.14*AZGP1 -0.09*SLC22A3
RS12	(BGN (thresholded)+COL3A1+ INHBA+ SPARC(thresholded))/4	(FLNC(thresholded) + GSN(thresholded)+GSTM2 +PPAP2B +TPM2)/5	TPX2	(FAM13C+ KLK2)/2	AZGP1, SLC22A3	1.13*ECM-0.85*Migration-0.34*PSA + 0.15*TPX2-0.15*AZGP1-0.08*SLC22A3
RS13	(BGN (thresholded)+COL3A1+ INHBA+ SPARC(thresholded))/4	(FLNC(thresholded) + GSN(thresholded)+GSTM2 +PPAP2B +TPM2)/5	TPX2	(FAM13C+ KLK2)/2	AZGP1, ERG, SLC22A3	1.12*ECM -0.83*Migration-0.33*PSA+ 0.17*TPX2 -0.14*AZGP1 +0.04*ERG-0.10*SLC22A3
RS14	(BGN (thresholded)+COL3A1+ INHBA+ SPARC(thresholded))/4	(FLNC(thresholded) + GSN(thresholded)+GSTM2 +PPAP2B +TPM2)/5	TPX2	(FAM13C+ KLK2)/2	AR, AZGP1, ERG, SLC22A3	1.13*ECM -0.83*Migration-0.35*PSA+ 0.16*TPX2+0.15*AR -0.15*AZGP1 +0.03*ERG-0.10*SLC22A3
RS15	(BGN (thresholded)+COL3A1+ INHBA+ SPARC(thresholded))/4	(FLNC(thresholded) + GSN(thresholded)+GSTM2 +PPAP2B +TPM2)/5		KLK2	AR, ERG, SLC22A3	1.30*ECM -1.20*Migration-0.52*KLK2+ 0.09*AR +0.05*ERG -0.06*SLC22A3
RS16	(BGN (thresholded)+COL3A1+ INHBA+ SPARC(thresholded))/4	(C7+ FLNC(thresholded)+ GSN(thresholded)+GSTM1)/4		KLK2	AR, ERG, SLC22A3	1.23*ECM -1.02*Migration-0.46*KLK2+ 0.09*AR+0.07*ERG -0.09*SLC22A3
RS17	(BGN+COL1A1+5FRP4)/3	(FLNC+GSN+GSTM1+ TPM2)/4	TPX2	(FAM13C+ KLK2)/2	AR, AZGP1, ERG, SLC22A3,SRD5A2	0.63*ECM-0.12*Migration-0.44*PSA+ 0.19*TPX2-0.02*AR-0.15*AZGP1+0.06*ERG-0.13*SLC22A3-0.33*SRD5A2

RS Model	ECM (Stromal Response)	Migration (Cellular Organization)	Prolif.	Androgen (PSA)	Other	Algorithm
RS18	(BGN+COL1A1+SFRP4)/3	(FLNC+GSN+GSTM1+TPM2)/4	TPX2	(FAM13C+KLK2)/2	AR, ERG, SLC22A3, SRD5A2	0.63*ECM-0.17*Migration-0.52*PSA+0.19*TPX2-0.07*AR+0.09*ERG-0.14*SLC22A3-0.36*SRD5A2
RS19	(BGN+COL1A1+SFRP4)/3	(FLNC+GSN+GSTM1+TPM2)/4	.	(FAM13C+KLK2)/2	AR, AZGP1, ERG, SLC22A3, SRD5A2	0.72*ECM-0.24*Migration-0.51*PSA+0.03*AR-0.15*AZGP1+0.04*ERG-0.12*SLC22A3-0.32*SRD5A2
RS20	(BGN+COL1A1+SFRP4)/3	(FLNC+GSN+PPAP2B+TPM2)/4	TPX2	(FAM13C+KLK2)/2	(Stress: GSTM1+GSTM2) AZGP1, SLC22A3, SRD5A2	0.72*ECM-0.26*Migration-0.45*PSA+0.15*TPX2+0.02*Stress-0.16*AZGP1-0.06*SLC22A3-0.30*SRD5A2
RS21	(BGN+COL1A1+SFRP4)/3	(FLNC+GSN+PPAP2B+TPM2)/4	TPX2	(FAM13C+KLK2)/2	AZGP1, SLC22A3, SRD5A2	0.68*ECM-0.19*Migration-0.43*PSA+0.16*TPX2-0.18*AZGP1-0.07*SLC22A3-0.31*SRD5A2
RS22	(BGN+COL1A1+SFRP4)/3		TPX2	(FAM13C+KLK2)/2	(Stress: GSTM1+GSTM2) AZGP1, SLC22A3, SRD5A2	0.62*ECM-0.46*PSA+0.18*TPX2-0.07*Stress-0.18*AZGP1-0.08*SLC22A3-0.34*SRD5A2
RS23	(BGN+COL1A1+SFRP4)/3	(FLNC+GSN+GSTM2+TPM2)/4	TPX2	(FAM13C+KLK2)/2	AR, AZGP1, ERG, SRD5A2	0.73*ECM-0.26*Migration-0.45*PSA+0.17*TPX2+0.02*AR-0.17*AZGP1+0.03*ERG-0.29*SRD5A2
RS24	(BGN+COL1A1+SFRP4)/3	(FLNC+GSN+GSTM1+GSTM2+PPAP2B+TPM2)/6	TPX2	(FAM13C+KLK2)/2	AZGP1, SLC22A3, SRD5A2	0.52*ECM-0.23*Migration-0.30*PSA+0.14*TPX2-0.17*AZGP1-0.07*SLC22A3-0.27*SRD5A2

RS Model	ECM (Stromal Response)	Migration (Cellular Organization)	Prolif.	Androgen (PSA)	Other	Algorithm
RS25	{BGN+COL1A1+SFRP4}/3	{FLNC+GSN+TPM2}/3	TPX2	{FAM13C+KLK2}/2	AZGP1, GSTM2, SRD5A2	0.72*ECM-0.14*Migration-0.45*PSA+0.16*TPX2 - 0.17*AZGP11-0.14*GSTM2-0.28*SRD5A2
RS26	{1.581*BGN+1.371*COL1A1+0.469*SFRP4}/3	{0.489*FLNC+1.512*GSN+1.264*TPM2}/3	TPX2 (thresholded)	{1.267*FAM13C+2.158*KLK2}/2	AZGP1, GSTM2, SRD5A2 (thresholded)	0.735*ECM-0.368*Migration-0.352*PSA+0.094*TPX2 - 0.226*AZGP11-0.145*GSTM2-0.351*SRD5A2
RS27	{1.581*BGN+1.371*COL1A1+0.469*SFRP4}/3 =0.527*BGN+0.457*COL1A1+0.156*SFRP4	{0.489*FLNC+1.512*GSN+1.264*TPM2}/3]+ {0.145*GSTM2/0.368} =0.163*FLNC+0.504*GSN+0.421*TPM2+0.394*GSTM2	TPX2 (thresholded)	{1.267*FAM13C+2.158*KLK2}/2]+ {0.226*AZGP1/0.352}+ {0.351*SRD5A2Thresh/0.352} =0.634*FAM13C+1.079*KLK2+0.642*AZGP1+0.997SRD5A2Thresh		0.735*ECM-0.368*Migration-0.352*PSA+0.095*TPX2

[00156] Table 5A shows the standardized odds ratio of each of the RS models using the data from the original Gene ID study described in Example 1 for time to cR and for upgrading and upstaging and the combination of significant upgrading and upstaging. Table 5B shows the performance of each of the RS models using the data from the CC Companion (Cohorts 2 and 3) study for upgrading and upstaging and the combination of significant upgrading and upstaging. In this context, “upgrading” refers to an increase in Gleason grade from 3+3 or 3+4 at biopsy to greater than or equal to 3+4 at radical prostatectomy. “Significant upgrading” in this context refers to upgrading from Gleason grade 3+3 or 3+4 at biopsy to equal to or greater than 4+3 at radical prostatectomy.

[00157] In addition, the gene groups used in the RS25 model were evaluated alone and in various combinations. Table 6A shows the results of this analysis using the data from the Gene Identification study and Table 6B shows the results of this analysis using the data from Cohorts 2 and 3 of the CC Companion Study.

[00158] The gene expression for some genes may be thresholded, for example SRD5A2 Thresh = 5.5 if SRD5A2 <5.5 or SRD5A2 if SRD5A2 ≥5.5 and TPX2 Thresh = 5.0 if TPX2 <5.0 or TPX2 if TPX2 ≥5.0, wherein the gene symbols represent normalized gene expression values.

[00159] The unscaled RS scores derived from Table 4 can also be rescaled to be between 0 and 100. For example, RS27 can be rescaled to be between 0 and 100 as follows:

[00160] RS (scaled) = 0 if $13.4 \times (RSu+10.5) < 0$; $13.4 \times (RSu+10.5)$ if $0 \leq 13.4 \times (RSu+10.5) \leq 100$; or 100 if $13.4 \times (RSu+10.5) > 100$.

[00161] Using the scaled RS, patients can be classified into low, intermediate, and high RS groups using pre-specified cut-points defined below in Table B. These cut-points define the boundaries between low and intermediate RS groups and between intermediate and high RS groups. The cutpoints were derived from the discovery study with the intent of identifying substantial proportions of patients who on average had clinically meaningful low or high risk of aggressive disease. The scaled RS is rounded to the nearest integer before the cut-points defining RS groups are applied.

TABLE B.

RS Group	Risk Score
Low	Less than 16
Intermediate	Greater than or equal to 16 and less than 30
High	Greater than or equal to 30

Table 5A

RS	N	Upgrading		Significant Upgrading		Upstaging		Significant Upgrading or Upstaging	
		OR	95% CI	OR	95% CI	OR	95% CI	OR	95% CI
RS0	280	1.72	(1.22,2.41)	7.51	(4.37,12.9)	2.01	(1.41,2.88)	2.91	(1.95,4.34)
RS1	287	1.73	(1.21,2.48)	5.98	(3.30,10.8)	1.99	(1.40,2.82)	2.68	(1.80,3.97)
RS2	287	1.72	(1.19,2.48)	5.89	(3.18,10.9)	2.02	(1.42,2.86)	2.67	(1.80,3.95)
RS3	287	1.71	(1.20,2.45)	6.30	(3.66,10.8)	1.96	(1.38,2.80)	2.69	(1.84,3.93)
RS4	287	1.69	(1.18,2.42)	6.06	(3.48,10.5)	1.99	(1.40,2.82)	2.65	(1.82,3.86)
RS5	288	1.78	(1.21,2.62)	5.60	(3.56,8.81)	2.24	(1.59,3.15)	2.87	(1.93,4.28)
RS6	287	1.94	(1.37,2.74)	10.16	(5.82,17.8)	2.07	(1.48,2.91)	3.11	(2.07,4.67)
RS7	288	1.91	(1.34,2.71)	9.34	(5.25,16.6)	2.06	(1.47,2.89)	3.01	(2.02,4.48)
RS8	289	1.80	(1.27,2.55)	7.49	(4.02,14.0)	2.09	(1.49,2.92)	2.86	(1.97,4.14)
RS9	288	2.00	(1.39,2.89)	9.56	(5.06,18.0)	1.99	(1.42,2.79)	3.09	(2.08,4.60)
RS10	287	1.94	(1.37,2.75)	10.12	(5.79,17.7)	2.09	(1.49,2.94)	3.14	(2.08,4.74)
RS11	288	2.09	(1.43,3.05)	9.46	(5.18,17.3)	2.17	(1.54,3.05)	3.42	(2.24,5.23)
RS12	287	2.10	(1.45,3.04)	10.41	(5.92,18.3)	2.17	(1.55,3.06)	3.52	(2.30,5.40)
RS13	287	2.10	(1.44,3.05)	9.40	(5.50,16.1)	2.20	(1.55,3.13)	3.50	(2.25,5.43)
RS14	287	2.06	(1.42,2.99)	9.71	(5.65,16.7)	2.18	(1.55,3.08)	3.53	(2.29,5.44)
RS15	288	1.92	(1.32,2.78)	7.93	(4.56,13.8)	2.12	(1.51,2.99)	3.25	(2.20,4.80)
RS16	288	1.76	(1.23,2.52)	7.10	(4.12,12.2)	1.99	(1.41,2.82)	2.94	(1.98,4.38)
RS17	286	2.23	(1.52,3.27)	7.52	(4.18,13.5)	2.91	(1.93,4.38)	4.48	(2.72,7.38)
RS18	286	2.12	(1.46,3.08)	7.04	(3.87,12.8)	2.89	(1.91,4.37)	4.30	(2.62,7.06)
RS19	287	2.14	(1.46,3.13)	6.90	(3.80,12.5)	2.88	(1.96,4.23)	4.20	(2.66,6.63)
RS20	286	2.30	(1.55,3.42)	8.41	(4.65,15.2)	2.90	(1.98,4.25)	4.78	(3.00,7.61)
RS21	287	2.36	(1.59,3.52)	8.83	(4.87,16.0)	2.63	(1.76,3.94)	4.93	(3.06,7.93)
RS22	286	2.16	(1.48,3.15)	7.57	(4.14,13.8)	2.90	(1.96,4.27)	4.39	(2.75,7.01)
RS23	287	2.26	(1.53,3.35)	7.46	(4.24,13.1)	2.80	(1.85,4.24)	4.79	(2.98,7.68)
RS24	286	2.21	(1.50,3.24)	8.01	(4.38,14.7)	2.93	(1.99,4.31)	4.62	(2.89,7.39)
RS25	287	2.25	(1.53,3.31)	7.70	(4.25,14.0)	2.76	(1.83,4.16)	4.76	(2.99,7.58)
RS26	287	2.23	(1.51,3.29)	6.67	(3.52,12.7)	2.64	(1.81,3.86)	4.01	(2.56,6.28)
RS27	287	2.23	(1.51,3.29)	6.67	(3.52,12.7)	2.64	(1.81,3.86)	4.01	(2.56,6.28)

Table SB

Model	Upgrading			Significant Upgrading			Upstaging			Significant Upgrading or Upstaging		
	N	Std OR	95% CI	Std OR	95% CI	Std OR	95% CI	Std OR	95% CI	Std OR	95% CI	95% CI
RS0	166	1.16	(0.84,1.58)	2.45	(1.61,3.73)	2.42	(1.61,3.62)	3	(1.98,4.56)			
RS1	167	1.05	(0.77,1.43)	2.46	(1.63,3.71)	2.38	(1.61,3.53)	3.36	(2.18,5.18)			
RS2	167	1.04	(0.76,1.42)	2.45	(1.63,3.69)	2.34	(1.58,3.46)	3.25	(2.12,4.99)			
RS3	167	1.04	(0.76,1.41)	2.56	(1.69,3.89)	2.28	(1.55,3.36)	3.27	(2.13,5.03)			
RS4	167	1.03	(0.75,1.40)	2.54	(1.68,3.86)	2.23	(1.52,3.27)	3.16	(2.07,4.82)			
RS5	167	1.02	(0.75,1.39)	1.89	(1.28,2.78)	1.77	(1.23,2.55)	2.21	(1.52,3.20)			
RS6	167	1.08	(0.79,1.48)	2.49	(1.64,3.79)	2.42	(1.62,3.62)	3.22	(2.09,4.96)			
RS7	167	1.03	(0.75,1.40)	2.31	(1.54,3.48)	2.28	(1.54,3.38)	2.97	(1.96,4.51)			
RS8	167	0.94	(0.69,1.28)	2.34	(1.55,3.53)	2.31	(1.56,3.43)	2.87	(1.91,4.30)			
RS9	167	1.02	(0.75,1.39)	2.19	(1.47,3.27)	2.22	(1.51,3.27)	2.77	(1.85,4.14)			
RS10	167	1.08	(0.79,1.48)	2.49	(1.63,3.78)	2.41	(1.61,3.61)	3.22	(2.09,4.95)			
RS11	167	0.99	(0.73,1.35)	2.18	(1.46,3.24)	2.17	(1.48,3.19)	2.83	(1.88,4.25)			
RS12	167	1.06	(0.78,1.45)	2.36	(1.57,3.56)	2.34	(1.57,3.48)	3.12	(2.04,4.78)			
RS13	166	1.01	(0.74,1.37)	2.17	(1.45,3.23)	2.41	(1.61,3.60)	2.99	(1.97,4.54)			
RS14	166	1.03	(0.76,1.41)	2.22	(1.48,3.31)	2.33	(1.57,3.46)	2.95	(1.94,4.47)			
RS15	166	1	(0.73,1.36)	1.98	(1.34,2.92)	2.12	(1.44,3.12)	2.58	(1.74,3.84)			
RS16	166	0.94	(0.69,1.28)	1.7	(1.16,2.48)	2.07	(1.41,3.03)	2.24	(1.54,3.25)			
RS17	165	0.98	(0.72,1.34)	1.96	(1.33,2.89)	2.63	(1.73,3.98)	3.02	(1.99,4.60)			
RS18	165	0.97	(0.71,1.33)	1.86	(1.26,2.73)	2.71	(1.78,4.13)	3.01	(1.98,4.56)			
RS19	165	0.93	(0.68,1.27)	1.86	(1.27,2.72)	2.4	(1.61,3.58)	2.75	(1.84,4.10)			
RS20	166	1.07	(0.78,1.46)	2.2	(1.48,3.29)	2.47	(1.65,3.69)	3.1	(2.04,4.72)			
RS21	166	1.06	(0.77,1.45)	2.2	(1.47,3.28)	2.48	(1.65,3.71)	3.11	(2.04,4.74)			
RS22	166	1.04	(0.76,1.43)	2.21	(1.48,3.29)	2.47	(1.65,3.70)	3.14	(2.05,4.79)			
RS23	165	1.02	(0.75,1.40)	2.01	(1.36,2.97)	2.52	(1.67,3.79)	2.94	(1.95,4.44)			
RS24	166	1.04	(0.76,1.42)	2.18	(1.46,3.26)	2.52	(1.68,3.78)	3.14	(2.06,4.80)			
RS25	166	1.04	(0.76,1.42)	2.11	(1.42,3.13)	2.45	(1.64,3.67)	3	(1.98,4.54)			

RS26	166	0.99	(0.72,1.35)	2.05	(1.38, 3.04)	2.43	(1.63, 3.65)	2.82	(1.88, 4.21)
RS27	166	0.99	(0.72,1.35)	2.05	(1.38, 3.04)	2.43	(1.63, 3.65)	2.82	(1.88, 4.21)

Table 6A

Model	Time to cR			Upgrading			Significant Upgrading			Significant Upgrading or Upstaging		
	N	Std HR	N	Std OR	95% CI	Std	OR	95% CI	Std	OR	95% CI	Std
RS25	428	2.82	232	2.09	(1.41,3.10)	7.35	(3.87,14.0)	2.55	(1.63,4.00)	4.46	(2.72,7.32)	
Stromal	430	2.05	234	1.32	(0.95,1.84)	3.08	(1.84,5.14)	1.6	(1.12,2.30)	1.95	(1.35,2.82)	
Cellular Organization	430	1.67	234	1.67	(1.16,2.39)	2.83	(1.63,4.90)	1.38	(0.96,1.99)	2.06	(1.37,3.10)	
PSA	430	1.89	234	0.96	(0.70,1.32)	1.38	(0.72,2.63)	1.47	(1.06,2.03)	1.25	(0.83,1.88)	
ECM Cellular Organization	430	2.6	234	2	(1.37,2.93)	11.5	(5.84,22.7)	1.98	(1.34,2.93)	4.01	(2.44,6.58)	
ECM PSA	430	2.45	234	1.17	(0.85,1.61)	2.46	(1.44,4.21)	1.7	(1.21,2.39)	1.76	(1.22,2.53)	
Cellular Organization PSA	430	2.04	234	1.3	(0.92,1.82)	2.52	(1.23,5.16)	1.63	(1.13,2.36)	1.85	(1.19,2.87)	
ECM Cellular Organization TPX2	429	2.61	233	1.89	(1.31,2.72)	11.3	(5.46,23.5)	1.94	(1.31,2.87)	3.99	(2.44,6.54)	
ECM PSA TPX2	429	2.42	233	1.24	(0.90,1.71)	3.25	(1.91,5.51)	1.75	(1.22,2.49)	2.08	(1.45,2.98)	
Cellular Organization PSA TPX2	429	2.04	233	1.33	(0.95,1.86)	3.2	(1.74,5.90)	1.69	(1.17,2.44)	2.21	(1.45,3.37)	
ECM Cellular Organization GSTM2	430	2.67	234	2.03	(1.39,2.96)	11.3	(5.72,22.3)	2.17	(1.43,3.30)	4.35	(2.50,7.58)	
ECM PSA GSTM2	430	2.86	234	1.48	(1.05,2.09)	4.45	(2.03,9.76)	2.2	(1.45,3.34)	2.66	(1.64,4.31)	
Cellular Organization PSA GSTM2	430	2.25	234	1.34	(0.94,1.90)	2.52	(1.18,5.38)	1.92	(1.29,2.84)	2.02	(1.20,3.39)	
ECM Cellular Organization GSTM2	428	2.72	232	2.38	(1.58,3.57)	11.5	(6.02,21.8)	2.48	(1.58,3.87)	5.22	(2.97,9.17)	
TPX2 AZGP1 SRD5A2	428	2.8	232	2.03	(1.38,3.00)	6.65	(3.52,12.6)	2.6	(1.65,4.09)	4.26	(2.58,7.02)	
ECM PSA GSTM2 TPX2 AZGP1 SRD5A2												
Cellular Organization PSA GSTM2 TPX2	428	2.38	232	1.92	(1.28,2.88)	3.63	(2.14,6.15)	2.6	(1.64,4.12)	3.49	(2.08,5.83)	
AZGP1 SRD5A2												

Table 6B

Model	Upgrading						Significant Upgrading						Upstaging						Significant Upgrading or Upstaging					
	Upgrading			Significant Upgrading			Upstaging			Significant Upgrading			Upstaging			Significant Upgrading or Upstaging			Upgrading			Significant Upgrading		
	N	Std OR	95% CI	Std OR	95% CI	Std OR	N	Std OR	95% CI	Std OR	95% CI	Std OR	N	Std OR	95% CI	Std OR	95% CI	Std OR	N	Std OR	95% CI	Std OR	95% CI	Std OR
RS25	166	1.04	(0.76,1.42)	2.11	(1.42,3.13)	2.45	166	1.04	(0.76,1.42)	2.11	(1.42,3.13)	2.45	166	1.04	(0.76,1.42)	2.11	(1.42,3.13)	2.45	166	1.04	(0.76,1.42)	2.11	(1.42,3.13)	2.45
Stromal	166	0.99	(0.73,1.35)	2.19	(1.45,3.32)	1.65	166	0.99	(0.73,1.35)	2.19	(1.45,3.32)	1.65	166	0.99	(0.73,1.35)	2.19	(1.45,3.32)	1.65	166	0.99	(0.73,1.35)	2.19	(1.45,3.32)	1.65
Cellular Organization	167	1.06	(0.77,1.44)	0.93	(0.64,1.36)	1.49	167	1.06	(0.77,1.44)	0.93	(0.64,1.36)	1.49	167	1.06	(0.77,1.44)	0.93	(0.64,1.36)	1.49	167	1.06	(0.77,1.44)	0.93	(0.64,1.36)	1.49
PSA	167	1.04	(0.76,1.42)	1.68	(1.16,2.44)	1.78	167	1.04	(0.76,1.42)	1.68	(1.16,2.44)	1.78	167	1.04	(0.76,1.42)	1.68	(1.16,2.44)	1.78	167	1.04	(0.76,1.42)	1.68	(1.16,2.44)	1.78
ECM Cellular Organization	166	1.04	(0.76,1.42)	1.96	(1.32,2.91)	2.32	166	1.04	(0.76,1.42)	1.96	(1.32,2.91)	2.32	166	1.04	(0.76,1.42)	1.96	(1.32,2.91)	2.32	166	1.04	(0.76,1.42)	1.96	(1.32,2.91)	2.32
ECM PSA	166	1.02	(0.75,1.39)	2.14	(1.44,3.20)	1.84	166	1.02	(0.75,1.39)	2.14	(1.44,3.20)	1.84	166	1.02	(0.75,1.39)	2.14	(1.44,3.20)	1.84	166	1.02	(0.75,1.39)	2.14	(1.44,3.20)	1.84
Cellular Organization PSA	167	1.07	(0.78,1.46)	1.36	(0.94,1.97)	2.06	167	1.07	(0.78,1.46)	1.36	(0.94,1.97)	2.06	167	1.07	(0.78,1.46)	1.36	(0.94,1.97)	2.06	167	1.07	(0.78,1.46)	1.36	(0.94,1.97)	2.06
ECM Cellular Organization TPX2	166	1.15	(0.84,1.58)	2.24	(1.49,3.37)	2.55	166	1.15	(0.84,1.58)	2.24	(1.49,3.37)	2.55	166	1.15	(0.84,1.58)	2.24	(1.49,3.37)	2.55	166	1.15	(0.84,1.58)	2.24	(1.49,3.37)	2.55
ECM PSA TPX2	166	1.2	(0.88,1.65)	2.66	(1.71,4.13)	2.28	166	1.2	(0.88,1.65)	2.66	(1.71,4.13)	2.28	166	1.2	(0.88,1.65)	2.66	(1.71,4.13)	2.28	166	1.2	(0.88,1.65)	2.66	(1.71,4.13)	2.28
Cellular Organization PSA TPX2	167	1.3	(0.95,1.79)	1.77	(1.21,2.60)	2.42	167	1.3	(0.95,1.79)	1.77	(1.21,2.60)	2.42	167	1.3	(0.95,1.79)	1.77	(1.21,2.60)	2.42	167	1.3	(0.95,1.79)	1.77	(1.21,2.60)	2.42
ECM Cellular Organization GSTM2	166	0.96	(0.70,1.30)	1.76	(1.20,2.57)	2.12	166	0.96	(0.70,1.30)	1.76	(1.20,2.57)	2.12	166	0.96	(0.70,1.30)	1.76	(1.20,2.57)	2.12	166	0.96	(0.70,1.30)	1.76	(1.20,2.57)	2.12
ECM PSA GSTM2	166	0.91	(0.67,1.24)	1.69	(1.16,2.46)	1.85	166	0.91	(0.67,1.24)	1.69	(1.16,2.46)	1.85	166	0.91	(0.67,1.24)	1.69	(1.16,2.46)	1.85	166	0.91	(0.67,1.24)	1.69	(1.16,2.46)	1.85
Cellular Organization PSA GSTM2	167	0.89	(0.65,1.22)	1.13	(0.78,1.62)	1.72	167	0.89	(0.65,1.22)	1.13	(0.78,1.62)	1.72	167	0.89	(0.65,1.22)	1.13	(0.78,1.62)	1.72	167	0.89	(0.65,1.22)	1.13	(0.78,1.62)	1.72
ECM Cellular Organization GSTM2 TPX2 AZGP1 SRD5A2	166	1.04	(0.76,1.42)	2.14	(1.44,3.20)	2.47	166	1.04	(0.76,1.42)	2.14	(1.44,3.20)	2.47	166	1.04	(0.76,1.42)	2.14	(1.44,3.20)	2.47	166	1.04	(0.76,1.42)	2.14	(1.44,3.20)	2.47
ECM PSA GSTM2 TPX2 AZGP1 SRD5A2	166	1.03	(0.75,1.41)	2.11	(1.42,3.13)	2.39	166	1.03	(0.75,1.41)	2.11	(1.42,3.13)	2.39	166	1.03	(0.75,1.41)	2.11	(1.42,3.13)	2.39	166	1.03	(0.75,1.41)	2.11	(1.42,3.13)	2.39
Cellular Organization PSA GSTM2 TPX2 AZGP1 SRD5A2	167	1.07	(0.78,1.46)	1.84	(1.26,2.68)	2.22	167	1.07	(0.78,1.46)	1.84	(1.26,2.68)	2.22	167	1.07	(0.78,1.46)	1.84	(1.26,2.68)	2.22	167	1.07	(0.78,1.46)	1.84	(1.26,2.68)	2.22

EXAMPLE 3: CLIQUE STACK ANALYSIS TO IDENTIFY CO-EXPRESSED GENES

[00162] The purpose of the gene clique stacks method described in this Example was to find a set of co-expressed (or surrogate) biomarkers that can be used to reliably predict outcome as well or better than the genes disclosed above. The method used to identify the co-expressed markers is illustrated in Figure 4. The set of co-expressed biomarkers were obtained by seeding the maximal clique enumeration (MCE) with curated biomarkers extracted from the scientific literature. The maximal clique enumeration (MCE) method [Bron et al, 1973] aggregates genes into tightly co-expressed groups such that all of the genes in the group have a similar expression profile. When all of the genes in a group satisfy a minimal similarity condition, the group is called a clique. When a clique is as large as possible without admitting any 'dissimilar' genes into the clique, then the clique is said to be maximal. Using the MCE method, all maximal cliques are searched within a dataset. Using this method, almost any degree of overlap between the maximal cliques can be found, as long as the overlap is supported by the data. Maximal clique enumeration has been shown [Borate et al, 2009] to be an effective way of identifying co-expressed gene modules (CGMs).

1. Definitions

[00163] The following table defines a few terms commonly used in the gene clique stack analyses.

Table 7

Term	Definition
Node	The abundance of a gene (for the purposes of CGM analysis)
Edge	A line connecting two nodes, indicating co-expression of the two nodes
Graph	A collection of nodes and edges
Clique	A graph with an edge connecting all pair-wise combinations of nodes in the graph
maximal clique	A clique that is not contained in any other clique
Stack	A graph obtained by merging at least two cliques or stacks such that the overlap between the two cliques or stacks exceeds some user-defined threshold.
gene expression profile	A two-dimensional matrix, with genes listed down the rows and samples listed across the columns. Each (i,j) entry in the matrix corresponds to relative mRNA abundance for gene i and sample j.

2. Examples of cliques and stacks

[00164] Figure 5 shows a family of three different graphs. A graph consists of nodes (numbered) and connecting edges (lines). Figure 5(a) is not a clique because there is no edge connecting nodes 3 and 4. Figure 5(b) is a clique because there is an edge connecting all pair-

wise combinations of nodes in the graph. Figure 5(c) is a clique, but not a maximal clique because it is contained in clique (b). Given a graph with connecting edges, the MCE algorithm will systematically list all of maximal cliques with 3 or more nodes. For example, the graph in Figure 6 has two maximal cliques: 1-2-3-4-5 and 1-2-3-4-6.

[00165] When based on gene expression data, there are typically large numbers of maximal cliques that are very similar to one another. These maximal cliques can be merged into stacks of maximal cliques. The stacks are the final gene modules of interest and generally are far fewer in number than are the maximal cliques. Figure 7 schematically illustrates stacking of two maximal cliques.

3. Seeding

[00166] For the purposes of finding surrogate co-expressed markers, biomarkers from the literature can be identified and then used to seed the MCE and stacking algorithms. The basic idea is as follows: for each seed, compute a set of maximal cliques (using the parallel MCE algorithm). Then stack the maximal cliques obtained for each seed, yielding a set of seeded stacks. Finally, stack the seeded stacks to obtain a “stack of seeded stacks.” The stack of seeded stacks is an approximation to the stacks that would be obtained by using the conventional (i.e. unseeded) MCE/stacking algorithms. The method used to identify genes that co-express with the genes disclosed above illustrated in Figure 4 and is described in more detail below.

3.1 Seeded MCE algorithm (steps 1-4)

[00167] 1. The process begins by identifying an appropriate set, S_s , of seeding genes. In the instant case, the seeding genes were selected from the gene subsets disclosed above.

[00168] 2. With the seeding genes specified, select a measure of correlation, $R(g_1, g_2)$, between the gene expression profiles of any two genes, g_1, g_2 , along with a correlation threshold below which g_1, g_2 , can be considered uncorrelated. For each seeding gene s in the seeding set S_s , find all gene pairs (s, g) in the dataset such that $R(s, g)$ is greater than or equal to the correlation threshold. Let G_s be the union of s and the set of all genes correlated with s . For the instant study, the Spearman coefficient was used as the measure of correlation and 0.7 as the correlation threshold.

[00169] 3. Compute the correlation coefficient for each pair-wise combination of genes (g_i, g_j) in G_s . Let X_s be the set of all gene pairs for which $R(g_i, g_j)$ is greater than or equal to the correlation threshold. If the genes were plotted as in Figure 5, there would be an edge (line) between each pair of genes in X_s .

[00170] 4. Run the MCE algorithm, as described in Schmidt et al (J. Parallel Distrib. Comput. 69 (2009) 417-428) on the gene pairs X_s for each seeding gene.

3.2 Seeded stacking algorithm (steps 5-6)

[00171] The purpose of stacking is to reduce the number of cliques down to a manageable number of gene modules (stacks). Continuing with steps 5 and 6 of Figure 4:

[00172] 5. For each seeding gene, sort cliques from largest to smallest, i.e. most number of nodes to smallest number of nodes. From the remaining cliques, find the clique with the greatest overlap. If the overlap exceeds a user-specified threshold T , merge the two cliques together to form the first stack. Resort the cliques and stack(s) from largest to smallest and repeat the overlap test and merging. Repeat the process until no new merges occur.

[00173] 6. One now has a set of stacks for each seeding gene. In the final step, all of the seeded stacks are combined into one set of stacks, σ . As the final computation, all of the stacks in σ are stacked, just as in step 5. This stack of stacks is the set of gene modules used for the instant study.

[00174] Genes that were shown to co-express with genes identified by this method are shown in Tables 8-11. "Stack ID" in the Tables is simply an index to enumerate the stacks and "probeWt" refers to the probe weight, or the number of times a probe (gene) appears in the stack.

Table 8

StackID	Coexpressed Gene	ProbeWt	SeedingGene	StackID	Coexpressed Gene	ProbeWt	SeedingGene	StackID	Coexpressed Gene	ProbeWt	SeedingGene
1	SLCO2B1	1	BGN	1	SPARC	1	SPARC	1	SPARC	1	COL4A1
1	LHFP	1	BGN	1	COL4A1	1	SPARC	1	COL4A1	1	COL4A1
1	ENG	1	BGN	1	COL4A2	1	SPARC	1	HTRA1	1	COL4A1
2	LHFP	1	BGN	2	COL3A1	1	SPARC	2	COL4A1	3	COL4A1
2	THY1	1	BGN	2	SPARC	1	SPARC	2	NID1	3	COL4A1
2	ENG	1	BGN	2	COL4A1	1	SPARC	2	CD93	2	COL4A1
3	COL1A1	1	BGN	2	VCAN	1	SPARC	2	FBN1	2	COL4A1
3	THY1	1	BGN	2	FN1	1	SPARC	2	COL1A1	1	COL4A1
3	ENG	1	BGN	3	HEG1	3	SPARC	2	MCAM	1	COL4A1
4	COL1A1	1	BGN	3	MEF2C	3	SPARC	2	SPARC	1	COL4A1
4	PDGFRB	1	BGN	3	RG55	2	SPARC	3	COL1A2	4	COL4A1
4	FMNL3	1	BGN	3	KDR	2	SPARC	3	COL4A1	4	COL4A1
5	SLCO2B1	1	BGN	3	LAMA4	1	SPARC	3	VCAN	2	COL4A1
5	LHFP	1	BGN	3	SPARC	1	SPARC	3	FN1	2	COL4A1
5	COL3A1	1	BGN	4	COL3A1	5	SPARC	3	COL1A1	2	COL4A1
6	THY1	1	BGN	4	SPARC	5	SPARC	3	NID1	2	COL4A1
6	LHFP	1	BGN	4	COL1A1	3	SPARC	3	HTRA1	1	COL4A1
6	COL3A1	1	BGN	4	COL1A2	2	SPARC	3	COL6A3	1	COL4A1
7	THY1	1	BGN	4	BGN	2	SPARC				
7	COL1A1	1	BGN	4	PDGFRB	2	SPARC	1	INHBA	1	INHBA
7	COL3A1	1	BGN	4	COL4A1	1	SPARC	1	STMN2	1	INHBA
8	BGN	7	BGN	4	IGFBP7	1	SPARC	1	COL10A1	1	INHBA
8	COL1A1	4	BGN	4	FBN1	1	SPARC				
8	COL3A1	4	BGN	5	SPARC	4	SPARC	1	THBS2	1	THBS2
8	FMNL3	4	BGN	5	PDGFRB	4	SPARC	1	COL3A1	1	THBS2
8	SLCO2B1	3	BGN	5	DPYSL2	3	SPARC	1	VCAN	1	THBS2

StackID	Coexpressed Gene	ProbeWt	SeedingGene	StackID	Coexpressed Gene	ProbeWt	SeedingGene	StackID	Coexpressed Gene	ProbeWt	SeedingGene	StackID	Coexpressed Gene	ProbeWt	SeedingGene
8	SPARC	3	BGN	5	FBN1	3	SPARC								
8	ENG	3	BGN	5	HEG1	2	SPARC								
8	PDGFRB	3	BGN	5	CDH11	2	SPARC								
8	THBS2	1	BGN	5	FBN5	2	SPARC								
1	THBS2	1	COL3A1	5	LAMA2	2	SPARC								
1	COL3A1	1	COL3A1	5	IGFBP7	2	SPARC								
1	VCAN	1	COL3A1	5	LAMA4	2	SPARC								
2	COL3A1	3	COL3A1	5	RGS5	1	SPARC								
2	SPARC	3	COL3A1	5	COL4A2	1	SPARC								
2	FN1	2	COL3A1	5	COL1A2	1	SPARC								
2	COL4A1	2	COL3A1	6	FBN1	7	SPARC								
2	VCAN	1	COL3A1	6	LAMA4	6	SPARC								
2	COL1A1	1	COL3A1	6	SGK269	5	SPARC								
2	FBN1	1	COL3A1	6	CDH11	5	SPARC								
3	COL1A2	3	COL3A1	6	DPYSL2	5	SPARC								
3	PDGFRB	3	COL3A1	6	LAMA2	5	SPARC								
3	IGFBP7	3	COL3A1	6	SPARC	4	SPARC								
3	FBN1	3	COL3A1	6	SULF1	4	SPARC								
3	CDH11	2	COL3A1	6	FBN5	3	SPARC								
3	AEBP1	2	COL3A1	6	LTBP1	3	SPARC								
3	COL3A1	1	COL3A1	6	EPB41L2	3	SPARC								
3	SPARC	1	COL3A1	6	MEF2C	3	SPARC								
4	COL3A1	5	COL3A1	6	FN1	2	SPARC								
4	BGN	4	COL3A1	6	EDIL3	2	SPARC								
4	COL1A1	3	COL3A1	6	COL3A1	1	SPARC								
4	SPARC	3	COL3A1	6	IGFBP7	1	SPARC								
4	FMN13	2	COL3A1	6	HEG1	1	SPARC								

StackID	Coexpressed Gene	ProbeWt	SeedingGene	StackID	Coexpressed Gene	ProbeWt	SeedingGene	StackID	Coexpressed Gene	ProbeWt	SeedingGene
4	PDGFRB	2	COL3A1								
4	COL1A2	1	COL3A1								
4	THY1	1	COL3A1								
4	THBS2	1	COL3A1								

Table 9

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
1	DDR2	26870	C7	1	MYH11	168	GSTM2	1	PPAP2B	15794	SRD5A2
1	SPARCL1	25953	C7	1	TGFB3	163	GSTM2	1	VWA5A	12616	SRD5A2
1	FAT4	24985	C7	1	RBMS3	162	GSTM2	1	SPON1	12395	SRD5A2
1	SYNE1	24825	C7	1	FHL1	161	GSTM2	1	FAT4	12218	SRD5A2
1	SLC8A1	24327	C7	1	MYLK	158	GSTM2	1	SSPN	12126	SRD5A2
1	MEIS1	23197	C7	1	CACHD1	155	GSTM2	1	MXK	11552	SRD5A2
1	PRRX1	22847	C7	1	TIMP3	154	GSTM2	1	PRRX1	11061	SRD5A2
1	CACHD1	22236	C7	1	SYNM	152	GSTM2	1	LOC645954	10811	SRD5A2
1	DPYSL3	20623	C7	1	NEXN	147	GSTM2	1	SYNM	10654	SRD5A2
1	LTBP1	20345	C7	1	MYL9	142	GSTM2	1	ANXA6	10330	SRD5A2
1	SGK269	19461	C7	1	CRYAB	141	GSTM2	1	PDE5A	10011	SRD5A2
1	EDNRA	19280	C7	1	VWA5A	131	GSTM2	1	TSHZ3	9588	SRD5A2
1	TRPC4	18689	C7	1	AOX1	130	GSTM2	1	GSN	9505	SRD5A2
1	TIMP3	18674	C7	1	FLNC	127	GSTM2	1	NID2	9503	SRD5A2
1	TGFB3	18367	C7	1	PPAP2B	125	GSTM2	1	CLU	9304	SRD5A2
1	ZEB1	18355	C7	1	GSTM2	118	GSTM2	1	TPM2	8659	SRD5A2
1	C1S	16871	C7	1	C21orf63	101	GSTM2	1	FBLN1	8068	SRD5A2
1	ABCC9	16562	C7	1	POPC2	72	GSTM2	1	PARVA	7949	SRD5A2
1	PCDH18	14936	C7	1	TPM2	66	GSTM2	1	SPOCK3	7772	SRD5A2
1	C7	14789	C7	1	CDC42EP3	60	GSTM2	1	PCDH18	7514	SRD5A2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
1	PDGFC	14748	C7	1	CCDC69	58	GSTM2	1	ILK	7078	SRD5A2				
1	PTPLAD2	13590	C7	1	CRISPLD2	52	GSTM2	1	ITIH5	6903	SRD5A2				
1	VCL	13332	C7	1	GBP2	47	GSTM2	1	ADCY5	6374	SRD5A2				
1	MMP2	13107	C7	1	ADCY5	44	GSTM2	1	CRYAB	6219	SRD5A2				
1	FERMT2	12681	C7	1	MATN2	40	GSTM2	1	RBMS3	6108	SRD5A2				
1	EPB41L2	12335	C7	1	AOC3	38	GSTM2	1	AOX1	4943	SRD5A2				
1	PRNP	12133	C7	1	ACACB	36	GSTM2	1	WWTR1	4789	SRD5A2				
1	FBN1	11965	C7	1	RND3	28	GSTM2	1	AOC3	4121	SRD5A2				
1	GLT8D2	11954	C7	1	CLIP4	26	GSTM2	1	CAP2	4091	SRD5A2				
1	DSE	11888	C7	1	POBEC3C	20	GSTM2	1	MAP1B	3917	SRD5A2				
1	SCN7A	11384	C7	1	CAV2	18	GSTM2	1	OGN	3893	SRD5A2				
1	PPAP2B	11121	C7	1	TRIP10	17	GSTM2	1	PLN	3581	SRD5A2				
1	PGR	10566	C7	1	TCF21	11	GSTM2	1	CHL2	2857	SRD5A2				
1	PALID	10240	C7	1	CAMK2G	11	GSTM2	1	MATN2	2808	SRD5A2				
1	CNTN1	10113	C7	1	GSTM5P1	9	GSTM2	1	ADRA1A	2694	SRD5A2				
1	SERPING1	9800	C7	1	ACS3	9	GSTM2	1	BOC	2401	SRD5A2				
1	DKK3	9279	C7	1	GSTM4	7	GSTM2	1	ANGPT1	2290	SRD5A2				
1	CCND2	9131	C7	1	GSTP1	5	GSTM2	1	POPDC2	2205	SRD5A2				
1	MSRB3	8502	C7	1	GSTM1	3	GSTM2	1	FGF2	2162	SRD5A2				
1	LAMA4	8477	C7	1	GSTM3	2	GSTM2	1	TCF21	1996	SRD5A2				
1	RBMS3	8425	C7	1	GSTM2P1	2	GSTM2	1	LOC283904	1983	SRD5A2				
1	FBLN1	7968	C7	1	TGFB3	1	GSTM2	1	DNAJB5	1773	SRD5A2				
1	EPHA3	6930	C7	1	FTO	1	IGF1	1	TSPAN2	1731	SRD5A2				
1	ACTA2	6824	C7	1	UTP11L	1	IGF1	1	GSTM5	1635	SRD5A2				
1	ADAM22	6791	C7	1	SGCB	1	IGF1	1	RGN	1594	SRD5A2				
1	WWTR1	6611	C7	2	CHP	14	IGF1	1	PDLIM7	1503	SRD5A2				
1	HEPH	6406	C7	2	RP2	14	IGF1	1	MTF	1481	SRD5A2				

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
1	TIMP2	6219	C7	2	SPRYD4	14	IGF1	1	BNC2	1300	SRD5A2	1	BNC2	1300	SRD5A2
1	CLIC4	6151	C7	2	SGCB	13	IGF1	1	SCN7A	1274	SRD5A2	1	SCN7A	1274	SRD5A2
1	ATP2B4	5897	C7	2	INMT	13	IGF1	1	GPM6B	1202	SRD5A2	1	GPM6B	1202	SRD5A2
1	TNS1	5842	C7	2	IGF1	12	IGF1	1	ARHGAP20	1193	SRD5A2	1	ARHGAP20	1193	SRD5A2
1	PDGFRA	5802	C7	2	ARPP19	9	IGF1	1	PDZRN4	1190	SRD5A2	1	PDZRN4	1190	SRD5A2
1	ITGA1	5781	C7	2	MOC53	9	IGF1	1	PCP4	1107	SRD5A2	1	PCP4	1107	SRD5A2
1	RHOJ	5103	C7	2	KATNAL1	8	IGF1	1	ANO5	987	SRD5A2	1	ANO5	987	SRD5A2
1	COL14A1	5063	C7	2	C3orf33	8	IGF1	1	C6orf186	930	SRD5A2	1	C6orf186	930	SRD5A2
1	CALD1	4828	C7	2	SLC16A4	7	IGF1	1	ARHGAP10	793	SRD5A2	1	ARHGAP10	793	SRD5A2
1	DCN	4825	C7	2	FTO	7	IGF1	1	CLIP4	775	SRD5A2	1	CLIP4	775	SRD5A2
1	IRAK3	4476	C7	2	SNX27	6	IGF1	1	CCDC69	733	SRD5A2	1	CCDC69	733	SRD5A2
1	MATN2	4448	C7	2	C1orf55	5	IGF1	1	SLC24A3	673	SRD5A2	1	SLC24A3	673	SRD5A2
1	KIT	4329	C7	2	C1orf174	4	IGF1	1	ACS3	668	SRD5A2	1	ACS3	668	SRD5A2
1	NEXN	4257	C7	2	SNTN	4	IGF1	1	IL33	611	SRD5A2	1	IL33	611	SRD5A2
1	ZEB2	3798	C7	2	MCART6	4	IGF1	1	CAMK2G	519	SRD5A2	1	CAMK2G	519	SRD5A2
1	COL6A3	3679	C7	2	OTUD3	4	IGF1	1	PTPLA	505	SRD5A2	1	PTPLA	505	SRD5A2
1	NID2	3678	C7	2	ADAMTS4	4	IGF1	1	EFEMP1	493	SRD5A2	1	EFEMP1	493	SRD5A2
1	PRICKLE2	3671	C7	2	FEZ1	4	IGF1	1	KIT	470	SRD5A2	1	KIT	470	SRD5A2
1	OGN	3418	C7	2	SPATA5	4	IGF1	1	ODZ3	428	SRD5A2	1	ODZ3	428	SRD5A2
1	SSPN	3142	C7	2	ZNRF3	4	IGF1	1	MKGPRF	390	SRD5A2	1	MKGPRF	390	SRD5A2
1	SORBS1	3126	C7	2	C1orf229	4	IGF1	1	C21orf63	383	SRD5A2	1	C21orf63	383	SRD5A2
1	PDE5A	2963	C7	2	STX2	4	IGF1	1	CRISPLD2	322	SRD5A2	1	CRISPLD2	322	SRD5A2
1	LOC732446	2925	C7	2	PURB	4	IGF1	1	MYADM	314	SRD5A2	1	MYADM	314	SRD5A2
1	FCHSD2	2741	C7	2	BVES	4	IGF1	1	C7	278	SRD5A2	1	C7	278	SRD5A2
1	PMP22	2609	C7	2	DTX3L	4	IGF1	1	PDGFRA	219	SRD5A2	1	PDGFRA	219	SRD5A2
1	TRPC1	2519	C7	2	ZNFT13	4	IGF1	1	EYA1	199	SRD5A2	1	EYA1	199	SRD5A2
1	ANXA6	2353	C7	2	DSCR3	4	IGF1	1	ATP1A2	174	SRD5A2	1	ATP1A2	174	SRD5A2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
1	SPON1	2278	C7	2	SLC35F1	4	IGF1	1	ACACB	173	SRD5A2	1	ACACB	173	SRD5A2
1	FBLN5	2115	C7	2	C22orf25	4	IGF1	1	NTSE	168	SRD5A2	1	NTSE	168	SRD5A2
1	CHRD1	1996	C7	2	STK4	4	IGF1	1	GPR124	166	SRD5A2	1	GPR124	166	SRD5A2
1	MEF2C	1980	C7	2	EIF5A2	4	IGF1	1	LOC652799	165	SRD5A2	1	LOC652799	165	SRD5A2
1	EFEMP1	1939	C7	2	SUPT7L	4	IGF1	1	LICH2	123	SRD5A2	1	LICH2	123	SRD5A2
1	JAZF1	1748	C7	2	C10orf78	4	IGF1	1	PYGM	100	SRD5A2	1	PYGM	100	SRD5A2
1	DNAJB4	1636	C7	2	ANKS4B	4	IGF1	1	GSTM2	92	SRD5A2	1	GSTM2	92	SRD5A2
1	ARHGEF6	1594	C7	2	C1orf151	4	IGF1	1	KCNAB1	90	SRD5A2	1	KCNAB1	90	SRD5A2
1	MFAP4	1503	C7	2	RPL32P3	4	IGF1	1	HHIP	82	SRD5A2	1	HHIP	82	SRD5A2
1	LOC652799	1470	C7	2	SEC62	4	IGF1	1	ALDH1A2	70	SRD5A2	1	ALDH1A2	70	SRD5A2
1	PREX2	1464	C7	2	D8R1	4	IGF1	1	PRDM5	63	SRD5A2	1	PRDM5	63	SRD5A2
1	MAN1A1	1433	C7	2	FLJ39639	4	IGF1	1	ABCA8	59	SRD5A2	1	ABCA8	59	SRD5A2
1	TCF21	1224	C7	2	ZNF543	4	IGF1	1	MAML2	51	SRD5A2	1	MAML2	51	SRD5A2
1	CRIM1	1181	C7	2	FRRS1	4	IGF1	1	PAK3	38	SRD5A2	1	PAK3	38	SRD5A2
1	A2M	1168	C7	2	TATDN3	4	IGF1	1	SNAI2	35	SRD5A2	1	SNAI2	35	SRD5A2
1	DPYSL2	1029	C7	2	WDR55	4	IGF1	1	UST	27	SRD5A2	1	UST	27	SRD5A2
1	GPM6B	993	C7	2	KIAA1737	4	IGF1	1	TMLHE	21	SRD5A2	1	TMLHE	21	SRD5A2
1	PLN	970	C7	2	APOBEC3F	4	IGF1	1	ACTC1	15	SRD5A2	1	ACTC1	15	SRD5A2
1	IL33	942	C7	2	RNF7	4	IGF1	1	C5orf4	8	SRD5A2	1	C5orf4	8	SRD5A2
1	CCDC80	889	C7	2	SIKE1	4	IGF1	1	GSTM5P1	4	SRD5A2	1	GSTM5P1	4	SRD5A2
1	LMO3	852	C7	2	HSP90B3P	4	IGF1	1	GSTM4	3	SRD5A2	1	GSTM4	3	SRD5A2
1	SEC23A	765	C7	2	GNS	4	IGF1	1	PKD4	2	SRD5A2	1	PKD4	2	SRD5A2
1	MOXD1	708	C7	2	C1orf212	4	IGF1	1	TGFB3	2	SRD5A2	1	TGFB3	2	SRD5A2
1	SPOCK3	622	C7	2	ZNF70	4	IGF1	1	GSTM1	1	SRD5A2	1	GSTM1	1	SRD5A2
1	HEG1	608	C7	2	TMEM127	4	IGF1	1	LOC728846	1	TGFB11	1	LOC728846	1	TGFB11
1	LUM	589	C7	2	ALDH1B1	4	IGF1	1	CLIP3	1	TGFB11	1	CLIP3	1	TGFB11
1	C7orf58	566	C7	2	HP1BP3	4	IGF1	1	EMILIN1	1	TGFB11	1	EMILIN1	1	TGFB11

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
1	CDC42EP3	539	C7	2	APOE6	4	IGF1	2	CLIP3	1	TGFB111
1	CPVL	524	C7	2	MALL	4	IGF1	2	MRC2	1	TGFB111
1	CPA3	421	C7	2	C11orf17	4	IGF1	2	MEG3	1	TGFB111
1	SLIT2	417	C7	2	LOC729199	4	IGF1	3	MRC2	1	TGFB111
1	KLHL5	376	C7	2	RELL1	4	IGF1	3	LCAT	1	TGFB111
1	HLF	322	C7	2	PELI1	4	IGF1	3	MEG3	1	TGFB111
1	PLXDC2	313	C7	2	ASB6	4	IGF1	4	LD83	18	TGFB111
1	CAP2	301	C7	2	C2orf18	4	IGF1	4	TGFB111	15	TGFB111
1	FXYD6	291	C7	2	PSTPIP2	4	IGF1	4	ASB2	11	TGFB111
1	ECM2	272	C7	2	CLEC7A	4	IGF1	4	CLIP3	11	TGFB111
1	SRD5A2	245	C7	2	RAB22A	4	IGF1	4	ITGA7	10	TGFB111
1	MBNL1	245	C7	2	LOC643770	4	IGF1	4	JPH2	10	TGFB111
1	LAMA2	169	C7	2	LOC100129502	4	IGF1	4	RUSC2	10	TGFB111
1	IL6ST	166	C7	2	ZCCHC4	4	IGF1	4	HRNP3	8	TGFB111
1	PODN	112	C7	2	PNMA2	4	IGF1	4	LIMS2	8	TGFB111
1	ATRNL1	110	C7	2	PIGW	4	IGF1	4	CSPG4	7	TGFB111
1	DOCK11	60	C7	2	SLC25A32	4	IGF1	4	NLGN3	5	TGFB111
1	FGL2	56	C7	2	CLCC1	4	IGF1	4	ADAM33	3	TGFB111
1	SPRY2	12	C7	2	KIAA0513	4	IGF1	4	NHSL2	3	TGFB111
1	OLFML1	12	C7	2	SS18	4	IGF1	4	SYDE1	2	TGFB111
1	NEGR1	4	C7	2	CECR1	4	IGF1	4	RASL12	2	TGFB111
1	IGFBP5	1	C7	2	ZNF490	4	IGF1	4	LOC90586	2	TGFB111
1	SORBS1	1	DES	2	PDE12	4	IGF1	4	GNAZ	1	TGFB111
1	CACNA1C	1	DES	2	C10orf76	4	IGF1	4	TMEM35	1	TGFB111
1	DES	1	DES	2	CCL22	4	IGF1	4	LCAT	1	TGFB111
2	ITIH5	1	DES	2	RRN3P1	4	IGF1	4	LOC728846	1	TGFB111
2	ANXA6	1	DES	2	LOC100127925	4	IGF1	4	SLC24A3	1	TGFB111

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
2	ATP1A2	1	DES	2	SC4MOL	4	IGF1	5	MRGPRF	381	TGFB11I				
3	ITIH5	1	DES	2	AP4E1	4	IGF1	5	PDLIM7	362	TGFB11I				
3	DES	1	DES	2	APOLD1	4	IGF1	5	AOC3	321	TGFB11I				
3	ANXA6	1	DES	2	ARSB	4	IGF1	5	ADCY5	317	TGFB11I				
4	TPM1	1	DES	2	ZNF264	4	IGF1	5	KANK2	306	TGFB11I				
4	DES	1	DES	2	SLC30A6	4	IGF1	5	SLC24A3	292	TGFB11I				
4	CES1	1	DES	2	METTL7A	4	IGF1	5	MYL9	287	TGFB11I				
5	TAGLN	72309	DES	2	PARD6B	4	IGF1	5	FLNC	275	TGFB11I				
5	FLNA	72305	DES	2	STOM	4	IGF1	5	TGFB11I	253	TGFB11I				
5	TNS1	72049	DES	2	CYP20A1	4	IGF1	5	ITGA7	222	TGFB11I				
5	CNN1	69837	DES	2	LVZ	4	IGF1	5	DES	216	TGFB11I				
5	ACTA2	68389	DES	2	ATP1B4	4	IGF1	5	FLNA	214	TGFB11I				
5	CHRD1	67725	DES	2	SCD5	4	IGF1	5	EFEMP2	206	TGFB11I				
5	DPYSL3	67225	DES	2	CEP170L	4	IGF1	5	TAGLN	184	TGFB11I				
5	M5R83	66488	DES	2	NUDT19	4	IGF1	5	RASL12	163	TGFB11I				
5	VCL	65707	DES	2	TXNL4B	4	IGF1	5	GAS6	163	TGFB11I				
5	CCND2	65291	DES	2	APPL1	4	IGF1	5	KCNMB1	163	TGFB11I				
5	SLC8A1	65217	DES	2	OSBP2	4	IGF1	5	SMTN	157	TGFB11I				
5	MEIS1	65097	DES	2	VMA21	4	IGF1	5	GPR124	140	TGFB11I				
5	ATP2B4	64428	DES	2	NEF2	4	IGF1	5	COL6A1	133	TGFB11I				
5	DDR2	64293	DES	2	ZNF772	4	IGF1	5	DNAJB5	127	TGFB11I				
5	LMOD1	64271	DES	2	LOC646973	4	IGF1	5	COL6A2	124	TGFB11I				
5	SORBS1	63359	DES	2	LOC100128096	4	IGF1	5	TPM2	121	TGFB11I				
5	KCNMB1	61499	DES	2	MOAP1	4	IGF1	5	WFDC1	121	TGFB11I				
5	PGR	60803	DES	2	HIGD1A	4	IGF1	5	TNS1	112	TGFB11I				
5	RBPMS	59947	DES	2	DISC2	4	IGF1	5	DKK3	111	TGFB11I				
5	FLNC	59840	DES	2	CYCS	4	IGF1	5	HSPB8	108	TGFB11I				

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
5	MYLK	58329	DES	2	ZSCAN22	4	IGF1	5	TSPAN18	103	TGFB111
5	FHL1	58303	DES	2	LOC646127	4	IGF1	5	MYH11	102	TGFB111
5	FZD7	56889	DES	2	RRP15	4	IGF1	5	GEFT	90	TGFB111
5	EDNRA	56620	DES	2	LOC100130357	4	IGF1	5	ITIH5	81	TGFB111
5	DKK3	56591	DES	2	YES1	4	IGF1	5	PYGM	81	TGFB111
5	DES	54990	DES	2	MTFMT	4	IGF1	5	MCAM	78	TGFB111
5	PGM5	54713	DES	2	JOSD1	4	IGF1	5	MRV11	75	TGFB111
5	LOC729468	53979	DES	2	RHOF	4	IGF1	5	MYLK	68	TGFB111
5	SYNE1	53386	DES	2	LINS4	4	IGF1	5	CNN1	63	TGFB111
5	PGMSP2	53378	DES	2	LOC729142	4	IGF1	5	RBPMS2	63	TGFB111
5	SPARCL1	52082	DES	2	GNG4	4	IGF1	5	ATP1A2	58	TGFB111
5	ACTG2	51556	DES	2	HGPD	4	IGF1	5	LIMS2	58	TGFB111
5	TRPC4	51205	DES	2	FBXW2	4	IGF1	5	LMOD1	56	TGFB111
5	CAV1	49615	DES	2	NUP43	4	IGF1	5	GNAO1	46	TGFB111
5	GNAL	49292	DES	2	WDR5B	4	IGF1	5	LGALS1	43	TGFB111
5	TIMP3	48293	DES	2	ANGEL2	4	IGF1	5	DAAM2	41	TGFB111
5	ABCC9	46190	DES	2	SGTB	4	IGF1	5	MRC2	39	TGFB111
5	MRV11	44926	DES	2	MAPK1IP1L	4	IGF1	5	HRNBP3	38	TGFB111
5	ACTN1	44120	DES	2	ZSCAN29	4	IGF1	5	ASB2	36	TGFB111
5	PALLD	43624	DES	2	FXC1	4	IGF1	5	CLIP3	25	TGFB111
5	SERPINF1	43602	DES	2	NQO1	4	IGF1	5	C16orf45	22	TGFB111
5	JAZF1	42715	DES	2	MOBK1A	4	IGF1	5	DBNDD2	20	TGFB111
5	KANK2	42364	DES	2	ANAPC16	4	IGF1	5	RUSC2	19	TGFB111
5	HSPB8	41435	DES	2	C16orf63	4	IGF1	5	RARRES2	18	TGFB111
5	MYL9	37460	DES	2	TBCCD1	4	IGF1	5	ADRA1A	18	TGFB111
5	PRNP	33800	DES	2	DLEU2	4	IGF1	5	TINAGL1	17	TGFB111
5	TSPAN18	33287	DES	2	CARD8	4	IGF1	5	SYNM	17	TGFB111

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
5	FRMD6	32935	DES	2	LOC100130236	4	IGF1	5	TMEM35	14	TGFB11
5	CSRPI	32471	DES	2	LOC100130442	4	IGF1	5	COP22	12	TGFB11
5	HEPH	32337	DES	2	CAMIG	4	IGF1	5	LTBP4	12	TGFB11
5	NEXN	29867	DES	2	ZBTB3	4	IGF1	5	SCARA3	11	TGFB11
5	PRICKLE2	29746	DES	2	ZNF445	4	IGF1	5	NR2F1	11	TGFB11
5	PPAP2B	28983	DES	2	CASP8	4	IGF1	5	PCDH10	11	TGFB11
5	MYH11	28923	DES	2	RAB21	4	IGF1	5	RAB34	10	TGFB11
5	PDGFC	28732	DES	2	ZC3HAV1L	4	IGF1	5	FOXF1	8	TGFB11
5	TPM1	27766	DES	2	SC5D1	4	IGF1	5	TCF7L1	7	TGFB11
5	SVIL	27521	DES	2	KILLIN	4	IGF1	5	KIRREL	6	TGFB11
5	LOC732446	27335	DES	2	MTX3	4	IGF1	5	DACT1	6	TGFB11
5	MEIS2	25944	DES	2	KCNE4	4	IGF1	5	ZNF516	5	TGFB11
5	CALD1	25386	DES	2	GM2A	4	IGF1	5	EMILIN1	4	TGFB11
5	CNTN1	25377	DES	2	LOC401588	4	IGF1	5	DCHS1	4	TGFB11
5	FERMT2	25146	DES	2	C8orf79	4	IGF1	5	EHBP1L1	3	TGFB11
5	CLU	24888	DES	2	KIAA0754	4	IGF1	5	SYDE1	2	TGFB11
5	SPON1	23171	DES	2	SMU1	4	IGF1	5	PPP1R14A	2	TGFB11
5	TGFBR3	23018	DES	2	TSPYL1	4	IGF1	5	SMOC1	2	TGFB11
5	CACHD1	22496	DES	2	SPRED1	4	IGF1	5	JPH2	1	TGFB11
5	TPM2	22108	DES	2	LOC100128997	4	IGF1	5	MICALL1	1	TGFB11
5	GSN	22102	DES	2	LOC729652	4	IGF1	5	LCAT	1	TGFB11
5	NID2	21240	DES	2	TRAPPC2	4	IGF1	5	HSPB6	1	TGFB11
5	MYOCD	21178	DES	2	KCTD10	4	IGF1	1	FLNA	33418	TPM2
5	MXK	20028	DES	2	DUSP19	4	IGF1	1	TAGLN	33391	TPM2
5	EYA4	19967	DES	2	CCDC122	4	IGF1	1	TNSI	32975	TPM2
5	LOC100127983	18208	DES	2	NXN	4	IGF1	1	CNN1	32489	TPM2
5	ANXA6	16600	DES	2	ZNF283	4	IGF1	1	CHRD1	31765	TPM2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
5	HLF	16262	DES	2	SPATS2L	4	IGF1	1	LMOD1	31568	TPM2
5	VWASA	16175	DES	2	TRIM5	4	IGF1	1	MYLK	31444	TPM2
5	SRD5A2	16145	DES	2	HAUS3	4	IGF1	1	ACTA2	31310	TPM2
5	SYNM	15943	DES	2	UTP11L	4	IGF1	1	ACTG2	30665	TPM2
5	CDC42EP3	14001	DES	2	SLC30A5	4	IGF1	1	KCNMB1	30331	TPM2
5	AOC3	13787	DES	2	MBOAT1	4	IGF1	1	MSRB3	30007	TPM2
5	TIMP2	13760	DES	2	TERF2	4	IGF1	1	SORBS1	29926	TPM2
5	ILK	13444	DES	2	VPS33A	4	IGF1	1	DPYSL3	29802	TPM2
5	ADCY5	13346	DES	2	SENP5	4	IGF1	1	DES	29158	TPM2
5	PARVA	13266	DES	2	EVIS	4	IGF1	1	VCL	29088	TPM2
5	FBLN1	12617	DES	2	NDUFC2	4	IGF1	1	SLC8A1	29075	TPM2
5	LOC645954	12259	DES	2	ZBTB8A	4	IGF1	1	CCND7	28780	TPM2
5	FAT4	12247	DES	2	ST8SIA4	4	IGF1	1	MEIS1	28764	TPM2
5	ITIH5	11490	DES	2	C7orf64	4	IGF1	1	PGM5	28584	TPM2
5	COL6A3	10595	DES	2	MED18	4	IGF1	1	ATP2B4	28495	TPM2
5	TSHZ3	10118	DES	2	MPV17L	4	IGF1	1	LOC729468	28204	TPM2
5	MCAM	8671	DES	2	C1orf210	4	IGF1	1	FHL1	28101	TPM2
5	MAP1B	8478	DES	2	LIN7C	4	IGF1	1	FLNC	27926	TPM2
5	WFDC1	7000	DES	2	KCNJ11	4	IGF1	1	PGM5P2	27789	TPM2
5	PDE5A	6648	DES	2	COX18	4	IGF1	1	HSPB8	27438	TPM2
5	TLN1	5948	DES	2	PCBD2	4	IGF1	1	DDR2	26679	TPM2
5	PDIM7	5715	DES	2	SPAST	4	IGF1	1	PGR	26409	TPM2
5	SPOCK3	5657	DES	2	CYP4V2	4	IGF1	1	MRV11	25979	TPM2
5	BOC	5611	DES	2	LRTOMT	4	IGF1	1	DKK3	25603	TPM2
5	CRYAB	5555	DES	2	IMPAD1	3	IGF1	1	RBPMS	24576	TPM2
5	PMP22	4795	DES	2	UBXN2B	3	IGF1	1	MYH11	24353	TPM2
5	ADRA1A	4611	DES	2	CSorf33	3	IGF1	1	FZD7	24298	TPM2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
5	FGF2	4439	DES	2	FOXJ3	3	IGF1	1	TPM2	23458	TPM2
5	CELF2	4392	DES	2	PPP1R15B	3	IGF1	1	GNAL	23091	TPM2
5	MMP2	4243	DES	2	GNAI3	2	IGF1	1	MYL9	22987	TPM2
5	WWTR1	3966	DES	2	SAR1B	2	IGF1	1	JAZF1	21665	TPM2
5	CAP2	3592	DES	2	SERPINB9	2	IGF1	1	CAV1	21569	TPM2
5	LOC100129846	3236	DES	2	PTGIS	2	IGF1	1	KANK2	21564	TPM2
5	RBMS3	3165	DES	2	C3orf70	2	IGF1	1	EDNRA	20876	TPM2
5	AOX1	3042	DES	2	RUNDC2B	2	IGF1	1	SPARCL1	20468	TPM2
5	MFAP4	3011	DES	2	SYT11	1	IGF1	1	TRPC4	19698	TPM2
5	TCF21	2881	DES	1	CPXM2	1	ITGA7	1	TSPAN18	18763	TPM2
5	MATN2	2851	DES	1	MRV1	1	ITGA7	1	ACTN1	18284	TPM2
5	MIRGPRF	2724	DES	1	ITGA7	1	ITGA7	1	TIMP3	18017	TPM2
5	POPDC2	2704	DES	2	ADCY5	661	ITGA7	1	ABCC9	17793	TPM2
5	CFL2	2404	DES	2	MIRGPRF	652	ITGA7	1	SYNE1	17659	TPM2
5	LOC283904	2374	DES	2	PDLIM7	649	ITGA7	1	SERPINF1	17306	TPM2
5	PRELP	2253	DES	2	FLNC	627	ITGA7	1	PALLD	16659	TPM2
5	CCDC69	2088	DES	2	KANK2	624	ITGA7	1	PRICKLE2	16570	TPM2
5	PLN	2046	DES	2	MYL9	611	ITGA7	1	CSRP1	15853	TPM2
5	DNAJB5	1956	DES	2	AOC3	602	ITGA7	1	HEPH	14646	TPM2
5	GPR124	1851	DES	2	FLNA	540	ITGA7	1	NEXN	13548	TPM2
5	GAS6	1830	DES	2	TAGLN	527	ITGA7	1	MYOCD	13479	TPM2
5	TSPAN2	1830	DES	2	KCNMB1	492	ITGA7	1	MEIS2	13043	TPM2
5	ANGPT1	1797	DES	2	DES	491	ITGA7	1	TPM1	12988	TPM2
5	MFGE8	1766	DES	2	ITGA7	481	ITGA7	1	SPON1	12334	TPM2
5	ITGA1	1682	DES	2	SIC24A3	434	ITGA7	1	EYA4	12112	TPM2
5	GSTM5	1596	DES	2	TNS1	423	ITGA7	1	HLF	11972	TPM2
5	MYADM	1579	DES	2	TSPAN18	364	ITGA7	1	SYNM	11833	TPM2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
5	CES1	1511	DES	2	MCAM	351	ITGA7	1	SVIL	11249	TPM2				
5	CAMK2G	1453	DES	2	TPM2	322	ITGA7	1	FRMD6	10974	TPM2				
5	PCP4	1361	DES	2	MYLK	322	ITGA7	1	CNTN1	10796	TPM2				
5	SLC24A3	1275	DES	2	HSP88	317	ITGA7	1	CLU	10687	TPM2				
5	RGN	1215	DES	2	MVH11	317	ITGA7	1	LOC100127983	10582	TPM2				
5	KCNMA1	1050	DES	2	MRV11	314	ITGA7	1	PRNP	10088	TPM2				
5	PDZRN4	876	DES	2	LMOD1	301	ITGA7	1	MKX	9903	TPM2				
5	ARHGAP10	867	DES	2	CNN1	288	ITGA7	1	CALD1	9712	TPM2				
5	C6orf186	841	DES	2	ITIH5	287	ITGA7	1	FERMT2	9315	TPM2				
5	ARHGAP20	828	DES	2	DNAJB5	282	ITGA7	1	NID2	9290	TPM2				
5	FXYD6	826	DES	2	CHRD11	264	ITGA7	1	ITIH5	8936	TPM2				
5	PTGER2	802	DES	2	EFEMP2	256	ITGA7	1	PDGFC	8919	TPM2				
5	SLC12A4	721	DES	2	ATP1A2	239	ITGA7	1	LOC732446	8793	TPM2				
5	NID1	670	DES	2	SMTN	238	ITGA7	1	LOC645954	8764	TPM2				
5	ITGA9	568	DES	2	GAS6	231	ITGA7	1	ADCY5	8698	TPM2				
5	SMTN	558	DES	2	WFDC1	222	ITGA7	1	AOC3	8557	TPM2				
5	TCEAL2	557	DES	2	TGFB111	220	ITGA7	1	SRD5A2	8415	TPM2				
5	COL6A1	499	DES	2	GPR124	206	ITGA7	1	GSN	7427	TPM2				
5	ITGA5	475	DES	2	NID2	204	ITGA7	1	WFDC1	6345	TPM2				
5	ATP1A2	417	DES	2	ADRA1A	197	ITGA7	1	VWA5A	6297	TPM2				
5	C21orf63	408	DES	2	PYGM	189	ITGA7	1	ILK	6243	TPM2				
5	EFEMP2	389	DES	2	RASL12	186	ITGA7	1	TGFB3	5718	TPM2				
5	PTPLA	366	DES	2	BOC	184	ITGA7	1	CDC42EP3	5544	TPM2				
5	ST5	364	DES	2	FZD7	174	ITGA7	1	TSHZ3	5478	TPM2				
5	JAM3	350	DES	2	ACTG2	172	ITGA7	1	FAT4	4923	TPM2				
5	ITGA7	333	DES	2	PRICKLE2	157	ITGA7	1	PARVA	4922	TPM2				
5	LPP	320	DES	2	GEFT	156	ITGA7	1	MCAM	4880	TPM2				

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
5	COL6A2	302	DES	2	COL6A1	142	ITGA7	1	PDLIM7	4753	TPM2	1	PDLIM7	4753	TPM2
5	ODZ3	294	DES	2	PGM5	133	ITGA7	1	ADRA1A	4540	TPM2	1	ADRA1A	4540	TPM2
5	PLEKHO1	266	DES	2	SYNM	132	ITGA7	1	ANXA6	4499	TPM2	1	ANXA6	4499	TPM2
5	PYGM	249	DES	2	FHL1	126	ITGA7	1	FBLN1	4133	TPM2	1	FBLN1	4133	TPM2
5	TINAGL1	239	DES	2	HEPH	112	ITGA7	1	BOC	3515	TPM2	1	BOC	3515	TPM2
5	PCDH10	238	DES	2	COL6A2	110	ITGA7	1	COL6A3	3490	TPM2	1	COL6A3	3490	TPM2
5	PNMA1	232	DES	2	LOC729468	109	ITGA7	1	CRYAB	3436	TPM2	1	CRYAB	3436	TPM2
5	ACACB	221	DES	2	MYOCD	101	ITGA7	1	SPOCK3	3141	TPM2	1	SPOCK3	3141	TPM2
5	RASL12	213	DES	2	ACTA2	66	ITGA7	1	PDE5A	2530	TPM2	1	PDE5A	2530	TPM2
5	LARGE	182	DES	2	RBPMS2	62	ITGA7	1	MAP18	2406	TPM2	1	MAP18	2406	TPM2
5	GEFT	181	DES	2	LIMS2	53	ITGA7	1	FGF2	2375	TPM2	1	FGF2	2375	TPM2
5	NCS1	176	DES	2	GNAO1	45	ITGA7	1	LOC100129846	2231	TPM2	1	LOC100129846	2231	TPM2
5	TRANK1	173	DES	2	ASB2	44	ITGA7	1	MARGPRF	2029	TPM2	1	MARGPRF	2029	TPM2
5	FGFR1	166	DES	2	HRNP3	43	ITGA7	1	DNAJB5	2029	TPM2	1	DNAJB5	2029	TPM2
5	AHNAK2	164	DES	2	PODPC2	41	ITGA7	1	LOC283904	2007	TPM2	1	LOC283904	2007	TPM2
5	LGALS1	156	DES	2	DAAM2	38	ITGA7	1	PODPC2	1965	TPM2	1	PODPC2	1965	TPM2
5	RRAS	133	DES	2	ODZ3	34	ITGA7	1	TCF21	1785	TPM2	1	TCF21	1785	TPM2
5	C2orf40	132	DES	2	PDZRN4	33	ITGA7	1	TLN1	1720	TPM2	1	TLN1	1720	TPM2
5	TGFB1I1	126	DES	2	C6orf186	30	ITGA7	1	CELF2	1700	TPM2	1	CELF2	1700	TPM2
5	RAB34	95	DES	2	ITGA9	28	ITGA7	1	AOX1	1459	TPM2	1	AOX1	1459	TPM2
5	PTRF	94	DES	2	NID1	27	ITGA7	1	SLC24A3	1296	TPM2	1	SLC24A3	1296	TPM2
5	SCHIP1	91	DES	2	C16orf45	22	ITGA7	1	CDC69	1287	TPM2	1	CDC69	1287	TPM2
5	GSTM2	87	DES	2	RUSC2	22	ITGA7	1	ANGPT1	1256	TPM2	1	ANGPT1	1256	TPM2
5	MAOB	49	DES	2	TMEM35	19	ITGA7	1	PCP4	1226	TPM2	1	PCP4	1226	TPM2
5	MASP1	48	DES	2	CLIP3	19	ITGA7	1	BNC2	1170	TPM2	1	BNC2	1170	TPM2
5	TRIP10	45	DES	2	MRC2	19	ITGA7	1	PDZRN4	1069	TPM2	1	PDZRN4	1069	TPM2
5	RARRES2	40	DES	2	TINAGL1	17	ITGA7	1	RGN	1065	TPM2	1	RGN	1065	TPM2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
5	RBPMS2	37	DES	2	DBNDD2	17	ITGA7	1	CES1	1060	TPM2
5	APOBEC3C	30	DES	2	ITGB3	11	ITGA7	1	GPR124	917	TPM2
5	COPZ2	29	DES	2	LDB3	9	ITGA7	1	GAS6	888	TPM2
5	CACNA1C	21	DES	2	ITGA5	9	ITGA7	1	CHL2	871	TPM2
5	GNAO1	16	DES	2	NCS1	9	ITGA7	1	CAMK2G	869	TPM2
5	UST	12	DES	2	FOXF1	8	ITGA7	1	ARHGAP20	850	TPM2
5	ACTC1	12	DES	2	DACT1	7	ITGA7	1	GSTM5	794	TPM2
5	CES4	11	DES	2	CSPG4	6	ITGA7	1	CAP2	752	TPM2
5	ID4	11	DES	2	JPH2	6	ITGA7	1	PRELP	693	TPM2
5	C16orf45	10	DES	2	ZNF516	6	ITGA7	1	SMTN	540	TPM2
5	LIMS2	9	DES	2	KIRREL	3	ITGA7	1	FXYD6	533	TPM2
5	GSTM5P1	6	DES	2	NHSL2	3	ITGA7	1	TSPAN2	500	TPM2
5	GSTM4	5	DES	2	LCAT	2	ITGA7	1	KCNMA1	488	TPM2
5	CBX7	3	DES	2	FABP3	2	ITGA7	1	PTGER2	429	TPM2
5	PPP1R14A	3	DES	2	GNAZ	1	ITGA7	1	TCEAL2	425	TPM2
5	FABP3	3	DES	2	P2RX1	1	ITGA7	1	MYADM	402	TPM2
5	GSTM1	2	DES	1	SLC8A1	47139	SRD5A2	1	JAM3	360	TPM2
5	GSTM2P1	2	DES	1	LOC729468	47056	SRD5A2	1	COL6A1	354	TPM2
5	HSPB6	1	DES	1	DPYSL3	47002	SRD5A2	1	ATP1A2	339	TPM2
1	GSTM5P1	4	GSTM1	1	ACTA2	46967	SRD5A2	1	SLC12A4	327	TPM2
1	GSTM2	4	GSTM1	1	PGM5	46874	SRD5A2	1	ITGA5	325	TPM2
1	GSTM4	4	GSTM1	1	MEIS1	46871	SRD5A2	1	ITGA9	301	TPM2
1	GSTM5	4	GSTM1	1	ACTG2	46703	SRD5A2	1	ITGA7	300	TPM2
1	SPOCK3	3	GSTM1	1	PGM5P2	46699	SRD5A2	1	EFEMP2	298	TPM2
1	PGM5	3	GSTM1	1	MSRB3	46428	SRD5A2	1	PYGM	254	TPM2
1	HSPB8	3	GSTM1	1	TAGLN	46404	SRD5A2	1	COL6A2	248	TPM2
1	AOX1	2	GSTM1	1	FLNA	46365	SRD5A2	1	ARHGAP10	227	TPM2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
1	CSRPI	2	GSTM1	1	VCL	46278	SRD5A2	1	PNMA1	211	TPM2
1	FLNC	2	GSTM1	1	CNN1	45892	SRD5A2	1	RASL12	207	TPM2
1	DES	2	GSTM1	1	CHRD1	45879	SRD5A2	1	GEFT	194	TPM2
1	GSTM2P1	2	GSTM1	1	TNS1	45774	SRD5A2	1	PTPLA	183	TPM2
1	GSTM1	2	GSTM1	1	ATP2B4	45519	SRD5A2	1	CRISPLD2	181	TPM2
1	CAV1	1	GSTM1	1	LMOD1	44299	SRD5A2	1	ACSS3	177	TPM2
1	SRD5A2	1	GSTM1	1	PGR	44126	SRD5A2	1	AHNAK2	175	TPM2
1	GSTM3	1	GSTM1	1	SORBS1	43839	SRD5A2	1	ST5	175	TPM2
1	LOC729468	1	GSTM1	1	CCND2	43401	SRD5A2	1	PLEKHO1	165	TPM2
1	EVA4	1	GSTM1	1	DOR2	43389	SRD5A2	1	LARGE	164	TPM2
1	PGM5P2	1	GSTM1	1	EDNRA	42947	SRD5A2	1	C21orf63	151	TPM2
1	CSRPI	364	GSTM2	1	FHL1	41382	SRD5A2	1	TINAGL1	150	TPM2
1	CAV1	358	GSTM2	1	KCNMB1	41204	SRD5A2	1	ACACB	138	TPM2
1	TNS1	358	GSTM2	1	TRPC4	40884	SRD5A2	1	LGALS1	136	TPM2
1	ATP2B4	356	GSTM2	1	SYNE1	40118	SRD5A2	1	TGFB111	126	TPM2
1	MEIS2	352	GSTM2	1	CAV1	39836	SRD5A2	1	ITGB3	123	TPM2
1	FLNA	350	GSTM2	1	SPARCL1	39359	SRD5A2	1	RRAS	123	TPM2
1	TAGLN	350	GSTM2	1	RBPMS	38414	SRD5A2	1	NCS1	107	TPM2
1	GNAL	350	GSTM2	1	FZD7	34246	SRD5A2	1	PTRF	94	TPM2
1	DPYSL3	348	GSTM2	1	SRD5A2	33968	SRD5A2	1	LPP	91	TPM2
1	MEIS1	347	GSTM2	1	DNK3	33963	SRD5A2	1	C2orf40	90	TPM2
1	TRPC4	345	GSTM2	1	JAZF1	33635	SRD5A2	1	MAOB	59	TPM2
1	CCND2	325	GSTM2	1	MYLK	33158	SRD5A2	1	GSTM2	51	TPM2
1	SYNE1	321	GSTM2	1	ABCC9	33072	SRD5A2	1	TRIP10	48	TPM2
1	EDNRA	317	GSTM2	1	GNAL	32392	SRD5A2	1	ALDH1A2	43	TPM2
1	ACTA2	313	GSTM2	1	PALLD	31713	SRD5A2	1	RARRES2	36	TPM2
1	PALLD	310	GSTM2	1	FLNC	29309	SRD5A2	1	COPZ2	34	TPM2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
1	FRMD6	309	GSTM2	1	PRICKLE2	29168	SRD5A2	1	APOBEC3C	34	TPM2
1	PGM5	301	GSTM2	1	MRVI1	28467	SRD5A2	1	RBPMS2	33	TPM2
1	HSPB8	293	GSTM2	1	TIMP3	28313	SRD5A2	1	DBNDD2	31	TPM2
1	ACTG2	287	GSTM2	1	FRMD6	28108	SRD5A2	1	GNAO1	19	TPM2
1	CNN1	286	GSTM2	1	PRNP	28106	SRD5A2	1	ACTC1	15	TPM2
1	PGMSP2	285	GSTM2	1	HSPB8	26756	SRD5A2	1	CES4	11	TPM2
1	SLC8A1	282	GSTM2	1	PDGFC	26571	SRD5A2	1	C16orf45	10	TPM2
1	LOC729468	276	GSTM2	1	CNTN1	26148	SRD5A2	1	GSTP1	8	TPM2
1	PRICKLE2	275	GSTM2	1	EYA4	26129	SRD5A2	1	UST	5	TPM2
1	SRD5A2	275	GSTM2	1	MEIS2	25616	SRD5A2	1	GSTM4	4	TPM2
1	RBPMS	275	GSTM2	1	MYOCD	25477	SRD5A2	1	GSTM5P1	4	TPM2
1	PDGFC	270	GSTM2	1	NEXN	25068	SRD5A2	1	CBX7	3	TPM2
1	EYA4	270	GSTM2	1	CACHD1	25049	SRD5A2	1	PPP1R14A	3	TPM2
1	MYOCD	262	GSTM2	1	FERMT2	24208	SRD5A2	1	FABP3	3	TPM2
1	CALD1	255	GSTM2	1	LOC100127983	23635	SRD5A2	1	C15orf51	1	TPM2
1	KCNMB1	250	GSTM2	1	TPM1	22943	SRD5A2	1	GSTM2P1	1	TPM2
1	ACTN1	227	GSTM2	1	CALD1	22765	SRD5A2				
1	FZD7	216	GSTM2	1	SERPINF1	22186	SRD5A2				
1	LOC100127983	216	GSTM2	1	CSRPI	21728	SRD5A2				
1	DKK3	209	GSTM2	1	ACTN1	21590	SRD5A2				
1	GSTM5	207	GSTM2	1	HLF	21402	SRD5A2				
1	CHRD1	204	GSTM2	1	DES	20952	SRD5A2				
1	SORBS1	202	GSTM2	1	MYL9	19970	SRD5A2				
1	SPOCK3	202	GSTM2	1	HEPH	19688	SRD5A2				
1	JAZF1	189	GSTM2	1	TSPAN18	19099	SRD5A2				
1	LMOD1	180	GSTM2	1	SVIL	18819	SRD5A2				
1	DES	172	GSTM2	1	TGFB3	18423	SRD5A2				

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
				1	MYH11	18066	SRD5A2				
				1	KANK2	17638	SRD5A2				
				1	CDC42EP3	16173	SRD5A2				

Table 10

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
1	NCAPH	9758	CDC20	1	GPAT2	1997	CDC20	1	SUSD2	3824	MYBL2
1	CDC20	9758	CDC20	1	AVPR1B	1991	CDC20	1	TGM6	3767	MYBL2
1	IQGAP3	9758	CDC20	1	MGC50722	1990	CDC20	1	CDC43	3765	MYBL2
1	ESPL1	9674	CDC20	1	AQP12A	1983	CDC20	1	C20orf151	3706	MYBL2
1	CENPA	9671	CDC20	1	C6orf222	1965	CDC20	1	C11orf41	3650	MYBL2
1	POC1A	9671	CDC20	1	PRAMEF19	1965	CDC20	1	C9orf98	3636	MYBL2
1	KIF188	9328	CDC20	1	PRAMEF18	1965	CDC20	1	KRT24	3589	MYBL2
1	WDR62	9316	CDC20	1	SLC5A9	1965	CDC20	1	ABCC12	3582	MYBL2
1	TROAP	9178	CDC20	1	FCN3	1955	CDC20	1	B3GNT4	3569	MYBL2
1	ADAMTS7	8987	CDC20	1	GCM2	1910	CDC20	1	AZ11	3556	MYBL2
1	PKMYT1	8875	CDC20	1	ADORA3	1862	CDC20	1	RLTPR	3427	MYBL2
1	SLC2A6	8875	CDC20	1	PLA2G2F	1821	CDC20	1	KIF24	3264	MYBL2
1	FNDCl	8554	CDC20	1	C6orf25	1765	CDC20	1	DERL3	3232	MYBL2
1	FAM64A	8346	CDC20	1	CDC45	1681	CDC20	1	LIPE	3221	MYBL2
1	FAM131B	8322	CDC20	1	AGXT	1529	CDC20	1	TTL19	3196	MYBL2
1	PNLDC1	8135	CDC20	1	KIF25	1507	CDC20	1	SEC1	3196	MYBL2
1	KIFC1	7598	CDC20	1	ZDHHC19	1507	CDC20	1	ADAM8	3185	MYBL2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Co-expressed Gene	ProbeWt	Seeding Gene
1	C9orf100	7547	CDC20	1	APLN	1374	CDC20	1	SLC25A19	3136	MYBL2
1	RPS6KL1	7527	CDC20	1	TACC3	1220	CDC20	1	PRSS27	3136	MYBL2
1	MRAP	7521	CDC20	1	TK1	1063	CDC20	1	ODF3L2	3094	MYBL2
1	AURKB	7224	CDC20	1	C15orf42	1052	CDC20	1	ODZ4	3034	MYBL2
1	C2orf54	7150	CDC20	1	FANCA	990	CDC20	1	RAD54L	2936	MYBL2
1	TMEM163	6853	CDC20	1	GINS4	932	CDC20	1	KCNE1L	2936	MYBL2
1	KRBA1	6846	CDC20	1	MCM10	757	CDC20	1	SBF1P1	2915	MYBL2
1	ZMYND10	6825	CDC20	1	CYB561D1	748	CDC20	1	AIPL1	2868	MYBL2
1	LOC541473	6824	CDC20	1	FUT5	687	CDC20	1	UNC13A	2862	MYBL2
1	SLC6A1	6669	CDC20	1	POLQ	632	CDC20	1	REM2	2832	MYBL2
1	DOX1	6601	CDC20	1	LOC643988	621	CDC20	1	KIFC1	2808	MYBL2
1	BAI2	6583	CDC20	1	RAD51	601	CDC20	1	TSNAXIP1	2799	MYBL2
1	EME1	6533	CDC20	1	DGAT2	582	CDC20	1	LOC390660	2767	MYBL2
1	CICP3	6481	CDC20	1	KIF24	566	CDC20	1	SLC6A12	2762	MYBL2
1	PPFIA4	6480	CDC20	1	CDCA3	442	CDC20	1	WDR16	2723	MYBL2
1	PADH1	6458	CDC20	1	CLSPN	381	CDC20	1	ACR	2710	MYBL2
1	SSPO	6431	CDC20	1	ESYT3	356	CDC20	1	TMPRSS13	2672	MYBL2
1	GABRB2	6422	CDC20	1	EXO1	278	CDC20	1	C15orf42	2659	MYBL2
1	IRF5	6399	CDC20	1	CDCA2	186	CDC20	1	DNMT3B	2649	MYBL2
1	NXPH1	6399	CDC20	1	CKAP2L	159	CDC20	1	UNC13D	2610	MYBL2
1	ZIC1	6346	CDC20	1	FOXM1	157	CDC20	1	SYT5	2544	MYBL2
1	SLC6A20	6336	CDC20	1	FEN1	136	CDC20	1	PAX2	2462	MYBL2
1	PKD1L1	6281	CDC20	1	UHRF1	125	CDC20	1	PRCD	2426	MYBL2
1	BIRC5	6278	CDC20	1	KIF20A	110	CDC20	1	PPFIA3	2421	MYBL2
1	AQP10	6251	CDC20	1	ESCO2	107	CDC20	1	GCGR	2338	MYBL2
1	ABCA4	6216	CDC20	1	CA2	100	CDC20	1	CACNG3	2289	MYBL2
1	TFR2	6181	CDC20	1	PLK1	84	CDC20	1	LAIR2	2233	MYBL2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Co-expressed Gene	ProbeWt	Seeding Gene
1	LOC646070	6179	CDC20	1	PTTG1	64	CDC20	1	MCM10	2178	MYBL2
1	CSPG5	6165	CDC20	1	KIF14	53	CDC20	1	C2orf54	2172	MYBL2
1	CENPM	6124	CDC20	1	CIT	42	CDC20	1	LOC400419	2138	MYBL2
1	EFNA3	6100	CDC20	1	FAM54A	39	CDC20	1	RINL	2136	MYBL2
1	GPC2	6078	CDC20	1	CDC48	28	CDC20	1	DKFZp451A211	2118	MYBL2
1	HYAL3	6047	CDC20	1	DEPDC1B	12	CDC20	1	LAMA1	2060	MYBL2
1	CELA3B	6031	CDC20	1	MYBL2	12208	MYBL2	1	C9orf169	2060	MYBL2
1	LOC100287112	6015	CDC20	1	BIRC5	12208	MYBL2	1	CATSPER1	2001	MYBL2
1	SRCR84D	5999	CDC20	1	TROAP	12208	MYBL2	1	OPCML	1896	MYBL2
1	DNAJB3	5993	CDC20	1	ESPL1	12190	MYBL2	1	C9orf50	1852	MYBL2
1	PADI3	5989	CDC20	1	WDR62	12032	MYBL2	1	DOC2G6P	1760	MYBL2
1	PAX8	5971	CDC20	1	KIF18B	12015	MYBL2	1	TACC3	1685	MYBL2
1	AIM1L	5971	CDC20	1	FAM64A	11915	MYBL2	1	APOBEC3A	1632	MYBL2
1	FAM131C	5915	CDC20	1	PKMYT1	11774	MYBL2	1	LOC728307	1606	MYBL2
1	PRRT4	5915	CDC20	1	SLC2A6	11774	MYBL2	1	PDIA2	1572	MYBL2
1	MLXIPL	5915	CDC20	1	GTSE1	11356	MYBL2	1	LTB4R2	1419	MYBL2
1	E2F1	5912	CDC20	1	E2F1	11062	MYBL2	1	OIP5	1393	MYBL2
1	E2F7	5893	CDC20	1	AURKB	11010	MYBL2	1	ORC1L	1340	MYBL2
1	RAD54L	5888	CDC20	1	RNFT2	10720	MYBL2	1	GSG2	1268	MYBL2
1	C1orf81	5813	CDC20	1	CENPM	10651	MYBL2	1	FSD1	1256	MYBL2
1	NFKBIL2	5742	CDC20	1	CENPA	10628	MYBL2	1	CDC25C	1228	MYBL2
1	LOC729061	5728	CDC20	1	POCIA	10289	MYBL2	1	KSR2	1183	MYBL2
1	TAS1R3	5671	CDC20	1	FDXR	10285	MYBL2	1	DGAT2	1183	MYBL2
1	VWA3B	5643	CDC20	1	NFKBIL2	10214	MYBL2	1	KIF2C	1180	MYBL2
1	MYBL2	5565	CDC20	1	E2F7	10195	MYBL2	1	RAD51	1178	MYBL2
1	TTL6	5531	CDC20	1	C9orf100	10103	MYBL2	1	FNDCC8	1178	MYBL2
1	LOC100130097	5525	CDC20	1	CDH24	10094	MYBL2	1	RAB31L1	991	MYBL2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Co-expressed Gene	ProbeWt	Seeding Gene
1	CHRG	5491	CDC20	1	ABCB9	10079	MYBL2	1	UHRF1	936	MYBL2
1	TTBK1	5491	CDC20	1	NDUFA4L2	9961	MYBL2	1	ENO4	855	MYBL2
1	TRIM46	5491	CDC20	1	ADAMTS7	9614	MYBL2	1	C10orf105	780	MYBL2
1	MST1R	5491	CDC20	1	MAST1	9313	MYBL2	1	NEIL3	733	MYBL2
1	EXOC3L	5474	CDC20	1	GABBR2	9262	MYBL2	1	PPBP	672	MYBL2
1	TH	5474	CDC20	1	MYH7B	8759	MYBL2	1	PROCA1	671	MYBL2
1	CHST1	5474	CDC20	1	DNAH3	8637	MYBL2	1	TMEM132A	647	MYBL2
1	LOC442676	5439	CDC20	1	TLL6	8619	MYBL2	1	DHRS2	548	MYBL2
1	CNTN2	5435	CDC20	1	ZFX2	8592	MYBL2	1	PLK1	523	MYBL2
1	DPYSL5	5435	CDC20	1	CDC20	8589	MYBL2	1	GIN54	485	MYBL2
1	C3orf20	5368	CDC20	1	RASAL1	8452	MYBL2	1	CEL	480	MYBL2
1	NPC1L1	5291	CDC20	1	NCAPH	8273	MYBL2	1	ZNF367	406	MYBL2
1	CICP5	5281	CDC20	1	IQGAP3	8245	MYBL2	1	FOXM1	402	MYBL2
1	KLRG2	5275	CDC20	1	DNAH2	8219	MYBL2	1	POLQ	319	MYBL2
1	CCDC108	5275	CDC20	1	LOC400499	8151	MYBL2	1	ADAM12	312	MYBL2
1	HL288	5217	CDC20	1	CHST1	8037	MYBL2	1	SEMA7A	284	MYBL2
1	CELSR3	5166	CDC20	1	ATP4A	7868	MYBL2	1	HOXB5	137	MYBL2
1	RNFT2	5138	CDC20	1	TH	7731	MYBL2	1	EXO1	115	MYBL2
1	C17orf53	5114	CDC20	1	EXOC3L	7603	MYBL2	1	KIF4A	114	MYBL2
1	TRPC2	5095	CDC20	1	E2F8	7590	MYBL2	1	FEN1	112	MYBL2
1	KCNA1	5078	CDC20	1	MMP11	7465	MYBL2	1	CLSPN	107	MYBL2
1	C8G	4946	CDC20	1	CELP	7344	MYBL2	1	CIT	94	MYBL2
1	COL11A1	4685	CDC20	1	CDC45	7024	MYBL2	1	CDCA2	85	MYBL2
1	C1orf222	4673	CDC20	1	FAM131B	6981	MYBL2	1	KIF4B	68	MYBL2
1	SLC6A12	4633	CDC20	1	C14orf73	6896	MYBL2	1	PIK3R5	56	MYBL2
1	HCN3	4608	CDC20	1	FBXW9	6802	MYBL2	1	KIF20A	52	MYBL2
1	GTSE1	4528	CDC20	1	PLEKHG6	6725	MYBL2	1	ZWINT	31	MYBL2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Co-expressed Gene	ProbeWt	Seeding Gene
1	ORC1L	4497	CDC20	1	FNDC1	6720	MYBL2	1	SPAG5	19	MYBL2
1	STX1A	4475	CDC20	1	SEZ6	6515	MYBL2	1	ERCC6L	17	MYBL2
1	MFSD2A	4451	CDC20	1	FCHO1	6413	MYBL2	1	TPX2	11	TPX2
1	BEST4	4389	CDC20	1	APLN	6402	MYBL2	1	TOP2A	11	TPX2
1	CACNA1E	4299	CDC20	1	ALAS2	6382	MYBL2	1	NUSAP1	10	TPX2
1	KLHDC7A	4297	CDC20	1	V5X1	6360	MYBL2	1	MELK	7	TPX2
1	MAPK15	4272	CDC20	1	LOC197350	6312	MYBL2	1	RACGAP1	6	TPX2
1	GHRHR	4211	CDC20	1	DPF1	6205	MYBL2	1	NCAPG	4	TPX2
1	KEL	4155	CDC20	1	CDC45	6026	MYBL2	1	MKI67	4	TPX2
1	C2orf62	4113	CDC20	1	C11orf9	6020	MYBL2	1	CDKN3	4	TPX2
1	ANXA9	4063	CDC20	1	EME1	6010	MYBL2	1	PRC1	4	TPX2
1	RAET1G	4059	CDC20	1	ADAMTS13	5896	MYBL2	1	ARHGAP11B	3	TPX2
1	GPR88	3913	CDC20	1	TMEM145	5896	MYBL2	1	KIAA0101	3	TPX2
1	F12	3749	CDC20	1	C8G	5840	MYBL2	1	ANLN	3	TPX2
1	LYPD1	3681	CDC20	1	CBX2	5838	MYBL2	1	FAM111B	2	TPX2
1	C2orf70	3665	CDC20	1	TMEM210	5659	MYBL2	1	RRM2	1	TPX2
1	ABC89	3638	CDC20	1	CDC135	5593	MYBL2	1	KIF11	1	TPX2
1	MSLN	3589	CDC20	1	ADAMTS14	5571	MYBL2	1	PRR11	1	TPX2
1	CDC25C	3573	CDC20	1	ITGA2B	5337	MYBL2	1	CENPF	1	TPX2
1	CELA3A	3551	CDC20	1	POLD1	5286	MYBL2	2	MKI67	41	TPX2
1	AQP12B	3551	CDC20	1	PNLDC1	5146	MYBL2	2	CASC5	39	TPX2
1	NEU4	3551	CDC20	1	UCP3	5123	MYBL2	2	ASPM	38	TPX2
1	KIF2C	3541	CDC20	1	FANCA	5068	MYBL2	2	KIF4A	36	TPX2
1	NEIL3	3426	CDC20	1	MSLN	5061	MYBL2	2	DLGAP5	36	TPX2
1	NUDT17	3399	CDC20	1	TEPP	4930	MYBL2	2	KIF48	36	TPX2
1	ULRP2	3395	CDC20	1	LRR16B	4901	MYBL2	2	TPX2	33	TPX2
1	KIF17	3341	CDC20	1	CACNA1F	4901	MYBL2	2	KIF14	31	TPX2

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Co-expressed Gene	ProbeWt	Seeding Gene
1	ARHGEF19	3340	CDC20	1	EFNB3	4887	MYBL2	2	EXO1	31	TPX2
1	CYP4A22	3317	CDC20	1	MYBPC2	4851	MYBL2	2	SKA3	30	TPX2
1	CYP4A11	3317	CDC20	1	FUT6	4847	MYBL2	2	SPAG5	27	TPX2
1	SCNN1D	3311	CDC20	1	CDH15	4847	MYBL2	2	CIT	27	TPX2
1	FRMD1	3219	CDC20	1	HAL	4809	MYBL2	2	BUB1	26	TPX2
1	FAM179A	3194	CDC20	1	PGA3	4720	MYBL2	2	CDKN3	26	TPX2
1	NDUFA4L2	3109	CDC20	1	PGA4	4720	MYBL2	2	CENPF	25	TPX2
1	LCE2D	2984	CDC20	1	CL7orf53	4717	MYBL2	2	MELK	20	TPX2
1	ODZ4	2936	CDC20	1	UMODL1	4713	MYBL2	2	ANLN	19	TPX2
1	ABCC12	2809	CDC20	1	OTOG	4690	MYBL2	2	BUB1B	18	TPX2
1	DPF1	2750	CDC20	1	D8H	4661	MYBL2	2	UBE2C	17	TPX2
1	CDH24	2653	CDC20	1	POM121IP	4629	MYBL2	2	CEP55	16	TPX2
1	LOC154449	2641	CDC20	1	DNAJB13	4394	MYBL2	2	KIF20A	15	TPX2
1	KIF21B	2534	CDC20	1	TK1	4360	MYBL2	2	DEPDC1B	15	TPX2
1	SEMA5B	2499	CDC20	1	C9orf117	4336	MYBL2	2	DTL	14	TPX2
1	PSORS1C2	2497	CDC20	1	RHBDL1	4308	MYBL2	2	UBE2T	13	TPX2
1	FCRL4	2434	CDC20	1	MUC5B	4283	MYBL2	2	NCAPG	13	TPX2
1	FUT6	2313	CDC20	1	SPAG4	4276	MYBL2	2	PBK	13	TPX2
1	TRAIP	2258	CDC20	1	GOLGA7B	4111	MYBL2	2	DIAPH3	10	TPX2
1	E2F8	2232	CDC20	1	APOB48R	4107	MYBL2	2	KIF23	6	TPX2
1	SLC38A3	2199	CDC20	1	IQCD	3984	MYBL2	2	FOXMI	5	TPX2
1	CBX2	2174	CDC20	1	FUT5	3977	MYBL2	2	RRM2	3	TPX2
1	CDC45	2130	CDC20	1	AIFM3	3973	MYBL2	2	SGO11	2	TPX2
1	DUSP5p	2080	CDC20	1	LOC390595	3868	MYBL2	2	PLK1	2	TPX2
				1	CYP27B1	3833	MYBL2	2	CCNA2	2	TPX2
								2	CDK1	2	TPX2
								2	NUSAP1	1	TPX2

Table II

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
1	NNT	1	DUSP1	1	FOS	14	FOS	3	ZFP36	4	GADD45B
1	RNF180	1	DUSP1	1	BTG2	14	FOS	3	SOC3	4	GADD45B
1	PCDH18	1	DUSP1	1	CSRP1	13	FOS	3	RHOB	3	GADD45B
2	RNF180	1	DUSP1	1	ZFP36	13	FOS	3	BHLHE40	3	GADD45B
2	DUSP1	1	DUSP1	1	JUNB	9	FOS	3	FOS	2	GADD45B
2	PCDH18	1	DUSP1	1	NR4A3	7	FOS	3	FOSL2	2	GADD45B
3	ACTB	1	DUSP1	1	FOSB	7	FOS	3	BTG2	2	GADD45B
3	RHOB	1	DUSP1	1	SIK1	6	FOS	3	NR4A3	2	GADD45B
3	DUSP1	1	DUSP1	1	BHLHE40	6	FOS	3	FOSB	1	GADD45B
4	ACTB	1	DUSP1	1	RHOB	5	FOS	3	IRF1	1	GADD45B
4	DUSP1	1	DUSP1	1	TIPARP	5	FOS	1	FOSL2	1	ZFP36
4	CRTAP	1	DUSP1	1	KLF6	5	FOS	1	HBEGF	1	ZFP36
5	RNF180	1	DUSP1	1	MC11	5	FOS	1	BHLHE40	1	ZFP36
5	DUSP1	1	DUSP1	1	NR4A1	4	FOS	2	HBEGF	1	ZFP36
5	CRTAP	1	DUSP1	1	EGR1	4	FOS	2	NR4A3	1	ZFP36
5	PAM	1	DUSP1	1	NR4A2	4	FOS	2	BHLHE40	1	ZFP36
6	DUSP1	8	DUSP1	1	GADD45B	3	FOS	3	CSRP1	53	ZFP36
6	NR4A1	7	DUSP1	1	SOC3	2	FOS	3	ZFP36	49	ZFP36
6	FOS	7	DUSP1	1	NFKBIZ	1	FOS	3	JUNB	29	ZFP36
6	EGR1	5	DUSP1	2	FOS	24	FOS	3	FOS	26	ZFP36
6	BTG2	5	DUSP1	2	FOSB	22	FOS	3	BHLHE40	24	ZFP36
6	FOSB	5	DUSP1	2	EGR1	20	FOS	3	BTG2	24	ZFP36
6	JUN	4	DUSP1	2	NR4A1	19	FOS	3	FOSB	20	ZFP36
6	NR4A2	3	DUSP1	2	BTG2	18	FOS	3	NR4A3	18	ZFP36

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
6	TIPARP	3	DUSP1	2	ZFP36	12	FOS	3	SOC3	18	ZFP36				
6	CYR61	3	DUSP1	2	CSRNP1	11	FOS	3	EGR1	16	ZFP36				
6	ATF3	2	DUSP1	2	CYR61	10	FOS	3	RHOB	16	ZFP36				
6	RHOB	2	DUSP1	2	DUSP1	8	FOS	3	FOSL2	15	ZFP36				
6	NEDD9	2	DUSP1	2	ATF3	8	FOS	3	NR4A1	15	ZFP36				
6	MCL1	1	DUSP1	2	IER2	7	FOS	3	GADD45B	10	ZFP36				
6	RASD1	1	DUSP1	2	RHOB	6	FOS	3	MYADM	9	ZFP36				
1	JUNB	1	EGR1	2	TIPARP	6	FOS	3	KLF6	8	ZFP36				
1	TIPARP	1	EGR1	2	NR4A2	6	FOS	3	CYR61	8	ZFP36				
1	BTG2	1	EGR1	2	JUN	6	FOS	3	EGR3	8	ZFP36				
2	JUNB	1	EGR1	2	JUNB	6	FOS	3	EMP1	8	ZFP36				
2	BTG2	1	EGR1	2	EGR3	5	FOS	3	LMNA	7	ZFP36				
2	EGR1	1	EGR1	2	NR4A3	4	FOS	3	TIPARP	7	ZFP36				
3	KLF4	1	EGR1	2	KLF6	3	FOS	3	NR4A2	7	ZFP36				
3	FOSB	1	EGR1	2	PPP1R15A	2	FOS	3	MCL1	6	ZFP36				
3	EGR1	1	EGR1	2	NEDD9	2	FOS	3	SIK1	6	ZFP36				
4	FOSB	1	EGR1	2	KLF4	2	FOS	3	ATF3	6	ZFP36				
4	CSRNP1	1	EGR1	2	EGR2	2	FOS	3	CEBPD	5	ZFP36				
4	EGR1	1	EGR1	2	MCL1	1	FOS	3	IER3	5	ZFP36				
5	EGR1	35	EGR1	1	EMP1	7	GADD45B	3	IER2	5	ZFP36				
5	FOS	30	EGR1	1	BHLHE40	7	GADD45B	3	MAFF	4	ZFP36				
5	NR4A1	25	EGR1	1	SOC3	7	GADD45B	3	IRF1	4	ZFP36				
5	FOSB	23	EGR1	1	NR4A3	4	GADD45B	3	RNF122	4	ZFP36				
5	BTG2	22	EGR1	1	FOSL2	3	GADD45B	3	SRF	3	ZFP36				
5	CYR61	20	EGR1	1	GADD45B	3	GADD45B	3	ERRF1	2	ZFP36				
5	ZFP36	18	EGR1	1	RNF122	3	GADD45B	3	SLC25A25	2	ZFP36				
5	CSRNP1	17	EGR1	1	KLF10	3	GADD45B	3	CDKN1A	2	ZFP36				

StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene	StackID	Coexpressed Gene	ProbeWt	Seeding Gene
5	NP4A3	13	EGR1	1	CSRNP1	3	GADD45B	3	EGR2	2	ZFP36
5	EGR3	13	EGR1	1	SLC2A3	2	GADD45B	3	KLF4	1	ZFP36
5	KLF6	12	EGR1	1	ZFP36	1	GADD45B				
5	RHOB	11	EGR1	2	FOSB	2	GADD45B				
5	DUSP1	10	EGR1	2	NR4A1	2	GADD45B				
5	ATF3	9	EGR1	2	FOS	2	GADD45B				
5	JUN	9	EGR1	2	GADD45B	2	GADD45B				
5	TIPARP	8	EGR1	2	BTG2	2	GADD45B				
5	NFKBIZ	7	EGR1	2	NR4A3	2	GADD45B				
5	NR4A2	7	EGR1	2	JUNB	2	GADD45B				
5	JUNB	7	EGR1	2	EGR1	2	GADD45B				
5	IER2	7	EGR1	2	CSRNP1	2	GADD45B				
5	MCL1	4	EGR1	2	ZFP36	2	GADD45B				
5	KLF4	4	EGR1	2	RHOB	1	GADD45B				
5	EGR2	4	EGR1	2	EGR3	1	GADD45B				
5	NEDD9	2	EGR1	2	ATF3	1	GADD45B				
5	SRF	2	EGR1	3	GADD45B	4	GADD45B				
5	GADD45B	1	EGR1	3	JUNB	4	GADD45B				
5	TRIB1	1	EGR1	3	CSRNP1	4	GADD45B				

Example 4: Prospective Validation Study of RS27

Study Design and Statistical Methods

[00175] The algorithm RS27 in Table 4 was tested in a prospective clinical validation study that included 395 evaluable patients who had surgery for their prostate cancer between 1997 and 2010 at the University of California, San Francisco (UCSF). The patients had Low or Intermediate risk (by CAPRA) for clinically localized prostate cancer who might have been reasonable candidates for active surveillance but underwent RP at UCSF within 6 months of the diagnosis of prostate cancer by biopsy. No randomization for patient selection was performed. For each patient, prostate biopsy samples from one fixed, paraffin-embedded tissue (FPET) block containing one or more tumor-containing needle cores was evaluated.

[00176] To investigate if there is a significant relationship between RS27 or any component of RS27 and adverse pathology at RP, multivariable and univariable multinomial logistic regression models were used and p-values from likelihood-ratio (LR) tests of the null hypothesis that the odds ratio (OR) is one were reported. The multinomial logistic model was also used to calculate estimates with 95% confidence intervals of the probability of high-grade or non-organ confined disease. To evaluate the relationship between RS27, baseline covariates, and combinations of these factors with high grade or non-organ confined disease, multivariable and univariable binary logistic regression models were used and p-values from likelihood-ratio tests of the null hypothesis that the odds ratio (OR) is one were reported.

[00177] The primary endpoint was formulated as follows:

Table 12: Clinical Endpoint – RP Grade and Stage

RP Gleason Score	Pathologic T2 Stage	Pathologic T3 Stage
$\leq 3+3$	1	2
3+4	3	4
Major pattern 4 or minor pattern 5, or tertiary pattern 5	5	6

where Gleason Score $\leq 3+3$ and pT2 (denoted “1”) is the reference category and all other categories (2-6) are compared to the reference category.

[00178] Cell combinations of Table 12 evaluated in binary logistic regression models include the following:

Cells 2, 4, 6 vs. 1, 3, 5: Non-organ-confined disease

Cells 5, 6 vs. 1, 2, 3, 4: High-grade disease

Cells 2, 4, 5, 6 vs. 1 and 3: High-grade or non-organ-confined disease

RS27 Algorithm

[00179] RS27 on a scale from 0 to 100 was derived from reference-normalized gene expression measurements as follows.

[00180] Unscaled RS27 (RS27u) was defined as in Table 4:

[00181] $RS27u = 0.735 \times \text{ECM (Stromal Response) group} - 0.368 \times \text{Migration (Cellular Organization) group} - 0.352 \times \text{PSA (Androgen) group} + 0.095 \times \text{Proliferation (TPX2)}$

[00182] Where:

[00183] $\text{ECM (Stromal Response) group score} = 0.527 \times \text{BGN} + 0.457 \times \text{COL1A1} + 0.156 \times \text{SFRP4}$

[00184] $\text{Migration (Cellular Organization) group score} = 0.163 \times \text{FLNC} + 0.504 \times \text{GSN} + 0.421 \times \text{TPM2} + 0.394 \times \text{GSTM2}$

[00185] $\text{PSA (Androgen) group score} = 0.634 \times \text{FAM13C} + 1.079 \times \text{KLK2} + 0.642 \times \text{AZGP1} + 0.997 \times \text{SRD5A2 Thresh}$

[00186] $\text{Proliferation (TPX2) score} = \text{TPX2 Thresh}$

[00187] where the thresholded gene scores for SRD5A2 and TPX2 are calculated as follows:

[00188] $\text{SRD5A2 Thresh} = \begin{cases} 5.5 & \text{if SRD5A2} < 5.5 \\ \text{SRD5A2} & \text{otherwise} \end{cases}$

[00189] $\text{TPX2 Thresh} = \begin{cases} 5.0 & \text{if TPX2} < 5.0 \\ \text{TPX2} & \text{otherwise} \end{cases}$

[00190] RS27u is then rescaled to be between 0 and 100 as follows:

[00191] $RS27(\text{scaled}) = \begin{cases} 0 & \text{if } 13.4 \times (RS27u + 10.5) < 0 \\ 13.4 \times (RS27u + 10.5) & \text{if } 0 \leq 13.4 \times (RS27u + 10.5) \leq 100 \\ 100 & \text{if } 13.4 \times (RS27u + 10.5) > 100 \end{cases}$

[00192] Patients were classified into low, intermediate, and high RS27 groups using pre-specified cut-points defined in Table 13 below. These cut-points defined the boundaries between low and intermediate RS27 groups and between intermediate and high RS27 groups. The cutpoints were derived from the discovery study with the intent of identifying substantial proportions of patients who on average had clinically meaningful low or high risk of adverse pathology. The RS27 was rounded to the nearest integer before the cut-points defining the RS27 groups were applied.

Table 13

RS27 Group	Score
Low	Less than 16

Intermediate	Greater than or equal to 16 and less than 30
High	Greater than or equal to 30

Assay Methods

[00193] Paraffin from the samples was removed with Shandon Xylene substitute (Thermo Scientific, Kalamazoo, MI). Nucleic acids were isolated using the Agencourt® FormaPure® XP kit (Beckman Coulter, Beverly, MA).

[00194] The amount of RNA was determined using the Quant-iT™ RiboGreen® RNA Assay kit (Invitrogen™, Carlsbad, CA). Quantitated RNA was converted to complementary DNA using the Omniscript® RT kit (Qiagen, Valencia, CA) and combined with the reverse primers for the 12 genes of RS27 and 5 normalization genes (ARF1, ATP5E, CLTC, GPS1, PGK1) as shown in Table A. The reaction was incubated at 37°C for 60 minutes and then inactivated at 93°C for 5 minutes.

[00195] The cDNA was preamplified using a custom TaqMan® PreAmp Master Mix made for Genomic Health, Inc. by Life Technologies (Carlsbad, CA) and the forward and reverse primers for all targets as shown in Table A. The reaction was placed in a thermocycler (DNA Engine® PTC 200G, Bio-Rad, Hercules, CA) and incubated under the following conditions: A) 95°C for 15 sec; B) 60°C for 4 min; C) 95°C for 15 sec; and D) steps B and C were repeated 8 times. The amplified product was then mixed with the forward and reverse primers and probes for each of the targets as shown in Table A and the QuantiTect® Primer Assay master mix (Qiagen, Valencia, CA) and amplified for 45 cycles in a LightCycler® 480 (Roche Applied Science, Indianapolis, IN). The level of expression was calculated using the crossing-point (Cp) method.

Results

[00196] RS27 significantly predicted for adverse pathology, non-organ-confined disease, high-grade disease, and high-grade or non-organ-confined disease, and adds value beyond biopsy Gleason Score as shown in Tables 14, 15, 16, and 17, respectively.

Table 14 Prediction of Adverse Pathology

Variable	LR Chi-Square	DF	P-value
RS27 Score	19.31	5	0.002
Central Biopsy Gleason Score 3+4 vs 3+3	32.86	5	<0.001

Results obtained from the multivariable multinomial logistic model for cells 2, 3, 4, 5, and 6 vs 1 in Table 12.

DF = degrees of freedom

Table 15 Prediction of Non-organ-confined disease

Model	Variables	Odds Ratio	95% Confidence Interval	DF	LR Chi-Square	P-value
Univariable Model						
	RS27	2.20	(1.46, 3.31)	1	14.44	<0.001
Multivariable Model						
	RS27	1.93	(1.25, 2.96)	1	8.97	0.003
	Central Biopsy Gleason Score 3+4 vs 3+3	1.79	(1.04, 3.10)	1	4.23	0.040

Results obtained from univariable and multivariable binary logistic regression models for cells 2, 4, 6 vs. 1, 3, 5 in Table 12

Odds ratio for RS27 was per 20 unit increase

Table 16 Prediction of high-grade disease

Model	Variables	Odds Ratio	95% Confidence Interval	DF	LR Chi-Square	P-value
Univariable Model						
	RS27	2.48	(1.60, 3.85)	1	16.78	<0.001
Multivariable Model						
	RS27	2.32	(1.46, 3.67)	1	12.92	<0.001
	Central Biopsy Gleason Score 3+4 vs 3+3	1.36	(0.75, 2.47)	1	0.98	0.322

Results obtained from univariable and multivariable binary logistic regression models for cells 5, 6 vs. 1-4 in Table 12

Odds ratio for RS27 was per 20 unit increase

Table 17 Prediction of high-grade or non-organ-confined disease

Model	Variables	Odds Ratio	95% Confidence Interval	DF	LR Chi-Square	P-value
Univariable Model						
	RS27	2.23	(1.52, 3.27)	1	17.77	<0.001
Multivariable Model						
	RS27	1.93	(1.30, 2.88)	1	10.70	0.001

	Central Biopsy Gleason Score 3+4 vs 3+3	1.94	(1.17, 3.21)	1	6.45	0.011
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*Results obtained from univariable and multivariable binary logistic regression models for cells 2, 4, 5, 6 vs. 1, 3 in Table 12

Odds ratio for RS17 was per 20 unit increase

[00197] In addition, RS27 predicted adverse pathology beyond conventional clinical/pathology treatment factors as shown in Table 18.

Table 18 Prediction of Adverse Pathology Beyond Conventional Clinical/Pathology Treatment Factors

Variable	LR Chi-Square	DF	P-value
RS27	21.46	5	<0.001
Original Biopsy Gleason Score	22.77	5	<0.001
RS27	19.31	5	0.002
Central Biopsy Gleason Score	32.86	5	<0.001
RS27	30.09	5	<0.001
Clin T2 v. T1	11.94	5	0.036
RS27	30.17	5	<0.001
Baseline PSA (ng/ml) <10 v. ≥10	10.44	5	0.064
RS27	30.75	5	<0.001
Continuous PSA	15.17	5	0.010
RS27	26.36	5	<0.001
Age	19.05	5	0.002
RS27	29.20	5	<0.001
Pct Core Positive	4.75	5	0.447

[00198] When added to conventional clinical/pathology tools such as CAPRA, RS27 further refined the risk of high grade or non-organ-confined disease. Using CAPRA alone, 5% of patients were identified as having greater than 85% probability of being free from high-grade

or non-organ-confined disease compared to 22% of patients identified of being free from high-grade or non-organ-confined disease using RS27 in addition to CAPRA (Figure 8).

[00199] When added to conventional clinical/pathology tools such as AUA (D'Amico et al., *JAMA* 280:969-974, 1998), RS27 further refined the risk of high grade or non-organ-confined disease. As shown in Figure 9, using AUA alone, 0% of patients are identified as having greater than 80% probability of being free from high-grade or non-organ-confined disease compared to 27% of patients identified of being free from high-grade or non-organ-confined disease using GPS in addition to AUA.

[00200] Individual genes and gene groups of RS27 were also associated with adverse pathology, high-grade disease, non-organ-confined disease, and high-grade or non-organ-confined disease, in univariable analyses as shown in Tables 19, 20, 21, and 22, respectively.

Table 19 Association of Genes and Gene Groups with Adverse Pathology, Univariable Analyses

Genes and Gene Groups	LR Chi-Square	DF	P-value
BGN	7.11	5	0.213
COL1A1	7.88	5	0.163
SFRP4	8.87	5	0.114
FLNC	12.26	5	0.031
GSN	5.73	5	0.333
GSTM2	1.84	5	0.870
TPM2	18.33	5	0.003
AZGP1	22.87	5	<0.001
KLK2	5.97	5	0.309
FAM13C1	21.55	5	<0.001
SRD5A2	9.10	5	0.105
SRD5A2 Thresholded	9.25	5	0.099
TPX2	14.26	5	0.014
TPX2 Thresholded	23.34	5	<0.001

Ref Gene Average	3.27	5	0.659
Stromal Group Score	9.84	5	0.080
Cellular Organization Group Score	8.04	5	0.154
Androgen Group Score	29.46	5	<0.001
Proliferation Group Score	23.34	5	<0.001
GPS	29.98	5	<0.001

Table 20 Association of Genes and Gene Groups with High-Grade Disease, Univariable Analyses

Gene	Chi-Square	DF	P-value	OR	(95% CI)
BGN	3.67	1	0.055	1.46	(0.99, 2.15)
COL1A1	2.33	1	0.127	1.32	(0.93, 1.87)
SFRP4	6.08	1	0.014	1.33	(1.06, 1.67)
FLNC	3.04	1	0.081	0.77	(0.57, 1.03)
GSN	0.14	1	0.710	0.94	(0.67, 1.32)
GSTM2	0.03	1	0.870	0.97	(0.69, 1.37)
TPM2	2.85	1	0.091	0.76	(0.56, 1.04)
AZGP1	12.69	1	<0.001	0.58	(0.42, 0.79)
KLK2	3.50	1	0.061	0.62	(0.38, 1.02)
FAM13C1	9.29	1	0.002	0.51	(0.33, 0.79)
SRD5A2	3.26	1	0.071	0.76	(0.56, 1.02)
SRD5A2 Thresholded	2.70	1	0.100	0.75	(0.53, 1.06)
TPX2	1.72	1	0.190	1.21	(0.91, 1.59)
TPX2 Thresholded	7.38	1	0.007	1.93	(1.20, 3.11)
Ref Gene Average	1.18	1	0.277	0.86	(0.65, 1.14)
Stromal Response Group Score	4.92	1	0.027	1.49	(1.05, 2.12)
Cellular Organization Group Score	1.12	1	0.290	0.87	(0.66, 1.13)
Androgen Group Score	15.07	1	<0.001	0.69	(0.58, 0.83)
Proliferation Group Score	7.38	1	0.007	1.93	(1.20, 3.11)

Table 21 Association of Genes and Gene Groups with Non-Organ-Confined Disease, Univariable Analyses

Gene	Chi-Square	DF	p-value	Odds Ratio	95% CI
BGN	2.58	1	0.109	1.34	(0.94, 1.91)
COL1A1	2.90	1	0.089	1.33	(0.96, 1.83)
SFRP4	4.39	1	0.036	1.25	(1.01, 1.54)
FLNC	0.34	1	0.560	0.92	(0.70, 1.21)
GSN	0.27	1	0.603	0.92	(0.67, 1.26)
GSTM2	0.16	1	0.693	0.94	(0.68, 1.29)
TPM2	0.51	1	0.473	0.9	(0.67, 1.20)
AZGP1	12.48	1	<0.001	0.59	(0.44, 0.80)
KLK2	2.10	1	0.148	0.71	(0.45, 1.12)
FAM13C1	12.42	1	0.000	0.48	(0.32, 0.73)
SRD5A2	2.42	1	0.120	0.8	(0.60, 1.06)
SRD5A2 Thresholded	2.65	1	0.103	0.77	(0.56, 1.06)
TPX2	6.38	1	0.012	1.39	(1.08, 1.81)
TPX2 Thresholded	6.51	1	0.011	1.82	(1.14, 2.89)
Ref Gene Average	0.33	1	0.563	0.93	(0.73, 1.19)
Stromal Response Group Score	4.24	1	0.040	1.41	(1.02, 1.95)
Cellular Organization Group Score	0.45	1	0.504	0.92	(0.72, 1.18)
Androgen Group Score	14.64	1	<0.001	0.71	(0.60, 0.85)
Proliferation Group Score	6.51	1	0.011	1.82	(1.14, 2.89)

Table 22 Association of Genes and Gene Groups with High-Grade or Non-Organ-Confined Disease, Univariable Analyses

Gene	Chi-Square	DF	p-value	Odds Ratio	95% CI
BGN	3.15	1	0.076	1.33	(0.97, 1.84)
COL1A1	1.96	1	0.162	1.23	(0.92, 1.65)
SFRP4	7.08	1	0.008	1.29	(1.07, 1.55)
FLNC	2.36	1	0.125	0.83	(0.65, 1.06)
GSN	0.45	1	0.503	0.91	(0.68, 1.20)
GSTM2	0.49	1	0.484	0.9	(0.68, 1.20)
TPM2	2.24	1	0.135	0.82	(0.63, 1.06)
AZGP1	12.20	1	0.001	0.61	(0.46, 0.82)
KLK2	2.18	1	0.140	0.73	(0.48, 1.11)

Gene	Chi-Square	DF	p-value	Odds Ratio	95% CI
FAM13C1	11.13	1	0.001	0.53	(0.37, 0.78)
SRD5A2	4.36	1	0.037	0.76	(0.59, 0.98)
SRD5A2 Thresholded	4.63	1	0.032	0.73	(0.55, 0.98)
TPX2	3.50	1	0.062	1.25	(0.99, 1.58)
TPX2 Thresholded	5.86	1	0.016	1.73	(1.09, 2.74)
Ref Gene Average	0.68	1	0.041	0.91	(0.73, 1.14)
Stromal Response Group Score	4.59	1	0.032	1.37	(1.03, 1.84)
Cellular Organization Group Score	1.54	1	0.215	0.87	(0.70, 1.08)
Androgen Group Score	16.56	1	<0.001	0.72	(0.61, 0.84)
Proliferation Group Score	5.86	1	0.016	1.73	(1.09, 2.74)

Example 5: RS27 Adds Value Beyond PTEN/TMPRSS2-ERG Status in Predicting Clinical Recurrence

[00201] PTEN mutation and TMPRSS2-ERG fusion genes are commonly associated with poor prognosis in prostate cancer. Here, RS27 was analyzed to determine whether it can provide value beyond PTEN/TMPRSS2-ERG status in predicting clinical recurrence.

[00202] PTEN and TMPRSS2-ERG fusion expression levels obtained in the gene identification study described in Example 1 above and in U.S. Pub. No. 20120028264 were used to stratify patients into PTEN low and PTEN normal groups. PTEN and TMPRSS2-ERG ("T2-ERG") status of the patients were found as follows:

Table 23. Distribution of PTEN Expression by T2-ERG Status

	T2-ERG Negative (53%)	T2-ERG Positive (47%)
Median PTEN	8.9	8.7
25% PTEN	8.7	8.4

[00203] A cutpoint for "PTEN low" was made at ≤ 8.5 , which included approximately 13% of T2-ERG negative patients and 28% of T2-ERG positive patients. PTEN normal was defined as > 8.5 .

[00204] Univariable Cox Proportional Hazards was applied to evaluate the association between PTEN status and time to clinical recurrence (cR). Figure 10 and Table 24 show that PTEN low patients have a higher risk of recurrence compared to PTEN normal patients.

[00205]

Table 24.

Chi Sq	P-value	HR	95% CI
12.44	<0.001	0.38	(0.22,0.65)

[00206] When the patients were further stratified into PTEN low/T2-ERG negative ("category 0"), PTEN low/T2-ERG positive ("category 1"), PTEN normal/T2-ERG negative ("category 2"), and PTEN normal/T2-ERG positive ("category 3"), both PTEN low categories had the lowest recurrence rates compared to PTEN normal patients as shown in Figure 11 and Table 25.

Table 25.

PTEN/T2-ERG categories	CHISQ	P-VALUE	95% CI
Cat 1 v 0	0.93	0.34	(0.28,1.55)
Cat 2 v 0	11.80	<0.01	(0.12,0.56)
Cat 3 v 0	7.05	0.01	(0.16,0.78)

[00207] The tables below summarize the results of a multivariable model with PTEN/T2-ERG status (Table 26) or PTEN status (Table 27), RS27, and biopsy Gleason Score (Bx GS), demonstrating that RS27 adds value beyond PTEN and T2-ERG markers and Biopsy GS in predicting clinical recurrence.

Table 26.

VARIABLE	DF	CHISQ	P-VALUE	HR	95% CI
RS27	1	64.13	<0.01	1.07	(1.05,1.09)
PTEN/T2-ERG Status	3	1.59	0.66		
PTEN/T2-ERG (Cat 1 v. 0)	1	0.06	0.80	0.91	(0.41,1.98)
PTEN/T2-ERG (Cat 2 v. 0)	1	1.17	0.28	0.65	(0.29,1.42)
PTEN/T2-ERG (Cat 3 v. 0)	1	0.14	0.71	0.86	(0.39,1.89)
BX GS	2	7.19	0.03		
Bx GS (7 v. 6)	1	6.86	0.01	0.40	(0.20,0.79)
Bx GS (8+ v. 6)	1	1.35	0.24	0.69	(0.36,1.29)

Table 27.

VARIABLE	DF	CHISQ	P-VALUE	HR	95% CI
GPS	1	66.67	<0.01	1.07	(1.05,1.09)
PTEN Status	1	0.86	0.35	0.78	(0.46,1.32)
BX GS	2	6.43	0.04		
Bx GS (7 v. 6)	1	6.12	0.01	0.42	(0.21,0.84)
Bx GS (8+ v. 6)	1	1.15	0.28	0.71	(0.37,1.33)

TABLE A

Organism Symbol	Sequence ID	Forward Primer Sequence	SEQ ID NO	Reverse Primer Sequence	SEQ ID NO	Probe Sequence
ALDH1A2	NM_170696.1	CACGCTCTGCTCTCTGCT	1	GACGCTGGCTCACTTTGTAT	2	TCCTGTAGGGCCCAAGCTCTGAGG
ANPEP	NM_001150.2	CCACCTTGGACCAAGTAAAGC	5	TCACAGCTCACCTGGTAGGA	6	CTCCCAACACGCTGAACCCG
AR	NM_000444.2	CGACTTACCGCACCTGAT	9	TGACACAAGTGGACTGGGATA	10	ACCATGCCCAAGGGTACCACA
ARF1	NM_001658.2	CAGTAGAGATCCCGCAACT	13	ACAAGCACATGGCTATGAA	14	CTTGCTCTGGGTACCCCTGCA
ASPN	NM_017680.4	CATTGCCACTCAACTCTAA	17	ATTGTAGTGTCCAGGGCTCT	18	TATCCCTTTGGAGACCTTGCTTG
ATP5E	NM_006885.2	CCGCTTTCGCTACAGCAT	21	TGGGAGTTCGGATGTAGCTG	22	TCCAGCCGTCTCCAGTAGGGCCAC
AZGP1	NM_001185.2	GAGCGACGCTAGGAAGCA	25	CAGGAAGGGCAGCTACTGG	26	TCAGATCCCACTATGCCCTCAA
BGN	NM_001711.3	GAGCTCCGCAAGGATGAC	29	CTGTGTTTACCAAGGACGA	30	CAAGGGTCTCCAGCACTCTAGGC
BIN1	NM_004305.1	CTGCAAAAGGGAACAAGAG	33	CGTGGTTGACTCTGATCTCG	34	CTCGCCTCCAGATGGCTCCG
BNIP6	NM_001718.4	GTGACAGACCTTGTTCACCT	37	CTAGTTGGCCGCAAGCAC	38	TGAACCCCGAGTATGTCCCAACAG
C7	NM_000587.2	ATGCTGAGTGTGAGCGGG	41	AGGCTTATGCTGGTGACAG	42	ATGCTCTGCCCCCTGCACTCAGC
CADM1	NM_014333.2	CCACCAACCATCTTACCATC	45	GATCCACTGCCCTGATCG	46	TCTTCACTGCTCGGGAACTCTGT
CD276	NM_001024736.1	CCAAAGGATGCGATACACAG	49	GGATGACTTGGGAATCATGTC	50	CCACTGTGAGCCCTTATTTCTCCAATG
CD44	NM_000610.3	GGCACCACTGCTTATGAAGG	53	GATGCTCATGGTGAATGAGG	54	ACTGGAACCCAGCAAGCACACCTC
CDKN2C	NM_001255.2	AGTGACCTGCACTGGCTGCT	57	GGTCTCTTGGCTTGGCT	58	CCAATGCAACCCCTGCGGCTGGC
CLTC	NM_004859.1	TGAAGGGAACTGCCCTTGCA	61	TGCTCTTACCAAGCACTCCAGC	62	TGGCTGCCAAAGAGGCACTCCGGGT
COL1A1	NM_000089.2	ACCGTATGGAGAGCCACAG	65	TGACTACAGGATCAGGCTTTC	66	TCTACATGCTGTACCCAAAGCCA
COL1A2	NM_000089.2	GTGGCCATCAGCTGACG	69	CAGTGGTAGGTGATGTTCTGGGA	70	TCTGCGCTGATGTCCACG
COL3A1	NM_000090.3	CAGCAAGAAGTGGTATAGGCT	73	AACTGGCTGCCAGCATG	74	TCTCTAGCCAGACGCTGTTTCTGTCCCTTG
COL4A1	NM_001845.4	GGAGGTCTGGACCTGCTG	77	ACCAGGACTGCCACGTTG	78	CTCTGTGCCCCCAAGGTGTCAAAG
COL5A2	NM_000393.3	ACAAAGGCTCCCAAGGAT	81	GAGTCCCAAGGAAGACCTGCT	82	CTCCTTTGACACCAAGGATGCCAT
COL6A1	NM_001848.2	GGTGAGGAACCAAGGT	85	GGCTGGAGTCCCACTCTG	86	CCAGGAATCTCTGTAGCACCAAGC
COL8A1	NM_001850.3	GGAGACCCCTGGTGAAGCTG	89	TCTCCAGGACACCAACG	90	CTTCTCTTCCCTGATCACCTGCG
CSF1	NM_000757.3	TGGTCTCCAGGGCTTCT	93	CCCTGTAAACCTGATCCC	94	CCTAAGGGAGAGCCAGGAATCCCA
CSRP1	NM_004078.1	TGCAGCGGCTGATTGACA	97	CAACTGTCTCTGCTACAACTCA	98	TCAGATGGAGACCTGTGCCAAATTACA
CYP3A5	NM_000777.2	ACCAAGACCCCTGCCCTCT	101	GAGGGGTGGAGTGATGT	102	CCACCTTCTCCAGGGACCCCTTAG
DES	NM_001927.3	TCATTGCCAGTATGGAGATG	105	GACAGGCTTGGCTTCTCTG	106	TCCCGCTCAAGTTCTCACCAT
DRP4	NM_001935.3	ACTTCTCACTGGCCGACG	109	GCCTCACCTTCTGTTGGT	110	TGAACCAAGGAGTTTCTGACCACGC
DUSP1	NM_004417.2	GTCTGGGATCGGGAAGT	113	GTACTCCACCCGGATACAG	114	CGGCTATTCACACACTTGACACGC
EGR1	NM_001964.2	AGACATDAGCTCTGGTTCA	117	GACAAACACCTTCTCTCCAG	118	CGAGGCCATTGACTTACATAGACTCCA
EGR3	NM_004430.2	GTCCCGCTGCAGATCTCT	121	CTCCAGCTTAGGGTAGTGTCCAT	122	CGATCCTTCTCTCACTCGCCCA
ERG	NM_004449.3	CCAATGTGGATGAATGAGGTG	125	TGCTTGAGAAGAGGTGAGGT	126	ACCCAGTCTCACCTTCTGCCCCACC
F3R	NM_001992.2	CCAAACACTAGGCTCCCA	129	CTCCGCCAGGCTTCTTAGT	130	AGCCATATGCCCTTCTCATCTGGGC
FAM107A	NM_007177.2	AAGAGCAAAACCATCAGG	133	GCAGGGTTCATTGAGCAC	134	CCCGGGCTCAACTACTACTACTGT
FAM13C	NM_198215.2	TCTGCCAGGCCCTTCCAC	137	AGGAGCTGGGTGTACGAGA	138	TCTCGAGGCTCCCAAGGGCCCCG
FAP	NM_004460.2	ATCTTCAAAGCGGAGAGCG	141	GCTGGATACCAATGCTCTG	142	TCTGACTTCTCCGTTGGCTCTG
FLNC	NM_001458.4	GTGGCTACGTTGGGTAC	145	GACAGGCGAAACATCTCTG	146	AGCCACTGCAAAACATAGCTTCATCA
FN1	NM_002026.2	CAGGACAATGGTGAAGGT	149	TGATGGTGTACTCTGCCAGC	150	ATGTGCTGTAGCTACCTGCCCCAC
		GGAAATGACACGCTGAAGGT	153	ACACGGTAGCCCGGTCACT	154	ACTCTCAGCGGTGTCCACATGAT

Original Symbol	Sequence ID	Forward Primer Sequence	SEQ ID NO	Reverse Primer Sequence	SEQ ID NO	Probe Sequence
FOS	NM_005252.2	CGAGCCCTTTGATGACTTCT	157	GGAGGGGCTGTCTCAGA	158	TCCAGCATCATCCAGGCCAG
GADD45B	NM_015675.1	ACCTCGACAGACCACT	161	TGGAGTTATGGGTACAGA	162	TGGAGTTATGGGTACAGA
GPM6B	NM_001001994.1	ATGTGCTTGGAGTGGCT	165	TGTAGAACTAAACACGGCA	166	TGGAGTTATGGGTACAGA
GPS1	NM_004127.4	AGTACAAGCAGCTGCCAAG	169	GCAGCTCAGGGAAGTACA	170	CGCTCTGTGGCTTCTTTGATCA
GSN	NM_000177.1	CTTCTGCTAAGCGGTACATCGA	173	GGCTCAAAGCTTGTCTCAC	174	ACCGACCAATGGGATCGGC
GSTM1	NM_000561.1	AAGCTATGAGGAAAGAGTACACGAT	177	GGCCAGCTTGAATTTTCA	178	TCAGCCACTGGCTTCTGTCTAATCAGGAG
GSTM2	NM_000848.2	CTGAGGCACTCCCTGAAT	181	CCAGAAACCACTGGCTGCTT	182	CTGAAGCTCTACTACACAGTTTCTGGG
HIF	NM_002126.4	CACCTGCGAGGTGCTCTGAG	185	GGTACCTAGGACGAGAGTGA	186	TAAGTATCTGCCCTCCAGGTGGC
IGF1	NM_000618.1	TCCGAGCTGTGATCTAAGGA	189	CGGACAGAGCGAGCTGACTT	190	TGATTTGGCGACCCCTCAAGGCTG
IGFBP2	NM_000597.1	GTGGACAGCACCATGAACA	193	CTTCATACCCGACTTGAGG	194	CTCCGGCCAGCAGCTGCCCTC
IGFBP6	NM_002178.1	TGAACCCGACAGACCAACAG	197	GTCTTGGACCCCGCAGAAT	198	ATCCAGGCACCTCTACCACGCCCTC
IL6ST	NM_002184.2	GGCTTAATGTTCCAGATCCT	201	AAATTTGGCTTGGAGGAG	202	CATATGCCAGTGGTCACTCACA
INHBA	NM_002192.1	GTGCCGAGCCATATAGCA	205	CGGTAGTGGTTGATGACTTTGA	206	ACGTCCGGGTCTCAGCTGCTCTCC
ITGA7	NM_002206.1	GATATGATGGTGGCTGCTTG	209	AGAACTTCCATTCCCAACAT	210	CAGCCAGGACCTGGCCATCCG
JUN	NM_002228.2	GACTGCAAGATGGAACGA	213	TAGCCATAAGGTCCGCTCTC	214	CTATGACGATGCCCTCAACGGCTC
KLK2	NM_005551.3	AGTCTGGATTGTGGAGG	217	GTACACAGCCACTGCC	218	TTGGAACTGCTCTCACACTCCCA
KRT15	NM_002275.2	GCCTGGTCTTCAGCAAGAC	221	CTTGCTGGTCTGGATCATTTTC	222	TGAACTGCTGCTGCTGCTCAAGCA
KRT5	NM_000424.2	TCAGTGGAGAGGAGTTGGA	225	TGCCATATCCAGAGAAACA	226	CCAGTCAACATCTGTTGTCAAGCA
LAMB3	NM_000228.1	ACTGACCAAGCCTGAGACT	229	GTACACATTCAGCATTTTCA	230	CCACTGCCATCTCTCCCGGAGT
LGALS3	NM_002306.1	AGCGGAATTTGGCAGACAT	233	CTTGAGGTTTGGTCTTCCA	234	ACCCAGATAACGCATCATGAGCGA
MMP11	NM_005940.2	CCTGAGGCTGCAACATACC	237	TACAATGGCTTTGAGGATAGCA	238	ATCTCTGTAAGCCCTTTTGGCAGC
MYBL2	NM_002466.1	GCCGAGATGCCAAGATG	241	CTTTGATGGTAGAGTTCCAGTGATTC	242	CAGCTTCTGCTCTCCCTGGCA
NFAT5	NM_006599.2	CTGAACCCCTCTCTCTGTC	245	AGGAACGATGGCGAGGT	246	CGAATACAGTCCCGTGGGATTC
OLFML3	NM_020190.2	TCAGAACTGAGGCCGACAC	249	CCAGATAGTCTACCTCCGCT	250	CAGACGATCCAGTCTCCCGGAGAT
PAGE4	NM_007003.2	GAATCTAGCAAGAGAACCA	253	GTCTTGATCGGAGGTGT	254	CCAACTGACAATCAGGATATTGAAGCTGG
PGK1	NM_000281.1	AGGCCAGTTGCTGTAGAACTCAA	257	CTGGGCTTACACAGTCTTCA	258	TCTCTGCTGGCAAGGATGTTCTGTTC
PPAP2B	NM_003713.3	ACAAGCACCATCCCACTGA	261	CTGGGCTTACACAGTCTTCA	262	ACCCAGGCTCTTGAAGCAAACTCT
PPP1R12A	NM_002480.1	CGSCAAGGGTTGATATAGA	265	TGCTGGCATCTCTAAGCA	266	CGGTCTCTCTCTCTTCCAGCTGC
PRKCA	NM_002737.1	CAAGCAATTCGCTCATCAATGT	269	GTAAATCCGCGCTCTCTCT	270	CAGCTCTCGGGAATGGATCACT
SDC1	NM_002997.1	GAATGAGCAGGGGTTCT	273	AGGAGCTAGCGGAGAACCTG	274	CTCTGAGCGCTCCATCCAAAG
SFRP4	NM_003014.2	TACAGGATGAGGCTGGC	277	GTGTAGGGCAAGGGGC	278	CTGGGACAGCTCTATGAAGCCA
SHMT2	NM_005412.4	AGCGGTGCTAGAGCTTGTA	281	ATGGCACTTCGGTCTCCA	282	CCATCACTGCCAAAGAAACACCTG
SLC22A3	NM_021977.2	ATGCTCAGCGAGTTTGACCT	285	CAGGATGCTTGGGTGAG	286	CAGCATCCAGGATTTGACACAGAC
SMAD4	NM_005359.3	GGACATTACTGCCCTGTTCA	289	ACCACTACTCAGGACGAGTGA	290	TGCTCCAGCCTCCCTTCCATTC
SRC	NM_003118.1	TCTCCCTGTACACTGGCAGTTC	293	AGCTCGGTGGGAGAGTGA	294	TGACCCAGCAGCCCTTGAAGG
SRD6A2	NM_005417.3	TGAGGAGTGGTATTTGGCAAGA	297	CTCTCGGTCTCTGCACTTA	298	AACCGCTGAGCTCCCTGCTGGTG
STAT5B	NM_003348.2	GTAGGTCTCTGGGTTCTG	301	CCAGTGGTGGTATGCTTCA	302	AGAACCACTCAGAAATCCCGAGC
STAT5B	NM_012448.1	CCAGTGGTGGTATGCTTCA	305	GGTACCATCTTTGTGTGG	306	CAGCCAGGACAACATGCGACGG
TGFB11	NM_01042454.1	GCTACTTTGAGCGCTTCTCG	309	GGTACCATCTTTGTGTGG	310	CAAGTCTGCTGCTTCTGCAACCCAGC
THBS2	NM_003247.2	CAAGACTGGCTACATCAGAGTCTTAGTG	313	CAGCGAGTGTGGTATAGTAGG	314	TGAGTCTGCTGCTGCTTCTTCTCAT
TNFRSF10B	NM_003842.2	CTCTGAGACAGTGTCTCGATGACT	317	CCATGAGGCCCAACTTCT	318	CAGACTTGGTGGCTTCTTGAATCC

Official Symbol	Sequence ID	SEQ ID NO	Forward Primer Sequence	SEQ ID NO	Reverse Primer Sequence	SEQ ID NO	Probe Sequence
TPM2	NM_213674.1	321	AGGAGATGCAGCTGAAGGAG	322	CCACCTCTTCATATTGCGG	323	CCAGCACATGGCTGAGGATTGAG
TPX2	NM_012112.2	325	TCAGCTGTGAGCTGGGATA	326	ACGGTCCTAGGTTTGAGGTTAAGA	327	CAGGTCCCATGCGGGCG
TUBB2A	NM_001069.1	329	CGAGGACGAGGCTTAAAAAC	330	ACCATGCTTGAGGACAACAG	331	TCTCAGATCAATGTCGTCATCCTTAGTGAA
UBE2T	NM_014176.1	333	TGTTCTCAAAATTGGCCACCAA	334	AGAGGTCAACACAGTTGCCGA	335	AGGTGCTTGAGAGACCATCCCTCAA
VCL	NM_003373.2	337	GATACCACAACTCCCATCAAGCT	338	TCCCTGTTAGGGCCATCAG	339	AGTGGCAGCCACGGCGGCG
ZFP36	NM_003407.1	341	CATTAAACCACACTCCCTGA	342	CCCCACCACATCATGAATCT	343	CAGTCCCCCAAGTGTGCAAGCTC

Gene Symbol	SEQ ID NO	Annotation
GPS1	172	AGTAAAGCAGGCTGCCAAGTGCCCTGCTGGCTTCCTTTCATCACTGTGACTTCCCTGAGCTGC
GSN	176	CTCTGCTAAGCGGTACATGAGAGCGGACCCAGCCCAATGGGATGGGAGCCCAATCAAGTGGTGAAGCAAGGCTTTGAGCC
GSTM1	180	AAGCTATGAGGAAAGAGTACATGAGGAGCGCTCTGATTATGACAAGCCAGTGGCTGATGAATAATCAAGCTGGGCC
GSTM2	184	CTGAGGCACTCCCTGAATGCTGAAGCTTACTACAGCTTTCTGGGAAAGCAGCCATGGTTCTTTGG
HLF	188	CACCCTGCAGGTGCTGAGACTAAGTGTCTGCCCCCTCAGGTGGGATCACTCTGCTCTAGGTACC
IGF1	192	TCCGAGCTGTATCTAAGGAGGCTGGAGATGTTGGCCACCCCTCAAGCTGCCAAGTCAGCTCGCTCTGTCCG
IGFBP2	196	GTGGACAGCACCATGAACATGTTGGCGGGGGAGGAGTCTGGCCCGGAAGCCCTCAAGTCGGGTATGAGG
IGFBP6	200	TGAACGACAGAGAACACAGAGGATCCAGGCCTCTACCAAGCTCCAGCCCAATCTCGGGGTGTCCAAGAC
IL5ST	204	GGCCTAATGTTCCAGATCTTCAAGAGTCAATTGCCAGTGGTCACTCACACTCTCTCCAAAGCAACAATTT
INHBA	208	GTGCCAGCCCATATAGACGACGCTCGGGTCTCACTGCTCTCCACTCAACAGTCAATCAACCACTACCG
ITGA7	212	GATATGATTGGTCTGCTTTTGTGCTCAGCCAGGACCTGGCCATCCGGATGAGTTGGGGAATGGAAATTTCT
JUN	216	GACTGCAAGATGGAAACACCTTCTATGACGATGCCCTCAACGCCCTGCTCCCGTCCGAGAGGACCTTATGGCTA
KLK2	220	AGTCTCGGATTGTGGAGGCTGGGAGTGTGAGAAAGCATTCCTCAACCTGGCAGGTGGCTGTGTACA
KRT15	224	GCTTGGTCTTCAGCAAGACTGAGGAGCTGAACAAAGGTGGCTCCAAACAGAAATGATCCAGACCAAGCAAG
KRT5	228	TCAGTGGAGAAAGAGTTGGACCAAGTCAACATCTCTGTGTCAAGCAGAGTGTTCCTCTGGATATGGCA
LAMB3	232	ACTGACCAAGCTGAGACTACTGACCCAGTATGGCAGTGGCAGTGAATGCTGCAAGTGTGAC
LGALS3	236	AGCGAAATGGCAGACAATTTTCGCTCCATGATGCTGCTGCTGGTCTGGAACCCCAACCCCTCAAG
MMF11	240	CCTGGAGCTGCAACATACCTCAATCTCTCCAGCGGATCTCTCTGAAGCCCTTTCCGAGCACTCTCTCCAAAGCCATTGTA
MYBL2	244	GCCGAGATGCCAAGATGTTCCAGGAGGACAGACAATGCTGTGAAGATCACTGGAATCTACCATCAAAAG
NFAT5	248	CTGAACCCCTCTCTGTCACCGAATCACTCCCGTGGAGTTCCCTCCACCTCGCCATCTGCTTTCT
OLFML3	252	TCAGAACTGAGCGGACACATCTCGGAGAGTGGATGCTCTGGAGCGGGAGGTAGACTATCTGG
PAGE4	256	GAATCTCAGCAAGAGAACCAACCACTGACAATGAGATTTGCTCAAGCAGCCCTGGTGGCTGCTGCTAGT
PGK1	260	AGAGCAGTTGCTAGAACTCAATCTCTGCTGGCAAGGATGTTCTGTTCTTGAAGCACTGTAGGCCAG
PPAP2B	264	ACAAGCACCCTCCAGTGATGTTCTGGCAGGATTTCTCAAGCAGCCCTGGTGGCTGCTGCTAGT
PPP1R12A	268	CGGCAAGGGTTGATATAGAAGCAGCTGGAAGAGAAAGGATGCTTAGAGATGCCAGGCA
PRKCA	272	CAAGCAATGCGTCATCAATGTCGCCAGCTCTGGGAATGGATCACTGAGAGAGGGGGGCGGATTTAC
SDC1	276	GAATTTGACGAGGGTGTCTTGGCAGAGCTGGCTCTGAGCGCTCCATCCAAAGGCCAGGTTCTCCGTTAGCTCT
SFRP4	280	TACAGATGAGGCTGGGCTATGCTGGGACAGCTATGTAAGCCATGTGCCCTTGGCTTACAAAC
SHMT2	284	AGCGGTGCTAGAGCTTGTATCCATCACTGCCAACAGAACACCTGCTGGAGACCGAAGTCCCAT
SIL22A3	288	ATCGTCAGCGAGTTTGACCTGTCTGTGTCATGGGTGGATGCTGGAGCTCAACCAAGCCATCTTG
SMAD4	292	GGACATTACTGGCCTGTTCAATGAGCTTGCTCCAGCCCTCCATTTCCATCATCTCTGCTGAGTATGGT
SPARC	296	TCITCCCTGTACACTGGCAGTTTGGCCAGCTGGACAGCCCACTGAGGGTACCTCTCCACACCGAGCT
SRC	300	TGAGGAGTGGTATTTTGGCAAGTACCAAGCGGGAGTCAGAGGGTACTGCTCAATGCAGAGACCCGAGAG
SRD5A2	304	GTAGGTCTCTGCGCTTCTGCAAGTGGCTGGGATTTCTGAGTGTGTCTGTCTAGAGTTTACTCTACCTTCCAGGGA
STAT5B	308	CCAGTGGTGGTGTCTGAGAGCCAGGACCAATGCGAGCCCACTGCTCCGACACAAAGTGGTACC
TGFBH1	312	GCTACTTTGAGCGCTTCTGCCAAGTATGGCTTCTGCAACCGCCCATCCGACACAAAGTGGTACC
THBS2	316	CAAGACTGGCTACATCAGACTTATGCTATGAGGAAACAGGCTATGGCAGACTCAGGACCTATCTATGACCAACCTAGGCTG
TNFRSF10B	320	CTCTGAGACAGTCTTGATGACTTTGAGACTTGGTGCCCTTTGACTCTGGAGCCGCTCATGAGGAAGTTGGGCTCATGG
TPM2	324	AGGAGATGCAGCTGAAGAGGCCAAGCACA TCGCTGAGGATTCAGACCGCAATATGAAGAGGTGG
TPX2	328	TCAGCTGTGAGCTGGGATACCGCGCAATGGGACCTGCTCTTAACCTCAAACTAGGACCGT
TUBB2A	332	CGAGCAGAGGCTTAAAACTCTCAGATCAATGCTGCTAGTGAACCTCTGTTGCTCTCAAGCATGGT
UBE2T	336	TGTTCTAAATGGCAACCAAGGTGCTTGGAGACCATCTCCATCACTCGCAACTGTGTGACCTCT

Original Symbol	SEQ ID NO.	Amplicon Sequence
VCL	340	GATACACAACCTCCCATCAAGCTGTGGCAGTGGCAGCCACGGGCGCTCCTGATGGCGCTAACAGGGA
ZFP36	344	CATTAAACCACTCCCTGACCTGACGCTGGGGGACGTCCCAAGTGTGCAAGCTCAGTATTCATGATGGTGGGGG

CLAIMS

1. A method of predicting a likelihood of a clinical outcome for a patient with prostate cancer, comprising:
 - (a) measuring levels of RNA transcripts of each of genes FAM13C, KLK2, AZGP1, and SRD5A2 in a biological sample containing cancer cells obtained from the patient;
 - (b) normalizing the levels of the RNA transcripts to obtain normalized RNA expression levels;
 - (c) calculating an androgen group score for the patient from the normalized RNA expression levels; and
 - (d) predicting the likelihood of a clinical outcome for the patient based on the androgen group score, wherein decreased normalized RNA expression of each of FAM13C, KLK2, AZGP1, and SRD5A2 correlates with an increased likelihood of a negative clinical outcome.
2. The method of claim 1, wherein the androgen group score equals $0.634 \times \text{FAM13C} + 1.079 \times \text{KLK2} + 0.642 \times \text{AZGP1} + 0.997 \times \text{SRD5A2}$ Threshold score, wherein the gene symbols in the equation represent the normalized RNA expression levels of the respective genes, and wherein the SRD5A2 Threshold score equals 5.5 if SRD5A2 is less than 5.5 and equals SRD5A2 if SRD5A2 is 5.5 or greater.
3. The method of claim 1 or claim 2, wherein the levels of the RNA transcripts of FAM13C, KLK2, AZGP1, and SRD5A2 are normalized relative to the levels of the RNA transcripts of ARF1, ATP5E, CLTC, GPS1, and PGK1.
4. The method of any one of claims 1-3, wherein the clinical outcome is upgrading of prostate cancer.
5. The method of any one of claims 1-3, wherein the clinical outcome is upstaging of prostate cancer.
6. The method of any one of claims 1-3, wherein the clinical outcome is recurrence of prostate cancer.

7. The method of any one of claims 1-3, wherein the clinical outcome is non-organ-confined disease.
8. The method of any one of claims 1-3, wherein the clinical outcome is adverse pathology.
9. The method of any one of claims 1-8, wherein the levels of the RNA transcripts are measured by a method comprising quantitative polymerase chain reaction.
10. The method of any one of claims 1-9, further comprising creating a report summarizing the likelihood of a clinical outcome.
11. The method of any one of claims 1-10, wherein the biological sample containing cancer cells is fixed, paraffin-embedded, or fresh, or frozen.
12. The method of any one of claims 1-11, wherein part (d) comprises predicting the likelihood of a clinical outcome for the patient based on the androgen group score of the sample as compared to androgen group scores for reference prostate cancer samples.
13. A method of predicting a likelihood of a clinical outcome for a patient with prostate cancer, comprising:
 - (a) measuring levels of RNA transcripts of each of genes FAM13C, KLK2, AZGP1, and SRD5A2, and of at least one of BGN, COL1A1, SFRP4, FLNC, GSN, GSTM2, and TPM2 in a biological sample comprising cancer cells obtained from the patient;
 - (b) normalizing the levels of the RNA transcripts to obtain normalized RNA expression levels;
 - (c) calculating a stromal response group, a cellular organization group, and an androgen group score for the patient from the normalized RNA expression levels, wherein the genes are grouped into stromal response, cellular organization, and androgen groups as follows:
 - a) stromal response group: BGN, COL1A1, and SFRP4,

b) cellular organization group: FLNC, GSN, GSTM2, and TPM2, and
 c) androgen group: FAM13C, KLK2, AZGP1, and SRD5A2; and
 (d) predicting the likelihood of a clinical outcome for the patient based on the stromal response group, cellular organization group, and androgen group scores, wherein increased normalized RNA expression levels of BGN, COL1A1, and SFRP4 correlate with an increased likelihood of a negative clinical outcome, and wherein decreased normalized RNA expression levels of FLNC, GSN, GSTM2, TPM2, FAM13C, KLK2, AZGP1, and SRD5A2 correlate with an increased likelihood of a negative clinical outcome.

14. The method of claim 13, further comprising determining a normalized TPX2 RNA expression level, wherein the normalized TPX2 RNA expression level is used to calculate a proliferation group score, and wherein an increased normalized TPX2 RNA expression level correlates with an increased likelihood of a negative clinical outcome.

15. The method of claim 13 or 14, wherein the androgen group score equals $0.634 \times \text{FAM13C} + 1.079 \times \text{KLK2} + 0.642 \times \text{AZGP1} + 0.997 \times \text{SRD5A2 Threshold score}$, wherein the gene symbols in the equation represent the normalized RNA expression levels of the respective genes, and wherein the SRD5A2 Threshold score equals 5.5 if SRD5A2 is less than 5.5 and equals SRD5A2 if SRD5A2 is 5.5 or greater.

16. The method of any one of claims 13-15, wherein the stromal response group score equals $0.527 \times \text{BGN} + 0.457 \times \text{COL1A1} + 0.156 \times \text{SFRP4}$, wherein the gene symbols in the equation represent the normalized RNA expression levels of the respective genes.

17. The method of any one of claims 13-16, wherein the cellular organization group score equals $0.163 \times \text{FLNC} + 0.504 \times \text{GSN} + 0.421 \times \text{TPM2} + 0.394 \times \text{GSTM2}$, wherein the gene symbols in the equation represent the normalized RNA expression levels of the respective genes.

18. The method of any one of claims 14-17, wherein the proliferation group score equals TPX2 Threshold score, wherein TPX2 Threshold score is 5.0 if $\text{TPX2} < 5.0$

and TPX2 otherwise, wherein the gene symbols in the equation represent the normalized RNA expression levels of the respective genes.

19. The method of any one of claims 13-18, wherein the levels of the RNA transcripts are normalized relative to the levels of the RNA transcripts of ARF1, ATP5E, CLTC, GPS1, and PGK1.
20. The method of any one of claims 13-19, wherein the clinical outcome is upgrading of prostate cancer.
21. The method of any one of claims 13-19, wherein the clinical outcome is upstaging of prostate cancer.
22. The method of any one of claims 13-19, wherein the clinical outcome is recurrence of prostate cancer.
23. The method of any one of claims 13-19, wherein the clinical outcome is non-organ-confined disease.
24. The method of any one of claims 13-19, wherein the clinical outcome is adverse pathology.
25. The method of any one of claims 13-24, wherein the levels of the RNA transcripts are measured by a method comprising quantitative polymerase chain reaction.
26. The method of any one of claims 13-25, further comprising creating a report summarizing the likelihood of a clinical outcome.
27. The method of any one of claims 13-26, wherein the biological sample containing cancer cells is fixed, paraffin-embedded, or fresh, or frozen.
28. The method of any one of claims 13-27, wherein part (d) comprises predicting the likelihood of a clinical outcome for the patient based on the stromal response,

cellular organization, and androgen group scores of the sample as compared to stromal response, cellular organization, and androgen group scores for reference prostate cancer samples.

29. A method of predicting a likelihood of a clinical outcome for a patient with prostate cancer, comprising:

(a) determining levels of RNA transcripts of each of the following genes: BGN, COL1A1, COL3A1, SFRP4, FLNC, GSN, TPM2, GSTM2, FAM13C, KLK2, AZGP1, SRD5A2, and TPX2 in a biological sample containing cancer cells obtained from the patient;

(b) normalizing the levels of the RNA transcripts to obtain normalized RNA expression levels;

(c) calculating a quantitative score (QS) for the patient, wherein the QS is calculated as follows:

$$\begin{aligned} \text{QS} = & 0.735 \times \text{stromal response group score} \\ & -0.368 \times \text{cellular organization group score} \\ & -0.352 \times \text{androgen group score} \\ & + 0.095 \times \text{proliferation group score,} \end{aligned}$$

wherein

$$\begin{aligned} \text{the stromal response group score} = & 0.527 \times \text{BGN} + 0.457 \times \text{COL1A1} + \\ & 0.156 \times \text{SFRP4,} \end{aligned}$$

$$\begin{aligned} \text{the cellular organization group score} = & 0.163 \times \text{FLNC} + 0.504 \times \text{GSN} \\ & + 0.421 \times \text{TPM2} + 0.394 \times \text{GSTM2,} \end{aligned}$$

$$\begin{aligned} \text{the androgen group score} = & 0.634 \times \text{FAM13C} + 1.079 \times \text{KLK2} + 0.642 \\ & \times \text{AZGP1} + 0.997 \times \text{SRD5A2 Threshold score,} \end{aligned}$$

$$\text{the proliferation score} = \text{TPX2 Threshold score,}$$

wherein

$$\begin{aligned} \text{the SRD5A2 Threshold Score} = & \{5.5 \text{ if } \text{SRD5A2} < 5.5 \text{ and } \text{SFD5A2} \\ & \text{otherwise}\}, \text{ and} \end{aligned}$$

$$\text{the TPX Threshold score} = \{5.0 \text{ if } \text{TPX2} < 5.0 \text{ and } \text{TPX2 otherwise}\},$$

wherein the gene symbols in the equations represent the normalized RNA expression levels of the respective genes; and

(d) predicting a likelihood of a clinical outcome for the patient based on the quantitative score.

30. The method of claim 29, wherein the clinical outcome is upgrading of prostate cancer.
31. The method of claim 29, wherein the clinical outcome is upstaging of prostate cancer.
32. The method of claim 29, wherein the clinical outcome is recurrence of prostate cancer.
33. The method of claim 29, wherein the clinical outcome is non-organ-confined disease.
34. The method of claim 29, wherein the clinical outcome is adverse pathology.
35. The method of any one of claims 29-34, wherein the levels of the RNA transcripts are measured by a method comprising quantitative polymerase chain reaction.
36. The method of any one of claims 29-35, further comprising creating a report summarizing the likelihood of a clinical outcome.
37. The method of any one of claims 29-36, wherein the biological sample containing cancer cells is fixed, paraffin-embedded, or fresh, or frozen.