

**POLYPEPTIDE-BASED SHUTTLE AGENTS FOR IMPROVING THE TRANSDUCTION EFFICIENCY OF
POLYPEPTIDE CARGOS TO THE CYTOSOL OF TARGET EUKARYOTIC CELLS, USES THEREOF, METHODS
AND KITS RELATING TO SAME**

5 The present description relates to synthetic peptides useful for increasing the transduction efficiency of polypeptide cargos to the cytosol of target eukaryotic cells. More specifically, the present description relates to synthetic peptides and polypeptide-based shuttle agents comprising an endosome leakage domain (ELD) operably linked to a cell penetrating domain (CPD), or an ELD operably linked to a histidine-rich domain and a CPD.

10 **BACKGROUND**

Cell delivery technologies to transport large molecules inside eukaryotic cells have a wide range of applications, particularly in the biopharmaceutical industry. While some soluble chemical substances (e.g., small molecule drugs) may passively diffuse through the eukaryotic cell membrane, larger cargos (e.g., biologics, polynucleotides, and polypeptides) require the help of shuttle agents to reach their intracellular targets.

15 An area that would greatly benefit from advances in cell delivery technologies is the field of cell therapy, which has made enormous leaps over the last two decades. Deciphering the different growth factors and molecular cues that govern cell expansion, differentiation and reprogramming open the door to many therapeutic possibilities for the treatment of unmet medical needs. For example, induction of pluripotent stem cells directly from adult cells, direct cell conversion (trans-differentiation), and genome editing (Zinc finger nuclease, TALEN™ and CRISPR/Cas9 technologies) are examples of methods that have been developed to maximize the therapeutic value of cells for clinical applications. Presently, the production of cells with high therapeutic activity usually requires *ex vivo* manipulations, mainly achieved by viral transduction, raising important safety and economical concerns for human applications. The ability to directly deliver active proteins such as transcription factors or artificial nucleases, inside these cells, may advantageously circumvent the safety concerns and regulatory hurdles associated with more risky gene transfer methods.

25 In this regard, polypeptide-based transduction agents may be useful for introducing purified recombinant proteins directly into target cells, for example, to help bypass safety concerns regarding the introduction of foreign DNA. Lipid- or cationic polymer-based transduction agents exist, but introduce safety concerns regarding chemical toxicity and efficiency, which hamper their use in human therapy. Protein transduction approaches involving fusing a recombinant protein cargo directly to a cell-penetrating peptide (e.g., HIV transactivating protein TAT) require large amounts of the recombinant protein and often fail to deliver the cargo to the proper subcellular location, leading to massive endosomal trapping and eventual degradation. Several endosomal membrane disrupting peptides have been developed to try and facilitate the escape of endosomally-trapped cargos to the cytosol. However, many of these endosomolytic peptides are intended to alleviate endosomal entrapment of cargos that have already been delivered intracellularly, and do not by themselves aid in the initial step of shuttling the cargos intracellularly across

the plasma membrane (Salomone et al., 2012; Salomone et al., 2013; Erazo-Oliveras et al., 2014; Fasoli et al., 2014). Thus, there is a need for improved shuttle agents capable of increasing the transduction efficiency of polypeptide cargos, and delivering the cargos to the cytosol of target eukaryotic cells.

The present description refers to a number of documents, the content of which is herein incorporated by reference in their entirety.

SUMMARY

The present description stems from the surprising discovery that synthetic peptides comprising an endosome leakage domain (ELD) operably linked to a cell penetrating domain (CPD) and optionally a histidine-rich domain, have the ability to increase the proportion of cells that can be transduced with a polypeptide cargo of interest, without the synthetic peptide being covalently bound to the polypeptide cargo. Following successful transduction, the synthetic peptides may facilitate the ability of endosomally-trapped polypeptide cargos to gain access to the cytosol, and optionally be targeted to various subcellular compartments (e.g., the nucleus).

Accordingly, the present description may relate to the following aspects:

- (1) A synthetic peptide comprising an endosome leakage domain (ELD) operably linked to a cell penetrating domain (CPD), or an ELD operably linked to a histidine-rich domain and a CPD.
- (2) A polypeptide-based shuttle agent comprising an endosome leakage domain (ELD) operably linked to a cell penetrating domain (CPD), or an ELD operably linked to a histidine-rich domain and a CPD, for use in increasing the transduction efficiency of an independent polypeptide cargo to the cytosol of a target eukaryotic cell.
- (3) The synthetic peptide or polypeptide-based shuttle agent of (1) or (2), wherein the synthetic peptide or polypeptide-based shuttle agent: (a) comprises a minimum length of 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30 amino acid residues and a maximum length of 35, 40, 45, 50, 55, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, or 150 amino acid residues; (b) has a predicted net charge of at least +6, +7, +8, +9, +10, +11, +12, +13, +14, or +15 at physiological pH; (c) is soluble in aqueous solution; or (d) any combination of (a) to (c).
- (4) The synthetic peptide or polypeptide-based shuttle agent of any one of (1) to (3), wherein: (a) the ELD is or is from: an endosomolytic peptide; an antimicrobial peptide (AMP); a linear cationic alpha-helical antimicrobial peptide; a Cecropin-A/Melittin hybrid (CM series) peptide; pH-dependent membrane active peptide (PAMP); a peptide amphiphile; a peptide derived from the N terminus of the HA2 subunit of influenza hemagglutinin (HA); CM18; Diphtheria toxin T domain (DT); GALA; PEA; INF-7; LAH4; HGP; H5WYG; HA2; EB1; VSVG; *Pseudomonas* toxin; melittin; KALA; JST-1; C(LLKK)₃C; G(LLKK)₃G; or any combination thereof; (b) the CPD is or is from: a cell-penetrating peptide or the protein transduction domain from a cell-penetrating peptide; TAT; PTD4; Penetratin (Antennapedia); pVEC; M918; Pep-1; Pep-2; Xentry; arginine stretch; transportan;

SynB1; SynB3; or any combination thereof; (c) the histidine-rich domain is a stretch of at least 3, at least 4, at least 5, or at least 6 amino acids comprising at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, or at least 90% histidine residues; and/or comprises at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, or at least 9 consecutive histidine residues; or
 5 (d) any combination of (a) to (c).

(5) The synthetic peptide or polypeptide-based shuttle agent of any one of (1) to (4), wherein the synthetic peptide or polypeptide-based shuttle agent comprises: (a) an ELD comprising the amino acid sequence of any one of **SEQ ID NOs: 1-15, 63, or 64**, or a variant or fragment thereof having endosomolytic activity; (b) a CPD comprising the amino acid sequence of any one of **SEQ ID NOs: 16-27 or 65**, or a variant or fragment thereof
 10 having cell penetrating activity; (c) a histidine-rich domain having at least 2, at least 3, at least 4, at least 5, or at least 6 consecutive histidine residues; (d) of any combination of (a) to (c).

(6) The synthetic peptide or polypeptide-based shuttle agent of any one of (1) to (5), wherein the domains are operably linked via one or more linker domains.

(7) The synthetic peptide or polypeptide-based shuttle agent of any one of (1) to (6), wherein the synthetic peptide or polypeptide-based shuttle agent comprises at least two different types of CPDs and/or ELDs.
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(8) The synthetic peptide or polypeptide-based shuttle agent of any one of (1) to (7), wherein the synthetic peptide or polypeptide-based shuttle agent comprises: (a) an ELD which is CM18, KALA, or C(LLKK)₃C having the amino acid sequence of **SEQ ID NO: 1, 14, or 63**, or a variant thereof having at least 85%, 90%, or 95% identity to **SEQ ID NO: 1** and having endosomolytic activity; (b) a CPD which is TAT or PTD4 having the amino acid sequence of **SEQ ID NO: 17 or 65**, or a variant thereof having at least 85%, 90%, or 95% identity
 20 to **SEQ ID NO: 17 or 65**, and having cell penetrating activity; or Penetratin having the amino acid sequence of **SEQ ID NO: 18**, or a variant thereof having at least 85%, 90%, or 95% identity to **SEQ ID NO: 18** and having cell penetrating activity; (c) a histidine-rich domain comprising at least 6 consecutive histidine residues; or (d) any combination of (a) to (c).

(9) The synthetic peptide or polypeptide-based shuttle agent of any one of (1) to (8), wherein the synthetic peptide or polypeptide-based shuttle agent comprises or consists of the amino acid sequence of any one of **SEQ ID NOs: 57-59, 66-73, or 82-102**, or a functional variant thereof having at least 85%, 90%, or 95% identity to any one of **SEQ ID NOs: 57-59, 66-73, or 82-102**.
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(10) The synthetic peptide or polypeptide-based shuttle agent of any one of (1) to (9), wherein the synthetic peptide or polypeptide-based shuttle agent is non-toxic and/or is metabolizable.
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(11) A composition comprising: (a) the synthetic peptide or polypeptide-based shuttle agent as defined in any one of (1) to (10), and a further independent synthetic peptide comprising a histidine-rich domain and a CPD; and/or (b) a cocktail of at least 2, at least 3, at least 4, or at least 5 different types of the synthetic peptides or polypeptide-based shuttle agents as defined in any one of (1) to (10).

- (12) Use of the synthetic peptide, polypeptide-based shuttle agent, or composition as defined in any one of (1) to (11), for delivering an independent polypeptide cargo to the cytosol of a target eukaryotic cell.
- (13) A method for increasing the transduction efficiency of a polypeptide cargo to the cytosol of a target eukaryotic cell, the method comprising contacting the target eukaryotic cell with the synthetic peptide, polypeptide-based shuttle agent, or composition as defined in any one of (1) to (11), and the polypeptide cargo.
- (14) A kit for increasing the transduction efficiency of a polypeptide cargo to the cytosol of a target eukaryotic cell, the kit comprising the synthetic peptide, polypeptide-based shuttle agent, or composition as defined in any one of (1) to (11), and a suitable container.
- (15) The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of any one of (1) to (14), for use in increasing the transduction efficiency of a polypeptide cargo to the cytosol of a target eukaryotic cell in the presence of serum.
- (16) The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of any one of (2) to (15), wherein the polypeptide cargo: (a) comprises or lacks a CPD or a CPD as defined in (4)(b); (b) is a recombinant protein; (c) comprises a subcellular targeting domain; (d) is complexed with a DNA and/or RNA molecule; or (e) any combination of (a) to (d).
- (17) The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of (16), wherein the subcellular targeting domain is: (a) a nuclear localization signal (NLS); (b) a nucleolar signal sequence; (c) a mitochondrial signal sequence; or (d) a peroxisome signal sequence.
- (18) The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of (17), wherein: (a) the NLS is from: E1a, T-Ag, c-myc, T-Ag, op-T-NLS, Vp3, nucleoplasmin, histone 2B, Xenopus N1, PARP, PDX-1, QKI-5, HCDA, H2B, v-Rel, Amida, RanBP3, Pho4p, LEF-1, TCF-1, BDV-P, TR2, SOX9, or Max; (b) the nucleolar signal sequence is from BIRC5 or RECQL4; (c) the mitochondrial signal sequence is from Tim9 or Yeast cytochrome c oxidase subunit IV; or (d) the peroxisome signal sequence is from PTS1.
- (19) The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of any one of (2) to (18), wherein the polypeptide cargo is a transcription factor, a nuclease, a cytokine, a hormone, a growth factor, or an antibody.
- (20) The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of (19), wherein: (a) the transcription factor is: HOXB4, NUP98-HOXA9, Oct3/4, Sox2, Sox9, Klf4, c-Myc, MyoD, Pdx1, Ngn3, MafA, Blimp-1, Eomes, T-bet, FOXO3A, NF-YA, SALL4, ISL1, FoxA1, Nanog, Esrrb, Lin28, HIF1-alpha, Hlf, Runx1t1, Pbx1, Lmo2, Zfp37, Prdm5, Bcl-6, or any combination thereof; and/or the nuclease is: an RNA-guided endonuclease, a CRISPR endonuclease, a type I CRISPR endonuclease, a type II CRISPR endonuclease, a type III CRISPR endonuclease, a type IV CRISPR endonuclease, a type V CRISPR endonuclease, a type VI CRISPR endonuclease, CRISPR associated protein 9 (Cas9), Cpf1, a zinc-finger

nuclease (ZFN), a Transcription activator-like effector nucleases (TALENs), a homing endonuclease, a meganuclease, or any combination thereof.

- (21) The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of any one of (1) to (20), for use in cell therapy, genome editing, adoptive cell transfer, and/or regenerative medicine.
- 5 (22) The shuttle agent, shuttle system, composition, use, method, or kit of any one of (2) to (21), wherein the target eukaryotic cell is a stem cell, a primary cell, an immune cell, a T cell, or a dendritic cell.
- (23) A eukaryotic cell comprising the synthetic peptide or polypeptide-based shuttle agent as defined in any one of (1) to (10), or the composition of (11).
- (24) The eukaryotic cell of (23), wherein said cell further comprises an independent polypeptide cargo delivered intracellularly by said synthetic peptide or polypeptide-based shuttle agent.
- 10 (25) A method for delivering an independent polypeptide cargo to the cytosol of a target eukaryotic cell, said method comprising contacting said target eukaryotic cell with the synthetic peptide or polypeptide-based shuttle agent as defined in any one of (1) to (10), or the composition of (11); and an independent polypeptide cargo to be delivered intracellularly by said synthetic peptide or polypeptide-based shuttle agent.
- 15 (26) The eukaryotic cell of (23) or (24), or the method of (25), wherein said independent polypeptide cargo is as defined in any one of (16) to (20).
- (27) The eukaryotic cell of (24) or (26), or the method of (25) or (26), wherein said independent polypeptide cargo is as defined in any one of (16) to (20).
- (28) The eukaryotic cell of (23), (24), (26) or (27), or the method of (25), (26), or (27), wherein said eukaryotic cell is an animal cell, a mammalian cell, a human cell, a stem cell, a primary cell, an immune cell, a T cell, or a dendritic cell.
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In some aspects, the present description may relate to one or more of the following items:

- 1. A method for increasing the transduction efficiency of an independent polypeptide cargo to the cytosol of a target eukaryotic cell, said method comprising contacting said target eukaryotic cell with a synthetic peptide and said independent polypeptide cargo, wherein said synthetic peptide:
 - (a) comprises an endosome leakage domain (ELD), or a variant or fragment thereof having endosomolytic activity, operably linked to a cell penetrating domain (CPD), wherein said ELD comprises the amino acid sequence of any one of SEQ ID NOs: 1-15, 63, or 64;
 - (b) is not covalently bound to said independent polypeptide cargo;
 - 30 (c) has an overall length of between 20 and 100 amino acid residues;
 - (d) has a net charge of at least +6 at physiological pH; and
 - (e) is soluble in aqueous solution at physiological pH,

wherein said CPD enables intracellular delivery of said synthetic peptide, and said ELD enables escape of endosomally trapped independent polypeptide cargo to the cytosol of the target eukaryotic cell.

2. The method of item 1, wherein said synthetic peptide has an overall length of between 20 and 70 amino acid residues.
3. The method of item 1, wherein said CPD comprises the amino acid sequence of any one of **SEQ ID NOs: 16-27 or 65**, or is a variant or fragment thereof having cell penetrating activity.
4. The method of item 1, wherein said synthetic peptide further comprises a histidine-rich domain consisting of a stretch of at least 6 amino acids comprising at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, or at least 90% histidine residues; and/or comprises at least 2, at least 3, at least 4, at least 5, or at least 6 consecutive histidine residues.
5. The method of item 1, wherein said ELD variant or ELD fragment has at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, or at least 95% sequence identity to any one of **SEQ ID NOs: 1-15, 63, or 64**.
6. The method of item 3, wherein said CPD variant or CPD fragment has at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, or at least 95% sequence identity to any one of **SEQ ID NOs: 16-27 or 65**.
7. The method of item 1, wherein said ELD and CPD are operably linked via one or more linker domains.
8. The method of item 1, wherein said synthetic peptide is chemically synthesized without an N-terminal methionine residue.
9. The method of item 1, wherein the synthetic peptide comprises the amino acid sequence of any one of **SEQ ID NOs: 57-59, 66-72, or 82-102**, or a functional variant thereof having at least 70%, at least 85%, at least 90%, or at least 95% identity to any one of **SEQ ID NOs: 57-59, 66-72, or 82-102**.
10. The method of item 1, wherein said independent polypeptide cargo is a recombinant protein lacking a CPD.
11. The method of item 1, wherein said independent polypeptide cargo is a transcription factor, a nuclease, a cytokine, a hormone, a growth factor, or an antibody.
12. The method of item 11, wherein:
 - (b) said transcription factor is: HOXB4, NUP98-HOXA9, Oct3/4, Sox2, Sox9, Klf4, c-Myc, MyoD, Pdx1, Ngn3, MafA, Blimp-1, Eomes, T-bet, FOXO3A, NF-YA, SALL4, ISL1, FoxA1, Nanog, Esrrb, Lin28, HIF1-alpha, Hlf, Runx1t1, Pbx1, Lmo2, Zfp37, Prdm5, Bcl-6, or any combination thereof; or
 - (b) said nuclease is an RNA-guided endonuclease, a CRISPR endonuclease, a type I CRISPR endonuclease, a type II CRISPR endonuclease, a type III CRISPR endonuclease, a type IV CRISPR endonuclease, a type V CRISPR endonuclease, a type VI CRISPR endonuclease, CRISPR associated protein 9 (Cas9), Cpf1, a zinc-finger nuclease (ZFNs), a Transcription activator-like effector nucleases (TALENs), a homing endonuclease, or a meganuclease.
13. The method of item 11, wherein said nuclease is Cas9 or Cpf1.
14. The method of item 13, wherein said nuclease further comprises a guide RNA, a crRNA, a tracrRNA, or both a crRNA and a tracrRNA.

15. The method of item 1, wherein said independent polypeptide cargo comprises a nuclear localization signal or a further nuclear localization signal.
16. The method of item 15, wherein said independent polypeptide cargo is a transcription factor or a nuclease.
- 5 17. The method of item 16 wherein:
- (a) said transcription factor is: HOXB4, NUP98-HOXA9, Oct3/4, Sox2, Sox9, Klf4, c-Myc, MyoD, Pdx1, Ngn3, MafA, Blimp-1, Eomes, T-bet, FOXO3A, NF-YA, SALL4, ISL1, FoxA1, Nanog, Esrrb, Lin28, HIF1-alpha, Hlf, Runx1t1, Pbx1, Lmo2, Zfp37, Prdm5, Bcl-6, or any combination thereof; or
- 10 (b) said nuclease is an RNA-guided endonuclease, a CRISPR endonuclease, a type I CRISPR endonuclease, a type II CRISPR endonuclease, a type III CRISPR endonuclease, a type IV CRISPR endonuclease, a type V CRISPR endonuclease, a type VI CRISPR endonuclease, CRISPR associated protein 9 (Cas9), Cpf1, a zinc-finger nuclease (ZFNs), a Transcription activator-like effector nucleases (TALENs), a homing endonuclease, or a meganuclease.
18. The method of item 17, wherein said nuclease is Cas9 or Cpf1.
- 15 19. The method of item 18, wherein said nuclease further comprises a guide RNA.
20. The method of item 1, wherein said cell is stem cell, a primary cell, an immune cell, a T cell, or a dendritic cell.
21. A method for increasing the transduction efficiency of an independent polypeptide cargo to the cytosol of a target eukaryotic cell, said method comprising contacting said target eukaryotic cell with a synthetic peptide and said
- 20 independent polypeptide cargo, wherein said synthetic peptide:
- (a) comprises an endosome leakage domain (ELD) operably linked to a cell penetrating domain (CPD), wherein said ELD is an endosomolytic peptide which is, or is derived from: a linear cationic alpha-helical antimicrobial peptide; a Cecropin-A/Melittin hybrid (CM series) peptide; pH-dependent membrane active peptide (PAMP); a peptide amphiphile; a peptide derived from the N terminus of the
- 25 HA2 subunit of influenza hemagglutinin (HA); CM18; Diphtheria toxin T domain (DT); GALA; PEA; INF-7; LAH4; HGP; H5WYG; HA2; EB1; VSVG; *Pseudomonas* toxin; melittin; KALA; JST-1; C(LLKK)₃C; or G(LLKK)₃G;
- (b) is not covalently bound to said independent polypeptide cargo;
- (c) has an overall length of between 20 and 100 amino acid residues;
- 30 (d) has a net charge of at least +6 at physiological pH; and
- (e) is soluble in aqueous solution at physiological pH,
- wherein said CPD enables intracellular delivery of said synthetic peptide, and said ELD enables escape of endosomally trapped independent polypeptide cargo to the cytosol of the target eukaryotic cell.

22. The method of item 21, wherein said CPD is, or is derived from: a cell-penetrating peptide or the protein transduction domain from a cell-penetrating peptide; TAT; PTD4; Penetratin (Antennapedia); pVEC; M918; Pep-1; Pep-2; Xentry; arginine stretch; transportan; SynB1; SynB3; or any combination thereof.
- 5 23. The method of item 21, wherein said synthetic peptide further comprises a histidine-rich domain consisting of a stretch of at least 3 amino acids comprising at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, or at least 90% histidine residues; and/or comprises at least 2, at least 3, at least 4, at least 5, or at least 6 consecutive histidine residues.
24. The method of item 21, wherein said ELD and CPD are operably linked via one or more linker domains.
- 10 25. The method of item 21, wherein said independent polypeptide cargo is a transcription factor, a nuclease, a cytokine, a hormone, a growth factor, or an antibody.
26. The method of item 25, wherein said transcription factor is: HOXB4, NUP98-HOXA9, Oct3/4, Sox2, Klf4, c-Myc, MyoD, Pdx1, Ngn3, MafA, Blimp-1, Eomes, T-bet, FOXO3A, NF-YA, SALL4, ISL1, FoxA1, Nanog, Esrrb, Lin28, HIF1-alpha, Hlf, Runx1t1, Pbx1, Lmo2, Zfp37, Prdm5, Bcl-6, or any combination thereof.
- 15 27. The method of item 25, wherein said nuclease is an RNA-guided endonuclease, a CRISPR endonuclease, a type I CRISPR endonuclease, a type II CRISPR endonuclease, a type III CRISPR endonuclease, a type IV CRISPR endonuclease, a type V CRISPR endonuclease, a type VI CRISPR endonuclease, CRISPR associated protein 9 (Cas9), Cpf1, a zinc-finger nuclease (ZFNs), a Transcription activator-like effector nucleases (TALENs), a homing endonuclease, or a meganuclease.
- 20 28. The method of item 25, wherein said nuclease is Cas9 or Cpf1.
29. A method for increasing the transduction efficiency of an independent polypeptide cargo to the cytosol of a target eukaryotic cell, said method comprising contacting said target eukaryotic cell with a synthetic peptide and said independent polypeptide cargo which is not covalently bound to said synthetic peptide, wherein said synthetic peptide comprises an endosome leakage domain (ELD) operably linked to a cell penetrating domain, or an ELD
- 25 operably linked to a CPD and a histidine-rich domain, wherein:
- (a) said ELD comprises the amino acid sequence of any one of **SEQ ID NOs: 1-15, 63, or 64**;
 - (b) said CPD comprises the amino acid sequence of any one of **SEQ ID NOs: 16-27 or 65**; and
 - (c) said histidine-rich domain comprises at least two consecutive histidine residues.
30. A method for delivering a CRISPR associated protein 9 (Cas9) to the nucleus of a target eukaryotic cell, said method comprising contacting said eukaryotic cell with a Cas9 recombinant protein comprising a nuclear localization signal, and a separate synthetic peptide shuttle agent less than 100 residues in length and comprising an endosome leakage domain (ELD) operably linked to a cell penetrating domain, or an ELD operably linked to a CPD and a histidine-rich domain, wherein:
- 30 (a) said ELD comprises the amino acid sequence of any one of **SEQ ID NOs: 1-15, 63, or 64**;

- (b) said CPD comprises the amino acid sequence of any one of **SEQ ID NOs: 16-27 or 65**; and
- (c) said histidine-rich domain comprises at least two consecutive histidine residues.

General Definitions

5 Headings, and other identifiers, e.g., (a), (b), (i), (ii), etc., are presented merely for ease of reading the specification and claims. The use of headings or other identifiers in the specification or claims does not necessarily require the steps or elements be performed in alphabetical or numerical order or the order in which they are presented.

The use of the word “a” or “an” when used in conjunction with the term “comprising” in the claims and/or the
10 specification may mean “one” but it is also consistent with the meaning of “one or more”, “at least one”, and “one or more than one”.

The term “**about**” is used to indicate that a value includes the standard deviation of error for the device or method being employed to determine the value. In general, the terminology “about” is meant to designate a possible variation of up to 10%. Therefore, a variation of 1, 2, 3, 4, 5, 6, 7, 8, 9 and 10% of a value is included in the term
15 “about”. Unless indicated otherwise, use of the term “about” before a range applies to both ends of the range.

As used in this specification and claim(s), the words “**comprising**” (and any form of comprising, such as “comprise” and “comprises”), “having” (and any form of having, such as “have” and “has”), “including” (and any form of including, such as “includes” and “include”) or “containing” (and any form of containing, such as “contains” and “contain”) are inclusive or open-ended and do not exclude additional, un-recited elements or method steps.

20 As used herein, “**protein**” or “**polypeptide**” means any peptide-linked chain of amino acids, which may or may not comprise any type of modification (e.g., post-translational modifications such as acetylation, phosphorylation, glycosylation, sulfatation, sumoylation, prenylation, ubiquitination, etc).

As used herein, the expression “**is or is from**” or “**is from**” comprises functional variants of a given protein domain (CPD or ELD), such as conservative amino acid substitutions, deletions, modifications, as well as
25 variants or function derivatives, which do not abrogate the activity of the protein domain. Other objects, advantages and features of the present description will become more apparent upon reading of the following non-restrictive description of specific embodiments thereof, given by way of example only with reference to the accompanying drawings.

30 BRIEF DESCRIPTION OF THE DRAWINGS

In the appended drawings:

Figure 1 shows a typical result of a calcein endosomal escape assay in which HEK293A cells were loaded with the fluorescent dye calcein (“100 μ M calcein”), and were then treated (or not) with a shuttle agent that facilitates

endosomal escape of the calcein ("100 μ M calcein + CM18-TAT 5 μ M"). Panel **A** shows the results of a fluorescence microscopy experiment, while panel **B** shows the results of a flow cytometry experiment.

Figure 2 shows the results of a calcein endosomal escape flow cytometry assay in which HeLa cells were loaded with calcein ("calcein 100 μ M"), and were then treated with increasing concentrations of the shuttle agent CM18-TAT-Cys (labeled "CM18-TAT").

Figures 3 and 4 show the results of calcein endosomal escape flow cytometry assays in which HeLa cells (Figure 3) or primary myoblasts (Figure 4) were loaded with calcein ("calcein 100 μ M"), and were then treated with 5 μ M or 8 μ M of the shuttle agents CM18-TAT-Cys or CM18-Penetratin-Cys (labeled "CM18-TAT" and "CM18-Penetratin", respectively).

Figure 5 shows the results of a GFP transduction experiment visualized by fluorescence microscopy in which a GFP cargo protein was co-incubated with 0, 3 or 5 μ M of CM18-TAT-Cys (labeled "CM18-TAT"), and then exposed to HeLa cells. The cells were observed by bright field (upper panels) and fluorescence microscopy (lower panels).

Figure 6 shows the results of a GFP transduction efficiency experiment in which GFP cargo protein (10 μ M) was co-incubated with different concentrations of CM18-TAT-Cys (labeled "CM18-TAT"), prior to being exposed to HeLa cells. Cells were evaluated by flow cytometry and the percentage of fluorescent (GFP-positive) cell is shown in panel **A**, and corresponding cell toxicity data is shown in panel **B**.

Figure 7 shows the results of a GFP transduction efficiency experiment in which different concentrations of GFP cargo protein (10, 5 or 1 μ M) were co-incubated with either 5 μ M of CM18-TAT-Cys (panel A, labeled "CM18TAT"), or 2.5 μ M of dCM18-TAT-Cys (panel B, labeled "dCM18TAT"), prior to being exposed to HeLa cells. Cells were evaluated by flow cytometry and the percentages of fluorescent (GFP-positive) cells are shown.

Figures 8 and 9 show the results of GFP transduction efficiency experiments in which GFP cargo protein (10 μ M) was co-incubated with different concentrations and combinations of CM18-TAT-Cys (labeled "CM18TAT"), CM18-Penetratin-Cys (labeled "CM18penetratin"), and dimers of each (dCM18-TAT-Cys (labeled "dCM18TAT"), dCM18-Penetratin-Cys (labeled "dCM18penetratin"), prior to being exposed to HeLa cells. Cells were evaluated by flow cytometry and the percentages of fluorescent (GFP-positive) cells are shown.

Figure 10 shows typical results of a TAT-GFP transduction experiment in which TAT-GFP cargo protein (5 μ M) was co-incubated with 3 μ M of CM18-TAT-Cys (labeled "CM18-TAT"), prior to being exposed to HeLa cells. Cells and GFP fluorescence were visualized by bright field and fluorescence microscopy at 10x and 40x magnifications. Arrows indicate the endosome delivery of TAT-GFP in the absence of CM18-TAT-Cys, as well as its nuclear delivery in the presence of CM18-TAT-Cys.

Figure 11 shows the results of a TAT-GFP transduction efficiency experiment in which TAT-GFP cargo protein (5 μ M) was co-incubated with different concentrations of CM18-TAT-Cys (labeled "CM18TAT"), prior to being

exposed to HeLa cells. Cells were evaluated by flow cytometry and the percentage of fluorescent (GFP-positive) cell is shown in panel **A**, and corresponding cell toxicity data is shown in panel **B**.

Figure 12 shows typical results of a GFP-NLS transduction experiment in which GFP-NLS cargo protein (5 μ M) was co-incubated with 5 μ M of CM18-TAT-Cys (labeled "CM18-TAT"), prior to being exposed to HeLa cells for 5 minutes. Cells and GFP fluorescence were visualized by bright field and fluorescence microscopy at 10x, 20x, and 40x magnifications. Arrows indicate areas of nuclear delivery of GFP-NLS.

Figure 13 shows the results of a GFP-NLS transduction efficiency experiment in which GFP-NLS cargo protein (5 μ M) was co-incubated with different concentrations of CM18-TAT-Cys (labeled "CM18TAT"), prior to being exposed to HeLa cells. Cells were evaluated by flow cytometry and the percentage of fluorescent (GFP-positive) cell is shown in panel **A**, and corresponding cell toxicity data is shown in panel **B**.

Figures 14 and 15 show the results of GFP-NLS transduction efficiency experiments in which GFP-NLS cargo protein (5 μ M) was co-incubated with different concentrations and combinations of CM18-TAT (labeled "CM18TAT"), CM18-Penetratin (labeled "CM18penetratin"), and dimers of each (dCM18-TAT-Cys, dCM18-Penetratin-Cys; labeled "dCM18TAT" and "dCM18penetratin", respectively), prior to being exposed to HeLa cells. Cells were evaluated by flow cytometry and the percentages of fluorescent (GFP-positive) cells are shown.

Figure 16 shows the results of a GFP-NLS transduction efficiency experiment in which GFP-NLS cargo protein (5 μ M) was co-incubated with either CM18-TAT-Cys (3.5 μ M, labeled "CM18TAT") alone or with dCM18-Penetratin-Cys (1 μ M, labeled "dCM18pen") for 5 minutes or 1 hour in plain DMEM media ("DMEM") or DMEM media containing 10% FBS ("FBS"), before being subjected to flow cytometry analysis. The percentages of fluorescent (GFP-positive) cells are shown. Cells that were not treated with shuttle agent or GFP-NLS ("ctrl"), and cells that were treated with GFP-NLS without shuttle agent ("GFP-NLS 5 μ M") were used as controls.

Figure 17 shows the results of a GFP-NLS transduction efficiency experiment in which GFP-NLS cargo protein (5 μ M) was co-incubated with or without 1 μ M CM18-TAT-Cys (labeled "CM18TAT"), prior to being exposed to THP-1 cells. Cells were evaluated by flow cytometry and the percentage of fluorescent (GFP-positive) cells is shown in panel **A**, and corresponding cell toxicity data is shown in panel **B**.

Figure 18 shows the results of a transduction efficiency experiment in which the cargo protein, FITC-labeled anti-tubulin antibody (0.5 μ M), was co-incubated with 5 μ M of CM18-TAT-Cys (labeled "CM18-TAT"), prior to being exposed to HeLa cells. Functional antibody delivery was visualized by bright field (20x) and fluorescence microscopy (20x and 40x), in which fluorescent tubulin fibers in the cytoplasm were visualized.

Figure 19 shows the results of an FITC-labeled anti-tubulin antibody transduction efficiency experiment in which the antibody cargo protein (0.5 μ M) was co-incubated with 3.5 μ M of CM18-TAT-Cys (labeled "CM18TAT"), CM18-Penetratin-Cys (labeled "CM18pen") or dCM18-Penetratin-Cys (labeled "dCM18pen"), or a combination of 3.5 μ M of CM18-TAT-Cys and 0.5 μ M of dCM18-Penetratin-Cys, prior to being exposed to HeLa cells. Cells were

evaluated by flow cytometry and the percentage of fluorescent (FITC-positive) cell is shown in panel **A**, and corresponding cell toxicity data is shown in panel **B**.

Figure 20 shows the results of DNA transfection efficiency experiment in which plasmid DNA (pEGFP) was labeled with a Cy5TM dye was co-incubated with 0, 0.05, 0.5, or 5 μ M of CM18-TAT-Cys (labeled "CM18-TAT"), prior to being exposed to HEK293A cells. Flow cytometry analysis allowed quantification of Cy5TM emission (corresponding to DNA intracellular delivery; y-axis) and GFP emission (corresponding to successful nuclear delivery of DNA; percentage indicated above each bar).

Figure 21 shows the results of a GFP-NLS transduction efficiency experiment in which the GFP-NLS cargo protein (5 μ M) was co-incubated with 1, 3, or 5 μ M of CM18-TAT-Cys (labeled "CM18TAT"), of His-CM18-TAT (labeled "His-CM18TAT"), prior to being exposed to HeLa cells. Cells were evaluated by flow cytometry and the percentage of fluorescent (GFP-positive) cell is shown in panel **A**, and corresponding cell toxicity data is shown in panel **B**.

Figure 22 shows the results of a transduction efficiency experiment in which GFP-NLS cargo protein was intracellularly delivered using the shuttle His-CM18-PTD4 in HeLa cells. GFP-NLS transduction efficiency was evaluated by flow cytometry and the percentage of GFP fluorescent cells ("Pos cells (%)"), as well as corresponding cell viability data ("viability (%)") are shown. Panel **A** shows a comparison of GFP-NLS transduction efficiencies using different transduction protocols (Protocol A vs. B). Panel **B** shows the effect of using different concentrations of the shuttle His-CM18-PTD4 when using Protocol B.

Figures 23-26 are microscopy images showing the results of transduction experiments in which GFP-NLS (**Figures 23, 24A, 24B, 25 and 26**) or FITC-labeled anti-tubulin antibody (**Figure 24C and 24D**) cargo protein was intracellularly delivered with the shuttle His-CM18-PTD4 in HeLa cells. The bright field and fluorescence images of living cells are shown in **Figures 23, 24 and 26**. In **Figure 25**, the cells were fixed, permeabilized and subjected to immuno-labelling with an anti-GFP antibody and a fluorescent secondary antibody. White triangle windows indicate examples of areas of co-labelling between nuclei (DAPI) and GFP-NLS signals. **Figure 26** shows images captured by confocal microscopy.

Figure 27 shows microscopy images of a kinetic (time-course) transduction experiment in HeLa cells, where the fluorescence of GFP-NLS cargo protein was tracked after 45, 75, 100, and 120 seconds following intracellular delivery with the shuttle His-CM18-PTD4. The diffuse cytoplasmic fluorescence pattern observed after 45 seconds (panel **A**) gradually becomes a more concentrated nuclear pattern at 120 seconds (panel **D**).

Figure 28 shows microscopy images of co-delivery transduction experiment in which two cargo proteins (GFP-NLS and mCherryTM-NLS) are simultaneously delivered intracellularly by the shuttle His-CM18-PTD4 in HeLa cells. Cells and fluorescent signals were visualized by (**A**) bright field and (**B-D**) fluorescence microscopy. White triangle windows indicate examples of areas of co-labelling between nuclei (DAPI) and GFP-NLS or mCherryTM.

Figure 29 shows the results of GFP-NLS transduction efficiency experiments in HeLa cells using different shuttle agents or single-domain/control peptides. GFP-NLS transduction efficiency was evaluated by flow cytometry and the percentage of GFP fluorescent cells ("Pos cells (%)"), as well as corresponding cell viability data ("viability (%)") are shown in panels A, B, D-G and I. In panels A and D-F, cells were exposed to the cargo/shuttle agent for 10 seconds. In panel I, cells were exposed to the cargo/shuttle agent for 1 minute. In panels B, C, G and H, cells were exposed to the cargo/shuttle agent for 1, 2, or 5 min. "Relative fluorescence intensity (FL1-A)" or "Signal intensity" corresponds to the mean of all fluorescence intensities from each cell with a GFP fluorescent signal after GFP-NLS fluorescent protein delivery with the shuttle agent. Panel D shows the results of a control experiment in which only single-domain peptides (ELD or CDP) or the peptide His-PTD4 (His-CPD) were used for the GFP-NLS transduction, instead of the multi-domain shuttle agents.

Figure 30 shows microscopy images of HeLa cells transduced with GFP-NLS using the shuttle agent **(A)** TAT-KALA, **(B)** His-CM18-PTD4, **(C)** His-C(LLKK)₃C-PTD4, **(D)** PTD4-KALA, **(E)** EB1-PTD4, and **(F)** His-CM18-PTD4-His. The insets in the bottom row panels show the results of corresponding flow cytometry analyses, indicating the percentage of cells exhibiting GFP fluorescence.

Figure 31 shows the results of a transduction efficiency experiment in which GFP-NLS cargo protein was intracellularly delivered using the shuttle His-CM18-PTD4 in THP-1 cells using different Protocols (Protocol A vs C). GFP-NLS transduction efficiency was evaluated by flow cytometry and the percentage of GFP fluorescent cells ("Pos cells (%)"), as well as corresponding cell viability data ("viability (%)") are shown. "Ctrl" corresponds to THP-1 cells exposed to GFP-NLS cargo protein in the absence of a shuttle agent.

Figure 32 shows microscopy images of THP-1 cells transduced with GFP-NLS cargo protein using the shuttle His-CM18-PTD4. Images captured under at 4x, 10x and 40x magnifications are shown in panels **A-C**, respectively. White triangle windows in panel **C** indicate examples of areas of co-labelling between cells (bright field) and GFP-NLS fluorescence. Panel **D** shows the results of corresponding flow cytometry analyses, indicating the percentage of cells exhibiting GFP fluorescence.

Figure 33 shows microscopy images of THP-1 cells transduced with GFP-NLS cargo protein using the shuttle His-CM18-PTD4. White triangle windows indicate examples of areas of co-labelling between cells (bright field; panels **A** and **B**), and GFP-NLS fluorescence (panels **C** and **D**).

Figure 34 shows the results of GFP-NLS transduction efficiency experiments in THP-1 cells using the shuttle TAT-KALA, His-CM18-PTD4, or His-C(LLKK)₃C-PTD4. The cargo protein/shuttle agents were exposed to the THP-1 cells for 15, 30, 60 or 120 seconds. GFP-NLS transduction efficiency was evaluated by flow cytometry and the percentage of GFP fluorescent cells ("Pos cells (%)"), as well as corresponding cell viability data ("viability (%)") are shown in panel **A**. In panel **B**, "Relative fluorescence intensity (FL1-A)" corresponds to the mean of all fluorescence intensities from each cell with a GFP fluorescent signal after GFP-NLS fluorescent protein delivery with the shuttle agent.

Figure 35 shows the results of transduction efficiency experiments in which THP-1 cells were exposed daily to GFP-NLS cargo in the presence of a shuttle agent for 2.5 hours. His-CM18-PTD4 was used in panels **A-E**, and His-C(LLKK)₃C-PTD4 was used in panel **F**. GFP-NLS transduction efficiency was determined by flow cytometry at Day 1 or Day 3, and the results are expressed as the percentage of GFP fluorescent cells ("Pos cells (%)"), as well as corresponding cell viability data ("viability (%)") in panels **A-C** and **F**. Panel **D** shows the metabolic activity index of the THP-1 cells after 1, 2, 4, and 24h, and panel **E** shows the metabolic activity index of the THP-1 cells after 1 to 4 days, for cells exposed to the His-CM18-PTD4 shuttle.

Figure 36 shows a comparison of the GFP-NLS transduction efficiencies in a plurality of different types of cells (e.g., adherent and suspension, as well as cell lines and primary cells) using the shuttle His-CM18-PTD4, as measured by flow cytometry. The results are expressed as the percentage of GFP fluorescent cells ("Pos cells (%)"), as well as corresponding cell viability data ("viability (%)").

Figure 37 shows fluorescence microscopy images of different types of cells (**A-H**) transduced with GFP-NLS cargo using the shuttle His-CM18-PTD4. GFP fluorescence was visualized by fluorescence microscopy at a 10x magnification. The results of parallel flow cytometry experiments are also provided in the insets (viability and percentage of GFP-fluorescing cells).

Figure 38 shows fluorescence microscopy images of primary human myoblasts transduced with GFP-NLS using the shuttle His-CM18-PTD4. Cells were fixed and permeabilized prior to immuno-labelling GFP-NLS with an anti-GFP antibody and a fluorescent secondary antibody. Immuno-labelled GFP is shown in panel **A**, and this image is overlaid with nuclei (DAPI) labelling in panel **B**.

Figure 39 shows a schematic layout (**A**, **B** and **C**) and sample fluorescence images (**D** and **E**) of a transfection plasmid surrogate assay used to evaluate the activity of intracellularly delivered CRISPR/Cas9-NLS complex. (**A**) At Day 1, cells are transfected with an expression plasmid encoding the fluorescent proteins mCherryTM and GFP, with a STOP codon separating their two open reading frames. Transfection of the cells with the expression plasmid results in only mCherryTM expression (**D**). A CRISPR/Cas9-NLS complex, which has been designed/programmed to cleave the plasmid DNA at the STOP codon, is then delivered intracellularly to the transfected cells expressing mCherryTM, resulting double-stranded cleavage of the plasmid DNA at the STOP codon (**B**). In a fraction of the cells, random non-homologous DNA repair of the cleaved plasmid occurs and results in removal of the STOP codon (**C**), and thus GFP expression and fluorescence (**E**). White triangle windows indicate examples of areas of co-labelling of mCherryTM and GFP fluorescence.

Figure 40 shows fluorescence microscopy images of HeLa cells expressing mCherryTM and GFP, indicating CRISPR/Cas9-NLS-mediated cleavage of plasmid surrogate DNA. In panels **A-D**, HeLa cells were co-transfected with three plasmids: the plasmid surrogate as described in the brief description of **Figure 39**, and two other expression plasmids encoding the Cas9-NLS protein and crRNA/tracrRNAs, respectively. CRISPR/Cas9-mediated cleavage of the plasmid surrogate at the STOP codon, and subsequent DNA repair by the cell, enables expression of

GFP (panels **B** and **D**) in addition to mCherry™ (panels **A** and **C**). In panels **E-H**, HeLa cells were transfected with the plasmid surrogate and then transduced with an active CRISPR/Cas9-NLS complex using the shuttle His-CM18-PTD4. CRISPR/Cas9-NLS-mediated cleavage of the plasmid surrogate at the STOP codon, and subsequent DNA repair by the cell, enables expression of GFP (panels **F** and **H**) in addition to mCherry™ (panels **E** and **G**).

5 **Figure 41A** shows the products of a DNA cleavage assay (T7E1 assay) separated by agarose gel electrophoresis, which is used to measure CRISPR/Cas9-mediated cleavage of cellular genomic DNA. HeLa cells were transduced with a CRISPR-Cas9-NLS complex programmed to cleave the PPIB gene. The presence of the cleavage product framed in white boxes 1 and 2, indicates cleavage of the PPIB gene by the CRISPR-Cas9-NLS complex, which was delivered intracellularly using the shuttle His-C(LLKK)₃C-PTD4 (lane **B**) or with a lipidic transfection agent used as a positive control (lane **D**). This cleavage product is absent in negative controls (lanes **A** and **C**).

Figure 41B shows the products of a DNA cleavage assay (T7E1 assay) separated by agarose gel electrophoresis, which is used to measure CRISPR/Cas9-mediated cleavage of cellular genomic DNA (PPIB DNA sequences). The left panel shows the cleavage product of the amplified PPIB DNA sequence by the CRISPR/Cas9 complex after the delivery of the complex with the shuttle agent His-CM18-PTD4 in HeLa cells. The right panel shows amplified DNA sequence before the T7E1 digestion procedure as a negative control.

Figure 41C shows the products of a DNA cleavage assay (T7E1 assay) separated by agarose gel electrophoresis, which is used to measure CRISPR/Cas9-mediated cleavage of cellular genomic DNA (PPIB DNA sequences). The left panel shows the amplified PPIB DNA sequence after incubation of the HeLa cells with the Cas9/RNAs complex in presence of a lipidic transfection agent (DharmaFect™ transfection reagent # T-20XX-01) (positive control). The right panel shows amplified DNA sequence before the T7E1 digestion procedure as a negative control. **Figures 42-44** show the transcriptional activity of THP-1 cells that have been transduced with the transcription factor HOXB4 using different concentrations of the shuttle His-CM18-PTD4 and different cargo/shuttle exposure times. Successful intra-nuclear delivery of HOXB4 was determined by monitoring mRNA levels of a target gene by real-time PCR, and the results are normalized against those in the negative control (HOXB4 without shuttle agent) and expressed as "Fold over control" (left bars). Total cellular RNA (ng/μL) was quantified and used a marker for cell viability (right bars). "Ø" or "Ctrl" means "no treatment"; "TF" means "Transcription Factor alone"; "FS" means "shuttle alone".

Figure 45 shows fluorescence microscopy images of HeLa cells transduced with wild-type HOXB4 cargo using the shuttle His-CM18-PTD4. After a 30-minute incubation to allow transduced HOXB4-WT to accumulate in the nucleus, the cells were fixed, permeabilized and HOXB4-WT was labelled using a primary anti-HOXB4 monoclonal antibody and a fluorescent secondary antibody (panels **B** and **D**). Nuclei were labelled with DAPI (panels **A** and **C**). White triangle windows indicate examples of areas of co-labelling between nuclei and HOXB4 – compare **A** vs **B** (x20 magnification), and **C** vs **D** (x40 magnification).

Figure 46 shows the products of a DNA cleavage assay separated by agarose gel electrophoresis, which is used to measure CRISPR/Cas9-mediated cleavage of cellular genomic DNA (HPTR sequence) after intracellular delivery of the complex with different shuttle agents. Panel **A** shows the results with the shuttle agents: His-CM18-PTD4, His-CM18-PTD4-His, and His-C(LLKK)3C-PTD4 in HeLa cells. Panel **B** shows the results with His-CM18-PTD4-His and His-CM18-L2-PTD4 in Jurkat cells. Negative controls (lane 4 in panels A and B) show amplified HPTR DNA sequence after incubation of the cells with the CRISPR/Cas9 complex without the presence of the shuttle agent. Positive controls (lane 5 in panels A and B) show the amplified HPTR DNA sequence after incubation of the cells with the Cas9/RNAs complex in presence of a commercial lipidic transfection agent.

Figure 47 shows the transcriptional activity of THP-1 cells that have been transduced with the transcription factor HOXB4 using the shuttle agents His-CM18-PTD4, TAT-KALA, EB1-PTD4, His-C(LLKK)3C-PTD4 and His-CM18-PTD4-His. Successful intra-nuclear delivery of HOXB4 was determined by monitoring mRNA levels of a target gene by real-time PCR, and the results were normalized against those in the negative control (HOXB4 without shuttle agent) and expressed as "Fold over control" (left bars). Total cellular RNA (ng/ μ L) was quantified and used a marker for cell viability (right bars). "Ø" or "Ctrl" means "no treatment"; "TF" means "Transcription Factor alone"; "FS" means "shuttle alone".

Figure 48 shows *in vivo* GFP-NLS delivery in rat parietal cortex by His-CM18-PTD4. Briefly, GFP-NLS (20 μ M) was injected in the parietal cortex of rat in presence of the shuttle agent His-CM18-PTD4 (20 μ M) for 10 min. Dorso-ventral rat brain slices were collected and analysed by fluorescence microscopy at **(A)** 4x, **(C)** 10x and **(D)** 20x magnifications. The injection site is located in the deepest layers of the parietal cortex (PCx). In presence of the His-CM18-PTD4 shuttle agent, the GFP-NLS diffused in cell nuclei of the PCx, of the Corpus Callus (Cc) and of the striatum (Str) (white curves mark limitations between brains structures). Panel **B** shows the stereotaxic coordinates of the injection site (black arrows) from the rat brain atlas of Franklin and Paxinos. The injection of GFP-NLS in presence of His-CM18-PTD4 was performed on the left part of the brain, and the negative control (injection of GFP-NLS alone), was done on the contralateral site. The black circle and connected black lines in panel **B** show the areas observed in the fluorescent pictures (A, C and D).

SEQUENCE LISTING

This application contains a Sequence Listing in computer readable form entitled Sequence_Listing.txt, created April 3, 2016 having a size of about 57 kb. The computer readable form is incorporated herein by reference.

SEQ ID NO:	Description	SEQ ID NO:	Description
1	CM18	53	Mitochondrial signal sequence from 18S rRNA
2	Diphtheria toxin T domain (DT)	54	Peroxisome signal sequence – PTS1
3	GALA	55	Nucleolar signal sequence from BIRC5
4	PEA	56	Nucleolar signal sequence from RECQL4
5	INF-7	57	CM18-TAT
6	LAH4	58	CM18-Penetratin

7	HGP	59	His-CM18-TAT
8	H5WYG	60	GFP
9	HA2	61	TAT-GFP
10	EB1	62	GFP-NLS
11	VSVG	63	C(LLKK) ₃ C
12	Pseudomonas toxin	64	G(LLKK) ₃ G
13	Melittin	65	PTD4
14	KALA	66	TAT-CM18
15	JST-1	67	TAT-KALA
16	SP	68	His-CM18-PTD4
17	TAT	69	His-CM18-9Arg
18	Penetratin (Antennapedia)	70	His-CM18-Transportan
19	pVEC	71	His-LAH4-PTD4
20	M918	72	His-C(LLKK) ₃ C-PTD4
21	Pep-1	73	mCherry TM -NLS
22	Pep-2	74	Cas9-NLS
23	Xentry	75	crRNA (Example 13.3)
24	Arginine stretch	76	tracrRNA (Example 13.3)
25	Transportan	77	Feldan tracrRNA (Example 13.5, 13.6)
26	SynB1	78	PPIB crRNA (Example 13.5)
27	SynB3	79	Dharmacon tracrRNA (Example 13.5)
28	E1a	80	HOXB4-WT
29	SV40 T-Ag	81	His-PTD4
30	c-myc	82	PTD4-KALA
31	Op-T-NLS	83	9Arg-KALA
32	Vp3	84	Pep1-KALA
33	Nucleoplasmin	85	Xentry-KALA
34	Histone 2B NLS	86	SynB3-KALA
35	Xenopus N1	87	VSVG-PTD4
36	PARP	88	EB1-PTD4
37	PDX-1	89	JST-PTD4
38	QKI-5	90	CM18-PTD4
39	HCDA	91	6Cys-CM18-PTD4
40	H2B	92	CM18-L1-PTD4
41	v-Rel	93	CM18-L2-PTD4
42	Amida	94	CM18-L3-PTD4
43	RanBP3	95	His-CM18-TAT
44	Pho4p	96	His-CM18-PTD4-6Cys
45	LEF-1	97	3His-CM18-PTD4
46	TCF-1	98	12His-CM18-PTD4
47	BDV-P	99	HA-CM18-PTD4
48	TR2	100	3HA-CM18-PTD4
49	SOX9	101	CM18-His-PTD4
50	Max	102	His-CM18-PTD4-His
51	Mitochondrial signal sequence from Tim9	103	HPRT crRNA (Example 13.6)
52	Mitochondrial signal sequence from Yeast cytochrome c oxidase subunit IV		

DETAILED DESCRIPTION

The present description stems from the surprising discovery that multi-domain synthetic peptides comprising an endosome leakage domain (ELD) operably linked to a cell penetrating domain (CPD) can significantly increase the transduction efficiency of an independent polypeptide cargo to the cytosol of eukaryotic target cells. In contrast, this increase in transduction efficiency was not found using independent single-domain peptides containing only an ELD, or only a CPD used alone or together (i.e., in a mixture of separate single-domain peptides). Accordingly, in some aspects the present description relates to a polypeptide-based shuttle agent comprising an endosome leakage domain (ELD) operably linked to a cell penetrating domain (CPD), or an ELD operably linked to a histidine-rich domain and a CPD, for use in increasing the transduction efficiency of an independent polypeptide cargo to the cytosol of a target eukaryotic cell.

Synthetic peptides and polypeptide-based shuttle agents

As used herein, the term "**synthetic**" used in expressions such as "synthetic peptide" or "synthetic polypeptide" is intended to refer to non-naturally occurring molecules that can be produced *in vitro* (e.g., synthesized chemically and/or produced using recombinant DNA technology). The purities of various synthetic preparations may be assessed by for example high-performance liquid chromatography analysis and mass spectroscopy. Chemical synthesis approaches may be advantageous over cellular expression systems (e.g., yeast or bacteria protein expression systems), as they may preclude the need for extensive recombinant protein purification steps (e.g., required for clinical use). In contrast, longer synthetic polypeptides may be more complicated and/or costly to produce via chemical synthesis approaches and such polypeptides may be more advantageously produced using cellular expression systems. In some embodiments, the peptides or shuttle agent of the present description may be chemically synthesized (e.g., solid- or liquid phase peptide synthesis), as opposed to expressed from a recombinant host cell. In some embodiments, the peptides or shuttle agent of the present description may lack an N-terminal methionine residue. A person of skill in the art may adapt a synthetic peptide or shuttle agent of the present description by using one or more modified amino acids (e.g., non-naturally-occurring amino acids), or by chemically modifying the synthetic peptide or shuttle agent of the present description, to suit particular needs of stability or other needs.

The expression "**polypeptide-based**" when used here in the context of a shuttle agent of the present description, is intended to distinguish the presently described shuttle agents from non-polypeptide or non-protein-based shuttle agents such as lipid- or cationic polymer-based transduction agents, which are often associated with increased cellular toxicity and may not be suitable for use in human therapy.

As used herein, the expression "**increasing transduction efficiency**" refers to the ability of a shuttle agent (e.g., a polypeptide-based shuttle agent of the present description) to improve the percentage or proportion of a population of target cells into which a cargo of interest (e.g., a polypeptide cargo) is delivered intracellularly across

the plasma membrane. Immunofluorescence microscopy, flow cytometry, and other suitable methods may be used to assess cargo transduction efficiency. In some embodiments, a shuttle agent of the present description may enable a transduction efficiency of at least 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, or 85%, for example as measure by immunofluorescence microscopy, flow cytometry, FACS, and other suitable methods. In some embodiments, a shuttle agent of the present description may enable one of the aforementioned transduction efficiencies together with a cell viability of at least 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, or 95%, for example as measure by the assay described in **Example 3.3a**, or by another suitable assay known in the art.

As used herein, the term **"independent"** is generally intended refer to molecules or agents which are not covalently bound to one another. For example, the expression **"independent polypeptide cargo"** is intended to refer to a polypeptide cargo to be delivered intracellularly that is not covalently bound (e.g., not fused) to a shuttle agent of the present description. In some aspects, having shuttle agents that are independent of (not fused to) a polypeptide cargo may be advantageous by providing increased shuttle agent versatility -- e.g., not being required to re-engineer a new fusion protein for different polypeptide cargoes, and/or being able to readily vary the ratio of shuttle agent to cargo (as opposed to being limited to a 1:1 ratio in the case of a fusion protein).

In addition to increasing target cell transduction efficiency, shuttle agents of the present description may facilitate the delivery of a cargo of interest (e.g., a polypeptide cargo) to the cytosol of target cells. In this regard, efficiently delivering an extracellular cargo to the cytosol of a target cell using approaches based on cell penetrating peptides can be challenging, as the cargo often becomes trapped in intracellular endosomes after crossing the plasma membrane, which may limit its intracellular availability and may result in its eventual metabolic degradation. For example, use of the protein transduction domain from the HIV-1 Tat protein has been reported to result in massive sequestration of the cargo into intracellular vesicles. In some aspects, shuttle agents of the present description may facilitate the ability of endosomally-trapped cargo to escape from the endosome and gain access to the cytoplasmic compartment. In this regard, the expression **"to the cytosol"** in the phrase "increasing the transduction efficiency of an independent polypeptide cargo to the cytosol," is intended to refer to the ability of shuttle agents of the present description to allow an intracellularly delivered cargo of interest to escape endosomal entrapment and gain access to the cytoplasmic compartment. After a cargo of interest has gained access to the cytosol, it may be subsequently targeted to various subcellular compartments (e.g., nucleus, nucleolus, mitochondria, peroxisome). In some embodiments, the expression "to the cytosol" is thus intended to encompass not only cytosolic delivery, but also delivery to other subcellular compartments that first require the cargo to gain access to the cytoplasmic compartment.

As used herein, a **"domain"** or "protein domain" generally refers to a part of a protein having a particular functionality or function. Some domains conserve their function when separated from the rest of the protein, and thus can be used in a modular fashion. By combining such domains from different proteins of viral, bacterial, or eukaryotic

origin, it becomes possible in accordance with the present description to not only design multi-domain polypeptide-based shuttle agents that are able to deliver a cargo intracellularly, but also enable the cargo to escape endosomes and reach the cytoplasmic compartment.

5 The modular characteristic of many protein domains can provide flexibility in terms of their placement within the shuttle agents of the present description. However, some domains may perform better when engineered at certain positions of the shuttle agent (e.g., at the N- or C-terminal region, or therebetween). The position of the domain within its endogenous protein is sometimes an indicator of where the domain should be engineered within the shuttle agent, and of what type/length of linker should be used. Standard recombinant DNA techniques can be used by the skilled person to manipulate the placement and/or number of the domains within the shuttle agents of the present description in view of the present disclosure. Furthermore, assays disclosed herein, as well as others known in the art, can be used to assess the functionality of each of the domains within the context of the shuttle agents (e.g., their ability to facilitate cell penetration across the plasma membrane, endosome escape, and/or access to the cytosol). Standard methods can also be used to assess whether the domains of the shuttle agent affect the activity of the cargo to be delivered intracellularly. In this regard, the expression "**operably linked**" as used herein refers to the ability of the domains to carry out their intended function(s) (e.g., cell penetration, endosome escape, and/or subcellular targeting) within the context of the shuttle agents of the present description. For greater clarity, the expression "operably linked" is meant to define a functional connection between two or more domains without being limited to a particular order or distance between same.

20 In some embodiments, synthetic peptide or polypeptide-based shuttle agent of the present description may comprise a minimum length of 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30 amino acid residues and a maximum length of 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, or 150 amino acid residues. In some embodiments, shorter synthetic peptide or polypeptide-based shuttle agents are particularly advantageous because they may be more easily synthesized and purified by chemical synthesis approaches, which may be more suitable for clinical use (as opposed to recombinant proteins that must be purified from cellular expression systems). While numbers and ranges in the present description are often listed as multiples of 5, the present description should not be so limited. For example, the maximum length described herein should be understood as also encompassing a length of 36, 37, 38...51, 62, etc., in the present description, and that their non-listing herein is only for the sake of brevity. The same reasoning applies to the % of identities listed herein (e.g., 86%, 87%...93%...), the percentages of histidine residues, etc.

30 In some embodiments, synthetic peptide or polypeptide-based shuttle agent of the present description may comprise a predicted net charge of at least +5, +6, +7, at least +8, at least +9, at least +10, at least +11, at least +12, at least +13, at least +14, or at least +15 at physiological pH. These positive charges are generally conferred by the greater presence of positively-charged lysine and/or arginine residues, as opposed to negatively charged aspartate and/or glutamate residues.

In some embodiments, synthetic peptide or polypeptide-based shuttle agent of the present description may be soluble in aqueous solution (e.g., at physiological pH), which facilitates their use in for example cell culture media to delivery cargoes intracellularly to live cells.

5 In some embodiments, synthetic peptide or polypeptide-based shuttle agent of the present description may comprise or consist of the amino acid sequence of any one of **SEQ ID NOs: 57-59, 66-72, or 82-102**, or a functional variant thereof having at least 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81% 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, or 95% identity to any one of **SEQ ID NOs: 57-59, 66-72, or 82-102**.

10 In some embodiments, synthetic peptide or polypeptide-based shuttle agents of the present description may comprise oligomers (e.g., dimers, trimers, etc.) of a synthetic peptide or polypeptide-based shuttle agent as defined herein. Such oligomers may be constructed by covalently binding the same or different types of shuttle agent monomers (e.g., using disulfide bridges to link cysteine residues introduced into the monomer sequences).

In some embodiments, the synthetic peptide or polypeptide-based shuttle agent of the present description may comprise an N-terminal and/or a C-terminal cysteine residue.

15

Endosome leakage domains (ELDs)

In some aspects, synthetic peptide or polypeptide-based shuttle agents of the present description may comprise an endosome leakage domain (ELD) for facilitating endosome escape and access to the cytoplasmic compartment. As used herein, the expression "**endosome leakage domain**" refers to a sequence of amino acids
20 which confers the ability of endosomally-trapped macromolecules to gain access to the cytoplasmic compartment. Without being bound by theory, endosome leakage domains are short sequences (often derived from viral or bacterial peptides), which are believed to induce destabilization of the endosomal membrane and liberation of the endosome contents into the cytoplasm. As used herein, the expression "**endosomolytic peptide**" is intended to refer to this general class of peptides having endosomal membrane-destabilizing properties. Accordingly, in some
25 embodiments, synthetic peptide or polypeptide-based shuttle agents of the present description may comprise an ELD which is an endosomolytic peptide. The activity of such peptides may be assessed for example using the calcein endosome escape assays described in **Example 2**.

In some embodiments, the ELD may be a peptide that disrupts membranes at acidic pH, such as pH-dependent membrane active peptide (PMAP) or a pH-dependent lytic peptide. For example, the peptides GALA and
30 INF-7 are amphiphilic peptides that form alpha helixes when a drop in pH modifies the charge of the amino acids which they contain. More particularly, without being bound by theory, it is suggested that ELDs such as GALA induce endosomal leakage by forming pores and flip-flop of membrane lipids following conformational change due to a decrease in pH (Kakudo, Chaki et al., 2004, Li, Nicol et al., 2004). In contrast, it is suggested that ELDs such as INF-7 induce endosomal leakage by accumulating in and destabilizing the endosomal membrane (El-Sayed, Futaki et al.,

2009). Accordingly in the course of endosome maturation, the concomitant decline in pH causes a change in the conformation of the peptide and this destabilizes the endosome membrane leading to the liberation of the endosome contents. The same principle is thought to apply to the toxin A of *Pseudomonas* (Varkouhi, Scholte et al., 2011). Following a decline in pH, the conformation of the domain of translocation of the toxin changes, allowing its insertion
5 into the endosome membrane where it forms pores (London 1992, O'Keefe 1992). This eventually favors endosome destabilization and translocation of the complex outside of the endosome. The above described ELDs are encompassed within the ELDs of the present description, as well as other mechanisms of endosome leakage whose mechanisms of action may be less well defined.

In some embodiments, the ELD may be an antimicrobial peptide (AMP) such as a linear cationic alpha-helical antimicrobial peptide (AMP). These peptides play a key role in the innate immune response due to their ability
10 to strongly interact with bacterial membranes. Without being bound by theory, these peptides are thought to assume a disordered state in aqueous solution, but adopt an alpha-helical secondary structure in hydrophobic environments. The latter conformation thought to contribute to their typical concentration-dependent membrane-disrupting properties. When accumulated in endosomes at a certain concentrations, some antimicrobial peptides may induce
15 endosomal leakage.

In some embodiments, the ELD may be an antimicrobial peptide (AMP) such as Cecropin-A/Melittin hybrid (CM series) peptide. Such peptides are thought to be among the smallest and most effective AMP-derived peptides with membrane-disrupting ability. Cecropins are a family of antimicrobial peptides with membrane-perturbing abilities against both Gram-positive and Gram-negative bacteria. Cecropin A (CA), the first identified antibacterial peptide, is
20 composed of 37 amino acids with a linear structure. Melittin (M), a peptide of 26 amino acids, is a cell membrane lytic factor found in bee venom. Cecropin-melittin hybrid peptides have been shown to produce short efficient antibiotic peptides without cytotoxicity for eukaryotic cells (i.e., non-hemolytic), a desirable property in any antibacterial agent. These chimeric peptides were constructed from various combinations of the hydrophilic N-terminal domain of Cecropin A with the hydrophobic N-terminal domain of Melittin, and have been tested on bacterial model systems.
25 Two 26-mers, CA(1-13)M(1-13) and CA(1-8) M(1-18) (Boman et al., 1989), have been shown to demonstrate a wider spectrum and improved potency of natural Cecropin A without the cytotoxic effects of melittin.

In an effort to produce shorter CM series peptides, the authors of Andreu et al., 1992 constructed hybrid peptides such as the 26-mer (CA(1-8)M(1-18)), and compared them with a 20-mer (CA(1-8)M(1-12)), a 18-mer (CA(1-8)M(1-10)) and six 15-mers ((CA(1-7)M(1-8), CA(1-7)M(2-9), CA(1-7)M(3-10), CA(1-7)M(4-11), CA(1-7)M(5-12), and CA(1-7)M(6-13)). The 20 and 18-mers maintained similar activity comparatively to CA(1-8)M(1-18). Among
30 the six 15-mers, CA(1-7)M(1-8) showed low antibacterial activity, but the other five showed similar antibiotic potency compared to the 26-mer without hemolytic effect. Accordingly, in some embodiments, synthetic peptide or polypeptide-based shuttle agents of the present description may comprise an ELD which is or is from CM series peptide variants, such as those described above.

In some embodiments, the ELD may be the CM series peptide CM18 composed of residues 1–7 of Cecropin-A (KWKLFFKIGAVLKVLTTG) fused to residues 2–12 of Melittin (YGRKKRRQRRR), [C(1–7)M(2–12)]. When fused to the cell penetrating peptide TAT, CM18 was shown to independently cross the plasma membrane and destabilize the endosomal membrane, allowing some endosomally-trapped cargos to be released to the cytosol (Salomone et al., 2012). However, the use of a CM18-TAT11 peptide fused to a fluorophore (atto-633) in some of the author's experiments, raises uncertainty as to the contribution of the peptide versus the fluorophore, as the use of fluorophores themselves have been shown to contribute to endosomolysis -- e.g., via photochemical disruption of the endosomal membrane (Erazo-Oliveras et al., 2013).

In some embodiments, the ELD may be CM18 having the amino acid sequence of **SEQ ID NO: 1**, or a variant thereof having at least 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, or 95% identity to **SEQ ID NO: 1** and having endosomolytic activity.

In some embodiments, the ELD may be a peptide derived from the N terminus of the HA2 subunit of influenza hemagglutinin (HA), which may also cause endosomal membrane destabilization when accumulated in the endosome.

In some embodiments, synthetic peptide or polypeptide-based shuttle agents of the present description may comprise an ELD which is or is from an ELD set forth in **Table A**, or a variant thereof having endosome escape activity and/or pH-dependent membrane disrupting activity.

Table A: Examples of endosome leakage domains

Name	Amino acid sequence	SEQ ID NO:	Reference(s)
CM18	KWKLFFKIGAVLKVLTTG	1	(Salomone, Cardarelli et al., 2012)
Diphtheria toxin T domain (DT)	VGSSLSCINLDWDVIRDKTKTKIESLKEHGIK NKMSESPNKTVSEKAKQYLEEFHQTALEHP ELSELKTVTGTNPVFAGANYAAWAVNVAQVID SETADNLEKTTAALSILPGIGSVMGIADGAVHH NTEEIVAQSIALLSLMVAQAIPLVGELVDIGFAA YNFVESIINLFQVVHNSYNRPAYSPG	2	(Uherek, Fominaya et al., 1998, Glover, Ng et al., 2009)
GALA	WEAALAEALAEALAEHLAEALAEALAA	3	(Parente, Nir et al., 1990) (Li, Nicol et al., 2004)
PEA	VLGNPAKHDLIDKPTVISHRLHFEGGSLAAL TAHQACHLPLETFTRHRQPRGWEQLEQCGYP VQRLVALYLAARLSWNQVDQVIRNALASPGS GGDLGEAIREQPEQARLALT	4	(Fominaya and Wels 1996)
INF-7	GLFEAIEGFIENGWEGMIDGWYGC	5	(El-Sayed, Futaki et al., 2009)
LAH4	KKALLALALHHLAHLALHLALALKKA	6	(Kichler, Mason et al., 2006) Kichler et al., 2003
HGP	LLGRRGWEVLKYWWNLLQYWSQEL	7	(Zhang, Cui et al., 2006)

H5WYG	GLFHAIAHFIHGGWHGLIHGWYG	8	(Midoux, Kichler et al., 1998)
HA2	GLFGAAGFIENGWEGMIDGWYG	9	(Lorieau, Louis et al., 2010)
EB1	LIRLWSHLIHIWFQNRRLKWKKK	10	(Amand, Norden et al., 2012)
VSVG	KFTIVFPHNQKGNWKNVPSNYHYCP	11	(Schuster, Wu et al., 1999)
<i>Pseudomonas</i> toxin	EGGSLAALTAHQACHLPLETFTRHRQPRGWE QLEQCGYPVQRLVALYLAARLSWNQVDQVIR NALASPGSGGDLGEAIREQPEQARLALTAAA ESERFVRQGTGNDEAGANAD	12	(Fominaya, Uherek et al., 1998)
Melittin	GIGAVLKVLTTGLPALISWIKRKRQQ	13	(Tan, Chen et al., 2012)
KALA	WEAKLAKALAKALAKHLAKALAKALACEA	14	(Wyman, Nicol et al., 1997)
JST-1	GLFEALLELLESLWELLLEA	15	(Gottschalk, Sparrow et al., 1996)
C(LLKK) ₃ C	CLLKLLKKLLKKC	63	(Luan et al., 2014)
G(LLKK) ₃ G	GLLKLLKKLLKKG	64	(Luan et al., 2014)

In some embodiments, shuttle agents of the present description may comprise one or more ELD or type of ELD. More particularly, they can comprise at least 2, at least 3, at least 4, at least 5, or more ELDs. In some embodiments, the shuttle agents can comprise between 1 and 10 ELDs, between 1 and 9 ELDs, between 1 and 8 ELDs, between 1 and 7 ELDs, between 1 and 6 ELDs, between 1 and 5 ELDs, between 1 and 4 ELDs, between 1 and 3 ELDs, etc.

In some embodiments, the order or placement of the ELD relative to the other domains (CPD, histidine-rich domains) within the shuttle agents of the present description may be varied provided the shuttling ability of the shuttle agent is retained.

In some embodiments, the ELD may be a variant or fragment of any one those listed in **Table A**, and having endosomolytic activity. In some embodiments, the ELD may comprise or consist of the amino acid sequence of any one of **SEQ ID NOs: 1-15, 63, or 64**, or a sequence which is at least 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, or 95% identical to any one of **SEQ ID NOs: 1-15, 63, or 64**, and having endosomolytic activity.

Cell penetration domains (CPDs)

In some aspects, the shuttle agents of the present description may comprise a cell penetration domain (CPD). As used herein, the expression "**cell penetration domain**" refers to a sequence of amino acids which confers the ability of a macromolecule (e.g., peptide or protein) containing the CPD to be transduced into a cell.

In some embodiments, the CPD may be (or may be from) a cell-penetrating peptide or the protein transduction domain of a cell-penetrating peptide. Cell-penetrating peptides can serve as carriers to successfully

deliver a variety of cargos intracellularly (e.g., polynucleotides, polypeptides, small molecule compounds or other macromolecules/compounds that are otherwise membrane-impermeable). Cell-penetrating peptides often include short peptides rich in basic amino acids that, once fused (or otherwise operably linked) to a macromolecule, mediate its internalization inside cells (Shaw, Catchpole et al., 2008). The first cell-penetrating peptide was identified by analyzing the cell penetration ability of the HIV-1 trans-activator of transcription (Tat) protein (Green and Loewenstein 1988, Vives, Brodin et al., 1997). This protein contains a short hydrophilic amino acid sequence, named "TAT", which promotes its insertion within the plasma membrane and the formation of pores. Since this discovery, many other cell-penetrating peptides have been described. In this regard, in some embodiments, the CPD can be a cell-penetrating peptide as listed in **Table B**, or a variant thereof having cell-penetrating activity.

Table B: Examples of cell-penetrating peptides

Name	Amino acid sequence	SEQ ID NO:	Reference(s)
SP	AAVALLPAVLLALLAP	16	(Mahlum, Mandal et al., 2007)
TAT	YGRKKRRQRRR	17	(Green and Loewenstein 1988, Fawell, Seery et al., 1994, Vives, Brodin et al., 1997)
Penetratin (Antennapedia)	RQIKIWFQNRRMKWKK	18	(Perez, Joliot et al., 1992)
pVEC	LLILRRRIRKQAHASK	19	(Elmqvist, Lindgren et al., 2001)
M918	MVTVLFRRLRIRACGPPRVRV	20	(El-Andaloussi, Johansson et al., 2007)
Pep-1	KETWWETWWTEWSQPKKKRKV	21	(Morris, Depollier et al., 2001)
Pep-2	KETWFETWFTEWSQPKKKRKV	22	(Morris, Chaloin et al., 2004)
Xentry	LCLRPVG	23	(Montrose, Yang et al., 2013)
Arginine stretch	RRRRRRRRR	24	(Zhou, Wu et al., 2009)
Transportan	WTLNSAGYLLGKINLKALAALAKKIL	25	(Hallbrink, Floren et al., 2001)
SynB1	RGGRLSYSRRRFSTSTGR	26	(Drin, Cottin et al., 2003)
SynB3	RRLSYSRRRF	27	(Drin, Cottin et al., 2003)
PTD4	YARAAARQARA	65	(Ho et al, 2001)

Without being bound by theory, cell-penetrating peptides are thought to interact with the cell plasma membrane before crossing by pinocytosis or endocytosis. In the case of the TAT peptide, its hydrophilic nature and charge are thought to promote its insertion within the plasma membrane and the formation of a pore (Hercé and Garcia 2007). Alpha helix motifs within hydrophobic peptides (such as SP) are also thought to form pores within plasma membranes (Veatch, Liu et al., 2004).

In some embodiments, shuttle agents of the present description may comprise one or more CPD or type of CPD. More particularly, they may comprise at least 2, at least 3, at least 4, or at least 5 or more CPDs. In some embodiments, the shuttle agents can comprise between 1 and 10 CPDs, between 1 and 6 CPDs, between 1 and 5 CPDs, between 1 and 4 CPDs, between 1 and 3 CPDs, etc.

In some embodiments, the CPD may be TAT having the amino acid sequence of **SEQ ID NO: 17**, or a variant thereof having at least 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%,

85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, or 95% identity to **SEQ ID NO: 17** and having cell penetrating activity; or Penetratin having the amino acid sequence of **SEQ ID NO: 18**, or a variant thereof having at least 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, or 95% identity to **SEQ ID NO: 18** and having cell
5 penetrating activity.

In some embodiments, the CPD may be PTD4 having the amino acid sequence of **SEQ ID NO: 65**, or a variant thereof having at least 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, or 95% identity to **SEQ ID NO: 65**.

In some embodiments, the order or placement of the CPD relative to the other domains (ELD, histidine-rich
10 domains) within the shuttle agents of the present description may be varied provided the shuttling ability of the shuttle agent is retained.

In some embodiments, the CPD may be a variant or fragment of any one those listed in **Table B**, and having cell penetrating activity. In some embodiments, the CPD may comprise or consist of the amino acid sequence of any one of **SEQ ID NOs: 16-27 or 65**, or a sequence which is at least 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%,
15 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 85%, 90%, 91%, 92%, 93%, 94%, or 95% identical to any one of **SEQ ID NOs: 16-27 or 65**, and having cell penetrating activity.

Histidine-rich domains

In some aspects, the shuttle agents of the present description may comprise a histidine-rich domain. In
20 other embodiments, the shuttle agents of the present description may be combined/used together with a further independent synthetic peptide comprising or consisting essentially of a histidine-rich domain and a CPD (e.g., but lacking an ELD). This latter approach may provide the added advantage of allowing the concentration of the histidine-rich domain to be varied or controlled independently from the concentration of the ELD and the CPD contained in the shuttle agent. Without being bound by theory, the histidine-rich domain may act as a proton sponge in the endosome,
25 providing another mechanism of endosomal membrane destabilization.

In some embodiments, the histidine-rich domain may be a stretch of at least 2, at least 3, at least 4, at least 5, or at least 6 amino acids comprising at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, or at least 90% histidine residues. In some embodiments, the histidine-rich domain may comprise at least 2, at least 3, at least 4 at least 5, at
30 least 6, at least 7, at least 8, or at least 9 consecutive histidine residues. Without being bound by theory, the histidine-rich domain in the shuttle agent may act as a proton sponge in the endosome through protonation of their imidazole groups under acidic conditions of the endosomes, providing another mechanism of endosomal membrane destabilization and thus further facilitating the ability of endosomally-trapped cargos to gain access to the cytosol. In some embodiments, the histidine-rich domain may be located at the N or C terminus of the synthetic peptide or

shuttle agent. In some embodiments, the histidine-rich domain may be located N-terminal or C terminal to the CPD and/or ELD.

In some embodiments, the order or placement of the histidine-rich domain relative to the other domains (CPD, ELD) within the shuttle agents of the present description may be varied provided the shuttling ability of the shuttle agent is retained. In some embodiments, the shuttle agents of the present description may comprise more than one histidine-rich domain (e.g., histidine-rich domains at the amino and carboxyl termini).

Linkers

In some embodiments, **suitable linkers** (e.g., flexible polypeptide linkers) can be used to operably connect the domains (CPDs, ELDs, or histidine-rich domains) to one another within the context of synthetic peptides and shuttle agents of the present description. In some embodiments, linkers may be formed by adding sequences of small hydrophobic amino acids without rotatory potential (such as glycine) and polar serine residues that confer stability and flexibility. Linkers may be soft and allow the domains of the shuttle agents to move. In some embodiments, prolines may be avoided since they can add significant conformational rigidity. In some embodiments, the linkers may be serine/glycine-rich linkers (e.g., GGS, GGS GGGS, GGS GGGS GGGS, or the like). In some embodiments, the use shuttle agents comprising a suitable linker may be advantageous for delivering an independent polypeptide cargo to suspension cells, rather than to adherent cells.

Cargos

In some aspects, the synthetic peptide or polypeptide-based shuttle agent of the present description may be useful for delivering an independent cargo (e.g., a polypeptide cargo) to the cytosol of a target eukaryotic cell. In some embodiments, the polypeptide cargo may be fused to one or more CPDs to further facilitate intracellular delivery. In some embodiments, the CPD fused to the polypeptide cargo may be the same or different from the CPD of the shuttle agent of the present description. Such fusion proteins may be constructed using standard recombinant technology. In some embodiments, the independent polypeptide cargo may be fused, complexed with, or covalently bound to a second biologically active cargo (e.g., a biologically active polypeptide or compound). Alternatively or simultaneously, the polypeptide cargo may comprise a subcellular targeting domain.

In some embodiments, the polypeptide cargo must be delivered to the nucleus for it to carry out its intended biological effect. One such example is when the cargo is a polypeptide intended for nuclear delivery (e.g., a transcription factor). In this regard, studies on the mechanisms of translocation of viral DNA have led to the identification of nuclear localization signals (NLSs). The NLS sequences are recognized by proteins (importins α and β), which act as transporters and mediators of translocation across the nuclear envelope. NLSs are generally enriched in charged amino acids such as arginine, histidine, and lysine, conferring a positive charge which is partially responsible for their recognition by importins. Accordingly, in some embodiments, the polypeptide cargo may

comprise an NLS for facilitating nuclear delivery, such as one or more of the NLSs as listed in **Table C**, or a variant thereof having nuclear targeting activity. Of course, it is understood that, in certain embodiments, the polypeptide cargo may comprise its natural NLS.

Table C: Nuclear localization signals

Name	Amino acid sequence	SEQ ID NO:	Reference(s)
E1a	KRPRP	28	(Kohler, Gorlich et al., 2001)
SV40 T-Ag	PKKKRKV	29	(Lanford, Kanda et al., 1986)
c-myc	PAAKRVKLD	30	(Makkerh, Dingwall et al., 1996)
Op-T-NLS	SSDDEATADSQHAAPPKKKKRKV	31	(Chan and Jans 1999)
Vp3	KKKKRK	32	(Nakanishi, Shum et al., 2002)
Nucleoplasmin	KRPAATKKAGQAKKKK	33	(Fanara, Hodel et al., 2000)
Histone 2B NLS	DGKKRKRSRK	34	(Moreland, Langevin et al., 1987)
Xenopus N1	VRKKRKTEESPLKDKDAKSKQE	35	(Kleinschmidt and Seiter 1988)
PARP	KRKGDEVGVDECAKSKK	36	(Schreiber, Molinete et al., 1992)
PDX-1	RRMKWKK	37	(Moede, Leibiger et al., 1999)
QKI-5	RVHPYQR	38	(Wu, Zhou et al., 1999)
HCDA	KRPACTLKPECVQQLLVCSQEAKK	39	(Somasekaram, Jarmuz et al., 1999)
H2B	GKKRSKA	40	(Moreland, Langevin et al., 1987)
v-Rel	KAKRQR	41	(Gilmore and Temin 1988)
Amida	RKRRR	42	(Irie, Yamagata et al., 2000)
RanBP3	PPVKRERTS	43	(Welch, Franke et al., 1999)
Pho4p	PYLNRKRGKP	44	(Welch, Franke et al., 1999)
LEF-1	KKKKRKREK	45	(Prieve and Waterman 1999)
TCF-1	KKKRRSREK	46	(Prieve and Waterman 1999)
BDV-P	PRPRKIPR	47	(Shoya, Kobayashi et al., 1998)
TR2	KDCVINKHHRNRCQYCRLQR	48	(Yu, Lee et al., 1998)
SOX9	PRRRK	49	(Sudbeck and Scherer 1997)
Max	PQSRKKLR	50	(Kato, Lee et al., 1992)

5

Once delivered to the cytoplasm, **recombinant** proteins are exposed to protein trafficking system of eukaryotic cells. Indeed, all proteins are synthesized in the cell's cytoplasm and are then redistributed to their final subcellular localization by a system of transport based on small amino acid sequences recognized by shuttle proteins (Karniely and Pines 2005, Stojanovski, Bohnert et al., 2012). In addition to NLSs, other localization sequences can mediate subcellular targeting to various organelles following intracellular delivery of the polypeptide cargos of the present description. Accordingly, in some embodiments, polypeptide cargos of the present description may comprise a subcellular localization signal for facilitating delivery of the shuttle agent and cargo to specific organelles, such as one or more of the sequences as listed in **Table D**, or a variant thereof having corresponding subcellular targeting activity.

15

Table D: Subcellular localization signals

Name	Amino acid sequence	SEQ ID NO:	Reference(s)
Mitochondrial signal sequence from Tim9	NLVERCFTD	51	(Milenkovic, Ramming et al., 2009)

Mitochondrial signal sequence from Yeast cytochrome c oxidase subunit IV	MLSLRQSIRFFK	52	(Hurt, Pesold-Hurt et al., 1985)
Mitochondrial signal sequence from 18S rRNA	MLISRCKWSRFPGNQR	53	(Bejarano and Gonzalez 1999)
Peroxisome signal sequence – PTS1	SKL	54	(Gould, Keller et al., 1989)
Nucleolar signal sequence from BIRC5	MQRKPTIRRKNLRLRRK	55	(Scott, Boisvert et al., 2010)
Nucleolar signal sequence from RECQL4	KQAWKQKWRKK	56	(Scott, Boisvert et al., 2010)

In some embodiments, the cargo can be a biologically active compound such as a biologically active (recombinant) polypeptide (e.g., a transcription factor, a cytokine, or a nuclease) intended for intracellular delivery. As used herein, the expression “**biologically active**” refers to the ability of a compound to mediate a structural, regulatory, and/or biochemical function when introduced in a target cell.

In some embodiments, the cargo may be a recombinant polypeptide intended for nuclear delivery, such as a transcription factor. In some embodiments, the transcription factor can be HOXB4 (Lu, Feng et al., 2007), NUP98-HOXA9 (Takeda, Goolsby et al., 2006), Oct3/4, Sox2, Sox9, Klf4, c-Myc (Takahashi and Yamanaka 2006), MyoD (Sung, Mun et al., 2013), Pdx1, Ngn3 and MafA (Akinci, Banga et al., 2012), Blimp-1 (Lin, Chou et al., 2013), Eomes, T-bet (Gordon, Chaix et al., 2012), FOXO3A (Warr, Binnewies et al., 2013), NF-YA (Dolfini, Minuzzo et al., 2012), SALL4 (Aguila, Liao et al., 2011), ISL1 (Fonoudi, Yeganeh et al., 2013), FoxA1 (Tan, Xie et al., 2010), Nanog, Esrrb, Lin28 (Buganim et al., 2014), HIF1-alpha (Lord-Dufour et al., 2009), Hlf, Runx1t1, Pbx1, Lmo2, Zfp37, Prdm5 (Riddell et al., 2014), or Bcl-6 (Ichii, Sakamoto et al., 2004).

In some embodiments, the cargo may be a recombinant polypeptide intended for nuclear delivery, such as a nuclease useful for genome editing technologies. In some embodiments, the nuclease may be an RNA-guided endonuclease, a CRISPR endonuclease, a type I CRISPR endonuclease, a type II CRISPR endonuclease, a type III CRISPR endonuclease, a type IV CRISPR endonuclease, a type V CRISPR endonuclease, a type VI CRISPR endonuclease, CRISPR associated protein 9 (Cas9), Cpf1 (Zetsche et al., 2015), a zinc-finger nuclease (ZFN), a Transcription activator-like effector nuclease (TALEN) (Cox et al., 2015), a homing endonuclease, a meganuclease, or any combination thereof. Other nucleases not explicitly mentioned here may nevertheless be encompassed in the present description. In some embodiments, the nuclease may be fused to a nuclear localization signal (e.g., Cas9-NLS; Cpf1-NLS; ZFN-NLS; TALEN-NLS). In some embodiments, the nuclease may be complexed with a nucleic acid (e.g., one or more guide RNAs, a crRNA, a tracrRNAs, or both a crRNA and a tracrRNA). In some embodiments, the nuclease may possess DNA or RNA-binding activity, but may lack the ability to cleave DNA.

In some embodiments, the shuttle agents of the present description may be used for intracellular delivery (e.g., nuclear delivery) of one or more CRISPR endonucleases, for example one or more of the CRISPR endonucleases described below.

Type I and its subtypes A, B, C, D, E, F and I, including their respective Cas1, Cas2, Cas3, Cas4, Cas6, Cas7 and Cas8 proteins, and the signature homologs and subunits of these Cas proteins including Cse1, Cse2, Cas7, Cas5, and Cas6e subunits in *E. coli* (type I-E) and Csy1, Csy2, Csy3, and Cas6f in *Pseudomonas aeruginosa* (type I-F) (Wiedenheft et al., 2011; Makarova et al, 2011). Type II and its subtypes A, B, C, including their respective Cas1, Cas2 and Cas9 proteins, and the signature homologs and subunits of these Cas proteins including Csn complexes (Makarova et al, 2011). Type III and its subtypes A, B and MTH326-like module, including their respective Cas1, Cas2, Cas6 and Cas10 proteins, and the signature homologs and subunits of these Cas proteins including Csm and CMR complexes (Makarova et al, 2011). Type IV represents the Csf3 family of Cas proteins. Members of this family show up near CRISPR repeats in *Acidithiobacillus ferrooxidans* ATCC 23270, *Azoarcus* sp. (strain *Ebn1*), and *Rhodoferrax ferrireducens* (strain DSM 15236/ATCC BAA-621/T118). In the latter two species, the CRISPR/Cas locus is found on a plasmid. Type V and its subtypes have only recently been discovered and include Cpf1, C2c1, and C2c3. Type VI includes the enzyme C2c2, which reported shares little homology to known sequences.

In some embodiments, the shuttle agents of the present description may be used in conjunction with one or more of the nucleases, endonucleases, RNA-guided endonuclease, CRISPR endonuclease described above, for a variety of applications, such as those described herein. CRISPR systems interact with their respective nucleic acids, such as DNA binding, RNA binding, helicase, and nuclease motifs (Makarova et al, 2013; Barrangou & Marraffini, 2014). CRISPR systems may be used for different genome editing applications including:

- a Cas-mediated genome editing method conducting to non-homologous end-joining (NHEJ) and/or Homologous-directed recombination (HDR) (Cong et al, 2013);
- a catalytically dead Cas (dCas) that can repress and /or activate transcription initiation when bound to promoter sequences, to one or several gRNA(s) and to a RNA polymerase with or without a complex formation with others protein partners (Bikard et al, 2013);
- a catalytically dead Cas (dCas) that can also be fused to different functional proteins domains as a method to bring enzymatic activities at specific sites of the genome including transcription repression, transcription activation, chromatin remodeling, fluorescent reporter, histone modification, recombinase system acetylation, methylation, ubiquitylation, phosphorylation, sumoylation, ribosylation and citrullination (Gilbert et al, 2013).

The person of ordinary skill in the art will understand that the present shuttle agents, although exemplified with Cas9 in the present examples, may be used with other nucleases as described herein. Thus, nucleases such as Cpf1, Cas9, and variants of such nucleases or others, are encompassed by the present description. It should be understood that, in one aspect, the present description may broadly cover any cargo having nuclease activity, such as an RNA-guided endonuclease, or variants thereof (e.g., those that can bind to DNA or RNA, but have lost their nuclease activity; or those that have been fused to a transcription factor).

In some embodiments, the polypeptide cargo may be a cytokine such as a chemokine, an interferon, an interleukin, a lymphokine, or a tumour necrosis factor. In some embodiments, the polypeptide cargo may be a hormone or growth factor. In some embodiments, the cargo may be an antibody (e.g., a labelled antibody). In some embodiments, the cargo can be a detectable label (fluorescent polypeptide or reporter enzyme) that is intended for intracellular delivery, for example, for research and/or diagnostic purposes.

In some embodiments, the cargo may be a globular protein or a fibrous protein. In some embodiments, the cargo may have a molecule weight of any one of about 5, 10, 15, 20, 25, 30, 35, 40, 45, to 50 to about 150, 200, 250, 300, 350, 400, 450, 500 kDa or more. In some embodiments, the cargo may have a molecule weight of between about 20 to 200 kDa.

Non-toxic, metabolizable synthetic peptides and shuttle agents

In some embodiments, synthetic peptides and shuttle agents of the present description may be non-toxic to the intended target eukaryotic cells at concentrations up to 50 μ M, 45 μ M, 40 μ M, 35 μ M, 30 μ M, 25 μ M, 20 μ M, 15 μ M, 10 μ M, 9 μ M, 8 μ M, 7 μ M, 6 μ M, 5 μ M, 4 μ M, 3 μ M, 2 μ M, 1 μ M, 0.5 μ M, 0.1 μ M, or 0.05 μ M. Cellular toxicity of shuttle agents of the present description may be measured using any suitable method. Furthermore, transduction protocols may be adapted (e.g., concentrations of shuttle and/or cargo used, shuttle/cargo exposure times, exposure in the presence or absence of serum), to reduce or minimize toxicity of the shuttle agents, and/or to improve/maximize transfection efficiency.

In some embodiments, synthetic peptides and shuttle agents of the present description may be readily metabolizable by intended target eukaryotic cells. For example, the synthetic peptides and shuttle agents may consist entirely or essentially of peptides or polypeptides, for which the target eukaryotic cells possess the cellular machinery to metabolize/degrade. Indeed, the intracellular half-life of the synthetic peptides and polypeptide-based shuttle agents of the present description is expected to be much lower than the half-life of foreign organic compounds such as fluorophores. However, fluorophores can be toxic and must be investigated before they can be safely used clinically (Alford et al., 2009). In some embodiments, synthetic peptides and shuttle agents of the present description may be suitable for clinical use. In some embodiments, the synthetic peptides and shuttle agents of the present description may avoid the use of domains or compounds for which toxicity is uncertain or has not been ruled out.

Cocktails

In some embodiments, the present description relates to a composition comprising a cocktail of at least 2, at least 3, at least 4, or at least 5 different types of the synthetic peptides or polypeptide-based shuttle agents as defined herein. In some embodiments, combining different types of synthetic peptides or polypeptide-based shuttle agents (e.g., different shuttle agents comprising different types of CPDs) may provide increased versatility for delivering different polypeptide cargos intracellularly. Furthermore, without being bound by theory, combining lower

concentrations of different types of shuttle agents may help reduce cellular toxicity associated with using a single type of shuttle agent (e.g., at higher concentrations).

Methods, kits, uses and cells

5 In some embodiments, the present description relates to a method for increasing the transduction efficiency of a polypeptide cargo to the cytosol of a target eukaryotic cell. The method may comprise contacting the target eukaryotic cell with the synthetic peptide, polypeptide-based shuttle agent, or composition as defined herein, and the polypeptide cargo. In some embodiments, the synthetic peptide, polypeptide-based shuttle agent, or composition may be pre-incubated with the polypeptide cargo to form a mixture, prior to exposing the target eukaryotic cell to that mixture. In some embodiments, the type of CPD may be selected based on the amino acid sequence of the polypeptide cargo to be delivered intracellularly. In other embodiments, the type of CPD and ELD may be selected to take into account the amino acid sequence of the polypeptide cargo to be delivered intracellularly, the type of cell, the type of tissue, etc.

15 In some embodiments, the method may comprise multiple treatments of the target cells with the synthetic peptide, polypeptide-based shuttle agent, or composition (e.g., 1, 2, 3, 4 or more times per day, and/or on a pre-determined schedule). In such cases, lower concentrations of the synthetic peptide, polypeptide-based shuttle agent, or composition may be advisable (e.g., for reduced toxicity). In some embodiments, the cells may be suspension cells or adherent cells. In some embodiments, the person of skill in the art will be able to adapt the teachings of the present description using different combinations of shuttles, domains, uses and methods to suit particular needs of delivering a polypeptide cargo to particular cells with a desired viability.

20 In some embodiments, the methods of the present description may apply to methods of delivering a polypeptide cargo intracellularly to a cell *in vivo*. Such methods may be accomplished by parenteral administration or direct injection into a tissue, organ, or system.

25 In some embodiments, the synthetic peptide, polypeptide-based shuttle agent, or composition, and the polypeptide cargo may be exposed to the target cell in the presence or absence of serum. In some embodiments, the method may be suitable for clinical or therapeutic use.

In some embodiments, the present description relates to a kit for increasing the transduction efficiency of a polypeptide cargo to the cytosol of a target eukaryotic cell. The kit may comprise the synthetic peptide, polypeptide-based shuttle agent, or composition as defined herein, and a suitable container.

30 In some embodiments, the target eukaryotic cells may be an animal cell, a mammalian cell, or a human cell. In some embodiments, the target eukaryotic cells may be a stem cell (e.g., embryonic stem cells, pluripotent stem cells, induced pluripotent stem cells, neural stem cells, mesenchymal stem cells, hematopoietic stem cells, peripheral blood stem cells), primary cells (e.g., myoblasts, fibroblasts), or an immune cell (e.g., T cells, dendritic cells, antigen presenting cells). In some embodiments, the present description relates to an isolated cell comprising a synthetic

peptide or polypeptide-based shuttle agent as defined herein. In some embodiments, the cell may be a protein-induced pluripotent stem cell. It will be understood that cells that are often resistant or not amenable to protein transduction may be interesting candidates for the synthetic peptides or polypeptide-based shuttle agents of the present description.

5

Other objects, advantages and features of the present description will become more apparent upon reading of the following non-restrictive description of specific embodiments thereof, given by way of example only with reference to the accompanying drawings.

10 **EXAMPLES****Example 1:****Materials and Methods****1.1 Materials**

All chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA or Oakville, ON, Canada) or equivalent grade from BioShop Canada Inc. (Mississauga, ON, Canada) or VWR (Ville Mont-Royal, QC, Canada), unless otherwise noted.

1.2 Reagents**Table 1.1:** Reagents

Material	Company	City, Province-State, Country
RPMI 1640 media	Sigma-Aldrich	Oakville, ON, Canada
DMEM	Sigma-Aldrich	Oakville, ON, Canada
Fetal bovine serum (FBS)	NorthBio	Toronto, ON, Canada
L-glutamine-Penicillin-Streptomycin	Sigma-Aldrich	Oakville, ON, Canada
Trypsin-EDTA solution	Sigma-Aldrich	Oakville, ON, Canada
pEGFP-C1	CLONTECH Laboratories	Palo Alto, CA, USA
FITC-Antibody α-tubulin	Abcam ab64503	Cambridge, MA, USA
ITS	Invitrogen/41400-045	Burlington, ON, Canada
FGF 2	Feldan Bio/1D-07-017	Quebec, QC, Canada
Dexamethasone	Sigma-Aldrich/D8893	Oakville, ON, Canada
Bovine serum albumin (BSA)	Sigma-Aldrich/A-1933	Oakville, ON, Canada
MB1 media	GE Healthcare HyClone	Logan, Utah, USA
Calcein	Sigma-Aldrich/ C0875	Oakville, ON, Canada
HisTrap™ FF column	GE Healthcare	Baie d'Urfe, QC, Canada
Q Sepharose™	GE Healthcare	Baie d'Urfe, QC, Canada
SP Sepharose™	GE Healthcare	Baie d'Urfe, QC, Canada
Amicon Ultra centrifugal filters	EMD Millipore	Etobicoke, ON Canada
Label IT® Cy®5 kit	Mirus Bio LLC	Madison, WI, USA
Calf serum	NorthBio	Toronto, ON, Canada
beta-mercaptoethanol	Sigma-Aldrich	Oakville, ON, Canada

IL-2	Feldan Bio/ rhIL-2 Research	Quebec, QC, Canada
Rezazurine sodium salt	Sigma-Aldrich/R7017-1G	Oakville, ON, Canada
Anti-HOXB4 monoclonal antibody	Novus Bio #NBP2-37257	Oakville, ON, Canada
Alexa™-594 Anti-Mouse	Abcam #150116	Toronto, ON, Canada
Fluoroshield™ with DAPI	Sigma #F6057	Oakville, ON, Canada
GFP Monoclonal antibody	Feldan Bio #A017	Quebec, QC, Canada
Phusion™ High-Fidelity DNA polymerase	(NEB #M0530S)	Whitby, ON, Canada
Edit-R™ Synthetic crRNA Positive Controls	(Dharmacon #U-007000-05)	Ottawa, ON, Canada
T7 Endonuclease I	(NEB, Cat #M0302S)	Whitby, ON, Canada
FastFect™ transfection reagent	(Feldan Bio # 9K-010-0001)	Quebec, QC, Canada

1.3 Cell lines

HeLa, HEK293A, HEK293T, THP-1, CHO, NIH3T3, CA46, Balb3T3 and HT2 cells were obtained from American Type Culture Collection (Manassas, VA, USA) and cultured following the manufacturer's instructions.

5 Myoblasts are primary human cells kindly provided by Professor J.P. Tremblay (Université Laval, Quebec, Canada).

Table 1.2: Cell lines and culture conditions

Cell lines	Description	ATCC/others	Culture media	Serum	Additives
HeLa (adherent cells)	Human cervical carcinoma cells	ATCC™ CCL-2	DMEM	10% FBS	L-glutamine 2 mM Penicillin 100 units Streptomycin 100 µg/mL
HEK 293A (adherent cells)	Human embryonic Epithelial kidney cells	ATCC™ CRL-1573	DMEM	10% FBS	L-glutamine 2 mM Penicillin 100 units Streptomycin 100 µg/mL
HEK 293T (adherent cells)	Human embryonic Epithelial kidney cells	ATCC™ CRL-3216	DMEM	10% FBS	L-glutamine 2 mM Penicillin 100 units Streptomycin 100 µg/mL
THP-1 (suspension cells)	Acute monocytic leukemia	ATCC™ TIB202	RPMI 1640	10% FBS	2- mercaptoethanol 0.05 mM L-glutamine 2 mM Penicillin 100 units Streptomycin 100 µg/mL
Myoblasts (primary adherent cells)	Human (13 months) myoblasts	Kindly provided by Professor JP Tremblay	MB1	15% FBS	ITS 1x, FGF 2 10 ng/mL, Dexamethasone 0.39 µg/mL, BSA 0.5mg/mL, MB1 85%
CHO (adherent cells)	Chinese hamster ovary cells	ATCC™ CCL-61	DMEM	10% FBS	L-glutamine 2 mM Penicillin 100 units Streptomycin 100 µg/mL
NIH3T3 (adherent cells)	Fibroblasts	ATCC™ CRL-1658	DMEM	10% Calf serum	L-glutamine 2 mM Penicillin 100 units Streptomycin 100 µg/mL

HT2 (suspension cells)	T lymphocytes	ATCC™ CRL-1841	RPMI 1640	10% FBS	200 IU/mL IL-2 β-mercaptoethanol 0.05 mM L-glutamine 2 mM Penicillin 100 units Streptomycin 100 µg/mL
CA46 (suspension cells)	Homo sapiens Burkitt's lymphoma	ATCC™ CRL-1648	RPMI 1640	20% FBS	L-glutamine 2 mM Penicillin 100 units Streptomycin 100 µg/mL
Balb3T3 (adherent cells)	Fibroblasts	ATCC™ CCL-163	DMEM	10% Calf serum	L-glutamine 2 mM Penicillin 100 units Streptomycin 100 µg/mL
Jurkat (suspension cells)	Human T cells	ATCC™ TIB-152	RPMI 1640	10% FBS	L-glutamine 2 mM Penicillin 100 units Streptomycin 100 µg/mL

FBS: Fetal bovine serum

1.4 Protein purification

Fusion proteins were expressed in bacteria (*E. coli* BL21DE3) under standard conditions using an isopropyl β-D-1-thiogalactopyranoside (IPTG) inducible vector containing a T5 promoter. Culture media contained 24 g yeast extract, 12 g tryptone, 4 mL glycerol, 2.3 g KH₂PO₄, and 12.5 g K₂HPO₄ per liter. Bacterial broth was incubated at 37°C under agitation with appropriate antibiotic (e.g., ampicillin). Expression was induced at optical density (600 nm) between 0.5 and 0.6 with a final concentration of 1 mM IPTG for 3 hours at 30°C. Bacteria were recuperated following centrifugation at 5000 RPM and bacterial pellets were stored at -20°C.

Bacterial pellets were resuspended in Tris buffer (Tris 25 mM pH 7.5, NaCl 100mM, imidazole 5 mM) with phenylmethylsulfonyl fluoride (PMSF) 1 mM, and lysed by passing 3 times through the homogenizer Panda 2K™ at 1000 bar. The solution was centrifuged at 15000 RPM, 4°C for 30 minutes. Supernatants were collected and filtered with a 0.22 µm filtration device.

Solubilized proteins were loaded, using a FPLC (AKTA Explorer 100R), on HisTrap™ FF column previously equilibrated with 5 column volumes (CV) of Tris buffer. The column was washed with 30 column volumes (CV) of Tris buffer supplemented with 0.1% Triton™ X-114 followed with 30 CV of Tris buffer with imidazole 40 mM. Proteins were eluted with 5 CV of Tris buffer with 350 mM Imidazole and collected. Collected fractions corresponding to specific proteins were determined by standard denaturing SDS-PAGE.

Purified proteins were diluted in Tris 20 mM at the desired pH according to the protein's pI and loaded on an appropriate ion exchange column (Q Sepharose™ or SP Sepharose™) previously equilibrated with 5 CV of Tris 20 mM, NaCl 30 mM. The column was washed with 10 CV of Tris 20 mM, NaCl 30 mM and proteins were eluted with a NaCl gradient until 1 M on 15 CV. Collected fractions corresponding to specific proteins were determined by standard denaturing SDS-PAGE. Purified proteins were then washed and concentrated in PBS 1X on Amicon Ultra™ centrifugal filters 10,000 MWCO. Protein concentration was evaluated using a standard Bradford assay.

1.5 Synthetic peptides and shuttle agents

All peptides used in this study were purchased from GLBiochem (Shanghai, China) and their purities were confirmed by high-performance liquid chromatography analysis and mass spectroscopy. In some cases, chimeric peptides were synthesized to contain a C-terminal cysteine residue to allow the preparation of peptide dimers. These dimeric peptides were directly synthesized with a disulfide bridge between the C-terminal cysteines of two monomers. The amino acid sequences and characteristics of each of the synthetic peptides and shuttle agents tested in the present examples are summarized in **Table 1.3**.

Table 1.3: Synthetic peptides and shuttle agents tested

Domain(s)	Peptide or Shuttle agent	Amino acid (a.a.) sequence [SEQ ID NO; not including C-terminal Cys, unless indicated with an *]	a.a.	MW (kDa)	pI	Charge	Hydrophobicity index
ELD	CM18	KWKLFKKIGAVLKVLTTG [1]	18	2.03	10.60	5+ / 0-	0.350
	C(LLKK) ₃ C	CLLKKLLKLLKKC [63]	14	1.69	10.05	6+ / 0-	0.314
	LAH4	KKALLALALHHLAHLALALAKKA [6]	26	2.78	10.48	4+ / 0-	0.923
	KALA	WEAKLAKALAKALAKHLAKALAKACEA [14]	30	3.13	9.9	7+ / 2-	0.283
CPD	TAT-cys	YGRKKRRQRRRC [17]	12	1.66	12.01	8+ / 0-	-3.125
	Penetratin-cys	RQIKIWFQNRRMKWKKC [18]	17	2.35	11.75	7+ / 0-	-1.482
	PTD4	YARAAARQARA [65]	11	1.2	11.72	3+ / 0-	-0.682
His-PTD4	His-PTD4	HHHHHHYARAAARQARA [81]	17	2.03	11.71	3+ / 0-	-1.57
CPD-ELD	TAT-CM18	YGRKKRRQRRRCWKLFKKIGAVLKVLTTG [66]	30	3.68	12.02	13+ / 0-	-1.041
	TAT-KALA	YGRKKRRQRRRCWEAKLAKALAKALAKHLAKALAKACEA [67]	42	4.67	11.46	15+ / 2-	-0.768
	PTD4-KALA	YARAAARQARAWWEAKLAKALAKALAKHLAKALAKACEA [82]	41	4.32	10.46	10+ / 2-	0.024
	9Arg-KALA	RRRRRRRRRWEAKLAKALAKALAKHLAKALAKACEA [83]	39	4.54	12.11	16+ / 2-	-0.821
	Pep1-KALA	KETWWETWWTEWSQPKKKRKVWEAKLAKALAKHLAKALAKACEA [84]	51	5.62	10.01	13+ / 5-	-0.673
	Xentry-KALA	LCLRPVGWEAKLAKALAKALAKHLAKALAKACEA [85]	37	3.87	9.93	8+ / 2-	0.441
	SynB3-KALA	RRLSYSRRRFWEAKLAKALAKALAKHLAKALAKACEA [86]	40	4.51	11.12	12+ / 2-	-0.258
ELD-CPD	CM18-TAT-Cys	KWKLFKKIGAVLKVLTTGYGRKKRRQRRRC [57]	30	3.67	12.02	13+ / 0-	-1.04
	CM18-Penetratin-Cys	KWKLFKKIGAVLKVLTTGRQIKIWFQNRRMKWKKC [58]	35	4.36	11.36	12+ / 0-	-0.54
	dCM18-TAT-Cys (CM18-TAT-cys dimer)	KWKLFKKIGAVLKVLTTGYGRKKRRQRRRC [57]	60	7.34	12.16	26+ / 0-	-1.04
	dCM18-Penetratin-Cys (CM18-Penetratin-Cys dimer)	KWKLFKKIGAVLKVLTTGRQIKIWFQNRRMKWKKC [58]	70	8.72	12.05	24+ / 0-	-0.54
	VSVG-PTD4	KFTIVFPHNQKGNWKNVPSNYHYCPYARAAARQARA [87]	36	4.2	10.3	6+ / 0-	-0.89

	EB1-PTD4	LIRLWSHLIHWQNRRLKWKKKYARAAA RQARA [88]	34	4.29	12.31	10+ / 0-	-0.647
	JST-PTD4	GLFEALLELLESLWELLLEAYARAAARQAR A [89]	31	3.49	4.65	5+ / 3-	0.435
	CM18-PTD4	KWKLFKKIGAVLKVLTTGYARAAARQARA [90]	29	3.217	11.76	8+ / 0-	-0.041
	6Cys-CM18-PTD4	CCCCCKWKLFKKIGAVLKVLTTGYARAA ARQARA [91]	35	3.835	9.7	8+ / 0-	0.394
	CM18-L1-PTD4	KWKLFKKIGAVLKVLTTGGGSYARAAARQ ARA [92]	32	3.42	11.76	8+ / 0-	-0.087
	CM18-L2-PTD4	KWKLFKKIGAVLKVLTTGGSGGGSYARA AARQARA [93]	36	3.68	11.76	8+ / 0-	-0.133
	CM18-L3-PTD4	KWKLFKKIGAVLKVLTTGGSGGGSGGG SGYARAAARQARA [94]	41	3.99	11.76	8+ / 0	-0.176
His-ELD-CPD	Met-His-CM18-TAT- Cys	MHHHHHKWKLFKKIGAVLKVLTTGYGRK KRRQRRRC [59*]	37	4.63	12.02	13+ / 0-	-1.311
	His-CM18-TAT	HHHHHKWKLFKKIGAVLKVLTTGYGRKK RRQRR [95]	35	4.4	12.31	13+ / 0-	-1.208
	His-CM18-PTD4	HHHHHKWKLFKKIGAVLKVLTTGYARAA ARQARA [68]	35	4.039	11.76	8+ / 0-	-0.583
	His-CM18-PTD4-6Cys	HHHHHKWKLFKKIGAVLKVLTTGYARAA ARQARACCCCC [96*]	41	4.659	9.7	8+ / 0-	-0.132
	His-CM18-9Arg	HHHHHKWKLFKKIGAVLKVLTTGRRRRR RRRR [69]	33	4.26	12.91	14+ / 0-	-1.618
	His-CM18-Transportan	HHHHHKWKLFKKIGAVLKVLTTGGWTLN SAGYLLKINKALAAALAKKIL [70]	50	5.62	10.6	9+ / 0-	0.092
	His-LAH4-PTD4	HHHHHKKALLALALHHLALHLALALAL KAYARAAARQARA [71]	43	4.78	11.75	7+ / 0-	-0.63
	His-C(LLKK) ₃ C-PTD4	HHHHHCLLKKLLKLLKKCYARAAARQA RA [72]	31	3.56	11.21	9+ / 0-	-0.827
	3His-CM18-PTD4	HHHKWKLFKKIGAVLKVLTTGYARAAARQ ARA [97]	32	3.63	11.76	8+ / 0-	-0.338
	12His-CM18-PTD4	HHHHHHHHHHHKWKLFKKIGAVLKVLTT GYARAAARQARA [98]	41	4.86	11.76	8+ / 0-	-0.966
	HA-CM18-PTD4	HHHAHHHKWKLFKKIGAVLKVLTTGYARA AARQARA [99]	36	4.11	11.76	8+ / 0-	-0.517
	3HA-CM18-PTD4	HAHAHAHAKWKLFKKIGAVLKVLTTGYA RAAARQARA [100]	38	4.25	11.76	8+ / 0-	-0.395
ELD-His-CPD	CM18-His-PTD4	KWKLFKKIGAVLKVLTTGHHHHHHYARAA ARQARA [101]	35	4.04	11.76	8+ / 0-	-0.583
His-ELD-CPD-His	His-CM18-PTD4-His	HHHHHKWKLFKKIGAVLKVLTTGYARAA ARQARAHHHHH [102]	41	4.86	11.76	8+ / 0-	-0.966

Results computed using the ProtParam™ online tool available from ExPASy™ Bioinformatics Resource Portal
(<http://web.expasy.org/cgi-bin/protparam/protparam>)

MW: Molecular weight

pI: Isoelectric point

5 Charge: Total number of positively (+) and negatively (-) charged residues

Example 2:**Peptide shuttle agents facilitate escape of endosomally-trapped calcein****2.1 Endosome escape assays**

5 Microscopy-based and flow cytometry-based fluorescence assays were developed to study endosome leakage and to determine whether the addition of the shuttle agents facilitates endosome leakage of the polypeptide cargo.

2.1.1 Endosomal leakage visualization by microscopy

10 Calcein is a membrane-impermeable fluorescent molecule that is readily internalized by cells when administered to the extracellular medium. Its fluorescence is pH-dependent and calcein self-quenches at higher concentrations. Once internalized, calcein becomes sequestered at high concentrations in cell endosomes and can be visualized by fluorescence microscopy as a punctate pattern. Following endosomal leakage, calcein is released to the cell cytoplasm and this release can be visualized by fluorescence microscopy as a diffuse pattern.

15 One day before the calcein assay was performed, mammalian cells (e.g., HeLa, HEK293A, or myoblasts) in exponential growth phase were harvested and plated in a 24-well plate (80,000 cells per well). The cells were allowed to attach by incubating overnight in appropriate growth media, as described in **Example 1**. The next day, the media was removed and replaced with 300 μ L of fresh media without FBS containing 62.5 μ g/mL (100 μ M) of calcein, except for HEK293A (250 μ g/mL, 400 μ M). At the same time, the shuttle agent(s) to be tested was added at a predetermined concentration. The plate was incubated at 37°C for 30 minutes. The cells were washed with 1x PBS (37°C) and fresh media containing FBS was added. The plate was incubated at 37°C for 2.5 hours. The cells were washed three times and were visualized by phase contrast and fluorescence microscopy (IX81™, Olympus).

20 A typical result is shown in **Figure 1A**, in which untreated HEK293A cells loaded with calcein ("100 μ M calcein") show a low intensity, punctate fluorescent pattern when visualized by fluorescence microscopy (upper left panel). In contrast, HeLa cells treated with a shuttle agent that facilitates endosomal escape of calcein ("100 μ M calcein + CM18-TAT 5 μ M") show a higher intensity, more diffuse fluorescence pattern in a greater proportion of cells (upper right panel).

2.1.2 Endosomal leakage quantification by flow cytometry

30 In addition to microscopy, flow cytometry allows a more quantitative analysis of the endosomal leakage as the fluorescence intensity signal increases once the calcein is released in the cytoplasm. Calcein fluorescence is optimal at physiological pH (e.g., in the cytosol), as compared to the acidic environment of the endosome.

One day before the calcein assay was performed, mammalian cells (e.g., HeLa, HEK293, or myoblasts) in exponential growth phase were harvested and plated in a 96-well plate (20,000 cells per well). The cells were

allowed to attach by incubating overnight in appropriate growth media, as described in **Example 1**. The next day, the media in wells was removed and replaced with 50 μ L of fresh media without serum containing 62.5 μ g/mL (100 μ M) of calcein, except for HEK293A (250 μ g/mL, 400 μ M). At the same time, the shuttle agent(s) to be tested was added at a predetermined concentration. The plate was incubated at 37°C for 30 minutes. The cells were washed with 1x PBS (37°C) and fresh media containing 5-10% serum was added. The plate was incubated at 37°C for 2.5 hours. The cells were washed with 1x PBS and detached using trypsinization. Trypsinization was stopped by addition of appropriate growth media, and calcein fluorescence was quantified using flow cytometry (Accuri C6, Becton, Dickinson and Company (BD)).

Untreated calcein-loaded cells were used as a control to distinguish cells having a baseline of fluorescence due to endosomally-trapped calcein from cells having increased fluorescence due to release of calcein from endosomes. Fluorescence signal means ("mean counts") were analyzed for endosomal escape quantification. In some cases, the "Mean Factor" was calculated, which corresponds to the fold-increase of the mean counts relative to control (untreated calcein-loaded cells). Also, the events scanned by flow cytometry corresponding to cells (size and granularity) were analyzed. The cellular mortality was monitored with the percentage of cells in the total events scanned. When it became lower than the control, it was considered that the number of cellular debris was increasing due to toxicity and the assay was discarded.

A typical result is shown in **Figure 1B**, in which an increase in fluorescence intensity (right-shift) is observed for calcein-loaded HeLa cells treated with a shuttle agent that facilitates endosomal escape ("Calcein 100 μ M + CM18-TAT 5 μ M", right panel), as compared to untreated calcein-loaded HeLa cells ("Calcein 100 μ M", left panel). The increase in calcein fluorescence is caused by the increase in pH associated with the release of calcein from the endosome (acidic) to the cytoplasm (physiological).

2.2 Results from endosome escape assays

2.2.1 *HeLa cells*

HeLa cells were cultured and tested in the endosomal escape assays as described in **Example 2.1**. The results of flow cytometry analyses are summarized below. In each case, the flow cytometry results were also confirmed by fluorescence microscopy (data not shown).

Table 2.1: CM18-Penetratin-Cys v. Controls in HeLa cells

Domains	Peptide	Cells	Concentration (μ M)	Mean Counts ($\pm$ St. Dev.; n=3)	Mean Factor
-	No peptide	HeLa	0	55 359 $\pm$ 6844	1.0
ELD	CM18	HeLa	5	46 564 $\pm$ 9618	0.8
CPD	TAT-Cys	HeLa	5	74 961 $\pm$ 9337	1.3
	Penetratin-Cys	HeLa	5	59 551 $\pm$ 7119	1.1
ELD + CPD	CM18 + TAT-Cys	HeLa	5 + 5	64 333 $\pm$ 6198	1.2
	CM18 + Penetratin-Cys	HeLa	5 + 5	40 976 $\pm$ 8167	0.7
ELD-CPD	CM18-Penetratin-Cys	HeLa	5	262 066 $\pm$ 28 146	4.7

Table 2.2: CM18-TAT-Cys v. Control in HeLa cells

Domains	Peptide	Cells	Concentration (μ M)	Mean counts (n=3)	Stand. dev.	Mean Factor
-	No peptide	HeLa	0	53 369	4192	1.0
ELD-CPD	CM18-TAT-Cys	HeLa	5	306 572	46 564	5.7

5 The results in **Tables 2.1 and 2.2** show that treating calcein-loaded HeLa cells with the shuttle agents CM18-Penetratin-Cys and CM18-TAT-Cys (having the domain structure ELD-CPD) results in increased mean cellular calcein fluorescence intensity, as compared to untreated control cells or cells treated with single-domain peptides used alone (CM18, TAT-Cys, Penetratin-Cys) or together (CM18 + TAT-Cys, CM18 + Penetratin-Cys). These results suggest that CM18-Penetratin-Cys and CM18-TAT-Cys facilitate escape of endosomally-trapped calcein, but that

10 single domain peptides (used alone or together) do not.

Table 2.3: Dose response of CM18-TAT-Cys in HeLa cells, data from **Figure 2**

Domains	Peptide	Cells	Concentration (μ M)	Mean counts (n=3)	Stand. dev.	Mean Factor
-	No peptide ("calcein 100 μ M")	HeLa	0	63 872	11 587	1.0
ELD-CPD	CM18-TAT-Cys	HeLa	1	86 919	39 165	1.4
	CM18-TAT-Cys	HeLa	2	137 887	13 119	2.2
	CM18-TAT-Cys	HeLa	3	174 327	11 519	2.7
	CM18-TAT-Cys	HeLa	4	290 548	16 593	4.5
	CM18-TAT-Cys	HeLa	5	383 685	5578	6.0

Table 2.4: Dose response of CM18-TAT-Cys in HeLa cells

Domains	Peptide	Cells	Concentration (μ M)	Mean counts (n=3)	Stand. dev.	Mean Factor
-	No peptide	HeLa	0	81 013	14 213	1.0
ELD-CPD	CM18-TAT-Cys	HeLa	3	170 652	63 848	2.1
	CM18-TAT-Cys	HeLa	4	251 799	33 880	3.1
	CM18-TAT-Cys	HeLa	5	335 324	10 651	4.1

Table 2.5: Dose response of CM18-TAT-Cys and CM18-Penetratin-Cys in HeLa cells, data from **Figure 3**

Domains	Peptide	Cells	Concentration (μ M)	Mean counts (n=3)	Stand. dev.	Mean Factor
-	No peptide	HeLa	0	62 503	23 752	1.0
ELD-CPD	CM18-TAT-Cys	HeLa	5	187 180	8593	3.0
	CM18-TAT-Cys	HeLa	8	321 873	36 512	5.1
	CM18-Penetratin-Cys	HeLa	5	134 506	2992	2.2
	CM18-Penetratin-Cys	HeLa	8	174 233	56 922	2.8

5 The results in **Tables 2.3 (Figure 2), 2.4, and 2.5 (Figure 3)** suggest that CM18-TAT-Cys and CM18-Penetratin-Cys facilitate escape of endosomally-trapped calcein in HeLa cells in a dose-dependent manner. In some cases, concentrations of CM18-TAT-Cys or CM18-Penetratin-Cys above 10 μ M were associated with an increase in cell toxicity in HeLa cells.

10 **Table 2.6:** Dimers v. monomers of CM18-TAT-Cys and CM18-Penetratin-Cys in HeLa cells

Domains	Peptide	Cells	Concentration (μ M)	Mean counts (n=4)	Stand. dev.	Mean Factor
-	No peptide	HeLa	0	60 239	9860	1.0
ELD-CPD	CM18-TAT-Cys	HeLa	4	128 461	25 742	2.1
	CM18-Penetratin-Cys	HeLa	4	116 873	3543	1.9
ELD-CPD dimer	dCM18-TAT-Cys	HeLa	2	79 380	4297	1.3
	dCM18-Penetratin-Cys	HeLa	2	128 363	8754	2.1

Table 2.7: Monomers v. dimers of CM18-TAT-Cys and CM18-Penetratin-Cys in HeLa cells

Domains	Peptide	Cells	Concentration (μM)	Mean counts (n=3)	Stand. dev.	Mean Factor
-	No peptide	HeLa	0	55 834	1336	1.0
ELD-CPD	CM18-TAT-Cys	HeLa	4	159 042	16 867	2.8
ELD-CPD dimer	dCM18-TAT-Cys	HeLa	2	174 274	9 553	3.1

The results in **Table 2.6 and 2.7** suggest that shuttle peptide dimers (which are molecules comprising more than one ELD and CPD) are able to facilitate calcein endosomal escape levels that are comparable to the corresponding monomers.

2.2.3 HEK293A cells

To examine the effects of the shuttle agents on a different cell line, HEK293A cells were cultured and tested in the endosomal escape assays as described in **Example 2.1**. The results of flow cytometry analyses are summarized below in **Table 2.8** and in **Figure 1B**.

Table 2.8: CM18-TAT-Cys in HEK293A cells

Domains	Peptide	Cells	Concentration (μM)	Mean counts (n=2)	Stand. dev.	Mean Factor
-	No peptide	HEK293A	0	165 819	7693	1.0
ELD-CPD	CM18-TAT-Cys	HEK293A	0.5	196 182	17 224	1.2
	CM18-TAT-Cys	HEK293A	5	629 783	1424	3.8

The results in **Table 2.8 and in Figure 1B** show that treating calcein-loaded HEK293A cells with the shuttle agent CM18-TAT-Cys results in increased mean cellular calcein fluorescence intensity, as compared to untreated control cells.

2.2.2 Myoblasts

To examine the effects of the shuttle agents on primary cells, primary myoblast cells were cultured and tested in the endosomal escape assays as described in **Example 2.1**. The results of flow cytometry analyses are summarized below in **Tables 2.9 and 2.10**, and in **Figure 4**. In each case, the flow cytometry results were also confirmed by fluorescence microscopy.

Table 2.9: Dose response of CM18-TAT-Cys in primary myoblasts, data from **Figure 4**

Domains	Peptide	Cells	Peptide Conc. (μM)	Mean counts (n=3)	Stand. dev.	Mean Factor
-	No peptide; no calcein ("Cells")	Myoblasts	0	863	61	n/a
-	No peptide ("Calcein 100 μM ")	Myoblasts	0	38 111	13 715	1.0
ELD-CPD	CM18-TAT-Cys	Myoblasts	5	79 826	12 050	2.1
	CM18-TAT-Cys	Myoblasts	8	91 421	10 846	2.4

Table 2.10: Dose response of CM18-TAT-Cys in primary myoblasts

Domains	Peptide	Cells	Peptide Conc. (μM)	Mean counts (n=3)	Stand. dev.	Mean Factor
-	No peptide	Myoblasts	0	31 071	21 075	1.0
ELD-CPD	CM18-TAT-Cys	Myoblasts	5	91 618	10 535	2.9
	CM18-TAT-Cys	Myoblasts	7.5	95 289	11 266	3.1

5 The results in **Table 2.9** (shown graphically in **Figure 4**) and **Table 2.10** suggest that CM18-TAT-Cys facilitates escape of endosomally-trapped calcein in a dose-dependent manner in primary myoblasts. Concentrations of CM18-TAT-Cys above 10 μM were associated with an increase in cell toxicity in myoblast cells, as for HeLa cells.

Table 2.11: Monomers v. dimers CM18-TAT-Cys and CM18-Penetratin-Cys in primary myoblasts

Domains	Peptide	Cells	Concentration (μM)	Mean counts	Stand. dev.	Mean Factor
-	No peptide	Myoblasts	0	30 175	4687	1.0
ELD-CPD	CM18-TAT-Cys	Myoblasts	5	88 686	19 481	2.9
ELD-CPD dimer	dCM18-TAT-Cys	Myoblasts	2.5	64 864	1264	2.1
ELD-CPD	CM18-Penetratin-Cys	Myoblasts	5	65 636	3288	2.2
ELD-CPD dimer	dCM18-Penetratin-Cys	Myoblasts	2.5	71 547	10 975	2.4

10 The results in **Table 2.11** suggest that shuttle peptide dimers are able to facilitate calcein endosomal escape levels that are comparable to the corresponding monomers in primary myoblasts.

Example 3:

Peptide shuttle agents increase GFP transduction efficiency

3.1 Protein transduction assay

One day before the transduction assay was performed, mammalian cells (e.g., HEK293, CHO, HeLa, THP-1, and myoblasts) in exponential growth phase were harvested and plated in a 96-well plate (20,000 cells per well). The cells were incubated overnight in appropriate growth media containing FBS (see **Example 1**). The next day, in separate sterile 1.5 mL tubes, cargo protein at 0.5 to 10 μ M (GFP, TAT-GFP, GFP-NLS, or FITC-labeled anti-tubulin antibody) was pre-mixed (pre-incubated) for 10 min at 37°C with shuttle agents (0.5 to 5 μ M) in 50 μ L of fresh medium without serum (unless otherwise specified). GFP, GFP-NLS and TAT-GFP are recombinant proteins developed and produced by Feldan (see **Example 3.4** below). FITC-labeled anti-tubulin antibody was purchased from Abcam (ab64503). The media in wells was removed and the cells were washed three times with freshly prepared phosphate buffered saline (PBS) previously warmed at 37°C. The cells were incubated with the cargo protein/shuttle agent mixture at 37°C for 5 or 60 min. After the incubation, the cells were quickly washed three times with freshly prepared PBS and/or heparin (0.5 mg/mL) previously warmed at 37°C. The washes with heparin were required for human THP-1 blood cells to avoid undesired cell membrane-bound protein background in subsequent analyses (microscopy and flow cytometry). The cells were finally incubated in 50 μ L of fresh medium with serum at 37°C before analysis.

3.2 Fluorescence microscopy analysis

The delivery of fluorescent protein cargo in cytosolic and nuclear cell compartments was observed with an Olympus IX70™ microscope (Japan) equipped with a fluorescence lamp (Model U-LH100HGAP0) and different filters. The Olympus filter U-MF2™ (C54942-Exc495/Em510) was used to observe GFP and FITC-labeled antibody fluorescent signals. The Olympus filter HQ-TR™ (V-N41004-Exc555-60/Em645-75) was used to observe mCherry™ and GFP antibody fluorescent signals. The Olympus filter U-MWU2™ (Exc330/Em385) was used to observe DAPI or Blue Hoechst fluorescent signals. The cells incubated in 50 μ L of fresh medium were directly observed by microscopy (Bright-field and fluorescence) at different power fields (4x to 40x). The cells were observed using a CoolSNAP-PRO™ camera (Series A02D874021) and images were acquired using the Image-Proplus™ software.

3.2a Cell immuno-labelling

Adherent cells were plated on a sterile glass strip at 1.5×10^5 cells per well in a 24-plate well and incubated overnight at 37°C. For fixation, cells were incubated in 500 μ L per well of formaldehyde (3.7% v/v) for 15 minutes at room temperature, and washed 3 times for 5 minutes with PBS. For permeabilization, cells were incubated in 500 μ L per well of Triton™ X-100 (0.2%) for 10 minutes at room temperature, and washed 3 times for 5 minutes with PBS. For blocking, cells were incubated in 500 μ L per well of PBS containing 1% BSA (PBS/BSA) for 60 minutes at room temperature. Primary mouse monoclonal antibody was diluted PBS/BSA (1%). Cells were incubated in 30 μ L of primary antibody overnight at 4°C. Cells were washed 3 times for 5 minutes with PBS. Secondary antibody was

diluted in PBS/BSA (1%) and cells were incubated in 250 μ L of secondary antibody 30 minutes at room temperature in the dark. Cells were washed 3 times for 5 minutes with PBS. Glass strips containing the cells were mounted on microscope glass slides with 10 μ L of the mounting medium Fluoroshield™ with DAPI.

5 3.3 Flow cytometry analysis:

The fluorescence of GFP was quantified using flow cytometry (Accuri C6, Becton, Dickinson and Company (BD)). Untreated cells were used to establish a baseline in order to quantify the increased fluorescence due to the internalization of the fluorescent protein in treated cells. The percentage of cells with a fluorescence signal above the maximum fluorescence of untreated cells, "mean %" or "Pos cells (%)", is used to identify positive fluorescent cells. "Relative fluorescence intensity (FL1-A)" corresponds to the mean of all fluorescence intensities from each cell with a fluorescent signal after fluorescent protein delivery with the shuttle agent. Also, the events scanned by flow cytometry corresponding to cells (size and granularity) were analyzed. The cellular toxicity (% cell viability) was monitored comparing the percentage of cells in the total events scanned of treated cells comparatively to untreated cells.

15 3.3a Viability analysis

The viability of cells was assessed with a rezazurine test. Rezazurine is a sodium salt colorant that is converted from blue to pink by mitochondrial enzymes in metabolically active cells. This colorimetric conversion, which only occurs in viable cells, can be measured by spectroscopy analysis in order to quantify the percentage of viable cells. The stock solution of rezazurine was prepared in water at 1 mg/100 mL and stored at 4°C. 25 μ L of the stock solution was added to each well of a 96-well plate, and cells were incubated at 37°C for one hour before spectrometry analysis. The incubation time used for the rezazurine enzymatic reaction depended on the quantity of cells and the volume of medium used in the wells.

25 3.4 Construction and amino acid sequence of GFP

The GFP-encoding gene was cloned in a T5 bacterial expression vector to express a GFP protein containing a 6x histidine tag and a serine/glycine rich linker in the N-terminal end, and a serine/glycine rich linker and a stop codon (-) at the C-terminal end. Recombinant GFP protein was purified as described in **Example 1.4**. The sequence of the GFP construct was:

30

MHHHHHGGGSGGGSGGASTGTGIRMVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDAT YGKLTLLKFICTTGKLPVPWPTLVTTLTYGVCFSRYPDHMKQHDFKSAPEGYVQERTIFFKDDG NYKTRAEVKFEGDTLVNRIELKGIDFKEDGNILGHKLEYNNSHNVYIMADKQKNGIKVNFKIRHN IEDGSVQLADHYQNTPIGDGPVLLPDNHYLSTQSALSKDPNEKRDHMLLEFVTAAGITLGMDEL YKGGSGGGSGGSGWIRASSGGREIS- [SEQ ID NO: 60]
--

35 (MW= 31.46 kDa; pI=6.19)
Serine/glycine rich linkers are in bold
GFP sequence is underlined

3.5 GFP transduction by CM18-TAT-Cys in HeLa cells: Fluorescence microscopy

HeLa cells were cultured and tested in the protein transduction assay described in **Example 3.1**. Briefly, GFP recombinant protein was co-incubated with 0, 3 or 5 μM of CM18-TAT, and then exposed to HeLa cells for 1 hour. The cells were observed by bright field and fluorescence microscopy as described in **Example 3.2**. The results presented in **Figure 5** show that GFP was delivered intracellularly to HeLa cells in the presence of the shuttle agent CM18-TAT.

3.6 GFP transduction by shuttle agents in HeLa cells: Dose responses (CM18-TAT-Cys, dCM18-TAT-Cys, GFP) and cell viability

HeLa cells were cultured and tested in the protein transduction assay described in **Examples 3.1-3.3**. Briefly, GFP recombinant protein was co-incubated with different concentrations of CM18-TAT-Cys or dimerized CM18-TAT-Cys (dCM18-TAT-Cys), and then exposed to HeLa cells for 1 hour. The results are shown in **Table 3.1** and **Figure 6**.

Table 3.1: Dose response (CM18-TAT) and cell viability, data from **Figure 6A** and **6B**

Shuttle	Cells	Concentration (μM)	Figure 6A Mean (%) (n=3)	Standard deviation	Figure 6B Cell viability (%) ($\pm$ St. Dev.; n=3)
CM18-TAT-Cys	HeLa	0	0.69	0.12	95 $\pm$ 4
	HeLa	0.5	8.67	0.96	88.4 $\pm$ 6
	HeLa	1	20.03	2.55	90 $\pm$ 6
	HeLa	3	31.06	5.28	91 $\pm$ 5
	HeLa	5	36.91	4.33	90 $\pm$ 7

Table 3.1 and **Figure 6A** show the results of flow cytometry analysis of the fluorescence intensity of HeLa cells transduced with GFP (5 μM) without or with 5, 3, 1, and 0.5 μM of CM18-TAT-Cys. Corresponding cellular toxicity data are presented in **Table 3.1** and in **Figure 6B**. These results suggest that the shuttle agent CM18-TAT-Cys increases the transduction efficiency of GFP in a dose-dependent manner.

Table 3.2: Dose response (GFP), data from **Figures 7A** and **7B**

Shuttle	Cells	Conc. of shuttle agent (μM)	Conc. of GFP (μM)	Mean (%) (n=3)	Standard deviation
Control	HeLa	0	10	0.93	0.08
CM18-TAT-Cys	HeLa	5	10	37.1	4.29
	HeLa	5	5	21.1	2.19
	HeLa	5	1	8.56	1.91

Control	HeLa	0	10	0.91	0.09
dCM18-TAT-Cys	HeLa	2.5	10	34.2	3.42
	HeLa	2.5	5	22.2	3.17
	HeLa	2.5	1	9.38	2.11

Table 3.2 and **Figure 7** show the results of flow cytometry analysis of the fluorescence intensity of HeLa cells transduced with different concentrations of GFP (1 to 10 μ M) without or with 5 μ M of CM18-TAT-Cys (**Figure 7A**) or 2.5 μ M dCM18-TAT-Cys (**Figure 7B**).

5

3.7 GFP transduction in HeLa cells: Dose responses of CM18-TAT-Cys and CM18-Penetratin-Cys, and dimers thereof

HeLa cells were cultured and tested in the protein transduction assay described in **Example 3.1**. Briefly, GFP recombinant protein (5 μ M) was co-incubated with different concentrations and combinations of CM18-TAT-Cys, CM18-Penetratin-Cys, and dimers of each (dCM18-TAT-Cys, dCM18-Penetratin-Cys), and then exposed to HeLa cells for 1 hour. The cells were subjected to flow cytometry analysis as described in **Example 3.3**. The results are shown in **Table 3.3** and **Figure 8**, as well as in **Table 3.4** and **Figure 9**.

10

Table 3.3: Data in **Figure 8**

No. in Figure 8	Shuttle agent	Cells	Concentration (μ M)	Mean (%) (n=3)	Standard deviation
Control ("ctrl")	No shuttle	HeLa	0	0.43	0.08
1	CM18-TAT-Cys	HeLa	0.5	8.75	0.63
2	dCM18-TAT-Cys	HeLa	0.5	8.86	1.03
3	CM18-Penetratin-Cys	HeLa	3	0.59	0.11
4	dCM18-Penetratin-Cys	HeLa	3	0.73	0.08
1 + 3	CM18-TAT-Cys + CM18-Penetratin-Cys	HeLa	0.5 3	19.52	2.18
2 + 3	dCM18-TAT-Cys + CM18-Penetratin-Cys	HeLa	0.5 3	22.44	3.29
1 + 4	CM18-TAT-Cys + dCM18-Penetratin-Cys	HeLa	0.5 3	18.73	1.55
2 + 4	dCM18-TAT-Cys + dCM18-Penetratin-Cys	HeLa	0.5 3	17.19	1.93

15

The results in **Table 3.3** and **Figure 8** show that the transduction efficiency of GFP is increased in HeLa cells using the shuttle agents CM18-TAT-Cys and dCM18-TAT-Cys (see bars "1" and "2" in **Figure 8**). Although no GFP intracellular delivery was observed using CM18-Penetratin-Cys or dCM18-Penetratin-Cys alone (see bars "3" or "4" in **Figure 8**), combination of CM18-TAT-Cys with CM18-Penetratin-Cys (monomer or dimer) improved GFP protein delivery (see four right-most bars in **Figure 8**).

20

Table 3.4: Data in Figure 9

No. in Figure 9	Shuttle	Cells	Concentration (μ M)	Mean (%) (n=3)	Standard deviation
Control ("ctrl")	No shuttle	HeLa	0	0.51	0.07
1	CM18-TAT-Cys	HeLa	1	20.19	2.19
2	dCM18-TAT-Cys	HeLa	1	18.43	1.89
3	CM18-Penetratin-Cys	HeLa	3	0.81	0.07
4	dCM18-Penetratin-Cys	HeLa	3	0.92	0.08
1 + 3	CM18-TAT-Cys + CM18-Penetratin-Cys	HeLa	1 3	30.19	3.44
2 + 3	dCM18-TAT-Cys + CM18-Penetratin-Cys	HeLa	1 3	22.36	2.46
1 + 4	CM18-TAT-Cys + dCM18-Penetratin-Cys	HeLa	1 3	26.47	2.25
2 + 4	dCM18-TAT-Cys + dCM18-Penetratin-Cys	HeLa	1 3	21.44	3.11

The results in **Table 3.4** and **Figure 9** show that the transduction efficiency of GFP is increased in HeLa cells using the shuttle agents CM18-TAT-Cys and dCM18-TAT-Cys (see bars "1" and "2" in **Figure 9**). Although no GFP intracellular delivery was observed using CM18-Penetratin-Cys or dCM18-Penetratin-Cys alone (see bars "3" or "4" in **Figure 9**), combination of CM18-TAT-Cys with CM18-Penetratin-Cys (monomer or dimer) improved GFP protein delivery (see four right-most bars in **Figure 9**).

3.8 GFP transduction by shuttle agents in HeLa cells: Controls

HeLa cells were cultured and tested in the protein transduction assay described in **Example 3.1**. Briefly, GFP recombinant protein (5 μ M) was co-incubated with 5 μ M of each of the following peptide(s): TAT-Cys; CM18; Penetratin-Cys; TAT-Cys + CM18; Penetratin-Cys + CM18; and CM18-TAT-Cys, and then exposed to HeLa cells for 1 hour. GFP fluorescence was visualized by bright field and fluorescence microscopy. The microscopy results (data not shown) showed that GFP was successfully delivered intracellularly using CM18-TAT-Cys. However, GFP was not successfully delivered intracellularly using single-domain peptides used alone (CM18, TAT-Cys, Penetratin-Cys) or together (CM18 + TAT-Cys, CM18 + Penetratin-Cys). These results are consistent with those presented in **Tables 2.1 and 2.2** with respect to the calcein endosome escape assays.

Example 4:

Peptide shuttle agents increase TAT-GFP transduction efficiency

The experiments in **Example 3** showed the ability of shuttle agents to deliver GFP intracellularly. The experiments presented in this example show that the shuttle agents can also increase the intracellular delivery of a GFP cargo protein that is fused to a CPD (TAT-GFP).

4.1 Construction and amino acid sequence of TAT-GFP

Construction was performed as described in **Example 3.4**, except that a TAT sequence was cloned between the 6x histidine tag and the GFP sequences. The 6x histidine tag, TAT, GFP and a stop codon (-) are separated by serine/glycine rich linkers. The recombinant TAT-GFP protein was purified as described in **Example 1.4**. The sequence of the TAT-GFP construct was:

```
MHHHHHHGGGSGGGSGGGSGGASTGTGRKKRRQRRRPPQGGGSGGGSGGGTGIRMVSKGEELFTGV
VPIILVELDGDVNGHKFSVSGEGEGDATYGLTLKFICTGKLPVPWPTLVTTLTLYGVQCFSRYPDH
MKQHDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDTLVNRIELKGIDFKEDGNILGHKLEY
NYNSHNVYIMADKQKNGIKVNFKIRHNIEDGSVQLADHYQONTPIGDGPVLLPDNHYLSTQSALSK
DPNEKRDMVLLFEFVTAAGITLGMDELYKGGSGGGSGGGSGWIRASSGGREIS-
```

[SEQ ID NO: 61]

(MW= 34.06 kDa ; pI=8.36)

TAT sequence is underlined

Serine/glycine rich linkers are in bold

4.2 TAT-GFP transduction by CM18-TAT-Cys in HeLa cells: Visualisation by fluorescence microscopy

HeLa cells were cultured and tested in the protein transduction assay described in **Example 3.1**. Briefly, TAT-GFP recombinant protein (5 μ M) was co-incubated with 3 μ M of CM18-TAT-Cys and then exposed to HeLa cells for 1 hour. Cells and GFP fluorescence were visualized by bright field and fluorescence microscopy (as described in **Example 3.2**) at 10x and 40x magnifications, and sample results are shown in **Figure 10**. The microscopy results revealed that in the absence of CM18-TAT-Cys, TAT-GFP shows a low intensity, endosomal distribution as reported in the literature. In contrast, TAT-GFP is delivered to the cytoplasm and to the nucleus in the presence of the shuttle agent CM18-TAT-Cys. Without being bound by theory, the TAT peptide itself may act as a nuclear localization signal (NLS), explaining the nuclear localization of TAT-GFP. These results show that CM18-TAT-Cys is able to increase TAT-GFP transduction efficiency and allow endosomally-trapped TAT-GFP to gain access to the cytoplasmic and nuclear compartments.

4.3 TAT-GFP transduction by CM18-TAT-Cys in HeLa cells: Dose responses and viability of cells transduced

HeLa cells were cultured and tested in the protein transduction assay described in **Example 3.1**. Briefly, TAT-GFP-Cys recombinant protein (5 μ M) was co-incubated with different concentrations of CM18-TAT-Cys (0, 0.5, 1, 3, or 5 μ M) and then exposed to HeLa cells for 1 hour. The cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 4.3** and **Figure 11A**. Corresponding cellular toxicity data are presented in **Figure 11B**.

Table 4.3: Data from Figure 11A and 11B

Shuttle agent	Cells	Concentration (μ M)	Figure 11A		Figure 11B
			Mean (%) (n=3)	Standard deviation	Cell viability (%) ($\pm$ St. Dev.; n= 3)
CM18-TAT-Cys	HeLa	0	11.79 ¹	1.16	100
	HeLa	0.5	10.19	1.94	84.36 $\pm$ 5
	HeLa	1	14.46	2.59	89.26 $\pm$ 5.26
	HeLa	3	28.12	3.27	93.18 $\pm$ 6.28
	HeLa	5	35.5 ²	3.59	95.14 $\pm$ 5.28

¹ The fluorescence was mostly endosomal, as confirmed by fluorescence microscopy.

² Fluorescence was more diffuse and also nuclear, as confirmed by fluorescence microscopy.

5

Example 5:

Peptide shuttle agents increase GFP-NLS transduction efficiency and nuclear localization

The experiments in **Examples 3 and 4** showed the ability of shuttle agents to deliver GFP and TAT-GFP intracellularly. The experiments presented in this example show that the shuttle agents can facilitate nuclear delivery of a GFP protein cargo fused to a nuclear localization signal (NLS).

10

5.1 Construction and amino acid sequence of GFP-NLS

Construction was performed as described in **Example 3.4**, except that an optimized NLS sequence was cloned between the GFP sequence and the stop codon (-). The NLS sequence is separated from the GFP sequence and the stop codon by two serine/glycine rich linkers. The recombinant GFP-NLS protein was purified as described in **Example 1.4**. The sequence of the GFP-NLS construct was:

15

```

MHHHHHHGGGSGGGSGGSGASTGIRMVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDATY
GKLTLLKFICTTGKLPVPWPTLVTTLTLYGVQCFSRYPDHMKQHDFFKSAMPEGYVQERTIFFKDDG
NYKTRAEVKFEGDTLVNRIELKGIDFKEDGNILGHKLEYNNSHNVYIMADKQKNGIKVNFKIRH
NIEDGSVQLADHYQNTPIGDGPVLLPDNHYLSTQSALSKDPNEKRDHMLLEFVTAAGITLGM
ELYKGGSGGGSGGGSGWIRASSGGRSSDDEATADSQHAAPPKKKRKVGGSGGGSGGGSGGGGRGTE
IS- [SEQ ID NO: 62]

```

20

(MW = 34.85 kDa; pI = 6.46)

NLS sequence is underlined

25

Serine/glycine rich linkers are in bold

5.2 Nuclear delivery of GFP-NLS by CM18-TAT-Cys in HeLa cells in 5 minutes: Visualisation by fluorescence microscopy

HeLa cells were cultured and tested in the protein transduction assay described in **Example 3.1**. Briefly, GFP-NLS recombinant protein (5 μ M) was co-incubated with 5 μ M of CM18-TAT-Cys, and then exposed to HeLa cells. GFP fluorescence was visualized by bright field and fluorescence microscopy after 5 minutes (as described in **Example 3.2**) at 10x, 20x and 40x magnifications, and sample results are shown in **Figure 12**. The microscopy

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results revealed that GFP-NLS is efficiently delivered to the nucleus in the presence of the shuttle agent CM18-TAT-Cys, after only 5 minutes of incubation.

5.3 GFP-NLS transduction by CM18-TAT-Cys in HeLa cells: Dose responses and viability of cells transduced

HeLa cells were cultured and tested in the protein transduction assay described in **Example 3.1**. GFP-NLS recombinant protein (5 μ M) was co-incubated with 0, 0.5, 1, 3, or 5 μ M of CM18-TAT-Cys, and then exposed to HeLa cells for 1 hour. The cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 5.1** and **Figure 13A**. Corresponding cellular toxicity data are presented in **Figure 13B**.

Table 5.1: Data from Figure 13A and 13B

Shuttle agent	Cells	Concentration (μ M)	Figure 13A		Figure 13B
			Mean (%) (n=3)	Standard deviation	Cell viability (%) ($\pm$ St. Dev.; n= 3)
CM18-TAT-Cys	HeLa	0	0.90	0.12	100
	HeLa	0.5	9.81	1.63	87.6 $\pm$ 4
	HeLa	1	18.42	2.47	93 $\pm$ 8
	HeLa	3	28.09	3.24	94 $\pm$ 5
	HeLa	5	32.26	4.79	93 $\pm$ 4

These results show that CM18-TAT-Cys is able to increase GFP-NLS transduction efficiency in HeLa cells in a dose-dependent manner.

5.4 GFP-NLS transduction by CM18-TAT-Cys, CM18-Penetratin-Cys, and dimers thereof in HeLa cells

HeLa cells were cultured and tested in the protein transduction assay described in **Example 3.1**. GFP-NLS recombinant protein (5 μ M) was co-incubated with different concentrations and combinations of CM18-TAT-Cys, CM18-Penetratin-Cys, and dimers of each (dCM18-TAT-Cys, dCM18-Penetratin-Cys), and then exposed to HeLa cells for 1 hour. The cells were subjected to flow cytometry analysis as described in **Example 3.3**. The results are shown in **Tables 5.2 and 5.3**, and in **Figures 14 and 15**.

Table 5.2: Data in Figure 14

No. in Figure 14	Shuttle agent	Cells	Concentration (μ M)	Mean (%) (n=3)	Standard deviation
ctrl	No shuttle	HeLa	0	0.41	0.10
1	CM18-TAT-Cys	HeLa	0.5	7.64	0.85
2	dCM18-TAT-Cys	HeLa	0.5	8.29	0.91
3	CM18-Penetratin-Cys	HeLa	3	0.43	0.08
4	dCM18-Penetratin-Cys	HeLa	3	0.85	0.07

1 + 3	CM18-TAT-Cys + CM18-Penetratin-Cys	HeLa	0.5 3	21.1	2.47
2 + 3	dCM18-TAT-Cys + CM18-Penetratin-Cys	HeLa	0.5 3	19.22	2.73
1 + 4	CM18-TAT-Cys + dCM18-Penetratin-Cys	HeLa	0.5 3	23.44	2.51
2 + 4	dCM18-TAT-Cys + dCM18-Penetratin-Cys	HeLa	0.5 3	19.47	2.16

Table 5.3: Data in Figure 15

No. in Figure 15	Shuttle agent	Cells	Concentration (μ M)	Mean (%) (n=3)	Standard deviation
ctrl	No shuttle	HeLa	0	0.44	0.12
1	CM18-TAT-Cys	HeLa	1	15.56	2.24
2	dCM18-TAT-Cys	HeLa	1	17.83	2.13
3	CM18-Penetratin-Cys	HeLa	3	0.68	0.05
4	dCM18-Penetratin-Cys	HeLa	3	0.84	0.07
1 + 3	CM18-TAT-Cys + CM18-Penetratin-Cys	HeLa	1 3	27.26	3.61
2 + 3	dCM18-TAT-Cys + CM18-Penetratin-Cys	HeLa	1 3	25.47	3.77
1 + 4	CM18-TAT-Cys + dCM18-Penetratin-Cys	HeLa	1 3	31.47	4.59
2 + 4	dCM18-TAT-Cys + dCM18-Penetratin-Cys	HeLa	1 3	28.74	2.93

The results in **Tables 5.2 and 5.3** and **Figures 14 and 15** show that the transduction efficiency of GFP-NLS is increased in HeLa cells using the shuttle agents CM18-TAT-Cys and dCM18-TAT-Cys (see bars "1" and "2" in **Figures 14 and 15**). Although no GFP-NLS intracellular delivery was observed using CM18-Penetratin-Cys or dCM18-Penetratin-Cys alone (see bars "3" and "4" in **Figures 14 and 15**), combination of CM18-TAT-Cys with CM18-Penetratin-Cys (monomer or dimer) improved GFP-NLS intracellular delivery (see four right-most bars in **Figures 14 and 15**).

5.5 GFP-NLS transduction by shuttle agents in HeLa cells: 5 min v. 1 h incubation; with or without FBS

HeLa cells were cultured and tested in the protein transduction assay described in **Example 3.1**. GFP-NLS recombinant protein (5 μ M) was co-incubated with either CM18-TAT-Cys (3.5 μ M) alone or with dCM18-Penetratin-Cys (1 μ M). Cells were incubated for 5 minutes or 1 hour in plain DMEM media ("DMEM") or DMEM media containing 10% FBS ("FBS"), before being subjected to flow cytometry analysis as described in **Example 3.3**. The results are shown in **Table 5.4**, and in **Figure 16**. Cells that were not treated with shuttle agent or GFP-NLS ("ctrl"), and cells that were treated with GFP-NLS without shuttle agent ("GFP-NLS 5 μ M") were used as controls.

Table 5.4: Data in Figure 16

Shuttle	No. in Fig. 16	Cells	Medium	Incubation time	Shuttle Conc. (μ M)	Mean (%) (n=3)	Standard deviation
No shuttle (Ctrl)	1	HeLa	DMEM	1h	0	0.59	0.09
GFP-NLS alone	2	HeLa	DMEM	1h	0	1.19	0.31
CM18-TAT-Cys	3	HeLa	DMEM	1 h	3.5	20.69	1.19
	4	HeLa	FBS	1 h	3.5	13.20	0.82
CM18-TAT-Cys	5	HeLa	DMEM	5 min	3.5	20.45	4.26
	6	HeLa	FBS	5 min	3.5	10.83	1.25
No shuttle (Ctrl)	1	HeLa	DMEM	1h	0	0.53	0.11
GFP-NLS alone	2	HeLa	DMEM	1h	0	1.25	0.40
CM18-TAT-Cys + dCM18-Penetratin-Cys	3	HeLa	DMEM	1 h	3.5 1	27.90	2.42
	4	HeLa	FBS	1 h	3.5 1	8.35	0.46
CM18-TAT-Cys + dCM18-Penetratin-Cys	5	HeLa	DMEM	5 min	3.5 1	24.10	2.76
	6	HeLa	FBS	5 min	3.5 1	5.02	0.72

The results in **Table 5.4** and **Figure 16** show that the addition of even a relatively low amount of the dimer dCM18-Penetratin-Cys (1 μ M; "dCM18pen") to the CM18-TAT-Cys monomer improved GFP-NLS transduction efficiency. Interestingly, intracellular GFP-NLS delivery was achieved in as little as 5 minutes of incubation, and delivery was still achievable (although reduced) in the presence of FBS.

5.6 GFP-NLS transduction by shuttle agents in THP-1 suspension cells

The ability of the shuttle agents to deliver GFP-NLS intracellularly was tested in THP-1 cells, which is an acute monocytic leukemia cell line that grows in suspension. THP-1 cells were cultured (see **Example 1**) and tested in the protein transduction assay described in **Example 3.1**. GFP-NLS recombinant protein (5 μ M) was co-incubated with or without 1 μ M CM18-TAT-Cys, and exposed to the THP-1 cells for 5 minutes, before being subjected to flow cytometry analysis as described in **Example 3.3**. The results are shown in **Table 5.5** and in **Figure 17A**. Corresponding cellular toxicity data are presented in **Figure 17B**.

Table 5.5: Data in Figure 17A and 17B

Shuttle	Cells	Shuttle Conc. (μ M)	Figure 17A Mean (%) (n=3)	Standard deviation	Figure 17B Cell viability (%) ($\pm$ St. Dev.; n= 3)
No shuttle (Ctrl)	THP-1	0	1.23	0.16	95 $\pm$ 4
GFP-NLS alone		0	2.49	0.37	96 $\pm$ 3
CM18-TAT-Cys		1	38.1	4.16	85 $\pm$ 6

The results in **Table 5.5** and **Figure 17** demonstrate the ability of the shuttle agents to deliver protein cargo intracellularly to a human monocytic cell line grown in suspension.

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Example 6:

Peptide shuttle agents increase transduction efficiency of an FITC-labeled anti-tubulin antibody

The experiments in **Examples 3-5** showed the ability of shuttle agents to increase the transduction efficiency of GFP, TAT-GFP, and GFP-NLS. The experiments presented in this example show that the shuttle agents can also deliver a larger protein cargo: an FITC-labeled anti-tubulin antibody. The FITC-labeled anti-tubulin antibody was purchased from (Abcam, ab64503) and has an estimated molecular weight of 150 KDa. The delivery and microscopy protocols are described in **Example 3**.

6.1 Transduction of a functional antibody by CM18-TAT-Cys in HeLa cells: Visualization by microscopy

FITC-labeled anti-tubulin antibody (0.5 μ M) was co-incubated with 5 μ M of CM18-TAT-Cys and exposed to HeLa cells for 1 hour. Antibody delivery was visualized by bright field (20x) and fluorescence microscopy (20x and 40x). As shown in **Figure 18**, fluorescent tubulin fibers in the cytoplasm were visualized, demonstrating the functionality of the antibody inside the cell.

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6.2 Transduction of a functional antibody by CM18-TAT-Cys, CM18-Penetratin-Cys, and dimers in HeLa cells: Flow cytometry

HeLa cells were cultured and tested in the protein transduction assay described in **Example 3.1**. FITC-labeled anti-tubulin antibody (0.5 μ M) was co-incubated with 3.5 μ M of CM18-TAT-Cys, CM18-Penetratin-Cys or dCM18-Penetratin-Cys, or a combination of 3.5 μ M of CM18-TAT-Cys and 0.5 μ M of dCM18-Penetratin-Cys, and exposed to HeLa cells for 1 hour. The cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 6.1** and **Figure 19A**. Corresponding cellular toxicity data are presented in **Figure 19B**.

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Table 6.1: Data from Figure 19A and 19B

Domains	Shuttle agent	Cells	Shuttle Conc. (μ M)	Figure 19A Mean (%) (n=3)	Standard deviation	Figure 19B Cell viability (%) ($\pm$ St. Dev.; n= 3)
-	No shuttle ("Ctrl")	HeLa	0	0.9	0.06	98 $\pm$ 1.0
-	Antibody alone ("antibody")	HeLa	0	2.66	0.61	96 $\pm$ 3.4
ELD-CPD	CM18-TAT-Cys	HeLa	3.5	36.56	4.06	95 $\pm$ 4.06
	CM18-Penetratin-Cys	HeLa	3.5	53.05	9.5	73 $\pm$ 9.5
ELD-CPD dimer	dCM18-Penetratin-Cys	HeLa	3.5	50.23	9.12	74 $\pm$ 9.0
ELD-CPD + ELD-CPD dimer	CM18-TAT-Cys + dCM18-Penetratin-Cys	HeLa	3.5 0.5	47.19	8.5	93 $\pm$ 8.5

The results in Table 6.1 and Figures 18 and 19 show that both CM18-TAT-Cys and CM18-Penetratin-Cys facilitate intracellular delivery of an FITC-labeled anti-tubulin antibody. In contrast to the results with GFP, TAT-GFP, and GFP-NLS in Examples 3-5, CM18-Penetratin-Cys was able to deliver the antibody cargo intracellularly when used alone (without CM18-TAT-Cys). However, combination of CM18-TAT-Cys and dCM18-Penetratin-Cys allowed for higher intracellular delivery as compared with CM18-TAT-Cys alone, and with less cell toxicity as compared to CM18-Penetratin-Cys and dCM18-Penetratin-Cys (see Figure 19A and 19B).

Example 7:

CM18-TAT-Cys enables intracellular plasmid DNA delivery but poor plasmid expression

The ability of the CM18-TAT-Cys shuttle agent to deliver plasmid DNA intracellularly was tested in this example on HEK293A cells using a plasmid encoding GFP.

7.1 Transfection assay in HEK293A cells

One day before the transfection assay was performed, mammalian cells (HEK293A) in exponential growth phase were harvested and plated in a 24-well plate (50,000 cells per well). The cells were incubated overnight in appropriate growth media containing FBS. The next day, in separate sterile 1.5 mL tubes, pEGFP labeled with a Cy5™ fluorochrome was mixed for 10 min at 37°C with CM18-TAT-Cys (0.05, 0.5, or 5 μ M) in fresh PBS at a final 100 μ L volume. The media in wells was removed and the cells were quickly washed three times with PBS and 500 μ L of warm media without FBS was added. The pEGFP and CM18-TAT-Cys solution was added to the cells and incubated at 37°C for 4 hours. After the incubation, cells were washed with PBS and fresh media containing FBS was added. Cells were incubated at 37°C before being subjected to flow cytometry analysis as described in Example 3.

7.2 Plasmid DNA delivery with CM18-TAT-Cys

Plasmid DNA (pEGFP) was labeled with a Cy5TM dye following the manufacturer's instructions (Mirus Bio LLC). Cy5TM Moiety did not influence transfection efficiency when compared to unlabelled plasmid using standard transfection protocol (data not shown). Flow cytometry analysis allowed quantification of Cy5TM emission, corresponding to DNA intracellular delivery, and GFP emission, corresponding to successful nuclear delivery, DNA transcription and protein expression. The results are shown in **Table 7.1** and in **Figure 20**.

Table 7.1: Data from Figure 20

Sample	DNA (ng)	Cy5 TM fluorescence		GFP expression	
		Mean Cy5 TM signal (n=3)	Standard deviation	Mean (% of cells with GFP signal; n=3)	Standard deviation
pEGFP-Cy5 alone	500	914	0	0.0%	n/a
CM18-TAT-Cys, 0.05 μ M	500	1450	120	0.0%	n/a
CM18-TAT-Cys, 0.5 μ M	500	8362	294	0.0%	n/a
CM18-TAT-Cys, 5 μ M	500	140 497	3977	0.1%	n/a

The results shown in **Table 7.1** and in **Figure 20** show that CM18-TAT-Cys was able to increase the intracellular delivery the plasmid DNA when used at 0.05, 0.5 and 5 μ M concentrations, as compared to cell incubated with DNA alone ("pEGFP-Cy5"). However, no expression of GFP was detected in the cells, which suggests that very little of the plasmid DNA gained access to the cytoplasmic compartment, allowing nuclear localization. Without being bound by theory, it is possible that the plasmid DNA was massively sequestered in endosomes, preventing escape to the cytoplasmic compartment. Salomone et al., 2013 reported the use of a CM18-TAT11 hybrid peptide to deliver plasmid DNA intracellularly. They used the luciferase enzyme reporter assay to assess transfection efficiency, which may not be ideal for quantifying the efficiency of cytoplasmic/nuclear delivery, as the proportion of plasmid DNA that is successfully released from endosomes and delivered to the nucleus may be overestimated due to the potent activity of the luciferase enzyme. In this regard, the authors of Salomone et al., 2013 even noted that the expression of luciferase occurs together with a massive entrapment of (naked) DNA molecules into vesicles, which is consistent with the results shown in **Table 7.1** and in **Figure 20**.

Example 8:

Addition of a histidine-rich domain to shuttle agents further improves GFP-NLS transduction efficiency

8.1 GFP-NLS transduction by His-CM18-TAT-Cys in HeLa cells: Visualization by microscopy

GFP-NLS (5 μ M; see **Example 5**) was co-incubated with 5 μ M of CM18-TAT-Cys or His-CM18-TAT and exposed to HeLa cells for 1 hour. Nuclear fluorescence of intracellularly delivered GFP-NLS was confirmed by fluorescence microscopy (data not shown), indicating successful delivery of GFP-NLS to the nucleus.

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8.2 GFP-NLS transduction by His-CM18-TAT in HeLa cells: Flow cytometry

HeLa cells were cultured and tested in the protein transduction assay described in **Example 3.1**. GFP-NLS (5 μ M) was co-incubated with 0, 1, 3, or 5 μ M of CM18-TAT-Cys or His-CM18-TAT, and exposed to HeLa cells for 1 hour. The cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 8.1** and **Figure 21A**. Corresponding cellular toxicity data are presented in **Figure 21B**.

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Table 8.1: Data from Figure 21A and 21B

Shuttle agent	Cells	Shuttle Conc. (μ M)	Figure 21A		Figure 21B
			Mean (%) cell with GFP signal (n=3)	Standard deviation	Cell viability (%) ($\pm$ St. Dev.; n= 3)
Ctrl (no shuttle, no GFP-NLS)	HeLa	0	0.63	0.10	96 $\pm$ 3.17
GFP-NLS alone		0	0.93	0.26	97 $\pm$ 2.05
CM18-TAT-Cys		5	20.54	3.51	81 $\pm$ 6.34
		3	15.66	2.18	89 $\pm$ 5.37
		1	8.64	1.11	94 $\pm$ 4.28
Ctrl (no shuttle, no GFP-NLS)	HeLa	0	0.51	0.28	95 $\pm$ 4.19
GFP-NLS alone		0	1.07	0.42	96 $\pm$ 3.16
His-CM18-TAT		5	41.38	4.59	86 $\pm$ 4.59
		3	29.58	3.61	91 $\pm$ 5.18
		1	8.45	1.83	95 $\pm$ 3.55

Strikingly, the results in **Table 8.1** and in **Figure 21** show that His-CM18-TAT was able to increase GFP-NLS protein transduction efficiency by about 2-fold at 3 μ M and 5 μ M concentrations, as compared to CM18-TAT-Cys. These results suggest that adding a histidine-rich domain to a shuttle agent comprising an ELD and CPD, may significantly increase its polypeptide cargo transduction efficiency. Alternatively or in parallel, combining the shuttle agents with a further independent synthetic peptide containing a histidine-rich domain fused to a CPD (but lacking an ELD) may provide a similar advantage for protein transduction, with the added advantage of allowing the concentration of the histidine-rich domain to be varied or controlled independently from the concentration of the shuttle agent. Without being bound by theory, the histidine-rich domain may act as a proton sponge in the endosome, providing another mechanism of endosomal membrane destabilization.

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Example 9:**His-CM18-PTD4 increases transduction efficiency and nuclear delivery of GFP-NLS, mCherryTM-NLS and FITC-labeled anti-tubulin antibody**5 **9.1 Protein transduction protocols****Protocol A: Protein transduction assay for delivery in cell culture medium**

One day before the transduction assay was performed, cells in exponential growth phase were harvested and plated in a 96-well plate (20,000 cells per well). The cells were incubated overnight in appropriate growth media containing FBS (see **Example 1**). The next day, in separate sterile 1.5-mL tubes, cargo protein at the desired concentration was pre-mixed (pre-incubated) for 10 min at 37°C with the desired concentration of shuttle agents in 50 µL of fresh serum-free medium (unless otherwise specified). The media in wells was removed and the cells were washed one to three times (depending on the type of cells used) with PBS previously warmed at 37°C. The cells were incubated with the cargo protein/shuttle agent mixture at 37°C for the desired length of time. After the incubation, the cells were washed three times with PBS and/or heparin (0.5 mg/mL) previously warmed at 37°C. The washes with heparin were used for human THP-1 blood cells to avoid undesired cell membrane-bound protein background in subsequent analyses (microscopy and flow cytometry). The cells were finally incubated in 50 µL of fresh medium with serum at 37°C before analysis.

20 **Protocol B: Protein transduction assay for adherent cells in PBS**

One day before the transduction assay was performed, cells in exponential growth phase were harvested and plated in a 96-well plate (20,000 cells per well). The cells were incubated overnight in appropriate growth media containing serum (see **Example 1**). The next day, in separate sterile 1.5-mL tubes, shuttle agents were diluted in sterile distilled water at room temperature (if the cargo is or comprised a nucleic acid, nuclease-free water was used). Cargo protein(s) were then added to the shuttle agents and, if necessary, sterile PBS was added to obtain the desired concentrations of shuttle agent and cargo in a sufficient final volume to cover the cells (e.g., 10 to 100 µL per well for a 96-well plate). The shuttle agent/cargo mixture was then immediately used for experiments. At least three controls were included for each experiment, including: (1) shuttle agent alone (e.g., at highest concentration tested); (2) cargo alone; and (3) without any cargo or shuttle agent. The media in wells was removed, cells were washed once with PBS previously warmed at 37°C, and the shuttle agent/cargo mixture was then added to cover all cells for the desired length of time. The shuttle agent/cargo mixture in wells was removed, the cells were washed once with PBS, and fresh complete medium was added. Before analysis, the cells were washed once with PBS and fresh complete medium was added.

Protocol C: Protein transduction assay for suspension cells in PBS

One day before the transduction assay was performed, suspension cells in exponential growth phase were harvested and plated in a 96-well plate (20,000 cells per well). The cells were incubated overnight in appropriate growth media containing serum (see **Example 1**). The next day, in separate sterile 1.5-mL tubes, shuttle agents were diluted in sterile distilled water at room temperature (if the cargo is or comprised a nucleic acid, nuclease-free water was used). Cargo protein(s) were then added to the shuttle agents and, if necessary, sterile PBS or cell culture medium (serum-free) was added to obtain the desired concentrations of shuttle agent and cargo in a sufficient final volume to resuspend the cells (e.g., 10 to 100 μ L per well in a 96-well plate). The shuttle agent/cargo mixture was then immediately used for experiments. At least three controls were included for each experiment, including: (1) shuttle agent alone (e.g., at highest concentration tested); (2) cargo alone; and (3) without any cargo or shuttle agent. The cells were centrifuged for 2 minutes at 400g, the medium was then removed and the cells were resuspended in PBS previously warmed at 37°C. The cells were centrifuged again 2 minutes at 400g, the PBS removed, and the cells were resuspended in the shuttle agent/cargo mixture. After the desired incubation time, 100 μ L of complete medium was added directly on the cells. Cells were centrifuged for 2 minutes at 400g and the medium was removed. The pellet was resuspended and washed in 200 μ L of PBS previously warmed at 37°C. After another centrifugation, the PBS was removed and the cells were resuspended in 100 μ L of complete medium. The last two steps were repeated one time before analysis.

9.2 GFP-NLS transduction by His-CM18-PTD4 in HeLa cells using Protocol A or B: Flow cytometry

To compare the effects of different protocols on shuttle agent transduction efficiency, HeLa cells were cultured and tested in the protein transduction assays using Protocol A or B as described in **Example 9.1**. Briefly, GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 10 μ M of His-CM18-PTD4 and exposed to HeLa cells for 1 hour using Protocol A, or was co-incubated with 35 μ M of His-CM18-PTD4 and exposed to HeLa cells for 10 seconds using Protocol B. The cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 9.1** and **Figure 22A**. ("Pos cells (%)") is the percentage of cells emanating a GFP signal).

Table 9.1: Comparison of Protein Transduction Protocols A and B: Data from **Figure 22A**

Protocol	Shuttle	Cells	Conc. of shuttle (μ M)	Conc. of GFP-NLS (μ M)	Mean % cells with GFP signal ($\pm$ St. Dev.; n=3)	Cell viability (%) ($\pm$ St. Dev.; n=3)
B	None ("Ctrl")	HeLa	0	5	0.53 $\pm$ 0.07	100
A	His-CM18-PTD4	HeLa	10	5	25.4 $\pm$ 3.6	96.4 $\pm$ 2.7
B	His-CM18-PTD4	HeLa	35	5	78.3 $\pm$ 5.3	94.6 $\pm$ 0.4

The above results show that higher protein transduction efficiency for the cargo GFP-NLS using the shuttle agent His-CM18-PTD4 was obtained using Protocol B, as compared to Protocol A.

9.3 GFP-NLS transduction by His-CM18-PTD4 in HeLa cells using Protocol B: Flow cytometry

A dose response experiment was performed to evaluate the effect of His-CM18-PTD4 concentration on protein transduction efficiency. HeLa cells were cultured and tested in the protein transduction assay described in Protocol B of **Example 9.1**. Briefly, GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 0, 50, 35, 25, or 10 μ M of His-CM18-PTD4, and then exposed to HeLa cells for 10 seconds. The cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 9.2** and **Figure 22B**.

Table 9.2: Dose response of shuttle agent using Protocol B: Data from **Figure 22B**

Protocol	Shuttle	Cells	Conc. of shuttle (μM)	Conc. of GFP-NLS (μM)	Mean % cells with GFP signal (± St. Dev.; n=3)	Cell viability (%) (± St. Dev.; n=3)
B	None (“Ctrl”)	HeLa	0	5	0.13 ± 0.1	100 ± 0
	His-CM18-PTD4		50	5	73.2 ± 5.2	69.2 ± 2.7
			35	5	77.7 ± 7.8	79.6 ± 5.9
			25	5	62.1 ± 6.1	95.3 ± 3.7
			10	5	25.3 ± 3.6	96.3 ± 2.3

The above results show that His-CM18-PTD4 is able to increase GFP-NLS transduction efficiency in HeLa cells in a dose-dependent manner.

9.4 GFP-NLS transduction by His-CM18-PTD4 in HeLa cells using Protocol B: Visualization by microscopy

GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 35 μ M of His-CM18-PTD4, and then exposed to HeLa cells for 10 seconds using Protocol B as described in **Example 9.1**. The cells were then subjected to fluorescence microscopy analysis as described in **Examples 3.2 and 3.2a**.

For the sample results shown in **Figures 23 and 24**, GFP fluorescence of the HeLa cells was immediately visualized by bright field and fluorescence microscopy at 4x, 20x and 40x magnifications after the final washing step.

In **Figure 23**, the upper panels in A, B and C show nuclei labelling (DAPI) at 4x, 20x and 40x magnifications, respectively, while the lower panels show corresponding GFP-NLS fluorescence. In panel C, white triangle windows indicate examples of areas of co-labelling between nuclei (DAPI) and GFP-NLS signals. In panel D, the upper and bottom panels show sample bright field images of the HeLa cells, and the middle panel shows the results of a corresponding FACS analysis (performed as described in **Example 3.3**), which indicates the percentage of cells in a 96-plate with a GFP signal. No significant GFP fluorescence was observed in negative control samples (i.e., cells exposed to GFP-NLS without any shuttle agent; data not shown).

Figure 24 shows bright field (panel A) and fluorescent images (panel B). The inset in panel B shows the results of a corresponding FACS analysis (performed as described in **Example 3.3**), which indicates the percentage

of cells in a 96-plate well with a GFP signal. No significant GFP fluorescence was observed in negative control samples (i.e., cells exposed to GFP-NLS without any shuttle agent; data not shown).

For the sample results shown in **Figure 25**, the HeLa cells were fixed, permeabilized and subjected to immuno-labelling as described in **Example 3.2a** before visualization by fluorescence microscopy as described in **Example 3.2**. GFP-NLS was labelled using a primary mouse monoclonal anti-GFP antibody (Feldan, #A017) and a secondary goat anti-mouse AlexaTM-594 antibody (Abcam #150116). The upper panels in **Figure 25** show nuclei labelling (DAPI), and the lower panels show corresponding labelling for GFP-NLS. Panels A and B show sample images at 20x and 40x magnifications, respectively. White triangle windows indicate examples of areas of co-labelling between nuclei and GFP-NLS. No significant GFP-NLS labelling was observed in negative control samples (i.e., cells exposed to GFP-NLS without any shuttle agent; data not shown).

Figure 26 shows sample images captured with confocal microscopy at 63x magnification of living cells. Panel A of **Figure 26** shows a bright field image, while panel B shows the corresponding fluorescent GFP-NLS. Panel C is an overlay between the images in panels A and B. No significant GFP-NLS fluorescence was observed in negative control samples (i.e., cells exposed to GFP-NLS without any shuttle agent; data not shown).

9.4a FITC-labeled anti-tubulin antibody transduction by His-CM18-PTD4 in HeLa cells using Protocol B: Visualization by microscopy

FITC-labeled anti-tubulin antibody (0.5 μ M; Abcam, ab64503) was co-incubated with 50 μ M of His-CM18-PTD4, and then exposed to HeLa cells for 10 seconds using Protocol B as described in **Example 9.1**. The cells were then subjected to fluorescence microscopy analysis as described in **Examples 3.2 and 3.2a**, wherein the FITC fluorescence of the anti-tubulin antibody in the HeLa cells was immediately visualized by bright field and fluorescence microscopy at 20x magnification after the final washing step. Sample results are shown in **Figure 24C and 24D**. No significant FITC fluorescence was observed in negative control samples (i.e., cells exposed to the FITC-labeled anti-tubulin antibody without any shuttle agent; data not shown).

Overall, the results in **Examples 9.4 and 9.4a** show that GFP-NLS and FITC-labeled anti-tubulin antibody cargos are successfully transduced and delivered to the nucleus and/or the cytosol of HeLa cells in the presence of the shuttle agent His-CM18-PTD4.

9.5 GFP-NLS kinetic transduction by His-CM18-PTD4 in HeLa cells: Visualization by microscopy

GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 50 μ M of His-CM18-PTD4, and then exposed to HeLa cells for 10 seconds using Protocol B as described in **Example 9.1**. After a washing step, the GFP fluorescence of the HeLa cells was immediately visualized by fluorescence microscopy (**Example 3.2**) at 20x magnification after different intervals of time. Typical results are shown in **Figure 27**, in which fluorescence microscopy images were captured after 45, 75, 100, and 120 seconds (see panels A, B, C and D, respectively).

As shown in **Figure 27A**, diffuse cellular GFP fluorescence was generally observed after 45 seconds, with areas of lower GFP fluorescence in the nucleus in many cells. These results suggest predominantly cytoplasmic and low nuclear distribution of the GFP-NLS delivered intracellularly via the shuttle agent after 45 seconds. **Figures 27B-27D** show the gradual redistribution of GFP fluorescence to the cell nuclei at 75 seconds (panel B), 100 seconds (panel C), and 120 seconds (panel D) following exposure to the His-CM18-PTD4 shuttle agent and GFP-NLS cargo. No significant cellular GFP fluorescence was observed in negative control samples (i.e., cells exposed to GFP-NLS without any shuttle agent; data not shown).

The results in **Example 9.5** show that GFP-NLS is successfully delivered to the nucleus of HeLa cells in the presence of the shuttle agent His-CM18-PTD4 by 2 minutes.

9.6 GFP-NLS and mCherryTM-NLS co-transduction by His-CM18-PTD4 in HeLa cells: Visualization by microscopy

mCherryTM-NLS recombinant protein was constructed, expressed and purified from a bacterial expression system as described in **Example 1.4**. The sequence of the mCherryTM-NLS recombinant protein was:

```
MHHHHHHGGGGSGGGSGGASTGIRMVSKCEDNMAIIKEFMRFKVHMEGSVNGHEFEIEGEG
EGRPYEGTQTAKLKVTKGGPLPFAWDILSPQFMYGSKAYVKHPADIPDYLKLSFPEGFKWERV
MNFEDGGVVTVTQDSSLQDGEFIYKVKLRGTNFPDSDGQVMQKKTMGWEASSERMPEDGALKG
EIKQRLKLKDGGHYDAEVKTTYKAKKPVQLPGAYNVNICKLDITSHNEDYTIVEQYERAEGRHS
TGGMDELYKGGSGGGSGGGSGWIRASSGGRSSDDEATADSQHAAPPKKKRKVGGSGGGSGGGG
GGGRGTEIS [SEQ ID NO: 73]
```

(MW = 34.71 kDa; pI = 6.68)

NLS sequence is underlined

Serine/glycine rich linkers are in bold

GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) and mCherryTM-NLS recombinant protein (5 μ M) were co-incubated together with 35 μ M of His-CM18-PTD4, and then exposed to HeLa cells for 10 seconds using Protocol B as described in **Example 9.1**. After washing steps, the cells were immediately visualized by bright field and fluorescence microscopy at 20x magnifications as described in **Example 3.2**. Sample results are shown in **Figure 28**, in which corresponding images showing bright field (panel A), DAPI fluorescence (panel B), GFP-NLS fluorescence (panel C), and mCherryTM-NLS fluorescence (panel D) are shown. White triangle windows indicate examples of areas of co-labelling between GFP-NLS and mCherryTM fluorescence signals in cell nuclei. No significant cellular GFP or mCherryTM fluorescence was observed in negative control samples (i.e., cells exposed to GFP-NLS or mCherryTM without any shuttle agent; data not shown).

These results show that GFP-NLS and mCherryTM-NLS are successfully delivered together to the nucleus in HeLa cells in the presence of the shuttle agent His-CM18-PTD4.

9.7 GFP-NLS transduction by His-CM18-PTD4 in THP-1 suspension cells: Flow cytometry

The ability of the His-CM18-PTD4 to deliver GFP-NLS in the nuclei of suspension cells was tested using THP-1 cells. THP-1 cells were cultured and tested in the protein transduction assays using Protocols A and C as described in **Example 9.1**. GFP-NLS (5 μ M; see **Example 5.1**) was co-incubated with 1 μ M of His-CM18-PTD4 and exposed to THP-1 cells for 1 hour (Protocol A), or was co-incubated with 5 μ M of His-CM18-PTD4 and exposed to THP-1 cells for 15 seconds (Protocol C). The cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 9.3** and in **Figure 31**.

Table 9.3: Data from **Figure 31**

Protocol	Shuttle	Cells	Conc. of shuttle (μM)	Conc. of GFP-NLS (μM)	Mean % cells with GFP signal (± St. Dev.; n=3)	Cell viability (%) (± St. Dev.; n=3)
C	No shuttle ("Ctrl")	THP-1	0	5	0.2 ± 0.03	99.1 ± 0.7
A	His-CM18-PTD4		1	5	14.2 ± 2.2	96.9 ± 3.6
C	His-CM18-PTD4		0.5	5	34.9 ± 3.8	82.1 ± 2.7
			5	5	64.1 ± 1.6	64.0 ± 4.1

9.8 GFP-NLS transduction by His-CM18-PTD4 in THP-1 cells: Visualization by microscopy

GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 5 μ M of His-CM18-PTD4, and then exposed to THP-1 cells for 15 seconds using Protocol C as described in **Example 9.1**. The cells were subjected to microscopy visualization as described in **Example 3.2**.

For the sample results shown in **Figure 32**, GFP fluorescence of the HeLa cells was immediately visualized by bright field (upper panels) and fluorescence (lower panels) microscopy at 4x, 10x and 40x magnifications (panels A-C, respectively) after the final washing step. White triangle windows in panel C indicate examples of areas of co-labelling between bright field and fluorescence images. Panel D shows typical results of a corresponding FACS analysis (performed as described in **Example 3.3**), which indicates the percentage of cells in a 96-plate well with a GFP signal. Additional results are shown in **Figure 33**, in which panels A and B show bright field images, and panels C and D show corresponding fluorescence images. White triangle windows indicate examples of areas of co-labelling between panels A and C, as well as panels B and D. The right-most panel shows typical results of a corresponding FACS analysis (performed as described in **Example 3.3**), which indicates the percentage of cells in a 96-plate well with a GFP signal.

No significant cellular GFP fluorescence was observed in negative control samples (i.e., cells exposed to GFP-NLS without any shuttle agent; data not shown).

The results in this example show that GFP-NLS is successfully delivered intracellularly in THP-1 cells in the presence of the shuttle agent His-CM18-PTD4.

Example 10:

Different multi-domain shuttle agents, but not single-domain peptides, successfully transduce GFP-NLS in HeLa and THP-1 cells

5 **10.1 GFP-NLS transduction by different shuttle agents in HeLa cells: Flow cytometry**

HeLa cells were cultured and tested in the protein transduction assays using Protocol B as described in **Example 9.1**. Briefly, GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 50 μ M of different shuttle agents and exposed to the HeLa cells for 10 seconds. The cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 10.1** and **Figure 29A**. "Pos cells (%)" is the mean percentages of all cells that emanate a GFP signal. The negative control ("Ctrl") corresponds to cells that were

10 incubated with GFP-NLS recombinant protein (5 μ M) without any shuttle agent.

Table 10.1: Data from Figure 29A

Protocol	Shuttle agent	Cells	Conc. of shuttle (μM)	Conc. of GFP-NLS (μM)	Mean % cells with GFP signal (± St. Dev.; n=3)	Cell viability (%) (± St. Dev.; n=3)
B	No shuttle (“ctrl”)	HeLa	0	5	0	100
	His-CM18-TAT	HeLa	50		55.5 ± 3.6	35.2 ± 5.7
	His-CM18-Transportan (TPT)	HeLa			33.2 ± 2.8	41.3 ± 3.3
	TAT-KALA	HeLa			56.3 ± 3.6	95.6 ± 4.3
	His-CM18-PTD4	HeLa			68 ± 2.2	92 ± 3.6
	His-CM18-9Arg	HeLa			57.2 ± 3.9	45.8 ± 5.4
	TAT-CM18	HeLa			39.4 ± 3.9	23.5 ± 1.1
	His-C(LLKK) ₃ C-PTD4	HeLa			76 ± 3.8	95 ± 2.7
	His-LAH4-PTD4	HeLa			63 ± 1.64	98 ± 1.5
	PTD4-KALA	HeLa			73.4 ± 4.12	91.4 ± 3.67

15 **10.2 GFP-NLS transduction by different shuttle agents with varying incubation times in HeLa cells: Flow cytometry**

HeLa cells were cultured and tested in the protein transduction assays using Protocol B as described in **Example 9.1**. Briefly, GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 10 μ M of TAT-KALA, His-CM18-PTD4, or His-C(LLKK)₃C-PTD4 for 1, 2, or 5 minutes. After the final washing step, the cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 10.2** and **Figure 29B**.

20 "Pos cells (%)" is the mean percentages of all cells that emanate a GFP signal. The negative control ("Ctrl") corresponds to cells that were incubated with GFP-NLS recombinant protein (5 μ M) without any shuttle agent.

Table 10.2: Data from Figure 29B

Protocol	Shuttle agent	Cells	Conc. of shuttle (μ M)	Incubation time	Mean % cells with GFP signal ($\pm$ St. Dev.; n=3)	Cell viability (%) ($\pm$ St. Dev.; n=3)
-	No shuttle ("Ctrl")	HeLa	0	5 min.	$0 \pm n/a$	97.5 ± 1.7
B	TAT-KALA	HeLa	10	1 min.	83.7 ± 3.5	93.5 ± 2.7
				2 min.	86.2 ± 4.3	92.1 ± 3.1
				5 min.	68.1 ± 3.0	86 ± 4.4
	His-CM18-PTD4	HeLa	10	1 min.	50.6 ± 3.5	97.6 ± 2.7
				2 min.	74 ± 3.3	80.9 ± 3.2
				5 min.	82.7 ± 5.0	66.2 ± 4.4
	His-C(LLKK) ₃ C-PTD4	HeLa	10	1 min.	51.1 ± 3.5	99.5 ± 2.7
				2 min.	77.8 ± 4.3	94.3 ± 3.2
				5 min.	86.4 ± 4.0	80.8 ± 4.4

10.3 GFP-NLS transduction by TAT-KALA, His-CM18-PTD4 and His-C(LLKK)₃C-PTD4 with varying incubation times in HeLa cells: Flow cytometry

5 HeLa cells were cultured and tested in the protein transduction assays using Protocol C as described in **Example 9.1**. Briefly, GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 5 μ M of TAT-KALA, His-CM18-PTD4, or His-C(LLKK)₃C-PTD4 for 1, 2, or 5 minutes. After the final washing step, the cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 10.3** and **Figure 29C**. The negative control ("Ctrl") corresponds to cells that were incubated with GFP-NLS recombinant protein (5 μ M)

10 without any shuttle agent.

Table 10.3: Data from Figure 29C

Protocol	Shuttle agent	Cells	Conc. of shuttle (μ M)	Incubation time	Relative fluorescence intensity (FL1-A) (n=3)	St. Dev.
	No shuttle ("Ctrl")		0	5 min.	8903	501
C	TAT-KALA	HeLa	10	1 min.	216 367	13 863.48
				2 min.	506 158	14 536.28
				5 min.	78 010	2 463.96
	His-CM18-PTD4	HeLa	10	1 min.	524 151	12 366.48
				2 min.	755 624	26 933.16
				5 min.	173 930	15 567.33
	His-C(LLKK) ₃ C-PTD4	HeLa	10	1 min.	208 968	23 669.19
				2 min.	262 411.5	19 836.84
				5 min.	129 890	16 693.29

10.4 GFP-NLS transduction by different shuttle agents in HeLa cells: Flow cytometry

HeLa cells were cultured and tested in the protein transduction assays using Protocol B as described in **Example 9.1**. Briefly, GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 50 μ M of different shuttle agents (see **Table 1.3** for amino acid sequences and properties) and exposed to the HeLa cells for 10 seconds. The cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Tables 10.3a & 10.3b** and **Figure 29E & 29F**. "Pos cells (%)" is the mean percentages of all cells that emanate a GFP signal. The negative control ("Ctrl") corresponds to cells that were incubated with GFP-NLS recombinant protein (5 μ M) without any shuttle agent.

Table 10.3a: Data from Figure 29E

Domain structure	Shuttle agent	Conc. of shuttle (μ M)	Conc. of GFP-NLS (μ M)	Mean % cells with GFP signal ($\pm$ St. Dev.; n=3)	Cell viability (%) ($\pm$ St. Dev.; n=3)
-	No shuttle ("Ctrl")	0	5	0	100
ELD-CPD	VSVG-PTD4	50	5	3.5 $\pm$ 1.1	100
	EB1-PTD4			75.8 $\pm$ 8.26	39 $\pm$ 5.6
	JST-PTD4			0.84 $\pm$ 0.69	98.9 $\pm$ 0.57
His-ELD-CPD	His-C(LLKK) ₃ C-PTD4	50	5	76 $\pm$ 3.8	95 $\pm$ 2.7
	His-LAH4-PTD4			63 $\pm$ 1.64	98 $\pm$ 1.5
	His-CM18-PTD4			68 $\pm$ 2.2	92 $\pm$ 3.6
	His-CM18-TAT			55.5 $\pm$ 3.6	35.2 $\pm$ 5.7
	His-CM18-TAT-Cys*			49.3 $\pm$ 4.1	41.4 $\pm$ 3.91
	His-CM18-9Arg			57.2 $\pm$ 3.93	45.8 $\pm$ 3.53
	His-CM18-Transportan (TPT)			33.2 $\pm$ 2.82	41.3 $\pm$ 3.29

* Not shown in Figure 29E

Table 10.3b: Data from Figure 29F

Domain structure	Shuttle agent	Conc. of shuttle (μ M)	Conc. of GFP-NLS (μ M)	Mean % cells with GFP signal ($\pm$ St. Dev.; n=3)	Cell viability (%) ($\pm$ St. Dev.; n=3)
-	No shuttle ("Ctrl")	0	5	0	100
CPD-ELD	TAT-CM18	50	5	39.4 $\pm$ 3.9	23.5 $\pm$ 1.1
	TAT-KALA			56.3 $\pm$ 3.6	95.6 $\pm$ 4.3
	PTD4-KALA			73.4 $\pm$ 4.12	91.4 $\pm$ 3.67
	9Arg-KALA			7.8 $\pm$ 1.53	62.8 $\pm$ 5.11
	Pep1-KALA			17.2 $\pm$ 3.07	94.7 $\pm$ 3.77
	Xentry-KALA			19.4 $\pm$ 1.01	98.3 $\pm$ 0.64
	SynB3-KALA			14.3 $\pm$ 2.37	91.1 $\pm$ 0.82

- HeLa cells were cultured and tested in the protein transduction assays using Protocol B as described in **Example 9.1**. Briefly, GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 10 μ M of TAT-KALA, His-CM18-PTD4, or His-C(LLKK)₃C-PTD4 for 1, 2, or 5 minutes. After the final washing step, the cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Tables 10.3c & 10.3b** and **Figure 29G and 29H**. "Pos cells (%)" is the mean percentages of all cells that emanate a GFP signal. The negative control ("Ctrl") corresponds to cells that were incubated with GFP-NLS recombinant protein (5 μ M) without any shuttle agent.

Table 10.3c: Data from Figure 29G

Domain structure	Shuttle agent	Conc. of shuttle (μ M)	Conc. of GFP-NLS (μ M)	Incubation time (min)	Mean % cells with GFP signal ($\pm$ St. Dev.; n=3)	Cell viability (%) ($\pm$ St. Dev.; n=3)
-	No shuttle ("Ctrl")	0	5	5	0 $\pm$ n/a	98.3 $\pm$ 0.9
CPD-ELD	PTD4-KALA	10	5	1	64.6 $\pm$ 4.3	96.2 $\pm$ 3.0
				2	78.8 $\pm$ 3.6	75.3 $\pm$ 3.8
				5	71.4 $\pm$ 4.2	82.4 $\pm$ 4.7
ELD-CPD	EB1-PTD4	10	5	1	76.3 $\pm$ 3.5	61.7 $\pm$ 2.7
				2	79.0 $\pm$ 3.3	56.6 $\pm$ 3.2
				5	71.1 $\pm$ 5.0	55.8 $\pm$ 4.4
His-ELD-CPD-His	His-CM18-PTD4-His	10	5	1	68.6 $\pm$ 3.5	68.1 $\pm$ 2.7
				2	74.1 $\pm$ 4.3	61.6 $\pm$ 3.2
				5	59.8 $\pm$ 4.0	41.2 $\pm$ 4.4

10

Table 10.3d: Data from Figure 29H

Domain structure	Shuttle agent	Conc. of shuttle (μ M)	Conc. of GFP-NLS (μ M)	Incubation time (min)	Relative Fluorescence Intensity (FL1-A) ($\pm$ St. Dev.; n=3)
-	No shuttle ("Ctrl")	0	5	5	8903 $\pm$ 501.37
CPD-ELD	PTD4-KALA	10	5	1	190 287 $\pm$ 9445
				2	386 480 $\pm$ 17 229
				5	241 230 $\pm$ 14 229
ELD-CPD	EB1-PTD4	10	5	1	178 000 $\pm$ 11 934
				2	277 476 $\pm$ 25 319
				5	376 555 $\pm$ 16 075
His-ELD-CPD-His	His-CM18-PTD4-His	10	5	1	204 338 $\pm$ 22 673
				2	307 329 $\pm$ 19 618
				5	619 964 $\pm$ 17 411

The shuttle agent CM18-PTD4 was used as a model to demonstrate the modular nature of the individual protein domains, as well as their ability to be modified. More particularly, the presence or absence of: an N-terminal cysteine residue ("Cys"); different flexible linkers between the ELD and CPD domains ("L1": GGS; "L2": GGS GGGS; and "L3": GGS GGGS GGGS) and different lengths, positions, and variants to histidine-rich domains; were studied.

5 HeLa cells were cultured and tested in the protein transduction assays using Protocol B as described in **Example 9.1**. Briefly, GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 20 μ M of different shuttle peptide variants (see **Table 1.3** for amino acid sequences and properties) of the shuttle agent His-CM18-PTD4 for 1 minute. After the final washing step, the cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 10.3e** and **Figure 29I**. "Pos cells (%)" is the mean percentages of all cells that emanate a GFP signal. The negative control ("Ctrl") corresponds to cells that were
10 incubated with GFP-NLS recombinant protein (5 μ M) without any shuttle agent.

Table 10.3e: Data from Figure 29I

Domain structure	Shuttle agent	Conc. of shuttle (μ M)	Conc. of GFP-NLS (μ M)	Mean % cells with GFP signal ($\pm$ St. Dev.; n=3)	Cell viability (%) ($\pm$ St. Dev.; n=3)
-	No shuttle ("Ctrl")	0	5	0	99.6 $\pm$ 0.12
ELD-CPD	CM18-PTD4	20	5	47.6 $\pm$ 2.6	33.9 $\pm$ 3.7
	Cys-CM18-PTD4			36.6 $\pm$ 2.3	78.7 $\pm$ 3.1
	CM18-L1-PTD4			48.5 $\pm$ 3.0	50.1 $\pm$ 3.8
	CM18-L2-PTD4			45.5 $\pm$ 6.5	64.0 $\pm$ 1.3
	CM18-L3-PTD4			39.0 $\pm$ 2.7	71.9 $\pm$ 6.0
His-ELD-CPD	His-CM18-PTD4	20	5	60.3 $\pm$ 3.2	81.6 $\pm$ 4.5
	His-CM18-PTD4-6Cys			41.3 $\pm$ 4.28	62 $\pm$ 5.76
	Met-His-CM18-PTD4-Cys			45.6 $\pm$ 3.88	54.9 $\pm$ 3.45
	3His-CM18-PTD4			39.4 $\pm$ 0.5	39.2 $\pm$ 3.3
	12His-CM18-PTD4			36.9 $\pm$ 4.3	33.4 $\pm$ 4.3
	HA-CM18-PTD4			42.3 $\pm$ 4.2	68.3 $\pm$ 4.1
	3HA-CM18-PTD4			37.2 $\pm$ 3.9	43.6 $\pm$ 2.8
ELD-His-CPD	CM18-His-PTD4	20	5	61.7 $\pm$ 1.8	57.7 $\pm$ 4.2
His-ELD-CPD-His	His-CM18-PTD4-His	20	5	68.0 $\pm$ 6.0	78.6 $\pm$ 1.1

15 These results show that variations in a given shuttle (e.g., CM18-PTD4) may be used to modulate the degree of transduction efficiency and cell viability of the given shuttle. More particularly, the addition of an N-terminal cysteine residue to CM18-PTD4 (see Cys-CM18-PTD4), decreased GFP-NLS transduction efficiency by 11% (from 47.6% to 36.6%), but increased cell viability from 33.9% to 78.7%. Introduction of flexible linker domains (L1, L2, and L3) of different lengths between the CM18 and PTD4 domains did not result in a dramatic loss of transduction efficiency, but increased cell viability (see CM18-L1-PTD4, CM18-L2-PTD4, and CM18-L3-PTD4). Finally, variations

to the amino acid sequences and/or positions of the histidine-rich domain(s) did not result in a complete loss of transduction efficiency and cell viability of His-CM18-PTD4 (see 3His-CM18-PTD4, 12His-CM18-PTD4, HA-CM18-PTD4, 3HA-CM18-PTD4, CM18-His-PTD4, and His-CM18-PTD4-His). Of note, adding a second histidine-rich domain at the C terminus of His-CM18-PTD4 (i.e., His-CM18-PTD4-His) increased transduction efficiency from 60% to 68% with similar cell viability.

10.5 Lack of GFP-NLS transduction by single-domain peptides or a His-CPD peptide in HeLa cells: Flow cytometry

HeLa cells were cultured and tested in the protein transduction assays using Protocol B as described in **Example 9.1**. Briefly, GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 50 μ M of different single-domain peptides (TAT; PTD4; Penetratin; CM18; C(LLKK)₃C; KALA) or the two-domain peptide His-PTD4 (lacking an ELD), and exposed to the HeLa cells for 10 seconds. After the final washing step, the cells were subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 10.4** and **Figure 29D**. "Pos cells (%)" is the mean percentages of all cells that emanate a GFP signal. The negative control ("Ctrl") corresponds to cells that were incubated with GFP-NLS recombinant protein (5 μ M) without any single-domain peptide or shuttle agent.

Table 10.4: Data from **Figure 29D**

Protocol	Domain	Single-domain peptide	Cells	Conc. of shuttle (μ M)	Conc. of GFP-NLS (μ M)	Mean % cells with GFP signal ($\pm$ St. Dev.; n=3)	Cell viability (%) ($\pm$ St. Dev.; n=3)
B	-	No peptide ("Ctrl")	HeLa	0	5	0.1 $\pm$ 0.02	98.3 $\pm$ 0.59
	CPD	TAT	HeLa	50	5	1.1 $\pm$ 0.27	94.6 $\pm$ 0.44
		PTD4				1.1 $\pm$ 0.06	94 $\pm$ 4.5
		Penetratin (Pen)				3.6 $\pm$ 0.1	96 $\pm$ 0.6
	ELD	CM18	HeLa	50	5	2.9 $\pm$ 0.2	95 $\pm$ 1.2
		C(LLKK) ₃ C				1.1 $\pm$ 0.57	61.8 $\pm$ 0.1
		KALA				1.4 $\pm$ 0.13	84 $\pm$ 0.7
	His-CPD	His-PTD4	HeLa	50	5	1.04 $\pm$ 0.12	96.5 $\pm$ 0.28

These results show that the single-domain peptides TAT, PTD4, Penetratin, CM18, C(LLKK)₃C, KALA, or the two-domain peptide His-PTD4 (lacking an ELD), are not able to successfully transduce GFP-NLS in HeLa cells.

10.6 GFP-NLS transduction by TAT-KALA, His-CM18-PTD4, His-C(LLKK)₃C-PTD4, PTD4-KALA, EB1-PTD4, and His-CM18-PTD4-His in HeLa cells: Visualization by microscopy

GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 50 μ M of shuttle agent, and then exposed to HeLa cells for 10 seconds using Protocol B as described in **Example 9.1**. The cells were visualized by microscopy as described in **Example 3.2**, after an incubation time of 2 minutes.

For the sample results shown in **Figure 30**, GFP fluorescence of the HeLa cells was immediately visualized by bright field (bottom row panels) and fluorescence (upper and middle row panels) microscopy at 20x or 40x magnifications after the final washing step. The results with the shuttle agents TAT-KALA, His-CM18-PTD4, and His-C(LLKK)₃C-PTD4 are shown in panels A, B and C, respectively. The results with the shuttle agents PTD4-KALA, EB1-PTD4, and His-CM18-PTD4-His are shown in panels D, E and F, respectively. The insets in the bottom row panels show the results of corresponding FACS analyses (performed as described in **Example 3.3**), which indicates the percentage of cells in a 96-plate well with a GFP signal. No significant cellular GFP fluorescence was observed in negative control samples (i.e., cells exposed to GFP-NLS without any shuttle agent; data not shown).

10.7 GFP-NLS transduction by TAT-KALA, His-CM18-PTD4 and His-C(LLKK)₃C-PTD4 with varying incubation times in THP-1 cells: Flow cytometry

THP-1 cells were cultured and tested in the protein transduction assays using Protocol C as described in **Example 9.1**. Briefly, GFP-NLS recombinant protein (5 μ M; see **Example 5.1**) was co-incubated with 1 μ M of TAT-KALA, His-CM18-PTD4, or His-C(LLKK)₃C-PTD4 for 15, 30, 60, or 120 seconds. After the final washing step, the cells were subjected to flow cytometry analysis as described in **Example 3.3**. The mean percentages of cells emanating a GFP signal ("Pos cells (%)") are shown in **Table 10.4** and in **Figure 34A**. The mean fluorescence intensity is shown in **Table 10.5** and **Figure 34B**. The negative control ("Ctrl") corresponds to cells that were incubated with GFP-NLS recombinant protein (5 μ M) without any shuttle agent.

Table 10.4: Data from Figure 34A

Protocol	Shuttle agent	Cells	Conc. of shuttle (μ M)	Conc. of GFP-NLS (μ M)	Incubation time (sec.)	Mean % cells with GFP signal ($\pm$ St. Dev.; n=3)	Cell viability (%) ($\pm$ St. Dev.; n=3)
C	No shuttle ("Ctrl")	THP-1	0	5	120	1.12 $\pm$ 0.27	97.3 $\pm$ 1.55
	TAT-KALA	THP-1	1	5	15	47 $\pm$ 3.5	84.6 $\pm$ 2.7
					30	52.9 $\pm$ 1.3	70.3 $\pm$ 3.2
					60	70.1 $\pm$ 2.0	82.7 $\pm$ 1.4
					120	82.1 $\pm$ 2.5	46.3 $\pm$ 4.9
	His-CM18-PTD4	THP-1	1	5	15	23.7 $\pm$ 0.2	90 $\pm$ 3.0
					30	53 $\pm$ 0.3	89 $\pm$ 1.1
					60	69.6 $\pm$ 4.2	85.3 $\pm$ 3.6
					120	89 $\pm$ 0.8	74.3 $\pm$ 3.2
	His-C(LLKK) ₃ C-PTD4	THP-1	1	5	15	38.4 $\pm$ 0.3	85.2 $\pm$ 2.8
					30	42.3 $\pm$ 4.2	86 $\pm$ 2.0
					60	64.5 $\pm$ 1.0	86.9 $\pm$ 3.8
					120	78.7 $\pm$ 0.3	79.6 $\pm$ 2.8

Table 10.5: Data from Figure 34B

Protocol	Shuttle agent	Cells	Conc. of shuttle (μ M)	Incubation time (sec.)	Relative fluorescence intensity (FL1-A) (n = 3)	Standard Deviation
C	No shuttle ("Ctrl")	THP-1	0	120	217	23.09
	TAT-KALA	THP-1	1	15	6 455.12	333.48
				30	8 106.81	436.28
				60	13 286.2	463.96
				120	27 464.92	2 366.48
	His-CM18-PTD4	THP-1	1	15	5 605.45	933.16
				30	25 076.41	5 567.33
				60	34 046.94	3 669.19
				120	55 613.48	9 836.84
	His-C(LLKK) ₃ C-PTD4	THP-1	1	15	5 475.12	693.29
				30	5 755.8	635.18
				60	8 267.38	733.29
				120	21 165.06	209.37

Example 11:**Repeated daily treatments with low concentrations of shuttle agent in the presence of serum results in GFP-NLS transduction in THP-1 cells****11.1 GFP-NLS transduction with His-CM18-PTD4 or His-C(LLKK)₃C-PTD4 in THP-1 cells: Flow cytometry**

THP-1 cells were cultured and tested in the protein transduction assay using Protocol A as described in **Example 9.1**, but with the following modifications. GFP-NLS recombinant protein (5, 2.5, or 1 μ M; see **Example 5.1**) was co-incubated with 0.5 or 0.8 μ M of His-CM18-PTD4, or with 0.8 μ M of His-C(LLKK)₃C-PTD4, and then exposed to THP-1 cells each day for 150 min in the presence of cell culture medium containing serum. Cells were washed and subjected to flow cytometry analysis as described in **Example 3.3** after 1 or 3 days of repeated exposure to the shuttle agent/cargo. The results are shown in **Table 11.1** and in **Figure 35A, B, C and F**. The negative control ("Ctrl") corresponds to cells that were incubated with GFP-NLS recombinant protein (5 μ M) without any shuttle agent.

Table 11.1: Data from Figure 35A, B, C and F

Figure	Shuttle agent	Cells	Conc. of shuttle (μM)	Conc. of GFP-NLS (μM)	Exposure to shuttle/cargo (days)	Mean % cells with GFP signal (± St. Dev.; n=3)	Cell viability (%) (± St. Dev.; n=3)
35A	No shuttle (Ctrl)	THP-1	0	5	0	0.15 ± 0.04	98.7 ± 0.1
	His-CM18-PTD4		0.5	5	1	12.1 ± 1.5	98.2 ± 2.4
					3	73.4 ± 1.1	84.3 ± 3.8
35B	No shuttle (Ctrl)	THP-1	0	5	0	0.36 ± 0.09	97.1 ± 1.2
	His-CM18-PTD4		0.8	2.5	1	12.2 ± 0.9	92.3 ± 1.9
					3	62.4 ± 3.5	68.5 ± 2.2
35C	No shuttle (Ctrl)	THP-1	0	5	0	0.28 ± 0.05	96.4 ± 2.0
	His-CM18-PTD4		0.8	1	1	1.6 ± 0.2	98.4 ± 6.4
					3	6.5 ± 0.9	80.6 ± 4.6
35F	No shuttle (Ctrl)	THP-1	0	5	0	0.62 ± 0.11	96.3 ± 1.4
	His-C(LLKK) ₃ C-PTD4		0.8	1	1	1.8 ± 0.2	97.2 ± 2.2
					3	6.6 ± 0.8	76.6 ± 3.4

The viability of THP-1 cells repeatedly exposed to His-CM18-PTD4 and GFP-NLS was determined as described in **Example 3.3a**. The results are shown in **Tables 11.2 and 11.3** and in **Figure 35D and 35E**. The results in **Table 11.2** and **Figure 35D** show the metabolic activity index of the THP-1 cells after 1, 2, 4, and 24h, and the results in **Table 11.3** and **Figure 35E** show the metabolic activity index of the THP-1 cells after 1 to 4 days.

Table 11.2: Data from Figure 35D

Shuttle agent	Cells	Conc. of shuttle (μ M)	Conc. of GFP-NLS (μ M)	Mean metabolic activity index ($\pm$ St. Dev.; n=3) (Exposure to shuttle/cargo)			
				1h	2h	4h	24h
No shuttle (Ctrl)	THP-1	0	5	40810 $\pm$ 757.39	38223 $\pm$ 238.66	44058 $\pm$ 320.23	42362 $\pm$ 333.80
His-CM18-PTD4	THP-1	0.5	5	9974 $\pm$ 1749.85	9707 $\pm$ 1259.82	3619 $\pm$ 2247.54	2559 $\pm$ 528.50
		1	5	42915 $\pm$ 259.67	41386 $\pm$ 670.66	44806 $\pm$ 824.71	43112 $\pm$ 634.56

Table 11.3: Data from Figure 35E

Shuttle agent	Cells	Conc. of shuttle (μ M)	Conc. of GFP-NLS (μ M)	Mean metabolic activity index ($\pm$ St. Dev.; n=3) (Exposure to shuttle/cargo)			
				1 day	2 days	3 days	4 days
No shuttle (Ctrl)	THP-1	0	5	44684 $\pm$ 283.27	43389 $\pm$ 642.47	45312 $\pm$ 963.40	43697 $\pm$ 1233
His-CM18-PTD4	THP-1	0.5	5	44665 $\pm$ 310.3	42664 $\pm$ 398.46	43927 $\pm$ 3511.54	43919 $\pm$ 4452.25
		0.8	5	44531 $\pm$ 176.66	43667 $\pm$ 421.66	44586 $\pm$ 383.68	44122 $\pm$ 239.98
		1	5	41386 $\pm$ 670.66	36422 $\pm$ 495.01	27965 $\pm$ 165.33	22564 $\pm$ 931.28

- 5 The results in **Example 11** show that repeated daily (or chronic) treatments with relatively low concentrations of His-CM18-PTD4 or His-C(LLKK)₃C-PTD4 in the presence of serum result in intracellular delivery of GFP-NLS in THP-1 cells. The results also suggest that the dosages of the shuttle agents and the cargo can be independently adjusted to improve cargo transduction efficiency and/or cell viability.

10 **Example 12:**

His-CM18-PTD4 increases transduction efficiency and nuclear delivery of GFP-NLS in a plurality of cell lines

12.1 **GFP-NLS transduction with His-CM18-PTD4 in different adherent & suspension cells: Flow cytometry**

- 15 The ability of the shuttle agent His-CM18-PTD4 to deliver GFP-NLS to the nuclei of different adherent and suspension cells using Protocols B (adherent cells) or C (suspension cells) as described in **Example 9.1** was examined. The cell lines tested included: HeLa, Balb3T3, HEK 293T, CHO, NIH3T3, Myoblasts, Jurkat, THP-1, CA46, and HT2 cells, which were cultured as described in **Example 1**. GFP-NLS (5 μ M; see **Example 5.1**) was co-incubated with 35 μ M of His-CM18-PTD4 and exposed to adherent cells for 10 seconds (Protocol B), or was co-

incubated with 5 μM of His-CM18-PTD4 and exposed to suspension cells for 15 seconds (Protocol C). Cells were washed and subjected to flow cytometry analysis as described in **Example 3.3**. Results are shown in **Table 12.1** and **Figure 36**. "Pos cells (%)" is the mean percentages of all cells that emanate a GFP signal.

Table 12.1: Data from **Figure 36**

Shuttle agent	Protocol	Conc. of shuttle (μM)	Conc. of GFP-NLS (μM)	Cells	Mean % cells with GFP signal ($\pm$ St. Dev.; n=3)	Cell viability (%) ($\pm$ St. Dev.; n=3)
His-CM18-PTD4	B	35	5	HeLa	72.3 $\pm$ 5.3	94.6 $\pm$ 0.4
				Balb3T3	40.2 $\pm$ 3.1	98.4 $\pm$ 0.6
				HEK 293T	55.3 $\pm$ 0.2	95.3 $\pm$ 1.2
				CHO	53.7 $\pm$ 4.6	92.8 $\pm$ 0.1
				NIH3T3	35.4 $\pm$ 3.9	3.3 $\pm$ 5.4
				Myoblasts	25.6 $\pm$ 2.6	23.5 $\pm$ 1.1
	C	5	5	Jurkat	30.7 $\pm$ 2.2	73.6 $\pm$ 0.7
				THP-1	64.1 $\pm$ 1.6	64.1 $\pm$ 4.5
				CA46	24.4 $\pm$ 0.6	71.6 $\pm$ 1.0
				HT2	30.5 $\pm$ 2.5	90.6 $\pm$ 1.5

12.2 GFP-NLS transduction with His-CM18-PTD4 in several adherent and suspension cells: visualization by microscopy

GFP-NLS recombinant protein (5 μM ; see **Example 5.1**) was co-incubated with 35 μM of His-CM18-PTD4 and exposed to adherent cells for 10 seconds using Protocol A, or was co-incubated with 5 μM of His-CM18-PTD4 and exposed to suspension cells for 15 seconds using Protocol B, as described in **Example 9.1**. After washing the cells, GFP fluorescence was visualized by bright field and fluorescence microscopy. Sample images captured at 10x magnifications showing GFP fluorescence are shown in **Figure 37** for (A) 293T, (B) Balb3T3, (C) CHO, (D) Myoblasts, (E) Jurkat, (F) CA46, (G) HT2, and (H) NIH3T3 cells. The insets show corresponding flow cytometry results performed as described in **Example 3.3**, indicating the percentage of GFP-NLS-positive cells. No significant cellular GFP fluorescence was observed in negative control samples (i.e., cells exposed to GFP-NLS without any shuttle agent; data not shown).

Nuclear localization of the GFP-NLS was further confirmed in fixed and permeabilized myoblasts using cell immuno-labelling as described in **Example 3.2a**. GFP-NLS was labeled using a primary mouse monoclonal anti-GFP antibody (Feldan, #A017) and a secondary goat anti-mouse AlexaTM-594 antibody (Abcam #150116). Nuclei were labelled with DAPI. Sample results for primary human myoblast cells are shown in **Figure 38**, in which GFP immuno-labelling is shown in panel A, and an overlay of the GFP immuno-labelling and DAPI labelling is shown in panel B. No significant cellular GFP labelling was observed in negative control samples (i.e., cells exposed to GFP-NLS without any shuttle agent; data not shown).

The microscopy results revealed that GFP-NLS is successfully delivered to the nucleus of all the tested cells using the shuttle agent His-CM18-PTD4.

Example 13:**His-CM18-PTD4 enables transduction of a CRISPR/Cas9-NLS system and genome editing in Hela cells****13.1 Cas9-NLS recombinant protein**

Cas9-NLS recombinant protein was constructed, expressed and purified from a bacterial expression system as described in **Example 1.4**. The sequence of the Cas9-NLS recombinant protein produced was:

```

MDKKYSIGLDIGTNSVGWAVITDEYKVPSSKKFKVLGNTDRHSIKKNLIGALLFDSGETAEATRLKR
TARRRYTRRNRI CYLQEIFSNEMAKVDDSFHRLSEESFLVEEDKKHERHPIFGNIVDEVAYHEKY
PTIYHLRKKLV DSTDKADLR LIYLALAHMIKFRGHFLIEGDLNPDNSDVKLF IQLVQTYNQLFEE
NPINASGVDAKAIL SARLSKSRRL ENLIAQLPGEKKNGLFGNLIALSLGLTPNFKSNFDLAEDAKL
QLSKD TYDDDLNLLAQIGDQYADFLAAKNLSDAI LLSDI LRVNTEITKAPLSASMIKRYDEHHQ
DLTLLKALVRQQLPEKYKEIFFDQSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEELLVKLNRE
DLLRKQRTFDNGSI PHQIHLGELHAILRRQEDFY PFLKDNREKIEKILTFRI PYVVGPLARGNSRF
AWMTRKSEETITPWNFEFEEVDKGASAQSFIERMTNFDKNLPNEKVLPHKSLLEYFTVYNELTKVK
YVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKIECFDSVEISGVEDRFNASLGTY
HDLLKIIKDKDFLDNEENEDILEDIVLTTLTFEDREMIERLKYAHLFDDKVMKQLKRRRYTGWG
RLSRKLINGIRDKQSGKTI LDFLKSDGFANRNF MQLIHDDSLTFKEDIQKAQVSGQGDSLHEHIAN
LAGSPA I KKGILQTVKVVDLVKVMGRHKPENIVI EMARENQTTQKGQNSRERMKRIE EGIKELG
SQILKEHPVENTQLQNEKLYLYYLQNGRDMYVDQELDINRLSDYDVDHIVPQSFLKDDSIDNKVLT
RSDKNRGKSDNVPSEEVVKMKNYWRQLLNAKLITQRKFDNLTKAERGGLSELDKAGFIKRQLVET
RQITKHVAQILDSRMNTKYDENDKLIREVKVITLKSCLVSDFRKDFQFYKVR EINNYHHAHDAYLN
AVVGTA L I KKP KLESEFVYGDYKVYDVRKMI AKSEQEI GKATAKYFFYSNIMNFFKTEITLANGE
IRKRPLIETNGETGEIVWDKGRDFATVRKVL SMPQVNI VKKTEVQTGGFSKESI LPKRNSDKLIAR
KKDWD PKKYGGFDSPTVAYSVLVAKVEKGSKKLKS VKELLGITIMERS SF EKNPIDFLEAKGYK
EVK KDL I I KLPKYSLFELENGRKRMLASAGELQKGNELALPSKYVNFY LASHYEKLKGS PEDNEQ
KQLFVEQHKHYLDEIIEQISEFSKRVI LADANLDKVL SAYNKH RDKPIREQAENI IHLFTLTNLGA
PAAFKYFDTTIDRKRYTSTKEVL DATLIHQSI TGLYETRIDLSQLGGDGRSSDDEATADSQHAAP
PKKKRKVGGSGGGSGGGSGGGRHHHHHH [SEQ ID NO: 74]
(MW = 162.9 kDa; pI = 9.05)
NLS sequence is underlined
Serine/glycine rich linkers are in bold

```

13.2 Transfection plasmid surrogate assay

This assay enables one to visually identify cells that have been successfully delivered an active CRISPR/Cas9 complex. As shown in **Figure 39A**, the assay involves transfecting cells with an expression plasmid DNA encoding the fluorescent proteins mCherry™ and GFP, with a STOP codon separating their two open reading frames. Transfection of the cells with the expression plasmid results in mCherry™ expression, but no GFP expression (**Figure 39B**). A CRISPR/Cas9 complex, which has been designed/programmed to cleave the plasmid DNA at the STOP codon, is then delivered intracellularly to the transfected cells expressing mCherry™ (**Figure 39D**). Successful transduction of an active CRISPR/Cas9 complex results in the CRISPR/Cas9 complex cleaving the plasmid DNA at the STOP codon (**Figure 39C**). In a fraction of the cells, random non-homologous DNA repair of the cleaved plasmid occurs and results in removal of the STOP codon, and thus GFP expression and fluorescence (**Figure 39E**).

On Day 1 of the transfection plasmid surrogate assay, DNA plasmids for different experimental conditions (250 ng) are diluted in DMEM (50 μ L) in separate sterile 1.5-mL tubes, vortexed and briefly centrifuged. In separate sterile 1.5-mL tubes, Fastfect™ transfection reagent was diluted in DMEM (50 μ L) with no serum and no antibiotics at a ratio of 3:1 (3 μ L of Fastfect™ transfection reagent for 1 μ g of DNA) and then quickly vortexed and briefly centrifuged. The Fastfect™/DMEM mixture was then added to the DNA mix and quickly vortexed and briefly centrifuged. The Fastfect™/DMEM/DNA mixture is then incubated for 15-20 min at room temperature, before being added to the cells (100 μ L per well). The cells are then incubated at 37°C and 5% CO₂ for 5h. The media is then changed for complete medium (with serum) and further incubated at 37°C and 5% CO₂ for 24-48h. The cells are then visualized under fluorescent microscopy to view the mCherry™ signal.

13.3 His-CM18-PTD4 - mediated CRISPR/Cas9-NLS system delivery and cleavage of plasmid DNA

RNAs (crRNA & tracrRNA) were designed to target a nucleotide sequence of the EMX1 gene, containing a STOP codon between the mCherry™ and GFP coding sequences in the plasmid of **Example 13.2**. The sequences of the crRNA and tracrRNA used were as follows:

- crRNA [SEQ ID NO: 75]: 5' - GAGUCCGAGCAGAAGAAGAAGUUUUAGAGCUAUGCUGUUUUG - 3' - tracrRNA [SEQ ID NO: 76]: 5' - AAACAGCAUAGCAAGUUAAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGUCU - 3'
--

HeLa cells were cultured and subjected to the transfection plasmid surrogate assay as described in **Example 13.2**. On Day 1, the HeLa cells were transfected with a plasmid surrogate encoding the mCherry™ protein as shown in **Figure 39A**. On Day 2, a mix of Cas9-NLS recombinant protein (2 μ M; see **Example 13.1**) and RNAs (crRNA & tracrRNA; 2 μ M; see above) were co-incubated with 50 μ M of His-CM18-PTD4, and the mixture (CRISPR/Cas9 complex) was exposed to HeLa cells for 10 seconds using Protocol B as described in **Example 9.1**. Double-stranded plasmid DNA cleavage by the CRISPR/Cas9 complex at the STOP codon between the mCherry™ and GFP coding sequences (**Figure 39B**), and subsequent non-homologous repair by the cell in some cases results in removal of the STOP codon (**Figure 39C**), thereby allowing expression of both the mCherry™ and GFP fluorescent proteins in the same cell on Day 3 (**Figure 39D-E**). White triangle windows in panels D and E indicate examples of areas of co-labelling between mCherry™ and GFP.

As a positive control for the CRISPR/Cas9-NLS system, HeLa cells were cultured and co-transfected with three plasmids: the plasmid surrogate (as described in **Example 13.2**) and other expression plasmids encoding the Cas9-NLS protein (**Example 13.1**) and the crRNA/tracrRNAs (**Example 13.3**). Typical fluorescence microscopy results are shown in **Figure 40A-D**. Panels A and B show cells 24 hours post-transfection, while panels C and D show cells 72 hours post-transfection.

Figure 40E-H shows the results of a parallel transfection plasmid surrogate assay performed using 35 μ M of the shuttle His-CM18-PTD4, as described for **Figure 39**. Panels E and F show cells 24 hours post-transduction, while panels G and H show cells 48 hours post-transduction. Panels E and G show mCherryTM fluorescence, and panels F and H show GFP fluorescence, the latter resulting from removal of the STOP codon by the transduced CRISPR/Cas9-NLS complex and subsequent non-homologous repair by the cell. No significant cellular GFP fluorescence was observed in negative control samples (i.e., cells exposed to CRISPR/Cas9-NLS complex without any shuttle agent; data not shown).

13.4 T7E1 assay

The T7E1 assay was performed with the Edit-RTM Synthetic crRNA Positive Controls (Dharmacon #U-007000-05) and the T7 Endonuclease I (NEB, Cat #M0302S). After the delivery of the CRISPR/Cas9 complex, cells were lysed in 100 μ L of PhusionTM High-Fidelity DNA polymerase (NEB #M0530S) laboratory with additives. The cells were incubated for 15-30 minutes at 56°C, followed by deactivation for 5 minutes at 96°C. The plate was briefly centrifuged to collect the liquid at bottom of the wells. 50- μ L PCR samples were set up for each sample to be analyzed. The PCR samples were heated to 95°C for 10 minutes and then slowly (>15 minutes) cooled to room temperature. PCR product (~5 μ L) was then separated on an agarose gel (2%) to confirm amplification. 15 μ L of each reaction was incubated with T7E1 nuclease for 25 minutes at 37 °C. Immediately, the entire reaction volume was run with the appropriate gel loading buffer on an agarose gel (2%).

13.5 His-CM18-PTD4 and His-C(LLKK)₃C-PTD4 -mediated CRISPR/Cas9-NLS system delivery and cleavage of genomic PPIB sequence

A mix composed of a Cas9-NLS recombinant protein (25 nM; **Example 13.1**) and crRNA/tracrRNA (50 nM; see below) targeting a nucleotide sequence of the PPIB gene were co-incubated with 10 μ M of His-CM18-PTD4 or His-C(LLKK)₃C-PTD4, and incubated with HeLa cells for 16h in medium without serum using Protocol A as described in **Example 9.1**.

The sequences of the crRNA and tracrRNAs constructed and their targets were:

- Feldan tracrRNA [SEQ ID NO: 77]: 5' - AAACAGCAUAGCAAGUUAUAAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGUCU - 3' - PPIB crRNA [SEQ ID NO: 78]: 5' - GUGUAUUUUGACCUACGAAUGUUUUAGAGCUAUGCUGUUUUG - 3' - Dharmacon tracrRNA [SEQ ID NO: 79]: 5' - AACAGCAUAGCAAGUUAUAAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUUUUU - 3'
--

After 16h, HeLa cells were washed with PBS and incubated in medium with serum for 48h. HeLa cells were harvested to proceed with the T7E1 protocol assay as described in **Example 13.4**.

Figure 41A shows an agarose gel with the PPIB DNA sequences after PCR amplification. Lane **A** shows the amplified PPIB DNA sequence in HeLa cells without any treatment (i.e., no shuttle or Cas9/RNAs complex). Lanes **B**: The two bands framed in white box #1 are the cleavage product of the PPIB DNA sequence by the CRIPR/Cas9 complex after the delivery of the complex with the shuttle His-C(LLKK)₃C-PTD4. Lane **C**: These bands show the amplified PPIB DNA sequence after incubation of the HeLa cells with the Cas9/RNAs complex without shuttle (negative control). Lane **D**: The bands framed in white box #2 show the amplified PPIB DNA sequence after incubation of the HeLa cells with the Cas9/RNAs complex in presence of a lipidic transfection agent (DharmaFect™ tranfection reagent # T-20XX-01) (positive control). Similar results were obtained using the shuttle His-CM18-PTD4 (data not shown).

Figure 41B shows an agarose gel with the PPIB DNA sequences after PCR amplification. The left panel shows the cleavage product of the amplified PPIB DNA sequence by the CRIPR/Cas9 complex after the delivery of the complex with the shuttle agent His-CM18-PTD4 in HeLa cells. The right panel shows amplified DNA sequence before the T7E1 digestion procedure as a negative control.

Figure 41C shows an agarose gel with the PPIB DNA sequences after PCR amplification. The left panel shows the amplified PPIB DNA sequence after incubation of the HeLa cells with the Cas9/RNAs complex in presence of a lipidic transfection agent (DharmaFect™ tranfection reagent # T-20XX-01) (positive control). The right panel shows amplified DNA sequence before the T7E1 digestion procedure as a negative control.

These results show that the shuttle agents His-CM18-PTD4 and His-C(LLKK)₃C-PTD4 successfully deliver a functional CRISPR/Cas9 complex to the nucleus of HeLa cells, and that this delivery results in CRISPR/Cas9-mediated cleavage of genomic DNA.

13.6 CRISPR/Cas9-NLS system delivery by different shuttle agents, and cleavage of genomic HPTR sequence in HeLa and Jurkat cells

A mix composed of a Cas9-NLS recombinant protein (2.5 μM; **Example 13.1**) and crRNA/tracrRNA (2 μM; see below) targeting a nucleotide sequence of the HPTR gene were co-incubated with 35 μM of His-CM18-PTD4, His-CM18-PTD4-His, His-C(LLKK)₃C-PTD4, or EB1-PTD4, and incubated with HeLa or Jurkat cells for 2 minutes in PBS using Protocol B as described in **Example 9.1**.

The sequences of the crRNA and tracrRNAs constructed and their targets were:

```
- Feldan tracrRNA [SEQ ID NO: 77]:
  5' - AAACAGCAUAGCAAGUAAAAUAAGGCUAGUCGCUUAUACAACUUGAAAAAGUGGCACCGAGUCGGUGCU - 3'

- HPRT crRNA [SEQ ID NO: 103]:
  5' - AAUUAUGGGGAUUACUAGGAGUUUUAGAGCUAUGCU - 3'
```

After 2 minutes, cells were washed with PBS and incubated in medium with serum for 48h. Cells were harvested to proceed with the T7E1 protocol assay as described in **Example 13.4**. **Figure 46** shows an agarose gel

with the HPTR DNA sequences after PCR amplification and the cleavage product of the amplified HPTR DNA sequence by the CRISPR/Cas9 complex after the delivery of the complex with the different shuttle agents. Panel **A** shows the results with the shuttle agents: His-CM18-PTD4, His-CM18-PTD4-His, and His-C(LLKK)₃C-PTD4 in HeLa cells. Panel **B** shows the results with His-CM18-PTD4 and His-CM18-L2-PTD4 in Jurkat cells. Negative controls (lanes 4) show amplified HPTR DNA sequence after incubation of the cells with the CRISPR/Cas9 complex without the presence of the shuttle agent. Positive controls (lane 5 in panels A and B) show the amplified HPTR DNA sequence after incubation of the cells with the Cas9/RNAs complex in presence of a lipidic transfection agent (Lipofectamine® RNAiMAX™ Transfection Reagent ThermoFisher Product No. 13778100).

These results show that different polypeptide shuttle agents of the present description may successfully deliver a functional CRISPR/Cas9 complex to the nucleus of HeLa and Jurkat cells, and that this delivery results in CRISPR/Cas9-mediated cleavage of genomic DNA.

Example 14:

His-CM18-PTD4 enables transduction of the transcription factor HOXB4 in THP-1 cells

14.1 HOXB4-WT recombinant protein

Human HOXB4 recombinant protein was constructed, expressed and purified from a bacterial expression system as described in **Example 1.4**. The sequence of the HOXB4-WT recombinant protein produced was:

MHHHHHHMAMSSFLINSNYVDPKFPPCEEYSQSDYLPSDHSPGYYAGGQRRESSFQPEAGFGRRAA
CTVQRYPPPPPPPPGLSPRAPAPPAGALLPEPGQRC EAVSSPPPPPCAQNPLHPSPSHSACK
EPVVYPWMRKVHVSTVNPNYAGGEPKRSRTAYTRQQVLELEKEFHYNRYLTRRRRVEIAHALCLSE
RQIKIWFQNRMRMKWKDHLKLPNTKIRSGGAAGSAGGPPGRPNGGPRAL **[SEQ ID NO: 80]**

(MW = 28.54 kDa; pI = 9.89)

The initiator methionine and the 6x Histidine tag are shown in bold.

14.2 Real-Time Polymerase Chain Reaction (rt-PCR)

Control and treated cells are transferred to separate sterile 1.5-mL tubes and centrifuged for 5 minutes at 300g. The cell pellets are resuspended in appropriate buffer to lyse the cells. RNAase-free 70% ethanol is then added followed by mixing by pipetting. The lysates are transferred to an RNeasy™ Mini spin column and centrifuged 30 seconds at 13000 RPM. After several washes with appropriate buffers and centrifugation steps, the eluates are collected in sterile 1.5-mL tubes on ice, and the RNA quantity in each tube is then quantified with a spectrophotometer. For DNase treatment, 2 µg of RNA is diluted in 15 µL of RNase-free water. 1.75 µL of 10X DNase buffer and 0.75 µL of DNase is then added, followed by incubation at 37°C for 15 minutes. For reverse transcriptase treatment, 0.88 µL of EDTA (50 nM) is added, followed by incubation at 75°C for 5 minutes. In a PCR tube, 0.5 µg of DNase-treated RNA is mixed with 4 µL of iScript™ Reverse transcription Supermix (5X) and 20 µL of nuclease-free water. The mix is incubated in a PCR machine with the following program: 5 min at 25°C, 30 min at

42°C and 5 min at 85°C. Newly synthesized cDNA is transferred in sterile 1.5-mL tubes and diluted in 2 µL of nuclease-free water. 18 µL per well of a qPCR machine (CFX-96™) mix is then added in a PCR plate for analysis.

14.3 HOXB4-WT transduction by His-CM18-PTD4 in THP-1 cells: Dose responses and viability

5 THP-1 cells were cultured and tested in the protein transduction assay using Protocol A as described in **Example 9.1**. Briefly, THP-1 cells were plated at 30 000 cells/well one day before transduction. HOXB4-WT recombinant protein (0.3, 0.9, or 1.5 µM; **Example 14.1**) was co-incubated with different concentrations of His-CM18-PTD4 (0, 0.5, 7.5, 0.8 or 1 µM) and then exposed to THP-1 cells for 2.5 hours in the presence of serum. The cells were subjected to real time-PCR analysis as described in **Example 14.2** to measure the mRNA levels of a target gene as a marker for HOXB4 activity, which was then normalized to the target gene mRNA levels detected in the negative control cells (no treatment), to obtain a "Fold over control" value. Total RNA levels (ng/µL) were also measured as a marker for cell viability. Results are shown in **Table 14.1** and **Figure 42**.

Table 14.1: Data from **Figure 42**

Cargo / shuttle agent (Fig. 41)	Cells	Conc. of shuttle (µM)	Conc. of HOXB4-WT (µM)	Fold over control (mean ± St. Dev)	Total RNA in ng/µL (mean ± St. Dev)
No treatment ("Ø")	THP-1	0	0	1 ± 0.1	263 ± 0.4
HOXB4-WT alone ("TF")	THP-1	0	1.5	4.3 ± 0.1	271 ± 6.0
His-CM18-PTD4 alone ("FS")	THP-1	1	0	2.7 ± 0.3	252 ± 10.7
His-CM18-PTD4 + HOXB4-WT	THP-1	0.5	0.3	2.7 ± 0.6	255 ± 3.9
			0.9	4.3 ± 2.1	239 ± 17.5
			1.5	3.8 ± 0.7	269 ± 6.4
His-CM18-PTD4 + HOXB4-WT	THP-1	0.75	0.3	4.2 ± 1.2	248 ± 28
			0.9	5.7 ± 2.5	245 ± 31
			1.5	7.5 ± 2.8	230 ± 3.3
His-CM18-PTD4 + HOXB4-WT	THP-1	0.8	0.3	9.1 ± 2.7	274 ± 4.4
			0.9	16.4 ± 1.7	272 ± 12.5
			1.5	22.7 ± 3.2	282 ± 4.7
His-CM18-PTD4 + HOXB4-WT	THP-1	0.9	0.3	10.2 ± 2.5	280 ± 11.3
			0.9	18.7 ± 3.1	281 ± 9.2
			1.5	26.1 ± 3.5	253 ± 7.1
His-CM18-PTD4 + HOXB4-WT	THP-1	1	0.3	10.5 ± 0.7	184 ± 12.3
			0.9	17 ± 3.7	168 ± 16.2
			1.5	24.5 ± 3.9	154 ± 4.7

15 These results show that exposing THP-1 cells to a mixture of the shuttle agent His-CM18-PTD4 and the transcription factor HOXB4-WT for 2.5 hours in the presence of serum results in a dose-dependent increase in

mRNA transcription of the target gene. These results suggest that HOXB4-WT is successfully delivered in an active form to the nucleus of THP-1 cells, where it can mediate transcriptional activation.

14.4 HOXB4-WT transduction by His-CM18-PTD4 in THP-1 cells: Time course and viability (0 to 48 hours)

THP-1 cells were cultured and tested in the protein transduction assay using Protocol A as described in **Example 9.1**. Briefly, THP-1 cells were plated at 30 000 cells/well one day before the first time course experiment. HOXB4-WT recombinant protein (1.5 μ M; **Example 14.1**) was co-incubated with His-CM18-PTD4 (0.8 μ M) and then exposed to THP-1 cells for 0, 2.5, 4, 24 or 48 hours in presence of serum. The cells were subjected to real time-PCR analysis as described in **Example 14.2** to measure mRNA levels of a target gene as a marker for HOXB4 activity, which was then normalized to the target gene mRNA levels detected in the negative control cells (no treatment), to obtain a "Fold over control" value. Total RNA levels (ng/ μ L) were also measured as a marker for cell viability. Results are shown in **Table 14.2** and **Figure 43**.

Table 14.2: Data from **Figure 43**

Cargo / shuttle agent (Fig. 43)	Cells	Conc. of shuttle (μ M)	Conc. of HOXB4-WT (μ M)	Exposure time (hours)	Fold over control (mean $\pm$ St. Dev)	Total RNA in ng/ μ L (mean $\pm$ St. Dev)
No treatment ("Ctrl")	THP-1	0	0	-	1 $\pm$ 0.1	180 $\pm$ 0.4
HOXB4-WT alone ("TF")	THP-1	0	1.5	2.5 h	3.4 $\pm$ 0.3	129 $\pm$ 10.7
His-CM18-PTD4 alone ("FS")	THP-1	0.8	0	2.5 h	1.2 $\pm$ 0.14	184 $\pm$ 6.0
His-CM18-PTD4 + HOXB4-WT	THP-1	0.8	1.5	48 h	0.27 $\pm$ 0.1	58 $\pm$ 11.2
				24 h	0.8 $\pm$ 0.14	74 $\pm$ 9.2
				4 h	5.6 $\pm$ 1.2	94 $\pm$ 7.1
				2.5 h	9.1 $\pm$ 1.2	146 $\pm$ 11.6
				0	3.9 $\pm$ 0.4	167 $\pm$ 13

14.5 HOXB4-WT transduction by His-CM18-PTD4 in THP-1 cells: Time course and viability (0 to 4 hours)

THP-1 cells were cultured and tested in the protein transduction assay using Protocol A as described in **Example 9.1**. Briefly, THP-1 cells were plated at 30 000 cells/well one day before the first time course experiment. HOXB4-WT recombinant protein (0.3 μ M; **Example 14.1**) was co-incubated with His-CM18-PTD4 (0.8 μ M) and then exposed to THP-1 cells for 0, 0.5, 1, 2, 2.5, 3 or 4 hours in presence of serum. The cells were subjected to real time-PCR analysis as described in **Example 14.2** to measure mRNA levels of a target gene as a marker for HOXB4 activity, which was then normalized to target gene mRNA levels detected in the negative control cells (no treatment), to obtain a "Fold over control" value. Total RNA levels (ng/ μ L) were also measured as a marker for cell viability. Results are shown in **Table 14.3** and **Figure 44**.

Table 14.3: Data from Figure 44

Cargo / shuttle agent (Fig. 42)	Cells	Conc. of shuttle (μ M)	Conc. of HOXB4-WT (μ M)	Exposure time (hours)	Fold over control (mean $\pm$ St. Dev)	Total RNA in ng/ μ L (mean $\pm$ St. Dev)
No treatment ("Ctrl")	THP-1	0	0	-	1 $\pm$ 0.1	289 $\pm$ 9.2
His-CM18-PTD4 alone ("FS")	THP-1	0	0.3	2.5 h	2.5 $\pm$ 0.2	260 $\pm$ 7.1
HOXB4-WT alone ("TF")	THP-1	0.8	0	2.5 h	1 $\pm$ 0.14	264 $\pm$ 12.3
His-CM18-PTD4 + HOXB4-WT	THP-1	0.8	0.3	4 h	1.2 $\pm$ 0.1	198 $\pm$ 6.0
				3 h	1.3 $\pm$ 0.21	268 $\pm$ 12.5
				2.5 h	2 $\pm$ 0.3	275 $\pm$ 4.7
				2 h	2.2 $\pm$ 0.2	269 $\pm$ 12.5
				1	9.7 $\pm$ 2.6	268 $\pm$ 3.9
				0.5	23.1 $\pm$ 2.0	266 $\pm$ 17.5
				0	4 $\pm$ 0.5	217 $\pm$ 6.4

14.6 HOXB4-WT transduction by His-CM18-PTD4 in HeLa cells: immuno-labelling and visualization by microscopy

5 Recombinant HOXB4-WT transcription factor (25 μ M; **Example 14.1**) was co-incubated with 35 μ M of His-CM18-PTD4 and exposed to HeLa cells for 10 seconds using Protocol B as described in **Example 9.1**. After a 30-minute incubation to allow transduced HOXB4-WT to accumulate in the nucleus, the cells were fixed, permeabilized and immuno-labelled as described in **Example 3.2a**. HOXB4-WT was labelled using a primary mouse anti-HOXB4 monoclonal antibody (Novus Bio #NBP2-37257) diluted 1/500, and a secondary anti-mouse antibody AlexaTM-594

10 (Abcam #150116) diluted 1/1000. Nuclei were labelled with DAPI. The cells were visualized by bright field and fluorescence microscopy at 20x and 40x magnifications as described in **Example 3.2**, and sample results are shown in **Figure 45**. Co-localization was observed between nuclei labelling (panels A and C) and HOXB4-WT labelling (panels B and D), indicating that HOXB4-WT was successfully delivered to the nucleus after 30 min in the presence of the shuttle agent His-CM18-PTD4. White triangle windows show examples of areas of co-localization between the

15 nuclei (DAPI) and HOXB4-WT immuno-labels.

14.7 HOXB4-WT transduction by different shuttle agents in THP-1 cells: Dose responses and viability

THP-1 cells were cultured and tested in the protein transduction assay using Protocol A as described in **Example 9.1**. Briefly, THP-1 cells were plated at 30 000 cells/well one day before the first time course experiment.

20 HOXB4-WT recombinant protein (1.5 μ M; **Example 14.1**) co-incubated with the shuttle agents His-CM18-PTD4, TAT-KALA, EB1-PTD4, His-C(LLKK)₃C-PTD4 and His-CM18-PTD4-His at 0.8 μ M, and then exposed to THP-1 cells for 2.5 hours in presence of serum. The cells were subjected to real time-PCR analysis as described in **Example**

14.2 to measure mRNA levels of a target gene as a marker for HOXB4 activity, which was then normalized to target gene mRNA levels detected in the negative control cells (no treatment), to obtain a "Fold over control" value. Total RNA levels (ng/ μ L) were also measured as a marker for cell viability. Results are shown in **Table 14.4** and **Figure 47**.

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Table 14.4: Data from **Figure 47**

Cargo / shuttle agent	Shuttle conc. (μ M)	HOXB4-WT Conc. (μ M)	Exposure time	Fold over control (mean $\pm$ St. Dev)	Total RNA in ng/ μ L (mean $\pm$ St. Dev)
No treatment ("Ctrl")	0	0	-	1 $\pm$ 0.09	240.3 $\pm$ 8.9
His-CM18-PTD4 alone ("FS")	0	1.5	2.5 h	2.5 $\pm$ 0.3	303.9 $\pm$ 7.6
HOXB4-WT alone ("TF")	0.8	0	2.5 h	1 $\pm$ 0.11	251.9 $\pm$ 11.9
His-CM18-PTD4 + HOXB4-WT	0.8	1.5	2.5 h	44.5 $\pm$ 0.09	182 $\pm$ 5.97
TAT-KALA + HOXB4-WT				5.1 $\pm$ 0.21	222.4 $\pm$ 12.5
EB1-PTD4 + HOXB4-WT				6.4 $\pm$ 0.3	240.4 $\pm$ 4.71
His-C(LLKK)3C-PTD4 + HOXB4-WT				9.8 $\pm$ 0.19	175.3 $\pm$ 11.25
His-CM18-PTD4-His + HOXB4-WT				28.1 $\pm$ 2.61	91.4 $\pm$ 3.92

Example 15:***In vivo* GFP-NLS delivery in rat parietal cortex by His-CM18-PTD4**

10 The ability of the shuttle agent His-CM18-PTD4 to deliver GFP-NLS *in vivo* in the nuclei of rat brain cells was tested.

In separate sterile 1.5-mL tubes, shuttle agent His-CM18-PTD4 was diluted in sterile distilled water at room temperature. GFP-NLS, used as cargo protein, was then added to the shuttle agent and, if necessary, sterile PBS was added to obtain the desired concentrations of shuttle agent and cargo in a sufficient final volume for injection in
 15 rat brain (e.g., 5 μ L per each injection brain site). The shuttle agent/cargo mixture was then immediately used for experiments. One negative control was included for the experiment, which corresponds to the injection of the GFP-NLS alone.

Bilateral injections were performed in the parietal cortex of three rats. In the left parietal cortex (ipsilateral), a mix composed of the shuttle agent (20 μ M) and the GFP-NLS (20 μ M) was injected, and in the right parietal cortex

(contralateral), only the GFP-NLS (20 μ M) was injected as a negative control. For surgical procedures, mice were anesthetized with isoflurane. Then the animal was placed in a stereotaxic frame, and the skull surface was exposed. Two holes were drilled at the appropriate sites to allow bilateral infusion of the shuttle/cargo mix or GFP-NLS alone (20 μ M) with 5- μ L Hamilton syringe. Antero-posterior (AP), lateral (L), and dorso-ventral (DV) coordinates were taken relative to the bregma: (a) AP +0.48 mm, L $\pm$ 3 mm, V – 5 mm; (b) AP – 2 mm, L $\pm$ 1.3 mm, V – 1.5 mm; (c) AP – 2.6 mm, L $\pm$ 1.5 mm, V – 1.5 mm. The infused volume of the shuttle/cargo mix or cargo alone was 5 μ L per injection site and the injection was performed for 10 minutes. After that, experimenter waited 1 min before removing the needle from the brain. All measures were taken before, during, and after surgery to minimize animal pain and discomfort. Animals were sacrificed by perfusion with paraformaldehyde (4%) 2 h after surgery, and brain were collected and prepared for microscopy analysis. Experimental procedures were approved by the Animal Care Committee in line with guidelines from the Canadian Council on Animal Care.

Dorso-ventral rat brain slices were collected and analysed by fluorescence microscopy and results are shown in **Figure 48** at (A) 4x, (C) 10x and (D) 20x magnifications. The injection site is located in the deepest layers of the parietal cortex (PCx). In the presence of the His-CM18-PTD4 shuttle, the GFP-NLS diffused in cell nuclei of the PCx, of the Corpus Callus (Cc) and of the striatum (Str) (White curves mean limitations between brains structures). Panel B shows the stereotaxic coordinates of the injection site (black arrows) from the rat brain atlas of Franklin and Paxinos. The injection of GFP-NLS in presence of His-CM18-PTD4 was performed on the left part of the brain, and the negative control (an injection of GFP-NLS alone), was done on the contralateral site. The black circle and connected black lines in panel B show the areas observed in the fluorescent pictures (A, C and D).

This experiment demonstrated the cell delivery of the cargo GFP-NLS after its stereotaxic injection in the rat parietal cortex in the presence of the shuttle agent His-CM18-PTD4. Results show the delivery of the GFP-NLS in the nucleus of cells from the deeper layers of the parietal cortex (injection site) to the corpus callus and the dorsal level of the striatum (putamen). In contrast, the negative control in which GFP-NLS is only detectable locally around the injection site. This experiment shows that shuttle agent induced nuclear delivery of the cargo in the injection site (parietal cortex) and its diffusion through both neighboring brain areas (corpus callus and striatum rat brain).

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CLAIMS:

1. A synthetic peptide comprising an endosome leakage domain (ELD) operably linked to a cell penetrating domain (CPD), or an ELD operably linked to a histidine-rich domain and a CPD.
- 5 2. A polypeptide-based shuttle agent comprising an endosome leakage domain (ELD) operably linked to a cell penetrating domain (CPD), or an ELD operably linked to a histidine-rich domain and a CPD, for use in increasing the transduction efficiency of an independent polypeptide cargo to the cytosol of a target eukaryotic cell.
- 10 3. The synthetic peptide or polypeptide-based shuttle agent of claim 1 or 2, wherein said synthetic peptide or polypeptide-based shuttle agent:
 - (a) comprises a minimum length of 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30 amino acid residues and a maximum length of 35, 40, 45, 50, 55, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, or 150 amino acid residues;
 - 15 (b) has a predicted net charge of at least +6, +7, +8, +9, +10, +11, +12, +13, +14, or +15 at physiological pH;
 - (c) is soluble in aqueous solution; or
 - (d) any combination of (a) to (c).
- 20 4. The synthetic peptide or polypeptide-based shuttle agent of any one of claims 1 to 3, wherein:
 - (a) said ELD is or is from: an endosomolytic peptide; an antimicrobial peptide (AMP); a linear cationic alpha-helical antimicrobial peptide; a Cecropin-A/Melittin hybrid (CM series) peptide; pH-dependent membrane active peptide (PAMP); a peptide amphiphile; a peptide derived from the N terminus of the HA2 subunit of influenza hemagglutinin (HA); CM18; Diphtheria toxin T domain (DT); GALA; PEA;
 - 25 INF-7; LAH4; HGP; H5WYG; HA2; EB1; VSVG; *Pseudomonas* toxin; melittin; KALA; JST-1; C(LLKK)₃C; G(LLKK)₃G; or any combination thereof;
 - (b) said CPD is or is from: a cell-penetrating peptide or the protein transduction domain from a cell-penetrating peptide; TAT; PTD4; Penetratin (Antennapedia); pVEC; M918; Pep-1; Pep-2; Xentry; arginine stretch; transportan; SynB1; SynB3; or any combination thereof;
 - 30 (c) said histidine-rich domain is a stretch of at least 3, at least 4, at least 5, or at least 6 amino acids comprising at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, or at least 90% histidine residues; and/or comprises at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, or at least 9 consecutive histidine residues; or
 - (d) any combination of (a) to (c).

5. The synthetic peptide or polypeptide-based shuttle agent of any one of claims 1 to 4, wherein said synthetic peptide or polypeptide-based shuttle agent comprises:
- (a) an ELD comprising the amino acid sequence of any one of **SEQ ID NOs: 1-15, 63, or 64**, or a variant or fragment thereof having endosomolytic activity;
 - (b) a CPD comprising the amino acid sequence of any one of **SEQ ID NOs: 16-27 or 65**, or a variant or fragment thereof having cell penetrating activity;
 - (c) a histidine-rich domain having at least 2, at least 3, at least 4, at least 5, or at least 6 consecutive histidine residues; or
 - (d) of any combination of (a) to (c).
6. The synthetic peptide or polypeptide-based shuttle agent of any one of claims 1 to 5, wherein said domains are operably linked via one or more linker domains.
7. The synthetic peptide or polypeptide-based shuttle agent of any one of claims 1 to 6, wherein said synthetic peptide or polypeptide-based shuttle agent comprises at least two different types of CPDs and/or ELDs.
8. The synthetic peptide or polypeptide-based shuttle agent of any one of claims 1 to 7, wherein said synthetic peptide or polypeptide-based shuttle agent comprises:
- (a) an ELD which is CM18, KALA, or C(LLKK)₃C having the amino acid sequence of **SEQ ID NO: 1, 14, or 63**, or a variant thereof having at least 85%, 90%, or 95% identity to **SEQ ID NO: 1, 14, or 63**, and having endosomolytic activity;
 - (b) a CPD which is TAT or PTD4 having the amino acid sequence of **SEQ ID NO: 17 or 65**, or a variant thereof having at least 85%, 90%, or 95% identity to **SEQ ID NO: 17 or 65** and having cell penetrating activity; or Penetratin having the amino acid sequence of **SEQ ID NO: 18**, or a variant thereof having at least 85%, 90%, or 95% identity to **SEQ ID NO: 18** and having cell penetrating activity;
 - (c) a histidine-rich domain comprising at least 6 consecutive histidine residues; or
 - (d) any combination of (a) to (c).
9. The synthetic peptide or polypeptide-based shuttle agent of any one of claims 1 to 8, wherein said synthetic peptide or polypeptide-based shuttle agent comprises or consists of the amino acid sequence of any one of **SEQ ID NOs: 57-59, 66-72, or 82-102**, or a functional variant thereof having at least 85%, 90%, or 95% identity to any one of **SEQ ID NOs: 57-59, 66-72, or 82-102**.

10. The synthetic peptide or polypeptide-based shuttle agent of any one of claims 1 to 9, wherein said synthetic peptide or polypeptide-based shuttle agent is non-toxic and/or is metabolizable.
11. A composition comprising:
- 5 (a) the synthetic peptide or polypeptide-based shuttle agent as defined in any one of claims 1 to 10, and a further independent synthetic peptide comprising a histidine-rich domain and a CPD; and/or
- (b) a cocktail of at least 2, at least 3, at least 4, or at least 5 different types of the synthetic peptides or polypeptide-based shuttle agents as defined in any one of claims 1 to 10.
- 10 12. Use of the synthetic peptide, polypeptide-based shuttle agent, or composition as defined in any one of claims 1 to 11, for delivering an independent polypeptide cargo to the cytosol of a target eukaryotic cell.
13. A method for increasing the transduction efficiency of a polypeptide cargo to the cytosol of a target eukaryotic cell, said method comprising contacting the target eukaryotic cell with the synthetic peptide, polypeptide-
- 15 based shuttle agent, or composition as defined in any one of claims 1 to 11, and the polypeptide cargo.
14. A kit for increasing the transduction efficiency of a polypeptide cargo to the cytosol of a target eukaryotic cell, said kit comprising the synthetic peptide, polypeptide-based shuttle agent, or composition as defined in any one of claims 1 to 11, and a suitable container.
- 20 15. The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of any one of claims 1 to 14, for use in increasing the transduction efficiency of a polypeptide cargo to the cytosol of a target eukaryotic cell in the presence of serum.
- 25 16. The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of any one of claims 2 to 15, wherein said polypeptide cargo:
- (a) comprises or lacks a CPD or a CPD as defined in claim 4(b);
- (b) is a recombinant protein;
- (c) comprises a subcellular targeting domain;
- 30 (d) is complexed with a DNA and/or RNA molecule; or
- (e) any combination of (a) to (d).
17. The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of claim 16, wherein said subcellular targeting domain is:

- (a) a nuclear localization signal (NLS);
- (b) a nucleolar signal sequence;
- (c) a mitochondrial signal sequence; or
- (d) a peroxisome signal sequence.

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18. The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of claim 17, wherein:

- (a) said NLS is from: E1a, T-Ag, c-myc, T-Ag, op-T-NLS, Vp3, nucleoplasmin, histone 2B, Xenopus N1, PARP, PDX-1, QKI-5, HCDA, H2B, v-Rel, Amida, RanBP3, Pho4p, LEF-1, TCF-1, BDV-P, TR2, SOX9, or Max;
- (b) said nucleolar signal sequence is from BIRC5 or RECQL4;
- (c) said mitochondrial signal sequence is from Tim9 or Yeast cytochrome c oxidase subunit IV; or
- (d) said peroxisome signal sequence is from PTS1.

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19. The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of any one of claims 2 to 18, wherein said polypeptide cargo is a transcription factor, a nuclease, a cytokine, a hormone, a growth factor, or an antibody.

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20. The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of claim 19, wherein:

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- (a) said transcription factor is: HOXB4, NUP98-HOXA9, Oct3/4, Sox2, Sox9, Klf4, c-Myc, MyoD, Pdx1, Ngn3, MafA, Blimp-1, Eomes, T-bet, FOXO3A, NF-YA, SALL4, ISL1, FoxA1, Nanog, Esrrb, Lin28, HIF1-alpha, Hlf, Runx1t1, Pbx1, Lmo2, Zfp37, Prdm5, Bcl-6, or any combination thereof; and/or
- (b) said nuclease is: an RNA-guided endonuclease, a CRISPR endonuclease, a type I CRISPR endonuclease, a type II CRISPR endonuclease, a type III CRISPR endonuclease, a type IV CRISPR endonuclease, a type V CRISPR endonuclease, a type VI CRISPR endonuclease, CRISPR associated protein 9 (Cas9), Cpf1, a zinc-finger nuclease (ZFNs), a Transcription activator-like effector nucleases (TALENs), a homing endonuclease, a meganuclease, or any combination thereof.

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21. The synthetic peptide, polypeptide-based shuttle agent, composition, use, method or kit of any one of claims 1 to 20, for use in cell therapy, genome editing, adoptive cell transfer, and/or regenerative medicine.

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22. The shuttle agent, shuttle system, composition, use, method, or kit of any one of claims 2 to 21, wherein said target eukaryotic cell is a stem cell, a primary cell, an immune cell, a T cell, or a dendritic cell.

23. A eukaryotic cell comprising the synthetic peptide or polypeptide-based shuttle agent as defined in any one of claims 1 to 10, or the composition of claim 11.

5 24. The eukaryotic cell of claim 23, wherein said cell further comprises an independent polypeptide cargo delivered intracellularly by said synthetic peptide or polypeptide-based shuttle agent.

25. A method for delivering an independent polypeptide cargo to the cytosol of a target eukaryotic cell, said method comprising contacting said target eukaryotic cell with the synthetic peptide or polypeptide-based shuttle agent as defined in any one of claims 1 to 10, or the composition of claim 11; and an independent polypeptide cargo to be delivered intracellularly by said synthetic peptide or polypeptide-based shuttle agent.

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26. The eukaryotic cell of claim 23 or 24, or the method of claim 25, wherein said independent polypeptide cargo is as defined in any one of claims 16 to 20.

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27. The eukaryotic cell of claim 24 or 26, or the method of claim 25 or 26, wherein said independent polypeptide cargo is as defined in any one of claims 16 to 20.

28. The eukaryotic cell of claim 23, 24, 26 or 27, or the method of claim 25, 26, or 27, wherein said eukaryotic cell is an animal cell, a mammalian cell, a human cell, a stem cell, a primary cell, an immune cell, a T cell, or a dendritic cell.

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