

CD3-DELTA A/EPSILON HETERODIMER SPECIFIC ANTIBODIES**CROSS-REFERENCE TO RELATED APPLICATIONS**

[0001] This application claims priority benefit of the filing date of US Provisional Patent Application No. 62/610,764, filed on December 27, 2017, the disclosure of which application is herein incorporated by reference in its entirety.

FIELD OF THE INVENTION

[0002] The present invention concerns novel human CD3 antigen-binding polypeptides and their preparation and use in the treatment and/or diagnosis of various diseases, as well as bispecific antibody molecules capable of activating immune effector cells and their use in diagnosis and/or treatment of various diseases.

BACKGROUND

[0003] The body's immune system serves as a defense against infection, injury and cancer. Two separate but interrelated systems, humoral and cellular immune systems, work together to protect the body. The humoral system is mediated by soluble factors, named antibodies, which neutralize products recognized as being foreign by the body. In contrast, the cellular system involves cells, such as T cells and macrophages, which remove and neutralize foreign invaders.

[0004] The activation of T cells is critical for the stimulation of immune responses. T cells exhibit immunological specificity and direct most of the cellular immune responses. Although T cells do not secrete antibodies, they are required for the secretion of antibodies by B lymphocytes. T cell activation requires the participation of a number of cell surface molecules, such as the T cell receptor complex, and CD4 or CD8 molecules. The antigen-specific T cell receptor (TcR) is composed of a disulfide-linked heterodimer, membrane glycoprotein with chains, alpha and beta (α and β), or gamma and delta (γ and δ). The TcR is non-covalently linked with a complex of invariant proteins, designated CD3.

[0005] T cells are known to exert potent antitumor effects in numerous experimental settings. Antibodies capable of effectively recruiting T cells against tumor cells have been available as bispecific antibodies, for example directed to tumor-associated antigens (TAAs) and agonistic T-cell membrane proteins, such as the TCR/CD3 complex and CD28. These bispecific antibodies are capable of activating T cells, irrespective of their TCR specificity, resulting in specific lysis of cells carrying the respective TAAs.

[0006] However, while anti-CD3 bispecific antibodies can redirect T-cell-mediated lysis toward malignant cells, clinical trials with CD3-based bsAbs have shown high toxicity in patients. Non-specific T-cell activation from bsAbs can occur in an antigen-independent manner due to the Fc/Fc

receptor (FcR) interaction, or in an antigen-dependent manner when antigen is expressed on both normal and tumor cells. Both mechanisms may have been responsible for the toxicity observed in prior clinical studies. (See for example, Link et al. (1998) *Int. J. Cancer* 77(2):251-6; Durben et al. *Molecular Therapy* (2015); 23 4, 648–655). Because of the resulting cytokine release syndrome, there have been significant blocks to the development of these antibodies for therapeutic purposes.

[0007] The interaction of the T cell receptor (TCR) with its peptide-MHC ligand determines the activity of a T cell. The binding characteristics of this interaction has been studied in great detail and shown to control T cell function. The strength and nature of the TCR-peptide/MHC interaction determines whether T cells exert effector functions or are inactivated and deleted. Antibodies against CD3 activate T cells by changing the conformation of the CD3ε chain and depending on the epitope may have either agonistic or antagonistic effects on T cells (Yoon et al., 1994 *Immunity* 1:563-569). In light of the significant side-effects of many T cell agonists it may be preferred to maintain potent anti-tumor effects while reducing the release of pro-inflammatory cytokines. However, partial agonistic anti-CD3 antibodies may alter the CD3ε chain sub-optimally resulting in ineffective signaling, and most anti-CD3 antibodies are full agonists for both pathways. It is unclear whether these effector functions can be separated. Many existing anti-CD3 antibodies (for example SP-34, UCHT1, OKT3) have affinities in the range of 1-50 nM KD, however this may not be optimal for therapeutic use.

[0008] CD3 specific antibodies, and bispecific antibodies derived therefrom are provided by the invention.

Publications

[0009] CD3 antibodies are disclosed, for example, in U.S. Pat. Nos. 5,585,097; 5,929,212; 5,968,509; 6,706,265; 6,750,325; 7,381,803; 7,728,114. Bispecific antibodies with CD3 binding specificity are disclosed, for example, in U.S. Pat. Nos. 7,262,276; 7,635,472; 7,862,813; and 8,236,308, each herein specifically incorporated by reference. CD3 binding antibody sequences are provided in co-pending application PCT US2017/038377, herein specifically incorporated by reference.

SUMMARY

[0010] Compositions and methods of use thereof are provided for antibodies that bind to and activate signaling through CD3, e.g. activation of CD3⁺ T cells. The antibodies are characterized by binding to the CD3 epitope bound by the F2B antibody, which may be referred to herein as the F2B epitope. The F2B antibody comprises the set of CDR sequences of SEQ ID NO:1, and the fixed light chain sequences of SEQ ID NO:19. In some embodiments the antibody that binds to the F2B epitope comprises a heavy chain variable region sequence other than a sequence set forth in SEQ ID NO:1-18.

- [0011] Antibodies binding to the F2B epitope provide significant benefits in terms of biological activity. The antibodies minimize toxic cytokine release while maintaining effective tumor cell lysis. In some embodiments, anti-CD3 antibodies binding to the F2B epitope are characterized by a reduced propensity to induce cytokine release, upon binding to a competent T cell, e.g. for release of IL-2 and IFN γ . Without being bound by the theory, it is believed that binding to the specific epitope recognized by F2B provides for the unique and beneficial properties of antibodies described herein.
- [0012] Antibodies binding to the F2B epitope may be selected for a binding affinity for CD3 ranging from around about 10^{-6} to around about 10^{-11} . Anti-CD3 antibodies that have affinities (KD) of 50 nM or greater, 100 nM or greater, 500 nM or greater, or 1 μ M or greater can be desirable to more closely mimic the TCR/MHC interaction and minimize toxic cytokine release while maintaining effective tumor cell lysis. Antibodies may be selected that induce a cytokine release that is not more than about 200% of the maximum cytokine release observed with the F2B antibody, and may be not more than about 150%, not more than 125%, not more than 100%, and may be less than the maximum observed for F2B in a comparative assay.
- [0013] Antibodies may be selected that induce maximum IL-2 and IL-10 release in a comparative *in vitro* assay of not more than 20%, not more than 30%, and not more than 50% of the maximum release of a control anti-CD3 antibody, such as OKT-3 or TNB-383B.
- [0014] The F2B epitope is characterized by binding to at least one residue selected from CD3 epsilon (SEQ ID NO:23): K73 and S83; and CD3 delta (SEQ ID NO:24) K82 and C93. In some embodiments the epitope comprises the region of CD3 epsilon defined by K73, N74, I75, G76, S77, D78, E79, D80, H81, L82, S83. In some embodiments the epitope comprises one or both of K73 and S83. In some embodiments the epitope comprises the region of CD3 delta defined by K82, E83, S84, T85, V86, Q87, V88, H89, Y90, R91, M92, C93. In some embodiments the epitope comprises one or both of K82 and C93. In some embodiments the F2B epitope comprises a conformational epitope involving residues of both CD3 delta and CD3 epsilon. In some embodiments the conformational epitope comprises each of residues CD3 $\square$ K73 and S83; CD3 $\square$ K82 and C93. In some embodiments antibodies that bind to the F2B epitope are not cross-reactive with cynomolgus CD3 protein.
- [0015] In some embodiments, an antibody that binds to the F2B epitope is determined by competition assays between the antibodies disclosed herein and other antibodies. In some embodiments, the antibodies bind to a specific residue of CD3 that reduces cytokine release.
- [0016] In some embodiments, bispecific or multispecific antibodies are provided, which comprise at least a heavy chain variable region from an antibody that binds to the F2B epitope. Bispecific antibodies comprise at least the heavy chain variable region of an antibody specific for a protein other than CD3, and may comprise a heavy and light chain variable region. In some such embodiments, the second antibody specifically binds to a tumor associated antigen, a targeting antigen, e.g. integrins, *etc.*, a pathogen antigen, a checkpoint protein, and the like. Various formats of bispecific antibodies

are within the ambit of the invention, including without limitation single chain polypeptides, two chain polypeptides, three chain polypeptides, four chain polypeptides, and multiples thereof.

[0017] In some embodiments, an F2B epitope-binding antibody of the invention comprises a CD3-binding variable region, paired with a light chain. In some embodiments the light chain comprises the variable region sequence set forth in SEQ ID NO:19, or a variable region comprising the set of CDR sequences in SEQ ID NO:19 and framework sequences. Various Fc sequences find use, including without limitation human IgG1, IgG2a, IgG2b, IgG3, IgG4, etc. In some embodiments, the second arm of the bispecific antibody comprises a variable region that specifically binds to a tumor-associated antigen. In some embodiments, the second arm of the bispecific antibody comprises a variable region that specifically binds to BCMA.

[0018] In other embodiments, pharmaceutical compositions are provided, comprising at least a CD3-binding VH domain of the invention, e.g. a monospecific, bispecific, etc. antibody or antibody-like protein comprising at least a CD3-binding VH domain of the invention; and a pharmaceutically acceptable excipient. The composition may be lyophilized, suspended in solution, etc. and may be provided in a unit dose formulation.

[0019] In some embodiments, a method is provided for treatment of cancer, the method comprising administering to an individual in need thereof an effective dose of a mono-specific, bi-specific, *etc.* antibody of the invention. Where the antibody is bispecific, a second antigen-binding site may specifically bind a tumor antigen, a checkpoint protein, etc. In various embodiments, the cancer is selected from the group consisting of ovarian cancer, breast cancer, gastrointestinal, brain cancer, head and neck cancer, prostate cancer, colon cancer, lung cancer, leukemia, lymphoma, sarcoma, carcinoma, neural cell tumors, squamous cell carcinomas, germ cell tumors, metastases, undifferentiated tumors, seminomas, melanomas, myelomas, neuroblastomas, mixed cell tumors, and neoplasias caused by infectious agents.

[0020] In some embodiments, a method is provided for treatment of infectious disease, the method comprising administering to an individual in need thereof an effective dose of a mono-specific, bi-specific, etc. antibody of the invention. Where the antibody is bispecific, a second antigen-binding site may specifically bind a pathogen antigen, e.g. bacteria, viruses or parasites.

[0021] In other embodiments, a method is provided for the production of a bispecific antibody of the present invention comprising expressing the antibody sequences, e.g. one or more light chain encoding sequences, one or more heavy chain encoding sequences, in a single host cell. In various embodiments, the host cell may be a prokaryotic or a eukaryotic cell, such as a mammalian cell.

[0022] Aspects of the invention include methods for producing an antigen-binding protein as described herein, comprising growing a host cell under conditions permissive for expression of the protein, and isolating the protein from the cell and/or a cell culture medium.

- [0023] Aspects of the invention include methods of treatment, comprising administering to an individual an effective dose of an antigen-binding protein as described herein, or a pharmaceutical composition as described herein.
- [0024] Aspects of the invention relate to use of an antigen-binding protein as described herein in the preparation of a medicament for the treatment of a disease.
- [0025] Aspects of the invention include antigen-binding proteins as described herein for use in the treatment of a disease.
- [0026] In some embodiments, a method or use involves a human subject (e.g., a individual who is a human).

BRIEF DESCRIPTION OF THE DRAWINGS

- [0027] The invention is best understood from the following detailed description when read in conjunction with the accompanying drawings. The patent or application file contains at least one drawing executed in color. Copies of this patent or patent application publication with color drawing(s) will be provided by the Office upon request and payment of the necessary fee. It is emphasized that, according to common practice, the various features of the drawings are not to-scale. On the contrary, the dimensions of the various features are arbitrarily expanded or reduced for clarity. Included in the drawings are the following figures.
- [0028] **FIGS. 1A-1C.** **FIG. 1A** show an alignment of CDR1, 2 and 3 regions of members of antibody family 2 of SEQ ID NO:1-18, which specifically bind to human CD3, corresponding to residues 26-33; 51-58; and 97-112. **FIG. 1B** shows the CDR1, 2 and 3 regions of the fixed light chain (SEQ ID NO:19); and an exemplary anti-BCMA sequence (SEQ ID NO:20 and SEQ ID NO:21). **FIG. 1C** provides the CDR sequences of a reference anti-CD3 antibody (SEQ ID NO:22), ID 304704.
- [0029] **FIG. 2.** Schematic representation of molecule TNB-383B with the anti-CD3 arm (CD3_F2B or ID:312557) and a high affinity anti-BCMA arm.
- [0030] **FIG. 3.** Dose-response curves of cytokine release by PBMCs treated with TNB-383B and a positive control. Pre-cultured PBMCs were stimulated with a positive control (black crosses) or with TNB-383B (black square) at increasing concentrations. The positive control in this experiment was a bispecific anti-CD3/anti-BCMA antibody (TNB-384B). The anti-CD3 arm of this bispecific antibody recognizes a different epitope on human CD3 with high affinity ($K_D=30\text{nM}$, also named), but the anti-BCMA arm is identical to the anti-BCMA arm of TNB-383B. This positive control showed a similar cytokine secretion profile as OKT3 (data not shown).
- [0031] **FIG. 4.** Activation profile of T cell subsets after overnight stimulation with TNB-383B and a positive control (TNB-384B). Responses at plateau concentrations of positive control (132 ng/ml, black) and TNB-383B (1320 ng/ml, pattern) are shown. Cells were analyzed after a 24 hour incubation step as described in Example 1. CD69 expression was analyzed (percentage positive cells

and mean fluorescence intensity (MFI) of positive cells) upon gating on T cell subsets. Graphs show the median values and ranges obtained from three donors.

[0032] **FIG. 5.** Combined sequence coverage of DEPC labeled CD3 delta and CD3 epsilon after proteolytic digests and analysis by LC-MS/MS. Shaded sequences indicate presence of peptides with DEPC modifications. Uncovered sequences are buried and not accessible to DEPC modification.

[0033] **FIG. 6.** Number of Endo-GluC derived peptides with significant decrease of DEPC labeling in the presence of mAb F2B. Peptides with an at least 15-fold difference in degree of labeling are mapped onto the location of the CD3 delta (D) and CD3 epsilon (E) ECDs, respectively (shaded).

[0034] **FIG. 7.** CD3 epsilon derived proteolytic peptides and their impact on DEPC labeling. In bold and underlined are residues that were found to be labeled. The biggest impact was observed for Lys 73 and Lysine 85.

[0035] **FIG. 8.** Epitopes of mAb F2B of the CD3 delta subunit identified by DEPC labeling.

[0036] **FIG. 9.** Ribbon drawing of the X-ray structure derived CD3 delta/epsilon complex. Residues important for interaction with mAb F2B are highlighted through space filling.

[0037] **FIG. 10.** Mapping of CD3 epsilon epitopes obtained with DEPC labeling.

DETAILED DESCRIPTION

[0038] To facilitate an understanding of the invention, a number of terms are defined below.

[0039] Before the present active agents and methods are described, it is to be understood that this invention is not limited to the particular methodology, products, apparatus and factors described, as such methods, apparatus and formulations may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by appended claims.

[0040] It must be noted that as used herein and in the appended claims, the singular forms "a," "and," and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a drug candidate" refers to one or mixtures of such candidates, and reference to "the method" includes reference to equivalent steps and methods known to those skilled in the art, and so forth.

[0041] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. All publications mentioned herein are incorporated herein by reference for the purpose of describing and disclosing devices, formulations and methodologies which are described in the publication and which might be used in connection with the presently described invention.

[0042] Where a range of values is provided, it is understood that each intervening value, to the tenth of the unit of the lower limit unless the context clearly dictates otherwise, between the upper and lower limit of that range and any other stated or intervening value in that stated range is encompassed within the invention. The upper and lower limits of these smaller ranges may independently be

included in the smaller ranges is also encompassed within the invention, subject to any specifically excluded limit in the stated range. Where the stated range includes one or both of the limits, ranges excluding either both of those included limits are also included in the invention.

[0043] In the following description, numerous specific details are set forth to provide a more thorough understanding of the present invention. However, it will be apparent to one of skill in the art that the present invention may be practiced without one or more of these specific details. In other instances, well-known features and procedures well known to those skilled in the art have not been described in order to avoid obscuring the invention.

[0044] Generally, conventional methods of protein synthesis, recombinant cell culture and protein isolation, and recombinant DNA techniques within the skill of the art are employed in the present invention. Such techniques are explained fully in the literature, see, *e.g.*, Maniatis, Fritsch & Sambrook, *Molecular Cloning: A Laboratory Manual* (1982); Sambrook, Russell and Sambrook, *Molecular Cloning: A Laboratory Manual* (2001); Harlow, Lane and Harlow, *Using Antibodies: A Laboratory Manual: Portable Protocol No. I*, Cold Spring Harbor Laboratory (1998); and Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory; (1988).

Definitions

[0045] By “comprising” it is meant that the recited elements are required in the composition/method/kit, but other elements may be included to form the composition/method/kit etc. within the scope of the claim.

[0046] By “consisting essentially of”, it is meant a limitation of the scope of composition or method described to the specified materials or steps that do not materially affect the basic and novel characteristic(s) of the subject invention.

[0047] By “consisting of”, it is meant the exclusion from the composition, method, or kit of any element, step, or ingredient not specified in the claim.

[0048] The terms "treatment", "treating" and the like are used herein to generally mean obtaining a desired pharmacologic and/or physiologic effect. The effect may be prophylactic in terms of completely or partially preventing a disease or symptom thereof and/or may be therapeutic in terms of a partial or complete cure for a disease and/or adverse effect attributable to the disease. "Treatment" as used herein covers any treatment of a disease in a mammal, and includes: (a) preventing the disease from occurring in a subject which may be predisposed to the disease but has not yet been diagnosed as having it; (b) inhibiting the disease, *i.e.*, arresting its development; or (c) relieving the disease, *i.e.*, causing regression of the disease. The therapeutic agent may be administered before, during or after the onset of disease or injury. The treatment of ongoing disease, where the treatment stabilizes or reduces the undesirable clinical symptoms of the patient, is of particular interest. Such treatment is desirably performed prior to complete loss of function in the affected tissues. The subject therapy may

be administered during the symptomatic stage of the disease, and in some cases after the symptomatic stage of the disease.

[0049] A "therapeutically effective amount" is intended for an amount of active agent which is necessary to impart therapeutic benefit to a subject. For example, a "therapeutically effective amount" is an amount which induces, ameliorates or otherwise causes an improvement in the pathological symptoms, disease progression or physiological conditions associated with a disease or which improves resistance to a disorder.

[0050] The terms "subject," "individual," and "patient" are used interchangeably herein to refer to a mammal being assessed for treatment and/or being treated. In an embodiment, the mammal is a human. The terms "subject," "individual," and "patient" encompass, without limitation, individuals having cancer, individuals with autoimmune diseases, with pathogen infections, and the like. Subjects may be human, but also include other mammals, particularly those mammals useful as laboratory models for human disease, *e.g.* mouse, rat, *etc.*

[0051] The terms "cancer," "neoplasm," and "tumor" are used interchangeably herein to refer to cells which exhibit autonomous, unregulated growth, such that they exhibit an aberrant growth phenotype characterized by a significant loss of control over cell proliferation. Cells of interest for detection, analysis, or treatment in the present application include precancerous (*e.g.*, benign), malignant, pre-metastatic, metastatic, and non-metastatic cells. Cancers of virtually every tissue are known. The phrase "cancer burden" refers to the quantum of cancer cells or cancer volume in a subject. Reducing cancer burden accordingly refers to reducing the number of cancer cells or the cancer volume in a subject. The term "cancer cell" as used herein refers to any cell that is a cancer cell or is derived from a cancer cell *e.g.* clone of a cancer cell. Many types of cancers are known to those of skill in the art, including solid tumors such as carcinomas, sarcomas, glioblastomas, melanomas, lymphomas, myelomas, *etc.*, and circulating cancers such as leukemias, including specifically B cell leukemias, T cell leukemias, *etc.* Examples of cancer include but are not limited to, ovarian cancer, breast cancer, colon cancer, lung cancer, prostate cancer, hepatocellular cancer, gastric cancer, pancreatic cancer, cervical cancer, ovarian cancer, liver cancer, bladder cancer, cancer of the urinary tract, thyroid cancer, renal cancer, carcinoma, melanoma, head and neck cancer, and brain cancer.

[0052] "Antibody-dependent cell-mediated cytotoxicity" and "ADCC" refer to a cell-mediated reaction in which nonspecific cytotoxic cells that express Fc receptors, such as natural killer cells, neutrophils, and macrophages, recognize bound antibody on a target cell and cause lysis of the target cell. ADCC activity may be assessed using methods, such as those described in U.S. Pat. No. 5,821,337. ADCP refers to antibody dependent cell-mediated phagocytosis.

[0053] "Effector cells" are leukocytes which express one or more constant region receptors and perform effector functions.

- [0054] A "cytokine" is a protein released by one cell to act on another cell as an intercellular mediator. Cytokines of interest include, without limitation, cytokines released from activated T cells, for example IL-2, IFN γ , etc.
- [0055] "Non-immunogenic" refers to a material that does not initiate, provoke or enhance an immune response where the immune response includes the adaptive and/or innate immune responses.
- [0056] The term "isolated" means that the material is removed from its original environment (e.g., the natural environment if it is naturally occurring). For example, a naturally-occurring polynucleotide or polypeptide present in a living animal is not isolated, but the same polynucleotide or polypeptide, separated from some or all of the coexisting materials in the natural system, is isolated. Such polynucleotides could be part of a vector and/or such polynucleotides or polypeptides could be part of a composition, and still be isolated in that such vector or composition is not part of its natural environment.
- [0057] "Pharmaceutically acceptable excipient" means an excipient that is useful in preparing a pharmaceutical composition that is generally safe, non-toxic, and desirable, and includes excipients that are acceptable for veterinary use as well as for human pharmaceutical use. Such excipients can be solid, liquid, semisolid, or, in the case of an aerosol composition, gaseous.
- [0058] "Pharmaceutically acceptable salts and esters" means salts and esters that are pharmaceutically acceptable and have the desired pharmacological properties. Such salts include salts that can be formed where acidic protons present in the compounds are capable of reacting with inorganic or organic bases. Suitable inorganic salts include those formed with the alkali metals, *e.g.*, sodium and potassium, magnesium, calcium, and aluminum. Suitable organic salts include those formed with organic bases such as the amine bases, *e.g.*, ethanolamine, diethanolamine, triethanolamine, tromethamine, N methylglucamine, and the like. Such salts also include acid addition salts formed with inorganic acids (*e.g.*, hydrochloric and hydrobromic acids) and organic acids (*e.g.*, acetic acid, citric acid, maleic acid, and the alkane- and arene-sulfonic acids such as methanesulfonic acid and benzenesulfonic acid). Pharmaceutically acceptable esters include esters formed from carboxy, sulfonyloxy, and phosphonoxy groups present in the compounds, *e.g.*, C₁₋₆ alkyl esters. When there are two acidic groups present, a pharmaceutically acceptable salt or ester can be a mono-acid-mono-salt or ester or a di-salt or ester; and similarly where there are more than two acidic groups present, some or all of such groups can be salified or esterified. Compounds named in this invention can be present in unsalified or unesterified form, or in salified and/or esterified form, and the naming of such compounds is intended to include both the original (unsalified and unesterified) compound and its pharmaceutically acceptable salts and esters. Also, certain compounds named in this invention may be present in more than one stereoisomeric form, and the naming of such compounds is intended to include all single stereoisomers and all mixtures (whether racemic or otherwise) of such stereoisomers.

[0059] The terms "pharmaceutically acceptable", "physiologically tolerable" and grammatical variations thereof, as they refer to compositions, carriers, diluents and reagents, are used interchangeably and represent that the materials are capable of administration to or upon a human without the production of undesirable physiological effects to a degree that would prohibit administration of the composition.

[0060] "Homology" between two sequences is determined by sequence identity. If two sequences, which are to be compared with each other, differ in length, sequence identity preferably relates to the percentage of the nucleotide residues of the shorter sequence which are identical with the nucleotide residues of the longer sequence. Sequence identity can be determined conventionally with the use of computer programs such as the Bestfit program (Wisconsin Sequence Analysis Package, Version 8 for Unix, Genetics Computer Group, University Research Park, 575 Science Drive Madison, Wis. 53711). Bestfit utilizes the local homology algorithm of Smith and Waterman, *Advances in Applied Mathematics* 2 (1981), 482-489, in order to find the segment having the highest sequence identity between two sequences. When using Bestfit or another sequence alignment program to determine whether a particular sequence has for instance 95% identity with a reference sequence of the present invention, the parameters are preferably so adjusted that the percentage of identity is calculated over the entire length of the reference sequence and that homology gaps of up to 5% of the total number of the nucleotides in the reference sequence are permitted. When using Bestfit, the so-called optional parameters are preferably left at their preset ("default") values. The deviations appearing in the comparison between a given sequence and the above-described sequences of the invention may be caused for instance by addition, deletion, substitution, insertion or recombination. Such a sequence comparison can preferably also be carried out with the program "fasta20u66" (version 2.0u66, September 1998 by William R. Pearson and the University of Virginia; see also W. R. Pearson (1990), *Methods in Enzymology* 183, 63-98, appended examples and <http://workbench.sdsc.edu/>). For this purpose, the "default" parameter settings may be used.

[0061] "Variant" refers to polypeptides having amino acid sequences that differ to some extent from a native sequence polypeptide. Ordinarily, amino acid sequence variants will possess at least about 80% sequence identity, more preferably, at least about 90% homologous by sequence. The amino acid sequence variants may possess substitutions, deletions, and/or insertions at certain positions within the reference amino acid sequence.

[0062] The term "vector," as used herein, is intended to refer to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments may be ligated. Another type of vector is a viral vector, wherein additional DNA segments may be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (e.g., non-episomal mammalian vectors) can be integrated into the genome of

a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operably linked. Such vectors are referred to herein as "recombinant expression vectors" (or simply, "recombinant vectors"). In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" may be used interchangeably as the plasmid is the most commonly used form of vector.

[0063] The term "host cell" (or "recombinant host cell"), as used herein, is intended to refer to a cell that has been genetically altered, or is capable of being genetically altered by introduction of an exogenous polynucleotide, such as a recombinant plasmid or vector. It should be understood that such terms are intended to refer not only to the particular subject cell but to the progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term "host cell" as used herein.

[0064] "Binding affinity" generally refers to the strength of the sum total of noncovalent interactions between a single binding site of a molecule (e.g., an antibody or other binding molecule) and its binding partner (e.g., an antigen or receptor). The affinity of a molecule X for its partner Y can generally be represented by the dissociation constant (K_d). Affinity can be measured by common methods known in the art, including those described herein. Low-affinity antibodies bind antigen (or receptor) weakly and tend to dissociate readily, whereas high-affinity antibodies bind antigen (or receptor) more tightly and remain bound longer.

[0065] Unless specifically indicated to the contrary, the term "conjugate" as described and claimed herein is defined as a heterogeneous molecule formed by the covalent attachment of one or more antibody fragment(s) to one or more polymer molecule(s), wherein the heterogeneous molecule is water soluble, i.e. soluble in physiological fluids such as blood, and wherein the heterogeneous molecule is free of any structured aggregate. A conjugate of interest is PEG. In the context of the foregoing definition, the term "structured aggregate" refers to (1) any aggregate of molecules in aqueous solution having a spheroid or spheroid shell structure, such that the heterogeneous molecule is not in a micelle or other emulsion structure, and is not anchored to a lipid bilayer, vesicle or liposome; and (2) any aggregate of molecules in solid or insolubilized form, such as a chromatography bead matrix, that does not release the heterogeneous molecule into solution upon contact with an aqueous phase. Accordingly, the term "conjugate" as defined herein encompasses the aforementioned heterogeneous molecule in a precipitate, sediment, bioerodible matrix or other solid capable of releasing the heterogeneous molecule into aqueous solution upon hydration of the solid.

[0066] The word "label" when used herein refers to a detectable compound or composition which is conjugated directly or indirectly to the antibody. The label may itself be detectable by itself (e.g., radioisotope labels or fluorescent labels) or, in the case of an enzymatic label, may catalyze chemical alteration of a substrate compound or composition which is detectable.

[0067] By "solid phase" is meant a non-aqueous matrix to which the antibody of the present invention can adhere. Examples of solid phases encompassed herein include those formed partially or entirely of glass (e.g. controlled pore glass), polysaccharides (e.g., agarose), polyacrylamides, polystyrene, polyvinyl alcohol and silicones. In certain embodiments, depending on the context, the solid phase can comprise the well of an assay plate; in others it is a purification column (e.g. an affinity chromatography column). This term also includes a discontinuous solid phase of discrete particles, such as those described in U.S. Pat. No. 4,275,149.

[0068] Antibodies, also referred to as immunoglobulins, conventionally comprise at least one heavy chain and one light, where the amino terminal domain of the heavy and light chains is variable in sequence, hence is commonly referred to as a variable region domain, or a variable heavy (VH) or variable light (VL) domain. The two domains conventionally associate to form a specific binding region, although as well be discussed here, specific binding can also be obtained with heavy chain only variable sequences, and a variety of non-natural configurations of antibodies are known and used in the art.

[0069] A "functional" or "biologically active" antibody or antigen-binding molecule (including heavy chain only antibodies and bispecific three-chain antibody-like molecules (TCAs) herein) is one capable of exerting one or more of its natural activities in structural, regulatory, biochemical or biophysical events. For example, a functional antibody or other binding molecule, e.g. TCA, may have the ability to specifically bind an antigen and the binding may in turn elicit or alter a cellular or molecular event such as signaling transduction or enzymatic activity. A functional antibody or other binding molecule, e.g. TCA, may also block ligand activation of a receptor or act as an agonist or antagonist. The capability of an antibody or other binding molecule, e.g. TCA, to exert one or more of its natural activities depends on several factors, including proper folding and assembly of the polypeptide chains.

[0070] The term "antibody" herein is used in the broadest sense and specifically covers monoclonal antibodies, polyclonal antibodies, monomers, dimers, multimers, multispecific antibodies (e.g., bispecific antibodies), heavy chain only antibodies, three chain antibodies, single chain Fv, nanobodies, etc., and also include antibody fragments, so long as they exhibit the desired biological activity (Miller et al (2003) Jour. of Immunology 170:4854-4861). Antibodies may be murine, human, humanized, chimeric, or derived from other species.

[0071] The term antibody may reference a full-length heavy chain, a full length light chain, an intact immunoglobulin molecule; or an immunologically active portion of any of these polypeptides, i.e., a polypeptide that comprises an antigen binding site that immunospecifically binds an antigen of a target of interest or part thereof, such targets including but not limited to, cancer cell or cells that produce autoimmune antibodies associated with an autoimmune disease. The immunoglobulin disclosed herein can be of any type (e.g., IgG, IgE, IgM, IgD, and IgA), class (e.g., IgG1, IgG2, IgG3, IgG4, IgA1 and IgA2) or subclass of immunoglobulin molecule, including engineered subclasses with

altered Fc portions that provide for reduced or enhanced effector cell activity. The immunoglobulins can be derived from any species. In one aspect, the immunoglobulin is of largely human origin.

[0072] The term “variable” refers to the fact that certain portions of the variable domains differ extensively in sequence among antibodies and are used in the binding and specificity of each particular antibody for its particular antigen. However, the variability is not evenly distributed throughout the variable domains of antibodies. It is concentrated in three segments called hypervariable regions both in the light chain and the heavy chain variable domains. The more highly conserved portions of variable domains are called the framework regions (FRs). The variable domains of native heavy and light chains each comprise four FRs, largely adopting a beta-sheet configuration, connected by three hypervariable regions, which form loops connecting, and in some cases forming part of, the beta-sheet structure. The hypervariable regions in each chain are held together in close proximity by the FRs and, with the hypervariable regions from the other chain, contribute to the formation of the antigen-binding site of antibodies (see Kabat et al (1991) Sequences of Proteins of Immunological Interest, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, Md.). The constant domains are not involved directly in binding an antibody to an antigen, but exhibit various effector functions, such as participation of the antibody in antibody dependent cellular cytotoxicity (ADCC).

[0073] The term “hypervariable region” when used herein refers to the amino acid residues of an antibody which are responsible for antigen-binding. The hypervariable region may comprise amino acid residues from a “complementarity determining region” or “CDR”, and/or those residues from a “hypervariable loop”. “Framework Region” or “FR” residues are those variable domain residues other than the hypervariable region residues as herein defined.

[0074] Exemplary CDR designations are shown herein, however one of skill in the art will understand that a number of definitions of the CDRs are commonly in use, including the Kabat definition (see “Zhao et al. A germline knowledge based computational approach for determining antibody complementarity determining regions.” Mol Immunol. 2010;47:694–700), which is based on sequence variability and is the most commonly used. The Chothia definition is based on the location of the structural loop regions (Chothia et al. “Conformations of immunoglobulin hypervariable regions.” Nature. 1989; 342:877–883). Alternative CDR definitions of interest include, without limitation, those disclosed by Honegger, “Yet another numbering scheme for immunoglobulin variable domains: an automatic modeling and analysis tool.” J Mol Biol. 2001;309:657–670; Ofra et al. “Automated identification of complementarity determining regions (CDRs) reveals peculiar characteristics of CDRs and B cell epitopes.” J Immunol. 2008;181:6230–6235; Almagro “Identification of differences in the specificity-determining residues of antibodies that recognize antigens of different size: implications for the rational design of antibody repertoires.” J Mol Recognit. 2004;17:132–143; and Padlan et al. “Identification of specificity-determining residues in antibodies.” Faseb J. 1995;9:133–139., each of which is herein specifically incorporated by reference.

[0075] The term “monoclonal antibody” as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to polyclonal antibody preparations, which include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. In addition to their specificity, the monoclonal antibodies are advantageous in that they may be synthesized uncontaminated by other antibodies. The modifier “monoclonal” indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method.

[0076] The antibodies herein specifically include “chimeric” antibodies in which a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies, so long as they exhibit the desired biological activity (U.S. Pat. No. 4,816,567; and Morrison et al (1984) Proc. Natl. Acad. Sci. USA, 81:6851-6855). Chimeric antibodies of interest herein include “primatized” antibodies comprising variable domain antigen-binding sequences derived from a non-human primate (e.g., Old World Monkey, Ape, etc.) and human constant region sequences.

[0077] An “intact antibody chain” as used herein is one comprising a full length variable region and a full length constant region (Fc). An intact “conventional” antibody comprises an intact light chain and an intact heavy chain, as well as a light chain constant domain (CL) and heavy chain constant domains, CH1, hinge, CH2 and CH3 for secreted IgG. Other isotypes, such as IgM or IgA may have different CH domains. The constant domains may be native sequence constant domains (e.g., human native sequence constant domains) or amino acid sequence variants thereof. The intact antibody may have one or more “effector functions” which refer to those biological activities attributable to the Fc constant region (a native sequence Fc region or amino acid sequence variant Fc region) of an antibody. Examples of antibody effector functions include C1q binding; complement dependent cytotoxicity; Fc receptor binding; antibody-dependent cell-mediated cytotoxicity (ADCC); phagocytosis; and down regulation of cell surface receptors. Constant region variants include those that alter the effector profile, binding to Fc receptors, and the like.

[0078] Depending on the amino acid sequence of the Fc (constant domain) of their heavy chains, antibodies and various antigen-binding proteins can be provided as different classes. There are five major classes of heavy chain Fc regions: IgA, IgD, IgE, IgG, and IgM, and several of these may be further divided into “subclasses” (isotypes), e.g., IgG1, IgG2, IgG3, IgG4, IgA, and IgA2. The Fc

constant domains that correspond to the different classes of antibodies may be referenced as α , δ , ϵ , γ , and μ , respectively. The subunit structures and three-dimensional configurations of different classes of immunoglobulins are well known. Ig forms include hinge-modifications or hingeless forms (Roux et al (1998) *J. Immunol.* 161:4083-4090; Lund et al (2000) *Eur. J. Biochem.* 267:7246-7256; US 2005/0048572; US 2004/0229310). The light chains of antibodies from any vertebrate species can be assigned to one of two types, called κ and λ , based on the amino acid sequences of their constant domains.

[0079] A “functional Fc region” possesses an “effector function” of a native-sequence Fc region. Exemplary effector functions include C1q binding; CDC; Fc-receptor binding; ADCC; ADCP; down-regulation of cell-surface receptors (e.g., B-cell receptor), etc. Such effector functions generally require the Fc region to be interact with a receptor, e.g. the Fc γ RI; Fc γ RIIA; Fc γ RIIB1; Fc γ RIIB2; Fc γ RIIIA; Fc γ RIIIB receptors, and the low affinity FcRn receptor; and can be assessed using various assays as disclosed, for example, in definitions herein. A “dead” Fc is one that has been mutagenized to retain activity with respect to, for example, prolonging serum half-life, but which does not activate a high affinity Fc receptor.

[0080] A “native-sequence Fc region” comprises an amino acid sequence identical to the amino acid sequence of an Fc region found in nature. Native-sequence human Fc regions include, for example, a native-sequence human IgG1 Fc region (non-A and A allotypes); native-sequence human IgG2 Fc region; native-sequence human IgG3 Fc region; and native-sequence human IgG4 Fc region, as well as naturally occurring variants thereof.

[0081] A “variant Fc region” comprises an amino acid sequence that differs from that of a native-sequence Fc region by virtue of at least one amino acid modification, preferably one or more amino acid substitution(s). Preferably, the variant Fc region has at least one amino acid substitution compared to a native-sequence Fc region or to the Fc region of a parent polypeptide, e.g., from about one to about ten amino acid substitutions, and preferably from about one to about five amino acid substitutions in a native-sequence Fc region or in the Fc region of the parent polypeptide. The variant Fc region herein will preferably possess at least about 80% homology with a native-sequence Fc region and/or with an Fc region of a parent polypeptide, and most preferably at least about 90% homology therewith, more preferably at least about 95% homology therewith.

[0082] Variant Fc sequences may include three amino acid substitutions in the CH2 region to reduce Fc γ RI binding at EU index positions 234, 235, and 237 (see Duncan et al., (1988) *Nature* 332:563). Two amino acid substitutions in the complement C1q binding site at EU index positions 330 and 331 reduce complement fixation (see Tao et al., *J. Exp. Med.* 178:661 (1993) and Canfield and Morrison, *J. Exp. Med.* 173:1483 (1991)). Substitution into human IgG1 of IgG2 residues at positions 233-236 and IgG4 residues at positions 327, 330 and 331 greatly reduces ADCC and CDC (see, for example, Armour KL. *et al.*, 1999 *Eur J Immunol.* 29(8):2613-24; and Shields RL. *et al.*, 2001. *J Biol Chem.* 276(9):6591-604). Other Fc variants are possible, including without limitation one in which a region

capable of forming a disulfide bond is deleted, or in which certain amino acid residues are eliminated at the N-terminal end of a native Fc form or a methionine residue is added thereto. Thus, in one embodiment of the invention, one or more Fc portions of the scFc molecule can comprise one or more mutations in the hinge region to eliminate disulfide bonding. In yet another embodiment, the hinge region of an Fc can be removed entirely. In still another embodiment, the molecule can comprise an Fc variant.

[0083] Further, an Fc variant can be constructed to remove or substantially reduce effector functions by substituting, deleting or adding amino acid residues to effect complement binding or Fc receptor binding. For example, and not limitation, a deletion may occur in a complement-binding site, such as a C1q-binding site. Techniques of preparing such sequence derivatives of the immunoglobulin Fc fragment are disclosed in International Patent Publication Nos. WO 97/34631 and WO 96/32478. In addition, the Fc domain may be modified by phosphorylation, sulfation, acylation, glycosylation, methylation, farnesylation, acetylation, amidation, and the like.

[0084] The Fc may be in the form of having native sugar chains, increased sugar chains compared to a native form or decreased sugar chains compared to the native form, or may be in an aglycosylated or deglycosylated form. The increase, decrease, removal or other modification of the sugar chains may be achieved by methods common in the art, such as a chemical method, an enzymatic method or by expressing it in a genetically engineered production cell line. Such cell lines can include microorganisms, e.g. *Pichia Pastoris*, and mammalian cell line, e.g. CHO cells, that naturally express glycosylating enzymes. Further, microorganisms or cells can be engineered to express glycosylating enzymes, or can be rendered unable to express glycosylation enzymes (See e.g., Hamilton, et al., *Science*, 313:1441 (2006); Kanda, et al, *J. Biotechnology*, 130:300 (2007); Kitagawa, et al., *J. Biol. Chem.*, 269 (27): 17872 (1994); Ujita-Lee et al., *J. Biol. Chem.*, 264 (23): 13848 (1989); Imai-Nishiya, et al, *BMC Biotechnology* 7:84 (2007); and WO 07/055916). As one example of a cell engineered to have altered sialylation activity, the alpha-2,6-sialyltransferase 1 gene has been engineered into Chinese Hamster Ovary cells and into sf9 cells. Antibodies expressed by these engineered cells are thus sialylated by the exogenous gene product. A further method for obtaining Fc molecules having a modified amount of sugar residues compared to a plurality of native molecules includes separating said plurality of molecules into glycosylated and non-glycosylated fractions, for example, using lectin affinity chromatography (See e.g., WO 07/117505). The presence of particular glycosylation moieties has been shown to alter the function of Immunoglobulins. For example, the removal of sugar chains from an Fc molecule results in a sharp decrease in binding affinity to the C1q part of the first complement component C1 and a decrease or loss in antibody-dependent cell-mediated cytotoxicity (ADCC) or complement-dependent cytotoxicity (CDC), thereby not inducing unnecessary immune responses in vivo. Additional important modifications include sialylation and fucosylation: the presence of sialic acid in IgG has been correlated with anti-inflammatory activity

(See e.g., Kaneko, et al, Science 313:760 (2006)), whereas removal of fucose from the IgG leads to enhanced ADCC activity (See e.g., Shoj-Hosaka, et al, J. Biochem., 140:777 (2006)).

[0085] In alternative embodiments, antibodies of the invention may have an Fc sequence with enhanced effector functions, e.g. by increasing their binding capacities to FcγRIIIA and increasing ADCC activity. For example, fucose attached to the N-linked glycan at Asn-297 of Fc sterically hinders the interaction of Fc with FcγRIIIA, and removal of fucose by glyco-engineering can increase the binding to FcγRIIIA, which translates into >50-fold higher ADCC activity compared with wild type IgG1 controls. Protein engineering, through amino acid mutations in the Fc portion of IgG1, has generated multiple variants that increase the affinity of Fc binding to FcγRIIIA. Notably, the triple alanine mutant S298A/E333A/K334A displays 2-fold increase binding to FcγRIIIA and ADCC function. S239D/I332E (2X) and S239D/I332E/A330L (3X) variants have a significant increase in binding affinity to FcγRIIIA and augmentation of ADCC capacity *in vitro* and *in vivo*. Other Fc variants identified by yeast display also showed the improved binding to FcγRIIIA and enhanced tumor cell killing in mouse xenograft models. See, for example Liu et al. (2014) JBC 289(6):3571-90, herein specifically incorporated by reference.

[0086] The term “Fc-region-comprising antibody” refers to an antibody that comprises an Fc region. The C-terminal lysine (residue 447 according to the EU numbering system) of the Fc region may be removed, for example, during purification of the antibody or by recombinant engineering the nucleic acid encoding the antibody. Accordingly, an antibody having an Fc region according to this invention can comprise an antibody with or without K447.

[0087] “Fv” is the minimum antibody fragment, which contains a complete antigen-recognition and antigen-binding site. The CD3 binding antibodies of the invention comprise a dimer of one heavy chain and one light chain variable domain in tight, non-covalent association; however additional antibodies, e.g. for use in a multi-specific configuration, may comprise a VH in the absence of a VL sequence. Even a single variable domain (or half of an Fv comprising only three hypervariable regions specific for an antigen) has the ability to recognize and bind antigen, although the affinity may be lower than that of two domain binding site.

[0088] The Fab fragment also contains the constant domain of the light chain and the first constant domain (CH1) of the heavy chain. Fab' fragments differ from Fab fragments by the addition of a few residues at the carboxy terminus of the heavy chain CH1 domain including one or more cysteines from the antibody hinge region. Fab'-SH is the designation herein for Fab' in which the cysteine residue(s) of the constant domains bear at least one free thiol group. F(ab')₂ antibody fragments originally were produced as pairs of Fab' fragments which have hinge cysteines between them. Other chemical couplings of antibody fragments are also known.

[0089] “Humanized” forms of non-human (e.g., rodent) antibodies, including single chain antibodies, are chimeric antibodies (including single chain antibodies) that contain minimal sequence derived from non-human immunoglobulin. See, for example, Jones et al, (1986) Nature 321:522-525; Chothia

et al (1989) *Nature* 342:877; Riechmann et al (1992) *J. Mol. Biol.* 224, 487-499; Foote and Winter, (1992) *J. Mol. Biol.* 224:487-499; Presta et al (1993) *J. Immunol.* 151, 2623-2632; Werther et al (1996) *J. Immunol. Methods* 157:4986-4995; and Presta et al (2001) *Thromb. Haemost.* 85:379-389. For further details, see U.S. Pat. Nos. 5,225,539; 6,548,640; 6,982,321; 5,585,089; 5,693,761; 6,407,213; Jones et al (1986) *Nature*, 321:522-525; and Riechmann et al (1988) *Nature* 332:323-329.

[0090] The term “single chain antibody” as used herein means a single polypeptide chain containing one or more antigen binding domains that bind an epitope of an antigen, where such domains are derived from or have sequence identity with the variable region of an antibody heavy or light chain. Parts of such variable region may be encoded by V_H or V_L gene segments, D and J_H gene segments, or J_L gene segments. The variable region may be encoded by rearranged V_HDJ_H, V_LDJ_H, V_HJ_L, or V_LJ_L gene segments. V-, D- and J-gene segments may be derived from humans and various animals including birds, fish, sharks, mammals, rodents, non-human primates, camels, lamas, rabbits and the like.

[0091] The term “compete” when used in the context of antibodies that compete for the same epitope means competition between antibodies as determined by an assay in which the antibody (e.g., antibody or immunologically functional fragment thereof) being tested prevents or inhibits (e.g., reduces) specific binding of a reference antibody (e.g., a ligand, or a reference antibody) to a common antigen (e.g., CD3 or a fragment thereof). Numerous types of competitive binding assays can be used to determine if one antibody competes with another, for example: solid phase direct or indirect radioimmunoassay (RIA), solid phase direct or indirect enzyme immunoassay (EIA), sandwich competition assay (see, e.g., Stahli et al., 1983, *Methods in Enzymology* 9:242-253); solid phase direct biotin-avidin EIA (see, e.g., Kirkland et al., 1986, *J. Immunol.* 137:3614-3619) solid phase direct labeled assay, solid phase direct labeled sandwich assay (see, e.g., Harlow and Lane, 1988, *Antibodies, A Laboratory Manual*, Cold Spring Harbor Press); solid phase direct label RIA using 1-125 label (see, e.g., Morel et al., 1988, *Molec. Immunol.* 25:7-15); solid phase direct biotin-avidin EIA (see, e.g., Cheung, et al., 1990, *Virology* 176:546-552); and direct labeled RIA (Moldenhauer et al., 1990, *Scand. J. Immunol.* 32:77-82).

[0092] Typically, such an assay involves the use of purified antigen bound to a solid surface or cells bearing either of these, an unlabelled test antibody and a labeled reference antibody. Competitive inhibition is measured by determining the amount of label bound to the solid surface or cells in the presence of the test antibody. Usually the test antibody is present in excess. Antibodies identified by competition assay (competing antibodies) include antibodies binding to the same epitope as the reference antibodies and antibodies binding to an adjacent epitope sufficiently proximal to the epitope bound by the reference antibody for steric hindrance to occur. Additional details regarding methods for determining competitive binding are provided in the examples herein. Usually, when a competing antibody is present in excess, it will inhibit (e.g., reduce) specific binding of a reference antibody to a common antigen by at least 40-45%, 45-50%, 50-55%, 55-60%, 60-65%, 65-70%, 70-75% or 75% or

more. In some instances, binding is inhibited by at least 80-85%, 85-90%, 90-95%, 95-97%, or 97% or more.

[0093] The term “epitope” includes any determinant capable being bound by an antibody, such as an F2B antibody. An epitope is a region of an antigen that is bound by an antibody that targets that antigen, and when the antigen is a protein, includes specific amino acids that directly contact the antibody. Epitope determinants can include chemically active surface groupings of molecules such as amino acids, sugar side chains, phosphoryl or sulfonyl groups, and can have specific three dimensional structural characteristics, and/or specific charge characteristics. Generally, antibodies specific for a particular target antigen will preferentially recognize an epitope on the target antigen in a complex mixture of proteins and/or macromolecules.

[0094] The CD3-binding antibodies of the invention find particular utility in multi-specific configurations, which include without limitation bispecific antibodies, trifunctional antibodies, etc. A large variety of methods and protein configurations are known and use in bispecific monoclonal antibodies (BsMAB), tri-specific antibodies, etc.

[0095] First-generation BsMAbs consisted of two heavy and two light chains, one each from two different antibodies. The two Fab regions are directed against two antigens. The Fc region is made up from the two heavy chains and forms the third binding site with the Fc receptor on immune cells (see for example Lindhofer et al., *The Journal of Immunology*, Vol 155, p 219-225, 1995). The antibodies may be from the same or different species. For example, cell lines expressing rat and mouse antibodies secrete functional bispecific Ab because of preferential species-restricted heavy and light chain pairing. In other embodiments the Fc regions are designed to only fit together in specific ways.

[0096] Other types of bispecific antibodies include chemically linked Fabs, consisting only of the Fab regions. Two chemically linked Fab or Fab2 fragments form an artificial antibody that binds to two different antigens, making it a type of bispecific antibody. Antigen-binding fragments (Fab or Fab2) of two different monoclonal antibodies are produced and linked by chemical means like a thioether (see Glennie, M J et al., *Journal of immunology* 139, p 2367-75, 1987; Peter Borchmann et al., *Blood*, Vol. 100, No. 9, p 3101-3107, 2002).

[0097] Various other methods for the production of multivalent artificial antibodies have been developed by recombinantly fusing variable domains of two antibodies. A single-chain variable fragment (scFv) is a fusion protein of the variable regions of the heavy (VH) and light chains (VL) of immunoglobulins, connected with a short linker peptide of ten to about 25 amino acids. The linker is usually rich in glycine for flexibility, as well as serine or threonine for solubility, and can either connect the N-terminus of the VH with the C-terminus of the VL, or vice versa. Bispecific single-chain variable fragments (di-scFvs, bi-scFvs) can be engineered by linking two scFvs with different specificities. A single peptide chain with two VH and two VL regions is produced, yielding bivalent scFvs.

[0098] Bispecific tandem scFvs are also known as bi-specific T-cell engagers (BiTEs). Bispecific scFvs can be created with linker peptides that are too short for the two variable regions to fold together (about five amino acids), forcing scFvs to dimerize. This type is known as diabodies (Adams et al., *British journal of cancer* 77, p 1405-12, 1998). The Dual-Affinity Re-Targeting (DART) platform technology (MacroGenics, Rockville, Md.). This fusion protein technology uses two single-chain variable fragments (scFvs) of different antibodies on a single peptide chain of about 55 kilodaltons. SCORPION Therapeutics (Emergent Biosolutions, Inc., Seattle, Wash.) combines two antigen-binding domains in a single chain protein. One binding domain is on the C-terminus and a second binding domain on the N-terminus of an effector domain, based on immunoglobulin Fc regions.

[0099] Tetravalent and bispecific antibody-like proteins also include DVD-Igs which are engineered from two monoclonal antibodies (Wu, C. et al., *Nature Biotechnology*, 25, p 1290-1297, 2007). To construct the DVD-Ig molecule, the V domains of the two mAbs are fused in tandem by a short linker (TVAAP) with the variable domain of the first antibody light (VL) chain at the N terminus, followed by the other antibodies VL and Ck to form the DVD-Ig protein light chain. Similarly, the variable regions of the heavy (VH) chain of the two mAbs are fused in tandem by a short linker (ASTKGP) with the first antibody at the N terminus, followed by the other antibody and the heavy chain constant domains to form the DVD-Ig protein heavy chain (VH1/VL1). All light chain and heavy chain constant domains are preserved in the DVD-Ig design, as they are critical for the formation of a disulfide-linked full IgG-like molecule. Co-transfection of mammalian cells with expression vectors encoding the DVD-Ig light chain and heavy chain leads to the secretion of a single species of an IgG-like molecule with molecular weight of approximately 200 kDa. This molecule has now four binding sites, 2 from each mAb.

[00100] The term "bispecific three-chain antibody like molecule" or "TCA" is used herein to refer to antibody-like molecules comprising, consisting essentially of, or consisting of three polypeptide subunits, two of which comprise, consist essentially of, or consist of one heavy and one light chain of a monoclonal antibody, or functional antigen-binding fragments of such antibody chains, comprising an antigen-binding region and at least one CH domain. This heavy chain/light chain pair has binding specificity for a first antigen. The third polypeptide subunit comprises, consists essentially of, or consists of a heavy chain only antibody comprising an Fc portion comprising CH2 and/or CH3 and/or CH4 domains, in the absence of a CH1 domain, and an antigen binding domain that binds an epitope of a second antigen or a different epitope of the first antigen, where such binding domain is derived from or has sequence identity with the variable region of an antibody heavy or light chain. Parts of such variable region may be encoded by V_H and/or V_L gene segments, D and J_H gene segments, or J_L gene segments. The variable region may be encoded by rearranged V_HDJ_H , V_LDJ_H , V_HJ_L , or V_LJ_L gene segments.

[00101] A TCA protein makes use of a heavy chain only antibody” or “heavy chain antibody” or “heavy chain polypeptide” as used herein means a single chain antibody comprising heavy chain constant regions CH2 and/or CH3 and/or CH4 but no CH1 domain. In one embodiment, the heavy chain antibody is composed of an antigen-binding domain, at least part of a hinge region and CH2 and CH3 domains. In another embodiment, the heavy chain antibody is composed of an antigen-binding domain, at least part of a hinge region and a CH2 domain. In a further embodiment, the heavy chain antibody is composed of an antigen-binding domain, at least part of a hinge region and a CH3 domain. Heavy chain antibodies in which the CH2 and/or CH3 domain is truncated are also included herein. In a further embodiment the heavy chain is composed of an antigen binding domain, and at least one CH (CH1, CH2, CH3, or CH4) domain but no hinge region. The heavy chain only antibody can be in the form of a dimer, in which two heavy chains are disulfide bonded other otherwise, covalently or non-covalently attached with each other. The heavy chain antibody may belong to the IgG subclass, but antibodies belonging to other subclasses, such as IgM, IgA, IgD and IgE subclass, are also included herein. In a particular embodiment, the heavy chain antibody is of the IgG1, IgG2, IgG3, or IgG4 subtype, in particular IgG1 subtype.

[00102] Heavy chain antibodies constitute about one fourth of the IgG antibodies produced by the camelids, e.g. camels and llamas (Hamers-Casterman C., et al. *Nature*. 363, 446-448 (1993)). These antibodies are formed by two heavy chains but are devoid of light chains. As a consequence, the variable antigen binding part is referred to as the VHH domain and it represents the smallest naturally occurring, intact, antigen-binding site, being only around 120 amino acids in length (Desmyter, A., et al. *J. Biol. Chem.* 276, 26285-26290 (2001)). Heavy chain antibodies with a high specificity and affinity can be generated against a variety of antigens through immunization (van der Linden, R. H., et al. *Biochim. Biophys. Acta*. 1431, 37-46 (1999)) and the VHH portion can be readily cloned and expressed in yeast (Frenken, L. G. J., et al. *J. Biotechnol.* 78, 11-21 (2000)). Their levels of expression, solubility and stability are significantly higher than those of classical F(ab) or Fv fragments (Ghahroudi, M. A. et al. *FEBS Lett.* 414, 521-526 (1997)). Sharks have also been shown to have a single VH-like domain in their antibodies termed VNAR. (Nuttall et al. *Eur. J. Biochem.* 270, 3543-3554 (2003); Nuttall et al. *Function and Bioinformatics* 55, 187-197 (2004); Dooley et al., *Molecular Immunology* 40, 25-33 (2003)).

[00103] An antibody or antigen-binding molecule, including the heavy chain only antibodies and bispecific three-chain antibody-like molecules (TCAs) herein, “which binds” an antigen of interest, is one that binds the antigen with sufficient affinity such that the antibody or binding molecule is useful as a diagnostic and/or therapeutic agent in targeting the antigen, and does not significantly cross-react with other proteins. In such embodiments, the extent of binding of the antibody or other binding molecule to a non-targeted antigen will be no more than 10% as determined by fluorescence activated cell sorting (FACS) analysis or radioimmunoprecipitation (RIA).

PROTEINS

[00104] The present invention provides a family of closely related antibodies that bind to and activate signaling through CD3, e.g. activation of CD3⁺ T cells. The antibodies within the family comprise a set of CDR sequences as defined herein, and are exemplified by the provided VH sequences of SEQ ID NO:1-18. The family of antibodies provides a number of benefits that contribute to utility as clinically therapeutic agent(s). The antibodies within the family include members with a range of binding affinities, allowing the selection of a specific sequence with a desired affinity. The ability to fine tune affinity is of particular importance to manage the level of CD3 activation in an individual being treated, and thereby reduce toxicity. For example, if low abundant tumor antigens (less than 10,000 molecules per cell) are targeted, it is anticipated that high affinity CD3 binders (<30 nM) are preferred. If highly abundant tumor antigens (more than 50,000 molecules per cell) are targeted, CD3 binders with low affinities (>50 nM) are preferred. Separately evaluated from affinity can be the propensity of the antibody to induce release of cytokines when bound to a T cell, e.g. release of IL-2, IFN γ , etc., where reduced cytokine release may be desirable.

[00105] In another aspect, antibodies are provided that compete with one of the exemplified antibodies or functional fragments binding to the epitope described herein for specific binding to CD3. Such antibodies can also bind to the same epitope as one of the herein exemplified antibodies, or an overlapping epitope. Antibodies and fragments that compete with or bind to the same epitope as the exemplified antibodies are expected to show similar functional properties. The exemplified antibodies and fragments include those described above, including those with the heavy and light chains, variable region domains and CDRs shown in FIG. 1.

[00106] Such competing antibodies may bind to the F2B epitope, but comprise a set of CDR sequences other than those set forth in SEQ ID NO:1-18. The CDR sequences of the heavy chain may be substantially similar but not identical to the CDR sequences set forth in SEQ ID NO:1-18, e.g. may comprise 1 amino acid substitution in a CDR sequence, 1 amino acid substitutions, 3 amino acid substitutions, or more, where the changes may be found in 1 CDR sequence, 2 CDR sequences, or 3 CDR sequences. The light chain sequence may comprise the set of CDR sequences set forth in SEQ ID NO:19; or may comprise a different set of CDR sequences.

[00107] Residues directly involved in binding to an epitope or covered by an antibody can be identified from scanning results. These residues can thus provide an indication of the domains or regions of CD3 that contain the binding region(s) to which antibodies bind.

[00108] The F2B epitope is characterized by binding to at least one residue selected from CD3 epsilon (SEQ ID NO:23): K73 and S83; and CD3 delta (SEQ ID NO:24) K82 and C93. In some embodiments the epitope comprises the region of CD3 epsilon defined by K73, N74, I75, G76, S77, D78, E79, D80, H81, L82, S83. In some embodiments the epitope comprises one or both of K73 and S83. In some embodiments the epitope comprises the region of CD3 delta defined by K82, E83, S84, T85, V86, Q87, V88, H89, Y90, R91, M92, C93. In some embodiments the epitope comprises one or both of

K82 and C93. In some embodiments the F2B epitope comprises a conformational epitope involving residues of both CD3 delta and CD3 epsilon. In some embodiments the conformational epitope comprises each of residues CD3ε K73 and S83; CD3δ K82 and C93. In some embodiments antibodies that bind to the F2B epitope are not cross-reactive with cynomolgus CD3 protein. A suitable antibody may be selected from those provided herein for development and use, including without limitation use as a bispecific antibody. Determination of affinity for a candidate protein can be performed using methods known in the art, e.g. Biacore measurements, etc. Members of the antibody family may have an affinity for CD3 with a K_d of from about 10⁻⁶ to around about 10⁻¹¹, including without limitation: from about 10⁻⁶ to around about 10⁻¹⁰; from about 10⁻⁶ to around about 10⁻⁹; from about 10⁻⁶ to around about 10⁻⁸; from about 10⁻⁸ to around about 10⁻¹¹; from about 10⁻⁸ to around about 10⁻¹⁰; from about 10⁻⁸ to around about 10⁻⁹; from about 10⁻⁹ to around about 10⁻¹¹; from about 10⁻⁹ to around about 10⁻¹⁰; or any value within these ranges. The affinity selection may be confirmed with a biological assessment for activation of T cells in, for example, and in vitro or pre-clinical model, and assessment of potential toxicity. Determination of cytokine release can be evaluated using any convenient method, including without limitation the assays described in the examples.

[00109] Engagement of the T cell receptor (TCR), either by binding MH-peptide complexes or anti-TCR/CD3 antibodies, initiates T cell activation. Examples of anti-TCR/CD3 antibodies that activate T cells are OKT3 and UCHT1. These anti-CD3 antibodies cross-compete for binding to CD3 on T cells and are routinely used in T cell activation assays. Anti-CD3 antibodies of this invention cross-compete with OKT3 for binding to human CD3. Depending on the binding affinity for CD3 and epitope on CD3, anti-CD3 antibodies activated T cells with different functional outcomes. In vitro incubation of human T cells with low affinity anti-CD3 antibodies resulted in incomplete activation of T cells, low IL-2 and IL-10 production. In contrast, high-affinity CD3 binders activated T cells to produce significantly more IL-2 and other cytokines. The low-affinity anti-CD3 antibodies are considered partial agonists that selectively induce some effector functions, potent tumor killing and CD69 upregulation, while failing to induce others, such as IL-2 and IL-10 production. The strength of the interaction with CD3 and the epitope recognized resulted in qualitatively different activation of T cells. Maximal cytokine production of T cells activated by low-affinity anti-CD3 antibodies was lower than maximal activation by high-affinity anti-CD3 antibodies. In some embodiments, an antibody of the invention results in a lower release of one or both of IL-2 and IL-10 when combined with T cells in an activation assay when compared to a reference anti-CD3 antibody in the same assay, where the reference antibody can be ID 304703 (SEQ ID NO:22) or an antibody of equivalent affinity. The maximal release of IL-2 and/or IL-10 can be less than about 75% of the release by the reference antibody, less than about 50% of the release by the reference antibody, less than about 25% of the release by the reference antibody, and may be less than about 10% of the release by a reference antibody.

[00110] In some embodiments of the invention, bispecific or multispecific antibodies are provided, which may have any of the configurations discussed herein, including without limitation a three chain bispecific. Bispecific antibodies comprise at least the heavy chain variable region of an antibody specific for a protein other than CD3, and may comprise a heavy and light chain variable region. In some such embodiments, the second antibody specificity binds to a tumor associated antigen, a targeting antigen, *e.g.* integrins, *etc.*, a pathogen antigen, a checkpoint protein, and the like. Various formats of bispecific antibodies are within the ambit of the invention, including without limitation single chain polypeptides, two chain polypeptides, three chain polypeptides, four chain polypeptides, and multiples thereof.

[00111] The family of CD3 specific antibodies comprise a VH domain, comprising CDR1, CDR2 and CDR3 sequences in a human VH framework. The CDR sequences may be situated, as an example, in the region of around amino acid residues 26-33; 51-58; and 97-112 for CDR1, CDR2 and CDR3, respectively, of the provided exemplary variable region sequences set forth in SEQ ID NO:1-18. It will be understood by one of skill in the art that the CDR sequences may be in different position if a different framework sequence is selected, although generally the order of the sequences will remain the same.

[00112] The CDR sequences for a family 2 antibody may have the following sequence formulas. An X indicates a variable amino acid, which may be specific amino acids as indicated below.

CDR1

G₁ F₂ T₃ F₄ X₅ X₆ Y₇ A₈

where:

X₅ may be any amino acid; in some embodiments X₅ is D, A or H; in some embodiments X₅ is D.

X₆ may be any amino acid; in some embodiments X₆ is D or N; in some embodiments D₆ is D.

In some embodiments a CDR1 sequence of a family 2 anti-CD3 antibody comprises the sequence set forth in any of SEQ ID NO:1-18, residues 26-33.

CDR2

I₁' S₂' W₃' N₄' S₅' G₆' S₇' I₈'

[00113] In some embodiments a CDR2 sequence of a family 2 anti-CD3 antibody comprises the sequence set forth in any of SEQ ID NO:1-18, residues 51-58.

CDR3

A₁" K₂" D₃" S₄" R₅" G₆" Y₇" G₈" X₉" Y₁₀" X₁₁" X₁₂" G₁₃" G₁₂" A₁₅" Y₁₆"

where:

X₉" may be any amino acid, in some embodiments X₉" is D or S; in some embodiments X₉" is D;

X₁₁" may be any amino acid, in some embodiments X₁₁" is R or S;

X₁₂" may be any amino acid, in some embodiments X₁₂" is L or R.

[00114] In some embodiments a CD3 sequence of a family 2 anti-CD3 antibody has the formula A K D S R G Y G D Y X₁₁" X₁₂" G G A Y where X₁₁" and X₁₂" are as defined above. In some embodiments a CDR3 sequence of a family 2 anti-CD3 antibody comprises the sequence set forth in any of SEQ ID NO:1-18, residues 97-112.

[00115] In some embodiments the CD3-binding VH domain is paired with a light chain variable region domain. In some such embodiments the light chain is a fixed light chain. In some embodiments the light chain comprises a VL domain with CDR1, CDR2 and CDR3 sequences in a human VL framework. The CDR sequences may be those of SEQ ID NO:19. In some embodiments, the CDR1 sequence comprises amino acid residues 27-32; 50-52; 89-97 for CDR1, CDR2, CDR3, respectively.

[00116] In some embodiments the CDR sequences of a family 2 antibody have a sequence with at least 85% identity, at least 90% identity, at least 95% identity, at least 99% identity relative to a CDR sequence or set of CDR sequences in any one of SEQ ID NO:1-18. In some embodiments a CDR sequence of the invention comprises one, two, three or more amino acid substitutions relative to a CDR sequence or set of CDR sequences in any one of SEQ ID NO:1-18. In some embodiments said amino acid substitution(s) are one or more of position 5 or 10 of CDR1, position 2, 6 or 7 of CDR2, position 1, 8, 9 or 10 of CDR3, relative to the formulas provided above.

[00117] Where a protein of the invention is a bispecific antibody, one binding moiety, i.e. VH/VL combination or VH only, is specific for human CD3 while the other arm may be specific for target cells, including cancer cells, such as cells of ovarian, breast, gastrointestinal, brain, head and neck, prostate, colon, and lung cancers, and the like, as well as hematologic tumors such as B-cell tumors, including leukemias, lymphomas, sarcomas, carcinomas, neural cell tumors, squamous cell carcinomas, germ cell tumors, metastases, undifferentiated tumors, seminomas, melanomas, myelomas, neuroblastomas, mixed cell tumors, neoplasias caused by infectious agents, and other malignancies, cells infected with a pathogen, autoreactive cells causing inflammation and/or autoimmunity. The non-CD3 moiety can also be specific for an immune regulatory protein, as will be described herein.

[00118] Tumor-associated antigens (TAAs) are relatively restricted to tumor cells, whereas tumor-specific antigens (TSAs) are unique to tumor cells. TSAs and TAAs typically are portions of intracellular molecules expressed on the cell surface as part of the major histocompatibility complex.

[00119] Tissue specific differentiation antigens are molecules present on tumor cells and their normal cell counterparts. Tumor-associated antigens known to be recognized by therapeutic mAbs fall into several different categories. Hematopoietic differentiation antigens are glycoproteins that are usually associated with cluster of differentiation (CD) groupings and include CD20, CD30, CD33 and CD52. Cell surface differentiation antigens are a diverse group of glycoproteins and carbohydrates that are found on the surface of both normal and tumor cells. Antigens that are involved in growth and differentiation signaling are often growth factors and growth factor receptors. Growth factors that are targets for antibodies in cancer patients include CEA, epidermal growth factor receptor (EGFR; also known as ERBB1), ERBB2 (also known as HER2), ERBB3, MET (also known as HGFR), insulin-like growth factor 1 receptor (IGF1R), ephrin receptor A3 (EPHA3), tumor necrosis factor (TNF)-related apoptosis-inducing ligand receptor 1 (TRAILR1; also known as TNFRSF10A), TRAILR2 (also known as TNFRSF10B) and receptor activator of nuclear factor- κ B ligand (RANKL; also known as TNFSF11). Antigens involved in angiogenesis are usually proteins or growth factors that support the formation of new microvasculature, including vascular endothelial growth factor (VEGF), VEGF receptor (VEGFR), integrin α V β 3 and integrin α 5 β 1. Tumor stroma and the extracellular matrix are indispensable support structures for a tumor. Stromal and extracellular matrix antigens that are therapeutic targets include fibroblast activation protein (FAP) and tenascin.

[00120] Examples of therapeutic antibodies useful in bispecific configurations include, without limitation, rituximab; Ibritumomab; tiuxetan; tositumomab; Brentuximab; vedotin; Gemtuzumab; ozogamicin; Alemtuzumab; IGN101; adecatumumab; Labetuzumab; huA33; Pemtumomab; oregovomab; CC49 (minretumomab); cG250; J591; MOv18; MORAb-003 (farletuzumab); 3F8, ch14.18; KW-2871; hu3S193; IgN311; Bevacizumab; IM-2C6; CDP791; Etaracizumab; Volociximab; Cetuximab, panitumumab, nimotuzumab; 806; Trastuzumab; pertuzumab; MM-121; AMG 102, METMAB; SCH 900105; AVE1642, IMC-A12, MK-0646, R1507; CP 751871; KB004; IIIA4; Mapatumumab (HGS-ETR1); HGS-ETR2; CS-1008; Denosumab; Sibrotuzumab; F19; and 81C6.

[00121] The immune-checkpoint receptors that have been most actively studied in the context of clinical cancer immunotherapy, cytotoxic T-lymphocyte-associated antigen 4 (CTLA4; also known as CD152) and programmed cell death protein 1 (PD1; also known as CD279) - are both inhibitory receptors. The clinical activity of antibodies that block either of these receptors implies that antitumor immunity can be enhanced at multiple levels and that combinatorial strategies can be intelligently designed, guided by mechanistic considerations and preclinical models.

[00122] The two ligands for PD1 are PD1 ligand 1 (PDL1; also known as B7-H1 and CD274) and PDL2 (also known as B7-DC and CD273). PDL1 is expressed on cancer cells and through binding to its receptor PD1 on T cells it inhibits T cell activation/function.

[00123] Lymphocyte activation gene 3 (LAG3; also known as CD223), 2B4 (also known as CD244), B and T lymphocyte attenuator (BTLA; also known as CD272), T cell membrane protein 3 (TIM3;

also known as HAVcr2), adenosine A2a receptor (A2aR) and the family of killer inhibitory receptors have each been associated with the inhibition of lymphocyte activity and in some cases the induction of lymphocyte anergy. Antibody targeting of these receptors can be used in the methods of the invention.

[00124] Agents that agonize an immune costimulatory molecule are also useful in the methods of the invention. Such agents include agonists of CD40 and OX40. CD40 is a costimulatory protein found on antigen presenting cells (APCs) and is required for their activation. These APCs include phagocytes (macrophages and dendritic cells) and B cells. CD40 is part of the TNF receptor family. The primary activating signaling molecules for CD40 are IFN γ and CD40 ligand (CD40L). Stimulation through CD40 activates macrophages.

[00125] Anti CCR4 (CD194) antibodies of interest include humanized monoclonal antibodies directed against C-C chemokine receptor 4 (CCR4) with potential anti-inflammatory and antineoplastic activities. CCR2 is expressed on inflammatory macrophages that can be found in various inflammatory conditions, e.g. rheumatoid arthritis; and have also been identified as expressed on tumor promoting macrophages. CCR2 is also expressed on regulatory T cells, and the CCR2 ligand, CCL2, mediates recruitment of regulatory T cells into tumors. Regulatory T cells suppress a response for anti-tumor T cells and thus their inhibition or depletion is desired.

PRODUCING PROTEINS OF THE INVENTION

[00126] Although antibodies can be prepared by chemical synthesis, they are typically produced by methods of recombinant DNA technology, such as co-expression of all the chains making up the protein in a single recombinant host cell, or co-expression of a heavy chain polypeptide and an antibody, e.g. a human antibody. In addition, the antibody heavy and light chains can also be expressed using a single polycistronic expression vector. Purification of individual polypeptides is achieved using standard protein purification technologies such as affinity (protein A) chromatography, size exclusion chromatography and/or hydrophobic interaction chromatography. Bispecifics are sufficiently different in size and hydrophobicity that purification can be performed using standard procedures.

[00127] The amount of antibody and heavy chain polypeptide produced in a single host cell can be minimized through engineering of constant regions of the antibody and the heavy chain such that homodimerization is favored over heterodimerization, e.g. by introducing self-complementary interactions (see e.g. WO 98/50431 for possibilities, such as “protuberance-into-cavity” strategies (see WO 96/27011)). It is therefore another aspect of the present invention to provide a method for producing a bispecific in a recombinant host, the method including the step of: expressing in a recombinant host cell a nucleic acid sequences encoding at least two heavy chain polypeptides, wherein said heavy chain polypeptides differ in their constant regions sufficiently to reduce or prevent homodimer formation but increase bispecific formation.

[00128] Where the protein comprises three chains, e.g. FlicAbs, they may be produced by co-expression of the three chains (2 heavy chains and one light chain) making up the molecule in a single recombinant host cell.

[00129] For recombinant production of the proteins herein, one or more nucleic acids encoding all chains, e.g. 2, 3, 4, etc. are isolated and inserted into a replicable vector for further cloning (amplification of the DNA) or for expression. Many vectors are available. The vector components generally include, but are not limited to, one or more of the following: a signal sequence, an origin of replication, one or more marker genes, an enhancer element, a promoter, and a transcription termination sequence.

[00130] In a preferred embodiment, the host cell according to the method of the invention is capable of high-level expression of human immunoglobulin, i.e. at least 1 pg/cell/day, preferably at least 10 pg/cell/day and even more preferably at least 20 pg/cell/day or more without the need for amplification of the nucleic acid molecules encoding the single chains in said host cell.

PHARMACEUTICAL COMPOSITION

[00131] It is another aspect of the present invention to provide pharmaceutical compositions comprising one or more proteins of the present invention in admixture with a suitable pharmaceutically acceptable carrier. Pharmaceutically acceptable carriers as used herein are exemplified, but not limited to, adjuvants, solid carriers, water, buffers, or other carriers used in the art to hold therapeutic components, or combinations thereof.

[00132] Therapeutic formulations of the proteins used in accordance with the present invention are prepared for storage by mixing proteins having the desired degree of purity with optional pharmaceutically acceptable carriers, excipients or stabilizers (see, e.g. Remington's Pharmaceutical Sciences 16th edition, Osol, A. Ed. (1980)), such as in the form of lyophilized formulations or aqueous solutions. Acceptable carriers, excipients, or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid and methionine; preservatives (such as octadecyldimethylbenzyl ammonium chloride; hexamethonium chloride; benzalkonium chloride, benzethonium chloride; phenol, butyl or benzyl alcohol; alkyl parabens such as methyl or propyl paraben; catechol; resorcinol; cyclohexanol; 3-pentanol; and m-cresol); low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, histidine, arginine, or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrans; chelating agents such as EDTA; sugars such as sucrose, mannitol, trehalose or sorbitol; salt-forming counter-ions such as sodium; metal complexes (e.g. Zn-protein complexes); and/or non-ionic surfactants such as TWEEN™, PLURONICS™ or polyethylene glycol (PEG).

[00133] Anti-CD3 antibody formulations are disclosed, for example, in U.S. Patent Publication No. 20070065437, the entire disclosure is expressly incorporated by reference herein. Similar formulations can be used for the proteins of the present invention. The main components of such formulations are a pH buffering agent effective in the range of 3.0 to 6.2, a salt, a surfactant, and an effective amount of a bispecific with anti-CD3 specificity.

METHODS OF USE

[00134] Methods are provided for treating or reducing disease, including without limitation infection, autoimmune disease, primary or metastatic cancer, etc. in a regimen comprising contacting the targeted cells with an antigen-binding composition of the invention, particularly where the antigen-binding composition is a multi-specific antibody suitable for the condition being treated, e.g. where one binding moiety specifically binds to a tumor associated antigen for treatment of the relevant cancer cells; a binding moiety specific for a pathogen of interest for treatment of the relevant infection, and the like. Such methods include administering to a subject in need of treatment a therapeutically effective amount or an effective dose of the agents of the invention, including without limitation combinations of the reagent with a chemotherapeutic drug, radiation therapy, or surgery.

[00135] Effective doses of the compositions of the present invention for the treatment of disease vary depending upon many different factors, including means of administration, target site, physiological state of the patient, whether the patient is human or an animal, other medications administered, and whether treatment is prophylactic or therapeutic. Usually, the patient is a human, but nonhuman mammals may also be treated, e.g. companion animals such as dogs, cats, horses, *etc.*, laboratory mammals such as rabbits, mice, rats, *etc.*, and the like. Treatment dosages can be titrated to optimize safety and efficacy.

[00136] Dosage levels can be readily determined by the ordinarily skilled clinician, and can be modified as required, e.g., as required to modify a subject's response to therapy. The amount of active ingredient that can be combined with the carrier materials to produce a single dosage form varies depending upon the host treated and the particular mode of administration. Dosage unit forms generally contain between from about 1 mg to about 500 mg of an active ingredient.

[00137] In some embodiments, the therapeutic dosage the agent may range from about 0.0001 to 100 mg/kg, and more usually 0.01 to 5 mg/kg, of the host body weight. For example dosages can be 1 mg/kg body weight or 10 mg/kg body weight or within the range of 1-10 mg/kg. An exemplary treatment regime entails administration once every two weeks or once a month or once every 3 to 6 months. Therapeutic entities of the present invention are usually administered on multiple occasions. Intervals between single dosages can be weekly, monthly or yearly. Intervals can also be irregular as indicated by measuring blood levels of the therapeutic entity in the patient. Alternatively, therapeutic entities of the present invention can be administered as a sustained release formulation, in which case

less frequent administration is required. Dosage and frequency vary depending on the half-life of the polypeptide in the patient.

[00138] In prophylactic applications, a relatively low dosage may be administered at relatively infrequent intervals over a long period of time. Some patients continue to receive treatment for the rest of their lives. In other therapeutic applications, a relatively high dosage at relatively short intervals is sometimes required until progression of the disease is reduced or terminated, and preferably until the patient shows partial or complete amelioration of symptoms of disease. Thereafter, the patent can be administered a prophylactic regime.

[00139] In still other embodiments, methods of the present invention include treating, reducing or preventing tumor growth, tumor metastasis or tumor invasion of cancers including carcinomas, hematologic cancers such as leukemias and lymphomas, melanomas, sarcomas, gliomas, *etc.* For prophylactic applications, pharmaceutical compositions or medicaments are administered to a patient susceptible to, or otherwise at risk of disease in an amount sufficient to eliminate or reduce the risk, lessen the severity, or delay the outset of the disease, including biochemical, histologic and/or behavioral symptoms of the disease, its complications and intermediate pathological phenotypes presenting during development of the disease.

[00140] Compositions for the treatment of disease can be administered by parenteral, topical, intravenous, intratumoral, oral, subcutaneous, intraarterial, intracranial, intraperitoneal, intranasal or intramuscular means. A typical route of administration is intravenous or intratumoral, although other routes can be equally effective.

[00141] Typically, compositions are prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid vehicles prior to injection can also be prepared. The preparation also can be emulsified or encapsulated in liposomes or micro particles such as polylactide, polyglycolide, or copolymer for enhanced adjuvant effect, as discussed above. Langer, Science 249: 1527, 1990 and Hanes, Advanced Drug Delivery Reviews 28: 97-119, 1997. The agents of this invention can be administered in the form of a depot injection or implant preparation which can be formulated in such a manner as to permit a sustained or pulsatile release of the active ingredient. The pharmaceutical compositions are generally formulated as sterile, substantially isotonic and in full compliance with all Good Manufacturing Practice (GMP) regulations of the U.S. Food and Drug Administration.

[00142] Toxicity of the proteins described herein can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, *e.g.*, by determining the LD₅₀ (the dose lethal to 50% of the population) or the LD₁₀₀ (the dose lethal to 100% of the population). The dose ratio between toxic and therapeutic effect is the therapeutic index. The data obtained from these cell culture assays and animal studies can be used in formulating a dosage range that is not toxic for use in human. The dosage of the proteins described herein lies preferably within a range of circulating concentrations that include the effective dose with little or no toxicity. The dosage can vary within

this range depending upon the dosage form employed and the route of administration utilized. The exact formulation, route of administration and dosage can be chosen by the individual physician in view of the patient's condition.

[00143] The pharmaceutical compositions can be administered in a variety of unit dosage forms depending upon the method of administration. For example, unit dosage forms suitable for oral administration include, but are not limited to, powder, tablets, pills, capsules and lozenges. It is recognized that compositions of the invention when administered orally, should be protected from digestion. This is typically accomplished either by complexing the molecules with a composition to render them resistant to acidic and enzymatic hydrolysis, or by packaging the molecules in an appropriately resistant carrier, such as a liposome or a protection barrier. Means of protecting agents from digestion are well known in the art.

[00144] The compositions for administration will commonly comprise an antibody or other ablative agent dissolved in a pharmaceutically acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers can be used, e.g., buffered saline and the like. These solutions are sterile and generally free of undesirable matter. These compositions may be sterilized by conventional, well known sterilization techniques. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions such as pH adjusting and buffering agents, toxicity adjusting agents and the like, e.g., sodium acetate, sodium chloride, potassium chloride, calcium chloride, sodium lactate and the like. The concentration of active agent in these formulations can vary widely, and will be selected primarily based on fluid volumes, viscosities, body weight and the like in accordance with the particular mode of administration selected and the patient's needs (e.g., Remington's Pharmaceutical Science (15th ed., 1980) and Goodman & Gillman, The Pharmacological Basis of Therapeutics (Hardman et al., eds., 1996)).

[00145] Also within the scope of the invention are kits comprising the active agents and formulations thereof, of the invention and instructions for use. The kit can further contain a least one additional reagent, e.g. a chemotherapeutic drug, *etc.* Kits typically include a label indicating the intended use of the contents of the kit. The term label includes any writing, or recorded material supplied on or with the kit, or which otherwise accompanies the kit.

[00146] The compositions can be administered for therapeutic treatment. Compositions are administered to a patient in an amount sufficient to substantially ablate targeted cells, as described above. An amount adequate to accomplish this is defined as a "therapeutically effective dose", which may provide for an improvement in overall survival rates. Single or multiple administrations of the compositions may be administered depending on the dosage and frequency as required and tolerated by the patient. The particular dose required for a treatment will depend upon the medical condition and history of the mammal, as well as other factors such as age, weight, gender, administration route, efficiency, etc.

[00147] The invention now being fully described, it will be apparent to one of ordinary skill in the art that various changes and modifications can be made without departing from the spirit or scope of the invention.

EXAMPLES

Example 1: Genetically Engineered Rats Expressing Heavy Chain-Only Antibodies

[00148] A human IgH locus was constructed and assembled in several parts, which involved the modification and joining of rat C region genes, which were then joined downstream of human V_H6-D-J_H region. Two BACs with separate clusters of human V_H genes were then co-injected with a BAC encoding the assembled (human V_H6-D-J_H-rat C) fragment.

[00149] Transgenic rats carrying artificial heavy chain immunoglobulin loci in unrearranged configuration were generated. The included constant region genes encode IgM, IgD, IgG2b, IgE, IgA and 3' enhancer. RT-PCR and serum analysis (ELISA) of transgenic rats revealed productive rearrangement of transgenic immunoglobulin loci and expression of heavy chain only antibodies of various isotypes in serum. Transgenic rats were cross-bred with rats with mutated endogenous heavy chain and light chain loci previously described in US patent publication 2009/0098134 A1. Analysis of such animals demonstrated inactivation of rat immunoglobulin heavy and light chain expression and high level expression of heavy chain antibodies with variable regions encoded by human V, D, and J genes. Immunization of transgenic rats resulted in production of high titer serum responses of antigen-specific heavy chain antibodies. These transgenic rats expressing heavy chain antibodies with a human VDJ region were called UniRats.

Example 2: Genetically Engineered Rats Expressing Fixed Light Chain Antibodies

[00150] Transgenic human antibody repertoires were generated from H-chains with diverse (V_H-D-J_H)_n rearrangement in combination with a unique L-chain. For this a rearranged L-chain, human Vk-Jk1-Ck, was integrated in the rat germline by DNA microinjection and the obtained transgenic animals were bred with a previously described rat strain that expresses a human H-chain repertoire naturally (Osborn *et al.*, 2013). This new rat strain was named OmniFlic.

[00151] Immunizations of OmniFlic rats, using many different antigens, produced high levels of antigen-specific IgG similar to other transgenic rats carrying the same IgH locus. Repertoire analysis by RT-PCR identified highly variable V_H-gene rearrangements at high transcript and protein levels. In addition, only one L-chain product, also expressed at high level, was identified.

[00152] Antigen-specific binders from OmniFlic were obtained by NGS and selection from cDNA libraries (yeast, *E.coli*, phage), which upon sequencing identified diverse H-chain transcripts. For the expression in mammalian cells hypermutated H-chain constructs were transfected in combination with the original transgenic Igk sequence. In this rearranged Vk-Jk1-Ck no mutational changes were

allowed and always the same L-chain was expressed with various H-chain products to generate monoclonal human IgG.

Example 3: Generation of Antigen-Specific Antibodies in Transgenic Rats

[00153] For the generation of antigen-specific heavy chain antibodies in rats, genetically engineered rats were immunized in two ways.

[00154] *Immunization with recombinant extracellular domains of PD-L1 and BCMA.* Recombinant extracellular domains of PD-L1 and BCMA were purchased from R&D Systems and were diluted with sterile saline and combined with adjuvant. Immunogens were either combined with Complete Freund's Adjuvant (CFA) and Incomplete Freund's Adjuvant (IFA) or Titermax and Ribi adjuvants. The first immunization (priming) with immunogen in CFA or Titermax was administered in the left and right legs. After the first immunization with immunogens in CFA two more immunizations in IFA (boosters) or 4 more immunizations in Ribi and one more in Titermax were administered in each leg. This sequence of immunizations leads to the development of B cells producing high affinity antibodies. The immunogen concentrations were 10 microgram per leg. Serum was collected from rats at the final bleed to determine serum titers.

[00155] For the generation of anti-human CD3 $\delta\epsilon$ antibodies genetically engineered rats were immunized using DNA-based immunization protocols. OmniFlic rats were immunized with human and cynomolgus CD3-epsilon/delta constructs at Aldevron, Inc. (Fargo, ND) using the GENOVAC Antibody Technology. Draining lymph nodes were harvested after the final boost and RNA isolated. Following cDNA synthesis, the IgH heavy chain antibody repertoire was characterized by Next Generation Sequencing and proprietary software. Candidate antigen-specific VH sequences showing evidence of antigen-specific positive selection were selected. Several hundred VH sequences encoding FlicAbs were selected for gene assembly and cloned into an expression vector. Subsequently, fully human FlicAb IgG1 antibodies were expressed in HEK cells for analysis by flow cytometry and ELISA. Human FlicAbs were tested for binding to primary human T cells and Jurkat cells by flow cytometry. In addition, human FlicAbs were tested using recombinant CD3 $\delta\epsilon$ proteins in ELISA. All FlicAbs with positive binding for human T cells are listed in FIG. 1. Selected sequences were further characterized in T cell activation assays.

Example 4: Analysis of Activation of Treg Cells

[00156] CD69 is a cell surface marker on T cells that is upregulated upon stimulation. In this experiment, peripheral blood mononuclear cells (PBMC) were isolated from buffy coats using Biocol (1.077 g/ml density) and standard methods. Isolated PBMCs were pre-cultured for 48 hours at 2×10^7 cells/ml in complete medium, washed and re-suspended at 10^6 cells/ml in complete medium. These cells were incubated for 24 hours in FACS tubes (BD Falcon Corning) coated with recombinant BCMA protein. Tubes were pre-coated overnight with recombinant BCMA protein at 10 microgram/ml.

[00157] Cells were washed and stained with antibodies specific for different subsets of T cells, namely: (1) CD4 positive T cells (T4-anti-huCD4-ECD), (2) CD8 positive T cells (anti-huCD8-AF700), and (3) Treg cells are defined by positivity for CD4, CD25, and the intracellular marker Foxp3 (anti-huCD25-PECy7, anti-Foxp3-AF647, anti-huCD25-PECy7, all antibodies were obtained from Beckman). Cells were analyzed on a Cytosflex flow cytometer using the appropriate templates. In contrast to currently available anti-CD3 bi-specific molecules, TNB-383B preferentially activates CD4+ and CD8+ T-cells over Treg cells. Preventing activation of Treg cells, an immunosuppressive cell type, could bolster CD8+ T cell functions and increase immune destruction of tumor targets. Data is shown in FIGS. 3 and 4.

Example 5: Expression and purification of CD3 delta/epsilon Fc fusion protein

[00158] Recombinant antigens CD3 epsilon (SEQ ID NO:23, extracellular domain (ECD) residues 22 – 105) and CD3 delta (SEQ ID NO:24, ECD residues 23-126), shown in FIG. 5, were cloned in frame with mouse IgG1 Fc, transiently co-expressed, and purified from CHO cell culture medium. A C-terminal His-tag added to the CD3 epsilon subunit was used for affinity capture of the CD3 delta/epsilon complex by IMAC using standard protocols and elution with imidazole. A second purification affinity tag with the sequence EPEA (C-tag) was added to the C-terminus of the CD3 delta subunit.

Example 6: Epitope mapping of mAb CD3 F2B

[00159] *Labeling of surface residues.* Epitope mapping was achieved by surface residue labeling with diethylpyrocarbonate DEPC (reference) which covalently reacts with accessible side chains of amino acids histidine, lysine, tyrosine, cysteine, serine, threonine and the free N-terminal amino group. DEPC labeling was carried out at a molar ratio of antibody: antigen of 30:1 to drive complex formation. The DEPC-labeled antibody-antigen complex was captured on Capture Select C-tag affinity matrix (Thermo Fisher Scientific) and excess antibody was removed by extensive washing. Affinity-resin bound CD3 delta/epsilon was subjected to standard reduction and alkylation methods, prior to digestion with trypsin, chymotrypsin and endopeptidase Glu-C. Released peptides were analyzed by LC-MS/MS. An identical DEPC labeling experiment was carried out in the absence of

CD3 antibody F2B, followed by affinity capture, reduction/alkylation and protease digests. Each digest was carried out in triplicate. Mass-spectrometry derived sequence coverage in the absence of mAb CD3 F2B was high and is shown in FIG. 6.

[00160] *DEPC labeling of CD3delta/epsilon is impacted by binding of mAb CD3 F2B (SEQ ID NO:1 heavy chain and SEQ ID NO:19 light chain).* Peptides obtained from the 3 proteolytic digests were analyzed for significantly reduced DEPC-modified residues due to presence of mAb CD3F2B during labeling. FIG. 7 shows the impacted peptides obtained from the digest with Endopeptidase Glu-C as a representative example.

[00161] The impact on labeling of amino acid residues is considered significant if a) the decrease in DEPC incorporation is larger than 15 -fold and b) is detected in at least 2 of the 3 digests. Impacted CD3 epsilon peptides are shown in FIG. 7. Consistent and significant labeling impact was observed for Lys73 in both the chymotryptic and Glu-C peptides. In addition, Glu-C peptides I57-E86 and D80-E86 provide further evidence that the epitope for mAb F2B extends to at least Lysine 85 within the epsilon chain.

[00162] Impacted CD3 delta peptide were also found. One label-impacted sequence stretch was common to each of the three digest sets. These data suggest an epitope of mAb F2B from Lysine 82 to Cysteine 93. FIG. 8 depicts epitopes identified in the CD3 delta protein (shaded). FIG. 9 shows the crystal structure of CD3 delta/epsilon (PDB ID code 1XIW, Arnett K. et.al, Proc Natl Acad Sci U S A. 2004; 101(46): 16268–16273) with impacted residues in each CD3 subunit shown in space filling mode.

Example 7: CD3 Family 2 antibodies recognize an epitope distinct from OKT3 and SP34

[00163] The same epitope mapping approach was used to confirm known interactions between the therapeutic antibody OKT3 and CD3 delta/epsilon (Salmeron, A., Sanchez-Madrid, F., Ursa, M. A., Fresno, M. & Alarcon, B. (1991) J. Immunol. 147, 3047-3052.). The cross-reactive antibody SP-34 has also been characterized (US patent 8,236,308, 2012). This antibody recognizes an epitope within the extended E-F-loop of CD3 epsilon and does not require CD3 delta for binding.

[00164] Our dataset confirms that CD3 antibody F2B binds a different epitope, compared to OKT3 and SP-34. FIG. 10 shows the alignment of human and cynomolgus ECDs. CD3 mAbs provided herein bind to a loop that is absent in the cynomolgus homolog, which further explains why those antibodies show no monkey cross reactivity to the CD3 complex. In addition, neither OKT3 nor SP-34 bind directly with CD3 delta.

[00165] The examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the present invention, and are not intended to limit the scope of what the inventors regard as their invention nor are they intended to represent that the experiments below are all or the only experiments performed. Efforts have been made to ensure

accuracy with respect to numbers used (e.g. amounts, temperature, etc.) but some experimental errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, molecular weight is weight average molecular weight, temperature is in degrees Centigrade, and pressure is at or near atmospheric.

[00166] While the present invention has been described with reference to the specific embodiments thereof, it should be understood by those skilled in the art that various changes may be made and equivalents may be substituted without departing from the true spirit and scope of the invention. In addition, many modifications may be made to adapt a particular situation, material, composition of matter, process, process step or steps, to the objective, spirit and scope of the present invention. All such modifications are intended to be within the scope of the claims appended hereto.