WHAT IS CLAIMED IS:

1. A method for large scale production of an adenovirus, the method comprising: culturing in a serum-free suspension culture PER.C6 cells infected with an adenovirus which comprises a murine pre-proendothelin promoter, thereby producing the adenovirus.

2. The method of claim 1, wherein said adenovirus is selected from the group consisting of a non-replicating adenovirus and a conditionally replicating adenovirus.

3. The method of claim 2, wherein said non-replicating adenovirus comprises a polynucleotide which comprises a fas-chimera transgene transcriptionally linked to said murine pre-proendothelin promoter.

4. The method of claim 2, wherein said conditionally replicating adenovirus is transcriptionally linked to said murine pre-proendothelin promoter.

5. The method of claim 2, wherein said non-replicating adenovirus comprises a polynucleotide which comprises an anti-angiogenic transgene transcriptionally linked to said murine pre-proendothelin promoter.

6. The method of claim 2, wherein said non-replicating adenovirus comprises a polynucleotide which comprises a pro-angiogenic transgene transcriptionally linked to said murine pre-proendothelin promoter.

7. The method of claim 2, wherein said non-replicating adenovirus comprises a polynucleotide which comprises a suicide transgene transcriptionally linked to said murine pre-proendothelin promoter.

8. The method of claim 2 or 4, wherein said conditionally replicating adenovirus transcriptionally linked to said murine pre-proendothelin promoter is devoid of non-viral heterologous sequences encoding pro- or anti-angiogenic agents.

9. The method of claim 7, wherein said suicide transgene comprises a thymidine kinase.

10. The method of claim 1, wherein said adenovirus further comprises a heterologous nucleic acid sequence encoding a therapeutic agent operably linked to said murine pre-proendothelin promoter.

11. The method of claim 10, wherein said heterologous nucleic acid sequence comprises an apoptotic gene.

12. The method of any one of claims 1-7, further comprising recovering virus from said cells following said culturing.

13. The method of claim 12, wherein said recovering is effected at a point of harvest (POH) of 3-4 days post infection and an MOI of 5.

14. The method of any one of claims 1-7, wherein said culturing is effected at a 5-100 L volume.

15. The method of claim 14, wherein said culturing is effected at a 25 L volume.

16. The method of claim 15, wherein said culturing is effected at a 50 L volume.

17. The method of claim 15, wherein said culturing is effected at a 100 L volume.

18. The method of any one of claims 1-7, wherein said culturing is effected using a disposable bag.

19. The method of claim 12, wherein said recovering is effected by subjecting said cells to a detergent lysis.

20. The method of claim 19, wherein said detergent comprises Triton X-100.

21. The method of claim 19, further comprising removing cellular DNA and cell debris so as to obtain a clear feedstock.

22. The method of claim 21, wherein said feedstock is subjected to Tangential Flow Filtration (TFF).

23. The method of claim 22, further comprising obtaining a viral pellet and subjecting the viral pellet to anion exchange chromatography and size exclusion chromatography.

24. The method of claim 3, wherein said fas-chimera transgene comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 2.

25. The method of claim 3, wherein said fas-chimera transgene comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 3.

26. The method of claim 3, wherein said fas-chimera transgene comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 4.

27. The method of any one of claims 1-7, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 5.

28. The method of any one of claims 1-7, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 6.

29. The method of claim 28, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having at least two copies of said nucleotide sequence as set forth in SEQ ID NO: 6.

30. The method of any one of claims 1-7, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 8.

31. The method of any one of claims 1-5, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 7.

32. The method of any one of claims 1-7, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 13.

33. The method of any one of claims 1-7, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 12.

34. The method of any one of claims 3, 5, 6 or 7, wherein said non-replicating adenovirus vector is an adenovirus 5 vector.

35. The method of claim 3, wherein said adenovirus 5 vector comprises a nucleic acid sequence as set forth in SEQ ID NO: 9 or 10.

36. A method for large scale production of an adenovirus, the method comprising: culturing in a serum-free suspension culture PER.C6 cells infected with an

adenovirus which comprises a nucleic acid sequence as set forth in SEQ ID NO: 9 or 10, thereby producing the adenovirus.

37. A method of producing an adenovirus, the method comprising, culturing PER.C6 cells infected with an adenovirus which comprises a murine pre-proendothelin promoter in an adherent culture under conditions suitable for viral propagation, thereby producing the adenovirus.

38. The method of claim 37, wherein said adenovirus is selected from the group consisting of a non-replicating adenovirus and a conditionally replicating adenovirus.

39. The method of claim 38, wherein said non-replicating adenovirus comprises a polynucleotide which comprises a fas-chimera transgene transcriptionally linked to said murine pre-proendothelin promoter.

40. The method of claim 38, wherein said conditionally replicating adenovirus is transcriptionally linked to said murine pre-proendothelin promoter.

41. The method of claim 38, wherein said non-replicating adenovirus comprises a polynucleotide which comprises an anti-angiogenic transgene transcriptionally linked to said murine pre-proendothelin promoter.

42. The method of claim 38, wherein said non-replicating adenovirus comprises a polynucleotide which comprises a pro-angiogenic transgene transcriptionally linked to said murine pre-proendothelin promoter.

43. The method of claim 38, wherein said non-replicating adenovirus comprises a polynucleotide which comprises a suicide transgene transcriptionally linked to said murine pre-proendothelin promoter.

44. The method of claim 40, wherein said conditionally replicating adenovirus transcriptionally linked to said murine pre-proendothelin promoter is devoid of non-viral heterologous sequences encoding pro- or anti-angiogenic agents.

45. The method of claim 43, wherein said suicide transgene comprises a thymidine kinase.

46. The method of claim 37, wherein said adenovirus further comprises a heterologous nucleic acid sequence encoding a therapeutic agent operably linked to said murine pre-proendothelin promoter.

47. The method of claim 46, wherein said heterologous nucleic acid sequence comprises an apoptotic gene.

48. The method of any one of claims 37-42, wherein said conditions comprise serum.

49. The method of any one of claims 37-42, further comprising recovering the virus from said cells following said culturing.

50. The method of claim 49, wherein said recovering is effected at a point of harvest (POH) of 3-4 days post infection.

51. The method of claim 49, wherein said recovering is effected by freeze-thaw releasing of the virus.

52. The method of claim 51, further comprising removing cellular DNA and cell debris so as to obtain a clear feedstock by ultracentrifugation.

53. The method of claim 52, further comprising centrifuging said clear feedstock on a CsCl gradient.

54. The method of claim 53, further comprising removing said CsCl using a Sephadex desalting column.

55. The method of claim 39, wherein said fas-chimera transgene comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 2.

56. The method of claim 39, wherein said fas-chimera transgene comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 3.

57. The method of claim 39, wherein said fas-chimera transgene comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 4.

58. The method of claim 37, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 5.

59. The method of claim 37, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 6.

60. The method of claim 59, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having at least two copies of said nucleotide sequence as set forth in SEQ ID NO: 6.

61. The method of claim 37, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 8.

62. The method of claim 37, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 7.

63. The method of claim 37, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 13.

64. The method of claim 37, wherein said murine pre-pro endothelin promoter comprises a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 12.

65. The method of claim 38, wherein said non-replicating adenovirus vector is an adenovirus 5 vector.

66. The method of claim 39, wherein said adenovirus 5 vector comprises a nucleic acid sequence as set forth in SEQ ID NO: 9 or 10.

67. A method of producing an adenovirus, the method comprising, culturing PER.C6 cells infected with an adenovirus comprising a nucleic acid sequence as set forth in SEQ ID NO: 9 or 10 in an adherent culture under conditions suitable for viral propagation, thereby producing the adenovirus.

68. A viral preparation generated according to the method of any one of claims 1-36 and exhibiting an ion exchange and size exclusion chromatography traces of Figures 7A-B and product profile of Table 6.

69. A viral preparation generated according to the method of any one of claims 37-67 and having a product profile of Table 3.

70. A pharmaceutical composition comprising as an active ingredient the viral preparation of claim 68 or 69.

71. A method of reducing angiogenesis in a subject in need thereof, the method comprising administering to the subject a therapeutically effective amount of the viral preparation of claim 68 or 69, thereby reducing angiogenesis in the subject.

72. The method of claim 71, wherein the subject has a solid tumor.

73. The method of claim 71, wherein said administering comprises intravenous administration.

WHAT IS CLAIMED IS:

1. A method of reducing angiogenesis in a subject in need thereof, the method comprising administering to the subject a therapeutically effective amount of the viral preparation generated according to the method for large scale production of an adenovirus, the method comprising: culturing in a serum-free suspension culture PER.C6 cells infected with an adenovirus which comprises a murine pre-proendothelin promoter, thereby producing the adenovirus, and exhibiting an ion exchange and size exclusion chromatography traces of Figures 7A-B and product profile of Table 6, thereby reducing angiogenesis in a subject.

2. The method of claim 1, wherein the subject has a solid tumor.

3. The method of claim 1, wherein said administering comprises intravenous administration.