

APOC-II MIMETIC PEPTIDES

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

[0001] This invention was created in the performance of Cooperative Research and Development Agreement No. HL-CR-16-005 with the National Institutes of Health, an Agency of the Department of Health and Human Services. The Government of the United States has certain rights in this invention.

SEQUENCE LISTING

[0002] The instant application contains a Sequence Listing which has been submitted via EFS-Web and is hereby incorporated herein by reference in its entirety. Said ASCII copy, created on Month XX, 2018, is named XXXXXUS_sequencelisting.txt, and is X,XXX,XXX bytes in size.

BACKGROUND

[0003] Hypertriglyceridemia is typically defined as serum triglyceride levels over 150 mg/dL (Berglund et al., *J. Clin. Endocrinol. Metab.* 97:2969-2989, 2012). Patients with moderate increases in triglycerides are at risk for cardiovascular disease (Budoff, *Am J Cardiol* 118:138-45, 2016). Patients with severe hypertriglyceridemia are additionally at risk for acute pancreatitis (Viljoen and Wierzbicki, *Expert Rev Cardiovasc Ther* 10:505-514, 2012).

[0004] Hypertriglyceridemia is most commonly secondary to obesity, diabetes mellitus, pregnancy, alcohol and a wide variety of drugs (Anderson et al., *Pancreatology* 9:252-257, 2009; Dominguez-Munoz et al., *Int. J. Pancreatol.* 10:261-267, 1991). Triglycerides are enriched in chylomicrons and very low density lipoprotein (VLDL), and lipolysis by lipoprotein lipase (LPL) is a critical step in the catabolism of these triglyceride-rich lipoprotein particles. Rarely, hypertriglyceridemia is the result of genetic defects in LPL or apoC-II, an obligate activator of LPL (Fojo et al., *J. Intern. Med.* 231:669-677, 1992) or in other genes.

[0005] Fibrates and supplements rich in omega-3 polyunsaturated fatty acids, such as fish oils, are the main treatment for hypertriglyceridemia (Berglund et al., *J. Clin. Endocrinol. Metab.* 97:2969-2989, 2012). However, it has not been clearly demonstrated that these agents are efficacious in lowering the risks associated with elevated triglycerides, such as cardiovascular disease (Budoff, *Am J Cardiol* 118:138-45, 2016).

[0006] There is, therefore, a need for new therapeutic agents for treatment of hypertriglyceridemia, both common forms of hypertriglyceridemia and hypertriglyceridemia caused by genetic defects.

SUMMARY

[0007] We have designed and produced apoC-II mimetic peptides that possess the ability to lower the triglyceride level both *in vitro* and *in vivo*. These peptides can be used for treating hypertriglyceridemia.

[0008] In typical embodiments, the apoC-II mimetic peptide is a multihelical peptide in which one or more of the helical domains is amphipathic, conferring on the peptide the ability to bind to lipids and/or to the surface of lipoproteins, and another of the helical domains activates LPL. In certain embodiments, the apoC-II mimetic peptide is a bihelical peptide comprising a first helical domain, a hinge region, and a second helical domain in order from N-terminus to C-terminus, in which the first helical domain is amphipathic and the second helical domain activates LPL. We have found that a variety of amphipathic helices are suitable, with some modeled on amphipathic helices of known apolipoproteins, such as apoC-II. We have also found that the peptides' ability to lower triglyceride is not limited to activation of LPL, allowing their use in disorders in which LPL is diminished or absent, and that they can displace apoC-III from the surface of lipoproteins, allowing their use in disorders in which apoC-III levels are elevated.

[0009] Accordingly, in a first aspect, provided herein is an isolated apoC-II mimetic peptide of no more than 50 amino acids, comprising from N-terminus to C-terminus: a first helical domain, a hinge region, and a second helical domain, wherein the first helical domain is amphipathic, wherein the apoC-II mimetic peptide is not a peptide of SEQ ID NO: 56.

[0010] In some embodiments, the first domain of the isolated apoC-II mimetic peptide is native helix 1 of apoC-II. In some other embodiments, the first domain of the isolated apoC-II mimetic peptide is a variant of helix 1 of apoC-II. In some embodiments, the first domain of the isolated apoC-II mimetic peptide is native helix 2 of apoC-II. In some other embodiments, the first domain of the isolated apoC-II mimetic peptide is a variant of helix 2 of apoC-II.

[0011] In some embodiments, the variant of helix 2 of apoC-II comprises elongation of helix 2 of ApoC-II. In various embodiments, the helix 2 of apoC-II is elongated by 1-10 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is elongated by 1 amino acid at the N-terminus. In some of these embodiments, the amino acid is aspartic acid.

In certain embodiments, the helix 2 of apoC-II is elongated by 2 amino acids at the N-terminus. In some of these embodiments, the amino acids are lysine and alanine from N-terminus to C-terminus. In certain embodiments, the helix 2 of apoC-II is elongated by 5 amino acids at the N-terminus. In some of these embodiments, the helix 2 of apoC-II is elongated by the native upstream amino acid sequence of human apoC-II, or a mutant of native upstream amino acid sequence of human apoC-II. In specific embodiments, the variant of helix 2 of apoC-II comprises an aspartic acid at position 40. In specific embodiments, the variant of helix 2 of apoC-II comprises a tyrosine or a tryptophan at position 41. In specific embodiments, the variant of helix 2 of apoC-II comprises a leucine at position 42. In specific embodiments, the variant of helix 2 of apoC-II comprises a lysine or an arginine at position 43. In specific embodiments, the variant of helix 2 of apoC-II comprises an alanine or a glutamic acid at position 44. In particular embodiments, the variant of helix 2 of apoC-II comprises an aspartic acid at position 40, a tyrosine at position 41, a leucine at position 42, a lysine at position 43, and a glutamic acid at position 44.

[0012] In some embodiments, the variant of helix 2 of apoC-II comprises truncation of helix 2 of apoC-II. In some of these embodiments, the amino acid residues 45-46 of native helix 2 of apoC-II are deleted. In some of these embodiments, the amino acid residues 45-48 of native helix 2 of apoC-II are deleted. In some of these embodiments, the amino acid residues 45-49 of native helix 2 of apoC-II are deleted. In some of these embodiments, the amino acid residues 45-50 of native helix 2 of apoC-II are deleted. In some of these embodiments, the amino acid residues 45-53 of native helix 2 of apoC-II are deleted.

[0013] In some embodiments, the variant of helix 2 of apoC-II comprises at least one mutation of helix 2 of apoC-II. In some of these embodiments, the mutation is an amino acid substitution. In certain embodiments, an original amino acid of helix 2 of apoC-II is substituted by a natural amino acid. In certain embodiments, an original amino acid of helix 2 of apoC-II is substituted by an unnatural amino acid. In certain embodiments, an original amino acid of helix 2 of apoC-II is substituted by an amino acid analog. In specific embodiments, the amino acid substitution is at position 45. In some of these embodiments, the valine at position 45 is substituted by phenylalanine. In specific embodiments, the amino acid substitution is at position 46. In some of these embodiments, the aspartic acid at position 46 is substituted by a phenylalanine. In some of these embodiments, the aspartic acid at position 46 is substituted by a lysine. In some of these embodiments, the aspartic acid at position 46 is substituted by an aminoisobutyric acid. In specific embodiments, the amino

acid substitution is at position 48. In some of these embodiments, the lysine at position 48 is substituted by an arginine. In specific embodiments, the amino acid substitution is at position 49. In some of these embodiments, the leucine at position 49 is substituted by a lysine. In specific embodiments, the amino acid substitution is at position 50. In some of these embodiments, the arginine at position 50 is substituted by a lysine. In specific embodiments, the amino acid substitution is at position 53. In some of these embodiments, the tyrosine at position 53 is substituted by a leucine. In specific embodiments, the amino acid substitution is at position 54. In some of these embodiments, the serine at position 54 is substituted by a glutamic acid. In some of these embodiments, the serine at position 54 is substituted by a lysine. In some of these embodiments, the serine at position 54 is substituted by an aspartic acid. In specific embodiments, the amino acid substitution is at position 55. In some of these embodiments, the lysine at position 55 is substituted by an arginine. In specific embodiments, the amino acid substitution is at position 56. In some of these embodiments, the serine at position 56 is substituted by a phenylalanine. In some of these embodiments, the serine at position 56 is substituted by a lysine. In some of these embodiments, the serine at position 56 is substituted by an alanine. In some of these embodiments, the serine at position 56 is substituted by an aminoisobutyric acid. In specific embodiments, the amino acid substitution is at position 57. In some of these embodiments, the threonine at position 57 is substituted by a phenylalanine. In specific embodiments, the variant of helix 2 of apoC-II comprises three amino acid substitutions, wherein the amino acid substitutions are at position 46, position 54, and position 56. In some of these embodiments, the aspartic acid at position 46 is substituted by a phenylalanine, the serine at position 54 is substituted by a glutamic acid, and the serine at position 56 is substituted by a phenylalanine.

[0014] In some embodiments, the variant of helix 2 of apoC-II comprises at least one chemical modification. In certain embodiments, the chemical modification is a covalent linkage of a fatty acid. In various embodiments, the fatty acid comprises 4 to 26 carbons. In specific embodiments, the fatty acid comprises 10 carbons. In specific embodiments, the fatty acid comprises 12 carbons. In specific embodiments, the fatty acid comprises 14 carbons. In specific embodiments, the fatty acid comprises 16 carbons. In specific embodiments, the fatty acid comprises 18 carbons. In specific embodiments, the fatty acid comprises 20 carbons. In some embodiments, the fatty acid is saturated. In some other embodiments, the fatty acid is unsaturated. In certain embodiments, the fatty acid is linked to the N-terminal amino acid. In

certain embodiments, the fatty acid is linked to an amino acid side group. In certain embodiments, the fatty acid is linked to the ϵ -amine of a lysine residue.

[0015] In some embodiments, the hinge region of the isolated apoC-II mimetic peptide comprises 5-10 amino acids. In certain embodiments, the hinge region comprises 7 amino acids. In specific embodiments, the hinge region comprises a proline. In some of these embodiments, the hinge region comprises a proline at position 58. In specific embodiments, the hinge region comprises an alanine. In some of these embodiments, the hinge region comprises an alanine at position 58. In specific embodiments, the hinge region comprises a norleucine. In some of these embodiments, the hinge region comprises a norleucine at position 60. In specific embodiments, the hinge region comprises a lysine. In some of these embodiments, the hinge region comprises a lysine at position 60. In specific embodiments, the hinge region comprises a valine. In some of these embodiments, the hinge region comprises a valine at position 60. In specific embodiments, the hinge region comprises a leucine. In some of these embodiments, the hinge region comprises a leucine at position 60. In specific embodiments, the hinge region comprises a methionine. In some of these embodiments, the hinge region comprises a methionine at position 60.

[0016] In some embodiments, the hinge region comprises at least one chemical modification. In certain embodiments, the chemical modification is a covalent linkage of a fatty acid. In various embodiments, the fatty acid comprises 4 to 26 carbons. In specific embodiments, the fatty acid comprises 10 carbons. In specific embodiments, the fatty acid comprises 12 carbons. In specific embodiments, the fatty acid comprises 14 carbons. In specific embodiments, the fatty acid comprises 16 carbons. In specific embodiments, the fatty acid comprises 18 carbons. In specific embodiments, the fatty acid comprises 20 carbons. In some embodiments, the fatty acid is saturated. In some other embodiments, the fatty acid is unsaturated. In certain embodiments, the fatty acid is linked to an amino acid side group. In certain embodiments, the fatty acid is linked to the ϵ -amine of a lysine residue.

[0017] In various embodiments, the hinge region allows the first helical domain and the second helical domain to retain a nearly straight conformation, wherein the second domain bends away from the first domain over an angle of no more than about 20°.

[0018] In some embodiments, the second domain of the isolated apoC-II mimetic peptide is native helix 3 of apoC-II. In some other embodiments, the second domain of the isolated apoC-II mimetic peptide is a variant of helix 3 of apoC-II.

[0019] In some embodiments, the variant of helix 3 of apoC-II comprises elongation of helix 3 of apoC-II. In various embodiments, the helix 3 of apoC-II is elongated by 1-10 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 4 amino acids at the C-terminus. In specific embodiments, the helix 3 of apoC-II is elongated by amino acids lysine, glycine, glutamic acid, and glutamic acid from N-terminus to C-terminus.

[0020] In some embodiments, the variant of helix 3 of apoC-II comprises at least one mutation of helix 3 of apoC-II. In some of these embodiments, the mutation is an amino acid substitution. In certain embodiments, an original amino acid of helix 3 of apoC-II is substituted by a natural amino acid. In certain embodiments, an original amino acid of helix 3 of apoC-II is substituted by an unnatural amino acid. In certain embodiments, an original amino acid of helix 3 of apoC-II is substituted by an amino acid analog. In specific embodiments, the amino acid substitution is at position 70. In some of these embodiments, the glutamine at position 70 is substituted by an arginine. In some of these embodiments, the glutamine at position 70 is substituted by a lysine.

[0021] In some embodiments, the variant of helix 3 of apoC-II comprises at least one chemical modification. In certain embodiments, the chemical modification is a covalent linkage of a fatty acid. In various embodiments, the fatty acid comprises 4 to 26 carbons. In specific embodiments, the fatty acid comprises 10 carbons. In specific embodiments, the fatty acid comprises 12 carbons. In specific embodiments, the fatty acid comprises 14 carbons. In specific embodiments, the fatty acid comprises 16 carbons. In specific embodiments, the fatty acid comprises 18 carbons. In specific embodiments, the fatty acid comprises 20 carbons. In some embodiments, the fatty acid is saturated. In some other embodiments, the fatty acid is unsaturated. In certain embodiments, the fatty acid is linked to the C-terminal amino acid. In certain embodiments, the fatty acid is linked to an amino acid side group. In certain embodiments, the fatty acid is linked to the ϵ -amine of a lysine residue. In certain embodiments, the C-terminal amino acid is modified by a C-terminal amide.

[0022] In various embodiments, the isolated apoC-II mimetic peptide further comprises a purification tag. In some of these embodiments, the purification tag is a polyhistidine-tag, a myc-tag, or an HA-tag.

[0023] In various embodiments, the first domain of the apoC-II mimetic peptide has an affinity for binding to lipoproteins. In various embodiments, the second domain is capable of activating lipoprotein lipase (LPL). In some embodiments, the peptide is capable of activating lipolysis by LPL. In some embodiments, the peptide is capable of displacing apoC-III in

lipoproteins. In some embodiments, the peptide is capable of reducing triglyceride (TG) level *in vivo*. In some of these embodiments, the peptide is capable of reducing elevated TG level caused by LPL deficiency. In some of these embodiments, the peptide is capable of reducing elevated TG level caused by elevated apoC-III. In some of these embodiments, the peptide is capable of reducing elevated TG level caused by apoC-II deficiency. In some of these embodiments, the peptide is capable of reducing post-prandial elevated TG level.

[0024] In specific embodiments, the peptide has the sequence set forth in SEQ ID NO: 6.

[0025] In specific embodiments, the peptide has the sequence set forth in SEQ ID NO: 9.

[0026] In another aspect, provided herein is an isolated apoC-II mimetic peptide of no more than 30 amino acids, consisting of a variant of helix 3 of apoC-II.

[0027] In some embodiments, the variant of helix 3 of apoC-II comprises elongation of helix 3 of apoC-II. In various embodiments, the helix 3 of apoC-II is elongated by 1-10 amino acids at the N-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 6 amino acids at the N-terminus. In specific embodiments, the variant of helix 3 of apoC-II comprises a lysine or methionine at position 60. In various embodiments, the helix 3 of apoC-II is elongated by 1-10 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 4 amino acids at the C-terminus. In specific embodiments, the helix 3 of apoC-II is elongated by amino acids lysine, glycine, glutamic acid, and glutamic acid from N-terminus to C-terminus.

[0028] In some embodiments, the variant of helix 3 of apoC-II comprises at least one mutation of helix 3 of apoC-II. In some of these embodiments, the mutation is an amino acid substitution. In certain embodiments, an original amino acid of helix 3 of apoC-II is substituted by a natural amino acid. In certain embodiments, an original amino acid of helix 3 of apoC-II is substituted by an unnatural amino acid. In certain embodiments, an original amino acid of helix 3 of apoC-II is substituted by an amino acid analog.

[0029] In some embodiments, the variant of helix 3 of apoC-II comprises at least one chemical modification. In certain embodiments, the chemical modification is a covalent linkage of a fatty acid. In various embodiments, the fatty acid comprises 4 to 26 carbons. In specific embodiments, the fatty acid comprises 10 carbons. In specific embodiments, the fatty acid comprises 12 carbons. In specific embodiments, the fatty acid comprises 14 carbons. In specific embodiments, the fatty acid comprises 16 carbons. In specific embodiments, the fatty acid comprises 18 carbons. In specific embodiments, the fatty acid comprises 20 carbons. In some embodiments, the fatty acid is saturated. In some other embodiments, the fatty acid is

unsaturated. In certain embodiments, the fatty acid is linked to the N-terminal amino acid. In certain embodiments, the fatty acid is linked to the C-terminal amino acid. In certain embodiments, the fatty acid is linked to an amino acid side group. In certain embodiments, the fatty acid is linked to the ϵ -amine of a lysine residue.

[0030] In various embodiments, the isolated apoC-II mimetic peptide further comprises a purification tag. In some of these embodiments, the purification tag is a polyhistidine-tag, a myc-tag, or an HA-tag.

[0031] In various embodiments, the variant of helix 3 of apoC-II has an affinity for binding to lipoproteins. In various embodiments, the variant of helix 3 of apoC-II is capable of activating lipoprotein lipase (LPL). In some embodiments, the peptide is capable of activating lipolysis by LPL. In some embodiments, the peptide is capable of displacing apoC-III in lipoproteins. In some embodiments, the peptide is capable of reducing triglyceride (TG) level *in vivo*. In some of these embodiments, the peptide is capable of reducing elevated TG level caused by LPL deficiency. In some of these embodiments, the peptide is capable of reducing elevated TG level caused by elevated apoC-III. In some of these embodiments, the peptide is capable of reducing elevated TG level caused by apoC-II deficiency. In some of these embodiments, the peptide is capable of reducing post-prandial elevated TG level.

[0032] In another aspect, provided herein is a pharmaceutical composition comprising the apoC-II mimetic peptide and a pharmaceutically acceptable carrier. In some embodiments, the pharmaceutical composition is suitable for subcutaneous injection.

[0033] In another aspect, provided herein is a method of treating hypertriglyceridemia in a patient, comprising administering to the patient an effective amount of the apoC-II mimetic peptide or the pharmaceutical composition comprising the apoC-II mimetic peptide. In some embodiments, the hypertriglyceridemia is associated with obesity. In some embodiments, the hypertriglyceridemia is associated with diabetes mellitus. In some embodiments, the hypertriglyceridemia is associated with alcohol consumption. In some embodiments, the hypertriglyceridemia is associated with medication.

[0034] In some embodiments, the hypertriglyceridemia is caused by LPL deficiency. In certain embodiments, the hypertriglyceridemia is familial lipoprotein lipase deficiency. In some embodiments, the LPL deficiency is caused by a mutation the LPL gene. In certain embodiments, the mutation leads to reduced LPL enzyme activity. In certain embodiments, the mutation leads to absent LPL enzyme activity. In some embodiments, the LPL deficiency is diagnosed by the absence of LPL activity in serum of the patient. In some embodiments, the

mutation is detected by DNA sequence analysis. In some embodiments, the hypertriglyceridemia is caused by apoC-II deficiency. In some embodiments, the hypertriglyceridemia is caused by elevated apoC-III.

[0035] In certain embodiments, the pre-treatment serum triglyceride (TG) concentration of the patient is between 150 mg/dL and 199 mg/dL. In certain embodiments, the pre-treatment serum triglyceride (TG) concentration of the patient is between 200 mg/dL to 499 mg/dL. In certain embodiments, the pre-treatment serum triglyceride (TG) concentration of the patient is between 500 mg/dL to 999 mg/dL. In certain embodiments, the pre-treatment serum triglyceride (TG) concentration of the patient is between 1000 mg/dL and 1999 mg/dL. In certain embodiments, the pre-treatment serum triglyceride (TG) concentration of the patient is equal to or higher than 2000 mg/dL.

[0036] In specific embodiments, the patient has developed or is at risk for acute pancreatitis. In specific embodiments, the patient has developed or is at risk for acute cardiovascular disease.

[0037] In another aspect, also provided herein is a method of making the apoC-II mimetic peptide, comprising producing the peptide recombinantly.

[0038] In yet another aspect, also provided herein is a method of making the apoC-II mimetic peptide, comprising producing the peptide by chemical synthesis.

BRIEF DESCRIPTION OF THE DRAWINGS

[0039] These and other features, aspects, and advantages of the present invention will become better understood with regard to the following description, and accompanying drawings, where:

[0040] FIG. 1 shows the homology of amino acid residues 40-58 of mature human apoC-II protein across various species.

[0041] FIG. 2A and FIG. 2B present helical wheel plots of amino acid residues 40-57 of native apoC-II and a variant Delta4', with FIG. 2A representing the native apoC-II and FIG. 2B representing the variant, Delta4'. The size and color of balls indicate the degree of charge and hydrophobicity. The first number in the center of the helical wheel indicates hydrophobic moment. The second number in the center of the helical wheel and the arrow indicate the angle of hydrophobic moment.

[0042] **FIG. 3A and FIG. 3B** present helical wheel plots of amino acid residues 40-57 of native apoC-II and a variant Delta5', with FIG. 3A representing the native apoC-II and FIG. 3B representing the variant, Delta5'.

[0043] **FIG. 4A and FIG. 4B** present helical wheel plots of amino acid residues 40-57 of native apoC-II and a variant Delta6', with FIG. 4A representing the native apoC-II and FIG. 4B representing the variant, Delta6'.

[0044] **FIG. 5** shows examples of chemical modification of the variant Delta4'. Mono-acyl-Delta4' includes a stearic acid modified N-terminus. Di-acyl-Delta4' includes a stearic acid modified N-terminus and a stearic acid modified leucine residue at position 49.

[0045] **FIG. 6** presents the results of *in vitro* LPL assay of exemplary apoC-II mimetic peptides and variants with chemical modifications using apoC-II deficient patient sera as substrate. Native full-length human apoC-II protein was used as control.

[0046] **FIG. 7** presents the results of *in vitro* LPL assay of exemplary apoC-II mimetic peptides and variants with chemical modifications using Intralipid as substrate. Native full-length human apoC-II protein was used as control.

[0047] **FIG. 8** presents the results of *in vitro* LPL assay of exemplary apoC-II mimetic peptide Delta4, Delta4 variants, and variants of the third helix of apoC-II using apoC-II deficient patient sera as substrate.

[0048] **FIG. 9** shows the results of *in vitro* LPL assay of variants of the third helix of apoC-II using apoC-II deficient patient sera as substrate.

[0049] **FIG. 10** shows the results of *in vitro* LPL assay of variants of exemplary apoC-II mimetic peptide Delta4 using apoC-II deficient patient sera as substrate.

[0050] **FIG. 11** shows the results of *in vitro* LPL assay of variants of exemplary apoC-II mimetic peptide Delta4 using apoC-II deficient patient sera as substrate.

[0051] **FIG. 12** presents the results of *in vitro* LPL assay of exemplary apoC-II mimetic peptide Delta4 and its variants using apoC-II deficient patient sera as substrate. The concentration of apoC-II mimetic peptide ranges from 100 pM to 10 μ M. Native full-length human apoC-II protein was used as control.

[0052] **FIG. 13** presents the results of *in vitro* LPL assay of truncation variants of exemplary apoC-II mimetic peptide Delta4 using apoC-II deficient patient sera as substrate. The concentration of apoC-II mimetic peptide ranges from 10 pM to 100 μ M.

[0053] **FIG. 14** presents the results of *in vitro* LPL assay of variants of exemplary apoC-II mimetic peptide Delta5 and the exemplary apoC-II mimetic peptide Delta6, using apoC-II deficient patient sera as substrate. The concentration of apoC-II mimetic peptide ranges from 100 pM to 10 μ M.

[0054] **FIG. 15A, FIG. 15B, FIG. 15C, and FIG. 15D** show the percentage change of LPL activity in an *in vitro* LPL assay of exemplary apoC-II mimetic peptide Delta6 and its variants using hypertriglyceridemia patient sera as substrate. The triglyceride concentration of the patient sera in FIG. 15A, FIG. 15B, FIG. 15C, and FIG. 15D are 1084 mg/dL, 1224 mg/dL, 1442 mg/dL, and 1669 mg/dL respectively.

[0055] **FIG. 16** presents the serum triglyceride (TG) level of apoC-II knockout mice after subcutaneous injection of apoC-II mimetic peptide Delta4. Saline was used as control.

[0056] **FIG. 17** presents the total cholesterol (TC) level of apoC-II knockout mice after subcutaneous injection of apoC-II mimetic peptide Delta4. Saline was used as control.

[0057] **FIG. 18** presents the serum triglyceride (TG) level of apoC-II knockout mice after intraperitoneal injection of apoC-II mimetic peptide Delta4. Saline was used as control.

[0058] **FIG. 19** presents the total cholesterol (TC) level of apoC-II knockout mice after intraperitoneal injection of apoC-II mimetic peptide Delta4. Saline was used as control.

[0059] **FIG. 20** shows the percentage change of serum triglyceride (TG) level of wild-type mice after vegetable oil gavage in the presence or absence of intraperitoneal injection of apoC-II mimetic peptide. The tested apoC-II mimetic peptides included Delta4, Delta4b and Delta4b-3. Saline was used as control.

[0060] **FIG. 21** presents the serum triglyceride (TG) level of apoC-II knockout mice after intraperitoneal injection of apoC-II mimetic peptide Delta6. Saline was used as control.

[0061] **FIG. 22** presents the total cholesterol (TC) level of apoC-II knockout mice after intraperitoneal injection of apoC-II mimetic peptide Delta6. Saline was used as control.

[0062] **FIG. 23** presents the serum triglyceride (TG) level of apoC-II knockout mice after intraperitoneal injection of different doses of apoC-II mimetic peptide Delta6. Saline was used as control. The percentage of the serum TG level at different time points after the injection compared to the serum TG level at the time of injection was calculated.

[0063] **FIG. 24** presents the percentage change of serum triglyceride (TG) of apoC-II knockout mice after intraperitoneal injection of Delta6 variants. Saline was used as control.

The percentage change of the serum TG compared to the serum TG at the time of injection was calculated.

[0064] **FIG. 25** shows apoC-III displacement by the apoC-II mimetic peptide Delta6 in VLDL and chylomicrons isolated from human plasma.

[0065] **FIG. 26** shows apoC-III displacement by variants of the apoC-II mimetic peptide Delta6 in VLDL isolated from human plasma.

[0066] **FIG. 27** shows the results of *in vitro* LPL assay of apoC-II mimetic peptides Delta4 and Delta6 using hypertriglyceridemia patient sera with high apoC-III levels as substrate.

[0067] **FIG. 28** shows apoC-III displacement by apoC-II-a peptide in VLDL isolated from human plasma (TG = 291mg/dL).

[0068] **FIG. 29A and FIG. 29B** show the dose-dependent displacement of apoC-III by apoC-II-a peptide analyzed by mass spectrometry (MALDI-TOF), with FIG. 29A showing that apoC-II-a selectively displaces apoC-III and apoC-I but not apoC-II, and FIG. 29B showing that the binding of apoC-II-a peptide to VLDL is approximately proportional to its concentration.

[0069] **FIG. 30** shows apoC-III displacement by apoC-II-a peptide in chylomicrons isolated from human plasma.

[0070] **FIG. 31** presents results from an LPL activation assay with Intralipid as substrate and recombinant apoC-III as an inhibitor. ApoC-II-a peptide overcomes the inhibition by apoC-III.

[0071] **FIG. 32** presents results from an LPL activation assay with high TG human plasma as substrate and recombinant apoC-III as an inhibitor. ApoC-II-a peptide overcomes the inhibition by apoC-III.

[0072] **FIG. 33** graphs results from an LPL activation assay using apoC-III transgenic (tg) mice plasma as substrate. The TG concentrations of ApoC-III tg1, ApoC-III tg2, and ApoC-III tg3 mice are 323 mg/dL, 497 mg/dL, and 478 mg/dL, respectively. ApoC-II-a peptide overcomes the inhibitory effect of elevated expression of apoC-III in all three transgenic mice plasma samples.

[0073] **FIG. 34A and FIG. 34B** show 24 hour serum post-prandial TG levels in mice after an oral gavage with vegetable oil in the presence or absence of the LPL inhibitor Triton WR1339, and in the presence or absence of apoC-II-a peptide, with FIG. 34A focusing on the

TG concentration range of 0-100 mg/dL, and FIG. 34B showing the TG concentration range of 0-4500 mg/dL.

[0074] FIG. 35A and FIG. 35B show the percentage change of serum TG level of wild-type mice after Intralipid injection with or without the intraperitoneal injection of the Delta6PV peptide in the presence or absence of the LPL inhibitor Triton, with FIG. 35A showing the timeline of injection and FIG. 35B showing the percentage change of serum TG level after intraperitoneal injection of Delta6PV in the presence or absence of Triton. Saline was used as control.

[0075] FIG. 36A, FIG. 36B, and FIG. 36C show the percentage change of serum TG level of apoC-II knockout mice after injection of apoC-II mimetic peptide Delta6PV, with FIG. 36A showing the timeline of injection, FIG. 36B showing the percentage change of serum TG level after intraperitoneal injection of Delta6PV, and FIG. 36C showing the percentage change of serum TG level after subcutaneous injection of the Delta6PV. Saline was used as control. SM represents the conditions where Delta6PV was mixed with sphingomyelin (SM), a component of cell membrane.

[0076] FIG. 37A and FIG. 37B show the percentage change of serum TG level of apoC-II knockout mice after Intralipid injection with or without the intraperitoneal injection of Delta6PV peptide, with FIG. 37A showing the timeline of injection and FIG. 37B showing the percentage change of serum TG level after intraperitoneal injection of Delta6PV at 5 μ mole/kg and 10 μ mole/kg (B. W.). Saline was used as control.

[0077] FIG. 38A, and FIG. 38B show the percentage change of serum TG level of apoC-III transgenic mice after injection of apoC-II mimetic peptide Delta6PV, with FIG. 38A showing the timeline of injection, FIG. 38B showing the percentage change of serum TG level after intraperitoneal injection of Delta6PV at 1 μ mole/kg, 5 μ mole/kg, and 10 μ mole/kg (B. W.). Saline was used as control.

[0078] FIG. 39 shows the percentage change of serum TG level of apoC-III transgenic mice after Intralipid injection with or without the intraperitoneal injection of Delta6PV peptide at 5 μ mole/kg and 10 μ mole/kg (B. W.). Saline was used as control.

[0079] FIG. 40 shows the percentage change of serum TG level of apoC-II knockout mice after multiple injections of apoC-II mimetic peptide Delta6PV at 0.25 μ mole/kg and 1 μ mole/kg (B. W.). The arrows indicate the time points of Delta6PV injection. Saline was used as control.

[0080] **FIG. 41** presents the results of SEC-FPLC separation of various lipid fractions in the 1 μ mole/kg (B. W.) Delta6PV administered apoC-II knockout mice. The level of lipoprotein particles is represented by phospholipid.

[0081] **FIG. 42** presents the results of SEC-FPLC separation of various lipid fractions in the 1 μ mole/kg Delta6PV administered apoC-II knockout mice. The level of lipoprotein particles is represented by TG.

[0082] **FIG. 43A and FIG. 43B** show the percentage change of serum TG level of apoC-III transgenic mice after multiple injections of apoC-II mimetic peptide Delta6PV, with **FIG. 43A** showing the timeline of injections and **FIG. 43B** showing the percentage change of serum TG level after intraperitoneal injection of Delta6PV at 5 μ mole/kg (B. W.). The arrows indicate the time points of Delta6PV injection. Saline was used as control.

[0083] **FIG. 44** shows the percentage change of serum TG level of apoC-III transgenic mice after multiple injections of apoC-II mimetic peptide Delta6PV at 5 μ mole/kg (B. W.). The arrows indicate the time points of Delta6PV injection. Saline was used as control.

[0084] **FIG. 45A and FIG. 45B** show the apoC-III level after Delta6PV injections in apoC-III transgenic mice, as measured by ELISA, with **FIG. 45A** showing the apoC-III level in VLDL particles on Day 1 and Day 5 before and 3 hours after injection and **FIG. 45B** showing the apoC-III level in HDL particles on Day 1 and Day 5 before and 3 hours after injection.

[0085] **FIG. 46A and FIG. 46B** show the percentage change of serum TG level of wild-type mice after Intralipid injection with or without multiple intraperitoneal injections of Delta6PV peptide, with **FIG. 46A** showing the timeline of injection and **FIG. 46B** showing the percentage change of serum TG level after intraperitoneal injection of Delta6PV at 0.25 μ mole/kg and 1 μ mole/kg (B. W.). Saline was used as control.

[0086] **FIG. 47** shows the ratio of serum TG in the obese monkey model with or without intravenous injection of Delta6PV peptide at 1 mg/kg and 5 mg/kg (B. W.) as compared to the baseline level. Saline was used as control.

[0087] **FIG. 48A, FIG. 48B, and FIG. 48C** show the percentages of VLDL, LDL, and HDL in the obese monkey model with or without intravenous injection of Delta6PV peptide at 1 mg/kg and 5 mg/kg (B. W.) as compared to the baseline levels, with **FIG. 48A** showing the percentage of VLDL in the obese monkey model as compared to the baseline level, **FIG. 48B** showing the percentage of LDL in the obese monkey model as compared to the baseline level, and **FIG. 48C** showing the percentage of HDL in the obese monkey model as compared to the baseline level.

[0088] **FIG. 49** shows apoC-III displacement by Delta6PV peptide in VLDL isolated from human plasma.

[0089] **FIG. 50** presents results from an LPL activation assay with high TG human plasma as substrate and recombinant apoC-III as an inhibitor. Delta6PV peptide overcomes the inhibition of apoC-III.

[0090] **FIG. 51** represents the results of *in vitro* LPL assay of Delta6 and Delta6PV peptides using apoC-II deficient patient sera as substrate.

[0091] **FIG. 52** represents the results of *in vitro* LPL assay of apoC-II mimetic peptides Delta6PV, Delta6-T18F-PV, Delta6-T18F-PV-AIB7, 17 and Delta13-31-PV using apoC-II deficient patient sera as substrate.

[0092] **FIG. 53** presents the percentage change of serum TG in apoC-II knockout mice after intraperitoneal injection of Delta6 variants, Delta6PV and Delta6-T18F-PV. Saline was used as control. The percentage change of the serum TG compared to the serum TG at the time of injection was calculated.

[0093] **FIG. 54A, FIG. 54B, and FIG. 54C** present helical wheel and net plots of the apoC-II mimetic peptide, apoC-II-a. FIG. 54A is a helical wheel plot of the 18A helix. The size of balls and color indicate the degree of charge and hydrophobicity (yellow, nonpolar; green, polar/uncharged; pink, acidic; blue, basic). FIG. 54B is a helical wheel plot of the third helix of apoC-II. FIG. 54C is a helical net plot of the apoC-II-a peptide. ApoC-II-a has the sequence of DWLKAFYDKVAEKLKEAFPAMSTYTGTGIFTDQVLSVLKGEE (SEQ ID NO: 56). The central region indicates the position of the hydrophobic face.

[0094] **FIG. 55** shows the study design for testing post-prandial hypertriglyceridemia in mice.

[0095] **FIG. 56** shows the study design for testing post-prandial hypertriglyceridemia in mice in the presence of LPL inhibitor.

DETAILED DESCRIPTION

Definitions

[0096] Unless defined otherwise, all technical and scientific terms used herein have the meaning commonly understood by one of ordinary skill in the art to which the invention pertains.

[0097] The term “amino acid” refers to natural amino acids, unnatural amino acids, and amino acid analogs. Unless otherwise indicated, the term “amino acid” includes both D and L stereoisomers if the respective structure allows such stereoisomeric forms.

[0098] Natural amino acids include alanine (Ala or A), arginine (Arg or R), asparagine (Asn or N), aspartic acid (Asp or D), cysteine (Cys or C), glutamine (Gln or Q), glutamic acid (Glu or E), glycine (Gly or G), histidine (His or H), isoleucine (Ile or I), leucine (Leu or L), Lysine (Lys or K), methionine (Met or M), phenylalanine (Phe or F), proline (Pro or P), serine (Ser or S), threonine (Thr or T), tryptophan (Trp or W), tyrosine (Tyr or Y) and valine (Val or V).

[0099] Unnatural amino acids, or non-natural amino acid include, but are not limited to, azetidinecarboxylic acid, 2-aminoadipic acid, 3-aminoadipic acid, beta-alanine, naphthylalanine (“naph”), aminopropionic acid, 2-aminobutyric acid, 4-aminobutyric acid, 6-aminocaproic acid, 2-aminoheptanoic acid, 2-aminoisobutyric acid, 3-aminoisobutyric acid, 2-aminopimelic acid, tertiary-butylglycine (“tBuG”), 2,4-diaminoisobutyric acid, desmosine, 2,2'-diaminopimelic acid, 2,3-diaminopropionic acid, N-ethylglycine, N-ethylasparagine, homoproline (“hPro” or “homoP”), hydroxylysine, allo-hydroxylysine, 3-hydroxyproline (“3Hyp”), 4-hydroxyproline (“4Hyp”), isodesmosine, allo-isoleucine, N-methylalanine (“MeAla” or “Nime”), N-alkylglycine (“NAG”) including N-methylglycine, N-methylisoleucine, N-alkylpentylglycine (“NAPG”) including N-methylpentylglycine. N-methylvaline, naphthylalanine, norvaline (“Norval”), norleucine (“Norleu”), octylglycine (“OctG”), ornithine (“Orn”), pentylglycine (“pG” or “PGly”), pipecolic acid, thioproline (“ThioP” or “tPro”), homoLysine (“hLys”), and homoArginine (“hArg”).

[00100] The term “amino acid analog” refers to a natural or unnatural amino acid where one or more of the C-terminal carboxy group, the N-terminal amino group and side-chain functional group has been chemically blocked, reversibly or irreversibly, or otherwise modified to another functional group. For example, aspartic acid-(beta-methyl ester) is an amino acid analog of aspartic acid; N-ethylglycine is an amino acid analog of glycine; or alanine carboxamide is an amino acid analog of alanine. Other amino acid analogs include methionine sulfoxide, methionine sulfone, S-(carboxymethyl)-cysteine, S-(carboxymethyl)-cysteine sulfoxide and S-(carboxymethyl)-cysteine sulfone.

[00101] As used herein, the term “peptide” refers a short polymer of amino acids linked together by peptide bonds. In contrast to other amino acid polymers (e.g., proteins, polypeptides, etc.), peptides are of about 75 amino acids or less in length. A peptide can comprise natural amino acids, non-natural amino acids, amino acid analogs, and/or modified amino acids. A peptide can be a subsequence of naturally occurring protein or a non-natural (synthetic) sequence.

[00102] As used herein, the term “mutant peptide” refers to a variant of a peptide having a distinct amino acid sequence from the most common variant occurring in nature, referred to as the “wild-type” sequence. A mutant peptide can comprise one or more amino acid substitution, deletion, or insertion as compared to the wild-type sequence. A mutant peptide can be a subsequence of a mutant protein or polypeptide (e.g., a subsequence of a naturally-occurring protein that is not the most common sequence in nature), or can be a peptide that is not a subsequence of a naturally occurring protein or polypeptide. For example, a “mutant apoC-II peptide” can be a subsequence of a mutant version of apoC-II or can be distinct sequence not found in naturally-occurring apoC-II proteins.

[00103] As used herein, the term “synthetic peptide” refers to a peptide having a distinct amino acid sequence from those found in natural peptides and/or proteins. A synthetic protein is not a subsequence of a naturally occurring protein, either the wild-type (i.e., most abundant) or mutant versions thereof. For example, a “synthetic apoC-II peptide” is not a subsequence of naturally occurring apoC-II. A “synthetic peptide,” as used herein, can be produced or synthesized by any suitable method (e.g., recombinant expression, chemical synthesis, enzymatic synthesis, etc.).

[00104] The terms “peptide mimetic” or “peptidomimetic” refer to a peptide-like molecule that emulates a sequence derived from a protein or peptide. A peptide mimetic or peptidomimetic can contain amino acids and/or non-amino acid components. Examples of peptidomimetics include chemically modified peptides, peptoids (side groups are appended to the nitrogen atom of the peptide backbone, rather than to the α -carbons), β -peptides (amino group bonded to the β carbon rather than the α -carbon), etc. Chemical modification includes one or more modifications at amino acid side groups, α -carbon atoms, terminal amine group, or terminal carboxy group. A chemical modification can be adding chemical moieties, creating new bonds, or removing chemical moieties. Modifications at amino acid side groups include, without limitation, acylation of lysine ϵ -amino groups, N-alkylation of arginine, histidine, or lysine, alkylation of glutamic or aspartic carboxylic acid groups, lactam formation via cyclization of lysine ϵ -amino groups with glutamic or aspartic acid side group carboxyl groups, hydrocarbon “stapling” (e.g., to stabilize alpha-helix conformations), and deamidation of glutamine or asparagine. Modifications of the terminal amine group include, without limitation, the desamino, N-lower alkyl, N-di-lower alkyl, constrained alkyls (e.g. branched, cyclic, fused, adamantyl) and N-acyl modifications. Modifications of the terminal carboxy group include, without limitation, the amide, lower alkyl amide, constrained alkyls

(e.g. branched, cyclic, fused, adamantyl) alkyl, dialkyl amide, and lower alkyl ester modifications. Lower alkyl is C1-C4 alkyl. Furthermore, one or more side groups, or terminal groups, can be protected by protective groups known to the ordinarily skilled peptide chemist. The α -carbon of an amino acid can be mono- or dimethylated.

[00105] As used herein, a “conservative” amino acid substitution refers to the substitution of an amino acid in a peptide or polypeptide with another amino acid having similar chemical properties, such as size or charge. For purposes of the present disclosure, each of the following eight groups contains amino acids that are conservative substitutions for one another:

- 1) Alanine (A) and Glycine (G);
- 2) Aspartic acid (D) and Glutamic acid (E);
- 3) Asparagine (N) and Glutamine (Q);
- 4) Arginine (R) and Lysine (K);
- 5) Isoleucine (I), Leucine (L), Methionine (M), and Valine (V);
- 6) Phenylalanine (F), Tyrosine (Y), and Tryptophan (W);
- 7) Serine (S) and Threonine (T); and
- 8) Cysteine (C) and Methionine (M).

[00106] Naturally occurring residues can be divided into classes based on common side group properties, for example: polar positive (histidine (H), lysine (K), and arginine (R)); polar negative (aspartic acid (D), glutamic acid (E)); polar neutral (serine (S), threonine (T), asparagine (N), glutamine (Q)); non-polar aliphatic (alanine (A), valine (V), leucine (L), isoleucine (I), methionine (M)); non-polar aromatic (phenylalanine (F), tyrosine (Y), tryptophan (W)); proline and glycine; and cysteine. As used herein, a “semi-conservative” amino acid substitution refers to the substitution of an amino acid in a peptide or polypeptide with another amino acid within the same class.

[00107] In some embodiments, unless otherwise specified, a conservative or semi-conservative amino acid substitution can also encompass non-naturally occurring amino acid residues that have similar chemical properties to the natural residue. These non-natural residues are typically incorporated by chemical peptide synthesis rather than by synthesis in biological systems. These include, but are not limited to, peptidomimetics and other reversed or inverted forms of amino acid moieties. Embodiments herein include natural amino acids, non-natural amino acids, and amino acid analogs. For example, nor-leucine can be used to substitute methionine.

[00108] Non-conservative substitutions can involve the exchange of a member of one class for a member from another class.

[00109] As used herein, the term “sequence identity” refers to the degree to which two polymer sequences (e.g., peptide, polypeptide, nucleic acid, etc.) have the same sequential composition of monomer subunits. The term “sequence similarity” refers to the degree with which two polymer sequences (e.g., peptide, polypeptide, nucleic acid, etc.) differ only by conservative and/or semi-conservative amino acid substitutions. The “percent sequence identity” (or “percent sequence similarity”) is calculated by: (1) comparing two optimally aligned sequences over a window of comparison (e.g., the length of the longer sequence, the length of the shorter sequence, a specified window, etc.), (2) determining the number of positions containing identical (or similar) monomers (e.g., same amino acids occurs in both sequences, similar amino acid occurs in both sequences) to yield the number of matched positions, (3) dividing the number of matched positions by the total number of positions in the comparison window (e.g., the length of the longer sequence, the length of the shorter sequence, a specified window), and (4) multiplying the result by 100 to yield the percent sequence identity or percent sequence similarity. For example, if peptides A and B are both 20 amino acids in length and have identical amino acids at all but 1 position, then peptide A and peptide B have 95% sequence identity. If the amino acids at the non-identical position shared the same biophysical characteristics (e.g., both were acidic), then peptide A and peptide B would have 100% sequence similarity. As another example, if peptide C is 20 amino acids in length and peptide D is 15 amino acids in length, and 14 out of 15 amino acids in peptide D are identical to those of a portion of peptide C, then peptides C and D have 70% sequence identity, but peptide D has 93.3% sequence identity to an optimal comparison window of peptide C. For the purpose of calculating “percent sequence identity” (or “percent sequence similarity”) herein, any gaps in aligned sequences are treated as mismatches at that position.

[00110] As used herein, the term “grouped residues” refers to a set of amino acids within a peptide, polypeptide, or protein that are physically positioned together in three dimensional space. The grouped residues may or may not be sequential in the primary sequence of the peptide, polypeptide, or protein. The residues can be grouped in a globular domain, can be present on the same surface, or can be presented on the same end or side of a secondary structure (e.g., alpha helix) or tertiary structure within a peptide, polypeptide, or protein. In some embodiments, in addition to being physically positioned together, the grouped residues

also exhibit some degree of similarity in residue characteristics (e.g., size, polarity charge, etc.).

[00111] As used herein, “hydrophobic moment (μ_H)” refers to a measure of the amphipathicity of a helix. The degree of amphipathicity (i.e., degree of asymmetry of hydrophobicity) in the multi-domain peptides or peptide analogs can be quantified by calculating the hydrophobic moment (μ_H) of each of the amphipathic α -helical domains. Methods for calculating μ_H for a particular peptide sequence are well-known in the art, and are described, for example in Eisenberg et al., *Faraday Symp. Chem. Soc.* 17: 109-120, 1982; Eisenberg et al., *PNAS* 81:140-144, 1984; and Eisenberg et al., *J. Mol. Biol.* 179:125-142, 1984. The actual μ_H obtained for a particular peptide sequence will depend on the total number of amino acid residues composing the peptide. The amphipathicities of peptides of different lengths can be directly compared by way of the mean hydrophobic moment. The mean hydrophobic moment per residue can be obtained by dividing μ_H by the number of residues in the peptide.

[00112] Unless otherwise specified, “apoC-II” (synonymously, “apoC2”) refers to human apolipoprotein C-II. Residues are numbered herein according to their position in the mature 79 amino acid form of human apoC-II protein following cleavage of the signal sequence. The mature apolipoprotein C-II protein is a 79 amino acid protein that includes three helices: helix 1, residues 17-38; helix 2, residues 45-57; and helix 3, residues 65-74/75 (Zdunek et al., *Biochemistry* 42: 1872-1889, 2003). The lipase-activating region of apoC-II has previously been localized to the C-terminal domain of the sequence, from about residue 56, whereas the N-terminal domain (residues 1-50) of the sequence is involved in lipid binding. Nucleic acid and protein sequences for apoC-II orthologues from a variety of species are publicly available. The GenBank NCBI accession number for the human apoC-II precursor is NP_000474. The amino acid sequences of mature human apoC-II, of amino acid residues 40-79 of human apoC-II, and of native helices 1-3 of human apoC-II are shown in Table 1 below.

Table 1		
Name	Sequence	SEQ ID NO
Native ApoC-II (full length)	TQQPQQDEMPSPFTLTQVKESLSSYWESAKTAAQN LYEKTYLPAVDEKLRDLYSKSTAAMSTYTGIFTDQ VLSVLKGEE	SEQ ID NO: 1
Native ApoC-II (a.a.40 – a.a.79)	TYLPAVDEKLRDLYSKSTAAMSTYTGIFTDQVLSVL KGEE	SEQ ID NO: 2
Native ApoC-II (helix 1)	QVKESLSSYWESAKTAAQNLYE	SEQ ID NO: 3
Native ApoC-II (helix 2)	VDEKLRDLYSKST	SEQ ID NO: 4
Native ApoC-II (helix 3)	GIFTDQVLSVL	SEQ ID NO: 5

[00113] Unless otherwise specified, “LPL” refers to lipoprotein lipase. The GenBank NCBI accession number for human LPL protein is NP_000228.

[00114] As used herein, the term “subject” broadly refers to any animal, including but not limited to, human and non-human animals (e.g., dogs, cats, cows, horses, sheep, pigs, poultry, fish, crustaceans, etc.). As used herein, the term “patient” typically refers to a subject that is being treated for a disease or condition.

[00115] As used herein, the term “effective amount” refers to the amount of a composition (e.g., a synthetic peptide) sufficient to effect beneficial or desired results. An effective amount can be administered in one or more administrations, applications or dosages and is not intended to be limited to a particular formulation or administration route.

[00116] As used herein, the terms “administration” and “administering” refer to the act of giving a drug, prodrug, or other agent, or therapeutic treatment (e.g., synthetic peptide) to a subject or *in vivo*, *in vitro*, or *ex vivo* cells, tissues, and organs. Exemplary routes of administration to the human body can be through space under the arachnoid membrane of the brain or spinal cord (intrathecal), the eyes (ophthalmic), mouth (oral), skin (topical or transdermal), nose (nasal), lungs (inhalant), oral mucosa (buccal or lingual), ear, rectal, vaginal, by injection (e.g., intravenously, subcutaneously, intratumorally, intraperitoneally, etc.) and the like.

[00117] As used herein, the terms “co-administration” and “co-administering” refer to the administration of at least two agent(s) (e.g., multiple synthetic peptide or a synthetic peptide and another therapeutic agent) or therapies to a subject. In some embodiments, the co-administration of two or more agents or therapies is concurrent. In other embodiments, a first

agent/therapy is administered prior to a second agent/therapy. Those of skill in the art understand that the formulations and/or routes of administration of the various agents or therapies used can vary. The appropriate dosage for co-administration can be readily determined by one skilled in the art. In some embodiments, when agents or therapies are coadministered, the respective agents or therapies are administered at lower dosages than appropriate for their administration alone. Thus, co-administration is especially desirable in embodiments where the co-administration of the agents or therapies lowers the requisite dosage of a potentially harmful (e.g., toxic) agent(s), and/or when co-administration of two or more agents results in sensitization of a subject to beneficial effects of one of the agents via co-administration of the other agent.

[00118] As used herein, the term “treatment” means an approach to obtaining a beneficial or intended clinical result. The beneficial or intended clinical result can include alleviation of symptoms, a reduction in the severity of the disease, inhibiting an underlying cause of a disease or condition, steadying diseases in a non-advanced state, delaying the progress of a disease, and/or improvement or alleviation of disease conditions.

[00119] As used herein, the term “pharmaceutical composition” refers to the combination of an active agent (e.g., apoC-II mimetic peptide) with a carrier, inert or active, making the composition especially suitable for therapeutic or diagnostic use *in vitro*, *in vivo* or *ex vivo*.

[00120] The terms “pharmaceutically acceptable” or “pharmacologically acceptable,” as used herein, refer to compositions that do not substantially produce adverse reactions, e.g., toxic, allergic, or immunological reactions, when administered to a subject.

[00121] As used herein, the term “pharmaceutically acceptable carrier” refers to any of the standard pharmaceutical carriers including, but not limited to, phosphate buffered saline solution, water, emulsions (e.g., such as an oil/water or water/oil emulsions), glycerol, liquid polyethylene glycols, aprotic solvents such as dimethylsulfoxide, N-methylpyrrolidone and mixtures thereof, and various types of wetting agents, solubilizing agents, anti-oxidants, bulking agents, protein carriers such as albumins, any and all solvents, dispersion media, coatings, sodium lauryl sulfate, isotonic and absorption delaying agents, disintegrants (e.g., potato starch or sodium starch glycolate), and the like. The compositions also can include stabilizers and preservatives. For examples of carriers, stabilizers and adjuvants, *see, e.g., Martin, Remington's Pharmaceutical Sciences, 21th Ed., Mack Publ. Co., Easton, Pa. (2005)*, incorporated herein by reference in its entirety.

1.1. ApoC-II Mimetic Peptide

[00122] In a first aspect, disclosed herein is an isolated apoC-II mimetic peptide.

[00123] In some embodiments, the isolated apoC-II mimetic peptide is a multihelical peptide comprising a plurality of helical domains. In some multihelical embodiments, one or more of the helical domains are amphipathic and can bind to lipids and/or to the surface of lipoproteins. In certain embodiments, isolated the apoC-II mimetic peptide is a bihelical peptide comprising two helical domains. In some bihelical embodiments, at least one helical domain is amphipathic. In some bihelical embodiments, one helical domain is amphipathic, and the other helical domain is Type G helix found in globular proteins.

[00124] In some embodiments, the isolated apoC-II mimetic peptide is a single helical peptide. In some of these embodiments, the helical domain is amphipathic. In some other of these embodiments, the helical domain is Type G helix.

[00125] In various embodiments, the isolated apoC-II mimetic peptide is no more than 75 amino acids. In certain embodiments, the isolated apoC-II mimetic peptide is no more than 70 amino acids. In certain embodiments, the isolated apoC-II mimetic peptide is no more than 60 amino acids. In certain embodiments, the isolated apoC-II mimetic peptide is no more than 50 amino acids. In certain embodiments, the isolated apoC-II mimetic peptide is no more than 40 amino acids. In certain embodiments, the isolated apoC-II mimetic peptide is no more than 30 amino acids. In certain embodiments, the isolated apoC-II mimetic peptide is no more than 20 amino acids.

[00126] In typical embodiments, the isolated apoC-II mimetic peptide comprises a first helical domain, a hinge region, and a second helical domain.

[00127] In some embodiments, the isolated apoC-II mimetic peptide comprises a first helical domain, a hinge region, and a second helical domain in order from N-terminus to C-terminus. In some of these embodiments, the first helical domain is amphipathic. In some of these embodiments, the second helical domain forms a globular helix. In certain of these embodiments, the first helical domain is amphipathic and the second helical domain forms a globular helix.

[00128] In some embodiments, the first domain has an affinity for binding to lipoproteins. In some embodiments, the second domain is capable of activating lipoprotein lipase (LPL). In certain embodiments, the first domain has an affinity for binding to lipoproteins, and the second domain is capable of activating lipoprotein lipase (LPL).

[00129] In certain embodiments, the isolated apoC-II mimetic peptide is capable of activating lipolysis by LPL. In certain embodiments, the isolated apoC-II mimetic peptide is capable of displacing apoC-III from lipoproteins.

[00130] In various embodiments, the isolated apoC-II mimetic peptide is capable of reducing triglyceride (TG) level *in vivo*. In some of these embodiments, the peptide is capable of reducing elevated TG level caused by LPL deficiency. In some of these embodiments, the peptide is capable of reducing elevated TG level caused by elevated apoC-III. In some of these embodiments, the peptide is capable of reducing elevated TG level caused by apoC-II deficiency. In yet some of these embodiments, the peptide is capable of reducing post-prandial elevated TG level.

[00131] In various embodiments, the isolated apoC-II mimetic peptide has an amino acid sequence set forth in any one of SEQ ID NOs: 6-52 (Table 2). In some embodiments, the isolated apoC-II mimetic peptide has 90% sequence similarity to an amino acid sequence set forth in any one of SEQ ID NOs: 6-52. In some embodiments, the isolated apoC-II mimetic peptide has 91%, 92%, 93%, 94%, or 95% sequence similarity to an amino acid sequence set forth in any one of SEQ ID NOs: 6-52. In some embodiments, the isolated apoC-II mimetic peptide has 96%, 97%, 98% or 99% sequence similarity to an amino acid sequence set forth in any one of SEQ ID NOs: 6-52.

1.1.1. The First Helical Domain

[00132] In various embodiments, the first helical domain has an affinity for binding to lipoproteins. In some embodiments, the first helical domain is amphipathic. In certain of these embodiments, the helical domain has a hydrophobic moment score of about 6 to about 15, such as about 7 to about 15, about 8 to about 15, about 9 to about 15, about 10 to about 15, about 11 to about 15, about 12 to about 15, about 13 to about 15, about 14 to about 15, about 8 to about 14, about 9 to about 14, about 10 to about 14, about 11 to about 14, about 12 to about 14, about 13 to about 14, about 8 to about 13, about 9 to about 13, about 10 to about 13, about 11 to about 13, about 12 to about 13, about 8 to about 12, about 9 to about 12, about 10 to about 12, about 11 to about 12, about 8 to about 11, about 9 to about 11, about 10 to about 11, about 8 to about 10, about 9 to about 10, or about 8 to about 9. The hydrophobic moment of peptide can be calculated with online tools, such as rzlab.ucr.edu/scripts/wheel/wheel.cgi.

[00133] In some embodiments, the first helical domain of the isolated apoC-II mimetic peptide is native helix 1 of apoC-II protein. Helix 1 of apoC-II protein consists of amino acid residues 17-38 (SEQ ID NO: 3), numbered according to their position in the mature 79 amino acid apoC-II protein. In some other embodiments, the first helical domain of the isolated apoC-II mimetic peptide is a variant of helix 1 of apoC-II protein.

[00134] In some embodiments, the first helical domain of the isolated apoC-II mimetic peptide is native helix 2 of apoC-II protein. Helix 2 of apoC-II protein consists of amino acid residues 45-57 (SEQ ID NO: 4), numbered according to their position in the mature 79 amino acid apoC-II protein. In some other embodiments, the first helical domain of the isolated apoC-II mimetic peptide is a variant of helix 2 of apoC-II protein. In certain embodiments, the variant of helix 2 of apoC-II protein comprises at least one modification of the native helix 2 of apoC-II protein. In certain embodiments, the variant of helix 2 of apoC-II protein comprises a plurality of modifications of the native helix 2 of apoC-II protein. In various embodiments, each modification is independently selected from elongations, truncations, mutations, and/or chemical modifications.

[00135] In some other embodiments, the first helical domain of the isolated apoC-II mimetic peptide is a variant of an amphipathic helix found on apolipoproteins other than apoC-II, or from other proteins that can bind lipids.

1.1.1.1. Elongation of Helix 2 of ApoC-II

[00136] In certain embodiments, the variant of helix 2 of apoC-II comprises an elongation of helix 2 of apoC-II. In various embodiments, the elongation increases the amphipathicity of the variant of helix 2 of apoC-II compared to the native helix 2 of apoC-II. In some embodiments, the elongation increases the affinity of the variant of helix 2 of apoC-II for binding to lipoproteins.

[00137] In some embodiments, the helix 2 of apoC-II is elongated at the N-terminus. In various embodiments, the helix 2 of apoC-II is elongated by 1-20 amino acids at the N-terminus. In some of these embodiments, the helix 2 of apoC-II is elongated by 1-10 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is elongated by 1 amino acid at the N-terminus. In certain embodiments, the helix 2 of apoC-II is elongated by 2 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is elongated by 3 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is

elongated by 4 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is elongated by 5 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is elongated by 6 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is elongated by 7 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is elongated by 8 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is elongated by 9 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is elongated by 10 amino acids at the N-terminus.

[00138] In specific embodiments, when the helix 2 of apoC-II is elongated by 1 amino acid at the N-terminus, the elongating amino acid is aspartic acid.

[00139] In specific embodiments, when the helix 2 of apoC-II is elongated by 2 amino acids at the N-terminus, the elongating amino acids are lysine and alanine, from N-terminus to C-terminus.

[00140] In typical embodiments, the helix 2 of apoC-II is elongated by 5 amino acids at the N-terminus. In some embodiments, the helix 2 of apoC-II is elongated by the native upstream amino acid sequence of human apoC-II. In some embodiments, the helix 2 of apoC-II is elongated by a mutant of native upstream amino acid sequence of human apoC-II. In some embodiments, the amino acids are at positions 40 to 44, numbered according to their position in the mature 79 amino acid human apoC-II protein. In some embodiments, the variant of helix 2 of apoC-II comprises an aspartic acid at position 40. In some embodiments, the variant of helix 2 of apoC-II comprises a tyrosine or a tryptophan at position 41. In some embodiments, the variant of helix 2 of apoC-II comprises a leucine at position 42. In some embodiments, the variant of helix 2 of apoC-II comprises a lysine or an arginine at position 43. In some embodiments, the variant of helix 2 of apoC-II comprises an alanine or a glutamic acid at position 44.

[00141] In specific embodiments, when the helix 2 of apoC-II is elongated by 5 amino acids at the N-terminus, the variant of helix 2 of apoC-II comprises an aspartic acid at position 40, a tyrosine at position 41, a leucine at position 42, a lysine at position 43, and a glutamic acid at position 44.

[00142] In specific embodiments, when the helix 2 of apoC-II is elongated by 5 amino acids at the N-terminus, the variant of helix 2 of apoC-II comprises an aspartic acid at position 40, a tyrosine at position 41, a leucine at position 42, a lysine at position 43, and an alanine at position 44.

1.1.1.2. Truncation of Helix 2 of ApoC-II

[00143] In certain embodiments, the variant of helix 2 of apoC-II comprises a truncation of helix 2 of apoC-II. In various embodiments, the truncation increases the amphipathicity of the variant of helix 2 of apoC-II compared to the native helix 2 of apoC-II. In some embodiments, the truncation increases the affinity of the variant of helix 2 of apoC-II for binding to lipoproteins.

[00144] In some embodiments, the helix 2 of apoC-II is truncated at the N-terminus. In various embodiments, the helix 2 of apoC-II is truncated by 1-12 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is truncated by 1 amino acid at the N-terminus. In certain embodiments, the helix 2 of apoC-II is truncated by 2 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is truncated by 3 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is truncated by 4 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is truncated by 5 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is truncated by 6 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is truncated by 7 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is truncated by 8 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is truncated by 9 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is truncated by 10 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is truncated by 11 amino acids at the N-terminus. In certain embodiments, the helix 2 of apoC-II is truncated by 12 amino acids at the N-terminus.

[00145] In specific embodiments, when the helix 2 of apoC-II is truncated by 2 amino acids at the N-terminus, the amino acid residues 45-46 of native helix 2 of apoC-II are deleted.

[00146] In specific embodiments, when the helix 2 of apoC-II is truncated by 4 amino acids at the N-terminus, the amino acid residues 45-48 of native helix 2 of apoC-II are deleted.

[00147] In specific embodiments, when the helix 2 of apoC-II is truncated by 5 amino acids at the N-terminus, the amino acid residues 45-49 of native helix 2 of apoC-II are deleted.

[00148] In specific embodiments, when the helix 2 of apoC-II is truncated by 6 amino acids at the N-terminus, the amino acid residues 45-50 of native helix 2 of apoC-II are deleted.

[00149] In specific embodiments, when the helix 2 of apoC-II is truncated by 9 amino acids at the N-terminus, the amino acid residues 45-53 of native helix 2 of apoC-II are deleted.

1.1.1.3. Mutation of Helix 2 of ApoC-II

[00150] In certain embodiments, the variant of helix 2 of apoC-II comprises at least one mutation of helix 2 of apoC-II. In various embodiments, the mutation increases the amphipathicity of the variant of helix 2 of apoC-II compared to the native helix 2 of apoC-II. In some embodiments, the mutation increases the affinity of the variant of helix 2 of apoC-II for binding to lipoproteins.

[00151] In certain embodiments, the variant of helix 2 of apoC-II comprises one mutation of helix 2 of apoC-II. In certain embodiments, the variant of helix 2 of apoC-II comprises two mutations of helix 2 of apoC-II. In certain embodiments, the variant of helix 2 of apoC-II comprises three mutations of helix 2 of apoC-II. In certain embodiments, the variant of helix 2 of apoC-II comprises four mutations of helix 2 of apoC-II. In certain embodiments, the variant of helix 2 of apoC-II comprises five mutations of helix 2 of apoC-II. In certain embodiments, the variant of helix 2 of apoC-II comprises more than five mutations of helix 2 of apoC-II.

[00152] In some embodiments, the mutation is an amino acid substitution. In some embodiments, the mutation is an amino acid insertion. In some embodiments, the mutation is an amino acid deletion.

[00153] In some embodiments, an original amino acid of helix 2 of apoC-II is substituted by a natural amino acid. In some embodiments, an original amino acid of helix 2 of apoC-II is substituted by an unnatural amino acid. In some embodiments, an original amino acid of helix 2 of apoC-II is substituted by an amino acid analog. In some embodiments, an original amino acid of helix 2 of apoC-II is substituted by a chemically modified amino acid.

[00154] In various embodiments, the amino acid substitution is a conservative or semi-conservative substitution. In some embodiments, the amino acid substitution has minimal impact on the activity and/or structure of the resultant peptide. In certain embodiments, the

amino acid substitution maintains the structure of the peptide backbone in the area of the substitution, for example, as a helical conformation. In certain embodiments, the amino acid substitution maintains the charge or hydrophobicity of the molecule at the target site. In certain embodiments, the amino acid substitution maintains the bulk of the amino acid side group.

[00155] In various embodiments, the amino acid substitution is a non-conservative substitution. In some embodiments, the amino acid substitution produces significant changes in the peptide property. In certain embodiments, a hydrophilic residue is substituted by a hydrophobic residue. In certain other embodiments, a hydrophobic residue is substituted by a hydrophilic residue. In certain embodiments, a cysteine or proline is substituted by another residue. In certain other embodiments, a non-cysteine or non-proline is substituted by a cysteine or proline. In certain embodiments, a residue having an electropositive side group is substituted by an electronegative residue. In certain other embodiments, a residue having an electronegative side group is substituted by an electropositive residue. In certain embodiments, a residue having a bulky side group is substituted by a residue not having a side group. In certain other embodiments, a residue not having a side group is substituted by a residue having a bulky side group.

[00156] In certain embodiments, the amino acid substitution is at position 45. In some embodiments, the valine at position 45 is substituted by a phenylalanine.

[00157] In certain embodiments, the amino acid substitution is at position 46. In some embodiments, the aspartic acid at position 46 is substituted by a phenylalanine. In some embodiments, the aspartic acid at position 46 is substituted by a lysine. In some embodiments, the aspartic acid at position 46 is substituted by an aminoisobutyric acid.

[00158] In certain embodiments, the amino acid substitution is at position 47.

[00159] In certain embodiments, the amino acid substitution is at position 48. In some embodiments, the lysine at position 48 is substituted by an arginine.

[00160] In certain embodiments, the amino acid substitution is at position 49. In some embodiments, the leucine at position 49 is substituted by a lysine. In some of these embodiments, the lysine is modified by fatty acid.

[00161] In certain embodiments, the amino acid substitution is at position 50. In some embodiments, the arginine at position 50 is substituted by a lysine.

[00162] In certain embodiments, the amino acid substitution is at position 51.

- [00163]** In certain embodiments, the amino acid substitution is at position 52.
- [00164]** In certain embodiments, the amino acid substitution is at position 53. In some embodiments, the tyrosine at position 53 is substituted by a leucine.
- [00165]** In certain embodiments, the amino acid substitution is at position 54. In some embodiments, the serine at position 54 is substituted by a glutamic acid. In some embodiments, the serine at position 54 is substituted by a lysine. In some embodiments, the serine at position 54 is substituted by an aspartic acid.
- [00166]** In certain embodiments, the amino acid substitution is at position 55. In some embodiments, the lysine at position 55 is substituted by an arginine.
- [00167]** In certain embodiments, the amino acid substitution is at position 56. In some embodiments, the serine at position 56 is substituted by a phenylalanine. In some embodiments, the serine at position 56 is substituted by a lysine. In some embodiments, the serine at position 56 is substituted by an alanine. In some embodiments, the serine at position 56 is substituted by an aminoisobutyric acid.
- [00168]** In certain embodiments, the amino acid substitution is at position 57. In some embodiments, the threonine at position 57 is substituted by a phenylalanine.
- [00169]** In specific embodiments, the variant of helix 2 of apoC-II comprises two amino acid substitutions, and the amino acid substitutions are at position 46 and position 56. In some of these embodiments, the aspartic acid at position 46 is substituted by a phenylalanine and the serine at position 56 is substituted by a phenylalanine.
- [00170]** In specific embodiments, the variant of helix 2 of apoC-II comprises three amino acid substitutions, and the amino acid substitutions are at position 46, position 54, and position 56. In some of these embodiments, the aspartic acid at position 46 is substituted by a phenylalanine, the serine at position 54 is substituted by a glutamic acid, and the serine at position 56 is substituted by a phenylalanine.

1.1.1.4. Chemical Modification of the Variant of Helix 2 of ApoC-II

- [00171]** In certain embodiments, the variant of helix 2 of apoC-II comprises at least one chemical modification. In various embodiments, the chemical modification increases the amphipathicity of the variant of helix 2 of apoC-II compared to the native helix 2 of apoC-II. In some embodiments, the chemical modification increases the affinity of the variant of helix 2 of apoC-II for binding to lipoproteins.

[00172] In some embodiments, the chemical modification is at the N-terminus of the first domain. In various embodiments, the amino-end of the N-terminus can be modified by conjugation with various functional groups. Neutralization of the terminal charge of synthetic peptide mimics of apolipoproteins has been shown to increase their lipid affinity (Yancey et al., *Biochem.* 34:7955-7965, 1995; Venkatachalapathi et al., *Protein: Structure, Function and Genetics* 15:349-359, 1993). For example, acetylation of the amino terminal end of amphipathic peptides increases the lipid affinity of the peptide (Mishra et al., *J. Biol. Chem.* 269:7185-7191, 1994). Other possible end modifications are described, for example, in Brouillette et al., *Biochem. Biophys. Acta* 1256: 103-129, 1995; Mishra et al., *J. Biol. Chem.* 269:7185-7191, 1994; and Mishra et al., *J. Biol. Chem.* 270:1602-1611, 1995.

[00173] In some embodiments, the chemical modification is at an amino acid side group of the first domain. Modifications at amino acid side groups include, without limitation, acylation of lysine ϵ -amino groups, N-alkylation of arginine, histidine, or lysine, alkylation of glutamic or aspartic carboxylic acid groups, lactam formation via cyclization of lysine ϵ -amino groups with glutamic or aspartic acid side group carboxyl groups, hydrocarbon “stapling” (e.g., to stabilize alpha-helix conformations), and deamidation of glutamine or asparagine. In some embodiments, the first domain is modified by replacement of one or more side groups with other side groups such as alkyl, lower alkyl, cyclic 4-, 5-, 6-, to 7-membered alkyl, amide, amide lower alkyl, amide di(lower alkyl), lower alkoxy, hydroxy, carboxy and the lower ester derivatives thereof, and with 4-, 5-, 6-, to 7-membered heterocyclics. For example, proline analogs can be made in which the ring size of the proline residue is changed from a 5-membered ring to a 4-, 6-, or 7-membered ring. Cyclic groups can be saturated or unsaturated, and if unsaturated, can be aromatic or non-aromatic. Heterocyclic groups can contain one or more nitrogen, oxygen, and/or sulphur heteroatoms. Examples of such groups include furazanyl, furyl, imidazolidinyl, imidazolyl, imidazolinyl, isothiazolyl, isoxazolyl, morpholinyl (e.g., morpholino), oxazolyl, piperazinyl (e.g., 1-piperazinyl), piperidyl (e.g., 1-piperidyl, piperidino), pyranyl, pyrazinyl, pyrazolidinyl, pyrazolinyl, pyrazolyl, pyridazinyl, pyridyl, pyrimidinyl, pyrrolidinyl (e.g., 1-pyrrolidinyl), pyrrolinyl, pyrrolyl, thiadiazolyl, thiazolyl, thienyl, thiomorpholinyl (e.g., thiomorpholino), and triazolyl groups. These heterocyclic groups can be substituted or unsubstituted. Where a group is substituted, the substituent can be alkyl, alkoxy, halogen, oxygen, or substituted or unsubstituted phenyl. Peptides, as well as peptide analogs and mimetics, can also be covalently bound to one or more of a variety of nonproteinaceous polymers, for example,

polyethylene glycol, polypropylene glycol, or polyoxyalkenes, as described in U.S. Pat. Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192; and 4,179,337.

[00174] In certain embodiments, the chemical modification is a non-covalent modification. In certain other embodiments, the chemical modification is covalently linked to the modified amino acid. In various embodiments, the chemical modification can be phosphorylation, glycosylation, or lipidation.

[00175] In some embodiments, the chemical modification is a covalent linkage of a fatty acid. In certain embodiments, the fatty acid is saturated. In certain other embodiments, the fatty acid is unsaturated.

[00176] In various embodiments, the fatty acid comprises 2 to 30 carbons, such as 4 to 26 carbons, 6 to 24 carbons, 10 to 20 carbons, 12 to 18 carbons, and 14 to 16 carbons. In certain embodiments, the fatty acid comprises 6 carbons. In certain embodiments, the fatty acid comprises 7 carbons. In certain embodiments, the fatty acid comprises 8 carbons. In certain embodiments, the fatty acid comprises 9 carbons. In certain embodiments, the fatty acid comprises 10 carbons. In certain embodiments, the fatty acid comprises 11 carbons. In certain embodiments, the fatty acid comprises 12 carbons. In certain embodiments, the fatty acid comprises 13 carbons. In certain embodiments, the fatty acid comprises 14 carbons. In certain embodiments, the fatty acid comprises 15 carbons. In certain embodiments, the fatty acid comprises 16 carbons. In certain embodiments, the fatty acid comprises 17 carbons. In certain embodiments, the fatty acid comprises 18 carbons. In certain embodiments, the fatty acid comprises 19 carbons. In certain embodiments, the fatty acid comprises 20 carbons. In certain embodiments, the fatty acid comprises 21 carbons. In certain embodiments, the fatty acid comprises 22 carbons. In certain embodiments, the fatty acid comprises 23 carbons. In certain embodiments, the fatty acid comprises 24 carbons.

[00177] In some embodiments, the unsaturated fatty acid has 1, 2, 3, 4, 5, or 6 double bonds. In certain embodiments, the unsaturated fatty acid is arachidonic acid. In certain embodiments, the unsaturated fatty acid is linoleic acid. In certain embodiments, the unsaturated fatty acid is oleic acid. In certain embodiments, the unsaturated fatty acid is palmitoleic acid. In certain embodiments, the unsaturated fatty acid is linolenic acid. In certain embodiments, the unsaturated fatty acid is eicosapentaenoic acid. In certain embodiments, the unsaturated fatty acid is docosahexaenoic acid.

[00178] In specific embodiments, the fatty acid is palmitic acid. In specific embodiments, the fatty acid is stearic acid.

[00179] In certain embodiments, the isolated apoC-II mimetic peptide comprises a stearic acid covalently linked to the N-terminal amino acid. In some of these embodiments, the N-terminal amino acid is aspartic acid.

[00180] In certain embodiments, the isolated apoC-II mimetic peptide comprises a stearic acid covalently linked to the side group of a lysine residue. In specific embodiments, when the isolated apoC-II mimetic peptide comprises a lysine at position 46, the stearic acid is covalently linked to the lysine residue at position 46. In specific embodiments, when the isolated apoC-II mimetic peptide comprises a lysine at position 49, the stearic acid is covalently linked to the lysine residue at position 49. In specific embodiments, when the isolated apoC-II mimetic peptide comprises a lysine at position 56, the stearic acid is covalently linked to the lysine residue at position 56.

1.1.2. The Hinge Region

[00181] In some embodiments, the isolated apoC-II mimetic peptide comprises a hinge region connecting the first helical domain and the second helical domain. In certain embodiments, the hinge region functionally separates the first helical domain and the second helical domain. In various embodiments, the hinge region helps the second domain, which has the LPL activation activity, to attach itself to the micelle surface of lipoprotein. In certain embodiments, the hinge region allows the first helical domain and the second helical domain to retain a nearly straight conformation, wherein the second domain bends away from the first domain over an angle of no more than about 20°, such as no more than about 15°, no more than about 10°, or no more than about 5°.

1.1.2.1. Amino Acid Composition of the Hinge Region

[00182] In various embodiments, the hinge region comprises 3-15 amino acids, such as 4-12 amino acids, 5-10 amino acids, 6-9 amino acids, or 7-8 amino acids. In certain embodiments, the hinge region comprises 3 amino acids. In certain embodiments, the hinge region comprises 4 amino acids. In certain embodiments, the hinge region comprises 5 amino acids. In certain embodiments, the hinge region comprises 6 amino acids. In certain embodiments, the hinge region comprises 7 amino acids. In certain embodiments, the hinge region comprises 8 amino acids. In certain embodiments, the hinge region comprises 9 amino acids. In certain embodiments, the hinge region comprises 10 amino acids. In certain embodiments, the hinge region comprises 11 amino acids. In certain embodiments, the hinge

region comprises 12 amino acids. In certain embodiments, the hinge region comprises 13 amino acids. In certain embodiments, the hinge region comprises 14 amino acids. In certain embodiments, the hinge region comprises 15 amino acids.

[00183] In some embodiments, the hinge region comprises a natural amino acid. In some embodiments, the hinge region comprises an unnatural amino acid. In some embodiments, the hinge region comprises an amino acid analog.

[00184] In various embodiments, the hinge region comprises a proline. In some embodiments, the hinge region comprises a hydroxyl proline. Other suitable amino acids (such as glycine, serine, threonine, and alanine) that functionally separate the two helical domains can also be used.

[00185] In some embodiments, the hinge region comprises a proline. In some of these embodiments, the hinge region comprises a proline at position 58. In certain embodiments, the hinge region comprises an alanine. In some of these embodiments, the hinge region comprises an alanine at position 58. In some of these embodiments, the hinge region comprises an alanine at position 59. In some embodiments, the hinge region comprises a norleucine. In some of these embodiments, the hinge region comprises a norleucine at position 60. In some embodiments, the hinge region comprises a lysine. In some of these embodiments, the hinge region comprises a lysine at position 60. In some embodiments, the hinge region comprises a methionine. In some of these embodiments, the hinge region comprises a methionine at position 60. In some embodiments, the hinge region comprises a serine. In some of these embodiments, the hinge region comprises a serine at position 61. In some embodiments, the hinge region comprises a threonine. In some of these embodiments, the hinge region comprises a threonine at position 62. In some of these embodiments, the hinge region comprises a threonine at position 64. In some embodiments, the hinge region comprises a tyrosine. In some of these embodiments, the hinge region comprises a tyrosine at position 63.

1.1.2.2. Chemical Modification of the Hinge Region

[00186] In certain embodiments, the hinge region comprises at least one chemical modification.

[00187] In some embodiments, the chemical modification is at an amino acid side group of the hinge region. Modifications at amino acid side groups include, without limitation,

acylation of lysine ϵ -amino groups, N-alkylation of arginine, histidine, or lysine, alkylation of glutamic or aspartic carboxylic acid groups, lactam formation via cyclization of lysine ϵ -amino groups with glutamic or aspartic acid side group carboxyl groups, hydrocarbon “stapling” (e.g., to stabilize alpha-helix conformations), and deamidation of glutamine or asparagine. In some embodiments, the hinge region is modified by replacement of one or more side groups with other side groups such as alkyl, lower alkyl, cyclic 4-, 5-, 6-, to 7-membered alkyl, amide, amide lower alkyl, amide di(lower alkyl), lower alkoxy, hydroxy, carboxy and the lower ester derivatives thereof, and with 4-, 5-, 6-, to 7-membered heterocyclics. For example, proline analogs can be made in which the ring size of the proline residue is changed from a 5-membered ring to a 4-, 6-, or 7-membered ring. Cyclic groups can be saturated or unsaturated, and if unsaturated, can be aromatic or non-aromatic. Heterocyclic groups can contain one or more nitrogen, oxygen, and/or sulphur heteroatoms. Examples of such groups include furazanyl, furyl, imidazolidinyl, imidazolyl, imidazolinyl, isothiazolyl, isoxazolyl, morpholinyl (e.g., morpholino), oxazolyl, piperazinyl (e.g., 1-piperazinyl), piperidyl (e.g., 1-piperidyl, piperidino), pyranyl, pyrazinyl, pyrazolidinyl, pyrazolinyl, pyrazolyl, pyridazinyl, pyridyl, pyrimidinyl, pyrrolidinyl (e.g., 1-pyrrolidinyl), pyrrolinyl, pyrrolyl, thiadiazolyl, thiazolyl, thienyl, thiomorpholinyl (e.g., thiomorpholino), and triazolyl groups. These heterocyclic groups can be substituted or unsubstituted. Where a group is substituted, the substituent can be alkyl, alkoxy, halogen, oxygen, or substituted or unsubstituted phenyl. Peptides, as well as peptide analogs and mimetics, can also be covalently bound to one or more of a variety of nonproteinaceous polymers, for example, polyethylene glycol, polypropylene glycol, or polyoxyalkenes, as described in U.S. Pat. Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192; and 4,179,337.

[00188] In certain embodiments, the chemical modification is a non-covalent modification. In certain other embodiments, the chemical modification is covalently linked. In various embodiments, the chemical modification can be phosphorylation, glycosylation, or lipidation.

[00189] In some embodiments, the chemical modification is a covalent linkage of a fatty acid. In certain embodiments, the fatty acid is saturated. In certain other embodiments, the fatty acid is unsaturated.

[00190] In various embodiments, the fatty acid comprises 2 to 30 carbons, such as 4 to 26 carbons, 6 to 24 carbons, 10 to 20 carbons, 12 to 18 carbons, and 14 to 16 carbons. In certain embodiments, the fatty acid comprises 6 carbons. In certain embodiments, the fatty acid comprises 7 carbons. In certain embodiments, the fatty acid comprises 8 carbons. In certain

embodiments, the fatty acid comprises 9 carbons. In certain embodiments, the fatty acid comprises 10 carbons. In certain embodiments, the fatty acid comprises 11 carbons. In certain embodiments, the fatty acid comprises 12 carbons. In certain embodiments, the fatty acid comprises 13 carbons. In certain embodiments, the fatty acid comprises 14 carbons. In certain embodiments, the fatty acid comprises 15 carbons. In certain embodiments, the fatty acid comprises 16 carbons. In certain embodiments, the fatty acid comprises 17 carbons. In certain embodiments, the fatty acid comprises 18 carbons. In certain embodiments, the fatty acid comprises 19 carbons. In certain embodiments, the fatty acid comprises 20 carbons. In certain embodiments, the fatty acid comprises 21 carbons. In certain embodiments, the fatty acid comprises 22 carbons. In certain embodiments, the fatty acid comprises 23 carbons. In certain embodiments, the fatty acid comprises 24 carbons.

[00191] In some embodiments, the unsaturated fatty acid has 1, 2, 3, 4, 5, or 6 double bonds. In certain embodiments, the unsaturated fatty acid is arachidonic acid. In certain embodiments, the unsaturated fatty acid is linoleic acid. In certain embodiments, the unsaturated fatty acid is oleic acid. In certain embodiments, the unsaturated fatty acid is palmitoleic acid. In certain embodiments, the unsaturated fatty acid is linolenic acid. In certain embodiments, the unsaturated fatty acid is eicosapentaenoic acid. In certain embodiments, the unsaturated fatty acid is docosahexaenoic acid.

[00192] In specific embodiments, the fatty acid is palmitic acid. In specific embodiments, the fatty acid is stearic acid.

[00193] In certain embodiments, the isolated apoC-II mimetic peptide comprises a palmitic acid covalently linked to the side group of the lysine residue. In specific embodiments, when the isolated apoC-II mimetic peptide comprises a lysine at position 60, the palmitic acid is covalently linked to the lysine residue at position 60.

1.1.3. The Second Helical Domain

[00194] In some embodiments, the second helical domain of the isolated apoC-II mimetic peptide is native helix 3 of apoC-II protein. Helix 3 of apoC-II protein consists of amino acid residues 65-74/75 (SEQ ID NO: 5), numbered according to their position in the mature 79 amino acid apoC-II protein. In some other embodiments, the second helical domain of the isolated apoC-II mimetic peptide is a variant of helix 3 of apoC-II protein.

[00195] In certain embodiments, the variant of helix 3 of apoC-II protein comprises modifications of the native helix 3 of apoC-II protein. In various embodiments, the modifications include elongations, truncations, mutations, and chemical modifications.

1.1.3.1. Elongation of Helix 3 of ApoC-II

[00196] In certain embodiments, the variant of helix 3 of apoC-II comprises elongation of helix 3 of apoC-II. In some embodiments, the elongation increases the ability of variant of helix 3 of apoC-II for activating lipolysis by LPL.

[00197] In some embodiments, the helix 3 of apoC-II is elongated at the C-terminus. In various embodiments, the helix 3 of apoC-II is elongated by 1-20 amino acids at the C-terminus. In some of these embodiments, the helix 3 of apoC-II is elongated by 1-10 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 1 amino acid at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 2 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 3 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 4 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 5 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 6 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 7 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 8 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 9 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 10 amino acids at the C-terminus.

[00198] In specific embodiments, when the helix 3 of apoC-II is elongated by 4 amino acids at the C-terminus, the amino acids are lysine, glycine, glutamic acid, and glutamic acid from N-terminus to C-terminus.

[00199] In specific embodiments, when the helix 3 of apoC-II is elongated by 4 amino acids at the C-terminus, the amino acids are arginine, glycine, glutamic acid, and glutamic acid from N-terminus to C-terminus.

1.1.3.2. Mutation of Helix 3 of ApoC-II

[00200] In certain embodiments, the variant of helix 3 of apoC-II comprises at least one mutation of helix 3 of apoC-II. In some embodiments, the mutation increases the ability of variant of helix 3 of apoC-II for activating lipolysis by LPL.

[00201] In certain embodiments, the variant of helix 3 of apoC-II comprises one mutation of helix 3 of apoC-II. In certain embodiments, the variant of helix 3 of apoC-II comprises two mutations of helix 3 of apoC-II. In certain embodiments, the variant of helix 3 of apoC-II comprises three mutations of helix 3 of apoC-II. In certain embodiments, the variant of helix 3 of apoC-II comprises four mutations of helix 3 of apoC-II. In certain embodiments, the variant of helix 3 of apoC-II comprises five mutations of helix 3 of apoC-II. In certain embodiments, the variant of helix 3 of apoC-II comprises more than five mutations of helix 3 of apoC-II.

[00202] In some embodiments, the mutation is an amino acid substitution. In some embodiments, the mutation is an amino acid insertion. In some embodiments, the mutation is an amino acid deletion.

[00203] In some embodiments, an original amino acid of helix 3 of apoC-II is substituted by a natural amino acid. In some embodiments, an original amino acid of helix 3 of apoC-II is substituted by an unnatural amino acid. In some embodiments, an original amino acid of helix 3 of apoC-II is substituted by an amino acid analog.

[00204] In various embodiments, the amino acid substitution is a conservative or semi-conservative substitution. In some embodiments, the amino acid substitution has minimal impact on the activity and/or structure of the resultant peptide. In certain embodiments, the amino acid substitution maintains the structure of the peptide backbone in the area of the substitution, for example, as a helical conformation. In certain embodiments, the amino acid substitution maintains the charge or hydrophobicity of the molecule at the target site. In certain embodiments, the amino acid substitution maintains the bulk of the amino acid side group.

[00205] In various embodiments, the amino acid substitution is a non-conservative substitution. In some embodiments, the amino acid substitution produces significant changes in the peptide property. In certain embodiments, a hydrophilic residue is substituted by a hydrophobic residue. In certain other embodiments, a hydrophobic residue is substituted by a hydrophilic residue. In certain embodiments, a cysteine or proline is substituted by another

residue. In certain other embodiments, a non-cysteine or non-proline is substituted by a cysteine or proline. In certain embodiments, a residue having an electropositive side group is substituted by an electronegative residue. In certain other embodiments, a residue having an electronegative side group is substituted by an electropositive residue. In certain embodiments, a residue having a bulky side group is substituted by a residue not having a side group. In certain other embodiments, a residue not having a side group is substituted by a residue having a bulky side group.

[00206] In certain embodiments, the amino acid substitution is at position 70. In some of these embodiments, the glutamine at position 70 is substituted by an arginine. In some other of these embodiments, the glutamine at position 70 is substituted by a lysine.

1.1.3.3. Chemical Modification of the Variant of Helix 3 of ApoC-II

[00207] In certain embodiments, the variant of helix 3 of apoC-II comprises at least one chemical modification. In some embodiments, the chemical modification increases the ability of the variant of helix 3 to activate lipolysis by LPL.

[00208] In some embodiments, the chemical modification is at the C-terminus of the second domain. In various embodiments, the carboxyl-end of the C-terminus can be modified by conjugation with various functional groups. Neutralization of the terminal charge of synthetic peptide mimics of apolipoproteins has been shown to increase their lipid affinity (Yancey et al., *Biochem.* 34:7955-7965, 1995; Venkatachalapathi et al., *Protein: Structure, Function and Genetics* 15:349-359, 1993). For example, acetylation of the amino terminal end of amphipathic peptides increases the lipid affinity of the peptide (Mishra et al., *J. Biol. Chem.* 269:7185-7191, 1994). Other possible end modifications are described, for example, in Brouillette et al., *Biochem. Biophys. Acta* 1256: 103-129, 1995; Mishra et al., *J. Biol. Chem.* 269:7185-7191, 1994; and Mishra et al., *J. Biol. Chem.* 270:1602-1611, 1995.

[00209] In some embodiments, the chemical modification is at an amino acid side group of an amino acid in the second domain. Modifications at amino acid side groups include, without limitation, acylation of lysine ϵ -amino groups, N-alkylation of arginine, histidine, or lysine, alkylation of glutamic or aspartic carboxylic acid groups, lactam formation via cyclization of lysine ϵ -amino groups with glutamic or aspartic acid side group carboxyl groups, hydrocarbon “stapling” (e.g., to stabilize alpha-helix conformations), and deamidation of glutamine or asparagine. In some embodiments, the second domain is modified by

replacement of one or more side groups with other side groups such as alkyl, lower alkyl, cyclic 4-, 5-, 6-, to 7-membered alkyl, amide, amide lower alkyl, amide di(lower alkyl), lower alkoxy, hydroxy, carboxy and the lower ester derivatives thereof, and with 4-, 5-, 6-, to 7-membered heterocyclics. For example, proline analogs can be made in which the ring size of the proline residue is changed from a 5-membered ring to a 4-, 6-, or 7-membered ring. Cyclic groups can be saturated or unsaturated, and if unsaturated, can be aromatic or non-aromatic. Heterocyclic groups can contain one or more nitrogen, oxygen, and/or sulphur heteroatoms. Examples of such groups include furazanyl, furyl, imidazolidinyl, imidazolyl, imidazolinyl, isothiazolyl, isoxazolyl, morpholinyl (e.g., morpholino), oxazolyl, piperazinyl (e.g., 1-piperazinyl), piperidyl (e.g., 1-piperidyl, piperidino), pyranyl, pyrazinyl, pyrazolidinyl, pyrazolinyl, pyrazolyl, pyridazinyl, pyridyl, pyrimidinyl, pyrrolidinyl (e.g., 1-pyrrolidinyl), pyrrolinyl, pyrrolyl, thiadiazolyl, thiazolyl, thienyl, thiomorpholinyl (e.g., thiomorpholino), and triazolyl groups. These heterocyclic groups can be substituted or unsubstituted. Where a group is substituted, the substituent can be alkyl, alkoxy, halogen, oxygen, or substituted or unsubstituted phenyl. Peptides, as well as peptide analogs and mimetics, can also be covalently bound to one or more of a variety of nonproteinaceous polymers, for example, polyethylene glycol, polypropylene glycol, or polyoxyalkenes, as described in U.S. Pat. Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192; and 4,179,337.

[00210] In certain embodiments, the chemical modification is a non-covalent modification. In certain other embodiments, the chemical modification is covalently linked. In various embodiments, the chemical modification can be phosphorylation, glycosylation, or lipidation.

[00211] In some embodiments, the chemical modification is a covalent linkage of a fatty acid. In certain embodiments, the fatty acid is saturated. In certain other embodiments, the fatty acid is unsaturated.

[00212] In various embodiments, the fatty acid comprises 2 to 30 carbons, such as 4 to 26 carbons, 6 to 24 carbons, 10 to 20 carbons, 12 to 18 carbons, and 14 to 16 carbons. In certain embodiments, the fatty acid comprises 6 carbons. In certain embodiments, the fatty acid comprises 7 carbons. In certain embodiments, the fatty acid comprises 8 carbons. In certain embodiments, the fatty acid comprises 9 carbons. In certain embodiments, the fatty acid comprises 10 carbons. In certain embodiments, the fatty acid comprises 11 carbons. In certain embodiments, the fatty acid comprises 12 carbons. In certain embodiments, the fatty acid comprises 13 carbons. In certain embodiments, the fatty acid comprises 14 carbons. In certain embodiments, the fatty acid comprises 15 carbons. In certain embodiments, the fatty acid

comprises 16 carbons. In certain embodiments, the fatty acid comprises 17 carbons. In certain embodiments, the fatty acid comprises 18 carbons. In certain embodiments, the fatty acid comprises 19 carbons. In certain embodiments, the fatty acid comprises 20 carbons. In certain embodiments, the fatty acid comprises 21 carbons. In certain embodiments, the fatty acid comprises 22 carbons. In certain embodiments, the fatty acid comprises 23 carbons. In certain embodiments, the fatty acid comprises 24 carbons.

[00213] In some embodiments, the unsaturated fatty acid has 1, 2, 3, 4, 5, or 6 double bonds. In certain embodiments, the unsaturated fatty acid is arachidonic acid. In certain embodiments, the unsaturated fatty acid is linoleic acid. In certain embodiments, the unsaturated fatty acid is oleic acid. In certain embodiments, the unsaturated fatty acid is palmitoleic acid. In certain embodiments, the unsaturated fatty acid is linolenic acid. In certain embodiments, the unsaturated fatty acid is eicosapentaenoic acid. In certain embodiments, the unsaturated fatty acid is docosahexaenoic acid.

[00214] In specific embodiments, the fatty acid is palmitic acid. In specific embodiments, the fatty acid is stearic acid.

[00215] In certain embodiments, the C-terminal amino acid is modified by a C-terminal amide. In some of these embodiments, the C-terminal amino acid is glutamic acid.

1.1.4. Single Domain ApoC-II Mimetic Peptide

[00216] In some embodiments, the apoC-II mimetic peptide consists of one helical domain. In some of these embodiments, the helical domain is amphipathic. In some other of these embodiments, the helical domain is a globular helix. In certain embodiments, the apoC-II mimetic peptide is native helix 3 of apoC-II protein. Helix 3 of apoC-II protein consists of amino acid residues 65-74/75 (SEQ ID NO: 5), numbered according to their position in the mature 79 amino acid apoC-II protein. In certain other embodiments, the apoC-II mimetic peptide is a variant of helix 3 of apoC-II.

[00217] In some embodiments, the variant of helix 3 of apoC-II is no more than 40 amino acids, such as no more than 30 amino acids, or no more than 20 amino acids. In some embodiments, the variant of helix 3 of apoC-II comprises modifications of the native helix 3 of apoC-II. In various embodiments, the modifications include elongations, truncations, mutations, and chemical modifications.

1.1.4.1. Elongation of Helix 3 of ApoC-II

[00218] In certain embodiments, the variant of helix 3 of apoC-II comprises elongation of helix 3 of apoC-II. In some embodiments, the elongation increases the affinity of the variant of helix 3 for binding to lipoproteins. In some embodiments, the elongation increases the ability of variant of helix 3 of apoC-II for activating lipolysis by LPL.

[00219] In some embodiments, the helix 3 of apoC-II is elongated at the N-terminus. In various embodiments, the helix 3 of apoC-II is elongated by 1-20 amino acids at the N-terminus. In some of these embodiments, the helix 3 of apoC-II is elongated by 1-10 amino acids at the N-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 1 amino acid at the N-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 2 amino acids at the N-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 3 amino acids at the N-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 4 amino acids at the N-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 5 amino acids at the N-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 6 amino acids at the N-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 7 amino acids at the N-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 8 amino acids at the N-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 9 amino acids at the N-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 10 amino acids at the N-terminus.

[00220] In some embodiments, the helix 3 of apoC-II is elongated at the C-terminus. In various embodiments, the helix 3 of apoC-II is elongated by 1-20 amino acids at the C-terminus. In some of these embodiments, the helix 3 of apoC-II is elongated by 1-10 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 1 amino acid at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 2 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 3 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 4 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 5 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 6 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 7 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 8 amino acids at the C-terminus. In certain embodiments, the helix

3 of apoC-II is elongated by 9 amino acids at the C-terminus. In certain embodiments, the helix 3 of apoC-II is elongated by 10 amino acids at the C-terminus.

[00221] In certain embodiments, the helix 3 of apoC-II is elongated at both the N-terminus and the C-terminus. In specific embodiments, the helix 3 of apoC-II is elongated at the N-terminus by 6 amino acids and at the C-terminus by 4 amino acids. In some of these embodiments, when the helix 3 of apoC-II is elongated by 4 amino acids at the C-terminus, the amino acids are arginine, glycine, glutamic acid, and glutamic acid from N-terminus to C-terminus. In some of these embodiments, when the helix 3 of apoC-II is elongated by 6 amino acids at the N-terminus, the amino acids are alanine, methionine, serine, threonine, tyrosine, and threonine from N-terminus to C-terminus. In some other of these embodiments, when the helix 3 of apoC-II is elongated by 6 amino acids at the N-terminus, the amino acids are alanine, lysine, serine, threonine, tyrosine, and threonine from N-terminus to C-terminus.

1.1.4.2. Mutation of Helix 3 of ApoC-II

[00222] In certain embodiments, the variant of helix 3 of apoC-II comprises at least one mutation of helix 3 of apoC-II. In some embodiments, the mutation increases the affinity of the variant of helix 3 for binding to lipoproteins. In some embodiments, the mutation increases the ability of variant of helix 3 of apoC-II for activating lipolysis by LPL.

[00223] In certain embodiments, the variant of helix 3 of apoC-II comprises one mutation of helix 3 of apoC-II. In certain embodiments, the variant of helix 3 of apoC-II comprises two mutations of helix 3 of apoC-II. In certain embodiments, the variant of helix 3 of apoC-II comprises three mutations of helix 3 of apoC-II. In certain embodiments, the variant of helix 3 of apoC-II comprises four mutations of helix 3 of apoC-II. In certain embodiments, the variant of helix 3 of apoC-II comprises five mutations of helix 3 of apoC-II. In certain embodiments, the variant of helix 3 of apoC-II comprises more than five mutations of helix 3 of apoC-II.

[00224] In some embodiments, the mutation is an amino acid substitution. In some embodiments, the mutation is an amino acid insertion. In some embodiments, the mutation is an amino acid deletion.

[00225] In some embodiments, an original amino acid of helix 3 of apoC-II is substituted by a natural amino acid. In some embodiments, an original amino acid of helix 3 of apoC-II is

substituted by an unnatural amino acid. In some embodiments, an original amino acid of helix 3 of apoC-II is substituted by an amino acid analog.

[00226] In various embodiments, the amino acid substitution is a conservative or semi-conservative substitution. In some embodiments, the amino acid substitution has minimal impact on the activity and/or structure of the resultant peptide. In certain embodiments, the amino acid substitution maintains the structure of the peptide backbone in the area of the substitution, for example, as a helical conformation. In certain embodiments, the amino acid substitution maintains the charge or hydrophobicity of the molecule at the target site. In certain embodiments, the amino acid substitution maintains the bulk of the amino acid side group.

[00227] In various embodiments, the amino acid substitution is a non-conservative substitution. In some embodiments, the amino acid substitution produces significant changes in the peptide property. In certain embodiments, a hydrophilic residue is substituted by a hydrophobic residue. In certain other embodiments, a hydrophobic residue is substituted by a hydrophilic residue. In certain embodiments, a cysteine or proline is substituted by another residue. In certain other embodiments, a non-cysteine or non-proline is substituted by a cysteine or proline. In certain embodiments, a residue having an electropositive side group is substituted by an electronegative residue. In certain other embodiments, a residue having an electronegative side group is substituted by an electropositive residue. In certain embodiments, a residue having a bulky side group is substituted by a residue not having a side group. In certain other embodiments, a residue not having a side group is substituted by a residue having a bulky side group.

1.1.4.3. Chemical Modification of the Variant of Helix 3 of ApoC-II

[00228] In certain embodiments, the variant of helix 3 of apoC-II comprises at least one chemical modification. In some embodiments, the chemical modification increases the affinity of the variant of helix 3 for binding to lipoproteins. In some embodiments, the chemical modification increases the ability of the variant of helix 3 for activating lipolysis by LPL.

[00229] In some embodiments, the chemical modification is at the N-terminus of the variant of helix 3 of apoC-II. In some embodiments, the chemical modification is at the C-terminus of the variant of helix 3 of apoC-II. In various embodiments, the amino-end of the

N-terminus and/or at the carboxyl-end of the C-terminus can be modified by conjugation with various functional groups. Neutralization of the terminal charge of synthetic peptide mimics of apolipoproteins has been shown to increase their lipid affinity (Yancey et al., *Biochem.* 34:7955-7965, 1995; Venkatachalapathi et al., *Protein: Structure, Function and Genetics* 15:349-359, 1993). For example, acetylation of the amino terminal end of amphipathic peptides increases the lipid affinity of the peptide (Mishra et al., *J. Biol. Chem.* 269:7185-7191, 1994). Other possible end modifications are described, for example, in Brouillette et al., *Biochem. Biophys. Acta* 1256: 103-129, 1995; Mishra et al., *J. Biol. Chem.* 269:7185-7191, 1994; and Mishra et al., *J. Biol. Chem.* 270:1602-1611, 1995.

[00230] In some embodiments, the chemical modification is at an amino acid side group of the variant of helix 3 of apoC-II. Modifications at amino acid side groups include, without limitation, acylation of lysine ϵ -amino groups, N-alkylation of arginine, histidine, or lysine, alkylation of glutamic or aspartic carboxylic acid groups, lactam formation via cyclization of lysine ϵ -amino groups with glutamic or aspartic acid side group carboxyl groups, hydrocarbon “stapling” (e.g., to stabilize alpha-helix conformations), and deamidation of glutamine or asparagine. In some embodiments, the variant of helix 3 of apoC-II is modified by replacement of one or more side groups with other side groups such as alkyl, lower alkyl, cyclic 4-, 5-, 6-, to 7-membered alkyl, amide, amide lower alkyl, amide di(lower alkyl), lower alkoxy, hydroxy, carboxy and the lower ester derivatives thereof, and with 4-, 5-, 6-, to 7-membered heterocyclics. For example, proline analogs can be made in which the ring size of the proline residue is changed from a 5-membered ring to a 4-, 6-, or 7-membered ring. Cyclic groups can be saturated or unsaturated, and if unsaturated, can be aromatic or non-aromatic. Heterocyclic groups can contain one or more nitrogen, oxygen, and/or sulphur heteroatoms. Examples of such groups include furazanyl, furyl, imidazolidinyl, imidazolyl, imidazolinyl, isothiazolyl, isoxazolyl, morpholinyl (e.g., morpholino), oxazolyl, piperazinyl (e.g., 1-piperazinyl), piperidyl (e.g., 1-piperidyl, piperidino), pyranyl, pyrazinyl, pyrazolidinyl, pyrazolinyl, pyrazolyl, pyridazinyl, pyridyl, pyrimidinyl, pyrrolidinyl (e.g., 1-pyrrolidinyl), pyrrolinyl, pyrrolyl, thiadiazolyl, thiazolyl, thienyl, thiomorpholinyl (e.g., thiomorpholino), and triazolyl groups. These heterocyclic groups can be substituted or unsubstituted. Where a group is substituted, the substituent can be alkyl, alkoxy, halogen, oxygen, or substituted or unsubstituted phenyl. Peptides, as well as peptide analogs and mimetics, can also be covalently bound to one or more of a variety of nonproteinaceous polymers, for example,

polyethylene glycol, polypropylene glycol, or polyoxyalkenes, as described in U.S. Pat. Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192; and 4,179,337.

[00231] In certain embodiments, the chemical modification is non-covalent modification. In certain other embodiments, the chemical modification is covalent linkage. In various embodiments, the chemical modification can be phosphorylation, glycosylation, or lipidation.

[00232] In some embodiments, the chemical modification is a covalent linkage of a fatty acid. In certain embodiments, the fatty acid is saturated. In certain other embodiments, the fatty acid is unsaturated.

[00233] In various embodiments, the fatty acid comprises 2 to 30 carbons, such as 4 to 26 carbons, 6 to 24 carbons, 10 to 20 carbons, 12 to 18 carbons, and 14 to 16 carbons. In certain embodiments, the fatty acid comprises 6 carbons. In certain embodiments, the fatty acid comprises 7 carbons. In certain embodiments, the fatty acid comprises 8 carbons. In certain embodiments, the fatty acid comprises 9 carbons. In certain embodiments, the fatty acid comprises 10 carbons. In certain embodiments, the fatty acid comprises 11 carbons. In certain embodiments, the fatty acid comprises 12 carbons. In certain embodiments, the fatty acid comprises 13 carbons. In certain embodiments, the fatty acid comprises 14 carbons. In certain embodiments, the fatty acid comprises 15 carbons. In certain embodiments, the fatty acid comprises 16 carbons. In certain embodiments, the fatty acid comprises 17 carbons. In certain embodiments, the fatty acid comprises 18 carbons. In certain embodiments, the fatty acid comprises 19 carbons. In certain embodiments, the fatty acid comprises 20 carbons. In certain embodiments, the fatty acid comprises 21 carbons. In certain embodiments, the fatty acid comprises 22 carbons. In certain embodiments, the fatty acid comprises 23 carbons. In certain embodiments, the fatty acid comprises 24 carbons.

[00234] In some embodiments, the unsaturated fatty acid has 1, 2, 3, 4, 5, or 6 double bonds. In certain embodiments, the unsaturated fatty acid is arachidonic acid. In certain embodiments, the unsaturated fatty acid is linoleic acid. In certain embodiments, the unsaturated fatty acid is oleic acid. In certain embodiments, the unsaturated fatty acid is palmitoleic acid. In certain embodiments, the unsaturated fatty acid is linolenic acid. In certain embodiments, the unsaturated fatty acid is eicosapentaenoic acid. In certain embodiments, the unsaturated fatty acid is docosahexaenoic acid.

[00235] In specific embodiments, the fatty acid is palmitic acid. In specific embodiments, the fatty acid is stearic acid.

[00236] In certain embodiments, the isolated apoC-II mimetic peptide comprises a stearic acid covalently linked to the N-terminal amino acid. In some of these embodiments, the N-terminal amino acid is alanine.

[00237] In certain embodiments, the isolated apoC-II mimetic peptide comprises a stearic acid covalently linked to the side group of the lysine residue. In specific embodiments, when the isolated apoC-II mimetic peptide comprises a lysine at position 60, the stearic acid is covalently linked to the lysine residue. In specific embodiments, when the isolated apoC-II mimetic peptide comprises a lysine at position 76, the stearic acid is covalently linked to the lysine residue.

1.1.5. Preparation of ApoC-II Mimetic Peptide

[00238] Also disclosed herein are methods for producing the isolated apoC-II mimetic peptide.

1.1.5.1. Recombinant Synthesis

[00239] In certain embodiments, the isolated apoC-II mimetic peptide is produced recombinantly, for example using bacterial, yeast, or eukaryotic expression systems.

[00240] For recombinant production, a polynucleotide sequence encoding the single or multi-domain peptide is inserted into an appropriate expression vehicle, that is, a vector which contains the necessary elements for the transcription and translation of the inserted coding sequence, or in the case of an RNA viral vector, the necessary elements for replication and translation. The expression vehicle is then transfected into a suitable target cell which will express the single or multi-domain peptide. Depending on the expression system used, the expressed peptide is then isolated by procedures well-established in the art. Methods for recombinant protein and peptide production are well known in the art.

[00241] To increase efficiency of production, the polynucleotide can be designed to encode multiple units of the single or multi-domain peptide separated by enzymatic cleavage sites. The resulting polypeptide can be cleaved (e.g., by treatment with the appropriate enzyme) in order to recover the peptide units. This can increase the yield of peptides driven by a single promoter. In some embodiments, a polycistronic polynucleotide can be designed so that a single mRNA is transcribed which encodes multiple peptides, each coding region operatively linked to a cap-independent translation control sequence, for example, an internal ribosome entry site (IRES). When used in appropriate viral expression systems, the translation of each

peptide encoded by the mRNA is directed internally in the transcript, for example, by the IRES. Thus, the polycistronic construct directs the transcription of a single, large polycistronic mRNA which, in turn, directs the translation of multiple, individual peptides. This approach eliminates the production and enzymatic processing of polypeptides and can significantly increase yield of peptide driven by a single promoter.

[00242] A variety of host-expression vector systems can be utilized to express the peptides described herein. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage DNA or plasmid DNA expression vectors containing an appropriate coding sequence; yeast or filamentous fungi transformed with recombinant yeast or fungi expression vectors containing an appropriate coding sequence; insect cell systems infected with recombinant virus expression vectors (e.g., baculovirus) containing an appropriate coding sequence; plant cell systems infected with recombinant virus expression vectors (e.g., cauliflower mosaic virus (CaMV) or tobacco mosaic virus (TMV)) or transformed with recombinant plasmid expression vectors (e.g., Ti plasmid) containing an appropriate coding sequence; or animal cell systems.

[00243] The expression elements of the expression systems vary in their strength and specificities. Depending on the host/vector system utilized, any of a number of suitable transcription and translation elements, including constitutive and inducible promoters, can be used in the expression vector. For example, when cloning in bacterial systems, inducible promoters such as pL of bacteriophage λ , plac, ptrp, ptac (ptrp-lac hybrid promoter) and the like can be used. When cloning in insect cell systems, promoters such as the baculovirus polyhedron promoter can be used. When cloning in plant cell systems, promoters derived from the genome of plant cells (e.g., heat shock promoters, the promoter for the small subunit of RUBISCO, the promoter for the chlorophyll a/b binding protein) or from plant viruses (e.g., the 35S RNA promoter of CaMV, the coat protein promoter of TMV) can be used. When cloning in mammalian cell systems, promoters derived from the genome of mammalian cells (e.g., metallothionein promoter) or from mammalian viruses (e.g., the adenovirus late promoter, the vaccinia virus 7.5 K promoter) can be used.

1.1.5.2. Chemical Synthesis

[00244] In certain other embodiments, the isolated apoC-II mimetic peptide is produced by chemical synthesis. In some embodiments, the peptide is produced using liquid phase peptide

synthesis techniques. In some other embodiments, the peptide is produced using solid phase peptide synthesis techniques.

[00245] Peptides having either the D- or L-configuration can be synthesized by automated solid phase procedures well known in the art. Suitable syntheses can be performed by utilizing “Boc” or “Fmoc” procedures. Techniques and procedures for solid phase synthesis are well-known in the art. The single and multi-domain peptides can also be prepared by way of segment condensation, as described, for example, in Liu et al., *Tetrahedron Lett.* 37:933-936, 1996; Baca et al., *J. Am. Chem. Soc.* 117: 1881-1887, 1995; Tam et al., *Int. J. Peptide Protein Res.* 45:209-216, 1995; Schnolzer and Kent, *Science* 256:221-225, 1992; Liu and Tam, *J. Am. Chem. Soc.* 116:4149-4153, 1994; Liu and Tam, *Proc. Natl. Acad. Sci. USA* 91:6584-6588, 1994; and Yamashiro and Li, *Int. J. Peptide Protein Res.* 31:322-334, 1988). This is particularly the case with glycine containing peptides. Other methods useful for synthesizing the single and multi-domain peptides of the disclosure are described in Nakagawa et al., *J. Am. Chem. Soc.* 107:7087-7092, 1985.

[00246] Additional exemplary techniques known to those of ordinary skill in the art of peptide and peptide analog synthesis are taught by Bodanszky, M. and Bodanszky, A., *The Practice of Peptide Synthesis*, Springer Verlag, New York, 1994; and by Jones, J., *Amino Acid and Peptide Synthesis*, 2nd ed., Oxford University Press, 2002. The Bodanszky and Jones references detail the parameters and techniques for activating and coupling amino acids and amino acid derivatives. Moreover, the references teach how to select, use and remove various useful functional and protecting groups.

[00247] Peptides having either the D- or L-configuration can also be purchased from commercial suppliers of synthetic peptides. Such suppliers include, for example, Advanced ChemTech (Louisville, KY), Applied Biosystems (Foster City, CA), Bachem (Torrance, CA), Anaspec (San Jose, CA), and Cell Essentials (Boston, MA).

1.1.6. Purification of ApoC-II Mimetic Peptide

[00248] The peptides or peptide analogs of the disclosure can be purified by many techniques well known in the art, such as reverse phase chromatography, high performance liquid chromatography, ion exchange chromatography, size exclusion chromatography, affinity chromatography, gel electrophoresis, and the like. The actual conditions used to purify a particular single or multi-domain peptide or peptide analog will depend, in part, on

synthesis strategy and on factors such as net charge, hydrophobicity, hydrophilicity, and the like, and will be apparent to those of ordinary skill in the art.

[00249] In various embodiments, the isolated apoC-II mimetic peptide further comprises a purification tag. In some embodiments, the purification tag is a polyhistidine-tag, a myc-tag, or an HA-tag.

1.2. Pharmaceutical Composition

[00250] Also provided herein are pharmaceutical compositions comprising one or more isolated apoC-II mimetic peptides described herein and a pharmaceutically acceptable carrier.

[00251] Any carrier which can supply an active peptide (e.g., without destroying or damaging the peptide within the carrier) is a suitable carrier, and such carriers are well known in the art. The apoC-II mimetic peptide can be formulated, e.g., using any formulation currently used to formulate other therapeutic peptides, such as insulins, GLP-1 agonists, and all approved peptides disclosed in the THPdb database of FDA approved therapeutic peptides and proteins (crdd.osdd.net/raghava/thpdb/).

[00252] The apoC-II mimetic peptide based pharmaceutical composition can be in the form of a solid, semi-solid or liquid dosage form: such as powder, solution, elixir, syrup, suspension, cream, drops, paste and spray. As those skilled in the art can recognize, depending on the chosen route of administration the composition form is determined.

[00253] In some embodiments, compositions are formulated for administration by any suitable route, including but not limited to, orally (e.g., such as in the form of tablets, capsules, granules or powders), sublingually, buccally, parenterally (such as by subcutaneous, intravenous, intramuscular, intradermal, or intrasternal injection or infusion (e.g., as sterile injectable aqueous or non-aqueous solutions or suspensions, etc.)), nasally (including administration to the nasal membranes, such as by inhalation spray), topically (such as in the form of a cream or ointment), transdermally (such as by transdermal patch), rectally (such as in the form of suppositories), or by surgical implantation at a particular site, etc.

[00254] In certain embodiments, the pharmaceutical compositions are formulated to be suitable for subcutaneous injection.

1.3. Methods of Treatment

[00255] Also provided herein are methods for treating dyslipidemic and vascular disorders, including but not limited to hyperlipidemia, hyperlipoproteinemia, hypercholesterolemia,

hypertriglyceridemia, HDL deficiency, coronary artery disease, atherosclerosis, thrombotic stroke, peripheral vascular disease, restenosis, acute coronary syndrome, and reperfusion myocardial injury.

[00256] In some embodiments, the methods comprise administering the peptide or the pharmaceutical composition as described herein to a patient with hypertriglyceridemia.

[00257] In some embodiments, the patient is at risk of developing hypertriglyceridemia.

[00258] In various embodiments, the patient has hypertriglyceridemia, based on fasting plasma or serum TG concentration above a certain level. In some embodiments, the patient has mild hypertriglyceridemia, defined as pre-treatment serum triglyceride (TG) concentration between 150 mg/dL and 199 mg/dL. In some embodiments, the patient has moderate hypertriglyceridemia, defined as pre-treatment serum triglyceride (TG) concentration between 200 mg/dL and 999 mg/dL, such as between 200 mg/dL to 499 mg/dL and between 500 mg/dL to 999 mg/dL. In some embodiments, the patient has severe hypertriglyceridemia, defined as pre-treatment serum triglyceride (TG) concentration between 1000 mg/dL and 1999 mg/dL. In some embodiments, the patient has very severe hypertriglyceridemia, defined as pre-treatment serum triglyceride (TG) concentration equal to or higher than 2000 mg/dL.

[00259] In some embodiments, the patient has above-normal LDL-c levels. In some other embodiments, the patient has below-normal levels of HDL-c.

[00260] In certain embodiments, the hypertriglyceridemia is caused by LPL deficiency. In some of these embodiments, the patient's hypertriglyceridemia is familial lipoprotein lipase deficiency.

[00261] In some embodiments, the LPL deficiency is caused by a gene mutation. In some of these embodiments, the gene mutation is detected by DNA sequence analysis. In various embodiments, the LPL deficiency is caused by a mutation in the LPL, APOC2, APOA5, GPIHBP1, or LMF1 gene.

[00262] In certain embodiments, the LPL deficiency is caused by a mutation in the LPL gene. In some of these embodiments, the mutation leads to reduced LPL enzyme activity. In some of these embodiments, the mutation leads to absent LPL enzyme activity.

[00263] In certain embodiments, the patient's hypertriglyceridemia is monogenic. In certain other embodiments, the patient's hypertriglyceridemia is polygenic. In certain embodiments, the mutations are present homozygous state. In certain embodiments, the mutations are present in heterozygous state.

[00264] In some embodiments, the LPL deficiency is diagnosed by the absence of LPL activity in serum of the patient.

[00265] In certain embodiments, the hypertriglyceridemia is caused by apoC-II deficiency. In certain embodiments, the hypertriglyceridemia is caused by elevated apoC-III.

[00266] In some embodiments, the patient has diabetes. In some embodiments, the patient has metabolic syndrome. In some of these embodiments, the metabolic syndrome is obesity. In some embodiments, the patient has pancreatitis. In some of these embodiments, the patient has acute pancreatitis. In some embodiments, the patient has steatosis or steatohepatitis. In some of these embodiments, the steatosis or steatohepatitis is alcohol-related. In some of these embodiments, the steatosis or steatohepatitis is non-alcoholic. In some embodiments, the patient has cardiovascular disease. In some of these embodiments, the patient has acute cardiovascular disease. In some embodiments, the patient has atherosclerosis. In some embodiments, the patient has taken medication that can lead to hypertriglyceridemia.

[00267] In certain embodiments, the patient is an adult. In certain other embodiments, the patient is a child.

[00268] In various embodiments, the peptide or the pharmaceutical composition is administered in an amount, on a schedule, and for a duration sufficient to reduce blood triglyceride level of the patient. In some embodiments, the polypeptide is administered in an amount, on a schedule, and for a duration sufficient to decrease triglyceride levels by at least 5%, 10%, 15%, 20%, 25%, or more as compared to levels just prior to initiation of treatment. In certain embodiments, the polypeptide is administered in an amount, on a dosage schedule, and for a duration sufficient to decrease triglyceride levels by at least 30%, 35%, 40%, 45%, 50%, or more. In particular embodiments, the polypeptide is administered in an amount, on a schedule, and for a time sufficient to reduce triglyceride levels by at least 55%, 60%, 65%, 70%, or more.

[00269] The treatment can consist of a single dose or a plurality of doses over a period of time.

EXAMPLES

[00270] Below are examples of specific embodiments for carrying out the present invention. The examples are offered for illustrative purposes only, and are not intended to limit the scope of the present invention in any way. Efforts have been made to ensure

accuracy with respect to numbers used (e.g., amounts, temperatures, etc.), but some experimental error and deviation should, of course, be allowed for.

[00271] The practice of the present invention will employ, unless otherwise indicated, conventional methods of protein chemistry, biochemistry, recombinant DNA techniques and pharmacology, within the skill of the art. Such techniques are explained fully in the literature.

Methods

[00272] LPL assay

[00273] Preparation of LPL: frozen stock of 10 μ L of 0.5 mg/mL bovine LPL was used and diluted 1:100 (10 μ L + 990 μ L of PBS).

[00274] Preparation of 20% Intralipid: 2000 mg/dL TG Intralipid was used to prepare 3 mg/dL TG Intralipid by diluting 1.5 μ L of the stock with 998.5 μ L PBS, or by diluting the stock 1:10 first and then mixing 15 μ L of the diluted stock with 985 μ L PBS. When apoC-II deficient human serum was used as a substrate, a frozen 10 μ L aliquot of 1711 mg/dL (TG) was diluted by adding 990 μ L PBS (1:100 dilution).

[00275] Preparation of free fatty acid (NEFA) assay: NEFA reagents were purchased from Wako Diagnostics, Wako Life Sciences, Inc. Reagent A was prepared by mixing Coloring Reagent A with Solvent A (225 μ L per well). Reagent B was prepared by mixing Coloring Reagent B with Solvent B (75 μ L per well). 5 nmol oleic acid standard was prepared by diluting 20 μ L of 1 mM oleic acid stock (WAKO NEFA standard solution) with 180 μ L of 0.2% BSA.

[00276] LPL activity assay: 10 μ L of Intralipid, human plasma sample, or mouse plasma sample was added in each well using repeater pipette. 10 μ L (or 15 μ L in the wells with no LPL control) of PBS and 10 μ L of 1% BSA was added in each well using repeater pipette. 10 μ L of peptides were added to corresponding wells in vertical triplicates (lowest concentration is on a left, highest is on the right). The plate was spun down at 1000 RPM for 10 seconds to mix and to remove the solutions from the wall of each well. The plate was put on ice and 5 μ L of LPL was added using repeater pipette. The plate was spun down again at 1000 RPM for 10 seconds. The plate was covered and put back on ice for an incubation of 30 min. The plate was then moved to 37°C and incubated for 1 hour.

[00277] Free fatty acids (NEFA) assay: 45 μ L of 5 nmol standard and 0 nmol control (0.2% BSA) were added to corresponding wells. 225 μ L of reagent A was added in each well using the reagent tray and 12-channel pipette. The plate was incubated for 10 min before 75

μL of reagent B was added to each well using the reagent tray and 12-channel pipette. The plate was not covered due to the high volume. The absorbance at 550 nm and 660 nm was measured.

[00278] ApoC-III displacement study

[00279] VLDL or chylomicrons isolated from human plasma were incubated with an apoC-II mimetic peptide for 1 hour at 37°C and washed 3 times with PBS using spin filter with 100 kDa cutoff membrane. Supernatants were brought to the original volume with PBS and loaded to 4-12% Bis-TRIS SDS PAGE gel. ApoC-I, apoC-II, apoC-III, and apoE bands were identified by their corresponding size.

[00280] Inhibition of ApoC-III study

[00281] LPL assay was performed using Intralipid, high TG human plasma (1:50 dilution), or apoC-III transgenic mouse plasma (1:50 dilution). When Intralipid was used as a substrate, 25 or 50 μM recombinant ApoC-III was added as an inhibitor. When the high TG human plasma was used as a substrate, 25 or 50 μM recombinant ApoC-III was added as an inhibitor with or without pre-incubation at 37°C for 1 hour. An apoC-II mimetic peptide was added at various concentrations.

[00282] In vivo study

[00283] C57Bl/6 mice (wild-type), apoC-II knockout (apoC-II KO) mice, and apoC-III transgenic mice were used for *in vivo* study. The C57Bl/6 mice (wild-type) were purchased from Taconic Biosciences Inc. The apoC-II KO mice were created as described in Sakurai et al., *J Pharmacol Exp Ther* 356:341–353, 2016. The apoC-III transgenic mice were generated by overexpressing human apoC-III in C57Bl/6 mice. See Qu et al., *J. Lipid Res.* 48:1476–1487, 2007.

[00284] Mice were fasted overnight (approximately 12 hours) before the study and 6 hours during the study, the next day. The apoC-II mimetic peptides were synthetically produced and free of biological contamination (not available as FDA approved compounds/treatment). Peptides were produced and used in sterile conditions and filtered by 0.2 nm filter before injections in a single dose 1-10 μmol/kg body weight (B. W.) administered either subcutaneous injection (S.Q. or S.C.), intraperitoneal injection (I.P.) or intravenous injection (I.V.) (in a sterile, pharmaceutical grade PBS solution at a volume ≤200 μL) as described in Sakurai et al., *J Pharmacol Exp Ther* 356:341–353, 2016.

[00285] In vivo Intralipid study

[00286] In some experiments, each animal received 20% Intralipid (FDA approved, Fresenius Kabi, Uppsala, Sweden) in a single dose either by oral gavage (using special sterile single-use non-toxic animal feeding needles) in a dose 10-20 $\mu\text{L}/\text{gram}$ body weight (B.W.) (Tuzcu et al. *Drug Chem Toxicol* 37: 261-267, 2014; Kusminski et al. *Nature Medicine* 18: 1539-1549, 2012), or by intraperitoneal injection in a single bolus of 0.1 – 1.0 mL (Mahadero et al. *American J Surgery* 164: 45-50, 1992), or by intragastric administration of a dietetic commercially-available liquid oil (food grade vegetable, corn, soy or olive oil) in a single dose 10-20 $\mu\text{L}/\text{gram}$ body weight. These oils were a source of TG - when absorbed TG tends to be broken down within 3 h (peak in 30 min) (Kritchevsky *Nutritional Biochemistry* 6:172-178, 1995). Blood samples (20-30 μL) were obtained from the retro-orbital plexus at time points: -1 h, 0 h, 1 h, 3 h, 6 h and 24 h for measurements of lipids. Mice were returned to metabolic cages after the 6 h time point where the food was put back. Once the study was completed (after 24 hours from the fat administration), mice were euthanized and organs collected.

[00287] In some cases, mice were returned to regular cages and housed for another 2-3 weeks for a complete recovery, before being studied in another experiment. There was 1 control group that received a sterile pharmaceutical grade saline in place of peptide.

[00288] The study design is presented on FIG. 55.

[00289] In vivo Triton WR1339 study

[00290] The ability of apolipoprotein mimetic peptides to normalize hypertriglyceridemia in mice treated with an LPL inhibitor, Tyloxapol, in presence of high TG provided by 20% Intralipid, was assessed.

[00291] Mice were fasted overnight (approximately 12 hours) before the study and 6 hours during the study, the next day. The apo mimetic peptides were synthetically produced and free of biological contamination (not available as FDA approved compounds/treatment). Peptides were produced and used in sterile conditions and filtered by 0.2 nm filter before injections in a single dose 1-5 $\mu\text{mol}/\text{kg}$ B.W. either S.C. or I.P. or I.V. (in a sterile, pharmaceutical grade PBS solution at a volume ≤ 200 μL). Next, 10% Tyloxapol (Triton WR-1339, T-0307, Sigma-Aldrich) was injected I.V. in a single dose of 5 $\mu\text{L}/\text{gram}$ B.W., as described earlier (Zhang Y.L. et al., *J Biol Chem.* 279: 19362-19374, 2004; Abe C. et al., *J. Nutr.* 137: 345–350, 2007). Tyloxapol is a nonionic detergent that inhibits LPL and hence clearance of triglycerides from the serum (Rasouli M. et al., *J Clin. Diagn. Research.* 10: BF01-BF05,

2016). Finally, each animal received 20% Intralipid (FDA approved, Fresenius Kabi, Uppsala, Sweden) in a single dose either by oral gavage (using special sterile single-use non-toxic animal feeding needles) — in a dose 10-20 μL /gram B.W. (Tuzcu K. et al., *Drug Chem Toxicol.*, 37: 261-267, 2014, Kusminski C. et al., *Nature Medicine*. 18: 1539-1549, 2012), or by intraperitoneal injection — in a single bolus of 0.1-1.0 mL (Mahadero G. et al., *American J Surgery*. 164: 45-50, 1992). Blood samples (20-30 μL) were obtained from the retro-orbital plexus at time points: -1 h, 0 h, 1 h, 3 h, 6 h and 24 h for measurements of lipids. Mice were returned to metabolic cages after the 6 h time point where the food was put back. Once the study was completed (after 24 hours from the Intralipid administration), mice were euthanized and organs collected.

[00292] In some cases, mice were returned to regular cages and housed for another 2-3 weeks for a complete recovery, before being studied in another experiment. There were 2 control groups, both receiving a sterile pharmaceutical grade saline in place of either peptide or Triton WR1339).

[00293] The study design is presented in FIG. 56.

[00294] In vivo studies in the obese monkey model

[00295] Monkeys with a TG level greater than 2 mmol/L were selected, trained and assigned into 3 groups (control, 1 mg/kg Delta6PV, and 5 mg/kg Delta6PV). The desired dose was injected intravenously into the animals at 0 h time point. The monkeys were monitored over the course of 72 hours and blood samples at different time points were collected for biomarker analysis.

Example 1: Peptide Design

[00296] Apolipoprotein C-II (apoC-II) is a physiologic activator of lipoprotein lipase (LPL) and it can promote the hydrolysis of triglyceride in lipoproteins. A lack of functional apoC-II leads to severe hypertriglyceridemia, similar to what is seen for a genetic deficiency of LPL (Breckenridge et al., *N Engl J Med*. 298: 1265-1273, 1978).

[00297] The mature form of human apoC-II consists of 79 amino acid residues. The secondary structure of apoC-II has been determined by various measures, with residues 17-38 designated as helix 1, residues 45-57 designated as helix 2, and residues 65-74/75 designated as helix 3 (Zdunek et al., *Biochemistry* 42: 1872-1889, 2003). The C-terminal one-third of apoC-II, which is needed for activation of LPL, is more conserved between different species than the N-terminal two thirds of apoC-II, which is needed for lipid binding (Shen et al., *Gene*

254:189-198, 2000). The homology of amino acid residues 40-58 of mature human apoC-II protein across various species is shown on FIG. 1.

[00298] In order to make a clinically useful LPL activating peptide, we designed single helical and bihelical apoC-II mimetic peptides. We introduced various modifications, such as elongations, truncations, mutations, and chemical modifications to the helices of native human apoC-II protein.

[00299] Some modifications, such as amino acid substitutions, increased the hydrophobic moment of the native amino acid sequence of apoC-II. For example, as shown in FIG. 2A and FIG. 2B, the T to D substitution at position 40, the P to K substitution at position 43, the D to F substitution at position 46, and the S to F substitution at position 56 increased the hydrophobic moment of amino acid residues 40-57 from 6.04 to 9.62. As shown in FIG. 3A and FIG. 3B, the T to D substitution at position 40, the P to K substitution at position 43, the A to E substitution at position 44, the D to F substitution at position 46, and the S to F substitution at position 56 increased the hydrophobic moment of amino acid residues 40-57 from 6.04 to 11.08. FIG. 4A and FIG. 4B show that a T to D substitution at position 40, the P to K substitution at position 43, the A to E substitution at position 44, the D to F substitution at position 46, the S to E substitution at position 54, and the S to F substitution at position 56 increased the hydrophobic moment of amino acid residues 40-57 from 6.04 to 12.48.

[00300] Chemical modifications were also introduced to increase the association of the apoC-II mimetic peptides for lipid particles and to enhance other therapeutic characteristics of the apoC-II mimetic peptides, such as increased cell permeability and/or prolonged biological half-life. FIG. 5 shows the examples of apoC-II mimetic peptides modified by stearic acid.

[00301] A list of apoC-II mimetic peptides and their sequences is shown in Table 2 below, where “nL” indicates norleucine and “Aib” indicates aminoisobutyric acid.

Table 2		
Name	Sequence	SEQ ID NO
Delta6	DYLKEVFEKLRDLYEKFTAAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 6
Delta6V	DYLKEVFEKLRDLYEKFTAAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 7
Delta6L	DYLKEVFEKLRDLYEKFTAAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 8
Delta6PV	DYLKEVFEKLRDLYEKFTPAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 9
Delta6PL	DYLKEVFEKLRDLYEKFTPAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 10
Delta6MP	DYLKEVFEKLRDLYEKFTPAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 11
Delta6MPK-4R	DYLREVFERLRDLYERKTPAnLSTYTGIFTDQVLSV RGE	SEQ ID NO: 12
Delta6-NH	DYLKEVFEKLRDLYEKFTAAnLSTYTGIFTDQVLSV LKGE _{NH2}	SEQ ID NO: 13
Delta6b-P	DYLKEVFEKLRDLYEKK _{C18} TPAnLSTYTGIFTDQVLS VLKGE	SEQ ID NO: 14
Delta6b-A	DYLKEVFEKLRDLYEKK _{C18} TAAAnLSTYTGIFTDQVLS SVLKGE	SEQ ID NO: 15
Delta4	DYLKAVFEKLRDLYSKFTAAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 16
Delta4 S15E	DYLKAVFEKLRDLYEKFTAAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 17
Delta4 T18F	DYLKAVFEKLRDLYSKFFAAAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 18
Delta4 Y24S	DYLKAVFEKLRDLYSKFTAAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 19
Delta4 Q31R	DYLKAVFEKLRDLYSKFTAAnLSTYTGIFTDRVLSV LKGE	SEQ ID NO: 20
Delta4 Q31K	DYLKAVFEKLRDLYSKFTAAnLSTYTGIFTDKVLSV LKGE	SEQ ID NO: 21
Delta4 Truncated	KRDLYEKKFAAnLSTYTGIFTDQVLSVLKGE	SEQ ID NO: 22
mono-acyl-Delta4	C ₁₈ DYLKAVFEKLRDLYSKFTAAnLSTYTGIFTDQVLS VLKGE	SEQ ID NO: 23
bi-acyl-Delta4	C ₁₈ DYLKAVFEKK _{C18} RDLYSKFTAAnLSTYTGIFTDQ VLSVLKGE	SEQ ID NO: 24
Delta4b	DYLKAVFEKLRDLYSKFTPAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 25
Delta4b S15E	DYLKAVFEKLRDLYEKFTPAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 26
Delta4b S15K	DYLKAVFEKLRDLYKKFTPAnLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 27

Table 2		
Name	Sequence	SEQ ID NO
Delta4b R11K	DYLKAVFEKLRDLYSKFTPAMSTYTGIFTDQVLSVLK GEE	SEQ ID NO: 28
Delta4b Y2W	DWLKAVFEKLRDLYSKFTPAMSTYTGIFTDQVLSV LKGE	SEQ ID NO: 29
Delta4b T18F	DYLKAVFEKLRDLYSKFFPAMSTYTGIFTDQVLSVL KGE	SEQ ID NO: 30
Delta4b V6F/F7Y/Y14L/ F17A/T18F	DYLKAFYEKLRDLLSKAFPAMSTYTGIFTDQVLSVL KGE	SEQ ID NO: 31
Delta4b-1	DYLKAVK _{C18} EKLRDLYSKFTPAMSTYTGIFTDQVLS VLKGE	SEQ ID NO: 32
Delta4b-2	DYLKAVFEKK _{C18} RDLYSKFTPAMSTYTGIFTDQVLS VLKGE	SEQ ID NO: 33
Delta4b-3	DYLKAVFEKLRDLYSKK _{C18} TPAMSTYTGIFTDQVLS VLKGE	SEQ ID NO: 34
Delta4c	DYLKAVFEKLRDLYSKFTAAC _{C16} STYTGIFTDQVLS VLKGE	SEQ ID NO: 35
Delta4c trunc3	KAVFEKLRDLYSKFTAAC _{C16} STYTGIFTDQVLSVLK GEE	SEQ ID NO: 36
Delta4c trunc4	DVFEKLRDLYSKFTAAC _{C16} STYTGIFTDQVLSVLK GEE	SEQ ID NO: 37
Delta4c trunc7	EKLRDLYSKFTAAC _{C16} STYTGIFTDQVLSVLKGE	SEQ ID NO: 38
Delta4c trunc10	RDLYSKFTAAC _{C16} STYTGIFTDQVLSVLKGE	SEQ ID NO: 39
Delta4c trunc11	DLYSKFTAAC _{C16} STYTGIFTDQVLSVLKGE	SEQ ID NO: 40
Delta4c trunc14	DKFTAAC _{C16} STYTGIFTDQVLSVLKGE	SEQ ID NO: 41
Delta5	DYLKEVFEKLRDLYSKFTAANLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 42
Delta5b Y2W	DWLKAVFEKLRDLYEKFTPAMSTYTGIFTDQVLSV LKGE	SEQ ID NO: 43
Delta5b T18F	DYLKAVFEKLRDLYEKFFPAMSTYTGIFTDQVLSVL KGE	SEQ ID NO: 44
Delta5b Y2W/T18F	DWLKAVFEKLRDLYEKFFPAMSTYTGIFTDQVLSV LKGE	SEQ ID NO: 45
Delta5b Y2W M21NL	DWLKAVFEKLRDLYEKFTPANLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 46
Delta5b T18F M21NL	DYLKAVFEKLRDLYEKFFPANLSTYTGIFTDQVLSVL KGE	SEQ ID NO: 47
Delta5b Y2W/T18F M21NL	DWLKAVFEKLRDLYEKFFPANLSTYTGIFTDQVLSV LKGE	SEQ ID NO: 48
C2thirdhelix	AMSTYTGIFTDQVLSVLKGE	SEQ ID NO: 49
C2thirdhelixStearylpos1	_{C18} AMSTYTGIFTDQVLSVLKGE	SEQ ID NO: 50
C2thirdhelix	AK _{C18} STYTGIFTDQVLSVLKGE	SEQ ID NO: 51

Table 2		
Name	Sequence	SEQ ID NO
Stearylpos2		
C2thridhelix Stearylpos17	AMSTYTGIFTDQVLSVLK _{C18} GEE	SEQ ID NO: 52
Delta6T18F-PV	DYLKEVFELKRDLYEKFFPAVSTYTGIFTDQVLSVL KGEE	SEQ ID NO: 53
Delta6T18F-PV- AIB7,17	DYLKEVAibEKLKRDLYEKAibFPAVSTYTGIFTDQVL SVLKGEE	SEQ ID NO: 54
Delta13-31-PV	DKVKEFLSEYWEKAKEFAPAVSTYTGIFTDQVLSVL KGEE	SEQ ID NO: 55

Example 2: Exemplary ApoC-II mimetic peptides with amino acid substitutions promote TG hydrolysis *in vitro*

[00302] To determine the possible impact of apoC-II mimetic peptides on hydrolysis of TG, we assessed the effect of the peptides using an *in vitro* lipolysis assay.

[00303] LPL assay with apoC-II deficient patient sera showed that exemplary apoC-II mimetic peptides Delta4, Delta5, and Delta6, as well as Delta4 variants mono-acyl-Delta4 and bi-acyl-Delta4, were capable of promoting TG hydrolysis in the presence of LPL. At lower concentrations, most of the tested mimetic peptides, such as Delta4, Delta5, and Delta6, worked more efficiently than the native apoC-II protein (FIG. 6).

[00304] LPL assay using Intralipid as substrate confirmed that all apoC-II mimetic peptide tested were capable of promoting TG hydrolysis in the presence of LPL. The apoC-II mimetic peptides Delta4 and Delta5 worked more efficiently than the native apoC-II protein at concentrations of between about 1 nM to about 10 μ M (FIG. 7).

[00305] These results show that the exemplary apoC-II mimetic peptides Delta4, Delta5, and Delta6 are capable of promoting TG hydrolysis *in vitro*.

Example 3: Variants of Delta4 and third helix of apoC-II promote TG hydrolysis *in vitro*

[00306] The hinge region between helix 2 and helix 3 of apoC-II protein allows the two helices to retain a relatively straight conformation. The helix 3 bends away from helix2 uniformly in different directions over an angle of no more than about 20°. This conformation is believed to help helix 3, which contains the LPL activating domain, to attach to the micelle surface of lipoprotein. Incorporation of a proline at position 58 of the hinge domain preserves

and accentuates the conformation. Variants of Delta4, such as Delta4b, Delta4b-1, and Delta4b S54E, comprise a proline at position 58.

[00307] The apoC-II mimetic peptide C2thirdhelix comprises amino acid residues 59-79 of apoC-II protein. C2thirdhelixStearylpos1, C2thirdhelixStearylpos2, and C2thirdhelixStearylpos17 comprise a stearic acid modification of C2thirdhelix at different amino acid residues.

[00308] As shown in FIG. 8, most of the above-mentioned apoC-II mimetic peptides exhibited the ability to promote TG hydrolysis in the presence of LPL when apoC-II deficient patient sera were used as substrate. A closer look at the results of C2thirdhelix, C2thirdhelixStearylpos1, C2thirdhelixStearylpos2, and C2thirdhelixStearylpos17 revealed that lipidation enhanced the ability of C2thirdhelix to promote TG hydrolysis (FIG. 9). A comparison of the apoC-II mimetic peptides Delta4, Delta4b, and Delta4b-3 revealed that incorporation of proline at position 58 and the stearic acid modification at the lysine residue at position 56 enhanced the TG clearance ability of the exemplary apoC-II mimetic peptide Delta4 (FIG. 10). FIG. 11 confirmed that lipidation of apoC-II mimetic peptide Delta4b enhanced its ability to enhance lipolysis, especially at lower concentrations.

[00309] We further tested the apoC-II mimetic peptides in a wide range of concentrations from 100 pM to 10 μ M. The apoC-II mimetic peptides, Delta4b-1, Delta4b-2, Delta4b-3, Delta4b, Delta4b T57F, and Delta4, promoted TG hydrolysis in a dose-dependent manner. All of the apoC-II mimetic peptides tested in this experiment worked more efficiently than the full length apoC-II protein (FIG. 12).

[00310] Delta4c peptide comprised a palmitic acid modified lysine (K) at position 60, and its derivatives included truncations at the C-terminal by 3, 4, 7, 10, 11, or 14 amino acids. In an LPL assay with apoC-II deficient patient sera as substrate and over a range of peptide concentrations from 10 pM to 100 μ M, Delta4c and its truncation variants promoted TG hydrolysis (FIG. 13).

[00311] Together, these results show that variants of exemplary apoC-II mimetic peptide Delta4 and variants of third helix of apoC-II can promote TG hydrolysis *in vitro*.

Example 4: Variants of Delta5 promote TG hydrolysis *in vitro*

[00312] Additional amino acid substitutions were made to apoC-II mimetic peptide Delta5b, which comprises a proline at position 58 and a methionine at position 60.

[00313] LPL assay was performed using apoC-II deficient patient sera as substrate. The peptides were tested in a range of concentrations from 100 pM to 10 nM. As shown in FIG. 14, all variants of Delta5 with the indicated amino acid substitutions and Delta6 exhibited the ability to promote TG hydrolysis in a dose dependent matter.

[00314] These results show that variants of exemplary apoC-II mimetic peptide Delta5 are capable of promoting TG hydrolysis *in vitro*.

Example 5: Variants of Delta6 promote TG hydrolysis *in vitro*

[00315] The exemplary apoC-II mimetic peptide Delta6 was further modified to be more suitable for recombinant expression. Delta6L comprises a leucine (L) residue at position 60. Delta6PV comprises a proline (P) at position 58 and a valine (V) at position 60. Delta6PV comprises a proline (P) at position 58 and a leucine (V) at position 60. Delta6T18F-PV further comprises a phenylalanine (F) at position 57 compared to Delta6PV. Delta6T18F-PV-AIB7,17 further comprises an aminoisobutyric acid (Aib) at positions 46 and 56 and a phenylalanine (F) at position 57 compared to Delta6PV.

[00316] As shown in FIGs. 15A-D, Delta6 peptide and its variants Delta6L, Delta6PV, and Delta6PL activated LPL *in vitro* when hypertriglyceridemia patient sera were used as substrate. The triglyceride concentrations of the patient sera ranged from about 1000 mg/dL to about 1700 mg/dL.

[00317] We further tested the activity Delta6 peptide and its variant Delta6PV in a wide range of concentrations from 10 pM to 10 μ M. ApoC-II deficient patient sera were used as substrate. As shown in FIG. 51, Delta6 and Delta6PV activated LPL *in vitro* more efficiently than the full length apoC-II protein.

[00318] Additionally, Delta 6 variants Delta6PV, Delta6T18F-PV, and Delta6T18F-PV-AIB7,17 activated LPL *in vitro* with serum samples from apoC-II deficient patients (FIG. 52).

[00319] We generated an apoC-II mimetic peptide, Delta13-31-PV, which comprises a variant of helix 1 of apoC-II. Delta13-31-PV has the sequence of

DKVKEFLSEYWEKAKEFAPAVSTYTGIFTDQVLSVLKGEE (SEQ ID NO: 55), with the amino acid substitutions to helix 1 of apoC-II underlined. The Delta13-31-PV peptide promoted TG hydrolysis *in vitro* in a dose-dependent manner (FIG. 52).

[00320] These results show that variants of exemplary apoC-II mimetic peptide Delta6 and the apoC-II mimetic peptide comprising the variant of helix 1 of apoC-II are capable of promoting TG hydrolysis *in vitro*.

Example 6: Delta4 and its variants enhanced *in vivo* lipolysis

[00321] ApoC-II is a cofactor for lipoprotein lipase, an enzyme that hydrolyzes triglycerides. ApoC-II deficiency results in hypertriglyceridemia in human subjects. As described in Sakurai et al., *J Pharmacol Exp Ther* 356:341–353, 2016, we created apoC-II knockout mice that had high plasma TG.

[00322] ApoC-II mimetic peptide Delta4 was injected subcutaneously into apoC-II knockout mice and the plasma lipids were monitored over time (FIG. 16). Injection of Delta4 resulted in a rapid and marked reduction of plasma TG. The subcutaneous injection of Delta4 also led to a decrease in total cholesterol level (FIG. 17).

[00323] The apoC-II knockout mice were also injected with Delta4 peptide intraperitoneally (FIGs 18 and 19). Similarly, the Delta4 injection led to a marked reduction of plasma TG with a maximum decrease after about 3 hours. The intraperitoneal injection also led to a decrease in total cholesterol level.

[00324] Delta4 derivatives Delta4b and Delta4b-3 were also tested for their ability to promote TG hydrolysis *in vivo* (FIG. 20). After an oral gavage with vegetable oil, serum TG increased in C57BL/6 mice by about 4.5-fold by 3 h. When mice were intraperitoneally injected 30 min before gavage with an apoC-II mimetic peptide Delta4, Delta4b, or Delta4b-3, the TG increase was significantly reduced.

[00325] These results show that apoC-II mimetic peptide Delta4 and its variants have the ability to enhance *in vivo* lipolysis.

Example 7: Delta6 and its variants enhanced *in vivo* lipolysis

[00326] ApoC-II mimetic peptide Delta6 was injected intraperitoneally into apoC-II knockout mice and the plasma lipids were monitored over time (FIG. 21). Injection of Delta6 resulted in a rapid and marked reduction of plasma TG. The intraperitoneal injection of Delta6 also led to a decrease in the total cholesterol level (FIG. 22).

[00327] As shown in FIG. 23, three different doses of Delta6 peptide (0.25, 0.5, and 1 $\mu\text{mol/kg}$ of body weight) were injected intraperitoneally into apoC-II knockout mice and the plasma TG was monitored over time. All doses of Delta6 led to a rapid and marked reduction of plasma TG.

[00328] Variants of Delta6, Delta6b-A, Delta6b-P, and Delta6-NH were also tested for their ability to enhance *in vivo* lipolysis at the concentration of 0.25 $\mu\text{mol/kg}$ of body weight.

All Delta6 variants resulted in a significant reduction of plasma TG following their intraperitoneal injection in apoC-II knockout mice (FIG. 24).

[00329] As shown in FIG. 53, Delta6 variants Delta6PV and Delta6-T18F-PV were injected intraperitoneally into apoC-II knockout mice at 0.1 $\mu\text{mol/kg}$ of body weight and the plasma TG was monitored over time. Both Delta6PV and Delta6-T18F-PV led to a rapid reduction of plasma TG in the apoC-II knockout mice with a maximum reduction at about 6 hours after the peptide injection.

[00330] These results show that apoC-II mimetic peptide Delta6 and its variants have the ability to enhance *in vivo* lipolysis.

Example 8: ApoC-III displacement by ApoC-II mimetic peptides

[00331] ApoC-III is commonly associated with triglyceride rich lipoproteins and has been identified as a potent modulator of plasma TG levels. ApoC-III regulates plasma TG levels through a variety of mechanisms, including suppression of lipoprotein lipase (LPL) activity and LPL-independent effects on TG clearance. We investigated the effect of apoC-II mimetic peptides on displacement of apoC-III.

[00332] Incubation of VLDL and chylomicrons isolated from human plasma with different concentrations of apoC-II mimetic peptide Delta6 showed that Delta6 had a higher affinity for the VLDL and chylomicrons than the apolipoproteins (FIG. 25). Similarly, incubation of VLDL isolated from human plasma with Delta6 and its derivatives, Delta6-A, Delat6-P, and Delta6-NH, showed that Delta6 and the derivatives were capable of displacing apoC-III (FIG. 26).

[00333] As shown in FIG. 27, *in vitro* LPL assay was performed with hypertriglyceridemia patient sera. ApoC-II mimetic peptides Delta4 and Delta6 increased the LPL activity in patients with high levels of apoC-III protein.

[00334] These results show that apoC-II mimetic peptides Delta4, Delat6 and their variants are capable of displacing apoC-III and enhancing *in vitro* lipolysis in patient sera with high apoC-III protein.

Example 9: ApoC-III displacement by ApoC-II-a peptide

[00335] We investigated the effect of apoC-II-a peptide on displacement of apoC-III.

[00336] Incubation of VLDL isolated from human plasma with exogenous apoC-II-a peptide showed that apoC-II-a has a higher affinity for the VLDL than the apolipoproteins (FIG. 28). MALDI-TOF mass spectrometry analysis showed that apoC-II-a peptide selectively

replaced apoC-III and apoC-I, but not apoC-II (FIG. 29A). Interestingly, the binding of the apoC-II-a peptide to VLDL is almost proportional to its concentration (FIG. 29B).

[00337] Incubation of chylomicrons isolated from human plasma with exogenous apoC-II-a peptide showed that apoC-II-a has a higher affinity for chylomicrons than the apolipoproteins (FIG. 30).

[00338] LPL assay was performed using Intralipid as a substrate. As shown in FIG. 31, apoC-III inhibited the production of free fatty acid (NEFA). Addition of apoC-II-a peptide overcame the inhibitory effect of apoC-III and facilitated lipolysis in a dose-dependent manner.

[00339] LPL assay was also performed using high TG human plasma as a substrate in a 1:50 dilution. The results are shown on FIG. 32. When 50 μ M apoC-III recombinant protein was added, the free fatty acid (NEFA) production dropped from about 3.2 μ M to about 1.7 μ M. ApoC-II-a peptide was added in various concentrations. At a concentration of 0.01 μ M, it started to affect lipolysis. At a concentration of about 0.2 μ M, it overcame the inhibitory effect of apoC-III. Higher concentrations of apoC-II-a further facilitated lipolysis. Pre-incubation of apoC-III with human plasma did not change the effect of apoC-II-a peptide.

[00340] We also tested plasma from transgenic mice over-expressing human apoC-III. The apoC-III in the plasma suppressed lipolysis, consistent with the known ability of high concentrations of apoC-III to cause hypertriglyceridemia. In the LPL assay using the plasma from apoC-III transgenic mice, apoC-II-a peptide overcame the inhibitory effect of elevated expression of apoC-III in all three transgenic mice plasma samples (FIG. 33).

Example 10: ApoC-II-a blunts post-prandial hypertriglyceridemia in mice by mechanisms additional to lipoprotein lipase activation.

[00341] Post-prandial hypertriglyceridemia is an important cardiovascular disease (CVD) risk factor. We recently described an apoC-II mimetic peptide (apoC-II-a) that activates LPL and lowers serum TG in apoC-II-KO mice. To investigate apoC-II-a as a therapy for post-prandial hypertriglyceridemia, we investigated its effect on several mouse models after a fat challenge test.

[00342] After an oral gavage with vegetable oil (10 μ L/gram), serum TG increased by 3 h in C57BL/6 mice by at least 5-fold but when mice were injected 30 min before gavage with apoC-II-a (1 μ mol/kg; IP or SQ), the post-prandial TG rise was almost completely blunted. The post-prandial TG level in mice serum is shown in FIGs. 34A and 34B. Similar results were found in apoE-KO mice, indicating that the effect of apoC-II-a is independent of apoE

(not shown). ApoC-II-a did not appear to work at the level of TG absorption, because it also rapidly accelerated serum clearance of TG after IP injection of 20% Intralipid. Addition of exogenous LPL to serum samples collected after IP injection with Intralipid revealed that lipoprotein particles from mice treated with apoC-II-a were better substrates for LPL.

[00343] The rise in serum TG after IP injection of Intralipid was only partially blocked by about 50% when mice were co-injected with the LPL inhibitor Triton WR1339, indicating that apoC-II-a works additionally by an LPL-independent mechanism. FIGs. 34A and 34B show the comparison of the post-prandial TG in mice serum with or without the Triton co-injection. Incubation of human serum with exogenous apoC-II-a at doses comparable to that achieved *in vivo* in mice showed that apoC-II-a binds to all lipoproteins, including HDL and causes the displacement of apoC-I and apoC-III. Furthermore, the *in vitro* inhibition of LPL from the addition of exogenous apoC-III could be overcome by adding apoC-II-a at a molar ratio of at least 1:10 compared to apoC-III.

[00344] In summary, the apoC-II-a mimetic peptide at relatively low doses can accelerate the clearance of post-prandial TG after a fat load. It does so in part by activating LPL, as expected. Unexpectedly, however, we found that apoC-II-a also reduces triglyceride levels by relieving the inhibition of LPL by apoC-III, and possibly also by blocking the other known LPL-independent effects of apoC-III, and possibly apoC-I, in delaying the post-prandial clearance of TG.

Example 11: Delta6PV enhanced *in vivo* lipolysis in mouse models (single-dose studies)

[00345] The effect of the Delta6PV peptide on *in vivo* lipolysis was investigated in wild type, apoC-II knockout, and apoC-III transgenic mice with single-dose studies.

[00346] After an intraperitoneal injection of Intralipid, serum TG increased significantly in wild-type mice. Co-injection of LPL inhibitor Triton further increased the serum TG level to more than 1000-fold at the 6 h time point. When mice were intraperitoneally injected with the apoC-II mimetic peptide Delta6PV, the TG increase was significantly reduced (FIG. 35B). Delta6PV peptide reversed the inhibition of LPL by Triton.

[00347] The apoC-II mimetic peptide Delta6PV was injected intraperitoneally (FIG. 36B) or subcutaneously (FIG. 36C) into apoC-II knockout mice and the plasma TG level was monitored over time. Injection of Delta6PV resulted in a rapid and marked reduction of plasma TG.

[00348] After an intraperitoneal injection with Intralipid, serum TG increased in apoC-II knockout mice by about 7-fold 6 hours after the injection. When mice were intraperitoneally injected with the Delta6PV peptide 1 hour after the injection of Intralipid, the TG increase was significantly reduced (FIG. 37B).

[00349] The Delta6PV peptide was injected intraperitoneally into apoC-III transgenic mice and the plasma TG level was monitored over time. Injection of Delta6PV resulted in a rapid and marked reduction of plasma TG with a maximum decrease after about 6 hours after the injection (FIG. 38B).

[00350] After an intraperitoneal injection with Intralipid, serum TG increased in apoC-III transgenic mice by about 4-fold 6 hours after the injection. When mice were intraperitoneally injected with the Delta6PV peptide 1 hour after the injection of Intralipid, the TG increase was significantly reduced (FIG. 39).

[00351] The single-dose studies show that the apoC-II mimetic peptide Delta6PV has the ability to enhance *in vivo* lipolysis in mouse models.

Example 12: Delta6PV enhanced *in vivo* lipolysis in mouse models (repeat-dose studies)

[00352] Wild-type mice, apoC-II knockout mice, and apoC-III transgenic mice were injected with the apoC-II mimetic peptide Delta6PV multiple times by intraperitoneal injection.

[00353] The Delta6PV peptide was injected intraperitoneally into apoC-II knockout mice at 24 h intervals for 6 days and the plasma TG level was measured at 0 h, 3 h and 6 h on Day 1 and 0 h, 3 h, and 6 h on Day 6. Repeat injection of Delta6PV resulted in a marked and lasting reduction of plasma TG (FIG. 40). SEC-FPLC (size exclusion chromatography - fast protein liquid chromatography) were run on the pooled plasma samples of 0 h and 6 h on Day 1 and 0 h and 6 h on Day 6 from mice treated with Delta6PV peptide at 1 μ mole/kg (B. W.). As shown in FIG. 41, the level VLDL decreased 6 hours after injection of the Delta6PV peptide, while the level of HDL increased, as measured by the phospholipid level. The level of TG also decreased in the VLDL pool (FIG. 42).

[00354] The Delta6PV peptide was injected intraperitoneally into apoC-III transgenic mice at 24 h intervals for 5 days and the plasma lipids were monitored on Day 1 and Day 5 as indicated on FIG. 43A. Repeat injection of Delta6PV resulted in lower plasma TG 1 h, 3 h, and 6 h after the peptide injection on Day 1 and Day 5 as compared to the control conditions (FIG. 43B and FIG. 44). The plasma samples of 0 h and 3 h on Day 1 and Day 5 from mice

treated with 5 μ mole/kg (B. W.) Delta6PV peptide were applied to SEC-FPLC to separate VLDL and HDL populations. The level of apoC-III on VLDL and HDL were measured by ELISA. The apoC-III decreased on both VLDL and HDL in the Delta6PV-treated mice (FIG. 45A and FIG. 45B). This result indicates that Delta6PV reduced the apoC-III level on VLDL and HDL by displacement.

[00355] After an intraperitoneal injection with Intralipid, serum TG increased in wild-type mice by more than 40-fold by 3 hours after the injection. When mice were intraperitoneally injected with the Delta6PV peptide 1 hour after the injection of Intralipid, the TG increase was significantly reduced (FIG. 46B).

[00356] The repeat-dose studies show that the apoC-II mimetic peptide Delta6PV has the ability to enhance *in vivo* lipolysis in mouse models.

Example 13: Delta6PV enhanced *in vivo* lipolysis in the obese monkey model

[00357] ApoC-II mimetic peptide Delta6PV was injected intravenously into obese monkeys with a plasma TG level of greater than 2 mmol/L and the plasma lipids were monitored over time. A single injection of Delta6PV peptide led to a significant reduction of plasma TG level measured at 24 h, 48 h, and 72 h after the injection (FIG. 47). The effect of the Delta6PV peptide in the obese monkey model is dose-dependent.

[00358] The injection of Delta6PV peptide also led to a marked reduction of the VLDL level (FIG. 48A), although the levels of LDL and HDL did not change significantly (FIG. 48B and FIG. 48C).

[00359] In summary, the apoC-II mimetic peptide Delta6PV has the ability to enhance *in vivo* lipolysis and reduce the VLDL level the obese monkey model.

Example 14: ApoC-III displacement by Delta6PV peptide

[00360] We investigated the effect of Delta6PV peptide on displacement of apoC-III.

[00361] Incubation of VLDL isolated from human plasma with Delta6PV peptide showed that Delta6PV peptide has a higher affinity for the VLDL than the apolipoproteins. As shown in FIG. 49, Delta6PV peptide displaced apoC-III in a dose-dependent matter. The apoC-II in VLDL was also displaced by the Delta6PV peptide.

[00362] LPL assay was also performed using human apoC-II deficient plasma as a substrate. The results are shown on FIG. 50. When 25 μ M apoC-III recombinant protein was added, the free fatty acid (NEFA) production dropped from about 1.5 μ M to about 0.8 μ M.

Delta6 and Delta6PV peptides were added in various concentrations. Addition of Delta6 and Delta6PV peptides overcame the inhibitory effect of apoC-III and facilitated lipolysis.

[00363] These results show that the Delta6PV peptide is capable of displacing apoC-III and enhancing *in vitro* lipolysis.

[00364] While the invention has been particularly shown and described with reference to a preferred embodiment and various alternate embodiments, it will be understood by persons skilled in the relevant art that various changes in form and details can be made therein without departing from the spirit and scope of the invention.

[00365] All references, issued patents and patent applications cited within the body of the instant specification are hereby incorporated by reference in their entirety, for all purposes.

CLAIMS

1. An isolated apoC-II mimetic peptide of no more than 50 amino acids, comprising from N-terminus to C-terminus a first helical domain, a hinge region, and a second helical domain,
wherein the first helical domain is amphipathic, and wherein the apoC-II mimetic peptide is not a peptide of SEQ ID NO: 56.
2. The peptide of claim 1, wherein the first domain is native helix 1 of apoC-II.
3. The peptide of claim 1, wherein the first domain is a variant of helix 1 of apoC-II.
4. The peptide of claim 1, wherein the first domain is native helix 2 of apoC-II.
5. The peptide of claim 1, wherein the first domain is a variant of helix 2 of apoC-II.
6. The peptide of claim 5, wherein the variant of helix 2 of apoC-II comprises elongation of helix 2 of ApoC-II.
7. The peptide of claim 6, wherein the helix 2 of apoC-II is elongated by 1-10 amino acids at the N-terminus.
8. The peptide of claim 7, wherein the helix 2 of apoC-II is elongated by 1 amino acid at the N-terminus.
9. The peptide of claim 8, wherein the amino acid is aspartic acid.
10. The peptide of claim 7, wherein the helix 2 of apoC-II is elongated by 2 amino acids at the N-terminus.
11. The peptide of claim 10, wherein the amino acids are lysine and alanine from N-terminus to C-terminus.
12. The peptide of claim 7, wherein the helix 2 of apoC-II is elongated by 5 amino acids at the N-terminus.
13. The peptide of claim 12, wherein the helix 2 of apoC-II is elongated by the native upstream amino acid sequence of human apoC-II, or a mutant of native upstream amino acid sequence of human apoC-II.
14. The peptide of claim 12 or claim 13, wherein the variant of helix 2 of apoC-II comprises an aspartic acid at position 40.

- 15.** The peptide of any one of claims 12 to 14, wherein the variant of helix 2 of apoC-II comprises a tyrosine or a tryptophan at position 41.
- 16.** The peptide of any one of claims 12 to 15, wherein the variant of helix 2 of apoC-II comprises a leucine at position 42.
- 17.** The peptide of any one of claims 12 to 16, wherein the variant of helix 2 of apoC-II comprises a lysine or an arginine at position 43.
- 18.** The peptide of any one of claims 12 to 17, wherein the variant of helix 2 of apoC-II comprises an alanine or a glutamic acid at position 44.
- 19.** The peptide any one of claims 12 to 18, wherein the variant of helix 2 of apoC-II comprises an aspartic acid at position 40, a tyrosine at position 41, a leucine at position 42, a lysine at position 43, and a glutamic acid at position 44.
- 20.** The peptide of claim 5, wherein the variant of helix 2 of apoC-II comprises truncation of helix 2 of apoC-II.
- 21.** The peptide of claim 20, wherein the amino acid residues 45-46 of native helix 2 of apoC-II are deleted.
- 22.** The peptide of claim 20, wherein the amino acid residues 45-48 of native helix 2 of apoC-II are deleted.
- 23.** The peptide of claim 20, wherein the amino acid residues 45-49 of native helix 2 of apoC-II are deleted.
- 24.** The peptide of claim 20, wherein the amino acid residues 45-50 of native helix 2 of apoC-II are deleted.
- 25.** The peptide of claim 20, wherein the amino acid residues 45-53 of native helix 2 of apoC-II are deleted.
- 26.** The peptide of any one of claims 5 to 25, wherein the variant of helix 2 of apoC-II comprises at least one mutation of helix 2 of apoC-II.
- 27.** The peptide of claim 26, wherein the mutation is an amino acid substitution.
- 28.** The peptide of claim 27, wherein an original amino acid of helix 2 of apoC-II is substituted by a natural amino acid.

- 29.** The peptide of claim 27, wherein an original amino acid of helix 2 of apoC-II is substituted by an unnatural amino acid.
- 30.** The peptide of claim 27, wherein an original amino acid of helix 2 of apoC-II is substituted by an amino acid analog.
- 31.** The peptide of any one of claims 27 to 30, wherein the amino acid substitution is at position 45.
- 32.** The peptide of claim 31, wherein the valine at position 45 is substituted by phenylalanine.
- 33.** The peptide of any one of claims 27 to 30, wherein the amino acid substitution is at position 46.
- 34.** The peptide of claim 33, wherein the aspartic acid at position 46 is substituted by a phenylalanine.
- 35.** The peptide of claim 33, wherein the aspartic acid at position 46 is substituted by a lysine.
- 36.** The peptide of claim 33, wherein the aspartic acid at position 46 is substituted by an aminoisobutyric acid.
- 37.** The peptide of any one of claims 27 to 30, wherein the amino acid substitution is at position 48.
- 38.** The peptide of claim 37, wherein the lysine at position 48 is substituted by an arginine.
- 39.** The peptide of any one of claims 27 to 30, wherein the amino acid substitution is at position 49.
- 40.** The peptide of claim 39, wherein the leucine at position 49 is substituted by a lysine.
- 41.** The peptide of any one of claims 27 to 30, wherein the amino acid substitution is at position 50.
- 42.** The peptide of claim 41, wherein the arginine at position 50 is substituted by a lysine.
- 43.** The peptide of any one of claims 27 to 30, wherein the amino acid substitution is at position 53.
- 44.** The peptide of claim 43, wherein the tyrosine at position 53 is substituted by a leucine.
- 45.** The peptide of any one of claims 27 to 30, wherein the amino acid substitution is at position 54.

- 46.** The peptide of claim 45, wherein the serine at position 54 is substituted by a glutamic acid.
- 47.** The peptide of claim 45, wherein the serine at position 54 is substituted by a lysine.
- 48.** The peptide of claim 45, wherein the serine at position 54 is substituted by an aspartic acid.
- 49.** The peptide of any one of claims 27 to 30, wherein the amino acid substitution is at position 55.
- 50.** The peptide of claim 49, wherein the lysine at position 55 is substituted by an arginine.
- 51.** The peptide of any one of claims 27 to 30, wherein the amino acid substitution is at position 56.
- 52.** The peptide of claim 51, wherein the serine at position 56 is substituted by a phenylalanine.
- 53.** The peptide of claim 51, wherein the serine at position 56 is substituted by a lysine.
- 54.** The peptide of claim 51, wherein the serine at position 56 is substituted by an alanine.
- 55.** The peptide of claim 51, wherein the serine at position 56 is substituted by an aminoisobutyric acid.
- 56.** The peptide of any one of claims 27 to 30, wherein the amino acid substitution is at position 57.
- 57.** The peptide of claim 56, wherein the threonine at position 57 is substituted by a phenylalanine.
- 58.** The peptide of any one of claims 27 to 30, wherein the variant of helix 2 of apoC-II comprises three amino acid substitutions, wherein the amino acid substitutions are at position 46, position 54, and position 56.
- 59.** The peptide of claim 58, wherein the aspartic acid at position 46 is substituted by a phenylalanine, the serine at position 54 is substituted by a glutamic acid, and the serine at position 56 is substituted by a phenylalanine.
- 60.** The peptide of any one of claims 5 to 59, wherein the variant of helix 2 of apoC-II comprises at least one chemical modification.

61. The peptide of claim 60, wherein the chemical modification is a covalent linkage of a fatty acid.
62. The peptide of claim 61, wherein the fatty acid comprises 4 to 26 carbons.
63. The peptide of claim 62, wherein the fatty acid comprises 10 carbons.
64. The peptide of claim 62, wherein the fatty acid comprises 12 carbons.
65. The peptide of claim 62, wherein the fatty acid comprises 14 carbons.
66. The peptide of claim 62, wherein the fatty acid comprises 16 carbons.
67. The peptide of claim 62, wherein the fatty acid comprises 18 carbons.
68. The peptide of claim 62, wherein the fatty acid comprises 20 carbons.
69. The peptide of any one of claims 61 to 68, wherein the fatty acid is saturated.
70. The peptide of any one of claims 61 to 68, wherein the fatty acid is unsaturated.
71. The peptide of any one of claims 61 to 70, wherein the fatty acid is linked to the N-terminal amino acid.
72. The peptide of any one of claims 61 to 71, wherein the fatty acid is linked to an amino acid side group.
73. The peptide of claim 72, wherein the fatty acid is linked to the ϵ -amine of a lysine residue.
74. The peptide of any of the preceding claims, wherein the hinge region comprises 5-10 amino acids.
75. The peptide of claim 74, wherein the hinge region comprises 7 amino acids.
76. The peptide of claim 74, wherein the hinge region comprises a proline.
77. The peptide of claim 76, wherein the hinge region comprises a proline at position 58.
78. The peptide of claim 74, wherein the hinge region comprises an alanine.
79. The peptide of claim 78, wherein the hinge region comprises an alanine at position 58.
80. The peptide of claim 74, wherein the hinge region comprises a norleucine.
81. The peptide of claim 80, wherein the hinge region comprises a norleucine at position 60.
82. The peptide of claim 74, wherein the hinge region comprises a lysine.

- 83.** The peptide of claim 82, wherein the hinge region comprises a lysine at position 60.
- 84.** The peptide of claim 74, wherein the hinge region comprises a valine.
- 85.** The peptide of claim 84, wherein the hinge region comprises a valine at position 60.
- 86.** The peptide of claim 74, wherein the hinge region comprises a leucine.
- 87.** The peptide of claim 86, wherein the hinge region comprises a leucine at position 60.
- 88.** The peptide of claim 74, wherein the hinge region comprises a methionine.
- 89.** The peptide of claim 88, wherein the hinge region comprises a methionine at position 60.
- 90.** The peptide of any one of claims 74 to 89, wherein the hinge region comprises at least one chemical modification.
- 91.** The peptide claim 90, wherein the chemical modification is a covalent linkage of a fatty acid.
- 92.** The peptide claim 91, wherein the fatty acid comprises 4 to 26 carbons.
- 93.** The peptide claim 92, wherein the fatty acid comprises 10 carbons.
- 94.** The peptide claim 92, wherein the fatty acid comprises 12 carbons.
- 95.** The peptide claim 92, wherein the fatty acid comprises 14 carbons.
- 96.** The peptide claim 92, wherein the fatty acid comprises 16 carbons.
- 97.** The peptide claim 92, wherein the fatty acid comprises 18 carbons.
- 98.** The peptide claim 92, wherein the fatty acid comprises 20 carbons.
- 99.** The peptide of any one of claims 91 to 98, wherein the fatty acid is saturated.
- 100.** The peptide of any one of claims 91 to 98, wherein the fatty acid is unsaturated.
- 101.** The peptide of any one of claims 91 to 100, wherein the fatty acid is linked to an amino acid side group.
- 102.** The peptide of claim 101, wherein the fatty acid is linked to the ϵ -amine of a lysine residue.
- 103.** The peptide of any of the preceding claims, wherein the hinge region allows the first helical domain and the second helical domain to retain a nearly straight conformation,

wherein the second domain bends away from the first domain over an angle of no more than about 20°.

104. The peptide of any of the preceding claims, wherein the second domain is native helix 3 of apoC-II.

105. The peptide of any one of claims 1 to 103, wherein the second domain is a variant of helix 3 of apoC-II.

106. The peptide of claim 105, wherein the variant of helix 3 of apoC-II comprises elongation of helix 3 of apoC-II.

107. The peptide of claim 106, wherein the helix 3 of apoC-II is elongated by 1-10 amino acids at the C-terminus.

108. The peptide of claim 107, wherein the helix 3 of apoC-II is elongated by 4 amino acids at the C-terminus.

109. The peptide of claim 108, wherein the helix 3 of apoC-II is elongated by amino acids lysine, glycine, glutamic acid, and glutamic acid from N-terminus to C-terminus.

110. The peptide of any one of claims 105 to 109, wherein the variant of helix 3 of apoC-II comprises at least one mutation of helix 3 of apoC-II.

111. The peptide of claim 110, wherein the mutation is an amino acid substitution.

112. The peptide of claim 111, wherein an original amino acid of helix 3 of apoC-II is substituted by a natural amino acid.

113. The peptide of claim 111, wherein an original amino acid of helix 3 of apoC-II is substituted by an unnatural amino acid.

114. The peptide of claim 111, wherein an original amino acid of helix 3 of apoC-II is substituted by an amino acid analog.

115. The peptide of any one of claims 111 to 114, wherein the amino acid substitution is at position 70.

116. The peptide of claim 115, wherein the glutamine at position 70 is substituted by an arginine.

- 117.** The peptide of claim 115, wherein the glutamine at position 70 is substituted by a lysine.
- 118.** The peptide of any one of claims 105 to 117, wherein the variant of helix 3 of apoC-II comprises at least one chemical modification.
- 119.** The peptide of claim 118, wherein the chemical modification is a covalent linkage of a fatty acid.
- 120.** The peptide of claim 119, wherein the fatty acid comprises 4 to 26 carbons.
- 121.** The peptide of claim 120, wherein the fatty acid comprises 10 carbons.
- 122.** The peptide of claim 120, wherein the fatty acid comprises 12 carbons.
- 123.** The peptide of claim 120, wherein the fatty acid comprises 14 carbons.
- 124.** The peptide of claim 120, wherein the fatty acid comprises 16 carbons.
- 125.** The peptide of claim 120, wherein the fatty acid comprises 18 carbons.
- 126.** The peptide of claim 120, wherein the fatty acid comprises 20 carbons.
- 127.** The peptide of any one of claims 119 to 126, wherein the fatty acid is saturated.
- 128.** The peptide of any one of claims 119 to 126, wherein the fatty acid is unsaturated.
- 129.** The peptide of any one of claims 119 to 128, wherein the fatty acid is linked to the C-terminal amino acid.
- 130.** The peptide of any one of claims 119 to 129, wherein the fatty acid is linked to an amino acid side group.
- 131.** The peptide of claim 130, wherein the fatty acid is linked to the ϵ -amine of a lysine residue.
- 132.** The peptide of any one of claims 119 to 131, wherein the C-terminal amino acid is modified by a C-terminal amide.
- 133.** The peptide of any of the preceding claims, wherein the isolated apoC-II mimetic peptide further comprises a purification tag.
- 134.** The peptide of claim 133, wherein the purification tag is a polyhistidine-tag, a myc-tag, or an HA-tag.

- 135.** The peptide of any of the preceding claims, wherein the first domain has an affinity for binding to lipoproteins.
- 136.** The peptide of any of the preceding claims, wherein the second domain is capable of activating lipoprotein lipase (LPL).
- 137.** The peptide of claim 135 or claim 136, wherein the peptide is capable of activating lipolysis by LPL.
- 138.** The peptide of claim 135 or claim 136, wherein the peptide is capable of displacing apoC-III in lipoproteins.
- 139.** The peptide of any of the preceding claims, wherein the peptide is capable of reducing triglyceride (TG) level *in vivo*.
- 140.** The peptide of claim 139, wherein the peptide is capable of reducing elevated TG level caused by LPL deficiency.
- 141.** The peptide of claim 139, wherein the peptide is capable of reducing elevated TG level caused by elevated apoC-III.
- 142.** The peptide of claim 139, wherein the peptide is capable of reducing elevated TG level caused by apoC-II deficiency.
- 143.** The peptide of claim 139, wherein the peptide is capable of reducing post-prandial elevated TG level.
- 144.** The isolated apoC-II mimetic peptide of claim 1, wherein the peptide has the sequence set forth in SEQ ID NO: 6.
- 145.** The isolated apoC-II mimetic peptide of claim 1, wherein the peptide has the sequence set forth in SEQ ID NO: 9.
- 146.** An isolated apoC-II mimetic peptide of no more than 30 amino acids, consisting of a variant of helix 3 of apoC-II.
- 147.** The peptide of claim 146, wherein the variant of helix 3 of apoC-II comprises elongation of helix 3 of apoC-II.
- 148.** The peptide of claim 147, wherein the helix 3 of apoC-II is elongated by 1-10 amino acids at the N-terminus.

- 149.** The peptide of claim 148, wherein the helix 3 of apoC-II is elongated by 6 amino acids at the N-terminus.
- 150.** The peptide of claim 149, wherein the variant of helix 3 of apoC-II comprises a lysine or methionine at position 60.
- 151.** The peptide of anyone of claims 147 to 150, wherein the helix 3 of apoC-II is elongated by 1-10 amino acids at the C-terminus.
- 152.** The peptide of claim 151, wherein the helix 3 of apoC-II is elongated by 4 amino acids at the C-terminus.
- 153.** The peptide of claim 152, wherein the helix 3 of apoC-II is elongated by amino acids lysine, glycine, glutamic acid, and glutamic acid from N-terminus to C-terminus.
- 154.** The peptide of any one of claims 146 to 153, wherein the variant of helix 3 of apoC-II comprises at least one mutation of helix 3 of apoC-II.
- 155.** The peptide of claim 154, wherein the mutation is an amino acid substitution.
- 156.** The peptide of claim 155, wherein an original amino acid of helix 3 of apoC-II is substituted by a natural amino acid.
- 157.** The peptide of claim 155, wherein an original amino acid of helix 3 of apoC-II is substituted by an unnatural amino acid.
- 158.** The peptide of claim 155, wherein an original amino acid of helix 3 of apoC-II is substituted by an amino acid analog.
- 159.** The peptide of any one of claims 146 to 158, wherein the variant of helix 3 of apoC-II comprises at least one chemical modification.
- 160.** The peptide of claim 159, wherein the chemical modification is a covalent linkage of a fatty acid.
- 161.** The peptide of claim 160, wherein the fatty acid comprises 4 to 26 carbons.
- 162.** The peptide of claim 161, wherein the fatty acid comprises 10 carbons.
- 163.** The peptide of claim 161, wherein the fatty acid comprises 12 carbons.
- 164.** The peptide of claim 161, wherein the fatty acid comprises 14 carbons.
- 165.** The peptide of claim 161, wherein the fatty acid comprises 16 carbons.

- 166.** The peptide of claim 161, wherein the fatty acid comprises 18 carbons.
- 167.** The peptide of claim 161, wherein the fatty acid comprises 20 carbons.
- 168.** The peptide of any one of claims 160 to 167, wherein the fatty acid is saturated.
- 169.** The peptide of any one of claims 160 to 167, wherein the fatty acid is unsaturated.
- 170.** The peptide of any one of claims 160 to 169, wherein the fatty acid is linked to the N-terminal amino acid.
- 171.** The peptide of any one of claims 160 to 169, wherein the fatty acid is linked to the C-terminal amino acid.
- 172.** The peptide of any one of claims 160 to 171, wherein the fatty acid is linked to an amino acid side group.
- 173.** The peptide of claim 172, wherein the fatty acid is linked to the ϵ -amine of a lysine residue.
- 174.** The peptide of any one of claims 146 to 173, wherein the isolated apoC-II mimetic peptide further comprises a purification tag.
- 175.** The peptide of claim 174, wherein the purification tag is a polyhistidine-tag, a myc-tag, or an HA-tag.
- 176.** The peptide of any one of claims 146 to 175, wherein the variant of helix 3 of apoC-II has an affinity for binding to lipoproteins.
- 177.** The peptide of any one of claims 146 to 176, wherein the variant of helix 3 of apoC-II is capable of activating lipoprotein lipase (LPL).
- 178.** The peptide of claim 176 or claim 177, wherein the peptide is capable of activating lipolysis by LPL.
- 179.** The peptide of claim 176 or claim 177, wherein the peptide is capable of displacing apoC-III in lipoproteins.
- 180.** The peptide of any one of claims 146 to 179, wherein the peptide is capable of reducing triglyceride (TG) level *in vivo*.
- 181.** The peptide of claim 180, wherein the peptide is capable of reducing elevated TG level caused by LPL deficiency.

- 182.** The peptide of claim 180, wherein the peptide is capable of reducing elevated TG level caused by elevated apoC-III.
- 183.** The peptide of claim 180, wherein the peptide is capable of reducing elevated TG level caused by apoC-II deficiency.
- 184.** The peptide of claim 180, wherein the peptide is capable of reducing post-prandial elevated TG level.
- 185.** A pharmaceutical composition comprising the peptide of any of the preceding claims and a pharmaceutically acceptable carrier.
- 186.** The pharmaceutical composition of claim 185, wherein the pharmaceutical composition is suitable for subcutaneous injection.
- 187.** A method of treating hypertriglyceridemia in a patient, comprising administering to the patient an effective amount of the peptide or the pharmaceutical composition of any of the preceding claims.
- 188.** The method of claim 187, wherein the hypertriglyceridemia is associated with obesity.
- 189.** The method of claim 187, wherein the hypertriglyceridemia is associated with diabetes mellitus.
- 190.** The method of claim 187, wherein the hypertriglyceridemia is associated with alcohol consumption.
- 191.** The method of claim 187, wherein the hypertriglyceridemia is associated with medication.
- 192.** The method of claim 187, wherein the hypertriglyceridemia is caused by LPL deficiency.
- 193.** The method of claim 192, wherein the hypertriglyceridemia is familial lipoprotein lipase deficiency.
- 194.** The method of claim 192 or 193, wherein the LPL deficiency is caused by a mutation the LPL gene.
- 195.** The method of claim 194, wherein the mutation leads to reduced LPL enzyme activity.
- 196.** The method of claim 194, wherein the mutation leads to absent LPL enzyme activity.

- 197.** The method of any one of claims 192 to 196, wherein the LPL deficiency is diagnosed by the absence of LPL activity in serum of the patient.
- 198.** The method of any one of claims 194 to 196, wherein the mutation is detected by DNA sequence analysis.
- 199.** The method of claim 187, wherein the hypertriglyceridemia is caused by apoC-II deficiency.
- 200.** The method of claim 187, wherein the hypertriglyceridemia is caused by elevated apoC-III.
- 201.** The method of any one of claims 187 to 200, wherein the pre-treatment serum triglyceride (TG) concentration of the patient is between 150 mg/dL and 199 mg/dL.
- 202.** The method of any one of claims 187 to 200, wherein the pre-treatment serum triglyceride (TG) concentration of the patient is between 200 mg/dL to 499 mg/dL.
- 203.** The method of any one of claims 187 to 200, wherein the pre-treatment serum triglyceride (TG) concentration of the patient is between 500 mg/dL to 999 mg/dL.
- 204.** The method of any one of claims 187 to 200, wherein the pre-treatment serum triglyceride (TG) concentration of the patient is between 1000 mg/dL and 1999 mg/dL.
- 205.** The method of any one of claims 187 to 200, wherein the pre-treatment serum triglyceride (TG) concentration of the patient is equal to or higher than 2000 mg/dL.
- 206.** The method of any one of claims 187 to 205, wherein the patient has developed or is at risk for acute pancreatitis.
- 207.** The method of any one of claims 187 to 206, wherein the patient has developed or is at risk for acute cardiovascular disease.
- 208.** A method of making the peptide of any one of claims 1 to 184, comprising producing the peptide recombinantly.
- 209.** A method of making the peptide of any one of claims 1 to 184, comprising producing the peptide by chemical synthesis.