ORAL CARE COMPOSITION FOR ALLEVIATING DENTIN HYPERESTHESIA

BACKGROUND OF THE INVENTION

1. Field of the Invention

The present invention relates to a composition for oral care, and more specifically, the present invention relates to oral care composition for preventing or alleviating dentin hyperesthesia by inducing physiological remineralization of defects in dentin constituting a tooth.

2. Description of the Related Art

Dentin hyperesthesia, commonly referred to as 'sensitive dentin', is a common symptom experienced by 8% to 57% of the adult population. In particular, the case of periodontal disease, which is the most common disease in Korea, 72.5% to 98% of patients suffer from 'sensitive dentin' (Source: National Health Information Portal Medical Information; http://health.cdc.go.kr/health/Main.do).

Dentin hyperesthesia can be defined as a pain caused by the exposure of dentinal tubules transmitting all external stimuli to the pulp nerve. This makes the pulp nerve to be more sensitive to the same stimulus. Although there are no nerves in the dentin itself, but we can perceive the changes because the cold temperature stimulus is transmitted through the dentinal tubules to the nerves inside the pulp.

In the dentin, which occupies the most part of the tooth, there are dentinal tubules that extend from the pulp to the enamel. These tubules are filled with liquid. The diameter increases toward the pulp and has a dense structure. Because of these distinct structures, external stimuli can be transmitted quickly to the pulp nerve. If the dentin surface is damaged and the number of exposed dentinal tubules increases, it may cause a more sensitive reaction to the same stimulus than usual.

Currently, there are two ways for dentin hyperesthesia, depending on the principle of action. One is to interfere with the signal transmission of the nerve that transmits pain, and the other is to block the exposed dentinal tubules to alleviate the symptoms.

Dipotassium phosphate (K_2HPO_4) has been widely used in a method for interfering with the signal transmission of nerves that transmit pain. However, dipotassium phosphate has a low pain-blocking effect and must be used repeatedly, and it is not an effective treatment method because it limits the sense of chewing.

Next, calcium hydrogen phosphate ($CaHPO_4$), fluorine, oxalate, arginine (amino acid), and calcium carbonate ($CaCO_3$) are used to block the dentinal tubules. However, since sealing off the dentinal tubule also uses a foreign material, different from the dentin, there would be a gap created in the peripheral

boundary area between dentin and the foreign materials. The nerve would be exposed again after the foreign material being separated from the sealing, and it would cause a recurrence of sensitive dentin.

Currently, there are many products containing fluoride for oral cleansers to alleviate dentin hyperesthesia (dentin hypersensitivity or sensitive teeth). Because fluoride coats teeth and has strong binding power with calcium components present in saliva to seal off the exposed dentinal tubules to relieve aching symptoms. However, fluoride is known to have side effects that can damage the human immune system or cause arthritis, backache, and osteoporosis when used for a long time.

SUMMARY OF THE INVENTION

Embodiments of the present inventive concepts may provide oral care composition for alleviating dentin hyperesthesia, comprising a peptide including an amino acid sequence of the following Formula 1:

K-Y-R1-R2-R3-R4-R5-R6-R7-R8 (Formula 1)

wherein R1 is arginine(R), lysine(K) or glutamine(Q);

R2 is arginine(R) or glutamine(Q);

R3, R4, and R5 are arginine(R) or lysine(K), respectively;

R6 is asparagine(N) or serine(S); and

R7 and R8 are lysine(K) or tyrosine(Y), respectively.

Embodiments of the present inventive concepts may also provide a composition for oral care comprising a peptide including any one amino acid sequence of SEQ ID NOS: 1 to 96.

Embodiments of the present inventive concepts, 0.00005-0.00015 parts by weight of the peptide may be included based on 100 parts by weight of the composition.

Embodiments of the present inventive concepts, 0.0545-0.0555 parts by weight of cetylpyridinium chloride may be included based on 100 parts by weight of the composition.

Embodiments of the present inventive concepts, the composition for oral care may include 85-87 parts by weight of purified water, 1.7-2.9 parts by weight of surfactant, and 0.0045-0.0055 parts by weight of citric acid hydrate based on 100 parts by weight of the composition.

Embodiments of the present inventive concepts, the surfactant may be poloxamer and/or polysorbate 20.

Embodiments of the present inventive concepts, the surfactant may include 12-14% by weight of the poloxamer 407 and 86-88% by weight of the polysorbate 20.

Embodiments of the present inventive concepts, the oral care composition may include 9-11 parts by weight of a humectant based on 100 parts by weight of the composition.

Embodiments of the present inventive concepts, the humectant may be D-sorbitol solution and/or concentrated

glycerin.

Embodiments of the present inventive concepts, the humectant may include 45-55% by weight of the D-sorbitol solution and 45-55% by weight of the concentrated glycerin.

Effect of the invention

The present inventive concepts may provide oral care composition that prevents or alleviates dentin hyperesthesia by sealing off exposed dentinal tubule defects through physiological remineralization of dentin.

Other aspects, advantages, and salient features of the embodiments will become apparent to those skilled in the art from the following detailed description, which, taken in conjunction with the annexed drawings, discloses exemplary embodiments of the disclosure.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1A is a graph showing the results of comparing the effect of the respective groups of the peptide included in the oral care composition for alleviating dentin hyperesthesia according to an embodiment of the present invention on the expression of DSPP, which is an odontoblast differentiation marker gene.

FIG. 1B is a graph showing the results of comparing the

expression levels of the odontoblast differentiation marker Dspp gene in MDPC-23 cells treated with the peptides included in the oral care composition for alleviating dentin hyperesthesia according to an embodiment of the present invention.

FIG. 1C is a graph showing the results of comparing expression levels of odontoblast differentiation marker genes, Dspp, Dmp1, and Nestin in MDPC-23 cells treated with peptides of Group 11 and Group 12 included in the oral care composition for alleviating dentin hyperesthesia according to an embodiment of the present invention.

FIG. 1D is a graph showing the results of evaluating cytotoxicity of the peptides included in the oral care composition for alleviating dentin hyperesthesia according to an embodiment of the present invention.

FIG. 2 shows the permeability of oral care composition for alleviating dentin hyperesthesia according to an embodiment of the present invention. A fluorescence dyeing reagent (Rhodamine B) was mixed and treated for 1 minute on the tooth exposed to the dentinal tubules and then observed with a fluorescence microscope.

FIG. 3 shows the results of comparing the sealing ability of the dentinal tubule with the oral care composition for alleviating dentin hyperesthesia according to the embodiment of the present invention(example 2), comparative test example 2-1,

and comparative test example 2-2.

A-C shows the dentinal tubules treated only with purified water (comparative test example 2-1), D-F is composition without the peptide comparing with the embodiment of the present invention (comparative test example 2-2), and G-I shows the dentinal tubules treated with the oral care composition for alleviating dentin hyperesthesia according to the embodiment of the present invention (scale bar: A, D, G, 100 μm; B, E, H, 20 μm; C, F, I, 10 μm). In each case, the dentin slices exposed the dentinal tubules were treated once a day for 1 minute and then immersed in artificial saliva. And after repeating this process for 2 weeks, the ability to seal off the dentin tubes was observed with a scanning electron microscope.

FIG. 4 is an enlarged picture of which the dentinal tubules are sealed off by treating the oral care composition for alleviating dentin hyperesthesia according to an embodiment of the present invention. It shows remineralization in the sealed off dentinal tubules and dentin surfaces.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

Reference will now be made in detail to embodiments, examples of which are illustrated in the accompanying drawings, wherein like reference numerals refer to like elements throughout. In this regard, the present embodiments may have

different forms and should not be construed as being limited to the descriptions set forth herein. Unless otherwise defined, all terms including technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skills in the art to which this disclosure belongs. It will be further understood that terms, such as those defined in commonly used dictionaries, should be interpreted as having a meaning that is consistent with their meaning in the context of the relevant art and will not be interpreted in an idealized or overly formal sense unless expressly so defined herein. In addition, terms to be described later are defined in consideration of contributions in the present disclosures, which may vary according to the intention of the user or practice.

The disclosure now will be described more fully hereinafter with reference to the accompanying drawings, in which illustrative embodiments of the disclosure are shown. This disclosure may, however, be embodied in many different forms and should not be construed as limited to the embodiments set forth herein; rather, these embodiments are provided so that this disclosure will be thorough and complete, and will fully convey the scope of the disclosure to those skilled in the art. However, as it is presented as an example, the present invention is not limited thereto and the present invention is defined only by the scope of the claims which will be described later.

It will be understood that the terms "comprises" and/or "comprising," when used in this specification, specify the presence of stated components, this means that it may contain more components, rather than exclude other components, unless there is a particularly contrary article.

Hereinafter, embodiments of the present invention are described in more detail.

An odontoblast may refer to a cell that synthesizes and secretes proteins and polysaccharides composing the matrix of the dentin. It is a columnar cell that is in contact with the predentin (uncalcified dentin) and forms a cell layer at the periphery of the dental pulp. And, it is a differentiated cell(becoming a cell derived from the mesenchymal ectoderm) involved in the calcification of dentin. At the developmental stage, an odontoblast faces the enamel among the cells of the dental papilla.

A peptide, included in the oral care composition for alleviating dentin hyperesthesia according to an embodiment of the present invention (hereinafter, 'odontoblast differentiation promoting peptide'), does not exhibit cytotoxicity, but it is possible to increase the expression level of the odontoblast differentiation marker genes DSPP, Dmp1 and Nestin. When transplanted *in vivo* with pulp tissue cells, the pulp tissue cells may form a dentin/dentin-like tissue.

Odontoblast differentiation promoting peptide includes mutant peptides having a sequence different from the amino acid sequence constituting the amino acid sequence and at least one amino acid residue, as long as it can promote dentin regeneration or treat dentin hyperesthesia.

Amino acid exchanges in proteins and polypeptides, which do not generally alter the molecular activity, are known in the art. The most commonly occurring exchanges are amino acid residues Ala/Ser, Val/Ile, Asp/Glu, Thr/Ser, Ala/Gly, Ala/Thr, Ser/Asn, Ala/Val, Ser/Gly, Thy/Phe, Ala/Pro, Lys/Arg, Asp/Asn, Leu/Ile, Leu/Val, Ala/Glu, Asp/Gly, in both directions. The peptide may include peptides that have improved structural stability against heat, pH, etc., or improved ability to promote regeneration of dentin or dental pulp due to alteration or modification of the amino acid sequence.

For example, although glutamine which is an acidic amino acid at position 3 of the peptide of SEQ ID NO: 1 of the present invention is substituted with a basic amino acid, lysine or arginine, the effects of the peptide of the present invention may be obtained as it is; although arginine which is a basic amino acid at position 4 or 5 of the peptide of SEQ ID NO: 1 is substituted with an acidic amino acid glutamine or a basic amino acid lysine, the effects of the peptide of the present invention may be obtained as it is; although lysine which is a basic amino

acid at position 6, 7, or 9 of the peptide of SEQ ID NO: 1 is substituted with a basic amino acid arginine or an aromatic amino acid tyrosine, the effects of the peptide of the present invention may be obtained as it is; although asparagine which is an acidic amino acid at position 8 of the peptide of SEQ ID NO: 1 is substituted with a neutral amino acid serine, the effects of the peptide of the present invention may be obtained as it is; and although tyrosine which is an aromatic amino acid at position 10 of the peptide of SEQ ID NO: 1 is substituted with a basic amino acid lysine, the effects of the peptide of the present invention may be obtained as it is.

As such, although the acidic amino acids, basic amino acids, or aromatic amino acids constituting the peptide of the present invention are substituted with amino acids having the same properties, or substituted with different acidic amino acids, basic amino acids, neutral amino acids, or aromatic amino acids, respectively, the effects of the peptide of the present invention may be obtained as it is. Therefore, it is apparent that a peptide variant having a sequence including one or more amino acid residues different from those of the amino acid sequence constituting the peptide of the present invention is also included in the scope of the peptide of the present invention.

Further, although arbitrary amino acids are added at the N-

terminus or C-terminus of the peptide of the prevention, the effects of the peptide of the present invention may be obtained as it is. Therefore, a peptide prepared by adding arbitrary amino acids at the N-terminus or C-terminus of the peptide of the present invention is also included in the scope of the peptide of the present invention. For example, a peptide prepared by adding 1 to 300 amino acids at the N-terminus or C-terminus of the peptide of the present invention may be exemplified, for another example, a peptide prepared by adding 1 to 100 amino acids at the N-terminus or C-terminus of the peptide of the present invention may be exemplified, and for still another example, a peptide prepared by adding 1 to 24 amino acids at the N-terminus or C-terminus of the peptide of the present invention may be exemplified.

The mRNA of the DSPP gene in MDPC-23 cells treated with the odontoblast differentiation promoting peptide, compared to the mRNA level of the DSPP gene in MDPC-23 cells (control) not treated with the odontoblast differentiation promoting peptide, was all 1.3 times or more (Tables 13 to 24).

As reported up to now, it is known that as the mRNA level of DSPP is increased, odontoblast differentiation and dentin regeneration are promoted, and therefore, it can be seen that 128 kinds of peptides increases the mRNA level of Dspp gene, which in turn may exhibit the effect of promoting odontoblast

differentiation and dentin regeneration (Taduru Sreenath et al., THE JOURNAL OF BIOLOGICAL CHEMISTRY, Vol. 278, No. 27, Issue of July 4, pp. 24874-24880, 2003; William T. Butler et al., Connective Tissue Research, 44(Suppl. 1): 171-178, 2003).

The peptide included in the oral care composition for alleviating dentin hyperesthesia may be used in a single form of the peptide or in a polypeptide form of 2 or more repeats of the peptide, and the peptide may also be used in a complex form of a drug having a therapeutic effect on dentin or dental pulp diseases linked at the N-terminus or C-terminus of the peptide.

Example 1:

Synthesis of Peptides for Promoting Odontoblast Differentiation

The present inventors synthesized a peptide (SEQ ID NO: 1) showing the effect of promoting regeneration of dentin or dental pulp tissues by a 9-fluorenylmethyloxycarbonyl (Fmoc) method, and they synthesized peptides of respective groups (Tables 1 to 12) by substituting the amino acids of the synthesized peptide.

N-KYQRRKKNKY-C (SEQ ID NO: 1)

First, peptides of Group 1 were synthesized by using the peptide of SEQ ID NO: 1 or by substituting any amino acid at positions 5 to 7 of the peptide of SEQ ID NO: 1 with lysine or

arginine (Table 1).

[Table 1]

Peptides of Group 1

SEQ ID NO:	Amino acid sequence(N-C)
1	KYQRRKKNKY
2	KYQRRKRNKY
3	KYQRRRKNKY
4	KYQRRRRNKY
5	KYQRKKKNKY
6	KYQRKRKNKY
7	KYQRKKRNKY
8	KYQRKRRNKY

Next, peptides of Group 2 were synthesized by substituting any amino acid at positions 5 to 7 of the peptide of SEQ ID NO: 1 with lysine or arginine or by substituting an amino acid at position 8 of the peptide of SEQ ID NO: 1 with serine (Table 2).

[Table 2]

Peptides of Group 2

SEQ ID NO:	Amino acid sequence(N-C)

9	KYQRRKKSKY
10	KYQRRKRSKY
11	KYQRRRKSKY
12	KYQRRRRSKY
13	KYQRKKKSKY
14	KYQRKRKSKY
15	KYQRKKRSKY
16	KYQRKRRSKY

Next, peptides of Group 3 were synthesized by substituting any amino acid at positions 5 to 7 of the peptide of SEQ ID NO: 1 with lysine or arginine or by substituting an amino acid at position 9 of the peptide of SEQ ID NO: 1 with tyrosine (Table 3).

[Table 3]

Peptides of Group 3

SEQ ID NO:	Amino acid sequence(N-C)
17	KYQRRKKNYK
18	KYQRRKRNYK
19	KYQRRRKNYK
20	KYQRRRRNYK

21	KYQRKKKNYK
22	KYQRKRKNYK
23	KYQRKKRNYK
24	KYQRKRRNYK

Next, peptides of Group 4 were synthesized by substituting any amino acid at positions 5 to 7 of the peptide of SEQ ID NO: 1 with lysine or arginine, by substituting an amino acid at position 8 of the peptide of SEQ ID NO: 1 with serine, by substituting an amino acid at position 9 of the peptide of SEQ ID NO: 1 with tyrosine, or by substituting an amino acid at position 10 of the peptide of SEQ ID NO: 1 with lysine (Table 4).

[Table 4]

Peptides of Group 4

SEQ ID NO:	Amino acid sequence(N-C)
25	KYQRRKKSYK
26	KYQRRKRSYK
27	KYQRRRKSYK
28	KYQRRRRSYK
29	KYQRKKKSYK

30	KYQRKRKSYK
31	KYQRKKRSYK
32	KYQRKRRSYK

Next, peptides of Group 5 were synthesized by substituting an amino acid at position 3 of the peptide of SEQ ID NO: 1 with arginine, by substituting an amino acid at position 4 of the peptide of SEQ ID NO: 1 with glutamine, or by substituting any amino acid at positions 5 to 7 of the peptide of SEQ ID NO: 1 with lysine or arginine (Table 5).

[Table 5]

Peptides of Group 5

SEQ ID NO:	Amino acid sequence(N-C)
33	KYRQRKKNKY
34	KYRQRKRNKY
35	KYRQRRKNKY
36	KYRQRRRNKY
37	KYRQKKKNKY
38	KYRQKRKNKY
39	KYRQKKRNKY
40	KYRQKRRNKY

Next, peptides of Group 6 were synthesized by substituting an amino acid at position 3 of the peptide of SEQ ID NO: 1 with arginine, by substituting an amino acid at position 4 of the peptide of SEQ ID NO: 1 with glutamine, by substituting any amino acid at positions 5 to 7 of the peptide of SEQ ID NO: 1 with lysine or arginine, or by substituting an amino acid at position 8 of the peptide of SEQ ID NO: 1 with serine (Table 6).

[Table 6]

Peptides of Group 6

SEQ ID NO:	Amino acid sequence(N-C)
41	KYRQRKKSKY
42	KYRQRKRSKY
43	KYRQRRKSKY
44	KYRQRRRSKY
45	KYRQKKKSKY
46	KYRQKRKSKY
47	KYRQKKRSKY
48	KYRQKRRSKY

Next, peptides of Group 7 were synthesized by substituting an amino acid at position 3 of the peptide of SEQ ID NO: 1 with arginine, by substituting an amino acid at position 4 of the

peptide of SEQ ID NO: 1 with glutamine, by substituting any amino acid at positions 5 to 7 of the peptide of SEQ ID NO: 1 with lysine or arginine, by substituting an amino acid at position 9 of the peptide of SEQ ID NO: 1 with tyrosine, or by substituting an amino acid at position 10 of the peptide of SEQ ID NO: 1 with lysine (Table 7).

[Table 7]

Peptides of Group 7

SEQ ID NO:	Amino acid sequence(N-C)
49	KYRQRKKNYK
50	KYRQRKRNYK
51	KYRQRRKNYK
52	KYRQRRRNYK
53	KYRQKKKNYK
54	KYRQKRKNYK
55	KYRQKKRNYK
56	KYRQKRRNYK

Next, peptides of Group 8 were synthesized by substituting an amino acid at position 3 of the peptide of SEQ ID NO: 1 with arginine, by substituting an amino acid at position 4 of the peptide of SEQ ID NO: 1 with glutamine, by substituting any amino acid at positions 5 to 7 of the peptide of SEQ ID NO: 1

with lysine or arginine, by substituting an amino acid at position 8 of the peptide of SEQ ID NO: 1 with serine, by substituting an amino acid at position 9 of the peptide of SEQ ID NO: 1 with tyrosine, or by substituting an amino acid at position 10 of the peptide of SEQ ID NO: 1 with lysine (Table 8).

[Table 8]

Peptides of Group 8

SEQ ID NO:	Amino acid sequence(N-C)
57	KYRQRKKSYK
58	KYRQRKRSYK
59	KYRQRRKSYK
60	KYRQRRRSYK
61	KYRQKKKSYK
62	KYRQKRKSYK
63	KYRQKKRSYK
64	KYRQKRRSYK

Next, peptides of Group 9 were synthesized by substituting an amino acid at position 3 of the peptide of SEQ ID NO: 1 with lysine, by substituting an amino acid at position 4 of the peptide of SEQ ID NO: 1 with glutamine, or by substituting any

amino acid at positions 5 to 7 of the peptide of SEQ ID NO: 1 with lysine or arginine (Table 9).

[Table 9]

Peptides of Group 9

SEQ ID NO:	Amino acid sequence(N-C)
65	KYKQRKKNKY
66	KYKQRKRNKY
67	KYKQRRKNKY
68	KYKQRRRNKY
69	KYKQKKKNKY
70	KYKQKRKNKY
71	KYKQKKRNKY
72	KYKQKRRNKY

Next, peptides of Group 10 were synthesized by substituting an amino acid at position 3 of the peptide of SEQ ID NO: 1 with lysine, by substituting an amino acid at position 4 of the peptide of SEQ ID NO: 1 with glutamine, by substituting any amino acid at positions 5 to 7 of the peptide of SEQ ID NO: 1 with lysine or arginine, or by substituting an amino acid at position 8 of the peptide of SEQ ID NO: 1 with serine (Table 10).

[Table 10]

Peptides of Group 10

SEQ ID NO:	Amino acid sequence(N-C)
73	KYKQRKKSKY
74	KYKQRKRSKY
75	KYKQRRKSKY
76	KYKQRRRSKY
77	KYKQKKKSKY
78	KYKQKRKSKY
79	KYKQKKRSKY
80	KYKQKRRSKY

Next, peptides of Group 11 were synthesized by substituting an amino acid at position 3 of the peptide of SEQ ID NO: 1 with lysine, by substituting an amino acid at position 4 of the peptide of SEQ ID NO: 1 with glutamine, by substituting any amino acid at positions 5 to 7 of the peptide of SEQ ID NO: 1 with lysine or arginine, by substituting an amino acid at position 9 of the peptide of SEQ ID NO: 1 with tyrosine, or by substituting an amino acid at position 10 of the peptide of SEQ ID NO: 1 with lysine (Table 11).

[Table 11]

Peptides of Group 11

SEQ ID NO:	Amino acid sequence(N-C)
81	KYKQRKKNYK
82	KYKQRKRNYK
83	KYKQRRKNYK
84	KYKQRRRNYK
85	KYKQKKKNYK
86	KYKQKRKNYK
87	KYKQKKRNYK
88	KYKQKRRNYK

Lastly, peptides of Group 12 were synthesized by substituting an amino acid at position 3 of the peptide of SEQ ID NO: 1 with lysine, by substituting an amino acid at position 4 of the peptide of SEQ ID NO: 1 with glutamine, by substituting any amino acid at positions 5 to 7 of the peptide of SEQ ID NO: 1 with lysine or arginine, by substituting an amino acid at position 8 of the peptide of SEQ ID NO: 1 with serine, by substituting an amino acid at position 9 of the peptide of SEQ ID NO: 1 with tyrosine, or by substituting an amino acid at position 10 of the peptide of SEQ ID NO: 1 with lysine (Table 12).

[Table 12]

Peptides of Group 12

SEQ ID NO:	Amino acid sequence(N-C)
89	KYKQRKKSYK
90	KYKQRKRSYK
91	KYKQRRKSYK
92	KYKQRRRSYK
93	KYKQKKKSYK
94	KYKQKRKSYK
95	KYKQKKRSYK
96	KYKQKRRSYK

Example 2: Verification of the effect of promoting regeneration of dentin using the odontoblasts.

Example 2-1: Validation of the effect of peptides on the activity of the DSPP (dentin sialophosphoprotein) promoter.

First, MDPC-23 cells, which are mouse-derived odontoblasts, were cultured in DMEM medium containing 10% FBS, 5% CO_2 and 37°C.

Next, the cultured MDPC-23 cells were dispensed into a 24-well plate at 5×10^4 cells per well, incubated for 24 hours. And then using Lipofectamine Plus™ reagent, the cultured cells were transformed by introducing a recombinant vector(pGL3 vector) –

which the DSPP promoter and luciferase gene were introduced. The transformed MDPC-23 cells were treated with the peptides of groups 1 to 12 synthesized in Example 1, respectively, and cultured for 48 hours. Then luciferase activity was measured, and the average level was compared (Fig. 1a). As a control, transformed MDPC-23 cells without an odontoblast differentiation promoting peptide were used.

FIG. 1A is a graph showing the effect of each peptide provided in the present invention on the expression of DSPP, an odontoblast differentiation marker gene, for each group. As shown in FIG. 1A, each peptide provided in the present invention showed a value of about 1.3 times or more of the luciferase activity level measured in the control group as a whole, but the difference was shown for each group, and the peptide of group 12 was the highest. And the next highest level of luciferase activity was from group 11 peptide.

Therefore, it was verified that the peptides provided by the present invention exhibit an effect of activating the DSPP promoter.

Example 2-2: Verification of the effect of peptides on the expression level of the DSPP gene, an odontoblast differentiation marker gene

The MDPC-23 cells cultured in Example 2-1 were treated with the peptides of each group synthesized in Example 1, then cultured for 48 hours. The mRNA level of the DSPP, an odontoblast differentiation marker gene, expressed in the MDPC-23 cells were measured, and the measured mRNA level of each DSPP gene was converted into a relative ratio to the mRNA level of the DSPP gene measured in control (Tables 13 to 24). In addition, the average value of the mRNA level of the DSPP gene measured according to the peptides of each group was compared (Fig. 1b). At this time, as a control, MDPC-23 cells that were not treated with the peptide promoting differentiation of odontoblast were used.

The expression level of the DSPP gene was measured through RT-PCR and real-time PCR analysis: Specifically, total RNA was extracted from the MDPC-23 cells using TRIzol reagent. 2 μg of the total RNA, 1 μl of reverse transcriptase, and 0.5 μg of oligo (oligo; dT) were used to synthesize cDNA. The synthesized cDNA was used in a real-time polymerase chain reaction. The real-time polymerase chain reaction was performed on an ABI PRISM 7500 sequence detection system (Applied Biosystems) and an SYBR GREEN PCR Master Mix (Takara, Japan). The real-time polymerase chain reaction was performed under conditions of 94°C, 1 min; 95°C, 15 sec; 60°C, 34 sec for 40 cycles. Results

were analyzed by a comparative cycle threshold (CT) method. At this time, the Gapdh gene was used as the internal control group, and the measured value was repeated three times. The mean value and standard deviation value thereof were used.

Dspp_F: 5'-ATTCCGGTTCCCCAGTTAGTA-3'(SEQ ID NO: 97)

Dspp_R: 5'-CTGTTGCTAGTGGTGCTGTT-3'(SEQ ID NO: 98)

Gapdh_F: 5'-AGGTCGGTGTGAACGGATTTG-3'(SEQ ID NO: 99)

Gapdh_R: 5'-TGTAGACCATGTAGTTGAGGTCA-3'(SEQ ID NO: 100)

[Table 13]

Effects of peptides of group 1 on mRNA level of Dspp gene

	mRNA level of Dspp gene	
SEQ ID NO:	Mean	Standard deviation
1	1.754	0.132
2	1.646	0.092
3	1.464	0.221
4	1.855	0.102
5	1.639	0.057
6	1.746	0.091
7	1.864	0.132
8	1.639	0.032

[Table 14]

Effects of peptides of group 2 on mRNA level of Dspp gene

	mRNA level of Dspp gene	
SEQ ID NO:	Mean	Standard deviation
9	1.854	0.032
10	1.746	0.052
11	1.639	0.201
12	1.548	0.027
13	1.685	0.077
14	1.846	0.141
15	1.964	0.279
16	1.739	0.092

[Table 15]

Effects of peptides of group 3 on mRNA level of Dspp gene

	mRNA level of Dspp gene	
SEQ ID NO:	Mean	Standard deviation
17	2.117	0.209
18	2.319	0.092
19	1.931	0.102
20	2.553	0.099

21	1.893	0.132
22	2.412	0.072
23	2.171	0.281
24	2.212	0.111

[Table 16]

Effects of peptides of group 4 on mRNA level of Dspp gene

	mRNA level of Dspp gene	
SEQ ID NO:	Mean	Standard deviation
25	2.371	0.089
26	2.193	0.052
27	1.993	0.202
28	2.453	0.192
29	1.883	0.101
30	2.512	0.209
31	2.371	0.139
32	2.219	0.302

[Table 17]

Effects of peptides of group 5 on mRNA level of Dspp gene

	mRNA level of Dspp gene	
SEQ ID NO:	Mean	Standard

		deviation
33	1.712	0.091
34	1.931	0.172
35	1.983	0.102
36	2.319	0.292
37	1.597	0.301
38	2.116	0.211
39	1.712	0.191
40	2.219	0.212

[Table 18]

Effects of peptides of group 6 on mRNA level of Dspp gene

	mRNA level of Dspp gene	
SEQ ID NO:	Mean	Standard deviation
41	1.546	0.091
42	1.586	0.103
43	1.669	0.095
44	1.793	0.203
45	1.532	0.31
46	1.887	0.077
47	1.697	0.009
48	1.558	0.201

[Table 19]

Effects of peptides of group 7 on mRNA level of Dspp gene

	mRNA level of Dspp gene	
SEQ ID NO:	Mean	Standard deviation
49	1.923	0.192
50	1.887	0.007
51	1.601	0.082
52	2.019	0.135
53	1.592	0.222
54	1.437	0.341
55	1.663	0.094
56	1.701	0.109

[Table 20]

Effects of peptides of group 8 on mRNA level of Dspp gene

	mRNA level of Dspp gene	
SEQ ID NO:	Mean	Standard deviation
57	2.039	0.082
58	1.998	0.172
59	1.792	0.007

60	2.107	0.201
61	2.301	0.019
62	1.672	0.308
63	1.769	0.085
64	1.967	0.039

[Table 21]

Effects of peptides of group 9 on mRNA level of Dspp gene

	mRNA level of Dspp gene	
SEQ ID NO:	Mean	Standard deviation
65	1.723	0.072
66	1.627	0.291
67	1.777	0.027
68	1.432	0.41
69	2.011	0.081
70	1.927	0.105
71	1.879	0.06
72	2.011	0.009

[Table 22]

Effects of peptides of group 10 on mRNA level of Dspp gene

	mRNA level of Dspp gene

SEQ ID NO:	Mean	Standard deviation
73	2.035	0.021
74	2.011	0.063
75	1.997	0.059
76	2.351	0.109
77	1.729	0.111
78	2.635	0.091
79	2.231	0.077
80	1.837	0.201

[Table 23]

Effects of peptides of group 11 on mRNA level of Dspp gene

	mRNA level of Dspp gene	
SEQ ID NO:	Mean	Standard deviation
81	3.092	0.152
82	3.361	0.098
83	3.572	0.209
84	3.702	0.301
85	3.67	0.088
86	3.705	0.137
87	3.888	0.072

88	4.021	0.301

[Table 24]

Effects of peptides of group 12 on mRNA level of Dspp gene

	mRNA level of Dspp gene	
SEQ ID NO:	Mean	Standard deviation
89	4.211	0.413
90	4.811	0.302
91	4.362	0.182
92	4.211	0.287
93	4.525	0.25
94	3.836	0.099
95	4.62	0.401
96	5.211	0.371

As shown in Tables 13 to 24, compared to the mRNA level of the DSPP gene measured in the control group, it was confirmed that the mRNA levels of the DSPP gene of the experimental group treated with the peptide were all 1.3 times or more. In particular, it was confirmed that all the peptides of group 11 showed a value of 3 times or more in the mRNA level of the DSPP gene, and all peptides of group 12 showed a value of 3.8 times

or more in the mRNA level of the DSPP gene.

In addition, FIG. 1B is a graph comparing the expression level of the DSPP gene, an odontoblast differentiation marker, in MDPC-23 cells treated with an odontoblast differentiation promoting peptide. As shown in FIG. 1B, when the peptide for promoting differentiation of odontoblast was treated, the mRNA level of the DSPP gene, which is a marker for odontoblast differentiation, was increased. Similar to that of FIG. 1A, it was confirmed that a value of about 1.3 times or more compared to the level of DSPP gene mRNA was measured in the control group.

Example 2-3: Verification of the effect of the peptide on the expression level of the odontoblast differentiation marker genes DSPP, Dmp1, and Nestin genes

From the results of Example 2-2, it was confirmed that the odontoblast differentiation promoting peptide could increase the mRNA level of the DSPP gene, and in particular, the peptides of groups 11 and 12 can increase the mRNA level of the DSPP gene by at least 3 times or more.

Accordingly, it was confirmed whether the peptides of groups 11 and 12 can also increase the mRNA levels of the Dmp1 and Nestin genes, which are other odontoblast differentiation

marker genes.

The following primers were used with the method from Example 2-2. The peptides of groups 11 and 12 were used, thereby affecting the expression levels of Dmp1 and Nestin genes. The effect of the differentiation promoting peptide was measured, and the average level was compared (Fig. 1C). At this time, as a control group, MDPC-23 cells without a peptide promoting differentiation of odontoblast were used.

Dmp1_F: 5'-CATTCTCCTTGTGTTCCTTTGGG-3'(SEQ ID NO 101)

Dmp1_R: 5'-TGTGGTCACTATTTGCCTGTG-3'(SEQ ID NO 102)

Nestin_F: 5'-CCCTGAAGTCGAGGAGCTG-3'(SEQ ID NO 103)

Nestin_R: 5'-CTGCTGCACCTCTAAGCGA-3'(SEQ ID NO 104)

FIG. 1C is a graph showing the results of comparing the expression levels of the odontoblast differentiation marker DSPP, Dmp1, and Nestin genes in MDPC-23 cells treated with the peptides of groups 11 and 12. As shown in Fig. 1c, when the odontoblast differentiation promoting peptide was treated, the expression levels of the odontoblast differentiation marker DSPP, Dmp1, and Nestin genes were all increased, but the level of increase for each gene was different. It was confirmed that the peptide of group 12 was more effective.

Since each differentiation marker gene is known to be

involved in the differentiation of odontoblasts and dentin calcification, the peptides provided in the present invention were analyzed whether they promote dentin regeneration.

Example 2-4: Evaluation of cytotoxicity of peptides on pulp tissue cells

Human dental pulp cells were separated from wisdom teeth of 10 adults (aged 18-22) at the School of Dentistry, Seoul National University. In detail, all experiments were performed after the approval of the Institutional Review Board and obtaining informed consent from patients. Wisdom teeth were fractured according to a method of Jung HS et al. (J Mol Histol.(2011)) to expose the dental pulps, and dental pulp tissues were separated with forceps. Each of the separated dental pulp tissues was chopped into small pieces with a razor blade, put in a 60-mm dish, covered with a coverslip, and then cultured in a Dulbecco’s modified Eagle’s medium.

Next, the obtained dental pulp tissue cells were dispensed into a 96-well plate, so the number of cells per well was to be about 3×10^3, cultured for 24 hours. Then the peptides of groups 11 or 12 were treated at a concentration of 10 or 50 μg/ml. And it was incubated again for 1, 3, or 5 days. The cultured cells were washed with PBS, 20 μl of MTT solution was added, and then reacted at about 37°C for 4 hours. After the reaction was

completed, the MTT solution was removed, 100 μl of DMSO was added, and absorbance was measured at a wavelength of 540 nm (Fig. 1D). At this time, as a control, pulp tissue cells cultured without the peptide were used.

FIG. 1D is a graph showing the cytotoxicity of a peptide that promotes the differentiation of oblasts to dental pulp tissue cells. As shown in FIG. 1D, it was confirmed that the survival rate of pulp tissue cells was at the same level as that of the control group even when the odontoblast differentiation promoting peptide was added.

Example 3: Preparation of oral care composition for alleviating dentin hyperesthesia

Step 1:

Add poloxamer 407 to purified water and stir in a stirrer for about 30 minutes (Stirring conditions: PADDLE 15~20 rpm, DISPERSE 400~500 rpm)

Step 2:

Add potassium sorbate, cetylpyridinium chloride, xylitol, acesulfame potassium colorant (blue No. 1), D-sorbitol solution, and concentrated glycerin and stir in a stirrer for about 30 minutes (Stirring condition: PADDLE 15~20 rpm, DISPERSE 400~ 500 rpm)

Step 3:

Add polysorbate 20 (Tween 20), *Scutellaria Baicalensis* root extract, green tea extract, chamomile extract, rosemary extract, and mint flavor (HF-3585) by heating and stir in a stirrer for about 30 minutes (stirring condition: PADDLE 15~20 rpm, DISPERSE 400~500 rpm)

Step 4:

After mixing about 0.0001% of the odontoblast differentiation promoting peptide (SEQ ID NO: 96), citric acid hydrate is added to adjust pH 5.5 to 6.0

[Table 25]

oral care composition for alleviating dentin hyperesthesia according to Example 3

Component		Ingredient	Content (Wt%)
1	Solvent	Purified water	86.25
	Surfactant	Poloxamer407	0.3
	Odontoblast differentiation promoting peptide	Peptide (SEQ ID NO: 96)	0.0001
2	Preservative	Potassium	0.1

		sorbate	
	Staple	Cetylpyridinium chloride	0.055
	Sweetening agent	Xylitol	1
		Acesulfame potassium	0.05
	pH Adjuster	Citric acid hydrate	0.005
	Coloring agent	Blue 1(CI 42090)	0.00025
	Humectant	D-Sorbitol Solution	5
		Concentrated glycerin	5
3	Surfactant	Polysorbate(Tween 20)	2
	Flavoring agent	Scutellaria Baicalensis root extract	0.01
		green tea extract	0.01
		chamomile	0.01

		extract	
		rosemary extract	0.01
		mint flavor (HF-3585)	0.2
	Total		100

Preparing Compositions of Comparative Example

Comparative Example 3-1:

Prepared purified water of the same volume as in Example 3

Comparative Example 3-2:

Among the ingredients of Example 3, all ingredients other than those that did not contain an odontoblast differentiation promoting peptide (SEQ ID NO: 96) were prepared to be contained the same.

Test Example 1:

Observation of the dentinal tubule permeability of the oral care composition for alleviating dentin hyperesthesia according to Example 3.

A. Cut the tooth to expose the dentinal tubules

Cut the crown of the extracted person's tooth horizontally with a diamond saw to expose the dentinal tubules, and then wash

twice for about 5 minutes with a phosphate buffer solution.

B. Cleaning amputated tooth

The previously cut tooth was reacted with 0.5 M ethylenediaminetetraacetic acid (EDTA, pH 7.4) solution for about 5 minutes and then washed twice for about 5 minutes with a phosphate buffer solution.

C. Addition of a fluorescent dyeing reagent to the oral care composition for alleviating dentin hyperesthesia according to Example 3

Added 0.1% of the fluorescent dyeing reagent to the oral care composition for alleviating dentin hyperesthesia containing the odontoblast differentiation promoting peptide (SEQ ID NO: 96), mixed well, and then reacted the cut tooth exposed to the dentinal tubules for about 1 minute.

D. Observation of Penetration of oral care composition for alleviating dentin hyperesthesia

The reacted cut tooth was washed twice in a phosphate buffer solution for about 5 minutes and then cut lengthwise to a thickness of about 0.5 mm so that the dentinal tubules of the cut tooth looked long using a diamond saw, and the degree of penetration was observed with a fluorescence microscope (Fig. 2)

Test Example 2:

Observation of the sealing ability of the dentinal tubules

of the oral care composition for alleviating dentin hyperesthesia according to Example 3

A. Preparation of artificial saliva

The composition of artificial saliva is shown in Table 26 below.

※ The purified water was added to the final concentration of each component in Table 2 and mixed, and potassium phosphate (K2HPO4) was added last.

※ The pH of artificial saliva is measured near 7.2, similar to human saliva.

[Table 26]

Ingredient	**concentration (mM)**
$CaCl_2$	0.7
$Mgcl_2$	0.2
K_2HPO_4	4
KCl	30
NaN_3	0.3
HEPES	20

B. Making dentinal tuble specimens

The extracted human tooth was cut horizontally using a diamond saw to make a 1mm thick dentin specimen with exposed dentinal tubules.

※ The dentin specimen was reacted for about 5 minutes in a 32% phosphoric acid solution to expose the dentinal tubules completely, and then washed three times with purified water for about 5 minutes. Then, the dentin specimen was washed 6 times in an ultrasonic cleaner for about 5 minutes to expose the dentinal tubules completely.

Thereafter, washed three times with a phosphate buffer solution and stored.

C. Observation of the sealing ability of the dentinal tubules of the oral care composition for alleviating dentin hyperesthesia

Using the oral care composition for alleviating dentin hyperesthesia according to Example 3, the specimen was reacted for about 1 minute to the dentinal tubule specimen and then reacted to the artificial saliva for about 24 hours.

After repeating this process for 2 weeks, washed three times with distilled water, dried, and observed the degree of dentinal tubule blockade with a scanning electron microscope (S-

4700, HITACHI, Tokyo, Japan) (FIG. 3, G-I).

Comparative Test Example 2-1:

Using the purified water prepared in Comparative Example 3-1, reacted for about 1 minute in the dentinal tubule specimen, and then reacted for about 24 hours in artificial saliva.

After repeating this process for 2 weeks, the specimens were washed with distilled water 3 times, dried, and observed the degree of dentinal tubule blockade with a scanning electron microscope (Fig. 3, A-C).

Comparative Test Example 2-2:

Using the composition for oral care prepared in Comparative Example 3-2, reacted for about 1 minute on the dentinal tubule specimen, and then reacted for about 24 hours with artificial saliva.

After repeating this process for 2 weeks, the specimens were washed with distilled water 3 times, dried, and observed the degree of dentinal tubule blockade with a scanning electron microscope (Fig. 3, D-F).

According to Test Example 1, as a result of observing the dentinal tubule permeability of the oral care composition for alleviating dentin hyperesthesia according to Example 3 with a

fluorescence microscope, as shown in FIG. 2, in the case of the tooth treated with the composition, fluorescence was strongly observed on the dentin surface. In addition, penetration of the fluorescent staining reagent was observed along the lower side of the exposed dentinal tubules.

Next, the results of comparing Test Example 2 and Comparative Test Examples 2-1 and 2-2 are as shown in FIG. 3. FIG. 3 is a set of images comparing the sealing ability of the dentinal tubules of composition for oral care composition for alleviating dentin hyperesthesia according to Example 3, Comparative Example 3-1, and Comparative Example 3-2. And in more detail, A-C shows the dentinal tubules of the dentin treated only with purified water (Comparative Example 3-1), and D-F shows the oral care composition without odontoblast differentiation promoting peptide (Comparative Example 3-2), and G-I shows the dentinal tubules reacted with the composition including a peptide for oral care that prevents or alleviates dentin hyperesthesia according to an embodiment of the present invention. One (size bar: A, D, G, 100 μm; B, E, H, 20 μm; C, F, I, 10 μm).

As can be seen from FIG. 3, it could be observed that the dentinal tubules were blocked by remineralization in the dentinal tubules by reacting with the composition including a peptide for oral care that prevents or alleviates dentin

hyperesthesia according to an embodiment of the present invention.

FIG. 4 is an enlarged image of the dentinal tubules blocked by the composition, including a peptide for oral care that prevents or alleviates dentin hyperesthesia according to an embodiment of the present invention and shows the results of remineralization in the blocked dentinal tubules and dentin surfaces.

Referring to FIG. 4, the appearance of the dentinal tubules according to Experimental Example 3 can be observed in more detail, and it can be seen that remineralization has occurred in both the dentinal tubules and the dentin surface. The composition, including a peptide for oral care that prevents or alleviates dentin hyperesthesia according to an embodiment of the present invention. It forms a thin film on the dentin and at the same time, strongly binds to the phosphate-calcium ions present in the dentinal tubules and saliva, which remineralizes the exposed dentinal tubules and dentin surfaces. In other words, the composition including a peptide for oral care that prevents or alleviates dentin hyperesthesia according to an embodiment of the present invention induces remineralization not only on the surface of the exposed dentinal tubules but also inside the dentinal tubule, thereby exhibiting the effect of reducing and/or preventing symptoms of ache.

While one or more embodiments of the present invention have been described with reference to the figures, it will be understood by those of ordinary skill in the art that various changes in form and details may be made therein without departing from the spirit and scope of the present invention.

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CLAIMS

1. Oral care composition for alleviating dentin hyperesthesia, comprising a peptide consisting of an amino acid sequence of the following Formula 1:

K-Y-R1-R2-R3-R4-R5-R6-R7-R8 (Formula 1)

wherein R1 is arginine(R), lysine(K) or glutamine(Q);

R2 is arginine(R) or glutamine(Q);

R3, R4, and R5 are arginine(R) or lysine(K), respectively;

R6 is asparagine(N) or serine(S); and

R7 and R8 are lysine(K) or tyrosine(Y), respectively,

wherein said oral care composition includes 0.00005-0.00015 parts by weight of the peptide, 85-87 parts by weight of purified water, 1.7-2.9 parts by weight of surfactant, and 0.0045-0.0055 parts by weight of citric acid hydrate based on 100 parts by weight,

wherein said oral care composition forms a thin film on the surface of said dentin and induces remineralization on said surface of said dentin and dentinal tubule by binding with phosphate-calcium ions present in said dentinal tubules and in saliva.

2. The composition of claim 1, wherein said peptide is any one amino acid sequence of SEQ ID NOS: 1 to 96.

3. The composition of claim 1, wherein said oral care composition comprises 0.0545-0.555 parts by weight of

cetylpyridinium chloride on 100 parts by weight.

4. The composition of claim 1, wherein said surfactant is poloxamer and/or polysorbate 20.

5. The composition of claim 1, wherein said surfactant includes 12-14% by weight of the poloxamer 407 and 86-88% by weight of the polysorbate 20.

6. The composition of claim 1, wherein said oral care composition includes 9-11 parts by weight of a humectant based on 100 parts by weight.

7. The composition of claim 6, wherein said humectant is D-sorbitol solution and/or concentrated glycerin.

8. The composition of claim 7, wherein said humectant comprises 45-55% by weight of said D-sorbitol solution and 45-55% by weight of said concentrated glycerin.

9. A method of alleviating dentin hyperesthesia in a subject in need thereof, administering the oral care composition according to claim 1, to dentin surface of said subject.