

THE USE OF *BACILLUS* PB6 FOR THE PROPHYLAXIS OR TREATMENT OF GASTROINTESTINAL AND IMMUNO-RELATED DISEASES

This application claims priority to United States Patent Application Serial No. 60/740,518, filed November 29, 2005.

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Background of the Invention

The invention relates generally to the administration of bacteria to treat gastrointestinal disease and, more specifically, to the administration of bacteria of a strain of *Bacillus amyloliquefaciens* to treat antibiotic associated diarrhea (AAD) and *Clostridium difficile* associated disease (CDAD).

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The term antibiotic-associated diarrhea refers to a benign, self-limited diarrhea, following the use of antimicrobials. Typically no pathogens are identified and the diarrhea is due to changes in the composition and function of the intestinal flora. Most patients respond to supportive measures and discontinuation of antibiotics.

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The prolonged use of multiple antibiotics, especially broad-spectrum agents with poor intestinal absorption or high biliary excretion, induces a change in the composition and function of the intestinal flora and therefore results in a higher incidence of AAD.^{1,2} The degree of alteration will be influenced by the ability of the normal flora to resist colonization and the type of antibiotic used. A decrease in the colonic anaerobic flora interferes with carbohydrate and bile acid metabolism. Osmotic or secretory diarrhea may occur. Overgrowth of opportunistic pathogens takes place as a result of microbiologic and metabolic alterations.

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C difficile, an anaerobic gram-positive rod, accounts for 15% to 20% of all AAD cases. In particular this organism can be isolated in a great number of AAD cases with evidence of colitis and in all those with pseudomembranes. It is widely present in the environment, may survive for a considerable time, and is transmitted by the fecal-oral route to susceptible individuals. It is considered part of the normal flora of infants and can be isolated in about 5% of healthy adults and in up to one third of asymptomatic or colonized, hospitalized patients.

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The clinical manifestations of AAD may vary from mild diarrhea to fulminant colitis.³ The severity of *C difficile* colitis appears to be influenced by a myriad of factors including age, comorbidity, host's immune response, and the use of antiperistaltic agents. Interestingly, bacterial genotype and toxin production appear to play minimal roles.⁴ The cardinal symptom of the disease is diarrhea that commonly develops during treatment but may appear as late as 8 weeks after discontinuation of antibiotics. In most cases of AAD,

patients present with loose stools, minimal signs of colitis, and no constitutional symptoms. The diarrhea promptly responds to supportive measures and withdrawal of the antimicrobial agent.

Clostridium difficile was first described in 1935⁵, but it was not associated with antibiotic-related diarrhea until the late 1970's. *Clostridium difficile* is a spore-forming gram-positive anaerobic *Bacillus* that produces at least two exotoxins: toxin A, primarily an enterotoxin, and toxin B, a cytotoxin. The organism causes gastro-intestinal infections in humans that range in severity from asymptomatic colonization to severe diarrhea, pseudomembranous colitis (PMC), toxic megacolon, colonic perforation, and death^{6,7,8}. The first step in development of *C. difficile* colonization is disruption of the normal flora of the colon, usually caused by antibiotics or, in unusual cases, by antineoplastic or immunosuppressive drugs^{9,10}. Colonization occurs by the fecal-oral route; ingested spores of *C. difficile* survive the gastric acid barrier and germinate in the colon^{11,12}. Symptoms of CDAD may start on the first day of antibiotic therapy or up to several weeks after antibiotic therapy is stopped¹³. The following two factors recently have been shown to increase the probability of symptomatic disease in patients who acquire *C. difficile* colonization in hospital; the severity of other illnesses, and reduced levels of serum IgG antibody to toxin A¹⁴. These results suggest that pre-existing anti-toxin A antibody may ameliorate severity of disease and that immunization might be efficacious in preventing and controlling nosocomial CDAD.

For clarity, we define patients as having *C. difficile*-associated disease (CDAD) if they display symptomatic illness caused by *C. difficile*. Detection of the presence of a *C. difficile* toxin in the stool of patients with diarrhea has been the most generally accepted method of diagnosis.

Clostridium difficile is the cause of approximately 25% of all cases of antibiotic-associated diarrhea¹⁵. Most cases of *C. difficile*-associated disease occur in hospitals or long-term care facilities (rate of 25-60 per 100,000 occupied bed-days), causing more than 300,000 cases per year in the US and similar rates estimated for many European countries. It can add up to two weeks to the length of the hospitalization, at an additional cost of \$6,000 - \$10,000 per case^{16,17,18,19,20,21}

Diarrhea may resolve spontaneously in patients with CDAD once the inciting antibiotic has been withdrawn, and for some patients with mild disease no specific therapy may be necessary^{22,23}. However, the standard practice is to treat almost all symptomatic patients with the antibiotics vancomycin or metronidazole. Although metronidazole is not

currently approved by the FDA for the treatment of CDAD, it is widely used as first-line therapy due to the higher cost of vancomycin and concerns over the emergence of vancomycin resistant bacteria. Because metronidazole effectively disrupts normal enteric flora, it also predisposes patients to colonization with metronidazole resistant enterococci²⁴. Oral metronidazole (250 mg 4 times per day or 500 mg 3 times per day) for 10-14 d is usually adequate. Oral vancomycin hydrochloride (125 mg 4 times per day) for 10-14 d is indicated for those who cannot tolerate oral metronidazole, those in whom metronidazole therapy fails, pregnant patients, and, perhaps, severely ill patients. The first relapse/recurrence of *Clostridium difficile* colitis can be treated with another 10- to 14-d course of oral metronidazole or vancomycin.

Treatment of CDAD with vancomycin or metronidazole propagates a vicious cycle by altering normal, protective flora of the gut. Consequently 20% of treated patients with an initial episode have a recurrence of CDAD, usually within two weeks after discontinuation of therapy^{25,26,27,28}. A further benefit of removing antibiotics from the treatment regimen of CDAD is a reduction in selective pressure for bacterial resistance. Vancomycin and metronidazole have been clearly shown to select for resistant gram-positive cocci, such as VRE. Retrospective epidemiological studies have linked intestinal VRE colonization with the use of broad-spectrum antibiotics such as the cephalosporins, fluoroquinolones and metronidazole²⁹. Intestinal VRE colonization provides a reservoir for this pathogen within the hospital. Many strains of VRE are multiresistant, leaving few options for treatment of life-threatening systemic infections. *C. difficile* patients, perhaps because of their prior antibiotic exposure, appear to be especially susceptible to VRE colonization and infection^{30,31}. Management of VRE colonization is a critical component of hospital infection control practices. Therefore, therapeutic strategies that reduce the risk of VRE colonization both in the general patient population and in *C. difficile* patients are highly desirable. The potential emergence of vancomycin and metronidazole resistant *C. difficile* presents an additional risk for the use of antibiotics to treat this disease. Currently, the occurrence of antibiotic resistant *C. difficile* is sporadic but has been reported in up to 12% of clinical isolates³².

Summary of the Invention

[0010] The invention relates to the prophylaxis of a bowel condition, such as antibiotic associated diarrhea, *Clostridium difficile* acquired diarrhea, inflammatory bowel disease, and gastro-intestinal disease, by administering an effective amount of a *Bacillus* bacteria that produces lipopeptides. The *Bacillus* bacteria may be administered as a probiotic and may be combined with other probiotics, such as inulin.

Using biochemical methods, and specifically API 50 CBH/L, a preferred *Bacillus* was putatively identified as *Bacillus subtilis*. Using 16S rRNA, the preferred *Bacillus* was also putatively identified as *Bacillus subtilis*. Using *gyrA*, the preferred *Bacillus* was putatively identified as *Bacillus amyloliquefaciens*. Prior to the *gyrA* assay, the preferred *Bacillus* was deposited with the ATCC and identified as *Bacillus subtilis*.

The invention also relates to an isolated bacteria strain having the 16S rRNA sequence of SEQ ID NO. 1, to an isolated bacterial strain having 90% homology, 80% homology, 70% homology, 60% homology and 50% homology to SEQ ID NO. 1.

The invention also relates to an isolated bacteria strain having the partial *gyrA* sequence of SEQ ID NO. 2, to an isolated bacterial strain having 90% homology, 80% homology, 70% homology, 60% homology and 50% homology to SEQ ID NO. 2.

The invention also relates to an isolated bacteria strain having the partial *gyrA* sequence of SEQ ID NO. 3, to an isolated bacterial strain having 90% homology, 80% homology, 70% homology, 60% homology and 50% homology to SEQ ID NO. 3.

The invention further relates to a *Bacillus* bacteria of the strain identified as ATCC strain PTA-6737.

The preferred *Bacillus* PB6 can be positioned for several unmet medical needs due to its versatile and unique characteristics.

Brief Description of the Figures

Fig. 1 is a photograph of the antagonistic effect of *Bacillus* PB6 (1) against *C. perfringens* ATCC13124 (2) and *C. difficile* ATCC9689 (3).

Fig 2 is a photograph of the antagonistic effect of *Bacillus* PB6 on *Clostridium difficile* NAP1/027.

Fig. 3 is a photograph of the antagonistic effect of *Bacillus* PB6 on *Campylobacter jejuni* ATCC55918.

Fig. 4 is a drawing of the chemical structure of surfactin.

Fig. 5 is a chart of the % survival in hamsters suffering from CDAD with different treatments.

Fig. 6 is the 1466-bp 16S rRNA gene sequence (nucleotide position of 27-1492) of *Bacillus* PB6.

5 Fig. 7 is the 1023-bp partial *gyrA* sequence (nucleotide position of 43-1065) of *Bacillus* PB6.

Fig. 8 is the 801-bp consensus sequence obtained from partial *gyrA* sequencing of *Bacillus* PB6.

10 Fig. 9 is a gel of the detection of a PCR product (1650-bp) encoding for hemolysin BL; Lane 1, the GeneRuler™ mass ladders (3000, 2000, 1500, 1200-bp); Lane 2, *Bacillus* PB6; Lane 3, *Escherichia coli* ATCC 25922; Lane 4, *B. cereus* ATCC 49064; Lane 5, *B. cereus* ATCC 11778. No amplified band corresponding to a 1650-bp PCR product was detected in any of the lanes except for lanes 4 and 5.

15 Fig. 10 is a gel of the detection of a PCR product (1437-bp) encoding for non-hemolytic enterotoxin (Nhe). Lane 1, the GeneRuler™ mass ladders (3000, 2000, 1500, 1200-bp); Lane 2, *Bacillus* PB6; Lane 3, *Escherichia coli* ATCC 25922; Lane 4, *B. cereus* ATCC 49064; Lane 5, *B. cereus* ATCC 11778. No amplified band corresponding to a 1437-bp PCR product was detected in any of the lanes.

20 Fig. 11 is a gel of the detection of a PCR product (1400-bp) encoding for enterotoxin K (EntK); Lane 1, *Bacillus* PB6; Lane 3, *Escherichia coli* ATCC 25922; Lane 4, *B. cereus* ATCC 49064; Lane 5, *B. cereus* ATCC 11778; Lane 6, GeneRuler™ mass ladders (3000, 2000, 1500, 1200-bp); no amplified band corresponding to a 1400-bp PCR product was detected in any of the lanes.

25 Fig. 12 is a photograph showing the lack of antagonistic effect of *Bacillus cereus* (1) against *C. perfringens* ATCC13124 (2) and *C. difficile* ATCC9689 (3).

Fig. 13 is a photograph of the antagonistic effect of *Bacillus* PB6 against *Campylobacter jejuni* ATCC 33291.

Fig. 14 is a photograph showing the lack of antagonistic effect of *Bacillus cereus* against *Campylobacter jejuni* ATCC 33291.

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Detailed Description of Preferred Embodiments

PB6 is a proprietary bacterial strain that was isolated from nature and has not been genetically modified. Using a ribotyping technique this bacteria was identified as being a

strain of *Bacillus subtilis*. DNA:DNA hybridization studies indicate that *Bacillus* PB6 strain may more likely be a *Bacillus amyloliquefaciens*, which will be further described below.

As used in this specification, the term prophylaxis means a medical or public health procedure whose purpose is to prevent rather than treat or cure a condition. As used in this specification, the term treatment means a medical or public health procedure whose purpose is to treat or cure a condition. As used in this specification, the term synergistic compound means a compound which enhances the prophylactic effect or treatment efficacy of a *Bacillus* bacterium administered for the prophylaxis or treatment of a disease or health condition. As used in this specification, lipopeptides are molecules that contain both lipids and proteins and include surface-active molecules containing several amino acids and one or more fatty acids. Surfactins, iturins, mycosubtilins, baillomycins, bacillopeptins, fengycins, and plipastatins are examples of lipopeptides.

15 EXAMPLE 1 – EFFICACY OF *BACILLUS* PB6 AGAINST *C. DIFFICILE*, AAD AND CDAD

The antagonistic properties of *Bacillus* PB6 were tested against *C. perfringens* ATCC13124 and *C. difficile* ATCC9689.

20 *Bacillus* PB6 had antagonistic effect against *C. perfringens* ATCC13124 and *C. difficile* ATCC9689. A clear zone was observed at the intersections of the streak-lines on the plate for both species. An example of the test plate is depicted in Fig. 1.

Bacillus PB6 also had antagonistic effect against *C. difficile* NAP1/027. This *C. difficile* strain is linked to several highly dangerous outbreaks and shows resistance to antibiotics. An example of the test plate is depicted in Fig. 2.

25 In order to determine the antimicrobial effect of the secondary metabolites of *Bacillus* PB6, the bacteria was fermented and the fermentation product was extracted by diethyl ether. The organic layer was separated, concentrated *in vacuo* and redissolved in DMSO for screening.

30 The minimum inhibitory concentration (MIC) of the extract was 2.5 – 5 µg/ml against *C. perfringens* and 5 – 10 µg/ml against *C. difficile* (Table 1).

Table 1: Results of the screening with the crude extract of *Bacillus* PB6 against *C. perfringens*, *C. difficile* and *C. jejuni*

	MIC (µg/ml)		
	<i>C. perfringens</i> ATCC13124	<i>C. difficile</i> ATCC9689	<i>C. jejuni</i> ATCC 33291
Crude extract	2.5-5	5-10	25-100

It was also proven that *Bacillus* PB6 inhibits the growth of *Campylobacter jejuni* *in vitro*. An example of the test plate is shown in Fig. 3. The MIC of an ether extract of the fermentation product of *Bacillus* PB6 against *C. jejuni* was 25-100 µg/ml.

Campylobacter jejuni and *Helicobacter pylori* are very closely related and therefore it is probable that *Bacillus* PB6 is also active against *Helicobacter pylori*.

Furthermore, in literature can be found that *Bacillus* bacteria (e.g. *Bacillus subtilis*) possess activity against *Helicobacter pylori*.³³

Further research of the crude extract showed that the molecule responsible for the activity against *Clostridium* was the cyclic lipopeptide surfactin (Fig. 4).

When the activity of pure surfactin (either purified from our fermentation or purchased from Sigma) was determined against *Clostridium*, a higher MIC was found. The MIC against *C. perfringens* proved to be 10 – 25 µg/ml. It is remarkable that the pure active compound is less active than the crude fermentation extract. This is likely due to a co-factor(s) present in the extract that enhances the activity of surfactin. This has been shown by experiments where the MIC of surfactin was compared with the MIC of surfactin in combination with an inactive compound. This inactive compound was isolated from the same extract we isolated surfactin from. The MIC of this combination was between 1 and 10 µg/ml.

Experiments have shown that, when the filtrates of *Bacillus* PB6 were treated with pepsin and trypsin, the activity against *Clostridium sp.* decreased significantly. Since pepsin and trypsin are enzymes produced in the mucosal lining of the stomach and the pancreas, it is clear that oral administration of surfactin will lead to a significant loss of the activity. Other experiments, where surfactin has been incubated at 37°C for 30, 60 and 90 minutes with 0.1 N HCl, showed that acidic conditions (as in the stomach) lead to a loss in activity (Table 2). Therefore, administration of *Bacillus* PB6 (eventually as spores) is more effective to obtain effective concentrations of surfactin or other lipopeptides in the intestine.

Table 2: Results of the influence of acidic incubation on the antibacterial activity of surfactin against *Clostridium perfringens*

	MIC (µg/ml) (<i>C. perfringens</i>)			
	No HCL	30' HCl	60' HCl	90' HCl
surfactin	1-10	10-25	25-50	25-50

Besides the production of antimicrobial secondary metabolites, *Bacillus* PB6 spores are able to germinate in the intestine and thus can suppress pathogens by competitive exclusion.

Bacillus, more specifically *Bacillus subtilis*, remains one of the most potent and beneficial of all health-promoting and immune-stimulating bacteria. According to several clinical studies documented in medical research reports, the cell wall components of ingested *Bacillus* are able to activate nearly all systems of the human immune defense, including the activation of at least three specific antibodies (IgM, IgG and IgA) which are highly effective against many of the harmful viruses, fungi and bacterial pathogens which regularly attempt to invade and infect the human system. *Bacillus subtilis* has also been shown to stimulate B and T lymphocytes and macrophages. Also evidence has been provided that *Bacillus subtilis* spores may exert an immunomodulatory effect *in vivo*. And an increased response of plaque-forming cells to T dependent antigens has been described after exposure to spores as well as an enhancement of different phagocytes' functions.^{34,35,36,37,38,39,40,41}

In a study (Table 3) an elevated degree in phagocytosis was observed in broilers fed with different levels of *B. amyloliquefaciens* PB6 compared to the antibiotic and negative controls.

From this we can conclude that also *Bacillus amyloliquefaciens* possesses immuno-stimulating properties.

Table 3. Influence of *Bacillus* PB6 on immune response in male broilers

Treatment	Phagocytosis
Negative control	6.59
Positive control (100 mg/kg zinc bacitracin)	6.23
PB6 (10 ⁷ CFU/T)	11.82
PB6 (10 ⁸ CFU/T)	8.85

The filtrates from *Bacillus* PB6 were evaluated for the presence of hemolytic, non-hemolytic enterotoxins and enterotoxin K using commercially available immunological

assays (TECRA and Oxoid). The same filtrates were further subjected to cytotoxicity tests on Vero and HEp-2 cell lines. Finally, PCR-based methods were used to confirm for the presence of genes with possible enterotoxigenic capacity in *Bacillus* PB6. There was no immuno-cross reactivity observed with Hbl or Nhe enterotoxins and antibodies in the two commercial immunoassays. No cytotoxicity was also observed with the Vero and HEp-2 cell assays. *Bacillus* PB6 strain does not produce the hemolytic, non-hemolytic enterotoxins and enterotoxin K under the same conditions that allowed detection for a known toxigenic strain of *B. cereus*.

The *in vivo* toxicity of *Bacillus* PB6 has also been tested. Therefore a spray-dried product containing 10^{10} cfu/g of *Bacillus* PB6 was made from the fermentation product.

Toxicity Study of Spray-dried *B. amyloliquefaciens* in Wistar Rats

A first study was designed and conducted to determine the acute oral toxicity of the spray-dried *Bacillus* PB6 product (10^{10} cfu/g) in Wistar rats. A total of 5 male and 5 female animals were administered an oral dose of 5000 mg/kg (as a suspension in double distilled water and using a dose volume of 10 ml/kg). The control group consisted of 5 male and 5 female animals that received double distilled water at a dose of 10 ml/kg. No mortality was observed in the 5000 mg/kg treated animals. 50 % of the animals receiving the *Bacillus* PB6 product were more active in comparison with the control group. None of the animals treated with *Bacillus* PB6 suffered from diarrhea. After necroscopy, no pathological changes were seen in any organ in both groups. Thus the maximum non-lethal dose (LD_0) and LD_{50} of the orally administered *Bacillus* PB6 product in Wistar rats was found to be greater than 5000 mg/kg.

Conclusion: Maximum non-lethal dose (LD_0) and LD_{50} of *B. PB6* dry (10^{10} cfu/g) in Wistar rat by oral route was found to be greater than 5000 mg/kg.

Repeated Dose 28-day Oral Toxicity of *B. PB6* in Wistar Rats

This study was designed and conducted to determine repeated dose (28 days) oral toxicity of *B. PB6* (10^{10} cfu/g) in Wistar rats. In each group 6 male and 6 female animals were administered oral doses of 250, 500 or 1000 mg/kg for 28 days. The control vehicle group also consisted of 6 male and 6 female animals, which received doubled distilled water at a dose of 10 ml/kg for 28 days. These groups were sacrificed on day 29.

The study also consisted of two reversible groups for control vehicle and high dose which each had 6 male and 6 female animals. The high dose group received medication up

to day 28 and was without treatment from day 29 to 42 and sacrificed on day 43. The control vehicle group received distilled water at a dose of 10ml/kg up to day 28 and was without treatment from day 29 to day 42 and sacrificed on day 43.

Conclusion: No Observed Adverse Event Level (NOAEL) for *B. PB6* (10^{10} cfu/g) in 28 days toxicity trials in rats is found to be greater than 1000 mg/kg.

Local Irritancy Study (Dermal and Eye) of *B. PB6* in New-Zealand White Rabbit

A total of 3 New-Zealand White Rabbits (either sex) – Same rabbits were used for dermal and eye irritation in this study. A minimum gap of 5 days was kept between both studies. 1 g of *B. PB6* (10^{10} cfu/g) was applied as a paste to the skin for dermal irritation and 100µl of 10% *B. PB6* suspension instilled into left eye for eye irritation.

Test Item PB6 paste was applied to skin at dorso-lateral area after removal of hairs in 3 rabbits. Suspension of 10% PB6 was instilled in the eye of 3 rabbits. The test item was removed from skin of the animals 4 hours post-application. After instillation of test item in left eye, the eyelids were held together for 2-3 seconds.

The rabbits were observed and scored for dermal irritancy at 1, 24, 48 and 72 hours after removal of test item PB6. For eye irritancy scoring was carried out at the same time points post-instillation of PB6 suspension.

Conclusions: Dermal : No irritation was observed, when 1g of *B. PB6* (10^{10} cfu/g) was applied as a paste.

Eye : No irritation was observed, when 100µl of 10% suspension of *B. PB6* (10^{10} cfu/g) was instilled.

Erythrocyte Micronucleus Assay of *B. amyloliquefaciens* PB6 in Mice

This study was designed and conducted to detect the damage induced by the test substance to the chromosomes or the mitotic apparatus of Swiss albino mice. A total of 5 male and 5 female animals were administered oral dose of 2500 and 5000mg/kg, the control vehicle group consisted of 5 male and 5 female mice which received double distilled water at a dose of 10ml/kg orally. The positive control group (5 male + 5 female) received cyclophosphamide orally at a dose of 40 mg/kg.

The animals were sacrificed by excess of CO₂ at respective time points (control group, 2500 mg/kg PB6 cyclophosphamide at 24h and 5000 mg/kg PB6 at both 24 and 48 h) and both femora were removed and bone marrow smears made on slides, stained with

Giemsa and May-Greunwald stain, viewed under microscope for the incidence of micronucleus by counting 2000 immature erythrocytes.

The percentage of immature among total (immature + mature) erythrocytes is also determined for each animal by counting at least 200 erythrocytes.

- 5 Conclusion: *B. PB6* (10^{10} cfu/g) at a dose of 2500 & 5000 mg/kg did not significantly induce micro nucleated polychromatic erythrocytes in mice.

Determination of the *in vivo* efficacy of orally administered *Bacillus PB6* in the treatment of Golden Syrian hamsters suffering from CDAD.

10 Study design

- Forty-two male Golden Syrian hamsters were obtained from the National Centre for Laboratory Animal Sciences, NIN (Hyderabad, India). At the beginning of the treatment period, the animals were 12 to 14 weeks old. Upon their arrival at the test facility, the animals were given a complete clinical examination under the supervision of a veterinarian to ensure that they were in good condition. The animals were acclimatized to the study conditions for a period of at least 7 days. Body weights were recorded before allocation of the animals into the study groups at the start of the trial. The animals were housed individually in polycarbonate cages (290x22x140 mm, LxWxH). The animal room and test room conditions were set as follows: temperature: $22 \pm 4^\circ\text{C}$, relative humidity: $50 \pm 20\%$, light/dark cycle: 12hr/12hr (light 07.00-19.00) and ventilation: approximately 7 cycles/hour of filtered, non-recycled air. All animals had free access to Hamster Pellet Feed (NIN, Hyderabad) and Aquaguard purified water *ad libitum*.
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- Animals were allocated to one of seven study groups (A to G). In groups A to E *Clostridium difficile* associated diarrhea was induced by administering 10000 CFU's of *Clostridium difficile* ATCC 9689 orally on day 0, followed by a subcutaneous injection in the trunk region just behind the ears of clindamycin 100 mg/Kg on day 1. Group A received no further treatment.
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- Group B was treated once daily from days 2 to 6 with vancomycin 50 mg/Kg by oral gavage. Groups C, D and E were treated with PB6 at a dose of 1.5×10^8 , 1.5×10^7 and 1.5×10^6 CFU/Kg respectively, 3 times daily with 4 hours interdose (first dose at 9.30 am) from day 1 to 6 by oral gavage. On day 1 the first dose of PB6 was given 1 hour after the injection of clindamycin. In groups F and G the animals were given PB6 at a dose of 1.5×10^9 CFU/Kg, 3 times daily with 4 hours interdose (first dose at 9.30 am) from day 1 to 6 by oral gavage. Day 6 was the last day of treatment. Twice daily, up to day 15,
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observations were made for clinical signs and mortality. Signs of diarrhea were scored as being mild, moderate or severe. Body weights of animals were recorded on day 0, 7 and 14. For groups A to E faeces were tested on days 1,2 and 7 for the presence of *Clostridium* toxin A and B using Immunocards (Meridian life sciences). On day 1 fecal samples were taken before clindamycin administration. On days 2 and 7 fecal samples were taken between the second and third treatment dose.

Test preparations

A Culti-loop (Oxoid, Basingstoke, England) containing *Clostridium difficile* ATCC 9689 was inoculated according to the manufacturers instructions and the broth was diluted with saline to obtain 10000 CFU/ml. Clindamycin hydrochloride (Pharmacia, Puurs –Belgium) and vancomycin (Neon Laboratories, Bombay, India) were suspended in double distilled water to a concentration of 10 and 50 mg/ml respectively. Dose volumes were 10 and 1 ml/Kg bodyweight for clindamycin and vancomycin respectively. PB6 Dry (Kemin Consumer Care, Des Moines, USA), a *Bacillus* 'PB6' fermentation broth dried on a malto- and cyclodextrin carrier, was suspended in double distilled water to concentrations of 1.5×10^8 , 1.5×10^6 and 1.5×10^5 CFU/ml. Dose volume was 10 ml/Kg bodyweight. Fresh preparations were made prior to each administration. Clindamycin, vancomycin and PB6 were administered on the basis of the last individual body weight taken. The preparations were stirred vigorously before each dosing.

Results and discussion

About 6 hours after clindamycin was administered mild diarrhea was observed in 3 animals of the no treatment group and in 1 animal of the group to be treated with vancomycin. In groups C, D,E, where the animals received their first dose of PB6 about 1 hour after the administration of clindamycin, none of the animals showed any sign of diarrhea that same day. The number of animals suffering from diarrhea and the severity of diarrhea evolved differently between treatment groups the following 2 days (Table 4). All groups in which *Clostridium difficile* associated diarrhea was induced showed signs of diarrhea. The intensity of diarrhea was less in the group treated with vancomycin and the group treated with the high dose of PB6. In the no treatment group as well as in the low and mid dose PB6 groups the stool was very watery and whole of the abdominal area was wet.

Table 4. Number of animals with diarrhea and their score for days 1 to 3, in the groups in which CDAD was induced.

Treatment	Day 1		Day 2		Day 3	
	number	score	number	score	number	score
No treatment	3	+	6	+(1) ++ (5)	6	++ (5) +++ (1)
Vancomycin	1	+	2	++	2	+++
PB6 high	0		3	+(2) ++ (1)	4	+(1) ++ (3)
PB6 medium	0		5	+(4) ++ (1)	6	+(1) ++ (2) +++ (3)
PB6 low	0		5	++	6	+(1) ++ (2) +++ (3)

+: mild ++: moderate +++: severe

- 5 At the end of day 3 all animals of groups in which CDAD was induced were still alive but several of them were showing signs of severe diarrhea. On day 4, three days after clindamycin administration, the first animals died. At the end of the treatment period, on day 6, all hamsters in the no treatment group had died. Survival was highest in the vancomycin and PB6 high dose treatment groups, where 4 out of 6 had survived (Fig. 5).
- 10 On day 7 a decreased average body weight was observed in all groups in which CDAD was induced. There was a clear inverse dose response relation with PB6 concerning this decrease in weight (Table 5). The animals receiving low dose of PB6, on average lost 3 times more weight than the animals which received the high dose. Weight loss was least with vancomycin treatment. The average body weight in the two groups in
- 15 which CDAD was not induced had slightly increased over the same time period.

Table 5. Average body weight (g) and body weight gain (%).

Treatment	Average body weight (g)		Average body weight difference (%)
	Day 0	Day 7	
CDAD Induced			
No treatment	219.0 ± 5.1 (n=6)	*	*
Vancomycin	218.0 ± 4.6 (n=6)	215.8 ^b ± 1.7 (n=4)	-1.7 ^b ± 1.8
PB6 high	218.8 ± 4.0 (n=6)	206.5 ^c ± 5.7 (n=4)	-5.0 ^b ± 2.1
PB6 medium	217.8 ± 3.2 (n=6)	189.0 ^d ± 2.7 (n=3)	-12.9 ^c ± 0.7
PB6 low	217.2 ± 4.5 (n=6)	176.5 ^e ± 2.1 (n=2)	-17.9 ^d ± 1.2
No CDAD induced			
PB6 high	217.8 ± 2.3 (n=6)	230.0 ^a ± 7.6 (n=6)	5.6 ^a ± 3.1
PB6 high	219.2 ± 3.9 (n=6)	225.8 ^a ± 4.8 (n=6)	3.1 ^a ± 3.2

*no data, all animals had died

^{a,b,c,d,e} values within the same column not bearing a common superscript are significantly different (P<0.05, Least Significant Difference).

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The presence of clostridium toxin A and B in fecal samples of the animals in which CDAD was induced, was checked on days 1, 2 and 7. As expected, groups showing high mortality and a high decrease of average body weight also had a high percentage of animals tested positive for these toxins with again an inverse dose response relation with PB6.

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Table 6. Number of animals tested positive for the presence of *C difficile* toxins A or B in their faeces.

Treatment	Day 1		Day 2		Day 7	
	Positive	Tested	Positive	Tested	Positive	Tested
No treatment	3	6	5	6	*	*
Vancomycin	0	6	2	6	1	4
PB6 high	0	6	3	6	2	4
PB6 medium	0	6	3	6	2	3
PB6 low	0	6	5	6	2	2

*no data, all animals had died

5 Conclusion

This study was designed and conducted to evaluate the efficacy of *B. PB6* in treating CDAD in the hamster model of clindamycin-induced CDAD, a well-established and highly sensitive model for this infection. Hamsters that were not treated after induction of CDAD all got severe diarrhea, lost weight and died within five days.

- 10 Treatment with PB6 resulted in a dose related response. Diarrhea, body weight reduction and mortality were least with the highest dose of PB6. At the stop of treatment the highest dose of PB6 showed to have been equally efficient as vancomycin in helping hamsters to survive clindamycin-induced CDAD.

15 Evaluation of the Efficacious Components of the Orally Administered *Bacillus* PB6

- The new dietary concept of probiotics (such as inulin) is gaining popularity among nutritionists, physicians, food manufactures and consumers. The most widely accepted definition of a probiotic food is: "A probiotic is a selectively fermented ingredient that
- 20 allows specific changes, both in the composition and activity of the gastrointestinal microflora that confers benefits upon the host well-being and health." To be classified as a probiotic ingredient in food products, that ingredient must 1) resist digestion (hydrolysis) by alimentary enzymes in the stomach and small intestine; 2) enter the colon chemically intact and subsequently undergo partial or complete fermentation; and 3) stimulate active
- 25 growth and/or activity of health-stimulating intestinal bacteria. Health benefits of probiotic-stimulated bacteria growth are wide-ranging and include such benefits as positive effects on colon cancer and pathogens, decreased triacylglycerols, increased

absorption of Ca and Mg, and increased stool frequency and fecal weight. Inulin is a polydisperse $\beta(2 \rightarrow 1)$ fructan with chain lengths ranging from 2 to 60 units. The unique linkage between the fructose molecules of inulin and oligofructose distinguish them from typical carbohydrates. They resist digestion by human alimentary enzymes and absorption in the small intestine but are hydrolyzed and fermented by colonic microflora and therefore classified as a probiotic dietary fiber.^{42,43,44,45,46}

The fact that probiotics stimulate the active growth and/or activity of health stimulating intestinal bacteria, make them very interesting products to be administered together with probiotics such as *Bacillus* PB6.

In addition, *Bacillus amyloliquefaciens* PB6 does not inhibit the growth of "healthy" bacteria from the gut flora such as *LactoBacillus* spp. and *Bifidobacterium* spp.

Materials and Methods

Identification of isolated PB6 bacteria

The PB6 *Bacillus* strain isolated is a Gram-positive rod and possesses catalase. Malachite Green staining confirmed that this microorganism possesses endospores and is a spore-former. This microorganism is also a "swarmer" as upon extended incubation period, it tends to spread over the entire agar surface. In terms of relatedness to known *Bacillus* spp., the *Bacillus* strain isolated has 92.0 % ID. Based on the API biochemical profiles, the PB6 *Bacillus* strain was putatively identified as *B. subtilis*. The PB6 *Bacillus* strain was therefore deposited as ATCC - PTA 6737 and named there *Bacillus subtilis*. United States Patent Application Serial No. 10/306,365, filed November 27, 2002, is incorporated herein by this reference.

16 rRNA gene sequencing

The nearly complete 16 rRNA sequence of *Bacillus* PB6 was completed and aligned (SEQ ID NO.1, Fig. 6).

GyrA sequencing

Partial gyrA sequences of PB6 obtained by two different groups are shown in Fig. 7 (SEQ ID No. 2) and Fig. 8 (SEQ ID NO. 3).

DNA:DNA hybridizations

Hybridizations were performed under stringent conditions (at 40 °C) according to a modification of the method described by Ezaki *et al.*⁴⁷ The DNA homology percentages are the mean of minimum 4 hybridizations. The value given between brackets is the

difference between the reciprocal values. With this technique, the average standard deviation is 14 units.⁴⁸ The results are presented in Table 7.

Table 7 - %DNA homology

	% DNA homology		
<i>Bacillus</i> PB6	100		
<i>B. amyloliquefaciens</i> LMG 9814 ^T	75 (13)	100	
<i>B. velezensis</i> LMG 22478 ^T	90 (6)	80 (8)	100

A DNA homology above 70%, the generally accepted limit for species delineation,⁴⁹ is found between *Bacillus* PB6, LMG 9814^T and LMG 22478^T.

From these results, it appears that *B. velezensis* and *B. amyloliquefaciens* belong to the same genospecies and are therefore subjective synonyms such that in applying nomenclatural rule 42 the oldest legitimate name should be retained, i.e. *B. amyloliquefaciens*; and that *Bacillus* PB6 (ATCC – PTA 6737) may more properly be categorized to the species *B. amyloliquefaciens*.

Testing of antagonistic properties of *Bacillus amyloliquefaciens* by the streak-line assay

Bacillus amyloliquefaciens PB6 was inoculated as a straight line on Tryptone Soy blood plates (Oxoid, Belgium), after 24 hours of incubation at 37°C in aerobic conditions, the different indicator strains were inoculated perpendicularly to the *Bacillus* PB6 culture. Plates inoculated with *Clostridia* species were incubated in anaerobic conditions using Anaerogen Pak (Oxoid, Belgium). After overnight incubation at 37°C, antagonistic effects were evaluated by the appearance of clear zones surrounding the junctions of the streak-lines indicating the inhibitory effects of one organism against the other.

Extraction and antimicrobial screening of the secondary metabolites of *Bacillus* PB6.

Bacillus PB6 was grown on Trypton Soya Agar plates supplemented with 5% sheep blood (Oxoid, Belgium) for 24 hours at 37°C. This culture was used to inoculate 100ml Tryptic Soy broth supplemented with 0.6% yeast extract (Oxoid, Belgium). After incubation for 24 hours in a shaking incubator (100 rpm) at 37°C, the broth was mixed (3 times) with equal amounts of diethyl ether (Acros, Belgium). After extraction of the metabolites, both layers were separated and the ether fraction was collected and centrifuged at 4000 rpm for 5 min.

Afterwards, the solvent was removed *in vacuo* using a rotating evaporator. The residue was weighed and dissolved in dimethylsulfoxide (Acros, Belgium) resulting in a final concentration of 10000µg/ml crude extract. Further dilutions (500, 250, 100, 50, 25 and 10µg/ml) were made in a mixture of DMSO/water with ratio 1/11. Finally, 25µl of each dilution was pipetted into the wells of the microtiter plates (Labsystems, Finland).

The bacterial strains *Clostridium perfringens* ATCC13124 (C1600L, Oxoid, Belgium) and *Clostridium difficile* ATCC9689 (C1610L, Oxoid, Belgium) were purchased as freeze-dried culti-loops and brought into culture according to the manufacturer's instructions. From both cultures a McFarland standard ($A_{625nm} = 0.100$) was prepared in Anaerobic Basal broth (Oxoid, Belgium). 250µl of this standard was added to 10ml of fresh Anaerobic Basal broth, and 225µl of this medium was pipetted into the wells. Yielding a final cell density of 5×10^5 cfu/ml in each well. The microtiter plates were incubated in anaerobic conditions for 18 hours (*C. perfringens*) and 48 hours (*C. difficile*) using Anaerogen Compact (Oxoid, Belgium) in an airtight plastic bag. Before and after the incubation period, optical density (OD) of each well was measured using the Bioscreen C analyzer (Labsystems, Finland). White light (Wide Band) was used to measure the OD. Tests were done in duplicate and also control of the medium, medium plus inoculum (negative control) and a positive control; vancomycin (Fluka, Belgium) at three concentrations (0.1, 0.5 and 1.0µg/ml) was included in the test batch. Minimum inhibition concentration (MIC) was defined as the lowest concentration where no growth occurred or where no increase of OD was detected.

Determination of the enhancing effect of a co-factor on the activity of surfactin

The ether extract of the *B. amyloliquefaciens* PB6 fermentation product was separated by preparative TLC (Kieselgel 60, 20 x 20 cm, 2mm layer thickness). The eluent used was hexane/acetone (30/70). The surfactins were isolated in the zone with Rf 0 to 0.33. The MIC against *C. perfringens* of the surfactins isolated from this zone was between 10 and 25 µg/ml. In another zone (Rf 0.33 to 0.76) one or more products were found that were not active against *C. perfringens* (MIC > 100 µg/ml). When a combination was tested of the products of both zones, with a final concentration of both extracts of 10 µg/ml each, there was no growth of *C. perfringens*. The MIC of surfactins decreased to below 10 µg/ml when combined with these products that were not active on their own.

Determination of the effect of acidic treatment on the antimicrobial activity of surfactin

A solution was prepared of 750 µg/ml surfactin (Sigma) in acetonitrile/ 0.1 N HCl (1/1; v/v). 0.5 ml of this solution was incubated at 37 °C for 30', 60' or 90', after which 0.25 ml of a 0.1 N NaHCO₃ solution was added and the mixture was mixed using a vortex shaker. All solutions were screened for activity against *Clostridium perfringens*.

Determination of enterotoxin production by *Bacillus* PB6

Bacterial strains and culture conditions. *Bacillus* PB6 was grown in 100-ml volume of Tryptic Soy Broth (Becton Dickenson and Company, Cockeysville, MD) supplemented with 0.6% yeast extract (Oxoid Limited, England) (TSBYE) and incubated at 37°C in a shaker incubator set at 100 rpm. One toxin-producing strain of *B. cereus* ATCC 11778 was also grown in TSBYE at 37°C in a shaker incubator. Similarly, non toxin-producing strains of *B. cereus* ATCC 49064 and *Escherichia coli* ATCC 25922 were also grown in TSBYE at 37°C under aerobic conditions. All bacterial strains used in this study were transferred weekly to fresh TSBYE and then kept in a 4°C refrigerator as working culture. Freshly grown strains were re-suspended in 40% glycerol and kept at –80°C freezer for long-term storage.

Enterotoxin production. A 1-ml volume of overnight test culture was inoculated into 50 ml of Brain Heart Infusion (BHI) supplemented with 1% glucose (BHIG) and incubated in a shaking incubator (100 rpm) for 6 h at 32°C³¹. Bacterial cells were precipitated by centrifugation at 5000 X g for 10 min and the supernatant was collected for cytotoxicity studies of Vero cells³¹. The proteins in the supernatant were then concentrated ten-fold using up to 80% saturated ammonium sulfate solution (561g per liter)³³. After centrifugation at 10000 X g for 20 min, the supernatant was decanted and protein-pellet was re-suspended in 2.5 ml of phosphate buffer (20 mM; pH 6.8). Residual salts of ammonium sulfate were removed by dialysis against the same buffer at 4°C for 6 h. The final volume of the dialyzed protein solution was adjusted to one-tenth of the original volume (5 ml) using the phosphate buffer (20 mM; pH 6.8)³¹.

Emetic toxin production. A 1-ml volume of overnight test culture was inoculated into 50 ml of Brain Heart Infusion (BHI) supplemented with 1% glucose (BHIG) and incubated for 6 h at 32°C in a shaker incubator set at 250 rpm. Bacterial cells were precipitated by centrifugation at 2000 X g for 10 min at 4°C³¹. The supernatant was collected and then autoclaved at 121°C for 15 min to remove heat-labile enterotoxins²⁰.

The heat-treated filtrate with possibly the heat-stable emetic toxin was collected for the vacuolation assay on HEp-2 cells²⁰.

Preparation of Vero and HEp-2 cells. African green monkey kidney cells (Vero) or HEp-2 (human carcinoma of the larynx) were maintained as monolayer cultures in 30 ml of Medium 199 with Earle's modified salts (MEM) containing 2 mM L-glutamine, 10 mM sodium bicarbonate, 1% non-essential amino acids, 100 UI/ml penicillin, 0.1 mg/ml streptomycin, and 3 ml fetal calf serum (10%). The leucine-free medium (MEM) was prepared on the basis of minimum essential medium (Gibco), which also contained 1.8 mM CaCl₂, 0.4 mM MgCl₂, 5.0 mM KCl, 0.12 M NaCl, 3.2 mM NaH₂PO₄, and 20 mM Hepes (pH 7.7). The Vero or HEp-2 cells were incubated in MEM at 5% CO₂ at 37°C. Confluent monolayer cultures of Vero or HEp-2 cells were sub-cultured by discarding the media before washing with 5 ml of PBS (pH 7.7). Vero or HEp-2 cells were then detached from the culture flask by the addition of 2 ml of trypsin solution (0.25% trypsin; 0.025% EDTA). The levels of loosen Vero or HEp-2 cells were determined microscopically before the addition of MEM medium (8 ml) to prevent further effect of trypsin. A 5-ml aliquot of freshly trypsinised Vero or HEp-2 cells were then transferred to new culture flask containing 15 ml of MEM for incubation at 37°C under 5% CO₂.

TECRA[®] *Bacillus* Diarrheal Enterotoxin (BDE) Visual Immunoassay. All components of the test kit (TECRA International Pte Ltd, Chatswood, NSW, Australia) were kept at 20 – 25°C prior to testing of samples for BDE. As stated by the manufacturer, microtiter wells containing high affinity antibodies specific for BDE were pre-soaked with wash solution, provided in the kit and allowed to stand for 10 min at 20 – 25°C. The wells were emptied before aliquots containing 200-µl volume of test samples and controls (positive and negative) were transferred into individual wells. The wells were incubated at 37°C for 2 h. The wells were washed 4 times before an aliquot of 200 µl of conjugate was added to each well and incubated at 25°C for 1 h. Each well was washed 5 times before 200 µl of substrate was added to each well and incubated at 25°C for 30 min. After 30 min, the colorimetric development for each well was compared against the TECRA Color Card provided in the test kit.

Oxoid *Bacillus cereus* Enterotoxin Reversed Passive Latex Agglutination (BCET-RPLA). The test was developed for the detection of diarrheal enterotoxin of *Bacillus cereus* by reversed passive agglutination (RPLA) (Unipath, Basingstoke, UK). Polystyrene latex particles are sensitized with purified antiserum from rabbits immunized

with diarrheal enterotoxin from *B. cereus*. The kit also provided the both positive (enterotoxin) and negative (latex particles without specific *B. cereus* anti-enterotoxin) controls. Aliquots containing 25 µl of diluent and test or control (positive and negative) samples were dispensed successively into 2 different sets of V-well microtiter plates.

- 5 Solutions containing 25 µl of sensitized latex and latex control were then dispensed into the first and second set of V-well microtiter plates, respectively. Each well was examined for agglutination after 20 – 24 h of incubation at 25°C.

- Measurement of cytotoxicity. Freshly trypsinised Vero cells were re-suspended in 30 ml of leucine-free medium (MEM). One milliliter of the Vero cell suspension was
10 transferred to each of the 24 wells (~5 X 10⁶ cells per well). Cells were washed once with 1 ml of leucine-free MEM and incubated for 2 h at 37°C under 5% CO₂. After 2 h, the growth medium was removed and each well was washed once with 1 ml of leucine-free MEM. A 1-ml volume of preheated (37°C) leucine-free MEM was added to each well, followed by 50 µl of supernatant or filtrate of *Bacillus amyloliquefaciens* PB6, *B. cereus*
15 ATCC 11778, *B. cereus* ATCC 49064 and *E. coli* ATCC 25922 immediately thereafter. The inoculated wells were incubated at 37°C with 5% CO₂ for 2 h. After 2 h of incubation, supernatant or filtrate-treated Vero cells were washed once with 1 ml of pre-heated (37°C) leucine-free MEM. A volume of 300 µl of solution containing radioactive-labeled isotope (16 µl ¹⁴C-leucine in 8 ml of leucine-free MEM) (Perkin Elmer Asia,
20 Singapore) was added to each of the well. ¹⁴C-leucine labeled Vero cells were incubated at 37°C without CO₂ for 1 h and thereafter the growth medium was discarded. Subsequently, 1-ml aliquots of solution containing 5% trichloroacetic acid were added to each well containing ¹⁴C-leucine labeled Vero cells before incubating at 25°C for 10 min. After 10 min of incubation, the contents in each well were washed twice with 1 ml of 5%
25 trichloroacetic acid-solution. A 300-µl volume of 0.1 M KOH was then added to each well and incubated at 25°C for another 10 min. After 10 min of incubation, 2-ml volume of scintillation liquid was added to each well. Finally, all contents from each well were transferred into scintillation tubes. All scintillation tubes were agitated for 1 min before performing radioactivity counts. The percent inhibition of ¹⁴C leucine uptake by Vero
30 cells is calculated using the following formula.

Percent inhibition of ¹⁴C leucine uptake = [(cpm for Vero cells without toxin added – cpm for test sample)] / [cpm for Vero cells without toxin added] X 100%. The cpm used for calculating percent inhibition of ¹⁴C leucine uptake by Vero cells has to be subtracted by

the value for background counts (~ 30 - 60 cpm). No toxin is present if the inhibition of ^{14}C leucine uptake is less than 20% after the ten-fold concentration. Each assay was conducted in duplicates. The results are presented in Tables 8 and 9.

5 Table 8: Effect of bacterial filtrate on Vero and HEp-2 Cell Lines

Bacteria	Vero cells		HEp-2 cells	
	Cytotoxic effect		Vacuole response	
	1 h	24 h	6 h	24 h
<i>Bacillus amyloliquefaciens</i> PB6	- ^a	-	-	-
<i>Escherichia coli</i> ATCC 25922	-	-	-	-
<i>Bacillus cereus</i> ATCC 11778	-	-	-	-
<i>Bacillus cereus</i> ATCC 49064	++ ^b	++++	-	+++

^a "-" indicates no cytotoxic effect or presence of vacuoles observed.

^b "+" indicates destruction of Vero cell or vacuole production in HEp-2 cells have occurred.

10 Table 9: % inhibition of ^{14}C -leucine isotope uptake

Concentrated filtrates of bacteria	% inhibition ^a
<i>Bacillus amyloliquefaciens</i> PB6 ^b	1
<i>Escherichia coli</i> ATCC 25922 ^c	1
<i>Bacillus cereus</i> ATCC 11778 ^d	17
<i>Bacillus cereus</i> ATCC 49064 ^e	100

^aToxin from the tested strain is considered negative if the inhibition of ^{14}C leucine uptake is less than 20% after ten-fold concentration. (SCAN report).

^bTest microorganisms,

^cnegative and ^epositive controls and

^dnon-toxin producing strain.

HEp-2 Cells Vacuolation Assay. A 25- μl volume containing filtrates from test and control cultures were serially diluted (2-fold) in 0.15 M NaCl solution and dispensed across wells of a 96-well tissue culture plate (Gibco Ltd, Uxbridge, UK). Volumes containing 100 μl of freshly trypsinized HEp-2 cells were added to each well and incubated for 24 h at 37°C. All vacuolation assays were performed in duplicates. Microscopic examination for the presence of vacuole formation in HEp-2 cells was conducted at the 6th and 24th hour-intervals.

PCR-based methods. PCR primers were developed for the non-haemolytic enterotoxin (*NheB-nheC*)^{25,26}, hemolysin BL (*HblD-hbLA*)^{10,11}, and enterotoxin K (*EntK*)^{27,28}. The sequences of the *NheB-nheC* primers were 5' CGGTTTCATC-TGTTGCGACAGC 3' and 3'GTCCTCGTGTTCGTCTTC- AGC 5'. The primer sequences for *HblD-hbLA* were 5' CGCT- CAAGAACAAAAAGTAGG 3' and 3' TCCCTAATGT- CTAAATGTTTCCTC 5'. The forward and reverse primers for *EntK*

were 5'GAATTACGTTGGCGAATC3' and 3'CGGG- CGGATGGGA 5', respectively. Both sample PCR and positive control tubes containing DNA of *Bacillus amyloliquefaciens* PB6 and toxigenic strains of *B. cereus* ATCC 11778 and *B. cereus* ATCC 49064 were placed into a Perkin-Elmer thermal cycler (GeneAmp PCR System 9600, Perkin-Elmer Corp., Norwalk, CT) with initial set point temperature of 90°C. The conditions for thermal cycling were 1 cycle at 94°C for 2 min followed by a 38-cycle temperature cycling routine of 94°C for 15 s, and 70°C for 3 min. Following amplification, a final extension at 72°C for 7 min was performed. The PCR amplification step required approximately 3 h to complete. Aliquots (15 µl) of each reaction product and 15 µl DNA standard ladder (500, 1031, 2000 and 3000 base-pairs) were then electrophoresed at 180 volts for 1 h through a 2% agarose gel (Life Technologies Inc., Gaithersburg, MD) containing 0.1 µg/ml ethidium bromide (Sigma Chemical Co., St. Louis, MO). After electrophoresis, all gels were viewed using a UV-transilluminator (wavelength - 302 nm) (UVP Model White /2 UV; UVP Inc., Upland, CA) and then photographed using a Polaroid MP-4 Camera (Polaroid Corp., Cambridge, MA)(See Fig. 9, Fig. 10 and Fig. 11).

EXAMPLE 2 - DETERMINATION OF THE ANTIMICROBIAL PROPERTIES
OF METABOLITES OF DIFFERENT PROBIOTICS AND *BACILLUS*
AMYLOLIQUEFACIENS PB6 AGAINST *CLOSTRIDIUM PERFRINGENS* AND
CLOSTRIDIUM DIFFICILE VIA BROTH MICRODILUTION METHOD

In this study we investigated the anticlostridial properties of the metabolites from *Bacillus* PB6 and four commercially available probiotics: Bactisubtil® (Sanofi – synthelabo), Pcenterol® (Biodiphar), Bioplus 2B (Miavit GmbH) and Biosporinum (Dniprofarm). Small-scale fermentations were setup with the species isolated from the different probiotics. Ether extracts of the fermentation broth, containing the metabolites, were screened against *C. perfringens* ATCC13124 and *C. difficile* ATCC9689 using broth microdilution method. The metabolites of *B. PB6* showed significant anticlostridial properties. Minimum inhibition concentrations were situated between 2.5 and 5.0µg/ml towards *C. perfringens* and between 5.0 and 10.0µg/ml towards *C. difficile*. The ether extracts of the fermentation from Bactisubtil®, Pcenterol®, Bioplus 2B and Biosporinum did not have any significant antibacterial activity against both clostridia species.

Although *Clostridium* species are ubiquitous in nature, their principle habitats are the soil and the intestinal tracts of many animals and humans. The widespread occurrence

of *C. perfringens*, including its spores, in soil samples almost guarantees the frequent presence of this organism on surfaces exposed to dust contamination, including many food items.⁵⁰ *C. perfringens* is also the species most commonly isolated from human clinical specimens, excluding faeces. It is encountered in a wide variety of clinical settings ranging from simple contamination of wounds to traumatic myonecrosis, intra-abdominal sepsis, intravascular haemolysis, aspiration pneumonia, necrotising pneumonia etc.⁵¹

C. difficile is a major cause of antibiotic-associated diarrhea (AAD) and is also the most frequently identified cause of hospital-acquired diarrhea. In *C. difficile*-associated disease (CDAD), the primary initiating event involves the disruption of the protective intestinal flora during treatment with antibiotics. As the level of antibiotic drops below inhibitory concentrations, nosocomial pathogens such as *C. difficile* are able to grow. Colonization occurs by the faecal-oral route, ingested spores survive the gastric acid barrier and start to germinate in the colon.^{52,53} Toxigenic as well as nontoxigenic isolates are capable of forming spores and existing in the hospital environment. As a result, either type can infect the colon and utilize the nutrients that are available because the lack of competition by the normal flora. Whether the organism attaches to the colonic wall is not clear, but it is more likely that the organism grows throughout the lumen of the colon. Toxigenic strains produce and release toxins A and/or B as the cells grow and lyse. This activity, along with the inflammatory response, result in the histopathological events leading to *C. difficile*-associated diarrhea and colitis.^{54,55,56}

A "probiotic" by generally accepted definition, is a "live microbial" feed or food supplement which beneficially affects the host by improving its intestinal microbial balance. But how does a probiotic work? The effect of probiotics on the intestinal ecosystem impacts in some beneficial way on the consumer. A number of potential benefits arising from changes to the intestinal milieu through probiotics have been proposed, including: increased resistance to infectious diseases, particularly of the intestine, decreased duration of diarrhea, reduction in blood pressure, reduction in serum cholesterol concentration, reduction in allergy etc.⁵⁷

The comparative study described in this paper was set up in order to compare the antimicrobial properties of different probiotic fermentation extracts towards clostridia species

Methods and Materials

Bacillus PB6 and the *Bacillus cereus* strain isolated from Bactisubtil[®] (Sanofi-synthelabo) were grown on Trypton Soya Agar plates supplemented with 5% sheep blood (Oxoid, Belgium) for 24 hours at 37°C. The feed probiotic Bioplus 2B contained two
5 different species, *Bacillus licheniformis* DSM5749 and *Bacillus subtilis* DSM5750. Both species were also grown on Trypton Soya Agar plates supplemented with 5% sheep blood (Oxoid, Belgium) for 24 hours at 37°C. A mixture of these cultures was used to inoculate 100ml Tryptic Soy broth supplemented with 0.6% yeast extract (Oxoid, Belgium). Biosporinum, also contains two species, i.e. *Bacillus licheniformis* and *Bacillus subtilis*.
10 This product is marketed as lyophilised cultures in glass vials. The content of one vial was resuspended in broth, this mixture was used to inoculate 100ml Tryptic Soy broth supplemented with 0.6% yeast extract (Oxoid, Belgium). After incubation of all probiotics for 24 hours in a shaking incubator (100 rpm) at 37°C, the broth was mixed (3 times) with equal amounts of dichthyl ether (Acros, Belgium). After extraction of the metabolites, both
15 layers were separated and the ether fraction was collected and centrifuged at 4000 rpm for 5 min.

Afterwards, the solvent was removed *in vacuo* using a rotating evaporator. The residue was weighed and dissolved in dimethylsulfoxide (Acros, Belgium) resulting in a final concentration of 10000µg/ml crude extract. Further dilutions (500, 250, 100, 50, 25
20 and 10µg/ml) were made in a mixture of DMSO/water with ratio 1/12. Finally, 25µl of each dilution was pipetted into the wells of the microtiter plates (Labsystems, Finland). *Saccharomyces boulardii* used in Pcenterol[®] (Biodiphar) was grown on Sabouraud dextrose agar (Oxoid, Belgium) for 48 hours at 37°C. This culture was used to inoculate 100ml Sabouraud liquid medium (Oxoid, Belgium). After incubation for 2 days in a
25 shaking incubator (100 rpm) at 37°C, the same procedure was followed to extract the metabolites.

The bacterial strains *Clostridium perfringens* ATCC13124 (C1600L, Oxoid, Belgium) and *Clostridium difficile* ATCC9689 (C1610L, Oxoid, Belgium) were purchased as freeze-dried culti-loops and brought into culture according to the manufacturer's
30 instructions. From both cultures a McFarland standard ($A_{625nm} = 0.100$) was prepared in Anaerobic Basal broth (Oxoid, Belgium). 250µl of this standard was added to 10ml of fresh Anaerobic Basal broth, and 225µl of this medium was pipetted into the wells. Yielding a final cell density of 5×10^5 cfu/ml in each well. The microtiter plates were incubated in anaerobic conditions for 18 hours (*C. perfringens*) and 48 hours (*C. difficile*)

using Anaerogen Compact (Oxoid, Belgium) in an airtight plastic bag. Before and after the incubation period, optical density (OD) of each well was measured using the Bioscreen C analyser (Labsystems, Finland). White light (Wide Band) was used to measure the OD. Tests were done in duplo and also control of the medium, medium plus inoculum (negative control) and a positive control; vancomycin (Fluka, Belgium) at three concentrations (0.1, 0.5 and 1.0 µg/ml) was included in the test batch. Minimum inhibition concentration (MIC) was defined as the lowest concentration where no growth occurred or where no increase of OD was detected.

Results and Discussion

Strong increase of OD was noticed in the negative control wells for both species. No growth occurred in the wells containing 1.0 and 0.5 µg/ml vancomycin, 0.1 µg/ml vancomycin did not inhibit the growth of *C. perfringens*. In the wells containing 1 and 2.5 µg/ml PB6-extract, normal growth of *C. perfringens* occurred. From 5 µg/ml on, no growth of *C. perfringens* could be noticed. MIC of the PB6 crude fermentation extract against *C. perfringens* was between 2.5 and 5.0 µg/ml. Also for *C. difficile* a strong increase of OD was noticed in the negative control wells. No growth occurred in the wells containing 1.0 and 0.5 µg/ml vancomycin, 0.1 µg/ml of vancomycin did not inhibit the growth. In the wells containing 1, 2.5 and 5 µg/ml crude PB6-extract, normal growth of *C. difficile* occurred. From 10 µg/ml on, no growth could be detected. MIC of the ether extract of PB6 metabolites against *C. difficile* was between 5.0 and 10 µg/ml. The ether extracts from Bactisubtil® (*B. cereus*), Perenterol® (*S. boulardii*), Bioplus 2B (*Bacillus licheniformis* DSM5749 and *Bacillus subtilis* DSM5750) and Biosporinum (*Bacillus licheniformis* sp. and *Bacillus subtilis* sp.) did not have any significant antibacterial activity for all concentrations tested. MIC was above 50 µg/ml for both clostridia species tested. These results can be found in Table 10.

Table 10: Results of the screening with the crude extracts against *C. perfringens* and *C. difficile*

	MIC (µg/ml)				
	<i>Bacillus</i> PB6	Bactisubtil®	Perenterol®	Bioplus 2B	Biosporinum
<i>C. perfringens</i> ATCC13124	2.5-5	> 50	> 50	> 50	> 50
<i>C. difficile</i> ATCC9689	5-10	> 50	> 50	> 50	> 50

5 Conclusion

We investigated the anticlostridial properties of ether extracts from *B. PB6*, Bactisubtil® (*B. cereus*), Perenterol® (*S. boulardii*), Bioplus 2B (*B. licheniformis* DSM5749 and *B. subtilis* DSM5750) and Biosporinum (*Bacillus licheniformis* sp. and *Bacillus subtilis* sp.). Lab-scale fermentations with these species were set up and the ether
 10 extracts were screened against *C. perfringens* ATCC13124 and *C. difficile* ATCC9689 using broth microdilution techniques. The metabolites of *B. PB6* possess strong anticlostridial properties, MIC are situated between 2.5 and 5.0 µg/ml towards *C. perfringens* and between 5.0 and 10.0 µg/ml towards *C. difficile*. The ether extracts from Bactisubtil®, Perenterol®, Bioplus 2B and Biosporinum fermentations did not have any
 15 significant effect against *C. perfringens* ATCC13124 and *C. difficile* ATCC9689.

EXAMPLE 3 – EFFICACY OF *BACILLUS* PB6 AGAINST IBD

It is known that surfactin inhibits the activity of cytosolic PLA₂, an enzyme centrally involved in many inflammatory processes.⁵⁸ The inhibition of inflammatory
 20 processes makes surfactin producing probiotics very interesting for the treatment of inflammatory diseases, such as Inflammatory Bowel Disease (IBD).

Inflammatory bowel disease refers to two chronic diseases that cause inflammation of the intestines: ulcerative colitis (UC) and Crohn's disease (CD). Although the diseases have some features in common, there are some important differences.

25 CD is a chronic inflammation of the intestinal wall, typically affecting the full thickness of the intestinal wall. Most commonly, it occurs in the lowest portion of the small intestine (ileum) and the large intestine, but it can occur in any part of the digestive tract from the mouth to the anus and the skin around the anus.

In recent decades, CD has become more common both in western and developing countries. It occurs roughly equally in both sexes, and is more common among Jewish people. Most cases begin before the age of 30; the majority starts between the ages of 14 and 24.

5 In each person, the disease affects specific areas of the intestine, sometimes with normal areas (skip areas) sandwiched between the affected zones. In about 35% of CD sufferers, only the ileum is affected. In about 20%, only the large intestine is affected. In about 45% of patients, both the ileum and the large intestine are affected.

The causes of CD are unknown. Research has focused on three main possibilities:
10 a dysfunction of the immune system, infection, and diet.

The most common early symptoms of CD are chronic diarrhea, abdominal pain, fever, loss of appetite, and weight loss. Symptoms differ among CD patients, but there are four common patterns:

- inflammation with pain and tenderness in the right lower part of the abdomen;
- 15 • recurring acute intestinal obstructions that cause severe painful spasms of the intestinal wall, swelling of the abdomen, constipation, and vomiting;
- inflammation and chronic partial intestinal obstruction causing malnutrition and chronic debility;
- abnormal fistulas and abscesses that often cause fever, painful masses in the
20 abdomen, and severe weight loss.

UC is a chronic disease in which the large intestine becomes inflamed and ulcerated, leading to episodes of bloody diarrhea, abdominal cramps, and fever. The disease can start at any age, but usually begins between the ages of 15 and 30.

Unlike CD, UC does not usually affect the full thickness of the intestine, and does not affect the small intestine. The disease usually begins in the rectum or the sigmoid
25 colon, and eventually spreads partially or completely through the large intestine. In some patients, most of the large intestine is affected early on.

About 10% of patients who appear to have UC only suffer a single attack. However, a proportion of such patients may actually be suffering from an undetected
30 infection, rather than true UC. For most patients, UC is a chronic disease that waxes and wanes over time. The causes of UC remain unknown. Heredity and over-active immune responses in the intestine are thought to be contributing factors.

Determination of the *in vivo* efficacy of orally administered *Bacillus* PB6 in the treatment of TNBS-induced colitis in rats (a model for colitis in humans).

In this experiment, the efficacy of PB6 was studied against colitis in rats induced
5 by the rectal administration of 2,4,5-trinitrobenzene sulfonic acid (TNBS).

Study design

Two consecutive trials were done. Male Wistar rats were obtained from the National Centre for Laboratory Animal Sciences (Hyderabad, India) (trial 1) or from the Department of Animal Medicine, TANUVAS (Chennai, India) (trial 2). At the beginning
10 of the treatment period, the animals were 10 to 12 weeks old. Upon their arrival at the test facility, the animals were given a complete clinical examination under the supervision of a veterinarian to ensure that they were in good condition. The animals were acclimatised to the study conditions for a period of at least 7 days. Per trial, animals were randomised and allocated to one of the study groups. The animals were housed per study group in
15 polycarbonate cages (421x290x190 mm, LxWxH). The animal room and test room conditions were set as follows: temperature: $22 \pm 3^{\circ}\text{C}$, relative humidity: $50 \pm 20\%$, light/dark cycle: 12hr/12hr (light 07.00-19.00) and ventilation: approximately 7 cycles/hour of filtered, non-recycled air. All animals had free access (except for the overnight fasting prior to TNBS administration) to rat pellet feed from the National
20 Institute of Nutrition (NIN, Hyderabad, India) and Aquaguard purified water *ad libitum*. In both trials the day of induction of colitis was set as day 1. Colitis was induced, after an overnight fast, using a single intrarectal administration of TNBS at 100 mg/Kg body weight, 8 cm proximal to the anus. The colitis-negative control groups were given saline intrarectally (0.5 ml per animal once) on day 1. Colitis-negative and colitis-positive
25 control groups were given distilled water orally at 10 ml/kg, 3 times daily with 4 h inter dosing, starting on day 1 and up to and including day 7. In the first trial, groups with TNBS-induced colitis were treated with PB6 (1.5×10^8 CFU/Kg or 1.5×10^9 CFU/Kg respectively), 3 times daily with 4 h inter dosing, starting on day 1 and up to and including day 7; with mesalazine (250 mg/Kg/day), starting on day 1 and up to and including day 7;
30 or with infliximab (3 mg/Kg) as a single dose on day 1. The second trial partially repeated the first one concerning the PB6 (1.5×10^8 CFU/Kg), mesalazine and infliximab treatments and included an additional treatment with *S. boulardii* (1.5×10^8 CFU/Kg), 3 times daily with 4 h inter dosing, starting on day 1 and up to and including day 7. The first or only (in case of infliximab) dose of treatment was given within 2 (distilled water, PB6, *S. boulardii*

and mesalazine) or 3 (infliximab) hours after administration of TNBS. Except for infliximab, which was injected intravenously⁵⁹, all treatments were administered by gavage. Twice daily observations were made for clinical signs and mortality. Body weights of animals were recorded on days 1, 4 and 7. On day 8 animals were sacrificed and a 5 cm long segment of the colon (from 10 to 5 cm proximal to the anus) was excised. These segments were opened longitudinally. Contents were removed by washing with saline and gross morphology was scored using the following scale: 0 - no ulcers or inflammation, 1 - no ulcers only local hyperaemia, 2 - ulceration without hyperaemia, 3 - ulceration and inflammation at one site only, 4 - two or more sites of ulceration and inflammation, and 5 - ulceration extending more than 2 cm. The weight of each 5 cm colonic segment was also recorded to assess inflammatory induced edema.

Test preparations

TNBS 5% (w/v) in water (Sigma-aldrich, St Louis, USA) was diluted to a 2.5% solution with ethanol 50%. Dose volume was 4 ml/Kg body weight. PB6 Dry (Kemin Health, Des Moines, USA), a *Bacillus* 'PB6' fermentation broth dried on a malto- and cyclodextrin carrier, was suspended in distilled water to concentrations of 1.5×10^7 and 1.5×10^8 CFU/ml. Dose volume was 10 ml/Kg body weight. *Saccharomyces boulardii* (Enterol[®], Biodiphar, Brussel, Belgium) was suspended in distilled water to a concentration of 1.5×10^7 CFU/ml. Dose volume was 10 ml/Kg body weight. Mesalazine (Mesacol[®], Sun Pharmaceutical Ind. Ltd, Mumbai, India) tablets were powdered using pestle and mortar and a solution in distilled water was prepared containing 25 mg 5-aminosalicylic acid per ml. Dose volume was 10 ml/Kg body weight. Remicade[®] (Infliximab) (Centocor B.V., Leiden, The Netherlands) was first reconstituted with 10 ml water for injection and was further diluted to 2 mg/ml concentration using saline. Dose volume employed was 1.5 ml/Kg body weight. All body weight dependant doses were administered on the basis of the last individual body weight taken. Fresh preparations were made prior to each administration. The preparations were stirred vigorously before each dosing.

Results and discussion

In the first trial one of the rats treated with Infliximab died on day 2. This animal had been given a second injection on day 1 because at the first one drug solution oozed out. Some animals showed mild signs of diarrhea after induction of colitis with TNBS. The number of observations of diarrhea recorded (and the number of animals affected) in the different treatment groups from day 1 up to and including day 7 were in the first trial 0,

22 (2), 4 (1), 0, 0 and 8 (1) for colitis-negative control, colitis-positive control, PB6 $3 \times 1.5 \times 10^8$ CFU/Kg/day, PB6 $3 \times 1.5 \times 10^9$ CFU/Kg/day, mesalazine 250 mg/Kg/day and infliximab 3 mg/Kg single dose respectively; and in the second trial 0, 42 (4), 10 (1), 34 (3), 12 (1) and 24 (2) for colitis-negative control, colitis-positive control, PB6 $3 \times 1.5 \times 10^8$ CFU/Kg/day, *S. boulardii* $3 \times 1.5 \times 10^8$ CFU/Kg/day, mesalazine 250 mg/Kg/day and infliximab 3 mg/Kg single dose respectively. The colitis-positive control groups showed substantial body weight loss accompanying the colitis. In trial 1 this loss was higher than in trial 2, what might be related to a difference in age and growth rate at the moment of induction of colitis. The lower body weight in general at the start and the higher body weight gain in terms of percentage over the trial period of the colitis-negative control group in trial 1 probably indicate that these animals were on average somewhat younger than those in trial 2. While common treatments in trial 1 all significantly suppressed the negative effect of colitis on body weight gain, in trial 2 this was only the case with PB6 treatment (Table 11).

Table 11. Average body weight (g) and average body weight gain (%) with their standard deviation.

	Av. Body weight (g)			Av. Body weight gain (%)
<u>Trial 1</u>	<u>Day 1</u>	<u>Day 4</u>	<u>Day 7</u>	<u>Day 1 to 7</u>
Colitis-negative control	178 ± 17	191 ± 13 *	203 ± 13 *	14.3 ± 4.5 *
Colitis-positive control	182 ± 15	167 ± 16	159 ± 18	-13.0 ± 4.5
PB6 1.5×10^8 CFU/Kg	180 ± 13	186 ± 13	195 ± 10 *	8.8 ± 8.0 *
PB6 1.5×10^9 CFU/Kg	180 ± 16	187 ± 15	199 ± 17 *	10.3 ± 1.1 *
Mesalazine	181 ± 11	187 ± 10	196 ± 10 *	8.8 ± 1.4 *
Infliximab	180 ± 14	183 ± 7	186 ± 6 *	1.3 ± 5.4 *
<u>Trial 2</u>	<u>Day 1</u>	<u>Day 4</u>	<u>Day 7</u>	<u>Day 1 to 7</u>
Colitis-negative control	207 ± 20	210 ± 19	214 ± 17	3.5 ± 1.8 *
Colitis-positive control	208 ± 21	196 ± 21	193 ± 19	-7.5 ± 3.2
PB6 1.5×10^8 CFU/Kg	206 ± 23	206 ± 23	212 ± 26	2.9 ± 3.1 *
<i>S. boulardii</i>	207 ± 17	196 ± 16	195 ± 24	-5.7 ± 5.6
Mesalazine	207 ± 12	199 ± 18	201 ± 20	-3.3 ± 5.5
Infliximab	208 ± 15	197 ± 14	197 ± 13	-5.2 ± 3.4

*Significantly different from the positive control group (Dunnett, $P < 0.05$).

The greater impact of colitis on the animals in trial 1 probably left more room for improvement by any of the treatments. PB6 and mesalazine always resulted in a colon segment weight and a gross morphology score for the colon wall that could be clearly distinguished from those of the colitis-positive control group (Table 12).

5

Table 12. Average gross morphology score for the colon wall and average wet weight of a 5 cm colon segment.

	Gross morphology score	Wet weight (g)
<u>Trial 1</u>		
IBD-negative control	0.2 ± 0.4 *	0.378 ± 0.047 *
IBD-positive control	3.8 ± 0.8	1.406 ± 0.209
PB6 1.5 × 10 ⁸ CFU/Kg (post-induction)	1.0 ± 0.0 *	0.443 ± 0.063 *
PB6 1.5 × 10 ⁹ CFU/Kg (post-induction)	0.6 ± 0.5 *	0.513 ± 0.317 *
Mesalazine	0.8 ± 0.4 *	0.435 ± 0.052 *
Infliximab	3.0 ± 1.4	1.055 ± 0.490
<u>Trial 2</u>		
IBD-negative control	0.0 ± 0.0 *	0.274 ± 0.049 *
IBD-positive control	3.4 ± 1.1	0.832 ± 0.216
PB6 1.5 × 10 ⁸ CFU/Kg	0.6 ± 0.5 *	0.280 ± 0.039 *
<i>S. boulardii</i>	2.8 ± 1.3	0.735 ± 0.221
Mesalazine	1.2 ± 0.8 *	0.445 ± 0.145 *
Infliximab	2.0 ± 0.7	0.657 ± 0.076

*Significantly different from the positive control group (Dunnett, P<0.05).

10 The health status of the colon wall in rats with TNBS induced colitis treated with PB6 and Mesacol was macroscopically the same as that in colitis-free rats. For an unknown reason mesalazine treatment was somewhat less effective in trial 2, resulting in some ulcerations. Visual examinations of the longitudinally opened colon segments clearly show the ulcerations and areas of necrotic tissue present in the positive control, the *S.*

15 *boulardii*, and the infliximab treatment groups and the absence thereof in the PB6 groups and the trial 1 mesalazine group. In trial 1 one of the rats treated with infliximab had a gross morphology score of 1 and a colon segment weight of 0.402 g. These data suggest that in this particular animal the infliximab treatment was successful or that the induction of colitis was not. At the end of the treatment period the average body weight gain and

20 colon segment weight data of rats treated with infliximab were intermediary to those of the colitis-negative and the colitis-positive control groups in both trials. This probably indicates that there was some effect of infliximab, although not statistically significant in

these trials. In a similar trial in rats where infliximab was administered at the same dose of 3 mg/Kg i.v. 2 days before induction of colitis with TNBS, it was shown to have some effect on colon gross morphology but not to reduce the level of edema significantly below that of the colitis-positive control group.¹⁹ In the colon segments of rats treated with PB6 there were no ulcerations. Only hyperaemia was observed in the majority of these segments. PB6 clearly attenuates inflammation in the rat colon wall as induced by intrarectal administration of TNBS. The efficacy of a 7 days post-induction treatment with $3 \times 1.5 \times 10^9$ CFU/Kg/day observed in a previous trial was confirmed in trial 1.²⁰ While a 7 days post-induction treatment with PB6 $3 \times 8 \times 10^7$ CFU/Kg/day from the same previous trial was concluded to be ineffective, an increase of the CFU/Kg/day to $3 \times 1.5 \times 10^8$ proved in both current trials to be sufficient to reduce inflammation to an extent that the data of this treatment could no longer be statistically discerned from those of the colitis-negative control group.

Conclusion

This study was designed and conducted to confirm and document the efficacy of *Bacillus* 'PB6' against 2,4,6 - trinitrobenzenesulfonic acid (TNBS)-induced colitis in rats observed in a previous trial as well as to compare its efficacy with that of *S. boulardii* (probiotic), mesalazine and infliximab (standard drugs). This rat model is well established, reliable and widely used to examine the efficiency of drugs aimed at treating IBD. Without any treatment after induction of colitis several rats showed mild signs of diarrhea and on average kept losing weight throughout the remaining trial period. The gross morphology of the intestinal wall of their colon was characterized by inflammation, ulceration and even necrosis. Treatment with PB6 3 times 1.5×10^8 CFU/Kg/day or 3 times 1.5×10^9 CFU/Kg/day for 7 days resulted in a colon wall health status that was statistically identical to that of the negative control group and that of the group treated with the standard drug mesalazine, an anti-inflammatory.

EXAMPLE 4 - ANTIMICROBIAL PROPERTIES OF THE METABOLITES OF PB6

Methods and Materials

Antagonistic properties of *Bacillus* PB6 and *Bacillus cereus* isolated from Bactisubtil (Sanofi -- synthelabo) were tested against different indicator strains e.g. *C. perfringens* ATCC13124, *C. difficile* ATCC9689 and *Campylobacter jejuni* ATCC 33291.

Bacillus cereus and *Bacillus* PB6 were each suspended in 5ml of sterile saline. Using a swab a single streak of the probiotic suspensions on Tryptone Soy agar plates

(Oxoid, Belgium) was made and the plates were incubated at 37°C for 24 hours in aerobic conditions.

Afterwards a suspension of the different indicator strains were inoculated perpendicularly to both *Bacillus* cultures with a swab and incubated for 24 hours in aerobic conditions. Plates inoculated with *Clostridia perfringens* were incubated for 24 hours, plates inoculated with *C. difficile*, were incubated for 48 hours in anaerobic conditions using Anaerogen Pak (Oxoid, Belgium). A suspension of a 48 hours *C. jejuni* ATCC33291 culture was used to make 4 streaks perpendicularly to the probiotic cultures. Plates inoculated with *Campylobacter species* were incubated in micro-aerobic conditions (CampyGen, Oxoid) at 37°C for 48 hours. The streak lines must not touch one another. Following incubation at 37°C, antagonistic effects were evaluated by the appearance of clear zones surrounding the junctions of the streak-lines indicating the inhibitory effect of one organism against the other.

Results and Discussion

PB6 had an antagonistic effect against *C. perfringens* ATCC13124 and *C. difficile* ATCC9689. A clear zone could be observed at the intersections of the streak-lines on the plate for both species. The antagonistic effect against *C. perfringens* is clearly visible due to the haemolytic characteristics of this species. An example is depicted in Fig. 1. Although not as clear on this picture, a significant clear zone at the intersection of PB6 and *C. difficile* cultures was also noticed.

Bacillus cereus (Bactisubtil) had no antagonistic effect against *C. perfringens* ATCC13124 and *C. difficile* ATCC9689. No clear zone could be observed at the intersections of the streak-lines on the plate. An example of the test plate is depicted in Fig. 12.

Bacillus PB6 had an antagonistic effect against *C. jejuni* ATCC 33291. Clear zones can be observed at the intersections of the streak-lines on the plate. An example of the test plate is depicted in Fig. 13.

Bacillus cereus (Bactisubtil) had no antagonistic effect against *C. jejuni* as can be observed in Fig. 14.

Conclusion

Bacillus PB6 isolated from nature, clearly has strong antagonistic properties towards *C. perfringens* ATCC13124, *C. difficile* ATCC9689 and *C. jejuni* ATCC 33291. On the contrary, no effect could be observed for other human pathogenic bacteria tested.

Bacillus cereus (Bactisubtil) did not show any antagonistic effect towards *C. perfringens* ATCC13124, *C. difficile* ATCC9689 and other tested microorganisms.

Viable cells of the PB6 strain have been deposited with the American Type Culture Collection ("ATCC"), 10801 University Blvd., Manassas, Va., 20110-2209, U.S.A., on
5 May 27, 2005, and assigned accession number PTA-6737. The deposit was made in accordance with 37 C.F.R. 1.801-1.809

The foregoing description and drawings comprise illustrative embodiments of the present inventions. The foregoing embodiments and the methods described herein may
10 vary based on the ability, experience, and preference of those skilled in the art. Merely listing the steps of the method in a certain order does not constitute any limitation on the order of the steps of the method. The foregoing description and drawings merely explain and illustrate the invention, and the invention is not limited thereto, except insofar as the claims are so limited. Those skilled in the art that have the disclosure before them will be
15 able to make modifications and variations therein without departing from the scope of the invention.

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- ⁵⁶ Dzink J., Bartlett J.G. 1980. *In vitro* susceptibility of *C. difficile* isolates from patients with antibiotic-associated diarrhea or colitis. *Antimicrob. Agents Chemother.* 17:695-698.
- ⁵⁷ Tannock G.W. 1999. Probiotics: A critical review. Horizon Scientific Press.
- ⁵⁸ Kim, K. *et al.* Suppression of inflammatory responses by surfactin, a selective inhibitor of platelet cytosolic phospholipase A₂. *Biochem. Pharmacol.* 1998; 55: 975 – 985.

Claims

1. Use of a *Bacillus subtilis* bacteria of the strain identified as ATCC strain PTA-6737 in the manufacture of a medicament for the prophylaxis or treatment of inflammatory bowel disease.
2. Use according to claim 1, wherein the *Bacillus* bacteria is combined with inulin or a probiotic.
3. A *Bacillus subtilis* bacteria of the strain identified as ATCC strain PTA-6737 for use in the treatment or prophylaxis of inflammatory bowel disease.
4. A *Bacillus subtilis* bacteria according to claim 3 in combination with inulin or a probiotic for use according to claim 3.
5. Use of an isolated bacterial strain having the 16S rRNA sequence of SEQ ID NO. 1 in the manufacture of a medicament for the prophylaxis or treatment of inflammatory bowel disease.
6. An isolated bacterial strain having the 16S rRNA sequence of SEQ ID NO. 1 for use in the treatment or prophylaxis of inflammatory bowel disease.

KEM 121 Sequence Listing.ST25.txt
SEQUENCE LISTING

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