

Method for Improving the Dissolution Profile of a Biologically Active Material**Field of the Invention**

The present invention relates to methods for improving the dissolution profile of a biologically active material. The invention also relates to biologically active materials in particulate form produced by said methods, compositions comprising such materials, medicaments produced using said biologically active materials in particulate form and/or compositions, and to methods of treatment of an animal, including man, using a therapeutically effective amount of said biologically active materials administered by way of said medicaments.

Background

Poor bioavailability is a significant problem encountered in the development of therapeutic compositions, particularly those materials containing a biologically active material that is poorly soluble in water at physiological pH. An active material's bioavailability is the degree to which the active material becomes available to the target tissue in the body after systemic administration through, for example, oral or intravenous means. Many factors affect bioavailability, including the form of dosage and the solubility and dissolution rate of the active material.

Poorly and slowly water-soluble materials tend to be eliminated from the gastrointestinal tract before being absorbed into the circulation. In addition, poorly soluble active agents tend to be disfavored or even unsafe for intravenous administration due to the risk of particles of agent blocking blood flow through capillaries.

It is known that the rate of dissolution of a particulate drug will increase with increasing surface area. One way of increasing surface area is decreasing particle size. Consequently, methods of making finely divided or sized drugs have been studied with a view to controlling the size and size range of drug particles for pharmaceutical compositions.

For example, dry milling techniques have been used to reduce particle size and hence influence drug absorption. However, in conventional dry milling the limit of fineness is reached generally in the region of about 100 microns (100,000 nm), at which point material cakes on the milling chamber and prevents any further diminution of particle size. Alternatively, wet grinding may be employed to reduce particle size, but flocculation restricts the lower particle size limit to approximately 10 microns (10,000 nm). The wet milling process, however, is prone to contamination, thereby leading to a bias in the pharmaceutical art against wet milling. Another alternative milling technique, commercial airjet milling, has provided particles ranging in average size from as low as about 1 to about 50 microns (1,000-50,000 nm).

There are several approaches currently used to formulate poorly soluble active agents. One approach is to prepare the active agent as a soluble salt. Where this approach cannot be employed, alternate (usually physical) approaches are employed to improve the solubility of the active agent. Alternate approaches generally subject the active agent to physical conditions that change the agent's physical and or chemical properties to improve its solubility. These include process technologies such as micronization, modification of crystal or polymorphic structure, development of oil based solutions, use of co-solvents, surface stabilizers or complexing agents, micro-emulsions, super critical fluid and production of solid dispersions or solutions. More than one of these processes may be used in combination to improve formulation of a particular therapeutic material. Many of these approaches commonly convert a drug into an amorphous state, which generally leads to a higher dissolution rate. However, formulation approaches that result in the production of amorphous material are not common in commercial formulations due to concerns relating to stability and the potential for material to re-crystallize.

These techniques for preparing such pharmaceutical compositions tend to be complex. By way of example, a principal technical difficulty encountered with emulsion polymerization is the removal of contaminants, such as unreacted monomers or initiators (which may have undesirable levels of toxicity), at the end of the manufacturing process.

Another method of providing reduced particle size is the formation of pharmaceutical drug microcapsules, which techniques include micronizing, polymerisation and co-dispersion. However, these techniques suffer from a number of disadvantages including at least the inability to produce sufficiently small particles such as those obtained by milling, and the presence of co-solvents and/or contaminants such as toxic monomers which are difficult to remove, leading to expensive manufacturing processes.

Over the last decade, intense scientific investigation has been carried out to improve the solubility of active agents by converting the agents to ultra fine powders by methods such as milling and grinding. These techniques may be used to increase the dissolution rate of a particulate solid by increasing the overall surface area and decreasing the mean particle size.

US Patent 6,634,576 discloses examples of wet-milling a solid substrate, such as a pharmaceutically active compound, to produce a "synergetic co-mixture".

International Patent Application PCT/AU2005/001977 (Nanoparticle Composition(s) and Method for Synthesis Thereof) describes, *inter alia*, a method comprising the step of contacting a precursor compound with a co-reactant under mechanochemical synthesis conditions wherein a solid-state chemical reaction between the precursor compound and the co-reactant produces therapeutically active nanoparticles dispersed in a carrier matrix. Mechanochemical synthesis, as discussed in International Patent Application PCT/AU2005/001977, refers to the use of mechanical energy to activate, initiate or promote a chemical reaction, a crystal structure transformation or a phase change in a material or a mixture of materials, for example by

agitating a reaction mixture in the presence of a milling media to transfer mechanical energy to the reaction mixture, and includes without limitation "mechanochemical activation", "mechanochemical processing", "reactive milling", and related processes.

International Patent Application PCT/AU2007/000910 (Methods for the preparation of biologically active compounds in nanoparticulate form) describes, *inter alia*, a method for dry milling raloxifene with lactose and NaCl which produced nanoparticulate raloxifene without significant aggregation problems. One limitation of this method is an upper limit to the drug content that can be successfully milled to produce nanoparticles. For some drugs that require a high dose this limitation may restrict the options available for the production of a commercially viable dosage form.

The present invention provides methods for improving the dissolution profile of a biologically active material which ameliorate some of the problems attendant with prior technologies, or provides an alternative thereto.

One example of a therapeutic area where this technology could be applied in is the area of acute pain management. Many pain medications such as meloxicam (marketed as Mobic® by pharmaceutical company Boehringer Ingelheim) provides pain relief for chronic pain, but must be taken on a daily basis to maintain an effective therapeutic level.

Meloxicam is a poorly water soluble drug which is only slowly absorbed by the body (Tmax is 4-5 hours), so a method such as the present invention which provides for improved dissolution, will likely provide much faster absorption resulting in a more rapid onset of the therapeutic effect. Meloxicam also has a long half life (15-20 hours) that means it only need be taken once a day. By using a method such as the present invention, which provides faster absorption, a drug such as meloxicam, could be transformed from a chronic pain drug to an acute pain drug. For meloxicam this would provide a medication that could provide therapeutic relief for acute pain, with the advantage of sustained pain relief over 24 hours.

Meloxicam also has sub-optimal bioavailability at 89% for an oral capsule, compared with an IV dosage form. A component of this sub optimal bioavailability is also likely due to the poor water solubility of this drug. If the low solubility does contribute to this sub optimal bioavailability, the improvement of the dissolution of this drug with a method such as the present invention could provide scope to produce a dosage form with a lower active dose whilst still providing the effective therapeutic dose.

Although the background to the present invention is discussed in the context of improving the bioavailability of materials that are poorly or slowly water soluble, the applications of the methods of the present invention are not limited to such, as is evident from the following description of the invention.

Further, although the background to the present invention is largely discussed in the context of improving the bioavailability of therapeutic or pharmaceutical compounds, the applications of the methods of the present invention are clearly not limited to such. For example, as is evident

from the following description, applications of the methods of the present invention include but are not limited to: nutraceutical and nutritional compounds, complementary medicinal compounds, veterinary therapeutic applications and agricultural chemical applications, such as pesticide, fungicide or herbicide.

Furthermore, an application of the current invention would be to materials which contain a biologically active compound such as, but not limited to a therapeutic or pharmaceutical compound, a nutraceutical or nutrient, a complementary medicinal product such as active components in plant or other naturally occurring material, a veterinary therapeutic compound or an agricultural compound such as a pesticide, fungicide or herbicide. Specific examples would be the spice turmeric that contains the active compound curcumin, or flax seed that contains the nutrient ALA an omega-3 fatty acid. As these specific examples indicate this invention could be applied to, but not limited to, a range of natural products such as seeds, cocoa and cocoa solids, coffee, herbs, spices, other plant materials or food materials that contain a biologically active compound. The application of this invention to these types of materials would enable greater availability of the active compound in the materials when used in the relevant application. For example where material subject to this invention is orally ingested the active would be more bioavailable.

Summary of the Invention

In one aspect the present invention is directed to the unexpected finding that the dissolution profile of biologically active materials can be improved by dry milling solid biologically active material to a particle size of greater than 1 μ m. In one surprising aspect of the invention, the dissolution profile of a biologically active material can be improved without substantially reducing the particle size of the material or reducing the material to nanoparticulate form. In another surprising aspect of the invention, the material retains its crystalline structure and is not amorphous, yet the dissolution profile of the biologically active material is improved. In another surprising aspect of the invention, the dissolution profile of a biologically active material is improved without the need for a surfactant or stabiliser. In another surprising aspect of the invention, the dissolution profile of a biologically active material is improved without the need for a disintegrant to be present during the milling process.

Thus, in a first aspect the invention comprises a method for improving the dissolution profile of a biologically active material, comprising the steps of: dry milling a solid biologically active material and a millable grinding matrix in a mill comprising a plurality of milling bodies, for a time period sufficient to produce particles of the biologically active material dispersed in an at least partially milled grinding material.

In one preferred embodiment, the particles have an average particle size equal or greater than 1 μ m determined on a particle number basis. More preferably, the average particle size of the biologically active material may be reduced by a factor selected from the group consisting of:

less than 5%, less than 10%, less than 20%, less than 30%, less than 40%, less than 50%, less than 60%, less than 70%, less than 80%, less than 90 %, less than 95% and less than 99%. Even more preferably, the average particle size falls within the range selected from the group consisting of: 1-1000 μ m, 1-500 μ m, 1-300 μ m, 1-200 μ m, 1-150 μ m, 1-100 μ m, 1-50 μ m, 1-20 μ m, 1-10 μ m, 1-7.5 μ m, 1-5 μ m and 1-2 μ m.

In another preferred embodiment, the particles have a median particle size selected from the group consisting of: equal or greater than 1 μ m; and equal or greater than 2 μ m, wherein the median particle size is determined on a particle volume basis. More preferably, the percentage of particles with an average particle size greater than 1 μ m on a particle volume basis is a percentage selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 100%. Alternatively, the percentage of particles with an average particle size greater than 2 μ m on a particle volume basis is a percentage selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 100%.

In another preferred embodiment, the median particle size may be reduced by a factor selected from the group consisting of: less than 5%, less than 10%, less than 20%, less than 30%, less than 40%, less than 50%, less than 60%, less than 70%, less than 80%, less than 90 %, less than 95% and less than 99%.

In another preferred embodiment, the median particle size falls within the range selected from the group consisting of: 1-1000 μ m, 1-500 μ m, 1-300 μ m, 1-200 μ m, 1-150 μ m, 1-100 μ m, 1-50 μ m, 1-20 μ m, 1-10 μ m, 1-7.5 μ m, 1-5 μ m 1-2 μ m, 2-1000 μ m, 2-500 μ m, 2-300 μ m, 2-200 μ m, 2-150 μ m, 2-100 μ m, 2-50 μ m, 2-20 μ m, 2-10 μ m, 2-7.5 μ m and 2-5 μ m.

In another preferred embodiment, the crystallinity profile of the biologically active material is selected from the group consisting of: at least 50% of the biologically active material is crystalline, at least 60% of the biologically active material is crystalline, at least 70% of the biologically active material is crystalline, at least 75% of the biologically active material is crystalline, at least 85% of the biologically active material is crystalline, at least 90% of the biologically active material is crystalline, at least 95% of the biologically active material is crystalline and at least 98% of the biologically active material is crystalline. More preferably, the crystallinity profile of the biologically active material is substantially equal to the crystallinity profile of the biologically active material before the material was subjected to the method as described herein.

In another preferred embodiment, the amorphous content of the biologically active material is selected from the group consisting of: less than 50% of the biologically active material is amorphous, less than 40% of the biologically active material is amorphous, less than 30% of the biologically active material is amorphous, less than 25% of the biologically active material is amorphous, less than 15% of the biologically active material is amorphous, less than 10% of the biologically active material is amorphous, less than 5% of the biologically active material is amorphous and less than 2% of the biologically active material is amorphous. Preferably, the

biologically active material has no significant increase in amorphous content after subjecting the material to the method as described herein.

In another preferred embodiment, the milling time period is a range selected from the group consisting of: between 10 minutes and 2 hours, between 10 minutes and 1 hour, between 10 minutes and 45 minutes, between 10 minutes and 30 minutes, between 5 minutes and 30 minutes, between 5 minutes and 20 minutes, between 2 minutes and 10 minutes, between 2 minutes and 5 minutes, between 1 minutes and 20 minutes, between 1 minute and 10 minutes, and between 1 minute and 5 minutes.

In another preferred embodiment, the milling medium is selected from the group consisting of: ceramics, glasses, polymers, ferromagnetics and metals. Preferably, the milling medium is steel balls having a diameter selected from the group consisting of: between 1 and 20 mm, between 2 and 15 mm and between 3 and 10 mm. In another preferred embodiment, the milling medium is zirconium oxide balls having a diameter selected from the group consisting of: between 1 and 20 mm, between 2 and 15 mm and between 3 and 10 mm. Preferably, the dry milling apparatus is a mill selected from the group consisting of: attritor mills (horizontal or vertical), nutating mills, tower mills, pearl mills, planetary mills, vibratory mills, eccentric vibratory mills, gravity-dependent-type ball mills, rod mills, roller mills and crusher mills. Preferably the milling medium within the milling apparatus is mechanically agitated by 1, 2 or 3 rotating shafts. Preferably, the method is configured to produce the biologically active material in a continuous fashion. Preferably, the total combined amount of biologically active material and grinding matrix in the mill at any given time is equal to or greater than a mass selected from the group consisting of: 200grams, 500grams, 1kg, 2kg, 5kg, 10kg, 20kg, 30kg, 50kg, 75kg, 100kg, 150kg, 200kg. Preferably, the total combined amount of biologically active material and grinding matrix is less than 2000kg.

In another preferred embodiment, the biologically active material is selected from the group consisting of: fungicides, pesticides, herbicides, seed treatments, cosmeceuticals, cosmetics, complementary medicines, natural products, vitamins, nutrients, nutraceuticals, pharmaceutical actives, biologics, amino acids, proteins, peptides, nucleotides, nucleic acids additives, foods and food ingredients and analogs, homologs and first order derivatives thereof. Preferably, the biologically active material is selected from the group consisting of: anti-obesity drugs, central nervous system stimulants, carotenoids, corticosteroids, elastase inhibitors, anti-fungals, oncology therapies, anti-emetics, analgesics, cardiovascular agents, anti-inflammatory agents, such as NSAIDs and COX-2 inhibitors, anthelmintics, anti-arrhythmic agents, antibiotics (including penicillins), anticoagulants, antidepressants, antidiabetic agents, antiepileptics, antihistamines, antihypertensive agents, antimuscarinic agents, antimycobacterial agents, antineoplastic agents, immunosuppressants, antithyroid agents, antiviral agents, anxiolytics, sedatives (hypnotics and neuroleptics), astringents, alpha-adrenergic receptor blocking agents, beta-adrenoceptor blocking agents, blood products and substitutes, cardiac inotropic agents,

contrast media, cough suppressants (expectorants and mucolytics), diagnostic agents, diagnostic imaging agents, diuretics, dopaminergics (anti-parkinsonian agents), haemostatics, immunological agents, lipid regulating agents, muscle relaxants, parasympathomimetics, parathyroid calcitonin and biphosphonates, prostaglandins, radio-pharmaceuticals, sex hormones (including steroids), anti-allergic agents, stimulants and anoretics, sympathomimetics, thyroid agents, vasodilators, and xanthines.

Preferably, the biologically active material is selected from the group consisting of: indomethacin, diclofenac, naproxen, meloxicam, metaxalone, cyclosporin A, progesterone, celecoxib, cilostazol, ciprofloxacin, 2,4-dichlorophenoxyacetic acid, anthraquinone, creatine monohydrate, glyphosate, halosulfuron, mancozeb, metsulfuron, salbutamol, sulphur, tribenuran and estradiol or any salt or derivative thereof.

In another preferred embodiment, the grinding matrix is a single matrix or is a mixture of two or more matrices in any proportion. Preferably, the major components of the grinding matrix are selected from the group consisting of: mannitol, sorbitol, Isomalt, xylitol, maltitol, lactitol, erythritol, arabitol, ribitol, glucose, fructose, mannose, galactose, anhydrous lactose, lactose monohydrate, sucrose, maltose, trehalose, maltodextrins, dextrin, Inulin, dextrans, polydextrose, starch, wheat flour, corn flour, rice flour, rice starch, tapioca flour, tapioca starch, potato flour, potato starch, other flours and starches, milk powder, skim milk powders, other milk solids and derivatives, soy flour, soy meal or other soy products, cellulose, microcrystalline cellulose, microcrystalline cellulose based co blended materials, pregelatinized (or partially) starch, HPMC, CMC, HPC, citric acid, tartaric acid, malic acid, maleic acid, fumaric acid, ascorbic acid, succinic acid, sodium citrate, sodium tartrate, sodium malate, sodium ascorbate, potassium citrate, potassium tartrate, potassium malate, potassium ascorbate, sodium carbonate, potassium carbonate, magnesium carbonate, sodium bicarbonate, potassium bicarbonate and calcium carbonate, dibasic calcium phosphate, tribasic calcium phosphate, sodium sulfate, sodium chloride, sodium metabisulfite, sodium thiosulfate, ammonium chloride, Glauber's salt, ammonium carbonate, sodium bisulfate, magnesium sulfate, potash alum, potassium chloride, sodium hydrogen sulfate, sodium hydroxide, crystalline hydroxides, hydrogen carbonates, ammonium chloride, methylamine hydrochloride, ammonium bromide, silica, thermal silica, alumina, titanium dioxide, talc, chalk, mica, kaolin, bentonite, hectorite, magnesium trisilicate, clay based materials or aluminium silicates, sodium lauryl sulfate, sodium stearyl sulfate, sodium cetyl sulfate, sodium cetostearyl sulfate, sodium docusate, sodium deoxycholate, N-lauroylsarcosine sodium salt, glyceryl monostearate, glycerol distearate, glyceryl palmitostearate, glyceryl behenate, glyceryl caprylate, glyceryl oleate, benzalkonium chloride, CTAB, CTAC, Cetrimide, cetylpyridinium chloride, cetylpyridinium bromide, benzethonium chloride, PEG 40 stearate, PEG 100 stearate, poloxamer 188, poloxamer 407, poloxamer 338, polyoxyl 2 stearyl ether, polyoxyl 100 stearyl ether, polyoxyl 20 stearyl ether, polyoxyl 10 stearyl ether, polyoxyl 20 cetyl ether, polysorbate 20, polysorbate

40, polysorbate 60, polysorbate 61, polysorbate 65, polysorbate 80, polyoxyl 35 castor oil, polyoxyl 40 castor oil, polyoxyl 60 castor oil, polyoxyl 100 castor oil, polyoxyl 200 castor oil, polyoxyl 40 hydrogenated castor oil, polyoxyl 60 hydrogenated castor oil, polyoxyl 100 hydrogenated castor oil, polyoxyl 200 hydrogenated castor oil, cetostearyl alcohol, macrogel 15 hydroxystearate, sorbitan monopalmitate, sorbitan monostearate, sorbitan trioleate, Sucrose Palmitate, Sucrose Stearate, Sucrose Distearate, Sucrose laurate, Glycocholic acid, sodium Glycholate, Cholic Acid, Sodium Cholate, Sodium Deoxycholate, Deoxycholic acid, Sodium taurocholate, taurocholic acid, Sodium taurodeoxycholate, taurodeoxycholic acid, soy lecithin, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, phosphatidylinositol, PEG4000, PEG6000, PEG8000, PEG10000, PEG20000, alkyl naphthalene sulfonate condensate/Lignosulfonate blend, Calcium Dodecylbenzene Sulfonate, Sodium Dodecylbenzene Sulfonate, Diisopropyl naphthalenesulphonate, erythritol distearate, Naphthalene Sulfonate Formaldehyde Condensate, nonylphenol ethoxylate (poe-30), Tristyrylphenol Ethoxylate, Polyoxyethylene (15) tallowalkylamines, sodium alkyl naphthalene sulfonate, sodium alkyl naphthalene sulfonate condensate, sodium alkylbenzene sulfonate, sodium isopropyl naphthalene sulfonate, Sodium Methyl Naphthalene Formaldehyde Sulfonate, sodium n-butyl naphthalene sulfonate, tridecyl alcohol ethoxylate (poe-18), Triethanolamine isodecanol phosphate ester, Triethanolamine tristyrylphosphate ester, Tristyrylphenol Ethoxylate Sulfate, Bis(2-hydroxyethyl)tallowalkylamines.

Preferably, the concentration of the single (or first) material is selected from the group consisting of: 5-99% w/w, 10-95% w/w, 15-85% w/w, of 20-80% w/w, 25-75% w/w, 30-60% w/w, 40-50% w/w.

Preferably, the concentration of the second or subsequent material is selected from the group consisting of: 5-50% w/w, 5-40% w/w, 5-30% w/w, of 5-20% w/w, 10-40% w/w, 10-30% w/w, 10-20% w/w, 20-40% w/w, or 20-30% w/w or if the second or subsequent material is a surfactant or water soluble polymer the concentration is selected from 0.1-10% w/w, 0.1-5% w/w, 0.1-2.5% w/w, of 0.1-2% w/w, 0.1-1%, 0.5-5% w/w, 0.5-3% w/w, 0.5-2% w/w, 0.5-1.5%, 0.5-1% w/w, of 0.75-1.25% w/w, 0.75-1% and 1% w/w.

Preferably, the grinding matrix is selected from the group consisting of:

- (a) lactose monohydrate or lactose monohydrate combined with at least one material selected from the group consisting of: xylitol; lactose anhydrous; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000,

sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

- (b) lactose anhydrous or lactose anhydrous combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18;

Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrilphosphate ester; Tristyrilphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

- (c) mannitol or mannitol combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrilphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrilphosphate ester; Tristyrilphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.
- (d) Sucrose or sucrose combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338,

sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

- (e) Glucose or glucose combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

- (f) Sodium chloride or sodium chloride combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.
- (g) xylitol or xylitol combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407,

Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

- (h) Tartaric acid or tartaric acid combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

- (i) microcrystalline cellulose or microcrystalline cellulose combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; mannitol; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.
- (j) Kaolin combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338,

Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

- (k) Talc combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

Preferably, the grinding matrix is selected from the group consisting of: a material considered to be Generally Regarded as Safe (GRAS) for pharmaceutical products; a material considered acceptable for use in an agricultural formulation; and a material considered acceptable for use in a veterinary formulation.

In another preferred embodiment, a milling aid is used or a combination of milling aids. Preferably, the milling aid is selected from the group consisting of: colloidal silica, a surfactant, a polymer, a stearic acid and derivatives thereof. Preferably, the surfactant is selected from the group consisting of: polyoxyethylene alkyl ethers, polyoxyethylene stearates, polyethylene glycols (PEG), poloxamers, poloxamines, sarcosine based surfactants, polysorbates, aliphatic alcohols, alkyl and aryl sulfates, alkyl and aryl polyether sulfonates and other sulfate surfactants, trimethyl ammonium based surfactants, lecithin and other phospholipids, bile salts, polyoxyethylene castor oil derivatives, polyoxyethylene sorbitan fatty acid esters, Sorbitan fatty acid esters, Sucrose fatty acid esters, alkyl glucopyranosides, alkyl maltopyranosides, glycerol fatty acid esters, Alkyl Benzene Sulphonic Acids, Alkyl Ether Carboxylic Acids, Alkyl and aryl Phosphate esters, Alkyl and aryl Sulphate esters, Alkyl and aryl Sulphonic acids, Alkyl Phenol Phosphates esters, Alkyl Phenol Sulphates esters, Alkyl and Aryl Phosphates, Alkyl Polysaccharides, Alkylamine Ethoxylates, Alkyl-Naphthalene Sulphonates formaldehyde condensates, Sulfosuccinates, lignosulfonates, Ceto-Oleyl Alcohol Ethoxylates, Condensed Naphthalene Sulphonates, Dialkyl and Alkyl Naphthalene Sulphonates, Di-alkyl Sulphosuccinates, Ethoxylated nonylphenols, Ethylene Glycol Esters, Fatty Alcohol Alkoxylates, Hydrogenated tallowalkylamines, Mono-alkyl Sulphosuccinamates, Nonyl Phenol Ethoxylates, Sodium Oleyl N-methyl Taurate, Tallowalkylamines, linear and branched dodecylbenzene sulfonic acids

Preferably, the surfactant is selected from the group consisting of: sodium lauryl sulfate, sodium stearyl sulfate, sodium cetyl sulfate, sodium cetostearyl sulfate, sodium docusate, sodium deoxycholate, N-lauroylsarcosine sodium salt, glyceryl monostearate, glycerol distearate, glyceryl palmitostearate, glyceryl behenate, glyceryl caprylate, glyceryl oleate, benzalkonium chloride, CTAB, CTAC, Cetrimide, cetylpyridinium chloride, cetylpyridinium bromide, benzethonium chloride, PEG 40 stearate, PEG 100 stearate, poloxamer 188, poloxamer 407, poloxamer 338, polyoxyl 2 stearyl ether, polyoxyl 100 stearyl ether, polyoxyl 20 stearyl ether, polyoxyl 10 stearyl ether, polyoxyl 20 cetyl ether, polysorbate 20, polysorbate 40, polysorbate 60, polysorbate 61, polysorbate 65, polysorbate 80, polyoxyl 35 castor oil, polyoxyl 40 castor oil, polyoxyl 60 castor oil, polyoxyl 100 castor oil, polyoxyl 200 castor oil, polyoxyl 40 hydrogenated castor oil, polyoxyl 60 hydrogenated castor oil, polyoxyl 100 hydrogenated castor oil, polyoxyl 200 hydrogenated castor oil, cetostearyl alcohol, macrogel 15 hydroxystearate, sorbitan monopalmitate, sorbitan monostearate, sorbitan trioleate, Sucrose Palmitate, Sucrose Stearate, Sucrose Distearate, Sucrose laurate, Glycocholic acid, sodium Glycholate, Cholic

Acid, Sodium Cholate, Sodium Deoxycholate, Deoxycholic acid, Sodium taurocholate, taurocholic acid, Sodium taurodeoxycholate, taurodeoxycholic acid, soy lecithin, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, phosphatidylinositol, PEG4000, PEG6000, PEG8000, PEG10000, PEG20000, alkyl naphthalene sulfonate condensate/Lignosulfonate blend, Calcium Dodecylbenzene Sulfonate, Sodium Dodecylbenzene Sulfonate, Diisopropyl naphthaenesulphonate, erythritol distearate, Naphthalene Sulfonate Formaldehyde Condensate, nonylphenol ethoxylate (poe-30), Tristyrylphenol Ethoxylate, Polyoxyethylene (15) tallowalkylamines, sodium alkyl naphthalene sulfonate, sodium alkyl naphthalene sulfonate condensate, sodium alkylbenzene sulfonate, sodium isopropyl naphthalene sulfonate, Sodium Methyl Naphthalene Formaldehyde Sulfonate, sodium n-butyl naphthalene sulfonate, tridecyl alcohol ethoxylate (poe-18), Triethanolamine isodecanol phosphate ester, Triethanolamine tristyrylphosphate ester, Tristyrylphenol Ethoxylate Sulfate, Bis(2-hydroxyethyl)tallowalkylamines. Preferably the polymer is selected from the list of: polyvinylpyrrolidones (PVP), polyvinylalcohol, Acrylic acid based polymers and copolymers of acrylic acid.

Preferably, the milling aid has a concentration selected from the group consisting of: 0.1-10 % w/w, 0.1-5 % w/w, 0.1-2.5 % w/w, of 0.1-2% w/w, 0.1-1 %, 0.5-5% w/w, 0.5-3% w/w, 0.5-2% w/w, 0.5-1.5%, 0.5-1 % w/w, of 0.75-1.25 % w/w, 0.75-1% and 1% w/w.

Preferably the biologically active ingredient is milled with lactose monohydrate; mannitol; glucose; microcrystalline cellulose; tartaric acid; or lactose monohydrate and sodium dodecyl sulfate.

Preferably, Diclofenac is milled with lactose mono-hydrate. Preferably, Meloxicam is milled with mannitol. Preferably, Diclofenac is milled with mannitol. Preferably, Meloxicam is milled with glucose. Preferably, Diclofenac is milled with glucose. . Preferably, Meloxicam is milled with microcrystalline cellulose. Preferably, diclofenac in microcrystalline cellulose. Preferably, Meloxicam is milled with Tartaric acid. Preferably, Meloxicam is milled with lactose monohydrate. Preferably, Meloxicam is milled with mannitol. Preferably, Diclofenac is milled with lactose mono-hydrate and sodium dodecyl sulfate. Preferably, Meloxicam is milled with lactose monohydrate and sodium dodecyl sulfate.

In another preferred embodiment, a facilitating agent or combination of facilitating agents is used. Preferably, the facilitating agent is selected from the group consisting of: surface stabilizers, binding agents, filling agents, lubricating agents, sweeteners, flavouring agents, preservatives, buffers, wetting agents, disintegrants, effervescent agents, agents that may form part of a medicament, including a solid dosage form and other excipient required for specific drug delivery. Preferably, the facilitating agent is added during dry milling. Preferably, the facilitating agent is added to the milled biologically active material and grinding matrix and further processed in a mechanofusion process. Mechanofusion milling causes mechanical energy to be applied to powders or mixtures of particles in the micrometre and nanometre. The

reasons for including facilitating agents include, but are not limited to providing better dispersibility, control of agglomeration, the release or retention of the active particles from the delivery matrix. Examples of facilitating agents include, but are not limited to stearic acid, magnesium stearate, calcium stearate, sodium stearyl fumarate, sodium stearyl lactylate, zinc stearate, sodium stearate or lithium stearate, other solid state fatty acids such as oleic acid, lauric acid, palmitic acid, erucic acid, behenic acid, or derivatives (such as esters and salts), amino acids such as leucine, isoleucine, lysine, valine, methionine, phenylalanine, aspartame or acesulfame K. In a preferred aspect of manufacturing this formulation the facilitating agent is added to the milled mixture of biologically active material and co-grinding matrix and further processed in another milling device such as Mechonofusion, Cyclomixing, or impact milling such as ball milling, jet milling, or milling using a high pressure homogeniser, or combinations thereof. In a highly preferred aspect the facilitating agent is added to the milling of the mixture of biologically active material and co-grinding matrix as some time before the end of the milling process.

Preferably, the facilitating agent is added to the dry milling at a time selected from the group consisting of: with 1-5 % of the total milling time remaining, with 1-10 % of the total milling time remaining, with 1-20 % of the total milling time remaining, with 1-30 % of the total milling time remaining, with 2-5% of the total milling time remaining, with 2-10% of the total milling time remaining, with 5-20% of the total milling time remaining and with 5-20% of the total milling time remaining.

In another preferred embodiment, a disintegrant is selected from the group consisting of: crosslinked PVP, cross linked carmellose and sodium starch glycolate.

In another preferred embodiment, the dissolution profile of the measurement sample or prototype formulation thereof is improved by a factor selected from the group consisting of: wherein X is reached in 10 minutes, wherein X is reached within 10-20 minutes, wherein X is reached within 10-30 mins, wherein X is reached within 10-40 mins, wherein X is reached within 10-50 mins, wherein X is reached within 20-30 mins, wherein X is reached within 20-40 mins, wherein X is reached within 20-50 mins, wherein X is reached within 30-40 mins, wherein X is reached within 30-50 mins and wherein X is reached within 40-50 mins, wherein X is defined as the concentration equal to the dissolution concentration achieved by a control sample or prototype formulation thereof of the biologically active material or compound after 60 minutes.

In another preferred embodiment, the dissolution profile of the measurement sample or prototype formulation thereof is improved by a factor selected from the group consisting of: wherein Y is reached in 5 minutes, wherein Y is reached within 10 minutes, wherein Y is reached within 10-15 mins, wherein Y is reached within 10-20 mins, wherein Y is reached within 10-25 mins, wherein Y is reached within 15-20 mins, wherein Y is reached within 15-25 mins, wherein Y is reached within 20-25 mins, wherein Y is defined as the concentration equal

to the dissolution concentration achieved by a control sample (or prototype formulation thereof) of the biologically active material or compound after 30 minutes.

In a second aspect the invention comprises a biologically active material produced by the method described herein and composition comprising the biologically active material as described herein. Preferably, the particles have an average particle size equal or greater than 1 μ m determined on a particle number average basis. Preferably, the average particle size of the biologically active material has been reduced by a factor selected from the group consisting of: less than 5%, less than 10%, less than 20%, less than 30%, less than 40%, less than 50%, less than 60%, less than 70%, less than 80%, less than 90%, less than 95% and less than 99%. Preferably, the average particle size falls within the range selected from the group consisting of: 1-1000 μ m, 1-500 μ m, 1-300 μ m, 1-200 μ m, 1-150 μ m, 1-100 μ m, 1-50 μ m, 1-20 μ m, 1-10 μ m, 1-7.5 μ m, 1-5 μ m and 1-2 μ m. Preferably, the particles have a median particle size selected from the group consisting of: equal or greater than 1 μ m; and equal or greater than 2 μ m, wherein the median particle size is determined on a particle volume basis. Preferably, the percentage of particles with an average particle size greater than 1 μ m on a particle volume basis is a percentage selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 100%. Preferably, the percentage of particles with an average particle size greater than 2 μ m on a particle volume basis is a percentage selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 100%. Preferably, the median particle size has been reduced by a factor selected from the group consisting of: less than 5%, less than 10%, less than 20%, less than 30%, less than 40%, less than 50%, less than 60%, less than 70%, less than 80%, less than 90 %, less than 95% and less than 99%. Preferably, the median particle size falls within the range selected from the group consisting of: 1-1000 μ m, 1-500 μ m, 1-300 μ m, 1-200 μ m, 1-150 μ m, 1-100 μ m, 1-50 μ m, 1-20 μ m, 1-10 μ m, 1-7.5 μ m, 1-5 μ m 1-2 μ m, 2-1000 μ m, 2-500 μ m, 2-300 μ m, 2-200 μ m, 2-150 μ m, 2-100 μ m, 2-50 μ m, 2-20 μ m, 2-10 μ m, 2-7.5 μ m and 2-5 μ m. Preferably, the crystallinity profile of the biologically active material is selected from the group consisting of: at least 50% of the biologically active material is crystalline, at least 60% of the biologically active material is crystalline, at least 70% of the biologically active material is crystalline, at least 75% of the biologically active material is crystalline, at least 85% of the biologically active material is crystalline, at least 90% of the biologically active material is crystalline, at least 95% of the biologically active material is crystalline and at least 98% of the biologically active material is crystalline. Preferably, the crystallinity profile of the biologically active material is substantially equal to the crystallinity profile of the biologically active material before the material was subject to the method described herein. Preferably, the amorphous content of the biologically active material is selected from the group consisting of: less than 50% of the biologically active material is amorphous, less than 40% of the biologically active material is amorphous, less than 30% of the biologically active material is amorphous, less than 25% of the biologically active material is amorphous, less than 15% of the biologically active material is amorphous, less than

10% of the biologically active material is amorphous, less than 5% of the biologically active material is amorphous and less than 2% of the biologically active material is amorphous. Preferably, the biologically active material has had no significant increase in amorphous content following subjecting the material to the method as described herein. Preferably, the biologically active material is selected from the group consisting of: fungicides, pesticides, herbicides, nutraceuticals, pharmaceutical actives, biologics, amino acids, proteins, peptides, nucleotides, nucleic acids and analogs, homologs and first order derivatives thereof. Preferably, the biologically active material is selected from the group consisting of: anti-obesity drugs, central nervous system stimulants, carotenoids, corticosteroids, elastase inhibitors, anti-fungals, oncology therapies, anti-emetics, analgesics, cardiovascular agents, anti-inflammatory agents, such as NSAIDs and COX-2 inhibitors, anthelmintics, anti-arrhythmic agents, antibiotics (including penicillins), anticoagulants, antidepressants, antidiabetic agents, antiepileptics, antihistamines, antihypertensive agents, antimuscarinic agents, antimycobacterial agents, antineoplastic agents, immunosuppressants, antithyroid agents, antiviral agents, anxiolytics, sedatives (hypnotics and neuroleptics), astringents, alpha-adrenergic receptor blocking agents, beta-adrenoceptor blocking agents, blood products and substitutes, cardiac inotropic agents, contrast media, cough suppressants (expectorants and mucolytics), diagnostic agents, diagnostic imaging agents, diuretics, dopaminergics (anti-parkinsonian agents), haemostatics, immunological agents, lipid regulating agents, muscle relaxants, parasympathomimetics, parathyroid calcitonin and biphosphonates, prostaglandins, radio-pharmaceuticals, sex hormones (including steroids), anti-allergic agents, stimulants and anoretics, sympathomimetics, thyroid agents, vasodilators, and xanthines. Preferably, the biologically active material is selected from the group consisting of: indomethacin, diclofenac, naproxen, meloxicam, metaxalone, cyclosporin A, progesterone celecoxib, cilostazol, ciprofloxacin, 2,4-dichlorophenoxyacetic acid, anthraquinone, creatine monohydrate, glyphosate, halosulfuron, mancozeb, metsulfuron, salbutamol, sulphur, tribenuran and estradiol or any salt or derivative thereof.

In one preferred embodiment, the invention comprises compositions comprising the biologically active ingredient together with a grinding matrix, a mixture of grinding matrix materials, milling aids, mixtures of milling aids, facilitating agents and/or mixtures of facilitating agents as described herein, in concentrations and ratios as described herein under the methods of the invention.

In a third aspect the invention comprises a pharmaceutical composition comprising a biologically active material produced by the method described herein and compositions described herein. Preferably, the invention comprises pharmaceutical compositions comprising the biologically active ingredient together with a grinding matrix, a mixture of grinding matrix materials, milling aids, mixtures of milling aids, facilitating agents and/or mixtures of facilitating agents as described herein, in concentrations and ratios as described herein under the

methods of the invention. Preferably, the particles have an average particle size equal or greater than 1 μ m determined on a particle number basis. Preferably, the average particle size of the biologically active material has been reduced by a factor selected from the group consisting of: less than 5%, less than 10%, less than 20%, less than 30%, less than 40%, less than 50%, less than 60%, less than 70%, less than 80%, less than 90 %, less than 95% and less than 99%. Preferably, the average particle size falls within the range selected from the group consisting of: 1-1000 μ m, 1-500 μ m, 1-300 μ m, 1-200 μ m, 1-150 μ m, 1-100 μ m, 1-50 μ m, 1-20 μ m, 1-10 μ m, 1-7.5 μ m, 1-5 μ m and 1-2 μ m. Preferably, the particles have a median particle size selected from the group consisting of: equal or greater than 1 μ m; and equal or greater than 2 μ m, wherein the median particle size is determined on a particle volume basis. Preferably, the percentage of particles with an average particle size greater than 1 μ m on a particle volume basis is a percentage selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 100%. Preferably, the percentage of particles with an average particle size greater than 2 μ m on a particle volume basis is a percentage selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 100%. Preferably, the median particle size has been reduced by a factor selected from the group consisting of: less than 5%, less than 10%, less than 20%, less than 30%, less than 40%, less than 50%, less than 60%, less than 70%, less than 80%, less than 90 %, less than 95% and less than 99%. Preferably, the median particle size falls within the range selected from the group consisting of: 1-1000 μ m, 1-500 μ m, 1-300 μ m, 1-200 μ m, 1-150 μ m, 1-100 μ m, 1-50 μ m, 1-20 μ m, 1-10 μ m, 1-7.5 μ m, 1-5 μ m 1-2 μ m, 2-1000 μ m, 2-500 μ m, 2-300 μ m, 2-200 μ m, 2-150 μ m, 2-100 μ m, 2-50 μ m, 2-20 μ m, 2-10 μ m, 2-7.5 μ m and 2-5 μ m. Preferably, the crystallinity profile of the biologically active material is selected from the group consisting of: at least 50% of the biologically active material is crystalline, at least 60% of the biologically active material is crystalline, at least 70% of the biologically active material is crystalline, at least 75% of the biologically active material is crystalline, at least 85% of the biologically active material is crystalline, at least 90% of the biologically active material is crystalline, at least 95% of the biologically active material is crystalline and at least 98% of the biologically active material is crystalline. Preferably, the crystallinity profile of the biologically active material is substantially equal to the crystallinity profile of the biologically active material before the material was subject to the method as described herein. Preferably, the amorphous content of the biologically active material is selected from the group consisting of: less than 50% of the biologically active material is amorphous, less than 40% of the biologically active material is amorphous, less than 30% of the biologically active material is amorphous, less than 25% of the biologically active material is amorphous, less than 15% of the biologically active material is amorphous, less than 10% of the biologically active material is amorphous, less than 5% of the biologically active material is amorphous and less than 2% of the biologically active material is amorphous. Preferably, the biologically active material has no significant increase in amorphous content after subjecting the material to the method as described herein. Preferably,

the biologically active material is selected from the group consisting of: new chemical entities, pharmaceutical actives, biologics, amino acids, proteins, peptides, nucleotides, nucleic acids and analogs, homologs and first order derivatives thereof. Preferably, the biologically active material is selected from the group consisting of: anti-obesity drugs, central nervous system stimulants, carotenoids, corticosteroids, elastase inhibitors, anti-fungals, oncology therapies, anti-emetics, analgesics, cardiovascular agents, anti-inflammatory agents, such as NSAIDs and COX-2 inhibitors, anthelmintics, anti-arrhythmic agents, antibiotics (including penicillins), anticoagulants, antidepressants, antidiabetic agents, antiepileptics, antihistamines, antihypertensive agents, antimuscarinic agents, antimycobacterial agents, antineoplastic agents, immunosuppressants, antithyroid agents, antiviral agents, anxiolytics, sedatives (hypnotics and neuroleptics), astringents, alpha-adrenergic receptor blocking agents, beta-adrenoceptor blocking agents, blood products and substitutes, cardiac inotropic agents, contrast media, cough suppressants (expectorants and mucolytics), diagnostic agents, diagnostic imaging agents, diuretics, dopaminergics (anti-parkinsonian agents), haemostatics, immunological agents, lipid regulating agents, muscle relaxants, parasympathomimetics, parathyroid calcitonin and biphosphonates, prostaglandins, radio-pharmaceuticals, sex hormones (including steroids), anti-allergic agents, stimulants and anoretics, sympathomimetics, thyroid agents, vasodilators, and xanthines. Preferably, the biologically active material is selected from the group consisting of: indomethacin, diclofenac, naproxen, meloxicam, metaxalone, cyclosporin A, progesterone celecoxib, cilostazol, ciprofloxacin, 2,4-dichlorophenoxyacetic acid, anthraquinone, creatine monohydrate, glyphosate, halosulfuron, mancozeb, metsulfuron, salbutamol, sulphur, tribenuran and estradiol or any salt or derivative thereof..

Preferably cosmeceuticals, cosmetics, complementary medicines, natural products, vitamins, nutrients and nutraceuticals are selected from the group consisting of: Glycolic acids, Lactic acids, Carrageenan, Almonds, Mahogany wood, Andrographis Paniculata, Aniseed, Anthemis nobilis (chamomile), Apricot kernel, leaves of bearberry, leaves of cranberry, leaves of blueberry, leaves of pear trees, beta-carotene, black elderberry, black raspberry, black walnut shell, blackberry, bladderwrack, bletilla striata, borage seed, boysenberry, brazil nut, burdock root, butcher's broom extract, calamine, calcium gluconate, calendula, carnosic acid, Cantella asiatica, charcoal, chaste tree fruit, Chicory root extract, chitosan, choline, cichorium intybus, clematis vitalba, coffea Arabica, coumarin, crithmum maritimum, curcumin, coffee, cocoa, cocoa powder, cocoa nibs, cocoa mass, cocoa liquor, cocoa products, dogwood, Echinacea, echium lycopsis, anise, atragalus, bilberry, bitter orange, black cohosh, cat's claw, chamomile, chasteberry, cranberry, dandelion, Echinacea, ephedra, European elder Epilobium angustifolium, horse chestnut, cloves, evening primrose, fennel seed, fenugreek, feverfew, flaxseed, fumaria officinalis, garlic, geranium, ginger, ginkgo, ginseng, goldenseal, grape seed, green tea, guava, hawthorn, hayflower, hazelnut, helichrysum, hoodia, horseradish, mulbe

italicum, hibiscus, hierochloe odorata, hops, horse chestnut, ilex paraguariensis, indian gooseberry, irish moss, juniper berry, kudzu root, lady's thistle, lavender, lemongrass, lentius edodes, licorice, longifolene, loquat, lotus seed, luffa cylindrica, lupine, maroinberry, marjoram, meadowsweet, milk vetch root, mimosa tenuiflora, mistletoe, mulberry, noni, kelp, oatmeal, oregano, papaya, parsley, peony root, pomegranate, pongamia glabra seed, pongamia pinnata, quinoa seed, red raspberry, rose hip, rosemary, sage, saw palmetto, soy bean, szechuan peppercorn, tephrosia purpurea, terminalia catappa, terminalia sericea, thunder god vine, thyme, turmeric, valeriana officinalis, walnuts, white tea leaf, yam, witch hazel, wormwood, yarrow, valerian, yohimbe, mangosteen, sour sob, goji berry, spirulina and durian skin.

In a fourth aspect the invention comprises a method of treating a human in need of such treatment comprising the step of administering to the human an effective amount of a pharmaceutical composition as described herein.

In a fifth aspect the invention comprises a method for manufacturing a pharmaceutical composition as described herein comprising the step of combining a therapeutically effective amount of a biologically active material prepared by a method described herein together with a pharmaceutically acceptable carrier to produce a pharmaceutically acceptable dosage form.

In a sixth aspect the invention comprises a method for manufacturing a veterinary product comprising the step of combining a therapeutically effective amount of the biologically active material prepared by a method as described herein together with an acceptable excipient to produce a dosage form acceptable for veterinary use.

In a seventh aspect the invention comprises a method for manufacturing an agricultural product comprising the step of combining an effective amount of the biologically active material prepared by a method described herein together with acceptable excipients to produce a formulation such as, but not limited to a water dispersible granule, wettable granule, dry flowable granule or soluble granule that is used to prepare a solution for use in agricultural applications. Preferably, the product is selected from the group consisting of: herbicides, pesticides, seed treatments, herbicide safeners, plant growth regulators and fungicides. The methods of the invention can be used to increase the dissolution of the biologically active material particles in water or other solvents, resulting in better, faster or more complete preparation and mixing. This will result in a more consistent product performance such as better weed, disease and pest control and other practical benefits such as faster machinery, tank and sprayer cleanout, less rinsate, and a reduced impact on the environment.

In a future aspect the invention comprises a method for manufacturing an agricultural product comprising the step of combining an effective amount of the biologically active material prepared by a method described herein together with acceptable excipients to produce a formulation such as, but not limited to a water dispersible granule, wettable granule, wettable powder or a powder for seed treatment that is used to prepare a dry powder or particle suspension for use in agricultural applications. Preferably, the product is selected from the

group consisting of: herbicides, pesticides, seed treatments, herbicide safeners, plant growth regulators and fungicides. Another preferred aspect of the method of invention would be to produce powders that have active particles with a high surface area. Such powders would provide better performance in areas such as seed treatment where dry powders are applied to seeds as fungicides, herbicide safeners, plant growth regulators and other treatments. The higher surface area would provide more activity per mass of active used. In another preferred aspect actives such as pesticides, fungicides and seed treatments subject to the method of invention are formulated to produce suspensions of the actives when added to water or other solvents. As these suspensions will have particles of very small size and high surface area they will possess at least three highly desirable traits. The first is that small particles with high surface area will adhere better to surfaces such as leafs and other foliage that the suspension is applied to. This will result in better rain fastness and a longer period of activity. The second aspect is that smaller particles with a higher surface area deliver superior coverage per unit mass of active applied. For example, if 100 particles are needed on a leaf and if the particle diameter is reduced to one third of the former diameter by the methods of this invention, then the dosage can be reduced to about 11% of the former dosage, resulting in lower cost, less residue on harvested crops, and mitigation of environmental impact. In the third aspect the smaller particles will deliver better bioavailability. With many low solubility actives, such as fungicides and pesticides the particles that adhere to plant material slowly dissolve over days and weeks providing continued protection from disease and pests. With this method of invention able to deliver better bioavailability in many circumstances it will be possible to reduce the amount of active that needs to be applied. As with the second aspect such an outcome would lower costs, minimize residues and mitigate environmental impact. In a highly preferred aspect of the invention the powder produced in the milling process would be subject to a process such as wet or dry granulation that makes the powder free flowing and low in dust content yet easily dispersible once in water or other solvent.

Preferably the biologically active material is a herbicide, pesticide, seed treatment, herbicide safener, plant growth regulator or fungicide selected from the group consisting of: 2-phenylphenol, 8-hydroxyquinoline sulfate, acibenzolar, allyl alcohol, azoxystrobin, basic benomyl, benzalkonium chloride, biphenyl, blasticidin-S, Bordeaux mixture, Boscalid, Burgundy mixture, butylamine, Cadendazim, calcium polysulfide, Captan, carbamate fungicides, carbendazim, carvone, chloropicrin, chlorothalonil, ciclopirox, clotrimazole, conazole fungicides, Copper hydroxide, copper oxychloride, copper sulfate, copper(II) carbonate, copper(II) sulfate, cresol, cryprodinil, cuprous oxide, cycloheximide, Cymoxanil, DBCP, dehydroacetic acid, dicarboximide fungicides, difenoconazole, dimethomorph, diphenylamine, disulfiram, ethoxyquin, famoxadone, fenamidone, Fludioxonil, formaldehyde, fosetyl, Fosetyl-aluminium, furfural, griseofulvin, hexachlorobenzene, hexachlorobutadiene, hexachlorophene, hexaconazole, imazalil, Imidacloprid, iodomethane, Iprodione, Lime sulfur, mancozeb, mercuric

chloride, mercuric oxide, mercurous chloride, Metalaxyl, metam, methyl bromide, methyl isothiocyanate, metiram, natamycin, nystatin, organotin fungicides, oxythioquinox, pencycuron, pentachlorophenol, phenylmercury acetate, potassium thiocyanate, procymidone, propiconazole, propineb, pyraclostrobin, pyrazole fungicides, pyridine fungicides, pyrimethanil, pyrimidine fungicides, pyrrole fungicides, quinoline fungicides, quinone fungicides, sodium azide, streptomycin, sulfur, Tebucanazole, thiabendazole, thiomersal, tolnaftate, Tolyfluanid, triadimorsol, tributyltin oxide, Trifloxystrobin, triflumuron, Undecylenic acid, urea fungicides, vinclozolin, Ziram, 3-dihydro-3-methyl-1, 3-thiazol-2-ylidene-xylylene, 4-D esters, 4-DB esters, 4-parathion methyl, Acetamiprid, acetonitrile, acrinathrin, alachlor, allethrin, alpha-cypermethrin, Aluminium phosphide, amitraz, anilophos, azaconazole, azinphos-ethyl, azinphos-methyl, benalaxyl, benfluralin, benfuracarb, benfuresate, bensulide, benzoximate, benzoylprop-ethyl, betacyfluthrin, beta-cypermethrin, bifenox, bifenthrin, binapacryl, bioallethrin, bioallethrin S, bioresmethrin, biteranol, Brodifacoum, bromophos, bromopropylate, bromoxynil, bromoxynil esters, bupirimate, buprofezin, butacarboxim, butachlor, butamifos, butoxycarboxin, butralin, butylate, calcium sulfate, cambda-cyhalothrin, carbetamide, carboxin, chlordimeform, chlorfenvinphos, chlorflurazuron, chlormephos, chlornitrofen, chlorobenzilate, chlorophoxim, chloropropylate, chlorpropham, Chlorpyrifos, chlorpyrifos-methyl, cinmethylin, clethodim, clomazone, clopyralid esters, CMPP esters, cyanophos, cycloate, cycloprothrin, cycloxydim, cyfluthrin, cyhalothrin, cypermethrin, cyphenothrin, cyproconazole, deltamethrin, demeton-S-methyl, desmedipham, dichlorprop esters, dichlorvos, diclofop-methyldiethyl, dicofol, difenoconazole, dimethachlor, dimethomorph, diniconazole, dinitramine, dinobuton, dioxabenzafos, dioxacarb, disulfoton, ditalimfos, dodemorph, dodine, edifenphos, emamectin, emperthrin, endosulfan, EPNethiofencarb, epoxyconazole, esfenvalerate, ethalfluralin, ethofumesate, ethoprophos, ethoxyethyl, etofenprox, etridiazole, etrimphos, Famoxadone, fenamiphos, fenarimol, fenazaquin, fenitrothion, fenobucarb, fenoxapropethyl, fenoxycarb, fenpropathrin, fenpropidin, fenpropimorph, fenthion, fenvalerate, fluazifop, fluazifop-P, fluchloralin, flucythrinate, flufenoxim, flufenoxuron, flumetralin, fluorodifen, fluoroglycofen ethyl, fluoroxyppyr esters, flurecol butyl, flurochloralin, flusilazole, formothion, gamma-HCH, haloxyfop, haloxyfop-methyl, hexaflumuron, hydroprene, imibenconazole, indoxacarb, ioxynil esters, isofenphos, isoprocarb, isopropalin, isoxathion, malathion, maneb, MCPA esters, mecoprop-P esters, mephospholan, Metaldehyde, methidathion, Methomyl, methoprene, methoxychlor, metolachlor, mevinphos, monalide, myclobutanil, N-2, napropamide, nitrofen, nuarimol, oxadiazon, oxycarboxin, oxyfluorfen, penconazole, pendimethalin, permethrin, phenisopham, phenmedipham, phenothrin, phenthoate, phosalone, phosfolan, phosmet, picloram esters, pirimicarb, pirimiphos-ethyl, pirimiphos-methyl, pretilachlor, prochloraz, profenofos, profluralin, promecarb, propachlor, propanil, propaphos, propaquizafop, propargite, propetamphos, pymetrozine, pyrachlofos, pyridate, pyrifenox, quinalphos, quizalofop-P, resmethrin, Spinetoram J, Spinetoram L, Spinosad A, Spinosad B,

tau-fluvalinate, tebuconazole, Tebufenozide, tefluthrin, temephos, terbufos, tetrachlorinphos, tetraconazole, tetradifon, tetramethrin, Thiamethoxam, tolclofos-methyl, tralomethrin, triadimefon, triadimenol, triazophos, triclopyr esters, tridemorph, tridiphane, triflumizole, trifluralin, xylylcarb, 3-dihydro-3-methyl-1, 3-thiazol-2-ylidene-xylydene, 4-D esters, 4-DB esters, 4-parathion methyl, Acetamiprid, acetochlor, aclonifen, acrinathrin, alachlor, allethrin, alpha-cypermethrin, Aluminium phosphide, amitraz, anilophos, azaconazole, azinphos-ethyl, azinphos-methyl, benalaxyl, benfluralin, benfuracarb, benfuresate, bensulide, benzoximate, benzoylprop-ethyl, betacyfluthrin, beta-cypermethrin, bifenox, bifenthrin, binapacryl, bioallethrin, bioallethrin S, bioresmethrin, biteranol, Brodifacoum, bromophos, bromopropylate, bromoxynil, bromoxynil esters, bupirimate, buprofezin, Butacarboxim, butachlor, butamifos, butoxycarboxin, butralin, butylate, calcium sulfate, cambda-cyhalothrin, carbetamide, carboxin, chlordimeform, chlorfenvinphos, chlorflurazuron, chlormephos, chlornitrofen, chlorobenzilate, chlorophoxim, chloropropylate, chlorpropham, Chlorpyrifos, chlorpyrifos-methyl, cinmethylin, clethodim, clomazone, clopyralid esters, CMPP esters, cyanophos, cycloate, cycloprothrin, cycloxydim, cyfluthrin, cyhalothrin, cypermethrin, cyphenothrin, cyproconazole, deltamethrin, demeton-S-methyl, desmedipham, dichlorprop esters, dichlorvos, diclofop-methyldiethyl, dicofol, dimethachlor, dimethomorph, diniconazole, dinitramine, dinobuton, dioxabenzafos, dioxacarb, disulfoton, ditalimfos, dodemorph, dodine, edifenphos, emamectin, empenthrin, endosulfan, EPNethiofencarb, epoxyconazole, esfenvalerate, ethalfluralin, ethofumesate, ethoprophos, ethoxyethyl, ethoxyquin, etofenprox, etridiazole, etrimphos, fenamiphos, fenarimol, fenazaquin, fenitrothion, fenobucarb, fenoxapropethyl, fenoxycarb, fenpropathrin, fenpropidin, fenpropimorph, fenthion, fenvalerate, fluazifop, fluazifop-P, fluchloralin, flucythrinate, flufenoxim, flufenoxuron, flumetralin, fluorodifen, fluoroglycofen ethyl, fluoroxypyr esters, flurecol butyl, flurochloralin, flusilazole, formothion, gamma-HCH, haloxyfop, haloxyfop-methyl, hexaflumuron, hydroprene, imibenconazole, indoxacarb, ioxynil esters, isofenphos, isoprocarb, isopropalin, isoxathion, malathion, maneb, MCPA esters, mecoprop-P esters, mephospholan, Metaldehyde, methidathion, Methomyl, methoprene, methoxychlor, mevinphos, monalide, myclobutanil, myclobutanil, N-2, napropamide, nitrofen, nuarimol, oxadiazon, oxycarboxin, oxyfluorfen, penconazole, permethrin, phenisopham, phenmedipham, phenothrin, phenthoate, phosalone, phosfolan, phosmet, picloram esters, pirimicarb, pirimiphos-ethyl, pirimiphos-methyl, pretilachlor, prochloraz, profenofos, profluralin, promecarb, propachlor, propanil, propaphos, propaquizafop, propargite, propetamphos, pymetrozine, pyridate, pyrifenox, quinalphos, quizalofop-P, resmethrin, Spinetoram J, Spinetoram L, Spinosad A, Spinosad B, tau-fluvalinate, Tebufenozide, tefluthrin, temephos, terbufos, tetrachlorinphos, tetraconazole, tetradifon, tetramethrin, Thiamethoxam, tolclofos-methyl, tralomethrin, triadimenol, triazophos, triclopyr esters, tridemorph, tridiphane, triflumizole, trifluralin, xylylcarb and any combination thereof.

In an eighth aspect the invention comprises a method for manufacturing of a pharmaceutical formulation comprising the step of combining an effective amount of the biologically active material prepared by a method described herein together with acceptable excipients to produce a formulation that can deliver a therapeutically effective amount of active to the pulmonary or nasal area. Such a formulation could be, but is not limited to a dry powder formulation for oral inhalation to the lungs or a formulation for nasal inhalation. Preferably the method for manufacturing such a formulation uses lactose, mannitol, sucrose, sorbitol, xylitol or other sugars or polyols as the co-grinding matrix together with surfactant such as, but not limited to lecithin, DPPC (dipalmitoyl phosphatidylcholine), PG (phosphatidylglycerol), dipalmitoyl phosphatidyl ethanolamine (DPPE), dipalmitoyl phosphatidylinositol (DPPI) or other phospholipid. The particle size of the material produced by the invention disclosed herein results in the materials being readily aerosolized and suitable for methods of delivery to a subject in need thereof, including pulmonary and nasal delivery methods.

While the method of the present invention has particular application in the preparation of poorly water-soluble biologically active materials, the scope of the invention is not limited thereto. For example, the method of the present invention enables production of highly water-soluble biologically active materials. Such materials may exhibit advantages over conventional materials by way of, for example, more rapid therapeutic action or lower dose. In contrast, wet grinding techniques utilizing water (or other comparably polar solvents) are incapable of being applied to such materials, as the particles dissolve appreciably in the solvent.

Other aspects and advantages of the invention will become apparent to those skilled in the art from a review of the ensuing description.

Brief Description of the Drawings

Figure 1 shows the particle size distribution of Meloxicam milled in Lactose for 1 minute (B) or 2 minutes (C), respectively, compared to the particle size distribution of commercially available Meloxicam (A).

Figure 2 shows the dissolution of Meloxicam milled in Lactose for 1 minute (B) or 2 minutes (C), respectively, compared to the dissolution of commercially available Meloxicam (A).

Figure 3 shows the particle size distribution of Diclofenac milled in Lactose for 1 minute (B) or 2 minutes (C), respectively, compared to the particle size distribution of commercially obtained Diclofenac (A).

Figure 4 shows the dissolution of Diclofenac milled in Lactose for 1 minute (B) or 2 minutes (C), respectively, compared to the dissolution of commercially available Diclofenac (A).

Figure 5 shows the Differential Scanning Calorimetry (DSC) traces of mannitol, 10% meloxicam milled in mannitol for 2 minutes (example 3) and 20% meloxicam milled in mannitol for 2 minutes (example 11).

Figure 6 shows the XRD spectra of Meloxicam (A), milled lactose monohydrate (B), Meloxicam milled in Lactose at 20% for 2 minutes (example 10) (C) and Meloxicam milled in Lactose with 1% SDS at 50% for 10 minutes (example 17) (D).

Figure 7 shows the XRD spectra of Meloxicam (A), mannitol (B), a physical mixture of 20 % Meloxicam in Lactose (C) and Meloxicam milled in mannitol at 20% for 2 minutes (example 11) (D).

Figure 8 shows the XRD spectra of Diclofenac milled in Lactose with 1% SDS at 20% for 10 minutes (A), Diclofenac milled in Lactose with 1% SDS at 30% for 10 minutes (example 12) (B), Diclofenac milled in Lactose with 1% SDS at 40% for 10 minutes (example 13) (C) and Diclofenac milled in Lactose with 1% SDS at 50% for 10 minutes (example 14) (D).

Figure 9 shows the XRD spectra of a physical mixture of 20% Diclofenac in Lactose with 1% SDS (A), 30% Diclofenac in Lactose with 1% SDS (B), 40% Diclofenac in Lactose with 1% SDS (C) and 50% Diclofenac in Lactose with 1% SDS (D).

Figure 10 shows the XRD spectra of a Diclofenac acid (A), Lactose monohydrate (B) and milled Lactose monohydrate (C).

Figure 11 shows the XRD spectra of a Meloxicam (A), a physical mixture of 50% Meloxicam in Lactose with 1% SDS (B) and milled Lactose monohydrate (C).

Detailed Description of the Invention

General

Those skilled in the art will appreciate that the invention described herein is susceptible to variations and modifications other than those specifically described. It is to be understood that the invention includes all such variations and modifications. The invention also includes all of the steps, features, compositions and materials referred to or indicated in the specification, individually or collectively and any and all combinations or any two or more of the steps or features.

The present invention is not to be limited in scope by the specific embodiments described herein, which are intended for the purpose of exemplification only. Functionally equivalent products, compositions and methods are clearly within the scope of the invention as described herein.

The invention described herein may include one or more ranges of values (e.g. size, concentration etc). A range of values will be understood to include all values within the range, including the values defining the range, and values adjacent to the range that lead to the same or substantially the same outcome as the values immediately adjacent to that value which defines the boundary to the range.

The entire disclosures of all publications (including patents, patent applications, journal articles, laboratory manuals, books, or other documents) cited herein are hereby incorporated by

reference. Inclusion does not constitute an admission is made that any of the references constitute prior art or are part of the common general knowledge of those working in the field to which this invention relates.

Throughout this specification, unless the context requires otherwise, the word "comprise" or variations, such as "comprises" or "comprising" will be understood to imply the inclusion of a stated integer, or group of integers, but not the exclusion of any other integers or group of integers. It is also noted that in this disclosure, and particularly in the claims and/or paragraphs, terms such as "comprises", "comprised", "comprising" and the like can have the meaning attributed to it in US Patent law; e.g., they can mean "includes", "included", "including", and the like.

"Therapeutically effective amount" as used herein with respect to methods of treatment and in particular drug dosage, shall mean that dosage that provides the specific pharmacological response for which the drug is administered in a significant number of subjects in need of such treatment. It is emphasized that "therapeutically effective amount," administered to a particular subject in a particular instance will not always be effective in treating the diseases described herein, even though such dosage is deemed a "therapeutically effective amount" by those skilled in the art. It is to be further understood that drug dosages are, in particular instances, measured as oral dosages, or with reference to drug levels as measured in blood.

The term "inhibit" is defined to include its generally accepted meaning which includes prohibiting, preventing, restraining, and lowering, stopping, or reversing progression or severity, and such action on a resultant symptom. As such the present invention includes both medical therapeutic and prophylactic administration, as appropriate.

The term "biologically active material" is defined to mean a biologically active compound or a substance which comprises a biologically active compound. In this definition, a compound is generally taken to mean a distinct chemical entity where a chemical formula or formulas can be used to describe the substance. Such compounds would generally, but not necessarily be identified in the literature by a unique classification system such as a CAS number. Some compounds may be more complex and have a mixed chemical structure. For such compounds they may only have a empirical formula or be qualitatively identified. A compound would generally be a pure material, although it would be expected that up to 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% of the substance could be other impurities and the like. Examples of biologically active compounds are, but not limited to, pharmaceutical actives, fungicides, pesticides, herbicides, nutraceuticals, cosmeceuticals, cosmetics, complementary medicines, natural products, vitamins, nutrients, biologics, amino acids, proteins, peptides, nucleotides, nucleic acids. A substance that contains a biologically active compound is any substance which has as one of its components a biologically active compound... Examples of substances containing biologically active compounds are, but not limited to, pharmaceutical formulations and products, cosmetic formulations and products, industrial formulations and products,

agricultural formulations and products, foods, seeds, cocoa and cocoa solids, coffee, herbs, spices, other plant materials, minerals, animal products, shells and other skeletal material.

Any of the terms, "biological(ly) active", "active", "active material" shall have the same meaning as biologically active material.

The term "grinding matrix" is defined as any inert substance that a biologically active material can or is combined with and milled. The terms "co-grinding matrix" and "matrix" are interchangeable with "grinding matrix".

Particle Size

There are a wide range of techniques that can be utilized to characterize the particle size of a material. Those skilled in the art also understand that almost all these techniques do not physically measure the actually particle size, as one might measure something with a ruler, but measure a physical phenomena which is interpreted to indicate a particle size. As part of the interpretation process some assumptions need to be made to enable mathematical calculations to be made. These assumptions deliver results such as an equivalent spherical particle size, or a hydrodynamic radius.

Amongst these various methods, two types of measurements are most commonly used. Photon correlation spectroscopy (PCS), also known as 'dynamic light scattering' (DLS) is commonly used to measure particles with a size less than 10 micron. Typically this measurement yields an equivalent hydrodynamic radius often expressed as the average size of a number distribution. The other common particle size measurement is laser diffraction which is commonly used to measure particle size from 100 nm to 2000 micron. This technique calculates a volume distribution of equivalent spherical particles that can be expressed using descriptors such as the median particle size or the % of particles under a given size.

Those skilled in the art recognize that different characterization techniques such as photon correlation spectroscopy and laser diffraction measure different properties of a particle ensemble. As a result multiple techniques will give multiple answers to the question, "what is the particle size." In theory one could convert and compare the various parameters each technique measures, however, for real world particle systems this is not practical. As a result the particle size used to describe this invention will be given as two different sets of values that each relate to these two common measurement techniques, such that measurements could be made with either technique and then evaluated against the description of this invention.

For measurements made using a photo correlation spectroscopy instrument, or an equivalent method known in the art, the term "number average particle size" is defined as the average particle diameter as determined on a number basis.

For measurements made using a laser diffraction instrument, or an equivalent method known in the art, the term "median particle size" is defined as the median particle diameter as determined on an equivalent spherical particle volume basis. Where the term median is used, it

is understood to describe the particle size that divides the population in half such that 50% of the population is greater than or less than this size. The median particle size is often written as D50, D(0.50) or D[0.5] or similar. As used herein D50, D(0.50) or D[0.5] or similar shall be taken to mean 'median particle size'.

The term "Dx of the particle size distribution" refers to the xth percentile of the distribution; thus, D90 refers to the 90th percentile, D95 refers to the 95th percentile, and so forth. Taking D90 as an example this can often be written as, D(0.90) or D[0.9] or similar. With respect to the median particle size and Dx an upper case D or lowercase d are interchangeable and have the same meaning. Another commonly used way of describing a particle size distribution measured by laser diffraction, or an equivalent method known in the art, is to describe what % of a distribution is under or over a nominated size. The term "percentage less than" also written as "%<" is defined as the percentage, by volume, of a particle size distribution under a nominated size -for example the % < 1000 nm. The term "percentage greater than" also written as "%>" is defined as the percentage, by volume, of a particle size distribution over a nominated size -for example the % > 1000 nm.

The particle size used to describe this invention should be taken to mean the particle size as measured at or shortly before the time of use. For example, the particle size is measured 2 months after the material is subject to the milling method of this invention. In a preferred form, the particle size is measured at a time selected from the group consisting of: 1 day after milling, 2 days after milling, 5 days after milling, 1 month after milling, 2 months after milling, 3 months after milling, 4 months after milling, 5 months after milling, 6 months after milling, 1 year after milling, 2 years after milling, 5 years after milling.

For many of the materials subject to the methods of this invention the particle size can be easily measured. Where the active material has poor water solubility and the matrix it is milled in has good water solubility the powder can simply be dispersed in an aqueous solvent. In this scenario the matrix dissolves leaving the active material dispersed in the solvent. This suspension can then be measured by techniques such as PCS or laser diffraction.

Suitable methods to measure an accurate particle size where the active material has substantive aqueous solubility or the matrix has low solubility in a water based dispersant are outlined below.

1. In the circumstance where insoluble matrix such as microcrystalline cellulose prevents the measurement of the active material, separation techniques such as filtration or centrifugation could be used to separate the insoluble matrix from the active material particles. Other ancillary techniques would also be required to determine if any active material was removed by the separation technique so that this could be taken into account.
2. In the case where the active material is too soluble in water other solvents could be evaluated for the measurement of particle size. Where a solvent could be found that

active material is poorly soluble in but is a good solvent for the matrix a measurement would be relatively straight forward. If such a solvent is difficult to find another approach would be to measure the ensemble of matrix and active material in a solvent (such as iso-octane) which both are insoluble in. Then the powder would be measured in another solvent where the active material is soluble but the matrix is not. Thus with a measurement of the matrix particle size and a measurement of the size of the matrix and active material together an understanding of the active material particle size can be obtained.

3. In some circumstances image analysis could be used to obtain information about the particle size distribution of the active material. Suitable image measurement techniques might include transmission electron microscopy (TEM), scanning electron microscopy (SEM), optical microscopy and confocal microscopy. In addition to these standard techniques some additional technique would be required to be used in parallel to differentiate the active material and matrix particles. Depending on the chemical makeup of the materials involved possible techniques could be elemental analysis, raman spectroscopy, FTIR spectroscopy or fluorescence spectroscopy.

Other Definitions

Throughout this specification, unless the context requires otherwise, the phrase “dry mill” or variations, such as “dry milling”, should be understood to refer to milling in at least the substantial absence of liquids. If liquids are present, they are present in such amounts that the contents of the mill retain the characteristics of a dry powder.

“Flowable” means a powder having physical characteristics rendering it suitable for further processing using typical equipment used for the manufacture of pharmaceutical compositions and formulations.

Other definitions for selected terms used herein may be found within the detailed description of the invention and apply throughout. Unless otherwise defined, all other scientific and technical terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which the invention belongs.

The term “millable” means that the grinding matrix is capable of being physically degraded under the dry milling conditions of the method of the invention. In one embodiment of the invention, the milled grinding matrix is of a comparable particle size to the biologically active material. In another embodiment of the invention the particle size of the matrix is substantially reduced but not as small as the biologically active material

Other definitions for selected terms used herein may be found within the detailed description of the invention and apply throughout. Unless otherwise defined, all other scientific and technical terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which the invention belongs.

Specific

In one embodiment, the present invention is directed to a method for improving the dissolution profile of a biologically active material, the method comprising the step of:

dry milling a mixture of a solid biologically active material and a millable grinding matrix, in a mill comprising a plurality of milling bodies, to produce particles of a biologically active material dispersed in at least partially milled grinding matrix.

The mixture of active material and matrix may then be separated from the milling bodies and removed from the mill.

In one aspect the mixture of active material and matrix is then further processed. In another aspect, the grinding matrix is separated from the particles of biologically active material. In a further aspect, at least a portion of the milled grinding matrix is separated from the particulate biologically active material.

The milling bodies are essentially resistant to fracture and erosion in the dry milling process. The quantity of the grinding matrix relative to the quantity of biologically active material in particulate form, and the extent of milling of the grinding matrix, is sufficient to improve the dissolution profile of the active material milled.

The present invention also relates to biologically active materials produced by said methods, to medicaments produced using said biologically active materials and to methods of treatment of an animal, including man, using a therapeutically effective amount of said biologically active materials administered by way of said medicaments.

Improving the dissolution profile

The present invention leads to the improved dissolution profile. An improved dissolution profile has significant advantages including the improvement of bioavailability of the biologically active material *in vivo*.

Preferably, the improved dissolution profile is observed *in vitro*. Alternatively, the improved dissolution profile is observed *in vivo* by the observation of an improved bioavailability profile.

Standard methods for determining the dissolution profile of a material *in vitro* are available in the art. A suitable method to determine an improved dissolution profile *in vitro* may include determining the concentration of the sample material in a solution over a period of time and comparing the results from the sample material to a control sample. An observation that peak solution concentration for the sample material was achieved in less time than the control sample would indicate (assuming it is statistically significant), that the sample material has an improved dissolution profile.

The measurement sample is herein defined as the mixture of biologically active material with grinding matrix and/or other additives that has been subject to the processes of the invention described here. Herein a control sample is defined as a physical mixture (not subject to the processes described in this invention) of the components in the measurement sample with the

same relative proportions of active, matrix and/or additive as the measurement sample. For the purposes of the dissolution testing a prototype formulation of the measurement sample could also be used. In this case the control sample would be formulated in the same way.

Standard methods for determining the improved dissolution profile of a material *in vivo* are available in the art. A suitable method to determine an improved dissolution profile in a human may be after delivering the dose to measure the rate of active material absorption by measuring the plasma concentration of the sample compound over a period of time and comparing the results from the sample compound to a control. An observation that peak plasma concentration for the sample compound was achieved in less time than the control would indicate (assuming it is statistically significant) that the sample compound has improved bioavailability and an improved dissolution profile.

Preferably, the improved dissolution profile is observed at a relevant gastrointestinal pH, when it is observed *in vitro*. Preferably, the improved dissolution profile is observed at a pH which is favourable at indicating improvements in dissolution when comparing the measurement sample to the control compound.

Suitable methods for quantifying the concentration of a compound in an *in vitro* sample or an *in vivo* sample are widely available in the art. Suitable methods could include the use of spectroscopy or radioisotope labeling. In one preferred embodiment the method of quantification of dissolution is determined in a solution with a pH selected from the group consisting of: pH 1, pH 2, pH 3, pH 4, pH 5, pH 6, pH 7, pH 7.3, pH 7.4, pH 8, pH 9, pH 10, pH 11, pH 12, pH 13, pH 14 or a pH with 0.5 of a pH unit of any of this group.

Crystallization Profile

Methods for determining the crystallinity profile of the biologically active material are widely available in the art. Suitable methods may include X-ray diffraction, differential scanning calorimetry, raman or IR spectroscopy.

Amorphicity Profile

Methods for determining the amorphous content of the biologically active material are widely available in the art. Suitable methods may include X-ray diffraction, differential scanning calorimetry, raman or IR spectroscopy.

Grinding Matrix

As will be described subsequently, selection of an appropriate grinding matrix affords particular advantageous applications of the method of the present invention.

A highly advantageous application of the method of the invention is the use of a water-soluble grinding matrix in conjunction with a poorly water-soluble biologically active material. This

affords at least two advantages. The first being when the powder containing the biologically active material is placed into water – such as the ingestion of the powder as part of an oral medication - the matrix dissolves, releasing the particulate active material such that there is maximum surface area exposed to solution, thereby allowing a rapid dissolution of the active compound. The second key advantage is the ability, if required, to remove or partially remove the matrix prior to further processing or formulation.

Another advantageous application of the method of the invention is the use of a water-insoluble grinding matrix, particularly in the area of agricultural use, when a biologically active material such as a fungicide is commonly delivered as part of a dry powder or a suspension. The presence of a water insoluble matrix will afford benefits such as increased rain fastness.

Without wishing to be bound by theory, it is believed that the physical degradation (including but not limited to particle size reduction) of the millable grinding matrix affords the advantage of the invention, by acting as a more effective diluent than grinding matrix of a larger particle size. Again, as will be described subsequently, a highly advantageous aspect of the present invention is that certain grinding matrixes appropriate for use in the method of the invention are also appropriate for use in a medicament. The present invention encompasses methods for the production of a medicament incorporating both the biologically active material and the grinding matrix or in some cases the biologically active material and a portion of the grinding matrix, medicaments so produced, and methods of treatment of an animal, including man, using a therapeutically effective amount of said biologically active materials by way of said medicaments.

Analogously, as will be described subsequently, a highly advantageous aspect of the present invention is that certain grinding matrixes appropriate for use in the method of the invention are also appropriate for use in a carrier for an agricultural chemical, such as a pesticide, fungicide, or herbicide. The present invention encompasses methods for the production of an agricultural chemical composition incorporating both the biologically active material in particulate form and the grinding matrix, or in some cases the biologically active material, and a portion of the grinding matrix, and agricultural chemical compositions so produced. The medicament may include only the biologically active material together with the milled grinding matrix or, more preferably, the biologically active material and milled grinding matrix may be combined with one or more pharmaceutically acceptable carriers, as well as any desired excipients or other like agents commonly used in the preparation of medicaments.

Analogously, the agricultural chemical composition may include only the biologically active material together with the milled grinding matrix or, more preferably, the biologically active materials and milled grinding matrix may be combined with one or more carriers, as well as any desired excipients or other like agents commonly used in the preparation of agricultural chemical compositions.

In one particular form of the invention, the grinding matrix is both appropriate for use in a medicament and readily separable from the biologically active material by methods not dependent on particle size. Such grinding matrixes are described in the following detailed description of the invention. Such grinding matrixes are highly advantageous in that they afford significant flexibility in the extent to which the grinding matrix may be incorporated with the biologically active material into a medicament.

In a highly preferred form, the grinding matrix is harder than the biologically active material, and is thus capable of improving the dissolution profile of the active material under the dry milling conditions of the invention. Again without wishing to be bound by theory, under these circumstances it is believed that the millable grinding matrix affords the advantage of the present invention through a second route, with the smaller particles of grinding matrix produced under the dry milling conditions enabling greater interaction with the biologically active material. The quantity of the grinding matrix relative to the quantity of biologically active material, and the extent of physical degradation of the grinding matrix, is sufficient to improve the dissolution profile of the milled biologically active material. The grinding matrix is not generally selected to be chemically reactive with the biologically active material under the milling conditions of the invention, excepting for example, where the matrix is deliberately chosen to undergo a mechanico-chemical reaction. Such a reaction might be the conversion of a free base or acid to a salt or vice versa.

As stated above, the method of the present invention requires the grinding matrix to be milled with the biologically active material; that is, the grinding matrix will physically degrade under the dry milling conditions of the invention to facilitate the formation and retention of particulates of the biologically active material with improved dissolution profiles. The precise extent of degradation required will depend on certain properties of the grinding matrix and the biologically active material, the ratio of biologically active material to grinding matrix, and the particle size distribution of the particles comprising the biologically active material.

The physical properties of the grinding matrix necessary to achieve the requisite degradation are dependent on the precise milling conditions. For example, a harder grinding matrix may degrade to a sufficient extent provided [it is subjected to] more vigorous dry milling conditions. Physical properties of the grinding matrix relevant to the extent that the agent will degrade under dry milling conditions include hardness, friability, as measured by indicia such as hardness, fracture toughness and brittleness index.

A low hardness (typically a Mohs Hardness less than 7) of the biologically active material is desirable to ensure fracture of the particles during processing, so that composite microstructures develop during milling. Preferably, the hardness is less than 3 as determined using the Mohs Hardness scale.

Preferably, the grinding matrix is of low abrasivity. Low abrasivity is desirable to minimise contamination of the mixture of the biologically active material in the grinding matrix by the

milling bodies and/or the milling chamber of the media mill. An indirect indication of the abrasivity can be obtained by measuring the level of milling-based contaminants.

Preferably, the grinding matrix has a low tendency to agglomerate during dry milling. While it is difficult to objectively quantify the tendency to agglomerate during milling, it is possible to obtain a subjective measure by observing the level of "caking" of the grinding matrix on the milling bodies and the milling chamber of the media mill as dry milling progresses.

The grinding matrix may be an inorganic or organic substance.

In one embodiment, the grinding matrix is selected from the following, either as a single substance or a combination of two or more substances: Polyols (sugar alcohols) for example (but not limited to) mannitol, sorbitol, isomalt, xylitol, maltitol, lactitol, erythritol, arabitol, ribitol, monosaccharides for example (but not limited to) glucose, fructose, mannose, galactose, disaccharides and trisaccharides for example (but not limited to) anhydrous lactose, lactose monohydrate, sucrose, maltose, trehalose, polysaccharides for example (but not limited to) maltodextrins, dextrin, Inulin, dextrans, polydextrose, other carbohydrates for example (but not limited to) starch, wheat flour, corn flour, rice flour, rice starch, tapioca flour, tapioca starch, potato flour, potato starch, other flours and starches, , soy flour, soy meal or other soy products, cellulose, microcrystalline cellulose, microcrystalline cellulose based co blended excipients, chemically modified excipients such as pregelatinized (or partially) starch, modified celluloses such as HPMC, CMC, HPC, enteric polymer coatings such as hypromellose phthalate, cellulose acetate phthalate (Aquacoat®), polyvinyl acetate phthalate (Sureteric®), hypromellose acetate succinate (AQOAT®), and polymethacrylates (Eudragit® and Acryl-EZE®), Milk products for example (but not limited to) milk powder, skim milk powders, other milk solids and derivatives, other functional Excipients, organic acids for example (but not limited to) citric acid, tartaric acid, malic acid, maleic acid fumaric acid , ascorbic acid, succinic acid, the conjugate salt of organic acids for example (but not limited to) sodium citrate, sodium tartrate, sodium malate, sodium ascorbate, potassium citrate, potassium tartrate, potassium malate, potassium ascorbate, inorganics such as sodium carbonate, potassium carbonate, magnesium carbonate, sodium bicarbonate, potassium bicarbonate and calcium carbonate. dibasic calcium phosphate, tribasic calcium phosphate, sodium sulfate, sodium chloride, sodium metabisulfite, sodium thiosulfate, ammonium chloride, Glauber's salt, ammonium carbonate, sodium bisulfate, magnesium sulfate, potash alum, potassium chloride, sodium hydrogen sulfate, sodium hydroxide, crystalline hydroxides, hydrogen carbonates, hydrogen carbonates of pharmaceutical acceptable alkali metals, such as but not limited by, sodium, potassium, lithium, calcium, and barium, ammonium salts (or salts of volatile amines), for example (but not limited to) ammonium chloride, methylamine hydrochloride, ammonium bromide, other inorganics for example (but not limited to), thermal silica, chalk, mica, silica, alumina, titanium dioxide, talc, kaolin, bentonite, hectorite, magnesium trisilicate, other clay or clay derivatives or aluminium silicates, a surfactant for example (but not limited to) sodium

lauryl sulfate, sodium stearyl sulfate, sodium cetyl sulfate, sodium cetostearyl sulfate, sodium docusate, sodium deoxycholate, N-lauroylsarcosine sodium salt, glyceryl monostearate, glycerol distearate glyceryl palmitostearate, glyceryl behenate, glyceryl caprylate, glyceryl oleate, benzalkonium chloride, CTAB, CTAC, Cetrimide, cetylpyridinium chloride, cetylpyridinium bromide, benzethonium chloride, PEG 40 stearate, PEG 100 stearate, poloxamer 188, poloxamer 338, poloxamer 407 polyoxyl 2 stearyl ether, polyoxyl 100 stearyl ether, polyoxyl 20 stearyl ether, polyoxyl 10 stearyl ether, polyoxyl 20 cetyl ether, polysorbate 20, polysorbate 40, polysorbate 60, polysorbate 61, polysorbate 65, polysorbate 80, polyoxyl 35 castor oil, polyoxyl 40 castor oil, polyoxyl 60 castor oil, polyoxyl 100 castor oil, polyoxyl 200 castor oil, polyoxyl 40 hydrogenated castor oil, polyoxyl 60 hydrogenated castor oil, polyoxyl 100 hydrogenated castor oil, polyoxyl 200 hydrogenated castor oil, cetostearyl alcohol, macrogel 15 hydroxystearate, sorbitan monopalmitate, sorbitan monostearate, sorbitan trioleate, Sucrose Palmitate, Sucrose Stearate, Sucrose Distearate, Sucrose laurate, Glycocholic acid, sodium Glycholate, Cholic Acid, Sodium Cholate, Sodium Deoxycholate, Deoxycholic acid, Sodium taurocholate, taurocholic acid, Sodium taurodeoxycholate, taurodeoxycholic acid, soy lecithin, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, phosphatidylinositol, PEG4000, PEG6000, PEG8000, PEG10000, PEG20000, alkyl naphthalene sulfonate condensate/Lignosulfonate blend, Calcium Dodecylbenzene Sulfonate, Sodium Dodecylbenzene Sulfonate, Diisopropyl naphthaenesulphonate, erythritol distearate, Naphthalene Sulfonate Formaldehyde Condensate, nonylphenol ethoxylate (poe-30), Tristyrylphenol Ethoxylate, Polyoxyethylene (15) tallowalkylamines, sodium alkyl naphthalene sulfonate, sodium alkyl naphthalene sulfonate condensate, sodium alkylbenzene sulfonate, sodium isopropyl naphthalene sulfonate, Sodium Methyl Naphthalene Formaldehyde Sulfonate, sodium n-butyl naphthalene sulfonate, tridecyl alcohol ethoxylate (poe-18), Triethanolamine isodecanol phosphate ester, Triethanolamine tristyrylphosphate ester, Tristyrylphenol Ethoxylate Sulfate, Bis(2-hydroxyethyl)tallowalkylamines.

In a preferred embodiment, the grinding matrix is a matrix that is considered GRAS (generally regarded as safe) by persons skilled in the pharmaceutical arts.

In another preferred aspect a combination of two or more suitable matrices, such as those listed above, can be used as the grinding matrix to provide improved properties such as the reduction of caking, and greater improvement of particle size reduction. Combination matrices may also be advantageous when the matrices have different solubility's allowing the removal or partial removal of one matrix, while leaving the other or part of the other to provide encapsulation or partial encapsulation of the biologically active material.

Another highly preferred aspect of the method is the inclusion of a suitable milling aid in the matrix to improve milling performance. Improvements to milling performance would be things such as, but not limited to, a reduction in caking or higher recovery of powder from the mill.

Examples of suitable milling aids include surfactants, polymers and inorganics such as silica (including colloidal silica), aluminium silicates and clays.

There are a wide range of surfactants that will make suitable milling aids. The highly preferred form is where the surfactant is a solid, or can be manufactured into a solid. Preferably, the surfactant is selected from the group consisting of: polyoxyethylene alkyl ethers, polyoxyethylene stearates, polyethylene glycols (PEG), poloxamers, poloxamines, sarcosine based surfactants, polysorbates, aliphatic alcohols, alkyl and aryl sulfates, alkyl and aryl polyether sulfonates and other sulfate surfactants, trimethyl ammonium based surfactants, lecithin and other phospholipids, bile salts, polyoxyethylene castor oil derivatives, polyoxyethylene sorbitan fatty acid esters, Sorbitan fatty acid esters, Sucrose fatty acid esters, alkyl glucopyranosides, alkyl maltopyranosides, glycerol fatty acid esters, Alkyl Benzene Sulphonic Acids, Alkyl Ether Carboxylic Acids, Alkyl and aryl Phosphate esters, Alkyl and aryl Sulphate esters, Alkyl and aryl Sulphonic acids, Alkyl Phenol Phosphates esters, Alkyl Phenol Sulphates esters, Alkyl and Aryl Phosphates, Alkyl Polysaccharides, Alkylamine Ethoxylates, Alkyl-Naphthalene Sulphonates formaldehyde condensates, Sulfosuccinates, lignosulfonates, Ceto-Oleyl Alcohol Ethoxylates, Condensed Naphthalene Sulphonates, Dialkyl and Alkyl Naphthalene Sulphonates, Di-alkyl Sulphosuccinates, Ethoxylated nonylphenols, Ethylene Glycol Esters, Fatty Alcohol Alkoxylates, Hydrogenated tallowalkylamines, Mono-alkyl Sulphosuccinamates, Nonyl Phenol Ethoxylates, Sodium Oleyl N-methyl Taurate, Tallowalkylamines, linear and branched dodecylbenzene sulfonic acids.

Preferably, the surfactant is selected from the group consisting of: sodium lauryl sulfate, sodium stearyl sulfate, sodium cetyl sulfate, sodium cetostearyl sulfate, sodium docusate, sodium deoxycholate, N-lauroylsarcosine sodium salt, glyceryl monostearate, glycerol distearate, glyceryl palmitostearate, glyceryl behenate, glyceryl caprylate, glyceryl oleate, benzalkonium chloride, CTAB, CTAC, Cetrimide, cetylpyridinium chloride, cetylpyridinium bromide, benzethonium chloride, PEG 40 stearate, PEG 100 stearate, poloxamer 188, poloxamer 338, poloxamer 407 polyoxyl 2 stearyl ether, polyoxyl 100 stearyl ether, polyoxyl 20 stearyl ether, polyoxyl 10 stearyl ether, polyoxyl 20 cetyl ether, polysorbate 20, polysorbate 40, polysorbate 60, polysorbate 61, polysorbate 65, polysorbate 80, polyoxyl 35 castor oil, polyoxyl 40 castor oil, polyoxyl 60 castor oil, polyoxyl 100 castor oil, polyoxyl 200 castor oil, polyoxyl 40 hydrogenated castor oil, polyoxyl 60 hydrogenated castor oil, polyoxyl 100 hydrogenated castor oil, polyoxyl 200 hydrogenated castor oil, cetostearyl alcohol, macrogel 15 hydroxystearate, sorbitan monopalmitate, sorbitan monostearate, sorbitan trioleate, Sucrose Palmitate, Sucrose Stearate, Sucrose Distearate, Sucrose laurate, Glycocholic acid, sodium Glycholate, Cholic Acid, Sodium Cholate, Sodium Deoxycholate, Deoxycholic acid, Sodium taurocholate, taurocholic acid, Sodium taurodeoxycholate, taurodeoxycholic acid, soy lecithin, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, phosphatidylinositol, PEG4000, PEG6000, PEG8000, PEG10000, PEG20000, alkyl naphthalene sulfonate

condensate/Lignosulfonate blend, Calcium Dodecylbenzene Sulfonate, Sodium Dodecylbenzene Sulfonate, Diisopropyl naphthaenesulphonate, erythritol distearate, Naphthalene Sulfonate Formaldehyde Condensate, nonylphenol ethoxylate (poe-30), Tristyrylphenol Ethoxylate, Polyoxyethylene (15) tallowalkylamines, sodium alkyl naphthalene sulfonate, sodium alkyl naphthalene sulfonate condensate, sodium alkylbenzene sulfonate, sodium isopropyl naphthalene sulfonate, Sodium Methyl Naphthalene Formaldehyde Sulfonate, sodium n-butyl naphthalene sulfonate, tridecyl alcohol ethoxylate (poe-18), Triethanolamine isodecanol phosphate ester, Triethanolamine tristyrylphosphate ester, Tristyrylphenol Ethoxylate Sulfate, Bis(2-hydroxyethyl)tallowalkylamines.

Preferably the polymer is selected from the list of: polyvinylpyrrolidones (PVP), polyvinylalcohol, Acrylic acid based polymers and copolymers of acrylic acid

Preferably, the milling aid has a concentration selected from the group consisting of: 0.1-10% w/w, 0.1-5% w/w, 0.1-2.5% w/w, of 0.1-2% w/w, 0.1-1%, 0.5-5% w/w, 0.5-3% w/w, 0.5-2% w/w, 0.5-1.5%, 0.5-1% w/w, of 0.75-1.25% w/w, 0.75-1% and 1% w/w.

Milling bodies

In the method of the present invention, the milling bodies are preferably chemically inert and rigid. The term "chemically-inert", as used herein, means that the milling bodies do not react chemically with the biologically active material or the grinding matrix.

As described above, the milling bodies are essentially resistant to fracture and erosion in the milling process.

The milling bodies are desirably provided in the form of bodies which may have any of a variety of smooth, regular shapes, flat or curved surfaces, and lacking sharp or raised edges. For example, suitable milling bodies can be in the form of bodies having ellipsoidal, ovoid, spherical or right cylindrical shapes. Preferably, the milling bodies are provided in the form of one or more of beads, balls, spheres, rods, right cylinders, drums or radius-end right cylinders (i.e., right cylinders having hemispherical bases with the same radius as the cylinder).

Depending on the nature of the biologically active material and the grinding matrix, the milling media bodies desirably have an effective mean particle diameter (i.e. "particle size") between about 0.1 and 30 mm, more preferably between about 1 and about 15 mm, still more preferably between about 3 and 10 mm.

The milling bodies may comprise various substances such as ceramic, glass, metal or polymeric compositions, in a particulate form. Suitable metal milling bodies are typically spherical and generally have good hardness (i.e. RHC 60-70), roundness, high wear resistance, and narrow size distribution and can include, for example, balls fabricated from type 52100 chrome steel, type 316 or 440C stainless steel or type 1065 high carbon steel.

Preferred ceramics, for example, can be selected from a wide array of ceramics desirably having sufficient hardness and resistance to fracture to enable them to avoid being chipped or crushed during milling and also having sufficiently high density. Suitable densities for milling media can range from about 1 to 15 g/cm³, preferably from about 1 to 8 g/cm³. Preferred ceramics can be selected from steatite, aluminum oxide, zirconium oxide, zirconia-silica, yttria-stabilized zirconium oxide, magnesia-stabilized zirconium oxide, silicon nitride, silicon carbide, cobalt-stabilized tungsten carbide, and the like, as well as mixtures thereof.

Preferred glass milling media are spherical (e.g. beads), have a narrow size distribution, are durable, and include, for example, lead-free soda lime glass and borosilicate glass. Polymeric milling media are preferably substantially spherical and can be selected from a wide array of polymeric resins having sufficient hardness and friability to enable them to avoid being chipped or crushed during milling, abrasion-resistance to minimize attrition resulting in contamination of the product, and freedom from impurities such as metals, solvents, and residual monomers.

Preferred polymeric resins, for example, can be selected from crosslinked polystyrenes, such as polystyrene crosslinked with divinylbenzene, styrene copolymers, polyacrylates such as polymethylmethacrylate, polycarbonates, polyacetals, vinyl chloride polymers and copolymers, polyurethanes, polyamides, high density polyethylenes, polypropylenes, and the like. The use of polymeric milling media to grind materials down to a very small particle size (as opposed to mechanochemical synthesis) is disclosed, for example, in U.S. patents 5,478,705 and 5,500,331. Polymeric resins typically have densities ranging from about 0.8 to 3.0 g/cm³. Higher density polymeric resins are preferred. Alternatively, the milling media can be composite particles comprising dense core particles having a polymeric resin adhered thereon. Core particles can be selected from substances known to be useful as milling media, for example, glass, alumina, zirconia silica, zirconium oxide, stainless steel, and the like. Preferred core substances have densities greater than about 2.5 g/cm³.

In one embodiment of the invention, the milling media are formed from a ferromagnetic substance, thereby facilitating removal of contaminants arising from wear of the milling media by the use of magnetic separation techniques.

Each type of milling body has its own advantages. For example, metals have the highest specific gravities, which increase grinding efficiency due to increased impact energy. Metal costs range from low to high, but metal contamination of final product can be an issue. Glasses are advantageous from the standpoint of low cost and the availability of small bead sizes as low as 0.004 mm. However, the specific gravity of glasses is lower than other media and significantly more milling time is required. Finally, ceramics are advantageous from the standpoint of low wear and contamination, ease of cleaning, and high hardness.

Dry Milling

In the dry milling process of the present invention, the biologically active material and grinding matrix, in the form of crystals, powders, or the like, are combined in suitable proportions with the plurality of milling bodies in a milling chamber that is mechanically agitated (i.e. with or without stirring) for a predetermined period of time at a predetermined intensity of agitation. Typically, a milling apparatus is used to impart motion to the milling bodies by the external application of agitation, whereby various translational, rotational or inversion motions or combinations thereof are applied to the milling chamber and its contents, or by the internal application of agitation through a rotating shaft terminating in a blade, propeller, impeller or paddle or by a combination of both actions.

During milling, motion imparted to the milling bodies can result in application of shearing forces as well as multiple impacts or collisions having significant intensity between milling bodies and particles of the biologically active material and grinding matrix. The nature and intensity of the forces applied by the milling bodies to the biologically active material and the grinding matrix is influenced by a wide variety of processing parameters including: the type of milling apparatus; the intensity of the forces generated, the kinematic aspects of the process; the size, density, shape, and composition of the milling bodies; the weight ratio of the biologically active material and grinding matrix mixture to the milling bodies; the duration of milling; the physical properties of both the biologically active material and the grinding matrix; the atmosphere present during activation; and others.

Advantageously, the media mill is capable of repeatedly or continuously applying mechanical compressive forces and shear stress to the biologically active material and the grinding matrix. Suitable media mills include but are not limited to the following: high-energy ball, sand, bead or pearl mills, basket mill, planetary mill, vibratory action ball mill, multi-axial shaker/mixer, stirred ball mill, horizontal small media mill, multi-ring pulverizing mill, and the like, including small milling media. The milling apparatus also can contain one or more rotating shafts.

In a preferred form of the invention, the dry milling is performed in a ball mill. Throughout the remainder of the specification reference will be made to dry milling being carried out by way of a ball mill. Examples of this type of mill are attritor mills, nutating mills, tower mills, planetary mills, vibratory mills and gravity-dependent-type ball mills. It will be appreciated that dry milling in accordance with the method of the invention may also be achieved by any suitable means other than ball milling. For example, dry milling may also be achieved using jet mills, rod mills, roller mills or crusher mills.

Biologically active material

The biologically active material includes active compounds, including compounds for veterinary and human use such as but not limited to, pharmaceutical actives, nutraceuticals, cosmeceuticals, cosmetics, complementary medicines, natural products, vitamins, nutrients,

biologics, amino acids, proteins, peptides, nucleotides, nucleic acids. and agricultural compounds such as pesticides, herbicides and fungicides, germinating agents and the like.

Other biologically active materials include, but are not limited to, foods, seeds, cocoa and cocoa solids, coffee, herbs, spices, other plant materials, minerals, animal products, shells and other skeletal material.

In a preferred form of the invention, the biologically active material is an organic compound. In a highly preferred form of the invention, the biologically active material is an organic, therapeutically active compound for veterinary or human use.

In a preferred form of the invention, the biologically active material is an inorganic compound.

In a highly preferred form of the invention, the biologically active material is sulphur, copper hydroxide, an organometallic complex or copper oxychloride..

The biologically active material is ordinarily a material for which one of skill in the art desires improved dissolution properties. The biologically active material may be a conventional active agent or drug, although the process of the invention may be employed on formulations or agents that already have reduced particle size compared to their conventional form.

Biologically active materials suitable for use in the invention include actives, biologics, amino acids, proteins, peptides, nucleotides, nucleic acids, and analogs, homologs and first order derivatives thereof. The biologically active material can be selected from a variety of known classes of drugs, including, but not limited to: anti-obesity drugs, central nervous system stimulants, carotenoids, corticosteroids, elastase inhibitors, anti-fungals, oncology therapies, anti-emetics, analgesics, cardiovascular agents, anti-inflammatory agents, such as NSAIDs and COX-2 inhibitors, anthelmintics, anti-arrhythmic agents, antibiotics (including penicillins), anticoagulants, antidepressants, antidiabetic agents, antiepileptics, antihistamines, antihypertensive agents, antimuscarinic agents, antimycobacterial agents, antineoplastic agents, immunosuppressants, antithyroid agents, antiviral agents, anxiolytics, sedatives (hypnotics and neuroleptics), astringents, alpha-adrenergic receptor blocking agents, beta-adrenoceptor blocking agents, blood products and substitutes, cardiac inotropic agents, contrast media, cough suppressants (expectorants and mucolytics), diagnostic agents, diagnostic imaging agents, diuretics, dopaminergics (anti-Parkinsonian agents), haemostatics, immunological agents, lipid regulating agents, muscle relaxants, parasympathomimetics, parathyroid calcitonin and biphosphonates, prostaglandins, radio-pharmaceuticals, sex hormones (including steroids), anti-allergic agents, stimulants and anoretics, sympathomimetics, thyroid agents, vasodilators, and xanthines.

A description of these classes of active agents and a listing of species within each class can be found in Martindale's The Extra Pharmacopoeia, 31st Edition (The Pharmaceutical Press, London, 1996), specifically incorporated by reference. Another source of active agents is the Physicians Desk Reference (60th Ed., pub. 2005), familiar to those of skill in the art. The active agents are commercially available and/or can be prepared by techniques known in the art.

An exhaustive list of drugs for which the methods of the invention are suitable would be burdensomely long for this specification; however, reference to the general pharmacopoeia listed above would allow one of skill in the art to select virtually any drug to which the method of the invention may be applied.

In addition it is also expected that new chemical entities (NCE) and other actives for which the methods of the invention are suitable will be created or become commercially available in the future.

Notwithstanding the general applicability of the method of the invention, more specific examples of biologically active materials include, but are not limited to: haloperidol (dopamine antagonist), DL isoproterenol hydrochloride (β -adrenergic agonist), terfenadine (H1-antagonist), propranolol hydrochloride (β -adrenergic antagonist), desipramine hydrochloride (antidepressant), sildenafil citrate, tadalafil and vardenafil. Minor analgesics (cyclooxygenase inhibitors), fenamic acids, Piroxicam, Cox-2 inhibitors, and Naproxen, and others, may all benefit from being prepared.

As discussed in the context of the background to the invention, biologically active materials that are poorly water soluble at gastrointestinal pH will particularly benefit from being prepared, and the method of the present invention is particularly advantageously applied to materials that are poorly water soluble at gastrointestinal pH.

Such materials include, but are not limited to: albendazole, albendazole sulfoxide, alfaxalone, acetyl digoxin, acyclovir analogs, alprostadil, aminofostin, anipamil, antithrombin III, atenolol, azidothymidine, beclobrate, beclomethasone, belomycin, benzocaine and derivatives, beta carotene, beta endorphin, beta interferon, bezafibrate, binovum, biperiden, bromazepam, bromocryptine, bucindolol, buflomedil, bupivacaine, busulfan, cadralazine, camptothecin, canthaxanthin, captopril, carbamazepine, carboprost, cefalexin, cefalotin, cefamandole, cefazedone, ceftiofur, cefuroxime, cefixime, cefoperazone, cefotaxime, cefoxitin, cefsulodin, ceftiofur, chlorambucil, chromoglycinic acid, ciclonate, ciglitazone, clonidine, cortexolone, corticosterone, cortisol, cortisone, cyclophosphamide, cyclosporin A and other cyclosporins, cytarabine, desocryptin, desogestrel, dexamethasone esters such as the acetate, dezocine, diazepam, diclofenac, dideoxyadenosine, dideoxyinosine, digitoxin, digoxin, dihydroergotamine, dihydroergotoxin, diltiazem, dopamine antagonists, doxorubicin, econazole, endralazine, enkephalin, enalapril, epoprostenol, estradiol, estramustine, etofibrate, etoposide, factor ix, factor viii, felbamate, fenbendazole, fenofibrate, fexofenadine, flunarizin, flurbiprofen, 5-fluorouracil, flurazepam, fosfomycin, fosmidomycin, furosemide, gallopamil, gamma interferon, gentamicin, gepirone, glimepiride, glipizide, griseofulvin, haptoglobulin, hepatitis B vaccine, hydralazine, hydrochlorothiazide, hydrocortisone, ibuprofen, ibuprofen, indinavir, indomethacin, iodinated aromatic x-ray contrast agents such as iodamide, ipratropium bromide, ketoconazole, ketoprofen, ketotifen, ketotifen fumarate, K-strophanthin, labetalol, lactobacillus vaccine, lidocaine, lidoflazine, lisuride, lisuride hydrogen maleate, lorazepam, lovastatin,

mefenamic acid, melphalan, memantin, mesulergin, metergoline, methotrexate, methyl digoxin, methylprednisolone, metronidazole, metisoprenol, metipranolol, metkephamide, metolazone, metoprolol, metoprolol tartrate, miconazole, miconazole nitrate, minoxidil, misonidazol, molsidomin, nadolol, nafiverine, nafazatrom, naproxen, natural insulins, nesapidil, nicardipine, nicorandil, nifedipine, niludipin, nimodipine, nitrazepam, nitrendipine, nitrocamptothecin, 9-nitrocamptothecin, olanzapine, oxazepam, oxprenolol, oxytetracycline, penicillins such as penicillin G benethamine, penecillin O, phenylbutazone, picotamide, pindolol, piposulfan, piretanide, piribedil, piroxicam, pirprofen, plasminogenici activator, prednisolone, prednisone, pregnenolone, procabacin, procaterol, progesterone, proinsulin, propafenone, propanolol, propentofyllin, propofol, propranolol, raloxifene, rifapentin, simvastatin, semi-synthetic insulins, sobrerol, somastotine and its derivatives, somatropin, stilamine, sulfinalol hydrochloride, sulfinpyrazone, suloctidil, suprofen, sulproston, synthetic insulins, talinolol, taxol, taxotere, testosterone, testosterone propionate, testosterone undecanoate, tetracane HI, tiaramide HCl, tolmetin, tranilast, triquilar, tromantadine HCl, urokinase, valium, verapamil, vidarabine, vidarabine phosphate sodium salt, vinblastine, vinburin, vincamine, vincristine, vindesine, vinpocetine, vitamin A, vitamin E succinate, and x-ray contrast agents. Drugs can be neutral species or basic or acidic as well as salts of an acid or base. Specifically the chemical makeup and the functional groups, including an acid or base group, are generally not the determinant factor, excepting a possible chemical reaction with a specific matrix, for the successful creation of a biologically active substance with improved dissolution. This invention is not limited to any drug specific class, application type, chemical type or function grouping. Rather the suitability of a biologically active material for use in this invention is primarily determined by the mechanical properties of the material. In addition, some biologically active materials may have the benefit of absorption through the skin if presented in a particle formulation. Such biologically active materials include, but are not limited to, Voltaren (diclofenac), rofecoxib, and ibuprofen.

Conveniently, the biologically active material is capable of withstanding temperatures that are typical in uncooled dry milling, which may exceed 80 °C. Therefore, materials with a melting point about 80 °C or greater are highly suitable. For biologically active materials with lower melting points, the media mill may be cooled, thereby allowing materials with significantly lower melting temperatures to be processed according to the method of the invention. For instance, a simple water-cooled mill will keep temperatures below 50 °C, or chilled water could be used to further lower the milling temperature. Those skilled in the art will understand that a high energy ball mill could be designed to run at any temperature between say -30 to 200 °C. For some biologically active materials it may be advantageous to control the milling temperature to temperatures significantly below the melting points of the biologically active materials.

The biologically active material is obtained in a conventional form commercially and/or prepared by techniques known in the art.

It is preferred, but not essential, that the particle size of the biologically active material be less than about 1000 μm , as determined by sieve analysis. If the coarse particle size of the biologically active material is greater than about 1000 μm , then it is preferred that the particles of the biologically active material substrate be reduced in size to less than 1000 μm using another standard milling method.

Processed biologically active material

Preferably, the biologically active materials, which have been subject to the methods of the invention, comprises particles of biologically active material of an average particle size diameter equal or greater than 1 μm , determined on a particle number basis.

Preferably, the biologically active materials, which have been subject to the methods of the invention, comprises particles of biologically active material of a median particle size diameter equal or greater than 1 μm , determined on a particle volume basis.

These sizes refer to particles either fully dispersed or partially agglomerated.

Agglomerates of biologically active material after processing

Agglomerates comprising particles of biologically active material, said particles having a particle size within the ranges specified above, should be understood to fall within the scope of the present invention. Agglomerates comprising particles of biologically active material, said agglomerates having a total agglomerate size within the ranges specified above, should be understood to fall within the scope of the present invention.

Agglomerates comprising particles of biologically active material, should be understood to fall within the scope of the present invention if at the time of use, or further processing, the particle size of the agglomerate is within the ranges specified above.

Processing Time

Preferably, the biologically active material and the grinding matrix are dry milled for the shortest time necessary to form the mixture of the biologically active material in the grinding matrix such that the active material has improved dissolution to minimise any possible contamination from the media mill and/or the plurality of milling bodies. This time varies greatly, depending on the biologically active material and the grinding matrix, and may range from as short as 1 minute to several hours. Dry milling times in excess of 2 hours may lead to degradation of the biologically active material and an increased level of undesirable contaminants.

Suitable rates of agitation and total milling times are adjusted for the type and size of milling apparatus as well as the milling media, the weight ratio of the biologically active material and grinding matrix mixture to the plurality of milling bodies, the chemical and physical properties of the biologically active material and grinding matrix, and other parameters that may be optimized empirically.

Inclusion of the grinding matrix with the biologically active material and separation of the grinding matrix from the biologically active material

In a preferred aspect, the grinding matrix is not separated from the biologically active material but is maintained with the biologically active material in the final product. Preferably the grinding matrix is considered to be Generally Regarded as Safe (GRAS) for pharmaceutical products.

In an alternative aspect, the grinding matrix is separated from the biologically active material. In one aspect, where the grinding matrix is not fully milled, the unmilled grinding matrix is separated from the biologically active material. In a further aspect, at least a portion of the milled grinding matrix is separated from the biologically active material.

Any portion of the grinding matrix may be removed, including but not limited to 10%, 25%, 50%, 75%, or substantially all of the grinding matrix.

In some embodiments of the invention, a significant portion of the milled grinding matrix may comprise particles of a size similar to and/or smaller than the particles comprising the biologically active material. Where the portion of the milled grinding matrix to be separated from the particles comprising the biologically active material comprises particles of a size similar to and/or smaller than the particles comprising the biologically active material, separation techniques based on size distribution are inapplicable.

In these circumstances, the method of the present invention may involve separation of at least a portion of the milled grinding matrix from the biologically active material by techniques including but not limited to electrostatic separation, magnetic separation, centrifugation (density separation), hydrodynamic separation, froth flotation.

Advantageously, the step of removing at least a portion of the milled grinding matrix from the biologically active material may be performed through means such as selective dissolution, washing, or sublimation.

An advantageous aspect of the invention would be the use of grinding matrix that has two or more components where at least one component is water soluble and at least one component has low solubility in water. In this case washing can be used to remove the matrix component soluble in water leaving the biologically active material encapsulated in the remaining matrix components. In a highly advantageous aspect of the invention the matrix with low solubility is a functional excipient.

A highly advantageous aspect of the present invention is that certain grinding matrixes appropriate for use in the method of the invention (in that they physically degrade to the desired extent under dry milling conditions) are also pharmaceutically acceptable and thus appropriate for use in a medicament. Where the method of the present invention does not involve complete separation of the grinding matrix from the biologically active material, the present invention encompasses methods for the production of a medicament incorporating both the biologically active material and at least a portion of the milled grinding matrix, medicaments so produced

and methods of treatment of an animal, including man, using a therapeutically effective amount of said biologically active materials by way of said medicaments.

The medicament may include only the biologically active material and the grinding matrix or, more preferably, the biologically active materials and grinding matrix may be combined with one or more pharmaceutically acceptable carriers, as well as any desired excipients or other like agents commonly used in the preparation of medicaments.

Analogously, a highly advantageous aspect of the present invention is that certain grinding matrixes appropriate for use in the method of the invention (in that they physically degrade to a desirable extent under dry milling conditions) are also appropriate for use in an agricultural chemical composition. Where the method of the present invention does not involve complete separation of the grinding matrix from the biologically active material, the present invention encompasses methods for the production of a agricultural chemical composition incorporating both the biologically active material and at least a portion of the milled grinding matrix, agricultural chemical composition so produced and methods of use of such compositions.

The agricultural chemical composition may include only the biologically active material and the grinding matrix or, more preferably, the biologically active materials and grinding matrix may be combined with one or more acceptable carriers, as well as any desired excipients or other like agents commonly used in the preparation of agricultural chemical compositions.

In one particular form of the invention, the grinding matrix is both appropriate for use in a medicament and readily separable from the biologically active material by methods not dependent on particle size. Such grinding matrixes are described in the following detailed description of the invention. Such grinding matrixes are highly advantageous in that they afford significant flexibility in the extent to which the grinding matrix may be incorporated with the biologically active material into a medicament.

The mixture of biologically active material and grinding matrix may then be separated from the milling bodies and removed from the mill.

In one embodiment, the grinding matrix is separated from the mixture of biologically active material and grinding matrix. Where the grinding matrix is not fully milled, the unmilled grinding matrix is separated from the biologically active material. In a further aspect, at least a portion of the milled grinding matrix is separated from the biologically active material.

The milling bodies are essentially resistant to fracture and erosion in the dry milling process.

The quantity of the grinding matrix relative to the quantity of biologically active material, and the extent of milling of the grinding matrix, is sufficient to provide improved dissolution of the biologically active material.

The grinding matrix is neither chemically nor mechanically reactive with the pharmaceutical material under the dry milling conditions of the method of the invention except, for example, where the matrix is deliberately chosen to undergo a mechanico-chemical reaction. Such a reaction might be the conversion of a free base or acid to a salt or vice versa.

Preferably, the medicament is a solid dosage form, however, other dosage forms may be prepared by those of ordinary skill in the art.

In one form, after the step of separating said mixture of biologically active material and grinding matrix from the plurality of milling bodies, and before the step of using said mixture of biologically active material and grinding matrix in the manufacture of a medicament, the method may comprise the step of:

removing a portion of the grinding matrix from said mixture of biologically active material and grinding matrix to provide a mixture enriched in the biologically active material; and the step of using said mixture of biologically active material and grinding matrix in the manufacture of a medicament, more particularly comprises the step of using the mixture of biologically active material and grinding matrix enriched in the biologically active material form in the manufacture of a medicament.

The present invention includes medicaments manufactured by said methods, and methods for the treatment of an animal, including man, by the administration of a therapeutically effective amount of the biologically active materials by way of said medicaments.

In another embodiment of the invention, a facilitating agent or a combination of facilitating agents is also comprised in the mixture to be milled. Such facilitating agents appropriate for use in the invention include diluents, surfactants, polymers, binding agents, filling agents, lubricating agents, sweeteners, flavouring agents, preservatives, buffers, wetting agents, disintegrants, effervescent agents and agents that may form part of a medicament, including a solid dosage form, or other excipients required for other specific drug delivery, such as the agents and media listed below under the heading *Medicinal and Pharmaceutical Compositions*, or any combination thereof.

Biologically active materials and compositions

The present invention encompasses pharmaceutically acceptable materials produced according to the methods of the present invention, compositions including such materials, including compositions comprising such materials together with the grinding matrix, with at least a portion of the grinding matrix or separated from the grinding matrix.

The pharmaceutically acceptable materials within the compositions of the invention are present at a concentration of between about 0.1% and about 99.0% by weight. Preferably, the concentration of pharmaceutically acceptable materials within the compositions will be about 5% to about 80% by weight, while concentrations of 10% to about 50% by weight are highly preferred. Desirably, the concentration will be in the range of about 10 to 15% by weight, 15 to 20% by weight, 20 to 25% by weight, 25 to 30% by weight, 30 to 35% by weight, 35 to 40% by weight, 40 to 45% by weight, 45 to 50% by weight, 50 to 55% by weight, 55 to 60% by weight, 60 to 65% by weight, 65 to 70% by weight, 70 to 75% by weight or 75 to 80% by weight for the composition prior to any later removal (if desired) of any portion of the grinding matrix. Where

part or all of the grinding matrix has been removed, the relative concentration of pharmaceutically acceptable materials in the composition may be considerably higher depending on the amount of the grinding matrix that is removed. For example, if all of the grinding matrix is removed the concentration of particles in the preparation may approach 100% by weight (subject to the presence of facilitating agents).

Compositions produced according to the present invention are not limited to the inclusion of a single species of pharmaceutically acceptable materials. More than one species of pharmaceutically acceptable materials may therefore be present in the composition. Where more than one species of pharmaceutically acceptable materials is present, the composition so formed may either be prepared in a dry milling step, or the pharmaceutically acceptable materials may be prepared separately and then combined to form a single composition.

Medicaments

The medicaments of the present invention may include the pharmaceutically acceptable material, optionally together with the grinding matrix or at least a portion of the grinding matrix, combined with one or more pharmaceutically acceptable carriers, as well as other agents commonly used in the preparation of pharmaceutically acceptable compositions.

As used herein "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like that are physiologically compatible. Preferably, the carrier is suitable for parenteral administration, intravenous, intraperitoneal, intramuscular, sublingual, pulmonary, transdermal or oral administration. Pharmaceutically acceptable carriers include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. The use of such media and agents for the manufacture of medicaments is well known in the art. Except insofar as any conventional media or agent is incompatible with the pharmaceutically acceptable material, use thereof in the manufacture of a pharmaceutical composition according to the invention is contemplated.

Pharmaceutical acceptable carriers according to the invention may include one or more of the following examples:

- (1) surfactants and polymers, including, but not limited to polyethylene glycol (PEG), polyvinylpyrrolidone (PVP), polyvinylalcohol, crospovidone, polyvinylpyrrolidone-polyvinylacetate copolymer, cellulose derivatives, hydroxypropylmethyl cellulose, hydroxypropyl cellulose, carboxymethylethyl cellulose, hydroxypropylmethyl cellulose phthalate, polyacrylates and polymethacrylates, urea, sugars, polyols, and their polymers, emulsifiers, sugar gum, starch, organic acids and their salts, vinyl pyrrolidone and vinyl acetate; and or
- (2) binding agents such as various celluloses and cross-linked polyvinylpyrrolidone, microcrystalline cellulose; and or

- (3) filling agents such as lactose monohydrate, lactose anhydrous, microcrystalline cellulose and various starches; and or
- (4) lubricating agents such as agents that act on the flowability of the powder to be compressed, including colloidal silicon dioxide, talc, stearic acid, magnesium stearate, calcium stearate, silica gel; and or
- (5) sweeteners such as any natural or artificial sweetener including sucrose, xylitol, sodium saccharin, cyclamate, aspartame, and acsulfame K; and or
- (6) flavouring agents; and or
- (7) preservatives such as potassium sorbate, methylparaben, propylparaben, benzoic acid and its salts, other esters of parahydroxybenzoic acid such as butylparaben, alcohols such as ethyl or benzyl alcohol, phenolic chemicals such as phenol, or quarternary compounds such as benzalkonium chloride; and or
- (8) buffers; and or
- (9) Diluents such as pharmaceutically acceptable inert fillers, such as microcrystalline cellulose, lactose, dibasic calcium phosphate, saccharides, and/or mixtures of any of the foregoing; and or
- (10) wetting agents such as corn starch, potato starch, maize starch, and modified starches, croscarmellose sodium, crosspovidone, sodium starch glycolate, and mixtures thereof; and or
- (11) disintegrants; and or
- (12) effervescent agents such as effervescent couples such as an organic acid (e.g., citric, tartaric, malic, fumaric, adipic, succinic, and alginic acids and anhydrides and acid salts), or a carbonate (e.g. sodium carbonate, potassium carbonate, magnesium carbonate, sodium glycine carbonate, L-lysine carbonate, and arginine carbonate) or bicarbonate (e.g. sodium bicarbonate or potassium bicarbonate); and or
- (13) other pharmaceutically acceptable excipients.

Medicaments of the invention suitable for use in animals and in particular in man typically must be sterile and stable under the conditions of manufacture and storage. The medicaments of the invention comprising the biologically active material can be formulated as a solid, a solution, a microemulsion, a liposome, or other ordered structures suitable to high drug concentration. Actual dosage levels of the biologically active material in the medicament of the invention may be varied in accordance with the nature of the biologically active material, as well as the potential increased efficacy due to the advantages of providing and administering the biologically active material (e.g., increased solubility, more rapid dissolution, increased surface area of the biologically active material, etc.). Thus as used herein "therapeutically effective amount" will refer to an amount of biologically active material required to effect a therapeutic response in an animal. Amounts effective for such a use will depend on: the desired

therapeutic effect; the route of administration; the potency of the biologically active material; the desired duration of treatment; the stage and severity of the disease being treated; the weight and general state of health of the patient; and the judgment of the prescribing physician.

In another embodiment, the biologically active material, optionally together with the grinding matrix or at least a portion of the grinding matrix, of the invention may be combined into a medicament with another biologically active material, or even the same biologically active material. In the latter embodiment, a medicament may be achieved which provides for different release characteristics – early release from the biologically active material, and later release from a larger average size biologically active material.

Modes of administration of medicaments comprising biologically active materials

Medicaments of the invention can be administered to animals, including man, in any pharmaceutically acceptable manner, such as orally, rectally, pulmonary, intravaginally, locally (powders, ointments or drops), transdermal, parenteral administration, intravenous, intraperitoneal, intramuscular, sublingual or as a buccal or nasal spray

Solid dosage forms for oral administration include capsules, tablets, pills, powders, pellets, and granules. Further, incorporating any of the normally employed excipients, such as those previously listed, and generally 5-95% of the biologically active agent, and more preferably at a concentration of 10%-75% will form a pharmaceutically acceptable non-toxic oral composition.

Medicaments of the invention may be parenterally administered as a solution of the biologically active agent suspended in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers may be used, e.g. water, buffered water, 0.4% saline, 0.3% glycine, hyaluronic acid and the like. These compositions may be sterilized by conventional, well known sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as is, or lyophilized, the lyophilized preparation being combined with a sterile solution prior to administration.

For aerosol administration, medicaments of the invention are preferably supplied along with a surfactant or polymer and propellant. The surfactant or polymer must, of course, be non-toxic, and preferably soluble in the propellant. Representative of such agents are the esters or partial esters of fatty acids containing from 6 to 22 carbon atoms, such as caproic, octanoic, lauric, palmitic, stearic, linoleic, linolenic, olesteric and oleic acids with an aliphatic polyhydric alcohol or its cyclic anhydride. Mixed esters, such as mixed or natural glycerides may be employed. The surfactant or polymer may constitute 0.1%-20% by weight of the composition, preferably 0.25-5%. The balance of the composition is ordinarily propellant. A carrier can also be included, as desired, as with, e.g., lecithin for intranasal delivery.

Medicaments of the invention may also be administered via liposomes, which serve to target the active agent to a particular tissue, such as lymphoid tissue, or targeted selectively to cells. Liposomes include emulsions, foams, micelles, insoluble monolayers, liquid crystals,

phospholipid dispersions, lamellar layers and the like. In these preparations the composite microstructure composition is incorporated as part of a liposome, alone or in conjunction with a molecule that binds to or with other therapeutic or immunogenic compositions.

As described above, the biologically active material can be formulated into a solid dosage form (e.g., for oral or suppository administration), together with the grinding matrix or at least a portion of it. In this case there may be little or no need to add stabilizing agents since the grinding matrix may effectively act as a solid-state stabilizer.

However, if the biologically active material is to be utilized in a liquid suspension, the particles comprising the biologically active material may require further stabilization once the solid carrier has been substantially removed to ensure the elimination, or at least minimisation of particle agglomeration.

Therapeutic uses

Therapeutic uses of the medicaments of the invention include pain relief, anti-inflammatory, migraine, asthma, and other disorders that require the active agent to be administered with a high bioavailability.

One of the main areas when rapid bioavailability of a biologically active material is required is in the relief of pain. The minor analgesics, such as cyclooxygenase inhibitors (aspirin related drugs) may be prepared as medicaments according to the present invention.

Medicaments of the invention may also be used for treatment of eye disorders. That is, the biologically active material may be formulated for administration on the eye as an aqueous suspension in physiological saline, or a gel. In addition, the biologically active material may be prepared in a powder form for administration via the nose for rapid central nervous system penetration.

Treatment of cardiovascular disease may also benefit from biologically active materials according to the invention, such as treatment of angina pectoris and, in particular, molsidomine may benefit from better bioavailability.

Other therapeutic uses of the medicaments of the present invention include treatment of hair loss, sexual dysfunction, or dermal treatment of psoriasis.

The present invention will now be described with reference to the following non-limiting Examples. The description of the Examples is in no way limiting on the preceding paragraphs of this specification, but is provided for exemplification of the methods and compositions of the invention.

Examples

It will be apparent to persons skilled in the milling and pharmaceutical arts that numerous enhancements and modifications can be made to the above described processes without departing from the basic inventive concepts. For example, in some applications the biologically

active material may be pretreated and supplied to the process in the pretreated form. All such modifications and enhancements are considered to be within the scope of the present invention, the nature of which is to be determined from the foregoing description and the appended claims. Furthermore, the following Examples are provided for illustrative purposes only, and are not intended to limit the scope of the processes or compositions of the invention.

The following materials were used in the examples: Meloxicam (Dayang, China), Diclofenac (Unique, India), Lactose monohydrate (Capsulac 60, Meggle, Germany), Mannitol (Sigma-Aldrich, US), Tartaric Acid (BDH, UK), Sorbitol (Sigma-Aldrich, US), Glucose (Ajax Finechem, Australia), Microcrystalline Cellulose (Sigma-Aldrich, US).

A Union Process attritor mill (model 1HD, 110mL milling chamber), fitted with a 4 arm rotating shaft, was used to conduct the milling experiments. Steel balls (5/16", 300g) were used as grinding media in the milling experiments. The mill was loaded through the loading port, with dry materials and matrices added initially, followed by the grinding media. The milling process was conducted at room temperature with the shaft rotating at 500 rpm. Upon completion of milling, the milled powder was discharged from the mill and sieved to remove grinding media.

The particle size distribution (PSD) was determined using a Malvern Mastersizer 2000 fitted with a Malvern Hydro 2000S pump unit. Dispersant used (0.01M HCl, RI: 1.33). Measurement settings used: Measurement Time: 12 secs, Measurement cycles: 3. Result generated by averaging the 3 measurements. Meloxicam specific conditions: Refractive index (RI): 1.73, absorption: 0.01. Diclofenac specific conditions: RI: 1.69, absorption: 0.01. Samples were prepared by adding 200mg of milled powder to 5.0mL of a 1% PVP solution in 0.01M hydrochloric acid (HCl), vortexing for 1 min, then sonicating with a horn for 1 min until samples dispersed. From this solution enough was added into the dispersant to attain a desired obscuration level of the red laser of $\approx 2.0\%$.

Dissolution behaviour of milled materials as well as unmilled controls were determined using an automated Varian 7025 dissolution unit fitted with a Cary 50 Tablet UV visible spectrometer. Dissolution settings used were according to USP 2 with stirrer speed at 100 rpm. Meloxicam specific conditions: wavelength $\lambda=362\text{nm}$, pH 6.1 (10mM Phosphate buffer), standard sized gelatine capsules contained 15mg Meloxicam, for example, a capsule prepared from a 10wt% Meloxicam milling required 150mg milled powder. Diclofenac specific conditions: wavelength $\lambda=276\text{nm}$, pH 5.75 (10mM Citrate buffer), standard sized gelatine capsules contained 20mg Diclofenac, for example, a capsule prepared from a 10wt% Diclofenac milling required 200mg milled powder. Capsules of milled materials were filled using Profill® equipment. Un-milled control samples were prepared by hand-filling appropriately sized capsules. Each dissolution result was obtained by averaging results from 3 capsules. Quantitative results are given as the time to reach X and Y.: X is defined as the concentration equal to the dissolution concentration achieved by a control sample (or prototype formulation thereof) of the biologically active material or compound after 60 minutes. Y is defined as the concentration equal to the

dissolution concentration achieved by a control sample (or prototype formulation thereof) of the biologically active material or compound after 30 minutes.

Powder X-Ray diffraction (XRD) patterns were measured with a Diffractometer D 5000, Kristalloflex (Siemens). The measurement range was from 5-18 degrees 2-Theta. The slit width was set to 2 mm and the cathode ray tube was operated at 40 kV and 35 mA. Measurements were recorded at room temperature. The recorded traces were subsequently processed using Bruker EVA software to obtain the diffraction pattern.

DSC traces were measured using a TA instruments DSC Q10. The data was obtained using a heating rate of 10 °C/min under nitrogen flow. Aluminium Tzero open pans were used for the measurements.

Example 1. 10% Meloxicam in Lactose mono-hydrate:

A mixture of Meloxicam (0.60g) and Lactose monohydrate (5.40g) was milled for either 1 (B) or 2 (C) minutes. PSDs of the milled products and unmilled material (A) are shown in Figure 1. The dissolution behaviour is shown in Figure 2. Results are summarised in Table 1 together with results obtained for an un-milled control (A), prepared by physically mixing Meloxicam (0.40g) and Lactose monohydrate (3.60g) in a vial until the appearance was homogenous.

Figure 1 shows that after 1 minute of milling the particle size is reduced by about half. After another minute of milling the particle size has further reduced but is still mostly in the range of 1-10 micron. In contrast to this the dissolution of the material milled for 1 minute is only slightly faster than the unmilled control sample. The dissolution at 2 minutes is dramatically improved over both the 1 minute and unmilled material. In Table 1 the median size and quantitative assessment of the dissolution are shown. According to the measures X and Y (set out above) the material milled for 2 minutes has a much improved dissolution compared with both the unmilled and the milled for 1 minute sample.

As the change in size of material from 1 to 2 mins is of the same order as the change in size from unmilled to 1 minute the primary reason for the improved dissolution for the 2 minute sample cannot be particle size reduction.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	8.79 µm	30	60
1 min (B)	4.86 µm	23	45
2 min (C)	2.53 µm	8	11

Table 1.

Example 2. 10% Diclofenac in Lactose mono-hydrate:

A mixture of Diclofenac (0.60g) and lactose monohydrate (5.40g) was milled for either 1 (B) or 2 (C) minutes. PSDs of the milled products and unmilled material (A) are shown in Figure 3. The dissolution behaviour is shown in Figure 4. Results are summarised in Table 2 together with results obtained for an un-milled control (A), prepared by physically mixing Diclofenac (0.40g) and Lactose monohydrate (3.60g) in a vial until the appearance was homogenous.

The data for Diclofenac milled in lactose monohydrate is very similar to the data in Example 1. Figure 3 shows that after 1 minute of milling the particle size is reduced by just over 50%. After another minute of milling the particle size has reduced a little more giving two milled materials in the range 2-4 micron. Again in contrast to this the dissolution of the material milled for 1 minute is only slightly faster than the unmilled control sample. The dissolution at 2 minutes is dramatically improved over both the 1 minute and unmilled material. In Table 1 the median size and quantitative assessment of the dissolution are shown. According to the measures X and Y (set out above) the material milled for 2 minutes has a much improved dissolution compared with both the unmilled and the milled for 1 minute sample.

As the size of the material from 1 to 2 mins is quite similar this size difference cannot be the primary reason for the improved dissolution for the 2 minute sample.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	9.50 μm	30	60
1 min (B)	4.09 μm	18	29
2 min (C)	2.57 μm	8	10

Table 2.

Example 3. 10% Meloxicam in Mannitol:

A mixture of Meloxicam (0.60g) and Mannitol (5.40g) was milled for either 1 (B) or 2 (C) minutes. PSDs of the milled products and unmilled material (A) were measured as well as the dissolution behaviour. Results are summarised in Table 3. The un-milled control (A) was prepared by physically mixing Meloxicam (0.40g) and Mannitol (3.60g) in a vial until the appearance was homogenous.

The PSD shows that the material milled for 1 and 2 minutes has a reduced size compared with the unmilled material, but the size reduction is not dramatic. According to the dissolution measures X and Y both materials have a much improved dissolution rate compared with the unmilled sample. This data also shows that once enough milling energy has been input to

deliver the improved dissolution (1 minute milling), further size reduction (2 minutes) has little impact on the dissolution rate.

In Figure 5 a DSC trace of material milled for 2 minutes is shown compared with the DSC trace of manitol. The trace only shows one melt other than mannitol at approximately 240°C being the normal melting point of meloxicam. This DSC trace shows no indication of any amorphous material or other forms of meloxicam being present. This indicates the meloxicam has retained its crystallinity during the milling process.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	8.79 μm	30	60
1 min (B)	3.80 μm	10	12
2 min (C)	2.19 μm	8	9

Table 3.

Example 4 10% Diclofenac in Mannitol:

A mixture of Diclofenac (0.60g) and Mannitol (5.40g) was milled for either 1 (B) or 2 (C) minutes. PSDs of the milled products and unmilled material (A) were measured as well as the dissolution behaviour. Results are summarised in Table 4. The un-milled control (A) was prepared by physically mixing Diclofenac (0.40g) and Mannitol (3.60g) in a vial until the appearance was homogenous.

The PSD shows that the material milled for 1 and 2 minutes has a reduced size compared with the unmilled material, but the size is still in the range 1-10 microns. According to the dissolution measures X and Y both materials have a much improved dissolution rate compared with the unmilled sample. Again the data also shows that once enough milling energy has been input to deliver the improved dissolution (1 minute milling), further size reduction (2 minutes) has little impact on the dissolution rate.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	9.50 μm	30	60
1 min (B)	2.20 μm	8	11
2 min (C)	1.23 μm	8	11

Table 4.

Example 5 10% Meloxicam in Glucose:

A mixture of Meloxicam (0.60g) and Glucose (5.40g) was milled for either 1 (B) or 2 (C) minutes. PSDs of the milled products and unmilled material (A) were measured as well as the dissolution behaviour. Results are summarised in Table 5. The un-milled control (A) was prepared by physically mixing Meloxicam (0.40g) and Glucose (3.60g) in a vial until the appearance was homogenous.

The PSD shows that the material milled for 1 and 2 minutes has a reduced size compared with the unmilled material. There is about a 50% reduction from unmilled to 1 minute and about another 50% reduction from 1 minute to 2 minutes. According to the dissolution measures X and Y both milled materials have a much improved dissolution rate compared with the unmilled sample. Again the data shows that the improved dissolution is independent of the final particle size, instead most improvement has come from the milling of the active with the grinding matrix.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	8.79 μm	30	60
1 min (B)	4.04 μm	9	10
2 min (C)	1.61 μm	7	8

Table 5.

Example 6. 10% Diclofenac in Glucose:

A mixture of Diclofenac (0.60g) and Glucose (5.40g) was milled for either 1 (B) or 2 (C) minutes. PSDs of the milled products and unmilled material (A) were measured as well as the dissolution behaviour. Results are summarised in Table 6. The un-milled control (A) was prepared by physically mixing Diclofenac (0.40g) and Glucose (3.60g) in a vial until the appearance was homogenous.

The PSD shows that the material milled for 1 and 2 minutes has a reduced size compared with the unmilled material, There is about a 60% reduction from unmilled to 1 minute and about another 30% reduction from 1 minute to 2 minutes. According to the dissolution measures X and Y the material milled for 1 minute has a greatly improved dissolution rate compared with the unmilled sample. The material milled for 2 minutes has a much slower dissolution rate compared with sample B and is only slightly improved compared with the unmilled material even though the particle size is smaller.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	9.50 μm	30	60
1 min (B)	3.13 μm	15	24
2 min (C)	1.97 μm	25	55

Table 6.

Example 7. 10% Meloxicam in microcrystalline Cellulose:

A mixture of Meloxicam (0.60g) and microcrystalline Cellulose (5.40g) was milled for either 1 (B) or 2 (C) minutes. No PSD was measured due to interference from insoluble excipient. Dissolution behaviour of milled products and unmilled material (A) were measured. Results are summarised in Table 7. The un-milled control (A) was prepared by physically mixing Meloxicam (0.40g) and microcrystalline Cellulose (3.60g) in a vial until the appearance was homogenous.

According to the dissolution measures X and Y both milled materials have an improved dissolution rate compared with the unmilled sample.

Milling Time	Time to Reach Concentration	
	Y (min)	X (min)
Un-milled (A)	30	60
1 min (B)	10	14
2 min (C)	9	10

Table 7.

Example 8. 10% Diclofenac in microcrystalline Cellulose:

A mixture of Diclofenac (0.60g) and microcrystalline Cellulose (5.40g) was milled for either 1 (B) or 2 (C) minutes. No PSD was measured due to interference from insoluble excipient. Dissolution behaviour of milled products and unmilled material (A) were measured. Results are summarised in Table 7. The un-milled control (A) was prepared by physically mixing Diclofenac (0.40g) and microcrystalline Cellulose (3.60g) in a vial until the appearance was homogenous. According to the dissolution measures X and Y both milled materials have an improved dissolution rate compared with the unmilled sample.

Milling Time	Time to Reach Concentration	
	Y (min)	X (min)
Un-milled (A)	30	60
1 min (B)	18	28
2 min (C)	24	31

Table 8.

Example 9. 10% Meloxicam in Tartaric acid:

A mixture of Meloxicam (0.60g) and Tartaric acid (5.40g) was milled for either 1 (B) or 2 (C) minutes. PSDs of the milled products and unmilled material (A) were measured as well as dissolution behaviour[#]. Results summarised in Table 9. The un-milled control (A) was prepared by physically mixing Meloxicam (0.40g) and Tartaric acid (3.60g) in a vial until the appearance was homogenous.

The PSD shows that the material milled for 1 and 2 minutes has a reduced size compared with the unmilled material, There is about a 40% reduction from unmilled to 1 minute and about another 40% reduction from 1 minute to 2 minutes. According to the dissolution measures X and Y both milled materials have a much improved dissolution rate compared with the unmilled sample. The dissolution data indicates that both milled materials have very fast dissolution even though the size reduction upon milling is not large.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	8.79 μm	30	60
1 min (B)	5.10 μm	9	11
2 min (C)	3.03 μm	8	9

Table 9.

[#]Dissolution test measured in 100mM phosphate buffer at pH 5.8.

Example 10. 20% Meloxicam in Lactose mono-hydrate:

A mixture of Meloxicam (1.20g) and Lactose monohydrate (4.80g) was milled for either 1 (B) or 2 (C) minutes. PSDs of the milled products and unmilled material (A) were measured as well as dissolution behaviour. Results summarised in Table 10. The un-milled control (A) was prepared

by physically mixing Meloxicam (0.80g) and Lactose monohydrate (3.20g) in a vial until the appearance was homogenous.

The PSD shows that the material milled for 1 and 2 minutes has a reduced size compared with the unmilled material. According to the dissolution measures X and Y both milled materials have an improved dissolution rate compared with the unmilled sample.

In Figure 6 the XRD spectra of the material milled for 2 minutes is shown. The spectra of pure meloxicam and pure milled lactose are also shown. These show that most meloxicam peaks are obscured by the lactose spectra. The clearest meloxicam peak is located at 2 theta 15°. For the material milled for 2 mins this peak is small (due to only 20 % meloxicam) but evidence of the presence of crystalline meloxicam after milling. The spectra also indicate that the lactose is still crystalline after milling as well.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	8.79 μm	30	60
1 min (B)	5.72 μm	14	26
2 min (C)	3.52 μm	17	20

Table 10.

Example 11. 20% Meloxicam in Mannitol:

A mixture of Meloxicam (1.20g) and Mannitol (4.80g) was milled for either 1 (B) or 2 (C) minutes. PSDs of the milled products and unmilled material (A) were measured as well as dissolution behaviour. Results are summarised in Table 11. The un-milled control (A) was prepared by physically mixing Meloxicam (0.80g) and Mannitol (3.20g) in a vial until the appearance was homogenous.

The PSD shows that the material milled for 1 and 2 minutes has a reduced size compared with the unmilled material. The level of size reduction compared with the material milled at 10% (example 3) is the same. The dissolution rate for the material milled at 20% is slightly slower than the rate for material milled at 10% (example 3) but the rate is still a good improvement over that of the unmilled material. Again this data would indicate that the improvement in dissolution observed is not primarily a function of particle size.

In Figure 5 a DSC trace of material milled for 2 minutes is shown compared with the DSC trace of mannitol. The trace only shows one melt other than mannitol at approximately 240°C being the normal melting point of meloxicam. This DSC trace shows no indication of any amorphous

material or other forms of meloxicam being present. This indicates the meloxicam has retained its crystallinity during the milling process.

In Figure 7 the XRD spectra of the material milled for 2 minutes is shown. The spectra of pure meloxicam, pure mannitol and a 20% physical mixture of meloxicam in mannitol are also shown. These show that most meloxicam peaks are obscured by the mannitol spectra. The clearest meloxicam peak is located at 2 theta 13°. The spectra indicate that both the meloxicam and mannitol are still crystalline after milling.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	8.79 μm	30	60
1 min (B)	3.53 μm	14	22
2 min (C)	2.39 μm	18	21

Table 11:

Example 12. 30% Diclofenac in 69% Lactose mono-hydrate and 1% Sodium dodecyl sulfate:

A mixture of Diclofenac (1.80g), Lactose monohydrate (4.14g) and Sodium dodecyl sulfate (SDS) (0.06g) was milled for 10 minutes (B). PSDs of the milled product and unmilled material (A) were measured as well as dissolution behaviour. Results are summarised in Table 12. The un-milled control (A) was prepared by physically mixing Diclofenac (1.20g), Lactose monohydrate (2.76g) and SDS (0.04g) in a vial until the appearance was homogenous.

At the higher API content 1% SDS has been used as a milling aid to help provide good flow during milling. The same concentration of SDS was also include in the unmilled control sample for dissolution measurements so that any improvement in the dissolution due to the SDS is accounted for. At this API concentration the milling time has also been extended to provide more milling energy. The PSD achieved here is similar to the 2 minute sample from example 2 (10%) and the dissolution measures X and Y have also shown a similar level of improved dissolution. This example demonstrates that the improved dissolution through the synergistic milling of API and grinding matrix is achieved at higher API levels.

In Figure 8 the XRD spectra of the diclofenac milled at various weight percentages from 20-50 % is shown. The 20 % material was produced in the same way as this example only with different amounts of diclofenac and lactose so as to achieve 20 % w/w diclofenac overall. In Figure 9 spectra of unmilled physical mixtures of the same compositions are shown as a comparison. In Figure 10 spectra are also shown for pure diclofenac, pure lactose and pure milled lactose. Figure 10 indicates there are unobscured peaks located at 2 theta 11°, 15° and

a partially obscured peak at 28° . When these peaks are compared between Figure 8 (milled) and Figure 9 (physical mixture) the spectra indicates that the material procuded by this example is still crystalline after milling.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	9.50 μm	30	60
10 min (B)	2.75 μm	12	13

Table 12.

Example 13. 40% Diclofenac in 59% Lactose mono-hydrate and 1% Sodium dodecyl sulfate:

A mixture of Diclofenac (2.40g), Lactose monohydrate (3.54g) and Sodium dodecyl sulfate (SDS) (0.06g) was milled for 10 minutes (B). PSDs of the milled product and unmilled material (A) were measured as well as dissolution behaviour. Results are summarised in Table 13. The un-milled control (A) was prepared by physically mixing Diclofenac (1.60g), Lactose monohydrate (2.36g) and SDS (0.04g) in a vial until the appearance was homogenous.

At this API concentration the PSD achieved is slightly coarser compare to example 12 (30%). The dissolution measures X and Y showed improved dissolution.

In Figure 8 the XRD spectra of the diclofenac milled at various weight percentages from 20-50 % is shown. The 20 % material was produced in the same way as example 12 only with different amounts of diclofenac and lactose so as to achieve 20 % w/w diclofenac overall. In Figure 9 spectra of unmilled physical mixtures of the same compositions are shown as a comparison. In Figure 10 spectra are also shown for pure diclofenac, pure lactose and pure milled lactose. Figure 10 indicates there are unobscured peaks located at 2 theta 11° , 15° and a partially obscured peak at 28° . When these peaks are compared between Figure 8 (milled) and Figure 9 (physical mixture) the spectra indicates that the material procuded by this example is still crystalline after milling.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	9.50 μm	30	60
10 min (B)	4.45 μm	18	25

Table 13.

Example 14. 50% Diclofenac in 49% Lactose mono-hydrate and 1% Sodium dodecyl sulfate:

A mixture of Diclofenac (3.00g), Lactose monohydrate (2.94g) and Sodium dodecyl sulfate (SDS) (0.06g) was milled for 10 minutes (B). PSDs of the milled product and unmilled material (A) were measured as well as dissolution behaviour. Results are summarised in Table 14. The un-milled control (A) was prepared by physically mixing Diclofenac (2.00g), Lactose monohydrate (1.96g) and SDS (0.04g) in a vial until the appearance was homogenous.

At this API concentration the PSD achieved is slightly coarser compare to example 12 (30%) and example 13 (40%). The dissolution measures X and Y still clearly indicate improved dissolution. This example demonstrates that the improved dissolution through the synergistic milling of API and grinding matrix is achieved at API levels up to at least 50%.

In Figure 8 the XRD spectra of the diclofenac milled at various weight percentages from 20-50 % is shown. The 20 % material was produced in the same way as example 12 only with different amounts of diclofenac and lactose so as to achieve 20 % w/w diclofenac overall. In Figure 9 spectra of unmilled physical mixtures of the same compositions are shown as a comparison. In Figure 10 spectra are also shown for pure diclofenac, pure lactose and pure milled lactose. Figure 10 indicates there are unobscured peaks located at 2 theta 11°, 15° and a partially obscured peak at 28°. When these peaks are compared between Figure 8 (milled) and Figure 9 (physical mixture) the spectra indicates that the material produced by this example is still crystalline after milling.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	9.50 µm	30	60
10 min (B)	5.65 µm	23	33

Table 14.

Example 15. 30% Meloxicam in 69% Lactose mono-hydrate and 1% Sodium dodecyl sulfate:

A mixture of Meloxicam (1.80g), Lactose monohydrate (4.14g) and Sodium dodecyl sulfate (SDS) (0.06g) was milled for 10 minutes (B). PSDs of the milled product and unmilled material (A) were measured as well as dissolution behaviour. Results are summarised in Table 15. The un-milled control (A) was prepared by physically mixing Meloxicam (1.20g), Lactose monohydrate (2.76g) and SDS (0.04g) in a vial until the appearance was homogenous.

Like the Diclofenac examples at higher API content 1% SDS has also been used as a milling aid with the high Meloxicam content millings to help provide good flow during milling. The same concentration of SDS was also include in the unmilled control sample for dissolution

measurements so that any improvement in the dissolution due to the SDS is accounted for. At this API concentration the milling time has also been extended to provide more milling energy. The PSD achieved here is slightly larger than the 2 minute sample from example 1 (10%). The dissolution measures X and Y show slightly more improvement in the dissolution compared with the 2 minute sample of example 1.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	8.79 μm	30	60
10 min (B)	3.69 μm	6	7

Table 15.

Example 16. 40% Meloxicam in 59% Lactose mono-hydrate and 1% Sodium dodecyl sulfate:

A mixture of Meloxicam (2.40g), Lactose monohydrate (3.54g) and Sodium dodecyl sulfate (SDS) (0.06g) was milled for 10 minutes (B). PSDs of the milled product and unmilled material (A) were measured as well as dissolution behaviour. Results are summarised in Table 16. The un-milled control (A) was prepared by physically mixing Meloxicam (1.60g), Lactose monohydrate (2.36g) and SDS (0.04g) in a vial until the appearance was homogenous.

The PSD achieved here is slightly larger than 30% sample (example 15) but the dissolution measures X and Y are virtually the same, again indicating strongly improved dissolution.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	8.79 μm	30	60
10 min (B)	4.91 μm	7	8

Table 16.

Example 17. 50% Meloxicam in 49% Lactose mono-hydrate and 1% Sodium dodecyl sulfate:

A mixture of Meloxicam (3.00g), Lactose monohydrate (2.94g) and Sodium dodecyl sulfate (SDS) (0.06g) was milled for 10 minutes (B). PSDs of the milled product and unmilled material (A) were measured as well as dissolution behaviour. Results are summarised in Table 17. The un-milled control (A) was prepared by physically mixing Meloxicam (2.00g), Lactose monohydrate (1.96g) and SDS (0.04g) in a vial until the appearance was homogenous.

The PSD achieved here is slightly larger than 40% sample (example 16) and is only slightly smaller than the unmilled material. The dissolution measures X and Y are very similar to the 30 and 40%, again indicating strongly improved dissolution. This series of millings at high Meloxicam content (example 15,16,17) clearly demonstrates that improved dissolution by synergistic milling of API with a grinding matrix is possible to at least 50 %. The PSD distributions for this series also indicate that the improved dissolution observed from this process is independent of particle size. From 30% to 50% the PSD almost doubles yet the dissolution has remained relatively constant indicating little or no influence from particle size.

In Figure 6 the XRD spectra of the material is shown (Spectra D). The spectra of pure meloxicam and pure milled lactose are also shown. These show that most meloxicam peaks are obscured by the lactose spectra. The clearest meloxicam peak is located at 2 theta 15°. In Figure 11 the spectra of a physical mixture of the material milled is also shown. The spectra indicates the presence of crystalline meloxicam after milling. The spectra also indicate that the lactose is still crystalline after milling as well.

Milling Time	Size	Time to Reach Concentration	
		Y (min)	X (min)
Un-milled (A)	8.79 μm	30	60
10 min (B)	6.22 μm	10	13

Table 17.

CLAIMS:

1. A method for improving the dissolution profile of a biologically active material, comprising the steps of:
dry milling a solid biologically active material and a millable grinding matrix in a mill comprising a plurality of milling bodies, for a time period sufficient to produce particles of the biologically active material dispersed in an at least partially milled grinding matrix,
wherein the particles have an average particle size falling within the range of 1 – 200 μm determined on a particle number basis,
wherein the average particle size of the biologically active material has been reduced by less than 70% during dry milling and the dissolution of the biologically active material is improved such that a dissolution concentration of X is achieved in 10-30 minutes and X is the concentration equal to the dissolution concentration of the biologically active material prior to dry milling after 60 minutes,
wherein the particle size of the millable grinding matrix is reduced by the dry milling,
wherein the solid biologically active compound and the millable grinding matrix are milled with a facilitating agent,
wherein the millable grinding matrix is selected from the group consisting of: lactose anhydrous, lactose monohydrate and mannitol,
wherein the time period sufficient is a range selected between 5 minutes and 8 hours,
wherein the milling bodies are balls having a diameter between 1mm and 20 mm, and
wherein the biologically active material is selected from the group consisting of: anti-obesity drugs, central nervous system stimulants, carotenoids, corticosteroids, elastase inhibitors, anti-fungals, oncology therapies, anti-emetics, analgesics, cardiovascular agents, anti-inflammatory agents, such as NSAIDs and COX-2 inhibitors, anthelmintics, anti-arrhythmic agents, antibiotics, penicillins, anticoagulants, antidepressants, antidiabetic agents, antiepileptics, antihistamines, antihypertensive agents, antimuscarinic agents, antimycobacterial agents, antineoplastic agents, immunosuppressants, antithyroid agents, antiviral agents, anxiolytics, sedatives (hypnotics and neuroleptics), astringents, alpha-adrenergic receptor blocking agents, beta-adrenoceptor blocking agents, blood products and substitutes, cardiac

inotropic agents, contrast media, cough suppressants, expectorants, mucolytics, diagnostic agents, diagnostic imaging agents, diuretics, dopaminergics, anti-parkinsonian agents, haemostatics, immunological agents, lipid regulating agents, muscle relaxants, parasympathomimetics, parathyroid calcitonin and biphosphonates, prostaglandins, radio-pharmaceuticals, sex hormones, steroids, anti-allergic agents, stimulants and anoretics, sympathomimetics, thyroid agents, vasodilators, and xanthines.

2. The method of claim 1, wherein the average particle size of the biologically active material has been reduced by a factor selected from the group consisting of: less than 5%, less than 10%, less than 20%, less than 30%, less than 40%, less than 50%, and less than 60%.
3. The method of any one of the claims 1 or 2, wherein the average particle size falls within the range selected from the group consisting of: 1-150 μ m, 1-100 μ m, 1-50 μ m, 1-20 μ m, 1-10 μ m, 1-7.5 μ m, 1-5 μ m and 1-2 μ m.
4. A method for improving the dissolution profile of a biologically active material, comprising the steps of:
dry milling a solid biologically active material and a millable grinding matrix in a mill comprising a plurality of milling bodies, for a time period sufficient to produce particles of the biologically active material dispersed in an at least partially milled grinding matrix,
wherein wherein the particles have a median particle size falling within the range of 1 – 200 μ m determined on a particle volume basis,
wherein the average particle size of the biologically active material has been reduced by less than 70% and the dissolution of the biologically active material is improved such that a dissolution concentration of X is achieved in 10-30 minutes and X is the concentration equal to the dissolution concentration of the biologically active material prior to dry milling after 60 minutes,
wherein the particle size of the millable grinding matrix is reduced by the dry milling,
wherein the solid biologically active compound and the millable grinding matrix are milled with a facilitating agent,

wherein the millable grinding matrix is selected from the group consisting of: lactose anhydrous, lactose monohydrate and mannitol,

wherein the time period sufficient is a range selected between 5 minutes and 8 hours,

wherein the milling bodies are balls having a diameter between 1mm and 20 mm, and

wherein the biologically active material is selected from the group consisting of: anti-obesity drugs, central nervous system stimulants, carotenoids, corticosteroids, elastase inhibitors, anti-fungals, oncology therapies, anti-emetics, analgesics, cardiovascular agents, anti-inflammatory agents, such as NSAIDs and COX-2 inhibitors, anthelmintics, anti-arrhythmic agents, antibiotics, penicillins, anticoagulants, antidepressants, antidiabetic agents, antiepileptics, antihistamines, antihypertensive agents, antimuscarinic agents, antimycobacterial agents, antineoplastic agents, immunosuppressants, antithyroid agents, antiviral agents, anxiolytics, sedatives (hypnotics and neuroleptics), astringents, alpha-adrenergic receptor blocking agents, beta-adrenoceptor blocking agents, blood products and substitutes, cardiac inotropic agents, contrast media, cough suppressants, expectorants, mucolytics, diagnostic agents, diagnostic imaging agents, diuretics, dopaminergics, anti-parkinsonian agents, haemostatics, immunological agents, lipid regulating agents, muscle relaxants, parasympathomimetics, parathyroid calcitonin and biphosphonates, prostaglandins, radio-pharmaceuticals, sex hormones, steroids, anti-allergic agents, stimulants and anoretics, sympathomimetics, thyroid agents, vasodilators, and xanthines.

5. The method of claim 4 wherein the percentage of particles greater than 1 μm on a particle volume basis is a percentage selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 100%.
6. The method of claim 4 wherein the percentage of particles greater than 2 μm on a particle volume basis is a percentage selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 100%.
7. The method of any preceding claim, wherein the facilitating agent is a surfactant.

8. The method of any preceding claim, wherein the surfactant is sodium lauryl sulphate.
9. The method of any one of claims 1 to 7, wherein X is reached within 10-20 minutes.
10. The method of any one of claims 1 to 8, wherein the dissolution of the biologically active material is improved such that a dissolution concentration of Y is achieved in 10-15 minutes and Y is the concentration equal to the dissolution concentration of the biologically active material prior to dry milling after 30 minutes.
11. A method of any one of claims 2 to 9, wherein the median particle size has been reduced by a factor selected from the group consisting of: less than 5%, less than 10%, less than 20%, less than 30%, less than 40%, less than 50%, less than 60%, less than 70%, less than 80%, less than 90 %, less than 95% and less than 99%.
12. The method of any one of claims 2 to 10, wherein the median particle size falls within the range selected from the group consisting of: 11-150 μ m, 1-100 μ m, 1-50 μ m, 1-20 μ m, 1-10 μ m, 1-7.5 μ m, 1-5 μ m 1-2 μ m, 2-1000 μ m, 2-500 μ m, 2-300 μ m, 2-200 μ m, 2-150 μ m, 2-100 μ m, 2-50 μ m, 2-20 μ m, 2-10 μ m, 2-7.5 μ m and 2-5 μ m.
13. The method of any preceding claim, wherein the crystallinity profile of the biologically active material is selected from the group consisting of: at least 50% of the biologically active material is crystalline, at least 60% of the biologically active material is crystalline, at least 70% of the biologically active material is crystalline, at least 75% of the biologically active material is crystalline, at least 85% of the biologically active material is crystalline, at least 90% of the biologically active material is crystalline, at least 95% of the biologically active material is crystalline and at least 98% of the biologically active material is crystalline.
14. The method of any preceding claim, wherein the crystallinity profile of the biologically active material is substantially equal to the crystallinity profile of

the biologically active material before the material was subject to the method of any one of the preceding claims.

15. The method of any preceding claim, wherein the amorphous content of the biological active material is selected from the group consisting of: less than 50% of the biologically active material is amorphous, less than 40% of the biologically active material is amorphous, less than 30% of the biologically active material is amorphous, less than 25% of the biologically active material is amorphous, less than 15% of the biologically active material is amorphous, less than 10% of the biologically active material is amorphous, less than 5% of the biologically active material is amorphous and less than 2% of the biologically active material is amorphous.
16. The method of any preceding claim, wherein the biologically active material has no significant increase in amorphous content before subjecting the material to the method of any one of the preceding claims.
17. The method of any preceding claim, wherein the milling time period is a range selected from the group consisting of: between 10 minutes and 2 hours, between 10 minutes and 1 hour, between 10 minutes and 45 minutes, between 10 minutes and 30 minutes, between 5 minutes and 30 minutes, between 5 minutes and 20 minutes, between 2 minutes and 10 minutes, between 2 minutes and 5 minutes, between 1 minute and 20 minutes, between 1 minute and 10 minutes, and between 1 minute and 5 minutes.
18. The method of any preceding claim, wherein the dry milling is undertaken in a mechanically agitated attritor mill (horizontal or vertical), vibratory mill or nutating mill, wherein the milling bodies are steel balls having a diameter selected from the group consisting of: between 2 and 15 mm and between 3 and 10 mm.
19. The method of any preceding claim, wherein the total combined amount of biologically active material and grinding matrix in the mill at any given time is equal to or greater than a mass selected from the group consisting of: 200 grams, 500 grams, 1 kg, 2kg, 5 kg, 10 kg, 20 kg, 30 kg, 50 kg, 75 kg, 100kg, 150 kg, 200 kg.

20. The method of any preceding claim, wherein the biologically active material is selected from the group consisting of: indomethacin, diclofenac, naproxen, meloxicam, metaxalone, cyclosporin A, progesterone, celecoxib, cilostazol, ciprofloxacin, 2,4-dichlorophenoxyacetic acid, anthraquinone, creatine monohydrate, glyphosate, halosulfuron, mancozeb, metsulfuron, salbutamol, sulphur, tribenuran and estradiol or any salt or derivative thereof.
21. The method of any preceding claim, wherein a milling aid or combination of milling aids is used and wherein the milling aid is selected from the group consisting of: colloidal silica, a polymer, a stearic acid and derivatives thereof.
22. The method of any one of claims 21, wherein the milling aid has a concentration selected from the group consisting of: 0.1 -10 % w/w, 0.1 -5 % w/w, 0.1 -2.5 % w/w, of 0.1 – 2% w/w, 0.1 -1 %, 0.5 -5% w/w, 0.5 -3% w/w, 0.5 -2% w/w, 0.5 – 1.5%, 0.5 -1 % w/w, of 0.75 – 1.25 % w/w, 0.75 -1% and 1% w/w.
23. The method of any preceding claim, wherein the facilitating agent is added during dry milling at a time selected from the group consisting of: with 100% of the total milling time remaining, with 1-5 % of the total milling time remaining, with 1-10 % of the total milling time remaining, with 1-20 % of the total milling time remaining, with 1-30 % of the total milling time remaining, with 2-5% of the total milling time remaining, with 2-10% of the total milling time remaining and with 5-20% of the total milling time remaining.
24. The method of any preceding claim, wherein the dissolution profile of a measurement sample or prototype formulation thereof is improved by a factor selected from the group consisting of: wherein X is reached in 10 minutes, wherein X is reached within 10-20 minutes, wherein X is reached within 10-30 mins, wherein X is reached within 10-40 mins, wherein X is reached within 10-50 mins, wherein X is reached within 20-30 mins, wherein X is reached within 20-40 mins, wherein X is reached within 20-50 mins, wherein X is reached within 30-40 mins, wherein X is reached within 30-50 mins and wherein X is reached within 40-50 mins, wherein X is defined as the

concentration equal to the dissolution concentration achieved by a control sample or prototype formulation thereof of the biologically active material or compound after 60 minutes.

25. The method of any preceding claim, wherein the dissolution profile of a measurement sample or prototype formulation thereof is improved by a factor selected from the group consisting of: wherein Y is reached in 5 minutes, wherein Y is reached within 10 minutes, wherein Y is reached within 10-15 mins, wherein Y is reached within 10-20 mins, wherein Y is reached within 10-25 mins, wherein Y is reached within 15-20 mins, wherein Y is reached within 15-25 mins, wherein Y is reached within 20-25 mins, wherein Y is defined as the concentration equal to the dissolution concentration achieved by a control sample (or prototype formulation thereof) of the biologically active material or compound after 30 minutes.
26. A composition comprising a biologically active material produced by the method of any one of claims 1 to 25.
27. A composition produced by the method for improving the dissolution profile of a biologically active material, comprising the steps of:
dry milling a solid biologically active material and a millable grinding matrix in a mill comprising a plurality of milling bodies, for a time period sufficient to produce particles of the biologically active material dispersed in an at least partially milled grinding matrix,
wherein the particles have an average particle size falling within the range of 1 – 200 μm determined on a particle number basis,
wherein the average particle size of the biologically active material has been reduced by less than 70% during dry milling and the dissolution of the biologically active material is improved such that a dissolution concentration of X is achieved in 10-30 minutes and X is the concentration equal to the dissolution concentration of the biologically active material prior to dry milling after 60 minutes,
wherein the particle size of the millable grinding matrix is reduced by the dry milling,
wherein the solid biologically active compound and the millable grinding matrix are milled with a facilitating agent,

wherein the millable grinding matrix is selected from the group consisting of: lactose anhydrous, lactose monohydrate and mannitol,

wherein the time period sufficient is a range selected between 5 minutes and 8 hours,

wherein the milling bodies are balls having a diameter between 1mm and 20 mm, and

wherein the biologically active material is selected from the group consisting of: anti-obesity drugs, central nervous system stimulants, carotenoids, corticosteroids, elastase inhibitors, anti-fungals, oncology therapies, anti-emetics, analgesics, cardiovascular agents, anti-inflammatory agents, such as NSAIDs and COX-2 inhibitors, anthelmintics, anti-arrhythmic agents, antibiotics, penicillins, anticoagulants, antidepressants, antidiabetic agents, antiepileptics, antihistamines, antihypertensive agents, antimuscarinic agents, antimycobacterial agents, antineoplastic agents, immunosuppressants, antithyroid agents, antiviral agents, anxiolytics, sedatives (hypnotics and neuroleptics), astringents, alpha-adrenergic receptor blocking agents, beta-adrenoceptor blocking agents, blood products and substitutes, cardiac inotropic agents, contrast media, cough suppressants, expectorants, mucolytics, diagnostic agents, diagnostic imaging agents, diuretics, dopaminergics, anti-parkinsonian agents, haemostatics, immunological agents, lipid regulating agents, muscle relaxants, parasympathomimetics, parathyroid calcitonin and biphosphonates, prostaglandins, radio-pharmaceuticals, sex hormones, steroids, anti-allergic agents, stimulants and anoretics, sympathomimetics, thyroid agents, vasodilators, and xanthines.

28. A composition of claim 27, wherein the average particle size of the biologically active material has been reduced by a factor selected from the group consisting of: less than 5%, less than 10%, less than 20%, less than 30%, less than 40%, less than 50%, less than 60%, less than 70%, less than 80%, less than 90 %, less than 95% and less than 99%.

29. A composition of any one of the claims 27 or 28, wherein the average particle size falls within the range selected from the group consisting of: 1-150 μ m, 1-100 μ m, 1-50 μ m, 1-20 μ m, 1-10 μ m, 1-7.5 μ m, 1-5 μ m and 1-2 μ m.

30. A composition produced by a method for improving the dissolution profile of a biologically active material, comprising the steps of:.
- dry milling a solid biologically active material and a millable grinding matrix in a mill comprising a plurality of milling bodies, for a time period sufficient to produce particles of the biologically active material dispersed in an at least partially milled grinding matrix,
- wherein wherein the particles have a median particle size falling within the range of 1 – 200 μm determined on a particle volume basis,
- wherein the average particle size of the biologically active material has been reduced by less than 70% during dry milling and the dissolution of the biologically active material is improved such that a dissolution concentration of X is achieved in 10-30 minutes and X is the concentration equal to the dissolution concentration of the biologically active material prior to dry milling after 60 minutes,
- wherein the particle size of the millable grinding matrix is reduced by the dry milling,
- wherein the solid biologically active compound and the millable grinding matrix are milled with a facilitating agent,
- wherein the millable grinding matrix is selected from the group consisting of: lactose anhydrous, lactose monohydrate and mannitol,
- wherein the time period sufficient is a range selected between 5 minutes and 8 hours,
- wherein the milling bodies are balls having a diameter between 1mm and 20 mm, and
- wherein the biologically active material is selected from the group consisting of: fungicides, pesticides, herbicides, seed treatments, cosmeceuticals, cosmetics, complementary medicines, natural products, vitamins, nutrients, nutraceuticals, pharmaceutical actives, biologics, amino acids, proteins, peptides, nucleotides, nucleic acids, additives, foods and food ingredients and analogs, homologs and first order derivatives thereof.
31. A composition of claim 30, wherein the percentage of particles greater than 1 μm on a particle volume basis is a percentage selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 95%, and 100%.

32. A composition of claim 30, wherein the percentage of particles greater than 2 μm on a particle volume basis is a percentage selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 95% and 100%.
33. A composition of any one of claims 30 to 32, wherein the median particle size has been reduced by a factor selected from the group consisting of: less than 5%, less than 10%, less than 20%, less than 30%, less than 40%, less than 50%, less than 60%, less than 70%, less than 80%, less than 90 %, less than 95% and less than 99%.
34. A composition of any one of claims 30 to 33, wherein the median particle size falls within the range selected from the group consisting of: 1-150 μm , 1-100 μm , 1-50 μm , 1-20 μm , 1-10 μm , 1-7.5 μm , 1-5 μm 1-2 μm , 2-1000 μm , 2-500 μm , 2-300 μm , 2-200 μm , 2-150 μm , 2-100 μm , 2-50 μm , 2-20 μm , 2-10 μm , 2-7.5 μm and 2-5 μm .
35. A composition of any one of claims 30 to 34, wherein the crystallinity profile of the biologically active material is selected from the group consisting of: at least 50% of the biologically active material is crystalline, at least 60% of the biologically active material is crystalline, at least 70% of the biologically active material is crystalline, at least 75% of the biologically active material is crystalline, at least 85% of the biologically active material is crystalline, at least 90% of the biologically active material is crystalline, at least 95% of the biologically active material is crystalline and at least 98% of the biologically active material is crystalline.
36. The composition of any one of claims 30 to 35, wherein the crystallinity profile of the biologically active material is substantially equal to the crystallinity profile of the biologically active material before the material was subject to the method of any one of the preceding claims 1 to 25.
37. A composition of any one of claims 30 to 36, wherein the amorphous content of the biologically active material is selected from the group consisting of: less than 50% of the biologically active material is amorphous, less than 40% of the biologically active material is amorphous, less than 30% of the biologically active material is amorphous, less than 25% of the biologically active material

is amorphous, less than 15% of the biologically active material is amorphous, less than 10% of the biologically active material is amorphous, less than 5% of the biologically active material is amorphous and less than 2% of the biologically active material is amorphous.

38. A composition of any one of claims 30 to 37, wherein the biologically active material has had no significant increase in amorphous content following subjecting the material to the method of any one of the preceding claims.
39. The composition of any one of claims 30 to 38, wherein the facilitating agent is a surfactant.
40. The composition of any one of claims 30 to 39, wherein the surfactant is sodium lauryl sulphate.
41. A pharmaceutical composition comprising a biologically active material produced by the method of any one of claims 1 to 25 together with a pharmaceutically acceptable carrier.
42. Use of an effective amount of a pharmaceutical composition of claim 41 in the preparation of a medicament for the treatment of a medical disease, condition or disorder.
43. Use of a biologically active material produced by the method of any one of claims 1-25 in the manufacture of a medicament for the treatment of a human in need of such treatment.
44. A method for manufacturing a pharmaceutical composition of claim 41 comprising the step of combining a therapeutically effective amount of a biologically active material prepared by a method according to any one of the claims 1 to 25 together with a pharmaceutically acceptable carrier to produce a pharmaceutically acceptable dosage form.
45. A method for manufacturing a veterinary product comprising the step of combining a therapeutically effective amount of the biologically active material prepared by a method of any one of the claims of 1 to 25 together with an acceptable excipient to produce a dosage form acceptable for veterinary use.

46. A method for manufacturing an agricultural product comprising the step of combining an effective amount of the biologically active material prepared by a method of any one of the claims of 1 to 25 together with an acceptable excipient.