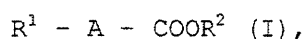

Process for reacting a carboxylic acid ester

5 The invention relates to a process for reacting a carboxylic acid ester of the formula (I)



10 wherein R^1 is selected from the group comprising hydrogen, $-CH_2OH$, $-CHO$, $-COOR^3$, $-CH_2SH$, $-CH_2OR^3$ and $-CH_2NH_2$,

15 wherein R^2 is selected from the group comprising hydrogen and alkyl, preferably methyl, ethyl and propyl,

wherein R^3 is selected from the group comprising hydrogen and alkyl, preferably methyl, ethyl and propyl,

20 and wherein A is a substituted, unsubstituted, linear, branched and/or cyclic alkylene, alkenylene, arylene or aralkylene radical having at least 4, more preferably 6, even more preferably 8, carbons,

25 by means of a cell, comprising the step of

a) contacting the cell with said carboxylic acid ester in an aqueous solution,

30 wherein the cell is a recombinant cell which has reduced activity of a polypeptide having SEQ ID NO 2 or a variant thereof over the wild-type cell, and to a cell suitable therefor, and to uses thereof.

Biotechnology is concerned with producing *inter alia* fine chemicals, employing various organisms that possess interesting synthesizing capabilities. Biotechnological processes have a number of advantages over conventional chemical processes. Firstly, they usually dispense entirely with harmful substances such as heavy metal-based catalysts and reaction conditions involving extreme pH, pressure and temperature values. Furthermore, the infrastructure required for the biotechnological process can often be set up with lower costs and safety measures. Selectivity and specificity of biotechnologically relevant enzymes often exceed those of chemical catalysts considerably, and formation of undesired secondary products which frequently are difficult to remove from the product and which would necessarily be produced in a synthesis using organic synthesis processes can thus be reduced. Finally, biotechnologically relevant organisms in many cases accept as reactants compounds like complex carbohydrates which may be derived from renewable raw materials. As a result, a producing enterprise may reduce its dependence on fossil raw materials such as petroleum and natural gas.

Establishing biotechnological processes, however, entails considerable difficulties which result in only very few substances being biotechnologically produced on an industrial scale these days, the main problem being that a cell with a desired synthesizing activity has not only the one enzyme responsible for said synthesizing activity but rather thousands of enzymes which co-exist in the same cell and compete with one another for substrates or even catalyse completely opposing reactions. Thus in the genome of *Escherichia coli* alone, approx. 80 polypeptides are encoded which have been identified by bioinformatics methods as hydrolases, i.e. enzymes which cleave particular bonds with consumption of a water molecule. Indeed, the conditions under which an enzyme is produced by the cell and the

reactions catalysed by said enzyme and the substrates involved in said reactions have been elucidated exhaustively only in a few cases. It is therefore in many cases not possible to specifically select an enzyme for catalysing a particular reaction.

Accordingly, when employing cells as biocatalysts rather than chemical or isolated biological catalysts, there is also always the risk of a product or intermediate produced by an enzyme equipped with a desired activity or even the original reactant being converted by another enzyme into an unwanted secondary product. Whether this will happen and which of the numerous enzymes will have said unwanted activity in this case is impossible to predict in spite of technical advances in the field of bioinformatics.

It is not unlikely, especially with chemically reactive substances desired in the industry as reactive reactants for producing more complex products, that said substances react inside the cell with essential components of the organism and thus have a toxic action. If this is the case, the ability of the organism to grow and synthesize will be impaired, and ultimately the cell will die, without the developer being able to immediately recognize said toxicity. Likewise, the organism that will tolerate a chemically reactive substance as well as the tolerated concentration of the latter cannot be predicted.

In processes involving a plurality of reactions catalysed in each case by an enzyme, the complexity of the system makes the search for factors limiting yield or purity more difficult. If the product yield is too low, the reason for this may be the concentration of one of the enzymes present being too low, although that enzyme would not be known from among possible enzymes, that is to say owing to insufficient synthesizing capacity, the reactant is not reacted within the intended time frame or prior to degradation by competing

enzymes. Alternatively, it is quite possible that an enzyme, although detectably present as a polypeptide in the cell, does not have the folding essential to the activity in that particular cell or that a hitherto unknown cofactor which
5 is, however, essential to the activity is missing. Equally, as mentioned previously, the metabolic product may be toxic to the cell or broken down.

A person skilled in the art who would like to establish or
10 improve a biotechnological process is thus confronted with numerous possible starting points, but in most cases is not provided by the prior art with any specific and executable instruction as to which of these starting points he needs to start from in order to achieve the objective.

15 Carboxylic acid esters constitute a group of compounds that are in high demand in the industry and that, either themselves or by way of processed products, are used as pharmaceuticals, cosmetics, plastics and the like.

20 Processing, however, frequently requires not only an ester function, which has to be introduced into a precursor first, but also further derivatization being carried out on the ester without hydrolysing the ester function in the process
25 or subsequently. The latter is not a trivial accomplishment to be achieved, since many carboxylic acid esters tend to hydrolyse even in the absence of enzymes catalysing such reactions, in particular in aqueous solutions and at pH values greatly deviating from the neutral point.

30 An important possibility of further derivatizing carboxylic acid esters is oxidization of alkyl chains present therein. This first produces an alcohol which either is used as such or may be further oxidized to the aldehyde or ketone. The
35 aldehyde or ketone may be either reductively aminated or further oxidized to the carboxylic acid which in turn may be esterified again, if required.

This multiplicity of possible reactions, many of which are catalysed by endogenous enzymes, i.e. enzymes that are naturally present in an organism, indicates that the problem of uncontrolled secondary product formation or metabolizing is particularly serious for reacting carboxylic acid esters by means of biotechnological processes.

An example of a carboxylic acid ester that is in high demand in the industry and that is customarily prepared starting from hydrocarbons present in petroleum is methyl 12-amino-dodecanoate (ALSME [12-Aminolaurinsäuremethylester]). ALSME is an important starting material in the preparation of polymers, for example for producing nylon-based line systems. ALSME has previously been produced in a low-yield process, starting from fossil raw materials.

A promising new way of biotechnologically producing ALS or ALSME is described in WO 2009/077461. This involves methyl dodecanoate being oxidized by a monooxygenase in a first step, and reacting the resultant aldehyde by means of a transaminase to give ALSME. A disadvantage of this process is the production of secondary products, for example the dicarboxylic acid, which can be removed from the desired product, ALSME, only with difficulties. This reduces the yield and makes recycling of hydrophobic solvents and hydrophobic cation exchangers, which may be used according to PCT/EP2011/071491 for removing the product from the aqueous reaction mixture, more difficult, at the expense of efficient resource usage.

Against this background, it is an object of the invention to provide a biotechnological process for reacting carboxylic acid esters which is as efficient as possible with regard to yield, carbon balance and/or nitrogen balance and/or purity.

Another object of the invention is to provide a biotechnological process for reacting carboxylic acid esters to give aminated carboxylic acid esters, which is as efficient as possible with regard to yield, carbon balance and/or nitrogen balance, re-usability of agents used and/or purity of the product. In this connection, an efficient carbon balance and/or nitrogen balance preferably means that the proportion of the carbon and/or nitrogen fed in the form of suitable substrates to a cell for reacting a carboxylic acid ester is as high as possible in the desired final product, instead of being reacted to give products other than the desired ones, for example.

Another object of the invention is to improve the ability of a multi-phase reaction mixture to be worked up from the reaction of a carboxylic acid ester, particularly with regard to re-usability of hydrophobic solvents and liquid cation exchangers used for work-up, and with regard to phase formation and separation in a biphasic system comprising an aqueous phase in which reaction of the carboxylic acid ester takes place and an organic phase with organic solvents and/or liquid cation exchangers.

These and further objects are achieved by the subject matter of the present application and in particular also by the subject matter of the enclosed independent claims, with the dependent claims giving rise to embodiments.

In one example, the problem addressed by the disclosure is solved by a process for reacting a carboxylic acid ester of the formula (I)

$$R^1 - A - COOR^2 \quad (I),$$

wherein R^1 is selected from the group comprising hydrogen, $-CH_2OH$, $-CHO$, $-COOR^3$, $-CH_2SH$, $-CH_2OR^3$ and $-CH_2NH_2$,

5 wherein R^2 is selected from the group comprising hydrogen and alkyl, preferably methyl, ethyl and propyl,

wherein R^3 is selected from the group comprising hydrogen and alkyl, preferably methyl, ethyl and propyl,

10

and wherein A is a substituted, unsubstituted, linear, branched and/or cyclic alkylene, alkenylene, arylene or aralkylene radical having at least 4, more preferably 6, even more preferably 8, carbons,

15

by means of a cell, comprising the step of

a) contacting the cell with said carboxylic acid ester in an aqueous solution,

20

wherein the cell is a recombinant cell which has reduced activity of a polypeptide comprising SEQ ID NO 2 or a variant thereof over the wild-type cell.

25 In another example, the problem is solved by a process further comprising the step of

b) contacting the aqueous solution with a hydrophobic organic solution comprising a cation exchanger.

30

In another example, the problem addressed by the disclosure is solved by use of a knockout of a gene coding for a polypeptide comprising SEQ ID NO 2 or a variant thereof as part of the genetic make-up of a recombinant cell for
35 increasing production of a carboxylic acid ester of the formula (I)



wherein R^1 is selected from the group comprising
hydrogen, $-CH_2OH$, $-CHO$, $-COOR^3$, $-CH_2SH$, $-CH_2OR^3$ and
5 $-CH_2NH_2$,

wherein R^2 is selected from the group comprising alkyl,
preferably methyl, ethyl and propyl,

10 wherein R^3 is selected from the group comprising hydrogen
and alkyl, preferably methyl, ethyl and propyl,

and wherein A is a substituted, unsubstituted, linear,
branched and/or cyclic alkylene, alkenylene, arylene or
15 aralkylene radical having at least 4, more preferably 6,
even more preferably 8, carbons, in particular an
alkylene radical having at least four, more preferably
6, even more preferably 8, carbons,

20 over the corresponding wild-type cell.

In another example, the problem addressed by the disclosure
is solved by using a recombinant cell which has reduced
activity of a polypeptide having SEQ ID NO 2 or a variant
25 thereof over the wild-type cell for reacting a carboxylic
acid ester of the formula (I)



30 wherein R^1 is selected from the group comprising
hydrogen, $-CH_2OH$, $-CHO$, $-COOR^3$, $-CH_2SH$, $-CH_2OR^3$ and
 $-CH_2NH_2$,

wherein R^2 is selected from the group comprising hydrogen
35 and alkyl, preferably methyl, ethyl and propyl,

wherein R^3 is selected from the group comprising hydrogen and alkyl, preferably methyl, ethyl and propyl,

5 and wherein A is a substituted, unsubstituted, linear, branched and/or cyclic alkylene, alkenylene, arylene or aralkylene radical having at least 4, more preferably 6, even more preferably 8, carbons, preferably an alkylene radical having at least eight carbons.

10 In another example, the problem is solved by a process or a use, wherein A is a saturated alkylene radical, preferably an alkylene radical of the formula $-(CH_2)_n-$, where n is at least 4.

15 In another example, the problem is solved by a process, wherein R^1 is selected from the group comprising hydrogen, $-CH_2OH$, $-CHO$ and $-CH_2NH_2$.

In another example, the problem addressed by the disclosure
20 is solved by a cell expressing a recombinant alkane hydroxylase, wherein the activity of a polypeptide comprising SEQ ID NO 2 or a variant thereof is reduced over the wild-type cell.

25 In a preferred example, the problem is solved by a cell, wherein the alkane hydroxylase is an alkane hydroxylase from the group comprising AlkB monooxygenases and a cytochrome P450 monooxygenase of the CYP153 family.

30 In one aspect, the present disclosure refers to a process for transforming a carboxylic acid ester of the formula (I) into a second carboxylic acid ester:



35 where

R¹ is hydrogen, -CH₂OH, -CHO, -COOR³, -CH₂SH, -CH₂OR³ and -CH₂NH₂,

R² is an alkyl group,

R³ is hydrogen or an alkyl group, and

5 A is an unsubstituted, a linear, or an unsubstituted and linear alkylene radical having at least 10 carbons,

the process comprising:

10 contacting a recombinant cell with the carboxylic acid ester of the formula (I) in an aqueous solution to convert the carboxylic acid ester of the formula (I) to the second carboxylic acid ester,

15 wherein the recombinant cell has a reduced activity of a polypeptide comprising SEQ ID NO: 2 or a polypeptide having at least 70% homology to SEQ ID NO: 2 and having essentially the same enzymatic activity as the polypeptide comprising SEQ ID NO: 2 compared to an activity in a corresponding wild-type cell due to knock-out of a gene encoding the polypeptide comprising SEQ ID
20 NO: 2 or the polypeptide having at least 70% homology to SEQ ID NO: 2 and having essentially the same enzymatic activity as the polypeptide comprising SEQ ID NO: 2,

25 the recombinant cell comprises a polypeptide having at least 80% sequence identity to consecutive 230 amino acids of SEQ ID NO: 15 and having essentially the same enzymatic activity as *Pseudomonas putida* AlkL, where the polypeptide is optionally overexpressed,

30 the recombinant cell has been transferred to express a transaminase, *Pseudomonas putida* AlkBGT, and an alanine dehydrogenase,

the recombinant cell has a reduced activity of at least one enzyme catalyzing any of reactions of fatty acid β -oxidation compared to an activity in the wild-type cell due to knock-out, and

35 the at least one enzyme catalyzing any of reactions of fatty acid β -oxidation is selected from the group consisting of fatty acid-CoA ligase, acyl-CoA

dehydrogenase, 2,4-dienoyl-CoA reductase, enoyl-CoA hydratase, 3-ketoacyl-CoA thiolase, and a fatty acid importer.

5 In one example, the problem addressed by the disclosure is solved by a reaction mixture comprising the cell according to the examples in aqueous solution and a carboxylic acid ester of the formula (I)

10
$$R^1 - A - COOR^2 \text{ (I),}$$

wherein R^1 is selected from the group comprising hydrogen, $-CH_2OH$, $-CHO$, $-COOR^3$, $-CH_2SH$, $-CH_2OR^3$ and $-CH_2NH_2$,

5 wherein R^2 is selected from the group comprising alkyl, preferably methyl, ethyl and propyl,

wherein R^3 is selected from the group comprising hydrogen and alkyl, preferably methyl, ethyl and propyl,

10 wherein A is a substituted, unsubstituted, linear, branched and/or cyclic alkylene, alkenylene, arylene or aralkylene radical having at least 4, more preferably 6, even more preferably 8, carbons.

15 In another example, the problem is solved by the reaction mixture,

wherein A is an alkylene radical having at least four carbons, in particular a saturated alkylene radical, 20 preferably an alkylene radical of the formula $-(CH_2)_n-$, where n is at least 4,

wherein R^1 is preferably selected from the group comprising hydrogen, $-CH_2OH$, $-CHO$ and $-CH_2NH_2$.

25

In another example, the problem is solved by a reaction mixture further comprising a hydrophobic organic solution containing a cation exchanger.

30 In a preferred example, the problem is solved by a process, a cell or a use, wherein the cell furthermore expresses a transaminase.

In another preferred example, the problem is solved by a 35 process, a cell or a use, wherein the cell furthermore expresses an alanine dehydrogenase.

In another preferred example, the problem is solved by a process, a cell or a use, wherein the cell furthermore has a protein of the AlkL family.

5 In another preferred example, the problem is solved by a process, a cell or a use, wherein the cell has an activity, which is reduced over its wild type, of at least one enzyme catalysing any of the reactions of fatty acid β -oxidation, wherein the enzyme is preferably one from the group
10 comprising fatty acid-CoA ligase, acyl-CoA dehydrogenase, 2,4-dienoyl-CoA reductase, enoyl-CoA hydratase and 3-ketoacyl-CoA thiolase, a fatty acid importer or variants thereof, particularly preferably FadL or a variant thereof.

15 In another preferred example, the problem is solved by a process, a cell or a use, wherein the cell has and/or overexpresses at least one enzyme from the group comprising alkane hydroxylase, alcohol dehydrogenase, transaminase, alanine dehydrogenase and protein of the AlkL family in
20 recombinant form.

In another preferred example, the problem is solved by a process, a cell or a use, wherein the activity of a polypeptide comprising SEQ ID NO 2 or a variant thereof is
25 reduced over the wild-type cell due to knockout of a gene coding for a polypeptide comprising SEQ ID NO 2 or a variant thereof.

In another aspect, the present disclosure refers to a method
30 for producing a carboxylic acid ester of the formula (I') from a substrate carboxylic acid ester:



where

35

R¹ is hydrogen, -CH₂OH, -CHO, -COOR³, -CH₂SH, -CH₂OR³ or -CH₂NH₂,

R² is an alkyl group,

R³ is hydrogen or an alkyl group, and

5 A is an unsubstituted, a linear, or an unsubstituted and linear alkylene radical having at least 10 carbons, the method comprising:

contacting a recombinant cell with the substrate
carboxylic acid ester in an aqueous solution to convert
10 the substrate carboxylic acid ester to the carboxylic acid ester of the formula (I'),

wherein the recombinant cell comprises a knockout of a gene encoding a polypeptide comprising SEQ ID NO: 2 or a polypeptide having at least 70% homology to SEQ ID NO:2
15 and having essentially the same enzymatic activity as the polypeptide comprising SEQ ID NO:2,

the recombinant cell comprises a polypeptide having at least 80% sequence identity to consecutive 230 amino acids of SEQ ID NO: 15 and having essentially the same
20 enzymatic activity as *Pseudomonas putida* AlkL, where the polypeptide is optionally overexpressed,

the recombinant cell has been transformed to express a transaminase, *Pseudomonas putida* AlkBGT, and an alanine dehydrogenase,

25 the production of the carboxylic acid ester of the formula (I') is increased over a corresponding wild-type cell of the recombinant cell,

the recombinant cell has a reduced activity of at least one enzyme catalyzing any of reactions of fatty acid β-oxidation compared to an activity in the wild-type cell
30 due to knockout, and

the at least one enzyme catalyzing any of reactions of fatty acid β-oxidation is selected from the group consisting of fatty acid-CoA ligase, acyl-CoA dehydrogenase, 2,4-dienoyl-CoA reductase, enoyl-CoA hydratase, 3-ketoacyl-CoA thiolase, and a fatty acid
35 importer.

The invention is based on the inventors surprisingly finding that a recombinant cell which has reduced activity of a polypeptide comprising SEQ ID NO 2 in particular or a variant thereof over the wild-type cell is suitable for
5 reacting carboxylic acid esters in that yield, carbon balance and/or nitrogen balance and purity of the products produced therefrom are surprisingly higher than with a cell that has the same activity as the wild-type cell with regard

to the polypeptide comprising SEQ ID NO 2. This is true, for example, for reactions in which the carboxylic acid ester is oxidized by using at least one alkane hydroxylase and optionally further enzymes.

5

The invention is based on the inventors furthermore surprisingly finding that a recombinant cell which has reduced activity of a polypeptide comprising SEQ ID NO 2 or a variant thereof over the wild-type cell is suitable for
10 reacting carboxylic acid esters particularly in that organic solvents and liquid cation exchangers used for working up the reaction product from the reaction of the carboxylic acid ester may be recovered particularly efficiently and/or frequently by recycling to be used again, and in that
15 removal of the aqueous reaction mixture from a hydrophobic solution comprising organic solvent and/or liquid cation exchanger is improved.

The invention relates to a process for reacting a carboxylic
20 acid ester by means of a cell, wherein the reaction may be any chemical reaction which utilizes the carboxylic acid ester of interest as reactant and in which hydrolysis or preliminary hydrolysis of the carboxylic acid ester or of a compound derived therefrom could impair the yield, carbon
25 balance and/or nitrogen balance and/or purity of the product to be produced therefrom. The cell of the invention and the process of the invention are particularly suitable for reactions in which a different chemical function than the carboxylate group is reacted, for example a terminal alkyl
30 group, and in which the aim is to retain the carboxylate group. According to the invention, however, it is also possible to carry out according to the invention reactions such as transesterifications of the carboxylate group, in which the aim is to avoid a preliminary and unspecific
35 reaction of the carboxylate group. The teaching according to the invention is also suitable for reactions in which a compound containing the carboxylate group is not itself the

reactant to be reacted but which require only said compound to be present and stable over a relatively long period, for example if said compound is an inducer or activator of an enzyme having an activity which is essential to the process.

5

To carry out the invention, it is essential for the cell according to the invention or cell used in a process according to the invention to be a recombinant cell that has reduced activity of a polypeptide comprising SEQ ID NO 2 or
10 a variant thereof over the wild-type cell. Said sequence codes for BioH, an enzyme known for its ability to convert, as part of the biosynthesis of biotin together with another enzyme, BioC, alanine and/or acetate into pimeloyl-CoA (Barker, D. F., and Campbell, A. M. (1980) *J. Bacteriol.*
15 143, 789-800). In *Escherichia coli*, BioH is encoded by a nucleotide sequence comprising SEQ ID NO 1. In a preferred embodiment, the activity of the polypeptide comprising SEQ ID NO 2 is reduced due to knockout, for example partial deletion, or other measures of reducing expression of a
20 nucleotide sequence comprising SEQ ID NO 1 or a variant thereof.

With the development of modern genetic, microbiological and molecular-biological methods, the skilled worker has
25 numerous tools at his disposal which he may use to routinely measure and influence the activity of polypeptides present in living cells. The activity of an enzyme which is in the form of a suspension, a pellet, or may have been taken in processed form from a cell culture may be determined by
30 using and analyzing enzymatic standard assays, as described in textbooks, for example Cornish-Bowden, 1995. An assay for determining the activity of the polypeptide comprising SEQ ID NO 1 or a variant thereof is described in X. Xie et al. (2007) *Metabolic Engineering* 9; 379-386.

35

Routinely applicable processes for reducing the activity of an enzyme in a cell, for example by random mutagenesis of

cells by exposure to radioactive radiation followed by concentrating or screening the mutants, by site-directed introduction of point mutations or by interruption of the reading frame or deletion of a part of the reading frame of
5 a gene chromosomally integrated into a cell, which gene codes for an active enzyme, have also been described in the prior art, for example in Maniatis et al (1989) or in Fuchs & Schlegl (2007), and may routinely be performed by a person skilled in the art. It is also possible to reduce activity
10 on the basis of RNA interference (Tuschl, 2001) or using specific inhibitors. In a preferred embodiment, the wording "wherein the cell" has, "over its wild type, reduced activity" of a polypeptide, as used herein, means that the activity of the polypeptide in the modified cell is reduced
15 compared to the activity of the same enzyme in a wild-type cell. In a preferred embodiment, the relative reduction is, in the order of increasing preference, 5, 10, 20, 40, 50, 75, 90, 95, 99 or more, per cent of the activity. In a particularly preferred embodiment, activity of the enzyme is
20 not detectable anymore over the background.

Particular preference is given to reducing the activity of the polypeptide comprising SEQ ID NO 2 or a variant thereof by a knockout. In a preferred embodiment, the term
25 "knockout", as used herein, means any measure that reduces the activity of the polypeptide comprising SEQ ID NO 2 or a variant thereof permanently and irreversibly, in particular also in progeny of corresponding cells, preferably by interrupting the reading frame of the sequence coding for
30 SEQ ID NO 2 or a variant thereof, by deleting at least part of the sequence coding for SEQ ID NO 2 or a variant thereof, which causes a loss of the enzyme activity of the encoded polypeptide but does not interrupt the reading frame, or by mutating a nucleotide sequence essential to expression, for
35 example a promoter, a ribosome binding site or the like. Measures of preparing cells having reduced activities of specific polypeptides are routine procedures available to a

person skilled in the art and are sufficiently well described in the prior art, for example in Kamionka et al. (2005) Appl Environ Microbiol. 2005 February; 71(2): 728-733, Geng et al. (2009), Journal of Biomedicine and
5 Biotechnology Volume 2009, Article ID 646380, doi:10.1155/2009/646380, and Murphy (2009) Methods Mol Biol. 2011;765:27-42. Also suitable are commercially available kits, for example the TargeTron™ Gene Knockout System from Sigma Aldrich.

10

The invention relates to reacting a carboxylic acid ester of the formula (I)



15

wherein R^1 is selected from the group comprising hydrogen, $-CH_2OH$, $-CHO$, $-COOR^3$, $-CH_2SH$, $-CH_2OR^3$ and $-CH_2NH_2$, wherein R^2 is selected from the group comprising alkyl, preferably methyl, ethyl and propyl, wherein R^3 is selected from the
20 group comprising hydrogen and alkyl, preferably methyl, ethyl and propyl, and wherein A is a substituted, unsubstituted, linear, branched and/or cyclic alkylene, alkenylene, arylene or aralkylene radical having at least 4, more preferably 6, even more preferably 8, carbons. The
25 alkylene radical A may be any, in particular linear, branched or cyclic, alkylene radical, provided that it comprises at least four carbons. In a particularly preferred embodiment, A is an alkylene chain of the formula $-(CH_2)_n-$, where n may be 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16,
30 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29 or 30. In a most preferred embodiment, A represents an alkylene radical of the formula $-(CH_2)_n-$, where n is from 4 to 24, more preferably 4 to 22, and most preferably 4 to 10, R_1 is hydrogen, $-CH_2OH$, $-CHO$, or $-COOH$, most preferably hydrogen,
35 and R^2 is methyl or ethyl.

In connection with reactants of the carboxylic acid esters reacted according to the invention and all other compounds described herein, a compound described by structural features, for example a chemical formula, as a rule equally
5 denotes protonated and deprotonated or other dissociated compounds. For example, the term "acetate" is understood as meaning equally the protonated form, i. e. acetic acid, and the dissociated form, i. e. $\text{CH}_3\text{-COO}^-$.

- 10 The cell to be employed according to the invention is preferably a whole cell catalyst in the form of a metabolically active cell which has an enzymatic activity required for reacting the carboxylic acid ester, particularly preferably by expressing a recombinant enzyme.
- 15 In a further preferred embodiment, the cell is a lysate, an extract or another preparation of the cell, which has at least one enzymatic activity.

With regard to the choice of organism, the cell utilizable
20 according to the invention is not subject to any restrictions, as long as it is cultivable, stable and accessible to processes for attenuating enzyme activities, for example knockouts. Thus it may equally be a prokaryotic or eukaryotic cell. In the case of a eukaryotic cell,
25 particular preference is given to unicellular eukaryotes, particularly yeasts such as *Saccharomyces cerevisiae*, *Candida tropicalis*, *Candida albicans* and *Pichia pastoris*. In the case of prokaryotic cells, it may be, for example, a bacterium selected from the group comprising *Magnetococcus*,
30 *Mariprofundus*, *Acetobacter*, *Acetobacterium*, *Acidiphilium*, *Afipia*, *Ahrensia*, *Asticcacaulis*, *Aurantimonas*, *Azorhizobium*, *Azospirillum*, *Bacillus*, *Bartonella*, *tribocorum*, *Beijerinckia*, *Bradyrhizobium*, *Brevundimonas*, *subvibrioides*, *Brucella*, *Caulobacter*, *Chelativorans*, *Citreicella*,
35 *Citromicrobium*, *Clostridium*, *Corynebacterium*, *Dinoroseobacter*, *Erythrobacter*, *Fulvimarina*, *Gluconacetobacter*, *Granulibacter*, *Hirschia*, *Hoeflea*,

- Hyphomicrobium, Hyphomonas, Ketogulonicigenium, Labrenzia,
 Loktanella, Magnetospirillum, Maricaulis, Maritimibacter,
 Mesorhizobium, Methylobacterium, Methylocystis,
 Methylosinus, Nitrobacter, Novosphingobium, Oceanibulbus,
 5 Oceanicaulis, Oceanicola, Ochrobactrum, Octadecabacter,
 Oligotropha, Paracoccus, Parvibaculum, Parvularcula,
 Pelagibaca, Phaeobacter, Phenylobacterium, Polymorphum,
 Pseudovibrio, Rhodobacter, Rhodomicrobium, Rhodopseudomonas,
 Rhodospirillum, Roseibium, Roseobacter, Roseomonas,
 10 Roseovarius, Ruegeria, Sagittula, Silicibacter, Sphingobium,
 Sphingomonas, Sphingopyxis, Starkeya, Sulfitobacter,
 Thalassiobium, Xanthobacter, Zymomonas, Agrobacterium,
 Rhizobium, Sinorhizobium, Anaplasma, Ehrlichia,
 Neorickettsia, Orientia, Rickettsia, Wolbachia, Bordetella,
 15 Burkholderia, Cupriavidus, Taiwanensis, Lautropia,
 Limnobacter, Polynucleobacter, Ralstonia, Chromobacterium,
 Eikenella, corrodens, Basfia, Kingella, Laribacter,
 Lutiella, Neisseria, Simonsiella, Achromobacter, Acidovorax,
 Alicyclophilus, Aromatoleum, Azoarcus, Comamonas,
 20 Dechloromonas, Delftia, Gallionella, Herbaspirillum,
 Herminiimonas, Hylemonella, Janthinobacterium, Leptothrix,
 Methylibium, Methylobacillus, Methylophilales,
 Methyloversatilis, Methylovorus, Nitrosomonas, Nitrosospira,
 Oxalobacter, Parasutterella, Polaromonas, Polaromonas,
 25 Pusillimonas, Rhodoferax, Rubrivivax, Sideroxydans,
 Sutterella, wadsworthensis, Taylorella, Thauera,
 Thiobacillus, Thiomonas, Variovorax, Verminephrobacter,
 Anaeromyxobacter, Bdellovibrio, bacteriovorus, Bilophila,
 Desulfarculus, Desulfatibacillum, Desulfobacca,
 30 Desulfobacterium, Desulfobulbus, Desulfococcus,
 Desulfohalobium, Desulfitobacterium, Desulfomicrobium,
 Desulfonatronospira, Desulfotalea, Desulfovibrio,
 Desulfuromonas, Geobacter, Haliangium, Hippea, Lawsonia,
 Myxococcus, Pelobacter, Plesiocystis, Sorangium,
 35 Stigmatella, Syntrophobacter, Syntrophus, Arcobacter,
 Caminibacter, Campylobacter, Helicobacter, Nitratifractor,
 Nitratiruptor, Sulfuricurvum, Sulfurimonas,

Sulfurospirillum, Sulfurovum, Wolinella, Buchnera,
Blochmannia, Hamiltonella, Regiella, Riesia, Citrobacter,
Cronobacter, Dickeya, Edwardsiella, Enterobacter, Erwinia,
Escherichia, Klebsiella, Pantoea, Pectobacterium, Proteus,
5 Providencia, Rahnella, Salmonella, Serratia, Shigella,
Sodalis, Wigglesworthia, Glossina, Xenorhabdus, Yersinia,
Acidithiobacillus, Acinetobacter, Aeromonas, Alcanivorax,
Alkalilimnicola, Allochromatium, Alteromonadales,
Alteromonas, Baumannia, Beggiatoa, Bermanella, Carsonella,
10 Ruthia, Vesicomysocius, Cardiobacterium, Chromohalobacter,
Colwellia, Congregibacter, Coxiella, Dichelobacter,
Endoriftia, Enhydrobacter, Ferrimonas, Francisella,
Glaciecola, Hahella, Halomonas, Halorhodospira,
Halothiobacillus, Idiomarina, Kangiella, Legionella,
15 Marinobacter, Marinomonas, Methylobacter, Methylococcus,
Methylomicrobium, Methylophaga, Moraxella, Moritella,
Neptuniibacter, Nitrococcus, Pseudoalteromonas,
Psychrobacter, Psychromonas, Reinekea, Rickettsiella,
Saccharophagus, Shewanella, Succinatimonas, Teredinibacter,
20 Thioalkalimicrobium, Thioalkalivibrio, Thiomicrospira,
Tolumonas, Vibrionales, Actinobacillus, Aggregatibacter,
Gallibacterium, Haemophilus, Histophilus, Mannheimia,
Pasteurella, Azotobacter, Cellvibrio, Pseudomonas,
Aliivibrio, Grimontia, Photobacterium, Photobacterium,
25 Vibrio, Pseudoxanthomonas, Stenotrophomonas, Xanthomonas,
Xylella, Borrelia, Brachyspira, Leptospira, Spirochaeta,
Treponema, Hodgkinia, Puniceispirillum, Liberibacter,
Pelagibacter, Odysseella, Accumulibacter, in particular B.
subtilis, B. megaterium, C. glutamicum, E. coli, Pseudomonas
30 sp., Pseudomonas fluorescens, Pseudomonas putida,
Pseudomonas stutzeri, Acinetobacter sp., Burkholderia sp.,
Burkholderia thailandensis, cyanobacteria, Klebsiella sp.,
Klebsiella oxytoca, Salmonella sp., Rhizobium sp. and
Rhizobium meliloti. In a particularly preferred embodiment,
35 it is an enterobacterium, most preferably Escherichia coli.

The process of the invention requires contacting the cell with the carboxylic acid ester in an aqueous solution. In a preferred embodiment, the term "contacting", as used herein, means that the cell according to the invention comes into
5 direct contact with the particular agent, for example the carboxylic acid ester or a liquid cation exchanger, in particular without any physical barriers such as porous membranes or the like in between. In the simplest case, contacting comprises adding the agent, for example the
10 carboxylic acid ester or the liquid cation exchanger, to an aqueous solution containing the cell.

Aqueous solutions that may be used are any water-based solutions suitable for maintaining or culturing the cell
15 and/or its activity that is required for reacting the carboxylic acid ester. These include equally culture media for microorganisms, including complete media such as LB media, minimal media such as M9 media, and selective media, for example those containing a high salt concentration and
20 therefore enable only halophilic or at least halotolerant organisms to grow. A particularly preferred medium is a minimal medium that contains very few components which can readily be removed from the product of the reaction of the carboxylic acid ester, in order to facilitate work-up of
25 said product.

If the product of the intended reaction is sufficiently hydrophobic and has an appropriate charge, it is possible to extract it in a step b) by contacting the aqueous solution
30 with a hydrophobic organic solution comprising a cation exchanger. Appropriate procedures are described in the international application PCT/EP2011/071491 or in the European application EP12181153.3. Briefly, the aqueous solution may be contacted with a hydrophobic solution
35 comprising a fatty acid ester as solvent and a fatty acid, preferably an unsaturated fatty acid, during or after the reaction.

The requirements of the cell, the reaction and necessary enzymes must be taken into account when setting the temperature and conditions in step a). The temperature requirements of various biotechnologically important cells can be found in textbooks of microbiology and molecular biology, for example Fuchs/Schlegel, Allgemeine Mikrobiologie [General Microbiology], 2008, or may be determined by culturing experiments within routine procedures. In a preferred embodiment, the pH of the aqueous culture medium at the time of contacting is between 4 and 9, more preferably between 4.5 and 8.5, most preferably between 6.5 and 7.5. In another preferred embodiment, the temperature is between 0 and 45 °C, more preferably between 15 and 40 °C, most preferably between 20 and 37 °C.

Preference is given to using for the process of the invention a cell expressing a recombinant alkane hydroxylase, wherein the activity of a polypeptide comprising SEQ ID NO 2 or a variant thereof is reduced over the wild-type cell. In a preferred embodiment, the alkane hydroxylase is a cytochrome P450 monooxygenase of the CYP153 family. In a preferred embodiment, the term "cytochrome P450 monooxygenase of the CYP153 family" means a cytosolic oxidase which is part of a 3-component system comprising furthermore a ferredoxin and a ferredoxin reductase, with an alkane-binding site and the ability to hydroxylate alkanes. In a particularly preferred embodiment, the enzyme has at least 80, preferably 90, most preferably 95 or 99, per cent sequence identity with cytochrome P450 monooxygenase of the CYP153 family from *Alcanivorax borkumensis* SK2 (database code YP_691921), or the enzyme has a polypeptide sequence which has at least 80, preferably 90, most preferably 95 or 99, per cent sequence identity with cytochrome P450 monooxygenase of the CYP153 family from *Alcanivorax borkumensis* SK2 (database code YP_691921), and additionally has alkane hydroxylase activity. The database codes stated

here, as throughout the application, refer to the NCB (National Center for Biotechnology Information, Bethesda, USA)I databases, more precisely the version available online on 8 November 2012. In a preferred embodiment, the term
5 "alkane hydroxylase activity", as used herein, means the ability to catalyse the hydroxylation of alkanes or unsubstituted linear alkyl radicals comprising at least six, preferably twelve, carbon radicals. In another preferred embodiment, the term "cytochrome P450 monooxygenase of the
10 CYP153 family" means a non-membrane-bound oxidase which comprises a binding site for alkanes, unsubstituted linear alkyl radicals comprising at least five, preferably twelve, carbon radicals or monohydroxylated alkanes, and the polypeptide chain of which comprises the motif
15 LL(I/L)(V/I)GGNDTTRN. In a preferred embodiment, a "cytochrome P450 monooxygenase of the CYP153 family", as used herein, is a cytochrome P450 monooxygenase of the CYP153 family from *Alcanivorax borkumensis* SK2 (database code YP_691921) or a variant which preferably has alkane
20 hydroxylase activity.

The enzymes used according to the invention are preferably recombinant enzymes. In a preferred embodiment, the term "recombinant", as used herein, means that the corresponding
25 nucleic acid molecule is not present in the natural cell and/or was produced using genetic engineering methods. In a preferred embodiment, a protein is said to be recombinant if the corresponding polypeptide is encoded by a recombinant nucleic acid. In a preferred embodiment, a recombinant cell,
30 as used herein, means a cell which has at least one recombinant nucleic acid or one recombinant polypeptide. A person skilled in the art is familiar with processes suitable for producing recombinant molecules or cells, for example those described in Sambrook/Fritsch/Maniatis (1989):
35 Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, 2nd edition. Recombinant enzymes are preferably overexpressed, for example by using pET- or pGEX-

vector systems which are known to a person skilled in the art.

To supply cytochrome P450 monooxygenase of the CYP153 family
5 with electrons from the reducing agent, preferably NADH, in
an optimal way, preference is given to the cell expressing
said monooxygenase together with ferredoxin reductase and
ferredoxin, both of which interact functionally with said
monooxygenase. The polypeptides may be isolated or, when
10 using a whole cell catalyst, co-expressed polypeptides or
polypeptides fused N- or C-terminally to the cytochrome P450
monooxygenase of the CYP153 family. A person skilled in the
art can readily determine whether a ferredoxin reductase or
a ferredoxin interacts functionally with a given cytochrome
15 P450 monooxygenase of the CYP153 family by whether the
reducing agent is oxidized in the presence of an alkane
substrate and the three polypeptides. Alternatively, the
enzyme assay described by Scheps, D., Malca, H., Hoffmann,
B., Nestl, B. M, and Hauer, B. (2011) *Org. Biomol. Chem.*, 9,
20 6727, may be used, which shows a distinct increase in the
reaction rate in the case of functionally interacting
polypeptides. In a particularly preferred embodiment,
cytochrome P450 monooxygenase of the CYP153 family,
ferredoxin and ferredoxin reductase are from the same
25 organism. In a particularly preferred embodiment, ferredoxin
reductase is that from *Alcanivorax borkumensis* SK2 (database
code YP_691923) or is a variant thereof, ferredoxin is that
from *Alcanivorax borkumensis* SK2 (database code YP_691920)
or is a variant thereof, and cytochrome P450 monooxygenase
30 of the CYP153 family is that from *Alcanivorax borkumensis*
SK2 (database code YP_691921) or is a variant thereof.

In another preferred embodiment, the alkane hydroxylase is
an AlkB monooxygenase. AlkB is an oxidoreductase first known
35 from the *Pseudomonas putida* Gp01 AlkBGT system, which is
dependent on another two polypeptides, AlkG and AlkT. AlkT
is characterized as FAD-dependent rubredoxin reductase that

passes on electrons from NADH to AlkG. AlkG is a rubredoxin, an iron-containing redox protein that acts as a direct electron donor for AlkB. In a preferred embodiment, the term "AlkB monooxygenase" means a polypeptide with a sequence
5 homology of at least, in the order of increasing preference, 75, 80, 85, 90, 92, 94, 96, 98 or 99 % to the sequence of *Pseudomonas putida* Gp1 AlkB (database code: CAB54050.1; this database code, like all the others used in the application, is from the prior art, namely from the NCBI
10 database, more precisely the release available online on 15 October 2012), which polypeptide is capable of oxidizing alkanes. In a particularly preferred embodiment, the AlkB monooxygenase is an alkane-oxidizing oxidoreductase which functionally acts together with the *Pseudomonas putida* Gp1
15 AlkG (CAB54052.1) and AlkT (CAB54063.1) polypeptides. For optimal supply of AlkB alkane hydroxylase with electrons, preference is given to the cell expressing the monooxygenase together with auxiliary proteins that functionally interact with it, preferably AlkG and/or AlkT or respective variants
20 thereof, which, in a particularly preferred embodiment, are again *Pseudomonas putida* Gp1 AlkG (CAB54052.1) and AlkT (CAB54063.1) polypeptides.

The ability of the cell of the invention or the cell used in
25 the process of the invention to oxidize substrates may be enhanced by said cell expressing an alcohol dehydrogenase as an alternative to or additionally to the alkane hydroxylase. In a preferred embodiment, the term "alcohol dehydrogenase", as used herein, means an enzyme that oxidizes an aldehyde or
30 ketone to the corresponding primary or secondary alcohol. Examples include the alcohol dehydrogenases of *Ralstonia eutropha* (ACB78191.1), *Lactobacillus brevis* (YP_795183.1), *Lactobacillus kefir* (ACF95832.1), from horse liver, of *Paracoccus pantotrophus* (ACB78182.1) and *Sphingobium*
35 *yanoikuyae* (EU427523.1), and also the respective variants thereof.

The use of a whole cell catalyst may give rise to the problem of a substrate having to be contacted with an enzyme located intracellularly for the desired reaction to occur. In the case of long-chain alkanes and derivatives thereof, preference is given to the whole cell catalyst having a polypeptide of the AlkL family. In a preferred embodiment, a "polypeptide of the AlkL family", as used herein, is a polypeptide which, over a length of 230 consecutive amino acids, has at least 80, preferably 90, even more preferably 90% sequence identity to *Pseudomonas putida* AlkL (database code CAB69081) or a variant of *Pseudomonas putida* AlkL and preferably the ability to support importing long-chain alkanes into the interior of a cell. In another embodiment, a "polypeptide of the AlkL family", as used herein, is a polypeptide located in the outer membrane of a Gram-negative bacterium, which has the sequence motif DXWAPAXQ(V/A)GXR, where X is a proteinogenic amino acid, and preferably additionally is *Pseudomonas putida* AlkL (database code CAB69081) or a variant thereof. Exemplary members of the AlkL family include AlkL from *Pseudomonas putida* (database code CAB69081), *Marinobacter aquaeolei* VT8 (database code YP_957722), *Oceanicaulis alexandrii* HTCC2633 (database code ZP_00953584), *Marinobacter manganoxydans* MnI7-9 (database code ZP_09158756), *Caulobacter* sp. K31 (database code YP_001672217), *Pseudomonas oleovorans* (database code Q00595), and variants thereof.

The teaching of the present invention may be performed not only by using macromolecules with the exact amino acid or nucleic acid sequence to which reference is made herein, or not only by using a cell having an activity of a polypeptide with the exact amino acid sequence to which reference is made herein, which activity is reduced relatively to the particular wild type, but also by using a variant of such macromolecules or a cell having an activity of a variant of said polypeptide, which may be obtained by deletion, addition or substitution of one or more than one amino acid

or nucleic acid, which activity is reduced relatively to the particular wild type of the particular cell. In a preferred embodiment, the term "variant" of a nucleic acid sequence or an amino acid sequence, used synonymously and interchangeably with the term "homologue" hereinbelow, means, as used herein, a different nucleic acid or amino acid sequence which has a homology, used synonymously with identity herein, of 70, 75, 80, 85, 90, 92, 94, 96, 98, 99 or more, per cent with regard to the corresponding original wild-type nucleic acid or amino acid sequence, wherein preferably amino acids other than those forming the catalytically active site or those essential to structure or folding have been deleted or substituted, or else merely conservative substitutions have taken place, for example glutamate for aspartate or leucine for valine. The prior art describes algorithms which may be used for calculating the degree of homology of two sequences, for example Arthur Lesk (2008), Introduction to bioinformatics, 3rd edition. In another more preferred embodiment of the present invention, the variant of an amino acid sequence or nucleic acid sequence has, preferably in addition to the sequence homology mentioned above, essentially the same enzymatic activity of the wild-type molecule or the original molecule. For example, a variant of a polypeptide which has protease enzymatic activity has the same or essentially the same proteolytic activity as the polypeptide enzyme, i.e. the ability to catalyse hydrolysis of a peptide bond. In a particular embodiment, the term "essentially the same enzymatic activity" means an activity with regard to the substrates of the wild-type polypeptide that is markedly above background activity or/and differs by less than 3, more preferably 2, even more preferably an order of magnitude from the K_M and/or k_{cat} values of the wild-type polypeptide with regard to the same substrates. In another preferred embodiment, the term "variant" of a nucleic acid or amino acid sequence comprises at least one active part/or fragment of said nucleic acid or amino acid sequence. In

another preferred embodiment, the term "active part", as used herein, means an amino acid sequence or a nucleic acid sequence, which is shorter than the full-length amino acid sequence or which codes for a shorter than the full-length amino acid sequence, with the amino acid sequence or the encoded amino acid sequence with a shorter length than the wild-type amino acid sequence having essentially the same enzymatic activity as the wild-type polypeptide or a variant thereof, for example as protease. In a particular embodiment, the term "variant" of a nucleic acid comprises a nucleic acid, the complementary strand of which binds to the wild-type nucleic acid, preferably under stringent conditions. The stringency of the hybridization reaction can readily be determined by a person skilled in the art and generally depends on the length of the probe, the temperatures during washing and the salt concentration. Generally, longer probes require higher temperatures for hybridizing, whereas low temperatures are adequate for shorter probes. Whether hybridization takes place generally depends on the ability of the denatured DNA to anneal to complementary strands in its surroundings, specifically below the melting temperature. The stringency of hybridization reactions and corresponding conditions are described in more detail in F. M. Ausubel (1995), Current Protocols in Molecular Biology. John Wiley & Sons, Inc. Instructions for the identification of DNA sequences by means of hybridization are found by the person skilled in the art, *inter alia*, in the handbook "The DIG System Users Guide for Filter Hybridization" of Boehringer Mannheim GmbH (Mannheim, Germany, 1993) and in Liebl et al. (International Journal of Systematic Bacteriology 41: 255-260 (1991)). In a preferred embodiment, hybridization takes place under stringent conditions, that is to say only hybrids are formed in which probe and target sequence, i.e. the polynucleotides treated with the probe, are at least 70% identical. It is known that the stringency of the hybridization including the washing steps is influenced or determined by varying the

buffer composition, the temperature and the salt concentration. The hybridization reaction is in general carried out with a relatively low stringency in comparison to the washing steps (Hybaid Hybridisation Guide, Hybaid Limited, Teddington, UK, 1996). For the hybridization reaction, it is possible, for example, to employ a buffer corresponding to 5x SSC buffer at a temperature of about 50°C - 68°C. Here, probes can also hybridize with polynucleotides that have less than 70% identity to the sequence of the probe. Such hybrids are less stable and are removed by washing under stringent conditions. This can be achieved, for example, by lowering the salt concentration to 2x SSC and optionally subsequently 0.5x SSC (The DIG System User's Guide for Filter Hybridisation, Boehringer Mannheim, Mannheim, Germany, 1995), where a temperature of, in the order of increasing preference, about 50°C - 68°C, about 52°C - 68°C, about 54°C - 68°C, about 56°C - 68°C, about 58°C - 68°C, about 60°C - 68°C, about 62°C - 68°C, about 64°C - 68°C, about 66°C - 68°C is adjusted. Temperature ranges of about 64°C - 68°C or about 66°C - 68°C are preferred. It is optionally possible to lower the salt concentration to a concentration corresponding to 0.2x SSC or 0.1x SSC. By increasing the hybridization temperature stepwise in steps of about 1 - 2°C from 50°C to 68°C polynucleotide fragments may be isolated that have, for example, in the order of increasing preference, at least 70% or at least 80% or at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% identity to the sequence of the nucleic acid molecule employed. Further instructions for hybridization are obtainable on the market in the form of "kits" (e.g. DIG Easy Hyb from Roche Diagnostics GmbH, Mannheim, Germany, catalogue no. 1603558). In a preferred embodiment, the term "variant" of a nucleic acid, as used herein, comprises any nucleic acid sequence which codes for the same amino acid sequence as the original

nucleic acid or a variant of this amino acid sequence within the bounds of the degeneracy of the genetic code.

In a preferred embodiment, the cell used according to the invention has a reduced activity, over its wild type, of at least one enzyme catalysing any of the reactions of fatty acid β -oxidation, which enzyme is preferably from the group comprising fatty acid-CoA ligase, acyl-CoA dehydrogenase, 2,4-dienoyl-CoA reductase, enoyl-CoA hydratase and 3-ketoacyl-CoA thiolase, a fatty acid importer, or variants thereof, and particularly preferably is FadL or a variant thereof. β -Oxidation of fatty acids is a widespread metabolic pathway which allows prokaryotic and equally eukaryotic organisms to oxidize fatty acids and to render the chemical energy stored therein metabolically available. It starts in a wider sense with a fatty acid being taken up into the cell, in the case of *E. coli* through the transporter FadL which passes it through the outer and inner membranes of the Gram-negative bacterial cell, and the FadD gene product which releases the fatty acid by way of the CoA ester into the cytosol. There, if required by the conditions, the fatty acid is first oxidized at the β -position of the fatty acid-CoA ester by an acyl-CoA dehydrogenase, FadE in the case of *E. coli*. Alternatively, a similar molecule may also be formed from a diunsaturated fatty acid by reduction by means of a 2,4-dienoyl-CoA reductase, FadH in *E. coli*. A multifunctional enzyme, enoyl-CoA hydratase/3-hydroxyacyl-CoA dehydrogenase, FadB in *E. coli*, then catalyses hydratization with the formation of the secondary alcohol which is subsequently oxidized to the ketone. In the last step, a 3-ketoacyl-CoA thiolase, FadA in the case of *E. coli*, catalyses cleavage of ketoacyl-CoA, resulting in acetyl-CoA and a fatty acid-CoA ester that is shorter by two carbons compared to the starting molecule being released. If the latter is not acetyl-CoA as well, it may be fed again into the β -oxidation cycle and shortened with oxidation. FadR, a regulator of the Fad operon

comprising the genes required for the break-down of fatty acids, is also involved in regulating fatty acid β -oxidation, but would not catalyse a β -oxidation reaction. In a preferred embodiment, the term "enzyme catalysing any of the reactions of fatty acid β -oxidation" means any enzyme which interacts directly with the fatty acid substrate or a molecule produced therefrom on the pathway to acetyl-CoA, preferably recognizing it as substrate, and catalyses its conversion to a metabolite that is closer to acetyl-CoA on this catabolic pathway, preferably including the fatty acid importer which takes up the fatty acid into the cell. According to the definition above, these enzymes include, for example, acyl-CoA dehydrogenase, since it interacts with the fatty acid-CoA ester and catalyses its conversion to enoyl-CoA which is closer to acetyl-CoA than the fatty acid-CoA ester on the β -oxidation metabolic pathway. In a particularly preferred embodiment, the term "enzyme catalysing any of the reactions of fatty acid β -oxidation", as used herein, means any enzyme from the group comprising the gene products FadA, FadB, FadD, FadL and FadE from *E. coli* and/or their variants or homologues from other organisms. The gene products FadA, FadB, FadD, FadL and FadE from *E. coli* and also variants and homologues from numerous other biotechnologically utilizable organisms and their nucleic acid and polypeptide sequences are described in the prior art, for example FadA under accession number AP009048.1, FadB under accession number BAE77457.1, FadD under accession number BAA15609.1, FadE under accession number BAA77891.2 and FadL under accession number BAA16205.1.

In another preferred embodiment, the cell of the invention or cell used in the process of the invention expresses a transaminase. In a preferred embodiment, the term "transaminase", as used herein, means an enzyme catalysing the transfer of α -amino groups from a donor molecule, preferably an amino acid, to an acceptor molecule,

preferably an α -ketocarboxylic acid. For example, use may be made of a transaminase from the group comprising 3HMU_A, AAD41041.1, AAK15486.1, ABE03917.1, ADR60699.1, ADR61066.1, ADR62525.1, AEL07495.1, CAZ86955.1, EFW82310.1, EFW87681.1, 5 EGC99983.1, EGD03176.1, EGE58369.1, EGH06681.1, EGH08331.1, EGH24301.1, EGH32343.1, EGH46412.1, EGH55033.1, EGH62152.1, EGH67339.1, EGH70821.1, EGH71404.1, EGH78772.1, EGH85312.1, EGH97105.1, EGP57596.1, NP_102850.1, NP_106560.1, NP_248912.1, NP_248990.1, NP_354026.2, NP_421926.1, 10 NP_637699.1, NP_642792.1, NP_744329.1, NP_744732.1, NP_747283.1, NP_795039.1, NP_901695.1 (, XP_002943905.1, YP_001021095.1, YP_001059677.1, YP_001061726.1, YP_001066961.1, YP_001074671.1, YP_001120907.1, YP_001140117.1, YP_001170616.1, YP_001185848.1, 15 YP_001188121.1, YP_001233688.1, YP_001268866.1, YP_001270391.1, YP_001345703.1, YP_001412573.1, YP_001417624.1, YP_001526058.1, YP_001579295.1, YP_001581170.1, YP_001668026.1, YP_001669478.1, YP_001671460.1, YP_001685569.1, YP_001747156.1, 20 YP_001749732.1, YP_001765463.1, YP_001766294.1, YP_001790770.1, YP_001808775.1, YP_001809596.1, YP_001859758.1, YP_001888405.1, YP_001903233.1, YP_001977571.1, YP_002229759.1, YP_002231363.1, YP_002280472.1, YP_002297678.1, YP_002543874.1, 25 YP_002549011.1, YP_002796201.1, YP_002801960.1, YP_002875335.1, YP_002897523.1, YP_002912290.1, YP_002974935.1, YP_003060891.1, YP_003264235.1, YP_003552364.1, YP_003578319.1, YP_003591946.1, YP_003607814.1, YP_003641922.1, YP_003674025.1, 30 YP_003692877.1, YP_003755112.1, YP_003896973.1, YP_003907026.1, YP_003912421.1, YP_004086766.1, YP_004142571.1, YP_004147141.1, YP_004228105.1, YP_004278247.1, YP_004305252.1, YP_004356916.1, YP_004361407.1, YP_004378186.1, YP_004379856.1, 35 YP_004390782.1, YP_004472442.1, YP_004590892.1, YP_004612414.1, YP_004676537.1, YP_004693233.1, YP_004701580.1, YP_004701637.1, YP_004704442.1, YP_108931.1,

	YP_110490.1,	YP_168667.1,	YP_237931.1,	YP_260624.1,
	YP_262985.1,	YP_271307.1,	YP_276987.1,	YP_334171.1,
	YP_337172.1,	YP_350660.1,	YP_351134.1,	YP_364386.1,
	YP_366340.1,	YP_369710.1,	YP_370582.1,	YP_426342.1,
5	YP_440141.1,	YP_442361.1,	YP_468848.1,	YP_521636.1,
	YP_554363.1,	YP_608454.1,	YP_610700.1,	YP_614980.1,
	YP_622254.1,	YP_625753.1,	YP_680590.1,	YP_751687.1,
	YP_767071.1,	YP_774090.1,	YP_774932.1,	YP_788372.1,
	YP_858562.1,	YP_928515.1,	YP_983084.1,	YP_995622.1,
10	ZP_00948889.1,	ZP_00954344.1,	ZP_00959736.1,	ZP_00998881.1,
	ZP_01011725.1,	ZP_01037109.1,	ZP_01058030.1,	ZP_01076707.1,
	ZP_01103959.1,	ZP_01167926.1,	ZP_01224713.1,	ZP_01442907.1,
	ZP_01446892.1,	ZP_01550953.1,	ZP_01625518.1,	ZP_01745731.1,
	ZP_01750280.1,	ZP_01754305.1,	ZP_01763880.1,	ZP_01769626.1,
15	ZP_01865961.1,	ZP_01881393.1,	ZP_01901558.1,	ZP_02145337.1,
	ZP_02151268.1,	ZP_02152332.1,	ZP_02167267.1,	ZP_02190082.1,
	ZP_02242934.1,	ZP_02360937.1,	ZP_02367056.1,	ZP_02385477.1,
	ZP_02456487.1,	ZP_02883670.1,	ZP_03263915.1,	ZP_03263990.1,
	ZP_03400081.1,	ZP_03452573.1,	ZP_03456092.1,	ZP_03517291.1,
20	ZP_03529055.1,	ZP_03571515.1,	ZP_03572809.1,	ZP_03587785.1,
	ZP_03588560.1,	ZP_03697266.1,	ZP_03697962.1,	ZP_04521092.1,
	ZP_04590693.1,	ZP_04890914.1,	ZP_04891982.1,	ZP_04893793.1,
	ZP_04902131.1,	ZP_04905327.1,	ZP_04941068.1,	ZP_04944536.1,
	ZP_04945255.1,	ZP_04959332.1,	ZP_04964181.1,	ZP_05053721.1,
25	ZP_05063588.1,	ZP_05073059.1,	ZP_05077806.1,	ZP_05082750.1,
	ZP_05091128.1,	ZP_05095488.1,	ZP_05101701.1,	ZP_05116783.1,
	ZP_05121836.1,	ZP_05127756.1,	ZP_05637806.1,	ZP_05742087.1,
	ZP_05783548.1,	ZP_05786246.1,	ZP_05843149.1,	ZP_05945960.1,
	ZP_06459045.1,	ZP_06487195.1,	ZP_06492453.1,	ZP_06493162.1,
30	ZP_06703644.1,	ZP_06731146.1,	ZP_06839371.1,	ZP_07007312.1,
	ZP_07266194.1,	ZP_07374050.1,	ZP_07662787.1,	ZP_07778196.1,
	ZP_07797983.1,	ZP_08099459.1,	ZP_08138203.1,	ZP_08141719.1,
	ZP_08142973.1,	ZP_08177102.1,	ZP_08185821.1,	ZP_08186468.1,
	ZP_08208888.1,	ZP_08266590.1,	ZP_08402041.1,	ZP_08406891.1,
35	ZP_08522175.1,	ZP_08527488.1,	ZP_08631252.1,	ZP_08636687.).

In another preferred embodiment, the cell of the invention or used in the process of the invention expresses an alanine dehydrogenase. In a preferred embodiment, the term "alanine dehydrogenase", as used herein, means an enzyme catalysing the conversion of L-alanine, with consumption of water and NAD⁺, to pyruvate, ammonia and NADH. For example, use may be made of the alanine dehydrogenases from the group comprising alanine dehydrogenase of *Bacillus subtilis* (database code L20916), *Rhizobium leguminosarum* (database code CP001622), *Vibrio proteolyticus* (database code AF070716), *Mycobacterium tuberculosis* (database code X63069), *Enterobacter aerogenes* (database code AB013821), EGR93259.1, YP_003654745.1, YP_003651439.1, YP_003637111.1, YP_003631815.1, YP_001327051.1, YP_001262560.1, YP_886996.1, YP_882850.1, YP_704410.1, YP_703508.1, ZP_08624689.1, YP_001230376.1, P17557.1, P17556.1, CCB94892.1, CCB73698.1, YP_001168635.1, YP_004668736.1, YP_004569425.1, YP_003513168.1, YP_004561169.1, ZP_08554945.1, YP_400777.1, ZP_08311476.1, ZP_08310170.1, ZP_08267322.1, ZP_08263846.1, ZP_07898723.1, YP_149301.1, YP_148605.1, YP_004340432.1, EFT09946.1, EFS80513.1, EFS51332.1, EFS42459.1, YP_003060895.1, YP_003059033.1, ZP_03305373.1, YP_847214.1, YP_004095847.1, YP_003338282.1, YP_003337256.1, YP_355846.1, YP_253131.1, ZP_08197563.1, ZP_08196283.1, ADW06447.1, YP_734091.1, NP_372233.1, NP_102173.1, ZP_08170259.1, EGD36706.1, EGD32748.1, ZP_08155540.1, YP_004142849.1, YP_002417649.1, YP_001301040.1, YP_002992892.1, YP_081348.1, YP_080482.1, YP_002476349.1, ZP_08115025.1, ZP_08114403.1, YP_003552869.1, YP_002358112.1, YP_575010.1, YP_477594.1, YP_474564.1, YP_130399.1, YP_129373.1, YP_123314.1, NP_810467.1, NP_646469.1, NP_626044.1, NP_391071.1, ZP_08086822.1, ZP_08084776.1, ZP_08083119.1, ZP_08020768.1, ZP_08013590.1, ZP_08011832.1, YP_003783744.1, YP_002781576.1, YP_002780533.1, ZP_02195873.1, NP_797482.1, ZP_07645051.1, ZP_07643260.1, ZP_06611917.1, AAT40119.1, ZP_07864946.1, YP_004068409.1, YP_002796203.1, YP_002774420.1, YP_003600348.1, YP_003599946.1,

YP_003565624.1, YP_003565223.1, YP_335198.1, YP_423850.1,
YP_155059.1, ZP_07843538.1, ZP_07841226.1, ZP_06928932.1,
ZP_05692073.1, ZP_05687006.1, ZP_04867480.1, YP_775531.1,
CBE70214.1, ZP_07721182.1, ZP_04302850.1, ZP_04298961.1,
5 ZP_04287684.1, ZP_04277177.1, ZP_04248389.1, ZP_04235899.1,
ZP_02159718.1, ZP_02152178.1, YP_003974610.1,
YP_003546595.1, YP_002317127.1, ZP_07313778.1,
ZP_07302778.1, ZP_07298850.1, CBK69442.1, YP_003413835.1,
YP_003595089.1, ZP_06807811.1, YP_003582455.1,
10 YP_003464731.1, YP_003496397.1, YP_003421918.1, CBL07274.1,
CBK64956.1, YP_003508515.1, AAL87460.1, AAC23579.1,
AAC23578.1, AAC23577.1, ACU78652.1, YP_003471439.1,
YP_003452777.1, ZP_06384971.1, ACY25368.1, ABC26869.1,
AAP44334.1, EEZ80018.1, ZP_05110458.1, 1PJB_A,
15 ZP_04717201.1, ZP_04689103.1, CAO90307.1, CAM75354.1,
CAA44791.1, BAA77513.1, EGR96638.1, EGL90046.1,
YP_004510847.1, ZP_08450330.1, YP_003387804.1,
YP_003058152.1, EFS74272.1, EFS67128.1, ZP_06844564.1,
YP_826658.1, YP_001195249.1, YP_003095978.1, YP_469292.1,
20 YP_004442054.1, YP_004461174.1, YP_004055616.1,
YP_003576656.1, YP_003094537.1, YP_001295973.1, AEE71143.1,
YP_004447480.1, YP_003761844.1, YP_040853.1, YP_003154888.1,
YP_003142045.1, YP_002280953.1, NP_371963.1, NP_422368.1,
EGC98966.1, EGC76398.1, YP_004263661.1, YP_004252039.1,
25 YP_679036.1, YP_499973.1, ZP_08054972.1, ZP_08053009.1,
ZP_04067276.1, ZP_03968868.1, ZP_03963857.1, ZP_03933079.1,
ZP_03497046.1, ZP_06668924.1, ZP_06667106.1, ZP_06324464.1,
ZP_06196777.1, ZP_05114159.1, ZP_05083968.1, ZP_05070370.1,
ZP_05030022.1, ZP_04673064.1, ZP_03517011.1, ZP_03505783.1,
30 XP_001310698.1, ABK27691.1 or CAB59281.2. It is advantageous
in the case of the cell expressing an alanine dehydrogenase
to add a sufficient amount of an inorganic nitrogen source,
preferably an ammonium salt such as ammonium chloride or
ammonium sulphate, to the aqueous solution. A method of
35 increasing the concentration of alanine is described in
EP12162846.5.

One aspect of the present invention provides for the use of a knockout of a gene coding for a polypeptide comprising SEQ ID NO 2 or a variant thereof as part of the genetic make-up of a recombinant cell for increasing production of a carboxylic acid ester of the formula (I). This means that the knockout of a gene coding for a polypeptide comprising SEQ ID NO 2 or a variant thereof as a feature of the cell was carried out for the purpose of increasing the yield, carbon and/or nitrogen balance and/or purity of the product of reacting the carboxylic acid ester of the formula (I).

In a most preferred embodiment, the invention provides for a cell which is an *E. coli* cell having a knockout of the polypeptide comprising SEQ ID NO 2 or a variant thereof and of the polypeptide FadL in the cell's genome, with said cell furthermore expressing an alkane hydroxylase, preferably *Pseudomonas putida* AlkB, a transaminase, preferably the transaminase of *Chromobacterium violaceum* ATCC 12472, an alanine dehydrogenase, preferably *Bacillus subtilis* alanine dehydrogenase, and *Pseudomonas putida* AlkL. In another most preferred embodiment, said cell is contacted with a fatty acid ester, preferably methyl dodecanoate, in an aqueous solution.

Figures:

Figure 1 depicts the omega-amino-dodecanoic acid productivity of various strains.

Example 1

30

Inactivation of *bioH* in *E. coli* W3110 and BW25113

A new plasmid was needed to specifically knock out the *bioH* gene (b3412, SEQ ID 1) using the basis vector pKO3_E933. This is based on the pKO3_E933 vector (SEQ ID 14), with 500 bp being inserted upstream (SEQ ID 3) and 500 bp being inserted downstream of the *bioH* gene (SEQ ID 4), separated

by a *PspXI* cleavage site (CCTCGAGG). The finished plasmid is internally referred to as AHp-LL-42 (SEQ ID 5). To finish, the following oligonucleotides were used:

- 5 o-LL-314, SEQ ID 6 5'-
CCGGGGATCGCGGCCCGGCTTCGCTATCCCATTGGCAGT-3'
o-LL-315, SEQ ID 7 5'-
CCTCTGCTTCAACGCCCTCGAGGCATCCGCTATTGTTCTCTTTTGACTTACAAGGATG-
3'
10 o-LL-316, SEQ ID 8 5'-GCGTTGAAGCAGAGGTTGTAGGTG-3'
o-LL-317, SEQ ID 9 5'-
TAGAGGATCGCGGCCCAAAGTGGCAAGGCAGCTTTATGC-3'

The 500bp regions were provided using the above primers from
15 available chromosomal DNA of *E. coli* W3110 by means of
polymerase chain reaction (PCR), with o-LL-314 + o-LL-315
being used for the *upstream* region (SEQ ID 3) and o-LL-316 +
o-LL-317 being used for the *downstream* region (SEQ ID 4).
The following parameters were applied to the PCR:

- 20 Initial denaturation: 98 °C, 10 s
30 x Denaturation: 98 °C, 10 s
30 x Annealing: 57.9/58.8/59.7/60.6/61.4/62.3/63.2/64.1 °C
(temperature gradient)
25 30 x Elongation: 72 °C, 20 s
Final elongation: 72 °C, 4 min

For multiplication, the 2x Phusion HF Master Mix from New
England Biolabs (NEB, M0531S) was used according to the
30 manufacturer's information. Depending on the degree of
purity, the PCR products were directly column-purified
(*QiaQuick PCR Purification Kit*, Qiagen, Hilden) or purified
via an agarose gel and extracted (*QiaQuick Gel Extraction
Kit*, Qiagen, Hilden, Germany). PCR, agarose gel
35 electrophoresis, ethidium bromide staining of the DNA and
determination of PCR fragment sizes were carried out in the

manner known to the skilled worker. It was possible in both cases to provide PCR fragments of the expected size.

5 The purified PCR products were cloned into the *NotI*-cut
pKO3_E933 vector (SEQ ID 14) by means of recombination using
the In-Fusion HD Cloning Kit according to the manufacturer's
instruction (Clontech Laboratories Inc., Mountain View, CA,
USA). Chemically competent *E.coli* DH10 β (New England
Biolabs, Frankfurt, Germany) were transformed in the manner
10 known to the skilled worker. Correct insertion of the target
sequences was checked by restriction analysis, and
authenticity of the introduced sequences was confirmed by
DNA sequencing. The resulting vector was referred to as
AHP-LL-42 (SEQ ID 5).

15

The strain *E.coli* W3110 Δ *bioH* was constructed with the aid
of the vector AHP-LL-42 (SEQ ID 5) by methods known to the
skilled worker (see Link AJ, Phillips D, Church GM.
J.Bacteriol. 1997. 179(20)). The strains used in the
20 experiments, BW25113 and BW25113 Δ *bioH* (JW3375), were
purchased commercially as part of the Keio collection (see
Construction of Escherichia coli K-12 in-frame, single-gene
knockout mutants: the Keio collection, Tomoya Baba, Takeshi
Ara, Miki Hasegawa, Yuki Takai, Yoshiko Okumura, Miki Baba,
25 Kirill A Datsenko, Masaru Tomita, Barry L Wanner and
Hirotada Mori, Molecular Systems Biology (2006), 2006 EMBO
and Nature Publishing Group). The strain constructed was
subsequently referred to as:

30

Strain	Description
AHs-LL-56	<i>E. coli</i> W3110 Δ <i>bioH</i> , clone 1.1

The DNA sequence of *bioH* after deletion in W3110 is
indicated in SEQ ID 10.

35

Example 2

Production of methyl amino-dodecanoate by *E. coli* W3110 containing a deletion in the *bioH* gene by employing expression vectors for the genes *Bacillus subtilis* *ald* and
5 *Chromobacterium violaceum* *Cv_2025* in combination with an expression vector for the *alkB*, *alkG*, *alkT* and *alkL* genes of the *Pseudomonas putida* *alk* operon.

The *E. coli* strains containing expression vectors for the
10 genes *Bacillus subtilis* *ald* (coding for an alanine dehydrogenase, codon-optimized for *E. coli*), *Chromobacterium violaceum* *Cv_2025* (coding for a transaminase, codon-optimized for *E. coli*) in combination with an expression
15 vector for the *alkB*, *alkG*, *alkT* (coding for an alkane monooxygenase, a rubredoxin and a rubredoxin reductase) and *alkL* (coding for a membrane/transport protein) genes of the *Pseudomonas putida* *alk* operon were generated by preparing
electro-competent *E. coli* W3110 Δ *bioH* cells and the corresponding *E. coli* W3110 control strain. This was carried
20 out in a manner known to the skilled worker. The strains were prepared as described in Example 1. They were transformed with plasmids *pBT10_alkL* (SEQ ID 11 or WO/2011/131420 and Seq ID NO: 8 listed there) and
pJ294[alaDH_Bs(co)TA_Cv(co)] (SEQ ID 12 or Example 1 of
25 WO/2013/024114 and SEQ ID No. 17 listed there) and plated on LB-agar plates containing kanamycin (50 μ g/ml) and ampicillin (100 μ g/ml). Transformants were checked with regard to the presence of the correct plasmids by plasmid preparation and analytical restriction analysis. In this
30 way, the following strains were constructed:

Strain background

Plasmids present

W3110	<i>pBT10_alkL</i> <i>pJ294[alaDH Bs(co)TA Cv(co)]</i>
W3110 Δ <i>bioH</i>	<i>pBT10_alkL</i> <i>pJ294[alaDH Bs(co)TA Cv(co)]</i>

The strains were subjected to fed-batch fermentation, in order to analyze the capacity for producing methyl hydroxy-dodecanoate, methyl oxo-dodecanoate, methyl carboxy-dodecanoate and methyl amino-dodecanoate from methyl dodecanoate. This was carried out using an 8-fold parallel fermentation system from DASGIP.

The fermentation was carried out using 1l-reactors. The pH probes were calibrated by means of a two-point calibration using standard solutions of pH 4.0 and pH 7.0. The reactors were charged with 300 ml of tap water and autoclaved at 121°C for 20 min in order to ensure sterility. The pO₂ probes were then polarized on the DASGIP system overnight (at least for 6 h). The next morning, the water was removed at the clean bench and replaced with 300 ml of high cell-density medium containing 100 mg/l ampicillin, 50 mg/l kanamycin and 5 mg/l tetracycline. This was followed by calibrating the pO₂ probes using a one-point calibration (stirrer: 400 rpm / gassing: 10 sL/h air) and cleaning the feed, correcting agent and inducer lines by means of Clean-in-Place. For this purpose, the tubings were rinsed with 70% ethanol, followed by 1 M NaOH, then with sterile deionized water and finally filled with the respective media.

The ALS- and ALSME-producing *E. coli* strains were first grown from the respective frozen stocks in LB medium (25 ml in a 100 ml baffled flask) containing 100 mg/l ampicillin at 37°C and 200 rpm overnight for approx. 18 h. In each case 2 ml of the cultures were then transferred to high cell-density medium (glucose 15 g/l (30 ml/l of a separately autoclaved 500 g/l stock solution containing 1% MgSO₄·7H₂O and 2.2% NH₄Cl), (NH₄)₂SO₄ 1.76 g/l, K₂HPO₄ 19.08 g/l, KH₂PO₄ 12.5 g/l, yeast extract 6.66 g/l, trisodium citrate dihydrate 2.24 g/l, ammonium iron citrate solution 17 ml/l of a separately autoclaved 1% strength stock solution, trace element solution 5 ml/l separately autoclaved stock solution (HCl (37%) 36.50 g/l, MnCl₂·4H₂O 1.91 g/l, ZnSO₄·7H₂O 1.87 g/l, ethylenediaminetetraacetic acid dihydrate 0.84 g/l,

H₃BO₃ 0.30 g/l, Na₂MoO₄*2H₂O 0.25 g/l, CaCl₂*2H₂O 4.70 g/l, FeSO₄*7H₂O 17.80 g/l, CuCl₂*2H₂O 0.15 g/l)) (per strain 25 ml in a 100 ml baffled flask) containing 100 mg/l ampicillin, 50 mg/l kanamycin and 5 mg/l tetracycline and incubated at 5 37°C / 200 rpm for another 5.5 h.

The reactors were inoculated with an optical density of 0.1 by drawing a corresponding volume of the pre-culture into a 5 ml syringe (under sterile conditions) and inoculating the reactors by means of a cannula via a septum overlaid with 10 70% ethanol.

The following standard program was used:

DO Control		pH Control	
Preset	0%	Preset	0 ml/h
P	0.1	P	5
Ti	300 s	Ti	200 s
min	0%	min	0 ml/h
max	100%	max	40 ml/h

15

N (rotation)	from	to	XO ₂ (gas mixture)	from	to	F (gas flow)	from	to
Growth and biotrans- formation	0%	30%	Growth and biotrans- formation	0%	100%	Growth and biotrans- formation	15%	80%
	400 rpm	1500 rpm		21%	21%		6 sL/h	72 sL/h

Script	
Trigger set	31% DO (1/60h)
Induction IPTG	2 h after feed start
Feed trigger	50% DO
Feed rate	3 [ml/h]

The experiment carried out can be divided into two phases, growth, during which the cells ought to reach a particular optical density, and subsequent biotransformation, during which, after addition of the substrate methyl dodecanoate, a reaction by enzymes formed during expression to give amino-dodecanoate ought to take place. The pH values were adjusted one-sidedly with ammonia (12.5%) to pH 6.8. During growth and biotransformation, dissolved oxygen (DO) in the culture was controlled via stirrer revolution and gassing rate to be at 30%. Fermentation was carried out by way of fed batch, with feed start, 5 g/lh glucose feed (500 g/l glucose containing 1% $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ and 2.2% NH_4Cl), being triggered via a DO peak. Also, with feed start, the temperature was lowered from 37°C before to 30°C. Expression of transaminase, alanine dehydrogenase and fatty acid reductase was induced by automated addition of IPTG (1 mM) 2 h after feed start. The alk genes were induced by manually adding DCPK (0.025 Vol.-%) 10 h after feed start. The optical density of the culture broths was determined before the start of biotransformation.

The biotransformation phase was started 14 h after feed start. For this purpose, 150 ml of a mixture of methyl dodecanoate and the ion exchanger oleic acid (technical grade, 90%) were added as batch to the fermentation broth. In order to provide an amino group donor for the transaminase, 5 ml of a 3 M ammonium sulphate solution were added to the fermentation broth half an hour before the start of biotransformation. Samples were taken by removing 2 ml of fermentation broth from the tank and diluting a portion thereof 1/20 in an acetone/HCl mixture ($c(\text{HCl}) = 0.1 \text{ mol/l}$) and extracting it. Samples were taken from all reactors 1 h, 2 h, 3 h, 4 h, 5 h, 7.5 h, 10.5 h, 19.5 h and 21 h after the start of biotransformation. The transfer rates for oxygen ($\text{OTR} = \text{oxygen transfer rate}$) and carbon

(CTR = carbon transfer rate) were determined by off-gas analysis on the DASGIP systems during fermentation. Fermentation was stopped 21 h after the start of biotransformation. Stirrer, gassing, and temperature and pH controls were switched off and the tanks were left standing still for 5-10 minutes.

To quantify DDS (C12-di-carboxylic acid), DDSME (C12-di-carboxylic acid methyl ester), LS (dodecanoic acid [Laurinsäure]), LSME (methyl dodecanoate [Laurinsäure-Methylester]), HLS (omega-hydroxy-dodecanoic acid [omega-Hydroxy-Laurinsäure]), HLSME (methyl omega-hydroxy-dodecanoate [omega-Hydroxy-Laurinsäure-Methylester]), OLS (omega-oxo-dodecanoic acid [omega-Oxo-Laurinsäure]), OLSME (methyl omega-oxo-dodecanoate [omega-Oxo-Laurinsäure-Methylester]), ALS (omega-amino-dodecanoic acid [omega-Amino-Laurinsäure]) and ALSME (methyl omega-amino-dodecanoate [omega-Amino-Laurinsäure-Methylester]) in fermentation samples, samples were removed during culturing. These samples were prepared for analysis. (see LC-ESI/MS²-based quantification of products).

LC-ESI/MS²-based quantification of products

Quantification of ALS, ALSME, DDS, DDSME, LS, LSME, HLS, HLSME, OLS and OLSME in fermentation samples was carried out by means of LC-ESI/MS² on the basis of an external calibration for all analytes (0.1 - 50 mg/L) and by using the internal standards amino-undecanoic acid (AUD, for HLS, DDS, OLS, HLSME, OLSME), d4-ALSME (for ALSME), ¹³C-DDSME (for DDSME), d3-LS (for LS) and d3-LSME (for LSME).

The following instruments are used:

- HPLC system 1260 (Agilent; Böblingen, Germany) with autosampler (G1367E), binary pump (G1312B) and thermostatted column (G1316A)
- Mass spectrometer TripelQuad 6410 (Agilent; Böblingen, Germany) with ESI source

- HPLC column: Kinetex C18, 100 x 2.1 mm, particle size: 2.6 μm , pore size 100 $\text{\AA}$ (Phenomenex; Aschaffenburg, Germany)
- Pre-column: KrudKatcher Ultra HPLC In-Line Filter; 0.5 μm filter depth and 0.004 mm inner diameter (Phenomenex; Aschaffenburg, Germany)

The samples were prepared by pipetting 1900 μl of solvent (80% (v/v) ACN, 20% double-distilled H_2O (v/v), + 0.1% formic acid) and 100 μl of sample into a 2 ml reaction vessel. The mixture was vortexed for approx. 10 seconds and then centrifuged at approx. 13 000 rpm for 5 min. The clear supernatant was removed using a pipette and analysed after appropriate dilution with a diluent (80% (v/v) ACN, 20% double-distilled H_2O (v/v), + 0.1% formic acid). In each case, 100 μl of ISTD were added to 900 μl of sample (10 μl with a sample volume of 90 μl).

HPLC separation was carried out using the above-mentioned column and pre-column. The injection volume is 0.7 μl , the column temperature is 50°C, the flow rate is 0.6 ml/min. The mobile phase consists of eluent A (0.1% strength (v/v) aqueous formic acid) and eluent B (acetonitrile with 0.1% (v/v) formic acid). The following gradient profile was utilized:

25

Time [min]	Eluent A [%]	Eluent B [%]
0	77	23
0.3	77	23
0.4	40	60
2.5	40	60
2.6	2	98
5.5	2	98
5.6	77	23
9	77	23

ESI-MS² analysis was carried out in positive mode with the following parameters of the ESI source:

- Gas temperature 280°C
- Gas flow 11 l/min
- Nebulizer pressure 50 psi
- Capillary voltage 4000 V

5

Detection and quantification of the compounds ALS, ALSME, DDS, DDSME, HLS, HLSME, OLS, OLSME were carried out with the following MRM parameters, with in each case one product ion being utilized as qualifier and one being utilized as

10

quantifier:

Analyte	Precursor ion [m/z]	Product ion [m/z]	Dwell time [ms]	Collision energy [eV]
DDSME	245.2	167.1	25	6
DDSME	245.2	149.1	50	8
HLSME	231.3	181.2	15	2
HLSME	231.3	163.2	25	5
DDS	231.2	213.2	50	0
DDS	231.2	149.1	25	9
ALSME	230.3	198.1	25	10
ALSME	230.3	163.2	15	10
OLSME	229.2	197.2	50	0
OLSME	229.2	161.1	25	5
HLS	217.2	181.2	35	0
HLS	217.2	163.1	20	4
OLS	215.2	161.2	25	0
OLS	215.2	95.2	60	13

The analytes LS and LSME were detected in SIM mode (m/z 201 and 215).

15

Results

20

Reduced formation of the free acids amino-dodecanoic acid, dodecanedioic acid and dodecanoic acid after knockout of *bioH* in *E. coli* W3110

5

The strain with *bioH* knockout exhibited markedly reduced formation of the free acids amino-dodecanoic acid, dodecanedioic acid and dodecanoic acid compared to the control strain with intact *bioH*. The ratios of the absolute end titres of dodecanedioic acid (DDS) and dodecanedioic acid methyl ester (DDSME), amino-dodecanoic acid (ALS) and methyl amino-dodecanoate (ALSME), and dodecanoic acid (LS) and methyl dodecanoate (LSME) were calculated and are expressed in per cent.

15

	End titre in g/l					
	ALS	ALSME	DDS	DDSME	LS	LSME
W3110	0.12	11.77	5.02	7.27	0.03	1.91
W3110 Δ <i>bioH</i>	0.01	11.82	0.23	2.35	0.03	12.92

	Free acid/methyl ester ratio in per cent		
	ALS/ALSME	DDS/DDSME	LS/LSME
W3110	0.98	69.12	1.79
W3110 Δ <i>bioH</i>	0.10	9.74	0.24

	Free acid/methyl ester ratio in per cent		
	ALS/ALSME	DDS/DDSME	LS/LSME
W3110 to W3110 Δ <i>bioH</i>	9.75	7.10	7.57

20

The effect of the *bioH* knockout on the formation of free acids became clearly visible and was between a reduction by around a factor of 7.10 (DDS/DDSME) up to a factor of 9.75 (ALS/ALSME).

25

Improved product/secondary product ratio after knockout of *bioH* in *E. coli* W3110

5 The absolute titres attained after a fixed time (= end of biotransformation) were evaluated. The strain background W3110 Δ *bioH* exhibited a markedly improved ratio of products (ALS and ALSME) to major secondary products (DDS and DDSME), with the final product titre remaining the same. The ratio increased from 49.17 % ALS (ME) to 82.10 % ALS (ME).

10

	Concentration in g/l		Ratio of ALS (ME) to ALS (ME) + DDS (ME) in per cent
	ALS + ALSME	DDS + DDSME	
W3110	11.89	12.29	49.17
W3110 Δ <i>bioH</i>	11.83	2.58	82.10

Example 3

15

Production of methyl amino-dodecanoate by an *E. coli* strain having a deletion in the *bioH* gene by employing an expression vector for the genes *Bacillus subtilis ald* and *Chromobacterium violaceum Cv_2025* and the *alkB*, *alkG*, *alkT* and *alkL* genes of the *Pseudomonas putida alk* operon

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The *E. coli* strains containing an expression vector for the genes *Bacillus subtilis ald* (coding for an alanine dehydrogenase, codon-optimized for *E. coli*), *Chromobacterium violaceum Cv_2025* (coding for a transaminase, codon-optimized for *C. tropicalis*), *alkB*, *alkG*, *alkT* (coding for an alkane monooxygenase, a rubredoxin and a rubredoxin reductase) and *alkL* (coding for a membrane/transport protein) of the *Pseudomonas putida alk* operon were generated by preparing electro-competent *E. coli* BW25113 Δ *bioH* cells and the corresponding *E. coli* BW25113 control strain. This took place in a manner known to the skilled worker. The

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strains are from the commercially available Keio collection. They were transformed with plasmid pACYC184{MCS2.0}[alkST_BFGL][alaDH_Bs(co) {PspXI} TAcv(ct)] (SEQ ID 13 and Example 1 of WO/2013/024114 and SEQ ID No. 17 listed there) and plated on LB-agar plates containing chloramphenicol (50 µg/ml). Transformants were checked with regard to the presence of the correct plasmids by plasmid preparation and analytical restriction analysis. In this way, the following strains were constructed:

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Strain background Plasmids present

BW25113	pACYC184{MCS2.0}[alkST_BFGL][alaDH_Bs(co) {PspXI} TAcv(ct)]
BW25113 <i>ΔbioH</i>	pACYC184{MCS2.0}[alkST_BFGL][alaDH_Bs(co) {PspXI} TAcv(ct)]

The strains were subjected to fed-batch fermentation, in order to analyze the capacity for producing methyl hydroxy-dodecanoate, methyl oxo-dodecanoate, methyl carboxy-dodecanoate and methyl amino-dodecanoate from methyl dodecanoate. This was carried out using an 8-fold parallel fermentation system from DASGIP. Further experimental procedures were exactly as described in Example 2.

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Results

Reduced formation of the free acids amino-dodecanoic acid, dodecanedioic acid, dodecanoic acid and hydroxy-dodecanoic acid after knockout of *bioH* in *E. coli* BW25113

The strain with *bioH* knockout exhibited markedly reduced formation of the free acids amino-dodecanoic acid, dodecanedioic acid, dodecanoic acid and hydroxy-dodecanoic acid compared to the control strain with intact *bioH*, in part below the detection limit. The ratios of the absolute end titres of dodecanedioic acid (DDS) and dodecanedioic

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acid methyl ester (DDSME), amino-dodecanoic acid (ALS) and methyl amino-dodecanoate (ALSME), hydroxy-dodecanoic acid (HLS) and methyl hydroxy-dodecanoate (HLSME), and dodecanoic acid (LS) and methyl dodecanoate (LSME) were calculated and are expressed in per cent.

	Titre in g/l							
	ALS	ALSME	DDS	DDSME	HLS	HLSME	LS	LSME
BW25113	0.16	10.80	2.75	1.40	0.72	15.01	0.48	40.77
BW25113 $\Delta bioH$	n.d.	15.86	0.23	3.95	n.d.	12.32	n.d.	28.49

The DDS/DDSME ratio is the only one that can be presented mathematically:

	DDS/DDSME in per cent
BW25113	196.94
BW25113 $\Delta bioH$	5.87

The *bioH* knockout reduced DDS formation by a factor of 33.5, with all other free acids being below the lower detection limit.

Extended oxidation phase after knockout of *bioH* in *E. coli* BW25113

It turned out that, compared to the control strain, the initial oxidation performance does not decline in the *bioH*-knockout strain but remains nearly constant with respect to ALSME formation throughout the process.

	Process time	ALSME
	[h]	[g/l]
	1	0.26
	2	1.16
BW25113	3	2.35

	19	10.51
	21.25	10.80
BW25113 Δ bioH	1	0.38
	2	1.27
	3	2.48
	19	14.94
	21.25	15.86

Figure 1 shows: The *bioH* knockout effected an increase in productivity by 47% based on the end titre under the same conditions.

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Increased rate of formation of methyl amino-dodecanoate after knockout of *bioH* in *E. coli* BW25113

- 10 The absolute titres attained after a fixed time (= end of biotransformation) were evaluated. Here, the knockout strain displayed a distinctly higher rate of product formation of 0.75 g of ALSME per litre and hour over 0.51 g of ALSME per litre and hour in the wild-type strain.

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	Process time [h]	ALSME [g/l]
BW25113	21.25	10.80
BW25113 Δ bioH	21.25	15.86

- 20 **Improved product/secondary product ratio after knockout of *bioH* in *E. coli* BW25113**

- 25 The absolute titres attained after a fixed time (= end of biotransformation) were evaluated. Here, the knockout strain background displayed a markedly improved ratio of product (ALS and ALSME) to major secondary product (DDS and DDSME).

	Titre in g/l			
	ALS	ALSME	DDS	DDSME
BW25113	0.16	10.80	2.75	1.40
BW25113 <i>ΔbioH</i>	0.00	15.86	0.23	3.95

	ALS + ALSME [g/l]	DDS + DDSME [g/l]
BW25113	10.96	4.15
BW25113 <i>ΔbioH</i>	15.86	4.18

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	ALS (ME) / DDS (ME)
BW25113	2.64
BW25113 <i>ΔbioH</i>	3.80

The ratio increased by 43.9 % due to *bioH* knockout. The product titre increased by 43.9 %, with the secondary-product titre remaining the same.

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Increased glucose yield coefficient ($Y_{P/S}$) for production of amino-dodecanoic acid and methyl amino-dodecanoate after knockout of *bioH* in *E. coli* BW25113

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The final cumulated absolute titres of amino-dodecanoic acid and methyl amino-dodecanoate were evaluated in comparison to the amount of glucose used.

	ALS + ALSME [g/l]	Glucose consumption [g]	g/l ALS (ME) per g of glucose
BW25113	10.96	52.95	0.21

BW25113 Δ <i>bioH</i>	15.86	52.55	0.30
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The glucose yield coefficient increased from 0.21 g/l ALS(ME) per gram of glucose by 46 % to 0.30 g/l ALS(ME) per gram of glucose as a result of the *bioH* knockout.

Patent Claims

1. A process for transforming a carboxylic acid ester of
5 the formula (I) into a second carboxylic acid ester:



where

10 R^1 is hydrogen, $-CH_2OH$, $-CHO$, $-COOR^3$, $-CH_2SH$, $-CH_2OR^3$ and
 $-CH_2NH_2$,

R^2 is an alkyl group,

15 R^3 is hydrogen or an alkyl group, and

A is an unsubstituted, a linear, or an unsubstituted and
linear alkylene radical having at least 10 carbons,

20 the process comprising:

contacting a recombinant cell with the carboxylic acid
ester of the formula (I) in an aqueous solution to
convert the carboxylic acid ester of the formula (I) to
the second carboxylic acid ester,

25 wherein the recombinant cell has a reduced activity of a
polypeptide comprising SEQ ID NO: 2 or a polypeptide
having at least 70% homology to SEQ ID NO: 2 and having
essentially the same enzymatic activity as the
30 polypeptide comprising SEQ ID NO: 2 compared to an
activity in a corresponding wild-type cell due to knock-
out of a gene encoding the polypeptide comprising SEQ ID
NO: 2 or the polypeptide having at least 70% homology to

SEQ ID NO: 2 and having essentially the same enzymatic activity as the polypeptide comprising SEQ ID NO: 2, the recombinant cell comprises a polypeptide having at least 80% sequence identity to consecutive 230 amino acids of SEQ ID NO: 15 and having essentially the same enzymatic activity as *Pseudomonas putida* AlkL, where the polypeptide is optionally overexpressed, the recombinant cell has been transferred to express a transaminase, *Pseudomonas putida* AlkBGT, and an alanine dehydrogenase, the recombinant cell has a reduced activity of at least one enzyme catalyzing any of reactions of fatty acid β -oxidation compared to an activity in the wild-type cell due to knock-out, and the at least one enzyme catalyzing any of reactions of fatty acid β -oxidation is selected from the group consisting of fatty acid-CoA ligase, acyl-CoA dehydrogenase, 2,4-dienoyl-CoA reductase, enoyl-CoA hydratase, 3-ketoacyl-CoA thiolase, and a fatty acid importer.

2. The process according to Claim 1, further comprising:

contacting the aqueous solution with a hydrophobic organic solution comprising a cation exchanger.

3. The process of claim 1, wherein R^2 is methyl, ethyl or propyl, R^3 is methyl, ethyl or propyl, and A is an unsubstituted, a linear, or an unsubstituted and linear alkylene radical having 11 carbons.

4. The process according claim 1, wherein A is an unsubstituted and linear alkylene radical.

5. The process according to claim 1, wherein R^1 is hydrogen, $-\text{CH}_2\text{OH}$, $-\text{CHO}$ and $-\text{CH}_2\text{NH}_2$.

6. The process according to claim 1, wherein A is an alkylene radical of formula $-(CH_2)_n-$, where n is at least 10.

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7. The process according to claim 1, wherein the second carboxylic acid ester has the formula (I'):



where

10 R^1 is hydrogen, $-CH_2OH$, $-CHO$, $-COOR^3$, $-CH_2SH$, $-CH_2OR^3$ and $-CH_2NH_2$,

R^2 is an alkyl group,

R^3 is hydrogen or an alkyl group, and

A is an unsubstituted, a linear, or an unsubstituted and
15 linear alkylene radical having at least 10 carbons.

8. The process according to claim 1, wherein the carboxylic acid ester of the formula (I) is methyl dodecanoate.

- 20 9. The process according to claim 1, wherein the second carboxylic acid ester comprises methyl 12-amino-dodecanoate.

10. The process according to claim 1, wherein the at least
25 one enzyme catalyzing any of reactions of fatty acid β -oxidation comprises at least one of FadA, FadB, FadD, FadL, and FadE from *Escherichia coli*.

11. The process according to claim 1,
30 wherein R^1 is hydrogen, $-CH_2OH$, $-CHO$ or $-CH_2NH_2$,
 R^2 is methyl, ethyl or propyl, and
A is an alkylene radical of formula $-(CH_2)_n-$, where n is at least 10.

- 35 12. The process according to claim 1,
wherein the cell has a reduced activity of a polypeptide comprising SEQ ID NO: 2 or a polypeptide having at least

90% homology to SEQ ID NO:2 and having essentially the same enzymatic activity as the polypeptide comprising SEQ ID NO:2 compared to an activity in a corresponding wild-type cell due to knockout of a gene encoding the polypeptide comprising SEQ ID NO:2 or the polypeptide having at least 90% homology to SEQ ID NO:2 and having essentially the same enzymatic activity as the polypeptide comprising SEQ ID NO:2, and the recombinant cell comprises a polypeptide having at least 90% sequence identity to consecutive 230 amino acids of SEQ ID NO: 15 and having essentially the same enzymatic activity as *Pseudomonas putida* AlkL.

13. The process according to claim 1, wherein the cell has a reduced activity of a polypeptide comprising SEQ ID NO: 2 or a polypeptide having at least 96% homology to SEQ ID NO:2 and having essentially the same enzymatic activity as the polypeptide comprising SEQ ID NO:2 compared to an activity in a corresponding wild-type cell due to knockout of a gene encoding the polypeptide comprising SEQ ID NO:2 or the polypeptide having at least 96% homology to SEQ ID NO:2 and having essentially the same enzymatic activity as the polypeptide comprising SEQ ID NO:2, and the recombinant cell comprises a polypeptide having at least 90% sequence identity to consecutive 230 amino acids of SEQ ID NO: 15 and having essentially the same enzymatic activity as *Pseudomonas putida* AlkL.

14. The process according to claim 1, wherein the carboxylic acid ester of the formula (I) is methyl dodecanoate, and the second carboxylic acid ester comprises methyl 12-amino-dodecanoate.

15. The process according to claim 14, wherein the recombinant cell is *Escherichia coli*, the transaminase

is *Chromobacterium violaceum* Cv_2025, and the alanine dehydrogenase is *Bacillus subtilis* ald.

16. The process according to claim 15, further comprising:
5 after the contacting, extracting the second carboxylic acid ester from the aqueous solution by contacting the aqueous solution with a hydrophobic organic solution comprising a cation exchanger.
- 10 17. The process according to claim 16, wherein the recombinant cell comprises a polypeptide having the consecutive 230 amino acids of SEQ ID NO: 15.
- 15 18. A method for producing a carboxylic acid ester of the formula (I') from a substrate carboxylic acid ester:



where

- 20 R^1 is hydrogen, $-CH_2OH$, $-CHO$, $-COOR^3$, $-CH_2SH$, $-CH_2OR^3$ or $-CH_2NH_2$,
 R^2 is an alkyl group,
 R^3 is hydrogen or an alkyl group, and
A is an unsubstituted, a linear, or an unsubstituted and
25 linear alkylene radical having at least 10 carbons,
the method comprising:
contacting a recombinant cell with the substrate
carboxylic acid ester in an aqueous solution to convert
the substrate carboxylic acid ester to the carboxylic
30 acid ester of the formula (I'),
wherein the recombinant cell comprises a knockout of a
gene encoding a polypeptide comprising SEQ ID NO: 2 or a
polypeptide having at least 70% homology to SEQ ID NO:2
and having essentially the same enzymatic activity as
35 the polypeptide comprising SEQ ID NO:2,
the recombinant cell comprises a polypeptide having at
least 80% sequence identity to consecutive 230 amino

acids of SEQ ID NO: 15 and having essentially the same enzymatic activity as *Pseudomonas putida* AlkL, where the polypeptide is optionally overexpressed,

the recombinant cell has been transformed to express a transaminase, *Pseudomonas putida* AlkBGT, and an alanine dehydrogenase,

the production of the carboxylic acid ester of the formula (I') is increased over a corresponding wild-type cell of the recombinant cell,

the recombinant cell has a reduced activity of at least one enzyme catalyzing any of reactions of fatty acid β -oxidation compared to an activity in the wild-type cell due to knockout, and

the at least one enzyme catalyzing any of reactions of fatty acid β -oxidation is selected from the group consisting of fatty acid-CoA ligase, acyl-CoA dehydrogenase, 2,4-dienoyl-CoA reductase, enoyl-CoA hydratase, 3-ketoacyl-CoA thiolase, and a fatty acid importer.

19. The method according to claim 18,

wherein the recombinant cell comprises a knockout of a gene encoding a polypeptide comprising SEQ ID NO: 2 or a polypeptide having at least 90% homology to SEQ ID NO:2 and having essentially the same enzymatic activity as the polypeptide comprising SEQ ID NO:2, and

the recombinant cell comprises a polypeptide having at least 90% sequence identity to consecutive 230 amino acids of SEQ ID NO: 15 and having essentially the same enzymatic activity as *Pseudomonas putida* AlkL.

20. The method according to claim 18,

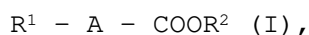
wherein the recombinant cell comprises a knockout of a gene encoding a polypeptide comprising SEQ ID NO: 2 or a polypeptide having at least 96% homology to SEQ ID NO:2 and having essentially the same enzymatic activity as the polypeptide comprising SEQ ID NO:2, and

the recombinant cell comprises a polypeptide having at least 90% sequence identity to consecutive 230 amino acids of SEQ ID NO: 15 and having essentially the same enzymatic activity as *Pseudomonas putida* AlkL.

Process for reacting a carboxylic acid ester

Abstract

The invention relates to a process for reacting a carboxylic
5 acid ester of the formula (I)



wherein R^1 is selected from the group comprising
10 hydrogen, $-CH_2OH$, $-CHO$, $-COOR^3$, $-CH_2SH$, $-CH_2OR^3$ and
 $-CH_2NH_2$,

wherein R^2 is selected from the group comprising alkyl,
preferably methyl, ethyl and propyl,
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wherein R^3 is selected from the group comprising hydrogen
and alkyl, preferably methyl, ethyl and propyl,

and wherein A is a substituted, unsubstituted, linear,
20 branched and/or cyclic alkylene, alkenylene, arylene or
aralkylene radical having at least 4, more preferably 6,
even more preferably 8, carbons,

by means of a cell, comprising the step of
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a) contacting the cell with said carboxylic acid
ester in an aqueous solution,

wherein the cell is a recombinant cell which has reduced
30 activity of a polypeptide comprising SEQ ID NO 2 or a
variant thereof over the wild-type cell, and to a cell
suitable therefor, and to uses thereof.

(No suitable figure)