HEPATITIS B TREATMENT VACCINE BASE ON INACTIVATED WHOLE RECOMBINANT HANSENULA POLYMORPHA CELLS WHICH EXPRESSES HBSAG AND HBCAG

CROSS-REFERENCE TO RELATED PATENT APPLICATION

This application claims the benefit of priority to China Patent Application No. 201610176389.4, filed on March 25, 2016 in People's Republic of China. The entire content of the above identified application is incorporated herein by reference.

Some references, which may include patents, patent applications and various publications, may be cited and discussed in the description of this disclosure. The citation and/or discussion of such references is provided merely to clarify the description of the present disclosure and is not an admission that any such reference is "prior art" to the disclosure described herein. All references cited and discussed in this specification are incorporated herein by reference in their entireties and to the same extent as if each reference was individually incorporated by reference.

FIELD OF THE DISCLOSURE

The present disclosure relates to a field of genetic engineering, and more particularly to a hepatitis B vaccine including HBsAg and HBcAg expressed by inactivated whole recombinant Hansenula polymorpha cells.

BACKGROUND OF THE DISCLOSURE

HBV (hepatitis B virus, HBV) infection is a serious public health problem. According to the World Health Organization (WHO), about 20 million people have been infected with HBV within 6 billion people worldwide, of which 350 million people have chronic HBV infection; about 1 million people die each year from liver failure, cirrhosis and primary hepatocellular carcinoma (liver cancer) caused by HBV infection. Liver cancer patients worldwide, more than 75% is caused by HBV. China is an endemic area of HBV infection. The Ministry of Health of China incorporated hepatitis B vaccine into planned immunization management in 1992. In December

2001, the State Council of China officially approved the immunization of hepatitis B vaccine into children's programs, requiring all neonates in all provinces and autonomous regions vaccinated hepatitis B vaccine for free (except for a small fee) since 2002. According to the plan immunization regulations formulated in March 2005, from June 1, 2005, offered free hepatitis B vaccine to all neonates. After nearly 15 years of efforts, the general population in China, especially children under the age of 15, HBV infection rate has significantly decreased. According to the national hepatitis B serological surveys in 2006, the carrying rate of hepatitis B surface antigen (HBsAg) in the whole population has decreased from 9.75% to 7.18% since 1992, and the hepatitis B surface antigen carrying rate in children under 5 years old has been decreased from 9.67% to 0.96%. Based on projections, about 93 million people are suffering from chronic HBV infection in China, including about 20 million patients with chronic hepatitis B. It is estimated that liver cirrhosis and liver cancer due to HBV cause nearly 300,000 death cases each year, wherein new hepatitis B are about 0.5 to 1 million cases. Therefore, HBV disease is an important factor that endangers people's health, hinders social development, and affects social stability for a long time. It is a serious public health problem in people-oriented society, and also a priority major health issue in China. The prevalence of HBV infection in Chinese population is high, which brings a heavy economic burden to the country. According to the survey, the annual direct and indirect medical expense for chronic hepatitis B (including liver cirrhosis and liver cancer) in China is about 680 billion. Hepatitis B vaccine immunization prevention is the most effective way to reduce the burden of disease.

Gene recombination technology is the core technology of modern biotechnology; also is the mainly technology of the large-scale production of hepatitis B vaccine, and the only technology of virus-like particle hepatitis B virus surface antigen (HBsAg VLP). The applicant has been involved in the research and development of Hansenula polymorpha (H. polymorpha) recombinant hepatitis B vaccine since 1995. In 1998 to 2002, in Dalian Gaoxin Bio-Pharmaceutical Co., Ltd., the H. polymorpha recombinant HBsAg-adr2 hepatitis B vaccine was developed. The yield of HBsAg VLP pure stock solution was 40 mg/L, and it has been approved since

2002 in China. In 2003 to 2006, the applicant assisted Beijing Tiantan Biological Products Co., Ltd. to develop recombinant H. polymorpha HBsAg-adr2 hepatitis B vaccine. The pure stock solution of HBsAg VLP has a yield of 85 mg/liter or more; it has been submitted to the National New Drug Review in 2015, and the China patent publication number based on the vaccine is CN104232661A.

The pathogenesis of hepatitis B identified according to the prior art is as follows: After HBV infection, the HBV carriers can be generally divided into immune tolerance phase, immune clearance phase, and residual or inactive phase. The immune tolerance phase is characterized by high levels of HBV replication, positive serum HBsAg and HBeAg, high HBV DNA level (>10^5 copies/ml), normal alanine aminotransferase level (ALT), and no obvious abnormalities in liver histology. The immune clearance phase is characterized by serum HBV DNA level >10^5 copies/ml, but generally lower than the immune tolerance phase, normal or intermittently elevated aspartate aminotransferase (AST) level, and necrotic inflammation shown in liver histology. The residual or inactive phase is characterized by HBeAg-negative, anti-HBe-positive, could not be detected (PCR assay) or below the lower limit of detection, level normal, and no obvious inflammation in liver histology. However, HBV infection in adolescents and adults generally does not start from the immune tolerance phase, but initial from the immune clearance phase, which is manifested as acute hepatitis B, of which only 5%-10% develop chronic hepatitis B. However, the exact pathogenesis is still unknown.

Anti-hepatitis B virus treatment is currently the main treatment for hepatitis B virus infection and hepatitis B patients. At present, anti-hepatitis B virus drugs mainly include interferon-based immunomodulators and nucleotide analogues against HBV DNA polymerase. Although they have certain curative effects, they are not satisfactory, and most patients cannot be cured. Interferon can induce HBsAg seroclearance or seroconversion in few patients, but its high cost, need to be injected, and has certain side effects. Nucleotide analogs act on HBV DNA polymerase, which only inhibits viral replication, does not completely eliminate HBV DNA and cccDNA, and easily lead to viral resistance mutation by long-term therapy.

Therefore, in order to completely eliminate HBV and cccDNA, to develop a new and more effectively of HBV hepatitis B vaccine is urgent need. The low immune rejection of liver transplantation indicates that the human liver is an immune-tolerant organ. Liver is the target organ of hepatitis B virus (HBV) infection, so that the immune tolerance to HBV in liver is a major feature. Reversal of HBV immune tolerance is the base for the development of immunotherapy vaccine for chronic hepatitis B (CHB) patients. HBV immune tolerance is not only reflected anti-HBV immune response failed to effectively eliminate the virus in local liver, leading to persistent infection, but also reflected in the persistence of HBV, leading the systemic immune system with no response to the HBV, such as the patient in HBV immune tolerance phase is no response to HBsAg vaccine. This is also the main reason why current therapeutic vaccines are difficult to succeed in CHB patients. Liver-induced immune tolerance and its reversal mechanism will provide a theoretical basis for the development of hepatitis B vaccine.

In 2005, Bowen suggested that the immune environment in the liver is associated with induced tolerance, but still maintains the ability to maintain an effective response to pathogens. The mechanism of this dichotomy is still unclear. Recent data suggest that initial CD8+T (CTL) cell activation occurs in the liver, while pre-inflammatory (i.e. innate immune activation) increases the number of survival CTLs, makes CTLs more effectively, resulting in a liver immune response to eliminate the infected HBV. In the absence of inflammation in advance (ie, innate immunity is not activated, such as infants and young children), the function of CTL is impaired and the half-life of CTL is short, resulting in liver tolerance to HBV. However, initial HBV antigen encounter immunity induced high-efficiently activated HBV CTLs in the lymph nodes outside the liver, and then into the liver, which also increases the number of survival CTLs, makes CTLs more effectively, resulting in a liver immune response to eliminate the infected HBV. This immune mechanism for the two sites provides a theoretical basis for the development of subcutaneous and intramuscular injection of hepatitis B vaccine, which leads to liver immune response and HBV clearance.

According to the journal “A Whole Recombinant Yeast-Based Therapeutic Vaccine that is comprised of heat-inactivated, whole recombinant Saccharomyces cerevisiae yeast cells expressing disease-related antigens” of Thomas H. King (US) in 2014, which relates to a therapeutic vaccine based on a heat-inactivated, whole recombinant Saccharomyces cerevisiae yeast cells expressing disease-related antigen. The study pioneered a therapeutic vaccine platform that uses intracellular recombinantly expressed proteins as antigens and heat-inactivated whole Saccharomyces cerevisiae cells as adjuvant. In addition, the HBV antigen expressed by a hepatitis B therapeutic vaccine (which has been numbered as GS4774 under the platform) is an x-s-core antigen as fusion protein. This yeast vector provides multiple antigens into the MHC class I and class II antigen presentation pathways, stimulates potent CD4+ and CD8+ cell responses, and disrupts antigen tolerance in immunogenic mouse model. The yeast vector is also not easily being neutralized in body, and is therefore suitable for repeated administration to obtain long-term immunological stress, ideally eliminating chronic intracellular infections such as HCV and HBV.

According to the journal of Huang in 2010, β-Glucan particles (GPs) which are purified from Saccharomyces cerevisiae cell walls have >85% β1,3-d-glucan polymers, ~2% chitin, and <1% lipids and protein, with the rest being mostly ash and moisture. In *in vitro* T-cell proliferation assays, ovalbumin (OVA) was complexed into the hollow GP shells (GP-OVA) to as vaccine, and free OVA as control antigen. At concentrations from 0.03 μg/ml to 0.5 μg/ml, GP-OVA stimulated OT-I and OT-II T-cell proliferation. In contrast, free OVA failed to stimulate proliferation of either OT-I or OT-II T cells. In order to achieve similar stimulation effects of GP-OVA, 100 times or higher concentrations of free OVA were required. These results demonstrate that: (1) Virus-like particle GPS is an efficiently t agonist of the Dectin-1 receptor. (2) Compared with free OVA, antigens delivered in Virus-like particle GP-OVA were more efficiently processed and presented by DCs (dendritic cells).

In 2003 to 2005, American scientists reported the results of a series of studies on hepatitis B virus infection in chimpanzees. The mechanism controlling disease is the covalently closed circular DNA (cccDNA) of the hepatocyte nuclear HBV pool.

The HBV-specific CD8+T (CTL cells), which produce INF-γ, massively influx into liver and target to hepatocytes infected with HBV, and the cccDNA clearance and hepatocytes infected HBV reversion are related to the INF-γ produced by liver CD8+ T cells. These results suggest that cccDNA clearance is a two-step process mediated by cellular immune responses: In the first step is to reduce the pool of cccDNA molecules by more than 90% without cell damage, thereby eliminating the precursor of HBV-relaxed circular deoxyribonucleic acid, and the second step is to improve the process of destroying infected liver cells and trigger an immune reversion.

In 2014, Dr. Zeng Zhutian of the University of Science and Technology of China reported that, by hydrodynamic injection of HBV persistent mouse mimic the immune tolerance phase of chronically infected HBV patients, the combination therapy of IL-12 pretreatment with IL-12 and HBsAg VLP vaccine, which is called IL-12-based vaccine therapy, can effectively reverse HBV systemic immune tolerance, and lead to HBV clearance. The levels of follicular-like helper T cells (Tfh, which are in lymph node) and germinal center B cells (GC B) of HBV mice were significantly increased after undergoing IL-12-based vaccine therapy. Correspondingly, HBsAg-specific IgG-producing cells in spleen cells were also significantly increased, and most of the mice showed protective antibody anti-HBs in the serum of the late treatment. In addition, the ability of T cells to stimulate HBsAg stimulation in vitro was also significantly restored after IL-12 combination vaccine treatment. These results fully confirm that HBV-tolerant mice recover from the peripheral HBsAg vaccine during IL-12 combination therapy, suggesting that HBV-induced systemic immune tolerance has been reversed. Based on the related art described above, a hepatitis B therapeutic vaccine with better therapeutic efficacy has become an urgent issue to be solved in the art.

SUMMARY OF THE DISCLOSURE

In response to the above-referenced technical inadequacies, the present disclosure provides a hepatitis B therapeutic vaccine based on inactivated whole recombinant Hansenula polymorpha (H. polymorpha) cells, which expresses HBsAg

and HBcAg. The hepatitis B therapeutic vaccine uses the HBsAg and HBcAg expressed by the inactivated whole recombinant H. polymorpha cells as antigen. The inactivated whole recombinant H. polymorpha cell includes 19 HBsAg-specific CTL epitopes and 19 HBcAg-specific CTL epitopes, and the hepatitis B therapeutic vaccine uses the inactivated whole recombinant H. polymorpha cells as an adjuvant.

In certain embodiments, the HBsAg is adw subtype, and the DNA sequence of the HBsAg expressed by the recombinant H. polymorpha is shown as SEQ ID NO: 1, and the amino acid sequence of the HBsAg is shown as SEQ ID NO: 2.

In certain embodiments, the DNA sequence of the HBcAg expressed by the recombinant H. polymorpha is shown as SEQ ID NO: 3, and the amino acid sequence of the HBcAg is shown as SEQ ID NO: 4.

In certain embodiments, the HBsAg expressed by the recombinant H. polymorpha is a virus-like particle structure, which is formed by inserting HBsAgs into H. polymorpha lipid, 9 to 12 among the 14 cysteic acids of the HBsAg form disulfide bonds, and the HBcAg expressed by the recombinant H. polymorpha is a virus-like particle structure.

In certain embodiments, the conditions of inactivation of the inactivated whole recombinant H. polymorpha are: inactivation temperature from 52°C to 56°C, and inactivation time from 1 hour to 3 hours.

In certain embodiments, the HBsAg expressed by the inactivated whole recombinant H. polymorpha cells includes 19 CTL epitopes as follow: VLQAGFFLL, PFVQWFVGL, FLLTRILTI, WYWGPSLYSI, SLNFLGGSPV, FLGGSPVCL, LYSIVSPF, LYSIVSPFI, PFIPLLPIF, LLLCLIFLL, LLCLIFLLV, LLDYQGMLPV , LVLLDYQGML, VLLDYQGML, WLSLLVPFV, LLVPFVQWFV, GLSPTVWLSA, SIVSPFIPLL, and LLPIFFCLWV.

In certain embodiments, the HBcAg expressed by the inactivated whole recombinant H. polymorpha cells includes 19 CTL epitopes as follow: SFLPSDFF, FLPSDFFPSI, DFFPSIRDLL, FFPSIRDLL, SYVNVNMGL, SYVNVNMGLKI, YVNVNMG, YVNVNMGLK, WFHISCLTF, CLTFGRETV, VLEYLVSFGV, EYLVSFGVW, EYLVSFGVWI, AYRPPNAPI, AYRPPNAPIL, APILSTLPE,

ILSTLPETTV, STLPETTVVRR, and RGRSPRRRTP.

In certain embodiments, the dosage form of the hepatitis B therapeutic vaccine is selected from pre-filled injection solution, injection solution, and lyophilized powder injection.

In certain embodiments, the hepatitis B therapeutic vaccine further includes HBsAg stock solution or aluminum adjuvant HBsAg.

In one aspect, the present disclosure provides a recombinant H. polymorpha, which includes a DNA sequence of SEQ ID NO: 1. Preferably, the DNA sequence of SEQ ID NO: 1 is integrated into the genome of the recombinant H. polymorpha.

In one aspect, the present disclosure provides a recombinant H. polymorpha, which includes a DNA sequence of SEQ ID NO: 3. Preferably, the DNA sequence of SEQ ID NO: 3 is integrated into the genome of the recombinant H. polymorpha.

In certain embodiments, the host H. polymorpha cell line of the recombinant H. polymorpha is HU-11, and the accession number is CGMCC No.1218, and the disrupted DNA sequence of the orphanin-5-phosphate decarboxylase gene of the host H. polymorpha is shown in SEQ ID NO: 5.

The hepatitis B therapeutic vaccine of the present disclosure, based on the recombinant H. polymorpha contains 6-10 μg of HBsAg per 10^8 cells, the injection amount of HBsAg can be maximized and the amount of HBsAg injected can be maximized within the upper limit of the human injection of the existing recombinant H. polymorpha cells as an adjuvant, so as to enhance the reversal of immune tolerance status in patients with hepatitis B. The heat-inactivated whole recombinant H. polymorpha cells is the efficiently agonist of the Dectin-1 receptor of dendritic cells (DC, which are the most efficient antigen-presenting cells). In addition, the HBsAg expressed by recombinant H. polymorpha cells of the present disclosure has 19 specific CTL epitopes, and CTLs cells target HBV-infected hepatocytes and release IFN-γ: In the first step is to reduce the pool of cccDNA molecules more than 90% without hepatocyte damage, and the second step is to improve the process of destroying infected hepatocytes and trigger HBV immune reversion. In certain embodiments, the immunogenicity and reactivity of the preferred HBsAg are further

improved by expressing the DNA sequence (SEQ ID NO: 1) of preferably HBsAg, preferably 19 CTL epitopes in 21 CTLs. Moreover, the present disclosure is also preferred the optimized heat-inactivation process of the recombinant H. polymorpha cells, thereby ensuring the efficiency and safety of the vaccine.

While using the heat-inactivated recombinant H. polymorpha cells which expressing HBcAg and the heat-inactivated recombinant H. polymorpha cells which expressing HBsAg which were matched, the preferred code gene of the selected HBcAg (high immunoreactivity) was compared with the C2 genotype and the subtype double representative sequence AF100309.1, deletion of the 59st to 69st: ILCWGELMNLA a total of 11 amino acids. According cryo image reconstruction technique to determine the structure of HBcAg particles, HBcAg dimers each comprising four α helices, that is, each HBcAg subunit formed a dimer interface through two long α helices, the head and tail of amino acids at the 51st to 78st of the long α helix were restricted by the 50st and 79st of the conservative Pro, deeply buried inside the dimer, and not in the CTL epitope. The α helix is the so-called 3.6 helix, where each amino acid residue corresponds to a 100° turn in the helix. After the deletion of 59st to 69st bases: 11 amino acids of ILCWGELMNLA, the α helix which originally formed 5 rotations by the 51st to 78st, a total of 18 amino acids became the remaining 7 amino acids to form an α helix 2 rotations, while the other long α helix that formed the interface of the HBcAg dimer, the knot-like α helix at 82st-110st, ending at the 111st, Gly has not changed. Therefore, this present disclosure basically did not change the two long alpha helices to form a dimer interface, and the above-mentioned 11 amino acids deletion of HBcAg can still assemble into a virus-like particles (VLP) in recombinant H. polymorpha cells, and maintaining thermal stability.

The results of immunoreactivity assay showed that the immunoreactivity of the recombinant H. polymorpha HBcAg engineering strain with amino acids deletion (HC-40-25, 172 amino acids) was more than three times greater than the immunoreactivity of the recombinant H. polymorpha HBcAg engineering strain with full-length amino acids (HC-43, 183 amino acids). Based on the preferred CTL

epitopes, expression sequences and recombinant H. polymorpha engineering strains, the present disclosure also optimizes the heat inactivation process of recombinant H. polymorpha cells to ensure the efficacy and safety of the vaccine.

It is expected that the hepatitis B therapeutic vaccine based on the heat-inactivated whole recombinant H. polymorpha cells expressing HBsAg and HBcAg provided by the present invention will have greater immunogenicity and can be better used for the treatment of hepatitis B.

These and other aspects of the present disclosure will become apparent from the following description of the embodiment taken in conjunction with the following drawings and their captions, although variations and modifications therein may be affected without departing from the spirit and scope of the novel concepts of the disclosure.

BRIEF DESCRIPTION OF THE DRAWINGS

The present disclosure will become more fully understood from the detailed description and the accompanying drawings, in which:

FIG. 1 is a schematic view showing the construction process of plasmid pMPT-HBS-adw;

FIG. 2 is a physical map of plasmid pMPT-HBC;

FIG. 3 is an electrophoresis photograph of a PCR amplification product of the engineering strain HS-604-5 obtained by screening;

FIG. 4 is an electron micrograph of the pure stock solution of recombinant H. polymorpha recombinant HBsAg; and

FIG. 5 is a flow chart showing the steps of transformation and screening of recombinant H. polymorpha in the second embodiment of the recombinant H. polymorpha recombinant HBsAg engineering strain.

DETAILED DESCRIPTION OF THE EXEMPLARY EMBODIMENTS

The present disclosure is more particularly described in the following examples that are intended as illustrative only since numerous modifications and variations

therein will be apparent to those skilled in the art. Like numbers in the drawings indicate like components throughout the views. As used in the description herein and throughout the claims that follow, unless the context clearly dictates otherwise, the meaning of “a”, “an”, and “the” includes plural reference, and the meaning of “in” includes “in” and “on”. Titles or subtitles can be used herein for the convenience of a reader, which shall have no influence on the scope of the present disclosure.

The terms used herein generally have their ordinary meanings in the art. In the case of conflict, the present document, including any definitions given herein, will prevail. The same thing can be expressed in more than one way. Alternative language and synonyms can be used for any term(s) discussed herein, and no special significance is to be placed upon whether a term is elaborated or discussed herein. A recital of one or more synonyms does not exclude the use of other synonyms. The use of examples anywhere in this specification including examples of any terms is illustrative only, and in no way limits the scope and meaning of the present disclosure or of any exemplified term. Likewise, the present disclosure is not limited to various embodiments given herein. Numbering terms such as “first”, “second” or “third” can be used to describe various components, signals or the like, which are for distinguishing one component/signal from another one only, and are not intended to, nor should be construed to impose any substantive limitations on the components, signals or the like.

Development of the construction of the H. polymorpha intracellular plasmid pMPT-02:

The H. polymorpha expression system includes two main components:

(1) A vector system (plasmid) that initiates efficient expression of a foreign gene; and (2) a host cell having a specific selection marsker.

1.5 kb H. polymorpha MOX (methanol oxidase) promoter, 350 bp H. polymorpha MOX (methanol oxidase) terminator, 1.0 kb H. polymorpha autonomous replication sequence HARS, and 1.1 kb Saccharomyces cerevisiae uracil gene ScURA3 were tightly ligated by gene synthesis technology element, and then inserted

into the pBluescripII plasmid to construct a shuttle plasmid pMPT-02, which is the applicant's non-exclusive proprietary technology.

Development of the host cell using the uracil auxotrophic URA3-host cell line HU-11:

A recombinant H. polymorpha strain HU-11 (CGMCC No. 1218) in which the orotidine-5-phosphate decarboxylase gene (HURA3) was disrupted by homologous sequence-mediated homologous integration. Compared with the conventional auxotrophic host strains produced by mutagenesis, the recombinant H. polymorpha strain HU-11 has the characteristics of high genetic stability and low back mutation rate. It was convenient for genetic transformation and screening of recombinant strains, and maintains the wild-type strain. The physiological and biochemical characteristics were beneficial to the culture of recombinant strains and the high expression of foreign proteins, and have high industrial application value. The DNA sequencing result of the disrupted URA3 gene of the H. polymorpha host strain HU11 showed that the five bases of GAAGT were inserted into the 31st base. The insertion of five bases of GAAGT produces a frameshift mutation. The frameshift mutation resulted in a mutation in all of the 254 amino acid codes after the 11th position, and the mutation produced a total of 15 termination codes, indicating that the structural gene of URA3 is no longer re-expressible. The probability that the five bases GAAGT simultaneously produce a back reversion mutation was extremely small. The experimental test also proved that the back mutation rate of the host strain HU11 is zero. This low back-reversion mutation rate of the host strain was particularly advantageous for transformation screening. URA3-ogal deficiency host cell line HU-11 (CGMCC No. 1218) established by gene knock out technology was disclosed in the applicant's previously invention CN1651570A. The DNA sequence in which the disrupted decarboxylase gene (HURA3) is shown in SEQ ID NO: 5.

Preferred code gene design of recombinant H. polymorpha HBsAg-adw2:

The DNA sequence of HBsAg expression of the recombinant H. polymorpha

of the present disclosure is based on the HBsAg adw2 subtypes as shown in SEQ ID NO: 1. The amino acid sequence of the HBsAg is shown in SEQ ID NO: 2.

Construction of the H. polymorpha intracellular plasmids pMPT-HBS-adw and pMPT-HBC:

A synthetic nucleotide sequence according to the sequence of SEQ ID NO: 1 (hereinafter referred to as HBsAg adw2 gene) was constructed into a glycerol strain containing the HBsAg adw2 gene plasmid; the plasmid after correct sequencing was digested with EcoRI/BamHI, and then 701 bp DNA fragment was obtained.

The correct plasmid pMPT-02 was digested with EcoRI/BamHI, and the vector DNA obtained after the gelatinization was ligated to obtain the H. polymorpha intracellular plasmid pMPT-HBS-adw, and the plasmid pMPT- HBS-adw was transformed into *E. coli* Competent Cell JM109 (Code No. D9052), and then was cultured overnight by plating on. Single colonies were selected from the transformation plates, plasmid DNA was extracted and digested with EcoRI/BamHI, and the results of restriction enzyme digestion showed positive clones. Sequencing confirmed that the plasmid pMPT-HBS-adw was correct.

The HBsAg adw2 gene was inserted into the multiple cloning site of the H. polymorpha expression system intracellular plasmid pMPT-02: between EcoRI and BamHI. The full length of the plasmid pMPT-HBS-adw was 7665 bp. A schematic diagram of the construction process of plasmid pMPT-HBS-adw was shown in FIG. 1.

The plasmid pMPT-HBC based on the sequence shown in SEQ ID NO: 3 was constructed in the same method as in the construction of plasmid pMPT-HBS-adw. The physical map of the pMPT-HBC adw2 plasmid was shown in FIG. 2.

Construction of the recombinant H. polymorpha HBsAg engineering strain and the recombinant H. polymorpha HBcAg engineering strain:

In order to construct the recombinant H. polymorpha hepatitis B virus surface antigen (HBsAg) adw2 subtype engineering strain, the cell electroporation technology developed by the applicant was applied. The RC pulse: amplitude 1500V, capacitance

22 μF, and time constant 3-5ms electric shock 1 time, adopted the pMPT-HBS-adw plasmid transformed into H. polymorpha cells of the HU-11 strain (CGMCC No. 1218) from which the URA3-gene was knocked out. The single colony transformants were picked up on the MD selection culture plate and transferred to the MD liquid medium for continuous subculture. The adw2 subtype HBsAg gene and the corresponding regulatory components were multi-copy and heterologously integrated into the host H. polymorpha cell chromosome. After a single colony of more than one thousand transformant single colonies, the following three steps were screened:

(1) Clonal strains with large single colonies and fast cell growth have a high probability of multiple copies.

(2) The PCR technique was used to compare the electrophoretic band luminance of the HBsAg gene and the single copy number MOX (methanol oxidase) gene, and the HBsAg gene copy number was determined semi-quantitatively.

(3) The expression level of HBsAg released after methanol-induced and shake flask culture for 72 hours was detected.

The application of PCR technology to transformants screening was a new creation of this application. The multiple copies of the foreign gene HBsAg are determined and heterologously integrated in the H. polymorpha chromosome, while the MOX gene in the H. polymorpha chromosome was intact and not destroyed. They all play an important role and show unique advantages of the H. polymorpha expression system. A pair of primers were designed to simultaneously amplify the MOX gene (single copy) and the heterologous integrated HBsAg foreign gene (multicopy) in the H. polymorpha chromosome. By comparing the brightness of the bands of the amplified product in agarose gel electrophoresis, it was possible to roughly determine whether the HBsAg gene was multiple copies. This method was used for the preliminary screening of multi-copy strains of engineered HBsAg gene. The amplified HBsAg fragment was 800 bp in length and the amplified MOX fragment was 2000 bp in length.

Design using primer sequences:

primer forward: 5‘-TCAAAAGCGGTATGTCCTTCCACGT-’3

primer reverse: 5‘-TACTGCTGCCAGTGCACGGTG-’3

PCR product agarose gel electrophoresis: the amplified product of HBsAg gene of engineering strain was about 800 bp, and the amplification product of H. polymorpha single copy gene MOX gene was about 2000 bp. The recombinant H. polymorpha hepatitis B virus surface antigen (HBsAg) engineering strain obtained by final screening is numbered as HS604-5.

Using the plasmid pMPT-HBC, the electrophoresis photograph of the PCR amplification product of the engineered strain obtained via the same method by screening was shown in FIG. 3, wherein 1 was Marker. The recombinant H. polymorpha HBcAg engineering strain obtained by final screening is numbered as HBC-40-25.

Determination of intracellular HBsAg VLP expression in recombinant H. polymorpha HBsAg adw2 subtype engineering strain fermentation broth:

10 μg of adw2 subtype HBsAg hepatitis B surface antigen lyophilized standard provided by Tiantan Biotechnology was diluted with diluent to dilute to 1024 ng/mL, 512 ng/mL, 256 ng/mL, 128 ng/mL, 64 ng/mL, 32 ng/mL, 16 ng. /mL, 8ng/mL, 4ng/mL, 2ng/mL, 0ng/mL (diluent) a total of 11 standard points, and using radioimmunoassay kit to detect HBsAg reaction.

The obtained engineering strain (No. HS604-5) was subjected to 30 liters of pilot fermentation (batch number 20150422), and 1 mL 10 $OD_{600\ nm}$ was sampled after 87 hours of fermentation. After disrupting the cells with glass beads (cell disruption rate: 65%), the 200-fold diluted sample and the standard were separately reacted in the radioimmunoassay kit at the same time, and the expression of HBsAg antigen obtained by the γ-counter auto-completed curve was 126.9 (ng/mL). Based on the above, the expression level of intracellular HBsAg antigen in recombinant H. polymorpha was calculated as:

126.96 (ng/mL) ×200 ÷10×4.0×10^7×65%/ mL=9.8 μg/10^8 cells

An electron micrograph of the recombinant HBsAg pure stock solution of recombinant H. polymorpha was shown in FIG. 4. The results showed that the high purity, high concentration and virus-like particle (VLP) structure of recombinant HBsAg were stable.

The expression of HBcAgVLP in the fermentation broth of recombinant H. polymorpha HBcAg engineering strain determined by the same method was 8-12 μg/10^8 cells.

Expression of virus-like particles (VLP) in recombinant H. polymorpha HBcAg cells

The central rule of modern molecular biology is: DNA → RNA → amino acid primary sequence → protein molecule high structure → protein molecular function; symbol → indicates: decision.

Different genotypes of hepatitis B virus core antigen HBcAg protein molecule is composed of 183 or 185 amino acid residues to form a primary sequence, which determines the secondary structure, tertiary structure and quaternary structure of HBcAg.

Experiments show that HBcAg isolated from isolated HBV particles, infected cells, recombinant E. coli or recombinant yeast were found to be assembled into two sizes of particles: The T=3 type consists of 180 identical protein subunits with a molecular weight of 2l kD (i.e. 90 homo-dimers), with a diameter of 30 nm, and 90 spikes on the 4 surfaces. The T=4 type consists of 240 identical protein subunits with a molecular weight of 2l kD (i.e. 120 homo-dimers), with a diameter of 34 nm, and 120 spikes on the surface.

The transmission electron micrograph of the crude extract of the cell suspension of HBC-40-25 engineering strain of the present disclosure clearly shows HBcAg virus-like particles.

The above-described recombinant H. polymorpha HBcAg engineering strain (HC-40-25, 172 amino acids) with lacking amino acid is compared with six strong

positive strains HBC-17 of recombinant H. polymorpha HBcAg (183 amino acids) with full-length amino acid. (1 OD_{600nm}/ml cells induced by methanol culture for 3 days, disrupted by glass beads, 1000-fold diluted sample), and the immunogenicity (unit: NCU/mL) was tested as follows:

HBC-40-25	HBC-17	HBC-31	HBC-36	HBC-39	HBC-41	HBC-51
35.21	5.83	5.64	5.64	6.34	8.99	5.75

The immunoreactivity of HBC-40-25 strain was more than three times that of the other six strong positive strains.

Optimization of heat-inactivated recombinant H. polymorpha HBsAg and HBcAg cell conditions:

In order to determine the optimal conditions for the inactivated recombinant H. polymorpha, the following requirements should be met: (1) Reduce the survival rate of the inactivated recombinant H. polymorpha less than 5%. (2) Maintain a complete cellular structure to avoid leakage of heat-inactivated recombinant H. polymorpha intracellular antigenic substance. (3) Maintain the thermal stabilities of HBsAg virus-like particles (VLP) and HBcAg virus-like particles (VLP) expressed in recombinant H. polymorpha cells, so as to avoid the antigenicity decline. The above three requirements are the main condition of the production for the heat-inactivated recombinant H. polymorpha expressing HBsAg and HBcAg, and would provide the basis for the development of vaccine manufacturing and verification procedures. In addition, the thermal stability of HBsAg virus-like particle (VLP) expressed in recombinant H. polymorpha cells is the first issue to be solved. For this purpose, 16 sets of different temperature (50°C, 52°C, 54°C, 56°C and 58°C), and different time (1hr, 2hr ,and 3hr) of the heat-inactivation test of recombinant HBsAg H. polymorpha cells were designed and set at 20°C as the control group. During the detection: as the heat inactivation temperature and time increase, the solubility of the outer layer of the cell wall increases, the cell breakage rate increases, and the intracellular HBsAg VLP antigen reactivity peak multiplied at 52°C, 1 hr, while the intracellular HBcAg VLP antigen reactivity peak multiplied at 56°C, 1 hr. The extracellular HBsAg and HBcAg

antigens reactivity was extremely low during the temperature and time changes of the heat-inactivation assay, indicating that intracellular HBsAg VLP did not leak out, maintaining the heat-inactivated recombinant H. polymorpha cell structure. The survival rate of heat-inactivated cells was as low as 1/50,000 at 56 °C, 3 hr. Therefore, a basis for optimizing the conditions for heat inactivation of recombinant H. polymorpha cells was provided.

The HBsAg-specific and HBsAg-specific CTL cells trigger reversal of immunity without cell damage

The results of a series of studies on hepatitis B virus infection in chimpanzees indicated that, the HBV-specific CD8+T cells, which produce INF-γ and target to hepatocytes infected with HBV; in the first step is to reduce the pool of cccDNA molecules by more than 90% without cell damage, and the second step is to improve the process of destroying infected liver cells and trigger reversal of immunity.

Based on three experiments described CTL epitopes review, prediction and patented invention reported, HBV capsid antigen (Pre-S1-Pre-S2-HBsAg) has 23 CTL epitopes that did not repeat. The amino acid sequence of the HBsAg antigen of the present disclosure includes the following 19 CTL epitopes: VLQAGFFLL, PFVQWFVGL, FLLTRILTI, WYWGPSLYSI, SLNFLGGSPV, FLGGSPVCL, LYSIVSPF, LYSIVSPFI, PFIPLLPIF, LLLCLIFLL, LLCLIFLLV, LLDYQGMLPV , LVLLDYQGML, VLLDYQGML, WLSLLVPFV, LLVPFVQWFV, GLSPTVWLSA, SIVSPFIPLL, and LLPIFFCLWV.

The amino acid sequence of the HBcAg antigen of the present disclosure includes the following 19 CTL epitopes: SFLPSDFF, FLPSDFFPSI, DFFPSIRDLL, FFPSIRDLL, SYVNVNMGL, SYVNVNMGLKI, YVNVNMG, YVNVNMGLK, WFHISCLTF, CLTFGRETV, VLEYLVSFGV, EYLVSFGVW, EYLVSFGVWI, AYRPPNAPI, AYRPPNAPIL, APILSTLPE, ILSTLPETTV, STLPETTVVRR, and RGRSPRRRTP.

H. polymorpha recombinant hepatitis B vaccine product is preferred as

prefilled injection:

The routinely dispensed recombinant hepatitis B vaccine (yeast) pre-filled syringe was processed thermal stability test at 37°C for 45 days, and has proved that the relative in vitro relative efficacy (RP) of the vaccine met the requirements, while at the same storage condition, RP of routinely dispensed hepatitis B vaccine did not meet the requirements. Pre-filled syringe-packed recombinant hepatitis B vaccine (yeast) can be transported without refrigerated in a short time, stored and used.

The pre-filled syringe has 1 needle and 1 box, which is easy to use, easy to learn to use and essentials, a disposable syringe cannot be reused. Vaccination without a separate syringe can prevent from infection or infectious diseases spread caused by not completely sterilized glass syringe, the adverse effects caused by improper needle selection or the risk of disposable syringe being reused.

Full vaccination of pre-filled hepatitis B vaccine syringe has a good comprehensive cost-benefit ratio.

The following embodiments are intended to be illustrative, and not restrictive, and the scope of the present disclosure is not limited by the following embodiments.

First Embodiment

The pMPT-HBS-adw plasmid was constructed based on the sequence of SEQ ID NO: 1 (an expression vector comprising the sequence of the SEQ ID NO: 1). The construction of plasmid pMPT-HBS-adw includes the following steps:

The HBsAg adw2 gene was synthesized according to the DNA sequence of SEQ ID NO: 1; and the glycerol strain containing the HBsAg adw2 gene plasmid was constructed and named as MC407B-16.

The correctly sequenced MC407B-16 plasmid was digested with EcoRI/BamHI, and the digested product was used a TaKaRa PCR Fragment Recovery Kit (Code No. D301) to recover 701 bp DNA fragment called Inset DNA6.

The correct plasmid pMPT-02 was digested with EcoRI/BamHI, and the vector DNA obtained from the DNA recovery kit was called Vector DNA6.

Inset DNA6 was ligated to Vector DNA6 by using Solution of the TaKaRa DNA

Ligation Kit (Code No. D6022), and then thermally transformed into *E. coli* Competent Cell JM109 (Code No. D9052), and the cells were plated in the transformation plate and cultured overnight. Single colonies were selected from the transformation plate, and plasmid DNA was extracted. The plasmid DNA was digested with EcoRI/BamHI. The results showed that MC407A+B+C+D-77~80 were positive clones.

The plasmid MC407A+B+C+D-77 was sequenced respectively with primer RV-M, M13-47, MC407P1, MC407P2, MC407P3, MC407P4, MC407P5, MC407P6, MC407P7, MC407P8, MC407P9, MC407BF11, MC407BR11 to prove the plasmid pMPT-HBS-adw were correct.

Second Embodiment

Construction of a recombinant H. polymorpha HBsAg engineered strain.

Recombinant Hepatitis B Vaccine H. polymorpha transformation and screening illustration: The transformation and screening process of the recombinant H. polymorpha was shown in FIG. 5:

Specifically

1) The pMPT-HBS-adw plasmid was transformed into the URA3-auxotrophic H. polymorpha cell strain HU-11 (CGMCC No. 1218) of the host cell by cell electroporation. The culture medium was selected using a selection medium (MD liquid medium). The single colony transformants were picked up on the MD selection culture plate and transferred to the MD liquid medium for continuous subculture. The adw2 subtype HBsAg gene and the corresponding regulatory components were multi-copy and heterologously integrated into the host H. polymorpha cell chromosome.

2) Strain screening included the following steps

(1) Selecting a single colony of uracil prototrophic transformants

Colonies with rapid growth rate of bacteria were selected. PCR was used to detect the brightness of HBsAg gene bands. Colonies with a large number of copies were selected, and single colonies were shake-cultured in a selective medium, and successively subcultured for 20 to 400 generations;

(2) Screening multiple copies of heterologous integrated transformed clones

After subculture in step (1), and 72 hours of methanol-induced culture, the expression level of HBsAg released by the disruption of transformant cells was determined by radio immunoassay or radioimmunoassay (RIA);

(3) Screening out high-copy, high-expression clones of free plasmids

The clones screened by step (2) were cultured in YPD complete medium for 48 hours, and then transferred into a selection medium plate for cloning culture, and the HBsAg gene copy number was detected by quantitative PCR, and the expression level of HBsAg was detected by RIA.

(4) Based on the detection result of the step (3), the primary strain of the genetically stabilized recombinant H. polymorpha HBsAg engineering strain was selected.

The recombinant H. polymorpha HBcAg engineering strain was constructed and screened in the same method.

Third Embodiment 30 liters of pilot fermentation (The recombinant H. polymorpha HBcAg engineering and the recombinant strains of H. polymorpha HBsAg engineering strain fermentation process using the same method)

The main process:

1) Strain stored in liquid nitrogen was thawed by 200 ml seed medium, inoculated into the medium, divided into two 0.5L shake flasks, and cultured at 31°C for 22 hours as a first-class seed;

2) The primary seed was transferred into the secondary seed culture medium with 1600 ml seed medium, divided into six 1L shake flasks, and incubate at 31°C for 20 hours as a secondary seed;

3) 12L fermentation medium was adjusted to pH 5.5 and transferred into a 30L fermenter, and then the secondary seed was inoculated under growing at 30-31°C through two sources of glycerol and methanol; growth, de-repression and induction for the three phases, and co-culture 85 to 96 hours, the cells were harvested after 2-3 hours stopped induction. The frozen cells are homogenized.

Operation points:

(1) The feeding operation of the growth phase was going when the dissolved oxygen was consumed and the basal medium was consumed; the flow acceleration was gradually increased as the consumption of the basic medium increases, and the flow was added before 2-3 hours the dissolved oxygen was recovered.

(2) In the later stage of the growth phase, pay attention to the dissolved oxygen recovery, record the lowest value of dissolved oxygen, and start to flow when the dissolved oxygen rises to 70-80%, and enter the de-repression phase.

(3) After the later stage of the de-repression phase, the dissolved oxygen began to rise after the end of the flow. When the dissolved oxygen was raised to 70-80%, the methanol induction solution was added, and the methanol concentration is controlled at 3-5‰; the flow acceleration was controlled by the methanol detection flow controller.

(4) Stopping methanol addition before 2-3 hours the end of fermentation to reduce methanol residue during cell harvest.

Medium

1. Preparation of calcium chloride solution

11.33g $CaCl_2$ was accurately weighed and put it into a cleaned triangular flask, deionized water was appropriately added to dissolve and dilute to 200 ml.

2. Preparation of micro element solution

Accurately weighting the following reagents:

$(NH_4)_2Fe(SO_4)_2•6H_2O$	1000 mg
$CuSO_4•5H_2O$	80 mg
$ZnSO_4•7H_2O$	300 mg
$MnSO_4•H_2O$	400 mg
EDTA	1000 mg

The weighed reagent was placed in a cleaned triangular flask, dissolved in deionized water and dissolved to 200 ml.

3. Preparation of vitamin solution

Accurately weighting the following reagents:

d-Biotin	6 mg
Thiamin HCl	2000 mg

Biotin was first dissolved in 10 ml of 50% isopropanol, and then dissolved in Thiamin HCl, and then dissolved in an appropriate amount of deionized water to a volume of 100 ml.

4. Preparation of trace element solution

Accurately weighting the following reagents:

$NiSO_4 \cdot 6H_2O$	10 mg
$CoCl_2 \cdot 6H_2O$	10 mg
H_3BO_3	10 mg
$Na_2MoO_4 \cdot 2H_2O$	10 mg
KI	10 mg

The weighed reagent was placed in a cleaned triangular flask, and an appropriate amount of deionized solution was added to a volume of 50 ml.

The above four solutions were separately sterilized and filtered for use.

5. Preparation of seed salt solution

Accurately weighting the following reagents:

$NH_4H_2PO_4$	80 g
$MgSO_4 \cdot 7H_2O$	18 g
KCl	20 g
NaCl	2 g

The weighed reagent was placed in a cleaned triangular flask, dissolved in deionized water and dissolved to a volume of 1600 ml.

6. 27 g of glycerin was weighted in a 2000 ml flask, mixed with a salt solution of 360 mL, and made up to 1800 ml with deionized water. The same amount was dispensed into two 2000 ml flasks, and autoclaved at 110℃ for 30 minutes.

Two empty 500 ml triangle bottles, six 1000ml triangle bottles, a 100 ml graduated cylinder and a 500 ml graduated cylinder all were sterilized under 110℃, 30 minutes high pressure steam.

7. Primary seed medium

In the clean bench, 100 ml of each sterilized glycerin solution was taken aseptically, and added separately into two 500 ml sterilized flasks, and respectively added the following:

Calcium chloride solution	1 ml
Micro element solution	1 ml
Vitamin solution	0.5 ml
Trace element solution	0.25 ml

Shaking the above solution.

8. Secondary seed medium

1600 ml of sterilized glycerin solution was placed in a clean bench with sterile operation technique and placed in a 2000 ml sterilized triangle, and separately added:

Calcium chloride solution	16 ml
Micro element solution	16 ml
Vitamin solution	8 ml
Trace element solution	4 ml

9. Fermentation medium

The following reagents were accurately weighted and dissolved in 2000 ml of deionized water.

$NH_4H_2PO_4$	175 g
$MgSO_4 \cdot 7H_2O$	40 g
KCl	44 g
NaCl	4.4 g

520 g glycerin was weighted and added into a small 500 ml beaker. 10 ml defoamer was added into the beaker to sterilize, and then added:

Calcium chloride solution	175 ml
Micro element solution	175 ml
Vitamin solution	88 ml
Trace element solution	44 ml

10. Feed medium

87 g $NH_4H_2PO_4$, 260 g glycerin and 500 ml deionized water were added into

1000 ml flask, and then wrapped feed line and sterilized at 110°C for 30 minutes.

11. De-repression solution

1800 g glycerin and 660 ml deionized water were added into a 5000 ml flask, and then wrapped feed line and sterilized at 110°C for 30 minutes. 540 ml filter-sterilized salt solution was added after cooling.

12. Induction solution

400 ml glycerin was added into a 1000 ml flask, and then wrapped feed line and sterilized at 110°C for 30 minutes. 1600 ml methanol was added aseptically after cooling.

Fourth Embodiment

Purification

The recombinant H. polymorpha HBsAg engineering strain fermentation broth obtained from the third embodiment was harvested and the cells were washed. The detailed steps of purification can be found in References: Li Jin, Kong Yan. Recombinant Hepatitis B Vaccine Production Process. See Li Jin, Yu Yu, Dong Dexiang Editor: Biopharmaceutical Equipment And separation and purification techniques. 1st edition. Beijing: Chemical Industry Press, 2003: 348-349. The harvested cells can be crushed by a homogenizer to release HBsAg; the cell debris was removed by filtration with a 0.22 μm microporous filter; the small molecular impurities were removed by ultrafiltration with a 300K ultramicrofilter; and the HBsAg was extracted by silica gel adsorption treatment. Finally, it was purified by butyl agarose hydrophobic chromatography.

Fifth Embodiment

The optimal operating condition tests of the inactivated recombinant H. polymorpha cell.

In order to determine the optimal conditions for the inactivated recombinant H. polymorpha, the following requirements should be met:

(1) Reduce the survival rate of the inactivated recombinant H. polymorpha less

than 5%.

(2) Maintain a complete cellular structure to exert as an adjuvant with multi titer activity of the inactivated recombinant H. polymorpha.

(3) Maintain the virus-like particle (VLP) expressed in the inactivated recombinant H. polymorpha intact, so as to avoid the antigenicity decline.

The above three requirements are the main condition of the production for the inactivated recombinant H. polymorpha expressing HBsAg and HBcAg, and would provide the basis for the development of vaccine manufacturing and verification procedures. In addition, the thermal stability of the virus-like particle (VLP) expressed in the inactivated recombinant H. polymorpha is the first issue to be solved.

(1) Preparation of the inactivated recombinant H. polymorpha cell with optimal operating condition.

After the H. polymorpha engineering strain (strain number HS604-5) was cultured by fermentation or shake flask induction, the cells were washed with phosphate buffered saline (PBS) for three times by centrifugal process, and suspended the H. polymorpha in PBS for the volume calculation. The cells were counted using $OD_{600\ nm}$, diluted to 10 OD_{600nm}/ml with PBS, 2 ml per tube; each test group was provided with two test tubes which were disrupted group and not disrupted group, respectively; 16 test groups were required to prepare 32 tube sample tubes.

Place them in a set temperature water bath and thermally inactivate the recombinant H. polymorpha for a set time.

The inactivated recombinant H. polymorpha should be cultured for 3 days at 37°C in a chloramphenicol complete medium agar dish, and the survival rate was counted. The inactivated H. polymorpha is stored at 4°C for further use.

(2) H. polymorpha recombinant HBsAg (HS604-5 strain) cell heat-inactivation test group was established:

20°C room temperature group 1tube (control)

50°C	1 hour	2 hours	3 hours
52°C	1 hour	2 hours	3 hours
54°C	1 hour	2 hours	3 hours

56℃	1 hour	2 hours	3 hours
58℃	1 hour	2 hours	3 hours

Total is 16 test groups. After inactivation, the HBsAg antigen activity was detected by radioimmunoassay HBsAg reagent; 1:100 dilutions and 1:1000 dilutions, and double tubes were set. The test results were used to analyze and determine the optimal process conditions for the inactivation of H. polymorpha in this new hepatitis B vaccine.

(2) The heat-inactivation test groups of the recombinant H. polymorpha HBcAg engineering strain were established:

20°C room temperature group 1tube (control)

50℃	1 hour	2 hours	3 hours
52℃	1 hour	2 hours	3 hours
54℃	1 hour	2 hours	3 hours
56℃	1 hour	2 hours	3 hours
58℃	1 hour	2 hours	3 hours

Total is 16 test groups. After heat-inactivation, the HBcAg antigen activity was detected by radioimmunoassay HBcAg reagent; 1:100 dilutions and 1:1000 dilutions, and double tubes were set. The test results were used to analyze and determine the optimal process conditions for the inactivation of H. polymorpha in this new hepatitis B vaccine.

References

1. Qi Xiaoqiu, etc., the national population of hepatitis B virus epidemiology investigation report, the first edition of April 2011, People's Health Publishing House.

2. Bowen DG et.al,Intrahepatic immunity: a tale of two sites? Bowen DG et.al, Trends Immunol. 2005, 26(10):512-7.

3. Thomas H. King1,et.al,A Whole Recombinant Yeast-Based Therapeutic Vaccine Elicits HBV X, S and Core Specific T Cells in Mice and Activates Human T Cells Recognizing Epitopes Linked to Viral Clearance, 2014, POLS.

4. Haibin Huang et.al, Robust Stimulation of Humoral and Cellular Immune

Responses following Vaccination with Antigen-Loaded β-Glucan Particles, 2010, MBio.asm.org, 1(3): 1-7.

5. Robert Thimme et.al, CD8+ T Cells Mediate Viral Clearance and Disease Pathogenesis during Acute Hepatitis B Virus Infection, JOURNAL OF VIROLOGY, 2003, p.68–76.

6. Stefan F. Wieland et.al,Expansion and contraction of the hepatitis B virus transcriptional template in infected chimpanzees. Proc Natl Acad Sci U S A. 2004 Feb 17; 101(7): 2129–2134.

7. John M. Murray et.al,Dynamics of hepatitis B virus clearance in chimpanzees, 2005 Dec 6；Proc Natl Acad Sci U S A. 102(49): 17780–17785.

8. Thimme R**et.al**,CD8(+) T cells mediate viral clearance and disease pathogenesis during acute hepatitis B virus infection； J Virol. 2003 Jan; 77 (1):68-76.

9. Zeng Zhu-tian, Liver-induced systemic immune tolerance and its reversal, Ph.D thesis, the University of Science and Technology of China, 2014.

10. Applicant: Fudan University, Vaccine for controlling persistent infection of hepatitis B virus, 2009, Publication No. CN102038948A.

11. Florian K Bihl et.al**,** Simultaneous assessment of cytotoxic T lymphocyte responses against multiple viral infections by combined usage of optimal epitope matrices, anti- CD3 mAb T-cell expansion and "RecycleSpot"; Journal of Translational Medicine 2005, 3: 20 1-19.

12. Yuji Sobao et.al, Identification of hepatitis B virus-specific CTL epitopes presented by HLA-A*2402, the most common HLA class I allele in East Asia, Journal of Hepatology, 2001, 34: 922-929.

16. Applicant: Yuzhang Wu et al., Immunogen for preparation of therapeutic vaccines or drugs for treatment of hepatitis B and the producing method and use thereof, Publication No.CN1483736A.

17. Deng Xiaoyan, Research on Human Genome-wide Hepatitis B Virus Gene (Sub) Type Recombinant, Ph.D thesis of Chongqing Medical University, May 2012.

The foregoing description of the exemplary embodiments of the disclosure has been presented only for the purposes of illustration and description and is not intended to be exhaustive or to limit the disclosure to the precise forms disclosed. Many modifications and variations are possible in light of the above teaching.

The embodiments were chosen and described in order to explain the principles of the disclosure and their practical application so as to enable others skilled in the art to utilize the disclosure and various embodiments and with various modifications as are suited to the particular use contemplated. Alternative embodiments will become apparent to those skilled in the art to which the present disclosure pertains without departing from its spirit and scope.

CLAIMS

What is claimed is:

1. A hepatitis B therapeutic vaccine based on inactivated whole recombinant Hansenula polymorpha cells, which expresses HBsAg and HBcAg, wherein the amino acid sequence of the HBcAg expressed by the inactivated whole recombinant Hansenula polymorpha cells is shown as SEQ ID NO: 4, the DNA sequence of the HBcAg expressed by the inactivated whole recombinant Hansenula polymorpha cells is shown as SEQ ID NO: 3, the recombinant Hansenula polymorpha cell comprises 15-21 HBcAg-specific CTL epitopes, the intracellular expression level of the HBsAg in the inactivated whole recombinant Hansenula polymorpha cells is 6-10 μg HBsAg per 10^8 cells, the HBsAg has 16-21 HBsAg-specific CTL epitopes, and the hepatitis B therapeutic vaccine uses the inactivated whole recombinant Hansenula polymorpha cells as an adjuvant.

2. The hepatitis B therapeutic vaccine according to claim 1, wherein the HBsAg expressed by the inactivated whole recombinant Hansenula polymorpha cells is adw subtype, and the DNA sequence of the HBsAg expressed by the inactivated whole recombinant Hansenula polymorpha cells is shown as SEQ ID NO: 1.

3. The hepatitis B therapeutic vaccine according to claim 2, wherein the amino acid sequence of the HBsAg expressed by the inactivated whole recombinant Hansenula polymorpha cells is shown as SEQ ID NO: 2.

4. The hepatitis B therapeutic vaccine according to claim 1, wherein the HBsAg expressed by the inactivated whole recombinant Hansenula polymorpha cells is a virus-like particle structure, which is formed by inserting HBsAgs into Hansenula polymorpha lipid, 9 to 12 among the 14 cysteic acids of the HBsAg

form disulfide bonds, and the HBcAg expressed by the inactivated whole recombinant Hansenula polymorpha cells is a virus-like particle structure.

5. The hepatitis B therapeutic vaccine according to any one of claim 1 to 3, wherein the HBsAg expressed by the inactivated whole recombinant Hansenula polymorpha cells comprises 19 CTL epitopes as follow: VLQAGFFLL, PFVQWFVGL, FLLTRILTI, WYWGPSLYSI, SLNFLGGSPV, FLGGSPVCL, LYSIVSPF, LYSIVSPFI, PFIPLLPIF, LLLCLIFLL, LLCLIFLLV, LLDYQGMLPV , LVLLDYQGML, VLLDYQGML, WLSLLVPFV, LLVPFVQWFV, GLSPTVWLSA, SIVSPFIPLL, and LLPIFFCLWV.

6. The hepatitis B therapeutic vaccine according to any one of claim 1, 4, and 5, wherein the HBcAg expressed by the inactivated whole recombinant Hansenula polymorpha cells comprises 19 CTL epitopes as follow: SFLPSDFF, FLPSDFFPSI, DFFPSIRDLL, FFPSIRDLL, SYVNVNMGL, SYVNVNMGLKI, YVNVNMG, YVNVNMGLK, WFHISCLTF, CLTFGRETV, VLEYLVSFGV, EYLVSFGVW, EYLVSFGVWI, AYRPPNAPI, AYRPPNAPIL, APILSTLPE, ILSTLPETTV, STLPETTVVRR, and RGRSPRRRTP.

7. The hepatitis B therapeutic vaccine according to claim 1, wherein the host Hansenula polymorpha cell line of the inactivated whole recombinant Hansenula polymorpha cells is HU-11, and the accession number is CGMCC No.1218, and the disrupted DNA sequence of the orphanin-5-phosphate decarboxylase gene of the host Hansenula polymorpha is shown in SEQ ID NO: 5.

8. The hepatitis B therapeutic vaccine according to claim 1, wherein the dosage form of the hepatitis B therapeutic vaccine is selected from pre-filled injection solution, injection solution, or lyophilized powder injection.

9. The hepatitis B therapeutic vaccine according to claim 1, the hepatitis B therapeutic vaccine further comprises HBsAg stock solution or aluminum adjuvant HBsAg.

10. A recombinant Hansenula polymorpha, wherein the recombinant Hansenula polymorpha comprises a DNA sequence of SEQ ID NO: 3.

11. The recombinant Hansenula polymorpha according to claim 12, wherein the DNA sequence of SEQ ID NO: 3 is integrated into the genome of the recombinant Hansenula polymorpha.