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CORE-GLYCOSYLATED HCV ENVELOPE PROTEINS

FIELD OF THE INVENTION

The present invention relates to the general field of recombinant protein expression, to diagnosis of HCV infection, to treatment or prevention of HCV infection and to the prognosing/monitoring of the clinical efficiency of treatment of an individual with chronic hepatitis, or the prognosing/ monitoring of the natural disease.

More particularly, the present invention relates to the expression of hepatitis C virus envelope proteins in yeast, yeast strains for the expression of core-glycosylated viral envelope proteins, and the use in diagnosis, prophylaxis or therapy of HCV envelope proteins according to the present invention,

BACKGROUND OF THE INVENTION

Hepatitis C virus (HCV) infection is a major health problem in both developed and developing countries. It is estimated that about 1 to 5 % of the world population is affected by the virus. HCV infection appears to be the most important cause of transfusion-associated hepatitis and frequently progresses to chronic liver damage. Moreover, evidence exists implicating HCV in induction of hepatocellular carcinoma. Consequently, the demand for reliable diagnostic methods and effective therapeutic agents is high. Also sensitive and specific screening methods of HCV-contaminated blood-products and improved methods to culture HCV are needed.

HCV is a positive stranded RNA virus of approximately 9,600 bases which encode a single polyprotein precursor of about 3000 amino acids. Proteolytic cleavage of the precursor coupled to co- and posttranslational modifications has been shown to result in at least three structural and six non-structural proteins. Based on sequence homology, the structural proteins have been functionally assigned as one single core protein and two envelope glycoproteins: E1 and E2. The E1 protein consists of 192 amino acids and contains 4 to 5 N-glycosylation sites, depending on the HCV genotype. The E2 protein consists of 363 to 370 amino acids and contains 9 to 11 N-glycosylation sites, depending on the HCV genotype (for

reviews see: Major and Feinstone, 1997; Maertens and Stuyver, 1997). The E1 protein contains various variable domains (Maertens and Stuyver, 1997). The E2 protein contains three hypervariable domains, of which the major domain is located at the N-terminus of the protein (Maertens and Stuyver, 1997). The HCV glycoproteins localize predominantly in the ER where they are modified and assembled into oligomeric complexes.

In eukaryotes, sugar residues are commonly linked to four different amino acid residues. These amino acid residues are classified as O-linked (serine, threonine, and hydroxylysine) and N-linked (asparagine). The O-linked sugars are synthesized in the Golgi or rough Endoplasmic Reticulum (ER) from nucleotide sugars. The N-linked sugars are synthesized from a common precursor, and subsequently processed. It is believed that HCV envelope proteins are N-glycosylated. It is known in the art that addition of N-linked carbohydrate chains is important for stabilization of folding intermediates and thus for efficient folding, prevention of malfolding and degradation in the endoplasmic reticulum, oligomerization, biological activity, and transport of glycoproteins (see reviews by Rose et al., 1988; Doms et al., 1993; Helenius, 1994). The tripeptide sequences Asn-X-Ser and Asn-X-Thr (in which X can be any amino acid) on polypeptides are the consensus sites for binding N-linked oligosaccharides. After addition of the N-linked oligosaccharide to the polypeptide, the oligosaccharide is further processed into the complex type (containing N-acetylglucosamine, mannose, fucose, galactose and sialic acid) or the high-mannose type (containing N-acetylglucosamine and mannose). HCV envelope proteins are believed to be of the high-mannose type. N-linked oligosaccharide processing in yeast is very different from mammalian Golgi processing. In yeast the oligosaccharide chains are elongated in the Golgi through stepwise addition of mannose, leading to elaborate high mannose structures, referred to as hyperglycosylation. In contrast therewith, proteins expressed in prokaryotes are never glycosylated.

Patterns of high mannose-type glycosylation of proteins or peptides have been determined for a variety of eukaryotic cells. In mammalian cells, an average of 5 to 9 mannose units is linked to two N-acetylglucosamine moieties in a core-glycosylation-type oligosaccharide (the structure represented in short as Man(5-9)GlcNAc(2)). Core-glycosylation refers to a structure similar to the boxed structure in Figure 3 of Herscovics and Orleans (1993).

The methylotrophic yeast *Pichia pastoris* was reported to attach an average of 8 to 14 mannose units, i.e. Man(8-14)GlcNAc(2) per glycosylation site (Tschopp in EP0256421) and approximately 85% of the N-linked oligosaccharides are in the size range Man(8-14)GlcNAc(2) (Grinna and Tschopp 1989). Other researchers have published slightly

different oligosaccharide structures attached to heterologous proteins expressed in *P. pastoris*: Man(8-9)GlcNAc(2) (Montesino et al. 1998), Man(9-14)GlcNAc(2) or Man(9-15)GlcNAc(2) (Kalidas et al. 2001), and Man(8-18)GlcNAc(2) with a preponderence of Man(9-12)GlcNAc(2) and with the major overall oligosaccharide being Man(10)GlcNAc(2) (Miele et al. 1998). Trimble et al. (1991) reported an equal distribution of Man(8)GlcNAc(2) and Man(9)GlcNAc(2) in about 75% of the N-linked oligosaccharides with additionally 17% of the N-glycosylation sites being occupied by Man(10)GlcNAc(2) and the remaining 8% of the sites by Man(11)GlcNAc(2). Hyperglycosylation of a *P. pastoris*-expressed protein has been reported occasionally (Scorer et al. 1993).

Aspergillus niger is adding Man(5-10)GlcNAc(2) to N-glycosylation sites (Panchal and Wodzinski 1998).

The *Saccharomyces cerevisiae* glycosylation deficient mutant *mnn9* differs from wild-type *S. cerevisiae* in that *mnn9* cells produce glycosylated proteins with a modified oligosaccharide consisting of Man(9-13)GlcNAc(2) instead of hyperglycosylated proteins (Mackay et al. in US5,135,854 and Kniskern et al. in WO94/01132). Another *S. cerevisiae* mutant, *och1mnn9*, was reported to add Man(8)GlcNAc(2) to N-glycosylation sites in proteins (Yoshifumi et al. JP06277086).

Characteristic for *S. cerevisiae* (wild-type and *mnn9* mutant) core oligosaccharides is the presence of terminal α1,3-linked mannose residues (Montesino et al. 1998). Oligosaccharides attached to N-glycosylation sites of proteins expressed in *P. pastoris* or *S. cerevisiae och1mnn1* are devoid of such terminal α1,3-linked mannoses (Gellissen et al. 2000). Terminal α1,3-linked mannoses are considered to be allergenic (Jenkins et al. 1996). Therefor, proteins carrying on their oligosaccharides terminal α1,3-linked mannose residues are not suitable for diagnostic or therapeutic purposes.

The glycosylation pattern on proteins expressed in the methylotrophic yeast *Hansenula polymorpha* have, despite the use of this yeast for production of a considerable number of heterologous proteins (see Table 3 in Gellissen et al. 2000), not been studied in great detail. From the experiments of Janowicz et al. (1991) and Diminsky et al. (1997), it seems that *H. polymorpha* is not or only poorly glycosylating the large or small hepatitis B viral surface antigen (HBsAg). Most likely this is due to the fact that the HBsAg was expressed without signal peptide, thus preventing the produced HBsAg to enter the lumen of the endoplasmic reticulum (ER) and glycosylation. Limited addition of mono- or dihexoses to G-CSF (granulocyte colony stimulating factor) produced in *H. polymorpha* was reported

(Fischer et al. in WO00/40727). At the other hand, hyperglycosylation was observed of a heterologous α-galactosidase expressed in *H. polymorpha* cells (Fellinger et al. 1991).

To date, vaccination against disease has been proven to be the most cost effective and efficient method for controlling diseases. Despite promising results, efforts to develop an efficacious HCV vaccine, however, have been plagued with difficulties. A conditio sine qua non for vaccines is the induction of an immune response in patients. Consequently, HCV antigenic determinants should be identified, and administered to patients in a proper setting. Antigenic determinants can be divided in at least two forms, i.e. lineair and conformational epitopes. Conformational epitopes result from the folding of a molecule in a three-dimensional space, including co- and posttranslational modifications, such as glycosylation. In general, it is believed that conformational epitopes will realize the most efficacious vaccines, since they represent epitopes which resemble native-like HCV epitopes, and which may be better conserved than the actual linear amino acid sequence. Hence, the eventual degree of glycosylation of the HCV envelope proteins is of the utmost importance for generating native-like HCV antigenic determinants. However, there are seemingly insurmountable problems with culturing HCV, that result in only minute amounts of virions. In addition, there are vast problems with the expression and purification of recombinant proteins, that result in either low amounts of proteins, hyperglycosylated proteins, or proteins that are not glycosylated.

The HCV envelope proteins have been produced by recombinant techniques in *Escherichia coli*, insect cells, yeast cells and mammalian cells. However, expression in higher eukaryotes has been characterised by the difficulty of obtaining large amounts of antigens for eventual vaccine production. Expression in prokaryotes, such as *E. coli* results in HCV envelope proteins that are not glycosylated. Expression of HCV envelope proteins in yeast resulted in hyperglycosylation. As already demonstrated by Maertens et al. in WO 96/04385, the expression of HCV envelope protein E2 in *Saccharomyces cerevisiae* leads to proteins which are heavily glycosylated. This hyperglycosylation leads to shielding of protein epitopes. Although Mustilli et al. (1999) claims that expression of HCV E2 in *S. cerevisiae* results in core-glycosylation, the results of the intracellularly expressed material demonstrate that part of it is at least hyperglycosylated, while the correct processing of the remainder of this material has not been shown. Moreover, the hyperglycosylation observed by Mustilli et al. (1999) could only be prevented in the presence of tunicamycin, an inhibitor of glycosylation, and this does thus not reflect glycosylation occurring under normal, natural growth conditions. The need for HCV envelope proteins derived from an intracellular source

is well accepted (Maertens et al. in WO 96/04385, Heile et al., 2000). This need is further exemplified by the poor reactivity of the secreted yeast derived E2 with sera of chimpanzee immunized with mammalian cell culture derived E2 proteins as evidenced in Figure 5 of Mustilli et al (1999). This is further documented by Rosa et al (1996) who show that immunization with yeast derived HCV envelope proteins fails to protect from challenge.

Consequently, there is a need for efficient expression systems resulting in large and cost-effective amounts of proteins that at the same time have a native-like glycosylation pattern devoid of terminal α1,3-linked mannoses. In particular, such systems are needed for production of HCV envelope proteins.

SUMMARY OF THE INVENTION

A first aspect of the current invention relates to an isolated HCV envelope protein or a fragment thereof comprising at least one N-glycosylation site, said protein or fragment thereof characterized in that it is the product of expression in a eukaryotic cell and further characterized in that on average up to 80 % of the N-glycosylation sites are core-glycosylated. In particular, more than 70 % of said core-glycosylated sites are glycosylated with an oligomannose with a structure defined by Man(8 to 10)-GlcNAc(2). Furthermore, the ratio of the oligomannose with structure Man(7)-GlcNAc(2) over the oligomannose with structure Man(8)-GlcNAc(2) is less than or equal to 0.45. More specifically said oligomannoses contain less than 10 % terminal α1,3 mannose. The eukaryotic cell expressing said isolated HCV envelope protein or part thereof can be a yeast cell such as a *Hansenula* cell.

A further aspect of the current invention relates to an isolated HCV envelope protein or part thereof according to the invention which is derived from a protein comprising an avian lysozyme leader peptide or a functional variant thereof joined to said HCV envelope protein or fragment thereof. More specifically, said isolated HCV envelope protein or part thereof is derived from a protein characterized by the structure

$CL\text{-}[(A1)_a - (PS1)_b - (A2)_c]\text{-}HCVENV\text{-}[(A3)_d - (PS2)_e - (A4)_f]$

wherein:

CL is an avian lysozyme leader peptide or a functional equivalent thereof,

A1, A2, A3 and A4 are adaptor peptides which can be different or the same,
PS1 and PS2 are processing sites which can be the different or the same,
HCVENV is a HCV envelope protein or a part thereof,
a, b, c, d, e and f are 0 or 1, and
wherein, optionally, A1 and/or A2 are part of PS1 and/or wherein A3 and/or A4 are part of PS2.

Another aspect of the invention covers an isolated HCV envelope protein or fragment thereof according to the invention which is comprised in a structure chosen from the group consisting of monomers, homodimers, heterodimers, homo-oligomers and hetero-oligomers. Alternatively, said isolated HCV envelope protein or fragment thereof according to the invention which is comprised in a virus-like particle. More specifically, any of the isolated HCV envelope protein or fragment thereof according to the invention may comprise cysteines of which the cysteine thiol-groups are chemically modified.

Specific aspects of the invention relate to isolated HCV envelope proteins or fragment thereofs according to the invention which are antigenic or immunogenic and/or which comprise a T-cell stimulating epitope.

A further aspect relates to a composition comprising an isolated HCV envelope protein or fragment thereof according to the invention. Said composition may further comprise a pharmaceutically acceptable carrier and can be a medicament or a vaccine.

The invention also relates to a method for producing the isolated HCV envelope protein or fragment thereof according to the invention.

Another method of the invention is a method for the detection of the presence of anti-HCV antibodies in a sample suspected to comprise anti-HCV antibodies, said method comprising:

(i) contacting a HCV envelope protein or part thereof according to any of claims 1 to 15 with said sample under conditions allowing complexation of said HCV envelope protein or part thereof with said anti-HCV antibodies,
(ii) detecting the complex formed in (i), and
(iii) inferring from (ii) the presence of said anti-HCV antibodies in said sample.

More specifically, said method may comprise step (i) in which said contacting is occurring under competitive conditions. In particular, said methods may utilize a solid support to which said HCV envelope protein or part thereof is attached.

The invention further relates to a diagnostic kit for the detection of the presence of anti-HCV antibodies in a sample suspected to comprise anti-HCV antibodies, said kit comprising a HCV envelope protein or part thereof according to the invention. More specifically, said kit may comprise said HCV envelope protein or part thereof attached to a solid support.

The invention also relates to a medicament or a vaccine comprising a HCV envelope protein or part thereof according to the invention.

Also covered by the invention is a pharmaceutical composition for inducing a HCV-specific immune response in a mammal, said composition comprising an effective amount of a HCV envelope protein or part thereof according to the invention and, optionally, a pharmaceutically acceptable adjuvant. Said pharmaceutical composition may alternatively be capable of inducing HCV-specific antibodies in a mammal or of inducing a T-cell function in a mammal. Furthermore, said pharmaceutical composition may be a prophylactic composition or a therapeutic composition. In particular, said mammal is a human.

FIGURE LEGENDS

Figure 1. Schematic map of the vector pGEMT-E1sH6RB which has the sequence as defined in SEQ ID NO:6.

Figure 2. Schematic map of the vector pCHH-Hir which has the sequence as defined in SEQ ID NO:9.

Figure 3. Schematic map of the vector pFPMT121 which has the sequence as defined in SEQ ID NO:12.

Figure 4. Schematic map of the vector pFPMT-CHH-E1-H6 which has the sequence as defined in SEQ ID NO:13.

Figure 5. Schematic map of the vector pFPMT-MFa-E1-H6 which has the sequence as defined in SEQ ID NO:16.

Figure 6. Schematic map of the vector pUC18-FMD-MFa-E1-H6 which has the sequence as defined in SEQ ID NO:17.

Figure 7. Schematic map of the vector pUC18-FMD-CL-E1-H6 which has the sequence as defined in SEQ ID NO:20.

Figure 8. Schematic map of the vector pFPMT-CL-E1-H6 which has the sequence as defined in SEQ ID NO:21.

Figure 9. Schematic map of the vector pSP72E2H6 which has the sequence as defined in SEQ ID NO:22.

Figure 10. Schematic map of the vector pMPT121 which has the sequence as defined in SEQ ID NO:23.

Figure 11. Schematic map of the vector pFPMT-MFa-E2-H6 which has the sequence as defined in SEQ ID NO:24.

Figure 12. Schematic map of the vector pMPT-MFa-E2-H6 which has the sequence as defined in SEQ ID NO:25.

Figure 13. Schematic map of the vector pMF30 which has the sequence as defined in SEQ ID NO:28.

Figure 14. Schematic map of the vector pFPMT-CL-E2-H6 which has the sequence as defined in SEQ ID NO:32.

Figure 15. Schematic map of the vector pUC18-FMD-CL-E1 which has the sequence as defined in SEQ ID NO:35.

Figure 16. Schematic map of the vector pFPMT-CL-E1 which has the sequence as defined in SEQ ID NO:36.

Figure 17. Schematic map of the vector pUC18-FMD-CL-H6-E1-K-H6 which has the sequence as defined in SEQ ID NO:39.

Figure 18. Schematic map of the vector pFPMT-CL-H6-K-E1 which has the sequence as defined in SEQ ID NO:40.

Figure 19. Schematic map of the vector pYIG5 which has the sequence as defined in SEQ ID NO:41.

Figure 20. Schematic map of the vector pYIG5E1H6 which has the sequence as defined in SEQ ID NO:42.

Figure 21. Schematic map of the vector pSY1 which has the sequence as defined in SEQ ID NO:43.

Figure 22. Schematic map of the vector pSY1aMFE1sH6a which has the sequence as defined in SEQ ID NO:44.

Figure 23. Schematic map of the vector pBSK-E2sH6 which has the sequence as defined in SEQ ID NO:45.

Figure 24. Schematic map of the vector pYIG5HCCL-22aH6 which has the sequence as defined in SEQ ID NO:46.

Figure 25. Schematic map of the vector pYYIGSE2H6 which has the sequence as defined in SEQ ID NO:47.

Figure 26. Schematic map of the vector pYIG7 which has the sequence as defined in SEQ ID NO:48.

Figure 27. Schematic map of the vector pYIG7E1 which has the sequence as defined in SEQ ID NO:49.

Figure 28. Schematic map of the vector pSY1YIG7E1s which has the sequence as defined in SEQ ID NO:50.

Figure 29. Schematic map of the vector pPICZalphaA which has the sequence as defined in SEQ ID NO:51.

Figure 30. Schematic map of the vector pPICZalphaD' which has the sequence as defined in SEQ ID NO:52.

Figure 31. Schematic map of the vector pPICZalphaE' which has the sequence as defined in SEQ ID NO:53.

Figure 32. Schematic map of the vector pPICZalphaD'E1sH6 which has the sequence as defined in SEQ ID NO:58.

Figure 33. Schematic map of the vector pPICZalphaE'E1sH6 which has the sequence as defined in SEQ ID NO:59.

Figure 34. Schematic map of the vector pPICZalphaD'E2sH6 which has the sequence as defined in SEQ ID NO:60.

Figure 35. Schematic map of the vector pPICZalphaE'E2sH6 which has the sequence as defined in SEQ ID NO:61.

Figure 36. Schematic map of the vector pUC18MFa which has the sequence as defined in SEQ ID NO:62.

Figure 37. Elution profile of size exclusion chromatography of IMAC-purified E2-H6 protein expressed from the MFα-E2-H6-expressing *Hansenula polymorpha* (see Example 15). The X-axis indicates the elution volume (in mL). The vertical lines through the elution profile indicate the fractions collected. "P1"= pooled fractions 4 to 9, "P2"= pooled fractions 30 to 35, and "P3"= pooled fractions 37 to 44. The Y-axis indicates absorbance given in mAU (milli absorbance units). The X-axis indicates the elution volume in mL.

Figure 38. The different pools and fractions collected after size exclusion chromatography (see Figure 37) were analyzed by non-reducing SDS-PAGE followed by silver staining of the polyacrylamide gel. The analyzed pools ("P1", "P2", and "P3") and fractions (16 to 26) are indicated on top of the picture of the silver-stained gel. At the left (lane "M") are indicated the sizes of the molecular mass markers.

Figure 39. Fractions 17 to 23 of the size exclusion chromatographic step as shown in Figure 37 were pooled and alkylated. Thereafter, the protein material was subjected to Endo H treatment for deglycosylation. Untreated material and Endo H-treated material were separated on an SDS-PAGE gel and blotted to a PVDF membrane. The blot was stained with amido black.

Lane 1: Alkylated E2-H6 before Endo H-treatment

Lane 2: Alkylated E2-H6 after Endo H-treatment.

Figure 40. Western-blot analysis of cell lysates of E1 expressed in *Saccharomyces cerevisiae.* The Western-blot was developed using the E1-specific monoclonal antibody IGH 201.

Lanes 1-4: expression product after 2, 3, 5 or 7 days expression, respectively, in a *Saccharomyces* clone transformed with pSY1YIG7E1s (SEQ ID NO:50, Figure 28) comprising the nucleotide sequence encoding the chicken lysozyme leader peptide joined to E1-H6.

Lanes 5-7: expression product after 2, 3 or 5 days expression, respectively, in a *Saccharomyces* clone transformed with pSY1aMFE1sH6aYIG1 (SEQ ID NO:44, Figure 22) comprising the nucleotide sequence encoding the α-mating factor leader peptide joined to E1-H6.

Lane 8: molecular weight markers with sizes as indicated.

Lane 9: purified E1s produced by HCV-recombinant vaccinia virus-infected mammalian cells.

Figure 41. Analysis of the immobilized metal ion affinity chromatography (IMAC)-purified E2-H6 protein expressed by and processed from CL-E2-H6 to E2-H6 by *H. polymorpha* (see Example 17). Proteins in different wash fractions (lanes 2 to 4) and elution fractions (lanes 5 to 7) were analyzed by reducing SDS-PAGE followed by silver staining of the gel (**A**, top picture) or by western blot using using a specific monoclonal antibody directed against E2 (**B**, bottom picture). The sizes of the molecular mass markers are indicated at the left.

Figure 42. Elution profile of the first IMAC chromatography step on a Ni-IDA column (Chelating Sepharose FF loaded with Ni^{2+}, Pharmacia) for the purification of the sulfonated H6-K-E1 protein produced by *H. polymorpha* (see Example 18). The column was equilibrated with buffer A (50 mM phosphate, 6 M GuHCl, 1 % Empigen BB (v/v), pH 7.2) supplemented with 20 mM imidazole. After sample application, the column was washed sequentially with buffer A containing 20 mM and 50 mM imidazole, respectively (as indicated on chromatogram). A further washing and elution step of the His-tagged products was performed by the sequential application of buffer B (PBS, 1% empigen BB, pH 7.2) supplemented with 50 mM imidazole and 200 mM imidazole respectively (as indicated on chromatogram). Following fractions were pooled: the wash pool 1 (fractions 8 to 11, wash with 50 mM imidazole). The eluted material was collected as separate fractions 63 to 72 or an elution pool (fractions 63 to 69) was made. The Y-axis indicates absorbance given in mAU (milli absorbance units). The X-axis indicates the elution volume in mL

Figure 43. Analysis of the IMAC-purified H6-K-E1 protein (see Figure 42) expressed by and processed from CL-H6-K-E1 to H6-K-E1 by *H. polymorpha*. Proteins in the wash pool 1 (lane 12) and elution fractions 63 to 72 (lanes 2 to 11) were analyzed by reducing SDS-PAGE followed by silver staining of the gel (**A**, top picture). Proteins present in the sample before IMAC (lane 2), in the flow-through pool (lane 4), in wash pool 1 (lane 5) and in the elution pool (lane 6) were analyzed by western blot using a specific monoclonal antibody directed against E1 (**B**, bottom picture; no sample was loaded in lane 3). The sizes of the molecular mass markers (lanes M) are indicated at the left.

Figure 44. Elution profile of the second IMAC chromatography step on a Ni-IDA column (Chelating Sepharose FF loaded with Ni^{2+}, Pharmacia) for the purification of E1 resulting from the *in vitro* processing of H6-K-E1 (purification: see Figure 42) with Endo Lys-C. The

flow through was collected in different fractions (1 to 40) that were screened for the presence of E1s-products. The fractions (7 to 28), containing intact E1 processed from H6-K-E1 were pooled. The Y-axis indicates absorbance given in mAU (milli absorbance units). The X-axis indicates the elution volume in mL

Figure 45. Western-blot analysis indicating specific E1s proteins bands reacting with biotinylated heparin (see also Example 19). E1s preparations purified from HCV-recombinant vaccinia virus-infected mammalian cell culture or expressed by *H. polymorpha* were analyzed. The panel right from the vertical line shows a Western-blot developed with the biotinylated E1 specific monoclonal IGH 200. The panel left from the vertical line shows a Western-blot developed with biotinylated heparin. From these results it is concluded that mainly the lower-glycosylated E1s has high affinity for heparin.

Lanes M: molecular weight marker (molecular weights indicated at the left).

Lanes 1: E1s from mammalian cells and alkylated during isolation.

Lanes 2: E1s-H6 expressed by *H. polymorpha* and sulphonated during isolation.

Lanes 3: E1s-H6 expressed by *H. polymorpha* and alkylated during isolation.

Lanes 4: same material as loaded in lane 2 but treated with dithiotreitol to convert the sulphonated Cys-thiol groups to Cys-thiol.

Figure 46. Size exclusion chromatography (SEC) profile of the purified *H. polymorpha*-expressed E2-H6 in its sulphonated form, submitted to a run in PBS, 3% betain to force virus-like particle formation by exchange of Empigen BB for betain. The pooled fractions containing the VLPs used for further study are indicated by "↔". The Y-axis indicates absorbance given in mAU (milli absorbance units). The X-axis indicates the elution volume in mL. See also Example 20.

Figure 47. Size exclusion chromatography (SEC) profile of the purified *H. polymorpha*-expressed E2-H6 in its alkylated form, submitted to a run in PBS, 3% betain to force virus-like particle formation by exchange of Empigen BB for betain. The pooled fractions containing the VLPs are indicated by "↔". The Y-axis indicates absorbance given in mAU (milli absorbance units). The X-axis indicates the elution volume in mL. See also Example 20.

Figure 48. Size exclusion chromatography (SEC) profile of the purified *H. polymorpha*-expressed E1 in its sulphonated form, submitted to a run in PBS, 3% betain to force virus-like particle formation by exchange of Empigen BB for betain. The pooled fractions containing the VLPs are indicated by "↔". The Y-axis indicates absorbance given in mAU (milli absorbance units). The X-axis indicates the elution volume in mL. See also Example 20.

Figure 49. Size exclusion chromatography (SEC) profile of the purified *H. polymorpha*-expressed E1 in its alkylated form, submitted to a run in PBS, 3% betain to force virus-like particle formation by exchange of Empigen BB for betain. The pooled fractions containing the VLPs are indicated by "↔". The Y-axis indicates absorbance given in mAU (milli absorbance units). The X-axis indicates the elution volume in mL. See also Example 20.

Figure 50. SDS-PAGE (under reducing conditions) and western blot analysis of VLPs as isolated after size exclusion chromatography (SEC) as described in Figures 48 and 49. Left panel: silver-stained SDS-PAGE gel. Right panel: western blot using a specific monoclonal antibody directed against E1 (IGH201). Lanes 1: molecular weight markers (molecular weights indicated at the left); lanes 2: pool of VLPs containing sulphonated E1 (cfr. Figure 48); lanes 3: pool of VLPs containing alkylated E1 (cfr. Figure 49). See also Example 20.

Figure 51. E1 produced in mammalian cells ("M") or *Hansenula*-produced E1 ("H") were coated on a ELISA solid support to determine the end point titer of antibodies present in sera after vaccination of mice with E1 produced in mammalian cells (top panel), or after vaccination of mice with *Hansenula*-produced E1 (bottom panel). The horizontal bar represents the mean antibody titer. The end-point titers (fold-dilution) are indicated on the Y-axis. See also Example 22.

Figure 52. *Hansenula*-produced E1 was alkylated ("A") or sulphonated ("S") and coated on a ELISA solid support to determine the end point titer of antibodies present in sera after vaccination of mice with Hansenula-produced E1 that was alkylated (top panel), or after vaccination of mice with *Hansenula*-produced E1 that was sulphonated (bottom panel). The horizontal bar represents the mean antibody titer. The end-point titers (fold-dilution) are indicated on the Y-axis. See also Example 23.

Figure 53. HCV E1 produced by HCV-recombinant vaccinia virus-infected mammalian cells and HCV E1 produced by *H. polymorpha* were coated directly to ELISA plates. End point titers of antibodies were deteremined in sera of chimpanzees vaccinated with E1 produced by mammalian cells (top panel) and of murine monoclonal antibodies raised against E1 produced by mammalian cells (bottom panel). Chimpanzees Yoran and Marti were prophylactically vaccinated. Chimpanzees Ton, Phil, Marcel, Peggy and Femma were therapeutically vaccinated. Black filled bars: ELISA plate coated with E1 produced by mammalian cells. Open bars: ELISA plate coated with E1 produced by *Hansenula*. The end-point titers (fold-dilution) are indicated on the Y-axis. See also Example 24.

Figure 54. Fluorophore-assisted carbohydrate gelelectrophoresis of oligosaccharides released from E1 produced by recombinant vaccinia virus-infected mammalian cells and from E1-H6 protein produced by *Hansenula*.

Lane 1: Glucose ladder standard with indication at the left of the number of monosaccharides (3 to 10, indicated by G3 to G10).

Lane 2: 25 µg N-linked oligosaccharides released from (alkylated) E1 produced by mammalian cells.

Lane 3: 25 µg N- linked oligosaccharides released from (alkylated) E1-H6 produced by *Hansenula*.

Lane 4: 100 pmoles maltotetraose.

See also Example 25.

Figure 55. This figure shows the simplified structures of the reference oligomannoses Man-9 (Figure 55.A), Man-8 (Figure 55.B), Man-7 (Figure 55.C), Man-6 (Figure 55.D) and Man-5 (Figure 55.E). "Man" = mannose; "GlcNAc" = N-acetylglucosamine; "α" = α-linkage between 2 mannoses; "β" = β-linkage between 2 mannoses; "(1-3)", "(1-4)" and "(1-6)" = (1-3), (1-4) and (1-6) linkage between 2 mannoses, respectively. The brackets in Figure 55.B and 55.C indicate that the 2 and 1, respectively, mannose residue(s) to the left of the bracket are coupled in an α (1-2) bond to 2 and 1, respectively, of the 3 mannose residues right from the bracket. See also Example 26.

Figure 56. This figure shows a higher oligomannose consisting of 10 mannose moieties coupled to chitobiose. Each terminal mannose residue is linked by an α 1-3 bond to a non-terminal mannose residue. The thin upward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by α 1-2 Mannosidase (none for this oligomannose), the thick upward or leftward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by α Mannosidase after removal of the α 1-2-linked mannoses (not applicable to this oligomannose) and the empty downward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by β Mannosidase after removal of the α-linked mannoses. See also Example 26.

Figure 57. This figure shows a higher oligomannose consisting of 9 mannose moieties coupled to chitobiose. In this oligomannose, one terminal mannose residue is linked by an α 1-2 bond to the non-terminal mannose residue. The thin upward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by α 1-2 Mannosidase, the thick upward or leftward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by α Mannosidase after removal of the α 1-2-linked mannoses and the empty downward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by β Mannosidase after removal of the α-linked mannoses. See also Example 26.

Figure 58. This figure shows the reference higher oligomannose Man-9 consisting of 9 mannose moieties coupled to chitobiose. In this oligomannose, all terminal mannose residues are linked by an α 1-2 bond to a non-terminal mannose residue. The thin upward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by α 1-2 Mannosidase, the thick upward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by α Mannosidase after removal of the α 1-2-linked mannoses and the empty downward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by β Mannosidase after removal of the α-linked mannoses. See also Example 26.

Figure 59. This figure shows the higher oligomannose Man-8 consisting of 8 mannose moieties coupled to chitobiose. In this oligomannose, all terminal mannose residues are linked by an α 1-3 or an α 1-6 bond to a non-terminal mannose residue which renders this structure fully resistant to cleavage by α 1-2 Mannosidase. The thick upward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by α Mannosidase and the empty

downward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by β Mannosidase after removal of the α-linked mannoses. See also Example 26.

Figure 60. This figure shows the higher oligomannose Man-7 consisting of 7 mannose moieties coupled to chitobiose. In this oligomannose, all terminal mannose residues are linked by an α 1-3 bond to a non-terminal mannose residue which renders this structure fully resistant to cleavage by α 1-2 Mannosidase. The thick upward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by α Mannosidase and the empty downward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by β Mannosidase after removal of the α-linked mannoses. See also Example 26.

Figure 61. This figure shows a higher oligomannose consisting of 9 mannose moieties coupled to chitobiose. In this oligomannose, one terminal mannose residue is linked by an α 1-2 bond to the non-terminal mannose residue. The thin upward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by α 1-2 Mannosidase, the thick upward or leftward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by α Mannosidase after removal of the α 1-2-linked mannoses and the empty downward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by β Mannosidase after removal of the α-linked mannoses. See also Example 26.

Figure 62. This figure shows the putative glucose-containing oligosaccharide consisting of 1 or 2 glucose moieties and 8 mannose moieties coupled to chitobiose. In this oligosaccharide, either one of the terminal α 1-2-linked mannose residues of the A- or B-branch ("A →" and "B →" in the Figure) is carrying one or two glucose residues, as indicated by (Glc)Glc to the left of the of the bracket. The thin upward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by α 1-2 Mannosidase given that no glucose is attached to the terminal mannose residue. The thick upward or leftward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by α Mannosidase after removal of the α 1-2-linked mannoses and the empty downward pointing arrow indicates the oligosaccharide bonds which are prone to cleavage by β Mannosidase after removal of the α-linked mannoses. An overview of the possible reaction products is given in Table 10 of Example 26.

Figure 63. The reaction products of Man-9 after overnight incubation with or without exoglycosidases were separated on a TSK gel-Amide-80 (0.46 x 25 cm, Tosoh Biosep) column coupled to a Waters Alliance HPLC station. Separation of the oligosaccharides was carried out at ambient temperature at 1.0 mL/min. Solvent A consisted of 0.1% acetic acid in acetonitrile and solvent B consisted of 0.2 % acetic acid-0.2 % triethylamine in water. Separation of 2-AB labeled oligosaccharides was carried out using 28 % B isocratic for 5 column volumes followed by a linear increase to 45 % B over fifteen column volumes. The composition of the elution solvent is indicate on the right Y-axis as % solvent B in solvent A (v/v). The elution time is indicated in minutes on the X-axis. The left Y-axis indicates fluorescence of eluting 2-aminobenzamide (2-AB)-labeled oligosaccharides. The excitation wavelength of 2-AB is 330 nm, the emission wavelength 420 nm.

Trace 1 ("1") of the chromatogram shows the elution of Man-9 incubated overnight without exoglycosidases. Trace 2 ("2") shows the elution of a mixture of Man-5 and Man-6 after overnight incubation of Man-9 with α 1-2 Mannosidase. Traces 3 and 4 ("3" and "4") show the elution of 4'-β-mannosyl chitobiose after 1 h and overnight incubation, respectively, of Man-9 with α-Mannosidase. Trace 5 ("5") shows the elution of chitobiose after overnight incubation of Man-9 with α- and β-Mannosidase. Traces 1-5 are represented as overlays, as such their respective baselines are not all on the zero level. Trace 6 ("6") indicates the applied solvent gradient.

The peaks, when present, indicated by the letters A to K on top of the Figure represent: A, chitobiose; B, 4'-β-mannosyl-chitobiose; C, Man-2; D, Man-3; E, Man-4; F, Man-5; G, Man-6; H, Man-7; I, Man-7; J, Man-8; and K, Man-10. See also Example 26.

Figure 64. The reaction products of the oligosaccharides derived from *Saccharomyces*-produced E1s after overnight incubation with or without exoglycosidases were separated on a TSK gel-Amide-80 (0.46 x 25 cm, Tosoh Biosep) column coupled to a Waters Alliance HPLC station. Separation of the oligosaccharides was carried out at ambient temperature at 1.0 mL/min. Solvent A consisted of 0.1% acetic acid in acetonitrile and solvent B consisted of 0.2 % acetic acid-0.2 % triethylamine in water. Separation of 2-AB labeled oligosaccharides was carried out using 28 % B isocratic for 5 column volumes followed by a linear increase to 45 % B over fifteen column volumes. The composition of the elution solvent is indicate on the right Y-axis as % solvent B in solvent A (v/v). The elution time is indicated in minutes on the X-axis. The left Y-axis indicates fluorescence of eluting 2-

aminobenzamide (2-AB)-labeled oligosaccharides. The excitation wavelength of 2-AB is 330 nm, the emission wavelength 420 nm.

Trace 1 ("1") of the chromatogram shows the elution of oligosaccharides derived from *Saccharomyces*-produced E1s incubated overnight without exoglycosidases. Trace 2 ("2") shows the elution of oligosaccharides derived from *Saccharomyces*-produced E1s after overnight incubation of Man-9 with α 1-2 Mannosidase. Traces 3 and 4 ("3" and "4") show the elution of oligosaccharides derived from *Saccharomyces*-produced E1s after 1 h and overnight incubation, respectively, with α-Mannosidase. Trace 5 ("5") shows the elution of oligosaccharides derived from *Saccharomyces*-produced E1s after overnight incubation with α- and β-Mannosidase. Traces 1-5 are represented as overlays, as such their respective baselines are not all on the zero level. Trace 6 ("6") indicates the applied solvent gradient.

The peaks, when present, indicated by the letters A to K on top of the Figure represent: A, chitobiose; B, 4'-β-mannosyl-chitobiose; C, Man-2; D, Man-3; E, Man-4; F, Man-5; G, Man-6; H, Man-7; I, Man-7; J, Man-8; and K, Man-10. See also Example 26.

Figure 65. The reaction products of the oligosaccharides derived from E1s produced in vaccinia-transfected mammalian cells after overnight incubation with or without exoglycosidases were separated on a TSK gel-Amide-80 (0.46 x 25 cm, Tosoh Biosep) column coupled to a Waters Alliance HPLC station. Separation of the oligosaccharides was carried out at ambient temperature at 1.0 mL/min. Solvent A consisted of 0.1% acetic acid in acetonitrile and solvent B consisted of 0.2 % acetic acid-0.2 % triethylamine in water. Separation of 2-AB labeled oligosaccharides was carried out using 28 % B isocratic for 5 column volumes followed by a linear increase to 45 % B over fifteen column volumes. The composition of the elution solvent is indicate on the right Y-axis as % solvent B in solvent A (v/v). The elution time is indicated in minutes on the X-axis. The left Y-axis indicates fluorescence of eluting 2-aminobenzamide (2-AB)-labeled oligosaccharides. The excitation wavelength of 2-AB is 330 nm, the emission wavelength 420 nm.

Trace 1 ("1") of the chromatogram shows the elution of oligosaccharides derived from E1s produced in vaccinia-transfected mammalian cells incubated overnight without exoglycosidases. Trace 2 ("2") shows the elution of oligosaccharides derived from E1s produced in vaccinia-transfected mammalian cells after overnight incubation of Man-9 with α 1-2 Mannosidase. Traces 3 and 4 ("3" and "4") show the elution of oligosaccharides derived from E1s produced in vaccinia-transfected mammalian cells after 1 h and overnight

incubation, respectively, with α-Mannosidase. Trace 5 ("5") shows the elution of oligosaccharides derived from E1s produced in vaccinia-transfected mammalian cells after overnight incubation with α- and β-Mannosidase. Traces 1-5 are represented as overlays, as such their respective baselines are not all on the zero level. Trace 6 ("6") indicates the applied solvent gradient.

The peaks, when present, indicated by the letters A to K on top of the Figure represent: A, chitobiose; B, 4'-β-mannosyl-chitobiose; C, Man-2; D, Man-3; E, Man-4; F, Man-5; G, Man-6; H, Man-7; I, Man-7; J, Man-8; and K, Man-10. See also Example 26.

Figure 66. The reaction products of the oligosaccharides derived from *Hansenula*-produced E1s after overnight incubation with or without exoglycosidases were separated on a TSK gel-Amide-80 (0.46 x 25 cm, Tosoh Biosep) column coupled to a Waters Alliance HPLC station. Separation of the oligosaccharides was carried out at ambient temperature at 1.0 mL/min. Solvent A consisted of 0.1% acetic acid in acetonitrile and solvent B consisted of 0.2 % acetic acid-0.2 % triethylamine in water. Separation of 2-AB labeled oligosaccharides was carried out using 28 % B isocratic for 5 column volumes followed by a linear increase to 45 % B over fifteen column volumes. The composition of the elution solvent is indicate on the right Y-axis as % solvent B in solvent A (v/v). The elution time is indicated in minutes on the X-axis. The left Y-axis indicates fluorescence of eluting 2-aminobenzamide (2-AB)-labeled oligosaccharides. The excitation wavelength of 2-AB is 330 nm, the emission wavelength 420 nm.

Trace 1 ("1") of the chromatogram shows the elution of oligosaccharides derived from *Hansenula*-produced E1s incubated overnight without exoglycosidases. Trace 2 ("2") shows the elution of oligosaccharides derived from *Hansenula*-produced E1s after overnight incubation of Man-9 with α 1-2 Mannosidase. Traces 3 and 4 ("3" and "4") show the elution of oligosaccharides derived from *Hansenula*-produced E1s after overnight incubation with α-Mannosidase. Trace 5 ("5") shows the elution of oligosaccharides derived from *Hansenula* -produced E1s after overnight incubation with α- and β-Mannosidase. Traces 1-5 are represented as overlays, as such their respective baselines are not all on the zero level. Trace 6 ("6") indicates the applied solvent gradient.

The peaks, when present, indicated by the letters A to K on top of the Figure represent: A, chitobiose; B, 4'-β-mannosyl-chitobiose; C, Man-2; D, Man-3; E, Man-4; F, Man-5; G, Man-6; H, Man-7; I, Man-7; J, Man-8; and K, Man-10. See also Example 26.

Figure 67. SDS-PAGE analysis and Coomassie Brilliant Blue staining of E1 proteins produced by *Hansenula* and by a HCV-recombinant vaccinia virus-infected mammalian cells. Lane 1: molecular weight markers with molecular weights indicated at the left; Lane 2: alkylated E1s produced by *Hansenula polymorpha* (10 μg); Lane 3: alkylated E1s produced by *Hansenula polymorpha* (5 μg); Lane 4: alkylated E1s produced by *Hansenula polymorpha* (2.5 μg); Lane 5: alkylated E1s produced by HCV-recombinant vaccinia virus-infected vero cells (10 μg); Lane 6: alkylated E1s produced by HCV-recombinant vaccinia virus-infected vero cells (5 μg); Lane 7: alkylated E1s produced by HCV-recombinant vaccinia virus-infected vero cells (2.5 μg). See also Example 27.

Figure 68. Sequence of HCV E2-H6 protein (SEQ ID NO:5) with indication of tryptic fragments (boxed sequences) of the deglycosylated protein. The glycosylated Asn-residues are converted into Asp-residues by the PNGase F enzyme and are indicated with a "*" under the sequence. The Asn-residues are prone to proteolytic cleavage by the Asp-N endoproteinase. The possible N-glycosylation sites in E2-H6 (SEQ ID NO:5) are N_{417}, N_{423}, N_{430}, N_{448}, N_{478}, N_{532}, N_{540}, N_{556}, N_{576}, N_{623} and N_{645} according to the numbering in the HCV polyprotein; these sites are numbered N_{34}, N_{40}, N_{47}, N_{65}, N_{95}, N_{149}, N_{157}, N_{173}, N_{193}, N_{240} and N_{262} in this figure. See also Example 28.

DETAILED DESCRIPTION OF THE INVENTION

In work leading to the present invention, it was observed that expression of glycosylated HCV envelope proteins in *Saccharomyces cerevisiae*, *Pichia pastoris* and *Hansenula polymorpha* was possible by expression of said HCV envelope proteins as proteins comprising a signal peptide sequence joined to said HCV envelope proteins. The glycosylation patterns of the HCV envelope proteins expressed in these three yeast species were, however, very different (see Examples 6, 10, 13 and 25). More specifically the *S. cerevisiae* (glycosylation deficient mutant)- and *H. polymorpha*-expressed HCV envelope proteins were glycosylated in a manner resembling core-glycosylation. The HCV envelope proteins expressed in *Pichia pastoris* were hyperglycosylated despite earlier reports that proteins expressed in this yeast are normally not hyperglycosylated (Gellissen et al. 2000, Sugrue et al. 1997).

Upon further analysis of the glycosylation patterns of HCV proteins produced in *S. cerevisiae* (glycosylation deficient strain), *H. polymorpha* and in HCV-recombinant vaccinia virus-infected mammalian cells, it was surprisingly found that the *Hansenula*-produced HCV envelope proteins displayed a glycosylation pattern which is very advantageous for diagnostic, prophylactic and therapeutic application of these HCV envelope proteins (see Examples 21-24 and 26-29). This unexpected finding is reflected in the different aspects and embodiments of the present invention as presented below.

A first aspect of the invention is related to an isolated HCV envelope protein or a fragment thereof comprising at least one N-glycosylation site, said protein or fragment thereof characterized in that it is the product of expression in a eukaryotic cell and further characterized in that on average up to 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, or 80% of the N-glycosylated sites are core-glycosylated. More specific thereto, more than 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94% or 95 % of the N-glycosylated sites are glycosylated with an oligomannose with a structure defined by Man(8 to 10)-GlcNAc(2). More specific to any of the above N-glycosylation characteristics, the ratio of sites core-glycosylated with an oligomannose with structure Man(7)-GlcNAc(2) over the sites core-glycosylated with an oligomannose with structure Man(8)-GlcNAc(2) is less than or equal to

0.15, 0.2, 0.25, 0.30, 0.35, 0.40, 0.44, 0.45, or 0.50. Further more specific to any of the above N-glycosylation characteristics, said oligomannoses contain less than 20%, 19%, 18%, 17%, 16%, 15%, 14%, 13%, 12%, 11%, 10%, 9%, 8%, 7%, 6%, or 5% terminal α1,3 mannose.

With "N-glycosylated sites glycosylated with an oligomannose with a structure defined by Man(8 to 10)-GlcNAc(2)" is meant that said N-glycosylated sites are glycosylated with either one of Man(8)-GlcNAc(2), Man(9)-GlcNAc(2), or Man(10)-GlcNAc(2).

It will be clear that the same N-glycosylation site in two proteins may be occupied by a different oligomannose.

The term "protein" refers to a polymer of amino acids and does not refer to a specific length of the product; thus, peptides, oligopeptides, and polypeptides are included within the definition of protein. This term also does not refer to or exclude post-expression modifications of the protein, for example, glycosylations, acetylations, phosphorylations and the like. Included within the definition are, for example, polypeptides containing one or more analogues of an amino acid (including, for example, unnatural amino acids, PNA, etc.), polypeptides with substituted linkages, as well as other modifications known in the art, both naturally occurring and non-naturally occurring.

With "pre-pro-protein" or "pre-protein" is, when used herein, meant a protein comprising a pre-pro-sequence joined to a protein of interest or a protein comprising a pro-sequence joined to a protein of interest, respectively. As alternatives for "pre-sequence", the terms "signal sequence", "signal peptide", "leader peptide", or "leader sequence" are used; all refer to an amino acid sequence that targets a pre-protein to the rough endoplasmic reticulum (ER) which is a prerequisite for (N-)glycosylation. The "signal sequence", "signal peptide", "leader peptide", or "leader sequence" is cleaved off, i.e. "removed" from the protein comprising the signal sequence joined to a protein of interest, at the on the luminal side of this ER by host specific proteases referred to as signal peptidases. Likewise, a pre-pro-protein is converted to a pro-protein upon translocation to the lumen of the ER. Depending on the nature of the "pro" amino acid sequence, it can or can not be removed by the host cell expressing the pre-pro-protein. A well known pre-pro-amino acid sequence is the α mating factor pre-pro-sequence of the *S. cerevisiae* α mating factor.

With "HCV envelope protein" is meant a HCV E1 or HCV E2 envelope protein or a part thereof whereby said proteins may be derived from a HCV strain of any genotype. More specifically, HCVENV is chosen from the group of amino acid sequences consisting of SEQ

ID NOs:85 to 98, amino acid sequences which are at least 90% identical to SEQ ID NOs:85 to98, and fragments of any thereof. As "identical" amino acids are considered the groups of conserved amino acids as described above, i.e. the group consisting of Met, Ile, Leu and Val; the group consisting of Arg, Lys and His; the group consisting of Phe, Trp and Tyr; the group consisting of Asp and Glu; the group consisting of Asn and Gln; the group consisting of Cys, Ser and Thr; and the group consisting of Ala and Gly.

More specifically, the term "HCV envelope proteins" relates to a polypeptide or an analogue thereof (e.g. mimotopes) comprising an amino acid sequence (and/or amino acid analogues) defining at least one HCV epitope of either the E1 or the E2 region, in addition to a glycosylation site. These envelope proteins may be both monomeric, hetero-oligomeric or homo-oligomeric forms of recombinantly expressed envelope proteins. Typically, the sequences defining the epitope correspond to the amino acid sequences of either the E1 or the E2 region of HCV (either identically or via substitutions of analogues of the native amino acid residue that do not destroy the epitope).

It will be understood that the HCV epitope may co-locate with the glycosylation site.

In general, the epitope-defining sequence will be 3 or 4 amino acids in length, more typically, 5, 6, or 7 amino acids in length, more typically 8 or 9 amino acids in length, and even more typically 10 or more amino acids in length. With respect to conformational epitopes, the length of the epitope-defining sequence can be subject to wide variations, since it is believed that these epitopes are formed by the three-dimensional shape of the antigen (e.g. folding). Thus, the amino acids defining the epitope can be relatively few in number, but widely dispersed along the length of the molecule being brought into the correct epitope conformation via folding. The portions of the antigen between the residues defining the epitope may not be critical to the conformational structure of the epitope. For example, deletion or substitution of these intervening sequences may not affect the conformational epitope provided sequences critical to epitope conformation are maintained (e.g. cysteines involved in disulfide bonding, glycosylation sites, etc.). A conformational epitope may also be formed by 2 or more essential regions of subunits of a homo-oligomer or hetero-oligomer.

As used herein, an epitope of a designated polypeptide denotes epitopes with the same amino acid sequence as the epitope in the designated polypeptide, and immunologic equivalents thereof. Such equivalents also include strain, subtype (=genotype), or type(group)-specific variants, e.g. of the currently known sequences or strains belonging to genotypes 1a, 1b, 1c, 1d, 1e, 1f, 2a, 2b, 2c, 2d, 2e, 2f, 2g, 2h, 2i, 3a, 3b, 3c, 3d, 3e, 3f, 3g, 4a, 4b, 4c, 4d, 4e, 4f, 4g, 4h, 4i, 4j, 4k, 4l, 5a, 5b, 6a, 6b, 6c, 7a, 7b, 7c, 8a, 8b, 9a, 9b, 10a, 11 (and subtypes thereof), 12 (and

subtypes thereof) or 13 (and subtypes thereof) or any other newly defined HCV (sub)type. It is to be understood that the amino acids constituting the epitope need not be part of a linear sequence, but may be interspersed by any number of amino acids, thus forming a conformational epitope.

The HCV antigens of the present invention comprise conformational epitopes from the E1 and/or E2 (envelope) domains of HCV. The E1 domain, which is believed to correspond to the viral envelope protein, is currently estimated to span amino acids 192-383 of the HCV polyprotein (Hijikata et al., 1991). Upon expression in a mammalian system (glycosylated), it is believed to have an approximate molecular weight of 35 kDa as determined via SDS-PAGE. The E2 protein, previously called NS1, is believed to span amino acids 384-809 or 384-746 (Grakoui et al., 1993) of the HCV polyprotein and also to be an envelope protein. Upon expression in a vaccinia system (glycosylated), it is believed to have an apparent gel molecular weight of about 72 kDa. It is understood that these protein endpoints are approximations (e.g. the carboxy terminal end of E2 could lie somewhere in the 730-820 amino acid region, e.g. ending at amino acid 730, 735, 740, 742, 744, 745, preferably 746, 747, 748, 750, 760, 770, 780, 790, 800, 809, 810, 820). The E2 protein may also be expressed together with E1, and/or core (aa 1-191), and/or P7 (aa 747-809), and/or NS2 (aa 810-1026), and/or NS3 (aa 1027-1657), and/or NS4A (aa 1658-1711) and/or NS4B (aa 1712-1972) and/or NS5A (aa 1973-2420), and/or NS5B (aa 2421-3011), and/or any part of any of these HCV proteins different from E2 . Likewise, the E1 protein may also be expressed together with the E2, and/or core (aa 1-191), and/or P7 (aa 747-809), and/or NS2 (aa 810-1026), and/or NS3 (aa 1027-1657), and/or NS4A (aa 1658-1711) and/or NS4B (aa 1712-1972), and/or NS5A (aa 1973-2420) , and/or NS5B (aa 2421-3011), and/or any part of any of these HCV proteins different from E1. Expression together with these other HCV proteins may be important for obtaining the correct protein folding.

The term "E1" as used herein also includes analogs and truncated forms that are immunologically cross-reactive with natural E1, and includes E1 proteins of genotypes 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13 or any other newly identified HCV type or subtype. The term 'E2' as used herein also includes analogs and truncated forms that are immunologically cross-reactive with natural E2, and includes E2 proteins of genotypes 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13 or any other newly identified HCV type or subtype. For example, insertions of multiple codons between codon 383 and 384, as well as deletions of amino acids 384-387 have been reported by Kato et al. (1992). It is thus also understood that the isolates used in the examples section of the present invention were not intended to limit the scope of the invention and that

any HCV isolate from type 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13 or any other new genotype of HCV is a suitable source of E1 and/or E2 sequence for the practice of the present invention. Similarly, as described above, the HCV proteins that are co-expressed with the HCV envelope proteins of the present invention, can be derived from any HCV type, thus also from the same type as the HCV envelope proteins of the present invention.

"E1/E2" as used herein refers to an oligomeric form of envelope proteins containing at least one E1 component and at least one E2 component.

The term "specific oligomeric" E1 and/or E2 and/or E1/E2 envelope proteins refers to all possible oligomeric forms of recombinantly expressed E1 and/or E2 envelope proteins which are not aggregates. E1 and/or E2 specific oligomeric envelope proteins are also referred to as homo-oligomeric E1 or E2 envelope proteins (see below). The term 'single or specific oligomeric' E1 and/or E2 and/or E1/E2 envelope proteins refers to single monomeric E1 or E2 proteins (single in the strict sense of the word) as well as specific oligomeric E1 and/or E2 and/or E1/E2 recombinantly expressed proteins. These single or specific oligomeric envelope proteins according to the present invention can be further defined by the following formula $(E1)_x(E2)_y$ wherein x can be a number between 0 and 100, and y can be a number between 0 and 100, provided that x and y are not both 0. With x=1 and y=0 said envelope proteins include monomeric E1.

The term "homo-oligomer" as used herein refers to a complex of E1 or E2 containing more than one E1 or E2 monomer, e.g. E1/E1 dimers, E1/E1/E1 trimers or E1/E1/E1/E1 tetramers and E2/E2 dimers, E2/E2/E2 trimers or E2/E2/E2/E2 tetramers, E1 pentamers and hexamers, E2 pentamers and hexamers or any higher-order homo-oligomers of E1 or E2 are all 'homo-oligomers' within the scope of this definition. The oligomers may contain one, two, or several different monomers of E1 or E2 obtained from different types or subtypes of hepatitis C virus including for example those described by Maertens et al. in WO94/25601 and WO96/13590, both by the present applicants. Such mixed oligomers are still homo-oligomers within the scope of this invention, and may allow more universal diagnosis, prophylaxis or treatment of HCV.

The E1 and E2 antigens used in the present invention may be full-length viral proteins, substantially full-length versions thereof, or functional fragments thereof (e.g. fragments comprising at least one epitope and/or glycosylation site). Furthermore, the HCV antigens of the present invention can also include other sequences that do not block or prevent the formation of the conformational epitope of interest. The presence or absence of a conformational epitope can be readily determined through screening the antigen of interest with an antibody (polyclonal

serum or monoclonal to the conformational epitope) and comparing its reactivity to that of a denatured version of the antigen which retains only linear epitopes (if any). In such screening using polyclonal antibodies, it may be advantageous to adsorb the polyclonal serum first with the denatured antigen and see if it retains antibodies to the antigen of interest.

The HCV proteins of the present invention may be glycosylated. Glycosylated proteins intend proteins that contain one or more carbohydrate groups, in particular sugar groups. In general, all eukaryotic cells are able to glycosylate proteins. After alignment of the different envelope protein sequences of HCV genotypes, it may be inferred that not all 6 glycosylation sites on the HCV E1 protein are required for proper folding and reactivity. It is further known that the glycosylation site at position 325 is not modified by N-glycosylation (Fournillier-Jacob et al. 1996, Meunier et al. 1999). Furthermore, HCV subtype 1b E1 protein contains 6 glycosylation sites, but some of these glycosylation sites are absent in certain other (sub)types. The fourth carbohydrate motif (on Asn250), present in types 1b, 6a, 7, 8, and 9, is absent in all other types know today. This sugar-addition motif may be mutated to yield a type 1b E1 protein with improved reactivity. Also, the type 2b sequences show an extra glycosylation site in the V5 region (on Asn299). The isolate S83, belonging to genotype 2c, even lacks the first carbohydrate motif in the V1 region (on Asn), while it is present on all other isolates (Stuyver et al., 1994). However, even among the completely conserved sugar-addition motifs, the presence of the carbohydrate may not be required for folding, but may have a role in evasion of immune surveillance. Thus, the identification of the role of glycosylation can be further tested by mutagenesis of the glycosylation motifs. Mutagenesis of a glycosylation motif (NXS or NXT sequences) can be achieved by either mutating the codons for N, S, or T, in such a way that these codons encode amino acids different from N in the case of N, and/or amino acids different from S or T in the case of S and in the case of T. Alternatively, the X position may be mutated into P, since it is known that NPS or NPT are not frequently modified with carbohydrates. After establishing which carbohydrate-addition motifs are required for folding and/or reactivity and which are not, combinations of such mutations may be made. Such experiments have been described extensively by Maertens et al. in Example 8 of WO96/04385, which is included herein specifically by reference.

The term glycosylation as used in the present invention refers to N-glycsoylation unless otherwise specified.

In particular, the present invention relates to HCV envelope proteins, or parts thereof that are core-glycosylated. In this respect, the term "core-glycosylation" refers to a structure "similar" to the structure as depicted in the boxed structure in Figure 3 of Herscovics and

Orlean (1993). Thus, the carbohydrate structure referred to contains 10 to 11 monosaccharides. Notably, said disclosure is herein incorporated by reference. The term "similar" intends that not more than 4 additional monosaccharide has been added to the structure or that not more than about 3 monosaccharides have been removed from the structure. Consequently, the core-glycosylation carbohydrate structure referred to in the current invention consists minimally of 7 and maximally of 15 monosaccharides and can consist of 8, 9, 10, 11, 12, 13 or 14 monosaccharides. The monosaccharides connoted are preferentially glucose, mannose or N-acetyl glucosamine.

An alternative aspect of the current invention relates to an isolated HCV envelope protein or a fragment thereof comprising at least one N-glycosylation site, said protein or fragment thereof characterized in that it is the product of expression in a eukaryotic cell and further characterized in that more than 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94% or 95 % of the N-glycosylated sites are glycosylated with an oligomannose with a structure defined by Man(8 to 10)-GlcNAc(2). More specific to the above N-glycosylation characteristic, the ratio of sites core-glycosylated with an oligomannose with structure Man(7)-GlcNAc(2) over the sites core-glycosylated with an oligomannose with structure Man(8)-GlcNAc(2) is less than or equal to 0.15, 0.2, 0.25, 0.30, 0.35, 0.40, 0.44, 0.45, or 0.50. Further more specific to any of the above N-glycosylation characteristics, said oligomannoses contain less than 20%, 19%, 18%, 17%, 16%, 15%, 14%, 13%, 12%, 11%, 10%, 9%, 8%, 7%, 6%, or 5% terminal α1,3 mannose.

Another alternative aspect of the invention is related to an isolated HCV envelope protein or a fragment thereof comprising at least one N-glycosylation site, said protein or fragment thereof characterized in that it is the product of expression in a eukaryotic cell and further characterized in that N-glycosylated sites are occupied by oligomannoses wherein the ratio of the oligomannoses with structure Man(7)-GlcNAc(2) over the oligomannoses with structure Man(8)-GlcNAc(2) is less than or equal to 0.15, 0.2, 0.25, 0.30, 0.35, 0.40, 0.44, 0.45, or 0.50. Further more specific to the above N-glycosylation characteristics, said oligomannoses contain less than 20%, 19%, 18%, 17%, 16%, 15%, 14%, 13%, 12%, 11%, 10%, 9%, 8%, 7%, 6%, or 5% terminal α1,3 mannose.

In another alternative aspect of the current invention is covered an isolated HCV envelope protein or a fragment thereof comprising at least one N-glycosylation site, said protein or fragment thereof characterized in that it is the product of expression in a non-mammalian eukaryotic cell and further characterized in that the number of N-glycosylated sites is at least 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, or 15%, less than the number of N-glycosylated sites in the same protein or fragment thereof expressed from a vaccinia virus in a eukaryotic cell liable of being infected with said vaccinia virus. More particular to the above N-glycosylation characteristic, up to 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, or 80% of said N-glycosylated sites are core-glycosylated. More specifically, more than 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94% or 95 % of the N-glycosylated sites are glycosylated with an oligomannose with a structure defined by Man(8 to 10)-GlcNAc(2). More specific to any of the above N-glycosylation characteristics, the ratio of sites core-glycosylated with an oligomannose with structure Man(7)-GlcNAc(2) over the sites core-glycosylated with an oligomannose with structure Man(8)-GlcNAc(2) is less than or equal to 0.15, 0.2, 0.25, 0.30, 0.35, 0.40, 0.45, or 0.50. Further more specific to any of the above N-glycosylation characteristics, said oligomannoses contain less than 20%, 19%, 18%, 17%, 16%, 15%, 14%, 13%, 12%, 11%, 10%, 9%, 8%, 7%, 6%, or 5% terminal α1,3 mannose.

In another aspect of the invention, the isolated HCV envelope protein or part thereof according to the invention is the product of expression in a yeast cell. More particularly, the isolated HCV envelope protein or part thereof according to the invention is the product of expression in a cell of strains of *Saccharomyces*, such as *Saccharomyces cerevisiae*, *Saccharomyces kluyveri*, or *Saccharomyces uvarum*, *Schizosaccharomyces*, such as *Schizosaccharomyces pombe*, *Kluyveromyces*, such as *Kluyveromyces lactis*, *Yarrowia*, such as *Yarrowia lipolytica*, *Hansenula*, such as *Hansenula polymorpha*, *Pichia*, such as *Pichia pastoris*, *Aspergillus species*, *Neurospora*, such as *Neurospora crassa*, or *Schwanniomyces*, such as *Schwanniomyces occidentalis*, or mutant cells derived from any thereof. More specifically, the isolated HCV envelope protein or part thereof according to the invention is the product of expression in a *Hansenula* cell. Even more specifically, the isolated HCV

envelope protein or part thereof according to the invention is the product of expression in a yeast, e.g. *Hansenula*, cell in the absence of glycosylation inhibitors such as tunicamycin.

In another aspect of the invention the isolated HCV envelope protein or part thereof according to the invention is derived from a protein comprising an avian lysozyme leader peptide or a functional variant thereof joined to said HCV envelope protein or fragment thereof. More specifically, the isolated HCV envelope protein or part thereof according to the invention is derived from a protein characterized by the structure

$CL\text{-}[(A1)_a - (PS1)_b - (A2)_c]\text{-}HCVENV\text{-}[(A3)_d - (PS2)_e - (A4)_f]$

wherein:

CL is an avian lysozyme leader peptide or a functional equivalent thereof,

A1, A2, A3 and A4 are adaptor peptides which can be different or the same,

PS1 and PS2 are processing sites which can be the different or the same,

HCVENV is a HCV envelope protein or a part thereof,

a, b, c, d, e and f are 0 or 1, and

wherein, optionally, A1 and/or A2 are part of PS1 and/or wherein A3 and/or A4 are part of PS2.

With "an avian leader peptide or a functional equivalent thereof joined to a HCV envelope protein or a part thereof" is meant that the C-terminal amino acid of said leader peptide is covalently linked via a peptide bond to the N-terminal amino acid of said HCV envelope protein or part thereof. Alternatively, the C-terminal amino acid of said leader peptide is separated from the N-terminal amino acid of said HCV envelope protein or part thereof by a peptide or protein. Said peptide or protein may have the structure $-[(A1)_a - (PS1)_b - (A2)_c]$ as defined above.

The derivation of the HCV envelope protein of interest from the protein comprising an avian lysozyme leader peptide or a functional equivalent thereof joined to an HCV envelope protein or a part thereof or of the protein characterized by the structure $CL\text{-}[(A1)_a - (PS1)_b - (A2)_c]\text{-}HCVENV\text{-}[(A3)_d - (PS2)_e - (A4)_f]$ can be performed *in vivo* by the proteolytic machinery of the cells in which the pre-protein protein is expressed. More specifically, the step consisting of removal of the avian leader peptide is preferably performed *in vivo* by the proteolytic machinery of the cells in which the pre-protein is expressed. Derivation may, however, also be performed solely *in vitro* after and/or during isolation and/or purification of

the pre-protein and/or protein from the cells expressing the pre-protein and/or from the culture fluid in which the cells expressing the pre-protein are grown. Alternatively, said *in vivo* derivation is performed in combination with said *in vitro* derivation. Derivation of the HCV protein of interest from a recombinantly expressed pre-protein can further comprise the use of (an) proteolytic enzyme(s) in a polishing step wherein all or most of the contaminating proteins co-present with the protein of interest are degraded and wherein the protein of interest is resistant to the polishing proteolytic enzyme(s). Derivation and polishing are not mutually exclusive processes and may be obtained by using the same single proteolytic enzyme. As an example is given here the HCV E1s protein of HCV genotype 1b (SEQ ID NO:2) which is devoid of Lys-residues. By digesting of a protein extract containing said HCV E1 proteins with the Endoproteinase Lys-C (endo-lys C), the E1 proteins will not be degraded whereas contaminating proteins containing one or more Lys-residues are degraded. Such a process may significantly simplify or enhance isolation and/or purification of the HCV E1 proteins. Furthermore, by including in a pre-protein an additional Lys-residue, e.g. between a leader peptide and a HCV E1 protein, the additional advantageous possibility of correct *in vitro* separation of the leader peptide from the HCV E1 pre-protein is obtainable. Other HCV E1 proteins may comprise a Lys-residue at either one or more of the positions 4, 40, 42, 44, 61, 65 or 179 (wherein position 1 is the first, N-terminal natural amino acid of the E1 protein, i.e. position 192 in the HCV polyprotein). In order to enable the use of endo-lys C as described above, said Lys-residues may be mutated into another amino acid residue, preferably into an Arg-residue.

With a "correctly removed" leader peptide is meant that said leader peptide is removed from the protein comprising the signal sequence joined to a protein of interest with high efficiency, i.e. a large number of pre-(pro-)proteins is converted to (pro-)proteins, and with high fidelity, i.e. only the pre-amino acid sequence is removed and not any amino acids of the protein of interest joined to said pre-amino acid sequence. With "removal of a leader peptide with high efficiency" is meant that at least about 40%, but more preferentially about 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98% or even 99% of the pre-proteins is converted to the protein from which the pre-sequence is removed. Alternatively, if a substantial part of the expressed pre-proteins is not converted to the protein from which the pre-sequence is removed, these pre-proteins may still be purified or be removed during purification.

With "functional equivalent of the avian lysozyme (CL) leader peptide" is meant a CL leader peptide wherein one or more amino acids have been substituted for another amino acid and whereby said substitution is a conservative amino acid substitution. With "conservative amino acid substitution" is meant a substitution of an amino acid belonging to a group of conserved amino acids with another amino acid belonging to the same group of conserved amino acids. As groups of conserved amino acids are considered: the group consisting of Met, Ile, Leu and Val; the group consisting of Arg, Lys and His; the group consisting of Phe, Trp and Tyr; the group consisting of Asp and Glu; the group consisting of Asn and Gln; the group consisting of Cys, Ser and Thr; and the group consisting of Ala and Gly. An exemplary conservative amino acid substitution in the CL leader peptide is the naturally variation at position 6, the amino acid at this position being either Val or Ile; another variation occurs at position 17, the amino acid at this position being, amongst others, Leu or Pro (see SEQ ID NO:1). The resulting CL leader peptides are thus to be considered as functional equivalents. Other functional equivalents of the CL leader peptides include those leader peptides reproducing the same technical aspects as the CL leader peptides as described throughout the current invention, including deletion variants and insertion variants.

With "A" or "adaptor peptide" is meant a peptide (e.g. 1 to 30 amino acids) or a protein which may serve as a linker between e.g. a leader peptide and a processing site (PS), a leader peptide and a protein of interest, a PS and a protein of interest, and/or a protein of interest and a PS; and/or may serve as a linker N- or C-terminal of e.g. a leader peptide, a PS or a protein of interest. The adaptor peptide "A" may have a certain three-dimensional structure, e.g. an α-helical or β-sheet structure or a combination thereof. Alternatively the three-dimensional structure of A is not well defined, e.g. a coiled-coil structure. The adaptor A may be part of e.g. a pre-sequence, a pro-sequence, a protein of interest sequence or a processing site. The adaptor A may serve as a tag enhancing or enabling detection and/or purification and/or processing of the protein of which A is a part. One examples of an A peptide is the his-tag peptide (HHHHHH; SEQ ID NO:63) Hn wherein n usually is six, but may be 7, 8, 9, 10, 11, or 12. Other examples of A-peptides include the peptides EEGEPK (Kjeldsen et al. in WO98/28429; SEQ ID NO:64) or EEAEPK (Kjeldsen et al. in WO97/22706; SEQ ID NO:65) which, when present at the N-terminal of the a protein of interest, were reported to increase fermentation yield but also to protect the N-terminus of the protein of interest against processing by dipeptidyl aminopeptidase and thus resulting in a homogenous N-terminus of the polypeptide. At the same time, *in vitro* maturation of the protein of interest, i.e. removal of said peptides EEGEPK (SEQ ID NO:64) and EEAEPK

(SEQ ID NO:65) from the protein of interest can be achieved by using e.g. endo-lys C which cleaves C-terminal of the Lys-residue in said peptides. Said peptides thus serve the function of adaptor peptide (A) as well as processing site (PS), (see below). Adaptor peptides are given in SEQ ID NOs:63-65, 70-72 and 74-82. Another example of an adaptor peptide is the G4S immunosilent linker. Other examples of adaptor peptides or adaptor proteins are listed in Table 2 of Stevens (Stevens et al. 2000).

With "PS" or "processing site" is meant a specific protein processing or processable site. Said processing may occur enzymatically or chemically. Examples of processing sites prone to specific enzymatic processing include IEGR↓X (SEQ ID NO:66), IDGR↓X (SEQ ID NO:67), AEGR↓X (SEQ ID NO:68), all recognized by and cleaved between the Arg and Xaa (any amino acid) residues as indicated by the "↓" by the bovine factor Xa protease (Nagai, K. and Thogersen, H. C. 1984). Another example of a PS site is a dibasic site, e.g. Arg-Arg, Lys-Lys, Arg-Lys or Lys-Arg, which is cleavable by the yeast Kex2 protease (Julius, D. et al. 1984). The PS site may also be a monobasic Lys-site. Said monobasic Lys-PS-site may also be included at the C-terminus of an A peptide. Examples of A adaptor peptides comprising a C-terminal monobasic Lys-PS-site are given by SEQ ID NOs:64-65 and 74-76. Exoproteolytic removal of a His-tag (HHHHHH; SEQ ID NO:63) is possible by using the dipeptidyl aminopeptidase I (DAPase) alone or in combination with glutamine cyclotransferase (Qcyclase) and pyroglutamic aminopeptidase (pGAPase) (Pedersen, J. et al. 1999). Said exopeptidases comprising a recombinant His-tag (allowing removal of the peptidase from the reaction mixture by immobilize metal-affinity chromatography, IMAC) are commercially available, e.g. as the TAGZyme System of Unizyme Laboratories (Hørsholm, DK). With "processing" is thus generally meant any method or procedure whereby a protein is specifically cleaved or cleavable at at least one processing site when said processing site is present in said protein. A PS may be prone to endoproteolytic cleavage or may be prone to exproteolytic cleavage, in any case the cleavage is specific, i.e. does not extend to sites other than the sites recognized by the processing proteolytic enzyme. A number of PS sites are given in SEQ ID NOs:66-68 and 83-84.

The versatility of the $[(A1/3)_{a/d}$ - $(PS1/2)_{b/e}$ – $(A2/4)_{c/f}]$ structure as outlined above is demonstrated by means of some examples. In a first example, said structure is present at the C-terminal end of a protein of interest comprised in a pre-protein and wherein A3 is the "VIEGR" peptide (SEQ ID NO:69) which is overlapping with the factor Xa "IEGRX" PS site (SEQ ID NO:66) and wherein X=A4 is the histidine-tag (SEQ ID NO:63) (d, e and f thus are

all 1 in this case). The HCV protein of interest can (optionally) be purified by IMAC. After processing with factor Xa, the (optionally purified) HCV protein of interest will carry at its C-terminus a processed PS site which is "IEGR" (SEQ ID NO:70). Variant processed factor Xa processing site, can be IDGR (SEQ ID NO:71) or AEGR (SEQ ID NO:72). In a further example, the $[(A1/3)_{a/d}$ - $(PS1/2)_{b/e}$ – $(A2/4)_{c/f}]$ structure is present at the N-terminus of the HCV protein of interest. Furthermore, A1 is the histidine-tag (SEQ ID NO:63), PS is the factor Xa recognition site (any of SEQ ID NOs:66-68) wherein X is the protein of interest, and wherein a=b=1 and c=0. Upon correct removal of a leader peptide, e.g. by the host cell, the resulting HCV protein of interest can be purified by IMAC (optional). After processing with factor Xa, the protein of interest will be devoid of the $[(A1)_a$ - $(PS1)_b$ – $(A2)_c]$ structure.

It will furthermore be clear that any of A1, A2, A3, A4, PS1 and PS2, when present, may be present in a repeat structure. Such a repeat structure, when present, is in this context still counted as 1, i.e. a, b, c, d, e, or f are 1 even if e.g. A1 is occurring as e.g. 2 repeats (A1-A1).

Yet another aspect of the current invention relates to any of the isolated HCV envelope protein or fragment thereof according to the invention in which the cysteine thiol-groups are chemically modified.

Yet another aspect of the current invention relates to any of the isolated HCV envelope protein or fragment thereof according to the invention which is antigenic.

Yet another aspect of the current invention relates to any of the isolated HCV envelope protein or fragment thereof according to the invention which is immunogenic.

Yet another aspect of the current invention relates to any of the isolated HCV envelope protein or fragment thereof according to the invention which comprises a T-cell stimulating epitope.

Another aspect of the current invention relates to any of the isolated HCV envelope protein or fragment thereof according to the invention which is comprised in a structure chosen from the group consisting of monomers, homodimers, heterodimers, homo-oligomers and hetero-oligomers.

Yet another aspect of the current invention relates to any of the isolated HCV envelope protein or fragment thereof according to the invention which is comprised in a virus-like particle.

In the HCV envelope proteins or parts thereof as described herein comprising at least one cysteine residue, but preferably 2 or more cysteine residues, the cysteine thiol-groups can

be irreversibly protected by chemical or enzymatic means. In particular, "irreversible protection" or "irreversible blocking" by chemical means refers to alkylation, preferably alkylation of the HCV envelope proteins by means of alkylating agents, such as, for example, active halogens, ethylenimine or N-(iodoethyl)trifluoro-acetamide. In this respect, it is to be understood that alkylation of cysteine thiol-groups refers to the replacement of the thiol-hydrogen by $(CH_2)_nR$, in which n is 0, 1, 2, 3 or 4 and R= H, COOH, NH_2, $CONH_2$, phenyl, or any derivative thereof. Alkylation can be performed by any method known in the art, such as, for example, active halogens $X(CH_2)_nR$ in which X is a halogen such as I, Br, Cl or F. Examples of active halogens are methyliodide, iodoacetic acid, iodoacetamide, and 2-bromoethylamine. Other methods of alkylation include the use of NEM (N-ethylmaleimide) or Biotin-NEM, a mixture thereof, or ethylenimine or N-(iodoethyl)trifluoroacetamide both resulting in substitution of -H by $-CH_2-CH_2-NH_2$ (Hermanson, G. T. 1996). The term "alkylating agents" as used herein refers to compounds which are able to perform alkylation as described herein. Such alkylations finally result in a modified cysteine, which can mimic other aminoacids. Alkylation by an ethylenimine results in a structure resembling lysine, in such a way that new cleavage sites for trypsine are introduced (Hermanson, G. T. 1996). Similarly, the usage of methyliodide results in an amino acid resembling methionine, while the usage of iodoacetate and iodoacetamide results in amino acids resembling glutamic acid and glutamine, respectively. In analogy, these amino acids are preferably used in direct mutation of cysteine. Therefore, the present invention pertains to HCV envelope proteins as described herein, wherein at least one cysteine residue of the HCV envelope protein as described herein is mutated to a natural amino acid, preferentially to methionine, glutamic acid, glutamine or lysine. The term "mutated" refers to site-directed mutagenesis of nucleic acids encoding these amino acids, ie to the well kown methods in the art, such as, for example, site-directed mutagenesis by means of PCR or via oligonucleotide-mediated mutagenesis as described in (Sambrook, J. et al. 1989). It should be understood that for the Examples section of the present invention, alkylation refers to the use of iodo-acetamide as an alkylating agent unless otherwise specified.

It is further understood that in the purification procedure, the cysteine thiol-groups of the HCV proteins or the parts thereof of the present invention can be reversibly protected. The purpose of reversible protection is to stabilize the HCV protein or part thereof. Especially, after reversible protection the sulfur-containing functional group (eg thiols and disulfides) is retained in a non-reactive condition. The sulfur-containing functional group is thus unable to

react with other compounds, e.g. have lost their tendency of forming or exchanging disulfide bonds, such as, for example

R_1-SH + R_2-SH ---X---> R_1-S-S-R_2 ;

R_1-S-S-R_2 + R_3-SH ---X---> R_1-S-S-R_3 + R_2-SH ;

R_1-S-S-R_2 + R_3-S-S-R_4 ---X---> R_1-S-S-R_3 + R_2-S-S-R_4.

The described reactions between thiols and/or disulphide residues are not limited to intermolecular processes, but may also occur intramolecularly.

The term "reversible protection" or "reversible blocking" as used herein contemplates covalently binding of modification agents to the cysteine thiol-groups, as well as manipulating the environment of the HCV protein such, that the redox state of the cysteine thiol-groups remains unaffected throughout subsequent steps of the purification procedure (shielding). Reversible protection of the cysteine thiol-groups can be carried out chemically or enzymatically.

The term "reversible protection by enzymatical means" as used herein contemplates reversible protection mediated by enzymes, such as for example acyl-transferases, e.g. acyl-transferases that are involved in catalysing thio-esterification, such as palmitoyl acyltransferase (see below).

The term "reversible protection by chemical means" as used herein contemplates reversible protection:

1. by modification agents that reversibly modify cysteinyls such as for example by sulphonation and thio-esterification;

   Sulphonation is a reaction where thiol or cysteines involved in disulfide bridges are modified to *S*-sulfonate: RSH → RS-SO_3^- (Darbre, A. 1986) or RS-SR→ 2 RS-SO_3^- (sulfitolysis; (Kumar, N. et al. 1986)). Reagents for sulfonation are e.g. Na_2SO_3, or sodium tetrathionate. The latter reagents for sulfonation are used in a concentration of 10-200 mM, and more preferentially in a concentration of 50-200 mM. Optionally sulfonation can be performed in the presence of a catalysator such as, for example Cu^{2+} (100 µM-1 mM) or cysteine (1-10 mM).

   The reaction can be performed under protein denaturing as well as native conditions (Kumar, N. et al. 1985, Kumar, N. et al. 1986).

   Thioester bond formation, or thio-esterification is characterised by:

   RSH + R'COX → RS-COR'

   in which X is preferentially a halogenide in the compound R'CO-X.

2. by modification agents that reversibly modify the cysteinyls of the present invention such as, for example, by heavy metals, in particular Zn^{2+}, Cd^{2+}, mono-, dithio- and disulfide-compounds (e.g. aryl- and alkylmethanethiosulfonate, dithiopyridine, dithiomorpholine, dihydrolipoamide, Ellmann reagent, aldrothiol™ (Aldrich) (Rein, A. et al. 1996), dithiocarbamates), or thiolation agents (e.g. gluthathion, N-Acetyl cysteine, cysteineamine). Dithiocarbamate comprise a broad class of molecules possessing an $R_1R_2NC(S)SR_3$ functional group, which gives them the ability to react with sulphydryl groups. Thiol containing compounds are preferentially used in a concentration of 0.1-50 mM, more preferentially in a concentration of 1-50 mM, and even more preferentially in a concentration of 10-50 mM;
3. by the presence of modification agents that preserve the thiol status (stabilise), in particular antioxidantia, such as for example DTT, dihydroascorbate, vitamins and derivates, mannitol, amino acids, peptides and derivates (e.g. histidine, ergothioneine, carnosine, methionine), gallates, hydroxyanisole, hydoxytoluene, hydroquinon, hydroxymethylphenol and their derivates in concentration range of 10 μM-10 mM, more preferentially in a concentration of 1-10 mM;
4. by thiol stabilising conditions such as, for example, (i) cofactors as metal ions (Zn^{2+}, Mg^{2+}), ATP, (ii) pH control (e.g. for proteins in most cases pH ~5 or pH is preferentially thiol pK_a -2; e.g. for peptides purified by Reversed Phase Chromatography at pH ~2).

Combinations of reversible protection as described in (1), (2), (3) and (4) may result in similarly pure and refolded HCV proteins. In effect, combination compounds can be used, such as, for example Z103 (Zn carnosine), preferentially in a concentration of 1-10 mM. It should be clear that reversible protection also refers to, besides the modification groups or shielding described above, any cysteinyl protection method which may be reversed enzymatically or chemically, without disrupting the peptide backbone. In this respect, the present invention specifically refers to peptides prepared by classical chemical synthesis (see above), in which, for example, thioester bounds are cleaved by thioesterase, basic buffer conditions (Beekman, N. J. et al. 1997) or by hydroxylamine treatment (Vingerhoeds, M. H. et al. 1996).

Thiol containing HCV proteins can be purified, for example, on affinity chromatography resins which contain (1) a cleavable connector arm containing a disulfide bond (e.g. immobilised 5,5' dithiobis(2-nitrobenzoic acid) (Jayabaskaran, C. et al. 1987) and covalent chromatography on activated thiol-Sepharose 4B (Pharmacia)) or (2) a aminohexanoyl-4-aminophenylarsine as immobilised ligand. The latter affinity matrix has

been used for the purification of proteins, which are subject to redox regulation and dithiol proteins that are targets for oxidative stress (Kalef, E. et al. 1993).

Reversible protection may also be used to increase the solubilisation and extraction of peptides (Pomroy, N. C. and Deber, C. M. 1998).

The reversible protection and thiol stabilizing compounds may be presented under a monomeric, polymeric or liposomic form.

The removal of the reversibly protection state of the cysteine residues can chemically or enzymatically accomplished by e.g.:

- a reductant, in particular DTT, DTE, 2-mercaptoethanol, dithionite, $SnCl_2$, sodium borohydride, hydroxylamine, TCEP, in particular in a concentration of 1-200 mM, more preferentially in a concentration of 50-200 mM;
- removal of the thiol stabilising conditions or agents by e.g. pH increase;
- enzymes, in particular thioesterases, glutaredoxine, thioredoxine, in particular in a concentration of 0.01-5 μM, even more particular in a concentration range of 0.1-5 μM.;
- combinations of the above described chemical and/or enzymatical conditions.

The removal of the reversibly protection state of the cysteine residues can be carried out *in vitro* or *in vivo*, e.g. in a cell or in an individual.

It will be appreciated that in the purification procedure, the cysteine residues may or may not be irreversibly blocked, or replaced by any reversible modification agent, as listed above.

A reductant according to the present invention is any agent which achieves reduction of the sulfur in cysteine residues, e.g. "S-S" disulfide bridges, desulphonation of the cysteine residue ($RS\text{-}SO_3^- \rightarrow RSH$). An antioxidant is any reagent which preserves the thiol status or minimises "S-S" formation and/or exchanges. Reduction of the "S-S" disulfide bridges is a chemical reaction whereby the disulfides are reduced to thiol (-SH). The disulfide bridge breaking agents and methods disclosed by Maertens et al. in WO 96/04385 are hereby incorporated by reference in the present description. "S-S" Reduction can be obtained by (1) enzymatic cascade pathways or by (2) reducing compounds. Enzymes like thioredoxin, glutaredoxin are known to be involved in the in vivo reduction of disulfides and have also been shown to be effective in reducing "S-S" bridges in vitro. Disulfide bonds are rapidly cleaved by reduced thioredoxin at pH 7.0, with an apparent second order rate that is around 10^4 times larger than the corresponding rate constant for the reaction with DTT. The reduction

kinetic can be dramatically increased by preincubation the protein solution with 1 mM DTT or dihydrolipoamide (Holmgren, A. 1979). Thiol compounds able to reduce protein disulfide bridges are for instance Dithiothreitol (DTT), Dithioerythritol (DTE), β-mercaptoethanol, thiocarbamates, bis(2-mercaptoethyl) sulfone and N,N'-bis(mercaptoacetyl)hydrazine, and sodium-dithionite. Reducing agents without thiol groups like ascorbate or stannous chloride ($SnCl_2$), which have been shown to be very useful in the reduction of disulfide bridges in monoclonal antibodies (Thakur, M. L. et al. 1991), may also be used for the reduction of HCV proteins. In addition, changes in pH values may influence the redox status of HCV proteins. Sodium borohydride treatment has been shown to be effective for the reduction of disulfide bridges in peptides (Gailit, J. 1993). Tris (2-carboxyethyl)phosphine (TCEP) is able to reduce disulfides at low pH (Burns, J. et al. 1991). Selenol catalyses the reduction of disulfide to thiols when DTT or sodium borohydride is used as reductant. Selenocysteamine, a commercially available diselenide, was used as precursor of the catalyst (Singh, R. and Kats,. L. 1995).

The term "immunogenic" refers to the ability of a protein or a substance to produce an immune response. The immune response is the total response of a body to the introduction of an antigen, including antibody formation, cellular immunity, hypersensitivity, or immunological tolerance. Cellular immunity refers to a T-helper cell- and/or CTL-response.

The term "antigenic" refers to the ability of a protein or a substance to causes the formation of an antibody or to elicit a cellular response.

The expression "T-cell stimulating epitope" according to the present invention refers to an epitope capable of stimulating T-cells or CTL-cells, respectively. A T-helper cell stimulating epitope may be selected by monitoring the lymphoproliferative response towards polypeptides containing in their amino acid sequence a (putative) T-cell stimulating epitope. Said lymphoproliferative response may be measured by either a T-helper assay comprising *in vitro* stimulation of peripheral blood mononuclear cells (PMBCs) from patient sera with varying concentrations of peptides to be tested for T-cell stimulating activity and counting the amount of radiolabelled thymidine uptake. A CTL-stimulating epitope may be selected by means of a cytotoxic T-cell (CTL) assay measuring the lytic activity of cytotoxic cells using ^{51}Cr release. Proliferation is considered positive when the stimulation index (mean cpm of antigen-stimulated cultures/mean cpm of controle cultures) is more than 1, preferably more than 2, most preferably more than 3.

Another aspect of the invention refers to a composition comprising an isolated HCV envelope protein or fragment thereof according to the invention. Said composition may further comprise a pharmaceutically acceptable carrier and can be a medicament or a vaccine.

A further aspect of the invention covers a medicament or a vaccine comprising a HCV envelope protein or part thereof according to the invention.

Yet another aspect of the invention comprises a pharmaceutical composition for inducing a HCV-specific immune response in a mammal, said composition comprising an effective amount of a HCV envelope protein or part thereof according to the invention and, optionally, a pharamaceutically acceptable adjuvant. Said pharmaceutical composition comprising an effective amount of a HCV envelope protein or part thereof according to the invention may also be capable of inducing HCV-specific antibodies in a mammal, or capable of inducing a T-cell function in a mammal. Said pharmaceutical compostion comprising an effective amount of a HCV envelope protein or part thereof according to the invention may be prophylactic composition or a therapeutic composition. In a specific embodiment said mammal is a human.

A "mammal" is to be understood as any member of the higher vertebrate class Mammalia, including humans; characterized by live birth, body hair, and mammary glands in the female that secrete milk for feeding the young. Mammals thus also include non-human primates and trimera mice (Zauberman et al. 1999).

A "vaccine" or "medicament" is a composition capable of eliciting protection against a disease, whether partial or complete, whether against acute or chronic disease; in this case the vaccine or medicament is a prophylactic vaccine or medicament. A vaccine or medicament may also be useful for treatment of an already ill individual, in which case it is called a therapeutic vaccine or medicament. Likewise, a pharmaceutical composition can be used for either prophylactic and/or therapeutic purposes in which cases it is a prophylactic and/or therapeutic composition, respectively.

The HCV envelope proteins of the present invention can be used as such, in a biotinylated form (as explained in WO 93/18054) and/or complexed to *Neutralite Avidin* (Molecular Probes Inc., Eugene, OR, USA), avidin or streptavidin. It should also be noted that "a vaccine" or "a medicament" may comprise, in addition to an active substance, a "pharmaceutically acceptable carrier" or "pharmaceutically acceptable adjuvant" which may

be a suitable excipient, diluent, carrier and/or adjuvant which, by themselves, do not induce the production of antibodies harmful to the individual receiving the composition nor do they elicit protection. Suitable carriers are typically large slowly metabolized macromolecules such as proteins, polysaccharides, polylactic acids, polyglycolic acids, polymeric amino acids, amino acid copolymers and inactive virus particles. Such carriers are well known to those skilled in the art. Preferred adjuvants to enhance effectiveness of the composition include, but are not limited to: aluminium hydroxide, aluminium in combination with 3-0-deacylated monophosphoryl lipid A as described in WO 93/19780, aluminium phosphate as described in WO 93/24148, N-acetyl-muramyl-L-threonyl-D-isoglutamine as described in U.S. Patent N° 4,606,918, N-acetyl-normuramyl-L-alanyl-D-isoglutamine, N-acetylmuramyl-L-alanyl-D-isoglutamyl-L-alanine2-(1'2'dipalmitoyl-sn-glycero-3-hydroxyphosphoryloxy) ethylamine, RIBI (ImmunoChem Research Inc., Hamilton, MT, USA) which contains monophosphoryl lipid A, detoxified endotoxin, trehalose-6,6-dimycolate, and cell wall skeleton (MPL + TDM + CWS) in a 2% squalene/Tween 80 emulsion. Any of the three components MPL, TDM or CWS may also be used alone or combined 2 by 2. The MPL may also be replaced by its synthetic analogue referred to as RC-529. Additionally, adjuvants such as Stimulon (Cambridge Bioscience, Worcester, MA, USA), SAF-1 (Syntex) or bacterial DNA-based adjuvants such as ISS (Dynavax) or CpG (Coley Pharmaceuticals) may be used, as well as adjuvants such as combinations between QS21 and 3-de-O-acetylated monophosphoryl lipid A (WO94/00153), or MF-59 (Chiron), or poly[di(carboxylatophenoxy) phosphazene] based adjuvants (Virus Research Institute), or blockcopolymer based adjuvants such as Optivax (Vaxcel, Cythx) or inulin-based adjuvants, such as Algammulin and GammaInulin (Anutech), Incomplete Freund's Adjuvant (IFA) or Gerbu preparations (Gerbu Biotechnik). It is to be understood that Complete Freund's Adjuvant (CFA) may be used for non-human applications and research purposes as well. "A vaccine composition" may further contain excipients and diluents, which are inherently non-toxic and non-therapeutic, such as water, saline, glycerol, ethanol, wetting or emulsifying agents, pH buffering substances, preservatives, and the like. Typically, a vaccine composition is prepared as an injectable, either as a liquid solution or suspension. Injection may be subcutaneous, intramuscular, intravenous, intraperitoneal, intrathecal, intradermal. Other types of administration comprise implantation, suppositories, oral ingestion, enteric application, inhalation, aerosolization or nasal spray or drops. Solid forms, suitable for solution on, or suspension in, liquid vehicles prior to injection may also be prepared. The preparation may also be emulsified or encapsulated in liposomes for enhancing adjuvant effect. The polypeptides may also be incorporated into Immune Stimulating

Complexes together with saponins, for example Quil A (ISCOMS). Vaccine compositions comprise an effective amount of an active substance, as well as any other of the above-mentioned components. "Effective amount" of an active substance means that the administration of that amount to an individual, either in a single dose or as part of a series, is effective for prevention or treatment of a disease or for inducing a desired effect. This amount varies depending upon the health and physical condition of the individual to be treated, the taxonomic group of the individual to be treated (e.g. human, non-human primate, primate, etc.), the capacity of the individual's immune system to mount an effective immune response, the degree of protection desired, the formulation of the vaccine, the treating doctor's assessment, the strain of the infecting pathogen and other relevant factors. It is expected that the amount will fall in a relatively broad range that can be determined through routine trials. Usually, the amount will vary from 0.01 to 1000 μg/dose, more particularly from 0.1 to 100 μg/dose. Dosage treatment may be a single dose schedule or a multiple dose schedule. The vaccine may be administered in conjunction with other immunoregulatory agents.

Another aspect of the current invention relates to a method for producing the isolated HCV envelope protein or fragment thereof according to the invention.

Said method for producing an HCV envelope protein or part thereof is e.g. comprising transformation of a host cell with a recombinant nucleic acid or vector comprising an open reading frame encoding said HCV envelope protein or part thereof, and wherein said host cell is capable of expressing said HCV envelope protein or part thereof. Said method may further comprise cultivation of said host cells in a suitable medium to obtain expression of said protein, isolation of the expressed protein from a culture of said host cells, or from said host cells. Said isolation may include one or more of (i) lysis of said host cells in the presence of a chaotropic agent, (ii) chemical modification of the cysteine thiol-groups in the isolated proteins wherein said chemical modification may be reversible or irreversible and (iii) heparin affinity chromatography.

Exemplary "chaotropic agents" are guanidinium chloride and urea. In general, a chaotropic agent is a chemical that can disrupt the hydrogen bonding structure of water. In concentrated solutions they can denature proteins because they reduce the hydrophobic effect

With "recombinant nucleic acid" is intended a nucleic acid of natural or synthetic origin which has been subjected to at least one recombinant DNA technical manipulation such as restriction enzyme digestion, PCR, ligation, dephosphorylation, phosphorylation, mutagenesis,

adaptation of codons for expression in a heterologous cell etc. In general, a recombinant nucleic acid is a fragment of a naturally occurring nucleic acid or comprises at least two nucleic acid fragments not naturally associated or is a fully synthetic nucleic acid.

The terms "polynucleotide", "polynucleic acid", "nucleic acid sequence", "nucleotide sequence", "nucleic acid molecule", "oligonucleotide", "probe" or "primer", when used herein refer to nucleotides, either ribonucleotides, deoxyribonucleotides, peptide nucleotides or locked nucleotides, or a combination thereof, in a polymeric form of any length or any shape (e.g. branched DNA). Said terms furthermore include double-stranded (ds) and single-stranded (ss) polynucleotides as well as triple-stranded polynucleotides. Said terms also include known nucleotide modifications such as methylation, cyclization and 'caps' and substitution of one or more of the naturally occurring nucleotides with an analog such as inosine or with non-amplifiable monomers such as HEG (hexethylene glycol).
Ribonucleotides are denoted as NTPs, deoxyribonucleotides as dNTPs and dideoxyribonucleotides as ddNTPs.

Nucleotides can generally be labeled radioactively, chemiluminescently, fluorescently, phosphorescently or with infrared dyes or with a surface-enhanced Raman label or plasmon resonant particle (PRP).

Said terms "polynucleotide", "polynucleic acid", "nucleic acid sequence", "nucleotide sequence", "nucleic acid molecule", "oligonucleotide", "probe" or "primer" also encompass peptide nucleic acids (PNAs), a DNA analogue in which the backbone is a pseudopeptide consisting of N-(2-aminoethyl)-glycine units rather than a sugar. PNAs mimic the behavior of DNA and bind complementary nucleic acid strands. The neutral backbone of PNA results in stronger binding and greater specificity than normally achieved. In addition, the unique chemical, physical and biological properties of PNA have been exploited to produce powerful biomolecular tools, antisense and antigene agents, molecular probes and biosensors. PNA probes can generally be shorter than DNA probes and are generally from 6 to 20 bases in length and more optimally from 12 to 18 bases in length (Nielsen, P. E. 2001). Said terms further encompass locked nucleic acids (LNAs) which are RNA derivatives in which the ribose ring is constrained by a methylene linkage between the 2'-oxygen and the 4'-carbon. LNAs display unprecedented binding affinity towards DNA or RNA target sequences. LNA nucleotides can be oligomerized and can be incorporated in chimeric or mix-meric LNA/DNA or LNA/RNA molecules. LNAs seem to be nontoxic for cultured cells (Orum, H. and Wengel, J. 2001, Wahlestedt, C. et al. 2000). In general, chimeras or mix-mers of any of DNA, RNA, PNA and LNA are considered as well as any of these wherein thymine is replaced by uracil.

It is clear from the above that the present invention also relates to the use of a core-glycosylated HCV envelope proteins according to the invention or a composition according to the invention for the manufacture of an HCV vaccine composition. In particular, the present invention relates to the use of a core-glycosylated HCV envelope protein according to the invention for inducing immunity against HCV in chronic HCV carriers. More in particular, the present invention relates to the use of a core-glycosylated HCV envelope protein as defined herein for inducing immunity against HCV in chronic HCV carriers prior to, simultaneously to or after any other therapy, such as, for example, the well-known interferon therapy either or not in combination with the administration of small drugs treating HCV, such as, for example, ribavirin. Such composition may also be employed before or after liver transplantation, or after presumed infection, such as, for example, needle-stick injury.

Another aspect of the invention relates to a method for the detection of the presence of anti-HCV antibodies in a sample suspected to comprise anti-HCV antibodies, said method comprising:

(i) contacting a HCV envelope protein or part thereof according to the invention with said sample under conditions allowing complexation of said HCV envelope protein or part thereof with said anti-HCV antibodies,

(ii) detecting the complex formed in (i), and

(iii) inferring from (ii) the presence of said anti-HCV antibodies in said sample.

In a specific embodiment, the contacting in step (i) of said method is occurring under competitive conditions. In another specific embodiment to said method, said HCV envelope protein or part thereof is attached to a solid support. In a further embodiment, said sample suspected to comprise anti-HCV antibodies is a biological sample.

A further aspect of the invention relates to a diagnostic kit for the detection of the presence of anti-HCV antibodies in a sample suspected to comprise anti-HCV antibodies, said kit comprising a HCV envelope protein or part thereof according to the invention. In a specific embodiment thereto, said HCV envelope protein or part thereof is attached to a solid support. In a further embodiment, said sample suspected to comprise anti-HCV antibodies is a biological sample.

The term "biological sample" as used herein, refers to a sample of tissue or fluid isolated from an individual, including but not limited to, for example, serum, plasma, lymph fluid, the external sections of the skin, respiratory-, intestinal- or genito-urinary tracts, oocytes, tears, saliva, milk, blood cells, tumors, organs, gastric secretions, mucus, spinal cord fluid, external secretions such as, for example, excrement, urine, sperm, and the like.

The HCV envelope proteins of the present invention, or the parts thereof, are particularly suited for incorporation into methods, such as immunoassay methods, for the detection of HCV, and/or genotyping of HCV, for prognosing/monitoring of HCV disease, or as a therapeutic agent.

The methods, such as immunoassay methods, according to the present invention utilize the HCV envelope proteins of the present invention that maintain linear (in case of peptides) and conformational epitopes, recognized by antibodies in the sera from individuals infected with HCV. The HCV E1 and E2 antigens of the present invention may be employed in virtually any assay format that employs a known antigen to detect antibodies. Of course, a format that denatures the HCV conformational epitope should be avoided or adapted. A common feature of all of these assays is that the antigen is contacted with the body component suspected of containing HCV antibodies under conditions that permit the antigen to bind to any such antibody present in the component. Such conditions will typically be physiologic temperature, pH and ionic strenght using an excess of antigen. The incubation of the antigen with the specimen is followed by detection of immune complexes comprised of the antigen.

Design of an immunoassays is subject to a great deal of variation, and many formats are known in the art. Protocols may, for example, use solid supports, or immunoprecipitation. Most assays involve the use of labeled antibody or polypeptide; the labels may be, for example, enzymatic, fluorescent, chemiluminescent, radioactive, or dye molecules. Assays which amplify the signals from the immune complex are also known; examples of which are assays which utilize biotin and avidin or streptavidin, and enzyme-labeled and mediated immunoassays, such as ELISA and RIA assays.

An immunoassay may be, without limitation, in a heterogeneous or in a homogeneous format, and of a standard or competitive type. In a heterogeneous format, the polypeptide is typically bound to a solid matrix or support to facilitate separation of the sample from the polypeptide after incubation. Examples of solid supports that can be used are nitrocellulose (e.g., in membrane or microtiter well form), polyvinyl chloride (e.g., in sheets or microtiter wells), polystyrene latex (e.g., in beads or microtiter plates, polyvinylidine fluoride (known as Immunolon™), diazotized paper, nylon membranes, activated beads, and Protein A beads. For

example, Dynatech Immunolon™ 1 or Immunlon™ 2 microtiter plates can be used in the heterogeneous format. The solid support containing the antigenic polypeptides is typically washed after separating it from the test sample, and prior to detection of bound antibodies. Both standard and competitive formats are know in the art.

In a homogeneous format, the test sample is incubated with the combination of antigens in solution. For example, it may be under conditions that will precipitate any antigen-antibody complexes which are formed. Both standard and competitive formats for these assays are known in the art.

In a standard format, the amount of antibodies, such as anti-HCV antibodies, in the antibody-antigen complexes is directly monitored. This may be accomplished by determining whether labeled anti-xenogeneic (e.g. anti-human) antibodies which recognize an epitope on said antibodies, such as said anti-HCV antibodies, will bind due to complex formation. In a competitive format, the amount of said antibodies, such as said anti-HCV antibodies, in a sample is deduced by monitoring the competitive effect on the binding of a known amount of (labeled) antibody (or other competing ligand) or antigen in the complex.

Antigen-antibody complexes can be detected by any of a number of known techniques, depending on the format. For example, unlabeled antibodies such as anti-HCV antibodies in the complex may be detected using a conjugate of anti-xenogeneic Ig complexed with a label (e.g. an enzyme label).

In an immunoprecipitation or agglutination assay format the reaction between an antigen and an antibody forms a protein cluster that precipitates from the solution or suspension and forms a visible layer or film of precipitate. If no antibody is present in the test specimen or sample, no such precipitate is formed.

The HCV envelope proteins, or specific parts thereof of the present invention comprised of conformational epitopes will typically be packaged in the form of a kit for use in these immunoassays. The kit will normally contain in separate containers the native HCV antigen, control antibody formulations (positive and/or negative), labeled antibody when the assay format requires the same and signal generating reagents (e.g. enzyme substrate) if the label does not generate a signal directly. The native HCV antigen may be already bound to a solid matrix or separate with reagents for binding it to the matrix. Instructions (e.g. written, tape, CD-ROM, etc.) for carrying out the assay usually will be included in the kit.

The solid phase selected can include polymeric or glass beads, nitrocellulose, microparticles, microwells of a reaction tray, test tubes and magnetic beads. The signal generating compound can include an enzyme, a luminescent compound, a chromogen, a

radioactive element and a chemiluminescent compound. Examples of enzymes include alkaline phosphatase, horseradish peroxidase and beta-galactosidase. Examples of enhancer compounds include biotin, anti-biotin and avidin. Examples of enhancer compounds binding members include biotin, anti-biotin and avidin. In order to block the effects of rheumatoid factor-like substances, the test sample is subjected to conditions sufficient to block the effect of rheumatoid factor-like substances. These conditions comprise contacting the test sample with a quantity of anti-human IgG to form a mixture, and incubating the mixture for a time and under conditions sufficient to form a reaction mixture product substantially free of rheumatoid factor-like substance.

In particular, the present invention relates to the use of an HCV envelope protein or part thereof according to the invention for the preparation of a diagnostic kit.

Since the a core-glycosylated HCV envelope proteins according to the present invention are highly immunogenic, and stimulate both the humoral and cellular immune response, the present invention further relates also to a kit for detecting HCV related T cell response, comprising the oligomeric particle or the purified single HCV envelope protein of the instant invention. HCV T cell response can for example be measured as described by Leroux-Roels et al. in WO95/12677.

A further aspect of the invention relates to a method of inducing a HCV-specific immune response in a mammal, said method comprising administering to said mammal an effective amount of a HCV envelope protein or part thereof according to the invention optionally comprising a pharmaceutically acceptable adjuvant. Said method comprising administering to said mammal an effective amount of a HCV envelope protein or part thereof according to the invention may also be used for inducing HCV-specific antibodies in a mammal or for inducing a specific T-cell function in a mammal. In said methods, said administering may be for prophylactic purposes, i.e. prophylactic administering or for therapeutic purposes, i.e. therapeutic administering.

Yet another aspect of the invention relates to a method of immunizing a mammal, said method comprising administering to said mammal an effective amount of a HCV envelope protein or part thereof according to the invention optionally comprising a pharmaceutically acceptable adjuvant.

The current invention also relates to a method of treating a mammal infected with HCV, said method comprising administering to said mammal an effective amount of a HCV envelope protein or part thereof according to the invention optionally comprising a pharmaceutically acceptable adjuvant.

Any of the above described aspects of the invention or the embodiments specific to said aspects is also applicable generally to proteins of interest which are the product of expression in a eukaryotic cells and which are further characterized by the same glycosylation properties as described above for the two different HCV envelope proteins.

More particularly, the invention thus relates to an isolated protein of interest or a fragment thereof comprising at least one N-glycosylation site, said protein or fragment thereof characterized in that it is the product of expression in a eukaryotic cell and further characterized in that on average up to 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, or 80% of the N-glycosylated sites are core-glycosylated. More specific thereto, more than 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94% or 95 % of the N-glycosylated sites are glycosylated with an oligomannose with a structure defined by Man(8 to 10)-GlcNAc(2). More specific to any of the above N-glycosylation characteristics, the ratio of sites core-glycosylated with an oligomannose with structure Man(7)-GlcNAc(2) over the sites core-glycosylated with an oligomannose with structure Man(8)-GlcNAc(2) is less than or equal to 0.15, 0.2, 0.25, 0.30, 0.35, 0.40, 0.45, or 0.50. Further more specific to any of the above N-glycosylation characteristics, said oligomannoses contain less than 20%, 19%, 18%, 17%, 16%, 15%, 14%, 13%, 12%, 11%, 10%, 9%, 8%, 7%, 6%, or 5% terminal α1,3 mannose.

Another alternative aspect of the invention is related to an isolated protein of interest or a fragment thereof comprising at least one N-glycosylation site, said protein or fragment thereof characterized in that it is the product of expression in a eukaryotic cell and further characterized in that N-glycosylated sites are occupied by oligomannoses wherein the ratio of the oligomannoses with structure Man(7)-GlcNAc(2) over the oligomannoses with structure Man(8)-GlcNAc(2) is less than or equal to 0.15, 0.2, 0.25, 0.30, 0.35, 0.40, 0.44, 0.45, or 0.50. Further more specific to the above N-glycosylation characteristics, said oligomannoses contain less than 20%, 19%, 18%, 17%, 16%, 15%, 14%, 13%, 12%, 11%, 10%, 9%, 8%, 7%, 6%, or 5% terminal α1,3 mannose.

In particular, said isolated protein of interest or fragment thereof is the product of expression in a yeast cell such as a *Hansenula* cell. Said isolated protein of interest or fragment thereof can e.g. be a viral envelope protein or a fragment thereof such as a HCV envelope protein or HBV (hepatitis B) envelope protein, or fragments thereto. Other examplary viral envelope proteins include the HIV (human immunodeficiency virus) envelope protein gp120 and viral envelope proteins of a virus belonging to the Flavirideae. In general, said isolated protein of interest or fragment thereof can be any protein needing the N-glycosylation characteristics of the current invention.

With "HCV-recombinant vaccinia virus" is meant a vaccinia virus comprising a nucleic acid sequence encoding a HCV protein or part thereof.

The terms "HCV virus-like particle formed of a HCV envelope protein" "oligomeric particles formed of HCV envelope proteins" are herein defined as structures of a specific nature and shape containing several basic units of the HCV E1 and/or E2 envelope proteins, which on their own are thought to consist of one or two E1 and/or E2 monomers, respectively. It should be clear that the particles of the present invention are defined to be devoid of infectious HCV RNA genomes. The particles of the present invention can be higher-order particles of spherical nature which can be empty, consisting of a shell of envelope proteins in which lipids, detergents, the HCV core protein, or adjuvant molecules can be incorporated. The latter particles can also be encapsulated by liposomes or apolipoproteins, such as, for example, apolipoprotein B or low density lipoproteins, or by any other means of targeting said particles to a specific organ or tissue. In this case, such empty spherical particles are often referred to as "virus-like particles" or VLPs. Alternatively, the higher-order particles can be solid spherical structures, in which the complete sphere consists of HCV E1 or E2 envelope protein oligomers, in which lipids, detergents, the HCV core protein, or adjuvant molecules can be additionally incorporated, or which in turn may be themselves encapsulated by liposomes or apolipoproteins, such as, for example, apolipoprotein B, low density lipoproteins, or by any other means of targeting said particles to a specific organ or tissue, e.g. asialoglycoproteins. The particles can also consist of smaller structures (compared to the empty or solid spherical structures indicated above) which are usually round (see further)-shaped and which usually do not contain more than a single layer of HCV envelope proteins. A typical example of such smaller particles are rosette-like structures which consist of a lower number of HCV envelope proteins, usually between 4 and 16. A specific example of the latter includes the smaller particles obtained with E1s in 0.2% CHAPS as exemplified herein which

apparently contain 8-10 monomers of E1s. Such rosette-like structures are usually organized in a plane and are round-shaped, e.g. in the form of a wheel. Again lipids, detergents, the HCV core protein, or adjuvant molecules can be additionally incorporated, or the smaller particles may be encapsulated by liposomes or apolipoproteins, such as, for example, apolipoprotein B or low density lipoproteins, or by any other means of targeting said particles to a specific organ or tissue. Smaller particles may also form small spherical or globular structures consisting of a similar smaller number of HCV E1 or E2 envelope proteins in which lipids, detergents, the HCV core protein, or adjuvant molecules could be additionally incorporated, or which in turn may be encapsulated by liposomes or apolipoproteins, such as, for example, apolipoprotein B or low density lipoproteins, or by any other means of targeting said particles to a specific organ or tissue. The size (i.e. the diameter) of the above-defined particles, as measured by the well-known-in-the-art dynamic light scattering techniques (see further in examples section), is usually between 1 to 100 nm, more preferentially between 2 to 70 nm, even more preferentially between 2 and 40 nm, between 3 to 20 nm, between 5 to 16 nm, between 7 to 14 nm or between 8 to 12 nm.

In particular, the present invention relates to a method for purifying core glycosylated hepatitis C virus (HCV) envelope proteins, or any part thereof, suitable for use in an immunoassay or vaccine, which method comprising:

(i) growing Hansenula or Saccharomyces glycosylation minus strains transformed with an envelope gene encoding an HCV E1 and/or HCV E2 protein, or any part thereof, in a suitable culture medium;

(ii) causing expression of said HCV E1 and/or HCV E2 gene, or any part thereof; and

(iii) purifying said core-glycosylated HCV E1 and/or HCV E2 protein, or any part thereof, from said cell culture.

The invention further pertains to a method for purifying core-glycosylated hepatitis C virus (HCV) envelope proteins, or any part thereof, suitable for use in an immunoassay or vaccine, which method comprising:

(i) growing Hansenula or Saccharomyces glycosylation minus strains transformed with an envelope gene encoding an HCV E1 and/or HCV E2 protein, or any part thereof, in a suitable culture medium;

(ii) causing expression of said HCV E1 and/or HCV E2 gene, or any part thereof; and

(iii) purifying said intracellularly expressed core-glycosylated HCV E1 and/or HCV E2 protein, or any part thereof, upon lysing the transformed host cell.

The invention further pertains to a method for purifying core-glycosylated hepatitis C virus (HCV) envelope proteins, or any part thereof, suitable for use in an immunoassay or vaccine, which method comprising:

(i) growing Hansenula or Saccharomyces glycosylation minus strains transformed with an envelope gene encoding an HCV E1 and/or HCV E2 protein, or any part thereof, in a suitable culture medium, in which said HCV E1 and/or HCV E2 protein, or any part thereof, comprises at least two Cys-amino acids;

(ii) causing expression of said HCV E1 and/or HCV E2 gene, or any part thereof; and

(iii) purifying said core-glycosylated HCV E1 and/or HCV E2 protein, or any part thereof, in which said Cys-amino acids are reversibly protected by chemical and/or enzymatic means, from said culture.

The invention further pertains to a method for purifying core-glycosylated hepatitis C virus (HCV) envelope proteins, or any part thereof, suitable for use in an immunoassay or vaccine, which method comprising:

(i) growing Hansenula or Saccharomyces glycosylation minus strains transformed with an envelope gene encoding an HCV E1 and/or HCV E2 protein, or any part thereof, in a suitable culture medium, in which said HCV E1 and/or HCV E2 protein, or any part thereof, comprises at least two Cys-amino acids;

(ii) causing expression of said HCV E1 and/or HCV E2 gene, or any part thereof; and,

(iii) purifying said intra-cellulary expressed core-glycosylated HCV E1 and/or HCV E2 protein, or any part thereof, upon lysing the transformed host cell, in which said Cys-amino acids are reversibly protected by chemical and/or enzymatic means.

The present invention specifically relates to a method for purifying recombinant core-glycosylated HCV yeast proteins, or any part thereof, as described herein, in which said purification includes heparin affinity chromatography.

Hence, the present invention also relates to a method for purifying recombinant core-glycosylated HCV yeast proteins, or any part thereof, as described above, in which said chemical means is sulfonation.

Hence, the present invention also relates to a method for purifying recombinant core-glycosylated HCV yeast proteins, or any part thereof, as described above, in which said reversibly protection of Cys-amino acids is exchanged for an irreversible protection by chemical and/or enzymatic means.

Hence, the present invention also relates to a method for purifying recombinant core-glycosylated HCV yeast proteins, or any part thereof, as described above, in which said irreversible protection by chemical means is iodo-acetamide.

Hence, the present invention also relates to a method for purifying recombinant core-glycosylated HCV yeast proteins, or any part thereof, as described above, in which said irreversible protection by chemical means is NEM or Biotin-NEM or a mixture thereof.

The present invention also relates to a composition as defined above which also comprises HCV core, E1, E2, P7, NS2, NS3, NS4A, NS4B, NS5A and/or NS5B protein, or parts thereof. The core-glycosylated proteins E1, E2, and/or E1/E2 of the present invention may, for example, be combined with other HCV antigens, such as, for example, core, P7, NS3, NS4A, NS4B, NS5A and/or NS5B. The purification of these NS3 proteins will preferentially include a reversible modification of the cysteine residues, and even more preferentially sulfonation of cysteines. Methods to obtain such a reversible modification, including sulfonation have been described for NS3 proteins in Maertens et al. (PCT/EP99/02547). It should be stressed that the whole content, including all the definitions, of the latter document is incorporated by reference in the present application.

Also, the present invention relates to the use of a core-glycosylated envelope protein as described herein for inducing immunity against HCV, characterized in that said core-glycosylated envelope protein is used as part of a series of time and compounds. In this regard, it is to be understood that the term "a series of time and compounds" refers to administering with time intervals to an individual the compounds used for eliciting an immune response. The latter compounds may comprise any of the following components: a core-glycosylated envelope protein, HCV DNA vaccine composition, HCV polypeptides.
In this respect, a series comprises administering, either:

(i) an HCV antigen, such as, for example, a core-glycosylated envelope protein, with time intervals, or

(ii) an HCV antigen, such as, for example, a core-glycosylated envelope protein in combination with a HCV DNA vaccine composition, in which said core-glycosylated envelope protein oligomeric particles and said HCV DNAvaccine composition, can be administered simultaneously, or at different time intervals, including at alternating time intervals, or

(iii) either (i) or (ii), possibly in combination with other HCV peptides, with time intervals.

In this regard, it should be clear that a HCV DNA vaccine composition comprises nucleic acids encoding HCV envelope peptide, including E1-, E2-, E1/E2-peptides, NS3 peptide, other HCV peptides, or parts of said peptides. Moreover, it is to be understood that said HCV peptides comprises HCV envelope peptides, including E1-, E2-, E1/E2-peptides, other HCV peptides, or parts thereof. The term "other HCV peptides" refers to any HCV peptide or fragment thereof. In item (ii) of the above scheme, the HCV DNA vaccine composition comprises preferentially nucleic acids encoding HCV envelope peptides. In item (ii) of the above scheme, the HCV DNA vaccine composition consists even more preferentially of nucleic acids encoding HCV envelope peptides, possibly in combination with a HCV-NS3 DNA vaccine composition. In this regard, it should be clear that an HCV DNA vaccine composition comprises a plasmid vector comprising a polynucleotide sequence encoding an HCV peptide as described above, operably linked to transcription regulatory elements. As used herein, a "plasmid vector" refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. Preferred vectors are those capable of autonomous replication and/or expression of nucleic acids to which they have been linked. In general, but not limited to those, plasmid vectors are circular double stranded DNA loops which, in their vector form, are not bound to the chromosome. As used herein, a "polynucleotide sequence" refers to polynucleotides such as deoxyribonucleic acid (DNA), and, where appropriate, ribonucleic acid (RNA). The term should also be understood to include, as equivalents, analogs of either RNA or DNA made from nucleotide analogs, and single (sense or antisense) and double-stranded polynucleotides. As used herein, the term "transcription regulatory elements" refers to a nucleotide sequence which contains essential regulatory elements, such that upon introduction into a living vertebrate cell it is able to direct the cellular machinery to produce translation products encoded by the polynucleotide. The term "operably linked" refers to a juxtaposition wherein the components are configured so as to perform their usual function. Thus, transcription regulatory elements operably linked to a nucleotide sequence are capable of effecting the expression of said nucleotide sequence.

Those skilled in the art can appreciate that different transcriptional promoters, terminators, carrier vectors or specific gene sequences may be used succesfully.
Alternatively, the DNA vaccine may be delivered through a live vector such as adenovirus, canary pox virus, MVA, and the like.

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The present invention is illustrated by the Examples as set forth below. These Examples are merely illustrative and are not construed to restrict or limit the invention in any way.

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EXAMPLES

EXAMPLE 1

CONSTRUCTION OF pFPMT-MFα-E1-H6 SHUTTLE VECTOR

Plasmids for *Hansenula polymorpha* transformation were constructed as follows. The pFPMT-MFα-E1-H6 shuttle vector has been constructed in a multi-step procedure. Intially the nucleic acid sequence encoding the HCV E1s protein (SEQ ID NO:2) was cloned after a CHH leader sequence (CHH = *Carcinus maenas* hyperglycemic hormone) which was subsequently changed for a MFα leader sequence (MFα = *Saccharomyces cerevisiae* α-mating factor).

At first a pUC18 derivative has been constructed harboring the CHH-E1-H6 unit as a *Eco*RI/*Bam*HI fragment by the seamless cloning method (Padgett, K. A. and Sorge, J. A. 1996). Thereto, the E1s-H6-encoding DNA fragment and the pCHH-Hir-derived acceptor plasmid were generated by PCR as described below.

Generation of E1s-H6-encoding DNA fragment

The E1-H6 DNA fragment (coding for HCV type 1b E1s protein consisting of the amino acids 192 to 326 of E1s elongated with 6 His-residues; SEQ ID NO:5) was isolated by PCR from the plasmid pGEMTE1sH6 (SEQ ID NO:6; Figure 1). The following primers were used thereto:

- CHHE1-F:5′-agttactcttca.**aggtatgaggtgcgcaacgtgtccg**-3′ (SEQ ID NO:7);

  The *Eam*1104I site is underlined, the dot marks the cleavage site. The bold printed bases are complementary to those of primer CHH-links. The non-marked bases anneal within the start region of E1 (192-326) in sense direction; and

- CHHE1-R: 5′- agttactcttca.**cagggatcctccttaatggtgatggtggtggtgcc**-3′ (SEQ ID NO: 8);

  The *Eam*1104I site is underlined, the dot marks the cleavage site. The bold printed bases are complementary to those of primer MF30-rechts. The bases forming the *Bam*HI site

usefull for later cloning procedures are printed in italics. The non-marked bases anneal in antisense direction within the end of the *E1-H6* unit, including the stop codon and three additional bases between the stop codon and the *Bam*HI site.

The reaction mixture was constituted as follows: total volume of 50 µL containing 20 ng of *Eco*311-linearized pGEMTE1sH6, each 0.2 µM of primers CHHE1-F and CHHE1-R, dNTP's (each at 0.2 µM), 1 x buffer 2 (Expand Long Template PCR System; Boehringer; Cat No 1681 834), 2.5 U polymerase mix (Expand Long Template PCR System; Boehringer; Cat No 1681 834).

Program 1 was used, said program consisting of the following steps:

1. denaturation: 5min 95°C;
2. 10 cycles of 30 sec denaturation at 95°C, 30 sec annealing at 65°C, and 130 sec elongation at 68°C
3. termination at 4°C.

Then 5 µL 10 x buffer 2 (Expand Long Template PCR System; Boeringer; Cat No 1681 834), 40 µL H_2O, and 5 µL of [dATP, dGTP, and dTTP (2mM each); 10mM 5-methyl-dCTP] were added to the sample derived from program 1, and further amplification was performed following program 2 consisting of the following steps:

1. denataruation: 5 min at 95°C
2. 5 cycles of 45 sec denaturation at 95°C, 30 sec annealing at 65°C, and 130 sec at 68°C
3. termination at 4°C.

Generation of pCHH-Hir-derived acceptor plasmid

The acceptor fragment was made by PCR from the pCHH-Hir plasmid (SEQ ID NO:9; Figure 2) and consists of almost the complete pCHH-Hir plasmid, except that the Hir-coding sequence is not present in the PCR product. Following primers were used for this PCR:

1. CHH-links: 5'-agtta<u>ctcttc</u>a.cctcttttccaacgggtgtgtag-3'
   (SEQ ID NO:10);
   The *Eam*1104I site is underlined, the dot marks the cleavage site. The bold printed bases are complementary to those of primer CHHE1-F. The non-marked bases anneal within the end of the CHH sequence in antisense direction; and
2. MF30-rechts: 5'-agtca<u>ctcttc</u>a.ctgcaggcatgcaagcttggcg-3'
   (SEQ ID NO:11);
   The *Eam*1104I site is underlined, the dot marks the cleavage site. The bold printed bases are complementary to those of primer CHHE1-R. The non-marked bases anneal within the

pUC18 sequences behind the cloned *CHH-Hirudin HL20* of pCHH-Hir, pointing away from the insert.

The reaction mixture was constituted as follows: total volume of 50 µL containing 20 ng of *Asp*718I-linearized pCHH-Hir, each 0.2 µM of primers CHH-links and MF30-rechts, dNTP's (each at 0.2 µM), 1 x buffer 2 (Expand Long Template PCR System; Boeringer; Cat No 1681 834), 2.5 U polymerase mix (Expand Long Template PCR System; Boeringer; Cat No 1681 834).

Program 1 was as described above was used.

Then 5 µL 10 x buffer 2 (Expand Long Template PCR System; Boeringer; Cat No 1681 834), 40 µL H_2O, and 5 µL of [dATP, dGTP, and dTTP (2mM each); 10mM 5-methyl-dCTP] were added to the sample derived from program 1, and further amplification was performed following program 2 as described above.

Generation of vector pCHHE1

The E1s-H6-encoding DNA fragment and the pCHH-Hir-derived acceptor plasmid generated by PCR as described above were purified using the PCR product purification kit (Qiagen) according to the supplier's specifications. Subsequently the purified fragments were digested separately with *Eam*1104I. Subsequently, the E1s-H6 DNA fragment was ligated into the pCHH-Hir-derived acceptor plasmid using T4 ligase (Boehringer) following the specifications of the supplier.

E. coli XL-Gold cells were transformed with the ligation mixture and the plasmid DNA of several ampicillin-resistant colonies were analyzed by digestion with *Eco*RI and *Bam*HI. A positive clone was selected and denominated as pCHHE1.

Generation of vector pFPMT-CHH-E1H6

The *Eco*RI/*Bam*HI fragment of pCHHE1 was ligated with the *Eco*RI/*Bam*HI digested vector pFPMT121 (SEQ ID NO:12; Figure 3). T4 ligase (Boehringer) was used according to the supplier's instructions. The ligation mixture was used to transform *E. coli* DH5αF'cells. Several transformants were analyzed on restriction pattern of the plasmid DNA and a positive clone was withheld which was denominated pFPMT-CHH-E1H6 (SEQ ID NO:13; Figure 4).

Generation of pFPMT-MFα-E1-H6

Finally the shuttle vector pFPMT-MFα-E1-H6 was generated by ligation of three fragments, said fragments being:

1. the 6.961 kb *Eco*RI/*Bam*HI digested pFPMT121 (SEQ ID NO:12; Figure 3),
2. the 0.245 *Eco*RI/*Hind*III fragment of pUC18-MFa (SEQ ID NO:62; Figure 36), and
3. the 0.442 kb *Hind*III/*Bam*HI fragment of a 0.454 kb PCR product derived from pFPMT-CHH-E1H6.

The 0.454 kb PCR product giving rise to fragment No.3 was obtained by PCR using the following primers:

1. primer MFa-E1 f-Hi:
   5′-aggggtaagcttggataaaaggtatgaggtgcgcaacgtgtccgggatgt-3′ (SEQ ID NO:14); and
2. primer E1 back-Bam:
   5′-agttacggatccttaatggtgatggtggtggtgccagttcat-3′
   (SEQ ID NO:15).

The reaction mixture was constituted as follows: Reaction mixture volume 50 µL, pFPMT-CHH-E1-H6 (*Eco*RI-linearized; 15 ng/µL), 0.5 µL; primer MFa-E1 f-Hi (50 µM), 0.25 µL; primer E1 back-Bam (50 µM), 0.25 µL; dNTP′s (all at 2mM), 5 µL; DMSO, 5µL; H_2O, 33.5 µL; Expand Long Template PCR System (Boeringer Mannheim; Cat No 1681 834) Buffer 2 (10 x concentrated), 5 µL; Expand Long Template PCR System Polymerase mixture (1 U/µL), 0.5 µL.

The PCR program consisting of the following steps was used:

1. denaturation: 5 min at 95°C
2. 29 cycles of 45 sec denaturation at 95°C, 45 sec annealing at 55°C, and 40 sec elongation at 68°C
3. termination at 4°C.

Based on the primers used, the resulting 0.454kb PCR product contained the codons of E1(192-326) followed by six histidine codons and a "taa" stop codon, upstream flanked by the 22 3′-terminal base pairs of the MFα prepro sequence (including the cloning relevant *Hind*III site plus a six base pairs overhang) and downstream flanked by a (cloning relevant) *Bam*HI site and a six base pairs overhang.

For the ligation reaction, T4 DNA ligase (Boehringer Mannheim) has been used according to the supplier's conditions (sample volume 20 µL).

E.coli HB101 cells were transformed with the ligation mixture and positive clones withheld after restriction analysis of the plasmids isolated from several transformants. A positive plasmid was selected and denominated as pFPMT-MFα-E1-H6 (SEQ ID NO:16; Figure 5).

EXAMPLE 2

CONSTRUCTION OF pFPMT-CL-E1-H6 SHUTTLE VECTOR

Plasmids for *Hansenula polymorpha* transformation were constructed as follows. The pFPMT-CL-E1-H6 shuttle vector was constructed in three steps starting from pFPMT-MFα-E1-H6 (SEQ ID NO:16, Figure 5).

In a first step, the MFα-E1-H6 reading frame of pFPMT-MFα-E1-H6 was subcloned into the pUC18 vector. Therefore a 1.798kb *SalI/Bam*HI fragment of pFPMT-MFα-E1-H6 (containing the FMD promotor plus MFα-E1-H6) was ligated to the *SalI/Bam*HI vector fragment of pUC18 with T4 ligase (Boehringer) accordig to the supplier's conditions. This resulted in plasmid that is depicted in Figure 6 (SEQ ID NO:17), and further denominated as pMa12-1 (pUC18-FMD-MFα-E1-H6). The ligation mixture was used to transform *E.coli* DH5αF' cells. Several ampicillin-resistant colonies were picked and analyzed by restriction enzyme digestion of plasmid DNA isolated from the picked clones. A positive clone was further analyzed by determining the DNA sequence of the MFα-E1-H6 coding sequence. A correct clone was used for PCR directed mutagenesis to replace the MFα pre-pro-sequence with the codons of the avian lysozyme pre-sequence ("CL"; corresponding to amino acids 1 to 18 of avian lysozyme; SEQ ID NO:1). The principle of the applied PCR-directed mutagenesis method is based on the amplification of an entire plasmid with the desired alterations located at the 5'-ends of the primers. In downstream steps, the ends of the linear PCR product are modified prior to self-ligation resulting in the desired altered plasmid.

The following primers were used for the PCR reaction:

1. primer CL hin: 5'-tgcttcctaccactagcagcactaggatatgaggtgcgcaacgtgtccggg-3' (SEQ ID NO:18);

2. primer CL her neu: 5′-tag*tactagt*attagtaggcttcgcatgaattcccgatgaaggcagagagcg-3′ (SEQ ID NO:19).

The underlined 5′ regions of the primers contain the codons of about half of the avian lysozyme pre-sequence. Primer CL her neu includes a *Spe*I restriction site (italic). The non-underlined regions of the primers anneal with the codons for amino acid residues 192 to 199 of E1 (CL hin) or the with the "atg" start codon over the *Eco*RI site up to position -19 (counted from the *Eco*RI site) of *FMD* promoter. The primers are designed to amplify the complete pMa12-1 thereby replacing the codons of the MF*a* pre-pro-sequence with the codons of the avian lysozyme pre sequence.

The reaction mixture was constituted as follows: pUC18-FMD-Mfα-E1-H6 (pMa12-1; 1.3 ng/μL), 1 μL; primer CL hin (100 μM), 2μL; primer CL her neu (100 μM), 2 μL; dNTP's (all at 2.5mM), 8 μL; H_2O, 76 μL; Expand Long Template PCR System (Boeringer; Cat No 1681 834) Buffer 2 (10 x concentrated), 10μL; Expand Long Template PCR System Polymerase mixture (1 U/μL), 0.75 μL.

The PCR program consisting of the following steps was applied:

1. denaturation: 15 min at 95°C
2. 35 cycles of 30 sec denaturation at 95°C, 1 min annealing at 60°C, and 1 min elongation at 72°C
3. termination at 4°C.

The resulting PCR product was checked by agarose gel electrophoresis for its correct size (3.5 kb). Thereafter the 3'-A overhangs form the PCR product were removed by a T4 polymerase reaction resulting in blunt ends with 3'- and 5'-OH-groups. Therefore, the PCR product was treated with T4 polymerase (Boehringer; 1 U/μL): to the remaining 95 μL of PCR reaction mix were added 1 μL T4 polymerase and 4 μL dNTP's (all at 2.5 mM). The sample was incubated for 20 min at 37°C. Subsequently, the DNA was precipitated with ethanol and taken up in 16 μL H_2O.

Subsequently 5'-phosphates were added to the blunt-ended PCR product by a kinase reaction. Therefore, to the 16 μL blunt-ended PCR product were added 1 μL T4 polynucleotide kinase (Boehringer; 1U/μL), 2 μL 10-fold concentrated T4 polynucleotide kinase reaction buffer (Boehringer), and 1 μL ATP (10mM). The sample was incubated for 30 min at 37°C.

Subsequently the DNA was applied onto a 1% agarose gel and the correct product band was isolated by means of the gel extraction kit (Qiagen) according to the supplier's conditions. Fifty (50) ng of the purified product was then self-ligated by use of T4 ligase (Boehringer)

according to the supplier's conditions. After 72 h incubation at 16°C, the DNA in the ligation mix was precipitated with ethanol and dissolved in 20 µL water.

E.coli DH5α-F' cells were subsequently transformed with 10 µL of the ligation sample. The plasmid DNA of several ampicillin-resistant clones was checked by means of restriction enzyme digestion. A positive clone was withheld and denominated p27d-3 (pUC18-FMD-CL-E1-H6, SEQ ID NO:20, Figure 7). Subsequently the CL-E1-H6 reading frame was verified by DNA sequencing.

In a last step the pFPMT-CL-E1-H6 shuttle vector was constructed as described below. The 0.486kb *Eco*RI/*Bam*HI fragment of p27d-3 (harboring CL-E1(192-326)-H6) was ligated with *Eco*RI/*Bam*HI-digested pFPMT121 (SEQ ID NO:12, Figure 3). For the reaction, T4 ligase (Boehringer) has been used according to the supplier's recommendations. The DNA in the ligation sample was precipitated with ethanol and dissolved in 10 µL H_2O. *E. coli* DH5α F' cells were transformed with 10 µL of the ligation sample, and the plasmid DNA of several ampicillin-resistant colonies were analyzed by digestion with *Eco*RI and *Bam*HI. Plasmid clone p37-5 (pFPMT-CL-E1-H6; SEQ ID NO:21, Figure 8) showed the desired fragment sizes of 0.486kb and 6.961kb. The correct sequence of *CL-E1-H6* of p37-5 was verified by sequencing.

EXAMPLE 3

CONSTRUCTION OF pFPMT-MFα-E2-H6 AND pMPT-MFα-E2-H6 SHUTTLE VECTORS

Plasmids for *Hansenula polymorpha* transformation were constructed as follows. The DNA sequence encoding the MFα-E2s (amino acids 384-673 of HCV E2)-VIEGR-His6 (SEQ ID NO:5) was isolated as a 1.331kb *Eco*RI/*Bgl*II fragment from plasmid pSP72E2H6 (SEQ ID NO:22, Figure 9).This fragment was ligated with either the *Eco*RI/*Bgl*II-digested vectors pFPMT121 (SEQ ID NO:12, Figure C+2) or pMPT121 (SEQ ID NO:23, Figure 10) using T4 DNA ligase (Boehringer Mannheim) according to the supplier's recommendations. After transformation of *E. coli* and checking of plasmid DNA isolated from different transformants by restriction enzyme digestion, positive clones were withheld and the resulting shuttle

vectors are denominated pFPMT-MFα-E2-H6 (SEQ ID NO:22, Figure 11) and pMPT-MFα-E2-H6 (SEQ ID NO:23, Figure 12), respectively.

EXAMPLE 4

CONSTRUCTION OF pFPMT-CL-E2-H6 SHUTTLE VECTOR

The shuttle vector pFPMT-CL-E2-H6 was assembled in a three-step procedure. An intermediate construct was prepared in which the E2 coding sequence was cloned behind the signal sequence of α-amylase of *Schwanniomyces accidentalis*. This was done by the seamless cloning method (Padgett, K. A. and Sorge, J. A. 1996).

Generation of E2s-H6 encoding DNA fragment

At first the DNA sequence encoding E2-H6 (amino acids 384 to 673 of HCV E2 extended with the linker peptide "VIEGR" and with 6 His residues, SEQ ID NO:5) was amplified from the pSP72E2H6 plasmid (SEQ ID NO:24, Figure 11) by PCR. The used primers were denoted MF30E2/F and MF30E2/R and have the following sequences:

- primer MF30E2/F: 5′-agtcactcttca.aggcatacccgcgtgtcaggaggg-3′ (SEQ ID NO:26; the *Eam*1104I site is underlined, the dot marks the enzyme's cleavage site; the last codon of the *S. occidentalis* signal sequence is printed in bold; the non- marked bases anneal with the codons of E2 (amino acids 384-390 of HCV E2);
- primer MF30E2/R: 5′-agtcactcttca.cag*ggatcc*ttagtgatggtggtgatg-3′ (SEQ ID NO:27; the *Eam*1104I site is underlined, the dot marks the enzyme's cleavage site; the bold printed bases are complementary to the bold printed bases of primer MF30-Rechts (see below); a *Bam*HI site to be introduced into the construct is printed in italic; the non- marked sequence anneals with the stop codon and the six terminal His codons of E2 (384-673)-VIEGR-H6 (SEQ ID NO:5).

The reaction mixture was constituted as follows: total volume of 50 µL containing 20 ng of the 1.33kb *Eco*RI/*Bgl*II fragment of pSP72E2H6, each 0.2 µM of primers MF30E2/F and MF30E2/R, dNTP's (each 0.2 µM), 1x buffer 2 (Expand Long Template PCR System;

Boeringer; Cat No 1681 834), 2.5 U polymerase mix (Expand Long Template PCR System; Boeringer; Cat No 1681 834).

The PCR program 3 consisting of the following steps was used:

1. denaturation: 5 min at 95°C
2. 10 cycles of 30 sec denaturation at 95°C, 30 sec annealing at 65°C, and 1 min elongation at 68°C
3. termination at 4°C.

Then 10 µL 10 x buffer 2 (Expand Long Template PCR System; Boeringer; Cat No 1681 834), 40 µL H_2O, and 5 µL of [dATP, dGTP, and dTTP (2mM each); 10mM 5-methyl-dCTP] have been added to the sample derived from PCR program 3, and it has been continued with PCR program 4 consisting of the following steps:

1. denaturation: 5 min at 95°C
2. 5 cycles of 45 sec denaturation at 95°C, 30 sec annealing at 65°C, and 1 min elongation at 68°C
3. termination at 4°C.

Generation of pMF30-derived acceptor plasmid

The second fragment originated from the plasmid pMF30 (SEQ ID NO:28, Figure 13), the amplicon was almost the complete pMF30 plasmid excluding the codons of the mature α-amylase of *S.occidentalis*, modifications relevant for cloning were introduced by primer design. The following set of primers was used:

- primer MF30-Links:
  5′-agtcactcttca.cctcttgtcaaaaataatcggttgag-3′ (SEQ ID NO:29; the *Eam*1104I site is underlined, the dot marks the enzyme′s cleavage site; the bold printed "cct" is complementary to the bold printed "agg" of primer MF30E2/F (see above); the non-marked and the bold printed bases anneal with the 26 terminal bases of the codons of the α-Amylase of *S. occidentalis* in pMF30);
- primer MF30-Rechts: 5′-agtcactcttca.ctgcaggcatgcaagcttggcg-3′ (SEQ ID NO:11; the *Eam*1104I site is underlined, the dot marks the enzyme′s cleavage site; the bold printed "ctg" is complementary to the bold printed "cag" of primer MF30E2/R (see above); the non-marked bases anneal with pUC18 sequences downstream of the stop codon of the α-Amylase of *S. occidentalis* in pMF30).

The reaction mixture was constituted as follows: total volume of 50 µL containing 20 ng of the *Bgl*II-linearized pMF30, each 0.2 µM of primers MF30-Links and MF30-Rechts, dNTP's (each 0.2 µM), 1x buffer 1 (Expand Long Template PCR System; Boeringer; Cat No 1681 834), 2.5 U polymerase mix (Expand Long Template PCR System; Boeringer; Cat No 1681 834). The same PCR programs (programs 3 and 4) as described above were used, except for the elongation times which were extended from 1 minute to 4 minutes in both programs.

Generation of vector pAMY-E2

The E2s-H6 encoding DNA fragment and pMF30-derived acceptor plasmid obtained by PCR were controlled on their respective size by gel electrophoresis on a 1 % agarose gel. The PCR products were purified with a PCR product purification kit (Qiagen) according to the supplier's instructions. Subsequently the purified fragments were digested separately with *Eam*11004I. Ligation of the E2s-H6 fragment with the pMF30-derived acceptor plasmid was performed by using T4 ligase (Boehringer) according to the supplier's recommendations. The ligation mixture was used to transform *E.coli* DH5αF'cells and the plasmid DNA of several clones was analyzed by *Eco*RI/*Bam*HI digestion. A positive clone was selected, its plasmid further denominated as pAMY-E2, and utilized for further modifications as described below.

Generation of vector pUC18-CL-E2-H6

The pAMY-E2 was subjected to PCR-directed mutagenesis in order to replace the codons of the α-amylase signal sequence with the codons of the avian lysozyme pre sequence. This is further denominated as "CL", corresponding to the first 18 amino acids of avian lysozyme ORF (SEQ ID NO:1). For this mutagenesis following primers were used:

- primer CL2 hin:
  5'-tgcttcctaccactagcagcactaggacatacccgcgtgtcaggaggggcag-3' (SEQ ID NO:30); and
- primer CL2 her:
  5'-tagtactagtattagtaggcttcgcatggaattcactggccgtcgtttta-caacgtc-3' (SEQ ID NO:31).

The underlined 5'-regions of the primers contain the DNA sequence of about half of the avian lysozyme pre sequence. Primer CL2 her includes *Spe*I (italic) and *Eco*RI (italic, double underlined) restriction sites. The non-underlined regions of the primers anneal with the codons of amino acid residues 384 to 392 of E2 (CL2 hin) or the with the "atg" start codon

over the *Eco*RI site up to position -19 (counted from the *Eco*RI site) of *FMD* promoter. The primers are designed to amplify the complete pAMY-E2 vector thereby replacing the codons of the α-amylase signal sequence with the codons of the avian lysozyme pre-sequence.

The PCR reaction was performed according to the following program:

1. denaturation: 15 min at 95°C
2. 35 cycles of 30 sec denaturation at 95°C, 1 min annealing at 60°C, and 1 min elongation at 72°C
3. termination at 4°C.

The following reaction mixture was used: pAMY-E2 (1 ng/μL), 1μL; primer CL2 hin (100 μM), 2 μL; primer CL2 her (100 μM), 2 μL; dNTP's (2.5mM each), 8 μL; H_2O, 76 μL; Expand Long Template PCR System (Boeringer; Cat No 1681 834) Buffer 2 (10 x concentrated), 10 μL; Expand Long Template PCR System Polymerase mixture (1U/μL), 0.75 μL.

The resulting PCR product was checked by gel electrophoresis on a 1% agarose gel. Prior to ligation the PCR fragment was modified as follows. The 3'-A overhangs were removed by T4 polymerase resulting in blunt ends with 3'- and 5'-OH-groups. Thereto 1 μL T4 polymerase (Boehringer, 1U/μL) was added to the residual 95 μL PCR reaction mixture along with 4 μL dNTP's (2.5 mM each). The sample was incubated for 20 min at 37°c. Subsequently the DNA was precipitated with ethanol and dissolved in 16 μL deionized water. This was followed by a kinase treatment to add 5'-phosphates to the blunt-ended PCR product. To the 16 μL dissolved blunt-ended PCR product were added 1 μL T4 polynucleotide kinase (Boehringer, 1U/μL), 2 μL 10-fold concentrated T4 polynucleotide kinase reaction buffer (Boehringer) and 1 μL ATP (10 mM). The sample was incubated for 30 min at 37°C.

The kinase treated sample was subsequently separated on a 1% agarose gel. The product band was isolated. The DNA was extracted from the agarose slice by means of the Gel Extraction kit (Qiagen) according to the supplier's recommendations. Fifty (50) ng of the purified product was then self-ligated by use of T4 ligase (Boehringer) according to the supplier's conditions. After 16 h incubation at 16°C, the DNA in the ligation mix was precipitated with ethanol and dissolved in 20 μL H_2O (ligation sample).

E.coli DH5αF' cells were transformed with 10 μL of the ligation sample. Several ampicillin-resistant clones were further characterized via restriction analysis of the isolated plasmid DNA. A positive clone was denominated as pUC18-CL-E2-H6 and was used for further modifications as described below.

Generation of shuttle vector pFPMT-CL-E2-H6

A 0.966 kb *Eco*RI/*Bam*HI fragment was isolated from pUC18-CL-E2-H6 (harboring CL-E2(384 – 673)-VIEGR-H6) and was ligated into the *Eco*RI/*Bam*HI-digested pFPMT121 (SEQ ID NO:12, Figure 3). For the reaction, T4 ligase (Boehringer) was used according to the supplier's conditions. The ligation sample was precipitated with ethanol and dissolved in 10 µL water. This was used to transform *E.coli* DH5αF' cells, a positive clone was withheld after restriction analysis and the respective plasmid is denominated pFPMT-CL-E2-H6 (SEQ ID NO:32, Figure 14).

EXAMPLE 5

CONSTRUCTION OF pFPMT-CL-K-H6-E1 SHUTTLE VECTOR

The construction of the shuttle vector was comprised of two steps.

In a first step the pUC18-FMD-CL-H6-K-E1-H6 construct was constructed by site-directed mutagenesis. The pUC18-FMD-CL-E1-H6 was used as template (SEQ ID NO:20; Figure 7). The following primers were used:

-Primer H6K hin neu: 5'-catcacaaatatgaggtgcgcaacgtgtccgggatgtac-3' (SEQ ID NO:37).

-Primer H6KRK her neu:

5'-gtgatggtggtgtcctagtgctgctagtggtaggaagcatag-3' (SEQ ID NO:38).

(The bases providing additional codons are underlined.)

The PCR reaction mixture was constituted as follows: pUC18-FMD-CL-E1-H6 (2 ng/µL), 1µL; primer H6K hin neu (100 µM), 2µL; primer H6KRK her neu (100 µM), 2µL; dNTP's (2.5 mM each), 8µL; H_2O, 76 µL; Expand Long Template PCR System (Boeringer; Cat No 1681 834) Buffer 2 (10 x concentrated), 10 µL; Expand Long Template PCR System Polymerase mixture (1 U/µL), 0.75 µL.

The PCR program used consisted of the following steps:

- denaturation step: 15 min at 95°C
- 35 cycles of 30 sec denaturation at 95°C, 1 min annealing at 60°C, and 5 min elongation at 72°C

- termination at 4°C.

An aliquot of the PCR sample was analyzed on a 1% agarose gel to check its size, which was correct (~4.2 kb).

Thereafter the 3'-A overhangs from the PCR product were removed by a T4 polymerase reaction resulting in blunt ends with 3'- and 5'-OH groups. Therefore, to the remaining 95 µL of the PCR reaction were added 1 µL T4 polymerase (Boehringer; 1 U/µL) and 4 µL dNTP's (2.5mM each). The sample was incubated for 20 min at 37°C. Subsequently, the DNA in the sample was precipitated with ethanol and dissolved in 16 µL H_2O.

Subsequently 5'-phosphates were added to the blunt-ended PCR product by a kinase reaction. Therefore, to the 16 µL dissolved blunt-ended PCR product were added 1 µL T4 polynucleotide kinase (Boehringer; 1 U/µL), 2 µL 10-fold concentrated T4 polynucleotide kinase reaction buffer (Boehringer), and 1 µL ATP (10 mM). The sample was incubated for 30 min at 37°C.

Subsequently the sample was applied onto a 1% agarose gel and the correct product band was isolated, by means of the gel extraction kit (Qiagen) according to the supplier's conditions. Fifty (50) ng of the purified product has then been self-ligated by use of T4 ligase (Boehringer) according to the supplier's recommendations. After 72 h incubation at 16°C the DNA in the ligation sample was precipitated with ethanol and dissolved in 10 µL water.

E.coli DH5αF' cells were transformed with 5 µL of the ligation sample. The plasmid DNA of several ampicillin-resitant colonies was analyzed by restriction enzyme digestion, a positive clone was withheld and the corresponding plasmid denominated: pUC18-FMD-CL-H6-E1-K-H6 (SEQ ID NO:39, Figure 17).

In a second step the transfer vector was constructed by a two-fragment ligation. In the following construction fragments with *Bcl*I cohesive ends were involved. Since *Bcl*I can cleave its site only on unmethylated DNA, an *E. coli dam*⁻ strain was transformed with the involved plasmids pUC18-FMD-CL-H6-K-E1-H6 (SEQ ID NO:39, Figure 17) and pFPMT-CL-E1 (SEQ ID NO:36, Figure 16). From each transformation, an ampicillin-resistant colony was picked, grown in a liquid culture and the unmethylated plasmid DNAs were prepared for the further use. The 1.273kb *Bcl*I/*Hin*dIII fragment of the unmethylated plasmid pUC18-FMD-CL-H6-K-E1-H6 (harbouring the *FMD* promoter, the codons of the CL-H6-K unit, and the start of E1) and the 6.057kb *Bcl*I/*Hin*dIII fragment of plasmid pFPMT-CL-E1 (harbouring the missing part of the E1 reading frame starting from the *Bcl*I site, without C-terminal His tag, as well as the pFPMT121-located elements except for the *FMD* promoter) were prepared

and ligated together for 72 h at 16°C by use of T4 ligase (Boehringer) in a total volume of 20 µL according to the supplier's specifications. Subsequently, the ligation mixture was placed on a piece of nitrocellulose membrane floating on sterile deionized water in order to desalt the ligation mixture (incubation for 30 min at room temperature). *E. coli* TOP10 cells were transformed by electroporation with 5 µL of the desalted sample. The plasmid DNA of several resulting ampicillin-resistant colonies was analyzed by restriction enzyme digestion. A positive clone was withheld and denominated pFPMT-CL-H6-K-E1 (SEQ ID NO:40, Figure 18).

EXAMPLE 6

TRANSFORMATION OF *HANSENULA POLYMORPHA* AND SELECTION OF TRANSFORMANTS

H.polymorpha strain RB11 was been transformed (PEG-mediated DNA uptake protocol essentially as described by (Klebe, R. J. et al. 1983) with the modification of (Roggenkamp, R. et al. 1986) with the different parental shuttle vectors as described in Examples 1 to 5. For each transformation, 72 uracil-prototrophic colonies were selected and used for strain generation by the following procedure. For each colony, a 2 mL liquid culture was inoculated and grown in test tubes for 48h (37°C; 160 rpm; angle 45°) in selective medium (YNB/glucose, Difco). This step is defined as the first passaging step. A 150 µL aliquot of the cultures of the first passaging step were used to inoculate 2 mL fresh YNB/glucose medium. Again, the cultures have been incubated as described above (second passaging step). Together, eight of such passaging steps were carried out. Aliquots of the cultures after the third and the eighth passaging steps were used to inoculate 2 mL of non-selective YPD medium (Difco). After 48 h of incubation at 37°C (160 rpm; angle 45°; the so-called first stabilization step), 150 µL aliquots of these YPD cultures have been used to inoculate fresh 2 mL YPD cultures which were incubated as described above (second stabilization step). Aliquots of the cultures of the second stabilization step were then streaked on plates containing selective YNB/agar. These plates were incubated for four days until macroscopic colonies became visible. A well-defined single colony of each separation was defined as strain and used for further expression analysis.

Expression analysis was performed on small-scale shake flask cultures. A colony was picked from the above mentioned YNB/agar plate and inoculated in 2 mL YPD and incubated for 48 h as mentioned above. This 2 mL-aliquot was used as seed culture for 20 mL shake flask culture. YPGlycerol (1%) was used as medium and the shake flask was incubated on a rotary shaker (200 rpm, 37°C). After 48 h of growth 1 % MeOH was added to the culture for induction of the expression cassette. At different time intervals cell pellets of 1 mL aliquots were collected and stored at –20°C until further analysis. Specific protein expression was analyzed by SDS-PAGE/ Western blotting. Therefore cell pellets were solubilized in sample-buffer (TrisHCl – SDS) and incubated for > 15 minutes at 95°C. Proteins were separated on a 15% polyacryl-amide gel and blotted (wet-blot; bicarbonate buffer) onto nitrocellulose membranes. Blots were developed using a specific murine anti-E1 (IGH 201) or murine anti-E2 (IGH 216, described by Maertens et al. in WO96/04385) as first antibody, Rabbit-Anti-Mouse- AP was used as second antibody. Staining was performed with NBT-BCIP.
Positive strains were withheld for further investigation.
Five of these positive clones were used in a shake flask expression experiment. A colony of the respective strain was picked from YNB plate and used to inoculate 2 mL YPD. These cultures were incubated as described above. This cell suspension was used to inoculate a second seed culture of 100 mL YPD medium in a 500 mL shake flask. This shake flask was incubated on a rotary shaker for 48 h at 37°C and 200 rpm. A 25 mL aliquot of this seed culture was used to inoculate 250 mL YPGlycerol (1%) medium and was incubated in a baffled 2-l shake flask under the above described conditions. 48 h after inoculation 1 % MeOH (promotor induction) was added and the shake flasks were further incubated under the above described conditions. 24 h post induction, the experiment was stopped and cell pellets collected by centrifugation. The expression level of the five different clones was analyzed by SDS-PAGE / Western blotting (conditions as above). A titration series of each clone was loaded onto the gel and the most productive strain was selected for further fermentation and purification trials.
Surprisingly, *H. polymorpha*, a yeast strain closely related to *Pichia pastoris* (Gellissen, G. 2000), is able to express HCV proteins essentially without hyperglycosylation and thus with sugar moieties comparable in size to the HCV envelope proteins expressed by HCV-recombinant vaccinia virus-infected mammalian cells.

The *Hansenula polymorpha* strain RB11 was deposited on April 19, 2002 under the conditions of the Budapest Treaty at the Mycothèque de l'UCL (MUCL), Université

Catholique de Louvain, Laboratoire de mycologie, Place Croix du Sud 3 bte 6, B-1348 Louvain-la-Neuve, Belgium and has the MUCL accession number MUCL43805.

EXAMPLE 7

CONSTRUCTION OF pSY1aMFE1sH6a VECTOR

The *S. cerevisiae* expression plasmid was constructed as follows. An E1-coding sequence was isolated as a *NsI*1/*Eco*52I fragment from pGEMT-E1sH6 (SEQ ID NO:6, Figure 1) which was made blunt-ended (using T4 DNA polymerase) and cloned in the pYIG5 vector (SEQ ID NO:41, Figure 19) using T4 DNA ligase (Boehringer) according to the supplier's specifications. The cloning was such that the E1s-H6 encoding fragment was joined directly and in frame to the αMF-coding sequence. The ligation mixture was transformed in *E.coli* DH5αF'cells. Subsequently, the plasmid DNA of several ampicilin resistant clones was analyzed by restriction digestion and a positive clone was withheld and denominated as pYIG5E1H6 (ICCG3470; SEQ ID NO:42, Figure 20).

The expression cassette (containing the αMF-sequence and the E1s-coding region with a His-tag) was transferred as a *BamH*I fragment (2790 bp) of pYIG5E1H6 into the *BamH*I-digested *E .coli/S. cerevisiae* pSY1 shuttle vector (SEQ ID NO:21, Figure 43). The ligation was performed with T4 DNA ligase (Boehringer) according to supplier's conditions. The ligation mix was transformed to *E .coli* DH5αF' cells, and the plasmid DNA of several ampicilin resistant colonies was analyzed by restriction enzyme digestion. A positive clone was withheld and denominated pSY1aMFE1sH6a (ICCG3479; SEQ ID NO:44, Figure 22).

EXAMPLE 8

CONSTRUCTION OF pSYYIGSE2H6 VECTOR

The *S. cerevisiae* expression plasmid pSYYIGSE2H6 was constructed as follows. An E2 coding sequence was isolated as a *SalI/Kpn*I fragment from pBSK-E2sH6 (SEQ ID NO:45, Figure 23) which was made blunt-ended (using T4 DNA polymerase) and subsequently

cloned in the pYIG5 vector (SEQ ID NO:41, Figure 19) using T4 DNA ligase (Boehringer) according to the supplier's specifications. The cloning was such that the E2-H6 encoding fragment was joined directly and in frame to the αMF-coding sequence. The ligation mixture was then transformed to *E. coli* DH5αF' cells, the plasmid DNA of several ampicilin resistant clones was analyzed by restriction digestion and a positive clone withheld and denominated as pYIG5HCCL-22aH6 (ICCG2424; SEQ ID NO:46, Figure 24).

The expression cassette (containing the αMF-sequence and the E2 (384 – 673) coding region with a His-tag) was transferred as a *BamHI* fragment (3281 bp) of pYIG5HCCL-22aH6 into the *BamHI* opened *E. coli/S. cerevisiae* pSY1 shuttle vector (SEQ ID NO:43, Figure 21). The ligation was performed with T4 DNA ligase (Boehringer) according to supplier's conditions. The ligation mix was transformed to *E. coli* DH5αF' cells and the plasmid DNA of several ampicilin resistant colonies was analyzed by restriction enzyme digestion. A restriction positive clone was withheld and denominated pSYYIGSE2H6 (ICCG2466; SEQ ID NO:47, Figure 25).

EXAMPLE 9

CONSTRUCTION OF pSY1YIG7E1s VECTOR

The *S. cerevisiae* expression plasmid pSY1YIG7E1s was constructed as follows. An E1 coding sequence was isolated as a *NsiI/Eco*52I fragment from pGEMT-E1s (SEQ ID NO:6, Figure 1) which was made blunt-ended and cloned into the pYIG7 vector (SEQ ID NO:48, Figure 26) using T4 DNA ligase (Boehringer) according to the supplier's specifications. The cloning was such that the E1-encoding fragment was joined directly and in frame to the αMF-coding sequence. The ligation mixture was transformed to *E. coli* DH5αF' cells, the plasmid DNA of several ampicilin resistant clones analyzed by restriction digestion and a positive clone withheld and denominated as pYIG7E1 (SEQ ID NO:49, Figure 27).

The expression cassette (containing the CL leader sequence and the E1 (192-326) coding region) was transferred as a *BamHI* fragment (2790 bp) of pYIG7E1 into the *BamHI*-digested *E. coli/S. cerevisiae* pSY1 shuttle vector (SEQ ID NO:43, Figure 21). The ligation was performed with T4 DNA ligase (Boehringer) according to supplier's conditions. The ligation mix was transformed to *E. coli* DH5αF' cells and the plasmid DNA of several ampicilin

resistant colonies was analyzed by restriction enzyme digestion. A positive clone was withheld and denominated pSY1YIG7E1s (SEQ ID NO:50, Figure 28).

EXAMPLE 10

TRANSFORMATION OF *SACCHAROMYCES CEREVISIAE* AND SELECTION OF TRANSFORMANTS

In order to overcome hyper-glycosylation problems, often reported for proteins over-expressed in *Saccharomyces cerevisiae,* a mutant screening was set-up. This screening was based on the method of Ballou (Ballou, L. et al. 1991), whereby spontaneous recessive orthovanadate-resistant mutants were selected. Initial strain selection was performed based on the glycosylation pattern of invertase, as observed after native gel electrophoresis. A strain, reduced in glycosylation capabilities, was withheld for further recombinant protein expression experiments and denominated strain IYCC155. The nature of mutation has not been further studied.

Said glycosylation-deficient strain IYCC155 was transformed with the plasmids as described in Examples 7 to 9 essentially by to the lithium acetate method as described by Elble (Elble, R. 1992). Several Ura complemented strains were picked from a selective YNB + 2 % agar plate (Difco) and used to inoculate 2ml YNB+2%glucose. These cultures were incubated for 72 h, 37°C, 200 rpm on orbital shaker, and the culture supernatant and intracellular fractions were analysed for expression of E1 by western blot developed with a E1 specific murine monoclonal antibody (IGH 201). A high producing clone was withheld for further experiments.

The expression of proteins in the *S. cerivisiae* glycosylation deficient mutant used here is hampered by the suboptimal growth characteristics of such strains which leads to a lower biomass yield and thus a lower yield of the desired proteins compared to wild-type *S. cerivisiae* strains. The yield of the desired proteins was still substantially higher than in mammalian cells.

EXAMPLE 11

CONSTRUCTION OF pPICZalphaD'E1sH6 AND pPICZalphaE'E1sH6 VECTORS

The shuttle vector pPICZalphaE'E1sH6 was constructed starting from the pPICZalphaA vector (Invitrogen; SEQ ID NO:51, Figure 29). In a first step said vector was adapted in order to enable cloning of the E1 coding sequence directly behind the cleavage site of the KEX2 or STE13 processing proteases, respectively. Therefore pPICZalphaA was digested with *Xho*I and *NotI*. The digest was separated on a 1% agarose gel and the 3519 kb fragment (major part of vector) was isolated and purified by means of a gel extraction kit (Qiagen). This fragment was then ligated using T4 polymerase (Boehringer) according to the supplier's conditions in presence of specific oligonucleotides yielding pPICZalphaD' (SEQ ID NO:52, Figure 30) or pPICZalphaE' (SEQ ID NO:53, Figure 31).

The following oligonucleotides were used:

- for constructing pPICZalphaD':

8822: 5'-TCGAGAAAAGGGGCCCGAATTCGCATGC-3' (SEQ ID NO:54); and

8823: 5'-GGCCGCATGCGAATTCGGGCCCCTTTTC-3' (SEQ ID NO:55)

which yield, after annealing, the linker oligonucleotide:

```
TCGAGAAAAGGGGCCCGAATTCGCATGC      (SEQ ID NO:54)
    CTTTTCCCCGGGCTTAAGCGTACGCCGG  (SEQ ID NO:55)
```

- for constructing pPICZalphaE'

8649: 5'-TCGAGAAAAGAGAGGCTGAAGCCTGCAGCATATGC-3' (SEQ ID NO: 56)

8650: 5'-GGCCGCATATGCTGCAGGCTTCAGCCTCTCTTTTC-3' (SEQ ID NO:57)

which yield, after annealing, the linker oligonucleotide:

```
TCGAGAAAAGAGAGGCTGAAGCCTGCAGCATATGC       (SEQ ID NO:56)
    CTTTTCTCTCCGACTTCGGACGTCGTATACGCCGG    (SEQ ID NO:57)
```

These shuttle vectors pPICZalphaD' and pPICZalphaE' have newly introduced cloning sites directly behind the cleavage site of the respective processing proteases, KEX2 and STE13.

The E1-H6 coding sequence was isolated as a *NsI*1/*Eco*52I fragment from pGEMT-E1sH6 (SEQ ID NO:6, Figure 1). The fragment was purified using a gel extraction kit (Qiagen) after separation of the digest on a 1% agarose gel. The resulting fragment was made blunt-ended

(using T4 DNA polymerase) and ligated into either pPICZalphaD' or pPICZalphaE' directly behind the respective processing protease cleavage site.

The ligation mixtures were transformed to *E. coli* TOP10F' cells and plasmid DNA of several zeocin resistant colonies analyzed by restriction enzyme digestion. Positive clones were withheld and denominated pPICZalphaD'E1sH6 (ICCG3694; SEQ ID NO:58, Figure 32) and pPICZalphaE'E1sH6 (ICCG3475; SEQ ID NO:59, Figure 33), respectively.

EXAMPLE 12

CONSTRUCTION OF pPICZalphaD'E2sH6 AND pPICZalphaE'E2sH6 VECTORS

The shuttle vectors pPICZalphaD' and pPICZalphaE' were constructed as described in Example 11.

The E2-H6 coding sequence was isolated as a *SalI/Kpn*I fragment from pBSK-E2sH6 (SEQ ID NO:45, Figure 23). The fragment was purified with a gel extraction kit (Qiagen) after separation of the digest on a 1% agarose gel. The resulting fragment was made blunt-ended (using T4 DNA polymerase) and ligated into either pPICZalphaD' or pPICZalphaE' directly behind the respective processing protease cleavage site.

The ligation mixture was transformed to *E. coli* TOP10F' cellls and the plasmid DNA of several zeocin resistant colonies was analyzed by restriction enzyme digestion. Positive clone were withheld and denominated pPICZalphaD'E2sH6 (ICCG3692; SEQ ID NO:60, Figure 34) and pPICZalphaE'E2sH6 (ICGG3476; SEQ ID NO:61, Figure 35), respectively.

EXAMPLE 13

TRANSFORMATION OF *PICHIA PASTORIS* AND SELECTION OF TRANSFORMANTS

The *P. pastoris* shuttle plasmids as described in Examples 11 and 12 were transformed to *P. pastoris* cells according to the supplier's conditions (Invitrogen). An E1- and an E2-producing strain were withheld for further characterization.

The HCV envelope proteins were expressed in *P. pastoris*, a yeast strain well known for the fact that hyperglycosylation is normally absent (Gellissen, G. 2000) and previously used to express dengue virus E protein as GST fusion (Sugrue, R. J. et al. 1997). Remarkably, the resulting *P. pastoris*-expressed HCV envelope proteins displayed a comparable glycosylation as is observed in wild-type *Saccharomyces* strains. More specifically, the HCV envelope proteins produced by *P. pastoris* are hyperglycosylated (based on the molecular weight of the expression products detected in western-blots of proteins isolated from transformed *P. pastoris* cells).

EXAMPLE 14

CULTURE CONDITIONS FOR *SACCHAROMYCES CEREVISIAE*, *HANSENULA POLYMORPHA* AND *PICHIA PASTORIS*

Saccharomyces cerevisiae

Cell banking

Of the selected recombinant clone a master cell bank and working cell bank were prepared. Cryo-vials were prepared from a mid-exponentially grown shake flask culture (incubation conditions as for fermentation seed cultures, see below). Glycerol was added (50 % final conc.) as a cryoprotectant.

Fermentation

Seed cultures were started from a cryo-preserved working cell bank vial and grown in 500 mL medium (YNB supplemented with 2 % sucrose, Difco) in a 2 L Erlenmeyer shake flasks at 37°C, 200 rpm for 48h.

Fermentations were typically performed in Biostat C fermentors with a working volume of 15 L (B.Braun Int., Melsungen, Germany). The fermentation medium contained 1% Yeast Extract, 2% Peptone and 2 % sucrose as carbon source. Poly-ethylene glycol was used as anti-foam agent.

Temperature, pH and dissolved oxygen were typically controlled during the fermentation, applicable set-points are summarised in Table 1. Dissolved oxygen was cascade controlled by

agitation/aeration. pH was controlled by addition of NaOH (0.5 M) or H_3PO_4 solution (8.5 %).

Table 1. Typical parameter settings for *S. cerevisiae* fermentations

Parameter	**set-point**
Temperature	33 – 37 °C
pH	4.2 – 5.0
DO (growth phase)	10 – 40 % air saturation
DO (induction)	0 – 5 %
aeration	0.5 – 1.8 vvm*
agitation	150 – 900 rpm

* volume replacement per minute

The fermentation was started by the addition of 10 % seed-culture. During the growth phase the sucrose concentration was monitored off-line by HPLC analysis (Polysphere Column OAKC Merck).

During the growth phase the dissolved oxygen was controlled by cascade control (agitation/aeration). After complete metabolisation of sucrose the heterologous protein production was driven by the endogenous produced ethanol supplemented with stepwise addition of EtOH in order to maintain the concentration at approximately 0.5 % (off-line HPLC analysis, polyspher OAKC column) During this induction phase the dissolved oxygen was controlled below 5% air-saturation, by manual adjustment of airflow rate and agitator speed.

Typically the fermentation was harvested 48 to 72 h post induction by concentration via tangential flow filtration followed by centrifugation of the concentrated cell suspension to obtain cell pellets. If not analyzed immediately, cell pellets were stored at –70°C.

Hansenula polymorpha

Cell banking

Of the selected recombinant clone a master cell bank and working cell bank were prepared.

Cryo-vials were prepared from a mid-exponentially grown shake flask culture (incubation conditions as for fermentation seed cultures, see below). Glycerol was added (50 % final conc.) as a cryoprotectant.

Fermentation

Seed cultures were started from a cryo-preserved (-70°C) working cell bank vial and grown in 500 mL medium (YPD, Difco) in a 2 L Erlenmeyer shake flasks at 37°C, 200 rpm for 48h. Fermentations were typically performed in Biostat C fermentors with a working volume of 15 L (B.Braun Int., Melsungen, Germany). The fermentation medium contained 1% Yeast Extract, 2% Peptone and 1% glycerol as carbon source. Poly-ethylene glycol was used as anti-foam agent.

Temperature, pH, air-in and dissolved oxygen were typically controlled during the fermentation, applicable set-points are summarised in Table 2. Dissolved oxygen was controlled by agitation. pH was controlled by addition of NaOH (0.5 M) or H_3PO_4 solution (8.5 %).

Table 2. Typical parameter settings for *H. polymorpha* fermentations

Parameter	**set-point**
Temperature	30 – 40 °C
pH	4.2 – 5.0
DO	10 – 40 % air saturation
aeration	0.5 – 1.8 vvm*
agitation	150 – 900 rpm

* volume replacement per minute

The fermentation was started by the addition of 10 % seed-culture. During the growth phase the glycerol concentration was monitored off-line (Polysphere Column OAKC Merck) and 24 h after complete glycerol consumption 1% methanol was added in order to induce the heterologous protein expression. The fermentation was harvested 24 h post induction by concentration via tangential flow filtration followed by centrifugation of the concentrated cell suspension to obtain cell pellets. If not analyzed immediately, cell pellets were stored at – 70°C.

Pichia pastoris

Small scale protein production experiments with recombinant *Pichia pastoris* were set up in shake flask cultures. Seed cultures were grown overnight in YPD medium (Difco). Initial medium pH was corrected to 4.5. Shake flasks were incubated on a rotary shaker at 200 – 250 rpm, 37°C.

The small scale production was typically performed at 500 mL scale in 2 L shake flasks and were started with a 10 % inoculation in expression medium, containing 1% Yeast extract, 2 % Peptone (both Difco), and 2 % glycerol as carbon source. Incubation conditions were as for the seed culture. Induction was started by addition of 1 % MeOH approximately 72 h after inoculation. The cells were collected 24 h post induction by centrifugation. If not analyzed immediately, cell pellets were stored at –70°C.

EXAMPLE 15

LEADER PEPTIDE REMOVAL FROM MFα-E1-H6 AND MFα-E2-H6 PROTEINS EXPRESSED IN SELECTED YEAST CELLS

The expression products in *Hansenula polymorpha* and a *Saccharomyces cerevisiae* glycosylation minus strain of the HCV E1 and E2 protein constructs with the α-mating factor (αMF) leader sequence of *S. cerevisiae* were further analyzed. Since both genotype 1b HCV E1s (aa 192-326) and HCV E2s (aa 383-673 extended by the VIEGR (SEQ ID NO:69)-sequence) were expressed as C-terminal his-tagged (H6, HHHHHH, SEQ ID NO:63; said HCV proteins are furtheron in this Example denoted as αMF-E1-H6 and αMF-E2-H6) proteins, a rapid and efficient purification of the expressed products after guanidinium chloride (GuHCl)-solubilization of the yeast cells was performed on Ni-IDA (Ni-iminodiacetic acid). In brief, cell pellets were resuspended in 50 mM phosphate, 6M GuHCl, pH 7.4 (9 vol/g cells). Proteins were sulfonated overnight at room temperature (RT) in the presence of 320 mM (4% w/v) sodium sulfite and 65 mM (2% w/v) sodium tetrathionate. The lysate was cleared after a freeze-thaw cycle by centrifugation (10.000 *g*, 30 min, 4°C) and Empigen (Albright &Wilson, UK) and imidazole were added to the supernatant to final concentrations of 1% (w/v) and 20 mM, respectively. The sample was filtrated (0.22 μM) and loaded on a Ni-IDA Sepharose FF column, which was equilibrated with 50 mM phosphate,

6M GuHCl, 1% Empigen (buffer A) supplemented with 20 mM imidazole. The column was washed sequentially with buffer A containing 20 mM and 50 mM imidazole, respectively, till absorbance at 280 nm reached baseline level. The his-tagged products were eluted by applying buffer D, 50 mM phosphate, 6M GuHCl, 0.2 % (for E1) or 1 % (for E2) Empigen, 200 mM imidazole. The eluted materials were analyzed by SDS-PAGE and western-blot using a specific monoclonal antibodies directed against E1 (IGH201), or E2 (IGH212).

The E1-products were immediately analyzed by Edman degradation.

Since at this stage, SDS-PAGE revealed already a very complex picture of protein bands for HCV E2, a further fractionation by size exclusion chromatography was performed. The Ni-IDA eluate was concentrated by ultrafiltration (MWCO 10 kDa, centriplus, Amicon, Millipore) and loaded on Superdex G200 (10/30 or 16/60; Pharmacia) in PBS, 1% Empigen or PBS, 3% Empigen. Elution fractions, containing E2 products, with a Mr between ~80 kDa and ~45 kDa, i.e. fractions 17-23 of the elution profile in Figure 37 based on the migration on SDS-PAGE (Figure 38), were pooled and alkylated (incubation with 10 mM DTT 3h at RT followed by incubation with 30 mM iodo-acetamide for 3 hours at RT). Samples for amino-terminal sequencing were treated with Endo H (Roche Biochemicals) or left untreated. The glycosylated and deglycosylated E2 products were blotted on PVDF-membranes for amino-terminal sequencing. An amido-black stained blot of glycosylated and deglycosylated E2 is shown in Figure 39.

The sequencing of both E1 and E2 purified products lead to the disappointing observation that removal of the signal sequence from the HCV envelope proteins is occurring only partially (see Table 3). In addition, the majority of the side products (degradation products and products still containing the leader sequence or part thereof) are glycosylated. This glycosylation resides even in part on the non-cleaved fragment of the signal sequence which contains also an N-glycosylation site. These sites can be mutated in order to result in less glycosylated side products. However, even more problematic is the finding that some alternatively cleaved products have only 1 to 4 amino acids difference compared to the desired intact envelope protein. Consequently, purification of the correctly processed product is virtually impossible due to the lack of sufficiently discriminating biochemical characteristics between the different expression products. Several of the degradation products may be a result of a Kex-2 like cleavage (e.g. the cleavage observed after aa 196 of E1 which is a cleavage after an arginine), which is also required for the cleavage of the α-mating factor leader and which can thus not be blocked without disturbing this essential process.

A high E1 producing clone derived from transformation of *S. cerevisiae* IYCC155 with pSY1YIG7E1s (SEQ ID NO:50; Figure 28) was compared with a high producing clone derived from transformation of *S. cerevisiae* IYCC155 with pSY1aMFE1sH6aYIG1E1s (SEQ ID NO:44; Figure 22). The intracellular expression of the E1 protein was evaluated after 2 up to 7 days after induction, and this by means of Western-blot using the E1 specific monoclonal antibody (IGH 201). As can be judged from Figure 40, maximal expression was observed after 2 days for both strains but the expression patterns for both strains are completely different. Expression with the α-mating factor leader results in a very complex pattern of bands, which is a consequence from the fact that the processing of the leader is not efficient. This leads to several expression products with a different amino-terminus and of which some are modified by 1 to 5 N-glycosylations. However, for the E1 expressed with the CL leader a limited number of distinct bands is visible which reflects the high level of correct CL leader removal and the fact that only this correctly processed material may be modified by N-glycosylation (1 to 5 chains), as observed for *Hansenula*-derived E1 expressed with the same CL leader (see Example 16).

The hybridoma cell line producting the monoclonal antibody directed against E1 (IGH201) was deposited on March 12, 1998 under the conditions of the Budapest Treaty at the European Collection of Cell Cultures, Centre for Applied Microbiology & Research, Salisbury, Wiltshire SP4 0JG, UK, and has the accession number ECACC 98031216). The monoclonal antibody directed against E2 (IGH212) has been described as antibody 12D11F2 in Example 7.4 by Maertens et al. in WO96/04385.

Table 3. Identification of N-termini of αMF-E1-H6 and αMF-E2-H6 proteins expressed in *S. cerevisiae* or *H. polymorpha*. Based on the N-terminal sequencing the amount of N-termini of the mature E1-H6 and E2-H6 proteins could be estimated ("mature" indicating correct removal of the αMF signal sequence). The total amount of protein products was calculated as pmol of protein based on the intensity of the peaks recovered by Edman degradation. Subsequently, for each specific protein (i.e. for each 'detected N-terminus') the mol % versus the total was estimated.

Yeast	αMF-E1-H6	αMF-E2-VIEGR-H6
S. cerevisiae	Experiment 1: - 16% of proteins still containing αMF sequences - 18% of proteins cleaved between aa 195 and 196 of E1 - 66% of proteins with correctly removed αMF Experiment 2 - 18% of proteins still containing αMF sequences - 33% of proteins cleaved between aa 195 and 196 of E1 - 8% of other proteins other E1 cleavage products - 44% of proteins with correctly removed αMF	/
H. polymorpha	- 64% of proteins still containing αMF sequences - 6% of proteins cleaved between aa 192 and 193 of E1 - 30% of proteins with correctly removed αMF	- 75% of proteins still containing αMF sequences - 25% of proteins with correctly removed αMF

EXAMPLE 16

EXPRESSION OF AN E1 CONSTRUCT IN YEAST SUITABLE FOR LARGE SCALE PRODUCTION AND PURIFICATION

Several other leader sequences were used to replace the *S. cerevisiae* αMF leader peptide including CHH (leader sequence of *Carcinus maenas* hyperglycemic hormone), Amy1 (leader

sequence of amylase from *S. occidentalis*), Gam1 (leader sequence of glucoamylase from *S. occidentalis*), Phy5 (leader sequence from fungal phytase), pho1 (leader sequence from acid phosphatase from *Pichia pastoris*) and CL (leader of avian lysozyme C, 1,4-beta-N-acetylmuramidase C) and linked to E1-H6 (i.e. E1 with C-terminal his-tag). All constructs were expressed in *Hansenula polymorpha* and each of the resulting cell lysates was subjected to western blot analysis. This allowed already to conclude that the extent of removal of the leader or signal sequence or peptide was extremely low, except for the construct wherein CL is used as leader peptide. This was confirmed for the CHH-E1-H6 construct by Edman-degradation of Ni-IDA purified material: no correctly cleaved product could be detected although several different sequences were recovered (see Table 4).

Table 4. Identification of N-termini of CHH-E1-H6 proteins expressed in *H. polymorpha*, based on N-terminal amino acid sequencing of different protein bands after separation by SDS-PAGE and blotting to a PVDF membrane.

Molecular size	Identified N-termini
45 kD	starts at amino acid 27 of CHH leader = only pre-sequence cleaved, pro-sequence still attached
26 kD	- partially starts at amino acid 1 of CHH leader = no removal of pre-pro-sequence - partially starts at amino acid 9 of CHH leader = product of alternative translation starting at second AUG codon
24 kD	- partially starts at amino acid 1 of CHH leader = no removal of pre-pro-sequence - partially starts at amino acid 9 of CHH leader = product of alternative translation starting at second AUG codon

As mentioned already, the western-blots of the cell lysates revealed a pattern of E1 specific protein bands, indicative for a higher degree of correct removal of the CL leader peptide. This is surprising since this leader is not derived from a yeast. Amino acid sequencing by Edman degradation of GuHCl solubilized and Ni-IDA purified material indeed confirmed that 84% of the E1 proteins is correctly cleaved and the material is essentially free of degradation products. Still 16% of non-processed material is present but since this material is non-glycosylated it can be easily removed from the mixture allowing specific enrichment of correctly cleaved and glycosylated E1. Such a method for enrichment may be an affinity

chromatography on lectins, other alternatives are also given in Example 19. Alternatively, the higher hydrophobic character of the non-glycosylated material may be used to select and optimize other enrichment procedures. The correct removal of the CL leader peptide from the CL-E1-H6 protein was further confirmed by mass spectrometry which also confirmed that up to 4 out of the 5 N-glycosylation sites of genotype 1b E1s can be occupied, whereby the sequence NNSS (amino acids 233 to 236; SEQ ID NO:73) are considered to be a single N-glycosylation site.

EXAMPLE 17

PURIFICATION AND BIOCHEMICAL CHARACTERIZATION OF THE HCV E2 PROTEIN EXPRESSED IN *HANSENULA POLYMORPHA* FROM THE CL-E2-H6 ENCODING CONSTRUCT

The efficiency of removal of the CL leader peptide from CL-E2-VIEGR-H6 (furtheron in this Example denoted as "CL-E2-H6") protein expressed in *Hansenula polymorpha* was analyzed. Since the HCV E2s (aa 383-673) was expressed as a his-tagged protein, a rapid and efficient purification of the expressed protein after GuHCl-solubilization of collected cells was performed on Ni-IDA. In brief, cell pellets were resuspended in 30 mM phosphate, 6 M GuHCl, pH 7.2 (9 mL buffer/g cells). The protein was sulfonated overnight at room temperature in the presence of 320 mM (4% w/v) sodium sulfite and 65 mM (2% w/v) sodium tetrathionate. The lysate was cleared after a freeze-thaw cycle by centrifugation (10.000 *g*, 30 min, 4°C). Empigen BB (Albright & Wilson) and imidazole were added to a final concentration of 1% (w/v) and 20 mM, respectively. All further chromatographic steps were executed on an Åkta FPLC workstation (Pharmacia). The sample was filtrated through a 0.22 µm pore size membrane (cellulose acetate) and loaded on a Ni-IDA column (Chelating Sepharose FF loaded with Ni^{2+}, Pharmacia), which was equilibrated with 50 mM phosphate, 6 M GuHCl, 1 % Empigen BB, pH 7.2 (buffer A) supplemented with 20 mM imidazole. The column was washed sequentially with buffer A containing 20 mM and 50 mM imidazole, respectively, till the absorbance at 280 nm reached the baseline level. The his-tagged products were eluted by applying buffer D, 50 mM phosphate, 6 M GuHCl, 0.2 % Empigen BB (pH 7.2), 200 mM imidazole. The purified materials were analysed by SDS-PAGE and western-blot using a specific monoclonal antibody directed against E2 (IGH212) (Figure 41). The

IMAC-purified E2-H6 protein was also subjected to N-terminal sequencing by Edman degradation. Thereto proteins were treated with N-glycosidase F (Roche) (0.2 U/µg E2, 1 h incubation at 37°C in PBS/3% empigen BB) or left untreated. The glycosylated and deglycosylated E2-H6 proteins were subjected to SDS-PAGE and blotted on a PVDF-membrane for amino acid sequencing (analysis was performed on a PROCISE™ 492 protein sequencer, Applied Biosystems). Since at this stage, SDS-PAGE revealed some degradation products, a further fractionation by size exclusion chromatography was performed. Hereto, the Ni-IDA eluate was concentrated by ultrafiltration (MWCO 10 kDa, centriplus, Amicon, Millipore) and loaded on a Superdex G200 (Pharmacia) in PBS, 1% Empigen BB. Elution fractions, containing mainly intact E2s related products with a Mr between ~30 kDa and ~70 kDa based on the migration on SDS-PAGE, were pooled and eventually alkylated (incubation with 5 mM DTT for 30 minutes at 37°C, followed by incubation with 20 mM iodoacetamide for 30 minutes at 37°C). The possible presence of degradation products after IMAC purification can thus be overcome by a further fractionation of the intact product by means of size exclusion chromatography. An unexpectedly good result was obtained. Based on the N-terminal sequencing the amount of E2 product from which the CL leader peptide is removed could be estimated. The total amount of protein products is calculated as pmol of protein based on the intensity of the peaks recovered by Edman degradation. Subsequently, for each specific protein (i.e. for each 'detected N-terminus') the mol % versus the total is estimated. In the current experiment, only the correct N-terminus of E2-H6 was detected and other variants of E2-H6 lacking amino acid of the E2 protein or containing N-terminal amino acids not comprised in the E2 protein were absent. In conclusion, the E2-H6 protein expressed by *H. polymorpha* as CL-E2-H6 protein was isolated without any further *in vitro* processing as a > 95 % correctly cleaved protein. This is in sharp contrast with the fidelity of leader peptide removal by *H. polymorpha* of the αMF-E2-H6 protein to the E2-H6 protein, which was estimated to occur in 25 % of the isolated proteins (see Table 3).

EXAMPLE 18

PURIFICATION AND BIOCHEMICAL CHARACTERIZATION OF THE HCV E1 PROTEIN EXPRESSED IN *HANSENULA POLYMORPHA* FROM THE CL-H6-K-E1 ENCODING CONSTRUCT AND *IN VITRO* PROCESSING OF H6-CONTAINING PROTEINS

The efficiency of removal of the CL leader peptide from the CL-H6-K-E1 protein expressed in *H. polymorpha* was analyzed, as well as the efficiency of subsequent in vitro processing in order to remove the H6 (his-tag)-adaptor peptide and the Endo Lys-C processing site. Since the HCV E1s (aa 192-326) was expressed as a N-terminal His-K-tagged protein CL-H6-K-E1, a rapid and efficient purification could be performed as described in Example 17. The elution profile of the IMAC-chromatographic purification of H6-K-E1 (and possibly residual CL-H6-K-E1) proteins is shown in Figure 42. After SDS-PAGE and silver staining of the gel and western-blot analysis using a specific monoclonal antibody directed against E1 (IGH201) (Figure 43), the elution fractions (63-69) containing the recombinant E1s products were pooled ('IMAC pool') and subjected to an overnight Endoproteinase Lys-C (Roche) treatment (enzyme/substrate ratio of 1/50 (w/w), 37 °C) in order to remove the H6-K-fusion tail. Removal of non-processed fusion product was performed by a negative IMAC chromatography step on a Ni-IDA column whereby Endo-Lys-C-processed proteins are collected in the flow-through fraction. Hereto the Endoproteinase Lys-C digested protein sample was applied on a Ni-IDA column after a 10-fold dilution with 10 mM $NaH_2PO_4.3H_2O$, 1 % (v/v) Empigen B, pH 7.2 (buffer B) followed by washing with buffer B till the absorbance at 280 nm reached the baseline level. The flow through was collected in different fractions (1-40) that were screened for the presence of E1s-products (Figure 44). The fractions (7-28), containing intact E1 from which the N-terminal H6-K (and possibly residual CL-H6-K) tail is removed (with a Mr between ~15 kDa and ~30 kDa based on the migration on SDS-PAGE followed by silver staining or western blot analysis using a specific monoclonal antibody directed against E1 (IGH201), were pooled and eventually alkylated (incubation with 5 mM DTT for 30 minutes at 37°C, followed by incubation with 20 mM iodoacetamide for 30 minutes at 37°C).

This material was subjected to N-terminal sequencing (Edman degradation). Hereto, protein samples were treated with N-glycosidase F (Roche) (0.2U/µg E1, 1h incubation at 37 °C in PBS/3% empigen BB) or left untreated. The glycosylated and deglycosylated E1 proteins were then separated by SDS-PAGE and blotted on a PVDF-membrane for further analysis by Edman degradation (analysis was performed on a PROCISE™ 492 protein sequencer, Applied Biosystems). Based on the N-terminal sequencing the amount of correctly processed E1 product could be estimated (processing includes correct cleavage of the H6-K-sequence). The total amount of protein products is calculated as pmol of protein based on the intensity of the peaks recovered by Edman degradation. Subsequently, for each specific protein (i.e. for

each 'detected N-terminus') the mol % versus the total is estimated. In the current experiment, only the correct N-terminus of E1 was detected and not the N-termini of other processing variants of H6-K-E1. Based thereon, *in vitro* processing by Endo Lys-C of the H6-K-E1 E1 (and possibly residual CL-H6-K-E1) protein to the E1 protein was estimated to occur with a fidelity of more than 95 %.

EXAMPLE 19

SPECIFIC REMOVAL OF LOW-GLYCOSYLATED FORMS OF HCV E1 BY HEPARIN

In order to find specific purification steps for HCV envelope proteins from yeast cells binding with heparin was evaluated. Heparin is known to bind to several viruses and consequently binding to the HCV envelope has already been suggested (Garson, J. A. et al. 1999). In order to analyze this potential binding, heparin was biotinylated and interaction with HCV E1 analyzed in microtiterplates coated with either sulfonated HCV E1 from *H. polymorpha*, alkylated HCV E1 from *H. polymorpha* (both produced as described in Example 16) and alkylated HCV E1 from a culture of mammalian cells transfected with a vaccinia expression vector. Surprisingly, a strong binding could only be observed with sulfonated HCV E1 from *H. polymorpha*, while binding with HCV E1 from mammalian cell culture was completely absent. By means of western-blot we could show that this binding was specific for the lower molecular weight bands of the HCV E1 protein mixture (Figure 45), corresponding to low-glycosylated mature HCV E1s. Figure 45 also reveals that sulfonation is not essential for heparin binding since upon removal of this sulfonation binding is still observed for the low molecular weight E1 (lane 4). Alternatively, alkylation is reducing this binding substantially, however, this may be caused by the specific alkylation agent (iodo-acetamide) used in this example. This finding further demonstrated the industrial applicability of the CL-HCV-envelope expression cassettes for yeast since we specifically can enrich HCV E1 preparations towards a preparation with HCV E1 proteins with a higher degree of glycosylation (i.e. more glycosylation sites occupied).

EXAMPLE 20

FORMATION AND ANALYSIS OF VIRUS-LIKE PARTICLES (VLPs)

Conversion of the HCV E1 and E2 envelope proteins expressed in *H. polymorpha* (Examples 16 to 18) to VLPs was done essentially as described by Depla et al. in WO99/67285 and by Bosman et al. in WO01/30815. Briefly, after cultivation of the transformed *H. polymorpha* cells during which the HCV envelope proteins were expressed, cells were harvested, lysed in GuHCl and sulphonated as described in Example 17. His-tagged proteins were subsequently purified by IMAC and concentrated by ultrafiltration as described in Example 17.

VLP-formation of HCV envelope proteins with sulphonated Cys-thiol groups

The concentrated HCV envelope proteins sulphonated during the isolation procedure were not subjected to a reducing treatment and loaded on a size-exclusion chromatograpy column (Superdex G200, Pharmacia) equilibrated with PBS, 1 % (v/v) Empigen. The eluted fractions were analyzed by SDS-PAGE and western blotting. The fractions with a relative Mr ~29-~15 kD (based on SDS-PAGE migration) were pooled, concentrated and loaded on Superdex G200, equilibrated with PBS, 3% (w/v) betain, to enforce virus like particle formation (VLP). The fractions were pooled, concentrated and desalted to PBS, 0.5% (w/v) betain.

VLP-formation of HCV envelope proteins with irreversibly modified Cys-thiol groups

The concentrated HCV envelope proteins sulphonated during the isolation procedure were subjected to a reducing treatment (incubation in the presence of 5 mM DTT in PBS) to convert the sulphonated Cys-thiol groups to free Cys-thiol groups. Irreversible Cys-thiol modification was performed by (i) incubation for 30 min in the presence of 20 mM iodoacetamide, or by (ii) incubation for 30 min in the presence of 5 mM N-ethylmaleimide (NEM) and 15 mM biotin-N-ethylmaleimide. The proteins were subsequently loaded on a size-exclusion chromatograpy column (Superdex G200, Pharmacia) equilibrated with PBS, 1 % (v/v) Empigen in case of iodoacetamide-blocking, or with PBS, 0.2 % CHAPS in case of blocking with NEM and biotin-NEM. The eluted fractions were analyzed by SDS-PAGE and Western blotting. The fractions with a relative Mr ~29-~15 kD (based on SDS-PAGE migration) were pooled, concentrated and, to force virus-like particle formation, loaded on a Superdex G200 column equilibrated with PBS, 3% (w/v) betain. The fractions were pooled, concentrated and desalted to PBS, 0.5% (w/v) betain in case of iodoacetamide-blocking, or with PBS, 0.05 % CHAPS in case of blocking with NEM and biotin-NEM.

VLP-formation of HCV envelope proteins with reversibly modified Cys-thiol groups

The concentrated HCV envelope proteins sulphonated during the isolation procedure were subjected to a reducing treatment (incubation in the presence of 5 mM DTT in PBS) to convert the sulphonated Cys-thiol groups to free Cys-thiol groups. Reversible Cys-thiol modification was performed by incubation for 30 min in the presence of dithiodipyridine (DTDP), dithiocarbamate (DTC) or cysteine. The proteins were subsequently loaded on a size-exclusion chromatograpy column (Superdex G200, Pharmacia) equilibrated with PBS, 1 % (v/v) Empigen. The eluted fractions were analyzed by SDS-PAGE and Western blotting. The fractions with a relative Mr ~29-~15 kD (based on SDS-PAGE migration) were pooled, concentrated and loaded on Superdex G200, equilibrated with PBS, 3% (w/v) betain, to enforce virus like particle formation (VLP). The fractions were pooled, concentrated and desalted to PBS, 0.5% (w/v) betain.

The elution profiles of size-exclusion chromatography in PBS, 3% (w/v) betain to obtain VLPs of *H. polymorpha*-expressed E2-H6 are shown in Figure 46 (sulphonated) and Figure 47 (alkylated with iodoacetamide).

The elution profiles of size-exclusion chromatography in PBS, 3% (w/v) betain to obtain VLPs of *H. polymorpha*-expressed E1 are shown in Figure 48 (sulphonated) and Figure 49 (alkylated with iodoacetamide). The resulting VLPs were analyzed by SDS-PAGE and western blotting as shown in Figure 50.

Size-analysis of VLPs formed by *H. polymorpha*-expressed HCV envelope proteins

The VLP particle size was determined by Dynamic Light Scattering. For the light-scattering experiments, a particle-size analyzer (Model Zetasizer 1000 HS, Malvern Instruments Ltd., Malvern, Worcester UK) was used which was controlled by photon correlation spectroscopy (PCS) software. Photon correlation spectroscopy or dynamic light scattering (DLS) is an optical method that measures brownian motion and relates this to the size of particles. Light from a continuous, visible laser beam is directed through an ensemble of macromolecules or particles in suspension and moving under brownian motion. Some of the laser light is scattered by the particles and this scattered light is measured by a photomultiplier. Fluctuations in the intensity of scattered light are converted into electrical pulses which are fed into a correlator. This generates the autocorrelation function which is passed to a computer where the appropriate data analysis is performed. The laser used was a 10 mW

monochromatic coherent He-Ne laser with a fixed wavelength of 633 nm. For each sample, three to six consecutive measurements were taken.
The results of these experiments are summarized in Table 5.

Table 5. Results of dynamic light scattering analysis on the indicated VLP-compositions of HCV envelope proteins expressed by *H. polymorpha*. The VLP particle sizes are given as mean diameter of the particles.

Cys-thiol modification	E1-H6	E2-VIEGR-H6	E1
sulphonation	25-45 nm	20 nm	20-26 nm
alkylation (iodoacetamide)	23-56 nm	20-56 nm	21-25 nm

The observation that sulphonated HCV E1 derived from *H. polymorpha* still forms particles with a size in the same range as alkylated HCV E1 from *Hansenula* is surprising. Such an effect was not expected since the high (up to 8 Cys-thiol groups can be modified on HCV E1) net increase of negative charges as a consequence of sulphonation should induce an ionic repulsion between the subunits. The other reversible cysteine modifying agents tested also allowed particle formation, the HCV E1 produced in this way, however, proved to be less stable than the sulphonated material, resulting in disulfide-based aggregation of the HCV E1. In order to use these other reversible blockers, further optimization of the conditions is required.

EXAMPLE 21

ANTIGENIC EQUIVALENCE OF *HANSENULA*-PRODUCED HCV E1-H6 AND HCV E1 PRODUCED BY VACCINIA-INFECTED MAMMALIAN CELLS

The reactivity of *Hansenula*-produced HCV E1-H6 with sera from HCV chronic carriers was compared to the reactivity of HCV E1 produced by HCV-recombinant vaccinia virus-infected mammalian cells as described by Depla et al. in WO 99/67285. Both HCV-E1 preparations tested consisted of VLP's wherein the HCV E1 proteins were alkylated with NEM and biotin-NEM. The reactivities of both HCV E1 VLP-preparations with sera from HCV chronic

carriers was determined by ELISA. The results are summarized in Table 6. As can be derived from Table 6, no differences in reactivity were noted between HCV E1 expressed in HCV-recombinant vaccinia virus-infected mammalian cells and HCV E1 expressed in *H. polymorpha*.

Table 6. Antigenicity of E1 produced in a mammalian cell culture or produced in *H. polymorpha* were evaluated on a panel of sera from human HCV chronic carriers. For this purpose biotinylated E1 was bound to streptavidin coated ELISA plates. Thereafter human sera were added at a 1/20 dilution and bound immunoglobulins from the sera bound to E1 were detected with a rabbit-anti-human IgG-Fc specific secondary antibody labeled with peroxidase. Results are expressed as OD-values. The average values are the averages of the OD-values of all serum samples tested.

Serum	*Hansenula*	mammalian	Serum	*Hansenula*	mammalian
17766	1.218	1.159	55337	1.591	1.416
17767	1.513	1.363	55348	1.392	1.261
17777	0.806	0.626	55340	1.202	0.959
17784	1.592	1.527	55342	1.599	1.477
17785	1.508	1.439	55345	1.266	1.428
17794	1.724	1.597	55349	1.329	1.137
17798	1.132	0.989	55350	1.486	1.422
17801	1.636	1.504	55352	0.722	1.329
17805	1.053	0.944	55353	1.065	1.157
17810	1.134	0.999	55354	1.118	1.092
17819	1.404	1.24	55355	0.754	0.677
17820	1.308	1.4	55362	1.43	1.349
17826	1.163	1.009	55365	1.612	1.608
17827	1.668	1.652	55368	0.972	0.959
17849	1.595	1.317	55369	1.506	1.377
55333	1.217	1.168	average	1.313	1.245

EXAMPLE 22

IMMUNOGENIC EQUIVALENCE OF *HANSENULA*-PRODUCED HCV E1-H6 AND HCV E1 PRODUCED BY VACCINIA-INFECTED MAMMALIAN CELLS

The immunogenecity of *Hansenula*-produced HCV E1-H6 was compared to the immunogenecity of HCV E1 produced by HCV-recombinant vaccinia virus-infected mammalian cells as described by Depla et al. in WO99/67285. Both HCV-E1 preparations tested consisted of VLP's wherein the HCV E1 proteins were alkylated with iodoacetamide. Both VLP preparations were formulated with alum and injected in Balb/c mice (3 intramuscular/subcutaneous injections with a three week interval between each and each consisting of 5 µg E1 in 125 µl containing 0.13% Alhydrogel, Superfos, Denmark). Mice were bled ten days after the third immunization.

Results of this experiment are shown in Figure 51. For the top part of Figure 51, antibodies raised following immunization with VLPs of E1 produced in mammalian cells were determined. Antibody titers were determined by ELISA (see Example 21) wherein either E1 produced in mammalian cells ("M") or *Hansenula*-produced E1 ("H") were coated directly on the ELISA solid support whereafter the ELISA plates were blocked with casein. For the bottom part of Figure 51, antibodies raised following immunization with VLPs of Hansenula-produced E1 were determined. Antibody titers were determined by ELISA (see Example 21) wherein either E1 produced in mammalian cells ("M") or Hansenula-produced E1 ("H") were coated directly on the ELISA solid support whereafter the ELISA plates were blocked with casein.

The antibody titers determined were end point titers. The end point titer is determined as the dilution of serum resulting in an OD (as determined by ELISA) equal to two times the mean of the background of the assay.

Figure 51 shows that no significant differences were observed between the immunogenic properties of both E1-compositions and that the determined antibody titers are independent of the antigen used in the ELISA to perform the end point titration.

The yeast-derived HCV E1 induced upon vaccination a protective response similar to the protective response obtained upon vaccination with alkylated HCV E1 derived from mammalian cell culture. The latter response was able to prevent chronic evolution of HCV after an acute infection.

EXAMPLE 23

ANTIGENIC AND IMMUNOGENIC PROFILE OF *HANSENULA*-PRODUCED HCV E1-H6 WHICH IS SULPHONATED

The reactivity of *Hansenula*-produced HCV E1-H6 with sera from HCV chronic carriers was compared to the reactivity of HCV E1 produced by HCV-recombinant vaccinia virus-infected mammalian cells as described by Depla et al. in WO99/67285. Both HCV-E1 preparations tested consisted of VLP's wherein the *Hansenula*-produced HCV E1 proteins were sulphonated and the HCV E1 produced by mammalian cells was alkylated. The results are given in Table 7. Although the overall (average) reactivity was identical, some major differences were noted for individual sera. This implies that the sulphonated material presents at least some of its epitopes in a way different from alkylated HCV E1.

The immunogenecity of *Hansenula*-produced HCV E1-H6 which was sulphonated was compared to the immunogenecity of *Hansenula*-produced HCV E1-H6 which was alkylated. Both HCV-E1 preparations tested consisted of VLP's. Both VLP preparations were formulated with alum and injected in Balb/c mice (3 intramuscular/subcutaneous injections with a three week interval between each and each consisting of 5 μg E1 in 125 μl containing 0.13% Alhydrogel, Superfos, Denmark). Mice were bled ten days after the third immunization.

Antibody titers were determined similarly as described in Example 22. Surprisingly, immunization with sulphonated material resulted in higher antibody titers, regardless of the antigen used in ELISA to assess these titers (Figure 51; top panel: titration of antibodies raised against alkylated E1; bottom panel: titration of antibodies raised against sulphonated E1; "A": alkylated E1 coated on ELISA plate; "S": sulphonated E1 coated on ELISA plate). However, in this experiment individual titers are different dependent on the antigen used for analysis which confirms the observation noted with sera from HCV patients. Consequently, HCV E1 wherein the cysteine thiol-gorups are modified in a reversible way may be more immunogenic and thus have an increased potency as a vaccine protecting against HCV (chronic infection). In addition thereto, induction of a response to neo-epitopes induced by irreversible blocking is less likely to occur.

Table 7. Antigenicity of alkylated E1 (produced in mammalian cell culture) or sulphonated E1-H6 (produced in *H. polymorpha*) was evaluated on a panel of sera from human HCV chronic carriers ("patient sera") and a panel of control sera ("blood donor sera"). To this purpose E1 was bound to ELISA plates, after which the plates were further saturated with casein. Human sera were added at a 1/20 dilution and bound immunoglobulins were detected with a rabbit-anti-human IgG-Fc specific secondary antibody labeled with peroxidase. Results are expressed as OD-values. The average values are the averages of the OD-values of all serum samples tested.

patient sera

sernr	*Hansenula*	**Mammalian**
17766	0.646	0.333
17777	0.46	0.447
17785	0.74	0.417
17794	1.446	1.487
17801	0.71	0.902
17819	0.312	0.539
17827	1.596	1.576
17849	0.586	0.964
55333	0.69	0.534
55338	0.461	0.233
55340	0.106	0.084
55345	1.474	1.258
55352	1.008	0.668
55355	0.453	0.444
55362	0.362	0.717
55369	0.24	0.452
average	0.706	0.691

blood donor sera

sernr	*Hansenula*	**mammalian**
F500	0.055	0.054
F504	0.05	0.05
F508	0.05	0.054
F510	0.05	0.058
F511	0.05	0.051
F512	0.051	0.057
F513	0.051	0.052
F527	0.057	0.054
average	0.052	0.054

EXAMPLE 24

IDENTICAL ANTIGENIC REACTIVITY OF *HANSENULA*-PRODUCED HCV E1-H6 AND HCV E1 PRODUCED BY VACCINIA-INFECTED MAMMALIAN CELLS WITH SERA FROM VACCINATED CHIMPANZEES

The reactivities of the E1 produced by HCV-recombinant vaccinia virus-infected mammalian cells and the E1-H6 produced by *Hansenula* (both alkylated) with sera from vaccinated chimpanzees and with monoclonal antibodies were compared. Thereto, said E1 proteins were coated directly to ELISA plates followed by saturation of the plates with casein. The end point titers of antibodies binding the E1 proteins coated to the ELISA plates was determined for chimpanzee sera and for specific murine monoclonal antibodies, all obtained from animals immunized with E1 produced by mammalian cells. End point titer determination was done as described in Example 22. The murine monoclonal antibodies used were IGH201 (see Example 15), IGH198 (IGH198 = 23C12 in Maertens et al. in WO96/04385), IGH203 (IGH203 = 15G6 in Maertens et al. in WO96/04385) and IGH202 (IGH202 = 3F3 in Maertens et al. in WO99/50301).

As can be derived from Figure 53, the reactivities of 7 different chimpanzee are identical when tested with E1 protein produced by either *Hansenula* or mammalian cells. The reactivities of the monoclonal antibodies against HCV E1 are also almost equal. Two of the chimpanzees (Yoran and Marti) were involved in a prophylactic vaccine study and were able to clear an acute infection upon challenge while a control animal did not clear the infection. The five other chimpanzees (Ton, Phil, Marcel, Peggy, Femma) were involved in therapeutic vaccination studies and showed a reduction in liver damage, as measured by ALT in serum and/or histological activity index on liver biopsy, upon the HCV E1 immunizations.

The results obtained in this experiment are clearly different from the findings of Mustilli and coworkers (Mustilli, A. C. et al. 1999) who expressed the HCV E2 protein both in *Saccharomyces cerevisiae* and *Kluyveromyces lactis*. The purified yeast-produced E2 was, however, different from the HCV E2 produced by mammalian (CHO) cells in that a lower reactivity was observed with sera from chimpanzees immunized with HCV E2 produced by mammalian cells while reactivity with monoclonal antibodies was higher for the yeast-produced HCV E2.

EXAMPLE 25

GLYCOPROFILING OF HCV E1 BY FLUOROPHORE-ASSISTED CARBOHYDRATE ELECTROPHORESIS (FACE)

The glycosylation profiles were compared of *Hansenula*-produced HCV E1 and HCV E1 produced by HCV-recombinant vaccinia virus-infected mammalian cells as described by Depla et al. in WO99/67285. This was done by means of fluorophore-assisted carbohydrate electrophoresis (FACE). Thereto, oligosaccharides were released from E1s produced by mammalian cells or *Hansenula* by peptide-N-glycosidase (PNGase F) and labelled with ANTS (the E1 proteins were alkylated with iodoacetamide prior to PNGase F digestion). ANTS-labeled oligosaccharides were separated by PAGE on a 21% polyacrylamide gel at a current of 15 mA at 4°C for 2-3 h. From Figure 54, it was concluded that the oligosaccharides on E1 produced by mammalian cells and E1-H6 produced by *Hansenula* migrate like oligomaltose with a degree of polymerization between 7 and 11 monosaccharides. This indicates that the *Hansenula* expression system surprisingly leads to an E1 protein which is not hyperglycosylated and which has sugar chains with a length similar to the sugar chains added to E1 proteins produced in mammalian cells.

EXAMPLE 26.

SEQUENCING OF N-LINKED OLIGOSACCHARADE DERIVED FROM *SACCHAROMYCES*- AND *HANSENULA*-PRODUCED E1s AND FROM E1s PRODUCED BY HCV-RECOMBINANT VACCINIA VIRUS-INFECTED MAMMALIAN CELLS.

E1s (225 µg) purified from a *Saccharomyces* or *Hansenula* culture (see Examples 15-16) or from a culture of RK13 cells infected with a HCV-recombinant vaccinia virus (see Maertens et al. in WO96/04385) and present in PBS, 0.5% betaine was diluted with deglycosilation incubation buffer (50 mM Na_2HPO_4, 0.75 % Nonidet P-40) to a final concentration of 140 µg/mL. The pH of the solution was adjusted to pH 5.5 with concentrated H_3PO_4. To this solution was added 2 U PNGase F (Peptide-N^4-(acetyl-*beta*-glucosaminyl)-asparagine amidase of *Flavobacterium meningosepticum*; EC 3.5.1.52; obtained from Roche) and the sample was incubated overnight at 37°C. After overnight incubation, the pH of the solution

was adjusted to pH 5.5 with concentrated H_3PO_4. Proteins and oligosaccharides were subsequently precipitated by adding 4 volumes of acetone (at –20°C), and the mixtures were incubated at –20°C for 15 min. Samples were centrifuged at 4°C at 13000 rpm during 5 min. The acetone supernatant was discarded and the pellet was incubated at –20°C for 1 hour after adding 150 µL ice-cold 60% methanol. The methanol supernatant containing the released oligosaccharides was collected and dried by rotary evaporation (SpeedVac).

The dried E1s glycans as well as reference oligosaccharides (all from Glyko, Bicester, UK; see Figure 55) Man-9 (11 monosaccharide units), Man-8 (10 monosaccharide units), Man-7 (9 monosaccharide units), Man-6 (8 monosaccharide units) and Man-5 (7 monosaccharide units) were dissolved in 5 µL 2-aminobenzamide (2-AB) labeling reagent (± 0.35 M 2-AB + ± 1 M $NaCNBH_3$ in 30 % HOAc / 70 % DMSO) to obtain a final glycan concentration between 5 and 100 µM. The glycan solution was then incubated for 2 hours at 65°C. After 30 minutes, the sample was mixed by vortexing. After conjugation, the excess of 2-AB was removed as follows. The sample was diluted with 16-µL purified water (MilliQ) and applied to a Sephadex G-10 column (diameter of 1 cm, height of 1.2 cm, Amersham Biosciences; coupled to a VacElut system, Varian) after the column was pulled dry.

The labeled oligosaccharides were eluted by applying 2 x 100-µL purified water (MilliQ) to the column. The eluates of the reference carbohydrates (Man-9, Man-8, Man-7 and Man-6) were dried and stored at –70°C until HPLC analysis. The eluates of the E1s samples as well as the Man-9 reference glycan were distributed over 4 numbered PCR tubes and dried. The reactions as outlined in Table 8 were performed, all reactions were allowed to proceed overnight at 37°C, except for the reaction in tube 3 which was terminated after 1 h. The final concentration of the exoglycosidase enzymes (all obtained from Glyko, Bicester, UK) used were: for α1-2 Mannosidase (*Aspergillus saitoi*): 2 mU/mL; for α-Mannosidase (Jack Bean): 50 U/mL; and for β-Mannosidase (*Helix pomatia*): 4 U/mL.

Table 8. Overview of reaction mixes for sequencing of oligosaccharides.

	Tube 1	Tube 2	Tube 3	Tube 4	Tube 5
E1s (pmol)	400	400	400	400	400
α 1-2 Mannosidase (μL)	-	4	-	-	-
α Mannosidase (μL)	-	-	5	5	5
β Mannosidase (μL)	-	-	-	-	4
Incubation buffer 4x (μL)	5	5	5	5	5
MilliQ H_2O (μL)	15	11	10	10	6

Figure 56 shows a higher oligomannose consisting of 10 mannose moieties coupled to chitobiose. Each terminal mannose residue is linked by an α 1-3 bond to a non-terminal mannose residue. The oligomannose of Figure 56 is fully resistant to cleavage by the exoglycosidase α 1-2 Mannosidase. Long-term (overnight) incubation of the oligomannose of Figure 56 with the exoglycosidase α-Mannosidase will result in cleavage of all α-linkages (α 1-2, α 1-3, α 1-6), but not of the β-linkages. The resulting oligosaccharide will thus be 4'-β-mannosyl chitobiose. This 4'-β-mannosyl chitobiose moiety can be converted to mannose and chitobiose through the action of the exoglycosidase β-Mannosidase. According to the specifications of the supplier (Glyko), α-Mannosidase converts the reference oligosaccharide Man-6 (see Figure 55.D) to 4'-β-mannosyl chitobiose completely and the further conversion thereof to mannose and chitobiose by β-Mannosidase is also reported to be complete.

Figure 57 shows a higher oligomannose consisting of 9 mannose moieties coupled to chitobiose. In this oligomannose, one terminal mannose residue is linked by an α 1-2 bond to the non-terminal mannose residue. Upon action of the exoglycosidase α 1-2 Mannosidase, said α 1-2-linked mannose will be removed. Upon subsequent action of α-Mannosidase and β-Mannosidase, the reaction products as described for the oligomannose of Figure 56 will be obtained. According to the specifications of the supplier (Glyko), α 1-2 Mannosidase is capable of converting the reference oligosaccharides Man-9 and Man-6 to Man-5 (see Figure 55) with an efficiency of >90 %.

Figure 58 shows the reference higher oligomannose Man-9 consisting of 9 mannose moieties coupled to chitobiose. In this oligomannose, all terminal mannose residue is linked by an α 1-

2 bond to a non-terminal mannose residue. Upon action of the exoglycosidase α 1-2 Mannosidase, Man-9 will thus be converted to Man-5, according to the specification of the supplier by > 90%. Subsequent digestion with α-Mannosidase will convert Man-5 to 4'-β-mannosyl chitobiose.

The contents of the different reaction tubes as indicated in Table 8 were dried in a centrifugal vacuum evaporator or in a lyophilizer and stored at –70°C until HPLC analysis. Before applying to the column, each sample (E1s and reference) was dissolved in 25 µL water and loaded on a TSK gel-Amide-80 (0.46 x 25 cm, Tosoh Biosep) column coupled to a Waters Alliance HPLC station.

Separation of the oligosaccharides was carried out at ambient temperature at 1.0 mL/min. Solvent A consisted of 0.1% acetic acid in acetonitrile and solvent B consisted of 0.2 % acetic acid-0.2 % triethylamine in water. Separation of 2-AB labeled oligosaccharides was carried out using 28 % B isocratic for 5 column volumes followed by a linear increase to 45 % B over fifteen column volumes.

The reference oligosaccharide Man-6 is eluting at 53±1 min, Man-7 is eluting at 59±1 min, Man-8 at 67±2 min, and Man-9 at 70±1 min; 4'-β-mannosyl chitobiose is eluting at 10±1 min and chitobiose at 6±1 min (not shown). This is exemplified for the reaction products of Man-9 after overnight incubation without exoglycosidases (trace 1 of the chromatogram in Figure 63; Man-9 only), after overnight incubation with α 1-2 Mannosidase (trace 2 of the chromatogram in Figure 63; mixture of Man-5 and Man-6), after 1 hr or overnight incubation with α-Mannosidase (traces 3 and 4, respectively, of the chromatogram in Figure 63; 4'-β-mannosyl chitobiose only), and after overnight incubation with α- and β-Mannosidase (trace 5 of the chromatogram in Figure 63; chitobiose only). Trace 6 of the chromatogram in Figure 63 is indicating the applied solvent gradient.

The products of the reaction of the oligosaccharides of *Saccharomyces*-produced E1s (obtained after PNGaseF treatment) without exoglycosidases were mainly four carbohydrates present eluting at 59±1 min (15%), 67±1 min (45%), 70±1 min (25%) and 75±1 min (15%). The overal content of Man(8)-GlcNAc(2) and Man(9)GlcNAc(2) in *Saccharomyces*-produced E1s was ~65 %. In the reaction with α 1-2 Mannosidase, only the carbohydrates with retention time 70±1 min has disappeared. The intensity of the carbohydrate with retention time 75±1 min remained the same and the intensity of carbohydrate with retention time 67±1 min was increased. This means that not all terminal mannose units have the α(1-2)

configuration. After overnight incubation with α Mannosidase all carbohydrate chains were reduced to the 4'-β-mannosyl chitobiose moiety. This means that the carbohydrate is high mannose and all mannose residues except one have the α configuration. Reduction of this 4'-β-mannosyl chitobiose moiety to chitobiose was apparent after overnight incubation with β-mannosidase. The resulting chromatograms are depicted in Figure 64 which were obtained under the same conditions as described for the chromatograms of Figure 63. The results are summarized in Table 9.

The same experiments were repeated with E1s produced by vaccinia-infected cells and surprisingly showed a completely different picture. In the reaction without enzymes, a complex mixture of carbohydrates was present (see Figure 65 and Table 9). The overall content of monosaccharide(8)-GlcNAc(2) and monosaccharide(9)-GlcNAc(2) was 37 %. After reaction with α 1-2 Mannosidase, the carbohydrates with retention times 70±1 and 59±1 min have disappeared. After overnight incubation with with α Mannosidase, a substantial amount of monosaccharide(6)-GlcNAc(2) remained in addition to the 4'-β-mannosyl chitobiose product. This is an indication that one of the oligosaccharide branches is resistant to α Mannosidase degradation. This can be explained by the presence of 1 or 2 glucose-residues linked to a Manα(1-2)-terminated branch of the N-linked oligosaccharide. A putative structure of such a glucose-containing oligosaccharide is depicted in Figure 62. The possible reaction products of glucose-containting oligosaccharides is given in Table 10). As no Man-7-equivalent oligosaccharide (i.e. oligosaccharide consisting of 9 monosaccharides) is remaining after the α 1-2 Mannosidase reaction, these glucose residues are most likely linked to the B-branch of the oligosaccharide structure given in Figure 62. It can, however, not be excluded that both the A- and B-branches of the oligosaccharide in Figure 62 are partially terminated by glucose.

Reduction of the 4'-β-mannosyl chitobiose moiety to chitobiose was apparent after overnight incubation with β Mannosidase. The resulting chromatograms are depicted in Figure 65 which were obtained under the same conditions as described for the chromatograms of Figures 63-64.

The obtained results are summarized in Table 9.

The same experiments were repeated *Hansenula*-produced E1s and surprisingly showed a completely different picture. In the reaction without enzymes, mainly two carbohydrates

with retention time 67±2 min and 70±1 min were present corresponding to respectively Man-8 and Man-9. The overal content of Man(8)-GlcNAc(2) and Man(9)GlcNAc(2) in *Hansenula*-produced E1s was ~90 %. After reaction with α 1-2 Mannosidase, carbohydrates were reduced to mainly Man-5 with retention time 45±1 min and a Man-6 with retention time 53±1 min. After overnight incubation with with α Mannosidase all carbohydrate chains were reduced to the 4'-β-mannosyl chitobiose moiety. This means that the carbohydrate is high mannose and all mannose residues except one have the α configuration. Reduction of this 4'-β-mannosyl chitobiose moiety to chitobiose was apparent after overnight incubation with β Mannosidase. The resulting chromatograms are depicted in Figure 66 which were obtained under the same conditions as described for the chromatograms of Figures 63-65. The obtained results are summarized in Table 9.

Table 9. Oligomannoses resulting from digestion of oligomannoses derived from *Saccharomyces* ("Sc")-, and *Hansenula* ("Hp")-produced E1s as well as from E1s produced by HCV-recombinant vaccinia-infected mammalian cells ("Vac"). Indicated are the different oligomannoses with their chromatographic retention times ("Rt", in minutes) and the percentage of a given oligomannose relative to the total oligomannose content (top rows) as well as the most likely structure for each observed oligomannose indicated with reference to any of Figures 55-62. Oligomannoses with terminal α 1-3 mannoses have been marked with a "°". Oligomannoses with terminal glucoses have been marked with a "*", for some reference is made to "from 62*", meaning that the structure can be derived from the structure given in Figure 62. "1" is the 'reaction' without exoglycosidases, "2" is the reaction with α 1-2 Mannosidase. The oligomannose with retention time 45±1 min is supposedly an oligomannose comprising 5 mannose residues linked to chitobiose. The oligomannose with retention time 75±1 min is supposedly an oligomannose comprising 10 mannose residues linked to chitobiose.

	(Man-5) Rt: 45±1	Man-6 Rt: 53±1	Man-7 Rt: 59±1	Man-8 Rt: 67±1	Man-9 Rt: 70±1	(Man-10) Rt: 75±1
Sc 1 structure	/	1 % 55.D	14 % 55.C	42 % 59°	23 % 57° 61°	18 % 56°
Sc 2 structure	17 % 55.E	/	8 % 60°	50 % 59°	6 % 57° 61°	16 % 56°
Vac 1 structure	3 % 55.E	32 % from 62* & Table 10	20 % 55.C	23 % from 62* & Table 10	14 % 62* & Table 10	5 % 62* & Table 10
Vac 1 structure	74 % 55.E	2 % from 62* & Table 10	/	20 % from 62* & Table 10	4 % 62* & Table 10	/ 62* & Table 10
Hp 1 structure	/	2 % 55.D	7 % 55.C	54 % 55.B	36 % 58	/
Hp 2 structure	80 % 55.E	20 % 55.D	/	/	/	/

Table 10. Products of glucose-containing N-linked oligosaccharides after reaction with α 1-2 Mannosidase or α 1-2 Mannosidase and α Mannosidase.

	Man-equivalent	α 1-2 Mannosidase product (1)	α Mannosidase product (after (1))
Branch A + 2 Glc	Man-10	Man-8	Man-6
Branch A + 1 Glc	Man-9	Man-7	Man-6
Branch A no Glc	Man-8	Man-5	4'-β-mannosyl chitobiose
Branch B + 2 Glc	Man-10	Man-9	Man-6
Branch B + 1 Glc	Man-9	Man-8	Man-6
Branch B no Glc	Man-8	Man-5	4'-β-mannosyl chitobiose

EXAMPLE 27

OCCUPATION OF N-GLYCOSYLATION SITES IN RECOMBINANT HCV E1

Dependent on the amount of occupied N-glycosylation sites, E1s shows a different migration behavior on SDS-PAGE analysis. Based on this property, the average amount of occupied N-glycosylation sites in the E1 product could be estimated. Hereto, samples of the purified E1-product were subjected to SDS-PAGE and Coomassie Brilliant Blue staining (Figure 67) and further analyzed by means of the ImageMaster 1D Prime Software packet (Pharmacia). In brief, the gel was scanned and for each specific protein band and the % of occurrence (its intensity in relation to the total intensity of the different bands whereby the total of all bands is 100 %) was estimated (Table 11). It should be noted that each specific protein band is representing E1s-molecules with the same number of occupied N-glycosylation sites

The obtained results indicated that the main part (> 90 %) of the *Hansenula*-produced E1-product has 1 or more N-glycosylation sites less occupied than the E1s obtained from the vaccinia expression system (as described by Maertens et al. in WO96/04385). Assuming that in the vaccinia derived E1 product all the N-glycosylation sites are occupied (in which the

sequence "NNSS" (SEQ ID NO:73) on position 233-236 of E1 is considered as one glycosylation site), it is quite safe to conclude that the average number of occupied N-glycosylation sites in the *Hansenula*-expressed E1 protein does not exceed 80 % of the total available N-glycosylation sites.

Table 11. Estimation of the average number of occupied N-glycosylation sites in the E1 proteins obtained from the *Hansenula polymorpha* and vaccinia/vero expression systems by means of SDS-PAGE and Coomassie Brilliant Blue staining intensity analysis. The protein bands are indicated by their molecular weight. See Figure 67.

Alkylated E1s	**% occurrence (relative intensity)**	
Hansenula polymorpha	Experiment 1	Experiment 2
MW 29	9	8
MW 25	25	27
MW 21	38	42
MW 18	27	22
MW 14-15	not quantified	not quantified
Vaccinia/ Vero		
MW 29	100	100

EXAMPLE 28

OCCUPATION OF N-GLYCOSYLATION SITES IN RECOMBINANT HCV E2

Two-hundred (200) µg of the E2-H6 protein produced by *Hansenula* was deglycosylated with PNGaseF. The deglycosylated E2s-H6 was loaded on a mini-gel (10 µg/lane). The protein bands were digested with trypsine and endo Asp-N. The masses of the resulting peptides were determined by Maldi-MS (dried droplet and thin layer method).

This method can be used to determine the degree of N-glycosylation: during the deglycosylation with the enzyme PNG-ase F the complete sugar chain is cleaved off, and at

the same time the asparagine (N) is hydrolyzed to aspartic acid (D). The mass difference between these two amino acids is 1 Da which can be determined by mass spectrometry. Additionally, the hydrolysis of N to D creates new cleavage sites for the Asp-N enzyme.

Possible glycosylation sites in E2s are N_{417}, N_{423}, N_{430}, N_{448}, N_{478}, N_{532}, N_{540}, N_{556}, N_{576}, N_{623} and N_{645} (see Figure 68). Maldi-MS analysis showed for each of these glycosylation sites that N-glycosylation was not complete, because after deglycosylation with PNGase F, peptides were found with either N or D at the glycosylation site (mass difference 1 Da). The ratio of the number of D-residues over the number of N-residues gives an indication of the average occupation, over all E2 proteins expressed by Hansenula and present in the analyzed sample, of a single N-glycosylation site by a sugar chain. These results are summarized in Tables 12 to 14.

From these results it was calculated that on average each glycosylation site was glycosylated for approximately **54 %.**

Table 12. Percent glycosylation determined from tryptic peptides containing one N-glycosylation site.

N-glycosylation site	Glycosylated	Not glycosylated
N430	**60 %**	40 %
N448	**50 %**	50 %
N556	**80 %**	20 %
N576	**90 %**	10 %
N623	**20 %**	80 %
N645	**10 %**	90 %

Table 13. Percent glycosylation determined from tryptic peptides containing two N-glycosylation sites.

N-glycosylation sites	Both sites glycosylated	1 out of 2 sites glycosylated	No site glycosylated	Calculated for individual N
N417 and N423	70 %	25 %	5 %	**85 %**
N532 and N540	0 %	80 %	20 %	**40 %**

Table 14. Percent glycosylation determined for N478 in Asp-N digest.

N-glycosylation site	Glycosylated	Not glycosylated
N478	**35 %**	65 %

EXAMPLE 29

REACTIVITY OF BLOOD DONOR SERA WITH HCV E1 PRODUCED BY *SACCHAROMYCES* OR *HANSENULA*

E1s-H6 produced by *Saccharomyces* (expressed with the α-MF leader) and E1s-H6 produced by *Hansenula* (expressed with the CL leader) were both purified as described in Examples 15 and 16 and were finally subjected to alkylation and VLP formation as described in Example 20. Both proteins were directly adsorbed on a microtiterplate at 0.5 µg/mL (1h, 37°C) and after blocking of the plate (PBS-0.1% casein, 1h, 37°C) sera from HCV-screened and negative blood donors were incubated at a dilution of 1/20 (PBS-0.5% casein, 10% (w/v) sucrose, 0.2% (v/v) Triton X-705, 1h, 37°C). Finally binding was detected using a secondary rabbit anti-human IgG-F_c specific antiserum coupled to peroxidase (Dako, Denmark) at a dilution of 1/50000 (PBS-0.1% casein, 1h, RT) followed by color development. Between all steps plates were washed 3 times with PBS-0.05% (w/v) Tween-20. For comparison the sera were also analyzed in an identical way on mammalian cell derived E1s, produced and purified as described by Depla et al. in WO99/67285.

The cut-off for this ELISA was set at 2 times the mean of the background (i.e. the reactivity of all sera in an identical set-up but with streptavidin adsorbed to the wells).

From Table 15 it can be judged that many (75 %) sera show reactivity above the cut-off towards the *Saccharomyces*-produced E1 while only a few sera (6 %) show some reactivity above cut-off with the *Hansenula*-produced E1. This difference in reactivity was attributed to the presence of terminal α 1-3 mannoses linked to α 1-2 mannoses on the *Saccharomyces*-produced E1 as evidenced in Example 26. Young and coworkers (1998) already previously indicated that this type of mannoses is also responsible for reactivity of human sera with *Saccharomyces* derived mannan. In order to further confirm that the reactivity on *Saccharomyces*-derived E1 can be attributed to this type of mannose residues, the ELISA on *Saccharomyces*-produced E1 was repeated with dilutions of blood donor sera preincubated

(1h at 37°C) with 1 or 5 mg/mL mannan (Sigma) added to the dilution buffer. As can be judged from Table 16, preincubation with mannan reduced the reactivity of this E1 with blood donor sera in a concentration dependent way to background levels for all but one (F556) sera analyzed. (mean OD is reduced from 0.24 without competition by mannan to 0.06, using 5 mg mannan/mL).

Table 15. Reactivity of E1 produced in Hansenula, Saccharomyces and mammalian cells. The reactivities above the cut-off value are indicated in black shaded cells.

serum nr	*Hansenula* E1	*Saccharomyces* E1	Mammalian E1		blank
F552	0,18	0,224	0,056		0,05
F553	0,062	0,449	0,056		0,052
F555	0,06	0,079	0,054		0,051
F556	0,073	0,679	0,054		0,051
F557	0,059	0,173	0,053		0,05
F558	0,066	0,232	0,06		0,058
F559	0,084	0,309	0,056		0,053
F560	0,062	0,338	0,052		0,052
F562	0,056	0,128	0,053		0,053
F563	0,064	0,181	0,059		0,056
F570	0,056	0,135	0,054		0,055
F571	0,06	0,209	0,054		0,055
F572	0,061	0,427	0,055		0,056
F575	0,079	0,104	0,062		0,056
F576	0,061	0,144	0,058		0,057
F577	0,063	0,224	0,055		0,058
F578	0,089	0,131	0,057		0,061
F581	0,064	0,098	0,061		0,057
F594	0,055	0,116	0,056		0,057
F595	0,059	0,539	0,057		0,058
F598	0,076	0,311	0,056		0,059
F450	0,205	0,078	0,099		0,059
F453	0,059	0,128	0,057		0,06
F456	0,058	0,121	0,056		0,06
F458	0,055	0,088	0,054		0,054
F459	0,054	0,069	0,056		0,054
F463	0,055	0,083	0,054		0,056
F466	0,086	0,208	0,071		0,094
F467	0,066	0,344	0,055		0,055
F469	0,059	0,074	0,057		0,057
F470	0,074	0,222	0,056		0,056
F473	0,094	0,807	0,054		0,057
F479	0,06	0,075	0,06		0,051
F480	0,053	0,305	0,056		0,053
F481	0,059	0,395	0,071		0,052
F488	0,063	0,467	0,059		0,053
average	0,072	0,242	0,058	cut-off	0,113
# of sera above cut-off	2/36	27/36	0/36		
% of sera above cut-off	6	75			

Table 16. Reactivity of E1 produced in Saccharomyces cells as in Table 15 but in the presence of 5 mg mannan/mL. The reactivities above the cut-off value are indicated in black shaded cells.

Serum nr	Mannan concentration		
	0 mg/mL	1 mg/mL	5 mg/mL
F552	0,207	0,128	0,103
F553	0,487	0,098	0,050
F555	0,066	0,044	0,041
F556	0,769	0,540	0,372
F557	0,158	0,094	0,088
F558	0,250	0,076	0,046
F559	0,300	0,077	0,066
F560	0,356	0,088	0,044
F562	0,122	0,106	0,089
F563	0,164	0,091	0,049
F570	0,110	0,043	0,040
F571	0,212	0,057	0,042
F572	0,464	0,087	0,043
F575	0,095	0,081	0,062
F576	0,138	0,042	0,043
F577	0,216	0,042	0,041
F578	0,125	0,100	0,093
F581	0,083	0,064	0,042
F594	0,102	0,044	0,041
F595	0,520	0,088	0,044
F598	0,340	0,054	0,042
F450	0,053	0,060	0,053
F453	0,116	0,049	0,044
F456	0,112	0,050	0,043
F458	0,086	0,051	0,042
F459	0,054	0,044	0,042
F463	0,078	0,043	0,041
F466	0,172	0,111	0,085
F467	0,420	0,117	0,049
F469	0,053	0,043	0,041
F470	0,220	0,070	0,061
F473	0,924	0,183	0,063
F479	0,059	0,049	0,043
F480	0,281	0,155	0,054
F481	0,355	0,042	0,046
F488	0,474	0,090	0,046
average	0,243	0,089	0,062

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CLAIMS

1. An isolated HCV envelope protein or a fragment thereof comprising at least one N-glycosylation site, said protein or fragment thereof characterized in that it is the product of expression in a eukaryotic cell and further characterized in that on average up to 80 % of the N-glycosylation sites are core-glycosylated.

2. The isolated HCV envelope protein or fragment thereof according to claim 1 wherein more than 70 % of said core-glycosylated sites are glycosylated with an oligomannose containing 8 to 10 mannoses.

3. The isolated HCV envelope protein or fragment thereof according to any of claims 1 or 2 wherein the ratio of sites core-glycosylated with an oligomannose with a structure defined by Man(7)-GlcNAc(2) over the sites core-glycosylated with an oligomannose with a structure defined by Man(8)-GlcNAc(2) is less than or equal to 0.45.

4. The isolated HCV envelope protein or fragment thereof according to any of claims 1 to 3 wherein said oligomannoses contain less than 10 % terminal α1,3 mannose.

5. The isolated HCV envelope protein or part thereof according to any of claims 1 to 4 which is the product of expression in a yeast cell.

6. The isolated HCV envelope protein or part thereof according to claim 5 wherein said yeast cell is a *Hansenula* cell.

7. The isolated HCV envelope protein or part thereof according to any of claims 1 to 6 which is derived from a protein comprising an avian lysozyme leader peptide or a functional variant thereof joined to said HCV envelope protein or fragment thereof.

8. The isolated HCV envelope protein or part thereof according to claim 7 which is derived from a protein characterized by the structure

   $CL-[(A1)_a - (PS1)_b - (A2)_c]-HCVENV-[(A3)_d - (PS2)_e - (A4)_f]$

   wherein:

CL is an avian lysozyme leader peptide or a functional equivalent thereof,
A1, A2, A3 and A4 are adaptor peptides which can be different or the same,
PS1 and PS2 are processing sites which can be the different or the same,
HCVENV is a HCV envelope protein or a part thereof,
a, b, c, d, e and f are 0 or 1, and
wherein, optionally, A1 and/or A2 are part of PS1 and/or wherein A3 and/or A4 are part of PS2.

9. The isolated HCV envelope protein or fragment thereof according to claim 8 wherein said avian lysozyme leader peptide CL has an amino acid sequence defined by SEQ ID NO:1.

10. The isolated HCV envelope protein or fragment thereof according to claim 8 wherein A has an amino acid sequence chosen from SEQ ID NOs:63-65, 70-72 and 74-82, wherein PS has an amino acid sequence chosen from SEQ ID NOs:66-68 and 83-84 or wherein PS is a dibasic site such as Lys-Lys, Arg-Arg, Lys-Arg and Arg-Lys or a monobasic site such as Lys, and wherein HCVENV is chosen from SEQ ID NOs:85-98 and fragments thereof.

11. The isolated HCV envelope protein or fragment thereof according to any of claims 1 to 10 which is comprised in a structure chosen from the group consisting of monomers, homodimers, heterodimers, homo-oligomers and hetero-oligomers.

12. The isolated HCV envelope protein or fragment thereof according to any of claims 1 to 10 which is comprised in a virus-like particle.

13. The isolated HCV envelope protein or fragment thereof according to any of claims 1 to 12 in which the cysteine thiol-groups are chemically modified.

14. The isolated HCV envelope protein or fragment thereof according to any of claims 1 to 13 which is antigenic.

15. The isolated HCV envelope protein or fragment thereof according to any of claims 1 to 13 which is immunogenic.

16. The isolated HCV envelope protein or fragment thereof according to any of claims 1 to 13 which comprises a T-cell epitope.

17. A composition comprising an isolated HCV envelope protein or fragment thereof according to any of claims 1 to 16.

18. The composition according to claim 17 further comprising a pharmaceutically acceptable carrier and which is a medicament.

19. The composition according to claim 17 further comprising a pharmaceutically acceptable carrier and which is a vaccine.

20. A method for producing the isolated HCV envelope protein or fragment thereof according to any of claims 1 to 16.

21. A method for the detection of the presence of anti-HCV antibodies in a sample suspected to comprise anti-HCV antibodies, said method comprising:
(i) contacting a HCV envelope protein or part thereof according to any of claims 1 to 16 with said sample under conditions allowing complexation of said HCV envelope protein or part thereof with said anti-HCV antibodies,
(ii) detecting the complex formed in (i), and
(iii) inferring from (ii) the presence of said anti-HCV antibodies in said sample.

22. The method according to claim 21 wherein said contacting in step (i) is occurring under competitive conditions.

23. The method according to claim 21 wherein said HCV envelope protein or part thereof is attached to a solid support.

24. A diagnostic kit for the detection of the presence of anti-HCV antibodies in a sample suspected to comprise anti-HCV antibodies, said kit comprising a HCV envelope protein or part thereof according to any of claims 1 to 16.

25. The diagnostic kit according to claim 24 wherein said HCV envelope protein or part thereof is attached to a solid support.

26. A medicament comprising a HCV envelope protein or part thereof according to any of claims 1 to 16.

27. A vaccine comprising a HCV envelope protein or part thereof according to any of claims 1 to 16.

28. A pharmaceutical composition for inducing a HCV-specific immune response in a mammal, said composition comprising an effective amount of a HCV envelope protein or part thereof according to any of claims 1 to 16 and, optionally, a pharmaceutically acceptable adjuvant.

29. A pharmaceutical composition for inducing HCV-specific antibodies in a mammal, said composition comprising an effective amount of a HCV envelope protein or part thereof according to any of claims 1 to 16 and, optionally, a pharmaceutically acceptable adjuvant.

30. A pharmaceutical composition for inducing a T-cell function in a mammal, said composition comprising an effective amount of a HCV envelope protein or part thereof according to any of claims 1 to 16 and, optionally, a pharmaceutically acceptable adjuvant.

31. The pharmaceutical composition according to any of claims 28 to 30 which is a prophylactic composition.

32. The pharmaceutical composition according to any of claims 28 to 30 which is a therapeutic composition.

33. The pharmaceutical composition according to any of claims 28 to 32 wherein said mammal is a human.