MODIFYING THE EXPRESSION OF THE FSH BETA GENE BY HOMOLOGOUS RECOMBINATION

Field of the Invention

This invention relates to genomic DNA.

Background of the Invention

Current approaches to treating disease with therapeutic proteins include both administration of proteins produced *in vitro* and gene therapy. *In vitro* production of a protein generally involves the introduction of exogenous DNA coding for the protein of interest into appropriate host cells in culture. Gene therapy methods, on the other hand, involve administering to a patient genetically engineered cells, plasmids, or viruses that contain a sequence encoding the therapeutic protein of interest.

Certain therapeutic proteins may also be produced by altering the expression of their endogenous genes in a desired manner with gene targeting techniques. See, e.g., U.S. Patent Nos. 5,641,670, 5,733,761, and 5,272,071; WO 91/06666; WO 91/06667; and WO 90/11354, all of which are incorporated by reference in their entirety.

Summary of the Invention

The present invention is based upon the identification and sequencing of genomic DNA 5′ to the coding sequences of the human follicle-stimulating hormone ß ("FSHß") gene. This DNA can be used, for example, in a DNA construct that alters (e.g., increases) expression of an endogenous FSHß gene in a mammalian cell upon integration into the genome of the cell via homologous recombination. "Endogenous FSHβ gene" refers to a genomic (i.e., chromosomal) copy of a gene that encodes FSHβ. The construct contains a targeting sequence including or derived from the newly disclosed 5′ noncoding sequence,

and a transcriptional regulatory sequence. The transcriptional regulatory sequence preferably differs in sequence from the transcriptional regulatory sequence of the endogenous FSHβ gene. The targeting sequence directs the integration of the regulatory sequence into a region witthin or upstream of the FSHβ-coding sequences of the target gene such that the regulatory sequence becomes operatively linked to the endogenous coding sequence. By "operatively linked" is meant that the regulatory sequence can direct expression of the endogenous FSHβ-coding sequence. The construct may additionally contain a selectable marker gene to facilitate selection of cells that have stably integrated the construct, and/or another coding sequence operatively linked to a promoter.

In one embodiment, the DNA construct contains: (a) a targeting sequence, (b) a regulatory sequence, (c) an exon, and (d) a splice-donor site. The targeting sequence directs the integration of itself and elements (b) - (d) into a region within or upstream of the FSHβ-coding sequences of the target gene. Once integrated, element (b) can direct transcription of elements (c) and (d) and all downstream coding sequences of the endogenous gene. In the construct, the exon is generally 3′ of the regulatory sequence, and the splice-donor site is at the 3′ end of the exon.

In another embodiment, the DNA construct comprises: (a) a targeting sequence, (b) a regulatory sequence, (c) an exon, (d) a splice-donor site, (e) an intron, and (f) a splice-acceptor site, wherein the targeting sequence directs the integration of itself and elements (b) - (f) such that elements (b) - (f) are within or upstream of the endogenous gene. The regulatory sequence then directs production of a transcript that includes not only elements (c) - (f), but also the endogenous FSHß coding sequence. Preferably, the intron and the splice-acceptor

site are situated in the construct downstream from the splice-donor site.

The targeting sequence is homologous to a pre-selected target site in the genome with which homologous recombination is to occur. It contains at least 20 (e.g., at least 30, 50, 100, or 1000) contiguous nucleotides from SEQ ID NO:4, which corresponds to nucleotides -7454 to -1417 of human FSHß genomic sequence (numbering relative to the translation start site), or SEQ ID NO:5, which corresponds to nucleotides -696 to -155 of human FSHß genomic sequence. By "homologous" is meant that the targeting sequence is identical or sufficiently similar to its genomic target site so that the targeting sequence and target site can undergo homologous recombination within a human cell. A small percentage of basepair mismatches is acceptable, as long as homologous recombination can occur at a useful frequency. To facilitate homologous recombination, the targeting sequence is preferably at least about 20 (e.g., at least 50, 100, 250, 400, or 1,000) base pairs ("bp") long. The targeting sequence can also include genomic sequences from outside the region covered by SEQ ID NO:4 or 5, so long as it includes at least 20 nucleotides from within one of the two regions. For example, additional targeting sequence could be derived from the sequence lying between SEQ ID NO:4 and the transcription initiation sequence of the FSHß gene.

Due to polymorphism that may exist at the FSHβ genetic locus, minor variations in the nucleotide composition of any given genomic target site may occur in any given mammalian species. Targeting sequences that correspond to such polymorphic variants (particularly human polymorphic variants) of SEQ ID NO:4 or 5 are within the scope of this invention.

Upon homologous recombination, the regulatory sequence of the construct is integrated into a pre-selected region upstream of the coding sequence of a FSHβ gene in a chromosome of a cell. The resulting new transcription unit containing the construct-derived regulatory sequence alters the expression of the target FSHβ gene. The FSHβ protein so produced may be identical in sequence to the FSHβ protein encoded by the unaltered, endogenous gene, or may contain additional, substituted, or fewer amino acid residues as compared to the wild type FSHβ protein, due to changes introduced as a result of homologous recombination.

Altering gene expression encompasses activating (or causing to be expressed) a gene which is normally silent (i.e., essentially unexpressed) in the cell as obtained, increasing or decreasing the expression level of a gene, and changing the regulation pattern of a gene such that the pattern is different from that in the cell as obtained. "Cell as obtained" refers to the cell prior to homologous recombination.

Also within the scope of the invention is a method of using the present DNA construct to alter expression of an endogenous FSHβ gene in a mammalian cell. This method includes the steps of (i) introducing the DNA construct into the mammalian cell, (ii) maintaining the cell under conditions that permit homologous recombination to occur between the construct and a genomic target site homologous to the targeting sequence, to produce a homologously recombinant cell; and (iii) maintaining the homologously recombinant cell under conditions that permit expression of the FSHβ-coding sequence under the control of the construct-derived regulatory sequence. At least a part of the genomic target site is 5' to the coding sequence of an endogenous FSHβ gene. That is, the genomic target

site can contain coding sequence as well as 5' non-coding sequence.

The invention also features transfected or infected cells in which the construct has undergone homologous recombination with genomic DNA upstream of the endogenous ATG initiation codon in one or both alleles of the endogenous FSHβ gene. Such transfected or infected cells, also called homologously recombinant cells, have an altered FSHβ expression pattern. These cells are particularly useful for *in vitro* FSHβ production and for delivering FSHβ via gene therapy. Methods of making and using such cells are also embraced by the invention. The cells can be of vertebrate origin such as mammalian (e.g., human, non-human primate, cow, pig, horse, goat, sheep, cat, dog, rabbit, mouse, guinea pig, hamster, or rat) origin.

The invention further relates to a method of producing a mammalian FSHβ protein *in vitro* or *in vivo* by introducing the above-described construct into the genome of a host cell via homologous recombination. The homologously recombinant cell is then maintained under conditions that allow transcription, translation, and optionally, secretion of the FSHβ protein.

The invention also features isolated nucleic acids comprising a sequence of at least 20 (e.g., at least 30, 50, 100, 200, or 1000) contiguous nucleotides of SEQ ID NO:4, or at least 20 (e.g., at least 30, 50, 100, or 200) contiguous nucleotides of SEQ ID NO:5, or of a similar-sized portion of a sequence identical to SEQ ID NO:4 or 5 except for polymorphic variations or other minor variations (e.g., less than 5% of the sequence) which do not prevent homologous recombination with the target sequence.

In one embodiment, the isolated nucleic acid of the invention includes a contiguous 100 bp block of SEQ

ID NO:4 or 5. For example, the isolated DNA can contain nucleotides 1 to 100, 101 to 200, 201 to 300, 301 to 400, 401 to 500, 501 to 600, 601 to 700, 701 to 800, 801 to 900, 901 to 1000, 1001 to 1100, 1101 to 1200, 1201 to 1300, 1301 to 1400, 1401 to 1500, 1501 to 1600, 1601 to 1700, 1701 to 1800, 1801 to 1900, 1901 to 2000, 2001 to 2100, 2101 to 2200, 2201 to 2300, 2301 to 2400, 2401 to 2500, 2501 to 2600, 2601 to 2700, 2701 to 2800, 2801 to 2900, 2901 to 3000, 3001 to 3100, 3101 to 3200, 3201 to 3300, 3301 to 3400, 3401 to 3500, 3501 to 3600, 3601 to 3700, 3701 to 3800 3801 to 3900, 3901 to 4000, 4001 to 4100, 4101 to 4200, 4201 to 4300, 4301 to 4400, 4401 to 4500, 4501 to 4600, 4601 to 4700, 4701 to 4800, 4801 to 4900, 4901 to 5000, 5001 to 5100, 5101 to 5200, 5201 to 5300, 5301 to 5400, 5401 to 5500, 5501 to 5600, 5601 to 5700, 5701 to 5800, 5801 to 5900, 5901 to 6000, or 5939 to 6038 of SEQ ID NO:4 or its complement. Alternatively, the isolated nucleic acid of the invention can include nucleotides 1 to 100, 101 to 200, 201 to 300, 301 to 400, 401 to 500, or 443 to 542 of SEQ ID NO:5. These blocks of SEQ ID NO:4 or 5 or their complements are useful as targeting sequences in the constructs of the invention.

In the isolated DNA, the contiguous nucleotide sequence is not linked to a sequence encoding full-length FSHß, or at least not linked in the same configuration (i.e., separated by the same sequence) as in any native genome. The term "isolated DNA", as used herein, thus does not denote a chromosome or large piece of genomic DNA (as might be incorporated into a cosmid or yeast artificial chromosome) that includes not only part or all of SEQ ID NO:4 or 5, but also an intact FSHβ-coding sequence and all of the sequence which lies between the FSHß coding sequence and the sequence corresponding to SEQ ID NO:4 or 5 as it exists in the genome of a cell. It does include, but is not limited to, a DNA (i) which

is incorporated into a plasmid or virus; or (ii) which exists as a separate molecule independent of other sequences, e.g., a fragment produced by polymerase chain reaction ("PCR") or restriction endonuclease treatment. The isolated DNA preferably does not contain a sequence which encodes intact FSHβ precursor (i.e., FSHß complete with its endogenous secretion signal peptide).

The invention also includes isolated DNA comprising a strand which contains a sequence that is at least 100 (e.g., at least 200, 400, or 1000) nucleotides in length and that hybridizes under either highly stringent or moderately stringent conditions with SEQ ID NO:4 or 5, or the complement of SEQ ID NO:4 or 5. The sequence is not linked to a FSHβ-coding sequence, or at least not linked in the same configuration as occurs in any native genome. By moderately stringent conditions is meant hybridization at 50°C in Church buffer (7% SDS, 0.5% $NaHPO_4$, 1 M EDTA, 1% bovine serum albumin) and washing at 50°C in 2X SSC. Highly stringent conditions are defined as hybridization at 42°C in the presence of 50% formamide; a first wash at 65°C with 2X SSC containing 1% SDS; followed by a second wash at 65°C with 0.1X SSC.

Also embraced by the invention is isolated DNA comprising a strand which contains a sequence that (i) is at least 100 (e.g., at least 200, 400, or 1000) nucleotides in length and (ii) shares at least 80% sequence (e.g., 85%, 90%, 95%, or 98%) identity with a segment of equal length from SEQ ID NO:4 or 5, or from the complement of SEQ ID NO:4 or 5. The sequence is not linked to a FSHβ-coding sequence, or at least not linked in the same configuration as occurs in any native genome.

Where a particular polypeptide or nucleic acid molecule is said to have a specific percent identity or conservation to a reference polypeptide or nucleic acid molecule, the percent identity or conservation is deter-

mined by the algorithm of Myers and Miller, CABIOS (1989-), which is embodied in the ALIGN program (version 2.0), or its equivalent, using a gap length penalty of 12 and a gap penalty of 4 where such parameters are required. All other parameters are set to their default positions. Access to ALIGN is readily available. See, e.g., http://www2.igh.cnrs.fr/bin/align-guess.cgi on the Internet.

The invention also features a method of delivering FSHß to an animal (e.g., a mammal such as a human, non-human primate, cow, pig, horse, goat, sheep, cat, dog, rabbit, mouse, guinea pig, hamster, or rat) by providing a cell whose endogenous FSHß gene has been activated as described herein, and implanting the cell in the animal, where the cell secretes FSHß. Also included in the invention is a method of producing FSHß by providing a cell whose endogenous FSHß gene has been activated as described herein, and culturing the cell *in vitro* under conditions which permit the cell to express and secrete FSHß.

The isolated DNA of the invention can be used, for example, as a source of an upstream PCR primer for use (when combined with a suitable downstream primer) in obtaining the regulatory and/or coding regions of an endogenous FSHß gene, or as a hybridization probe for indicating the presence of chromosome 11 in a preparation of human chromosomes. It can also be used, as described below, in a method for altering the expression of an endogenous FSHß gene in a vertebrate cell.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Exemplary methods and materials are described below, although methods and materials similar or equivalent to those described herein can also be used in the practice or testing of the present invention.

All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. The materials, methods, and examples are illustrative only and not intended to be limiting.

Other features and advantages of the invention will be apparent from the following detailed description, and from the claims.

Brief Description of the Drawings

Fig. 1 is a schematic diagram showing the genomic structure of the human FSHβ gene.

Fig. 2 is a schematic diagram showing the genomic region of the human FSHβ gene (top) encompassed by the insert (bottom) of plasmid pHFB2. The three bars in the middle represent genomic regions of the gene whose sequences have been published.

Fig. 3 a representation of a partial sequence (SEQ ID NO:1) of a human FSHβ gene, including 7,454 nucleotides of the sequence 5' to the ATG initiation codon. Also shown is a partial polypeptide sequence (SEQ ID NO:2) encoded by the coding sequence. Published sequences are underlined. "SD" and "SA" stand for splice-donor site and splice-acceptor site, respectively. "Mature" denotes the beginning of a mature FSHβ protein.

Fig. 4 is a schematic diagram showing a construct of the invention. The construct contains a first targeting sequence (1); an amplifiable marker gene (AM); a selectable marker gene (SM); a regulatory sequence; a CAP site; a sequence identical to the first, non-coding exon of a human FSHβ gene; an unpaired splice-donor site (SD); and a second targeting sequence (2). The black boxes represent coding DNA and the stippled boxes represent untranslated sequences.

Figs. 5-7 are schematic diagrams illustrating three constructs of the invention. The constructs differ in the size of the aldolase 5' UTS inserted into the plasmid. These constructs include a sequence encoding a glycoprotein α-subunit (i.e, FSHα) linked to a cytomegalovirus ("CMV") promoter. The abbreviations shown in the figure are: "UTS" for untranslated sequence; "amp" for ampicillin; "Ori" for replication origin,; "SD" for splice-donor site; "HSV TK" for herpes simplex virus thymidine kinase gene; "DHFR" for dihydrofolate reductase; "HBV" for hepatitis B virus; and "hGH" for human growth hormone.

Fig. 8 is a bar graph of FSH production from HT-1080 cells transfected with pGA308 (Fig. 5) and selected for growth in the presence of various concentrations of methotrexate.

Fig. 9 is a representation of SEQ ID NO:4, a sequence upstream of a human FSHß transcription start site.

Fig. 10 is a representation of SEQ ID NO:5, a sequence upstream of a human FSHß transcription start site.

Fig. 11 is a representation of a first targeting sequence (SEQ ID NO: 6) used in a construct of the invention.

Fig. 12 is a representation of a second targeting sequence (SEQ ID NO: 5) used in a construct of the invention.

Detailed Description

The present invention is based on the discovery of the nucleotide composition of sequences upstream to the coding sequence of a human FSHβ gene.

FSH is a gonadotrophin which plays an essential role in the maintenance and development of oocytes and

spermatozoa in normal reproductive physiology. FSH possesses two subunits, α and β, the latter being responsible for FSH's biological specificity.

The human FSHβ gene encodes a 129 amino acid precursor protein containing a 16 amino acid signal peptide. The gene contains three exons and two introns, with the first exon being a non-coding exon. The genomic map of the human FSHβ gene is shown in Fig. 1. The map is constructed based on published sequences (HUMFSHBQ1, GenBank accession numbers M54912, M38644, M21219, and M18536) that correspond to three separate genomic segments (Fig. 1). The first segment is 720 bp long and contains 530 bp of nontranscribed upstream sequences, exon 1 (63 bp; non-coding), and 127 bp of intron 1. The second segment begins at position -152 and ends at position +367 (all positions referred to herein are relative to the translational initiation site, unless specified otherwise). This segment includes 146 bp of intron 1, exon 2 (165 bp), and 208 bp of intron 2. The third segment contains 102 bp of intron 2 and exon 3, and extends 1,480 bp past the translational stop codon.

Specific Sequences 5' to a FSHβ Coding Sequence and Their Use in Altering Endogenous FSHß Gene Expression

To obtain genomic DNA containing sequence upstream to a FSHβ gene, a human leukocyte genomic library in lambda EMBL3 (Clontech catalog # HL1006d) was screened with a 40 bp oligonucleotide probe, BETA2. This probe is derived from 23 bp of exon 1 and 17 bp of intron 1, and has the following sequence:

5' TTGGCATCTACCGTTTTCAAGTGGTGACAGCTACTTTTGA 3'

(SEQ ID NO:3)

Approximately one million recombinant phage were screened with the radiolabelled BETA2 probe. One phage plaque, designated clone 8-1-1-1, was isolated. The 7.6 kb HindIII-KpnI fragment from phage 8-1-1-1 was subcloned

into pBluescript II SK+ (Stratagene, La Jolla, CA) to produce a plasmid containing about 6.6 kb of upstream sequences, exon 1, intron 1, exon 2, and 9 bp of intron 2 (Fig. 2). The plasmid was designated pHFB2.

The pHFB2 plasmid was sequenced by the Sanger method. The sequence data sets were aligned to obtain the complete sequence of the entire phage 8-1-1-1 insert. This nucleotide sequence (SEQ ID NO:1) is shown in Fig. 3.

The insert was shown to encompass a 7,622 bp region of the FSHß gene, starting at position -7,454 (Fig. 3). The sequences encompassing positions -7,454 to -1,417 (6,038 bp of the upstream sequence; SEQ ID NO:4) and positions -696 to -155 (542 bp of intron 1; SEQ ID NO:5) have not been reported previously.

To alter the expression of an endogenous FSHß gene, the general approach shown in Fig. 4 was used. Nucleotides 3860 to 5784 of SEQ ID NO:4 served as the first (5') targeting sequence, while SEQ ID NO:5 served as the second (3') targeting sequence. DNA fragments containing these sequences were then subcloned into plasmids to produce targeting constructs pGA308, pGA301, and pGA307, which are illustrated in Figs. 5-7, respectively. These plasmids each contain about a 3.2 Kb 5' targeting sequence and about a 0.5 Kb 3' targeting sequence.

HT-1080 cells were separately transfected with each of the plasmids and placed under G418 selection. After approximately 14 days, G418 resistant colonies in 6-well plates were counted. In addition, the conditioned medium in each well was screened for GA-FSH expression by ELISA. Cells exhibiting GA-FSH production were trypsinized and counted. The cells were then diluted and plated in 96-well plates to generate clones. After about two weeks of culture, clonal cell populations were screened for GA-FSH production by ELISA. Colonies found to

produce GA-FSH were expanded in culture and stored or further analyzed. Table 1 summarizes the endogenous gene activation frequency and other observations from the above cloning procedure.

The cells transfected with pGA308 were studied in more detail. Fig. 8 indicates the range of FSH production achieved in pGA308-transfected HT1080 cells cultured in media having various concentrations of methotrexate. The bar labeled "0.2 (cloned)" represents the FSH production from a cell line cloned by limiting dilution of cells resistant to 0.2 μM methotrexate. The results graphed in Fig. 8 clearly indicate that higher concentrations of methotrexate can yield cell lines that produce at least 50 $\mu g/10^6$ cells in a day.

Table 1

Plasmid Transfected	Total No. G418-Resistant Colonies	Total No. FSHß Gene Activation Events	Activation Frequency	Total No. Clonal Cell Lines Isolated	Average FSHß Production (ng/10^6 cells in 24 hours)
pGA301	38012	3	1/12671	11	465
pGA307	31068	3	1/10356	20	450
pGA308	27474	4	1/6869	16	521

General Methodologies

Alteration of Endogenous FSHß Expression

Using the above-described FSHβ upstream sequences, one can alter the expression of an endogenous human FSHβ gene by a method as generally described in U.S. Patent No. 5,641,670. One strategy is shown in Fig. 4. In this strategy, a targeting construct is designed to include a first targeting sequence homologous to a first target site upstream of the gene, an amplifiable marker gene, a selectable marker gene, a regulatory region, a CAP site, an exon, an unpaired splice-donor site, and a second targeting sequence corresponding to a second target site downstream of the first target site, and terminating either within or upstream of the FSHβ-coding sequence. In this strategy, the first and second target sites are immediately adjacent in the chromosome prior to homologous recombination, but such configuration is not required (see also below). Homologously recombinant cells will produce an mRNA precursor which corresponds to the exogenous exon and splice-donor site, and any sequence between the splice donor site and the transcription termination sequence of the FSHβ gene, including the FSHβ introns, exons, and 3′ untranslated region (Fig. 4). Splicing of this message results in a mRNA in which the exogenous exon is fused to exon 2 of the endogenous FSHβ gene. Translation of the mRNA produces a precursor FSHβ.

Other approaches can also be employed. For example, the first and/or second target sites can be in the first intron of the FSHβ gene. Alternatively, the DNA construct may be designed to include, from 5′ to 3′, a first targeting sequence, an amplifiable marker gene, a selectable marker gene, a regulatory region, a CAP site, an exon, a splice-donor site, an intron, a splice-acceptor site, and a second targeting sequence. For this

strategy, the 5' end of the second target site is preferably less than 40 bp upstream of the normal FSHβ transcriptional start site, in order to avoid undesired ATG start codons. A mRNA precursor produced from the homologously recombined locus will include the exogenous exon, the exogenous splice-donor site, the exogenous intron, the exogenous splice-acceptor site, and any sequences between the exogenous splice acceptor site and the transcription termination site of the endogenous FSHβ gene. Splicing of this transcript will generate a mRNA which can be translated to produce a precursor of human FSHβ, having either the normal FSHβ secretion signal sequence or a genetically engineered secretion signal sequence. The size of the exogenous intron and thus the position of the exogenous regulatory region relative to the coding region of the endogenous gene can be varied to optimize the function of the regulatory region.

In any activation strategy, the first and second target sites need not be immediately adjacent or even be near each other. When they are not immediately adjacent to each other, a portion of the FSHβ gene's normal upstream region and/or a portion of the coding region would be deleted upon homologous recombination.

If desired, the product of the activated FSHβ gene can be produced in a cell type that expresses a human glycoprotein α-subunit (FSHα) gene, the product of which forms a heterodimer with the product of the FSHβ gene. This may be a naturally occurring cell strain or cell line. Alternatively, the human glycoprotein α-subunit gene (Genbank sequence HUMGLYCA1) can be co-expressed with the product of the FSHβ gene, with such co-expression accomplished by expression of the human glycoprotein α-subunit gene or cDNA under the control of a suitable promoter, or by activation of the human glycoprotein α-subunit gene through the methods described herein.

By way of example, a sequence coding for a glycoprotein α-subunit can be included in the DNA construct. This coding sequence is placed under the transcriptional control of a regulatory sequence that has a nucleotide composition that may be identical to or different from that of the regulatory sequence that is to direct expression of the endogenous FSHβ gene. Figs. 5-7 illustrate examples of such constructs.

The DNA Construct

The DNA construct of the invention includes at least a targeting sequence and a regulatory sequence. It may additionally contain an exon; or an exon and an unpaired splice-donor site; or an exon, splice donor site, intron, and splice acceptor site. The exon, if present, is 3′ of the regulatory sequence, and the unpaired splice-donor site is at the 3′ end of the exon. The intron and splice acceptor site, if present, are 3′ of the splice donor site. In addition, there can be multiple exons and introns (with appropriate splice donor and acceptor sites) preceding (i.e., 5′ to) the exon flanked by the unpaired splice-donor site. The DNA in the construct is referred to as exogenous, since the DNA is not an original part of the genome of a host cell. Exogenous DNA may possess sequences identical to or different from portions of the endogenous genomic DNA present in the cell prior to transfection or infection by viral vector. As used herein, "transfection" means introduction of plasmid into a cell by chemical and physical means such as calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, electroporation, microinjection, microprojectiles, or biolistic-mediated uptake. As used herein "infection" means introduction of viral nucleic acid into a cell by virus infection. The various elements included in the DNA construct of the invention are described in detail below.

The DNA construct can also include *cis*-acting or *trans*-acting viral sequences (e.g., packaging signals), thereby enabling delivery of the construct into the nucleus of a cell via infection by a viral vector. Where necessary, the DNA construct can be disengaged from various steps of a virus life cycle, such as integrase-mediated integration in retroviruses or episome maintenance. Disengagement can be accomplished by appropriate deletions or mutations of viral sequences, such as a deletion of the integrase coding region in a retrovirus vector. Additional details regarding the construction and use of viral vectors are found in Robbins et al., Pharmacol. Ther. 80:35-47, 1998; and Gunzburg et al., Mol. Med. Today 1:410-417, 1995, herein incorporated by reference.

Targeting Sequences

Targeting sequences permit homologous recombination of a desired sequence into a selected site in the host genome. Targeting sequences are homologous to (i.e., able to homologously recombine with) their respective target sites in the host genome.

A circular DNA construct can employ a single targeting sequence, or two or more separate targeting sequences. A linear DNA construct may contain two or more separate targeting sequences. The target site to which a given targeting sequence is homologous can reside within an exon and/or intron of the FSHβ gene, upstream of and immediately adjacent to the FSHβ-coding region, or upstream of and at a distance from the FSHβ-coding region.

The first of the two targeting sequences in the construct (or the entire targeting sequence, if there is only one targeting sequence in the construct) is at least in part derived from the newly disclosed genomic regions upstream of the FSHβ-coding sequences. This targeting sequence contains a portion of SEQ ID NO:1, e.g., at least 20 consecutive nucleotides from the sequence cor-

responding to positions -7,454 to -1,417 (SEQ ID NO:4) or to positions -696 to -155 (SEQ ID NO:5). The second of the two targeting sequences in the construct may target a genomic region upstream of the coding sequence (e.g., also contain a portion of SEQ ID NO:4 or 5), or target an exon or intron of the gene.

The targeting sequence(s) may additionally include sequence derived from a previously known region of the FSHβ gene, including those described herein, as well as regions further upstream which are structurally uncharacterized but can be mapped by one skilled in the art.

Genomic fragments that can be used as targeting sequences can be identified by their ability to hybridize to a probe containing all or a portion of SEQ ID NO:4 or 5. Such a probe can be generated by PCR using primers derived from SEQ ID NO:1.

The Regulatory Sequence

The regulatory sequence of the DNA construct can contain one or more promoters (e.g., a constitutive, tissue-specific, or inducible promoter), enhancers, scaffold-attachment regions or matrix attachment sites, negative regulatory elements, transcription factor binding sites, or combinations of these elements.

The regulatory sequence can be derived from a eukaryotic (e.g., mammalian) or viral genome. Useful regulatory sequences include, but are not limited to, those that regulate the expression of SV40 early or late genes, cytomegalovirus genes, and adenovirus major late genes. They also include regulatory regions derived from genes encoding mouse metallothionein-I, elongation factor-1α, collagen (e.g., collagen Iα1, collagen Iα2, and collagen IV), actin (e.g., γ-actin), immunoglobulin, HMG-CoA reductase, glyceraldehyde phosphate dehydrogenase, 3-phosphoglyceratekinase, collagenase, stromelysin, fibronectin, vimentin, plasminogen activator inhibitor I, thym-

osin β4, tissue inhibitors of metalloproteinase, ribosomal proteins, major histocompatibility complex molecules, and human leukocyte antigens.

The regulatory sequence preferably contains transcription factor binding site, such as a TATA Box, CCAAT Box, AP1, Sp1, or a NF-κB binding site.

Marker Genes

If desired, the construct can include a sequence encoding a desired polypeptide, operatively linked to its own promoter. An example of this would be a selectable marker gene, which can be used to facilitate the identification of a targeting event. An amplifiable marker gene can also be used to facilitate selection of cells having co-amplified flanking DNA sequences. Cells containing amplified copies of the amplifiable marker gene can be identified by growth in the presence of an agent that selects for the expression of the amplifiable gene. The activated endogenous gene will typically be amplified in tandem with the amplified selectable marker gene. Cells containing multiple copies of the activated endogenous gene may produce very high levels of FSHβ and are thus useful for *in vitro* protein production and gene therapy.

The selectable and amplifiable marker genes do not have to lie immediately adjacent to each other. The amplifiable marker gene and selectable marker gene can be the same gene. One or both of the marker genes can be situated in the intron of the DNA construct. Suitable amplifiable marker genes and selectable marker genes are described in U.S. Patent No. 5,641,670.

The Exogenous Exon

The DNA construct may further contain an exon, i.e., a DNA sequence that is copied into RNA and is present in a mature mRNA molecule. The exon in the construct is referred to herein as an exogenous or construct-deriv-

ed exon. The exogenous exon can be non-coding, like the first exon of the human FSHβ gene, and in fact can optionally be identical in sequence to the latter exon. Alternatively, the exogenous exon encodes one or more amino acid residues, or partially encodes an amino acid residue (i.e., contains one or two nucleotides of a codon). When the exon contains a coding sequence, the DNA construct should be designed such that, upon transcription and splicing, the reading frame of the resulting mRNA is in-frame with the coding region of the target FSHβ gene. That is, the exogenous exon is spliced to an endogenous exon in a manner that does not change the appropriate reading frame of the portion of the mRNA derived from the endogenous exon.

The inclusion of a coding exon in the DNA construct allows the production of a fusion protein that contains both endogenous FSHβ protein sequence and exogenous protein sequence. Such a hybrid protein may combine the structural, enzymatic, or ligand- or receptor-binding properties from two or more proteins into one polypeptide. For example, the exogenous exon can encode a cell membrane anchor, a signal peptide to improve cellular secretion, a leader sequence, an enzymatic region, a cofactor binding region, or an epitope tag to facilitate purification of the FSHβ hybrid protein produced from the recombined gene locus.

The Splice-Donor Site

The exogenous exon is flanked at its 3′ end by a splice-donor site. A splice-donor site is a sequence which directs the splicing of one exon of an RNA transcript to the splice-acceptor site of another exon of the RNA transcript. Typically, the first exon lies 5′ of the second exon, and the splice-donor site located at the 3′ end of the first exon is paired with a splice-acceptor site on the 5′ side of the second exon. Splice-donor

sites have a characteristic consensus sequence represented as (A/C)AGGURAGU (where R denotes a purine), with the GU in the fourth and fifth positions being required (Jackson, Nucleic Acids Research 19:3715-3798, 1991). The first three bases of the splice-donor consensus site are the last three bases of the exon: i.e., they are not spliced out. Splice-donor sites are functionally defined by their ability to effect the appropriate reaction within the mRNA splicing pathway.

By way of example, the splice-donor site can be placed immediately adjacent and 3' to an ATG codon when the presence of one or more intervening nucleotides is not required for the exogenous exon to be in-frame with the second exon of the targeted gene. When the exogenous exon encodes one or more amino acids in-frame with the coding sequence of the targeted gene, the splice-donor site may preferably be placed immediately adjacent to the exogenous coding sequence on its 3' side.

The splice-donor site flanking the exogenous exon is unpaired in the construct, i.e., in the construct itself there is no accompanying splice-acceptor site downstream of the splice-donor site to which the latter can be spliced. Following homologous recombination into the target site upstream of the FSHβ coding sequence, what was the construct's unpaired splice-donor site is functionally paired with an endogenous splice-acceptor site of an endogenous exon of FSHβ. Processing of the transcript produced from the homologously recombined FSHβ gene results in splicing of the exogenous exon to the splice-acceptor site of an endogenous exon.

The construct of the invention can also include a splice-acceptor site. This site, in conjunction with a splice-donor site, directs the splicing of one exon to another exon. Splice-acceptor sites have a characteristic sequence represented as $(Y)_{10}$NYAG (SEQ ID NO:7), where

Y denotes any pyrimidine and N denotes any nucleotide (Jackson, Nucleic Acids Research 19:3715-3798, 1991).

Introns

The DNA construct may optionally contain an intron. An intron is a sequence of one or more nucleotides lying between a splice-donor site and a splice-acceptor site, and is removed, by splicing, from a precursor RNA molecule in the formation of a mature mRNA molecule.

The CAP Site

The DNA construct can optionally contain a CAP site. A CAP site is a specific transcription start site which is associated with and utilized by the regulatory region. This CAP site is located at a position relative to the regulatory sequence in the construct such that following homologous recombination, the regulatory sequence directs synthesis of a transcript that begins at the CAP site. Alternatively, no CAP site is included in the construct, and the transcriptional apparatus will locate by default an appropriate site in the targeted gene to be utilized as a CAP site.

Additional DNA elements

The construct may additionally contain sequences which affect the structure or stability of the RNA or protein produced by homologous recombination. Optionally, the DNA construct can include a bacterial origin of replication and bacterial antibiotic resistance markers or other selectable markers, which allow for large-scale plasmid propagation in bacteria or any other suitable cloning/host system.

All of the above-described elements of the DNA construct are operatively linked or functionally placed with respect to each other. That is, upon homologous recombination between the construct and the targeted genomic DNA, the regulatory sequence can direct the production of a primary RNA transcript which initiates at a

CAP site (optionally included in the construct) and includes (i) sequence corresponding to the exon and splice-donor site of the construct, if they are present, and (ii) sequence lying between that splice-donor site and the endogenous gene's transcription stop site. The latter sequence may include the FSHβ gene's endogenous regulatory region as well as sequences neighboring that region that are normally not transcribed. In an operatively linked configuration, the splice-donor site of the targeting construct directs a splicing event to a splice-acceptor site flanking one of the exons of the endogenous FSHβ gene, such that a desired protein can be produced from the fully spliced mature transcript. The splice-acceptor site can be endogenous, such that the splicing event is directed to an endogenous exon. In another embodiment where the splice-acceptor site is included in the targeting construct, the splicing event removes the exogenous intron introduced by the targeting construct.

The order of elements in the DNA construct can vary. Where the construct is a circular plasmid or viral vector, the relative order of elements in the resulting structure can be, for example: a targeting sequence, plasmid DNA (comprised of sequences used for the selection and/or replication of the targeting plasmid in a microbial or other suitable host), selectable marker(s), a regulatory sequence, an exon, and an unpaired splice-donor site.

Where the construct is linear, the order can be, for example: a first targeting sequence, a selectable marker gene, a regulatory sequence, an exon, a splice-donor site, and a second targeting sequence; or, in the alternative, a first targeting sequence, a regulatory sequence, an exon, a splice-donor site, a selectable marker gene, and a second targeting sequence. The order of the elements can also be: a first targeting sequence,

a selectable marker, a regulatory sequence, an exon, a splice-donor site, an intron, a splice-acceptor site, optionally an internal ribosomal entry site, and second targeting sequence.

Alternatively, the order can be: a first targeting sequence, a first selectable marker gene, a regulatory sequence, an exon, a splice-donor site, a second targeting sequence, and a second selectable marker gene; or, a first targeting sequence, a regulatory sequence, an exon, a splice-donor site, a first selectable marker gene, a second targeting sequence, and a second selectable marker gene. Recombination between the targeting sequences flanking the first selectable marker with homologous sequences in the host genome results in the targeted integration of the first selectable marker, while the second selectable marker is not integrated. Desired transfected or infected cells are those that are stably transfected or infected with the first, but not second, selectable marker. Such cells can be selected for by growth in a medium containing an agent which selects for expression of the first marker and another agent which selects against the second marker. Transfected or infected cells that have improperly integrated the targeting construct by a mechanism other than homologous recombination would be expected to express the second marker gene and will thereby be killed in the medium.

A positively selectable marker is sometimes included in the construct to allow for the selection of cells containing amplified copies of that marker. In this embodiment, the order of construct components can be, for example: a first targeting sequence, an amplifiable positively selectable marker, a second selectable marker (optional), a regulatory sequence, an exon, a splice-donor site, and a second targeting DNA sequence.

The various elements of the construct can be obtained from natural sources (e.g., genomic DNA), or can be produced using genetic engineering techniques or synthetic processes. The regulatory region, CAP site, exon, splice-donor site, and optional intron and splice acceptor site of the construct can be isolated as a complete unit from, e.g., the human elongation factor-1α (Genbank sequence HUMEF1A) gene or the cytomegalovirus (Genbank sequence HEHCMVP1) immediate early region. These components can also be isolated from separate genes.

Transfection or Infection and Homologous Recombination

The DNA construct of the invention can be introduced into the cell, such as a primary, secondary, or immortalized cell, as a single DNA construct, or as separate DNA sequences which become incorporated into the chromosomal or nuclear DNA of a transfected or infected cell. The DNA can be introduced as a linear, double-stranded (with or without single-stranded regions at one or both ends), single-stranded, or circular molecule. The DNA construct or its RNA equivalent can also be introduced as a viral nucleic acid.

When the construct is introduced into host cells in two separate DNA fragments, the two fragments share DNA sequence homology (overlap) at the 3′ end of one fragment and the 5′ end of the other, while one carries a first targeting sequence and the other carries a second targeting sequence. Upon introduction into a cell, the two fragments can undergo homologous recombination to form a single molecule with the first and second targeting sequences flanking the region of overlap between the two original fragments. The product molecule is then in a form suitable for homologous recombination with the cellular target sites. More than two fragments can be used, with each of them designed such that they will undergo homologous recombination with each other to ul-

timately form a product suitable for homologous recombination with the cellular target sites as described above.

The DNA construct of the invention, if not containing a selectable marker itself, can be co-transfected or co-infected with another construct that contains such a marker. A targeting plasmid may be cleaved with a restriction enzyme at one or more sites to create a linear or gapped molecule prior to transfection or infection. The resulting free DNA ends increase the frequency of the desired homologous recombination event. In addition, the free DNA ends may be treated with an exonuclease to create overhanging 5' or 3' single-stranded DNA ends (e.g., at least 30 nucleotides in length, and preferably 100-1000 nucleotides in length) to increase the frequency of the desired homologous recombination event. In this embodiment, homologous recombination between the targeting sequence and the genomic target will result in two copies of the targeting sequences, flanking the elements contained within the introduced plasmid.

The DNA constructs may be transfected into cells (preferably *in vitro*) by a variety of physical or chemical methods, including electroporation, microinjection, microprojectile bombardment, calcium phosphate precipitation, liposome delivery, or polybrene- or DEAE dextran-mediated transfection.

The transfected or infected cell is maintained under conditions which permit homologous recombination, as described in the art (see, e.g., Capecchi, Science 24:1288-1292, 1989). By "transfected cell" is meant a cell into which (or into an ancestor of which) a DNA molecule has been introduced by a means other than using a viral vector. By "infected cell" is meant a cell into which (or into an ancestor of which) a DNA or RNA molecule has been introduced using a viral vector. Viruses

known to be useful as vectors include adenovirus, adeno-associated virus, Herpes virus, mumps virus, poliovirus, lentivirus, retroviruses, Sindbis virus, and vaccinia viruses such as canary pox virus. When the homologously recombinant cell is maintained under conditions sufficient to permit transcription of the DNA, the regulatory region introduced by the DNA construct will alter transcription of the FSHβ gene.

Homologously recombinant cells (i.e., cells that have undergone the desired homologous recombination) can be identified by phenotypic screening or by analyzing the culture supernatant in enzyme-linked immunosorbent assays (ELISA) for FSHβ. Commercial ELISA kits for detecting FSHβ are available from Accurate Chemical and Scientific (Westbury, NY). Homologously recombinant cells can also be identified by Southern and Northern analyses or by polymerase chain reaction (PCR) screening.

As used herein, the term "primary cells" includes (i) cells present in a suspension of cells isolated from a vertebrate tissue source (prior to their being plated, i.e., attached to a tissue culture substrate such as a dish or flask), (ii) cells present in an explant derived from tissue, (iii) cells plated for the first time, and (iv) cell suspensions derived from these plated cells. Primary cells can also be cells as they naturally occur within a human or an animal.

Secondary cells are cells at all subsequent steps in culturing. That is, the first time that plated primary cells are removed from the culture substrate and replated (passaged), they are referred to herein as secondary cells, as are all cells in subsequent passages. Secondary cell strains consist of secondary cells which have been passaged one or more times. Secondary cells typically exhibit a finite number of mean population doublings in culture and the property of con-

tact-inhibited, anchorage-dependent growth (anchorage-dependence does not apply to cells that are propagated in suspension culture). Primary and secondary cells are not immortalized.

Immortalized cells are cell lines (as opposed to cell strains, with the designation "strain" reserved for primary and secondary cells) that exhibit an apparently unlimited lifespan in culture.

Cells selected for transfection or infection can fall into four types or categories: (i) cells which do not, as obtained, make or contain more than trace amounts of the FSHβ protein, (ii) cells which make or contain the protein but in quantities other than those desired (such as, in quantities less than the level which is physiologically normal for the type of cells as obtained), (iii) cells which make the protein at a level which is physiologically normal for the type of cells as obtained, but are to be augmented or enhanced in their content or production, and (iv) cells in which it is desirable to change the pattern of regulation or induction of a gene encoding the protein.

Primary, secondary and immortalized cells to be transfected or infected by the present method can be obtained from a variety of tissues and include all appropriate cell types which can be maintained in culture. For example, suitable primary and secondary cells include fibroblasts, keratinocytes, epithelial cells (e.g., mammary epithelial cells, intestinal epithelial cells), endothelial cells, glial cells, neural cells, formed elements of the blood (e.g., lymphocytes, bone marrow cells), muscle cells, and precursors of these somatic cell types. Where the homologously recombinant cells are to be used in gene therapy, primary cells are preferably obtained from the individual to whom the transfected or infected primary or secondary cells are to be adminis-

tered. However, primary cells can be obtained from a donor (i.e., an individual other than the recipient) of the same species.

Examples of immortalized human cell lines useful for protein production or gene therapy include, but are not limited to, 2780AD ovarian carcinoma cells (Van der Blick et al., Cancer Res., 48:5927-5932, 1988), A549 (American Type Culture Collection ("ATCC") CCL 185), BeWo (ATCC CCL 98), Bowes Melanoma cells (ATCC CRL 9607), CCRF-CEM (ATCC CCL 119), CCRF-HSB-2 (ATCC CCL 120.1), COLO201 (ATCC CCL 224), COLO205 (ATCC CCL 222), COLO 320DM (ATCC CCL 220), COLO 320HSR (ATCC CCL 220.1), Daudi cells (ATCC CCL 213), Detroit 562 (ATCC CCL 138), HeLa cells and derivatives of HeLa cells (ATCC CCL 2, 2.1 and 2.2), HCT116 (ATCC CCL 247), HL-60 cells (ATCC CCL 240), HT1080 cells (ATCC CCL 121), IMR-32 (ATCC CCL 127), Jurkat cells (ATCC TIB 152), K-562 leukemia cells (ATCC CCL 243), KB carcinoma cells (ATCC CCL 17), KG-1 (ATCC CCL 246), KG-1a (ATCC CCL 246.1), LS123 (ATCC CCL 255), LS17-4T (ATCC CCL CL-188), LS180 (ATCC CCL CL-187), MCF-7 breast cancer cells (ATCC BTH 22), MOLT-4 cells (ATCC CRL 1582), Namalwa cells (ATCC CRL 1432), NCI-H498 (ATCC CCL 254), NCI-H508 (ATCC CCL 253), NCI-H548 (ATCC CCL 249), NCI-H716 (ATCC CCL 251), NCI-H747 (ATCC CCL 252), NCI-H1-688 (ATCC CCL 257), NCI-H2126 (ATCC CCL 256), Raji cells (ATCC CCL 86), RD (ATCC CCL 136), RPMI 2650 (ATCC CCL 30), RPMI 8226 cells (ATCC CCL 155), SNU-C2A (ATCC CCL 250.1), SNU-C2B (ATCC CCL 250), SW-13 (ATCC CCL 105), SW48 (ATCC CCL 231), SW403 (ATCC CCL 230), SW480 (ATCC CCL 227), SW620 (ATCC CCL 227), SW837 (ATCC CCL 235), SW948 (ATCC CCL 237), SW1116 (ATCC CCL 233), SW1417 (ATCC CCL 238), SW1463 (ATCC CCL 234), T84 (ATCC CCL 248), U-937 cells (ATCC CRL 1593), WiDr (ATCC CCL 218), and WI-38VA13 subline 2R4 cells (ATCC CLL 75.1), as well as heterohybridoma cells produced by fusion of human cells

and cells of another species. Secondary human fibroblast strains, such as WI-38 (ATCC CCL 75) and MRC-5 (ATCC CCL 171), may be used. In addition, primary, secondary, or immortalized human cells, as well as primary, secondary, or immortalized cells from other species, can be used for *in vitro* protein production or gene therapy.

FSHβ-expressing Cells

Homologously recombinant cells of the invention express FSHβ at desired levels and are useful for *in vitro* production of FSHβ and gene therapy.

Protein Production

Homologously recombinant cells according to this invention can be used for *in vitro* production of FSHß. The cells are maintained under conditions, as described in the art, which result in expression of proteins. The FSHß protein may be purified from cell lysates or cell supernatants. A pharmaceutical composition containing the FSHß protein can be delivered to a human or an animal by conventional pharmaceutical routes known in the art (e.g., oral, intravenous, intramuscular, intranasal, pulmonary, transmucosal, intradermal, transdermal, rectal, intrathecal, subcutaneous, intraperitoneal, or intralesional). Oral administration may require use of a strategy for protecting the protein from degradation in the gastrointestinal tract: e.g., by encapsulation in polymeric microcapsules.

Gene Therapy

Homologously recombinant cells of the present invention are useful as populations of homologously recombinant cell lines, as populations of homologously recombinant primary or secondary cells, as homologously recombinant clonal cell strains or lines, as homologously recombinant heterogenous cell strains or lines, and as cell mixtures in which at least one representative cell of one of the four preceding categories of homologously

recombinant cells is present. Such cells may be used in a delivery system for treating infertility, for enhancing fertility in a human or animal, or for treating any other conditions treatable with FSHß.

Homologously recombinant primary cells, clonal cell strains or heterogenous cell strains are administered to an individual in whom the abnormal or undesirable condition is to be treated or prevented, in sufficient quantity and by an appropriate route, to express or make available the protein or exogenous DNA at physiologically relevant levels. A physiologically relevant level is one which either approximates the level at which the product is normally produced in the body or results in improvement of the abnormal or undesirable condition. If the cells are syngeneic with respect to a immunocompetent recipient, the cells can be administered or implanted intravenously, intraarterially, subcutaneously, intraperitoneally, intraomentally, subrenal capsularly, intrathecally, intracranially, or intramuscularly.

If the cells are not syngeneic and the recipient is immunocompetent, the homologously recombinant cells to be administered can be enclosed in one or more semipermeable barrier devices. The permeability properties of the device are such that the cells are prevented from leaving the device upon implantation into a subject, but the therapeutic protein is freely permeable and can leave the barrier device and enter the local space surrounding the implant or enter the systemic circulation. See, e.g., U.S. Patent Nos. 5,641,670, 5,470,731, 5,620,883, 5,487,737, and co-owned U.S. Patent Application entitled "Delivery of Therapeutic Proteins" (inventors: Justin C. Lamsa and Douglas A. Treco), filed April 16, 1999, all herein incorporated by reference. The barrier device can be implanted at any appropriate site: e.g., intraperito-

neally, intrathecally, subcutaneously, intramuscularly, within the kidney capsule, or within the omentum.

Barrier devices are particularly useful and allow homologously recombinant immortalized cells, homologously recombinant cells from another species (homologously recombinant xenogeneic cells), or cells from a nonhistocompatibility-matched donor (homologously recombinant allogeneic cells) to be implanted for treatment of a subject. The devices retain cells in a fixed position *in vivo*, while protecting the cells from the host's immune system. Barrier devices also allow convenient short-term (i.e., transient) therapy by allowing ready removal of the cells when the treatment regimen is to be halted for any reason. Transfected or infected xenogeneic and allogeneic cells may also be used in the absence of barrier devices for short-term gene therapy. In that case, the FSHß produced by the cells will be delivered *in vivo* until the cells are rejected by the host's immune system.

A number of synthetic, semisynthetic, or natural filtration membranes can be used for this purpose, including, but not limited to, cellulose, cellulose acetate, nitrocellulose, polysulfone, polyvinylidene difluoride, polyvinyl chloride polymers and polymers of polyvinyl chloride derivatives. Barrier devices can be utilized to allow primary, secondary, or immortalized cells from another species to be used for gene therapy in humans.

Another type of device useful in the gene therapy of the invention is an implantable collagen matrix in which the cells are embedded. Such a device, which can contain beads to which the cells attach, is described in WO 97/15195, herein incorporated by reference.

The number of cells needed for a given dose or implantation depends on several factors, including the expression level of the protein, the size and condition of the host animal, and the limitations associated with

the implantation procedure. Usually the number of cells implanted in an adult human or other similarly-sized animal is in the range of 1×10^4 to 5×10^{10}, and preferably 1×10^8 to 1×10^9. If desired, they may be implanted at multiple sites in the patient, either at one time or over a period of months or years. The dosage may be repeated as needed.

Other Embodiments

It is to be understood that while the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not to limit the scope of the invention, which is defined by the scope of the appended claims.

Other aspects, advantages, and modifications are within the scope of the following claims.

What is claimed is:

1. A DNA construct that alters expression of an endogenous FSHβ gene in a mammalian cell upon integration into the genome of the cell via homologous recombination, the construct comprising a targeting sequence containing at least 20 contiguous nucleotides from SEQ ID NO:4 or 5 and a transcriptional regulatory sequence.

2. The DNA construct of claim 1, wherein the construct further comprises an exon and a splice-donor site.

3. The DNA construct of claim 2, wherein the construct further comprises, downstream from the splice-donor site, an intron and a splice-acceptor site.

4. The DNA construct of claim 1, wherein the construct further comprises a selectable marker gene.

5. The DNA construct of claim 1, wherein the target sequence contains at least 50 contiguous nucleotides from SEQ ID NO:4 or 5.

6. An isolated nucleic acid comprising at least 20 contiguous nucleotides of SEQ ID NO:4 or its complement.

7. The isolated nucleic acid of claim 6, wherein the isolated nucleic acid comprises at least 50 contiguous nucleotides of SEQ ID NO:4 or its complement.

8. The isolated nucleic acid of claim 6, wherein the isolated nucleic acid comprises at least 100 contiguous nucleotides of SEQ ID NO:4 or its complement.

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9. The isolated nucleic acid of claim 6, wherein the isolated nucleic acid comprises at least 200 contiguous nucleotides of SEQ ID NO:4 or its complement.

10. The isolated nucleic acid of claim 6, wherein the isolated nucleic acid comprises at least 500 contiguous nucleotides of SEQ ID NO:4 or its complement.

11. The isolated nucleic acid of claim 6, wherein the isolated nucleic acid comprises at least 1000 contiguous nucleotides of SEQ ID NO:4 or its complement.

12. The isolated nucleic acid of claim 6, wherein the isolated nucleic acid comprises SEQ ID NO:4 or its complement.

13. An isolated nucleic acid comprising at least 20 contiguous nucleotides of SEQ ID NO:5 or its complement.

14. The isolated nucleic acid of claim 13, wherein the isolated nucleic acid comprises at least 50 contiguous nucleotides of SEQ ID NO:5 or its complement.

15. The isolated nucleic acid of claim 13, wherein the isolated nucleic acid comprises at least 100 contiguous nucleotides of SEQ ID NO:5 or its complement.

16. The isolated nucleic acid of claim 13, wherein the isolated nucleic acid comprises at least 200 contiguous nucleotides of SEQ ID NO:5 or its complement.

17. The isolated nucleic acid of claim 13, wherein the isolated nucleic acid comprises SEQ ID NO:5 or its complement.

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18. An isolated DNA comprising a strand that comprises a nucleotide sequence that (i) is at least 100 nucleotides in length and (ii) hybridizes under highly stringent conditions with SEQ ID NO:4 or 5, or the complement of SEQ ID NO:4 or 5.

19. The isolated DNA of claim 18, wherein the nucleotide sequence is at least 200 nucleotides in length.

20. The isolated DNA of claim 18, wherein the nucleotide sequence is at least 400 nucleotides in length.

21. The isolated DNA of claim 18, wherein the nucleotide sequence is at least 1,000 nucleotides in length.

22. An isolated DNA comprising a strand that comprises a nucleotide sequence that (i) is at least 100 nucleotides in length and (ii) shares at least 80% sequence identity with a fragment of SEQ ID NO:4 or 5 having the same length as the nucleotide sequence.

23. The isolated DNA of claim 22, wherein the nucleotide sequence is at least 200 nucleotides in length.

24. The isolated DNA of claim 22, wherein the nucleotide sequence is at least 400 nucleotides in length.

25. The isolated DNA of claim 22, wherein the nucleotide sequence is at least 1,000 nucleotides in length.

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26. A homologously recombinant cell stably transfected with the DNA construct of claim 1, the DNA construct having undergone homologous recombination with genomic DNA upstream of the ATG initiation codon of an endogenous FSHβ coding sequence.

27. A homologously recombinant cell stably transfected with the DNA construct of claim 2, the DNA construct having undergone homologous recombination with genomic DNA upstream of the ATG initiation codon of an endogenous FSHβ coding sequence.

28. A homologously recombinant cell stably transfected with the DNA construct of claim 3, the DNA construct having undergone homologous recombination with genomic DNA upstream of the ATG initiation codon of an endogenous FSHβ coding sequence.

29. A homologously recombinant cell stably transfected with the DNA construct of claim 4, the DNA construct having undergone homologous recombination with genomic DNA upstream of the ATG initiation codon of an endogenous FSHβ coding sequence.

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30. A method of altering expression of an endogenous FSHβ gene in a mammalian cell, the method comprising

introducing the DNA construct of claim 1 into the cell;

maintaining the cell under conditions which permit homologous recombination to occur between the construct and a genomic target site homologous to the targeting sequence, to produce a homologously recombinant cell; and

maintaining the homologously recombinant cell under conditions which permit expression of the FSHβ coding sequence under the control of the transcriptional regulatory sequence.

31. A method of delivering FSHß to an animal, comprising

providing the cell of claim 26, and

implanting the cell in the animal, wherein the cell secretes FSHß.

32. A method of delivering FSHß to an animal, comprising

providing the cell of claim 27, and

implanting the cell in the animal, wherein the cell secretes FSHß.

33. A method of delivering FSHß to an animal, comprising

providing the cell of claim 28, and

implanting the cell in the animal, wherein the cell secretes FSHß.

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34. A method of delivering FSHß to an animal, comprising

providing the cell of claim 29, and

implanting the cell in the animal, wherein the cell secretes FSHß.

35. A method of producing FSHß, comprising

providing the cell of claim 26, and

culturing the cell *in vitro* under conditions which permit the cell to express and secrete FSHß.

36. A method of producing FSHß, comprising

providing the cell of claim 27, and

culturing the cell *in vitro* under conditions which permit the cell to express and secrete FSHß.

37. A method of producing FSHß, comprising

providing the cell of claim 28, and

culturing the cell *in vitro* under conditions which permit the cell to express and secrete FSHß.

38. A method of producing FSHß, comprising

providing the cell of claim 29, and

culturing the cell *in vitro* under conditions which permit the cell to express and secrete FSHß.

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