ENGINEERING THE PATHWAY FOR SUCCINATE PRODUCTION

CROSS-REFERENCE TO RELATED APPLICATIONS

(001) This application claims the priority of the U.S. Provisional Application Serial No. 61/166,093, filed on April 2, 2009. This application is also a continuation-in-part of U.S. Application Serial No. 12/529,826, filed September 3, 2009, which is the U.S. national stage application of PCT/US2008/057439, filed March 19, 2009, which claims the benefit of U.S. Provisional Application Serial No. 60/895,806, filed March 20, 2007, the disclosure of each of which is hereby incorporated by reference in its entirety, including all figures, tables and amino acid or nucleic acid sequences.

GOVERNMENT SUPPORT

(002) This invention was made with government support under a grant awarded from the Department of Energy under grant number USDOE-DE FG02-96ER20222 and Department of Energy in conjunction with the United States Department of Agriculture under grant number USDA & DOE Biomass RDI DE FG36-04GO14019. The government has certain rights in the invention.

BACKGROUND OF THE INVENTION

(003) The present invention is in the field of production of succinic acid from renewable biological feedstocks using microbial biocatalysts. This invention discloses the genetic modifications to the biocatalysts that are useful in achieving high efficiency for succinic acid production. More specifically, this invention provides genetically modified biocatalysts that are suitable for the production of succinic acid from renewable feedstocks in commercially significant quantities.

(004) A 2004 U.S. Department of Energy report entitled "Top value added chemicals from biomass" has identified twelve building block chemicals that can be produced from renewable feed stocks. The twelve sugar-based building blocks are 1,4-diacids (succinic, fumaric and maleic), 2,5-furan dicarboxylic acid, 3-hydroxy propionic acid, aspartic acid, glucaric acid, glutamic acid, itaconic acid, levulinic acid, 3-hydroxybutyrolactone, glycerol, sorbitol, and xylitol/arabinitol. The fermentative production of these building block chemicals from renewable feedstocks will become increasingly competitive as petroleum prices increase.

(005) These building block chemicals are molecules with multiple functional groups that possess the potential to be transformed into new families of useful molecules. These twelve building block chemicals can be subsequently converted to a number of high-value bio-based chemicals or materials. For example, succinate can serve as a substrate for transformation into plastics, solvents, and other chemicals currently made from petroleum (Lee *et al.*, 2004; Lee *et al.*, 2005; McKinlay *et al.*, 2007; Wendisch *et al.*, 2006; Zeikus *et al.*, 1999). Many bacteria have been described with the natural ability to produce succinate as a major fermentation product (Table 1). However, complex processes, expensive growth media and long incubation times are often required to produce succinic acid from these naturally occurring succinic acid producing microorganisms.

(006) A variety of genetic approaches have previously been used to engineer *Escherichia coli* strains for succinate production with varying degrees of success (Table 1). In most studies, titers achieved were low and complex medium ingredients such as yeast extract or corn steep liquor was required. *E. coli* strain NZN111 produced 108 mM succinate with a molar yield of 0.98 mol succinate per mol of metabolized glucose (Chatterjee *et al.*, 2001; Millard *et al.*, 1996; Stols and Donnelly, 1997). This strain was engineered by inactivating two genes (*pflB* encoding pyruvate-formate lyase and *ldhA* encoding lactate dehydrogenase), and over-expressing two *E. coli* genes, malate dehydrogenase (*mdh*) and phosphoenol pyruvate carboxylase (*ppc*), from multicopy plasmids. *E. coli* strain HL27659k was engineered by mutating succinate dehydrogenase (*sdhAB*), phosphate acetyltransferase (*pta*), acetate kinase (*ackA*), pyruvate oxidase (*poxB*), glucose transporter (*ptsG*), and the isocitrate lyase repressor (*iclR*). This strain produced less than 100 mM succinate and required oxygen-limited fermentation conditions (Cox *et al.*, 2006; Lin *et al.*, 2005a, 2005b, 2005c; Yun *et al.*, 2005). Analysis of metabolism *in silico* has been used to design gene knockouts to create a pathway in *E. coli* that is analogous to the native succinate pathway in *Mannheimia succiniciproducens* (Lee *et al.*, 2005 and 2006). The resulting strain, however, produced very little succinate. Andersson *et al.* (2007) have reported the highest levels of succinate production by engineered *E. coli* (339 mM) containing only native genes.

(007) Other researchers have pursued alternative approaches that express heterologous genes in *E. coli*. The *Rhizobium eteloti* pyruvate carboxylase (*pyc*) was over-expressed from a multicopy plasmid to direct carbon flow to succinate. (Gokarn *et al.*, 2000; Vemuri *et al.*, 2002a, 2002b). Strain SBS550MG was constructed by inactivating the isocitrate lyase repressor (*iclR*), *adhE*, *ldhA*, and *ackA*, and over-expressing the *Bacillus subtilis citZ* (citrate synthase) and *R. etli pyc* from a multi-copy plasmid (Sanchez *et al.*, 2005a). With this strain, 160 mM succinate was produced from glucose with a molar yield of 1.6.

(008) More complex processes have also been investigated for succinate production (Table 1). Many of these processes include an aerobic growth phase followed by an anaerobic production phase. The anaerobic phase is often supplied with carbon dioxide, hydrogen, or both (Andersson *et al.*, 2007; Sanchez *et al.*, 2005a and 2005b; Sanchez *et al.*, 2006; U.S. Patent 5,869,301; Vemuri *et al.*, 2002a and 2002b). In a recent study with a native succinate producer, *A. succiniciproducens*, electro dialysis, sparging with CO_2, cell recycle, and batch feeding were combined for the production of succinic acid from glucose at high yield, titer and productivity (Meynial-Salles *et al.*, 2007).

(009) The majority by far of scientific knowledge of *E. coli* is derived from investigations in complex medium such as Luria broth rather than mineral salts medium using low concentrations of sugar substrates (typically 0.2% w/v; 11 mM) rather than the 5% (w/v) glucose (278 mM) and 10% (w/v) glucose (555 mM) used in the studies reported herein. Large amounts of sugar are required to produce commercially significant levels of product. Previous researchers have described the construction of many *E. coli* derivatives for succinate production in complex medium (Table 1). With complex medium, rational design based on primary pathways has been reasonably successful for academic demonstrations of metabolic engineering. However, the use of complex nutrients for production of bacterial fermentation products increases the cost of materials, the cost of purification, and the cost associated with waste disposal. Use of mineral salts medium without complex media components should be much more cost-effective.

(010) *E. coli* C grows well in NBS mineral salts medium containing glucose and produces a mixture of lactate, acetate, ethanol and succinate as fermentation products (Fig. 1A; Table 4).

In contrast to other studies with *E. coli* (Table 1), the studies reported herein have focused on the development of strains that are able to convert high level of sugars into succinate using mineral salts medium to minimize the costs of materials, succinate purification, and waste disposal.

(011) One aspect of the invention provides various strains of *E. coli*, that produce succinate at high titers and yields in mineral salts media during simple, pH-controlled, batch fermentations without the need for heterologous genes or plasmids. The inventors have surprisingly identified a number of target genes useful in genetic manipulation of biocatalysts for achieving high efficiency for succinic acid production.

BRIEF SUMMARY

(012) It is an objective of the present embodiments disclosed herein to provide a method for obtaining a biocatalyst for manufacturing succinic acid using biological feedstocks. The method for obtaining the biocatalysts for succinic acid production from biological feedstocks combines rational genetic manipulations and the process of metabolic evolution.

(013) In generating a biocatalysts for succinic acid production from biological feedstocks, using a rational design derived from our existing knowledge about the microbial metabolic pathways, a set of genes within the bacterial chromosome are inactivated followed by the process of metabolic evolution to select a strain with desirable phenotype.

(013a) In one aspect, there is provided an *Escherichia coli* bacterial cell comprising increased levels of phosphoenol pyruvate carboxykinase (*pck*) gene transcripts and one or more genetic modifications in one or more genes selected from the group consisting of *ptsG*, *ptsH* and *ptsI*, said transcripts encoding active phosphoenol pyruvate carboxykinase (PCK), said cell exhibiting increased PCK activity and wherein said one or more genetic modifications inactivate the function of the gene product encoded by *ptsG*, *ptsH* and *ptsI* and is selected from deleting the entire coding region of the gene, deleting a portion of the coding region of the gene, introducing a frame shift mutation in the coding region of the gene, introducing a missense mutation in the gene, insertion or introducing a stop codon in the coding region of the gene or combinations thereof.

(013b) In another aspect, there is provided a genetically modified *Escherichia coli* bacterial cell, wherein said genetically modified bacterial cell is NRRL B-50279, NRRL B-50280, or NRRL B-50281.

(014) In one embodiment of the present disclosure, the mutation of the genes in the bacterial chromosome is accomplished without introducing any exogenous genetic material.

(015) In another embodiment of the present disclosure the mutation of the endogenous genes are accomplished either by introducing point mutation or by introducing a stop codon in the open reading frame of the endogenous gene.

(016) In another embodiment of the present disclosure the entire open reading frame of the endogenous gene is deleted from the chromosomal DNA.

(017) In certain preferred embodiments of the disclosure, the expression of certain endogenous genes is significantly increased. In one embodiment of the present disclosure, the transcription of the endogenous gene is increased by means of introducing certain mutations in the promoter region of the endogenous genes. In other embodiment of the disclosure, the transcription of the endogenous gene is enhanced by means of reliving the repressive control of the target gene.

(018) In certain embodiment of the present disclosure, an exogenous nucleotide sequence may be introduced to inactivate a target gene for the purpose of selecting a bacterial strain with a mutated gene with desirable phenotype. In the most preferred embodiment of the present disclosure, the exogenous nucleotide sequence introduced into the microbial genome is subsequently removed in a seamless fashion without leaving behind any residual exogenous nucleotide sequence.

(019) The rationally designed genetic manipulations can all be done in a single stage or in multiple stages in which a single genetic change is accomplished at one time. The microbial strain resulting from genetic manipulations may be subjected to metabolic evolution in order to improve the yield, titer and volumetric productivity of the desired organic acid. In the most preferred embodiment of the present disclosure, the rational genetic manipulations are done in stages and the process of metabolic evolution is carried out in between the stages of genetic manipulation.

(020) In one embodiment of the present disclosure, the spontaneous mutations that occur during the metabolic evolution are identified through sequencing appropriate regions of the chromosomal DNA. In yet another embodiment of the present disclosure, the mutations that occur during the metabolic evolution are identified by measuring the activities of suspect enzyme.

(021) In one embodiment of the present disclosure, based on rational designing, one or more of the genes coding for the proteins known to function in the fermentative pathways are inactivated through one or more mutations.

(022) It yet another embodiment of the present disclosure, the genes which are functional homologues of the genes coding for the proteins functioning in the fermentative pathway are inactivated beside the genes coding for the proteins directly involved in the fermentative pathway.

(023) In one embodiment of the present disclosure the genes functioning within the TCA cycle are genetically manipulated so that there is an increased flow of carbon towards succinic acid production. In one embodiment of the present disclosure, the carbon flow through reductive arm of the TCA cycle is enhanced. In another embodiment of the present disclosure, the carbon flow through oxidative arm of the TCA cycle is genetically manipulated. In another embodiment of the present disclosure, the carbon flow to succinic acid is improved through genetic manipulation of glyoxalate bypass pathway closely associated with TCA cycle.

(024) In yet another embodiment of the present disclosure, the carbon flow from TCA to other metabolic pathways within the cell is blocked so that the carbon pool within the cell is funneled towards the succinic acid production.

(025) In another embodiment of the present disclosure, the carbon flow into the TCA cycle is enhanced through genetic manipulation leading to one of more carboxylating enzymes within the cell. In one embodiment of the present disclosure, the expression of one or more carboxylating enzyme is achieved through the genetic manipulation of the promoter region or by relieving the repression of the gene expression.

(026) In another embodiment of the present disclosure, the phosphoenol pyruvate pool within the cell is conserved by mutating those genes involved in the carbon uptake pathway requiring phosphoenol pyruvate. In one embodiment of the present disclosure, the phosphotransferase system for carbon uptake is inactivated in order to conserve the phosphoenol pyruvate available for the operation of the TCA cycle.

(027) The present disclosure illustrates a number of targets that can be genetically manipulated to achieve an increased succinic acid production. All these various targets described in the present disclosure can be genetically manipulated to achieve an improved

succinic acid production. In the most preferred embodiment, a minimum number of targets are selected for genetic manipulation to achieve a desirable rate of succinic acid production.

(028) In the preferred embodiment of the present disclosure, the biocatalysts are selected for their ability to produce succinic acid at high titer, yield and volumetric productivity. In the most preferred embodiment of the present disclosure, a biocatalyst capable of producing at least 1.0 mole of succinic acid for every one mole of carbon source consumed is preferred.

(029) In another most preferred embodiment of the present disclosure, the biocatalyst is selected during metabolic evolution for its ability to produce at least 1.0 mole of succinic acid for every mole of carbon source consumed in a mineral salt medium and coupling the succinic acid production to microbial growth.

(030) In yet another embodiment of the present disclosure, biocatalysts capable of producing succinic acid using glycerol as a feed stock are provided.

(031) Additional advantage of this disclosure will become readily apparent from the ensuing detailed description of the disclosure and the examples provided.

BRIEF DESCRIPTION OF THE DRAWINGS

(032) **Figures 1A-1B.** Fermentation of glucose to succinate. **Fig. 1A** shows the standard pathway for fermentation of glucose by *E. coli*. This pathway has been redrawn from Unden and Kleefeld (2004). Bold arrows represent central fermentative pathways. Crosses represent the gene deletions performed in this study to engineer KJ012 (*ldhA, adhE, ackA*). Genes and enzymes: *ldhA*, lactate dehydrogenase; *pflB*, pyruvate-formate lyase; *focA*, formate transporter; *pta*, phosphate acetyltransferase; *ackA*, acetate kinase; *adhE*, alcohol dehydrogenase; *ppc*, phosphoenolpyruvate carboxylase; *pdh*, pyruvate dehydrogenase complex; *gltA*, citrate synthase; *mdh*, malate dehydrogenase; *fumA*, *fumB*, and *fumC*, fumarase isozymes; *frdABCD*, fumarate reductase; *fdh*, formate dehydrogenase; *icd*, isocitrate dehydrogenase; *acs*, acetyl~CoA synthetase; *mgsA*, methylglyoxal synthase; *poxB*, pyruvate oxidase; *aldA*, aldehyde dehydrogenase; and *aldB*, aldehyde dehydrogenase. **Fig. 1B** shows the coupling of ATP production and growth to succinate production in engineered strains of

E. coli for a standard pathway for glucose fermentation. Solid arrows connect NADH pools. Dotted arrows connect NAD^+ pools. During glycolysis under anaerobic conditions, growth is obligately coupled to the production of ATP and the oxidation of NADH.

(033) Figures 2A-2D. Potential carboxylation pathways for succinate production by *E. coli*. Genes encoding key carboxylating enzymes are shown in bold. **Fig. 2A** shows the reaction catalyzed by phosphoenolpyruvate (PEP) carboxylase enzyme (PPC). No ATP is produced from the carboxylation of phosphoenolpyruvate (PEP) by the PPC enzyme. This is regarded as the primary route for succinate production by *E. coli* during glucose fermentation. **Fig. 2B** shows the NADH-dependent malic enzyme. Energy is conserved during the production of ATP from ADP and PEP by pyruvate kinase (*pykA* or *pykF*). Malic enzyme (*sfcA*) catalyzes an NADH-linked, reductive carboxylation to produce malate. **Fig. 2C** shows the NADPH-dependent malic enzyme. Energy is conserved during the production of ATP from ADP and PEP by pyruvate kinase (*pykA* or *pykF*). Malic enzyme (*maeB*) catalyzes an NADPH-linked, reductive carboxylation to produce malate. **Fig. 2D** shows the reaction catalyzed by PEP carboxykinase (PCK). Energy is conserved by the production of ATP during the carboxylation of PEP to produce oxaloacetic acid.

(034) Figures 3A-3C. Growth during metabolic evolution of KJ012 to produce KJ017, KJ032, and KJ060. Strain KJ012 was sequentially transferred in NBS medium containing 5% (w/v) (Fig. 3A) and 10% (w/v) (Fig. 3B) glucose, respectively to produce KJ017. After deletion of *focA* and *pflB*, the resulting strain (KJ032) was initially subcultured in medium supplemented with acetate (Fig. 3C). Acetate levels were decreased and subsequently eliminated during further transfers to produce KJ060. Broken line represents fermentation by KJ017 without acetate, added for comparison. Symbol: • = optical density at OD_{550nm}.

(035) Figures 4A-4F. Summary of fermentation products during the metabolic evolution of strains for succinate production. Cultures were supplemented with sodium acetate as indicated. Black arrows represent the transition between fermentation conditions as indicated by text. No formate and only small amounts of lactate were detected during metabolic evolution of KJ032. No formate and lactate were detected during metabolic evolution of KJ070 and KJ072. Metabolic evolution of KJ012 to KJ017 in the medium containing 5% w/v

glucose (Fig 4A) and in the medium containing 10% w/v glucose (Fig. 4B). Metabolic evolution of KJ032 to KJ060 in a medium containing 5% w/v glucose (Fig. 4C) and 10% w/v glucose (Fig. 4D). Metabolic evolution of KJ070 to KJ071 in the medium containing 10% glucose (Fig. 4E). Metabolic evolution of KJ072 to KJ073 in the medium containing 10% glucose (Fig. 4F). Symbols for Fig 4A – 4F: ┆, succinate; □, formate; Δ, acetate; ▲, malate; ♦, lactate; and ▼, pyruvate.

(036) Figure 5. Diagram summarizing steps in the genetic engineering and metabolic evolution of *E. coli* C as a biocatalyst for succinate production. This process represents 261 serial transfers providing over 2000 generations of growth-based selection. Clones were isolated from the final culture of each regimen and assigned strain designations, shown in parenthesis in Table 4.

(037) Figure 6. Standard pathway for the fermentation of glucose-6-phosphate with associated pathways showing the genes that have been deliberately deleted in constructs engineered for succinate production. Solid arrows represent central fermentative pathways. Dashed arrow represents the microaerophilic pathway for the oxidation of pyruvate to acetate. Dotted arrows show pathways including glyoxylate bypass that normally function during aerobic metabolism. Boxed crosses represent the three initial gene deletions (*ldhA, adhE, ackA*) that were used to construct KJ012 and KJ017. Plain crosses mark additional genes that were deleted during the construction of KJ017 derivatives: KJ032 (*ldhA, adhE, ackA, focA, pflB*), and KJ070 (*ldhA, adhE, ackA, focA, pflB, mgsA*), and KJ072 (*ldhA, adhE, ackA, focA, pflB, mgsA, poxB*). Genes and enzymes: *ldhA*, lactate dehydrogenase; *focA*, formate transporter; *pflB*, pyruvate-formate lyase; *pta*, phosphate acetyltransferase; *ackA*, acetate kinase; *adhE*, alcohol dehydrogenase; *ppc*, phosphoenolpyruvate carboxylase; *pdh*, pyruvate dehydrogenase complex; *gltA*, citrate synthase; *mdh*, malate dehydrogenase; *fumA*, *fumB*, and *fumC*, fumarase isozymes; *frdABCD*, fumarate reductase; *fdh*, formate dehydrogenase; *mgsA*, methylglyoxal synthase; *gloAB*, glyoxylase I and II; *poxB*, pyruvate oxidase; *aceA*, isocitrate lyase; *aceB*, malate synthase; *acnAB*, aconitase; and *acs*, acetyl~CoA synthetase.

(038) Figures 7A-7C. Production of succinate and malate in mineral salts media with 10% glucose (w/v) by derivatives of *E. coli* C. Fig. 7A shows succinate production by KJ060 in AM1 medium. Fig. 7B shows succinate production by KJ073 in AM1 medium. Fig. 7C shows production of malate by KJ071 in NBS medium. Fermentations were inoculated at a level of 33 mg DCW l^{-1}. Symbols for Figure 7A-7C: ○, glucose; ●, succinate; ■, malate; Δ, cell mass.

(039) Figure 8. Steps involved in the construction of plasmid pLOI4162. Short solid arrows associated with pEL04 and pLOI4152 represent primers used for DNA amplification.

(040) Figure 9. Succinate production from glucose-6-phosphate in KJ073. The *pck* gene encoding phosphoenolpyruvate carboxykinase, the primary carboxylating enzyme involved in succinate production in this study, is shown in reverse type. Solid arrows indicate reactions expected to be functional during anaerobic fermentation of glucose. Solid crosses indicate deleted genes. Boxed crosses represent key deletions used to construct initial strain for succinate production, KJ017 (*ldhA, adhE, ackA*). The dashed line represents oxidation of pyruvate to acetate by PoxB, a process that is typically functional only under microaerophilic conditions. The dotted lines indicate reactions that are primarily associated with aerobic metabolism. Genes and enzymes: *ldhA*, lactate dehydrogenase; *pflB*, pyruvate-formate lyase; *focA*, formate transporter; *pta*, phosphate acetyltransferase; *ackA*, acetate kinase; *adhE*, alcohol dehydrogenase; *pck*, phosphoenolpyruvate carboxykinase; *pdh*, pyruvate dehydrogenase complex; *gltA*, citrate synthase; *mdh*, malate dehydrogenase; *fumA*, *fumB*, and *fumC*, fumarase isozymes; *frdABCD*, fumarate reductase; *fdh*, formate dehydrogenase; *icd*, isocitrate dehydrogenase; *acs*, acetyl~CoA synthetase; *mgsA*, methylglyoxal synthase; *poxB*, pyruvate oxidase; *aldA*, aldehyde dehydrogenase; and *aldB*, aldehyde dehydrogenase. The *tdcE* gene (pyruvate formate-lyase, homologous to *pflB*) and *tcdD* gene (propionate kinase, homologous to *ackA*) are shown in parenthesis and are typically expressed during threonine degradation.

(041) Figure 10. Expanded portion of metabolism illustrating the pathways of additional genes that have been deleted (solid crosses). Succinate and acetate are principal products (boxed) from KJ073 fermentations. Genes and enzymes: *citDEF*, citrate lyase; *gltA*, citrate synthase; *aspC*, aspartate aminotransferase; *pck*, phosphoenolpyruvate carboxykinase; *sfcA*,

NAD^+-linked malic enzyme; *fumA* & *fumB*, fumarase; *frdABCD*, fumarate reductase; *pykA* & *pykF*, pyruvate kinase; *tdcE*, pyruvate formate-lyase (homologue of *pflB*); *pta,* phosphate transacetylase; *tcdD*, acetate kinase (homologue of *ackA*).

(042) Figure 11. Glucose fermentation in *E. coli* strains engineered for succinate production. The 5 solid stars indicate metabolic steps that have been blocked by constructing deletions. The 2 open stars indicate metabolic steps that have been blocked by mutations acquired during metabolic evolution. Dotted arrows indicate non-functional or weakly functional activities in succinate-producing mutants (KJ060 and KJ073). Bold arrows in the vertical direction indicate the primary pathway for succinate production in KJ060 and KJ073. This pathway is functionally equivalent to that of succinate-producing rumen bacteria. The two genes *galP* and *pck* were transcriptionally activated during strain development. A deletion in *ptsI* was acquired during growth-based selection (open star), inactivating the native glucose phosphotransferase system for uptake and phosphorylation. GalP was subsequently found to serve as the primary transporter for glucose with ATP-dependent phosphorylation by glucokinase (*glk*).

(043) Figures 12A–12C. Comparison of transcript abundance of the engineered succinate-producing strains. FIG 12A shows the relative abundance of transcripts for the genes *pck, ppc, sfcA* and *maeB* in ATCC8739, KJ012, KJ017, KJ060, KJ071 and KJ073 strains of *E. coli*. FIG 12B shows the relative abundance of transcripts for the genes related to glucose utilization namely *cyaA, crp, ptsG, galP* and *glk* in ATCC8739, KJ012, KJ017, KJ060, KJ071 and KJ073 strains of *E. coli*. FIG 12C shows the relative abundance of transcripts for the genes *ptsI* and *crr* gene in the ATCC8739, KJ012, KJ017, KJ060, KJ071 and KJ073 strains of *E. coli*.

(044) Figure 13. Anaerobic metabolism of *E. coli* using the mixed acid fermentation pathway. (Bock & Sawers, 1996). The native mixed acid pathway is shown with black arrows. Additional reactions for glucose uptake, carboxylation, and acetyl-CoA synthesis are shown with dotted arrows. Dotted bold arrows indicate new metabolic steps that have been recruited for succinate production in *E. coli* mutants. Reactions that have been blocked by gene deletions or point mutations are marked with an X. The *pck** indicates a novel mutation

that de-repressed phosphoenolpyruvate carboxykinase, increasing activity and allowing this enzyme to serve as the primary route for oxaloacetate production. Pyruvate (boxed) appears at two sites in this diagram but exists as a single intracellular pool.

(045) Figure 14. Generally recognized pathways for the glycerol catabolism combined with mixed acid fermentation (anaerobic) in wild type *E. coli*. These pathways are based on a combination of the most current reviews in EcoSal, data available in Ecocyc, and a review of primary literature. (Bachler *et al.*, 2005; Berman and Lin, 1971; Bock and Sawers, 1996; Erni *et al.*, 2006; Gutknecht *et al.*, *2001*; Jin *et al.*,*1983*; Keseler *et al.*, *2005*; Lin, 1996; Tang *et al.*, *1982*). Bold arrows indicate the pathways generally regarded as dominant for glycerol catabolism and for mixed acid fermentation. Thin arrows show a pathway regarded as cryptic and nonfunctional in wild type *E. coli* (Jin *et al.*, *1983*; Tang *et al.*, *1982*) involving dihydroxyacetone as an intermediate. This pathway is thought to function only in mutants in which *glpK* is inactive. (Jin *et al.*, *1983*; Tang *et al.*, *1982*). A thin arrow is also shown for *glpD*, the dehydrogenase thought to function during aerobic metabolism as a replacement for *glpABC* (anaerobic metabolism). Abbreviations: DHA, dihydroxyacetone; DHAP, dihydroxyacetone 3- phosphate; PEP, phosphoenolpyruvate; G3P, glycerol 3-phosphate; and GA3P, glyceraldehydes 3-phosphate. PEP is boxed to indicate a common pool.

(046) Figure 15. Novel *E. coli* pathway for the anaerobic production of succinate from glycerol in mineral salts medium. Bold arrows indicate the primary route for the anaerobic catabolism of glycerol in strains such as XZ721 containing three core mutations for succinate production. Dotted arrows show pathways that have been blocked by mutations in *pflB* and *ptsI*. In this pathway, mutational activation of phosphoenolpyruvate carboxykinase (*pck**) allows this enzyme to serve as the dominant carboxylation step for the production of oxaloacetate. Unlike the alternative carboxylating enzyme, phosphoenolpyruvate carboxylase (PPC), PCK conserves energy to produce additional ATP for biosynthesis.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

(047) As used in the present invention, the term "titer" means the molar concentration of particular compound in the fermentation broth. Thus in the fermentation process for the production of succinic acid according to the present invention, a succinic acid titer of 100

mM would mean that the fermentation broth at the time of measurement contained 100 mMoles of succinic acid per liter of the fermentation broth.

(048) As used in the present invention, the term "yield" refers to the moles of particular compound produced per mole of the feedstock consumed during the fermentation process. Thus in the fermentative process for the production of succinic acid using glucose as the feedstock, the term yield refers to the number of moles of succinic acid produced per mole of glucose consumed.

(049) As used in the present invention, the term "volumetric productivity" refers to the amount of particular compound in grams produced per unit volume per unit time. Thus a volumetric productivity value of 0.9 g L^{-1} h^{-1} for succinic acid would mean that 0.9 gram succinic acid is accumulated in one liter of fermentation broth during an hour of growth.

(050) The terms "titer," "yield," and "volumetric productivity" as used in this invention also include "normalized titer," "normalized yield," and "normalized volumetric productivity." In the determination of the normalized titer, normalized yield, and normalized volumetric productivity, the volume of the neutralizing reagents added to the fermentation vessel in order to maintain the pH of the growth medium is also taken into consideration.

(051) The terms "genetically engineered" or "genetically modified" as used herein refers to the practice of altering the expression of one or more enzymes in the microorganisms through manipulating the genomic DNA of the microorganisms.

(052) The present invention provides a process for the production of succinic acid in commercially significant quantities from the carbon compounds by genetically modified bacterial strains (GMBS). Disclosed in this present invention are the microorganisms suitable for the production of succinic acid through fermentative process.

(053) As used in the present invention, the term "gene" includes the open reading frame of the gene as well as the upstream and downstream regulatory sequences. The upstream regulatory region is also referred as the promoter region of the gene. The downstream regulatory region is also referred as the terminator sequence region.

(054) As used in the present invention, the term mutation refers to genetic modifications done to the gene including the open reading frame, upstream regulatory region and downstream regulatory region. The gene mutations result either an up regulation or a down regulation or complete inhibition of the transcription of the open reading frame of the gene. The gene mutations are achieved either by deleting the entire coding region of the gene or a portion of the coding nucleotide sequence or by introducing a frame shift mutation, a missense mutation, and insertion, or by introducing a stop codon or combinations thereof.

(055) As used in this invention, the term "exogenous" is intended to mean that a molecule or an activity derived from outside of a cell is introduced into the host microbial organism. In the case an exogenous nucleic acid molecule introduced into the microbial cell, the introduced nucleic acid may exist as an independent plasmid or may get integrated into the host chromosomal DNA. The exogenous nucleic acid coding for a protein may be introduced into the microbial cell in an expressible form with its own regulatory sequences such as promoter and terminator sequences. Alternatively, the exogenous nucleic acid molecule may get integrated into the host chromosomal DNA and may be under the control of the host regulatory sequences.

(056) The term "endogenous" refers to the molecules and activity that are present within the host cell. When used in reference to a biosynthetic activity, the term "exogenous" refers to an activity that is introduced into the host reference organism. The source can be, for example, a homologous or heterologous encoding nucleic acid that expresses the referenced activity following introduction into the host microbial organism. If the nucleic acid coding for a protein is obtained from the same species of the microbial organism, it is referred as homologous DNA. If the nucleic acid derived from a different microbial species, it is referred as heterologous DNA. Irrespective of the nature of the DNA, whether it is homologous or heterologous, when introduced into a host cell, the DNA as well as the activity derived form that introduced DNA is referred as exogenous. Therefore, exogenous expression of an encoding nucleic acid of the invention can utilize either or both heterologous and homologous encoding nucleic acid.

(057) The present invention provides GMBS showing impressive titers, high yield and significant volumetric productivity for succinic acid when grown under fermentative conditions in minimal salt medium containing a carbon source as the substrate for fermentation process. The microorganisms of the subject invention can be employed in a single step production process using various sugars such as hexoses, pentoses, disaccharides and other carbon compounds such as glycerol.

(058) In the present invention, unique and advantageous combinations of gene mutations have been employed to direct the carbon flow to succinic acid production. In addition, the succinic acid production is coupled with microbial growth which in turn is coupled to cellular ATP level and redox balance.

(059) The term "redox balance" refers to the ability of the cell to maintain the appropriate ratio of NADH to NAD^+. In other words, the cells are able to oxidize the NADH so that there is enough NAD^+ to oxidize the carbohydrate substrates during the anaerobic fermentative growth. During aerobic growth, the NAD^+ pool is regenerated through oxidative phosphorylation involving NADH. However, under anacrobic growth condition the regeneration of NAD^+ pool is achieved only by means of manipulating the flow of carbon through various metabolic pathways inside the cell which could oxidize NADH.

(060) In one embodiment of the present invention, the genetic modifications involve only the manipulation of genes within the native genome of the microorganisms. In that embodiment of the present invention, no exogenous genetic material such as plasmid bearing antibiotic resistance genes or any other exogenous nucleotide sequences coding for certain enzyme proteins is introduced into the bacterial strains used as a biocatalysts for succinic acid production.

(061) The recombinant microorganisms suitable for this present invention are derived from a number of bacterial families, preferably from the Enterobacteriaceae family. The suitable microorganisms are selected form the genera *Escherichia*, *Erwinia*, *Providencia*, and *Serratia*. The genus *Escherichia* is particularly preferred. Within the genus *Escherichia*, the species *Escherichia coli* is particularly preferred. Any one strain of *E. coli* such as *E. coli* B, *E. coli* C, *E. coli* W, or the like is useful for the present invention.

(062) *E. coli* strains capable of producing organic acids in significant quantities are well known in the art. For example, the U. S. Patent Application Publication No. 2009/0148914 provides strains of *E. coli* as a biocatalyst for the production of chemically pure acetate and/or pyruvate. The U. S. Patent Application Publication No. 2007/0037265 and U.S. Patent 7,629,162 provide derivatives of *E. coli* K011 strain constructed for the production of lactic acid. International Patent Application published under the Patent Cooperation Treaty No. WO 2008/115958 provides microorganism engineered to produce succinate and malate in minimal mineral salt medium containing glucose as a source of carbon in pH-controlled batch fermentation.

(063) In some other embodiments of the invention, bacteria that can be modified according to the present invention include, but are not limited to, *Gluconobacter oxydans, Gluconobacter asaii, Achromobacter delmarvae, Achromobacter viscosus, Achromobacter lacticum, Agrobacterium tumefaciens, Agrobacterium radiobacter, Alcaligenes faecalis, Arthrobacter citreus, Arthrobacter tumescens, Arthrobacter paraffineus, Arthrobacter hydrocarboglutamicus, Arthrobacter oxydans, Aureobacterium saperdae, Azotobacter indicus, Brevibacterium ammoniagenes, divaricatum, Brevibacterium lactofermentum, Brevibacterium flavum, Brevibacterium globosum, Brevibacterium fuscum, Brevibacterium ketoglutamicum, Brevibacterium helcolum, Brevibacterium pusillum, Brevibacterium testaceum, Brevibacterium roseum, Brevibacterium immariophilium, Brevibacterium linens, Brevibacterium protopharmiae, Corynebacterium acetophilum, Corynebacterium glutamicum, Corynebacterium callunae, Corynebacterium acetoacidophilum, Corynebacterium acetoglutamicum, Enterobacter aerogenes, Erwinia amylovora, Erwinia carotovora, Erwinia herbicola, Erwinia chrysanthemi, Flavobacterium peregrinum, Flavobacterium fucatum, Flavobacterium aurantinum, Flavobacterium rhenanum, Flavobacterium sewanense, Flavobacterium breve, Flavobacterium meningosepticum, Micrococcus sp.* CCM825, *Morganella morganii, Nocardia opaca, Nocardia rugosa, Planococcus eucinatus, Proteus rettgeri, Propionibacterium shermanii, Pseudomonas synxantha, Pseudomonas azotoformans, Pseudomonas fluorescens, Pseudomonas ovalis, Pseudomonas stutzeri, Pseudomonas acidovolans, Pseudomonas mucidolens, Pseudomonas testosteroni, Pseudomonas aeruginosa, Rhodococcus erythropolis, Rhodococcus*

rhodochrous, Rhodococcus sp. ATCC 15592, *Rhodococcus sp.* ATCC 19070, *Sporosarcina ureae, Staphylococcus aureus, Vibrio metschnikovii, Vibrio tyrogenes, Actinomadura madurae, Actinomyces violaceochromogenes, Kitasatosporia parulosa, Streptomyces coelicolor, Streptomyces flavelus, Streptomyces griseolus, Streptomyces lividans, Streptomyces olivaceus, Streptomyces tanashiensis, Streptomyces virginiae, Streptomyces antibioticus, Streptomyces cacaoi, Streptomyces lavendulae, Streptomyces viridochromogenes, Aeromonas salmonicida, Bacillus subtilis, Bacillus licheniformis, Bacillus amyloliqyefaciens, Bacillus coagulans, Bacillus pumilus, Bacillus circulans, Bacillus thiaminolyticus, Escherichia freundii, Microbacterium ammoniaphilum, Serratia marcescens, Salmonella typhimurium, Salmonella schottmulleri, Xanthomonas citri* and so forth.

(064) The microorganisms suitable for the practice of present invention can be grown aerobically (in the presence of oxygen) or anaerobically (in the complete absence of oxygen) or micro aerobically (with a minimal amount of oxygen supply). In the preferred embodiment of the present invention, the microorganism selected for the production of succinic acid is grown in an anaerobic condition. Alternatively, the microorganisms suitable for the present invention can be grown in a dual-phase growth regime, wherein the microorganism is initially grown in aerobic growth condition to reach a certain level of cell growth before transferring it to the anaerobic growth condition to achieve the production succinic acid in commercially significant quantities. During the dual-phase growth for the production of succinic acid by the microorganisms of the present invention, production and the accumulation of the succinic acid occurs during the anaerobic fermentative growth phase.

(065) The present invention combines the technique of specific genetic modifications with the process of metabolic evolution to obtain strains showing high yield, titer and volumetric productivity for succinic acid production under anaerobic growth condition in the mineral salt medium with a carbohydrate substrate.

(066) The microbial strains obtained from genetic manipulations would have the expected genotype for the production of succinic acids. However, their growth rate in the minimal mineral salt medium or the their ability to produce succinic acid at the required yield, titer and volumetric productivity may not allow us to use these genetically modified

microorganism as a biocatalyst for the commercial production of succinic acid through large scale fermentation process. Therefore, the genetically modified microbial strains obtained from genetic modifications are subsequently grown in mineral salt medium with a carbohydrate source for several generations to select a clone with very high yield for succinic acid production. This process for the growth-based selection of a clone with most preferred phenotype is referred as metabolic evolution. During the metabolic evolution, the genetically modified strain is repeatedly transferred into fresh minimal medium for a period of time to obtain a clone in which the spontaneous mutations that occurred during metabolic evolution results in a clone that exhibits fast cell growth, rapid consumption of different carbon sources, ability to use multiple sugars simultaneously, ability to tolerate toxic chemicals in the carbon source and high production yield and productivity of the desired organic acid coupled with the low production of other organic acids.

(067) During the metabolic evolution, attention is paid to select the clone with the desirable phenotypes. A clone resulting from the metabolic evolution showing a very good growth rate in mineral medium supplemented with a carbon source but that has not improved in the yield of the desired organic acid is not a desirable clone.

(068) During the process of metabolic evolution using certain selective pressure to force the organism to acquire certain desirable phenotype, two possible changes could occur. The organism could simply adapt itself to the selective pressure and show a changed phenotype. Alternatively, the organism might undergo certain genetic changes under selective pressure and exhibit a changed phenotype permanently. When there was only an adaptation and there is no genetic change, the organism reverts back to its original phenotype once the selection pressure is relieved. These organisms are referred to as "adapted" organisms. These "adapted" microorganisms would revert back to their original phenotype when the selection pressure is removed. The "adapted" microorganisms have to undergo another fresh round of metabolic evolution under selection pressure to show a changed phenotype. On the other hand, when there is an accompanying genetic change, the changed phenotype will continue to exist even when there is no selection pressure. Metabolic evolution accompanied by a certain genetic change is desirable. The microorganism acquiring a stable genetic change during metabolic evolution can be easily identified by means of growing the microorganism in the

original growth medium without any selection pressure for some time before transferring it to the fresh medium with the selection pressure. If these organisms are able to show good growth and the expected phenotype without any lag period, the organism is considered to have acquired a changed genotype during metabolic evolution.

(069) The basis of genetic change gained during the metabolic evolution can be determined by sequencing appropriate regions within the chromosomal DNA of the organism and comparing the sequence data with that of the parent strain. The DNA sequence data can be obtained by means of following the techniques well known in the art. For example, appropriate regions of the chromosomal DNA of the metabolically evolved strain can be obtained through polymerase chain reaction and the product obtained through polymerase chain reaction can be sequenced by using appropriate sequencing primers.

(070) The wild type *E. coli* strains obtained from culture collections such as ATCC (American Type Culture Collection) can be genetically engineered and subsequently metabolically evolved to obtain a strain with an enhanced ability to produce succinic acid in commercially significant amounts.

(071) The genetic manipulations can be done in several different stages accompanied by metabolic evolution in between the stages of genetic manipulations. The genomic manipulations involve either altering the endogenous DNA sequences or completely removing specific DNA sequences from the genomic DNA. The genetic manipulations may also involve inserting a foreign DNA sequence within the genomic DNA sequence of the microorganism. Certain embodiments of the present invention, the genetic manipulations are accomplished by means of removing specific DNA sequences from the genomic DNA of the microorganisms without introducing any foreign DNA. Certain genetic manipulations necessary to inactivate the expression of a gene coding for a particular protein product requires an insertion of a foreign DNA sequence into the genome of the microorganism to select a clone with the desired genetic modification. For example, exogenous antibiotic marker genes can be used to insertionally inactivate the endogenous genes and to select the clone with desired genotype. In one embodiment of the present invention, the introduced

exogenous DNA sequences are ultimately removed from the genomic DNA of the microorganism so that the microorganism at the end of the genetic engineering process would have no exogenous DNA in its original genomic DNA. Various genetic engineering techniques necessary for accomplishing the objectives of the preferred embodiment of the present invention have been described in detail in two different scientific publications (Jantama *et al.*, 2008a; Jantama *et al.*, 2008b). The published U.S. Patent Applications with numbers US 2007/0037265 and US 2009/0148914 and the International patent application published under the Patent Cooperation Treaty with International Publication Number WO 2008/115958 also describe the genetic engineering techniques useful in practicing various embodiments of this present invention. These scientific publications as well as patent documents are herein incorporated by reference for the purpose of providing the details for genetic engineering techniques useful for the present invention.

(072) In order to make the microorganism to produce succinic acid in significant quantities, various enzymes involved in a number of microbial metabolic pathways including glycolytic pathway, tricarboxylic acid cycle (also known as Krebs cycle or TCA cycle) and glyoxylate shunt can be manipulated using a variety of genetic engineering techniques described in the scientific and patent literature cited and incorporated by references in the paragraph above. The details about various microbial metabolic pathways can be found in the standard biochemistry text books such as Principles of Biochemistry, by Lehninger and Biochemistry by Lubert Stryer. The Biochemical pathways poster by G. Michael available from Sigma Chemical Company in St. Louis, MO, USA also provides details about various biochemical pathways with in a bacterial cell.

(073) During the aerobic growth, the microbial carbon metabolism involves glycolysis, tricarboxylic acid cycle and oxidative phosphorylation. The reduced enzyme co-factors such as NADPH and NADH are regenerated by the operation of oxidative phosphorylation accompanied by ATP production required for cell growth. Under anaerobic growth condition for the production of succinic acid in the preferred embodiment of the present invention, the regeneration of reduced cofactors NADPH and NADH is accomplished by directing the carbon flow into the tricarboxylic acid cycle and eliminating all of the fermentative pathways for regeneration of $NADP^+$ and NAD^+.

(074) Depending on the type of organic acid preferred, the metabolic pathways are specifically engineered so that the microorganism produces a particular organic acid of our choice. The microorganisms are capable of synthesizing a number of organic acids including lactic acid, acetic acid, malic acid, pyruvic acid, formic acid and succinic acid. Thus in developing a biocatalyst for the production of succinic acid, the pathways for production of acetic acid, lactic acid, pyruvic acid, and formic acid are blocked and the carbon flow to succinic acid production is facilitated through manipulating one or more enzymes involved in the carbon metabolism within the cell. The list of the enzymes that are active in the microbial fermentative pathway which can be manipulated using the known genetic engineering techniques includes, but not limited to, isocitrate synthetase (*aceA*), malate synthase (*aceB*), the glyoxylate shunt operon (*aceBAK*), acetate kinase-phosphotransacetylase (*ackA-pta*); aconitase hydratase 1 and 2 (*acnA and acnB*); acetyl-CoA synthetase (*acs*); citrate lyase (*citDEF*); alcohol dehydrogenase (*adhE*); citrate synthase (*citZ*); fumarate reductase (*frd*); lactate dehydrogenases (*ldh*); malate dehydrogenase (*mdh*); aceBAK operon repressor (*iclR*); phosphoenol pyruvate carboxylase (*pepC*); pyruvate formate lyase (*pfl*); pyruvate oxidase (*poxB*); pyruvate carboxy kinase (*pck);* and pyruvate carboxylase (*pyc*) (Figures 1, 6, and 9).

(075) Glycolysis of carbon sources results in the production of phosphoenol pyruvate (PEP). PEP is further metabolized by mixed acid pathway. As used in the present invention, the term "mixed acid pathway" refers to the flow of carbon from PEP through both tricarboxylic acid cycle and various fermentative pathways that are operational under anaerobic conditions. Under anaerobic conditions, at least four different fermentative pathways for the metabolism of pyruvate are recognizable. The pyruvate may be reduced to lactate using the NADH and thereby producing NAD^+ to maintain the redox balance of the cell necessary for the continuous metabolism of carbon source. The acetyl-CoA derived from pyruvate may also be reduced to produce ethanol accompanied by the oxidation of NADH to produce NAD^+. Pyruvate may also be converted into formate or acetate as shown in Figure 1A.

(076) Within the TCA cycle, two different arms are recognized. In one arm of the TCA cycle referred as the oxidative arm encompassing the carbon flow from oxaloacetate to succinic acid through isocitrate, the $NADP^+$ is utilized to oxidize isocitrate with the resulting

formation of NADPH. In the other arm of the TCA cycle referred as the reductive arm of the TCA cycle encompassing the flow of carbon from oxaloacetate to succinic acid through malate and fumarate, the NADH is oxidized to produce NAD^+ and thereby helping the cell to maintain the redox balance.

(077) In one embodiment of the present invention, the carbon flow from PEP through fermentative pathways is prevented by mean of inactivating the genes coding for the enzymes involved in the fermentative pathway. The enzymes suitable for blocking the carbon flow through fermentative pathway include *ldhA, pflB, adhE, pta, ackA*, and *poxB*. The elimination of one or more of these genes is expected to reduce the carbon flow from PEP through the fermentative pathway. Inactivation of all these six genes is expected to block the carbon flow through fermentative pathway totally. In another aspect of the present invention, the *mgsA* gene coding for the methylglyoxal synthase (*mgsA*) responsible for the conversion of methylglyoxal to lactic acid is inactivated beside the inactivation of six other genes involved in the fermentative pathway.

(078) In yet another embodiment of the present invention, the functional homologues of the genes involved in the fermentative pathway are also inactivated besides inactivating the genes well known to be involved in one or other fermentative pathway. A propionate kinase with acetate kinase activity is encoded by the *tdcD* gene which is produced only for the degradation of threonine. However, during the anaerobic growth with 10% (w/v) glucose, the expression of *tdcD* could functionally replace *ackA*. In addition, the adjacent *tdcE* gene in the same operon is similar to *pflB* and encodes α-ketobutryate formate lyase with pyruvate formate-lyase activity. In one aspect of the present invention, the *tdcDE* genes are inactivated to prevent the entry of carbon into fermentative pathway and to assure the flow of carbon into the TCA cycle.

(079) In another embodiment of the present invention, besides blocking the carbon flow through fermentative pathways, the carbon flow within the TCA cycle is altered so that there is carbon flow directed towards the production of succinic acid. In one aspect of the present invention the manipulation of carbon flow within the TCA cycle is achieved by means of up-regulating the expression of one or more genes. In yet another aspect of the present invention,

one or more genes functioning within TCA cycle may be inactivated to facilitate an increased carbon flow to succinic acid.

(080) In a preferred aspect of the present invention, the gene *mdh* encoding for malate dehydrogenase is up regulated to improve the conversion of malate to fumarate and succinate. The flow of the carbon from oxaloacetate to succinic acid through malate and fumarate is referred as the reductive arm of the TCA cycle. The flow of carbon through this reductive arm of the TCA cycle from oxaloacetic acid to succinic acid would consume two moles of NADH for every mole of succinic acid produced and thereby help in maintaining the redox balance of the cell under anaerobic condition. In other words, the up-regulation of *mdh* would help in regenerating the NAD^+ required to maintain the redox balance of the cell. The up-regulation of *mdh* gene expression can be achieved by means of replacing the native promoter for *mdh* gene with some other strong promoter sequence or alternatively by means of mutating the promoter region of the *mdh* gene so that there is an increase transcription of *mdh* gene. Alternatively, additional copies of the *mdh* gene can be added to the strain. In a preferred embodiment of the present invention, the up regulation of *mdh* gene expression is achieved by means of genetically manipulating its promoter region.

(081) In the regular operation of TCA cycle, succinic acid is produced through the operation of oxidative of arm of the TCA cycle. The flow of carbon from oxaloacetate to succinic acid through citrate, cis-aconitate, isocitrate, α-ketoglutarate, and succinyl-CoA is referred as the oxidative arm of the TCA cycle. The succinic acid can also be produced through the operation of glyoxylate bypass. During the operation of glyoxylate bypass, by the action of isocitrate lyase, succinate and glyoxylate are produced from isocitrate. The succinate thus produced from the operation of oxidative arm of the TCA cycle or from the operation of glyoxylate bypass, can be acted upon by succinate dehydrogenase (*sdh*) to yield fumaric acid and then malic acid. Therefore, in yet another embodiment of the present invention, gene inactivation can be used to prevent the dehydrogenation of the succinate in order to increase the intracellular succinic acid production.

(082) In yet another aspect of the present invention, the carbon flow through the glyoxylate bypass can be manipulated to achieve an increase in the succinic acid production. Isocitrate

lyase enzyme catalyzes the cleavage of isocitrate to glyoxylate and succinate. Isocitrate lyase is coded by the *aceBAK* operon. The isocitrate lyase activity is suppressed by *iclR* genes. In other words, the expression of *iclR* gene prevents the operation of glyoxylate shunt. In one aspect of the present invention, the *iclR* gene is inactivated beside the inactivation of the genes involved in the fermentative metabolism.

(083) In yet another embodiment of the present invention, besides preventing the operation of the fermentative pathways and increasing the flow of carbon within the TCA cycle towards succinic acid production through genetic manipulations, the outward carbon flow from the TCA cycle to other metabolic pathways can also be blocked through genetic means to increase the succinic acid production. For example, the flow of carbon from the TCA cycle into amino acid metabolism can be blocked in order to improve the carbon flow towards succinic acid. The aspartate aminotransferase gene (*aspC*) transfers the amino group from glutamic acid to oxaloacetic acid in the synthesis of aspartic acid and thereby facilitates the outward flow of carbon from the TCA cycle. In one aspect of the present invention, the inactivation of the *aspC* gene is followed to block the outward flow of carbon from the TCA cycle in order to improve the carbon flow from oxaloacetate towards succinic acid production either through the oxidative or reductive arm of the TCA cycle.

(084) The other outward flow of the carbon from TCA cycle occurs from malate. The decarboxylation of malate by malic enzyme (*sfcA*) results in the production of pyruvate. In one aspect of the present invention, the gene coding for the *sfcA* gene is inactivated to curtail the outward flow of carbon from TCA cycle. In yet another aspect of the present invention, both *aspC* and *sfcA* genes are inactivated to prevent the outward flow of carbon from TCA cycle so as to enhance the succinic acid accumulation.

(085) In yet another aspect of the present invention, the outward flow of carbon from TCA cycle is prevented by inactivating the citrate lyase gene (*citDEF*) responsible for the cleavage of citric acid into oxaloacetate and acetate.

(086) Besides discovering that inactivating the genes involved in the fermentative pathways and their functional analogues, preventing the carbon flow out of the TCA cycle and regulating the carbon flow towards succinic acid within the TCA cycle could improve the

succinic acid yield in the microbial fermentation, the present invention has also surprisingly discovered that the growth coupled succinic acid yield can further be improved by genetic manipulations of carboxylating enzymes within the microbial cells. While characterizing the changes that occurred during the metabolic evolution through conducting genetic and enzyme analysis, the inventors of the present invention have unexpectedly discovered that the carboxylating enzymes within the cell could be yet another target for genetic manipulation to achieve an improved succinic acid yield.

(087) The glycolytic intermediates phosphoenol pyruvate (PEP) and pyruvic acid can be carboxylated to improve the carbon flow into the TCA cycle. Under normal conditions, the carbon entry into the TCA cycle is accomplished by the action of citrate synthase which combines the acetyl-CoA derived from pyruvate with oxaloacetate, an intermediate in TCA cycle, to produce citric acid. By means of improving the efficiency of one or more carboxylating enzymes present within the cell, it is possible to carboxylate phosphoenol pyruvate and pyruvate to oxaloacetate, a TCA cycle intermediate (Figure 2). The oxaloacetate thus produced from the carboxylating reaction can be further reduced through the reductive arm of the TCA cycle to produce succinic acid.

(088) The present invention provides a method for manipulating the carboxylating enzymes present within the cell as a method to increase the succinic acid yield during anaerobic fermentative growth. It is well known in the art that by means of introducing pyruvate carboxylase (*pyc*) from an exogenous source it is possible to carboxylate pyruvate to oxaloacetic acid. The microbial strains well suited for genetic manipulations such as *E. coli* do not have the *pyc* gene. The *pyc* genes derived from other bacterial species such as *Rhizopium elti* and *Lactobacillus lacti* can be introduced into the genetically modified *E. coli* strains to improve succinic acid production.

(089) Four different endogenous carboxylating enzymes are known in *E. coli*. Two of these enzymes are responsible for carboxylating phosphoenol pyruvate and two other enzymes are responsible for the carboxylation of pyruvate derived from phosphoenol pyruvate by the action of pyruvate kinase enzyme. The enzyme phosphoenol pyruvate carboxylase (*ppc*) carboxylates phosphoenol pyruvate to oxaloacetate which could enter into reductive arm of the TCA cycle to produce succinate. The second carboxylating enzyme phosphoenol pyruvate

carboxykinase (*pck*) also carboxylates phosphoenol pyruvate to produce oxaloacetate, but normally catalyzes the reverse reaction as it is not expressed in the presence of glucose. The two other carboxylating enzymes namely NADH-linked maleic enzyme (*maeB*) and the NADPH-linked maleic enzyme (*maeA / sfcA*) carboxylate pyruvic acid to malic acid. The *maeB* and *sfcA* enzymes carboxylates the pyruvate derived from phosphoenol pyruvate by the action of pyruvate kinase.

(090) Any one of the four carboxylating enzymes present in the cell can be genetically manipulated to increase its enzymatic activity in order to improve the carbon flow from glycolytic cycle intermediates into the TCA cycle. Of the four native carboxylating enzymes present in *E.coli*, the PPC-catalyzed reaction is strongly favored. Energy contained in PEP is lost in this reaction with the release of inorganic phosphate. The other three carboxylating enzymes, namely *pck*, *maeA* and *sfcA* (*maeB*), are not expected to function during the fermentative growth using glucose as the substrate as these three carboxylating enzymes are repressed by glucose. These three carboxylating enzymes are thought to function in the reverse direction during gluconeogenesis when the cells are oxidatively metabolizing organic acids.

(091) In this invention, the inventors have surprisingly discovered that gluconeogenic PEP carboxykinase (PCK) can be genetically manipulated to improve the flow of carbon into the TCA cycle. The recruitment of *pck* as the primary pathway for succinic acid production in *E. coli* was surprising and is in contrast to our current understanding on the functional role of *pck*. Previous studies have shown that increased expression of *E. coli* and *Actinobacillus. succinogenes pck* had no effect on succinate production (Kim *et al.*, 2004; Millard *et al.*, 1996). A recent study has demonstrated that increased expression of *E. coli pck* is detrimental for growth in minimal medium, decreasing the growth rate, the rate of glucose metabolism, and the yield of succinate. [Kwon *et al.*, 2008). The advantage in improving the activity of *pck* lies in the fact that this enzyme while carboxylating phosphoenol pyruvate to oxaloacetate, results in the production of a molecule of ATP for every molecule of oxaloacetate produced. An increase in the ATP yield would increase the growth rate of the cells.

(092) The comparative analysis of phosphoenol pyruvate carboxykinase activity in a number of *E. coli* strains constructed during this invention has revealed that the increase in the PCK enzyme activity during the metabolic evolution results from an increase in the transcriptional activity of the *pck* gene. In those strains showing an improvement in the cell growth-coupled succinate production, there is an increase in the abundance of the *pck* transcript. In fact, the results of the present invention have established a positive correlation between an increase in the *pck* transcript level and an increase in the PCK enzyme activity and growth-coupled succinic acid production.

(093) The recruitment of the native gluconeogenic *pck* for fermentative succinate production can be achieved by any mutation that positively affects the transcription of the *pck* gene. An increase in the level of PCK activity can be achieved by means of expressing the *pck* gene in a multicopy plasmid with a native promoter or any other promoter sequence which is known to increase the gene's expression. Another way to increase the expression of the *pck* gene within the cell is to integrate additional copies of the *pck* gene using transposons. In another embodiment of the present invention, the native promoter of the *pck* gene can be replaced by some other promoter elements known to enhance the level of activity. An increased expression of *pck* gene can also be achieved either by mutation in the promoter region of the gene or by genetic manipulation of the regulatory elements that are known to interact with the promoter region of the *pck* gene. The gene coding for a regulator protein of the *pck* gene can be mutated or deleted or overexpressed in some way in order to increase the expression of *pck* gene. The results of the present invention have indicated that a single point mutation (G to A transition at position - 64 relative to the ATG start codon of *pck* gene) could increase the transcription of the *pck* gene accompanied by a corresponding increase in the phosphoenol pyruvate carboxykinase enzyme activity. A similar increase in the *pck* gene expression can also achieved by genetically manipulating the genes coding for the proteins known to regulate the expression of *pck* gene. For example, Cra protein has been shown to activate the expression of *pck* gene in *E. coli* (Saier and Ramseier, 196). Similarly the *csrA* system (comprising *csrA*, *csrB*, *csrC*, *csrD*, *uvrY* or *barA*) has also been reported to regulate the level of *pck* and other genes involved in glucose metabolism by altering mRNA stability (Babitzke and Romeo, 2007; Pernestig *et al.*, 2003; Suzuki K *et al.*, 2002).

(094) Yet another genetic approach of the present invention to increase the growth-coupled succinic acid production during the anaerobic fermentation process is concerned with the conservation of energy expended in sugar uptake by the biocatalysts.

(095) The microorganisms take up the sugars through a set of transporter proteins located on the cytoplasmic membrane (Jojima *et al.*, 2010). The microbial sugar transporters fall within three major categories. The largest group of sugar transporters in the bacteria is known as ATP binding cassette (ABC) transporters. As the name implies, the ABC transporters require a molecule of ATP for every molecule of sugar transported into the bacterial cell. XylFGH is an ABC transporter for the transport of xylose, a pentose sugar, into the cell. AraFGH is an ABC transporter for the transport of arabinose, yet another pentose sugar.

(096) The second type of bacterial sugar transporters are grouped under Major Facilitator Super family (MFS). Within the MFS sugar transporters, two different categories of transporter are recognized. MFS includes H^+- linked symporters, Na^+ - linked symporters-antiporters and uniporters. The uniporters are simple facilitators for the sugar transport and require a molecule of ATP for every molecule of sugar transported into the cell. The trans-membrane protein Glf in *E. coli* is an example of uniporter. The H^+ - symporters require a proton and a molecule of ATP for every sugar molecule transported into the cell. The GalP protein in *E. coli* is a symporter for the transport of galactose, a hexose sugar, into the cell. GalP is a very well characterized symporter with 12 trans-membrane loops. GalP is also reported to have the ability to transport glucose across the cell membrane. AraE is a proton-linked symporter for the transport of arabinose across the cell membrane. Similarly XylE protein is a proton-linked symporter for the transport of xylose.

(097) The third sugar transporter primarily responsible for the uptake of hexose sugars such as glucose is known as the phosphoenolpyruvate: carbohydrate phosphotransferase system (PTS). Transfer of the phosphoryl group from phosphoenolpyruvate (PEP) catalyzed by PTS drives the transport and phosphorylation of glucose and results in the formation of glucose 6-phosphate and pyruvic acid inside the cell. PTS generated pyruvic acid is apparently not recycled to PEP under aerobic culture conditions where glucose is the sole source of carbon. Rather, pyruvate is oxidized by way of the tricarboxylic acid cycle to carbon dioxide. Thus,

for the transport of every single molecule of glucose, a molecule of PEP is consumed. In terms of cellular bioenergetics, the transport of sugars through PTS is an energy intensive process. Therefore in cells growing anaerobically, where there is a need to conserve the phosphoenolpyruvate content within the cells for the production of industrially useful chemicals, it is desirable to replace the PTS with some other sugar transporters not requiring a molecule of PEP for every molecule of sugar transported into the cell.

(098) Besides these genes directly involved in the glycolysis, tricarboxylic acid cycle and glyoxylate shunt of microbial metabolic pathways, genetic manipulation of the genes involved in the uptake of carbon compounds useful as a source of energy for the synthesis of succinic acid can also be manipulated either to enhance the carbon uptake or to enhance the efficiency of energy utilization in organic acid production. For example the elimination of the glucose uptake by the phosphotransferase system (PTS) could help in reducing the energy spent on glucose uptake into the microbial cell. The energy conserved by manipulating the PTS can be channeled to improve the efficiency of organic acid production. The phosphotransferase system genes *ptsH* and *ptsG* can be manipulated to conserve the energy in glucose uptake and thereby improve the efficiency of succinic acid production by microorganism. Thus by mining the data available in the area of microbial metabolic pathways, one can delete a set of genes so as to block most of the metabolic pathways and channel the carbon flow to the production of succinic acid with great efficiency.

(099) PTS is comprised of two cytoplasmic components namely EI and HPr and a membrane-bound component EII. *E. coli* contains at least 15 different EII complexes. Each EII component is specific to a sugar type to be transported and contains two hydrophobic integral membrane domains (C and D) and two hydrophilic domains (A and B). These four domains together are responsible for the transport and phosphorylation of the sugar molecules. EI protein transfers the phosphate group from PEP to HPr protein. EII protein transfers the phosphate group from phosphorylated HPr protein to the sugar molecule.

(0100) EI is encoded by the *ptsI* gene. HPr is encoded by the *ptsH* gene. The glucose-specific EII complex of enteric bacteria consists of two distinct proteins namely, $EIIA^{Glc}$ encoded by the gene *crr* and the membrane-associated protein $EIICB^{Glc}$ encoded by the gene *ptsG*. The PTS mediated sugar transport can be inhibited by means of deleting one of these genes

coding for the proteins associated with PTS. Functional replacement of PTS by alternative phosphoenolpyruvate-independent uptake and phosphorylation activities is one of the genetic approaches for achieving significant improvements in product yield from glucose and productivity for several classes of metabolites.

(0101) With the inhibition the PTS-mediated glucose uptake, other systems for glucose uptake can be activated to assure the continued availability of glucose within the cell for the production of the industrially useful chemicals. For example, the *glf* gene coding for glucose permease, a glucose uniporter, has been shown to substitute for the loss of PTS mediated glucose uptake. Similarly the over expression of *galP* and *glk* genes are reported to enhance the glucose uptake and phosphorylation in the *pts*⁻ strain of *E. coli*. GalP is a symporter for the uptake of galactose, a hexose sugar. GalP has been reported to transport glucose in the *pts*⁻ strain. The significance of GalP mediated glucose uptake is evidence by the fact that the inactivation of *galP* gene in the *pts*⁻ mutant is found to be lethal (Yi *et al.*, 2003). Glk is necessary to achieve the phosphorylation of the glucose molecule before it can enter into glycolysis. The expression of the GalP protein in the *pts*⁻ strain can be achieved either by expressing an exogenous gene under a constitutive promoter or by means of relieving the repression of the *galP* expression through mutations in genes coding for the repressor of the *galP* gene such as *galS* and *galR*.

(0102) As described above, the succinic acid production using microbial catalysts can be achieved by means of genetic manipulation accompanied by metabolic evolution. The genetic changes that occur during the metabolic evolution can be identified through biochemical and genetic analysis. The present invention have surprisingly discovered that mutations occurring in the genes for the phosphotransferase system and carboxylating enzymes present within the cell could positively contribute to the increase in the succinic acid production. These newly discovered targets for genetic manipulations can be combined with the other targets in the glycolytic, tricarboxylic acid and fermentative pathways in several different ways to generate biocatalysts with high efficiency for succinic acid production. It is also highly desirable to identify a set of a minimal number of target genes for genetic manipulation in order to achieve the best succinic acid producing strains.

(0103) The invention also provides genetic approaches to enhance glycerol utilization in succinic acid production. The glycerol uptake is mediated by the protein coded by *glpF* gene. Once taken into the cell, glycerol is oxidized by the protein coded by *gldA* gene to produce dihydroxy acetone (DHA). The DHA is phosphorylated to dihydroxy acetone phosphate (DHAP) by the proteins coded by *dhaKLM* operon. The phosphorylation of DHA to DHAP by the proteins coded by *dhaKLM* is dependent on the availability of the phosphoenol pyruvate (PEP) pool. Since the phosphorylation of DHA requires PEP, it depletes the PEP available for PCK which directs the flow of carbon from PEP into the TCA cycle in order to assure proper redox balance required to achieve high succinic acid yield. The present invention has surprisingly found that preventing the flow of glycerol through the *gldA* and *dhaKLM* pathways in a bacterial cell having an increased PCK enzymatic activity could enhance the succinic acid yield using glycerol as the carbon source. The present invention has also surprisingly discovered that further improvement in succinic acid yield using glycerol as the carbon source can be achieved by means of preventing the carbon flow through fermentative pathways in a bacterial cell with improved PCK enzyme activity and deletions in the *gldA* gene and *dhaKLM* operon.

(0104) The following examples are provided as way of illustrating the present invention. These inventions in no way limit the scope of this invention. A person experienced in the field of industrial microbiology would be able to practice the present invention in several different embodiments without violating the spirit of the present invention.

EXPERIMENTAL SECTION

General remarks

Strains, media and growth conditions

(0105) New derivatives of *E. coli* C (ATCC 8739) were developed for succinate production using a unique combination of gene deletions coupled with growth-based selection. The various strain of *E. coli* developed in the present invention have been deposited with ARS Culture Collection with accession numbers as shown in Table 2.

(0106) The microbial organism of the present invention can be grown in a number of different culture media well known in the field of microbiology. For example, the wild type and mutant strains of *E. coli* are grown in Luria-Bertani (LB) medium containing 1% (w/v) tryptone, 0.5% (w/v) yeast extract, and 0.5% (w/v) NaCl. For the commercial production of the organic acid using fermentative process involving genetically modified microorganism as biocatalyst, a minimal mineral salt medium supplemented with a carbon source is preferred. The use of a minimal mineral salt medium as opposed to a rich medium like LB medium reduces the cost for the production of organic acids in a commercial scale.

(0107) The minimal mineral mediums suitable for the present invention include NBS medium (Causey *et al.*, 2007) and AMI medium (Martinez *et al.*, 2007). The NBS medium contains 1 mM betaine, 25.72 mM KH_2PO_4, 28.71 mM K_2HPO_4, 26.50 mM $(NH_4)2HPO_4$, 1 mM $MgSO_4.7H2O$, 0.1 mM $CaCl_2.2H_20$, 0.15 mM Thiamine HCl, 5.92 μM $FeCl_36H_20$, 0.84 μM $COCl_2.6H2O$, 0.59 μM $CuCl_2.2H_2O$, 1.47 μM $ZnCl_2$, 0.83 μM Na_2MoO_4 $2H_2O$, and 0.81 μM H_3BO_3. The AM1 medium contains 1 mM betaine, 19.92 mM $(NH_4)2HPO_4$, 7.56 mM $NH_4H_2PO_4$, 1 .5 mM $MgSO_4.7H2O$, 1.0 mM Betaine-KCl, 8.88μM $FeCl_36H_20$, 1,26 μM $COCl_2.6H2O$, 0.88 μM $CuCl_2.2H_2O$, 2.20 μM $ZnCl_2$, 1.24 μM $Na_2MoO_42H_2O$, 1.21 μM H_3BO_3 and 2,50 μM $MnCl_24H_2O$. The trace elements are prepared as a 1000X stock and contained the following components: 1.6 g/L $FeCl_3$, 0.2 g/L $CoCl_2•6H_2O$, 0.1 g/L $CuCl_2$, 0.2 g/L $ZnCl_2•4H_2O$, 0.2 g/L $NaMoO_4$, 0.05 g/L H_3BO_3, and 0.33 g/L $MnCl_2•4H_2O$.

(0108) The mineral medium for microbial production of organic acid is supplemented with a carbon source. The carbon sources useful in the present invention include but are not limited to pentose sugars like xylose, hexose sugars like glucose, fructose, and galactose and glycerol. The carbon source can also be satisfied by providing a combination of different sugars such as a combination of glucose and xylose. The carbon source can also be derived from a hydrolysis of starch or lignocellulose. The hydrolysis of complex carbohydrates such as starch and lignocelluloses can be achieved either by using thermo-chemical conversion processes or enzymatic methods well known in the art. The preferred carbon source for the industrial production of organic acid using microbial fermentation is lignocellulosic hydrolysate derived from the hydrolysis of agricultural or forestry wastes. The lignocellulosic hydrolysate may further be fractionated to yield a hexose-enriched and a

pentose-enriched fraction and those fractions can serve as the source of carbon for the commercial production of the organic acids using microbial fermentation process. The lignocellulosic hydrolysate can further be detoxified to remove certain chemicals such as furfural which are found to be toxic to a number of microbial organisms above certain concentrations.

(0109) Bacterial strains, plasmids, and primers used in this study are listed in Tables 3, 7, 9, 14 and 15 and are explained in detail at the appropriate places in the specification below. During strain construction, cultures were grown aerobically at 30, 37, or 39°C in Luria broth (10 g l^{-1} Difco tryptone, 5 g l^{-1} Difco yeast extract and 5 g l^{-1} NaCl) containing 2% (w/v) glucose or 5% (w/v) arabinose. No genes encoding antibiotic resistance, plasmids, or foreign genes are present in the final strains developed for succinate production. However, during the early stages of construction of the different strains, various antibiotic resistance markers were used. The antibiotics such as ampicillin (50 mg l^{-1}), kanamycin (50 mg l^{-1}), or chloramphenicol (40 mg l^{-1}) were added as needed for antibiotic selection process.

Fermentations

(0110) Seed cultures and fermentations were grown at 37°C, 100 rpm in NBS or AM1 mineral salts medium containing glucose, 100 mM $KHCO_3$ and 1 mM betaine HCl. In some experiments, corn steep liquor was used. It is a byproduct from the corn wet-milling industry. When compared to the yeast extract and peptone, it is an inexpensive source of vitamins and trace elements.

(0111) For fermentative succinate production, strains were grown without antibiotics at 37°C in NBS mineral salts medium (Causey *et al.*, 2004) supplemented with 10% (w/v) glucose and 100 mM potassium bicarbonate unless stated otherwise. Pre-inocula for fermentation were grown by transferring fresh colonies into a 250 ml flask (100 ml NBS medium, 2% glucose). After 16 h (37°C, 120 rpm), this culture was diluted into a small fermentation vessel containing 300 ml NBS medium (10% glucose, 100 mM potassium bicarbonate) to provide an inoculum of 0.033 g cell dry wt (CDW) l^{-1}.

(0112) Since the accumulation of organic acids in the growth medium tends to decrease the pH of the medium, it is necessary to add appropriate neutralizing agents as required to the culture medium. The pH of the culture vessel can be continuously monitored using a pH probe, and appropriate base can be added to maintain the pH of the growth medium around neutral pH. The bases suitable for maintaining the pH of the microbial culture include, but not limited to, NaOH, KOH, NH_4SO_4, Na_2CO_3, $NaHCO_3$, and NH_4CO_3. The bases suitable for this purpose can be used alone or in combination.

(0113) In certain experiments, fermentations were automatically maintained at pH 7.0 by adding base containing additional CO_2 (2.4 M potassium carbonate in 1.2 M potassium hydroxide). Subsequently, pH was maintained by adding a 1:1 mixture of 3M K_2CO_3 and 6N KOH. Fermentation vessels were sealed except for a 16 gauge needle which served as a vent for sample removal. Anaerobiosis was rapidly achieved during growth with added bicarbonate serving to ensure an atmosphere of CO_2.

Genetic methods

(0114) Methods for chromosomal deletions, integration, and removal of antibiotic resistance genes have been previously described (Datsenko and Wanner, 2000; Grabar *et al.*, 2006; Posfai *et al.*, 1997; Zhou *et al.*, 2006).

(0115) In the construction of bacterial strains used in the present invention, chromosomal genes were deleted seamlessly without leaving segments of foreign DNA as described previously. (Jantama *et al.*, 2008a, 2008b; Zhang *et al.*, 2007). Red recombinase technology (Gene Bridges GmbH, Dresden, Germany) was used to facilitate chromosomal integration. Plasmids and primers used during construction are listed in Tables 3, 7, 9, 14, and 15. Plasmids and primers used in the construction of the strains KJ012, KJ017, KJ032, KJ060, KJ070, KJ071, KJ072, KJ073, and SZ204 are summarized in Table 3. Sense primers contain sequences corresponding to the N-terminus of each targeted gene (boldface type) followed by 20 bp (underlined) corresponding to the FRT-*kan*-FRT cassette. Anti-sense primers contain sequences corresponding to the C-terminal region of each targeted gene (boldface type) followed by 20 bp (underlined) corresponding to the cassette. Amplified DNA fragments were electroporated into *E. coli* strains harboring *Red* recombinase (pKD46). In resulting

recombinants, the FRT-*kan*-FRT cassette replaced the deleted region of the target gene by homologous recombination (double-crossover event). The resistance gene (FRT-*kan*-FRT) was subsequently excised from the chromosome with FLP recombinase using plasmid pFT-A, leaving a scar region containing one FRT site. Chromosomal deletions and integrations were verified by testing for antibiotic markers, PCR analysis, and analysis of fermentation products. Generalized P1 phage transduction (Miller, 1992) was used to transfer the Δ*focA-pflB*::FRT-*kan*-FRT mutation from strain SZ204 into strain KJ017 to produce KJ032.

Deletion of FRT markers in the *adhE*, *ldhA*, and *focA-pflB* regions

(0116) The strategy used to make sequential gene deletions and remove the FRT markers from the *adhE, ldhA* and *focA-pflB* loci has been described previously (Datsenko and Wanner, 2000; Grabar *et al.*, 2006; Jantama *et al.*, 2008b; Zhang *et al.*, 2007). Plasmid pLOI4151 was used as a source of a *cat-sacB* cassette and Red recombinase (pKD46) was used to facilitate double-crossover, homologous recombination events. Chloramphenicol resistance was used to select for integration. Growth with sucrose was used to select for loss of *sacB*. With this approach, successive deletions were constructed to produce derivatives of KJ079 that eliminated all FRT sites. Primers and plasmids used in the removal of FRT markers from the *adhE, ldhA* and *focA-pflB* loci are listed in Table 3.

(0117) To remove the FRT site in the Δ*adhE* region, hybrid primers (WM*adhE*A/C) for Δ*adhE*::FRT target region were designed to contain approximately 50 bp of homology to the 5' and 3' regions of Δ*adhE*::FRT site and 20 bp corresponding to *cat-sacB* gene from pLOI4151. These primers were used for PCR amplification of the *cat-sacB* cassette using pLOI4151 as a template. The resulting PCR product was used to replace the FRT site in Δ*adhE* region with a *cat-sacB* cassette by a double-crossover, homologous recombination event with selection for resistance to chloramphenicol, to produce TG200.

(0118) The *adhE* gene and surrounding sequence were amplified from *E. coli* C using up/down*adhE* primers. The PCR product containing *ych*E'-*adh*E-*ych*G' (3.44 kb) was cloned into pCR2.1-TOPO, yielding pLOI4413. A second set of primers (IO-*adhE*up/down) was used to amplify the inside-out product with pLOI4413 as a template and *Pfu* polymerase to yield a blunt-ended product in which a 2.6 kb internal segment of *adhE* sequence was

deleted. This inside-out PCR product was kinase-treated and self-ligated, resulting in pLOI4419. The PCR product amplified from pLOI4419 (up/down*adhE* primers) was used to replace the *cat-sacB* cassette in TG200 with the desired chromosomal sequence by another double, homologous recombination event, with sucrose selection for loss of *sacB*. The resulting strain was designated TG201 (KJ079 with the FRT removed from Δ*adhE* region).

(0119) The FRT sites in the Δ*ldhA* and Δ(*focA-pflB*) regions were removed in a manner analogous to that used to delete the *adhE*::FRT site. Additional primer sets (*ldhA*A/C and IO-*ldhA*up/down) used to remove the FRT site in Δ*ldhA* are included in Table 7 together with the corresponding plasmids (pLOI4430 and pLOI4432). Strain TG202 was produced by replacing this region in TG201 with the PCR product from pLOI4151 (WM*ldhA*A/C primers). The *cat-sacB* cassette in TG202 was replaced with the PCR product from pLOI4432 (*ldhA*A/C primers) with sucrose selection for loss of sacB to produce TG203.

(0120) Primer sets (up*focA*/Mid*pflA* and IO-*ycaO*up/IO-mid*pflA*down) and corresponding plasmids (pLOI4415 and pLOI4421) used to remove the FRT site in Δ(*focA-pflB*) are included in Table 7. Strain TG204 was produced by replacing this region in TG203 with the PCR product from pLOI4151 (WM*pflB*A/C primers). The *cat-sacB* cassette in TG204 was replaced with the PCR product from pLOI4421 (up*focA*/Mid*pflA* primers) with sucrose selection for loss of *sacB* to produce KJ091. KJ091 is a derivative of KJ073 in which all FRT sites have been removed from the Δ*adhE*, Δ*ldhA* and Δ*focA-pflB* regions of the chromosome.

Removal of FRT site in *ackA* region and construction of *citF*, *sfcA*, and *pta-ackA* gene deletions

(0121) To eliminate the FRT site in the *ackA* region of KJ073, plasmids containing sequences of the desired mutation were constructed as follows. *E. coli* C genomic DNA was used as the template for PCR amplification of *ackA* with the JM*ackA*F1/R1 primers that bind approximately 200 bp upstream and downstream of the *ackA* gene. The linear product was cloned into pCR2.1-TOPO (Invitrogen, Carlsbad, CA) to produce pLOI4158. Plasmid pLOI4158 was then used as a template for inside-out PCR with JM*ackA*up1/down1 primers and *Pfu* polymerase to yield a blunt-ended product that lacks an 808-bp internal segment of *ackA*. The *Pac*I-flanked *cat-sacB* cassette (*Sma*I/*Sfo*I fragment from pLOI4162) was then

ligated into the blunt PCR product to produce pLOI4159. Plasmid pLOI4159 served as a template for PCR amplification (JM*ackA*F1/R1 primers). This PCR product was used to replace the FRT site in the *ackA* region of KJ073 by double-crossover homologous recombination, with selection for chloramphenicol resistance. The resulting clone was designated KJ076.

(0122) Plasmid pLOI4159 was also digested with *Pac*I to remove the *cat-sacB* cassette and self-ligated to produce pLOI4160, retaining the 18-bp translational stop sequence. Plasmid pLOI4160 served as a PCR template (JM*ackA*F1/R1 primers). This amplified fragment was used to replace the *cat-sacB* cassette in KJ076 by double-crossover homologous recombination with selection for loss of *sacB*. After removal of pKD46 by growth at elevated temperature, the resulting strain was designated KJ079. In this strain, the deleted region has been replaced by the 18-bp translational stop sequence.

(0123) The strategy used above to remove the FRT site from the *ackA* region was employed to make sequential deletions of *citF*, *sfcA* and *pta-ackA* and to replace the deleted regions with the 18-bp translational stop sequence. Additional primer sets (*citF*up/down and *citF*2/3) used to construct the *citF* deletion are included in Table 7 together with the corresponding plasmids (pLOI4629, pLOI4630, and pLOI4631). The resulting strain was designated KJ104.

(0124) The *sfcA* gene was deleted from strains KJ104 and KJ110, resulting in strains designated KJ119 and KJ122, respectively. Additional primer sets (*sfcA*up/down and *sfcA*1/2) used to construct the *sfcA* deletions are included in Table 7 together with the corresponding plasmids (pLOI4283, pLOI4284, and pLOI4285).

(0125) The *ackA-pta* operon (including the synthetic translational stop sequence) was deleted from KJ122 to produce strain KJ134. Additional primer sets (*ackA*up/*pta*down and *ackA*2/*pta*2) used to construct this deletion are included in Table 7 together with the corresponding plasmids (pLOI4710, pLOI4711, and pLOI4712). Strain KJ134 does not contain any FRT sites or foreign genes.

Construction of pLOI4162 containing a *cat-sacB* cassette for markerless gene deletions

(0126) To facilitate the sequential deletion of chromosomal DNA, plasmid pLOI4162 (Fig. 8) was constructed with a removable *cat-sacB* cassette and the option to include an 18-bp segment of synthetic DNA with stop codons in all reading frames. This plasmid is composed of synthetic sequences and parts of plasmids pLOI2228 (Martinez-Morales *et al.*, 1999), pLOI2511 (Underwood *et al.*, 2002), and pEL04 (Lee *et al.*, 2001; Thomason et al, 2005). Using pEL04 as a template, inside-out PCR was performed with the JMpEL04F1/R1 primers to eliminate unwanted *Sma*I and *BamH*I sites between the *cat* and *sacB* genes. The amplified product was digested with *Bgl*II (within both primers) and self-ligated to produce pLOI4152. Plasmid pLOI4131 was constructed by ligation of the FRT-*cat*-FRT fragment (Klenow-treated *Ban*I, *Cla*I) from pLOI2228 into compatible sites of pLOI2511 (Klenow-treated *Nhe*I, *Cla*I). Plasmid pLOI4131 was subsequently digested with *EcoR*I and self-ligated to remove the FRT-*cat*-FRT fragment to produce pLOI4145, retaining single *Kas*I and *Xma*I sites. A polylinker segment (SfPBXPS) was prepared by annealing complementary oligonucleotides (SfPBXPSsense and SfPBXPScomp). After digestion with *Kas*I and *Xma*I, this segment was ligated into corresponding sites of pLOI4145 to produce pLOI4153. The modified *cat-sacB* cassette in pLOI4152 was amplified by PCR using the JM*catsacB*up3/down3 primer set. After digestion with *Bam*HI and *Xho*I, this cassette was ligated into corresponding sites of pLOI4153 to produce pLOI4146. To create an 18-bp region (5'GCCTAATTAATTAATCCC3') (SEQ ID NO: 1) with stop codons in all six reading frames, pLOI4146 was digested with *Pac*I and self-ligated to produce pLOI4154 (not shown), removing the *cat-sacB* cassette. Two additional bases (T and A) were inserted between the *Sfo*I and *Pac*I sites of pLOI4154 using mutagenic primers (JM4161sense/comp) and linear plasmid amplification to produce pLOI4161. Finally, the *Pac*I digested fragment from pLOI4146 containing the *cat-sacB* cassette was ligated into the *Pac*I-digested site of pLOI4161 to produce pLOI4162 (GenBank accession EU531506).

Deletion of *mgsA* and *poxB* genes

(0127) A modified method was developed to delete *E. coli* chromosomal genes using a two-step homologous recombination process (Thomason *et al.*, 2005). With this method, no antibiotic genes or scar sequences remain on the chromosome after gene deletion. In the first recombination, part of the target gene was replaced by a DNA cassette containing a

chloramphenicol resistance gene (*cat*) and a levansucrase gene (*sacB*). In the second recombination, the *cat-sacB* cassette was replaced with native sequences omitting the region of deletion. Cells containing the *sacB* gene accumulate levan during incubation with sucrose and are killed. Surviving recombinants are highly enriched for loss of the *cat-sacB* cassette.

(0128) A cassette was constructed to facilitate gene deletions. The *cat-sacB* region was amplified from pEL04 (Lee *et al.*, 2001; Thomason *et al.*, 2005) by PCR using the JM*catsacB* primer set (Table 3), digested with *Nhe*I, and ligated into the corresponding site of pLOI3421 to produce pLOI4151. The *cat-sacB* cassette was amplified by PCR using pLOI4151 (template) and the *cat*-up2/*sacB*-down2 primer set (*EcoR*V site included in each primer), digested with *EcoR*V, and used in subsequent ligations.

(0129) The *mgsA* gene and neighboring 500 bp regions (*yccT'-mgsA-helD'*, 1435 bp) were amplified using primer set *mgsA*-up/down and cloned into the pCR2.1-TOPO vector (Invitrogen) to produce plasmid pLOI4228. A 1000-fold diluted preparation of this plasmid DNA served as a template for inside-out amplification using the *mgsA*-1/2 primer set (both primers within the *mgsA* gene and facing outward). The resulting 4958 bp fragment containing the replicon was ligated to the amplified, *EcoR*V-digested *cat-sacB* cassette from pLOI4151 to produce pLOI4229. This 4958 bp fragment was also used to construct a second plasmid, pLOI4230 (phosphorylation and self-ligation). In pLOI4230, the central region of *mgsA* is absent (*yccT'-mgsA'-mgsA'' – hel).*

(0130) After digestion of pLOI4229 and pLOI4230 with *Xmn*I (within the vector), each served as a template for amplification using the *mgsA*-up/down primer set to produce the linear DNA fragments for integration step I (*yccT'-mgsA'- cat-sacB- mgsA''- helD'*) and step II (*yccT'-mgsA'-mgsA''- helD'*), respectively. After electroporation of the step I fragment into KJ060 containing pKD46 (Red recombinase) and 2 h of incubation at 30°C to allow expression and segregation, recombinants were selected for chloramphenicol (40 mg l^{-1}) and ampicillin (20 mg l^{-1}) resistance on plates (30°C, 18 h). Three clones were chosen, grown in Luria broth with ampicillin and 5% w/v arabinose, and prepared for electroporation. After electroporation with the step II fragment, cells were incubated at 37°C for 4 h and transferred into a 250-ml flask containing 100 ml of modified LB (100 mM MOPS buffer added and

NaCl omitted) containing 10% sucrose. After overnight incubation (37°C), clones were selected on modified LB plates (no NaCl; 100 mM MOPS added) containing 6% sucrose (39°C, 16 h). Resulting clones were tested for loss of ampicillin and chloramphenicol resistance. Construction was further confirmed by PCR analysis. A clone lacking the *mgsA* gene was selected and designated KJ070.

(0131) The *poxB* gene was deleted from KJ071 in a manner analogous to that used to delete the *mgsA* gene. Additional primer sets (*poxB*-up/down and *poxB*-1/2) used to construct the *poxB* deletion are included in Table 3 together with the corresponding plasmids (pLOI4274, pLOI4275, and pLOI4276). The resulting strain was designated KJ072.

Construction of gene deletions in *tdcDE*, and *aspC*

(0132) The *tdcDE* gene and neighboring 1000 bp regions (*tdcG'-tdcFED-tdcC'*, 5325 bp) were amplified using *tdcDE*up/down primers and cloned into the pCR2.1-TOPO vector to produce plasmid pLOI4515. A 1000-fold diluted preparation of this plasmid DNA served as a template for inside-out amplification using the *tdcDE*F7/R7 primers (both primers within the *tdcDE* gene and facing outward). The resulting 6861 bp fragment containing the replicon was ligated to the amplified, *SmaI/Sfo*I-digested *cat-sacB* cassette from pLOI4162 (JM*catsacB*up3/down3 primers) to produce pLOI4516. This 6861 bp fragment was also used to construct a second plasmid, pLOI4517 (kinase treated, self-ligation) containing a deletion of *tcdD* and *tdcE*. The PCR fragments amplified from pLOI4516 and pLOI4517 (*tdcDE*up/down primers) were used to replace *tdcDE* region in KJ091. The resulting clones were tested for loss of ampicillin and chloramphenicol resistance.

(0133) The *aspC* gene was deleted from KJ104 in a manner analogous to that used to delete the *tdcDE* gene. Additional primer sets (*aspC*up/down and *aspC*1/2) used to construct the *aspC* deletion are included in Table 7 together with the corresponding plasmids (pLOI4280, pLOI4281, and pLOI4282). The resulting strain was designated KJ110. Neither KJ098, nor KJ110 contain any intervening sequence within the respective deleted regions (*tdcDE* and *aspC*).

Construction of gene deletions in *gldA* and *dhaKLM*

(0134) The *E. coli* strains XZ464, XZ465, and XZ466 were derived from the *E. coli* strain XZ632 using the genetic methods described above. The relevant characteristic of the *E. coli* strains XZ464, XZ465, XZ466 and XZ632 are provided in the Table 15.

Real time RT-PCR analysis

(0135) Real time RT-PCR was used to measure message RNA levels as described previously (Jarboe *et al.*, 2008). Cells were grown in NBS medium with 5% or 10% glucose and harvested during mid-log growth by swirling in a dry ice/ethanol bath, followed by centrifugation and storage at -80°C in RNALater (Qiagen, Valencia CA) until purification. RNA purification was performed with RNeasy Mini columns (Qiagen), followed by digestion with DNaseI (Invitrogen). Reverse transcription with Superscript II (Invitrogen, Carlsbad CA) used 50 ng total RNA as template. Real-time PCR was performed in a Bio-Rad iCycler with SYBR Green RT-PCR mix (Bio-Rad, Hercules CA). RNA was checked for genomic DNA contamination by running a RT-PCR in the absence of reverse transcription. Transcript abundance was estimated using genomic DNA as a standard and expression levels were normalized by the *birA* gene, a transcriptional repressor (Jarboe *et al.*, 2008). RT-PCR primers used for *pck* and *birA* are listed in Table 3.

Sequencing of *pck* region

(0136) In order to know whether there was any mutation in the *pck* gene of KJ073, the coding region and promoter region (about 800 bp in front of coding region) of *pck* gene in both KJ012 and KJ073 were amplified by PfuUltra High Fidelity DNA Polymerase (Stratagene; Wilmington, DE). Primer set *pck*-F/R was used to amplify the coding region through the transcriptional terminator (Table 9). Primer set *pck*-2/3 was used to amplify the promoter region. DNA sequencing was provided by the University of Florida Interdisciplinary Center for Biotechnology Research (with Applied Biosystems autosequencers).

Metabolic evolution

(0137) Cells from the pH controlled fermentations were serially transferred at 24 hours to encourage metabolic evolution though growth-based selection. The inoculum, approximately

1/100 of the volume of new media, was added directly to pre-warmed, fresh media to a starting OD_{550} of 0.05. Clones with improved fermentation characteristics were isolated. The metabolic evolution strategy was applied to improve yield of succinate production.

Analyses

(0138) Cell growth: Cell mass was estimated from the optical density at 550 nm (OD 1.0 = 333 mg of cell dry weight l^{-1}) with a Bausch & Lomb Spectronic 70 spectrophotometer.

(0139) The production of the organic acid by the genetically engineered microorganism can be confirmed and quantified by using appropriate techniques well known in the art. For example, HPLC techniques can be used to measure the quantity of the organic acid produced by the selected clone. The HPLC technology is also helpful in determining the purity of the organic acid produced by the selected clones. Organic acids and sugars were determined by using high performance liquid chromatography (Grabar *et al.*, 2006; Zhang *et al.*, 2007).

(0140) Succinic acid and other organic acids present in the fermentation broth were analyzed on Agilent 1200 HPLC apparatus with BioRad Aminex HPX-87H column. BioRad Microguard Cation H^+ was used as a guard column. The standards for HPLC analysis were prepared in 0.008N sulfuric acid. The HPLC column temperature was maintained at 50°C. Sulfuric acid at 0.008N concentration was used as a mobile phase at the flow rate of 0.6 ml/min. Quantification of various components was done by measuring their absorption at 210 nm.

Enzyme assay

(0141) Cells were harvested by centrifugation (8,000 x g for 5 min at 4°C) during mid-log growth, washed with cold 100 mM Tris-HCl (pH 7.0) buffer, and resuspended in the same buffer (1 ml). After cellular disruption using a Fast Prep-24 (MP Biomedicals, Solon, OH), preparations were clarified by centrifugation (13,000 x g for 15 min). Protein was measured by the BCA method (Pierce, Rockford, IL) using bovine serum albumin as a standard.

(0142) PEP carboxylase activity was measured as described by Canovas and Kornberg (1969). The reaction mixture contained 100 mM Tris-HCl buffer (pH8.0), 10 mM $MgCl_2$, 1

mM DTT, 25 mM $NaHCO_3$, 0.2 mM NADH, 20 U malate dehydrogenase, and crude extract. The assay mixture was incubated for 15 min at 37°C to activate the enzyme, after which the reaction was started by addition of 10 mM PEP.

(0143) PEP carboxykinase activity was measured as described by Van der Werf *et al*, (1997). The reaction mixture contained 100 mM MES buffer (pH6.6), 10 mM $MgCl_2$, 75 mM $NaHCO_3$, 5 mM $MnCl_2$, 50 mM ADP, 1 mM DTT, 0.2 mM NADH, 20 U malate dehydrogenase, and crude extract. The reaction was started by addition of 10 mM PEP.

(0144) Malic enzyme activity (carboxylation direction) was measured as described by Stols and Donnelly (1997). The reaction mixture contained 100 mM Tris-HCl buffer (pH7.5), 25 mM $NaHCO_3$, 1 mM $MnCl_2$, 1 mM DTT, 0.2 mM NADH, and crude extract. The reaction was started by addition of 25 mM pyruvate. This assay method was unsuitable for measurement of SfcA activity in wild type *E. coli* due to presence of lactate dehydrogenase.

(0145) The activity of ß-galactosidase was measured as described by Miller (1992). In all assays, one unit of activity represents the amount of enzyme required to oxidize or reduce 1 nmol of substrate per minute.

Example 1

Construction of KJ073 strain for succinate production

(0146) In the construction of a strain suitable for succinic acid production in minimal medium containing 5% glucose, both a rational design approach and metabolic evolution were followed. By inspection of Fig. 1 illustrating the generally accepted standard fermentation pathways for *E. coli*, a rational design for the metabolic engineering of strains producing succinate was devised in which insertional inactivations were made in genes encoding the terminal steps for all alternative products: lactate (*ldhA*), ethanol (*adhE*) and acetate (*ackA*). Results from this metabolic engineering by rational design were completely unexpected. The KJ012 (Δ*ldhA*::FRT Δ*adhE*::FRT Δ*ackA*::FRT) strain resulting from the insertional inactivation of *ldhA*, *adhE* and *ack* genes grew very poorly under anaerobic conditions in mineral salts medium containing 5% glucose (278 mM) and produced acetate instead of succinate as the primary fermentation product. Counter to expectations from rational design, succinate remained as a minor product. Molar yields of succinate based on

metabolized glucose were unchanged as a result of these mutations. The succinate yield was found to be 0.2 mol succinate per mol glucose both for the parent and KJ012 strains during fermentation in NBS mineral salts medium containing 5% glucose. We confirmed that NBS mineral salts medium contains all mineral nutrients needed for the growth of KJ012 by incubating under aerobic conditions (aerobic shaken flask; 5% glucose). In aerobic shaken flasks, cell yields for KJ012 were 5-fold higher than during anaerobic growth and 75% that of the *E. coli* C (parent) during anaerobic growth. These results also confirmed that all central biosynthetic pathways remain functional in KJ012.

(0147) When complex nutrients were present (Luria broth), fermentative succinate production by KJ012 increased 20-fold as compared to KJ012 in minimal salts medium and the molar yield for succinate increased by 3.5-fold. Clearly, rational design based on primary pathways is better suited to academic demonstrations or to design processes intended for use with complex nutrients.

(0148) The basis for the poor growth, poor succinate production, and the increase in acetate production by KJ012 during anaerobic metabolism in mineral salts medium is unknown. These are unexpected consequences that resulted from metabolic engineering using rational design based on standard pathway charts. In minimal medium, rationale designs for metabolic engineering clearly are not predictable. The resulting strain, KJ012, was inferior to the parent in growth and no better than the parent for succinate production. KJ012(Δ*ldhA*::FRT Δ*adhE*::FRT Δ*ackA*::FRT) grew poorly in comparison to the parent *E. coli* C, exhibited lower rates of succinate production, and provide no better molar yields (Table 4).

(0149) Despite these results, serial transfer of this strain was tried as a method to co-select improved growth and succinate production based on the following rationale. The primary pathway for the glucose fermentation into succinate (Fig. 1A and Fig. 2A) is generally thought to use phosphoenolpyruvate carboxylase (*ppc*) for the carboxylation step (Unden and Kleefeld, 2004; Fraenkel 1996; Keseler *et al.*, 2005; Millard *et al.*, 1996; Gottschalk, 1985; Karp *et al.*, 2007). This carboxylating enzyme does not conserve the high energy phosphate in phosphoenolpyruvate and reduces the net ATP available for growth. Alternative pathways for succinate production can be envisioned using the repertoire of known *E. coli* genes that could increase ATP yields and could thereby increase growth (Fig. 1A; Fig. 2B, 2C and 2D).

However, none of these alternative routes has been shown to function for succinate production during fermentation with native strains of *E. coli*. Key enzymes in these alternative routes are repressed by glucose and are normally active during gluconeogenesis. Typically, levels of these gluconeogenic enzymes vary inversely with the availability of glucose and other metabolites (Goldie and Sanwal, 1980a; Wright and Sanwal, 1969; Sanwal and Smando, 1969a) and function in the reverse direction, decarboxylation (Keseler *et al.*, 2005; Oh *et al.*, 2002; Kao *et al.*, 2005; Stols and Donnelly, 1997; Samuelov *et al.*, 1991; Sanwal, 1970a; Delbaere *et al.*, 2004; Goldie and Sanwal, 1980b; Sanwal and Smando, 1969b; Sanwal 1970b).

(0150) The key enzyme for one of these, NADH-linked malic enzyme (*sfcA*) (Fig. 2B), has been demonstrated to be capable of increasing succinate production in *E. coli* but required overexpression from a plasmid to do so (Stols and Donnelly, 1997). However, none of these alternative pathways would be expected to be functional in native strains of *E. coli* or KJ012 during anaerobic growth with high levels of glucose. Serial transfers of KJ012 with selection for improved growth offered an opportunity to select for mutational activation of alternative routes for succinate production (Fig. 1B) that maintained redox balance and increased ATP yields.

(0151) Metabolic evolution of KJ012 was carried out by sequentially subculturing under various regimens using small, pH-controlled fermentors to improve the growth. KJ012 was serially transferred in NBS glucose medium under fermentative conditions as rapidly as growth permitted (Fig. 3A; Fig. 4A; Fig. 5). Growth remained slow with 4-5 days of required incubation between the first 9 transfers, then dramatically increased allowing transfers to be made at 24 h-intervals. This event was accompanied by an increase in acetate (Fig 4A) with little improvement in succinate production. After 27 transfer (60 days), KOH was replaced with a 1:1 mixture of 3M K_2CO_3 and 6N KOH to provide additional carbon dioxide (100 mM initially added to all NBS mineral salts medium). Continuing transfers led to improvements in succinate production. A total of 40 transfers were made in 5% glucose (227 mM), followed by another 40 transfers in 10% glucose (555 mM). During the transfers in 10% glucose, succinate yields remained approximately 0.7 mol per mole of glucose metabolized with lactate, acetate, and formate as co-products (Table 4). This yield was 3-fold higher than

E. coli C and KJ012 strains. A clone was isolated and designated KJ017. Selection for improvements in the growth of KJ012 to produce KJ017 co-selected for improvements in succinate production (rate, titer, and molar yield).

(0152) Succinate produced by *E. coli* using the pathway generally regarded as the native fermentation pathway based on phosphoenolpyruvate carboxylase (*ppc*) wastes the energy of phosphoenolpyruvate by producing inorganic phosphate. One ATP is lost per succinate produced by this pathway (Fig. 1; Fig. 6). Conserving this energy as ATP by using alternative enzyme systems represents an opportunity to increase cell growth and co-select for increased succinate production. Based on known genes in *E. coli*, three other enzyme routes for succinate production were envisioned that would conserve ATP and could thereby increase growth (Fig. 1; Fig. 6). However, all carboxylation steps in these alternative routes are thought to function in the reverse direction (decarboxylation) primarily for gluconeogenesis during growth on substrates such as organic acids (Keseler *et al.*, 2005; Oh *et al.*, 2002; Kao *et al.*, 2005; Stols and Donnelly, 1997; Samuelov *et al.*, 1991; Sanwal, 1970a; Delbaere *et al.*, 2004; Goldie and Sanwal, 1980b; Sanwal and Smando, 1969b; Sanwal, 1970b). To test the hypothesis that growth-based selection to develop KJ017 has indeed activated one or more of these alternative routes, the activities of the four carboxylation enzymes were compared in wild type strain ATCC 8739, KJ012 (deleted in primary fermentation pathways), and the metabolically evolved strain KJ017 (Table 5). PEP carboxylase activities were similarly low in all, 20 to 30 U (mg protein)$^{-1}$. NADH-linked malic enzyme activity (SfcA; carboxylation direction) was also low and NADPH-linked malic enzyme activity (MaeB; carboxylation direction) was undetectable. In contrast, PEP carboxykinase activity was increased by 4-fold in KJ017 as compared to KJ012, consistent with a hypothesis that selection for improved growth would co-select for increased production of succinate through increasing ATP yields, a consequence of increasing the expression of an energy-conserving route for succinate production.

(0153) Further growth-based selections and additional gene deletions were used to construct many additional strains with further improvements in growth and succinate production (Fig. 3 and Fig. 4).

(0154) During growth with 10% (w/v) glucose, unwanted co-products (acetate, formate, and lactate) were abundant in fermentations with KJ017 (Δ*ldhA*::FRT Δ*adhE*::FRT Δ*ackA*::FRT) despite the deletion of genes encoding the primary lactate dehydrogenase (*ldhA*) and acetate kinase (*ackA*) activities (Table 4). Production of lactate and acetate could also result in higher ATP yields, a basis for growth-based selection (Fig. 1A).

(0155) The gene encoding pyruvate formatelyase (*pflB*) was deleted from KJ017 to eliminate the loss of reductant as formate and an excess acetyl~CoA, a potential source of acetate. The upstream formate transporter (*focA*) in this operon was also deleted. As expected, this deleted strain (KJ032) did not grow without acetate confirming that this is the primary route for acetyl~CoA production in KJ017 (Fig. 3C). Deletion of *pflB* is well-known to cause acetate auxotrophy under anaerobic conditions (Sawers and Bock, 1988). Growth and succinate production by KJ032 were restored by the addition of 20 mM acetate (Fig. 3C, Fig. 4C, and Fig. 5). Production of formate and acetate were substantially reduced as a result of *pflB* (and *focA*) deletion. Although this strain required acetate for growth, additional acetate was also produced during fermentation. The same phenomenon was previously reported for *pflB*-deleted strains during the construction of *E. coli* K-12 biocatalysts for pyruvate production (Causey et al., 2004). Lactate levels were also reduced in KJ032 (Table 4; Fig. 4C). Subsequent transfers were accompanied by improvements in growth and succinate production. Added acetate was reduced, inocula size was reduced, and glucose concentration was doubled (10% w/v) during subsequent transfers (Figs. 3D and 4D). After further transfers, acetate was omitted and a strain was developed that was no longer auxotrophic for acetate, presumably due to increased expression of another gene. However, succinate yields declined upon elimination of added acetate while malate and acetate levels increased. A clone was isolated from the last transfer and designated, KJ060 (Δ*ldhA*::FRT Δ*adhE*::FRT Δ*ackA*::FRT Δ*focA-pflB*::FRT). This strain produced 1 mole of succinate per mole of glucose metabolized in NBS mineral salts medium with 10% glucose.

(0156) The small amount of lactate present in the fermentation broths of various strains is presumed to originate from the methylglyoxal synthase pathway (Fig. 6; Grabar *et al.*, 2006). Although this represents a small loss of yield, lactate production by this pathway is indicative

of methylglyoxal accumulation, an inhibitor of both growth and glycolysis (Egyud and Szent-Gyorgyi, 1966; Grabar *et al.*, 2006; Hopper and Cooper, 1971). Production of methylglyoxal and lactate were eliminated by deleting the *mgsA* gene (methylglyoxal synthase) in KJ060 to produce KJ070 (Δ*ldhA*::FRT Δ*adhE*::FRT Δ*ackA*::FRT Δ*focA-pflB*::FRT Δ*mgsA*). Strain KJ070 was initially subcultured in 5% (w/v) glucose (Fig. 3E, Fig. 4E, and Fig. 5). Deletion of *mgsA* is presumed to have increased glycolytic flux as evidenced by the accumulation of pyruvate in the medium (Table 4). This increase in glycolytic flux may also be responsible for the further decline in the succinate/malate ratio due to increased production of oxaloacetate, an allosteric inhibitor of fumarate reductase (*Iverson* et al., 2002; Sanwal, 1970c).

(0157) At transfer 21, glucose was doubled to 10% (w/v) and transfers continued. This higher level of glucose and subsequent transfers resulted in a new strain that was isolated from the final subculture and designated KJ071 (Δ*ldhA*::FRT Δ*adhE*::FRT Δ*ackA*::FRT Δ*focA-pflB*::FRT Δ*mgsA*). This strain may be useful for malate production.

(0158) Although conversion of glucose to acetate is redox neutral, partitioning of carbon to acetate decreases the yield of succinate and malate. Pyruvate oxidase (*poxB*) represents a potential source of acetate and CO_2 during incubation under microaerophilic conditions (Causey *et al.*, 2004). Although it should not function to oxidize pyruvate under anaerobic condition, *poxB* was targeted for gene deletion (Fig. 6). As expected, deletion of *poxB* to produce KJ072 (Δ*ldhA*::FRT Δ*adhE*::FRT Δ*ackA*::FRT Δ*focA-pflB*::FRT Δ*mgsA* Δ*poxB*) did not reduce acetate production indicating that alternative pathways are involved in acetate production. However, eliminating *poxB* resulted in unexpected changes in fermentation products, including an increase in succinate (Table 4; Fig. 4F). The mechanism for this improvement in succinate production is unknown but may be related to other activities of pyruvate oxidase such as acetoin production, decarboxylation, and carboligation (Ajl and Werkman, 1948; Chang and Cronan, 2000).

(0159) Strain KJ072 was subjected to 40 further rounds of metabolic evolution in AM1 medium, a lower salt medium, with 10% (w/v) glucose (Table 4; Fig. 3F, Fig. 4F, and Fig. 5). Improvements in growth, cell yield and succinate production were observed during these

transfers. Malate, pyruvate and acetate levels also increased. A clone was isolated from the final transfer and designated KJ073 (Δ*ldhA*::FRT Δ*adhE*::FRT Δ*ackA*::FRT Δ*pflB*::FRT Δ*mgsA* Δ*poxB*).

(0160) The KJ073 strain retained the phosphoenolpyruvate carboxykinase route for carboxylation (Table 5). In vitro activity of this strain was 45-fold higher than that of KJ012 and 10-fold higher than KJ017 providing further evidence for the tight coupling of energy conservation to succinate production and growth and further establishing the basis used for selection.

(0161) Large increases in PEP carboxykinase activity were well correlated with improvements in cell yield and succinate production (Table 5). From KJ012 to KJ017 (metabolic evolution), PEP carboxykinase activity increased 4.3-fold, cell yield increased 5.7-fold, and succinate production increased 8.8-fold. From KJ017 to KJ060 (deletion of *pflB* followed by metabolic evolution), PEP carboxykinase activity increased 12-fold, cell yield increased 1.3-fold, and succinate production increased 2.6-fold. From KJ060 to KJ071 (deletion of *mgsA* followed by metabolic evolution), PEP carboxykinase activity decreased by 92%, cell yield decreased by 45%, and succinate production decreased by 63%. From KJ071 to KJ073 (deletion of *poxB* followed by metabolic evolution), PEP carboxykinase activity, cell yield, and succinate production were restored to levels equivalent to KJ060.

(0162) The *pck* and surrounding regions were cloned from KJ012 and KJ073, and sequenced. No changes were found in the coding region. Absent post-translational modifications, the catalytic properties of the enzyme should be unchanged. A single mutation was detected in the *pck* promoter region, G to A at -64 bp site relative to the translation start site. This mutation was behind the transcription start site which is -139 bp site relative to the translational start site.

(0163) Previous investigators have noted that the kinetic parameters of phosphoenolpyruvate carboxylase (*ppc*) and phosphoenolpyruvate carboxykinase (*pck*) may have important effects on carboxylation and succinate production (Millard *et al.*, 1996; Kim *et al.*, 2004). The Km towards bicarbonate for *E. coli* phosphoenolpyruvate carboxylase (*ppc*) is 0.15 mM (Morikawa *et al.*, 1980), 9-fold lower (13 mM) than *E. coli* phosphoenolpyruvate

carboxykinase (*pck*) (Krebs and Bridger, 1980). Although overexpressing *pck* from *E. coli* using multi-copy plasmid increased phosphoenolpyruvate carboxykinase activity by 50-fold, it was reported to have no effect on succinate production (Millard *et al.*, 1996). Succinate production was also not increased when phosphoenolpyruvate carboxykinase from *Anaerobiospirillum succiniciproducens* was overexpressed in *E. coli* K12 (Kim *et al.*, 2004). This enzyme also has a high Km (30 mM) for bicarbonate (Laivenieks *et al.*, 1997). However, when *A. succiniciproducens pck* was overexpressed in a *ppc* mutant of *E. coli* K12, succinate production was increased 6.5-fold (Kim *et al.*, 2004). In KJ017 and subsequent derivatives, phosphoenolpyruvate carboxykinase is clearly the dominant carboxylating activity even in the presence of functional native phosphoenolpyruvate carboxylase.

(0164) Results from enzyme measurements of *E. coli* C were quite surprising. The enzyme generally regarded as the dominant carboxylating activity for succinate production by native *E. coli* (phosphoenolpyruvate carboxylase; *ppc*) during growth (Unden and Kleefeld, 2004; Fraenkel, 1996; Keseler et al, 2005; Millard *et al.*, 1996; Gottschalk, 1985; Karp *et al.*, 2007) was not the most active enzyme *in vitro* for E. *coli* C. Thus the generally accepted metabolic pathways for *E. coli* (Unden and Kleefeld, 2004; Fraenkel, 1996; Sanchez *et al.*, 2006; Cox *et al.*, 2006; Vemuri *et al.*, 2002a; Wang *et al.*, 2006; Sanchez *et al.*, 2005ab; Gokarn *et al.*, 2000; Karp *et al.*, 2007) upon which rational design of metabolic engineering and estimates of metabolic flux are typically based may not accurately reflect metabolism in all strains. Under substrate-saturating conditions *in vitro*, phosphoenolpyruvate carboxykinase activity was the most active. In *E. coli* K12, activities for both phosphoenolpyruvate carboxylase and phosphoenolpyruvate carboxykinase were reported to be equal *in vitro* (140 nm min^{-1} mg^{-1} cell protein; Van der Werf *et al.*, 1997) with the former serving as the primary route to succinate.

(0165) Previous studies showed that the overexpression of a native *ppc* gene in *E. coli* resulted in higher specific succinate production (Millard *et al.*, 2000), higher specific growth rate, and lower specific acetate production due to more carboxylation of PEP to replenish TCA cycle intermediates (Farmer and Liao, 1997). However, since PEP is required for the glucose transport system, overexpressing *ppc* also decreases the glucose uptake rate by 15-40% without significantly increasing succinate yield (per glucose) as compared to an isogenic

control (Chao and Liao, 1993; Gokarn *et al.*, 2000). This failure of the native phosphoenolpyruvate carboxylase to increase succinate yields diverted most research attention to a new metabolic design, over expression of the PYC (pyruvate carboxylase) from *Lactobacillus lactis* or *Rhizobium etli* as the carboxylating step (Vemuri *et al.*, 2002ab; Gokarn *et al.*, 2000; Lin *et al.*, 2005a, 2005b, 2005c) rather than pursuing further work with the native repertoire of *E. coli* genes.

(0166) Rumen bacteria such as *Actinobacillus succinogenes* produce succinate as a primary product during glucose fermentation using the energy conserving phosphoenolpyruvate carboxykinase for carboxylation (Kim et al., 2004; McKinlay *et al.*, 2005; McKinlay and Vieille, 2008). Reported activities for this organism are 5-fold those of KJ017 and half of that obtained by continued growth-based selection (metabolic evolution) of KJ073. Thus by using a combination of metabolic engineering (*ldhA adhE ackA*) and metabolic evolution (growth-based selection for increased efficiency of ATP production), the studies reported herein demonstrate the development of succinate-producing strains of *E. coli* that resemble a rumen organism such as *A. succinogenes* by using only the native repertoire of *E. coli* genes. Despite prior reports that over expression of the *E. coli* phosphoenolpyruvate carboxykinase (*pck*) is not helpful for succinate production in the absence of a mutation in phosphoenolpyruvate synthase (Chao and Liao, 1993; Kim *et al.*, 2004; Gokarn *et al.*, 2000; Millard *et al.*, 1996), KJ017 and derivatives have been metabolically evolved to use phosphoenolpyruvate carboxykinase as the primary route for succinate and malate production.

(0167) Fig. 7 shows batch fermentations with KJ060 and KJ073, the two best biocatalysts for succinate production. Although growth was completed within the initial 48 h of incubation, succinate production continued for 96 h. One-third of succinate production occurred in the absence of cell growth. These strains produced succinate titers of 668-733 mM, with a molar yield of 1.2-1.6 based on glucose metabolized. With AM1 medium, yields were typically higher than with NBS mineral salts medium. Acetate, malate, and pyruvate accumulated as undesirable co-products and detracted from the potential yield of succinate (Table 4). The maximum theoretical yield of succinate from glucose and CO_2 (excess) is 1.71 mol per mole glucose based on the following equation:

$C_6H_{12}O_6 + 6\ CO_2 \rightarrow 12\ C_4H_6O_4 + 6\ H_2O$. However, there is no direct succinate pathway in *E. coli* that achieves this yield (Fig. 6).

(0168) Although this study primarily focused on the conversion of glucose to succinate. It is well known that *E. coli* has the native ability to metabolize all hexose and pentose sugars that are constituents of plant cell walls (*Asghari et al.*, 1996; Underwood *et al.*, 2004). Some strains of *E. coli* can also metabolize sucrose (Moniruzzaman *et al.*, 1997). Strain KJ073 was tested for utilization of 2% sugars of hexoses and pentoses in serum tubes. In all cases, these sugars were converted primarily to succinate. Strain KJ073 also metabolized glycerol to succinate. During incubation with 2% glycerol, 143 mM glycerol was metabolized to produce 127 mM succinate with a molar yield of 0.89, or 89% of the maximum theoretical yield for succinic acid using glycerol as the carbon source for bacterial growth.

(0169) The fermentative metabolism of *E. coli* has been shown to be remarkably adaptable. Derivatives were engineered and evolved to circumvent numerous deletions of genes concerned with native fermentation pathways and increase fluxes through remaining enzymes to maintain redox balance, increase the efficiency of ATP production, and increase growth. Though much more challenging, cells can make such adaptive changes in mineral salts media while balancing carbon partitioning to provide all biosynthetic needs. After eliminating the primary routes for NADH oxidation (lactate dehydrogenase, alcohol dehydrogenase) and acetate production (acetate kinase), growth and ATP production remain linked to NADH oxidation and the production of malate or succinate for redox balance (Fig. 1B). Anaerobic growth-based selections ensure redox balance and select for increased efficiency and increased rates of ATP production, the basis for increased growth. This selection for redox balance and ATP production cannot readily distinguish between malate and succinate as end products, since the precursors of both serve as electron acceptors.

(0170) Deletion of *pflB*, the primary *source* of acetyl~CoA during anaerobic growth, resulted in an auxotrophic requirement for acetate (Sawers and Bock, 1988). This requirement was eliminated through metabolic evolution, presumably due to increased production of acetyl~CoA by other routes such as pyruvate dehydrogenase (de Graef *et al.*, 1999). The metabolic source of the acetate or acetyl~CoA that replaced this auxotrophic need is

unknown. The shift to higher succinate production after a *poxB* deletion was also surprising. Little change in the acetate level was observed indicating that either this enzyme was a minor source of acetate or that it was functionally replaced by other routes for acetate production. After deletion of *poxB*, succinate was again produced as the dominant dicarboxylic acid. This shift in metabolic products accompanying poxB deletion was unexpected.

(0171) With the best strains for succinate production, KJ060 and KJ073, malate and acetate remained as abundant co-products (Table 4; Figs. 4D and 4F). Elimination of these represents a further opportunity to increase yield.

(0172) All previously engineered *E. coli* developed for succinate production have used complex media and plasmids with antibiotics for maintenance. Most have achieved only low titers of succinate in simple batch fermentations, requiring more complex processes to achieve high titers (Table 1). Other investigators have also used heterologous genes and complicated processes that include sparging with gas (CO_2, H_2, O_2 or air) and dual aerobic and anaerobic process steps. A variety of genetic approaches have been reported that increase succinate production from glucose by recombinant *E. coli* in complex medium. This complexity of process and nutrients would be expected to increase the cost of construction, materials, purification, and waste disposal. Complex media containing vitamins, amino acids, and other macromolecular precursors may mask potential regulatory problems in metabolism and biosynthesis that were created by metabolic engineering. In our initial construct, growth and sugar metabolism were very poor in mineral salts medium but were very robust in complex medium (Luria broth). In contrast, strains KJ060 and KJ073 produced high titers of succinate (600-700 mM) in simple batch fermentations (10% sugar) using mineral salts medium without any complex nutrients or foreign genes.

Example 2

Construction of KJ134 strain for succinate production

(0173) As described in the Example 1 above, the central anaerobic fermentation genes in *E. coli* C wild type were sequentially deleted by the strategy of Datsenko & Wanner (2000) with PCR products and removable antibiotic markers (by using FRT recognition sites and FLP recombinase). These constructions in combination with metabolic evolution (growth-based

selection for increased efficiency of ATP production) were used to select for a mutant strain that recruited the energy-conserving, phosphoenolpyruvate carboxykinase (*pck*) to increase growth and succinate production (Fig. 9). The resulting strain, KJ073, produced 1.2 moles of succinate per mole of metabolized glucose (Jantama *et al.*, 2008a) and uses a succinate pathway quite analogous to the rumen bacteria, *Actinobacillus succinogenes* (van der Werf *et al.*, 1997) and *Mannheimia succiniciproducens* (Song *et al.*, 2007). However, methods used to construct these gene deletions left a single 82 to 85 nucleotide-long genetic scar or FRT site in the region of each deleted gene (*ackA, ldhA, adhE, ackA, focA-pflB*). These FRT sites served as recognition sites for FLP recombinase (Storici *et al.*, 1999) during removal of the antibiotic genes. All of these extraneous sequences were sequentially removed from KJ073 and replaced with native DNA with only the desired gene deletion using methods that have been described previously (Grabar *et al.*, 2006; Zhang *et al.*. 2007; Jantama *et al.*, 2008b). The resulting strain, KJ091, contains specific deletions in *ackA, ldhA, adhE, focA-pflB, ackA, mgsA,* and *poxB* and lacks all FRT sites present in the KJ073 strain. The KJ091 strain is devoid of all foreign and synthetic DNA except for an 18-bp translational stop sequence within *ackA*. Succinate production by strain KJ091 was equivalent to that of KJ073 strain (Table 8). This strain was used as the parent for further improvements in succinate production.

(0174) In the first step for further improvement of KJ091 strain, attention was paid to reduce the acetate production. During the anaerobic fermentation of glucose by *E. coli*, pyruvate formate-lyase (*pflB*) serves as the primary source of acetyl~CoA, the precursor of acetyl~P, and acetate kinase (*ackA*) serves as the primary route for acetate production from acetyl~P (Karp *et al.*, 2007; Kessler & Knappe, 1996). The abundance of acetate as a fermentation product in strains KJ073 and KJ091 was surprising since these strains contain deletions in both *ackA* and *pflB* (Fig. 9). This residual acetate at the end of fermentation represents a potential opportunity to further redirect metabolism for improved succinate yield.

(0175) A related enzyme with acetate kinase (and proprionate kinase) activity is encoded by *tdcD* but is typically produced only for the degradation of threonine (Hesslinger *et al.*, 1998; Reed *et al.*, 2003). It is possible that mutations occurring during selection have increased expression of *tdcD* as illustrated in Fig. 10. During anaerobic growth with 10% (w/v)

glucose, expression of *tdcD* could functionally replace *ackA*, increasing the production of acetate from acetyl~P. The adjacent *tdcE* gene in the same operon is similar to *pflB* and encodes a pyruvate (and a-ketobutyrate) formatelyase activity that is co-expressed during threonine degradation (Hesslinger *et al.*, 1998). It is possible that increased expression of this gene during anaerobic growth with 10% (w/v) glucose could increase the production of acetyl~CoA, the immediate precursor of acetyl~P, and waste reductant as formate (Fig. 10). Both *tdcD* and *tdcE* (adjacent) were simultaneously deleted from KJ091 to produce KJ098. Deletion of these two genes reduced acetate production by half and increased succinate yield by 10% in KJ098 in comparison to KJ091, establishing the importance of this unexpected pathway in diverting carbon flow away from succinate. The level of pyruvate produced by KJ098 also declined by 40%, an intermediate that would be predicted to increase upon elimination of two alternative routes for pyruvate metabolism, pyruvate formate-lyase activity (*tdcD*) and acetate kinase activity (*tdcE*). The significant decrease in the pyruvate production is an unexpected result of tdcD / tdcE gene deletions. The mechanisms responsible for the reduction in pyruvate, and the increase in succinate which resulted from the simultaneous deletion of *tdcD* and *tdcE* are unknown.

(0176) Although KJ098 represents a significant improvement over KJ091, further reduction in acetate levels and further increases in succinate yields may be possible. Under anaerobic conditions, oxaloacetate is partitioned into the reduced product (malate) and oxidized intermediate (citrate) (Fig. 9). Citrate can be converted back to oxaloacetate and acetate by citrate lyase (*citDEF*) to recycle the intracellular OAA pool (Fig. 10) for other metabolic functions (Nilekani *et al.*, 1983). Expression of significant amounts of citrate lyase is associated with growth on citrate (Lutgens and Gottschalk, 1980; Kulla and Gottschalk, 1977). Citrate lyase is a multi-enzyme complex made up of three different polypeptide chains. The a or large subunit is a citrate-ACP transferase that catalyzes the first step. The ß or medium subunit is a citryl-ACP lyase that catalyzes the second step. The γ or small subunit acts as an acyl-carrier protein and also carries the prosthetic group components. All three subunits are required for the reaction to take place (Quentmeier *et al.*, 1987). The deletion of genes encoding one or more of these subunits would eliminate citrate lyase activity and may further reduce the level of acetate during succinate production. The *citF* gene was deleted from KJ098 to produce KJ104. This deletion, however, had no effect on acetate production

or other succinate yield (Table 8). Since deletion of *citF* did not cause any reduction in acetate, this intermediate is presumed to arise from other pathways. For unknown reasons, deletion of *citF* adversely affected the growth of KJ104 (reduced cell yield by 22%) and increased the level of pyruvate at the end of fermentation by almost 50% in comparison to KJ098. However, the succinate yield, titer, average productivity, and acetate levels with KJ104 were comparable to those with KJ098 (Table 8).

(0177) Aspartate aminotransferase (*aspC*) is a multifunctional enzyme that catalyzes the synthesis of aspartate, phenylalanine and other compounds by transamination. In the reaction, L-aspartate is synthesized from oxaloacetate, an intermediate from PEP carboxylation, by a transamination reaction with L-glutamate. Besides being a protein constituent, aspartate participates in several other biosynthetic pathways. About 27 percent of the cellular nitrogen has been estimated to flow through aspartate (Reitzer, 2004). Aspartate biosynthesis and succinate production share a common intracellular pool of oxaloacetate. Deletion of *aspC* could lead to increased succinate production but may also create an auxotrophic requirement that prevents anaerobic growth in minimal salts medium such as AM1.

(0178) This aspartate aminotransferase gene (*aspC*) was deleted from KJ104 to produce KJ110. Unexpectedly, the deletion of *aspC* had no effect on succinate yield or cell yield in KJ110 as compared to KJ104 (Table 8). Thus aspartase does not appear to divert significant levels of oxaloacetate away from succinate production in our strain. Alternative enzymes appear to be available that replace the biosynthetic needs formerly catalyzed by aspartate aminotransferase.

(0179) Significant amounts of pyruvate are present at the end of fermentation with KJ104 and other strains of *E. coli* engineered for succinate production (Table 8). This pyruvate represents an unwanted product and a further opportunity to increase succinate yield. This high level of pyruvate in fermentation broth could result from the decarboxylation of malate to pyruvate by malic enzyme (*sfc*A) as illustrated in Fig. 10. This enzyme is thought to function primarily during gluconeogenesis (Unden and Kleefeld, 2004; Stols and Donnelly, 1997; Oh *et al.*, 2002) rather than during the anaerobic catabolism of glucose. Although reductive carboxylation of pyruvate to form malate is thermodynamically favored, the kinetic

parameters of this enzyme favor the dehydrogenation and decarboxylation under physiological conditions (Stols and Donnelly, 1997). Over-expression of this enzyme to carboxylate pyruvate has been previously used as a basis to construct strains for succinate production of *E. coli* (Stols and Donnelly 1997).

(0180) If malic enzyme (*sfcA*) is carboxylating in KJ104 (and related strains) and contributing to succinate production, deletion of this gene would be expected to reduce succinate yield and increase the levels of other products such as pyruvate. Alternatively, if malic enzyme (*scfA*) is decarboxylating in KJ104 and diverting malate to pyruvate, deleting the gene encoding this enzyme would be expected to increase succinate yields and decrease the levels of pyruvate. Unexpectedly, deletion of the *sfcA* gene from KJ104 to produce KJ119 had no measurable effect on succinate production, growth, pyruvate levels, etc (Table 8) in comparison to KJ104. These results clearly demonstrated that malic enzyme (*sfcA*) is unimportant for succinate production in KJ104 and related strains. This result is in sharp contrast to the succinate-producing strains developed by Stols and Donnelly (1997) in which increased production of malic enzyme was used as the primary route for succinate production.

(0181) Although no significant benefits were observed from either a *sfcA* deletion or an *aspC* deletion in KJ104, studies were carried out to test the effect of deleting both genes in combination. This was done by deleting the *sfcA* gene in KJ110 to produce KJ122 and expected to see no benefit. However, the combined deletion of both *sfcA* and *aspC* (strain KJ122) resulted in an unexpected increase in succinate yield and titer with a small reduction in acetate (Table 8), in comparison to the parent strain KJ110 and related strains (KJ104 and KJ119). The combined deletion (*aspC* and *sfcA*) in KJ122 resulted in 18% increases in succinate yield, 24% increase in succinate titer, and 24% increase average productivity as compared to KJ104. Although the mechanism is unknown, it is possible that single mutations were ineffective because they were compensated in part by increased flow through the remaining enzyme activity, malic enzyme or aspartate aminotransferase (Fig. 10), dampening any potential benefit. The increase in succinate yield and titer are presumed to result from an increase in the availability of oxaloacetate allowing a larger fraction to proceed to succinate. Malate levels also remained extremely low.

(0182) Strain KJ122 (Table 8) produced 1.5 mol succinate per mole of glucose, 88% of the maximum theoretical yield (1.71 mol per mol glucose). To produce this high level of succinate and fully reduce malate production, additional reductant was required. Although the source of this additional reductant is unknown, these results are consistent with an increase in pyruvate flow through pyruvate dehydrogenase. This enzyme is thought to function primarily during aerobic metabolism (Guest *et al.*, 1989) but has also been reported to function at low levels during fermentation (de Graef *et al.*, 1999).

(0183) KJ122 produced excellent succinate yields (1.5 mol mol^{-1} glucose) plus smaller amounts of acetate and pyruvate. The maximum theoretical yield for succinate is 1.71 mol mol^{-1} glucose and these 3-carbon intermediates represent an opportunity to further increase yield. Pyruvate is presumed to accumulate from glycolysis as a metabolic overflow and may be related to acetate accumulation. Acetyl~CoA is an allosteric regulator of many enzymes. The source of acetate and acetate kinase activity is unknown since genes encoding the two primary activities for acetate kinase (*tdcD* and *ackA*) have been deleted (Fig. 9 and Fig. 10). Assuming that the acetate is produced from acetyl~P, the product of phosphotransacetylase, a further deletion was constructed in KJ122 to inactivate the *pta* gene. The resulting strain, KJ134, produced near theoretical level of succinate (Table 8). In this strain, pyruvate and acetate levels were substantially reduced. Volumetric productivity was also reduced by 17%. Succinate yields with strain KJ134 are equal or better than all other strains regardless of the complexity of fermentation processes, media, or growth conditions.

Example 3

Recruitment of gluconeogenic PEP carboxykinase (pck) for succinate production

(0184) *E. coli* has the metabolic potential for four native carboxylation pathways that could be used to produce succinate (Figs. 2A–2D). The carboxylation of phosphoenolpyruvate (PEP) to oxaloacetate (OAA) by phosphoenolpyruvate carboxylase (*ppc*) is recognized as the primary pathway for the fermentative production of succinate in *E. coli* (Fig. 2A). (Fraenkel DG, 1996; Unden & Kleefeld, 2004; Karp *et al.*, 2007). This reaction is essentially irreversible due to the energy loss associated with the release of inorganic phosphate. The three other carboxylation reactions are normally repressed by high concentration of glucose

in the medium and also reported to function in the reverse direction for gluconeogenesis. (Samuelov *et al.*, 1991; Oh *et al.*, 2002; Stols & Donnelly, 1997). The second and third carboxylation pathways (Figs. 2B and 2C) use NADH-linked and NAPDH-linked malic enzymes (*sfcA* and *maeB*), respectively, to catalyze the reversible carboxylation of pyruvate to malate. Both pathways allow energy to be conserved as ATP during the pyruvate formation from PEP. The fourth pathway uses PEP carboxykinase (*pck*) for the reversible carboxylation of PEP to OAA with the conservation of energy as ATP (Fig. 2D). Although PEP carboxykinase (PCK) typically functions only during gluconeogenesis in *E. coli*, an analogous PEP carboxykinase is present at very high levels in succinate-producing rumen bacteria where it serves as the primary PEP carboxylation activity. (Van der Werf *et al.*,1997).

(0185) *E. coli* strain KJ073 was metabolically engineered by both targeted gene deletion and evolution for high growth rate and succinate production. (Jantama *et al.*,2008a). Genes encoding each of the four carboxylating enzymes (*ppc*, *sfcA*, *maeB* and *pck*) were individually deleted to identify the primary pathway for succinate production in the engineered strain KJ073 (Table 10). Deletion of the *ppc* gene encoding the primary carboxylating step in native *E. coli* (XZ320) did not alter succinate production. Deletion of the genes encoding malic enzymes (*sfcA* and *maeB*) to produce XZ341 and XZ396, respectively, also had no effect on succinate production consistent with their primary function in gluconeogenesis. Deletion of *pck* encoding PEP carboxykinase (XZ332), however, dramatically reduced cell growth, sugar metabolism, succinate production, and succinate yield. Complementation of the deletion mutation in strain XZ332 with the entire *pck* gene from strain KJ073 (plasmid pLOI4677) substantially improved succinate production to near that of KJ073 (Table 10).

(0186) Together, these results demonstrate that the gluconeogenic PEP carboxykinase was recruited during metabolic evolution to serve as the primary carboxylation reaction for fermentative succinate production in KJ073. Unlike the PEP carboxylase (Fig. 2A), PEP carboxykinase conserves energy providing an additional ATP (Fig. 2D). Since fermentative growth and ATP production are closely linked, the increase in ATP yield with PEP

carboxykinase (Fig. 2D) would provide a competitive advantage during growth-based selection at the metabolic evolution steps leading to strain KJ073.

(0187) The increase in PEP carboxykinase activity correlated with an increase in *pck* mRNA abundance (Table 11; Fig. 12A). Transcript levels for *ppc*, *sfcA* and *maeB* were essentially unchanged while *pck* transcript abundance increased in KJ073 as compared to the wild type, in excellent agreement with the enhanced PEP carboxykinase activity (Table 11).

(0188) Sequencing the *pck* gene revealed no difference in the coding or terminator regions of the *pck* gene from KJ073. However, a single point mutation (G to A transition at position –64 relative to the ATG start codon) was found in the upstream promoter region of *pck* gene in KJ073(Table 11). All six strains were sequenced to identify the origin of this mutation during strain improvement. Although PEP carboxykinase activity was 4-fold higher in KJ017 than in KJ012, the G to A mutation was absent. This point mutation in the promoter region of pck gene in KJ073 strain is referred as *pck** mutation (high PEP carboxykinase activity). This *pck** mutation was present only in KJ060 and KJ073 strains showing high PEP carboxykinase activity and absent in KJ071. Loss of this mutation in KJ071 was accompanied by a 10-fold decrease in PEP carboxykinase activity to a level equivalent to that of KJ017.

(0189) Introduction of the *pck** point mutation (G to A) into KJ017, and KJ071 increased PEP carboxykinase activity by 9-fold (Table 11). Restoring the wild type *pck* gene (A to G) in strains KJ060 and KJ073 yield the strains XZ622 and XZ624 respectively. Both XZ622 and XZ624 showed a decreased PEP carboxykinase activity by almost 8-fold (Table 12). Together, these results demonstrate that the increase in PCK activity that occurred during the metabolic evolution of KJ017 to produce KJ060 is due to a single nucleotide change in *pck* (G to A at –64 relative to the ATG start). Absence of this mutation in KJ017 suggests that the initial 4-fold increase in PEP carboxykinase activity observed in this strain is due to other yet to be defined change(s). Apparently, recruitment of the native gluconeogenic PCK for fermentative succinate production resulted from multiple mutations that affected transcription of the *pck* gene.

(0190) The 4-fold increase in PEP carboxykinase activity that occurred during the development of KJ017 from KJ012 did not result from a mutation in *pck* and may involve a

mutation in a transcriptional regulator. The Cra protein has been shown to activate the expression of *pck* in *E. coli*. (Saier & Ramseier, 1996). However, no mutation was found in the *cra* gene or upstream region. Deletion of *cra* in KJ017 (XZ626) and KJ060 (XZ627) did not affect PEP carboxykinase activity (Table 12).

(0191) The *csrA* system has also been reported to regulate the level of *pck* and other genes involved in glucose metabolism by altering mRNA stability. (Pernestig *et al.*, 2003). However, no mutation was found in the sequences of genes involved in this regulatory system (*csrA, csrB, csrC, csrD, uvrY* or *barA*).

(0192) *E. coli* PEP carboxykinase activity is subject to glucose catabolite repression. (Goldie 1984). Two Crp-binding sites have been identified in the promoter (Ramseier *et al* 1995), quite distant from the point mutation in KJ060 and KJ073. Genes associated with catabolite repression (*cyaA, crp*) and glucose uptake by the phosphotransferase system (*ptsH, ptsI, crr, ptsG*) were sequenced (upstream region through terminator). Only one mutation was found, a frame-shift mutation in the carboxy-terminal region of *ptsI* (single base deletion at position 1,673) in strains KJ060, KJ071, and KJ073. Since this deletion was absent in KJ017, it cannot be responsible for the initial 4-fold increase in PEP carboxykinase activity in this strain (Table 12). Deletion of *ptsI* in KJ017 (XZ613) and in KJ060 (XZ615) did not affect PEP carboxykinase activity.

(0193) Potential changes in the catabolite repression of *pck* were investigated further by growing *E. coli* ATCC 8739, KJ012, KJ017, and KJ060 aerobically in LB medium with and without 5% glucose (Table 13). PEP carboxykinase activity has been reported to be maximal during late exponential phase of growth (Goldie, 1984) and this was confirmed in our strains. In both ATCC 8739 and the starting strain for metabolic evolution (KJ012), PEP carboxykinase activities were strongly repressed (>70%) by the presence of glucose. Glucose-mediated repression was reversed by the addition of cyclic-AMP. In contrast, PEP carboxykinase activity was 4-fold higher in KJ017 (than in KJ012) and was unaffected by the addition of glucose indicating a loss of catabolite repression. PEP carboxykinase activity was even higher in KJ060 (5-fold that of KJ017) during growth in the absence of glucose, and increased further when glucose was included. In KJ060 (and KJ073), glucose catabolite repression of *pck* has been replaced with glucose activation (Table 13). Transcript abundance

of *cyaA*, *crp*, *ptsG*, *ptsI* and *crr* were compared among these strains (Figs. 12B and 12C). Transcripts for *cyaA*, *crp*, and *ptsG* were generally higher in KJ017, KJ060, KJ071, and KJ073 than in KJ012 and ATCC 8739. Elevated levels of these proteins could mask catabolite repression by increasing the level of cyclic-AMP.

(0194) The loss of catabolite repression in these mutants was not limited to *pck* expression. ß-galactosidase activity was reduced by half in ATCC 8739 and KJ012 during growth with glucose (Table 13). This activity is normally repressed by glucose but was relatively unaffected in KJ017 and KJ060, consistent with general loss of Crp-mediated glucose regulation.

(0195) Deletion of the *crp* gene in KJ012 and KJ017 reduced the level of PEP carboxykinase activity in the resulting strains (XZ642 and XZ643, respectively) to that of the glucose-repressed parent, ATCC 8739 (Table 13). In these *crp*-deleted strains, PEP carboxykinase activity was not affected by the addition of glucose or cyclic AMP. Thus Crp may potentially be an essential regulatory element for the 4-fold increase in *pck* expression observed in KJ017 and its derivatives. This Crp-mediated 4-fold increase in *pck* expression together with the 10-fold increase in expression associated with the upstream *pck* mutation can account for the changes in the levels of PEP carboxykinase activity observed during the development of KJ060 and KJ073.

(0196) Other genes associated with cAMP-CRP regulation were also sequenced (Table 13) such as the predicted adenylate cyclase *ygiF*, a transcriptional co-activator for Crp *sxy*, the cAMP phosphodiesterase *cpdA* and the global regulator of carbohydrate metabolism *mlc*. (Keseler *et a., 2005*; Cameron & Redfield, 2006; Imamura *et al.*, 1996; Plumbridge, 2002). However, none of these genes associated with cAMP=CRP regulations were found to have any mutation in the upstream, coding or terminator regions.

(0197) The gene *ptsI* encodes PEP-protein phosphotransferase, a general (non sugar-specific) component of the phosphoenolpyruvate-dependent sugar phosphotransferase system. This major carbohydrate active-transport system catalyzes the phosphorylation of incoming sugar substrates concomitantly with their translocation across the cell membrane. PEP-protein

phosphotransferase transfers the phosphoryl group from phosphoenolpyruvate (PEP) to the phosphoryl carrier protein (HPr). (*See, e.g.*, UniProtKB, Accession Number P08839).

(0198) A frame-shift mutation in *ptsI* (single base-pair deletion of position 1673 bp) was found to have occurred during the metabolic evolution of KJ060 from KJ017. To investigate the significance of this mutation, the carboxy-terminal 175 bp of *ptsI* was deleted in KJ017 and KJ060 to produce XZ613 and XZ615, respectively. The *ptsI* deletion in KJ060 (XZ615) had no effect on growth and PEP carboxykinase activities as expected. Deletion of the *ptsI* carboxy-terminus in KJ017 to produce XZ613 resulted in an inability to grow on NBS mineral salts containing glucose, consistent with loss of the primary uptake system for glucose (PEP-dependent phosphotransferase system). After several days of incubation, cultures of XZ613 began to grow which are presumed to be derivatives that have activated alternative glucose transport system(s). (Karp *et al.*, 2007).

(0199) The *ptsI* mutation found in KJ060, KJ071, and KJ073 would be expected to inactivate the PEP-dependent phosphotransferase system for glucose. Alternative glucose uptake systems such as GalP have been shown to restore glucose uptake in *pts* mutants. (Wang *et a.l*, 2006; Yi *et al.*, 2003). Expression of *galP* was increased by 5-fold to 20-fold in these improved strains (Fig. 12B) as compared to the KJ012 and ATCC 8739, with a smaller increase in glucokinase (*glk*). Replacing the mutant *ptsI* gene in KJ060 with a functional wild type gene (XZ616) was detrimental, dramatically reducing growth in NBS-glucose medium. This detrimental effect may result from the depletion of PEP, a required substrate for wild-type glucose transport. A functional PEP-dependent phosphotransferase system would compete with PEP carboxykinase, decreasing the PEP pool available for redox balance, ATP production, and succinate production. Thus the loss of the PEP-dependent phosphotransferase system for glucose can be viewed as a beneficial event during strain development provided alternative transport systems such as GalP (and glucokinase) were available.

(0200) Carbon fluxes at the PEP node serve to limit the amount of succinate produced for redox balance during the anaerobic fermentation of glucose (Fig. 13). In addition, these fluxes must provide sufficient energy (ATP) for growth, maintenance, and precursors for biosynthesis. Rumen bacteria that produce succinate as the dominant product use an energy-

conserving PEP carboxykinase for OAA production (Fig. 2D). (Van der Werf *et al.*, 1997; Kim *et al.*, 2004). Native strains of *E. coli* produce succinate as a minor product and use PEP carboxylase (*ppc*) (Fig. 2A). Unlike PEP carboxykinase, PEP carboxylase is essentially irreversible due to the energy loss associated with release of inorganic phosphate. Previous studies have shown that overexpression of the native *ppc* gene in *E. coli* resulted in higher specific succinate production (Millard *et al.*, 1996) and higher specific growth rate due to increased carboxylation of PEP to oxaloacetate. (Farmer & Liao, 1997). However, since PEP is required for the native glucose transport system, over expressing *ppc* also decreased the rate of glucose uptake without significantly increasing succinate yield. (Chao & Liao, 1993; Gokarn *et al.*, 2000). This failure of the native PEP carboxylase to increase succinate yields diverted most research attention to a new metabolic design, over expression of the pyruvate carboxylase from *Lactobacillus lactis* or *Rhizobium etli* as the carboxylating step (Sanchez *et al.*,2005b; Gokarn *et al.*, 2000) and over expression of PEP carboxykinase from *Actinobacillus succinogenes* (Kim *et al.*, *2004*) for the development of industrial biocatalysts, rather than pursuing further work with the native repertoire of *E. coli* genes.

(0201) Native *E. coli* strains have three alternative carboxylation pathways (Figs. 2B –2D) that typically function only in the reverse direction for gluconeogenesis. (Samuelov *et al.*, *1991*; Kao *et al.*, 2005; Oh *et al.*, *2002*; Stols & Donnelly, *1997*). All three would allow the conservation of energy from PEP as ATP. With succinate as the sole route for NADH oxidation, growth-based selection (metabolic evolution) resulted in strains KJ017, KJ060, KJ071, and KJ073 with improvements in growth (cell yield) and succinate production. In these strains, the gluconeogenic PEP carboxykinase (*pck*) was recruited to serve as the primary carboxylation activity by transcriptional activation. The recruitment of PEP carboxykinase as the primary pathway for succinate production in *E. coli* was surprising. Many studies have shown that increased expression of *E. coli pck* had no effect on succinate production. (Gokarn, *et al.*, 2001; Vemuri *et al.*, 2002; Millard *et al.*, *1996*; Gokarn *et al.*, 2000; Hong & Lee, 2001). A recent study demonstrated that increased expression of *E. coli pck* was detrimental for growth in minimal medium, decreasing the growth rate, the rate of glucose metabolism, and the yield of succinate. (Kwon *et al.*, 2008).

(0202) Increased expression of *pck* in *E. coli* resulted in increased carbon flow into the 4-carbon intermediate OAA for succinate production (redox balance), increased succinate production, and increased the net production of ATP for growth and maintenance. At least three events contributed to increased transcription of *pck*: 1) loss of Crp-mediated glucose-repression; 2) gain of glucose-activation; and 3) a single base change in the upstream region of *pck*. Each of these events provided a basis for selection through metabolic evolution by increasing the level of PEP carboxykinase, increasing the flow of carbon into succinate, and increasing the conservation of metabolic energy as ATP. The combined action of these genetic events resulted in high levels of PEP carboxykinase (>7000 U (mg protein)$^{-1}$) in KJ060 and KJ073, equivalent to rumen bacteria that have evolved to produce succinate as the primary fermentation product. (VanderWerf *et al,. 1997*).

(0203) Additional energy-conserving changes were found in strains KJ060, KJ071, and KJ073 which may potentially assist in the recruitment of PEP carboxykinase as the primary route for fermentative succinate production. The PEP-dependent phosphotransferase system is the primary glucose uptake system in native strains of *E. coli*. During transport, half of the PEP produced from glucose is used for uptake and phosphorylation, limiting metabolic options for redox balance and ATP production. The improved strains contained a frame-shift mutation in *ptsI* that inactivated this uptake system. Expression of *galP* encoding a proton symporter which can transport glucose was increased by up to 20-fold. Increased expression of GalP and the native glucokinase can functionally replace the glucose phosphotransferase system by using ATP rather than PEP for phosphorylation. (Wang *et al.*, 2006; Yi *et al.*, 2003; Flores *et al.*, 2007). This exchange respects an energy-efficient way to increase the pool size of PEP available for carboxylation and redox balance using succinate (Fig. 11). All improved strains directed more than half of glucose carbon into 4-carbon products (malate plus succinate) and required more than half of the PEP for redox balance. Pyruvate can be converted to PEP by ATP with the formation of PPi and AMP but energy is wasted by this process. Thus the *ptsI* mutation and expression of *galP* increased the energy efficiency of metabolizing glucose to succinate, providing a growth advantage during metabolic evolution.

(0204) Eliminating alternative routes for NADH oxidation other than the succinate together with metabolic evolution with selection for improvements in growth resulted in succinate-

producing strains of *E. coli* that are the functional equivalents of succinate-producing rumen bacteria such as *Actinobacillus succinogenes* and *Mannheimia succiniciproducens*. (VanderWerf *et al.*, 1997; Kim *et al.*, 2007; Martin, 1994; McKinlay *et al.*, 2008). OAA is produced using an energy-conserving PEP-carboxykinase. PEP is conserved to eliminate the need for energy-expensive regeneration (2-ATP equivalents) by using glucose permeases (and glucokinase) rather than the PEP-dependent phosphotransferase system for glucose uptake. Note that among rumen bacteria producing other fermentation products, the phosphotransferase system is widely used for glucose uptake. (Martin, 1994). The most promising *E. coli* strains for succinate production, KJ060 and KJ073, produced approximately 700 mM succinate with a yield of 1.2 mol succinate per mol glucose (Jantama *et al.*, 2008a), comparable to the best natural succinate-producing rumen bacteria, *Actinobacillus succinogenes*. (Guettler *et al.*, U.S. Pat. No. 5,505,004).

(0205) Energy conserving strategies that improved succinate production from glucose in *E. coli* could also be applied to other important problems in strain engineering. Glycerol is becoming an inexpensive feedstock due to global increase in bio-diesel production. Being more reduced than glucose, each glycerol could be converted to succinate and maintain redox balance. However, no net ATP would be produced during glycerol metabolism to succinate using the native energy-wasting carboxylation activity, PEP carboxylase. This problem should be solved by using the energy-conserving PEP carboxykinase and allow the net formation of 1 ATP per succinate. In addition, the carboxylation product, OAA, is an important intermediate in cell metabolism that serves as a precursor for many other important fermentation products such as malic acid, fumaric acid, aspartic acid, lysine, threonine, methionine, among others. One of skill in the art will appreciate that the energy conservation strategies illustrated herein may be used to develop and improve biocatalysts for the production of many industrially important chemicals.

Example 4

Re-engineering *E. coli* for succinate production in mineral salts medium

(0206) Strains used in this study are listed in Table 16. Plasmids and primers used during construction of various strains developed during the course of this invention are listed in Table 15.

(0207) Examination of the mixed acid pathway in *E. coli* indicates three primary routes for NADH oxidation leading to succinate, lactate, and ethanol (Fig. 14). Deleting genes that eliminated these competing routes for NADH oxidation was insufficient to direct carbon flow to succinate (Table 16). With the exception of a small increase after deletion of *adhE*, individual deletions of these genes decreased succinate production and yield during fermentation in NBS mineral salts medium with little effect on cell yield as compared to the parent strain (ATCC 8739). Deletion of both alternative NADH oxidation pathways (*ldhA* with *adhE* or *ldhA* with *pflB*) substantially reduced growth, succinate yield, and succinate production in NBS mineral salts medium.

(0208) Quite different results were observed in Luria broth (Table 16). A modest increase in succinate yield was found in all strains that contained a deletion in *ldhA* (alone or in combination). Again, growth was reduced in double mutants and triple mutants. None of these deletions were sufficient to redirect substantial amounts of glucose carbon to succinate in either mineral salts medium or complex medium. Deleted strains constructed for succinate production based on observational analysis of the native mixed acid pathway in *E. coli* were less productive than expected in complex medium, and unsuccessful in NBS mineral salts medium.

(0209) Our previous investigations (Jantama *et al*, 2008a; Jantama *et al*, 2008b) discovered a promoter mutation (G to A at –64 relative to the ATG start; denoted *pck**) that increased phosphoenolpyruvate carboxykinase (PCK) activity and increased succinate production from glucose in the presence of many additional mutations. This was surprising because *pck* is typically repressed by glucose (Goldie, 1984) and has been shown to function primarily in gluconeogenesis during the aerobic metabolism of organic acids. (Kao *et al.*, 2005; Keseler *et al.*, *2005*; Oh *et al.*. 2002; Unden & Kleefeld, 2004; Wu *et al.*, 2007). To examine this further, the *pck* gene (ribosomal binding site, coding and terminator region) was amplified and cloned into pCR2.1-TOPO to produce pLOI4677 with *pck* expression under control of the *lac* promoter. This plasmid was transformed into ATCC 8739. Also, the native *pck* gene in ATCC 8739 was replaced with the mutant *pck** gene to construct XZ632. Glucose fermentation was examined in both strains using NBS mineral salts medium.

(0210) Introduction of *pck** resulted in a modest increase in succinate yield and production as compared to isogenic strains containing the native *pck* gene (Table 16). PCK activity was found enhanced over 8-fold both in the XZ632 stain and in the parent strain having a plasmid containing *pck** gene under the IPTG-induced promoter when compared to the parent strain. Upon chromosomal integration, the single copy *pck** was almost as effective for PCK production as the multi-copy plasmid containing *pck** gene under the IPTG-induced promoter. A modest improvement in succinate yield and production was observed for both strains with elevated PCK activity as compared to the parent ATCC 8739 strain (Table 17). However, increased PCK activity alone was insufficient to redirect glucose carbon to succinate.

(0211) The *pck** mutation was tested in combination with other mutations that eliminated pathways for NADH oxidation (Table 18). The combined action of mutations to eliminate competing routes for NADH oxidation and increased PCK activity were also insufficient to substantially redirect glucose carbon to succinate.

(0212) The phosphoenolpyruvate-dependent phosphotransferase system is the primary mechanism for glucose uptake in *E. coli* and an integral part of the glucose catabolite repression system. (Keseler *et al.*, 2005; Postma *et al.*, 1996). As described in the Example 3 above, a frame-shift mutation within the carboxyl end of *ptsI* (single base-pair deletion at 1673 bp position) was discovered in the succinate producing strains KJ060, KJ071 and KJ073 as a result of metabolic evolution. This mutation would be expected to disrupt function of the phosphotransferase system. (Postma *et al.*, 1996). In this mutant, glucose uptake was functionally replaced by *galP* (galactose permease) and *glk* (glucokinase) (Hernandez-Montalvo *et al.*, 2003; Keseler *et al.*, 2005). To investigate the effect of the *ptsI* disruption on succinate production, the carboxy-terminal 175 bp of *ptsI* was deleted in wild type *E. coli* ATCC 8739 to obtain strain XZ650. Surprisingly, succinate production and yield were decreased by this mutation in comparison to the parent (Table 18).

(0213) Two approaches were used to investigate the effect of combining the *ptsI* truncation with high levels of PCK using the *pck** mutation or the native *pck* over-expressed from the *lac* promoter (plasmid pLOI4677). In combination with the *ptsI* truncation, both approaches

resulted in a dramatic increase in succinate production (Table 18). Strain XZ647 (*pck**, Δ*ptsI*) produced 216 mmol succinate in NBS mineral salts medium with 5% glucose with a yield of 0.89 mol succinate per mol glucose, 4.7-fold higher than wild type *E. coli* ATCC 8739 (Table 18). With both XZ647 and XZ650 (pLOI4677), formate, ethanol and small amounts of lactate remained as minor side products. These two changes, inactivation of the phosphoenolpyruvate-dependent phosphotransferase system and elevated levels of PCK activity, represent the core changes required to effectively redirect glucose metabolism to succinate in NBS mineral salts medium without modifying any genes directly concerned with fermentative redox balance.

(0214) Further experiments confirmed that a wide variety of mutations in the phosphotransferase system increase succinate in a *pck** background. Disruption of the PEP-dependent phosphotransferase system for glucose at any step increases the flow of carbon into succinate, increasing both titer and yield.

(0215) The PEP-dependent phosphotransferase system shuttles phosphate from PEP to PtsI, then PtsH, then PtsG, and then to glucose to form glucose 6-phosphate. Table 19 shows that in a strain containing the pck* mutation that increases the level of phosphoenolpyruvate carboxykinase, a second mutation in any one of the *Pts* genes involved in this phosphate relay system (*ptsI, ptsH or ptsG*) results in a similar dramatic shift in carbon flow into succinate as the dominant fermentation product. In this regard, deletion of the whole *ptsI* gene or a truncation of the carboxyterminus of the *ptsI* gene was superior to the *ptsG* and *ptsH* deletions. The truncation of the ptsI carboxyterminus produced the highest yield and titer of succinate, slightly better than the complete deletion of ptsI. Other mutations such as insertions, deletions, and frame-shifts that lead to inactive gene products would be expected to have similar effects.

(0216) Although strain XZ647 (*pck** Δ*ptsI*) produced succinate as the dominant fermentation product, significant levels of unwanted co-products (lactate, ethanol, formate, and acetate) were also produced (Table 18). Deletion of either *adhE* (XZ723) or *pflB* (XZ721) eliminated the production of ethanol. Production of acetate and formate were substantially reduced or eliminated only by deletion of *pflB*. The resulting strain, (XZ721) produced high levels of succinate with a molar yield of over 1.2 mol succinate per mol glucose.

(0217) Succinate typically represents a minor product of glucose fermentation in *E. coli*. Most of the glucose carbon is converted to ethanol and lactate with smaller amounts of formate and acetate using alternative NADH-oxidizing pathways (Fig. 14). Derivatives of *E. coli* have been constructed to improve succinate production for more than a decade with variable success. (Donnelly *et al.*, U.S. Pat. No. 5,770,435; Gokarn *et al.*, 2000; Gokarn *et al.*, U.S. Pat. No. 6,455,284; Millard *et al.*, 1996; San *et al.*, U.S. Pat. No. 7,223,567; Sanchez *et* al, 2005b; Sanchez *et al*, 2005a; Stols & Donnelly, 1997; Vemuri *et al.*, 2002a; Wu *et al.*, 2007). The strategy used to construct these strains has typically focused on the elimination of competing pathways for NADH oxidation. (Donnelly *et al.*, U.S. Pat. No. 5,770,435; Gokarn *et al.*, U.S. Pat. No. 6,455,284; San *et al.*, U.S. Pat. No. 7,223,567; Sanchez *et al.*, 2005b; Sanchez *et al.* 2005a; Vemuri *et al.*, 2002a; Wu *et al.*, 2007). Target genes for deletion were selected primarily based on inspection of the pathway (Fig. 14). However, successes with this strategy have been limited to complex medium and two-step (aerobic growth phase followed by anaerobic production phase) processes. (Donnelly *et al.*, U.S. Pat. No. 5,770,435; Gokarn *et al.*, U.S. Pat. No. 6,455,284; Millard *et al.*, 1996; San *et al.*, U.S. Pat. No. 7,223,567; Sanchez *et al.*, 2005b; Sanchez *et al.*, 2005a; Vemuri *et al.*, 2002a; Wu *et al.*, 2007).

(0218) In complex medium such as Luria broth, most of the biosynthetic needs for growth are supplied by intermediates and building block molecules in the nutrients (yeast extract and tryptone). In this medium, deletion of genes based on observation of the fermentation pathway itself generally improved succinate production (Fig. 14; Table 16). Deletion of (*ldhA*) and all combinations of target genes including *ldhA* resulted in increased succinate production and increased succinate yield per glucose during fermentation in Luria broth (Table 16).

(0219) With a mineral salts medium such as NBS or AM1 broth, however, deletion of the same target genes in the mixed acid fermentation pathway was not helpful. Deletions of most were detrimental for both succinate production and succinate yield. With the exception of a small increase after the deletion of *adhE*, single gene deletions and combinations of gene deletions in the mixed acid fermentation pathway reduced succinate production and yields in

NBS mineral salts medium (Table 17). KJ012 (ATCC 8739 Δ*ldhA* Δ*adhE* Δ*ackA*), the genetic equivalent of a strain patented for two-step (aerobic growth phase followed by anaerobic production phase) succinate production in complex medium (San *et al.*, U.S. Pat. No. 7,223,567) produced less succinate than the wild type parent in NBS mineral salts medium. Based on these results, we concluded that rational selection of target genes for deletion based on observations of pathways is an unreliable predictor of success for improvements in succinate production when using mineral salts medium. In mineral salts medium, carbon must be precisely partitioned between energy generation, fermentation, and the biosynthesis of building block molecules needed for cell growth.

(0220) As the results shown above indicate this invention provides a new strategy to construct strains for succinate production in mineral salts medium. No mutations were required in genes encoding the *E. coli* mixed acid fermentation pathway (Fig. 14) to substantially redirect glucose carbon to succinate. In this invention, the combination of two core changes in peripheral pathways resulted in a five-fold increase in succinate yield. The two core changes that are required for succinate production are: 1) increased expression of the energy conserving (gluconeogenic) phosphoenolpyruvate carboxykinase to replace the native fermentative phosphoenolpyruvate carboxylase (energy wasting) and 2) replacement of the glucose phosphoenolpyruvate-dependent phosphotransferase system with an alternative permease such as *galP* and ATP-dependent phosphorylation (*glk*). Together, these changes increased net ATP production for growth, increased the pool of phosphoenolpyruvate available for carboxylation, and increased succinate production.

(0221) Additional mutations in genes in the mixed acid fermentation pathway such as the deletion of *pflB* and others represent opportunities for modest further increases in succinate yield and production. Note that acetyl-CoA is an essential metabolite for biosynthesis that is produced primarily by *pflB* during fermentative growth. This function is presumed to be replaced in *pflB* mutants by native expression of the pyruvate dehydrogenase complex (*aceEF, lpd*), an enzyme that typically serves as the dominant route for acetyl-CoA production during oxidative metabolism. (Kim *et al.*, 2007). The resulting succinate pathway in *E. coli* strains optimally engineered for succinate production in mineral salts medium (Fig. 14) is functionally similar to the native pathway that evolved in succinate-producing rumen

bacteria. (Kim *et al.*, 2004; Lee *et al.*, 2002; Lee *et al.*, 2006; Samuelov *et al.*, *1991*; VanderWerf *et al.*, 1997).

Example 5

Succinate Production from Glycerol

(0222) Despite the difference in redox properties and transport mechanism for glycerol and glucose, we have discovered that the same mutations enabling the conversion of glucose into succinate in mineral salt medium can also be used to effectively redirect glycerol metabolism into succinate production at 80% of the maximum efficiency for the conversion of glycerol to succinate (0.8 mol of succinate produced per mol glycerol consumed).

(0223) During the course of these studies, we have discovered that *gldA* and the PEP-dependent phosphotransferase pathway previously regarded as cryptic comprise an important functional route for glycerol catabolism (Fig. 15). In both pathways, glycerol enters the cells by facilitated diffusion. Within the cell, part of the glycerol is immediately phosphorylated followed by oxidation to DHAP, the pathway that is widely regarded as the standard pathway for glycerol metabolism in *E. coli*. We have discovered that at least 1/3 of the glycerol is first reduced by *gldA* to DHA and subsequently phosphorylated by the PEP-dependent phosphotransferase system (*ptsH, ptsI*) acting within the cytoplasm to yield DHAP. DHAP serves as the common entry point to central metabolism for both uptake and activation systems.

(0224) Table 20 clearly demonstrates the effectiveness of combining the core mutations identified for succinate production with glucose for succinate production from glycerol in mineral salts medium (NBS) without the addition of complex nutrients. Although deletion of *pflB* alone reduced succinate production below that of the wild type parent, the combination of the *pflB* deletion with the promoter mutation in *pck* (*pck**; transcriptional activation) doubled succinate yield from 0.25 mol/mol glycerol to 0.5 mol/mol glycerol. Subsequent addition of a mutation in *ptsI* further increased the succinate yield to 0.8 mol of succinate / mol glycerol, 80% of the maximum theoretical yield. Similar results were also found for the deletion of other key genes (*gldA, ptsH, ptsI, dhaKL, dhaM*) that disrupt the use of the

phosphorelay system for phosphorylation (Table 20). These results were unexpected. The resulting pathway for glycerol conversion is shown in Fig. 15.

(0225) Fig. 15 shows a combination of the generally accepted pathway for glycerol catabolism (Lin, 1996) and the mixed acid fermentation pathway (Bock and Sawers, 1996) in wild type *E. coli*. In this pathway, all ATP produced would be consumed by glycerol phosphorylation if succinate is produced as a sole product. No ATP would be available for growth. Based on Fig. 15, deletion of *pflB* would be expected to increase succinate production by increasing the availability of reductant and intermediates. In contrast to this expectation, a decrease in succinate production was observed with the deletion of *pflB* gene (Table 20). Based on this pathway, mutational activation of the *pck* gene that normally functions only during gluconeogenesis may be anticipated to increase the flow of phosphoenolpyruvate into succinate by competing with pyruvate kinases based solely on the prior observation in this patent application that this increase in PCK expression was beneficial for sugar fermentation into succinate, otherwise this would be unexpected. The utilization of phosphoenolpyruvate carboxykinase (*pck*) instead of phosphoenolpyruvate carboxylase (*ppc*) has the added advantage of conserving energy from phosphoenolpyruvate as an additional ATP which can be used for biosynthesis.

(0226) Although a mutation in *ptsI* was beneficial for the diversion of carbon from glucose to succinate, this could not have been predicted to be of any benefit since the only glycerol uptake system recognized as functional in wild type *E. coli* (*glpF*) does not involve the phosphoenolpyruvate phosphotransferases system. (Keseler *et al.*, 2005; Lin, 1996.). The only known involvement of the phosphotransferase system in glycerol metabolism is within a minor pathway thought to be inactive in wild type *E. coli*. (Jin *et al.*, 1983:; Lin, 1996; Tang *et al.*, 1982). This minor pathway first reduces intracellular glycerol to dihydroxyacetone (DHA) with GldA then uses PtsH (*ptsH*) and EI (*ptsI*) as phosphate carriers to couple phosphoenolpyruvate to the phosphorylation of intracellular DHA to produce dihydroxyacetone-phosphate (DHAP). This pathway is thought to function only in mutant strains in which *glpK* is inactive. (Gutknecht *et al.*, 2001; Keseler *et al.*, 2005). Contrary to all expectations based on the generally accepted pathways for glycerol metabolism, a mutation of *ptsI* significantly increased the production of succinate from glycerol. These results

indicate that the glycerol dehydrogenase (*gldA*) and PTS phospho-relay system (phosphoenolpyruvate, *PtsH, PtsI*) are of unexpected importance for the catabolism of glycerol to dihydroxyacetone phosphate (DHAP) during the anaerobic fermentation of glycerol in NBS mineral salts medium. Based on the large increase in succinate yield, this dihydroxyacetone (DHA) pathway can be estimated to account for at least one-third of the glycerol flux into glycolysis. The deletion in *ptsI* inactivated this pathway and increased the availability of phosphoenolpyruvate for succinate production. Together, these three core mutations (*pck**, *ptsI*, *pflB*) effectively and unexpectedly redirected carbon flow from glycerol to succinate at 80% of the maximum theoretical yield during anaerobic fermentation in mineral salts medium (Fig. 15). In addition, the use of this pathway results in an important increase in ATP production, facilitating growth.

Table 1. Comparison of succinate production by microbial biocatalysts [a]

Organism	Medium/Condition	Succinate Titer (mM)[b]	Succinate Yield (mol/mol)	Reference
E. coli KJ060 (*ldhA adhE ackA focA pflB*)	100 g/l glucose AM1 with 10 g/l $NaHCO_3$, simple batch fermentation, 120 h incubation, pH maintained with 1:1 mixture of 6M KOH+3M K_2CO_3	733 [0.90]	1.41	This paper
E. coli KJ073 (*ldhA adhE ackA focA pflB mgsA poxB*)	100 g/l glucose AM1 with 10 g/l $NaHCO_3$, simple batch fermentation, 96 h incubation, pH maintained with 1:1 mixture of 6M KOH+3M K_2CO_3	668 [0.82]	1.20	This paper
E. coli KJ060 (*ldhA adhE ackA focA pflB*) high inoculum (200 mg CDW l^{-1})	100 g/l glucose AM1 with 10 g/l $NaHCO_3$, simple batch fermentation, 120 h incubation, pH maintained with 1:1 mixture of 6M KOH+3M K_2CO_3	622 [0.61]	1.61	This paper
Actinobacillus succinogenes FZ53	130 g/l glucose supplemented with 15 g/l CSL and 5 g/l YE, 80 g/l $MgCO_3$, anaerobic batch fermentation, 78 h incubation	898 [1.36]	1.25	Guettler et al., 1996a
E. coli *AFP111* (pflAB, ldhA, ptsG) Rhizobium etli pyc *overexpressed*	40 g/l glucose (90 g total glucose) in medium supplemented with 20 g/l tryptone, 10 g/l YE and 40 g/l $MgCO_3$, dual phase-fed batch fermentation, 76 h incubation	841 [1.31]	1.68	Vemuri et al., 2002ab
Anaerobiospirillum succiniciproducens *ATCC 53488*	120 g/l glucose in peptone/YE based medium, integrated membrane-bioreactor-electrodialysis with CO_2 sparging, 150 h incubation	703 [0.55]	1.35	Meynial-Salles et al., 2007
Actinobacillus succinogenes	100 g/l glucose supplemented with 15 g/l CSL and YE, 80 g/l $MgCO_3$, anaerobic batch fermentation, CO_2 sparging, 39 h	678 [2.05]	1.37	Guettler et al., 1996b

Table 1. Comparison of succinate production by microbial biocatalysts [a]

Organism	Medium/Condition	Succinate Titer (mM)[b]	Succinate Yield (mol/mol)	Reference
130Z	incubation			
E. coli *HL27659k/pKK313* (iclR sdhAB ackA-pta poxB, pstG) Sorghum vulgare pepc *overexpressed*	106 g/l glucose in medium supplemented with 20 g/l tryptone, 32 g/l YE and 2 g/l $NaHCO_3$, fed batch fermentation under complete aerobic condition, 59 h incubation	499 [1.00]	0.85	Lin et al., 2005d
Anaerobiospirillum succiniciproducens *ATCC 53488*	50 g/l glucose and 10 g/l CSL, CO_2 sparging and 300 mM Na_2CO_3, batch fermentation, 24 h incubation	426 [2.09]	1.37	Glassner and Datta, 1992
Mannheimia succiniciproducens (ldhA pflB pta-ackA)	63 g/L glucose in MMH3 (yeast extract based medium), fed batch fermentation, 0.25 vol/vol/min CO_2 sparging, 30 h incubation	444 [1.75]	1.16	Lee et al., 2006
Bacterial Isolate 130Z ATCC 55618	50 g/l glucose supplemented with 1% CSL, 0.6% YE, and 2 g/l $MgCO_3$ neutralized with 10 N NaOH, 0.3 atm of CO_2 , 29.5 h incubation	388 [1.55]	1.40	Guettler et al., 1998
E. coli SBS550MG (*ldhA adhE iclR E.. ackA-pta*), *L. lactis pyc* *Bacillus subtilis citZ*	20 g/l glucose (100 g total glucose) LB supplemented with 1 g/l $NaHCO_3$, 200 mg/l ampicillin, and 1mM IPTG. 100% CO_2 at 1L/min STP headspace, repeated fed-batch fermentation, 95 h incubation	339 [0.42]	1.61[c]	Sanchez et al., 2005a; Cox et al., 2006
E. coli AFP184 (*pflB ldhA pts*)	102 g/l glucose supplemented with 15 g/l CSL, dual phase aerobic growth and anaerobic production, sparging with air followed by CO_2, 32 h incubation	339 [1.27]	0.72[c]	Andersson et al., 2007
Actinobacillus succinogenes	70 g/l glucose with flour hydrolysate and 5 g/l YE, anaerobic batch fermentation	302	1.18	Du et al., 2007

Table 1. Comparison of succinate production by microbial biocatalysts [a]

Organism	Medium/Condition	Succinate Titer (mM)[b]	Succinate Yield (mol/mol)	Reference
ATCC 55618	with 4% inoculum, 65 h incubation	[0.55]		
Anaerobiospirillum succiniciproducens ATCC 53488	50 g/l glucose, 2% CSL, and 25 ppm tryptophan, neutralized with 5.5 M $NaCO_3$, saturated medium of 0.3 atm partial pressure of CO_2, 29.5 h incubation	289 [1.16]	1.04	Guettler et al., 1998
Succinivibrio dextrinosolvens *ATCC 19716*	15 g/l of each CSL and YE, 100 g/l glucose, and 80 g/l $MgCO_3$, batch fermentation, 36 h.	226 [0.74]	NR	Guettler et al., 1998
Corynebacterium glutanicum *R*	40 g/l glucose (121 g total glucose) in Defined mineral salt medium with 400 mM $NaHCO_3$, fed batch fermentation, 6 h incubation	195 [3.83]	0.29	Okino et al., 2005
Prevotella ruminocola *ATCC 19188*	15 g/l of each CSL and YE, 100 g/l glucose, and 80 g/l $MgCO_3$, batch fermentation, 36 h incubation	160 [0.52]	NR	Guettler et al., 1998
E. coli SBS550MG (*ldhA adhE iclR ackA-pta*), *L. lactis pyc* *Bacillus subtilis citZ*	20 g/l glucose LB supplemented with 1 g/l $NaHCO_3$, 200 mg/l ampicillin, and 1mM IPTG. 100% CO_2 at 1L/min STP headspace, batch fermentation, 24 h. incubation	162.6 [0.80]	1.61[c]	Sanchez et al., 2005a; Cox et al., 2006
Mannheimia succiniciproducens MBEL55E KCTC 0769BP	18 g/L glucose in MH4 (YE based medium) supplemented with 119 mM $NaHCO_3$, a continuous-cell-recycle membrane reactor with the CO_2 partial pressure of 101.3 kPa gas (100% CO_2), 6 h incubation	144 [2.83]	1.44	Song et al., 2007
E. coli SBS110MG (*ldhA adhE*),	20 g/l glucose LB supplemented with1.5 g/l $NaHCO_3$ and 0.5g	130	1.24[c]	Sanchez et al., 2005a;

Table 1. Comparison of succinate production by microbial biocatalysts [a]

Organism	Medium/Condition	Succinate Titer (mM)[b]	Succinate Yield (mol/mol)	Reference
Lactococcus lactis pyc	$MgCO_3$, 200 mg/l ampicillin, and 1mM IPTG. Dual phase with 100% CO_2 at 1L/min STP headspace, 168 h incubation	[0.09]		Sanchez et al., 2006
E. coli NZN111 (W1485 *pflB ldhA*), *E. coli sfcA* overexpressed	20 g/l glucose LB supplemented with 0.5 g $MgCO_3$, 1.5 g/l NaOAc, 0.1 g/l ampicillin, and 10 μM IPTG, 44 h incubation, sealed serum tube.	108 [0.22]	0.98 [c]	Stols et al., 1997
E. coli JCL1208, *E. coli ppc* overexpressed	11 g/l glucose LB supplemented with 0.15 g $MgCO_3$, 0.1 g/l carbenicillin, and 0.1 mM IPTG, 44 h incubation, anoxic CO_2 charging at 1 atm headspace, 18 h incubation	91 [0.60]	0.44 [c]	Millard et al., 1996
E. coli GJT – *Sorghum pepC*	40 g/l glucose LB supplemented with 27.78 g/l $MgCO_3$, simple batch fermentation in sealed airtight flask	80 [no data]	0.42 [c]	Lin et al., 2005c
E. coli HL51276k (*iclR icd sdhAB ackA-pta poxB, pstG*), *Sorghum* sp. *pepC* S8D mutation	10.8 g/l glucose LB supplemented with 2g/l $NaHCO_3$, 50 mg/l kanamycin, 1 mM IPTG, aerobic batch reactor, 50 h incubation	68 [0.16]	1.09 [c]	Lin et al., 2005b
E. coli SBS880MG (*ldhA adhE ΔfdhF*), *L. lactis pyc*	20 g/l glucose LB supplemented with1.5 g/l $NaHCO_3$ and 0.5g $MgCO_3$, 200 mg/l ampicillin, and 1mM IPTG. Dual phase with 100% CO_2 headspace, 168 h incubation	60 [0.04]	0.94 [c]	Sanchez et al., 2005b

[a] Abbreviations: CSL, corn steep liquor; YE, yeast extract; NR, not reported.

[b] Average volumetric productivity is shown in brackets [g l^{-1} h^{-1}] beneath succinate titer.

[c] The molar yield was calculated based on the production of succinate from metabolized sugar during both aerobic and anaerobic conditions. Biomass was generated predominantly during aerobic growth. Succinate was produced primarily during anaerobic incubation with CO_2, H_2, or a mixture of both.

Table 2. List of bacterial strains that have been deposited with the ARS culture collection.

Culture	Strain Designations	Deposit Date
KJ012	B-50022	March 15, 2007
KJ017	B-50023	March 15, 2007
KJ032	B-50024	March 15, 2007
KJ060	B-50025	March 15, 2007
KJ070	B-50026	March 15, 2007
KJ071	B-50027	March 15, 2007
KJ072	B-50028	March 15, 2007
KJ073	B-50029	March 15, 2007
KJ091	B-50110	February 20, 2008
KJ098	B-50111	February 20, 2008
KJ104	B-50112	February 20, 2008
KJ110	B-50113	February 20, 2008
KJ119	B-50114	February 20, 2008
KJ122	B-50115	February 20, 2008
KJ134	B-50116	February 20, 2008
XZ320	NRRL B-50267	March 17, 2009
XZ332	NRRL B-50268	March 17, 2009
XZ341	NRRL B-50269	March 17, 2009
XZ396	NRRL B-50270	March 17, 2009
XZ468	NRRL B-50271	March 17, 2009
XZ469	NRRL B-50272	March 17, 2009
XZ470	NRRL B-50273	March 17, 2009
XZ613	NRRL B-50274	March 17, 2009
XZ615	NRRL B-50275	March 17, 2009
XZ616	NRRL B-50276	March 17, 2009
XZ618	NRRL B-50277	March 17, 2009
XZ620	NRRL B-50278	March 17, 2009
XZ647	NRRL B-50279	March 17, 2009
XZ721	NRRL B-50280	March 17, 2009
XZ723	NRRL B-50281	March 17, 2009

Table 3. *Escherichia coli* strains, plasmids, and primers used in this study

***Escherichia coli* Strains**		
Relevant Characteristics		**Sources**
Strain C	Wild type (ATCC 8739)	ATCC
KJ012	strain C, Δ*ldhA*::FRT Δ*adhE*::FRT Δ*ackA*::FRT	This study
KJ017	KJ012, improved strain selected from 10% glucose, NBS	This study
KJ032	KJ017, Δ*ldhA*::FRT Δ*adhE*::FRT Δ*ackA*::FRT Δ(*focA-pflB*)::FRT	This study
KJ060	KJ032, improved strain selected from 10% glucose without initial acetate, NBS	This study
KJ070	KJ060, Δ*mgsA*	This study
KJ071	KJ070, improved strain selected from 10% glucose , NBS	This study
KJ072	KJ071, Δ*poxB*	This study
KJ073	KJ072, improved strain selected from 10% glucose, AM1	This study
SZ204	Δ(*focA-pflB*)::FRT-*kan*-FRT	Zhou, 2003
Plasmids		
pKD4	*bla* FRT-*kan*-FRT	Datsenko, 2000
pKD46	*bla* γ β *exo* (Red recombinase), temperature-conditional replicon	Datsenko, 2000
pFT-A	*bla flp* temperature-conditional replicon and FLP recombinase	Posfai, 1997
pEL04	*cat-sacB* targeting cassette	Lee, 2001 Thomason, 2005
pLOI3421	1.8 kbp *Sma*I fragment containing *aac*	Wood, 2005
pLOI4151	*bla cat; cat-sacB* cassette	This study
pCR2.1-TOPO	*bla kan*; TOPO TA cloning vector	Invitrogen

Table 3. *Escherichia coli* strains, plasmids, and primers used in this study		
pLOI4228	*bla kan*; *yccT'-mgsA-helD'* (PCR) from *E.coli* C cloned into pCR2.1-TOPO vector	This study
pLOI4229	*cat-sacB* cassette PCR amplified from pLOI4151 (*EcoR*V digested) cloned into *mgsA* in pLOI4228	This study
pLOI4230	PCR fragment amplified from pLOI4228 (using *mgsA*-1/2 primers), kinase treated, then self-ligation	This study
pLOI4274	*bla kan*; *poxB* (PCR) from *E.coli* C cloned into pCR2.1-TOPO vector	This study
pLOI4275	*cat-sacB* cassette PCR amplified from pLOI4151 (*EcoR*V digested) cloned into *poxB* of pLOI4274	This study
pLOI4276	PCR fragment amplified from pLOI4274 (using *poxB*-1/2 primers), kinase treated, then self-ligation	This study
Primer sets		
ldhA	5'**ATGAACTCGCCGTTTTATAGCACAAAACAGTACG ACAAGAAGTAC**GTGTAGGCTGGAGCTGCTTC3' (SEQ ID NO: 2) 5'**TTAAACCAGTTCGTTCGGGCAGGTTTCGCCTTTT TCCAGATTGCT**CATATGAATATCCTCCTTAG3' (SEQ ID NO: 3)	This study
adhE	5'**ATGGCTGTTACTAATGTCGCTGAACTTAACGCAC TCGTAGAGCGT**GTGTAGGCTGGAGCTGCTTC3' (SEQ ID NO: 4) 5'**TTAAGCGGATTTTTTCGCTTTTTTCTCAGCTTTAG CCGGAGCAGC**CATATGAATATCCTCCTTAG3' (SEQ ID NO: 5)	Zhou, 2003
ackA	5'**ATGTCGAGTAAGTTAGTACTGGTTCTGAACTGCG GTAGTTCTTCA**GTGTAGGCTGGAGCTGCTTC3' (SEQ ID NO: 6) 5'**TCAGGCAGTCAGGCGGCTCGCGTCTTGCGCGATA ACCAGTTCTTC**CATATGAATATCCTCCTTAG3' (SEQ ID NO: 7)	Zhou, 2003

Table 3. *Escherichia coli* strains, plasmids, and primers used in this study		
focA-pflB	5'**TTACTCCGTATTTGCATAAAAACCATGCGAGTTA CGGGCCTATAA**GTGTAGGCTGGAGCTGCTTC3' (SEQ ID NO: 8) 5'**ATAGATTGAGTGAAGGTACGAGTAATAACGTCCT GCTGCTGTTCT**CATATGAATATCCTCCTTAG3' (SEQ ID NO: 9)	This study
JM*catsacB*	5'**TTAGCTAGCATGTGACGGAAGATCACTTCG**3' (SEQ ID NO: 10) 5'**CCGCTAGCATCAAAGGGAAAACTGTCCATAT**3' (SEQ ID NO: 11)	This study
cat-up2/*sacB*-down2	5'**AGAGAGGATATCTGTGACGGAAGATCACTTCG**3' (SEQ ID NO: 12) 5'**AGAGAGGATATCGAATTGATCCGGTGGATGAC**3' (SEQ ID NO: 13)	This study
mgsA-up/down	5'**CAGCTCATCAACCAGGTCAA**3' (SEQ ID NO: 14) 5'**AAAAGCCGTCACGTTATTGG**3' (SEQ ID NO: 15)	This study
mgsA-1/2	5'**AGCGTTATCTCGCGGACCGT3'** (SEQ ID NO: 16) 5'**AAGTGCGAGTCGTCAGTTCC**3' (SEQ ID NO: 17)	This study
poxB-up/down	5'**AAGCAATAACGTTCCGGTTG**3' (SEQ ID NO: 18) 5'**CCACTTTATCCAGCGGTAGC**3' (SEQ ID NO: 19)	This study
poxB-1/2	5'**GACGCGGTGATGAAGTGAT**3' (SEQ ID NO: 20) 5'**TTTGGCGATATAAGCTGCAA**3' (SEQ ID NO: 21)	This study
pck-F/R	5'**TTGGCTAAGG AGCAGTGAAA TGCGCGTTA**3' (SEQ ID NO: 22) 5'**CACGACAAAA GAAGGGTAAA TAAAC**3' (SEQ ID NO: 23)	This study
pck-2/3	5'**TTGTTAACGCGCATTTCACT**3' (SEQ ID NO: 24) 5'**GCGATAGCGGCTACTGTCAT**3' (SEQ ID NO: 25)	This study
pck (RT-PCR)	**5'GACGATACCACTCGCGAT3'** (SEQ ID NO: 26) **5'GTCGACAACGAACAGACGT3'** (SEQ ID NO: 27)	This study

Table 3. *Escherichia coli* strains, plasmids, and primers used in this study		
birA (RT-PCR)	**5'ATCGTGATGGCGGAAGT3'** (SEQ ID NO: 28) **5'CTTGCGATCCTGCAGATAG3'** (SEQ ID NO: 29)	This study

Table 4. Fermentation of glucose in mineral salts medium by mutant strains of *E. coli*

Strain[a]	Culture Conditions	Media, Glucose %(	Cell Yield[b] (g/L)	Succinate Yield[c]		Av. Vol. Prod [d] (g/L/h)	Fermentation Products (mM) [e]					
				mol/mol	g/g		Suc	Mal	Pyr	Ace	Lac	For
E. coli C wild type [f]	0.1 OD_{550}, 0.1 mM 0.2 betaine	5%, NBS	2.0±0.2	0.19±0.02	0.12	0.12±0.01	49±3	-[g]	33±10	152±30	98±24	262±19
KJ012 [f]	0.1 OD_{550}, 0.1 mM betaine	5% NBS	0.3±0.1	0.20±0.01	0.13	0.04±0.01	6±0.4	-	-	26±1	-	-
KJ012	0.1 OD_{550}, 0.1 mM betaine shaken flask [h]	5% NBS +MOPS	1.5	0.10	0.06	0.02	10	-	-	226	-	16
KJ012	0.1 OD_{550} Luria Broth	5% LB	1.5	0.70	0.50	0.09	108	-	-	61	<2	14
KJ012 (*ldhA, ackA, adhE*)	1st TF: No betaine, 0.1 OD_{550}, 120 h transfers	5%, NBS	0.3	0.13	0.09	0.072	6	-	-	26	<2	-
	3rd TF: 2 mM betaine, 0.1 OD_{550}, 96 h transfers	5%, NBS	0.7	0.28	0.18	0.128	26	-	-	71	<2	-
	40th TF: 1 mM betaine, 0.1 OD_{550}, 24 h transfers, 3M K_2CO_3+6N KOH	5%, NBS	2.3	0.73	0.48	0.251	204	-	-	179	<2	151

Table 4. Fermentation of glucose in mineral salts medium by mutant strains of *E. coli*

Strain[a]	Culture Conditions	Media, Glucose %(	Cell Yield[b] (g/L)	Succinate Yield[c]		Av. Vol. Prod [d] (g/L/h)	Fermentation Products (mM) [e]					
				mol/mol	g/g		Suc	Mal	Pyr	Ace	Lac	For
(KJ017)	40th TF: 1 mM betaine, 0.1 OD_{550}, 24 h transfers, 3M K_2CO_3+6N KOH	10%, NBS	1.7	0.74	0.49	0.354	288	-	-	181	38	199
KJ032 (*ldhA, ackA, adhE, focA, pflB*)	2nd TF: 1 mM betaine, 0.1 OD_{550}, 48 h transfers, 20 mM NaOAc, 3M K_2CO_3+6N KOH	5%, NBS	1.0	1.47	0.97	0.260	212	-	-	44	-	-
	15th TF: 1 mM betaine, 0.01 OD_{550}, 24h transfers, 5 mM NaOAc, 3M K_2CO_3+6N KOH	10%, NBS	1.4	1.07	0.71	0.736	596	331	9	170	<2	-
(KJ060)	5th TF: 1 mM betaine, 0.01 OD_{550}, 24 h transfers, No NaOAc, 3M K_2CO_3+6N KOH	10%, NBS	1.4	1.04	0.69	0.711	579	318	9	161	<2	-
KJ070 (*ldhA, ackA, adhE, focA, pflB,*	1st TF: 1 mM betaine, 0.01 OD_{550}, 24 h TF, 3M K_2CO_3+6N KOH,	5%, NBS	1.0	1.06	0.70	0.361	294	219	25	102	-	-

Table 4. Fermentation of glucose in mineral salts medium by mutant strains of *E. coli*

Strain[a]	Culture Conditions	Media, Glucose %(	Cell Yield[b] (g/L)	Succinate Yield[c]		Av. Vol. Prod[d] (g/L/h)	Fermentation Products (mM)[e]					
				mol/mol	g/g		Suc	Mal	Pyr	Ace	Lac	For
mgsA) (KJ071)	50th TF: 1 mM betaine, 0.01 OD_{550}, 24 h transfers, 3M K_2CO_3+6N KOH	10%, NBS	1.1	0.71	0.47	0.419	341	626	<2	76	-	-
KJ072 (*ldhA, ackA, adhE, focA* *pflB, mgsA, poxB*) (KJ073)	2nd TF: 1 mM betaine, 0.01 OD_{550}, 24 h transfers, 3M K_2CO_3+6N KOH	10%, NBS	1.3	0.97	0.64	0.663	539	186	<2	95	-	-
	6th TF: 1 mM betaine, 0.01 OD_{550}, 24 h transfers, 3M K_2CO_3+6N KOH	10%, AM1	1.2	1.34	0.88	0.733	596	38	4	112	-	-
	45th TF: 1mM betaine, 0.01 OD_{550}, 24 h transfers, 3M K_2CO_3+6N KOH	10%, AM1	1.5	1.26	0.83	0.858	699	313	103	172	-	-
KJ073[f]	1mM betaine, 3M K_2CO_3+6N KOH, 0.01 OD_{550} inoculum	10%, AM1	2.3 ±0.1	1.20 ±0.09	0.77 ±0.03	0.82 ±0.01	668 ±8	118 ±13	55 ±22	183 ±27	-	-
KJ060[f]	1mM betaine, 3M K_2CO_3+6N KOH, 0.01 OD_{550} inoculum	10%, AM1	2.2 ±0.1	1.41 ±0.07	0.92 ±0.05	0.90 ±0.04	733 ±39	39 ±17	-	250 ±36	2±1	-

Table 4. Fermentation of glucose in mineral salts medium by mutant strains of *E. coli*

Strain[a]	Culture Conditions	Media, Glucose %(	Cell Yield[b] (g/L)	Succinate Yield[c]		Av. Vol. Prod [d] (g/L/h)	Fermentation Products (mM) [e]					
				mol/mol	g/g		Suc	Mal	Pyr	Ace	Lac	For
KJ060 [f]	1mM betaine, 3M K_2CO_3+6N KOH, 0.60 OD_{550} inoculum	10%, AM1	2.2 ±0.1	1.61 ±0.12	1.05 ±0.09	0.77 ±0.04	622 ±8	17 ±5	1.5 ±1	180 ±13	2±1	-
KJ071 [f]	1mM betaine, 3M K_2CO_3+6N KOH, 0.01 OD_{550} inoculum	10%, NBS	1.5 ±0.0	0.78 ±0.02	0.53 ±0.01	0.33 ±0.04	280 ±7	516 ±14	58 ±15	64 ±9	-	-

[a] Clones were isolated from the fermentation broth at various points and assigned strain numbers, indicated by numbers in parenthesis.

[b] Cell yield estimated from optical density (3 OD_{550nm} = 1 g l^{-1} CDW).

[c] Succinate yields were calculated based on glucose metabolized.

[d] Average volumetric productivity was calculated for total incubation time.

[e] Abbreviations: suc, succinate; mal, malate; pyr, pyruvate; ace, acetate; lac, lacate; for, formate.

[f] Average of 3 or more fermentations with standard deviations.

[g] Dash indicates absence of product.

[h] Aerobic shaken flask (100 rpm; 100 ml NBS, 250-ml flask.

Table 5. Comparison of carboxylation enzyme activities in different strains						
Enzyme	Specific activity [nmol min^{-1} (mg protein)$^{-1}$]					
	E.coli C	*E.coli* KJ012	*E.coli* KJ017	*E.coli* KJ073	*E.coli* K12 [a]	*Actinobacillus succinogenes* [a]
PEP carboxylase	20±2	25±2	17±1	27±2	140	10
PEP carboxykinase	295±23	162±11	700±68	7341±462	140	4,700
Malic enzyme (NADH, carboxylation)	ND [b]	5±2	12±4	12±3	Unknown	Unknown
Malic enzyme (NADPH carboxylation)	<1	<1	<1	<1	Unknown	Unknown

[a] data was from van der Werf et al., 1997

[b] Unable to measure in wild type *E. coli* C due to presence of lactate dehydrogenase

Table 6. Composition of media (excluding carbon source).

Component	Concentration (mmol L^{-1})	
	[a]NBS + 1 mM betaine	AM1+1 mM betaine
KH_2PO_4	25.72	0
K_2HPO_4	28.71	0
$(NH_4)_2HPO_4$	26.50	19.92
$NH_4H_2PO_4$	0	7.56
Total PO_4	80.93	27.48
Total N	53.01	47.39
[b]**Total K**	84.13	1.00
$MgSO_4\,7H_2O$	1.00	1.50
$CaCl_2\,2H_2O$	0.10	0
Thiamine HCl	0.015	0
Betaine-KCl	1.00	1.00
	(μmol L^{-1}) [c]	
$FeCl_3\,6H_2O$	5.92	8.88
$CoCl_2\,6H_2O$	0.84	1.26
$CuCl_2\,2H_2O$	0.59	0.88
$ZnCl_2$	1.47	2.20
$Na_2MoO_4\,2H_2O$	0.83	1.24
H_3BO_3	0.81	1.21
$MnCl_2\,4H_2O_2$	0	2.50
Total Salts	12.5 g L^{-1}	4.1 g L^{-1}

[a] NBS + 1 mM betaine: NBS media amended with betaine (1 mM).

[b] Calculation includes KOH used to neutralize betaine-HCl stock.

[c] Trace metal stock (1000X) was prepared in 120 mM HCl.

Table 7. *Escherichia coli* strains, plasmids, and primers used in herein

***Escherichia coli* Strains**		
Relevant Characteristics		Sources
Strain B		
KJ073	Δ*ldhA*::FRT Δ*adhE*::FRT Δ(*focA-pflB*)::FRT Δ*ackA*::FRT Δ*mgsA* Δ*poxB*	Jantama et al., 2008
KJ076	KJ073, Δ*ackA*::*cat-sacB*, translational stop sequence	Disclosed herein
KJ079	KJ073, Δ*ackA*::translational stop sequence	Disclosed herein
TG200	KJ079, Δ*adhE*::*cat-sacB*	Disclosed herein
TG201	TG200, Δ*adhE*	Disclosed herein
TG202	TG201, Δ*ldhA*::*cat-sacB*	Disclosed herein
TG203	TG202, Δ*ldhA*	Disclosed herein
TG204	TG203, Δ (*focA-pflB*)::*cat-sacB*	Disclosed herein
KJ091	TG204, Δ(*focA-pflB*)	Disclosed herein
KJ098	KJ091, Δ*tdcDE*	Disclosed herein
KJ104	KJ098, Δ*citF*	Disclosed herein
KJ110	KJ104, Δ*aspC*	Disclosed herein
KJ119	KJ104, Δ*sfcA*	Disclosed herein

Table 7. *Escherichia coli* strains, plasmids, and primers used in herein		
KJ122	KJ110, Δ*sfcA*	Disclosed herein
KJ134	KJ122, Δ*ackA-pta*	Disclosed herein
Plasmids		
pKD46	*Bla γ β exo* (red recombinase), temperature-conditional replicon	Datsenko, 2000
pEL04	*cat-sacB* cassette	Lee, 2001 Thomason, 2005
pLOI2228	*cat ;* FRT-*cat*-FRT cassette	Martinez-Morales et al., 1999
pLOI2511	*bla kan;* FRT-*kan*-FRT cassette	Underwood et al., 2002
pLOI4131	*bla ;* ligation of pLOI2228 (*Ban*I digested, Klenow treated, *Cla*I digested FRT-*cat*-FRT cassette) and pLOI2511 (*Nhe*I digested, Klenow treated, *Cla*I digested)	Disclosed herein
pLOI4145	*bla; Eco*RI digested pLOI4131, self-ligation	Disclosed herein
pLOI4146	*bla cat;* ligation of *cat-sacB* cassette PCR amplified (using JMcatsacBup3/down3 primers) from pLOI4152, *BamHI*/*Xho*I digested and *BamHI*/*Xho*I digested pLOI4153	Disclosed herein
pLOI4151	*bla cat; cat-sacB* cassette	Jantama et al., 2008
pLOI4152	*cat-sacB* cassette ; PCR amplified cassette from pEL04 (using JMpEL04F1/R1 primers), *Bgl*II digestion and self-ligation	Disclosed herein
pLOI4153	*bla ;* ligation of pLOI4145 (*Kas*I/*Xma*I digested) and *Kas*I/*Xma*I digested SfPBXPS polylinker (annealing of complementary oligonucleotides SfPBXPSsense/SfPBXPScomp)	Disclosed herein
pLOI4154	*Pac*I digested pLOI4146, self ligation	Disclosed herein

Table 7. *Escherichia coli* strains, plasmids, and primers used in herein		
pLOI4161	*bla cat; cat-sacB* cassette	Disclosed herein
pLOI4162	*bla cat;* ligation of *cat-sacB* cassette (*Pac*I digested) from pLOI4146 and *Pac*I digested pLOI4161	Disclosed herein
pCR2.1-TOPO	*bla kan*; TOPO TA cloning vector	Invitrogen
pLOI4158	*bla kan*; *ackA* (PCR) from *E.coli* C (using JMackA-F1/R1 primers) cloned into pCR2.1-TOPO vector	Disclosed herein
pLOI4159	*SmaI/Sfo*I *digested cat-sacB* cassette from pLOI4162 cloned into the PCR amplified inside-out product from pLOI4158 (using JMackAup1/down1)	Disclosed herein
pLOI4160	*Pac*I digestion of pLOI4159, then self-ligated	Disclosed herein
pLOI4515	*bla kan*; *tdcG'-tdcFED-tdcC'* (PCR) from *E.coli* C (using *tdcDE*-up/down primers) cloned into pCR2.1-TOPO vector	Disclosed herein
pLOI4516	*SmaI/Sfo*I *digested cat-sacB* cassette from pLOI4162 cloned into the PCR amplified inside-out product from pLOI4515 (using *tdcDE*-F7/R7 primers)	Disclosed herein
pLOI4517	PCR fragment amplified inside-out product from pLOI415 (using *tdcDE*-F7/R7 primers), kinase treated, then self-ligated	Disclosed herein
pLOI4629	*bla kan*; *citF* (PCR) from *E.coli* C (using *citF*-up2/down2 primers) cloned into pCR2.1-TOPO vector	Disclosed herein
pLOI4630	*SmaI/Sfo*I *digested cat-sacB* cassette from pLOI4162 cloned into the PCR amplified inside-out product from pLOI4629 (using *citF*-2/3 primers)	Disclosed herein
pLOI4631	*Pac*I digestion of pLOI4630, then self-ligated	Disclosed herein
pLOI4280	*bla kan*; *aspC* (PCR) from *E.coli* C (using *aspC*-up/down primers) cloned into pCR2.1-TOPO vector	Disclosed herein
pLOI4281	*SmaI/Sfo*I *digested cat-sacB* cassette from pLOI4162 cloned into the PCR amplified inside-out product from pLOI4280 (using *aspC*-1/2 primers)	Disclosed herein

Table 7. *Escherichia coli* strains, plasmids, and primers used in herein		
pLOI4282	PCR fragment amplified inside-out product from pLOI4280 (using *aspC*-1/2 primers), kinase treated, then self-ligated	Disclosed herein
pLOI4283	*bla kan*; *sfcA* (PCR) from *E.coli* C (using *sfcA*-up/down primers) cloned into pCR2.1-TOPO vector	Disclosed herein
pLOI4284	*SmaI/SfoI digested cat-sacB* cassette from pLOI4162 cloned into the PCR amplified inside-out product from pLOI4283 (using *sfcA*-1/2 primers)	Disclosed herein
pLOI4285	*Pac*I digestion of pLOI4284, then self-ligated	Disclosed herein
pLOI4710	*bla kan*; *ackA-pta* (PCR) from *E.coli* C (using *ackA*-up/*pta*-down primers) cloned into pCR2.1-TOPO vector	Disclosed herein
pLOI4711	*SmaI/SfoI digested cat-sacB* cassette from pLOI4162 cloned into the PCR amplified inside-out product from pLOI4710 (using *ackA-2/pta*-2 primers)	Disclosed herein
pLOI4712	*Pac*I digestion of pLOI4711, then self-ligated	Disclosed herein
pLOI4413	*bla kan*; *ychE'-adhE-ychG'* (PCR) from *E.coli* C (using up/down-*adhE* primers) cloned into pCR2.1-TOPO vector	Disclosed herein
pLOI4419	PCR fragment amplified inside-out product from pLOI4413 (IO-*adhE*-up/down using primers), kinase treated, then self-ligated	Disclosed herein
pLOI4415	*bla kan*; *ycaO'-focA-pflB-pflA'* (PCR) from *E.coli* C (using up-*focA*/Mid-*pflA* primers) cloned into pCR2.1-TOPO vector	Disclosed herein
pLOI4421	PCR fragment amplified inside-out product from pLOI4415 (using IO-*ycaO*-up/IO-mid*pflB*-down primers), kinase treated, then self-ligated	Disclosed herein
pLOI4430	*bla kan*; *hslJ'-ldhA-ydbH'* (PCR) from *E.coli* C (using *ldhA*-A/C primers) cloned into pCR2.1-TOPO vector	Disclosed herein
pLOI4432	PCR fragment amplified inside-out product from pLOI4424 (using IO-*ldhA*-up/down primers), kinase treated, then self-ligated	Disclosed herein
Primer sets		

Table 7. *Escherichia coli* strains, plasmids, and primers used in herein		
JM4161 sense/ comp	5'ACCGCATCAGGCGCCTAATTAATTAATCCCGG3' (SEQ ID NO: 30) 5'CCGGGATTAATTAATTAGGCGCCTGATGCGGT3' (SEQ ID NO: 31)	Disclosed herein
JMpEL04F 1/R1	5'CAGCAGATCTAAGTAAATCGCGCGGGTTTG3' (SEQ ID NO: 32) 5'CAGCAGATCTAGCGGCTATTTAACGACCCT3' (SEQ ID NO: 33)	Disclosed herein
JM*ackA*-F1/R1	5'GCCTGAAGGCCTAAGTAGTA3' (SEQ ID NO: 34) 5'GCACGATAGTCGTAGTCTGA3' (SEQ ID NO: 35)	Disclosed herein
JmackA up1/down1	5'GTTGAGCGCTTCGCTGTGAG3' (SEQ ID NO: 36) 5'GCCGCAATGGTTCGTGAACT3' (SEQ ID NO: 37)	Disclosed herein
JmcatsacB up3/down3	5'CTCACCTCGAGTGTGACGGAAGATCACTTCG3' (SEQ ID NO: 38) 5'GTGCAGGATCCATCAAAGGGAAAACTGTCCATAT3' (SEQ ID NO: 39)	Disclosed herein
SfPBXPS sense/comp	5'ATGTAGGCGCCATTAATTAATGGATCCACTATCTCGAGATTAATTAATCCCGGGACTAT3' (SEQ ID NO: 40) 5'ATAGTCCCGGGATTAATTAATCTCGAGATAGTGGATCCATTAATTAATGGCGCCTACAT3' (SEQ ID NO: 41)	Disclosed herein
WM*adhE* A/C	5'ATGGCTGTTACTAATGTCGCTGAACTTAACGCACTCGTAGAGCGTCGGCACGTAAGAGGTTCCAA3' (SEQ ID NO: 42) 5'TTAAGCGGATTTTTTCGCTTTTTTCTCAGCTTTAGCCGGAGCAGCACACTGCTTCCGGTAGTCAA3' (SEQ ID NO: 43)	Disclosed herein
WM*ldhA* A/C	5'ATGAAACTCGCCGTTTATAGCACAAAACAGTACGACAAGAAGTACGGCACGTAAGAGGTTCCAA3' (SEQ ID NO: 44) 5'TTAAACCAGTTCGTTCGGGCAGGTTTCGCCTTTTTCCAGATTGCTACACTGCTTCCGGTAGTCAA3' (SEQ ID NO: 45)	Disclosed herein

Table 7. *Escherichia coli* strains, plasmids, and primers used in herein		
WM*pflB* A/C	5'TTACTCCGTATTTGCATAAAAACCATGCGAGTTACGGGC CTATAACGGCACGTAAGAGGTTCCAA3' (SEQ ID NO: 46) 5'TTACATAGATTGAGTGAAGGTACGAGTAATAACGTCCTG CTGCTGTTCTACACTGCTTCCGGTAGTCAA3' (SEQ ID NO: 47)	Disclosed herein
tdcDE-up/down	5'CGCCGACAGAGTAATAGGTT3' (SEQ ID NO: 48) 5'TGATGAGCTACCTGGTATGG3' (SEQ ID NO: 49)	Disclosed herein
tdcDE-F7/R7	5'CGATGCGGTGGCCAATTAAG3' (SEQ ID NO: 50) 5'GACGACGTGCTGGATTACGA3' (SEQ ID NO: 51)	Disclosed herein
citF-up2/down2	5'GGGTATTCAGGCGTTCGATA3' (SEQ ID NO: 52) 5'GCCCGAGAGGATGACTATGT3' (SEQ ID NO: 53)	Disclosed herein
citF-2/3	5'GGTGATCGATGTTGTGCATC3' (SEQ ID NO: 54) 5'CCCGTTCTTGTCGTTGAGAT3' (SEQ ID NO: 55)	Disclosed herein
IO-*adhE*-up/down	5'GCTGCTCCGGCTAAAGCTGA3' (SEQ ID NO: 56) 5'ACGCTCTACGAGTGCGTTAA3' (SEQ ID NO: 57)	Disclosed herein
up-*focA*/Mid-*pflB*	5'AGATCGCCAGCCGCTGCAAT3' (SEQ ID NO: 58) 5'AACCGTTGGTGTCCAGACAG3' (SEQ ID NO: 59)	Disclosed herein
IO-*ycaO*-up/IO-mid*pflB*-down	5'GCCTACATTGCGTAGGCTAT3' (SEQ ID NO: 60) 5'GCAGCAGGACGTTATTACTC3' (SEQ ID NO: 61)	Disclosed herein
ldhA-A/C	5'ATGAAACTCGCCGTTTATAG3' (SEQ ID NO: 62) 5'TTAAACCAGTTCGTTGCCC3' (SEQ ID NO: 63)	Disclosed herein
IO-*ldhA*-up/down	5'CGTTCGATCCGTATCCAAGT3' (SEQ ID NO: 64) 5'AGGCTGGAACTCGGACTACT3' (SEQ ID NO: 65)	Disclosed herein
aspC-up/down	5'TCCATCGCTTACACCAAATC3' (SEQ ID NO: 66) 5'TGGGGGATGACGTGATATTT3' (SEQ ID NO: 67)	Disclosed herein

Table 7. *Escherichia coli* strains, plasmids, and primers used in herein		
aspC-1/2	5'AGATAACATGGCTCCGCTGT3' (SEQ ID NO: 68) 5'AGGAGCGGCGGTAATGTTC3' (SEQ ID NO: 69)	Disclosed herein
sfcA-up/down	5'CTATGCTTGATCGGCAACCT3' (SEQ ID NO: 70) 5'ACGATCGCCTGGTTTTAATG3' (SEQ ID NO: 71)	Disclosed herein
sfcA-1/2	5'TACCGCCGTACCTCCATCTA3' (SEQ ID NO: 72) 5'CGTAAGGGATATAAAGCGAACG3' (SEQ ID NO: 73)	Disclosed herein
ackA-up/*pta*-down	5'CGGGACAACGTTCAAAACAT3' (SEQ ID NO: 74) 5'ATTGCCCATCTTCTTGTTGG3' (SEQ ID NO: 75)	Disclosed herein
ackA-2/*pta*-2	5'AACTACCGCAGTTCAGAACCA3' (SEQ ID NO: 76) 5'TCTGAACACCGGTAACACCA3' (SEQ ID NO: 77)	Disclosed herein

Table 8. Fermentation of glucose in mineral salts AM1 medium by mutant strains of *E. coli*

Strain	Culture Conditions	Media, Glucose (w/v)	Cell Yield[a] (g/L)	Succinate Yield[b]		Av. Vol. Prod [c] (g/L/h)	Fermentation Products (mM) [d,e,f]					
				mol/mol	g/g		Suc	Mal	Pyr	Ace	Lac	For
KJ073	1mM betaine, 3M K_2CO_3+6N KOH (1:1) 0.01 OD_{550} inoculum	10%, AM1	2.3 ±0.1	1.20 ±0.09	0.77 ±0.03	0.82 ±0.01	668 ±8	118 ±13	55 ±22	183 ±27	-	-
KJ091	1mM betaine, 3M K_2CO_3+6N KOH (1:1) 0.01 OD_{550} inoculum	10%, AM1	2.2 ±0.1	1.19 ±0.02	0.78 ±0.01	0.84 ±0.01	687 ±3	109 ±3	72 ±5	155 ±6	-	
KJ098	1mM betaine, 3M K_2CO_3+6N KOH (1:1) 0.01 OD_{550} inoculum	10%, AM1	2.3 ±0.1	1.30 ±0.04	0.85 ±0.02	0.79 +0.01	644 ±9	-	42 ±8	88 ±1	-	-
KJ104	1mM betaine, 3M K_2CO_3+6N KOH (4:1) 0.01 OD_{550} inoculum	10%, AM1	1.8 ±0.1	1.31 ±0.01	0.86 ±0.01	0.78 ±0.03	634 ±25	5 ±1	78 ±5	90 ±10	-	-
KJ104	1mM betaine, 3M K_2CO_3+6N KOH (6:1) 0.01 OD_{550} inoculum	10%, AM1	1.9 ±0.1	1.30 ±0.01	0.85 ±0.01	0.77 ±0.01	625 ±4	3 ±2	94 ±5	81 ±2	-	-
KJ110	1mM betaine, 3M K_2CO_3+6N KOH (4:1) 0.01 OD_{550} inoculum	10%, AM1	2.0 ±0.1	1.28 ±0.02	0.84 ±0.01	0.79 ±0.01	640 ±10	4 ±1	76 ±6	106 ±11	-	-
KJ119	1mM betaine, 3M K_2CO_3+6N KOH (4:1) 0.01 OD_{550} inoculum	10%, AM1	2.0 ±0.1	1.33 ±0.07	0.87 ±0.01	0.82 ±0.01	672 ±10	4 ±0	64 ±18	95 ±14	-	-

KJ122	1mM betaine, 3M K_2CO_3+6N KOH (4:1) 0.01 OD_{550} inoculum	10%, AM1	2.3 ±0.1	1.50 ±0.02	0.98 ±0.01	0.92 ±0.01	750 ±1	0 ±0	122 ±21	94 ±13	-	-
KJ122	1mM betaine, 3M K_2CO_3+6N KOH (6:1) 0.01 OD_{550} inoculum	10%, AM1	2.0 ±0.2	1.54 ±0.02	1.01 ±0.01	0.97 ±0.04	787 ±35	6 ±3	59 ±6	110 ±7	-	-
KJ122	1mM betaine, 3M K_2CO_3+6N KOH (6:1) 0.15 OD_{550} inoculum	10%, AM1	2.1 ±0.1	1.57 ±0.09	1.03 ±0.06	0.93 ±0.06	756 ±49	0 ±0	124 ±13	122 ±9	-	-
KJ134	1mM betaine, 3M K_2CO_3+6N KOH (6:1) 0.01 OD_{550} inoculum	10%, AM1	2.3 ±0.1	1.70[g] ±0.03	1.11 ±0.02	0.83 ±0.02	674 ±15	13 ±2	22 ±9	37 ±5	-	

[a] Cell yield estimated from optical density (3 OD_{550nm} = 1 g l^{-1} CDW).

[b] Succinate yields were calculated based on glucose metabolized.

[c] Average volumetric productivity was calculated for total incubation time.

[d] Abbreviations: suc, succinate; mal, malate; pyr, pyruvate; ace, acetate; lac, lacate; for, formate.

[e] Ethanol (153±39 mM) was present only in broth from *E. coli* C.

[f] All data represent an average of 3 or more fermentations with standard deviations.

[g] Additional products were also found despite near theoretical yields of succinate. Based on total products, coproducts represented 11%.

Table 9. Sources and characteristics of *E. coli* strains and plasmids used in this study

	Relevant characteristics	Source or reference
Strains		
ATCC 8739	Wild type	Lab collection
KJ012	Δ*ackA*, Δ *ldhA*, Δ *adhE*	Jantama et al. 2008
KJ017	Δ*ackA*, Δ *ldhA*, Δ*adhE*	Jantama et al. 2008
KJ060	Δ*ackA*, Δ *ldhA*, Δ*adhE*, Δ*pflB*	Jantama et al. 2008
KJ071	Δ*ackA*, Δ*ldhA*, Δ*adhE*, Δ *pflB*, Δ*mgsA*	Jantama et al. 2008
KJ073	Δ*ackA*, Δ*ldhA*, Δ *adhE*, Δ*pflB*, Δ *mgsA*, Δ*poxB*	Jantama et al. 2008
XZ320	KJ073, Δ*ppc*	This study
XZ332	KJ073, Δ*pck*	This study
XZ341	KJ073, Δ*sfcA*	This study
XZ396	KJ073, Δ*maeB*	This study
XZ613	KJ017, Δ*ptsI*	This study
XZ615	KJ060, Δ*ptsI*	This study
XZ616	KJ060, restored *ptsI* to wild type	This study
XZ618	KJ017, *pck**	This study
XZ620	KJ071, *pck**	This study
XZ622	KJ060, restored *pck* to wild type	This study
XZ624	KJ073, restored *pck* to wild type	This study
XZ626	KJ017, Δ*cra*	This study
XZ627	KJ060, Δ*cra*	This study
XZ642	KJ012, Δ*crp*	This study
XZ643	KJ017, Δ*crp*	This study

Plasmids		
pLOI4677	*bla kan*; *pck* (including ribosomal binding site, coding and terminator fragment) from *E.coli* ATCC8739 cloned into pCR2.1-TOPO vector	This study
ptsI mutation change		
pLOI4734	*bla kan*; *ptsI* (*ptsI*-D-up/D-down) from *E.coli* ATCC8739 cloned into pCR2.1-TOPO vector	This study
pLOI4735	*cat-sacB* cassette (*Sma*I-*Sfo*I fragment of pLOI4162) cloned into *ptsI* of pLOI4734	This study
pck mutation change		
pLOI4736	*bla kan*; *pck-P* (pck-Pro-up/Pro-down) from *E.coli* ATCC873 cloned into pCR2.1-TOPO vector	This study
pLOI4737	*cat-sacB* cassette (*Sma*I-*Sfo*I fragment of pLOI4162) cloned into *pck-P* of pLOI4736 (pck-Pro-1/Pro-2)	This study
Primers		
pck mutation change		
pck-Pro-up	CACGGTAGCAACAACATTGC (SEQ ID NO: 78)	This study
pck-Pro-down	AGAAAGCGTCGACAACGAAC (SEQ ID NO:79)	
pck-Pro-1	ATGCGCGTTAACAATGGTTT (SEQ ID NO: 80)	
pck-Pro-2	ATGGATAACGTTGAACTTTC (SEQ ID NO: 81)	
ptsI mutation change		
ptsI-D-up	CGCATTATGTTCCCGATGAT (SEQ ID NO: 82)	This study
ptsI-D-down	GCCTTTCAGTTCAACGGTGT (SEQ ID NO: 83)	
ptsI-D-1	CGGCCCAATTTACTGCTTAG (SEQ ID NO: 84)	
ptsI-D-2	ATCCCCAGCAACAGAAGTGT (SEQ ID NO: 85)	
pck sequencing		

pck-F	TTGGCTAAGGAGCAGTGAAATGCGCGTTA (SEQ ID NO: 86)	This study
pck-R	CACGACAAAAGAAGGGTAAATAAAC (SEQ ID NO: 87)	
pck-2	TTGTTAACGCGCATTTCACT (SEQ ID NO: 88)	
pck-3	GCGATAGCGGCTACTGTCAT (SEQ ID NO: 89)	

*pck** represents mutated form of *pck containing, a* G to A transition at -64 (ATG).

All strains are of derivatives of *E. coli* ATCC8739.

Table 10. Pck is critical for glucose fermentation to succinate by derivatives of KJ073

Strain*	Genetic modification	Glucose consumed	Succinate (mM)	Yield† (mol/mol)	Acetate (mM)	Pyruvate (mM)
KJ073	none	301	364	1.21	113	3
XZ320	KJ073, Δ*ppc*	278	348	1.25	100	-
XZ341	KJ073, Δ*sfcA*	278	339	1.22	106	-
XZ396	KJ073, Δ*maeB*	278	337	1.22	116	0
XZ332	KJ073, Δ*pck*	92	42	0.46	28	32
XZ332 (pLOI4677)	Δ*pck*, *pck* over-expression	273	260	0.95	78	32

*All fermentations were performed in NBS medium with 5% glucose and 100 mM potassium bicarbonate.

†Yield was calculated as mol succinate produced per mol glucose metabolized.

Table 11. Comparison of carboxylation enzymes in engineered *E. coli* strains

Strain	Cell yield (g/L)	Succinate productivity (g/L/h)	Enzyme activity* [nmol min^{-1} (mg protein) $^{-1}$]				*pck* mRNA	G to A change
			Pck	Ppc	SfcA	MaeB		
E.coli ATCC	2.0	0.12	295	20	ND	<1	1.0	No
KJ012‡	0.3	0.04	162	25	5	<1	0.2	No
KJ017	1.7	0.35	700	17	12	<1	2.7	No
KJ060	2.2	0.90	8,363	30	11	<1	7.2	Yes
KJ071	1.5	0.33	679	23	10	<1	2.5	No
KJ073	2.3	0.82	7,341	27	12	<1	8.4	Yes
E.coli K12†	ND	ND	140	140	ND	ND	ND	ND
Actinobacillus Succinogenes†	ND	ND	4700	10	ND	ND	ND	ND

*Pck, PEP carboxykinase; Ppc, PEP carboxylase; SfcA, NADH-linked malic enzyme; MaeB, NADPH-linked malic enzyme; ND, not determined.

†Data from Vanderwerf *et al.*, *Arch Microbiol* 167:332-342

‡ Poor growth

Table 12. Effects of mutations in *pck, ptsI, and cra* on PEP carboxykinase activity.

Strain	Genotype	PEP-carboxykinase (U / mg protein)
KJ017†	*pck*$^+$, *ptsI*$^+$, *cra*$^-$	700
XZ618	KJ017, *pck**	6,419
XZ613	KJ017,)*ptsI*	614
XZ626	KJ017,)*cra*	508
KJ060‡§	*pck**, *ptsI*∇, - *cra*$^+$	8,363
XZ622‡	KJ060, - *pck*$^+$	1,103
XZ615	KJ060,)*ptsI*	10,309
XZ627	KJ060,)*cra*	7181
KJ071†§	*ptsI*∇,– *pck*$^+$, *cra*$^+$	679
XZ620	KJ071, *pck**	6,193
KJ073‡§	*pck**, *ptsI*∇, - *cra*$^+$	7,341
XZ624‡	KJ073, – *pck*$^+$	904

†The *pck* in KJ017 and KJ071 was mutated (G to A) to *pck** in XZ618 and XZ620, respectively.

‡The *pck** mutation in KJ060 and KJ073 was restored to wild type (A to G, - *pck*$^+$) in XZ622 and XZ624.

§The abbreviation *ptsI*∇ refers to a frame-shift mutation, a single base deletion in the carboxy-terminal region.

Table 13. Effect of glucose and cAMP on PEP carboxykinase (*pck*) and β-galactosidase (*lacZ*) activities*

Strain	PEP carboxykinase activity†				β-galactosidase activity ($\times 10^3$)†		
	5% Glu	no Glu	ratio‡	Glu + cAMP	5% Glu	no Glu	ratio‡
8739	141	476	3.4	553	5.0	15.0	3.0
KJ012	146	462	3.2	737	7.9	19.0	2.4
KJ017	587	624	1.1	997	14.2	19.0	1.3
KJ060	5574	2777	0.5	4253	18.4	20.0	1.1
XZ642	118	113	1.0	112	-	-	-
XZ643	116	124	1.0	118	-	-	-

*Cultures were grown aerobically in shaken flasks with Luria broth.

†Activity is the average of two repeats and is expressed as U (mg protein)$^{-1}$.

‡Ratio is the activity without glucose to the activity with 5% glucose.

Table 14. Plasmids and primers used in Example 3	
Plasmids	
pCR2.1-TOPO	*bla kan*; TOPO TA cloning vector
pLOI4162	*bla cat;* plasmid to provide *cat-sacB* cassette
***sfcA* deletion**	
pLOI4283	*bla kan*; *sfcA* (PCR) from ATCC 8739 cloned into pCR2.1-TOPO vector
pLOI4284	*cat-sacB* cassette (*Sma*I-*Sfo*I fragment of pLOI4162) cloned into *sfcA* of pLOI4284
pLOI4285	*Pac*I digestion of pLOI4284, and self-ligated
***ppc* deletion**	
pLOI4264	*bla kan*; *ppc* (PCR) from ATCC 8739 cloned into pCR2.1-TOPO vector
pLOI4265	*cat-sacB* cassette (*Sma*I-*Sfo*I fragment of pLOI4162) cloned into *ppc* of pLOI4264
pLOI4266	*Pac*I digestion of pLOI4265, and self-ligated
***pck* deletion**	
pLOI4641	*bla kan*; *pck* (PCR) from ATCC 8739 cloned into pCR2.1-TOPO vector
pLOI4642	*cat-sacB* cassette (*Sma*I-*Sfo*I fragment of pLOI4162) cloned into *ppc* of pLOI4641
pLOI4643	*Pac*I digestion of pLOI4642, and self-ligated
***maeB* deletion**	
pLOI4728	*bla kan*; *maeB* (PCR) from ATCC 8739 cloned into pCR2.1-TOPO vector
pLOI4729	*cat-sacB* cassette (*Sma*I-*Sfo*I fragment of pLOI4162) cloned into *maeB* of pLOI4728
pLOI4730	*Pac*I digestion of pLOI4729, and self-ligated
ptsI mutation change	
pLOI4734	*bla kan*; *ptsI* (*ptsI*-D-up/D-down) from ATCC 8739 cloned into PCR2.1-TOPO vector
pLOI4735	*cat-sacB* cassette (*Sma*I-*Sfo*I fragment of pLOI4162) cloned into *ptsI* of pLOI4734
***pck* mutation**	

change	
pLOI4736	*bla kan*; *pck-P* (pck-Pro-up/Pro-down) from ATCC 8739 cloned into PCR2.1-TOPO vector
pLOI4737	*cat-sacB* cassette (*Sma*I-*Sfo*I fragment of pLOI4162) cloned into *pck-P* of pLOI4736 (pck-Pro-1/Pro-2)
Primers for gene deletions	
***sfcA* deletion**	
sfcA-up	CTATGCTTGATCGGCAACCT (SEQ ID NO: 90)
sfcA-down	ACGATCGCCTGGTTTTAATG (SEQ ID NO: 91)
sfcA-1	TACCGCCGTACCTCCATCTA (SEQ ID NO: 92)
sfcA-2	CGTAAGGGATATAAAGCGAACG (SEQ ID NO: 93)
***ppc* deletion**	
ppc-up	TCAAACGATGCCCAACTGTA (SEQ ID NO: 94)
ppc-down	TTTAATCCGCTTCGGAAAGA (SEQ ID NO: 95)
ppc-1	GTCACTATTGCCGGGATTGC (SEQ ID NO: 96)
ppc-2	CAATGCGGAATATTGTTCGT (SEQ ID NO: 97)
***pck* deletion**	
pck-up	TCCGGGCAGTAGTATTTTGC (SEQ ID NO: 98)
pck-down	ATGGCTGGATCAAAGTCAGC (SEQ ID NO: 99)
pck-1	CCTGGCGAAACTGTTTATCG (SEQ ID NO: 100)
pck-2	TTGTTAACGCGCATTTCACT (SEQ ID NO: 101)
***maeB* deletion**	
maeB-up	GCATCCTGGGGATGATAATG (SEQ ID NO: 102)
maeB-down	TTTCTTCGCCAGTTCCTCAC (SEQ ID NO: 103)
maeB-1	AACCCAACCGCTGTAATTTTT (SEQ ID NO: 104)
maeB-2	CTGGAACTGGAAATTCATGG (SEQ ID NO: 105)
***pck* mutation change**	

pck-Pro-up	CACGGTAGCAACAACATTGC (SEQ ID NO: 106)
pck-Pro-down	AGAAAGCGTCGACAACGAAC (SEQ ID NO: 107)
pck-Pro-1	ATGCGCGTTAACAATGGTTT (SEQ ID NO: 108)
pck-Pro-2	ATGGATAACGTTGAACTTTC (SEQ ID NO: 109)
***ptsI* mutation change**	
ptsI-D-up	CGCATTATGTTCCCGATGAT (SEQ ID NO: 110)
ptsI-D-down	GCCTTTCAGTTCAACGGTGT (SEQ ID NO: 111)
ptsI-D-1	CGGCCCAATTTACTGCTTAG (SEQ ID NO: 112)
ptsI-D-2	ATCCCCAGCAACAGAAGTGT (SEQ ID NO: 113)
Primers for sequencing*	
pck-F	TTGGCTAAGGAGCAGTGAAATGCGCGTTA (SEQ ID NO: 114)
pck-R	CACGACAAAAGAAGGGTAAATAAAC (SEQ ID NO: 115)
pck-2	TTGTTAACGCGCATTTCACT (SEQ ID NO: 116)
pck-3	GCGATAGCGGCTACTGTCAT (SEQ ID NO: 117)
cra-up	GCGGTAAGCTTGATGCATTT (SEQ ID NO: 118)
cra-down	CTTCCCCGGTTAACAGTCCT (SEQ ID NO: 119)
ptsHI-up1	TCATCGGGTGAGCGTTATTT (SEQ ID NO: 120)
ptsHI-down1	TGACCGTCCAGCGTAATAGC (SEQ ID NO: 121)
ptsHI-up2	CATCCTGGGCCTGAAGATTA (SEQ ID NO: 122)
ptsHI-down2	AGCAATACCATCACCAACGA (SEQ ID NO: 123)
crr-up	CCCGCGCATTAAGAAGATTA (SEQ ID NO: 124)
crr-down	CTCATCAGTGGCTTGCTGAA (SEQ ID NO: 125)
ptsG-up	GAAGAACTGGCGCAGGTAAC (SEQ ID NO: 126)
ptsG-down	AAGGAAACGCCGTTAATCCT (SEQ ID NO: 127)
cyaA-up	TCGCCATCAACTTGTCTTTG (SEQ ID NO: 128)
cyaA-down	AAAGGCGATGAGTGGATTTG (SEQ ID NO: 129)
crp-S-up	TGAGTTGCCGTCCATTAAAA (SEQ ID NO: 130)

crp-S-down	AATCGTAATTCGCCAAGCAT (SEQ ID NO: 131)
cpdA-S-up	GAAGTGTGTTCAAGCCAGCA (SEQ ID NO: 132)
cpdA-S-down	AGGACAATGGATTCCAGCAG (SEQ ID NO: 133)
ygiF-S-up	ATCAGTGTCGCTACGCAAAG (SEQ ID NO: 134)
ygiF-S-down	GCTGTCCTGCACAAAATCAC (SEQ ID NO: 135)
sxy-S-up	TTTACTTGCTGCGGATGAGA (SEQ ID NO: 136)
sxy-S-down	TATCTCAGCCCTCGGTGCTC (SEQ ID NO: 137)
csrA-up	CAGCGTTAGCCAGTGTGAAA (SEQ ID NO: 138)
csrA-down	ACGCCTCTTACGAGTGCTTC (SEQ ID NO: 139)
csrB-S-up	CTGTAGGAGATCGCCAGGAA (SEQ ID NO: 140)
csrB-S-down	TCTAACAAATCGTGCATTCG (SEQ ID NO: 141)
csrC-S-up	GCCATACGCTTTGTGAGACA (SEQ ID NO: 142)
csrC-S-down	AGTCACGCCCAATGGAATAG (SEQ ID NO: 143)
csrD-S-up1	ATGTGCATGATGGATTGGAA (SEQ ID NO: 144)
csrD-S-down1	CGGTATCCTGACCACTACGC (SEQ ID NO: 145)
csrD-S-up2	GTGATTTTGCTGCGCTGTTA (SEQ ID NO: 146)
csrD-S-down2	ACAAGGCGCAAAAATCATCT (SEQ ID NO: 147)
uvrY-s-up1	CCTCGTCATGTTGCAATGAA (SEQ ID NO: 148)
uvrY-s-down1	TATCATCGCGTAGCAAAACG (SEQ ID NO: 149)
barA-s-up1	TTTTGCTTCGCTGCTGTAAA (SEQ ID NO: 150)
barA-s-down1	TCAGGCACGTCGCTTTTAAT (SEQ ID NO: 151)
barA-s-up2	CGCGATCACCTGAATACGAT (SEQ ID NO: 152)
barA-s-down2	CTGGCTGGACGTTCGATAAC (SEQ ID NO: 153)
barA-s-up3	TGGCCTATGTCGAACCAAAC (SEQ ID NO: 154)
mlc-s-up1	CTGGCAAATAACCCGAATGT (SEQ ID NO: 155)
mlc-s-down1	CCAGGGCATCTTTATTACGC (SEQ ID NO: 156)
mlc-s-up2	TGAAACTGAAGCCTGGCACT (SEQ ID NO: 157)

*The primers for sequencing were used to amplify the upstream, coding and terminator fragment of all genes.

Table 15. Sources and characteristics of *E. coli* strains, plasmids and primers used in this study

	Relevant characteristics	Source or reference
Strains		
ATCC 8739	Wild type	Lab collection
KJ012	Δ*ldhA*, Δ*adhE*, Δ*ackA*	Jantama et al., 2008a
XZ02	Δ*ldhA*	This study
XZ04	Δ*ldhA*, Δ*pflB*	This study
XZ14	Δ*ldhA*, Δ*adhE*	This study
XZ15	Δ*adhE*	This study
XZ17	Δ*pflB*	This study
XZ464	*pck**, Δ*gldA*	This study
XZ465	*pck**, Δ*dhaKL*	This study
XZ466	*pck**, Δ*dhaM*	This study
XZ468	*pck**, Δ*ptsH*	This study
XZ469	*pck**, Δ*ptsI-W*	This study
XZ470	*pck**, Δ*ptsG*	This study
XZ629	Δ*ldhA*, Δ*adhE*, Δ*ackA*, *pck**	This study
XZ632	*pck**	This study
XZ635	Δ*ldhA*, *pck**	This study
XZ638	Δ*ldhA*, Δ*adhE*, *pck**	This study

XZ639	Δ*adhE*, *pck**	This study
XZ640	Δ*pflB*, *pck**	This study
XZ641	Δ*ldhA*, Δ*pflB*, *pck**	This study
XZ647	*pck**, Δ*ptsI*	This study
XZ650	Δ*ptsI*	This study
XZ721	*pck**, Δ*ptsI*, Δ*pflB*	This study
XZ723	*pck**, Δ*ptsI*, Δ*adhE*	This study
Plasmids		
pLOI4162	*bla*, *cat sacB* cassette	Jantama et al., 2008b
***ldhA* deletion**		
pLOI4652	*bla kan*; *ldhA* (PCR) from *E. coli* cloned into pCR2.1-TOPO vector	This study
pLOI4653	*cat-sacB* cassette cloned into *ldhA* of pLOI4652	This study
pLOI4655	*Pac*I digestion of pLOI4653, and self-ligated	This study
***pflB* deletion**		
pLOI4667	*bla kan*; *pflB* (PCR) from *E.coli* cloned into pCR2.1-TOPO vector	This study
pLOI4668	*cat-sacB* cassette cloned into *pflB* of pLOI4667	This study
pLOI4669	*Pac*I digestion of pLOI4668, and self-ligated	This study
***adhE* deletion**		
pLOI4707	*bla kan*; *adhE* (PCR) from *E.coli* cloned into pCR2.1-TOPO vector	This study
pLOI4708	*cat-sacB* cassette cloned into *adhE* of pLOI4707	This study
pLOI4709	*Pac*I digestion of pLOI4707, and self-ligated	This study

***pck* promoter change**		
pLOI4736	*bla kan*; *pck-P* from *E.coli* cloned into pCR2.1-TOPO vector	This study
pLOI4737	*cat-sacB* cassette cloned into *pck-P* of pLOI4736	This study
pLOI4752	*trc* promoter cloned into *pck-P* of pLOI4736	This study
***ptsI* deletion**		
pLOI4734	*bla kan*; *ptsI* from *E.coli* cloned into pCR2.1-TOPO vector	This study
pLOI4735	*cat-sacB* cassette cloned into *ptsI* of pLOI4734	This study
pLOI4735B	*Pac*I digestion of pLOI4735, and self-ligated	This study
***ptsG* deletion**		
pLOI4683	*bla kan*; *ptsG* from *E.coli* cloned into PCR2.1-TOPO vector	This study
pLOI4684	*cat-sacB* cassette (*Sma*I-*Sfo*I fragment of pLOI4162) cloned into *ptsG* of pLOI4683	This study
pLOI4685	*Pac*I digestion of pLOI4684, and self-ligated	This study
***gldA* deletion**		
pLOI4296	*bla kan*; *gldA* from *E.coli* cloned into PCR2.1-TOPO vector	This work
pLOI4297	*cat-sacB* cassette from pLOI4151 cloned into *gldA* of pLOI4296	This work
pLOI4298	PCR fragment amplified from pLOI4296 (using gldA -1/ gldA -2), kinase treated, and then self-ligated	This work
Primers		
***ldhA* deletion**		
XZ-ldhA-up	GATAACGGAGATCGGGAATG (SEQ ID NO: 158)	This study

XZ-ldhA-down	CTTTGGCTGTCAGTTCACCA (SEQ ID NO: 159)	
XZ-ldhA-1	TCTGGAAAAAGGCGAAACCT (SEQ ID NO: 160)	
XZ-ldhA-2	TTTGTGCTATAAACGGCGAGT (SEQ ID NO: 161)	
***pflB* deletion**		
PflB-up2	TGTCCGAGCTTAATGAAAAGTT(SEQ ID NO: 162)	This study
PflB-down2	CGAGTAATAACGTCCTGCTGCT (SEQ ID NO: 163)	
PflB-5	AAACGGGTAACACCCCAGAC (SEQ ID NO: 164)	
PflB-6	CGGAGTGTAAACGTCGAACA (SEQ ID NO: 165)	
***adhE* deletion**		
adhE-up	CATGCTAATGTAGCCACCAAA (SEQ ID NO: 166)	This study
adhE-down	TTGCACCACCATCCAGATAA (SEQ ID NO: 167)	
adhE-1	TCCGGCTAAAGCTGAGAAAA (SEQ ID NO: 168)	
adhE-2	GTGCGTTAAGTTCAGCGACA (SEQ ID NO: 169)	
***pck* mutation change**		
pck-Pro-up	CACGGTAGCAACAACATTGC (SEQ ID NO: 170)	This study
pck-Pro-down	AGAAAGCGTCGACAACGAAC (SEQ ID NO: 171)	
pck-Pro-1	ATGCGCGTTA ACAATGGTTT (SEQ ID NO: 172)	
pck-Pro-2	ATGGATAACG TTGAACTTTC (SEQ ID NO: 173)	
pck-P-2	TTCACTGCTC CTTAGCCAAT (SEQ ID NO: 174)	
***ptsI* deletion**		
ptsI-D-up	CGCATTATGTTCCCGATGAT (SEQ ID NO: 175)	This study
ptsI-D-down	GCCTTTCAGTTCAACGGTGT (SEQ ID NO: 176)	
ptsI-D-1	CGGCCCAATTTACTGCTTAG (SEQ ID NO: 177)	
ptsI-D-2	ATCCCCAGCAACAGAAGTGT (SEQ ID NO: 178)	

***ptsG* deletion**		
ptsG-up	GAAGAACTGGCGCAGGTAAC (SEQ ID NO: 179)	This study
ptsG-down	AAGGAAACGCCGTTAATCCT (SEQ ID NO: 180)	
ptsG-1	CCTGAAAACCGAGATGGATG (SEQ ID NO: 181)	
ptsG-2	CATCAGCGATTTACCGACCT (SEQ ID NO: 182)	
***ptsH* deletion**		
ptsH-D-up	ATGTTCCAGCAAGAAGTTACCATTACCGCTCCG AACGGTCTGCAC GTGTAGGCTGGAGCTGCTTC (SEQ ID NO: 183)	This study
ptsH-D-down	TTACTCGAGTTCCGCCATCAGTTTAACCAGATG TTCAACCGCTTT CATATGAATATCCTCCTTAG (SEQ ID NO: 184)	
***ptsI-W* deletion**		
ptsI-D-up	ATGATTTCAGGCATTTTAGCATCCCCGGGTATCG CTTTCGGTAAA GTGTAGGCTGGAGCTGCTTC (SEQ ID NO: 185)	This study
ptsI-D-down	TTAGCAGATTGTTTTTTCTTCAATGAACTTGTTA ACCAGCGTCAT CATATGAATATCCTCCTTAG (SEQ ID NO: 186)	
***gldA* deletion**		
gldA-up	TGTATATAGCGCCGCACAAG (SEQ ID NO: 187)	This study
gldA-down	GATGGCAAGGTTGGTATTGG (SEQ ID NO: 188)	
gldA-1	ACCAGTACGGTCAGCGTTTC (SEQ ID NO: 189)	
gldA-2	ACCCGGTGATTGAATAATGC (SEQ ID NO: 190)	
***dhaKL* deletion**		
dhaKL-D-up	ATGAAAAAATTGATCAATGATGTGCAAGACGT ACTGGACGAACAA TGTAGGCTGGAGCTGCTTC (SEQ ID NO: 191)	This study

dhaKL-D-down	TTACTCTTTTGCGGCTAACGCCAACATTTGCA TCATAAACATCAC CATATGAATATCCTCCTTAG (SEQ ID NO: 192)	
***dhaM* deletion**		
dhaM-D-up	GTGATGGTAAACCTGGTCATAGTTTCACATA GCAGCCGACTGGGATGTAGGCTGGAGCTGCTTC (SEQ ID NO: 193)	This study
dhaM-D-down	TTAACCCTGACGGTTGAAACGTTGCGTTTTA ACGTCCAGCGTTAGATATGAATATCCTCCTTAG (SEQ ID NO: 194)	

Table 16. The effects of inactivating alternative NADH oxidizing pathways on succinate production from glucose using NBS mineral salts medium and complex medium (Luria broth)

Strains	Genetic modification	Media	Time (days)	Growth Cell yield (g/L)	Gluc used (mmol)	Suc Yield[1]	Fermentation products (mmol) [2]						
							Suc	Ace	Mal	Pyr	Lac	For	EtOH
ATCC 8739	Wild type	NBS	3	3.0	278	0.16	46	151	-	-	110	318	133
XZ02	Δ*ldhA*	NBS	3	2.9	278	0.15 (-6%)	43	165	-	-	11	353	199
XZ15	Δ*adhE*	NBS	3	2.7	278	0.18 (+12%)	50	84	-	-	352	102	-
XZ17	Δ*pflB*	NBS	3	2.2	278	0.09 (-44%)	26	11	-	-	460	-	-
XZ14	Δ*ldhA*, Δ*adhE*	NBS	6	0.5	57	0.12 (-25%)	6	48	-	-	-	44	-
XZ04	Δ*ldhA*,Δ*pflB*	NBS	6	0.5	39	0.13 (-19%)	5	22	-	-	-	22	12
KJ012	Δ*ldhA*, Δ*adhE*, Δ*ackA*	NBS	6	0.3	46	0.13 (-19%)	6	26	-	-	-	-	-
ATCC 8739	Wild type	LB	2	2.0	257	0.13	34	138	-	-	214	161	127
XZ02	Δ*ldhA*	LB	2	1.8	257	0.21 (+61%)	55	224	-	-	17	152	206

XZ15	Δ*adhE*	LB	2	1.9	268	0.11 (-15%)	30	43	-	-	459	-	-
XZ17	Δ*pflB*	LB	2	2.0	261	0.09 (-31%)	24	8	-	-	469	-	-
XZ14	Δ*ldhA*, Δ*adhE*	LB	6	0.3	31	0.23 (+77%)	7	45	-	-	-	17	-
XZ04	Δ*ldhA*, Δ*pflB*	LB	6	1.2	150	0.33 (+154%)	48	9	-	86	35	23	89
KJ012	Δ*ldhA*, Δ*adhE*, Δ*ackA*	LB	6	1.5	154	0.70 (+438%)	108	61	-	-	-	14	-

[1] Succinate yield was calculated as moles of succinate produced per mol glucose metabolized. The effects of inactivating alternative NADH oxidizing pathway(s) on succinate yield are shown in the parentheses. A negative sign (–) indicates a decrease in yield and a positive sign (+) indicates an increase in yield as compared to that of the wild type under the same conditions.

[2]Fermentations were carried out in either NBS mineral salts medium or Luria broth medium with 5% glucose and 100 mM potassium bicarbonate (37°C, pH 7.0, 150 rpm). Abbreviations: Suc, succinate; Ace, acetate; Mal, malate; Pyr, pyruvate; Lac, lactate; For, formate; EtOH, ethanol.

Table 17. The effects of increasing PCK activity on succinate production and yield from glucose in NBS mineral salts medium

Strains	Genetic modification[1]	Growth		PCK activity [3]	Glu used (mmol)	Suc Yield[4]	Fermentation products (mmol) [5]						
		Day	Cell yield (g/L)				Suc	Ace	Mal	Pyr	Lac	For	EtOH
ATCC 8739	Wild type	3	3.0	203	278	0.16	46	151	-	-	110	318	133
ATCC 8739	pLOI4677 [2]	3	2.8	1782	278	0.27	73	141	-	24	112	266	116
XZ632	*pck**	3	2.9	1631	278	0.22	61	161	-	-	88	302	152
XZ635	Δ*ldhA*, *pck**	3	2.7	1697	278	0.22	60	151	-	-	13	327	178
XZ639	Δ*adhE*, *pck**	3	2.7	1608	278	0.25	70	80	-	-	329	99	-
XZ640	Δ*pflB*, *pck**	3	2.5	1623	278	0.14	39	12	-	-	448	-	-
XZ638	Δ*ldhA*, Δ*adhE*, *pck**	6	0.8	917	66	0.24	16	71	-	-	-	79	-
XZ641	Δ*ldhA*, Δ*pflB*, *pck**	6	0.8	932	61	0.39	24	9	-	-	-	2	15

[1] The abbreviation, *pck**, denotes mutated form of *pck* (G to A at –64 relative to the ATG start).

[2] The *pck* gene (ribosomal binding site, coding and terminator region) was over-expressed in a high-copy plasmid.

[3] PCK activity expressed as nmol min^{-1} (mg protein)$^{-1}$

[4] Succinate yield was calculated as moles of succinate produced per mol glucose metabolized.

[5] Fermentations were carried out in NBS mineral salts medium with 5% glucose and 100 mM potassium bicarbonate (37°C, pH 7.0, 150 rpm). Abbreviations: Suc, succinate; Ace, acetate; Mal, malate; Pyr, pyruvate; Lac, lactate; For, formate; EtOH, ethanol

Table 18. Effects of combining a mutation in *ptsI*, mutations that disrupted the ethanol pathway, and increased expression of *pck* on succinate production in NBS mineral salts medium containing 5% glucose.

Strains	Genetic modification [1]	Growth		Gluc used (mmol)	Suc Yield [3]	Fermentation products (mmol) [4]						
		Day	Cell yield			Suc	Ace	Mal	Pyr	Lac	For	EtOH
ATCC 8739	Wild type ATCC 8739	3	3.0	278	0.16	46	151	-	-	110	318	133
XZ632	*pck**	3	2.9	278	0.22	61	161	-	-	88	302	152
XZ650	Δ*ptsI*	3	2.6	259	0.14	35	131	-	28	130	273	119
XZ647	*pck**, Δ*ptsI*	3	2.5	242	0.89	216	174	5	-	5	199	22
XZ650	Δ*ptsI*, pLOI4677 [2]	3	2.4	224	0.75	167	167	3	2	8	224	52
XZ721	*pck**, Δ*ptsI*, Δ*pflB*	3	2.3	262	1.25	327	70	-	-	29	-	-
XZ723	*pck**, Δ*ptsI*, Δ*adhE*	3	2.4	259	0.86	222	175	-	-	55	171	-

[1] The abbreviation, *pck**, denotes mutated form of *pck* (G to A at –64 relative to the ATG start).

[2] The *pck* gene (ribosomal binding site, coding and terminator region) was over-expressed in a high-copy plasmid

[3] Succinate yield was calculated as moles of succinate produced per mol glucose metabolized.

[4] Fermentations were carried out in NBS mineral salts medium with 5% glucose and 100 mM potassium bicarbonate (37°C, pH 7.0, 150 rpm). Abbreviations: Suc, succinate; Ace, acetate; Mal, malate; Pyr, pyruvate; Lac, lactate; For, formate; EtOH, ethanol.

Table 19. Effects of combining a mutation in phosphoenolpyruvate-dependent phosphotransferase pathway, and increased expression of *pck* on succinate production in NBS mineral salts medium containing 5% glucose

Strains	Genetic modification [1]	Growth		Gluc used (mmol)	Suc Yield [2]	Fermentation products (mmol) [3]						
		Day	Cell yield			Suc	Ace	Mal	Pyr	Lac	For	EtOH
XZ632	*pck**	3	2.9	278	0.22	61	161	-	-	88	302	152
XZ647	*pck**, Δ*ptsI*	3	2.5	242	0.89	216	174	5	-	5	199	22
XZ469	*pck**, Δ*ptsI-W*	3	3.0	245	0.84	206	143	-	-	3	164	31
XZ468	*pck**, Δ*ptsH*	3	2.6	266	0.67	179	151	-	-	17	242	69
XZ470	*pck**, Δ*ptsG*	3	2.5	269	0.63	170	175	-	-	32	245	63

[1] The abbreviation, *pck**, denotes mutated form of *pck* (G to A at –64 relative to the ATG start). Δ*ptsI* denotes deletion of the carboxy-terminal 175 bp of *ptsI* gene, while Δ*ptsI-W* denotes deletion of the whole *ptsI* gene.

[2] Succinate yield was calculated as mole of succinate produced per mol glucose metabolized.

[3] Fermentations were carried out in NBS medium with 5% glucose and 100 mM potassium bicarbonate at 37°C, pH 7.0, 150 rpm.

Abbreviations: Suc, succinate; Ace, acetate; Mal, malate; Pyr, pyruvate; Lac, lactate; For, formate; EtOH, ethanol.

Table 20. Succinate production from glycerol by engineered *E. coli* strains in NBS mineral salts medium

Strains	Genetic modification	Time (Day)	Cell mass (g/l)	Glycerol used (mmol)	Fermentation products (mmol) [1]					
					Suc	Suc yield	For	EtOH	Lac	Ace
ATCC 8739	Wild type	6	0.55	153	38	0.25	110	81	-	16
XZ17	*ΔpflB*	6	0.3	48	5	0.11	-	-	20	5
XZ640	*ΔpflB, pck**	6	0.4	79	37	0.48	-	-	16	10
XZ721	*ΔpflB, pck*, ΔptsI*	6	0.5	128	102	0.8	-	-	6	12
XZ632	*pck**	6	0.7	149	64	0.44	83	46	-	20
XZ647	*pck*, ΔptsI*	6	0.4	89	63	0.71	25	5	-	13
XZ464	*pck*, ΔgldA*	6	0.43	125	95	0.77	35	8	-	16
XZ465	*pck*, ΔdhaKL*	6	0.47	150	112	0.75	37	11	-	15
XZ466	*pck*, ΔdhaM*	6	0.5	152	108	0.71	30	9	-	14
XZ468	*pck*, ΔptsH*	6	0.5	125	85	0.68	32	14	-	10

Fermentations were carried out in NBS medium with 5% glycerol and 100 mM potassium bicarbonate at 37°C, pH 7.0, 150 rpm. Succinate yield was calculated based on mol succinate produced per mol glycerol consumed. Abbreviations: Suc, succinate; For, formate; EtOH, ethanol; Lac, lactate; Ace, acetate.

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CLAIMS

We claim:

1. An *Escherichia coli* bacterial cell comprising increased levels of phosphoenol pyruvate carboxykinase (*pck*) gene transcripts and one or more genetic modifications in one or more genes selected from the group consisting of *ptsG*, *ptsH* and *ptsI*, said transcripts encoding active phosphoenol pyruvate carboxykinase (PCK), said cell exhibiting increased PCK activity and wherein said one or more genetic modifications inactivate the function of the gene product encoded by *ptsG*, *ptsH* and *ptsI* and is selected from deleting the entire coding region of the gene, deleting a portion of the coding region of the gene, introducing a frame shift mutation in the coding region of the gene, introducing a missense mutation in the gene, insertion or introducing a stop codon in the coding region of the gene or combinations thereof.

2. The bacterial cell of claim 1, wherein increased levels of said *pck* transcripts result from replacement of native promoter sequences with altered promoter sequences of the *pck* gene, wherein the altered promoter sequence causes an increase in the *pck* transcription.

3. The bacterial cell of claim 1, wherein said increased levels of said *pck* transcripts result from one or more mutations in the promoter region of the *pck* gene.

4. The bacterial cell of claim 3, wherein said one or more mutations are point mutations comprising replacement of nucleotide G with nucleotide A at position 64 up stream of the *pck* gene start codon.

5. The bacterial cell of claim 1, wherein increased levels of said *pck* transcripts result from replacement of the native promoter sequence with an exogenous promoter sequence.

6. The bacterial cell of claim 5, wherein said exogenous promoter is a constitutive promoter.

7. The bacterial cell of claim 5 wherein said exogenous promoter is an inducible promoter.

8. The bacterial cell of claim 7, wherein the said inducible promoter is a *lac* promoter.

9. The bacterial cell of claim 1, further comprising a genetic modification leading to the inactivation of gene expression in one or more genes selected from the group consisting of *adhE* and *pflB*.

10. A genetically modified *Escherichia coli* bacterial cell, wherein said genetically modified bacterial cell is NRRL B-50279, NRRL B-50280, or NRRL B-50281.

11. A method of producing succinic acid comprising:

(a) Culturing a bacterial cell of claim 1;

(b) Providing a carbon source;

(c) Allowing said bacteria to metabolize said carbon source; and

(d) Isolating succinic acid

12. The method of claim 11, wherein said bacterial cell is cultured in anaerobic condition, aerobic condition, microaerobic condition or combinations thereof.

13. The method of claim 11, wherein said bacterial cell is cultured in a minimal salt growth medium.

14. The method of claim 13, wherein said minimal salt growth medium comprises between 2% and 20% (w/v) carbon source.

15. The method of claim 11, wherein the said carbon source is glucose, fructose, xylose, arabinose, galactose, mannose, rhamnose, sucrose, cellobiose, hemicelluloses, glycerol or combination thereof.

16. The method of claim 11, wherein said culturing is a batch process or fed batch process.

17. The method of claim 11, wherein said culturing is a continuous process.

18. The bacterial cell according to any one of claims 1, 2, 3-8 or 9, wherein said *pck* transcripts are endogenous *pck* transcripts.