TITLE OF THE INVENTION

METHODS AND COMPOSITIONS FOR TREATING VIRAL DISEASES

CROSS REFERENCE TO RELATED APPLICATIONS

This application claims the priority of U.S. Provisional Appl. Ser. No. 61/567,491, filed December 6, 2011, the entire disclosure of which is incorporated herein by reference.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH

This invention was made with Government support under contract AI079406 awarded by the National Institutes of Health. The Government has certain rights in this invention.

FIELD OF THE INVENTION

The present invention relates to virology. More specifically, the invention relates to treatment of viral diseases through inhibition of interactions of host cellular clathrin adaptor proteins and viral YXX<l> or dileucine motifs or their regulation.

BACKGROUND OF THE INVENTION

Human infections with Lentiviridae, such as HIV, and Flaviviridae, such as HCV, pose significant challenges to global health. Although potent anti-HCV drugs are in clinical development and response rates to interferon-based regimens have improved with the inclusion of protease inhibitors **(Pl),** resistance, drug-drug interactions, and cumulative toxicity continue to pose challenges. More effective antiviral strategies are therefore still in need to combat the HCV pandemics. In addition, novel antiviral strategies are needed for inclusion in salvage regimens for treating HIV in patients failing highly active antiretroviral therapy (HAART) due to resistant virus.

SUMMARY OF THE INVENTION

Briefly described, embodiments of this disclosure include compounds, compositions, pharmaceutical compositions, and methods of treating a host with a viral infection. In an embodiment, susceptible viruses according to the present disclosure include viruses comprising at least one protein comprising a YXX<l> motif or a dileucine motif, referred to herein as "clathrin AP binding viruses," Flaviviridae, Lentiviridae,

HCV, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV, methods of treating replication of such virus in a host, methods of inhibiting the binding of a viral structural protein comprising the YXX<l> or dileucine motifs to host µ subunits of clathrin AP1-AP4 complexes, such as AP2Ml, APlMl, AP3Ml, or AP4Ml, methods of treating viral hepaitits-related liver fibrosis in a host, and the like.

One exemplary method of treating a host with a viral infection from clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV, among others, may include: administering to the host a therapeutically effective amount of an inhibiting agent to reduce the viral load in the host. In an embodiment, the inhibiting agent is selected from the group consisting of agents that competitively inhibit binding between a viral protein comprising the YXX<l> or dileucine motifs and a host protein selected from the group consisting of the µ subunits of clathrin AP1-AP4; and agents that inhibit host protein kinase activity of kinases that modulate the activity of host proteins selected from the group consisting of µ subunits of clathrin AP1-AP4. One exemplary method of treating viral infection of clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co• infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV, in a host, among others, may include: administering an inhibiting agent to the host having such viral infection or infections. In an embodiment, the inhibiting agent is selected from the group consisting of agents that competitively inhibit binding between a viral structural protein comprising the YXX<l> or dileucine motifs and a host protein selected from the group consisting of AP2Ml, and agents that inhibit host protein kinase activity of kinases that modulate the activity of host proteins selected from the group consisting of AP2M l and other µ subunits of clathrin AP complexes. In some embodiments, these inhibitors inhibit GAK (cyclin G-associated kinase) or AAK l (adaptor-associated kinase 1), which include compounds such as erlotinib, sunitinib, or PKC-412. Thus, inhibitory agents are of two classes: (1) competitive inhibitors of binding; and (2) agents that inhibit host protein kinase activity of kinases that modulate the activity of host proteins such as AP2Ml and/or other µ subunits of clathrin AP complexes.

One exemplary method of inhibiting the binding of the YXX<l> or dileucine motifs to host polypeptides, among others, includes administering an inhibiting agent to the host

having a viral infection. In an embodiment, the inhibiting agent is selected from the group consisting of agents that competitively inhibit binding between a viral structural protein comprising the YXX<l> or dileucine motifs and a host protein selected from AP2M 1 or other µ subunits of clathrin AP complexes, and agents that inhibit protein kinase activity of kinases that modulate the activity of host proteins selected from the group consisting of AP2Ml and/or other µ subunits of clathrin AP complexes.

One exemplary pharmaceutical composition for treating a host having a viral infection of clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co• infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV, among others, may include an inhibiting agent. In an embodiment, the inhibiting agent is selected from the group consisting of agents that competitively inhibit binding between a viral structural protein comprising the YXX <l> or dileucine motifs and a host protein selected from AP2Ml and/or other µ subunits of clathrin AP complexes, and agents that inhibit protein kinase activity of kinases that modulate the activity of host proteins secreted from the group consisting of AP2M l and/or other µ subunits of clathrin AP complexes.

One exemplary composition, among others, includes an inhibiting agent. In an embodiment, the inhibiting agent is selected from the group consisting of agents that competitively inhibit binding between a viral protein comprising the YXX<l> or dileucine motif and a host protein selected from the group consisting of AP2Ml and/or other µ subunits of clathrin AP complexes, and agents that inhibit protein kinase activity of kinases that modulate the activity of host proteins selected from the group consisting of AP2Ml and/or other µ subunits of clathrin AP complexes.

In one aspect, the present invention provides a method of treating HCV-HIV co-infection through administration of an effective amount of an agent that inhibits binding of HCV and HIV to host proteins selected from the group consisting of AP2M 1 and other µ subunits of clathrin AP complexes. In one embodiment, the inhibiting agent inhibits AAK l or GAK. In another embodiment, the agent is selected from the group consisting of erlotinib, sunitinib, and PKC-412. In another embodiment, the agent prevents binding of a Lentiviridae or Flaviviridae structural protein containing a YXX<l> or dileucine motif to a host protein selected from the group consisting of AP2M l and/or other µ subunits of clathrin AP complexes. In still another embodiment, the agent inhibits AAK.

In another aspect, the invention provides a method of inhibiting infections by at least a first virus other than HCV, another member of Flaviviridae, or a member of Lentiviridae, or combinations thereof, through administration to a patient in need an effective amount of an agent that inhibits binding of a structural protein from a virus containing a YXX <l> or dileucine motif to a host protein selected from the group consisting of AP2Ml and/or other µ subunits of clathrin AP complexes. In one embodiment, the agent inhibits AAK l or GAK. In another embodiment, the agent is selected from the group consisting of erlotinib, sunitinib, and PKC-412.

In another aspect, the present invention provides a method of screenmg for antiviral agents through contacting a susceptible virus as described herein or viral protein with a candidate agent and a protein selected from the group consisting of AP2Ml and/or other µ subunits of clathrin AP complexes, and determining an effect of the candidate agent on the binding of virus to the host protein.

In another aspect, the present invention provides a method of screenmg for antiviral agents through contacting a susceptible virus as described herein with a candidate agent and a cell expressing a protein selected from the group consisting of AP2Ml and other µ subunits of clathrin AP complexes, and determining the effect of the candidate agent on viral assembly or budding. In one embodiment, the candidate agent inhibits the activity of AAK l or GAK.

In another aspect, the present invention provides a method of inhibiting assembly or budding of clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co• infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV, in a host cell through contacting the host cell with an agent that prevents binding of a viral structural protein containing a YXX <l> or dileucine motif to a host protein selected from the group consisting of AP2M l and other µ subunits of clathrin AP complexes. In one embodiment, the agent blocks binding of the viral structural protein to the host protein. In another embodiment, the agent competes with the viral structural protein for binding to the host protein. In another embodiment, the agent competes with the host protein for binding to the viral structural protein. In still another embodiment, the method involves screening of AAK l and GAK inhibitors for non-Flaviviridae viruses.

BRIEF DESCRIPTION OF THE DRAWINGS

The following drawings form part of the present specification and are included to further demonstrate certain aspects of the invention. The invention may be better understood by reference to one or more of these drawings in combination with the detailed description of specific embodiments presented herein:

FIG. 1: Shows a viral core with a YXX<l> motif that binds AP2M l. (A) Schematics of core. The location of the identified motif is indicated. (B)-(C) Consensus sequences of YXX<l> motifs from representative human (B) and viral (C) proteins. (D) The consensus sequence of all HCV isolates, clones used in this study, and engineered core mutants. (E) Fluorescent images from a microfluidic chip and schematics. Left: AP2Ml-V5-his was anchored to the device surface via its interaction with anti-His antibodies and labeled with anti-V5-FITC antibodies. Middle: T7-tagged core or NS3 were incubated with surface bound AP2M 1 and labeled with anti-T7-Cy3 antibodies. Interactions were trapped mechanically by MITOMI. Cy3 signal representing bound viral protein is shown following a wash. Right: an overlay of the Cy3 and FITC signals, representing bound viral prey to human bait ratio. F. *In vitro* binding curves of core-T7 and NS3-T7 to surface bound AP2Ml. Y axis represents bound viral protein to surface bound AP2M l ratio.

FIG. 2: Shows a viral core binding AP2Ml in cells and in the context of HCV infection. (A) Schematics of the PCAs format. A and B represent prey and bait proteins, and GLucl/2 are fragments of *Gaussia* luciferase. (B) Cells were cotransfected with combinations of plasmids indicated below the graph with those indicated in the legend. Y axis represents luminescence relative to the core-AP2M l signal. The banded bar on the right represents AP2M l binding to the host cargo protein TFR. (C) Core-AP2Ml binding in the presence of free AP2Ml, core or NESI. Y axis represents luminescence ratio (the average luminescence signal detected in cells transfected with Glucl -AP2M 1 and Gluc2-core divided by the average of luminescence measured in control wells transfected with Glucl-AP2M l and an empty Gluc2 vector with those transfected with Gluc2-core and an empty Glucl vector relative to core-AP2Ml binding in the presence of empty PUC19. (D) Immunoprecipitations (IPs) in membranes of HCV infected cells. Left panels: Anti• AP2Ml antibodies or IgG were used for IP. Membranes were immunoblotted with anti• phospho-AP2Ml, anti-AP2Ml, anti-core, and anti-actin antibodies. Cal-A represents calyculin-A. Right panels: Anti-core antibodies or IgG were used for IP. Membranes were

immunoblotted with anti-core, anti-AP2M l, and anti-actin antibodies. (E) Representative confocal IF microscopy images of AP2Ml and core in Huh-7.5 cells 3 days postelectroporation with J6/JFH HCV RNA. Data represent means±s.d. (error bars) from three independent experiments in triplicates (n>20 in E). * p <0.05, ** p <0.01, *** p<0.001.

FIG. 3: Shows a core YXX<l> motif mediating AP2M l binding and viral assembly, functionally interchangeable with other YXX<l> signals. (A) Binding of wild type (WT) and core mutants to AP2M l by PCAs. Y axis represents luminescence ratio (the average luminescence signal detected in cells transfected with Glucl-AP2Ml and the various forms of Gluc2-core divided by the average of luminescence measured in control wells transfected with Glucl-AP2Ml and an empty Gluc2 vector with those transfected with the respective Gluc2-core and an empty Glucl vector) relative to WT core-AP2M l binding. (B) HCV RNA replication by *Renilla* luciferase assays at 6 hr (white) and 72 hr (black) postelectroporation with J6/JFH(p7-Rluc2A) harboring the corresponding mutations. El-E2 is an assembly defective control. GNN is a replication defective polymerase mutant. (C) Extracellular infectivity by luciferase assays in naive Huh-7.5 cells infected with supernatants derived from the electroporated cells. (D) Intracellular infectivity by luciferase assays in naive Huh-7.5 cells infected with clarified cell lysates derived from the electroporated cells. (E) Intra- and extracellular infectivity titers measured by limiting dilution assays. TCIDso is a 50% tissue culture infectious dose. (F) Viral RNA release into the culture supernatant at 72 hr postelectroporation measured by qRT-PCR. (G) HCV core protein release into the culture supernatant at 72 hr postelectroporation determined by ELISA relative to WT control. (H) Levels of core protein by western analysis in lysates prepared from cells infected with virus harboring the corresponding mutations. Means and s.d. (error bars) of results from at least three independent experiments in triplicates are shown. The dashed horizontal lines represent background levels of luciferase activity. RLU is relative light units. * p <0.05, ** p <0.01, *** p<0.001.

FIG. 4: Shows inhibition of HCV assembly by AP2M l depletion. (A) AP2Ml protein levels by quantitative western analysis in stable clones harboring shRNA lentiviral constructs targeting the AP2M l gene and a non-targeting (NT) sequence. A representative membrane and combined data from three independent measurements are shown. Y axis represents AP2Ml to actin protein ratio relative to NT control. (B)

AP2Ml/S18 RNA ratio by qRT-PCR in selected stable clones relative to NT control. (C) The indicated clones were electroporated with J6/JFH(p7-Rluc2A). HCV RNA replication in these clones by luciferase assays at 9 hr (white) and 72 hr (black) postelectroporation. (D) Extracellular infectivity measured by luciferase assays in naive cells inoculated with supernatants derived from the various stable cell clones. (E) Intracellular infectivity by luciferase assays in naive Huh-7 .5 cells infected with clarified cell lysates derived from the electroporated cells. (F) Intra- and extracellular infectivity titers measured by limiting dilution assays. TCIDso is a 50% tissue culture infectious dose. (G) Viral RNA release into the culture supernatant at 72 hr postelectroporation measured by qRT-PCR. (H) HCV core protein release into the culture supernatant at 72 hr postelectroporation, as determined by ELISA. (I) Infectious virus production relative to NT control (top panel) and levels of AP2M l protein by a western blot analysis (bottom panel) in cells concurrently transduced with lentiviruses expressing shAP2Ml and shRNA resistant WT AP2Ml cDNA (AP2M l-WT). Means and s.d. (error bars) of results from at least three independent experiments are shown. RLU is relative light units. * p <0.05, ** p <0.01, *** p<0.001.

FIG. 5: Shows that disruption of core-AP2Ml binding abolishes recruitment of AP2Ml to LD and alters the sub-cellular localization of core and its colocalization with E2. Quantitative confocal immunofluorescence (IF) analysis of the sub-cellular localization of core and AP2M l and core-E2 colocalization in Huh-7.5 cells. (A) A representative merged image of endogenous AP2Ml (blue) and the LD marker, Bodipy (green), in naive Huh-7.5 cells. (B)-(D) Merged images of Huh-7.5 cells infected with J6/JFH HCV stained for core (red), the LD marker, Bodipy (green), and AP2M l (blue). (E) Percent colocalization of the indicated signals in naive (white) or infected (black) cells by a quantitative colocalization analysis of (A)-(D). (F) A four channel merged image. The yellow arrows in the inset indicate colocalization of core and AP2M l to LD. (G)-(J) Representative merged images and quantitative colocalization analysis of AP2M l (red) and the lipid marker, LipidTOX (blue), in Huh-7.5 cells transfected with plasmids expressing AP2M l-mCherry alone (G) or AP2Ml-mCherry with WT core (H) or core Y136A mutant (I). Core expression (green) in the cells shown in panels (H) and (I) is demonstrated in the respective bottom panels. (K)-(P) Representative merged images and quantitative colocalization analysis of core (red) and (K) Bodipy (green), demonstrating increased localization of core to LD in Huh-7.5 cells electroporated with J6/JFH HCV

RNA harboring the Y136A core mutation (right panel) compared with WT core (left panel). (L) Bodipy (green) in control (NT) cells (left panel) or AP2Ml depleted (right panel) Huh-7.5 electroporated with J6/JFH HCV RNA , showing a dramatic localization of core to LD in AP2M1 depleted cells. (M). TGN46 (green), demonstrating decreased localization of core to TGN in Huh-7.5 cells electroporated with J6/JFH RNA harboring the Y136A core mutation (right panel) compared with WT core (left panel). (N) TGN46 (green) in control (NT) cells (left panel) or AP2M 1 depleted (right panel) Huh-7.5 electroporated with J6/JFH HCV RNA, showing decreased localization of core to TGN in AP2Ml depleted cells. (0) E2 (green), demonstrating decreased colocalization of core and E2 in Huh-7.5 cells electroporated with J6/JFH RNA harboring the Y136A core mutation (right panel) compared with WT core (left panel). (P) E2 (green) in control (NT) cells (left panel) or AP2Ml depleted (right panel) Huh-7.5 electroporated with J6/JFH HCV RNA, showing decreased colocalization of core and E2 in AP2Ml depleted cells. Representative images at x60 magnification are shown. Graphs represent quantitative colocalization analysis of Z stacks using Manders' coefficients. Values indicate mean M2 values represented as percent colocalization (the fraction of green intensity that coincides with red intensity or in the case of FIGS. (G)-(1), the fraction of blue intensity that coincides with red intensity)± s.d. (error bars); n = 10-15. * p <0.05, ** p <0.01, *** p<0.001.

FIG. 6: Shows that AAK l and GAK regulate core-AP2Ml binding and are involved in HCV assembly. (A) Regulatory mechanisms of AP2Ml binding to host cargo proteins harboring YXX<l> signals. (B) Binding of core to wild type and T156A AP2M l mutant by PCAs (black) and microfluidics (white). (C)-(E) Huh-7.5 were transfected with plasmids encoding WT or T156A AP2Ml mutant and electroporated with J6/JFH(p7-Rluc2A) 48 hr posttransfection. (C) Cellular viability by alamarBlue-based assays at 48 hr posttransfection relative to WT AP2Ml control. (D) HCV RNA replication in cells overexpressing WT or T156A AP2Ml mutant by luciferase assays at 6 hr (black) and 72 hr (white) postelectroporation with J6/JFH(p7-Rluc2A). (E) Extracellular (black) and intracellular (white) infectivity by luciferase assays in naive Huh-7.5 cells infected with supernatants or cell lysates derived from the indicated cells, respectively, relative to WT control. (F)-(G) Huh7.5 cells were transfected with the corresponding siRNAs. (F) Ratio of AAK l (left) or GAK (right) to S18 RNA in these cells relative to NT sequences by qRT-PCR. (G) Quantitative western analysis. Numbers represent AAK1 (top) or GAK

(bottom) to actin protein ratios relative to NT control. (H) Core-AP2M l binding by PCAs in Huh-7.5 cells depleted for AAK l or GAK by siRNAs. Y axis represents luminescence ratio (the average luminescence signal detected in cells transfected with Glucl -AP2M 1 and Gluc2-core divided by the average of luminescence measured in NT cells transfected with Glucl-AP2Ml and an empty Gluc2 vector with those transfected with Gluc2-core and an empty Glucl vector) relative to NT control. (1)-(K) AAK l or GAK depleted cells were electroporated with J6/JFH(p7-Rluc2A). (I) Cellular viability by alamarBlue-based assays in depleted cells relative to NT control. (J) HCV RNA replication in these cells by luciferase assays at 6 hr (black) and 72 hr (white) postelectroporation. (K) Extracellular (black) and intracellular (white) infectivity by luciferase assays in naive Huh-7.5 cells infected with supematants or cell lysates derived from the indicated cells, respectively, relative to NT control. (L) Core binding to AAK l and GAK by PCAs. Y axis represents luminescence ratio relative to core-AP2Ml binding. Data represent means and s.d. (error bars) from at least two experiments in triplicates. RLU is relative light units. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

FIG. 7: Pharmacological inhibition of core-AP2Ml binding and HCV assembly. (A) AP2M l regulators and the discovered inhibitors. (B) The inhibitors' Kds of binding to AAK l or GAK (Karaman *et al., Nat Biotechnol* 26:127-132, 2008). IC50s for these compounds effect on core-AP2Ml binding, and EC50s for their effect on extracellular infectivity, intracellular infectivity, and viral infection with cell culture grown HCV (HCVcc). (C) Inhibition of core-AP2Ml binding by the compounds measured by PCAs. (D)-(G) Huh-7.5 cells electroporated with J6/JFH(p7-Rluc2A) were treated daily with either erlotinib, sunitinib or PKC-412 for 3 days. Supematants and cell lysates were harvested at 72 hr and used to inoculate naive Huh-7.5 cells. Dose response curves of the inhibitors' effects on extracellular (D) and intracellular (E) infectivity relative to untreated controls. These compounds had no effect on HCV RNA replication (F) or cellular viability (G) by luciferase and AlamarBlue-based assays, respectively (GNN is a replication-defective polymerase mutant). (H) The effect of the inhibitors on AP2Ml phosphorylation by western analysis of cell lysates harvested following electroporation with J6/JFH(p7-Rluc2A) and treatment with the compounds in the presence of Calyculin A (Cal-A). Representative membranes blotted with anti-phopho• AP2Ml (p-AP2M l) and anti-actin antibodies, and quantitative analysis from 3 experiments are shown. Y axis represents pAP2Ml/actin protein ratio relative to

untreated controls. (I) The inhibitors' effect on viral infection (black) and cellular viability (grey) in cells infected with HCVcc following 72 hr of daily treatment relative to untreated controls. Data represent means and s.d. (error bars) from at least three experiments in triplicates. RLU is relative light units. * p <0.05, ** p <0.01, *** p<0.001.

FIG. 8: Transient depletion of AP2M l by pooled siRNAs inhibits HCV assembly. (A) AP2M l/S18 RNA ratio measured by qRT-PCR in Huh-7.5 cells transfected with a pool of four siRNAs (ON-TARGETplus SMARTpools, Dharmacon) targeting AP2M l or a pool of non-targeting (NT) sequences at 48 hr posttransfection relative to NT contols. (B) AP2Ml protein levels by quantitative western analysis in cells 48 hr posttransfection with the corresponding pooled siRNAs. Numbers represent AP2Ml to actin protein ratio relative to the NT control. (C) Cellular viability by alamarBlue assays 48 hr post siRNAs transfections relative to NT control. (D) Cells were electroporated with J6/JFH(p7-Rluc2A) at 48 hr following transfection with the indicated pooled siRNAs. HCV RNA replication in these cells by luciferase assays at 6 hr (black) and 72 hr (white) postelectroporation. (E) Extracellular (black) and intracellular (white) infectivity measured in naive Huh-7.5 cells infected with supematants or clarified cell lysates derived from electroporated cells harboring the indicated siRNAs by luciferase assays, respectively. (F) Infectious virus production measured by limiting dilution assays. (G) Extracellular (black) and intracellular (white) infectivity measured by focus formation assays in naive Huh-7 .5 cells infected with supematants or clarified cell lysates derived from Huh-7.5 cells transiently depleted for AP2M l by siRNAs and infected with culture grown J6/JFH virus (titer: $1.2x10^5$ $TCID_{50}$/ml). Results are relative to NT controls. Means ± s.d. (error bars) of results from at least two independent experiments are shown. RLU is relative light units. TCIDso is 50% tissue culture infectious dose.

FIG. 9: Shows that AAK l and GAK depletion or pharmacological inhibition abrogates HIV-1 replication in TZM-b l cells (CXCR4-positive HeLa cells that express CD4 and CCR5 and also contain integrated reporter genes for luciferase and *E. coli* • galactosidase, both under the control of an HIV long-terminal repeat sequence (tat gene)). (A) HIV replication in AAKl or GAK depleted cells relative to NT control. (B) The antiviral effect of AAK l and GAK inhibitors on HIV replication relative to untreated control.

FIG. 10: Shows Microfluidic-based protein-protein binding assay. Three individual unit cells (out of hundreds in a microfluidic device) are shown in this scheme.

(A) Compartments and micromechanical valves. A valve is created where a control channel crosses a flow channel. The resulting thin membrane in the junction between the two channels can be deflected by hydraulic actuation. Using multiplexed valve systems allows a large number of elastomeric microvalves to perform complex fluidic manipulations within these devices.

(B) Experimental protocol. ® represents surface bound biotinylated anti-histidine antibodies (shown in B l and B2). @ represents surface bound FITC-labeled bait human protein (shown in B3). ® represents Cy3-labeled prey viral protein (shown in B4 and B5). 1) The microfluidic device was bonded to a glass slide and subjected to surface patterning that resulted in a circular area coated with biotinylated anti-histidine antibodies within each unit cell (see c). 2) VS-his-tagged bait human proteins were expressed off the chip using *in vitro* transcription/ translation (TNT) mixture and were loaded into the device. 3) These proteins bound to the surface anti-his antibodies. 4) T7-tagged viral proteins were expressed off the chip by the same mammalian *in vitro* TNT mixture in the presence of microsomal membranes and loaded into the device along with FITC-labeled anti-V5 and Cy3-labeled anti-T7 antibodies. 5) The "sandwich valves" were closed to allow incubation of the viral protein with the human proteins and their labeling with the respective fluorescent antibodies. 6) MITOMI was then performed by actuation of the "button membrane" facilitating trapping surface-bound complexes while expelling any solution phase molecules. The "sandwich valves" were opened followed by a brief wash to remove untrapped unbound material. 7) The device was scanned by an array scanner. Trapped viral protein and surface bound human proteins were detected. The ratio of bound viral protein to expressed human protein was calculated for each data point by measuring the median signal of Cy3 to median signal of FITC (represented by 0, shown in B7).

(C) Surface patterning. 1) Accessible surface area was derivatized by flowing a solution of biotinylated BSA **(i!l)** through all flow channels. 2) A Neutravidin solution **(.)** was loaded. 3) The "button" membrane was activated. 4) All remaining accessible surface area except for a circular area of 60 µm masked by the button was passivated with biotinylated solution **(l!!).** 5) The "button" membrane was opened. 6) A solution of biotinylated-anti-his antibodies (was loaded allowing specific functionalization of the previously masked circular area 7) *In vitro* expressed human protein ( ) bound to the

biotinylated-antibodies coating the discrete circular area. Each of the described steps was followed by a PBS wash.

FIG. 11: Shows that the core's YXX<l> mutations do not affect HCV RNA replication by qRT-PCR assays. HCV RNA replication, by qRT-PCR in Huh-7.5 cells 72 hr following electroporation with J6/JFH(p7-Rluc2A) harboring the corresponding core's YXX<l> mutations relative to WT control. El-E2 is an assembly defective control. GNN is a replication defective polymerase mutant. Means and s.d. (error bars) of results from two independent experiments in triplicates are shown.

FIG. 12: Shows that AP2M l depletion has no effect on HCV RNA replication by qRT-PCR assays. HCV RNA replication, by qRT-PCR in Huh-7.5 cells harboring the corresponding shRNAs 72 hr following electroporation with J6/JFH(p7-Rluc2A) relative to NT control. Means and s.d. (error bars) of results from two independent experiments in triplicates are shown.

FIG. 13: Shows transient depletion of AP2Ml by pooled siRNAs inhibits HCV assembly. (A) AP2M l/Sl8 RNA ratio measured by qRT-PCR in Huh-7.5 cells transfected with a pool of four siRNAs (ON-TARGETplus SMARTpools, Dharmacon) targeting AP2M l or a pool of non-targeting (NT) sequences at 48 hr posttransfection relative to NT contols. (B) AP2M l protein levels by quantitative Western analysis in cells 48 hr posttransfection with the corresponding pooled siRNAs. Numbers represent AP2Ml to actin protein ratio relative to the NT control. (C) Cellular viability by alamarBlue assays 48 hr post siRNAs transfections relative to NT control. (D) Cells were electroporated with J6/JFH(p7-Rluc2A) at 48 hr following transfection with the indicated pooled siRNAs. HCV RNA replication in these cells by luciferase assays at 6hr (black) and 72hr (white) postelectroporation. (E) Extracellular (black) and intracellular (white) infectivity measured in naive Huh-7 .5 cells infected with supernatants or clarified cell lysates derived from electroporated cells harboring the indicated siRNAs by luciferase assays, respectively. (F) Infectious virus production measured by limiting dilution assays. (G) Extracellular (black) and intracellular (white) infectivity measured by focus formation assays in naive Huh-7 .5 cells infected with supernatants or clarified cell lysates derived from Huh-7.5 cells transiently depleted for AP2M l by siRNAs and infected with culture grown J6/JFH virus (titer: $1.2xl0^5$ $TCID_{50}$/ml). Results are relative to NT controls. Means ± s.d. (error bars) of results from at least two independent experiments are shown. RLU is relative light units. $TCID_{50}$ is 50% tissue culture infectious dose.

FIG. 14: Shows that a Y136A core mutation does not seem to affect core localization to ER membranes in HCV infected cells. (A) quantitative confocal immunofluorescence (IF) analysis for localization of core to the ER membrane in Huh-7.5 cells infected with culture grown HCV. (A) and (B) are representative merged images of core (red) and the ER marker, calreticulin (green), demonstrating a comparable partial localization of core to the ER membrane in Huh-7.5 cells infected with WT virus (A) or with virus harboring the Y136A core mutation (B). Representative images at x60 magnification are shown. (C) Colocalization analysis of Z stacks using Manders' coefficients (with a higher value representing more colocalization). Values indicate mean M2 values represented as percent colocalization (the fraction of green intensity that coincides with red intensity ± s.d. (error bars); n = 10-15.

DETAILED DESCRIPTION

Before the present disclosure is described in greater detail, it is to be understood that this disclosure is not limited to particular embodiments described, and the embodiment of the invention as such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the present disclosure will be limited only by the appended claims.

As will be apparent to those of skill in the art upon reading this disclosure, each of the individual embodiments described and illustrated herein has discrete components and features which may be readily separated from or combined with the features of any of the other several embodiments without departing from the scope or spirit of the present disclosure. Any recited method can be carried out in the order of events recited or in any other order that is logically possible.

Before the embodiments of the present disclosure are described in detail, it is to be understood that, unless otherwise indicated, the present disclosure is not limited to particular materials, reagents, reaction materials, manufacturing processes, or the like, as such can vary. It is also to be understood that the terminology used herein is for purposes of describing particular embodiments only, and is not intended to be limiting. It is also possible in the present disclosure that steps can be executed in different sequence where this is logically possible. As used in the specification and the appended claims, the singular forms "a," "an," and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a compound" includes a plurality of

compounds. In this specification and in the claims that follow, reference will be made to a number of terms that shall be defined to have the following meanings unless a contrary intention is apparent.

Definitions

In describing and claiming the disclosed subject matter, the following terminology will be used in accordance with the definitions set forth below.

By "Flaviviridae virus" is meant any virus of the Flaviviridae family, including those viruses that infect humans and non-human animals. The polynucleotide and polypeptides sequences encoding these viruses are well known in the art, and may be found at NCBI's GenBank database, e.g., as Genbank Accession numbers NC_004102, AB031663, D11355, D11168, AJ238800, NC_001809, NC_001437, NC_004355, NC_004119, NC_003996, NC_003690, NC_003687, NC_003675, NC_003676, NC_003218, NC_001563, NC_000943, NC_003679, NC_003678, NC_003677, NC_002657, NC_002032, and NC_001461, the contents of which database entries are incorporated by references herein in their entirety.

As used herein, the terms "treatment," "treating," and "treat" are defined as acting upon a disease, disorder, or condition with an agent to reduce or ameliorate the pharmacologic and/or physiologic effects of the disease, disorder, or condition and/or its symptoms. "Treatment," as used herein, covers any treatment of a disease in a host (e.g., a mammal, typically a human or non-human animal of veterinary interest), and includes: (a) reducing the risk of occurrence of the disease in a subject determined to be predisposed to the disease but not yet diagnosed as infected with the disease, (b) impeding the development of the disease, and (c) relieving the disease, i.e., causing regression of the disease and/or relieving one or more disease symptoms. "Treatment" is also meant to encompass delivery of an inhibiting agent to provide a pharmacologic effect, even in the absence of a disease or condition. For example, "treatment" encompasses delivery of a disease or pathogen inhibiting agent that provides for enhanced or desirable effects in the subject (e.g., reduction of pathogen load, reduction of disease symptoms, etc.).

As used herein, the terms "prophylactically treat" or "prophylactically treating" refer to completely or partially preventing a disease or symptom thereof and/or may be therapeutic in terms of a partial or complete cure for a disease and/or adverse effect attributable to the disease.

As used herein, the term "host," "subject," "patient," or "organism" includes

humans and mammals (e.g., mice, rats, pigs, cats, dogs, and horses). Typical hosts to which compounds of the present disclosure may be administered will be mammals, particularly primates, especially humans. For veterinary applications, a wide variety of subjects will be suitable, e.g., livestock such as cattle, sheep, goats, cows, swine, and the like; poultry such as chickens, ducks, geese, turkeys, and the like; and domesticated animals, particularly pets such as dogs and cats. For diagnostic or research applications, a wide variety of mammals may be suitable subjects, including rodents (e.g., mice, rats, hamsters), rabbits, primates, and swine, such as inbred pigs and the like. The term "living host" refers to a host as noted above or another organism that is alive. The term "living host" refers to the entire host or organism and not just a part excised (e.g., a liver or other organ) from the living host.

The term "isolated compound" means a compound which has been substantially separated from, or enriched relative to, other compounds with which it occurs in nature. Isolated compounds are usually at least about 80%, at least 90% pure, at least 98% pure, or at least about 99% pure, by weight. The present disclosure is meant to include diastereomers, as well as their racemic and resolved, enantiomerically pure forms and pharmaceutically acceptable salts thereof.

The term "unit dosage form," as used herein, refers to physically discrete units suitable as unitary dosages for human and/or animal subjects, each unit containing a predetermined quantity of a compound (e.g., an antiviral compound, as described herein) calculated in an amount sufficient to produce the desired effect in association with a pharmaceutically acceptable diluent, carrier, or vehicle. The specifications for unit dosage forms depend on the particular compound employed, the route and frequency of administration, the effect to be achieved, and the pharmacodynamics associated with each compound in the host.

A "pharmaceutically acceptable excipient," "pharmaceutically acceptable diluent," "pharmaceutically acceptable carrier," or "pharmaceutically acceptable adjuvant" means an excipient, diluent, carrier, and/or adjuvant that is useful in preparing a pharmaceutical composition that is generally safe, non-toxic, and neither biologically nor otherwise undesirable, and includes an excipient, diluent, carrier, and adjuvant that are acceptable for veterinary use and/or human pharmaceutical use. "A pharmaceutically acceptable excipient, diluent, carrier and/or adjuvant" as used in the specification and claims includes one and more such excipients, diluents, carriers, and adjuvants.

As used herein, a "pharmaceutical composition" 1s meant to encompass a composition suitable for administration to a subject, such as a mammal, especially a human. In general a "pharmaceutical composition" is sterile, and preferably free of contaminants that are capable of eliciting an undesirable response within the subject (e.g., the compound(s) in the pharmaceutical composition is pharmaceutical grade). Pharmaceutical compositions can be designed for administration to subjects or patients in need thereof via a number of different routes of administration including oral, intravenous, buccal, rectal, parenteral, intraperitoneal, intradermal, intracheal, intramuscular, subcutaneous, inhalational, and the like.

The term "therapeutically effective amount" as used herein refers to that amount of an embodiment of the agent (which may be referred to as a compound, an inhibitory agent, and/or a drug) being administered that will relieve to some extent one or more of the symptoms of the disease, i.e., infection, being treated, and/or that amount that will prevent, to some extent, one or more of the symptoms of the disease, i.e., infection, that the host being treated has or is at risk of developing.

Viruses are small intracellular infectious agents that replicate via use of the host cell machinery. Virus assembly and budding of Flaviviridae remain poorly understood, although recent data suggest that the endocytic pathway may be involved, such as with Hepatitis C Virus (HCV). Host proteins are involved in these processes, although prior to the present disclosure, host proteins involved in mediating late assembly and budding were not targeted as antiviral agents. The present disclosure, however, provides antiviral methods and compositions that target host proteins for late assembly and budding. In particular, host clathrin adaptor proteins and their regulators involved in the endocytic or secretory pathways are effective targets. The role of clathrin adaptor proteins and their regulators in infectious Flaviviridae production was previously unknown. The present disclosure demonstrates that host clathrin adaptor proteins, such as AP2M l, APlM l, AP3Ml, and AP4Ml, and their regulatoryo proteins, such as, but not limited to AAK l, and GAK are suitable targets for antiviral therapies. In addition, the present disclosure provides a novel viral amino acid motif that mediates interaction with a host protein.

A conserved YXX<l> motif was discovered within the core protein of HCV, a 191-amino acid membrane protein that forms the viral capsid (FIG. 1). This YXX<l> motif is conserved across all HCV isolates available in databases to date. This motif was also discovered in structural proteins from other Flaviviridae and other viruses. This YXX<l> motif conforms with the YXX<l> consensus sorting signal within membrane host cargo

proteins, recognized by the μ subunit of clathrin adaptor protein (AP) complexes (FIG. la). AP2M l mediates clathrin-dependent endocytosis, the process by which cargo proteins are sorted into clathrin-coated pits destined for fusion with early endosomes. Recognition of the YXX<l> or dileucine motifs within retroviral proteins by AP2Ml or APlMl has been shown to be involved in mediating assembly and budding of retroviruses (FIG. lb) (Batonick *et al., Virology* 342:190-200, 2005; Camus *et al., Malec Biol Cell* 18:3193-3203, 2007; Berlioz-Torrent *et al., J Virol* 73:1350-1361, 1999; Byland *et al., Malec Biol Cell* 18:414-425, 2007; Wyss *et al., J Virol* 75:2982-2992, 2001; Ohno *et al., Virology* 238:305-315, 1997; Boge *et al., J Biol Chem* 273:15773-15778, 1998; Egan *et al., J Viral* 70:6547-6556, 1996; Rowell *et al., J Immunol* 155:473-488, 1995; Lodge *et al., EMBO J* 16:695-705, 1997; Deschambeault *et al., J Viral* 73:5010-5017, 1999). However, the role of AAK l and GAK in HIV infection has not been studied, and these mechanisms have not been targeted pharmacologically. Accordingly, the present disclosure provides methods and compositions for treating or preventing viral infection from HCV, Flaviviridae, Flaviviridae other than HCV, HIV, Lentiviridae other than HIV, clathrin AP binding viruses, clathrin AP binding other than Flaviviridae, clathrin AP binding viruses other than HCV, or co-infections such as HCV/HIV co• infections.

Inhibition of the interaction between the viral YXX<l> motif and host adaptor proteinis, such as, but not limited to AP2M l, results in reduced infectious virus production of HCV, Flaviviridae, Flaviviridae other than HCV, clathrin AP binding viruses, clathrin AP binding viruses other than Flaviviridae, clathrin AP binding viruses other than HCV, or co-infections such as HCV/HIV co-infections. In addition, inhibition of the activity of proteins that regulate clathrin adaptor proteins may result in reduced infectious virus production of HCV, Flaviviridae, Flaviviridae other than HCV, clathrin AP binding viruses, clathrin AP binding viruses other than Flaviviridae, clathrin AP binding viruses other than HCV, or co-infections such as HCV/HIV co-infections. Thus, approaches designed to disrupt the viral YXX<l> motif s interaction with clathrin adaptor proteins, such as AP2M l and the like, may be useful for inhibiting infectious virus production.

Regulation of the activity of many proteins occurs through the action of protein kinases and protein phophatases. GAK and AAK l are two protein kinases that modulate the activity of AP2M l and other μ subunits of clathrin AP complexes, indicating that administration of protein kinase inhibitors such as erlotinib, sunitinib, or PKC-412 may

abolish the interaction of AP2M 1 and other μ subunits of clathrin AP complexes with viral YXX<l> or dileucine motifs, thereby inhibiting viral infection from HCV, Flaviviridae, Flaviviridae other than HCV, clathrin AP binding viruses, clathrin AP binding viruses other than Flaviviridae, clathrin AP binding viruses other than HCV, HIV, Lentiviridae other than HIV, or co-infections such as HCV/HIV co-infections, as described in more detail below.

Embodiments of the present disclosure therefore provide methods of treating a viral infection from clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV, or any other enveloped virus that hijacks AP2Ml other μ subunits of clathrin AP complexes/AAK l/GAK, compositions (including inhibiting agents and combinations of such agents with other antiviral therapies) for treating an infection by such viruses, and the like. In particular, embodiments of the present invention provide for methods of treating infections caused by clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV, and compositions for treating these infections or any other enveloped virus that binds clathrin adaptor proteins, such as AP2M l, or is regulated by AAK l or GAK.

Embodiments of the present disclosure provide methods of prophylactically treating a viral infection from clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV, compositions (including inhibiting agents and combinations of such agents with other antiviral therapies) for prophylactically treating an infection by such viruses, and the like. In particular, embodiments of the present invention provide for methods of prophylactically treating HCV and/or HIV, and compositions for treating HCV and/or HIV. In an alternative embodiment, the present invention provides for methods for treating HCV/HIV infection or Flaviviridae other than HCV. While the discussion herein may describe the invention with respect to HCV, it should be noted that the disclosure relates to not only HCV, but also to clathrin AP binding viruses, Flaviviridae, Lentiviridae, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV, and compositions for treating these infections or any other enveloped

virus that binds AP2M lor other µ subunits of clathrin AP complexes or is regulated by AAK l or GAK, or any other enveloped virus that binds clathrin adaptor proteins, such as AP2Ml and the like, or other µ subunits of clathrin AP complexes and this interaction is regulated by AAK l/GAK.

In view of the recognition that viral proteins comprising a YXX<l> or dileucine motif can mediate interaction with host proteins, embodiments of the present invention provide compositions (including pharmaceutical compositions) including an inhibiting agent that can be used to treat a host infected by a virus of the Flaviviridae family of viruses. In particular, the inhibiting agent can be used to treat hosts infected with viral infection from clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV, or patients suffering from coinfections, in particular HCV and HIV or Flaviviridae other than HCV. Accordingly, two types of inhibiting agents find use herein: (1) those that inhibit interaction between a viral protein comprising the YXX<l> or dileucine motif and host clathrin adaptor proteins, such as AP2M 1 andother µ subunits of clathrin AP complexes; and (2) those that inhibit kinase activity that regulates host proteins. In some embodiments, the inhibiting agent inhibits AAK l or GAK.

As described in the Examples, it has been observed that the HCV YXX<l> motif in a structural protein binds to the host protein AP2M 1. This interaction has been used to identify inhibiting agents that interfere with the interaction and thus may be candidates for clinical development as drugs for treating viral infection from clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co-infections, such as HCV/HIV co• infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV.

Inhibiting agents described herein are useful in the treatment of viral infections, where the virus is a YXX<l> motif-containing virus, and Y is a tyrosine residue, X is any amino acid, and <l> is a bulky hydrophobic residue. These can include, but are not limited to, phenylalanine, methionine, leucine, isoleucine, and valine. Inhibiting agents described herein are useful in the treatment of viral infections, where the virus comprises a dileucine motif-containing virus. Dileucine motifs may include, but are not limited to, leucine-leucine, isoleucine-leucine, leucine-isoleucine, [D/E]XXXL[L/1], or DXXLL (where X is any amino acid). YXX<l> or dileucine motif-containing viruses include, among others, Lentiviridae, such as HIV, and Flaviviridae family viruses. Exemplary

Flaviviridae include, but are not limited to, flaviviruses, pestiviruses, and hepatitis C viruses. Other YXX<l> or dileucine motif-containing viruses include yellow fever virus (YFV); Dengue virus, including Dengue types 1-4; Japanese Encephalitis virus; Murray Valley Encephalitis virus; St. Louis Encephalitis virus; West Nile virus; tick-borne encephalitis virus; Hepatitis C virus (HCV); Kunjin virus; Central European encephalitis virus; Russian spring-summer encephalitis virus; Powassan virus; Kyasanur Forest disease virus; Ilheus virus; Apoi virus; GB virus A and B; Louping ill virus and Omsk hemorrhagic fever virus.

In an embodiment, inhibiting agents (e.g., anti-HCV agents) for use in inhibiting HCV replication and treating HCV infection, are of particular interest. Flaviviridae other than HCV and enveloped viruses other than Flaviviridae are also of interest. The HCV contemplated by the disclosure may be of any genotype (genotype 1, 2, 3, 4, 5, 6, and the like), as well as subtypes of an HCV genotype (e.g., la, lb, 2a, 2b, 3a, etc.). Because HCV genotype 1 is typically the most difficult to treat, the methods and compositions of the invention for treating infections by HCV genotype 1 and genotype 1 subtypes are of particular interest. In an embodiment, HCV co-infections are of interest. In particular, HCV/HIV co-infections are of interest.

While the specification below refers to HCV, such a reference is only for clarity and is not intended to limit the disclosure as described in more detail below to HCV. As noted above, the methods and compositions of the invention can be applied to any virus possessing a YXX<l> or dileucine motif in a structural protein (e.g., viral infection from HCV, Flaviviridae, Flaviviridae other than HCV, clathrin AP binding viruses, clathrin AP binding viruses other than Flaviviridae, clathrin AP binding viruses other than HCV, co• infections such as HCV/HIV co-infections), Lentiviridae or HIV. The compositions and methods can also be applied to co-infections, and infections with Flaviviridae other than HCV, and enveloped viruses other than Flaviviridae.

The instant disclosure also describes *in vitro* cell-free methods of identifying agents (inhibiting agents) that modulate binding between a viral protein containing a YXX<l> or dileucine motif and a host clathrin adaptor protein such as AP2Ml or other µ subunits of clathrin AP complexes. A test agent that inhibits binding of YXX<l> or dileucine motif-containing viral proteins to host proteins including clathrin adatpor proteins, such as AP2M l or other µ subunits of clathrin AP complexes, can be further tested for its ability to inhibit viral replication in a cell-based assay. For example, a test agent of interest can be contacted with a mammalian cell that harbors all or part of an

HCV genome, and the effect of the test agent on HCV replication can be determined. Suitable cells include mammalian liver cells that are permissive for HCV replication, e.g., an immortalized human hepatocyte cell line that is permissive for HCV. For example, a suitable mammalian cell is Huh7 hepatocyte or a subclone of Huh7 hepatocyte, e.g., Huh-7.5. Suitable cell lines are described in, e.g., Blight *et al*. (J *Viral* 76:13001, 2002) and Zhang *et al*. (J *Viral* 78:1448, 2004). In an embodiment, the HCV genome in the cell comprises a reporter, e.g., a nucleotide sequence encoding luciferase, a fluorescent protein, or other protein that provides a detectable signal; and determining the effect, if any, of the test agent on HCV replication is achieved by detection of a signal from the reporter. Other viral assay systems are known in the art.

In one embodiment, the test agents are organic moieties. In this embodiment, as is generally described in WO 94/24314, which is incorporated herein by reference, test agents are synthesized from a series of substrates that can be chemically modified. "Chemically modified" herein includes traditional chemical reactions, as well as enzymatic reactions. These substrates generally include, but are not limited to, alkyl groups (including alkanes, alkenes, alkynes, and heteroalkyl), aryl groups (including arenes and heteroaryl), alcohols, ethers, amines, aldehydes, ketones, acids, esters, amides, cyclic compounds, heterocyclic compounds (including punnes, pyrimidines, benzodiazepines, beta-lactams, tetracyclines, cephalosporins, and carbohydrates), steroids (including estrogens, androgens, cortisone, ecodysone, etc.), alkaloids (including ergots, vinca, curare, pyrollizdine, and mitomycines), organometallic compounds, hetero-atom bearing compounds, amino acids, and nucleosides. Chemical (including enzymatic) reactions may be done on the moieties to form new substrates or candidate agents which may then be tested using the present methods.

Thus, in specific embodiments, a test agent of interest (e.g., an inhibiting agent of the invention) inhibits viral replication by at least about 5%, at least about 10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, or at least about 90%, or more, compared to the level of HCV replication in the absence of the test agent.

In particular, embodiments of the present invention include inhibiting agents that inhibit binding of viral proteins containing a YXX<l> or dileucine motif to host clathrin adaptor proteins such as AP2M 1 or other μ subunits of clathrin AP complexes by at least about 5%, at least about 10%, at least about 15%, at least about 20%, at least about 25%,

at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, or at least about 90%, or more, compared to the binding of viral proteins containing a YXX<l> or dileucine motif to host clathrin adaptor proteins such as AP2M1 or other µ subunits of clathrin AP complexes in the absence of the test agent.

In yet another embodiment, the inhibiting agent is one that inhibits binding of a viral protein containing a YXX<l> or dileucine motif to host clathrin adaptor proteins such as AP2Ml or other µ subunits of clathrin AP complexes with a 50% inhibitory concentration (IC_{50}) of about 100 µM to 50 µM, about 50 µM to 25 µM, about 25 µM to 10 µM, about 10 µM to 5 µM, about 5 µM to 1 µM, about 1 µM to 500 nM, about 500 nM to 400 nM, about 400 nM to 300 nM, about 300 nM to 250 nM, about 250 nM to 200 nM, about 200 nM to 150 nM, about 150 nM to 100 nM, about 100 nM to 50 nM, about 50 nM to 30 nM, about 30 nM to 25 nM, about 25 nM to 20 nM, about 20 nM to 15 nM, about 15 nM to 10 nM, about 10 nM to 5 nM, or less than about 5 nM.

In still yet another embodiment, the inhibiting agent inhibits binding of at least one viral protein containing a YXX<l> or dileucine motif to host clathrin adaptor proteins such as AP2Ml or other µ subunits of clathrin AP complexes. In an embodiment, the inhibiting agent inhibits binding of at least one viral protein containing a YXX<l> or dileucine motif to host proteins such as AP2Ml or other µ subunits of clathrin AP complexes with an ICso of less than about 500 nM, e.g., in some embodiments, the inhibiting agent inhibits binding of at least one viral protein containing a YXX<l> or dileucine motif to host structural proteins such as AP2M1 or other µ subunits of clathrin AP complexes with an ICso of about 500 nM to 400 nM, about 400 nM to 300 nM, about 300 nM to 250 nM, about 250 nM to 200 nM, about 200 nM to 150 nM, about 150 nM to 100 nM, about 100 nM to 50 nM, about 50 nM to 30 nM, about 30 nM to 25 nM, about 25 nM to 20 nM, about 20 nM to 15 nM, about 15 nM to 10 nM, about 10 nM to 5 nM, or less than about 5 nM.

In still yet another embodiment, the inhibiting agent, when contacted with a virus•infected cell (e.g., an BCV-infected liver cell), inhibits viral replication in the cell by at least about 5%, at least about 10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, or at least about 90%, or more, compared to the level of viral replication in a viral-infected cell not contacted with the inhibiting agent.

In still yet another embodiment, the inhibiting agent, when contacted with an virus-infected cell (e.g., an BCV-infected liver cell), reduces the amount of infectious viral particles produced by the infected cell by at least about 5%, at least about 10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, or at least about 90%, or more, compared to the number of infectious viral particles produced by the cell not contacted with the inhibiting agent.

In still yet another embodiment, in addition to determining the effect of a test agent on inhibition of viral proteins containing a YXX <l> or dileucine motif to host clathrin adaptor proteins such as AP2Ml or other µ subunits of clathrin AP complexes, test agents are assessed for any cytotoxic activity they may exhibit toward a living eukaryotic cell, using well-known assays, such as trypan blue dye exclusion, an MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide) assay, and the like. Agents that do not exhibit significant cytotoxic activity may be considered preferred agents for further development as drugs.

In still yet another embodiment, the inhibiting agent, when administered in one or more doses to an individual infected with a virus as described herein (e.g., a human), reduces the viral load in the individual by at least about 5%, at least about 10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, or at least about 90%, or more, compared to the viral load in the individual not treated with the inhibiting agent.

Inhibiting Agents

Embodiments of the present disclosure provide for inhibiting agents that can be used to treat a host infected by a virus of the Flaviviridae family of viruses. In particular, Flaviviridae other than HCV. In particular, the inhibiting agent can be used to treat hosts infected with Hepatitis C virus or coinfections, in particular HCV + HIV. The inhibiting agent can be in one or more of the following groups: inhibitors of protein kinases AAK1 or GAK, which regulate viral protein-clathrin adaptor, such as AP2M 1 (or other µ subunits of clathrin AP complexes), binding and mediate virus assembly and/or budding. Since AP2M 1 and other µ subunits of clathrin AP complexes mediate various stages in the life cycles of multiple viruses, inhibition of AP2M1 and other µ subunits of clathrin

AP complexes, such as by inhibition of its regulatory protein kinases or phosphatases, may affect or inhibit multiple viruses. Accordingly, the protein kinases that regulate these host proteins may be functionally relevant in other viral families. While not exclusively specific (like most compounds), the discovered protein kinase inhibitors bind AAK1 or GAK with high affinities compared to their other targets and have no effect on HCV RNA replication, demonstrating a relatively good selectivity.

Other agents that find use as inhibitors of GAK and AAK1 include, but are not limited to, RNAi, antisense, ribozymes, or small molecules that compete with erlotinib, sunitinib, or PKC-412 for binding to GAK or AAK1. In addition, inhibitors that are able to compete with YXX<l> or dileucine motifs for binding to host proteins include, but are not limited to, binding agents such as antibodies directed to YXX<l> or dileucine or against AP2M1 or other µ subunits of clathrin AP complexes. In addition, dominant-negative binding proteins or aptamers can inhibit GAK or AAK1. In another embodiment, decoy receptors or polypeptides corresponding to the YXX<l> or dileucine binding site from AP2M1 other µ subunits of clathrin AP complexes can inhibit GAK or AAK1.

Embodiments of the present invention include salts of the inhibiting agents. Embodiments of the present invention include prodrugs of the inhibiting agents. Embodiments of the invention include in compositions, pharmaceutical compositions, liquid compositions, gel compositions, and the like, each containing an inhibiting agent identified herein, and each of these compositions, in an embodiment, can be in the form of a controlled release or a sustained release formulation. In an embodiment, the inhibiting agent can be used in combination with another agent used to treat a Flaviviridae family viral infection, and as previously noted, either of the agents (the inhibiting agent and the other agent) or both of the agents can be in the form of a controlled release or a sustained release. In some instances, the term "inhibiting agent" may be referred to as an active agent or drug.

Pharmaceutical Formulations and Routes of Administration

Embodiments of the present disclosure include one or more inhibiting agents identified herein and formulated with one or more pharmaceutically acceptable excipients, diluents, carriers and/or adjuvants. In addition, embodiments of the present invention include such inhibiting agents formulated with one or more pharmaceutically acceptable auxiliary substances. In particular, one or more inhibiting agents can be

formulated with one or more pharmaceutically acceptable excipients, diluents, carriers, and/or adjuvants to provide an embodiment of a composition of the invention.

In an embodiment, the inhibiting agent can be combined with another antiviral agent to prepare a composition of the invention, and the composition can include one or more pharmaceutically acceptable excipients, diluents, carriers and/or adjuvants.

In an embodiment, an inhibiting agent that inhibits binding of a protein containing a YXX<l> or dileucine motif to a host clathrin adaptor protein such as AP2Ml or other µ subunits of clathrin AP complexes (referred to below as "a subject active agent" or "drug") can be formulated with one or more pharmaceutically acceptable excipients, diluents, carriers, and/or adjuvants.

A wide variety of pharmaceutically acceptable excipients are known in the art. Pharmaceutically acceptable excipients have been amply described in a variety of publications, including, for example, Gennaro (2000), "Remington: The Science and Practice of Pharmacy"; Pharmaceutical Dosage Forms and Drug Delivery Systems (1999); and Handbook of Pharmaceutical Excipients (2000).

The pharmaceutically acceptable excipients, such as vehicles, adjuvants, carriers or diluents, are readily available to the public. Moreover, pharmaceutically acceptable auxiliary substances, such as pH adjusting and buffering agents, tonicity adjusting agents, stabilizers, wetting agents, and the like, are readily available to the public.

In an embodiment of the present disclosure, the inhibiting agent is administered to the host using any means capable of resulting in the desired effect (e.g., reduction in viral load, reduction in liver fibrosis, increase in liver function, and the like). Thus, the inhibiting agent can be incorporated into a variety of formulations for therapeutic administration. For example, the inhibiting agent can be formulated into pharmaceutical compositions by combination with appropriate, pharmaceutically acceptable carriers or diluents, and may be formulated into preparations in solid, semi-solid, liquid, or gaseous forms, such as tablets, capsules, powders, granules, ointments, solutions, suppositories, injections, inhalants, and aerosols.

In pharmaceutical dosage forms, the inhibiting agent may be administered in the form of its pharmaceutically acceptable salts, or a subject active agent may be used alone or in appropriate association, as well as in combination, with other pharmaceutically active compounds. The following methods and excipients are merely exemplary and are in no way limiting.

For oral preparations, the inhibiting agent can be used alone or in combination with appropriate additives to make tablets, powders, granules or capsules, for example, with conventional additives, such as lactose, mannitol, corn starch, or potato starch; with binders, such as crystalline cellulose, cellulose derivatives, acacia, corn starch, or gelatins; with disintegrators, such as corn starch, potato starch, or sodium carboxymethylcellulose; with lubricants, such as talc or magnesium stearate; and if desired, with diluents, buffering agents, moistening agents, preservatives, and flavoring agents.

Embodiments of the inhibiting agent can be formulated into preparations for injection by dissolving, suspending, or emulsifying them in an aqueous or nonaqueous solvent, such as vegetable or other similar oils, synthetic aliphatic acid glycerides, esters of higher aliphatic acids, or propylene glycol; and if desired, with conventional additives such as solubilizers, isotonic agents, suspending agents, emulsifying agents, stabilizers, and preservatives.

Embodiments of the inhibiting agent can be utilized in aerosol formulation to be administered via inhalation. Embodiments of the inhibiting agent can be formulated into pressurized acceptable propellants such as dichlorodifluoromethane, propane, nitrogen and the like.

Furthermore, embodiments of the inhibiting agent can be made into suppositories by mixing with a variety of bases such as emulsifying bases or water-soluble bases. Embodiments of the inhibiting agent can be administered rectally via a suppository. The suppository can include vehicles such as cocoa butter, carbowaxes, and polyethylene glycols, which melt at body temperature, yet are solidified at room temperature.

Unit dosage forms for oral or rectal administration, such as syrups, elixirs, and suspensions, may be provided wherein each dosage unit, for example, teaspoonful, tablespoonful, tablet, or suppository, contains a predetermined amount of the composition containing one or more inhibiting agents. Similarly, unit dosage forms for injection or intravenous administration may comprise the inhibiting agent in a composition as a solution in sterile water, normal saline, or another pharmaceutically acceptable carrier.

Embodiments of the inhibiting agent can be formulated in an injectable composition in accordance with the invention. Typically, injectable compositions are prepared as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid vehicles prior to injection may also be prepared. The preparation

may also be emulsified or the active ingredient (inhibiting agent) encapsulated in liposome vehicles in accordance with the invention.

In an embodiment, the inhibiting agent is formulated for delivery by a continuous delivery system. The term "continuous delivery system" is used interchangeably herein with "controlled delivery system" and encompasses continuous (e.g., controlled) delivery devices (e.g., pumps) in combination with catheters, injection devices, and the like, a wide variety of which are known in the art.

Mechanical or electromechanical infusion pumps can also be suitable for use with the present disclosure. Examples of such devices include those described in, for example, U.S. Pat. Nos. 4,692,147; 4,360,019; 4,487,603; 4,360,019; 4,725,852; 5,820,589; 5,643,207; 6,198,966; and the like. In general, delivery of the inhibiting agent can be accomplished using any of a variety of refillable pump systems. Pumps provide consistent, controlled release over time. In some embodiments, the inhibiting agent is in a liquid formulation in a drug-impermeable reservoir, and is delivered in a continuous fashion to the individual.

In one embodiment, the drug delivery system is an at least partially implantable device. The implantable device can be implanted at any suitable implantation site using methods and devices well known in the art. An implantation site is a site within the body of a subject at which a drug delivery device is introduced and positioned. Implantation sites include, but are not necessarily limited to, a subdermal, subcutaneous, intramuscular, or other suitable site within a subject's body. Subcutaneous implantation sites are used in some embodiments because of convenience in implantation and removal of the drug delivery device.

Drug release devices suitable for use in the disclosure may be based on any of a variety of modes of operation. For example, the drug release device can be based upon a diffusive system, a convective system, or an erodible system (e.g., an erosion-based system). For example, the drug release device can be an electrochemical pump, osmotic pump, an electroosmotic pump, a vapor pressure pump, or osmotic bursting matrix, e.g., where the drug is incorporated into a polymer and the polymer provides for release of drug formulation concomitant with degradation of a drug-impregnated polymeric material (e.g., a biodegradable, drug-impregnated polymeric material). In other embodiments, the drug release device is based upon an electrodiffusion system, an electrolytic pump, an effervescent pump, a piezoelectric pump, a hydrolytic system, etc.

Drug release devices based upon a mechanical or electromechanical infusion pump can also be suitable for use with the present disclosure. Examples of such devices include those described in, for example, U.S. Pat. Nos. 4,692,147; 4,360,019; 4,487,603; 4,360,019; 4,725,852, and the like. In general, a subject treatment method can be accomplished using any of a variety of refillable, non-exchangeable pump systems. Pumps and other convective systems are generally preferred due to their generally more consistent, controlled release over time. Osmotic pumps are used in some embodiments due to their combined advantages of more consistent controlled release and relatively small size (see, e.g., PCT published application no. WO 97/27840 and U.S. Pat. Nos. 5,985,305 and 5,728,396). Exemplary osmotically-driven devices suitable for use in the disclosure include, but are not necessarily limited to, those described in U.S. Pat. Nos. 3,760,984; 3,845,770; 3,916,899; 3,923,426; 3,987,790; 3,995,631; 3,916,899; 4,016,880; 4,036,228; 4,111,202; 4,111,203; 4,203,440; 4,203,442; 4,210,139; 4,327,725; 4,627,850; 4,865,845; 5,057,318; 5,059,423; 5,112,614; 5,137,727; 5,234,692; 5,234,693; 5,728,396; and the like.

In some embodiments, the drug delivery device is an implantable device. The drug delivery device can be implanted at any suitable implantation site using methods and devices well known in the art. As noted herein, an implantation site is a site within the body of a subject at which a drug delivery device is introduced and positioned. Implantation sites include, but are not necessarily limited to a subdermal, subcutaneous, intramuscular, or other suitable site within a subject's body.

In some embodiments, an active agent is delivered using an implantable drug delivery system, e.g., a system that is programmable to provide for administration of the agent. Exemplary programmable, implantable systems include implantable infusion pumps. Exemplary implantable infusion pumps, or devices useful in connection with such pumps, are described in, for example, U.S. Pat. Nos. 4,350,155; 5,443,450; 5,814,019; 5,976,109; 6,017,328; 6,171,276; 6,241,704; 6,464,687; 6,475,180; and 6,512,954. A further exemplary device that can be adapted for the present disclosure is the Synchromed Infusion pump (Medtronic).

Suitable excipient vehicles for the inhibiting agent are, for example, water, saline, dextrose, glycerol, ethanol, or the like, and combinations thereof. In addition, if desired, the vehicle may contain minor amounts of auxiliary substances such as wetting or emulsifying agents or pH buffering agents. Methods of preparing such dosage forms are known, or will be apparent upon consideration of this disclosure, to those skilled in the

art. See, e.g., Remington's Pharmaceutical Sciences, 17th edition, 1985. The composition or formulation to be administered will, in any event, contain a quantity of the inhibiting agent adequate to achieve the desired state in the subject being treated.

Compositions of the present invention include those that comprise a sustained-release or controlled release matrix. In addition, embodiments of the present invention can be used in conjunction with other treatments that use sustained-release formulations. As used herein, a sustained-release matrix is a matrix made of materials, usually polymers, which are degradable by enzymatic or acid-based hydrolysis or by dissolution. Once inserted into the body, the matrix is acted upon by enzymes and body fluids. A sustained-release matrix desirably is chosen from biocompatible materials such as liposomes, polylactides (polylactic acid), polyglycolide (polymer of glycolic acid), polylactide co-glycolide (copolymers of lactic acid and glycolic acid), polyanhydrides, poly(ortho)esters, polypeptides, hyaluronic acid, collagen, chondroitin sulfate, carboxcylic acids, fatty acids, phospholipids, polysaccharides, nucleic acids, polyamino acids, amino acids such as phenylalanine, tyrosine, isoleucine, polynucleotides, polyvinyl propylene, polyvinylpyrrolidone, and silicone. Illustrative biodegradable matrices include a polylactide matrix, a polyglycolide matrix, and a polylactide co-glycolide (co-polymers of lactic acid and glycolic acid) matrix.

In another embodiment, the pharmaceutical composition of the present disclosure (as well as combination compositions) is delivered in a controlled release system. For example, the inhibiting agent may be administered using intravenous infusion, an implantable osmotic pump, a transdermal patch, liposomes, or other modes of administration. In one embodiment, a pump may be used (Sefton 1987; Buchwald *et al.* 1980; Saudek *et al.* 1989). In another embodiment, polymeric materials are used. In yet another embodiment a controlled release system is placed in proximity of the therapeutic target, i.e., the liver, thus requiring only a fraction of the systemic dose. In yet another embodiment, a controlled release system is placed in proximity of the therapeutic target, thus requiring only a fraction of the systemic. Other controlled release systems are discussed in the review by Langer (1990).

In another embodiment, the compositions of the present invention (as well as combination compositions separately or together) include those formed by impregnation of an inhibiting agent described herein into absorptive materials, such as sutures, bandages, and gauze, or coated onto the surface of solid phase materials, such as surgical

staples, zippers and catheters to deliver the compositions. Other delivery systems of this type will be readily apparent to those skilled in the art in view of the instant disclosure.

Treatment Methods

Embodiments of the present invention include methods of treating an infection by viral infection from clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV. In particular, inhibiting agents described herein can be used to treat an infection by a virus of the Flaviviridae family of viruses. In an embodiment, the present disclosure provides a method of treating a host infected with a virus as described above by administering to the host a therapeutically effective amount of an inhibiting agent in one or more doses, to reduce the viral load in the host.

Embodiments of the present invention include methods of prophylactically treating an infection by a viral infection from clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV. In particular, inhibiting agents described herein can be used to prophylactically treat an infection by a virus of the Flaviviridae family of viruses. In an embodiment, the present disclosure provides a method of prophylactically treating a host infected with a virus from the Flaviviridae family of viruses by administering to the host a therapeutically effective amount of an inhibiting agent in one or more doses, to reduce the viral load in the host. In an embodiment, a method of prophylactically treating a host infected with a virus from the Flaviviridae family of viruses, the method comprising administering to the host a therapeutically effective amount of an inhibiting agent to reduce the viral load in the host. Additional details regarding clemizole and dosing of clemizole is described above.

In an embodiment, inhibiting agents described herein are used in combination with another agent (e.g., an antiviral agent) to treat an infection with viral infection from clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV. In an embodiment, inhibiting agents described herein are used in combination with another agent (e.g., an antiviral agent) to prophylactically treat an infection with a virus from the Flaviviridae

family of viruses. Embodiments of the method involve administering to an individual in need thereof one or more inhibiting agents that inhibit binding of viral proteins containing a YXX<l> or dileucine motif to host clathrin adaptor proteins such as AP2Ml or other μ subunits of clathrin AP complexes. In an embodiment, the present invention provides methods of treating a flavivirus infection, e.g., an HCV infection, and methods of reducing liver fibrosis that may occur as sequelae of an HCV infection.

In an embodiment, the inhibiting agent includes one or more of inhibiting agents described above. In various embodiments, the inhibiting agent is a protein kinase inhibitor, such as inhibitors of AAK l or GAK, which regulates viral protein-AP2M l binding or binding of a viral protein to other μ subunits of clathrin AP complexes and mediate virus assembly and/or budding, or variants of these proteins thereof.

In an embodiment, an effective amount of the inhibiting agent is an amount that, when administered in one or more doses to a host (e.g., human) in need thereof, reduces viral load in the individual by at least about 10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 55%, at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, or at least about 90%, or more, compared to the viral load in the individual not treated with the inhibiting agent.

Viral load can be measured by measuring the titer or level of virus in serum. These methods include, but are not limited to, a quantitative polymerase chain reaction (PCR) and a branched DNA (bDNA) test. Quantitative assays for measuring the viral load (titer) of HCV RNA have been developed. Many such assays are available commercially, including a quantitative reverse transcription PCR (qRT-PCR) (Amplicor HCV Monitor.TM., Roche Molecular Systems, New Jersey); and a branched DNA signal amplification assay [Quantiplex™ HCV RNA Assay (bDNA), Chiron Corp., Emeryville, CA.]. See, e.g., Gretch *et al*. (1995). Also of interest is a nucleic acid test (NAT) sold by Chiron Corporation under the trade name ProcleixTM, which NAT simultaneously tests for the presence of HIV-1 and HCV (Vargo *et al.* 2002).

In some embodiments, an effective amount of the inhibiting agent is an amount that, when administered in one or more doses to a host (e.g., human) in need thereof, increases liver function in the individual by at least about 10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 55%, at least about 60%, at

least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, or at least about 90%, or more, compared to the liver function in the individual not treated with the inhibiting agent.

In some embodiments, an effective amount of the inhibiting agent is an amount that, when administered in one or more doses to a host (e.g., a human) in need thereof, reduces liver fibrosis in the host by at least about 10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 55%, at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, or at least about 90%, or more, compared to the degree of liver fibrosis in the individual not treated with the inhibiting agent.

Embodiments of the present disclosure provide methods, inhibiting agents, and pharmaceutical formulations useful in the treatment of patients suffering from a viral infection from clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV. In one embodiment, the patient is co-infected with Flaviviridae other than HCV. In one embodiment, the patient is infected with HCV but is not known to be infected with another virus, including, but not limited to, HIV. In another embodiment, the patient is infected with HCV and one or more additional viruses, including, but not limited to, HIV. In one embodiment, the patient is treated for a viral infection by administering only a single inhibiting agent as described herein as useful in the treatment of HCV infection. In another embodiment, the patient is treated for a viral infection by administering both an inhibiting agent described herein as useful in the treatment of HCV infection as well as one or more additional agents known to be useful in the treatment of viral infection, including, but not limited to drugs for the treatment of HIV, such as Atripla, Complera, Combivir, Retrovir, Truvada, Viracept, Fuzeon, Selzentry, Isentress, or the like. In one embodiment, the one or more additional agents does not include a CCR-5 antagonist. In another embodiment, the one or more additional agents does include a CCR-5 antagonist, and the patient is infected with HCV but not known to be infected (or is not infected) with HIV.

Dosages

Embodiments of the inhibiting agent can be administered to a host in one or more doses. In an embodiment, the inhibiting agent can be administered in an amount of about 10 mg to 1000 mg per dose, e.g., about 10 mg to 20 mg, about 20 mg to 25 mg, about 25 mg to 50 mg, about 50 mg to 75 mg, about 75 mg to 100 mg, about 100 mg to 125 mg, about 125 mg to 150 mg, about 150 mg to 175 mg, about 175 mg to 200 mg, about 200 mg to 225 mg, about 225 mg to 250 mg, about 250 mg to 300 mg, about 300 mg to 350 mg, about 350 mg to 400 mg, about 400 mg to 450 mg, about 450 mg to 500 mg, about 500 mg to 750 mg, or about 750 mg to 1000 mg per dose.

In an embodiment, the amount of the inhibiting agent per dose is determined on a per body weight basis. For example, in an embodiment, the inhibiting agent can be administered in an amount of about 0.5 mg/kg to 100 mg/kg, e.g., about 0.5 mg/kg to 1 mg/kg, about 1 mg/kg to 2 mg/kg, about 2 mg/kg to 3 mg/kg, about 3 mg/kg to 5 mg/kg, about 5 mg/kg to 7 mg/kg, about 7 mg/kg to about 10 mg/kg, about 10 mg/kg to 15 mg/kg, about 15 mg/kg to 20 mg/kg, about 20 mg/kg to 25 mg/kg, about 25 mg/kg to 30 mg/kg, about 30 mg/kg to 40 mg/kg, about 40 mg/kg to 50 mg/kg, about 50 mg/kg to 60 mg/kg, about 60 mg/kg to 70 mg/kg, about 70 mg/kg to 80 mg/kg, about 80 mg/kg to 90 mg/kg, or about 90 mg/kg to 100 mg/kg, or more than about 100 mg/kg.

Those of skill will readily appreciate that dose levels can vary as a function of the specific inhibiting agent administered, the severity of the symptoms and the susceptibility of the subject to side effects. Preferred dosages for a given compound are readily determinable by those of skill in the art by a variety of means.

In an embodiment, multiple doses of the inhibiting agent are administered. The frequency of administration of the inhibiting agent can vary depending on any of a variety of factors, e.g., severity of the symptoms, and the like. For example, in an embodiment, the inhibiting agent is administered once per month, twice per month, three times per month, every other week (qow), once per week (qw), twice per week (biw), three times per week (tiw), four times per week, five times per week, six times per week, every other day (qod), daily (qd), twice a day (qid), or three times a day (tid). As discussed above, in an embodiment, the inhibiting agent is administered continuously.

The duration of administration of the inhibiting agent, e.g., the period of time over which the inhibiting agent is administered, can vary, depending on any of a variety of factors, e.g., patient response, etc. For example, the inhibiting agent can be administered over a period of time of about one day to one week, about two weeks to four weeks, about

one month to two months, about two months to four months, about four months to six months, about six months to eight months, about eight months to 1 year, about 1 year to 2 years, or about 2 years to 4 years, or more.

Routes of Administration

Embodiments of the present invention provide methods and compositions for the administration of the inhibiting agent to a host (e.g., a human) using any available method and route suitable for drug delivery, including in vivo and ex vivo methods, as well as systemic and localized routes of administration.

Routes of administration include intranasal, intramuscular, intratracheal, subcutaneous, intradermal, topical application, intravenous, rectal, nasal, oral, and other enteral and parenteral routes of administration. Routes of administration may be combined, if desired, or adjusted depending upon the agent and/or the desired effect. An active agent can be administered in a single dose or in multiple doses.

Embodiments of the inhibiting agent can be administered to a host using available conventional methods and routes suitable for delivery of conventional drugs, including systemic or localized routes. In general, routes of administration contemplated by the disclosure include, but are not limited to, enteral, parenteral, or inhalational routes.

Parenteral routes of administration other than inhalation administration include, but are not limited to, topical, transdermal, subcutaneous, intramuscular, intraorbital, intracapsular, intraspinal, intrasternal, and intravenous routes, i.e., any route of administration other than through the alimentary canal. Parenteral administration can be conducted to effect systemic or local delivery of the inhibiting agent. Where systemic delivery is desired, administration typically involves invasive or systemically absorbed topical or mucosal administration of pharmaceutical preparations.

The inhibiting agent can also be delivered to the subject by enteral administration. Enteral routes of administration include, but are not limited to, oral and rectal (e.g., using a suppository) delivery.

Methods of administration of the inhibiting agent through the skin or mucosa include, but are not limited to, topical application of a suitable pharmaceutical preparation, transdermal transmission, injection and epidermal administration. For transdermal transmission, absorption promoters or iontophoresis are suitable methods. Iontophoretic transmission may be accomplished using commercially available "patches"

that deliver their product continuously via electric pulses through unbroken skin for periods of several days or more.

Combination Therapies

Embodiments of the present invention include methods, inhibiting agents, and pharmaceutical formulations for the treatment of viral infection. Embodiments of the inhibiting agents and pharmaceutical formulations useful in the methods of the present disclosure can be employed in combination with other antiviral agents to treat viral infection. In an embodiment, in accordance with the methods of the present invention, an inhibiting agent that is used to treat a host infected by viral infection from clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV is used in combination with one or more other antiviral agents to treat the infection. In an embodiment, in accordance with the methods of the present invention, an inhibiting agent that prevents the binding of at least one viral protein containing a YXX<l> or dileucine motif to host proteins such as AP2Ml or other µ subunits of clathrin AP complexes(also referred to herein as an "HCV YXX<l> or dileucine antagonist") can be used in combination with one or more other antiviral agents to treat viral infection.

For instance, current medical practice to treat HCV infection typically employs combination therapy with ribavirin (such as Rebetol or Copegus), either an interferon• alpha (such as interferon alpha 2b) or pegylated interferon (such as Pegasys, marketed by Roche, or PEG-Intron, marketed by Schering Plough), and a protease inhibitor. In accordance with the methods of the present disclosure, an inhibiting compound can be used in combination with these standard therapies to treat HCV infection.

A number of HCV protease and polymerase inhibitors are either approved or in development for the treatment of HCV infection, and in accordance with the methods of the present disclosure, co-administration of an inhibiting agent that prevents the binding of at least one viral protein containing a YXX<l> or dileucine motif to host proteins such as AP2Ml or other µ subunits of clathrin AP complexes and an HCV protease inhibitor can be efficacious in the treatment of HCV. In one embodiment, an interferon alpha and/or a nucleoside analog such as ribavirin is/are also employed in this combination therapy. Suitable HCV protease inhibitors include, but are not limited to, telaprevir (VX-950, Vertex), BILN 2061 and BI12202 (Boehringer Ingelheim), boceprevir (SCH 503034,

Schering Plough), ITMN191 (Roche/InterMune/Array BioPharma), MK-7009 (Merck), TMC435350 (Tibotec/Medivir), ACH-1095 and ACH-806 (Achillion/Gilead), and other inhibitors of NS3/NS4A protease, including, but not limited to, compounds in development by Presidio.

In accordance with the methods of the present disclosure, co-administration of an inhibiting agent that prevents the binding of at least one viral protein containing a YXX<l> or dileucine motif to host proteins such as AP2M 1 or other µ subunits of clathrin AP complexes and an HCV RNA polymerase inhibitor can be efficacious in the treatment of HCV. In one embodiment, an interferon alpha and/or a nucleoside analog such as ribavirin and/or an HCV protease inhibitor is/are also employed in this combination therapy. Suitable HCV RNA polymerase inhibitors include, but are not limited to, valopicitabine (NM283, Idenix/Novartis), HCV-796 (Wyeth/ViroPharma), R l626 (Roche), R7128 (Roche/Pharmasset), GS-9190 (Gilead), MK-0608 (Merck), PSI-6130 (Pharmasset), and PFE-868,554 (PFE). In an embodiment, the method provides combination treatments with agents that inhibit AAK l and GAK.

A number of toll-like receptor (TLR) agonists are m development for the treatment of HCV infection, and in accordance with the methods of the present disclosure, co-administration of a YXX<l> or dileucine antagonist that prevents the binding of at least one viral protein containing a YXX<l> or dileucine motif to host proteins such as AP2Ml or other µ subunits of clathrin AP complexes and a TLR agonist can be efficacious in the treatment of HCV. In one embodiment, an interferon alpha and/or a nucleoside analog such as ribavirin and/or an HCV protease inhibitor and/or an HCV RNA polymerase inhibitor is/are also employed in this combination therapy. Suitable TLR agonists include, but are not limited to, TLR7 agonists [i.e., ANA245 and ANA975 (Anadys/Novartis)] and TLR9 agonists [i.e., Actilon (Coley) and IM0-2125 (Idera)].

A number of thiazolide derivatives are in development for the treatment of HCV infection, and in accordance with the methods of the present disclosure, co-administration of an antagonist that prevents the binding of at least one viral protein containing a YXX<l> or dileucine motif to host proteins such as AP2M 1 or other µ subunits of clathrin AP complexes, and a thiazolide, including, but not limited to, Nitazoxanide (Alinia, or other sustained release formulations of nitazoxanide or other thiazolides, Romark Laboratories) can be efficacious in the treatment of HCV. In an embodiment, an interferon alpha and/or a nucleoside analog such as ribavirin and/or an HCV protease inhibitor and/or an HCV

RNA polymerase inhibitor and/or a TLR agonist is/are also employed in this combination therapy.

In another embodiment of the methods of the present disclosure, co-administration of an inhibiting agent that prevents the binding of at least one viral protein containing a YXX<l> or dileucine motif to host proteins such as AP2M 1 or other μ subunits of clathrin AP complexes, and a cyclophilin inhibitor [i.e., NIM-811 (Novartis) and DEBI0-025 (Debiopharm)] and/or an alpha-glucosidase inhibitor [i.e., Celgosivir (Migenix)] and/or one or more agents from one or more of the other classes of HCV therapeutic agents discussed herein is used to treat HCV infection.

Other agents that can be used in combination with inhibiting agents of the present disclosure that prevent the binding of at least one viral protein containing a YXX<l> or dileucine motif to host proteins such as AP2Ml or other μ subunits of clathrin AP complexes include (i) agents targeting NS5A, including, but not limited to, A-831 (Arrow Therapeutics), AZD2836 (Astra Zeneca), and agents in development by XTL/Presidio or BMS (see PCT publications WO 2006/133326 and WO 2008/021928, incorporated herein by reference); (ii) agents targeting TBC1D20 and/or NS5A's interaction with TBC1D20 (see PCT publication WO 2007/018692 and U.S. patent application Ser. No. 11/844,993, incorporated herein by reference), (iii) agents targeting NS4B's GTPase activity (see PCT publication WO 2005/032329 and US patent application publication 2006/0199174, incorporated herein by reference); (iv) agents inhibiting membrane association mediated by the HCV amphipathic helices, such as those found in NS5A, NS4B, and NS5B (see PCT publication WO 2002/089731, supra), (v) agents targeting PIP2 or BAAPP domains in HCV proteins, such as those found in NS4B and NS5A (see U.S. provisional patent application 60/057,188, supra); (vi) agents targeting HCV entry, assembly, or release, including antibodies to co-receptors; (vii) agents targeting HCV NS3 helicase; (viii) siRNAs, shRNAs, antisense RNAs, or other RNA-based molecules targeting sequences in HCV; (ix) agents targeting microRNA122 or other microRNAs modulating HCV replication; (x) agents targeting PD-1, PD-Ll, or PD-L2 interactions or pathways (see US patent application publications 2008/0118511, 2007/0065427, 2007/0122378, incorporated herein by reference); and (xi) agents targeting HCV amphipathic helix function, such as AH2 inhibitors.

In another embodiment of the present disclosure, an inhibiting agent that prevents the binding of at least one viral protein containing a YXX<l> or dileucine motif to host proteins such as AP2Ml or other μ subunits of clathrin AP complexes is used in

combination with one or more drugs capable of treating an HIV infection to treat a patient that is co-infected with HIV and HCV. In another embodiment of the present disclosure, an inhibiting agent that prevents the binding of at least one viral protein containing a YXX<l> or dileucine motif to host proteins such as AP2M 1 or other μ subunits of clathrin AP complexes is used in combination with one or more drugs capable of treating an HBV infection to treat a patient that is co-infected with HBV and HCV. In an embodiment, an inhibiting agent that prevents the binding of at least one viral protein containing a YXX<l> or dileucine motif to host proteins such as AP2M 1 or other μ subunits of clathrin AP complexes is used in combination with a PD-Ll inhibitor to treat a viral infection.

As mentioned above, embodiments of the present include the administration of an inhibiting agent identified herein (or by using an embodiment of the screen of the invention) in conjunction with at least one additional therapeutic agent to treat a viral infection. Suitable additional therapeutic agents include, but are not limited to, ribavirin; a nucleoside analog (e.g., levovirin, viramidine, etc.); an NS3 inhibitor; an NS5 inhibitor; an interferon; and a side effect management agent.

In an embodiment, the at least one additional suitable therapeutic agent includes ribavirin. Ribavirin, 1- -D-ribofuranosyl- lH-1,2,4-triazole-3-carboxamide, available from ICN Pharmaceuticals, Inc., Costa Mesa, Calif., is described in the Merck Index, compound No. 8199, Eleventh Edition. Its manufacture and formulation is described in U.S. Pat. No. 4,211,771. The disclosure also contemplates use of derivatives of ribavirin (see, e.g., U.S. Pat. No. 6,277,830).

In an embodiment, the at least one additional suitable therapeutic agent includes levovirin. Levovirin is the L-enantiomer of ribavirin, and exhibits the property of enhancing a Th1 immune response over a Th2 immune response. Levovirin 1s manufactured by ICN Pharmaceuticals.

In an embodiment, the at least one additional suitable therapeutic agent includes viramidine. Viramidine is a 3-carboxamidine derivative of ribavirin, and acts as a prodrug of ribavirin. It is efficiently converted to ribavirin by adenosine deaminases.

Nucleoside analogs that are suitable for use in a combination therapy include, but are not limited to, ribavirin, levovirin, viramidine, isatoribine, an L-ribofuranosyl nucleoside as disclosed in U.S. Pat. No. 5,559,101 and encompassed by Formula I of U.S. Pat. No. 5,559,101 (e.g., 1--L-ribofuranosyluracil, l--L-ribofuranosyl-5-fluorouracil, 1- -L-ribofuranosylcytosine, 9- -L-ribofuranosyladenine, 9- -L-ribofuranosylhypoxanthine, 9- -L-ribofuranosylguanine, 9- -L-ribofuranosyl-6-

thioguanine, 2-amino-a-L-ribofuranl[l ',2':4,5]oxazoline, 0_2,0ranhydro- l-a-L• ribofuranosyluracil, 1-a-L-ribofuranosyluracil, 1-(2,3,5-tri-O-benzoyl-a-ribofuranosyl)-4-thiouracil, 1-a-L-ribofuranosylcytosine, l-a-L-ribofuranosyl-4-thiouracil, l-a-L-ribofuranosyl-5-fluorouracil, 2-amino--L-arabinofurano[l',2':4,5]oxazoline, 0_2,0_2-anhydro--L-arabinofuranosyluracil, 2'-deoxy- -L-uridine, 3'5'-Di-O-benzoyl-2'deoxy-4-thio -L-uridine, 2'-deoxy- -L-cytidine, 2'-deoxy--L-4-thiouridine, 2'-deoxy- -L• thymidine, 2'-deoxy- -L-5-fluorouridine, 2',3'-dideoxy- -L-uridine, 2'-deoxy- -L-5-fluorouridine, and 2'-deoxy--L-inosine); a compound as disclosed in U.S. Pat. No. 6,423,695 and encompassed by Formula I of U.S. Pat. No. 6,423,695; a compound as disclosed in U.S. Patent Publication No. 2002/0058635, and encompassed by Formula 1 of U.S. Patent Publication No. 2002/0058635; a nucleoside analog as disclosed in WO 01/90121 A2 (Idenix); a nucleoside analog as disclosed in WO 02/069903 A2 (Biocryst Pharmaceuticals Inc.); a nucleoside analog as disclosed in WO 02/057287 A2 or WO 02/057425 A2 (both Merck/Isis); and the like.

In an embodiment, the at least one additional suitable therapeutic agent can include HCV NS3 inhibitors. Suitable HCV non-structural protein-3 (NS3) inhibitors include, but are not limited to, a tri-peptide as disclosed in U.S. Pat. Nos. 6,642,204; 6,534,523; 6,420,380; 6,410,531; 6,329,417; 6,329,379; and 6,323,180 (Boehringer• Ingelheim); a compound as disclosed in U.S. Pat. No. 6,143,715 (Boehringer-Ingelheim); a macrocyclic compound as disclosed in U.S. Pat. No. 6,608,027 (Boehringer-Ingelheim); an NS3 inhibitor as disclosed in U.S. Pat. Nos. 6,617,309; 6,608,067; and 6,265,380 (Vertex Pharmaceuticals); an azapeptide compound as disclosed in U.S. Pat. No. 6,624,290 (Schering); a compound as disclosed in U.S. Pat. No. 5,990,276 (Schering); a compound as disclosed in Pause *et al.* (2003); NS3 inhibitor BILN 2061 (Boehringer-Ingelheim; Lamarre *et al.* (2002), and Lamarre *et al.* (2003); NS3 inhibitor VX-950 (Vertex Pharmaceuticals; Kwong *et al.* (2003); NS3 inhibitor SCH6 (Abib *et al.* (2003); Program and Abstracts of the 54th Annual Meeting of the American Association for the Study of Liver Diseases (AASLD, 2003); any of the NS3 protease inhibitors disclosed in WO 99/07733, WO 99/07734, WO 00/09558, WO 00/09543, WO 00/59929, or WO 02/060926 (e.g., compounds 2, 3, 5, 6, 8, 10, 11, 18, 19, 29, 30, 31, 32, 33, 37, 38, 55, 59, 71, 91, 103, 104, 105, 112, 113, 114, 115, 116, 120, 122, 123, 124, 125, 126, and 127 disclosed in the table of pages 224-226 in WO 02/060926); an NS3 protease inhibitor as disclosed in any one of U.S. Patent Publication Nos. 2003/019067, 2003/0187018, and 2003/0186895; and the like.

In an embodiment, the NS3 inhibitor used in a combination therapy of the invention is a member of the class of specific NS3 inhibitors, e.g., NS3 inhibitors that inhibit NS3 serine protease activity and that do not show significant inhibitory activity against other serine proteases such as human leukocyte elastase, porcine pancreatic elastase, or bovine pancreatic chymotrypsin, or cysteine proteases such as human liver cathepsin B.

In an embodiment, the at least one additional suitable therapeutic agent includes NS5B inhibitors. Suitable HCV non-structural protein-5 (NS5; RNA-dependent RNA polymerase) inhibitors include, but are not limited to, a compound as disclosed in U.S. Pat. No. 6,479,508 (Boehringer-Ingelheim); a compound as disclosed in any of International Patent Application Nos. PCT/CA02/01127, PCT/CA02/01128, and PCT/CA02/01129, all filed on Jul. 18, 2002 by Boehringer Ingelheim; a compound as disclosed in U.S. Pat. No. 6,440,985 (ViroPharma); a compound as disclosed in WO 01/47883, e.g., JTK-003 (Japan Tobacco); a dinucleotide analog as disclosed in Zhong *et al.* (2003); a benzothiadiazine compound as disclosed in Dhanak *et al* . (2002); an NS5B inhibitor as disclosed in WO 02/100846 A l or WO 02/100851 A2 (both Shire); an NS5B inhibitor as disclosed in WO 01/85172 A l or WO 02/098424 A l (both Glaxo SmithKline); an NS5B inhibitor as disclosed in WO 00/06529 or WO 02/06246 A l (both Merck); an NS5B inhibitor as disclosed in WO 03/000254 (Japan Tobacco); an NS5B inhibitor as disclosed in EP 1 256,628 A2 (Agouron); JTK-002 (Japan Tobacco); JTK-109 (Japan Tobacco); and the like.

In an embodiment, the NS5 inhibitor used in the combination therapies of the invention is a member of the class of specific NS5 inhibitors, e.g., NS5 inhibitors that inhibit NS5 RNA-dependent RNA polymerase and that lack significant inhibitory effects toward other RNA dependent RNA polymerases and toward DNA dependent RNA polymerases.

In an embodiment, the at least one additional therapeutic agent is an interferon, e.g., interferon-alpha (IFN-a). Any known IFN-a. can be used in the treatment methods of the invention. The term "interferon-alpha" as used herein refers to a family of related polypeptides that inhibit viral replication and cellular proliferation and modulate immune response. The term "IFN-a" includes naturally occurring IFN-a; synthetic IFN-a; derivatized IFN-a (e.g., PEGylated IFN-a, glycosylated IFN-a, and the like); and analogs of naturally occurring or synthetic IFN-a; essentially any IFN-a that has antiviral properties, as described for naturally occurring IFN-a.

Suitable a intelferons include, but are not limited to, naturally-occurring IFN-a (including, but not limited to, naturally occurring IFN-a2a, IFN-a2b); recombinant intelferon a-2b such as Intron-A interferon available from Schering Corporation, Kenilworth, N.J.; recombinant intelferon a-2a such as Roferon interferon available from Hoffmann-La Roche, Nutley, N.J.; recombinant intelferon a-2C such as Berofor a2 intelferon available from Boehringer Ingelheim Pharmaceutical, Inc., Ridgefield, Conn.; intelferon a-n l, a purified blend of natural a interferons such as Sumiferon available from Sumitomo, Japan or as Wellferon interferon a-n l (INS) available from the Glaxo• Wellcome Ltd., London, Great Britain; and interferon a-n3 a mixture of natural a intelferons made by Intelferon Sciences and available from the Purdue Frederick Co., Norwalk, Conn., under the Alferon tradename.

The term "IFN-a" also encompasses consensus IFN-a. Consensus IFN-a (also referred to as "CIFN" and "IFN-con" and "consensus intelferon") encompasses, but is not limited to, the amino acid sequences designated IFN-con $_{1,}$ IFN-con $_{2}$ and IFN-con $_{3}$ which are disclosed in U.S. Pat. Nos. 4,695,623 and 4,897,471; and consensus interferon as defined by determination of a consensus sequence of naturally occurring interferon alphas (e.g., lnfergen.RTM., InterMune, Inc., Brisbane, CA.). IFN-con $_{1}$ is the consensus intelferon agent in the lnfergenTM alfacon-1 product. The lnfergenTM consensus intelferon product is referred to herein by its brand name (lnfergenTM) or by its generic name (interferon alfacon-1). DNA sequences encoding IFN-con may be synthesized as described in the aforementioned patents or other standard methods. In an embodiment, the at least one additional therapeutic agent is CIFN.

In an embodiment, fusion polypeptides comprising an IFN-a and a heterologous polypeptide can also be used in the combination therapies of the invention. Suitable IFN-a fusion polypeptides include, but are not limited to, Albuferon-alphaTM [a fusion product of human albumin and IFN-a; Human Genome Sciences; see, e.g., Osborn *et al.* (2002)]. Also suitable for use in the present disclosure are gene-shuffled forms of IFN-a. See, e.g., Masci *et al.* (2003). Other suitable interferons include), Multiferon (Viragen), Medusa Interferon (Flamel Technology), Locteron (Octopus), and Omega Intelferon (lntarcia/Boehringer Ingelheim).

The term "IFN-a" also encompasses derivatives of IFN-a that are derivatized (e.g., are chemically modified relative to the naturally occurring peptide) to alter certain properties such as serum half-life. As such, the term "IFN-a" includes glycosylated IFN• a; IFN-a derivatized with polyethylene glycol ("PEGylated IFN-a"); and the like.

PEGylated IFN-a, and methods for making same, is discussed in, e.g., U.S. Pat. Nos. 5,382,657; 5,981,709; and 5,951,974. PEGylated IFN-a encompasses conjugates of PEG and any of the above-described IFN-a molecules, including, but not limited to, PEG conjugated to interferon alpha-2a (Roferon, Hoffman La-Roche, Nutley, NJ.), interferon alpha 2b (Intron, Schering-Plough, Madison, NJ.), interferon alpha-2c (Berofor Alpha, Boehringer Ingelheim, Ingelheim, Germany); and consensus interferon as defined by determination of a consensus sequence of naturally occurring interferon alphas (Infergen™, InterMune, Inc., Brisbane, CA).

In an embodiment, the IFN-a polypeptides can be modified with one or more polyethylene glycol moieties, i.e., PEGylated. The PEG molecule of a PEGylated IFN-a polypeptide is conjugated to one or more amino acid side chains of the IFN-a polypeptide. In an embodiment, the PEGylated IFN-a contains a PEG moiety on only one amino acid. In another embodiment, the PEGylated IFN-a contains a PEG moiety on two or more amino acids, e.g., the IFN-a contains a PEG moiety attached to two, three, four, five, six, seven, eight, nine, or ten different amino acid residues. IFN-a may be coupled directly to PEG (i.e., without a linking group) through an amino group, a sulfhydryl group, a hydroxyl group, or a carboxyl group.

To determine the optimum combination of an inhibiting agent, such as PKC-412, erlotinib, sunitinib, and the like, with other anti-HCV agents, HCV replication assays and/or animal studies can be performed in the presence of various combinations of the various anti-HCV agents. Increased inhibition of replication in the presence of an additional agent (above that observed with monotherapy) is evidence for the potential benefit of the combination therapy.

For example, HCV replication assays employing a luciferase reporter-linked HCV genome in the presence of various combinations of PKC-412, erlotinib, sunitinib, and the like. In such assays, luciferase activity is directly proportional to HCV RNA genome replication.

In an embodiment, side effect management agents can be used in the treatment methods of the invention, and these include agents that are effective in pain management; agents that ameliorate gastrointestinal discomfort; analgesics, anti-inflammatories, antipsychotics, antineurotics, anxiolytics, and hematopoietic agents. In addition, embodiments of the invention contemplate the use of any compound for palliative care of patients suffering from pain or any other side effect in the course of treatment with a subject therapy. Exemplary palliative agents include acetaminophen, ibuprofen, other

NSAIDs, H2 blockers, and antacids. In an embodiment, the disclosure provides a method of treatment with agents that inhibit GAK and agents that inhibit AAK l. In an embodiment, such co-treatment provides synergistic effects, such as antiviral effects.

Hosts Suitable for Treatment

Hosts suitable for treatment with an embodiment of the inhibiting agent or an embodiment of the method include hosts who are infected with viral infection from clathrin AP binding viruses, Flaviviridae, Lentiviridae, HCV, HIV, co-infections, such as HCV/HIV co-infections, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV. As used herein, the term Flaviviridae includes any member of the family Flaviviridae, including, but not limited to, Dengue virus, including Dengue virus 1, Dengue virus 2, Dengue virus 3, Dengue virus 4 (see, e.g., GenBank Accession Nos. M23027, M l9197, A34774, and Ml4931); Yellow Fever Virus; West Nile Virus; Japanese Encephalitis Virus; St. Louis Encephalitis Virus; Bovine Viral Diarrhea Virus (BVDV); and Hepatitis C Virus (HCV); and any serotype, strain, genotype, subtype, quasispecies, or isolate of any of the foregoing. Where the Flaviviridae is HCV, the HCV is any of a number of genotypes, subtypes, or quasispecies, including, e.g., genotype 1, including la and lb, 2, 3, 4, 6, etc. and subtypes (e.g., 2a, 2b, 3a, 4a, 4c, etc.), and quasispecies.

Hosts suitable for treatment with embodiments of the present invention include treatment failure patients. The term "treatment failure patients" (or "treatment failures") as used herein generally refers to BCV-infected patients who failed to respond to previous therapy for HCV (referred to as "non-responders") or who initially responded to previous therapy, but in whom the therapeutic response was not maintained (referred to as "relapsers"). The previous therapy generally can include treatment with any antiviral agent other than an inhibiting agent of the present disclosure.

Hosts suitable for treatment with embodiments of the present disclosure include individuals who have been clinically diagnosed as infected with HCV. Individuals who are infected with HCV can be identified by detecting HCV RNA in their blood, and/or having an anti-HCV antibody in their serum.

Individuals who are clinically diagnosed as infected with HCV include naive individuals (e.g., individuals not previously treated for HCV).

Hosts suitable for treatment with embodiments of the present disclosure include individuals who have any detectable HCV titer. The patient may be infected with any

BCV genotype (genotype 1, including la and lb, 2, 3, 4, 6, etc. and subtypes (e.g., 2a, 2b, 3a, etc.)), particularly a difficult to treat genotype such as BCV genotype 1 and particular BCV subtypes and quasispecies.

Also suitable for treatment are BCV-positive hosts (as described above) who exhibit severe fibrosis or early cirrhosis (non-decompensated, Child's-Pugh class A or less), or more advanced cirrhosis (decompensated, Child's-Pugh class B or C) due to chronic BCV infection and who are viremic despite prior antiviral treatment, or who have a contraindication to therapy with a known antiviral agent.

In an embodiment, BCV-positive hosts with stage 3 or 4 liver fibrosis according to the METAVIR scoring system, which is known in the art, are suitable for treatment with the methods of the present disclosure. In another embodiment, hosts suitable for treatment with embodiments of the present disclosure are patients with decompensated cirrhosis with clinical manifestations, including patients with far-advanced liver cirrhosis, including those awaiting liver transplantation. In still another embodiment, hosts suitable for treatment with embodiments of the present disclosure include patients with milder degrees of fibrosis including those with early fibrosis (stages 1 and 2 in the METAVIR, Ludwig, and Scheuer scoring systems; or stages 1, 2, or 3 in the Ishak scoring system, all of which are known in the art). In an embodiment, the method finds use in treatment of BCV post liver transplantation for BCV induced hepatocellular carcmoma. In an embodiment, this serves to prevent re-infection of the graft.

In an embodiment of the present disclosure, to help optimally select patients most likely to benefit from therapy, as well as to monitor efficacy of therapy--especially in the face of potential drug resistant mutant viruses--the use of appropriate diagnostic tests provided by the present invention can be of great benefit. For example, assessing the sensitivity of the specific virus found in a given patient to the contemplated therapy can help identify the best match between candidate patient and the corresponding appropriate therapy.

In an embodiment, the method provides treating patients infected with viruses as described herein, particularly those infected with BCV who also are afflicted with cancer, such as hepatic cancer.

Assays

The present disclosure also provides in vitro and cell-based methods of screening for antiviral agents. In one embodiment, the disclosure provides methods for detecting

interaction between the YXX<l> motif and host proteins. In one embodiment, the present disclosure provides a binding assay to detect proteins or agents that inhibit binding of YXX<l> or dileucine motifs to AP2Ml or other µ subunits of clathrin AP complexes. For instance, the disclosure provides a method in which candidate agents as described herein are contacted with a Flaviviridae and AP2Ml or other µ subunits of clathrin AP complexes. Binding of the Flaviviridae to the proteins is detected as is known in the art. Reduced activity in the presence of a candidate agent relative to controls indicates identification of a binding inhibitor. In another embodiment, cell-based binding assays are used. In another embodiment, cell assays are used to screen for the effects on viral assembly or budding. In this embodiment, candidate agents are contacted with a Flaviviridae and a cell expressing AP2M l or other µ subunits of clathrin AP complexes. Cellular effects, such as viral budding or assembly, or binding between the Flaviviridae and cell can be assayed by methods known in the art. In one embodiment, a replication assay is used. An example of a replication assay is outlined in U.S. patent application publication 2011/0052536 A l, which is expressly incorporated herein by reference. Reduced binding, budding, or assembly and the like, of the virus in the presence of the candidate agent relative to controls indicates identification of a binding inhibitor or antiviral agent. In some embodiments, in vitro microfluidics affinity assay, as known in the art, is also used to enable detection of weak and transient protein interactions.

In one embodiment, protein-fragment complementation assays, as described herein and as are known in the art, are used to validate interaction between core and AP2Ml. In another embodiment, a co-immunprecipitation assay is used to identify interaction between the proteins.

EXAMPLES

The following examples are included to demonstrate preferred embodiments of the invention and to set forth a clear understanding of the principles of the disclosure. It should be appreciated by those of skill in the art that the techniques disclosed in the examples which follow represent techniques discovered by the inventors to function well in the practice of the invention, and thus can be considered to constitute preferred modes for its practice. However, those of skill in the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or similar result without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents

which are both chemically and physiologically related may be substituted for the agents described herein while the same or similar results would be achieved. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention as defined by the appended claims. .

Many variations and modifications may be made to the above-described embodiments of the disclosure without departing substantially from the spirit and principles of the disclosure. All such modifications and variations are intended to be included herein within the scope of this disclosure.

Example 1

Identification of a YXX<I> motif within HCV core

Inspection of the primary sequence of the HCV core protein reveals a conserved YIP(V/L) motif within the second domain (D2) of the protein (FIGS. lA, D). This motif conforms to the YXX<l> sorting signal consensus recognized by AP2Ml (Ohno, *J Cell Sci* 119:3719-3721, 2006; Nakatsu *et al*., *Cell Struct Funct* 28:419-429, 2003; Owen *et al*., *Ann Rev Cell Devel Biol* 20:153-191, 2004).

Core binds AP2Ml

Interactions of sorting signals with clathrin adaptors and endocytic components are typically weak (Kd of binding at a µM range), transient (Nakatsu *et al*., *Cell Struct Funct* 28:419-429, 2003; Aguilar *et al., J Biol Chem* 276:13145-13152, 2001), and involve membrane proteins, thus difficult to study by standard technologies (Cusick *et al*., *Hum Mal Genet* 2005; Bailer *Curr Opin Microbial* 12:453-459, 2009). To determine whether HCV core binds AP2Ml, proteomic platforms that overcome these challenges were used. *In vitro* microfluidics affinity assays are based on mechanical trapping of molecular interactions (MITOMI), which eliminates the off-rate problem facing current platforms, and thus allows studying weak and transient interactions, with nanoliter protein consumption (Maerkl *et al., Science* 315:233-237, 2007; Einav *et al* ., *Nat Biotech* 26:1019-1027, 2008; Gerber *et al., Nat Meth* 6:71-74, 2009). A microfluidics format that enables a high fidelity analysis of protein-protein interactions (P-Pis) was used (Gerber *et al., Nat Meth* 6:71-74, 2009). *In vitro* protein expression in the presence of microsomal membranes and binding experiments with MITOMI were performed essentially as described (Maerkl *et al*., *Science* 315:233-237, 2007; Einav *et al., Nat Biotech* 26:1019-1027, 2008; Gerber *et al* ., *Nat Meth* 6:71-74, 2009) (FIG. 9). These assays detected binding of AP2Ml to core. The degree of binding correlated with increasing core

concentration (FIG. lF). Background binding of AP2M l to a control HCV protein, NS3, was 4-20 fold lower and did not increase significantly with increased protein concentration.

To determine whether the core-AP2M l interaction occurs in cells, protein-fragment complementation assays (PCAs) based on reversible reconstitution of a *Gaussia* princeps luciferase reporter was used (FIG. 2A). The current format was optimized for improved signal, thus providing a highly sensitive means for measuring challenging P-Pls (Cassonnet *et al., Nat Meth* 8:990-992, 2011). Significant luciferase signal was detected in Huh-7.5 cells cotransfected with plasmids encoding the two reporter fragments fused to the prey and bait proteins (GLucl-AP2Ml and GLuc2-core). Background levels of binding were detected in cells cotransfected with either the GLucl-AP2Ml or GLuc2-core constructs and the empty reciprocal vector, the two empty GLuc vectors, or GLucl fused to three unrelated proteins (SPIRE, RACl, and ARPC). The apparent affinity of AP2Ml to core was higher than to transferrin receptor (TFR), a host cargo protein harboring a YXX<l> signal, known to be recognized by AP2Ml (Gminard *et al., Traffic* 5, 181-193, 2004) (FIG. 2B). Binding was not genotype-specific, as core proteins derived from either the 2a (Lindenbach *et al., Science* 309:623-626, 2005) or lb (Lohmann *et al*., *Science* 285: 110-113, 1999) genotypes demonstrated comparable levels of AP2Ml binding. Furthermore, there were no significant differences in core binding to the two isoforms of AP2M l(a/b) (FIG. 2B). Comparable results were demonstrated in Huh-7.5 (human hepatoma derived) cells, representing the most relevant cell model (FIG. 2B), and 293T cells (data not shown). Binding appeared specific, as increasing concentrations of free core or AP2M l, but not nuclear export signal-interacting protein (NESI), a control protein involved in mediating hepatitis D virus assembly (Wang *et al., J Virol* 79:8113-8120, 2005), progressively decreased core-AP2M l binding (FIG. 2C).

To determine whether core binds AP2Ml in the context of authentic HCV infection, co-immunoprecipitation assays in membrane fractions prepared from Huh-7.5 cells infected with cell culture-grown HCV (J6/JFH) were performed (Lindenbach *et al., Science* 309:623-626, 2005). AP2Ml could bring down core when anti-AP2Ml antibodies but not lgG controls were added to the membrane lysates. Binding was significantly augmented by calyculin A [an inhibitor of AP2Ml dephosphorylation, which "locks" AP2Ml in its YXX<l> binding active conformation (Ricotta *et al., J Biol Chem* 283:5510-5517, 2008)] (FIG. 2D). Binding in reciprocal conditions was similarly demonstrated (FIG. 2D). Colocalization of core with AP2Ml in Huh-7.5 cells 72 hr

following electroporation with J6/JFH HCV RNA was also investigated. Quantitative confocal immunofluorescence (IF) analysis revealed extensive colocalization of core and AP2Ml in these cells (with 67±6% colocalization of AP2M l stained puncta with core) (FIG. 2E).

Core's YXX<I> motif in AP2Ml binding and HCV assembly

Using a series of point mutations (FIG. lD), whether AP2Ml binding is mediated by core's YXX<l> motif was tested. A Y136A core mutation reduced AP2Ml binding measured by PCAs by -10 fold compared with wild type (WT) core, whereas a V(<l>)139A mutation caused less inhibition of binding (FIG. 3A). To study the role of core's YXX<l> motif in HCV infection, these mutations were introduced into the J6/JFH(p7-Rluc2A) HCV genome - a *Renilla* luciferase-containing reporter virus that replicates and produces high titers of virus in Huh-7.5 cells (Murray *et al., J Viral* 81:10220-10231, 2007). Cells were electroporated with *in vitro* transcribed RNA generated from each construct. HCV RNA replication of these viral mutants was comparable to that of the WT virus, as measured by luciferase reporter gene-linked assays (FIG. 3B), and qRT-PCR (FIG. 11 and 12). In contrast, a polymerase-defective mutant, J6/JFH(p7-Rluc2A)-GNN, did not replicate. Luciferase assays in naive cells inoculated with supernatants derived from cells electroporated with viral genome harboring the Y136A core mutation measured undetectable levels of extracellular infectivity. The V139A mutation decreased infectivity by -1.5 logs compared to WT virus (FIG. 3C). Intracellular infectivity, measured in naive cells infected with clarified supernatants derived from lysed electroporated cells, mirrored the diminished extracellular infectivity (FIG. 3D), suggesting that core's YXX<l> motif mediates virions assembly and not release. Essentially no infectious virus was produced either intra- or extracellularly by assembly (El-E2) or replication (GNN) defective controls. lnfectivity titers of WT virus measured by limiting dilution assays were comparable to those previously reported with this reporter system (Kopp *et al., J Viral* 84:1666-1673, 2010). Consistent with the luciferase assays described above, while the V139A core mutation decreased the extra- and intracellular infectivity titers by a -1-1.5 log compared with WT virus, an undetectable level of infectivity titers was measured with virus harboring the Y136A core mutation or El-E2 deletion (FIG. 3E). The effect of core mutations on infectivity correlated with their effect on AP2M l binding. To exclude the possibility that core's mutations affected infectivity by causing particle disassembly or production of defective core protein- and RNA-containing particles, production of noninfectious particles was determined.

Detectable levels of HCV RNA and core protein release were measured in supernatants of cells harboring replicating genomes by qRT-PCR and ELISA assays, respectively, as described (Murray *et al ., J Viral* 81:10220-10231, 2007; Kopp *J Viral* 84:1666-1673, 2010) (FIG. 3F, 3G). Nevertheless, the levels released by the Y136A and V139A core mutants were not significantly higher than those released by the assembly-defective El• E2 mutant, suggesting that noninfectious particles were not produced. Core expression was not affected by the mutations, as western analysis (FIG. 3H) and fluorescence microscopy (data is not shown) demonstrated protein expression at WT levels. Reversion of the infectivity phenotype was detected by luciferase assays in Huh-7.5 cells infected with HCV harboring the Y136A and V139A core mutations following two weeks of passaging. This reversion coincided with the emergence of primary-site revertants by sequencing analysis. These results provide additional evidence for the requirement of maintaining a functional YXX<l> motif for supporting HCV replication. Together, these data suggest that the AP2M 1 binding motif within core is required for viral assembly *in vitro.*

Core's YXX<l> motif is functionally interchangeable with other YXX<l> sorting signals

To determine whether core's YXX<l> motif is functionally interchangeable with homologous signals, a V139L mutation, thus "swapping" the genotype 2a core's sequence with that of genotype 1b was introduced. Similarly, the YTPL and YRRL sequences, known to mediate binding of HLA-DM (Ohno *et al ., J Bioll Chem* 273:25915-25921, 1998) and thrombopoietin receptor (Hitchcock *et al ., Blood* 112:2222-2231, 2008) to AP2Ml, respectively, were used to substitute the core's YXX<l> sequence (FIG. lD). Binding of core harboring these sequences to AP2M 1 was either comparable to or greater than that of WT core, as determined by PCAs (FIG. 3A). These mutations had no effect on HCV RNA replication (FIG. 3B). In correlation with their biochemical phenotype, the intracellular and extracellular infectivity of the V139L and YTPL core mutants were comparable to that of WT virus (FIG. 3C, 3D). This functional interchangeability supports that core's YXX<l> motifs exerts its function via interactions with host cell proteins. Despite its efficient binding to AP2M l (FIG. 3A) and stability by western analysis (FIG. 3H), the YRRL mutant did not produce detectable levels of infectious virus (FIG. 3C, 3D). Interestingly, this mutant released HCV RNA and core protein into supernatants of electroporated cells at levels comparable to that of WT core, likely reflecting production of noninfectious particles (FIG. 3F, 3G). The YRRL mutant may

thus impact other function of core in infectious virus production that is independent of its binding to AP2M 1.

AP2Ml in HCV assembly

The functional relevance of AP2Ml to the HCV life cycle was determined. Stable Huh-7.5 clones harboring short hairpin RNA (shRNA) lentiviral constructs targeting distinct regions in the AP2Ml gene or a non-targeting (NT) sequence were established. Effective suppression of AP2Ml levels was achieved (FIG. 4A, 4B), without apparent cytostatic or cytotoxic effects. The effect of AP2M l depletion on infectious virus production was studied in these clones following electroporation with J6/JFH(p7-Rluc2A) RNA. AP2Ml knockdown had no effect on HCV RNA replication as measured in these stable clones by luciferase assays (FIG. 4C) and qRT-PCR 72 hr following electroporation (data not shown). Supematants collected at 72 hr postelectroporation were used to inoculate naive Huh-7 .5 cells followed by luciferase assays at 72 hr postinoculation. As shown in FIG. 4D, AP2M l depletion reduced extracellular infectivity by >2 logs compared with NT control. Intracellular infectivity, measured in naive cells infected with clarified supematants derived from lysed electroporated cells, mirrored the diminished extracellular infectivity(FIG. 4E) and correlated with the degree of AP2M l depletion. Measurements of intra- or extracellular infectivity titers by limiting dilution assays demonstrated consistent results (FIG. 4F). AP2Ml depletion did not increase production of noninfectious particles, as suggested by the levels of HCV RNA and core protein release measured in supematants of cells by qRT-PCR and ELISA assays, respectively (FIG. 4G, 4H). Similar effects on infectious virus production were demonstrated in Huh-7.5 cells transiently depleted for AP2M l by siRNAs and either electroporated with the J6/JFH(p7-Rluc2A) RNA or infected with culture grown J6/JFH virus (titer: $1.2x10^5$ $TCID_{50}$/ml) (Lindenbach *et al., Science* 309:623-626, 2005). The effects of silencing endogenous AP2Ml on infectious virus production were rescued by ectopic expression of shRNA-resistant WT AP2M l harboring a wobble mutation within the site targeted by the shRNA, largely excluding the possibility of off-target effects causing the observed phenotype (FIG. 41). The stable and transient RNAi approaches thus both suggest that AP2Ml is important for efficient HCV assembly.

Disruption of core-AP2Ml binding abolishes recruitment of AP2Ml to LD, alters the sub-cellular localization of core, and core colocalization with E2

To test the hypothesis that the defect in HCV assembly resulting from YXX<l> core mutations or AP2Ml silencing correlates with alterations in the sub-cellular localization

of AP2Ml and/or core, a quantitative confocal immunofluorescence (IF) analysis was performed. 10-15 randomly chosen cells were analyzed for each category for the degree of localization of core or AP2M l to various intracellular compartments using ImageJ (JACoP) software and Manders' Colocalization Coefficients (MCC). The latter were chosen, as they strictly measure co-occurrence independent of signal proportionality (Dunn *et al ., Am J Physiol-Cell Physiol* 300:C723-C742, 2011). Endogenous AP2Ml minimally colocalized with the LD marker, Bodipy, in naive Huh-7.5 cells, with 8.2% of LD staining positive for AP2M l (FIG. 5A, 5E). In contrast, infection with J6/JFH virus appeared to significantly increase the localization of AP2Ml to LD, with 40±8% of LD being AP2Ml positive (FIG. 5B, 5E) (p-value=0.0006). Similarly, and as previously described (Kopp *et al ., J. Viral.* 84:1666-1673, 2010; Boulant *et al ., Journal of General Virology* 88:2204-2213, 2007; Coller *et al., PLoS pathogens* 8:e1002466-e1002466, 2012), core was partially localized to LD (37±14% of core positive LD) (FIG. 5C, 5E). Furthermore, the partial colocalization of core with AP2Ml occurred in part on LD (FIG. 5D, 5F).

To test whether core is involved in mediating the increased localization of AP2M l to LD measured in HCV infected cells, AP2Ml-mCherry was overexpressed either alone or in combination with WT core or Y136A core mutant by transfecting Huh-7.5 cells. LD were labeled with HCS LipidTOX (lnvitrogen). Similarly to naive cells, AP2M l was minimally localized to LD in Huh-7.5 cells overexpressing AP2Ml alone (8±2% of LD positive for AP2M l) (FIG. 5G, 5J). In contrast, as in infected cells, when co-expressed with WT core, AP2M l appeared to significantly accumulate at LD, with 51.8±20% of LD being AP2M l positive (p-value=4.8x10-^{5}) (FIG. 5H, 5J). No such increase in colocalization was demonstrated, however, when AP2M l was co-expressed with core harboring the Y136A mutation (with 10% AP2Ml positive LD) (p-value=0.001) (FIG. 51, 5J), suggesting that core's YXX<l> motif may mediate recruitment of AP2M l to LD.

The effect of disruption of the core-AP2M l interaction on core localization to LD, ER, and TGN and its colocalization with the E2 envelope protein in cells electroporated with J6/JFH HCV RNA was tested. As previously shown, core localized to all these intracellular compartments (Kopp *et al ., J. Viral.* 84, 1666-1673, 2010; Boulant *et al., Journal of General Virology* 88, 2204-2213, 2007) (with percent colocalization ranging from 30 to 45%). Interestingly, localization of core harboring the Y136A mutation to LD was significantly greater than that of WT core (78±13% vs. 32±16%, respectively, p• value=2.7x10-6) (FIG. 5K). Similarly, the percent colocalization of core with the LD

marker was dramatically increased from 25±10% in NT cells to 87±6.6% following silencing of AP2M l (p-value= l.55x10-^{8}) (FIG. 5L). While neither the Y l36A core mutation nor AP2M 1 depletion had an apparent effect on core colocalization with the ER marker, calreticulin (Figure 14), both were associated with a significant decrease in core colocalization with the TGN marker, TGN46 (FIG. 5M, 5N) and the E2 envelope protein (FIG. 50, 5P).

Together, these results suggest that core's interaction with AP2M l facilitates recruitment of AP2M l to LD and that the observed defect in HCV assembly following disruption of the core-AP2M l interaction is associated with accumulation of core on LD, decreased core colocalization with E2, and impaired core trafficking to TGN.

AAKl and GAK regulate core-AP2Ml binding and are involved in HCV assembly

Phosphorylation of T156 within AP2Ml by the serine/threonine kinases AAK l and GAK (Ricotta *et al*., *Journal of Cell Biology* 156, 791-795, 2002; Korolchuk *et al*., *Traffic* 3, 428-439, 2002; Zhang *et al., Traffic* 6, 1103-1113, 2005) stimulates binding of AP2Ml to cargo protein tyrosine signals and is transient due to dephosphorylation by PP2A (Ricotta *et al*., *Journal of Biological Chemistry* 283, 5510-5517, 2008) (FIG. 6A). Indeed, calyculin A (a PP2A inhibitor) augmented the binding of core to AP2M l (FIG. 2D) and a T156A AP2Ml mutation impaired it (FIG. 6B). To study the effect of overexpression of AP2M l harboring the T156A mutation on infectious HCV production, Huh-7.5 cells were transfected with plasmids encoding either WT or AP2Ml T156A mutant. 48 hr posttransfection cells were electroporated with J6/JFH(p7-Rluc2A) HCV RNA and subjected to HCV RNA replication and infectivity assays, as described above. Overexpression of the T156A AP2Ml mutant had no effect on cellular viability and was dispensable for HCV RNA replication, yet significantly reduced extra-and intracellular infectivity compared with WT AP2Ml (FIG. 6C-E), consistent with a dominant negative effect on HCV assembly.

To study the hypothesis that AAK l and GAK are involved in regulating the interaction of AP2M l with core, binding experiments were performed in Huh-7.5 cells depleted for AAK l or GAK by siRNAs (FIG. 6F, 6G). Depletion of either AAK l or GAK significantly decreased core-AP2M l binding compared to NT, as measured by PCAs (FIG. 6H), with no apparent cytotoxic effect (data not shown). AAK l and GAK depleted cells were then electroporated with J6/JFH(p7-Rluc2A) HCV RNA. While depletion of either AAK l or GAK had no cytotoxic effect and was dispensable for HCV RNA replication, it significantly reduced intracellular and extracellular infectivity (FIG. 61-K).

The effect of AAK l and GAK on HCV assembly did not result from their direct binding to core (FIG. 6L). These results provide evidence for the involvement of AAK l and GAK in the regulation of core-AP2M l binding and in mediating HCV assembly. Moreover, they validate the importance of AP2Ml for efficient HCV assembly by a dominant-interfering approach, thus supporting the RNAi data.

Pharmacological inhibitors of AAKl and GAK disrupt core-AP2Ml binding and HCV assembly

Analysis of heat maps and affinity assays of kinase inhibitors (Karaman *et al., Nat Biotech* 26, 127-132, 2008) revealed compounds, such as sunitinib and PKC-412, which bind AAK l, and erlotinib which binds GAK, with high affinities (nM range) (Karaman *et al., Nat Biotech* 26, 127-132, 2008) (FIG. 7A, 7B). These compounds inhibited core• AP2Ml binding in a dose-dependent manner, as determined by PCAs (FIG. 7C), with half maximal inhibitory concentrations (IC50s) of -0.04-0.2µM. When used to treat Huh-7.5 cells electroporated with the J6/JFH(p7-Rluc2A) HCV genome, these compounds had a dramatic dose-dependent effect on extra- (FIG. 7D) and intracellular infectivity (FIG. 7E) at 72 hr (with half maximal effective concentration (EC50s) of 0.15-1.8µM) (FIG. 7B). There was no effect on HCV RNA replication and no apparent cellular toxicity (FIG. 7F, 7G). Indeed, the inhibitory effect on core-AP2M l binding and infectivity was associated with reductions in phospho-AP2M l levels in the relevant cells by western analysis (FIG. 7H,). Last, these compounds significantly inhibited viral infection in Huh-7.5 cells infected with tissue culture grown HCV (titer: $6.3x10^5$ $TCID_{50}$/ml) (FIG. 71, 7B,). These results provide pharmacological validation for the involvement of AAK l and GAK in regulating the core-AP2M l interaction and for AP2M l's role in HCV assembly. Furthermore, they provide candidate compounds targeting assembly.

Example 2

AAKl and GAK depletion or pharmacological inhibition abrogate HIV-1 replication

There is a substantial body of evidence to support that interactions of APlM l and AP2Ml with HIV Gag and Env (gp41) mediate several critical steps along the assembly/release pathway (Batonick *et al., Virology* 342: 190-200, 2005; Camus *et al., Malec Biol Cell* 18:3193-3203, 2007; Berlioz-Torrent *et al., J Virol* 73:1350-1361, 1999; Byland *et al., Malec Biol Cell* 18:414-425, 2007; Wyss *et al., J Viral* 75:2982-2992,

2001; Ohno *et al*., *Virology* 238:305-315, 1997; Boge *et al.*, *J Biol Chem* 273:15773-15778, 1998; Egan *et al*., *J Virol* 70:6547-6556, 1996; Rowell *et al.*, *J Immunol* 155:473-488, 1995; Lodge *et al*., *EMBO J* 16:695-705, 1997; Deschambeault *et al*., *J Viral* 73:5010-5017, 1999). Most importantly, a tyrosine motif within Env mediates cell-to-cell spread (Gminard *et al*., *Traffic* 5, 181-193, 2004; Lindenbach *et al*., *Science* 309, 623-626, 2005), a mechanism thought to account for ongoing HIV replication despite ART (Sigal *et al.*, *Nature* 477, 95-98, 2011), possibly via APlM l which sorts to basolateral membranes. However, the role of AAK l and GAK in HIV infection has not been studied, and these mechanisms have not been targeted pharmacologically.

To test the hypothesis that AAK l and GAK are important for HIV infection, HeLa-derived TZM-b l cells were used, which express CD4, CCR5 and CXCR4, as well as -galactosidase and firefly luciferase reporter genes responsive to HIV transcription (Wei *et al.*, *Antimicrobial agents and chemotherapy* 46:1896-1905, 2002). Cells were transfected with siRNAs targeting AAK l, GAK or a NT sequence followed by infection with the infectious HIV-1 NL4-3 clone (Adachi *et al.*, *J Viral* 59:284-291, 1986), as described (Zhou *et al.*, *Cell Host & Microbe* 4:495-504, 2008). Luciferase activity was measured at 96 hr postinfection. AAK l and GAK depletion significantly inhibited HIV replication (Fig. 10). To test the effect of pharmacological inhibitors of AAK l and GAK against HIV, HIV-1 infected HeLa-derived TZM-b l cells were treated daily with serial dilutions of sunitinib, erlotinib or PKC-412 for 4 days followed by luciferase assays. A significant dose response antiviral effect was observed with no effect on viability (Fig. 10). EC_{50} of erlotininb was 1.4±0.3(P value - 0.0058), sunitinib - 0.8±0.4 (P value - 0.1), and PKC-412 - 8.5±4 (P value - 0.1). Since these studies were conducted 96 hr postinfection, they assessed all stages of infection from entry to release and spread (Zhou *et al.*, *Cell Host & Microbe* 4:495-504, 2008), and therefore suggest that AAK l and GAK are important for overall HIV replication.

Methods

Plasmids. ORFs encoding AP2M l, GAK, AAK l, TFR, SPIRE, RACl, ARPC, and NESI were picked from the Human ORFeome library of cDNA clones (Rual *et al.*, *Genome research* 14, 2128-2135, 2004) (Open biosystems) and recombined into either pcDNA-Dest40 (for C-terminal V5-his tagging), pGLuc (for *Gaussia Princeps* luciferase fragment (Gluc) tagging) (Cassonnet *et al*., *Nat Methods* 8, 990-992, 2011), and/or pCherry (for mCherry fluorescence protein tagging) vectors using gateway technology

(Invitrogen). ORFs encoding T7-tagged full length core and NS3 were amplified from described vectors (Blight *et al., Science* 290, 1972-1974, 2000) and ligated into pcDNA3.1 (Invitrogen). pFL-J6/JFH(p7-Rluc2A) (Murray *et al., J Viral* 81, 10220-10231, 2007) was a gift from Dr. C. M. Rice (Tscherne *et al., J Viral* 80, 1734-1741, 2006). The YXX<l> core mutations and AP2Ml mutations were introduced into these plasmids by site-directed mutagenesis (using the QuikChange kit (Stratagene)).

Antibodies and compounds. See Tables 1 and 2.

RNAi and rescue of gene silencing. 40-100 nM individual or pooled siRNAs (FIG. 8) were transfected into cells using silMPORTER (Upstate, Millipore) 48 hr prior to HCV RNA electroporation. Five individual MISSION Lentiviral Transduction Particles (Sigma) harboring short hairpin RNAs (shRNAs) targeting various sites in the AP2Ml RNA and a control shRNA were used to transduce Huh-7.5 cells according to the manufacturer's protocol. RNAi reagents used in this study are summarized in Table 3. AP2Ml rescue was performed by transduction of Huh-7.5 cells stably depleted for AP2Ml with lentiviruses expressing shRNA-resistant AP2Ml 48 hr prior to electroporation with HCV genome.

Microfluidics affinity assays. Device fabrication and design were done essentially as described (Maerkl *et al., Science* 315, 233-237, 2007). VS-his-tagged human proteins and T7-tagged viral proteins were expressed off the chip by mammalian *in vitro* transcription/translation mixture (TNT) (Promega) (in the presence of microsomal membranes). The device was subjected to surface patterning, resulting in a circular area coated with biotinylated anti-his antibodies within each unit cell (Einav *et al., Nature Biotechnolog y* 26, 1019-1027, 2008; Gerber *et al., Nature methods* 6, 71-74, 2009). Protein-protein binding experiments were performed as described (Gerber *et al., Nature methods* 6, 71-74, 2009). Briefly, human bait proteins were loaded into the device and bound to the surface anti-his antibodies. Viral and human proteins were incubated in the chip and labeled with anti-T7-Cy3 and anti-V5-FITC antibodies, respectively. Interactions were trapped mechanically by MITOMI (Maerkl *et al., Science* 315, 233-237, 2007; Einav *et al., Nature Biotechnolog y* 26, 1019-1027, 2008; Gerber *et al., Nature methods* 6, 71-74, 2009). After a brief wash to remove untrapped unbound material, the trapped protein complexes were detected by an array scanner (Tecan). The ratio of bound viral prey to expressed human bait protein was calculated for each data point by measunng the ratio of median signal of Cy3 to median signal of FITC. Protein concentration in lysates were determined by quantitative western analysis against

standard curves of T7-tagged proteins. Experiments were conducted at least three times, each time with >20 replicates. See FIG. 10 for a detailed protocol.

Cell cultures. Huh-7.5 cells and 293T cells were maintained m Dulbecco's modified minimal essential medium (Gibco) supplemented with 1% L-glutamine (Gibco), 1% penicillin, 1% streptomycin (Gibco), IX nonessential amino acids (Gibco), and 10% fetal bovine serum (Omega Scientific). Cell lines were passaged three times a week after treatment with 0.05% trypsin-0.02% EDTA and seeding at a dilution of 1:4.

Protein-fragment complementation assays. Binding assays were performed essentially as described (Cassonnet *et al*., *Nat Methods.* 8, 990-992, 2011). Combinations of plasmids encoding prey (A) and bait (B) proteins, each fused to a fragment of the *Gaussia Princeps* luciferase protein (Glucl and Gluc2) or control vectors were cotransfected into 293T or Huh-7 .5 cells plated in 96-well plates in triplicates. At 24 hr posttransfection, cells were lysed and subjected to standard luciferase reporter gene assays using *Renilla* luciferase assays system (Promega). Results were expressed either as luminescence or luminescence ratio. The latter represents the average luminescence signal detected in cells transfected with Glucl-A and Gluc2-B divided by the average of luminescence measured in control wells transfected with Glucl-A and an empty Gluc2 vector with those transfected with Gluc2-B and an empty Glucl vector. Competition assays and studies designed to determine inhibitory effect of compounds or siRNAs were performed as above, except that binding was measured in the presence of excess free proteins, the inhibitors, or siRNAs, respectively. Experiments were conducted at least three times in triplicates.

Co-immunoprecipitations. Membranes were prepared from -20x10^6 Huh-7.5 cells infected with HCV (J6/JFH) (Lindenbach, *et al., Science* 309, 623-626, 2005), as previously described (Einav *et al* ., *J Viral* 78, 11288-11295, 2004). Briefly, cells were collected by trypsinization, washed once with PBS and resuspended in HME buffer (20 mM HEPES [pH 7.4], 1 mM EDTA, 2 mM MgC12), supplemented with phenylmethylsulfonyl fluoride to a final concentration of 1 mM and a protease inhibitors cocktail (Sigma). Cells were lysed by two cycles of freeze-thaw in dry ice-ethanol and then passaged through a 27.5-gauge needle 10 times. Nuclei were removed by centrifugation at 250 xg for 10 min, and the postnuclear supernatant was subjected to ultracentrifugation at 100,000 xg for 30 min to obtain the membrane preparation. All steps were done at 4°C. Total membrane proteins were diluted in lml TDB buffer (2.5% Triton X-100, 25 mM triethanolamine-Cl [pH 8.6], 20 mM NaCl, 5mM EDTA, 0.2%

NaN3) (Einav *et al ., J Viral* 78, 11288-11295, 2004) and incubated for 30 min at 37°C with 100 nM calyculin-A or DMSO. Due to the weak and transient nature of the interactions, 25 mM dithiobis-succinimidyl-propionate (DSP) cross-linker (Pierce) was added to allow covalent binding of the already bound interacting proteins. Samples were incubated for 2 hr on ice. 1 M Tris was added to stop the DSP activity. Lysates were then clarified by 10 min centrifugation at 1OOOxg, followed by 30 min ultracentrifugation of the supernatants at 100,000xg. Membrane pellets were resuspended in 100 1 HME buffer (20mM NaHEpes (PH 7.4), lmM EDTA (Ph 8), 2mM MgC12), and TDB buffer was added for a final volume of 1 ml. Samples were incubated overnight with either anti-AP2Ml antibodies, anti-core antibodies or IgG controls, and protein G magnetic beads (Millipore). Immunoprecipitates were analyzed by western blotting. Experiments were conducted twice in duplicates.

In vitro transcription of viral RNA and transfection. HCV RNA was generated and delivered into Huh-7.5 cells, as previously described (Lindenbach *et al., Science* 309, 623-626, 2005; Murray *et al., J Viral* 81, 10220-10231, 2007). Briefly, RNA was synthesized from Xbal linearized J6/JFH(p7-Rluc2A) template using the T7 MEGAscript kit according to the manufacturer's protocol (Ambion). Reaction mixtures were incubated for 3 hr at 37°C and then subjected to DNase treatment for 15 min at 37°C. Viral RNA was purified using the RNeasy kit (Qiagen). RNA was quantified by absorbance at 260 nm, and its quality was verified by agarose gel electrophoresis. Subconfluent Huh-7.5 cells were trypsinized and collected by centrifugation at 700g for 5 min. The cells were then washed three times in ice-cold RNase-free PBS (BioWhittaker) and resuspended at $1.5*10^7$ cells/ml in PBS. 5 µg of the *in vitro* transcribed wild type or J6/JFH(p7-Rluc2A) mutant RNA was mixed with 400 µl of washed Huh-7.5 cells in a 2mm-gap cuvette (BTX) and immediately pulsed (0.82 kV, five 99 µs pulses) with a BTX-830 electroporator. After a 15 min recovery at 25°C, cells were diluted in 30ml of prewarmed growth medium and plated into 96, 24, 6-well and PlOO tissue culture plates.

HCV RNA replication by luciferase assays. HCV RNA replication was measured at 6-9 hr, and 72 hr postelectroporation, as described (Murray *et al., J Viral* 81, 10220-10231, 2007). Electroporated cells plated in quadruplicates in 96-well plates were washed twice with PBS and lysed with 30µ1 of *Renilla* lysis buffer (Promega). Following 15 min of shaking at RT, luciferase activity was quantified using a *Renilla* luciferase substrate (Promega) and a Tecan luminometer (Tecan) according to the manufacturers' protocols.

HIV replication assays. HeLa-derived TZM-b l cells, which express CD4, CCR5 and CXCR4, as well as -galactosidase and firefly luciferase reporter genes responsive to HIV transcription (Wei *et al., Antimicrobial agents and chemotherapy* 46, 1896-1905, 2002) were used in these assays. Cells were transfected with siRNAs targeting AAK l, GAK or a NT sequence followed by infection with the infectious HIV-1 NL4-3 clone (Adachi *et al., J Viral* 59, 284-291, 1986), as described (Zhou *et al., Cell Host & Microbe* 4, 495-504, 2008). Luciferase activity was measured at 96 hr postinfection. To test the effect of pharmacological inhibitors of AAK l and GAK against HIV, HIV-1 infected HeLa-derived TZM-b l cells were treated daily with serial dilutions of sunitinib, erlotinib or PKC-412 for 4 days followed by luciferase assays.

Extracellular infectivity. Culture supernatants of Huh-7.5 cells electroporated with J6/JFH(p7-Rluc2A) RNA and plated in PlOO dishes were harvested at 72 hr postelectroporation, clarified (with a 0.22-µm-pore size filter) and used to infect naive Huh-7.5 cells for 72 hr in triplicates before lysis in *Renilla* lysis buffer (Promega). Luciferase activity in 20µ1 of cell lysates was quantified as described above. To determine the effect of erlotinib, sunitinib, and PKC-412 on infectious virus production, electroporated cells were grown in the presence of the inhibitors with daily medium changes for 72 hr prior to collection of supernatants or cell lysates. Results represent loglO RLU values per lOcm tissue culture dish. Experiments were repeated three times, each time with quadruplicates.

Intracellular infectivity assays. As described by Murray *et al.* (J *Viral* 81, 10220-10231, 2007), 72 hr postelectroporation with J6/JFH(p7-Rluc2A) RNA cells were trypsinized, collected by centrifugation, resuspended in 500µ1 medium, lysed by freeze• thaw cycles, and pelleted twice at 3,650xg. Clarified supernatants diluted in complete medium were used to inoculate naive Huh-7.5 cells in triplicates, followed by lysis and luciferase assays at 72 hr. Results represent log lO RLU values per lOcm tissue culture dish. Experiments were repeated three times, each time with quadruplicates.

Virus titration. Extracellular and Intracellular titers were determined by limiting dilution assays based on immunohistochemical staining for core. 50% tissue culture infectious dose ($TCID_{50}$) was calculated, as described (Lindenbach *et al., Science* 309, 623-626, 2005). Results are expressed as TCIDso /ml. Minimal titers measured with the El-E2 mutant were used for background subtraction.

Core ELISA. The concentration of released core protein was measured in clarified cell culture supernatants harvested at 72 hr postelectroporation by ELISA (Cell

Biolabs) against standard curves of recombinant core antigen, according to the manufacturer's instructions.

Viability assays. Following 24, 48, and 72 hrs of treatment with inhibitory compounds or silencing with siRNAs, cells were incubated for 2-4 hrs at 37°C in the presence of 10% AlamarBlue reagent (TREK Diagnostic Systems), as described (Einav *et al., Nature Biotechnolog y* 26, 1019-1027, 2008). Plates were then scanned and fluorescence was detected by using FLEXstation II 384 (Molecular Devices, Inc.).

Infection studies. $6x10^3$ Huh-7.5 cells seeded in 96-well plates were infected in triplicates with cell culture-grown HCV J6/JFH (titer: $1.2x10^5$ $TCID_{50}$/ml). Extracellular and intracellular infectivity were measured by focus formation assays (Lindenbach *et al ., Science* 309, 623-626, 2005) in naive Huh-7.5 cells infected with supematants or clarified cell lysates derived from the infected cells at 72 hr postinfection.

To determine the antiviral effect of erlotinib, sunitinib, and PKC-412, $6x10^3$ Huh-7.5 cells seeded in 96-well plates were infected in triplicates with cell culture-grown HCV (J6/JFH(p7-Rluc2A)) (titer: $6.3x10^5$ $TCIDs_0$/ml) with an MOI *(multiplicity of infection)* of 0.1. Six hours after infection and daily thereafter, cells were washed and medium was replaced with medium containing serial dilutions of the inhibitory compounds. At 72 hr, samples were subjected to viability assays, followed by standard luciferase assays (Promega).

Analysis of revertants. Huh-7.5 cells electroporated with J6/JFH(p7-Rluc2A) harboring the Y136A or V139A mutations were propagated. Culture supematants were harvested every few days and used to inoculate naive Huh-7.5 cells for determination of extracellular infectivity by luciferase assays 72 hr following inoculation. Total cellular RNA was extracted from clones demonstrating reversion of the infectivity phenotypone (at day 8 postelectroporation for Y136A clones and day 14 for V139A mutant clones) using TRizol reagent (lnvitrogen). Reverse transcription reaction and PCR amplification were performed using Superscript One-Step reverse transcriptase PCR (RT-PCR) kit (lnvitrogen). A -1-kb segment harboring the core coding sequence was amplified. The PCR products were purified from agarose gels by Ultra Clean 15 DNA purification kit (MoBio) and subjected to automatic sequencing on an ABI Prism 377 DNA sequencer (Sequetech). Presence of primary site reversion was confirmed by two independent sequences using forward and reverse oligos.

RNA extraction and qRT-PCR. Total RNA was isolated from cells or lml cell culture supematants using TRizol (lnvitrogen) or QiaAmp UltraSens kit (Qiagen),

respectively. qRT-PCRs mixtures were assembled in triplicates using 0.5µg or 4µ1 RNA and High-Capacity RNA-to-cDNA (Applied Biosystems). TaqMan reagents are listed in Table 4. Amplification and analysis were pelformed using StepOnePlus Real-Time-PCR system (Applied Biosystems). Sl8 was used as a control.

Western blot. Cell lysates were subjected to western analyses using primary antibodies followed by HRP-conjugated secondary antibodies. Bands intensity was quantified using NIH Image.

Quantitative immunofluorescence confocal microscopy. Huh-7 .5 cells were electroporated with J6/JFH HCV RNA or infected with culture grown J6/JFH virus, seeded onto coverslips, and incubated for 72 hr at 37°C. Cells were fixed with 4% paraformaldehyde in PBS, washed with PBS, permeabilized with 0.1% Triton X-100 in PBS for 5 min, and blocked for 1 hr in PBS containing 1% BSA. Fixed cells were incubated with primary antibodies against either the core protein, AP2M l, TGN46, calreticulin or E2 at room temperature for 1 hr (see Table 1). Secondary antibodies (goat anti-rabbit Alexa Fluor 488, goat anti-mouse Alexa Fluor 594, chicken anti-goat IgG Alexa Fluor 647, and goat anti-human Alexa Fluor 647) (Invitrogen) were incubated for 1 hr at room temperature. LD staining with Bodipy-488/503 (Invitrogen) was performed as described (Kopp *et al., J Viral* 84, 1666-1673, 2010). Cover slips were then washed three times in PBS, and mounted with ProLong Gold antifade reagent (Invitrogen). Alternatively, Huh-7.5 cells were co-transfected with plasmids expressing AP2M l-YFP and/or Core-mCherry. LD were stained with HCS LipidTOX (Invitrogen). All slides were analyzed using Zeiss LSM 510 confocal microscope. Colocalization was quantified in 10-15 randomly chosen cells from each sample using ImageJ (JACoP) colocalization Software and Manders' Colocalization Coefficients (MCC). Threshold values were determined using auto_threshold (plugin, ImageJ). Only pixels whose red and green intensity values were both above their respective thresholds were considered to be pixels with colocalized probes. MCCs were then calculated as the fractions of total fluorescence in the region of interest that occurs in these "colocal" pixels (with a higher value representing more colocalization). M2 coefficient values represented as mean percent colocalization are shown.

Statistical analysis. IC50s and EC50s were measured by fitting data to a three parameter logistic curve, as described (Einav *et al., Nature Biotechnolog y* 26, 1019-1027, 2008). P-values were calculated using one-tailed unpaired Student's t test.

Example 3

The effect of erlotinib, sunitinib, and PKC-412 on AP2Ml phosphorylation.

To determine the effect of the inhibitory compounds on AP2M l phosphorylation, Huh-7.5 cells harboring J6/JFH(p7-Rluc2A) RNA were treated daily with various concentrations of the compounds or with DMSO. Since AP2M l phosphorylation is transient (due to the activity of the phosphatase PP2A (Ricotta *et al., J Biol Chem* 283:5510-5517, 2008), to allow capturing of the phosphorylated AP2M l state, lysates were prepared at 72hr following a 30 min incubation of the cells with 1OOnM of the PP2A inhibitor, calyculin A (Cal-A) or a DMSO control. Samples were then subjected to SDS-PAGE and blotting with antibodies targeting either the phosphorylated AP2M 1 form or actin. Indeed, significantly lower ratios of phosphorylated-AP2M l to actin were measured in lysates prepared from cells treated with either sunitinib, erlotinib or PKC-412, compared with the DMSO control (Fig. 6H). Phospho-AP2M l to actin ratios were progressively decreased by increasing concentrations of sunitinib and erlotinib in a dose-dependent manner. These results suggest that AAK l or GAK are potently inhibited by these compounds in Huh-7.5 cells harboring infectious HCV.

The effect of the compounds on HCV RNA replication, intracellular infectivity, extracellular infectivity, and HCV replication.

To determine the effect of erlotinib, sunitinib, and PKC-412 on infectious virus production, Huh-7.5 cells were electroporated with J6/JFH(p7-Rluc2A) HCV genome (Murray *et al., J Viral* 81:10220-10231, 2007), plated in 6-well plates, and treated daily with serial dilutions of the compounds or DMSO. At 72 hr postelectroporation, cellular viability was measured by alamarBlue-based assays followed by luciferase assays for determination of HCV RNA replication. Cell culture supematants and lysates were collected at 72 hr from parallel samples and used to inoculate naive Huh-7 .5 cells for determination of extracellular and intracellular infectivity, respectively. Luciferase assays were performed in these cells at 72 hr post inoculation.

To determine the effect of these compounds on HCV replication, $6x10^3$ Huh-7.5 cells seeded in 96-well plates were infected in triplicates with cell culture-grown HCV (J6/JFH(p7-Rluc2A)) tittered at $6.3x10^5$ $TCIDs_0$/ml with an MOI *(multiplicity of infection)* of 0.1. At 6 hr postelectroporation, cells were washed and medium was

replaced with serial dilutions of the inhibitory compounds. Cells were treated daily for 72 hr and then subjected to viability assays followed by standard luciferase assays.

Alternatively, $6xl0^3$ Huh-7.5 cells seeded in 96-well plates were infected m triplicates with cell culture-grown HCV J6/JFH (titer: $l.2x\,l0^5$ $TCID_{50}$/ml with an MOI of 0.1, and subjected to focus formation assays, as described (Lindenbach et al., *Science* 309:623-626, 2005). Following 72 hr of daily treatment with the compounds, cells were fixed in 4% formaldehyde and permeabilized with saponin. HCV core protein was detected with primary anti-core monoclonal and secondary goat anti-mouse Alexa 594-conjugated antibodies. Foci were counted under an inverted microscope.

Experiments were repeated twice, each with 4 replicates. Signal was normalized to samples grown in the presence of the corresponding concentration of DMSO. EC50s were measured by fitting data to a three parameter logistic curve using the formula Y=a+(b-a)/(l+ lO"(X-c)) (a, b and c represent minimum binding, maximum binding and logEC50, respectively)(BioDataFit, Chang Bioscience, Inc).

Table 1. Antibodies used in this study

Antibodies	Source
Biotinylated anti-penta-his	Qiagen
FITC-conjugated anti-VS	Invitrogen
Phycoerythrin-conjugated anti-TI tag	Abeam
Rabbit Anti-AAK I	Abeam
Rabbit anti-AP2Ml	Santa Cruz Biotechnology
Goat anti-AP2Ml	Santa Cruz Biotechnology
Rabbit anti-phospho-AP2Ml (Tl56)	Cell signaling
Rabbit anti-TGN46	Abeam
Rabbit anti-calreticulin	Enzo Life Sciences
Mouse anti-GAK	MBL international corporation
Mouse anti-core	Austral Biologicals
Human anti-E2 (CBH5)	Dr. Steven K Foung
Mouseanti-actin	Sigma
HRP-conjugated anti-mouse IgG	Cell signaling
HRP-conjugated anti-rabbit IgG	Cell signaling

Table 2. Compounds used in this study

Compound	Source
PKC-412	Sigma
Sunitinib malate	Sigma 15
Erlotinib	LC Laboratories
Calyculin A	Cell Signaling

Table 3. RNAi used in this study

RNAi type	Target	Catalogue#	Source	Sequence
ON-TARGETplus SMARTpools	AP2Ml	LQ-008170-00-0002	Dhannacon	
	AP2Ml	J-004233-05	Dhannacon	GUUAAGCGGUCCAACAUUU
	AP2Ml	J-004233-06	Dhannacon	GCGAGAGGGUAUCAAGUAU
	AP2Ml	J-004233-07	Dhannacon	AGUUUGAGCUUAUGAGGUA
	AP2Ml	J-004233-08	Dhannacon	GAACCGAAGCUGAACUACA
	Non-targeting	D-001810-10-05	Dhannacon	Non Available
siRNAs	AAKl	Ref #4	Ambion	GGUGUGCAAGAGAGAAAUCtt
	GAK	Ref #5	Dhannacon	AACGAAGGAACAGCUGAUUCA
	Non-targeting	D-001210-01-05	Dhannacon	Non Available
MISSION shRNAs	AP2Ml	TRCN0000060238 ("238")	Sigma	CCGGGTGGTCATCAAGTCCAACTTTCTCGA GAAAGTTGGACTTGATGACCACTTTTTG
	AP2Ml	TRCN0000060239 ("239")	Sigma	CCGGCACCAGCTTCTTCCACGTTAACTCGA GTTAACGTGGAAGAAGCTGGTGTTTTTG
	AP2Ml	TRCN0000060241 ("241")	Sigma	CCGGGCTGGATGAGATTCTAGACTTCTCGA GAAGTCTAGAATCTCATCCAGCTTTTTG
	AP2Ml	TRCN0000060242 ("242")	Sigma	CCGGCATTTATGAAACTCGCTGCTACTCGAG TAGCAGCGAGTTTCATAAATGTTTTTG
	Non-targeting	SHC002V	Sigma	Non Available

Table 4. Taqman probes and primers used in this study

Primer name	**Sequence or** assay **catalogue** #
HCV Forward	CTTCACGCAGAAAGCGTCTA
HCV Reverse	CAAGCACCCTATCAGGCAGT
HCV Taqman MGB probe	[6FAM]-TATGAGTGTCGTGCAGCCTC-[MGB-NFQ]
AP2Ml	HS01037584_ml
GAK	HSOO 178347_ml
AAKl	HS00208618_ml
Sl8	hs999999_ml

All reagents listed in this table were purchased from Applied Biosystems, Inc (ABI).

CLAIMS

Claim 1. Use of an agent in the manufacture of a medicament for use in treating HCV- HIV coinfection, wherein the agent comprises sunitinib or pharmaceutically acceptable salt of sunitinib.

Claim 2. The use of claim 1, wherein said sunitinib or pharmaceutically acceptable salt of sunitinib inhibits binding of the HCV and HIV viruses to a host µ subunit of clathrin adaptor protein (AP) complexes comprising µ subunits of clathrin AP1,AP2, AP3 and AP4 complexes.

Claim 3. The use of claim 2, wherein said µ subunit is selected from the group consisting of AP2M1, AP1M1, AP3M1 and AP4M1.

Claim 4. The use of claim 1,wherein sunitinib or pharmaceutically acceptable salt of sunitinib inhibits AAK1 or GAK.

Claim 5. The use of claim 1, wherein said agent further comprises erlotinib, PCK-412, or pharmaceutically acceptable salts or combinations thereof.

Claim 6. Use of an agent in the manufacture of an inhibitor of infections by Lentiviridae, HIV, HCV/HIV co-infection, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV, wherein said agent comprises sunitinib or pharmaceutically acceptable salt of sunitinib.

Claim 7. The use of claim 6, wherein said sunitinib or pharmaceutically acceptable salt of sunitinib inhibits binding of a protein from said virus comprising YXXΦ or dileucine motif to a host µ subunit of clathrin adaptor protein (AP) complexes, and wherein said µ subunit is selected from the group consisting of µ subunits of clathrin AP1, AP2, AP3 and AP4 complexes.

Claim 8. The use of claim 7, wherein said µ subunit is selected from the group consisting of AP2M1, AP1M1, AP1M3 and AP4M1.

Claim 9. The use of claim 6, wherein said sunitinib or pharmaceutically acceptable salt of sunitinib inhibits AAK1 or GAK.

Claim 10. The use of claim 6, wherein said agent further comprises erlotinib, PKC-412 or pharmaceutically acceptable salts or combinations thereof.

Claim 11. A method of screening for antiviral agents comprising:

contacting a clathrin AP binding virus, Flaviviridae, Lentiviridae, HCV, HIV, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding viruses other than HCV or protein thereof with a candidate agent and a host µ subunit of clathrin adaptor protein (AP) complexes, and determining an effect of said candidate agent on the binding of

said clathrin AP binding virus, Flaviviridae, Lentiviridae, HCV, HIV, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding virus other than HCV to said µ subunit of clathrin adaptor protein (AP) complexes.

Claim 12. A method of screening for antiviral agents comprising:

contacting a clathrin AP binding virus, Flaviviridae, Lentiviridae, HCV, HIV, Flaviviridae other than HCV, clathrin AP binding viruses other than Flaviviridae, or clathrin AP binding virus other than HCV virus with a candidate agent and a cell expressing a host µ subunit of clathrin adaptor protein (AP) complexes and determining the effect of said candidate agent on viral assembly or budding.

Claim 13. The method of claim 11, wherein said candidate agent inhibits the activity of AAK l or GAK.

Claim 14. Use of an agent in the manufacture of an inhibitor of assembly or budding of a Flaviviridae, a virus other than a Flaviviridae, or a virus other than HCV in a host cell, wherein said agent comprises sunitinib or a pharmaceutically acceptable salts of sunitinib, whereby said assembly or budding is inhibited.

Claim 15. The method of claim 14, wherein said sunitinib or a pharmaceutically acceptable salt of sunitinib blocks binding of a viral structural protein to a host protein.

Claim 16. The method of claim 14, wherein said sunitinib or a pharmaceutically acceptable salt of sunitinib competes with a viral protein for binding to a host protein.

Claim 17. The method of claim 14, wherein said sunitinib or a pharmaceutically acceptable salt of sunitinib competes with a host protein for binding to a viral protein.

Claim 18. The use of claim 6, wherein sunitinib or a pharmaceutically acceptable salt of sunitinib inhibits infection by a Flaviviridae other than HCV in a patient.

Claim 19. The use of claim 18, wherein Flaviviridae other than HCV comprises a Dengue virus.

Claim 20. The use of claim 14, wherein said agent further comprises erlotinib, PKC-412 or pharmaceutically acceptable salts or combinations thereof.