METHOD OF REDUCING THE EFFECT OF DIRECT AND MEDIATED INTERFERENCE CURRENT IN AN ELECTROCHEMICAL TEST STRIP

FIELD OF THE INVENTION

[0001] The present invention is related, in general to methods of reducing the effect of interfering compounds on measurements taken by analyte measurement systems and, more particularly, to a method of reducing the effects of direct interference currents and mediated interference currents in a glucose monitoring system using an electrochemical strip having electrodes with regions coated with active reagent and regions coated with inactive reagent.

BACKGROUND OF INVENTION

[0002] In many cases, an electrochemical glucose measuring system may have an elevated oxidation current due to the oxidation of interfering compounds commonly found in physiological fluids such as, for example, acetaminophen, ascorbic acid, bilirubin, dopamine, gentisic acid, glutathione, levodopa, methyldopa, tolazimide, tolbutamide, and uric acid. The accuracy of glucose meters may, therefore, be improved by reducing or eliminating the portion of the oxidation current generated by interfering compounds. Ideally, there should be no oxidation current generated from any of the interfering compounds so that the entire oxidation current would depend only on the glucose concentration.

[0003] It is, therefore, desirable to improve the accuracy of electrochemical sensors in the presence of potentially interfering compounds such as, for example, ascorbate, urate, and, acetaminophen, commonly found in physiological fluids. Examples of analytes for such electrochemical sensors may include glucose, lactate, and fructosamine. Although glucose will be the main analyte discussed, it will be obvious to one skilled in the art that the invention set forth herein may also be used with other analytes.

[0004] Oxidation current may be generated in several ways. In particular, desirable oxidation current results from the interaction of the redox mediator with the analyte of interest (e.g., glucose) while undesirable oxidation current

is generally comprised of interfering compounds being oxidized at the electrode surface and by interaction with the redox mediator. For example, some interfering compounds (e.g., acetominophen) are oxidized at the electrode surface. Other interfering compounds (e.g., ascorbic acid) are oxidized by chemical reaction with the redox mediator. This oxidation of the interfering compound in a glucose measuring system causes the measured oxidation current to be dependent on the concentration of both the glucose and any interfering compound. Therefore, in the situation where the concentration of interfering compound oxidizes as efficiently as glucose and the interferent concentration is high relative to the glucose concentration, the measurement of the glucose concentration would be improved by reducing or eliminating the contribution of the interfering compounds to the total oxidation current.

[0005] One known strategy that can be used to decrease the effects of interfering compounds is to use a negatively charged membrane to cover the working electrode. As an example, a sulfonated fluoropolymer such as NAFION™ may be used to repel all negatively charged chemicals. In general, most interfering compounds such as ascorbate and urate have a negative charge, thus, the negatively charged membrane prevents the negatively charged interfering compounds from reaching the electrode surface and being oxidized at that surface. However, this technique is not always successful since some interfering compounds such as acetaminophen do not have a net negative charge, and thus, can pass through a negatively charged membrane. Nor would this technique reduce the oxidation current resulting from the interaction of interfering compounds with some redox mediators. The use of a negatively charged membrane on the working electrode could also prevent some commonly used redox mediators, such as ferricyanide, from passing through the negatively charged membrane to exchange electrons with the electrode.

[0006] Another known strategy that can be used to decrease the effects of interfering compounds is to use a size selective membrane on top of the working electrode. As an example, a 100 Dalton exclusion membrane such as cellulose acetate may be used to cover the working electrode to exclude all chemicals with a molecular weight greater than 100 Daltons. In general, most

interfering compounds have a molecular weight greater than 100 Daltons, and thus, are excluded from being oxidized at the electrode surface. However, such selective membranes typically make the test strip more complicated to manufacture and increase the test time because the oxidized glucose must diffuse through the selective membrane to get to the electrode.

[0007] Another strategy that can be used to decrease the effects of interfering compounds is to use a redox mediator with a low redox potential, for example, between about –300mV and +100 mV (when measured with respect to a saturated calomel electrode). Because the redox mediator has a low redox potential, the voltage applied to the working electrode may also be relatively low which, in turn, decreases the rate at which interfering compounds are oxidized by the working electrode. Examples of redox mediators having a relatively low redox potential include osmium bipyridyl complexes, ferrocene derivatives, and quinone derivatives. A disadvantage of this strategy is that redox mediators having a relatively low potential are often difficult to synthesize, unstable and have a low water solubility.

[0008] Another known strategy that can be used to decrease the effects of interfering compounds is to use a dummy electrode which is coated with a redox mediator. In some instances the dummy electrode may also be coated with an inert protein or deactivated redox enzyme. The purpose of the dummy electrode is to oxidize the interfering compound at the electrode surface and/or to oxidize the redox mediator reduced by the interfering compound. In this strategy, the current measured at the dummy electrode is subtracted from the total oxidizing current measured at the working electrode to remove the interference effect. A disadvantage of this strategy is that it requires that the test strip include an additional electrode and electrical connection (i.e., the dummy electrode) which cannot be used to measure glucose. The inclusion of dummy electrode is an inefficient use of an electrode in a glucose measuring system.

SUMMARY OF INVENTION

[0009] The present invention is directed to a method of reducing interferences in an electrochemical sensor wherein the method includes the step of

measuring a first current at a first working electrode, the first working electrode being covered by an active reagent layer, the step of measuring a second current at a second working electrode, the second working electrode being covered by an inactive reagent layer and the step of calculating a corrected current value representative of a glucose concentration using a ratio of an active area of the first working electrode to an inactive area of the second working electrode.

[00010] The present invention is further directed to a method of reducing interferences in an electrochemical sensor wherein the method includes the step of measuring a first current at a first working electrode, the first working electrode being covered by an active reagent layer, the step of measuring a second current at a second working electrode, wherein the active reagent layer is disposed on an active region of the second working electrode and an inactive region of the second working electrode is covered by an inactive reagent layer and the step of calculating a corrected current value representative of a glucose concentration using a ratio of an active region on the first and second working electrodes and an inactive region on the second working electrode.

BRIEF DESCRIPTION OF DRAWINGS

[00011] A better understanding of the features and advantages of the present invention will be obtained by reference to the following detailed description that sets forth illustrative embodiments, in which the principles of the invention are utilized, and the accompanying drawings, of which:

[00012] Figure 1 is an exploded perspective view of a test strip according to an exemplary embodiment of the present invention;

[00013] Figure 2 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 1 including a conductive layer and an insulation layer;

[00014] Figure 3 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 1, wherein the

position of an active and an inactive reagent layer do not touch each other and are illustrated with the insulation and conductive layer;

[00015] Figure 4 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 1, wherein the position of the active and the inactive reagent layer are immediately adjacent to each other and are illustrated with the insulation and conductive layer;

[00016] Figure 5 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 1, wherein the position of the active and the inactive reagent layer that overlap with each other and are illustrated with the insulation and conductive layer;

[00017] Figure 6 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 1, wherein the active and the inactive reagent layer do not touch each other and are illustrated with the conductive layer;

[00018] Figure 7 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 1, wherein the active and the inactive reagent layer are immediately adjacent to each other and are illustrated with the conductive layer;

[00019] Figure 8 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 1, wherein the active and the inactive reagent layer overlap with each other and are illustrated with the conductive layer;

[00020] Figure 9 is a simplified schematic showing a meter interfacing with a test strip that has a first contact, second contact, and reference contact disposed on a substrate;

[00021] Figure 10 is a simplified schematic showing a meter interfacing with a test strip that has a first contact and a second contact disposed on a substrate and a reference contact which is orientated in a facing orientation with the first contact and second contact;

[00022] Figure 11 is a graph showing the effects of gamma radiation on precision for test strips tested at a 20 mg/dL glucose concentration;

[00023] Figure 12 is a graph showing the effects of gamma radiation on precision for test strips tested at a 50 mg/dL glucose concentration;

[00024] Figure 13 is a graph showing the effects of gamma radiation on precision for test strips tested at a 100 mg/dL glucose concentration;

[00025] Figure 14 is a graph showing the effects of gamma radiation on precision for test strips tested at a 300 mg/dL glucose concentration;

[00026] Figure 15 is a graph showing the effects of gamma radiation on precision for test strips tested at a 500 mg/dL glucose concentration;

[00027] Figure 16 is a graph showing the effects of gentisic acid on accuracy for test strips tested at a 70 mg/dL glucose concentration;

[00028] Figure 17 is a graph showing the effects of gentisic acid on accuracy for test strips tested at a 240 mg/dL glucose concentration;

[00029] Figure 18 is a graph showing the effects of uric acid on accuracy for test strips tested at a 70 mg/dL glucose concentration;

[00030] Figure 19 is a graph showing the effects of uric acid on accuracy for test strips tested at a 240 mg/dL glucose concentration;

[00031] Figure 20 a simplified plane view of a distal portion of a test showing a modified cutout that allows the area of a second working electrode to be increased;

[00032] Figure 21 is an exploded perspective view of a test strip according to another exemplary embodiment of the present invention;

[00033] Figure 22 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 21, wherein the position of an active and an inactive reagent layer do not touch each other and are illustrated with the insulation and conductive layer;

[00034] Figure 23 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 21, wherein the position of the active and the inactive reagent layer are immediately adjacent to each other and are illustrated with the insulation and conductive layer;

[00035] Figure 24 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 21, wherein the position of the active and the inactive reagent layer that overlap with each other and are illustrated with the insulation and conductive layer;

[00036] Figure 25 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 21, wherein the active

and the inactive reagent layer do not touch each other and are illustrated with the conductive layer;

[00037] Figure 26 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 21, wherein the active and the inactive reagent layer are immediately adjacent to each other and are illustrated with the conductive layer; and

[00038] Figure 27 is a simplified plane view of a distal portion of a test strip according to the embodiment of the present invention illustrated in Figure 21, wherein the active and the inactive reagent layer overlap with each other and are illustrated with the conductive layer;

DETAILED DESCRIPTION OF THE INVENTION

[00039] The invention described herein includes a test strip to improve the accuracy of a glucose measurement in the presence of interfering compounds. Under certain circumstances, a type of interfering compound may develop in the test strip itself before bodily fluid such as, for example, blood is added. An example this type of interfering compound may be a reduced mediator (e.g. ferrocyanide) which develops from the conversion of an oxidized mediator (e.g. ferricyanide). This causes the background signal to increase which, in turn, decreases the accuracy of the test strip measurement. It should be noted that in this circumstance the interfering compound develops in the test strip itself as opposed to being provided to the test strip in the form of a bodily fluid.

[00040] Typically, an oxidized mediator is disposed on a working electrode with the intent that the oxidized mediator will be stable and not transition over to the reduced redox state. The generation of reduced mediator causes the background signal to increase for electrochemical sensors which use an oxidation current to correlate with the glucose concentration. In general, ferricyanide (e.g. oxidized mediator) tends to become reduced over time to the reduced redox state. Ferricyanide generally transitions to the reduced redox state more rapidly when exposed to environmental conditions which include but are not limited to, basic pH, elevated temperature, elevated humidity, bright light condition, electron beam radiation, and gamma radiation.

[00041] Recently, a lance and a test strip have been integrated into a single medical device. These integrated medical devices can be employed, along with an associated meter, to monitor various analytes, including glucose. Depending on the situation, test strips can be designed to monitor analytes in an episodic single-use format, semi-continuous format, or continuous format. The integration of the lance and the test strip simplifies a monitoring procedure by eliminating the need for a user to coordinate the extraction of a bodily fluid from a sample site with the subsequent transfer of that bodily fluid to the test strip. In such a case, the lance and test strip must be sterilized together so as to mitigate the risk of infection.

[00042] Ionizing radiation may be used to sterilize test strips with a lance. Possible sources of ionizing radiation are electron beam, gamma, and x-ray. However, one of the challenges in sterilizing a test strip is to provide a sufficiently high intensity of radiation such that a sufficiently high proportion of microorganisms are neutralized for an entire package of test strips, while at the same time not adversely affecting the reagent layer. Typically, a batch or package of test strips are exposed to an ionizing radiation dose ranging from about 10 KGy to about 50 KGy. For the case using e-beam sterilization, the energy of the incident e-beam source can range from about 3 MeV to about 12 MeV. The impingent ionizing radiation may often have some non-uniformities in its intensity causing a particular portion of the package to receive more ionizing radiation than another portion of the package. Experiments have shown that both gamma radiation and electron beam radiation cause the background signal of the electrochemical sensors to increase. Furthermore, the relatively non-uniform nature of the radiation causes the background signal to increase in a non-uniform nature for a sterilized batch of test strips. This causes the precision to decrease when testing a particular batch of sterilized glucose test strips. In addition, the decrease in precision is exacerbated at the low glucose concentration range (e.g. about 20 mg/dL to about 100 mg/dL) because the proportion of reduced mediator is relatively high with respect to the low glucose concentration range.

[00043] Figure 1 shows an exploded perspective view of a test strip 800 that is designed to compensate for the variations in increased background potentially caused by the conversion of oxidized mediator to reduced mediator. In the embodiment of the present invention illustrated in Figure 1, an electrochemical test strip 800, which may be used for measuring glucose concentration in bodily fluids such as blood or

interstitial fluid, includes a first working electrode 808, a second working electrode 806, and a reference electrode 810. An active reagent layer 820 is disposed on first working electrode 808 and reference electrode 810 where active reagent layer 820 completely covers first working electrode 808 and at least partially covers reference electrode 810. An inactive reagent layer 818 is disposed on second working electrode 806.

[00044] In an embodiment of this invention, active reagent layer 820 may include, for example, glucose oxidase and a mediator such as, for example, ferricyanide. Inactive reagent layer 818 may include a mediator, but no active enzymes which are specific for the analyte of interest. Because ferricyanide has a redox potential of approximately 400 mV (when measured with respect to a saturated calomel electrode) at a carbon electrode, the introduction of a bodily fluid e.g., blood may generate a significant oxidation of interferents by the redox mediator and /or the working electrode generating a significant undesirable oxidation current. Therefore, the oxidation current measured at first working electrode 808 will be a superposition of oxidation current sources: a first, desirable, oxidation current generated by the oxidation of glucose; a second, undesirable, direct oxidation of interferents at the electrode (direct interference current); and a third, undesirable, indirect oxidation of interferents via a mediator (mediated interference current). The oxidation current measured at second working electrode 806 will also be a superposition of oxidation current sources similar to first working electrode 808, but the first, desirable, oxidation current should not occur because there is no enzyme present on second working electrode 806. Because the oxidation current measured at second working electrode 806 depends only on interferents, and the oxidation current measured at first working electrode 808 depends on glucose and interferents, it is possible to calculate a corrected glucose current which is independent to the effects of interfering compounds oxidized at first working electrode 808 and second working electrode 806. In such a case, the current density of first working electrode 808 is subtracted from the current density of the second working electrode 806 to calculate the corrected glucose current density G where

$$G = WE_1 - WE_2 \qquad \text{(Eq 8)}$$

where WE_1 is the current density at first working electrode 808 and WE_2 is the current density at second working electrode 806.

[00045] In an alternative embodiment to this invention, the interferent oxidation current density at second working electrode 806 may be slightly different than the interferent oxidation current density at first working electrode 808 because there is no enzyme on second working electrode 806. In such a case, a constant K can be used to correct for such non-idealities in the current measurements. Equation 9 shows how constant K would modify the previously described Equation 8.

$$G = WE_1 - (k \text{ x } WE_2) \qquad \text{(Eq 8)}$$

where K can range from about 0.5 to about 1.5.

[00046] Test strip 800 includes a substrate 50, a conductive layer 802, an insulation layer 804, inactive reagent layer 818, active reagent layer 820, an adhesive layer 830, and a top layer 824. Test strip 800 may be manufactured by sequentially printing five layers which are conductive layer 802, insulation layer 804, inactive reagent layer 818, active reagent layer 820, and adhesive layer 830 onto substrate 50. Top layer 824 may be assembled by a lamination process. Test strip 800 further includes a first side 54, a second side 56, a distal portion 58, and a proximal portion 60.

[00047] In one embodiment of the present invention, substrate 50 is an electrically insulating material such as plastic, glass, ceramic, and the like. In an embodiment of this invention, substrate 50 may be a plastic such as, for example, nylon, polycarbonate, polyimide, polyvinylchloride, polyethylene, polypropylene, PETG, or polyester. More particularly the polyester may be, for example Melinex ® ST328 which is manufactured by DuPont Teijin Films. Substrate 50 may also include an acrylic coating which is applied to one or both sides to improve ink adhesion.

[00048] The first layer deposited on substrate 50 is conductive layer 802 which includes first working electrode 808, second working electrode 806, reference electrode 810, and strip detection bar 17. In accordance with the present invention, a screen mesh with an emulsion pattern may be used to deposit a material such as, for example, a conductive carbon ink in a defined geometry as illustrated in Figure 1. Conductive layer 802 may be disposed on substrate 50 by using screen printing, rotogravure printing, sputtering, evaporation, electroless plating, ink jetting,

sublimation, chemical vapor deposition, and the like. Suitable materials which may be used for conductive layer 802 are Au, Pd, Ir, Pt, Rh, stainless steel, doped tin oxide, carbon, and the like. In an embodiment of this invention, the carbon ink layer may have a height between 1 and 100 microns, more particularly between 5 and 25 microns, and yet even more particularly at approximately 13 microns. The height of conductive layer 802 can vary depending on the desired resistance and conductivity of the printed material.

[00049] A first contact 814, a second contact 812, and a reference contact 816 may be used to electrically interface with a meter. This allows the meter to electrically communicate to first working electrode 808, second working electrode 806, and reference electrode 810 via, respective, first contact 814, second contact 812, and reference contact 816.

[00050] The second layer deposited on substrate 50 is insulation layer 804. Insulation layer 804 is disposed on at least a portion of conductive layer 802 as shown in Figures 1 and 2. Figure 2 is a simplified plane view of distal portion 58 of test strip 800 which highlights the position of first working electrode 808, second working electrode 806, and reference electrode 810 with respect to insulation layer 804. Insulation layer 804 further includes a cutout 18 which may have a rectangular shaped structure as shown in Figure 1 and 2. Cutout 18 exposes a portions of first working electrode 808, second working electrode 806, and reference electrode 810 which can be wetted with liquid. Cutout 18 includes a cutout width W20 and a cutout length L26. Cutout width W20 corresponds to a width of second working electrode 806, reference electrode 810, and first working electrode 808 as illustrated in Figure 2. In an embodiment of this invention, cutout width W20 may range from about 0.7 mm to about 1.4 mm, and cutout length L26 may range from about 0.4 mm and about 3.4 mm.

[00051] In one embodiment of the present invention, second working electrode 806 and first working electrode 808 have a respective length of L20 and L21 which may be the same and range from about 0.1 mm to about 0.8 mm. Reference electrode 810 may have a length L24 which may range from about 0.2 mm to about 1.6 mm. In accordance with the present invention, electrode spacing S1 is a distance between second working electrode 806 and reference electrode 810; and between reference electrode 810 and first working electrode 808 which may range from about 0.2 mm to about 0.6 mm.

[00052] In an alternative embodiment of the present invention, an area of first working electrode 808 may be different than an area of second working electrode 806. A ratio of first working electrode 808 area:second working electrode 806 area may range from about 1:1 to about 1:3. Under certain situations, the reduction in background can be improved by increasing the relative area of second working electrode 806. The area of second working electrode 806 may be increased by modifying the geometry of a cutout 6008 as shown in Figure 20.

[00053] Figure 2 shows that strip 800 may be cut along incision line A-A' after it is fully laminated as illustrated in Figure 1. In the process of cutting test strip 800 along incision line A-A' as illustrated in Figure 1, a sample inlet 52 is created in which a liquid sample can be applied for dosing test strip 800.

[00054] Figures 3 to 5 are a simplified plane view of distal portion 58 of test strip 800 according to the embodiment of the present invention illustrated in Figure 1, which show various positions of active reagent layer 820 and inactive reagent layer 818 with respect to each other. Figures 6 to 8, which correspond to Figures 3 to 5 respectively, do not show insulation layer 804 to help demonstrate more clearly the relationship between the conductive layer 802, active reagent layer 820, and inactive reagent layer 818.

[00055] Test strip 800 may have inactive reagent layer 818 disposed on second working electrode 806 such that it completely covers second working electrode 806 as is illustrated in Figures 3 to 5. In one embodiment of this invention, inactive reagent layer 818 completely covers second working electrode 806, but does not touch reference electrode 810 as is illustrated in Figures 3 and 4. In another embodiment of this invention, inactive reagent layer 818 completely covers second working electrode 806 and at least partially covers reference electrode 810 as is illustrated in Figure 5.

[00056] In an embodiment of this invention, inactive reagent layer 818 includes at least an oxidized mediator, such as ferricyanide, and may optionally include an inert protein or inactivated enzyme. Inactive reagent layer 818 may further include a citrate buffer at pH 6, a polyvinyl alcohol, a polyvinyl pyrrolidone-vinyl acetate, a Dow Corning DC1500 antifoam, a hydroxyethyl cellulose (Natrosol 250G, Hercules), and a surface modified silica (Cab-o-sil TS 610, Cabot) having both hydrophilic and hydrophobic domains. Examples of oxidized mediators may be ferricyanide, ferricinium complexes, quinone complexes, and osmium complexes. Examples of

inert protein may be crotein or albumin (e.g. bovine or human). Examples of inactivated enzyme may be the apo form of PQQ-glucose dehydrogenase (where PQQ is an acronym for pyrrolo-quinoline-quinone) or apo glucose oxidase (e.g. enzyme with no active site). Enzyme may also be deactivated or sufficiently attenuated by heat treatment or by treatment with denaturing agents such as urea. Because inactive reagent layer 818 does not include an active enzyme, the oxidation current measured at second working electrode 806 is not proportional to the glucose concentration. For this reason, one skilled in the art may refer to second working electrode 806 as a dummy electrode.

[00057] In an embodiment of this invention, the inert protein or deactivated enzyme in inactive reagent layer 818 may act as a stabilizer for the mediator. The inert protein or deactivated enzyme may shield the mediator during the drying process at elevated temperature. In addition, the inert protein or deactivated enzyme may act as a desiccant which helps protect the mediator from moisture that may potentially destabilize the mediator.

[00058] Test strip 800 has active reagent layer 820 disposed on first working electrode 808 as illustrated in Figures 3 to 5. In another embodiment of this invention, active reagent layer 820 completely covers first working electrode 808, but does not touch reference electrode 810. In another embodiment of this invention, active reagent layer 820 completely covers first working electrode 808 and at least partially covers reference electrode 810 as illustrated in Figures 3 to 5.

[00059] In an embodiment of this invention, active reagent layer 820 includes at least an oxidized mediator, and an enzyme. Active reagent layer 820 may further include a citrate buffer at pH 6, a polyvinyl alcohol, a polyvinyl pyrrolidone-vinyl acetate, a Dow Corning DC1500 antifoam, a hydroxyethyl cellulose (Natrosol 250G, Hercules), and a surface modified silica (Cab-o-sil TS 610, Cabot) having both hydrophilic and hydrophobic domains. Examples of oxidized mediators may be ferricyanide, ferricinium complexes, quinone complexes, and osmium complexes. Examples of the enzyme may be glucose oxidase, glucose dehydrogenase using a PQQ co-factor, and glucose dehydrogenase using a nicotinamide adenine dinucleotide co-factor. Because active reagent layer 820 does include the enzyme, the oxidation current measured at first working electrode 808 is proportional to the glucose concentration.

[00060] It should be noted that if screen printing were used for depositing both inactive reagent layer 818 and active reagent layer 820, then two separate screen printing steps would be required to deposit the respective reagent layers onto the appropriate electrode(s). It should be noted that screen printing is not well-suited for printing two discrete reagents on the same screen. The squeegee motion during printing may cause the two respective reagents to mix during the screen printing process. Figure 3 shows an embodiment of this invention which has inactive reagent layer 818 disposed on second working electrode 806, and active reagent layer 820 disposed on first working electrode 808 and reference electrode 810. In this embodiment, inactive reagent layer 818 does not touch or overlap with active reagent layer 820. Because the area of second working electrode 806, first working electrode 808 and reference electrode 810 is relatively small, it can be difficult to sequentially align and coat inactive reagent layer 818 and active reagent layer 820, respectively, with the desired yield. It should also be noted that relatively small electrode areas (e.g. about 0.6 mm^2) are preferred because this allows the volume of liquid sample required for a test strip to be small.

[00061] In an embodiment of this invention, inactive reagent layer 820 is printed first and then dried at an elevated temperature. Active reagent layer 818 is then subsequently printed followed by another drying step at an elevated temperature as described in International Application serial number PCT/GB/03004708 which is hereby incorporated by reference herein. Because active reagent layer 818 is deposited second, it is exposed to only one drying step as opposed to the two drying steps for inactive reagent layer 820. This helps stabilize both mediator and enzyme within active reagent layer 818 because under certain conditions enzymes can degrade with continued exposure to elevated temperatures.

[00062] In an embodiment of this invention, Figure 4 shows inactive reagent layer 818 disposed on second working electrode 806, and active reagent layer 820 disposed on first working electrode 808 and reference electrode 810. In this embodiment inactive reagent layer 818 and active reagent layer 820 are immediately adjacent to each other. In such a case, the inactive reagent layer 818 and active reagent layer 820 would touch, but typically not overlap with each other to any significant extent. Although the printing process targets the alignment such that inactive reagent layer 818 and active reagent layer 820 are immediately adjacent to each other, normal manufacturing variation will cause some overlap to occur with a certain frequency between inactive

reagent layer 818 and active reagent layer 820. Likewise, such variation will also cause inactive reagent layer 818 to sometimes not touch active reagent layer 820. Because inactive reagent layer 818 was allowed to touch or not touch active reagent layer 820 and the operation of the method of the invention still works to reduce the variation in the background in either circumstance, the yield of acceptable test strips was improved.

[00063] It should be noted that the overlap of inactive reagent layer 818 with active reagent layer 820 does not affect the glucose measurement as long as the enzyme from active reagent layer 820 cannot diffuse, to any significant extent in the time allowed for the measurement (i.e. about 5 seconds or less), to second working electrode 806. If enzyme were to diffuse to second working electrode 806, then first working electrode 808 would measure a glucose current in addition to the non-enzyme specific currents. This would prevent test strip 800 from effectively reducing the background signal.

[00064] It should also be noted that if the overlap of inactive reagent layer 818 with active reagent layer 820 were to occur on reference electrode 810 that this would not affect the glucose measurement. In such a case, the amount of enzyme and/or oxidized mediator on reference electrode 810 will increase, but should not affect the glucose measurement or the background correction algorithm.

[00065] Yet another embodiment of this invention which improves upon the method of coating inactive reagent layer 818 and active reagent layer 820 is shown in Figure 5 Inactive reagent layer 818 may be coated such that it completely covers second working electrode 806 and a portion of reference electrode 810. Similarly, active reagent layer 820 may be coated such that it completely covers first working electrode 808 and at least a portion of reference electrode 810. In an embodiment of this invention, the printing process can target the alignment such that inactive reagent layer 818 and active reagent layer 820 substantially overlap with each other on reference electrode 810 at an overlap zone 822. In such a case, inactive reagent layer 818 and active reagent layer 820 may mix with each other at overlap zone 822. Because the length of both inactive reagent layer 818 and active reagent layer 820 was further increased compared to the embodiment described in Figure 4, the alignment and coating of active reagent layer 820 and inactive reagent layer 818 to first working electrode 808 and second working electrode 806 was yet further improved.

[00066] It should be noted that second working electrode 806 (e.g. dummy electrode) is located on distal portion 58 of test strip 800 as illustrated in Figures 1 to 5. This causes the physiological fluid to sequentially wet in the following order - second working electrode 806, reference electrode 810, and then first working electrode 808. Test strip 800 was purposefully designed to have inactive reagent layer 818 (which contains no enzyme) upstream of active reagent layer 820 (which does contain enzyme). This reduces the possibility of enzyme being present at both second working electrode 806 and first working electrode 808. If active reagent layer 820, which contains enzyme, was coated over second working electrode 806, and no enzyme were present over first working electrode 808 then it would be possible that some enzyme could be swept to first working electrode 808 from second working electrode 806. The presence of a significant amount of enzyme on first working electrode 808 would prevent the background signal from being reduced through the use of the dummy electrode format.

[00067] In an embodiment of this invention, top layer 824 may be in the form of an integrated lance 826 as shown in Figure 1. In such an embodiment, top layer 824 may include a lance 826 which is located at distal portion 58. Lance 826, which may also be referred to as a penetration member, may be adapted to pierce a user's skin and draw blood into test strip 800 such that second working electrode 806, first working electrode 808, and reference electrode 810 are wetted. Top layer 824 is adhered to test strip 800 by adhesive layer 830. This adhesive layer 830 can be a heat seal or a pressure sensitive adhesive. Lance 826 includes a lancet base 832 that terminates at distal portion 58 of assembled test strip 800. Lance 826 may be made with either an insulating material such as plastic, glass, and silicon, or a conducting material such as stainless steel and gold. For the case in which top layer 824 is conductive, top layer 824 may also be used as a reference electrode 810 which is orientated with a facing relationship to second working electrode 806 and first working electrode 808. Further descriptions of integrated medical devices that use an integrated lance can be found in International Application No. PCT/GB01/05634 and U.S. Patent Application No. 10/143,399. In addition, lance 826 can be fabricated, for example, by a progressive die-stamping technique, as disclosed in the aforementioned International Application No. PCT/GB01/05634 and U.S. Patent Application No. 10/143,399.

[00068] In an embodiment of the present invention, adhesive layer 830 has a height of about 70 to 110 microns. Adhesive layer 830 may include a double sided pressure sensitive adhesive, a UV cured adhesive, heat activated adhesive, thermosetting plastic, or other adhesive known to those skilled in the art. As a non-limiting example, adhesive layer 830 may be formed by screen printing a pressure sensitive adhesive such as, for example, a water based acrylic copolymer pressure sensitive adhesive which is commercially available from Tape Specialties LTD in Tring, Herts, United Kingdom (part#A6435).

[00069] In a method of this invention, the background variations are reduced by subtracting a first current from first working electrode 808 from a second current from second working electrode 806. To initiate a test, a sample is applied to sample inlet 52 which allows a current to be measured at second working electrode 806 and first working electrode 808. Because second working electrode 806 does not have a glucose oxidizing enzyme disposed thereon, a magnitude of an oxidation current at second working electrode 806 is proportional to an amount of interfering compounds present on test strip 800 and also an amount of interfering compounds originating from the sample. This allows a corrected current value to be calculated using a difference between first working electrode 808 and second working electrode 806 to reduce the effects of interfering compounds present in the sample and also for interfering compounds that may be present on test strip 800.

[00070] Figure 9 is a simplified schematic showing a meter 900 interfacing with test strip 800. Meter 900 has at least three electrical contacts that form an electrical connection to second working electrode 806, first working electrode 808, and reference electrode 810. In particular second contact 812 and reference contact 816 connect to first voltage source 910; first contact 814 and reference contact 816 connect to second voltage source 920. When performing a test, first voltage source 910 applies a first potential E1 between second working electrode 806 and reference electrode 810 and second voltage source 920 applies a second potential E2 between first working electrode 808 and reference electrode 810.

[00071] In one embodiment of this invention, first potential E1 and second potential E2 may be the same such as for example about +0.4 V. In another embodiment of this invention, first potential E1 and second potential E2 may be different. A sample of blood is applied such that second working electrode 806, first working electrode 808,

and reference electrode 810 are covered with blood. This allows second working electrode 806 and first working electrode 808 to measure a current which is proportional to glucose and/or non-enzyme specific sources. After about 5 seconds from the sample application, meter 900 measures an oxidation current for both second working electrode 806 and first working electrode 808.

[00072] Figure 10 is a simplified schematic showing a meter 900 interfacing with test strip 800. In contrast to Figure 9, top layer 824 is conductive and used as a reference electrode instead of reference electrode 810 which is disposed on substrate 50. More particularly, Figure 10 shows that top layer 824, in the form of a reference electrode, has a facing relationship with first working electrode 808 and second working electrode 806. In this case, meter 900 forms an electrical contact to top layer 824 instead of at reference contact 816 as is shown in Figure 1.

[00073] Figure 21 is an exploded perspective view of a test strip according to another embodiment of the present invention. The oxidation current measured at a first working electrode 100 will be a superposition of oxidation current sources: a first, desirable, oxidation current generated by the oxidation of glucose and a second, undesirable, oxidation current generated by the interferents. The oxidation of interferents may occur directly at first working electrode 100 and indirectly through a mediated mechanism via a redox mediator.

[00074] Second working electrode 102 has a geometric trace that has an active portion 102a which is coated with active reagent 820 and an inactive portion 102i which is coated with inactive reagent 818. The oxidation current sources measured at active portion 102a will be similar to first working electrode 100. Inactive portion 102i of second working electrode 102 will oxidize interferents and not oxidize glucose because there is no enzyme present. Further, inactive portion 102i will oxidize interferents directly at second working electrode 102 and indirectly through a mediated mechanism via a redox mediator. Because the oxidation current measured at inactive portion 102i does not depend on glucose and the area of inactive portion 102i is known, it is possible to calculate its contribution to the interferent oxidation current measured at second working electrode 102. In turn, using the interferent oxidation current calculated for inactive portion 102i and knowing the area of first working electrode 100 and the area of active portion 102a, it is possible to calculate a corrected glucose current which accounts for the effects of interfering compounds

oxidized at the electrode. It should be noted that in the present invention, inactive portion 102i helps correct the glucose current for direct and mediated interference oxidation.

[00075] An algorithm may, therefore be used to calculate a corrected glucose current that is independent of interferences. After dosing a sample onto test strip 1000, a constant potential is applied to first working electrode 100 and second working electrode 102 and a current is measured for both electrodes. At first working electrode 100 where active reagent layer 820 covers the entire electrode area, the following equation can be used to describe the components contributing to the oxidation current,

$$WE_1 = G + I_{1a} \qquad \text{(Eq 1)}$$

where WE_1 is the current density at the first working electrode, G is the current density due to glucose which is independent of interferences, and I_{1a} is the current density due to interferences oxidized at first working electrode 100 which is covered with active reagent 820.

[00076] At second working electrode 102 which is partially covered with active reagent 820 and inactive reagent 818, the following equation can be used to describe the components contributing to the oxidation current,

$$WE_2 = G + I_{2a} + I_{2i} \qquad \text{(Eq 2)}$$

where WE_2 is the current density at the second working electrode, I_{2a} is the current density due to interferences at the active portion 102a, and I_{2i} is the current density due to interferences at inactive portion 102i.

[00077] To reduce the effects of interferences, an equation is formulated which describes the relationship between the interferent current at active portion 102a and inactive portion 102i. It is approximated that the interferent oxidation current density measured at active portion 102a is the same as the current density measured at the inactive portion 102i. This relationship is further described by the following equation,

$$I_{2a} = \frac{A_{2a}}{A_{2i}} \times I_{2i} \qquad \text{(Eq 3a)}$$

where A_{2a} is the area of second working electrode covered with active reagent layer 820 and A_{2i} is the area of second working electrode covered with inactive reagent layer 818.

[00078] Inactive portion 102i can oxidize interferents, but not glucose because it is not coated with enzyme. Active portion 102a can oxidize glucose and interferents. Because it was experimentally found that inactive portion 102i oxidizes interferents in a manner proportional to the area of active portion 102a, it is possible to predict the proportion of interferent current measured overall at second working electrode 102. This allows the overall current measured at second working electrode 102 (i.e. WE_2) to be corrected by subtracting the contribution of the interferent current. In an embodiment of the present invention the ratio of A_{2i}: A_{2a} may be between about 0.5:1 to 5:1, and is preferably about 3:1. More details describing this mathematical algorithm for current correction will be described in the subsequent sections.

[00079] In an alternative embodiment of the present invention, I_{2a} may be different than I_{2i}. This may be ascribed to a more efficient or less efficient oxidation of interferents at the active portion 102a because of the presence of enzyme. For not well described reasons, it is possible that the presence of enzyme may affect the electrode's ability to oxidize mediator. This behavior may be phenomenologically modeled by re-writing Equation 3a to the following form,

$$I_{2a} = f \times I_{2i} \qquad \text{(Eq 3b)}$$

where f is a correction factor which incorporates the effects of the interferent oxidation efficiency of the active portion 102a to inactive portion 120i.

[00080] In an embodiment of the present invention, Equation 1, 2, and 3a may be manipulated to derive an equation that outputs a corrected glucose current density independent of interferences. It should be noted that the three equations (Equation 1, 2, and 3a) collectively have 4 unknowns which are G, I_{2i}, I_{2a}, and I_{1a}. However, I_{1a} and I_{2a} can be conservatively assumed to be equal because they are the same

conductive material and coated with the same active reagent layer 820. Equation 1 can be rearranged to the following form.

$$G = WE_1 - I_{1a} = WE_1 - I_{2a} \qquad \text{(Eq 4)}$$

Next, I_{2a} from Equation 3a can be substituted into Equation 4 to yield Equation 5.

$$G = WE_1 - \left[\frac{A_{2a}}{A_{2i}} \times I_{2i}\right] \qquad \text{(Eq 5)}$$

Next, Equation 1 and Equation 2 can be combined to yield Equation 6.

$$I_{2i} = WE_2 - WE_1 \qquad \text{(Eq 6)}$$

Next, I_{2i} from Equation 6 can be substituted into Equation 5 to yield Equation 7a.

$$G = WE_1 - \left\{\left(\frac{A_a}{A_i}\right) X \left(WE_2 - WE_1\right)\right\} \qquad \text{(Eq 7a)}$$

[00081] Equation 7a outputs a corrected glucose current density G which removes the effects of interferences requiring only the measured current density from first working electrode 100 and second working electrode 102 (i.e. WE_1 and WE_2), and a proportion of the coated to uncoated area of the second working electrode (i.e. $\frac{A_{2a}}{A_{2i}}$). In one embodiment of the present invention the proportion $\frac{A_{2a}}{A_{2i}}$ may be programmed into a glucose meter, in, for example, a read only memory. In another embodiment of the present invention, the proportion $\frac{A_{2a}}{A_{2i}}$ may be transferred to the meter via a calibration code chip which would may account for manufacturing variations in A_{2a} or A_{2i}.

[00082] In an alternative embodiment to the present invention Equation 1, 2, and 3b may be used when the interferent oxidation current density for active portion 102a is different from the interferent oxidation current density of inactive portion 102i. In such a case, an alternative correction Equation 7b is derived as shown below.

$$G = WE_1 - \{f \times (WE_2 - WE_1)\} \qquad \text{(Eq 7b)}$$

[00083] In another embodiment of the present invention, the corrected glucose current Equation 7a or 7b may be used by the meter only when a certain threshold is exceeded. For example, if WE_2 is about 10% or greater than WE_1, then the meter would use Equation 7a or 7b to correct for the current output. However, if WE_2 is about 10% or less than WE_1, the meter would simple take an average current value between WE_1 and WE_2 to improve the accuracy and precision of the measurement. The strategy of using Equation 7a or 7b only under certain situations where it is likely that a significant level of interferences are in the sample mitigates the risk of overcorrecting the measured glucose current. It should be noted that when WE_2 is sufficiently greater than WE_1 (e.g. about 20% or more), this is an indicator of having a sufficiently high concentration of interferents. In such a case, it may be desirable to output an error message instead of a glucose value because a very high level of interferents may cause a breakdown in the accuracy of Equation 7a or 7b.

[00084] Figure 21 shows an exploded perspective view of a test strip embodiment that is designed to compensate for variations in increased background caused by the conversion of oxidized mediator to reduced mediator. Test strip 1000 includes a substrate 50, a conductive layer 164, an insulation layer 106, an inactive reagent layer 818, a active reagent layer 818, an adhesive layer 830, and a top layer 824. Test strip 1000 further includes a distal end 58 and a proximal end 60. It should be noted that test strip 1000 is a modification of test strip 800 so that an active reagent coating 820 covers a portion of both a first working electrode 100 and a second working electrode 102. This allows for two glucose measurements to be made while at the same time allows for the correction of interferents which develop within test strip 1000 or are dosed into test strip 1000. Test strip 1000 would employ either Equation 7a or 7b for reducing the effect of interfering compounds or increased background. In contrast to test strip 800, test strip 1000 has a modification to conductive layer 164 and insulation

layer 106. Substrate 50, inactive reagent layer 818, active reagent layer 818, adhesive layer 830 and top layer 824 are similar in both shape and material for both test strip 1000 and test strip 800.

[00085] Figures 22 to 24 are a simplified plane view of distal portion 58 of test strip 1000, according to the embodiment of the present invention illustrated in Figure 21, which show various positions of active reagent layer 820 and inactive reagent layer 818 with respect to each other. Figures 25 to 27, which correspond to Figures 22 to 24 respectively, do not show insulation layer 804 to help demonstrate more clearly the relationship between the conductive layer 164, active reagent layer 820, and inactive reagent layer 818.

[00086] In test strip 1000, conductive layer 164 is disposed on substrate 50. Conductive layer 164 includes a first working electrode 100, a second working electrode 102, a reference electrode 104, a first contact 101, a second contact 103, a reference contact 105, a strip detection bar 17, as shown in Figure 21. In contrast to test strip 800, second working electrode 806 and first working electrode 102 has a C-shape.

[00087] Figure 22 is a simplified plane view of first working electrode 100, second working electrode 102, and reference electrode 104, insulation layer 106, inactive reagent layer 818, and active reagent layer 818. Insulation layer 106 includes a cutout 108 which defines the area of second working electrode 102 to have an inactive portion 102i and an active portion 102a. In this embodiment, inactive reagent layer 818 was disposed on inactive portion 102i and active reagent layer 818 was disposed on active portion 102a, first working electrode 100, and reference electrode 104. Figure 22 shows that inactive reagent layer 818 does not touch or overlap with active reagent layer 818.

[00088] Test strip 1000 differs from test strip 800 in that both inactive reagent layer 818 and active reagent layer 818 both coat a portion of second working electrode 102. This allows two glucose measurements to be performed while at the same time reduce the effects of background and/or interferences. One of the challenges with making test strip 1000 as shown in Figure 22 is that it can be difficult to sequentially align and coat the respective inactive reagent layer 818 and active reagent layer 818 so that they do not touch each other with the desired yield because the area of first working

electrode 100, second working electrode 102 and reference electrode 104 is relatively small.

[00089] In an embodiment of this invention, Figure 23 shows inactive reagent layer 818 disposed on inactive portion 102i, and active reagent layer 818 disposed on active portion 102a, first working electrode 100, and reference electrode 104. In this embodiment inactive reagent layer 818 and active reagent layer 818 are immediately adjacent to each other. In such an ideal case the inactive reagent layer 818 and active reagent layer 818 would touch, but not substantially overlap with each other. Although the printing process targets the alignment such that inactive reagent layer 818 and active reagent layer 818 are immediately adjacent to each other, normal manufacturing variation will cause some overlap to occur with a certain frequency between inactive reagent layer 818 and active reagent layer 818. Likewise, such variation will also cause inactive reagent layer 818 to not touch active reagent layer 818 at a certain frequency. Because inactive reagent layer 818 was allowed to touch or not touch active reagent layer 818, the yield of acceptable test strips was improved.

[00090] Yet another embodiment of this invention which improves upon the method of coating inactive reagent layer 818 and active reagent layer 818 is shown in Figure 24. Inactive reagent layer 818 may be coated such that it completely covers inactive portion 102i and a portion of reference electrode 104. Similarly, active reagent layer 818 may be coated such that it completely covers active portion 102a, first working electrode 100 and at least a portion of reference electrode 104. In an embodiment of this invention, the printing process can target the alignment such that inactive reagent layer 818 and active reagent layer 818 substantially overlap with each other on reference electrode 810 at an overlap zone 822. In such a case, inactive reagent layer 818 and active reagent layer 818 may mix with each other at overlap zone 822. Because the length of both inactive reagent layer 818 and active reagent layer 818 was further increased compared to the embodiment described in Figure 23, the alignment and coating of active reagent layer 818 and inactive reagent layer 818 was yet further improved in terms of manufacturing yield.

[00091] It is an advantage of this invention in that two reagent layers are used which helps reduce the effects of increased background. The ability to sufficiently compensate for varying levels of reduced mediator such as ferrocyanide in the test strip itself enables a high level of accuracy and precision to be achieved. There are

several factors that may influence the conversion of oxidized mediator to the reduced form during the manufacturing, testing, and storage process. Therefore, this allows for corrections to be made which account for manufacturing variations such as reagent layer height (within batch and batch-to-batch), heat seal adhesive manufacturing conditions, high temperature drying, packaging, and sterilization conditions. Because the correction accounts for these variation, a more robust process can be envisaged in which rigorous process controls are not needed to monitor and control such manufacturing variations. The measurement of background currents may also improve the stability of test strip to withstand adverse storage conditions such as high temperature and humidity. This may allow simpler cartridges to be designed for storing test strips which may not need a rigorous seal to withhold moisture

Example 1

[00092] Test strips 800 were prepared as illustrated in Figures 1 to 3a. Test strips 800 were tested in blood which were exposed to varying levels of sterilizing radiation. To test strips 800, they were electrically connected to a potentiostat which has the means to apply a constant potential of +0.4 volts between first working electrode 808 and reference electrode 810; and second working electrode 806 and the reference electrode 810. A sample of blood is applied to sample inlet 52 allowing the blood to wick into the sample receiving chamber and to wet first working electrode 808, reference electrode 810, and second working electrode 806. Active layer 820 becomes hydrated with blood and then generates ferrocyanide which may be proportional to the amount of glucose and/or interferent concentration present in the sample. In contrast, inactive layer 818 becomes hydrated with blood and does not generate additional ferrocyanide that was not present within inactive layer 818 before hydration. After about 5 seconds from the sample application to test strip 800, an oxidation of ferrocyanide and/or interferences are measured as a current for both the first working electrode 808 and second working electrode 806.

Example 2

[00093] Two batches of test strips were prepared to show that the use of inactive reagent layer 818 and active reagent layer 820 improved the overall precision for test strips sterilized by gamma radiation. Both batches of test strips were tested in a

similar manner as described in Example 1. The first test strip batch is test strip 800 and is referred to as Batch 1. The second test strip batch, which is referred to as Batch 2, is also similar to test strip 800, but does not include inactive reagent layer 818 and also has a modified active reagent layer which covers both first working electrode 808, second working electrode 806, and reference electrode 810. When testing Batch 1, the difference in current from first working electrode 808 and second working electrode 806 was used to calculate a corrected signal current which was then converted to a glucose concentration. When testing Batch 2, the current from second working electrode 806 and first working electrode 808 were summed together to determine a value which was then used to calculate an uncorrected glucose concentration. Before testing with blood, both Batch 1 and Batch 2 test strips were treated with 0 kGy and 25 kGy of gamma radiation. Next, the four test cases, which are Batch 1- 0 kGy, Batch 1 – 25 kGy, Batch 2 – 0 kGy, and Batch 2 – 25 kGy, were evaluated for precision by testing 24 test strips with blood for each test case at 5 glucose concentrations which was 20, 50, 100, 300, and 500 mg/dL.

[00094] Figures 11 to 15 show that Batch 1 test strips did not suffer from a degradation in precision after being sterilized with 25 kGy of gamma radiation. For all five glucose concentrations, the precision was substantially similar or better after sterilization for Batch 1 test strips. This shows that the use of active reagent layer 820 and inactive reagent layer 818 helps compensate for background levels of ferrocyanide produced during the sterilization process.

[00095] Figures 11 to 13 show that Batch 2 test strips did suffer from a degradation in precision after being sterilized with 25 kGy of gamma radiation. This control experiment verifies that there is a degradation in precision when not using the background reduction method of the present invention. Because Batch 2 test strip did not have inactive reagent layer 818, the background reduction method could not be implemented. Batch 2 test strips, did not suffer from a degradation in precision after being sterilized because relatively high glucose concentrations were tested (300 and 500 mg/dL) in which the effect of sterilization on precision is not as significant. In this case, the amount of ferrocyanide generated by glucose oxidase is significantly higher than ferrocyanide generated (e.g. by sterilization processes) before hydrating the test strip.

Example 3

[00096] Another batch of test strips, which is referred to as Batch 3, was prepared in a manner similar to test strip 800 except that second working electrode 806 was not coated with either active reagent layer 820 or inactive reagent layer 818. In this example, Batches 1 to 3 were tested to evaluate the overall accuracy in the presence of interfering compounds such as uric acid and gentisic acid.

[00097] Batch 1, Batch 2, and Batch 3 test strips were tested in blood at three concentrations of gentisic acid which were 0, 25, and 50 mg/dL. For each gentisic acid concentration, two glucose concentrations were tested which were 70 and 240 mg/dL. Figures 16 and 17 show that Batch 1 and Batch 3 test strips had an insignificant change (<10 mg/dL or 10%) in bias when testing them at 25 and 50 mg/dL gentisic acid concentration. In contrast, Batch 2 test strips had a significant change (>10 mg/dL or 10%) in bias when testing them at a 25 and a 50 mg/dL gentisic acid concentration. This shows that the use of second working electrode 806 not coated with enzyme allows for an effective correction of the glucose signal in the presence of high concentrations of gentisic acid.

[00098] Batch 1, Batch 2, and Batch 3 test strips were tested in blood at three concentrations of uric acid which were 0, 10, and 20 mg/dL. For each uric acid concentration, two glucose concentrations were tested which were 70 and 240 mg/dL. Figures 18 and 19 show that Batch 1 and Batch 3 test strips had an insignificant change (<10 mg/dL or 10%) in bias when testing them at 10 and 20 mg/dL uric acid concentration. In contrast, Batch 2 test strips had a significant change (>10 mg/dL or 10%) in bias when testing them at a 10 and a 20 mg/dL uric acid concentration. This shows that the use of second working electrode 806 not coated with enzyme allows for an effective correction of the glucose signal in the presence of high concentrations of uric acid.

WHAT IS CLAIMED IS:

1. A method of reducing interferences in an electrochemical sensor comprising:

measuring a first current at a first working electrode, said first working electrode being covered by an active reagent layer;

measuring a second current at a second working electrode, said second working electrode being covered by an inactive reagent layer; and

calculating a corrected current value representative of a glucose concentration using a ratio of an active area of said first working electrode to an inactive area of said second working electrode.

2. A method of reducing interferences in an electrochemical sensor comprising:

measuring a first current at a first working electrode, said first working electrode being covered by an active reagent layer;

measuring a second current at a second working electrode, wherein said active reagent layer is disposed on an active region of said second working electrode and an inactive region of said second working electrode is covered by an inactive reagent layer; and

calculating a corrected current value representative of a glucose concentration using a ratio of an active region on said first and second working electrodes and an inactive region on said second working electrode.