

Brassica Plants Yielding Oils With A Low Total Saturated Fatty Acid Content

CROSS-REFERENCE TO RELATED APPLICATION

This application claims the benefit of U.S. Provisional Application No. 61/287,985, filed December 18, 2009, and U.S. Provisional Application No. 61/295,049, filed January 14, 2010. The contents of the foregoing applications are
5 hereby incorporated by reference in their entireties.

TECHNICAL FIELD

This invention relates to *Brassica* plants, and more particularly, *Brassica* plants having modified alleles at fatty acyl-acyl carrier protein thioesterase A2 (FATA2) loci and/or fatty acyl-acyl carrier protein thioesterase B (FATB) loci and yielding an oil with
10 a low total saturated fatty acid content in combination with a typical, mid, or high oleic acid content.

BACKGROUND

In recent years, diets high in saturated fats have been associated with increased levels of cholesterol and increased risk of coronary heart disease. As such, current
15 dietary guidelines indicate that saturated fat intake should be no more than 10 percent of total calories. Based on a 2,000-calorie-a-day diet, this is about 20 grams of saturated fat a day. While canola oil typically contains only about 7% to 8% saturated fatty acids, a decrease in its saturated fatty acid content would improve the nutritional profile of the oil.

SUMMARY

This document is based on the discovery of mutant FATA2 and FATB alleles, and
20 use of such alleles in *Brassica* plants to control total saturated fatty acid content. As described herein, *Brassica* plants containing such alleles can produce oils with a low total saturated fatty acid content (i.e., 6% or less total saturates) or oils having very low saturates (i.e., having 3.6% or less total saturates). Such *Brassica* plants also can include
25 mutant fatty acid desaturase alleles to tailor the oleic acid and α -linolenic acid content to the desired end use of the oil. *Brassica* plants described herein are particularly useful for

producing canola oils for certain food applications as the plants are not genetically modified.

This document features *Brassica* plants (e.g., *Brassica napus*, *Brassica juncea*, or *Brassica rapa* plants) and progeny thereof (e.g., seeds) that include modified alleles at two or more different fatty acyl-acyl carrier protein thioesterase B (FATB) loci (e.g., three or four different loci), wherein each modified allele results in the production of a FATB polypeptide having reduced thioesterase activity relative to a corresponding wild-type FATB polypeptide. The plant can be an F₁ hybrid. A modified allele can include a nucleic acid encoding a truncated FATB polypeptide. A modified allele can include a nucleic acid encoding a FATB polypeptide having a deletion of a helix/4-stranded sheet (4HBT) domain or a portion thereof. A modified allele can include a nucleic acid encoding a FATB polypeptide having a non-conservative substitution of a residue affecting substrate specificity. A modified allele can include a nucleic acid encoding a FATB polypeptide having a non-conservative substitution of a residue affecting catalytic activity. Any of the modified alleles can be a mutant allele.

In some embodiments, the nucleic acid encoding a truncated FATB polypeptide includes a nucleotide sequence selected from the group consisting of: SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, and SEQ ID NO:4. In some embodiments, the plant contains nucleic acids having the nucleotide sequences set forth in SEQ ID NO:1 and SEQ ID NO:2; SEQ ID NO:1 and SEQ ID NO:3; SEQ ID NO:1 and SEQ ID NO:4; SEQ ID NO:1, SEQ ID NO:2, and SEQ ID NO:3; SEQ ID NO:1, SEQ ID NO:2, and SEQ ID NO:4; SEQ ID NO:1, SEQ ID NO:3, and SEQ ID NO:4; or SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, and SEQ ID NO:4.

A plant can produce seeds yielding an oil having a total saturates content of about 2.5 to 5.5%. The palmitic acid content of the oil can be about 1.5 to 3.5%. The stearic acid content of the oil can be about 0.5 to 2.5%. The oil can have an oleic acid content of about 78 to 80%, a linoleic acid content of about 8 to 10%, and an α -linolenic acid content of no more than about 4% (e.g., about 2 to 4%).

This document also features *Brassica* plants (e.g., *Brassica napus*, *Brassica juncea*, or *Brassica rapa* plants) and progeny thereof (e.g., seeds) that include a modified allele at a fatty acyl-ACP thioesterase A2 (FATA2) locus, wherein the modified allele

results in the production of a FATA2 polypeptide (e.g., FATA2b polypeptide) having reduced thioesterase activity relative to a corresponding wild-type FATA2 polypeptide. The modified allele can include a nucleic acid encoding a FATA2 polypeptide having a mutation in a region (SEQ ID NO:29) corresponding to amino acids 242 to 277 of an *Arabidopsis* FATA2 polypeptide. The FATA2 polypeptide can include a substitution of a leucine residue for proline at position 255. The plant can be an F₁ hybrid. Any of the modified alleles can be a mutant allele.

Any of the plants described herein further can include one or more modified (e.g., mutant) alleles at FAD2 loci. For example, a mutant allele at a FAD2 loci can include a nucleic acid encoding a FAD2 polypeptide having a lysine substituted for glutamic acid in a HECGH motif. A mutant allele at a FAD2 locus can include a nucleic acid encoding a FAD2 polypeptide having a glutamic acid substituted for glycine in a DRDYGILNKV motif or a histidine substituted for leucine in a KYLNNP motif. In some embodiments, the plant contains a mutant allele at two different FAD2 loci, a mutant allele including a nucleic acid encoding a FAD2 polypeptide having a lysine substituted for glutamic acid in a HECGH motif and a mutant allele including a nucleic acid encoding a FAD2 polypeptide having a glutamic acid substituted for glycine in a DRDYGILNKV motif or a histidine substituted for leucine in a KYLNNP motif.

Any of the plants described herein further can include modified alleles (e.g., mutant alleles) at two different FAD3 loci, wherein one of the modified alleles includes a nucleic acid encoding a FAD3A polypeptide having a cysteine substituted for arginine at position 275, and wherein one of the modified alleles includes a *fad3B* nucleic acid sequence having a mutation in an exon-intron splice site recognition sequence.

In another aspect, this document features *Brassica* plants (e.g., *Brassica napus*, *Brassica juncea*, or *Brassica rapa* plants) and progeny thereof (e.g., seeds) that include modified alleles at two or more different FATB loci (e.g., 3 or 4 different FATB loci), wherein each modified allele results in production of a FATB polypeptide having reduced thioesterase activity relative to a corresponding wild-type FATB polypeptide, and further includes a modified allele at a FAD2 locus, wherein the modified allele includes a nucleic acid encoding a FAD2 polypeptide having a lysine substituted for glutamic acid in a HECGH motif. The plant further can include a modified allele at a different FAD2 locus,

the modified allele including a nucleic acid encoding a FAD2 polypeptide having a glutamic acid substituted for glycine in a DRDYGILNKV motif or a histidine substituted for leucine in a KYLNNP motif. The FATB modified allele can include a nucleic acid encoding a truncated FATB polypeptide. The nucleic acid encoding the truncated FATB polypeptide can include a nucleotide sequence selected from the group consisting of: SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, and SEQ ID NO:4. For example, the plant can contain nucleic acids having the nucleotide sequences set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, and SEQ ID NO:4. The plant can be an F₁ hybrid. Any of the modified alleles can be a mutant allele.

In another aspect, this document features a method of producing an oil. The method includes crushing seeds produced from at least one *Brassica* plant described herein; and extracting the oil from the crushed seeds, the oil having, after refining, bleaching, and deodorizing, a total saturates content of about 2.5 to 5.5%. The oil further can include an eicosenoic acid content of about 1.6 to 2.3%. The oil further can include an oleic acid content of about 78 to 80%, a linoleic acid content of about 8 to 10%, and an α -linolenic acid content of about 2 to 4%.

This document also features a method for making a *Brassica* plant. The method includes crossing one or more first *Brassica* parent plants that contain a modified allele (e.g., mutant allele) at a FATB locus and one or more second *Brassica* parent plants that contain a modified allele (e.g., mutant allele) at a different FATB locus, wherein each modified allele results in the production of a FATB polypeptide having reduced thioesterase activity relative to a corresponding wild-type FATB polypeptide; and selecting, for one to five generations, for progeny plants having modified alleles at two or more different FATB loci thereby obtaining the *Brassica* plant.

In another aspect, this document features a method for making a *Brassica* plant. The method includes obtaining one or more first *Brassica* parent plants that contain modified alleles (e.g., mutant alleles) at two or more different FATB loci (e.g., three or four different FATB loci), wherein each modified allele results in the production of a FATB polypeptide having reduced thioesterase activity relative to a corresponding wild-type FATB polypeptide; obtaining one or more second *Brassica* parent plants containing a modified allele at a FAD2 locus, the modified allele including a nucleic acid encoding a

FAD2 polypeptide having a lysine substituted for glycine in a HECGH motif; crossing the one or more first *Brassica* parent plants and the one or more second *Brassica* parent plants; and selecting, for one to five generations, for progeny plants having modified alleles at two or more different FATB loci and a modified allele at the FAD2 locus
5 thereby obtaining the *Brassica* plant. Any of the modified alleles can be a mutant allele.

The document also features a method for making a *Brassica* plant. The method includes obtaining one or more first *Brassica* parent plants that contain modified alleles (e.g., mutant alleles) at two or more different FATB loci (e.g., three or four different FATB loci), wherein each modified allele results in the production of a FATB polypeptide
10 having reduced thioesterase activity relative to a corresponding wild-type FATB polypeptide; obtaining one or more second *Brassica* parent plants containing a modified allele (e.g., mutant allele) at a FATA2 locus (e.g., FATA2b locus), the modified allele including a nucleic acid encoding a FATA2 polypeptide having a mutation in a region (SEQ ID NO:29) corresponding to amino acids 242 to 277 of the *Arabidopsis* FATA2
15 polypeptide; crossing said one or more first *Brassica* parent plants and said one or more second *Brassica* parent plants; and selecting, for one to five generations, for progeny plants having modified (e.g., mutant) alleles at two or more different FATB loci and a modified (e.g., mutant) allele at the FADA2 locus thereby obtaining the *Brassica* plant. The first *Brassica* parent plant further can contain a mutant allele at a FAD2 locus and
20 mutant alleles at two different FAD3 loci, the FAD2 mutant allele including a nucleic acid encoding a FAD2 polypeptide having a lysine substituted for glutamic acid in a HECGH motif, wherein one of the FAD3 mutant alleles contains a nucleic acid encoding a FAD3A polypeptide having a cysteine substituted for arginine at position 275, and wherein one of the FAD3 mutant alleles contains a *fad3B* nucleic acid sequence having a
25 mutation in an exon-intron splice site recognition sequence.

In yet another aspect, this document features a canola oil having an oleic acid content of about 78 to 80%, a linoleic acid content of about 8 to 10%, an α -linolenic acid content of no more than about 4%, and an eicosenoic acid content of about 1.6 to 2.3%. The palmitic acid content can be about 1.5 to 3.5%. The stearic acid content can be about
30 0.5% to 2.5%. The eicosenoic acid content can be about 1.9 to 2.2%. The α -linolenic acid content can be about 2 to about 4%.

This document also features seeds of a *Brassica* plant that include a modified allele (e.g., mutant allele) at a FATA2 locus, the modified allele (e.g., mutant allele) contains a nucleic acid encoding a FATA2 polypeptide having a mutation in a region (SEQ ID NO:29) corresponding to amino acids 242 to 277 of the polypeptide, the seeds
5 yielding an oil having an oleic acid content of 78 to 80%, a linoleic acid content of about 8 to 10%, an α -linolenic acid content of no more than about 4%, and an eicosenoic acid content of 1.6 to 2.3%. The seeds can be F₂ seeds. The *Brassica* plant further can include modified (e.g., mutant) alleles at four different FATB loci and/or a modified (e.g., mutant) allele at a FAD2 locus and modified (e.g., mutant) alleles at two different FAD3 loci, the
10 FAD2 modified (e.g., mutant) allele can include a nucleic acid encoding a FAD2 polypeptide having a lysine substituted for glutamic acid in a HECGH motif, one of the FAD3 modified (e.g., mutant) alleles can include a nucleic acid encoding a FAD3A polypeptide having a cysteine substituted for arginine at position 275, and one of the FAD3 modified (e.g., mutant) alleles can include a *fad3B* nucleic acid sequence having a
15 mutation in an exon-intron splice site recognition sequence.

This document also features a canola oil having a total saturated fatty acid content of no more than about 3.7% and an oleic acid content of about 72 to 75%. The oil can have a palmitic acid content of about 2.2 to 2.4%. The oil can have a stearic acid content of about 0.5 to 0.8%. The oil can have an eicosenoic acid content of about 1.6 to 1.9%.
20 The total saturated fatty acid content can be about 3.4 to 3.7%.

In yet another aspect, this document features a plant cell of a plant described herein, wherein the plant cell includes one or more of the modified (e.g., mutant) alleles.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this
25 invention belongs. Although methods and materials similar or equivalent to those described herein can be used to practice the invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. In addition, the materials,
30 methods, and examples are illustrative only and not intended to be limiting.

Other features and advantages of the invention will be apparent from the following detailed description, and from the claims.

DESCRIPTION OF DRAWINGS

FIG. 1 is an alignment of the nucleotides sequences of the *Brassica rapa* FatA1 (Genbank Accession No. U17098), *Arabis thaliana* FatA1 (At3g25110; Genbank Accession No. NM_113415), *B. napus* FatA1 from 15.24, *B. napus* FatA2 from 15.24, *A. thaliana* FatA2 (At4g13050; Genbank Accession No. NM_117374), and *B. napus* pNL2 (Genbank Accession No. X73849). The black boxes indicate sequence differences compared to the consensus sequence developed from the alignment; the position marked '1' highlights the SNP unique to 15.24 in the *B. napus* FatA2b isoform and shows the C to T mutation (Pro to Leu) of 15.24. The position marked as '2', highlights a SNP which distinguishes the *B. napus* FatA2a and *B. napus* FatA2b isoforms from each other (see FIG. 4).

FIG. 2 is an alignment of the FatA2 nucleotide sequence from *Arabidopsis*, 15.24, and the 01OB240 parent. At the position labeled "1," the "C" to "T" SNP is unique to BnFatA2b sequence in 15.24 germplasm (labeled 15.24FatA2(1)). At the position labeled "2", the isoform differences between *B. napus* FatA2a and *B. napus* FatA2b are apparent (15.24FatA2(2) and OB240FatA2(1) are *B. napus* FatA2a isoforms, while 15.24FatA2(1) and OB240FatA2(2) are *B. napus* FatA2b isoforms). Differences in sequence are highlighted in black.

FIG. 3 is an alignment of the amino acid sequence of residues 242 to 277 of the *A. thaliana* FatA2 (GenBank Accession No. NP_193041.1) with the *B. napus* FatA2 from 15.24 and 01OB240. The FatA2 SNP in position "1" (C to T mutation) in 15.24 causes a Pro to Leu change, while the isoform difference at position "2" does not result in an amino acid change in isoforms BnFatA2a and BnFatA2b.

FIG. 4 is an alignment of the BnFatA2 and BnFatA2b sequences from the 01OB240 and 15.24 germplasm. Position "1" refers to the "C" to "T" SNP unique to 15.24 in the BnFatA2b sequences that correlate with the low saturate phenotype. See also FIGs. 1-3. Position "2" refers to the "2" positions in Figures 1, 2, and 3, and highlights a difference in sequence between the BnFatA2a and BnFatA2b isoforms.

Black boxes represent mismatches compared to the 01OB240 BnFatA2b.

Like reference symbols in the various drawings indicate like elements.

DETAILED DESCRIPTION

In general, this document provides *Brassica* plants, including *B. napus*, *B. juncea*,
5 and *B. rapa* species of *Brassica*, that yield seeds producing oils having a low total
saturated fatty acid content (i.e., 6% or less) or having very low saturates (i.e., having
3.6% or less). As used herein, total saturated fatty acid content refers to the total of
myristic acid (C14:0), palmitic acid (C16:0), stearic acid (C18:0), arachidic acid (C20:0),
behenic acid (C22:0), and lignoceric acid (C24:0). For example, *Brassica* plants
10 described herein can produce oils having a total saturated fatty acid content of about 2.5
to 5.5%, 3 to 5%, 3 to 4.5%, 3.25 to 3.75%, 3.0 to 3.5%, 3.6 to 5%, 4 to 5.5%, or 4 to 5%.
Oils having a low or no total saturated fatty acid content have improved nutritional
quality and can help consumers reduce their intake of saturated fatty acids.

As described herein, *Brassica* plants can be made that yield seed oils having a low
15 total saturated fatty acid content in combination with a typical (60%-70%), mid (71%-
80%), or high (>80%) oleic acid content. Such *Brassica* plants can produce seed oils
having a fatty acid content tailored to the desired end use of the oil (e.g., frying or food
applications). For example, *Brassica* plants can be produced that yield seeds having a
low total saturated fatty acid content, an oleic acid content of 60% to 70%, and an α -
20 linolenic acid content of 2% to 5%. Total polyunsaturates (i.e., total of linoleic acid and
 α -linolenic acid) in such seeds typically is <35%. Canola oils having such fatty acid
contents are particularly useful for frying applications due to the polyunsaturated content,
which is low enough to have improved oxidative stability for frying yet high enough to
impart the desired fried flavor to the food being fried, and are an improvement over
25 commodity type canola oils. The fatty acid content of commodity type canola oils
typically is about 6-8% total saturated fatty acids, 55 to 65% oleic acid, about 22 to 30%
linoleic acid, and about 7-10% α -linolenic acid.

Brassica plants also can be produced that yield seeds having a low total saturated
fatty acid content, mid oleic acid content (e.g., 71% to 80% oleic acid) and a low α -
30 linolenic acid content (e.g., 2% to 5.0%). Canola oils having such fatty acid contents have

an oxidative stability that is higher than oils with a lower oleic acid content or commodity type canola oils, and are useful for coating applications (e.g., spray-coatings), formulating food products, or other applications where shelf-life stability is desired. In addition, *Brassica* plants can be produced that yield seeds having a low total saturated fatty acid content, high oleic acid content (e.g., 81% to 90% oleic acid) and an α -linolenic acid content of 2 to 5%. Canola oils having a low total saturated fatty acid content, high oleic acid, and low α -linolenic acid content are particularly useful for food applications requiring high oxidative stability and a reduced saturated fatty acid content.

Brassica Plants

Brassica plants described herein have low levels of total saturated fatty acids in the seed oil as a result of reduced activity of fatty acyl-ACP thioesterase A2 (FATA2) and/or reduced activity of fatty acyl-ACP thioesterase B (FATB). It is understood that throughout the disclosure, reference to “plant” or “plants” includes progeny, i.e., descendants of a particular plant or plant line, as well as cells or tissues from the plant. Progeny of an instant plant include seeds formed on F₁, F₂, F₃, F₄ and subsequent generation plants, or seeds formed on BC₁, BC₂, BC₃, and subsequent generation plants. Seeds produced by a plant can be grown and then selfed (or outcrossed and selfed, or doubled through dihaploid) to obtain seeds homozygous for a mutant allele. The term “allele” or “alleles” refers to one or more alternative forms of a gene at a particular locus. As used herein, a “line” is a group of plants that display little or no genetic variation between individuals for at least one trait. Such lines may be created by several generations of self-pollination and selection, or vegetative propagation from a single parent using tissue or cell culture techniques. As used herein, the term “variety” refers to a line which is used for commercial production, and includes hybrid varieties and open-pollinated varieties.

Fatty acyl-ACP thioesterases hydrolyze acyl-ACPs in the chloroplast to release the newly synthesized fatty acid from ACP, effectively removing it from further chain elongation in the plastid. The free fatty acid can then leave the plastid, become bound to CoenzymeA (CoA) and enter the Kennedy pathway in the endoplasmic reticulum (ER) for triacylglycerol (TAG) biosynthesis. Members of the FATA family prefer oleoyl

(C18:1) ACP substrates with minor activity towards 18:0 and 16:0-ACPs, while members of the FATB family hydrolyze primarily saturated acyl-ACPs between 8 and 18 carbons in length. See Jones et al., Plant Cell 7:359-371 (1995); Ginalski and Rhchlewski, Nucleic Acids Res 31:3291-3292 (2003); and Voelker T in Genetic Engineering (Setlow, JK, ed) Vol 18, 111-133, Plenum Publishing Corp., New York (2003).

Reduced activity, including absence of detectable activity, of FATA2 or FATB can be achieved by modifying an endogenous *fatA2* or *fatB* allele. An endogenous *fatA2* or *fat3B* allele can be modified by, for example, mutagenesis or by using homologous recombination to replace an endogenous plant gene with a variant containing one or more mutations (e.g., produced using site-directed mutagenesis). See, e.g., Townsend et al., Nature 459:442-445 (2009); Tovkach et al., Plant J., 57:747-757 (2009); and Lloyd et al., Proc. Natl. Acad. Sci. USA, 102:2232-2237 (2005). Similarly, for other genes discussed herein, the endogenous allele can be modified by mutagenesis or by using homologous recombination to replace an endogenous gene with a variant. Modified alleles obtained through mutagenesis are referred to as mutant alleles herein.

Reduced activity, including absence of detectable activity, can be inferred from the decreased level of saturated fatty acids in the seed oil compared with seed oil from a corresponding control plant. Reduced activity also can be assessed in plant extracts using assays for fatty acyl-ACP hydrolysis. See, for example, Bonaventure et al., Plant Cell 15:1020-1033 (2003); and Eccleston and Ohlrogge, Plant Cell 10:613-622 (1998).

Genetic mutations can be introduced within a population of seeds or regenerable plant tissue using one or more mutagenic agents. Suitable mutagenic agents include, for example, ethyl methane sulfonate (EMS), methyl N-nitrosoguanidine (MNNG), ethidium bromide, diepoxybutane, ionizing radiation, x-rays, UV rays and other mutagens known in the art. In some embodiments, a combination of mutagens, such as EMS and MNNG, can be used to induce mutagenesis. The treated population, or a subsequent generation of that population, can be screened for reduced thioesterase activity that results from the mutation, e.g., by determining the fatty acid profile of the population and comparing it to a corresponding non-mutagenized population. Mutations can be in any portion of a gene, including coding sequence, intron sequence and regulatory elements, that render the resulting gene product non-functional or with reduced activity. Suitable types of

mutations include, for example, insertions or deletions of nucleotides, and transitions or transversions in the wild-type coding sequence. Such mutations can lead to deletion or insertion of amino acids, and conservative or non-conservative amino acid substitutions in the corresponding gene product. In some embodiments, the mutation is a nonsense mutation, which results in the introduction of a stop codon (TGA, TAA, or TAG) and production of a truncated polypeptide. In some embodiments, the mutation is a splice site mutation which alters or abolishes the correct splicing of the pre-mRNA sequence, resulting in a protein of different amino acid sequence than the wild type. For example, one or more exons may be skipped during RNA splicing, resulting in a protein lacking the amino acids encoded by the skipped exons. Alternatively, the reading frame may be altered by incorrect splicing, one or more introns may be retained, alternate splice donors or acceptors may be generated, or splicing may be initiated at an alternate position, or alternative polyadenylation signals may be generated. In some embodiments, more than one mutation or more than one type of mutation is introduced.

Insertions, deletions, or substitutions of amino acids in a coding sequence may, for example, disrupt the conformation of essential alpha-helical or beta-pleated sheet regions of the resulting gene product. Amino acid insertions, deletions, or substitutions also can disrupt binding, alter substrate specificity, or disrupt catalytic sites important for gene product activity. It is known in the art that the insertion or deletion of a larger number of contiguous amino acids is more likely to render the gene product non-functional, compared to a smaller number of inserted or deleted amino acids. Non-conservative amino acid substitutions may replace an amino acid of one class with an amino acid of a different class. Non-conservative substitutions may make a substantial change in the charge or hydrophobicity of the gene product. Non-conservative amino acid substitutions may also make a substantial change in the bulk of the residue side chain, e.g., substituting an alanine residue for an isoleucine residue.

Examples of non-conservative substitutions include the substitution of a basic amino acid for a non-polar amino acid, or a polar amino acid for an acidic amino acid. Because there are only 20 amino acids encoded in a gene, substitutions that result in reduced activity may be determined by routine experimentation, incorporating amino acids of a different class in the region of the gene product targeted for mutation.

In some embodiments, a *Brassica* plant contains a mutant allele at a FATA2 locus, wherein the mutant allele results in the production of a FATA2 polypeptide having reduced thioesterase activity relative to a corresponding wild-type FATA2 polypeptide. For example, the mutant allele can include a nucleic acid that encodes a FATA2 polypeptide having a non-conservative substitution within a helix/4-stranded sheet (4HBT) domain (also referred to as a hot-dog domain) or non-conservative substitution of a residue affecting catalytic activity or substrate specificity. For example, a *Brassica* plant can contain a mutant allele that includes a nucleic acid encoding a FATA2b polypeptide having a substitution in a region (SEQ ID NO:29) of the polypeptide corresponding to residues 242 to 277 of the FATA2 polypeptide (as numbered based on the alignment to the *Arabidopsis thaliana* FATA2 polypeptide set forth in GenBank Accession No. NP_193041.1, protein (SEQ ID NO:30); GenBank Accession No. NM_117374, mRNA). This region of FATA2 is highly conserved in *Arabidopsis* and *Brassica*. In addition, many residues in this region are conserved between FATA and FATB, including the aspartic acid at position 259, asparagine at position 263, histidine at position 265, valine at position 266, asparagine at position 268, and tyrosine at position 271 (as numbered based on the alignment to SEQ ID NO:30). See also FIG. 3. The asparagine at position 263 and histidine at position 265 are part of the catalytic triad, and the arginine at position 256 is involved in determining substrate specificity. See also Mayer and Shanklin, *BMC Plant Biology* 7:1-11 (2007). SEQ ID NO:31 sets forth the predicted amino acid sequence of the *Brassica* FATA2b polypeptide encoded by exons 2-6, and corresponding to residues 121 to 343 of the *A. thaliana* sequence set forth in SEQ ID NO:30. For example, the FATA2 polypeptide can have a substitution of a leucine residue for proline at the position corresponding to position 255 of the *Arabidopsis* FATA2 polypeptide (i.e., position 14 of SEQ ID NO:29 or position 135 of SEQ ID NO:31). The proline in the *B. napus* sequence corresponding to position 255 in *Arabidopsis* is conserved among *B. napus*, *B. rapa*, *B. juncea*, *Zea mays*, *Sorghum bicolor*, *Oryza sativa* Indica (rice), *Triticum aestivum*, *Glycine max*, *Jatropha* (tree species), *Carthamus tinctorius*, *Cuphea hookeriana*, *Iris tectorum*, *Perilla frutescens*, *Helianthus annuus*, *Garcinia mangostana*, *Picea sitchensis*, *Physcomitrella patens* subsp. *Patens*, *Elaeis guineensis*, *Vitis vinifera*, *Elaeis oleifera*, *Camellia oleifera*, *Arachis*

hypogaea, *Capsicum annuum*, *Cuphea hookeriana*, *Populus trichocarpa*, and *Diploknema butyracea*. As described in Example 2, the mutation at position 255 is associated with a low total saturated fatty acid phenotype, low stearic acid phenotype, low arachidic acid phenotype, and an increased eicosenoic acid phenotype. The stearic acid content phenotype is negatively correlated with the eicosenoic acid phenotype.

In some embodiments, the mutant allele at a FATA2 locus includes a nucleotide sequence having at least 90% (e.g., at least 91, 92, 93, 94, 95, 96, 97, 98, or 99%) sequence identity to the nucleotide sequence set forth in SEQ ID NO:28 or SEQ ID NO:32. The nucleotide sequences set forth in SEQ ID NOs:28 and 32 are representative nucleotide sequences from the *fatA2b* gene from *B. napus* line 15.24. As used herein, the term “sequence identity” refers to the degree of similarity between any given nucleic acid sequence and a target nucleic acid sequence. The degree of similarity is represented as percent sequence identity. Percent sequence identity is calculated by determining the number of matched positions in aligned nucleic acid sequences, dividing the number of matched positions by the total number of aligned nucleotides, and multiplying by 100. A matched position refers to a position in which identical nucleotides occur at the same position in aligned nucleic acid sequences. Percent sequence identity also can be determined for any amino acid sequence. To determine percent sequence identity, a target nucleic acid or amino acid sequence is compared to the identified nucleic acid or amino acid sequence using the BLAST 2 Sequences (Bl2seq) program from the stand-alone version of BLASTZ containing BLASTN version 2.0.14 and BLASTP version 2.0.14. This stand-alone version of BLASTZ can be obtained from Fish & Richardson’s web site (World Wide Web at “fr” dot “com” slash “blast”) or the U.S. government’s National Center for Biotechnology Information web site (World Wide Web at “ncbi” dot “nlm” dot “nih” dot “gov”). Instructions explaining how to use the Bl2seq program can be found in the readme file accompanying BLASTZ.

Bl2seq performs a comparison between two sequences using either the BLASTN or BLASTP algorithm. BLASTN is used to compare nucleic acid sequences, while BLASTP is used to compare amino acid sequences. To compare two nucleic acid sequences, the options are set as follows: -i is set to a file containing the first nucleic acid sequence to be compared (e.g., C:\seq1.txt); -j is set to a file containing the second

nucleic acid sequence to be compared (e.g., C:\seq2.txt); -p is set to blastn; -o is set to any desired file name (e.g., C:\output.txt); -q is set to -1; -r is set to 2; and all other options are left at their default setting. The following command will generate an output file containing a comparison between two sequences: C:\Bl2seq -i c:\seq1.txt -j
5 c:\seq2.txt -p blastn -o c:\output.txt -q -1 -r 2. If the target sequence shares homology with any portion of the identified sequence, then the designated output file will present those regions of homology as aligned sequences. If the target sequence does not share homology with any portion of the identified sequence, then the designated output file will not present aligned sequences.

10 Once aligned, a length is determined by counting the number of consecutive nucleotides from the target sequence presented in alignment with sequence from the identified sequence starting with any matched position and ending with any other matched position. A matched position is any position where an identical nucleotide is presented in both the target and identified sequence. Gaps presented in the target
15 sequence are not counted since gaps are not nucleotides. Likewise, gaps presented in the identified sequence are not counted since target sequence nucleotides are counted, not nucleotides from the identified sequence.

The percent identity over a particular length is determined by counting the number of matched positions over that length and dividing that number by the length
20 followed by multiplying the resulting value by 100. For example, if (i) a 500-base nucleic acid target sequence is compared to a subject nucleic acid sequence, (ii) the Bl2seq program presents 200 bases from the target sequence aligned with a region of the subject sequence where the first and last bases of that 200-base region are matches, and
(iii) the number of matches over those 200 aligned bases is 180, then the 500-base nucleic
25 acid target sequence contains a length of 200 and a sequence identity over that length of 90 % (i.e., $180 / 200 \times 100 = 90$).

It will be appreciated that different regions within a single nucleic acid target sequence that aligns with an identified sequence can each have their own percent identity. It is noted that the percent identity value is rounded to the nearest tenth. For example, 78.11, 78.12, 78.13, and 78.14 are rounded down to 78.1, while 78.15, 78.16, 78.17, 78.18, and 78.19 are rounded up to 78.2. It also is noted that the length value will always be an integer.

In some embodiments, a *Brassica* plant contains a mutant allele at a FATB locus, wherein the mutant allele results in the production of a FATB polypeptide having reduced thioesterase activity relative to a corresponding wild-type FATB polypeptide. In some embodiments, a *Brassica* plant contains mutant alleles at two or more different FATB loci. In some embodiments, a *Brassica* plant contains mutant alleles at three different FATB loci or contains mutant alleles at four different FATB loci. *Brassica napus* contains 6 different FATB isoforms (i.e., different forms of the FATB polypeptide at different loci), which are called isoforms 1-6 herein. SEQ ID NOs:18-21 and 26-27 set forth the nucleotide sequences encoding FATB isoforms 1-6, respectively, of *Brassica napus*. The nucleotide sequences set forth in SEQ ID NOs:18-21 and 26-27 have 82% to 95% sequence identity as measured by the ClustalW algorithm.

For example, a *Brassica* plant can have a mutation in a nucleotide sequence encoding FATB isoform 1, isoform 2, isoform 3, isoform 4, isoform 5, or isoform 6. In some embodiments, a plant can have a mutation in a nucleotide sequence encoding isoforms 1 and 2; 1 and 3; 1 and 4; 1 and 5; 1 and 6; 2 and 3; 2 and 4; 2 and 5; 2 and 6; 3 and 4; 3 and 5; 3 and 6; 4 and 5; 4 and 6; 5 and 6; 1, 2, and 3; 1, 2, and 4; 1, 2, and 5; 1, 2, and 6; 2, 3, and 4; 2, 3, and 5; 2, 3, and 6; 3, 4, and 5; 3, 5, and 6; 4, 5, and 6; 1, 2, 3, and 4; 1, 2, 3, and 5; 1, 2, 3, and 6; 1, 2, 4, and 6; 1, 3, 4 and 5; 1, 3, 4, and 6; 1, 4, 5, and 6; 2, 3, 4, and 5; 2, 3, 4 and 6; or 3, 4, 5, and 6. In some embodiments, a *Brassica* plant can have a mutation in nucleotide sequences encoding FATB isoforms 1, 2, and 3; 1, 2, and 4; 2, 3, and 4; or 1, 2, 3, and 4. In some embodiments, a mutation results in deletion of a 4HBT domain or a portion thereof of a FATB polypeptide. FATB polypeptides typically contain a tandem repeat of the 4HBT domain, where the N-terminal 4HBT domain contains residues affecting substrate specificity (e.g., two conserved methionines, a conserved lysine, a conserved valine, and a conserved serine) and the C-terminal 4HBT

domain contains residues affecting catalytic activity (e.g., a catalytic triad of a conserved asparagine, a conserved histidine, and a conserved cysteine) and substrate specificity (e.g., a conserved tryptophan). See Mayer and Shanklin, J. Biol. Chem. 280:3621-3627 (2005). In some embodiments, the mutation results in a non-conservative substitution of a residue in a 4HBT domain or a residue affecting substrate specificity. In some
5 embodiments, the mutation is a splice site mutation. In some embodiment, the mutation is a nonsense mutation in which a premature stop codon (TGA, TAA, or TAG) is introduced, resulting in the production of a truncated polypeptide.

SEQ ID NOs:1-4 set forth the nucleotide sequences encoding isoforms 1-4,
10 respectively, and containing exemplary nonsense mutations that result in truncated FATB polypeptides. SEQ ID NO:1 is the nucleotide sequence of isoform 1 having a mutation at position 154, which changes the codon from CAG to TAG. SEQ ID NO:2 is the nucleotide sequence of isoform 2 having a mutation at position 695, which changes the codon from CAG to TAG. SEQ ID NO:3 is the nucleotide sequence of isoform 3 having
15 a mutation at position 276, which changes the codon from TGG to TGA. SEQ ID NO:4 is the nucleotide sequence of isoform 4 having a mutation at position 336, which changes the codon from TGG to TGA.

Two or more different mutant FATB alleles may be combined in a plant by making a genetic cross between mutant lines. For example, a plant having a mutant allele
20 at a FATB locus encoding isoform 1 can be crossed or mated with a second plant having a mutant allele at a FATB locus encoding isoform 2. Seeds produced from the cross are planted and the resulting plants are selfed in order to obtain progeny seeds. These progeny seeds can be screened in order to identify those seeds carrying both mutant alleles. In some embodiments, progeny are selected over multiple generations (e.g., 2 to
25 5 generations) to obtain plants having mutant alleles at two different FATB loci. Similarly, a plant having mutant alleles at two or more different FATB isoforms can be crossed with a second plant having mutant alleles at two or more different FATB alleles, and progeny seeds can be screened to identify those seeds carrying mutant alleles at four or more different FATB loci. Again, progeny can be selected for multiple generations to
30 obtain the desired plant.

In some embodiments, a mutant allele at a FATA2 locus and mutant alleles at two or more (e.g., three or four) different FATB loci can be combined in a plant. For example, a plant having a mutant allele at a FATA2 locus can be crossed or mated with a second plant having mutant alleles at two or more different FATB loci. Seeds produced from the cross are planted and the resulting plants are selfed in order to obtain progeny seeds. These progeny seeds can be screened in order to identify those seeds carrying mutant FATA2 and FATB alleles. Progeny can be selected over multiple generations (e.g., 2 to 5 generations) to obtain plants having a mutant allele at a FATA2 locus and mutant alleles at two or more different FATB loci. As described herein, plants having a mutant allele at a FATA2b locus and mutant alleles at three or four different FATB loci have a low total saturated fatty acid content that is stable over different growing conditions, i.e., is less subject to variation due to warmer or colder temperatures during the growing season. Due to the differing substrate profiles of the FatB and FatA enzymes with respect to 16:0 and 18:0, respectively, plants having mutations in FatA2 and FatB loci exhibit a substantial reduction in amounts of both 16:0 and 18:0 in seed oil.

Brassica plants having mutant alleles at FATA2 and/or FATB loci also can include mutant alleles at loci controlling fatty acid desaturase activity such that the oleic acid and linolenic acid levels can be tailored to the end use of the oil. For example, such *Brassica* plants also can exhibit reduced activity of delta-15 desaturase (also known as FAD3), which is involved in the enzymatic conversion of linoleic acid to α -linolenic acid. The gene encoding delta-15 fatty acid desaturase is referred to as *fad3* in *Brassica* and *Arabidopsis*. Sequences of higher plant *fad3* genes are disclosed in Yadav et al., Plant Physiol., 103:467-476 (1993), WO 93/11245, and Arondel et al., Science, 258:1353-1355 (1992). Decreased activity, including absence of detectable activity, of delta-15 desaturase can be achieved by mutagenesis. Decreased activity, including absence of detectable activity, can be inferred from the decreased level of linolenic acid (product) and in some cases, increased level of linoleic acid (the substrate) in the plant compared with a corresponding control plant. For example, parent plants can contain the mutation from the APOLLO or STELLAR *B. napus* variety that confers low linolenic acid. The STELLAR and APOLLO varieties were developed at the University of Manitoba (Manitoba, Canada). In some embodiments, the parents contain the *fad3A* and/or *fad3B*

mutation from IMC02 that confer a low linolenic acid phenotype. IMC02 contains a mutation in both the *fad3A* and *fad3B* genes. The *fad3A* gene contains a C to T mutation at position 2565, numbered from the ATG in genomic DNA, resulting in the substitution of a cysteine for arginine at position 275 of the encoded FAD3A polypeptide. The *fad3B* gene contains a G to A mutation at position 3053 from ATG in genomic DNA, located in the exon-intron splice site recognition sequence. IMC02 was obtained from a cross of IMC01 x Westar. See Example 3 of U.S. Patent No. 5,750,827. IMC01 was deposited with the American Type Culture Collection (ATCC) under Accession No. 40579. IMC02 was deposited with the ATCC under Accession No. PTA-6221.

In some embodiments, a *Brassica* plant contains a mutant allele at a FATA2 locus and a mutant allele at a FAD3 locus. For example, a *Brassica* plant can contain a mutant allele at a FATA2 locus and a mutant allele at a FAD3 locus that contains a nucleic acid encoding a FAD3 polypeptide with a cysteine substituted for arginine at position 275 and/or a nucleic acid having a mutation in an exon-intron splice site recognition sequence. A *Brassica* plant also can contain mutant alleles at two or more different FATB loci (three or four different loci) and a FAD3 locus that contains a nucleic acid encoding a FAD3 polypeptide with a cysteine substituted for arginine at position 275 and/or a nucleic acid having a mutation in an exon-intron splice site recognition sequence. A *Brassica* plant also contain a mutant allele at a FATA2 locus, mutant alleles at two or more different FATB loci (three or four different loci) and a FAD3 locus that contains a nucleic acid encoding a FAD3 polypeptide with a cysteine substituted for arginine at position 275 and/or a nucleic acid having a mutation in an exon-intron splice site recognition sequence.

Brassica plants also can have decreased activity of a delta-12 desaturase, which is involved in the enzymatic conversion of oleic acid to linoleic acid, to confer a mid or high oleic acid content in the seed oil. *Brassica* plants can exhibit reduced activity of delta-12 desaturase (also known as FAD2) in combination with reduced activity of FATA2 and/or FATB. The sequences for the wild-type *fad2* genes from *B. napus* (termed the D form and the F form) are disclosed in WO 98/56239. A reduction in delta-12 desaturase activity, including absence of detectable activity, can be achieved by mutagenesis. Decreased delta-12 desaturase activity can be inferred from the decrease

level of linoleic acid (product) and increased level of oleic acid (substrate) in the plant compared with a corresponding control plant. Non-limiting examples of suitable *fad2* mutations include the G to A mutation at nucleotide 316 within the *fad2-D* gene, which results in the substitution of a lysine residue for glutamic acid in a HECGH (SEQ ID NO:5) motif. Such a mutation is found within the variety IMC129, which has been deposited with the ATCC under Accession No. 40811. Another suitable *fad2* mutation can be the T to A mutation at nucleotide 515 of the *fad2-F* gene, which results in the substitution of a histidine residue for leucine in a KYLNNP (SEQ ID NO:6) motif (amino acid 172 of the Fad2 F polypeptide). Such a mutation is found within the variety Q508. See U.S. Patent No. 6,342,658. Another example of a *fad2* mutation is the G to A mutation at nucleotide 908 of the *fad2-F* gene, which results in the substitution of a glutamic acid for glycine in the DRDYGILNKV (SEQ ID NO:7) motif (amino acid 303 of the Fad2 F polypeptide). Such a mutation is found within the variety Q4275, which has been deposited with the ATCC under Accession No. 97569. See U.S. Patent No. 6,342,658. Another example of a suitable *fad2* mutation can be the C to T mutation at nucleotide 1001 of the *fad2-F* gene (as numbered from the ATG), which results in the substitution of an isoleucine for threonine (amino acid 334 of the Fad2 F polypeptide). Such a mutation is found within the high oleic acid variety Q7415.

Typically, the presence of one of the *fad2-D* or *fad2-F* mutations confers a mid-oleic acid phenotype (e.g., 70-80% oleic acid) to the seed oil, while the presence of both *fad2-D* and *fad2-F* mutations confers a higher oleic acid phenotype (e.g., >80% oleic acid). For example, Q4275 contains the *fad2-D* mutation from IMC129 and a *fad2-F* mutation at amino acid 303. Q508 contains *fad2-D* mutation from IMC129 and a *fad2-F* mutation at amino acid 172. Q7415 contains the *fad2-D* mutation from IMC129 and a *fad2-F* mutation at amino acid 334. The presence of both *fad2* mutations in Q4275, Q508, and Q7415 confers a high oleic acid phenotype of greater than 80% oleic acid.

Thus, in some embodiments, a *Brassica* plant contains a mutant allele at a FATA2 locus (e.g., FATA2b locus) and a mutant allele at a FAD2 locus. For example, a *Brassica* plant can contain a mutant allele at a FATA2 locus and a mutant allele at a FAD2 locus described above. A *Brassica* plant also can contain mutant alleles at two or more different FATB loci (three or four different loci) and a FAD2 locus described

above. A *Brassica* plant can also contain a mutant allele at a FATA2 locus, mutant alleles at two or more different FATB loci (three or four different loci) and a mutant allele at a FAD2 locus described above. In some embodiments, a *Brassica* plant contains a mutant allele at a FATA2 locus, a mutant allele at a FAD2 locus, and a mutant allele at a FAD3 locus described above. A *Brassica* plant also can contain mutant alleles at two or more different FATB loci (three or four different loci), mutant alleles at FAD2 loci, and mutant alleles at FAD3 loci described above. A *Brassica* plant also contain a mutant allele at a FATA2 locus, mutant alleles at two or more different FATB loci (three or four different loci), mutant alleles at FAD2 loci, and mutant alleles at FAD3 loci described above.

Production of Hybrid Brassica Varieties

Hybrid *Brassica* varieties can be produced by preventing self-pollination of female parent plants (i.e., seed parents), permitting pollen from male parent plants to fertilize such female parent plants, and allowing F₁ hybrid seeds to form on the female plants. Self-pollination of female plants can be prevented by emasculating the flowers at an early stage of flower development. Alternatively, pollen formation can be prevented on the female parent plants using a form of male sterility. For example, male sterility can be cytoplasmic male sterility (CMS), nuclear male sterility, molecular male sterility wherein a transgene inhibits microsporogenesis and/or pollen formation, or be produced by self-incompatibility. Female parent plants containing CMS are particularly useful. CMS can be, for example of the *ogu* (Ogura), *nap*, *pol*, *tour*, or *mur* type. See, for example, Pellan-Delourme and Renard, 1987, Proc. 7th Int. Rapeseed Conf., Poznan, Poland, p. 199-203 and Pellan-Delourme and Renard, 1988, Genome 30:234-238, for a description of Ogura type CMS. See, Riungu and McVetty, 2003, Can. J. Plant Sci., 83:261-269 for a description of *nap*, *pol*, *tour*, and *mur* type CMS.

In embodiments in which the female parent plants are CMS, the male parent plants typically contain a fertility restorer gene to ensure that the F₁ hybrids are fertile. For example, when the female parent contains an Ogura type CMS, a male parent is used that contains a fertility restorer gene that can overcome the Ogura type CMS. Non-limiting examples of such fertility restorer genes include the Kosena type fertility restorer

gene (U.S. Patent No. 5,644,066) and Ogura fertility restorer genes (U.S. Patent Nos. 6,229,072 and 6,392,127). In other embodiments in which the female parents are CMS, male parents can be used that do not contain a fertility restorer. F₁ hybrids produced from such parents are male sterile. Male sterile hybrid seed can be inter-planted with male fertile seed to provide pollen for seed-set on the resulting male sterile plants.

The methods described herein can be used to form single-cross *Brassica* F₁ hybrids. In such embodiments, the parent plants can be grown as substantially homogeneous adjoining populations to facilitate natural cross-pollination from the male parent plants to the female parent plants. The F₁ seed formed on the female parent plants is selectively harvested by conventional means. One also can grow the two parent plants in bulk and harvest a blend of F₁ hybrid seed formed on the female parent and seed formed upon the male parent as the result of self-pollination. Alternatively, three-way crosses can be carried out wherein a single-cross F₁ hybrid is used as a female parent and is crossed with a different male parent that satisfies the fatty acid parameters for the female parent of the first cross. Here, assuming a bulk planting, the overall oleic acid content of the vegetable oil may be reduced over that of a single-cross hybrid; however, the seed yield will be further enhanced in view of the good agronomic performance of both parents when making the second cross. As another alternative, double-cross hybrids can be created wherein the F₁ progeny of two different single-crosses are themselves crossed. Self-incompatibility can be used to particular advantage to prevent self-pollination of female parents when forming a double-cross hybrid.

Hybrids described herein have good agronomic properties and exhibit hybrid vigor, which results in seed yields that exceed that of either parent used in the formation of the F₁ hybrid. For example, yield can be at least 10% (e.g., 10% to 20%, 10% to 15%, 15% to 20%, or 25% to 35%) above that of either one or both parents. In some embodiments, the yield exceeds that of open-pollinated spring canola varieties such as 46A65 (Pioneer) or Q2 (University of Alberta), when grown under similar growing conditions. For example, yield can be at least 10% (e.g., 10% to 15% or 15% to 20%) above that of an open-pollinated variety.

Hybrids described herein typically produce seeds having very low levels of glucosinolates (<30 µmol/gram of de-fatted meal at a moisture content of 8.5%). In

particular, hybrids can produce seeds having <20 μmol of glucosinolates/gram of de-fatted meal. As such, hybrids can incorporate mutations that confer low glucosinolate levels. See, for example, U.S. Patent No. 5,866,762. Glucosinolate levels can be determined in accordance with known techniques, including high performance liquid chromatography (HPLC), as described in ISO 9167-1:1992(E), for quantification of total, intact glucosinolates, and gas-liquid chromatography for quantification of trimethylsilyl (TMS) derivatives of extracted and purified desulfoglucosinolates. Both the HPLC and TMS methods for determining glucosinolate levels analyze de-fatted or oil-free meal.

Canola Oil

Brassica plants disclosed herein are useful for producing canola oils with low or no total saturated fatty acids. For example, oil obtained from seeds of *Brassica* plants described herein may have a total saturated fatty acid content of about 2.5 to 5.5%, 3 to 5%, 3 to 4.5%, 3.25 to 3.75%, 3.0 to 3.5%, 3.4 to 3.7%, 3.6 to 5%, 4 to 5.5%, 4 to 5%, or 4.25 to 5.25%. In some embodiments, an oil has a total saturated fatty acid content of about 4 to about 5.5%, an oleic acid content of about 60 to 70% (e.g., 62 to 68%, 63 to 67%, or 65 to 66%), and an α -linolenic acid content of about 2.5 to 5%. In some embodiments, an oil has a total saturated fatty acid content of about 2.5 to 5.5% (e.g., 4 to 5%), an oleic acid content of about 71 to 80% (e.g., 72 to 78%, 73 to 75%, 74 to 78%, or 75 to 80%) and an α -linolenic acid content of about 2 to 5.0% (e.g., 2.0 to 2.8%, 2.25 to 3%, 2.5 to 3%, 3 to 3.5%, 3.25% to 3.75%, 3.5 to 4%, 3.75 to 4.25%, 4 to 4.5%, 4.25 to 4.75%, 4.5 to 5%). In some embodiments, a canola oil can have a total saturated fatty acid content of 2.5 to 5.5%, an oleic acid content of 78 to 80%, and an α -linolenic acid content of no more than 4% (e.g., 2 to 4%). In some embodiments, an oil has a total saturated fatty acid content of about 3.5 to 5.5% (e.g., 4 to 5%), an oleic acid content of about 81 to 90% (e.g., 82 to 88% or 83 to 87% oleic acid) and an α -linolenic acid content of 2 to 5% (e.g., 2 to 3% or 3 to 5%). In some embodiments, an oil has a total saturated fatty acid content of no more than 3.7% (e.g., about 3.4 to 3.7% or 3.4 to 3.6%) and an oleic acid content of about 72 to 75%.

Low saturate oils described herein can have a palmitic acid content of about 1.5 to 3.5% (e.g., 2 to 3% or 2.2 to 2.4%). The stearic acid content of such oils can be about 0.5 to 2.5% (e.g., 0.5 to 0.8%, 1 to 2%, or 1.5 to 2.5%).

Oils described herein can have an eicosenoic acid content greater than 1.6%, e.g., 1.6 to 1.9%, 1.7 to 2.3%, 1.8 to 2.3%, or 1.9 to 2.3%, in addition to a low total saturates content.

Oils described herein can have a linoleic acid content of about 3 to 20%, e.g., 3.4 to 5%, 3.75 to 5%, 8 to 10%, 10 to 12%, 11 to 13%, 13 to 16%, or 14 to 18%, in addition to a low total saturates content.

Oils described herein have an erucic acid content of less than 2% (e.g., less than 1%, 0.5%, 0.2, or 0.1%) in additions to a low total saturates content.

The fatty acid composition of seeds can be determined by first crushing and extracting oil from seed samples (e.g., bulk seeds samples of 10 or more seeds). TAGs in the seed are hydrolyzed to produce free fatty acids, which then can be converted to fatty acid methyl esters and analyzed using techniques known to the skilled artisan, e.g., gas-liquid chromatography (GLC) according to AOCS Procedure Ce 1e-91. Near infrared (NIR) analysis can be performed on whole seed according to AOCS Procedure Am-192 (revised 1999)

Seeds harvested from plants described herein can be used to make a crude canola oil or a refined, bleached, and deodorized (RBD) canola oil with a low or no total saturated fatty acid content. Harvested canola seed can be crushed by techniques known in the art. The seed can be tempered by spraying the seed with water to raise the moisture to, for example, 8.5%. The tempered seed can be flaked using smooth roller with, for example, a gap setting of 0.23 to 0.27 mm. Heat may be applied to the flakes to deactivate enzymes, facilitate further cell rupturing, coalesce the oil droplets, or agglomerate protein particles in order to ease the extraction process. Typically, oil is removed from the heated canola flakes by a screw press to press out a major fraction of the oil from the flakes. The resulting press cake contains some residual oil.

Crude oil produced from the pressing operation typically is passed through a settling tank with a slotted wire drainage top to remove the solids expressed out with the oil in the screw pressing operation. The clarified oil can be passed through a plate and

frame filter to remove the remaining fine solid particles. Canola press cake produced from the screw pressing operation can be extracted with commercial n-Hexane. The canola oil recovered from the extraction process is combined with the clarified oil from the screw pressing operation, resulting in a blended crude oil.

5 Free fatty acids and gums typically are removed from the crude oil by heating in a batch refining tank to which food grade phosphoric acid has been added. The acid serves to convert the non-hydratable phosphatides to a hydratable form, and to chelate minor metals that are present in the crude oil. The phosphatides and the metal salts are removed from the oil along with the soapstock. The oil-acid mixture is treated with sodium
10 hydroxide solution to neutralize the free fatty acids and the phosphoric acid in the acid-oil mixture. The neutralized free fatty acids, phosphatides, etc. (soapstock) are drained off from the neutralized oil. A water wash may be done to further reduce the soap content of the oil. The oil may be bleached and deodorized before use, if desired, by techniques known in the art.

15 Oils obtained from plant described herein can have increased oxidative stability, which can be measured using, for example, an Oxidative Stability Index Instrument (e.g., from Omnion, Inc., Rockland, MA) according to AOCS Official Method Cd 12b-92 (revised 1993). Oxidative stability is often expressed in terms of "AOM" hours.

20 *Food Compositions*

This document also features food compositions containing the oils described above. For example, oils having a low (6% or less) or no (3.5% or less) total saturated fatty acid content in combination with a typical (60-70%), mid (71-80%), or high (>80%) oleic acid content can be used to replace or reduce the amount of saturated fatty acids and
25 hydrogenated oils (e.g., partially hydrogenated oils) in various food products such that the levels of saturated fatty acids and *trans* fatty acids are reduced in the food products. In particular, canola oils having a low total saturated fatty acid content and a mid or high oleic acid content in combination with a low linolenic acid content can be used to replace or reduce the amount of saturated fats and partially hydrogenated oils in processed or
30 packaged food products, including bakery products such as cookies, muffins, doughnuts, pastries (e.g., toaster pastries), pie fillings, pie crusts, pizza crusts, frostings, breads,

biscuits, and cakes, breakfast cereals, breakfast bars, puddings, and crackers.

For example, an oil described herein can be used to produce sandwich cookies that contain reduced saturated fatty acids and no or reduced levels of partially hydrogenated oils in the cookie and/or cr me filling. Such a cookie composition can include, for example, in addition to canola oil, flour, sweetener (e.g., sugar, molasses, honey, high fructose corn syrup, artificial sweetener such as sucralose, saccharine, aspartame, or acesulfame potassium, and combinations thereof), eggs, salt, flavorants (e.g., chocolate, vanilla, or lemon), a leavening agent (e.g., sodium bicarbonate or other baking acid such as monocalcium phosphate monohydrate, sodium aluminum sulfate, sodium acid pyrophosphate, sodium aluminum phosphate, dicalcium phosphate, glucanodelta lactone, or potassium hydrogen tartrate, or combinations thereof), and optionally, an emulsifier (e.g., mono- and diglycerides of fatty acids, propylene glycol mono- and diesters of fatty acids, glycerol-lactose esters of fatty acids, ethoxylated or succinylated mono- and diglycerides, lecithin, diacetyl tartaric acid esters or mono- and diglycerides, sucrose esters of glycerol, and combinations thereof). A cr me filling composition can include, in addition to canola oil, sweetener (e.g., powdered sugar, granulated sugar, honey, high fructose corn syrup, artificial sweetener, or combinations thereof), flavorant (e.g., vanilla, chocolate, or lemon), salt, and, optionally, emulsifier.

Canola oils (e.g., with low total saturated fatty acid content, low oleic acid, and low linolenic acid content) also are useful for frying applications due to the polyunsaturated content, which is low enough to have improved oxidative stability for frying yet high enough to impart the desired fried flavor to the food being fried. For example, canola oils can be used to produce fried foods such as snack chips (e.g., corn or potato chips), French fries, or other quick serve foods.

Oils described herein also can be used to formulate spray coatings for food products (e.g., cereals or snacks such as crackers). In some embodiments, the spray coating can include other vegetable oils such as sunflower, cottonseed, corn, or soybean oils. A spray coating also can include an antioxidant and/or a seasoning.

Oils described herein also can be use in the manufacturing of dressings, mayonnaises, and sauces to provide a reduction in the total saturated fat content of the product. The low saturate oil can be used as a base oil for creating structured fat

solutions such as microwave popcorn solid fats or canola butter formulations.

The invention will be further described in the following examples, which do not limit the scope of the invention described in the claims.

EXAMPLES

In the Tables described herein, the fatty acids are referred to by the length of the carbon chain and number of double bonds within the chain. For example, C140 refers to C14:0 or myristic acid; C160 refers to C16:0 or palmitic acid; C180 refers to C18:0 or stearic acid; C181 refers to C18:1 or oleic acid; C182 refers to C18:2 or linoleic acid; C183 refers to C18:3 or α -linolenic acid; C200 refers to C20:0 or arachidic acid; C201 refers to C20:1 or eicosenoic acid, C220 refers to C22:0 or behenic acid, C221 refers to C22:1 or erucic acid, C240 refers to C24:0 or lignoceric acid, and C241 refers to C24:1 or nervonic acid. "Total Sats" refers to the total of C140, C160, C180, C200, C220, and C240. Representative fatty acid profiles are provided for each of the specified samples.

Unless otherwise indicated, all percentages refer to wt % based on total wt% of fatty acids in the oil.

EXAMPLE 1

Brassica plant line 15.24

Plants producing an oil with a high oleic acid and low total saturated fatty acid content were obtained from crosses of plants designated 90A24 and plants designated 90I22. 90A24 plants were obtained from a cross between HIO 11-5, a high oleic acid selection from the IMC 129 lineage (ATCC Deposit No. 40811; U.S. Patent No. 5,863,589), and LS 6-5, a low saturated fatty acid selection from the IMC 144 lineage (ATCC Deposit No. 40813; U.S. Patent No. 5,668,299). 90I22 plants were obtained from a cross between LS 4-3, a low saturated fatty acid selection from the IMC 144 lineage (ATCC Deposit No. 40813) and D336, a low α -linolenic acid selection from the IMC 01 lineage (ATCC Deposit No. 40579; U.S. Patent No. 5,750,827). Table 1 contains the fatty acid profile for the LS6-5, LS4-3, and HIO 11-5 parent lines, as well as IMC 01.

TABLE 1
Seed Fatty Acid Profile of Parental Lines

Line	C140	C16 0	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
LS0004-3	0	3.01	0.00	1.27	66.75	20.03	6.08	0.45	1.31	0.11	0.26	0	0.14	0.12	5.12
LS0006-5	0	3.07	0.06	1.11	64.83	22.18	6.10	0.40	1.29	0.11	0.24	0	0.13	0.13	4.94
HIO011-5	0	3.79	0.24	1.91	78.60	7.86	4.64	0.71	1.44	0.00	0.39	0	0.23	0.00	7.04
IMC 01	0	4.81	0.31	2.48	61.9i	24.81	2.61	0.85	1.06	0.07	0.48	0	0.33	0.15	9.02

The F₁ generation progeny of crosses between 90A24 and 90I22 were designated 91AS. F₁ 91AS plants were self-pollinated to produce F₂ seeds, which were harvested and analyzed for fatty acid composition by gas chromatography (GC). F₂ seeds having a low linolenic acid content and high oleic acid content were planted and self-pollinated to produce F₃ seeds. The fatty acid composition of F₃ seeds was analyzed. F₃ seeds having a high oleic acid and low linolenic acid content were planted to generate F₃ plants, which were selfed to produce F₄ seeds. The fatty acid composition of F₄ seeds was analyzed by GC. F₄ seeds having a high oleic acid and low linolenic acid content were planted to generate F₄ plants, of which 8 plants were self-pollinated to produce F₅ seeds. The fatty acid composition of F₅ seeds was analyzed by GC (Table 2).

F₅ seeds from one of the lines designated 91AS51057 was selected based on a total saturated fatty acid level of 4.99%, with low palmitic acid of 2.64% and stearic acid of 1.33% (Table 2). This line also had a higher eicosenoic acid (C20:1) content of 1.73%. The seeds of this selection (F₅ 91AS51057) were planted to generate F₅ plants, which were selfed to produce F₆ seeds. F₆ seeds were harvested from three of five selfed plants. The fatty acid composition of F₆ seeds harvested from each of the three plants is shown in Table 3. Selfing and selection within the 91AS51057 line were continued for additional 5 generations. Table 4 provides the fatty acid composition for field harvested F₁₀ seeds from 22 lines of self-pollinated 91AS51057 plants. The total saturated fatty acid content ranged from 4.38 to 6.28%, oleic acid content ranged from 74.9 to 82.5%, and α -linolenic acid content ranged from 2.1 to 4.8%. The eicosenoic acid content ranged from 1.28% to 2.30%, with most 91AS51057 F₉ plants producing F₁₀ seeds having an eicosenoic acid content from 1.90% to 2.25%. See Table 4. Seed of four individual F₁₀ 91AS51057 lines (X723868, X723977, X724734, and X724738) were selected and their seeds planted in the field in individual isolation tents. The low total saturate line X724734 was designated as 15.24 based on its nursery field position of range 15, row 24, and used in future crosses to introduce traits for low saturates through the reduction of palmitic and stearic acids. Line 15.24 also retained the higher level of eicosinoic acid of 2.06% associated with the saturate reduction.

TABLE 2

Fatty Acid Composition of Field Harvested F₅ Seed from Self-pollinated Plants

TRIAL_ID	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
91AS51023	0.05	3.32	0.18	1.03	65.59	18.89	7.95	0.58	1.46	0.07	0.43	0.03	0.21	0.21	5.62
91AS51026	0.09	4.50	0.32	1.57	63.81	24.19	2.90	0.55	1.25	0.07	0.39	0.02	0.20	0.14	7.30
91AS51026	0.09	4.36	0.29	1.51	63.09	25.21	3.11	0.49	1.14	0.07	0.33	0.01	0.17	0.13	6.95
91AS51028	0.06	3.91	0.25	1.35	64.68	24.32	3.08	0.46	1.19	0.05	0.31	0.03	0.16	0.15	6.27
91AS51028	0.06	3.71	0.24	1.32	64.77	24.38	2.97	0.47	1.30	0.05	0.34	0.04	0.16	0.19	6.06
91AS51034	0.04	2.68	0.17	1.31	74.75	11.44	5.88	0.57	1.88	0.25	0.42	0.20	0.26	0.17	5.27
91AS51044	0.02	2.66	0.17	1.35	75.19	12.23	5.20	0.54	1.81	0.12	0.34	0.04	0.18	0.16	5.08
91AS51057	0.03	2.64	0.16	1.33	71.68	12.85	8.23	0.50	1.73	0.08	0.36	0.09	0.14	0.18	4.99

TABLE 3

Fatty Acid Composition of Field Harvested F₆ Generation Seed of 91AS51057

Line	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
91AS51057	0.02	2.98	0.13	2.30	78.00	9.12	2.67	0.97	2.00	0.11	0.65	0.06	0.45	0.53	7.37
91AS51057	0.03	2.86	0.14	1.41	73.94	12.02	5.74	0.61	1.95	0.10	0.43	0.05	0.25	0.49	5.58
91AS51057	0.02	2.89	0.13	2.07	76.29	10.06	3.35	0.92	2.17	0.13	0.65	0.06	0.49	0.76	7.05

TABLE 4
Fatty Acid Composition of Field Harvest F₁₀ Generation Seed of 91AS51057

Line	Sample Number	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
91AS51057	X723860	0.04	3.16	0.19	1.10	78.79	9.13	3.37	0.53	2.05	0.30	0.37	0.05	0.24	0.68	5.43
91AS51057	X723861	0.04	2.94	0.18	1.58	81.26	7.55	2.79	0.65	1.91	0.08	0.38	0.05	0.25	0.34	5.84
91AS51057	X723862	0.04	3.01	0.19	1.69	80.31	7.83	2.81	0.71	2.02	0.09	0.44	0.06	0.32	0.50	6.21
91AS51057	X723863	0.04	2.97	0.19	1.87	80.88	7.37	2.95	0.73	1.79	0.07	0.41	0.05	0.25	0.44	6.27
91AS51057	X723868	0.04	2.66	0.17	0.92	78.20	10.71	3.81	0.39	2.11	0.12	0.26	0.06	0.11	0.44	4.38
91AS51057	X723869	0.04	3.17	0.21	1.18	80.01	8.47	2.99	0.50	2.16	0.12	0.34	0.05	0.24	0.51	5.47
91AS51057	X723924	0.04	2.81	0.16	1.11	80.23	9.38	3.01	0.42	1.93	0.12	0.23	0.03	0.12	0.39	4.74
91AS51057	X723931	0.04	2.82	0.15	0.91	79.65	9.22	3.33	0.41	2.13	0.14	0.27	0.06	0.13	0.74	4.58
91AS51057	X723932	0.10	2.75	0.15	0.98	79.62	9.21	3.15	0.44	2.18	0.16	0.31	0.05	0.15	0.76	4.73
91AS51057	X723933	0.02	2.81	0.14	0.93	80.13	9.15	3.31	0.41	2.15	0.13	0.26	0.04	0.14	0.40	4.56
91AS51057	X723970	0.04	3.25	0.25	1.73	82.09	8.11	2.34	0.51	1.28	0.05	0.19	0.00	0.10	0.06	5.83
91AS51057	X723971	0.04	3.20	0.23	1.68	82.46	7.79	2.25	0.52	1.29	0.05	0.22	0.01	0.13	0.12	5.79
91AS51057	X723977	0.04	2.72	0.19	1.19	80.64	9.76	2.10	0.52	1.92	0.07	0.32	0.02	0.15	0.35	4.95
91AS51057	X723978	0.03	2.84	0.13	1.04	80.36	8.24	3.56	0.58	2.30	0.12	0.38	0.00	0.23	0.20	5.09
91AS51057	X723984	0.04	2.73	0.16	1.01	79.33	9.37	4.00	0.45	1.97	0.10	0.29	0.04	0.14	0.36	4.66
91AS51057	X724733	0.04	3.22	0.24	1.33	74.93	12.62	4.76	0.52	1.67	0.07	0.28	0.02	0.13	0.17	5.51
91AS51057	X724734	0.03	2.82	0.18	0.98	80.14	8.92	3.27	0.44	2.24	0.13	0.28	0.04	0.16	0.37	4.72
91AS51057	X724735	0.03	2.80	0.17	1.08	79.37	9.54	3.38	0.45	2.24	0.13	0.26	0.04	0.16	0.34	4.79
91AS51057	X724736	0.04	3.16	0.25	1.73	80.96	7.68	2.59	0.70	1.90	0.07	0.40	0.05	0.25	0.23	6.28
91AS51057	X724737	0.04	2.80	0.20	1.54	80.29	8.36	3.49	0.64	1.75	0.06	0.38	0.04	0.17	0.25	5.57

91AS51057	X724738	0.03	2.72	0.17	1.12	81.88	7.71	2.84	0.52	2.06	0.10	0.32	0.05	0.17	0.30	4.89
91AS51057	X724754	0.04	2.79	0.18	1.64	80.73	8.19	3.39	0.60	1.64	0.06	0.33	0.03	0.16	0.22	5.56
AVERAGE		0.04	2.92	0.19	1.29	80.1	8.83	3.16	0.53	1.94	0.11	0.31	0.04	0.18	0.37	5.27

EXAMPLE 2

Identification of FatA2 Mutation in 15.24 Plants

Genome mapping, map-based gene cloning, and direct-sequencing strategies were used to identify loci associated with the low total saturated fatty acid phenotype in the 15.24 lines described in Example 1. A DH (doubled haploid) population was developed from a cross between 15.24 and 01OB240, a B line used in the maintenance of cytoplasmic male sterile (CMS) A lines for hybrid production. The two parental lines were screened with 1066 SNP (single nucleotide polymorphism) markers using the MassARRAY platform (Sequenom Inc., San Diego, CA) to identify polymorphic SNP markers between the two parents; 179 polymorphic SNP markers were identified.

Single marker correlations between fatty acid components and SNP markers were carried out using the SAS program (SAS Institute 1988). A *Brassica napus* genetic linkage map was constructed using the Kosambi function in JoinMap 3.0 (Kyazma). Interval quantitative trait loci (QTL) mapping was done with MapQTL 4.0 (Kyazma). A LOD score > 3.0 was considered as threshold to declare the association intervals. For fine QTL mapping, a BC₃S (backcrossing self) population was developed from a cross between 15.24 and 01PR06RR.001B, a canola R (restorer) line. SNP haplotype blocks and recombinant/crossover events within the identified QTL interval were identified using MS Excel® program.

Comparative genome mapping was performed to locate the identified QTL in *Brassica napus* chromosomes and further identify the *Brassica rapa* BAC (Bacterial Artificial chromosome) clones encompassing the identified SNP markers and the candidate genes in the identified QTL interval for the low total saturated fatty acid using publicly available *Brassica* and *Arabidopsis* genome sequences, genes, genetic linkage maps, and other information from the world wide web at brassica.bbsrc.ac.uk/, and ncbi.nlm.nih.gov/.

A total of 148 DH lines were genotyped with 179 polymorphic SNP markers. QTL mapping identified a major QTL interval (5 cM) encompassing 7 SNP markers for saturated fatty acid content (C18:0 and C20:0). Fine mapping using 610 BC₃S₁ lines from a cross between 15.24 and 01PR06RR.001B, a canola restorer line, identified two SNP markers flanking a 1 cM QTL interval that was associated with the low total

saturated fatty acid phenotype. Comparative genome mapping located this QTL in the N3 chromosome and further identified a *FatA2* candidate within this QTL interval.

DNA from lines 15.24 and 01OB240 was used as a template to amplify *FatA* sequences. Resultant sequences were analyzed using BLAST (the Basic Local Alignment Search Tool) and MegAlign and EditSeq programs from DNASTAR/Lasergene 8.0 (DNASTAR, Inc). Isoforms of *FatA1* and *FatA2* were amplified and a representative sampling is shown in Figure 1. The Bn*FatA1* sequence from 15.24 is homologous the *B. rapa* *FatA1* and *A. thaliana* *FatA1* sequence, while the Bn*FatA2* sequence from 15.24 is homologous to the *AtFatA2* and *B. napus* pNL2 sequences. Two isoforms (or alleles) of *FatA2* were evident in the sequencing results and were named *FatA2a* and *FatA2b*. Differences between the sequences of these two isoforms are shown in Figure 4. Figures 1 and 2 show a representative nucleotide (position labeled “2;” only the *FatA2b* isoform is represented in Fig 1) where, in that position, *FatA2a* is a “C” and *FatA2b* is a “T” (summarized in Figure 4). The *FatA2* sequencing results indicated that within the *FatA2b* isoform sequences, 15.24 contained a single nucleotide polymorphism represented by position labeled “1” in Figures 1, 2 and 4. In 15.24, the *FatA2b* sequences contain a “C” to “T” mutation that was not present in the 01OB240 sequences (“1” in Figs 1,2, 4). The nucleotide substitution of position “1” in Figures 1 and 2 corresponds to position 942 of the *FatA2* coding sequence (numbering based on the *Arabidopsis thaliana* sequence set forth in GenBank Accession No. NM_117374.3) and results in the substitution of a leucine residue for proline at position 255 of the encoded protein. See SEQ ID NO:28 and SEQ ID NO:32, which provide representative nucleotide sequences of the *Brassica napus* *FatA2b* gene from 15.24. In Fig. 4, position 798 is marked at the “C” to “T” SNP that correlates with low saturate content in the 15.24 lines. SEQ ID NO:29 contains the amino acid sequence of residues 242 to 277 of a wild-type *B. napus* *FatA2* polypeptide. Position 14 of SEQ ID NO:29 (position 255 in the full-length amino acid sequence) is a leucine in the *FatA2* polypeptide from 15.24. SEQ ID NO:30 contains the wild-type *Arabidopsis* *FatA2* polypeptide. SEQ ID NO:31 contains the predicted amino acid sequence of the *B. napus* *FATA2b* polypeptide from exons 2-6.

FIG. 3 contains an alignment of the conserved region around position 255 in the *Arabidopsis* *FatA2* protein, and *Brassica* *FatA2* protein from 15.24 and 01OB240. The

proline at position 255 is conserved among *Brassica*, *Arabidopsis*, *B. napus*, *B. rapa*, *B. juncea*, *Zea mays*, *Sorghum bicolor*, *Oryza sativa* Indica (rice), *Triticum aestivum*, *Glycine max*, *Jatropha* (tree species), *Carthamus tinctorius*, *Cuphea hookeriana*, *Iris tectorum*, *Perilla frutescens*, *Helianthus annuus*, *Garcinia mangostana*, *Picea sitchensis*, *Physcomitrella patens* subsp. *Patens*, *Elaeis guineensis*, *Vitis vinifera*, *Elaeis oleifera*, *Camellia oleifera*, *Arachis hypogaea*, *Capsicum annuum*, *Cuphea hookeriana*, *Populus trichocarpa*, and *Diploknema butyracea*. Furthermore, many amino acids in the region spanning amino acids 242 to 277 are homologous in both FatA and FatB (see Fett/Lipid 100 (1998) 167-172) in *Arabidopsis* and *Brassica*.

FIG. 4 shows a portion of representative BnFatA2a and BnFatA2b sequences from 010B240 and 15.24 germplasm. The positions labeled "1" and "2" correspond to the "1" and "2" positions in FIGs. 1, 2 and 3.

Large scale screening of the parental lines (15.24 and 01010B240) as well as other germplasm populations (including IMC144, IMC129, Q508, and Q7415) indicated the FatA2 SNP was 15.24-specific and was statistically significantly associated with the low total saturated fatty acid phenotype (R-square = 0.28 for total saturated content, R-square = 0.489 for C18:0; R-square = 0.385 for C20:0) and increased eicosenoic acid content (R-square = 0.389). The FatA2 SNP1 mutation was not significantly associated with the percent C14:0 and C16:0 content of oil from 15.24 plants. However, it was found that the C18:0 content of oil from 15.24 plants was negatively correlated with C20:1 content (R-value = -0.61).

EXAMPLE 3

Brassica Line 15.36

Plants producing an oil with a high oleic acid and low total saturated fatty acid content were obtained from crosses of plants from lines A12.20010 and Q508. The A12.20010 line was obtained from a cross of a selection from the IMC144 lineage and a selection from the IMC129 lineage. Line Q508 is a high oleic acid line that contains a mutation in each of the *fad2* D and F genes. See Examples 5 and 7 of U.S. Patent No. 6,342,658.

Plants designated 92EP.1039 were selected on the basis of fatty acid composition from progeny of the A12.20010 x Q508 cross. 92EP.1039 plants were crossed with plants of Trojan, a commercially available Canadian spring canola variety. The F₁ generation progeny of 92EP.1039 and Trojan were designated 93PI. F₁ 93PI plants were self-pollinated to produce F₂ seeds, which were harvested and analyzed for fatty acid composition by GC.

F₂ seeds having a high oleic acid content were selected and planted to obtain F₂ plants. The F₂ plants were self-pollinated to produce F₃ seeds. The fatty acid composition of F₃ seeds was analyzed. Table 5 contains the fatty acid profile of 93PI21 F₃ seeds from 13 different F₂ plants. F₃ 93PI21 seeds having a low saturated fatty acid content were planted to generate F₃ plants, which were selfed to produce F₄ seeds. The fatty acid composition of F₄ 93PI21 seeds was analyzed by GC. Table 6 contains the fatty acid profile of F₄ 93PI21 seeds from thirteen different self-pollinated F₃ plants. The three 93PI21 plants (T7440796, T740797, and T740799) with the lowest total saturated fatty acid content were subjected to additional rounds of selfing and selection for low total saturated fatty acid content for 5 generations. The 93PI21 line T740799 was designated as 93PI41003 at the F₄ generation and advanced. Table 7 provides the fatty acid composition for F₈ seeds harvested from 24 self-pollinated F₇ generation 93PI41003 plants. The results indicate that total saturated fatty acid content ranged from 4.51% to 6.29%, oleic acid content ranged from 64 to 71%, and α -linolenic acid content ranged from 4.8 to 7.5%. The eicosenoic acid content ranged from 1.51% to 1.99%. The 93PI41003 F₈ plant line X727712 was renamed as line 15.36 based on its nursery field position of range 15, row 36, and had a total saturated fatty acid composition of 4.51%, with reduced palmitic acid of 2.65% and stearic acid of 0.94%. Line 15.36 was used in crosses to introduce low saturate traits to other genetic backgrounds

TABLE 5

Seed Fatty Acid Composition of F₃ Generation of 93PI21

Line	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
93PI21	0.04	3.15	0.22	1.77	80.06	6.95	4.23	0.75	1.77	0.08	0.43	0.04	0.33	0.19	6.46
93PI21	0.04	3.22	0.21	1.29	79.05	7.82	4.90	0.60	1.79	0.09	0.37	0.07	0.31	0.24	5.82
93PI21	0.03	3.32	0.28	1.69	77.63	8.95	4.31	0.73	1.88	0.09	0.47	0.07	0.35	0.20	6.59
93PI21	0.04	3.57	0.33	1.43	81.33	6.09	3.89	0.63	1.61	0.05	0.41	0.17	0.25	0.20	6.34
93PI21	0.05	3.47	0.34	1.38	80.70	6.28	4.85	0.58	1.55	0.05	0.35	0.05	0.22	0.13	6.05
93PI21	0.05	3.63	0.34	1.41	80.06	6.54	4.99	0.60	1.57	0.05	0.37	0.04	0.22	0.15	6.27
93PI21	0.03	3.14	0.25	1.33	77.85	8.98	4.98	0.59	1.80	0.07	0.40	0.15	0.24	0.19	5.72
93PI21	0.03	3.00	0.24	1.34	77.65	8.02	6.23	0.61	1.90	0.08	0.40	0.06	0.24	0.22	5.60
93PI21	0.06	3.66	0.38	1.73	77.25	8.83	4.87	0.72	1.53	0.06	0.44	0.00	0.31	0.16	6.91
93PI21	0.08	4.34	0.52	2.17	77.22	6.57	4.94	0.99	1.75	0.06	0.66	0.07	0.40	0.24	8.64
93PI21	0.05	3.49	0.39	1.71	85.90	2.86	2.94	0.64	1.32	0.04	0.32	0.00	0.22	0.15	6.43
93PI21	0.04	3.13	0.25	1.44	80.58	6.99	4.31	0.60	1.81	0.07	0.36	0.04	0.23	0.15	5.80
93PI21	0.05	4.21	0.24	1.66	73.40	14.31	2.83	0.67	1.45	0.04	0.45	0.03	0.50	0.16	7.54

TABLE 6

Seed Fatty Acid Composition of Field Grown F₄ Seed Generation of 93PI21

Line	Sample No.	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
93PI21	T738147	0.03	2.78	0.15	1.60	69.57	13.82	8.81	0.62	1.77	0.08	0.37	0.04	0.14	0.22	5.54
93PI21	T738149	0.04	2.87	0.17	1.47	71.02	11.74	9.63	0.57	1.75	0.07	0.35	0.00	0.12	0.21	5.42
93PI21	T738148	0.05	3.35	0.29	1.84	73.71	11.27	5.81	0.64	1.40	0.07	0.35	0.06	0.17	0.99	6.40
93PI21	T740387	0.04	3.28	0.22	1.68	65.96	15.57	9.38	0.62	1.89	0.14	0.46	0.06	0.29	0.41	6.36
93PI21	T740388	0.03	3.00	0.20	1.66	71.33	11.89	8.15	0.63	1.93	0.10	0.49	0.05	0.29	0.26	6.09

93PI21	T740389	0.03	2.72	0.20	1.42	75.27	8.72	7.90	0.57	2.06	0.10	0.46	0.06	0.22	0.27	5.42
93PI21	T740749	0.03	2.86	0.18	1.31	68.64	13.27	10.44	0.50	1.90	0.09	0.34	0.06	0.16	0.22	5.21
93PI21	T740797	0.03	2.99	0.21	1.23	72.19	10.92	9.37	0.48	1.78	0.07	0.33	0.04	0.14	0.22	5.20
93PI21	T740798	0.03	2.78	0.20	1.26	76.73	7.47	7.39	0.58	2.35	0.14	0.47	0.07	0.18	0.34	5.29
93PI21	T740799	0.03	3.03	0.22	1.19	72.63	11.46	8.18	0.49	1.76	0.11	0.37	0.05	0.17	0.34	5.27
93PI21	T738147	0.03	2.78	0.15	1.60	69.57	13.82	8.81	0.62	1.77	0.08	0.37	0.04	0.14	0.22	5.54
93PI21	T7381149	0.04	2.87	0.17	1.47	71.02	11.74	9.63	0.57	1.75	0.007	0.35	0.00	0.12	0.21	5.42
93PI21	T738148	0.05	3.35	0.29	1.84	73.71	11.27	5.81	0.64	1.40	0.07	0.35	0.06	0.17	0.99	6.40

TABLE 7

Fatty Acid Composition of F₉ Seeds from 93PI41003 Plants in Isolation Tents

RESCHID	SAMPLE ID	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
93PI41003	X723830	0.04	2.99	0.23	1.21	60.88	23.17	8.30	0.53	1.67	0.12	0.33	0.04	0.18	0.33	5.26
93PI41003	X723846	0.03	2.73	0.22	1.22	66.06	20.77	6.22	0.45	1.57	0.09	0.26	0.03	0.15	0.22	4.83
93PI41003	X723847	0.04	2.89	0.20	1.18	68.08	17.98	6.16	0.54	1.89	0.10	0.33	0.03	0.23	0.35	5.21
93PI41003	X723848	0.03	2.80	0.21	1.23	64.93	20.91	7.09	0.47	1.58	0.08	0.27	0.02	0.15	0.22	4.95
93PI41003	X723882	0.06	2.84	0.20	1.38	69.81	16.49	5.47	0.58	1.94	0.10	0.39	0.06	0.27	0.41	5.53
93PI41003	X723883	0.04	2.87	0.19	1.35	68.41	17.07	5.98	0.60	1.95	0.14	0.42	0.06	0.31	0.61	5.59
93PI41003	X723916	0.04	3.12	0.17	1.43	69.74	16.59	5.19	0.63	1.99	0.09	0.41	0.04	0.31	0.25	5.93
93PI41003	X723917	0.02	2.51	0.20	1.02	65.86	19.54	7.61	0.41	1.77	0.11	0.29	0.04	0.11	0.53	4.35
93PI41003	X723918	0.03	2.48	0.17	1.20	68.96	17.58	5.99	0.52	1.94	0.09	0.33	0.04	0.19	0.49	4.74
93PI41003	X723919	0.03	3.12	0.18	1.10	67.25	18.48	6.46	0.48	1.90	0.11	0.32	0.04	0.23	0.31	5.27
93PI41003	X724063	0.04	2.73	0.19	1.18	66.70	19.56	6.43	0.50	1.80	0.09	0.29	0.02	0.18	0.28	4.92
93PI41003	X724064	0.04	2.71	0.21	1.22	64.00	21.73	7.06	0.45	1.60	0.08	0.27	0.04	0.15	0.44	4.83
93PI41003	X724077	0.03	2.60	0.16	1.16	67.89	19.14	5.78	0.52	1.87	0.09	0.32	0.04	0.19	0.20	4.82
93PI41003	X724091	0.03	2.72	0.18	1.27	68.62	18.11	5.76	0.57	1.93	0.10	0.34	0.00	0.18	0.19	5.11
93PI41003	X724092	0.03	2.65	0.19	1.11	63.98	21.64	7.13	0.45	1.81	0.10	0.28	0.03	0.16	0.44	4.69
93PI41003	X724093	0.03	2.57	0.19	1.21	67.35	19.67	5.77	0.47	1.80	0.09	0.29	0.04	0.18	0.36	4.74
93PI41003	X724412	0.03	2.65	0.18	0.94	65.27	20.41	7.54	0.44	1.71	0.09	0.26	0.04	0.18	0.26	4.51
93PI41003	X724416	0.04	3.02	0.22	1.19	68.18	18.57	5.49	0.54	1.67	0.08	0.34	0.05	0.29	0.33	5.41
93PI41003	X724417	0.04	2.72	0.23	1.05	66.68	19.31	6.59	0.47	1.93	0.11	0.31	0.05	0.17	0.35	4.75
93PI41003	X724420	0.03	2.81	0.19	1.22	69.48	17.31	5.43	0.57	1.71	0.09	0.36	0.05	0.36	0.42	5.34
93PI41003	X724421	0.03	2.86	0.20	1.14	66.28	19.70	6.81	0.49	1.61	0.08	0.28	0.05	0.21	0.26	5.01

93PI41003	X724422	0.04	3.18	0.21	1.04	64.87	20.88	7.02	0.50	1.51	0.07	0.30	0.05	0.15	0.18	5.20
93PI41003	X724423	0.03	2.88	0.17	1.15	68.48	17.75	6.50	0.51	1.69	0.08	0.30	0.05	0.20	0.20	5.08
93PI41003	X724611	0.04	3.28	0.25	1.64	71.09	15.28	4.78	0.64	1.73	0.08	0.39	0.05	0.31	0.45	6.29

EXAMPLE 4

Cloning of *Brassica napus* FatB

Cloning of the *Brassica napus* Fat B gene was initiated by performing PCR with primers Fat B1 (5'-ATGAAGGTTAAACCAAACGCTCAGGC-3'; SEQ ID NO:8) and Fat B2 (5'-TGTTCTTCCTCTCACCACTTCAGC-3'; SEQ ID NO:9), respectively, using Westar genomic DNA as template and Taq polymerase (Qiagen). Each 50 µL reaction contained 0.5 µM primers, 1X Qiagen Taq polymerase buffer, 2.5U Taq polymerase, and 0.2mM dNTPs. The target was amplified using the following cycling conditions: 1 cycle of 94°C for 30 seconds; 5 cycles of 94°C for 10 seconds, 58°C for 30 seconds, and 72°C for 1 min. 30 secs; 5 cycles of 94°C for 10 seconds, 54°C for 30 seconds, and 72°C for 1 min. 30 secs; and 24 cycles of 94°C for 10 seconds, 51°C for 30 seconds, and 72°C for 1 min. 45 secs. Aliquots of the PCR reactions were run on an agarose gel and selected bands were excised; DNA was eluted from the bands using the Qiagen Qiaquick kit. The DNA eluate was subjected to a 'polishing' reaction to facilitate T/A cloning and then TOPO® T/A cloned using the TOPO® T/A® cloning kit (Invitrogen). Sequences were obtained for the clones then analyzed using BLAST to search for homology. One of the clones appeared to be a FatB.

PCR was repeated using Invitrogen Platinum® Pfx polymerase, its buffer, supplementary MgSO₄ at a final concentration of 2 mM, and IMC201 strain genomic DNA with cycling conditions as follows: 1 cycle of 94°C for 2 minutes; 5 cycles of 94°C for 10 seconds, 60°C for 30 seconds, and 72°C for 1 min. 20 secs; 5 cycles of 94°C for 10 seconds, 57°C for 30 seconds, and 72°C for 1 min. 30 secs; and 24 cycles of 94°C for 10 seconds, 54°C for 30 seconds, and 72°C for 1 min. 30 secs. The PCR product from this reaction also was Topo®T/A® cloned using the Topo® T/A® cloning system (Invitrogen).

A number of the clones that were sequenced showed homology to Fat B (SEQ ID NOS:10, 11, 12, 13), with 4 distinct isoforms of the gene identified. To obtain sequence of the start and stop regions of each gene, a 'walking' procedure was employed utilizing GenomeWalker™ kits (Clontech), according to manufacturer protocols. Based on the sequence information from the walking procedure, primers corresponding to 5' UTR and

3'UTR or near-stop codon regions of the FatB genes were designed. PCR was performed using IMC201 genomic DNA as template and two sets of primers in 50 μ L reactions containing 1X Platinum® Taq High Fidelity buffer; 2.5U Platinum® Taq High Fidelity polymerase; 0.2 mM dNTPs; 0.5 μ M primers; and 2 mM MgSO₄. Primers for the first reaction were 5'-CTTTGAACGCTCAGCTCCTCAGCC-3' (SEQ ID NO:14) and 5'-AAACGAACCAAAGAACCCATGTTTGC-3' (SEQ ID NO:15). Primers for the second reaction were 5'-CTTTGAAAGCTCATCTTCCTCGTC-3' (SEQ ID NO:16) and 5'-GGTTGCAAGGTAGCAGCAGGTACAG-3' (SEQ ID NO:17). The first reaction was performed under the following cycling conditions: 1 cycle of 94°C for 2 minutes; 5 cycles of 94°C for 10 seconds, 56°C for 40 seconds, and 68°C for 1 min. 30 secs; 30 cycles of 94°C for 10 seconds, 53°C for 30 seconds, and 68°C for 2 min. The second reaction was performed under the following cycling conditions: 1 cycle of 95°C for 2 minutes; 5 cycles of 94°C for 10 seconds, 58°C for 40 seconds, and 68°C for 2 min; and 30 cycles of 94°C for 10 seconds, 55°C for 30 seconds, and 68°C for 2 min. Both reaction sets produced bands with an expected size of ~1.6Kb.

To clone the DNA, PCR reactions were performed using 1 cycle of 94°C for 2 minutes, and 35 cycles of 94°C for 10 seconds, 58°C for 40 seconds, and 68°C for 2 min. The resultant bands were gel purified and run over Qiagen Qiex II columns to purify the DNA from the agarose gel. The DNA was Topo®T/A® cloned using the Invitrogen T/A® cloning system. The nucleotide sequences set forth in SEQ ID NOS:18-21 represent full-length (or near full-length) Fat B isoforms 1, 2, 3, and 4, respectively.

FatB isoforms 5 and 6 were identified as follows. Primers 5'-ACAGTGGATGATGCTTGACTC-3' (SEQ ID NO:22) and 5'-TAGTAATATACCTGTAAGTGG-3' (SEQ ID NO:23) were designed based on FatB sequences from *B. napus* 01OB240 and used to amplify *B. napus* genomic DNA from IMC201. The resulting products were cloned and sequenced, and a new Fat B partial length isoform was identified. Sequence walking was performed with GenomeWalker™ kits (Clontech). Primers 5'-TACGATGTAGTGTCCCAAGTTGTTG-3' (SEQ ID NO:24) and 5'-TTTCTGTGGTGTGTCAGTGTGTCT-3' (SEQ ID NO:25) were designed based on the sequence obtained through genome walking and used to amplify a contiguous ORF region of the new FatB isoform. PCR products were cloned and sequenced to identify

FatB isoforms 5 and 6 (SEQ ID NO:26 and SEQ ID NO:27). The six isoforms have 82 to 95% sequence identity as assessed with the ClustalW algorithm.

EXAMPLE 5

Mutant FATB Genes

A population of *B. napus* IMC201 seeds was subjected to chemical mutagenesis. The typical fatty acid composition of field grown IMC201 is 3.6% C16:0, 1.8% C18:0, 76% C18:1, 12.5% C18:2, 3% C18:3, 0.7% C20:0, 1.5% C20:1, 0.3% C22:0, 0% C22:1, with total saturates of 6.4%. Prior to mutagenesis, IMC201 seeds were pre-imbibed in 700 gm seed lots by soaking for 15 min then draining for 5 min at room temperature. This was repeated four times to soften the seed coat. The pre-imbibed seed then were treated with 4 mM methyl N-nitrosoguanidine (MNNG) for three hours. Following the treatment with MNNG, seeds were drained of the mutagen and rinsed with water for one hour. After removing the water, the seeds were treated with 52.5 mM ethyl methanesulfonate (EMS) for sixteen hours. Follow the treatment with EMS, the seeds were drained of mutagen and rinsed with water for one and half hours. This dual mutagen treatment produced an LD₅₀ with the seed population.

Approximately 200,000 treated seeds were planted in standard greenhouse potting soil and placed into environmentally controlled greenhouse. The plants were grown under sixteen hours of day light. At maturity, M2 seed was harvested from the plants and bulked together. The M2 generation was planted and leaf samples from the early, post-cotyledon stage of development from 8 plants were pooled and DNA was extracted from leaves of these plants. The leaf harvest, pooling and DNA extraction was repeated for approximately 32,000 plants, and resulted in approximately forty 96-well blocks containing mutagenized *B. napus* IMC201 DNA. This grouping of mutagenized DNA is referred to below as the DNA mutagenesis library.

The DNA mutagenesis library was screened to identify stop-codon containing FatB mutants. In general, PCR primers were designed to amplify a region of each FatB isoform. The reaction products were analyzed using temperature gradient capillary electrophoresis on a REVEAL™ instrument (Transgenomics Inc.), which allows PCR reactions containing heterogeneous PCR products to be distinguished from reactions

containing only homogeneous products, as would be the case if a single-nucleotide polymorphism (SNP) existed in genomic DNA from chemical mutagenesis and subsequent PCR amplification.

Individual seeds representing the primary hit of each M2 plant that was the source genomic DNA mix for this primary mutagenesis screen were sampled and genomic DNA was isolated in order to perform the isoform PCR. PCR reactions were performed using *B. napus* IMC201 genomic DNA in a 30 µL reaction containing 1X Platinum® Taq High Fidelity buffer; 2.0U Platinum™ Taq High Fidelity polymerase; 0.2 mM dNTPs; 0.5 µM primers; and 2 mM MgSO₄. Cycling conditions were as follows: 1 cycle of 95°C for 2 minutes followed by 34 cycles of 94°C for 6 seconds, 64°C for 40 seconds, and 68°C for 40 seconds. PCR products were sequenced and the sequences were compared to the wild-type sequence for each isoform.

The sequence comparisons indicated that mutations had been generated and mutant plants obtained for each of isoforms 1, 2, 3 and 4. The mutant sequences are shown in SEQ ID NOS: 1-4. SEQ ID NO:1 contains the nucleotide sequence of isoform 1 having a mutation at position 154, changing the codon from CAG to TAG. SEQ ID NO:2 contains the nucleotide sequence of isoform 2 having a mutation at position 695, changing the codon from CAG to TAG. SEQ ID NO:3 contains the nucleotide sequence of isoform 3 having a mutation at position 276, changing the codon from TGG to TGA. SEQ ID NO:4 contains the nucleotide sequence of isoform 4 having a mutation at position 336, changing the codon from TGG to TGA.

EXAMPLE 6

Brassica napus plants carrying combinations of mutant *Brassica* FatB genes

Brassica napus plants carrying different combinations of mutants in different FatB isoforms were generated in order to determine the effect of the various mutant *Brassica* FatB alleles described in Example 5 on the fatty acid composition of *Brassica napus* seed oil. Parent plants, each carrying one or more mutations in a different isoform were crossed in various ways, progeny were screened by DNA sequence analysis to identify the mutation(s) present, followed by self-pollination and DNA sequence analysis

to determine whether the mutations were present in the homozygous or heterozygous state.

Using this process, three *Brassica* plants were generated that carried mutant alleles of four FatB isoforms. Each of these plants was self pollinated, harvested and replanted in the greenhouse to create a population of 1,140 plants. All 1,140 plants were screened via DNA sequence analysis to determine whether the mutant alleles were present in the homozygous or heterozygous state at each of the FatB isoform loci. Progeny were identified that were homozygous for the following combinations of mutant FatB isoforms: FatB isoforms 1, 2 and 3; FatB isoforms 1, 2 and 4; FatB isoforms 2, 3 and 4; FatB isoforms 1, 3 and 4; and FatB isoforms 1, 2, 3 and 4.

Plants carrying combinations of mutant FatB isoforms were self pollinated and seeds were harvested. The resulting seeds were planted in growth chambers under two different temperature regimes, in order to assess the effect of the different combinations of mutant alleles on fatty acid composition. The IMC201 parent was used as a control in both temperature regimes.

The seeds were planted in Premier Pro-Mix BX potting soil (Premier Horticulture, Quebec, Canada) in four inch plastic pots. Planted seeds were watered and stratified at 5°C for 5 days and germinated at 20°C day temperature and 17°C night temperature (20/17) in Conviron ATC60 controlled-environment growth chambers (Controlled Environments, Winnipeg, MB). Each gene combination was randomized and replicated 10 times in each of two separate growth chambers. At flowering, one chamber was reduced to a diurnal temperature cycle of 14°C day temperature and 11°C night temperature (14/11) while the other remained at 20/17. The temperature treatments were imposed to identify the effects of temperature on fatty acid composition. Plants were watered five times per week and fertilized bi-weekly using a 20:20:20 (NPK) liquid fertilizer at a rate of 150 ppm. Plants were bagged individually to ensure self pollination and genetic purity of the seed. Seeds from each plant was harvested individually at physiological seed maturity. All plants were analyzed using PCR based assays to confirm the presence of the *FatB* mutant alleles at the expected loci as well as the presence of mutant alleles of fatty acid desaturase genes (*mFad3a*, *mFad3b* and *mFad2d*) from the IMC201 pedigree.

IMC201 was selected from a cross of 91AE.318 x IMC02. 91AE.318 is a sister or descendent of IMC129, which is described in U.S. Patent No. 5,668,299. IMC02 was obtained from a cross of IMC01 x Westar. See Example 3 of U.S. Patent No. 5,750,827. IMC02 contains a mutation in both the *fad3A* and *fad3B* genes. The *fad3A* gene contains a C to T mutation at position 2565 from ATG in genomic DNA, resulting in the substitution of a cysteine for arginine at position 275 of the Fad3A protein. The *fad3B* gene contains a G to A mutation at position 3053 from ATG in genomic DNA, located in the exon-intron splice site recognition sequence.

A modified method for gas chromatograph determination of fatty acid profile per the American Oil Chemist's Society protocol (AOCS, 2009) was used for sample evaluation. Vials were placed in a Hewlett-Packard 5890 Series II gas chromatograph (Hewlett-Packard, Palo Alto, CA) equipped with a fused silica capillary column (5 m x 0.180 mm and 0.20 μ m film thickness) packed with a polyethylene glycol based DB-Wax® for liquid phase separation (J&W Scientific, Folsom, CA). Hydrogen (H²) was used as the carrier gas at a flow rate of 2.5 mL/min and the column temperature was isothermal at 200°C. Seed from each plant was tested via this method in replicates of three.

Fatty acid data from plants grown under the different temperature regimes was analyzed in two ways. First, data was analyzed separately as different environments and then it was pooled and analyzed across environments. Data was analyzed in SAS (SAS Institute, 2003) using proc glm to estimate differences in mean fatty acid values. Table 8 contains the genotype, population size, mean value and standard deviation of palmitic, stearic and total saturated fatty acid of seeds produced by plants carrying various combinations of mutant FatB alleles grown in two environmental growth chambers set at different diurnal temperature regimes (20°C day/17°C night; 14°C day/11°C night) as discussed above. Genotypes preceded by Iso are mutant allele combinations and the numbers thereafter indicate the specific locus. Means with different letters are significantly different as determined by a Student-Newman-Keuls mean separation test.

TABLE 8

Across Environments											
Genotype	N	C16:0	s.d.	Genotype	n	C18:0	s.d.	Genotype	n	Total Sats	s.d.
IMC201	16	3.795 a	0.424	Iso 234	16	1.971 a	0.880	IMC201	16	6.757 a	0.925
Iso234	16	3.273 b	0.368	IMC201	16	1.831 ab	0.373	Iso234	16	6.542 a	1.549
Iso124	9	3.135 bc	0.109	Iso124	9	1.81 ab	0.195	Iso124	9	6.168 ab	0.338
Iso123	8	2.959 c	0.174	Iso123	8	1.628 ab	0.227	Iso123	8	5.719 bc	0.376
Iso1234	17	2.721 d	0.240	Iso1234	17	1.520 b	0.310	Iso1234	17	5.412 c	0.729

PCR screening showed that the *mFad2d* mutant allele from IMC129 was segregating in all of the FatB mutant combinations. It was found to be absent or heterozygous in 70% of the individuals screened. The effect of this allele was statistically significant for palmitic, stearic and total saturated fatty acid contents ($F=11.17$, $p=.0011$; $F=4.43$, $p=.0376$; $F=6.55$, $p=.0118$, respectively) in analyses comparing means across environments. Therefore, the number of copies of this allele (0, 1 or 2) was included as a covariate in ANOVA mean separation tests. Significant differences were discovered for mean values of seed palmitic and total saturated fatty acid content in analyses using data pooled across environments (Table 9).

All plants carrying mutant FatB alleles showed statistically significant reductions in seed palmitic acid relative to the IMC201 control with the largest reduction in plants carrying all 4 mutant alleles. Significant reductions in total saturated fatty acid were found in seeds produced by plants carrying mutant alleles 1, 2 and 3 (i.e., Iso 123 in Tables 9 and 10) as well as Iso 1234.

Statistically significant differences were discovered for mean stearic acid content when seeds produced in the different chambers under different temperature treatments were analyzed separately (Table 10, means with different letters are significantly different as determined by a Student-Newman-Keuls mean separation test). In the 20/17 environment, Iso 123, Iso 124 and Iso 1234 all showed significant reductions in stearic acid. Only Iso 1234 showed this reduction in the 14/11 environment. Reductions in total saturated fatty acid content for Iso 123, Iso 124 and Iso 1234 were significant in the 20/17 environment and all mutant allele combinations showed significant reductions in the 14/11 environment (Tables 9 and 10). Again, plants carrying all forms of the mutant allele combinations showed significant reductions in palmitic acid when data from environments was analyzed separately.

TABLE 9

20/17 Environment											
Genotype	N	C16:0	s.d.	Genotype	N	C18:0	s.d.	Genotype	n	Total Sats	s.d.
IMC201	8	3.971 a	0.292	Iso 234	7	2.771 a	0.807	Iso234	7	8.098 a	1.116
Iso234	7	3.614 b	0.106	IMC201	8	2.158 b	0.203	IMC201	8	7.465 b	0.244
Iso124	9	3.135 c	0.109	Iso124	9	1.810 c	0.195	Iso124	9	6.168 c	0.338
Iso123	4	2.979 cd	0.159	Iso123	4	1.806 c	0.111	Iso123	4	5.988 c	0.256
Iso1234	9	2.916 d	0.102	Iso1234	9	1.749 c	0.187	Iso1234	9	5.965 c	0.390
14/11 Environment											
Genotype	N	C16:0	s.d.	Genotype	N	C18:0	s.d.	Genotype	n	Total Sats	s.d.
IMC201	8	3.618 a	0.471	IMC201	8	1.504 a	0.195	IMC201	8	6.050 a	0.826
Iso234	9	3.007 b	0.317	Iso123	4	1.451 a	0.156	Iso123	4	5.450 b	0.268
Iso123	4	2.939 b	0.210	Iso234	9	1.349 ab	0.082	Iso234	9	5.331 b	0.305
Iso1234	8	2.501 c	0.119	Iso1234	8	1.262 b	0.197	Iso1234	8	4.791 c	0.463

The mean content of the three fatty acids reported here were significantly different between the environments (C16:0 $F=59.59$, $p<.0001$; C18:0 $F=83.42$, $p<.0001$; Total Sats $F=122.02$, $p<.0001$). The data indicate that a low temperature environment reduces the amount of these saturated fatty acids in the seed oil.

TABLE 10
Fatty Acid Profile of IMC201 and Plants With Mutant FatB Alleles

Genotype	Environment	14:0	16:0	16:1	18:0	18:1	18:2	18:3	20:0	20:1	20:2	22:0	22:1	24:0	24:1	Total Sats
IMC201	High (20/17)	0.05	4.26	0.20	2.06	78.16	10.15	2.02	0.84	1.32	0.05	0.44	0.02	0.25	0.18	7.90
IMC201	High (20/17)	0.05	4.06	0.20	2.21	75.83	12.34	2.49	0.78	1.25	0.06	0.36	0.02	0.19	0.15	7.65
IMC201	High (20/17)	0.05	3.79	0.18	2.34	77.38	10.98	2.25	0.84	1.30	0.06	0.40	0.02	0.23	0.20	7.64
IMC201	High (20/17)	0.05	3.99	0.19	2.16	76.33	12.22	2.27	0.77	1.24	0.05	0.36	0.02	0.17	0.18	7.50
IMC201	High (20/17)	0.05	4.30	0.22	1.86	77.13	11.37	2.37	0.69	1.23	0.06	0.36	0.03	0.17	0.17	7.43
IMC201	High (20/17)	0.05	4.34	0.22	1.84	76.58	11.93	2.43	0.68	1.20	0.06	0.34	0.02	0.16	0.17	7.40
IMC201	High (20/17)	0.05	4.03	0.19	2.05	76.20	12.56	2.31	0.69	1.21	0.06	0.34	0.02	0.15	0.15	7.30
IMC201	High (20/17)	0.05	3.90	0.19	2.16	75.80	12.94	2.42	0.68	1.22	0.06	0.30	0.02	0.15	0.13	7.22
IMC201	High (20/17)	0.03	3.41	0.13	2.46	76.72	12.12	2.37	0.72	1.35	0.07	0.30	0.01	0.18	0.12	7.10
IMC201	Low (14/11)	0.06	4.02	0.23	1.47	74.65	13.94	3.03	0.58	1.33	0.06	0.33	0.03	0.13	0.15	6.58
IMC201	Low (14/11)	0.05	3.97	0.22	1.49	75.43	13.43	2.74	0.58	1.36	0.06	0.34	0.03	0.11	0.18	6.54
IMC201	Low (14/11)	0.05	3.76	0.21	1.63	76.15	12.63	2.98	0.61	1.27	0.06	0.34	0.04	0.11	0.16	6.51
IMC201	Low (14/11)	0.04	3.84	0.21	1.42	75.88	12.93	2.93	0.57	1.43	0.07	0.36	0.02	0.12	0.19	6.35
IMC201	Low (14/11)	0.04	3.66	0.20	1.59	75.94	12.96	2.98	0.55	1.32	0.08	0.34	0.05	0.10	0.20	6.28
IMC201	Low (14/11)	0.05	3.67	0.20	1.62	76.61	12.52	2.96	0.37	1.32	0.05	0.31	0.03	0.11	0.20	6.13
IMC201	Low (14/11)	0.03	3.49	0.13	1.73	74.49	14.29	3.38	0.40	1.61	0.04	0.21	0.02	0.06	0.13	5.92
IMC201	Low (14/11)	0.02	2.53	0.16	1.09	75.76	15.24	3.20	0.14	1.29	0.08	0.24	0.02	0.07	0.16	4.08
Isol23	High (20/17)	0.04	3.20	0.26	1.82	76.55	12.40	3.05	0.68	1.21	0.06	0.35	0.02	0.17	0.18	6.26
Isol23	High (20/17)	0.03	2.85	0.27	1.97	78.31	10.98	2.78	0.76	1.17	0.04	0.38	0.04	0.25	0.15	6.25
Isol23	High (20/17)	0.04	2.96	0.24	1.95	77.09	12.09	3.06	0.68	1.15	0.06	0.33	0.02	0.16	0.18	6.10
Isol23	High (20/17)	0.04	2.82	0.32	1.75	74.68	14.64	2.96	0.69	1.17	0.05	0.38	0.01	0.25	0.26	5.92
Isol23	High (20/17)	0.04	2.94	0.27	1.70	76.32	13.20	3.21	0.57	1.11	0.06	0.29	0.01	0.13	0.16	5.67
Isol23	Low (14/11)	0.04	3.19	0.27	1.50	72.50	15.95	3.75	0.59	1.41	0.07	0.37	0.02	0.12	0.23	5.80
Isol23	Low (14/11)	0.05	2.89	0.30	1.64	75.34	13.62	3.80	0.52	1.12	0.06	0.30	0.04	0.11	0.21	5.51
Isol23	Low (14/11)	0.03	3.00	0.24	1.29	75.38	13.96	3.40	0.53	1.41	0.06	0.35	0.03	0.11	0.21	5.32
Isol23	Low (14/11)	0.03	2.68	0.25	1.37	76.24	12.90	3.65	0.59	1.43	0.06	0.38	0.02	0.14	0.26	5.18
Isol24	High (20/17)	0.04	3.23	0.28	2.13	72.73	16.29	2.74	0.72	1.07	0.05	0.37	0.02	0.16	0.16	6.65
Isol24	High (20/17)	0.04	3.17	0.27	2.01	72.62	16.76	2.55	0.71	1.09	0.05	0.37	0.02	0.17	0.16	6.48

Iso124	High (20/17)	0.04	3.12	0.24	1.87	78.55	10.92	2.43	0.74	1.24	0.06	0.39	0.02	0.21	0.17	6.37
Iso124	High (20/17)	0.04	3.19	0.25	1.82	71.84	17.19	2.96	0.67	1.15	0.06	0.37	0.02	0.19	0.26	6.27
Iso124	High (20/17)	0.05	3.15	0.32	1.82	77.52	11.88	2.65	0.68	1.18	0.06	0.36	0.00	0.18	0.16	6.22
Iso124	High (20/17)	0.04	3.20	0.28	1.89	67.27	22.04	2.95	0.62	1.02	0.06	0.31	0.01	0.13	0.17	6.20
Iso124	High (20/17)	0.04	3.24	0.26	1.57	66.46	22.91	2.93	0.62	1.15	0.07	0.35	0.03	0.14	0.23	5.97
Iso124	High (20/17)	0.04	2.98	0.22	1.59	78.96	11.00	2.76	0.61	1.15	0.05	0.34	0.02	0.15	0.15	5.70
Iso124	High (20/17)	0.04	2.93	0.24	1.60	78.65	11.12	2.87	0.62	1.21	0.05	0.33	0.02	0.15	0.18	5.65
Iso124	Low (14/11)	0.04	2.84	0.31	1.54	73.84	15.65	3.51	0.36	1.16	0.06	0.35	0.02	0.10	0.22	5.23
Iso234	High (20/17)	0.05	3.64	0.25	3.72	69.54	16.12	2.79	1.30	1.13	0.07	0.63	0.02	0.42	0.31	9.78
Iso234	High (20/17)	0.05	3.39	0.22	3.70	67.48	18.73	3.14	1.13	1.06	0.06	0.50	0.03	0.27	0.24	9.04
Iso234	High (20/17)	0.05	3.60	0.22	3.26	70.34	17.05	2.52	1.04	1.03	0.06	0.45	0.01	0.21	0.17	8.60
Iso234	High (20/17)	0.05	3.69	0.25	2.64	70.29	17.18	3.07	0.85	1.07	0.06	0.39	0.01	0.26	0.18	7.88
Iso234	High (20/17)	0.05	3.69	0.23	2.37	72.38	15.60	2.50	0.92	1.20	0.07	0.49	0.02	0.27	0.22	7.79
Iso234	High (20/17)	0.05	3.82	0.31	1.76	74.59	14.16	2.74	0.64	1.15	0.07	0.34	0.01	0.25	0.12	6.85
Iso234	High (20/17)	0.05	3.70	0.26	1.76	76.47	12.22	2.70	0.70	1.26	0.06	0.37	0.03	0.23	0.19	6.81
Iso234	High (20/17)	0.05	3.59	0.25	1.94	70.65	18.05	2.81	0.68	1.13	0.06	0.34	0.04	0.18	0.21	6.79
Iso234	Low (14/11)	0.06	3.71	0.32	1.27	66.07	21.66	4.24	0.35	1.32	0.07	0.43	0.06	0.16	0.29	5.98
Iso234	Low (14/11)	0.03	3.18	0.32	1.40	66.38	22.07	3.79	0.55	1.29	0.10	0.39	0.06	0.13	0.32	5.68
Iso234	Low (14/11)	0.03	3.28	0.29	1.40	66.93	23.28	2.44	0.46	1.13	0.06	0.29	0.04	0.12	0.24	5.59
Iso234	Low (14/11)	0.04	3.13	0.28	1.43	67.90	21.10	3.53	0.52	1.30	0.08	0.34	0.02	0.11	0.23	5.57
Iso234	Low (14/11)	0.04	3.05	0.27	1.30	68.17	20.89	3.63	0.50	1.33	0.07	0.35	0.04	0.12	0.24	5.36
Iso234	Low (14/11)	0.05	3.12	0.29	1.30	66.56	22.26	3.88	0.35	1.30	0.10	0.35	0.03	0.14	0.26	5.30
Iso234	Low (14/11)	0.02	3.12	0.30	1.33	69.56	20.66	2.59	0.33	1.28	0.08	0.34	0.04	0.11	0.24	5.25
Iso234	Low (14/11)	0.04	2.74	0.27	1.45	76.56	12.91	3.53	0.49	1.25	0.06	0.32	0.04	0.11	0.21	5.15
Iso234	Low (14/11)	0.04	2.93	0.25	1.18	70.80	18.54	3.58	0.49	1.40	0.07	0.34	0.02	0.11	0.26	5.09
Iso234	Low (14/11)	0.03	2.52	0.38	1.35	72.27	16.85	3.81	0.54	1.30	0.06	0.40	0.00	0.15	0.33	4.99
Iso1234	High (20/17)	0.04	3.07	0.26	2.09	69.61	18.91	2.87	0.88	1.18	0.07	0.52	0.03	0.23	0.23	6.84
Iso1234	High (20/17)	0.04	2.90	0.26	1.92	68.36	20.89	3.05	0.72	1.06	0.06	0.37	0.00	0.20	0.18	6.15
Iso1234	High (20/17)	0.04	2.92	0.26	1.75	73.39	15.92	2.90	0.75	1.17	0.06	0.42	0.03	0.20	0.21	6.07
Iso1234	High (20/17)	0.04	2.87	0.28	1.83	71.68	17.50	3.11	0.72	1.11	0.06	0.37	0.03	0.19	0.22	6.02
Iso1234	High (20/17)	0.04	3.01	0.26	1.54	71.11	18.51	2.66	0.71	1.19	0.07	0.44	0.03	0.20	0.23	5.94
Iso1234	High (20/17)	0.04	3.01	0.29	1.57	70.56	18.63	3.40	0.62	1.12	0.06	0.34	0.02	0.16	0.19	5.74
Iso1234	High (20/17)	0.04	2.79	0.27	1.80	70.89	18.95	2.88	0.63	1.06	0.06	0.31	0.02	0.15	0.15	5.74

Iso1234	High (20/17)	0.04	2.77	0.24	1.71	72.23	17.53	2.90	0.66	1.12	0.07	0.35	0.01	0.19	0.18	5.72
Iso1234	High (20/17)	0.04	2.89	0.28	1.53	67.27	22.56	3.11	0.57	1.03	0.07	0.31	0.01	0.14	0.19	5.47
Iso1234	Low (14/11)	0.04	2.61	0.29	1.36	68.44	20.55	3.93	0.63	1.24	0.08	0.46	0.03	0.14	0.20	5.24
Iso1234	Low (14/11)	0.02	2.51	0.27	1.42	69.76	19.44	3.75	0.64	1.27	0.10	0.44	0.02	0.15	0.23	5.17
Iso1234	Low (14/11)	0.03	2.54	0.26	1.33	64.33	25.15	3.73	0.56	1.22	0.09	0.40	0.01	0.12	0.24	4.97
Iso1234	Low (14/11)	0.04	2.68	0.27	1.36	70.70	18.79	3.82	0.38	1.19	0.05	0.38	0.01	0.12	0.21	4.96
Iso1234	Low (14/11)	0.03	2.47	0.29	1.31	68.06	21.43	3.63	0.59	1.35	0.07	0.40	0.02	0.11	0.27	4.89
Iso1234	Low (14/11)	0.03	2.53	0.29	1.29	65.90	23.37	3.93	0.53	1.30	0.08	0.39	0.02	0.10	0.24	4.86
Iso1234	Low (14/11)	0.03	2.39	0.27	1.24	70.56	18.96	3.64	0.59	1.38	0.08	0.43	0.03	0.15	0.26	4.82
Iso1234	Low (14/11)	0.04	2.48	0.28	1.34	71.51	18.58	3.57	0.34	1.14	0.08	0.34	0.02	0.10	0.19	4.63
Iso1234	Low (14/11)	0.02	2.27	0.14	0.76	74.15	13.60	6.46	0.32	1.67	0.12	0.23	0.04	0.07	0.16	3.66

EXAMPLE 7

Brassica plant lines 1764, 1975, and 2650

Lines 1764, 1975, and 2650 were selected from the mutagenized population of IMC201 seeds of Example 5 as follows. Three thousand bulk M₂ generation seeds were planted. Upon maturity, M₃ seed (2500 individuals) was harvested from 2500 M₂ plants and analyzed via GC. Table 11 provides the fatty acid profile of seed from three lines identified as having a low total saturates content in seed oil: 1764, 1975, and 2650. M₃ seeds of 1764, 1975, and 2650 were planted (100 per line) and the resulting plants were self pollinated. M₄ seeds were harvested from the plants and analyzed via GC (see Table 12).

TABLE 11

Fatty acid composition of M₃ generation seed from mutant lines exhibiting reduced saturated fatty acid content

Line	C14 0	C16 0	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
1764	0.05	3.30	0.31	1.65	76.30	13.40	2.00	0.668	1.46	0.06	0.38	0.02	0.28	0.15	6.32
1975	0.03	3.19	0.22	1.35	75.51	14.21	2.19	0.59	1.77	0.10	0.43	0.00	0.23	0.19	5.82
2650	0.04	3.00	0.12	3.79	77.77	8.59	2.056	1.42	1.68	0.08	0.74	0.02	0.45	0.26	9.44

TABLE 12

Fatty acid composition of M₄ generation seed from three mutant lines exhibiting reduced saturated fatty acid content

Line	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total sat
1764-06	0.05	3.06	0.34	1.94	76.89	12.57	1.99	0.70	1.34	0.05	0.39	0.00	0.23	0.45	6.37
1764-35	0.04	3.54	0.47	1.64	74.09	15.38	2.04	0.59	1.32	0.05	0.32	0.00	0.19	0.34	6.31
1764-43	0.04	3.06	0.32	1.88	75.24	14.26	1.86	0.75	1.58	0.07	0.45	0.03	0.28	0.18	6.46
1764-59	0.05	3.33	0.38	1.57	74.92	14.56	2.21	0.57	1.33	0.05	0.32	0.03	0.19	0.49	6.02
1764-91	0.05	3.11	0.34	1.77	75.83	13.70	2.15	0.67	1.37	0.05	0.38	0.02	0.24	0.32	6.21
1764-92	0.04	3.00	0.30	2.07	76.75	12.79	2.11	0.74	1.40	0.05	0.40	0.00	0.22	0.13	6.47
1764-95	0.06	3.38	0.40	1.62	74.11	15.17	2.18	0.63	1.36	0.06	0.37	0.03	0.22	0.43	6.27
1975-01	0.05	3.31	0.23	1.52	73.60	15.79	2.17	0.62	1.51	0.08	0.40	0.03	0.17	0.51	6.07
1975-04	0.02	3.04	0.16	1.74	77.28	12.64	2.08	0.66	1.54	0.06	0.36	0.00	0.17	0.24	6.00
1975-32	0.03	3.54	0.22	1.52	73.89	15.44	2.35	0.59	1.55	0.09	0.34	0.00	0.18	0.26	6.20
1975-65	0.03	3.18	0.16	1.71	75.16	14.26	2.22	0.63	1.64	0.09	0.36	0.00	0.16	0.39	6.07
1975-76	0.05	3.52	0.19	1.48	73.18	16.04	2.37	0.62	1.63	0.09	0.39	0.03	0.23	0.19	6.28
1975-84	0.04	3.12	0.14	1.68	75.57	14.07	2.35	0.64	1.61	0.09	0.35	0.00	0.20	0.12	6.03
1975-90	0.04	3.34	0.23	1.40	72.21	17.44	2.25	0.58	1.70	0.11	0.35	0.00	0.20	0.16	5.92

1975-96	0.04	3.13	0.17	1.99	76.43	12.99	2.05	0.76	1.60	0.07	0.40	0.00	0.23	0.13	6.55
1975-99	0.04	3.13	0.20	1.83	74.80	14.34	2.15	0.72	1.68	0.08	0.43	0.04	0.21	0.35	6.37
2650-20	0.06	2.81	0.13	4.08	74.24	11.71	2.29	1.38	1.84	0.11	0.62	0.05	0.38	0.31	9.32
2650-36	0.05	2.93	0.14	3.63	74.55	11.95	2.58	1.20	1.64	0.09	0.55	0.00	0.28	0.40	8.64
2650-45	0.06	3.02	0.14	3.74	75.16	11.27	2.49	1.19	1.58	0.08	0.51	0.00	0.26	0.52	8.77
IMC02-01	0.06	3.73	0.21	2.96	70.35	18.37	1.30	0.98	1.15	0.05	0.44	0.01	0.26	0.13	8.43
IMC02-02	0.05	3.64	0.23	2.85	70.86	17.82	1.28	0.98	1.21	0.05	0.48	0.02	0.29	0.26	8.27
IMC02-03	0.05	3.66	0.21	2.90	69.84	18.96	1.32	0.94	1.15	0.05	0.42	0.02	0.27	0.21	8.24
IMC02-04	0.05	3.62	0.23	3.06	68.94	19.37	1.38	1.01	1.20	0.06	0.49	0.00	0.31	0.28	8.54
IMC02-05	0.04	3.62	0.24	3.13	69.27	19.33	1.34	0.96	1.13	0.06	0.41	0.01	0.25	0.20	8.42
IMC02-06	0.05	3.87	0.25	3.74	70.11	17.17	1.40	1.21	1.14	0.06	0.58	0.00	0.34	0.09	9.79
IMC02-07	0.06	3.75	0.27	2.89	66.48	22.22	1.34	0.89	1.11	0.05	0.40	0.00	0.23	0.29	8.23
IMC02-08	0.06	3.71	0.25	2.83	69.87	18.87	1.25	0.95	1.16	0.05	0.43	0.00	0.27	0.30	8.26
IMC02-09	0.07	4.51	0.35	3.83	65.22	20.57	1.96	1.20	1.03	0.00	0.57	0.00	0.37	0.34	10.53
IMC02-10	0.05	3.66	0.25	2.77	68.23	20.84	1.27	0.90	1.17	0.05	0.41	0.00	0.25	0.16	8.03
IMC02-11	0.05	3.79	0.23	2.95	68.43	20.15	1.32	0.98	1.15	0.06	0.46	0.00	0.29	0.13	8.52
IMC02-12	0.06	3.72	0.25	2.78	68.35	20.50	1.30	0.90	1.15	0.05	0.42	0.00	0.26	0.25	8.14
IMC02-13	0.08	3.92	0.25	2.92	67.17	21.30	1.43	0.93	1.11	0.06	0.42	0.00	0.30	0.12	8.56
IMC02-14	0.05	3.64	0.23	3.09	71.73	16.73	1.36	1.05	1.19	0.05	0.51	0.00	0.28	0.09	8.62
IMC02-15	0.06	3.73	0.25	2.99	69.14	19.49	1.23	0.99	1.15	0.05	0.45	0.00	0.29	0.17	8.51
IMC02-16	0.06	3.76	0.24	2.81	69.05	19.89	1.21	0.94	1.17	0.05	0.43	0.00	0.27	0.14	8.25
IMC02-17	0.05	3.63	0.25	2.61	67.52	21.91	1.33	0.83	1.12	0.06	0.39	0.00	0.21	0.10	7.72
IMC02-18	0.05	3.66	0.22	3.19	71.15	17.32	1.25	1.06	1.16	0.05	0.51	0.00	0.29	0.11	8.76
IMC02-19	0.05	3.65	0.24	3.18	68.92	19.62	1.28	1.02	1.13	0.05	0.45	0.00	0.30	0.12	8.64
IMC02-20	0.05	3.71	0.26	2.79	66.85	22.13	1.55	0.87	1.10	0.06	0.41	0.00	0.22	0.00	8.05
IMC02Ave	0.05	3.75	0.24	3.01	68.87	19.63	1.36	0.98	1.14	0.05	0.45	0.00	0.28	0.17	8.52
IMC201-01	0.05	4.01	0.19	2.45	77.44	10.56	2.01	0.93	1.38	0.05	0.47	0.02	0.28	0.15	8.20
IMC201-02	0.05	3.94	0.18	2.44	77.52	10.55	2.09	0.92	1.38	0.05	0.46	0.02	0.26	0.15	8.07
IMC201-03	0.06	4.06	0.21	2.59	76.51	11.16	2.06	0.94	1.34	0.05	0.46	0.02	0.26	0.28	8.37

IMC201-04	0.06	4.02	0.21	2.46	76.25	11.61	2.21	0.87	1.32	0.05	0.42	0.00	0.23	0.29	8.05
IMC201-05	0.05	4.10	0.20	2.56	76.42	11.35	2.07	0.93	1.34	0.05	0.46	0.02	0.28	0.15	8.39
IMC201-06	0.05	4.05	0.21	2.50	76.36	11.51	2.08	0.91	1.37	0.05	0.45	0.03	0.26	0.16	8.23
IMC201-07	0.07	4.22	0.22	2.62	75.71	11.77	2.05	0.94	1.35	0.05	0.47	0.02	0.26	0.25	8.58
IMC201-08	0.05	3.64	0.18	2.63	77.81	10.20	2.02	0.96	1.47	0.06	0.47	0.02	0.31	0.17	8.07
IMC201-09	0.05	4.41	0.24	2.85	63.92	22.50	2.79	0.96	1.20	0.08	0.48	0.02	0.32	0.17	9.08
IMC201-10	0.05	4.03	0.18	2.48	77.12	10.69	2.17	0.90	1.33	0.05	0.45	0.00	0.23	0.31	8.15
IMC201Ave	0.06	4.05	0.20	2.56	75.51	12.19	2.16	0.93	1.35	0.05	0.46	0.02	0.27	0.21	8.32
Westar16-01	0.06	4.41	0.30	2.34	65.36	18.31	6.50	0.76	1.13	0.06	0.35	0.00	0.22	0.19	8.15
Westar16-02	0.06	4.25	0.26	2.37	67.28	16.80	6.24	0.75	1.13	0.05	0.35	0.02	0.20	0.24	7.99
Westar16-03	0.06	4.20	0.26	2.46	66.06	17.62	6.71	0.76	1.13	0.06	0.37	0.00	0.20	0.11	8.05
Westar16-04	0.07	4.52	0.29	2.54	64.75	18.82	6.53	0.74	1.04	0.06	0.34	0.00	0.19	0.11	8.40
Westar16-05	0.07	4.30	0.27	2.43	65.09	18.31	6.67	0.80	1.19	0.07	0.39	0.00	0.25	0.17	8.23
Westar16-06	0.08	4.54	0.30	2.39	65.63	17.74	6.44	0.81	1.15	0.06	0.39	0.00	0.25	0.21	8.46
Westar16-07	0.08	4.34	0.28	2.57	65.47	17.92	6.57	0.79	1.12	0.06	0.35	0.00	0.20	0.27	8.32
Westar16-08	0.07	4.37	0.28	2.18	64.49	19.54	6.61	0.64	1.05	0.06	0.28	0.00	0.15	0.28	7.70
Westar16-09	0.08	4.65	0.29	2.35	61.81	21.30	6.72	0.72	1.21	0.08	0.33	0.00	0.20	0.27	8.33
Westar16-10	0.06	4.26	0.25	2.54	67.17	16.96	5.85	0.80	1.17	0.06	0.38	0.00	0.22	0.28	8.27
Westar16Ave	0.07	4.39	0.28	2.42	65.31	18.33	6.48	0.76	1.13	0.06	0.35	0.00	0.21	0.21	8.19

EXAMPLE 8

DH line Salomon

A cross was made between 15.24 (Example 1) and 1764-92-05 (Example 7). A DH population was generated by collecting F₁ microspores from the cross, treating the microspores with colchicine, and propagating them *in vitro*. Plantlets formed *in vitro* from the microspores were moved to a greenhouse and inflorescences that formed were self pollinated. Seed was harvested from the DH₁ plants at maturity and analyzed for fatty acid profile. Seeds from those plants exhibiting reduced saturated fatty acid content were grown in the greenhouse and in the field. Table 13 contains the fatty acid profile of seeds produced by greenhouse-grown plants of a DH₁ population designated Salomon. Table 14 contains the fatty acid profile of seeds from three plants of DH line Salomon-05 grown in the field and re-coded to Salomon-005. The fatty acid profile of IMC111RR, a registered Canadian *B. napus* variety, is included as a control in Table 14. The field grown seed of individual plants of Salomon 005 had a range of 3.83% to 4.44% total saturates with 2.92% to 3.35% palmitic acid and 0.29% to 0.47% stearic acid. Line Salomon-005 demonstrated the lowest total saturated fatty acid profile of the DH lines in the greenhouse and in the field.

Table 15 contains the fatty acid profile of seeds from individual Salomon-005 plants, progeny of DH line Salomon, as grown in a growth chamber under the conditions described in Example 6. Under the high temperature environment (20/17), selfed plants of Salomon 005 had a total saturated fatty acid range of 4.13% to 4.67% with palmitic acid of 2.55 % to 2.70% and stearic acid of 1.05 to 0.78%. Seed from the same Salomon 005 DH₁ source when grown in a low temperature environment (14/11) had a total saturates of 3.45% to 3.93% with palmitic acid of 2.25% to 2.39% and stearic acid of 0.57% to 0.85%. The FATA2 mutation from 15.24 when combined with other low saturate mutations such as 1764, 1975, and 2650 can further reduce total saturates through the additive reduction of palmitic and stearic acids.

In the low 14/11 environment, Salomon-005-09 exhibited the lowest palmitic acid content, Salomon-005-05 exhibited the lowest stearic acid content, and Salomon-005-07 exhibited the lowest total saturated fatty acid content. Table 15 also contains the profile of individual plants of 15.24, IMC201, and F6 progeny of 1764-43-06 x 1975-90-14 (see

Example 10). The data indicate that a low temperature environment reduces the amount of saturated fatty acids in the seed oil.

Lines 1764, 1975 and 2650 are also crossed with 15.36 (Example 3) to generate progeny having reduced saturated fatty acid content.

TABLE 13

Seed Fatty acid composition of progeny of DH₁ line Salomon in the greenhouse

Line	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
Salomon-01	0.05	3.68	0.33	2.24	72.56	16.20	2.16	0.73	1.19	0.064	0.35	0.00	0.23	0.23	7.28
Salomon-02	0.04	2.66	0.17	1.60	72.33	14.38	4.28	0.67	2.05	0.16	0.30	0.00	0.24	0.13	6.49
Salomon-03	0.04	2.89	0.21	1.59	76.24	13.68	2.28	0.66	1.57	0.07	0.33	0.00	0.24	0.20	5.74
Salomon-04	0.05	3.17	0.19	1.40	79.15	9.70	3.47	0.56	1.52	0.06	0.29	0.00	0.24	0.21	5.70
Salomon-05	0.03	3.19	0.16	1.22	75.41	12.99	3.58	0.57	1.92	0.14	0.33	0.00	0.30	0.16	5.65
Salomon-06	0.05	3.67	0.24	1.53	76.15	12.12	3.49	0.66	1.53	0.06	0.32	0.00	0.18	0.00	6.40
Salomon-07	0.05	4.37	0.20	0.87	77.28	10.76	3.46	0.43	1.81	0.10	0.25	0.00	0.22	0.20	6.19
Salomon-08	0.05	4.19	0.25	1.29	78.05	10.35	2.99	0.59	1.65	0.08	0.34	0.00	0.19	0.00	6.64
Average	0.05	3.48	0.22	1.47	75.9	12.52	3.21	0.61	1.66	0.092	0.31	0.00	0.23	0.14	6.26

TABLE 14

Seed Fatty acid composition of DH₂ line Salomon-005 in the field

Line	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
Salomon-005	0.036	2.92	0	0.29	73.16	14.02	5.83	0.21	2.57	0.13	0.27	0.05	0.11	0.406	3.83
Salomon-005	0.036	2.85	0	0.55	74.17	13.24	5.74	0.27	2.44	0.12	0.28	0.02	0.02	0.268	3.99
Salomon-005	0.043	3.35	0	0.47	71.35	15.20	5.90	0.24	2.63	0.17	0.32	0.06	0.03	0.251	4.44
Average	0.038	3.04	0.0	0.44	72.89	14.15	5.82	0.24	2.55	0.14	0.29	0.04	0.05	0.308	4.09
IMC111RR	0.08	5.06	0.41	2.07	56.80	28.40	3.87	0.83	1.44	0.14	0.50	0.00	0.23	0.162	8.77
IMC111RR	0.09	5.38	0.50	2.09	56.61	28.38	3.50	0.81	1.41	0.13	0.50	0.01	0.53	0.083	9.40
IMC111RR	0.21	6.15	0.50	1.46	47.82	36.03	3.38	0.71	1.24	0.14	0.56	0.00	1.43	0.369	10.5

TABLE 15

Seed fatty acid profile of individual DH line Salomon-005 Plants, 15.24, IMC201, and F6 plants in the growth chamber

Genotype	Environment	14:0	16:0	16:1	18:0	18:1	18:2	18:3	20:0	20:1	20:2	22:0	22:1	24:0	24:1	Total Sats
Salomon-005-01	High 20/17	0.02	2.59	0.14	1.05	76.66	11.34	5.29	0.44	1.68	0.12	0.27	0.04	0.14	0.21	4.51
Salomon-005-02	High 20/17	0.02	2.64	0.13	0.93	76.44	12.61	4.65	0.37	1.56	0.13	0.23	0.04	0.11	0.15	4.31
Salomon-005-03	High 20/17	0.02	2.63	0.12	0.83	77.05	11.95	4.82	0.34	1.62	0.12	0.23	0.04	0.10	0.13	4.15
Salomon-005-04	High 20/17	0.02	2.57	0.12	0.84	77.73	11.43	4.60	0.35	1.68	0.13	0.24	0.04	0.11	0.14	4.13
Salomon-005-05	High 20/17	0.02	2.67	0.13	1.08	75.92	12.43	4.82	0.47	1.67	0.13	0.28	0.05	0.15	0.19	4.67
Salomon-005-06	High 20/17	0.02	2.56	0.13	1.03	76.63	12.18	4.84	0.40	1.56	0.12	0.25	0.04	0.11	0.13	4.37
Salomon-005-07	High 20/17	0.02	2.58	0.13	0.78	77.50	11.64	4.49	0.36	1.78	0.14	0.26	0.05	0.12	0.18	4.11
Salomon-005-08	High 20/17	0.02	2.70	0.14	0.90	76.60	11.80	4.92	0.41	1.73	0.14	0.27	0.04	0.13	0.20	4.44
Salomon-005-09	High 20/17	0.02	2.58	0.12	0.88	77.75	11.62	4.50	0.34	1.61	0.12	0.22	0.04	0.10	0.10	4.14
Salomon-005-10	High 20/17	0.02	2.46	0.13	0.99	77.92	11.20	4.55	0.41	1.62	0.13	0.26	0.04	0.13	0.16	4.28
Salomon-005-01	Low 14/11	0.02	2.27	0.12	0.68	73.66	13.53	6.86	0.32	1.83	0.13	0.25	0.06	0.06	0.21	3.59
Salomon-005-02	Low 14/11	0.02	2.39	0.14	0.85	74.61	13.00	6.54	0.34	1.48	0.05	0.25	0.05	0.07	0.18	3.93
Salomon-005-03	Low 14/11	0.02	2.39	0.14	0.74	73.41	14.26	6.47	0.32	1.66	0.13	0.24	0.03	0.05	0.15	3.76
Salomon-005-04	Low 14/11	0.02	2.37	0.15	0.68	73.55	14.02	6.52	0.31	1.71	0.12	0.24	0.05	0.06	0.19	3.69
Salomon-005-05	Low 14/11	0.01	2.33	0.11	0.57	72.96	15.04	6.19	0.27	1.84	0.16	0.23	0.04	0.06	0.19	3.47
Salomon-005-06	Low 14/11	0.02	2.32	0.14	0.84	73.64	13.54	6.96	0.32	1.59	0.10	0.25	0.06	0.07	0.16	3.82
Salomon-005-07	Low 14/11	0.02	2.31	0.12	0.60	72.14	15.60	6.54	0.25	1.78	0.14	0.21	0.05	0.06	0.17	3.45
Salomon-005-08	Low 14/11	0.02	2.39	0.14	0.61	72.72	14.76	6.38	0.30	1.97	0.14	0.24	0.05	0.07	0.21	3.64
Salomon-005-09	Low 14/11	0.02	2.25	0.14	0.73	74.30	13.27	6.75	0.31	1.66	0.10	0.23	0.04	0.05	0.15	3.60
Salomon-005-10	Low 14/11	0.03	2.30	0.14	0.81	74.10	13.40	6.91	0.13	1.60	0.05	0.24	0.06	0.06	0.18	3.57
F6-01	High 20/17	0.03	2.60	0.14	0.97	77.08	13.84	2.51	0.44	1.57	0.09	0.30	0.04	0.17	0.23	4.51
F6-02	High 20/17	0.03	2.66	0.16	1.08	75.93	14.82	2.56	0.46	1.55	0.09	0.29	0.03	0.14	0.18	4.68
F6-03	High 20/17	0.02	2.54	0.12	0.97	74.35	16.41	2.44	0.45	1.85	0.13	0.31	0.04	0.15	0.22	4.44

F6-04	High 20/17	0.03	2.59	0.16	1.17	77.17	13.62	2.55	0.50	1.48	0.08	0.29	0.03	0.15	0.20	4.72
F6-05	High 20/17	0.03	2.39	0.12	1.24	74.19	15.98	2.97	0.50	1.77	0.12	0.31	0.04	0.16	0.20	4.62
F6-06	High 20/17	0.03	2.46	0.12	1.30	74.78	15.28	2.97	0.53	1.72	0.11	0.32	0.05	0.14	0.21	4.77
F6-07	High 20/17	0.03	2.59	0.17	1.23	75.88	14.86	2.49	0.52	1.45	0.08	0.34	0.03	0.18	0.16	4.88
F6-08	High 20/17	0.03	2.43	0.13	1.35	74.57	15.91	2.65	0.53	1.59	0.11	0.31	0.03	0.19	0.16	4.84
F6-09	High 20/17	0.03	2.58	0.18	1.27	77.36	13.34	2.44	0.54	1.46	0.08	0.34	0.03	0.18	0.19	4.94
F6-10	High 20/17	0.03	2.31	0.12	1.28	75.12	14.90	2.99	0.53	1.84	0.12	0.33	0.04	0.17	0.23	4.65
F6-01	Low 14/11	0.02	2.47	0.14	0.92	73.90	16.63	3.30	0.39	1.51	0.10	0.27	0.03	0.10	0.22	4.17
F6-02	Low 14/11	0.02	2.34	0.14	0.88	75.11	15.79	3.16	0.37	1.56	0.09	0.25	0.03	0.08	0.18	3.94
F6-03	Low 14/11	0.02	2.38	0.12	0.91	74.76	15.89	3.28	0.37	1.57	0.11	0.28	0.03	0.09	0.19	4.04
F6-04	Low 14/11	0.02	2.35	0.15	0.97	74.66	16.22	3.15	0.39	1.50	0.09	0.26	0.03	0.07	0.17	4.06
F6-05	Low 14/11	0.03	2.50	0.17	0.98	74.94	15.83	3.10	0.37	1.42	0.06	0.27	0.05	0.08	0.19	4.23
F6-06	Low 14/11	0.02	2.45	0.14	0.91	74.36	16.44	3.10	0.36	1.52	0.07	0.27	0.06	0.08	0.20	4.10
F6-07	Low 14/11	0.03	2.49	0.15	0.94	75.38	15.37	3.45	0.25	1.42	0.06	0.17	0.04	0.07	0.18	3.94
F6-08	Low 14/11	0.02	2.34	0.14	0.89	74.17	16.578	3.21	0.37	1.67	0.10	0.25	0.04	0.07	0.17	3.94
F6-09	Low 14/11	0.03	2.69	0.23	1.10	69.80	20.52	2.73	0.46	1.59	0.13	0.32	0.08	0.12	0.23	4.71
F6-10	Low 14/11	0.02	2.44	0.16	0.92	73.55	16.87	3.39	0.38	1.60	0.09	0.28	0.04	0.07	0.19	4.12
IMC201-01	High 20/17	0.05	3.79	0.18	2.34	77.38	10.98	2.25	0.84	1.30	0.06	0.40	0.02	0.23	0.20	7.64
IMC201-02	High 20/17	0.05	4.30	0.22	1.86	77.13	11.37	2.37	0.69	1.23	0.06	0.36	0.03	0.17	0.17	7.43
IMC201-04	High 20/17	0.05	4.03	0.19	2.05	76.20	12.56	2.31	0.69	1.21	0.06	0.34	0.02	0.15	0.15	7.30
IMC201-05	High 20/17	0.05	4.34	0.22	1.84	76.58	11.93	2.43	0.68	1.20	0.06	0.34	0.02	0.16	0.17	7.40
IMC201-06	High 20/17	0.05	4.06	0.20	2.21	75.83	12.34	2.49	0.78	1.25	0.06	0.36	0.02	0.19	0.15	7.65
IMC201-07	High 20/17	0.05	3.99	0.19	2.16	76.33	12.22	2.27	0.77	1.24	0.05	0.36	0.02	0.17	0.18	7.50
IMC201-08	High 20/17	0.05	3.90	0.19	2.16	75.80	12.94	2.42	0.68	1.22	0.06	0.30	0.02	0.15	0.13	7.22
IMC201-09	High 20/17	0.03	3.41	0.13	2.46	76.72	12.12	2.37	0.72	1.35	0.07	0.30	0.01	0.18	0.12	7.10
IMC201-10	High 20/17	0.05	4.26	0.20	2.06	78.16	10.15	2.02	0.84	1.32	0.05	0.44	0.02	0.25	0.18	7.90
IMC201-01	Low 14/11	0.05	3.76	0.21	1.63	76.15	12.63	2.98	0.61	1.27	0.06	0.34	0.04	0.11	0.16	6.51

IMC201-02	Low 14/11	0.05	3.67	0.20	1.62	76.61	12.52	2.96	0.37	1.32	0.05	0.31	0.03	0.11	0.20	6.13
IMC201-04	Low 14/11	0.06	4.02	0.23	1.47	74.65	13.94	3.03	0.58	1.33	0.06	0.33	0.03	0.13	0.15	6.58
IMC201-05	Low 14/11	0.05	3.97	0.22	1.49	75.43	13.43	2.74	0.58	1.36	0.06	0.34	0.03	0.11	0.18	6.54
IMC201-06	Low 14/11	0.04	3.66	0.20	1.59	75.94	12.96	2.98	0.55	1.32	0.08	0.34	0.05	0.10	0.20	6.28
IMC201-07	Low 14/11	0.02	2.53	0.16	1.09	75.76	15.24	3.20	0.14	1.29	0.08	0.24	0.02	0.07	0.16	4.08
IMC201-08	Low 14/11	0.03	3.49	0.13	1.73	74.49	14.29	3.38	0.40	1.61	0.04	0.21	0.02	0.06	0.13	5.92
IMC201-10	Low 14/11	0.04	3.84	0.21	1.42	75.88	12.93	2.93	0.57	1.43	0.07	0.36	0.020	0.12	0.19	6.35
15.24-01	High 20/17	0.03	3.14	0.12	1.12	77.45	11.38	3.87	0.46	1.71	0.13	0.28	0.04	0.14	0.14	5.17
15.24-02	High 20/17	0.03	3.16	0.14	1.45	76.54	11.27	4.38	0.56	1.70	0.11	0.30	0.05	0.15	0.17	5.65
15.24-03	High 20/17	0.03	3.18	0.14	1.39	77.14	10.63	4.44	0.58	1.70	0.11	0.30	0.03	0.16	0.16	5.64
15.24-04	High 20/17	0.02	3.25	0.12	1.11	76.16	11.90	4.40	0.48	1.79	0.14	0.28	0.04	0.13	0.17	5.28
15.24-05	High 20/17	0.03	3.12	0.12	1.10	77.38	11.11	4.20	0.44	1.81	0.14	0.26	0.04	0.14	0.13	5.08
15.24-06	High 20/17	0.03	2.90	0.13	1.28	76.83	11.53	4.00	0.51	1.90	0.15	0.29	0.05	0.17	0.24	5.18
15.24-07	High 20/17	0.02	3.19	0.13	1.28	75.24	12.39	4.88	0.49	1.70	0.14	0.27	0.03	0.12	0.13	5.37
15.24-08	High 20/17	0.03	3.18	0.13	1.23	76.44	11.21	4.67	0.51	1.83	0.12	0.29	0.04	0.15	0.18	5.39
15.24-09	High 20/17	0.02	3.12	0.14	1.41	77.36	10.36	4.48	0.58	1.75	0.10	0.31	0.03	0.16	0.17	5.60
15.24-10	High 20/17	0.04	3.18	0.14	1.43	76.19	11.33	4.71	0.56	1.67	0.11	0.29	0.05	0.14	0.16	5.64
15.24-02	Low 14/11	0.04	3.09	0.15	0.64	75.62	11.81	5.84	0.37	1.76	0.11	0.27	0.07	0.09	0.16	4.49
15.24-03	Low 14/11	0.03	2.71	0.12	1.04	75.80	11.73	5.74	0.28	1.95	0.07	0.20	0.04	0.11	0.19	4.36
15.24-04	Low 14/11	0.02	2.85	0.11	0.97	76.63	10.60	5.58	0.45	2.00	0.12	0.33	0.06	0.08	0.22	4.69
15.24-06	Low 14/11	0.02	2.86	0.13	1.07	76.75	10.47	5.70	0.44	1.88	0.10	0.30	0.04	0.09	0.15	4.78
15.24-07	Low 14/11	0.03	3.05	0.14	1.22	75.85	11.13	5.99	0.48	1.47	0.11	0.29	0.04	0.07	0.14	5.14
15.24-08	Low 14/11	0.02	2.98	0.13	0.97	75.51	11.78	5.72	0.39	1.84	0.11	0.27	0.04	0.08	0.15	4.71
15.24-09	Low 14/11	0.02	2.98	0.13	1.00	75.11	12.02	5.81	0.42	1.81	0.12	0.28	0.04	0.08	0.19	4.78
15.24-10	Low 14/11	0.01	2.96	0.12	0.89	76.55	11.00	5.53	0.40	1.85	0.12	0.32	0.03	0.08	0.14	4.66

EXAMPLE 9

DH population Skechers

A DH population designated Skechers was obtained from a cross between 15.24 and 06SE-04GX-33. The 06SE-04GX-33 parent line was selected from progeny of a cross between 04GX-33 and 01NM.304. Line 04GX-33, which has an oleic acid content of about 80% and reduced saturated fatty acid content, was produced by crossing 01NM.304 and a European spring growth habit line 'Lila' and developing a DH population from the F₁ cross. Line 01NM.304 was developed from a DH population of an F₁ cross between IMC302 and Surpass 400. 06SE-04GX-33 seeds have a mean C14:0 content of 0.091%, a C16:0 content of 4.47%, a C16:1 content of 0.68%, a C18:0 content of 1.69%, a C18:1 content of 79.52%, a C18:2 content of 6.62%, a C18:3 content of 4.12%, a C20:0 content of 0.63%, a C20:1 content of 1.22%, a C22:0 content of 0.49%, a C22:1 content of 0.0%, a C24:0 content of 0.21%, and a C24:1 content of 0.24%.

This DH population was generated from the cross of 15.24 and 06SE-04GX-33 by collecting microspores, treating the microspores with colchicine, and propagating them *in vitro*. Plantlets formed *in vitro* from the microspores were moved to a greenhouse and inflorescences that formed were self pollinated. Seed was harvested from the DH₁ plants at maturity and analyzed for fatty acid profile via GC. Table 16 contains the fatty acid profile of seeds produced by plants grown in the greenhouse and in the field of DH lines selected from the Skechers population. The fatty acid profile of IMC111RR is included as a control in Table 16. Skechers-159 and Skechers-339 exhibited a low total saturated fatty acid profile in the greenhouse and in the field (Table 16).

TABLE 16

Fatty acid composition of seed of Skechers 339 and Skechers 159

Line	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
Greenhouse															
Skechers-339	0.04	2.86	0.20	1.11	84.53	4.40	3.61	0.48	1.98	0.12	0.27	0.00	0.23	0.17	4.98
Skechers-159	0.03	2.91	0.19	1.26	84.24	4.05	3.57	0.55	1.88	0.15	0.34	0.00	0.00	0.82	5.08
Field															
Skechers-339	0.00	2.55	0.12	0.94	82.64	5.07	5.44	0.39	2.11	0.16	0.28	0.04	0.14	0.13	4.29
Skechers-339	0.00	2.80	0.16	1.22	81.55	5.57	4.89	0.50	2.25	0.21	0.52	0.00	0.19	0.16	5.22
Skechers-339	0.000	3.01	0.22	1.04	79.43	7.39	5.12	0.46	2.21	0.20	0.55	0.04	0.17	0.18	5.23
Mean	0.00	2.79	0.17	1.07	81.20	6.01	5.15	0.45	2.19	0.19	0.44	0.03	0.17	0.16	4.91
Skechers-159	0.03	2.65	0.14	1.03	83.52	5.07	5.09	0.41	2.04	0.00	0.00	0.01	0.01	0.00	4.13
Skechers-159	0.03	2.60	0.15	0.97	82.93	4.80	5.52	0.39	2.16	0.13	0.33	0.00	0.00	0.01	4.32
Skechers-159	0.04	2.69	0.23	0.95	82.99	5.08	5.18	0.39	2.06	0.12	0.28	0.00	0.01	0.00	4.35
Skechers-159	0.04	2.59	0.15	0.90	80.65	5.50	5.48	0.36	2.08	0.12	2.12	0.00	0.00	0.00	6.01
Mean	0.04	2.63	0.17	0.96	82.52	5.11	5.32	0.39	2.08	0.09	0.68	0.00	0.01	0.01	4.70
IMC111RR	0.08	5.06	0.41	2.07	56.80	28.40	3.87	0.83	1.44	0.14	0.50	0.00	0.23	0.16	8.77
IMC111RR	0.09	5.38	0.50	2.09	56.61	28.38	3.50	0.81	1.41	0.13	0.50	0.01	0.53	0.08	9.40
IMC111RR	0.21	6.15	0.50	1.46	47.82	36.03	3.38	0.71	1.24	0.14	0.56	0.00	1.43	0.37	10.52

EXAMPLE 10

Line 1764-43-06 x 1975-90-14

A pedigree selection program was carried out with progeny of a cross of 1764-43-06 x 1975-90-14 over multiple cycles of single plant selections in the greenhouse for low total saturated fatty acid content in seeds. Table 17 contains the seed fatty acid profile of each parent used to make the F₁ cross. Table 18 contains the seed fatty acid profile of selections advanced through the F₆ generation. The mean seed fatty acid profiles of the inbred 01PR06RR.001B and the variety IMC201 are shown for comparison. Additional rounds of self-pollination and selection for low total saturated fatty acids can be performed.

TABLE 17

Fatty acid composition of seed of Lines 1975-90-14 and 1764-43-06

Line	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
1975-90-14	0.00	3.78	0.23	1.54	75.12	14.06	2.08	0.64	1.62	0.09	0.38	0.0	0.27	0.18	6.61
1764-43-06	0.039	3.28	0.31	2.40	75.45	12.97	1.96	0.90	1.54	0.08	0.48	0.0	0.42	0.17	7.52

TABLE 18

Seed Fatty acid composition of F2-F6 generations selected in progeny 1764-43-06 x 1975-90-14

Line	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
F2 seed															
E626033	0.063	4.33	0.63	1.59	63.33	22.87	4.04	0.63	1.50	0.13	0.39	0.00	0.26	0.25	7.26
E626088	0.051	3.41	0.26	1.58	72.76	16.44	2.16	0.64	1.68	0.10	0.39	0.00	0.23	0.30	6.30
E626134	0.042	3.40	0.23	1.66	73.61	15.71	1.96	0.70	1.69	0.09	0.42	0.03	0.26	0.20	6.48
E626082	0.05	3.50	0.26	1.69	72.38	16.58	2.05	0.69	1.61	0.09	0.41	0.00	0.24	0.45	6.58
01PR06RR.001B Mean	0.07	4.73	0.37	2.17	66.27	21.15	2.13	0.87	1.12	0.06	0.49	0.01	0.36	0.21	8.69
F3 seed															
E642092	0.05	3.57	0.32	1.06	60.40	27.06	4.14	0.46	1.85	0.17	0.43	0.00	0.20	0.29	5.77
E642105	0.03	2.98	0.16	1.67	74.64	14.52	2.39	0.67	1.88	0.10	0.39	0.04	0.28	0.24	6.02
E641751	0.04	3.16	0.19	1.40	73.53	15.88	2.57	0.57	1.74	0.12	0.35	0.00	0.23	0.23	5.75
E641767	0.04	2.99	0.18	1.46	72.85	16.25	2.59	0.59	1.92	0.14	0.39	0.06	0.27	0.26	5.74
E642058	0.02	3.56	0.31	1.26	70.79	18.60	2.65	0.51	1.59	0.12	0.30	0.00	0.18	0.11	5.84
E642706	0.00	2.95	0.20	1.49	72.76	16.92	2.58	0.60	1.59	0.11	0.30	0.00	0.23	0.27	5.57
E641983	0.03	3.21	0.23	1.62	71.50	17.62	2.51	0.63	1.74	0.12	0.33	0.00	0.22	0.23	6.05

20°/17°															
Seed from FTF647754:															
FTG603509	0.03	2.66	0.14	1.39	76.57	14.16	2.52	0.53	1.36	0.08	0.28	0.02	0.13	0.13	5.01
FTG603519	0.03	2.56	0.15	1.32	76.7	14.05	2.43	0.57	1.43	0.08	0.32	0.03	0.17	0.16	4.97
FTG603505	0.02	2.47	0.14	1.33	79.5	11.43	2.22	0.58	1.52	0.07	0.34	0.02	0.21	0.16	4.95
FTG603506	0.07	3.59	0.14	2.73	68.43	19.28	3.41	0.76	0.89	0.04	0.30	0.00	0.22	0.13	7.67
FTG603517	0.03	2.66	0.16	1.44	76.75	13.94	2.44	0.56	1.37	0.07	0.29	0.02	0.14	0.13	5.12
FTG603507	0.03	2.63	0.15	1.31	76.59	14.18	2.5	0.53	1.39	0.05	0.29	0.02	0.16	0.18	4.94
FTG603508	0.03	2.51	0.12	1.38	75.88	14.61	2.58	0.56	1.56	0.09	0.33	0.03	0.16	0.15	4.97
FTG603515	0.03	2.74	0.13	1.33	75.67	14.91	2.71	0.49	1.36	0.08	0.25	0.02	0.12	0.14	4.97
FTG603516	0.03	2.65	0.13	1.41	76.32	14.16	2.37	0.6	1.54	0.09	0.34	0.03	0.18	0.16	5.21
FTG603520	0.03	2.72	0.14	1.42	75.61	14.9	2.37	0.57	1.49	0.09	0.32	0.03	0.16	0.15	5.23

EXAMPLE 11

Seed Fatty Acid Profiles for Field-Grown Plants

Plants of 15.24, Salomon-03, Salomon-05, Salomon-07, an F6 selected line described in Example 10, Skechers-159 and Skecher-339 were grown in field plots in Aberdeen, SK, Canada. At maturity, seeds from each line were harvested and fatty acid content determined by GC analysis. The ranges of palmitic, stearic, oleic, linoleic, and linolenic acid content, and the range of total saturated fatty acids are shown in Table 19. The ranges for seed of line Q2 and Pioneer® variety 46A65 are shown for comparison.

TABLE 19

Fatty Acid Profiles for Field-Grown Plants

Genotype	C16:0	C18:0	C18:1	C18:2	C18:3	Total Sats
46A65	3.37-4.12	1.53-2.29	64.85-71.46	13.57-19.16	5.06-7.95	6.24-7.52
Q2	3.53-4.10	1.46-2.10	63.03-70.49	13.79-19.44	6.15-10.28	6.14-7.62
Salomon-07	3.44-4.20	0.71-0.81	73.68-76.74	11.76-13.24	3.66-4.08	4.96-5.97
Salomon-05	3.02-3.34	0.95-1.11	72.74-74.51	13.70-15.94	3.40-4.69	4.34-5.22
15.24	2.77-3.19	0.95-1.06	77.16-77.95	10.76-12.02	3.42-3.68	4.53-5.36
Selection from 1764-43-06 x 1975-90-14	2.42-2.73	0.97-1.29	71.07-73.56	15.65-18.80	2.75-2.91	4.21-5.19
Salomon-03	2.24-2.51	1.08-1.36	72.20-76.70	14.15-18.15	2.03-2.71	4.38-4.81
Skechers-339	2.38-2.84	0.91-1.28	79.93-86.50	3.95-4.90	3.23-4.90	4.03-5.23
Skechers-159	2.37-3.75	0.91-1.26	83.97-86.45	3.49-4.80	4.11-4.47	4.11-4.47

EXAMPLE 12

Radiation Mutagenesis (RMU) of 15.24 germplasm

About 30 grams (~8000 seeds) of M₀ seeds from an individual selected from the DH population of 15.24 x 01OB240 on the basis of low total saturates (see Example 2) were mutagenized using cesium irradiation at 45krad. About 1500 of the mutagenized seeds were planted in the greenhouse immediately after irradiation, about 500 of them developed into plants to produce M₁ seeds. About 840 M₁ seeds were planted and M₂ seed was harvested. M₂ seed was planted along with F₁ progeny plants of a cross of 15.24 x 01OB240 (designated control 1; M₀ seed) were also planted. The fatty acid composition of M₃ seeds produced by individual M₂ plants and control plants was

analyzed by GC. The results are shown in Table 20 under the M2 heading. The individual M₂ plant producing M₃ seeds with the lowest total saturates was 08AP-RMU-tray 3-18, which had 5.28% total saturates compared to 6.48% for control-1. The individual M₂ plant producing M₃ seeds with the lowest 16:0 was 08AP-RMU-tray 13-25, which had 2.55% 16:0 compared with 3.19% for control-1. The individual M₂ plant producing M₃ seeds with the lowest 18:0 was 08AP-RMU-tray 10-34, which had an 18:0 content of 0.93% compared with 1.7% for control-1. M₃ seed used to generate fatty acid profiles shown in Table 20 was planted from these three lines in the greenhouse.

M₄ plants derived from M₃ seed with low total saturates, 16:0, and 18:0, respectively, from each of the three groups were selected for use in crosses. Line M4-L1601-12 had a total saturates content of 5.28% in the M₃ generation and was selected from the 08AP-RMU-tray 3-18 lineage. A cross was made between plants of line M4-L1601-12 and a line containing the homozygous mutant alleles of Isoforms 1, 2, 3, 4 of FatB (described in Example 6). Seed fatty acid profiles from F₂ seeds for two F₁ individuals are shown in Table 20. Plants of lines M4-Lsat1-23 and M4-L1601-22 were crossed, and the fatty acid profile for seeds produced on an F₁ individual designated 09AP-RMU-003-06 are shown in Table 20. M4-Lsat1-23 and M4-L1601-22 were selected from the M₃ generation with total saturate of 5.02% and 16:0 of 2.43%. Plants of lines M4-L1601-12 X M4-D60-2-01 were crossed, and the fatty acid profile for seeds produced on an F₁ individual designated 09AP-RMU-012-2 are shown in Table 20. M4-L1601-12 X M4-D60-2-01 were selected from the M₃ generation with total saturates of 5.28% and 18:0 of 0.88%, respectively. Seeds from F₁ plants with low total saturated fatty acid content, low 16:0, and low 18:0 were grown for further pedigree selection breeding. Some plants were self-pollinated and used to generate DH populations for further selection. It is expected that total saturated fatty acid content in seeds produced on F₂ plants and on progeny of the DH populations will be lower than that in seeds produced on F₁ plants, due to genetic segregation for homozygosity for mutant alleles at loci that confer the low total saturates phenotype.

TABLE 20

Identifier	C140	C160	C161	C180	C181	C182	C183	C200	C201	C202	C220	C221	C240	C241	Total Sats
15.24 x 01OB240 (control 1)	0.04	3.19	0.14	1.7	78.32	10.51	2.23	0.73	2.03	0.13	0.41	0.05	0.39	0.13	6.45
M2															
08AP-RMU-tray13-25	0.03	2.55	0.1	1.68	78.86	10.86	2.14	0.7	2.16	0.14	0.35	0.01	0.3	0.13	5.61
08AP-RMU-tray10-34	0.02	3.08	0.02	0.93	79.4	10.51	2.24	0.61	2.11	0.15	0.4	0.05	0.34	0.14	5.39
08AP-RMU-tray3-18	0.02	2.96	0.1	1.16	80.49	9.68	2.07	0.56	2.12	0.14	0.35	0.04	0.23	0.08	5.28
M4															
M4-L1601-12	0	3.26	0	1.76	72.64	15.82	2.95	0.76	2.13	0.2	0.49	0	0	0	6.26
Salomon-05 (control 2)	0	2.49	0.11	1.67	77.95	10.05	4.25	0.64	1.93	0.12	0.32	0.05	0.23	0.2	5.58
F1 of RMU mutants x RMU mutants															
09AP-RMU-003-06 [M4-Lsat1-23 X M4- L1601-22]	0.03	2.66	0.06	1.47	79.34	10.6	2.11	0.67	2.06	0.15	0.36	0.07	0.25	0.2	5.42
09AP-RMU-012-2 [M4-L1601-12 X M4- D60-2-01]	0.03	2.79	0.11	1.44	77.36	12.4	2.36	0.64	1.93	0.14	0.38	0.05	0.21	0.15	5.5
F1 of RMU mutants x mutant FatB 1,2,3, 4															
09AP-RMU-008-07 [M4-L1601-12 X Iso1234	0.04	2.98	0.18	1.74	72.18	17.38	2.37	0.7	1.44	0.11	0.37	0.04	0.26	0.22	6.09
09AP-RMU-008-05 [M4-L1601-12 X Iso1234	0.02	2.74	0.17	2.08	74.02	15.78	2.14	0.74	1.41	0.11	0.35	0.02	0.24	0.2	6.17

EXAMPLE 13

Development of hybrid canola producing reduced saturated fat seed oil

A hybrid canola variety yielding seeds with a total saturated fatty acid content of less than 6% was produced by introducing genes from the low saturate line 15.24 into a commercially grown hybrid, Victory® v1035. Hybrid v1035 has an average oleic acid content of 65%. Plants of the line 15.24, and the inbreds 01PR06RR.001B and 95CB504, were planted in a greenhouse. Inbred 01PR06RR.001B is the male parent of v1035. Inbred 95CB504 is the B line female parent of v1035. Plants of 01PR06RR.001B and 15.24 were cross pollinated in the greenhouse as were 95CB504 and 15.24, as shown in Table 21.

TABLE 21

Female	x	Male
01PR06RR.001B (R-line)		15.24
95CB504 (B-line)		15.24

F₁ progeny from the cross of 95CB504 and 15.24 were backcrossed to 95CB04 to produce BC₁-B progeny, which were selfed (BC₁S). Plants with low total saturates were selected from the BC₁-B selfed progeny, and backcrossed to 95CB504 to produce BC₂-B progeny. F₁ progeny from the cross of 01PR06RR.001B and 15.24 were backcrossed to 01PR06RR.001B to produce BC₁-R progeny, which were selfed. Plants with low total saturates were selected from the BC₁-R selfed progeny, and backcrossed to 01PR06RR.001B to produce BC₂-R progeny. Backcrossing, selection, and self-pollination of the BC-B and BC-R progeny were continued for multiple generations. The 95CB504 male sterile A line, 00OA05, was converted to a low saturated phenotype in parallel with the conversion of the 95CB504 B line.

Hybrid seed was generated by hand, using BC₁S₃ generation plants of the 95CB504 B line as the female parent and BC₁S₃ generation plants of the 01PR06RR.001B R line as the male parent. The hybrid seed was grown at 5 locations x 4 replications in Western Canada. In the trial plot locations, some individual plants were bagged for self pollination (5 locations x 2 reps) and seeds harvested at maturity. The remaining plants were not bagged (5 locations x 4 reps) and seeds were harvested in bulk. As such, the bulk samples had some level of out crossing with non-low saturate fatty acid

lines in adjacent plots. Seeds from the individual and bulk samples were analyzed for fatty acid content. Seeds from control plants of line Q2, hybrid v1035 and commercial variety 46A65 were also harvested individually and in bulk..

Table 22 shows the fatty acid profile of the individually bagged samples and bulked samples for hybrid 1524 and controls. The results indicate that seed produced by Hybrid 1524 has a statistically significant decrease in 16:0 content and 18:0 content relative to the controls, and a statistically significant increase in 20:1 content relative to controls. In addition, seeds produced by Hybrid 1524 have a statistically significant decrease in total saturated fatty acid content relative to controls. The total saturated fatty acid content for individually bagged plants is about 5.7%, or about 0.8% less than the parent hybrid which lacks the FatA2 mutation contributed by line 15.24. The total saturated fatty acid content for bulk seed is about 5.9%, or more than 0.9% less than the parent hybrid which lacks the FatA2 mutation contributed by line 15.24.

TABLE 22
Seed Fatty Acid Profile

Mean C16:0	N	Line	Mean C18:0	N	Line
3.902 a	11	Q2	1.903 a	16	V1035Bulk
3.876 a	16	Q2Bulk	1.899 a	16	Q2Bulk
3.675 b	16	46A65Bulk	1.887 a	16	46A65Bulk
3.669 b	16	V1035Bulk	1.803 ab	9	46A65
3.594 bc	9	46A65	1.765 b	11	Q2
3.513 cd	10	V1035	1.744 b	10	V1035
3.414 de	16	H1524Bulk	1.405 c	16	H1524Bulk
3.344 e	10	H1524	1.283 d	10	H1524
Mean C20:1	N	Line	Mean Total Sats	N	Line
1.660 a	10	H1524	6.986 a	16	Q2Bulk
1.599 a	16	H1524Bulk	6.875 a	11	Q2
1.421 b	10	V1035	6.859 a	16	V1035Bulk
1.398 b	16	Q2Bulk	6.776 ab	16	46A65Bulk
1.336 b	16	V1035Bulk	6.601 b	9	46A65
1.332 b	16	46A65Bulk	6.568 b	10	V1035
1.331 b	9	46A65	5.911 c	16	H1524Bulk
1.265 b	11	Q2	5.704 d	10	H1524

Another hybrid canola variety yielding seeds with a low total saturated fatty acid content is produced by introducing genes from the low saturate line Skechers-339 into a commercially grown hybrid, using the backcrossing and selection program described above for v1035.

5 Another hybrid canola variety yielding seeds with a low total saturated fatty acid content is produced by crossing F₆ progeny of a cross of 1764-43-06 x 1975-90-14, selected for low total saturates, with the parent inbreds of a commercially grown hybrid. An A line, a B line and an R line are selected for low total saturates, using backcrossing and selection as described above for v1035.

10 Another hybrid canola variety yielding seeds with a low total saturated fatty acid content is produced by crossing Salomon-05, with the parent inbreds of a commercially grown hybrid. An A line, a B line and an R line are selected for low total saturates, using backcrossing and selection as described above for v1035.

15 Another hybrid canola variety yielding seeds with a low total saturated fatty acid content is produced by crossing Iso1234 with the parent inbreds of hybrid 1524. An A line, a B line and an R line are selected for low total saturates, using backcrossing and selection as described above for v1035. The resulting hybrid, designated Hybrid A2-1234, carries a mutant FatA2 allele and mutant FatB alleles at isoforms 1, 2, 3, and 4.

20 Another hybrid canola variety yielding seeds with a low total saturated fatty acid content is produced by crossing a variety homozygous for a mutant Fad2 allele and a mutant Fad3 allele with the parent inbreds of Hybrid A2-1234. An A line, a B line and an R line are selected for low total saturates, using backcrossing and selection as described above for v1035. The resulting hybrid carries a mutant FatA2 allele, mutant FatB alleles at isoforms 1, 2, 3, and 4, a mutant Fad2 allele, and a mutant Fad3 allele.

25 Another hybrid canola variety yielding seeds with a low total saturated fatty acid content is produced by introducing genes from the low saturate line 15.36 into a commercially grown hybrid, using the backcrossing and selection program described above for v1035.

OTHER EMBODIMENTS

5 It is to be understood that while the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims. Other aspects, advantages, and modifications are within the scope of the following claims.

WHAT IS CLAIMED IS:

1. A *Brassica* plant comprising a mutant allele at a fatty acyl-acyl-ACP thioesterase A2 (FATA2) locus, wherein said mutant allele results in the production of a FATA2 polypeptide having reduced thioesterase activity relative to a corresponding wild-type FATA2 polypeptide.
2. The plant of claim 1, wherein said mutant allele comprises a nucleic acid encoding a FATA2 polypeptide having a mutation in a region (SEQ ID NO:29) corresponding to amino acids 242 to 277 of the FATA2 polypeptide.
3. The plant of claim 2, wherein said FATA2 polypeptide comprises a substitution of a leucine residue for proline at position 255.
4. The plant of claim 1, said plant further comprising a mutant allele at a FAD2 locus, said mutant allele comprising a nucleic acid encoding a FAD2 polypeptide having a lysine substituted for glutamic acid in a HECGH motif.
5. The plant of claim 1, said plant further comprising a mutant allele at a FAD2 locus, said mutant allele comprising a nucleic acid encoding a FAD2 polypeptide having a glutamic acid substituted for glycine in the DRDYGILNKV motif or a histidine substituted for leucine in a KYLNNP motif.
6. The plant of claim 4, said plant further comprising a mutant allele at a different FAD2 locus, said mutant allele comprising a nucleic acid encoding a FAD2 polypeptide having a glutamic acid substituted for glycine in the DRDYGILNKV motif or a histidine substituted for leucine in a KYLNNP motif.
7. The plant of claim 1 or claim 6, said plant further comprising mutant alleles at two different FAD3 loci, wherein one of said mutant alleles comprises a nucleic acid encoding a FAD3A polypeptide having a cysteine substituted for arginine at position 275, and wherein one of said mutant alleles comprises a fad3B nucleic acid sequence having a mutation in an exon-intron splice site recognition sequence.
8. The plant of claim 1, said plant further comprising mutant alleles at four different FATB loci.
9. The plant of claim 8, wherein at least one of said FATB mutant alleles comprises a nucleic acid encoding a truncated FATB polypeptide.

10. The plant of claim 9, wherein said nucleic acid encoding said truncated FATB polypeptide comprises a nucleotide sequence selected from the group consisting of: SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, and SEQ ID NO:4.

11. The plant of claim 10, said plant comprising nucleic acids having the nucleotide sequences set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, and SEQ ID NO:4.

12. The plant of claim 1, wherein said plant is a *Brassica napus*, *Brassica juncea*, or *Brassica rapa* plant.

13. The plant of claim 1, wherein said plant is an F1 hybrid.

14. Progeny of the plant of claim 1, said progeny comprising said mutant allele.

15. Seeds of the plant of claim 1.

16. The plant of claim 1, further comprising mutant alleles at two or more different fatty acyl-acyl carrier protein thioesterase B (FATB) loci, wherein each of said FATB mutant alleles results in the production of a FATB polypeptide having reduced thioesterase activity relative to a corresponding wild-type FATB polypeptide.

17. The plant of claim 16, wherein at least one of said FATB mutant alleles comprises a nucleic acid encoding a truncated FATB polypeptide.

18. The plant of claim 16, wherein at least one of said mutant alleles comprises a nucleic acid encoding a FATB polypeptide having a deletion of a helix/4-stranded sheet (4HBT) domain or a portion thereof.

19. The plant of claim 16, wherein at least one of said FATB mutant alleles comprises a nucleic acid encoding a FATB polypeptide having a non-conservative substitution of a residue affecting substrate specificity.

20. The plant of claim 16, wherein at least one of said FATB mutant alleles comprises a nucleic acid encoding a FATB polypeptide having a non-conservative substitution of a residue affecting catalytic activity.

21. The plant of claim 17, wherein said nucleic acid encoding said truncated FATB polypeptide comprises a nucleotide sequence selected from the group consisting of: SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, and SEQ ID NO:4.

22. The plant of claim 16 comprising mutant alleles at three different FATB loci.

23. The plant of claim 16 comprising mutant alleles at four different FATB loci.

24. The plant of claim 21 comprising nucleic acids having the nucleotide sequences set forth in SEQ ID NO:1 and SEQ ID NO:2.

25. The plant of claim 21 comprising nucleic acids having the nucleotide sequences set forth in SEQ ID NO:1 and SEQ ID NO:3.
26. The plant of claim 21 comprising nucleic acids having the nucleotide sequences set forth in SEQ ID NO:1 and SEQ ID NO:4.
27. The plant of claim 21 comprising nucleic acids having the nucleotide sequences set forth in SEQ ID NO:1, SEQ ID NO:2, and SEQ ID NO:3.
28. The plant of claim 21 comprising nucleic acids having the nucleotide sequences set forth in SEQ ID NO:1, SEQ ID NO:2, and SEQ ID NO:4.
29. The plant of claim 21 comprising nucleic acids having the nucleotide sequences set forth in SEQ ID NO:1, SEQ ID NO:3, and SEQ ID NO:4.
30. The plant of claim 16 comprising nucleic acids having the nucleotide sequences set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, and SEQ ID NO:4.
31. The plant of claim 16, said plant producing seeds yielding an oil having a total saturates content of about 2.5% to 5.5%.
32. The plant of claim 16, said plant producing seeds yielding an oil having a palmitic acid content of about 1.5% to 3.5%.
33. The plant of claim 16, said plant producing seeds yielding an oil having a stearic acid content of about 0.5% to 2.5%.
34. The plant of claim 31, said oil further having an oleic acid content of about 78% to 80%, a linoleic acid content of about 8% to 10%, and an α -linolenic acid content of about 2% to 4%.
35. The plant of claim 16, said plant further comprising a mutant allele at a delta-12 fatty acid desaturase (FAD2) locus, said mutant allele comprising a nucleic acid encoding a FAD2 polypeptide having a lysine substituted for glutamic acid in a His-Glu-Cys-Gly-His motif.
36. The plant of claim 16, said plant further comprising a mutant allele at a FAD2 locus, said mutant allele comprising a nucleic acid encoding a FAD2 polypeptide having a glutamic acid substituted for glycine in the DRDYGILNKV motif or a histidine substituted for leucine in a KYLNNP motif.
37. The plant of claim 35, said plant further comprising a mutant allele at a different FAD2 locus, said mutant allele comprising a nucleic acid encoding a FAD2 polypeptide having a glutamic acid substituted for glycine in the DRDYGILNKV motif or a histidine substituted for leucine in a KYLNNP motif.

38. The plant of claim 16, said plant further comprising mutant alleles at two different FAD3 loci, wherein one of said mutant alleles comprises a nucleic acid encoding a FAD3A polypeptide having a cysteine substituted for arginine at position 275, and wherein one of said mutant alleles comprises a fad3B nucleic acid sequence having a mutation in an exon-intron splice site recognition sequence.

39. The plant of claim 16, wherein said plant is a *Brassica napus*, *Brassica juncea*, or *Brassica rapa* plant.

40. The plant of claim 16, wherein said plant is an F1 hybrid.

41. Progeny of the plant of claim 16, said progeny comprising said mutant alleles.

42. Seeds of the plant of claim 16.

43. A method of producing an oil, said method comprising

a) crushing seeds produced from at least one *Brassica* plant of claim 1 or claim 16;

and

b) extracting said oil from said crushed seeds, said oil having, after refining, bleaching, and deodorizing, a total saturates content of about 2.5% to 5.5%.

44. The method of claim 43, wherein said oil further comprises an eicosenoic acid content of about 1.6% to 2.3%.

45. The method of claim 43 or claim 44, wherein said oil further comprises an oleic acid content of about 78% to 80%, a linoleic acid content of about 8% to 10%, and an α -linolenic acid content of about 2% to 4%.

46. The method of any one of claims 43-45, said plant further comprising a mutant allele at a FAD2 locus, said mutant allele comprising a nucleic acid encoding a FAD2 polypeptide having a lysine substituted for glutamic acid in a His-Glu-Cys-Gly-His motif.

47. The method of claim 46, said plant further comprising a mutant allele at a different FAD2 locus, said mutant allele comprising a nucleic acid encoding a FAD2 polypeptide having a glutamic acid substituted for glycine in the DRDYGILNKV motif or a histidine substituted for leucine in a KYLNNP motif.

48. The method of claim 47, said plant further comprising mutant alleles at two different FAD3 loci, wherein one of said FAD3 mutant alleles comprises a nucleic acid encoding a FAD3A polypeptide having a cysteine substituted for arginine at position 275, and wherein one of said FAD3 mutant alleles comprises a fad3B nucleic acid sequence having a mutation in an exon-intron splice site recognition sequence.

49. A method for making a *Brassica* plant, said method comprising

a) obtaining one or more first *Brassica* parent plants comprising mutant alleles at two or more different FATB loci, wherein each said mutant allele results in the production of a FATB polypeptide having reduced thioesterase activity relative to a corresponding wildtype FATB polypeptide;

b) obtaining one or more second *Brassica* parent plants comprising a mutant allele at a FATA2 locus, said mutant allele comprising a nucleic acid encoding a FATA2 polypeptide having a mutation in a region (SEQ ID NO:29) corresponding to amino acids 242 to 277 of the polypeptide;

c) crossing said one or more first *Brassica* parent plants and said one or more second *Brassica* parent plants; and

d) selecting, for one to five generations, for progeny plants having mutant alleles at two or more different FATB loci and a mutant allele at said FATA2 locus thereby obtaining said *Brassica* plant.

50. The method of claim 49, wherein said first *Brassica* parent plant further comprises a mutant allele at a FAD2 locus, said mutant allele comprising a nucleic acid encoding a FAD2 polypeptide having a lysine substituted for glycine in a His-Glu-Cys-Gly-His motif.

51. The method of claim 49, wherein said first *Brassica* parent plant comprises mutant alleles at four different FATB loci.

52. The method of claim 51, wherein said first *Brassica* parent plant further comprises a mutant allele at a FAD2 locus and mutant alleles at two different FAD3 loci, said FAD2 mutant allele comprising a nucleic acid encoding a FAD2 polypeptide having a lysine substituted for glutamic acid in a His-Glu-Cys-Gly-His motif, wherein one of said FAD3 mutant alleles comprises a nucleic acid encoding a FAD3A polypeptide having a cysteine substituted for arginine at position 275, and wherein one of said FAD3 mutant alleles comprises a fad3B nucleic acid sequence having a mutation in an exon-intron splice site recognition sequence.

53. Seeds of a *Brassica* plant comprising a mutant allele at a FATA2 locus, said mutant allele comprising a nucleic acid encoding a FATA2 polypeptide having a mutation in a region (SEQ ID NO:29) corresponding to amino acids 242 to 277 of the polypeptide, said seeds yielding an oil having an oleic acid content of 78% to 80%, a linoleic acid content of

about 8% to 10%, an α -linolenic acid content of no more than about 4%, and an eicosenoic acid content of 1.6 to 2.3%.

54. The seeds of claim 53, wherein said seeds are F2 seeds.

55. The seeds of claim 54, wherein said *Brassica* plant further comprises mutant alleles at four different FATB loci.

56. The seeds of any one of claims 53-55, wherein said *Brassica* plant further comprises a mutant allele at a FAD2 locus and mutant alleles at two different FAD3 loci, said FAD2 mutant allele comprising a nucleic acid encoding a FAD2 polypeptide having a lysine substituted for glutamic acid in a His-Glu-Cys-Gly-His motif, wherein one of said FAD3 mutant alleles comprises a nucleic acid encoding a FAD3A polypeptide having a cysteine substituted for arginine at position 275, and wherein one of said FAD3 mutant alleles comprises a fad3B nucleic acid sequence having a mutation in an exon-intron splice site recognition sequence.