

MUTANT OF PORIN MONOMER, PROTEIN PORE, AND USE THEREOF

TECHNICAL FIELD

5 The present invention belongs to the technical field of characterization of target analyte properties, and particularly relates to a mutant of a porin monomer, a protein pore comprising same, and use thereof in the detection of a target analyte.

BACKGROUND

10 With the research on the structure and sequence of nucleic acids, nucleic acid sequencing technologies have been continuously developed, become the core field of life science research, and play a great driving role in the technical development in the fields of biology, chemistry, electricity, life science, medicine, and the like. It is one of the hot spots of the
15 post-Human Genome Project to develop a novel, rapid, accurate, low-cost, high-precision, and high-throughput nucleic acid sequencing technology by using nanopores.

Nanopore sequencing technology, also known as the fourth generation
20 sequencing technology, is a gene sequencing technology that uses a single-stranded nucleic acid molecule as a sequencing unit, utilizes a nanopore capable of providing an ion current channel, enables the single-stranded nucleic acid molecule to pass through the nanopore driven by electrophoresis, reduces the current of the nanopore when the nucleic
25 acid passes through the nanopore, and reads sequence information in real time for different generated signals.

The nanopore sequencing is mainly characterized in that the reading length is very long, the accuracy rate is relatively high, and most error regions
30 occur in homopolymeric oligonucleotide regions. The nanopore

sequencing can not only realize natural DNA and RNA sequencing, but also directly acquire base modification information about DNA and RNA. For example, methylated cytosine can be directly read by the nanopore sequencing, and bisulfite treatment on genomes is not needed in advance
5 like the second generation sequencing method, which greatly promotes the direct research on epigenetic phenomenon at the genome level. As a novel platform, the nanopore detection technology has the advantages of low cost, high throughput, no label, and the like.

10 Nanopore analysis technology originated from the invention of the Coulter counter and the single-channel current record technology. In 1976, Neher and Sakamann, Nobel Prize winners in Physiology and Medicine, utilized patch clamp technology to measure membrane potential and research membrane proteins and ion channels, thereby promoting the practical
15 application process of the nanopore sequencing technology. In 1996, Kasianowicz et al. proposed a new idea for DNA sequencing by using α -hemolysin, which is a milestone marker of single molecule sequencing by biological nanopores. Subsequently, the research reports on the biological nanopores such as MspA porin, bacteriophage Phi29 connector,
20 and the like enrich the research on the nanopore analysis technology. In 2001, Li et al. opened a new era of solid-state nanopore research. Limited by the development of the semiconductor and material industries, solid-state nanopore sequencing has progressed slowly.

25 One of the key points of the nanopore sequencing technology lies in the design of a special biological nanopore, in which a reading head structure formed in a constriction zone of a pore can cause the blockage of a pore channel current when a single-stranded nucleic acid (such as ssDNA) molecule passes through the nanopore, thereby transiently affecting the
30 intensity of the current flowing through the nanopore (the amplitude of the

current change affected by each base is different), and finally, a high-sensitivity electronic device detects these changes to identify the passed bases. At present, protein pores are used as nanopores for sequencing, and porins are mainly derived from *Escherichia coli*.

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Currently, the nanoporin is single, so it is necessary to develop an alternative nanoporin to achieve the nanopore sequencing technology. The porin is also closely related to sequencing precision, and the porin is also involved in a mode change in the interaction with a rate-controlling protein. Therefore, further optimizing the stability of an interaction interface between the porin and the rate-controlling protein has a positive effect on improving the consistency and stability of sequencing data. The accuracy rate of the nanopore sequencing technology also needs to be improved, and therefore, it is necessary to develop an improved nanoporin to further improve the resolution of the nanopore sequencing.

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SUMMARY

In order to solve the problems described above, embodiments of the present invention are intended to provide an alternative mutant of a porin monomer, a protein pore comprising same, and use thereof.

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In a first aspect, embodiments of the present invention provide a mutant of a porin monomer, wherein an amino acid of the mutant of the porin monomer comprises or consists of a sequence set forth in SEQ ID NO: 1 or a sequence having at least 99%, 98%, 97%, 96%, 95%, 90%, 80%, 70%, 60%, or 50% identity thereto, and the amino acid of the mutant of the porin monomer comprises mutations at one or more positions in T71, S75, or F76 corresponding to SEQ ID NO: 1; one or more of T71, S75, and F76 are specifically T71, S75, F76, T71 and S75, S75 and F76, T71 and S76, or T71, S75 and F76.

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Preferably, the amino acid of the mutant of the porin monomer comprises mutations at one or more positions corresponding to 62-175, 62-104, 68-175, 64-79, 71-76, or 69-76 of SEQ ID NO: 1.

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Preferably, the amino acid of the mutant of the porin monomer comprises: (1) an insertion, a deletion, and/or a substitution of an amino acid at one or more positions corresponding to K69, P70, T71, P72, A73, S74, S75, and F76 of SEQ ID NO: 1; (2) an insertion, a deletion, and/or a substitution of an amino acid at one or more positions corresponding to Q64, T65, G66, Q67, Y68, K69, P70, T71, P72, A73, S74, S75, F76, S77, T78, and S79 of SEQ ID NO: 1; (3) an insertion, a deletion, and/or a substitution of an amino acid at one or more positions corresponding to R62, D63, Y68, K69, T71, S75, F76, and E104 of SEQ ID NO: 1; or (4) an insertion, a deletion, and/or a substitution of an amino acid at one or more positions corresponding to Y68, K69, P70, T71, P72, A73, S74, S75, F76, E171, and D175 of SEQ ID NO: 1.

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In one embodiment, the amino acid mutation of the mutant of the porin monomer is selected from the group consisting of:

(a) mutations from amino acids KPTPASSF corresponding to positions 69-76 of SEQ ID NO: 1 to $M_1M_2M_3M_4M_5M_6M_7M_8$, wherein M_1 is selected from 0 to 3 of R, K, and H; M_2 is selected from 0 to 1 of P; M_3 is selected from 0 to 10 of S, G, C, U, T, M, A, V, L, and I; M_4 is selected from 0 to 1 of P; M_5 is selected from 0 to 5 of A, G, V, L, and I; M_6 is selected from 0 to 5 of S, C, U, T, and M; M_7 is selected from 0 to 10 of A, T, G, V, L, I, S, C, U, and M; M_8 is selected from 0 to 7 of Q, D, E, N, K, H, and R;

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(b) mutations from amino acids QTGQYKPTPASSFSTS corresponding to positions 64-79 of SEQ ID NO: 1 to $M_9M_{10}M_{11}M_{12}M_{13}M_{14}M_{15}M_{16}M_{17}M_{18}M_{19}M_{20}M_{21}M_{22}M_{23}M_{24}$, wherein M_9 is selected from 0 to 5 of L, G,

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A, V, and I; M₁₀ is selected from 0 to 5 of T, S, C, U, and M; M₁₁ is selected from 0 to 5 of G, A, V, L, and I; M₁₂ is selected from 0 to 4 of Q, D, E, and N; M₁₃ is selected from 0 to 3 of Y, F, and W; M₁₄ is selected from 0 to 3 of R, H, and K; M₁₅ is selected from 0 to 1 of P; M₁₆ is selected from 0 to 5 of S, C, U, T, and M; M₁₇ is selected from 0 to 1 of P; M₁₈ is selected from 0 to 5 of A, G, V, L, and I; M₁₉ is selected from 0 to 5 of S, C, U, T, and M; M₂₀ is selected from 0 to 5 of A, G, V, L, and I; M₂₁ is selected from 0 to 9 of N, D, E, Q, L, G, A, V, and I; M₂₂ is selected from 0 to 5 of S, C, U, T, and M; M₂₃ is selected from 0 to 5 of T, S, C, U, and M; M₂₄ is selected from 0 to 5 of A, G, V, L, and I;

(c) a mutation corresponding to position 62 of SEQ ID NO: 1 being 0 to 5 of S, C, U, T, and M; a mutation at position 63 being 0 to 5 of V, G, A, L, and I; a mutation at position 68 being 0 to 2 of F and W; a mutation at position 69 being 0 to 2 of R and H; a mutation at position 71 being 0 to 4 of S, C, U, and M; a mutation at position 75 being 0 to 5 of A, G, V, L, and I; a mutation at position 76 being 0 to 4 of Q, D, E, and N; a mutation at position 104 being 0 to 5 of V, G, A, L, and I; and

(d) mutations from amino acids YKPTPASSF corresponding to positions 68-76 of SEQ ID NO: 1 to M₂₅M₂₆M₂₇M₂₈M₂₉M₃₀M₃₁M₃₂M₃₃, a mutation at position 171 being 0 to 3 of N, D, and Q, and a mutation at position 175 being 0 to 3 of N, E, and Q, wherein M₂₅ is selected from 0 to 3 of F, Y, and W; M₂₆ is selected from 0 to 3 of R, H, and K; M₂₇ is selected from 0 to 1 of P; M₂₈ is selected from 0 to 5 of S, C, U, T, and M; M₂₉ is selected from 0 to 1 of P; M₃₀ is selected from 0 to 5 of A, G, V, L, and I; M₃₁ is selected from 0 to 5 of S, C, U, T, and M; M₃₂ is selected from 0 to 5 of A, G, V, L, and I; M₃₃ is selected from 0 to 4 of Q, D, E, and N.

In one embodiment, the amino acid mutation of the mutant of the porin monomer is selected from the group consisting of:

(a) mutations from amino acids KPTPASSF corresponding to positions

69-76 of SEQ ID NO: 1 to M₁M₂M₃M₄M₅M₆M₇M₈, wherein M₁ is selected from R, K, or H; M₂ is selected from P; M₃ is selected from S, G, C, U, T, M, A, V, L, or I; M₄ is selected from P; M₅ is selected from A, G, V, L, or I; M₆ is selected from S, C, U, T, or M; M₇ is selected from A, T, G, V, L, I, S, C, U, or M; M₈ is selected from Q, D, E, N, K, H, or R;

(b) mutations from amino acids QTGQYKPTPASSFSTS corresponding to positions 64-79 of SEQ ID NO: 1 to M₉M₁₀M₁₁M₁₂M₁₃M₁₄M₁₅M₁₆M₁₇M₁₈M₁₉M₂₀M₂₁M₂₂M₂₃M₂₄, wherein M₉ is selected from L, G, A, V, or I; M₁₀ is selected from T, S, C, U, or M; M₁₁ is selected from G, A, V, L, or I; M₁₂ is selected from Q, D, E, or N; M₁₃ is selected from Y, F, or W; M₁₄ is selected from R, H, or K; M₁₅ is selected from P; M₁₆ is selected from S, C, U, T, or M; M₁₇ is selected from P; M₁₈ is selected from A, G, V, L, or I; M₁₉ is selected from S, C, U, T, or M; M₂₀ is selected from A, G, V, L, or I; M₂₁ is selected from N, D, E, Q, L, G, A, V, or I; M₂₂ is selected from S, C, U, T, or M; M₂₃ is selected from T, S, C, U, or M; M₂₄ is selected from A, G, V, L, or I;

(c) a mutation from R62 corresponding to SEQ ID NO: 1 to R62S, R62C, R62U, R62T, or R62M; a mutation from D63 to D63V, D63G, D63A, D63L, or D63I; a mutation from Y68 to Y68F or Y68W; a mutation from K69 to K69R or K69H; a mutation from T71 to T71S, T71C, T71U, or T71M; a mutation from S75 to S75A, S75G, S75V, S75L, or S75I; a mutation from F76 to F76Q, F76D, F76E, or F76N; a mutation from E104 to E104V, E104G, E104A, E104L, or E104I; and

(d) mutations from amino acids YKPTPASSF corresponding to positions 68-76 of SEQ ID NO: 1 to M₂₅M₂₆M₂₇M₂₈M₂₉M₃₀M₃₁M₃₂M₃₃, a mutation from E171 to E171N, E171D, or E171Q, and a mutation from D175 to D175N, D175E, or D175Q, wherein M₂₅ is selected from F, Y, or W; M₂₆ is selected from R, H, or K; M₂₇ is selected from P; M₂₈ is selected from S, C, U, T, or M; M₂₉ is selected from P; M₃₀ is selected from A, G, V, L, or I; M₃₁ is selected from S, C, U, T, or M; M₃₂ is selected from A, G, V, L,

or I; M₃₃ is selected from Q, D, E, or N.

In one embodiment, the amino acid mutation of the mutant of the porin monomer is selected from the group consisting of:

- 5 (a) mutations from the amino acids KPTPASSF corresponding to positions 69-76 of SEQ ID NO: 1 to RPSPASAQ;
- (b) mutations from the amino acids KPTPASSF corresponding to positions 69-76 of SEQ ID NO: 1 to KPGPASTK;
- (c) mutations from the amino acids QTGQYKPTPASSFSTS corresponding to positions 64-79 of SEQ ID NO: 1 to LTGQYRPSPASANSTA;
- 10 (d) mutations from the amino acids QTGQYKPTPASSFSTS corresponding to positions 64-79 of SEQ ID NO: 1 to LTGQYRPSPASALSTA;
- (e) R62S, D63V, Y68F, K69R, T71S, S75A, F76Q, and E104V corresponding to SEQ ID NO: 1; and
- 15 (f) mutations from the amino acids YKPTPASSF corresponding to positions 68-76 of SEQ ID NO: 1 to FRPSPASAQ, and E171N and D175N corresponding to SEQ ID NO: 1.

In a second aspect, embodiments of the present invention provide a protein pore comprising at least one mutant of the porin monomer.

20 In a third aspect, embodiments of the present invention provide a complex for characterizing a target analyte, which comprises the protein pore and a rate-controlling protein bound thereto.

In a fourth aspect, embodiments of the present invention provide a nucleic acid encoding the mutant of the porin monomer, the protein pore, or the complex.

25 In a fifth aspect, embodiments of the present invention provide a vector or a genetically engineered host cell comprising the nucleic acid.

In a sixth aspect, embodiments of the present invention provide use of the mutant of the porin monomer or the protein pore, the complex, the nucleic acid, or the vector or host cell thereof in the detection of the presence,

absence, or one or more characteristics of a target analyte or in the preparation of a product for detecting the presence, absence, or one or more characteristics of a target analyte.

5 In a seventh aspect, embodiments of the present invention provide a method for producing a protein pore or a polypeptide thereof, comprising transforming the host cell with the vector, and inducing the host cell to express the protein pore or the polypeptide thereof.

10 In an eighth aspect, embodiments of the present invention provide a method for determining the presence, absence, or one or more characteristics of a target analyte, comprising:

- a. contacting the target analyte with the protein pore, the complex, or the protein pore in the complex, such that the target analyte moves relative to the protein pore; and
- 15 b. acquiring one or more measurements when the target analyte moves relative to the protein pore, thereby determining the presence, absence, or one or more characteristics of the target analyte.

20 In one embodiment, the method comprises: the target analyte interacting with the protein pore present in a membrane, such that the target analyte moves relative to the protein pore.

In one embodiment, the target analyte is a nucleic acid molecule.

25 In one embodiment, the method for determining the presence, absence, or one or more characteristics of a target analyte comprises coupling the target analyte to a membrane; and the target analyte interacting with the protein pore present in the membrane, such that the target analyte moves relative to the protein pore.

30 In a ninth aspect, embodiments of the present invention provide a kit for

determining the presence, absence, or one or more characteristics of a target analyte, comprising the mutant of the porin monomer, the protein pore, the complex, the nucleic acid, or the vector or host cell, and a component of the membrane.

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In a tenth aspect, embodiments of the present invention provide a device for determining the presence, absence, or one or more characteristics of a target analyte, comprising the protein pore or the complex, and the membrane.

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In one embodiment, the target analyte includes a polysaccharide, a metal ion, an inorganic salt, a polymer, an amino acid, a peptide, a protein, a nucleotide, an oligonucleotide, a polynucleotide, a dye, a drug, a diagnostic agent, an explosive, or an environmental contaminant;

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preferably, the target analyte comprises a polynucleotide;

more preferably, the polynucleotide comprises DNA or RNA; and/or, the one or more characteristics are selected from (i) a length of the polynucleotide; (ii) an identity of the polynucleotide; (iii) a sequence of the polynucleotide; (iv) a secondary structure of the polynucleotide; and (v) whether the polynucleotide is modified; and/or, the rate-controlling protein in the complex comprises a polynucleotide binding protein.

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BRIEF DESCRIPTION OF THE DRAWINGS

The drawings described are only schematic rather than restrictive.

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FIG. 1 shows the basic working principle of a nanopore according to one embodiment.

FIG. 2 shows a schematic diagram of DNA sequencing according to one embodiment.

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FIG. 3 shows a corresponding pore-blocking signal when a nucleotide

passes through a protein pore according to one embodiment.

FIGs. 4A, 4B, and 4C show a channel surface structure and a ribbon diagram model of a wild-type protein pore according to one embodiment. FIG. 4A is a side view of the surface structure model, FIG. 4B is a top view of the surface structure model, and FIG. 4C is the ribbon structure model.

FIGs. 5A and 5B show amino acid model diagrams of a mutant pore 1 according to one embodiment, wherein FIG. 5A is a top view and FIG. 5B is a side view.

FIG. 6 shows dimensional information about each portion of the mutant pore 1 according to one embodiment.

FIG. 7 shows a monomer amino acid model diagram of the porin mutant 1 according to one embodiment, (b) being the core amino acid composition of a constriction zone (eye loop) shown enlarged in (a).

FIG. 8 shows negative staining electron microscopy results for the porin mutant 1 according to one embodiment.

FIG. 9A shows a cryogenic electron micrograph of the porin mutant 1 according to one embodiment, and FIG. 9B shows 2D classification results.

FIGs. 10A and 10B show locally refined Fourier shell correlation (FSC) results of the porin mutant 1 according to one embodiment, wherein a is a rIn FSC unshielded pattern; b is a rIn FSC phase random shielding pattern; c is a rIn FSC correction pattern; d is a rIn FSC shielding pattern.

FIG. 11 shows an electron density diagram of the porin mutant 1 after three-dimensional reconstruction at a resolution of 2.2 Å by cryogenic electron microscopy according to one embodiment.

FIG. 12 shows an electron density map of the porin mutant 1 at a resolution of 2.2 Å according to one embodiment.

FIG. 13 shows the structure of a DNA construct, BS7-4C3-SE1, according to one embodiment.

FIG. 14 shows the structure of a DNA construct, BS7-4C3-PLT, according to one embodiment.

FIG. 15A shows an opening current and gated features of the mutant pore 1 at a voltage of ± 180 mV according to one embodiment.

5 FIG. 15B shows a scenario in which a nucleic acid passes through the pore of the mutant pore 1 at a voltage of +180 mV according to one embodiment.

FIGs. 16A and 16B show example current trajectories when helicase Mph-MP1-E105C/A362C controls the translocation of the DNA construct
10 BS7-4C3-PLT through the mutant pore 1 according to one embodiment.

FIG. 17 is an enlarged area display of a single signal of the embodiment in FIG. 16B.

FIG. 18A shows an opening current and gated features of a mutant pore 2 at a voltage of ± 180 mV according to one embodiment.

15 FIG. 18B shows a scenario in which a nucleic acid passes through the pore of the mutant pore 2 at a voltage of +180 mV according to one embodiment.

FIGs. 19A and 19B show example current trajectories when the helicase Mph-MP1-E105C/A362C controls the translocation of the DNA construct
20 BS7-4C3-PLT through the mutant pore 2 according to one embodiment.

FIG. 20 is an enlarged area display of a single signal of the embodiment in FIGs. 19A and B.

FIG. 21A shows an opening current and gated features of a mutant pore 3 at a voltage of ± 180 mV according to one embodiment.

25 FIG. 21B shows a scenario in which a nucleic acid passes through the pore of the mutant pore 3 at a voltage of +180 mV according to one embodiment.

FIGs. 22A and 22B show example current trajectories when the helicase Mph-MP1-E105C/A362C controls the translocation of the DNA construct
30 BS7-4C3-PLT through the mutant pore 3 according to one embodiment.

FIG. 23 is an enlarged area display of a single signal of the embodiment in FIG. 22A.

FIG. 24A shows an opening current and gated features of a mutant pore 4 at a voltage of ± 180 mV according to one embodiment.

5 FIG. 24B shows a scenario in which a nucleic acid passes through the pore of the mutant pore 4 at a voltage of +180 mV according to one embodiment.

FIGs. 25A and 25B show example current trajectories when the helicase Mph-MP1-E105C/A362C controls the translocation of the DNA construct
10 BS7-4C3-PLT through the mutant pore 4 according to one embodiment.

FIG. 26 is an enlarged area display of a single signal of the embodiment in FIGs. 25A and 25B.

FIG. 27 shows an opening current and gated features of a mutant pore 5 at a voltage of ± 180 mV according to one embodiment.

15 FIG. 28 shows an example current trajectory when the helicase Mph-MP1-E105C/A362C controls the translocation of the DNA construct BS7-4C3-PLT through the mutant pore 5 according to one embodiment.

FIG. 29 is an enlarged area display of a single signal of the embodiment in FIG. 28.

20 FIG. 30A shows an opening current and gated features of a mutant pore 6 at a voltage of ± 180 mV according to one embodiment.

FIG. 30B shows a scenario in which a nucleic acid passes through the pore of the mutant pore 6 at a voltage of +180 mV according to one embodiment.

25 FIGs. 31A and 31B show example current trajectories when the helicase Mph-MP1-E105C/A362C controls the translocation of the DNA construct BS7-4C3-PLT through the mutant pore 6 according to one embodiment.

FIG. 32 is an enlarged area display of a single signal of the embodiment in FIG. 31A.

30 FIG. 33 shows SDS-PAGE electrophoresis results of the mutant 1

according to one embodiment.

FIG. 34 shows a size exclusion chromatogram of the mutant 1 protein according to one embodiment.

5 DETAILED DESCRIPTION

It should be understood that unused applications of the disclosed products and methods may be adapted according to the particular needs in the art. It should also be understood that the terms used herein are for the purpose of describing particular embodiments of the present invention only, and are
10 not intended to be limiting.

In addition, as used in this specification and the claims, the singular forms “a”, “an”, and “the” include plural referents, unless otherwise specified clearly in the context. For example, reference to “a nucleotide” includes
15 two or more nucleotides, and reference to “a helicase” includes two or more helicases.

As used herein, the term “comprising” means that any of the listed elements must be included, and that other elements may also optionally be
20 included. “Consisting of...” means excluding all unlisted elements. Embodiments defined by each of these terms are within the scope of the present invention.

As used herein, a “nucleotide sequence”, “DNA sequence”, or “nucleic
25 acid molecule” refers to a polymeric form of nucleotides (ribonucleotides or deoxyribonucleotides) of any length. The term only refers to the primary structure of the molecule. Thus, the term includes double-stranded and single-stranded DNA and RNA.

30 The term “nucleic acid” as used herein refers to a single-stranded or

double-stranded covalently linked nucleotide sequence in which the 3' and 5' ends on each nucleotide are linked by phosphodiester bonds. A nucleotide may consist of deoxyribonucleotide bases or ribonucleotide bases. Nucleic acids may include DNA and RNA, and may be prepared synthetically *in vitro* or isolated from natural sources. Nucleic acids may further include modified DNA or RNA, such as methylated DNA or RNA, or RNA that has been subjected to post-translational modification, for example, 5'-capping with 7-methylguanosine, and 3'-end processing, such as cleavage and polyadenylation, and splicing. Nucleic acids may also include synthetic nucleic acids (XNA), such as a hexitol nucleic acid (HNA), a cyclohexene nucleic acid (CeNA), a threose nucleic acid (TNA), a glycerol nucleic acid (GNA), a locked nucleic acid (LNA), and a peptide nucleic acid (PNA). The size of a nucleic acid (or polynucleotide) is generally expressed in terms of the number of base pairs (bp) of a double-stranded polynucleotide, or in the case of a single-stranded polynucleotide, in terms of the number of nucleotides (nt). One thousand bp or nt equals one kilobase pair (kb). Polynucleotides of less than about 40 nucleotides in length are generally referred to as "oligonucleotides" and may comprise primers for use in DNA manipulation, for example, by polymerase chain reaction (PCR).

A polynucleotide, such as a nucleic acid, is a macromolecule comprising two or more nucleotides. The polynucleotide or nucleic acid may comprise any combination of any nucleotides. The nucleotides may be naturally occurring or synthetic. One or more nucleotides in the polynucleotide may be oxidized or methylated. One or more nucleotides in the polynucleotide may be damaged. For example, the polynucleotide may comprise a pyrimidine dimer. This dimer is generally associated with the damage caused by ultraviolet light and is the major cause of cutaneous melanoma. One or more nucleotides in the polynucleotide may be modified, for

example, with a conventional label or tag. The polynucleotide may comprise one or more nucleotides that are abasic (i.e., lack nucleobases), or lack nucleobases and sugars (i.e., C3).

5 The nucleotides in the polynucleotide may be linked to each other in any manner. The nucleotides are generally linked by glycosyl and phosphate groups thereof, as in the nucleic acid. The nucleotides may be linked by nucleobases thereof, as in the pyrimidine dimer.

10 The polynucleotide may be single-stranded or double-stranded. At least a portion of the polynucleotide is preferably double-stranded. The polynucleotide may be a nucleic acid, such as deoxyribonucleic acid (DNA) or ribonucleic acid (RNA). The polynucleotide may comprise an RNA strand that is hybridized to a DNA strand. The polynucleotide may
15 be any synthetic nucleic acid known in the art, such as a peptide nucleic acid (PNA), a glycerol nucleic acid (GNA), a threose nucleic acid (TNA), a locked nucleic acid (LNA), or other synthetic polymers having nucleotide side chains. The PNA backbone is composed of repeating N-(2-aminoethyl)-glycine units linked by peptide bonds. The GNA
20 backbone is composed of repeating ethylene glycol units linked by phosphodiester bonds. The TNA backbone is composed of repeating threose sugars linked together by phosphodiester bonds. LNA is formed from the ribonucleic acid described above and has an additional bridging structure linking the 2' oxygen and the 4' carbon in the ribose moiety.
25 Bridged nucleic acids (BNAs) are modified RNA nucleotides. They may also be referred to as restricted or inaccessible RNA13BNA monomers that may contain a 5-, 6-, or even 7-membered bridging structure and have a “fixed” C3'-endo sugar puckering structure. The bridging structure is synthetically introduced into the position 2',4' of the ribose to produce the
30 2',4'-BNA monomer.

The polynucleotide is most preferably ribonucleic acid (RNA) or deoxyribonucleic acid (DNA). The polynucleotide may be of any length. For example, the polynucleotide may be of at least 10, at least 50, at least 100, at least 150, at least 200, at least 250, at least 300, at least 400, or at least 500 nucleotides or nucleotide pairs in length. The polynucleotide may be of 1000 or more nucleotides or nucleotide pairs, 5000 or more nucleotides or nucleotide pairs, or 100000 or more nucleotides or nucleotide pairs in length.

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Any number of polynucleotides may be studied. For example, the methods of the embodiments may involve the characterization of 2, 3, 4, 5, 6, 7, 8, 9, 10, 20, 30, 50, 100, or more polynucleotides. If two or more polynucleotides are characterized, they may be different polynucleotides or the same polynucleotide.

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The polynucleotides may be naturally occurring or synthetic. For example, the method may be used to verify the sequence of the prepared oligonucleotides. The method is generally performed *in vitro*.

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In the context of the present disclosure, the term “amino acid” is used in its broadest sense and is meant to include organic compounds containing amine (NH₂) and carboxyl (COOH) functional groups as well as side chains unique to each amino acid (e.g., R groups). In some embodiments, the amino acid refers to naturally occurring L α -amino acids or residues. Commonly used single and three-letter abbreviations for the naturally occurring amino acids as used herein are as follows: A = Ala; C = Cys; D = Asp; E = Glu; F = Phe; G = Gly; H = His; I = Ile; K = Lys; L = Leu; M = Met; N = Asn; P = Pro; Q = Gln; R = Arg; S = Ser; T = Thr; V = Val; W = Trp; and Y = Tyr (Lehninger, A. L., (1975) BioChemistry, 2nd edition,

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pages 71-92, Worth Publishers, New York). The common term “amino acid” further includes D-amino acids, retro-inverso amino acids, chemically modified amino acids (such as amino acid analogs), naturally occurring amino acids that are not generally incorporated into proteins (such as norleucine), and chemically synthesized compounds (such as β -amino acids) that have properties to be characteristic of amino acids known in the art. For example, included within the definition of amino acids are analogs or mimetics of phenylalanine or proline that allow the same conformational restriction on peptide compounds as native Phe or Pro. Such analogs and mimetics are referred to herein as “functional equivalents” of the corresponding amino acids. Other examples of amino acids are listed in Roberts and Vellaccio, *The Peptides: Analysis, Synthesis, Biology*, edited by Gross and Meiehofer, Volume 5, page 341, Academic Press, Inc., N. Y. 1983, which is incorporated herein by reference.

The terms “protein,” “polypeptide”, and “peptide” are further used interchangeably herein and refer to a polymer of amino acid residues as well as a variant and synthetic analog of amino acid residues. Thus, these terms apply to amino acid polymers in which one or more amino acid residues are synthetic non-naturally occurring amino acids, such as chemical analogs of corresponding naturally occurring amino acids, as well as to naturally occurring amino acid polymers. The polypeptide may also be subjected to maturation or post-translational modification processes, which may include, but are not limited to: glycosylation, proteolytic cleavage, lipidation, signal peptide cleavage, propeptide cleavage, phosphorylation, and the like.

“Homologs” of a protein encompass peptides, oligopeptides, polypeptides, proteins, and enzymes having amino acid substitutions, deletions, and/or

insertions relative to the unmodified or wild-type protein in discussion and having similar biological and functional activities to the unmodified protein from which they are derived. As used herein, the term “amino acid identity” refers to the degree to which sequences are identical on an amino acid-to-amino acid basis in a comparison window. Thus, the “percent sequence identity” is calculated by the following steps: comparing two optimally aligned sequences in a comparison window; determining the number of positions in which amino acid residues (e.g., Ala, Pro, Ser, Thr, Gly, Val, Leu, Ile, Phe, Tyr, Trp, Lys, Arg, His, Asp, Glu, Asn, Gln, Cys, and Met) are identical in the two sequences to yield the number of matched positions; dividing the number of matched positions by the total number of positions in the comparison window (i.e., the window size); and multiplying the result by 100 to yield the percent sequence identity.

The sequence identity may also be a fragment or portion of a full-length polynucleotide or polypeptide. Thus, a sequence may have only 50% overall sequence identity to a full-length reference sequence, but the sequence of a particular region, domain, or subunit may have 80%, 90%, or up to 99% sequence identity to the reference sequence.

The term “wild-type” refers to a gene or gene product isolated from a naturally occurring source. The wild-type gene is a gene most commonly observed in a population and is therefore arbitrarily designed as the “normal” or “wild-type” form of the gene. Conversely, the term “modified,” “mutation”, or “variant” refers to a gene or gene product that exhibits sequence modification (e.g., substitution, truncation, or insertion), post-translational modification, and/or functional properties (e.g., altered characteristics) compared to the wild-type gene or gene product. It is noted that naturally occurring mutants may be isolated. These mutants are identified by the fact that they have altered characteristics compared to the

wild-type gene or gene product. Methods for introducing or substituting naturally occurring amino acids are well known in the art. For example, methionine (M) may be substituted with arginine (R) by replacing the codon of methionine (ATG) with the codon of arginine (CGT) at the relevant position in the polynucleotide encoding the mutated monomer. Methods for introducing or substituting non-naturally occurring amino acids are also well known in the art. For example, the non-naturally occurring amino acids may be introduced by including synthetic aminoacyl-tRNA in the IVTT system for expressing the mutated monomer. Alternatively, the non-naturally occurring amino acids may be introduced by expressing the mutated monomer in *Pseudomonas taeanensis* MS-3, which is auxotrophic for particular amino acids in the presence of synthetic (i.e., non-naturally occurring) analogs of those specific amino acids. If the mutated monomers are generated using partial peptide synthesis, they may also be generated by naked linkage. A conservative substitution is the replacement of amino acids to other amino acids having similar chemical structures, similar chemical properties, or similar side chain volumes. The amino acids introduced may have similar polarity, hydrophilicity, hydrophobicity, basicity, acidity, neutrality, or charge to the amino acids they replace. Alternatively, the conservative substitution may be the introduction of another aromatic or aliphatic amino acid in place of a pre-existing aromatic or aliphatic amino acid. Conservative amino acid changes are well known in the art and may be selected in accordance with the properties of the 20 major amino acids defined in Table 1 below. In the case of amino acids with similar polarity, this may also be determined with reference to the hydrophilicity scale of the amino acid side chains in Table 2.

Table 1. Chemical properties of amino acids

Ala,A	Aliphatic, hydrophobic,	Met,M	Hydrophobic and neutral
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	and neutral		
Cys,C	Polar, hydrophobic, and neutral	Asn,N	Polar, hydrophilic, and neutral
Asp,D	Polar, hydrophilic, and charged (-)	Pro,P	Hydrophobic and neutral
Glu,E	Polar, hydrophilic, and charged (-)	Gln,Q	Polar, hydrophilic, and neutral
Phe,F	Aromatic, hydrophobic, and neutral	Arg,R	Polar, hydrophilic, and charged (+)
Gly,G	Aliphatic and neutral	Ser,S	Polar, hydrophilic, and neutral
His,H	Aromatic, polar, hydrophilic, and charged (+)	Thr,T	Polar, hydrophilic, and neutral
Ile,I	Aliphatic, hydrophobic, and neutral	Val,V	Aliphatic, hydrophobic, and neutral
Lys,K	Polar, hydrophilic, and charged (+)	Trp,W	Aromatic, hydrophobic, and neutral
Leu,L	Aliphatic, hydrophobic, and neutral	Tyr,Y	Aromatic, polar, and hydrophobic

Table 2. Hydrophilicity scale

Side chain	Hydrophilicity
Ile,I	4.5
Val,V	4.2
Leu,L	3.8
Phe,F	2.8
Cys,C	2.5
Met,M	1.9
Ala,A	1.8
Gly,G	-0.4
Thr,T	-0.7
Ser,S	-0.8
Trp,W	-0.9
Tyr,Y	-1.3
Pro,P	-1.6
His,H	-3.2
Glu,E	-3.5
Gln,Q	-3.5
Asp,D	-3.5
Asn,N	-3.5
Lys,K	-3.9
Arg,R	-4.5

It is well known that conservative substitutions of amino acids with similar properties between each other, such as those in Table 3, do not generally affect the activity of peptide sequences.

Table 3. Conservative amino acid substitutions

Type	Amino acid
Aliphatic	Glycine (G), alanine (A), valine (V), leucine (L), and isoleucine (I)
Hydrated or sulfur/selenium-containing	Serine (S), cysteine (C), selenocysteine (U), threonine (T), and methionine (M)
Cyclic	Proline (P)
Aromatic	Phenylalanine (F), tyrosine (Y), and tryptophan (W)
Basic	Histidine (H), lysine (K), and arginine (R)
Acidic and amide	Aspartic acid (D), glutamic acid (E), asparagine (N), and glutamine (Q)

The mutated or modified protein, monomer, or peptide may also be chemically modified in any manner at any site. The mutated or modified monomer or peptide is preferably chemically modified by attachment of the molecule to one or more cysteines (cysteine linkage), attachment of the molecule to one or more lysines, attachment of the molecule to one or more non-natural amino acids, and enzymatic modification of epitopes or terminal modification. Suitable methods for performing such modifications are well known in the art. A mutant of the modified protein, monomer, or peptide may be chemically modified by attachment of any molecule. For example, the mutant of the modified protein, monomer, or peptide may be chemically modified by attachment of a dye or fluorophore. In some embodiments, the mutated or modified monomer or peptide is chemically modified with a molecular adapter that facilitates interaction between a

pore comprising a monomer or peptide and a target nucleotide or target polynucleotide sequence. The molecular adapter is preferably a cyclic molecule, a cyclodextrin, a substance capable of hybridizing, a DNA binding agent or intercalator, a peptide or peptide analog, a synthetic polymer, an aromatic planar molecule, a positively charged small molecule, or a small molecule capable of hydrogen bonding.

The presence of the adapter improves the host-guest chemistry of the pore and the nucleotide or polynucleotide sequence, thereby improving the sequencing capability of the pore formed by the mutated monomer. The principles of host-guest chemistry are well known in the art. The adapter has an effect on the physical or chemical properties of the pore, which improves the interaction between the pore and the nucleotide or polynucleotide sequence. The adapter may alter the charge of a barrel or channel of the pore, or specifically interact with or bind to the nucleotide or polynucleotide sequence, thereby facilitating the interaction between the nucleotide or polynucleotide sequence and the pore.

A “protein pore” is a transmembrane protein structure that defines a channel or pore that allows molecules and ions to translocate from one side of the membrane to the other side. The translocation of ionic substances through the pore may be driven by a potential difference applied to either side of the pore. A “nanopore” is a protein pore in which the smallest diameter of the channel through which molecules or ions pass is on the order of nanometers (10^{-9} meters). In some embodiments, the protein pore may be a transmembrane protein pore. The transmembrane protein structure of the protein pore may be essentially monomeric or oligomeric. Generally, the pore comprises a plurality of polypeptide subunits arranged around a central axis, thereby forming a protein-lined channel extending substantially perpendicular to the membrane in which the nanopore

resides. The number of polypeptide subunits is not limited. Generally, the number of subunits is from 5 to 30, suitably from 6 to 10. Alternatively, the number of subunits is not defined as in the case of perfringolysin or related large membrane pores. The protein subunit portions within the nanopore that form the protein-lined channel generally comprise a secondary structural motif that may include one or more transmembrane β -barrel and/or α -helix portions.

In one embodiment, the protein pore comprises one or more porin monomers. Each porin monomer may be derived from *Pseudomonas taeanensis*. In one embodiment, the protein pore comprises one or more mutants of the porin monomer (i.e., one or more mutated monomers of porins).

In one embodiment, the porin is derived from a wild-type protein, wild-type homolog, or mutant thereof in the biological world. The mutant may be a modified porin or a porin mutant. Modifications in the mutants include, but are not limited to, any one or more of the modifications disclosed herein or a combination of the modifications. In one embodiment, the wild-type protein in the biological world is a protein derived from *Pseudomonas taeanensis*.

In one embodiment, the porin homolog refers to a polypeptide having at least 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, 90%, 85%, 80%, 75%, 70%, 65%, 60%, 55%, or 50% complete sequence identity to a protein set forth in SEQ ID NO: 1.

In one embodiment, the porin homolog refers to a polynucleotide having at least 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, 90%, 85%, 80%, 75%, 70%, 65%, 60%, 55%, or 50% complete sequence identity to a

polynucleotide encoding a protein set forth in SEQ ID NO: 2. The polynucleotide sequence may comprise a sequence that differs from SEQ ID NO: 2 based on the degeneracy of the genetic code.

5 Polynucleotide sequences may be derived and replicated using standard methods in the art. Chromosomal DNA encoding the wild-type porin may be extracted from pore-producing organisms such as *Pseudomonas taeanensis*. A gene encoding the pore subunit may be amplified using PCR comprising specific primers. The amplified sequence may then be
10 subjected to site-directed mutagenesis. Suitable methods for the site-directed mutagenesis are known in the art and include, for example, combine chain reaction. The constructed polynucleotides encoding the embodiments may be prepared using techniques well known in the art, such as those described in Sambrook, J. and Russell, D., (2001) Molecular
15 Cloning A Laboratory Manual, 3rd Edition., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.

The resulting polynucleotide sequences may then be integrated into recombinant replicable vectors, such as cloning vectors. The vectors may
20 be used to replicate the polynucleotides in compatible host cells. Thus, the polynucleotide sequences may be prepared by introducing the polynucleotides into replicable vectors, introducing the vectors into compatible host cells, and allowing the growth of the host cells under conditions that cause the replication of vectors. The vectors may be
25 recovered from the host cells.

Basic working principle of nanopore or protein pore

In one embodiment, in a cavity 100 filled with electrolyte, an insulating film 102 having a nanoscale pore divides the cavity into 2 chambers, as
30 shown in FIG. 1. When a voltage is applied to the electrolyte chamber,

ions or other small molecule substances pass through the pore under the force of an electric field, resulting in a stable detectable ionic current. By controlling the size and surface characteristics of the nanopore, the applied voltage, and the solution conditions, different types of biomolecules may be detected.

Because the four types of bases, adenine (A), guanine (G), cytosine (C), and thymine (T), which form DNA have different molecular structures and volume sizes, when single-stranded DNA (ssDNA) passes through the nanoscale pore under the drive of a rate-controlling enzyme and the electric field, the difference of chemical properties of different bases leads to different amplitude changes in the current when it passes through the nanopore or protein pore, thereby obtaining the sequence information about the detected nucleic acid, such as DNA.

FIG. 2 shows a schematic diagram 200 of DNA sequencing. As shown in FIG. 2, in a typical nanopore/protein pore sequencing experiment, the nanopore is the only channel through which ions on both sides of the phospholipid membrane pass. Rate-controlling proteins, such as polynucleotide binding proteins, act as motor proteins for the nucleic acid molecules, such as DNA, and pull DNA strands to sequentially pass through the nanopore/protein pore in steps of a single nucleotide. Whenever a nucleotide passes through the nanopore/protein pore, a corresponding pore-blocking signal is recorded (FIG. 3). By analyzing the current signals associated with these sequences using a corresponding algorithm, sequence information about the nucleic acid molecules, such as DNA, may be deduced.

In the embodiments, the porin is screened from different species in nature (mainly bacteria and archaea) by bioinformatics means and evolutionary

perspectives. In one embodiment, the porin is derived from any organism, preferably from *Pseudomonas taeanensis*. By sequence analysis, the porin has an intact functional domain. A porin 3D structure model is predicted and analyzed by using a structural biology means, and a channel protein with a proper reading head architectural form is selected. Then, candidate channel proteins (or porins) are modified, tested, and optimized by means of genetic engineering, protein engineering, protein directed evolution, computer-aided protein design, and the like, and after several iterations, a plurality of homologous protein mutants, preferably six homologous protein mutants (different homologous protein scaffolds) are obtained, which have different signal characteristics and signal distribution patterns.

The porin in the embodiments may be applied to the fourth generation sequencing technology. In one embodiment, the porin is a nanoporin. In one embodiment, the porin may be applied to solid-state pores for sequencing.

In one embodiment, a new protein scaffold is employed to form a new constriction zone (reading head region) structure, thereby providing a novel mode of action during sequencing. The porins of the embodiments have good jump distribution and recombination efficiency with phospholipid membranes.

In one embodiment, a wild-type porin monomer is modified by gene mutation to form a mutant of the porin monomer. In one embodiment, an amino acid of the mutant of the porin monomer comprises a sequence set forth in SEQ ID NO: 1 or a sequence having at least 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, 90%, 85%, 80%, 75%, 70%, 65%, 60%, 55%, or 50% identity thereto, and the amino acid of the mutant of the porin monomer has mutations at one or more positions corresponding to

positions 62-79, 104, 170, and 175 of SEQ ID NO: 1.

5 In one embodiment, the mutation comprises an insertion, a deletion, and/or a substitution of an amino acid. In one embodiment, the mutations at one or more of positions 62-79, 104, 170, and 175 of SEQ ID NO: 1 are insertions, deletions, and/or substitutions of amino acids at one or more of positions 62-79, 104, 170, and 175 of SEQ ID NO: 1.

10 In one embodiment, the amino acid of the mutant of the porin monomer has mutations at one or more positions corresponding to (1) positions 69-76, (2) positions 64-79, (3) positions 62, 63, 68, 69, 71, 75, 76, and 104, or (4) positions 68-76, 171, and 175 of SEQ ID NO: 1.

15 In one embodiment, the amino acid of the mutant of the porin monomer has insertions, deletions, and/or substitutions of amino acids at one or more positions corresponding to (1) positions 69-76, (2) positions 64-79, (3) positions 62, 63, 68, 69, 71, 75, 76, and 104, or (4) positions 68-76, 171, and 175 of SEQ ID NO: 1.

20 In one embodiment, the amino acid of the mutant of the porin monomer has mutations only at positions 69-76 (i.e., K69, P70, T71, P72, A73, S74, S75, and F76) corresponding to SEQ ID NO: 1, or has insertions, deletions, and/or substitutions of amino acids at one or more positions.

25 In one embodiment, the amino acid of the mutant of the porin monomer has mutations only at positions 64-79 (i.e., Q64, T65, G66, Q67, Y68, K69, P70, T71, P72, A73, S74, S75, F76, S77, T78, and S79) corresponding to SEQ ID NO: 1, or has insertions, deletions, and/or substitutions of amino acids at one or more positions.

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In one embodiment, the amino acid of the mutant of the porin monomer has mutations only at positions R62, D63, Y68, K69, T71, S75, F76Q, and E104 corresponding to SEQ ID NO: 1, or has insertions, deletions, and/or substitutions of amino acids at one or more positions.

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In one embodiment, the amino acid of the mutant of the porin monomer has mutations only at positions 68-76 (i.e., Y68, K69, P70, T71, P72, A73, S74, S75, and F76), E171, and D175 corresponding to SEQ ID NO: 1, or has insertions, deletions, and/or substitutions of amino acids at one or more positions.

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In one embodiment, the position corresponding to SEQ ID NO: 1 means that regardless of whether the sequence numbering is changed by insertions or deletions of amino acids or by adopting a sequence having identity, the relative position is unchanged and the sequence numbering of SEQ ID NO: 1 may be still used. For example, Q64 corresponding to SEQ ID NO: 1 may be mutated to Q64L, and even if the sequence numbering of SEQ ID NO: 1 is changed or a sequence having the identity as defined herein to SEQ ID NO: 1 is adopted, the amino acid Q at position 64 corresponding to SEQ ID NO: 1 (even if this amino acid is not at position 64 in another sequence) may also be mutated to L, and still be within the scope of the present invention.

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In one embodiment, the amino acid of the mutant of the porin monomer consists of a sequence set forth in SEQ ID NO: 1 or a sequence having at least 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, 90%, 85%, 80%, 75%, 70%, 65%, 60%, 55%, or 50% identity thereto, and the amino acid of the mutant of the porin monomer has mutations at one or more positions corresponding to positions 62-79, 104, 170, and 175 of SEQ ID NO: 1.

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In one embodiment, the sequence set forth in SEQ ID NO: 1 of the porin monomer is derived from *Pseudomonas taeanensis*. The nucleotide sequence encoding the amino acid set forth in SEQ ID NO: 1 is set forth in SEQ ID NO: 2.

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In one embodiment, amino acids KPTPASSF at positions 69-76 are mutated to M₁M₂M₃M₄M₅M₆M₇M₈, wherein M₁ is selected from R, K, or H; M₂ is selected from P; M₃ is selected from S, G, C, U, T, M, A, V, L, or I; M₄ is selected from P; M₅ is selected from A, G, V, L, or I; M₆ is selected from S, C, U, T, or M; M₇ is selected from A, T, G, V, L, I, S, C, U, or M; M₈ is selected from Q, D, E, N, K, H, or R.

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In one embodiment, amino acids QTGQYKPTPASSFSTS corresponding to positions 64-79 of SEQ ID NO: 1 are mutated to M₉M₁₀M₁₁M₁₂M₁₃M₁₄M₁₅M₁₆ M₁₇M₁₈M₁₉M₂₀M₂₁M₂₂M₂₃M₂₄, wherein M₉ is selected from L, G, A, V, or I; M₁₀ is selected from T, S, C, U, or M; M₁₁ is selected from G, A, V, L, or I; M₁₂ is selected from Q, D, E, or N; M₁₃ is selected from Y, F, or W; M₁₄ is selected from R, H, or K; M₁₅ is selected from P; M₁₆ is selected from S, C, U, T, or M; M₁₇ is selected from P; M₁₈ is selected from A, G, V, L, or I; M₁₉ is selected from S, C, U, T, or M; M₂₀ is selected from A, G, V, L, or I; M₂₁ is selected from N, D, E, Q, L, G, A, V, or I; M₂₂ is selected from S, C, U, T, or M; M₂₃ is selected from T, S, C, U, or M; M₂₄ is selected from A, G, V, L, or I.

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In one embodiment, R62 corresponding to SEQ ID NO: 1 is mutated to R62S, R62C, R62U, R62T, or R62M; D63 is mutated to D63V, D63G, D63A, D63L, or D63I; Y68 is mutated to Y68F or Y68W; K69 is mutated to K69R or K69H; T71 is mutated to T71S, T71C, T71U, or T71M; S75 is mutated to S75A, S75G, S75V, S75L, or S75I; F76 is mutated to F76Q, F76D, F76E, or F76N; E104 is mutated to E104V, E104G, E104A, E104L,

25

30

or E104I.

In one embodiment, amino acids YKTPASSF corresponding to positions 68-76 of SEQ ID NO: 1 are mutated to M₂₅M₂₆M₂₇M₂₈M₂₉M₃₀M₃₁M₃₂M₃₃,
5 E171 is mutated to E171N, E171D, or E171Q, and D175 is mutated to D175N, D175E, or D175Q, wherein M₂₅ is selected from F, Y, or W; M₂₆ is selected from R, H, or K; M₂₇ is selected from P; M₂₈ is selected from S, C, U, T, or M; M₂₉ is selected from P; M₃₀ is selected from A, G, V, L, or I; M₃₁ is selected from S, C, U, T, or M; M₃₂ is selected from A, G, V, L,
10 or I; M₃₃ is selected from Q, D, E, or N.

In one embodiment, in the mutant of the porin monomer, the amino acid mutation is selected from the group consisting of:

- 15 (a) mutations from the amino acids KTPASSF corresponding to positions 69-76 of SEQ ID NO: 1 to RPSPASAQ;
- (b) mutations from the amino acids KTPASSF corresponding to positions 69-76 of SEQ ID NO: 1 to KPGPASTK;
- (c) mutations from the amino acids QTGQYKTPASSFSTS corresponding to positions 64-79 of SEQ ID NO: 1 to LTGQYRPSPASANSTA;
- 20 (d) mutations from the amino acids QTGQYKTPASSFSTS corresponding to positions 64-79 of SEQ ID NO: 1 to LTGQYRPSPASALSTA;
- (e) R62S, D63V, Y68F, K69R, T71S, S75A, F76Q, and E104V corresponding to SEQ ID NO: 1; and
- (f) mutations from the amino acids YKTPASSF corresponding to
25 positions 68-76 of SEQ ID NO: 1 to FRPSPASAQ, and E171N and D175N corresponding to SEQ ID NO: 1.

In one embodiment, the mutant of the porin monomer comprises or consists of an amino acid sequence set forth in SEQ ID NO: 24, SEQ ID
30 NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, or SEQ ID

NO: 30.

5 In one embodiment, the protein pore comprises at least one mutant of the porin monomer (or porin-mutated monomer). In one embodiment, the protein pore comprises at least two, three, four, five, six, seven, eight, nine, ten, or more mutants of the porin monomer. In one embodiment, the protein pore comprises at least two mutants of the porin monomer, and the mutants of the porin monomer may be identical or different. In one embodiment, the protein pore comprises two or more mutants of the porin monomer; preferably, the two or more mutants of the monomer are identical. In one embodiment, the protein pore has a pore channel diameter of a constriction zone of 0.7 nm to 2.2 nm, 0.9 nm to 1.6 nm, 1.4 nm to 1.6 nm, or 15.9 Å to 20.1 Å.

15 Provided is use of the mutant of the porin monomer or the protein pore comprising same in the detection of the presence, absence, or one or more characteristics of a target analyte. In one embodiment, the mutant of the porin monomer or the protein pore is used to detect the sequence of nucleic acid molecules, or to characterize the sequence of polynucleotides, such as sequencing polynucleotides, because they may distinguish different nucleotides with high sensitivity. The mutant of the porin monomer or the protein pore comprising same may distinguish four types of nucleotides in DNA and RNA, and even may distinguish between methylated and unmethylated nucleotides, with unexpectedly high resolution. The mutant of the porin monomer or the protein pore shows almost complete separation from all four types of DNA/RNA nucleotides. Deoxycytidine monophosphate (dCMP) and methyl-dCMP are further distinguished based on the dwell time in the protein pore and the current flowing through the protein pore.

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5 The mutant of the porin monomer or the protein pore may also distinguish
between different nucleotides under a range of conditions. In particular,
the mutant of the porin monomer or the protein pore distinguishes
nucleotides under conditions that are favorable for nucleic acid
characterization such as sequencing. By altering the applied potential, salt
concentrations, buffers, temperature, and the presence of additives such as
urea, betaine and DTT, the extent to which the mutant of the porin
monomer or the protein pore distinguishes between different nucleotides
may be controlled. This allows the functions of the mutant of the porin
monomer or the protein pore to be finely regulated and controlled,
especially during sequencing. The mutant of the porin monomer or the
protein pore may also be used to identify polynucleotide polymers by the
interaction with one or more monomers rather than on a
nucleotide-by-nucleotide basis.

15 The mutant of the porin monomer or the protein pore may be isolated,
substantially isolated, purified, or substantially purified. The mutant of the
porin monomer or the protein pore of the embodiments is isolated or
purified if it is completely free of any other components, such as
liposomes or other protein pores/porins. The mutant of the porin monomer
or the protein pore is substantially isolated if it is mixed with a carrier or
diluent that does not interfere with its intended use. For example, the
mutant of the porin monomer or the protein pore is substantially isolated
or substantially purified if it is present in a form comprising less than 10%,
less than 5%, less than 2%, or less than 1% of other components, such as
triblock copolymers, liposomes, or other protein pores/porins.
Alternatively, the mutant of the porin monomer or the protein pore may be
present in a membrane.

30 For example, the membrane is preferably an amphiphilic layer. The

amphiphilic layer is a layer formed of amphiphilic molecules, for example, phospholipids, which have hydrophilicity and lipophilicity. The amphiphilic molecules may be synthetic or naturally occurring. The amphiphilic layer may be a monolayer or a bilayer. The amphiphilic layer is generally planar. The amphiphilic layer may be curved. The amphiphilic layer may be supported. The membrane may be a lipid bilayer. The lipid bilayer is formed by two opposing layers of lipids. The two layers of the lipids are arranged such that their hydrophobic tail groups face each other to form a hydrophobic interior. The hydrophilic head groups of the lipids face outward towards the aqueous environment on each side of the bilayer. The membrane comprises a solid layer. The solid layer may be formed from organic and inorganic materials. If the membrane comprises a solid layer, the pore is generally present in the amphiphilic membrane or in a layer comprised within the solid layer, for example, in holes, wells, gaps, channels, grooves, or slits within the solid layer.

Characterization of analytes

Embodiments provide a method for determining the presence, absence, or one or more characteristics of a target analyte. The method involves contacting the target analyte with a mutant of a porin monomer or a protein pore, such that the target analyte moves relative to, e.g., through, the mutant of the porin monomer or the protein pore, and acquiring one or more measurements when the target analyte moves relative to the mutant of the porin monomer or the protein pore, thereby determining the presence, absence, or one or more characteristics of the target analyte. The target analyte may also be referred to as a template analyte or analyte of interest.

The target analyte is preferably a polysaccharide, a metal ion, an inorganic salt, a polymer, an amino acid, a peptide, a polypeptide, a protein, a

nucleotide, an oligonucleotide, a polynucleotide, a dye, a drug, a diagnostic agent, an explosive, or an environmental contaminant. The method may involve determining the presence, absence, or one or more characteristics of two or more target analytes of the same class, e.g., two or more proteins, two or more nucleotides, or two or more drugs. Alternatively, the method may involve determining the presence, absence, or one or more characteristics of two or more target analytes of different classes, e.g., one or more proteins, one or more nucleotides, and one or more drugs.

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The method comprises contacting the target analyte with a mutant of a porin monomer or a protein pore, such that the target analyte moves through the mutant of the porin monomer or the protein pore. The protein pore generally comprises at least 1, at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, or at least 10 porin-mutated monomers, for example, 7, 8, 9, or 10 monomers. The protein pore comprises identical monomers or different porin monomers, preferably 8 or 9 identical monomers. One or more, such as 2, 3, 4, 5, 6, 7, 8, 9, or 10, of the monomers are preferably chemically modified as discussed above. In one embodiment, the amino acid of each monomer comprises SEQ ID NO: 1 and mutants thereof as described above. In one embodiment, the amino acid of each monomer consists of SEQ ID NO: 1 and mutants thereof as described above.

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The method of the embodiments may measure two, three, four, five, or more characteristics of a polynucleotide. The one or more characteristics are preferably selected from (i) a length of the polynucleotide, (ii) an identity of the polynucleotide, (iii) a sequence of the polynucleotide, (iv) a secondary structure of the polynucleotide, and (v) whether the polynucleotide is modified. In one embodiment, any combination of (i) to

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(v) may be measured.

For (i), the length of the polynucleotide may be measured, for example, by determining the number of interactions between the polynucleotide and the mutant of the protein monomer/protein pore or the duration time of the interaction between the polynucleotide and the mutant of the protein monomer/protein pore.

For (ii), the identity of the polynucleotide may be measured in a variety of ways, and the identity of the polynucleotide may be measured in combination with or without measurement of the polynucleotide sequence. The former is simpler, and the polynucleotide is sequenced and thus identified. The latter may be done in several different ways. For example, the presence of a particular motif in the polynucleotide may be measured (without measuring the remaining sequence of the polynucleotide). Alternatively, the measurement of a particular electrical and/or optical signal in the method may identify that the polynucleotide is derived from a particular source.

For (iii), the sequence of the polynucleotide may be determined as previously described. Suitable sequencing methods, particularly those using electrical measurement methods, are described in Stoddart D et al., Proc Natl Acad Sci, 12; 106 (19) 7702-7, Lieberman KR et al., J Am Chem Soc., 2010; 132 (50) 17961-72, and International Application W02000/28312.

For (iv), the secondary structure may be measured using a variety of methods. For example, if the method involves an electrical measurement method, a change in dwell time or a change in current flowing through the pore may be used to measure the secondary structure. This allows regions

of single-stranded and double-stranded polynucleotides to be distinguished.

For (v), the presence or absence of any modification may be measured.

5 The method preferably comprises determining whether the polynucleotide is modified by methylation, by oxidation, by damage, with one or more proteins, with one or more labels or tags, or by the absence of bases or nucleobases and sugars. Particular modifications will result in specific interactions with the pore, which may be measured using the methods
10 described below. For example, methylcytosine may be distinguished from cytosine based on the current flowing through the pore during its interaction with each nucleotide.

The target polynucleotide is contacted with a mutant of a protein
15 monomer/protein pore, for example, a mutant of a protein monomer/protein pore as in the embodiments. The mutant of the protein monomer/protein pore is generally present in a membrane. Suitable membranes are as previously described. The method may be performed using any device suitable for studying a system of the membrane/protein
20 pore or mutant of the porin monomer, in which the mutant of the protein monomer/protein pore is present in the membrane. The method may be performed using any device suitable for use in transmembrane pore sensing. For example, the device comprises a chamber containing an aqueous solution and a barrier dividing the chamber into two parts. The
25 barrier generally has a hole in which a membrane containing a pore is formed. Alternatively, the barrier forms a membrane in which a mutant of a protein monomer/protein pore is present. The method may be performed using the device described in International Application No. PCT/GB08/000562 (WO 2008/102120).

30

Various different types of measurements may be performed. This includes, but is not limited to, electrical measurements and optical measurements. The electrical measurements include voltage measurements, capacitance measurements, current measurements, impedance measurements, tunneling measurements (Ivanov AP et al., Nano Lett., 2011, Jan 12; 11 (I): 279-85) and FET measurements (International Application TO 2005/124888). The optical measurements may be combined with the electrical measurements (Soni GV et al., Rev Sci Instrum., 2010, Jan; 81 (1) 014301). The measurement may be a transmembrane current measurement, for example, a measurement of an ionic current flowing through the pore. In one embodiment, the electrical measurements or optical measurements may employ conventional electrical measurements or optical measurements.

The electrical measurements may be performed using standard single-channel recording apparatus as described in Stoddart D et al., Proc Natl Acad Sci, 12; 106 (19) 7702-7, Lieberman KR et al., J Am Chem Soc., 2010; 132 (50) 17961-72, and International Application WO 2000/28312. Alternatively, the electrical measurements may be performed using multichannel systems, for example, as described in International Application W02009/077734 and International Application WO 2011/067559.

The method is preferably performed using a potential applied across the membrane. The applied potential may be a voltage potential. Alternatively, the applied potential may be a chemical potential. An example of the method is using a salt gradient across a membrane, such as an amphiphilic molecular layer. The salt gradient is disclosed in Holden et al., J Am Chem Soc., 2007, Jul 11; 129 (27): 8650-5. In some cases, the current flowing through a mutant of a protein monomer/protein pore when a polynucleotide moves relative to the mutant of the protein

monomer/protein pore is used to estimate or determine the sequence of the polynucleotide. This is strand sequencing.

5 The method may comprise measuring the current flowing through the pore when the polynucleotide moves relative to the pore. Therefore, the apparatus used in the method may also comprise circuitry capable of applying a potential and measuring an electrical signal through the membrane and the pore. The method may be performed using a patch clamp or a voltage clamp,

10 and may comprise measuring the current flowing through the pore when the polynucleotide moves relative to the pore. Suitable conditions for measuring ion currents through transmembrane protein pores are known in the art and are disclosed in the embodiments. The method is generally performed with a voltage applied across the membrane and the pore. The
15 voltage used is generally from +5 V to -5 V, for example, from +4 V to -4 V, from +3 V to -3 V, or from +2 V to -2 V. The voltage used is generally from -600 mV to +600 V or -400 mV to +400 mV. The voltage used is preferably in a range having a lower limit selected from -400 mV, -300
20 mV, -200 mV, -150 mV, -100 mV, -50 mV, -20 mV, and 0 mV, and an upper limit independently selected from +10 mV, +20 mV, +50 mV, +100 mV, +150 mV, +200 mV, +300 mV, and +400 mV. The voltage used is more preferably in the range of 100 mV to 240 mV and most preferably in the range of 120 mV to 220 mV. By using an increased applied potential,
25 the identification of different nucleotides by a pore may be increased.

The method is generally performed in the presence of any charge carrier, for example, a metal salt such as an alkali metal salt, a halide salt such as a chloride salt, for example, an alkali metal chloride salt. The charge carriers
30 may include an ionic liquid or an organic salt, such as

tetramethylammonium chloride, trimethylphenylammonium chloride, phenyltrimethylammonium chloride, or 1-ethyl-3-methylimidazolium chloride. In the exemplary device described above, the salt is present in the aqueous solution in the chamber. Potassium chloride (KCl), sodium chloride (NaCl), cesium chloride (CsCl), or a mixture of potassium ferrocyanide and potassium ferricyanide is generally used. KCl, NaCl, and the mixture of potassium ferrocyanide and potassium ferricyanide are preferred. The charge carriers may be asymmetric on the membrane. For example, the type and/or concentration of the charge carriers may be different on each side of the membrane.

The concentration of the salt may be saturated. The concentration of the salt may be 3 M or less, and is generally 0.1 to 2.5 M, 0.3 to 1.9 M, 0.5 to 1.8 M, 0.7 to 1.7 M, 0.9 to 1.6 M, or 1 to 1.4 M. The concentration of the salt is preferably 150 mM to 1 M. The method is preferably performed using a salt concentration of at least 0.3 M, for example, at least 0.4 M, at least 0.5 M, at least 0.6 M, at least 0.8 M, at least 1.0 M, at least 1.5 M, at least 2.0 M, at least 2.5 M, or at least 3.0 M. High salt concentrations provide a high signal-to-noise ratio and allow the presence of a nucleotide to be identified in the background of normal current fluctuations to be indicated by the current.

The method is generally performed in the presence of a buffer. In the exemplary device described above, the buffer is present in the aqueous solution in the chamber. Any buffer may be used in the method of the present invention. Generally, the buffer is a phosphate buffer. Other suitable buffers are HEPES or Tris-HCl buffers. The method is generally performed at a pH of 4.0 to 12.0, 4.5 to 10.0, 5.0 to 9.0, 5.5 to 8.8, 6.0 to 8.7, 7.0 to 8.8, or 7.5 to 8.5. The pH value used is preferably about 7.5.

The method may be performed at a temperature of 0 °C to 100 °C, 15 °C to 95 °C, 16 °C to 90 °C, 17 °C to 85 °C, 18 °C to 80 °C, 19 °C to 70 °C, or 20 °C to 60 °C. The method is generally performed at room temperature. The method is optionally performed at a temperature that supports enzyme functions, for example, about 37 °C.

In one embodiment, the method for determining the presence, absence, or one or more characteristics of a target analyte (e.g., a polynucleotide) comprises coupling the target analyte to a membrane; and the target analyte interacting (e.g., contacting) with the protein pore present in the membrane, such that the target analyte moves relative to the protein pore (e.g., passes through the protein pore). In one embodiment, the current through the protein pore is measured when the target analyte moves relative to the protein pore, thereby determining the presence, absence, or one or more characteristics of the target analyte (e.g., the sequence of the polynucleotide).

Polynucleotide binding protein

The characterization method of the embodiments preferably comprises contacting a polynucleotide with a polynucleotide binding protein, such that the protein controls the movement of the polynucleotide relative to, e.g., through, a mutant of a protein monomer/protein pore.

More preferably, the method comprises (a) contacting the polynucleotide with the mutant of the protein monomer/protein pore and the polynucleotide binding protein, such that the protein controls the movement of the polynucleotide relative to, e.g., through, the mutant of the protein monomer/protein pore, and (b) acquiring one or more measurements when the polynucleotide moves relative to the mutant of the protein monomer/protein pore, wherein the measurements are indicative of one or more characteristics of the polynucleotide, thereby characterizing

the polynucleotide.

5 More preferably, the method comprises (a) contacting the polynucleotide with the mutant of the protein monomer/protein pore and the polynucleotide binding protein, such that the protein controls the movement of the polynucleotide relative to, e.g., through, the mutant of the protein monomer/protein pore, and (b) measuring a current through the mutant of the protein monomer/protein pore when the polynucleotide moves relative to the mutant of the protein monomer/protein pore, wherein
10 the current is indicative of one or more characteristics of the polynucleotide, thereby characterizing the polynucleotide.

15 The polynucleotide binding protein may be any protein capable of binding a polynucleotide and controlling the movement thereof through a pore. The polynucleotide binding protein generally interacts with a polynucleotide and modifies at least one property of the polynucleotide. The protein may modify a polynucleotide by cleaving it to form individual nucleotides or short strands of nucleotides such as dinucleotides or trinucleotides. The protein may modify a polynucleotide by orienting it or moving it to a
20 specific position, i.e., controlling its movement.

The polynucleotide binding protein is preferably derived from a polynucleotide handling enzyme. The polynucleotide handling enzyme is a polypeptide that is capable of interacting with a polynucleotide and
25 modifying at least one property of the polynucleotide. The enzyme may modify a polynucleotide by cleaving it to form individual nucleotides or short strands of nucleotides such as dinucleotides or trinucleotides. The enzyme may modify a polynucleotide by orienting it or moving it to a specific position. The polynucleotide handling enzyme does not need to
30 exhibit enzymatic activity as long as it is capable of binding to a

polynucleotide and controlling its movement through a pore. For example, the enzyme may be modified to remove its enzymatic activity, or may be used under conditions that prevent it from acting as an enzyme.

5 The polynucleotide handling enzyme is preferably a polymerase, an exonuclease, a helicase, and a topoisomerase such as a gyrase. In one embodiment, the enzyme is preferably a helicase, such as Hel308Mbu, Hel308Csy, Hel308Tga, Hel308Mhu, Tral Eco, XPD Mbu, Dda, or variants thereof. Any helicase may be used in the embodiments.

10

In one embodiment, any number of helicases may be used. For example, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more helicases may be used. In some embodiments, different numbers of helicases may be used.

15 The method of the embodiments preferably comprises contacting a polynucleotide with two or more helicases. The two or more helicases are generally the same helicase. The two or more helicases may be different helicases.

20 The two or more helicases may be any combination of the helicases described above. The two or more helicases may be two or more Dda helicases. The two or more helicases may be one or more Dda helicases and one or more TrwC helicases. The two or more helicases may be different variants of the same helicase.

25

The two or more helicases are preferably linked to each other. The two or more helicases are more preferably covalently linked to each other. The helicases may be linked in any order and using any method.

30 Kit

The present invention further provides a kit for characterizing a target analyte (e.g., a target polynucleotide). The kit comprises a pore and components of a membrane in the embodiments. The membrane is preferably formed from the components. The pore is preferably present in the membrane. The kit may comprise the components of any of the membranes disclosed above (e.g., an amphiphilic layer or a triblock copolymer membrane). The kit may further comprise a polynucleotide binding protein. Any of the polynucleotide binding proteins discussed above may be used.

10

In one embodiment, the membrane is an amphiphilic layer, a solid layer, or a lipid bilayer.

15

The kit may further comprise one or more anchors for coupling the polynucleotide to the membrane.

20

The kit is preferably used for characterizing a double-stranded polynucleotide and preferably comprises a Y adapter and a hairpin loop adapter.

25

The Y adapter preferably has one or more helicases linked, and the hairpin loop adapter preferably has one or more molecular brakes linked. The Y adapter preferably comprises one or more first anchors for coupling the polynucleotide to the membrane, the hairpin loop adapter preferably comprises one or more second anchors for coupling the polynucleotide to the membrane, and the coupling strength of the hairpin loop adapter to the membrane is preferably greater than the coupling strength of the Y adapter to the membrane.

30

The kit may additionally comprise one or more other reagents or

instruments that enable any of the embodiments mentioned above to be performed. Such reagents or instruments include one or more of the following: suitable buffers (aqueous solutions), device for obtaining a sample from an individual (such as a vessel or instrument containing a needle), device for amplifying and/or expressing a polynucleotide, or voltage or patch clamp apparatus. The reagents may be present in the kit in a dry form, such that a fluid sample resuspends the reagents. Optionally, the kit may further comprise instructions to enable the kit to be used in the method of the present invention or details as to what organism may use the method.

Apparatus (or device)

The present invention further provides an apparatus for characterizing a target analyte (e.g., a target polynucleotide). The apparatus comprises single or multiple mutants of a protein monomer/protein pores, and single or multiple membranes. The mutant of the protein monomer/protein pore is preferably present in the membrane. The number of pores and membranes is preferably equal. Preferably, a single pore is present in each membrane.

Preferably, the apparatus further comprises instructions for implementing the method of the embodiments. The apparatus may be any conventional apparatus for analyte analysis, for example, an array or chip. Any of the embodiments discussed in combination with the method of the embodiments is equally applicable to the apparatus. The apparatus may further comprise any of the characteristics present in the kit described herein. The apparatus used in the embodiments may specifically be a gene sequencer, QNome-9604, from QitanTech.

The above-mentioned prior art is incorporated herein by reference in its entirety.

The following examples are intended to illustrate the present invention without limiting it.

5 Example 1

In the example, a wild-type porin was derived from *Pseudomonas taeanensis*, and the amino acid sequence of the wild-type porin was set forth in SEQ ID NO: 1, and the nucleotide sequence encoding this amino acid sequence was set forth in SEQ ID NO: 2. Mutant 1 of a porin monomer was a wild-type porin having mutations at positions 69-76 corresponding to SEQ ID NO: 1; specifically, KPTPASSF at positions 69-76 were replaced by RPSPASAQ. A protein pore comprising the mutant 1 of the porin monomer was mutant pore 1. The amino acid sequence of the mutant 1 of the protein monomer was set forth in SEQ ID NO: 24, and the nucleic acid sequence was set forth in SEQ ID NO: 25.

Example 2

In the example, a wild-type porin was derived from *Pseudomonas taeanensis*, and the amino acid sequence of the wild-type porin was set forth in SEQ ID NO: 1, and the nucleotide sequence encoding this amino acid sequence was set forth in SEQ ID NO: 2. Mutant 2 of a porin monomer was a wild-type porin having mutations at positions 69-76 corresponding to SEQ ID NO: 1; specifically, KPTPASSF at positions 69-76 were replaced by KPGPASTK. A protein pore comprising the mutant 2 of the porin monomer was mutant pore 2. The amino acid sequence of the mutant 2 of the protein monomer was set forth in SEQ ID NO: 26.

Example 3

30 In the example, a wild-type porin was derived from *Pseudomonas*

taeanensis, and the amino acid sequence of the wild-type porin was set forth in SEQ ID NO: 1, and the nucleotide sequence encoding this amino acid sequence was set forth in SEQ ID NO: 2. Mutant 3 of a porin monomer was a wild-type porin having mutations at positions 64-79 corresponding to SEQ ID NO: 1; specifically, QTGQYKPTPASSFSTS at positions 64-79 were replaced by LTGQYRSPASANSTA. A protein pore comprising the mutant 3 of the porin monomer was mutant pore 3. The amino acid sequence of the mutant 3 of the protein monomer was set forth in SEQ ID NO: 27.

10

Example 4

In the example, a wild-type porin was derived from *Pseudomonas taeanensis*, and the amino acid sequence of the wild-type porin was set forth in SEQ ID NO: 1, and the nucleotide sequence encoding this amino acid sequence was set forth in SEQ ID NO: 2. Mutant 4 of a porin monomer was a wild-type porin having mutations at positions 64-79 corresponding to SEQ ID NO: 1; specifically, QTGQYKPTPASSFSTS at positions 64-79 were replaced by LTGQYRSPASALSTA. A protein pore comprising the mutant 4 of the porin monomer was mutant pore 4. The amino acid sequence of the mutant 4 of the protein monomer was set forth in SEQ ID NO: 28.

20

Example 5

In the example, a wild-type porin was derived from *Pseudomonas taeanensis*, and the amino acid sequence of the wild-type porin was set forth in SEQ ID NO: 1, and the nucleotide sequence encoding this amino acid sequence was set forth in SEQ ID NO: 2. Mutant 5 of a porin monomer was a wild-type porin having mutations at the following positions corresponding to SEQ ID NO: 1: R62S, D63V, Y68F, K69R, T71S, S75A, F76Q, and E104V. A protein pore comprising the mutant 5 of

25

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the porin monomer was mutant pore 5. The amino acid sequence of the mutant 5 of the protein monomer was set forth in SEQ ID NO: 29.

Example 6

5 In the example, a wild-type porin was derived from *Pseudomonas taeanensis*, and the amino acid sequence of the wild-type porin was set forth in SEQ ID NO: 1, and the nucleotide sequence encoding this amino acid sequence was set forth in SEQ ID NO: 2. Mutant 6 of a porin monomer was a wild-type porin having mutations at positions 68-76, 171,
10 and 175 corresponding to SEQ ID NO: 1; specifically, YKPTPASSF at positions 68-76 were replaced by FRPSPASAQ, E at position 171 was replaced by N, and D at position 175 was replaced by N. A protein pore comprising the mutant 6 of the porin monomer was mutant pore 6. The amino acid sequence of the mutant 6 of the protein monomer was set forth
15 in SEQ ID NO: 30.

Example 7

The wild-type porin was subjected to homologous modeling by adopting SWISS MODEL, and the amino acid of the wild-type porin monomer was
20 set forth in SEQ ID NO: 1. FIG. 4A is a side view 400 of a predicted protein structure model, in which the darker portion shows a protein monomer 402. FIG. 4B is a top view 404 of the surface structure model, in which the darker portion shows a protein monomer 406. FIG. 4C is a ribbon structure model diagram 408, in which the darker portion shows a
25 protein monomer 410.

The mutant pore 1 was subjected to homologous modeling by adopting SWISS MODEL. FIGs. 5A and 5B show amino acid model diagrams of the mutant pore 1, in which the plus sign “+” indicates a water molecule.

30

FIG. 6 shows the dimension of each portion of the mutant 1 of the porin monomers, in which the maximum pore channel diameter of the constriction zone between the mutant of two porin monomers (i.e., two mutated porin monomers) 602 and 604 is 20.1 Å, followed by 17.2 Å, and the smallest diameter is 15.9 Å. The head-to-head distance and tail-to-tail distance of the two mutated porin monomers are 52.4 Å and 36.9 Å, respectively. The full length of the mutated porin monomer is 94.6 Å, and the height from the head to the pore channel of the constriction zone of the mutated porin monomer is 41.7 Å. The heights of the pore channel portions of the constriction zones are 12.2 Å and 4.3 Å as shown in FIG. 6.

FIG. 7 shows a monomer amino acid model of the porin mutant 1, and the enlarged diagram shows the amino acid composition of the constriction zone structure, i.e., Gln76, Ser74, and Ser71.

Example 8

Negative staining electron microscopy results for the porin mutant 1 are shown in FIG. 8. As can be seen from the negative staining EM results, particles of the mutant 1 were uniform with little aggregation, and many apparently correct protein particles could be seen.

A cryogenic electron micrograph and 2D classification results of the porin mutant 1 are shown in FIGs. 9A and 9B, in which the 2D results only show the better classification.

FIGs. 10A and 10B show locally refined Fourier shell correlation (FSC) results. The resolution results for different regions of the porin mutant 1 could be seen, and its final cryogenic electron microscopy single particle reconstruction resolution is 2.2 Å. The reconstruction resolution was determined based on the gold-standard FSC 0.143 criterion and the

high-resolution noise replacement. FIG. 11 shows an electron density diagram of the porin mutant 1 after three-dimensional reconstruction at a resolution of 2.2 Å by cryogenic electron microscopy. FIG. 12 shows an electron density map of the porin mutant 1 at a resolution of 2.2 Å. The map shows the eye-loop region of the channel and is overlaid on the final refined model.

Cryogenic electron microscopy data of the porin mutant 1 are shown in Table 4.

Table 4. Cryogenic electron microscopy data of porin mutant 1

Data collection	
Electron microscope apparatus	Titan Krios G3i
Voltage (kV)	300
Detector	Gatan K3
Enlarged	64k
Pixel size (Å)	1.08
Electron dose (e-/Å ²)	51.8
Defocus range (µm)	1.6-2.0
Micrographs	5361
Reconstruction	
Software	RELION3.1
Number of refined particles	835258
Symmetric	C9
Resolution range	2.1–2.8
Reconstructed resolution shielding 0.143 (after post-processing, Å)	2.2
Map sharpening factor B (Å ²)	-85

Example 9. Preparation of DNA Constructs

Two DNA constructs, BS7-4C3-SE1 and BS7-4C3-PLT, were prepared. The structure of BS7-4C3-SE1 is shown in FIG. 13, and the sequence information is shown below:

a: 30*C3

5 b: 5'-TTTTT TTTTT-3' (i.e., SEQ ID NO: 3)

c: rate-controlling protein

d: 4*C18

e: 5'-AATGT ACTTC GTTCA GTTAC GTATT GCT-3' (i.e., SEQ ID NO: 4)

10 f: 5'P-GC AATAC GTAAC TGAAC GAAGT
TCACTATCGCATTCTCATGA-3' (i.e., SEQ ID NO: 5)

g: cholesterol tag

h: 5'-TCATG AGAAT GCGAT AGTGA -3' (i.e., SEQ ID NO: 6)

15 i: 5'-AAAAA AAAAA AAAAA AAAAA AAAAA AAAAA AAAAA
AAAAA AAGCA ATACG TAACT GAACG AAGTA CATTA AAAAA
AAAAA AAAAA AAAA-3' (i.e., SEQ ID NO: 7)

j: 5'-ATCCT TTTTT TTTTT TTTTT TTTT-3' (i.e., SEQ ID NO: 8)

k: 5'-AATGT ACTTC GTTCA GTTAC GTATT GCTTT TTTTT TTTTT
TTTTT TTT-3' (i.e., SEQ ID NO: 9)

20 l: dSpacer

m: 5'-TTTTT TTTTT TTTTT TTTTT-3' (i.e., SEQ ID NO: 10)

The structure of BS7-4C3-PLT is shown in FIG. 14, and the sequence information is shown below:

25 a: 30*C3

b: 5'-TTTTT TTTTT-3' (i.e., SEQ ID NO: 11)

c: rate-controlling protein

d: 4*C18

30 e: 5'-AATGT ACTTC GTTCA GTTAC GTATT GCT-3' (i.e., SEQ ID NO: 12)

f: 5'-P-GC AATAC GTAAC TGAAC GAAGT
TCACTATCGCATTCTCATGA-3' (i.e., SEQ ID NO: 13)

g: cholesterol tag

h: 5'- TCATG AGAAT GCGAT AGTGA -3' (i.e., SEQ ID NO: 14)

5 i: 5'-AAAAAAAAAAAAAAAAAAAAAAAAAAAAA (i.e., SEQ ID NO:
15)/dSpacer/ AAAAAAAAAAAAAA (i.e., SEQ ID NO: 16)/dSpacer/
AAAAAAAAAAAAAAAAATCTCTGAATCTCTGAATCTCTGAATCTCTAA
AAAAAAAAAGAAAAAAAAAAAAAAAACAAAAAAAAAAAAAATAAAAA
AAAAAAAAAGCAATACGTAACGAAGTACATTAAAAAAAAAA
10 A (i.e., SEQ ID NO: 17)-3'

j: 5'-ATCCTTTTTTTTTTTAATGTACTTCGTTTCAGTTACGTATTGCT
-3' (i.e., SEQ ID NO: 18)

k:

5'-P-TTTTTTTTTTTTATTTTTTTTTTTTGTTTTTTTTTTTCTTTTTTT
15 TTTTAGAGATTCAGAGATTCAGAGATTCAGAGATTTTTTTTTTTTT
TT (i.e., SEQ ID NO: 19)/dSpacer/ TTTTTTTTTTTT (i.e., SEQ ID NO:
20)/iSpC3/ TTTTTTTTTTTTTTTTTTTTTTTTTTTT (i.e., SEQ ID NO:
21)-3'

C3, C18, dSpacer, and iSpC3 were sequences of markers introduced to
20 indicate the resolution characteristics of pore sequencing.

In this example, the rate-controlling protein c in FIGs. 13 and 14 is
helicase Mph-MP1-E105C/A362C (having mutations E105C/A362C), the
amino acid sequence is set forth in SEQ ID NO: 22, and the nucleic acid
25 sequence is set forth in SEQ ID NO: 23.

Example 10

The mutant pore 1 was used as a protein pore and detected by adopting a
single-pore sequencing technique. After the insertion of a single porin with
30 the amino acid sequence of the mutant 1 into a phospholipid bilayer, a

buffer (625 mM KCl, 10 mM HEPES at pH of 8.0, and 50 mM MgCl₂) flowed through the system to remove any excess nanopores of the mutant 1. The DNA construct, BS7-4C3-SE1 (data not shown) or BS7-4C3-PLT (with a final concentration of 1-2 nM), was added to the nanopore experimental system of the mutant 1. After mixing well, the buffer (625 mM KCl, 10 mM HEPES at pH of 8.0, and 50 mM MgCl₂) flowed through the system to remove any excess DNA construct BS7-4C3-SE1 or BS7-4C3-PLT. A premix of the helicase (Mph-MP1-E105C/A362C with a final concentration of 15 nM) and fuel (ATP with a final concentration of 3 mM) was then added to the nanopore experimental system of the single mutant 1, and the sequencing of the mutant 1 porin was monitored at a voltage of +180 mV.

The mutant pore 1 was opened at a voltage of ± 180 mV. FIG. 15A shows an opening current and gated features of the mutant pore 1 at a voltage of ± 180 mV. FIG. 15B shows a scenario in which a single-stranded nucleic acid passes through the pore of the mutant pore 1 at a voltage of +180 mV. The nucleic acid could pass through the pore. After the addition of the single-stranded nucleic acid, the downward line shows a signal of the nucleic acid passing through the pore.

The single-pore sequencing technique was used to sequence the DNA construct BS7-4C3-PLT through the mutant pore 1, and after the pore was embedded, the nucleic acid sequencing signal that appeared in the sequencing system was added. FIGs. 16A and 16B show example current trajectories when the helicase Mph-MP1-E105C/A362C controls the translocation of the DNA construct BS7-4C3-PLT through the mutant pore 1. Based on the signal characteristics, the mutant pore 1 could be used for nucleic acid sequencing.

FIG. 17 is an enlarged result of the current trajectory shown in a portion of FIG. 16B. The diagram (middle diagram) with dashed boxes and arrows shows the result of filtering the original signal (for the two trajectories, the y-axis coordinate = current (pA), and the x-axis coordinate = time (s)). The dotted arrow indicative portions show enlarged results of the current trajectory. The enlarged region display of this single signal further demonstrates that mutant pore 1 could be used for the nucleic acid sequencing.

Example 11

Similar to Example 10, Example 11 used the mutant pore 2 for the empty test and through-pore test.

FIG. 18A shows an opening current and gated features of the mutant pore 2 at a voltage of ± 180 mV. FIG. 18B shows a scenario in which a single-stranded nucleic acid passes through the pore of the mutant pore 2 at a voltage of +180 mV. The nucleic acid could pass through the pore. After the addition of the single-stranded nucleic acid, the downward line shows a signal of the nucleic acid passing through the pore.

The single-pore sequencing technique was used to sequence the DNA construct BS7-4C3-PLT through the mutant pore 2, and after the pore was embedded, the nucleic acid sequencing signal that appeared in the sequencing system was added. FIGs. 19A and 19B show example current trajectories when the helicase Mph-MP1-E105C/A362C controls the translocation of the DNA construct BS7-4C3-PLT through the mutant pore 2. According to the signal characteristics, the sequencing resolution, stability, signal consistency, and other related characteristics of the mutant pore 2 could be obtained. The pore had clear steps, significant jump distribution, and high-precision sequencing capability. From the signal

characteristics, the consistency of the sequencing signals was relatively high.

FIG. 20 shows an enlarged result of a portion of the current trajectory. The diagram with dashed boxes and arrows shows the result of filtering the original signal (for the two trajectories, the y-axis coordinate = current (pA), and the x-axis coordinate = time (s)). The dotted arrow indicative portions show enlarged results of the current trajectory. The enlarged region display of this single signal indicates that the mutant pore had a high resolution for the nucleic acid sequencing.

Example 12

Similar to Example 10, Example 12 used the mutant pore 3 for the empty test and through-pore test.

FIG. 21A shows an opening current and gated features of the mutant pore 3 at a voltage of ± 180 mV. FIG. 21B shows a scenario in which a single-stranded nucleic acid passes through the pore of the mutant pore 3 at a voltage of +180 mV. The nucleic acid could pass through the pore. After the addition of the single-stranded nucleic acid, the downward line shows a signal of the nucleic acid passing through the pore.

The single-pore sequencing technique was used to sequence the DNA construct BS7-4C3-PLT through the mutant pore 3, and after the pore was embedded, the nucleic acid sequencing signal that appeared in the sequencing system was added. FIGs. 22A and 22B show example current trajectories when the helicase Mph-MP1-E105C/A362C controls the translocation of the DNA construct BS7-4C3-PLT through the mutant pore 3. Based on the signal characteristics, the mutant pore 3 could be used for nucleic acid sequencing.

FIG. 23 is an enlarged result of the current trajectory shown in a portion of FIG. 22A. The diagram with dashed boxes and arrows shows the result of filtering the original signal (for the two trajectories, the y-axis coordinate = current (pA), and the x-axis coordinate = time (s)). The dotted arrow indicative portions show enlarged results of the current trajectory. The enlarged region display of this single signal further demonstrates that mutant pore 3 could be used for the nucleic acid sequencing.

10 Example 13

Similar to Example 10, Example 13 used the mutant pore 4 for the empty test and through-pore test.

FIG. 24A shows an opening current and gated features of the mutant pore 4 at a voltage of ± 180 mV. FIG. 24B shows a scenario in which a single-stranded nucleic acid passes through the pore of the mutant pore 4 at a voltage of +180 mV. The nucleic acid could pass through the pore.

The single-pore sequencing technique was used to sequence the DNA construct BS7-4C3-PLT through the mutant pore 4, and after the pore was embedded, the nucleic acid sequencing signal that appeared in the sequencing system was added. FIGs. 25A and 25B show example current trajectories when the helicase Mph-MP1-E105C/A362C controls the translocation of the DNA construct BS7-4C3-PLT through the mutant pore 4. Based on the signal characteristics, the mutant pore 4 could be used for nucleic acid sequencing.

FIG. 26 is an enlarged result of the current trajectory shown in portions of the examples in FIGs. 25A and 25B. The diagram with dashed boxes and arrows shows the result of filtering the original signal (for the two

trajectories, the y-axis coordinate = current (pA), and the x-axis coordinate = time (s)). The dotted arrow indicative portions show enlarged results of the current trajectory. The enlarged region display of this single signal further demonstrates that mutant pore 4 could be used for the nucleic acid sequencing.

Example 14

Similar to Example 10, Example 14 used the mutant pore 5 for the empty test and through-pore test.

FIG. 27 shows an opening current and gated features of a mutant pore 5 at a voltage of ± 180 mV.

The single-pore sequencing technique was used to sequence the DNA construct BS7-4C3-PLT through the mutant pore 5, and after the pore was embedded, the nucleic acid sequencing signal that appeared in the sequencing system was added. FIG. 28 shows an example current trajectory when the helicase Mph-MP1-E105C/A362C controls the translocation of the DNA construct BS7-4C3-PLT through the mutant pore 5. Based on the signal characteristics, the mutant pore 5 could be used for nucleic acid sequencing.

FIG. 29 is an enlarged result of the current trajectory shown in a portion of the example in FIG. 28. The diagram with dashed boxes and arrows shows the result of filtering the original signal (for the two trajectories, the y-axis coordinate = current (pA), and the x-axis coordinate = time (s)). The dotted arrow indicative portions show enlarged results of the current trajectory. The enlarged region display of this single signal further demonstrates that mutant pore 5 could be used for the nucleic acid sequencing.

Example 15

Similar to Example 10, Example 15 used the mutant pore 6 for the empty test and through-pore test.

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FIG. 30A shows an opening current and gated features of the mutant pore 6 at a voltage of ± 180 mV. FIG. 30B shows a scenario in which a single-stranded nucleic acid passes through the pore of the mutant pore 6 at a voltage of +180 mV. The nucleic acid could pass through the pore.

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After the addition of the single-stranded nucleic acid, the downward line shows a signal of the nucleic acid passing through the pore.

The single-pore sequencing technique was used to sequence the DNA construct BS7-4C3-PLT through the mutant pore 6, and after the pore was embedded, the nucleic acid sequencing signal that appeared in the sequencing system was added. FIGs. 31A and 31B show example current trajectories when the helicase Mph-MP1-E105C/A362C controls the translocation of the DNA construct BS7-4C3-PLT through the mutant pore 6. According to the signal characteristics, the sequencing resolution, stability, signal consistency, and other related characteristics of the mutant pore 6 could be obtained. The pore had clear steps, significant jump distribution, and high-precision sequencing capability. From the signal characteristics, the consistency of the sequencing signals was relatively high.

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FIG. 32 is an enlarged result of the current trajectory shown in a portion of FIG. 31A. The diagram with dashed boxes and arrows shows the result of filtering the original signal (for the two trajectories, the y-axis coordinate = current (pA), and the x-axis coordinate = time (s)). The dotted arrow indicative portions show enlarged results of the current trajectory. The

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enlarged region display of this single signal indicates that the mutant pore had a high resolution for the nucleic acid sequencing.

Example 16

5 A recombinant plasmid containing the nucleic acid sequence of the mutant 1 of the porin monomer (SEQ ID NO: 25) was transformed into BL21(DE3) competent cells by heat shock, and 0.5 mL of LB culture medium was added. The cells were cultured at 30 °C for 1 h, and then a proper amount of bacterial liquid was taken and coated on an
10 ampicillin-resistant solid LB plate. The plate was cultured at 37 °C overnight. Monoclonal colonies were picked the next day and inoculated into 50 mL of liquid LB culture medium containing ampicillin resistance, and the colonies were cultured at 37 °C overnight. The colonies were transferred to an ampicillin-resistant TB liquid culture medium at an
15 inoculation amount of 1% for expansion culture. The colonies were cultured at 37 °C and 220 rpm and continuously measured for OD600 values. When OD600 = 2.0-2.2, the culture liquid in the TB culture medium was cooled to 16 °C, and isopropyl β -D-thiogalactoside (IPTG) was added to induce the expression, such that the final concentration
20 reached 0.015 mM. After the expression was induced for 20-24 h, the bacteria were collected by centrifugation. The bacteria were resuspended in a crushing buffer and then crushed at high pressure. The mixed solution was purified by Ni-NTA affinity chromatography, and a target elution sample was collected. The mutants 2-6 of the porin monomer were purified
25 and obtained according to the method described above.

Illustratively, FIG. 33 shows protein purification results of the mutant 1, and SDS-PAGE electrophoresis results of the separated different components are shown in lanes 1-4. FIG. 34 shows an example of size
30 exclusion chromatography (SEC) of the mutant 1 protein (25 mL

superose-6 GE healthcare; x-axis coordinate = elution volume (mL), and the Y-axis coordinate = absorbance (mAu)).

Claims

1. A mutant of a porin monomer, wherein an amino acid of the mutant of the porin monomer comprises a sequence set forth in SEQ ID NO: 1 or a sequence having at least 99%, 98%, 97%, 96%, 95%, 90%, 80%, 70%,
5 60%, or 50% identity thereto, and the amino acid of the mutant of the porin monomer comprises mutations at one or more positions corresponding to T71, S75, or F76 of SEQ ID NO: 1.

2. The mutant of the porin monomer according to claim 1, wherein the
10 amino acid of the mutant of the porin monomer comprises mutations at one or more positions corresponding to 62-175, 62-104, 68-175, 64-79, 71-76, or 69-76 of SEQ ID NO: 1.

3. The mutant of the porin monomer according to claim 1 or 2, wherein the
15 amino acid of the mutant of the porin monomer comprises:
(1) an insertion, a deletion, and/or a substitution of an amino acid at one or more positions corresponding to K69, P70, T71, P72, A73, S74, S75, and F76 of SEQ ID NO: 1; (2) an insertion, a deletion, and/or a substitution of an amino acid at one or more positions corresponding to Q64, T65, G66,
20 Q67, Y68, K69, P70, T71, P72, A73, S74, S75, F76, S77, T78, and S79 of SEQ ID NO: 1; (3) an insertion, a deletion, and/or a substitution of an amino acid at one or more positions corresponding to R62, D63, Y68, K69, T71, S75, F76, and E104 of SEQ ID NO: 1; or (4) an insertion, a deletion, and/or a substitution of an amino acid at one or more positions
25 corresponding to Y68, K69, P70, T71, P72, A73, S74, S75, F76, E171, and D175 of SEQ ID NO: 1.

4. The mutant of the porin monomer according to any one of the preceding claims, wherein the sequence set forth in SEQ ID NO: 1 is derived from
30 *Pseudomonas taeanensis*.

5. The mutant of the porin monomer according to any one of the preceding claims, wherein the amino acid mutation of the mutant of the porin monomer is selected from the group consisting of:

- 5 (a) mutations from amino acids KPTPASSF corresponding to positions 69-76 of SEQ ID NO: 1 to $M_1M_2M_3M_4M_5M_6M_7M_8$, wherein M_1 is selected from 0 to 3 of R, K, and H; M_2 is selected from 0 to 1 of P; M_3 is selected from 0 to 10 of S, G, C, U, T, M, A, V, L, and I; M_4 is selected from 0 to 1 of P; M_5 is selected from 0 to 5 of A, G, V, L, and I; M_6 is selected from 0 to 5 of S, C, U, T, and M; M_7 is selected from 0 to 10 of A, T, G, V, L, I, S, C, U, and M; M_8 is selected from 0 to 7 of Q, D, E, N, K, H, and R;
- 10 (b) mutations from amino acids QTGQYKPTPASSFSTS corresponding to positions 64-79 of SEQ ID NO: 1 to $M_9M_{10}M_{11}M_{12}M_{13}M_{14}M_{15}M_{16}M_{17}M_{18}M_{19}M_{20}M_{21}M_{22}M_{23}M_{24}$, wherein M_9 is selected from 0 to 5 of L, G, A, V, and I; M_{10} is selected from 0 to 5 of T, S, C, U, and M; M_{11} is selected from 0 to 5 of G, A, V, L, and I; M_{12} is selected from 0 to 4 of Q, D, E, and N; M_{13} is selected from 0 to 3 of Y, F, and W; M_{14} is selected from 0 to 3 of R, H, and K; M_{15} is selected from 0 to 1 of P; M_{16} is selected from 0 to 5 of S, C, U, T, and M; M_{17} is selected from 0 to 1 of P;
- 15 M_{18} is selected from 0 to 5 of A, G, V, L, and I; M_{19} is selected from 0 to 5 of S, C, U, T, and M; M_{20} is selected from 0 to 5 of A, G, V, L, and I; M_{21} is selected from 0 to 9 of N, D, E, Q, L, G, A, V, and I; M_{22} is selected from 0 to 5 of S, C, U, T, and M; M_{23} is selected from 0 to 5 of T, S, C, U, and M; M_{24} is selected from 0 to 5 of A, G, V, L, and I;
- 20 (c) a mutation corresponding to position 62 of SEQ ID NO: 1 being 0 to 5 of S, C, U, T, and M; a mutation at position 63 being 0 to 5 of V, G, A, L, and I; a mutation at position 68 being 0 to 2 of F and W; a mutation at position 69 being 0 to 2 of R and H; a mutation at position 71 being 0 to 4 of S, C, U, and M; a mutation at position 75 being 0 to 5 of A, G, V, L, and I; a mutation at position 76 being 0 to 4 of Q, D, E, and N; a mutation
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at position 104 being 0 to 5 of V, G, A, L, and I; and

(d) mutations from amino acids YKPTPASSF corresponding to positions 68-76 of SEQ ID NO: 1 to M₂₅M₂₆M₂₇M₂₈M₂₉M₃₀M₃₁M₃₂M₃₃, a mutation at position 171 being 0 to 3 of N, D, and Q, and a mutation at position 175 being 0 to 3 of N, E, and Q, wherein M₂₅ is selected from 0 to 3 of F, Y, and W; M₂₆ is selected from 0 to 3 of R, H, and K; M₂₇ is selected from 0 to 1 of P; M₂₈ is selected from 0 to 5 of S, C, U, T, and M; M₂₉ is selected from 0 to 1 of P; M₃₀ is selected from 0 to 5 of A, G, V, L, and I; M₃₁ is selected from 0 to 5 of S, C, U, T, and M; M₃₂ is selected from 0 to 5 of A, G, V, L, and I; M₃₃ is selected from 0 to 4 of Q, D, E, and N.

6. The mutant of the porin monomer according to any one of the preceding claims, wherein the amino acid mutation of the mutant of the porin monomer is selected from the group consisting of:

(a) mutations from the amino acids KPTPASSF corresponding to positions 69-76 of SEQ ID NO: 1 to RPSPASAQ;

(b) mutations from the amino acids KPTPASSF corresponding to positions 69-76 of SEQ ID NO: 1 to KPGPASTK;

(c) mutations from the amino acids QTGQYKPTPASSFSTS corresponding to positions 64-79 of SEQ ID NO: 1 to LTGQYRPSPASANSTA;

(d) mutations from the amino acids QTGQYKPTPASSFSTS corresponding to positions 64-79 of SEQ ID NO: 1 to LTGQYRPSPASALSTA;

(e) R62S, D63V, Y68F, K69R, T71S, S75A, F76Q, and E104V corresponding to SEQ ID NO: 1; and

(f) mutations from the amino acids YKPTPASSF corresponding to positions 68-76 of SEQ ID NO: 1 to FRPSPASAQ, and E171N and D175N corresponding to SEQ ID NO: 1.

7. The mutant of the porin monomer according to any one of the preceding claims, wherein the mutant of the porin monomer comprises or consists of

an amino acid sequence set forth in SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, or SEQ ID NO: 30.

5 8. A protein pore comprising at least one mutant of the porin monomer according to any one of the preceding claims.

9. The protein pore according to claim 8, wherein the protein pore comprises at least two mutants of the porin monomer.

10 10. The protein pore according to any one of claims 8-9, wherein the protein pore has a pore channel diameter of a constriction zone of 0.7 nm to 2.2 nm, 0.9 nm to 1.6 nm, 1.4 nm to 1.6 nm, or 15.9 Å to 20.1 Å.

15 11. A complex for characterizing a target analyte, wherein the complex comprises the protein pore according to any one of claims 8-10 and a rate-controlling protein bound thereto.

20 12. A nucleic acid encoding the mutant of the porin monomer according to any one of claims 1-7, the protein pore according to any one of claims 8-10, or the complex according to claim 11.

13. The nucleic acid according to claim 12, wherein the nucleotide sequence of the porin monomer is a sequence set forth in SEQ ID NO: 2.

25 14. A vector or a genetically engineered host cell comprising the nucleic acid according to any one of claims 12-13.

30 15. Use of the mutant of the porin monomer according to any one of claims 1-7, the protein pore according to any one of claims 8-10, the complex according to claim 11, the nucleic acid according to any one of

claims 12-13, or the vector or host cell according to claim 14 in the detection of the presence, absence, or one or more characteristics of a target analyte or in the preparation of a product for detecting the presence, absence, or one or more characteristics of a target analyte.

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16. A method for producing a protein pore or a polypeptide thereof, comprising transforming the host cell according to claim 14 with the vector according to claim 14, and inducing the host cell to express the protein pore according to any one of claims 8-10 or the polypeptide thereof.

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17. A method for determining the presence, absence, or one or more characteristics of a target analyte, comprising:

a. contacting the target analyte with the protein pore according to any one of claims 8-10, the complex according to claim 11, or the protein pore in the complex according to claim 11, such that the target analyte moves relative to the protein pore; and

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b. acquiring one or more measurements when the target analyte moves relative to the protein pore, thereby determining the presence, absence, or one or more characteristics of the target analyte.

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18. The method according to claim 17, wherein the method comprises: the target analyte interacting with the protein pore present in a membrane, such that the target analyte moves relative to the protein pore.

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19. A kit for determining the presence, absence, or one or more characteristics of a target analyte, comprising the mutant of the porin monomer according to any one of claims 1-7, the protein pore according to any one of claims 8-10, the complex according to claim 11, the nucleic acid according to any one of claims 12-13, or the vector or host cell

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according to claim 14, and a component of the membrane according to claim 18.

5 20. A device for determining the presence, absence, or one or more characteristics of a target analyte, comprising the protein pore according to any one of claims 8-10 or the complex according to claim 11, and the membrane according to claim 18.

10 21. The use, method, kit, or device according to any one of claims 15-20, wherein the target analyte comprises a polysaccharide, a metal ion, an inorganic salt, a polymer, an amino acid, a peptide, a protein, a nucleotide, an oligonucleotide, a polynucleotide, a dye, a drug, a diagnostic agent, an explosive, or an environmental contaminant;
preferably, the target analyte comprises a polynucleotide;
15 more preferably, the polynucleotide comprises DNA or RNA; and/or, the one or more characteristics are selected from (i) a length of the polynucleotide; (ii) an identity of the polynucleotide; (iii) a sequence of the polynucleotide; (iv) a secondary structure of the polynucleotide; and (v) whether the polynucleotide is modified; and/or, the rate-controlling
20 protein in the complex comprises a polynucleotide binding protein.