

SINGLE CELL ANALYSIS OF TRANSPOSASE ACCESSIBLE CHROMATIN**CROSS-REFERENCE**

[0001] This application claims priority to U.S. Provisional Patent Application No. 62/511,905, filed May 26, 2017; U.S. Provisional Patent Application No. 62/527,529, filed June 30, 2017; and U.S. Provisional Patent Application No. 62/650,223, filed March 29, 2018, each of which is entirely incorporated herein by reference.

BACKGROUND

[0002] Samples may be processed for various purposes, such as identification of a type of sample or moiety within the sample. The sample may be a biological sample. The biological samples may be processed for various purposes, such as detection of a disease (e.g., cancer) or identification of a particular species. There are various approaches for processing samples, such as polymerase chain reaction (PCR) and sequencing.

[0003] Biological samples may be processed within various reaction environments, such as partitions. Partitions may be wells or droplets. Droplets or wells may be employed to process biological samples in a manner that enables the biological samples to be partitioned and processed separately. For example, such droplets may be fluidically isolated from other droplets, enabling accurate control of respective environments in the droplets.

[0004] Biological samples in partitions may be subjected to various processes, such as chemical processes or physical processes. Samples in partitions may be subjected to heating or cooling, or chemical reactions, such as to yield species that may be qualitatively or quantitatively processed.

[0005] Certain applications may benefit from the amplification or sequencing of species obtained from single cells obtained from a much larger population. In some cases, the single cells of interest may be quite rare.

SUMMARY

[0006] The present disclosure provides sample preparation techniques that allow sequencing of nucleic acids from single cells of interest.

[0007] In eukaryotic genomes, chromosomal DNA winds itself around histone proteins (i.e., “nucleosomes”), thereby forming a complex known as chromatin. The tight or loose packaging of chromatin contributes to the control of gene expression. Tightly packed chromatin (“closed chromatin”) is usually not permissive for gene expression while more loosely packaged, accessible regions of chromatin (“open chromatin”) is associated with the active transcription of gene products. Methods for probing genome-wide DNA accessibility have proven extremely

effective in identifying regulatory elements across a variety of cell types and quantifying changes that lead to both activation or repression of gene expression.

[0008] One such method is the Assay for Transposase Accessible Chromatin with high-throughput sequencing (ATAC-seq). The ATAC-seq method probes DNA accessibility with an artificial transposon, which inserts specific sequences into accessible regions of chromatin. Because the transposase can only insert sequences into accessible regions of chromatin not bound by transcription factors and/or nucleosomes, sequencing reads can be used to infer regions of increased chromatin accessibility.

[0009] Traditional approaches to the ATAC-seq methodology requires large pools of cells, processes cells in bulk, and result in data representative of an entire cell population, but lack information about cell-to-cell variation inherently present in a cell population (see, *e.g.*, Buenrostro, *et al.*, Curr. Protoc. Mol. Biol., 2015 Jan 5; 21.29.1–21.29.9). While single cell ATAC-seq (scATAC-seq) methods have been developed, they suffer from several limitations. For example, scATAC-seq methods that utilize sample pooling, cell indexing, and cell sorting (see, *e.g.*, Cusanovich, *et al.*, Science, 2015 May 22;348(6237):910-14) result in high variability and a low number of reads associated with any single cell. Other scATAC-seq methods that utilize a programmable microfluidic device to isolate single cells and perform scATAC-seq in nanoliter reaction chambers (see, *e.g.*, Buenrostro, *et al.*, Nature, 2015 Jul 23;523(7561):486-90) are limited by the throughput of the assay and may not generate personal epigenomic profiles on a timescale compatible with clinical decision-making.

[0010] In an example, the present disclosure provides a method for nucleic acid processing, comprising: (a) generating a partition comprising: (i) a biological particle comprising chromatin comprising a template nucleic acid molecule; (ii) a plurality of nucleic acid barcode molecules comprising a common barcode sequence; (iii) a plurality of transposon end nucleic acid molecules comprising a transposon end sequence; and (iv) a plurality of transposase molecules; (b) generating a plurality of template nucleic acid fragments with the aid of a transposase-nucleic acid complex comprising a transposase molecule of the plurality of transposase molecules and a transposon end oligonucleotide molecule of the plurality of transposon end oligonucleotide molecules; and (c) generating a barcoded nucleic acid fragment using a nucleic acid barcode molecule of the plurality of nucleic acid barcode molecules and a template nucleic acid fragment of the plurality of template nucleic acid fragments.

[0011] In some examples, the barcoded nucleic acid fragment is generated by ligation of the nucleic acid barcode molecule with the template nucleic acid fragment, or a derivative thereof.

- [0012]** In some examples, the barcoded nucleic acid fragment is generated by amplification of the template nucleic acid fragment, or a derivative thereof, using the nucleic acid barcode molecule as a primer.
- [0013]** In some examples, the transposase-nucleic acid complex is generated in the partition using transposase molecules of the plurality of transposase molecules and transposon end nucleic acid molecules of the plurality of transposon end nucleic acid molecules.
- [0014]** In some examples, the transposase-nucleic acid complex is partitioned into the partition.
- [0015]** In some examples, the plurality of nucleic acid barcode molecules are attached to a bead and wherein the partition further comprises the bead. In some examples, the nucleic acid barcode molecules are releasably attached to the bead. In some examples, the method further comprises releasing the plurality of nucleic acid barcode molecules from the bead. In some examples, the bead is a gel bead. In some examples, the gel bead is a degradable gel bead. In some examples, the degradable gel bead is degradable upon application of a stimulus. In some examples, the stimulus is a chemical stimulus. In some examples, the stimulus is a reducing agent. In some examples, the partition further comprises the chemical stimulus.
- [0016]** In some examples, prior to (b), the template nucleic acid molecule is released from the biological particle. In some examples, subsequent to (b), the plurality of template nucleic acid fragments is released from the biological particle.
- [0017]** In some examples, the biological particle is a cell. In some examples, the biological particle is a nucleus.
- [0018]** In some examples, the partition is an aqueous droplet in an emulsion. In some examples, the partition is a well.
- [0019]** In some examples, the method further comprises releasing or removing the barcoded nucleic acid fragment or a derivative thereof from the partition.
- [0020]** In some examples, the method further comprises sequencing the barcoded nucleic acid fragment, or a derivative thereof.
- [0021]** In some examples, the plurality of transposon end nucleic acid molecules further comprise a sequence complementary to a sequence of the plurality of nucleic acid barcode molecules. In some examples, the partition further comprises a plurality of second nucleic acid molecules comprising a transposon end sequence and a primer sequence, and wherein the transposase-nucleic acid complex comprises: (i) transposase molecules of the plurality of transposase molecules; (ii) a transposon end oligonucleotide molecule of the plurality of transposon end oligonucleotide molecules; and (iii) a second nucleic acid molecule of the

plurality of second nucleic acid molecules. In some examples, the plurality of nucleic acid barcode molecules are partially double-stranded and comprise (i) a first strand comprising the common barcode sequence and a sequence complementary to a sequence of the plurality of first nucleic acid molecules; and (ii) a second strand comprising a sequence complementary to the common barcode sequence. In some examples, the plurality of nucleic acid barcode molecules are single-stranded and comprise the common barcode sequence and a sequence complementary to a sequence in the plurality of transposon end nucleic acid molecules.

[0022] In some examples, the biological particle further comprises a template ribonucleic acid (RNA) molecule, and wherein the method further comprises generating a barcoded complementary deoxyribonucleic acid (cDNA) molecule from the template RNA molecule or a derivative thereof. In some examples, the barcoded cDNA molecule is generated in the partition. In some examples, the partition comprises a second plurality of nucleic acid barcode molecules comprising the common barcode sequence and a capture sequence. In some examples, the capture sequence is a poly-T sequence. In some examples, the capture sequence is a template switching oligo sequence and wherein the barcoded cDNA molecule is generated using a template switching reaction. In some examples, the plurality of nucleic acid barcode molecules are attached to a first bead, the second plurality of nucleic acid barcode molecules are attached to a second bead, and wherein the partition comprises the first bead and the second bead. In some examples, the first bead or the second bead is a magnetic bead. In some examples, the first bead is a gel bead and the second bead is a magnetic bead. In some examples, the gel bead comprises the magnetic bead. In some examples, the method further comprises degrading the gel bead. In some examples, the first bead is a magnetic bead and the second bead is a gel bead. In some examples, the gel bead comprises the magnetic bead. In some examples, the method further comprises degrading the gel bead. In some examples, the plurality of nucleic acid barcode molecules and the second plurality of nucleic acid barcode molecules are attached to a bead, and wherein the partition further comprises the bead. In some examples, the barcoded nucleic acid fragment and the barcoded cDNA molecule is released from the partition. In some examples, the method further comprises sequencing the barcoded nucleic acid fragment or a derivative thereof and the barcoded cDNA molecule or a derivative thereof.

[0023] In some examples, the method further comprises cleaving one or more mitochondrial nucleic acid fragments, or derivatives thereof, using (i) one or more guide ribonucleic acid molecules (gRNAs) targeted to the one or more mitochondrial nucleic acid fragments; and (ii) a clustered regularly interspaced short palindromic (CRISPR) associated

(Cas) nuclease. In some examples, the partition further comprises the one or more gRNAs and the Cas nuclease. In some examples, the Cas nuclease is Cas9.

[0024] In another example, the present disclosure provides a method of generating barcoded nucleic acid fragments, comprising: (a) providing: (i) a plurality of biological particles, an individual biological particle of the plurality of biological particles comprising chromatin comprising a template nucleic acid; (ii) a plurality of transposon end nucleic acid molecules comprising a transposon end sequence; and (iii) a plurality of transposase molecules; (b) generating a plurality of template nucleic acid fragments in a biological particle of the plurality of biological particles with the aid of a transposase-nucleic acid complex comprising a transposase molecule of the plurality of transposase molecules and a transposon end nucleic acid molecule of the plurality of transposon end nucleic acid molecules; (c) generating a partition comprising the biological particle comprising the plurality of template nucleic acid fragments and a plurality of nucleic acid barcode molecules comprising a common barcode sequence; and (d) generating a barcoded nucleic acid fragment using a nucleic acid barcode molecule of the plurality of nucleic acid barcode molecules and a template nucleic acid fragment of the plurality of template nucleic acid fragments.

[0025] In some examples, the barcoded nucleic acid fragment is generated by ligation of the nucleic acid barcode molecule with the template nucleic acid fragment, or a derivative thereof.

[0026] In some examples, the barcoded nucleic acid fragment is generated by nucleic acid amplification of the template nucleic acid fragment, or a derivative thereof, using the nucleic acid barcode molecule as a primer.

[0027] In some examples, the plurality of nucleic acid barcode molecules are attached to a bead and wherein the partition further comprises the bead. In some examples, the nucleic acid barcode molecules are releasably attached to the bead. In some examples, the method further comprises releasing the plurality of nucleic acid barcode molecules from the bead. In some examples, the bead is a gel bead. In some examples, the gel bead is a degradable gel bead. In some examples, the degradable gel bead is degradable upon application of a stimulus. In some examples, the stimulus is a chemical stimulus. In some examples, the stimulus is a reducing agent. In some examples, the partition further comprises the chemical stimulus.

[0028] In some examples, subsequent to (c), the template nucleic acid fragments are released from the biological particle.

[0029] In some examples, the biological particle is a cell. In some examples, the cell is a permeabilized cell.

[0030] In some examples, the biological particle is a nucleus. In some examples, the nucleus is a permeabilized nucleus.

[0031] In some examples, the partition is an aqueous droplet in an emulsion. In some examples, the partition is a well.

[0032] In some examples, the method further comprises releasing or removing the barcoded nucleic acid fragment or a derivative thereof from the partition.

[0033] In some examples, the method further comprises sequencing the barcoded nucleic acid fragment, or a derivative thereof.

[0034] In some examples, the plurality of transposon end nucleic acid molecules comprise a plurality of first nucleic acid molecules comprising a transposon end sequence and a sequence complementary to a sequence of the plurality of nucleic acid barcode molecules. In some examples, the plurality of transposon end nucleic acid molecules further comprise a plurality of second nucleic acid molecules comprising a transposon end sequence and a primer sequence. In some examples, the plurality of nucleic acid barcode molecules are partially double-stranded and comprise a first strand comprising the common barcode sequence and a sequence complementary to a sequence of the plurality of first nucleic acid molecules; and a second strand comprising a sequence complementary to the common barcode sequence. In some examples, the plurality of nucleic acid barcode molecules are single-stranded and comprise the common barcode sequence and a sequence complementary to a sequence of the plurality of first nucleic acid molecules.

In some examples, the biological particle further comprises a template RNA molecule, and wherein the method further comprises generating a barcoded cDNA molecule from the template RNA molecule or a derivative thereof. In some examples, the barcoded cDNA molecule is generated in the partition. In some examples, the partition comprises a second plurality of nucleic acid barcode molecules comprising the common barcode sequence and a capture sequence. In some examples, the capture sequence is a poly-T sequence. In some examples, the capture sequence is a template switching oligo sequence and wherein the barcoded cDNA molecule is generated using a template switching reaction. In some examples, the plurality of nucleic acid barcode molecules are attached to a first bead, the second plurality of nucleic acid barcode molecules are attached to a second bead, and wherein the partition comprises the first bead and the second bead. In some examples, the first bead or the second bead is a magnetic bead. In some examples, the first bead is a gel bead and the second bead is a magnetic bead. In some examples, the gel bead comprises the magnetic bead. In some examples, the method further comprises degrading the gel bead. In some examples, the first bead is a magnetic bead and the second bead is a gel bead. In some

examples, the gel bead comprises the magnetic bead. In some examples, the method further comprises degrading the gel bead. In some examples, the plurality of nucleic acid barcode molecules and the second plurality of nucleic acid barcode molecules are attached to a bead, and wherein the partition further comprises the bead. In some examples, the barcoded nucleic acid fragment and the barcoded cDNA molecule is released from the partition. In some examples, the method further comprises sequencing the barcoded nucleic acid fragment or a derivative thereof and the barcoded cDNA molecule or a derivative thereof.

[0035] In some examples, the method further comprises (a) lysing a plurality of cells in the presence of a cell lysis agent and a blocking agent; and (b) separating a plurality of nuclei from the plurality of lysed cells to generate the plurality of biological particles, wherein the blocking agent reduces the fraction of sequencing reads corresponding to a mitochondrial genome compared to sequencing reads obtained in the absence of the blocking agent. In some examples, the blocking agent is bovine serum albumin (BSA).

[0036] In some examples, the method further comprises cleaving one or more mitochondrial nucleic acid molecules or derivatives thereof, using (i) one or more guide ribonucleic acid molecules (gRNAs) targeted to the one or more mitochondrial nucleic acid fragments; and (ii) a clustered regularly interspaced short palindromic (CRISPR) associated (Cas) nuclease. In some examples, the Cas nuclease is Cas9. In some examples, the mitochondrial nucleic acid molecules are cleaved prior to (c). In some examples, the mitochondrial nucleic acid molecules are cleaved in the partition. In some examples, the partition comprises the Cas nuclease. In some examples, the partition further comprises the one or more gRNAs. In some examples, the mitochondrial nucleic acid molecules are cleaved subsequent to (d).

[0037a] In one aspect, the present disclosure provides a method of generating barcoded nucleic acid fragments, comprising:

(a) providing: (i) a plurality of cells or cell nuclei comprising chromatin comprising a template nucleic acid molecule; (ii) a plurality of oligonucleotide molecules comprising a transposon end sequence; and (iii) a plurality of transposase molecules;

(b) in said plurality of cells or cell nuclei, generating a plurality of template nucleic acid fragments with the aid of a plurality of transposase-nucleic acid complexes comprising a transposase molecule of said plurality of transposase molecules and an oligonucleotide molecule of said plurality of oligonucleotide molecules;

(c) partitioning said plurality of cells or cell nuclei into a plurality of partitions, each comprising (i) a cell or a cell nucleus of said plurality of cells or cell nuclei, said cell or cell nucleus comprising a template nucleic acid fragment of said plurality of template nucleic acid fragments, and (ii) a plurality of nucleic acid barcode molecules comprising a common barcode sequence; and

(d) in a partition of said plurality of partitions, generating a barcoded nucleic acid fragment using the nucleic acid barcode molecule of said plurality of nucleic acid barcode molecules

and a template nucleic acid fragment of said plurality of template nucleic acid fragments.

[0037] Additional examples and advantages of the present disclosure will become readily apparent to those skilled in this art from the following detailed description, wherein only illustrative examples of the present disclosure are shown and described. As will be realized, the present disclosure is capable of other and different examples, and its several details are capable of modifications in various obvious respects, all without departing from the disclosure. Accordingly, the drawings and description are to be regarded as illustrative in nature, and not as restrictive.

INCORPORATION BY REFERENCE

[0038] All publications, patents, and patent applications mentioned in this specification are herein incorporated by reference to the same extent as if each individual publication, patent, or patent application was specifically and individually indicated to be incorporated by reference. To

the extent publications and patents or patent applications incorporated by reference contradict the disclosure contained in the specification, the specification is intended to supersede and/or take precedence over any such contradictory material.

BRIEF DESCRIPTION OF THE DRAWINGS

[0040] The novel features of the invention are set forth with particularity in the appended claims. A better understanding of the features and advantages of the present invention will be obtained by reference to the following detailed description that sets forth illustrative embodiments, in which the principles of the invention are utilized, and the accompanying drawings (also “Figure” and “FIG.” herein), of which:

[0041] **FIG. 1** shows an example of a microfluidic channel structure for partitioning individual biological particles.

[0042] **FIG. 2** shows an example of a microfluidic channel structure for delivering barcode carrying beads to droplets.

[0043] **FIG. 3** shows an example of a microfluidic channel structure for co-partitioning biological particles and reagents.

[0044] **FIG. 4** shows an example of a microfluidic channel structure for the controlled partitioning of beads into discrete droplets.

[0045] **FIG. 5** shows an example of a microfluidic channel structure for increased droplet generation throughput.

[0046] **FIG. 6** shows another example of a microfluidic channel structure for increased droplet generation throughput.

[0047] **FIG. 7A** shows a cross-section view of another example of a microfluidic channel structure with a geometric feature for controlled partitioning. **FIG. 7B** shows a perspective view of the channel structure of **FIG. 7A**.

[0048] **FIG. 8** illustrates an example of a barcode carrying bead.

[0049] **FIG. 9** shows a computer system that is programmed or otherwise configured to implement methods provided herein.

[0050] **FIG. 10** illustrates an exemplary method to produce droplets wherein at least some of the droplets formed will comprise transposase molecules, a single cell, and a single gel bead comprising a forked adaptor.

[0051] **FIGs. 11A-11B** illustrate an exemplary method to produce forked adaptor flanked double-stranded template nucleic acid fragments. **FIG. 11A** illustrates an exemplary method for the in-partition transposition of sequencing adaptors into native chromatin while **FIG. 11B**

illustrates an exemplary method for the in-bulk production of a next-generation sequencing compatible library from the fragments produced in **FIG. 11A**.

[0052] **FIGs. 12A-12B** illustrate exemplary forked adaptors.

[0053] **FIG. 13** illustrates an exemplary method to produce droplets wherein at least some of the droplets formed will comprise transposase-nucleic acid complexes, a single cell, and a single gel bead comprising a forked adaptor.

[0054] **FIGs. 14A-14B** illustrate an alternative exemplary method to produce forked adaptor flanked double-stranded template nucleic acid fragments. **FIG. 14A** illustrates an exemplary method for the in-partition ligation of forked adaptors onto fragments of native chromatin generated by an in-partition transposition reaction. **FIG. 14B** illustrates an exemplary method for the in-bulk production of a next-generation sequencing compatible library from the fragments produced in **FIG. 14A**.

[0055] **FIGs. 15A-15B** illustrate additional exemplary forked adaptors and transposon end sequence containing oligonucleotides.

[0056] **FIG. 16** illustrates an exemplary method to produce droplets wherein at least some of the droplets formed will comprise transposase molecules, a single cell, and a single gel bead comprising a T7-containing adaptor.

[0057] **FIG. 17** illustrates an exemplary method to produce T7-containing adaptor flanked double-stranded template nucleic acid fragments.

[0058] **FIG. 18** illustrates an exemplary T7-containing barcoded adaptor.

[0059] **FIG. 19** illustrates an exemplary method to produce droplets wherein at least some of the droplets formed will comprise transposase molecules, a single cell, and a single gel bead comprising a barcoded adaptor.

[0060] **FIG. 20** illustrates an exemplary scheme for producing barcoded, adapter-flanked nucleic acid fragments.

[0061] **FIG. 21** illustrates an exemplary partially double-stranded barcode oligonucleotide releasably attached to a gel bead.

[0062] **FIG. 22** illustrates a random priming extension reaction scheme.

[0063] **FIG. 23** illustrates an exemplary method of inserting barcodes into a template nucleic acid.

[0064] **FIGs. 24A-24B** illustrate an exemplary transposase-nucleic acid complex showing a transposase, a first partially double-stranded oligonucleotide releasably attached to a gel bead, the first partially double-stranded oligonucleotide comprising a transposon end sequence, a barcode sequence, and a first primer sequence and a second partially double-stranded

oligonucleotide comprising a transposon end sequence and a second primer sequence. In **FIG. 24B**, the first and the second primer sequence are the same.

[0065] **FIGs. 25A-25B** illustrates an exemplary barcode oligonucleotides. **FIG. 25A** illustrates a partially double-stranded oligonucleotide releasably attached to a gel bead, the first strand comprising a transposon end sequence, a barcode sequence, and a first primer sequence and a second strand comprising a sequence complementary to the transposon end sequence. **FIG. 25B** illustrates a partially double-stranded oligonucleotide releasably attached to a gel bead, the first strand comprising a transposon end sequence and a barcode sequence and the second strand comprising a sequence complementary to the transposon end sequence

[0066] **FIG. 26** illustrates an exemplary transposase-nucleic acid complex showing a transposase, a first double-stranded oligonucleotide comprising a barcode sequence and a transposon end sequence releasably attached to a first gel bead and a second double-stranded oligonucleotide comprising a transposon end sequence releasably attached to a second gel bead.

[0067] **FIG. 27** illustrates an exemplary method to produce barcoded nucleic acid fragments suitable for next generation sequencing.

[0068] **FIGs. 28A-28B** illustrate an exemplary transposase-nucleic acid complex and an exemplary barcoded adaptor releasably attached to a gel bead. **FIG. 28A** illustrates an exemplary transposase-nucleic acid complex showing a transposase, a first double-stranded oligonucleotide comprising a transposon end sequence and a second double-stranded oligonucleotide comprising a transposon end sequence. **FIG. 28B** illustrates an exemplary barcoded adaptor comprising a transposon end sequence, a barcode sequence, and a primer sequence releasably attached to a gel bead.

[0069] **FIGs. 29A-29D** illustrate an exemplary transposase-nucleic acid complex and an exemplary barcoded adaptor, which can be releasably attached to a gel bead. **FIG. 29A** illustrates an exemplary transposase-nucleic acid complex showing a transposase, a first double-stranded oligonucleotide comprising a transposon end sequence and a first primer sequence and a second double-stranded oligonucleotide comprising a transposon end sequence and a second primer sequence. **FIG. 28B** illustrates an exemplary barcoded adaptor comprising an adapter sequence, a barcode sequence, and a sequence complementary to the first primer sequence. **FIGs. 28C-D** illustrates an exemplary barcoding scheme.

[0070] **FIGs. 30A-30B** illustrate an exemplary barcode oligonucleotide and combination bulk/in-partition barcoding scheme.

[0071] **FIG. 31** illustrates an exemplary in-partition transposition and barcoding scheme.

[0072] **FIGs. 32A-32C** illustrate an exemplary barcoding scheme. **FIG. 32A** illustrates an exemplary barcode oligonucleotide; **FIG. 32B** illustrates an exemplary combination of bulk/in-

partition barcoding scheme using CRISPR/Cas-9 mediated cleavage; **FIG. 32C** illustrates an exemplary in-partition barcoding scheme using CRISPR/Cas-9 mediated cleavage.

[0073] **FIGs. 33A-33C** illustrate an exemplary barcoding scheme. **FIG. 33A** illustrates an exemplary forked barcode oligonucleotide; **FIG. 33B** illustrates an exemplary combination of bulk/in-partition barcoding scheme using CRISPR/Cas-9 mediated cleavage; **FIG. 33C** illustrates an exemplary in-partition barcoding scheme using CRISPR/Cas-9 mediated cleavage.

[0074] **FIG. 34** illustrates barcoded magnetic beads which can be embedded within a gel bead comprising a barcoded oligonucleotide.

[0075] **FIG. 35** illustrates exemplary gel bead architectures for single cell ATAC-seq (sc-ATAC-seq) plus single cell RNA-seq (scRNA-seq) analyses. BC = barcode.

[0076] **FIGs. 36A-36B** illustrates an exemplary scATAC-seq protocol. **FIG. 36A** provides a schematic of the nuclei preparation of the scATAC-seq protocol. **FIG. 36B** illustrates approximate time required for steps in the scATAC-seq protocol.

[0077] **FIG. 37** illustrates three exemplary partially-double stranded barcode molecules suitable for use with a ligation-mediated ATAC-seq scheme as described in Example 9 and 18. In order to assess differences in the rate of barcode exchange between the three barcode molecules, two modifications were developed: a shorter R1 sequence (middle), as well as replacement of one nucleic acid in the R1 sequence with a uracil (bottom). Shorter R1 (7b); U in R1.

[0078] **FIGs. 38A-38C** illustrate exemplary barnyard plots of mixed human and mouse nuclei demonstrating sequencing reads assembled from scATAC-seq experiments using the barcode molecules in **FIG. 37**. **FIG. 38A** illustrates a barnyard plot using original R1 barcode molecules (“control”). **FIG. 38B** illustrates a barnyard plot using shorter R1 barcode molecules (“7b”). **FIG. 38C** illustrates a barnyard plot using the U-containing barcode molecules (“U40”). These barnyard plots represent separation between the mouse (light grey dots in the upper left quadrant) and human nucleic acid sequence reads (medium grey dots in the lower right quadrant) from noise (black dots in the lower left quadrant) and doublets (dark grey dots in the upper right quadrant).

[0079] **FIG. 39** illustrates an exemplary single stranded barcode molecule suitable for use with a linear amplification-mediated ATAC-seq scheme as described in Example 11. Primers were utilized at either 75 nM, 150nM, or 250 nM concentrations in the droplet.

[0080] **FIG. 40** shows a table comparing sequencing metrics generated from different replicate sample libraries (“A” and “B”) generated using either linear amplification or ligation. “A” replicates represent reactions done in the same conditions, but using a different user, while “B” replicates represent reactions done in the same conditions by the same user.

[0081] **FIGs. 41A-41B** illustrate exemplary amounts of detected mouse or human cells and the inferred doublet rate observed from the analysis of sequencing reads (**FIG. 40**) from a linear amplification ATAC-seq library (using 75 nM, 150nM, or 250 nM of barcode primer) or ligation-based ATAC-seq library (using the barcode molecules of **FIG. 37**). **FIG. 41A** illustrates differences in the number of mouse or human cells detected using either a linear amplification or ligation-based ATAC-seq method. **FIG. 41B** illustrate differences in inferred doublet rate detected using either a linear amplification or ligation-based ATAC-seq method.

[0082] **FIG. 42** illustrates a comparison of the sensitivity of sequencing reads obtained from various ATAC-seq libraries as measured by the median fragments per cell barcode. The sensitivity of the linear amplification and ligation methods of library preparation (36k reads/cell) as compared to the methods described in Buenrostro, *et al.*, Nature, 2015 Jul 23;523(7561):486-90 using a programmable microfluidics platform (Fluidigm).

[0083] **FIG. 43** illustrates a library prepared by ligation sequenced to increased depth (30M reads and 800M reads) compared to **FIGs. 40-42** demonstrate a significant increase in the sensitivity (as measured by median fragments per cell barcode) and reduced noise (as measured by fraction of non-duplicate wasted reads).

[0084] **FIG. 44** illustrates an exemplary comparison of the total noise (non-duplicate wasted reads (Non-dups")) and mitochondrial-based reads ("Mito")) in libraries prepared using linear amplification or ligation-based ATAC-seq methods.

[0085] **FIGs. 45A-45B** provide an exemplary illustration of the breakdown sequencing reads generated by different library preparation methods. **FIG. 45A** shows an illustration of the breakdown of reads generated by a linear amplification ATAC-seq library. **FIG. 45B** shows an illustration of the breakdown of reads generated by a ligation-based ATAC-seq library.

[0086] **FIGs. 46A-46B** illustrate an exemplary comparison of read pairs showing the periodicity of nucleosomes generated from ATAC-seq libraries prepared using either a linear amplification or ligation-based scheme. Nucleosome free fragments are typically observed below 200bp in length, fragments indicative of a nucleosome periodicity of one are approximately 200bp in length, fragments indicative of a nucleosome periodicity of two are approximately 400bp in length, , fragments indicative of a nucleosome periodicity of two are 600bp in length, and so forth. **FIG. 46A** illustrates insert lengths of fragments from a library prepared by linear amplification. **FIG. 46B** illustrates insert lengths of fragments from a library prepared by ligation.

[0087] **FIGs. 47A-47B** illustrate an exemplary enrichment of transcription start sites (TSS) or analysis of CTCF (CCCTC-binding factor) sites observed in ATAC-seq libraries prepared by linear amplification or ligation-based methodologies vs. a traditional bulk ATAC-seq library

preparation using 50,000 nuclei. **FIG. 47A** illustrates exemplary enrichment of TSS data. **FIG. 47B** illustrates enrichment of CTCF sites.

[0088] **FIG. 48** shows a table comparing exemplary sequencing metrics obtained from linear amplification-based ATAC-seq libraries prepared using different polymerases: a Phusion[®] DNA polymerase, a KAPA HiFi DNA polymerase (in combination with betaine), a Deep Vent[®] DNA polymerase, as well as a library prepared by ligation.

[0089] **FIG. 49** shows a table providing exemplary parameters of ~ 1,000 pL droplets vs. smaller ~ 400 pL droplets. The benefits of using a smaller droplet size include ~2-fold decrease in Poisson derived cell doublet rate and an ~2-fold increase in cell throughput.

[0090] **FIG. 50** illustrates protocols for ATAC-seq as described herein compared to data from (1) typical high quality traditional bulk ATAC-seq protocols; (2) Cusanovich, *et al.*, Science, 2015 May 22;348(6237):910-14; (3) Buenrostro, *et al.*, Nature, 2015 Jul 23;523(7561):486-90; (4) ideal sequencing metrics from an ATAC-seq experiment; (5) data obtained using the methods described herein (“10X”).

[0091] **FIGs. 51A-51B** shows the results of scRNA-seq and scATAC-seq analysis of nucleic acids in a peripheral blood mononuclear cell (PBMC) sample. **FIG. 51A** shows an exemplary scatterplot produced using t-Distributed Stochastic Neighbor Embedding (tSNE), allowing visualization of RNA transcripts of different subpopulations cell types in a peripheral blood mononuclear cell (PBMC) sample. **FIG. 51B** shows an exemplary scatterplot produced using t-Distributed Stochastic Neighbor Embedding (tSNE), allowing visualization of ATAC-seq data of different subpopulations cell types in a peripheral blood mononuclear cell (PBMC) sample.

DETAILED DESCRIPTION

[0092] While various embodiments of the invention have been shown and described herein, it will be obvious to those skilled in the art that such embodiments are provided by way of example only. Numerous variations, changes, and substitutions may occur to those skilled in the art without departing from the invention. It should be understood that various alternatives to the embodiments of the invention described herein may be employed.

[0093] Where values are described as ranges, it will be understood that such disclosure includes the disclosure of all possible sub-ranges within such ranges, as well as specific numerical values that fall within such ranges irrespective of whether a specific numerical value or specific sub-range is expressly stated.

[0094] The term “barcode,” as used herein, generally refers to a label, or identifier, that conveys or is capable of conveying information about an analyte. A barcode can be part of an

analyte. A barcode can be independent of an analyte. A barcode can be a tag attached to an analyte (e.g., nucleic acid molecule) or a combination of the tag in addition to an endogenous characteristic of the analyte (e.g., size of the analyte or end sequence(s)). A barcode may be unique. Barcodes can have a variety of different formats. For example, barcodes can include: polynucleotide barcodes; random nucleic acid and/or amino acid sequences; and synthetic nucleic acid and/or amino acid sequences. A barcode can be attached to an analyte in a reversible or irreversible manner. A barcode can be added to, for example, a fragment of a deoxyribonucleic acid (DNA) or ribonucleic acid (RNA) sample before, during, and/or after sequencing of the sample. Barcodes can allow for identification and/or quantification of individual sequencing-reads.

[0095] The term “real time,” as used herein, can refer to a response time of less than about 1 second, a tenth of a second, a hundredth of a second, a millisecond, or less. The response time may be greater than 1 second. In some instances, real time can refer to simultaneous or substantially simultaneous processing, detection or identification.

[0096] The term “subject,” as used herein, generally refers to an animal, such as a mammal (e.g., human) or avian (e.g., bird), or other organism, such as a plant. For example, the subject can be a vertebrate, a mammal, a rodent (e.g., a mouse), a primate, a simian or a human. Animals may include, but are not limited to, farm animals, sport animals, and pets. A subject can be a healthy or asymptomatic individual, an individual that has or is suspected of having a disease (e.g., cancer) or a pre-disposition to the disease, and/or an individual that is in need of therapy or suspected of needing therapy. A subject can be a patient. A subject can be a microorganism or microbe (e.g., bacteria, fungi, archaea, viruses).

[0097] The term “genome,” as used herein, generally refers to genomic information from a subject, which may be, for example, at least a portion or an entirety of a subject’s hereditary information. A genome can be encoded either in DNA or in RNA. A genome can comprise coding regions (e.g., that code for proteins) as well as non-coding regions. A genome can include the sequence of all chromosomes together in an organism. For example, the human genome ordinarily has a total of 46 chromosomes. The sequence of all of these together may constitute a human genome.

[0098] The terms “adaptor(s)”, “adapter(s)” and “tag(s)” may be used synonymously. An adaptor or tag can be coupled to a polynucleotide sequence to be “tagged” by any approach, including ligation, hybridization, or other approaches.

[0099] The term “sequencing,” as used herein, generally refers to methods and technologies for determining the sequence of nucleotide bases in one or more polynucleotides. The polynucleotides can be, for example, nucleic acid molecules such as deoxyribonucleic acid

(DNA) or ribonucleic acid (RNA), including variants or derivatives thereof (e.g., single stranded DNA). Sequencing can be performed by various systems currently available, such as, without limitation, a sequencing system by Illumina®, Pacific Biosciences (PacBio®), Oxford Nanopore®, or Life Technologies (Ion Torrent®). Alternatively or in addition, sequencing may be performed using nucleic acid amplification, polymerase chain reaction (PCR) (e.g., digital PCR, quantitative PCR, or real time PCR), or isothermal amplification. Such systems may provide a plurality of raw genetic data corresponding to the genetic information of a subject (e.g., human), as generated by the systems from a sample provided by the subject. In some examples, such systems provide sequencing reads (also “reads” herein). A read may include a string of nucleic acid bases corresponding to a sequence of a nucleic acid molecule that has been sequenced. In some situations, systems and methods provided herein may be used with proteomic information.

[00100] The term “bead,” as used herein, generally refers to a particle. The bead may be a solid or semi-solid particle. The bead may be a gel bead. The gel bead may include a polymer matrix (e.g., matrix formed by polymerization or cross-linking). The polymer matrix may include one or more polymers (e.g., polymers having different functional groups or repeat units). Polymers in the polymer matrix may be randomly arranged, such as in random copolymers, and/or have ordered structures, such as in block copolymers. Cross-linking can be via covalent, ionic, or inductive, interactions, or physical entanglement. The bead may be a macromolecule. The bead may be formed of nucleic acid molecules bound together. The bead may be formed via covalent or non-covalent assembly of molecules (e.g., macromolecules), such as monomers or polymers. Such polymers or monomers may be natural or synthetic. Such polymers or monomers may be or include, for example, nucleic acid molecules (e.g., DNA or RNA). The bead may be formed of a polymeric material. The bead may be magnetic or non-magnetic. The bead may be rigid. The bead may be flexible and/or compressible. The bead may be disruptable or dissolvable. The bead may be a solid particle (e.g., a metal-based particle including but not limited to iron oxide, gold or silver) covered with a coating comprising one or more polymers. Such coating may be disruptable or dissolvable.

[00101] The term “sample,” as used herein, generally refers to a biological sample of a subject. The biological sample may comprise any number of macromolecules, for example, cellular macromolecules. The sample may be a cell sample. The sample may be a cell line or cell culture sample. The sample can include one or more cells. The sample can include one or more microbes. The biological sample may be a nucleic acid sample or protein sample. The biological sample may also be a carbohydrate sample or a lipid sample. The biological sample may be derived from another sample. The sample may be a tissue sample, such as a biopsy, core

biopsy, needle aspirate, or fine needle aspirate. The sample may be a fluid sample, such as a blood sample, urine sample, or saliva sample. The sample may be a skin sample. The sample may be a cheek swab. The sample may be a plasma or serum sample. The sample may be a cell-free or cell free sample. A cell-free sample may include extracellular polynucleotides. Extracellular polynucleotides may be isolated from a bodily sample that may be selected from the group consisting of blood, plasma, serum, urine, saliva, mucosal excretions, sputum, stool and tears.

[00102] The term “biological particle,” as used herein, generally refers to a discrete biological system derived from a biological sample. The biological particle may be a macromolecule. The biological particle may be a small molecule. The biological particle may be a virus. The biological particle may be a cell or derivative of a cell. The biological particle may be an organelle. The biological particle may be a rare cell from a population of cells. The biological particle may be any type of cell, including without limitation prokaryotic cells, eukaryotic cells, bacterial, fungal, plant, mammalian, or other animal cell type, mycoplasmas, normal tissue cells, tumor cells, or any other cell type, whether derived from single cell or multicellular organisms. The biological particle may be a constituent of a cell. The biological particle may be or may include DNA, RNA, organelles, proteins, or any combination thereof. The biological particle may be or include a chromosome or other portion of a genome. The biological particle may be or may include a matrix (e.g., a gel or polymer matrix) comprising a cell or one or more constituents from a cell (e.g., cell bead), such as DNA, RNA, organelles, proteins, or any combination thereof, from the cell. The biological particle may be obtained from a tissue of a subject. The biological particle may be a hardened cell. Such hardened cell may or may not include a cell wall or cell membrane. The biological particle may include one or more constituents of a cell, but may not include other constituents of the cell. An example of such constituents is a nucleus or an organelle. A cell may be a live cell. The live cell may be capable of being cultured, for example, being cultured when enclosed in a gel or polymer matrix, or cultured when comprising a gel or polymer matrix.

[00103] The term “macromolecular constituent,” as used herein, generally refers to a macromolecule contained within or from a biological particle. The macromolecular constituent may comprise a nucleic acid. In some cases, the biological particle may be a macromolecule. The macromolecular constituent may comprise DNA. The macromolecular constituent may comprise RNA. The RNA may be coding or non-coding. The RNA may be messenger RNA (mRNA), ribosomal RNA (rRNA) or transfer RNA (tRNA), for example. The RNA may be a transcript. The RNA may be small RNA that are less than 200 nucleic acid bases in length, or large RNA that are greater than 200 nucleic acid bases in length. Small RNAs may include 5.8S

ribosomal RNA (rRNA), 5S rRNA, transfer RNA (tRNA), microRNA (miRNA), small interfering RNA (siRNA), small nucleolar RNA (snoRNAs), Piwi-interacting RNA (piRNA), tRNA-derived small RNA (tsRNA) and small rDNA-derived RNA (srRNA). The RNA may be double-stranded RNA or single-stranded RNA. The RNA may be circular RNA. The macromolecular constituent may comprise a protein. The macromolecular constituent may comprise a peptide. The macromolecular constituent may comprise a polypeptide.

[00104] The term “molecular tag,” as used herein, generally refers to a molecule capable of binding to a macromolecular constituent. The molecular tag may bind to the macromolecular constituent with high affinity. The molecular tag may bind to the macromolecular constituent with high specificity. The molecular tag may comprise a nucleotide sequence. The molecular tag may comprise a nucleic acid sequence. The nucleic acid sequence may be at least a portion or an entirety of the molecular tag. The molecular tag may be a nucleic acid molecule or may be part of a nucleic acid molecule. The molecular tag may be an oligonucleotide or a polypeptide. The molecular tag may comprise a DNA aptamer. The molecular tag may be or comprise a primer. The molecular tag may be, or comprise, a protein. The molecular tag may comprise a polypeptide. The molecular tag may be a barcode.

[00105] The term “partition,” as used herein, generally, refers to a space or volume that may be suitable to contain one or more species or conduct one or more reactions. A partition may be a physical compartment, such as a droplet or well. The partition may isolate space or volume from another space or volume. The droplet may be a first phase (e.g., aqueous phase) in a second phase (e.g., oil) immiscible with the first phase. The droplet may be a first phase in a second phase that does not phase separate from the first phase, such as, for example, a capsule or liposome in an aqueous phase. A partition may comprise one or more other (inner) partitions. In some cases, a partition may be a virtual compartment that can be defined and identified by an index (e.g., indexed libraries) across multiple and/or remote physical compartments. For example, a physical compartment may comprise a plurality of virtual compartments.

Single Cell Assay for Transposase Accessible Chromatin using Sequencing (ATAC-seq)

[00106] Disclosed herein, in some embodiments, are methods for nucleic acid processing. A method for nucleic acid processing may comprise generating a partition (e.g., a droplet or well) comprising: (i) a biological particle (e.g., a cell or nucleus) comprising chromatin comprising a template nucleic acid molecule; (ii) a plurality of nucleic acid barcode molecules comprising a common barcode sequence; (iii) a plurality of transposon end nucleic acid molecules comprising a transposon end sequence; and (iv) a plurality of transposase molecules. A plurality of template nucleic acid fragments may then be generated with the aid of a transposase-nucleic acid complex

comprising a transposase molecule of the plurality of transposase molecules and a transposon end oligonucleotide molecule of the plurality of transposon end oligonucleotide molecules. A barcoded nucleic acid fragment may then be generated using a nucleic acid barcode molecule of the plurality of nucleic acid barcode molecules and a template nucleic acid fragment of the plurality of template nucleic acid fragments.

[00107] The present disclosure also discloses a method of generating barcoded nucleic acid fragments, comprising providing: (i) a plurality of biological particles (e.g., cells or nuclei), an individual biological particle of the plurality of biological particles comprising chromatin comprising a template nucleic acid; (ii) a plurality of transposon end nucleic acid molecules comprising a transposon end sequence; and (iii) a plurality of transposase molecules. A plurality of template nucleic acid fragments may then be generated in a biological particle of the plurality of biological particles with the aid of a transposase-nucleic acid complex comprising a transposase molecule of the plurality of transposase molecules and a transposon end nucleic acid molecule of the plurality of transposon end nucleic acid molecules. A partition may then be generated, where the partition comprises the biological particle comprising the plurality of template nucleic acid fragments and a plurality of nucleic acid barcode molecules comprising a common barcode sequence. A barcoded nucleic acid fragment may then be generated using a nucleic acid barcode molecule of the plurality of nucleic acid barcode molecules and a template nucleic acid fragment of the plurality of template nucleic acid fragments.

[00108] In some embodiments, the transposase is a Tn5 transposase. In some embodiments, the transposase is a mutated, hyperactive Tn5 transposase. In some embodiments, the transposase is a Mu transposase. In some embodiments, the partitions disclosed in (a) further comprise cell lysis reagents; and/or (b) reagent and buffers necessary to carry out one or more reactions.

[00109] In some embodiments, a partition (e.g., a droplet or well) comprises a single cell and is processed according to the methods described herein. In some embodiments, a partition comprises a single cell and/or a single nucleus. The single cell and/or the single nucleus may be partitioned and processed according to the methods described herein. In some cases, the single nucleus may be a component of a cell. In some embodiments, a partition comprises chromatin from a single cell or single nucleus (e.g., a single chromosome or other portion of the genome) and is partitioned and processed according to the methods described herein. In some embodiments, the transposition reactions and methods described herein are performed in bulk and biological particles (e.g., nuclei/cells/chromatin from single cells) are then partitioned such that a plurality of partitions is singly occupied by a biological particle (e.g., a cell, cell nucleus, chromatin, or cell bead). For example, a plurality of biological particles may be partitioned into

a plurality of partitions such that partitions of the plurality of partitions comprise a single biological particle.

[00110] In some embodiments, the oligonucleotides described herein comprise a transposon end sequence. In some embodiments, the transposon end sequence is a Tn5 or modified Tn5 transposon end sequence. In some embodiments, the transposon end sequence is a Mu transposon end sequence. In some embodiments, the transposon end sequence has a sequence of:

AGATGTGTATAAGAGACA (SEQ ID NO: 1).

In some embodiments, the oligonucleotides described herein comprise an R1 sequencing priming region. In some embodiments, the R1 sequencing primer region has a sequence of

TCTACACTCTTTCCTACACGACGCTCTTCCGATCT (SEQ ID NO: 2),

or some portion thereof. In some embodiments, the R1 sequencing primer region has a sequence of

TCGTCGGCAGCGTCAGATGTGTATAAGAGACAG (SEQ ID NO: 3),

or some portion thereof. In some embodiments, the oligonucleotides described herein comprise a partial R1 sequence. In some embodiments, the partial R1 sequence is

ACTACACGACGCTCTTCCGATCT (SEQ ID NO: 4).

In some embodiments, the oligonucleotides described herein comprise an R2 sequencing priming region. In some embodiments, the R2 sequencing primer region has a sequence of

GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCT (SEQ ID NO: 5),

or some portion thereof. In some embodiments, the R2 sequencing primer region has a sequence of

GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAG (SEQ ID NO: 6),

or some portion thereof. In some embodiments, the oligonucleotides described herein comprise a T7 promoter sequence. In some embodiments, the T7 promoter sequence is

TAATACGACTCACTATAG (SEQ ID NO: 7).

In some embodiments, the oligonucleotides described herein comprise a region at least 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100% identical to any one of SEQ ID NO: 1-7. In some embodiments, the oligonucleotides described herein comprise a P5 sequence. In some embodiments, the oligonucleotides described herein comprise a P7 sequence. In some embodiments, the oligonucleotides described herein comprise a sample index sequence.

[00111] In some embodiments, the oligonucleotides described herein are attached to a solid support (e.g., a solid or semi-solid particle such as a bead). In some embodiments, the

oligonucleotides described herein are attached to a bead. In some embodiments, the bead is a gel bead. In some embodiments, the oligonucleotides described herein are releasably attached to a bead (e.g., a gel bead). In some embodiments, the oligonucleotides described herein are single-stranded and the first strand attached to a bead. In some embodiments, the oligonucleotides described herein are double-stranded or partially double-stranded molecules and the first strand is releasably attached to a bead. In some embodiments, the oligonucleotides described herein are double-stranded or partially double-stranded molecules and the second strand is releasably attached to a bead. In some embodiments, the oligonucleotides described herein are double-stranded or partially double-stranded molecules and both the first and the second strand are releasably attached to a bead or a collection of beads.

[00112] In some embodiments, the solid support (e.g., bead, such as a gel bead) comprises a plurality of first oligonucleotides and a plurality of second oligonucleotides. In some embodiments, the first oligonucleotides, the second oligonucleotides, or the combination thereof are releasably attached to a bead. In some embodiments, the first oligonucleotides, the second oligonucleotides, or the combination thereof are double-stranded or partially double-stranded molecules and the first strand is releasably attached to a bead. In some embodiments, the first oligonucleotides, the second oligonucleotides, or the combination thereof are double-stranded or partially double-stranded molecules and the second strand is releasably attached to a bead. In some embodiments, the first oligonucleotides, the second oligonucleotides, or the combination thereof are double-stranded or partially double-stranded molecules and the first strand and the second strand are releasably attached to a bead. In some embodiments, the oligonucleotides, the first oligonucleotides, the second oligonucleotides, or a combination thereof are bound to a magnetic particle. In some embodiments, the magnetic particle is embedded in a solid support (e.g., a bead, such as a gel bead).

[00113] In some embodiments, the first oligonucleotides are capable of coupling (e.g., by nucleic acid hybridization) to DNA molecules and the second oligonucleotides are capable of coupling (e.g., by nucleic acid hybridization) to RNA molecules (e.g., mRNA molecules). Examples of oligonucleotide architecture are provided in FIG. 35. In some embodiments, the first oligonucleotide comprises a P5 adaptor sequence, a barcode sequence, and an R1 sequence or partial R1 primer sequence. In some embodiments, the second oligonucleotide comprises a R1 sequence or partial R1 primer sequence, a barcode sequence, a unique molecular identifier (UMI) sequence, and a poly(dT) sequence. In some embodiments, the second oligonucleotide comprises a R1 sequence or partial R1 primer sequence, a barcode sequence, a unique molecular identifier (UMI) sequence, and a switch oligo.

[00114] In some embodiments, the first oligonucleotide comprises a P5 adaptor sequence, a barcode sequence, and an R1 sequence or partial R1 primer sequence and is partially double-stranded, and the second oligonucleotide comprises a R1 or partial R1 primer sequence, a barcode sequence, a unique molecular identifier (UMI) sequence, and a poly(dT) sequence. In some embodiments, the first oligonucleotide comprises a P5 adaptor sequence, a barcode sequence, and an R1 or partial R1 primer sequence and is single-stranded, and the second oligonucleotide comprises a R1 or partial R1 sequence, a barcode sequence, a unique molecular identifier (UMI) sequence, and a template switching oligo sequence.

Systems and Methods for Sample Compartmentalization

[00115] In an aspect, the systems and methods described herein provide for the compartmentalization, depositing, and/or partitioning of one or more particles (e.g., biological particles, macromolecular constituents of biological particles, beads, reagents, etc.) into discrete compartments or partitions (referred to interchangeably herein as partitions), where each partition maintains separation of its own contents from the contents of other partitions. The partition can be, for example, a well or a droplet in an emulsion. A partition may comprise one or more other partitions.

[00116] A partition may include one or more particles. A partition may include one or more types of particles. For example, a partition of the present disclosure may comprise one or more biological particles and/or macromolecular constituents thereof. For example, a partition may comprise one or more cells, nuclei, chromatins, and/or cell beads. For instance, a partition may comprise one or more cell beads. A cell bead can be a biological particle and/or one or more of its macromolecular constituents encased inside of a gel or polymer matrix, such as via polymerization of a droplet containing the biological particle and precursors capable of being polymerized or gelled. A partition may comprise one or more solid or semi-solid particles. For example, a partition may comprise one or more beads, such as one or more gel beads. In some cases, a partition may include a single gel bead, a single cell bead, or both a single cell bead and single gel bead. A partition may include one or more reagents (e.g., as described herein). Unique identifiers, such as barcodes, may be injected into the droplets previous to, subsequent to, or concurrently with droplet generation, such as via a microcapsule (e.g., bead), as described elsewhere herein. The barcodes may comprise nucleic acid sequences. Such nucleic acid barcode sequences may be components of nucleic acid barcode molecules, which may be coupled (e.g., releasably coupled) to a solid or semi-solid particle such as a bead. In some cases, a partition may be unoccupied. For example, a partition may not comprise a bead or a biological particle. Microfluidic devices comprising one or more channels (e.g., microfluidic

channel networks of a chip) can be utilized to generate partitions as described herein. For example, a first fluid and a second fluid that is immiscible with the first fluid may be flowed to a droplet generation junction, and a droplet of the first fluid may be generated within the second fluid. Alternative mechanisms may also be employed in the partitioning of individual biological particles, including porous membranes through which aqueous mixtures of cells are extruded into non-aqueous fluids.

[00117] The partitions can be flowable within fluid streams. The partitions may comprise, for example, micro-vesicles that have an outer barrier surrounding an inner fluid center or core. In some cases, the partitions may comprise a porous matrix that is capable of entraining and/or retaining materials within its matrix. The partitions can be droplets of a first phase within a second phase, wherein the first and second phases are immiscible. For example, the partitions can be droplets of aqueous fluid within a non-aqueous continuous phase (e.g., oil phase). In another example, the partitions can be droplets of a non-aqueous fluid within an aqueous phase. In some examples, the partitions may be provided in a water-in-oil emulsion or oil-in-water emulsion. A variety of different vessels are described in, for example, U.S. Patent Application Publication No. 2014/0155295, which is entirely incorporated herein by reference for all purposes. Emulsion systems for creating stable droplets in non-aqueous or oil continuous phases are described in, for example, U.S. Patent Application Publication No. 2010/0105112, which is entirely incorporated herein by reference for all purposes.

[00118] In the case of droplets in an emulsion, allocating individual particles to discrete partitions may in one non-limiting example be accomplished by introducing a flowing stream of particles in an aqueous fluid into a flowing stream of a non-aqueous fluid, such that droplets are generated at the junction of the two streams. Fluid properties (e.g., fluid flow rates, fluid viscosities, etc.), particle properties (e.g., volume fraction, particle size, particle concentration, etc.), microfluidic architectures (e.g., channel geometry, etc.), and other parameters may be adjusted to control the occupancy of the resulting partitions (e.g., number of biological particles per partition, number of beads per partition, etc.). For example, partition occupancy can be controlled by providing the aqueous stream at a certain concentration and/or flow rate of particles. To generate single biological particle partitions, the relative flow rates of the immiscible fluids can be selected such that, on average, the partitions may contain less than one biological particle per partition in order to ensure that those partitions that are occupied are primarily singly occupied. In some cases, partitions among a plurality of partitions may contain at most one biological particle (e.g., cell bead, chromatin, DNA, cell, cell nucleus, or other cellular material). In some embodiments, the various parameters (e.g., fluid properties, particle properties, microfluidic architectures, etc.) may be selected or adjusted such that a majority of

partitions are occupied, for example, allowing for only a small percentage of unoccupied partitions. The flows and channel architectures can be controlled as to ensure a given number of singly occupied partitions, less than a certain level of unoccupied partitions and/or less than a certain level of multiply occupied partitions.

[00119] FIG. 1 shows an example of a microfluidic channel structure **100** for partitioning individual biological particles (e.g., as described herein). The channel structure **100** can include channel segments **102**, **104**, **106** and **108** communicating at a channel junction **110**. In operation, a first aqueous fluid **112** that includes suspended biological particles (or cells) **114** may be transported along channel segment **102** into junction **110**, while a second fluid **116** that is immiscible with the aqueous fluid **112** is delivered to the junction **110** from each of channel segments **104** and **106** to create discrete droplets **118**, **120** of the first aqueous fluid **112** flowing into channel segment **108**, and flowing away from junction **110**. The channel segment **108** may be fluidically coupled to an outlet reservoir where the discrete droplets can be stored and/or harvested. A discrete droplet generated may include an individual biological particle **114** (such as droplets **118**). A discrete droplet generated may include more than one individual biological particle **114** (not shown in FIG. 1). A discrete droplet may contain no biological particle **114** (such as droplet **120**). Each discrete partition may maintain separation of its own contents (e.g., individual biological particle **114**) from the contents of other partitions.

[00120] The second fluid **116** can comprise an oil, such as a fluorinated oil, that includes a fluorosurfactant for stabilizing the resulting droplets, for example, inhibiting subsequent coalescence of the resulting droplets **118**, **120**. Examples of particularly useful partitioning fluids and fluorosurfactants are described, for example, in U.S. Patent Application Publication No. 2010/0105112, which is entirely incorporated herein by reference for all purposes.

[00121] As will be appreciated, the channel segments described herein may be coupled to any of a variety of different fluid sources or receiving components, including reservoirs, tubing, manifolds, or fluidic components of other systems. As will be appreciated, the microfluidic channel structure **100** may have other geometries. For example, a microfluidic channel structure can have more than one channel junction. For example, a microfluidic channel structure can have 2, 3, 4, or 5 channel segments each carrying particles (e.g., biological particles, cell beads, and/or solid or semi-solid particles such as gel beads) that meet at a channel junction. Fluid may be directed to flow along one or more channels or reservoirs via one or more fluid flow units. A fluid flow unit can comprise compressors (e.g., providing positive pressure), pumps (e.g., providing negative pressure), actuators, and the like to control flow of the fluid. Fluid may also or otherwise be controlled via applied pressure differentials, centrifugal force, electrokinetic pumping, vacuum, capillary or gravity flow, or the like.

[00122] The generated droplets may comprise two subsets of droplets: (1) occupied droplets **118**, containing one or more biological particles **114**, and (2) unoccupied droplets **120**, not containing any biological particles **114**. Occupied droplets **118** may comprise singly occupied droplets (having one biological particle) and multiply occupied droplets (having more than one biological particle). As described elsewhere herein, in some cases, the majority of occupied partitions can include no more than one biological particle per occupied partition and some of the generated partitions can be unoccupied (of any biological particle). In some cases, though, some of the occupied partitions may include more than one biological particle. In some cases, the partitioning process may be controlled such that fewer than about 25% of the occupied partitions contain more than one biological particle, and in many cases, fewer than about 20% of the occupied partitions have more than one biological particle, while in some cases, fewer than about 10% or even fewer than about 5% of the occupied partitions include more than one biological particle per partition.

[00123] In some cases, it may be desirable to minimize the creation of excessive numbers of empty partitions, such as to reduce costs and/or increase efficiency. While this minimization may be achieved by providing a sufficient number of biological particles (e.g., biological particles **114**) at the partitioning junction **110**, such as to ensure that at least one biological particle is encapsulated in a partition, the Poissonian distribution may expectedly increase the number of partitions that include multiple biological particles. As such, where singly occupied partitions are to be obtained, at most about 95%, 90%, 85%, 80%, 75%, 70%, 65%, 60%, 55%, 50%, 45%, 40%, 35%, 30%, 25%, 20%, 15%, 10%, 5% or less of the generated partitions can be unoccupied.

[00124] In some cases, the flow of one or more of the biological particles (e.g., in channel segment **102**), or other fluids directed into the partitioning junction (e.g., in channel segments **104**, **106**) can be controlled such that, in many cases, no more than about 50% of the generated partitions, no more than about 25% of the generated partitions, or no more than about 10% of the generated partitions are unoccupied. These flows can be controlled so as to present a non-Poissonian distribution of singly-occupied partitions while providing lower levels of unoccupied partitions. The above noted ranges of unoccupied partitions can be achieved while still providing any of the single occupancy rates described above. For example, in many cases, the use of the systems and methods described herein may result in partitions that have multiple occupancy rates of less than about 25%, less than about 20%, less than about 15%, less than about 10%, and in many cases, less than about 5%, while having unoccupied partitions of less than about 50%, less than about 40%, less than about 30%, less than about 20%, less than about 10%, less than about 5%, or less.

[00125] As will be appreciated, the above-described occupancy rates are also applicable to partitions that include both biological particles and additional reagents, including, but not limited to, microcapsules or beads (e.g., gel beads) carrying barcoded nucleic acid molecules (e.g., oligonucleotides) (described in relation to **FIG. 2**). The occupied partitions (e.g., at least about 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 95%, or 99% of the occupied partitions) can include both a microcapsule (e.g., bead) comprising barcoded nucleic acid molecules and a biological particle.

[00126] In another aspect, in addition to or as an alternative to droplet based partitioning, biological particles (e.g., as described herein) may be encapsulated within a microcapsule that comprises an outer shell, layer or porous matrix in which is entrained one or more individual biological particles or small groups of biological particles. The microcapsule may include other reagents. Encapsulation of biological particles may be performed by a variety of processes. Such processes may combine an aqueous fluid containing the biological particles with a polymeric precursor material that may be capable of being formed into a gel or other solid or semi-solid matrix upon application of a particular stimulus to the polymer precursor. Such stimuli can include, for example, thermal stimuli (e.g., either heating or cooling), photo-stimuli (e.g., through photo-curing), chemical stimuli (e.g., through crosslinking, polymerization initiation of the precursor (e.g., through added initiators)), mechanical stimuli, or a combination thereof.

[00127] Preparation of microcapsules comprising biological particles may be performed by a variety of methods. For example, air knife droplet or aerosol generators may be used to dispense droplets of precursor fluids into gelling solutions in order to form microcapsules that include individual biological particles or small groups of biological particles. Likewise, membrane based encapsulation systems may be used to generate microcapsules comprising encapsulated biological particles as described herein. Microfluidic systems of the present disclosure, such as that shown in **FIG. 1**, may be readily used in encapsulating cells as described herein. In particular, and with reference to **FIG. 1**, the aqueous fluid **112** comprising (i) the biological particles **114** and (ii) the polymer precursor material (not shown) is flowed into channel junction **110**, where it is partitioned into droplets **118**, **120** through the flow of non-aqueous fluid **116**. In the case of encapsulation methods, non-aqueous fluid **116** may also include an initiator (not shown) to cause polymerization and/or crosslinking of the polymer precursor to form the microcapsule that includes the entrained biological particles. Examples of polymer precursor/initiator pairs include those described in U.S. Patent Application Publication No. 2014/0378345, which is entirely incorporated herein by reference for all purposes.

[00128] For example, in the case where the polymer precursor material comprises a linear polymer material, such as a linear polyacrylamide, PEG, or other linear polymeric material, the

activation agent may comprise a cross-linking agent, or a chemical that activates a cross-linking agent within the formed droplets. Likewise, for polymer precursors that comprise polymerizable monomers, the activation agent may comprise a polymerization initiator. For example, in certain cases, where the polymer precursor comprises a mixture of acrylamide monomer with a N,N'-bis-(acryloyl)cystamine (BAC) comonomer, an agent such as tetraethylmethylenediamine (TEMED) may be provided within the second fluid streams **116** in channel segments **104** and **106**, which can initiate the copolymerization of the acrylamide and BAC into a cross-linked polymer network, or hydrogel.

[00129] Upon contact of the second fluid stream **116** with the first fluid stream **112** at junction **110**, during formation of droplets, the TEMED may diffuse from the second fluid **116** into the aqueous fluid **112** comprising the linear polyacrylamide, which will activate the crosslinking of the polyacrylamide within the droplets **118**, **120**, resulting in the formation of gel (e.g., hydrogel) microcapsules, as solid or semi-solid beads or particles entraining the biological particles (e.g., cells) **114**. Although described in terms of polyacrylamide encapsulation, other 'activatable' encapsulation compositions may also be employed in the context of the methods and compositions described herein. For example, formation of alginate droplets followed by exposure to divalent metal ions (e.g., Ca^{2+} ions), can be used as an encapsulation process using the described processes. Likewise, agarose droplets may also be transformed into capsules through temperature based gelling (e.g., upon cooling, etc.).

[00130] In some cases, encapsulated biological particles can be selectively releasable from the microcapsule, such as through passage of time or upon application of a particular stimulus, that degrades the microcapsule sufficiently to allow the biological particles (e.g., cell), or its other contents to be released from the microcapsule, such as into a partition (e.g., droplet). For example, in the case of the polyacrylamide polymer described above, degradation of the microcapsule may be accomplished through the introduction of an appropriate reducing agent, such as dithiothreitol (DTT) or the like, to cleave disulfide bonds that cross-link the polymer matrix. See, for example, U.S. Patent Application Publication No. 2014/0378345, which is entirely incorporated herein by reference for all purposes.

[00131] The biological particle can be subjected to other conditions sufficient to polymerize or gel the precursors. The conditions sufficient to polymerize or gel the precursors may comprise exposure to heating, cooling, electromagnetic radiation, and/or light. The conditions sufficient to polymerize or gel the precursors may comprise any conditions sufficient to polymerize or gel the precursors. Following polymerization or gelling, a polymer or gel may be formed around and/or within the biological particle. The polymer or gel may be diffusively permeable to chemical or biochemical reagents. The polymer or gel may be diffusively

impermeable to macromolecular constituents of the biological particle. In this manner, the polymer or gel may act to allow the biological particle to be subjected to chemical or biochemical operations while spatially confining the macromolecular constituents to a region of the droplet defined by the polymer or gel. The polymer or gel may include one or more of disulfide cross-linked polyacrylamide, agarose, alginate, polyvinyl alcohol, polyethylene glycol (PEG)-diacrylate, PEG-acrylate, PEG-thiol, PEG-azide, PEG-alkyne, other acrylates, chitosan, hyaluronic acid, collagen, fibrin, gelatin, or elastin. The polymer or gel may comprise any other polymer or gel. A cell comprising a polymer or gel matrix may be referred to as a “cell bead.”

[00132] The polymer or gel may be functionalized to bind to targeted analytes, such as nucleic acids, proteins, carbohydrates, lipids or other analytes. The polymer or gel may be polymerized or gelled via a passive mechanism. For example, polymerization or gelling may be brought about by a change in salinity, pH, temperature, or pressure. The polymer or gel may be stable in alkaline conditions or at elevated temperature. The polymer or gel may have mechanical properties similar to the mechanical properties of the bead. For instance, a polymer or gel matrix may be of a similar size to the bead. The polymer or gel may have a mechanical strength (e.g. tensile strength) similar to that of the bead. The polymer or gel may be of a lower density than an oil. The polymer or gel may be of a density that is roughly similar to that of a buffer. The polymer or gel may have a tunable pore size. The pore size may be chosen to, for instance, retain denatured nucleic acids. The pore size may be chosen to maintain diffusive permeability to exogenous chemicals such as sodium hydroxide (NaOH) and/or endogenous chemicals such as inhibitors. The polymer or gel may be biocompatible. The polymer or gel may maintain or enhance cell viability. The polymer or gel may be biochemically compatible. The polymer or gel may be polymerized and/or depolymerized thermally, chemically, enzymatically, and/or optically.

[00133] The polymer or gel may comprise poly(acrylamide-co-acrylic acid) crosslinked with disulfide linkages. The preparation of the polymer or gel may comprise a two-step reaction. In the first activation step, poly(acrylamide-co-acrylic acid) may be exposed to an acylating agent to convert carboxylic acids to esters. For instance, the poly(acrylamide-co-acrylic acid) may be exposed to 4-(4,6-dimethoxy-1,3,5-triazin-2-yl)-4-methylmorpholinium chloride (DMTMM). The polyacrylamide-co-acrylic acid may be exposed to other salts of 4-(4,6-dimethoxy-1,3,5-triazin-2-yl)-4-methylmorpholinium. In the second cross-linking step, the ester formed in the first step may be exposed to a disulfide crosslinking agent. For instance, the ester may be exposed to cystamine (2,2'-dithiobis(ethylamine)). Following the two steps, the biological particle may be surrounded by polyacrylamide strands linked together by disulfide bridges. In this manner, the biological particle may be encased inside of or comprise a gel or matrix (e.g.,

polymer matrix) to form a “cell bead.” A cell bead can contain biological particles (e.g., a cell) or macromolecular constituents (e.g., RNA, DNA, proteins, etc.) of biological particles. A cell bead may include a single cell or multiple cells, or a derivative of the single cell or multiple cells. For example after lysing and washing the cells, inhibitory components from cell lysates can be washed away and the macromolecular constituents can be bound as cell beads. Systems and methods disclosed herein can be applicable to both cell beads (and/or droplets or other partitions) containing biological particles and cell beads (and/or droplets or other partitions) containing macromolecular constituents of biological particles.

[00134] Encapsulated biological particles can provide certain potential advantages of being more storable and more portable than droplet-based partitioned biological particles.

Furthermore, in some cases, it may be desirable to allow biological particles to incubate for a select period of time before analysis, such as in order to characterize changes in such biological particles over time, either in the presence or absence of different stimuli. In such cases, encapsulation may allow for longer incubation than partitioning in emulsion droplets, although in some cases, droplet partitioned biological particles may also be incubated for different periods of time, e.g., at least 10 seconds, at least 30 seconds, at least 1 minute, at least 5 minutes, at least 10 minutes, at least 30 minutes, at least 1 hour, at least 2 hours, at least 5 hours, or at least 10 hours or more. The encapsulation of biological particles may constitute the partitioning of the biological particles into which other reagents are co-partitioned. Alternatively or in addition, encapsulated biological particles may be readily deposited into other partitions (e.g., droplets) as described above.

Beads

[00135] A partition may comprise one or more unique identifiers, such as one or more barcodes. Barcodes may be previously, subsequently, or concurrently delivered to the partitions that hold the compartmentalized or partitioned biological particle. For example, barcodes may be injected into droplets previous to, subsequent to, or concurrently with droplet generation. The delivery of the barcodes to a particular partition allows for the later attribution of the characteristics of the individual biological particle to the particular partition. Barcodes may be delivered, for example, as components of a nucleic acid molecule (e.g., an oligonucleotide). Barcodes may be delivered to a partition via any suitable mechanism. For example, barcoded nucleic acid molecules can be delivered to a partition such as a droplet or well via a microcapsule or bead. A microcapsule, in some instances, can comprise a bead. Beads are described in further detail below.

[00136] In some cases, barcoded nucleic acid molecules can be initially associated with the microcapsule or bead and then released from the microcapsule or bead. For example, a bead may comprise a plurality of nucleic acid barcode molecules releasably coupled thereto. Release of the barcoded nucleic acid molecules can be passive (e.g., by diffusion out of the microcapsule or bead). In addition or alternatively, release from the microcapsule or bead can be upon application of a stimulus which allows the barcoded nucleic acid nucleic acid molecules to dissociate or to be released from the microcapsule or bead. Such stimulus may disrupt the microcapsule or bead, an interaction that couples the barcoded nucleic acid molecules to or within the microcapsule or bead, or both. Such stimulus can include, for example, a thermal stimulus, photo-stimulus, chemical stimulus (e.g., change in pH or use of a reducing agent(s)), a mechanical stimulus, a radiation stimulus; a biological stimulus (e.g., enzyme), or any combination thereof.

[00137] FIG. 2 shows an example of a microfluidic channel structure **200** for delivering barcode carrying beads to droplets. The channel structure **200** can include channel segments **201**, **202**, **204**, **206** and **208** communicating at a channel junction **210**. In operation, the channel segment **201** may transport an aqueous fluid **212** that includes a plurality of beads **214** (e.g., beads comprising nucleic acid barcode molecules, oligonucleotides, or molecular tags) along the channel segment **201** into junction **210**. The plurality of beads **214** may be sourced from a suspension of beads. For example, the channel segment **201** may be connected to a reservoir comprising an aqueous suspension of beads **214**. The channel segment **202** may transport the aqueous fluid **212** that includes a plurality of biological particles **216** along the channel segment **202** into junction **210**. The plurality of biological particles **216** may be sourced from a suspension of biological particles. For example, the channel segment **202** may be connected to a reservoir comprising an aqueous suspension of biological particles **216**. In some instances, the aqueous fluid **212** in either the first channel segment **201** or the second channel segment **202**, or in both segments, can include one or more reagents, as further described below. A second fluid **218** that is immiscible with the aqueous fluid **212** (e.g., oil) can be delivered to the junction **210** from each of channel segments **204** and **206**. Upon meeting of the aqueous fluid **212** from each of channel segments **201** and **202** and the second fluid **218** from each of channel segments **204** and **206** at the channel junction **210**, the aqueous fluid **212** can be partitioned as discrete droplets **220** in the second fluid **218** and flow away from the junction **210** along channel segment **208**. The channel segment **208** may deliver the discrete droplets to an outlet reservoir fluidly coupled to the channel segment **208**, where they may be harvested.

[00138] As an alternative, the channel segments **201** and **202** may meet at another junction upstream of the junction **210**. At such junction, beads and biological particles may form a

mixture that is directed along another channel to the junction **210** to yield droplets **220**. The mixture may provide the beads and biological particles in an alternating fashion, such that, for example, a droplet comprises a single bead and a single biological particle.

[00139] Beads, biological particles, and droplets (e.g., droplets comprising beads and/or biological particles) may flow along channels at substantially regular flow profiles (e.g., at regular flow rates). Such regular flow profiles may permit a droplet to be formed such that it includes a single bead and a single biological particle. Such regular flow profiles may permit the droplets to have an occupancy (e.g., droplets having beads and biological particles) greater than 5%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, or 95%. Such regular flow profiles and devices that may be used to provide such regular flow profiles are provided in, for example, U.S. Patent Publication No. 2015/0292988, which is entirely incorporated herein by reference.

[00140] The second fluid **218** can comprise an oil, such as a fluorinated oil, that includes a fluorosurfactant for stabilizing the resulting droplets, for example, inhibiting subsequent coalescence of the resulting droplets **220**.

[00141] A discrete droplet that is generated may include an individual biological particle **216**. A discrete droplet that is generated may include a bead **214** comprising a barcode or other reagent. For example, bead **214** may comprise a plurality of nucleic acid barcode molecules each comprising a common barcode sequence. A discrete droplet generated may include both an individual biological particle and a barcode-carrying bead, such as droplets **220**. In some instances, a discrete droplet may include more than one individual biological particle or no biological particle. In some instances, a discrete droplet may include more than one bead or no bead. A discrete droplet may be unoccupied (e.g., no beads, no biological particles).

[00142] Beneficially, a discrete droplet partitioning a biological particle and a barcode carrying bead may effectively allow the attribution of the barcode to macromolecular constituents of the biological particle within the partition. The contents of a partition may remain discrete from the contents of other partitions.

[00143] As will be appreciated, the channel segments described herein may be coupled to any of a variety of different fluid sources or receiving components, including reservoirs, tubing, manifolds, or fluidic components of other systems. As will be appreciated, the microfluidic channel structure **200** may have other geometries. For example, a microfluidic channel structure can have more than one channel junctions. For example, a microfluidic channel structure can have 2, 3, 4, or 5 channel segments each carrying beads that meet at a channel junction. Fluid may be directed flow along one or more channels or reservoirs via one or more fluid flow units. A fluid flow unit can comprise compressors (e.g., providing positive pressure), pumps (e.g., providing negative pressure), actuators, and the like to control flow of the fluid. Fluid may also

or otherwise be controlled via applied pressure differentials, centrifugal force, electrokinetic pumping, vacuum, capillary or gravity flow, or the like.

[00144] A bead may be porous, non-porous, solid, semi-solid, semi-fluidic, fluidic, and/or a combination thereof. In some instances, a bead may be dissolvable, disruptable, and/or degradable. In some cases, a bead may not be degradable. In some cases, the bead may be a gel bead. A gel bead may be a hydrogel bead. A gel bead may be formed from molecular precursors, such as a polymeric or monomeric species. A semi-solid bead may be a liposomal bead. Solid beads may comprise metals including iron oxide, gold, and silver. In some cases, the bead may be a silica bead. In some cases, the bead can be rigid. In other cases, the bead may be flexible and/or compressible.

[00145] A bead may be of any suitable shape. Examples of bead shapes include, but are not limited to, spherical, non-spherical, oval, oblong, amorphous, circular, cylindrical, and variations thereof.

[00146] Beads may be of uniform size or heterogeneous size. In some cases, the diameter of a bead may be at least about 10 nanometers (nm), 100 nm, 500 nm, 1 micrometer (μm), 5 μm , 10 μm , 20 μm , 30 μm , 40 μm , 50 μm , 60 μm , 70 μm , 80 μm , 90 μm , 100 μm , 250 μm , 500 μm , 1 mm, or greater. In some cases, a bead may have a diameter of less than about 10 nm, 100 nm, 500 nm, 1 μm , 5 μm , 10 μm , 20 μm , 30 μm , 40 μm , 50 μm , 60 μm , 70 μm , 80 μm , 90 μm , 100 μm , 250 μm , 500 μm , 1 mm, or less. In some cases, a bead may have a diameter in the range of about 40-75 μm , 30-75 μm , 20-75 μm , 40-85 μm , 40-95 μm , 20-100 μm , 10-100 μm , 1-100 μm , 20-250 μm , or 20-500 μm .

[00147] In certain aspects, beads can be provided as a population or plurality of beads having a relatively monodisperse size distribution. Where it may be desirable to provide relatively consistent amounts of reagents within partitions, maintaining relatively consistent bead characteristics, such as size, can contribute to the overall consistency. In particular, the beads described herein may have size distributions that have a coefficient of variation in their cross-sectional dimensions of less than 50%, less than 40%, less than 30%, less than 20%, and in some cases less than 15%, less than 10%, less than 5%, or less.

[00148] A bead may comprise natural and/or synthetic materials. For example, a bead can comprise a natural polymer, a synthetic polymer or both natural and synthetic polymers. Examples of natural polymers include proteins and sugars such as deoxyribonucleic acid, rubber, cellulose, starch (e.g., amylose, amylopectin), proteins, enzymes, polysaccharides, silks, polyhydroxyalkanoates, chitosan, dextran, collagen, carrageenan, ispaghula, acacia, agar, gelatin, shellac, sterculia gum, xanthan gum, Corn sugar gum, guar gum, gum karaya, agarose, alginic acid, alginate, or natural polymers thereof. Examples of synthetic polymers include acrylics,

nylons, silicones, spandex, viscose rayon, polycarboxylic acids, polyvinyl acetate, polyacrylamide, polyacrylate, polyethylene glycol, polyurethanes, polylactic acid, silica, polystyrene, polyacrylonitrile, polybutadiene, polycarbonate, polyethylene, polyethylene terephthalate, poly(chlorotrifluoroethylene), poly(ethylene oxide), poly(ethylene terephthalate), polyethylene, polyisobutylene, poly(methyl methacrylate), poly(oxymethylene), polyformaldehyde, polypropylene, polystyrene, poly(tetrafluoroethylene), poly(vinyl acetate), poly(vinyl alcohol), poly(vinyl chloride), poly(vinylidene dichloride), poly(vinylidene difluoride), poly(vinyl fluoride) and/or combinations (e.g., co-polymers) thereof. Beads may also be formed from materials other than polymers, including lipids, micelles, ceramics, glass-ceramics, material composites, metals, other inorganic materials, and others.

[00149] In some instances, the bead may contain molecular precursors (e.g., monomers or polymers), which may form a polymer network via polymerization of the molecular precursors. In some cases, a precursor may be an already polymerized species capable of undergoing further polymerization via, for example, a chemical cross-linkage. In some cases, a precursor can comprise one or more of an acrylamide or a methacrylamide monomer, oligomer, or polymer. In some cases, the bead may comprise prepolymers, which are oligomers capable of further polymerization. For example, polyurethane beads may be prepared using prepolymers. In some cases, the bead may contain individual polymers that may be further polymerized together. In some cases, beads may be generated via polymerization of different precursors, such that they comprise mixed polymers, co-polymers, and/or block co-polymers. In some cases, the bead may comprise covalent or ionic bonds between polymeric precursors (e.g., monomers, oligomers, linear polymers), nucleic acid molecules (e.g., oligonucleotides), primers, and other entities. In some cases, the covalent bonds can be carbon-carbon bonds, thioether bonds, or carbon-heteroatom bonds.

[00150] Cross-linking may be permanent or reversible, depending upon the particular cross-linker used. Reversible cross-linking may allow for the polymer to linearize or dissociate under appropriate conditions. In some cases, reversible cross-linking may also allow for reversible attachment of a material bound to the surface of a bead. In some cases, a cross-linker may form disulfide linkages. In some cases, the chemical cross-linker forming disulfide linkages may be cystamine or a modified cystamine.

[00151] In some cases, disulfide linkages can be formed between molecular precursor units (e.g., monomers, oligomers, or linear polymers) or precursors incorporated into a bead and nucleic acid molecules (e.g., oligonucleotides). Cystamine (including modified cystamines), for example, is an organic agent comprising a disulfide bond that may be used as a crosslinker agent between individual monomeric or polymeric precursors of a bead. Polyacrylamide may be

polymerized in the presence of cystamine or a species comprising cystamine (e.g., a modified cystamine) to generate polyacrylamide gel beads comprising disulfide linkages (e.g., chemically degradable beads comprising chemically-reducible cross-linkers). The disulfide linkages may permit the bead to be degraded (or dissolved) upon exposure of the bead to a reducing agent.

[00152] In some cases, chitosan, a linear polysaccharide polymer, may be crosslinked with glutaraldehyde via hydrophilic chains to form a bead. Crosslinking of chitosan polymers may be achieved by chemical reactions that are initiated by heat, pressure, change in pH, and/or radiation.

[00153] In some cases, a bead may comprise an acrydite moiety, which in certain aspects may be used to attach one or more nucleic acid molecules (e.g., barcode sequence, barcoded nucleic acid molecule, barcoded oligonucleotide, primer, or other oligonucleotide) to the bead. In some cases, an acrydite moiety can refer to an acrydite analogue generated from the reaction of acrydite with one or more species, such as, the reaction of acrydite with other monomers and cross-linkers during a polymerization reaction. Acrydite moieties may be modified to form chemical bonds with a species to be attached, such as a nucleic acid molecule (e.g., barcode sequence, barcoded nucleic acid molecule, barcoded oligonucleotide, primer, or other oligonucleotide). Acrydite moieties may be modified with thiol groups capable of forming a disulfide bond or may be modified with groups already comprising a disulfide bond. The thiol or disulfide (via disulfide exchange) may be used as an anchor point for a species to be attached or another part of the acrydite moiety may be used for attachment. In some cases, attachment can be reversible, such that when the disulfide bond is broken (e.g., in the presence of a reducing agent), the attached species is released from the bead. In other cases, an acrydite moiety can comprise a reactive hydroxyl group that may be used for attachment.

[00154] Functionalization of beads for attachment of nucleic acid molecules (e.g., oligonucleotides) may be achieved through a wide range of different approaches, including activation of chemical groups within a polymer, incorporation of active or activatable functional groups in the polymer structure, or attachment at the pre-polymer or monomer stage in bead production.

[00155] For example, precursors (e.g., monomers, cross-linkers) that are polymerized to form a bead may comprise acrydite moieties, such that when a bead is generated, the bead also comprises acrydite moieties. The acrydite moieties can be attached to a nucleic acid molecule (e.g., oligonucleotide), which may include a priming sequence (e.g., a primer for amplifying target nucleic acids, random primer, primer sequence for messenger RNA) and/or one or more barcode sequences. The one more barcode sequences may include sequences that are the same for all nucleic acid molecules (e.g., nucleic acid barcode molecules) coupled to a given bead

and/or sequences that are different across all nucleic acid molecules coupled to the given bead. The nucleic acid molecule may be incorporated into the bead and/or may be attached to a surface of the bead.

[00156] In some cases, the nucleic acid molecule (e.g., nucleic acid barcode molecule) can comprise a functional sequence, for example, for attachment to a sequencing flow cell, such as, for example, a P5 sequence for Illumina® sequencing. In some cases, the nucleic acid molecule (e.g., nucleic acid barcode molecule) or derivative thereof (e.g., oligonucleotide or polynucleotide generated from the nucleic acid molecule) can comprise another functional sequence, such as, for example, a P7 sequence for attachment to a sequencing flow cell for Illumina sequencing. In some cases, the nucleic acid molecule can comprise a barcode sequence. In some cases, the primer can further comprise a unique molecular identifier (UMI). In some cases, the primer can comprise an R1 primer sequence for Illumina sequencing. In some cases, the primer can comprise an R2 primer sequence for Illumina sequencing. Examples of such nucleic acid molecules (e.g., oligonucleotides, polynucleotides, etc.) and uses thereof, as may be used with compositions, devices, methods and systems of the present disclosure, are provided in U.S. Patent Pub. Nos. 2014/0378345 and 2015/0376609, each of which is entirely incorporated herein by reference.

[00157] **FIG. 8** illustrates an example of a barcode carrying bead. A nucleic acid molecule **802**, such as an oligonucleotide, can be coupled to a bead **804** by a releasable linkage **806**, such as, for example, a disulfide linker. The same bead **804** may be coupled (e.g., via releasable linkage) to one or more other nucleic acid molecules **818**, **820**. The nucleic acid molecule **802** may be or comprise a barcode. As noted elsewhere herein, the structure of the barcode may comprise a number of sequence elements. The nucleic acid molecule **802** may comprise a functional sequence **808** that may be used in subsequent processing. For example, the functional sequence **808** may include one or more of a sequencer specific flow cell attachment sequence (e.g., a P5 sequence for Illumina® sequencing systems) and a sequencing primer sequence (e.g., a R1 primer for Illumina® sequencing systems). The nucleic acid molecule **802** may comprise a barcode sequence **810** for use in barcoding the sample (e.g., DNA, RNA, protein, etc.). In some cases, the barcode sequence **810** can be bead-specific such that the barcode sequence **810** is common to all nucleic acid molecules (e.g., including nucleic acid molecule **802**) coupled to the same bead **804**. Alternatively or in addition, the barcode sequence **810** can be partition-specific such that the barcode sequence **810** is common to all nucleic acid molecules coupled to one or more beads that are partitioned into the same partition. The nucleic acid molecule **802** may comprise a specific priming sequence **812**, such as an mRNA specific priming sequence (e.g., poly-T sequence), a targeted priming sequence, and/or a random priming sequence. The nucleic

acid molecule **802** may comprise an anchoring sequence **814** to ensure that the specific priming sequence **812** hybridizes at the sequence end (e.g., of the mRNA). For example, the anchoring sequence **814** can include a random short sequence of nucleotides, such as a 1-mer, 2-mer, 3-mer or longer sequence, which can ensure that a poly-T segment is more likely to hybridize at the sequence end of the poly-A tail of the mRNA.

[00158] The nucleic acid molecule **802** may comprise a unique molecular identifying sequence **816** (e.g., unique molecular identifier (UMI)). In some cases, the unique molecular identifying sequence **816** may comprise from about 5 to about 8 nucleotides. Alternatively, the unique molecular identifying sequence **816** may comprise less than about 5 or more than about 8 nucleotides. The unique molecular identifying sequence **816** may be a unique sequence that varies across individual nucleic acid molecules (e.g., **802**, **818**, **820**, etc.) coupled to a single bead (e.g., bead **804**). In some cases, the unique molecular identifying sequence **816** may be a random sequence (e.g., such as a random N-mer sequence). For example, the UMI may provide a unique identifier of the starting mRNA molecule that was captured, in order to allow quantitation of the number of original expressed RNA. As will be appreciated, although **FIG. 8** shows three nucleic acid molecules **802**, **818**, **820** coupled to the surface of the bead **804**, an individual bead may be coupled to any number of individual nucleic acid molecules, for example, from one to tens to hundreds of thousands or even millions of individual nucleic acid molecules. The respective barcodes for the individual nucleic acid molecules can comprise both common sequence segments or relatively common sequence segments (e.g., **808**, **810**, **812**, etc.) and variable or unique sequence segments (e.g., **816**) between different individual nucleic acid molecules coupled to the same bead.

[00159] In operation, a biological particle (e.g., cell, DNA, RNA, etc.) can be co-partitioned along with a barcode bearing bead **804**. The barcoded nucleic acid molecules **802**, **818**, **820** can be released from the bead **804** in the partition. By way of example, in the context of analyzing sample RNA, the poly-T segment (e.g., **812**) of one of the released nucleic acid molecules (e.g., **802**) can hybridize to the poly-A tail of a mRNA molecule. Reverse transcription may result in a cDNA transcript of the mRNA, but which transcript includes each of the sequence segments **808**, **810**, **816** of the nucleic acid molecule **802**. Because the nucleic acid molecule **802** comprises an anchoring sequence **814**, it will more likely hybridize to and prime reverse transcription at the sequence end of the poly-A tail of the mRNA. Within any given partition, all of the cDNA transcripts of the individual mRNA molecules may include a common barcode sequence segment **810**. However, the transcripts made from the different mRNA molecules within a given partition may vary at the unique molecular identifying sequence **812** segment (e.g., UMI segment). Beneficially, even following any subsequent amplification of the contents of a given

partition, the number of different UMIs can be indicative of the quantity of mRNA originating from a given partition, and thus from the biological particle (e.g., cell). As noted above, the transcripts can be amplified, cleaned up and sequenced to identify the sequence of the cDNA transcript of the mRNA, as well as to sequence the barcode segment and the UMI segment. While a poly-T primer sequence is described, other targeted or random priming sequences may also be used in priming the reverse transcription reaction. Likewise, although described as releasing the barcoded oligonucleotides into the partition, in some cases, the nucleic acid molecules bound to the bead (e.g., gel bead) may be used to hybridize and capture the mRNA on the solid phase of the bead, for example, in order to facilitate the separation of the RNA from other cell contents.

[00160] In some cases, precursors comprising a functional group that is reactive or capable of being activated such that it becomes reactive can be polymerized with other precursors to generate gel beads comprising the activated or activatable functional group. The functional group may then be used to attach additional species (e.g., disulfide linkers, primers, other oligonucleotides, etc.) to the gel beads. For example, some precursors comprising a carboxylic acid (COOH) group can co-polymerize with other precursors to form a gel bead that also comprises a COOH functional group. In some cases, acrylic acid (a species comprising free COOH groups), acrylamide, and bis(acryloyl)cystamine can be co-polymerized together to generate a gel bead comprising free COOH groups. The COOH groups of the gel bead can be activated (e.g., via 1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC) and N-Hydroxysuccinimide (NHS) or 4-(4,6-Dimethoxy-1,3,5-triazin-2-yl)-4-methylmorpholinium chloride (DMTMM)) such that they are reactive (e.g., reactive to amine functional groups where EDC/NHS or DMTMM are used for activation). The activated COOH groups can then react with an appropriate species (e.g., a species comprising an amine functional group where the carboxylic acid groups are activated to be reactive with an amine functional group) comprising a moiety to be linked to the bead.

[00161] Beads comprising disulfide linkages in their polymeric network may be functionalized with additional species via reduction of some of the disulfide linkages to free thiols. The disulfide linkages may be reduced via, for example, the action of a reducing agent (e.g., DTT, TCEP, etc.) to generate free thiol groups, without dissolution of the bead. Free thiols of the beads can then react with free thiols of a species or a species comprising another disulfide bond (e.g., via thiol-disulfide exchange) such that the species can be linked to the beads (e.g., via a generated disulfide bond). In some cases, free thiols of the beads may react with any other suitable group. For example, free thiols of the beads may react with species comprising an acrydite moiety. The free thiol groups of the beads can react with the acrydite via Michael

addition chemistry, such that the species comprising the acrydite is linked to the bead. In some cases, uncontrolled reactions can be prevented by inclusion of a thiol capping agent such as N-ethylmaleimide or iodoacetate.

[00162] Activation of disulfide linkages within a bead can be controlled such that only a small number of disulfide linkages are activated. Control may be exerted, for example, by controlling the concentration of a reducing agent used to generate free thiol groups and/or concentration of reagents used to form disulfide bonds in bead polymerization. In some cases, a low concentration (e.g., molecules of reducing agent:gel bead ratios of less than or equal to about 1:100,000,000,000, less than or equal to about 1:10,000,000,000, less than or equal to about 1:1,000,000,000, less than or equal to about 1:100,000,000, less than or equal to about 1:10,000,000, less than or equal to about 1:1,000,000, less than or equal to about 1:100,000, less than or equal to about 1:10,000) of reducing agent may be used for reduction. Controlling the number of disulfide linkages that are reduced to free thiols may be useful in ensuring bead structural integrity during functionalization. In some cases, optically-active agents, such as fluorescent dyes may be coupled to beads via free thiol groups of the beads and used to quantify the number of free thiols present in a bead and/or track a bead.

[00163] In some cases, addition of moieties to a gel bead after gel bead formation may be advantageous. For example, addition of an oligonucleotide (e.g., barcoded oligonucleotide) after gel bead formation may avoid loss of the species during chain transfer termination that can occur during polymerization. Moreover, smaller precursors (e.g., monomers or cross linkers that do not comprise side chain groups and linked moieties) may be used for polymerization and can be minimally hindered from growing chain ends due to viscous effects. In some cases, functionalization after gel bead synthesis can minimize exposure of species (e.g., oligonucleotides) to be loaded with potentially damaging agents (e.g., free radicals) and/or chemical environments. In some cases, the generated gel may possess an upper critical solution temperature (UCST) that can permit temperature driven swelling and collapse of a bead. Such functionality may aid in oligonucleotide (e.g., a primer) infiltration into the bead during subsequent functionalization of the bead with the oligonucleotide. Post-production functionalization may also be useful in controlling loading ratios of species in beads, such that, for example, the variability in loading ratio is minimized. Species loading may also be performed in a batch process such that a plurality of beads can be functionalized with the species in a single batch.

[00164] A bead injected or otherwise introduced into a partition may comprise releasably, cleavably, or reversibly attached barcodes. A bead injected or otherwise introduced into a

partition may comprise activatable barcodes. A bead injected or otherwise introduced into a partition may be degradable, disruptable, or dissolvable beads.

[00165] Barcodes can be releasably, cleavably or reversibly attached to the beads such that barcodes can be released or be releasable through cleavage of a linkage between the barcode molecule and the bead, or released through degradation of the underlying bead itself, allowing the barcodes to be accessed or be accessible by other reagents, or both. In non-limiting examples, cleavage may be achieved through reduction of di-sulfide bonds, use of restriction enzymes, photo-activated cleavage, or cleavage via other types of stimuli (e.g., chemical, thermal, pH, enzymatic, etc.) and/or reactions, such as described elsewhere herein. Releasable barcodes may sometimes be referred to as being activatable, in that they are available for reaction once released. Thus, for example, an activatable barcode may be activated by releasing the barcode from a bead (or other suitable type of partition described herein). Other activatable configurations are also envisioned in the context of the described methods and systems.

[00166] In addition to, or as an alternative to the cleavable linkages between the beads and the associated molecules, such as barcode containing nucleic acid molecules (e.g., barcoded oligonucleotides), the beads may be degradable, disruptable, or dissolvable spontaneously or upon exposure to one or more stimuli (e.g., temperature changes, pH changes, exposure to particular chemical species or phase, exposure to light, reducing agent, etc.). In some cases, a bead may be dissolvable, such that material components of the beads are solubilized when exposed to a particular chemical species or an environmental change, such as a change temperature or a change in pH. In some cases, a gel bead can be degraded or dissolved at elevated temperature and/or in basic conditions. In some cases, a bead may be thermally degradable such that when the bead is exposed to an appropriate change in temperature (e.g., heat), the bead degrades. Degradation or dissolution of a bead bound to a species (e.g., a nucleic acid molecule, e.g., barcoded oligonucleotide) may result in release of the species from the bead.

[00167] As will be appreciated from the above disclosure, the degradation of a bead may refer to the disassociation of a bound or entrained species from a bead, both with and without structurally degrading the physical bead itself. For example, the degradation of the bead may involve cleavage of a cleavable linkage via one or more species and/or methods described elsewhere herein. In another example, entrained species may be released from beads through osmotic pressure differences due to, for example, changing chemical environments. By way of example, alteration of bead pore sizes due to osmotic pressure differences can generally occur without structural degradation of the bead itself. In some cases, an increase in pore size due to osmotic swelling of a bead can permit the release of entrained species within the bead. In other

cases, osmotic shrinking of a bead may cause a bead to better retain an entrained species due to pore size contraction.

[00168] A degradable bead may be introduced into a partition, such as a droplet of an emulsion or a well, such that the bead degrades within the partition and any associated species (e.g., oligonucleotides) are released within the droplet when the appropriate stimulus is applied. The free species (e.g., oligonucleotides, nucleic acid molecules) may interact with other reagents contained in the partition. For example, a polyacrylamide bead comprising cystamine and linked, via a disulfide bond, to a barcode sequence, may be combined with a reducing agent within a droplet of a water-in-oil emulsion. Within the droplet, the reducing agent can break the various disulfide bonds, resulting in bead degradation and release of the barcode sequence into the aqueous, inner environment of the droplet. In another example, heating of a droplet comprising a bead-bound barcode sequence in basic solution may also result in bead degradation and release of the attached barcode sequence into the aqueous, inner environment of the droplet.

[00169] Any suitable number of molecular tag molecules (e.g., primer, barcoded oligonucleotide) can be associated with a bead such that, upon release from the bead, the molecular tag molecules (e.g., primer, e.g., barcoded oligonucleotide) are present in the partition at a pre-defined concentration. Such pre-defined concentration may be selected to facilitate certain reactions for generating a sequencing library, e.g., amplification, within the partition. In some cases, the pre-defined concentration of the primer can be limited by the process of producing nucleic acid molecule (e.g., oligonucleotide) bearing beads.

[00170] In some cases, beads can be non-covalently loaded with one or more reagents. The beads can be non-covalently loaded by, for instance, subjecting the beads to conditions sufficient to swell the beads, allowing sufficient time for the reagents to diffuse into the interiors of the beads, and subjecting the beads to conditions sufficient to de-swell the beads. The swelling of the beads may be accomplished, for instance, by placing the beads in a thermodynamically favorable solvent, subjecting the beads to a higher or lower temperature, subjecting the beads to a higher or lower ion concentration, and/or subjecting the beads to an electric field. The swelling of the beads may be accomplished by various swelling methods. The de-swelling of the beads may be accomplished, for instance, by transferring the beads in a thermodynamically unfavorable solvent, subjecting the beads to lower or high temperatures, subjecting the beads to a lower or higher ion concentration, and/or removing an electric field. The de-swelling of the beads may be accomplished by various de-swelling methods. Transferring the beads may cause pores in the bead to shrink. The shrinking may then hinder reagents within the beads from diffusing out of the interiors of the beads. The hindrance may be due to steric interactions between the reagents and the interiors of the beads. The transfer may be accomplished

microfluidically. For instance, the transfer may be achieved by moving the beads from one co-flowing solvent stream to a different co-flowing solvent stream. The swellability and/or pore size of the beads may be adjusted by changing the polymer composition of the bead.

[00171] In some cases, an acrydite moiety linked to a precursor, another species linked to a precursor, or a precursor itself can comprise a labile bond, such as chemically, thermally, or photo-sensitive bond e.g., disulfide bond, UV sensitive bond, or the like. Once acrydite moieties or other moieties comprising a labile bond are incorporated into a bead, the bead may also comprise the labile bond. The labile bond may be, for example, useful in reversibly linking (e.g., covalently linking) species (e.g., barcodes, primers, etc.) to a bead. In some cases, a thermally labile bond may include a nucleic acid hybridization based attachment, e.g., where an oligonucleotide is hybridized to a complementary sequence that is attached to the bead, such that thermal melting of the hybrid releases the oligonucleotide, e.g., a barcode containing sequence, from the bead or microcapsule.

[00172] The addition of multiple types of labile bonds to a gel bead may result in the generation of a bead capable of responding to varied stimuli. Each type of labile bond may be sensitive to an associated stimulus (e.g., chemical stimulus, light, temperature, enzymatic, etc.) such that release of species attached to a bead via each labile bond may be controlled by the application of the appropriate stimulus. Such functionality may be useful in controlled release of species from a gel bead. In some cases, another species comprising a labile bond may be linked to a gel bead after gel bead formation via, for example, an activated functional group of the gel bead as described above. As will be appreciated, barcodes that are releasably, cleavably or reversibly attached to the beads described herein include barcodes that are released or releasable through cleavage of a linkage between the barcode molecule and the bead, or that are released through degradation of the underlying bead itself, allowing the barcodes to be accessed or accessible by other reagents, or both.

[00173] The barcodes that are releasable as described herein may sometimes be referred to as being activatable, in that they are available for reaction once released. Thus, for example, an activatable barcode may be activated by releasing the barcode from a bead (or other suitable type of partition described herein). Other activatable configurations are also envisioned in the context of the described methods and systems.

[00174] In addition to thermally cleavable bonds, disulfide bonds and UV sensitive bonds, other non-limiting examples of labile bonds that may be coupled to a precursor or bead include an ester linkage (e.g., cleavable with an acid, a base, or hydroxylamine), a vicinal diol linkage (e.g., cleavable via sodium periodate), a Diels-Alder linkage (e.g., cleavable via heat), a sulfone linkage (e.g., cleavable via a base), a silyl ether linkage (e.g., cleavable via an acid), a glycosidic

linkage (e.g., cleavable via an amylase), a peptide linkage (e.g., cleavable via a protease), or a phosphodiester linkage (e.g., cleavable via a nuclease (e.g., DNAase)). A bond may be cleavable via other nucleic acid molecule targeting enzymes, such as restriction enzymes (e.g., restriction endonucleases), as described further below.

[00175] Species may be encapsulated in beads during bead generation (e.g., during polymerization of precursors). Such species may or may not participate in polymerization. Such species may be entered into polymerization reaction mixtures such that generated beads comprise the species upon bead formation. In some cases, such species may be added to the gel beads after formation. Such species may include, for example, nucleic acid molecules (e.g., oligonucleotides), reagents for a nucleic acid amplification reaction (e.g., primers, polymerases, dNTPs, co-factors (e.g., ionic co-factors), buffers) including those described herein, reagents for enzymatic reactions (e.g., enzymes, co-factors, substrates, buffers), reagents for nucleic acid modification reactions such as polymerization, ligation, or digestion, and/or reagents for template preparation (e.g., tagmentation) for one or more sequencing platforms (e.g., Nextera® for Illumina®). Such species may include one or more enzymes described herein, including without limitation, polymerase, reverse transcriptase, restriction enzymes (e.g., endonuclease), transposase, ligase, proteinase K, DNase, etc. Such species may include one or more reagents described elsewhere herein (e.g., lysis agents, inhibitors, inactivating agents, chelating agents, stimulus). Trapping of such species may be controlled by the polymer network density generated during polymerization of precursors, control of ionic charge within the gel bead (e.g., via ionic species linked to polymerized species), or by the release of other species. Encapsulated species may be released from a bead upon bead degradation and/or by application of a stimulus capable of releasing the species from the bead. Alternatively or in addition, species may be partitioned in a partition (e.g., droplet) during or subsequent to partition formation. Such species may include, without limitation, the abovementioned species that may also be encapsulated in a bead.

[00176] A degradable bead may comprise one or more species with a labile bond such that, when the bead/species is exposed to the appropriate stimuli, the bond is broken and the bead degrades. The labile bond may be a chemical bond (e.g., covalent bond, ionic bond) or may be another type of physical interaction (e.g., van der Waals interactions, dipole-dipole interactions, etc.). In some cases, a crosslinker used to generate a bead may comprise a labile bond. Upon exposure to the appropriate conditions, the labile bond can be broken and the bead degraded. For example, upon exposure of a polyacrylamide gel bead comprising cystamine crosslinkers to a reducing agent, the disulfide bonds of the cystamine can be broken and the bead degraded.

[00177] A degradable bead may be useful in more quickly releasing an attached species (e.g., a nucleic acid molecule, a barcode sequence, a primer, etc) from the bead when the appropriate stimulus is applied to the bead as compared to a bead that does not degrade. For example, for a species bound to an inner surface of a porous bead or in the case of an encapsulated species, the species may have greater mobility and accessibility to other species in solution upon degradation of the bead. In some cases, a species may also be attached to a degradable bead via a degradable linker (e.g., disulfide linker). The degradable linker may respond to the same stimuli as the degradable bead or the two degradable species may respond to different stimuli. For example, a barcode sequence may be attached, via a disulfide bond, to a polyacrylamide bead comprising cystamine. Upon exposure of the barcoded-bead to a reducing agent, the bead degrades and the barcode sequence is released upon breakage of both the disulfide linkage between the barcode sequence and the bead and the disulfide linkages of the cystamine in the bead.

[00178] As will be appreciated from the above disclosure, while referred to as degradation of a bead, in many instances as noted above, that degradation may refer to the disassociation of a bound or entrained species from a bead, both with and without structurally degrading the physical bead itself. For example, entrained species may be released from beads through osmotic pressure differences due to, for example, changing chemical environments. By way of example, alteration of bead pore sizes due to osmotic pressure differences can generally occur without structural degradation of the bead itself. In some cases, an increase in pore size due to osmotic swelling of a bead can permit the release of entrained species within the bead. In other cases, osmotic shrinking of a bead may cause a bead to better retain an entrained species due to pore size contraction.

[00179] Where degradable beads are provided, it may be beneficial to avoid exposing such beads to the stimulus or stimuli that cause such degradation prior to a given time, in order to, for example, avoid premature bead degradation and issues that arise from such degradation, including for example poor flow characteristics and aggregation. By way of example, where beads comprise reducible cross-linking groups, such as disulfide groups, it will be desirable to avoid contacting such beads with reducing agents, e.g., DTT or other disulfide cleaving reagents. In such cases, treatment to the beads described herein will, in some cases be provided free of reducing agents, such as DTT. Because reducing agents are often provided in commercial enzyme preparations, it may be desirable to provide reducing agent free (or DTT free) enzyme preparations in treating the beads described herein. Examples of such enzymes include, e.g., polymerase enzyme preparations, reverse transcriptase enzyme preparations, ligase enzyme preparations, as well as many other enzyme preparations that may be used to treat the beads described herein. The terms “reducing agent free” or “DTT free” preparations can refer to a

preparation having less than about 1/10th, less than about 1/50th, or even less than about 1/100th of the lower ranges for such materials used in degrading the beads. For example, for DTT, the reducing agent free preparation can have less than about 0.01 millimolar (mM), 0.005 mM, 0.001 mM DTT, 0.0005 mM DTT, or even less than about 0.0001 mM DTT. In many cases, the amount of DTT can be undetectable.

[00180] Numerous chemical triggers may be used to trigger the degradation of beads. Examples of these chemical changes may include, but are not limited to pH-mediated changes to the integrity of a component within the bead, degradation of a component of a bead via cleavage of cross-linked bonds, and depolymerization of a component of a bead.

[00181] In some embodiments, a bead may be formed from materials that comprise degradable chemical crosslinkers, such as BAC or cystamine. Degradation of such degradable crosslinkers may be accomplished through a number of mechanisms. In some examples, a bead may be contacted with a chemical degrading agent that may induce oxidation, reduction or other chemical changes. For example, a chemical degrading agent may be a reducing agent, such as dithiothreitol (DTT). Additional examples of reducing agents may include β -mercaptoethanol, (2S)-2-amino-1,4-dimercaptobutane (dithiobutylamine or DTBA), tris(2-carboxyethyl) phosphine (TCEP), or combinations thereof. A reducing agent may degrade the disulfide bonds formed between gel precursors forming the bead, and thus, degrade the bead. In other cases, a change in pH of a solution, such as an increase in pH, may trigger degradation of a bead. In other cases, exposure to an aqueous solution, such as water, may trigger hydrolytic degradation, and thus degradation of the bead. In some cases, any combination of stimuli may trigger degradation of a bead. For example, a change in pH may enable a chemical agent (e.g., DTT) to become an effective reducing agent.

[00182] Beads may also be induced to release their contents upon the application of a thermal stimulus. A change in temperature can cause a variety of changes to a bead. For example, heat can cause a solid bead to liquefy. A change in heat may cause melting of a bead such that a portion of the bead degrades. In other cases, heat may increase the internal pressure of the bead components such that the bead ruptures or explodes. Heat may also act upon heat-sensitive polymers used as materials to construct beads.

[00183] Any suitable agent may degrade beads. In some embodiments, changes in temperature or pH may be used to degrade thermo-sensitive or pH-sensitive bonds within beads. In some embodiments, chemical degrading agents may be used to degrade chemical bonds within beads by oxidation, reduction or other chemical changes. For example, a chemical degrading agent may be a reducing agent, such as DTT, wherein DTT may degrade the disulfide bonds formed between a crosslinker and gel precursors, thus degrading the bead. In some

embodiments, a reducing agent may be added to degrade the bead, which may or may not cause the bead to release its contents. Examples of reducing agents may include dithiothreitol (DTT), β -mercaptoethanol, (2S)-2-amino-1,4-dimercaptobutane (dithiobutylamine or DTBA), tris(2-carboxyethyl) phosphine (TCEP), or combinations thereof. The reducing agent may be present at a concentration of about 0.1mM, 0.5mM, 1mM, 5mM, 10mM. The reducing agent may be present at a concentration of at least about 0.1mM, 0.5mM, 1mM, 5mM, 10mM, or greater than 10 mM. The reducing agent may be present at concentration of at most about 10mM, 5mM, 1mM, 0.5mM, 0.1mM, or less.

[00184] Any suitable number of molecular tag molecules (e.g., primer, barcoded oligonucleotide) can be associated with a bead such that, upon release from the bead, the molecular tag molecules (e.g., primer, e.g., barcoded oligonucleotide) are present in the partition at a pre-defined concentration. Such pre-defined concentration may be selected to facilitate certain reactions for generating a sequencing library, e.g., amplification, within the partition. In some cases, the pre-defined concentration of the primer can be limited by the process of producing oligonucleotide bearing beads.

[00185] Although **FIG. 1** and **FIG. 2** have been described in terms of providing substantially singly occupied partitions, above, in certain cases, it may be desirable to provide multiply occupied partitions, e.g., containing two, three, four or more cells and/or microcapsules (e.g., beads) comprising barcoded nucleic acid molecules (e.g., oligonucleotides) within a single partition. Accordingly, as noted above, the flow characteristics of the biological particle and/or bead containing fluids and partitioning fluids may be controlled to provide for such multiply occupied partitions. In particular, the flow parameters may be controlled to provide a given occupancy rate at greater than about 50% of the partitions, greater than about 75%, and in some cases greater than about 80%, 90%, 95%, or higher.

[00186] In some cases, additional microcapsules can be used to deliver additional reagents to a partition. In such cases, it may be advantageous to introduce different beads into a common channel or droplet generation junction, from different bead sources (e.g., containing different associated reagents) through different channel inlets into such common channel or droplet generation junction (e.g., junction **210**). In such cases, the flow and frequency of the different beads into the channel or junction may be controlled to provide for a certain ratio of microcapsules from each source, while ensuring a given pairing or combination of such beads into a partition with a given number of biological particles (e.g., one biological particle and one bead per partition).

[00187] The partitions described herein may comprise small volumes, for example, less than about 10 microliters (μ L), 5 μ L, 1 μ L, 900 picoliters (pL), 800 pL, 700 pL, 600 pL, 500 pL,

400pL, 300 pL, 200 pL, 100pL, 50 pL, 20 pL, 10 pL, 1 pL, 500 nanoliters (nL), 100 nL, 50 nL, or less.

[00188] For example, in the case of droplet based partitions, the droplets may have overall volumes that are less than about 1000 pL, 900 pL, 800 pL, 700 pL, 600 pL, 500 pL, 400pL, 300 pL, 200 pL, 100pL, 50 pL, 20 pL, 10 pL, 1 pL, or less. Where co-partitioned with microcapsules, it will be appreciated that the sample fluid volume, e.g., including co-partitioned biological particles and/or beads, within the partitions may be less than about 90% of the above described volumes, less than about 80%, less than about 70%, less than about 60%, less than about 50%, less than about 40%, less than about 30%, less than about 20%, or less than about 10% of the above described volumes. **FIG. 49** shows exemplary parameters of ~ 1,000 pL droplets vs. smaller ~ 400 pL droplets. In some cases, the use of a smaller droplet size may result in a decrease in Poisson derived cell doublet rate. The use of a smaller droplet size may also result in an ~2-fold increase in cell throughput.

[00189] As is described elsewhere herein, partitioning species may generate a population or plurality of partitions. In such cases, any suitable number of partitions can be generated or otherwise provided. For example, at least about 1,000 partitions, at least about 5,000 partitions, at least about 10,000 partitions, at least about 50,000 partitions, at least about 100,000 partitions, at least about 500,000 partitions, at least about 1,000,000 partitions, at least about 5,000,000 partitions at least about 10,000,000 partitions, at least about 50,000,000 partitions, at least about 100,000,000 partitions, at least about 500,000,000 partitions, at least about 1,000,000,000 partitions, or more partitions can be generated or otherwise provided. Moreover, the plurality of partitions may comprise both unoccupied partitions (e.g., empty partitions) and occupied partitions.

Reagents

[00190] In accordance with certain aspects, biological particles may be partitioned along with lysis reagents in order to release the contents of the biological particles within the partition. In such cases, the lysis agents can be contacted with the biological particle suspension concurrently with, or immediately prior to, the introduction of the biological particles into the partitioning junction/droplet generation zone (e.g., junction **210**), such as through an additional channel or channels upstream of the channel junction. In accordance with other aspects, additionally or alternatively, biological particles may be partitioned along with other reagents, as will be described further below.

[00191] **FIG. 3** shows an example of a microfluidic channel structure **300** for co-partitioning biological particles and reagents. The channel structure **300** can include channel segments **301**,

302, 304, 306 and 308. Channel segments **301** and **302** communicate at a first channel junction **309**. Channel segments **302, 304, 306, and 308** communicate at a second channel junction **310**.

[00192] In an example operation, the channel segment **301** may transport an aqueous fluid **312** that includes a plurality of biological particles **314** along the channel segment **301** into the second junction **310**. As an alternative or in addition to, channel segment **301** may transport beads (e.g., gel beads). The beads may comprise barcode molecules.

[00193] For example, the channel segment **301** may be connected to a reservoir comprising an aqueous suspension of biological particles **314**. Upstream of, and immediately prior to reaching, the second junction **310**, the channel segment **301** may meet the channel segment **302** at the first junction **309**. The channel segment **302** may transport a plurality of reagents **315** (e.g., lysis agents) suspended in the aqueous fluid **312** along the channel segment **302** into the first junction **309**. For example, the channel segment **302** may be connected to a reservoir comprising the reagents **315**. After the first junction **309**, the aqueous fluid **312** in the channel segment **301** can carry both the biological particles **314** and the reagents **315** towards the second junction **310**. In some instances, the aqueous fluid **312** in the channel segment **301** can include one or more reagents, which can be the same or different reagents as the reagents **315**. A second fluid **316** that is immiscible with the aqueous fluid **312** (e.g., oil) can be delivered to the second junction **310** from each of channel segments **304** and **306**. Upon meeting of the aqueous fluid **312** from the channel segment **301** and the second fluid **316** from each of channel segments **304** and **306** at the second channel junction **310**, the aqueous fluid **312** can be partitioned as discrete droplets **318** in the second fluid **316** and flow away from the second junction **310** along channel segment **308**. The channel segment **308** may deliver the discrete droplets **318** to an outlet reservoir fluidly coupled to the channel segment **308**, where they may be harvested.

[00194] The second fluid **316** can comprise an oil, such as a fluorinated oil, that includes a fluorosurfactant for stabilizing the resulting droplets, for example, inhibiting subsequent coalescence of the resulting droplets **318**.

[00195] A discrete droplet generated may include an individual biological particle **314** and/or one or more reagents **315**. In some instances, a discrete droplet generated may include a barcode carrying bead (not shown), such as via other microfluidics structures described elsewhere herein. In some instances, a discrete droplet may be unoccupied (e.g., no reagents, no biological particles).

[00196] Beneficially, when lysis reagents and biological particles are co-partitioned, the lysis reagents can facilitate the release of the contents of the biological particles within the partition. The contents released in a partition may remain discrete from the contents of other partitions.

[00197] As will be appreciated, the channel segments described herein may be coupled to any of a variety of different fluid sources or receiving components, including reservoirs, tubing, manifolds, or fluidic components of other systems. As will be appreciated, the microfluidic channel structure **300** may have other geometries. For example, a microfluidic channel structure can have more than two channel junctions. For example, a microfluidic channel structure can have 2, 3, 4, 5 channel segments or more each carrying the same or different types of beads, reagents, and/or biological particles that meet at a channel junction. Fluid flow in each channel segment may be controlled to control the partitioning of the different elements into droplets. Fluid may be directed flow along one or more channels or reservoirs via one or more fluid flow units. A fluid flow unit can comprise compressors (e.g., providing positive pressure), pumps (e.g., providing negative pressure), actuators, and the like to control flow of the fluid. Fluid may also or otherwise be controlled via applied pressure differentials, centrifugal force, electrokinetic pumping, vacuum, capillary or gravity flow, or the like.

[00198] Examples of lysis agents include bioactive reagents, such as lysis enzymes that are used for lysis of different cell types, e.g., gram positive or negative bacteria, plants, yeast, mammalian, etc., such as lysozymes, achromopeptidase, lysostaphin, labiase, kitalase, lyticase, and a variety of other lysis enzymes available from, e.g., Sigma-Aldrich, Inc. (St Louis, MO), as well as other commercially available lysis enzymes. Other lysis agents may additionally or alternatively be co-partitioned with the biological particles to cause the release of the biological particles's contents into the partitions. For example, in some cases, surfactant-based lysis solutions may be used to lyse cells, although these may be less desirable for emulsion based systems where the surfactants can interfere with stable emulsions. In some cases, lysis solutions may include non-ionic surfactants such as, for example, TritonX-100 and Tween 20. In some cases, lysis solutions may include ionic surfactants such as, for example, sarcosyl and sodium dodecyl sulfate (SDS). Electroporation, thermal, acoustic or mechanical cellular disruption may also be used in certain cases, e.g., non-emulsion based partitioning such as encapsulation of biological particles that may be in addition to or in place of droplet partitioning, where any pore size of the encapsulate is sufficiently small to retain nucleic acid fragments of a given size, following cellular disruption.

[00199] Alternatively or in addition to the lysis agents co-partitioned with the biological particles described above, other reagents can also be co-partitioned with the biological particles, including, for example, DNase and RNase inactivating agents or inhibitors, such as proteinase K, chelating agents, such as EDTA, and other reagents employed in removing or otherwise reducing negative activity or impact of different cell lysate components on subsequent processing of nucleic acids. In addition, in the case of encapsulated biological particles, the biological

particles may be exposed to an appropriate stimulus to release the biological particles or their contents from a co-partitioned microcapsule. For example, in some cases, a chemical stimulus may be co-partitioned along with an encapsulated biological particle to allow for the degradation of the microcapsule and release of the cell or its contents into the larger partition. In some cases, this stimulus may be the same as the stimulus described elsewhere herein for release of nucleic acid molecules (e.g., oligonucleotides) from their respective microcapsule (e.g., bead). In alternative aspects, this may be a different and non-overlapping stimulus, in order to allow an encapsulated biological particle to be released into a partition at a different time from the release of nucleic acid molecules into the same partition.

[00200] Additional reagents may also be co-partitioned with the biological particles, such as endonucleases to fragment a biological particle's DNA, DNA polymerase enzymes and dNTPs used to amplify the biological particle's nucleic acid fragments and to attach the barcode molecular tags to the amplified fragments. Other enzymes may be co-partitioned, including without limitation, polymerase, transposase, ligase, proteinase K, DNase, etc. Additional reagents may also include reverse transcriptase enzymes, including enzymes with terminal transferase activity, primers and oligonucleotides, and switch oligonucleotides (also referred to herein as "switch oligos" or "template switching oligonucleotides") which can be used for template switching. In some cases, template switching can be used to increase the length of a cDNA. In some cases, template switching can be used to append a predefined nucleic acid sequence to the cDNA. In an example of template switching, cDNA can be generated from reverse transcription of a template, e.g., cellular mRNA, where a reverse transcriptase with terminal transferase activity can add additional nucleotides, e.g., polyC, to the cDNA in a template independent manner. Switch oligos can include sequences complementary to the additional nucleotides, e.g., polyG. The additional nucleotides (e.g., polyC) on the cDNA can hybridize to the additional nucleotides (e.g., polyG) on the switch oligo, whereby the switch oligo can be used by the reverse transcriptase as template to further extend the cDNA. Template switching oligonucleotides may comprise a hybridization region and a template region. The hybridization region can comprise any sequence capable of hybridizing to the target. In some cases, as previously described, the hybridization region comprises a series of G bases to complement the overhanging C bases at the 3' end of a cDNA molecule. The series of G bases may comprise 1 G base, 2 G bases, 3 G bases, 4 G bases, 5 G bases or more than 5 G bases. The template sequence can comprise any sequence to be incorporated into the cDNA. In some cases, the template region comprises at least 1 (e.g., at least 2, 3, 4, 5 or more) tag sequences and/or functional sequences. Switch oligos may comprise deoxyribonucleic acids; ribonucleic acids; modified nucleic acids including 2-Aminopurine, 2,6-Diaminopurine (2-Amino-dA), inverted

dT, 5-Methyl dC, 2'-deoxyInosine, Super T (5-hydroxybutynl-2'-deoxyuridine), Super G (8-aza-7-deazaguanosine), locked nucleic acids (LNAs), unlocked nucleic acids (UNAs, e.g., UNA-A, UNA-U, UNA-C, UNA-G), Iso-dG, Iso-dC, 2' Fluoro bases (e.g., Fluoro C, Fluoro U, Fluoro A, and Fluoro G), or any combination.

[00201] In some cases, the length of a switch oligo may be at least about 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 151, 152, 153, 154, 155, 156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 190, 191, 192, 193, 194, 195, 196, 197, 198, 199, 200, 201, 202, 203, 204, 205, 206, 207, 208, 209, 210, 211, 212, 213, 214, 215, 216, 217, 218, 219, 220, 221, 222, 223, 224, 225, 226, 227, 228, 229, 230, 231, 232, 233, 234, 235, 236, 237, 238, 239, 240, 241, 242, 243, 244, 245, 246, 247, 248, 249 or 250 nucleotides or longer.

[00202] In some cases, the length of a switch oligo may be at most about 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 151, 152, 153, 154, 155, 156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 190, 191, 192, 193, 194, 195, 196, 197, 198, 199, 200, 201, 202, 203, 204, 205, 206, 207, 208, 209, 210, 211, 212, 213, 214, 215, 216, 217, 218, 219, 220, 221, 222, 223, 224, 225, 226, 227, 228, 229, 230, 231, 232, 233, 234, 235, 236, 237, 238, 239, 240, 241, 242, 243, 244, 245, 246, 247, 248, 249 or 250 nucleotides.

[00203] Once the contents of the cells are released into their respective partitions, the macromolecular components (e.g., macromolecular constituents of biological particles, such as RNA, DNA, or proteins) contained therein may be further processed within the partitions. In accordance with the methods and systems described herein, the macromolecular component contents of individual biological particles can be provided with unique identifiers such that, upon

characterization of those macromolecular components they may be attributed as having been derived from the same biological particle or particles. The ability to attribute characteristics to individual biological particles or groups of biological particles is provided by the assignment of unique identifiers specifically to an individual biological particle or groups of biological particles. Unique identifiers, e.g., in the form of nucleic acid barcodes can be assigned or associated with individual biological particles or populations of biological particles, in order to tag or label the biological particle's macromolecular components (and as a result, its characteristics) with the unique identifiers. These unique identifiers can then be used to attribute the biological particle's components and characteristics to an individual biological particle or group of biological particles.

[00204] In some aspects, this is performed by co-partitioning the individual biological particle or groups of biological particles with the unique identifiers, such as described above (with reference to **FIG. 2**). In some aspects, the unique identifiers are provided in the form of nucleic acid molecules (e.g., oligonucleotides) that comprise nucleic acid barcode sequences that may be attached to or otherwise associated with the nucleic acid contents of individual biological particle, or to other components of the biological particle, and particularly to fragments of those nucleic acids. The nucleic acid molecules are partitioned such that as between nucleic acid molecules in a given partition, the nucleic acid barcode sequences contained therein are the same, but as between different partitions, the nucleic acid molecule can, and do have differing barcode sequences, or at least represent a large number of different barcode sequences across all of the partitions in a given analysis. In some aspects, only one nucleic acid barcode sequence can be associated with a given partition, although in some cases, two or more different barcode sequences may be present.

[00205] The nucleic acid barcode sequences can include from about 6 to about 20 or more nucleotides within the sequence of the nucleic acid molecules (e.g., oligonucleotides). The nucleic acid barcode sequences can include from about 6 to about 20, 30, 40, 50, 60, 70, 80, 90, 100 or more nucleotides. In some cases, the length of a barcode sequence may be about 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 nucleotides or longer. In some cases, the length of a barcode sequence may be at least about 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 nucleotides or longer. In some cases, the length of a barcode sequence may be at most about 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 nucleotides or shorter. These nucleotides may be completely contiguous, i.e., in a single stretch of adjacent nucleotides, or they may be separated into two or more separate subsequences that are separated by 1 or more nucleotides. In some cases, separated barcode subsequences can be from about 4 to about 16 nucleotides in length. In some cases, the barcode subsequence may be about 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16

nucleotides or longer. In some cases, the barcode subsequence may be at least about 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16 nucleotides or longer. In some cases, the barcode subsequence may be at most about 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16 nucleotides or shorter.

[00206] The co-partitioned nucleic acid molecules can also comprise other functional sequences useful in the processing of the nucleic acids from the co-partitioned biological particles. These sequences include, e.g., targeted or random/universal amplification primer sequences for amplifying the genomic DNA from the individual biological particles within the partitions while attaching the associated barcode sequences, sequencing primers or primer recognition sites, hybridization or probing sequences, e.g., for identification of presence of the sequences or for pulling down barcoded nucleic acids, or any of a number of other potential functional sequences. Other mechanisms of co-partitioning oligonucleotides may also be employed, including, e.g., coalescence of two or more droplets, where one droplet contains oligonucleotides, or microdispensing of oligonucleotides into partitions, e.g., droplets within microfluidic systems.

[00207] In an example, microcapsules, such as beads, are provided that each include large numbers of the above described barcoded nucleic acid molecules (e.g., barcoded oligonucleotides) releasably attached to the beads, where all of the nucleic acid molecules attached to a particular bead will include the same nucleic acid barcode sequence, but where a large number of diverse barcode sequences are represented across the population of beads used. In some embodiments, hydrogel beads, e.g., comprising polyacrylamide polymer matrices, are used as a solid support and delivery vehicle for the nucleic acid molecules into the partitions, as they are capable of carrying large numbers of nucleic acid molecules, and may be configured to release those nucleic acid molecules upon exposure to a particular stimulus, as described elsewhere herein. In some cases, the population of beads provides a diverse barcode sequence library that includes at least about 1,000 different barcode sequences, at least about 5,000 different barcode sequences, at least about 10,000 different barcode sequences, at least about 50,000 different barcode sequences, at least about 100,000 different barcode sequences, at least about 1,000,000 different barcode sequences, at least about 5,000,000 different barcode sequences, or at least about 10,000,000 different barcode sequences, or more. Additionally, each bead can be provided with large numbers of nucleic acid (e.g., oligonucleotide) molecules attached. In particular, the number of molecules of nucleic acid molecules including the barcode sequence on an individual bead can be at least about 1,000 nucleic acid molecules, at least about 5,000 nucleic acid molecules, at least about 10,000 nucleic acid molecules, at least about 50,000 nucleic acid molecules, at least about 100,000 nucleic acid molecules, at least about 500,000 nucleic acids, at least about 1,000,000 nucleic acid molecules, at least about 5,000,000 nucleic

acid molecules, at least about 10,000,000 nucleic acid molecules, at least about 50,000,000 nucleic acid molecules, at least about 100,000,000 nucleic acid molecules, at least about 250,000,000 nucleic acid molecules and in some cases at least about 1 billion nucleic acid molecules, or more. Nucleic acid molecules of a given bead can include identical (or common) barcode sequences, different barcode sequences, or a combination of both. Nucleic acid molecules of a given bead can include multiple sets of nucleic acid molecules. Nucleic acid molecules of a given set can include identical barcode sequences. The identical barcode sequences can be different from barcode sequences of nucleic acid molecules of another set.

[00208] Moreover, when the population of beads is partitioned, the resulting population of partitions can also include a diverse barcode library that includes at least about 1,000 different barcode sequences, at least about 5,000 different barcode sequences, at least about 10,000 different barcode sequences, at least at least about 50,000 different barcode sequences, at least about 100,000 different barcode sequences, at least about 1,000,000 different barcode sequences, at least about 5,000,000 different barcode sequences, or at least about 10,000,000 different barcode sequences. Additionally, each partition of the population can include at least about 1,000 nucleic acid molecules, at least about 5,000 nucleic acid molecules, at least about 10,000 nucleic acid molecules, at least about 50,000 nucleic acid molecules, at least about 100,000 nucleic acid molecules, at least about 500,000 nucleic acids, at least about 1,000,000 nucleic acid molecules, at least about 5,000,000 nucleic acid molecules, at least about 10,000,000 nucleic acid molecules, at least about 50,000,000 nucleic acid molecules, at least about 100,000,000 nucleic acid molecules, at least about 250,000,000 nucleic acid molecules and in some cases at least about 1 billion nucleic acid molecules.

[00209] In some cases, it may be desirable to incorporate multiple different barcodes within a given partition, either attached to a single or multiple beads within the partition. For example, in some cases, a mixed, but known set of barcode sequences may provide greater assurance of identification in the subsequent processing, e.g., by providing a stronger address or attribution of the barcodes to a given partition, as a duplicate or independent confirmation of the output from a given partition.

[00210] The nucleic acid molecules (e.g., oligonucleotides) are releasable from the beads upon the application of a particular stimulus to the beads. In some cases, the stimulus may be a photo-stimulus, e.g., through cleavage of a photo-labile linkage that releases the nucleic acid molecules. In other cases, a thermal stimulus may be used, where elevation of the temperature of the beads environment will result in cleavage of a linkage or other release of the nucleic acid molecules from the beads. In still other cases, a chemical stimulus can be used that cleaves a linkage of the nucleic acid molecules to the beads, or otherwise results in release of the nucleic

acid molecules from the beads. In one case, such compositions include the polyacrylamide matrices described above for encapsulation of biological particles, and may be degraded for release of the attached nucleic acid molecules through exposure to a reducing agent, such as DTT.

[00211] In some aspects, provided are systems and methods for controlled partitioning. Droplet size may be controlled by adjusting certain geometric features in channel architecture (e.g., microfluidics channel architecture). For example, an expansion angle, width, and/or length of a channel may be adjusted to control droplet size.

[00212] **FIG. 4** shows an example of a microfluidic channel structure for the controlled partitioning of beads into discrete droplets. A channel structure **400** can include a channel segment **402** communicating at a channel junction **406** (or intersection) with a reservoir **404**. The reservoir **404** can be a chamber. Any reference to “reservoir,” as used herein, can also refer to a “chamber.” In operation, an aqueous fluid **408** that includes suspended beads **412** may be transported along the channel segment **402** into the junction **406** to meet a second fluid **410** that is immiscible with the aqueous fluid **408** in the reservoir **404** to create droplets **416**, **418** of the aqueous fluid **408** flowing into the reservoir **404**. At the junction **406** where the aqueous fluid **408** and the second fluid **410** meet, droplets can form based on factors such as the hydrodynamic forces at the junction **406**, flow rates of the two fluids **408**, **410**, fluid properties, and certain geometric parameters (e.g., w , h_0 , α , etc.) of the channel structure **400**. A plurality of droplets can be collected in the reservoir **404** by continuously injecting the aqueous fluid **408** from the channel segment **402** through the junction **406**.

[00213] A discrete droplet generated may include a bead (e.g., as in occupied droplets **416**). Alternatively, a discrete droplet generated may include more than one bead. Alternatively, a discrete droplet generated may not include any beads (e.g., as in unoccupied droplet **418**). In some instances, a discrete droplet generated may contain one or more biological particles, as described elsewhere herein. In some instances, a discrete droplet generated may comprise one or more reagents, as described elsewhere herein.

[00214] In some instances, the aqueous fluid **408** can have a substantially uniform concentration or frequency of beads **412**. The beads **412** can be introduced into the channel segment **402** from a separate channel (not shown in **FIG. 4**). The frequency of beads **412** in the channel segment **402** may be controlled by controlling the frequency in which the beads **412** are introduced into the channel segment **402** and/or the relative flow rates of the fluids in the channel segment **402** and the separate channel. In some instances, the beads can be introduced into the channel segment **402** from a plurality of different channels, and the frequency controlled accordingly.

[00215] In some instances, the aqueous fluid **408** in the channel segment **402** can comprise biological particles (e.g., described with reference to **FIGS. 1** and **2**). In some instances, the aqueous fluid **408** can have a substantially uniform concentration or frequency of biological particles. As with the beads, the biological particles can be introduced into the channel segment **402** from a separate channel. The frequency or concentration of the biological particles in the aqueous fluid **408** in the channel segment **402** may be controlled by controlling the frequency in which the biological particles are introduced into the channel segment **402** and/or the relative flow rates of the fluids in the channel segment **402** and the separate channel. In some instances, the biological particles can be introduced into the channel segment **402** from a plurality of different channels, and the frequency controlled accordingly. In some instances, a first separate channel can introduce beads and a second separate channel can introduce biological particles into the channel segment **402**. The first separate channel introducing the beads may be upstream or downstream of the second separate channel introducing the biological particles.

[00216] The second fluid **410** can comprise an oil, such as a fluorinated oil, that includes a fluorosurfactant for stabilizing the resulting droplets, for example, inhibiting subsequent coalescence of the resulting droplets.

[00217] In some instances, the second fluid **410** may not be subjected to and/or directed to any flow in or out of the reservoir **404**. For example, the second fluid **410** may be substantially stationary in the reservoir **404**. In some instances, the second fluid **410** may be subjected to flow within the reservoir **404**, but not in or out of the reservoir **404**, such as via application of pressure to the reservoir **404** and/or as affected by the incoming flow of the aqueous fluid **408** at the junction **406**. Alternatively, the second fluid **410** may be subjected and/or directed to flow in or out of the reservoir **404**. For example, the reservoir **404** can be a channel directing the second fluid **410** from upstream to downstream, transporting the generated droplets.

[00218] The channel structure **400** at or near the junction **406** may have certain geometric features that at least partly determine the sizes of the droplets formed by the channel structure **400**. The channel segment **402** can have a height, h_0 and width, w , at or near the junction **406**. By way of example, the channel segment **402** can comprise a rectangular cross-section that leads to a reservoir **404** having a wider cross-section (such as in width or diameter). Alternatively, the cross-section of the channel segment **402** can be other shapes, such as a circular shape, trapezoidal shape, polygonal shape, or any other shapes. The top and bottom walls of the reservoir **404** at or near the junction **406** can be inclined at an expansion angle, α . The expansion angle, α , allows the tongue (portion of the aqueous fluid **408** leaving channel segment **402** at junction **406** and entering the reservoir **404** before droplet formation) to increase in depth and facilitate decrease in curvature of the intermediately formed droplet. Droplet size may decrease

with increasing expansion angle. The resulting droplet radius, R_d , may be predicted by the following equation for the aforementioned geometric parameters of h_0 , w , and α :

$$R_d \approx 0.44 \left(1 + 2.2 \sqrt{\tan \alpha} \frac{w}{h_0} \right) \frac{h_0}{\sqrt{\tan \alpha}}$$

[00219] By way of example, for a channel structure with $w = 21 \mu\text{m}$, $h = 21 \mu\text{m}$, and $\alpha = 3^\circ$, the predicted droplet size is $121 \mu\text{m}$. In another example, for a channel structure with $w = 25 \mu\text{m}$, $h = 25 \mu\text{m}$, and $\alpha = 5^\circ$, the predicted droplet size is $123 \mu\text{m}$. In another example, for a channel structure with $w = 28 \mu\text{m}$, $h = 28 \mu\text{m}$, and $\alpha = 7^\circ$, the predicted droplet size is $124 \mu\text{m}$.

[00220] In some instances, the expansion angle, α , may be between a range of from about 0.5° to about 4° , from about 0.1° to about 10° , or from about 0° to about 90° . For example, the expansion angle can be at least about 0.01° , 0.1° , 0.2° , 0.3° , 0.4° , 0.5° , 0.6° , 0.7° , 0.8° , 0.9° , 1° , 2° , 3° , 4° , 5° , 6° , 7° , 8° , 9° , 10° , 15° , 20° , 25° , 30° , 35° , 40° , 45° , 50° , 55° , 60° , 65° , 70° , 75° , 80° , 85° , or higher. In some instances, the expansion angle can be at most about 89° , 88° , 87° , 86° , 85° , 84° , 83° , 82° , 81° , 80° , 75° , 70° , 65° , 60° , 55° , 50° , 45° , 40° , 35° , 30° , 25° , 20° , 15° , 10° , 9° , 8° , 7° , 6° , 5° , 4° , 3° , 2° , 1° , 0.1° , 0.01° , or less. In some instances, the width, w , can be between a range of from about 100 micrometers (μm) to about 500 μm . In some instances, the width, w , can be between a range of from about 10 μm to about 200 μm . Alternatively, the width can be less than about 10 μm . Alternatively, the width can be greater than about 500 μm . In some instances, the flow rate of the aqueous fluid **408** entering the junction **406** can be between about 0.04 microliters (μL)/minute (min) and about 40 $\mu\text{L}/\text{min}$. In some instances, the flow rate of the aqueous fluid **408** entering the junction **406** can be between about 0.01 microliters (μL)/minute (min) and about 100 $\mu\text{L}/\text{min}$. Alternatively, the flow rate of the aqueous fluid **408** entering the junction **406** can be less than about 0.01 $\mu\text{L}/\text{min}$. Alternatively, the flow rate of the aqueous fluid **408** entering the junction **406** can be greater than about 40 $\mu\text{L}/\text{min}$, such as 45 $\mu\text{L}/\text{min}$, 50 $\mu\text{L}/\text{min}$, 55 $\mu\text{L}/\text{min}$, 60 $\mu\text{L}/\text{min}$, 65 $\mu\text{L}/\text{min}$, 70 $\mu\text{L}/\text{min}$, 75 $\mu\text{L}/\text{min}$, 80 $\mu\text{L}/\text{min}$, 85 $\mu\text{L}/\text{min}$, 90 $\mu\text{L}/\text{min}$, 95 $\mu\text{L}/\text{min}$, 100 $\mu\text{L}/\text{min}$, 110 $\mu\text{L}/\text{min}$, 120 $\mu\text{L}/\text{min}$, 130 $\mu\text{L}/\text{min}$, 140 $\mu\text{L}/\text{min}$, 150 $\mu\text{L}/\text{min}$, or greater. At lower flow rates, such as flow rates of about less than or equal to 10 microliters/minute, the droplet radius may not be dependent on the flow rate of the aqueous fluid **408** entering the junction **406**.

[00221] In some instances, at least about 50% of the droplets generated can have uniform size. In some instances, at least about 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, or greater of the droplets generated can have uniform size. Alternatively, less than about 50% of the droplets generated can have uniform size.

[00222] The throughput of droplet generation can be increased by increasing the points of generation, such as increasing the number of junctions (e.g., junction **406**) between aqueous fluid

408 channel segments (e.g., channel segment **402**) and the reservoir **404**. Alternatively or in addition, the throughput of droplet generation can be increased by increasing the flow rate of the aqueous fluid **408** in the channel segment **402**.

[00223] **FIG. 5** shows an example of a microfluidic channel structure for increased droplet generation throughput. A microfluidic channel structure **500** can comprise a plurality of channel segments **502** and a reservoir **504**. Each of the plurality of channel segments **502** may be in fluid communication with the reservoir **504**. The channel structure **500** can comprise a plurality of channel junctions **506** between the plurality of channel segments **502** and the reservoir **504**. Each channel junction can be a point of droplet generation. The channel segment **402** from the channel structure **400** in **FIG. 4** and any description to the components thereof may correspond to a given channel segment of the plurality of channel segments **502** in channel structure **500** and any description to the corresponding components thereof. The reservoir **404** from the channel structure **400** and any description to the components thereof may correspond to the reservoir **504** from the channel structure **500** and any description to the corresponding components thereof.

[00224] Each channel segment of the plurality of channel segments **502** may comprise an aqueous fluid **508** that includes suspended beads **512**. The reservoir **504** may comprise a second fluid **510** that is immiscible with the aqueous fluid **508**. In some instances, the second fluid **510** may not be subjected to and/or directed to any flow in or out of the reservoir **504**. For example, the second fluid **510** may be substantially stationary in the reservoir **504**. In some instances, the second fluid **510** may be subjected to flow within the reservoir **504**, but not in or out of the reservoir **504**, such as via application of pressure to the reservoir **504** and/or as affected by the incoming flow of the aqueous fluid **508** at the junctions. Alternatively, the second fluid **510** may be subjected and/or directed to flow in or out of the reservoir **504**. For example, the reservoir **504** can be a channel directing the second fluid **510** from upstream to downstream, transporting the generated droplets.

[00225] In operation, the aqueous fluid **508** that includes suspended beads **512** may be transported along the plurality of channel segments **502** into the plurality of junctions **506** to meet the second fluid **510** in the reservoir **504** to create droplets **516**, **518**. A droplet may form from each channel segment at each corresponding junction with the reservoir **504**. At the junction where the aqueous fluid **508** and the second fluid **510** meet, droplets can form based on factors such as the hydrodynamic forces at the junction, flow rates of the two fluids **508**, **510**, fluid properties, and certain geometric parameters (e.g., w , h_0 , α , etc.) of the channel structure **500**, as described elsewhere herein. A plurality of droplets can be collected in the reservoir **504** by continuously injecting the aqueous fluid **508** from the plurality of channel segments **502** through the plurality of junctions **506**. Throughput may significantly increase with the parallel

channel configuration of channel structure **500**. For example, a channel structure having five inlet channel segments comprising the aqueous fluid **508** may generate droplets five times as frequently than a channel structure having one inlet channel segment, provided that the fluid flow rate in the channel segments are substantially the same. The fluid flow rate in the different inlet channel segments may or may not be substantially the same. A channel structure may have as many parallel channel segments as is practical and allowed for the size of the reservoir. For example, the channel structure may have at least about 2, 3, 4, 5, 6, 7, 8, 9, 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 150, 500, 250, 300, 350, 400, 450, 500, 600, 700, 800, 900, 1000, 1500, 5000 or more parallel or substantially parallel channel segments.

[00226] The geometric parameters, w , h_0 , and α , may or may not be uniform for each of the channel segments in the plurality of channel segments **502**. For example, each channel segment may have the same or different widths at or near its respective channel junction with the reservoir **504**. For example, each channel segment may have the same or different height at or near its respective channel junction with the reservoir **504**. In another example, the reservoir **504** may have the same or different expansion angle at the different channel junctions with the plurality of channel segments **502**. When the geometric parameters are uniform, beneficially, droplet size may also be controlled to be uniform even with the increased throughput. In some instances, when it is desirable to have a different distribution of droplet sizes, the geometric parameters for the plurality of channel segments **502** may be varied accordingly.

[00227] In some instances, at least about 50% of the droplets generated can have uniform size. In some instances, at least about 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, or greater of the droplets generated can have uniform size. Alternatively, less than about 50% of the droplets generated can have uniform size.

[00228] **FIG. 6** shows another example of a microfluidic channel structure for increased droplet generation throughput. A microfluidic channel structure **600** can comprise a plurality of channel segments **602** arranged generally circularly around the perimeter of a reservoir **604**. Each of the plurality of channel segments **602** may be in fluid communication with the reservoir **604**. The channel structure **600** can comprise a plurality of channel junctions **606** between the plurality of channel segments **602** and the reservoir **604**. Each channel junction can be a point of droplet generation. The channel segment **402** from the channel structure **400** in **FIG. 2** and any description to the components thereof may correspond to a given channel segment of the plurality of channel segments **602** in channel structure **600** and any description to the corresponding components thereof. The reservoir **404** from the channel structure **400** and any description to the components thereof may correspond to the reservoir **604** from the channel structure **600** and any description to the corresponding components thereof.

[00229] Each channel segment of the plurality of channel segments **602** may comprise an aqueous fluid **608** that includes suspended beads **612**. The reservoir **604** may comprise a second fluid **610** that is immiscible with the aqueous fluid **608**. In some instances, the second fluid **610** may not be subjected to and/or directed to any flow in or out of the reservoir **604**. For example, the second fluid **610** may be substantially stationary in the reservoir **604**. In some instances, the second fluid **610** may be subjected to flow within the reservoir **604**, but not in or out of the reservoir **604**, such as via application of pressure to the reservoir **604** and/or as affected by the incoming flow of the aqueous fluid **608** at the junctions. Alternatively, the second fluid **610** may be subjected and/or directed to flow in or out of the reservoir **604**. For example, the reservoir **604** can be a channel directing the second fluid **610** from upstream to downstream, transporting the generated droplets.

[00230] In operation, the aqueous fluid **608** that includes suspended beads **612** may be transported along the plurality of channel segments **602** into the plurality of junctions **606** to meet the second fluid **610** in the reservoir **604** to create a plurality of droplets **616**. A droplet may form from each channel segment at each corresponding junction with the reservoir **604**. At the junction where the aqueous fluid **608** and the second fluid **610** meet, droplets can form based on factors such as the hydrodynamic forces at the junction, flow rates of the two fluids **608**, **610**, fluid properties, and certain geometric parameters (e.g., widths and heights of the channel segments **602**, expansion angle of the reservoir **604**, etc.) of the channel structure **600**, as described elsewhere herein. A plurality of droplets can be collected in the reservoir **604** by continuously injecting the aqueous fluid **608** from the plurality of channel segments **602** through the plurality of junctions **606**. Throughput may significantly increase with the substantially parallel channel configuration of the channel structure **600**. A channel structure may have as many substantially parallel channel segments as is practical and allowed for by the size of the reservoir. For example, the channel structure may have at least about 2, 3, 4, 5, 6, 7, 8, 9, 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250, 300, 350, 400, 450, 500, 600, 700, 800, 900, 1000, 1500, 5000 or more parallel or substantially parallel channel segments. The plurality of channel segments may be substantially evenly spaced apart, for example, around an edge or perimeter of the reservoir. Alternatively, the spacing of the plurality of channel segments may be uneven.

[00231] The reservoir **604** may have an expansion angle, α (not shown in **FIG. 6**) at or near each channel junction. Each channel segment of the plurality of channel segments **602** may have a width, w , and a height, h_0 , at or near the channel junction. The geometric parameters, w , h_0 , and α , may or may not be uniform for each of the channel segments in the plurality of channel segments **602**. For example, each channel segment may have the same or different widths at or near its respective channel junction with the reservoir **604**. For example, each channel segment

may have the same or different height at or near its respective channel junction with the reservoir **604**.

[00232] The reservoir **604** may have the same or different expansion angle at the different channel junctions with the plurality of channel segments **602**. For example, a circular reservoir (as shown in **FIG. 6**) may have a conical, dome-like, or hemispherical ceiling (e.g., top wall) to provide the same or substantially same expansion angle for each channel segments **602** at or near the plurality of channel junctions **606**. When the geometric parameters are uniform, beneficially, resulting droplet size may be controlled to be uniform even with the increased throughput. In some instances, when it is desirable to have a different distribution of droplet sizes, the geometric parameters for the plurality of channel segments **602** may be varied accordingly.

[00233] In some instances, at least about 50% of the droplets generated can have uniform size. In some instances, at least about 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, or greater of the droplets generated can have uniform size. Alternatively, less than about 50% of the droplets generated can have uniform size. The beads and/or biological particle injected into the droplets may or may not have uniform size.

[00234] **FIG. 7A** shows a cross-section view of another example of a microfluidic channel structure with a geometric feature for controlled partitioning. A channel structure **700** can include a channel segment **702** communicating at a channel junction **706** (or intersection) with a reservoir **704**. In some instances, the channel structure **700** and one or more of its components can correspond to the channel structure **100** and one or more of its components. **FIG. 7B** shows a perspective view of the channel structure **700** of **FIG. 7A**.

[00235] An aqueous fluid **712** comprising a plurality of particles **716** may be transported along the channel segment **702** into the junction **706** to meet a second fluid **714** (e.g., oil, etc.) that is immiscible with the aqueous fluid **712** in the reservoir **704** to create droplets **720** of the aqueous fluid **712** flowing into the reservoir **704**. At the junction **706** where the aqueous fluid **712** and the second fluid **714** meet, droplets can form based on factors such as the hydrodynamic forces at the junction **706**, relative flow rates of the two fluids **712**, **714**, fluid properties, and certain geometric parameters (e.g., Δh , etc.) of the channel structure **700**. A plurality of droplets can be collected in the reservoir **704** by continuously injecting the aqueous fluid **712** from the channel segment **702** at the junction **706**.

[00236] A discrete droplet generated may comprise one or more particles of the plurality of particles **716**. As described elsewhere herein, a particle may be any particle, such as a bead, cell bead, gel bead, biological particle, macromolecular constituents of biological particle, or other particles. Alternatively, a discrete droplet generated may not include any particles.

[00237] In some instances, the aqueous fluid 712 can have a substantially uniform concentration or frequency of particles 716. As described elsewhere herein (e.g., with reference to FIG. 4), the particles 716 (e.g., beads) can be introduced into the channel segment 702 from a separate channel (not shown in FIG. 7). The frequency of particles 716 in the channel segment 702 may be controlled by controlling the frequency in which the particles 716 are introduced into the channel segment 702 and/or the relative flow rates of the fluids in the channel segment 702 and the separate channel. In some instances, the particles 716 can be introduced into the channel segment 702 from a plurality of different channels, and the frequency controlled accordingly. In some instances, different particles may be introduced via separate channels. For example, a first separate channel can introduce beads and a second separate channel can introduce biological particles into the channel segment 702. The first separate channel introducing the beads may be upstream or downstream of the second separate channel introducing the biological particles.

[00238] In some instances, the second fluid 714 may not be subjected to and/or directed to any flow in or out of the reservoir 704. For example, the second fluid 714 may be substantially stationary in the reservoir 704. In some instances, the second fluid 714 may be subjected to flow within the reservoir 704, but not in or out of the reservoir 704, such as via application of pressure to the reservoir 704 and/or as affected by the incoming flow of the aqueous fluid 712 at the junction 706. Alternatively, the second fluid 714 may be subjected and/or directed to flow in or out of the reservoir 704. For example, the reservoir 704 can be a channel directing the second fluid 714 from upstream to downstream, transporting the generated droplets.

[00239] The channel structure 700 at or near the junction 706 may have certain geometric features that at least partly determine the sizes and/or shapes of the droplets formed by the channel structure 700. The channel segment 702 can have a first cross-section height, h_1 , and the reservoir 704 can have a second cross-section height, h_2 . The first cross-section height, h_1 , and the second cross-section height, h_2 , may be different, such that at the junction 706, there is a height difference of Δh . The second cross-section height, h_2 , may be greater than the first cross-section height, h_1 . In some instances, the reservoir may thereafter gradually increase in cross-section height, for example, the more distant it is from the junction 706. In some instances, the cross-section height of the reservoir may increase in accordance with expansion angle, β , at or near the junction 706. The height difference, Δh , and/or expansion angle, β , can allow the tongue (portion of the aqueous fluid 712 leaving channel segment 702 at junction 706 and entering the reservoir 704 before droplet formation) to increase in depth and facilitate decrease in curvature of the intermediately formed droplet. For example, droplet size may decrease with increasing height difference and/or increasing expansion angle.

[00240] The height difference, Δh , can be at least about 1 μm . Alternatively, the height difference can be at least about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, 30, 35, 40, 45, 50, 60, 70, 80, 90, 100, 200, 300, 400, 500 μm or more. Alternatively, the height difference can be at most about 500, 400, 300, 200, 100, 90, 80, 70, 60, 50, 45, 40, 35, 30, 25, 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1 μm or less. In some instances, the expansion angle, β , may be between a range of from about 0.5° to about 4° , from about 0.1° to about 10° , or from about 0° to about 90° . For example, the expansion angle can be at least about 0.01° , 0.1° , 0.2° , 0.3° , 0.4° , 0.5° , 0.6° , 0.7° , 0.8° , 0.9° , 1° , 2° , 3° , 4° , 5° , 6° , 7° , 8° , 9° , 10° , 15° , 20° , 25° , 30° , 35° , 40° , 45° , 50° , 55° , 60° , 65° , 70° , 75° , 80° , 85° , or higher. In some instances, the expansion angle can be at most about 89° , 88° , 87° , 86° , 85° , 84° , 83° , 82° , 81° , 80° , 75° , 70° , 65° , 60° , 55° , 50° , 45° , 40° , 35° , 30° , 25° , 20° , 15° , 10° , 9° , 8° , 7° , 6° , 5° , 4° , 3° , 2° , 1° , 0.1° , 0.01° , or less.

[00241] In some instances, the flow rate of the aqueous fluid **712** entering the junction **706** can be between about 0.04 microliters (μL)/minute (min) and about 40 $\mu\text{L}/\text{min}$. In some instances, the flow rate of the aqueous fluid **712** entering the junction **706** can be between about 0.01 microliters (μL)/minute (min) and about 100 $\mu\text{L}/\text{min}$. Alternatively, the flow rate of the aqueous fluid **712** entering the junction **706** can be less than about 0.01 $\mu\text{L}/\text{min}$. Alternatively, the flow rate of the aqueous fluid **712** entering the junction **706** can be greater than about 40 $\mu\text{L}/\text{min}$, such as 45 $\mu\text{L}/\text{min}$, 50 $\mu\text{L}/\text{min}$, 55 $\mu\text{L}/\text{min}$, 60 $\mu\text{L}/\text{min}$, 65 $\mu\text{L}/\text{min}$, 70 $\mu\text{L}/\text{min}$, 75 $\mu\text{L}/\text{min}$, 80 $\mu\text{L}/\text{min}$, 85 $\mu\text{L}/\text{min}$, 90 $\mu\text{L}/\text{min}$, 95 $\mu\text{L}/\text{min}$, 100 $\mu\text{L}/\text{min}$, 110 $\mu\text{L}/\text{min}$, 120 $\mu\text{L}/\text{min}$, 130 $\mu\text{L}/\text{min}$, 140 $\mu\text{L}/\text{min}$, 150 $\mu\text{L}/\text{min}$, or greater. At lower flow rates, such as flow rates of about less than or equal to 10 microliters/minute, the droplet radius may not be dependent on the flow rate of the aqueous fluid **712** entering the junction **706**. The second fluid **714** may be stationary, or substantially stationary, in the reservoir **704**. Alternatively, the second fluid **714** may be flowing, such as at the above flow rates described for the aqueous fluid **712**.

[00242] In some instances, at least about 50% of the droplets generated can have uniform size. In some instances, at least about 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, or greater of the droplets generated can have uniform size. Alternatively, less than about 50% of the droplets generated can have uniform size.

[00243] While **FIGs. 7A** and **7B** illustrate the height difference, Δh , being abrupt at the junction **706** (e.g., a step increase), the height difference may increase gradually (e.g., from about 0 μm to a maximum height difference). Alternatively, the height difference may decrease gradually (e.g., taper) from a maximum height difference. A gradual increase or decrease in height difference, as used herein, may refer to a continuous incremental increase or decrease in height difference, wherein an angle between any one differential segment of a height profile and

an immediately adjacent differential segment of the height profile is greater than 90° . For example, at the junction **706**, a bottom wall of the channel and a bottom wall of the reservoir can meet at an angle greater than 90° . Alternatively or in addition, a top wall (e.g., ceiling) of the channel and a top wall (e.g., ceiling) of the reservoir can meet an angle greater than 90° . A gradual increase or decrease may be linear or non-linear (e.g., exponential, sinusoidal, etc.). Alternatively or in addition, the height difference may variably increase and/or decrease linearly or non-linearly. While **FIGs. 7A** and **7B** illustrate the expanding reservoir cross-section height as linear (e.g., constant expansion angle, β), the cross-section height may expand non-linearly. For example, the reservoir may be defined at least partially by a dome-like (e.g., hemispherical) shape having variable expansion angles. The cross-section height may expand in any shape.

[00244] The channel networks, e.g., as described above or elsewhere herein, can be fluidly coupled to appropriate fluidic components. For example, the inlet channel segments are fluidly coupled to appropriate sources of the materials they are to deliver to a channel junction. These sources may include any of a variety of different fluidic components, from simple reservoirs defined in or connected to a body structure of a microfluidic device, to fluid conduits that deliver fluids from off-device sources, manifolds, fluid flow units (e.g., actuators, pumps, compressors) or the like. Likewise, the outlet channel segment (e.g., channel segment **208**, reservoir **604**, etc.) may be fluidly coupled to a receiving vessel or conduit for the partitioned cells for subsequent processing. Again, this may be a reservoir defined in the body of a microfluidic device, or it may be a fluidic conduit for delivering the partitioned cells to a subsequent process operation, instrument or component.

[00245] The methods and systems described herein may be used to greatly increase the efficiency of single cell applications and/or other applications receiving droplet-based input. For example, following the sorting of occupied cells and/or appropriately-sized cells, subsequent operations that can be performed can include generation of amplification products, purification (e.g., via solid phase reversible immobilization (SPRI)), further processing (e.g., shearing, ligation of functional sequences, and subsequent amplification (e.g., via PCR)). These operations may occur in bulk (e.g., outside the partition). In the case where a partition is a droplet in an emulsion, the emulsion can be broken and the contents of the droplet pooled for additional operations. Additional reagents that may be co-partitioned along with the barcode bearing bead may include oligonucleotides to block ribosomal RNA (rRNA) and nucleases to digest genomic DNA from cells. Alternatively, rRNA removal agents may be applied during additional processing operations. The configuration of the constructs generated by such a method can help minimize (or avoid) sequencing of the poly-T sequence during sequencing and/or sequence the 5' end of a polynucleotide sequence. The amplification products, for

example, first amplification products and/or second amplification products, may be subject to sequencing for sequence analysis. In some cases, amplification may be performed using the Partial Hairpin Amplification for Sequencing (PHASE) method.

[00246] A variety of applications require the evaluation of the presence and quantification of different biological particle or organism types within a population of biological particles, including, for example, microbiome analysis and characterization, environmental testing, food safety testing, epidemiological analysis, e.g., in tracing contamination or the like.

Methods and systems for reducing or removing mitochondrial DNA

[00247] Cells comprise both nuclear DNA (e.g., DNA comprising chromatin) and mitochondrial DNA. In the tagmentation methods described herein, both nuclear DNA and mitochondrial DNA may be susceptible to tagmentation. Depending on the cell type, a method for measuring accessible chromatin may yield a plurality of sequencing reads, where between about 20 and 80% of the sequencing reads may be attributable to mitochondrial DNA. Such reads may be discarded if nuclear DNA (e.g., accessible chromatin) is of interest. Techniques to either eliminate or reduce mitochondrial DNA prior to tagmentation or to remove or reduce tagmented mitochondrial DNA fragments prior to sequencing may result in enhancement in chromatin-attributable data.

[00248] The present disclosure further provides a method of enhancing nuclear DNA within a sample. The method may comprise reducing or removing mitochondrial DNA. In some cases, the method may comprise providing a sample comprising both mitochondrial DNA and nuclear DNA, and removing mitochondrial DNA from or reducing the relative amount of mitochondrial DNA in the sample prior to analyzing cells or nuclei within the sample for accessible chromatin (e.g., tagmentation). In some cases, the method may comprise providing a sample comprising both mitochondrial DNA and nuclear DNA; subjecting the sample to analysis for accessible chromatin (e.g., as described herein), thereby generating a processed sample comprising both tagmented mitochondrial DNA fragments and tagmented nuclear DNA fragments; and removing tagmented mitochondrial DNA fragments from the processed sample or reducing the relative amount of tagmented mitochondrial DNA fragments within the processed sample.

[00249] In some cases, whole cells may be used as the input material for an ATAC-seq analysis (e.g., as described herein). In some cases, nuclei may be used as the input material for an ATAC-seq analysis. For example, nuclei may be used as the input material where mitochondrial DNA is removed prior to tagmentation of genomic DNA. In this approach, whole cells may be gently lysed to remove their cytoplasmic membrane whilst leaving the nuclear membrane and nucleoprotein packaging of the genome intact. In some cases, a lysis buffer used

to lyse whole cells may comprise one or more of Tris-HCl, NaCl, magnesium ions (e.g., MgCl_2), and a lysis agent. A lysis agent may be, for example, NP40, Tween-20, Digitonin, Nonident P40, or DBDM. Mitochondria may be washed away along with other cellular debris during subsequent centrifugation steps to yield a suspension of nuclei.

[00250] In some cases, mitochondrial membranes may be lysed along with their cytoplasmic counterparts. This may result in mitochondrial DNA non-specifically binding to nuclear membranes during this step and thus being carried into a subsequent tagmentation reaction. A blocking agent (e.g., a blocking protein) may be used to prevent nonspecific binding of mitochondrial DNA to nuclei membranes during cell lysis. A blocking agent may not inhibit downstream biochemistry. In some cases, a blocking agent may be selected from the group consisting of, for example, a single purified protein (e.g., bovine serum albumin or casein), a single purified glycoprotein, a milk protein, fish gelatin, a normal serum (e.g., fetal calf serum, rabbit serum, goat serum, or the like), a buffer such as ThermoFisher Scientific's SuperBlock, and polyvinylpyrrolidone (PVP). For example, the blocking agent (e.g., blocking protein) may be bovine serum albumin (BSA), an inexpensive and readily available isolated protein. The blocking protein (e.g., BSA) may be added to a detergent formulation used during cell lysis. The blocking protein (e.g., BSA) may block available binding sites on nuclei membrane, thereby preventing nonspecific binding of mitochondrial DNA. The blocking agent (e.g., blocking protein) may be provided in any useful concentration. The blocking agent may be provided in a solution comprising a buffer. After providing a blocking agent to a sample (e.g., a solution, partition, or plurality of partitions, such as a plurality of aqueous droplets) comprising one or more cells and/or nuclei, the sample may be agitated and/or centrifuged one or more times, and/or undergo one or more washing steps. During subsequent processing (e.g., centrifugation), cell debris and mitochondrial DNA may be washed away (e.g., in a supernatant). This approach may reduce the amount of sequencing reads associated with mitochondrial DNA in an ATAC-seq library by at least about 5%. For example, the amount of sequencing reads associated with mitochondrial DNA may be reduced by at least about 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 99%, or by 100%. In an example, the amount of sequencing reads associated with mitochondrial DNA may be reduced by at least about 50%, such as from about 30% to about 60%. In some cases, the fraction of sequencing reads associated with mitochondrial DNA of a plurality of sequencing reads derived from a sample (e.g., a solution, partition, or plurality of partitions comprising one or more cells and/or nuclei) may be reduced to be less than about 60%. For example, the fraction of sequencing reads associated with mitochondrial DNA may be reduced to less than about 60%, 55%, 50%, 45%, 40%, 35%, 30%, 25%, 20%, 15%, 10%, 5%, 1%, or less of the total sequencing

reads derived from a sample. In an example, the fraction of sequencing reads associated with mitochondrial DNA may be reduced to between about 1-5% of the total sequencing reads derived from a sample.

[00251] In some cases, a method of generating barcoded nucleic acid fragments comprises: (a) lysing a plurality of cells in the presence of a cell lysis agent and a blocking agent to generate a plurality of lysed cells; (b) separating a plurality of nuclei from the plurality of lysed cells to generate a plurality of biological particles, where an individual biological particle (e.g., cell) comprises a non-template nucleic acid and chromatin comprising a template nucleic acid; (c) providing (i) a plurality of transposon end nucleic acid molecules comprising a transposon end sequence and (ii) a plurality of transposase nucleic acid molecules; (d) generating a plurality of template nucleic acid fragments in a biological particle of the plurality of biological particles with the aid of a transposase-nucleic acid complex comprising a transposase nucleic acid molecule of the plurality of transposase nucleic acid molecules and a transposon end nucleic acid molecule of the plurality of transposon end nucleic acid molecules; (e) generating a partition (e.g., a droplet or well) comprising the biological particle comprising the plurality of template nucleic acid fragments and a plurality of barcode oligonucleotide molecules (e.g., nucleic acid barcode molecules) comprising a barcode sequence; and (f) generating a barcoded template nucleic acid fragment using a barcode oligonucleotide molecule (e.g., nucleic acid barcode molecule) of the plurality of barcode oligonucleotide molecule and a template nucleic acid fragment of the plurality of template nucleic acid fragments. In some cases, the blocking agent may comprise bovine serum albumin (BSA). For example, the blocking agent may comprise a solution of 3% BSA in phosphate buffered saline. Lysing the plurality of cells may comprise providing a lysis buffer comprising the cell lysis agent to the plurality of cells. In some cases, the biological particles may be nuclei, which may be isolated from lysed cells by, e.g., washing the lysed cells. In some cases, the method further comprises generating sequencing reads. Application of the method may reduce the fraction of sequencing reads deriving from mitochondrial DNA relative to the total number of sequencing reads generated. In some cases, the template nucleic acid may comprise chromatin.

[00252] In some cases, a method of generating barcoded nucleic acid fragments comprises: (a) lysing a plurality of cells in the presence of a cell lysis agent and a blocking agent to generate a plurality of lysed cells; (b) separating a plurality of nuclei from the plurality of lysed cells to generate a plurality of biological particles, where an individual biological particle comprises non-template DNA molecules (e.g., mitochondrial DNA molecules), template DNA molecules, and template RNA molecules; (c) providing (i) a plurality of transposon end nucleic acid molecules comprising a transposon end sequence and (ii) a plurality of transposase nucleic acid

molecules; (d) generating a plurality of template DNA fragments in biological particles of the plurality of biological particles with the aid of a transposase-nucleic acid complex comprising a transposase nucleic acid molecule of the plurality of transposase nucleic acid molecules and a transposon end nucleic acid molecule of the plurality of transposon end nucleic acid molecules; (e) generating a partition (e.g., a droplet or well) comprising (i) the biological particle comprising the plurality of template nucleic acid fragments and template RNA molecules; (ii) a plurality of first barcode oligonucleotide molecules (e.g., first nucleic acid barcode molecules) comprising a barcode sequence; (iii) a plurality of second barcode oligonucleotide molecules (e.g., second nucleic acid molecules) comprising a barcode sequence and a capture sequence; and (iv) a plurality of reverse transcriptase molecules; (f) generating a barcoded template DNA fragment using a barcode oligonucleotide molecule of the plurality of first barcode oligonucleotide molecules and a template DNA fragment of the plurality of template DNA fragments; and (g) generating a barcoded cDNA molecule from the template RNA molecules by reverse transcription using a barcode oligonucleotide molecule of the plurality of second barcode oligonucleotide molecules. In some cases, the blocking agent may comprise bovine serum albumin (BSA). For example, the blocking agent may comprise a solution of 3% BSA in phosphate buffered saline. Lysing the plurality of cells may comprise providing a lysis buffer comprising the cell lysis agent to the plurality of cells. In some cases, the biological particles may be nuclei, which may be isolated from lysed cells by, e.g., washing the lysed cells. In some cases, the method further comprises generating sequencing reads. Application of the method may reduce the fraction of sequencing reads deriving from mitochondrial DNA relative to the total number of sequencing reads generated. In some cases, the template DNA molecules may comprise chromatin.

[00253] In some cases, a method of reducing or removing mitochondrial DNA from a sample may comprise providing a sample comprising a plurality of cells comprising nuclear DNA and mitochondrial DNA; subjecting the plurality of cells to analysis for accessible chromatin (e.g., tagmentation, as described herein), thereby generating tagmented mitochondrial DNA fragments and tagmented nuclear DNA fragments; and using an enzyme to deplete tagmented mitochondrial DNA fragments, thereby reducing the relative amount of tagmented mitochondrial DNA fragments within the total number of fragments. In some cases, the enzyme may be an endonuclease enzyme. For example, the enzyme may be an RNA-guided DNA endonuclease enzyme. The enzyme may be a clustered regularly interspaced short palindromic (CRISPR) associated protein (e.g., nuclease) such as Cas9. The enzyme (e.g., Cas9, such as recombinant Cas9) may be complexed with one or more guide RNAs (gRNAs). The Cas9-gRNA scheme may be used to target species for cleavage. For example, tagmented mitochondrial DNA

fragments may be cleaved so that mitochondrial-derived DNA fragments will not be processed in subsequent sequencing. In this scheme, a gRNA may be specifically designed to target a species of interest (e.g., a tagged mitochondrial DNA fragment). An enzyme (e.g., Cas9, such as recombinant Cas9) may be complexed with gRNAs to target loci of human mitochondrial chromosome. Application of the enzyme-gRNA system may result in a reduction in the fraction of mitochondrial DNA fragments relative to a total number of fragments. In an example, a plurality of gRNAs may be provided to a mixture comprising mitochondrial DNA fragments, where the plurality of gRNAs target different regions of mitochondrial DNA. For example, the plurality of gRNAs may target mitochondrial DNA (e.g., human mitochondrial chromosome) about every 250 base pairs.

[00254] In some cases, the amount of sequencing reads associated with mitochondrial DNA in an ATAC-seq library may be reduced by at least about 5%. For example, the amount of sequencing reads associated with mitochondrial DNA may be reduced by at least about 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, or more. In an example the amount of sequencing reads associated with mitochondrial DNA may be reduced by at least about 50%, such as from about 30% to about 60%. In some cases, the fraction of sequencing reads associated with mitochondrial DNA of a plurality of sequencing reads derived from a sample (e.g., a solution, partition, or plurality of partitions comprising one or more cells and/or nuclei) may be reduced to be less than about 60%. For example, the fraction of sequencing reads associated with mitochondrial DNA may be reduced to less than about 60%, 55%, 50%, 45%, 40%, 35%, 30%, 25%, 20%, 15%, 10%, 5%, 1%, or less of the total sequencing reads derived from a sample. In an example, the fraction of sequencing reads associated with mitochondrial DNA may be reduced to between about 1-5% of the total sequencing reads derived from a sample.

[00255] In some cases, ATAC-seq and RNA-seq methods may be combined with a method of reducing non-template (e.g., mitochondrial) fragments. For example, a method may comprise generating both barcoded template nucleic acid fragments and barcoded cDNA fragments (e.g., generated via reverse transcription of template RNA molecules) as well as using a blocking agent to reduce non-template fragments (e.g., as described herein). Alternatively or in addition, a method may comprise generating both barcoded template nucleic acid fragments and barcoded cDNA fragments (e.g., generated via reverse transcription of template RNA molecules) as well as using a CRISPR-associated protein to reduce non-template fragments (e.g., as described herein). In some cases, non-template fragments may comprise mitochondrial DNA. In some cases, non-template fragments may comprise mitochondrial RNA. In an example, a biological particle comprising a template DNA molecule, a non-template nucleic acid molecule comprising

ribosomal RNA (rRNA, e.g., mitochondrial rRNA), and a template RNA molecule is provided, and a barcoded cDNA is generated from the template RNA molecule by reverse transcription using a barcode oligonucleotide (e.g., nucleic acid barcode molecule) (e.g., as described herein). Guide RNA molecules (e.g., sgRNAs) targeted to specific non-template rRNA (e.g., specific regions of mitochondrial rRNA) or cDNA molecule generated from rRNA molecules may be used in combination with a CRISPR associated protein such as Cas9 to selectively deplete sequences associated with rRNA, e.g., upon sequencing a plurality of sequences derived from the biological particle. Therefore, the methods described herein may allow for selective depletion of fragments and/or sequences (e.g., sequence reads) associated with a non-template species such as a non-template rRNA and corresponding enhancement of fragments and/or sequences (e.g., sequence reads) associated with template DNA and RNA species.

[00256] In some cases, a method for nucleic acid processing comprises: (a) generating a partition (e.g., a droplet or well), wherein the partition comprises: (i) a biological particle (e.g., cell); (ii) a plurality of barcode oligonucleotide molecules (e.g., nucleic acid barcode molecules) comprising a common barcode sequence; (iii) a plurality of transposon end oligonucleotide molecules (e.g., transposon end nucleic acid molecules) comprising a transposon end sequence; and (iv) a plurality of transposase molecules, wherein the biological particle (e.g., cell) comprises a template nucleic acid and a non-template nucleic acid; (b) generating a plurality of template nucleic acid fragments and a plurality of non-template nucleic acid fragments (e.g., mitochondrial nucleic acid fragments) with the aid of a transposase-nucleic acid complex comprising a transposase molecule of the plurality of transposase molecules and a transposon end oligonucleotide molecule of the plurality of transposon end oligonucleotide molecules; (c) generating a barcoded template nucleic acid fragment using a barcode oligonucleotide molecule of the plurality of barcode oligonucleotide molecules and a template nucleic acid fragment of the plurality of template nucleic acid fragments; and (d) cleaving one or more non-template nucleic acid fragments of the plurality of non-template nucleic acid fragments, or derivatives thereof, using (i) one or more guide ribonucleic acid molecules (gRNAs) targeted to the one or more non-template nucleic acid fragments, and (ii) a clustered regularly interspaced short palindromic (CRISPR) associated (Cas) nuclease. In some cases, one or more barcoded non-template nucleic acid fragments are generated using barcode oligonucleotide molecules of the plurality of barcode oligonucleotide molecules and one or more non-template nucleic acid fragments (e.g., mitochondrial nucleic acid fragments) of the plurality of non-template nucleic acid fragments. In some cases, the method may reduce the amount of fragments and/or barcoded fragments comprising and/or deriving from non-template nucleic acids. For example, the method may reduce the total number of non-template nucleic acid fragments and barcoded non-

template nucleic acid fragments, e.g., in a mixture comprising one or more of template nucleic acid fragments, barcoded template nucleic acid fragments, non-template nucleic acid fragments, and barcoded non-template nucleic acid fragments. In some cases, the endonuclease may be Cas9, such as recombinant Cas9. In some cases, the non-template nucleic acid fragment may comprise a mitochondrial DNA fragment. In some cases, the template nucleic acid fragment may comprise a nuclear DNA fragment.

[00257] In some cases, a method of generating barcoded nucleic acid fragments comprises: (a) providing a plurality of biological particles (e.g., cells or nuclei), an individual biological particle (e.g., cell or nucleus) comprising a template nucleic acid (e.g., chromatin) and a non-template nucleic acid (e.g., mitochondrial nucleic acid); (b) generating a plurality of template nucleic acid fragments and a plurality of non-template nucleic acid fragments (e.g., mitochondrial nucleic acid fragments) in a biological particle of the plurality of biological particles with the aid of a transposase-nucleic acid complex comprising a transposase nucleic acid molecule and a transposon end nucleic acid molecule; (c) generating a partition (e.g., a droplet or well) comprising the biological particle comprising the plurality of template nucleic acid fragments and the plurality of non-template nucleic acid fragments, and a plurality of barcode oligonucleotide molecules (e.g., nucleic acid barcode molecules) comprising a barcode sequence; (d) generating a barcoded template nucleic acid fragment using a barcode oligonucleotide molecule of the plurality of barcode oligonucleotide molecules and a template nucleic acid fragment of the plurality of template nucleic acid fragments; and (e) cleaving one or more non-template DNA fragments of the plurality of non-template DNA fragments, or derivatives thereof, using (i) one or more guide ribonucleic acid molecules (gRNAs) targeted to the one or more non-template DNA fragments, and (ii) a clustered regularly interspaced short palindromic (CRISPR) associated (Cas) nuclease. In some cases, one or more barcoded non-template nucleic acid fragments are generated using barcode oligonucleotide molecules of the plurality of barcode oligonucleotide molecules and one or more non-template nucleic acid fragments (e.g., mitochondrial nucleic acid fragments) of the plurality of non-template nucleic acid fragments. In some cases, the method may reduce the amount of fragments and/or barcoded fragments comprising and/or deriving from non-template nucleic acids. For example, the method may reduce the total number of non-template nucleic acid fragments and barcoded non-template nucleic acid fragments, e.g., in a mixture comprising one or more of template nucleic acid fragments, barcoded template nucleic acid fragments, non-template nucleic acid fragments, and barcoded non-template nucleic acid fragments. In some cases, the endonuclease may be Cas9, such as recombinant Cas9. In some cases, the non-template nucleic acid fragment may comprise a mitochondrial DNA fragment. In some cases, the template nucleic acid fragment

may comprise chromatin. In some cases, a method of generating barcoded nucleic acid fragments comprises: (a) generating a partition (e.g., a droplet or well), wherein the partition comprises: (i) a biological particle (e.g., cell or nucleus), wherein the biological particle comprises template DNA molecules (e.g., chromatin), non-template DNA molecules (e.g., mitochondrial DNA molecules), and template RNA molecules; (ii) a plurality of first barcode oligonucleotide molecules (e.g., first nucleic acid barcode molecules) comprising a barcode sequence; (iii) a plurality of transposon end nucleic acid molecules comprising a transposon end sequence; (iv) a plurality of transposase nucleic acid molecules; (v) a plurality of second barcode oligonucleotide molecules (e.g., second nucleic acid barcode molecules) comprising a barcode sequence and a capture sequence; and (vi) a plurality of reverse transcriptase molecules; (b) generating a plurality of template DNA fragments and a plurality of non-template DNA fragments (e.g., mitochondrial nucleic acid fragments) with the aid of a transposase-nucleic acid complex comprising a transposase nucleic acid molecule of the plurality of transposase nucleic acid molecules and a transposon end nucleic acid molecule of the plurality of transposon end nucleic acid molecules; (c) generating a barcoded template DNA fragment using a barcode oligonucleotide molecule of the plurality of first barcode oligonucleotide molecules and a template DNA fragment of the plurality of template DNA fragments; (d) generating a barcoded cDNA molecule from the template RNA molecules by reverse transcription using a barcode oligonucleotide molecule of the plurality of second barcode oligonucleotide molecules; and (e) cleaving one or more non-template DNA fragments of the plurality of non-template DNA fragments, or derivatives thereof, using (i) one or more guide ribonucleic acid molecules (gRNAs) targeted to the one or more non-template DNA fragments, and (ii) a clustered regularly interspaced short palindromic (CRISPR) associated (Cas) nuclease. In some cases, one or more barcoded non-template DNA fragments are generated using barcode oligonucleotide molecules of the plurality of barcode oligonucleotide molecules and one or more non-template DNA fragments (e.g., mitochondrial DNA fragments) of the plurality of non-template DNA fragments. In some cases, the method may reduce the amount of fragments and/or barcoded fragments comprising and/or deriving from non-template DNA. For example, the method may reduce the total number of non-template DNA fragments and barcoded non-template DNA fragments, e.g., in a mixture comprising one or more of template DNA fragments, barcoded template DNA fragments, non-template DNA fragments, and barcoded non-template DNA fragments. In some cases, the endonuclease may be Cas9, such as recombinant Cas9. In some cases, the non-template DNA fragment may comprise a mitochondrial DNA fragment. In some cases, the template DNA fragment may comprise chromatin.

[00258] In some cases, a method of generating barcoded nucleic acid fragments comprises:

(a) providing a biological particle (e.g., cell) comprising template DNA molecules, non-template DNA molecules, and template RNA molecules; (b) generating a plurality of template DNA fragments and non-template DNA fragments in the biological particle with the aid of a transposase-nucleic acid complex comprising a transposase nucleic acid molecule and a transposon end nucleic acid molecule; (c) generating a partition (e.g., a droplet or well) comprising (i) the biological particle comprising template DNA fragments, non-template DNA fragments, and template RNA molecules; (ii) a plurality of first barcode oligonucleotide molecules (e.g., first nucleic acid barcode molecules) comprising a common barcode sequence; (iii) a plurality of second barcode oligonucleotide molecules (e.g., second nucleic acid molecules) comprising a barcode sequence and a capture sequence; and (iv) a plurality of reverse transcriptase molecules; (d) generating a barcoded template DNA fragment using a barcode oligonucleotide molecule of the plurality of first barcode oligonucleotide molecules and a template DNA fragment of the plurality of template DNA fragments; (e) generating a barcoded cDNA molecule from the template RNA molecules by reverse transcription using a barcode oligonucleotide molecule of the plurality of second barcode oligonucleotide molecules; and (f) cleaving one or more non-template DNA fragments of the plurality of non-template DNA fragments, or derivatives thereof, using (i) one or more guide ribonucleic acid molecules (gRNAs) targeted to the one or more non-template nucleic acid fragments, and (ii) a clustered regularly interspaced short palindromic (CRISPR) associated (Cas) nuclease. In some cases, one or more barcoded non-template DNA fragments are generated using barcode oligonucleotide molecules of the plurality of barcode oligonucleotide molecules and one or more non-template DNA fragments (e.g., mitochondrial DNA fragments) of the plurality of non-template DNA fragments. In some cases, the method may reduce the amount of fragments and/or barcoded fragments comprising and/or deriving from non-template DNA. For example, the method may reduce the total number of non-template DNA fragments and barcoded non-template DNA fragments, e.g., in a mixture comprising one or more of template DNA fragments, barcoded template DNA fragments, non-template DNA fragments, and barcoded non-template DNA fragments. In some cases, the endonuclease may be Cas9, such as recombinant Cas9. In some cases, the non-template DNA fragment may comprise a mitochondrial DNA fragment. In some cases, the template DNA fragment may comprise a nuclear DNA fragment.

[00259] In some cases, a method of generating barcoded nucleic acid fragments comprises:

(a) providing a partition (e.g., a droplet or well), wherein the partition comprises (i) a biological particle (e.g., cell or nucleus), (ii) a single solid or semi-solid particle (e.g., bead, such as a gel bead), and (iii) a plurality of transposase nucleic acid molecules, wherein the single biological

particle comprises a template nucleic acid molecule and a non-template nucleic acid molecule and wherein the single solid or semi-solid particle (e.g., bead) comprises a barcoded oligonucleotide (e.g., nucleic acid barcode molecule) releasably coupled thereto; (b) subjecting the partition to conditions sufficient to release the template nucleic acid molecule and the non-template nucleic acid molecule from the single biological particle; (c) subjecting the partition to conditions sufficient to release the barcoded oligonucleotide from the solid or semi-solid particle; (d) subjecting the partition to conditions sufficient to cause transposition of the barcoded oligonucleotides into the template nucleic acid molecule and the non-template nucleic acid molecule with the aid of a transposome complex generated from at least a subset of the plurality of transposase nucleic acid molecules; (e) fragmenting (i) the template nucleic acid molecule into a plurality of double-stranded template nucleic acid fragments comprising the barcoded oligonucleotides and (ii) the non-template nucleic acid molecule into a plurality of double-stranded non-template nucleic acid fragments comprising the barcoded oligonucleotides; and (f) cleaving one or more non-template nucleic acid fragments of the plurality of non-template nucleic acid fragments, or derivatives thereof, using (i) one or more guide ribonucleic acid molecules (gRNAs) targeted to the one or more non-template nucleic acid fragments, and (ii) a clustered regularly interspaced short palindromic (CRISPR) associated (Cas) nuclease. In some cases, the method may reduce the amount of fragments comprising and/or deriving from non-template nucleic acids. For example, the method may reduce the total number of non-template nucleic acid fragments, e.g., in a mixture comprising one or more of template nucleic acid fragments, barcoded template nucleic acid fragments, non-template nucleic acid fragments, and barcoded non-template nucleic acid fragments. In some cases, the endonuclease may be Cas9, such as recombinant Cas9. In some cases, the non-template nucleic acid fragment may comprise a mitochondrial DNA fragment. In some cases, the template nucleic acid fragment may comprise chromatin.

[00260] The present methods may have the effect of reducing or removing non-template nucleic acids (e.g., mitochondrial DNA) and/or enhancing accessible chromatin. The methods may be used with any cell type (e.g., as described herein). For example, the methods may be used with a human or a mammalian cell. The methods may also be used with any concentration of cells. For example, the methods of reducing or removing non-template nucleic acids (e.g., mitochondrial DNA) and/or enhancing accessible chromatin may be used with an isolated cell. In another example, the method of reducing or removing non-template nucleic acids (e.g., mitochondrial DNA) and/or enhancing accessible chromatin may be used with a plurality of cells, such as a plurality of cells within a plurality of droplets or a plurality of cells in a bulk solution. The method of reducing or removing non-template nucleic acids (e.g., mitochondrial

DNA) and/or enhancing accessible chromatin may be applied to any library type that requires a reducing in sequencing reads associated with non-template nucleic acids (e.g., mitochondrial DNA) or RNA, including RNA-seq.

[00261] A method of reducing non-template (e.g., mitochondrial) reads and/or enhancing reads associated with accessible chromatin relative to a total number of reads associated with a sample (e.g., a cell or a plurality of cells) may be applied to any of the methods described herein. For example, a blocking agent (e.g., BSA) may be provided to any sample described herein. As described above, use of a blocking agent may reduce or prevent non-specific adsorption of mitochondrial DNA to nuclei such that mitochondrial DNA can be washed away during nuclei isolation. Similarly, an endonuclease may be provided after any tagmentation process described herein to reduce the relative fraction of tagmented mitochondrial DNA in a sample comprising tagmented DNA.

[00262] Additional details and methods regarding processing nucleic acid samples and reducing non-template (e.g., mitochondrial) reads can be found in, for example: Gu et al. (Gu, W.; Crawford, E.D.; O'Donovan, B.D.; Wilson, M.R.; Chow, E.D.; Retallack, H., and DeRisi, J.L. "Depletion of Abundant Sequences by Hybridization (DASH): using Cas9 to remove unwanted high- abundance species in sequencing libraries and molecular counting applications," Genome Biology (2016) 17:41), Montefiori et al. (Montefiori, L.; Hernandez, L.; Zhang, Z.; Gilad, Y.; Ober, C.; Crawford, G.; Nobrega, M.; and Sakabe, N.J. "Reducing mitochondrial reads in ATAC-seq using CRISPR/Cas9," Scientific Reports (2017) 7:2451), and Corces et al. (Corces, M.R.; Trevino, A.E.; Hamilton, E.G.; Greenside, P.G.; Sinnott-Armstrong, N.A.; Vesuna, S.; Satpathy, A.T.; Rubin, A.J.; Montine, K.S.; Wu, B.; Kathiria, A.; Cho, S.W.; Mumbach, M.R.; Carter, A.C.; Kasowski, M.; Orloff, L.A.; Risca, V.I.; Kundaje, A.; Khavari, P.A.; Montine, T.J.; Greenleaf, W.J.; and Chang, H.Y. "An improved ATAC-seq protocol reduces background and enables interrogation of frozen tissues," Nature Methods (2017)), each of which is herein incorporated by reference in its entirety.

Kits

[00263] Also provided herein are kits for analyzing the accessible chromatin (e.g., for ATAC-seq) and/or RNA transcripts of individual cells or small populations of cells. The kits may include one or more of the following: one, two, three, four, five or more, up to all of partitioning fluids, including both aqueous buffers and non-aqueous partitioning fluids or oils, nucleic acid barcode libraries that are releasably associated with beads, as described herein, microfluidic devices, reagents for disrupting cells amplifying nucleic acids, and providing additional

functional sequences on fragments of cellular nucleic acids or replicates thereof, as well as instructions for using any of the foregoing in the methods described herein.

Computer systems

[00264] The present disclosure provides computer systems that are programmed to implement methods of the disclosure. **FIG. 9** shows a computer system **901** that is programmed or otherwise configured to, e.g., (i) control a microfluidics system (e.g., fluid flow), (ii) sort occupied droplets from unoccupied droplets, (iii) polymerize droplets, (iv) perform sequencing applications, (v) generate and maintain a library of DNA or cDNA fragments, and/or (vi) analyze areas of accessible chromatin. The computer system **901** can regulate various aspects of the present disclosure, such as, for example, e.g., regulating fluid flow rate in one or more channels in a microfluidic structure, regulating polymerization application units, regulating conditions for certain reactions described herein. The computer system **901** can be an electronic device of a user or a computer system that is remotely located with respect to the electronic device. The electronic device can be a mobile electronic device.

[00265] The computer system **901** includes a central processing unit (CPU, also “processor” and “computer processor” herein) **905**, which can be a single core or multi core processor, or a plurality of processors for parallel processing. The computer system **901** also includes memory or memory location **910** (e.g., random-access memory, read-only memory, flash memory), electronic storage unit **915** (e.g., hard disk), communication interface **920** (e.g., network adapter) for communicating with one or more other systems, and peripheral devices **925**, such as cache, other memory, data storage and/or electronic display adapters. The memory **910**, storage unit **915**, interface **920** and peripheral devices **925** are in communication with the CPU **905** through a communication bus (solid lines), such as a motherboard. The storage unit **915** can be a data storage unit (or data repository) for storing data. The computer system **901** can be operatively coupled to a computer network (“network”) **930** with the aid of the communication interface **920**. The network **930** can be the Internet, an internet and/or extranet, or an intranet and/or extranet that is in communication with the Internet. The network **930** in some cases is a telecommunication and/or data network. The network **930** can include one or more computer servers, which can enable distributed computing, such as cloud computing. The network **930**, in some cases with the aid of the computer system **901**, can implement a peer-to-peer network, which may enable devices coupled to the computer system **901** to behave as a client or a server.

[00266] The CPU **905** can execute a sequence of machine-readable instructions, which can be embodied in a program or software. The instructions may be stored in a memory location, such as the memory **910**. The instructions can be directed to the CPU **905**, which can subsequently

program or otherwise configure the CPU **905** to implement methods of the present disclosure. Examples of operations performed by the CPU **905** can include fetch, decode, execute, and writeback.

[00267] The CPU **905** can be part of a circuit, such as an integrated circuit. One or more other components of the system **901** can be included in the circuit. In some cases, the circuit is an application specific integrated circuit (ASIC).

[00268] The storage unit **915** can store files, such as drivers, libraries and saved programs. The storage unit **915** can store user data, e.g., user preferences and user programs. The computer system **901** in some cases can include one or more additional data storage units that are external to the computer system **901**, such as located on a remote server that is in communication with the computer system **901** through an intranet or the Internet.

[00269] The computer system **901** can communicate with one or more remote computer systems through the network **930**. For instance, the computer system **901** can communicate with a remote computer system of a user (e.g., operator). Examples of remote computer systems include personal computers (e.g., portable PC), slate or tablet PC's (e.g., Apple® iPad, Samsung® Galaxy Tab), telephones, Smart phones (e.g., Apple® iPhone, Android-enabled device, Blackberry®), or personal digital assistants. The user can access the computer system **901** via the network **930**.

[00270] Methods as described herein can be implemented by way of machine (e.g., computer processor) executable code stored on an electronic storage location of the computer system **901**, such as, for example, on the memory **910** or electronic storage unit **915**. The machine executable or machine readable code can be provided in the form of software. During use, the code can be executed by the processor **905**. In some cases, the code can be retrieved from the storage unit **915** and stored on the memory **910** for ready access by the processor **905**. In some situations, the electronic storage unit **915** can be precluded, and machine-executable instructions are stored on memory **910**.

[00271] The code can be pre-compiled and configured for use with a machine having a processor adapted to execute the code, or can be compiled during runtime. The code can be supplied in a programming language that can be selected to enable the code to execute in a pre-compiled or as-compiled fashion.

[00272] Aspects of the systems and methods provided herein, such as the computer system **901**, can be embodied in programming. Various aspects of the technology may be thought of as “products” or “articles of manufacture” typically in the form of machine (or processor) executable code and/or associated data that is carried on or embodied in a type of machine readable medium. Machine-executable code can be stored on an electronic storage unit, such as

memory (e.g., read-only memory, random-access memory, flash memory) or a hard disk.

“Storage” type media can include any or all of the tangible memory of the computers, processors or the like, or associated modules thereof, such as various semiconductor memories, tape drives, disk drives and the like, which may provide non-transitory storage at any time for the software programming. All or portions of the software may at times be communicated through the Internet or various other telecommunication networks. Such communications, for example, may enable loading of the software from one computer or processor into another, for example, from a management server or host computer into the computer platform of an application server. Thus, another type of media that may bear the software elements includes optical, electrical and electromagnetic waves, such as used across physical interfaces between local devices, through wired and optical landline networks and over various air-links. The physical elements that carry such waves, such as wired or wireless links, optical links or the like, also may be considered as media bearing the software. As used herein, unless restricted to non-transitory, tangible “storage” media, terms such as computer or machine “readable medium” refer to any medium that participates in providing instructions to a processor for execution.

[00273] Hence, a machine readable medium, such as computer-executable code, may take many forms, including but not limited to, a tangible storage medium, a carrier wave medium or physical transmission medium. Non-volatile storage media include, for example, optical or magnetic disks, such as any of the storage devices in any computer(s) or the like, such as may be used to implement the databases, etc. shown in the drawings. Volatile storage media include dynamic memory, such as main memory of such a computer platform. Tangible transmission media include coaxial cables; copper wire and fiber optics, including the wires that comprise a bus within a computer system. Carrier-wave transmission media may take the form of electric or electromagnetic signals, or acoustic or light waves such as those generated during radio frequency (RF) and infrared (IR) data communications. Common forms of computer-readable media therefore include for example: a floppy disk, a flexible disk, hard disk, magnetic tape, any other magnetic medium, a CD-ROM, DVD or DVD-ROM, any other optical medium, punch cards paper tape, any other physical storage medium with patterns of holes, a RAM, a ROM, a PROM and EPROM, a FLASH-EPROM, any other memory chip or cartridge, a carrier wave transporting data or instructions, cables or links transporting such a carrier wave, or any other medium from which a computer may read programming code and/or data. Many of these forms of computer readable media may be involved in carrying one or more sequences of one or more instructions to a processor for execution.

[00274] The computer system **901** can include or be in communication with an electronic display **935** that comprises a user interface (UI) **940** for providing, for example, results of

sequencing analysis, correlating sequencing reads to areas of accessible chromatin, etc.

Examples of UIs include, without limitation, a graphical user interface (GUI) and web-based user interface.

[00275] Methods and systems of the present disclosure can be implemented by way of one or more algorithms. An algorithm can be implemented by way of software upon execution by the central processing unit **905**. The algorithm can, for example, perform sequencing reactions, correlating sequencing reads to areas of accessible chromatin, etc.

[00276] Devices, systems, compositions and methods of the present disclosure may be used for various applications, such as, for example, processing a single analyte (e.g., RNA, DNA, or protein) or multiple analytes (e.g., DNA and RNA, DNA and protein, RNA and protein, or RNA, DNA and protein) from a single cell. For example, a biological particle (e.g., a cell or cell bead) is partitioned in a partition (e.g., droplet), and multiple analytes from the biological particle are processed for subsequent processing. The multiple analytes may be from the single cell. This may enable, for example, simultaneous proteomic, transcriptomic and genomic analysis of the cell.

Examples

[00277] Although one or more of the Examples herein make use of partitions that comprise droplets in an emulsion (e.g., droplet emulsion partitions), any of the above-described partitions (such as wells) can be utilized in the methods, systems, and compositions described below.

Example 1. Generation of Barcoded Nucleic Acid Fragments from Single Cells using Forked Adaptors Comprising Transposon End Sequences

[00278] A plurality of transposase molecules, a plurality of cells of interest (or a plurality of nuclei harvested from cells of interest, *e.g.*, by nonionic detergents such as NP-40 (IGEPAL CA-630) or Triton X-100), and a plurality of barcode oligonucleotides (e.g., nucleic acid barcode molecules) are partitioned such that at least some partitions (e.g., droplets or wells) comprise a plurality of transposase molecules, a single cell (or nucleus), and a plurality of barcode oligonucleotides comprising a sequencing primer sequence, a barcode sequence, and a transposon end sequence. In some embodiments, the plurality of barcode oligonucleotides are attached to a solid or semi-solid particle such as a gel bead and partitioned such that at least some partitions (e.g., droplets or wells) comprise transposase molecules, a single cell (or nucleus), and support (e.g., a single gel bead).

[00279] For example, in some embodiments, aqueous droplet emulsion partitions are generated as described herein such that at least some of the droplets formed will comprise transposase molecules, cell lysis reagents, a single cell, and a single bead (e.g., gel bead)

comprising a plurality of barcoded forked adaptor oligonucleotides. See, *e.g.*, **FIG. 10**. The cells are then lysed within the droplets in a manner that releases template nucleic acid molecules from cell nuclei into their respective droplets, but that substantially maintains native chromatin organization. Droplets are then generally processed as outlined in **FIG. 11A**.

[00280] Specifically, a droplet is subjected to conditions such that the barcoded forked adaptor oligonucleotides are released from the bead (*e.g.*, gel bead) into the aqueous droplet (*e.g.*, by depolymerization or degradation of the bead using a reducing agent, such as DTT). Although the forked adaptors can be prepared in a variety of different configurations, an exemplary forked adaptor is illustrated in **FIG. 12A** and shows a partially complementary double-stranded oligonucleotide comprising a first oligonucleotide strand releasably attached to a bead (*e.g.*, gel bead) and a second partially complementary oligonucleotide strand. With continued reference to **FIG. 12A**, the first strand comprises a transposon end sequence (“mosaic end” or “ME”), a barcode sequence (“BC”), and a sequencing primer sequence (“R1”). The partially complementary second strand comprises: (i) a region fully complementary to the transposon end sequence (“ME_{rc}”); (ii) a region fully complementary to the barcode sequence (“BC_{rc}”); and (iii) a primer sequence (“R2”) partially complementary to the first strand primer sequence.

[00281] In alternative embodiments (*e.g.*, **FIG. 12B**), the double-stranded forked adaptor of **FIG. 12A** further comprises: (a) a first oligonucleotide strand further comprising a P5 sequence releasably attached to the gel bead; and (b) a second partially complementary oligonucleotide strand further comprising an index sequence (“i7”) and a P7 sequence.

[00282] After forked adaptors are released from a bead (*e.g.*, gel bead) into a droplet, the droplet is subjected to conditions such that a transposase-nucleic acid complex is formed comprising a transposase molecule and two forked adaptors. See **FIG. 11A**, step 2. The droplets are then subjected to conditions such that the transposase-nucleic acid complexes integrate the transposon end sequences into the template nucleic acid and fragment the template nucleic acid into double-stranded template nucleic acid fragments flanked by the forked adaptors. See **FIG. 11A**, step 3. In alternative embodiments, cells (or nuclei) are permeabilized/permeable and the transposase-nucleic acid complexes enter the nucleus to fragment the template nucleic acid. Cells are then lysed to release double-stranded template nucleic acid fragments. Because the transposase-nucleic acid complex can only act on a nucleosome-free template, the double-stranded template nucleic acid fragments are representative of genome-wide areas of accessible chromatin in a single cell or nucleus.

[00283] The double-stranded template nucleic acid fragments are then collected from the droplets and processed in bulk to fill any gaps created from the transposition reaction and to

generate a library suitable for next generation high throughput sequencing (*e.g.*, subjecting the fragments, or derivatives thereof, to one or more reactions (*e.g.*, nucleic acid amplification) to add functional sequences to facilitate Illumina sequencing; see **FIG. 11B**). The fully constructed library is then sequenced according to any suitable sequencing protocol.

Example 2. Generation of Barcoded Nucleic Acid Fragments from Single Cells using Forked Adaptors and Transposase-Nucleic Acid Complexes

[00284] A plurality of transposase-nucleic acid complexes, a plurality of cells of interest (or a plurality of nuclei harvested from cells of interest), and a plurality of barcode oligonucleotides are partitioned such that at least some partitions (*e.g.*, droplets or wells) comprise a plurality of transposase-nucleic acid complexes, a single cell (or nucleus), and a plurality of barcode oligonucleotides (*e.g.*, nucleic acid barcode molecules) comprising a sequencing primer sequence and a barcode sequence. In some embodiments, the plurality of barcode oligonucleotides are attached to a solid or semi-solid particle (*e.g.*, a bead, such as a gel bead) and partitioned such that at least some partitions (*e.g.*, droplets or wells) comprise transposase-nucleic acid complexes, a single cell (or nucleus), and a single solid or semi-solid particle (*e.g.*, gel bead). In alternative embodiments, a plurality of transposase molecules and a plurality of transposon end sequence oligonucleotides are partitioned along with a single cell (or nucleus) and the barcode oligonucleotides and transposase-nucleic acid complexes are generated in the partition (*e.g.*, droplet or well).

[00285] For example, in some embodiments, droplet emulsion partitions are generated as described herein such that at least some of the droplets comprise transposase-nucleic acid complexes, cell lysis reagents, T4 DNA ligase, a single cell or cell nucleus, and a single bead (*e.g.*, gel bead) comprising a plurality of barcoded forked adapter oligonucleotides. See, *e.g.*, **FIG. 13**.

[00286] An individual transposase-nucleic acid complex comprises a transposase and a pair of double-stranded oligonucleotides each comprising a transposon end sequence (*e.g.*, an ME sequence). See **FIGs. 13-15**. In some embodiments, the double-stranded transposon-end sequence containing oligonucleotides further comprise a spacer sequence.

[00287] After droplet formation, single cells, if present, are lysed in a manner that releases template nucleic acid molecules from the nucleus into the droplet, but that substantially maintains native chromatin organization. Droplets are then generally processed as outlined in **FIG. 14A**.

[00288] Specifically, a droplet is subjected to conditions such that the forked adaptors are released from the bead (*e.g.*, gel bead) into the aqueous droplet (*e.g.*, by depolymerization or

degradation of the bead using a reducing agent, such as DTT). See **FIG. 14A**, step 1a. Although the forked adaptors can be prepared in a variety of different configurations, an exemplary forked adaptor is illustrated in **FIG. 15A** and shows a double-stranded oligonucleotide comprising a first oligonucleotide strand releasably attached to a bead (e.g., gel bead) and a second partially complementary oligonucleotide strand. With continued reference to **FIG. 15A**, the first strand comprises a barcode sequence (“BC”), a primer sequence (“R1”), and a transposon end sequence (“mosaic end” or “ME”). The partially complementary second strand comprises a region fully complementary to the barcode sequence (“BC_{rc}”), a primer sequence (“R2”) partially complementary to the first strand primer sequence, and a region fully complementary to the transposon end sequence (“ME_{rc}”). In some embodiments, the first strand further comprises a phosphorothioate linkage in the terminal nucleotide at the 3’ end. In other embodiments, the first strand comprises phosphorothioate linkages in the last 3-5 nucleotides at the 3’ end. In still other embodiments, the first strand comprises phosphorothioate linkages throughout the first strand. The first oligonucleotide strand may further comprise a P5 adapter sequence releasably attached to the bead (e.g., gel bead). The second partially complementary oligonucleotide strand may further comprise an index primer (“i7”) and an adaptor sequence different than the first strand (“P7”). Similarly, **FIG. 15B** shows a double-stranded oligonucleotide comprising a first oligonucleotide strand releasably attached to a bead (e.g., gel bead) and a second partially complementary oligonucleotide strand. With continued reference to **FIG. 15B**, the first strand comprises a barcode sequence (“BC”) and a primer sequence (“R1”). The partially complementary second strand comprises a region fully complementary to the barcode sequence (“BC_{rc}”) and a primer sequence (“R2”) partially complementary to the first strand primer sequence. In some embodiments, the first strand further comprises a phosphorothioate linkage in the terminal nucleotide at the 3’ end. In other embodiments, the first strand comprises phosphorothioate linkages in the last 3-5 nucleotides at the 3’ end. In still other embodiments, the first strand comprises phosphorothioate linkages throughout the first strand. The first oligonucleotide strand may further comprise a P5 adapter sequence releasably attached to the bead (e.g., gel bead). The second partially complementary oligonucleotide strand may further comprise an index primer (“i7”) and an adaptor sequence different than the first strand (“P7”).

[00289] After forked adaptors are released from a bead (e.g., gel bead) into a droplet, the droplet is subjected to conditions such that the transposase-nucleic acid complexes integrate the transposon end sequences into the template nucleic acid and fragment the template nucleic acid into double-stranded template nucleic acid fragments flanked by transposon end sequences. See **FIG. 14A**, step 1b. In alternative embodiments, cells (or nuclei) are permeabilized/permeable and the transposase-nucleic acid complexes enter the nuclei to fragment the template nucleic

acids therein. Cells, if present, are then lysed to release the double-stranded template nucleic acid fragments. Because the transposase-nucleic acid complex can only act on a nucleosome-free template, the double-stranded template nucleic acid fragments are representative of genome-wide areas of accessible chromatin in a single cell or nucleus. The forked adaptors are then ligated onto the ends of the double-stranded template nucleic acid fragments. See **FIG. 14A**, step 2.

[00290] The double-stranded template nucleic acid fragments are then collected from the droplets and processed in bulk to fill any gaps created from the transposition reaction and to generate a library suitable for next generation high throughput sequencing (*e.g.*, subjecting the fragments, or derivatives thereof, to one or more reactions (*e.g.*, nucleic acid amplification) to add functional sequences to facilitate Illumina sequencing; see **FIG. 14B**). The fully constructed library is then sequenced according to any suitable sequencing protocol. In some embodiments, custom sequencing primers directed against the spacer-ME sequence are utilized to avoid sequencing the barcode-spacer-ME region of the library.

Example 3. Generation of Barcoded Nucleic Acid Fragments from Single Cells using Adaptors Comprising a T7 Promoter Sequence

[00291] A plurality of transposase molecules, a plurality of cells of interest (or a plurality of nuclei harvested from cells of interest), and a plurality of barcode oligonucleotides are partitioned such that at least some partitions (*e.g.*, droplets or wells) comprise a plurality of transposase molecules, a single cell (or nucleus), and a plurality of barcode oligonucleotides (*e.g.*, nucleic acid barcode molecules) comprising a T7 promoter sequence, a sequencing primer sequence, a barcode sequence, and a transposon end sequence. In some embodiments, the plurality of barcode oligonucleotides are attached to a solid or semi-solid particle (*e.g.*, a bead, such as a gel bead) and partitioned such that at least some partitions (*e.g.*, droplets or wells) comprise transposase molecules, a single cell (or nucleus), and a single solid or semi-solid particle (*e.g.*, gel bead).

[00292] For example, in some embodiments, droplet emulsion partitions are generated as previously described such that at least some droplets (*e.g.*, aqueous droplets) that comprise transposase molecules, cell lysis reagents, a single cell (or nucleus), and a single bead (*e.g.*, gel bead) comprising partially double-stranded T7 promoter oligonucleotide adaptors. See, *e.g.*, **FIG. 16**. The cells, if present, are then lysed in a manner that releases template nucleic acid molecules from the nucleus into the droplet, but that substantially maintains native chromatin organization. Droplets are then generally processed as outlined in **FIG. 17**.

[00293] Specifically, a droplet (e.g., an aqueous droplet) is subjected to conditions such that the partially double-stranded adaptors are released from the bead (e.g., gel bead) into the droplet (e.g., by depolymerization or degradation of the bead using a reducing agent, such as DTT). See **FIG. 17**, step 1. Although the partially double-stranded adaptors can be prepared in a variety of different configurations, an exemplary partially double-stranded adaptor is illustrated in **FIG. 18** and shows a partially double-stranded oligonucleotide comprising a first oligonucleotide strand releasably attached to a bead (e.g., a gel bead) and a second, shorter complementary oligonucleotide strand. With continued reference to **FIG. 18**, the first strand comprises a transposon end (“mosaic end” or “ME”) sequence, a barcode sequence (“BC”), a partial sequencing primer sequence (“pR1” or “Partial R1”), and a T7 promoter sequence while the second oligonucleotide strand comprises a sequence fully complementary to the transposon end sequence (“ME_{rc}”).

[00294] After partially double-stranded adaptors are released from a bead (e.g., gel bead) into a droplet, the droplet is subjected to conditions such that a transposase-nucleic acid complex is formed comprising a transposase molecule and two partially double-stranded oligonucleotide adaptors. See **FIG. 17**, step 2. The droplet or plurality of droplets is then subjected to conditions such that the transposase-nucleic acid complexes integrate the adaptors into the template nucleic acid and produce double-stranded template nucleic acid fragments flanked by the partially double-stranded adaptors. See **FIG. 17**, step 3. In alternative embodiments, cells (or nuclei) are permeabilized/permeable and the transposase-nucleic acid complexes enter the nuclei to fragment the template nucleic acids therein. Cells, if present, are then lysed to release the double-stranded template nucleic acid fragments. Because the transposase-nucleic acid complex can only act on a nucleosome-free template, the double-stranded template nucleic acid fragments are representative of genome-wide areas of accessible chromatin in a single cell or nucleus.

[00295] The double-stranded template nucleic acid fragments are then collected from the droplets and processed in bulk to fill any gaps created from the transposition reaction. RNA is generated from the double-stranded template nucleic acid fragments using an *in vitro* transcription reaction and T7 RNA polymerase. RNA is collected and purified, followed by first and second strand cDNA synthesis. Double-stranded cDNA molecules are then further processed (including fragmentation and adaptor insertion by, e.g., a second transposase-mediated fragmentation) to generate a library suitable for next generation high throughput sequencing (e.g., subjecting the fragments, or derivatives thereof, to one or more reactions (e.g., nucleic acid amplification) to add functional sequences to facilitate Illumina sequencing). The fully constructed library is then sequenced according to any suitable sequencing protocol.

Example 4. Generation of Barcoded Nucleic Acid Fragments from Single Cells using Transposition of Sequencing Adaptors followed by Random Priming and Extension

[00296] A plurality of transposase molecules, a plurality of cells of interest (or a plurality of nuclei harvested from cells of interest), and a plurality of barcode oligonucleotides are partitioned such that at least some partitions (e.g., droplets or wells) comprise a plurality of transposase molecules, a single cell (or nucleus), and a plurality of barcode oligonucleotides (e.g., nucleic acid barcode molecules) comprising a sequencing primer sequence, a barcode sequence, and a transposon end sequence. In some embodiments, the plurality of barcode oligonucleotides are attached to a solid or semi-solid particle (e.g., a bead, such as a gel bead) and partitioned such that at least some partitions (e.g., droplets or wells) comprise transposase molecules, a single cell (or nucleus), and a single solid or semi-solid particle (e.g., gel bead).

[00297] For example, in some embodiments, droplet emulsion partitions are generated as previously described such that at least some droplets that comprise transposase molecules, cell lysis reagents, a single cell (or nucleus), and a single bead (e.g., gel bead) comprising partially double-stranded barcoded oligonucleotide adaptors. See, e.g., **FIG. 19**. The cells, if present, are then lysed in a manner that releases template nucleic acid molecules from the nucleus into the aqueous droplet, but that substantially maintains native chromatin organization. Droplets are then generally processed as outlined in **FIG. 20**.

[00298] Specifically, a droplet (e.g., an aqueous droplet) is subjected to conditions such that partially double-stranded adaptors are released from a bead (e.g., gel bead) into the droplet (e.g., by depolymerization or degradation of the bead using a reducing agent, such as DTT). See **FIG. 20**, step 1. Although the partially double-stranded adaptors can be prepared in a variety of different configurations, an exemplary partially double-stranded adaptor is illustrated in **FIG. 21** and shows a partially double-stranded oligonucleotide comprising a first oligonucleotide strand releasably attached to a bead (e.g., gel bead) and a second, shorter complementary oligonucleotide strand. With continued reference to **FIG. 21**, the first strand comprises a transposon end (“mosaic end” or “ME”) sequence, a barcode sequence (“BC”), and a partial sequencing primer sequence (“pR1” or “Partial R1”) while the second oligonucleotide strand comprises a sequence fully complementary to the transposon end sequence (“ME_{rc}”).

[00299] After partially double-stranded adaptors are released from a bead (e.g., gel bead) into a droplet (e.g., an aqueous droplet), the droplet is subjected to conditions such that a transposase-nucleic acid complex is formed comprising a transposase molecule and two partially double-stranded oligonucleotides. See **FIG. 20**, step 2. The droplets are then subjected to conditions such that the transposase-nucleic acid complexes integrate the adaptors into the template nucleic acid and produce double-stranded template nucleic acid fragments flanked by the partially

double-stranded adaptors. See **FIG. 20**, step 3. In alternative embodiments, cells (or nuclei) are permeabilized/permeable and the transposase-nucleic acid complexes enter the nucleus to fragment the template nucleic acid. Cells, if present, are then lysed to release the double-stranded template nucleic acid fragments. Because the transposase-nucleic acid complex can only act on a nucleosome-free template, the double-stranded template nucleic acid fragments are representative of genome-wide areas of accessible chromatin in a single cell or nucleus.

[00300] The double-stranded template nucleic acid fragments are then collected from the droplets and processed in bulk to generate a library suitable for next generation high throughput sequencing. In some embodiments, for example, double-stranded template nucleic acid fragments are processed in bulk in a random priming extension reaction. See, *e.g.*, **FIG. 22**. The random extension primer has a sequence of random nucleotides (N-mer) and, for example, can be attached to second PCR handle (*e.g.*, partial R2 sequence, see **FIG. 22**). The random extension primers are annealed to the double-stranded template nucleic acid fragments, or derivatives thereof, and extended (*e.g.*, **FIG. 22**). Reactions can then be cleaned-up and extension products subjected to one or more reactions (*e.g.*, nucleic acid amplification) to add functional sequences to facilitate Illumina sequencing (*e.g.*, **FIG. 22**). The fully constructed library is then sequenced according to any suitable sequencing protocol.

Example 5. Generation of Barcoded Nucleic Acid Fragments from Single Cells using Transposase-Nucleic Acid Complexes and Barcoded Adaptors

[00301] In some embodiments, artificial transposons are designed to insert sequences of interest into a target DNA molecule (*e.g.*, open chromatin). For example, an artificial transposon oligonucleotide comprising a barcode sequence and an adapter sequence is flanked by a transposon end sequence on each end of the oligonucleotide (see, *e.g.*, **FIG. 23**). A plurality of transposase molecules, a plurality of cells of interest (or a plurality of nuclei harvested from cells of interest), and a plurality of artificial transposon oligonucleotides are partitioned such that at least some partitions (*e.g.*, droplets or wells) comprise a plurality of artificial transposon oligonucleotides, a plurality of transposase molecules, and a single cell (or nucleus). In some embodiments, the plurality of artificial transposon oligonucleotides are attached to a solid or semi-solid particle (*e.g.*, bead, such as a gel bead) and partitioned such that at least some partitions (*e.g.*, droplets or wells) comprise a plurality of transposase molecules, a single cell (or nucleus), and a single solid or semi-solid particle (*e.g.*, gel bead). In other embodiments, a plurality of transposon nucleic acid complexes comprising an artificial transposon oligonucleotide comprising a barcode sequence and an adapter sequence flanked by transposon end sequenced are partitioned such that at least some partitions comprise a plurality of

transposon nucleic acid complexes and a single cell (or nucleus). In some embodiments, the plurality of artificial transposon oligonucleotides are attached to a solid or semi-solid particle (e.g., bead, such as a gel bead) and partitioned such that at least some partitions (e.g., droplets or wells) comprise a single cell (or nucleus) and a single solid or semi-solid particle (e.g., gel bead).

[00302] For example, in some embodiments, droplet emulsion partitions are generated as previously described such that at least some droplets that comprise transposase molecules, cell lysis reagents, a single cell (or nucleus), and a single bead (e.g., gel bead). The cells are then lysed to release template nucleic acid molecules from the nucleus into the aqueous droplet. Droplets are then generally processed as outlined in **FIG. 23**.

[00303] Specifically, a droplet (e.g., aqueous droplet) is subjected to conditions such that barcoded adaptors are released from a bead (e.g., gel bead) into the droplet (e.g., by depolymerization or degradation of the bead using a reducing agent, such as DTT). Although the barcoded adaptors can be prepared in a variety of different configurations, an exemplary barcoded adaptor is illustrated in **FIG. 23** and is a double-stranded oligonucleotide releasably attached to a bead (e.g., gel bead), wherein the barcoded adaptor comprises a pair of transposon end (“mosaic end” or “ME”) sequences flanking a barcode sequence (“BC”) and an adaptor sequence (“P5”).

[00304] After barcoded adaptors are released from a bead (e.g., gel bead) into a droplet, the droplet is subjected to conditions such that a transposase-nucleic acid complex is formed comprising a transposase molecule and a barcoded adaptor comprising a pair of transposon end sequences. See **FIG. 23**. The droplet or plurality of droplets is then subjected to conditions such that the transposase-nucleic acid complexes integrate the barcoded adaptors into the template nucleic acid. See **FIG. 23**. In alternative embodiments, cells (or nuclei) are permeabilized/permeable and the transposase-nucleic acid complexes enter the nuclei to perform the transposition reaction. Cells, if present, are then lysed to release the transposon-containing template nucleic acid fragments.

[00305] The barcode-transposed template nucleic acids are then collected from the droplets and processed in bulk to fragment the barcode-transposed template nucleic acids and to generate a library suitable for next generation high throughput sequencing (e.g., subjecting the fragments, or derivatives thereof, to one or more reactions (e.g., nucleic acid amplification) to add functional sequences to facilitate Illumina sequencing). The fully constructed library is then sequenced according to any suitable sequencing protocol.

[00306]

Example 6. Generation of Barcoded Nucleic Acid Fragments from Single Cells using Gel Bead-Functionalized Transposase-Nucleic Acid Complexes

[00307] A plurality of transposase nucleic acid complexes and a plurality of cells of interest (or a plurality of nuclei harvested from cells of interest) are partitioned such that at least some partitions (e.g., droplets or wells) comprise a single cell (or nucleus) and a plurality of transposase nucleic acid complexes comprising a transposase molecule and a barcode oligonucleotide (e.g., nucleic acid barcode molecule) comprising a sequencing primer sequence, a barcode sequence, and a transposon end sequence (see, *e.g.*, **FIGs. 24A-24B** and **25A-25B**). In some embodiments, the plurality of transposase nucleic acid complexes are attached to a solid or semi-solid particle (e.g., bead, such as a gel bead) (see, *e.g.*, **FIGs. 24A-24B** and **26**) and partitioned such that at least some partitions (e.g., droplets or wells) comprise a single cell (or nucleus) and a single solid or semi-solid particle (e.g., gel bead).

[00308] For example, in some embodiments, droplet emulsion partitions are generated as previously described such that at least some droplets (e.g., aqueous droplets) comprise cell lysis reagents, a single cell (or nucleus), and a single bead (e.g., gel bead) functionalized with a transposase-nucleic acid complex. The cells are then lysed in a manner that releases template nucleic acid molecules from the nuclei into the respective droplets, but that substantially maintains native chromatin organization.

[00309] Droplets (e.g., aqueous droplets) are then subjected to conditions such that the transposase-nucleic acid complexes are released from beads (e.g., gel beads) into the respective droplets (*e.g.*, by depolymerization or degradation of beads using a reducing agent, such as DTT). Although the transposase-nucleic acid complexes can be prepared in a variety of different configurations, a transposase-nucleic acid complex is illustrated in **FIG. 24A** and shows a complex comprising a transposase, a first partially double-stranded oligonucleotide, and a second partially double-stranded oligonucleotide. Continuing with the embodiment of **FIG. 24A**, the first partially double-stranded oligonucleotide comprises: (a) a first strand releasably attached to a bead (e.g., gel bead), wherein the first strand comprises a transposon end sequence ("ME"), a barcode sequence ("BC"), and a first sequencing primer sequence ("R1"); and (b) a second strand complementary to the transposon end sequence of the first oligonucleotide strand ("ME_{rc}"). With continued reference to **FIG. 24A**, the second partially double-stranded oligonucleotide comprises: (a) a first oligonucleotide strand comprising a transposon end sequence ("ME") and a second primer sequence ("R2"); and (b) a second strand complementary to the transposon end sequence ("ME_{rc}"). See also **FIG. 24B**, which comprises an identical first partially double-stranded oligonucleotide as the above described embodiment and a second partially double-stranded oligonucleotide comprising: (a) a first oligonucleotide strand

comprising a transposon end sequence (“ME”), a barcode sequence (“BC”), and the first primer sequence (“R1”); and (b) a second strand complementary to the transposon end sequence of the second oligonucleotide strand (“ME_{rc}”).

[00310] Alternatively, bead- (e.g., gel bead) functionalized transposase-nucleic acid complexes are prepared as illustrated in **FIG. 26**, which shows a complex comprising a transposase, a first partially double-stranded oligonucleotide and a second double-stranded oligonucleotide. In this embodiment, the first partially double-stranded oligonucleotide comprises: (a) a first strand releasably attached to a bead (e.g., gel bead), wherein the first strand comprises a transposon end sequence (“ME”) and a barcode sequence (“BC”) and (b) a second strand complementary to the transposon end sequence of the first oligonucleotide strand (“ME_{rc}”) while the second double-stranded oligonucleotide comprises: (a) a first strand releasably attached to a bead (e.g., gel bead), wherein the first strand comprises a transposon end sequence (“ME”) and (b) a second strand complementary to the first oligonucleotide strand (“ME_{rc}”). Alternative embodiments of **FIG. 26** comprise additional functional sequences, such as a sequencing primer sequence (e.g., R1 and/or R2) or an adapter sequence (e.g., P5 and/or P7).

[00311] In other embodiments, droplets are partitioned such that at least some droplets comprise cell lysis reagents, a plurality of transposase molecules, a single cell (or nucleus), and a single bead (e.g., gel bead) comprising a barcode oligonucleotide (e.g., nucleic acid barcode molecule) comprising a barcode sequence (“BC”) and a transposon end sequence (“ME”). See, e.g., **FIGs. 25A-25B**. Droplets are then subjected to conditions such that transposase nucleic acid complexes comprising a transposase molecule and a barcode oligonucleotide are formed in the partition.

[00312] After a transposase-nucleic acid complex is released from a bead (e.g., gel bead) into a droplet (or is formed in the partition in embodiments such as **FIGs. 25A-25B**), the droplet is subjected to conditions such that the transposase-nucleic acid complexes integrate the transposon end sequences into the template nucleic acid and fragments the template nucleic acid into double-stranded template nucleic acid fragments flanked by first and second partially double-stranded oligonucleotides. In alternative embodiments, cells (or nuclei) are permeabilized/permeable and the transposase-nucleic acid complexes enter the nuclei to fragment the template nucleic acids therein. Cells, if present, are then lysed to release the double-stranded template nucleic acid fragments. Because the transposase-nucleic acid complex can only act on a nucleosome-free template, the double-stranded template nucleic acid fragments are representative of genome-wide areas of accessible chromatin in a single cell or nucleus.

[00313] The double-stranded template nucleic acid fragments are then collected from the droplets and processed in bulk to fill any gaps created from the transposition reaction and to generate a library suitable for next generation high throughput sequencing (*e.g.*, subjecting the fragments, or derivatives thereof, to one or more reactions (*e.g.*, nucleic acid amplification) to add functional sequences to facilitate Illumina sequencing; see, *e.g.*, **FIG. 27**). The fully constructed library is then sequenced according to any suitable sequencing protocol.

Example 7. Generation of Barcoded Nucleic Acid Fragments from Single Cells using Transposase-Nucleic Acid Complexes and Barcoded Adaptors (Two-Step Approach)

[00314] A plurality of transposase nucleic acid complexes and a plurality of cells of interest (or a plurality of nuclei harvested from cells of interest) are partitioned such that at least some partitions (*e.g.*, droplets or wells) comprise a single cell (or nucleus), a plurality of transposase nucleic acid complexes comprising a transposon end sequence, and a plurality of barcoded oligonucleotides (*e.g.*, nucleic acid barcode molecules) comprising a barcode sequence and a sequencing primer sequence (see, *e.g.*, **FIGs. 28A-28B**). In some embodiments, the plurality of barcode oligonucleotides further comprise a transposon end sequence. In some embodiments, the plurality of barcode oligonucleotides are attached to a solid or semi-solid particle (*e.g.*, gel bead) (see, *e.g.*, **FIG. 28B**) and partitioned such that at least some partitions (*e.g.*, droplets or wells) comprise a plurality of transposase nucleic acid complexes, a single cell (or nucleus), and a single solid or semi-solid particle (*e.g.*, gel bead).

[00315] For example, in some embodiments, droplet emulsion partitions are generated as previously described such that at least some droplets (*e.g.*, aqueous droplets) comprise a transposase-nucleic acid complex comprising a transposase and a pair of double-stranded oligonucleotides, cell lysis reagents, a single cell (or nucleus), and a single bead (*e.g.*, gel bead) comprising a barcoded adaptor. Although the transposase-nucleic acid complexes can be prepared in a variety of different configurations, a transposase-nucleic acid complex is illustrated in **FIG. 28A** and shows a complex comprising a transposase, a first double-stranded oligonucleotide comprising a transposon end (“mosaic end” or “ME”) sequence, and a second double-stranded oligonucleotide comprising a transposon end (“mosaic end” or “ME”) sequence.

[00316] Cells, if present, are lysed in a manner that releases template nucleic acid molecules from nuclei into droplets (*e.g.*, aqueous droplets), but that substantially maintains native chromatin organization. Droplets are then subjected to conditions such that barcoded adaptors are released from beads (*e.g.*, gel beads) into the respective droplets. Although the barcoded adaptors can be prepared in a variety of different configurations, an exemplary barcoded adaptor is illustrated in **FIG. 28B** and shows a single-stranded oligonucleotide comprising a transposon

end (“mosaic end” or “ME”) sequence, a barcode sequence (“BC”), and a primer sequence (“R1”) releasably attached to a bead (e.g., gel bead).

[00317] After barcoded adaptors are released from a bead (e.g., gel bead) into a droplet, the droplet is subjected to conditions such that the transposase-nucleic acid complexes integrate the transposon end sequences into the template nucleic acid and fragment the template nucleic acid into double-stranded template nucleic acid fragments. In alternative embodiments, cells (or nuclei) are permeabilized/permeable and the transposase-nucleic acid complexes enter the nuclei to fragment the template nucleic acids therein. Cells, if present, are then lysed to release the double-stranded template nucleic acid fragments. Because the transposase-nucleic acid complex can only act on a nucleosome-free template, the double-stranded template nucleic acid fragments are representative of genome-wide areas of accessible chromatin in a single cell or nucleus. After transposition and fragmentation, a PCR reaction is performed to fill any gaps created from the transposition reaction and to add the barcoded adaptors to the ends of the double-stranded template nucleic acid fragments.

[00318] The double-stranded template nucleic acid fragments are then collected from the droplets and processed in bulk to generate a library suitable for next generation high throughput sequencing (e.g., subjecting the fragments, or derivatives thereof, to one or more reactions (e.g., nucleic acid amplification) to add functional sequences to facilitate Illumina sequencing). The fully constructed library is then sequenced according to any suitable sequencing protocol.

Example 8. Generation of Barcoded Nucleic Acid Fragments using Transposase-Nucleic Acid Complexes Assembled in a Single Reaction Step

[00319] Traditional tube-based implementations of Tn5-based tagmentation systems typically rely upon sample processing steps that take place in two independent reactions to generate the final transposase-fragmented nucleic acid sample. For example, in Reaction #1, oligonucleotide adaptors containing the Tn5 transposon end sequences and the Tn5 transposase enzyme are incubated to form a transposase-nucleic acid complex. Typically, magnesium (or other divalent cations) are omitted from the reaction buffer to keep the transposases catalytically inactive. In Reaction #2, the transposase-nucleic acid complex from Reaction #1 is combined with a target double-stranded DNA and an appropriate reaction buffer containing magnesium (or other divalent cations) to activate the transposase-nucleic acid complex and cause fragmentation of the target DNA and ligation of the adapter oligonucleotide sequences. While the above-described serial reaction workflow is straightforward, implementing a tagmentation reaction within a single reaction or reaction vessel (“one-pot reaction”) can be complicated.

[00320] A one-pot transposition reaction was performed on 50,000 HEK293T cells and compared to a traditional two-step tagmentation reaction. Sequencing libraries were prepared from each reaction type and sequenced on an Illumina sequencer. Sequencing data is presented in Table 1 below and demonstrates comparable sequencing metrics between the traditional and one-pot tagmentation methods.

Table 1. Results from One-Pot Sequencing Reactions

Sequencing metrics	Traditional	One-pot
Total gb	43	31
Library complexity	1.1	0.52
Median insert size	160	180
Unmapped fraction	0.014	0.034
Non-nuclear read fraction	0.53	0.26
Fraction of fragments on target	0.57	0.81
Mean duplication rate	1.5	1.8
Percent reads gt 10bp soft clipped	0.25	0.17

[00321] The transposition methods described herein can be utilized in a one-pot reaction as described above. These one-pot reactions can be done either in bulk or in discrete partitions, such as a well or droplet.

Example 9. Generation of Barcoded Nucleic Acid Fragments using Bulk Tagmentation and Barcoding by Ligation in Partitions (Variant 1)

[00322] Intact nuclei are harvested in bulk from cells in a cell population of interest in a manner that substantially maintains native chromatin organization (*e.g.*, using IGEPAL CA-630 mediated cell lysis). Nuclei are then incubated in the presence of a transposase-nucleic acid complex comprising a transposase molecule and two partially double-stranded adaptor oligonucleotides. See, *e.g.*, **FIG. 29A**. Alternatively, cells are permeable/permeabilized, allowing the transposase-nucleic acid complex to gain access to the nucleus. The first adapter oligonucleotide comprises a double stranded transposon end sequence (“ME”) and a single stranded Read1 sequencing primer sequence (“R1”) while the second adapter oligonucleotide comprises a double stranded transposon end sequence (“ME”) and a single stranded Read2 sequencing primer sequence (“R2”). See **FIG. 29A**. In some embodiments, the R1 and/or R2 sequencing primer in the first and/or second adapter oligonucleotide comprises a TruSeq R1 and/or R2 sequence, or a portion thereof. The transposase-nucleic acid complexes integrate the

adaptors into the template nucleic acid and produce template nucleic acid fragments flanked by the partially double-stranded adaptors. See **FIG. 29C**. Because the transposase-nucleic acid complex can only act on a nucleosome-free template, the fragmented template nucleic acid fragments are representative of genome-wide areas of accessible chromatin. In some embodiments, the transposase molecules are inactivated prior to further processing steps.

[00323] Nuclei (or cells) comprising the adapter-flanked template nucleic acid fragments are then partitioned into a plurality of partitions (e.g., droplets or wells) such that at least some partitions comprise (1) a single nucleus (or cell) comprising the adapter-flanked template nucleic acid fragments; and (2) a plurality of partially double-stranded barcode oligonucleotide molecules comprising a doubled stranded barcode sequence (“BC”), a double-stranded P5 adapter sequence (“P5”), and a single stranded sequence complementary to the Read 1 sequence (“R1_{rc}”). See **FIG. 29B**. In some embodiments, the partially double-stranded barcode oligonucleotide molecules are attached to a solid or semi-solid particle (e.g., a bead, such as a gel bead) and partitioned such that at least some partitions (e.g., droplets or wells) comprise (1) a single nucleus (or cell) and (2) a single solid or semi-solid particle (e.g., bead, such as a gel bead). In addition to the aforementioned components, in some embodiments, the plurality of partitions (e.g., droplets or wells) further comprises reagents (e.g., enzymes and buffers) that facilitate the reactions described below.

[00324] Single cell- or nucleus-containing partitions (e.g., droplets or wells) are then subjected to conditions to release adapter-flanked template nucleic acid fragments from the nuclei (e.g., cell lysis). In certain embodiments, where barcode oligonucleotides (e.g., nucleic acid barcode molecules) are attached to a bead (e.g., gel bead), partitions (e.g., droplets or wells) are subjected to conditions to cause release of the barcode oligonucleotide molecules from the bead (e.g., gel bead) (e.g., depolymerization or degradation of the beads, for example, using a reducing agent such as DTT). After release from single nuclei, the adapter-flanked template nucleic acid fragments are subjected to conditions to phosphorylate the 5' end of the Read 1 sequence (e.g., using T4 polynucleotide kinase) for subsequent ligation steps. After phosphorylation, the barcode oligonucleotide molecules are ligated onto the adapter-flanked template nucleic acid fragments using a suitable DNA ligase enzyme (e.g., T4 or *E. coli* DNA ligase) and the complementary Read 1 sequences in the barcode oligonucleotides and the adapter-flanked template nucleic acid fragments. See **FIG. 29C**.

[00325] After barcode ligation, gaps remaining from the transposition reaction are filled to produce barcoded, adapter-flanked template nucleic acid fragments. See **FIG. 29C**. The barcoded, adapter-flanked template nucleic acid fragments are then released from the partitions (e.g., droplets or wells) and processed in bulk to complete library preparation for next generation

high throughput sequencing (*e.g.*, to add sample index (SI) sequences (*e.g.*, “i7”) and/or further adapter sequences (*e.g.*, “P7”). In alternative embodiments, the gap filling reaction is completed in bulk after barcoded, adapter-flanked template nucleic acid fragments have been released from the partitions (*e.g.*, droplets or wells). The fully constructed library is then sequenced according to a suitable next-generation sequencing protocol (*e.g.*, Illumina sequencing).

Example 10. Generation of Barcoded Nucleic Acid Fragments using Tagmentation and Barcoding by Ligation in Partitions

[00326] Cells from a cell population of interest (or nuclei from cells in a cell population of interest) are partitioned into a plurality of partitions (*e.g.*, droplets or wells) such that at least some partitions comprise (1) a single cell (or a single nucleus) comprising a template nucleic acid; and (2) a plurality of partially double-stranded barcode oligonucleotide molecules (*e.g.*, nucleic acid barcode molecules) comprising a doubled stranded barcode sequence (“BC”), a doubled stranded P5 adapter sequence (“P5”), and a single stranded sequence complementary to a Read 1 sequence (“R1_{rc}”) (*e.g.*, **FIG. 29B**). In some embodiments, the partially double-stranded barcode oligonucleotide molecules are attached to a solid or semi-solid particle (*e.g.*, bead, such as a gel bead) and partitioned such that at least some partitions (*e.g.*, droplets or wells) comprise (1) a single cell (or a single nucleus) and (2) a single solid or semi-solid particle (*e.g.*, gel bead). In addition to the aforementioned components, in some embodiments, the plurality of partitions (*e.g.*, droplets or wells) further comprises reagents (*e.g.*, enzymes and buffers) that facilitate the reactions described below.

[00327] After partitioning into partitions (*e.g.*, droplets or wells), the single cells (or nuclei) are lysed to release the template genomic DNA in a manner that substantially maintains native chromatin organization. Partitions (*e.g.*, droplets or wells) are then subjected to conditions to generate a transposase-nucleic acid complex as described in Example 9 and shown in **FIG. 29A**. Alternatively, in some embodiments, a plurality of pre-formed transposase-nucleic acid complexes as shown in **FIG. 29A** are partitioned into the plurality of partitions (*e.g.*, droplets or wells). Partitions (*e.g.*, droplets or wells) are then subjected to conditions such that the transposase-nucleic acid complexes integrate the first and second adapter sequences into the template nucleic acid to generate double-stranded adapter-flanked template nucleic acid fragments. See **FIG. 29D**. Because the transposase-nucleic acid complex can only act on nucleosome-free DNA, the adapter-flanked template nucleic acid fragments are representative of genome-wide areas of accessible chromatin in a single cell or nucleus. Alternatively, in some embodiments, the tagmentation reaction is performed in intact nuclei, and the nuclei are lysed

after transposition to release the double-stranded adapter-flanked template nucleic acid fragments.

[00328] Samples are then processed generally as described in Example 9. In certain embodiments, where barcode oligonucleotides (e.g., nucleic acid barcode molecules) are attached to a solid or semi-solid particle (e.g., bead, such as a gel bead), droplets are subjected to conditions to cause release of the barcode oligonucleotide molecules from the solid or semi-solid particle (e.g., gel bead) (e.g., depolymerization or degradation of beads, for example, using a reducing agent such as DTT). In some embodiments, the transposase molecules are inactivated (e.g., by heat inactivation) prior to further processing steps. The adapter-flanked template nucleic acid fragments are subjected to conditions to phosphorylate the 5' end of the Read1 sequence (e.g., using T4 polynucleotide kinase) of the adapter-flanked template nucleic acid fragments. After phosphorylation, the barcode oligonucleotide molecules are ligated onto the adapter-flanked template nucleic acid fragments using a suitable DNA ligase enzyme (e.g., T4, 9°N, or *E. coli* DNA ligase) and the complementary Read1 sequences in the barcode oligonucleotides and the adapter-flanked template nucleic acid fragments. See **FIG. 29C**.

[00329] After barcode ligation, gaps remaining from the transposition reaction are filled to produce barcoded, adapter-flanked template nucleic acid fragments. See **FIG. 29C**. The barcoded, adapter-flanked template nucleic acid fragments are then released from the partitions (e.g., droplets or wells) and processed in bulk to complete library preparation for next generation high throughput sequencing (e.g., to add sample index (SI) sequences (e.g., "i7") and/or further adapter sequences (e.g., "P7")). In alternative embodiments, the gap filling reaction is completed in bulk after barcoded, adapter-flanked template nucleic acid fragments have been released from the droplets. The fully constructed library is then sequenced according to a suitable next-generation sequencing protocol (e.g., Illumina sequencing).

Example 11. Generation of Barcoded Nucleic Acid Fragments using Bulk Tagmentation and Barcoding by Linear Amplification in Partitions

[00330] Nuclei are harvested in bulk from cells in a cell population of interest in a manner that substantially maintains native chromatin organization. Alternatively, cells are permeabilized/permeable, allowing the transposase-nucleic acid complex to gain access to the nucleus. Nuclei (or permeabilized cells) are then incubated in the presence of a transposase-nucleic acid complex as described in Example 9. See **FIG. 29A**.

[00331] Nuclei (or cells) comprising the adapter-flanked template nucleic acid fragments are then partitioned into a plurality of partitions (e.g., droplets or wells) such that at least some partitions comprise (1) a single nucleus (or cell) comprising the adapter-flanked template nucleic

acid fragments; and (2) a plurality of single-stranded barcode oligonucleotide molecules (e.g., nucleic acid barcode molecules) comprising a transposon end sequence (“ME”), a Read1 sequence (“R1”), or a portion thereof, a barcode sequence (“BC”), and a P5 adapter sequence (“P5”). See **FIG. 30A**. In some embodiments, the single-stranded barcode oligonucleotide molecules are attached to a solid or semi-solid particle (e.g., a bead, such as a gel bead) and partitioned such that at least some partitions (e.g., droplets or wells) comprise (1) a single nucleus (or cell) comprising the adapter-flanked template nucleic acid fragments and (2) a single solid or semi-solid particle (e.g., gel bead). In addition to the aforementioned components, in some embodiments, the plurality of partitions (e.g., droplets or wells) further comprises reagents (e.g., enzymes and buffers) that facilitate the reactions described below.

[00332] Single cell- or nucleus-containing partitions (e.g., droplets or wells) are then subjected to conditions to release the adapter-flanked template nucleic acid fragments from the nuclei. After the adapter-flanked template nucleic acid fragments are released, gaps from the transposition reaction are filled with a suitable enzyme. See **FIG. 30B**. In certain embodiments, where barcode oligonucleotides (e.g., nucleic acid barcode molecules) are attached to a solid or semi-solid particle (e.g., bead, such as a gel bead), partitions (e.g., droplets or wells) are subjected to conditions to cause release of the barcode oligonucleotide molecules from the solid or semi-solid particle (e.g., bead, such as a gel bead) (e.g., depolymerization or degradation of beads, for example, using a reducing agent such as DTT). Gap-filled adapter-flanked template nucleic acid fragments are then subjected to a linear amplification reaction using the single-stranded barcode oligonucleotide molecules as primers to produce barcoded, adapter-flanked template nucleic acid fragments. See **FIG. 30B**.

[00333] The barcoded, adapter-flanked template nucleic acid fragments are then released from the partitions (e.g., droplets or wells) and processed in bulk to complete library preparation for next generation high throughput sequencing (e.g., to add sample index (SI) sequences (e.g., “i7”) and/or further adapter sequences (e.g., “P7”). The fully constructed library is then sequenced according to a suitable next-generation sequencing protocol (e.g., Illumina sequencing).

Example 12. Generation of Barcoded Nucleic Acid Fragments using Tagmentation and Barcoding by Linear Amplification in Partitions

[00334] Cells from a cell population of interest (or intact nuclei from cells in a cell population of interest) are partitioned into a plurality of partitions (e.g., droplets or wells) such that at least some partitions comprise (1) a single cell (or a single nucleus) comprising a template nucleic acid; and (2) a plurality of single-stranded barcode oligonucleotide molecules (e.g., nucleic acid

barcode molecules) comprising a transposon end sequence (“ME”), a Read1 sequence (“R1”), a barcode sequence (“BC”), and a P5 adapter sequence (“P5”). See, e.g., **FIG. 30A**. In some embodiments, the single-stranded barcode oligonucleotide molecules are attached to a solid or semi-solid particle (e.g., bead, such as a gel bead) and partitioned such that at least some partitions (e.g., droplets or wells) comprise (1) a single cell (or a single nucleus) and (2) a single solid or semi-solid particle (e.g., bead, such as a gel bead). In addition to the aforementioned components, in some embodiments, the plurality of partitions (e.g., droplets or wells) further comprises reagents (e.g., enzymes and buffers) that facilitate the reactions described below.

[00335] After partitioning into partitions (e.g., droplets or wells), the single cells (or nuclei) are lysed to release the template genomic DNA in a manner that substantially maintains native chromatin organization. In certain embodiments, where barcode oligonucleotides (e.g., nucleic acid barcode molecules) are attached to a solid or semi-solid particle (e.g., bead, such as a gel bead), partitions (e.g., droplets or wells) are subjected to conditions to cause release of the barcode oligonucleotide molecules from the solid or semi-solid particle (e.g., bead, such as a gel bead) (e.g., depolymerization or degradation of beads, for example, using a reducing agent such as DTT). Partitions (e.g., droplets or wells) are then subjected to conditions to generate a transposase-nucleic acid complex as described in Example 9 and shown in **FIG. 29A**.

Alternatively, in some embodiments, a plurality of pre-formed transposase-nucleic acid complexes as shown in **FIG. 29A** are partitioned into the plurality of partitions (e.g., droplets or wells). Partitions (e.g., droplets or wells) are then subjected to conditions such that the transposase-nucleic acid complexes integrate the first and second adapter sequences into the template nucleic acid to generate double-stranded adapter-flanked template nucleic acid fragments. Because the transposase-nucleic acid complex can only act on nucleosome-free DNA, the adapter-flanked template nucleic acid fragments are representative of genome-wide areas of accessible chromatin in a single cell or nucleus. Alternatively, in some embodiments, the tagmentation reaction is performed in intact nuclei, and the nuclei are lysed to release the double-stranded adapter-flanked template nucleic acid fragments.

[00336] Samples are then processed generally as described in Example 11. After tagmentation, gaps from the transposition reaction are filled with a suitable gap-filling enzyme. See **FIG. 31**. Gap-filled adapter-flanked template nucleic acid fragments are then subjected to a linear amplification reaction using the single-stranded barcode oligonucleotide molecules as primers to produce barcoded, adapter-flanked template nucleic acid fragments. See **FIG. 31**.

[00337] The barcoded, adapter-flanked template nucleic acid fragments are then released from the partitions (e.g., droplets or wells) and processed in bulk to complete library preparation for next generation high throughput sequencing (e.g., to add sample index (SI) sequences (e.g.,

“i7”) and/or further adapter sequences (*e.g.*, “P7”). The fully constructed library is then sequenced according to a suitable next-generation sequencing protocol (*e.g.*, Illumina sequencing).

Example 13. Generation of Barcoded Nucleic Acid Fragments using Bulk Tagmentation and CRISPR/Cas9 Cleavage in Partitions

[00338] Nuclei are harvested in bulk from cells in a cell population of interest in a manner that substantially maintains native chromatin organization. Alternatively, cells are permeabilized/permeable, allowing the transposase-nucleic acid complex to gain access to the nucleus. Nuclei are then incubated in the presence of a transposase-nucleic acid complex as described in Example 9. See **FIG. 29A**. In some embodiments, after transposition, the transposase is inactivated or dissociated from the adapter-flanked template nucleic acid fragments.

[00339] Nuclei (or cells) comprising the adapter-flanked template nucleic acid fragments are then partitioned into a plurality of partitions (*e.g.*, droplets or wells) such that at least some partitions comprise (1) a single nucleus comprising the adapter-flanked template nucleic acid fragments; (2) a plurality of double-stranded barcode oligonucleotide molecules (*e.g.*, nucleic acid barcode molecules) comprising a barcode sequence (BC) and a TruSeqR1 sequencing primer sequence (*e.g.*, **FIG. 32A**); and (3) a plurality of CRISPR/Cas9 complexes comprising a Cas9 nuclease and a synthetic guide RNA (gRNA) that targets the Read1/ME adapter sequence in the adapter-flanked template nucleic acid fragments. See **FIG. 32B**. In some embodiments, the double-stranded barcode oligonucleotide molecules (*e.g.*, nucleic acid barcode molecules) are attached to a solid or semi-solid particle (*e.g.*, bead, such as a gel bead) and partitioned such that at least some droplets comprise (1) a single nucleus; (2) a single solid or semi-solid particle (*e.g.*, gel bead); and (3) a plurality of CRISPR/Cas9 complexes. In addition to the aforementioned components, in some embodiments, the plurality of partitions (*e.g.*, droplets or wells) further comprises reagents (*e.g.*, enzymes and buffers) that facilitate the reactions described below.

[00340] Single nucleus-containing partitions (*e.g.*, droplets or wells) are then subjected to conditions to release the adapter-flanked template nucleic acid fragments from the nuclei. After the adapter-flanked template nucleic acid fragments are released, gaps from the transposition reaction are filled with a suitable gap-filling enzyme. Gap-filled adapter-flanked template nucleic acid fragments are then subjected to Cas9-mediated cleavage of the R1/ME adaptor, or some portion thereof. See **FIG. 32B**. In certain embodiments, where barcode oligonucleotides (*e.g.*, nucleic acid barcode molecules) are attached to a solid or semi-solid particle (*e.g.*, a bead, such as a gel bead), partitions (*e.g.*, droplets or wells) are subjected to conditions to cause release

of the barcode oligonucleotide molecules from the solid or semi-solid particle (e.g., gel bead) (e.g., depolymerization or degradation of beads, for example, using a reducing agent such as DTT). The barcode oligonucleotides are then ligated onto the R1 adapter-cleaved ends of the template nucleic acid fragments to produce barcoded, adapter-flanked template nucleic acid fragments. See **FIG. 32B**.

[00341] The barcoded, adapter-flanked template nucleic acid fragments are then released from the partitions (e.g., droplets or wells) and processed in bulk to complete library preparation for next generation high throughput sequencing (e.g., subjecting the fragments, or derivatives thereof, to one or more reactions (e.g., nucleic acid amplification) to add functional sequences to facilitate Illumina sequencing). In some embodiments, a second CRISPR/Cas9 mediated cleavage event using a synthetic guide RNA (gRNA) that targets the Read2/ME adapter sequence is performed either in the partitions or in bulk after release from the partitions. The fully constructed library is then sequenced according to any suitable sequencing protocol.

Example 14. Generation of Barcoded Nucleic Acid Fragments using Tagmentation and CRISPR/Cas9 Cleavage in Partitions

[00342] Cells from a cell population of interest (or intact nuclei from cells in a cell population of interest) are partitioned into a plurality of partitions (e.g., droplets or wells) such that at least some partitions comprise (1) a single cell (or a single nucleus) comprising a template nucleic acid; (2) a plurality of double-stranded barcode oligonucleotide molecules (e.g., nucleic acid barcode molecules) comprising a barcode sequence ("BC") and a TruSeqR1 sequencing primer sequence (e.g., **FIG. 32A**); and (3) a plurality of CRISPR/Cas9 complexes comprising a Cas9 nuclease and a synthetic guide RNA (gRNA) that targets the Read1/ME adapter sequence in the adapter-flanked template nucleic acid fragments. See, e.g., **FIG. 32C**. In some embodiments, the double-stranded barcode oligonucleotide molecules (e.g., nucleic acid barcode molecules) are attached to a solid or semi-solid particle (e.g., bead, such as a gel bead) and partitioned such that at least some partitions (e.g., droplets or wells) comprise (1) a single cell (or single nucleus); (2) a single solid or semi-solid particle (e.g., gel bead); and (3) a plurality of CRISPR/Cas9 complexes. In addition to the aforementioned components, in some embodiments, the plurality of partitions (e.g., droplets or wells) further comprises reagents (e.g., enzymes and buffers) that facilitate the reactions described below.

[00343] After partitioning into partitions (e.g., droplets or wells), the single cells (or nuclei) are lysed to release the template genomic DNA in a manner that substantially maintains native chromatin organization. Partitions (e.g., droplets or wells) are then subjected to conditions to generate a transposase-nucleic acid complex as described in Example 9 and shown in **FIG. 29A**.

Alternatively, in some embodiments, a plurality of pre-formed transposase-nucleic acid complexes as shown in **FIG. 29A** are partitioned into the plurality of partitions (e.g., droplets or wells). Partitions (E.g., droplets or wells) are then subjected to conditions such that the transposase-nucleic acid complexes integrate the first and second adapter sequences into the template nucleic acid to generate double-stranded adapter-flanked template nucleic acid fragments. Because the transposase-nucleic acid complex can only act on nucleosome-free DNA, the adapter-flanked template nucleic acid fragments are representative of genome-wide areas of accessible chromatin in a single cell or nucleus. Alternatively, in some embodiments, the tagmentation reaction is performed in intact nuclei, and the nuclei are lysed to release the double-stranded adapter-flanked template nucleic acid fragments.

[00344] Samples are then processed generally as described in Example 13. After tagmentation, gaps from the transposition reaction are filled with a suitable gap-filling enzyme. See **FIG. 32C**. Gap-filled adapter-flanked template nucleic acid fragments are then subjected to Cas9-mediated cleavage of the R1 adaptor. See **FIG. 32C**. In certain embodiments, where barcode oligonucleotides (e.g., nucleic acid barcode molecules) are attached to a solid or semi-solid particle (e.g., bead, such as a gel bead), partitions (e.g., droplets or wells) are subjected to conditions to cause release of the barcode oligonucleotide molecules from the solid or semi-solid particle (e.g., gel bead) (e.g., depolymerization or degradation of beads, for example, using a reducing agent such as DTT). The barcode oligonucleotides are then ligated onto the R1 adapter-cleaved ends of the template nucleic acid fragments to produce barcoded, adapter-flanked template nucleic acid fragments. See **FIG. 32C**.

[00345] The barcoded, adapter-flanked template nucleic acid fragments are then released from the partitions (e.g., droplets or wells) and processed in bulk to complete library preparation for next generation high throughput sequencing (e.g., subjecting the fragments, or derivatives thereof, to one or more reactions (e.g., nucleic acid amplification) to add functional sequences to facilitate Illumina sequencing). In some embodiments, a second CRISPR/Cas9 mediated cleavage event using a synthetic guide RNA (gRNA) that targets the Read2/ME adapter sequence is performed either in the partitions or in bulk after release from the partitions. The fully constructed library is then sequenced according to any suitable sequencing protocol.

Example 15. Generation of Barcoded Nucleic Acid Fragments using Bulk Tagmentation and CRISPR/Cas9 Cleavage in Partitions using Y-Adapters

[00346] Nuclei are harvested in bulk from cells in a cell population of interest in a manner that substantially maintains native chromatin organization. Alternatively, cells are permeabilized/permeable, allowing the transposase-nucleic acid complex to gain access to the

nucleus. Nuclei are then incubated in the presence of a transposase-nucleic acid complex as described in Example 9. See **FIG. 29A**. In some embodiments, after transposition, the transposase is inactivated or dissociated from the adapter-flanked template nucleic acid fragments.

[00347] Nuclei (or cell) comprising the adapter-flanked template nucleic acid fragments are then partitioned into a plurality of partitions (e.g., droplets or wells) such that at least some partitions comprise (1) a single nucleus comprising the adapter-flanked template nucleic acid fragments; (2) a plurality of Y-adaptor barcode oligonucleotide molecules (e.g., nucleic acid barcode molecules) comprising a barcode sequence (“BC”), a Read1 sequencing primer sequence (“R1”), and a Read2 sequencing primer sequence (“R2”), *e.g.*, **FIG. 33A**; (3) a first plurality of CRISPR/Cas9 complexes comprising a Cas9 nuclease and a synthetic guide RNA (gRNA) that targets the Read1/ME adapter sequence in the adapter-flanked template nucleic acid fragments; and (4) a second plurality of CRISPR/Cas9 complexes comprising a Cas9 nuclease and a synthetic guide RNA (gRNA) that targets the Read2/ME adapter sequence in the adapter-flanked template nucleic acid fragments. See **FIG. 33C**. In some embodiments, the Y-adaptor barcode oligonucleotide molecules are attached to a solid or semi-solid particle (e.g., a bead, such as a gel bead) (*e.g.*, **FIG. 15A**) and partitioned such that at least some partitions (e.g., droplets or wells) comprise (1) a single nucleus; (2) a single solid or semi-solid particle (e.g., bead, such as a gel bead); (3) the first plurality of CRISPR/Cas9 complexes; and (4) the second plurality of CRISPR/Cas9 complexes. In addition to the aforementioned components, in some embodiments, the plurality of partitions (e.g., droplets or wells) further comprises reagents (*e.g.*, enzymes and buffers) that facilitate the reactions described below.

[00348] Single nucleus containing partitions (e.g., droplets or wells) are then subjected to conditions to release the adapter-flanked template nucleic acid fragments from the nuclei. After the adapter-flanked template nucleic acid fragments are released, gaps from the transposition reaction are filled with a suitable gap-filling enzyme. Gap-filled adapter-flanked template nucleic acid fragments are then subjected to Cas9-mediated cleavage of the R1 and R2 adaptors, or a portion thereof. See **FIG. 33B**. In certain embodiments, where barcode oligonucleotides (e.g., nucleic acid barcode molecules) are attached to a solid or semi-solid particle (e.g., a bead, such as a gel bead), partitions (e.g., droplets or wells) are subjected to conditions to cause release of the barcode oligonucleotide molecules from the solid or semi-solid particle (e.g., gel bead) (*e.g.*, depolymerization or degradation of beads, for example, using a reducing agent such as DTT). The Y-adaptor barcode oligonucleotides are then ligated onto the R1/R2 adapter-cleaved ends of the template nucleic acid fragments to produce barcoded, adapter-flanked template nucleic acid fragments. See **FIG. 33B**.

[00349] The barcoded, adapter-flanked template nucleic acid fragments are then released from the partitions (e.g., droplets or wells) and processed in bulk to complete library preparation for next generation high throughput sequencing (e.g., subjecting the fragments, or derivatives thereof, to one or more reactions (e.g., nucleic acid amplification) to add functional sequences to facilitate Illumina sequencing). The fully constructed library is then sequenced according to any suitable sequencing protocol.

Example 16. Generation of Barcoded Nucleic Acid Fragments using Tagmentation and CRISPR/Cas9 Cleavage in Partitions using Y-Adapters

[00350] Cells from a cell population of interest (or intact nuclei from cells in a cell population of interest) are partitioned into a plurality of partitions (e.g., droplets or wells) such that at least some partitions comprise (1) a single cell (or a single nucleus) comprising a template nucleic acid; (2) a plurality of Y-adaptor barcode oligonucleotide molecules (e.g., nucleic acid barcode molecules) comprising a barcode sequence (“BC”), a Read1 sequencing primer sequence (“R1”), and a Read2 sequencing primer sequence (“R2”) (e.g., **FIG. 33A**); (3) a first plurality of CRISPR/Cas9 complexes comprising a Cas9 nuclease and a synthetic guide RNA (gRNA) that targets the Read1/ME adapter sequence in the adapter-flanked template nucleic acid fragments; and (4) a second plurality of CRISPR/Cas9 complexes comprising a Cas9 nuclease and a synthetic guide RNA (gRNA) that targets the Read2/ME adapter sequence in the adapter-flanked template nucleic acid fragments. See **FIG. 33C**. In some embodiments, the Y-adaptor barcode oligonucleotide molecules are attached to a solid or semi-solid particle (e.g., a bead, such as a gel bead) and partitioned such that at least some partitions (e.g., droplets or wells) comprise (1) a single cell (or single nucleus); (2) a single solid or semi-solid particle (e.g., gel bead); (3) the first plurality of CRISPR/Cas9 complexes; and (4) the second plurality of CRISPR/Cas9 complexes. In addition to the aforementioned components, in some embodiments, the plurality of partitions (e.g., droplets or wells) further comprises reagents (e.g., enzymes and buffers) that facilitate the reactions described below.

[00351] After partitioning into partitions (e.g., droplets or wells), the single cells (or nuclei) are lysed to release the template genomic DNA in a manner that substantially maintains native chromatin organization. Partitions (e.g., droplets or wells) are then subjected to conditions to generate a transposase-nucleic acid complex as described in Example 9 and shown in **FIG. 29A**. Alternatively, in some embodiments, a plurality of pre-formed transposase-nucleic acid complexes as shown in **FIG. 29A** are partitioned into the plurality of partitions (e.g., droplets or wells). Partitions (e.g., droplets or wells) are then subjected to conditions such that the transposase-nucleic acid complexes integrate the first and second adapter sequences into the

template nucleic acid to generate double-stranded adapter-flanked template nucleic acid fragments. Because the transposase-nucleic acid complex can only act on nucleosome-free DNA, the adapter-flanked template nucleic acid fragments are representative of genome-wide areas of accessible chromatin in a single cell or nucleus. Alternatively, in some embodiments, the tagmentation reaction is performed in intact nuclei, and the nuclei are lysed to release the double-stranded adapter-flanked template nucleic acid fragments.

[00352] Samples are then processed generally as described in Example 15. After tagmentation, gaps from the transposition reaction are filled with a suitable gap-filling enzyme. See **FIG. 33C**. Gap-filled adapter-flanked template nucleic acid fragments are then subjected to Cas9-mediated cleavage of the R1 and R2 adaptors, or a portion thereof. See **FIG. 33C**. In certain embodiments, where barcode oligonucleotides (e.g., nucleic acid barcode molecules) are attached to a solid or semi-solid particle (e.g., a bead, such as a gel bead), partitions (e.g., droplets or wells) are subjected to conditions to cause release of the barcode oligonucleotide molecules from the solid or semi-solid particle (e.g., gel bead) (e.g., depolymerization or degradation of beads, for example, using a reducing agent such as DTT). The Y-adaptor barcode oligonucleotides are then ligated onto the R1/R2 adapter-cleaved ends of the template nucleic acid fragments to produce barcoded, adapter-flanked template nucleic acid fragments. See **FIG. 33C**.

[00353] The barcoded, adapter-flanked template nucleic acid fragments are then released from the partitions (e.g., droplets or wells) and processed in bulk to complete library preparation for next generation high throughput sequencing (e.g., subjecting the fragments, or derivatives thereof, to one or more reactions (e.g., nucleic acid amplification) to add functional sequences to facilitate Illumina sequencing). The fully constructed library is then sequenced according to any suitable sequencing protocol.

Example 17. Generation of Barcoded Nucleic Acid Molecules for ATAC-seq and Barcoded cDNA from the Same Single Cell

[00354] Cells from a cell population of interest (or intact nuclei from cells in a cell population of interest) are partitioned into a plurality of partitions (e.g., droplets or wells) such that at least some partitions comprise (1) a single cell (or a single nucleus) comprising template genomic DNA molecules and template RNA molecules; (2) a plurality of first barcoded oligonucleotide molecules (e.g., first nucleic acid barcode molecules) comprising a barcode sequence; (3) a plurality of transposase molecules, (4) a plurality of second barcode oligonucleotide molecules (e.g., second nucleic acid barcode molecules) comprising a barcode sequence and a capture sequence; and (5) a plurality of reverse transcriptase molecules. In some embodiments, the

barcode sequence from the first barcoded oligonucleotide and the barcode sequence from the second barcoded oligonucleotide is the same. In some embodiments, the barcode sequence from the first barcoded oligonucleotide and the barcode sequence from the second barcoded oligonucleotide are different.

[00355] In some embodiments, the plurality of first barcoded oligonucleotides (e.g., first nucleic acid barcode molecules) and the plurality of second barcoded oligonucleotides (e.g., second nucleic acid barcode molecules) are attached to a solid or semi-solid particle (e.g., a bead, such as a gel bead) and partitioned such that at least some partitions (e.g., droplets or wells) comprise (1) a single cell (or single nucleus); (2) a single solid or semi-solid particle (e.g., gel bead) comprising the first and second plurality of barcoded oligonucleotides; (3) a plurality of transposase molecules; and (4) a plurality of reverse transcriptase molecules. In other embodiments, the plurality of first barcoded oligonucleotides (e.g., first nucleic acid barcode molecules) are attached to a first solid or semi-solid particle (e.g., bead, such as a gel bead) while the plurality of second barcoded oligonucleotides (e.g., second nucleic acid barcode molecules) are attached to a second solid or semi-solid particle (e.g., bead, such as a gel bead) and partitioned such that at least some partitions (e.g., droplets or wells) comprise (1) a single cell (or single nucleus); (2) a single first solid or semi-solid particle (e.g., gel bead); (2) a single second solid or semi-solid particle (e.g., gel bead); (3) a plurality of transposase molecules; and (4) a plurality of reverse transcriptase molecules.

[00356] In certain embodiments, the plurality of first barcoded oligonucleotides (e.g., first nucleic acid barcode molecules) are attached to a plurality of solid or semi-solid particles (e.g., beads, such as gel beads) while the plurality of second barcoded oligonucleotides (e.g., second nucleic acid barcode molecules) are attached to a plurality of magnetic particles (e.g., beads), wherein the plurality of magnetic particles are embedded within the solid or semi-solid particles (e.g., gel beads). Continuing these embodiments, the plurality of solid or semi-solid particles (e.g., gel beads) is partitioned such that at least some partitions (e.g., droplets or wells) comprise: (1) a single cell (or single nucleus); (2) a single solid or semi-solid particle (e.g., gel bead) comprising (i) a plurality of first barcoded oligonucleotides attached to the single solid or semi-solid particle (e.g., gel bead); and (ii) a plurality of magnetic particles embedded within the single solid or semi-solid particle (e.g., gel bead), wherein the magnetic particles comprise the second barcode oligonucleotide attached thereto; (3) a plurality of transposase molecules; and (4) a plurality of reverse transcriptase molecules. See, e.g., **FIG. 34**. Similarly, in other embodiments, the second barcode oligonucleotides (e.g., second nucleic acid barcode molecules) are attached to the solid or semi-solid particle (e.g., gel bead) while the first oligonucleotides (e.g., first nucleic acid barcode molecules) are attached to a plurality of magnetic particles

embedded within the solid or semi-solid particle (e.g., gel bead). See, e.g., **FIG. 34**. In addition to the aforementioned components, in some embodiments, the plurality of partitions (e.g., droplets or wells) further comprise reagents (e.g., enzymes and buffers) that facilitate the reactions described herein.

[00357] The first barcoded oligonucleotide and related nucleic acid processing steps can take on the structure of any of the aforementioned Examples and may include additional components as described therein. For instance, in some embodiments, the first barcoded oligonucleotide comprises a barcode sequence and a transposon end sequence (e.g., a ME sequence) and is, for example, (1) a forked adapter such as those described in Example 1, **FIGs. 12A-12B**; (2) a T7-containing oligonucleotide such as those described in Example 3, **FIG. 18**; or (3) a barcoded oligonucleotide such as those described in (i) Example 7, **FIG. 28B**; (ii) Example 11, **FIGs. 30A-30B**; and (iii) Example 12, **FIG. 31**. In other embodiments, the first barcoded oligonucleotide comprises a barcode sequence and is, for example, (1) a forked adapter such as those described in Example 2, **FIGs. 15A-15B** or Examples 15-16, **FIGs. 33A-33C**; or (2) a barcoded oligonucleotide such as those described in Examples 9 and 10, **FIGs. 29A-29C** or Examples 13-14, **FIGs. 32A-32C**. In some cases, the second barcoded oligonucleotide comprises a second barcode sequence and a capture sequence, where the capture sequence can be, for example, a poly-T sequence, a random primer sequence (e.g., a random hexamer), or a gene-specific sequence.

[00358] After partitioning into partitions (e.g., droplets or wells), the single cells (or nuclei) are lysed to release template genomic DNA and template RNA (e.g., cytoplasmic mRNA or nuclear mRNA) in a manner that substantially maintains native chromatin organization of the genomic DNA. In certain embodiments, where barcode oligonucleotides are attached to a solid or semi-solid particle (e.g., a bead, such as a gel bead), partitions (e.g., droplets or wells) are subjected to conditions to cause release of barcode oligonucleotide molecules from the solid or semi-solid particle (e.g., gel bead) (e.g., depolymerization or degradation of beads, for example, using a reducing agent such as DTT). Partitions (e.g., droplets or wells) are then subjected to conditions to generate a transposase-nucleic acid complex as described in the aforementioned examples. Alternatively, in some embodiments, a plurality of pre-formed transposase-nucleic acid complexes are partitioned into the plurality of partitions (e.g., droplets or wells). Partitions (e.g., droplets or wells) are then subjected to conditions such that the transposase-nucleic acid complexes generate double-stranded template genomic DNA fragments. As described above, the transposition reaction can take on the structure of any of the aforementioned Examples to generate double-stranded template genomic DNA fragments flanked by a wide variety of functional sequences and suitable for a number of downstream processing steps. For example, as

described herein, in some embodiments, the transposition reaction directly integrates the barcode sequence into the template genomic DNA fragments (*e.g.*, Example 1) while, in other embodiments, the barcode sequence is added to template genomic DNA fragments subsequent to the transposition reaction (such as by ligation, *e.g.*, Example 2). Because the transposase-nucleic acid complex can only act on nucleosome-free DNA, the template genomic DNA fragments are representative of genome-wide areas of accessible chromatin in a single cell or nucleus.

Alternatively, in some embodiments, the transposition reaction is performed in intact nuclei, and the nuclei are lysed to release the adapter-flanked template genomic DNA fragments.

Alternatively, in some embodiments, the transposition reaction is performed in bulk in intact nuclei and a single nucleus comprising template genomic DNA fragments is partitioned and processed as described above. In some embodiments, gaps from the transposition reaction are filled in-droplet with a suitable gap-filling enzyme. In other embodiments, a gap-filling reaction is performed in bulk after the double-stranded, barcoded adapter-flanked DNA fragments have been released from the partitions.

[00359] In some cases, partitions (*e.g.*, droplets or wells) may be subjected to conditions to capture template RNA (*e.g.*, messenger RNA, mRNA) on second solid or semi-solid particles using a capture sequence of second barcoded oligonucleotides of the plurality of second barcoded oligonucleotides. Alternatively, second barcoded oligonucleotides of the plurality of second barcoded oligonucleotides may be released from the second solid or semi-solid particles (*e.g.*, as described herein) and used to capture template RNA. Captured template RNA may be subsequently processed in bulk after removal from the partitions. Alternatively, in some cases, partitions (*e.g.*, droplets or wells) may be subjected to conditions to generate single-stranded, barcoded cDNA molecules from the template RNA using the capture sequence from the second barcode oligonucleotide to prime the reverse transcription reaction (*e.g.*, an oligo (dT) sequence). In some embodiments, second strand cDNA is produced (*e.g.*, through a template switching oligonucleotide or through random priming) to generate double-stranded, barcoded cDNA molecules. In some embodiments, the template switching oligonucleotide also comprises a barcode sequence such that both the 5' and 3' end of the cDNA comprise a barcode sequence. The barcode sequence on the 5' and 3' end can be the same barcode sequence or the 5' end can have a different barcode sequence than the 3' end. In other embodiments, the plurality of second barcode oligonucleotide molecules are omitted and replaced with plurality of second oligonucleotide molecules comprising a capture sequence and no barcode sequence. Continuing with these embodiments, first strand cDNA molecules are generated using the capture sequence while second strand cDNA is generated through use of a barcoded template switching oligonucleotide to barcode the 5' end of the template RNA. In some embodiments, an in-

partition (e.g., in-droplet or in-well) amplification reaction, such as linear amplification, is performed on the adapter-flanked DNA fragments, the barcoded cDNA molecules, or both the adapter-flanked DNA fragments and the barcoded cDNA molecules. In some embodiments, a barcode oligonucleotide is directly ligated onto the template RNA.

Exemplary Scheme 1 – scATAC-seq + 3' scRNA-seq using ligation

[00360] In an exemplary barcoding scheme, cells from a cell population of interest (or intact nuclei from cells in a cell population of interest) are partitioned into a plurality of partitions (e.g., droplets or wells) such that at least some partitions comprise: (1) a single cell (or a single nucleus) comprising template genomic DNA molecules and template RNA molecules (e.g., mRNA or nuclear pre-mRNA); (2) a solid or semi-solid particle (e.g., bead, such as a gel bead) comprising (i) a plurality of partially double-stranded nucleic acid barcode molecules as shown in **FIG. 29B** (see also **FIG. 35A**), and (ii) a plurality of nucleic acid barcode molecules as shown in **FIG. 35B**; (3) a plurality of transposase molecules; (4) a plurality of nucleic acid molecules comprising a transposase end sequence (e.g., **FIG. 29A**); and (5) a plurality of reverse transcriptase molecules. In some embodiments, the solid or semi-solid particle is a bead. In some embodiments, the bead is a gel bead. In some embodiments, the solid or semi-solid particle (e.g., gel bead) comprises the plurality of partially double-stranded nucleic acid barcode molecules of **FIG. 35A** attached thereto and a plurality of magnetic particles embedded within the solid or semi-solid particle (e.g., gel bead) (see, e.g., **FIG. 34**), wherein the plurality of magnetic particles comprises, attached thereto, the plurality of nucleic acid barcode molecules of **FIG. 35B**. In other embodiments, the solid or semi-solid particle (e.g., gel bead) comprises the plurality of nucleic acid barcode molecules of **FIG. 35B** attached thereto and a plurality of magnetic particles embedded within the solid or semi-solid particle (e.g., gel bead), wherein the plurality of magnetic particles comprises, attached thereto, the plurality of nucleic acid barcode molecules of **FIG. 35A**. In some embodiments, the barcode sequence in **FIG. 35A** and the barcode sequence **FIG. 35B** is the same. In some embodiments, the barcode sequence in **FIG. 35A** and the barcode sequence **FIG. 35B** comprise one or more barcode segments that are identical. In some embodiments, the barcode sequence in **FIG. 35A** and the barcode sequence **FIG. 35B** are different.

[00361] Continuing these embodiments, after partitioning, the single cells (or nuclei) are lysed to release template genomic DNA and template RNA (e.g., cytoplasmic mRNA or nuclear mRNA) in a manner that substantially maintains native chromatin organization of the genomic DNA. In certain embodiments, the plurality of nucleic acid barcode molecules of **FIG. 35A** and **FIG. 35B** are releasably attached to the solid or semi-solid particle (e.g., bead) and partitions (e.g., droplets or wells) are subjected to conditions to cause release of barcode oligonucleotide

molecules from the solid or semi-solid particle (e.g., bead) (e.g., depolymerization or degradation of a bead, for example, using a reducing agent such as DTT). Partitions (e.g., droplets or wells) are then subjected to conditions to generate a transposase-nucleic acid complex as described in the aforementioned examples. Alternatively, in some embodiments, a plurality of pre-formed transposase-nucleic acid complexes (e.g., **FIG. 29A**) are partitioned into the plurality of partitions (e.g., droplets or wells). Partitions are then subjected to conditions such that the transposase-nucleic acid complexes generate double-stranded template genomic DNA fragments. Alternatively, in some embodiments, the transposition reaction is performed in bulk in intact nuclei which are partitioned such that at least some partitions (e.g., droplets or wells) comprise (1) a single cell (or a single nucleus) comprising template genomic DNA fragments and template RNA molecules; (2) a solid or semi-solid particle (e.g., a bead, such as a gel bead) comprising (i) a plurality of partially double-stranded nucleic acid barcode molecules as shown in **FIG. 29B** (see also **FIG. 35A**), and (ii) a plurality of nucleic acid barcode molecules as shown in **FIG. 35B**; and (3) a plurality of reverse transcriptase molecules. In some embodiments, the nucleic acid barcode molecules of **FIG. 35A** or **FIG. 35B** are attached to a magnetic particle embedded within a solid or semi-solid particle (e.g., a bead, such as a gel bead), as previously described. After partitioning, the single nuclei are lysed to release template genomic DNA fragments and template RNA (e.g., cytoplasmic mRNA or nuclear mRNA) molecules. Continuing this embodiment, after fragmentation, template genomic DNA fragments are processed generally as outlined in **FIG. 29D** (or **FIG. 29C** for bulk tagmentation) by ligating a partially double-stranded nucleic acid barcode molecule as shown in **FIG. 29B** (see also **FIG. 35A**) to the template nucleic acid fragments (see, e.g., Examples 9 and 10). Using the reverse transcriptase, template RNA molecules comprising a poly-A tail (e.g., mRNA molecules) are processed with the plurality of nucleic acid barcode molecules of **FIG. 35B** to generate barcoded cDNA molecules as generally described elsewhere herein.

Exemplary Scheme 2 – scATAC-seq + 5' scRNA-seq using ligation

[00362] In another barcoding scheme, cells from a cell population of interest (or intact nuclei from cells in a cell population of interest) are partitioned into a plurality of partitions (e.g., droplets or wells) such that at least some partitions comprise: (1) a single cell (or a single nucleus) comprising template genomic DNA molecules and template RNA molecules (e.g., mRNA or nuclear pre-mRNA); (2) a solid or semi-solid particle (e.g., bead, such as a gel bead) comprising (i) a plurality of partially double-stranded nucleic acid barcode molecules as shown in **FIG. 29B** (see also **FIG. 35A**), and (ii) a plurality of nucleic acid barcode molecules as shown in **FIG. 35D**; (3) a plurality of transposase molecules; (4) a plurality of nucleic acid molecules comprising a transposase end sequence (e.g., **FIG. 29A**); (5) a plurality of nucleic acid

molecules comprising a poly-T sequence; and (6) a plurality of reverse transcriptase molecules. In some embodiments, the solid or semi-solid particle is a bead. In some embodiments, the bead is a gel bead. In some embodiments, the solid or semi-solid particle (e.g., gel bead) comprises the plurality of partially double-stranded nucleic acid barcode molecules of **FIG. 35A** attached thereto and a plurality of magnetic particles embedded within the solid or semi-solid particle (e.g., gel bead) (see, *e.g.*, **FIG. 34**), wherein the plurality of magnetic particles comprises, attached thereto, the plurality of nucleic acid barcode molecules of **FIG. 35D**. In other embodiments, the solid or semi-solid particle (e.g., gel bead) comprises the plurality of nucleic acid barcode molecules of **FIG. 35D** attached thereto and a plurality of magnetic particles embedded within the solid or semi-solid particle (e.g., gel bead), wherein the plurality of magnetic particles comprises, attached thereto, the plurality of nucleic acid barcode molecules of **FIG. 35A**. In some embodiments, the barcode sequence in **FIG. 35A** and the barcode sequence **FIG. 35D** is the same. In some embodiments, the barcode sequence in **FIG. 35A** and the barcode sequence **FIG. 35D** comprise one or more barcode segments that are identical. In some embodiments, the barcode sequence in **FIG. 35A** and the barcode sequence **FIG. 35D** are different.

[00363] Continuing these embodiments, after partitioning, the single cells (or nuclei) are lysed to release template genomic DNA and template RNA (*e.g.*, cytoplasmic mRNA or nuclear mRNA) in a manner that substantially maintains native chromatin organization of the genomic DNA. In certain embodiments, the plurality of nucleic acid barcode molecules of **FIG. 35A** and **FIG. 35D** are releasably attached to the solid or semi-solid particle (e.g., bead, such as a gel bead) and partitions (e.g., droplets or wells) are subjected to conditions to cause release of barcode oligonucleotide molecules from the bead (*e.g.*, depolymerization or degradation of a bead, for example, using a reducing agent such as DTT). Partitions are then subjected to conditions to generate a transposase-nucleic acid complex as described in the aforementioned examples. Alternatively, in some embodiments, a plurality of pre-formed transposase-nucleic acid complexes (*e.g.*, **FIG. 29A**) are partitioned into the plurality of partitions (e.g., droplets or wells). Partitions are then subjected to conditions such that the transposase-nucleic acid complexes generate double-stranded template genomic DNA fragments. Alternatively, in some embodiments, the transposition reaction is performed in bulk in intact nuclei which are partitioned such that at least some partitions comprise (1) a single cell (or a single nucleus) comprising template genomic DNA fragments and template RNA molecules; (2) a solid or semi-solid particle (e.g., a bead, such as a gel bead) comprising (i) a plurality of partially double-stranded nucleic acid barcode molecules as shown in **FIG. 29B** (see also **FIG. 35A**), and (ii) a plurality of nucleic acid barcode molecules as shown in **FIG. 35D**; (3) a plurality of nucleic acid

molecules comprising a poly-T sequence; and (4) a plurality of reverse transcriptase molecules. In some embodiments, the nucleic acid barcode molecules of **FIG. 35A** or **FIG. 35D** are attached to a magnetic particle embedded within a solid or semi-solid particle (e.g., gel bead), as previously described. After partitioning, the single nuclei are lysed to release template genomic DNA fragments and template RNA (e.g., cytoplasmic mRNA or nuclear mRNA) molecules. Continuing this embodiment, after fragmentation, template genomic DNA fragments are processed generally as outlined in **FIG. 29D** (or **FIG. 29C** for bulk tagmentation) by ligating a partially double-stranded nucleic acid barcode molecule as shown in **FIG. 29B** (see also **FIG. 35A**) to the template nucleic acid fragments (see, e.g., Examples 9 and 10). Template RNA molecules (e.g., mRNA molecules) are processed to generate 5' barcoded cDNA molecules. For example, a template RNA molecule comprising a poly-A tail (e.g., mRNA) and a nucleic acid molecule comprising a poly-T sequence are processed using the reverse transcriptase to generate a cDNA molecule. In an example of template switching, in some embodiments, an enzyme with terminal transferase activity (e.g., a reverse transcriptase with terminal transferase activity) adds additional nucleotides, (e.g., polyC), to the cDNA in a template independent manner. Switch oligonucleotides or switch oligonucleotide sequences can include sequences complementary to the additional nucleotides, e.g., polyG. The additional nucleotides (e.g., polyC) on the cDNA hybridize to the additional nucleotides (e.g., polyG) on the switch oligo, whereby the switch oligo can be used by the reverse transcriptase as template to further extend the cDNA. Thus, in some embodiments, the reverse transcriptase comprises terminal transferase activity and generates 5' barcoded cDNA fragments using the switch oligonucleotide sequence in the plurality of nucleic acid barcode molecules of **FIG. 35D**.

Exemplary Scheme 3 – scATAC-seq + 3' scRNA-seq using linear amplification

[00364] In certain barcoding schemes, cells from a cell population of interest (or intact nuclei from cells in a cell population of interest) are partitioned into a plurality of partitions (e.g., droplets) such that at least some partitions comprise: (1) a single cell (or a single nucleus) comprising template genomic DNA molecules and template RNA molecules (e.g., mRNA or nuclear pre-mRNA); (2) a solid or semi-solid particle (e.g., a bead, such as a gel bead) comprising (i) a plurality of single-stranded nucleic acid barcode molecules as shown in **FIG. 35C**, and (ii) a plurality of nucleic acid barcode molecules as shown in **FIG. 35B**; (3) a plurality of transposase molecules; (4) a plurality of nucleic acid molecules comprising a transposase end sequence (e.g., **FIG. 29A**); and (5) a plurality of reverse transcriptase molecules. In some embodiments, the solid or semi-solid particle is a bead. In some embodiments, the bead is a gel bead. In some embodiments, the solid or semi-solid particle (e.g., gel bead) comprises the plurality of single-stranded nucleic acid barcode molecules of **FIG. 35C** attached thereto and a

plurality of magnetic particles embedded within the solid or semi-solid particle (e.g., gel bead) (see, e.g., **FIG. 34**), wherein the plurality of magnetic particles comprises, attached thereto, the plurality of nucleic acid barcode molecules of **FIG. 35B**. In other embodiments, the solid or semi-solid particle (e.g., gel bead) comprises the plurality of nucleic acid barcode molecules of **FIG. 35B** attached thereto and a plurality of magnetic particles embedded within the solid or semi-solid particle (e.g., gel bead), wherein the plurality of magnetic particles comprises, attached thereto, the plurality of nucleic acid barcode molecules of **FIG. 35C**. In some embodiments, the barcode sequence in **FIG. 35C** and the barcode sequence **FIG. 35B** is the same. In some embodiments, the barcode sequence in **FIG. 35C** and the barcode sequence **FIG. 35B** comprise one or more barcode segments that are identical. In some embodiments, the barcode sequence in **FIG. 35C** and the barcode sequence **FIG. 35B** are different.

[00365] Continuing these embodiments, after partitioning, the single cells (or nuclei) are lysed to release template genomic DNA and template RNA (e.g., cytoplasmic mRNA or nuclear mRNA) in a manner that substantially maintains native chromatin organization of the genomic DNA. In certain embodiments, the plurality of nucleic acid barcode molecules of **FIG. 35C** and **FIG. 35D** are releasably attached to the solid or semi-solid particle (e.g., bead, such as a gel bead) and partitions (e.g., droplets or wells) are subjected to conditions to cause release of barcode oligonucleotide molecules from the solid or semi-solid particle (e.g., bead) (e.g., depolymerization or degradation of a bead, for example, using a reducing agent such as DTT). Partitions (e.g., droplets or wells) are then subjected to conditions to generate a transposase-nucleic acid complex as described in the aforementioned examples. Alternatively, in some embodiments, a plurality of pre-formed transposase-nucleic acid complexes (e.g., **FIG. 29A**) are partitioned into the plurality of partitions (e.g., droplets or wells). Partitions are then subjected to conditions such that the transposase-nucleic acid complexes generate double-stranded template genomic DNA fragments. Alternatively, in some embodiments, the transposition reaction is performed in bulk in intact nuclei which are partitioned such that at least some partitions (e.g., droplets or wells) comprise (1) a single cell (or a single nucleus) comprising template genomic DNA fragments and template RNA molecules; (2) a solid or semi-solid particle (e.g., bead, such as a gel bead) comprising (i) a plurality of single-stranded nucleic acid barcode molecules as shown in **FIG. 35C**, and (ii) a plurality of nucleic acid barcode molecules as shown in **FIG. 35D**; and (3) a plurality of reverse transcriptase molecules. In some embodiments, the nucleic acid barcode molecules of **FIG. 35C** or **FIG. 35D** are attached to a magnetic particle embedded within a solid or semi-solid particle (e.g., bead, such as a gel bead), as previously described. After partitioning, the single nuclei are lysed to release template genomic DNA fragments and template RNA (e.g., cytoplasmic mRNA or nuclear mRNA) molecules. Continuing this

embodiment, after fragmentation, template genomic DNA fragments are processed generally as outlined in **FIG. 31** (or **FIG. 30B** for bulk tagmentation) by performing a linear amplification of template nucleic acid fragments (which in some embodiments, are gap-filled prior to amplification) using nucleic acid barcode molecules of **FIG. 35C** (see, *e.g.*, Examples 11 and 12). Using the reverse transcriptase, template RNA molecules comprising a poly-A tail (*e.g.*, mRNA molecules) are processed with the plurality of nucleic acid barcode molecules of **FIG. 35B** to generate barcoded cDNA molecules as generally described elsewhere herein.

Exemplary Scheme 4 – scATAC-seq + 5' scRNA-seq using linear amplification

[00366] In another barcoding scheme, cells from a cell population of interest (or intact nuclei from cells in a cell population of interest) are partitioned into a plurality of partitions (*e.g.*, droplets or wells) such that at least some partitions comprise: (1) a single cell (or a single nucleus) comprising template genomic DNA molecules and template RNA molecules (*e.g.*, mRNA or nuclear pre-mRNA); (2) a solid or semi-solid particle (*e.g.*, bead, such as a gel bead) comprising (i) a plurality of single-stranded nucleic acid barcode molecules as shown in **FIG. 35C**, and (ii) a plurality of nucleic acid barcode molecules as shown in **FIG. 35D**; (3) a plurality of transposase molecules; (4) a plurality of nucleic acid molecules comprising a transposase end sequence (*e.g.*, **FIG. 29A**); (5) a plurality of nucleic acid molecules comprising a poly-T sequence; and (6) a plurality of reverse transcriptase molecules. In some embodiments, the solid or semi-solid particle is a bead. In some embodiments, the bead is a gel bead. In some embodiments, the solid or semi-solid particle (*e.g.*, gel bead) comprises the plurality of single-stranded nucleic acid barcode molecules of **FIG. 35C** attached thereto and a plurality of magnetic particles embedded within the solid or semi-solid particle (*e.g.*, gel bead) (see, *e.g.*, **FIG. 34**), wherein the plurality of magnetic particles comprises, attached thereto, the plurality of nucleic acid barcode molecules of **FIG. 35D**. In other embodiments, the solid or semi-solid particle (*e.g.*, gel bead) comprises the plurality of nucleic acid barcode molecules of **FIG. 35D** attached thereto and a plurality of magnetic particles embedded within the solid or semi-solid particle (*e.g.*, gel bead), wherein the plurality of magnetic particles comprises, attached thereto, the plurality of nucleic acid barcode molecules of **FIG. 35C**. In some embodiments, the barcode sequence in **FIG. 35C** and the barcode sequence **FIG. 35D** is the same. In some embodiments, the barcode sequence in **FIG. 35C** and the barcode sequence **FIG. 35D** comprise one or more barcode segments that are identical. In some embodiments, the barcode sequence in **FIG. 35C** and the barcode sequence **FIG. 35D** are different.

[00367] Continuing these embodiments, after partitioning, the single cells (or nuclei) are lysed to release template genomic DNA and template RNA (*e.g.*, cytoplasmic mRNA or nuclear mRNA) in a manner that substantially maintains native chromatin organization of the genomic

DNA. In certain embodiments, the plurality of nucleic acid barcode molecules of **FIG. 35C** and **FIG. 35D** are releasably attached to the solid or semi-solid particle (e.g., bead, such as a gel bead) and partitions (e.g., droplets or wells) are subjected to conditions to cause release of barcode oligonucleotide molecules from the solid or semi-solid particle (e.g., bead) (e.g., depolymerization or degradation of a bead, for example, using a reducing agent such as DTT). Partitions (e.g., droplets or wells) are then subjected to conditions to generate a transposase-nucleic acid complex as described in the aforementioned examples. Alternatively, in some embodiments, a plurality of pre-formed transposase-nucleic acid complexes (e.g., **FIG. 29A**) are partitioned into the plurality of partitions (e.g., droplets or wells). Partitions are then subjected to conditions such that the transposase-nucleic acid complexes generate double-stranded template genomic DNA fragments. Alternatively, in some embodiments, the transposition reaction is performed in bulk in intact nuclei which are partitioned such that at least some partitions comprise (1) a single cell (or a single nucleus) comprising template genomic DNA fragments and template RNA molecules; (2) a solid or semi-solid particle (e.g., bead, such as a gel bead) comprising (i) a plurality of single-stranded nucleic acid barcode molecules as shown in **FIG. 35C**, and (ii) a plurality of nucleic acid barcode molecules as shown in **FIG. 35D**; (3) a plurality of nucleic acid molecules comprising a poly-T sequence; and (4) a plurality of reverse transcriptase molecules. After partitioning, the single nuclei are lysed to release template genomic DNA fragments and template RNA (e.g., cytoplasmic mRNA or nuclear mRNA) molecules. Continuing this embodiment, after fragmentation, template genomic DNA fragments are processed generally as outlined in **FIG. 31** (or **FIG. 30B** for bulk tagmentation) by performing a linear amplification of template nucleic acid fragments (which in some embodiments, are gap-filled prior to amplification) using nucleic acid barcode molecules of **FIG. 35C** (see, e.g., Examples 11 and 12). Template RNA molecules (e.g., mRNA molecules) are processed to generate 5' barcoded cDNA molecules as previously described. For example, a template RNA molecule comprising a poly-A tail (e.g., mRNA) and a nucleic acid molecule comprising a poly-T sequence are processed using a reverse transcriptase (e.g., a reverse transcriptase with terminal transferase activity) to generate a cDNA molecule comprising additional nucleotides on the 5' end (e.g., poly-C). The additional nucleotides on the cDNA hybridize to the additional nucleotides (e.g., poly-G) on the switch oligo, whereby the switch oligonucleotide sequence in the plurality of nucleic acid barcode molecules of **FIG. 35D** are used to further extend the cDNA to generate the 5' barcoded cDNA molecules.

[00368] In the aforementioned embodiments, after barcoding, the barcoded, adapter-flanked DNA fragments and the barcoded cDNA molecules are then released from the partitions (e.g., droplets or wells) to provide a released mixture and processed in bulk to complete library

preparation for next generation high throughput sequencing (*e.g.*, subjecting the fragments, or derivatives thereof, to one or more reactions (*e.g.*, nucleic acid amplification) to add functional sequences to facilitate Illumina sequencing). In some embodiments, a first portion of the released mixture comprising the adapter-flanked DNA fragments and the barcoded cDNA is taken and processed in bulk to complete library preparation for the barcoded, adapter-flanked DNA fragments while a second portion of the released mixture is taken and processed in bulk to complete library preparation for the barcoded cDNA molecules. In other embodiments, a first portion of the partitions (*e.g.*, droplets or wells) comprising the barcoded, adapter-flanked DNA fragments and the barcoded cDNA molecules is taken and processed in bulk to complete library preparation for the barcoded, adapter-flanked DNA fragments while a second portion of the partitions (*e.g.*, droplets or wells) comprising the barcoded, adapter-flanked DNA fragments and the barcoded cDNA molecules is taken and processed in bulk to complete library preparation for the barcoded cDNA molecules. In embodiments that utilize a magnetic particle (*e.g.*, bead), the barcoded template molecules (*e.g.*, barcoded template DNA fragments or barcoded cDNA molecules, or derivatives thereof) attached thereto can be magnetically separated and further processed to complete library preparation. The fully constructed library or libraries are then sequenced according to a suitable next-generation sequencing protocol (*e.g.*, Illumina sequencing).

Example 18. Effect of R1 Sequence Modifications on Barcode Exchange

[00369] Nuclei from mice and humans were tagmented and barcoded by ligation (see, *e.g.*, Example 9 and **FIGs. 29A-29C**) with one of three types of partially double-stranded barcodes, mixed, and subsequently subjected to detection in order to assess the effect of each type of barcode on the ability to distinguish the different types of nuclei. The three types of barcodes included a barcode with a standard R1 sequence (**FIG. 37**; top), a barcode with a shortened R1 sequence compared to the standard R1 sequence (**FIG. 37**; middle) and a barcode comprising a uracil in the R1 sequence (**FIG. 37**; bottom).

[00370] Compared to the separation of the different cell types seen in the barcodes with standard R1 sequences, the barcodes with the shortened R1 sequence and barcodes with a uracil showed greater separation of mice and human nuclei, indicating a reduced rate of barcode exchange (**FIGs. 38A-38C**).

Example 19. Reduction of mitochondrial DNA reads using a blocking agent

[00371] A plurality of cells is provided in which each cell comprises a template nucleic acid (*e.g.*, nuclear DNA, such as chromatin) and a non-template nucleic acid (*e.g.*, mitochondrial DNA). The plurality of cells is lysed in the presence of a cell lysis agent and a blocking agent

(e.g., bovine serum albumin) to generate a plurality of lysed cells. A plurality of nucleic acid is then separated from the plurality of lysed cells to generate a plurality of biological particles (e.g., nuclei), where a given biological particle of the plurality of biological particles comprises non-template nucleic acid molecules (e.g., mitochondrial DNA molecules) and template nucleic acid molecules. The template nucleic acid molecules may comprise, for example, DNA molecules and RNA molecules. A plurality of template nucleic acid fragments may then be generated in biological particles of the plurality of biological particles with the aid of a transposase-nucleic acid complex comprising a transposase nucleic acid molecule and a transposon end nucleic acid molecule. A plurality of partitions may then be generated, where a given partition comprises (i) a single biological particle comprising the plurality of template nucleic acid fragments and template RNA molecules and (ii) a plurality of first barcode oligonucleotide molecules (e.g., first nucleic acid barcode molecules) comprising a barcode sequence. The partitions may further comprise (iii) a plurality of second barcode oligonucleotide molecules (e.g., second nucleic acid molecules) comprising a barcode sequence and a capture sequence; and (iv) a plurality of reverse transcriptase molecules. The partitions may be, for example, droplets or wells. Barcoded template DNA fragments may then be generated within the partitions using barcode oligonucleotide molecules of the plurality of first barcode oligonucleotide molecules and template nucleic acid fragments of the plurality of template DNA fragments. If template RNA molecules are present, barcoded cDNA molecules may be generated from the template RNA molecules by reverse transcription using barcode oligonucleotide molecules of the plurality of second barcode oligonucleotide molecules. In some cases, the biological particles may be nuclei, which may be isolated from lysed cells by, e.g., washing the lysed cells. In some cases, the method further comprises generating sequencing reads. Application of the method may reduce the fraction of sequencing reads deriving from non-template nucleic acid molecules (e.g., mitochondrial DNA molecules) relative to the total number of sequencing reads generated.

Example 20. Reduction of mitochondrial DNA reads in partitions

[00372] A plurality of partitions (e.g., droplets or wells) is provided. A given partition of the plurality of partitions comprises: (i) a single biological particle (e.g., cell) from a plurality of biological particles (e.g., cells), wherein the single biological particle comprises template nucleic acid molecules (e.g., nuclear DNA molecules, such as chromatin) and non-template DNA molecules (e.g., mitochondrial DNA molecules); (ii) a plurality of barcode oligonucleotide molecules (e.g., nucleic acid barcode molecules) comprising a barcode sequence; (iii) a plurality of transposon end oligonucleotide molecules comprising a transposon end sequence; and (iv) a plurality of transposase molecules. The given partition may further comprise reverse

transcriptases and a plurality of second barcode oligonucleotide molecules (e.g., second nucleic acid barcode molecules) comprising a barcode sequence and a capture sequence. Barcoded template nucleic acid fragments and barcoded non-template nucleic acid fragments may be generated as described in the preceding examples. One or more non-template nucleic acid fragments of the plurality of non-template DNA fragments, or derivatives thereof, may then be cleaved using (i) one or more guide ribonucleic acid molecules (gRNAs) targeted to the one or more non-template nucleic acid fragments, and (ii) a clustered regularly interspaced short palindromic (CRISPR) associated (Cas) nuclease. In some cases, the method may reduce the amount of fragments and/or barcoded fragments comprising and/or deriving from non-template nucleic acid molecules. For example, the method may reduce the total number of non-template nucleic acid fragments and barcoded non-template nucleic acid fragments, e.g., in a mixture comprising one or more of template nucleic acid fragments, barcoded template nucleic acid fragments, non-template nucleic acid fragments, and barcoded non-template nucleic acid fragments. In some cases, the endonuclease may be Cas9, such as recombinant Cas9. In some cases, the non-template nucleic acid fragment may comprise a mitochondrial DNA or RNA fragment. In some cases, the template nucleic acid fragment may comprise a nuclear DNA fragment.

Example 21. Comparison of linear amplification and ligation methods

[00373] Sample libraries may be generated using any of the methods described in the preceding examples. In some cases, a library may be generated using a linear amplification method. In other cases, a library may be generated using a ligation method. A linear amplification method may comprise, within a partition, generating template nucleic acid fragments and performing a gap filling extension process within the partition. Linear amplification may then be performed using, for example, a heat resistant polymerase. Barcodes may be incorporated into sequences comprising template nucleic acid sequences, or complements thereof. The barcoded nucleic acid fragments may then be released from their respective partitions into the bulk for additional processing including, for example, PCR. A ligation method may comprise, within a partition, generating template nucleic acid fragments and ligating barcodes to the fragments with a ligase such as a T4 DNA ligase, thereby generating barcoded nucleic acid fragments. The barcoded nucleic acid fragments may then be released from their respective partitions into the bulk for additional processing including, for example, gap filling and addition of additional sequences.

[00374] In order to compare linear amplification and ligation methods, various mouse and human cells were processed according to the methods described herein.

[00375] **FIG. 40** shows a table comparing sequencing metrics generated from different replicate sample libraries (“A” and “B”) generated using either linear amplification or ligation. “A” replicates represent reactions done in the same conditions, but using a different user, while “B” replicates represent reactions done in the same conditions by the same user.

[00376] **FIGs. 41A-41B** illustrate exemplary amounts of detected mouse or human cells and the inferred doublet rate observed from the analysis of sequencing reads (**FIG. 40**) from a linear amplification ATAC-seq library (using 75 nM, 150nM, or 250 nM of barcode primer) or ligation-based ATAC-seq library (using the barcode molecules of **FIG. 37**). **FIG. 41A** illustrates differences in the number of mouse or human cells detected using either a linear amplification or ligation-based ATAC-seq method. **FIG. 41B** illustrate differences in inferred doublet rate detected using either a linear amplification or ligation-based ATAC-seq method. As shown in **FIG. 41B**, linear amplification demonstrated a higher doublet rate than ligation.

[00377] **FIG. 42** illustrates a comparison of the sensitivity of sequencing reads obtained from various ATAC-seq libraries as measured by the median fragments per cell barcode. The sensitivity of the linear amplification and ligation methods of library preparation (36k reads/cell) is compared to the methods described in Buenrostro, *et al.*, Nature, 2015 Jul 23;523(7561):486-90 using a programmable microfluidics platform (Fluidigm). As shown in **FIG. 42**, ligation provides higher sensitivity than linear amplification.

[00378] As shown in **FIG. 43**, a library prepared by ligation sequenced to increased depth (30M reads to 800M reads) demonstrates a significant increase in the sensitivity (as measured by median fragments per cell barcode) and reduced noise (as measured by fraction of non-duplicate wasted reads) relative to the lower depth library.

[00379] **FIG. 48** shows a comparison of exemplary sequencing metrics obtained from linear amplification-based ATAC-seq libraries prepared using different polymerases: a Phusion[®] DNA polymerase, a KAPA HiFi DNA polymerase (in combination with betaine), a Deep Vent[®] DNA polymerase, as well as a library prepared by ligation.

“Wasted” reads

[00380] As shown in **FIG. 44**, non-duplicate “wasted” reads may be generated while performing the methods described herein. **FIG. 44** illustrates an exemplary comparison of the total noise (non-duplicate wasted reads (“Non-dups”) and mitochondrial-based reads (“Mito”)) in libraries prepared using linear amplification or ligation-based ATAC-seq methods. **FIGs. 45A-45B** provide an exemplary illustration of the breakdown sequencing reads generated by different library preparation methods. **FIG. 45A** shows an illustration of the breakdown of reads generated by a linear amplification ATAC-seq library. **FIG. 45B** shows an illustration of the breakdown of reads generated by a ligation-based ATAC-seq library. As described above, reads

attributable to mitochondrial nucleic acids may be removed by the use of blocking agents such as bovine serum albumin and/or through the use of Cas9 and targeted gRNAs.

Insert size distribution

[00381] Linear amplification and ligation library preparation methods were also compared to examine insert size distributions. **FIGs. 46A-46B** illustrate an exemplary comparison of read pairs showing the periodicity of nucleosomes generated from ATAC-seq libraries prepared using either a these schemes. Nucleosome free fragments are typically observed below 200bp in length, fragments indicative of a nucleosome periodicity of one are approximately 200bp in length, fragments indicative of a nucleosome periodicity of two are approximately 400bp in length, fragments indicative of a nucleosome periodicity of two are 600bp in length, and so forth.

Bulk enrichment

[00382] As shown in **FIGs. 47A-47B**, linear amplification shows a higher enrichment of transcription start sites (TSS) than ligation due to a bias for shorter insert fragments. A difference may also be observed between linear amplification and ligation methods for CTCF (CCCTC-binding factor) sites.

Example 22. Analysis of nucleic acids in peripheral blood mononuclear cell samples

[00383] Peripheral blood mononuclear cells (PBMCs) are subjected to single cell RNA-seq and ATAC-seq analyses as described herein.

[00384] **FIG. 51A** shows an exemplary scatterplot produced using t-Distributed Stochastic Neighbor Embedding (tSNE), allowing visualization of RNA transcripts of different subpopulations cell types in a peripheral blood mononuclear cell (PBMC) sample. **FIG. 51B** shows an exemplary scatterplot produced using t-Distributed Stochastic Neighbor Embedding (tSNE), allowing visualization of ATAC-seq data of different subpopulations cell types in a peripheral blood mononuclear cell (PBMC) sample.

Example 23. Comparison of ATAC-seq methods

[00385] **FIG. 50** illustrates protocols for ATAC-seq analyses as described herein compared to data from (1) typical high quality traditional bulk ATAC-seq protocols; (2) Cusanovich, *et al.*, Science, 2015 May 22;348(6237):910-14; (3) Buenrostro, *et al.*, Nature, 2015 Jul 23;523(7561):486-90; (4) ideal sequencing metrics from an ATAC-seq experiment; and (5) data obtained using the methods described herein ("10X").

[00386] While preferred embodiments of the present invention have been shown and described herein, it will be obvious to those skilled in the art that such embodiments are provided by way of example only. It is not intended that the invention be limited by the specific examples provided within the specification. While the invention has been described with reference to the

aforementioned specification, the descriptions and illustrations of the embodiments herein are not meant to be construed in a limiting sense. Numerous variations, changes, and substitutions will now occur to those skilled in the art without departing from the invention. Furthermore, it shall be understood that all aspects of the invention are not limited to the specific depictions, configurations or relative proportions set forth herein which depend upon a variety of conditions and variables. It should be understood that various alternatives to the embodiments of the invention described herein may be employed in practicing the invention. It is therefore contemplated that the invention shall also cover any such alternatives, modifications, variations or equivalents. It is intended that the following claims define the scope of the invention and that methods and structures within the scope of these claims and their equivalents be covered thereby.

WHAT IS CLAIMED IS:

1. A method of generating barcoded nucleic acid fragments, comprising:
 - (a) providing: (i) a plurality of cells or cell nuclei comprising chromatin comprising a template nucleic acid molecule; (ii) a plurality of oligonucleotide molecules comprising a transposon end sequence; and (iii) a plurality of transposase molecules;
 - (b) in said plurality of cells or cell nuclei, generating a plurality of template nucleic acid fragments with the aid of a plurality of transposase-nucleic acid complexes comprising a transposase molecule of said plurality of transposase molecules and an oligonucleotide molecule of said plurality of oligonucleotide molecules;
 - (c) partitioning said plurality of cells or cell nuclei into a plurality of partitions, each comprising (i) a cell or a cell nucleus of said plurality of cells or cell nuclei, said cell or cell nucleus comprising a template nucleic acid fragment of said plurality of template nucleic acid fragments, and (ii) a plurality of nucleic acid barcode molecules comprising a common barcode sequence; and
 - (d) in a partition of said plurality of partitions, generating a barcoded nucleic acid fragment using a nucleic acid barcode molecule of said plurality of nucleic acid barcode molecules and the template nucleic acid fragment of said plurality of template nucleic acid fragments.
2. The method of claim 1, wherein said barcoded nucleic acid fragment is generated by ligation of said nucleic acid barcode molecule with said template nucleic acid fragment, or a derivative thereof, or wherein said barcoded nucleic acid fragment is generated by nucleic acid amplification of said template nucleic acid fragment, or a derivative thereof, using said nucleic acid barcode molecule as a primer.
3. The method of claim 1 or claim 2, wherein said plurality of nucleic acid barcode molecules are attached to a bead and wherein said partition further comprises said bead.
4. The method of claim 3, wherein said nucleic acid barcode molecules are releasably attached to said bead.

5. The method of claim 4, further comprising releasing said plurality of nucleic acid barcode molecules from said bead.
6. The method of any one of claims 3 to 6, wherein said bead is a degradable gel bead, wherein said degradable gel bead is degradable upon application of a stimulus.
7. The method of any one of claims 1 to 6, wherein subsequent to (c), said template nucleic acid fragments are released from said cells or cell nuclei.
8. The method of any one of claims 1 to 7, wherein said partition is an aqueous droplet in an emulsion, or a well.
9. The method of any one of claims 1 to 8, further comprising: (i) releasing or removing said barcoded nucleic acid fragment or a derivative thereof from said partition, or (ii) sequencing said barcoded nucleic acid fragment, or a derivative thereof.
10. The method of any one of claims 1 to 9, wherein said plurality of oligonucleotide molecules comprise a plurality of first nucleic acid molecules comprising a transposon end sequence and a sequence complementary to a sequence of said plurality of nucleic acid barcode molecules, or wherein said cells or cell nuclei further comprises a template RNA molecule, and wherein the method further comprises generating a barcoded cDNA molecule from said template RNA molecule or a derivative thereof.
11. The method of claim 10, wherein said plurality of oligonucleotide molecules further comprise a plurality of second nucleic acid molecules comprising a transposon end sequence and a primer sequence, or wherein said plurality of nucleic acid barcode molecules are (A) partially double-stranded and comprise (i) a first strand comprising said common barcode sequence and a sequence complementary to a sequence of said plurality of first nucleic acid molecules; and (ii) a second strand comprising a sequence complementary to said common barcode sequence or (B) single-stranded and comprise said common barcode sequence and a sequence complementary to a sequence of said plurality of first nucleic acid molecules.

12. The method of claim 10, wherein said barcoded cDNA molecule is generated in said partition, or wherein said partition comprises a second plurality of nucleic acid barcode molecules comprising said common barcode sequence and a capture sequence.
13. The method of any one of claims 1 to 12, further comprising (a) lysing a plurality of cells in the presence of a cell lysis agent and a blocking agent; and (b) separating a plurality of nuclei from said plurality of lysed cells to generate said plurality of cells or cell nuclei, wherein said blocking agent reduces the fraction of sequencing reads corresponding to a mitochondrial genome compared to sequencing reads obtained in the absence of said blocking agent.
14. The method of any one of claims 1 to 13, further comprising cleaving one or more mitochondrial nucleic acid molecules or derivatives thereof, using (i) one or more guide ribonucleic acid molecules (gRNAs) targeted to said one or more mitochondrial nucleic acid fragments; and (ii) a clustered regularly interspaced short palindromic (CRISPR) associated (Cas) nuclease.